# **Chapter 13 Genetic Control of Metal Sequestration in Hyper-Accumulator Plants**

#### Shahida Shaheen, Qaisar Mahmood, Mahnoor Asif, and Rafiq Ahmad

**Abstract** Heavy metal contamination is an emergent environmental dilemma all over the world, posing serious threat to environment as well as human being by disturbing the ecological balance. There are a number of physical, chemical, and biological techniques applicable worldwide for wastewater treatment, but the phytoremediation techniques are the green, sustainable, and promising solutions to problem of environmental contamination. Studies revealed that there are certain hyper-accumulator genes present in plants, which make them more metal tolerant than non-hyper-accumulator plants species where those genes are absent. In addition, hyper-accumulator plants tackle with heavy metals by activating their responsive genes for chelation, trafficking, and sequestration. Therefore, studying such hyper-accumulator genes opens a gateway for the thorough understanding of phytoremediation techniques.

**Keywords** Hyper-accumulator • Non-hyper-accumulator • Phytoremediation • Contamination • Tolerant

# Abbreviations

Al	Aluminum		
BjMT	Brassica juncea metallothioneins		
Ca	Calcium		
CaM	Calmodulin		
CBL	Calcineurin B-like protein		
CIPK	Calcium-interacting protein kinase		
CRKs	Cysteine-rich receptor-like kinases		
DHAR	Dehydroascorbate reductase		
DNA	Deoxyribonucleic acid		
GR	Glutathione reductase		

S. Shaheen • Q. Mahmood (🖂) • M. Asif • R. Ahmad

Department of Environmental Sciences, COMSATS Institute of Information Technology, Abbottabad 22060, Pakistan

e-mail: mahmoodzju@gmail.com

GSH	Glutathione			
$H_2O_2$	Hydrogen peroxide			
Κ	Kalium (potassium)			
MAPK	Mitogen-activated protein kinase			
MDHAR	Monodehydroascorbate reductase			
MV	Methyl viologen			
MTs	Metallothioneins			
OSMT	Oryza sativa metallothioneins			
PCs	Phytochelatins			
RLKs	Receptor-like kinases			
ROS	Reactive oxygen species			
SOD	Superoxide dismutase			
tApx	Tobacco ascorbate peroxidase			

#### 13.1 Introduction

Wastewater released from industries makes human lives easier but brings heavy metals menace, which is disturbing the ecological balance. Heavy metals are nonbiodegradable chemical species which may accumulate in different plants parts and therefore cause threats to plants and human health [1]. There are a number of techniques developed over times for remediation of heavy metals, but natural treatment systems are more effective compared to a conventional treatment system. Phytoremediation or the use of living plants to remove heavy metals from soils and water bodies and is proposed as a cost-effective and environment-friendly way to clean up the contaminants [2, 3].

In nature, plants are tolerant towards some heavy metals to some extent and assimilate these as essential nutrients. Green plants can be categorized on the basis of plant-metal interaction as hyper-accumulating and non-accumulating plants. On the basis of adaptations against heavy metals exposure, plants are divided into four main categories, metal-tolerant species, metal-resistant species, metal-tolerant non-hyper-accumulator species, metal hyper-tolerant hyper-accumulator plants species [4]. Hyper-accumulators are plant species which are able to uptake, translocate, and accumulate metals in aboveground plant tissues. A hyper-accumulator should have an intensive root uptake system and faster root-to-shoot translocation. Roots uptake metal from the soil and transport them to the stems and into the leaves. As low concentration of trace metals are present in soil so high affinity transport system is used to accumulate metal ions. A number of transporter genes are involved in this process of metal transport [5].

In most of the plants, heavy metals interaction produces oxidative stress in the chloroplast and mitochondrial membranes. This oxidative stress produce of ROS species causes disruption of intercellular and extracellular membranous organelles, ion leakage, lipid peroxidation, and DNA strand cleavage [6–8]. Most of the heavy

metals are recalcitrant in nature thus causing serious damage to the environment. These are nonbiodegradable in nature but biologically can be transformed from more toxic to less toxic condition by their transformation of oxidation state and their conversion from more complex to simplest forms [9].

In nature, plants are capable of self-protection by the production of less toxic reactive compounds or by controlling metals transportation, accumulation, and metal binding with cell wall and vacuole [10, 11]. Many plants when expose to toxic concentration of metal ions try to avoid or decrease its uptake into root cells by limiting the metal ions to the apoplast, binding them to the cell wall or cellular exudates, or by reducing their long distance transportation. If this does not happened, then metals already in the cell adopted storage and detoxification strategies, along with metal transportation, chelation, trafficking, and sequestration into the vacuole. When these actions were completed, then plants trigger oxidative stress defense mechanism and synthesis of stress-related proteins and signaling molecules, such as heat-shock proteins, hormones, and reactive oxygen species [12]. This review has attempted a comprehensive description of plants mechanisms against heavy metals avoidance, transportation, accumulation, and detoxification of heavy metals contamination, and exploring the genetically based defense strategies adopted by plants against trace element excess.

#### **13.2** Avoidance Strategy in Plants

# 13.2.1 Extracellular Defense Strategy of Plants Against Heavy Metals

Plants possess different intrinsic and extrinsic defense strategies for tolerance or detoxification whenever faces the stressful conditions due to the high concentrations of heavy metals. Initially, regarding metal intoxication, plants implement avoidance strategy to prevent the arrival of stress via restricting metal removal from soil or eliminating it, and control metal entry into plant roots [13]. This can be attained by some mechanisms such as restriction of metals by mycorrhizal association, metal sequestration, or complication by releasing organic compound from root [14, 15].

For heavy metals prevention or reduction of its toxicity impacts, plants develop avoidance approach against HMs entrance. Plants adapted two main pathways by taking part in enhancing its complexity in roots vicinity. For the reduction of heavy metal toxicity, plants enhance the pH of rhizosphere which released anions of phosphate. Studies revealed that South American maize variety 3 released phosphate ions without toxicity while sensitive maize variety 5 showed toxicity symptoms under Al stress [16]. Studies revealed that under Cd stress, malate is secreted from sorghum (*Sorghum bicolor* L.) roots, and citrate is secreted from maize roots [17]. Studies accomplished the fact that root exudates in plants rhizosphere decrease the level of toxicity by activating HM-binding proteins which inhibit the HM uptake [18].

Similarly, oxalate released from the root apex facilitates the prevention of Cd from entering into tomato (*Lycopersicon esculentum* L.) roots, thus in the Cd-resistant tomato cultivar (Micro-Tom) these exudates promote Cd resistance. Genetic studies revealed that under Al exposure Al-tolerant higher plants produces more malic acid than sensitive genotypic plants species [19]. Thus, it is concluded that the tolerant plant species may have adopted precipitation as an avoidance mechanism for the prohibition of the HM.

#### **13.3** Signaling Strategy in Plant

#### 13.3.1 Signals Transduction in Plants

In all plants, reaction towards heavy metal stress involves a complex signal transduction system that is trigger by sensing the heavy metal and is characterize by the production of stress-related proteins and signaling molecules, and finally the transcriptional activation of particular metal-responsive genes to neutralize the stress [20].

The most significant signal transduction processes consist of the Cacalmodulin system, hormones, ROS signaling, and the mitogen-activated protein kinase (MAPK) phosphorylation flow, which activates stress-related genes [21]. There are two main types of plants signaling, i.e., extracellular signaling and intracellular signaling.

#### 13.3.2 Signaling Networks

The ROS network is highly dynamic for plants growth, development, and stress states thus producing ROS-signaling response effectively by ROS-scavenging and ROS-producing protein [22]. The production of ROS physiologically occurs as a by-product of biological reactions. During ROS production, P-450 and other cellular elements are released as a by-product [23]. Under chemical toxicity, ROS genes network is being regulated by cytochromes P-450 which slow down the ROS level in plants cells. For instance, the ROS gene network of *Arabidopsis thaliana* contains more than 150 genes for the maintenance of ROS level in plants [24]. Calcium-signaling network regulates the transmission of calcium signals through channels, pumps, and carriers that between cellular, subcellular, and extracellular parts of plants. Ca2+-binding proteins decoded and transmitted the information provided by calcium signaling for transcription by Ca2+-responsive promoter elements that ultimately regulate proteins phosphorylation [25].

#### 13.3.3 Extracellular Signaling

When plants exposed to multiple abiotic stress stimuli, it rapidly activates signaling proteins MAPKs. An extracellular signaling ROS system has been named as "the ROS wave" that covers about 8 cm/min distance. The concept of "ROS wave" is concerned with the perception or signaling of ROS produced in the plants [26]. In many signaling network, the most important thing is the presence of transmembrane proteins that act as receptor-like kinases (RLKs) and recognize signals with their extracellular kinase containing parts then transmit them through the intracellular kinase containing parts. RLKs manage developmental and hormone responses, stomata closing and opening and stress response, and resistance against bacterial and fungal pathogens [27, 28].

About 600 members of RLK gene family has been reported in Arabidopsis. In extracellular parts of plants, the RLK groups like CYSTEINE-RICH RECEPTOR-LIKE KINASES (CRKs) have two preserved cysteine domains (C-2x-C-8x-C; DUF26 domain). Various studies revealed that on the basis of transcriptional and phenotypic analysis of CRK mutants like their extracellular domain structure, phenotype, and genotypic expression, these could be concerned in apoplastic ROS signaling [29–32].

## 13.3.4 Intercellular Signaling

During intercellular signaling plants, information is transmitted in the form of mobile signals, including transcription factors and membrane-associated proteins. Generally, membrane-associated proteins are significant in transcription, as small RNAs and revealed intercellular movement through mobile peptides [33]. ROS are important mediators of developmental procedures in different organisms like prokaryotes, fungi, plants, and animals through redox-sensitive transcriptional regulator genes expression. In plants, regulation of peroxidase genes is possible by novel ROS-sensitive transcription factor, UPB1 [34]. Various organelles within the cell like chloroplasts, peroxisome, and mitochondria can generate reactive oxygen species under stress situations and donate to plant stress tolerance [35]. In case of calcium intercellular signaling, calcium signatures transformed the cellular levels of calcium [36]. Cell organelles like vacuoles, endoplasmic reticulum, mitochondria, and cell wall are the store houses of Ca2+ from where these are released when it is necessary by the plant cells [37]. Similarly, cell organelles surrounded by double membrane (e.g., mitochondria, chloroplasts, and nuclei) can generate Ca<sup>2+</sup> signals whenever posed to stress conditions [38].

#### 13.3.5 Calcium Signaling in Plants

When plants come across a number of physiological stimuli or stress, like low temperature, drought, salinity stress, and pathogen or herbivorous attacks, then free calcium ions of cytoplasm enormously activated [39]. The Ca<sup>2+</sup> are later on transcripted by intercellular reactions, mainly by Ca<sup>2+</sup> sensor proteins which have been preserved in all eukaryotic organisms, so that by activating complex downstream signals in reaction of developmental and environmental stimuli. The physical changes of Ca<sup>2+</sup> binding is measured by structural changes of sensor proteins in Ca<sup>2+</sup> relying proteins [40, 41]. In response to abiotic stress, calcium signaling is produced with the regulation of cell cycle. The equilibrium of Ca<sup>2+</sup> ions depends on the Ca<sup>2+</sup> deficiency, Ca<sup>2+</sup> transporters, efflux pumps, Ca<sup>2+</sup>/H<sup>+</sup> antiporters, Ca<sup>2+</sup> signatures, Ca<sup>2+</sup> memory, Ca<sup>2+</sup> sensor, and transducer proteins [42] (Fig. 13.1).



Fig. 13.1 Representation of Ca2+ signals under diverse abiotic stresses regulated by CAM, CRKS, CPKS, and CCaMK for the activation of regulatory genes

#### 13.3.6 Genes Involved in Calcium Signaling

Calcium and protein kinases significantly take part in signaling pathways against environmental stress in plants. The first calcium-dependent, calmodulinindependent protein kinase activities were reported in pea (*Pisum sativum*) extracts 20 years ago [43]. Various studies have shown that Ca-regulated proteins and kinases [44]. Such as CaM protein [45] neurin B-like 218 (CBL) 8 proteins, CDPK genes [46], and CBL-interacting protein kinase (CIPK) [47] are associated to abiotic stress response in plants.

By the activation of protein kinesis, calcium sensors or  $Ca^{2+}$ -binding proteins detect the high concentration of  $Ca^{2+}$ . In response to the genes, expression of regulatory proteins produced by protein kinesis can enhance transcription factors by phosphorylation and by changing the metabolism that ultimately results in producing phenotypic responses for enhancing stress tolerance [37]. Because of the rise of calcium level two types of genes, specifically up regulating or down regulating are overexpressed and produces stunt growth or death of plant cells. These consist of known quick stressresponsive genes in addition to genes of nonspecific function [48]. Recent studies show that calcium signaling plays a significant role in some pathways, for example, in *Arabidopsis*,  $Ca^{2+}$ -signaling pathway can also control a K<sup>+</sup> channel for low-K response in the presence of a blue light receptor phototropins, which successfully promotes growth and plant development [49]. Calcium signaling is correlated with the sucrosesignaling pathway that is an essential source of fructan synthesis [50]. Under abiotic stress, calcium signaling controls the cell cycle progression.

# 13.3.7 ROS Signaling in Plants

Plants can sense, transduce, and translate ROS signal into suitable cellular response with the assistance of redox-sensitive proteins, calcium mobilization, protein phosphorylation, and gene expression. ROS can be sense directly by some important signaling proteins such as a tyrosine phosphatase through oxidation of conserved cysteine residues [51]. ROS can also regulate many enzymes in signaling, such as protein phosphatases, protein kinases, and transcription factors [52], and transmit to other signal molecules and the ways forming part of the signaling network that regulate the response downstream of ROS [53]. Usually, the power, lifetime, and size of the ROS-signaling pool rely on balance between oxidant production and removal by the antioxidant. By using mutants that lack in key ROS-scavenging enzymes, Miller and coworkers determined a signaling pathway that is operated in cells in response to ROS accumulation [54]. In tomato leaves, ROS produced in cell walls of vascular bundle cells, as a result of wounding and produce  $H_2O_2$  from wound-inducible polygalacturonase which is acted as a second messenger for the activation of defense genes in mesophyll cells, but not for signaling pathway genes in vascular bundle cells [55]. Tracing or detoxification of the unnecessary ROS is achieved by well-organized antioxidative system that contains the nonenzymatic as well as enzymatic antioxidants [56].

## 13.3.8 Genes Involved in ROS Signaling

ROS are a sort of free radicals, reactive molecules, and ions that are obtained from  $O_2$ . It has been evaluated that about 1% of  $O_2$  used by plants is transformed to ROS [57]. The most frequently existing ROS are  $O_2$ ,  $O_2^{-}$ ,  $H_2O_2$ , and 'OH. As  $O_2$  is a completely nontoxic molecule as in its ground state it has two unpaired electrons with equidistant spin which form it paramagnetic and, therefore not likely to take part in reactions with organic molecules unless it is activated [58]. Naturally, in living organisms the production of ROS is responsible for the intracellular communication system that regulated the response to environmental stresses [59]. When the plants are under salinity stress, the target of the ROS is regulated by vesicle trafficking complexes [60].

At low concentration, ROS have been concerned as second messengers in intracellular signaling cascades that mediate several plant responses in plant cells with closing of stomata [53, 61, 62], automatic cell death [63, 64], gravitropism [65], and achievement of tolerance to both biotic and abiotic stresses [54, 66] (Fig. 13.2).

#### 13.3.9 Genetic Control of ROS Production

Studies revealed that under abiotic conditions signaling is produced due to oxidative stress, which result in activation of defense genes giving out specific adaptive responses [67]. The system involved in up regulation of mRNA due to ROS production can occur by redox-sensitive second messenger systems (e.g., MAP kinase activation) [68]. Plants genetics analysis showed that ROS signaling in Arabidopsis plants enhances the antioxidative defense by rising the antioxidative genes expression and activation of the genes of inducible stress proteins [69]. The specific effect of ROS-mediated signaling is related with the confirmation of definite genes expression. Some specific promoters and transcription factors have been recognized as a producer of oxidative stress-responsive elements [70]. Under chilling and salt stress, overexpression of a cytosolic APX-gene taken from pea (Pisum sativum L.) in transgenic tomato plants (Lycopersicon esculentum L.) improves the oxidative injury [71]. Likewise, tolerance against oxidative stress can also be improved by overexpression of the tApx genes in tobacco or in Arabidopsis [72]. Several studies revealed those in plants, under environmental stresses MDHAR show overexpression [73]. Gene expression analysis of wheat showed two varieties of tolerant wheat bHLHs (bHLH2: CA599618 and bHLH3: CJ685625) that have been affected by salinity [74]. Another gene family WRKY plays effective regulatory role in plants under biotic



**Fig. 13.2** Representation of the transduction pathways involved in ROS signaling in response to abiotic stresses. External stimuli produce ROS in chloroplast, mitochondria, and peroxisomes; these activated MAPK cascade. These mitogen-activated protein kinesis that regulate transcription factor and synthesis of genes (SFR, WRKY, bHLH, etc.) to overcome the negative effects of stress

and abiotic stress conditions like cold, drought, wounding, salinity, UV,  $H_2O_2$ , salicylic acid (SA), viral and bacterial attack [75, 76].

Overexpression of Arabidopsis MDHAR gene in tobacco enhanced salt tolerance and mitigated polyethylene glycol stress [77]. Tomato chloroplastic MDHAR overexpressed in transgenic Arabidopsis improved its tolerance to temperature and methyl viologen-related oxidative stresses [78]. Likewise, regulation of the geneencoding cytosolic DHAR was observed in *L. japonicas*, which was proved to be more tolerant to salt stress than other legumes. This increase of DHAR was associated with its action in AsA recovery in the apoplast [79]. Transgenic potato overexpressing Arabidopsis cytosolic AtDHAR1 proved higher tolerance to herbicide, drought, and salt stresses [80].

Kwon et al. [81] verified that simultaneous expression of Cu/Zn-SOD and APX genes in tobacco chloroplasts increased tolerance to methyl viologen (MV) stress relatively to expression of either of these genes alone. Similarly, improved

tolerance to multiple environmental stresses has been produced by instantaneous overexpression of the genes of SOD and APX in the chloroplasts [82–84] SOD and CAT in cytosol [85] and SOD and GR in cytosol [86]. Moreover, the instantaneous expression of numerous antioxidant enzymes, such as Cu/Zn-SOD, APX, and DHAR, in chloroplasts has shown to be more efficient than single or double expression for developing transgenic plants with improved tolerance to various environmental stresses [87]. So, in order to attain tolerance to multiple environmental stresses, increased importance is now given to produce transgenic plants overexpressing multiple antioxidants.

## **13.4 Detoxification Strategy in Plants**

#### 13.4.1 Detoxification Mechanism in Plants

Main detoxification mechanisms in plants are

- 1. Transportation to storage parts
- 2. Chelate formation
- 3. Compartmentalization in subcellular parts
- 4. Removal from the plant body [88].

## 13.4.2 Metal Transporters

Transport system of metal ions is very complex and miscellaneous. Metal transport system is involved in uptake of metal, its translocation to various plant organs and metal liberation in subcellular parts together with metal storage in vacuoles [89]. For long distance or intercellular transport in plants and subcellular compartmentalization of metals, low-molecular-weight chelators, such as glutathione, phytochelatins, histidine, or citrate, play a crucial role. By selective metal chelation and trafficking or by internal transporter selectivity, there is a requirement to make a distinction between metal cations of different elements.

A wide variety of transport proteins occurs that belongs to different families including

- The zinc-regulated transporter, iron-regulated transporter protein (ZIP) family
- The cation diffusion facilitator (CDF) family
- The P1B-type subfamily of P-type ATPases
- The natural resistance-associated macrophage protein (NRAMP) family
- The yellow-stripe 1-like (YSL) subfamily of the oligopeptide transporter (OPT) superfamily
- The copper transporter (COPT) family
- The Ca2+-sensitive cross complementer 1 (CCC1) family
- The iron-regulated protein (IREG) family



Fig. 13.3 Mechanisms involved in heavy metal transport

Other membrane protein families that have been found to involve in transition metal transport are the cation exchanger (CAX) family and three subfamilies of ATP-binding cassette (ABC) transporters, the multidrug resistance-associated proteins (MRP), the ABC transporters of the mitochondria (ATM), and the pleiotropic drug resistance (PDR) transporters [90, 91]. A metal transporter is involved in metal detoxification and metal hyper-accumulation [89].

Metal transporters are involved in:

- 1. Uptake from the soil to root
- 2. Translocation from the root to the shoot
- 3. Detoxification by storage in the vacuoles (Fig. 13.3)

## 13.4.3 Uptake from the Soil to Root

Transporter genes involved in cellular uptake of metals from soil have been identified by researchers. A number of ZIP transporters are found to be involved in Zn uptake across plasma membrane [89]. Fifteen potential ZIP genes may be identified in the *Arabidopsis thaliana* genome [92].

Transport properties of plant metal transporters that mediate metal entry into the cytoplasm, for example, the Zn transporter AtZIP1 have been analyzed upon heterologous expression in yeast by measuring metal uptake into yeast cells [93]. IRT1 is one of the most important members of ZIP family, involved in iron uptake from the soil [93]. Puig et al. [94] found that AtZIP2 and AtZIP4 are involved in cellular accumulation of zinc and copper. OsIRT1 and OsIRT2 have been proposed to contribute in Cd uptake [95]. Milner et al. [96] found that expression of AtZIP1 is localized to the root stele and is a vacuolar transporter while AtZIP1 expression was also found in the leaf vasculature and is localized to the plasma membrane. Functional studies with Arabidopsis AtZIP1 and AtZIP2 T-DNA knockout lines suggest that both transporters play a role in Mn (and possibly Zn) translocation from the root to the shoot. AtZIP1 may play a role in remobilizing Mn from the vacuole to the cytoplasm in root stellar cells and may contribute to radial movement to the xylem parenchyma. AtZIP2, on the other hand, may mediate Mn (and possibly Zn) uptake into root stellar cells, and thus also may contribute to Mn/Zn movement in the stele to the xylem parenchyma, for subsequent xylem loading and transport to the shoot [96].

#### 13.4.4 Translocation from the Root to the Shoot

In phytoextraction, transport of metals from root to shoot is of utmost important. The study of Zn and Cd hyper-accumulators provides the evidence of involving P-ATPase also called HMA (Heavy Metal transporting ATPase) as an important factor in their transport from the cystol of root cells into vascular tissues [97]. The HMAs divide into two groups: those transporting monovalent cations (Cu, Ag) and those transporting divalent cations (Pb, Cd) group [98]. Eight HMAs has been identified in Arabidopsis and *Oryza sativa* [99]. Analysis of the complete genomic sequence in Arabidopsis shows the division of these eight HMAs in two groups: HMA1–4 for the transport of Zn/Co/Cd/Pb and HMA5–8 for the transport of Cu/Ag [100], while HMA2 and HMA4, are involved, in the transport of Zn and Cd. HMA4 confers increased Cd tolerance when expressed in yeast [99].

Transport of cadmium from root to shoot is a control process and most of the Cd is stored in roots. To increase root-to-shoot translocation of Cd, transformation with the genes of high biomass responsible for high root-to-shoot translocation such as HMA4 can be done. In *A. thaliana*, HMA2 and HMA4 genes are involved in the transport of Zn and also nonessential Cd to the shoots [101, 102]. Studies show that both proteins encoding for Cd loading in xylem and are plasma membrane contained proteins [101]. The P1B-type ATPases (also known as HMAs) have a major role in translocation of metal ions against their electrochemical gradient by using ATP as energy. All living organisms including humans, yeast, and plants contain HMAs [100]. In Arabidopsis, HMA4 is found to be more expressed in vascular tissues of root, stem, and leaves. It has been characterized and its role in Cd detoxification has been confirmed in Arabidopsis [100]. HMA4 plays a role in xylem loading

of Zn and Cd, and hence in the control of translocation to shoots in *Arabidopsis halleri* and *Thlaspi caerulescens*. *A. halleri* plants (from a Cd-hyper-tolerant accession) with a lowered expression of HMA4 translocated less Zn from the root to the shoot and were more sensitive to Cd and Zn treatments [103].

Work of Courbot et al. [100] leads to conclusion that elevated expression of HMA4  $P_{1B}$ -type ATPase is an efficient mechanism for improving Cd/Zn tolerance in plants under conditions of Cd/Zn excess by maintaining low cellular Cd<sup>2+</sup> and Zn<sup>2+</sup> concentrations in the cytoplasm. HMA2 and HMA4 are the only P1B ATPases identified so far, which are predicted to have a long C-terminal domain. It is interesting that the CC dipeptides and the His-rich domains are found in the prolonged C termini of HMA2 and HMA4 and not in the N-terminal domain, where HMA domains are always found. The N-terminal end of HMA1 also harbors a poly-His domain [104] (Table 13.1).

## 13.4.5 Detoxification by Storage in the Vacuoles

In the hyper-accumulation of Zn, Ni, and Cd, an enhanced capacity of metal storage in leaf vacuoles seems to play an important role [98]. ABC transporter is involved in many physiological processes. Several members of ABC transporter are involved in vacuolar sequestration of metals. Hmt1 is found to be involved in transport of PC-Cd complexes in the vacuoles of *S. pombe* [89]. Some members of the MATE family were shown to function as cation antiporters that remove toxic compounds from the cytosol by exporting them out of the cell or sequestering them to vacuole [105].

Members of CDF family involved in the cytoplasmic efflux of metal cations from cytoplasm to organelles like Zn<sup>2+</sup>, Cd<sup>2+</sup>, and have been named MTP (metal tolerance protein). CDFs are highly expressed in *A. halleri* and *T. caerulescens*: MTP1, MTP, and MTP11. AtMTP1 suggested being involved in Zn tolerance and basal Zn accumulation in leaves. MTP11 and especially MTP8 are close homologues of ShMTP8 that give Mn tolerance when expressed in yeast and when ectopically overexpressed in *A. thaliana* [93] (Fig. 13.4).

Mechanism of metal transport in plant cell. Heavy metals enter into cytosol through metal transporter from cytosol into vacuole via metal transporters.

HM—High Metal LM—Low Metal ZIP—Zinc-regulated transporter, iron-regulated transporter protein NRAMP—Natural resistance-associated macrophage protein CAX—Cation exchanger ABC—ATP-binding cassette

MT-Metallothioneins

PC—Phytochelatins

-		-	
Type of stress	Genes involved	Plant species	References
Heat	AtCaM3	A. thaliana	Xuan et al. [145]
Heat	AtCaM7	A. thaliana	Lu et al. [146]
Heat	OsCAM1-1	O. sativa	Wu et al. [147]
Salt	GmCaM4/5	Glycine max	Park et al. [148]
Heat	TaCaM1-2	Triticum aestivum	Liu et al. [149]
Salt	AtCML8	A. thaliana	Park et al. [150]
ABA, droughts, salt	AtCML9	A. thaliana	Magnan et al. [151]
Salt	AtCML18/CaM15	A. thaliana	Yamaguchi et al. [152]
Heat, cold, ABA	AtCML24/TCH2	A. thaliana	Delk et al. [153]
ABA, salt	AtCML37/38/39	A. thaliana	Vanderbeld and Snedden [154]
ABA, drought	AtCML42	A. thaliana	Vadassery et al. [155]
Cold, heat, drought, Salt.ABA	OsMSR2	O. sativa	Xu et al. [156]
Heat	AtPP7	A. thaliana	Liu et al. [157]
Heat	AtCBK3/CRK1	A. thaliana	Liu et al. [158]
Cold, heat, Salt.ABA, H <sub>2</sub> O <sub>2</sub>	AtCRCK1	A. thaliana	Yang et al. [159]
Cold	AtCRLK1	A. thaliana	Yang et al. [160, 161]
ABA, H <sub>2</sub> O <sub>2</sub> , ROS, dehydration	OsCCaMK/DMI3	O. sativa	Shi et al. [162]
ABA, ROS	ZmCCaMK	Z. mays	Ma et al. [163]
ABA, salt	TaCCaMK	Pisumsativum	Pandey et al. [164]
Salinity	DHAR	Oryza sativa	Chen and Gallie [165]
Drought, ozone	DHAR	A. thaliana	Ushimaru et al. [166]
Cu, Zn	AtZIP2, AtZIP4	A. thaliana	Puig et al. [94]
Cd	OsIRT1, OsIRT2	O. sativa	Clemens et al. [95]
Mn	ShMTP	A. thaliana	Delhaize et al. [167]
Cu/Ag	HMA5-8	A. halleri	Courbot et al. [100]
Zn/Co/Cd/Pb	HMA1-4	A. halleri	Courbot et al. [100]

 Table 13.1
 Gene expression of various abiotic stress conditions in plants

# 13.4.6 Metal-Binding Genes

Plants have been authentically proved to minimize the harmful effects of metal toxicity, by pathways or methods relating to the binding of heavy metals to cell wall and its transporation [106, 107]. Generally, the synthesis of metal-binding peptides based on the production of metallothioneins and phytochelatins [108]. Usually, chelation is the most common intracellular system for the maintenance of low concentrations and detoxification of free metals in plant cytoplasm that can be achieved by thiol compounds (which contain sulfhydryl/thiol groups; such as a tripeptide glutathione, GSH,  $\gamma$ -Glu-Cys-Gly; phytochelatins, PCs; metallothioneins, MTs), and



Fig. 13.4 Mechanism of metal transport in plant cell. Heavy metals enter into cytosol through metal transporter from cytosol into vacuole via metal transporters

also by non-thiol compounds (such as organic acids and amino acids) [109–118]. Studies revealed that peptides that have either histidines (GHHPHG) 2 (HP) or cysteines (GCGCPCGCG) (CP) be engineered to Lam B and expressed on the surface of *E. coli*. Surface demonstrated that CP and HP enhanced the bioaccumulation fourfold and twofold [119].

#### 13.4.7 Phytochelatins

Phytochelatins are a family of cysteine-rich, thiol-reactive peptides that attach many toxic metals and metalloids, producing good messengers for genetically better phytoremediation system [120]. The general structure of PCs is (g-Glu-Cys) n-Gly, where n differs from 2 to 11 [121]. Practical descriptions of an unusual phytochelatin synthase, LjPCS3, of *Lotus japonicus*, have been acknowledged in an extensive variety of plant species and some microorganisms [122].

Phytochelatins plays an important role in biosynthesis and detoxification of heavy metals [122]. After production, PCs combine with heavy metal ions and make possible their transportation as complex into the vacuole, where they finally produce complexes of high molecular mass, which is the key method that utilizes to bind heavy metal ions in both plants and yeasts [122]. Genes concerned in the production of PCs are phytochelatin synthases, such as g-glutamyl cysteine trans peptidase [121].

Overexpression of phytochelatin synthase in *Arabidopsis thaliana* seedlings causes tolerance of arsenic, but hypersensitivity to cadmium and zinc [123]. The same fact was observed in other transgenic plants with diverse PCS genes show diverse phenotypes, including heavy metal tolerance (by or lacking accumulation) and hypersensitivity to heavy metal ions [124]. Overexpression of pytochelatin synthase in tobacco: distinctive effects of AtPCS1 and CePCS genes on plant response to cadmium [125]. Phytochelatin synthase of *Thlaspi caerulescens* increases tolerance and accretion of heavy metals when expressed in yeast and tobacco. PCs are a group of cysteine-rich, thiol-reactive peptides that combine many toxic metals and metalloids, thus producing best messenger for genetically improved phytoremediation pathways [120].

#### 13.4.8 Metallothioneins (MTs)

Apart from PCs that are the product of enzymatically formulated peptides, MTs are formulated resultantly by mRNA translation [126]. While PCs in plants may primarily deal with Cd detoxification, MTs appear to elaborate the attraction with a larger series of metals such as Cu, Zn, Cd, and As [127]. MTs demonstrated unusual properties and performance that depends on their presence in a type of plants and are extremely mottled in terms of their molecular characteristics and structural qualities [128]; they probably contain a number of various activities in plants than a few other living creatures. In plants, these ligands are concerned to negate the toxicity of HMs by cellular sequestration, homeostasis of intracellular metal ions, and metal transport modifications [129–131].

Additionally, MTs play an important role in HM detoxification, actively involved in cellular-related events including ROS scavenger [132], maintaining of the redox level [133], repair of plasma membrane [134], cell proliferation, and its growth and repair of damaged DNA [135]. There are numerous endogenous and exogenous agents other than HMs that are able to bring the synthesis and expression of MTs. Of these, osmotic stress, drought, intense temperatures, nutrient deficiency, release of different hormones, natural and dark-induced tissue decay, injuries, and viral infections can be mentioned [12, 127, 136].

Ectopically expressed MTs in transgenic plants are proved to increase their tolerance towards metal intoxication. Kumar et al. [137] showed that OSMT1e-p, a type 1 MT extracted from a salt-tolerant rice genotype (*Oryza sativa* L. cv. Pokkali), participated in tolerance for copper and zinc toxicity when ectopically expressed in transgenic tobacco. They evaluated that tobacco plants in which gene have been inserted possessed to hold more quantity of Cu<sup>2+</sup> and Zn<sup>2+</sup> in their roots or lower leaves, considerably decreasing the HMs ions transportation and quantity in leaves and harvestable plant parts. Zhigang et al. [138] accomplished that the ectopic expression of BjMT2, a metallothionein type 2 from *Brassica juncea*, in *Arabidopsis thaliana* enhanced copper and cadmium tolerance at the seedling phase but intensely decreased root growth when there was no heavy metal treatment. This tendency may propose that ectopic expression of MTs in transgenic plants may proceed in host plant in a nonspecific method and in a different way effect the organ growth.

## 13.5 Conclusion and Future Prospects

The present review outlines the impact of abiotic stresses on plants. Most of the investigations done so far mainly described the genetic investigation of plants against abiotic stress; this review involved in genetically based defense and detoxification pathways mainly Ca and ROS signaling, transportation, chelation, and detoxification has been discussed in detail. Under stress conditions, plants activate specific molecules which enhance plants tolerance and the development of defense mechanisms in it. It has been observed that the activation of defense genes cascade transmit various signals in cell organelles under various biotic stress conditions [40].

As abiotic condition produces oxidation stresses that overexpressed a number of stress-induced proteins, this review could provide fundamental information about antioxidant and regulatory genes production. As ROS have regulatory function as signaling molecules, this feature may open a gateway to physiological, molecular, and evolutionary research perspectives. Due to the importance of ROS, it is central to modern plant biology to obtain a comprehensive understanding of the processes where ROS have regulatory roles. Studies revealed that ROS signaling with ozone as a tool is significant for the transmission of distinct from of ROS signals to chromatin reformation and transcriptional regulation [140].

Therefore, the elaboration of transcriptomics and proteomics analysis will be more helpful in understanding the bioinformatics and mutant studies. It has been observed that ROS signals play an important role in intercellular Ca2+ signals, Ca2+ influx which can be regulated by various PAs and Spm4+ proteins [141]. There is need to highlight the mechanism and interrelation of Ca2+-efflux systems with ROS and AtMPK6-signaling under biotic and abiotic stresses along with the overexpression of regulatory genes against stress. Xing et al. [142] observed various proteins kinesis like MKK2-MPK4/MPK6 and MEKK1 activation in salt, cold, drought, and wounding stress can phosphorylate MPK4 which is also significant in abiotic stress signaling. Studies also revealed that ABA is a key hormone in inducing abiotic stress responses in plants like barley showed pronounced effect of ROS and PAs in salt-sensitive variety then salt-tolerant plants [143]. Likewise, ABA is an important hormone under biotic and abiotic stress in plants [144]. But the links between ABA and MAPKs under biotic and abiotic stresses has not yet been properly studied at cellular and subcellular levels in plants. Similarly, little work has been done on K+/Na+ -signaling pathways under biotic and abiotic stresses like Ca<sup>2+</sup> homeostasis. Therefore, plants responses towards multiple abiotic stresses would be another interesting area of future transgenic hyper-accumulator plants production.

In future, it is needed to identify molecular interaction of chelators with metal transporters. Further investigation is needed to determine more about functional significance and biological role of transporter genes, especially, overexpression in plants with greater biomass to increase their potential use in phytoremediation processes. Furthermore, these genes can provide better understanding in the analysis of gene regulation in metal-rich environment as well as metal-deficient environment. Similarly, silencing of transporter genes in edible crops may decrease metal bioaccumulation in food chain. Likewise, there is a greater area of exploration in terms of 3D structures of protein and functional analysis of the candidate genes. In future, gene cloning and plant transformation can be done to determine efficiency of metal transporter genes in transgenic plants. Thus, the application of powerful genetic and molecular techniques may surely be helpful in designing of hyper-accumulator transgenic plants for bioremediation.

#### References

- Annan K, Dickson RA, Amponsah IK, Nooni IK (2013) The heavy metal contents of some selected medicinal plants sampled from different geographical locations. Pharmacognosy Res 5(2):103
- Mirza N, Pervez A, Mahmood Q, Ahmed SS (2010) Phytoremediation of Arsenic (As) and Mercury (Hg) from contaminated soil. World Appl Sci J 8(1):113–118
- Favas PJ, Varun M, DSouza R, Paul MS (2014) Phytoremediation of soils contaminated with metals and metalloids at mining areas: potential of native flora. In: Hernández-Soriano MC (ed) Environmental risk assessment of soil contamination. InTech, Croatia, pp 485–517
- Shabani N, Sayadi MH (2012) Evaluation of heavy metals accumulation by two emergent macrophytes from the polluted soil: an experimental study. Environmentalist 32(1):91–98
- 5. Rascio N, Navari-Izzo F (2011) Heavy metal hyperaccumulating plants: how and why do they do it? And what makes them so interesting? Plant Sci 180(2):169–181
- 6. He J, Qin J, Long L, Ma Y, Li H, Li K, Jiang X, Liu T, Polle A, Liang Z et al (2011) Net cadmium flux and accumulation reveal tissue-specific oxidative stress and detoxification in Populus × canescens. Physiol Plant 143:50–63
- Carrasco-Gil S, Estebaranz-Yubero M, Medel-Cuesta D, Millan R, Hernandez LE (2012) Influence of nitrate fertilization on Hg uptake and oxidative stress parameters in alfalfa plants cultivated in a Hg-polluted soil. Environ Exp Bot 75:16–24
- Chen F, Gao J, Zhou Q (2012) Toxicity assessment of simulated urban runoff containing polycyclic musks and cadmium in Carassiusauratus using oxidative stress biomarkers. Environ Pollut 162:91–97
- 9. Garbisu C, Alkorta I (2001) Phytoextraction: a cost-effective plant-based technology for the removal of metals from the environment. Bioresour Technol 77(3):229–236
- Tang K, Zhan JC, Yang HR, Huang WD (2010) Changes of resveratrol and antioxidant enzymes during UV-induced plant defense response in peanut seedlings. J Plant Physiol 167:95–102
- Alvarez R, Hoyo AD, Garcia-Breijo F, Reig-Arminana J, del Campo EM, Guera A, Barreno E, Casano LM (2012) Different strategies to achieve Pb-tolerance by the two Trebouxia algae coexisting in the lichen Ramalinafarinacea. J Plant Physiol 169(18):1797–1806
- Manara A (2012) Plant responses to heavy metal toxicity. In: Furini A (ed) Plants and heavy metals. Springer briefs in molecular science. Springer, Dordrecht, pp 27–53
- 13. Viehweger K (2014) How plants cope with heavy metals. Bot Stud 55(1):35

- Patra M, Bhowmik N, Bandopadhyay B, Sharma A (2004) Comparison of mercury, lead and arsenic with respect to genotoxic effects on plant systems and the development of genetic tolerance. Environ Exp Bot 52(3):199–223
- Dalvi AA, Bhalerao SA (2013) Response of plants towards heavy metal toxicity: an overview of avoidance, tolerance and uptake mechanism. Ann Plant Sci 2(9):362–368
- Pellet DM, Grunes DL, Kochian LV (1995) Organic acid exudation as an aluminum-tolerance mechanism in maize (Zea mays L.). Planta 196(4):788–795
- Pinto AP, Simões I, Mota AM (2008) Cadmium impact on root exudates of sorghum and maize plants: a speciation study. J Plant Nutr 31(10):1746–1755
- Zhu XF, Zheng C, Hu YT et al (2011) Cadmium-induced oxalate secretion from root apex is associated with cadmium exclusion and resistance in Lycopersicon esulentum. Plant Cell Environ 34(7):1055–1064
- Yang LT, Qi YP, Jiang HX, Chen LS (2012) Roles of organic acid anion secretion in aluminium tolerance of higher plants. Biomed Res Int 2013:Article 173682
- 20. Maksymiec W (2007) Signaling responses in plants to heavy metal stress. Acta Physiol Plant 29(3):177–187
- 21. DalCorso G, Farinati S, Furini A (2010) Regulatory networks of cadmium stress in plants. Plant Signal Behav 5(6):663–667
- 22. Harir Y, Mittler R (2009) The ROS signaling network of cells. In: Reactive oxygen species in plant signaling. Springer, Berlin, pp 165–174
- Balaban RS, Nemoto S, Finkel T (2005) Mitochondria, oxidants, and aging. Cell 120(4): 483–495
- 24. Sharma P, Jha AB, Dubey RS, Pessarakli M (2012) Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. J Bot 2012:Article ID 217037
- Dodd AN, Kudla J, Sanders D (2010) The language of calcium signaling. Annu Rev Plant Biol 61:593–620
- Miller G, Schlauch K, Tam R et al (2009) The plant NADPH oxidase RBOHD mediates rapid systemic signaling in response to diverse stimuli. Sci Signal 2(84):ra45
- 27. Shiu SH, Bleecker AB (2003) Expansion of the receptor-like kinase/Pelle gene family and receptor-like proteins in Arabidopsis. Plant Physiol 132(2):530–543
- Lehti-Shiu MD, Zou C, Hanada K, Shiu SH (2009) Evolutionary history and stress regulation of plant receptor-like kinase/pelle genes. Plant Physiol 150(1):12–26
- 29. Wrzaczek M, Brosche M, Salojarvi J et al (2010) Transcriptional regulation of the CRK/ DUF26 group of receptor-like protein kinases by ozone and plant hormones in Arabidopsis. BMC Plant Biol 10:95
- 30. Idanheimo N, Gauthier A, Salojarvi J et al (2014) The Arabidopsis thaliana cysteine-rich receptor-like kinases CRK6 and CRK7 protect against apoplastic oxidative stress. Biochem Biophys Res Commun 445:457–462
- Wrzaczek M, Brosche M, Kangasjarvi J (2013) ROS signaling loops—production, perception, regulation. Curr Opin Plant Biol 16(5):575–582
- 32. Munne-Bosch S, Queval G, Foyer CH (2013) The impact of global change factors on redox signaling underpinning stress tolerance. Plant Physiol 161(1):5–19
- Van Norman JM, Breakfield NW, Benfey PN (2011) Intercellular communication during plant development. Plant Cell 23(3):855–864
- Tsukagoshi H, Busch W, Benfey PN (2010) Transcriptional regulation of ROS controls transition from proliferation to differentiation in the root. Cell 143(4):606–616
- Gill SS, Tuteja N (2010) Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. Plant Physiol Biochem 48(12):909–930
- Mahajan S, Tuteja N (2005) Cold, salinity and drought stresses: an overview. Arch Biochem Biophys 444:139–158
- Mahaja S, Sopoy SK, Tuteja N (2006) CBL-CIPK paradigm: role in calcium and stress signaling in plants. Proc Indian Nat Sci Acad 72:63–78

- Xiang Y, Huang Y, Xiong L (2007) Characterization of stress-responsive CIPK genes in rice for stress tolerance improvement. Plant Physiol 144(3):1416–1428
- Hashimoto K, Kudla J (2011) Calcium decoding mechanisms in plants. Biochimie 93(12):2054–2059
- 40. DeFalco TA, Chiasson D, Munro K, Kaiser BN, Snedden WA (2010) Characterization of GmCaMK1, a member of a soybean calmodulin-binding receptor-like kinase family. FEBS Lett 584(23):4717–4724
- Sanders D, Brownlee C, Harper JF (1999) Communicating with calcium. Plant Cell 11(4): 691–706
- 42. Tuteja N (2009) Integrated calcium signaling in plants. In: Baluška F, Mancuso S (eds) Signaling in plants. Springer, Berlin, pp 29–49
- Hetherington A, Trewavas A (1982) Calcium-dependent protein kinase in pea shoot membranes. FEBS Lett 145(1):67–71
- 44. Das R, Pandey GK (2010) Expressional analysis and role of calcium regulated kinases in abiotic stress signaling. Curr Genomics 11(1):2
- Yang T, Poovaiah BW (2003) Calcium/calmodulin-mediated signal network in plants. Trends Plant Sci 8(10):505–512
- 46. Gu Z, Ma B, Jiang Y, Chen Z, Su X, Zhang H (2008) Expression analysis of the calcineurin B-like gene family in rice (Oryza sativa L.) under environmental stresses. Gene 415(1):1–12
- 47. Tripathi V, Parasuraman B, Laxmi A, Chattopadhyay D (2009) CIPK6, a CBL-interacting protein kinase is required for development and salt tolerance in plants. Plant J 58(5): 778–790
- 48. Kaplan B, Davydov O, Knight H, Galon Y, Knight MR, Fluhr R, Fromm H (2006) Rapid transcriptome changes induced by cytosolic Ca2+ transients reveal ABRE-related sequences as Ca2+-responsive cis elements in Arabidopsis. Plant Cell 18:2733–2748
- 49. Harada A, Shimazaki KI (2006) Phototropins and blue light-dependent calcium signaling in higher plants. Photochem Photobiol 83(1):102–111
- Martinez-Noel G, Tognetti J, Nagaraj V, Wiemken A, Pontis H (2006) Calcium is essential for fructan synthesis induction mediated by sucrose in wheat. Planta 225:183–191
- Xiong L, Schumaker KS, Zhu JK (2002) Cell signaling during cold, drought, and salt stress. Plant Cell 14(1):165–183
- Cheng Y, Song C (2006) Hydrogen peroxide homeostasis and signaling in plant cells. Sci China C Life Sci 49(1):1–11
- Neill S, Desikan R, Hancock J (2002) Hydrogen peroxide signaling. Curr Opin Plant Biol 5(5):388–395
- Miller G, Shulaev V, Mittler R (2008) Reactive oxygen signaling and abiotic stress. Physiol Plant 133(3):481–489
- 55. Orozco-Cárdenas ML, Narváez-Vásquez J, Ryan CA (2001) Hydrogen peroxide acts as a second messenger for the induction of defense genes in tomato plants in response to wounding, systemin, and methyl jasmonate. Plant Cell 13(1):191
- Noctor G, Foyer CH (1998) Ascorbate and glutathione: keeping active oxygen under control. Annu Rev Plant Biol 49:249–279
- Asada K, Takahashi M (1987) Production and scavenging of active oxygen in photosynthesis. In: Kyle DJ, Osmond CB, Arntzen CJ (eds) Photoinhibition topics of photosynthesis. Elsevier, Amsterdam, pp 227–287
- Apel K, Hirt H (2004) Reactive oxygen species: metabolism, oxidative stress, and signal transduction. Annu Rev Plant Biol 55:373–399
- 59. Fedoroff N (2006) Redox regulatory mechanisms in cellular stress responses. Ann Bot 98(2):289–300
- Sanderfoot AA, Assaad FF, Raikhel NV (2000) The Arabidopsis genome. An abundance of soluble N-ethylmaleimide-sensitive factor adaptor protein receptors. Plant Physiol 124(4): 1558–1569
- Yan J, Tsuichihara N, Etoh T, Iwai S (2007) Reactive oxygen species and nitric oxide are involved in ABA inhibition of stomatal opening. Plant Cell Environ 30(10):1320–1325

- 62. Kwak JM, Mori IC, Pei ZM et al (2003) NADPH oxidase AtrobhD and AtrobhF genes function in ROS-dependent ABA signaling in Arabidopsis. EMBO J 22(11):2623–2633
- 63. Mittler R (2002) Oxidative stress, antioxidants and stress tolerance. Trends Plant Sci 7(9): 405–410
- Bethke PC, Jones RL (2001) Cell death of barley aleurone protoplasts is mediated by reactive oxygen species. Plant J 25(1):19–29
- Joo JH, Bae YS, Lee JS (2001) Role of auxin-induced reactive oxygen species in root gravitropism. Plant Physiol 126(3):1055–1060
- 66. Torres MA, Dangl JL, Jones JDG (2002) Arabidopsis gp91phox homologues Atrobhd and Atrobhf are required for accumulation of reactive oxygen intermediates in the plant defense response. Proc Natl Acad Sci U S A 99(1):517–522
- 67. Locato V, de Pinto MC, Paradiso A, de Gara L (2010) Reactive oxygen species and ascorbate glutathione interplay in signaling and stress responses. In: Gupta DS (ed) Reactive oxygen species and antioxidants in higher plants. CRC Press, New York, pp 45–64
- Jonak C, Nakagami H, Hirt H (2004) Heavy metal stress. Activation of distinct mitogenactivated protein kinase pathways by copper and cadmium. Plant Physiol 136(2):3276–3283
- Santos M, Gousseau H, Lister C, Foyer C, Creissen G, Mullineaux P (1996) Cytosolic ascorbate peroxidase from Arabidopsis thaliana L. is encoded by a small multigene family. Planta 198(1):64–69
- Goyer C, Haslekås M, Miginiac-Maslow M et al (2002) Isolation and characterization of a thioredoxin-dependent peroxidase from Chlamydomonas reinhardtii. Eur J Biochem 269(1): 272–282
- Wang Y, Wisniewski M, Meilan R, Cui M, Webb R, Fuchigami L (2005) Overexpression of cytosolic ascorbate peroxidase in tomato confers tolerance to chilling and salt stress. J Am Soc Hortic Sci 130(2):167–173
- 72. Yabuta Y, Motoki T, Yoshimura K, Takeda T, Ishikawa T, Shigeoka S (2002) Thylakoid membrane-bound ascorbate peroxidase is a limiting factor of antioxidative systems under photo-oxidative stress. Plant J 32(6):915–925
- 73. Sharma P, Dubey RS (2007) Involvement of oxidative stress and role of antioxidative defense system in growing rice seedlings exposed to toxic concentrations of aluminum. Plant Cell Rep 26(11):2027–2038
- 74. Li E, Chen L, Zeng C, Chen X, Yu N, Lai Q, Qin JG (2007) Growth, body composition, respiration and ambient ammonia nitrogen tolerance of the juvenile white shrimp, Litopenaeus vannamei, at different salinities. Aquaculture 265(1):385–390
- Rahaie M, Naghavi MR, Alizadeh H, Malboobi MA, Dimitrov K (2011) A novel DNA-based nanostructure for single molecule detection purposes. Int J Nanotechnol 8(6–7):458–470
- 76. Rahaie M, Gomarian M, Alizadeh H, Malboobi MA, Naghavi MR (2011) The expression analysis of transcription factors under long term salt stress in tolerant and susceptible wheat (Triticum aestivum L) genotypes using Reverse Northern Blot. Iranian J Crop Sci 13(3): 580–595
- 77. Eltayeb E, Kawano N, Badawi GH et al (2007) Overexpression of monodehydroascorbate reductase in transgenic tobacco confers enhanced tolerance to ozone, salt and polyethylene glycol stresses. Planta 225(5):1255–1264
- Li F, Wu QY, Sun YL, Wang LY, Yang XH, Meng QW (2010) Overexpression of chloroplastic monodehydroascorbate reductase enhanced tolerance to temperature and methyl viologenmediated oxidative stresses. Physiol Plant 139(4):421–434
- Rubio MI, Escrig I, Martinez-Cortina C, Lopez-Banet FJ, Sanz A (1994) Cadmium and nickel accumulation in rice plants: effects on mineral nutrition and possible interactions of abscisic and gibberellic acids. Plant Growth Regul 14:151–157
- Eltayeb AE, Yamamoto S, Habora MEE, Yin L, Tsujimoto H, Tanaka K (2011) Transgenic potato overexpressing Arabidopsis cytosolic AtDHAR1 showed higher tolerance to herbicide, drought and salt stresses. Breed Sci 61(1):3–10
- 81. Kwon SY, Jeong YJ, Lee HS, Kim JS, Cho KY, Allen RD, Kwak SS (2002) Enhanced tolerances of transgenic tobacco plants expressing both superoxide dismutase and ascorbate

peroxidase in chloroplasts against methyl viologen-mediated oxidative stress. Plant Cell Environ 25(7):873–882

- 82. Lim S, Kim YH, Kim SH et al (2007) Enhanced tolerance of transgenic sweetpotato plants that express both Cu Zn SOD and APX in chloroplasts to methyl viologen-mediated oxidative stress and chilling. Mol Breed 19(3):227–239
- Kwak SS, Lim S, Tang L, Kwon SY, Lee HS (2009) Enhanced tolerance of transgenic crops expressing both SOD and APX in chloroplasts to multiple environmental stress. In: Ashraf M, Ozturk M, Athar HR (eds) Salinity and water stress improving crop efficiency. Springer, Berlin, pp 197–203
- 84. Lim CJ, Hwang JE, Chen H, Hong JK, Yang KA, Choi MS et al (2007) Over-expression of the Arabidopsis DRE/CRT-binding transcription factor DREB2C enhances thermotolerance. Biochem Biophys Res Commun 362(2):431–436
- 85. Tseng MJ, Liu CW, Yiu JC (2008) Tolerance to sulfur dioxide in transgenic Chinese cabbage transformed with both the superoxide dismutase containing manganese and catalase genes of Escherichia coli. Sci Horticult 115(2):101–110
- Aono M, Saji H, Sakamoto A, Tanaka K, Kondo N, Tanaka K (1995) Paraquat tolerance of transgenic Nicotianatabacum with enhanced activities of glutathione reductase and superoxide dismutase. Plant Cell Physiol 36(8):1687–1691
- Lee YP, Kim SH, Bang JW, Lee HS, Kwak SS, Kwon SY (2007) Enhanced tolerance to oxidative stress in transgenic tobacco plants expressing three antioxidant enzymes in chloroplasts. Plant Cell Rep 26(5):591–598
- Eapen S, D'souza SF (2005) Prospects of genetic engineering of plants for phytoremediation of toxic metals. Biotechnol Adv 23(2):97–114
- Verbruggen N, LeDuc D, Vanek T (2009) Potential of plant genetic engineering for phytoremediation of toxic trace elements. Phytotechnologies solutions for sustainable land management. Eolss Publishers, Oxford
- 90. Hall JL, Williams LE (2003) Transition metal transporters in plants. J Exp Bot 54(393): 2601–2613
- Kim DY, Bovet L, Maeshima M, Martinoia E, Lee Y (2007) The ABC transporter AtPDR8 is a cadmium extrusion pump conferring heavy metal resistance. Plant J 50(2):207–218
- 92. Plaza S, Tearall KL, Zhao FJ, Buchner P, McGrath SP, Hawkesford MJ (2007) Expression and functional analysis of metal transporter genes in two contrasting ecotypes of the hyperaccumulator Thlaspicaerulescens. J Exp Bot 58(7):1717–1728
- 93. Kramer U, Talke IN, Hanikenne M (2007) Transition metal transport. FEBS Lett 581(12): 2263–2272
- Puig S, Andres-Colas NURIA, Garcia-Molina A, PeNarrubia L (2007) Copper and iron homeostasis in Arabidopsis: responses to metal deficiencies, interactions and biotechnological applications. Plant Cell Environ 30(3):271–290
- Clemens S, Aarts MG, Thomine S, Verbruggen N (2013) Plant science: the key to preventing slow cadmium poisoning. Trends Plant Sci 18(2):92–99
- 96. Milner MJ, Seamon J, Craft E, Kochian LV (2013) Transport properties of members of the ZIP family in plants and their role in Zn and Mn homeostasis. J Exp Bot 64(1):369–381
- 97. Kabata-Pendias A (2004) Soil–plant transfer of trace elements—an environmental issue. Geoderma 122(2):143–149
- Verbruggen N, Hermans C, Schat H (2009) Molecular mechanisms of metal hyperaccumulation in plants. New Phytol 181(4):759–776
- Won CKE, Cobbett CS (2009) HMA P-type ATPases are the major mechanism for root-toshoot Cd translocation in Arabidopsis thaliana. New Phytol 181(1):71–78
- 100. Courbot M, Willems G, Motte P, Arvidsson S, Roosens N, Saumitou-Laprade P, Verbruggen N (2007) A major quantitative trait locus for cadmium tolerance in Arabidopsis hallericolocalizes with HMA4, a gene encoding a heavy metal ATPase. Plant Physiol 144(2): 1052–1065
- 101. Siemianowski O, Barabasz A, Weremczuk A, Ruszczyńska A, Bulska E, Williams LE, Antosiewicz D (2013) Development of Zn-related necrosis in tobacco is enhanced by expressing AtHMA4 and depends on the apoplastic Zn levels. Plant Cell Environ 36(6):1093–1104

- 102. Menguer PK, Farthing E, Peaston KA, Ricachenevsky FK, Fett JP, Williams LE (2013) Functional analysis of the rice vacuolar zinc transporter OsMTP1. J Exp Bot 64(10): 2871–2883
- 103. Hanikenne M, Talke IN, Haydon MJ, Lanz C, Nolte A, Motte P, Krämer U (2008) Evolution of metal hyperaccumulation required cis-regulatory changes and triplication of HMA4. Nature 453(7193):391–395
- 104. Axelsen KB, Palmgren MG (2001) Inventory of the superfamily of P-type ion pumps in Arabidopsis. Plant Physiol 126(2):696–706
- 105. Delhaize E, Gruber BD, Pittman JK, White RG, Leung H, Miao Y, Richardson AE (2007) A role for the AtMTP11 gene of Arabidopsis in manganese transport and tolerance. Plant J 51(2):198–210
- 106. Mari S, Lebrun M (2006) Metal immobilization: where and how?. In: Tamas MJ, Martinoia E (eds) Molecular biology of metal homeostasis and detoxification. Springer, Berlin, pp 273–298
- 107. Bolan N, Kunhikrishnan A, Thangarajan R, Kumpiene J, Park J, Makino T, Kirkham MB, Scheckel K (2014) Remediation of heavy metal (loid) s contaminated soils—to mobilize or to immobilize? J Hazard Mater 266(2014):141–166
- 108. Kang SH, Singh S, Kim JY, Lee W, Mulchandani A, Chen W (2007) Bacteria metabolically engineered for enhanced phytochelatin production cadmium accumulation. Appl Environ Microbiol 73:6317–6320
- 109. Clemens S (2001) Molecular mechanisms of plant metal tolerance and homeostasis. Planta 212(4):475–486
- Mejáre M, Bülow L (2001) Metal-binding proteins and peptides in bioremediation and phytoremediation of heavy metals. Trends Biotechnol 19(2):67–73
- 111. Thomas M, Lieberman J, Lal A (2010) Desperately seeking microRNA targets. Nat Struct Mol Biol 17(10):1169–1174
- 112. Hassan Z, Aarts MG (2011) Opportunities and feasibilities for biotechnological improvement of Zn, Cd or Ni tolerance and accumulation in plants. Environ Exp Bot 72(1):53–63
- 113. Anjum NA, Srikanth K, Mohmood I, Sayeed I, Trindade T, Duarte AC et al (2014) Brain glutathione redox system significance for the control of silica-coated magnetite nanoparticles with or without mercury co-exposures mediated oxidative stress in European eel (Anguilla anguilla L.). Environ Sci Pollut Res 21(12):7746–7756
- 114. Anjum NA, Aref IM, Duarte AC, Pereira E, Ahmad I, Iqbal M (2014) Glutathione and proline can coordinately make plants withstand the joint attack of metal (loid) and salinity stresses. Front Plant Sci 5:662
- 115. Jozefczak M, Remans T, Vangronsveld J, Cuypers A (2012) Glutathione is a key player in metal-induced oxidative stress defenses. Int J Mol Sci 13(3):3145–3175
- 116. Seth CS, Remans T, Keunen E, Jozefczak M, Gielen H, Opdenakker K et al (2012) Phytoextraction of toxic metals: a central role for glutathione. Plant Cell Environ 35(2):334–346
- 117. Anjum NA, Hasanuzzaman M, Hossain MA, Thangavel P, Roychoudhury A, Gill SS, Rodrigo MAM, Adam V, Fujita M, Kizek R, Duarte AC, Pereira E, Ahmad I (2015) Jacks of metal/metalloid chelation trade in plants an overview. Front Plant Sci 6:192–209
- 118. Yuzefovych LV, Musiyenko SI, Wilson GL, Rachek LI (2013) Mitochondrial DNA damage and dysfunction, and oxidative stress are associated with endoplasmic reticulum stress, protein degradation and apoptosis in high fat diet-induced insulin resistance mice. PLoS One 8(1):e54059
- 119. Kotrba P, Doleckova L, de Lorenzo V, Ruml T (1999) Enhanced bioaccumulation of heavy metal ions by bacterial cells due to surface display of short metal binding peptides. Appl Environ Microbiol 65:1092–1098
- 120. Cobbett C, Meagher R (2002) Phytoremediation the Arabidopsis proteome. In: Somerville C (ed) Arabidopsis. Cold Spring Harbor Laboratory Press, New York, pp 1–22
- 121. Ramos J, Naya L, Gay M, Abian J et al (2008) Functional characterization of an unusual phytochelatin synthase, LjPCS3, of Lotus japonicus. Plant Physiol 148:536–545
- 122. Cobbett CS (2000) Phytochelatin biosynthesis and function in heavy-metal detoxification. Curr Opin Plant Biol 3(3):211–216

- 123. Sauge-Merle S, Cuine S, Carrier P, Lecomte-Pradines C, Luu DT, Peltier G (2003) Enhanced toxic metal accumulation in engineered bacterial cells expressing Arabidopsis thaliana phytochelatin synthase. Appl Environ Microbiol 69:490–494
- 124. Wojas S, Clemens S, SkŁodowska A, Antosiewicz DM (2010) Arsenic response of AtPCS1and CePCS-expressing plants—effects of external As (V) concentration on As-accumulation pattern and NPT metabolism. J Plant Physiol 167(3):169–175
- 125. Liu GY, Zhang YX, Chai TY (2011) Phytochelatin synthase of Thlaspi caerulescens enhanced tolerance and accumulation of heavy metals when expressed in yeast and tobacco. Plant Cell Rep 30:1067–1076
- 126. Verkleij JAC, Sneller FEC, Schat H (2003) Metallothioneins and phytochelatins: ecophysiological aspects. In: Abrol YP, Ahmad A (eds) Sulphur in plants. Springer, Dordrecht, pp 163–176
- 127. Yang Z, Chu C (2011) Towards understanding plant response to heavy metal stress. In: Shanker A (ed) Abiotic stress in plants—mechanisms and adaptations. InTech, Croatia, pp 59–78
- 128. Leszczyszyn OI, Imam HT, Blindauer CA (2013) Diversity and distribution of plant metallothioneins: a review of structure, properties and functions. Metallomics 5(9):1146–1169
- 129. Memon R, Aktoprakligil D, Zdemur A, Vertii A (2001) Heavy metal accumulation and detoxification mechanisms in plants. Turk J Bot 25(3):111–121
- 130. Guo JL, Xu LP, Su YC et al (2013) ScMT2-1-3, a metallothionein gene of sugarcane, plays an important role in the regulation of heavy metal tolerance/accumulation. Biomed Res Int 2013:904769
- 131. Kohler A, Blaudez D, Chalot M, Martin F (2004) Cloning and expression of multiple metallothioneins from hybrid poplar. New Phytol 164(1):83–93
- 132. Wong HL, Sakamoto T, Kawasaki T, Umemura K, Shimamoto K (2004) Down-regulation of metallothionein, a reactive oxygen scavenger, by the small GTPase OsRac1 in rice. Plant Physiol 135(3):1447–1456
- 133. Macovei L, Ventura M, Dona MF, Balestrazzi A, Carbonera D (2010) Effects of heavy metal treatments on metallothionein expression profiles in white poplar (Populusalba L.) cell suspension cultures. Fascicula Biol 18(2):274–279
- 134. Mishra S, Dubey RS (2006) Heavy metal uptake and detoxification mechanisms in plants. Int J Agric Res 1(2):122–141
- 135. Grennan AK (2011) Metallothioneins, a diverse protein family. Plant Physiol 155(4): 1750–1751
- Du J, Yang JL, Li CH (2012) Advances in metallotionein studies in forest trees. Plant Omics 5(1):46–51
- 137. Kumar SV, Tan SG, Quah SC, Yusoff K (2002) Isolation and characterization of seven tetranucleotide microsatellite loci in mungbean, Vigna radiata. Mol Ecol Notes 2(3):293–295
- 138. Zhigang A, Cuijie L, Yuangang Z, Yejie D, Wachter A, Gromes R, Rausch T (2006) Expression of BjMT2, a metallothionein 2 from Brassica juncea, increases copper and cadmium tolerance in Escherichia coli and Arabidopsis thaliana, but inhibits root elongation in Arabidopsis thaliana seedlings. J Exp Bot 57(14):3575–3582
- 139. Hossain MA, Piyatida P, da Silva JAT, Fujita M (2012) Molecular mechanism of heavy metal toxicity and tolerance in plants: central role of glutathione in detoxification of reactive oxygen species and methylglyoxal and in heavy metal chelation. J Bot 2012, Article ID 872875
- 140. Kangasjarvi S, Kangasjarvi J (2014) Towards understanding extracellular ROS sensory and signaling systems in plants. Adv Bot 2014:Article ID 538946
- 141. Pottosin I, Velarde-Buendía AM, Bose J, Zepeda-Jazo I, Shabala S, Dobrovinskaya O (2014) Cross-talk between reactive oxygen species and polyamines in regulation of ion transport across the plasma membrane: implications for plant adaptive responses. J Exp Bot 65(5):1271–1283
- 142. Xing Y, Jia W, Zhang J (2008) AtMKK1 mediates ABA-induced CAT1 expression and H2O2 production via AtMPK6-coupled signaling in Arabidopsis. Plant J 54(3):440–451

- 143. Velarde-Buendía AM, Shabala S, Cvikrova M, Dobrovinskaya O, Pottosin I (2012) Saltsensitive and salt-tolerant barley varieties differ in the extent of potentiation of the ROSinduced K+ efflux by polyamines. Plant Physiol Biochem 61:18–23
- 144. Hirayama T, Shinozaki K (2007) Perception and transduction of abscisic acid signals: keys to the function of the versatile plant hormone ABA. Trends Plant Sci 12(8):343–351
- 145. Xuan Y, Zhou S, Wang L, Cheng Y, Zhao L (2010) Nitric oxide functions as a signal and acts upstream of AtCaM3 in thermotolerance in Arabidopsis seedlings. Plant Physiol 153(4): 1895–1906
- 146. Liu HT, Sun DY, Zhou RG (2005) Ca2+ and AtCaM3 are involved in the expression of heat shock protein gene in Arabidopsis. Plant Cell Environ 28(10):1276–1284
- 147. Wu HC, Luo DL, Vignols F, Jinn TL (2012) Heat shock-induced biphasic Ca2+ signature and OsCaM1-1 nuclear localization mediate downstream signalling in acquisition of thermotolerance in rice (Oryza sativa L.). Plant Cell Environ 35(9):1543–1557
- 148. Park HC, Kim ML, Kang YH, Jeon JM, Yoo JH, Kim MC et al (2004) Pathogen-and NaClinduced expression of the SCaM-4 promoter is mediated in part by a GT-1 box that interacts with a GT-1-like transcription factor. Plant Physiol 135(4):2150–2161
- 149. Liu HT, Li B, Shang ZL, Li XZ, Mu RL, Sun DY et al (2003) Calmodulin is involved in heat shock signal transduction in wheat. Plant Physiol 132:1186–1195
- 150. Park HC, Park CY, Koo SC, Cheong MS, Kim KE, Kim MC, Chung WS (2010) AtCML8, a calmodulin-like protein, differentially activating CaM-dependent enzymes in Arabidopsis thaliana. Plant Cell Rep 29(11):1297–1304
- 151. Magnan F, Ranty B, Charpenteau M, Sotta B, Galaud JP, Aldon D (2008) Mutations in AtCML9, a calmodulin-like protein from Arabidopsis thaliana, alter plant responses to abiotic stress and abscisic acid. Plant J 56(4):575–589
- 152. Yamaguchi T, Aharon GS, Sottosanto JB, Blumwald E (2005) Vacuolar Na+/H+ antiportercation selectivity is regulated by calmodulin from within the vacuole in a Ca2+- and pHdependent manner. Proc Natl Acad Sci U S A 102(44):16107–16112
- 153. Delk NA, Johnson KA, Chowdhury NI, Braam J (2005) CML24, regulated in expression by diverse stimuli, encodes a potential Ca2+ sensor that functions in responses to abscisic acid, daylength, and ion stress. Plant Physiol 139:240–253
- 154. Vanderbeld B, Snedden WA (2007) Developmental and stimulus-induced expression patterns of Arabidopsis calmodulin-like genes CML37, CML38 and CML39. Plant Mol Biol 64: 683–697
- 155. Vadassery J, Reichelt M, Hause B, Gershenzon J, Boland W, Mithofer A (2012) CML42mediated calcium signaling coordinates responses to Spodopteraherbivory and abiotic stresses in Arabidopsis. Plant Physiol 159:1159–1175
- 156. Xu GY, Rocha PS, Wang ML, Xu ML, Cui YC, Li LY et al (2011) A novel rice calmodulinlike gene, OsMSR2, enhances drought and salt tolerance and increases ABA sensitivity in Arabidopsis. Planta 234:47–59
- 157. Liu HT, Li GL, Chang H, Sun DY, Zhou RG, Li B (2007) Calmodulin-binding protein phosphatase PP7 is involved in thermotolerance in Arabidopsis. Plant Cell Environ 30: 156–164
- 158. Liu HT, Gao F, Li GL, Han JL, Liu DL, Sun DY et al (2008) The calmodulin-binding protein kinase 3 is part of heat-shock signal transduction in Arabidopsis thaliana. Plant J 55: 760–773
- Yang T, Chaudhuri S, Yang L, Chen Y, Poovaiah BW (2004) Calcium/calmodulin Up-regulates a cytoplasmic receptor-like kinase in plants. J Biol Chem 279:42552–42559
- 160. Yang T, Poovaiah BW (2000) An early ethylene up-regulated gene encoding a calmodulinbinding protein involved in plant senescence and death. J Biol Chem 275:38467–38473
- 161. Yang T, Poovaiah BW (2000) Molecular and biochemical evidence for the involvement of calcium/calmodulin in auxin action. J Biol Chem 275:3137–3143
- 162. Shi B, Ni L, Zhang A, Cao J, Zhang H, Qin T et al (2012) OsDMI3 is a novel component of abscisic acid signaling in the induction of antioxidant defense in leaves of rice. Mol Plant 5:1359–1374

- 163. Ma FF, Lu R, Liu HY, Shi B, Zhang JH, Tan MP et al (2012) Nitric oxide-activated calcium/ calmodulin-dependent protein kinase regulates the abscisic acid-induced antioxidant defence in maize. J Exp Bot 63:4835–4847
- 164. Pandey S, Tiwari SB, Tyagi W, Reddy MK, Upadhyaya KC, Sopory SK (2002) A Ca2+/CaMdependent kinase from pea is stress regulated and in vitro phosphorylates a protein that binds to AtCaM5 promoter. Eur J Biochem 269:3193–3204
- Chen Z, Gallie DR (2006) Dehydroascorbate reductase affects leaf growth, development, and function. Plant Physiol 142:775–787
- 166. Ushimaru T, Nakagawa T, Fujioka Y, Daicho K, Naito M, Yamauchi Y, Murata N (2006) Transgenic Arabidopsis plants expressing the rice dehydroascorbate reductase gene are resistant to salt stress. J Plant Physiol 163(11):1179–1184
- 167. Delhaize E, Kataoka T, Hebb DM, White RG, Ryan PR (2003) Genes encoding proteins of the cation diffusion facilitator family that confer manganese tolerance. Plant Cell 15(5): 1131–1142