

Pathophysiologic Role of Autophagy in Human Airways

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Abstract Lung diseases are among the most common and widespread disorders worldwide. They refer to many different pathological conditions affecting the pulmonary system in acute or chronic forms, such as asthma, chronic obstructive pulmonary disease, infections, cystic fibrosis, lung cancer and many other breath complications. Environmental, epigenetic and genetic co-factors are responsible for these pathologies that can lead to respiratory failure, and, even, ultimately death. Increasing evidences have highlighted the implication of the autophagic pathways in the pathogenesis of lung diseases and, in some cases, the deregulated molecular mechanisms underlying autophagy may be considered as potential new therapeutic targets. This chapter summarizes recent advances in understanding the pathophysiological functions of autophagy and its possible roles in the causation and/or prevention of human lung diseases.

Abbreviations

AAT	Alpha-1-antitrypsin
AATD	Alpha-1-antitrypsin deficiency
ALI	Acute lung injury

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ALT-E	Alternaria-associated asthma
ARDS	Acute respiratory distress syndrome
Atg	Autophagy-related
ATP	Adenosine triphosphate
Bcl-2	B-cell lymphoma 2
BMP	Bone morphogenetic protein
BMPR2	BMP receptor type-II
BRAF	B-Raf proto-oncogene
CAV-1	Caveolin-1
CD274	Cluster of differentiation 274 (known as Programmed death-ligand 1, PD-L1 or B7 homolog 1, B7-H1)
CF	Cystic Fibrosis
CFTR	Cystic Fibrosis Transmembrane Conductance Regulator
COPD	Chronic obstructive pulmonary disease
CRC	Murine colorectal carcinoma
CS	Cigarette smoke
ECM	Extracellular matrix
EGFR	Epidermal growth factor receptor
Egr-1	Early growth response protein 1
EMT	Epithelial-to-mesenchymal transition
ER	Endoplasmic Reticulum
F508del-CFTR	Deletion of phenylalanine in position 508 of the CFTR
FEV1	Forced expiratory volume in 1 second
FF	Fibroblastic foci
FMD	Myofibroblast differentiation
FoxO3	Forkhead box O3
FOXP3	Forkhead box P3
H ₂ O ₂	Hydrogen peroxide
HDAC6	Histone deacetylase 6
HH	Hedgehog
HO-1	Heme oxygenase-1
IFN	Interferon
IFT20	Intraflagellar transport protein 20 homolog
IL	Interleukin
ILD	Interstitial lung disease
IPF	Idiopathic pulmonary fibrosis
KRAS	Kirsten rat sarcoma viral oncogene homolog
LC3 (MAP1LC3)	Microtubule-associated protein 1 light chain 3*
LPS	Lipopolysaccharide
MCC	Mucociliary clearance
MMP	Matrix metalloproteinases
mTOR	Mammalian target of rapamycin
MUC5AC	Mucin 5AC
MyD88	Myeloid differentiation primary response gene 88
NK	Natural killer

NO	Nitric oxide
NSCLC	Human non-small cell lung carcinoma
OFD1	Oral facial digital syndrome
p62/SQSTM1	Sequestosome 1
PAH	Pulmonary arterial hypertension
PARK2	Parkin RBR E3 ubiquitin protein ligase
PASMCs	Pulmonary artery smooth muscle cells
PH	Pulmonary hypertension
PI3K	Class III-phosphoinositide 3-kinase
PINK	PTEN-induced putative kinase
PTEN	Phosphatase and tensin homolog
ROS	Reactive oxygen species
Rtp801	Known as Redd1 (regulated in development and DNA damage responses 1)
SIRT6	Sirtuin 6
SNPs	Single Nucleotide Polymorphisms
STK11 (LKB1)	Serine/threonine kinase 11
TFEB	Transcription factor EB
TG2	Transglutaminase type 2
TGF- β 1	Transforming growth factor- β 1
Th	T helper
TLR4	Toll-like receptor 4
TSC	Tuberous sclerosis complex
WHO	World Health Organization
α -SMA	Smooth muscle- α actin

1 Introduction

Lung diseases are some of the most common medical conditions in the world. The lung has the principal aim to mediate gas exchange [60]. For this reason, the lung can be subjected to several insults, belonging to the environment (inspiration of foreign matter, particles, smoke), reactive oxygen species (ROS) production, biological origins (e.g., viruses, bacteria), changes in O₂ tension, and mechanical stresses (e.g., mechanical ventilation). It is possible to discriminate between diseases affecting: (I) the airways (asthma, chronic obstructive pulmonary disease, chronic bronchitis, emphysema, acute bronchitis and cystic fibrosis); (II) the interstitium (sarcoidosis, idiopathic pulmonary fibrosis, autoimmune diseases, pneumonias and pulmonary edemas); (III) the blood vessels (pulmonary embolism and hypertension); the pleura (pleural effusion, pneumothorax and mesothelioma); (IV) the chest wall (obesity hypoventilation syndrome and neuromuscular disorders). The development of lung diseases can be associated to both acute and chronic exposure to such insults. However, in most conditions, a favouring genetic is necessary [60]. Yet, the lung has

various inducible defence mechanisms to protect itself. First, constitutive and inducible stress protein and antioxidant defences; second, innate immune responses; third, pro- and anti-apoptotic mechanisms [84, 85, 103]. Several studies have recently pinpointed the emerging role of macroautophagy (more often and hereby referred to as autophagy) in lung homeostasis and diseases. Autophagy is a catabolic process that involves the sequential sequestration of cytoplasmic material within double-membraned vesicles (autophagosomes), the fusion of autophagosomes with lysosomes, and the degradation of autophagosomal cargoes (as well as of structural autophagosomal components) by lysosomal hydrolases [26]. Autophagy is mediated by a genetically encoded, evolutionary conserved machinery that is connected to most, if not all, major biochemical processes of the cell, including core metabolic circuitries as well as signal transduction pathways initiated by plasma membrane receptors [18]. Basically, autophagy responds to three major organismal needs: (1) it preserves cellular homeostasis in physiological conditions; (2) it plays a key role in cellular adaptation to stressful stimuli; and (3) it participates in the communication of states of the danger to the whole organism [21]. Indeed, autophagy continuously operates to mediate the disposal of potentially dangerous structures that may otherwise accumulate in the cytoplasm as a consequence of normal cellular activities, like old (and damaged) organelles or protein aggregates [64]. Moreover, the autophagic flux is highly responsive to situations in which intracellular or extracellular homeostasis is perturbed, which generally involves either an increased offer of autophagic substrates (as it occurs in the course of viral infection) or an increased need for autophagic functions or products (as it occurs in response to nutrient deprivation) [90]. In both these settings, proficient autophagic responses are required for the optimal adaptation of cells to stress, as demonstrated in experiments involving pharmacological inhibitors of autophagy or the depletion of essential components of the autophagic machinery [46]. Finally, autophagy is required for cells experiencing so-called “oncogenic stress” (i.e., the boost of cellular functions driven by activating mutations in one oncogene or loss-of-function mutation in one tumor suppressor gene) to become senescent (a cell-intrinsic oncosuppressive mechanism) while secreting immunostimulatory cytokines and expressing on their surface ligands for activatory natural killer (NK)-cell receptors (hence triggering a cell-extrinsic mechanism of tumor suppression) [55]. Along similar lines, cancer cells succumbing to a peculiar form of apoptosis known as “immunogenic cell death” are able to recruit antigen-presenting cells and hence trigger an adaptive immune response only if they secrete ATP as they die, a process that requires proficient autophagic responses [42, 45]. It should be noted that autophagy has also been causally implicated in some instances of cell death, especially in lower organisms like *Drosophila melanogaster* [13, 17]. However, in mammals autophagy mainly mediates robust cytoprotective functions, and – when cellular homeostasis is irremediably compromised – contributes to the maintenance of organismal homeostasis by playing a role in danger signalling. In line with this notion, defects in the autophagic machinery have been associated with a wide panel of human pathologies, including (but not limited to) malignant diseases, neurodegenerative disorders, as well as cardiovascular, renal and pulmonary conditions [86]. An accurate description of the autophagy pathway and its role in

immunity and inflammation has been provided in several previous chapters of this book; therefore, here we will focus on the impact of autophagic in the etiology and treatment of human pulmonary diseases.

2 Acute Lung Injury

Acute lung injury (ALI) and the acute respiratory distress syndrome (ARDS) describe clinical syndromes of acute respiratory failure with substantial morbidity and mortality. ALI is characterised by acute inflammation that causes disruption of the lung endothelial and epithelial barriers. The ALI cellular features include loss of alveolar–capillary membrane integrity, excessive transepithelial neutrophil migration, and release of pro-inflammatory, cytotoxic mediators. The treatment of ALI is predominantly based on ventilatory strategies [35]. However, prolonged exposure to high oxygen therapy (hyperoxia) can result in lung injury [7]. Few studies are present in the literature concerning the role of autophagy in ALI, even so these works support the hypothesis that activation of autophagy has a protective role in this disease. It has been demonstrated that prolonged hyperoxia, which causes characteristic lung injury in mice, induced the increase of LC3II expression. Moreover, in pulmonary epithelial cells, the genetic depletion of LC3 sensitizes the cells to hyperoxia-induced cell death suggesting that LC3 activation confers cytoprotection in oxygen-dependent cytotoxicity [93]. Besides, the involvement of mitophagy has also been identified. The ability to resist hyperoxia is proportional to PTEN-induced putative kinase 1 (PINK1) expression. In fact, the *Pink1*^{-/-} mice were more susceptible to hyperoxia when compared to wild-type mice. Furthermore, genetic deletion of PINK1 or PINK1 silencing in the lung endothelium cells increased susceptibility to hyperoxia *via* alterations in autophagy/mitophagy, proteasome activation, apoptosis and oxidant generation [108].

3 Chronic Obstructive Pulmonary Disease (COPD)

Chronic obstructive pulmonary disease (COPD) is a chronic inflammatory lung disease that causes breathing difficulty, cough, sputum production and dyspnoea. Emphysema and chronic bronchitis can contribute to COPD development. Emphysema is a condition resulting from a severe damage of air sacs (the alveoli). Chronic bronchitis is due to inflammation of the lining of the bronchial tubes. The lung damage that leads to COPD is caused by long-term exposure to irritating gases or particulate matter, most often from cigarette smoke (CS), air pollution or workplace exposure to dust, smoke or fumes. However, a genetic susceptibility to the disease should be considered as an important cofactor. Patients with COPD present increased risk of developing other pathologies, such as heart disease or lung cancer [53]. Multiple molecular mechanisms, not fully understood, participate to the

COPD evolution and, among others, the involvement of the autophagic pathway has been pointed out [3, 86]. In lung tissue from COPD patients, an increase of autophagic vacuoles as well as several autophagy markers (LC3, ATG4, ATG5/12, ATG7) expression has been detected [8]. These evidences are perhaps a result of defective autophagic flux. To corroborate this hypothesis, an increased accumulation of p62 and ubiquitinated proteins and a decreased expression levels of sirtuin 6 (SIRT6) have been evaluated in lung homogenates from COPD patients [92]. Kuwano and colleagues hypothesize that the insufficient autophagic clearance is involved in the accelerated cell senescence observed in COPD [16, 92]. The CS induces mitochondrial damage, accompanied by increased ROS production *in vitro*. The CS-induced mitophagy was inhibited by PINK1 and PARK2 knockdown, resulting in enhanced mitochondrial ROS production. Moreover, a decreased expression of PARK2 in COPD lungs compared with non-COPD lungs has been detected, suggesting that insufficient mitophagy is a part of the pathogenic sequence and cellular senescence of COPD [32]. In addition, a defective xenophagy has been observed in alveolar macrophages of smokers, suggesting that the deregulation of this selective process may contribute to recurrent infections [65]. In contrast, other findings indicate that autophagy has an opposite role in COPD favouring the pathological environment. It has been shown that Rtp801 (also known as Redd1) expression is increased in human emphysematous lungs and in lungs of mice exposed to CS, whereas Rtp801 knockout mice were protected against acute CS-induced lung injury. Rtp801 inhibits mammalian target of rapamycin (mTOR), by stabilizing the TSC1-TSC2 inhibitory complex. The inhibition of mTOR is linked to autophagy induction, but Rtp801 expression enhances oxidative stress-dependent cell death, amplifying the development of CS-induced lung injury [105]. Furthermore, the higher expression of autophagy proteins has been linked to lung epithelial cell death, airway dysfunction and emphysema in response to CS. Genetic depletion of LC3B *in vivo* (*Map1lc3B*^{-/-} mice) suppressed cell death and emphysematous airspace enlargement during chronic CS exposure compared to the wild type mice [9]. More recently, the same group demonstrated that mitophagy regulates necroptosis, which contributes to the COPD pathogenesis. Mice deficient for *Pink1* were protected against mitochondrial dysfunction, airspace enlargement and mucociliary clearance (MCC) disruption during CS exposure [63]. Interestingly, they identified the contribution of a novel selective autophagy-dependent pathway that regulates cilia length, “ciliophagy”, in the COPD pathophysiological evolution. Exposure to CS reduced cilia length and autophagy-impaired (*Beclin 1*^{+/-} or *Map1lc3B*^{-/-}) mice resisted to the CS-induced cilia shortening *via* a mechanism involving histone deacetylase 6 (HDAC6) [48]. Accordingly, it has been shown that autophagy negatively regulate ciliogenesis by the degradation of the essential ciliary protein IFT20 [70]. Conversely, Hedgehog (HH) signalling from primary cilia promotes autophagy [70] and autophagy promotes ciliogenesis by degrading OFD1 (oral facial digital syndrome) at centriolar satellites [95]. Further studies are necessary to clarify the dual relationship between these processes [101]. In conclusion, these studies illustrate that the contribution of autophagy in COPD pathophysiology is complex and show a context-specific role depending on the cell type and tissue as well as on the different stimuli involved.

4 Interstitial Lung Disease (ILD)

Interstitial lung disease (ILD) is a general category that includes all lung diseases affecting the interstitium, the tissue and space that extends throughout both lungs. Among them the most common are Sarcoidosis and Idiopathic pulmonary fibrosis (IPF). Sarcoidosis is a systemic inflammatory disease caused by persistent reaction toward a stimulus (virus or antigens) that continues even when it is physiologically cleared from the body. Lung interstitium fibrosis is the first symptom in patients with Sarcoidosis. Conversely, IPF is characterized by specific fibrosis at interstitial level due to the increased extracellular matrix (ECM) protein deposition and hyper activation of myofibroblasts [10].

Recently, reduced LC3II expression and p62 accumulation has been found in lung tissue from IPF patients [72]. The reduced expression of the transcription factor FoxO3a in IPF fibroblasts could be the cause for the reduction in the levels of LC3 protein as the expression of this latter is positively stimulated by FoxO3a [30].

Furthermore, in fibroblast of IPF patients, decreased expression in Beclin-1 protein and increased expression of the anti-apoptotic protein Bcl-2 have been found, confirming a defect in the autophagy pathway at different level [81]. Moreover, fibroblastic foci (FF), that are the starting point for fibrogenesis, are enriched in ubiquitinated proteins and p62, confirming the insufficient autophagy at the basis of IPF pathogenesis [3].

Autophagy inhibition is able to induce acceleration of epithelial cell senescence and fibroblast to myofibroblast differentiation (FMD), which have a critical role in IPF development [3]. Transforming growth factor- β 1 (TGF- β 1) is one of the essential mediators of fibrosis since it stimulates fibroblasts to produce fibronectin and the smooth muscle- α actin (α -SMA), which is a myofibroblast marker. Autophagy has been associated to fibrosis through TGF- β 1. In fact, genetic deletion of LC3 or Beclin 1 increases TGF- β 1 activity as well as *in vivo* treatment with Rapamycin can protect from fibrosis [72]. TGF- β 1 expression seems to be dependent on IL-17A, a proinflammatory cytokine involved in chronic inflammation and autoimmune disease. Blocking IL-17A might reduce the progression of fibrosis promoting the autophagic degradation of collagen [61].

Recently, lacking of matrix metalloproteinases-19 (MMP-19) has been associated with exacerbated fibrosis in the hyperplastic alveolar epithelium of IPF lungs [106]. Additionally, MMP-19-deficient mice exhibit diminished Atg4c protein expression, demonstrating a direct correlation between these two pathways [33]. Similar evidences from an independent group corroborate the role of autophagy in promoting FMD. In fact, Atg4b-deficient mice exhibited reduction in autophagic activity in lungs, collagen accumulation and increased protein levels of the myofibroblast biomarker α -SMA [6].

Pharmacological treatment with the alkaloid Barberine has been proposed for IPF monitoring because of its capacity to inhibit the activation of mTOR and to increase the expression of LC3 and Beclin 1 in an bleomycin *in vivo* model of airway-fibrosis [11]. Furthermore, the multiple tyrosine kinase inhibitor Nintedanib

has recently been approved for the treatment of IPS for its anti-fibrotic effect. It has been shown that Nintedanib is able to reduce the expression of ECM proteins, fibronectin and collagen as well as to induce a Beclin 1 dependent, ATG7 independent autophagy [76].

5 Asthma

Asthma is a chronic respiratory disease affecting 300 million people worldwide. Asthma manifests through several symptoms including wheezing, breathlessness, and chest tightness. Asthmatic airways are characterized by chronic inflammation, eosinophil infiltration, epithelial fibrosis, mucus hyperproduction, and goblet cell hyperplasia [20].

It is considered as chronic allergic inflammatory disease, mostly mediated by a Th2 response, but an initial Th1-type immune response seems to be the trigger for the subsequent Th2-type response [82]. Thus, Th2 hyperactivation leads to persistent airway inflammation and the occurring of asthma phenotype [38].

Emerging evidences suggest that activation of autophagy is associated with reduced lung function in asthmatic patients. In particular electron microscopy analysis of fibroblast and epithelial cells from asthmatic patients showed increased autophagic hallmarks “such as double membrane autophagosomes” compared to healthy patients [75]. Unfortunately, at present, the role of autophagy in asthma is still unclear.

A recent study demonstrated that two Single Nucleotide Polymorphisms (SNPs), namely rs12201458 and rs510432 were associated with childhood asthma. In particular rs510432 localises at the promoter of ATG5 gene and could increase its expression in nasal epithelium of acute asthmatics compared to stable asthmatics and non-asthmatic patients [58]. Another intronic SNP variant (rs12212740) in ATG5 gene was also shown to be associated with pre-bronchodilator forced expiratory volume in 1 second (FEV1) in asthmatic patients [75].

ATG5 is an essential player in the initiation of autophagy, but its role in asthma pathogenesis is controversial. On one hand ATG5 could help viral elimination through the activation of Xenophagy, and on the other hand it negatively regulates the antiviral properties of type I interferon (IFN) inhibiting innate anti-virus immune responses [36, 90]. Together with these findings, lungs from conditional *Atg7* knockout mice manifest hyper-responsiveness to cholinergic stimuli, which is a common sign of asthma and chronic inflammatory diseases [31]. Asthma severity has been directly correlated with the level of autophagic response in the sputum granulocytes, peripheral blood cells and peripheral blood eosinophils of severe and non-severe asthmatic patients [5].

Autophagy is also involved in the maintenance of intracellular ROS homeostasis, and it has been well established that oxidative stress is associated with asthma so that exhaled levels of hydrogen peroxide (H₂O₂) and nitric oxide (NO) are currently used as predictors of asthma severity [68].

Chronic asthma is characterized by excessive ECM deposition and proliferation of myofibroblasts, leading to fibrosis in the airway wall [79]. The accumulation of fibrotic tissue is mostly due to the production of collagen A1 and fibronectin by the primary human airway smooth muscle through a mechanism autophagy-dependent that involves the TGF β 1. This response is reverted by the silencing of the major key autophagy-inducing gene Atg5 and Atg7 [104].

As already mentioned, asthma is a pathology mostly driven by Th2-type cytokines. Among them, IL-13 is extensively produced in activated CD4⁺ Th2 lymphocytes and is overexpressed in the airway epithelium of asthmatic patients [47]. Here, IL-13 is thought to be responsible for epithelial hypertrophy, mucus hypersecretion, adventitial fibrosis and goblet cell hyperplasia [111]. It directly induces hypersecretion of mucin 5AC, oligomeric mucus/gel forming (MUC5AC) in airway epithelial cell and oxidant stress through a mechanism that is autophagy-dependent, as demonstrated *in vitro* by depletion of ATG5 or ATG14 in primary human tracheal bronchial epithelial cells [15].

Autophagy might be involved in the pathophysiology of Alternaria (ALT-E)-associated asthma. ALT-E is an outdoor allergen able to activate autophagy, which in turn stimulates epithelial cells to release IL-18 [67]. This latter when produced is able to stimulate Th2 differentiation from naïve CD4⁺ T-cells and IFN- γ production by Th1 cells. IL-18 level in serum of asthmatic patients might reflect the degree of disease exacerbation [94].

6 Cystic Fibrosis (CF)

Cystic Fibrosis (CF) is one of the most common lethal genetic diseases in Caucasian population. It is an autosomal recessive disease caused by mutation in the Cystic Fibrosis Transmembrane Conductance Regulator (CFTR) gene. Approximately 1 out of 20 Caucasians are carriers for mutation in this gene. Up to date over 2000 types of different mutations have been discovered and classified according to the degree of functional CFTR protein (<http://www.genet.sickkids.on.ca/StatisticsPage.html>; [27]). Among these, the most common one is the F508del-CFTR. Approximately 90% of CF patients have at least one F508del-CFTR allele, and about 70% are homozygous for it.

The CFTR channel is located at the apical surface of epithelial cells and it is deputized to move out Cl⁻ from the cell. Na⁺ passes through the membranes passively, increasing the movement of water by osmosis. Loss of functional CFTR expression is thought to alter this homeostatic balance through the epithelial layer, leading to net volume depletion of mucus, increased viscosity, and ineffective bacterial clearance [43, 78]. Recurrent pulmonary infections in turn induce an increased inflammatory response and signalling, thus starting a vicious cycle of mucus retention, infection, and inflammation. Since the CFTR is localized in many organs, CF symptoms could go from malabsorption at pancreatic level and gastrointestinal obstruction to male infertility and liver disease. Nevertheless, the main cause of

death remains persistent and untreatable pulmonary *Pseudomonas aeruginosa* infection.

Several recent studies have demonstrated an impairment of autophagy in CF. In fact, in epithelial cells, mutated/unfunctional CFTR causes increased ROS production with consequent increase in tissue transglutaminase type 2 (TG2) levels. TG2, in turn, leads to crosslinking of several targets including Beclin 1 [54, 57]. Beclin 1 interactome displaces from the Endoplasmic Reticulum (ER) leading to the sequestration of class III-phosphoinositide 3-kinase (PI3K) complex, accumulation of p62 with consequent inhibition of autophagosomes formation. The resulting accumulation of aggresomes leads to proteasome overload and may promote the accumulation of mutated CFTR in intracellular aggregates [54]. Restoration of Beclin 1 activity, depletion of p62 by genetic manipulation or treatment with autophagy-stimulatory proteostasis regulators, such as cystamine, functionally rescue the CFTR mutated protein at the apical surface of epithelial cells both *in vitro* and *in vivo* [54].

Heme oxygenases are enzymes involved in the catabolism of the heme ring to generate carbon monoxide, biliverdin-IX α , and ferrous iron. The inducible isoform Heme oxygenase-1 (HO-1) is activated in response to stress such as oxidative stress, hypoxia, heavy metals exposure and cytokines. HO-1, together with its enzymatic products, is able to inhibit apoptosis and related cell death pathways, conferring tissue protection in case of lung or vascular injury [66]. HO-1 could represent the link between CF and impaired autophagy since its expression is increased in human bronchial CF cells. This increase has been associated either to the reduction of apoptosis/injury during *P. aeruginosa* challenge either to the expression of inflammatory mediators [109]. Other evidences suggesting the cytoprotective role of HO-1 in CF showed that Lipopolysaccharide (LPS)-challenged CF macrophages fail to compartmentalize HO-1 to the cell surface and this mechanism seems to be dependent on the reduction in Caveolin-1 (CAV-1) expression [107]. In fact, when HO-1 localises at the plasma membrane, is able to form a complex with CAV-1, which in turn binds and detaches MyD88 from its complex with TLR4 thus terminating the cell death signal [99].

Autophagic clearance of bacteria (so-called Xenophagy) could also be impaired in case of disease, inducing increased bacterial infection that is one of the most frequent injuries in CF patients [90]. In fact it has been demonstrated that *Burkholderia cenocepacia* has the capacity to survive in F508del-CFTR macrophages since immediately after the engulfment, the bacteria resides on LC3-positive vacuoles that appear as arrested autophagosomes [98]. This capacity is directly correlated to the levels of p62, so that its depletion leads not only to a decreased bacterial survival in macrophages but also to the release of Beclin 1 from aggresomes allowing its recruitment to the *B. cenocepacia* vacuole and bacterial clearance via autophagy [2]. *B. cenocepacia* represents a serious threat for CF patients since the infection results in persistent lung inflammation and the bacteria are resistant to most of all available antibiotics [1].

Similar findings showed that pharmacological or molecular inhibition of autophagy reduces the clearance of intracellular *Pseudomonas aeruginosa* *in vitro* [37].

Treatment of CF mice with the mTOR inhibitor Rapamycin decreases bacterial burden in the lungs and drastically reduces signs of lung inflammation [1].

In a normal situation, autophagy can help not only removing polyubiquitinated protein but also controlling bacteria clearance; for these reasons novel strategies aimed at restoring autophagy are emerging as promising therapeutic approaches for CF patients [56].

7 Alpha-1-Antitrypsin Deficiency (AATD)

AATD is a hereditary disorder characterized by a low serum level of alpha-1-antitrypsin (AAT), a 52 kDa serine protease inhibitor, member of the serpin family [29]. AAT is essentially synthesized in the liver and secreted into the bloodstream, where it controls tissue degradation by the enzyme neutrophil elastase. The deficiency in AAT is associated with liver and lung disease due to the loss of anti-inflammatory and antiproteolytic functions. The majority of patients with AAT deficiency are homozygotes for a missense mutation (“PiZ mutation”: lysine replaces glutamic acid at position 342) that alters protein folding. Mutant AAT molecules polymerize and aggregate in the ER of hepatocytes, forming large intrahepatocytic globules, the characteristic features of this disease. The proteasome is responsible for degrading the soluble form of AAT by means of ER-associated degradation while autophagy is involved in disposal of insoluble AAT polymers and aggregates [74]. In fact, a significant accumulation of autophagic vacuoles was found *in vitro* and *in vivo* in liver cells from AATD patients as well as in PiZ mouse model [96, 97]. Whereas in absence of autophagy the degradation of AAT was retarded [39]. Moreover, it has been demonstrated that the stimulation of autophagy by carbamazepine or rapamycin treatment or by liver-directed gene transfer of transcription factor EB (TFEB), a gene regulating lysosomal function and autophagy [89], reduce the hepatic amount of AAT as well as the hepatic fibrosis in mice expressing mutant AAT [28, 41, 71]. Although these results should be corroborated, altogether indicate that autophagy exerts a protective role in AATD and open a real possibility to treat AATD with pro-autophagic molecules.

8 Pulmonary Hypertension (PH)

Pulmonary hypertension (PH) was first identified in 1891 by Ernst von Romberg. PH is a severe and progressive disease that consists in increased blood pressure of lung vasculature and, often, can be a complication of chronic lung disease [88].

Since 2008 the pathology has been classified, by the World Health Organization (WHO), in five groups on the basis of mechanisms underlying the pathogenesis of the multiple types of PH.

The role of autophagy in pulmonary hypertension has mainly been described in correlation with pulmonary arterial hypertension (PAH), WHO Group I.

Little is known about the aetiology of PH, one of the most frequent genetic mutations causing idiopathic inherited form of PH is found in the gene encoding bone morphogenetic protein (BMP) receptor type-II (BMPR2).

In PAH, the pulmonary artery smooth muscle cells (PASMCs) proliferate excessively and are resistant to apoptosis. Chloroquine, a known inhibitor of autophagy flux, has been described as a drug preventing experimental PAH progression. The induction of PAH, by monocrotaline, in rat is associated with increased autophagy and decreased BMPR2 protein expression. The inhibition of autophagy by chloroquine ameliorates the level of BMPR2, inhibits the proliferation and stimulates apoptosis of rat PASMCs [52]. A recent publication [50] confirms that the inhibition of autophagy, by overexpressing mTOR, is a promising therapeutic strategy against PAH.

However, the role of autophagy in PH is still unclear and controversial, in fact, its protective role has been described in the initial phase of the pathogenesis of PH. Histochemical analysis of samples obtained from human PH lungs and mouse exposed to chronic hypoxia, showed an increase in the lipidated form of LC3 and in *Egr-1*, which regulates LC3 expression. Moreover, *LC3^{-/-}* or *Egr-1^{-/-}*, but not *Beclin 1^{+/-}* mice are more susceptible to PH and *in vitro* LC3 knockdown cells showed an increase of hypoxic cell proliferation, suggesting a role for LC3 in the adaptation during vascular remodelling under hypoxia [49].

9 Autophagy in the Etiology of Lung Cancer

In most organs, including the lung, autophagy robustly counteracts malignant transformation, *i.e.*, the conversion of a healthy cell into a (pre-)neoplastic cell, and several mechanisms related to the ability of autophagy to preserve cellular or organismal homeostasis account for such a pronounced oncosuppressive activity [19]. Indeed, besides being required for oncogene-induced senescence and anticancer immunosurveillance (see above) [112], autophagy promotes the maintenance of genomic integrity by multiple mechanisms [25]. First, it mediates the degradation of damaged mitochondria, which are prone to overproduce genotoxic ROS and other redox active entities of endogenous and exogenous origin [22]. Second, proficient autophagic responses appear to be required for optimal DNA damage responses [59]. Third, autophagy is involved in the disposal of potentially oncogenic retrotransposons and micronuclei [80]. Moreover, autophagy generally mediates anti-inflammatory effects, and chronic inflammation is known to accelerate oncogenesis (at least in some tissues, including the lung) [14]. Finally, it has been proposed that autophagy is required for the preservation of normal tissue architecture, in particular at the level of the stem-cell compartment [23]. Although little is known on the deregulation of stem cells in pulmonary carcinogenesis, it cannot be excluded that autophagic defects may promote malignant transformation in the lung also via this mechanism [69]. Conversely, the ability of autophagy to preserve genomic and redox homeostasis seems very relevant in the context of lung tumorigenesis, which in a significant

proportion of cases is associated with tobacco smoking or exposure to environmental nanoparticles like asbestos crystals [65]. Indeed, the oncogenic effects of both smoking and asbestos have been linked to their ability to cause ROS overgeneration along with genetic/genomic defects and chronic inflammatory responses [12]. All these effects are limited, at least to some extent, by proficient autophagic responses.

Irrespective of the precise mechanisms whereby autophagy counteracts malignant transformation in the lung, various genetic interventions aimed at specifically disabling autophagy in the lungs have been shown to promote malignant transformation driven by several oncogenes, including mutated B-Raf proto-oncogene, serine/threonine kinase (*BRAF*) [91], epidermal growth factor receptor (*EGFR*) [100], Kirsten rat sarcoma viral oncogene homolog (*KRAS*) [24, 77]. Intriguingly enough, in one of these models, accelerated oncogenesis caused by the lung-specific inactivation of *ATG5* was linked to increased tumor-infiltration by immunosuppressive $CD4^+CD25^+FOXP3^+$ regulatory T cells [77]. Moreover, the concomitant bi-allelic inactivation of serine/threonine kinase 11 (*STK11*, best known as *LKB1*) and phosphatase and tensin homolog (*PTEN*), two tumor suppressor genes that inhibit autophagy [34, 87], has been shown to cause the formation of pulmonary squamous cell carcinomas that express high levels of the immunosuppressive molecule *CD274* (best known as *PD-L1*) [102]. These latter observations strongly corroborate the notion that autophagy mediates not only cell-intrinsic, but also cell-extrinsic oncosuppression.

9.1 Autophagy in the Progression of Lung Cancer

The capacity of autophagy to preserve cellular homeostasis is beneficial to healthy cells, but also beneficial to transformed cells. This implies that autophagy often (but not always) promotes tumor progression, i.e., the growth and evolution of a transformed cells into an ever more malignant cancer [62]. Indeed, malignant cells are often exposed to relatively adverse microenvironmental conditions, including a shortage of nutrients and oxygen (especially in poorly vascularized tumor areas), and autophagy is instrumental for these cells (as it is for their non-transformed counterparts) to cope with stress and proliferate. Along similar lines, the ability of autophagy to preserve stemness is beneficial for the host when it preserves normal tissue architecture, but detrimental when it sustains the malignant stem-cell compartment. Finally, autophagy supports the survival of malignant cells in key step of tumor progression, the so-called “epithelial-to-mesenchymal transition” (EMT). In this context, epithelial cancer cells “initially growing *in situ*” physically detach from ECM and become able to colonize surrounding tissues as well as distant organs. The EMT is required for all malignancies to become locally and distantly invasive, and critically relies on proficient autophagic responses [4]. In the presence of autophagic defects or pharmacological inhibitors of autophagy, indeed, malignant cells undergoing the EMT and detaching from the ECM, succumb to a form of regulated cell death often referred to as “anoikis” [73].

Corroborating these observations, the genetic and/or pharmacological inhibition of the autophagic machinery in established tumors has been shown to accelerate disease progression in various models of pulmonary oncogenesis, including (but not limited to) *BRAF*- and *KRAS*-driven tumorigenesis [24, 77, 91].

9.2 Autophagy in the Treatment of Lung Cancer

Autophagy provides malignant cells with an increased resistance to various perturbations of homeostasis, including the lack of nutrient and oxygen that cancer cells normally experience in poorly vascularized tumor areas, as well as the presence of xenobiotics like chemotherapeutic agents and physical stress conditions like irradiation. An abundant amount of literature demonstrates indeed that chemical inhibitors of autophagy as well as genetic interventions that compromise autophagic responses accelerate (rather than inhibit) the demise of malignant cells exposed to a wide panel of chemotherapeutics or to irradiation, both *in vitro* and *in vivo*. These observations provided a strong rationale to the development of combinatorial therapeutic strategies involving chemo- or radiotherapy given in combination with an inhibitor of autophagy [19].

Clinical grade highly specific chemical inhibitors of autophagy, however, have not yet been developed, and currently available molecules that can be used in the clinic, like chloroquine (a widely employed antimalarial agent) often operate as lysosomal inhibitors, i.e., they target several processes other than autophagy [83]. Moreover, concerns have been raised that inhibiting autophagy at the whole-body level may *de facto* favor malignant transformation in healthy tissues, reflecting the prominent oncosuppressive functions of autophagy in physiological conditions [51]. Finally, recent data highlight the differential role of autophagy in cancer therapy in immunocompromised *versus* immunocompetent hosts [44]. In this setting, the response to radiotherapy of human non-small cell lung carcinoma (NSCLC) or murine colorectal carcinoma (CRC) cells xenografted in nude mice was significantly improved when cells were rendered autophagy-deficient by the stable depletion of ATG5 or Beclin 1 [44]. However, when murine CRC cells were implanted in immunocompetent syngeneic mice, the stable knockdown of ATG5 compromised the therapeutic activity of irradiation, a defect that could be restored (at least in part) by the intratumoral administration of a chemical inhibitor of extracellular ATPases [44]. These findings demonstrate that inhibiting autophagy in immunocompetent hosts may prevent the elicitation of a therapeutically relevant immune response against dying cancer cells.

In summary, although autophagy generally (but not always) promote the progression of pulmonary malignancies and increases the resistance of lung cancer cells to chemo- and radiotherapeutic regimens, additional experiments are required to understand whether combinatorial treatments involving autophagy inhibitors constitute a clinically viable approach against pulmonary neoplasms. Similarly, further work is needed to clarify whether biomarkers of autophagy such as the expres-

sion levels of Beclin 1 or the lipidation of LC3 have a positive or negative prognostic/predictive value in patients with lung cancer, as preliminary results are rather controversial [40, 110].

10 Conclusions

Abundant evidences indicate that autophagy actively participates in a wide range of cellular responses to both physiologic- and pathologic-related events in the diverse tissues and cell types that constitute the lung system. Nevertheless, much is yet to be learnt about its biological relevance, functional targets, and role in development and disease. As described in this chapter, lungs are the first line of defence against several insults and associated diseases are growing both in number and chronicisation. A clear deregulation of the autophagic machinery has been highlighted in most of the lung diseases, suggesting that this process mainly exerts a defensive role. However, in some pathological contexts, it has been reported that the activation of the autophagic process contributes to damage. As a consequence, a detailed knowledge of the molecular mechanisms at the basis of autophagy in lung pathologies is required for the development of novel diagnostic tools and promising therapeutic strategies.

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References

1. Abdulrahman BA, Khweek AA, Akhter A et al (2011) Autophagy stimulation by rapamycin suppresses lung inflammation and infection by Burkholderia cenocepacia in a model of cystic fibrosis. *Autophagy* 7:1359–1370
2. Abdulrahman BA, Khweek AA, Akhter A et al (2013) Depletion of the ubiquitin-binding adaptor molecule SQSTM1/p62 from macrophages harboring cfr Δ F508 mutation improves the delivery of Burkholderia cenocepacia to the autophagic machinery. *J Biol Chem* 288: 2049–2058
3. Araya J, Hara H, Kuwano K (2013) Autophagy in the pathogenesis of pulmonary disease. *Intern Med* 52:2295–2303
4. Avivar-Valderas A, Bobrovnikova-Marjon E, Alan Diehl J, Bardeesy N, Debnath J, Aguirre-Ghiso JA (2013) Regulation of autophagy during ECM detachment is linked to a selective inhibition of mTORC1 by PERK. *Oncogene* 32:4932–4940. doi:10.1038/onc.2012.512
5. Ban GY, Pham DL, Trinh HK et al (2015) Autophagy mechanisms in sputum and peripheral blood cells of patients with severe asthma: a new therapeutic target. *Clin Exp Allergy*. doi:10.1111/cea.12585
6. Cabrera S, Maciel M, Herrera I et al (2015) Essential role for the ATG4B protease and autophagy in bleomycin-induced pulmonary fibrosis. *Autophagy* 11:670–684
7. Capellier G, Maupoil V, Boussat S, Laurent E, Neidhardt A (1999) Oxygen toxicity and tolerance. *Minerva Anestesiol* 65:388–392

8. Chen ZH, Kim HP, Sciruba FC, Lee SJ, Feghali-Bostwick C, Stolz DB, Dhir R, Landreneau RJ, Schuchert MJ, Yousem SA, Nakahira K, Pilewski JM, Lee JS, Zhang Y, Ryter SW, Choi AM (2008) Egr-1 regulates autophagy in cigarette smoke-induced chronic obstructive pulmonary disease. *PLoS One* 3, e3316. doi:[10.1371/journal.pone.0003316](https://doi.org/10.1371/journal.pone.0003316)
9. Chen ZH, Lam HC, Jin Y, Kim HP, Cao J, Lee SJ, Ifedigbo E, Parameswaran H, Ryter SW, Choi AM (2010) Autophagy protein microtubule-associated protein 1 light chain-3B (LC3B) activates extrinsic apoptosis during cigarette smoke-induced emphysema. *Proc Natl Acad Sci U S A* 107:18880–18885
10. Cheresch P, Kim SJ, Tulasiram S, Kamp DW (2013) Oxidative stress and pulmonary fibrosis. *Biochim Biophys Acta* 1832:1028–1040
11. Chitra P, Saiprasad G, Manikandan R, Sudhandiran G (2015) Berberine inhibits Smad and non-Smad signaling cascades and enhances autophagy against pulmonary fibrosis. *J Mol Med (Berl)* 93:1015–1031
12. Coussens LM, Zitvogel L, Palucka AK (2013) Neutralizing tumor-promoting chronic inflammation: a magic bullet? *Science* 339:286–291. doi:[10.1126/science.1232227](https://doi.org/10.1126/science.1232227)
13. Denton D, Shrivage B, Simin R, Mills K, Berry DL, Baehrecke EH, Kumar S (2009) Autophagy, not apoptosis, is essential for midgut cell death in *Drosophila*. *Curr Biol* 19:1741–1746
14. Deretic V, Saitoh T, Akira S (2013) Autophagy in infection, inflammation and immunity. *Nat Rev Immunol* 13:722–737. doi:[10.1038/nri3532](https://doi.org/10.1038/nri3532)
15. Dickinson JD, Alevy Y, Malvin NP et al (2015) IL13 activates autophagy to regulate secretion in airway epithelial cells. *Autophagy*. 2016;12(2):397–409
16. Fujii S, Hara H, Araya J, Takasaka N, Kojima J, Ito S, Minagawa S, Yumino Y, Ishikawa T, Numata T, Kawaishi M, Hirano J, Odaka M, Morikawa T, Nishimura S, Nakayama K, Kuwano K (2012) Insufficient autophagy promotes bronchial epithelial cell senescence in chronic obstructive pulmonary disease. *Oncoimmunology* 1:630–641
17. Galluzzi L, Bravo-San Pedro JM, Vitale I et al (2015) Essential versus accessory aspects of cell death: recommendations of the NCCD 2015. *Cell Death Differ* 22:58–73
18. Galluzzi L, Pietrocola F, Levine B, Kroemer G (2014) Metabolic control of autophagy. *Cell* 159:1263–1276
19. Galluzzi L, Pietrocola F, Bravo-San Pedro JM et al (2015) Autophagy in malignant transformation and cancer progression. *EMBO J* 34:856–880. doi:[10.15252/embo.201490784](https://doi.org/10.15252/embo.201490784)
20. Grainge C, Thomas PS, Mak JC, Benton MJ, Lim TK, Ko FW (2016) Asthma and chronic obstructive pulmonary disease. *Respirology*. doi: [10.1111/resp.12771](https://doi.org/10.1111/resp.12771). [Epub ahead of print]
21. Green DR, Levine B (2014) To be or not to be? How selective autophagy and cell death govern cell fate. *Cell* 157:65–75
22. Green DR, Galluzzi L, Kroemer G (2011) Mitochondria and the autophagy-inflammation-cell death axis in organismal aging. *Science* 333:1109–1112. doi:[10.1126/science.1201940](https://doi.org/10.1126/science.1201940)
23. Greim H, Kaden DA, Larson RA, Palermo CM, Rice JM, Ross D, Snyder R (2014) The bone marrow niche, stem cells, and leukemia: impact of drugs, chemicals, and the environment. *Ann N Y Acad Sci* 1310:7–31. doi:[10.1111/nyas.12362](https://doi.org/10.1111/nyas.12362)
24. Guo JY, Karsli-Uzunbas G, Mathew R et al (2013) Autophagy suppresses progression of K-ras-induced lung tumors to oncocytomas and maintains lipid homeostasis. *Genes Dev* 27:1447–1461
25. Hanahan D, Weinberg RA (2011) Hallmarks of cancer: the next generation. *Cell* 144:646–674
26. He C, Klionsky DJ (2009) Regulation mechanisms and signaling pathways of autophagy. *Annu Rev Genet* 43:67–93
27. Herrmann U, Dockter G, Lammert F (2010) Cystic fibrosis-associated liver disease. *Best Pract Res Clin Gastroenterol* 24:585–592
28. Hidvegi T, Ewing M, Hale P, Dippold C, Beckett C, Kemp C, Maurice N, Mukherjee A, Goldbach C, Watkins S, Michalopoulos G, Perlmutter DH (2010) An autophagy-enhancing drug promotes degradation of mutant alpha1-antitrypsin Z and reduces hepatic fibrosis. *Science* 329:229–232
29. Hunt JM, Tuder R (2012) Alpha 1 anti-trypsin: one protein, many functions. *Curr Mol Med* 12:827–835

30. Im J, Hergert P, Nho RS (2015) Reduced FoxO3a expression causes low autophagy in idiopathic pulmonary fibrosis fibroblasts on collagen matrices. *Am J Physiol Lung Cell Mol Physiol* 309:L552–L561
31. Inoue D, Kubo H, Taguchi K et al (2011) Inducible disruption of autophagy in the lung causes airway hyper-responsiveness. *Biochem Biophys Res Commun* 405:13–18
32. Ito S, Araya J, Kurita Y, Kobayashi K, Takasaka N, Yoshida M, Hara H, Minagawa S, Wakui H, Fujii S, Kojima J, Shimizu K, Numata T, Kawaishi M, Odaka M, Morikawa T, Harada T, Nishimura SL, Kaneko Y, Nakayama K, Kuwano K (2015) PARK2-mediated mitophagy is involved in regulation of HBEC senescence in COPD pathogenesis. *Autophagy* 11:547–559
33. Jara P, Calyeca J, Romero Y et al (2015) Matrix metalloproteinase (MMP)-19-deficient fibroblasts display a profibrotic phenotype. *Am J Physiol Lung Cell Mol Physiol* 308:L511–L522
34. Ji H, Ramsey MR, Hayes DN et al (2007) LKB1 modulates lung cancer differentiation and metastasis. *Nature* 448:807–810. doi:10.1038/nature06030
35. Johnson ER, Matthay MA (2010) Acute lung injury: epidemiology, pathogenesis, and treatment. *J Aerosol Med Pulm Drug Deliv* 23:243–252
36. Jounai N, Takeshita F, Kobiyama K et al (2007) The Atg5 Atg12 conjugate associates with innate antiviral immune responses. *Proc Natl Acad Sci U S A* 104:14050–14055
37. Junkins RD, Shen A, Rosen K et al (2013) Autophagy enhances bacterial clearance during *P. aeruginosa* lung infection. *PLoS One* 8, e72263. doi:10.1371/journal.pone.0072263
38. Jyothula SS, Eissa NT (2013) Autophagy and role in asthma. *Curr Opin Pulm Med* 19:30–35
39. Kamimoto T, Shoji S, Hidvegi T, Mizushima N, Umabayashi K, Perlmutter DH, Yoshimori T (2006) Intracellular inclusions containing mutant alpha1-antitrypsin Z are propagated in the absence of autophagic activity. *J Biol Chem* 281:4467–4476
40. Karpathiou G, Sivridis E, Koukourakis MI, Mikroulis D, Bouros D, Froudarakis ME, Giatromanolaki A (2011) Light-chain 3A autophagic activity and prognostic significance in non-small cell lung carcinomas. *Chest* 140:127–134. doi:10.1378/chest.10-1831
41. Kaushal S, Annamali M, Blomenkamp K, Rudnick D, Halloran D, Brunt EM, Teckman JH (2010) Rapamycin reduces intrahepatic alpha-1-antitrypsin mutant Z protein polymers and liver injury in a mouse model. *Exp Biol Med (Maywood)* 235:700–709
42. Kepp O, Senovilla L, Vitale I et al (2014) Consensus guidelines for the detection of immunogenic cell death. *Oncoimmunology* 3, e955691. doi:10.4161/21624011.2014.955691
43. Kerem B, Kerem E (1996) The molecular basis for disease variability in cystic fibrosis. *Eur J Hum Genet* 4:65–73
44. Ko A, Kanehisa A, Martins I et al (2014) Autophagy inhibition radiosensitizes in vitro, yet reduces radioresponses in vivo due to deficient immunogenic signalling. *Cell Death Differ* 21:92–99. doi:10.1038/cdd.2013.124
45. Kroemer G, Galluzzi L, Kepp O, Zitvogel L (2013) Immunogenic cell death in cancer therapy. *Annu Rev Immunol* 31:51–72
46. Kroemer G, Mariño G, Levine B (2010) Autophagy and the integrated stress response. *Mol Cell* 40:280–293
47. Kuperman DA, Huang X, Koth LL et al (2002) Direct effects of interleukin-13 on epithelial cells cause airway hyperreactivity and mucus overproduction in asthma. *Nat Med* 8: 885–889
48. Lam HC, Cloonan SM, Bhashyam AR, Haspel JA, Singh A, Sathirapongsasuti JF, Cervo M, Yao H, Chung AL, Mizumura K, An CH, Shan B, Franks JM, Haley KJ, Owen CA, Tesfaigzi Y, Washko GR, Quackenbush J, Silverman EK, Rahman I, Kim HP, Mahmood A, Biswal SS, Rytter SW, Choi AM (2013) Histone deacetylase 6-mediated selective autophagy regulates COPD-associated cilia dysfunction. *J Clin Invest* 123:5212–5230
49. Lee SJ, Smith A, Guo L, Alastalo TP, Li M, Sawada H et al (2011) Autophagic protein LC3B confers resistance against hypoxia-induced pulmonary hypertension. *Am J Respir Crit Care Med* 183:649–658. doi:10.1164/rccm.201005-0746OC
50. Li L, Wang X, Wang L et al (2015) Mammalian target of rapamycin overexpression antagonizes chronic hypoxia-triggered pulmonary arterial hypertension via the autophagic pathway. *Int J Mol Med* 36:316–322. doi:10.3892/ijmm.2015.2224

51. Liang C, Feng P, Ku B, Dotan I, Canaani D, Oh BH, Jung JU (2006) Autophagic and tumour suppressor activity of a novel Beclin1-binding protein UVRAG. *Nat Cell Biol* 8:688–699
52. Long L, Yang X, Southwood M, Lu J, Marciniak SJ, Dunmore BJ, Morrell NW (2013) Chloroquine prevents progression of experimental pulmonary hypertension via inhibition of autophagy and lysosomal bone morphogenetic protein type II receptor degradation. *Circ Res* 112:1159–1170
53. López-Campos JL, Tan W, Soriano JB (2015) Global burden of COPD. *Respiology*. doi:[10.1111/resp.12660](https://doi.org/10.1111/resp.12660)
54. Luciani A, Vilella VR, Esposito S et al (2010) Defective CFTR induces aggresome formation and lung inflammation in cystic fibrosis through ROS-mediated autophagy inhibition. *Nat Cell Biol* 12:863–875
55. Ma Y, Galluzzi L, Zitvogel L, Kroemer G (2013) Autophagy and cellular immune responses. *Immunity* 39:211–227
56. Maiuri L, De Stefano D, Raia V, Kroemer G (2015) The holy grail of cystic fibrosis research: pharmacological repair of the F508del-CFTR mutation. *Ann Transl Med* 3:S24. doi:[10.3978/j.issn.2305-5839.2015.02.32](https://doi.org/10.3978/j.issn.2305-5839.2015.02.32)
57. Maiuri L, Luciani A, Giardino I et al (2008) Tissue transglutaminase activation modulates inflammation in cystic fibrosis via PPARgamma down-regulation. *J Immunol* 180:7697–7705
58. Martin LJ, Gupta J, Jyothula SS et al (2012) Functional variant in the autophagy-related 5 gene promoter is associated with childhood asthma. *PLoS One* 7, e33454. doi:[10.1371/journal.pone.0033454](https://doi.org/10.1371/journal.pone.0033454)
59. Mathew R, Kongara S, Beaudoin B et al (2007) Autophagy suppresses tumor progression by limiting chromosomal instability. *Genes Dev* 21:1367–1381. doi:[10.1101/gad.1545107](https://doi.org/10.1101/gad.1545107)
60. Menzel DB, Amdur MO (1986) Toxic response of the respiratory system. In: Klaassen K, Amdur MO, Doull J (eds) *Casarett and Doull's toxicology: the basic science of poisons*, 3rd edn. Macmillan, New York, pp 330–358
61. Mi S, Li Z, Yang HZ et al (2011) Blocking IL-17A promotes the resolution of pulmonary inflammation and fibrosis via TGF-beta1-dependent and -independent mechanisms. *J Immunol* 187:3003–3014
62. Mihaylova MM, Shaw RJ (2011) The AMPK signalling pathway coordinates cell growth, autophagy and metabolism. *Nat Cell Biol* 13:1016–1023. doi:[10.1038/ncb2329](https://doi.org/10.1038/ncb2329)
63. Mizumura K, Cloonan SM, Nakahira K, Bhashyam AR, Cervo M, Kitada T, Glass K, Owen CA, Mahmood A, Washko GR, Hashimoto S, Ryter SW, Choi AM (2014) Mitophagy-dependent necroptosis contributes to the pathogenesis of COPD. *J Clin Invest* 124:3987–4003
64. Mizushima N, Komatsu M (2011) Autophagy: renovation of cells and tissues. *Cell* 147:728–741
65. Monick MM, Powers LS, Walters K, Lovan N, Zhang M, Gerke A, Hansdottir S, Hunninghake GW (2010) Identification of an autophagy defect in smokers' alveolar macrophages. *J Immunol* 185:5425–5435
66. Morse D, Lin L, Choi AM, Ryter SW (2009) Heme oxygenase-1, a critical arbitrator of cell death pathways in lung injury and disease. *Free Radic Biol Med* 47:1–12
67. Murai H, Okazaki S, Hayashi H et al (2015) *Alternaria* extract activates autophagy that induces IL-18 release from airway epithelial cells. *Biochem Biophys Res Commun* 464:969–974
68. Murata K, Fujimoto K, Kitaguchi Y et al (2014) Hydrogen peroxide content and pH of expired breath condensate from patients with asthma and COPD. *COPD* 11:81–87
69. Ojha R, Bhattacharyya S, Singh SK (2015) Autophagy in cancer stem cells: a potential link between chemoresistance, recurrence, and metastasis. *Biores Open Access* 4:97–108
70. Pampliega O, Orhon I, Patel B, Sridhar S, Díaz-Carretero A, Beau I, Codogno P, Satir BH, Satir P, Cuervo AM (2013) Functional interaction between autophagy and ciliogenesis. *Nature* 502:194–200
71. Pastore N, Blomenkamp K, Annunziata F et al (2013) Gene transfer of master autophagy regulator TFEB results in clearance of toxic protein and correction of hepatic disease in alpha-1-anti-trypsin deficiency. *EMBO Mol Med* 5:397–412

72. Patel AS, Lin L, Geyer A et al (2012) Autophagy in idiopathic pulmonary fibrosis. *PLoS One* 7, e41394. doi:[10.1371/journal.pone.0041394](https://doi.org/10.1371/journal.pone.0041394)
73. Peng YF, Shi YH, Ding ZB et al (2013) Autophagy inhibition suppresses pulmonary metastasis of HCC in mice via impairing anoikis resistance and colonization of HCC cells. *Autophagy* 9:2056–2068. doi:[10.4161/autophagy.26398](https://doi.org/10.4161/autophagy.26398)
74. Perlmutter DH (2009) Autophagic disposal of the aggregation-prone protein that causes liver inflammation and carcinogenesis in α -1-antitrypsin deficiency. *Cell Death Differ* 16:39–45
75. Poon AH, Chouiali F, Tse SM et al (2012) Genetic and histologic evidence for autophagy in asthma pathogenesis. *J Allergy Clin Immunol* 129:569–571
76. Rangarajan S, Kurundkar A, Kurundkar D et al (2015) Novel mechanisms for the anti-fibrotic action of nintedanib. *Am J Respir Cell Mol Biol*. 2016;54(1):51–9. doi: [10.1165/rmb.2014-0445OC](https://doi.org/10.1165/rmb.2014-0445OC)
77. Rao S, Tortola L, Perlot T et al (2014) A dual role for autophagy in a murine model of lung cancer. *Nat Commun* 5:3056. doi:[10.1038/ncomms4056](https://doi.org/10.1038/ncomms4056)
78. Ratjen F, Döring G (2003) Cystic fibrosis. *Lancet* 361:681–689
79. Redington AE (2000) Airway fibrosis in asthma: mechanisms, consequences, and potential for therapeutic intervention. *Monaldi Arch Chest Dis* 55:317–323
80. Rello-Varona S, Lissa D, Shen S et al (2012) Autophagic removal of micronuclei. *Cell Cycle* 11:170–176. doi:[10.4161/cc.11.1.18564](https://doi.org/10.4161/cc.11.1.18564)
81. Ricci A, Cherubini E, Scozzi D et al (2013) Decreased expression of autophagic beclin 1 protein in idiopathic pulmonary fibrosis fibroblasts. *J Cell Physiol* 228:1516–1524
82. Robinson DS, Hamid Q, Ying S et al (1992) Predominant Th2-like bronchoalveolar T-lymphocyte population in atopic asthma. *N Engl J Med* 326:298–304
83. Rubinsztein DC, Codogno P, Levine B (2012) Autophagy modulation as a potential therapeutic target for diverse diseases. *Nat Rev Drug Discov* 11:709–730. doi:[10.1038/nrd3802](https://doi.org/10.1038/nrd3802)
84. Ryter SW, Choi AM (2009) Heme oxygenase-1/carbon monoxide: from metabolism to molecular therapy. *Am J Respir Cell Mol Biol* 41:251–260
85. Ryter SW, Choi AMK (2010) Autophagy in the lung. *Proc Am Thorac Soc* 7:13–21
86. Ryter SW, Choi AMK (2015) Autophagy in lung disease pathogenesis and therapeutics. *Redox Biol* 4:215–225
87. Sansal I, Sellers WR (2004) The biology and clinical relevance of the PTEN tumor suppressor pathway. *J Clin Oncol* 22:2954–2963. doi:[10.1200/JCO.2004.02.141](https://doi.org/10.1200/JCO.2004.02.141)
88. Semenza GL (2011) Oxygen sensing, homeostasis, and disease. *N Engl J Med* 365:537–547
89. Settembre C, Di Malta C, Polito VA, Garcia Arencibia M et al (2011) TFEB links autophagy to lysosomal biogenesis. *Science* 332:1429–1433
90. Sica V, Galluzzi L, Bravo-San Pedro JM et al (2015) Organelle-specific initiation of autophagy. *Mol Cell* 59:522–539
91. Strohecker AM, Guo JY, Karsli-Uzunbas G et al (2013) Autophagy sustains mitochondrial glutamine metabolism and growth of BrafV600E-driven lung tumors. *Cancer Discov* 3:1272–1285. doi:[10.1158/2159-8290.CD-13-0397](https://doi.org/10.1158/2159-8290.CD-13-0397)
92. Takasaka N, Araya J, Hara H, Ito S, Kobayashi K, Kurita Y, Wakui H, Yoshii Y, Yumino Y, Fujii S, Minagawa S, Tsurushige C, Kojima J, Numata T, Shimizu K, Kawaiishi M, Kaneko Y, Kamiya N, Hirano J, Odaka M, Morikawa T, Nishimura SL, Nakayama K, Kuwano K (2014) Autophagy induction by SIRT6 through attenuation of insulin-like growth factor signaling is involved in the regulation of human bronchial epithelial cell senescence. *J Immunol* 192:958–968
93. Tanaka A, Jin Y, Lee SJ, Zhang M, Kim HP, Stolz DB, Ryter SW, Choi AM (2012) Hyperoxia-induced LC3B interacts with the Fas apoptotic pathway in epithelial cell death. *Am J Respir Cell Mol Biol* 46:507–514
94. Tanaka H, Miyazaki N, Oashi K et al (2001) IL-18 might reflect disease activity in mild and moderate asthma exacerbation. *J Allergy Clin Immunol* 107:331–336
95. Tang Z, Lin MG, Stowe TR, Chen S, Zhu M, Stearns T, Franco B, Zhong Q (2013) Autophagy promotes primary ciliogenesis by removing OFD1 from centriolar satellites. *Nature* 502:254–257

96. Teckman JH, An JK, Loethen S, Perlmutter DH (2002) Fasting in alpha1-antitrypsin deficient liver: constitutive activation of autophagy. *Am J Physiol* 283:G1156–G1165
97. Teckman JH, Perlmutter DH (2000) Retention of mutant alpha1-antitrypsin Z in endoplasmic reticulum is associated with an autophagic response. *Am J Physiol* 279:G961–G974
98. Valvano MA (2015) Intracellular survival of Burkholderia cepacia complex in phagocytic cells. *Can J Microbiol* 61:607–615
99. Wang XM, Kim HP, Nakahira K et al (2009) The heme oxygenase-1/carbon monoxide pathway suppresses TLR4 signaling by regulating the interaction of TLR4 with caveolin-1. *J Immunol* 182:3809–3818
100. Wei Y, Zou Z, Becker N et al (2013) EGFR-mediated Beclin 1 phosphorylation in autophagy suppression, tumor progression, and tumor chemoresistance. *Cell* 154:1269–1284
101. Wrighton KH (2013) Cytoskeleton: autophagy and ciliogenesis come together. *Nat Rev Mol Cell Biol* 14:687. doi:10.1038/nrm3686
102. Xu C, Fillmore CM, Koyama S et al (2014) Loss of Lkb1 and Pten leads to lung squamous cell carcinoma with elevated PD-L1 expression. *Cancer Cell* 25:590–604
103. Yao H, Rahman I (2011) Current concepts on oxidative/carbonyl stress, inflammation and epigenetics in pathogenesis of chronic obstructive pulmonary disease. *Toxicol Appl Pharmacol* 254:72–85
104. Yeganeh B, Serebrin A, Mutawe MM et al (2011) Autophagy regulates TGF-Beta1 induced fibrosis in human airway smooth muscle cells. *Am J Respir Crit Care Med* 183:A2110
105. Yoshida T, Mett I, Bhunia AK, Bowman J, Perez M, Zhang L, Gandjeva A, Zhen L, Chukwueke U, Mao T, Richter A, Brown E, Ashush H, Notkin N, Gelfand A, Thimmulappa RK, Rangasamy T, Sussan T, Cosgrove G, Mouded M, Shapiro SD, Petrache I, Biswal S, Feinstein E, Tudor RM (2010) Rtp801, a suppressor of mTOR signaling, is an essential mediator of cigarette smoke-induced pulmonary injury and emphysema. *Nat Med* 16:767–773
106. Yu G, Kovkarova-Naumovski E, Jara P et al (2012) Matrix metalloproteinase-19 is a key regulator of lung fibrosis in mice and humans. *Am J Respir Crit Care Med* 186:752–762
107. Zhang PX, Murray TS, Villella VR et al (2013) Reduced caveolin-1 promotes hyperinflammation due to abnormal heme oxygenase-1 localization in lipopolysaccharide-challenged macrophages with dysfunctional cystic fibrosis transmembrane conductance regulator. *J Immunol* 190:5196–5206
108. Zhang Y, Sauler M, Shinn AS, Gong H, Haslip M, Shan P, Mannam P, Lee PJ (2014) Endothelial PINK1 mediates the protective effects of NLRP3 deficiency during lethal oxidant injury. *J Immunol* 192:5296–5304. doi:10.4049/jimmunol.1400653
109. Zhou H, Lu F, Latham C, Zander DS, Visner GA (2004) Heme oxygenase-1 expression in human lungs with cystic fibrosis and cytoprotective effects against *Pseudomonas aeruginosa* in vitro. *Am J Respir Crit Care Med* 170:633–640
110. Zhou W, Yue C, Deng J et al (2013) Autophagic protein Beclin 1 serves as an independent positive prognostic biomarker for non-small cell lung cancer. *PLoS One* 8, e80338. doi:10.1371/journal.pone.0080338
111. Zhu Z, Homer RJ, Wang Z et al (1999) Pulmonary expression of interleukin-13 causes inflammation, mucus hypersecretion, subepithelial fibrosis, physiologic abnormalities, and eotaxin production. *J Clin Invest* 103:779–788
112. Zitvogel L, Galluzzi L, Smyth MJ, Kroemer G (2013) Mechanism of action of conventional and targeted anticancer therapies: reinstating immunosurveillance. *Immunity* 39:74–88