# The Crystalline State of Archaeological Bone Material

Wolfgang W. Schmahl, Balazs Kocsis, Anita Toncala, Dominika Wycisk, and Gisela Grupe

#### Abstract

Isotope studies on archaeological bone mineral require a validation of the investigated sample material. Diagenetic alteration or contaminated bone mineral should be recognized as such and not be used for conclusions requiring pristine material. X-ray diffraction (XRD) and Fourier Transform Infrared Spectroscopy (FTIR) provide two complementary tools to characterize the state of the bone mineral. While IR measurements are easy and rapid, their interpretation is still largely empirical. Modern XRD analysis is more demanding with respect to experiment and data evaluation, but it is based on rigorous theoretical modelling of the observed data. Our study involved both uncremated animal bone samples from the alpine region covering ages from 7600 to 550 years before present, as well as cremated human bone remains in comparison with experimentally cremated bovine bone. All samples were mechanically cleaned to remove soil, and inner and outer periosteal surfaces were mechanically removed. We avoided visually decomposed bones completely. The mineralogic state of the thus cleaned, uncremated samples showed only minor systematic changes with archaeologic age. The changes are most pronounced for the lattice parameter and crystalline domain size in the short dimension of the original bone-apatite platelets. The long direction corresponding to the crystallographic c-axis of the apatite appears almost unaffected. We conclude that in the investigated samples, there is only a minor diagenetic alteration of the original bone mineral, possibly related to exchange of carbonate by hydroxyl or fluorine.

W.W. Schmahl  $(\boxtimes) \cdot B$ . Kocsis

Sektion Kristallographie, Ludwig-Maximilians-Universität, Munich, Germany e-mail: [Wolfgang.W.Schmahl@lrz.uni-muenchen.de](mailto:Wolfgang.W.Schmahl@lrz.uni-muenchen.de)

A. Toncala • D. Wycisk • G. Grupe

Biozentrum, Ludwig-Maximilians-Universität, Martinsried, Germany

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From annealing experiments with bovine femur bone material at different temperatures for 1 h annealing time, we established calibration curves to be used to estimate cremation temperatures of bones based on FTIR spectra and X-ray diffractograms. While the evaluation of the diffractograms is rigorously based on a physical model, the evaluation of spectral components in FTIR spectra is empirical. The experiments indicate that original bone apatite contains little–if any– $OH^-$  such that it is carbonate-H<sub>2</sub>O-apatite (rather than hydroxyapatite), and with increasing temperature, water and carbonate leave the material, and from  $600 \degree C$ , hydroxyapatite is formed with increasing purity and crystallite size with increasing temperature. The analysis of some cremated bones from Urnfield Culture sites of Eching and Zuchering, southern Bavaria, clearly indicated that the archaeological cremated bones are inhomogeneous materials where different parts of the samples were subjected to different cremation temperatures and/or times at temperature within fractions of centimetres. This can be attributed to the conditions in the pile of burning logs and burning tissue. Nevertheless, a fair estimation of cremation temperatures is certainly possible even where the FTIR approach and the XRD approach still do not have a full mutual consistency. Future work needs to address the anatomical variability of original bone material within and between species in much more detail than is known at present. Beyond the well-defined temperature/time conditions in furnace annealing experiments, cremation experiments with bone material analyses must also be done in more realistic conditions as they occur in a funeral pyre.

## Introduction

The creation of a map of isotope signatures of archaeological vertebrate bone finds excavated along the Inn-Eisack-Adige passage via the Brenner Pass in the European Alps has been at the core of the initial period of the DFG Forschergruppe FOR1670 (see Introduction). This map is intended to function as a basis to reconstruct ancient migration, trade and culture transfer across this region by geochemical analysis of human bone remains. Certainly, any reliable interpretation of isotopic or microchemical signatures of archaeologic bone samples relies on the assumptions that the analysed material is the uncontaminated original material and that this material has been chemically preserved as it was during the life of the individual. The bioapatite bone mineral, however, represents a biogenic tissue component with a specific chemistry and nanostructure that is quite different from that of nominally similar inorganic apatite minerals that are in equilibrium with the ambient conditions where the bone is buried. This, aside from the trivial possibility of contamination with soil particles and the effects of the decomposition of the organic component of the bone, makes bioapatite susceptible to chemical and structural alterations occurring after the death of the individual as the material reacts towards equilibrium with the environment. To take precautions against these possible sources of error in isotope data, FOR1670 decided to screen the

archaeologic bone finds by mineralogic analysis based on X-ray diffraction and infrared spectroscopy. In a time period of about 1500 years in the investigated area, from the Bronze Age until Imperial Roman Times, i.e. 60 generations, cremating the deceased was the dominant burial custom (Grupe et al. [2015\)](#page-27-0). Thus, we performed some initial analyses on cremated bones to prepare for a deeper study in the second funding period of FOR1670.

#### The Bone Mineral

Bone is a composite material of nearly equal volumes of collagen and nanometre scale crystallites of a carbonated apatite mineral (Peters et al. [2000](#page-27-0)). The geologic mineral hydroxyapatite is rarely found, as the geologic apatite tends to incorporate  $F^-$  and  $Cl^-$  ions instead of  $OH^-$  to give an approximate formula  $Ca_{10}(PO_4)_6(F_{1-x-y}, Cl_x, OH_y)_2$ . Bone has a complex hierarchical structure (Wegst et al. [2015](#page-29-0); Reznikov et al. [2014;](#page-28-0) Schwarcz et al. [2014](#page-28-0); Weiner et al. [1999;](#page-29-0) Rho et al. [1998\)](#page-28-0) where the platelike nanocrystals of  $\sim$  2  $\times$  30  $\times$  40 nm size (Cuisinier et al. [1987](#page-26-0); Alexander et al. [2012\)](#page-26-0) provide stiffness and compressive strength to the hybrid composite.

The formation of the apatite mineral phase is a complex, biologically controlled process, where the mechanisms of nucleation, growth and growth limitation of the small crystallites are not yet clear. There are several models, including other calcium phosphate phases as precursors and growth from an amorphous phase (Landis and Jacquet [2013](#page-27-0)).

Chemically, the apatite mineral component of the mammal bone can be approximated as

$$
\left[Ca_{1-u-v}Na_{u}\right]_{10}\left[\left(PO_{4}\right)_{1-x-y}(CO_{3})_{x}(HPO_{4})_{y}\right]_{6}\left[OH_{1-z-w},\left(CO_{3}\right)_{z},\left(H_{2}O\right)_{w}\right]_{2}
$$

where the small potential vacancy concentration  $v$  on Ca-sites is

 $v = 3(x + y) - z + w - \frac{1}{2}u$ ; (x, y, z, u, v,  $w \ge 0$ ).

Carbonate (5–8 wt.%) is an essential component of the bone mineral, mostly substituting for  $[PO_4]^{3-}$ , but also for  $[OH]^{-}$  to a much lesser degree (LeGeros et al. [1967,](#page-27-0) [1968;](#page-27-0) Rey et al. [1989](#page-28-0), [1990](#page-28-0), [1995;](#page-28-0) Yi et al. [2013](#page-29-0)). The two different variants of  $[CO<sub>3</sub>]<sup>2</sup>$  substitution in the crystal structure (type A  $[CO<sub>3</sub>]<sup>2</sup>$   $\leftrightarrow$   $[PO<sub>4</sub>]<sup>3</sup>$ , type B  $[CO<sub>3</sub>]<sup>2-</sup> \leftrightarrow [OH]^-$ ) (Wopenka and Pasteris [2005](#page-29-0)) can be distinguished by IR spectroscopy (Balan et al. [2011](#page-26-0)). The presence of the acidic phosphate species  $[HPO<sub>4</sub>]$ <sup>2-</sup> is also important in the biomineral (Wang et al. [2013](#page-29-0); Rey et al. [2014\)](#page-28-0). For bone apatite, the presence of  $OH^-$  is low (Pasteris et al. [2004\)](#page-27-0), and water may reside in the OH position. A multitude of different schemes of coupled exchange to maintain charge balance can be suggested (e.g. Wopenka and Pasteris [2005](#page-29-0); de Leeuw [2010](#page-26-0)). In mammal bioapatite, small amounts of  $F^-$  replace OH<sup>-</sup> (notably in tooth enamel), and on the  $Ca^{2+}$  cation position voids (Wilson et al. [2005](#page-29-0)), Na<sup>+</sup> and  $Mg^{2+}$  can be substituted as well as traces of  $Sr^{2+}$  and  $Ba^{2+}$  (e.g. Elliot [2002;](#page-26-0) TurnerWalker [2008\)](#page-29-0). Sponheimer et al. ([2005\)](#page-29-0) report that herbivores accumulate higher levels of  $Sr^{2+}$  und  $Ba^{2+}$  than carnivores and omnivores.

Nuclear magnetic resonance investigations of Jäger et al. ([2006\)](#page-27-0) and Wang et al. [\(2013](#page-29-0)) indicate the existence of an amorphous surface layer of water-bearing carbonated Ca-phosphate (amorphous calcium carbonate, ACC) which coats a bioapatite nano-crystallite core. Together with the collagen and other organic/ cellular components, the mineral nanoparticles compose, right from the supramolecular scale, a hierarchical hybrid composite (Wegst et al. [2015;](#page-29-0) Reznikov et al. [2014;](#page-28-0) Schwarcz et al. [2014;](#page-28-0) Weiner et al. [1999;](#page-29-0) Rho et al. [1998\)](#page-28-0). As long as this compound structure remains intact, both the inorganic and the organic components are stabilized against chemical and biochemical attack (Gernaey et al. [2001;](#page-26-0) Turner-Walker [2008\)](#page-29-0). The taphonomic (biological postmortem) decomposition of the about 30 wt.-% organic components of the bone (Peters et al. [2000\)](#page-27-0) creates a high porosity and internal surface in the remaining mineral. This internal surface is reactive and promotes adsorption of ions and molecules from solutions in the soil. The triangular  $[CO<sub>3</sub>]<sup>2</sup>$  groups which are substituted into the crystal structure for the tetrahedral  $[PO<sub>4</sub>]$ <sup>3-</sup> complex or for the essentially spherical  $OH^-$  induce structural disorder related to the geometric mismatch and charge compensation mechanism. Both this structural misfit and the high surface/interface area increase the free energy of the bioapatite, and thus it has an increased solubility compared to inorganic hydroxyapatite, which is fairly insoluble (Hedges [2002](#page-27-0); Berna et al. [2004;](#page-26-0) Wopenka and Pasteris [2005;](#page-29-0) de Leeuw [2010\)](#page-26-0). In vivo this enhanced solubility plays a substantial role in the continual physiologic reconstruction of bone. Similarly, the increased free energy provides a driving force for diagenetic recrystallization (Nielsen-Marsh and Hedges [2000](#page-27-0); Nielsen-Marsh et al. [2000](#page-27-0)) and formation of larger crystals. Thus, secondary minerals may form such as newly grown abiotic inorganic hydroxyapatite (which is free of carbonate) and calcite, CaCO<sub>3</sub>, taking up the  $[CO<sub>3</sub>]<sup>2</sup>$  and associated Ca<sup>2+</sup> from the former bioapatite.

$$
[Ca_{1-u-v}Na_u]_{10} [(PO_4)_{1-x-y}(CO_3)_x(HPO_4)_y]_6 [OH_{1-z-w}, (CO_3)_z, (H_2O)_w]_2
$$

Bioapatite (Carbonate-Hydro-Hydroxyapatite)

$$
\rightarrow \sim s Ca_{10}(PO_4)_6(OH)_2 + (6x + 2z - 6y) CaCO_3 + 10u NaCaPO_4 + 2w H_2O + 6y CO_2
$$
  
Hydroxyapatite Calculate Calculate

with  $s \approx (1 - 2u - v - 0.6x - 0.2z + 0.6y)$ .

The NaCaPO<sub>4</sub> phase here is a nominal phase to account for the sodium. Those secondary minerals potentially incorporate ions present in the soil solution, such as  $Sr^{2+}$ , Pb<sup>2+</sup>, etc. (Tütken et al. [2008\)](#page-29-0), which changes the trace elements signatures of the sample.

While the bone mineral is comparatively stable against exposures at temperatures reached by cooking, boiling, or roasting, conditions above ca. 500  $\degree$ C profoundly change the bulk chemistry and crystalline consistence of the bioapatite (Rogers and Daniels [2002](#page-28-0); Enzo et al. [2007;](#page-26-0) Munro et al. [2008](#page-27-0); Piga et al. [2008](#page-28-0), [2009a](#page-28-0), [b](#page-28-0), [2013;](#page-28-0)

Harbeck et al. [2011;](#page-27-0) Galeano and Garcia-Lorenzo  $2014$ ). From about 500 °C, a significant recrystallization of the bioapatite and crystallite growth of hydroxyapatite sets in. Prehistoric cremation pyres could reach temperatures up to  $1000^{\circ}$ C (McKinley [2016](#page-27-0)). Ratios of stable isotopes of light elements contained in the atmosphere change considerably during heat treatment (Olsen et al. [2008;](#page-27-0) Schurr et al. [2008](#page-28-0)), while those of elements such as strontium and lead do not appear to be altered as long as there are no reactions with surrounding materials (Harbeck et al. [2011\)](#page-27-0).

The selected techniques for mineralogical characterization of the bone materials, X-ray diffraction (XRD) and Fourier Transform Infrared Spectroscopy (FT IR), are essentially complementary. To characterize the diagenetic recrystallization of bone apatite, Person et al. ([1995](#page-27-0)) defined their classical empirical "crystallinity index" (CI) on the basis of the width of X-ray diffraction lines. This index number became widely used (e.g. Hedges [2002](#page-27-0)). Another popular empirical index number is the infrared splitting factor (IRSF) by Weiner et al. ([1993](#page-29-0)) (see, e.g. Surovell and Stiner [2001](#page-29-0)). The utility of the simple IRSF to describe complex spectral features is debated in the literature (Sillen [1989;](#page-29-0) Shemesh [1990](#page-28-0); Weiner and Bar-Yosef [1990;](#page-29-0) Wright and Schwarcz [1996](#page-29-0); Trueman et al. [2008](#page-29-0); Tütken et al. [2008](#page-29-0)). According to Shinomiya et al. [\(1998\)](#page-28-0), the IRSF indicated chemical alterations already a few years after death. Sample preparation for XRD or FTIR measurement certainly has an influence on the CI/IRSF results (Surovell and Stiner [2001\)](#page-29-0). The simplifying indices for a "crystallinity" of the bone mineral do not correlate with modifications of chemical composition and also not with stable isotope data of the structural carbonate (e.g. Lee-Thorp and Sponheimer [2003;](#page-27-0) Pucéat et al. [2004;](#page-28-0) Trueman et al. [2008;](#page-29-0) Tütken et al. [2008](#page-29-0)). Moreover, physiological differences in bioapatite characteristics between different species, different skeletal elements and different age groups of the population can be expected and need to be considered in the future (Rey et al. [1991](#page-28-0), [2009](#page-28-0); Yerramshetty and Akkus [2008\)](#page-29-0). To put the IRSF on a more physical basis, Lebon et al. ([2008](#page-27-0)) suggested spectral decomposition by curve-fitting procedures on the relevant part of the infrared spectrum; these procedures were later improved by Vandecandelaere et al. ([2012](#page-29-0)) and Grunenwald et al. ([2014](#page-27-0)). Pedone et al. [\(2007\)](#page-27-0) and Yi et al. [\(2014\)](#page-29-0) contributed more rigorous considerations on the theory of phosphate and carbonate vibrations to be expected in bioapatite.

The X-ray diffraction (XRD) technique takes advantage of the periodic latticelike structure of crystalline materials, as it measures interference patterns of the X-ray waves scattered from the atoms or molecules in the material. Crystals are objects with a spatially periodic arrangement of the constituting atoms, ions or molecules, and for such materials, the interference maxima become sharp if the spatial range of the periodic order is wide. The concept of a crystal is an array of building blocks (called unit cells, Fig. [1\)](#page-5-0) which are periodically repeated in a spacefilling, lattice-like fashion. The positions of the X-ray diffraction peaks are defined by the unit-cell dimensions, the so-called lattice parameters. The peak intensities, explained in a nutshell, are related to the electron density distribution pattern in the unit cell averaged over time and space in the crystal. This pattern reflects the positions and electron densities of all the individual atoms or molecules in the crystal, superimposed by the space and time average (cf. for details e.g. to

<span id="page-5-0"></span>

Fig. 1 Crystal structure of hydroxyapatite in hexagonal setting after Wilson et al. [\(1999](#page-29-0)) and Tonegawa et al. [\(2010\)](#page-29-0). Some authors (Ikoma et al. [1999](#page-27-0); Tonegawa et al. [2010\)](#page-29-0) believe that monoclinic symmetry (space group P2<sub>1</sub>/c) gives a better description for well-crystalline hydroxyapatite. In bioapatite, the  $[PO_4]^{3-}$  is exchanged by  $[CO_3]^{2-}$  and  $[HPO_4]^{2-}$ , while the channel sites (OH<sup>-</sup> location of hydroxyapatite) contain  $[CO<sub>3</sub>]<sup>2</sup>$ , H<sub>2</sub>O, OH<sup>-</sup> and other ions. The Ca-sites are also occupied by minor amounts of  $Na<sup>+</sup>$  and by vacancies (white square). It is here where trace elements such as  $Sr^{2+}$  and  $Pb^{2+}$  reside

Pecharsky and Zavalij [2003](#page-27-0) and Mittemeijer and Welzel [2012\)](#page-27-0). For small crystallites, as is the case for bioapatite, there are fewer atoms contributing to the interference phenomenon, and thus the diffraction maxima widen inversely proportional to the coherence length of the periodic crystalline order (de Keijser et al. [1982\)](#page-26-0). The width and shape of the X-ray diffraction peak related to a particular lattice plane depend both on this coherence length (i.e. in the simplest case the crystallite size) perpendicular to that plane and on the variance of the lattice parameters in the sample, the so-called microstrain. Microstrain results from inhomogeneities in chemical composition, other lattice defects, and from mechanical strains on the scale of the crystallite size. In the case of bone, such mechanical strains can result from the composite structure with collagen. Note that the frequently used term "crystallinity" (Rogers et al. [2010](#page-28-0)) is a notion mingling effects of crystallite size, microstrains, random chemical substitution and presence of amorphous phases in a rather undefined way.

Deviations from the average structure give a very small intensity contribution in XRD, which we neglect here. Amorphous substances do not have a periodic structure; their diffraction pattern consists of diffuse intensity halos with maxima related to peaks in the spatial probability density distribution of the constituting atoms. In any case, amorphous, crystalline or "in between", the diffraction signal is the Fourier transform of the electron density self correlation function, which is straightforward to calculate (see Schmahl et al. [2016](#page-28-0) for more information). Thus,

modern XRD analysis is based on a rigorous calculation of the diffraction profile and algorithms fitting the model parameters to obtain the best-possible agreement with the observed diffraction profile (so-called Rietveld refinement method, Rietveld [1969;](#page-28-0) Rodriguez-Carvajal [1993;](#page-28-0) Rodriguez-Carvajal and Roisnel [2004;](#page-28-0) Pecharsky and Zavalij [2003](#page-27-0); Mittemeijer and Welzel [2012](#page-27-0)).

This quantitative treatment of the X-ray diffraction profile led to assessments of the crystallographic characteristics of archaeological and paleontological bone materials (Stathopoulou et al. [2008](#page-29-0); Piga et al. [2008,](#page-28-0) [2009a,](#page-28-0) [b,](#page-28-0) [2013](#page-28-0); Harbeck et al. [2011\)](#page-27-0). Stathopoulou et al. ([2008](#page-29-0)) examined Miocene and Pleistocene samples from the Aegean with the Rietveld technique and arrived at the conclusion that "Diagenetic trends, common to all these sites include a subtle but systematic decrease of the unitcell volume and a-axis of carbonate hydroxylapatite, as well as a parallel increase of the coherence length along the c-axis". Stathopoulou et al. ([2008](#page-29-0)) also established a correlation of X-ray diffraction data with IR data. Enzo et al. [\(2007\)](#page-26-0), Piga et al. [\(2008](#page-28-0), [2009a,](#page-28-0) [b](#page-28-0), [2013](#page-28-0)), Harbeck et al. [\(2011](#page-27-0)), and Galeano and Garcia-Lorenzo ([2014](#page-26-0)) applied modern full-profile analysis of X-ray diffractograms to evaluate effects of cremation on the bone mineral. From about 500  $\degree$ C, a significant recrystallization of the bioapatite and crystallite growth of hydroxyapatite sets in. Piga et al. ([2008](#page-28-0), [2009a](#page-28-0)) experimented with varying cremation times at different temperatures and found that between 600  $\degree$ C and 850  $\degree$ C, the transformation of the bioapatite to hydroxyapatite occurs essentially within the first 20 min of the heat treatment.

X-ray diffraction is less well suited to probe local molecular environments. Infrared spectroscopy, however, can distinguish, e.g. the two different variants of  $[CO<sub>3</sub>]<sup>2-</sup>$  substitution in the crystal structure (type A  $[CO<sub>3</sub>]<sup>2-</sup> \leftrightarrow [PO<sub>4</sub>]<sup>3-</sup>$ , type B  $[CO<sub>3</sub>]<sup>2-</sup> \leftrightarrow [OH]^-$ ) (Wopenka and Pasteris [2005](#page-29-0)). Infrared spectroscopy (IR) probes the frequencies of molecular scale vibrations in the material. Infrared photons are absorbed if their frequency is in resonance with a frequency of a molecular scale vibration in the sample. The frequency characteristics of the vibration depend on the precise local structural configuration of the vibrating entity in the sample. In a crystallographically well-ordered material, the vibrations in neighbouring unit cells are identical. In this case, the IR absorption peaks are well developed and rather sharp. In a disordered or amorphous material, the IR absorption signal is still present, but it broadens as the molecular vibration frequencies become locally different due to different molecular environments in the structure. This interaction is complex, however, as the frequencies shift both with changing chemical bond strengths and masses of vibrating bodies in different configurations or environments, but the signal also broadens in relation to the fading (damping) of the vibration after its initial excitation.

In any case, the IR spectrum reflects the time structure of local molecular scale environments of the vibrating entities in the sample, while the X-ray diffraction signal reflects the spatial periodicity in the atomistic structure. Accordingly, the two methods give complementary information. Atoms and molecules vibrate around their equilibrium configuration in all materials, regardless if the materials structure is crystallographically ordered or not. This gives an advantage to IR spectroscopy for the investigation of materials which are not spatially ordered. On the other hand, IR probes periodic long-range order ("crystallinity") only indirectly: by the effect of the crystal structure on the time structure of the vibrations. So far, there is no straightforward general way to calculate this indirect influence.

## Materials and Methods

Among the uncremated archaeological animal bone samples which were prepared for isotope studies in FOR1670 (see chapter "Isotopic Map of the Inn-Eisack-Adige-Brenner Passage and its Application to Prehistoric Human Cremations"), we selected 63 random samples for screening by X-ray diffraction. The samples were from the species Bos taurus, Sus domesticus and Cervus elaphus. Fresh bovine bone samples and fossil samples of a Miocene hippopotamus from Bavaria were obtained for comparison. A piece of compact bone was cut from the skeletal element, and the endosteal and periosteal surfaces were mechanically removed by grinding off. The sample was then ultrasonically washed in deionized  $H<sub>2</sub>O$  (35 kHz), where the water was changed every 5 min. The washing was repeated until the water remained clear. After drying in air, the bone piece was defatted for 5 h with diethylether in a Soxhlet and air dried again. Finally, the sample was homogenized to a fine powder.

For experimental cremation, we used thus prepared bovine femur powder and subjected it to heat treatment in air in a muffle. The samples were put into the preheated furnace for an exposure time of 1 h, after which they were removed and left to cool in air.

For an investigation archaeological bone samples from burials which were subjected to cremation as a burial rite, we selected long bones from the Bronze Age (Urnfield Culture) site of Eching (1300–800 BC) and a skull from the Zuchering site of the same culture, both in in southern Bavaria. All cremated bones were buried in urns; the urns were found in a broken state.

X-ray diffractograms were collected on a General Electric 3003 powder diffractometer in Bragg-Brentano reflection geometry. Cu-K $\alpha_1$  radiation was selected with a focussing monochromator in the primary beam. Data collection time for a complete diffractogram from 5 to 100 degrees  $2\theta$  amounted to 240 min with a 1D Meteor detector and exposure time of 1000 s for each frame. The instrumental resolution function was experimentally determined with a NIST  $LaB<sub>6</sub>$  standard. For data evaluation and Rietveld refinement, the FULLPROF code (Rodriguez-Carvajal [1993;](#page-28-0) Rodriguez-Carvajal and Roisnel [2004\)](#page-28-0) was employed. The Thompson-Cox-Hastings method for convolution of sample effects such as anisotropic size and isotropic microstrain broadening (Thompson et al. [1987\)](#page-29-0) with instrumental resolution (as determined with a NIST  $LaB<sub>6</sub>$  standard) was applied.

Infrared spectra were measured on a Bruker Equinox FTIR instrument with a resolution of 4 cm<sup>-1</sup>. A total of 128 scans was averaged resulting in a 2 min acquisition time per sample. The bone powder (as described above for XRD) was sieved through a 100 μm mesh, and 1 mg of resulting sample powder was manually mixed with 200 mg of KBr in a mortar and pelletized subsequently. Spectral decomposition into seven bands was performed in the range  $400-800$  cm<sup>-1</sup>. The seven bands were fitted with a Gaussian-Lorentzian sum function with fixed ratio of

3:7 each. For each band, the center position, the full width at half maximum (FWHM) and the peak area were fitted. Background was described with a cubic polynomial, but the function remained essentially flat.

## Results and Discussion

#### The Uncremated Mammal Bones

Due to the nanoscale dimension of the bone bioapatite particles, the diffraction peaks of original bone material are significantly broadened (Fig. 2). The broadening is anisotropic, as the (002) peak, corresponding to the crystallographic c-axis of the crystallites, is comparatively sharp while the peaks corresponding to the a- and b-directions are significantly broader. We parameterized the size as a second rank tensor, corresponding to an ellipsoid. The agreement between observed and calculated diffraction profile is excellent in view of the complexity of the system. Due to



Fig. 2 X-ray diffraction trace of the bone mineral from the femur of modern domestic cattle (Bos taurus) taken with Cu-k $\alpha_1$  radiation. Red dots: observed data points; black line: calculated XRD profile; blue line on bottom: difference of observed and calculated data. Blue vertical bars: positions of the diffraction peaks. The two enlarged regions show the most featured parts of the diffractogram. The XRD profile was fitted with an anisotropic size model for the coherently diffracting domain

the hexagonal symmetry, the XRD peaks which would have been able to discriminate different crystallite sizes in a- and b-directions directly overlap, so that a distinction is not possible with the current method. Correspondingly, quoted crystallite sizes correspond to an average of the directions in the a-b plane. For the fresh bovine bone, we arrive at 12.6 nm for the c-direction and 3.9 nm for the a-b plane. The estimated precision of these numbers from the Rietveld least-squares fit is in the order of 5 %. Most of the archaeological samples show a slightly reduced broadening such that the 211, 121, 112 and 030 peaks become more distinct (Fig. 3). This may indicate grain growth and/or dissolution of the finest-scaled fraction during burial. Most importantly, we observe calcite as a secondary phase in about 20 % of the investigated archaeological sample. Note that all investigated samples had visually very good quality to be selected for isotope analysis, and their surfaces were cleaned away to avoid contamination by soil. There is no indication of typical soil minerals such as clay minerals or quartz, other than calcite, such that the selection and cleaning procedure was effective. Only in 3 samples out of 63, the determined calcite fraction was  $\geq 2$  wt.%, with the maximum at 5 wt%. The



Fig. 3 X-ray diffraction trace of an archaeologic animal bone sample (Bos taurus, radius) from the Brixen Stufels, Hotel Dominik site. Cu-k $\alpha_1$  radiation. Red dots: observed data points; black line: calculated XRD profile; blue line on bottom: difference of observed and calculated data. Green vertical bars: positions of the diffraction peaks; top row: bone apatite; bottom row: calcite. The two enlarged regions show the most featured parts of the diffractogram. The XRD profile was fitted with an anisotropic size model for the coherently diffracting domain. In the enlarged region the different broadening of the 002 and the 030 peaks can be clearly seen (c.f. Fig. [5](#page-10-0))

<span id="page-10-0"></span>observed calcite itself shows a diffraction line broadening which indicates a crystallite size on the scale of 100 nm, which is a strong indicator that this calcite is not simply due to crystallization from infiltrating soil solution, for which micrometre-sized crystals would be expected. We examined an archaeological deer bone (metatarsus) in the SEM after careful deproteination (Fig. 4). The calcite is contained inside the bone fabric in the form of ideal rhombohedral crystals of the 100–500 nm size range.



Fig. 4 SEM micrograph of deproteinated archaeological deer bone (metatarsus) from the Brixen-Stufels "Hotel Dominik"  $4856$  archaeological site ( $\sim$  5th century BC). Note the ideal rhombohedral crystals of secondary calcite (arrows) distributed in the bone fabric



Fig. 5 Comparison of the  $25^{\circ}42^{\circ}$  2 $\theta$  range of X-ray diffractograms of fresh bone (Bos taurus, femur) and two archaeological bones (Sus domesticus, mandibula, Cervus elaphus, metatarsus III  $+IV$ ). Cu-k $\alpha_1$  radiation. *Blue dots*: observed data points; *red line*: calculated XRD profile; *black* line on bottom: difference of observed and calculated data. crosses: positions of the diffraction peaks; top row: bone apatite; bottom row: calcite



Fig. 6 Overview on crystallographic parameters of the uncremated archaeological samples of animal bones. (a) Unit-cell volume vs. archaeological (cultural) age group. The bubble size indicates the size along c of the coherently diffracting domain. There is only a weak correlation with archaeological age. (b) Lattice constants (unit-cell axes) vs. unit-cell volume. Red dots and blue triangles: unit cell c-axis and a-axis, respectively, of the archaeological bones. Yellow stars: modern bovine femur. Pink circles: Miocene (~20 million year old) fossil hippopotamus from Bavaria. Light blue circles: hydroxyapatite. Orange circles: fluorapatite

A closer comparison of the  $25^{\circ} - 42^{\circ}$  diffraction angle (2 $\theta$ ) range is shown in Fig. [5.](#page-10-0) There is a small but detectable sharpening of the XRD peaks of the archaeological material compared to the fresh bovine femur.

Figure 6 gives an overview on crystallographic parameters of the 63 examined archaeological samples. Figure 6a compares the apatite unit-cell volume and the crystallite size along the c-axis with the archaeological age group. We observe only a vague correlation of unit-cell volume decreasing with archaeological age, and the crystallite size shows no correlation with age. Clearly, this indicates that there are other factors contributing to the measured lattice parameter than just changes with age, related to possible loss of carbonate. These other factors may include humidity and soil pH at the archaeological site, which might affect the alteration reaction, and also intraspecies and across-species variations of the crystallographic parameters of the bone mineral, which still need to be examined in the future. Figure 6b displays the lattice parameters and unit-cell volume. The star in Fig. 6b marks the lattice parameters of the fresh bovine femur bone material. The pinkish circles mark the lattice parameters of a bone fragment of a fossil Miocene hippopotamus from Bavaria  $\sim$  20 million years old). The plot of Fig. 6b allows to conclude that the c-axis of the archaeological bone mineral is fairly constant while the a-axis lattice parameter varies in a systematic way. The data lie on a pseudo-linear trend (because the unitcell volume *V* scales with the lattice parameters as  $V = a^2 c \sin 120^\circ$ , and the parabolic relation between  $V$  and  $a$  is approximately linear in a small range of data), but interestingly, this trend line connects the fresh bone with the fossil bone. Synthetic hydroxyapatite also lies on this trend line (light-blue circle) and fluorapatite (orange circles) as well. We infer that the bone mineral undergoes a chemical alteration during burial, possibly by exchange of the carbonate with hydroxyl and fluorine. But this interpretation needs to be confirmed with future studies.



Figure 7 plots the unit-cell volume vs. the distribution width of the statistical variation of lattice parameters within each single sample (the so-called microstrain), which also gives a diffraction line broadening, which can be distinguished, however, from the size broadening to its different dependence on the scattering angle  $(tan\theta)$ compared to the size broadening  $(cos \theta)^{-1}$ . There is a correlation indicating that smaller unit cells, which appear to occur due to alteration by loss of carbonate, as discussed above, have a more inhomogeneous distribution of lattice parameters. This can be explained by an inhomogeneity in the progress of the alteration reaction within each bone sample. However, the plot also indicates that the three different species which were examined show different characteristic, with the wild form (Cervus elaphus) displaying a larger spread than the domesticated species (Sus and Bos).

Figure [8](#page-13-0) displays a compilation of crystallographic parameters and  ${}^{87}S/{}^{86}Sr$ isotope values for the 63 samples. It shows that the data set is fairly uniform with any correlations occurring in a quite random manner. We interpret this situation as due to the successful cleaning procedure, which removed potential contamination, and that in the selected material, the isotope ratios do not show any trends with the crystallographic parameters of the bone apatite.

#### Furnace Annealing (Experimental Cremation)

To simulate the effect of heat treatment on bone material, we performed annealing experiments with bovine femur material. Figure [9](#page-14-0)a gives an overview of the change of the bone mineral diffractograms as a function of annealing temperature; Fig. [9b](#page-14-0) and c show sections of the diffractograms as difference plots between measured and calculated profile. Below  $400^{\circ}$ C, the width of the peaks reduces only little, but we see small but significant changes of the lattice parameters (Fig. [10\)](#page-15-0). These could be the result of the breakdown of the collagen and the changes in the composite structure of the bone occurring accordingly. Between 400  $^{\circ}$ C and 600  $^{\circ}$ C, the trend in the lattice parameters is reversed compared to the lower temperatures, indicating that a different and so far unknown structural process sets in. From about  $600^{\circ}$ C upwards, there is a marked

<span id="page-13-0"></span>

Fig. 8 Scattergram of the most relevant crystallographic parameters (from Schmahl et al. [2016\)](#page-28-0)

increase in sharpness of the XRD peaks, which is related to a dramatic increase of the crystallite size and a progressive transformation from carbonated apatite to hydroxyapatite as a function of annealing temperature (Fig. [10](#page-15-0); Table [1\)](#page-16-0). Similar results have been obtained by Rogers and Daniels ([2002](#page-28-0)). CaO forms as a further reaction product of the bioapatite decomposition at 800 $\degree$ C and above.

Infrared spectra of the cremated bioapatite are shown in Figs. [11](#page-17-0), [12](#page-17-0) and [13](#page-18-0). Figure [11](#page-17-0) compares a typical IR spectrum of archaeological bone with those of two selected experimentally annealed bones. The archaeological bone–like fresh bone– shows carbonate vibration bands. Their intensity decreases with heat treatment, and they are absent in the material annealed at  $1000^{\circ}$ C, which shows the spectrum of hydroxyapatite. In Fig. [12,](#page-17-0) spectral changes with heat treatment are shown, in particular, the development of the  $OH^-$  stretching vibration at 3568 cm<sup>-1</sup> and the  $OH^-$  libration near 633 cm<sup>-1</sup> as a function of heat treatment. It is very evident that the contributions of  $OH^-$  to the spectra of untreated bone are quite insignificant and that they rise with heat treatment. This can also be seen in the FTIR spectra of Stiner et al. [\(1995](#page-29-0)). Further, the deproteinated untreated bone shows an intense contribution of  $H_2O$ , which diminishes with heat treatment and which is practically absent in the sample treated at  $1000\,^{\circ}$ C. This all points to the assumption that the bioapatite is a carbonate-H<sub>2</sub>O-apatite with little OH<sup>-</sup> (Rey et al. [1995;](#page-28-0) Loong et al. [2000](#page-27-0); Pasteris et al. [2004\)](#page-27-0) and it becomes hydroxyapatite with

<span id="page-14-0"></span>

Fig. 9 Overview of X-ray diffractograms of bovine femur bone mineral samples annealed for 1 h at different temperatures (experimental cremation). Note the dramatically increasing sharpness of the XRD peaks with annealing temperatures above  $600^{\circ}$ C corresponding to increasing crystallite size. Exemplary Rietveld fits are shown in (b) and (c) for treatment at 600 °C and 800 °C

heat treatment, losing CO<sub>2</sub> and perhaps some  $H_2O$  (CO<sub>3</sub><sup>2</sup> +  $H_2O \rightarrow CO_2 + 2OH^-$ ). At the same time, the high temperatures lead to an increased crystallite size by Ostwald-ripening, driven by the reduction of free energy by lowering the internal surface. The excess free energy due to the surface is proportional to  $(2\gamma/r)$  where  $\gamma$  is the specific surface energy and  $r$  is the crystallite size; thus, large crystallites grow at the expense of smaller crystallites.

Figure [13](#page-18-0) displays the spectral decomposition based on seven peaks in the 400–800 cm<sup>-1</sup> range which covers the  $v_4$  phosphate group bending modes and the mode at 633 cm<sup>-1</sup> which is attributed to OH<sup>-</sup> libration (Gonzalez-Diaz and Hidalgo [1976;](#page-27-0) Gonzalez-Diaz and Santos [1977;](#page-27-0) Vandecandelaere et al. [2012\)](#page-29-0). The most robust spectral parameter useful to correlate with cremation temperature is the full width at half maximum (Balan et al. [2011](#page-26-0)) of the band at 603 cm<sup> $^{-1}$ </sup>. It correlates linearly with cremation temperature (Fig. [14\)](#page-19-0). For uncremated bones and for low temperatures of heat treatment, the FTIR peaks are poorly developed and show a large overlap of the spectral features (Figs. [13](#page-18-0)a and [14](#page-19-0)). As the material gets more ordered while it approaches more stoichiometric chemistry with increasing sintering temperature, the peaks get increasingly narrower and well defined. The calibration equation is

<span id="page-15-0"></span>

Fig. 10 Size of the coherently diffracting domains in c- and a- and b-direction (crystallite sizes labelled "c-size" and "a-size", respectively) and lattice parameters for the experimentally annealed samples as a function of annealing temperature. Annealing time is 1 h

Experimentally annealed samples							
Annealing temperature $(^{\circ}C)$	$c(\AA)$	$a(\AA)$	$c$ -size (nm)	$(a,b)$ -size (nm)			
21	6.8854(5)	9.4318(7)	12.7	3.9			
200	6.8876(5)	9.4234(7)	11.7	4.1			
300	6.8932(5)	9.4218(7)	11.8	4.2			
400	6.8912(4)	9.4161(6)	11.7	4.3			
500	6.8921(4)	9.4231(5)	12.1	4.6			
600	6.8899(3)	9.4266(4)	12.7	6.1			
700	6.88718(11)	9.4303(12)	22.1	19.0			
800	6.88486(5)	9.42865(6)	81.2	75.8			
900	6.88616(6)	9.43106(7)	108.2	70.6			
1000	6.88601(5)	9.42822(5)	163.7	131.5			
Eching samples							
8 Eching 247	6.891953(18)	9.41861(20)	104	93			
10_Eching_626	6.8836(7)	9.44582(9)	128	44			
14_Eching_727	6.89629(12)	9.41812(17)	22.5	11.5			
17_Eching_346	6.88568(3)	9.42397(3)	113	101			
18 Eching 140	6.88682(3)	9.42269(4)	143	98			
19_Eching_219	6.88686(2)	9.42079(3)	188	130			
20_Eching_222	6.89439(14)	9.4198(2)	22.5	10.2			
Zuchering skull							
Outer_1	6.89796(6)	9.42178(8)	69	60			
Inner_1	6.89727(14)	9.4221(2)	36	22			
Outer <sub>2</sub>	6.89857(4)	9.42311(5)	139	147			
Inner <sub>2</sub>	6.89603(5)	9.42419(6)	173	108			
Outer 3	6.89873(5)	9.4215(6)	73	63			
Inner <sub>3</sub>	6.89832(16)	9.4211(2)	35	21			
Outer 4	6.89772(11)	9.42127(14)	36	27			
Inner_4	6.89885(18)	9.4185(2)	23	13			
Outer <sub>5</sub>	6.89797(8)	9.42159(11)	42	32			
Inner_5	6.9004(2)	9.4206(3)	22	12			

<span id="page-16-0"></span>Table 1 X-ray diffraction results on experimentally annealed samples and cremated archaeological samples

Numbers in brackets are standard deviations referring to the last digit(s). The estimated standard deviation for crystallite sizes is estimated as 5 %

$$
FWHM = -0.0231 \left[ \frac{\text{cm}^{-1}}{\text{°C}} \right] *T + 37.9 \left[ \text{cm}^{-1} \right] \tag{1}
$$

where  $T$  is the cremation temperature in  $\mathrm{C}$ .

<span id="page-17-0"></span>

Fig. 11 Comparison of FTIR spectra of uncremated archaeological and experimentally annealed samples with the most important vibrational bands indicated. Note the absence of carbonate bands in the 1000  $\degree$ C annealed samples and the absence of OH<sup>-</sup> libration in the uncremated sample and the 400 °C annealed sample



Fig. 12 Change of the FTIR spectra of degreased and deproteinated bovine (femur) bone material as a function of experimental annealing temperature (annealing time 1 h). Note the pronounced increase of the  $OH^-$  signals and the decrease of the  $H_2O$  signal with increasing treatment temperature

## Cremated Archaeological Samples

We exemplify results on cremated archaeological bone remains from the Bronze Age (Urnfield Culture) of southern Bavaria with long bones from the Eching site and a skull from the Zuchering site.

Figure [15](#page-19-0) shows the Fourier transform infrared spectra of seven samples from the Eching locality in the spectral range of the  $\nu_4$  phosphate group bending modes and the OH<sup>-</sup> libration mode at 633 cm<sup>-1</sup>. The seven samples can be separated into two groups. The first group consists of the samples 20\_Eching (black trace) and 14\_Eching (red trace) which show fairly similar FTIR spectra. Their  $\sim 567$  cm<sup>-1</sup> band is at lower frequencies than that of the second group, and the width of both the  $567$  cm<sup>-1</sup> band and the  $603$  cm<sup>-1</sup> band is broad; further, they do not show a well-

<span id="page-18-0"></span>

**Fig. 13** Spectral decomposition of the FTIR signal of the  $\nu_4$  [PO<sub>4</sub>]<sup>3-</sup> vibration bands and the OH<sup>-</sup> libration band in the 400 cm<sup>-1</sup> to 800 cm<sup>-1</sup> range into seven contributions for two exemplary states of experimental annealing of bovine femur bone mineral

differentiated hydroxyl libration peak at  $632 \text{ cm}^{-1}$ . The second group consists of the remaining five samples with narrower peaks for the phosphate bands and a welldifferentiated  $OH^-$  libration band. Correspondingly, the group 2 samples have seen higher cremation temperatures than the group 1 samples.

Table [2](#page-20-0) lists the results for the estimated cremation temperature based on Eq. [\(1](#page-14-0)) from the spectral decomposition of the P–O bands. For group 1, we estimate a cremation temperature around 750  $\degree$ C, while the group 2 samples are placed between  $1000^\circ$  and  $1150^\circ$ C.

As the XRD of the Eching samples (see below) gave strong indications that cremated bones are typically inhomogeneous with respect to the cremation state, we also investigated a cremated skull from the Zuchering site for which we had visually observed colour differences between the inner and outer side of the cranium. Five samples from the cranium were divided into the inner compact bone layer and the outer compact bone layer (the spongiosa was thus lost). Infrared spectra of the skull sample are shown in Fig. [16](#page-20-0); FWHM values of the  $\sim 603 \text{ cm}^{-1}$ band and corresponding calculated cremation temperatures are given in Table [2](#page-20-0). First of all, the IR data for the single skull are quite variant and indicate a range of cremation temperatures between 834 °C  $\pm$  45 °C and 1016 °C  $\pm$  20 °C, and there are significant differences between corresponding inner and outer layers, which are just millimetres apart. In contrast, the estimation of the cremation temperature

<span id="page-19-0"></span>

Fig. 14 Correlation of the full width at half maximum (FWHM) of the  $603 \text{ cm}^{-1}$  band vs. annealing temperature of experimental cremation (annealing time 1 h)



Fig. 15 FTIR spectra of archaeological cremated samples from the Urnfield Culture site of Eching (1300–800 bc). The phosphate  $\nu_4$  bending modes are highlighted with changes in the frequency (1) of the  $\sim$ 567 cm<sup>-1</sup> band, and the appearance and broadening of the OH<sup>-</sup> libration mode at  $633 \text{ cm}^{-1}$  are indicated (2)

<span id="page-20-0"></span>**Table 2** Full width at half maximum of the ~603 cm<sup>-1</sup>  $\nu_4$  [PO<sub>4</sub>] infrared absorption band for the archaeological cremated samples from the Eching and Zuchering Urnfield Culture sites, the corresponding calculated cremation temperature from Eq. [\(1](#page-14-0)), the cremation temperature estimated from XRD crystallite size (Table [1](#page-16-0); Fig. [10](#page-15-0)) and the cremation temperature estimate from the colour of the specimen on the basis of (Wahl [1981](#page-29-0))

	FWHM of the 603 cm <sup><math>-1</math></sup> P-O band	Calculated cremation temperature	Estimated cremation temperature from	Estimated cremation temperature after Wahl		
Sample	$\rm (cm^{-1})$	from Eq. 1 $(°C)$	XRD line width $(^{\circ}C)$	$(1981)$ (°C)		
20 Eching 222	20.1(1.2)	769(100)	700			
14_Eching_727	20.7(1.3)	743(100)	700			
8 Eching 247	14.4(3)	1015(15)	840			
10_Eching_626	14.3(2)	1020(10)	920			
17_Eching_346	11.6(7)	1136(100)	900			
18 Eching 140	12.74(1.8)	1087(10)	950			
19 Eching 219	11.6(2)	1136(10)	1030			
Zuchering skull						
Outer 1	16.6(7)	920(30)	780	550		
Inner 1	18.6(0.98)	834(45)	750	550		
Outer <sub>2</sub>	15.12(81)	985(35)	920	> 800		
Inner 2	14.4(4)	1016(20)	1020	> 800		
Outer 3	15.12(57)	984(25)	800	550-600		
Inner 3	15.0(5)	989(25)	740	550-600		
Outer <sub>4</sub>	17.6(8)	877(35)	740	500-550		
Inner 4	20(3)	773(167)	700	500-550		
Outer 5	16.5(7)	925(30)	750	550		
Inner 5	20(3)	773(130)	700	550		



Fig. 16 FTIR spectra of archaeological cremated skull from the Urnfield Culture site of Zuchering (1300–800 BC) covering the phosphate  $v_4$  bending modes and the OH<sup>-</sup> libration mode near 633 cm<sup>-1</sup>



Fig. 17 XRD patterns of the cremated archaeological samples of the Urnfield Culture Eching locality. Cu-K $\alpha_1$  radiation. The group 1 samples 14\_Eching and 20\_Eching show broad peaks, indicating smaller crystallize sizes and thus lower cremation temperatures than the other samples (group 2). The diffractogram of the 10\_Eching sample indicates a completely different material state than the other group 2 samples

based on the colour of the sample (Wahl [1981\)](#page-29-0) gives much lower temperatures. The estimate of the cremation temperature based on the FWHM of the  $~603 \text{ cm}^{-1}$  band may give slightly overestimated values as the correlation in Fig. [14](#page-19-0) may not really be linear and flatten off at temperatures  $>700$  °C. However, even a simple interpolation between the data points in Fig. [14](#page-19-0) with the FWHM values of the cremated samples indicates temperatures in excess of those based on the method of (Wahl [1981\)](#page-29-0).

The X-ray diffractograms of the Eching samples are compiled in Fig. 17. Here three groups can be distinguished. Like in the FTIR data above, the samples 14\_Eching and 20\_Eching experienced lower cremation temperatures as their XRD peaks are broad. Rietveld refinement (Fig. [18\)](#page-22-0) puts their crystallite sizes to 22 nm (Table [1\)](#page-16-0), which corresponds to 700  $\degree$ C cremation temperature, within limits of error well corresponding to the estimate derived from FTIR above. The group 2 samples have sharper lines with the exception of 10\_Eching. The latter forms a very particular case which cannot be treated with our temperature calibration for XRD (see below). The group 2 samples excluding 10 Eching show crystallite sizes in the order of 100–190 nm which puts them into the 800–1000  $\degree$ C range, again corresponding to the FTIR within bounds of error. A representative Rietveld analysis is shown in Fig. [19](#page-22-0). The apatite in the 10 Eching sample appears to have an entirely different structure from what has been encountered so far in fresh, uncremated archeologic and experimentally cremated bone samples. The observed diffraction trace can be fitted with a monoclinic apatite structure (Ikoma et al. [1999\)](#page-27-0), which is able to produce the flat-topped XRD peaks encountered here

<span id="page-22-0"></span>

Fig. 18 Section of the X-ray diffractogram of the 14 Eching sample. Red: observed profile. Cu-K $\alpha_1$  radiation. Black: calculated profile. Blue: difference curve. Green: markers for reflection positions. From top to bottom: calcite, quartz, apatite-I and apatite-II. To fit the observed profile with reasonable quality, two apatite phases with slightly different lattice parameters and different microstructural characteristics had to be used



Fig. 19 Section of the X-ray diffractogram of the 8\_Eching sample. Red: observed profile. Cu-K $\alpha_1$  radiation. Black: calculated profile. Blue: difference curve. Green: markers for reflection positions. To fit the observed profile with reasonable quality, two apatite phases with slightly different lattice parameters and different microstructural characteristics had to be used

(Fig. [20\)](#page-23-0). Quartz and calcite impurities from the sediment are also present, which were included in the fit. A possibility to explain the occurrence of the monoclinic symmetry of apatite may be a reaction with ashes or other materials at high temperature in the burning funeral pile. What is clear from the Rietveld refinements of the archaeological sample is that we get pathologic XRD profiles which are not as clearly interpretable as the profiles from fresh or experimentally annealed bone materials. The diffractograms clearly indicate an inhomogeneity in the sample in the sense that apatites with different lattice parameters, different crystallite sizes,

<span id="page-23-0"></span>

Fig. 20 Section of the X-ray diffractogram of the 10 Eching sample. Red: observed profile. Cu-K $\alpha_1$  radiation. *Black*: calculated profile. *Blue*: difference curve. *Green*: markers for reflection positions. From top to bottom: calcite, quartz, monoclinic apatite. To fit the observed profile with reasonable quality, monoclinic symmetry for apatite had to be used

and microstrain values are needed to model the diffractograms. The origin of inhomogeneity can be attributed to different degrees of cremation intensity. These can result from inhomogeneous temperatures produced by the burning wood logs and tissues, and also from differing lengths of time to which certain bone segments were exposed to the highest cremation temperatures. A similar observation has been made by Stiner et al. [\(1995](#page-29-0)) on paleolithic bone material.

Finally, we turn to the XRD results on the Zuchering skull samples. Diffractograms are shown in Fig. [21.](#page-24-0) The inner parts of samples 4 and 5 have the broadest lines corresponding to the lowest treatment temperatures. From the corresponding crystallite size of 22–23 nm obtained from Rietveld refinement (Table [1\)](#page-16-0), we estimate the cremation temperatures as  $700^{\circ}$  or slightly above, which is quite consistent with the IR data discussed above. Sample 2 has certainly experienced the highest cremation temperatures, and here we also have fair agreement between FTIR and XRD results. On the whole, we see that the IR calibration overestimates the cremation temperature compared to XRD; for the skull sample 3, we have the greatest discrepancies. There are several potential origins of

<span id="page-24-0"></span>

Fig. 21 Section of the X-ray diffractograms of the Zuchering skull sample comparing outer and inner compact bone layers of the cranium. Cu-K $\alpha_1$  radiation

difference obtained in cremation temperature when applied to archaeological cremated samples. First, our IR calibration is based on the assumption of a linear relationship between the FWHM of the  $\sim 603$  cm<sup>-1</sup> phosphate vibration (Fig. [14](#page-19-0)) and the annealing temperature. From the XRD evaluation of the experimentally annealed samples, we clearly see that the microstructural change in the material is strongly nonlinear in temperature, with a drastic change between the regimes below and above 600–700 °C (Fig. [10](#page-15-0)). Second, the archaeological cremated bones are clearly inhomogeneous, and the calibration so far assumes a well-defined average state. However, both the IR and the XRD data are at odds with the cremation temperatures estimated from the colour of the specimens after Wahl [\(1981](#page-29-0)).

### Concluding Discussion

Our study of uncremated mammal bone samples from the alpine region covers ages from 7600 to 550 years before present. Only those samples which were of the best quality selected for the isotope studies within FOR1670 were analysed. Visually decomposed bones were avoided completely, and also the surfaces of the bones were mechanically cleaned by grinding away soil and/or decomposed material. The mineralogic state of the thus prepared samples showed only minor signs of diagenetic alteration, and beside bone apatite, the only secondary phase was calcite. The most pronounced change with archeologic age resides in the lattice parameter and in the crystalline domain size in the short dimension of the original bone-apatite platelets; the long direction corresponding to the crystallographic c-axis of the apatite appears almost unaffected. From the data, it is clear that other factors such as anatomic variation of the original materials and conditions in the soil play a major role in the diagenetic change of the bone material, masking the changes with burial time. As judged from the lattice parameters, the diagenetic alteration of the original bone mineral, apart from the sight increase in crystallite size, is possibly related to exchange of carbonate by hydroxyl and fluorine.

We established calibration curves from annealing experiments with bovine femur bone material at different temperatures for 1 h annealing time, which were subsequently used to estimate cremation temperatures of archaeological bones based on FTIR spectra and X-ray diffractograms. The experiments indicate that the original bone apatite contains little–if any–OH<sup>-</sup> while carbonate and  $H_2O$  are present. Thus, the bone mineral is a carbonate-hydro-apatite rather than hydroxyapatite. With increasing annealing temperature, water and carbonate leave the material, and from 600  $\degree$ C, the bone mineral progressively reacts to form hydroxyapatite with increasing purity and crystallite size with increasing temperature. We analysed some cremated bones from the Urnfield Culture sites of Eching and Zuchering, southern Bavaria. The analyses clearly indicate that the archaeological cremated bones are inhomogeneous materials, where within fractions of centimetres, the different parts of the samples experienced different cremation temperatures and/or times at temperature. Thus, compared to annealing experiments in furnaces, the conditions within the pile of burning logs and burning tissue are strongly variable locally, in particular towards the outside of the pyre. As the urns typically contain only a small fraction of the skeleton, the selection of bone fragments for burial in the urn might preferentially pick such items which experienced inhomogeneous conditions. The estimated cremation temperatures based on the FTIR and XRD analytical approaches tentatively agree but are still not fully mutually consistent. The IR and XRD data are consistently at odds with the cremation temperatures estimated from the colour of the specimens after Wahl [\(1981](#page-29-0)). To discuss the discrepancies between the FTIR and the XRD approaches, we first need to consider that the archaeological cremated bones are clearly inhomogeneous and the calibration so far assumes that a well-defined average state exists. Secondly, our IR calibration so far assumes a linear model relation (Eq. [1](#page-14-0)) between the FWHM of the  $\sim 603$  cm<sup>-1</sup> phosphate vibration (Fig. [14](#page-19-0)) and the

<span id="page-26-0"></span>cremation temperature. From the XRD evaluation of the experimentally annealed samples, however, we clearly see that the microstructural change in the material is not simply proportional to temperature: there is a drastic change between the regimes below and above 600–700 °C (Fig. [10](#page-15-0)). Below this temperature, the IR signal is more sensitive to the change in material state, while in the temperature range above 600  $\degree$ C, it is the XRD signal which is more sensitive. In the future, more refined models can be produced on the basis of more annealing experiments made to obtain better statistics. Further, cremation experiments must be done in the conditions that exist in a funeral pyre to gain experience on temperature and burning time inhomogeneity. While the evaluation of the diffractograms today is rigorously based on a physical model, the evaluation of spectral components in FTIR spectra is still empirical, and results depend very much on empirical assumptions made on the spectral shape of any single band and the number of bands contributing to the spectral range of overlapping peaks. Future work also definitely needs to address the chemistry and anatomical variability of original bone material within and between species in much more detail than is known at present.

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## References

- Alexander B, Daulton TL, Genin GM, Lipner J, Pasteris JD, Wopenka B, Thomopoulos S (2012) The nanometre-scale physiology of bone: steric modelling and scanning transmission electron microscopy of collagen–mineral structure. J Royal Soc Interface 9:1774–1786
- Balan E, Delattre S, Roche D, Segalen L, Morin G, Guillaumet M, Blanchard M, Lazzeri M, Brouder C, Salje EKH (2011) Line-broadening effects in the powder infrared spectrum of apatite. Phys Chem Miner 38:111–122
- Berna F, Matthews A, Weiner S (2004) Solubilities of bone mineral from archaeological sites: the recrystallization window. J Archaeol Sci 31:867–882
- Cuisinier F, Bres EF, Hemmerle J, Voegel JC, Frank RM (1987) Transmission electron microscopy of lattice planes in human alveolar bone apatite crystals. Calcif Tissue Int 40:332–338
- de Keijser TH, Langford JI, Mittemeijer EJ, Vogels ABP (1982) Use of the Voigt function in a single-line method for the analysis of X-ray diffraction line broadening. J Appl Crystallogr 15:308–314
- de Leeuw NH (2010) Computer simulations of structures and properties of the biomaterial hydroxyapatite. J Mater Chem 20:5376–5389
- Elliot JC (2002) Calcium phosphate biominerals. In: Kohn MJ, Rakovan J, Hughes JM (Hrsg) Phosphates: geochemical, geobiological and material importance. Reviews in mineralogy and geochemistry, vol 48. Mineralogical Society of America, Washington, DC, pp 631–672
- Enzo S, Bazzoni M, Mazzarello V, Piga G, Bandiera P, Melis P (2007) A study by thermal treatment and X-ray powder diffraction on burnt fragmented bones from tombs II, IV and IX belonging to the hypogeic necropolis of "Sa Figu" near Ittiri, Sassari (Sardinia, Italy). J Archaeol Sci 34:1731–1737
- Galeano S, García-Lorenzo ML (2014) Bone mineral change during experimental calcination: an X-ray diffraction study. J Forensic Sci 59:1602–1606
- Gernaey AM, Waite ER, Collins MJ, Craig OE, Sokol RJ (2001) Survival and interpretation of archaeological proteins. In Brothwell DR, Pollard AM (Hrsg) Handbook of archaeological science. Wiley, Chichester: 323–329
- <span id="page-27-0"></span>Gonzalez-Diaz PF, Hidalgo A (1976) Infrared spectra of calcium apatites. Spectrochim Acta A Mol Spectrosc 32A:1119–1124
- Gonzalez-Diaz PF, Santos M (1977) On the hydroxyl ions in apatites. J Solid State Chem 22:193–199
- Grunenwald A, Keyser C, Sautereau AM, Crubézy E, Ludes B, Drouet C (2014) Novel contribution on the diagenetic physicochemical features of bone and teeth minerals, as substrates for ancient DNA typing. Anal Bioanal Chem 406:4691–4704
- Grupe G, Harbeck M, McGlynn GC (2015) Prähistorische Anthropologie. Springer, Berlin
- Harbeck M, Schleuder R, Schneider J, Wiechmann I, Schmahl WW, Grupe G (2011) Research potential and limitations of trace analyses of cremated remains. Forensic Sci Int 204:191–200
- Hedges REM (2002) Bone diagenesis: an overview of precesses. Archaeometry 44:319–328
- Ikoma T, Yamazaki A, Nakamura S, Akao M (1999) Preparation and structure refinement of monoclinic hydroxyapatite. J Solid State Chem 144:272–276
- Jäger C, Welzel T, Meyer-Zaika W, Epple M (2006) A solid-state NMR investigation of the structure of nanocrystalline hydroxyapatite. Magn Reson Chem 44:573–580
- Landis WJ, Jacquet R (2013) Association of calcium and phosphate ions with collagen in the mineralization of vertebrate tissues. Calcif Tissue Int 93:329–337
- Lebon M, Reiche I, Frohlich F, Bahain J, Falgueres C (2008) Characterization of archaeological burnt bones: contribution of a new analytical protocol based on derivative FTIR spectroscopy and curve fitting of the  $v_1$  and  $v_3$  PO<sub>4</sub>. Anal Bioanal Chem 392:1479–1488
- Lee-Thorp J, Sponheimer M (2003) Three case studies used to reassess the reliability of fossil bone and enamel isotope signals for paleodietary studies. J Anthropol Archaeol 23:208–216
- LeGeros RZ, Trautz OR, Legeros JP, Klein E, Shirra WP (1967) Apatite crystallites: effects of carbonate on morphology. Science 155:1409–1411
- LeGeros RZ, Trautz OR, Legeros JP, Klein E (1968) Carbonate substitution in the apatite structure. Bull Soc Chim Fr 4:1712–1718
- Loong CK, Rey C, Kuhn LT, Combes C, Wu Y, Chen SH, Glimcher MJ (2000) Evidence of hydroxyl-ion deficiency in bone apatites: an inelastic neutron-scattering study. Bone 26:599–602
- McKinley J (2016) Complexities of the ancient mortuary rite of cremation: an osteoarchaeological conundrum. In: Grupe G, McGlynn GC (eds) Isotopic landscapes in bioarchaeology. Springer, Berlin, pp. 17–41
- Mittemeijer EJ, Welzel U (eds) (2012) Modern diffraction methods. Wiley-VCH, Weinheim, 528p
- Munro LE, Longstaffe FJ, White CD (2008) Effects of heating on the carbon and oxygen-isotope compositions of structural carbonate in bioapatite from modern deer bone (Beyond documenting diagenesis: the fifth international bone diagenesis workshop). Palaeogeogr Palaeoclimatol Palaeoecol 266:142–150
- Nielsen-Marsh CM, Hedges REM (2000) Patterns of diagenesis in bone I: the effects of site environments. J Archaeol Sci 27:1139–1150
- Nielsen-Marsh CM, Gernaey A, Turner-Walker G, Hedges REM, Pike A, Collins MJ (2000) The chemical degradation of bone. In: Cox M, Mays S (eds) Human osteology in archaeology and forensic science. Greenwich Medical Media, London, pp. 439–454
- Olsen J, Heinemeier J, Bennike P, Krause C, Hornstrup KM, Thrane H (2008) Characterisation and blind testing of radiocarbon dating of cremated bone. J Archaeol Sci 35:791–800
- Pasteris JD, Wopenka B, Freeman JJ, Rogers K, Valsami-Jones E, van der Houwen AM, Silva MJ (2004) Lack of OH in nanocrystalline apatite as a function of degree of atomic order: implications for bone and biomaterials. Biomaterials 25:229–238
- Pecharsky VK, Zavalij PY (2003) Fundamentals of powder diffraction and structural characterization of materials. Springer, New York, 741p
- Pedone A, Corno M, Civalleri B, Malavasi G, Menziani C, Segrea U, Ugliengo P (2007) An ab initio parameterized interatomic force field for hydroxyapatite. J Mater Chem 17:2061–2068
- Person A, Bocherens H, Saliege JF, Paris F, Zeitoun V, Gerard M (1995) Early diagenetic evolution of bone phosphate—an X-ray diffractometry analysis. J Archaeol Sci 22:211–221
- Peters F, Schwarz K, Epple M (2000) The structure of bone studied with synchrotron X-ray diffraction, X-ray absorption spectroscopy and thermal analysis. Thermochim Acta 361:131–138
- <span id="page-28-0"></span>Piga G, Malgosa A, Thompson TJU, Enzo S (2008) A new calibration of the XRD technique for the study of archaeological burned human remains. J Archaeol Sci 35:2171–2178
- Piga G, Thompson TJU, Malgosa A, Enzo S (2009a) The potential of X-Ray diffraction in the analysis of burned remains from forensic contexts. J Forensic Sci 54:3534–3539
- Piga G, Santos-Cubedo A, Moya Sola S, Brunetti A, Malgosa A, Enzo S (2009b) An X-ray diffraction (XRD) and X-ray fluorescence (XRF) investigation in human and animal fossil bones from Holocene to Middle Triassic. J Arch Sci 36:857–1868
- Piga G, Solinas G, Thompson TJU, Brunetti A, Malgosa A, Enzo S (2013) Is X-ray diffraction able to distinguish between animal and human bones? J Archaeol Sci 40:778–785
- Pucéat E, Reynard B, Lécuyer C (2004) Can crystallinity be used to determine the degree of chemical alteration of biogenic apatites? Chem Geol 205:83–97
- Rey C, Collins B, Goehl T, Dickson IR, Glimcher MJ (1989) The carbonate environment in bone mineral: A resolution-enhanced fourier transform infrared spectroscopy study. Calcif Tissue Int 45:157–164
- Rey C, Combes C, Drouet C, Glimcher MJ (2009) Bone mineral: update on chemical composition and structure. Osteoporos Int 20:1013–1021
- Rey C, Combes C, Drouet C, Cazalbou S, Grossin D, Brouillet F, Sarda S (2014) Surface properties of biomimetic nanocrystalline apatites; applications in biomaterials. Prog Cryst Growth Charact Mater 60:63–73
- Rey C, Miquel JL, Facchini L, Legrand AP, Glimcher MJ (1995) Hydroxyl groups in bone mineral. Bone 16:583–586
- Rey C, Renugopalakrishnan V, Collins B, Glimcher MJ (1991) Fourier transform infrared spectroscopic study of carbonate ions in bone mineral during aging. Calcif Tissue Int 49:251–258
- Rey C, Shimizu M, Collins B, Glimcher MJ (1990) Resolution-enhanced fourier transform infrared spectroscopy study of the environment of phosphate ions in the early deposits of a solid phase of calcium-phosphate in bone and enamel, and their evolution with age. I: Investigations in the v4PO4 domain. Calcif Tissue Int 46:384–394
- Reznikov N, Shahar R, Weiner S (2014) Three-dimensional structure of human lamellar bone: the presence of two different materials and new insights into the hierarchical organization. Bone 59:93–104
- Rho J-Y, Kuhn-Spearing L, Zioupos P (1998) Mechanical properties and the hierarchical structure of bone. Med Eng Phys 20:92–102
- Rietveld H (1969) A profile refinement method for nuclear and magnetic structures. J Appl Crystallogr 2:65–71
- Rodriguez-Carvajal J (1993) Recent advances in magnetic structure determination by neutron powder diffraction. Phys B Condens Matter 192:55–69
- Rodriguez-Carvajal J, Roisnel T (2004) Line broadening analysis using FullProf: determination of microstructural properties. Mater Sci Forum 443:123–126
- Rogers KD, Beckett S, Kuhn S, Chamberlain A, Clement J (2010) Contrasting the crystallinity indicators of heated and diagenetically altered bone mineral. Palaeogeogr Palaeoclimatol Palaeoecol 296:125–129
- Rogers KD, Daniels P (2002) An X-ray diffraction study of the effects of heat treatment on bone mineral microstructure. Biomaterials 23:2577–2585
- Schmahl WW, Kocis B, Toncala A, Grupe GJ (2016) Mineralogic characterisation of archaeologic bone. In: Grupe G, McGlynn GC (eds) Isotopic landscapes in bioarchaeology. Springer, Berlin, pp. 17–41
- Schurr MR, Hayes RG, Cook DC (2008) Thermally induced changes in the stable carbon and nitrogen isotope ratios of charred bones. In: Schmidt CW, Symes SA (eds) The analysis of burned human remains. Academic Press, London, pp 95–108
- Schwarcz HP, McNally EA, Botton GA (2014) Dark-field transmission electron microscopy of cortical bone reveals details of extrafibrillar crystals. J Struct Biol 188:240–248
- Shemesh A (1990) Crystallinity and diagenesis of sedimentary apatites. Geochim Cosmochim Acta 54:2433–2438
- Shinomiya T, Shinomiya K, Orimoto C, Minami T, Tohno Y, Yamada MO (1998) In- and out-flows of elements in bones embedded in reference soils. Forensic Sci Int 98:109–118
- <span id="page-29-0"></span>Sillen A (1989) Diagenesis of the inorganic phase of cortical bone. In Price TD (Hrsg) The chemistry of prehistoric human bone. Cambridge University Press, New York, pp 211–229
- Sponheimer M, de Ruiter D, Lee-Thorp J, Späth A (2005) Sr/Ca and early hominin diets revisited: new data from modern and fossil tooth enamel. J Hum Evol 48:147–156
- Stathopoulou ET, Psycharis V, Chryssikos GD, Gionis V, Theodorou G (2008) Bone diagenesis: new data from infrered spectroscopy and X-ray diffraction. Palaeogeogr Palaeoclimatol Palaeoecol 266:168–174
- Stiner MC, Kuhn SL, Weiner S, Bar-Yosef O (1995) Differential burning, recrystallization, and fragmentation of archaeological bone. J Archaeol Sci 22:223–237
- Surovell TA, Stiner M (2001) Standardizing infra-red measures of bone mineral crystallinity: an experimental approach. J Archaeol Sci 28:633–642
- Thompson P, Cox DE, Hastings JB (1987) Rietveld refinement of Debye-Scherrer synchrotron X-ray data from  $Al_2O_3$ . J Appl Crystallogr 20:79–83
- Tonegawa T, Ikoma T, Yoshioka T, Hanagata N, Tanaka J (2010) Crystal structure refinement of A-type carbonate apatite by X-ray powder diffraction. J Mater Sci 45:2419–2426
- Trueman CN, Privat K, Field J (2008) Why do crystallinity values fail to predict the extent of diagenetic alteration of bone mineral? Palaeogeogr Palaeoclimatol Palaeoecol 266:160–167
- Tütken T, Vennemann TW, Pfretzschner H-U (2008) Early diagenesis of bone and tooth apatite in fluvial and marine settings: constraints from combined oxygen isotope, nitrogen and REE analysis. Palaeogeogr Palaeoclimatol Palaeoecol 266:254–268
- Turner-Walker G (2008) The chemical and microbial degradation of bones and teeth. In Pinhasi R, Mays S (Hrsg) Advances in human paleopathology. Wiley, Chichester, pp 3–29
- Vandecandelaere N, Rey C, Drouet C (2012) Biomimetic apatite-based biomaterials: on the critical impact of synthesis and post-synthesis parameters. J Mater Sci Mater Med 23:2593–2606
- Wahl J (1981) Beobachtungen zur Verbrennung menschlicher Leichname. Archäologisches Korrespondenzblatt 11:271–279
- Wang Y, Von Euw S, Fernandes FM, Cassaignon S, Selmane M, Laurent G, Pehau-Arnaudet G, Coelho C, Bonhomme-Coury L, Giraud-Guille M-M, Babonneau F, Azaïs T, Nassif N (2013) Water-mediated structuring of bone apatite. Nat Mater 12:1144–1153
- Wegst UGK, Bai H, Saiz E, Tomsia AP, Ritchie RO (2015) Bioinspired structural materials. Nat Mater 14:23–36
- Weiner S, Bar-Yosef O (1990) State of preservation of bones from the prehistoric sites in the near East: a survey. J Archaeol Sci 17:187–196
- Weiner S, Goldberg P, Bar-Yosef O (1993) Bone preservation in Kebara Cave, Israel using on-site Fourier-transform infrared spectroscopy. J Archaeol Sci 20:613–627
- Weiner S, Traub W, Wagner HD (1999) Lamellar bone: structure-function relations. J Struct Biol 126:241–255
- Wilson RM, Elliot JC, Dowker SEP (1999) Rietveld refinement of the crystallographic structure of human dental enamel apatites. Am Mineral 84:1406–1414
- Wilson RM, Elliton JC, Dowker SEP, Rodriguez-Lorenzo LM (2005) Rietveld refinements and spectroscopic studies of the structure of Ca-deficient apatite. Biomaterials 26:1317–1327
- Wopenka B, Pasteris JD (2005) A mineralogical perspective on the apatite in bone. Mater Sci Eng C 25:131–143
- Wright LE, Schwarcz HP (1996) Infrared and isotopic evidence for diagenesis of bone apatite at Dos Pilas, Guatemala: palaeodietary implications. J Archaeol Sci 23:933–944
- Yerramshetty JS, Akkus O (2008) The associations between mineral crystallinity and the mechanical properties of human cortical bone. Bone 42:476–482
- Yi H, Balan E, Gervais C, Ségalen L, Blanchard M, Lazzeri M (2014) Theoretical study of the local charge compensation and spectroscopic properties of B-type carbonate defects in apatite. Phys Chem Miner 4:347–359
- Yi H, Balan E, Gervais C, Segalen L, Fayon F, Roche D, Person A, Morin G, Guillaumet M, Blanchard M, Lazzeri M, Babonneau F (2013) A carbonate-fluoride defect model for carbonate-rich fluorapatite. Am Mineral 98:1066–1069