

Chapter 15

Gambogic Acid and Its Role in Chronic Diseases

Manoj K. Pandey, Deepkamal Karelia and Shantu G. Amin

Abstract Kokum, a spice derived from the fruit of the *Garcinia hanburyi* tree, is traditionally used in Ayurvedic medicines to facilitate digestion and to treat sores, dermatitis, diarrhoea, dysentery, and ear infection. One of the major active components of kokum is gambogic acid, also known as guttic acid, guttatic acid, beta-guttillactone, and beta-guttiferin. Gambogic acid's anti-proliferative, anti-bacterial; antioxidant and anti-inflammatory effects result from its modulation of numerous cell-signaling intermediates. This chapter discusses the sources, chemical components, mechanism of action, and disease targets of the kokum spice.

Keywords Nutraceuticals · Dietary agents · Gambogic acid · Kokum · Cancer · Signal transduction pathways

15.1 Introduction

Mother Nature has gifted us a variety of natural agents, including nutraceuticals. One of the well-known nutraceuticals is Gambogic acid (GA), which is a xanthonoid derived from the brownish or orange resin from *Garcinia hanburyi* (Fig. 15.1). *Garcinia hanburyi* is a small to medium-sized evergreen tree with smooth gray bark, and it is native to Cambodia, southern Vietnam, and Thailand. *Garcinia indica*, primarily of Indian origin, is known by many names: bindin, biran, bhirand, bhinda, kokum, katambi, panarpuli, ratamba, amsol, and tamal. In English language, it is commonly known as mangosteen, wild mangosteen, red mango, Hanbury's *Garcinia*, gambojia, gamboge, and Indian gamboge tree. Germans called this gummi-gutti.

The *Garcinia indica* seed contains 23–26 % oil, which is used in confectionery, medicines, and cosmetics. It is used in curries and other dishes as a slightly bitter spice, a souring agent, and as a substitute for tamarind.

M.K. Pandey (✉) · D. Karelia · S.G. Amin
Department of Pharmacology, College of Medicine, Pennsylvania State University, 500
University Dr., Hershey, PA 17033, USA
e-mail: mkp13@psu.edu

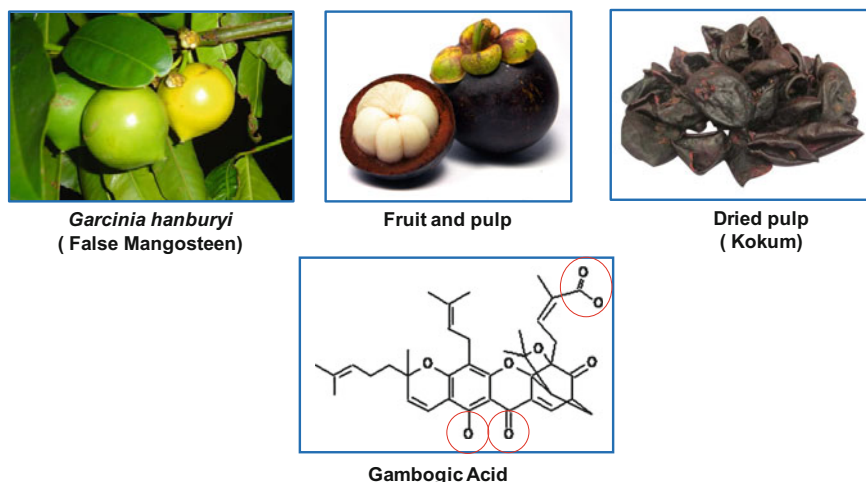


Fig. 15.1 Plant species and fruits by which gambogic acid is derived. *Highlighted circles* on GA structure indicate the most common sites for novel derivative generation

In traditional medicine, such as ayurveda, kokum is prescribed for edema, rheumatism, delayed menstruation, constipation and other bowel complaints, and intestinal parasites. The extract of *Garcinia cambogia* is used as an herbal appetite suppressant and weight-loss supplement.

In last decades, worldwide extensive studies have been performed on Gambogic acid to understand its full potential as therapeutic agents against variety of diseases including chronic diseases such as cancers which is summarized in following sections.

15.2 Physicochemical Properties of GA

GA is also chemically called as Guttic acid, Guttatic acid, beta-Guttillactone, and beta-Guttiferin. The molecular formula and weight of GA is $C_{38}H_{44}O_8$ and 628.76, respectively. The appearance of this xanthone is amorphous orange solid. The core of GA is known as xanthone core and contains a unique 4-oxatricyclo [4.3.1.0^{3,7}] decan-2-one scaffold [1, 2]. Earlier studies regarding its structural activity relationships (SAR) revealed that the C₄C bond of the α,β -unsaturated ketone in GA is critical for its antitumor activity, while the HOC(6), C(8)₄O, and C(30)OOH groups could tolerate a variety of modifications Fig. 15.1 [3, 4]. Along these lines various modifications have been performed to make GA as a better antitumor agent [5, 6].

15.3 Modulation of Cell Signaling Pathways by GA

Enthusiasm shown by researchers from around the globe clearly suggests that GA has been one of the “hot” nutraceuticals. GA has shown to be effective on different chronic diseases (Sect. 4.1 covers different chronic diseases), but its effect on cancer has been studied the most. Almost a decade ago our group showed that the anti-inflammatory and anticancer response of GA is associated with its inhibitory response on Nuclear Factor-Kappa B (NF- κ B) [7], since then plethora of studies suggest that GA regulates several key signaling pathways. GA exhibits anti-proliferative, antioxidant, and anti-inflammatory effects by modulating cell signaling pathways, enzymes, and molecular targets, such as epigenetic regulators, protein kinases, transcription factors, inflammatory biomarkers, and growth regulators. Through microarray analysis, GA modulates many gene products [8, 9] (Table 15.1).

15.3.1 GA Inhibits Signaling of Nuclear Factor-Kappa B (NF- κ B)

The transcription factor NF- κ B is one of the major mediators of inflammation and is linked with many diseases including cancer, diabetes, arthritis, and neurological disorders. Therefore, an agent that can suppress NF- κ B activation has potential for clinical use against various chronic illnesses. GA suppression of NF- κ B activation induced by TNF- α , LPS, and various agents [7, 10] leads to the suppression of NF- κ B regulated products, such as cyclooxygenase type 2 (COX-2), inducible nitric oxide synthase (iNOS), and survival proteins [7, 10, 11]. These actions give it great potential as a broad-spectrum clinical agent.

15.3.2 GA Inhibits Phosphatidylinositol 3'-Kinase/Protein Kinase B (PI3K/Akt)

Serine/threonine-specific protein kinase B, commonly designated Akt, is a central regulator of widely divergent cellular processes, including proliferation, differentiation, migration, survival, and metabolism [12, 13]. Akt is activated by a variety of stimuli, through growth factor receptors, in a PI3K-dependent manner [12, 13]. Frequently in human cancer, normal signaling along the Akt/PKB/phosphatase, and tensin homolog (PTEN) pathway is disrupted [14]. Akt plays important roles in development, progression, and resistance to chemotherapy in cells [12, 13]. Blocking Akt signaling can mediate apoptosis and inhibit the growth of tumor cells in vitro [14]. GA inhibits Akt activation, which leads to inhibition of tumor cell proliferation and survival [15–19].

Table 15.1 A list of molecular targets of Gambogic acid

Transcription factor
Nuclear factor—kappa B ↓
STAT-3 - ↓
STAT-5 ↓
Inflammatory cytokines
IL-6 ↓
Tumor necrosis factor alpha↓
Enzymes
Cyclooxygenase-2 ↓
Inducible nitric oxide synthase↓
Matrix metalloproteinase↓
Src homology 2 domain-containing tyrosine phosphate 2↓
Kinases
Focal adhesion kinase↓
Janus kinase ↓
Mitogen-activated protein kinase ↓
Protein kinase A↓
Protein kinase B↓
Protein kinase C↓
Growth factors
Vascular endothelial growth factor ↓
Receptors
Chemokine (C-X-C motif) receptor 4 ↓
Transferrin receptor ↓
Adhesion molecules
Endothelial leukocyte adhesion molecule-1
Intracellular adhesion molecule-1 ↓
Anti-apoptotic proteins
B-cell lymphoma protein-2 ↓
Bcl-xL ↓
Inhibitory apoptosis protein-1 ↓
Mcl-1 ↓
Survivin ↓
Others
Cyclin D1 ↓
Heat shock protein 90 ↓
Heat shock protein 70 ↑

15.3.3 GA Inhibits Mitogen-Activated Protein Kinase (MAPK)

MAPKs are evolutionarily conserved enzymes that play a key role in the inflammatory stimuli and environmental stresses that lead to the activation of three independent pathways: p44/42 MAPK extracellular signal-regulated kinases 1 and 2 (ERK1/ERK2), c-Jun N-terminal kinase, and p38 MAPK [20]. In vitro studies of several cancer cells showed that GA inhibits MAPK pathways [21]. Moreover, this phytochemical also inhibited ERK in HT-29, HepG2, KBM5, and NCI-H460 cancer cells [22–24].

15.3.4 GA Inhibits Src

The Src family of proteins consists of eight non-receptor tyrosine kinases characterized by a common structure [25]. Src kinases are involved in signal transduction pathways that are triggered by a variety of surface receptors, including receptors for tyrosine kinases, integrin, and antigens, as well as receptors coupled with the G-protein [25, 26]. As a consequence of changes observed in protein expression and kinase activity in cancer cells, the Src family has been implicated in the development of cancer [25, 26]. This prompted the design of specific inhibitors, the most common of which are adenine mimetics, to treat solid tumors and leukemia clinically [25, 26]. In addition, some of the Src kinases expressed in hematopoietic cells play pivotal roles in lymphocyte maturation and activation [25]. This finding encouraged the development of safe and effective Src-specific inhibitors that are currently in clinical trials as immune-suppressants for the treatment of immunological disorders [15, 27]. Separate research showing that GA inhibits Src in PC3, and K562 cells suggests that GA may also have clinical potential against cancers and immunological disorders in which Src plays a pivotal role [15, 16].

15.3.5 GA Inhibits Signal Transducer and Activator of Transcription-3 (STAT-3) Pathways

Proteins in the STAT family are among the best studied of the latent cytoplasmic signal-dependent transcription factors [28–30]. In vitro studies of the MCF-7, MCF-10A, U266, and MM1.s cell lines suggest that GA modulates the nuclear translocation and DNA binding of STAT3 and inhibits genes modulated by this transcription factor [28, 31].

15.3.6 GA Inhibits Chemokine X-Receptor 4 (CXCR4) and Downstream Signaling Pathways

Chemokine receptors belong to class A seven transmembrane G-protein-coupled receptors and consist of 350 amino acids on average. Primary receptors are defined as CXCR, CCR, CR, or CX3CR [32]. The function of atypical chemokine receptors is to modulate immune responses by scavenging, sequestration, buffering as well as intracellular transport of chemokines from inflammatory sites [33–35]. The receptor CXCR4 is expressed on almost all of the hematopoietic cells, embryonic pluripotent, and tissue-committed stem cells, allowing them to migrate and invade along CXCL12 gradients. In malignant cells the chemokine receptor that is most commonly found is the receptor CXCR4 [34, 36]. At least 23 different types of tumor cells from human cancers of epithelial, mesenchymal and hematopoietic origin express CXCR4 [37]. In cancer, CXCL12 plays a role in the mobilization and recruitment of these cells to the inflammatory tumor microenvironment, neo-angiogenic niches, supporting revascularization, tumor growth, and metastasis [32, 38]. Thus an inhibitor of CXCR4/CXCL12 axis will inhibit tumor metastasis, GA is one of those inhibitors. Recently, we showed that GA directly interacts with CXCR4 and inhibits the migration of multiple myeloma cells [17]. We further demonstrate that GA inhibits CXCR4 regulated pathways and suppresses the bone loss [17]. Overall, GA has a tremendous potential to be used a therapeutic agent.

15.3.7 GA Inhibits CBP/p300 Histone Acetyltransferase (HAT) and Histone Deacetylase (HDAC)

The process of histone acetylation and deacetylation in eukaryotic cells alters chromatin structure and thereby modulates gene expression [39]. HATs and HDACs are classes of enzymes that effect histone acetylation [40]. These enzymes can also acetylate and deacetylate several nonhistone substrates, which can have functional consequences. Altered HAT and HDAC activities can lead to several diseases, ranging from cancer to neurodegenerative disorders. Therefore, HAT and HDAC inhibitors are being developed as therapeutic agents. GA inhibits HAT and HDAC activity in A549 lung cancer cells [41]. These activities of GA demonstrate its great potential as a therapeutic candidate.

15.3.8 GA Inhibits the Activation of Focal Adhesion Kinase (FAK)

FAK is a 119- to 121-kDa non-receptor protein kinase widely expressed in various tissues and cell types [42]. Several studies showed that FAK plays an important role

in integrin signaling [43–45]. Once activated, whether by integrin or non-integrin stimuli, FAK binds to and activates several other molecules, such as Src, Src adaptor protein p130Cas, the growth factor receptor-bound protein 2 (Grb2), PI3K, and paxillin, and thus promotes signaling transduction [44, 46–50]. In a recent study, FAK was held responsible for uninhibited proliferation, protection from apoptosis, invasion, migration, adhesion, and spread, as well as tumor angiogenesis [46, 47]. Our group showed that GA modulates the tyrosine phosphorylation of FAK and subsequently induce apoptosis by downregulating Src, ERK, and Akt signaling in prostate cancer PC3 cells [16].

15.3.9 GA Inhibits iNOS

iNOS is expressed in a variety of cell types, particularly inflammatory cells, in response to diverse pro-inflammatory stimuli [51–53]. iNOS, which may be induced by bacterial LPS or its derivative lipid A, is expressed by a variety of solid tumors and generates high levels of nitric oxide inside tumor cells [10]. In vitro studies showed that GA inhibits LPS- and interferon-gamma-induced iNOS in RAW246.6 cells [10].

15.3.10 GA Induces the Production of Reactive Oxygen Species (ROS)

ROS have been linked with various cell signaling pathways [54, 55]. GA, induces the production of ROS [56].

15.3.11 GA Inhibits COX-2

Overexpression of COX-2 is associated with many cancers and is linked with tumor cell proliferation and suppression of apoptosis [57, 58]. Therefore, COX-2 inhibitors have great potential in the treatment of cancers and inflammatory conditions, as evidenced by the U. S. Food and Drug Administration's approval of celecoxib, a known COX-2 inhibitor, for the treatment of various inflammatory conditions [59]. GA, too, has been shown to inhibit COX-2 activation induced by TNF- α in KBM5 leukemic cells [7].

15.3.12 GA Inhibits Matrix Metalloproteinase 7 & 9 (MMP-7 & 9)

Also known as matrilysin, MMP-7 is a “minimal domain” MMP that exhibits proteolytic activity against components of the extracellular matrix [60–62]. MMP-7 is frequently overexpressed in human cancer tissues and is associated with cancer progression [63]. Therefore, MMP-7 inhibitors have great potential in the treatment of cancer [64]. The studies showed that GA inhibits the expression of MMP-7 in breast cancer cells like MDA-MB-231 and MDA-MB-435 [65, 66], further supporting the idea that GA may be effective against breast cancer in humans.

15.3.13 GA Inhibits Tubulin

Microtubules are a major component of the cytoskeleton. They are important in many cellular events and play a crucial role in cell division [67]. As such, microtubules are a highly attractive target for anticancer-drug design. Tubulin-binding agents, also called anti-microtubule or microtubule-targeted agents, are widely used chemotherapeutic drugs with a proven clinical efficacy against breast, lung, ovarian, prostate, and hematologic malignancies, as well as childhood cancers [68, 69]. Research has shown that GA belongs to this class of agents because it inhibits microtubule assembly and prevents cell division [20].

15.3.14 GA Inhibits Expression of Cyclin D1

The sequential transcriptional activation of cyclins, the regulatory subunits of cell cycle-specific kinases, is thought to regulate progress through the cell cycle [70]. Thus, cyclins are potential oncogenes, and overexpression of cyclin D1 or amplification at its genomic locus, 11q13, is commonly seen in breast cancer, head, and neck cancer, non-small-cell lung cancer, and mantle cell lymphoma [71, 72]. GA has been shown to inhibit Cyclin D1 in several cancers including leukemia and multiple myeloma [7, 31].

15.3.15 GA Induces Cleavage of Poly(ADP-Ribose) Polymerases (PARPs)

PARPs are cell signaling enzymes present in eukaryotes and are involved in poly(ADP ribosylation) of DNA-binding proteins [73]. Pharmacological degradation of PARP-1 may enhance the activity of antitumor drugs by inhibiting necrosis and

activating apoptosis [74]. In vitro studies have shown that GA induces PARP degradation and enhances apoptosis in T98 glioma, HeLa, non-small lung cancer A549 and NCI-H460, breast cancer, and multiple myeloma cells [7, 11, 17, 22, 24, 31, 66, 75–77].

15.3.16 GA Inhibits Tumor Necrosis Factor- α (TNF- α)

TNF- α is a vital member of the multifunctional superfamily of TNFs and plays important roles in immunity and cellular remodeling, as well as apoptosis and cell survival [78]. Because TNF- α is a key player in inflammation and cancer, several efforts are underway to develop therapeutic TNF- α antagonists. Two such antagonists are from the *Garcinia* species. At a dose of 5 μ M, both GA and cambogin inhibited the release of TNF- α by LPS-activated macrophages [79, 80], suggesting another mechanism for their antitumor activity.

15.3.17 GA Inhibits the Expression of Bcl-2 Family Proteins

The bcl-2 gene family consists of at least 25 genes that are proapoptotic or anti-apoptotic and share at least one of the four characteristic BH domains [81]. The anti-apoptotic protein bcl-2, which displays sequence homology in all four domains (i.e., BH1–BH4), promotes cell survival [82]. Increased expression of the bcl-2 protein commonly occurs in human malignancies and is associated with disease maintenance and progression, resistance to chemotherapy, and poor clinical outcome. Antisense oligonucleotides targeting bcl-2 have been shown to facilitate apoptosis in various tumor types [83]. Therefore, bcl-2 inhibitors have great potential in the treatment of cancer. In vitro and in vivo studies showed that GA inhibits bcl-2 expression in MGC-803, HL-60, MCF-7, A375M, SMMC-7721, BGC-823, Jeko-1, and K562 [15, 84–88].

15.3.18 GA Induces BID

Pro-apoptotic BID activates the multi-domain bcl-2 family members bcl-2-associated X protein (BAX) and bcl-2 homologous antagonist killer (BAK) [89]. Activation of either BAX or BAK produces an allosteric conformational change and releases cytochrome *c* [90]. This means that compounds that can induce BID could be very useful in the treatment of cancer. GA and its derivative GA3 are such inducers because these agents activate BID and induces apoptosis in cancer cells [91].

15.3.19 GA Induces BAD

BAD is proapoptotic and proliferative, suggesting that the cell cycle functions of the multi-domain bcl-2 family members [89]. BAD antagonizes both the cell cycle and anti-apoptotic functions of bcl-2 and bcl-xL through BH3 binding [89]. Overexpression of the BH3-only molecule BAD renders the cell unable to arrest in G0 and persistently activates cdk2 [92]. Previous study showed that GA in combination with nanoparticle Fe₃O₄ activates BAD and induces apoptosis in LOVO cells [93].

15.3.20 GA Inhibits Cytochrome c

Cytochrome *c*, an intermediate in apoptosis, is released by the mitochondria in response to proapoptotic stimuli. The studies have shown that GA, induces the expression of cytochrome *c* in colorectal cancer HT-29, bladder cancer T24 and UMUC3, breast cancer MDA-MB-231, and human hepatocellular carcinoma cells [94–96].

15.3.21 GA Induces the Activation of Caspase-3 and Caspase-9

Caspases play a central role in mediating various apoptotic responses. In vitro and in vivo research of GA has shown that it induces the activation of caspase-3 and caspase-9 in various cancer cells including glioma, osteosarcoma, non-small lung cancer, leukemia, lymphoma, breast cancer, pancreatic cancer, melanoma and multiple myeloma and, induces apoptosis [7, 11, 17, 22, 24, 31, 66, 75–77].

15.4 Role of GA in Chronic Diseases

Extensive studies from past one decade have shed light on GA's potential as anti-inflammatory and anticancer agents. So far the focus of the studies have been to identify the molecular targets by which GA exerts its effects, primarily on cancer cells. However, a very recent study showed that GA could be used as an anti-psoriatic agent [97]. Importantly, the molecular mechanism by which GA mediates its effect strongly suggest that it could be used for the prevention and treatment of many organ and tissue disorders, which are associated with inflammation and oxidative stress. GA alleviates oxidative stress, inflammation in chronic diseases and regulates inflammatory and pro-inflammatory pathways related with most chronic diseases.

15.4.1 Cardiovascular Diseases

Cardiovascular Diseases (CVDs), including heart disease, vascular disease and atherosclerosis, are the most critical current global health threat. Epidemiological and clinical trials have shown strong consistent relationships between the inflammation markers and risk of cardiovascular diseases [98]. It is widely appreciated that the key mechanisms in the development of CVDs are inflammation and oxidant stress, activation of pro-inflammatory cytokines, chronic transmural inflammation, and C reactive protein (CRP) [99]. Thus cytokines, other bioactive molecules, and cells that are characteristic of inflammation are believed to be involved in atherogenesis. An elegant recent study by Liu et al. [100] showed that GA inhibits pressure overload or isoproterenol infusion-induced cardiac hypertrophy and fibrosis, through the inhibition of the proteasome and the NF- κ B pathway, suggesting that GA treatment may provide a new strategy to treat cardiac hypertrophy and changes in myocardial NF- κ B signaling [100].

15.4.2 Rheumatoid Arthritis

Rheumatoid arthritis (RA) could give rise to a systemic chronic inflammatory disorder and may impact many organs and tissues but mainly attack flexible (synovial) joints [101]. It was reported that oxidative stress made an important contribution to joint destruction in RA [102]. ROS is a significant mediator that activates a variety of transcription factors including NF- κ B and AP-1, thus regulating the expression of over 500 different genes, such as growth factors, chemokines, cell cycle regulatory molecules, inflammatory cytokines, and anti-inflammatory molecules [103]. Therefore, transcription factors and genes, involved in inflammation and antioxidation, are suspected to play a crucial adjective function in RA. The main treatment of RA is to reduce arthritis reaction, inhibit disease development and irreversible bone destruction, protect the joints and muscle function, and ultimately achieve complete remission or low disease activity. Treatment principles include patient education, early treatment, and combination therapy [104, 105]. Drug therapy includes nonsteroidal anti-inflammatory drugs (NSAIDs), slow-acting antirheumatic drugs, immunosuppressive agents, immune and biological agents, and botanicals. NSAIDs are most common. Our earlier studies strongly suggest that GA is one of the NSAIDs with anti-inflammatory and antioxidant actions both in vivo and in vitro and could be used effectively as anti-RA agent. Recent studies of Cascao et al. [106] support our hypothesis. By using rat RA model, this group showed that GA inhibits RA by inhibiting the levels of cytokines and key inflammatory molecules [106].

15.4.3 Diabetes and Obesity

Type 2 diabetes is a chronic disease where cells have reduced insulin signaling, leading to hyperglycemia, and long-term complications, such as heart, kidney, and liver disease. Recently, more and more studies have shown the critical roles of oxidative stress and inflammatory reactions in the pathogenesis of diabetes. Studies have shown that AMP-activated protein kinase (AMPK) plays a key role in maintaining intracellular and whole-body energy homeostasis. Activation of AMPK has been shown to ameliorate the symptoms of type 2 diabetes and obesity. In vitro studies by Zhao et al. [107] demonstrate that GA, activates AMPK by increasing the phosphorylation of AMPK α and its downstream substrate ACC in various cell lines [107]. This group also showed that GA induced activation of AMPK was associated with increased intracellular ROS level. Collectively, these results suggest that GA may be a novel direct activator of AMPK and could be used as anti-diabetic agent. However, further studies are required to fully evaluate this function of GA.

15.4.4 Psoriasis

Psoriasis is a chronic inflammatory skin disease characterized by thick, red, and scaly lesions on any part of the body, which affects approximately 2 % of the population worldwide [108]. Many cytokines, including interleukin-23(IL-23), IL-17A, TNF- α , IL-6, IL-1 β , and IL-22, are also involved in the pathogenesis of psoriasis [109]. Along these lines a recent study showed that GA could be used as an anti-psoriatic agent [97].

15.4.5 Cancer

Inflammation plays key roles in all the ways of tumorigenesis and therapy response [110]. Activation and interaction between STAT3 and NF- κ B are very vital in the control of cancer cells and inflammatory cells [111]. TNF- α , VEGF, IL-10, MMP-2 and MMP-9, MCP, CD4+ T, AP-1, Akt, PPAR- γ , MAP kinases, and mTORC1 are also important linking factors between inflammation and cancer [111]. It has been shown that GA suppresses the growth of various cancer cells such as non-small cell lung cancer [112], human hepatocellular carcinoma [113], oral squamous cell carcinoma [114], human breast cancer [86], human malignant melanoma [115], human gastric carcinoma [116], and human leukemia cancer [7] and multiple myeloma [31, 117]. A variety of mechanisms have been proposed by which GA inhibits the proliferation of cancer cells and induces apoptosis. These include inhibition of antiapoptotic proteins Bcl-2 [86, 88] and survivin [118]; induction of apoptosis-associated proteins p53 [119], bax, and procaspase-3 [115]; activation of

c-jun-NH2-kinase, p38 [20], and GSK-3 β [15]; inhibition of topoisomerase II by binding to its ATPase domain [120], downregulation of the MDM2 oncogene and subsequent induction of p21 [119]; suppression of LPS induced COX-2 [10]; and downregulation of human telomerase reverse transcriptase [121]. It has also been shown that GA directly binds to *c-myc* [122], transferrin receptors [123], and CXCR4 [124]. Recently, a proteomic approach revealed that GA suppresses expression of 14-3-3 protein sigma and stathmin [116]. We have shown earlier that GA inhibits NF- κ B and its regulated gene products in human myeloid leukemia [7]; STAT3 and its regulated gene products in MM [31]. Most recently, we showed that GA interacts with CXCR4 and inhibits chemotaxis and osteoclastogenesis in MM [124]. Recently, it is shown that GA is a novel tissue specific proteasome inhibitor, with potency comparable to bortezomib [25]. In addition, recent studies have shown that GA is bioavailable, less toxic, effective, and inhibits development of tumors in animal models, and most importantly it has been approved for phase 2 clinical trial in solid tumors [4, 121, 125, 126]. Since, GA modulates the expression of proteins plays important role in survival, migration, invasion and chemoresistance of multiple myeloma cells (Fig. 15.2), we have been working on the development of GA as anti-myeloma agent.

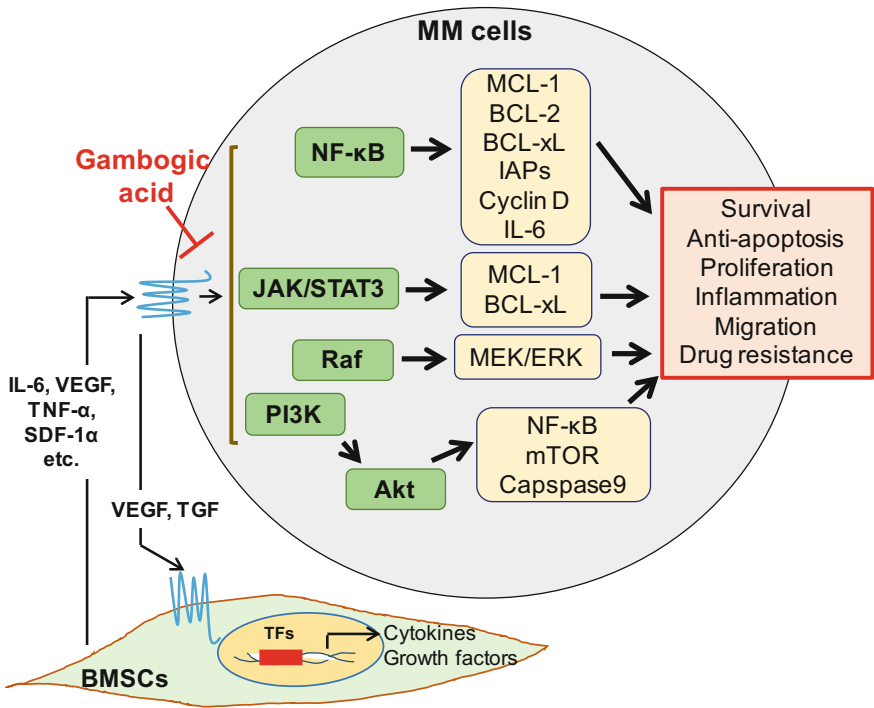


Fig. 15.2 Intimate relationship between multiple myeloma and bone marrow microenvironment. Bone marrow stromal cells secrete cytokine a growth factors, these growth factors activates several pathways in multiple myeloma. Gambogic acid targets these pathways

15.5 Biological Activities of GA in Animal Models

Besides the extensive *in vitro* demonstrations of GA's anti-proliferative effects, numerous other studies have evaluated its efficacy in various animal models *in vivo* (Table 15.2). The *in vivo* studies have investigated the effects of GA on tumor angiogenesis and the biomarkers COX-2 and VEGF in prostate carcinoma [16]. One group demonstrated that systemic administration of GA for 3 weeks to athymic mice bearing non-small lung NCI-H1993 xenografts significantly inhibited tumor growth [127]. Meanwhile, others have shown that GA can suppress the growth of cervical carcinoma [128], modulate the growth of colorectal cancer [129], modulate the growth of prostate cancer in rodents [16], inhibit the growth of human B-cell lymphoma by inducing proteasome inhibition [130] in nude mice, and inhibits hepatocellular carcinoma, multiple myeloma, bladder cancer tumor growth, in part by suppressing angiogenesis, and inducing apoptosis [7, 11, 17, 22, 24, 31, 66, 75–77]. More recent studies have evaluated GA's chemosensitizing effects [118]. Our group evaluated the chemosensitizing effect of GA in combination with paclitaxel, TNF- α , 5-FU on multiple myeloma [7]. Together, these *in vivo* animal studies clearly suggest GA's anticancer potential when administered either alone or in combination with currently employed chemotherapeutic agents.

Table 15.2 A list of studies describing antitumor effects of gambogic acid in animals

Tumor	Cell line	Route	Dose (mg/kg)	Model	References
Lung	SPC-A1	i.v.	4, 8	Xenograft	[121]
	NCI-H1993	i.p.	10, 20 or 30	Xenograft	[127]
	NCI-H1975	i.v.	8	Xenograft	[137]
	A549	i.p.	8, 16 and 32	Xenograft	[134]
Gastric cancer	BGC-823	i.v.	8	Xenograft	[85]
Cervical carcinoma	HeLa cells	i.p.	2	Xenograft	[128]
Chronic myeloid leukemia	KBM5	i.p.	3	Xenograft	[24]
Liver	HepG2	i.v.	1.5	Xenograft	[132]
Ovarian	SKOV3		1.0	Xenograft	[133]
Breast	MDA-MB-231	i.v.	4 and 8	Orthotopic	[94]
Hepatoma	H22	i.v.	2, 4 and 8	Xenograft	[125]
	H22	p.o.	12.5, 25, 30 and 50	Xenograft	[125]
Melanoma	SMMC-7721	i.v.	2, 4, and 8	Xenograft	[131, 136]
	B16-F10	i.v.	0.375, 0.75, and 1.5	Orthotopic	[135]
Colon	HT-29	i.v.	5, 10 and 20	Xenograft	[129]

15.6 Biological Activities of GA in Humans

There are no clinical studies so far in USA, however in China GA is in Phase II clinical trial [4, 121, 125, 126]. The plethora of studies clearly suggest GA's potential as anticancer agent. It the opportune time to seriously consider this xanthone in clinical trial especially in USA.

15.7 Conclusions

The spice derived from kokum, the fruit of *Garcinia indica*, is used in Indian cuisines and Ayurvedic medicine. The main component isolated from kokum is GA, which demonstrates thrust quencher, antioxidant, antimicrobial, antiulceration, and anticancer properties. Although GA is a potent, biologically active compound, only a number of studies are carried out in animals and none have been done in humans. Because of its diverse range of biological activity in vitro, more in vivo and clinical studies are warranted to establish its true usefulness as a clinical therapeutic agent in a variety of human diseases.

References

1. Guo Q et al (2006) Toxicological studies of gambogic acid and its potential targets in experimental animals. *Basic Clin Pharmacol Toxicol* 99(2):178–184
2. Noguer O, Villena J, Lorita J, Vilaro S, Reina M (2009) Syndecan-2 downregulation impairs angiogenesis in human microvascular endothelial cells. *Exp Cell Res* 315(5):795–808
3. Jang SW et al (2007) Gambogic amide, a selective agonist for TrkA receptor that possesses robust neurotrophic activity, prevents neuronal cell death. *Proc Natl Acad Sci USA* 104(41):16329–16334
4. Qi Q et al (2008) Studies on the toxicity of gambogic acid in rats. *J Ethnopharmacol* 117(3):433–438
5. Udvadia AJ, Linney E (2003) Windows into development: historic, current, and future perspectives on transgenic zebrafish. *Dev Biol* 256(1):1–17
6. Zhang HZ et al (2004) Discovery, characterization and SAR of gambogic acid as a potent apoptosis inducer by a HTS assay. *Bioorg Med Chem* 12(2):309–317
7. Pandey MK et al (2007) Gambogic acid, a novel ligand for transferrin receptor, potentiates TNF-induced apoptosis through modulation of the nuclear factor-kappaB signaling pathway. *Blood* 110(10):3517–3525
8. Li X et al (2013) Gambogic acid is a tissue-specific proteasome inhibitor in vitro and in vivo. *Cell Rep* 3(1):211–222
9. Wang Y et al (2014) Methyl jasmonate sensitizes human bladder cancer cells to gambogic acid-induced apoptosis through down-regulation of EZH2 expression by miR-101. *Br J Pharmacol* 171(3):618–635
10. Palempalli UD et al (2009) Gambogic acid covalently modifies IkappaB kinase-beta subunit to mediate suppression of lipopolysaccharide-induced activation of NF-kappaB in macrophages. *Biochem J* 419(2):401–409

11. Yang LJ, Chen Y (2013) New targets for the antitumor activity of gambogic acid in hematologic malignancies. *Acta Pharmacol Sin* 34(2):191–198
12. Franke TF (2008) PI3K/Akt: getting it right matters. *Oncogene* 27(50):6473–6488
13. Franke TF, Hornik CP, Segev L, Shostak GA, Sugimoto C (2003) PI3K/Akt and apoptosis: size matters. *Oncogene* 22(56):8983–8998
14. Fruman DA, Rommel C (2014) PI3K and cancer: lessons, challenges and opportunities. *Nat Rev Drug Discov* 13(2):140–156
15. Li R et al (2009) Gambogic acid induces G0/G1 arrest and apoptosis involving inhibition of SRC-3 and inactivation of Akt pathway in K562 leukemia cells. *Toxicology* 262(2):98–105
16. Yi T et al (2008) Gambogic acid inhibits angiogenesis and prostate tumor growth by suppressing vascular endothelial growth factor receptor 2 signaling. *Cancer Res* 68(6):1843–1850
17. Pandey MK et al (2014) Gambogic acid inhibits multiple myeloma mediated osteoclastogenesis through suppression of chemokine receptor CXCR4 signaling pathways. *Exp Hematol* 42(10):883–896
18. Yang Y, Sun X, Yang Y, Yang X, Zhu H, Dai S, Chen X, Zhang H, Guo Q, Song Y, Wang F, Cheng H, Sun X (2016) Gambogic acid enhances the radiosensitivity of human esophageal cancer cells by inducing reactive oxygen species via targeting Akt/mTOR pathway. *Tumour Biol* 37(2):1853–1862
19. Ma J et al (2015) Gambogic acid inhibits osteoclast formation and ovariectomy-induced osteoporosis by suppressing the JNK, p38 and Akt signalling pathways. *Biochem J* 469(3):399–408
20. Chen J et al (2008) Microtubule depolymerization and phosphorylation of c-Jun N-terminal kinase-1 and p38 were involved in gambogic acid induced cell cycle arrest and apoptosis in human breast carcinoma MCF-7 cells. *Life Sci* 83(3–4):103–109
21. Lu N et al (2007) Gambogic acid inhibits angiogenesis through suppressing vascular endothelial growth factor-induced tyrosine phosphorylation of KDR/Flk-1. *Cancer Lett* 258(1):80–89
22. Wang LH et al (2014) Gambogic acid synergistically potentiates cisplatin-induced apoptosis in non-small-cell lung cancer through suppressing NF- κ B and MAPK/HO-1 signalling. *Br J Cancer* 110(2):341–352
23. Yan F et al (2012) Gambogic acid induced mitochondrial-dependent apoptosis and referred to phospho-Erk1/2 and phospho-p38 MAPK in human hepatoma HepG2 cells. *Environ Toxicol Pharmacol* 33(2):181–190
24. Shi X et al (2014) Gambogic acid induces apoptosis in imatinib-resistant chronic myeloid leukemia cells via inducing proteasome inhibition and caspase-dependent Bcr-Abl downregulation. *Clin Cancer Res* 20(1):151–163
25. Parsons SJ, Parsons JT (2004) Src family kinases, key regulators of signal transduction. *Oncogene* 23(48):7906–7909
26. Benati D, Baldari CT (2008) SRC family kinases as potential therapeutic targets for malignancies and immunological disorders. *Curr Med Chem* 15(12):1154–1165
27. Aleshin A, Finn RS (2010) SRC: a century of science brought to the clinic. *Neoplasia* 12(8):599–607
28. Yu H, Jove R (2004) The STATs of cancer—new molecular targets come of age. *Nat Rev Cancer* 4(2):97–105
29. Turkson J, Jove R (2000) STAT proteins: novel molecular targets for cancer drug discovery. *Oncogene* 19(56):6613–6626
30. Cleveland CV (2004) Roles and regulation of stat family transcription factors in human breast cancer. *Am J Pathol* 165(5):1449–1460
31. Prasad S, Pandey MK, Yadav VR, Aggarwal BB (2011) Gambogic acid inhibits STAT3 phosphorylation through activation of protein tyrosine phosphatase SHP-1: potential role in proliferation and apoptosis. *Cancer Prev Res* 4(7):1084–1094
32. Pandey MK, Rastogi S, Kale VP, Gowda T, Amin SG (2014) Targeting CXCL12/CXCR4 axis in multiple myeloma. *J Hematol Thrombo Dis* 2:159

33. Bachelierie F et al (2014) International Union of Basic and Clinical Pharmacology. [corrected]. LXXXIX. Update on the extended family of chemokine receptors and introducing a new nomenclature for atypical chemokine receptors. *Pharmacol Rev* 66 (1):1–79
34. Murphy PM et al (2000) International union of pharmacology. XXII. Nomenclature for chemokine receptors. *Pharmacol Rev* 52(1):145–176
35. Hansell CA, Hurson CE, Nibbs RJ (2011) DARC and D6: silent partners in chemokine regulation? *Immunol Cell Biol* 89(2):197–206
36. Nakayama T et al (2003) Cutting edge: profile of chemokine receptor expression on human plasma cells accounts for their efficient recruitment to target tissues. *J Immunol* 170(3):1136–1140
37. Balkwill F (2004) Cancer and the chemokine network. *Nat Rev Cancer* 4(7):540–550
38. Teicher BA, Fricker SP (2010) CXCL12 (SDF-1)/CXCR4 pathway in cancer. *Clin Cancer Res* 16(11):2927–2931
39. Yang XJ, Seto E (2007) HATs and HDACs: from structure, function and regulation to novel strategies for therapy and prevention. *Oncogene* 26(37):5310–5318
40. Legube G, Trouche D (2003) Regulating histone acetyltransferases and deacetylases. *EMBO Rep* 4(10):944–947
41. Qi Q et al (2015) Involvement of RECK in gambogic acid induced anti-invasive effect in A549 human lung carcinoma cells. *Mol Carcinog* 54(Suppl 1):E13–E25
42. Abbi S, Guan JL (2002) Focal adhesion kinase: protein interactions and cellular functions. *Histol Histopathol* 17(4):1163–1171
43. Guan JL (2010) Integrin signaling through FAK in the regulation of mammary stem cells and breast cancer. *IUBMB Life* 62(4):268–276
44. Mitra SK, Schlaepfer DD (2006) Integrin-regulated FAK-Src signaling in normal and cancer cells. *Curr Opin Cell Biol* 18(5):516–523
45. Guan JL (1997) Role of focal adhesion kinase in integrin signaling. *Int J Biochem Cell Biol* 29(8–9):1085–1096
46. Sulzmaier FJ, Jean C, Schlaepfer DD (2014) FAK in cancer: mechanistic findings and clinical applications. *Nat Rev Cancer* 14(9):598–610
47. Tai YL, Chen LC, Shen TL (2015) Emerging roles of focal adhesion kinase in cancer. *BioMed Res Int* 2015:690690
48. You D et al (2015) FAK mediates a compensatory survival signal parallel to PI3K-AKT in PTEN-null T-ALL cells. *Cell Rep* 10(12):2055–2068
49. Hu YL et al (2014) FAK and paxillin dynamics at focal adhesions in the protrusions of migrating cells. *Sci Rep* 4:6024
50. Schlaepfer DD, Jones KC, Hunter T (1998) Multiple Grb2-mediated integrin-stimulated signaling pathways to ERK2/mitogen-activated protein kinase: summation of both c-Src- and focal adhesion kinase-initiated tyrosine phosphorylation events. *Mol Cell Biol* 18(5):2571–2585
51. Janakiram NB, Rao CV (2012) iNOS-selective inhibitors for cancer prevention: promise and progress. *Future Med Chem* 4(17):2193–2204
52. Kostourou V et al (2011) The role of tumour-derived iNOS in tumour progression and angiogenesis. *Br J Cancer* 104(1):83–90
53. Lechner M, Lirk P, Rieder J (2005) Inducible nitric oxide synthase (iNOS) in tumor biology: the two sides of the same coin. *Semin Cancer Biol* 15(4):277–289
54. Finkel T (2011) Signal transduction by reactive oxygen species. *J Cell Biol* 194(1):7–15
55. Schieber M, Chandel NS (2014) ROS function in redox signaling and oxidative stress. *CB* 24 (10):R453–R462
56. Geng J, Xiao S, Zheng Z, Song S, Zhang L (2013) Gambogic acid protects from endotoxin shock by suppressing pro-inflammatory factors in vivo and in vitro. *Inflammation research: official journal of the European Histamine Research Society...* [et al.] 62(2):165–172
57. Stasinopoulos I, Shah T, Penet MF, Krishnamachary B, Bhujwalla ZM (2013) COX-2 in cancer: Gordian knot or Achilles heel? *Front Pharmacol* 4:34

58. Greenhough A et al (2009) The COX-2/PGE2 pathway: key roles in the hallmarks of cancer and adaptation to the tumour microenvironment. *Carcinogenesis* 30(3):377–386
59. Tindall E (1999) Celecoxib for the treatment of pain and inflammation: the preclinical and clinical results. *J Am Osteopath Assoc* 99(11 Suppl):S13–S17
60. Page-McCaw A, Ewald AJ, Werb Z (2007) Matrix metalloproteinases and the regulation of tissue remodelling. *Nat Rev Mol Cell Biol* 8(3):221–233
61. Parks WC, Wilson CL, Lopez-Boado YS (2004) Matrix metalloproteinases as modulators of inflammation and innate immunity. *Nat Rev Immunol* 4(8):617–629
62. Egeblad M, Werb Z (2002) New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* 2(3):161–174
63. Gialeli C, Theocharis AD, Karamanos NK (2011) Roles of matrix metalloproteinases in cancer progression and their pharmacological targeting. *FEBS J* 278(1):16–27
64. Shay G, Lynch CC, Fingleton B (2015) Moving targets: emerging roles for MMPs in cancer progression and metastasis. *Matrix Biol* 44–46:200–206
65. Qi Q et al (2008) Involvement of matrix metalloproteinase 2 and 9 in gambogic acid induced suppression of MDA-MB-435 human breast carcinoma cell lung metastasis. *J Mol Med* 86(12):1367–1377
66. Qi Q et al (2008) Anti-invasive effect of gambogic acid in MDA-MB-231 human breast carcinoma cells. *Biochem Cell Biol* 86(5):386–395
67. Etienne-Manneville S (2010) From signaling pathways to microtubule dynamics: the key players. *Curr Opin Cell Biol* 22(1):104–111
68. Dumontet C, Jordan MA (2010) Microtubule-binding agents: a dynamic field of cancer therapeutics. *Nat Rev Drug Discov* 9(10):790–803
69. Jordan MA, Wilson L (2004) Microtubules as a target for anticancer drugs. *Nat Rev Cancer* 4(4):253–265
70. Hochegger H, Takeda S, Hunt T (2008) Cyclin-dependent kinases and cell-cycle transitions: does one fit all? *Nat Rev Mol Cell Biol* 9(11):910–916
71. Musgrove EA, Caldon CE, Barraclough J, Stone A, Sutherland RL (2011) Cyclin D as a therapeutic target in cancer. *Nat Rev Cancer* 11(8):558–572
72. Hosokawa Y, Arnold A (1998) Mechanism of cyclin D1 (CCND1, PRAD1) overexpression in human cancer cells: analysis of allele-specific expression. *Genes Chromosom Cancer* 22(1):66–71
73. Rouleau M, Patel A, Hendzel MJ, Kaufmann SH, Poirier GG (2010) PARP inhibition: PARP1 and beyond. *Nat Rev Cancer* 10(4):293–301
74. Helleday T, Petermann E, Lundin C, Hodgson B, Sharma RA (2008) DNA repair pathways as targets for cancer therapy. *Nat Rev Cancer* 8(3):193–204
75. Krajarng A et al (2015) Apoptosis induction associated with the ER stress response through up-regulation of JNK in HeLa cells by gambogic acid. *BMC Complement Altern Med* 15:26
76. Thida M, Kim DW, Tran TT, Pham MQ, Lee H, Kim I, Lee JW (2016) Gambogic acid induces apoptotic cell death in T98G glioma cells. *Bioorg Med Chem Lett* 26(3):1097–1101
77. Yang LJ et al (2012) Effects of gambogic acid on the activation of caspase-3 and downregulation of SIRT1 in RPMI-8226 multiple myeloma cells via the accumulation of ROS. *Oncol Lett* 3(5):1159–1165
78. Wang X, Lin Y (2008) Tumor necrosis factor and cancer, buddies or foes? *Acta Pharmacol Sin* 29(11):1275–1288
79. Lee JY, Lee BH, Lee JY (2015) Gambogic acid disrupts toll-like receptor4 activation by blocking lipopolysaccharides binding to myeloid differentiation factor 2. *Toxicol Res* 31(1):11–16
80. Liao CH, Sang S, Liang YC, Ho CT, Lin JK (2004) Suppression of inducible nitric oxide synthase and cyclooxygenase-2 in downregulating nuclear factor-kappa B pathway by Garcinol. *Mol Carcinog* 41(3):140–149
81. Youle RJ, Strasser A (2008) The BCL-2 protein family: opposing activities that mediate cell death. *Nat Rev Mol Cell Biol* 9(1):47–59

82. Juin P, Geneste O, Gautier F, Depil S, Campone M (2013) Decoding and unlocking the BCL-2 dependency of cancer cells. *Nat Rev Cancer* 13(7):455–465
83. Gleave ME, Monia BP (2005) Antisense therapy for cancer. *Nat Rev Cancer* 5(6):468–479
84. Xu J et al (2013) Gambogic acid induces mitochondria-dependent apoptosis by modulation of Bcl-2 and Bax in mantle cell lymphoma JeKo-1 cells. *Chin J Cancer Res* 25(2):183–191
85. Liu W et al (2005) Anticancer effect and apoptosis induction of gambogic acid in human gastric cancer line BGC-823. *World J Gastroenterol* 11(24):3655–3659
86. Gu H et al (2009) Gambogic acid reduced bcl-2 expression via p53 in human breast MCF-7 cancer cells. *J Cancer Res Clin Oncol* 135(12):1777–1782
87. Zhai D et al (2008) Gambogic acid is an antagonist of antiapoptotic Bcl-2 family proteins. *Mol Cancer Ther* 7(6):1639–1646
88. Zhao L, Guo QL, You QD, Wu ZQ, Gu HY (2004) Gambogic acid induces apoptosis and regulates expressions of Bax and Bcl-2 protein in human gastric carcinoma MGC-803 cells. *Biol Pharm Bull* 27(7):998–1003
89. Czabotar PE, Lessene G, Strasser A, Adams JM (2014) Control of apoptosis by the BCL-2 protein family: implications for physiology and therapy. *Nat Rev Mol Cell Biol* 15(1):49–63
90. Ma SB et al (2014) Bax targets mitochondria by distinct mechanisms before or during apoptotic cell death: a requirement for VDAC2 or Bak for efficient Bax apoptotic function. *Cell Death Differ* 21(12):1925–1935
91. Xie H et al (2009) GA3, a new gambogic acid derivative, exhibits potent antitumor activities in vitro via apoptosis-involved mechanisms. *Acta Pharmacol Sin* 30(3):346–354
92. Zinkel S, Gross A, Yang E (2006) BCL2 family in DNA damage and cell cycle control. *Cell Death Differ* 13(8):1351–1359
93. Fang L et al (2012) Synergistic effect of a combination of nanoparticulate Fe₃O₄ and gambogic acid on phosphatidylinositol 3-kinase/Akt/Bad pathway of LOVO cells. *Int J Nanomed* 7:4109–4118
94. Li C et al (2012) Gambogic acid promotes apoptosis and resistance to metastatic potential in MDA-MB-231 human breast carcinoma cells. *Biochem Cell Biol* 90(6):718–730
95. Ishaq M et al (2014) Gambogic acid induced oxidative stress dependent caspase activation regulates both apoptosis and autophagy by targeting various key molecules (NF-kappaB, Beclin-1, p62 and NBR1) in human bladder cancer cells. *Biochim Biophys Acta* 1840(12):3374–3384
96. Tang C et al (2009) Downregulation of survivin and activation of caspase-3 through the PI3K/Akt pathway in ursolic acid-induced HepG2 cell apoptosis. *Anticancer Drugs* 20(4):249–258
97. Wen J et al (2014) Gambogic acid exhibits anti-psoriatic efficacy through inhibition of angiogenesis and inflammation. *J Dermatol Sci* 74(3):242–250
98. Costa S, Reina-Couto M, Albino-Teixeira A, Sousa T (2016) Statins and oxidative stress in chronic heart failure. *Rev Port J Cardiol* 35(1):41–57
99. Urbietta Caceres VH et al (2011) Early experimental hypertension preserves the myocardial microvasculature but aggravates cardiac injury distal to chronic coronary artery obstruction. *Am J Physiol Heart Circ Physiol* 300(2):H693–H701
100. Liu S et al (2013) Gambogic acid suppresses pressure overload cardiac hypertrophy in rats. *Am J Cardiovasc Dis* 3(4):227–238
101. McInnes IB, Schett G (2011) The pathogenesis of rheumatoid arthritis. *N Engl J Med* 365(23):2205–2219
102. Wruck CJ et al (2011) Role of oxidative stress in rheumatoid arthritis: insights from the Nrf2-knockout mice. *Ann Rheum Dis* 70(5):844–850
103. Ray PD, Huang BW, Tsuji Y (2012) Reactive oxygen species (ROS) homeostasis and redox regulation in cellular signaling. *Cell Signal* 24(5):981–990
104. Kahlenberg JM, Fox DA (2011) Advances in the medical treatment of rheumatoid arthritis. *Hand Clin* 27(1):11–20
105. Forestier R et al (2009) Non-drug treatment (excluding surgery) in rheumatoid arthritis: clinical practice guidelines. *Joint Bone Spine* 76(6):691–698

106. Cascao R et al (2014) Potent anti-inflammatory and antiproliferative effects of gambogic acid in a rat model of antigen-induced arthritis. *Mediat Inflamm* 2014:195327
107. Zhao B, Shen H, Zhang L, Shen Y (2012) Gambogic acid activates AMP-activated protein kinase in mammalian cells. *Biochem Biophys Res Commun* 424(1):100–104
108. Gupta MA, Simpson FC, Gupta AK (2015) Psoriasis and sleep disorders: a systematic review. *Sleep Med Rev* 29:63–75
109. Coimbra S, Figueiredo A, Castro E, Rocha-Pereira P, Santos-Silva A (2012) The roles of cells and cytokines in the pathogenesis of psoriasis. *Int J Dermatol* 51(4):389–395; quiz 395–388
110. Aggarwal BB, Shishodia S, Sandur SK, Pandey MK, Sethi G (2006) Inflammation and cancer: how hot is the link? *Biochem Pharmacol* 72(11):1605–1621
111. Grivennikov SI, Karin M (2010) Dangerous liaisons: STAT3 and NF-kappaB collaboration and crosstalk in cancer. *Cytokine Growth Factor Rev* 21(1):11–19
112. Zhu X et al (2009) Mechanisms of gambogic acid-induced apoptosis in non-small cell lung cancer cells in relation to transferrin receptors. *J Chemother* 21(6):666–672
113. Mu R et al (2010) An oxidative analogue of gambogic acid-induced apoptosis of human hepatocellular carcinoma cell line HepG2 is involved in its anticancer activity in vitro. *Eur J Cancer Prev* 19(1):61–67
114. He D et al (2009) The NF-kappa B inhibitor, celastrol, could enhance the anti-cancer effect of gambogic acid on oral squamous cell carcinoma. *BMC Cancer* 9:343
115. Xu X et al (2009) Gambogic acid induces apoptosis by regulating the expression of Bax and Bcl-2 and enhancing caspase-3 activity in human malignant melanoma A375 cells. *Int J Dermatol* 48(2):186–192
116. Wang X et al (2009) Proteomic identification of molecular targets of gambogic acid: role of stathmin in hepatocellular carcinoma. *Proteomics* 9(2):242–253
117. Wang F et al (2014) Gambogic acid suppresses hypoxia-induced hypoxia-inducible factor-1alpha/vascular endothelial growth factor expression via inhibiting phosphatidylinositol 3-kinase/Akt/mammalian target protein of rapamycin pathway in multiple myeloma cells. *Cancer Sci* 105(8):1063–1070
118. Wang T et al (2008) Gambogic acid, a potent inhibitor of survivin, reverses docetaxel resistance in gastric cancer cells. *Cancer Lett* 262(2):214–222
119. Rong JJ et al (2010) Gambogic acid triggers DNA damage signaling that induces p53/p21 (Waf1/CIP1) activation through the ATR-Chk1 pathway. *Cancer Lett* 296(1):55–64
120. Qin Y et al (2007) Gambogic acid inhibits the catalytic activity of human topoisomerase IIalpha by binding to its ATPase domain. *Mol Cancer Ther* 6(9):2429–2440
121. Wu ZQ, Guo QL, You QD, Zhao L, Gu HY (2004) Gambogic acid inhibits proliferation of human lung carcinoma SPC-A1 cells in vivo and in vitro and represses telomerase activity and telomerase reverse transcriptase mRNA expression in the cells. *Biol Pharm Bull* 27(11):1769–1774
122. Yu J et al (2006) Repression of telomerase reverse transcriptase mRNA and hTERT promoter by gambogic acid in human gastric carcinoma cells. *Cancer Chemother Pharmacol* 58(4):434–443
123. Kasibhatla S et al (2005) A role for transferrin receptor in triggering apoptosis when targeted with gambogic acid. *Proc Natl Acad Sci USA* 102(34):12095–12100
124. Pandey MK, Kale VP, Song C, Sung SS, Sharma AK, Talamo G, Dovat S, Amin SG (2014) Gambogic acid inhibits multiple myeloma mediated osteoclastogenesis through suppression of chemokine receptor CXCR4 signaling pathways. *Exp Hematol* 42(10):883–896
125. Gu H et al (2008) Gambogic acid induced tumor cell apoptosis by T lymphocyte activation in H22 transplanted mice. *Int Immunopharmacol* 8(11):1493–1502
126. Chi Y et al (2013) An open-labeled, randomized, multicenter phase IIa study of gambogic acid injection for advanced malignant tumors. *Chin Med J* 126(9):1642–1646
127. Li D et al (2015) Antitumor activity of gambogic acid on NCI-H1993 xenografts via MET signaling pathway downregulation. *Oncol Lett* 10(5):2802–2806

128. Yue Q et al (2016) proteomic analysis revealed the important role of vimentin in human cervical carcinoma HeLa cells treated with gambogic acid. *MCP* 15(1):26–44
129. Huang GM, Sun Y, Ge X, Wan X, Li CB (2015) Gambogic acid induces apoptosis and inhibits colorectal tumor growth via mitochondrial pathways. *WJG* 21(20):6194–6205
130. Shi X et al (2015) Gambogic acid induces apoptosis in diffuse large B-cell lymphoma cells via inducing proteasome inhibition. *Sci Rep* 5:9694
131. Yang Y et al (2007) Differential apoptotic induction of gambogic acid, a novel anticancer natural product, on hepatoma cells and normal hepatocytes. *Cancer Lett* 256(2):259–266
132. Lu N et al (2013) Gambogic acid inhibits angiogenesis through inhibiting PHD2-VHL-HIF-1alpha pathway. *Eur J Pharm Sci* 49(2):220–226
133. Wang J, Yuan Z (2013) Gambogic acid sensitizes ovarian cancer cells to doxorubicin through ROS-mediated apoptosis. *Cell Biochem Biophys* 67(1):199–206
134. Li Q et al (2010) Gambogic acid inhibits proliferation of A549 cells through apoptosis-inducing and cell cycle arresting. *Biol Pharm Bull* 33(3):415–420
135. Zhao J et al (2008) Inhibition of alpha(4) integrin mediated adhesion was involved in the reduction of B16-F10 melanoma cells lung colonization in C57BL/6 mice treated with gambogic acid. *Eur J Pharmacol* 589(1–3):127–131
136. Guo QL, You QD, Wu ZQ, Yuan ST, Zhao L (2004) General gambogic acids inhibited growth of human hepatoma SMMC-7721 cells in vitro and in nude mice. *Acta Pharmacol Sin* 25(6):769–774
137. Wang C, Wang W, Wang C, Tang Y, Tian H (2015) Combined therapy with EGFR TKI and gambogic acid for overcoming resistance in -T790M mutant lung cancer. *Oncol Lett* 10(4):2063–2066