

Chapter 5

Sepsis-Induced Immune Suppression

Nicholas Csikesz and Nicholas S. Ward

Introduction

Newton's third law is commonly quoted as "for every action, there is an equal and opposite reaction." This concept was largely ignored in the initial theories regarding the body's response to overwhelming infection, which was characterized as a surge of inflammation that arises to eliminate invading pathogens, and may injure the host organism along with it. This leads to the concept that it may be possible to ameliorate the organ damage in sepsis by reining in this unmitigated inflammatory response. Indeed, many early preclinical studies showed some initial promise in mitigating sepsis mortality in animals by limiting the pro-inflammatory response. However, a large number of clinical trials in humans of a variety of anti-inflammatory therapies failed to demonstrate any improvement in the high mortality associated with sepsis [1].

In 1996, Roger Bone invoked Isaac Newton in a landmark editorial discussing possible explanations for the failure of anti-inflammatory therapies in sepsis [2]. Bone and others recognized that there was a growing body of evidence demonstrating immunosuppression following other forms of inflammation such as surgery or trauma. He reviewed the existing evidence showing that the immune response to sepsis is not nearly as straightforward as was originally held. In addition to the pro-inflammatory response to sepsis, there is a concomitant anti-inflammatory response. He postulated that the balance between these two intertwined responses largely

N. Csikesz (✉)

Alpert Medical School of Brown University, Rhode Island Hospital,
593 Eddy Street, Providence, RI 02903, USA
e-mail: ncsikesz@lifespan.org

N.S. Ward

Division of Pulmonary, Critical Care, and Sleep Medicine, Alpert/Brown Medical School,
Providence, RI, USA
e-mail: nward@lifespan.org

determines the outcome in sepsis and coined the term “Compensatory Anti-inflammatory Response Syndrome” (CARS) to describe the anti-inflammatory response.

Thus was created a more nuanced view of sepsis as a pathologic dysregulation of the immune system with both pro- and anti-inflammatory effects. Bone hypothesized that overwhelming infection disrupts the body’s normal homeostasis such that at different points within the time course of infection pro-inflammatory or anti-inflammatory forces may predominate, either of which can be pathologic and contribute to mortality. The most straightforward theory resulting from these ideas holds that the early course of sepsis is characterized by the traditional pro-inflammatory response, with capillary leak, organ dysfunction, and (if left unchecked) death. This is then followed by a period of immune suppression during which the body is susceptible to secondary infections (and subsequent death) before returning to homeostasis. The relative strength of the pro- and anti-inflammatory responses likely depends on the host and the pathogen, as well as on external interventions (i.e., medical care), resulting in multiple potential immunologic responses.

Two terms critical to the discussion of this topic are immunosuppression and immunoparalysis. For the purposes of this chapter, immunosuppression should be considered to be the active anti-inflammatory responses (such as increased secretion of anti-inflammatory cytokines, and increases in immune suppressor cell populations). Immunoparalysis should be considered the loss of any discernible function that occurs in some cell populations in this process, i.e., anergy. There is now overwhelming evidence demonstrating the clinical importance of sepsis-induced immune suppression along with the general idea of an initial pro-inflammatory state giving way to a later anti-inflammatory state. Almost 2/3 of deaths due to sepsis occur after hospital day 5, during a phase marked by an increase in the percentage of positive cultures due to normally nonpathogenic organisms [3]. Critically ill patients have also been shown to have high rates of reactivation of dormant viruses such as cytomegalovirus (CMV) and herpes simplex virus (HSV) [4, 5]. This chapter will review the current understanding of sepsis-induced immune suppression.

Cytokines in Sepsis-Induced Immune Suppression

Cytokines play a significant role in mediating the immune response of the body. While specific pathways of activation of individual cytokines are nuanced, many can be generally labeled as either pro-inflammatory or anti-inflammatory. Some well characterized pro-inflammatory cytokines include interleukin 1 (IL-1), IL-6, IL-12, tumor necrosis factor α (TNF α), and interferon γ (IFN γ). Anti-inflammatory cytokines include IL-4, IL-10, and transforming growth factor β (TGF β) although these cytokines can have proinflammatory effects in other conditions [6, 7]. Both pro- and anti-inflammatory cytokine production is stimulated early on in response to infection.

An illustrative study demonstrating this was done by Novotny et al. [8]. Measurements of IL-6 and IL-10 were taken from human patients with postoperative sepsis as well as in a murine model of septic peritonitis. The initial immune response in both study populations was characterized by concomitant increases in IL-6 and IL-10. Further, the relationship was exponential such that a linear increase in the pro-inflammatory cytokine IL-6 correlated with an exponential increase in the anti-inflammatory cytokine IL-10. This suggests that the body engages in an immediate attempt to rein in the pro-inflammatory response that is unleashed in response to infection.

IL-10 is currently thought to be the most important of the anti-inflammatory cytokines [6]. It was first characterized around 1990 when it was shown to regulate T-cell populations [9, 10]. It has now been established that IL-10 has multiple immunosuppressive roles, with most important being the downregulation of TNF [11]. In animal models of sepsis, the administration of IL-10 has been shown to have both positive [12–14] and negative [15, 16] effects on outcome, which likely depend on the time of administration and the severity of the infection. In one carefully done animal model, Ashare and colleagues [17] followed levels of pro-inflammatory and anti-inflammatory cytokines throughout the whole course of sepsis in mice. They found that bacterial levels in tissue correlated with IL-10 levels and that if the pro-inflammatory response was blocked by pretreatment with IL-1 receptor antagonist, bacterial levels were higher, as was mortality. Similarly, Song and colleagues [18] showed that blocking IL-10 activity early had no effect on mortality, whereas blocking it late (12 h) after sepsis improved mortality. This suggests that in the pro-inflammatory milieu of early sepsis, IL-10 does not have a major role, whereas in the later phase of disease when immunosuppression predominates, its effect is more pronounced.

The Role of Immune Cells in Sepsis-Induced Immunosuppression

Impaired Immune Cell Function and Programmed Cell Death in Sepsis

Activated by triggers such as antigens or inflammatory cytokines, immune cells speed the death and clearance of infectious organisms in sepsis. This immune protection comes at a cost however, as immune cell activity can lead to tissue and organ injury through the release of anti-infective products such as oxidants. As the sepsis inflammatory cascade develops, the body begins the process of inhibiting these immune cells through two processes, deactivation of immune cell function and apoptosis.

Some of the earliest studies in impaired immunity in states of inflammation noted impaired immune cell function manifest as anergy [19]. Later studies were able to characterize an array of immune cell dysfunction that accompanies severe inflammation (reviewed below). Cell death is a common process and can occur via two pathways, apoptosis or necrosis. Apoptosis, or programmed cell death, is a carefully regulated process by which the body can allow for cell turnover without inducing inflammation (as occurs with necrosis) [20]. Apoptosis of immune effector cells is an important mechanism by which the body regulates the intensity and duration of a pro-inflammatory state. Many animal and human studies in sepsis have shown extensive apoptosis of immune effector cells including B and CD4+ T lymphocytes, dendritic cells, and epithelial cells [21–24]. Additionally, the subsequent burden that clearing these apoptotic cells plays on the remaining immune cells is thought to be a major contributor to immunoparalysis.

Neutrophils

Neutrophils play a critical role in the body's response to infection, so it is not surprising that they are intricately involved in the balance between pro- and anti-inflammatory pathways in sepsis [25]. In contrast to many other immune cells, the apoptosis of neutrophils is down-regulated in sepsis [26, 27]. This was shown by Tamayo et al. in a prospective observational study of 80 septic patients and 25 healthy controls [27]. The rates of neutrophil apoptosis were decreased at 24 h, 5 days, and 12 days after diagnosis of sepsis in comparison to controls. There was no difference seen between survivors and non-survivors of sepsis.

Additionally, immature neutrophils are released in a large number from the bone marrow, resulting in the neutrophilia with bandemia seen in many patients presenting with sepsis [28]. These immature neutrophils are immunologically active. This was shown in a prospective observational study by Drifte et al. [28]. Whole blood from 33 ICU patients with sepsis, 12 ICU patients with SIRS, and 32 healthy volunteers was taken and immune function was assessed *in vitro*. Immature neutrophils were able to engage in phagocytosis and bacterial killing via production of reactive oxygen species, although less efficiently than mature neutrophils. Interestingly, immature neutrophils exhibited a more pro-inflammatory state, as evidenced by an elevated TNF-alpha/IL-10 ratio. This is important as mature neutrophils have been shown to be involved in the anti-inflammatory response of sepsis via the production of IL-10 [29]. This balance between a pro-inflammatory and anti-inflammatory state in neutrophils has been shown to predict the development of secondary infections that are so often the actual cause of mortality in sepsis. Stephan et al. studied *in vitro* neutrophil function in patients with sepsis 4 days after they had been admitted [30]. They found that those patients who subsequently developed a nosocomial infection exhibited impaired phagocytosis and bactericidal killing at day 4.

Antigen Presenting Cells

Dendritic Cells

Antigen presenting phagocytes, including dendritic cells and macrophages, play a critical role in the immune response to infection [20, 31, 32]. Tinsley et al. investigated dendritic cell populations in a murine model of sepsis [24]. They found a rapid proliferation in follicular dendritic cells in the first 36 h after infection that was followed by extensive apoptosis, resulting in a net decrease in dendritic cell numbers by 48 h. Fujita et al. identified a subset of regulatory dendritic cells that activate anti-inflammatory pathways via the secretion of IL-10 [32]. Additional studies have also identified a shift in phenotype of dendritic cells toward an anti-inflammatory pathway in patients with sepsis-induced immunosuppression [33, 34]. These findings are in keeping with the model of sepsis as a balance between pro- and anti-inflammatory immune responses.

Multiple studies have examined the importance of dendritic cells to the immune response to sepsis. In a straightforward study, Guisset et al. measured peripheral blood dendritic cell counts in patients with sepsis [35]. They found that an early decrease in dendritic cell numbers correlated strongly with subsequent mortality and hypothesized that this may be a useful prognostic biomarker in septic patients. Toliver-Kinsky et al. have extensively studied the ability of a dendritic cell growth factor, FLT3 ligand, to impact murine resistance to pseudomonas infection in burns. They have shown that FLT3 ligand improves murine resistance to infection via improved neutrophil function in a dendritic-cell-dependent manner [36, 37].

Monocytes/Macrophages

In contrast to dendritic cells, macrophage populations are not reduced in response to sepsis [31]. However, numerous studies have demonstrated that monocytes/macrophages effectively undergo "cellular reprogramming" with a transition from a pro-inflammatory, immune activating response to an immunosuppressing anti-inflammatory response [38, 39]. Indeed, the impaired monocyte response to lipopolysaccharide (LPS), the immunogenic cell membrane component of gram-negative bacteria, seen in sepsis, labeled "endotoxin tolerance," is considered to be a hallmark of the disorder. These alterations in monocyte function were shown in a study by Munoz et al. using plasma from patients in the ICU with sepsis or non-septic shock [40]. Monocytes isolated from patients with sepsis exhibited impaired release of IL-1, IL-6, and TNF-alpha in response to LPS exposure. Further, sepsis survivors recovered their capacity to respond to LPS exposure whereas non-survivors did not. Monneret et al., while investigating the anti-inflammatory response in sepsis, showed a down-regulation of HLA-receptor expression on monocytes which correlated strongly with levels of the anti-inflammatory cytokine IL-10 [41].

Lymphocytes

Natural Killer Cells

Lymphopenia is a frequent finding in sepsis with reductions in all lymphocyte subtypes [42]. In addition to a reduction in cell number, there is also evidence for cellular reprogramming similar to that seen in antigen presenting cells. Natural Killer (NK) cells are part of the innate immune response and were originally described based on their ability to kill leukemic cells [43]. They play an important role in the early response to infection, largely through production of IFN γ . Similarly to other lymphocyte populations, a reduction in NK cell numbers has been demonstrated in septic patients [42, 44]. A higher percentage of NK cells correlates with improved survival in septic patients [45]. Sepsis may result in NK cell tolerance to antigen stimulation, resulting in impaired production of IFN γ [44, 46, 47].

Chiche et al. demonstrated the clinical importance of this in a study of CMV reactivation in septic patients in the ICU [48]. Patients with sepsis admitted to the ICU were followed prospectively with serial monitoring of NK cell function while assessing for CMV reactivation. At baseline there was no difference in NK cell effector function between cases and controls. However, prior to CMV reactivation, there was a decrease in the ability of NK cells to secrete IFN-gamma (and elevations in serum IL-10 and IL-15 levels).

CD4+ T_H Cells

CD4+ helper T cells are typically divided into two distinct subtypes, Th1 and Th2, based on their pattern of cytokine secretion in response to stimulation. Th1 cells release pro-inflammatory cytokines including TNF-a, IFN-g, and IL-2, whereas Th2 cells release anti-inflammatory cytokines including IL-4 and IL-10 [6]. Sepsis has been shown in some studies to result in a shift toward a pro-Th2 response with the release of more anti-inflammatory cytokines, whereas other studies have shown an overall suppression of both Th1 and Th2 cells [6, 39]. These findings are consistent with the development of both immune suppression and immunoparalysis.

A third population of helper T cells, Th17 cells, has also been identified as playing an important role in sepsis-induced immune suppression. These cells are important in protecting against extracellular bacterial and fungal infections via secretion of IL-17 and IL-22. Th17 cells are also decreased in septic patients and loss of effective Th17 function is thought to contribute to the occurrence of secondary fungal infections in sepsis-induced immune suppression [39, 42, 49].

$\gamma\delta$ T Cells

$\gamma\delta$ T cells are present in high numbers in the intestinal mucosa and can be considered first line defenders against particular pathogens. In 2005, Venet et al. showed that $\gamma\delta$ T cells decrease in patients with sepsis [50]. In the largest study to date, Andreu-Ballester et al. studied 135 patients with sepsis from an emergency department and intensive care unit. All the $\gamma\delta$ T cell populations decreased significantly as the septic picture worsened. Almost 20% of patients died and the $\gamma\delta$ T cells were significantly reduced in those septic patients who died. In this study, $\gamma\delta$ T cells showed the largest decrease of any T cell population and the reduction correlated with sepsis severity [51].

Regulatory T Cells

Regulatory T cells (T_{Reg}), formerly known as suppressor T cells, are a subpopulation of CD4+ T cells that play an important role in modulating the immune response [39]. T_{Reg} levels are elevated in patients with sepsis compared to controls, and higher levels (along with more immunoparalysis) are seen in non-survivors compared to survivors [52, 53]. Several studies have attempted to elucidate the mechanisms by which T_{Reg} cells may directly contribute to immunoparalysis. T_{Reg} cells have been shown to redirect monocytes and macrophages into an anti-inflammatory alternative activating macrophages (AAM) pathway. This was partly through T_{Reg} production of IL-10, but also through a cytokine-independent pathway [54]. T_{Reg} cells also inhibit the memory $\gamma\delta$ T cell production of IFN- γ in response to antigen challenge [55].

Predicting Clinical Outcomes with Biomarkers of Sepsis-Induced Immunosuppression

There have been many efforts to study the timing and magnitude of the immunosuppressive response in relation to patient outcomes which would create effective biomarkers for prognosis and therapies [56]. In 1983, Keane et al. studied lymphocytes cultured from 31 patients with severe trauma. They found that, overall, lymphocyte response to stimulation with mitogens was markedly reduced from controls. Furthermore, responses were lower and the duration of suppression longer in those patients who became infected, and the suppression of response preceded the onset of infection. Extremely low responses were found in three patients who later died [19].

More commonly, studies of patient's monocytes and their own HLA receptor down regulation have shown promise as a biomarker [57–66]. Asadulla et al. studied 57 neurosurgical patients and found that HLA-DR expression was lower in 14 patients who developed infection, compared with patients with an uncomplicated

postoperative course [57]. Out of ten patients with less than 30% HLA-DR positive monocytes, nine developed infection. They hypothesized that the mechanism of this down regulation was high levels of endogenous cortisol as the effect coincided with high ACTH and cortisol concentrations and similar down regulation was seen in other patients who received high doses of exogenous corticosteroids. Subsequent studies supported the theory that the magnitude of HLA-DR receptor down regulation predicted a variety of other poor outcomes such as sepsis in liver transplant patients [58], however that study was confounded by exogenous steroids in some patients [60]. Allen et al. found HLA levels predicted sepsis in pediatric cardiac surgery patients [67]. In a small study of septic adults, Su et al. found that levels of HLA-DR positive monocytes <30% were more predictive of mortality than APACHE II scores [65].

The predictive power of monocyte deactivation has not been shown consistently, however and more recent studies have yielded different results. In 2003, three papers were published that seemed to contradict earlier findings. Hynninen et al. evaluated the HLA-DR expression of 61 patients with sepsis at admission and showed no predictive power of HLA expression for survival [61]. Another study of 70 septic patients also found no correlation between HLA expression and infectious or mortality outcomes [63]. Interestingly, this study showed that if patients' monocytes were stimulated with G-CSF ex-vivo, their HLA expression increased. The third study looked at 85 cardiac surgery patients. HLA expression was measured at presurgery, immediately after and 1 day later. Their data showed that while all patients' HLA levels declined after surgery, the magnitude of the response did not correlate with sepsis/SIRS, or other infectious complications [62]. Reasons for the different results are unclear but may be the result of small sample sizes, timing, or well-described variation caused by the different laboratory techniques used. In one study, the same samples were analyzed in two different labs and differed by as much as 20% [62].

Other studies have looked at anti-inflammatory cytokine levels as predictors of poor outcomes; most of these studies have been on human patients and bore mixed results. These data likely reflect the varied magnitudes and time courses of both pro and anti-inflammatory cytokine expression in real patients. In 1998, Doughty et al. sampled 53 pediatric ICU patients and found that high IL-10 levels correlated with three or more organ dysfunction and mortality [68]. Ahlstrom found no predictive value in IL-10 levels in patients with SIRS [64] but Simmons et al. found that IL-10 levels did correlate with mortality in a sample of 93 critically ill patients with acute renal failure [69]. Perhaps the most interesting data comes from two studies that looked at the ratio of IL-10 to TNF. In a large study of over 400 patients admitted to the hospital for fever, van Dissel et al. showed that a higher IL-10 to TNF ratio was predictive of mortality [70]. A similar study by Gogos et al. in a population of patients with mixed sepsis showed the same results [56].

A postmortem study by Boomer et al. compared patients who died in the intensive care unit (ICU) from severe sepsis with control patients who died of other causes. They found evidence of both immunosuppression and immunoparalysis in patients who died of sepsis. There was a marked change in the balance between suppressor

cell populations and immunogenic cell lines including splenic CD4, CD8, and HLA-DR cells. Additionally, they found that patients who died of sepsis had <10% of the levels of both pro-inflammatory cytokines (TNF- α , IFN- γ , IL-6) and anti-inflammatory (IL-10) cytokines compared to patients dying of other causes [71].

Potential Therapeutic Interventions in Sepsis-Induced Immune Suppression

Decades of research in sepsis-induced immunosuppression has fostered the concept that both the pro and anti-inflammatory responses are necessary for recovery from an overwhelming infection and that it is the imbalance of these forces that can lead to organ injury and death. This has led some investigators to explore manipulation of these systems to improve outcomes. While some animal studies in this field have shown positive results, many have not and it is clear that in a system as complicated as sepsis the timing and dose of any agent used to affect the degree of pro or anti-inflammatory response are crucial factors.

Some of the first agents used to manipulate the balance of immunosuppression were androgens and estrogens. The idea for hormonal therapy came from earlier studies showing that testosterone seemed to have a negative impact on sepsis and trauma outcomes and is believed to act through augmenting postinjury immunosuppression [72]. Two subsequent studies by the same investigators showed that administration of the estrogen-like drug DHEA reduced the immunosuppression and improved mortality in septic mice [73, 74]. By far, most of the studies that have tried to manipulate the balance of inflammation have involved using anti-inflammatory cytokines that are here reviewed.

Interleukin-10

In animal models of sepsis, the administration of IL10 has been shown to have both positive [12–14] and negative [15, 16] effects on outcome which likely are dependent on the time of administration and the severity of the infection. In one carefully done animal model, Ashare et al. followed levels of pro-inflammatory cytokines and anti-inflammatory cytokines throughout the whole course of sepsis in mice. They found that bacterial levels in tissue correlated with IL-10 levels and that if the complementary pro-inflammatory response was blocked by pretreatment with IL-1 receptor antagonist, bacterial levels were higher as was mortality [17]. Similarly, Song et al. showed that blocking IL-10 activity early had no effect on mortality, blocking it late (12 h) after sepsis improved mortality [18]. These studies help illustrate how IL10 helps maintain a careful balance of the immune system in inflammation; thus, manipulation of it is so dangerous.

Interleukin 7 (IL-7)

Interleukin 7 may be the immunomodulatory agent that is closest to clinical utility in sepsis as it has shown positive results in animal trials of sepsis as well as has been shown to be safe in clinical trials in humans for other disease processes. IL-7 plays a critical role in T cell function. Mutations in IL-7 are one of the causes of severe combined immunodeficiency (SCID) [75–77]. In a murine model of sepsis, IL-7 has been shown to prevent loss of T-cells by both decreasing T-cell apoptosis and increasing T-cell proliferation. This prevented the loss of delayed type hypersensitivity (DTH) response and improved overall survival [78, 79]. A phase I/IIA study in humans with HIV and persistent lymphopenia despite combination anti-retroviral therapy demonstrated sustained increases in CD4 and CD8 T cells without signs of a hyper-inflammatory response or other adverse effects [80].

Interleukin 15 (IL-15)

IL-15 is closely related to IL-7 [78]. It is an anti-apoptotic cytokine that is regarded as a promising immunomodulatory therapy in cancer [81]. In murine models of sepsis, it has been shown to block apoptosis through the BCL-2 pathway, resulting in improved IFN γ production and reversal of sepsis-induced immunosuppression, which leads to improved survival [82]. Technical challenges related to rapid renal clearance of its recombinant form have limited its efficacy in human studies to date, but further trials are ongoing [81].

Programmed Cell Death Receptor-1 (PD-1) and Programmed Cell Death Ligand-1 (PD-L1)

PD-1 and PD-L1 are part of a family of co-inhibitory cell surface molecules that have been studied as another promising immunomodulatory therapy. PD-1 is expressed by activated T-cells and PD-L1 by epithelial, endothelial, and antigen presenting cells. PD-1/PD-L1 binding is thought to be part of the negative feedback loop that is triggered by an activation of the immune system. In the presence of prolonged antigen presence, this may lead to T cell exhaustion. Blocking the PD-1/PD-L1 pathway has shown extreme promise in cancer immunotherapy in human trials [83, 84]. In studies of human septic patients, PD-1 and PD-L1 expression has been shown to be upregulated in T cells and monocytes respectively [85, 86]. Further, levels of expression of PD-1 and PD-L1 correlated with increased secondary infections and mortality [86]. In murine models of bacterial sepsis as well as in both primary and secondary fungal sepsis, blockade of this pathway has resulted in improved survival [87, 88]. Human studies of anti-PD-1 and anti-PD-L1 have not yet been performed.

Granulocyte Macrophage Colony-Stimulating Factor (GM-CSF)

Loss of immune cells is an important factor in sepsis-induced immune suppression. Stimulating immune cell production has been looked at as a therapeutic target. Initial studies focused on granulocyte stimulating factor (G-CSF), which resulted in a marked increase in neutrophil number, but no change in clinical outcomes [89, 90]. Given the loss of cell types other than neutrophil as well as the importance of non-neutrophil immune cells in fighting the secondary viral and fungal infections that are common in sepsis-induced immune suppression, subsequent studies have focused on inducing a broader immune response with GM-CSF. Two such studies in humans have shown promising results, and are also notable for their use of a biomarker-based approach for identifying patients in the immunosuppressed phase of sepsis [91, 92]. While neither study was powered to show a difference in mortality, both showed improvement in markers of immune function (specifically TNF- α production) in patients treated with GM-CSF.

Interferon-Gamma

IFN γ is a key downstream mediator activating the innate immune response. IFN γ levels are decreased in patients with sepsis-induced immune suppression. Administration of IFN γ in uncontrolled studies has been shown to restore immune function [93, 94]. At least five studies have examined the use of gamma interferon which has been shown in-vivo to reverse monocyte deactivation [95, 96]. Two very similar small trials were done on human subjects with sepsis [93, 97]. In both studies, subjects with sepsis and monocyte HLA-DR expression of 30% or less were given interferon gamma. Both groups reported increases in HLA-DR expression, usually after just one dose. One of the studies also examined the monocytes ex-vivo and showed that interferon improved monocyte cytokine production as well [93]. A third human trial was different in that it sought to study the effects of Interferon gamma regionally [98]. In this study, the authors selected 21 patients with severe trauma and alveolar macrophage dysfunction as determined by a bronchoalveolar lavage sample showing macrophage HLA-DR expression of 30% or less. Interferon gamma was administered via inhalation. They found about 50% of the subjects had an increase in their alveolar macrophage HLA-DR expression. These patients had a lower incidence of pneumonia but no other differences in outcomes. The small numbers and lack of a control population in all three of these studies limit the conclusions that can be drawn, especially since HLA-DR expression is known to increase as patients recover. A small randomized controlled trial of IFN γ in sepsis is ongoing ([ClinicalTrials.gov](https://clinicaltrials.gov) number: NCT01649921).

Conclusion

Sepsis-induced immune suppression is likely a major contributor to the morbidity and mortality associated with sepsis. It is characterized by a decrease in immune effector cell number as well as loss of function, which results in increased susceptibility to secondary infections. Potential therapies to augment the immune response show promise as a means to decrease sepsis-related mortality but large randomized controlled trials have not yet been done.

References

1. Freeman BD, Natanson C. Anti-inflammatory therapies in sepsis and septic shock. *Expert Opin Investig Drugs*. 2000;9(7):1651–63.
2. Bone RC. Sir Isaac Newton, sepsis, SIRS, and CARS. *Crit Care Med*. 1996;24(7):1125–8.
3. Otto GP, Sossdorf M, Claus RA, Rödel J, Menge K, Reinhart K, et al. The late phase of sepsis is characterized by an increased microbiological burden and death rate. *Crit Care*. 2011;15(4):R183.
4. Luyt CE, Combes A, Deback C, Aubriot-Lorton MH, Nieszkowska A, Trouillet JL, et al. Herpes simplex virus lung infection in patients undergoing prolonged mechanical ventilation. *Am J Respir Crit Care Med*. 2007;175(9):935–42.
5. Limaye AP, Kirby KA, Rubenfeld GD, Leisenring WM, Bulger EM, Neff MJ, et al. Cytomegalovirus reactivation in critically ill immunocompetent patients. *JAMA*. 2008;300(4):413–22.
6. Ward NS, Casserly B, Ayala A. The compensatory anti-inflammatory response syndrome (CARS) in critically ill patients. *Clin Chest Med*. 2008;29(4):617–25. viii
7. Schulte W, Bernhagen J, Bucala R. Cytokines in sepsis: potent immunoregulators and potential therapeutic targets—an updated view. *Mediators Inflamm*. 2013;2013:165974.
8. Novotny AR, Reim D, Assfalg V, Altmayr F, Friess HM, Emmanuel K, et al. Mixed antagonist response and sepsis severity-dependent dysbalance of pro- and anti-inflammatory responses at the onset of postoperative sepsis. *Immunobiology*. 2012;217(6):616–21.
9. MacNeil IA, Suda T, Moore KW, Mosmann TR, Zlotnik A. IL-10, a novel growth cofactor for mature and immature T cells. *J Immunol*. 1990;145(12):4167–73.
10. O’Garra A, Stapleton G, Dhar V, Pearce M, Schumacher J, Rugo H, et al. Production of cytokines by mouse B cells: B lymphomas and normal B cells produce interleukin 10. *Int Immunol*. 1990;2(9):821–32.
11. Oberholzer A, Oberholzer C, Moldawer LL. Interleukin-10: a complex role in the pathogenesis of sepsis syndromes and its potential as an anti-inflammatory drug. *Crit Care Med*. 2002;30(1 Supp):S58–63.
12. Berg DJ, Kuhn R, Rajewsky K, Müller W, Menon S, Davidson N, et al. Interleukin-10 is a central regulator of the response to LPS in murine models of endotoxemic shock and the Shwartzman reaction but not endotoxin tolerance. *J Clin Invest*. 1995;96(5):2339–47.
13. Howard M, Muchamuel T, Andrade S, Menon S. Interleukin 10 protects mice from lethal endotoxemia. *J Exp Med*. 1993;177(4):1205–8.
14. van der Poll T, Jansen PM, Montegut WJ, Braxton CC, Calvano SE, Stackpole SA, et al. Effects of IL-10 on systemic inflammatory responses during sublethal primate endotoxemia. *J Immunol*. 1997;158(4):1971–5.
15. Remick DG, Garg SJ, Newcomb DE, Wollenberg G, Huie TK, Bolgos GL. Exogenous interleukin-10 fails to decrease the mortality or morbidity of sepsis. *Crit Care Med*. 1998;26(5):895–904.

16. Steinhäuser ML, Hogaboam CM, Kunkel SL, Lukacs NW, Strieter RM, Standiford TJ. IL-10 is a major mediator of sepsis-induced impairment in lung antibacterial host defense. *J Immunol.* 1999;162(1):392–9.
17. Ashare A, Powers LS, Butler NS, Doerschug KC, Monick MM, Hunninghake GW. Anti-inflammatory response is associated with mortality and severity of infection in sepsis. *Am J Physiol Lung Cell Mol Physiol.* 2005;288(4):L633–40.
18. Song GY, Chung CS, Chaudry IH, Ayala A. What is the role of interleukin 10 in polymicrobial sepsis: anti-inflammatory agent or immunosuppressant? *Surgery.* 1999;126(2):378–83.
19. Keane RM, Birmingham W, Shatney CM, Winchurch RA, Munster AM. Prediction of sepsis in the multitraumatic patient by assays of lymphocyte responsiveness. *Surg Gynecol Obstet.* 1983;156(2):163–7.
20. Green DR, Beere HM. Apoptosis. Gone but not forgotten. *Nature.* 2000;405(6782):28–9.
21. Hotchkiss RS, Swanson PE, Freeman BD, Tinsley KW, Cobb JP, Matuschak GM, et al. Apoptotic cell death in patients with sepsis, shock, and multiple organ dysfunction. *Crit Care Med.* 1999;27(7):1230–51.
22. Hotchkiss RS, Schmiegl Jr RE, Swanson PE, Freeman BD, Tinsley KW, Cobb JP, et al. Rapid onset of intestinal epithelial and lymphocyte apoptotic cell death in patients with trauma and shock. *Crit Care Med.* 2000;28(9):3207–17.
23. Hotchkiss RS, Tinsley KW, Swanson PE, Schmiegl Jr RE, Hui JJ, Chang KC, et al. Sepsis-induced apoptosis causes progressive profound depletion of B and CD4+ T lymphocytes in humans. *J Immunol.* 2001;166(11):6952–63.
24. Tinsley KW, Grayson MH, Swanson PE, Drewry AM, Chang KC, Karl IE, et al. Sepsis induces apoptosis and profound depletion of splenic interdigitating and follicular dendritic cells. *J Immunol.* 2003;171(2):909–14.
25. Kovach MA, Standiford TJ. The function of neutrophils in sepsis. *Curr Opin Infect Dis.* 2012;25(3):321–7.
26. Fialkow L, Fochesatto Filho L, Bozzetti MC, Milani AR, Rodrigues Filho EM, Ladniuk RM, et al. Neutrophil apoptosis: a marker of disease severity in sepsis and sepsis-induced acute respiratory distress syndrome. *Crit Care.* 2006;10(6):R155.
27. Tamayo E, Gomez E, Bustamante J, Gómez-Herrerías JI, Fonteriz R, Bobillo F, et al. Evolution of neutrophil apoptosis in septic shock survivors and nonsurvivors. *J Crit Care.* 2012;27(4):415.e1–11.
28. Drifte G, Dunn-Siegrist I, Tissières P, Pugin J. Innate immune functions of immature neutrophils in patients with sepsis and severe systemic inflammatory response syndrome. *Crit Care Med.* 2013;41(3):820–32.
29. Kasten KR, Muenzer JT, Caldwell CC. Neutrophils are significant producers of IL-10 during sepsis. *Biochem Biophys Res Commun.* 2010;393(1):28–31.
30. Stephan F, Yang K, Tankovic J, Soussy CJ, Dhonneur G, Duvaldestin P, et al. Impairment of polymorphonuclear neutrophil functions precedes nosocomial infections in critically ill patients. *Crit Care Med.* 2002;30(2):315–22.
31. Hotchkiss RS, Tinsley KW, Swanson PE, Grayson MH, Osborne DF, Wagner TH, et al. Depletion of dendritic cells, but not macrophages, in patients with sepsis. *J Immunol.* 2002;168(5):2493–500.
32. Fujita S, Seino K, Sato K, Sato Y, Eizumi K, Yamashita N, et al. Regulatory dendritic cells act as regulators of acute lethal systemic inflammatory response. *Blood.* 2006;107(9):3656–64.
33. Poehlmann H, Schefold JC, Zuckermann-Becker H, Volk HD, Meisel C. Phenotype changes and impaired function of dendritic cell subsets in patients with sepsis: a prospective observational analysis. *Crit Care.* 2009;13(4):R119.
34. Pastille E, Didovic S, Brauckmann D, Rani M, Agrawal H, Schade FU, et al. Modulation of dendritic cell differentiation in the bone marrow mediates sustained immunosuppression after polymicrobial sepsis. *J Immunol.* 2011;186(2):977–86.
35. Guisset O, Dilhuydy MS, Thiebaut R, Lefèvre J, Camou F, Sarrat A, et al. Decrease in circulating dendritic cells predicts fatal outcome in septic shock. *Intensive Care Med.* 2007;33(1):148–52.

36. Toliver-Kinsky TE, Cui W, Murphey ED, Lin C, Sherwood ER. Enhancement of dendritic cell production by fms-like tyrosine kinase-3 ligand increases the resistance of mice to a burn wound infection. *J Immunol.* 2005;174(1):404–10.
37. Bohannon J, Fang G, Cui W, Sherwood E, Toliver-Kinsky T. Fms-like tyrosine kinase-3 ligand alters antigen-specific responses to infections after severe burn injury. *Shock.* 2009;32(4):435–41.
38. Cavaillon JM, Adib-Conquy M. Bench-to-bedside review: endotoxin tolerance as a model of leukocyte reprogramming in sepsis. *Crit Care.* 2006;10(5):233.
39. Hotchkiss RS, Monneret G, Payen D. Sepsis-induced immunosuppression: from cellular dysfunctions to immunotherapy. *Nat Rev Immunol.* 2013;13(12):862–74.
40. Munoz C, Carlet J, Fitting C, Misset B, Bleriot JP, Cavaillon JM. Dysregulation of in vitro cytokine production by monocytes during sepsis. *J Clin Invest.* 1991;88(5):1747–54.
41. Monneret G, Finck ME, Venet F, Debard AL, Bohé J, Bienvenu J, et al. The anti-inflammatory response dominates after septic shock: association of low monocyte HLA-DR expression and high interleukin-10 concentration. *Immunol Lett.* 2004;95(2):193–8.
42. Venet F, Davin F, Guignant C, Larue A, Cazalis MA, Darbon R, et al. Early assessment of leukocyte alterations at diagnosis of septic shock. *Shock.* 2010;34(4):358–63.
43. Chiche L, Forel JM, Thomas G, Farnarier C, Vely F, Bléry M, et al. The role of natural killer cells in sepsis. *J Biomed Biotechnol.* 2011;2011:986491.
44. Forel JM, Chiche L, Thomas G, Mancini J, Farnarier C, Cognet C, et al. Phenotype and functions of natural killer cells in critically-ill septic patients. *PLoS One.* 2012;7(12):e50446.
45. Giamarellos-Bourboulis EJ, Tsaganos T, Spyridaki E, Mouktaroudi M, Plachouras D, Vaki I, et al. Early changes of CD4-positive lymphocytes and NK cells in patients with severe Gram-negative sepsis. *Crit Care.* 2006;10(6):R166.
46. Souza-Fonseca-Guimaraes F, Parlato M, Fitting C, Cavaillon JM, Adib-Conquy M. NK cell tolerance to TLR agonists mediated by regulatory T cells after polymicrobial sepsis. *J Immunol.* 2012;188(12):5850–8.
47. Souza-Fonseca-Guimaraes F, Parlato M, Philippart F, Misset B, Cavaillon JM, Adib-Conquy M, et al. Toll-like receptors expression and interferon-gamma production by NK cells in human sepsis. *Crit Care.* 2012;16(5):R206.
48. Chiche L, Forel JM, Thomas G, Farnarier C, Cognet C, Guervilly C, et al. Interferon-gamma production by natural killer cells and cytomegalovirus in critically ill patients. *Crit Care Med.* 2012;40(12):3162–9.
49. Monneret G, Venet F, Kullberg BJ, Netea MG. ICU-acquired immunosuppression and the risk for secondary fungal infections. *Med Mycol.* 2011;49(Suppl 1):S17–23.
50. Venet F, Bohe J, Debard AL, Bienvenu J, Lepape A, Monneret G. Both percentage of gammadelta T lymphocytes and CD3 expression are reduced during septic shock. *Crit Care Med.* 2005;33(12):2836–40.
51. Andreu-Ballester JC, Tormo-Calandin C, Garcia-Ballesteros C, Pérez-Griera J, Amigó V, Almela-Quilis A, et al. Association of gammadelta T cells with disease severity and mortality in septic patients. *Clin Vaccine Immunol.* 2013;20(5):738–46.
52. Monneret G, Debard AL, Venet F, Bohe J, Hequet O, Bienvenu J, et al. Marked elevation of human circulating CD4+CD25+ regulatory T cells in sepsis-induced immunoparalysis. *Crit Care Med.* 2003;31(7):2068–71.
53. Venet F, Chung CS, Kherouf H, Geeraert A, Malcus C, Poitevin F, et al. Increased circulating regulatory T cells (CD4(+)CD25(+)CD127(-)) contribute to lymphocyte anergy in septic shock patients. *Intensive Care Med.* 2009;35(4):678–86.
54. Tiemessen MM, Jagger AL, Evans HG, van Herwijnen MJ, John S, Taams LS. CD4+CD25+Foxp3+ regulatory T cells induce alternative activation of human monocytes/macrophages. *Proc Natl Acad Sci U S A.* 2007;104(49):19446–51.
55. Li L, Wu CY. CD4+ CD25+ Treg cells inhibit human memory gammadelta T cells to produce IFN-gamma in response to M tuberculosis antigen ESAT-6. *Blood.* 2008;111(12):5629–36.
56. Gogos CA, Drosou E, Bassaris HP, Skoutelis A. Pro-versus anti-inflammatory cytokine profile in patients with severe sepsis: a marker for prognosis and future therapeutic options. *J Infect Dis.* 2000;181(1):176–80.

57. Asadullah K, Woiciechowsky C, Docke WD, Egerer K, Kox WJ, Vogel S, et al. Very low monocyte HLA-DR expression indicates high risk of infection—immunomonitoring for patients after neurosurgery and patients during high dose steroid therapy. *Eur J Emerg Med.* 1995;2(4):184–90.
58. van den Berk JM, Oldenburger RH, van den Berg AP, Klompmaker IJ, Mesander G, van Son WJ, et al. Low HLA-DR expression on monocytes as a prognostic marker for bacterial sepsis after liver transplantation. *Transplantation.* 1997;63(12):1846–8.
59. Denzel C, Riese J, Hohenberger W, Born G, Köckerling F, Tschalkowsky K, et al. Monitoring of immunotherapy by measuring monocyte HLA-DR expression and stimulated TNF α production during sepsis after liver transplantation. *Intensive Care Med.* 1998;24(12):1343–4.
60. Haveman JW, van den Berg AP, van den Berk JM, Mesander G, Slooff MJ, de Leij LH, et al. Low HLA-DR expression on peripheral blood monocytes predicts bacterial sepsis after liver transplantation: relation with prednisolone intake. *Transpl Infect Dis.* 1999;1(3):146–52.
61. Hynninen M, Pettila V, Takkunen O, Orko R, Jansson SE, Kuusela P, et al. Predictive value of monocyte histocompatibility leukocyte antigen-DR expression and plasma interleukin-4 and -10 levels in critically ill patients with sepsis. *Shock.* 2003;20(1):1–4.
62. Oczenski W, Krenn H, Jilch R, Watzka H, Waldenberger F, Köller U, et al. HLA-DR as a marker for increased risk for systemic inflammation and septic complications after cardiac surgery. *Intensive Care Med.* 2003;29(8):1253–7.
63. Perry SE, Mostafa SM, Wenstone R, Shenkin A, McLaughlin PJ. Is low monocyte HLA-DR expression helpful to predict outcome in severe sepsis? *Intensive Care Med.* 2003;29(8):1245–52.
64. Ahlstrom A, Hynninen M, Tallgren M, Kuusela P, Valtonen M, Orko R, et al. Predictive value of interleukins 6, 8 and 10, and low HLA-DR expression in acute renal failure. *Clin Nephrol.* 2004;61(2):103–10.
65. Su L, Zhou DY, Tang YQ, Wen Q, Bai T, Meng FS, et al. Clinical value of monitoring CD14+ monocyte human leukocyte antigen (locus) DR levels in the early stage of sepsis. *Zhongguo Wei Zhong Bing Ji Jiu Yi Xue.* 2006;18(11):677–9.
66. Zhang YT, Fang Q. Study on monocyte HLA-DR expression in critically ill patients after surgery. *Zhonghua Wai Ke Za Zhi.* 2006;44(21):1480–2.
67. Allen ML, Peters MJ, Goldman A, Elliott M, James I, Callard R, et al. Early postoperative monocyte deactivation predicts systemic inflammation and prolonged stay in pediatric cardiac intensive care. *Crit Care Med.* 2002;30(5):1140–5.
68. Doughty L, Carcillo JA, Kaplan S, Janosky J. The compensatory anti-inflammatory cytokine interleukin 10 response in pediatric sepsis-induced multiple organ failure. *Chest.* 1998;113(6):1625–31.
69. Simmons EM, Himmelfarb J, Sezer MT, Chertow GM, Mehta RL, Paganini EP, et al. Plasma cytokine levels predict mortality in patients with acute renal failure. *Kidney Int.* 2004;65(4):1357–65.
70. van Dissel JT, van Langevelde P, Westendorp RG, Kwappenberg K, Frolich M. Anti-inflammatory cytokine profile and mortality in febrile patients. *Lancet.* 1998;351(9107):950–3.
71. Boomer JS, To K, Chang KC, Takasu O, Osborne DF, Walton AH, et al. Immunosuppression in patients who die of sepsis and multiple organ failure. *JAMA.* 2011;306(23):2594–605.
72. Angele MK, Wichmann MW, Ayala A, Cioffi WG, Chaudry IH. Testosterone receptor blockade after hemorrhage in males. Restoration of the depressed immune functions and improved survival following subsequent sepsis. *Arch Surg.* 1997;132(11):1207–14.
73. Angele MK, Catania RA, Ayala A, Cioffi WG, Bland KI, Chaudry IH. Dehydroepiandrosterone: an inexpensive steroid hormone that decreases the mortality due to sepsis following trauma-induced hemorrhage. *Arch Surg.* 1998;133(12):1281–8.
74. Catania RA, Angele MK, Ayala A, Cioffi WG, Bland KI, Chaudry IH. Dehydroepiandrosterone restores immune function following trauma-haemorrhage by a direct effect on T lymphocytes. *Cytokine.* 1999;11(6):443–50.
75. Roifman CM, Zhang J, Chitayat D, Sharfe N. A partial deficiency of interleukin-7R α is sufficient to abrogate T-cell development and cause severe combined immunodeficiency. *Blood.* 2000;96(8):2803–7.

76. Puel A, Leonard WJ. Mutations in the gene for the IL-7 receptor result in T(-)B(+)NK(+) severe combined immunodeficiency disease. *Curr Opin Immunol.* 2000;12(4):468–73.
77. Puel A, Ziegler SF, Buckley RH, Leonard WJ. Defective IL7R expression in T(-)B(+)NK(+) severe combined immunodeficiency. *Nat Genet.* 1998;20(4):394–7.
78. Hutchins NA, Unsinger J, Hotchkiss RS, Ayala A. The new normal: immunomodulatory agents against sepsis immune suppression. *Trends Mol Med.* 2014;20(4):224–33.
79. Unsinger J, McGlynn M, Kasten KR, Hoekzema AS, Watanabe E, Muenzer JT, et al. IL-7 promotes T cell viability, trafficking, and functionality and improves survival in sepsis. *J Immunol.* 2010;184(7):3768–79.
80. Levy Y, Lacabaratz C, Weiss L, Viard JP, Goujard C, Lelièvre JD, et al. Enhanced T cell recovery in HIV-1-infected adults through IL-7 treatment. *J Clin Invest.* 2009;119(4):997–1007.
81. Ochoa MC, Mazzolini G, Hervas-Stubbs S, de Sanmamed MF, Berraondo P, Melero I. Interleukin-15 in gene therapy of cancer. *Curr Gene Ther.* 2013;13(1):15–30.
82. Inoue S, Unsinger J, Davis CG, Muenzer JT, Ferguson TA, Chang K, et al. IL-15 prevents apoptosis, reverses innate and adaptive immune dysfunction, and improves survival in sepsis. *J Immunol.* 2010;184(3):1401–9.
83. Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, DF MD, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med.* 2012;366(26):2443–54.
84. Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Engl J Med.* 2012;366(26):2455–65.
85. Zhang Y, Li J, Lou J, Zhou Y, Bo L, Zhu J, et al. Upregulation of programmed death-1 on T cells and programmed death ligand-1 on monocytes in septic shock patients. *Crit Care.* 2011;15(1):R70.
86. Guignant C, Lepape A, Huang X, Kherouf H, Denis L, Poitevin F, et al. Programmed death-1 levels correlate with increased mortality, nosocomial infection and immune dysfunctions in septic shock patients. *Crit Care.* 2011;15(2):R99.
87. Chang KC, Burnham CA, Compton SM, Rasche DP, Mazuski RJ, JS MD, et al. Blockade of the negative co-stimulatory molecules PD-1 and CTLA-4 improves survival in primary and secondary fungal sepsis. *Crit Care.* 2013;17(3):R85.
88. Zhang Y, Zhou Y, Lou J, Li J, Bo L, Zhu K, et al. PD-L1 blockade improves survival in experimental sepsis by inhibiting lymphocyte apoptosis and reversing monocyte dysfunction. *Crit Care.* 2010;14(6):R220.
89. Root RK, Lodato RF, Patrick W, Cade JF, Fotheringham N, Milwee S, et al. Multicenter, double-blind, placebo-controlled study of the use of filgrastim in patients hospitalized with pneumonia and severe sepsis. *Crit Care Med.* 2003;31(2):367–73.
90. Nelson S, Belknap SM, Carlson RW, Dale D, De Boisblanc B, Farkas S, et al. A randomized controlled trial of filgrastim as an adjunct to antibiotics for treatment of hospitalized patients with community-acquired pneumonia. CAP Study Group. *J Infect Dis.* 1998;178(4):1075–80.
91. Meisel C, Schefold JC, Pischowski R, Baumann T, Hetzger K, Gregor J, et al. Granulocyte-macrophage colony-stimulating factor to reverse sepsis-associated immunosuppression: a double-blind, randomized, placebo-controlled multicenter trial. *Am J Respir Crit Care Med.* 2009;180(7):640–8.
92. Hall MW, Knatz NL, Vetterly C, Tomarello S, Wewers MD, Volk HD, et al. Immunoparalysis and nosocomial infection in children with multiple organ dysfunction syndrome. *Intensive Care Med.* 2011;37(3):525–32.
93. Docke WD, Randow F, Syrbe U, Krausch D, Asadullah K, Reinke P, et al. Monocyte deactivation in septic patients: restoration by IFN-gamma treatment. *Nat Med.* 1997;3(6):678–81.
94. Nalos M, Santner-Nanan B, Parnell G, Tang B, McLean AS, Nanan R. Immune effects of interferon gamma in persistent staphylococcal sepsis. *Am J Respir Crit Care Med.* 2012;185(1):110–2.
95. Hershman MJ, Appel SH, Wellhausen SR, Sonnenfeld G, Polk Jr HC. Interferon-gamma treatment increases HLA-DR expression on monocytes in severely injured patients. *Clin Exp Immunol.* 1989;77(1):67–70.

96. Bundschuh DS, Barsig J, Hartung T, Randow F, Döcke WD, Volk HD, et al. Granulocyte-macrophage colony-stimulating factor and IFN-gamma restore the systemic TNF-alpha response to endotoxin in lipopolysaccharide-desensitized mice. *J Immunol.* 1997;158(6):2862–71.
97. Kox WJ, Bone RC, Krausch D, Döcke WD, Kox SN, Wauer H, et al. Interferon gamma-1b in the treatment of compensatory anti-inflammatory response syndrome. A new approach: proof of principle. *Arch Intern Med.* 1997;157(4):389–93.
98. Nakos G, Malamou-Mitsi VD, Lachana A, Karassavoglou A, Kitsiouli E, Agnandi N, et al. Immunoparalysis in patients with severe trauma and the effect of inhaled interferon-gamma. *Crit Care Med.* 2002;30(7):1488–94.