Chapter 8 Cancer Immunity and Immune Evasion Mechanisms

Stalin Chellappa, Einar M. Aandahl, and Kjetil Taskén

Abstract Understanding the role of the immune system in cancer development and progression is a challenging process. The collective efforts unequivocally show that the immune system is playing a dual role in promoting and inhibiting tumor development. The tumor microenvironment is highly infiltrated by immune cells, which includes innate (macrophages, mast cells, neutrophils, dendritic cells, and myeloidderived suppressor cells) and adaptive (T and B) cells. This diverse set of cells contributes to the secretion of different pro-inflammatory immune mediators creating a microenvironment that influences cancer growth in a pleiotropic manner. It is the composition of inflammatory mediators and the activation status of different immune cells that interact with the tumor to dictate either tumor regression or tumor progression. CD4+ and CD8+ T cells play a pivotal role in anticancer immunity. The CD4+ T cells are instrumental in eliminating cancer cells by secreting various cytokines and activating and recruiting other cell types such as macrophages and granulocytes. However, CD4+ T cell-mediated activation of CD8+ T cells and subsequent cytotoxic activity of the CD8+ T cells represent the major effector mechanism of antitumor immunity. Here, we review and discuss the current knowledge with respect to the functional role and prognostic significance of individual T cell subsets in various malignancies.

Keywords CD4+ T cells • CD4+ Th cell subsets • CD8+ CTLs • Tregs • GATA3 RORγt • T-bet • FOXP3 • Cytokines • Immunosuppression • Plasticity • Tumorinfiltrating T cells • Prognosis

S. Chellappa, Ph.D • E.M. Aandahl, M.D., Ph.D • K. Taskén, M.D., Ph.D (\boxtimes) Centre for Molecular Medicine Norway, Nordic EMBL Partnership, University of Oslo, Oslo, Norway e-mail: kjetil.tasken@ncmm.uio.no

[©] Springer International Publishing Switzerland 2017 195

L.A. Akslen, R.S. Watnick (eds.), *Biomarkers of the Tumor Microenvironment*, DOI 10.1007/978-3-319-39147-2_8

Cancer Immunoediting and Tumor Immune Evasion Mechanisms

While the role of the immune system in controlling microbial pathogens is well appreciated, the notion that the immune system can also control tumor development and progression has been a controversy for over a century. In 1909, Paul Ehrlich was the first to suggest that the immune system could protect the host from malignancies [\[1](#page-14-0)]. Nearly 50 years later, Thomas and Burnet predicted that adaptive immunity is responsible for preventing tumor formation and progression in an immunocompetent host and proposed the concept of cancer immunosurveillance [[2,](#page-14-1) [3\]](#page-14-2). However, due to the absence of experimental support, the cancer immunosurveillance concept was abandoned. This was largely due to the lack of mouse models with pure genetic backgrounds available at that time. By the 1990s, with improved genetically modified mouse models available, several seminal works have validated the role of cancer immunosurveillance in both chemically induced and spontaneous tumor models [\[4](#page-14-3)]. Multiple components of the immune system have been identified as having central roles in cancer immunosurveillance, such as T cells, B cells, natural killer (NK) cells and INFγ, and perforin [[4,](#page-14-3) [5](#page-14-4)]. Similarly, several experimental and clinical studies have confirmed the existence of cancer immunosurveillance (T cell-mediated cancer immunosurveillance is described in detail in the following sections) [[5\]](#page-14-4). Collectively, these findings suggest that cancer immunosurveillance can function as a microenvironmental tumor suppressor. However, despite the presence of an active cancer immunosurveillance process, many immunocompetent individuals still develop cancer. This paradox has been explained via seminal mice studies showing that the immune system not only eliminates but also reduces the immunogenicity of the tumor but also has the capability to promote tumor growth [[4\]](#page-14-3). This led to a significant revision of the original cancer immunosurveillance theory wherein Robert Schreiber and colleagues proposed a new concept termed "cancer immunoediting," which emphasized the cancer-promoting and cancer-suppressing role of the immune system during tumor growth [\[4](#page-14-3), [6\]](#page-14-5). Cancer immunoediting consists of three phases, elimination, equilibrium, and escape, and termed as "three Es of cancer immunoediting" [\[6](#page-14-5)]. The elimination phase represents the original concept of cancer immunosurveillance, in which the cooperative actions of innate and adaptive immunity eliminate the tumor before it is clinically manifest. Several studies suggest that the immune component required for the elimination of tumors depends on specific-tumor characteristics such as origin (spontaneous vs. carcinogen-induced), anatomical location, histology, and growth rate. During the elimination phase, rare tumor cell variants may survive and enter into an equilibrium state. In this period, tumor cells undergo antigenicity sculpting by immune cells applying a selective pressure leading to the survival of the fastest growing cells that escape elimination by the immune system. This process induces reduced immunogenicity and acquired resistance to immune effector cells. The equilibrium state is the longest phase, and it extends throughout the life of the host. The end stage of the equilibrium phase results in generation of several tumor clones with the most immunoevasive

Evasion strategy	Mechanism			
Impaired tumor antigen presentation	• Downregulation of tumor antigens or antigen-processing machinery (e.g., lack of LMP and TAP proteins) [8] • Downregulation of MHC genes [9]			
Impaired trafficking of immune cells into tumor microenvironment	• Epigenetic silencing of chemokine expression [10] • Lack of endothelial adhesion molecules $[11-13]$ • Physical barrier by stroma [14] • Lack of tumor antigens in lymphoidal organs [15]			
Immune cell dysfunction or subversion	• Immune suppression mediated by CD4+ FOXP3+ regulatory T cells (Tregs) and myeloid-derived suppressor cells (MDSCs) [16–20] • Secretion of suppressive cytokines (TGF- β , IL-10, etc.) [21–23] and other soluble immunosuppressive factors (prostaglandins, VEGF, RCAS1, extracellular adenosine, reactive oxygen, nitrogen species, etc.) $[24-28]$ • Expression of IDO in tumor cells leading to secretion of immunosuppressive tryptophan metabolites [29] • Induction of T cell tolerance by expressing cognate ligands for T cell checkpoint inhibitory receptors such as CTLA-4, PD-1, LAG-3, and Tim-3 $[30, 31]$ • Apoptosis of immune cells induced by tumor cell expression of CD95L (FasL) (tumor counterattack) [32] triggering CD95 (Fas)-mediated T cell apoptosis • Immune cell deviation and plasticity [33-36]			
Tumor cell resistance to apoptosis	• Abnormal expression of anti-apoptotic molecules (Bcl-2 and IAPs family protein) $[37]$ • Mutations or loss of pro-apoptotic molecules (TRAIL and CD95 receptors) $[37]$ • Interference with granzyme/perforin pathway [38, 39]			

Table 8.1 Tumor immune evasion mechanisms

mutations and epigenetic instability. These cells ultimately enter the escape phase and develop into visible tumors and successfully avoid immune destruction, which is now considered as an emerging hallmark of cancers as described by Hanahan and Weinberg [[7\]](#page-14-6). Tumor cells evade the protective immunity by several mechanisms as presented in Table [8.1.](#page-2-0) Currently, targeting one or more of these mechanisms clinically holds the most promising approach to improve antitumor immunity [\[24](#page-15-0)].

T Lymphocytes and Cancer Immunity

T cells are generally classified into two lineages: CD4+ T cells and CD8+ T cells. CD4+ T cells are further classified into CD4+ T helper cells (Th) that mediate tumor immunity and CD4 + FOXP3+ regulatory T cells (Tregs) that suppress antitumor immunity (described later). Naïve T cells that express a unique T cell receptor (TCR) on the surface develop through stringent positive and negative selection pathways in the thymus. T cells migrate through tissues and scan for cognate antigen peptide-MHC complex that activates their TCR, resulting in functional differentia-tion into a variety of subsets [[40\]](#page-16-0). Here we focus on conventional $TCR\alpha/\beta T$ cell subsets and their role in tumor immunity.

CD4+ T Cells in Anticancer Immunity

CD4+ Th cells are crucial in orchestrating humoral and cell-mediated immune responses [\[41\]](#page-16-1). However, their role in anticancer immunity is complex and reflects the diverse role of various CD4+ Th cell subsets [[33\]](#page-15-12). The CD4+ Th cell TCR recognizes antigenic epitopes in the form of 12–20-residue long peptides, presented by major histocompatibility complex II (MHC-II) expressed on professional antigen-presenting cells (APCs) which include dendritic cells (DCs), macrophages, and B cells [\[42\]](#page-16-2). Upon recognition of antigen on the APC surface by the TCR along with appropriate interaction of co-stimulatory receptors such as CD28 on T cells with ligands such as CD80/86 on APCs leads to naïve CD4+ Th cell activation [\[43\]](#page-16-3), which results in clonal expansion, triggered effector functions, and subsequent memory formation. During this period, the fate and functional specialization of activated CD4+ Th cells are largely dependent on the concentration and source of antigen, the type of APC engaged by CD4+ Th cells, the co-stimulatory receptors expressed by APCs, and, most importantly, the polarizing cytokine milieu of the microenvironment at the time of activation that drives the naïve CD4+ Th cells toward a particular Th cell subtype [[40](#page-16-0)]. Together, these polarizing factors contribute to the specific expression of key subset-defining transcriptional factors and the subsequent secretion of effector cytokines that defines the functional subsets of CD4+ Th cells [\[40\]](#page-16-0). The cytokines secreted by CD4+ Th cells then activate and recruit a variety of other immune effector cells that together define the type of immune response [[41\]](#page-16-1). Table [8.2](#page-3-0) summarizes the CD4+ Th cell subsets in the human and murine systems, the polarizing cytokines that drive their development, their master transcription factors, and the effector cytokines they secrete.

Th			
subset	Polarizing cytokine	Transcription factor	Effector cytokine
Th ₁	IL-12, IL-18, INF γ , $II - 27$	T-bet, STAT4	IL-2, IL-10, INF γ , TNF- α , TNF- β $(LT-\alpha)$, CCL2, CCL3
Th2	IL-4, IL-25, IL-33, TSLP	GATA3, IRF4, STAT6	IL-4, IL-5, IL-9, IL-10, IL-13, IL-21, IL-31, TNF- α
Th17	$TGF-\beta$, IL-1 β , IL-6, IL-21, IL-23	RORγt, RORα, IRF4 Batf, STAT3	IL-17A, IL-17F, IL-21, IL-22, IL-26 (human), TNF- α , CCL20
Th22	IL-6, IL-13, TNF- α	AHR, Batf, STAT3	IL-10, IL-13, IL-22, IL-21, TNF- α , IL-26 (human)
Th ₉	$TGF-\beta, IL-4$	PU.1, IRF4	$IL-9$, $IL-10$
Tfh	IL-6, IL-21	Bcl6, BATF, c-MAF	IL-4, IL-10, IL-12, IL-21, INFy

Table 8.2 CD4 + Th cell subsets: polarizing cytokines, master transcription factors, and effector cytokines

Conventional Role of CD4+ T Cells in CD8+ Cytotoxic T Cell (CTL) Responses

CD4+ Th cells play an essential role in priming, activation, and expansion of CTL responses, a concept known as $CD4+T$ cell help $[44-46]$ $[44-46]$. $CD4+T$ cell help is complex and involves multiple mechanisms broadly classified into direct and indirect help. During the primary immune response to the tumor, the major indirect help from activated CD4+ Th cell comes through CD40/CD40L interaction with APCs that leads to maturation of the APCs [\[47](#page-16-6)[–49](#page-16-7)]. This process provides all three necessary signals for CD8+ T cell activation, including antigen-mediated TCR triggering, co-stimulation, and stimulatory cytokines, most notably IL-12, which are all critically important for naïve antigen-specific CD8+ T cells to differentiate into CTLs. Alternatively, CD4+ Th cells can directly activate CTLs through CD40/CD40L [\[50](#page-16-8)]. Furthermore, activated CD4+ Th cells also directly help CTLs through secretion of IL-2, which supports growth and expansion [[51,](#page-16-9) [52](#page-16-10)]. Furthermore, secretion of INFγ by CD4+ Th1 cells upregulates the expression of MHC molecules on the surface of tumor cells leading to a feed-forward loop of enhanced CTL responses as well as CD4+ Th responses [\[53](#page-16-11)]. In addition to priming the primary CTL response, CD4+ Th cells also help during the post-priming stage that takes place at the tumor site [[54,](#page-16-12) [55\]](#page-16-13). Moreover, tumor-specific CD4+ Th cells have been shown to enhance the expansion of both low-avidity [[56\]](#page-16-14) and cognate [\[57](#page-16-15)] CTLs at the tumor site and enhance tumor rejection. In addition to their support to optimize CTL responses, CD4+ Th cells also play an essential role in generation and maintenance of memory CD8+ T cells during active CTL responses and homeostatic proliferation [\[58](#page-16-16), [59\]](#page-16-17). Hosts lacking CD4+ Th cells have been shown to have reduced number of CD8+ memory T cells and impaired secondary CD8+ T cell responses [\[60](#page-16-18)].

Unconventional Role of CD4+ T Cells in Tumor Immunity

CD4+ Th cell-mediated antitumor immunity is primarily thought to involve activation and maintenance of CTL responses. However, more recent studies have shown that CD4+ Th cells also play independent roles in antitumor immunity. Here we discuss the specific roles of different CD4+ Th cell subsets in antitumor immunity.

CD4+ Th1 Cells in Tumor Immunity

In 1986, Mossman and Coffman demonstrated that antigen-specific mouse CD4+ Th cells can be categorized into two types, Th1 and Th2, based on their pattern of cytokine production [\[61](#page-16-19)]. In 1991, Romagnani and colleagues discovered that human CD4+ Th clones specific for intracellular *Mycobacterium tuberculosis* were mostly Th1 cells, whereas the CD4+ Th clones specific for the extracellular helminth *Toxocara canis* were mainly Th2 cells [[62\]](#page-17-0). This firmly established the Th1/ Th2 paradigm in both human and mice. The Th1 lineage is controlled by the key

transcription factor T-bet and the key polarizing cytokine IL-12 [[40,](#page-16-0) [63](#page-17-1)[–65](#page-17-2)]. Th1 cells secrete a set of cytokines that includes IL-2, INF γ , and TNF- α and the chemo-kines CCL2 and CCL3 that attract macrophages (Table [8.2\)](#page-3-0), and they are best characterized for their role in clearance of intracellular pathogens such as viruses and their role in the pathogenesis of autoimmune conditions [\[66](#page-17-3)]. Th1 cells are considered to have potent antitumor activity due to their secretion of INFγ, IL-2, and CD40/CD40L co-stimulation to help initiate CD8+ T cell responses as described earlier [[58\]](#page-16-16). Several human Th1 cells can also mediate antitumor immunity independently of helping CTL responses. INFγ plays a crucial role in antitumor responses and acts directly on tumor cells as well as promoting immune cell responses against tumor cells [[67,](#page-17-4) [68](#page-17-5)]. Interestingly, an earlier study in mice demonstrated that Th1 cell-mediated INFγ secretion in the tumor microenvironment is essential for inhibiting angiogenesis and regression of tumors that do not express MHC-II [[69\]](#page-17-6). Similarly, a study of mouse B cell cancer suggests that Th1 cellmediated INFγ secretion in the tumor microenvironment is essential for eliminating MHC-II-negative tumor cells through activation of type 1 macrophages and angiogenic inhibitors like IP-10 [[70\]](#page-17-7). However, their mechanistic relevance in human cancer is yet to be determined. Furthermore, a key function of Th1-derived INFγ in tumor-bearing hosts is to substantially increase the IL-12 secretion by DCs, which serves to further polarize the naïve CD4+ T cells into a Th1 phenotype, thereby contributing to their own development and maintenance [[71\]](#page-17-8). In addition, secretion of cytokines and chemokines by Th1 cells also leads to recruitment and activation of pro-inflammatory type 1 macrophages (M1) and natural killer (NK) cells at the cancer site [[68,](#page-17-5) [72,](#page-17-9) [73\]](#page-17-10). The cytotoxic mediators secreted from type 1 macrophages and NK cells have multiple antitumor properties [[74,](#page-17-11) [75](#page-17-12)]. In line with this, patient studies show that the presence of Th1 cells and increased levels of their associated cytokines correlate with superior antitumor immunity and good clinical outcome in a majority of cancers [\[76](#page-17-13)]. Despite their potent antitumor role, Th1 cell functions are efficiently hindered by tumor cells by varying suppressive factors (Table [8.1](#page-2-0) and described later), and imbalance or alterations in Th1/Th2 ratio in many human cancers lead to poor clinical outcome [[77\]](#page-17-14). Owing to their importance, Th1 cells are also being utilized in clinical studies. Adoptive transfer of tumor antigen-specific Th1 cells in patients with metastatic melanoma [[78\]](#page-17-15) and metastatic cholangiocarcinoma [\[79](#page-17-16)] was recently shown to induce regression of the tumor for prolonged periods. In contrast, responses in melanoma patients that received only autologous, in vitro-expanded, tumor-infiltrating CD8+ T lymphocytes (TILs) [[80\]](#page-17-17) were found to be suboptimal and suggest the importance of inducing tumor antigen-specific Th1 cells for successful antitumor immunity.

CD4+ Th2 Cells in Tumor Immunity

CD4+ Th2 cells are recognized for their role in the host defense against extracellular parasites and their involvement in allergy and asthma. In both mice and humans, Th2 lineage commitment is controlled by the transcription factor GATA3 and exposure to the polarizing cytokine IL-4 in the microenvironment of APC-naïve CD4+ Th cells [\[40](#page-16-0), [81](#page-17-18), [82\]](#page-17-19). Th2 cells then produce their signature cytokines such as IL-4, IL-5, IL-13, and IL-10 (Table [8.2](#page-3-0)). These cytokines mutually antagonize the development of Th1 cells [\[40](#page-16-0), [64](#page-17-20)]. Th2 cells have been extensively studied for their role in antitumor immunity and in the context of disease progression and disease outcome. Initial studies from murine models and in vitro studies showed that IL-4 secreted from Th2 cells has a direct anti-angiogenic and tumoricidal activity [[83–](#page-17-21) [85\]](#page-18-0). IL-4 and IL-13 are critical for the recruitment of eosinophils and macrophages and in some cases neutrophils and CD8+ T cells to the tumor site and result in regression of tumor [\[86](#page-18-1)[–90](#page-18-2)]. Conversely, Th2 cytokines also interfere with antitumor activity, which is largely attributed to Th2 cytokines that antagonize the development of INFγ-secreting Th1 and CTLs at the cancer site. IL-4 and IL-13 have an anti-apoptotic role [\[91](#page-18-3)[–94](#page-18-4)], and IL-13 also has a pro-fibrotic role [[95,](#page-18-5) [96\]](#page-18-6) that may affect antitumor activity. Numerous studies indicate that the Th1/Th2 ratio is altered in a variety of cancers [\[76](#page-17-13), [77\]](#page-17-14). Initial murine studies suggested that both Th1 and Th2 cells contribute to antitumor immunity [[73,](#page-17-10) [97,](#page-18-7) [98\]](#page-18-8). However, the increased presence of Th2 cells was found to be pro-carcinogenic in many human cancers [[33,](#page-15-12) [76,](#page-17-13) [99](#page-18-9), [100](#page-18-10)]. These pro-tumorigenic roles of Th2 cells were proposed to be cancer specific rather than a global effect, as the Th1 response in these patients was not impaired [\[101](#page-18-11), [102\]](#page-18-12). Multiple tumor-derived factors may favor the development of Th2 cells. Tumor cell-derived IL-10 induces skewing toward Th2 cells and inhibits the maturation of dendritic cells (DCs), which effectively reduces the secretion of INFγ and IL-12 from T cells resulting in impaired antitumor activity [[103,](#page-19-0) [104\]](#page-19-1). Early reports demonstrated that human renal cell carcinoma and non-small cell lung cancer actively produced Th2-polarizing cytokines [[105,](#page-19-2) [106](#page-19-3)]. Pancreatic cancer, an aggressive malignancy, is typically infiltrated by Th2 cells [\[107](#page-19-4)]. A clinical study from pancreatic cancer patients showed that the skewing toward Th2 was primarily due to the secretion of thymic stromal lymphopoietin from cancer-associated fibroblasts that activate DCs to produce Th2-associated cytokines and polarize T cells toward Th2 cells [[108\]](#page-19-5). A similar mechanism was observed in mouse models of breast cancer $[109]$ $[109]$, and chronic gastritis $[110]$ $[110]$, which is the causative factor for gastric cancer. Similarly, studies in mice have shown that the expression of the human tumor antigen EpCAM strongly promotes Th2 skewing despite of the presence of strong Th1-polarizing conditions [\[111](#page-19-8)]. Thus, the involvement of Th2 cells in antitumor immunity is still controversial and that their effect may be context dependent.

CD4+ Th17 Cells in Tumor Immunity

In 2005, a third subset of CD4+ Th cells was identified in mice and named as Th17 cells based on the production of the cytokine IL-17 [\[112](#page-19-9), [113](#page-19-10)]. Two years later, the existence of Th17 cells was confirmed in the human immune system [[114,](#page-19-11) [115](#page-19-12)]. The development of Th17 cells are controlled by the master transcription factor RORγt and multiple polarizing cytokines [\[116–](#page-19-13)[118\]](#page-19-14) (Table [8.2\)](#page-3-0). Owing to their inflammatory properties, Th17 cells have been studied in a number of diseases both in mice and in humans and found to be important in the host defense against extracellular bacteria and fungi, but pathogenic in many inflammatory and autoimmune diseases [[34,](#page-15-17) [116,](#page-19-13) [119](#page-19-15), [120](#page-19-16)]. Th17 cells are shown to infiltrate several cancer types in both mice and humans [\[34](#page-15-17)]. However, their exact role in antitumor immunity is controversial and still elusive. Contradictory findings with respect to their role in antitumor immunity versus a pro-carcinogenic role may be due to the existence of multiple flavors of Th17 cells that are fostered by different cancerous cell types and mediators in the cancer microenvironment. Furthermore, the use of a variety of mouse tumor models adds complexity to this issue. Evidence for the role of Th17 cells in antitumor immunity came from studies with established models of B16 melanoma [[122\]](#page-19-17), and B16/F10 lung metastatic melanoma [\[123](#page-19-18)] in mice, in which adoptive transfer of in vitro-expanded, tumor antigen-specific Th17 cells induced regression of the cancer to a larger extent than Th1 cells transferred in a parallel experiment. The transfused Th17 cells were found to promote the infiltration of DCs and enhanced cross-antigen presentation to naïve CD8+ T cells as well as to induce the secretion of CCL20 from cancer-residing lung cells to further recruit CD8+ CTLs into the tumor site [\[123](#page-19-18)]. Therefore, the Th17 cells were proposed to have a synergistic function with CD8+ CTLs. In contrast, other tumor models in mice, which included leukemia [\[124\]](#page-20-0), cervical cancer [\[125\]](#page-20-1), non-small cell lung cancer [\[126](#page-20-2)], lung cancer [[127](#page-20-3)], and colon cancer [[128\]](#page-20-4), suggested that Th17 cell-secreted inflammatory cytokines in the tumor microenvironment promoted neutrophil recruitment and secretion of elastase, a pro-tumorigenic factor [[129\]](#page-20-5). They also promoted the secretion of pro-angiogenic factors and pro-inflammatory cytokines from tumor cells, which promote angiogenesis and cancer progression [[129\]](#page-20-5). Recent studies with genetically modified mice with colon cancer [[130\]](#page-20-6) and pancreatic cancer [[131\]](#page-20-7) showed that the preinvasive epithelial layer expressed large amounts of IL-17R that facilitated the infiltration of Th17 cells further substantiating the above findings. Subsequently, the IL-17A derived from Th17 cells triggered the oncogenic signal through the IL-17R-STAT3 pathway and accelerated the transformation of epithelial cells into invasive neoplasia. Recently, β-catenin signaling was also implicated in the development of Th17 cells in colon cancer [[132](#page-20-8)]. Similar dichotomous findings were observed in human cancer patients where infiltration of Th17 cells was positively associated with CD8+ T cell count and better survival in ovarian cancer [\[133](#page-20-9)] and esophageal cancer [[134\]](#page-20-10), whereas increasing evidence suggests the opposite in many solid tumors [[34,](#page-15-17) [76\]](#page-17-13).

Th17 cells are also found to be a major fraction of TILs in human cancers, attracted by tumor-derived RANTES and MCP-1 [\[135](#page-20-11), [136\]](#page-20-12). Human Th17 cells also undergo plasticity (secreting cytokines of other lineages) [\[117](#page-19-19), [120\]](#page-19-16). Interestingly, in vitro-expanded, tumor antigen-specific Th17 clones from melanoma and breast and colon cancer produced large amounts of polyfunctional cytokines including IL-8 and TNF- α , but not IL-2, IL-4, IL-12, or IL-23 [[135\]](#page-20-11). Furthermore, the same authors also suggested that Th17 cells can be converted into FOXP3-expressing, Treg, cells that produce IL-10 and TGF-β1, indicating a possible regulatory function [[137\]](#page-20-13). In contrast, other studies suggest that in vitroexpanded, tumor antigen-specific Th17 clones from colon cancer and ulcerative colitis mainly produced IL-2, TNF- α , INF γ , and GM-CSF and exhibited plasticity to convert into FOXP3- and INFγ-expressing cells with suppressive properties [\[129](#page-20-5), [133,](#page-20-9) [138\]](#page-20-14). These findings were contrasted by the proposed cytokine signature of freshly isolated Th17 cells from healthy patients [[139\]](#page-20-15) and argue that these differences may arise from in vitro induced changes or may reflect their actual function in the cancer microenvironment. The conversion of Th17 cells into Th1 cells is well documented in autoimmune diseases and cancer [\[117](#page-19-19), [120\]](#page-19-16). However, recent findings have shown that ex vivo-isolated Th17 cells from peripheral blood mononuclear cells (PBMCs) of human pancreatic cancer patients can also produce Th2 and Th17 cytokines [\[140](#page-20-16)]. Notably, these findings demonstrate that Th17 cells from human cancers not only correlate with IL-17 secretion but can also acquire Th1- or Th2-associated features. To summarize, Th17 cell-mediated antitumor immunity is due to enhancement of DC and CD8+ CTL function. However, Th17 cells also contribute to cancer-promoting inflammation and angiogenesis. Further, their plasticityassociated complexity in the tumor microenvironment may determine their pro-tumorigenic, suppressive, or anti-tumorigenic role that may influence cancer prognosis.

CD8+ Cytotoxic T Lymphocytes (CTLs) in Cancer Immunity

CTLs recognize their cognate antigen through binding of their TCR to antigen-MHC-I complex expressed on the surface of tumor cells. Th cells also provide help to CTL responses (see the section "Unconventional Role of CD4+ T Cells in Tumor Immunity"). CTLs potentially eliminate the tumor cells and have been shown to correlate with good prognosis in almost every type of human malignancy (Table [8.3\)](#page-9-0). CD8+ T cells use multiple mechanisms to kill tumor cells mediated by granzyme B, perforin, and the triggering of the Fas signaling pathway through Fas ligand (FasL). FasL expressed on CTLs binds to its cognate receptor on the tumor cell surface and induces apoptosis. Similarly, perforin secreted by activated CTLs forms pores on the surface of tumor cells that aid in directed delivery of granzyme B into the tumor cell that subsequently induces apoptosis. In addition, naïve CD8+ T cells also differentiate into different subsets such as Tc1 (Tbet+ Eomes+ INFγ+), Tc2 (GATA3+ IL-4+), and Tc17 (RORγt+ Tbet+ IL-17+) cells, which are driven by master transcription factors and polarizing cytokines similar to those described for Th1, Th2, and Th17 cells (Table [8.2\)](#page-3-0) and also produce key cytokines similar to that of Th subsets (Fig. [8.1\)](#page-10-0). Since type 1-, 2-, and 17-related cytokines are mainly produced by Th subsets rather than Tc subsets in the cancer microenvironment, their functional relevance is not yet clearly known. However, recent studies in mice suggest that T cells secrete INFy, but not IL-4 and TNF- α , in a directional way (at the immunological synapse) onto the target cell [[205\]](#page-24-0). It is possible that INFγ secreted by tumor-infiltrating Tc1 cells can have direct antitumor activity by enhancing MHC expression on cancer cells, inducing angiostatic effects, and also recruiting macrophages [[68\]](#page-17-5). The role of IL-4-secreting Tc2 cells in the cancer microenvironment is largely unknown, although a study from breast cancer [[206\]](#page-24-1) showed their association with cancer progression. In contrast to Tc1 cells, IL-17-secreting Tc17 cells were found

Cancer type	$CD8+T$ cells	$CD4+Th1$ cells	$CD4+$ Th ₂ cells	$CD4+Th17$ cells	CD4+Treg cells
Head and neck cancers	Good [141, 1421				Good [142]
Esophageal cancer	Good [143, 1441	Good [145]		Good [134]	
Lung cancer	Good [146]	Good [146]		Poor [147]	Poor [148]
Pancreatic cancer	Good [149, 150]		Poor [108]	Poor $[151]$	Poor $[151, 152]$
Distal bile duct cancer	Good [153a] Poor [153b]				Good [153a]
Breast cancer	Good [154]	Good [155]	Good [156]	Poor [157]	Poor [158, 159] Good [160]
Gastric cancer	Poor [161, 162]	Good [163]	Poor [163]	Good [164] Poor $[165]$	Good [165] Poor [166]
Hepatocellular carcinoma	Good [167, 1681 Poor $[168]$	Good [169]		Poor [170]	Poor [168, 171]
Colon cancer	Good $[172 - 178]$	Good $[172 - 174]$	None [173]	Poor [173, 179, 180]	Good [173, 180-1921 Poor [183] None [176]
Ovarian cancer	Good [184]	Good [185, 186]	Poor [186]	Good [133]	Good [187, 1881 Poor [189]
Renal cell carcinoma	Good [190]	Good [191]			Poor [191]
Prostate cancer	Good [192]				
Urothelial carcinoma	Good [193]				
Endometrial cancer	Good [194]				
Cervical cancer	Good [195] Poor [196]				
Melanoma	Good [197, 1981				None [199] Poor [200, 201]
Follicular and Hodgkin's lymphoma			Good [202]		Good [203, 2041 Poor [202]

Table 8.3 The association of tumor-infiltrating T cell subsets and prognosis

to be impaired in cytotoxic activity [[207,](#page-24-2) [208](#page-24-3)]. However, adoptive transfer studies in mouse tumor models have shown that Tc17 cells inhibited tumor growth, which was primarily associated with their plasticity to convert into Tc17/1 cells that produced INFγ along with IL-17A [\[209](#page-24-4)]. However, Tc17 cells identified in gastric cancer [\[161](#page-22-0)], hepatocellular cancer [\[210](#page-24-5)], distal bile duct cancer [\[153b](#page-21-0)], cervical cancer [\[196](#page-23-0)], breast cancer [[206\]](#page-24-1), and endometrial carcinoma [[211\]](#page-24-6) were primarily found to be less cytotoxic and promoted cancer. Especially in gastric [[161\]](#page-22-0) and cervical cancer [\[196](#page-23-0)], Tc17 cells were shown to promote angiogenesis and to recruit suppressor cells, including myeloid-derived suppressor cells (MSDCs) and Tregs. Therefore,

Fig. 8.1 Role of T cell subsets in antitumor immunity. Th1 cells express T-bet, INFγ, and IL-12. Th1 cell is superior in antitumor activity primarily due to activating APC, M1 macrophages, NK cells, and prime CTL (Tc1 cells) response. Both Th1 cells and Tc1 cells associate with good prognosis in many cancers and form a prominent antitumor axis in humans. Th2 cells express GATA3, IL-4, and IL-13 and contribute to cancer regression via recruiting eosinophils and neutrophils. Furthermore, cytokines produced by Th2 cells also contribute to angiogenesis, recruit M2 macrophages, and have an anti-apoptotic role. Their counterpart the Tc2 cells contribute to cancer regression through their cytotoxic activity, but their possible cancer-promoting features are not clearly known. Th17 cells contribute to cancer regression via activating APC and CTL. However, they may also contribute to cancer progression by various mechanisms. Similarly, their counterpart Tc17 also primarily contributes to cancer progression by recruiting suppressor cells into cancer stroma, mainly Tregs and MDSCs. Both Th17 and Tc17 cells contribute to angiogenesis. Tregs contribute to cancer progression by suppressing the effector functions of Th and Tc cell subsets. Tregs also largely accumulate at the cancer site, and their phenotypic heterogeneity and plasticity also contribute to pro-carcinogenic inflammation and cancer progression. Therefore, Th2, Th17, Tc17, and Treg subsets form a context-dependent axis in antitumor immunity in human malignancy

emerging results suggest that the cytotoxic activity of CD8+ T cells is context dependent, and under specific polarizing conditions, they may potentially lose their cytotoxic activity.

Tumor-Infiltrating T Cell Subsets and Their Prognostic Value

Despite the associations described above of various types of Th and Tc subsets with different cancers, the use of phenotyping of tumor-infiltrating T cell subsets as a prognostic marker is a complicated endeavor. In addition to the complex interactions in the tumor microenvironment, CD4+ Th cells in the tumor can be found in different maturation states such as activated, exhausted, or regulatory. Moreover, they may share phenotypic markers with other immune cells adding more complexity to analyses and interpretations of individual patient TIL profiles. Conflicting conclusions with respect to TIL phenotype could also potentially be due to differences in methodologies used, such as immunohistochemistry (IHC), multicolor flow cytometry, and polymerase chain reaction (PCR). Nonetheless, similar conclusions drawn for a particular cancer type by several groups substantiate the need for studying the link between Th cell subsets and prognosis and/or response to therapy. Here we summarize the prognostic value of analyzing the abundance of Th1, Th2, Th17, and CD8+ T cell subsets in several human malignancies (see Table [8.3](#page-9-0)). Th1 cells and CD8+ T cells are strongly associated with good prognosis in many human cancers including esophageal cancer [[143](#page-20-19)[–145](#page-21-2)], colon cancer [[172–](#page-22-11)[178](#page-22-12)], head and neck cancer [\[141](#page-20-17), [142\]](#page-20-18), lung cancer [\[146](#page-21-3)], pancreatic cancer [[149,](#page-21-6) [150](#page-21-7)], distal bile duct cancer [\[153a\]](#page-21-10), breast cancer [[154,](#page-21-11) [155\]](#page-21-12), gastric cancer [[163\]](#page-22-2), prostate cancer [[192\]](#page-23-2), urothelial cancer [[193\]](#page-23-12), ovarian cancer [\[184](#page-23-4)[–186\]](#page-23-6), endometrial cancer [\[194](#page-23-13)], cervical cancer [\[195](#page-23-14)], hepatocellular carcinoma [[167](#page-22-6)[–169](#page-22-8)], melanoma [[197,](#page-24-7) [198\]](#page-24-8), and renal cell carcinoma [\[190](#page-23-10), [191\]](#page-23-11). Despite this, the presence of CD8+ T cells has also been reported to associate with poor outcome, particularly in hepatocellular carcinoma, gastric cancer, distal bile duct cancer, and cervical cancer (Table [8.3](#page-9-0)), which is thought primarily to be due to conversion of CD8+ T cells into Tc17 cells [[153b,](#page-21-0) [161, 162](#page-22-0), [196\]](#page-23-0). In contrast to Th1 cells and CD8+ T cells, Th2 and Th17 cells correlate with either good or poor prognosis (Table [8.3\)](#page-9-0). Th17 cells have been associated with good prognosis in esophageal cancer [\[134](#page-20-10)], ovarian cancer [[133\]](#page-20-9), and gastric cancer [\[164](#page-22-3)] but correlated with poor prognosis in colon cancer [\[173](#page-22-14), [179\]](#page-22-15), lung cancer [\[147](#page-21-4)], pancreatic cancer [[151\]](#page-21-8), breast cancer [\[157\]](#page-21-14), gastric cancer [\[165\]](#page-22-4), and hepatocellular carcinoma [\[170\]](#page-22-9) (Table [8.3](#page-9-0)). Whereas the presence of Th2 cells is associated with good prognosis in breast cancer [\[156](#page-21-13)] and follicular and Hodgkin's lymphoma [[202](#page-24-12)], their presence associates with poor prognosis in pancreatic cancer [\[108](#page-19-5)], gastric cancer [\[163](#page-22-2)], and ovarian cancer [[186\]](#page-23-6), but does not appear to have an impact on colon cancer prognosis [\[173](#page-22-14)] (Table [8.3\)](#page-9-0). Interestingly, in gastric cancer accumulation of Th17 cells have been shown to associate with either good prognosis irrespective of the cancer stage $[164]$ $[164]$ or poor prognosis at early stage of the cancer $[165]$. These disparities could originate from differences in experimental setup and markers used to define Th17 and Th2 cells. Some of the abovementioned studies used only IL-17 as a predictor, investigating the CD4+ IL-17+ T cells. This may affect the results as other immune cell types including $\gamma \delta$ T cells, myeloid cells, and innate lymphoid cells (ILCs) can also produce IL-17 [[41](#page-16-1), [121\]](#page-19-20). In addition, as we described earlier (see the section "Unconventional Role of CD4+ T Cells in Tumor Immunity"), Th17 cells also undergo plasticity, and therefore the conflicting observation of Th17 cells and Th2 cells may also reflect the fundamental differences in the inflammatory tumor

microenvironment and stress the importance of well-delineated Th lineage analysis in these patients. In addition, Galon and colleagues earlier proposed a concept termed "immune contexture" in which the location and density of CD8+ T cells and CD4+ Th cells in both the invasive margin and intra-tumoral region predicted a favorable outcome in colorectal cancer patients [\[172](#page-22-11), [212\]](#page-24-15). Recently, this particular immune contexture has also been demonstrated in other cancer types [\[153a,](#page-21-10) [213](#page-24-16), [214\]](#page-24-17). These findings provide a framework to further standardize the studies that involve T cell subset association with prognosis in human cancer.

CD4+ T Cells Suppressing Antitumor Immunity

CD4 + FOXP3+ Regulatory T Cells (Tregs)

Tregs are crucial in the maintenance of peripheral tolerance and prevention of autoimmunity [[215](#page-24-18)]. The transcription factor FOXP3 is essential for the development and function of Tregs [\[215\]](#page-24-18). Various CD4+ Treg subsets have been identified in humans that can be broadly divided into thymus-derived (tTregs) and peripherally induced Tregs (pTregs). The essential function of Tregs is to suppress the activation, clonal expansion, and effector functions of various immune cells including CD4+ T cells, CD8+ T cells, natural killer T (NKTs) cells, and antigen-presenting cells (APCs) through a myriad of mechanisms [\[216](#page-25-0), [217\]](#page-25-1). Accumulating evidence suggests that Tregs are specifically attracted by chemokines secreted by cancer cells [[16\]](#page-15-3). Similarly, the cancer microenvironment provides a niche to strongly expand Tregs [[218\]](#page-25-2) where the Tregs contribute to the suppression of antitumor immunity initiated by Th cells, CTLs, and other innate immune cell tumors [[17\]](#page-15-18). The conversion of Th cells into pTreg cells has been suggested to account for the high number of Tregs in tumor tissue. However, recent findings using epigenetic analysis of Tregs from tumor sites from mice and human found that a significant proportion of intra-tumoral Tregs were of nTreg origin and suggested that Th to Treg conversion is only partly contributing to the expansion of the Treg population [\[219](#page-25-3)]. Apart from their suppressive function, IL-10 secreted by Tregs can also skew Th subset tumor into a Th2 phenotype, which is associated with poor prognosis in many tumor types (Table [8.3\)](#page-9-0). In addition, recent evidence from many solid tumors especially colon cancer [[183](#page-23-3), [220,](#page-25-4) [221](#page-25-5)], pancreatic cancer [[140](#page-20-16)], and breast cancer [\[222\]](#page-25-6) suggests that IL-17+ FOXP3+ Tregs retain their suppressive function but also contribute to Th17-associated inflammation, which is associated with poor prognosis in these tumor types (Table [8.3](#page-9-0)).

Tumor-Infiltrating Tregs and Their Prognostic Value

Tumor-infiltrating Tregs have been extensively studied and the prognostic value of their presence varies in different tumors. Tregs have been reported to correlate with poor outcome in colon cancer [[183\]](#page-23-3), lung cancer [[148,](#page-21-5) [223](#page-25-7)], pancreatic cancer [\[151](#page-21-8), [152\]](#page-21-9), breast cancer [\[158](#page-21-15), [159\]](#page-21-16), gastric cancer [[166\]](#page-22-5), ovarian cancer [[189\]](#page-23-9), renal cell carcinoma [\[191](#page-23-11)], and hepatocellular carcinoma [[168,](#page-22-7) [171](#page-22-10)] as well as melanoma and follicular and Hodgkin's lymphoma [[202\]](#page-24-12). In contrast, the presence of Tregs was found to be associated with good prognosis in colon cancer [[173,](#page-22-14) [181](#page-23-15), [182\]](#page-23-16), head and neck cancer [[142\]](#page-20-18), distal bile duct cancer [[153a](#page-21-10)], gastric cancer [\[165](#page-22-4)], ovarian cancer, and breast cancer [[160\]](#page-21-17), as well as follicular and Hodgkin's lymphoma [[203,](#page-24-13) [204\]](#page-24-14) (Table [8.3](#page-9-0)). Interestingly, associations with both good and poor prognosis were observed within the same cancer type for colon, breast, gastric, and ovarian cancer and Hodgkin's lymphoma (Table [8.3](#page-9-0)). Moreover, some studies have reported that the presence of Tregs has no impact in colon cancer and melanoma (Table [8.3\)](#page-9-0). These discrepancies in prognostic value may arise from the use of different markers to define Tregs. Other factors that may contribute to these discrepancies are the presence of tumor subtypes where the impact of antitumor immunity varies, tumor stage, and the location of the characterized Tregs (within the tumor tissue, at the margin of the tumor or in the inflamed tissue outside the tumor). Finally, the role of Tregs in cancer progression may also be dependent on whether the cancers were preceded, or stimulated, by inflammation. In addition, many of these studies have not reported Treg-suppressive function or their phenotypic plasticity. The positive impact of Tregs in some tumor types may reflect their anti-inflammatory role in suppressing tumor-promoting inflammation. Moreover, discrepancies within the same tumor type such as colon, breast, and gastric cancer may indicate that Tregs may predominantly share other Th lineage phenotypes, such as IL-17+ FOXP3+ Treg, which have been found to be the major Treg pool in colon, breast, and pancreatic cancer patients [\[140](#page-20-16), [183](#page-23-3), [222](#page-25-6)]. Nonetheless, these data suggest that the original view on Tregs in suppressing antitumor immunity is oversimplified and that Tregs may have multiple roles in influencing inflammation and shaping the tumor microenvironment as well as in suppressing antitumor immunity.

Conclusion

Experimental and clinical studies now indicate that T cells play a pivotal, albeit sometimes paradoxical role in shaping antitumor immunity (Fig. [8.1\)](#page-10-0). Nonetheless, the presence of Th1 and CTL cells is strongly associated with favorable outcomes in many tumor types and indicates that active cancer immunosurveillance is an integral part of many human malignancies. However, the potency of CTL function in several malignant tumors is generally compromised. The main factors contributing to tumor immune evasion include reduced MHC-I and MCH-II expression by tumor cells to eliminate the direct detection by CTLs, along with reduced help from CD4+ Th tumor cells. In addition, the differentiation of CD8+ T cells into less cytotoxic and pro-inflammatory subsets under polarizing conditions in the tumor microenvironment together with Treg-mediated immunosuppression at the cancer site contributes to the functional defect in tumor-specific Th1 cells and CTLs that ultimately lead to tumor progression. In addition, Th2, Th17, and Tregs are largely associated with poor outcome in many tumor types. The bifurcation of the pro- and

anti-tumorigenic nature of T cell subsets is too complex to predict, as it largely depends on cytokines secreted in the cancer microenvironment. To add to this complexity, recent reports suggest that T cells share different lineage-specific transcription factors and exhibit heterogeneity and plasticity. This may explain the paradoxical role of Th2, Th17, and Treg subsets observed, as many earlier studies assessed the prognostic value of individual subsets, but did not consider the potential of phenotypic plasticity. It is also inevitable that the location of T cells and the niche they share with other immune cells, cancer cells, and stromal cells along with their complex interactions dictate their functional status. An integrated picture of all these factors will shed more light on the role of T cells in cancer and enable us to better tailor T cell therapies in the future.

Acknowledgments We apologize to all authors whose work we were unable to cite due to space restrictions. Our work is supported by grants from Research Council of Norway (221938 to E.M. Aandahl; 204784 and 187615 to K. Taskén), Norwegian Cancer Society (741746 to E.M. Aandahl; 419544 to K. Taskén), South Eastern Norway Regional Health Authority (2010038 to E.M. Aandahl), and KG Jebsen Foundation (2012/21 and 2012/23 to K. Taskén and E.M. Aandahl). S. Chellappa is a post-doctoral fellow funded by The Research Council of Norway.

References

- 1. Ehrlich P. Über den jetzigen Stand der Karzinomforschung. Ned Tijdschr Geneeskd. 1909;5:273–90.
- 2. Burnet M. Cancer: a biological approach. III. Viruses associated with neoplastic conditions. IV. Practical applications. Br Med J. 1957;1(5023):841–7.
- 3. Thomas L. Cellular and humoral aspects of the hypersensitive states. H Lawrence, ed. New York:Hoeber-Harper; 1959.
- 4. Dunn GP, Bruce AT, Ikeda H, Old LJ, Schreiber RD. Cancer immunoediting: from immunosurveillance to tumor escape. Nat Immunol. 2002;3(11):991-8.
- 5. Kim R, Emi M, Tanabe K. Cancer immunoediting from immune surveillance to immune escape. Immunology. 2007;121(1):1–14.
- 6. Dunn GP, Old LJ, Schreiber RD. The three Es of cancer immunoediting. Annu Rev Immunol. 2004;22(1):329–60.
- 7. Hanahan D, Weinberg Robert A. Hallmarks of cancer: the next generation. Cell. 2011;144(5):646–74.
- 8. Khanna R. Tumour surveillance: missing peptides and MHC molecules. Immunol Cell Biol. 1998;76(1):20–6.
- 9. Bubenik J. MHC class I down-regulation: tumour escape from immune surveillance? (review). Int J Oncol. 2004;25(2):487–91.
- 10. Peng D, Kryczek I, Nagarsheth N, Zhao L, Wei S, Wang W, et al. Epigenetic silencing of TH1-type chemokines shapes tumour immunity and immunotherapy. Nature. 2015;527(7577):249–53.
- 11. Onrust SV, Hartl PM, Rosen SD, Hanahan D. Modulation of L-selectin ligand expression during an immune response accompanying tumorigenesis in transgenic mice. J Clin Invest. 1996;97(1):54–64.
- 12. Wu T-C. The role of vascular cell adhesion molecule-1 in tumor immune evasion. Cancer Res. 2007;67(13):6003–6.
- 13. Piali L, Fichtel A, Terpe HJ, Imhof BA, Gisler RH. Endothelial vascular cell adhesion molecule 1 expression is suppressed by melanoma and carcinoma. J Exp Med. 1995;181(2):811–6.
- 14. Turley SJ, Cremasco V, Astarita JL. Immunological hallmarks of stromal cells in the tumour microenvironment. Nat Rev Immunol. 2015;15(11):669–82.
- 15. Ochsenbein AF. Principles of tumor immunosurveillance and implications for immunotherapy. Cancer Gene Ther. 2002;9(12):1043–55.
- 16. Mailloux AW, Young MR. Regulatory T-cell trafficking: from thymic development to tumorinduced immune suppression. Crit Rev Immunol. 2010;30(5):435–47.
- 17. Savage PA, Malchow S, Leventhal DS. Basic principles of tumor-associated regulatory T cell biology. Trends Immunol. 2013;34(1):33–40.
- 18. Wolf D, Sopper S, Pircher A, Gastl G, Wolf AM. Treg(s) in cancer: friends or foe? J Cell Physiol. 2015;230(11):2598–605.
- 19. Marvel D, Gabrilovich DI. Myeloid-derived suppressor cells in the tumor microenvironment: expect the unexpected. J Clin Invest. 2015;125(9):3356–64.
- 20. Töpfer K, Kempe S, Müller N, Schmitz M, Bachmann M, Cartellieri M, et al. Tumor evasion from T cell surveillance. J Biomed Biotechnol. 2011;2011:918471.
- 21. Pickup M, Novitskiy S, Moses HL. The roles of TGF[beta] in the tumour microenvironment. Nat Rev Cancer. 2013;13(11):788–99.
- 22. Sato T, Terai M, Tamura Y, Alexeev V, Mastrangelo MJ, Selvan SR. Interleukin 10 in the tumor microenvironment: a target for anticancer immunotherapy. Immunol Res. 2011;51(2–3):170–82.
- 23. Germano G, Allavena P, Mantovani A. Cytokines as a key component of cancer-related inflammation. Cytokine. 2008;43(3):374–9.
- 24. Smyth MJ, Ngiow SF, Ribas A, Teng MWL. Combination cancer immunotherapies tailored to the tumour microenvironment. Nat Rev Clin Oncol. 2015;advance online publication.
- 25. Antonioli L, Blandizzi C, Pacher P, Hasko G. Immunity, inflammation and cancer: a leading role for adenosine. Nat Rev Cancer. 2013;13(12):842–57.
- 26. Brudvik KW, Tasken K. Modulation of T cell immune functions by the prostaglandin $E(2)$ cAMP pathway in chronic inflammatory states. Br J Pharmacol. 2012;166(2):411–9.
- 27. Sonoda K. RCAS1 is a promising therapeutic target against cancer: its multifunctional bioactivities and clinical significance. Expert Rev Obstet Gynecol. 2012;7(3):261–7.
- 28. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. Cell. 2010;140(6):883–99.
- 29. Platten M, Wick W, Van den Eynde BJ. Tryptophan catabolism in cancer: beyond IDO and tryptophan depletion. Cancer Res. 2012;72(21):5435–40.
- 30. Crespo J, Sun H, Welling TH, Tian Z, Zou W. T cell anergy, exhaustion, senescence, and stemness in the tumor microenvironment. Curr Opin Immunol. 2013;25(2):214–21.
- 31. Pauken KE, Wherry EJ. Overcoming T cell exhaustion in infection and cancer. Trends Immunol. 2015;36(4):265–76.
- 32. Peter ME, Hadji A, Murmann AE, Brockway S, Putzbach W, Pattanayak A, et al. The role of CD95 and CD95 ligand in cancer. Cell Death Differ. 2015;22(4):549–59.
- 33. Kim HJ, Cantor H. CD4 T-cell subsets and tumor immunity: the helpful and the not-sohelpful. Cancer Immunol Res. 2014;2(2):91-8.
- 34. Bailey SR, Nelson MH, Himes RA, Li Z, Mehrotra S, Paulos CM. Th17 cells in cancer: the ultimate identity crisis. Front Immunol. 2014;5:276.
- 35. Protti MP, De Monte L. Cross-talk within the tumor microenvironment mediates Th2-type inflammation in pancreatic cancer. OncoImmunology. 2012;1(1):89–91.
- 36. Ostuni R, Kratochvill F, Murray PJ, Natoli G. Macrophages and cancer: from mechanisms to therapeutic implications. Trends Immunol. 2015;36(4):229–39.
- 37. Fulda S. Tumor resistance to apoptosis. Int J Cancer. 2009;124(3):511–5.
- 38. Lehmann C, Zeis M, Schmitz N, Uharek L. Impaired binding of perforin on the surface of tumor cells is a cause of target cell resistance against cytotoxic effector cells. Blood. 2000;96(2):594–600.
- 39. Medema JP, de Jong J, Peltenburg LTC, Verdegaal EME, Gorter A, Bres SA, et al. Blockade of the granzyme B/perforin pathway through overexpression of the serine protease inhibitor

PI-9/SPI-6 constitutes a mechanism for immune escape by tumors. Proc Natl Acad Sci U S A. 2001;98(20):11515–20.

- 40. Zhu J, Yamane H, Paul WE. Differentiation of effector CD4 T cell populations. Annu Rev Immunol. 2010;28(1):445–89.
- 41. Annunziato F, Romagnani C, Romagnani S. The 3 major types of innate and adaptive cellmediated effector immunity. J Allergy Clin Immunol. 2015;135(3):626–35.
- 42. Rossjohn J, Gras S, Miles JJ, Turner SJ, Godfrey DI, McCluskey J. T cell antigen receptor recognition of antigen-presenting molecules. Annu Rev Immunol. 2015;33(1):169–200.
- 43. Chen L, Flies DB. Molecular mechanisms of T cell co-stimulation and co-inhibition. Nat Rev Immunol. 2013;13(4):227–42.
- 44. Keene JA, Forman J. Helper activity is required for the in vivo generation of cytotoxic T lymphocytes. J Exp Med. 1982;155(3):768–82.
- 45. Bennett SR, Carbone FR, Karamalis F, Miller JF, Heath WR. Induction of a CD8+ cytotoxic T lymphocyte response by cross-priming requires cognate CD4+ T cell help. J Exp Med. 1997;186(1):65–70.
- 46. Ossendorp F, Mengede E, Camps M, Filius R, Melief CJ. Specific T helper cell requirement for optimal induction of cytotoxic T lymphocytes against major histocompatibility complex class II negative tumors. J Exp Med. 1998;187(5):693–702.
- 47. Bennett SR, Carbone FR, Karamalis F, Flavell RA, Miller JF, Heath WR. Help for cytotoxic-T-cell responses is mediated by CD40 signalling. Nature. 1998;393(6684):478–80.
- 48. Schoenberger SP, Toes RE, van der Voort EI, Offringa R, Melief CJ. T-cell help for cytotoxic T lymphocytes is mediated by CD40-CD40L interactions. Nature. 1998;393(6684):480–3.
- 49. Ridge JP, Di Rosa F, Matzinger P. A conditioned dendritic cell can be a temporal bridge between a CD4+ T-helper and a T-killer cell. Nature. 1998;393(6684):474–8.
- 50. Bourgeois C, Rocha B, Tanchot C. A role for CD40 expression on CD8+ T cells in the generation of CD8+ T cell memory. Science. 2002;297(5589):2060–3.
- 51. Williams MA, Tyznik AJ, Bevan MJ. Interleukin-2 signals during priming are required for secondary expansion of CD8+ memory T cells. Nature. 2006;441(7095):890–3.
- 52. Tham EL, Shrikant P, Mescher MF. Activation-induced nonresponsiveness: a Th-dependent regulatory checkpoint in the CTL response. J Immunol. 2002;168(3):1190–7.
- 53. Mescher MF, Curtsinger JM, Agarwal P, Casey KA, Gerner M, Hammerbeck CD, et al. Signals required for programming effector and memory development by CD8+ T cells. Immunol Rev. 2006;211:81–92.
- 54. Baxevanis CN, Voutsas IF, Tsitsilonis OE, Gritzapis AD, Sotiriadou R, Papamichail M. Tumor-specific CD4+ T lymphocytes from cancer patients are required for optimal induction of cytotoxic T cells against the autologous tumor. J Immunol. 2000;164(7):3902–12.
- 55. Bos R, Sherman LA. CD4+ T-cell help in the tumor milieu is required for recruitment and cytolytic function of CD8+ T lymphocytes. Cancer Res. 2010;70(21):8368–77.
- 56. Wong SB, Bos R, Sherman LA. Tumor-specific CD4+ T cells render the tumor environment permissive for infiltration by low-avidity CD8+ T cells. J Immunol. 2008;180(5):3122–31.
- 57. Hwang ML, Lukens JR, Bullock TN. Cognate memory CD4+ T cells generated with dendritic cell priming influence the expansion, trafficking, and differentiation of secondary CD8+ T cells and enhance tumor control. J Immunol. 2007;179(9):5829–38.
- 58. Shedlock DJ, Shen H. Requirement for CD4 T cell help in generating functional CD8 T cell memory. Science. 2003;300(5617):337–9.
- 59. Hamilton SE, Wolkers MC, Schoenberger SP, Jameson SC. The generation of protective memory-like CD8+ T cells during homeostatic proliferation requires CD4+ T cells. Nat Immunol. 2006;7(5):475–81.
- 60. Belz GT, Wodarz D, Diaz G, Nowak MA, Doherty PC. Compromised influenza virus-specific CD8(+)-T-cell memory in CD4(+)-T-cell-deficient mice. J Virol. 2002;76(23):12388–93.
- 61. Mosmann TR, Cherwinski H, Bond MW, Giedlin MA, Coffman RL. Two types of murine helper T cell clone. I. Definition according to profiles of lymphokine activities and secreted proteins. J Immunol. 1986;136(7):2348–57.
- 62. Del Prete GF, De Carli M, Mastromauro C, Biagiotti R, Macchia D, Falagiani P, et al. Purified protein derivative of Mycobacterium tuberculosis and excretory-secretory antigen(s) of Toxocara canis expand in vitro human T cells with stable and opposite (type 1 T helper or type 2 T helper) profile of cytokine production. J Clin Invest. 1991;88(1):346–50.
- 63. Szabo SJ, Kim ST, Costa GL, Zhang X, Fathman CG, Glimcher LH. A novel transcription factor, T-bet, directs Th1 lineage commitment. Cell. 2000;100(6):655–69.
- 64. Kanno Y, Vahedi G, Hirahara K, Singleton K, O'Shea JJ. Transcriptional and epigenetic control of T helper cell specification: molecular mechanisms underlying commitment and plasticity. Annu Rev Immunol. 2012;30(1):707–31.
- 65. Hsieh CS, Macatonia SE, Tripp CS, Wolf SF, O'Garra A, Murphy KM. Development of TH1 CD4+ T cells through IL-12 produced by Listeria-induced macrophages. Science. 1993;260(5107):547–9.
- 66. Raphael I, Nalawade S, Eagar TN, Forsthuber TG. T cell subsets and their signature cytokines in autoimmune and inflammatory diseases. Cytokine. 2015;74(1):5–17.
- 67. Zaidi MR, Merlino G. The two faces of interferon-gamma in cancer. Clin Cancer Res. 2011;17(19):6118–24.
- 68. Schroder K, Hertzog PJ, Ravasi T, Hume DA. Interferon-γ: an overview of signals, mechanisms and functions. J Leukoc Biol. 2004;75(2):163–89.
- 69. Qin Z, Blankenstein T. CD4+ T cell—mediated tumor rejection involves inhibition of angiogenesis that is dependent on IFN gamma receptor expression by nonhematopoietic cells. Immunity. 2000;12(6):677–86.
- 70. Haabeth OA, Lorvik KB, Hammarstrom C, Donaldson IM, Haraldsen G, Bogen B, et al. Inflammation driven by tumour-specific Th1 cells protects against B-cell cancer. Nat Commun. 2011;2:240.
- 71. Palucka K, Banchereau J. Cancer immunotherapy via dendritic cells. Nat Rev Cancer. 2012;12(4):265–77.
- 72. Murray HW, Spitalny GL, Nathan CF. Activation of mouse peritoneal macrophages in vitro and in vivo by interferon-gamma. J Immunol. 1985;134(3):1619–22.
- 73. Hung K, Hayashi R, Lafond-Walker A, Lowenstein C, Pardoll D, Levitsky H. The central role of $CD4(+)$ T cells in the antitumor immune response. J Exp Med. 1998;188(12):2357–68.
- 74. Waldhauer I, Steinle A. NK cells and cancer immunosurveillance. Oncogene. 2008;27(45):5932–43.
- 75. Noy R, Pollard Jeffrey W. Tumor-associated macrophages: from mechanisms to therapy. Immunity. 2014;41(1):49–61.
- 76. Fridman WH, Pagès F, Sautès-Fridman C, Galon J. The immune contexture in human tumours: impact on clinical outcome. Nat Rev Cancer. 2012;12(4):298–306.
- 77. Shurin MR, Lu L, Kalinski P, Stewart-Akers AM, Lotze MT. Th1/Th2 balance in cancer, transplantation and pregnancy. Springer Semin Immunopathol. 1999;21(3):339–59.
- 78. Hunder NN, Wallen H, Cao J, Hendricks DW, Reilly JZ, Rodmyre R, et al. Treatment of metastatic melanoma with autologous CD4+ T cells against NY-ESO-1. N Engl J Med. 2008;358(25):2698–703.
- 79. Tran E, Turcotte S, Gros A, Robbins PF, YC L, Dudley ME, et al. Cancer immunotherapy based on mutation-specific CD4+ T cells in a patient with epithelial cancer. Science. 2014;344(6184):641–5.
- 80. Chandran SS, Paria BC, Srivastava AK, Rothermel LD, Stephens DJ, Dudley ME, et al. Persistence of CTL clones targeting melanocyte differentiation antigens was insufficient to mediate significant melanoma regression in humans. Clin Cancer Res. 2015;21(3):534–43.
- 81. Mosmann TR, Coffman RL. TH1 and TH2 cells: different patterns of lymphokine secretion lead to different functional properties. Annu Rev Immunol. 1989;7(1):145–73.
- 82. Zheng W, Flavell RA. The transcription factor GATA-3 is necessary and sufficient for Th2 cytokine gene expression in CD4 T cells. Cell. 1997;89(4):587–96.
- 83. Tepper RI, Pattengale PK, Leder P. Murine interleukin-4 displays potent anti-tumor activity in vivo. Cell. 1989;57(3):503–12.

8 Cancer Immunity and Immune Evasion Mechanisms

- 84. Volpert OV, Fong T, Koch AE, Peterson JD, Waltenbaugh C, Tepper RI, et al. Inhibition of angiogenesis by interleukin 4. J Exp Med. 1998;188(6):1039–46.
- 85. Shen Y, Fujimoto S. A tumor-specific Th2 clone initiating tumor rejection via primed CD8+ cytotoxic T-lymphocyte activation in mice. Cancer Res. 1996;56(21):5005–11.
- 86. Modesti A, D'Orazi G, Masuelli L, Modica A, Scarpa S, Bosco MC, et al. Ultrastructural evidence of the mechanisms responsible for interleukin-4-activated rejection of a spontaneous murine adenocarcinoma. Int J Cancer. 1993;53(6):988–93.
- 87. Musiani P, Allione A, Modica A, Lollini PL, Giovarelli M, Cavallo F, et al. Role of neutrophils and lymphocytes in inhibition of a mouse mammary adenocarcinoma engineered to release IL-2, IL-4, IL-7, IL-10, IFN-alpha, IFN-gamma, and TNF-alpha. Lab Investig. 1996;74(1):146–57.
- 88. Pericle F, Giovarelli M, Colombo MP, Ferrari G, Musiani P, Modesti A, et al. An efficient Th2-type memory follows CD8+ lymphocyte-driven and eosinophil-mediated rejection of a spontaneous mouse mammary adenocarcinoma engineered to release IL-4. J Immunol. 1994;153(12):5659–73.
- 89. Lebel-Binay S, Laguerre B, Quintin-Colonna F, Conjeaud H, Magazin M, Miloux B, et al. Experimental gene therapy of cancer using tumor cells engineered to secrete interleukin-13. Eur J Immunol. 1995;25(8):2340–8.
- 90. Mattes J, Hulett M, Xie W, Hogan S, Rothenberg ME, Foster P, et al. Immunotherapy of cytotoxic T cell-resistant tumors by T helper 2 cells: an eotaxin and STAT6-dependent process. J Exp Med. 2003;197(3):387–93.
- 91. Conticello C, Pedini F, Zeuner A, Patti M, Zerilli M, Stassi G, et al. IL-4 protects tumor cells from anti-CD95 and chemotherapeutic agents via up-regulation of antiapoptotic proteins. J Immunol. 2004;172(9):5467–77.
- 92. Terabe M, Park JM, Berzofsky JA. Role of IL-13 in regulation of anti-tumor immunity and tumor growth. Cancer Immunol Immunother. 2004;53(2):79–85.
- 93. Zhang WJ, Li BH, Yang XZ, Li PD, Yuan Q, Liu XH, et al. IL-4-induced Stat6 activities affect apoptosis and gene expression in breast cancer cells. Cytokine. 2008;42(1):39–47.
- 94. Aspord C, Pedroza-Gonzalez A, Gallegos M, Tindle S, Burton EC, Su D, et al. Breast cancer instructs dendritic cells to prime interleukin 13–secreting CD4+ T cells that facilitate tumor development. J Exp Med. 2007;204(5):1037–47.
- 95. Wynn TA. Fibrotic disease and the T(H)1/T(H)2 paradigm. Nat Rev Immunol. 2004;4(8):583–94.
- 96. Wynn TA. IL-13 effector functions. Annu Rev Immunol. 2003;21(1):425–56.
- 97. Schuler T, Qin Z, Ibe S, Noben-Trauth N, Blankenstein T. T helper cell type 1-associated and cytotoxic T lymphocyte-mediated tumor immunity is impaired in interleukin 4-deficient mice. J Exp Med. 1999;189(5):803–10.
- 98. Nishimura T, Iwakabe K, Sekimoto M, Ohmi Y, Yahata T, Nakui M, et al. Distinct role of antigen-specific T helper type 1 (Th1) and Th2 cells in tumor eradication in vivo. J Exp Med. 1999;190(5):617–27.
- 99. Wormann SM, Diakopoulos KN, Lesina M, Algul H. The immune network in pancreatic cancer development and progression. Oncogene. 2014;33(3):2956–67.
- 100. Kristensen VN, Vaske CJ, Ursini-Siegel J, Van Loo P, Nordgard SH, Sachidanandam R, et al. Integrated molecular profiles of invasive breast tumors and ductal carcinoma in situ (DCIS) reveal differential vascular and interleukin signaling. Proc Natl Acad Sci U S A. 2012;109(8):2802–7.
- 101. Tassi E, Gavazzi F, Albarello L, Senyukov V, Longhi R, Dellabona P, et al. Carcinoembryonic antigen-specific but not antiviral CD4+ T cell immunity is impaired in pancreatic carcinoma patients. J Immunol. 2008;181(9):6595–603.
- 102. Tatsumi T, Kierstead LS, Ranieri E, Gesualdo L, Schena FP, Finke JH, et al. Diseaseassociated bias in T helper type 1 (Th1)/Th2 CD4(+) T cell responses against MAGE-6 in HLA-DRB10401(+) patients with renal cell carcinoma or melanoma. J Exp Med. 2002;196(5):619–28.
- 103. Fiorentino DF, Zlotnik A, Vieira P, Mosmann TR, Howard M, Moore KW, et al. IL-10 acts on the antigen-presenting cell to inhibit cytokine production by Th1 cells. J Immunol. 1991;146(10):3444–51.
- 104. Steinbrink K, Wolfl M, Jonuleit H, Knop J, Enk AH. Induction of tolerance by IL-10-treated dendritic cells. J Immunol. 1997;159(10):4772–80.
- 105. Huang M, Wang J, Lee P, Sharma S, Mao JT, Meissner H, et al. Human non-small cell lung cancer cells express a type 2 cytokine pattern. Cancer Res. 1995;55(17):3847–53.
- 106. Maeurer MJ, Martin DM, Castelli C, Elder E, Leder G, Storkus WJ, et al. Host immune response in renal cell cancer: interleukin-4 (IL-4) and IL-10 mRNA are frequently detected in freshly collected tumor-infiltrating lymphocytes. Cancer Immunol Immunother. 1995;41(2):111–21.
- 107. Ochi A, Nguyen AH, Bedrosian AS, Mushlin HM, Zarbakhsh S, Barilla R, et al. MyD88 inhibition amplifies dendritic cell capacity to promote pancreatic carcinogenesis via Th2 cells. J Exp Med. 2012;209(9):1671–87.
- 108. De Monte L, Reni M, Tassi E, Clavenna D, Papa I, Recalde H, et al. Intratumor T helper type 2 cell infiltrate correlates with cancer-associated fibroblast thymic stromal lymphopoietin production and reduced survival in pancreatic cancer. J Exp Med. 2011;208(3):469–78.
- 109. Pedroza-Gonzalez A, Xu K, T-C W, Aspord C, Tindle S, Marches F, et al. Thymic stromal lymphopoietin fosters human breast tumor growth by promoting type 2 inflammation. J Exp Med. 2011;208(3):479–90.
- 110. Kido M, Tanaka J, Aoki N, Iwamoto S, Nishiura H, Chiba T, et al. Helicobacter pylori promotes the production of thymic stromal lymphopoietin by gastric epithelial cells and induces dendritic cell-mediated inflammatory Th2 responses. Infect Immun. 2010;78(1):108–14.
- 111. Ziegler A, Heidenreich R, Braumuller H, Wolburg H, Weidemann S, Mocikat R, et al. EpCAM, a human tumor-associated antigen promotes Th2 development and tumor immune evasion. Blood. 2009;113(15):3494–502.
- 112. Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, et al. Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. Nat Immunol. 2005;6(11):1123–32.
- 113. Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang YH, et al. A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. Nat Immunol. 2005;6(11):1133–41.
- 114. Acosta-Rodriguez EV, Rivino L, Geginat J, Jarrossay D, Gattorno M, Lanzavecchia A, et al. Surface phenotype and antigenic specificity of human interleukin 17-producing T helper memory cells. Nat Immunol. 2007;8(6):639–46.
- 115. Annunziato F, Cosmi L, Santarlasci V, Maggi L, Liotta F, Mazzinghi B, et al. Phenotypic and functional features of human Th17 cells. J Exp Med. 2007;204(8):1849–61.
- 116. Korn T, Bettelli E, Oukka M, Kuchroo VK. IL-17 and Th17 Cells. Annu Rev Immunol. 2009;27:485–517.
- 117. Annunziato F, Cosmi L, Liotta F, Maggi E, Romagnani S. Main features of human T helper 17 cells. Ann N Y Acad Sci. 2013;1284:66–70.
- 118. Annunziato F, Cosmi L, Liotta F, Maggi E, Romagnani S. The phenotype of human Th17 cells and their precursors, the cytokines that mediate their differentiation and the role of Th17 cells in inflammation. Int Immunol. 2008;20(11):1361–8.
- 119. Kleinewietfeld M, Hafler DA. The plasticity of human Treg and Th17 cells and its role in autoimmunity. Semin Immunol. 2013;25(4):305–12.
- 120. Sundrud MS, Trivigno C. Identity crisis of Th17 cells: many forms, many functions, many questions. Semin Immunol. 2013;25(4):263–72.
- 121. Jin W, Dong C. IL-17 cytokines in immunity and inflammation. Emerg Microbes Infect. 2013;2(9):e60.
- 122. Muranski P, Boni A, Antony PA, Cassard L, Irvine KR, Kaiser A, et al. Tumor-specific Th17 polarized cells eradicate large established melanoma. Blood. 2008;112(2):362–73.
- 123. Martin-Orozco N, Muranski P, Chung Y, Yang XO, Yamazaki T, Lu S, et al. T helper 17 cells promote cytotoxic T cell activation in tumor immunity. Immunity. 2009;31(5):787–98.
- 124. Cho BS, Lim JY, Yahng SA, Lee SE, Eom KS, Kim YJ, et al. Circulating IL-17 levels during the peri-transplant period as a predictor for early leukemia relapse after myeloablative allogeneic stem cell transplantation. Ann Hematol. 2012;91(3):439–48.
- 125. Tartour E, Fossiez F, Joyeux I, Galinha A, Gey A, Claret E, et al. Interleukin 17, a T-cellderived cytokine, promotes tumorigenicity of human cervical tumors in nude mice. Cancer Res. 1999;59(15):3698–704.
- 126. Numasaki M, Watanabe M, Suzuki T, Takahashi H, Nakamura A, McAllister F, et al. IL-17 enhances the net angiogenic activity and in vivo growth of human non-small cell lung cancer in SCID mice through promoting CXCR-2-dependent angiogenesis. J Immunol. 2005;175(9):6177–89.
- 127. Chang SH, Mirabolfathinejad SG, Katta H, Cumpian AM, Gong L, Caetano MS, et al. T helper 17 cells play a critical pathogenic role in lung cancer. Proc Natl Acad Sci U S A. 2014;111(15):5664–9.
- 128. De Simone V, Pallone F, Monteleone G, Stolfi C. Role of T(H)17 cytokines in the control of colorectal cancer. Oncoimmunology. 2013;2(12):e26617.
- 129. Wei S, Zhao E, Kryczek I, Zou W. Th17 cells have stem cell-like features and promote longterm immunity. OncoImmunology. 2012;1(4):516–9.
- 130. Wang K, Kim MK, Di Caro G, Wong J, Shalapour S, Wan J, et al. Interleukin-17 receptor a signaling in transformed enterocytes promotes early colorectal tumorigenesis. Immunity. 2014;41(6):1052–63.
- 131. McAllister F, Bailey JM, Alsina J, Nirschl CJ, Sharma R, Fan H, et al. Oncogenic KRAS activates a hematopoietic-to-epithelial IL-17 signaling axis in preinvasive pancreatic neoplasia. Cancer Cell. 2014;25(5):621–37.
- 132. Keerthivasan S, Aghajani K, Dose M, Molinero L, Khan MW, Venkateswaran V, et al. beta-Catenin promotes colitis and colon cancer through imprinting of proinflammatory properties in T cells. Sci Transl Med. 2014;6(225):225ra28.
- 133. Kryczek I, Banerjee M, Cheng P, Vatan L, Szeliga W, Wei S, et al. Phenotype, distribution, generation, and functional and clinical relevance of Th17 cells in the human tumor environments. Blood. 2009;114(6):1141–9.
- 134. Lv L, Pan K, Li XD, She KL, Zhao JJ, Wang W, et al. The accumulation and prognosis value of tumor infiltrating IL-17 producing cells in esophageal squamous cell carcinoma. PLoS One. 2011;6(3):e18219.
- 135. Su X, Ye J, Hsueh EC, Zhang Y, Hoft DF, Peng G. Tumor microenvironments direct the recruitment and expansion of human Th17 cells. J Immunol. 2010;184(3):1630–41.
- 136. De Simone V, Franze E, Ronchetti G, Colantoni A, Fantini MC, Di Fusco D, et al. Th17 type cytokines, IL-6 and TNF-alpha synergistically activate STAT3 and NF-kB to promote colorectal cancer cell growth. Oncogene. 2015;34(27):3493–503.
- 137. Ye J, Su X, Hsueh EC, Zhang Y, Koenig JM, Hoft DF, et al. Human tumor-infiltrating Th17 cells have the capacity to differentiate into IFN-gamma+ and FOXP3+ T cells with potent suppressive function. Eur J Immunol. 2011;41(4):936–51.
- 138. Kryczek I, Zhao E, Liu Y, Wang Y, Vatan L, Szeliga W, et al. Human TH17 cells are long-lived effector memory cells. Sci Transl Med. 2011;3(104):104ra0-ra0.
- 139. Liu H, Rohowsky-Kochan C. Regulation of IL-17 in human CCR6+ effector memory T cells. J Immunol. 2008;180(12):7948–57.
- 140. Chellappa S, Hugenschmidt H, Hagness M, Line PD, Labori KJ, Wiedswang G, et al. Regulatory T cells that co-express RORγt and FOXP3 are pro-inflammatory and immunosuppressive and expand in human pancreatic cancer. OncoImmunology. 2015;5(4):e1102828.
- 141. Shibuya TY, Nugyen N, McLaren CE, Li KT, Wei WZ, Kim S, et al. Clinical significance of poor CD3 response in head and neck cancer. Clin Cancer Res. 2002;8(3):745–51.
- 142. Badoual C, Hans S, Rodriguez J, Peyrard S, Klein C, Agueznay Nel H, et al. Prognostic value of tumor-infiltrating CD4+ T-cell subpopulations in head and neck cancers. Clin Cancer Res. 2006;12(2):465–72.
- 143. Cho Y, Miyamoto M, Kato K, Fukunaga A, Shichinohe T, Kawarada Y, et al. CD4+ and CD8+ T cells cooperate to improve prognosis of patients with esophageal squamous cell carcinoma. Cancer Res. 2003;63(7):1555–9.
- 144. Schumacher K, Haensch W, Roefzaad C, Schlag PM. Prognostic significance of activated $CD8(+)$ T cell infiltrations within esophageal carcinomas. Cancer Res. $2001:61(10):3932-6$.
- 145. van Sandick JW, Boermeester MA, Gisbertz SS, ten Berge IJ, Out TA, van der Pouw Kraan TC, et al. Lymphocyte subsets and T(h)1/T(h)2 immune responses in patients with adenocarcinoma of the oesophagus or oesophagogastric junction: relation to pTNM stage and clinical outcome. Cancer Immunol Immunother. 2003;52(10):617–24.
- 146. Dieu-Nosjean MC, Antoine M, Danel C, Heudes D, Wislez M, Poulot V, et al. Long-term survival for patients with non-small-cell lung cancer with intratumoral lymphoid structures. J Clin Oncol. 2008;26(27):4410–7.
- 147. Chen X, Wan J, Liu J, Xie W, Diao X, Xu J, et al. Increased IL-17-producing cells correlate with poor survival and lymphangiogenesis in NSCLC patients. Lung Cancer. 2010;69(3):348–54.
- 148. Tao H, Mimura Y, Aoe K, Kobayashi S, Yamamoto H, Matsuda E, et al. Prognostic potential of FOXP3 expression in non-small cell lung cancer cells combined with tumor-infiltrating regulatory T cells. Lung Cancer. 2012;75(1):95–101.
- 149. Fukunaga A, Miyamoto M, Cho Y, Murakami S, Kawarada Y, Oshikiri T, et al. CD8+ tumor-infiltrating lymphocytes together with CD4+ tumor-infiltrating lymphocytes and dendritic cells improve the prognosis of patients with pancreatic adenocarcinoma. Pancreas. 2004;28(1):e26–31.
- 150. Bazhin AV, Shevchenko I, Umansky V, Werner J, Karakhanova S. Two immune faces of pancreatic adenocarcinoma: possible implication for immunotherapy. Cancer Immunol Immunother. 2014;63(1):59–65.
- 151. Vizio B, Novarino A, Giacobino A, Cristiano C, Prati A, Ciuffreda L, et al. Potential plasticity of T regulatory cells in pancreatic carcinoma in relation to disease progression and outcome. Exp Ther Med. 2012;4(1):70–8.
- 152. Hiraoka N, Onozato K, Kosuge T, Hirohashi S. Prevalence of FOXP3+ regulatory T cells increases during the progression of pancreatic ductal adenocarcinoma and its premalignant lesions. Clin Cancer Res. 2006;12(18):5423–34.
- 153a. Goeppert B, Frauenschuh L, Zucknick M, Stenzinger A, Andrulis M, Klauschen F, et al. Prognostic impact of tumour-infiltrating immune cells on biliary tract cancer. Br J Cancer. 2013;109(10):2665–74.
- 153b. Chellappa S, Hugenschmidt H, Hagness M, Subramani S, Melum E, Line PD, et al. CD8+ T Cells That Coexpress RORγt and T-bet Are Functionally Impaired and Expand in Patients with Distal Bile Duct Cancer. J Immunol. 2017;198(4):1729–39.
- 154. Mahmoud SM, Paish EC, Powe DG, Macmillan RD, Grainge MJ, Lee AH, et al. Tumorinfiltrating CD8+ lymphocytes predict clinical outcome in breast cancer. J Clin Oncol. 2011;29(15):1949–55.
- 155. Teschendorff AE, Gomez S, Arenas A, El-Ashry D, Schmidt M, Gehrmann M, et al. Improved prognostic classification of breast cancer defined by antagonistic activation patterns of immune response pathway modules. BMC Cancer. 2010;10:604.
- 156. Yoon NK, Maresh EL, Shen D, Elshimali Y, Apple S, Horvath S, et al. Higher levels of GATA3 predict better survival in women with breast cancer. Hum Pathol. 2010;41(12):1794–801.
- 157. Chen WC, Lai YH, Chen HY, Guo HR, IJ S, Chen HH. Interleukin-17-producing cell infiltration in the breast cancer tumour microenvironment is a poor prognostic factor. Histopathology. 2013;63(2):225–33.
- 158. Bates GJ, Fox SB, Han C, Leek RD, Garcia JF, Harris AL, et al. Quantification of regulatory T cells enables the identification of high-risk breast cancer patients and those at risk of late relapse. J Clin Oncol. 2006;24(34):5373–80.
- 159. Gobert M, Treilleux I, Bendriss-Vermare N, Bachelot T, Goddard-Leon S, Arfi V, et al. Regulatory T cells recruited through CCL22/CCR4 are selectively activated in lymphoid infiltrates surrounding primary breast tumors and lead to an adverse clinical outcome. Cancer Res. 2009;69(5):2000–9.
- 160. West NR, Kost SE, Martin SD, Milne K, Deleeuw RJ, Nelson BH, et al. Tumour-infiltrating FOXP3(+) lymphocytes are associated with cytotoxic immune responses and good clinical outcome in oestrogen receptor-negative breast cancer. Br J Cancer. 2013;108(1):155–62.

8 Cancer Immunity and Immune Evasion Mechanisms

- 161. Zhuang Y, Peng LS, Zhao YL, Shi Y, Mao XH, Chen W, et al. CD8(+) T cells that produce interleukin-17 regulate myeloid-derived suppressor cells and are associated with survival time of patients with gastric cancer. Gastroenterology. 2012;143(4):951–62.e8.
- 162. Saito H, Yamada Y, Takaya S, Osaki T, Ikeguchi M. Clinical relevance of the number of interleukin-17-producing CD 8+ T cells in patients with gastric cancer. Surg Today. 2015;45(11):1429–35.
- 163. Ubukata H, Motohashi G, Tabuchi T, Nagata H, Konishi S, Tabuchi T. Evaluations of interferon-gamma/interleukin-4 ratio and neutrophil/lymphocyte ratio as prognostic indicators in gastric cancer patients. J Surg Oncol. 2010;102(7):742–7.
- 164. Chen JG, Xia JC, Liang XT, Pan K, Wang W, Lv L, et al. Intratumoral expression of IL-17 and its prognostic role in gastric adenocarcinoma patients. Int J Biol Sci. 2011;7(1):53–60.
- 165. Maruyama T, Kono K, Mizukami Y, Kawaguchi Y, Mimura K, Watanabe M, et al. Distribution of Th17 cells and FoxP3(+) regulatory T cells in tumor-infiltrating lymphocytes, tumor-draining lymph nodes and peripheral blood lymphocytes in patients with gastric cancer. Cancer Sci. 2010;101(9):1947–54.
- 166. Shen Z, Zhou S, Wang Y, Li RL, Zhong C, Liang C, et al. Higher intratumoral infiltrated Foxp3+ Treg numbers and Foxp3+/CD8+ ratio are associated with adverse prognosis in resectable gastric cancer. J Cancer Res Clin Oncol. 2010;136(10):1585–95.
- 167. Cai XY, Gao Q, Qiu SJ, Ye SL, ZQ W, Fan J, et al. Dendritic cell infiltration and prognosis of human hepatocellular carcinoma. J Cancer Res Clin Oncol. 2006;132(5):293–301.
- 168. Gao Q, Qiu SJ, Fan J, Zhou J, Wang XY, Xiao YS, et al. Intratumoral balance of regulatory and cytotoxic T cells is associated with prognosis of hepatocellular carcinoma after resection. J Clin Oncol. 2007;25(18):2586–93.
- 169. Gao Q, Wang XY, Qiu SJ, Zhou J, Shi YH, Zhang BH, et al. Tumor stroma reaction-related gene signature predicts clinical outcome in human hepatocellular carcinoma. Cancer Sci. 2011;102(8):1522–31.
- 170. Zhang JP, Yan J, Xu J, Pang XH, Chen MS, Li L, et al. Increased intratumoral IL-17 producing cells correlate with poor survival in hepatocellular carcinoma patients. J Hepatol. 2009;50(5):980–9.
- 171. Kobayashi N, Hiraoka N, Yamagami W, Ojima H, Kanai Y, Kosuge T, et al. FOXP3+ regulatory T cells affect the development and progression of hepatocarcinogenesis. Clin Cancer Res. 2007;13(3):902–11.
- 172. Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pages C, et al. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. Science. 2006;313(5795):1960–4.
- 173. Tosolini M, Kirilovsky A, Mlecnik B, Fredriksen T, Mauger S, Bindea G, et al. Clinical impact of different classes of infiltrating T cytotoxic and helper cells (Th1, th2, treg, th17) in patients with colorectal cancer. Cancer Res. 2011;71(4):1263–71.
- 174. Camus M, Tosolini M, Mlecnik B, Pages F, Kirilovsky A, Berger A, et al. Coordination of intratumoral immune reaction and human colorectal cancer recurrence. Cancer Res. 2009;69(6):2685–93.
- 175. Mlecnik B, Tosolini M, Kirilovsky A, Berger A, Bindea G, Meatchi T, et al. Histopathologicbased prognostic factors of colorectal cancers are associated with the state of the local immune reaction. J Clin Oncol. 2011;29(6):610–8.
- 176. Sinicrope FA, Rego RL, Ansell SM, Knutson KL, Foster NR, Sargent DJ. Intraepithelial effector (CD3+)/regulatory (FoxP3+) T-cell ratio predicts a clinical outcome of human colon carcinoma. Gastroenterology. 2009;137(4):1270–9.
- 177. Naito Y, Saito K, Shiiba K, Ohuchi A, Saigenji K, Nagura H, et al. CD8+ T cells infiltrated within cancer cell nests as a prognostic factor in human colorectal cancer. Cancer Res. 1998;58(16):3491–4.
- 178. Nosho K, Baba Y, Tanaka N, Shima K, Hayashi M, Meyerhardt JA, et al. Tumour-infiltrating T-cell subsets, molecular changes in colorectal cancer, and prognosis: cohort study and literature review. J Pathol. 2010;222(4):350–66.
- 179. Liu J, Duan Y, Cheng X, Chen X, Xie W, Long H, et al. IL-17 is associated with poor prognosis and promotes angiogenesis via stimulating VEGF production of cancer cells in colorectal carcinoma. Biochem Biophys Res Commun. 2011;407(2):348–54.
- 180. Yoshida N, Kinugasa T, Miyoshi H, Sato K, Yuge K, Ohchi T, et al. A high RORgammaT/ CD3 ratio is a strong prognostic factor for postoperative survival in advanced colorectal cancer: analysis of helper T cell lymphocytes (Th1, Th2, Th17 and regulatory T cells). Ann Surg Oncol. 2015;23(3):919–27.
- 181. Frey DM, Droeser RA, Viehl CT, Zlobec I, Lugli A, Zingg U, et al. High frequency of tumorinfiltrating FOXP3(+) regulatory T cells predicts improved survival in mismatch repairproficient colorectal cancer patients. Int J Cancer. 2010;126(11):2635–43.
- 182. Salama P, Phillips M, Grieu F, Morris M, Zeps N, Joseph D, et al. Tumor-infiltrating FOXP3+ T regulatory cells show strong prognostic significance in colorectal cancer. J Clin Oncol. 2009;27(2):186–92.
- 183. Blatner NR, Mulcahy MF, Dennis KL, Scholtens D, Bentrem DJ, Phillips JD, et al. Expression of RORγt marks a pathogenic regulatory T cell subset in human colon cancer. Sci Transl Med. 2012;4(164):164ra59.
- 184. Sato E, Olson SH, Ahn J, Bundy B, Nishikawa H, Qian F, et al. Intraepithelial CD8+ tumorinfiltrating lymphocytes and a high CD8+/regulatory T cell ratio are associated with favorable prognosis in ovarian cancer. Proc Natl Acad Sci U S A. 2005;102(51):18538–43.
- 185. Marth C, Fiegl H, Zeimet AG, Muller-Holzner E, Deibl M, Doppler W, et al. Interferongamma expression is an independent prognostic factor in ovarian cancer. Am J Obstet Gynecol. 2004;191(5):1598–605.
- 186. Kusuda T, Shigemasa K, Arihiro K, Fujii T, Nagai N, Ohama K. Relative expression levels of Th1 and Th2 cytokine mRNA are independent prognostic factors in patients with ovarian cancer. Oncol Rep. 2005;13(6):1153–8.
- 187. Milne K, Kobel M, Kalloger SE, Barnes RO, Gao D, Gilks CB, et al. Systematic analysis of immune infiltrates in high-grade serous ovarian cancer reveals CD20, FoxP3 and TIA-1 as positive prognostic factors. PLoS One. 2009;4(7):e6412.
- 188. Leffers N, Gooden MJ, de Jong RA, Hoogeboom BN, ten Hoor KA, Hollema H, et al. Prognostic significance of tumor-infiltrating T-lymphocytes in primary and metastatic lesions of advanced stage ovarian cancer. Cancer Immunol Immunother. 2009;58(3):449–59.
- 189. Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. Nat Med. 2004;10(9):942–9.
- 190. Nakano O, Sato M, Naito Y, Suzuki K, Orikasa S, Aizawa M, et al. Proliferative activity of intratumoral CD8(+) T-lymphocytes as a prognostic factor in human renal cell carcinoma: clinicopathologic demonstration of antitumor immunity. Cancer Res. 2001;61(13):5132–6.
- 191. Kondo T, Nakazawa H, Ito F, Hashimoto Y, Osaka Y, Futatsuyama K, et al. Favorable prognosis of renal cell carcinoma with increased expression of chemokines associated with a Th1-type immune response. Cancer Sci. 2006;97(8):780–6.
- 192. Karja V, Aaltomaa S, Lipponen P, Isotalo T, Talja M, Mokka R. Tumour-infiltrating lymphocytes: a prognostic factor of PSA-free survival in patients with local prostate carcinoma treated by radical prostatectomy. Anticancer Res. 2005;25(6C):4435–8.
- 193. Sharma P, Shen Y, Wen S, Yamada S, Jungbluth AA, Gnjatic S, et al. CD8 tumor-infiltrating lymphocytes are predictive of survival in muscle-invasive urothelial carcinoma. Proc Natl Acad Sci U S A. 2007;104(10):3967–72.
- 194. de Jong RA, Leffers N, Boezen HM, ten Hoor KA, van der Zee AG, Hollema H, et al. Presence of tumor-infiltrating lymphocytes is an independent prognostic factor in type I and II endometrial cancer. Gynecol Oncol. 2009;114(1):105–10.
- 195. Piersma SJ, Jordanova ES, van Poelgeest MI, Kwappenberg KM, van der Hulst JM, Drijfhout JW, et al. High number of intraepithelial CD8+ tumor-infiltrating lymphocytes is associated with the absence of lymph node metastases in patients with large early-stage cervical cancer. Cancer Res. 2007;67(1):354–61.
- 196. Zhang Y, Hou F, Liu X, Ma D, Zhang Y, Kong B, et al. Tc17 cells in patients with uterine cervical cancer. PLoS One. 2014;9(2):e86812.
- 197. Taylor RC, Patel A, Panageas KS, Busam KJ, Brady MS. Tumor-infiltrating lymphocytes predict sentinel lymph node positivity in patients with cutaneous melanoma. J Clin Oncol. 2007;25(7):869–75.
- 198. Clemente CG, Mihm MC Jr, Bufalino R, Zurrida S, Collini P, Cascinelli N. Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. Cancer. 1996;77(7):1303–10.
- 199. Ladanyi A, Mohos A, Somlai B, Liszkay G, Gilde K, Fejos Z, et al. FOXP3+ cell density in primary tumor has no prognostic impact in patients with cutaneous malignant melanoma. Pathol Oncol Res. 2010;16(3):303–9.
- 200. Mougiakakos D, Johansson CC, Trocme E, All-Ericsson C, Economou MA, Larsson O, et al. Intratumoral forkhead box P3-positive regulatory T cells predict poor survival in cyclooxygenase-2-positive uveal melanoma. Cancer. 2010;116(9):2224–33.
- 201. Miracco C, Mourmouras V, Biagioli M, Rubegni P, Mannucci S, Monciatti I, et al. Utility of tumour-infiltrating CD25+FOXP3+ regulatory T cell evaluation in predicting local recurrence in vertical growth phase cutaneous melanoma. Oncol Rep. 2007;18(5):1115–22.
- 202. Schreck S, Friebel D, Buettner M, Distel L, Grabenbauer G, Young LS, et al. Prognostic impact of tumour-infiltrating Th2 and regulatory T cells in classical Hodgkin lymphoma. Hematol Oncol. 2009;27(1):31–9.
- 203. Tzankov A, Meier C, Hirschmann P, Went P, Pileri SA, Dirnhofer S. Correlation of high numbers of intratumoral FOXP3+ regulatory T cells with improved survival in germinal centerlike diffuse large B-cell lymphoma, follicular lymphoma and classical Hodgkin's lymphoma. Haematologica. 2008;93(2):193–200.
- 204. Carreras J, Lopez-Guillermo A, Fox BC, Colomo L, Martinez A, Roncador G, et al. High numbers of tumor-infiltrating FOXP3-positive regulatory T cells are associated with improved overall survival in follicular lymphoma. Blood. 2006;108(9):2957–64.
- 205. Huse M, Lillemeier BF, Kuhns MS, Chen DS, Davis MM. T cells use two directionally distinct pathways for cytokine secretion. Nat Immunol. 2006;7(3):247–55.
- 206. Faghih Z, Rezaeifard S, Safaei A, Ghaderi A, Erfani N. IL-17 and IL-4 producing CD8+ T cells in tumor draining lymph nodes of breast cancer patients: positive association with tumor progression. Iran J Immunol. 2013;10(4):193–204.
- 207. Tsai JP, Lee MH, Hsu SC, Chen MY, Liu SJ, Chang JT, et al. CD4+ T cells disarm or delete cytotoxic T lymphocytes under IL-17-polarizing conditions. J Immunol. 2012;189(4):1671–9.
- 208. Huber M, Heink S, Grothe H, Guralnik A, Reinhard K, Elflein K, et al. A Th17-like developmental process leads to CD8(+) Tc17 cells with reduced cytotoxic activity. Eur J Immunol. 2009;39(7):1716–25.
- 209. Tajima M, Wakita D, Satoh T, Kitamura H, Nishimura T. IL-17/IFN-gamma double producing CD8+ T (Tc17/IFN-gamma) cells: a novel cytotoxic T-cell subset converted from Tc17 cells by IL-12. Int Immunol. 2011;23(12):751–9.
- 210. Kuang DM, Peng C, Zhao Q, Wu Y, Zhu LY, Wang J, et al. Tumor-activated monocytes promote expansion of IL-17-producing CD8+ T cells in hepatocellular carcinoma patients. J Immunol. 2010;185(3):1544–9.
- 211. Zhang W, Hou F, Zhang Y, Tian Y, Jiao J, Ma D, et al. Changes of Th17/Tc17 and Th17/Treg cells in endometrial carcinoma. Gynecol Oncol. 2014;132(3):599–605.
- 212. Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf Anna C, et al. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. Immunity. 2013;39(4):782–95.
- 213. YF X, Lu Y, Cheng H, Shi S, Xu J, Long J, et al. Abnormal distribution of peripheral lymphocyte subsets induced by PDAC modulates overall survival. Pancreatology. 2014;14(4):295–301.
- 214. Jochems C, Schlom J. Tumor-infiltrating immune cells and prognosis: the potential link between conventional cancer therapy and immunity. Exp Biol Med. 2011;236(5):567–79.
- 215. Sakaguchi S, Yamaguchi T, Nomura T, Ono M. Regulatory T cells and immune tolerance. Cell. 2008;133(5):775–87.
- 216. Sakaguchi S, Miyara M, Costantino CM, Hafler DA. FOXP3+ regulatory T cells in the human immune system. Nat Rev Immunol. 2010;10(7):490–500.
- 217. Shevach EM. Mechanisms of Foxp3+ T regulatory cell-mediated suppression. Immunity. 2009;30(5):636–45.
- 218. Banerjee A, Vasanthakumar A, Grigoriadis G. Modulating T regulatory cells in cancer: how close are we? Immunol Cell Biol. 2013;91(5):340–9.
- 219. Waight JD, Takai S, Marelli B, Qin G, Hance KW, Zhang D, et al. Cutting edge: epigenetic regulation of Foxp3 defines a stable population of CD4+ regulatory T cells in tumors from mice and humans. J Immunol. 2015;194(3):878–82.
- 220. Kryczek I, Wu K, Zhao E, Wei S, Vatan L, Szeliga W, et al. IL-17+ regulatory T cells in the microenvironments of chronic inflammation and cancer. J Immunol. 2011;186(7):4388–95.
- 221. Yang S, Wang B, Guan C, Wu B, Cai C, Wang M, et al. Foxp3+IL-17+ T cells promote development of cancer-initiating cells in colorectal cancer. J Leukoc Biol. 2011;89(1):85–91.
- 222. Thibaudin M, Chaix M, Boidot R, Végran F, Derangère V, Limagne E, et al. Human ectonucleotidase-expressing CD25high Th17 cells accumulate in breast cancer tumors and exert immunosuppressive functions. OncoImmunology. 2015;5(1):e1055444.
- 223. O'Callaghan DS, Rexhepaj E, Gately K, Coate L, Delaney D, O'Donnell DM, et al. Tumour islet Foxp3+ T-cell infiltration predicts poor outcome in nonsmall cell lung cancer. Eur Respir J. 2015;46(6):1762–72.