

Glucocorticoids: Inflammation and Immunity

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Abstract Glucocorticoids are universally prescribed as the drug of choice for the treatment of inflammatory and autoimmune disorders. These stress hormones act through their cognate glucocorticoid receptor to regulate transcription of various target genes. The mechanisms of glucocorticoid action are often cell type dependent and involve the regulation of thousands of genes. Glucocorticoids have a tremendous impact on the immune system during inflammation including effects on the plasticity, survival, and function of immune cells. This chapter highlights the dynamic effects of glucocorticoids in regards to both physiological and pathological conditions during inflammation. We address issues involving classical and alternative mechanisms of glucocorticoid inhibition, the effect on innate and adaptive immunity, glucocorticoid tissue-specific actions, and their role in target immune cells.

Keywords Glucocorticoid • Steroids • Stress hormones • Inflammation • Gene expression • Transrepression • Transactivation • Innate immunity • Adaptive immunity • Resistance

Introduction

Glucocorticoids are primary stress hormones that function throughout the body to regulate a diverse array of physiological systems. Glucocorticoids (GCs) derived their name from early observations of their effect in regulating glucose metabolism [1, 2]. Currently, the actions of this class of steroids extend beyond the mobilization of amino acids and gluconeogenesis and are known to play important roles in the control/regulation of various biological processes. In fact, glucocorticoids are required for life, as the absence of these stress hormones results in death prior to or at the time of birth. Glucocorticoids influence a number of physiological systems including the

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immune system [3] where in addition to exerting both pro- and anti-inflammatory actions, this stress hormone has a potent role in development and homeostasis of T lymphocytes [4]. Additionally, these stress hormones impact development [5], where glucocorticoids are known to play a key role in the maturation of the fetal lung [6]. Furthermore, glucocorticoids have a role in the brain where they have been shown to regulate arousal and cognitive functions controlled by the hippocampus, amygdala, and the frontal lobes of the brain [7]. The pleiotropic actions of glucocorticoids occur through binding to its cognate receptor, the glucocorticoid receptor (GR), which is expressed in nearly every cell in the body. Glucocorticoids act through its receptor to regulate transcription of various target genes, however as will be discussed later, non-genomic effects have also been described.

Human GR protein is encoded by the *NR3C1* gene located in chromosome 5 (5q31) and is a member of the nuclear receptor superfamily of ligand-dependent transcription factors [8]. Like other steroid receptors, GR is modular in structure containing an N-terminal regulatory domain (NTD), a central DNA-binding domain (DBD), a hinge region, and a C-terminal ligand-binding domain (LBD) [9]. In the absence of hormone, GR resides in the cytoplasm in a complex with other proteins including heat shock protein 90, heat shock protein 70, and FKBP52, the latter being an immunophilin molecule involved in protein folding and trafficking. Upon ligand binding, GR is released from its cytoplasmic complex and translocates into the nucleus where it interacts with specific targeting sequences termed glucocorticoid-response elements (GREs) to regulate thousands of genes. The nature of the GR-occupied GRE results in either induction or repression of target gene expression. Additionally, GR can undergo a conformational change upon binding to the GRE that leads to the recruitment of cofactors and/or coregulators to modulate, and thereby alter the transcriptional rates of target genes [10]. Along with the nature of the GRE and the recruitment of cofactors and/or coregulators, several other factors can also influence GRs ability to regulate gene transcription including chromatin structure, epigenetic regulators, and its physical interaction with other transcription factors such as nuclear factor kappa B (NF- κ B) and activator protein 1 (AP-1).

GR, while derived from a single gene, has multiple functionally distinct isoforms due to alternative splicing and translational initiation mechanisms [11]. Alternative splicing accounts for 2 discrete receptor isoforms (GR α and GR β) that differ at their carboxyl termini, while alternative translation initiation results in 8 additional receptor subtypes, each with a progressively shorter NTD. GR α has been the primary and most extensively studied glucocorticoid receptor, as the GR β splice variant does not bind GCs [12]. However, expression of GR β has been associated with glucocorticoid resistance and tissue specificity, as GR β has been shown to antagonize the activity of GR α [13, 14]. In contrast to GR β , the 8 unique translational isoforms of GR α have similar binding affinities for glucocorticoids and can interact with GREs. Similar to GR β , the expression of these various translational GR α isoforms varies widely across tissues. Thus, the existence of numerous GR isoforms is thought to be a major factor contributing to the diverse array of tissue-specific actions of glucocorticoids in the body.

Glucocorticoids are also known to have potent immunosuppressive and anti-inflammatory actions, thus being vital in the treatment of autoimmune and inflammatory diseases and are one of the most widely prescribed drugs in the world. In this review, we will focus on the how glucocorticoids modulate and interact with the immune system, along with its effect on combating inflammation. Additionally, we will discuss how glucocorticoids affect the response and behavior of different immune cells in the management of inflammatory diseases.

Inflammation as a Natural Host Defense Mechanism and Glucocorticoid Regulation

Inflammation is an innate defensive mechanism that protects us from damaging stimuli such as pathogens and harmful irritants. Inflammation is a complex biological process that initially involves increased blood flow and movement of immune cells, especially granulocytes and macrophages along with other molecular mediators, from the blood to the site of injury. This acute response sets the stage for the healing process by combating the initial source of inflammation through the removal of necrotic cells and damaged tissue in a coordinated response involving both the immune and vascular systems. As the inflammation process continues, a shift in the type of cells present at the site of injury results in the repair or healing of the tissue.

Classic signs of inflammation include pain, redness, swelling, warmth, and loss of function or immobility at the site of damage. Additionally, inflammation may result in more global symptoms such as fever, chills, fatigue, and general stiffness. Pain expressly plays an important role in the ability of glucocorticoids to regulate inflammation. Cytokines and inflammatory mediators released into the blood from the damage site activate peripheral pain receptors. Pain signals sent to the brain result in the activation of the hypothalamic–pituitary–adrenal (HPA) axis which, in turn, induces glucocorticoid secretion. GCs inhibit the synthesis of cytokines and inflammatory mediators to counter the extent of the inflammation. Despite the initial unfavorable effects of this condition, inflammation is extremely important and beneficial to human health as infections and wounds, or any damage tissue, would not heal without this homeostatic response.

As with all physiological responses in the body, inflammation needs to be regulated especially in conjunction with other host defense systems. Too little inflammation can result in progressive and detrimental tissue destruction, while excessive inflammation can lead to a host of diseases including allergies, autoimmune disorders, chronic inflammatory diseases, and even cancer. Glucocorticoids regulate and reduce the inflammatory response by entering cells and suppressing the transcription of proteins that promote inflammation. In the absence of glucocorticoids, persistent inflammation can lead to dysregulation of converging pro- and anti-inflammatory factors at the site of injury resulting in abnormalities and pathogenesis [15].

Since glucocorticoids are known to be essential for the regulation of the inflammatory response, they also act to reduce the extent of an overactive immune system. Thus, glucocorticoids are among the most widely prescribed drugs for the treatment of asthma, allergies, and autoimmune diseases. This class of steroid hormones initiates a multitude of diverse signaling pathways that hold inflammation in check and counter a rampant immune system, limiting the excessive damage that can occur to the host cells and surrounding tissue [16]. A major barrier in employing glucocorticoid therapy in the clinic has been our lack of understanding of the molecular mechanisms that resolves inflammation. At one level, the mechanism of glucocorticoid action in counteracting inflammation may appear simplistic as this class of drug (GC) acting on its receptor (GR) modulates gene transcription to inhibit the extent of inflammation. However, the heterogeneity of glucocorticoid receptor isoforms and the cell-type specific biological responses suggest that GR's ability to prevent inflammation is not a simple endeavor but a complex series of events. Thus, there are many ways glucocorticoids exert their anti-inflammatory effects.

Classical Mechanisms of Glucocorticoid Inhibition of Inflammation

Glucocorticoids utilize a variety of processes simultaneously to control inflammation; from the activation of anti-inflammatory genes, to suppressing proinflammatory cytokines and chemokines, to moderating key proinflammatory regulators such as NF- κ B and AP-1. Several fundamental mechanisms have been elucidated for these actions of GR. First, direct binding of GR to GREs in DNA can enhance the transcription of anti-inflammatory genes (transactivation). Glucocorticoid-induced transactivation of genes such as IL-10, IL-1 receptor antagonist, and mitogen-activated protein kinase phosphatase-1 (MKP-1) is known to increase gene expression and thus protein expression of these anti-inflammatory molecules [17, 18]. Second, direct binding of GR to negative GREs (nGRE) in DNA can suppress transcription (transrepression) of various proinflammatory cytokines and modulators such as iNOS, COX-2, IL-1 β , and TNF [17, 18]. Finally, GR binding directly to transcription factors like the p65 subunit of NF- κ B or AP-1 can prevent downstream transcription of proinflammatory mediators to control the extent of inflammation [19]. This latter mechanism of transcriptional repression, known as tethering, where GR does not directly bind DNA response sequences, has been shown to be key in GRs ability to regulate inflammation [20–22]. Additionally, cross-talk between GR and other transcription factors can occur through the binding to composite or overlapping response elements [23]. Interestingly, while the repression of NF- κ B by GR has long been considered a crucial determinant in reducing the expression of specific proinflammatory targets, a recent study by Altonsy et al. showed the cooperative association between the GR and NF- κ B enhanced the expression of TNFAIP3, a potent anti-inflammatory gene and inhibitor of NF- κ B, suggesting a greater complexity in the cross-talk of these two molecules [24].

GR binding directly to either GREs or nGREs that initiate or suppress gene expression of anti- or pro-inflammatory genes, respectively, is not the only mechanism to consider in GRs ability to control inflammation. While it has been suggested that the number of GREs present play a role in the activation or suppression of gene transcription, it has become increasingly clear that their proximity to the TATA box also is an important factor [25]. Additionally, the recruitment of various coactivators such as TIF2 and SRC-1, corepressors such as NCoR and SMRT, along with various other comodulators can interact with the GR-DNA complex enabling an additional level of transcriptional regulation in GR's ability to control inflammation [26, 27]. GR may regulate inflammation by reducing mRNA half-life through the GC responsive gene tristetraprolin (TTP) [28, 29]. GR can also be phosphorylated by various kinases that can affect its stability, its DNA binding capacity, its ability to translocate to the nucleus, and its interactions with other transcription factors and/or molecular chaperones [30]. Thus, the simple concept of one drug (GC) working on its receptor (GR) has evolved to comprise numerous multifaceted mechanisms to control and regulate inflammation.

Furthermore, glucocorticoids have been shown to regulate inflammation in the absence of DNA binding or interactions with other transcription factors. Nongenomic GC-GR interactions were shown to account for the cardioprotective effect of an acute high dose of corticosteroids resulting in the nontranscriptional activation of endothelial nitric oxide synthase (eNOS) [31]. eNOS has been shown to play an important role during inflammation in regulating the expression of proinflammatory molecules such as NF- κ B and cyclooxygenase-2 (Cox-2) [32, 33]. Additionally, nongenomic GC-GR mechanisms involving the activation/inhibition of various signaling pathways, including the p42 MAPK and MAPK ERK1/2 pathways, and the activation of proteins with SH3 domains such as Src and Ras that in turn activate the aforementioned kinase pathways, have been shown to occur [34]. Finally, a mechanism of GC anti-inflammatory action still in its infancy involves posttranscriptional gene regulation via RNA-binding proteins and microRNAs [35]. Interestingly, the role of these posttranscriptional gene regulation actions is thought not to function specifically in turning on or off genes, but to act more as a rheostat in controlling the appropriate amplitude and duration of the response [36].

As the modulation of gene transcription is the major consequence of glucocorticoid activity, the changes in gene transcription that occur directly via activation/repression of GC target genes, or through tethering to another transcription factor, are the most well studied means in controlling inflammation. However, GR can also modulate gene transcription indirectly through the consequences of the activation of the initial target gene. A classic example of this mode of GR regulation involves the glucocorticoid-induced leucine zipper (GILZ) gene. GILZ encodes for a potent anti-inflammatory protein with immunosuppressive and cell survival-promoting effects. GILZ was initially identified as a molecule that protected lymphocytes from TCR/CD3-activated cell death [37]. However, subsequently it was shown that GILZ itself did not directly protect lymphocytes from death, but inhibited the ability of the T cell receptor to induce interleukin-2/interleukin-2 receptor expression and NF- κ B activity [38]. Specifically, it was shown that GILZ inhibited NF- κ B nuclear translocation

and DNA binding via direct protein-to-protein interaction of GILZ with NF- κ B. Since these initial observations, GILZ has been shown to be a multifunctional protein that can inhibit key immune cell signaling pathways. Recently, GILZ was shown to regulate Th17 responses and to restrain IL-17-mediated skin inflammation [39]. While the anti-inflammatory and immune-modulatory effects of GILZ have been widely described [40, 41], the induction of this protein has also been associated with provoking apoptosis. GILZ expression in human neutrophils promoted apoptosis through the down-regulation of the myeloid leukemia cell differentiation protein Mcl-1, an antiapoptotic protein of the Bcl-2 family [42].

Alternative Mechanisms of Glucocorticoid Inhibition of Inflammation

The ability of glucocorticoids to regulate and control inflammation goes beyond simply regulating gene transcription of pro- and anti-inflammatory genes. Recently, a unique mechanism of action for anti-inflammatory effects of GCs was reported during the early phase of acute lung injury (ALI) [43]. These authors showed that glucocorticoids attenuate inflammation associated with ALI via up-regulation of the *SphK1* gene in macrophages. The up-regulation of sphingosine kinase 1 in the lung resulted in the synthesis of sphingosine 1 (S1P) that in turn binds to the S1P receptor type 1 (S1PR1) and triggers the Rho family-dependent reorganization of the cytoskeleton leading to enhanced barrier function of the endothelium. The protection afforded by glucocorticoids to enhance the barrier function through this mechanism prevents vascular leakage and the massive infiltration of immune cells into the lung as a way of controlling inflammation.

Interestingly, another recent study linked the action of glucocorticoids to the circadian clock to control time-of-day variations and magnitude of pulmonary inflammation [44]. In this study, the authors observed that pulmonary antibacterial responses of neutrophil recruitment via the chemokine and glucocorticoid responsive gene CXCL5 were modulated by a circadian clock mechanism within epithelial club (Clara) cells [45]. Intriguingly, adrenalectomy blocked this circadian neutrophil recruitment and rhythmic inflammatory responses afforded by CXCL5 upon intraperitoneal injection of LPS. Therefore, this study suggests that the therapeutic effects of glucocorticoids can depend on the local circadian circuit regulation of GR function.

Glucocorticoids have also been shown to suppress overactive inflammatory responses by induction of negative feedback regulators such as the interleukin-1 receptor-associated kinase M (IRAK-M; also known as IRAK3). IRAK-M is known to be a critical negative feedback regulator of Toll-like receptor/Interleukin (IL)-1 receptor (TLR/IL-1R) superfamily of signaling molecules that trigger increased expression of multiple inflammatory genes [46]. Miyata et al. have shown that glucocorticoids suppress bacteria-induced inflammation by directly binding to and up-regulating IRAK-M in airway macrophages and epithelial cells [47]. Additionally, these authors show that IRAK-M

depletion results in the enhanced expression of proinflammatory mediators. Thus, glucocorticoids can suppress an overactive inflammatory response via negative feedback to tightly control the inflammatory response and maintain homeostasis.

Overview of Glucocorticoids and the Immune System

The first medical use of glucocorticoids some 60 years ago was for the treatment of rheumatoid arthritis [48, 49]. Since then, glucocorticoids have remained the most commonly used anti-inflammatory and immunomodulatory agents. Their therapeutic activity is substantial in a wide spectrum of diseases, including acute and chronic inflammations, autoimmune disorders, organ transplantations, and the treatment of hematologic cancers [50]. Over the years, numerous publications have focused on glucocorticoids effects on the immune system, and much has been discovered about the molecular mechanism by which GCs act. As in other cells, GR is able to regulate gene expression both positively and negatively in immune cells [9, 18] and can control the inflammatory processes either by a direct binding with glucocorticoid-responsive sequences expressed in the promoters of target genes, or by binding other crucial transcription factors, thus inhibiting the propagation of proinflammatory signals [51]. Furthermore, recent studies have described discrepancies between the immunosuppressive and immunostimulatory effects of glucocorticoids [52, 53].

As described earlier, the inflammatory response is the first protective host response elicited by an injury prompting mobilization of the immune system. The inflammatory recruitment of immune cells neutralizes injurious stimuli and restores the function and structure of damaged tissues [54]. The initial manifestation is the release of intracellular contents after cellular necrosis within the inflammatory site that induces the activation of innate immune components. Through invariant pattern-recognition receptors (PRRs), innate immunity is promptly activated upon detection of conserved structures known as pathogen-associated molecular patterns (PAMPs) or damage-associated molecular patterns (DAMPs) [55]. Mast cells and resident macrophages exert different effector functions, one of which is the increased production of proinflammatory molecules such as interleukin-1 and TNF- α , free radicals, histamine, nitric oxide, prostaglandins, and leukotrienes. This increase in proinflammatory molecules results in vasodilatation, capillary permeability, growth of new blood vessels, and leukocyte migration into the inflamed region. The ability of these cells to generate a chemotactic gradient to recruit cells into the injured tissue is rapid; thus, granulocytes and monocyte migrate from blood into tissue within minutes of injury. Among granulocytes, neutrophils are the most important cells at this first stage because of their capacity to destroy invading microorganisms through phagocytosis and microbicidal activity. Pathogenic antigens are engulfed by resident dendritic cells that rapidly differentiate, migrate to lymph nodes, and present antigens to T and B lymphocytes, thus priming and propagating the adaptive immune components including cell-mediated immunity, cytokine production, antigen-specific antibody production, and immunological memory [56].

The resolution of acute inflammation is a dynamic, limited, and finely regulated process that depends upon the crosstalk between innate and adaptive compartments that restore homeostasis after the elimination of harmful agent. An excessive immune response that continues to counteract the persistence of the injurious stimulus triggers a domino effect that leads to chronic inflammation. For this purpose, in the management of numerous mechanisms that control the development and maintenance of inflammation and autoimmune diseases, glucocorticoids have been the most potent drugs of choice, affecting nearly every cell of the immune system, depending on their state of differentiation or activation [57, 58].

Glucocorticoid Effects on Innate Immunity

The innate immune system is the first line of defense that acts rapidly after encountering noxious agents without the reliance on antibody or other acquired responses. For this reason, the effects of glucocorticoids on innate immune cells must be immediate (in terms of minutes) thus contributing to the resolution of inflammation. Among innate immune cells, glucocorticoids strongly influence the plasticity, survival, and function of monocytes and macrophages according to the plasma GC concentration and the state of cell activation [59]. To enhance the clearance of pathogens, dead cells, and toxins, low GC concentrations enhance antigen uptake, scavenger function, and phagocytosis. For this purpose, the induction of the opsonins MFG-E8, Merck and protein S [60, 61], the up-regulation of mannose receptor MR/CD206 [62], the scavenger receptor CD163 [63], and the increase of IFN- γ -induced FcR [64] have been observed. Glucocorticoids target macrophages to ensure survival in response to LPS-induced sepsis and to suppress inflammation associated with contact allergy [65, 66]. These results suggest that low concentrations of glucocorticoids have an immune-stimulatory effect on macrophage function in the presence of inflammatory stimuli, whereas high concentrations of this stress hormone exert inhibitory functions on macrophages. High dose actions abrogate the production of proinflammatory mediators as numerous cytokines are down-regulated, the secretion of many chemokines is inhibited, the expression of adhesion molecules such as beta-2 integrin is reduced, and antigen presentation and expression of HLA molecules are decreased by GCs [53, 59, 67].

Consistent with the immunomodulatory properties, some studies have shown that steroid hormones induce highly phagocytic monocyte-derived macrophages. Glucocorticoid exposure functions to reprogram monocyte differentiation through changes in intracellular components that regulate cytoskeletal reorganization following adhesion. The enhanced phagocytic activity and increased expression of the anti-inflammatory cytokine IL-10 observed in these cells support the hypothesis that glucocorticoids do not cause a global suppression of macrophages effectors, but result in the differentiation of a specific anti-inflammatory phenotype which seems to be actively involved in the resolution of inflammatory conditions [68–70].

Furthermore, glucocorticoids exert many of their anti-inflammatory effects through the regulation of granulocyte trafficking. GCs can induce apoptosis and degranulation of basophils and eosinophils. However, at the same time GCs promote the survival and expansion of neutrophils increasing the release of bone marrow precursors [71–75]. In the presence of glucocorticoids, the flow and movement of granulocytes appear tightly regulated to that of monocytes. In an inflammatory scenario, endothelial cells increase the expression of adhesion molecules that bind to their cognate receptor on granulocytes thus allowing cellular transmigration into inflammatory sites. To reduce cellular infiltration, GCs promote shedding of L-selectin and E-selectin from neutrophils [76], suppress the synthesis of many chemokines including IL-8, Mip-1 β , Mip-3 β , Mcp-2, Mcp-3, Mcp-4, RANTES, TARC, and eotaxin, and increase IL-1RII expression, a decoy receptor which limit the deleterious effects of IL-1 [77].

Natural or synthetic glucocorticoids can also alter natural killer (NK) cell activity. Acting as regulatory cells, this homogenous population of innate lymphocytes interacts with various components of the immune system suppressing the immune response [78]. Nevertheless, glucocorticoid treatment is able to reduce NK cell cytolytic activity by the reduction of histone promoter acetylation for perforin and granzyme B. In contrast, glucocorticoids increase histone acetylation in regulatory regions for INF- γ and IL-6. The increase in histone acetylation is associated with increased proinflammatory cytokine mRNA and protein production upon cellular stimulation and epigenetic modifications [79]. These immunologic effects demonstrate how glucocorticoids epigenetically reduce NK cell cytolytic activity, while at the same time prime NK cells for proinflammatory cytokine production that can act as a powerful tool in cancer immunotherapy [80].

Coupling Innate to Adaptive Immunity

While innate immunity provides the first line of defense against pathogens, adaptive immunity, also known as acquired immunity, is also involved during inflammation. Adaptive immunity creates immunological memory after an initial exposure to antigen, resulting in an enhanced antigen-specific response upon subsequent exposure. It is now appreciated that the innate immune response shapes the acquired immune response. The link between the innate and adaptive components involves soluble cytokines and chemokines, and cellular interactions between antigen-specific lymphocytes and antigen-bearing dendritic cells (DCs). In response to a plethora of stimuli, DCs change from immature cells specialized for antigen capture, processing, and presentation, into mature cells that migrate to draining lymph nodes to interact with naive T cell. On this basis, it is evident that innate immune receptors on DCs play a pivotal role in determining the type of adaptive immune response triggered.

Impairment of DC maturation and function is one of the immunosuppressive effects of glucocorticoids [81]. Synthetic GC treatment interferes with the lifecycle

of DCs both in vitro and in vivo [82]. After in vitro maturation with LPS and CD40L, DCs treated with the synthetic glucocorticoid methylprednisolone exhibit enhanced antigen uptake, but down-regulated expression of CD80, CD86, and CD54, and decreased production of TNF- α , IL-6, and IL-12, thus inhibiting the induction of primary T cell responses. Similar results were observed when TNF- α was used to activate DCs [83]. In this study, a different synthetic glucocorticoid dexamethasone, inhibited DC expression of MHC I and II, costimulatory molecules (including B7.1 and B7.2), and the ICAM-1/LFA-1 complex, thus promoting the formation of tolerogenic DCs [82, 84–86]. Tolerogenic DCs are able to drive uncommitted T cells toward the Treg subtype [87] and promote the conversion of CD4⁺ T cells into IL-10 producing type 1 Tregs (Tr1) [88–90]. Moreover, tolerogenic DCs inhibit the proliferation of allospecific T cells [91, 92], preventing acute graft rejection in mice [93], decreasing the number of IFN- γ producing CD4⁺ T cells, and promoting NK cell function toward an alternative activated phenotype unable to secrete IFN- γ [94].

Besides their capacity to modulate and induce effector T cell responses, tolerogenic DCs are defined based on the expression of various surface markers such as Ig-like transcript (ILT) molecules [95], transcriptional regulators like glucocorticoid-induced leucine zipper (GILZ) [40, 96, 97], and enzymes such as retinaldehyde dehydrogenase (RALDH) [98] or NO synthetase-2 (NOS-2) [99], all contributing to their functional properties. Additionally, dendritic cells can also facilitate communication between the immune system and the endocrine system. A recent study described a novel DC population in the pituitary gland that produces cytokines, controls LPS-dependent ACTH secretion, and expresses factors for glucocorticoid release [100]. These data suggest that pituitary DCs relay an immune challenge (such as LPS) to the HPA axis by secreting proinflammatory cytokines, which stimulates the anterior pituitary gland to release ACTH. Therefore, DCs are distinguished not only by their role in linking the innate and adaptive immune responses but also in directing communication between the immune and endocrine systems.

Glucocorticoid Effects on Adaptive Immunity

Autoimmune diseases are associated with the generation of an adaptive immune response mounted against self-antigens. During development, most lymphocytes bearing high affinity receptors for self-antigens are deleted, but not all self-reactive lymphocytes are eliminated. The activity of self-reactive lymphocytes is regulated by peripheral tolerance, an active immunosuppressive process that involves clonal anergy and clonal suppression. A failure in peripheral tolerance allows the activation of self-reactive T or B cell clones, thus eliciting (or inducing) cell-mediated or humoral responses against self-antigen. As potent immunosuppressors, synthetic GCs are extensively used for the treatment of various autoimmune and chronic inflammatory conditions [101]. GCs target several aspects of adaptive immunity, including thymocyte maturation, as well as T and B cell proliferation, survival, and differentiation.

Glucocorticoids and Thymocyte Development

The thymus is the key organ for T cell maturation, and glucocorticoids play an important role in thymocyte selection and survival. The selection process drives immature CD4⁺CD8⁻ “double negative” thymocytes to CD4⁺CD8⁺TCR^{low} “double positive” thymocytes, which represent about 80% of the cells in the thymus. At this stage double positive thymocytes are extremely sensitive to glucocorticoid-induced apoptosis, but escape apoptosis when both TCR and GR signal simultaneously, according to the “mutual antagonism” model [102]. Thymocytes that are unable to process a functional TCR undergo GC-induced apoptosis, since the TCR signal cannot counteract GR signaling, while only thymocytes expressing a TCR with high affinity for self-peptides undergo negative selection due to the inability of GR signaling to overcome the strong TCR-dependent signal. Finally, only those thymocytes exerting a moderate avidity for self-antigens will survive, suggesting interplay between TCR and GC signals. The grade of affinity between TCR and self-peptides is crucial for the survival of a mature T cell repertoire expressing either the CD4 or CD8 receptor.

A number of *in vitro* experiments implicate glucocorticoids in regulating T cell number, survival, and TCR repertoire, although the *in vivo* evidence is still contradictory regarding the correlation between GR expression and thymocyte sensitivity to glucocorticoid-induced death. Adrenalectomy induces thymic hypertrophy [103], mice overexpressing GR in T cells exhibit a reduced number of double positive thymocytes, despite the fact that these cells express lower level of GR compared to thymocytes in other developmental stages. In GILZ-overexpressing transgenic mice, CD4⁺CD8⁺ thymocyte number is significantly decreased and *ex vivo* thymocyte apoptosis is increased [104]. In contrast, intrathymic T cell development and selection proceed normally in mice expressing antisense GR in the thymus and in fetal mice from GR-KO mice [105, 106]. However, these studies suggest the molecular and cellular mechanisms that regulate thymocyte maturation need further investigation.

Moreover, corticosterone synthesis has been suggested to occur in the thymus and there is debate about how locally produced GCs may regulate thymocyte development as well as may affect the initiation of age-associated thymic involution [107].

Glucocorticoid Function in T Cells

Although immature T cells are extremely sensitive to undergoing apoptosis, cell death can also occur in mature T cells either by a glucocorticoid-directed action, or by the involvement of factors that mediate activation-induced cell death, i.e. inhibiting IL-2-mediated activation. According to the activation state and the timing of hormone exposure, T cells can be sensitive or resistant to glucocorticoid-induced cell death. Moreover, mature T cells are susceptible to mutual antagonism between GR

and TCR. Mice lacking GR in T cells (GR^{LckCre}) or carrying a point mutation which inhibits GR dimerization and DNA binding (GR^{dim}) demonstrate that inhibition of activation-induced cell death depends on direct binding of the GR to two nGREs in the CD95 (APO-1/Fas) ligand promoter [108]. In contrast, overexpression of the SWI3-related gene (SRG3) protein in peripheral T cells renders them sensitive to GC-induced apoptosis through the GR–SRG3 complex formation, suggesting that SRG3 may play a critical role in controlling GC-mediated apoptosis of developing thymocytes. Studies have also shown that a dominant negative SRG3 decreases GC sensitivity in thymoma cells. In addition, mice overexpressing the SRG3 protein appear to be more susceptible to stress-induced deletion of peripheral T cells than WT mice, which may result in an immunosuppressive condition [109].

In addition inducing apoptosis, glucocorticoids also affect T cell polarization. Since endogenous or exogenous glucocorticoids attenuate IL-12 synthesis, T cell response is shifted from the Th1 to Th2 phenotype [110]. GCs inhibit both T-bet and GATA-3 transcriptional activity through two different mechanisms. T-bet does not only function as an activator of IFN- γ expression, but also interacts with the GATA-3 transcription factor, inhibiting Th2 cytokine gene expression. Consistent with these results, GCs inhibit both Th1 and Th2 master regulator factors, however long-term treatment favors Th2 expansion [111]. In addition, glucocorticoids induce T polarization toward Th2 phenotype through an increase of Itk expression, a Tec kinase inducing T helper 2 differentiation via negative regulation of T-bet [112, 113].

The effects of GCs on a third subset of IL-17-producing effector T helper cells, called Th17 cells, are still a matter of debate. Dexamethasone inhibits anti-CD3/anti-CD28-stimulated IFN- γ , IL-4, IL-17A, IL-17F, and IL-22 in various cell clones [114]. Interestingly, IL-17A and IL-17F, but not IL-22, lead to resistance of GC-induced apoptosis in in vitro-differentiated Th17 cells despite immunocytochemistry confirming glucocorticoid receptor translocation to the nucleus following treatment [114]. Mice lacking GILZ exhibit severe inflammation and a proinflammatory cytokine expression pattern in the imiquimod model of psoriasis, and DCs lacking GILZ produced greater IL-1, IL-23, and IL-6 in response to imiquimod stimulation in vitro [39]. These studies assessing glucocorticoid-dependent inhibition of IL-17 synthesis are in stark contrast with other studies describing Th17 sensitivity upon GC administration [115–117].

While the role of GC and Th17 is controversial, how glucocorticoids affect Treg function is much clearer. Both in humans and mice, treatment with dexamethasone increases the frequency of Treg cells, suggesting GC-mediated immune suppression is achieved, in part by enhancing Treg cell number or activity [118, 119] and by promoting the development of IL-10-producing T cells, an inducible peripheral Treg subpopulation [120]. There are several mechanisms that have been proposed to explain GC-mediated increase in Treg frequency. First, dexamethasone inhibits IL-2-mediated activation of T effector cells, increasing the proportion of Treg cells [118]; moreover, Treg cells were relatively more resistant to Dex-induced cell death and they were further protected by IL-2 [121]. Second, glucocorticoids synergize with TGF- β in FoxP3 induction [122, 123].

Glucocorticoid Function in B Cells

Circulating B lymphocytes are reduced by GC treatment, however not to the same extent as T cells [124]. This results from a reduction in splenic and lymph node B cell numbers. Furthermore, *in vivo* administration of dexamethasone to adrenalectomized mice reduced B cell numbers in both the spleen and bone marrow [125]. Studies on human leukemic lymphoblasts have shown that glucocorticoids have preferential apoptotic effects in certain lymphoid cell populations including B cell lymphomas [126]. B-lymphoblastic leukemia/lymphoma has been characterized in having increased expression of Bcl-2 resulting in resistance to glucocorticoid-induced apoptosis. Additionally, it was shown that deletion of GILZ in murine B lymphocytes leads to an accumulation of B cells in the bone marrow, blood, and lymphoid tissues due to impaired glucocorticoid-induced apoptosis. Since GILZ inhibits NF- κ B in B cells, increased nuclear translocation of p65 has been shown in GILZ-deficient cells resulting in an increase in Bcl-2 gene transcription [127].

Regarding the humoral immune response, glucocorticoids increase IgE synthesis, which is driven by the synergistic effects of hormones and IL-4 [128]. GC-induced increases in IgE synthesis support why systemic administration of corticosteroids does not interfere in skin prick tests to common allergens [128]. *In vivo* studies using mice deficient for either CD40L or CD40 lack serum IgE and fail to undergo isotype switching after immunization with T cell-dependent antigens [129, 130]. In addition, patients with X-linked hyper-IgM syndrome are deficient in CD40L and have low serum levels of IgG, IgA, and IgE due to impaired isotype switching [131]. To explain this effect of glucocorticoids, many studies suggest that glucocorticoid- and IL-4-induced IgE production is dependent on CD40L increased transcription and, thereby expression [132]. *In vitro* experiments demonstrated that agonist antibodies against CD40 mimic CD40L-dependent triggering of IL-4-driven isotype switching to IgE [132], while soluble CD40 inhibits IL-4 dependent IgE synthesis [133]. These results suggest that a rise in IgE production associated with glucocorticoid treatment is not clinically detrimental but presents additional immunomodulatory effects of corticosteroids on the T cell response.

Glucocorticoid Therapy and Resistance During Inflammation

The advantages of GC therapy in controlling and regulating inflammation are many, however due to the numerous signaling pathways glucocorticoids activate, consequences of this class of corticosteroids can also result in harmful side effects. GC-related side effects include musculoskeletal complications such as osteoporosis, hypertension, rapid weight gain, diabetes, glaucoma, peptic ulcer disease, and decelerated wound healing [134, 135]. Additionally, an increased risk of infection can occur resulting from a compromised immune system [136].

The successful resolution of acute inflammation afforded by glucocorticoids occurs by the delicate balance of pro- and anti-inflammatory molecules. However, long-term glucocorticoid therapy for the treatment of chronic inflammation typically results in reduced anti-inflammatory effects. These diminished effects can occur through a variety of ways including down-regulation of the GR itself [137, 138], defective GR binding and translocation (exemplified by GR^{dim}) [139, 140], GR nitrosylation by nitric oxide (NO) donors [141], and/or increased expression of GR β which competes with GR α for binding to GREs [142]. Recently, it was shown that hypoxia attenuates the anti-inflammatory effects of GCs by down-regulating GR along with inhibiting nuclear translocation [143]. Genetic factors may also contribute to GC resistance [144], such as the occurrence of polymorphisms in GR that may occur within families. A recent study by Mohamed et al. suggested a marked association of the glucocorticoid receptor 646 C>G polymorphism in resistance to GCs, resulting in severe bronchial asthma [145]. Finally, causative factors for glucocorticoid resistance go beyond defects in GR and include cigarette smoke, where in asthmatic patients who smoke, diminished anti-inflammatory actions in response to glucocorticoids were observed [146], viral infections where a study showed that rhinovirus infection can reduce GR nuclear translocation and GC function [147], and hypoxia observed at the site of inflammation that can impair GR transactivation [148].

Despite the well-known adverse side effects of prolonged GC treatment and the occurrence of GC resistance associated with long-term usage of glucocorticoids, these stress hormones remain the most effective treatment and commonly prescribed medication for controlling inflammation. The beneficial effects of GCs in treating anti-inflammatory and immunosuppressive disorders such as rheumatic diseases, allergy, asthma, and sepsis still outweigh their unfavorable consequences. Further research into the practice of GC therapy for combating inflammation to minimize harmful side effects and reduce the resistance associated with chronic treatment will be required to fully understand the pharmacological characteristics and biological actions of these stress hormones.

References

1. Munck A. Studies on the mode of action of glucocorticoids in rats. II. The effects in vivo and in vitro on net glucose uptake by isolated adipose tissue. *Biochim Biophys Acta*. 1962;57:318–26.
2. Munck A, Koritz SB. Studies on the mode of action of glucocorticoids in rats. I. Early effects of cortisol on blood glucose and on glucose entry into muscle, liver and adipose tissue. *Biochim Biophys Acta*. 1962;57:310–7.
3. Oppong E, Cato AC. Effects of Glucocorticoids in the Immune System. *Adv Exp Med Biol*. 2015;872:217–33.
4. Savino W, Mendes-da-Cruz DA, Lepletier A, Dardenne M. Hormonal control of T-cell development in health and disease. *Nat Rev Endocrinol*. 2016;12(2):77–89.
5. Singh RR, Cuffe JS, Moritz KM. Short- and long-term effects of exposure to natural and synthetic glucocorticoids during development. *Clin Exp Pharmacol Physiol*. 2012;39(11):979–89.
6. Grier DG, Halliday HL. Effects of glucocorticoids on fetal and neonatal lung development. *Treat Respir Med*. 2004;3(5):295–306.

7. Bellavance MA, Rivest S. The HPA-immune axis and the immunomodulatory actions of glucocorticoids in the brain. *Front Immunol.* 2014;5:136.
8. Evans RM. The steroid and thyroid hormone receptor superfamily. *Science.* 1988;240(4854):889–95.
9. Kumar R, Thompson EB. Gene regulation by the glucocorticoid receptor: structure: function relationship. *J Steroid Biochem Mol Biol.* 2005;94(5):383–94.
10. Lonard DM, O'Malley BW. Expanding functional diversity of the coactivators. *Trends Biochem Sci.* 2005;30(3):126–32.
11. Oakley RH, Cidlowski JA. The biology of the glucocorticoid receptor: new signaling mechanisms in health and disease. *J Allergy Clin Immunol.* 2013;132(5):1033–44.
12. Lewis-Tuffin LJ, Cidlowski JA. The physiology of human glucocorticoid receptor beta (hGRbeta) and glucocorticoid resistance. *Ann N Y Acad Sci.* 2006;1069:1–9.
13. Oakley RH, Jewell CM, Yudit MR, Bofetiado DM, Cidlowski JA. The dominant negative activity of the human glucocorticoid receptor beta isoform. Specificity and mechanisms of action. *J Biol Chem.* 1999;274(39):27857–66.
14. Oakley RH, Webster JC, Sar M, Parker Jr CR, Cidlowski JA. Expression and subcellular distribution of the beta-isoform of the human glucocorticoid receptor. *Endocrinology.* 1997;138(11):5028–38.
15. Coussens LM, Werb Z. Inflammation and cancer. *Nature.* 2002;420(6917):860–7.
16. Barnes PJ. Mechanisms and resistance in glucocorticoid control of inflammation. *J Steroid Biochem Mol Biol.* 2010;120(2-3):76–85.
17. Clark AR. Anti-inflammatory functions of glucocorticoid-induced genes. *Mol Cell Endocrinol.* 2007;275(1-2):79–97.
18. Smoak KA, Cidlowski JA. Mechanisms of glucocorticoid receptor signaling during inflammation. *Mech Ageing Dev.* 2004;125(10-11):697–706.
19. Gottlicher M, Heck S, Herrlich P. Transcriptional cross-talk, the second mode of steroid hormone receptor action. *J Mol Med (Berl).* 1998;76(7):480–9.
20. Barnes PJ. Corticosteroid effects on cell signalling. *Eur Respir J.* 2006;27(2):413–26.
21. De Bosscher K, Vanden Berghe W, Haegeman G. The interplay between the glucocorticoid receptor and nuclear factor-kappaB or activator protein-1: molecular mechanisms for gene repression. *Endocr Rev.* 2003;24(4):488–522.
22. De Bosscher K, Vanden Berghe W, Haegeman G. Cross-talk between nuclear receptors and nuclear factor kappaB. *Oncogene.* 2006;25(51):6868–86.
23. Ratman D, Vanden Berghe W, Dejager L, Libert C, Tavernier J, Beck IM, et al. How glucocorticoid receptors modulate the activity of other transcription factors: a scope beyond tethering. *Mol Cell Endocrinol.* 2013;380(1-2):41–54.
24. Altonsy MO, Sasse SK, Phang TL, Gerber AN. Context-dependent cooperation between nuclear factor kappaB (NF-kappaB) and the glucocorticoid receptor at a TNFAIP3 intronic enhancer: a mechanism to maintain negative feedback control of inflammation. *J Biol Chem.* 2014;289(12):8231–9.
25. Jantzen HM, Strahle U, Gloss B, Stewart F, Schmid W, Boshart M, et al. Cooperativity of glucocorticoid response elements located far upstream of the tyrosine aminotransferase gene. *Cell.* 1987;49(1):29–38.
26. Glass CK, Rosenfeld MG. The coregulator exchange in transcriptional functions of nuclear receptors. *Genes Dev.* 2000;14(2):121–41.
27. Simons Jr SS. Glucocorticoid receptor cofactors as therapeutic targets. *Curr Opin Pharmacol.* 2010;10(6):613–9.
28. Hahn RT, Hoppstadter J, Hirschfelder K, Hachenthal N, Diesel B, Kessler SM, et al. Downregulation of the glucocorticoid-induced leucine zipper (GILZ) promotes vascular inflammation. *Atherosclerosis.* 2014;234(2):391–400.
29. Smoak K, Cidlowski JA. Glucocorticoids regulate tristetraprolin synthesis and posttranscriptionally regulate tumor necrosis factor alpha inflammatory signaling. *Mol Cell Biol.* 2006;26(23):9126–35.
30. Weigel NL, Moore NL. Steroid receptor phosphorylation: a key modulator of multiple receptor functions. *Mol Endocrinol.* 2007;21(10):2311–9.

31. Hafezi-Moghadam A, Simoncini T, Yang Z, Limbourg FP, Plumier JC, Rebsamen MC, et al. Acute cardiovascular protective effects of corticosteroids are mediated by non-transcriptional activation of endothelial nitric oxide synthase. *Nat Med*. 2002;8(5):473–9.
32. Connelly L, Jacobs AT, Palacios-Callender M, Moncada S, Hobbs AJ. Macrophage endothelial nitric-oxide synthase autoregulates cellular activation and pro-inflammatory protein expression. *J Biol Chem*. 2003;278(29):26480–7.
33. Grumbach IM, Chen W, Mertens SA, Harrison DG. A negative feedback mechanism involving nitric oxide and nuclear factor kappa-B modulates endothelial nitric oxide synthase transcription. *J Mol Cell Cardiol*. 2005;39(4):595–603.
34. Mitre-Aguilar IB, Cabrera-Quintero AJ, Zentella-Dehesa A. Genomic and non-genomic effects of glucocorticoids: implications for breast cancer. *Int J Clin Exp Pathol*. 2015;8(1):1–10.
35. Stellato C. Posttranscriptional gene regulation: novel pathways for glucocorticoids' anti-inflammatory action. *Transl Med UniSa*. 2012;3:67–73.
36. Anderson P. Post-transcriptional regulons coordinate the initiation and resolution of inflammation. *Nat Rev Immunol*. 2010;10(1):24–35.
37. D'Adamo F, Zollo O, Moraca R, Ayroldi E, Bruscoli S, Bartoli A, et al. A new dexamethasone-induced gene of the leucine zipper family protects T lymphocytes from TCR/CD3-activated cell death. *Immunity*. 1997;7(6):803–12.
38. Ayroldi E, Migliorati G, Bruscoli S, Marchetti C, Zollo O, Cannarile L, et al. Modulation of T-cell activation by the glucocorticoid-induced leucine zipper factor via inhibition of nuclear factor kappaB. *Blood*. 2001;98(3):743–53.
39. Jones SA, Perera DN, Fan H, Russ BE, Harris J, Morand EF. GILZ regulates Th17 responses and restrains IL-17-mediated skin inflammation. *J Autoimmun*. 2015;61:73–80.
40. Ayroldi E, Riccardi C. Glucocorticoid-induced leucine zipper (GILZ): a new important mediator of glucocorticoid action. *FASEB J*. 2009;23(11):3649–58.
41. Ronchetti S, Migliorati G, Riccardi C. GILZ as a mediator of the anti-inflammatory effects of glucocorticoids. *Front Endocrinol (Lausanne)*. 2015;6:170.
42. Espinasse MA, Pepin A, Virault-Rocroy P, Szely N, Chollet-Martin S, Pallardy M, et al. Glucocorticoid-Induced Leucine Zipper Is Expressed in Human Neutrophils and Promotes Apoptosis through Mcl-1 Down-Regulation. *J Innate Immun*. 2016;8(1):81–96.
43. Vettorazzi S, Bode C, Dejager L, Frappart L, Shelest E, Klassen C, et al. Glucocorticoids limit acute lung inflammation in concert with inflammatory stimuli by induction of SphK1. *Nat Commun*. 2015;6:7796.
44. Gibbs J, Ince L, Matthews L, Mei J, Bell T, Yang N, et al. An epithelial circadian clock controls pulmonary inflammation and glucocorticoid action. *Nat Med*. 2014;20(8):919–26.
45. Smith JB, Herschman HR. Glucocorticoid-attenuated response genes encode intercellular mediators, including a new C-X-C chemokine. *J Biol Chem*. 1995;270(28):16756–65.
46. Loiarro M, Ruggiero V, Sette C. Targeting TLR/IL-1R signalling in human diseases. *Mediators Inflamm*. 2010;2010:674363.
47. Miyata M, Lee JY, Susuki-Miyata S, Wang WY, Xu H, Kai H, et al. Glucocorticoids suppress inflammation via the upregulation of negative regulator IRAK-M. *Nat Commun*. 2015;6:6062.
48. Bunim JJ. The clinical effects of cortisone and ACTH on rheumatic diseases. *Bull N Y Acad Med*. 1951;27(2):75–100.
49. Hench P. Effects of cortisone in the rheumatic diseases. *Lancet*. 1950;2(6634):483–4.
50. Rhen T, Cidlowski JA. Antiinflammatory action of glucocorticoids--new mechanisms for old drugs. *N Engl J Med*. 2005;353(16):1711–23.
51. De Bosscher K, Vanden Berghe W, Haegeman G. Mechanisms of anti-inflammatory action and of immunosuppression by glucocorticoids: negative interference of activated glucocorticoid receptor with transcription factors. *J Neuroimmunol*. 2000;109(1):16–22.
52. Zhong HJ, Wang HY, Yang C, Zhou JY, Jiang JX. Low concentrations of corticosterone exert stimulatory effects on macrophage function in a manner dependent on glucocorticoid receptors. *Int J Endocrinol*. 2013;2013:405127.
53. Zhou JY, Zhong HJ, Yang C, Yan J, Wang HY, Jiang JX. Corticosterone exerts immunostimulatory effects on macrophages via endoplasmic reticulum stress. *Br J Surg*. 2010;97(2):281–93.

54. Medzhitov R, Horng T. Transcriptional control of the inflammatory response. *Nat Rev Immunol.* 2009;9(10):692–703.
55. Akira S. Pathogen recognition by innate immunity and its signaling. *Proc Jpn Acad Ser B Phys Biol Sci.* 2009;85(4):143–56.
56. Pancer Z, Cooper MD. The evolution of adaptive immunity. *Annu Rev Immunol.* 2006;24:497–518.
57. Baschant U, Tuckermann J. The role of the glucocorticoid receptor in inflammation and immunity. *J Steroid Biochem Mol Biol.* 2010;120(2-3):69–75.
58. Coutinho AE, Chapman KE. The anti-inflammatory and immunosuppressive effects of glucocorticoids, recent developments and mechanistic insights. *Mol Cell Endocrinol.* 2011;335(1):2–13.
59. Lim HY, Muller N, Herold MJ, van den Brandt J, Reichardt HM. Glucocorticoids exert opposing effects on macrophage function dependent on their concentration. *Immunology.* 2007;122(1):47–53.
60. McColl A, Bourmazos S, Franz S, Perretti M, Morgan BP, Haslett C, et al. Glucocorticoids induce protein S-dependent phagocytosis of apoptotic neutrophils by human macrophages. *J Immunol.* 2009;183(3):2167–75.
61. McColl A, Michlewska S, Dransfield I, Rossi AG. Effects of glucocorticoids on apoptosis and clearance of apoptotic cells. *ScientificWorldJournal.* 2007;7:1165–81.
62. Roszer T. Understanding the mysterious M2 macrophage through activation markers and effector mechanisms. *Mediators Inflamm.* 2015;2015:816460.
63. Tang Z, Niven-Fairchild T, Tadesse S, Norwitz ER, Buhimschi CS, Buhimschi IA, et al. Glucocorticoids enhance CD163 expression in placental Hofbauer cells. *Endocrinology.* 2013;154(1):471–82.
64. Warren MK, Vogel SN. Opposing effects of glucocorticoids on interferon-gamma-induced murine macrophage Fc receptor and Ia antigen expression. *J Immunol.* 1985;134(4):2462–9.
65. Bhattacharyya S, Brown DE, Brewer JA, Vogt SK, Muglia LJ. Macrophage glucocorticoid receptors regulate Toll-like receptor 4-mediated inflammatory responses by selective inhibition of p38 MAP kinase. *Blood.* 2007;109(10):4313–9.
66. Tuckermann JP, Kleiman A, Moriggl R, Spanbroek R, Neumann A, Illing A, et al. Macrophages and neutrophils are the targets for immune suppression by glucocorticoids in contact allergy. *J Clin Invest.* 2007;117(5):1381–90.
67. Broug-Holub E, Kraal G. Dose- and time-dependent activation of rat alveolar macrophages by glucocorticoids. *Clin Exp Immunol.* 1996;104(2):332–6.
68. Ehrchen J, Steinmuller L, Barczyk K, Tenbrock K, Nacken W, Eisenacher M, et al. Glucocorticoids induce differentiation of a specifically activated, anti-inflammatory subtype of human monocytes. *Blood.* 2007;109(3):1265–74.
69. Giles KM, Ross K, Rossi AG, Hotchin NA, Haslett C, Dransfield I. Glucocorticoid augmentation of macrophage capacity for phagocytosis of apoptotic cells is associated with reduced p130Cas expression, loss of paxillin/pyk2 phosphorylation, and high levels of active Rac. *J Immunol.* 2001;167(2):976–86.
70. Heasman SJ, Giles KM, Ward C, Rossi AG, Haslett C, Dransfield I. Glucocorticoid-mediated regulation of granulocyte apoptosis and macrophage phagocytosis of apoptotic cells: implications for the resolution of inflammation. *J Endocrinol.* 2003;178(1):29–36.
71. Meagher LC, Cousin JM, Seckl JR, Haslett C. Opposing effects of glucocorticoids on the rate of apoptosis in neutrophilic and eosinophilic granulocytes. *J Immunol.* 1996;156(11):4422–8.
72. Cox G. Glucocorticoid treatment inhibits apoptosis in human neutrophils. Separation of survival and activation outcomes. *J Immunol.* 1995;154(9):4719–25.
73. Liles WC, Dale DC, Klebanoff SJ. Glucocorticoids inhibit apoptosis of human neutrophils. *Blood.* 1995;86(8):3181–8.
74. Yoshimura C, Miyamasu M, Nagase H, Iikura M, Yamaguchi M, Kawanami O, et al. Glucocorticoids induce basophil apoptosis. *J Allergy Clin Immunol.* 2001;108(2):215–20.

75. Jia W, Wu J, Jia H, Yang Y, Zhang X, Chen K, et al. The peripheral blood neutrophil-to-lymphocyte ratio is superior to the lymphocyte-to-monocyte ratio for predicting the long-term survival of triple-negative breast cancer patients. *PLoS One*. 2015;10(11), e0143061.
76. Weber PS, Toelboell T, Chang LC, Tirrell JD, Saama PM, Smith GW, et al. Mechanisms of glucocorticoid-induced down-regulation of neutrophil L-selectin in cattle: evidence for effects at the gene-expression level and primarily on blood neutrophils. *J Leukoc Biol*. 2004;75(5):815–27.
77. Re F, Muzio M, De Rossi M, Polentarutti N, Giri JG, Mantovani A, et al. The type II “receptor” as a decoy target for interleukin 1 in polymorphonuclear leukocytes: characterization of induction by dexamethasone and ligand binding properties of the released decoy receptor. *J Exp Med*. 1994;179(2):739–43.
78. Lunemann A, Lunemann JD, Munz C. Regulatory NK-cell functions in inflammation and autoimmunity. *Mol Med*. 2009;15(9-10):352–8.
79. Eddy JL, Krukowski K, Janusek L, Mathews HL. Glucocorticoids regulate natural killer cell function epigenetically. *Cell Immunol*. 2014;290(1):120–30.
80. Fauci AS, Dale DC, Balow JE. Glucocorticosteroid therapy: mechanisms of action and clinical considerations. *Ann Intern Med*. 1976;84(3):304–15.
81. Piemonti L, Monti P, Allavena P, Sironi M, Soldini L, Leone BE, et al. Glucocorticoids affect human dendritic cell differentiation and maturation. *J Immunol*. 1999;162(11):6473–81.
82. Woltman AM, de Fijter JW, Kamerling SW, Paul LC, Daha MR, van Kooten C. The effect of calcineurin inhibitors and corticosteroids on the differentiation of human dendritic cells. *Eur J Immunol*. 2000;30(7):1807–12.
83. Chabot V, Martin L, Meley D, Sensebe L, Baron C, Lebranchu Y, et al. Unexpected impairment of TNF-alpha-induced maturation of human dendritic cells in vitro by IL-4. *J Transl Med*. 2016;14(1):93.
84. Bros M, Jahrling F, Renzing A, Wiechmann N, Dang NA, Sutter A, et al. A newly established murine immature dendritic cell line can be differentiated into a mature state, but exerts tolerogenic function upon maturation in the presence of glucocorticoid. *Blood*. 2007;109(9):3820–9.
85. Zimmer A, Luce S, Gaignier F, Nony E, Naveau M, Biola-Vidamment A, et al. Identification of a new phenotype of tolerogenic human dendritic cells induced by fungal proteases from *Aspergillus oryzae*. *J Immunol*. 2011;186(7):3966–76.
86. Bosma BM, Metselaar HJ, Nagtzaam NM, de Haan R, Mancham S, van der Laan LJ, et al. Dexamethasone transforms lipopolysaccharide-stimulated human blood myeloid dendritic cells into myeloid dendritic cells that prime interleukin-10 production in T cells. *Immunology*. 2008;125(1):91–100.
87. Kapsenberg ML. Dendritic-cell control of pathogen-driven T-cell polarization. *Nat Rev Immunol*. 2003;3(12):984–93.
88. Calmette J, Ellouze M, Tran T, Karaki S, Ronin E, Capel F, et al. Glucocorticoid-induced leucine zipper enhanced expression in dendritic cells is sufficient to drive regulatory T cells expansion in vivo. *J Immunol*. 2014;193(12):5863–72.
89. Zimmer A, Bouley J, Le Mignon M, Pliquet E, Horiot S, Turfkruyer M, et al. A regulatory dendritic cell signature correlates with the clinical efficacy of allergen-specific sublingual immunotherapy. *J Allergy Clin Immunol*. 2012;129(4):1020–30.
90. Schlecht G, Leclerc C, Dadaglio G. Induction of CTL and nonpolarized Th cell responses by CD8alpha(+) and CD8alpha(-) dendritic cells. *J Immunol*. 2001;167(8):4215–21.
91. Wang Y, Kissenpfennig A, Mingueneau M, Richelme S, Perrin P, Chevrier S, et al. Th2 lymphoproliferative disorder of LatY136F mutant mice unfolds independently of TCR-MHC engagement and is insensitive to the action of Foxp3+ regulatory T cells. *J Immunol*. 2008;180(3):1565–75.
92. Kamanaka M, Kim ST, Wan YY, Sutterwala FS, Lara-Tejero M, Galan JE, et al. Expression of interleukin-10 in intestinal lymphocytes detected by an interleukin-10 reporter knockin tiger mouse. *Immunity*. 2006;25(6):941–52.
93. Inaba K, Inaba M, Romani N, Aya H, Deguchi M, Ikehara S, et al. Generation of large numbers of dendritic cells from mouse bone marrow cultures supplemented with granulocyte/macrophage colony-stimulating factor. *J Exp Med*. 1992;176(6):1693–702.

94. Spallanzani RG, Torres NI, Avila DE, Ziblat A, Iraolagoitia XL, Rossi LE, et al. Regulatory dendritic cells restrain NK Cell IFN- γ production through mechanisms involving NKp46, IL-10, and MHC Class I-specific inhibitory receptors. *J Immunol.* 2015;195(5):2141–8.
95. Wu J, Horuzsko A. Expression and function of immunoglobulin-like transcripts on tolerogenic dendritic cells. *Hum Immunol.* 2009;70(5):353–6.
96. Karaki S, Garcia G, Tcherakian C, Capel F, Tran T, Pallardy M, et al. Enhanced glucocorticoid-induced leucine zipper in dendritic cells induces allergen-specific regulatory CD4(+) T-cells in respiratory allergies. *Allergy.* 2014;69(5):624–31.
97. Benkhoucha M, Molnarfi N, Dunand-Sauthier I, Merkler D, Schneider G, Bruscoli S, et al. Hepatocyte growth factor limits autoimmune neuroinflammation via glucocorticoid-induced leucine zipper expression in dendritic cells. *J Immunol.* 2014;193(6):2743–52.
98. Vicente-Suarez I, Larange A, Reardon C, Matho M, Feau S, Chodaczek G, et al. Unique lamina propria stromal cells imprint the functional phenotype of mucosal dendritic cells. *Mucosal Immunol.* 2015;8(1):141–51.
99. Kania G, Siegert S, Behnke S, Prados-Rosales R, Casadevall A, Luscher TF, et al. Innate signaling promotes formation of regulatory nitric oxide-producing dendritic cells limiting T-cell expansion in experimental autoimmune myocarditis. *Circulation.* 2013;127(23):2285–94.
100. Glennon E, Kaunzner UW, Gagnidze K, McEwen BS, Bulloch K. Pituitary dendritic cells communicate immune pathogenic signals. *Brain Behav Immun.* 2015;50:232–40.
101. Kadmiel M, Cidlowski JA. Glucocorticoid receptor signaling in health and disease. *Trends Pharmacol Sci.* 2013;34(9):518–30.
102. Erlacher M, Knoflach M, Stec IE, Bock G, Wick G, Wieggers GJ. TCR signaling inhibits glucocorticoid-induced apoptosis in murine thymocytes depending on the stage of development. *Eur J Immunol.* 2005;35(11):3287–96.
103. Inomata T, Nakamura T. Influence of adrenalectomy on the development of the neonatal thymus in the rat. *Biol Neonate.* 1989;55(4-5):238–43.
104. Delfino DV, Agostini M, Spinicelli S, Vito P, Riccardi C. Decrease of Bcl-xL and augmentation of thymocyte apoptosis in GILZ overexpressing transgenic mice. *Blood.* 2004;104(13):4134–41.
105. Purton JF, Boyd RL, Cole TJ, Godfrey DI. Intrathymic T cell development and selection proceeds normally in the absence of glucocorticoid receptor signaling. *Immunity.* 2000;13(2):179–86.
106. Wieggers GJ, Kaufmann M, Tischner D, Villunger A. Shaping the T-cell repertoire: a matter of life and death. *Immunol Cell Biol.* 2011;89(1):33–9.
107. van den Brandt J, Luhder F, McPherson KG, de Graaf KL, Tischner D, Wiehr S, et al. Enhanced glucocorticoid receptor signaling in T cells impacts thymocyte apoptosis and adaptive immune responses. *Am J Pathol.* 2007;170(3):1041–53.
108. Baumann S, Dostert A, Novac N, Bauer A, Schmid W, Fas SC, et al. Glucocorticoids inhibit activation-induced cell death (AICD) via direct DNA-dependent repression of the CD95 ligand gene by a glucocorticoid receptor dimer. *Blood.* 2005;106(2):617–25.
109. Han S, Choi H, Ko MG, Choi YI, Sohn DH, Kim JK, et al. Peripheral T cells become sensitive to glucocorticoid- and stress-induced apoptosis in transgenic mice overexpressing SRG3. *J Immunol.* 2001;167(2):805–10.
110. Liberman AC, Antunica-Noguerol M, Ferraz-de-Paula V, Palermo-Neto J, Castro CN, Druker J, et al. Compound A, a dissociated glucocorticoid receptor modulator, inhibits T-bet (Th1) and induces GATA-3 (Th2) activity in immune cells. *PLoS One.* 2012;7(4), e35155.
111. Liberman AC, Druker J, Refojo D, Holsboer F, Arzt E. Glucocorticoids inhibit GATA-3 phosphorylation and activity in T cells. *FASEB J.* 2009;23(5):1558–71.
112. Petrillo MG, Fettucciari K, Montuschi P, Ronchetti S, Cari L, Migliorati G, et al. Transcriptional regulation of kinases downstream of the T cell receptor: another immunomodulatory mechanism of glucocorticoids. *BMC Pharmacol Toxicol.* 2014;15:35.
113. Miller AT, Wilcox HM, Lai Z, Berg LJ. Signaling through Itk promotes T helper 2 differentiation via negative regulation of T-bet. *Immunity.* 2004;21(1):67–80.
114. Banuelos J, Shin S, Cao Y, Bochner BS, Morales-Nebreda L, Budinger GR, et al. BCL-2 protects human and mouse Th17 cells from glucocorticoid-induced apoptosis. *Allergy.* 2016;71(5):640–50.

115. Momcilovic M, Miljkovic Z, Popadic D, Markovic M, Savic E, Ramic Z, et al. Methylprednisolone inhibits interleukin-17 and interferon-gamma expression by both naive and primed T cells. *BMC Immunol.* 2008;9:47.
116. Schewitz-Bowers LP, Lait PJ, Copland DA, Chen P, Wu W, Dhanda AD, et al. Glucocorticoid-resistant Th17 cells are selectively attenuated by cyclosporine A. *Proc Natl Acad Sci U S A.* 2015;112(13):4080–5.
117. Chambers ES, Nanzer AM, Pfeffer PE, Richards DF, Timms PM, Martineau AR, et al. Distinct endotypes of steroid-resistant asthma characterized by IL-17A(high) and IFN-gamma(high) immunophenotypes: potential benefits of calcitriol. *J Allergy Clin Immunol.* 2015;136(3):628–37. e4.
118. Chen X, Oppenheim JJ, Winkler-Pickett RT, Ortaldo JR, Howard OM. Glucocorticoid amplifies IL-2-dependent expansion of functional FoxP3(+)/CD4(+)/CD25(+) T regulatory cells in vivo and enhances their capacity to suppress EAE. *Eur J Immunol.* 2006;36(8):2139–49.
119. Suarez A, Lopez P, Gomez J, Gutierrez C. Enrichment of CD4+ CD25high T cell population in patients with systemic lupus erythematosus treated with glucocorticoids. *Ann Rheum Dis.* 2006;65(11):1512–7.
120. Karagiannidis C, Akdis M, Holopainen P, Woolley NJ, Hense G, Ruckert B, et al. Glucocorticoids upregulate FOXP3 expression and regulatory T cells in asthma. *J Allergy Clin Immunol.* 2004;114(6):1425–33.
121. Chen X, Murakami T, Oppenheim JJ, Howard OM. Differential response of murine CD4+CD25+ and CD4+CD25- T cells to dexamethasone-induced cell death. *Eur J Immunol.* 2004;34(3):859–69.
122. Bereshchenko O, Coppo M, Bruscoli S, Biagioli M, Cimino M, Frammartino T, et al. GILZ promotes production of peripherally induced Treg cells and mediates the crosstalk between glucocorticoids and TGF-beta signaling. *Cell Rep.* 2014;7(2):464–75.
123. Prado C, Gomez J, Lopez P, de Paz B, Gutierrez C, Suarez A. Dexamethasone upregulates FOXP3 expression without increasing regulatory activity. *Immunobiology.* 2011;216(3):386–92.
124. Slade JD, Hepburn B. Prednisone-induced alterations of circulating human lymphocyte subsets. *J Lab Clin Med.* 1983;101(3):479–87.
125. Gruver-Yates AL, Quinn MA, Cidlowski JA. Analysis of glucocorticoid receptors and their apoptotic response to dexamethasone in male murine B cells during development. *Endocrinology.* 2014;155(2):463–74.
126. Smith LK, Cidlowski JA. Glucocorticoid-induced apoptosis of healthy and malignant lymphocytes. *Prog Brain Res.* 2010;182:1–30.
127. Bruscoli S, Biagioli M, Sorcini D, Frammartino T, Cimino M, Sportoletti P, et al. Lack of glucocorticoid-induced leucine zipper (GILZ) deregulates B-cell survival and results in B-cell lymphocytosis in mice. *Blood.* 2015;126(15):1790–801.
128. Barnes PJ. Corticosteroids, IgE, and atopy. *J Clin Invest.* 2001;107(3):265–6.
129. Xu J, Foy TM, Laman JD, Elliott EA, Dunn JJ, Waldschmidt TJ, et al. Mice deficient for the CD40 ligand. *Immunity.* 1994;1(5):423–31.
130. Kawabe T, Naka T, Yoshida K, Tanaka T, Fujiwara H, Suematsu S, et al. The immune responses in CD40-deficient mice: impaired immunoglobulin class switching and germinal center formation. *Immunity.* 1994;1(3):167–78.
131. Callard RE, Smith SH, Herbert J, Morgan G, Padayachee M, Lederman S, et al. CD40 ligand (CD40L) expression and B cell function in agammaglobulinemia with normal or elevated levels of IgM (HIM). Comparison of X-linked, autosomal recessive, and non-X-linked forms of the disease, and obligate carriers. *J Immunol.* 1994;153(7):3295–306.
132. Jabara HH, Fu SM, Geha RS, Vercelli D. CD40 and IgE: synergism between anti-CD40 monoclonal antibody and interleukin 4 in the induction of IgE synthesis by highly purified human B cells. *J Exp Med.* 1990;172(6):1861–4.
133. Fanslow WC, Anderson DM, Grabstein KH, Clark EA, Cosman D, Armitage RJ. Soluble forms of CD40 inhibit biologic responses of human B cells. *J Immunol.* 1992;149(2):655–60.
134. Ozbakir B, Crielgaard BJ, Metselaar JM, Storm G, Lammers T. Liposomal corticosteroids for the treatment of inflammatory disorders and cancer. *J Control Release.* 2014;190:624–36.

135. Kleiman A, Tuckermann JP. Glucocorticoid receptor action in beneficial and side effects of steroid therapy: lessons from conditional knockout mice. *Mol Cell Endocrinol.* 2007;275(1-2):98–108.
136. Kulkarni NN, Gunnarsson HI, Yi Z, Gudmundsdottir S, Sigurjonsson OE, Agerberth B, et al. Glucocorticoid dexamethasone down-regulates basal and vitamin D3 induced cathelicidin expression in human monocytes and bronchial epithelial cell line. *Immunobiology.* 2016;221(2):245–52.
137. Cidlowski JA, Cidlowski NB. Regulation of glucocorticoid receptors by glucocorticoids in cultured HeLa S3 cells. *Endocrinology.* 1981;109(6):1975–82.
138. Okret S, Poellinger L, Dong Y, Gustafsson JA. Down-regulation of glucocorticoid receptor mRNA by glucocorticoid hormones and recognition by the receptor of a specific binding sequence within a receptor cDNA clone. *Proc Natl Acad Sci U S A.* 1986;83(16):5899–903.
139. Adams M, Meijer OC, Wang J, Bhargava A, Pearce D. Homodimerization of the glucocorticoid receptor is not essential for response element binding: activation of the phenylethanolamine N-methyltransferase gene by dimerization-defective mutants. *Mol Endocrinol.* 2003;17(12):2583–92.
140. Jewell CM, Scoltock AB, Hamel BL, Yudit MR, Cidlowski JA. Complex human glucocorticoid receptor dim mutations define glucocorticoid induced apoptotic resistance in bone cells. *Mol Endocrinol.* 2012;26(2):244–56.
141. Galigniana MD, Piwien-Pilipuk G, Assreuy J. Inhibition of glucocorticoid receptor binding by nitric oxide. *Mol Pharmacol.* 1999;55(2):317–23.
142. Webster JC, Oakley RH, Jewell CM, Cidlowski JA. Proinflammatory cytokines regulate human glucocorticoid receptor gene expression and lead to the accumulation of the dominant negative beta isoform: a mechanism for the generation of glucocorticoid resistance. *Proc Natl Acad Sci U S A.* 2001;98(12):6865–70.
143. Zhang P, Fang L, Wu H, Ding P, Shen Q, Liu R. Down-regulation of GRalpha expression and inhibition of its nuclear translocation by hypoxia. *Life Sci.* 2016;146:92–9.
144. Silverman MN, Sternberg EM. Glucocorticoid regulation of inflammation and its functional correlates: from HPA axis to glucocorticoid receptor dysfunction. *Ann N Y Acad Sci.* 2012;1261:55–63.
145. Mohamed NA, Abdel-Rehim AS, Farres MN, Muhammed HS. Influence of glucocorticoid receptor gene NR3C1 646 C>G polymorphism on glucocorticoid resistance in asthmatics: a preliminary study. *Cent Eur J Immunol.* 2015;40(3):325–30.
146. Hew M, Chung KF. Corticosteroid insensitivity in severe asthma: significance, mechanisms and aetiology. *Intern Med J.* 2010;40(5):323–34.
147. Papi A, Contoli M, Adcock IM, Bellettato C, Padovani A, Casolari P, et al. Rhinovirus infection causes steroid resistance in airway epithelium through nuclear factor kappaB and c-Jun N-terminal kinase activation. *J Allergy Clin Immunol.* 2013;132(5):1075–85. e6.
148. Kodama T, Shimizu N, Yoshikawa N, Makino Y, Ouchida R, Okamoto K, et al. Role of the glucocorticoid receptor for regulation of hypoxia-dependent gene expression. *J Biol Chem.* 2003;278(35):33384–91.