

Chapter 9

Nanoparticle Toxicity in Water, Soil, Microbes, Plant and Animals

Naureen S. Khan, Ashwini K. Dixit, and Rajendra Mehta

Abstract Nanotechnology is defined as the design, synthesis and application of materials, devices whose size and shape have been engineered at the nanoscale. The building blocks of nanotechnology are nanoparticles. Nanoparticles are broadly classified as natural nanoparticles and anthropogenic nanoparticles. This chapter presents first the definitions, principles and applications of nanomaterials. Then the toxicity of nanomaterials on soil, water, food, microbes, plants and animals is detailed.

Keywords Nanoparticles • Toxicity • Water • Soil • Microbes • Bacteria • Plants • Animals

9.1 Introduction

The building blocks of nanotechnology are nanoparticles (Biswas and Wu 2005). The term nanoparticles is derived from the word ‘nanos’ meaning tiny. Particles that are measured in nanoscale level, i.e. 0.1–100 nm in diameter, are termed as nanoparticles (Royal society and Royal Academy of Engineering 2004). They have been present on earth and used by mankind for millions of years (Nowack and Bucheli 2007). Nanotechnology is defined as the design, synthesis and application of materials, devices whose size and shape have been engineered at the nanoscale (Buzea et al. 2007). It has bridged the gap among various other existing technologies of present scenario. It has mingled chemistry, physics, biology, engineering and technology, bioinformatics and biotechnology in such a manner that has offered a novel

N.S. Khan (✉) • A.K. Dixit (✉)
Department of Botany, Guru Ghasidas Vishwavidyalaya,
495009 Bilaspur, Chhattisgarh, India
e-mail: nicks30khan@gmail.com; dixitak@live.com

R. Mehta
Department of Rural Technology, Guru Ghasidas Vishwavidyalaya,
495009 Bilaspur, Chhattisgarh, India
e-mail: drmehta21@gmail.com

concept of interdisciplinary research. Nanoparticles play a key role and are known as the backbone of this technology due to their distinguishing and incomparable properties. The small size offers a large surface area as compared to their bulk counter parts (Biswas and Wu 2005). The large surface to volume ratio, surface charge, geometry and state of existence decide their relative reactivity.

Nanoparticles are broadly classified as natural nanoparticles and manmade (anthropogenic) nanoparticles. Anthropogenic nanoparticles are further sub divided into unintentional and intentional/engineered nanoparticles. A variety of nanoparticles are introduced in the environment by various natural processes. Volcanic eruptions, forest fires, weathering phenomenon and soil erosion driven by water and wind are the sources of natural nanoparticles (Smita et al. 2012). Whereas, biological origin i.e. viral particles (Hogan et al. 2004), Pollen fragments (McMurry et al. 2000). Coal fired combustion systems and incinerators (Chang et al. 2004); automobiles and diesel powered vehicles (Kittelson 1998); welding processes industrial boilers, fire places, automobiles, diesel (Vincent and Clement 2000); trucks, and meat-cooking operations (Hildemann et al. 1991) are the sources of unintentional anthropogenic nanoparticles. The intentional/engineered nanoparticle i.e. carbon containing nanoparticles (fullerence, carbon nanotubes etc.) inorganic nanoparticles (metal nanoparticles like nano silver, nano gold, titanium dioxide nanoparticles, zinc oxide nanoparticles etc.), organic and inorganic nanohybrids (Sajid et al. 2015) are synthesized by following two approaches i.e. either top down or bottom up method.

The engineered nanoparticles have been exploited in every sector of society. They have become a compulsive content of today's smart products. Their rapidly increasing application in textiles, electronics, pharmaceuticals, cosmetics and environmental remediation is noticeable and calls for their impact assessment (Royal society and Royal Academy of Engineering 2004). Table 9.1 corroborates the application of different nanoforms.

In the present review we have taken an attempt to commemorate the toxicity imposed by the nanoparticles on every sphere of life forms. Brief information about toxic effect of nanoparticles on the environment, microbes, animals and plants has been discussed in this article. The possible preventive measures and future challenges are also mentioned.

9.2 Nanotechnology, an Old Concept

The concept of nanotechnology was proposed early in 1959 by a physicist Richard Feynman in a lecture-“There's Plenty of Room at the Bottom” at a meeting of the American Physical Society and was honored by Nobel prize for this valuable contribution. Whereas, the term nanotechnology was coined by Norio Taniguchi in 1974. Unknowingly, the people were using the nanoparticles in their work place. Evidences of nanotechnology's existence in the middle ages are drawn from the stained glasses of various old churches. The glass windows of European cathedrals

Table 9.1 Application of nanoparticles

Nanoparticle	Sector	Use	Product	Reference	
Silver (Ag)	Textile	As casuals	T-shirt, socks, sports clothing, shoe	Benn and Westerhoff (2008)	
		Food packaging:	As antimicrobial agent	Blue moon goods fresh box silver	Cushen et al. (2012)
				Nanoparticle food storage container	Alfadul and Elneshwy (2010)
			As antimicrobial agent	Nano Care Technology Ltd. Antibacterial	Bouwmeester et al. (2007)
		As antimicrobial agent	Sunriver industrial nanosilver fresh food bag	Huang et al. (2011)	
	Implants and medical devices	Dental hygiene	Toothpaste	Reidy et al. (2013)	
		Treatment of eye condition		Reidy et al. (2013)	
		Application in plastic		Reidy et al. (2013)	
		To produce surgical meshes		Reidy et al. (2013)	
		Vascular prosthesis		Reidy et al. (2013)	
		Ventricular drainage catheters		Reidy et al. (2013)	
		In surface coating of respirators		Cushen et al. (2012)	
		Due to plasmonic property used in	Biosensor	Austin et al. (2011)	
		In reducing secondary bacteremia	Acticoat	Reidy et al. (2013)	
		Consumer products	In water treatment process/		Reidy et al. (2013)
	Silver impregnated (<100 nm)		Water filter	Nowack et al. (2011)	
	Surface coatings, paints and washing machine			Reidy et al. (2013)	
				Reidy et al. (2013)	

(continued)

Table 9.1 (continued)

Nanoparticle	Sector	Use	Product	Reference
			Air filters	Reidy et al. (2013)
			Vaccum cleaners	Klaine et al. (2008)
			Phones	Klaine et al. (2008)
			Laptops	Klaine et al. (2008)
			Toys	Klaine et al. (2008)
			Baby products	Cerkez et al. (2012)
	Other products	In disinfection	Disinfecting spray	Wijnhoven et al. (2009)
		Imparting fragrance	Deodrants	Lorenz et al. (2011)
			Other cosmetics	Lorenz et al. (2011)
	As antifungal agent	Fungicide	Wright et al. (1999)	
TiO ₂	Domestic purpose	In cleaning items	Detergent	Kaida et al. (2004)
			Toothpaste	Kaida et al. (2004)
			Window panes	Remédios et al. (2012)
			Ceramic tiles	Remédios et al. (2012)
		Medical implants	Biosensor	Remédios et al. (2012)
	Commercial purpose	Displays	Television screen	Remédios et al. (2012)
			Computer monitors	Remédios et al. (2012)
		As photocatalyst:	Solar cells	Klaine et al. (2008)
			Batteries	Remédios et al. (2012)
	Food products			Remédios et al. (2012)
Environmental remediation	As decontaminant		Esterkin et al. (2005)	

(continued)

Table 9.1 (continued)

Nanoparticle	Sector	Use	Product	Reference
ZnO	Cosmetics	As UV light scattering additive	Sunscreen	Serpone, et al. (2007)
			Toothpaste	Serpone et al. (2007)
			Beauty products	Serpone et al. (2007)
		In electronic items		Song et al. (2010)
		In textiles		Dastjerdi and Montazer (2010)
		As essential ingredient in:	Antifouling paints	IPPIC (2012)
CuO	Electronics and technology		Lithium batteries	Sau et al. (2010)
Gold nanoparticles (Au)	Electronics	Used in conducting Inks/ Films		Klaine et al. (2008)
Iron oxide			Lipstick	Remédios et al. (2012)
		As detoxifying agent		Zhang (2003)
Alumina			Shampoo	Klaine et al. (2008)
CeO ₂ cerium dioxide		As combustion catalyst	Diesel fuel	Park et al. (2008)
			Solar cells	Serpone et al. (2007)
			Gas sensors	
			Oxygen pumps	Lin et al. (2006)
Carbon nanotubes	Commercial application	Polymer composites	Lubricants, machinable ceramics,	Remédios et al. (2012)
		Structural composites	Magnetic materials, military battle suits	Remédios et al. (2012)
		Conductive coatings		Remédios et al. (2012)
		Electromagnetic shielding		Remédios et al. (2012)
	Medical application	Orthopedic implants		Klaine et al. (2008)
		Automotive industries		Klaine et al. (2008)
	Others	In water purification system		Klaine et al. (2008)
		Adhesives		Klaine et al. (2008)

(continued)

Table 9.1 (continued)

Nanoparticle	Sector	Use	Product	Reference
Quantum dots	Electronics and technology		(Insulators Semiconductors Magnetic materials)	
			Solar cells	Royal society (2004)
			Photovoltaic cells	Royal society (2004)
			Security inks	Royal society (2004)
			Photonics	Royal society (2004)
			Telecommunication	Alivisatos et al. (2005)
	Medical application	In biological imaging	As markers	Hoet et al. (2004)
			As targeted therapeutics	Alivisatos et al. (2005)
Dendrimers	Medical devices	In drug delivery		Royal society (2004)
		As surface modifiers for enantioselective catalysis	Macrocapsules	Klaine et al. (2008)
			Nanolatex	Klaine et al. (2008)
			Coloured hlasses	Klaine et al. (2008)
			Chemical sensors	Klaine et al. (2008)
			Modified electrodes	Klaine et al. (2008)
			DNA transfecting agents	Klaine et al. (2008)
		As therapeutic agent	Hydrogels	Klaine et al. (2008)
DNA chips	Klaine et al. (2008)			
Nano composites	Vehicular application	Packaging auto- parts		Royal society (2004)
Carbon black			Plastic fillers	Hoet et al. (2004)
Fumed silica	Vehicular application	Used in car tyres and plastic fillers		Hoet et al. (2004)
Silica	Gene delivery			Ravi Kumar et al. (2004)

are live examples of this. The historical aspect of nanotechnology from the most primitive time to the middle age was commemorated by (Daniel and Astruc 2004). Due to very astonishing optical properties of nanoparticles they have been used since the fourth century, the prominent example of which is the dichroic glass of Lycurgus cup whereas, the use of nanomaterials in industrial sectors begins in twentieth century (Horikoshi and Serpone 2013). Use of silver nanoparticles as an anti-septic agent has been documented early in Rig veda. Various metals in nano form occupies a remarkable place in ayurvedic, sidhha and unani system of medicine due to their terrific potential in curing a wide range of diseases and abnormalities.

9.3 Distinguishing Properties of Nanoparticles

Before highlighting the hazardousness of nanoparticles the expected factors that are responsible for nanoparticle toxicity are size, shape, nature, reactivity, mobility, stability and surface chemical charge (Sajid et al. 2015). Property of the material changes in nanoparticle regime. Activity of the nanoparticles are strongly governed by their stability which is attained by the provision of ideal capping agents. Temperature, pressure and long term incubation governs the texture and consistency of the nano regime material due to the onset of aggregation or agglomeration among the nanopopulation.

9.4 Effect of Nanoparticles on the Environment

Environment harbors all the things that exist within itself. Any alteration in the environment gradually affects its state, constituents and vice-versa. Environment consists of two factors broadly classified as biotic and abiotic factor. Nanoparticles that are expelled in the environment either naturally or by anthropogenic means, Spontaneously through natural phenomenon or by engineered sources imparts/leads to some changes in them. The inhalation of atmospheric nanoparticle has caused 60,000 deaths per year reported by US Environmental Protection Agency (EPA) and also these Nanoparticles directly get transferred in to the brain Oberdorster et al. (2004) and Raloff (2003). This is due to the large surface area which has resulted in direct generation of reactive oxygen species that are known as harmful oxyradicals, having the potential to attack DNA, proteins and membranes too as a result of which the cell gets injured Brown et al. (2001). In addition they show affinity towards the transition metals and organic chemical pollutants that enhances the level of toxicity to a greater extent Cheng et al. (2004) Route of entry of nanoparticles varies with the type of habitat. Figure 9.1 reflects the possible route of nanoparticle exposure to various life form in brief. In terrestrial organisms they may enter via inhalation or ingestion Brigger et al. (2002) whereas, in case of the aquatic animals they enter via direct passage across gill and other external surface epithelia as reported by Moore

environmental hydroxyl radical concentration. Wilson et al. (2007) reported the increase in ozone depletion mechanism by these tiny creatures that may be attributed due to change in temperature of stratospheric layer as reported by Tromp et al. (2003), Smita et al. (2012). According to Strawa et al. 2010 nanoparticles take part in the coarse material formation which is the major cause of global warming in present scenario.

9.4.2 *Effect on Soil*

The ultimate sink of every component in environment is soil and so as for nanoparticles. These tiny tools end up their journey in the soil or become dormant for some time. Soil is a natural reservoir for a wide variety of micro flora as it offer conducive environment for growth and survival of biological indicators. In turn they offer nutrients in the form of enzyme that take part in decomposition of soil content and improve the quality of soil by executing this cyclic process. In a broad sense their exists mutualism. The long term dormancy period of nanoparticles may evoke serious issues on the health of its inhabitant i.e. contribute in reducing the fertility of soil thereby causing an imbalance in the soil ecosystem (Kennedy and Smith 1995). Common ENPs include the metal ENPs (elemental Ag, Au, Fe etc.), fullerenes (grouping Buckminster fullerenes, CNTs, nanocones etc.), metal oxides (TiO₂, CuO, FeO₂, ZnO, Al₂O₃, SiO₂ etc.), complex compounds (Co–Zn–Fe oxide), quantum dots often coated with a polymer e.g. cadmium–selenide (CdSe) and organic polymers (dendrimers, polystyrene, etc.) Dinesh et al. 2012. Fortner et al. (2005) performed toxicity study of C₆₀ aggregates on soil inhabiting *E. Coli* and *B. subtilis* bacteria. They found it to be toxic even at low concentration marked by reduced growth (>0.4 ppm). more over, this concentration has lowered the rate of aerobic respiration up to 4 ppm. Thus may interfere with soil development (pedogenesis) or behavior (e.g., crusting and dispersion) Klaine et al. (2008), Cameron et al. (1915).

9.4.3 *Effect of Nanoparticle on Water*

Domestic and Industrial wastes (may contain Nanoparticles in aggregate or colloidal form) expelled in the water bodies viz. ponds, rivers, sea etc. increases the threat of nanoparticle contamination. Discharges usually contains organic (protenaceous and polysaccharide from the microbes) and inorganic matter (heavy metal oxides) that make the water bodies unfit for use. It has been suggested that Ionic silver has found to be highly toxic that disrupts Na+K+– ATPase enzyme activity due to which active Na+ and Cl– uptake is blocked that causes Osmoregulation by fishes, leads to death of fishes thereby contaminate the water as studied by Tang and Wang (2004).

Nanoparticles may also accumulate on the surface microlayers of the oceans, presenting a route of aerosol exposure risk to marine birds and mammals, as well as the organisms living on the surface microlayer due to their hydrophobic nature. Kennedy et al. (2004), Simkiss K. (1990).

9.4.4 Effect of Nanoparticles on Food Chain

Starting from the primary producer (Plants) to that of the tertiary consumers (insects/ animals). At every successive level nano-particles reflects their involvement by being entered in their metabolic pathway through active and passive transport. Confirmatory reports have been published by Holbrook et al. (2008), Lin et al. (2009) that shows their harmful effect in the food chain. Their small size offers large surface area that imparts them high potential to carry toxic materials i.e. such as lipophilic pollutants and heavy metals Baun et al. (2008) to enter the system.

9.4.5 Effect of Nanoparticles on Agriculture Production

About 70 % of India's economy is based on agricultural practices. The formation of dust clouds has found to be highly hazardous. Burning of wood for cooking and other purposes and the incomplete combustion of biomass give rise to natural nanoparticle. Their existence in environment has resulted in brown cloud formation which has remarkably lowered the sunlight intensity in south Asia. These pollution clouds have been indulged in the reduction of monsoon season in India. The productivity of chief crop plants like rice, wheat and soybean has immensely hampered as reported by UNEP 2002. Asian brown clouds are found to be loaded with large amount of soot and black carbon. These complexly degradable materials contributes to increase in melting of glaciers thereby blocking the availability of water to various glacier fed rivers as commemorated by Gustafsson et al. (2009). Water scarcity contributes in low yield of food crops.

9.5 Effect of Nanoparticles on Microbes

Nanoparticles not only affected abiotic factors rather have also been observed to pose harmful effects on microscopic creatures too ranging from the prokaryotes to Eukaryotes. For example – Akhavan and Ghaderi (2010) investigated the toxicity of graphene and graphene oxide damages the cell membrane of both the Gram-negative bacteria *Escherichia coli* and the Gram-positive bacteria *Staphylococcus aureus* by their sharp edges. A long descriptive about nanoparticles impact on micro flora has been expressed in Table 9.2.

Table 9.2 Harmful effect of nanoparticles on microorganisms

Test organism	Nanoparticle	Toxic effects	Reference
Bacterial <i>Sp.</i>	C60 (water suspension)	Antibacterial to broad range	Lyon et al. (2005, 2006), and Sayes et al. (2004)
Bacterial <i>Sp.</i>	C60 (encapsulated in polyvinylpyrrolidone)	Antibacterial to broad range	Kai et al. (2003)
Gm +veBacterial <i>Sp.</i>	Carboxyfullerene (malonic acid derivative)	Bactericidal effect	Mashino et al. (1999) and Tsao et al. (2002).
Gm +veBacterial <i>Sp.</i>	Hydroxylated fullerene	Bactericidal effect	Rozhkov et al. (2003)
Gm –veBacterial <i>Sp.</i> (<i>E. Coli Sp.</i>)	C60 derivative with pyrrolidine groups	Inhibit its growth by interfering with energy metabolism	Mashino et al. (1999, 2003)
Gm +veBacterial <i>Sp.</i> (<i>Mycobacteria Sp.</i>)	Other derivative of C60	Inhibit its growth	Babynin et al. (2002) and Bosi et al. (2000)
Gm –veBacterial <i>Sp.</i> (<i>Salmonella typhimurium</i>)	Other derivative of C60: (carbon nanotube)	Antibacterial and antimutagenic effect	Babynin et al. (2002) and Bosi et al. (2000)
Gm –veBacterial <i>Sp.</i> (<i>E. Coli Sp.</i>)	(SWCNT)	Antibacterial(cell membrane damage)	Kang et al. (2007) and Wei et al. (2007)
Bacterial <i>Sp.</i>	(MWCNT)	Cytotoxic in nature	Biswas and Wu (2005)
Gm –veBacterial <i>Sp.</i> (<i>E. Coli Sp.</i>)	Metallic: quantum dots (uncoated)	Toxic causes oxidative damage to cell membrane	Kloepfer et al. (2005) and Hardman (2006)
Gm +veBacterial <i>Sp.</i> (<i>Bacillus Subtilis</i>)	Metallic: quantum dots (uncoated)	Toxic causes oxidative damage to cell membrane	Kloepfer et al. (2005) and Hardman (2006)
Gm –veBacterial <i>Sp.</i> (<i>E. Coli Sp.</i>)	Silver	Highly toxic (in environmental condition)	Chernousova and Epple (2013)
Bacterial <i>Sp.</i>	Silver	Bactericidal effect	Sondi and Salopek-Sondi (2004) and Morones et al. (2005)
Virus	Silver	Viricidal effect	Sondi and Salopek-Sondi (2004) and Morones et al. (2005)
Gm +veBacterial <i>Sp.</i> (<i>Staphylococcus aureus</i>)	Gold	Low toxic	Nyberg et al. (2008) and Goodman et al. (2004)
Gm –veBacterial <i>Sp.</i> (<i>E. Coli Sp.</i>)	Gold	Low toxic	Nyberg et al. (2008) and Goodman et al. (2004)
Gm –veBacterial <i>Sp.</i> (<i>Shewanella oneidensis</i>)	Metal oxides: magnetite	Low toxic	De Windt et al. (2006)

(continued)

Table 9.2 (continued)

Test organism	Nanoparticle	Toxic effects	Reference
Gm –veBacterial Sp. (<i>E. Coli Sp.</i>)	TiO ₂	Accelerates solar disinfection by photocatalytic activity and (ROS)	Rincon and Pulgarin (2004) and Wolfrum et al. (2002)
Gm +veBacterial Sp. (<i>Micrococcus luteus</i>)	TiO ₂	Accelerates solar disinfection by photocatalytic activity and (ROS)	Rincon and Pulgarin (2004) and Wolfrum et al. (2002)
Gm +veBacterial Sp. (<i>Bacillus Subtilis</i>)	TiO ₂	Accelerates solar disinfection by photocatalytic activity and (ROS)	Rincon and Pulgarin (2004) and Wolfrum et al. (2002)
Fungal Sp. (<i>Aspergillus niger</i>)	TiO ₂	Accelerates solar disinfection by photocatalytic activity and (ROS)	Rincon and Pulgarin (2004) and Wolfrum et al. (2002)
Gm +veBacterial Sp. (<i>Bacillus Subtilis</i>)	MgO	Antibacterial effect	Huang et al. (2005)
Gm +veBacterial Sp. (<i>Staphylococcus aureus</i>)	MgO	Antibacterial effect	Huang et al. (2005)
Gm –veBacterial Sp. (<i>E. Coli Sp.</i>)	CeO ₂	Antimicrobial effect	Thill et al. (2006)
Gm –veBacterial Sp. (<i>E. Coli Sp.</i>)	Zno (uncoated)	Highly toxic	Bondarenko et al. (2012)
Gm +veBacterial Sp. (<i>Bacillus Subtilis</i>)	Zno	Antibacterial activity	Sawai et al. (1995,1996)
Gm –veBacterial Sp. (<i>E. Coli Sp.</i>)	Zno	Antibacterial activity	Sawai et al. (1995,1996)
Gm –veBacterial Sp. (<i>E. Coli Sp.</i>)	ZnO (20 nm)	100 % mortality at 20 mg/ml of ZnO	Jiang et al. (2009)
Gm –veBacterial Sp. (<i>E. Coli Sp.</i>)	ZnO (10–30 nm)	Prevented growth at 500 mg/ml of ZnO	Premanathan et al. (2011)
Bacterial Sp.	SiO ₂	Mild toxicity due to (ROS) production	Adams et al. (2006)
Gm –veBacterial Sp. (<i>E. Coli Sp.</i>)	CuO (uncoated)	Highly toxic	Bondarenko et al. (2012)
Gm –veBacterial Sp.:- <i>P. aeruginosa</i>	CuO (80–160 nm)	Greatly affects the growth of these PGPR	Mahapatra et al. (2008)
<i>Klebsiella pneumoniae</i>	CuO (80–160 nm)	Greatly affects the growth of these PGPR	Mahapatra et al. (2008)

(continued)

Table 9.2 (continued)

Test organism	Nanoparticle	Toxic effects	Reference
<i>Salmonella Paratyphi</i>	CuO (80–160 nm)	Greatly affects the growth of these PGPR	Mahapatra et al. (2008)
<i>Shigella Strain</i>	CuO (80–160 nm)	Greatly affects the growth of these PGPR	Mahapatra et al. (2008)
<i>P. aeruginosa</i>	Iron oxide nanoparticles	Reacts with peroxides present in environment,	Saliba et al. (2006) and Mishra and Kumar (2009)
		Generates free radical, which are highly toxic	
<i>P. putida</i>	Iron oxide nanoparticles	Reacts with peroxides present in environment,	Saliba et al. (2006) and Mishra and Kumar (2009)
		Generates free radical, which are highly toxic	
<i>P. fluorescens</i>	Iron oxide nanoparticles	Reacts with peroxides present in environment,	Saliba et al. (2006) and Mishra and Kumar (2009)
		Generates free radical, which are highly toxic	

9.5.1 Effect of Nanoparticles on Nitrogen Fixing Bacteria (Cyanobacteria & Others)

Cyanobacteria are integral part of producer community as they are the largest nitrogen fixer globally due to the presence of nif gene and other ideal characteristics they are capable of fixing the atmospheric nitrogen themselves and serves as Biofertilizers thereby play a significant role in the growth of greenery. Due to the presence of oxygenic photosynthetic system they are capable of maintaining the equilibrium of earth atmosphere. These certain properties drag them in the list of essentials that vitally contribute for balancing the environment and nature. Brayner et al. (2010) performed the eco-toxicological investigation of ZnO Nanoparticles by taking *Anabaena flos-aquae* cyanobacteria and *Euglena gracilis* euglenoid microalgae as the experimental organism. ZnO Nanoparticles were synthesized by polyol process in which s trin-octylphosphine oxide (TOPO) and polyoxyethylene stearyl ether (Brij-76) were used as protective agents to control the particle size and shape. In the present investigation effect of ZnO nanoparticles on photosynthetic activity of the selected organisms was corroborated. In the case of *Anabaena flos-aquae*, addition of ZnO, ZnO-TOPO, and ZnO-Brij-76 induces stress which causes a progressive

decline in the photosynthetic rate. Among these three different types of nanoparticles only ZnO-TOPO causes a fall in photosynthetic rate followed by cell death. However, In case of *Euglena gracilis*, all the nanoparticles were found to be indulged in cell death which was confirmed by doing the TEM analysis of their ultra thin sections. It reveals that polysaccharides produced by *Anabaena flos-aquae* avoid particle internalization after contact with ZnO and ZnO-Brij- 76 nanoparticles. Ma et al. (2013) suggested that nanoparticles have been found to interfere in the metabolic pathway of nitrogen fixing bacteria as well.

9.5.2 Effect of Nanoparticles on Plant Growth Promoting Rhizobacteria

Plant growth promoting rhizobacteria (PGPR) like *P. aeruginosa*, *P. putida*, *P. fluorescens*, *B. subtilis* and soil Nitrogen cycle bacteria viz., nitrifying bacteria and denitrifying bacteria have shown varying degrees of inhibition when exposed to ENPs in pure culture conditions or aqueous suspensions Mishra and Kumar (2009). Metal oxide nanoparticles of Cu (80–160 nm) were tested for antibacterial activity against plant growth promoting *Klebsiella pneumoniae*, *P. aeruginosa*, *Salmonella paratyphi* and *Shigella strains* Mahapatra et al. (2008). Iron and copper based nanoparticles are presumed to react with peroxides present in the environment generating free radicals known to be highly toxic to microorganisms like *P. aeruginosa* Saliba et al. (2006).

9.5.3 Effect of Nanoparticles on Eukaryotes

Likewise, the prokaryotes nanoparticles adversely affects the growth and metabolism of eukaryotes. Kasemets et al. (2009) has shown the toxic effect of ZnO nanoparticles on the unicellular Fungi i. e. yeast (*Saccharomyces cerevisiae*). Mortimer et al. (2010) suggested the toxicity of ZnO nanoparticles against free living Protozoan Sp. (*Tetrahymena thermophila*).

9.6 Effect of Nanoparticles on Plants

9.6.1 Effect of Nanoparticles on Algae & Diatom (Lower Plants)

Due to the adsorption of TiO₂ nanoparticles on the surface of algae, its weight becomes double as a result of this its floating capacity decreases and low surface gets exposed to sunlight leads to a gradual fall in photosynthetic rate as suggested

by Navarro et al. (2008). In case of a green algae i.e. *Desmodesmus subspicatus*, TiO₂ nanoparticles were found to be toxic by Hund-Rinke and Simon (2006). ZnO nanoparticles showed toxicity against *Euglena gracilis* Euglinoid Brayner et al. (2010). Marine diatom *T. pseudonana* was found to be susceptible to ZnO nanoparticles and exhibit toxicity as reported by Miao et al. (2010).

9.6.2 Effect of Nanoparticles on Marine Phytoplanktons

Population growth studies on four marine phytoplanktons has been conducted by Miller et al. (2010) to assess the toxicity estimation of two metal oxide nanoparticle i.e. ZnO and TiO₂. Salty water bodies comprises a vast portion of earth and a repeated history of waste disposition has been in a common practice since ancient time, whether by accidental or intentional means. Marine phytoplanktons are adversely affected by engineered nanoparticles. In order to assess the sublethal effect of two selected metal NPs a modeling approach (Dynamic Energy Budget (DEB) framework) was employed in which They have found that ZnO nanoparticles depressed the growth rate of *Skeietonema marioni*, *Thalassiosira pseudonana*, *Dunaliella tertiolecta* and *Isochrysis galbana* by 50–70%. Reproduction and early stage developmental study has been conducted by Nielsen et al. (2008). They have selected a marine brown algae *Fucus serratus* to test against manufactured Carbon black nanoparticle. Inference clearly states a great changes. Carbon black nanoparticles when subjected to the sperm suspension and embryo culture gets agglomerate thereby causing a decline in the fertilization success at a concentration of 100 mg l⁻¹.

9.7 Effect of Nanoparticles on Higher Plants

Nanoparticles are found to be indulged in the formation of reactive oxygen species in some higher plants explained by Navarro et al. (2008). Accumulation of nanoparticle takes place on the leaf lamina and on stomatal tissues causes an imbalance in the equilibrium inside outside the leaf and ultimately in plants Da Silva et al. (2006). Accumulation of nanoparticle on photosynthetic surfaces causes shading effects, i.e. reduced sun light availability and hence reduced photosynthetic rate Smita et al. (2012). Carbon nanotubes diminished rice yields and made wheat more prone to other pollutants Wild and Jones (2009) due to the asbestos-like behaviour of carbon nanotubes. Also, aluminium nanoparticle have been identified to inhibit root growth in various economically important plant species by interacting with root surface Canas et al. (2008), Yang and Watts (2005).

9.7.1 Phytotoxicity by Nanoparticles

Y. K. Mushtaq (2011) demonstrated The effects of Fe₃O₄, TiO₂, and carbon nanoparticles on cucumber plants. These nanoparticles exhibits the potential to negatively affect seed germination rate, root elongation, and germination index. Another study performed by Kumari et al. (2011) on the harsh effect of ZnO nanoparticles supports their toxic nature. Growing use of ZnO nanoparticles in the daily consumer products calls for an alert after their significant investigation. Workers have done a depth study by determining the Mitotic index (MI), micronuclei index (MN index), chromosomal aberration index, and lipid peroxidation of the root cells of *A. cepa* (grown hydroponically). They have treated the model organism with four different concentrations of ZnO nanoparticles dispersions (25, 50, 75, and 100 gml⁻¹). It was observed that MI decreases with the increase in concentration of ZnO nanoparticles to the contrary MN index and chromosomal index increases steadily. in the ZnO nanoparticles treated cells, the total count of micronucleated cell were higher than control set up which corroborates the genotoxic effect of ZnO nanoparticles on plant community.

9.7.2 Genotoxicity by Nanoparticles

Atha and collaborators (2012) reported for the first time that copper oxide Nanoparticles damaged DNA in some agricultural and grassland plants (*Raphanus sativus*, *Lolium perenne*, and *Lolium rigidum*). Mutation occurs in the crop plants that blocks their growth and results in gradual fall in productivity. Through this fraction of investigation they have clarified that metal nanoparticles acts as a carrier/mediator for DNA damage in all the living community including mammalian cell, plants and even the microscopic beings i.e. bacteria (Table 9.3).

Table 9.3 Harmful effect of nanoparticles on plants

Nanoparticle	Plant name	Harmful effect	Reference
	ALGAE:		
ZnO	<i>Pseudokirchneriella subcapitata</i>	Toxic interfere in metabolic pathway	Aruoja et al. (2009)
ZnO	<i>Anabaena flos-aquae</i>	Inhibit photosynthetic activity	Brayner et al. (2010)
Ag Nanoparticles	<i>unknown</i>	Highly toxic	Albright and Wilson (1974)
ZnO	<i>Euglena gracilis</i>	Cause cell death	Brayner et al. (2010)
ZnO (20 nm)	Marine diatom: <i>T. pseudonana</i>	Found to be toxic	Miao et al. (2010)

(continued)

Table 9.3 (continued)

Nanoparticle	Plant name	Harmful effect	Reference
TiO ₂	Unknown	Adsorbs on algal cell surface, affecting its ability to float.	Navarro et al. (2008)
		Reduced sunlight availability for photosynthesis	Navarro et al. (2008)
TiO ₂	<i>Desmodemus subpicatus</i>	Found to be toxic	Hund-Rinke and Simon (2006)
ZnO	<i>Allium cepa</i>	Exert cytotoxic & genotoxic effect	Kumari et al. (2011)
		Lipid peroxidation	Kumari et al. (2011)
		Decrease of the mitotic index	Kumari et al. (2011)
		Increase of the micronuclei	Kumari et al. (2011)
		Increase of chromosomal aberration index	Kumari et al. (2011)
ZnO	unknown	Penetrate plant system and may interfere with	Anita K. Patlolla (2013)
		intracellular components causing damage to cell division	Anita K. Patlolla (2013)
ZnO	<i>Triticum aestivum</i>	Makes it more vulnerable to other pollutants	Wild and Jones (2009)
		Reduced its biomass	Du et al. (2011)
		Due to dissolution of nanoparticles to Zn ions, facilitation of toxic Zn uptake occurs	Du et al. (2011)
ZnO 2000 mg/l	<i>Rye grass</i>	Inhibit germination of seed	Lin and Xing (2007)
		Terminate root elongation	Lin and Xing (2007)
ZnO	<i>Lepidium sativum</i> (Garden cress)	Affects root elongation	Manzo et al. (2011)
ZnO	<i>Vicia faba</i> (Broad bean)	Cause genotoxicity	Manzo et al. (2011)
ZnO (20–30 nm) at 1 mg/l	Marine phytoplankton species:		
	<i>Skeletonema marioni</i>	Depresses growth rate by 50–75 %	Miller et al. (2010)
	<i>Thalassiosira pseudonana</i>	Depresses growth rate by 50–75 %	Miller et al. (2010)
	<i>Dunaliella tertiolecta</i>	Depresses growth rate by 50–75 %	Miller et al. (2010)
	<i>Isochrysis galbana</i>	Depresses growth rate by 50–75 %	Miller et al. (2010)

(continued)

Table 9.3 (continued)

Nanoparticle	Plant name	Harmful effect	Reference
Carbon nanotubes	Economically important plants	Inhibit root elongation	Canas et al. (2008) and Yang and Watts (2005)
Aluminium nanoparticles	Economically important plants	Inhibit root elongation	Canas et al. (2008) and Yang and Watts (2005)
CNanoparticles	<i>Oryzae Sativa (Rice)</i>	Diminishes yield	Wild and Jones (2009)
CuO	<i>Rhaphanus sativus</i>	Diminishes yield	Wild and Jones (2009)
CuO	<i>Lolium perenne</i>	Diminishes yield	Wild and Jones (2009)
CuO	<i>Lolium rigidum</i>	DNA damage, thus inhibits plant growth	Atha et al. (2012)
CuO (1000 mg/l)	<i>Cucurbita pepo</i>	Reduced emerging root length	Stampoulis et al. (2009)
Ag nanoparticles	<i>Cucurbita pepo</i>	Decrease plant biomass and transpiration	Remédios et al. (2012)
Yb ₂ O ₃	Higher Plant Sp.	Affects root elongation	Ma et al. (2010)
	<i>Cucumis sativus</i>	Yb ₂ O ₃ deposits were found in cytoplasm of root cells which are toxic in nature	Zhang et al. (2012)
CeO ₂	Higher Plant Sp.	Affects root elongation	Ma et al. (2010)
La ₂ O ₃	Higher Plant Sp.	Affects root elongation	Ma et al. (2010)
Gd ₂ O ₃	Higher Plant Sp.	Affects root elongation	Ma et al. (2010)

9.8 Effect of Nanoparticles on Aquatic Organisms

To assess the merits and demerits of engineered nanoparticles several comparative studies has been done since time immemorial. These documentation provide a thorough and in depth study of the other face of a coin say the negative impacts of nanomaterials. Nanoparticles are being exploited and used in diversified applications. And being a multifunctional paradigm researchers have warned to use this versatile tool in an ecofriendly manner. Deleterious effects of nanoparticles are being experienced by atmosphere and land than how could the water bodies that covers 71 % of the earth surface will be ignorant and its inhabitant as well carbon nanotubes and fullerenes found to be potentially harmful especially to benthic organisms Velzeboer et al. (2011). Several researchers have excavated the harmful effects of nanomaterials by following different types of approaches. In recent years Blinova et al. (2010), have performed a comparative analysis of acute toxicity test

of CuO and ZnO nanoparticles between crustaceans and protozoan population found in an artificially prepared fresh water ecosystem and a natural water body. In the study acute toxicity of crustaceans *Daphnia magna* and *Thamnocephalus platyurus* and protozoan *Tetrahymena thermophila* was determined.

9.9 Effect of Nanoparticles on Terrestrial Organisms

9.9.1 Effect of Nanoparticles on Terrestrial Invertebrates

Invertebrates cover a large group of animals ranging from top to bottom trophic levels of ecosystem. They are involved in various physiological phenomenon i.e. pollination, soil formation etc. Their occurrence differ from one habitat to the other and are considered as bioindicators (symbol of environmental change). Being an integral part of environment obviously this group of organisms encounters with the nanoparticles. In this section authors have tried to reflect the effect of nanoparticles on Annelids.

9.9.2 Effect of Nanoparticles on Annelids

Agricultural practices are the most important and widely used occupation cum source of income world wide. In order to earn the livelihood and growing demand of requirements one has to look for an effective and environmentally safe tool to reach the target of high productivity without compromising with the chemical based fertilizers and pesticides that has proved to be indulged in the formation of waste lands by successively decreasing the humus and fertility rate of the soil ultimately leading to a major cause of Bioaccumulation and biomagnification. Organic farming is the best alternative in which earthworms play the most significant role. Keeping in view the immense importance of earthworms in agriculture and zooming application of engineered nanoparticles in almost every sector of society studies has been conducted to see the effect of TiO₂ and ZnO nanoparticles by growing earthworms *Eisenia fetida* in an artificially created soil system by Hu et al. (2010). Distilled water at a concentration of, 0.1, 0.5, 1.0 or 5.0 g kg⁻¹ of Nanoparticles were prepared and earthworms were placed in that soil system for 7 days. By using Acute toxicity test and Comet assay concentration of Ti and Zn in earthworms was determined. It was observed that when doses of TiO₂ and ZnO were greater than 1.0 g kg⁻¹ Zn accumulated in higher concentration than Ti and this leads to hyperaccumulation of Zn in the mitochondria of its gut cells which causes DNA damage. The cellulose activity was also inhibited at the dose level of 5.0 g kg⁻¹ of ZnO

nanoparticles. Likewise, Khare et al. (2011) reported the toxicity of TiO₂ and ZnO nanoparticles less than 25 nm and 100 nm on the nematode (*Caenorhabditis elegans*). In this study toxicity of small and large sized nanoparticles of ZnO and TiO₂ were compared. It was found that small sized (25 nm) ZnO nanoparticles, were more toxic than that of the small (25 nm) and large sized (100 nm) TiO₂ nanoparticles.

9.9.3 Effect of Nanoparticles on Vertebrates

In the previous sections an attempt has been made to throw light upon the effect of nanoparticles on various abiotic and biotic factors of the environment. In this section the hazardous effect of inorganic nanoparticles i.e. ZnO has been demonstrated on two groups of vertebrates i.e. amphibians and mammals.

9.9.4 Effect of Nanoparticles on Amphibians

ZnO nanoparticles brought about developmental abnormalities in amphibians. Metallic nanoparticles have been found to cause deleterious effect up on encountered life forms. Among them ZnO, TiO₂, Fe₂O₃, and CuO nanomaterials (20–100 nm) were subjected to access their effect on amphibians in a study conducted by Nations et al. (2011a) on *Xenopus laevis* by employing the Frog Embryo Teratogenesis Assay Xenopus (FETAX) protocol. Experimental organism was subjected to dispersed nanomaterials at a concentration upto 1000 mg L⁻¹ for TiO₂, Fe₂O₃, CuO, and ZnO. It was observed that inspite of causing mortality to the test specie these causes developmental abnormalities among them Viz.gastrointestinal, spinal, and other abnormalities. However, at a concentration of 10.3 mg L⁻¹ of ZnO leads to total malformation. This study confers the harmful effects of ZnO nanoparticles. so its use and production needs to be checked for future perspective.

9.9.5 Effect of Nanoparticles on Mammals

Nanoparticles have been found to cause a number of serious problems such as cytotoxicity. Copper nanoparticles while incubated with zinc ions remains normal but when exposed to large sized ZnO nanoparticles. (at a non toxic concentration 6.25 µg/ml) its cytotoxic activity enhances gradually, also synergistic activity becomes strong. This is due to the presence of large sized ZnO nanoparticles which brings about the alteration in the human hepatoma cell line HepG2 Li et al. (2015) Table 9.4.

Table 9.4 Harmful effect of nanoparticles on animals

Test system	Nanoparticle	Harmful effect	Reference
Nematodes: (<i>Caenorhabditis elegans</i>)	TiO ₂	Decrease in reproduction potential	Roh et al. (2009)
		Increased enzyme induction & protein formation	Roh et al. (2009)
Largemouth bass (<i>Micropterus salmoides</i>)	FullerenceC ₆₀ (0.5–1 mg/l)	Significant lipid peroxidation in brain	
		Total glutathione levels were marginally depleted in gills	Oberdörster (2004)
<i>Daphnia magna</i>	C ₆₀ water suspension (9 mg/l)	High rate of mortality has been noticed	Zhu et al. (2006)
<i>Fathhead minnow</i>	C ₆₀ water suspension (2.5–5 mg/l)	The peroxisomal lipid transport protein PMP70 was significantly reduced in the minnow	Oberdörster et al. (2006)
Zebrafish (<i>Danio rerio</i>)	C ₆₀ (prepared with benzene, THF, and acetone)	Suffered delayed embryo & larval development	
	(1.5 mg/l)	Decreased survival and hatching rates	Zhu et al. (2007)
Eukaryotic unicellular protozoan:	Carbon nanotubes	Observed pericardial edema	Zhu et al. (2006)
<i>Stylonychia mytilus</i>	MWCNT sonicated (1 mg/l)	Colocalization of CNTs within the mitochondria,	Zhu et al. (2006)
		Damage to micronucleus, macronucleus, and membrane	Gimbert et al. (2007)
Rainbow trout (<i>Oncorhynchus mykiss</i>)	Single-walled sonicated	Significant increases of Na ⁺ K ⁺ -adenosine triphosphatase (ATPase) activity observed in the gills and intestine	Gimbert et al. (2007)
		Oxidative stress–linked effects observed	Gimbert et al. (2007)
<i>Zebrafish (Danio rerio)</i>	DWCNTs (240 mg/l)	Delay in hatching	Gulson and Wong (2006)
<i>Zebrafish (Danio rerio)</i>	Metallic copper (1.5 mg/l)	Gill injury and lethality observed	Seaman and Bertsch (2000)
		Acute toxicity observed	Seaman and Bertsch (2000)
Freshwater mussels (<i>Elliptio complanata</i>)	Quantum dots: cadmium telluride (CdTe)	Observed reduction in phagocytic activity	Karathanasis (1999)
		Lipid peroxidation increased at higher concentrations	Karathanasis (1999)
Mice	Graphene oxide [0–100 mg/L]	Down-regulation of genes, disturbance of cell energy metabolism, transduction	Wang et al. (2011)

(continued)

Table 9.4 (continued)

Test system	Nanoparticle	Harmful effect	Reference
Rats	SWCNT 1 or 5 mg kg ⁻¹	Multifocal macrophage containing granulomas in lung	Warheit et al. (2004)
Guinea pigs	MWCNT 2.5 mg in 0.5 ml	Caused 'organising pneumonitis', pulmonary lesions	Grubek-Jaworska et al. (2006)
Mice	SWCNT 2-mg	SWCNT produced some activation of the histocompatibility complex, in CD4 ⁺ /CD8 ⁺ T-cells	Koyama et al. (2006)
Rats	TiO ₂ particles, rods or dots 1 or 5 mg	Produced transient inflammatory and cell injury effects	Warheit et al. (2006)
Rats	Ultrafine cadmium oxide 70 mg m ⁻³	Increased percentage of neutrophils	Warheit et al. (2006)
		Multifocal alveolar inflammation	Warheit et al. (2006)
		Showed elevated blood Cd.	Takenaka et al. (2004)
Rats	Ultrafine metallic nickel 0.15,–2.54 mg m ⁻³	Increase in pulmonary nickel,	Serita et al. (1999)
		Increase in lung weight	Serita et al. (1999)
		Accumulation of foamy alveolar macrophages (AM),	Serita et al. (1999)
		Generated AM indicating alveolar lipoproteinosis,	Serita et al. (1999)
		Acute calcification of the degenerated AM.	Serita et al. (1999)
Mouse spermatogonial stem cell line	Silver nanoparticles 5–100 mg ml ⁻¹	Mitochondrial function are adversely affected,	
		Cells show increased LDH leakage	Braydich-Stolle et al. (2005)
Rat lung cells	Silver nanoparticles	Reduction in lung function and inflammatory lesions	Sung et al. (2008)
Sprague-Dawley rats		Silver nanoparticles accumulation in olfactory bulb and subsequent translocation to the brain	Kim et al. (2008a, b)
Rat liver cells	Silver nanoparticles	Cell leakage and reduction of mitochondrial function	Hussain et al. (2005)
Mouse fibroblast	Silver nanoparticles	50 µg/ml induced apoptosis to 43.4% of cells	Arora et al. (2009)

(continued)

Table 9.4 (continued)

Test system	Nanoparticle	Harmful effect	Reference
HUMAN:			
Skin	Silver nanoparticles	Argyria-irreversible pigmentation of skin & argyrosis- pigmentation of eyes	Chen and Schluessener (2007)
Epithelium layer	Silver nanoparticles	Severe damage	Boosalis, et al. (1987)
Skin	Silver nanoparticles	Discolouration of the skin	Armitage et al. (1996) and Greene and Su (1987)
Eye	Silver nanoparticles	Severe irritation	Panyala et al. (2008)
Nose	Silver nanoparticles	Severe irritation	Panyala et al. (2008)
Brain	Silver nanoparticles	Neurotoxic damage	Cheng et al. (2004)
Male reproductive system	Silver nanoparticles	Cross blood-testes barrier, deposit in the testes, adversely effects sperm cells	Borm and Kreyling (2004)
Blood cells	Silver nanoparticles	Intensive toxic effect on PBMCs & inhibits phytohaemagglutinin-induced Cytokine production	Shin et al. (2007)
Keratinocytes & fibroblasts	Acticoat, aquacel, Ag & contreet foam	Most significant cytotoxic effect	Panyala et al. (2008)
Cell	Ag nanoparticles >44.0 $\mu\text{g ml}^{-1}$	Necrotic in nature	Panyala et al. (2008)
Central nervous system (CNS)	Silver nanoparticles	Prolonged exposure leads to cerebral ataxia	Aaseth et al. (1981)
Respiratory system: (Lungs)	Silver nanoparticles	Produce surface radical and reactive oxygen specie, toxic to alveolar surface	Chen and Schluessener (2007)
(Lungs)	Silver nanoparticles	Induce oxidative stress in lungs epithelial cells	Limbach et al. (2007)
(Lungs)	Silver nanoparticles	Irritation in respiratory tract	Rosenman et al. (1987)
Brain	Silver nanoparticles	Manic depressive psychosis leads to ruptured aortic aneurysm, finally death	Dietl et al. (1984)
Skin	Silver nanoparticles	Cutaneous side effects: hypersensitivity reaction, allergic contact dermatitis, erythema multiforme	Fisher et al. (2003)

(continued)

Table 9.4 (continued)

Test system	Nanoparticle	Harmful effect	Reference
Lung	Pristine and MWCNTs	Alteration in its architecture, collapsed thick walled alveoli, presence of microhaemorrhagic foci	Coccini et al. (2013)
Human chest cavity	MWCNTs	Causes inflammation, production of scars/lesions, leads to mesothelioma	Poland et al. (2008)
Nucleus	C60 fullerence	Accumulate along nuclear membrane, penetrate and cause DNA damage	Porter et al. (2006)
Metabolic process in rats, human, mice	C60 fullerence	Reduce activity of hepatic enzymes i.e. glutathione	Iwata et al. (1998)
Blood and haematopoietic system	C60 fullerence	Triggered hemolysis of 40–50 % of cells	Bosi et al. (2004)
Human (HEK293) embryo kidney cell	SWCNTs – 0.78–200 $\mu\text{g}/\text{ml}^{-1}$	Inhibition of HEK293 cell proliferation, induced cell apoptosis, and decreased cellular adhesive ability	Cui et al. (2005)
Human bronchial epithelial cells(BEAS 2B)	TiO ₂ nanoparticles (10 or 20 nm)	Cause DNA damage, lipid peroxidation and damage to nucleus	Gurr et al. (2005)
Human lung cell	Carbon nanotubes	Inflammation and oxidative stress	Pulskamp et al. (2007)
Human lung cell	Carbon black	Inflammation and oxidative stress	Monteiller et al. (2007)
Human lung cell	Silica	Inflammation and oxidative stress	Lin et al. (2006)
Human lung cell	Metal and metal oxides	Inflammation and oxidative stress	Limbach et al. (2007)
Human cancer cells	Graphene oxide [0–100 mg/L]	Oxidative stress	Zhang et al. (2010)

9.10 Conclusion

Nanoparticles are the most promising and widely exploited resources of present time. Their surface properties makes them unique, thus they are being used in every sector of society. To make and prove this technology as a boon we need to take some simple measures. By taking care while handling the engineered nanomaterials, by properly studying the fate of the synthesized nanoparticle, also by implementing ecofriendly techniques that consumes minimum energy, labour and cost effective methods we can add beauty to our living world inspite of accelerating the pollution rate. Use of chemically engineered nanoparticles should be minimizes. Establishment of regulatory guidelines for nanoparticle synthesis is the urgent call, which should

be followed by both the developed and developing nations. Now it's the need of time that without delaying the slogan of "GO Green" should be practiced not in words but in work. One thing is to be kept in mind i.e. the life cycle of engineered material should be well studied and the large scale synthesis of them should be executed on the basis of their fate in the environment. In the quest of innovation and advancement environmental issues are becoming more serious and uncontrolled. So, let's shake a leg towards the betterment of the earth in which we are living in.

References

- Aaseth J, Olsen A, Halse J, Hovig T (1981) Argyria-tissue deposition of silver as selenide. *Scand J Clin Lab Invest* 41:247–251
- Adams LK, Lyon DY, Alvarez PJJ (2006) Comparative ecotoxicity of nanoscale TiO₂, SiO₂, and ZnO water suspensions. *Water Res* 40:3527–3532
- Akhavan O, Ghaderi E (2010) Toxicity of graphene and graphene oxide nano walls against bacteria. *ACS Nano* 4:5731–5736. doi:10.1021/nn101390x
- Albright LJ, Wilson EM (1974) Sub-lethal effects of several metallic salt- organic compounds combinations upon heterotrophic microflora of a natural water. *Water Res* 8:101–105
- Alfadul SM, Elneshwy AA (2010) Use of nanotechnology in food processing, packaging and safety—review. *Afr J Food Agric Nutr* 10:2719–2739
- Alivisatos AP, Gu W, Larabell C (2005) Quantum dots as cellular probes. *Annu Rev Biomed Eng* 7:55–76
- Armitage SA, White MA, Wilson HK (1996) The determination of silver in whole blood and its application to biological monitoring of occupationally exposed groups. *Ann Occup Hyg* 40:331–338
- Arora S, Jain J, Rajwade JM, Paknikar KM (2009) Interactions of silver nanoparticles with primary mouse fibroblasts and liver cells. *Toxicol Appl Pharmacol* 236:310–318
- Aruoja V, Dubourguier HC, Kasemets K, Kahru A (2009) Toxicity of nanoparticles of CuO, ZnO and TiO₂ to microalgae *Pseudokirchneriella subcapitata*. *Sci Total Environ* 407(4):1461–1468
- Atha DH, Wang H, Petersen EJ (2012) Copper oxide nanoparticle mediated DNA damage in terrestrial plant models. *Environ Sci Technol* 46(3):1819–1827
- Austin LA, Kang B, Yen CW, El-Sayed MA (2011) Nuclear targeted silver nanospheres perturb the cancer cell cycle differently than those of nanogold. *Bioconjug Chem* 22:2324–2331
- Babynin EV, Nuretdinov IA, Gubskaja VP, Barabanshchikov BI (2002) Study of mutagenic activity of fullerene and some of its derivatives using his? Reversions of *Salmonella typhimurium* as an example. *Russ J Genet* 38:359–363
- Baun A, Hartmann NB, Grieger K, Kisk KO (2008) Ecotoxicity of engineered nanoparticles to aquatic invertebrates: a brief review and recommendations for future toxicity testing. *Ecotoxicology* 17(5):387–395. doi:10.1007/s10646-008-0208-y
- Benn TM, Westerhoff P (2008) Nanoparticle silver released into water from commercially available sock fabrics. *Environ Sci Technol* 42:4133–4139
- Biswas P, Wu C (2005) Nanoparticles and the environment. *J Air Waste Manage Assoc* 55:708–746
- Blinova I, Ivask A, Heinlaan M, Mortimer M, Kahru A (2010) Ecotoxicity of nanoparticles of CuO and ZnO in natural water. *Environ Pollut* 158(1):41–47. doi:10.1016/j.envpol.2009.08.017
- Bondarenko O, Ivask A, Käkinen A, Kahru A (2012) Sub-toxic effects of CuO nanoparticles on bacteria: kinetics, role of Cu ions and possible mechanisms of action. *Environ Pollut* 169:81–89

- Boosalis MG, McCall JT, Ahrenhalz DH, Solem LH, McClain CJ (1987) Serum and urinary silver levels in thermal injury patients. *Surgery* 101:40–43
- Borm PJ, Kreyling WJ (2004) Toxicological hazards of inhaled nanoparticles-potential implications for drug delivery. *J Nanosci Nanotechnol* 4:521–551
- Bosi S, Da Ros T, Castellano S, Banfi E, Prato M (2000) Antimycobacterial activity of ionic fullerene derivatives. *Bioorg Med Chem Lett* 10:1043–1045
- Bosi S, Feruglio L, Da Ros T, Spalluto G, Gregoret B, Terdoslavich M, Decorti G, Passamonti S, Moro S, Prato M (2004) Hemolytic effects of water-soluble fullerene derivatives. *J Med Chem* 47:6711–6715
- Bouwmeester H, Dekkers S, Noordam M, Hagens W, Bulder A, de Heer C, ten Voorde I S, Wijnhoven S, Sips A (nd) Health risks of application of nanotechnologies and nanoparticles within the food production chain. *RIKILT/RIVM Report* 2007.014
- Braydich-Stolle L, Hussain S, Schlager JJ, Hofmann MC (2005) In vitro cytotoxicity of nanoparticles in mammalian germline stem cells. *Toxicol Sci* 88:412–419
- Brayner R, Ferrari-Iliou R, Brivois N, Djediat S, Benedetti MF, Fievet F (2010) ZnO nanoparticles: synthesis, characterization, and ecotoxicological studies. *Langmuir* 26(9):6522–6528
- Brigger I, Dubernet C, Couvreur P (2002) Nanoparticles in cancer therapy and diagnosis. *Adv Drug Deliv Rev* 54:631–51 PII: S0169-409X(02)00044-3
- Brown DM, Wilson MR, MacNee W, Stone V, Donaldson K (2001) Size dependent proinflammatory effects of ultrafine polystyrene particles: a role for surface area and oxidative stress in the enhanced activity of ultrafines. *Toxicol Appl Pharmacol* 175:191–199. doi:[10.1006/taap.2001.9240](https://doi.org/10.1006/taap.2001.9240)
- Buza C, Ivan I, Blandino P, Robbie K (2007) Nanomaterials and nanoparticles: sources and toxicity. *Biointerphases* 2(4):MR17–MR172. doi:[10.1116/1.2815690](https://doi.org/10.1116/1.2815690)
- Cameron FK (1915) Soil colloids and the soil solution. *J Phys Chem* 19(1):1–13. doi: [10.1021/j150154a001](https://doi.org/10.1021/j150154a001)
- Canas JE, Long M, Nations S, Vadan R, Dai L, Luo M, Ambikapathi R, Lee EH, Olszyk D (2008) Effects of functionalized and nonfunctionalized single-walled carbon nanotubes on root elongation of selected crop species. *Environ Toxicol Chem* 27:1922–1931
- Cerkez I, Kocer HB, Worley SD, Broughton RM, Huang TS (2012) Multifunctional cotton fabric: antimicrobial and durable press. *J Appl Polym Sc* 124(5):4230–4238. doi:[10.1002/app.35402](https://doi.org/10.1002/app.35402)
- Chang MCO, Chow JC, Watson JG, Hopke PK, Yi SM, England GC (2004) Measurement of ultra-fine particle size distributions from coal-, oil-, and gas-fired stationary combustion sources. *J Air Waste Manag Assoc* 54:1494–1505
- Chen X, Schluesener HJ (2007) Nanosilver: a nanoproduct in medical application. *Toxicol Lett* 176:1–12
- Cheng D, Yang J, Zhao Y (2004) Antibacterial materials of silver nanoparticles application in medical appliances and appliances for daily use. *Chin Med Equip J* 4:26–32
- Chernousova S, Epple M (2013) Silver as antibacterial agent: ion, nanoparticle, metal. *Angew Chem*. doi:[10.1002/anie.201205923](https://doi.org/10.1002/anie.201205923)
- Coccini T, Manzo L, Roda E (2013) Safety evaluation of engineered nanomaterials for health risk assessment: an experimental tiered testing approach using pristine and functionalized carbon nanotubes. *ISRN Toxicol* 2013, Article ID 825427, 13 pages
- Cui DX, Tian FR, Ozkan CS, Wang M, Gao HJ (2005) Effect of single wall carbon nanotubes on human HEK293 cells. *Toxicol Lett* 155:73–85
- Cushen M, Kerry J, Morris M, Cruz-Romero M, Cummins E (2012) Nanotechnologies in the food industry – recent developments, risks and regulation. *Trends Food Sci Tech* 24:30–46
- Da Silva LC, Oliva MA, Azevedo AA, DeAraujo JM (2006) Responses of resting a plant species to pollution from an iron pelletization factory. *Water Air Soil Pollut* 175:241–256. doi:[10.1007/s11270-006-9135-9](https://doi.org/10.1007/s11270-006-9135-9)
- Daniel MC, Astruc D (2004) Gold nanoparticles: assembly, supramolecular chemistry, quantum-size-related properties and applications towards biology, catalysis and nanotechnology. *Chem Rev* 104:293–346. doi:[10.1021/cr030698](https://doi.org/10.1021/cr030698)

- Dastjerdi R, Montazer M (2010) A review on the application of inorganic nano-structured materials in the modification of textiles: focus on anti-microbial properties. *Colloid Surf B* 79(1):5–18
- De Windt W, Boon N, Van den Bulcke J, Rubberecht L, Prata F, Mast J, Hennebel T, Verstraete W (2006) Biological control of the size and reactivity of catalytic Pd(0) produced by *Shewanella oneidensis*. *Anton Leeuw Int J Gen Mol Microbiol* 90:377–389
- Dietl HW, Anzil AP, Mehraein P (1984) Brain involvement in generalized argyria. *Clin Neuropathol* 3:32–38
- Dinesh R, Anandaraj M, Srinivasan V, Hamza S (2012) Engineered nanoparticles in the soil and their potential implications to microbial activity. *Geoderma* 173–174:19–27
- Du W, Sun Y, Ji R, Zhu J, Wu J, Guo H (2011) TiO₂ and ZnO nanoparticles negatively affect wheat growth and soil enzyme activities in agricultural soil. *J Environ Monit* 13(4):822–828
- Esterkin CR, Negro AC, Alfano OM, Cassano AE (2005) Air pollution remediation in a fixed bed photocatalytic reactor coated with TiO₂. *AIChE J* 51:2298–2310. doi:10.1002/aic.10472
- Fisher NM, Marsh E, Lazova R (2003) Scar-localized argyria secondary to silver sulfadiazine cream. *J Am Acad Dermatol* 49:730–732
- Fortner JD, Lyon DY, Sayes CM, Boyd AM, Falkner JC, Hotze EM, Alemany LB, Tao YJ, Guo W, Ausman KD, Colvin VL, Huges JB (2005) C60 in water: nanocrystal formation and microbial response. *Environ Sci Technol* 39:4307–4316
- Gimbert LJ, Hamon RE, Casey PS, Worsfold PJ (2007) Partitioning and stability of engineered ZnO nanoparticles in soil suspensions using field-flow fractionation. *Environ Chem* 4:8–10
- Goodman CM, McCusker CD, Yilmaz T, Rotello VM (2004) Toxicity of gold nanoparticles functionalized with cationic and anionic side chains. *Bioconjug Chem* 15:897–900
- Greene RM, Su WPD (1987) Argyria *Am Fam Physician* 36:151–154
- Grubek-Jaworska H, Nejman P, Czuminska K, Przybylowski T, Huczko A, Lange H, Bystrzejewski M, Baranowski P, Chazan R (2006) Preliminary results on the pathogenic effects of intratracheal exposure to one-dimensional nanocarbons. *Carbon* 44:1057–1063
- Gulson B, Wong H (2006) Stable isotope tracing—a way forward in nanotechnology. *Environ Health Perspect* 114:1486–1488
- Gurr JR, Wang ASS, Chen CH, Jan KY (2005) Ultrafine titanium dioxide particles in the absence of photoactivation can induce oxidative damage to human bronchial epithelial cells. *Toxicology* 213:66–73
- Gustafsson O, Kruså M, Zencak Z, Sheesley RJ, Granat L, Engström E, Praveen PS, Rao PS, Leck C, Rodhe H (2009) Brown clouds over South Asia: biomass or fossil fuel combustion? *Science* 323(5913):495–498
- Hardman R (2006) A toxicologic review of quantum dots: toxicity depends on physicochemical and environmental factors. *Environ Health Perspect* 114:165–172
- Hildemann LM, Markowski GR, Jones MC, Cass GR (1991) Submicrometer aerosol mass distributions of emissions from boilers, fireplaces, automobiles, diesel trucks, and meat-cooking operations. *Aerosol Sci Technol* 14:138–152
- Hoet PH, Bruske-Hohlfeld I, Salata OV (2004) Nanoparticles known and unknown health risks. *J Nanobiotechnol* 2:12. doi:10.1186/1477-3155-2-12
- Hogan CJ, Lee MH, Biswas P (2004) Capture of viral particles in soft X-ray-enhanced corona systems: charge distribution and transport characteristics. *Aero Sci Technol* 38:475–486
- Holbrook RD, Murphy KE, Morrow JB, Cole KD (2008) Trophic transfer of nanoparticles in a simplified invertebrate food web. *Nat Nanotechnol* 335–355. doi:10.1038/nnano.2008.110
- Horikoshi S, Serpone N (2013) Introduction to nanoparticles microwaves in nanoparticle synthesis. In: Satoshi H, Nick S (eds) *Microwaves in nanoparticle synthesis: fundamentals and applications*, 1st edn. Wiley-VCH Verlag GmbH & Co/KGaA, Berlin. ISBN ISBN-978-3-527- 64814-6
- Hu CW, Li M, Cui YB, Li DS, Chen J, Yang LY (2010) Toxicological effects of TiO₂ and ZnO nanoparticles in soil on earthworm *Eiseniafetida*. *Soil Biol Biochem* 42(4):586–591. doi:10.1016/j.soilbio.2009.12.007

- Hua NP, Kobayashi F, Iwasaka Y, Shi G, Naganuma T (2007) Detailed identification of desert originated bacteria carried by Asian dust storms to Japan. *Aerobiologia* 23(4):291–298. doi:10.1007/s10453-007-9076-9
- Huang L, Li DQ, Lin YJ, Wei M, Evans DG, Duan X (2005) Controllable preparation of Nano-MgO and investigation of its bactericidal properties. *J Inorg Biochem* 99:986–993
- Huang Y, Chen S, Bing X, Gao C, Wang T, Yuan B (2011) Nanosilver migrated into food—simulating solutions from commercially available food fresh containers. *Packag Technol Sci* 24:291–297
- Hund-Rinke K, Simon M (2006) Ecotoxic effect of photocatalytic active nanoparticles TiO₂ on algae and daphnids. *Environ Sci Pollut Res* 13:225–232
- Hussain S, Hess K, Gearhart J, Geiss K, Schlager J (2005) In vitro toxicity of nanoparticles in BRL 3A rat liver cells. *Toxicol In Vitro* 19:975–983
- IPPIC (International Paint and Printing Ink Council) (2012) <http://www.ippic.org/site/assets/docs/Public%20AFWG/IPPIC%20%20Zinc%20oxide%20-%20regulatory%20status%20-final%20Draft%20March%201%202012.pdf>
- Iwata N, Mukai T, Yamakoshi TN, Hara S, Yanase T, Shoji M, Endo T, Miyata N (1998) Effects of C₆₀, a fullerene, on the activities of glutathione s-transferase and glutathione-related enzymes in rodent and human livers. *Fuller Sci Technol* 6(2):213–226
- Jia G, Wang HF, Yan L, Wang X, Pei RJ, Yan T, Zhao YL, Guo XB (2005). Cytotoxicity of carbon nano materials: single wall nanotube, multi-wall nanotube and fullerene. *Environ Sci Technol* 39:1378–1383. doi:10.1021/es0487291
- Jiang W, Mashayekhi H, Xing B (2009) Bacterial toxicity comparison between nano and micro-scaled oxide particles. *Environ Pollut* 157(5):1619–1625
- Kai Y, Komazawa Y, Miyajima A, Miyata N, Yamakoshi Y (2003) Fullerene as a novel photoinduced antibiotic. *Fullerenes Nanotubes Carbon Nanostruct* 11:79–87
- Kaida T, Kobayashi K, Adachi M, Suzuki F (2004) Optical characteristics of titanium oxide interference film and the film laminated with oxides and their applications for cosmetics. *J Cosmet Sci* 55:219–220
- Kang S, Pinault M, Pfefferle LD, Elimelech M (2007) Single walled carbon nanotubes exhibit strong antimicrobial activity. *Langmuir* 23:8670–8673
- Karathanasis AD (1999) Subsurface migration of copper and zinc mediated by soil colloids. *Soil Sci Soc Am J* 63:830–838
- Kasemets K, Ivask A, Dubourguier HC, Kahru A (2009) Toxicity of Nanoparticles of ZnO, CuO and TiO₂ to yeast *Saccharomyces cerevisiae*. *Toxicology In Vitro* 23(6):1116–1122. pii/S0887233309001209
- Kennedy AC, Smith KL (1995) Soil microbial diversity and the sustainability of agricultural soils. *Plant Soil* 170:75–86
- Kennedy CB, Scott SD, Ferris FG (2004) Hydrothermal phase stabilization of 2-line Ferrihydrite by bacteria. *Chem Geol* 212(3–4):269–277. pii/S0009254104003158
- Khare P, Sonane M, Pandey R, Ali S, Gupta KC, Satish A (2011) Adverse effects of TiO₂ and ZnO nanoparticles in soil nematode, *Caenorhabditis elegans*. *J Biomed Nanotechnol* 7(1):116–117. <http://dx.doi.org/10.1166/jbn.2011.1229>
- Kim Y, Kim J, Cho H, Rha D, Kim J, Park J, Choi B, Lim R, Chang H, Chung Y, Kwon I, Jeong J, Han B, Yu I (2008a) Twenty-eight-day oral toxicity, genotoxicity, and gender related tissue distribution of silver nanoparticles in Sprague-Dawley rats. *Inhal Toxicol* 20:575–583
- Kim K, Sung W, Moon S, Choi J, Kim J, Lee D (2008b) Antifungal effect of silver nanoparticles on dermatophytes. *J Microbiol Biotechnol* 18:1482–1484
- Kittelson DB (1998) Engines and nanoparticles: a review. *J Aero Sci* 29:575–588
- Klaine SJ, Alvarez PJJ, Batley GE (2008) Nanomaterials in the environment: behavior, fate, bio-availability, and effects. *Environ Toxicol Chem* 27(9):1825–1851
- Kloepfer JA, Mielke RE, Nadeau JL (2005) Uptake of CdSe and CdSe/ZnS quantum dots into bacteria via purine-dependent mechanisms. *Appl Environ Microbiol* 71:2548–2557

- Koyama S, Endo M, Kim YA, Hayashi T, Yanagisawa T, Osaka K, Koyama H, Haniu H, Kuroiwa N (2006) Role of systemic T-cells and histopathological aspects after subcutaneous implantation of various carbon nanotubes in mice. *Carbon* 44:1079–1092
- Kumari M, Khan SS, Pakrashi S, Mukherjee A, Chandrasekaran N (2011) Cytogenetic and genotoxic effects of zinc oxide nanoparticles on root cells of *Allium cepa*. *J Hazard Mater* 190(1–3):613–621
- Li L, Cruz MLF, Connolly M, Conde E, Fernandez M, Schuster M, Navas JM (2015) The potentiation effect makes the difference: nontoxic concentration of ZnO nanoparticles enhance Cu nanoparticle toxicity In-vitro. *Sci Total Environ* 505:253–260. pii/S0048969714014582
- Limbach LK, Wick P, Manser P, Grass RN, Bruinink A, Stark WJ (2007) Exposure of engineered nanoparticles to human lung epithelial cells: influence of chemical composition and catalytic activity on oxidative stress. *Environ Sci Technol* 41:4158–4163
- Lin D, Xing B (2007) Phytotoxicity of nanoparticles: inhibition of seed germination and root growth. *Environ Pollut* 150(2):243–250
- Lin W, Huang YW, Zhou XD, Ma Y (2006) Toxicity of cerium nanoparticles in human lung cancer cells. *Int J Toxicol* 25:451–457
- Lin S, Reppert J, Hu Q, Hudson JS, Reid ML, Ratnikova TA, Rao AM, Luo H, Ke PC (2009) Uptake, translocation, and transmission of carbon nanomaterials in rice plants. *Small* 5:1128–1132. doi:10.1002/sml.200801556
- Lorenz C, Hagedorfer H, von Goetz N, Kaegi R, Gehrig R, Ulrich A, Scheringer M, Hungerbühler K (2011) Nanosized aerosols from consumer sprays: experimental analysis and exposure modeling for four commercial products. *J Nanopart Res* 13:3377–3391
- Lyon DY, Fortner JD, Sayes CM, Colvin VL, Hughes JB (2005) Bacterial cell association and antimicrobial activity of a C-60 water suspension. *Environ Toxicol Chem* 24:2757–2762
- Lyon DY, Adams LK, Falkner JC, Alvarez PJJ (2006) Antibacterial activity of fullerene water suspensions: effects of preparation method and particle size. *Environ Sci Technol* 40:4360–4366
- Ma Y, Kuang L, He X et al (2010) Effects of rare earth oxide nanoparticles on root elongation of plants. *Chemosphere* 78(3):273–279
- Ma H, Williams PL, Diamond SA (2013) Ecotoxicity of manufactured ZnO nanoparticles - a review. *Environ Pollut* 172:76–85. pii/S0269749112003958
- Mahapatra O, Bhagat M, Gopalakrishnan C, Arunachalam KD (2008) Ultrafine dispersed CuO nanoparticles and their antibacterial activity. *J Exp Nanosci* 3:185–193
- Manning MR, Lowe DC, Moss RC, Bodeker GE, Allan W (2005) Short-term variations in the oxidizing power of the atmosphere. *Nature* 436:1001–1004. doi:10.1038/nature03900
- Manzo S, Rocco A, Carotenuto R, De Luca Picione F, Miglietta M, Rametta G, Di Francia G (2011) Investigation of ZnO nanoparticles' ecotoxicological effects towards different soil organisms. *Environ Sci Pollut Res* 18(5):756–763
- Mashino T, Okuda K, Hirota T, Hirobe M, Nagano T, Mochizuki M (1999) Inhibition of *E. coli* growth by fullerene derivatives and inhibition mechanism. *Bioorg Med Chem Lett* 9:2959–2962
- Mashino T, Nishikawa D, Takahashi K, Usui N, Yamori T, Seki M, Endo T, Mochizuki M (2003) Antibacterial and antiproliferative activity of cationic fullerene derivatives. *Bioorg Med Chem Lett* 13:4395–4397
- Mashino T, Usui N, Okuda K, Hirota T, Mochizuki M (2003) Respiratory chain inhibition by fullerene derivatives: hydrogen peroxide production caused by fullerene derivatives and a respiratory chain system. *Bioorg Med Chem Lett* 11:1433–1438
- McMurry PH, Woo KS, Weber R, Chen DR, Pui DYH (2000) Size distributions of 3–10 nm atmospheric particles: implications for nucleation mechanisms. *Philos Trans R Soc Lond Ser a-Math Phys Eng Sci* 358:2625–2642
- Miao AJ, Zhang XY, Luo Z, Chen CS, Chin WC, Santschi PH, Quigg A (2010) Zinc oxide engineered nanoparticles: dissolution and toxicity to marine phytoplankton. *Environ Toxicol Chem* 29(12):2814–2822

- Miller RJ, Lenihan HS, Muller EB, Tseng N, Hanna SK, Keller AA (2010) Impacts of metal oxide nanoparticles on marine phytoplankton. *Environ Sci Technol* 44(19):7329–7334
- Mishra VK, Kumar A (2009) Impact of metal nanoparticles on plant growth promoting rhizobacteria. *Dig J Nanomater Biostruct* 4:587–592
- Monteiller C, Tran L, MacNee W, Faux S, Jones A, Miller B, Donaldson K (2007) The pro-inflammatory effects of low-toxicity low-solubility particles, nanoparticles and fine particles, on epithelial cells in vitro: the role of surface area. *Occup Environ Med* 64:609–615
- Moore MN (2006) Do nanoparticles present ecotoxicological risks for the health of the aquatic environment? Elsevier. doi:[10.1016/j.envint.2006.06.014](https://doi.org/10.1016/j.envint.2006.06.014)
- Morones JR, Elechiguerra JL, Camacho A, Holt K, Kouri JB, Ramirez JT, Yacaman MJ (2005) The bactericidal effect of silver nanoparticles. *Nanotechnology* 16:2346–2353
- Mortimer M, Kasemets K, Kahru A (2010) Toxicity of ZnO and CuO nanoparticles to ciliated protozoa *Tetrahymena thermophila*. *Toxicology* 269(2-3):182–189. doi:[10.1016/j.tox.2009.07.007](https://doi.org/10.1016/j.tox.2009.07.007)
- Mushtaq YK (2011) Effect of nanoscale Fe₃O₄, TiO₂ and carbon particles on cucumber seed germination. *J Environ Sci Health A* 46(14):1732–1735. doi:[10.1080/10934529.2011.633403](https://doi.org/10.1080/10934529.2011.633403)
- Nations S, Wages M, Canas JE, Maul J, Theodorakis C, Cobb GP (2011a) Acute effect of Fe₂O₃, TiO₂, ZnO and CuO nanomaterials on *Xenopus laevis*. *Chemosphere* 83(8):1053–1061. pii/S0045653511001019
- Navarro E, Baun A, Behra R, Hartmann NB, Filser J, Miao AJ, Quigg A, Santschi PH, Sigg L (2008) Environmental behavior and ecotoxicity of engineered nanoparticles to algae, plants, and fungi. *Ecotoxicol* 17:372–386
- Nielsen HD, Berry LS, Stone V, Burridge TR, Fernandes TF (2008) Interactions between carbon black nanoparticles and the brown algae *Fucus serratus*: inhibition of fertilization and zygotic development. *Nanotoxicol* 2:88–89. doi:[10.1080/17435390802109185](https://doi.org/10.1080/17435390802109185)
- Nowack B, Bucheli TD (2007) Review -occurrence, behavior and effects of nanoparticles in the environment. *Environ Pollut* 150:5–22. doi:[10.1016/j.envpol.2007.06.006](https://doi.org/10.1016/j.envpol.2007.06.006)
- Nowack B, Krug HF, Height M (2011) 120 years of nanosilver history: implications for policy makers. *Environ Sci Technol* 45(4):1177–1183
- Nyberg L, Turco RF, Nies L (2008) Assessing the impact of nanomaterials on anaerobic microbial communities. *Environ Sci Technol* 42:1938–1943
- Oberdorster E (2004) Manufactured nanomaterials (fullerenes, C₆₀) induce oxidative stress in the brain of juvenile largemouth bass. *Environ Health Perspect* 112:1058–1062. doi:[10.1289/ehp.7021](https://doi.org/10.1289/ehp.7021)
- Oberdörster E, Zhu S, Blickley TM, McClellan-Green P, Haasch ML (2006) Ecotoxicology of carbon-based engineered nanoparticles: effects of fullerene (C₆₀) on aquatic organisms. *Carbon* 44:1112–1120
- Panyala NR, Pena-Mendez EM, Havel J (2008) Silver or silver nanoparticles: a hazardous threat to the environment and human health? Review. *J Appl Biomed* 6:117–129
- Park B, Donaldson K, Duffin R, Tran L, Kelly F, Mudway I, Morin JP, Guest R, Jenkinson P, Samaras Z, Giannouli M, Kouridis H, Martin P (2008) Hazard and risk assessment of a nanoparticulate cerium oxide based diesel fuel additive—a case study. *Inhal Toxicol* 20:547–566
- Patlolla AK (2013) Environmental toxicity monitoring of nanomaterials using vicia faba GENE-TOX assay. *J Nanomed Nanotechnol* 4:e129. doi:[10.4172/2157-7439.1000e129](https://doi.org/10.4172/2157-7439.1000e129)
- Poland CA, Duffin R, Kinloch I et al (2008) Carbon nanotubes introduced into the abdominal cavity of mice show asbestos like pathogenicity in a pilot study. *Nat Nanotechnol* 3:423–428
- Porter AE, Muller K, Skepper J, Midgley P, Welland M (2006) Uptake of C₆₀ by human monocyte macrophages, its localization and implications for toxicity: studied by high resolution electron microscopy and electron tomography. *Acta Biomater* 2:409–419
- Premanathan M, Karthikeyan K, Jeyasubramanian K, Manivannan G (2011) Selective toxicity of ZnO nanoparticles toward Gram-positive bacteria and cancer cells by apoptosis through lipid peroxidation. *Nanomed Nanotechnol Biol Med* 7(2):184–192

- Pulskamp K, Diabate S, Krug HF (2007) Carbon nanotubes show no sign of acute toxicity but induce intracellular reactive oxygen species in dependence on contaminants. *Toxicol Lett* 168:58–74
- Raloff J (2003) Air sickness: how microscopic dust particles causes subtle but serious harm. *Sci News* 164(5):1–11. doi:[10.2307/3982189](https://doi.org/10.2307/3982189)
- Ravi Kumar M, Hellermann G, Lockey RF, Mohapatra S (2004) Nanoparticle mediated gene delivery: state of the art. *Expert Opin Biol Ther* 4(8):1213–1224. doi:[10.1517/14712598.4.8.1213](https://doi.org/10.1517/14712598.4.8.1213)
- Reidy B, Haase A, Andreas L, Kenneth A, Dawson K, Lynch I (2013) Mechanisms of silver nanoparticle release, transformation and toxicity: a critical review of current knowledge and recommendations for future studies and applications. *Materials* 6:2295–2350
- Remédios C, Rosário F, Bastos V (2012) Environmental nanoparticles interactions with plants: morphological, physiological and genotoxic aspects. Review Article. *J Bot Article ID* 751686, 8 pages
- Rincon A, Pulgarin C (2004) Effect of pH, inorganic ions, organic matter and H₂O₂ on E. coli K12 photocatalytic inactivation by TiO₂ implications in solar water disinfection. *Appl Catal B Environ* 51:283–302
- Roh J, Sim SJ, Yi J, Park K, Chung KH, Ryu D, Choi J (2009) Ecotoxicity of silver nanoparticles on the soil nematode *Caenorhabditis elegans* using functional ecotoxicogenomics. *Environ Sci Technol* 43:3933–3940
- Rosenman KD, Seixas N, Jacobs I (1987) Potential nephrotoxic effects of exposure to silver. *Br J Ind Med* 44:267–272
- Royal society and Royal Academy of Engineering (2004) Nanoscience and nanotechnologies: opportunities and uncertainties, RS policy document, 19/04. The Royal Society, London, p 113
- Rozhkov SP, Goryunov AS, Sukhanova GA, Borisova AG, Rozhkova NN, Andrievsky GV (2003) Protein interaction with hydrated C₍₆₀₎ fullerene in aqueous solutions. *Biochem Biophys Res Commun* 303:562–566
- Sajid M, Ilyas M, Basheer C, Tariq M, Daud M, Baig N, Shehzad F (2015) Impact of nanoparticles on human and environment: review of toxicity factors, exposures, control strategies, and future prospects. *Environ Sci Pollut Res* 22:4122–4143. doi:[10.1007/s11356-014-3994-1](https://doi.org/10.1007/s11356-014-3994-1)
- Saliba AM, de Assis MC, Nishi R, Raymond B, Marques EA, Lopes UG, Touqui L, Plotkowski MC (2006) Implications of oxidative stress in the cytotoxicity of *Pseudomonas aeruginosa* ExoU. *Microbes Infect* 8:450–459
- Sau TK, Rogach AL, Jäckel F, Klar TA, Feldmann J (2010) Properties and applications of colloidal nonspherical noble metal nanoparticles. *Adv Mater* 22(16):1805–1825
- Sawai J, Igarashi H, Hashimoto A, Kokugan T, Shimizu M (1995) Effect of ceramic powder slurry on spores of *Bacillus subtilis*. *J Chem Eng Jpn* 28:556–561
- Sawai J, Igarashi H, Hashimoto A, Kokugan T, Shimizu M (1996) Effect of particle size and heating temperature of ceramic powders on antibacterial activity of their slurries. *J Chem Eng Jpn* 29:251–256
- Sayes CM, Fortner JD, Guo W, Lyon D, Boyd AM, Ausman KD, Tao YJ, Sitharaman B, Wilson LJ, Hughes JB (2004) The differential cytotoxicity of water-soluble fullerenes. *Nano Lett* 4:1881–1887
- Seaman JC, Bertsch PM (2000) Selective colloid mobilization through surface-charge manipulation. *Environ Sci Technol* 34:3749–3755
- Serita F, Kyono H, Seki Y (1999) Pulmonary clearance and lesions in rats after a single inhalation of ultrafine metallic nickel at dose levels comparable to the threshold limit value. *Ind Health* 37:353–363
- Serpone N, Dondi D, Albini A (2007) Inorganic and organic UV filters: their role and efficacy in sunscreens and sun care products. *Inorg Chim Acta* 360:794–802
- Shin SH, Ye MK, Kim HS, Kang HS (2007) The effects of nano-silver on the proliferation and cytokine expression by peripheral blood mononuclear cells. *Int Immunopharmacol* 7:1813–1821

- Simkiss K (1990) Surface effects in ecotoxicology. *Funct Ecol* 4(3):303–308. StableURL:<http://www.jstor.org/stable/2389590>
- Smita S, Gupta SK, Bartonova A, Dusinska M, Gutleb AC, Rahman Q (2012) Nanoparticles in the environment: assessment using the causal diagram approach. *Environ Health* 11:1–11
- Sondi I, Salopek-Sondi B (2004) Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for Gram negative bacteria. *J Colloid Interf Sci* 275:177–182
- Song W, Zhang J, Guo J, Zhang J, Ding F, Li L, Sun Z (2010) Role of the dissolved zinc ion and reactive oxygen species in cytotoxicity of ZnO nanoparticles. *Toxicol Lett* 199(3):389–397
- Stampoulis D, Sinha SK, White JC (2009) Assay-dependent phytotoxicity of nanoparticles to plants. *Environ Sci Technol* 43(24):9473–9479
- Strawa AW, Kirchstetter TW, Hallar AG, Ban-Weiss GA, McLaughlin JP, Harley RA, Lunden MM (2010) Optical and physical properties of primary on-road vehicle particle emissions and their implications for climate change. *J Aerosol Sci* 41: 36–50, No. 1,(January 2010), ISSN0021-8502
- Sung J, Ji J, Yoon J, Kim D, Song M, Jeong J, Han B, Han J, Chung Y, Kim J, Kim T, Chang H, Lee E, Lee J, Yu I (2008) Lung function changes in Sprague-Dawley rats after prolonged inhalation exposure to silver nanoparticles. *Inhal Toxicol* 20:567–574
- Takenaka S, Karg E, Kreyling W, Lentner B, Schulz H, Ziesenis A, Schramel P, Heyder J (2004) Fate and toxic effects of inhaled ultrafine cadmium oxide particles in the rat lung. *Inhal Toxicol* 16:83–92
- Tang H, Wang D, Ge X (2004) Environmental nanopollutants (ENP) and aquatic micro interfacial processes. *Water Sci Technol* 50(12):103–109. ISSNPrint:0273–1223
- The Royal Society (2004) Nanoscience and nanotechnologies: opportunities and uncertainties. The Royal Society & The Royal Academy of Engineering, London. ISBN 0 85403 604 0
- Thill A, Spalla O, Chauvat F, Rose J, Auffan M, Flank AM (2006) Cytotoxicity of CeO₂ nanoparticles for *Escherichia coli*: a physico-chemical insight of the cytotoxicity mechanism. *Environ Sci Technol* 40:6151–6156
- Tromp TK, Shia RL, Allen M, Eiler JM, Yung YL (2003) Potential environmental impact of a hydrogen economy on the stratosphere. *Science* 300:1740–1742. doi:[10.1126/science.1085169](https://doi.org/10.1126/science.1085169)
- Tsao N, Luh TY, Chou CK, Chang TY, Wu JJ, Liu CC, Lei HY (2002) In vitro action of carboxy-fullerene. *J Antimicrob Chemother* 49:641–649
- UNEP Assessment Report (nd) The Asian brown cloud: climate and other environmental impacts. UNEP/DEWA/RS; 2002.02-3[<http://www.rrcap.unep.org/abc/impactstudy/>]
- Velzeboer I, Kupryianchik D, Peeters ETH, Koelmans AA (2011) Community effects of carbon nanotubes in aquatic sediments. *Environ Int* 37(6):1126–1130. pii/S0160412011000328
- Vincent JH, Clement CF (2000) Ultrafine particles in workplace atmospheres. *Philos Trans R Soc Lond Ser a-Math Phys Eng Sci* 358:2673–2682
- Wang K, Ruan J, Song H et al (2011) Biocompatibility of graphene oxide. *Nanoscale Res Lett* 6:1–8
- Warheit DB, Laurence BR et al (2004) Comparative pulmonary toxicity assessment of single-wall carbon nanotubes in rats'. *Toxicol Sci* 77:117–125
- Warheit DB, Webb TR, Sayes CM, Colvin VL, Reed KL (2006) Pulmonary instillation studies with nanoscale TiO₂ rods and dots in rats: toxicity is not dependent upon particle size and surface area. *Toxicol Sci* 91:227–236
- Wei W, Sethuraman A, Jin C, Monteiro-Riviere NA, Narayan RJ (2007) Biological properties of carbon nanotubes. *J Nanosci Nanotechnol* 7:1284–1297
- Wijnhoven SWP, Peijnenburg WJGM, Herberts CA, Hagens W, Oomen AG, Heugens EHV, Roszek B, Bisschops J, Gosens I, van de Meent D et al (2009) Nano-silver—a review of available data and knowledge gaps in human and environmental risk assessment. *Nanotoxicology* 3:109–138
- Wild E, Jones KC (2009) Novel method for the direct visualization of in vivo nanomaterials and chemical interactions in plants. *Environ Sci Technol* 43:5290–5294

- Wilson SR, Solomon KR, Tang X (2007) Changes in tropospheric composition and air quality due to stratospheric ozone depletion and climate change. *Photochem Photobiol Sci* 6:301. doi:[10.1039/B700022G](https://doi.org/10.1039/B700022G)
- Wolfrum EJ, Huang J, Blake DM, Maness PC, Huang Z, Fiest J, Jacoby WA (2002) Photocatalytic oxidation of bacteria, bacterial and fungal spores, and model biofilm components to carbon dioxide on titanium dioxide-coated surfaces. *Environ Sci Technol* 36:3412–3419
- Wright JB, Lam K, Hansen D, Burrell RE (1999) Efficacy of topical silver against fungal burn wound pathogens. *Am J Infect Control* 27:344–350
- Yang L, Watts DJ (2005) Particle surface characteristics may play an important role in phytotoxicity of alumina nanoparticles. *Toxicol Lett* 158:122–132
- Zhang W (2003) Nanoscale iron particles for environmental remediation: an overview. *J Nanopart Res* 5(3):323–332. doi:[10.1023/A:1025520116015](https://doi.org/10.1023/A:1025520116015)
- Zhang Y, Ali SF, Dervishi E et al (2010) Cytotoxicity effects of graphene and single-wall carbon nanotubes in neural pheochromocytoma-derived pc12 cells. *ACS Nano* 4:3181–3186
- Zhang P, Ma Y, Zhang Z et al (2012) Comparative toxicity of nanoparticulate/bulk Yb_2O_3 and YbCl_3 to cucumber (*Cucumis sativus*). *Environ Sci Technol* 46(3):1834–1841
- Zhu Y, Zhao Q, Li Y, Cal X, Li W (2006) The interaction and toxicity of multi-walled carbon nanotubes with *Stylonychia mytilus*. *J Nanosci Nanotechnol* 6:1357–1364
- Zhu X, Zhu L, Li Y, Duan Z, Chen W, Alvarez PJJ (2007) Developmental toxicity in zebrafish (*Danio rerio*) embryos after exposure to manufactured nanomaterials: buckminsterfullerene aggregates (nC60) and fullerol. *Environ Toxicol Chem* 26(5):976–979