

Yossra A. Suliman, Harsh Agrawal, and Daniel E. Furst

Importance of Therapeutic Trials

While therapeutic trials are essential when seeking guidance in treating diseases, relying solely on open label studies or case reports may be misleading due to selection bias, reporting bias, and the lack of control group. Thus, several SSc treatments were thought to be effective until investigated in a randomized case-control manner [1, 2]. Progress in the development and validation of outcome measures, together with improved insights on SSc pathogenesis, have opened the door to establishing therapies in SSc through well-designed controlled trials.

Epidemiological Considerations

Status of Scleroderma as a Rare Disease

The Orphan Drug and Rare Disease Act of 1983 encourages pharmaceutical companies to develop drugs for “rare diseases” that otherwise have a very low prevalence and for which drug development lacks profit motive. In the US

Yossra A Suliman and Harsh Agrawal have no disclosures.

Daniel E. Furst. Disclosures

Grant/Research Support: AbbVie, Actelion, Amgen, BMS, Gilead, GSK, NIH, Novartis, Pfizer, Roche/Genentech, UCB

Consultant: AbbVie, Actelion, Amgen, BMS, Cytos, Janssen, Gilead, GSK, NIH, Novartis, Pfizer, Roche/Genentech, UCB

Speaker's Bureau (CME ONLY): AbbVie, Actelion, and UCB.

Y.A. Suliman, MD, MSc

Department of Rheumatology and Rehabilitation, Assiut

University Hospital, Assiut, Egypt

e-mail: dr.yossra@gmail.com

H. Agrawal, MD, FACP

Department of Internal Medicine, Division of Cardiology,

Paul. L. Foster School of Medicine Texas Tech University,

El Paso, Texas, USA

D.E. Furst, MD (✉)

Department of Rheumatology, University of California, Los

Angeles, Los Angeles, CA, USA

regulatory environment, “rare disease” is defined as one affecting fewer than 200,000 Americans. Given that SSc falls in the category of “rare disease”, affecting about 1 in 5,000 [3], pharmaceutical companies have tax and patent incentives under the Orphan Drug and Rare Disease Act to develop drugs for this condition. In part due to the support from the abovementioned legislations, there have been 23 randomized clinical trials in SSc in the last 5 years compared to seven such trials between 1980 and 1986 [4, 8, 10].

Trial Design

Phase I–III

The principle focus of phase I trials is the safety of the tested treatment, adverse events (AE), serious adverse events (SAEs), and/or death. Even during this phase, placebo controls are necessary because only placebo controls will enable one to differentiate whether a sign or symptom is due to treatment-related adverse event or an SSc-related complication. Stopping rules during this phase are particularly important (although should be included in all trials of disease with severe consequences such as SSc). This is because it is unacceptable in some circumstances to continue the tested drug for patients who develop organ complications or nonresponders when there is available effective treatment for such organ involvement. On the other hand, it is possible to continue a drug tested in certain aspects of organ involvement when there is no known effective treatment.

Phase II trials are mainly focused on evaluating initial efficacy and establishing an appropriate dose for later trials, although safety must continue to be carefully monitored. This is also an opportunity to explore and validate clinical, laboratory and biomarker end points. End points used for clinical trials should be practical and fully validated; in SSc, the use of surrogate outcome measurements may be more feasible in selected cases. As in phase 1, there should be controls, usually placebo, to establish the true early efficacy and further safety of the drug.

Phase III trials involve more patients to establish efficacy at the chosen dose(s) and establish the safety profile of the therapy for more common adverse events. This phase should be controlled, whether placebo and/or positive controls.

Phase IV: Although drugs are carefully tested in the above three phases before being marketed, postmarketing studies establish the profile of the drug in a more general population, further establish the therapy's safety profile, and attempt early discovery of less common adverse events during long-term use.

Risk evaluation and mitigation strategies (REMS) are risk management strategies initiated by the Food and Drug Administration Amendments Act of 2007 ("FDAAA"), which gave FDA the authority to request a REMS from drug companies to make sure that the benefits of a drug or biological product continue to outweigh its risks. It was specifically tailored to make sure that there is a favorable risk: benefit ratio in larger populations and in general use. The FDA website provides a list of REMS with the currently approved drugs including biologics via REMS [5].

Characteristics of Outcome Measurements in SSc

OMERACT (outcome measure in rheumatologic clinical trials) is an initiative established by a group of rheumatologists, statisticians, and epidemiologists whose main objective is to improve outcome measures in rheumatology. Clinical trials in SSc should seek to evaluate outcomes in a thorough, valid manner; the OMERACT principles of truth, discrimination, and feasibility are one approach and are frequently used [6, 7]. Those include feasibility, face, content, criterion and construct validity, reproducibility/reliability, sensitivity to change, and ability to discriminate therapy from control; it includes patient involvement and a consideration of the context (e.g., comorbidities, other medications used, cultural factors) of the measure. Certain aspects of measurement validation are particularly important, as they are critical to trial design and the ability to discern treatment effects. This applies to discrimination and responsiveness to change.

Discrimination Discriminant validity was shown in some outcome measures in SSc clinical trials. FVC percent predicted could discriminate between cyclophosphamide-treated and placebo control groups as a measure of improvement in SSc-ILD (interstitial lung disease) [8]. Johnson et al. used Bayesian model analysis of uncommon diseases to identify MRSS as an outcome measure of skin tightness in SSc. Better mean outcomes of MRSS in MTX-treated group than placebo (94%) demonstrated the discriminant validity of MRSS in SSc [9]. Most recently, event-free survival was identified as the outcome measure in a study of long-term effects of treatment with *Hematopoietic stem cell*

transplantation (HSCT) vs. cyclophosphamide in SSc. Event-free survival (time from randomization until the occurrence of death or persistent major organ failure) could discriminate the significant survival in HSCT group than in control group after 4-year follow-up [10].

Responsiveness to Change In SSc, several outcome measures may not show any change in RCTs. The reason behind the lack of change in RCTs is that most of SSc disease modification trials have been negative, although some trials showed positive change – for example, cyclophosphamide, which improved FVC and skin score [11]. Outcomes like GIT 2.0, FVC, HAQ-DI, SF-36, 6MWD, MRSS, and RCS are responsive to change and were able to show some improvement in clinical trials [12–16], while others, such as oral aperture opening, handspan, and other biomarkers, did not show any change in response to treatment [17].

Overall Measures of Scleroderma

A group of SSc experts within OMERACT started the combined response index for SSc (CRISS) as an instrument to be used for clinical trials. In an effort to develop single measure composed of a set of domains which reflect organ involvement, CRISS conducted a Delphi exercise with expert review to distinguish 11 core set items to be considered in SSc clinical trials: soluble biomarkers, cardiac, digital ulcers, gastrointestinal, global health, health-related quality of life and function, musculoskeletal, pulmonary, RP, renal, and skin. Ongoing prospective study to test the validity of CRISS against OMERACT criteria is currently being undertaken. Further revision and definition of the final set of domains will be commenced based on obtained results [18].

Another overall outcome measure in SSc is the European scleroderma study group activity index (EscSG) which evaluates both clinical domains and specific laboratory values, including the MRSS, DLCO, and presence of scleredema, digital ulcers, arthritis, ESR, hypocomplementemia, and patient-reported worsening of the skin and vascular and cardiopulmonary symptoms [19–22]. Valentini et al. evaluated the validity of EScSG activity index; face, content, and construct validity was demonstrated [20]. Further assessment of the content and construct validity was conducted by Minier et al. [22] in a larger cohort of SSc patients. Responsiveness to change, however, has not yet been evaluated for EscSG activity index, and further validation steps are still warranted.

Khanna et al. developed a consensus of 22 points to consider for evidence-based clinical trial design in SSc. They entail establishing standards for more uniform clinical trial design and improved selection of outcome measures; they also outlined areas where further research is warranted [23]. Outcome measures used in SSc clinical trials are listed in Table 46.1.

Table 46.1 Outcome measures used in SSc clinical trials

Organ system	Valid	Partially validated	Used but not completely valid	Emerging
Cardiac	Congestive heart failure clinical exam [24] Pericardial disease (EKG, clinical exam, echocardiography) [25, 26]	Tissue Doppler echocardiography [26, 27] Cardiac MRI [28, 29] Right heart catheterization [30] Left heart catheterization [31] Borg dyspnea instrument [32] Scintigraphy [33, 34] Holter [35] EKG [36] Nt-pro-BNP [37]	Cardiac conduction blocks [38] Fixed defects on perfusional scintigraphy [39] Video densitometric alterations [40]	Speckle-tracking echocardiography [41], diffuse fibrosis imaging using magnetic resonance imaging [42] Absolute perfusion magnetic resonance imaging Troponins cardiac computerized tomography [43]
Digital ulcers	Total net ulcer burden [44] HAQ pain VAS Digital Ulcer [45] HAQ disability index SF-36 [45]	Active ulcer on fingertips on the volar surface [46]	Raynaud condition score [45] Cochin hand function scale [47] Michigan hand questionnaire [48]	New ulcers Time to healing of baseline vs. largest vs. cardinal ulcer Capillaroscopy [49] Thermography Arteriography [50] MRI [51] Doppler ultrasound Laser Doppler Transcutaneous tensiometry Granulation tissue color pictures Surface area measurement
Raynaud's	Raynaud's condition score [52, 53] Frequency of RP attacks [45] Duration of RP attacks [45] Patient global assessment [45] Physician global assessment [45] Digital ulcers	VAS or Likert [45] Pain VAS or HAQ [45]		Thermography [54] Laser Doppler imaging [55] Finger systolic pressure measurements [55] Nail fold capillaroscopy [55] Plethysmography cold challenge [55]
PAH	6-min walk test [56, 57] NYHA or WHO functional class [58, 59] Right heart catheterization [60, 61] Time to clinical worsening, survival [61]	SF-36, VAS, and patient global assessments [62] SHAQ-DI NT-pro-BNP < BNP [63, 64] Pulmonary function testing [62] Anticentromere antibody [65] Dyspnea scale: Borg, Mahler [66] Telangiectasia [67] Echocardiographic parameters of RV function: TASPE, right ventricular volume, atrial volume, E/A ratio, maximum velocity of tricuspid valve regurgitation, pulmonary valve acceleration time, right ventricular systolic pressure [68, 69]	EKG [65]	High-resolution computerized tomography Exercise right heart catheterization, positron emission tomography Magnetic resonance imaging Magnetic resonance angiography Bronchoalveolar lavage Encouraged 6-min walk test DETECT algorithm [69]

(continued)

Table 46.1 (continued)

Organ system	Valid	Partially validated	Used but not completely valid	Emerging
Interstitial lung disease (ILD)	Forced vital capacity [16, 70, 71] Total lung capacity [16] Diffusing capacity for carbon monoxide [16, 70, 72] HRCT [16, 70, 73, 74] Mahler dyspnea [16, 70, 75–77] VAS breathing [16, 70, 78]		Exercise oxygen desaturation [79] 6-min walking distance [70, 71, 80]	Reduced radiation HRCT [81] UCSD shortness of breath questionnaire [82]
Skin	Modified Rodnan skin score [83–85] UCLA skin score [84, 86] Kahaleh skin score [87]	Durometry [88] Skin biopsy [89] VAS [90] SHAQ [91]	Self-related VAS [92] Skin self-assessment questionnaire [93] Maximum oral aperture Hand mobility Grip strength Tendon friction rub Skin thickness progression score	Plicometry [94] Elastometry [95] Ultrasound [96, 97] Serum makers of connective tissue metabolism
Gastrointestinal tract	UCLA GIT2.0 [98, 99]. Upper gastrointestinal (UGI) endoscopy [100–102] Biopsy [103–105] Manometer [106–111] Barium [112–115] Hydrogen and methane breath tests [116, 117]	Small bowel follow-through [118]	EKG [119, 120]. SPECT [121] UGI endoscopic US [122] Anal endoscopic US [123, 124]	CT enterography MR enteroclysis (MREc) MR enterography (MREg) Video capsule (smartpill) [125] PROMIS® GI [126]
Renal	Creatinine [127, 128] Creatinine clearance (MDRD) [129, 130]		Proteinuria [131] Renal blood flow [132, 133]	
Functional status	HAQ-DI [62, 134–137] United Kingdom functional score [138, 139] SF-36 version 2 PCS [62, 134–137] PROMIS® physical function SF-36 version 2 MCS [62, 134–137] SF-6D [140].	SF-36 vitality scale [136, 143]	Fatigue VAS [137, 139] Pain VAS [136, 137, 139]. Sleep VAS [137, 141] Patient global assessment VAS	MOS sleep scale [142]
Joints	Cochin hand function [144, 146] HAMIS [144–146]	MSK ultrasound [147]	Tender joint count Swollen joint count Tendon friction rub Pain VAS Pt global VAS Physician global VAS ESR, CRP [146]	MRI [146]
Muscle	sysQ [148]		Manual muscle testing [149, 151] Electromyogram [149, 151] Creatine phosphokinase [127, 130] Muscle pain, tenderness [150]	

CHF congestive heart failure, *EKG* electrocardiogram, *MRI* magnetic resonance imaging, *NT-pro-BNP* N-terminal pro b-type natriuretic peptide, *HAQ* health assessment questionnaire, *VAS* visual analog scale, *SF-36* Medical Outcome Study Short-Form 36, *PCS* physical component summary, *MCS* mental component summary, *NYHA* New York Heart Association, *WHO* World Health Organization, *SHAQ* Scleroderma Health Assessment Questionnaire, *TAPSE* tricuspid annular plane systolic excursion, *E/A* ratio of the early (E) to late (A) ventricular filling velocities, *HRCT* high-resolution computed tomography, *UCSD* University of California San Diego shortness of breath questionnaire, *UCLA GIT 2.0* University of California Los Angeles gastrointestinal questionnaire, *EKG* electrocardiogram, *SPECT* single-photon emission computed tomography, *PROMIS* patient-reported outcome measurement information system, *MDRD* modification of diet in renal disease, *HAMIS* hand mobility in scleroderma, *SYSQ* systemic sclerosis questionnaire, *MOS sleep scale* medical outcomes study, *CRP* C reactive protein

The Role of Surrogate Measurements

A surrogate end point is defined as a measure of a treatment effect that correlates or reflects a change in a clinical end point. Additionally, a surrogate end point is expected to predict clinical benefit based on epidemiologic, therapeutic, or pathophysiologic evidence [152]. Scleroderma is a complex disease with high rates of morbidity and case-specific mortality [153]. However, the use of mortality as a primary outcome is not feasible and requires longer study duration (years).

Surrogate end points are adopted as potential markers for clinically relevant outcomes and their response to therapy. Improved insights into the pathophysiologic pathways of SSc, in addition to identifying key cellular and molecular targets, pave the way for potential organ (pathway)-specific markers. Clinically addressed outcomes usually reflect organ function or organ-related complication. Dyspnea scales and 6-min walk distance are used as surrogate for PAH [154, 155]. FVC and HRCT are surrogates for ILD progression [8, 156]. Time to clinical worsening was considered a surrogate marker of PAH worsening in a recent study by Pulido et al. where they assessed the effect of macitentan (dual endothelin receptor antagonist) in a randomized controlled trial. They reported that macitentan significantly reduced morbidity and mortality in PAH patients [157]. Gene expression signature in the skin and peripheral blood play a major role in understanding SSc pathogenesis, identifying potential biomarkers and therapeutic targets [158]. Gene expression signatures were tested by Milano et al.; inflammatory, proliferative, limited, and normal skin patterns were identified in clustered analysis of intrinsic genes [159]. Further analyses of those intrinsic genes for changes in response to treatment were assessed by Hinchcliff et al., and differential expression was shown in MRSSs of MMF-responsive patients in comparison to nonresponders [160]. Chung et al. showed differential gene expression in the skin of two SSc patients examined before and after imatinib treatment; they also identified an imatinib-responsive signature which was differentially expressed in dcSSc (early and late) in comparison to lcSSc and normal skin [161]. Genetic studies reveal the potential value of gene signatures as surrogate markers of fibrosis and response to treatment in SSc patients, in addition to their contribution to the growing innovative field of personalized translational medicine.

Measurement Error in SSc Outcomes

Demonstration of measurable effect by a treatment in a clinical trial is of great importance. Application of treatments and diagnostic tests relies on scores obtained by the measured variable. As noted above, validated measures should adhere

to the OMERACT principles or a similar approach. In a study by Pope et al. [85], of ten rheumatologist and ten Ssc patients, they found that the intraobserver reliability was better than the interobserver reliability for most variables examined. Czirják et al. [162] demonstrated that, with repeated teaching of rheumatologists, the coefficient of variation of the measure decreased from 54% to 32%, while the intraclass correlation coefficient (ICC) increased from 0.496 to the expert level of 0.722. Clinical trials in Ssc thus need a carefully validated and reliable measurement instrument to ensure accurate and clinically meaningful results. Further, training to reduce inter-investigator variability seems to improve the usefulness of some clinical surrogates.

Patient Selection

Sample Size

A limitation in clinical study design in SSc is sample size because SSc is an uncommon/rare disease, so it is hard to enroll sufficient patients to have statistical power for confidence in the results. In addition, sample size calculation is dependent on a change in validated clinically relevant measures as the primary outcome, which requires a sample size of adequate number of patients to detect the change in such an outcome. For example, an adequately powered clinical trial of cyclophosphamide versus placebo, using FVC as the primary outcome, required about 150 patients. To recruit an adequate number of SSc patients in such a clinical trial in a timely manner, multisite trial designs are often adopted. This, in turn, requires consideration of the negative aspects of multicenter design: heterogeneity among patients, increased variability in outcome measures, reduced reliability among participating sites, and high cost.

Sampling Frame

SSc is a multisystem disease with various possible phenotypes; the phenotypic variability starts with the skin which yields two distinct SSc subtypes: limited (lcSSc) and diffuse cutaneous subtypes (dcSSc). Pope et al. studied SSc patients with both SSc subtypes to calculate the baseline characteristics of commonly used outcome measures and to provide parameters for sample size calculations for SSc clinical trials. Multiple baseline characteristics were significantly different in patients with diffuse SSc in comparison to patients with limited SSc, including health assessment questionnaire (HAQ) disability score, functional Index, grip strength, skin score, and physician global assessment [163]. SSc trials to date choose to enroll patients with diffuse cutaneous disease because the primary outcomes often chosen (e.g., skin or

lung changes) change more quickly in this subtype, despite the fact that the limited subtype is more common – often 60–70% of SSc population [164]. This approach may change as serological subtyping becomes more clearly defined and differentiating [165] or as genetic signatures as a more reliable method for subtyping on a pathogenetic basis becomes validated [161]. The predominance of fibrotic and inflammatory pathways in dcSSc versus vasculopathy in lcSSc supports the dcSSc vs. lcSSc grouping. However, genotypes may differ within the same subtype, pointing to the potential for a different subgrouping [159]. The potential here, not yet proven, is that patient populations in clinical trials will have more uniform pathogenetic backgrounds and, thus, more uniform response to appropriately targeted therapies.

Thus, patient selection at baseline has a substantial effect on the outcome measured; in cases of mild to moderate ILD in SSc patients, dyspnea and decreased quality of life (QOL) may be minimal, and improvement with treatment is not practical, which is not the case in severe ILD patients. Similarly, a lower baseline renal function in a clinical trial may allow us to discern small changes to define progressive renal dysfunction progression. Subsequently, variability in baseline severity could influence the outcomes measured. Accordingly, a careful consideration of possible predictable baseline differences for defining inclusions into the study (e.g., disease duration, disease activity, medications) is appropriate, as is a plan to account for baseline differences during analysis.

Disease Duration

The preliminary ACR criteria, developed in 1980 [166] for SSc, overlook the early stages of disease, with consequent delay in treatment. Matucci-Cerinic et al. developed a consensus for very early diagnosis of systemic sclerosis (VEDOSS) in 2009 to detect early symptoms/signs of SSc before the evolution of full-blown SSc. They identified the presence of Raynaud's phenomenon (RP), abnormal capillaroscopic pattern, and abnormal laboratory values (antinuclear, anticentromere, and antitopoisomerase-I antibodies) as major criteria for VEDOSS diagnosis [167]. A recent Delphi exercise in 2011 also documented four symptoms/signs necessary for VEDOSS: Raynaud's phenomenon, puffy fingers turning to sclerodactyly, specific SSc autoantibodies, and abnormal capillaroscopy with SSc pattern [168]. The importance of early identification of such abnormalities is to detect and treat as early as possible with potential to delay progression to fully defined SSc and, perhaps, to alter the long-term course of the disease. The development of the 2013 ACR/EULAR SSc criteria [169] improved the ability to diagnose SSc patients early, yet only 44% of the VEDOSS population fulfilled the new ACR/EULAR criteria [170]. Recently, a

study by Bruni et al. [171] showed that digital lesions (ulcers and scars) are present among 26% of 110 VEDOSS patients and demonstrated significant correlation with gastrointestinal involvement in VEDOSS patients. This actually implied that these VEDOSS patients may have had vasculopathic aspects of SSc well before being seen and diagnosed as VEDOSS patients. It is far too early to consider using VEDOSS as a criterion for trial design, but it is possible that it will be an important consideration in the future.

Trial Design

In 1995, the ACR published guidelines for designing clinical trial in patients with scleroderma [172]. Since then there have been significant advances in diagnostic testing, pathophysiological understanding, and treatment of the disease. Clinical trials should be designed using validated outcome measures, and the use of the OMERACT principles can be used to guide the use of those measures [6]. EULAR has recently put forward some point to consider when designing clinical trials in scleroderma [23] see Table 46.2.

Data Analysis

Data analysis of studies is a complex and individualized process, and a complete discussion cannot be undertaken in this section. A few points to consider are:

- Consider consulting with an expert for help with designing the trial.
- Design of the trial and outcomes will determine how the analysis is conducted and vice versa.
- The analysis should be prespecified before the trial starts, although exploratory analyses and work on validation of outcomes in early trials are encouraged.
- Critical to all trials is trying to minimize bias by using control groups and, if at all possible, blinding the trial as well as randomization of allocation.
- Sample size and power calculations for all phase III trials will depend upon the primary outcome measure(s), treatment duration, expected responses in the groups, and desired alpha and beta levels, among other factors. However, not all studies need to have a power analysis done (e.g., safety analysis, pharmacokinetics, some early phase 2 studies, and dose response trials are examples where power analysis is less important).
- Statistical analysis for in between group comparisons should consider the probability of distributions of the results (i.e., parametric vs. nonparametric variables).
- Outcome variables should be defined, using validated measures whenever possible. The characteristics of the

Table 46.2 Issues in clinical trial design

Trial design (all trials should be ethically sound)	Order of credibility:
	Fully statistically powered, randomized, controlled, double-blind trials are considered gold standard
	Possible designs:
	Active comparator
	Post trial provision of beneficial treatment
	Crossover design
	Randomized withdrawal design
	Randomized placebo phase design
Multiple n-of-1 trials	
Duration	<i>6 weeks to 36 months but organ specific. For example:</i> (a) 3–6 months for PAH and surrogate hemodynamic responses (b) 4–6 months for digital ulcer healing (c) 3 months for Raynaud’s phenomenon (d) 6 weeks for GI tract-related symptoms like dyspepsia (e) 6 months to 2 years but usually 6–12 months for skin changes and pulmonary fibrosis
Bio sampling	Collection and storage of tissue, blood, and other material if possible should be strongly considered
Inclusion and exclusion criteria	1. Limited vs. diffuse disease and severity of disease 2. Demographics 3. Exclusion vs. inclusion of children 4. Disease duration, early (<3 years) vs. late (>3 years) 5. Excluding confounders; medications, similar disease, drug exposures, end organ damage
Data analysis	See below
Outcomes	As per OMERACT principles or similar approaches and use of validated outcome measures, as above
Surrogate outcomes	Outcomes other than mortality can be used as primary outcomes

outcomes should be considered, as they may determine the robustness of the data when not normally distributed and the power of the statistics to discriminate among therapies. In general, for example, dichotomous measures do not have as much discriminatory power as continuous measures. Continuous measures are more able to discriminate among therapies than other approaches. If the continuous measures are particularly variable, nominal, categorical, or dichotomous measures are preferable. The specific analyses available are myriad – from simple proportions tests, through ANOVA, through generalized linear regressions with many variations, through survival analyses, etc. This is a very important reason to consult early with your statistical colleagues.

- Missing data, from single variables through patient dropout, are an inevitable aspect of clinical trial design, and there are multiple methods of imputing missing data, from simple completer analysis, through nonresponder imputation, through averaging, and through general linear equation modeling. The method chosen should be chosen in advance
- Adverse event reporting is as important as reporting of benefit and should be considered before the trial begins, although the methodology of such reporting remains unsophisticated. Data safety monitoring should be considered for larger or multicenter trials.

- Criteria for early termination of the trial and interim analysis should be prespecified, if needed.

Conclusion

Clinical trials in scleroderma are inherently difficult because the disease is uncommon/rare, making recruitment problematic and requiring multisite trials; longer trials are also often needed. Partly in response to these difficulties, clinical trial methodology in SSc is evolving and has been improving. This chapter reviewed updated issues in trial design including factors such as epidemiology, phases of trial design, outcome measures, surrogate measures, patient selection, analysis, and updated guidelines for trial design.

References

1. Clements PJ, Seibold JR, Furst DE, Mayes M, White B, Wigley F, Weisman MD, Barr W, Moreland L, Medsger Jr TA, Steen V, Martin RW, Collier D, Weinstein A, Lally E, Varga J, Weiner SR, Andrews B, Abeles M, Wong WK. Semin high-dose versus low-dose D-penicillamine in early diffuse systemic sclerosis trial: lessons learned. *Arthritis Rheum.* 2004;33(4):249–63.
2. Furst DE, Clements PJ, Hillis S, Lachenbruch PA, Miller BL, Sterz MG, Paulus HE. Immunosuppression with chlorambucil, versus placebo, for scleroderma. Results of a three-year, parallel, randomized, double-blind study. *Arthritis Rheum.* 1989;32(5):584–93.

3. Thompson AE, Pope JE. Increased prevalence of scleroderma in southwestern Ontario: a cluster analysis. *J Rheumatol.* 2002;29:1867–73.
4. Matucci-Cerinic M1, Denton CP, Furst DE, Mayes MD, Hsu VM, Carpentier P, Wigley FM, Black CM, Fessler BJ, Merkel PA, Pope JE, Sweiss NJ, Doyle MK, Hellmich B, Medsger Jr TA, Morganti A, Kramer F, Korn JH, Seibold JR. Bosentan treatment of digital ulcers related to systemic sclerosis: results from the RAPIDS-2 randomised, double-blind, placebo-controlled trial. *Ann Rheum Dis.* 2011;70(1):32–8. doi:10.1136/ard.2010.130658. Epub 2010 Aug 30.
5. <http://www.fda.gov/Drugs/DrugSafety/PostmarketDrugSafetyInformationforPatientsandProviders/ucm111350.htm>.
6. Boers M1, Kirwan JR2, Wells G3, Beaton D4, Gossec L5, D'Agostino MA6, Conaghan PG7, Bingham Jr CO8, Brooks P9, Landewé R10, March L11, Simon LS12, Singh JA13, Strand V14, Tugwell P15. Developing core outcome measurement sets for clinical trials: OMERACT filter 2.0. *J Clin Epidemiol.* 2014;67(7):745–53. doi:10.1016/j.jclinepi.2013.11.013. Epub 2014 Feb 28.
7. Williamson PR, Altman DG, Blazeby JM, Clarke M, Devane D, Gargon E, et al. Developing core outcome sets for clinical trials: issues to consider trials. *J Clin Epidemiol.* 2014;67(7):745–53. doi:10.1016/j.jclinepi.2013.11.013. Epub 2014 Feb 28.
8. Tashkin DP, Elashoff R, Clements PJ, Goldin J, Roth MD, Furst DE, Arriola E, Silver R, Strange C, Bolster M, Seibold JR, Riley DJ, Hsu VM, Varga J, Schraufnagel DE, Theodore A, Simms R, Wise R, Wigley F, White B, Steen V, Read C, Mayes M, Parsley E, Mubarak K, Connolly MK, Golden J, Olman M, Fessler B, Rothfield N, Metersky M, Scleroderma Lung Study Research Group. Cyclophosphamide versus placebo in scleroderma lung disease. *N Engl J Med.* 2006;354(25):2655–66.
9. Johnson SR, Feldman BM, Pope JE, Tomlinson GA. Shifting our thinking about uncommon disease trials: the case of methotrexate in scleroderma. *J Rheumatol.* 2009;36(2):323–9. doi:10.3899/jrheum.071169.
10. van Laar JM, Farge D, Sont JK, Naraghi K, Marjanovic Z, Larghero J, Schuerwegh AJ, Marijt EW, Vonk MC, Schattenberg AV, Matucci-Cerinic M, Voskuyl AE, van de Loosdrecht AA, Daikeler T, Kötter I, Schmalzing M, Martin T, Lioure B, Weiner SM, Kreuter A, Deligny C, Durand JM, Emery P, Machold KP, Sarrot-Reynauld F, Warnatz K, Adoue DF, Constans J, Tony HP, Del Papa N, Fassas A, Himsel A, Launay D, Lo Monaco A, Philippe P, Quére I, Rich É, Westhovens R, Griffiths B, Saccardi R, van den Hoogen FH, Fibbe WE, Socié G, Gratwohl A, Tyndall A, EBMT/EULAR Scleroderma Study Group. Autologous hematopoietic stem cell transplantation vs intravenous pulse cyclophosphamide in diffuse cutaneous systemic sclerosis: a randomized clinical trial. *JAMA.* 2014;311(24):2490–8. doi:10.1001/jama.2014.6368.
11. Clements PJ, Roth MD, Elashoff R, Tashkin DP, Goldin J, Silver RM, Sterz M, Seibold JR, Schraufnagel D, Simms RW, Bolster M, Wise RA, Steen V, Mayes MD, Connelly K, Metersky M, Furst DE, Scleroderma Lung Study Group. Scleroderma lung study (SLS): differences in the presentation and course of patients with limited versus diffuse systemic sclerosis. *Ann Rheum Dis.* 2007;66(12):1641–7.
12. Frech TM, Khanna D, Maranian P, et al. Probiotics for the treatment of systemic sclerosis-associated gastrointestinal bloating/distention. *Clin Exp Rheumatol.* 2011;29:S22–5.
13. Khanna D, Furst DE, Clements PJ, Park GS, Hays RD, Jeonglim Y, Korn JH, Merkel PA, Naomi R, Wigley FM, Moreland LW, Richard S, Steen VD, Michael W, Mayes MD, Collier DH, Medsger Jr TA, Seibold JR, Relaxin Study Group, et al. Responsiveness of the SF-36 and the health assessment questionnaire disability index in a systemic sclerosis clinical trial. *J Rheumatol.* 2005;32:832–40.
14. Kaldas M1, Khanna PP, Furst DE, Clements PJ, Kee Wong W, Seibold JR, Postlethwaite AE, Khanna D, investigators of the human recombinant relaxin and oral bovine collagen clinical trials. Sensitivity to change of the modified Rodnan skin score in diffuse systemic sclerosis—assessment of individual body sites in two large randomized controlled trials. *Rheumatology (Oxford).* 2009;48(9):1143–6. doi:10.1093/rheumatology/kep202. Epub 2009 Jul 14.
15. Khanna PP, Maranian P, Gregory J, Khanna D. The minimally important difference and patient acceptable symptom state for the Raynaud's condition score in patients with Raynaud's phenomenon in a large randomised controlled clinical trial. *Ann Rheum Dis.* 2010;69:588–91.
16. Khanna D, Seibold JR, Wells A, Distler O, Allanore Y, Denton C, Furst DE. Systemic sclerosis-associated interstitial lung disease: lessons from clinical trials, outcome measures, and future study design. *Curr Rheumatol Rev.* 2010;6(2):138–44. PMID: 20676227. PMC2911794.
17. Merkel PA, Silliman NP, Clements PJ, Denton CP, Furst DE, Mayes MD, Pope JE, Polisson RP, Streisand JB, Seibold JR, Scleroderma Clinical Trials Consortium. Patterns and predictors of change in outcome measures in clinical trials in scleroderma: an individual patient meta-analysis of 629 subjects with diffuse cutaneous systemic sclerosis. *Arthritis Rheum.* 2012;64(10):3420–9. doi:10.1002/art.34427.
18. Khanna D1, Distler O, Avouac J, Behrens F, Clements PJ, Denton C, Foeldvari I, Giannini E, Huscher D, Kowal-Bielecka O, Lovell D, Matucci-Cerinic M, Mayes M, Merkel PA, Nash P, Opitz CF, Pittrow D, Rubin L, Seibold JR, Steen V, Strand CV, Tugwell PS, Varga J, Zink A, Furst DE; CRIS; EPOSS. Measures of response in clinical trials of systemic sclerosis: the Combined Response Index for Systemic Sclerosis (CRIS) and Outcome Measures in Pulmonary Arterial Hypertension related to Systemic Sclerosis (EPOSS). *J. Rheumatol.* 2009;36(10):2356–61. doi: 10.3899/jrheum.090372.
19. Valentini G, Della Rossa A, Bombardieri S, Bencivelli W, Silman AJ, D'Angelo S, et al. European multicenter study to define disease activity criteria for systemic sclerosis. II. Identification of disease activity variable and development of a preliminary activity index. *Ann Rheum Dis.* 2001;60:592–8.
20. Valentini G, Bencivelli W, Bombardieri S, D'Angelo S, Della Rossa A, Silman AJ, et al. European scleroderma study group to define disease activity criteria for systemic sclerosis. III. Assessment of the construct validity of the preliminary activity criteria. *Ann Rheum Dis.* 2003;62:ar023186.
21. Valentini G, D'Angelo S, Della Rossa A, Bencivelli W, Bombardieri S. European scleroderma study group to define disease activity criteria for systemic sclerosis IV. Assessment of skin thickening by modified rodnan skin score. *Ann Rheum Dis.* 2003;62:904–5.
22. Minier T, Nagy Z, Balint Z, Farkas H, Radics J, Kuma' novics G b, Czmply T, Simon D, Varju C, Nemeth P, Czirjak L. Construct validity evaluation of the European Scleroderma Study Group activity index, and investigation of possible new disease activity markers in systemic sclerosis. *Rheumatology.* 2010;49:1133–45.
23. Khanna D, Furst DE, Allanore Y, Bae S, Bodukam V, Clements PJ, Cutolo M, Czirjak L, Denton CP, Distler O, Walker UA, Matucci-Cerinic M, Müller-Ladner U, Seibold JR, Singh M, Tyndall A. Twenty-two points to consider for clinical trials in systemic sclerosis, based on EULAR standards. *Rheumatology (Oxford).* 2014. pii: keu288.
24. Owens GR, Follansbee WP. Cardiopulmonary manifestations of systemic sclerosis. *Chest.* 1987;91:118–27.
25. McWhorter JE, LeRoy EC. Pericardial disease in scleroderma (systemic sclerosis). *Am J Med.* 1974;57:566–75.

26. Smith JW, Clements PJ, Levisman J, Furst D, Ross M. Echocardiographic features of progressive systemic sclerosis (PSS). Correlation with hemodynamic and postmortem studies. *Am J Med.* 1979;66:28–33.
27. Antoniadis L, Sfikakis PP, Mavrikakis M. Glucocorticoid effects on myocardial performance in patients with systemic sclerosis. *Clin Exp Rheumatol.* 2001;19:431–7.
28. Hachulla AL, Launay D, Gaxotte V, et al. Cardiac magnetic resonance imaging in systemic sclerosis: a cross-sectional observational study of 52 patients. *Ann Rheum Dis.* 2009;68:1878–84.
29. Di Cesare E, Battisti S, Di Sibio A, et al. Early assessment of sub-clinical cardiac involvement in systemic sclerosis (SSc) using delayed enhancement cardiac magnetic resonance (CE-MRI). *Eur J Radiol.* 2013;82:e268–73.
30. Hoepfer MM, Bogaard HJ, Condliffe R, et al. Definitions and diagnosis of pulmonary hypertension. *J Am Coll Cardiol.* 2013;62:D42–50.
31. Halpern SD, Taichman DB. Misclassification of pulmonary hypertension due to reliance on pulmonary capillary wedge pressure rather than left ventricular end-diastolic pressure. *Chest.* 2009;136:37–43.
32. Chung L, Chen H, Khanna D, Steen VD. Dyspnea assessment and pulmonary hypertension in patients with systemic sclerosis: utility of the University of California, San Diego, Shortness of Breath Questionnaire. *Arthritis Care Res.* 2013;65:454–63.
33. Candell-Riera J, Armadans-Gil L, Simeon CP, et al. Comprehensive noninvasive assessment of cardiac involvement in limited systemic sclerosis. *Arthritis Rheum.* 1996;39:1138–45.
34. Steen VD, Follansbee WP, Conte CG, Medsger Jr TA. Thallium perfusion defects predict subsequent cardiac dysfunction in patients with systemic sclerosis. *Arthritis Rheum.* 1996;39:677–81.
35. Kostis JB, Seibold JR, Turkevich D, et al. Prognostic importance of cardiac arrhythmias in systemic sclerosis. *Am J Med.* 1988;84:1007–15.
36. Follansbee WP, Curtiss EI, Rahko PS, et al. The electrocardiogram in systemic sclerosis (scleroderma). Study of 102 consecutive cases with functional correlations and review of the literature. *Am J Med.* 1985;79:183–92.
37. Choi HJ, Shin YK, Lee HJ, et al. The clinical significance of serum N-terminal pro-brain natriuretic peptide in systemic sclerosis patients. *Clin Rheumatol.* 2008;27:437–42.
38. Assassi S, Del Junco D, Sutter K, et al. Clinical and genetic factors predictive of mortality in early systemic sclerosis. *Arthritis Rheum.* 2009;61:1403–11.
39. Kahan A, Devaux JY, Amor B, et al. Nifedipine and thallium-201 myocardial perfusion in progressive systemic sclerosis. *N Engl J Med.* 1986;314:1397–402.
40. Ferri C, Di Bello V, Martini A, et al. Heart involvement in systemic sclerosis: an ultrasonic tissue characterisation study. *Ann Rheum Dis.* 1998;57:296–302.
41. Geyer H, Caracciolo G, Abe H, et al. Assessment of myocardial mechanics using speckle tracking echocardiography: fundamentals and clinical applications. *J Am Soc Echocardiogr: Off Publ Am Soc Echocardiogr.* 2010;23:351–69; quiz 453–5.
42. Flett AS, Hayward MP, Ashworth MT, et al. Equilibrium contrast cardiovascular magnetic resonance for the measurement of diffuse myocardial fibrosis: preliminary validation in humans. *Circulation.* 2010;122:138–44.
43. Mok MY, Lau CS, Chiu SS, et al. Systemic sclerosis is an independent risk factor for increased coronary artery calcium deposition. *Arthritis Rheum.* 2011;63:1387–95.
44. Wigley FM, Wise RA, Seibold JR, et al. Intravenous iloprost infusion in patients with Raynaud phenomenon secondary to systemic sclerosis. A multicenter, placebo-controlled, double-blind study. *Ann Intern Med.* 1994;120:199–206.
45. Merkel PA, Herlyn K, Martin RW, et al. Measuring disease activity and functional status in patients with scleroderma and Raynaud's phenomenon. *Arthritis Rheum.* 2002;46:2410–20.
46. Baron M, Chung L, Gyger G, et al. Consensus opinion of a North American working group regarding the classification of digital ulcers in systemic sclerosis. *Clin Rheumatol.* 2014;33:207–14.
47. Rannou F, Poiraudeau S, Berezne A, et al. Assessing disability and quality of life in systemic sclerosis: construct validities of the Cochin Hand Function Scale, Health Assessment Questionnaire (HAQ), systemic sclerosis HAQ, and medical outcomes study 36-item short form health survey. *Arthritis Rheum.* 2007;57:94–102.
48. Impens AJ, Chung KC, Buch MH, et al. Influences of clinical features of systemic sclerosis (SSc) on the Michigan Hand Questionnaire (MHQ). *Arthritis Rheum.* 2006;54:S483.
49. Sebastiani M, Manfredi A, Colaci M, et al. Capillaroscopic skin ulcer risk index: a new prognostic tool for digital skin ulcer development in systemic sclerosis patients. *Arthritis Rheum.* 2009;61:688–94.
50. Hasegawa M, Nagai Y, Tamura A, Ishikawa O. Arteriographic evaluation of vascular changes of the extremities in patients with systemic sclerosis. *Br J Dermatol.* 2006;155:1159–64.
51. Allanore Y, Seror R, Chevrot A, Kahan A, Drape JL. Hand vascular involvement assessed by magnetic resonance angiography in systemic sclerosis. *Arthritis Rheum.* 2007;56:2747–54.
52. Black CM, Halkier-Sorensen L, Belch JJ, et al. Oral iloprost in Raynaud's phenomenon secondary to systemic sclerosis: a multicentre, placebo-controlled, dose-comparison study. *Br J Rheumatol.* 1998;37:952–60.
53. Wigley FM, Korn JH, Csuka ME, et al. Oral iloprost treatment in patients with Raynaud's phenomenon secondary to systemic sclerosis: a multicenter, placebo-controlled, double-blind study. *Arthritis Rheum.* 1998;41:670–7.
54. Chucker FD, Fowler RC, Hurley CW. Photoplethysmometry and thermography in Raynaud's disorders. A preliminary report. *Angiology.* 1973;24:612–8.
55. Herrick AL, Clark S. Quantifying digital vascular disease in patients with primary Raynaud's phenomenon and systemic sclerosis. *Ann Rheum Dis.* 1998;57:70–8.
56. Nickel N, Golpon H, Greer M, et al. The prognostic impact of follow-up assessments in patients with idiopathic pulmonary arterial hypertension. *Eur Respir J.* 2012;39:589–96.
57. Barst RJ, Rubin LJ, Long WA, et al. A comparison of continuous intravenous epoprostenol (prostacyclin) with conventional therapy for primary pulmonary hypertension. *N Engl J Med.* 1996;334:296–301.
58. Taichman DB, McGoon MD, Harhay MO, et al. Wide variation in clinicians' assessment of New York Heart Association/World Health Organization functional class in patients with pulmonary arterial hypertension. *Mayo Clin Proc.* 2009;84:586–92.
59. Galie N, Hoepfer MM, Humbert M, et al. Guidelines for the diagnosis and treatment of pulmonary hypertension: the task force for the diagnosis and treatment of pulmonary hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS), endorsed by the International Society of Heart and Lung Transplantation (ISHLT). *Eur Heart J.* 2009;30:2493–537.
60. Denton CP, Caires JB, Phillips GD, Wells AU, Black CM, Bois RM. Comparison of Doppler echocardiography and right heart catheterization to assess pulmonary hypertension in systemic sclerosis. *Br J Rheumatol.* 1997;36:239–43.
61. McLaughlin VV, Badesch DB, Delcroix M, et al. End points and clinical trial design in pulmonary arterial hypertension. *J Am Coll Cardiol.* 2009;54:S97–107.
62. Khanna D, Clements PJ, Furst DE, et al. Correlation of the degree of dyspnea with health-related quality of life, functional abilities,

- and diffusing capacity for carbon monoxide in patients with systemic sclerosis and active alveolitis: results from the Scleroderma Lung Study. *Arthritis Rheum.* 2005;52:592–600.
63. Williams MH, Handler CE, Akram R, et al. Role of N-terminal brain natriuretic peptide (N-TproBNP) in scleroderma-associated pulmonary arterial hypertension. *Eur Heart J.* 2006;27:1485–94.
 64. Allanore Y, Borderie D, Avouac J, et al. High N-terminal pro-brain natriuretic peptide levels and low diffusing capacity for carbon monoxide as independent predictors of the occurrence of precapillary pulmonary arterial hypertension in patients with systemic sclerosis. *Arthritis Rheum.* 2008;58:284–91.
 65. Thenappan T, Shah SJ, Rich S, Gomberg-Maitland M. A USA-based registry for pulmonary arterial hypertension: 1982–2006. *Eur Respir J.* 2007;30:1103–10.
 66. O'Donnell DE, Chau LK, Webb KA. Qualitative aspects of exertional dyspnea in patients with interstitial lung disease. *J Appl Physiol.* 1998;84:2000–9.
 67. Johnson SR, Fransen J, Khanna D, et al. Validation of potential classification criteria for systemic sclerosis. *Arthritis Care Res.* 2012;64:358–67.
 68. Hachulla E, Gressin V, Guillemin L, et al. Early detection of pulmonary arterial hypertension in systemic sclerosis: a French nationwide prospective multicenter study. *Arthritis Rheum.* 2005;52:3792–800.
 69. Coghlan JG, Denton CP, Grunig E, et al. Evidence-based detection of pulmonary arterial hypertension in systemic sclerosis: the DETECT study. *Ann Rheum Dis.* 2014;73:1340–9.
 70. Khanna D, Brown KK, Clements PJ, Elashoff R, Furst DE, Goldin J, Seibold JR, Silver RM, Tashkin DP, Wells AU. Systemic sclerosis-associated interstitial lung disease-proposed recommendations for future randomized clinical trials. *Clin Exp Rheumatol.* 2010;28(2 Suppl 58):S55–62.
 71. Bouros D, Wells AU, Nicholson AG, Colby TV, Polychronopoulos V, Pantelidis P, Haslam PL, Vassilakis DA, Black CM, du Bois RM. Histopathologic subsets of fibrosing alveolitis in patients with systemic sclerosis and their relationship to outcome. *Am J Respir Crit Care Med.* 2002;165(12):1581–6.
 72. Latsi PI, du Bois RM, Nicholson AG, Colby TV, Bisirtzoglou D, Nikolakopoulou A, Veeraraghavan S, Hansell DM, Wells AU. Fibrotic idiopathic interstitial pneumonia: the prognostic value of longitudinal functional trends. *Am J Respir Crit Care Med.* 2003;168(5):531–7.
 73. Goldin J, Elashoff R, Kim HJ, Yan X, Lynch D, Strollo D, Roth MD, Clements P, Furst DE, Khanna D, Vasunilashorn S, Li G, Tashkin DP. Treatment of scleroderma-interstitial lung disease with cyclophosphamide is associated with less progressive fibrosis on serial thoracic high-resolution CT scan than placebo: findings from the scleroderma lung study. *Chest.* 2009;136(5):1333–40. PMC2773360.
 74. Goldin JG, Lynch DA, Strollo DC, Suh RD, Schraufnagel DE, Clements PJ, Elashoff RM, Furst DE, Vasunilashorn S, McNitt-Gray MF, Brown MS, Roth MD, Tashkin DP, Scleroderma Lung Study Research G. High-resolution CT scan findings in patients with symptomatic scleroderma-related interstitial lung disease. *Chest.* 2008;134(2):358–67.
 75. Khanna D, Clements PJ, Furst DE, Chon Y, Elashoff R, Roth MD, Sterz MG, Chung J, FitzGerald JD, Seibold JR, Varga J, Theodore A, Wigley FM, Silver RM, Steen VD, Mayes MD, Connolly MK, Fessler BJ, Rothfield NF, Mubarak K, Molitor J, Tashkin DP, Scleroderma Lung Study G. Correlation of the degree of dyspnea with health-related quality of life, functional abilities, and diffusing capacity for carbon monoxide in patients with systemic sclerosis and active alveolitis: results from the Scleroderma Lung Study. *Arthritis Rheum.* 2005;52(2):592–600.
 76. Khanna D, Tseng CH, Furst DE, Clements PJ, Elashoff R, Roth M, Elashoff D, Tashkin DP, for Scleroderma Lung Study I. Minimally important differences in the Mahler's Transition Dyspnoea Index in a large randomized controlled trial – results from the Scleroderma Lung Study. *Rheumatology (Oxford).* 2009;48(12):1537–40. PMC2777487.
 77. Khanna D, Yan X, Tashkin DP, Furst DE, Elashoff R, Roth MD, Silver R, Strange C, Bolster M, Seibold JR, Riley DJ, Hsu VM, Varga J, Schraufnagel DE, Theodore A, Simms R, Wise R, Wigley F, White B, Steen V, Read C, Mayes M, Parsley E, Mubarak K, Connolly MK, Golden J, Olman M, Fessler B, Rothfield N, Metersky M, Clements PJ, Scleroderma Lung Study G. Impact of oral cyclophosphamide on health-related quality of life in patients with active scleroderma lung disease: results from the scleroderma lung study. *Arthritis Rheum.* 2007;56(5):1676–84.
 78. Steen VD, Medsger Jr TA. The value of the health assessment questionnaire and special patient-generated scales to demonstrate change in systemic sclerosis patients over time. *Arthritis Rheum.* 1997;40(11):1984–91.
 79. Swigris JJ, Zhou X, Wamboldt FS, du Bois R, Keith R, Fischer A, Cosgrove GP, Frankel SK, Curran-Everett D, Brown KK. Exercise peripheral oxygen saturation (SpO2) accurately reflects arterial oxygen saturation (SaO2) and predicts mortality in systemic sclerosis. *Thorax.* 2009;64(7):626–30. PMC3667987.
 80. Buch MH, Denton CP, Furst DE, Guillemin L, Rubin LJ, Wells AU, Matucci-Cerinic M, Riemekasten G, Emery P, Chadha-Boreham H, Charef P, Roux S, Black CM, Seibold JR. Submaximal exercise testing in the assessment of interstitial lung disease secondary to systemic sclerosis: reproducibility and correlations of the 6-min walk test. *Ann Rheum Dis.* 2007;66(2):169–73.
 81. Frauenfelder T, Winklehner A, Nguyen TD, Dobrota R, Baumuegger S, Maurer B, Distler O. Screening for interstitial lung disease in systemic sclerosis: performance of high-resolution CT with limited number of slices: a prospective study. *Ann Rheum Dis.* 2014;73:2069–73.
 82. Chung L, Chen H, Khanna D, Steen VD. [Dyspnea assessment and pulmonary hypertension in patients with systemic sclerosis: utility of the University of California, San Diego, Shortness of Breath Questionnaire.](#) *Arthritis Care Res (Hoboken).* 2013;65(3):454–63. doi:10.1002/acr.21827.
 83. Clements PJ, Lachenbruch PA, Seibold JR, et al. Skin thickness score in systemic sclerosis: an assessment of interobserver variability in 3 independent studies. *J Rheumatol.* 1993;20:1892–6.
 84. Clements P, Lachenbruch P, Siebold J, et al. Inter and intraobserver variability of total skin thickness score (modified Rodnan TSS) in systemic sclerosis. *J Rheumatol.* 1995;22:1281–5.
 85. Pope JE, Baron M, Bellamy N, et al. Variability of skin scores and clinical measurements in scleroderma. *J Rheumatol.* 1995;22:1271–6.
 86. Clements PJ, Lachenbruch PA, Ng SC, Simmons M, Sterz M, Furst DE. Skin score. A semiquantitative measure of cutaneous involvement that improves prediction of prognosis in systemic sclerosis. *Arthritis Rheum.* 1990;33:1256–63.
 87. Kahaleh MB, Sultany GL, Smith EA, Huffstutter JE, Loadholt CB, LeRoy EC. A modified scleroderma skin scoring method. *Clin Exp Rheumatol.* 1986;4:367–9.
 88. Merkel PA, Silliman NP, Denton CP, et al. Validity, reliability, and feasibility of durometer measurements of scleroderma skin disease in a multicenter treatment trial. *Arthritis Rheum.* 2008;59:699–705.
 89. Furst DE, Clements PJ, Steen VD, et al. The modified Rodnan skin score is an accurate reflection of skin biopsy thickness in systemic sclerosis. *J Rheumatol.* 1998;25:84–8.
 90. Kuhn A, Haust M, Ruland V, et al. Effect of bosentan on skin fibrosis in patients with systemic sclerosis: a prospective, open-label, non-comparative trial. *Rheumatology.* 2010;49:1336–45.
 91. Denton CP, Engelhart M, Tvede N, et al. An open-label pilot study of infliximab therapy in diffuse cutaneous systemic sclerosis. *Ann Rheum Dis.* 2009;68:1433–9.

92. Sandqvist G, Akesson A, Eklund M. Evaluation of paraffin bath treatment in patients with systemic sclerosis. *Disabil Rehabil*. 2004;26:981–7.
93. Muellegger RR, Hofer A, Salmhofer W, Soyer HP, Kerl H, Wolf P. Extended extracorporeal photochemotherapy with extracorporeal administration of 8-methoxypsoralen in systemic sclerosis. An Austrian single-center study. *Photodermatol Photoimmunol Photomed*. 2000;16:216–23.
94. Basso M, Filaci G, Cutolo M, et al. Long-term treatment of patients affected by systemic sclerosis with cyclosporin A. *Ann Ital Med Int: Organo Ufficiale Soc Ital Med Intern*. 2001;16:233–9.
95. Balbir-Gurman A, Denton CP, Nichols B, et al. Non-invasive measurement of biomechanical skin properties in systemic sclerosis. *Ann Rheum Dis*. 2002;61:237–41.
96. Scheja A, Akesson A. Comparison of high frequency (20 MHz) ultrasound and palpation for the assessment of skin involvement in systemic sclerosis (scleroderma). *Clin Exp Rheumatol*. 1997;15:283–8.
97. Akesson A, Hesselstrand R, Scheja A, Wildt M. Longitudinal development of skin involvement and reliability of high frequency ultrasound in systemic sclerosis. *Ann Rheum Dis*. 2004;63:791–6.
98. Khanna D, Hays RD, Maranian P, Seibold JR, Impens A, Mayes MD, et al. Reliability and validity of the University of California, Los Angeles scleroderma clinical trial consortium gastrointestinal tract instrument. *Arthritis Rheum*. 2009;61:1257–63.
99. Khanna D, Hays RD, Park GS, Braun-Moscovici Y, Mayes MD, McNearney TA, et al. Development of a preliminary scleroderma gastrointestinal tract 1.0 quality of life instrument. *Arthritis Rheum*. 2007;57:1280–6.
100. Hung EW, Mayes MD, Sharif R, Assassi S, Machicao VI, Hosing C, St Clair EW, Furst DE, Khanna D, Forman S, Mineishi S, Phillips K, Seibold JR, Bredeson C, Csuka ME, Nash RA, Wener MH, Simms R, Ballen K, Leclercq S, Storek J, Goldmuntz E, Welch B, Keyes-Elstein L, Castina S, Crofford LJ, Mcsweeney P, Sullivan KMJ. Gastric antral vascular ectasia and its clinical correlates in patients with early diffuse systemic sclerosis in the SCOT trial. *Rheumatology*. 2013;40(4):455–60. doi:10.3899/jrheum.121087. Epub 2013 Feb 15.
101. Ghrénassia E, Avouac J, Khanna D, Derk CT, Distler O, Suliman YA, Airo P, Carreira PE, Foti R, Granel B, Berezne A, Cabane J, Ingegnoli F, Rosato E, Caramaschi P, Hesselstrand R, Walker UA, Alegre-Sancho JJ, Zarrouk V, Agard C, Riccieri V, Schioppa E, Gladue H, Steen VD, Allanore Y. Prevalence, correlates and outcomes of gastric antral vascular ectasia in systemic sclerosis: a EUSTAR case-control study. *J Rheumatol*. 2014;41(1):99–105. doi:10.3899/jrheum.130386. Epub 2013 Dec 1.
102. Thonhofer R, Siegel C, Trummer M, Graninger W. Early endoscopy in systemic sclerosis without gastrointestinal symptoms. *Rheumatol Int*. 2012;32(1):165–8. doi:10.1007/s00296-010-1595-y.
103. Hendel L1, Hage E, Hendel J, Stentoft P. Omeprazole in the long-term treatment of severe gastro-oesophageal reflux disease in patients with systemic sclerosis. *Aliment Pharmacol Ther*. 1992;6(5):565–77.
104. Manetti M1, Milia AF, Benelli G, Messerini L, Matucci-Cerinic M, Ibba-Manneschi L. The gastric wall in systemic sclerosis patients: a morphological study. *Ital J Anat Embryol*. 2010;115(1–2):115–21.
105. Hendel L. Hydroxyproline in the oesophageal mucosa of patients with progressive systemic sclerosis during an omeprazole-induced healing of reflux oesophagitis. *Aliment Pharmacol Ther*. 1991;5(5):471–80.
106. Horikoshi T, Matsuzaki T, Sekiguchi T. Effect of H2-receptor antagonists cimetidine and famotidine on interdigestive gastric motor activity and lower esophageal sphincter pressure in progressive systemic sclerosis. *Intern Med (Tokyo, Japan)*. 1994;33(7):407–12.
107. Weihsrauch TR, Korting GW. Manometric assessment of oesophageal involvement in progressive systemic sclerosis, morphea and Raynaud's disease. *Br J Dermatol*. 1982;107(3):325–32.
108. Blom-Bülow B, Sundström G, Jonson B, Tylén U, Wollheim FA. Early changes in oesophageal function in progressive systemic sclerosis: a comparison of manometry and radiology. *Clin Physiol*. 1984;4(2):147–58.
109. Stentoft P, Hendel L, Aggestrup S. Esophageal manometry and pH-probe monitoring in the evaluation of gastroesophageal reflux in patients with progressive systemic sclerosis. *Scand J Gastroenterol*. 1987;22(4):499–504.
110. Ipsen P, Egekvist H, Aksglaede K, Zachariae H, Bjerring P, Thommesen P. Oesophageal manometry and video-radiology in patients with systemic sclerosis: a retrospective study of its clinical value. *Acta Derm Venereol*. 2000;80:130–3.
111. Mainie I, Tutuian R, Patel A, Castell DO. Regional esophageal dysfunction in scleroderma and achalasia using multichannel intraluminal impedance and manometry. *Dig Dis Sci*. 2008;53(1):210–6.
112. Owen JP, Muston HL, Goolamali SK. Absence of oesophageal mucosal folds in systemic sclerosis. *Clin Radiol*. 1979;30(5):489–92.
113. Dantas RO, Villanova MG, de Godoy RA. Esophageal dysfunction in patients with progressive systemic sclerosis and mixed connective tissue diseases. *Arq Gastroenterol*. 1985;22(3):122–6.
114. Montesi A, Pesaresi A, Cavalli ML, Ripa G, Candela M, Gabrielli A. Oropharyngeal and esophageal function in scleroderma. *Dysphagia*. 1991;6(4):219–23.
115. Sharma VK, Trilokraj T, Khaitan BK, Krishna SM. Profile of systemic sclerosis in a tertiary care center in North India. *Indian J Dermatol Venereol Leprol*. 2006;72(6):416–20.
116. Kaye SA, Lim SG, Taylor M, Patel S, Gillespie S, Black CM. Small bowel bacterial overgrowth in systemic sclerosis: detection using direct and indirect methods and treatment outcome. *Br J Rheumatol*. 1995;34(3):265–9.
117. Marie I, Ducrotté P, Denis P, Menard JF, Levesque H. Small intestinal bacterial overgrowth in systemic sclerosis. *Rheumatology (Oxford)*. 2009;48(10):1314–9.
118. Weston S, Thumshirn M, Wiste J, Camilleri M. Clinical and upper gastrointestinal motility features in systemic sclerosis and related disorders. *Am J Gastroenterol*. 1998;93(7):1085–9.
119. Wollaston DE, Xu X, Tokumaru O, Chen JD, McNearney TA. Patients with systemic sclerosis have unique and persistent alterations in gastric myoelectrical activity with acupressure to Neiguan point PC6. *J Rheumatol*. 2005;32(3):494–501.
120. Franck-Larsson K1, Hedenström H, Dahl R, Rönnblom A. Delayed gastric emptying in patients with diffuse versus limited systemic sclerosis, unrelated to gastrointestinal symptoms and myoelectric gastric activity. *Scand J Rheumatol*. 2003;32(6):348–55.
121. Mo J, Wang C, Wang S. Gastric emptying and intragastric distribution of liquid and solid meal in patients with systemic sclerosis. *Zhonghua Nei Ke Za Zhi*. 1996;35(8):530–2.
122. Zuber-Jerger I, Müller A, Kullmann F, Gelbmann CM, Endlicher E, Müller-Ladner U, Fleck M. Gastrointestinal manifestation of systemic sclerosis – thickening of the upper gastrointestinal wall detected by endoscopic ultrasound is a valid sign. *Rheumatology (Oxford)*. 2010;49(2):368–72. doi:10.1093/rheumatology/kep381. Epub 2009 Dec 14.
123. Bartosik I, Andréasson K, Starck M, Scheja A, Hesselstrand R. Vascular events are risk factors for anal incontinence in systemic sclerosis: a study of morphology and functional properties measured by anal endosonography and manometry. *Scand J Rheumatol*. 2014;43(5):391–7. doi:10.3109/03009742.2014.889210. Epub 2014 Apr 11.
124. Thoua NM, Schizas A, Forbes A, Denton CP, Emmanuel AV. Internal anal sphincter atrophy in patients with systemic

- sclerosis. *Rheumatology (Oxford)*. 2011;50(9):1596–602. doi:[10.1093/rheumatology/ker153](https://doi.org/10.1093/rheumatology/ker153). Epub 2011 Apr 18.
125. Marie I, Antoniotti M, Houivet E, Hachulla E, Maunoury V, Bienvenu B, Viennot S, Smail A, Duhaut P, Dupas JL, Dominique S, Hatron PY, Levesque H, Benichou J, Ducrotté P. Gastrointestinal mucosal abnormalities using videocapsule endoscopy in systemic sclerosis. *Aliment Pharmacol Ther*. 2014;40(2):189–99. doi:[10.1111/apt.12818](https://doi.org/10.1111/apt.12818). Epub 2014 Jun 2.
 126. Penn H, et al. Scleroderma renal crisis: patient characteristics and long-term outcomes. *QJM*. 2007;100(8):485–94.
 127. Nagaraja V, Hays RD, Khanna PP, Spiegel BM, Chang L, Melmed GY, Bolus R, Khanna D. Construct validity of the Patient Reported Outcomes Measurement Information System (PROMIS®) gastrointestinal symptom scales in systemic sclerosis. *Arthritis Care Res (Hoboken)*. 2014;66(11):1725–30.
 128. Scheja A, et al. Renal function is mostly preserved in patients with systemic sclerosis. *Scand J Rheumatol*. 2009;38:1–4.
 129. Mohamed RH, Zayed HS, Amin A. Renal disease in systemic sclerosis with normal serum creatinine. *Clin Rheumatol*. 2010;29(7):729–37. doi:[10.1007/s10067-010-1389-3](https://doi.org/10.1007/s10067-010-1389-3). Epub 2010 Feb 23.
 130. Kingdon EJ, Knight CJ, Dustan K, Irwin AG, Thomas M, Powis SH, Burns A, Hilson AJ, Black CM. Calculated glomerular filtration rate is a useful screening tool to identify scleroderma patients with renal impairment. *Rheumatology (Oxford)*. 2003;42(1):26–33.
 131. Seiberlich B, Hunzelmann N, Krieg T, Weber M, Schulze-Lohoff E. Intermediate molecular weight proteinuria and albuminuria identify scleroderma patients with increased morbidity. *Clin Nephrol*. 2008;70(2):110–7.
 132. Rivolta R1, Mascagni B, Berruti V, Quarto Di Palo F, Elli A, Scorza R, Castagnone D. Renal vascular damage in systemic sclerosis patients without clinical evidence of nephropathy. *Arthritis Rheum*. 1996;39(6):1030–4.
 133. Scorza R, Rivolta R, Mascagni B, Berruti V, Bazzi S, Castagnone D, di Quarto PF. Effect of iloprost infusion on the resistance index of renal vessels of patients with systemic sclerosis. *J Rheumatol*. 1997;24(10):1944–8.
 134. Khanna D, Furst DE, Clements PJ, et al. Responsiveness of the SF-36 and the health assessment questionnaire disability index in a systemic sclerosis clinical trial. *J Rheumatol*. 2005;32:832–40.
 135. Khanna D, Yan X, Tashkin DP, et al. Impact of oral cyclophosphamide on health-related quality of life in patients with active scleroderma lung disease: results from the scleroderma lung study. *Arthritis Rheum*. 2007;56:1676–84.
 136. Hudson M, Steele R, Lu Y, Thombs BD, Baron M. Work disability in systemic sclerosis. *J Rheumatol*. 2009;36:2481–6.
 137. Sekhon S, Pope J, Baron M. The minimally important difference in clinical practice for patient-centered outcomes including health assessment questionnaire, fatigue, pain, sleep, global visual analog scale, and SF-36 in scleroderma. *J Rheumatol*. 2010;37:591–8.
 138. Smyth AE, MacGregor AJ, Mukerjee D, Brough GM, Black CM, Denton CP. A cross-sectional comparison of three self-reported functional indices in scleroderma. *Rheumatology (Oxford)*. 2003;42:732–8.
 139. Sandqvist G, Scheja A, Hesselstrand R. Pain, fatigue and hand function closely correlated to work ability and employment status in systemic sclerosis. *Rheumatology (Oxford)*. 2010;49:1739–46.
 140. Khanna D, Furst DE, Wong WK, et al. Reliability, validity, and minimally important differences of the SF-6D in systemic sclerosis. *Qual Life Res*. 2007;16:1083–92.
 141. Milette K, Razykov I, Pope J, et al. Clinical correlates of sleep problems in systemic sclerosis: the prominent role of pain. *Rheumatology (Oxford)*. 2011;50(5):921–5.
 142. Frech T, Hays RD, Maranian P, Clements PJ, Furst DE, Khanna D. Prevalence and correlates of sleep disturbance in systemic sclerosis – results from the UCLA scleroderma quality of life study. *Rheumatology (Oxford)*. 2011;50:1280–7.
 143. Thombs BD, Hudson M, Bassel M, Taillefer SS, Baron M. Sociodemographic, disease, and symptom correlates of fatigue in systemic sclerosis: evidence from a sample of 659 Canadian Scleroderma Research Group Registry patients. *Arthritis Rheum*. 2009;61:966–73.
 144. Bongi SM, Del Rosso A, Galluccio F, Sigismondi F, Miniati I, Conforti ML, et al. Efficacy of connective tissue massage and Mc Mennell joint manipulation in the rehabilitative treatment of the hands in systemic sclerosis. *Clin Rheumatol*. 2009;28(10):1167–73.
 145. Sandqvist G, Eklund M. Hand Mobility in Scleroderma (HAMIS) test: the reliability of a novel hand function test. *Arthritis Care Res*. 2000;13(6):369–74.
 146. Clements PJ, Allanore Y, Khanna D, Singh M, Furst DE. Arthritis in systemic sclerosis: systematic review of the literature and suggestions for the performance of future clinical trials in systemic sclerosis. *Semin Arthritis Rheum*. 2012;41:801–14.
 147. Scheiman-Elazary A, Ranganath VK, Ben-Artzi A, Duan L, Kafaja S, Borazan NH, Woodworth T, Elashoff D, Clements P, Furst DE. Validation of musculoskeletal us of hands and wrists in patients with systemic sclerosis abstract. *Eular 2014. Annals of the Rheumatic Diseases* 73(Suppl 2):651–651.
 148. Ruof J, Brühlmann P, Michel BA, Stucki G. Development and validation of a self-administered systemic sclerosis questionnaire (SySQ). *Rheumatology*. 1999;38(6):535–42. doi:[10.1093/rheumatology/38.6.535](https://doi.org/10.1093/rheumatology/38.6.535).
 149. Ranque B, Bérezné A, Le-Guern V, Pagnoux C, Allanore Y, Launay D, Hachulla E, Authier FJ, Gherardi R, Kahan A, Cabane J, Guillevin L, Mouthon L. Myopathies related to systemic sclerosis: a case-control study of associated clinical and immunological features. *Scand J Rheumatol*. 2010;39(6):498–505. doi:[10.3109/03009741003774626](https://doi.org/10.3109/03009741003774626). Epub 2010 Aug 20.
 150. Au K, Mayes MD, Maranian P, Clements PJ, Khanna D, Steen VD, et al. Course of dermal ulcers and musculoskeletal involvement in systemic sclerosis patients in the scleroderma lung study. *Arthritis Care Res (Hoboken)*. 2010;62(12):1772–8. ms tenderness.
 151. Clements PJ, Furst DE, Champion DS, Bohan A, Harris R, Levy J, Paulus HE. Muscle disease in progressive systemic sclerosis: diagnostic and therapeutic considerations. *Arthritis Rheum*. 1978;21(1):62–71.
 152. Cohn JN. Introduction to surrogate markers. *Circulation*. 2004;109(25 Suppl 1):IV20–1.
 153. Varga J. Systemic sclerosis. An update. *Bull NYU Hosp Jt Dis*. 2008;66(3):198–202.
 154. Kabunga P, Coghlan G. Endothelin receptor antagonism: role in the treatment of pulmonary arterial hypertension related to scleroderma. *Drugs*. 2008;68(12):1635–45.
 155. Launay D, Sitbon O, Le Pavé J, Savale L, Tchérakian C, Yaïci A, Achouh L, Parent F, Jais X, Simonneau G, Humbert M. Long-term outcome of systemic sclerosis-associated pulmonary arterial hypertension treated with bosentan as first-line monotherapy followed or not by the addition of prostanoids or sildenafil. *Rheumatology (Oxford)*. 2010;49(3):490–500. doi:[10.1093/rheumatology/kep398](https://doi.org/10.1093/rheumatology/kep398). Epub 2009 Dec 16.
 156. Goh NS, Desai SR, Veeraraghavan S, Hansell DM, Copley SJ, Maher TM, Corte TJ, Sander CR, Ratoff J, Devaraj A, Bozovic G, Denton CP, Black CM, du Bois RM, Wells AU. Interstitial lung disease in systemic sclerosis: a simple staging system. *Am J Respir Crit Care Med*. 2008;177(11):1248–54. doi:[10.1164/rccm.200706-877OC](https://doi.org/10.1164/rccm.200706-877OC). Epub 2008 Mar 27.
 157. Pulido T, Adzerikho I, Channick RN, Delcroix M, Galiè N, Ghofrani HA, Jansa P, Jing ZC, Le Brun FO, Mehta S, Mittelholzer CM, Perchenet L, Sastry BK, Sitbon O, Souza R, Torbicki A, Zeng X, Rubin LJ, Simonneau G, SERAPHIN Investigators. Macitentan and morbidity and mortality in pulmonary arterial

- hypertension. *N Engl J Med*. 2013;369(9):809–18. doi:[10.1056/NEJMoa1213917](https://doi.org/10.1056/NEJMoa1213917).
158. Assassi S, Mayes MD. What does global gene expression profiling tell us about the pathogenesis of systemic sclerosis? *Curr Opin Rheumatol*. 2013;25(6):686–91. doi:[10.1097/01.bor.0000434672.77891.41](https://doi.org/10.1097/01.bor.0000434672.77891.41).
159. Milano A, Pendergrass SA, Sargent JL, George LK, McCalmont TH, Connolly MK, Whitfield ML. Molecular subsets in the gene expression signatures of scleroderma skin. *PLoS ONE*. 2008;16(7):e2696. doi:[10.1371/journal.pone.0002696](https://doi.org/10.1371/journal.pone.0002696).
160. Hinchcliff M, Huang CC, Wood TA, Matthew Mahoney J, Martyanov V, Bhattacharyya S, Tamaki Z, Lee J, Carns M, Podlasky S, Sirajuddin A, Shah SJ, Chang RW, Lafyatis R, Varga J, Whitfield ML. Molecular signatures in skin associated with clinical improvement during mycophenolate treatment in systemic sclerosis. *J Invest Dermatol*. 2013;133(8):1979–89. doi:[10.1038/jid.2013.130](https://doi.org/10.1038/jid.2013.130). Epub 2013 Mar 14002E.
161. Chung L, Fiorentino DF, Benbarak MJ, Adler AS, Mariano MM, Paniagua RT, Milano A, Connolly MK, Ratiner BD, Wiskocil RL, Whitfield ML, Chang HY, Robinson WH. Molecular framework for response to imatinib mesylate in systemic sclerosis. *Arthritis Rheum*. 2009;60(2):584–91. doi:[10.1002/art.24221](https://doi.org/10.1002/art.24221).
162. Czirjak L, Nagy Z, Aringer M, et al. The EUSTAR model for teaching and implementing the modified Rodnan skin score in systemic sclerosis. *Ann Rheum Dis*. 2007;66:966–9.
163. Pope JE, Bellamy N. Sample size calculations in scleroderma trials. *Clin Invest Med*. 1995;18(1):1–10.
164. Rosa JE, Soriano ER, Narvaez-Ponce L, et al. Incidence and prevalence of systemic sclerosis in a healthcare plan in Buenos Aires. *J Clin Rheumatol*. 2011;17:59–63.
165. Steen VD. Autoantibodies in systemic sclerosis. *Semin Arthritis Rheum*. 2005;35(1):35–42.
166. Preliminary criteria for the classification of systemic sclerosis (scleroderma). Subcommittee for scleroderma criteria of the American rheumatism association diagnostic and therapeutic criteria committee. *Arthritis Rheum* 1980;23:581–90.
167. Matucci-Cerinic M, Allanore Y, Czirják L, et al. The challenge of early systemic sclerosis for the EULAR Scleroderma Trial and Research group (EUSTAR) community. It is time to cut the Gordian knot and develop a prevention or rescue strategy. *Ann Rheum Dis*. 2009;68:1377–80.
168. Avouac J, Fransen J, Walker UA, et al. Preliminary criteria for the very early diagnosis of systemic sclerosis: results of a Delphi Consensus Study from EULAR Scleroderma Trials and Research Group. *Ann Rheum Dis*. 2011;70:476.
169. van den Hoogen F, Khanna D, Fransen J, et al. 2013 classification criteria for systemic sclerosis: an American College of Rheumatology/European League against Rheumatism collaborative initiative. *Arthritis Rheum*. 2013;65:2737–47.
170. Minier T, Guiducci S, Bellando-Randone S, et al. Preliminary analysis of the Very Early Diagnosis of Systemic Sclerosis (VEDOSS) EUSTAR multicentre study: evidence for puffy fingers as pivotal sign for the suspicion of systemic sclerosis. *Ann Rheum Dis*. 2013. doi:[10.1136/annrheumdis-2013-203716](https://doi.org/10.1136/annrheumdis-2013-203716) [Epub ahead of print].
171. Bruni, Serena Guiducci, Silvia Bellando-Randone, Gemma Lepri, Francesca Braschi, Ginevra Fiori, Francesca Bartoli, Francesca Peruzzi, Jelena Blagojevic and Marco Matucci-Cerinic. Concise report Digital ulcers as a sentinel sign for early internal organ involvement in very early systemic sclerosis. *Cosimo* doi:[10.1093/rheumatology/keu29](https://doi.org/10.1093/rheumatology/keu29).
172. White B, Bauer EA, Goldsmith LA, et al. Guidelines for clinical trials in systemic sclerosis (scleroderma). I. Disease-modifying interventions. The American College of Rheumatology Committee on Design and Outcomes in Clinical Trials in Systemic Sclerosis. *Arthritis Rheum*. 1995;38:351–60.