

Robert P. Dickson

Case Presentation

A 67 year-old man with no recent hospitalizations presents to the Emergency Department with shortness of breath. He has a history of ulcerative colitis and is currently treated with cyclosporine and prednisone 10 mg/day. He denies fevers, chills or sputum production. Pulse oximetry is 82% on room air. Initial chest x-ray and high-resolution CT scan of the chest are shown (Figs. 24.1 and 24.2). Over the next 24 h, he experiences progressive hypoxemia and respiratory distress despite supplemental oxygen and empiric antibiotic therapy for community-acquired pneumonia (ceftriaxone and azithromycin). The patient undergoes endotracheal intubation and mechanical ventilation is initiated.

Question Should the patient's antimicrobial regimen be changed? What diagnostic test should be performed?

Answer The patient's antimicrobial regimen should be expanded empirically to cover *Pneumocystis jirovecii* (e.g. trimethoprim-sulfamethoxazole) given (1) his risk factors (cyclosporine and corticosteroids), (2) his consistent CT scan (interstitial infiltrate with cystic

changes), (3) his hypoxemia disproportionate to radiographic infiltrate, (4) his lack of clinical response to an empiric regimen adequate for community-acquired pneumonia, and (5) the fact that empiric therapy does not compromise the diagnostic yield of subsequent bronchoscopy in the diagnosis of *Pneumocystis pneumonia* [1]. A lower respiratory tract specimen should be acquired, via bronchoscopy or mini-bronchoalveolar lavage (BAL); lavage fluid should be tested for gram stain and culture, respiratory virus polymerase chain reaction (PCR), fungal culture, galactomannan, acid-fast stain and culture, and *Pneumocystis* PCR.

The patient underwent flexible bronchoscopy, and a positive *Pneumocystis* PCR assay confirmed the diagnosis. The patient received intravenous trimethoprim-sulfamethoxazole, and oxygenation gradually improved over the next 5 days. The patient was ultimately extubated and recovered full lung function. After 21 days of treatment, the patient's trimethoprim-sulfamethoxazole was changed to the prophylactic dose (1 double-strength tablet [160/800] once daily) for the duration of his immunosuppression.

Principles of Management

Presentation

Pneumonia is a common and morbid complication of immunosuppression, whether due to primary immunodeficiency or, more commonly, secondary

R.P. Dickson
Medicine (Pulmonary and Critical Care), University
of Michigan Health System, Ann Arbor, MI, USA
e-mail: rodickso@med.umich.edu



Fig. 24.1 Chest X-ray

to a systemic disease process or its treatment. The presentation of pneumonia among immunosuppressed patients is often more subtle, indolent and atypical than among immunocompetent patients [2]; the same immune deficits that permit microbial reproduction in the lower respiratory tract can decrease the intensity of fever, sputum production, or radiographic infiltrates. Immunosuppressed patients are often vulnerable to competing or concurrent non-infectious lung processes such as cardiogenic edema (e.g. among patients receiving cardiotoxic chemotherapy or aggressive hydration with chemotherapeutic regimens), medication toxicity (e.g. among patients receiving bleomycin or methotrexate), radiation pneumonitis, or malignancy (e.g. Kaposi's Sarcoma among patients with Human Immunodeficiency Virus [HIV]/Acquired Immunodeficiency Syndrome [AIDS]).

Etiology

The presence and persistence of microbes in the respiratory tract are determined by the balance of microbial immigration, elimination and local microbial growth conditions [3, 4], all of which are altered in immunosuppressed patients. The microbiota of the upper respiratory tract (the primary source community for migration of microbes to the lungs [4, 5]) are altered by systemic immu-



Fig. 24.2 High-resolution CT scan

nosuppression, whether by underlying disease (e.g. HIV/AIDS) [6] or immune-suppressing medications [7]. Impairment of innate and adaptive immunity decreases the elimination rate of transient microbes, increasing the likelihood of persistent reproduction, and makes the microbial growth conditions of the lung environment more hospitable to dysregulated reproduction [3]. Each patient's specific constellation of immune deficits predisposes him/her to a select number of opportunistic pathogens (Table 24.1). Consideration of each patient's candidate pathogen profile is critical to the appropriate selection of empiric antimicrobial therapy. Despite the wide breadth of potential pathogens in this population, the most common culprits remain the bacteria and viruses responsible for community-acquired pneumonia (e.g. *Streptococcus pneumoniae*) [9], which should be covered by any empiric regimen. Coverage for atypical organisms (*Mycoplasma* spp., *L. pneumophila* and *C. pneumoniae*) is warranted in community-dwelling patients until a specific pathogen is identified.

Diagnosis

Chest x-rays are of notoriously poor sensitivity in identifying pneumonia among immunocompromised patients; in one large series, the major-

Table 24.1 Correspondence of immunodeficiency and susceptibility to respiratory pathogens

Immune defect		Disease examples	Iatrogenic examples	Organisms to suspect
Innate immunity	Neutrophil abundance	Leukemia Parvovirus infection Agranulocytosis	Chemotherapy Methotrexate Clozapine	Gram-negative bacilli <i>Staphylococcus</i> spp. Fungi (e.g. <i>Aspergillus</i> spp.)
	Neutrophil function	Chronic granulomatous disease Cirrhosis Uremia	Anti-TNF agents [8]	<i>Staphylococcus aureus</i> Fungi (e.g. <i>Aspergillus</i> spp.)
Adaptive immunity	T-cell abundance and function	HIV/AIDS Lymphoma Primary immunodeficiency	Chemotherapy Corticosteroids Calcineurin inhibitors Anti-T-cell antibodies	<i>Pneumocystis jirovecci</i> <i>Cryptococcus</i> spp. Intracellular bacteria (e.g. <i>Legionella</i> spp.) <i>M. tuberculosis</i> Viruses (<i>CMV</i> , <i>HSV</i> , <i>VZV</i>)
	B-cell abundance and function	Multiple myeloma Primary immunodeficiency	Rituxumab	Encapsulated bacteria: <i>S. pneumoniae</i> , <i>H. influenzae</i>

ity of neutropenic patients with infiltrates on thin-sliced CT scans had no detectable abnormality on chest radiograph [10]. High-resolution CT scan is often helpful for confirming the presence of infection, guiding site selection for bronchoalveolar lavage, and directing empiric therapy based on imaging characteristics. The presence of cavitation is associated with *Mycobacterium* spp., *Nocardia* spp., *Aspergillus* spp. and *P. jirovecci*; interstitial infiltrates suggest viral (e.g. Cytomegalovirus [CMV]) pneumonia or *Pneumocystis*; dense consolidation implies either bacterial pathogens or *Aspergillus* spp.. Serologic tests are of decreased utility in immunocompromised patients, especially in patients with impaired T-cell and B-cell immunity (Table 24.1), whereas antigen-based testing (e.g. *Streptococcus* and *Legionella* urinary antigens, *Cryptococcus* serum antigen testing) can be useful. An aggressive approach to sampling the lower respiratory tract (via bronchoscopy or miniature bronchoalveolar lavage [“mini-BAL”]) is warranted, as the spectrum of potential pathogens usually exceeds any reasonable empiric antimicrobial regimen. Depending on the patient’s degree and type of immunosuppression, lower respiratory tract specimens should be tested for gram stain and bacterial culture, fungal culture, acid fast stain and culture, respiratory viral PCR, CMV antigen, galactomannan,

Pneumocystis PCR. Recommended diagnostic tests by specimen site are listed in Table 24.2.

Empiric Treatment

Antimicrobial therapy should be given promptly in patients with suspected pneumonia. Unless lower respiratory tract specimens can be acquired immediately, therapy should not be delayed for the sake of increasing diagnostic yield. Empiric treatment of *Pneumocystis* does not compromise the yield of lower respiratory tract testing [1]. No single empiric regimen exists for immunocompromised pneumonia, given the diversity of immunocompromised conditions and associated infections (Table 24.1). A reasonable approach is to start with a regimen for community-acquired or healthcare-associated pneumonia as appropriate [11, 12], then expand according to the patient’s specific immune deficits and past microbiological data. This regimen should then be routinely reassessed for effectiveness based on the patient’s clinical response and the results of invasive microbiological testing. Empiric treatment of fungal pneumonia is rarely indicated for initial regimens but should be strongly considered in patients with clinical risk factors (e.g. prolonged neutropenia), consistent imaging (Fig. 24.3, a CT scan of a patient with aspergillosis) and lack of response to antibacterial therapy.

Table 24.2 Diagnostic testing in immunocompromised patients with suspected pneumonia

Specimen	Diagnostic tests
Bronchoalveolar lavage fluid	Cell count and differential
	Gram stain and bacterial culture
	Fungal stain and culture
	Acid-fast bacteria stain and culture
	Respiratory virus PCR
	<i>Pneumocystis jirovecci</i> PCR
	CMV antigen
	Galactomannan
Serum	Bacterial culture
	Fungal culture
	Acid-fast bacteria culture
	<i>Cryptococcus</i> antigen
	Galactomannan
	β -D-glucan
Urine	<i>Streptococcus</i> antigen
	<i>Legionella</i> antigen

Supportive Care

Unless otherwise contraindicated, immunocompromised patients with hypoxemic respiratory failure should be given a trial of noninvasive positive pressure ventilation (NIPPV) [13–15]. Corticosteroids are indicated for patients with HIV/AIDS and *P. jirovecci* pneumonia with room air PaO₂ under 70 or A-a gradient over 30 [16, 17], though data supporting their use in non-HIV patients with the same infection is weaker [18, 19]. Competing non-infectious diagnoses should be explored and potentially treated empirically (e.g. diuresis for infiltrates suggestive of cardiogenic edema).

Evidence Contour

Utility of Invasive Testing

Invasive sampling of the lower respiratory tract (by bronchoscopy with and without transbronchial biopsy, mini-BAL or open lung biopsy) is common in the diagnosis of pneumonia in immunocompromised patients, and wide practice variation

**Fig. 24.3** CT scan – aspergillosis

exists among modalities used. Among intubated patients, mini-BAL performs comparably to flexible bronchoscopy with lavage [20]. Transbronchial biopsy increases the yield of bronchoalveolar lavage, generally by distinguishing invasive fungal disease from colonization [21, 22]. Transbronchial biopsy is associated with elevated rates of pneumothorax when performed on mechanically ventilated patients (14–24%) [23, 24], though this risk must be weighed against those of alternative diagnostic maneuvers (e.g. open lung biopsy). BAL galactomannan has excellent sensitivity and specificity in the diagnosis of invasive aspergillosis [25], and it is undetermined what effect its adoption has had on the marginal yield of transbronchial biopsy. In one series of patients with hematologic malignancies and pulmonary infiltrates, open lung biopsy identified a diagnosis in 62% of cases and changed management in 57% of cases [26], though only 55% of these patients had previously undergone bronchoscopy and only 13% had undergone transbronchial biopsy.

Serum Indices of Infection

Serum tests for pneumonia in immunocompromised patients are an attractive arena for investigation, but no consensus exists regarding their utility, and in practice they rarely preclude invasive lung sampling. A serum galactomannan test is relatively specific (89%) for invasive aspergillosis

among immunocompromised patients but has poor sensitivity (71 %) [25]; a negative result does not exclude the diagnosis. By contrast, a commercially available beta-D-glucan assay is more sensitive than serum galactomannan but less specific [27]. A serum procalcitonin level below 0.5 ng/ml effectively excludes the presence of a bacterial infection in critically ill immunocompromised patients [28].

Noninvasive Ventilation

Though noninvasive positive pressure ventilation (NIPPV) is infrequently indicated for immunocompetent patients with pneumonia given the difficulty of managing secretions and the lack of rapid reversibility, two randomized controlled trials have demonstrated a clinical benefit to its use among immunocompromised patients. In a large (238 patient) study of patients immunosuppressed for solid organ transplantation with acute respiratory failure, patients who received NIPPV (as compared to standard treatment with supplemental oxygen) were less frequently intubated and experienced lower Intensive Care Unit (ICU) mortality [13]. In a second study of more broadly immunosuppressed patients with respiratory failure and clinical evidence of pneumonia, treatment with NIPPV resulted in less frequent endotracheal intubation and lower ICU mortality and overall mortality [15].

References

- O'Donnell WJ, Pieciak W, Chertow GM, Sanabria J, Lahive KC. Clearance of *Pneumocystis carinii* cysts in acute *P. carinii* pneumonia: assessment by serial sputum induction. *Chest*. 1998;114(5):1264–8.
- Sickles EA, Greene WH, Wiernik PH. Clinical presentation of infection in granulocytopenic patients. *Arch Intern Med*. 1975;135(5):715–9.
- Dickson RP, Erb-Downward JR, Huffnagle GB. Towards an ecology of the lung: new conceptual models of pulmonary microbiology and pneumonia pathogenesis. *Lancet Respir Med*. 2014;2(3):238–46. Pubmed Central PMCID: 4004084.
- Dickson RP, Martinez FJ, Huffnagle GB. The role of the microbiome in exacerbations of chronic lung diseases. *Lancet*. 2014;384(9944):691–702. Pubmed Central PMCID: 4166502.
- Venkataraman A, Bassis CM, Beck JM, Young VB, Curtis JL, Huffnagle GB, et al. Application of a neutral community model to assess structuring of the human lung microbiome. *MBio*. 2015;6(1). Pubmed Central PMCID: PMC4324308.
- Iwai S, Fei M, Huang D, Fong S, Subramanian A, Grieco K, et al. Oral and airway microbiota in HIV-infected pneumonia patients. *J Clin Microbiol*. 2012;50(9):2995–3002. Pubmed Central PMCID: PMC3421777.
- Diaz PI, Hong BY, Frias-Lopez J, Dupuy AK, Angeloni M, Abusleme L, et al. Transplantation-associated long-term immunosuppression promotes oral colonization by potentially opportunistic pathogens without impacting other members of the salivary bacteriome. *Clin Vaccine Immunol*. 2013;20(6):920–30. Pubmed Central PMCID: PMC3675961.
- Wright HL, Moots RJ, Bucknall RC, Edwards SW. Neutrophil function in inflammation and inflammatory diseases. *Rheumatology (Oxford)*. 2010;49(9):1618–31.
- Camps Serra M, Cervera C, Pumarola T, Moreno A, Perello R, Torres A, et al. Virological diagnosis in community-acquired pneumonia in immunocompromised patients. *Eur Respir J*. 2008;31(3):618–24.
- Heussel CP, Kauczor HU, Heussel G, Fischer B, Mildenerger P, Thelen M. Early detection of pneumonia in febrile neutropenic patients: use of thin-section CT. *AJR Am J Roentgenol*. 1997;169(5):1347–53.
- Guidelines for the management of adults with hospital-acquired, ventilator-associated, and healthcare-associated pneumonia. *Am J Respir Crit Care Med*. 2005;171(4):388–416.
- Mandell LA, Wunderink RG, Anzueto A, Bartlett JG, Campbell GD, Dean NC, et al. Infectious Diseases Society of America/American Thoracic Society consensus guidelines on the management of community-acquired pneumonia in adults. *Clin Infect Dis*. 2007;44 Suppl 2:S27–72.
- Antonelli M, Conti G, Bufi M, Costa MG, Lappa A, Rocco M, et al. Noninvasive ventilation for treatment of acute respiratory failure in patients undergoing solid organ transplantation: a randomized trial. *JAMA*. 2000;283(2):235–41.
- Girou E, Schortgen F, Delclaux C, Brun-Buisson C, Blot F, Lefort Y, et al. Association of noninvasive ventilation with nosocomial infections and survival in critically ill patients. *JAMA*. 2000;284(18):2361–7.
- Hilbert G, Gruson D, Vargas F, Valentino R, Gbikpi-Benissan G, Dupon M, et al. Noninvasive ventilation in immunosuppressed patients with pulmonary infiltrates, fever, and acute respiratory failure. *N Engl J Med*. 2001;344(7):481–7.
- Consensus statement on the use of corticosteroids as adjunctive therapy for pneumocystis pneumonia in the acquired immunodeficiency syndrome. The National Institutes of Health-University of California Expert Panel for Corticosteroids as Adjunctive Therapy for *Pneumocystis Pneumonia*. *N Engl J Med*. 1990;323(21):1500–4.
- Briel M, Bucher HC, Boscacci R, Furrer H. Adjunctive corticosteroids for *Pneumocystis jirovecii* pneumonia

- in patients with HIV-infection. *Cochrane Database Syst Rev.* 2006;3, CD006150.
18. Delclaux C, Zahar JR, Amraoui G, Leleu G, Lebargy F, Brochard L, et al. Corticosteroids as adjunctive therapy for severe *Pneumocystis carinii* pneumonia in non-human immunodeficiency virus-infected patients: retrospective study of 31 patients. *Clin Infect Dis.* 1999;29(3):670–2.
 19. Pareja JG, Garland R, Koziel H. Use of adjunctive corticosteroids in severe adult non-HIV *Pneumocystis carinii* pneumonia. *Chest.* 1998;113(5):1215–24.
 20. Tasbakan MS, Gurgun A, Basoglu OK, Ekren PK, Pullukcu H, Bacakoglu F. Comparison of bronchoalveolar lavage and mini-bronchoalveolar lavage in the diagnosis of pneumonia in immunocompromised patients. *Respiration.* 2011;81(3):229–35.
 21. Cazzadori A, Di Perri G, Todeschini G, Luzzati R, Boschiero L, Perona G, et al. Transbronchial biopsy in the diagnosis of pulmonary infiltrates in immunocompromised patients. *Chest.* 1995;107(1):101–6.
 22. Jain P, Sandur S, Meli Y, Arroliga AC, Stoller JK, Mehta AC. Role of flexible bronchoscopy in immunocompromised patients with lung infiltrates. *Chest.* 2004;125(2):712–22.
 23. Bulpa PA, Dive AM, Mertens L, Delos MA, Jamart J, Evrard PA, et al. Combined bronchoalveolar lavage and transbronchial lung biopsy: safety and yield in ventilated patients. *Eur Respir J.* 2003;21(3):489–94.
 24. O'Brien JD, Ettinger NA, Shevlin D, Kollef MH. Safety and yield of transbronchial biopsy in mechanically ventilated patients. *Crit Care Med.* 1997;25(3):440–6.
 25. Guo YL, Chen YQ, Wang K, Qin SM, Wu C, Kong JL. Accuracy of BAL galactomannan in diagnosing invasive aspergillosis: a bivariate metaanalysis and systematic review. *Chest.* 2010;138(4):817–24.
 26. White DA, Wong PW, Downey R. The utility of open lung biopsy in patients with hematologic malignancies. *Am J Respir Crit Care Med.* 2000;161(3 Pt 1):723–9.
 27. Sulahian A, Porcher R, Bergeron A, Touratier S, Raffoux E, Menotti J, et al. Use and limits of (1–3)-beta-d-glucan assay (Fungitell), compared to galactomannan determination (*Platelia Aspergillus*), for diagnosis of invasive aspergillosis. *J Clin Microbiol.* 2014;52(7):2328–33. Pubmed Central PMCID: PMC4097729.
 28. Bele N, Darmon M, Coquet I, Feugeas JP, Legriel S, Adaoui N, et al. Diagnostic accuracy of procalcitonin in critically ill immunocompromised patients. *BMC Infect Dis.* 2011;11:224. Pubmed Central PMCID: PMC3170614.