Chapter 8 Molecular Markers and Thyroid Nodule Evaluation

Trevor E. Angell, Matthew I. Kim, and Erik K. Alexander

Introduction

Thyroid nodules are common, and incidental detection is increasingly prevalent. While nodules are most often asymptomatic and benign, the guiding clinical concern is the risk of possible thyroid carcinoma. Numerous studies from large nodule populations confirm the risk of malignancy to be approximately 8–15%. Thus, the principle goal of thyroid nodule evaluation is to accurately, efficiently, and safely determine which nodules are benign and which are cancerous, ideally through a simple and cost-effective process. Over the past 30 years, this clinical care paradigm has evolved to address limitations while seeking to improve care.

The approach to thyroid nodule evaluation is multidisciplinary, integrating radiologic, cytologic, clinical, and molecular analysis. No single means of evaluation can effectively determine if a nodule represents benign or malignant disease with high accuracy. However, an integrated approach provides a more accurate means of cancer detection. Most recently, this accuracy has been improved through our understanding that specific molecular markers or molecular profiles of thyroid nodules correlate with benign or malignant disease. The clinical application of these markers has reduced unnecessary surgery while enhancing understanding of individualized prognosis.

Molecular analysis is a broad term, encompassing many different types of molecular testing. Specific mutations in cellular DNA reflect a base pair change or gene translocation that affects the function of an oncogenic pathway. Messenger RNA (mRNA) expression profiles identify patterns of multiple expressed genes that

T.E. Angell, MD • M.I. Kim • E.K. Alexander, MD (🖂)

Thyroid Section, Division of Endocrinology, Diabetes, and Hypertension, The Brigham & Women's Hospital and Harvard Medical School, 75 Francis Street, Room PBB-B4, Boston, MA 02115, USA e-mail: ekalexander@partners.org

[©] Springer International Publishing AG 2018

H. Gharib (ed.), *Thyroid Nodules*, Contemporary Endocrinology, DOI 10.1007/978-3-319-59474-3_8

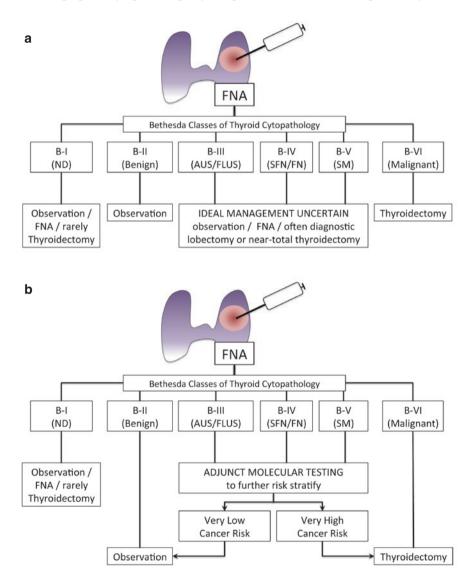
correspond to benign or malignant processes. MicroRNA are small non-translated modifiers of mRNA expression and translation, and individual or groups of microRNA may show differential levels of expression in benign or malignant processes. Finally, evaluation by immunohistochemistry (IHC) can determine the presence (or absence) at the cellular level of expressed proteins that are associated with benign or malignant disease. These four categories represent diverse and distinct mechanisms of molecular diagnostic testing, and each has shown the ability to assist in the clinical care of affected patients.

This chapter will review the typical approach to thyroid nodule evaluation. In euthyroid individuals, ultrasound (US) evaluation and ultrasound-guided FNA (UG-FNA) remain the principle first step of evaluation. However, a substantial limitation to this approach surrounds the 20–25% of nodules that prove to have indeterminate cytology. Traditionally, this finding has most often led to diagnostic surgery. The use of molecular markers has significantly addressed this shortcoming, allowing improved preoperative risk assessment (shown in Fig. 8.1). Through this, unnecessary surgery for benign nodules has been reduced. This chapter will conclude with discussion of forward-looking areas of necessary research, as we seek to expand our knowledge of this increasingly common disease.

Fig. 8.1 Management of a thyroid nodule based on fine needle aspiration biopsy results. (a) Historical management decisions for thyroid nodules: fine needle aspiration (FNA) biopsy provides a thyroid tissue specimen for cytopathologic evaluation and placement into one of six categories according to the Bethesda system for reporting thyroid cytopathology, which imply specific risk of malignancy. Benign and malignant cytologies are considered sufficiently accurate to recommend nodule observation or thyroidectomy, in these two situations, respectively. For nodules that are indeterminate, considered to be Bethesda classes III (AUS/FLUS), IV (SFN/FN), and V (suspicious for malignancy), the optimal management remains uncertain. In the former two categories, malignancy risk is low but not excluded, and observation may miss a clinically relevant malignancy, while surgical resection may be unnecessary. For nodules with Bethesda V cytology, malignant risk is often sufficiently high to warrant surgical resection (60-70%), but the extent of surgery (lobectomy versus near-total thyroidectomy) is uncertain. Nodules with non-diagnostic (ND) cytology are low risk (1-4% cancer risk) and can typically be managed non-operatively, though a subset of repeatedly ND nodules will be removed. (b) Current recommended schema of thyroid nodule management using adjunct molecular testing: for nodules with indeterminate cytologies, further evaluation with molecular testing allows for classification as low or high risk for malignancy. When test performance has been shown in prospective, multicenter, blinded validation studies to have a sufficiently high negative predictive value (NPV), indicating a very low chance of a "false-negative" or missed cancer, nodule observation can be selected in lieu of diagnostic lobectomy avoiding an unnecessary surgery. Conversely, when a test with very high positive predictive value (PPV) indicates the presence of cancer, thyroidectomy is indicated, and near-total thyroidectomy as initial surgery rather than diagnostic lobectomy may be appropriately performed. The management of ND, benign, and malignant nodules is unchanged with current molecular testing. B = Bethesda classification. Dx = diagnostic. AUS/FLUS = atypia of undetermined significance/follicular lesion on undetermined significance. SFN = suspicious for follicular neoplasm/follicular neoplasm. SM = suspicious for malignancy

The Approach to Thyroid Nodules

Most thyroid nodules are asymptomatic except as a palpable mass detected by the patient or medical practitioner, and may be discovered incidentally by cross-section imaging [1, 2]. Most recent large series of consecutive nodules suggest that approximately 8–15% of thyroid nodules will prove to be malignant [3–5]. Clinical factors such as sex, age [6], childhood exposure to ionizing radiation [7], and rarely symptoms or palpable lymphadenopathy are present that increase the possibility that a



thyroid nodule will be malignant. Thus, a full medical history and physical examination should be obtained on all patients who present for evaluation of a thyroid nodule.

Ultrasound is the optimal imaging technique for thyroid nodule evaluation. Beyond measurement of nodule size, sonographic features can assist with nodule risk assessment [8]. However, low to moderate inter-rater reproducibility of findings limits the precision and accuracy of interpretation [9]. For this reason, UG-FNA to obtain material for cytopathologic evaluation is recommended for most thyroid nodules larger than 1–2 cm [10]. Although there is high diagnostic accuracy of benign and malignant cytology, ~20–25% of aspirates will be cellularly sufficient but cytologically indeterminate. The Bethesda classification system effectively subdivides indeterminate cytologic findings into distinct categories associated with escalating risk of malignancy that improve the ability of clinicians to stratify the risk of thyroid malignancy [11–14], but inter-rater and intra-rater reliability of cytologic interpretation remains poor [15].

Because the risk of thyroid malignancy is generally low but not excluded in these cases, key considerations are whether such a nodule can be monitored conservatively or should be surgically removed and what extent of surgery (thyroid lobectomy or near-total thyroidectomy) is most appropriate [10, 16, 17]. These options carry risks and benefits, and the optimal management remains uncertain, in part, because of the limitations inherent to the clinical, sonographic, and cytologic assessments described above, indicating the need for synergistic forms of assessment, such as molecular testing, for such nodules.

Molecular analysis of nodular tissue has proved highly valuable. As price points for these diagnostic tests have fallen, cost-effectiveness analyses also suggest that the use of diagnostic molecular tests may reduce cost while improving quality-adjusted life years [18, 19]. Thus, the approach to cytologically indeterminate thyroid nodules is increasingly employing molecular diagnostic testing. Below, we provide a description of the molecular markers thus far evaluated for use in the care of patients with clinically relevant thyroid nodules.

Molecular Markers

A molecular marker assessed on fine needle aspiration material from a thyroid nodule biopsy may target one of many cellular constituents, such as expressed proteins, RNA, or DNA, measured either quantitatively or qualitatively using specific laboratory techniques. Some of the earliest investigations of molecular markers utilized IHC techniques on slides prepared from aspiration material to identify expressed proteins differentiating benign from malignant nodules. The identification of prevalent oncogenic mutations/translocations in thyroid cancer, including *BRAF*, *RAS*, and *RET/PTC*, led to intense focus on the DNA alterations in thyroid cancer, followed closely by advanced nucleic acid sequencing techniques allowing expression profiling of mRNA or microRNA, all of which have been investigated as molecular markers for thyroid nodule evaluation. Limitations inherent with the use of any single molecular marker have in some cases spawned the use of combinations of molecular tests improving test accuracy.

In each case, the process begins with identifying a possible molecular marker and creating a laboratory test to reliably detect it, which allows understanding of the test's analytic validity. How well the marker discriminates benign from malignant nodules, especially in clinical practice, determines the test's clinical validity. Whether or not knowing molecular marker status changes patient management when incorporated into clinical decision-making establishes its clinical utility and is important for determining the clinical value of the testing. Finally, the costeffectiveness of using the molecular tests compared to other management requires evaluation within any large healthcare framework. Each of these aspects must be considered in the evaluation of a molecular marker as it is applied to clinical medicine. While the molecular markers described in this chapter (Table 8.1) have largely shown consistent analytic performance, the clinical validity, clinical utility and costeffectiveness frequently remain unknown.

Galectin-3

Galectin-3 is a lectin that binds to cell surface glycoproteins and interacts with intracellular proteins regulating cellular functioning including cell growth, apoptosis, and malignant transformation [20]. Its presence has been measured by IHC using a monoclonal antibody targeting galectin-3 on formalin-fixed and paraffin-embedded (FFPE) cellblock preparations from thyroid nodule aspires. Early investigations found that galectin-3 expression correlated strongly with thyroid cancers compared to benign specimens [21, 22].

Table 8.1 Molecular markers Image: Comparison of the second s	Immunohistochemistry
	Galectin-3
	HBME-1
	CK19
	Mutations and gene rerrangements
	BRAF (V600E)
	RAS (H-RAS, K-RAS, N-RAS)
	RET/PTC (RET/PTC1, RET/PTC3)
	PAX8/PPARγ
	Asuragen miRInform [™] thyroid panel
	ThyroSeq v2 panel
	Gene expression and microarray analysis
	MicroRNA expression
	Afirma [®] gene expression classifier

Assessment of galectin-3 as a clinical test to improve diagnostic accuracy in cytologically indeterminate thyroid nodules has been performed. In a large multicenter prospective study, galectin-3 expression was assessed in 465 nodules with indeterminate cytology with histopathology available for blinded review and showed a sensitivity of 78% and specificity of 93%, yielding an 82% positive predictive value (PPV) and a 91% negative predictive value (NPV) in this population of patients [23]. While galectin-3 remains a marker of interest, there has not been more extensive validation of the test or robust data on how testing effects clinical care, and its initial performance has not been replicated. For these reasons, as well as the interpreter subjectivity surrounding its use, galectin-3 has shown greater promise in histopathologic assessment as opposed to cytologic assessment.

HBME-1

HBME-1 (Hector Battifora Mesothelial-1) is a monoclonal antibody targeting the microvilli of mesothelioma cells. However, the antigen recognized by HBME-1 has also been detected on differentiated thyroid cancer cells, leading to its development as a molecular marker for assessing malignancy risk. HBME-1 is measured by IHC on FFPE specimens. Preliminary studies evaluating HBME-1 on cytologically indeterminate thyroid nodules have shown moderate test performance, with sensitivity 79–94%, specificity 83–94%, PPV 84–94%, and NPV 80–94% [24, 25], but these have not been reproduced in prospective, multicenter, blinded investigations, and thus, validation of HBME-1 staining in such a population of indeterminate thyroid nodules remains to be seen.

СК19

Cytokeratin-19 (CK19) is a cytoskeleton component of epithelial cells that may be upregulated in well-differentiated thyroid cancer [26]. In one study of cytologically indeterminate nodules, most nodules ultimately proven to be papillary thyroid carcinoma stained positive for CK19; however, there was extensive overlap in CK19 expression between benign follicular adenomas and follicular thyroid carcinomas [27]. There are no high-quality data that support the sole use of CK19 as a single molecular marker at this time.

Combined Immunohistochemical Panels

Because of the limitations of individual protein markers, further studies have evaluated the potential for improved diagnostic accuracy when such markers are combined. An assessment of galectin-3 and HBME-1 together showed a PPV and NPV of 76.9% and 96.9% when the markers were concordant in their result [28]. Several studies have utilized the combinations of galectin-3, HBME-1, and CK19 [27, 29–31]. The largest of these evaluating 125 thyroid nodule FNA samples demonstrated an increase in sensitivity from 92% to 97% but a decrease in specificity from 96% to 80% compared to the performance of any marker in isolation [27]. Alternative targets have been included in separate panels of protein markers. One retrospective evaluation combining galectin-3, HBME-1, and CXCR4 showed little improvement compared to HBME-1 testing alone [25]. There have been no robust confirmatory data from large-scale clinical trials to validate these findings.

Mutations and Gene Rearrangements

The discovery of oncogenic mutations and gene rearrangements in thyroid cancers revealed that a significant percentage contain one of a small number of "driver" oncogenic events. Assessment of these mutations and translocations as a "rule-in" test for malignancy for indeterminate thyroid nodules soon followed. As our knowledge of the genetic landscape of thyroid cancer has grown, such molecular tests have evolved from detecting single mutations to assessing combinations of known genetic alterations.

BRAF

The *BRAF* gene encodes for the protein BRAF, a serine/threonine protein kinase in the MAPK pathway involved in many cellular processes including cell proliferation, differentiation, and apoptosis [32]. The most relevant *BRAF* mutation seen in thyroid cancers encodes for a mutated protein with a glutamic acid for valine substitution at position 600 (V600E) that causes unregulated kinase activation. BRAFV600E is the most common mutational event in classical PTC and is present in a significant minority of tumors classified as the follicular variant of papillary thyroid carcinoma (fvPTC) [33, 34]. Since this BRAF mutation is not present in benign thyroid disease, detection in FNA material essentially confirms the presence of thyroid cancer. BRAF mutations are less frequent in fvPTC and have a very low prevalence of 1-2% in follicular thyroid carcinoma (FTC). It lacks sufficient sensitivity to be a sole molecular marker for evaluating indeterminate nodules [35], especially because most BRAF-positive papillary thyroid carcinomas will demonstrate cytology that is "positive" or diagnostic of carcinoma (as opposed to cytologically indeterminate). Thyroid cancers harboring the BRAFV600E mutation are associated with a higher risk of extrathyroidal extension, lymph node metastasis, tumor recurrence, and patient mortality, and therefore, its presence may indicate the need for more aggressive initial therapy, though more data are needed to support specific treatment recommendations [35–37].

RAS

The *RAS* oncogene family is comprised of three genes (*H-RAS*, *K-RAS*, and *N-RAS*) that encode small GTPase proteins involved in signal transduction. Activating mutations of these genes stimulate the MAPK and phosphatidyl-3-inositol pathways that regulate cell growth, proliferation, differentiation, mobility, and survival [38]. *RAS* mutations are detected in up to 40% of differentiated thyroid cancers, occurring most often in fvPTC, noninvasive follicular thyroid neoplasms with papillary-like nuclear features (NIFT-P), and FTC [30], but are also present in benign thyroid lesions, such as follicular adenomas [39, 40] similarly making *RAS* status suboptimal as a single molecular marker. It may be that the presence of a *RAS* mutation portends a risk of malignant transformation, suggesting that resection of a thyroid nodule on the basis of a *RAS* mutation may still be appropriate [41]. However, data showing the stability and lack of malignant transformation of cytologically benign yet *RAS* mutation-positive thyroid nodules during long-term sonographic monitoring does not support this [42].

RET/PTC and PAX8/PPARy Translocations

The *RET* proto-oncogene encodes a receptor tyrosine kinase that has been found to undergo intrachromosomal gene rearrangements forming fusion genes in differentiated thyroid cancer. The *RET/PTC1* and *RET/PTC3* gene rearrangements produce constitutively active kinase activity that stimulates the MAPK and phosphatidyl-3-inositol pathways [43]. Because of its relative rarity compared to *BRAF* or *RAS* mutations and the overlap with benign thyroid neoplasms, testing for these translocations has limited value in isolation [44].

Another interchromosomal gene rearrangement linking the *PAX8* transcription factor gene with the nuclear hormone receptor gene *PPAR* γ produces the *PAX8*/*PPAR* γ fusion gene, possibly inhibiting the antiproliferative activity of the PPAR γ receptor. This gene rearrangement has been detected in 20–40% of follicular thyroid carcinomas and a lower percentage of Hurthle cell carcinomas, as well as follicular adenomas, but has insufficient sensitive or specificity as a molecular marker [45].

Combination Assessment of Oncogenes

Each of these single mutations or translocations has limited diagnostic values, but together genetic alterations in these genes are found in up to 70% of histologically proven thyroid cancers, indicating that if assessed together, better diagnostic discrimination may be achieved. In an early study, *BRAF*, *RAS*, *RET/PTC*, and *PAX8/PPAR* γ genes were assessed in 470 consecutive FNA samples with 55 of indeterminate cytology,

showing that while nearly all nodules testing positive were malignant, in some thyroid cancers, no mutations were detected [46]. This indicates potential value as a "rule-in" test to identify cancers but insufficient sensitivity to "rule-out" thyroid cancer.

These and similar results led to continued development and commercialization of a diagnostic test measuring relevant genetic alterations (about 17 in total) in the *BRAF*, *RAS*, *RET/PTC*, and *PAX8/PPAR* γ genes (marketed as miR*Inform* Thyroid, Asuragen, Inc.). Independent investigation and use in the larger clinical context have demonstrated positive and negative predictive values that were more variable and less robust than the initial report [47–52].

The results of the integrated genomic characterization of papillary thyroid cancer as part of The Cancer Genome Atlas (TCGA) expanded our knowledge of the somatic genetic landscape present in PTC and identified the driver mutation/translocation in 96% of PTC tumors studied [53]. Inclusion of these additional oncogenes may help identify more thyroid cancers and improve molecular testing of thyroid nodules. Informed by these data, a diagnostic test was produced using nextgeneration sequencing techniques to assess point mutations in 13 genes and 42 gene fusions (marketed as ThyroSeq v2). This testing involves additional aspiration material that is separately rinsed in a specific processing solution. Two prospective single-center unblinded studies evaluated its performance on FNA material from cytologically indeterminate thyroid nodules. In 143 consecutive FNA samples with cytology of SFN/FN and available histopathology after resection, ThyroSeq v2 showed a high NPV of 96% (CI 92–100%) and moderate PPV of 83% (CI 72–95%) [54]. In a similar study assessing the performance of ThyroSeq v2 on 98 nodules with cytology of AUS/FLUS, the NPV was similar (97.2% [CI 79-100%]) with again moderate PPV (76.9% [CI 61-93%]) [55]. The demonstration of NPV comparable to benign cytology and PPV similar to that found in the Bethesda V category for which surgically is typically recommended suggests this test may provide both "rule-out" and "rule-in" capability; however, confirmatory data from multicenter, blinded studies or broader clinical use remain lacking. There presently are no published data from a prospective, multicenter, blinded validation of this broader panel, thus limiting any understanding of its clinical validation or clinical utility at this time.

Microarray Expression

Microarray platforms are a method to rapidly determine the expression of all transcribed RNA at relative low cost, allowing for the development of diagnostic microarray panels assessing hundreds of expressed genes. Importantly, computational algorithms are necessary to analyze the expression patterns seen. These techniques may be replaced with next-generation sequencing platforms, but it remains a robust form of expression analysis.

MicroRNA

MicroRNA (miRNA) are 21–22 nucleotide segments of noncoding RNA that play a key role in posttranscriptional gene regulation through complementary binding to messenger RNA that regulates translation and degradation [56]. Overexpression of specific miRNAs has been shown in FTC versus follicular adenoma and PTC versus normal thyroid tissues [57, 58], and initial studies using different miRNA expression profiles on tissue and/or aspiration samples showed overall accuracy from 76 to 90% [59–61]. Combining the relative strengths found in miRNA expression classification and oncogene determination, a two-step testing protocol has been developed and studied (marketed as ThyGenX[®] and ThyraMIR[™] combination testing). 10-miRNA panel and 7-gene mutational panel were assessed on FNA biopsy material from 109 Bethesda III or IV nodules with available surgical pathology and demonstrated 94% [CI 85–98%]) NPV and 74% [CI 58–86%] PPV in a population with a 32% rate of malignancy [52]. Confirmation and validation in a large prospective sample representative of the general nodule population are needed.

RNA Gene Expression Classification

Since the majority of indeterminate nodules are referred for surgical resection and are ultimately proven benign, a novel paradigm was proposed to develop a test to identify benignity, thus effectively "ruling out" malignancy and preventing unnecessary surgery. The design of the Afirma[®] gene expression classifier (GEC) (marketed by Veracyte, Inc.) was based on this idea. This test measures 167 expressed RNAs from FNA tissue and uses trained computational algorithms to identify a profile highly correlated with a benign diagnosis when applied to indeterminate cytology nodules. After RNA expression profiling is completed, the result classifies a nodule as "benign" or "suspicious" [62]. A prospective, multicenter, blinded validation study evaluated the performance and clinical validity of the GEC in 265 indeterminate nodules greater than 1 cm with available histopathology [63]. In this study population with a rate of malignancy of 32%, NPV and PPV were 93% and 47%, respectively. The NPV for Bethesda V nodules was 85% and has generally been considered insufficient for "rule-out" testing. In contrast, NPV was 95% and 94% for nodules with AUS/FLUS and SFN/FN cytology, respectively, which is considered similar to the malignancy risk of a benign cytology and sufficient to recommend conservative management.

In a subsequent retrospective analysis of 339 indeterminate nodules from five academic centers, patients with an indeterminate nodule and "suspicious" GEC result underwent nodule resection in 121 of 148 (82%) cases, with malignancy confirmed in 53 (44%) nodules. By contrast, 4 of 174 patients with a "benign" GEC result were referred for surgery [64]. Taken together, these results show that when applied to clinical care, the Afirma GEC changes management decisions and reduces the need for diagnostic surgery.

The cost-effectiveness of clinical implementation of the GEC has been evaluated in several studies. Cost-effectiveness was demonstrated with statistical modeling [18] and separately by calculations based on rates of surgical resection for GECtested patients [65]. One subsequent single-center report did not find routine GEC testing to be cost-effective at a cancer prevalence of 24.5% [66]; however, routine GEC testing was projected to be cost-effective for third-party payers in the United States in a separate analysis [67].

Numerous additional independent retrospective reports have been published regarding the performance and clinical implications of GEC use in various practice settings [66, 68–78]. Assessing the reportedly high NPV and the ability to reasonably "rule out" the presence of malignancy has been difficult, since the vast majority of patients with a benign GEC result do not undergo resection. The small number of patients for whom nodule resection is performed likely represents a biased sample in which there was some additional risk that drove the decision to perform surgery. One study has compared the sonographic follow-up of benign GEC nodules to a large cohort of cytologically benign nodules with similar baseline characteristics and follow-up time [73]. In this analysis, there was no significant difference in nodule growth or suspicious features when reassessed sonographically at a mean of 13 months, suggesting comparable behavior of GEC-benign nodules to cytologically benign ones. To date, there has been no death attributed to a false-negative, GEC-benign, thyroid malignancy that was later detected.

The positive predictive value of the Afirma GEC has been the topic of more extensive study, with single-center, retrospective analyses reporting PPVs for Bethesda category III or IV nodules from 14 to 82%, though most fall within the range anticipated by the original validation trial [66, 68–72, 74–77]. Two studies selecting nodules with Hurthle cell cytologies (AUS with predominantly Hurthle cells and/or suspicious for Hurthle cell neoplasm) have detected a higher than expected rate of suspicious GEC results, and a low risk of malignancy in these nodules [68, 72], but this has not been found in all studies [76]. Whether these observations represent differences in underlying malignancy rate, selection bias in which patients with Hurthle cell cytology receive Afirma GEC testing, assay performance, variability of cytological interpretation, or physician/patient decision-making remains uncertain, but such factors likely contributed to these findings.

It is critical to appreciate that the likelihood of malignancy in a cytologically indeterminate and GEC suspicious nodule is related to the underlying probability of malignancy within the population tested. Differences in the population prevalence of thyroid cancer between institutions, interpretation of sonographic nodule appearance, selection of which nodules undergo FNA biopsy, risk of malignancy implied by an indeterminate result at a given location, selection of which patients undergo resection, and the interpretation of the histopathologic result will all influence the probability that such a nodule will prove malignant. This variability is inherent to the multidisciplinary evaluation of a thyroid nodule [8, 15] and is an immutable limitation of all molecular testing for thyroid nodules. Therefore, it is imperative to understand the PPV of a molecular test in the context of the likelihood that malignancy is present within a thyroid nodule prior to such testing in order to optimally interpret what the test result indicates regarding a patient's risk of harboring thyroid cancer.

Future Directions

The evaluation and management of thyroid nodules due to their inherent risk of thyroid cancer is likely to continue to increase in the future, highlighting the clinical need for continued progress toward adjuvant testing to assess the risk of malignancy when FNA cytology remains indeterminate. Thyroid cancer expression of programmed death ligand-1 (PD-L1), a cell surface protein that promotes inhibition of immune activation when binding to its ligand on effector immune cells, has recently been shown to correlate with aggressive behavior in PTC tumors [79], and IHC assessment of PD-L1 on FNA material may demonstrate value in differentiating benign from malignant nodules. Testing for mutations/translocations, miRNAs, and/ or mRNA expression profiles is likely to continue to improve and address limitations in diagnostic accuracy in future iterations of their development. Finally, novel molecular markers, such as measurement of circulating suppressor immune cells, circulating tumor cells, cell-free DNA, and other modalities, will continue to be explored.

Summary

The increasing availability of accurate, reliable, and affordable molecular diagnostic testing is changing the management of indeterminate thyroid nodules. Outcomes and cost-benefit analyses should favor limiting medical interventions to patients likely to obtain greatest benefit while simultaneously reducing test and treatment morbidity. As the number and types of tests continue to expand and their performance continues to improve, the regular use of molecular markers is increasingly becoming a standard practice [10, 16].

At present, there are few widely available options for the evaluation of nodules classified as indeterminate, of which the Veracyte Afirma[®] gene expression classifier and the 17 gene mutation panels have been the most independently studied and analyzed. Confirmation in large-scale, prospective, and blinded investigations will be critical to the full understanding of test performance of many molecular tests being introduced. Currently, publications are primarily based upon small, single-institution experiences, with suboptimal methodology, and data must be interpreted in the context of these limitations.

With widespread use of these tests, our understanding of their limitations also has become apparent. Population diversity and practice variation lead to substantial differences in test performance from site to site and best explain differences in published experiences. Such variation is a limitation that must be acknowledged and will never disappear. Separately, this field is increasingly seeking to better understand the meaning of all molecular changes. Increased and improved understanding of mutations that exist in benign thyroid nodules will help to expand our knowledge of when genetic alterations (base pair change or translocations) do not necessarily imply malignant transformation. Ongoing research in this field actively continues with the identification of new potential molecular markers and the publication of clinical trials demonstrating both the utility and limitations of these markers in clinical practice. After discovery, large-scale, prospective clinical validation trials are vitally important. Given the heterogeneity of thyroid cytology and histology, variation in populations, and use of a marker in clinical use, transparent clinical trials provide the optimal means for understanding molecular marker performance in a real-world setting. Indeed, large-scale validation should be viewed as a critical next step after molecular marker discovery.

The current, and future, approach to patients with a clinically relevant thyroid nodule will almost certainly include assessment of clinical, biochemical, radiologic, cytologic—and now molecular—variables as we determine the risk of thyroid malignancy and optimal thyroid nodule management.

Disclosure Statement Dr. Alexander has received research support from Veracyte, Inc. and Asuragen, Inc. He has served as a consultant or scientific advisory board member for Veracyte, Inc., Asuragen, Inc., and DiaSorin, Inc.

References

- Ahmed S, Horton KM, Jeffrey RB Jr, Sheth S, Fishman EK. Incidental thyroid nodules on chest CT: review of the literature and management suggestions. AJR. 2010;195(5):1066–71.
- Arslan N, Dehdashti F, Doherty GM, Lairmore TC, Brunt LM, Moley JF. Risk of malignancy in thyroid incidentalomas identified by fluorodeoxyglucose-positron emission tomography. Surgery. 2001;130(6):941–6.
- Gharib H, Goellner JR. Fine-needle aspiration biopsy of the thyroid: an appraisal. Ann Intern Med. 1993;118(4):282–9.
- 4. Mazzaferri EL. Management of a solitary thyroid nodule. N Engl J Med. 1993;328(8):553-9.
- Yassa L, Cibas ES, Benson CB, Frates MC, Doubilet PM, Gawande AA, Moore FD Jr, Kim BW, Nosé V, Marqusee E, Larsen PR, Alexander EK. Long-term assessment of a multidisciplinary approach to thyroid nodule diagnostic evaluation. Cancer. 2007;111(6):508–16.
- Kwong N, Medici M, Angell TE, Liu X, Marqusee E, Cibas ES, Krane JF, Barletta JA, Kim MI, Reed Larsen P, Alexander EK. The influence of patient age on thyroid nodule formation, multinodularity, and thyroid cancer risk. J Clin Endocrinol Metab. 2015;100(12):4434–40.
- Ron E, Lubin JH, Shore RE, Mabuchi K, Modan B, Pottern LM, Schneider AB, Tucker MA, Boice JD Jr. Thyroid cancer after exposure to external radiation: a pooled analysis of seven studies. Radiat Res. 1995;141(3):259–77.
- Moon WJ, Jung SL, Lee JH, Na DG, Baek JH, Lee YH, Kim J, Kim HS, Byun JS, Lee DH. Benign and malignant thyroid nodules: US differentiation – multicenter retrospective study. Radiology. 2008;247(3):762–70.
- 9. Jarlov AE, Nygard B, Hegedus L, Karstrup S, Hansen JM. Observer variation in ultrasound assessment of the thyroid gland. Br J Radiol. 1993;66(787):625–7.
- 10. Haugen BR, Alexander EK, Bible KC, Doherty GM, Mandel SJ, Nikiforov YE, Pacini F, Randolph GW, Sawka AM, Schlumberger M, Schuff KG, Sherman SI, Sosa JA, Steward DL, Tuttle RM, Wartofsky L. 2015 American Thyroid Association management guidelines for adult patients with thyroid nodules and differentiated thyroid cancer: the American Thyroid Association guidelines task force on thyroid nodules and differentiated thyroid cancer. Thyroid. 2016;26(1):1–133.

- Cibas ES, Ali SZ, NCI Thyroid FNA State of the Science Conference. The Bethesda system for reporting thyroid cytopathology. Am J Clin Pathol. 2009;132(5):658–65.
- 12. Ali SZ, Cibas ES. The Bethesda system for reporting thyroid cytopathology: definitions, criteria, and explanatory notes. New York: Springer; 2009.
- Piana S, Frasoldati A, Ferrari M, Valcavi R, Froio E, Barbieri V, Pedroni C, Gardini G. Is a fivecategory reporting scheme for thyroid fine needle aspiration cytology accurate? Experience of over 18,000 FNAs reported at the same institution during 1998–2007. Cytopathology. 2010;22(3):164–73.
- 14. Nayar R, Ivanovic M. The indeterminate thyroid fine-needle aspiration: experience from an academic center using terminology similar to that proposed in the 2007 National Cancer Institute Thyroid Fine Needle Aspiration State of the Science Conference. Cancer. 2009;117(3):195–202.
- Cibas ES, Baloch ZW, Fellegara G, LiVolsi VA, Raab SS, Rosai J, Diggans J, Friedman L, Kennedy GC, Kloos RT, Lanman RB, Mandel SJ, Sindy N, Steward DL, Zeiger MA, Haugen BR, Alexander EK. A prospective assessment defining the limitations of thyroid nodule pathologic evaluation. Ann Intern Med. 2013;159(5):325–32.
- 16. Gharib H, Papini E, Garber JR, Duick DS, Harrell RM, Hegedüs L, Paschke R, Valcavi R, Vitti P, AACE/ACE/AME Task Force on Thyroid Nodules. American Association of Clinical Endocrinologists, American College of Endocrinology, and Associazione Medici Endocrinologi medical guidelines for clinical practice for the diagnosis and management of thyroid nodules 2016 update. Endocr Pract. 2016;22(5):622–39.
- 17. Nikiforov YE, Seethala RR, Tallini G, Baloch ZW, Basolo F, Thompson LD, Barletta JA, Wenig BM, Al Ghuzlan A, Kakudo K, Giordano TJ, Alves VA, Khanafshar E, Asa SL, El-Naggar AK, Gooding WE, Hodak SP, Lloyd RV, Maytal G, Mete O, Nikiforova MN, Nosé V, Papotti M, Poller DN, Sadow PM, Tischler AS, Tuttle RM, Wall KB, VA LV, Randolph GW, Ghossein RA. Nomenclature revision for encapsulated follicular variant of papillary thyroid carcinoma: a paradigm shift to reduce overtreatment of indolent tumors. JAMA Oncol. 2016;2(8):1023–9.
- Li H, Robinson KA, Anton B, Saldanha IJ, Ladenson PW. Cost-effectiveness of a novel molecular test for cytologically indeterminate thyroid nodules. J Clin Endocrinol Metab. 2011;96(11):E1719–26.
- Yip L, Farris C, Kabaker AS, Hodak SP, Nikiforova MN, McCoy KL, Stang MT, Smith KJ, Nikiforov YE, Carty SE. Cost impact of molecular testing for indeterminate thyroid nodule fine-needle aspiration biopsies. J Clin Endocrinol Metab. 2012;97(6):1905–12.
- 20. Liu FT, Rabinovich GA. Galectins as modulators of tumour progression. Nat Rev Cancer. 2005;5(1):29–41.
- Chiu CG, Strugnell SS, Griffith OL, Jones SJ, Gown AM, Walker B, Nabi IR, Wiseman SM. Diagnostic utility of galectin-3 in thyroid cancer. Am J Pathol. 2010;176(5):2067–81.
- 22. Bartolazzi A, Gasbarri A, Papotti M, Bussolati G, Lucante T, Khan A, Inohara H, Marandino F, Orlandi F, Nardi F, Vecchione A, Tecce R, Larsson O, Thyroid Cancer Study Group. Application of an immunodiagnostic method for improving preoperative diagnosis of nodular thyroid lesions. Lancet. 2001;357(9269):1644–50.
- 23. Bartolazzi A, Orlandi F, Saggiorato E, Volante M, Arecco F, Rossetto R, Palestini N, Ghigo E, Papotti M, Bussolati G, Martegani MP, Pantellini F, Carpi A, Giovagnoli MR, Monti S, Toscano V, Sciacchitano S, Pennelli GM, Mian C, Pelizzo MR, Rugge M, Troncone G, Palombini L, Chiappetta G, Botti G, Vecchione A, Bellocco R, Italian Thyroid Cancer Study Group (ITCSG). Galectin-3-expression analysis in the surgical selection of follicular thyroid nodules with indeterminate fine-needle aspiration cytology: a prospective multicentre study. Lancet Oncol. 2008;9(6):543–9.
- Franco C, Martínez V, Allamand JP, Medina F, Glasinovic A, Osorio M, Schachter D. Molecular markers in thyroid fine-needle aspiration biopsy: a prospective study. Appl Immunohistochem Mol Morphol. 2009;17(3):211–5.
- 25. Torregrossa L, Faviana P, Filice ME, Materazzi G, Miccoli P, Vitti P, Fontanini G, Melillo RM, Santoro M, Basolo F. CXC chemokine receptor 4 immunodetection in the follicular variant of papillary thyroid carcinoma: comparison to galectin-3 and hector battifora mesothelial cell-1. Thyroid. 2010;20(5):495–504.

- 8 Molecular Markers and Thyroid Nodule Evaluation
- Moll R, Divo M, Langbein L. The human keratins: biology and pathology. Histochem Cell Biol. 2008;129(6):705–33.
- 27. Saggiorato E, De Pompa R, Volante M, Cappia S, Arecco F, Dei Tos AP, Orlandi F, Papotti M. Characterization of thyroid 'follicular neoplasms' in fine-needle aspiration cytological specimens using a panel of immunohistochemical markers: a proposal for clinical application. Endocr Relat Cancer. 2005;12(2):305–17.
- Fadda G, Rossi ED, Raffaelli M, Pontecorvi A, Sioletic S, Morassi F, Lombardi CP, Zannoni GF, Rindi G. Follicular thyroid neoplasms can be classified as low- and high-risk according to HBME-1 and Galectin-3 expression on liquid-based fine-needle cytology. Eur J Endocrinol. 2011;165(3):447–53.
- Saleh HA, Feng J, Tabassum F, Al-Zohaili O, Husain M, Giorgadze T. Differential expression of galectin-3, CK19, HBME1, and Ret oncoprotein in the diagnosis of thyroid neoplasms by fine needle aspiration biopsy. Cytojournal. 2009;6:18.
- 30. Cui W, Sang W, Zheng S, Ma Y, Liu X, Zhang W. Usefulness of cytokeratin-19, galectin-3, and Hector Battifora mesothelial-1 in the diagnosis of benign and malignant thyroid nodules. Clin Lab. 2012;58(7–8):673–80.
- Trimboli P, Guidobaldi L, Amendola S, Nasrollah N, Romanelli F, Attanasio D, Ramacciato G, Saggiorato E, Valabrega S, Crescenzi A. Galectin-3 and HBME-1 improve the accuracy of core biopsy in indeterminate thyroid nodules. Endocrine. 2016;52(1):39–45.
- 32. Garnett MJ, Marais R. Guilty as charged: B-RAF is a human oncogene. Cancer Cell. 2004;6(4):313–9.
- Xing M. BRAF mutation in papillary thyroid cancer: pathogenic role, molecular bases, and clinical implications. Endocr Rev. 2007;28(7):742–62.
- 34. Kebebew E, Weng J, Bauer J, Ranvier G, Clark OH, Duh QY, Shibru D, Bastian B, Griffin A. The prevalence and prognostic value of BRAF mutation in thyroid cancer. Ann Surg. 2007;246(3):466–70.
- 35. Xing M, Clark D, Guan H, Ji M, Dackiw A, Carson KA, Kim M, Tufaro A, Ladenson P, Zeiger M, Tufano R. BRAF mutation testing of thyroid fine-needle aspiration biopsy specimens for preoperative risk stratification in papillary thyroid cancer. J Clin Oncol. 2009;27(18):2977–82.
- 36. Xing M, Alzahrani AS, Carson KA, Shong YK, Kim TY, Viola D, Elisei R, Bendlová B, Yip L, Mian C, Vianello F, Tuttle RM, Robenshtok E, Fagin JA, Puxeddu E, Fugazzola L, Czarniecka A, Jarzab B, O'Neill CJ, Sywak MS, Lam AK, Riesco-Eizaguirre G, Santisteban P, Nakayama H, Clifton-Bligh R, Tallini G, Holt EH, Sýkorová V. Association between BRAF V600E mutation and recurrence of papillary thyroid cancer. J Clin Oncol. 2015;33(1):42–50.
- 37. Xing M, Alzahrani AS, Carson KA, Viola D, Elisei R, Bendlova B, Yip L, Mian C, Vianello F, Tuttle RM, Robenshtok E, Fagin JA, Puxeddu E, Fugazzola L, Czarniecka A, Jarzab B, O'Neill CJ, Sywak MS, Lam AK, Riesco-Eizaguirre G, Santisteban P, Nakayama H, Tufano RP, Pai SI, Zeiger MA, Westra WH, Clark DP, Clifton-Bligh R, Sidransky D, Ladenson PW, Sykorova V. Association between BRAF V600E mutation and mortality in patients with papillary thyroid cancer. JAMA. 2013;309(14):1493–501.
- 38. Repasky GA, Chenette EJ, Der CJ. Renewing the conspiracy theory debate: does Raf function alone to mediate Ras oncogenesis? Trends Cell Biol. 2004;14(11):639–47.
- Nikiforov YE, Nikiforova MN. Molecular genetics and diagnosis of thyroid cancer. Nat Rev Endocrinol. 2011;7(10):569–80.
- 40. Yoo S-K, Lee S, Kim S-j, Jee H-G, Kim B-A, Cho H, Song YS, Cho SW, Won J-K, Shin J-Y, Park DJ, Kim J-I, Lee KE, Park YJ, Seo J-S. Comprehensive analysis of the transcriptional and mutational landscape of follicular and papillary thyroid cancers. PLoS Genet. 2016;12(8):e1006239.
- 41. Nikiforov YE, Yip L, Nikiforova MN. New strategies in diagnosing cancer in thyroid nodules: impact of molecular markers. Clin Cancer Res. 2013;19(9):2283–8.
- 42. Medici M, Kwong N, Angell TE, Marqusee E, Kim MI, Frates MC, Benson CB, Cibas ES, Barletta JA, Krane JF, Ruan DT, Cho NL, Gawande AA, Moore FD Jr, Alexander EK. The variable phenotype and low-risk nature of RAS-positive thyroid nodules. BMC Med. 2015;13(1):184.

- Tallini G, Asa SL. RET oncogene activation in papillary thyroid carcinoma. Adv Anat Pathol. 2001;8(6):345–54.
- 44. Elisei R, Romei C, Vorontsova T, Cosci B, Veremeychik V, Kuchinskaya E, Basolo F, Demidchik EP, Miccoli P, Pinchera A, Pacini F. RET/PTC rearrangements in thyroid nodules: studies in irradiated and not irradiated, malignant and benign thyroid lesions in children and adults. J Clin Endocrinol Metab. 2001;86(7):3211–6.
- 45. Marques AR, Espadinha C, Catarino AL, Moniz S, Pereira T, Sobrinho LG, Leite V. Expression of PAX8-PPAR gamma 1 rearrangements in both follicular thyroid carcinomas and adenomas. J Clin Endocrinol Metab. 2002;87(8):3947–52.
- 46. Nikiforov YE, Steward DL, Robinson-Smith TM, Haugen BR, Klopper JP, Zhu Z, Fagin JA, Falciglia M, Weber K, Nikiforova MN. Molecular testing for mutations in improving the fineneedle aspiration diagnosis of thyroid nodules. J Clin Endocrinol Metab. 2009;94(6):2092–8.
- 47. Beaudenon-Huibregtse S, Alexander EK, Guttler RB, Hershman JM, Babu V, Blevins TC, Moore P, Andruss B, Labourier E. Centralized molecular testing for oncogenic gene mutations complements the local cytopathologic diagnosis of thyroid nodules. Thyroid. 2014;24(10):1479–87.
- 48. Cantara S, Capezzone M, Marchisotta S, Capuano S, Busonero G, Toti P, Di Santo A, Caruso G, Carli AF, Brilli L, Montanaro A, Pacini F. Impact of proto-oncogene mutation detection in cytological specimens from thyroid nodules improves the diagnostic accuracy of cytology. J Clin Endocrinol Metab. 2010;95(3):1365–9.
- 49. Nikiforov YE, Ohori NP, Hodak SP, Carty SE, LeBeau SO, Ferris RL, Yip L, Seethala RR, Tublin ME, Stang MT, Coyne C, Johnson JT, Stewart AF, Nikiforova MN. Impact of mutational testing on the diagnosis and management of patients with cytologically indeterminate thyroid nodules: a prospective analysis of 1056 FNA samples. J Clin Endocrinol Metab. 2011;96(11):3390–7.
- Eszlinger M, Krogdahl A, Münz S, Rehfeld C, Precht Jensen EM, Ferraz C, Bösenberg E, Drieschner N, Scholz M, Hegedüs L, Paschke R. Impact of molecular screening for point mutations and rearrangements in routine air-dried fine-needle aspiration samples of thyroid nodules. Thyroid. 2014;24(2):305–13.
- Eszlinger M, Piana S, Moll A, Bösenberg E, Bisagni A, Ciarrocchi A, Ragazzi M, Paschke R. Molecular testing of thyroid fine-needle aspirations improves presurgical diagnosis and supports the histologic identification of minimally invasive follicular thyroid carcinomas. Thyroid. 2015;25(4):401–9.
- 52. Labourier E, Shifrin A, Busseniers AE, Lupo MA, Manganelli ML, Andruss B, Wylie D, Beaudenon-Huibregtse S. Molecular testing for miRNA, mRNA, and DNA on fine-needle aspiration improves the preoperative diagnosis of thyroid nodules with indeterminate cytology. J Clin Endocrinol Metab. 2015;100(7):2743–50.
- Agrawal N, Akbani R, Aksoy BA, Ally A, Arachchi H, Asa SL, Auman JT, Balasundaram M. Cancer Genome Atlas Research Network. Integrated genomic characterization of papillary thyroid carcinoma. Cell. 2014;159(3):676–90.
- 54. Nikiforov YE, Carty SE, Chiosea SI, Coyne C, Duvvuri U, Ferris RL, Gooding WE, Hodak SP, LeBeau SO, Ohori NP, Seethala RR, Tublin ME, Yip L, Nikiforova MN. Highly accurate diagnosis of cancer in thyroid nodules with follicular neoplasm/suspicious for a follicular neoplasm cytology by ThyroSeq v2 next-generation sequencing assay. Cancer. 2014;120(23):3627–34.
- 55. Nikiforov YE, Carty SE, Chiosea SI, Coyne C, Duvvuri U, Ferris RL, Gooding WE, LeBeau SO, Ohori NP, Seethala RR, Tublin ME, Yip L, Nikiforova MN. Impact of the multi-gene ThyroSeq next-generation sequencing assay on cancer diagnosis in thyroid nodules with atypia of undetermined significance/follicular lesion of undetermined significance cytology. Thyroid. 2015;25(11):1217–23.
- 56. Bartels CL, Tsongalis GJ. MicroRNAs: novel biomarkers for human cancer. Clin Chem. 2009;55(4):623–31.
- 57. Weber F, Teresi RE, Broelsch CE, Frilling A, Eng CA. limited set of human MicroRNA is deregulated in follicular thyroid carcinoma. J Clin Endocrinol Metab. 2006;91(9):3584–91.

- 8 Molecular Markers and Thyroid Nodule Evaluation
- Pallante P, Visone R, Ferracin M, Ferraro A, Berlingieri MT, Troncone G, Chiappetta G, Liu CG, Santoro M, Negrini M, Croce CM, Fusco A. MicroRNA deregulation in human thyroid papillary carcinomas. Endocr Relat Cancer. 2006;13(2):497–508.
- 59. Keutgen XM, Filicori F, Crowley MJ, Wang Y, Scognamiglio T, Hoda R, Buitrago D, Cooper D, Zeiger MA, Zarnegar R, Elemento O, Fahey TJ 3rd. A panel of four miRNAs accurately differentiates malignant from benign indeterminate thyroid lesions on fine needle aspiration. Clin Cancer Res. 2012;18(7):2032–8.
- 60. Shen R, Liyanarachchi S, Li W, et al. MicroRNA signature in thyroid fine needle aspiration cytology applied to "atypia of undetermined significance" cases. Thyroid. 2012;22:9–16.
- 61. Kitano M, Rahbari R, Patterson EE, et al. Evaluation of candidate diagnostic microRNAs in thyroid fine-needle aspiration biopsy samples. Thyroid. 2012;22:285–91.
- 62. Chudova D, Wilde JI, Wang ET, Wang H, Rabbee N, Egidio CM, Reynolds J, Tom E, Pagan M, Rigl CT, Friedman L, Wang CC, Lanman RB, Zeiger M, Kebebew E, Rosai J, Fellegara G, LiVolsi VA, Kennedy GC. Molecular classification of thyroid nodules using high-dimensionality genomic data. J Clin Endocrinol Metab. 2010;95(12):5296–304.
- 63. Alexander EK, Kennedy GC, Baloch ZW, Cibas ES, Chudova D, Diggans J, Friedman L, Kloos RT, LiVolsi VA, Mandel SJ, Raab SS, Rosai J, Steward DL, Walsh PS, Wilde JI, Zeiger MA, Lanman RB, Haugen BR. Preoperative diagnosis of benign thyroid nodules with indeterminate cytology. N Engl J Med. 2012;367(8):705–15.
- 64. Alexander EK, Schorr M, Klopper J, Kim C, Sipos J, Nabhan F, Parker C, Steward DL, Mandel SJ, Haugen BR. Multicenter clinical experience with the Afirma gene expression classifier. J Clin Endocrinol Metab. 2014;99(1):119–25.
- 65. Duick DS, Klopper JP, Diggans JC, Friedman L, Kennedy GC, Lanman RB, McIver B. The impact of benign gene expression classifier test results on the endocrinologist-patient decision to operate on patients with thyroid nodules with indeterminate fine-needle aspiration cytopa-thology. Thyroid. 2012;22(10):996–1001.
- Wu JX, Lam R, Levin M, Rao J, Sullivan PS, Yeh MW. Effect of malignancy rates on costeffectiveness of routine gene expression classifier testing for indeterminate thyroid nodules. Surgery. 2016;159(1):118–26.
- 67. Lee L, How J, Tabah RJ, Mitmaker EJ. Cost-effectiveness of molecular testing for thyroid nodules with atypia of undetermined significance cytology. J Clin Endocrinol Metab. 2014;99(8):2674–82.
- 68. Harrell RM, Bimston DN. Surgical utility of Afirma: effects of high cancer prevalence and oncocytic cell types in patients with indeterminate thyroid cytology. Endocr Pract. 2014;20(4):364–9.
- 69. McIver B, Castro MR, Morris JC, Bernet V, Smallridge R, Henry M, Kosok L, Reddi H. An independent study of a gene expression classifier (Afirma[™]) in the evaluation of cytologically indeterminate thyroid nodules. J Clin Endocrinol Metab. 2014;99(11):4069–77.
- Lastra RR, Pramick MR, Crammer CJ, LiVolsi VA, Baloch ZW. Implications of a suspicious afirma test result in thyroid fine-needle aspiration cytology: an institutional experience. Cancer Cytopathol. 2014;122(10):737–44.
- Marti JL, Avadhani V, Donatelli LA, Niyogi S, Wang B, Wong RJ, Shaha AR, Ghossein RA, Lin O, Morris LG, Ho AS. Wide inter-institutional variation in performance of a molecular classifier for indeterminate thyroid nodules. Ann Surg Oncol. 2015;22(12):3996–4001.
- Brauner E, Holmes BJ, Krane JF, Nishino M, Zurakowski D, Hennessey JV, Faquin WC, Parangi S. Performance of the Afirma gene expression classifier in Hurthle cell thyroid nodules differs from other indeterminate thyroid nodules. Thyroid. 2015;25(7):789–96.
- Angell TE, Frates MC, Medici M, Liu X, Kwong N, Cibas ES, Kim MI, Marqusee E. Afirma benign thyroid nodules show similar growth to cytologically benign nodules during follow-up. J Clin Endocrinol Metab. 2015;100(11):E1477–83.
- 74. Yang SE, Sullivan PS, Zhang J, Govind R, Levin MR, Rao JY, Moatamed NA. Has Afirma gene expression classifier testing refined the indeterminate thyroid category in cytology? Cancer Cytopathol. 2016;124(2):100–9.

- Sacks WL, Bose S, Zumsteg ZS, Wong R, Shiao SL, Braunstein GD, Ho AS. Impact of Afirma gene expression classifier on cytopathology diagnosis and rate of thyroidectomy. Cancer Cytopathol. 2016;124(10):722–8.
- 76. Chaudhary S, Hou Y, Shen R, Hooda S, Li Z. Impact of the Afirma gene expression classifier result on the surgical management of thyroid nodules with category III/IV cytology and its correlation with surgical outcome. Acta Cytol. 2016;60(3):205–10.
- 77. Villabona CV, Mohan V, Arce KM, Diacovo J, Aggarwal A, Betancourt J, Amer H, Jose T, DeSantis P, Cabral J. Utility of ultrasound vs. gene expression classifier in thyroid nodules with atypia of undetermined significance. Endocr Pract. 2016;22(10):1199–203.
- Celik B, Whetsell CR, Nassar A. Afirma GEC and thyroid lesions: an institutional experience. Diagn Cytopathol. 2015;43(12):966–70.
- 79. Chowdhury S, Veyhl J, Jessa F, Polyakova O, Alenzi A, MacMillan C, Ralhan R, Walfish PG. Programmed death-ligand 1 overexpression is a prognostic marker for aggressive papillary thyroid cancer and its variants. Oncotarget. 2016;7(22):32318–28.