Chapter 1 A Short History of Chromosome Rearrangements and Gene Fusions in Cancer

Felix Mitelman

Contents

 Abstract The molecular characterization of recurrent chromosome aberrations in the early 1980s laid the foundation for gene fusion detection in cancer. This approach remained the unrivalled method to identify fusion genes for a quarter of a century and led to the detection of more than 700 neoplasia-associated fusion genes. The advancement of deep sequencing in the mid-2000s revolutionized the search for cytogenetically undetectable fusions, and such studies have dramatically changed the gene fusion landscape. A myriad of new gene fusions – more than 1,300 – the great majority involving previously unsuspected genes, have been identified by sequencing-based analyses during the past 10 years.

 Keywords Cytogenetics • Karyotype • Chromosome aberrations • Gene fusions • **Oncogenes**

1.1 Introduction

 One hundred years ago, Theodor Boveri in his famous book *Zur Frage der Entstehung maligner Tumoren* [1] proposed an idea that later became known as the somatic mutation theory of cancer, which essentially states that cancer originates in a single cell by a mitotic disturbance leading to chromosomal damage. The acquired genetic change is then propagated during subsequent mitoses to all descendants of

J.D. Rowley et al. (eds.), *Chromosomal Translocations and Genome Rearrangements in Cancer*, DOI 10.1007/978-3-319-19983-2_1

F. Mitelman (\boxtimes)

Department of Clinical Genetics, BMC C13, University of Lund, SE-221 84 Lund, Sweden e-mail: felix.mitelman@med.lu.se

[©] Springer International Publishing Switzerland 2015 3

the originally transformed cell. This concept is today the paradigmatic view of cancer pathogenesis, supported by a wealth of experimental evidence. It long remained a theoretical idea, however, which could not be examined critically until technical improvements in human cytogenetic analysis were made half a century later, culminating in the description of the normal human chromosome complement by Tjio and Levan in 1956 $[2]$.

The discovery only 4 years later by Nowell and Hungerford [3] of an acquired characteristic marker chromosome consistently seen in patients with chronic myelogenous leukaemia (CML), later designated the Philadelphia chromosome (Ph) after the city where it had been found, immediately provided strong support for the idea that chromosome aberrations indeed may play a major role in the initiation of the carcinogenic process. It was reasonable to assume that the specific chromosomal abnormality $-$ a perfect example of a somatic mutation in a haematopoietic stem cell – was the direct cause of the neoplastic state, i.e., the true verification of Boveri's somatic mutation theory. The discovery of the Ph chromosome greatly stimulated interest in cancer cytogenetics in the 1960s. However, the results obtained over the next decade were disappointing. Chromosome aberrations were detected in most tumours but no specific change comparable to the Ph was found. The abnormalities varied within the same tumour types and among patients, and at the end of the 1960s most scientists agreed that chromosome aberrations were secondary epiphenomena – not the cause, but the consequence, of neoplasia. The Ph was the exception to the rule that chromosome changes did not play any important pathogenetic role in carcinogenesis.

1.2 Chromosome Banding

 The situation changed dramatically in 1970 with the introduction of chromosome banding by Caspersson and co-workers [4]. Each chromosome, chromosome arm, and even chromosome region could now be precisely identified on the basis of its unique banding pattern, and hence aberrations that previously had not been possible to detect could now be visualized. The first characteristic cytogenetic changes in cancer cells discovered with the help of the new technique appeared in 1972 (see Mitelman and Heim [5] for a review of the early data): a $14q +$ marker chromosome in Burkitt lymphoma (BL), a deletion of the long arm of a chromosome 20 in polycythemia vera, +8 in acute myeloid leukaemia (AML), and −22 in meningiomas. The first balanced rearrangements were reported shortly afterwards. In 1973, Rowley first identified a reciprocal translocation between chromosomes 8 and 21, i.e., $t(8:21)(q22:q22)$, in the bone marrow cells of a patient with AML [6] and the very same year she showed that the Ph in CML originated through a $t(9;22)$ (q34;q11), not a deletion of the long arm of chromosome 22 as previously thought [7]. A steadily increasing number of characteristic, specific, sometimes even pathognomonic balanced rearrangements, in particular translocations, were soon described in various haematologic disorders and malignant lymphomas, including t(8;14)

5

 $(q24;q32)$, $t(2;8)(p11;q24)$, and $t(8;22)(q24;q11)$ in BL $[8-11]$, $t(15;17)(q22;q21)$ in acute promyelocytic leukaemia $[12]$, $t(4;11)(q21;q23)$ in acute lymphoblastic leukaemia $[13]$, t $(8:16)(p11:p13)$ in acute monocytic leukaemia $[14]$, and t $(14:18)$ $(q32;q21)$ in follicular lymphoma [15]. The first specific translocations in experimental neoplasms and, as it turned out, the perfect equivalents of the characteristic rearrangements in human BL were identified by Ohno et al. [16] in mouse plasmacytomas (MPC) by the end of the 1970s.

 The following decade saw a similar explosion of data emerging from studies of solid tumours, initially in particular among mesenchymal tumours. Several of the aberrations identified in the solid tumours were as specific as those previously found among haematologic malignancies, e.g., t(2;13)(q36;q14) in alveolar rhabdomyosarcoma [17], $t(11;22)(q24;q12)$ in Ewing sarcoma [18, 19], and $t(12;16)(q13;p11)$ in myxoid liposarcoma $[20]$. At this time, it also became clear that many benign tumours carried characteristic aberrations, including reciprocal translocations, e.g., t(3;8)(p21;q12) in salivary gland adenoma [21], t(3;12)(q27–28;q13–15) in lipoma [$22, 23$], and t($12;14$)(q $14;q24$) in uterine leiomyoma [$24-26$]. All published abnormal karyotypes in neoplasia detected by banding analyses are presented in Mitelman et al. [[27 \]](#page-7-0), and a comprehensive review of the presently known recurrent and specific chromosome aberrations may be found in Heim and Mitelman [28].

1.3 Recombinant DNA Technology

Technical developments in the late 1970s enabling the identification and characterization of genes in the breakpoints of chromosome rearrangements made it possible to elucidate the molecular consequences of the recurrent cancer-associated chromosome changes, and analyses in the early 1980s of the specific translocations in MPC, BL, and CML proved particularly pivotal for our understanding of how chromosome aberrations contribute to neoplastic transformation. When the different pieces of the puzzle were assembled, it became apparent that balanced rearrangements exert their effects by one of two mechanisms: Transcriptional up-regulation of an oncogene in one of the breakpoints through exchange of regulatory sequences in the other breakpoint, and the creation of a hybrid gene through fusion of parts of two genes, one in each breakpoint [29]. Deregulation of an oncogene by juxtaposition to a constitutively active gene region was predicted by Klein already in 1981 [30] and the principle was soon demonstrated in MPC and human BL. The breakpoints of the characteristic translocations in mice and humans were found to be located within or close to the *MYC* oncogene and one of the immunoglobulin heavyor light-chain genes (*IGH*, *IGK* or *IGL*). As a consequence of the translocations, the entire coding part of *MYC* is juxtaposed to one of the immunoglobulin genes, resulting in deregulation of *MYC* because the gene is now driven by regulatory elements of the immunoglobulin genes. The alternative mechanism – the creation of a fusion gene – was documented at the same time in CML with the demonstration that the Ph chromosome, i.e., the $der(22)t(9;22)(q34;q11)$, contains a fusion in which the 3'

part of the *ABL1* oncogene from 9q34 has become juxtaposed with the 5′ part of a gene from 22q11 called the *BCR* gene, resulting in the creation of an in-frame *BCR*/*ABL1* fusion transcript.

The first confirmation of the BL scenario in another B-cell neoplasm was the demonstration in 1984 that the t(14;18)(q32;q21) in follicular lymphoma results in overexpression of *BCL2* [31] due to its juxtaposition to the *IGH* locus, and in 1986 an analogous situation was established in T-cell acute lymphoblastic leukaemia in which regulatory elements of the T-cell receptor alpha *(TRA)* gene were found to deregulate the expression of *MYC* [32–34]; other 3' partner genes, e.g., *LYL1*, *TAL1*, *LMO1* ,and *LMO2* , involved in translocations involving *TRB* and *TRD* loci were soon identified in T-cell leukaemias/lymphomas carrying various translocations $[35-39]$. The CML scenario, i.e., the creation of a chimeric fusion gene, was firmly established in both haematologic malignancies and solid tumours in the early 1990s: *PML*/*RARA* in acute promyelocytic leukaemia with $t(15;17)(q22;q21)$ [40, 41], *RET/CCDC6* in thyroid carcinomas with inv(10)(q11q21) [42], *DEK/NUP214* in AML with t(6;9)(p22;q34) [43], *RUNX1|RUNX1T1* in AML with t(8;21)(q22;q22) [44], and *EWSR1* / *FLI1* in Ewing sarcoma with $t(11;22)(q24;q12)$ [45].

The molecular insights into the pathogenetic mechanisms of cancer-specific chromosome aberrations sparked an enormous interest in cancer cytogenetics as a powerful tool to locate and identify genes important in tumourigenesis. Further technical improvements during the 1980s, in particular the development of fluorescence in situ hybridization (FISH), multi-colour FISH, and the widespread adoption of the polymerase chain reaction (PCR), added a further sophistication to the analysis, and radically increased the precision in identifying new gene fusions [28]. This course of action – the genomic characterization of the breakpoints in cytogenetically detected specific balanced aberrations – remained the unrivalled method to identify fusion genes in cancer for a quarter of a century and led to the detection of more than 700 fusion genes (Table 1.1) caused by acquired translocations, inversions, and insertions characterizing various tumour entities [27].

	Haematologic disorders,	Solid tumours		
Year	including malignant lymphomas	Mesenchymal tumours	Epithelial tumours	Total ^a
1980-1989	19	Ω	Ω	19
1990-1994	55	8	6	69
1995-1999	101	20	18	140
2000–2004	162	43	20	220
2005-2009	247	41	109	394
2010-2014	379	245	975	1,598
Total ^a	674	299	1,080	2.038

 Table 1.1 Gene fusions in neoplasia reported 1980–2014, based on data contained in Mitelman et al. [27]

a The total numbers do not add up because each gene fusion is only counted once but may be found in distinct tumour entities

7

 There was a major limitation of this remarkably successful approach, however. It was in principle restricted to haematological malignancies and mesenchymal tumours, which typically have simple abnormalities, often seen as a sole anomaly. Malignant epithelial tumours, representing the dominant cause of human cancer morbidity and mortality, which characteristically have complex karyotypes with numerous numerical and structural abnormalities, were consequently not amenable for analysis. As a consequence, very few fusion genes were detected in carcinomas. By 2005, only 29 fusion genes were known in carcinomas, all organs combined, as compared to 56 in mesenchymal neoplasms and 272 in haematological malignancies. These quantitative differences led to the generally held view that fusion genes are not an important mechanism in carcinoma pathogenesis. Indirect evidence that fusion genes actually may play the same fundamental role in epithelial carcinogenesis as they do for the initiation of haematologic and mesenchymal neoplasms was presented by Mitelman et al. [46], and direct evidence clearly substantiating this view was soon produced with the help of new powerful technologies developed during the last decade.

1.4 Next-Generation Sequencing

 The breakthrough in the search for fusion genes by alternative methods to chromosome banding analysis followed by reverse transcriptase-PCR and Sanger sequencing was made by Chinnaiyan and coworkers in 2005 [\[47](#page-8-0)]. They took a bioinformatics approach to look for genes in prostate cancer that showed a very high expression in RNA microarray experiments, and demonstrated that two of the outlier genes – ERG and $ETVI$ – were frequently fused to the 5['] part of the prostate-specific androgen- regulated gene *TMPRSS2* . Subsequently other *ETS* family genes were found to be fused with *TMPRSS2* , and several other 5′ partner genes that activate *ETS* genes were also discovered [48]. The frequencies of the various fusions vary slightly in different patient series depending on the populations studied but altogether about 80 % of prostate cancers harbour one of the presently known fusion genes, the most common being *TMPRSS2/ERG*. Very soon afterwards, an *EML4* /*ALK* gene fusion was found in a subset of non-small cell lung cancer by screening a retroviral cDNA expression library from cancer samples [49]. The importance of these results in prostate and lung cancer cannot be overestimated. They showed, for the first time, that cytogenetically undetectable gene fusions may be a causative factor in a substantial fraction of common human cancers, and the findings underscored the need for high-resolution methods to be used in parallel with chromosome banding to characterize cancer genomes. The advancement of next-generation sequencing (NGS) at this time revolutionized the search for new fusion genes, enabling unprecedented opportunities to process thousands of tumours for systematic mutation and fusion gene discovery without any knowledge of the genetic constitution. The first report using the new sequencing technology to find fusion genes in cancer was presented by Stratton and co-workers in 2008 [50].

Numerous studies of common cancer types, such as carcinomas of the breast, lung, prostate, and uterus, quickly followed (e.g., $[51-57]$), and the results have dramatically changed the gene fusion landscape. A myriad of new gene fusions – more than 1,300 – the great majority involving previously unsuspected genes, have been identified with the help of NGS-based analysis $[27]$. Table [1.1](#page-3-0) shows the dramatic increase of gene fusions detected since 2010, in particular among malignant epithelial tumours, and Table 1.2 presents the distribution of all presently reported fusions among major neoplasia subtypes.

As can be seen from Table [1.1](#page-3-0), the total number of gene fusions now exceeds 2,000 and at least 65 $\%$ of these were identified by various sequencing technologies during the last 5 years. Clearly, the presently known gene fusions represent only the tip of an iceberg. Given the extraordinary rate at which The Cancer Genome Atlas (TCGA) project is generating cancer genomic data [[58 ,](#page-8-0) [59 \]](#page-8-0), a huge number of new

Diagnosis	Number of gene fusions	Number of genes involved in fusions			
Haematologic disorders Undifferentiated and biphenotypic leukaemia	24	32			
	267	339			
Acute myeloid leukaemia	50				
Myelodysplastic syndromes		59			
Chronic myeloproliferative disorders, including CML	68	84			
Acute lymphoblastic leukaemia	192	188			
Plasma cell neoplasms	20	23			
Mature B-cell neoplasms	179	195			
Mature T- and NK-cell neoplasms	28	37			
Hodgkin lymphoma	13	19			
Solid tumours					
Benign solid tumours					
Benign epithelial tumours	14	20			
Benign mesenchymal tumours	37	58			
Malignant solid tumours					
Respiratory system	373	596			
Digestive system	62	109			
Breast	343	578			
Female genital organs	95	185			
Male genital organs	142	209			
Urinary tract	55	98			
Endocrine system	22	28			
Nervous system	131	227			
Skin	12	24			
Bone	24	30			
Soft tissues	81	105			

 Table 1.2 Number of gene fusions and genes involved in fusions in major neoplasia subtypes, based on Mitelman et al. [27]

genomic rearrangements can be expected to be discovered within the next few years. It is important in this context to mention two notable differences between the fusion genes detected on the basis of cytogenetically identified aberrations and those so far identified by NGS. First, multiple NGS-detected fusion genes are generally found within the same tumour, e.g., more than 25 different fusions in one prostate cancer, and secondly, very few of the NGS-detected fusion genes have been found to be recurrent. A major challenge will be to verify by functional studies which of the alleged gene fusions are primary, pathogenetically important, and which are either secondary progressional changes or non-consequential "noise" abnormalities.

References

- 1. Boveri T (1914) Zur Frage der Entstehung maligner Tumoren. Gustav Fisher Verlag, Jena
- 2. Tjio JH, Levan A (1956) The chromosome number of man. Hereditas 42:1–6
- 3. Nowell PC, Hungerford DA (1960) A minute chromosome in human chronic granulocytic leukemia. Science 132:1497
- 4. Caspersson T, Zech L, Johansson C (1970) Differential binding of alkylating fluorochromes in human chromosomes. Exp Cell Res 60:315–319
- 5. Mitelman F, Heim S (2015) How it all began: cancer cytogenetics before sequencing. In: Heim S, Mitelman F (eds) Cancer cytogenetics, 4th edn. Wiley, New York
- 6. Rowley JD (1973) Identification of a translocation with quinacrine fluorescence in a patient with acute leukemia. Ann Genet 16:109–112
- 7. Rowley JD (1973) A new consistent chromosomal abnormality in chronic myelogenous leukaemia identified by quinacrine fluorescence and Giemsa staining. Nature 243:290–293
- 8. Zech L, Haglund U, Nilsson K, Klein G (1976) Characteristic chromosomal abnormalities in biopsies and lymphoid-cell lines from patients with Burkitt and non-Burkitt lymphomas. Int J Cancer 17:47–56
- 9. Berger R, Bernheim A, Weh H-J, Flandrin G, Daniel MT et al (1979) A new translocation in Burkitt's tumor cells. Hum Genet 53:111–112
- 10. Miyoshi I, Hiraki S, Kimura I, Miyamoto K, Sato J (1979) 2/8 translocation in a Japanese Burkitt's lymphoma. Experientia 35:742–743
- 11. van den Berghe H, Parloir C, Gosseye S, Englebienne V, Cornu G et al (1979) Variant translocation in Burkitt lymphoma. Cancer Genet Cytogenet 1:9–14
- 12. Rowley JD, Golomb HM, Dougherty C (1977) 15/17 translocation, a consistent chromosomal change in acute promyelocytic leukaemia. Lancet 1:549–550
- 13. Oshimura M, Freeman AI, Sandberg AA (1977) Chromosomes and causation of human cancer and leukemia. XXVI. Banding studies in acute lymphoblastic leukemia (ALL). Cancer 40:1161–1172
- 14. Mitelman F, Brandt L, Nilsson PG (1978) Relation among occupational exposure to potential mutagenic/carcinogenic agents, clinical findings, and bone marrow chromosomes in acute nonlymphocytic leukemia. Blood 52:1229–1237
- 15. Fukuhara S, Rowley JD, Variakojis D, Golomb HM (1979) Chromosome abnormalities in poorly differentiated lymphocytic lymphoma. Cancer Res 39:3119–3128
- 16. Ohno S, Babonits M, Wiener F, Spira J, Klein G et al (1979) Nonrandom chromosome changes involving the Ig gene-carrying chromosomes 12 and 6 in pristane-induced mouse plasmacytomas. Cell 18:1001–1007
- 17. Seidal T, Mark J, Hagmar B, Angervall L (1982) Alveolar rhabdomyosarcoma: a cytogenetic and correlated cytological and histological study. APMIS 90:345–354
- 18. Aurias A, Rimbaut C, Buffe D, Dubousset J, Mazabraud A (1983) Chromosomal translocations in Ewing's sarcoma. N Engl J Med 309:496–497
- 19. Turc-Carel C, Philip I, Berger M-P, Philip T, Lenoir GM (1983) Chromosomal translocations in Ewing's sarcoma. N Engl J Med 309:497–498
- 20. Limon J, Turc-Carel C, Dal Cin P, Rao U, Sandberg AA (1986) Recurrent chromosome translocations in liposarcoma. Cancer Genet Cytogenet 22:93–94
- 21. Mark J, Dahlenfors R, Ekedahl C, Stenman G (1980) The mixed salivary gland tumor a normally benign human neoplasm frequently showing specific chromosomal abnormalities. Cancer Genet Cytogenet 2:231–241
- 22. Heim S, Mandahl N, Kristoffersson U, Mitelman F, Rööser B et al (1986) Reciprocal translocation t(3;12)(q27;q13) in lipoma. Cancer Genet Cytogenet 23:301–304
- 23. Turc-Carel C, Dal Cin P, Rao U, Karakousis C, Sandberg AA (1986) Cytogenetic studies of adipose tissue tumors. I. A benign lipoma with reciprocal translocation $t(3;12)(q28;q14)$. Cancer Genet Cytogenet 23:283–289
- 24. Heim S, Nilbert M, Vanni R, Floderus U-M, Mandahl N et al (1988) A specific translocation, t(12;14)(q14-15;q23-24), characterizes a subgroup of uterine leiomyomas. Cancer Genet Cytogenet 32:13–17
- 25. Mark J, Havel G, Grepp C, Dahlenfors R, Wedell B (1988) Cytogenetical observations in human benign uterine leiomyomas. Anticancer Res 8:621–626
- 26. Turc-Carel C, Dal Cin P, Boghosian L, Terk-Zakarian J, Sandberg AA (1988) Consistent breakpoints in region 14q22-q24 in uterine leiomyoma. Cancer Genet Cytogenet 32:25–31
- 27. Mitelman F, Johansson B, Mertens F (2014) Mitelman database of chromosome aberrations and gene fusions in cancer.<http://cgap.nci.nih.gov/Chromosomes/Mitelman>
- 28. Heim S, Mitelman F (eds) (2015) Cancer cytogenetics, 4th edn. Wiley, New York
- 29. Mitelman F, Johansson B, Mertens F (2007) The impact of translocations and gene fusions on cancer causation. Nat Rev Cancer 7:233–245
- 30. Klein G (1981) The role of gene dosage and genetic transpositions in carcinogenesis. Nature 294:313–318
- 31. Tsujimoto Y, Finger LR, Yunis J, Nowell PC, Croce CM (1984) Cloning of the chromosome breakpoint of neoplastic B cells with the $t(14;18)$ chromosome translocation. Science 226:1097–1099
- 32. Erikson J, Finger L, Sun L, ar-Rushdi A, Nishikura K et al (1986) Deregulation of c-myc by translocation of the alfa-locus of the T-cell receptor in T-cell leukemias. Science 232:884–886
- 33. Mathieu-Mahul D, Sigaux F, Zhu C, Bernheim A, Mauchauffe M et al (1986) A t(8;14) (q24;q11) translocation in a T-cell leukemia (L1-ALL) with c-myc and TcR-alpha chain locus rearrangements. Int J Cancer 38:835–840
- 34. McKeithan TW, Shima EA, Le Beau MM, Minowada J, Rowley JD et al (1986) Molecular cloning of the breakpoint junction of a human chromosomal 8;14 translocation involving the T-cell receptor alpha-chain gene and sequences on the 3′ side of MYC. Proc Natl Acad Sci U S A 83:6636–6640
- 35. Boehm T, Baer R, Lavenir I, Forster A, Waters JJ et al (1988) The mechanism of chromosomal translocation t(11;14) involving the T-cell receptor Cdelta locus on human chromosome 14q11 and a transcribed region of chromosome 11p15. EMBO J 7:385–394
- 36. Mellentin JD, Smith SD, Cleary ML (1989) lyl-1, a novel gene altered by chromosomal translocation in T cell leukemia, codes for a protein with a helix-loop-helix DNA binding motif. Cell 58:77–83
- 37. Finger LR, Kagan J, Christopher G, Kurtzberg J, Hershfield MS et al (1989) Involvement of the TCL5 gene on human chromosome 1 in T-cell leukemia and melanoma. Proc Natl Acad Sci U S A 86:5039–5043
- 38. McGuire EA, Hockett RD, Pollock KM, Bartholdi MF, O'Brien SJ et al (1989) The t(11;14) (p15;q11) in a T-cell acute lymphoblastic leukemia cell line activates multiple transcripts, including Ttg-1, a gene encoding a potential zinc finger protein. Mol Cell Biol 9:2124-2132
- 39. Boehm T, Foroni L, Kaneko Y, Perutz MF, Rabbitts TH (1991) The rhombotin family of cysteine- rich LIM-domain oncogenes: distinct members are involved in T-cell translocations to human chromosomes 11p15 and 11p13. Proc Natl Acad Sci U S A 88:4367–4371
- 40. Lemons RS, Eilender D, Waldmann RA, Rebentisch M, Frej A-K et al (1990) Cloning and characterization of the t(15;17) translocation breakpoint region in acute promyelocytic leukemia. Gene Chromosome Cancer 2:79–87
- 41. de Thé H, Chomienne C, Lanotte M, Degos L, Dejean A (1990) The t(15;17) translocation of acute promyelocytic leukaemia fuses the retinoic acid receptor alpha gene to a novel transcribed locus. Nature 347:558–561
- 42. Pierotti MA, Santoro M, Jenkins RB, Sozzi G, Bongarzone I et al (1992) Characterization of an inversion on the long arm of chromosome 10 juxtaposing D10S170 and RET and creating the oncogenic sequence RET/PTC. Proc Natl Acad Sci U S A 89:1616–1620
- 43. von Lindern M, Fornerod M, van Baal S, Jaegle M, de Wit T et al (1992) The translocation $(6;9)$, associated with a specific subtype of acute myeloid leukemia, results in the fusion of two genes, dek and can, and the expression of a chimeric, leukemia-specific dek-can mRNA. Mol Cell Biol 12:1687–1697
- 44. Erickson P, Gao J, Chang K-S, Look T, Whisenant E et al (1992) Identification of breakpoints in t(8;21) acute myelogenous leukemia and isolation of a fusion transcript, AML1/ETO, with similarity to Drosophila segmentation gene, runt. Blood 80:1825–1831
- 45. Delattre O, Zucman J, Plougastel B, Desmaze C, Melot T et al (1992) Gene fusion with an ETS DNA-binding domain caused by chromosome translocation in human tumours. Nature 359:162–165
- 46. Mitelman F, Johansson B, Mertens F (2004) Fusion genes and rearranged genes as a linear function of chromosome aberrations in cancer. Nat Genet 36:331–334
- 47. Tomlins SA, Rhodes DR, Perner S, Dhanasekaran SM, Mehra R et al (2005) Recurrent fusion of TMPRSS2 and ETS transcription factor genes in prostate cancer. Science 310:644–648
- 48. Rubin MA, Maher CA, Chinnaiyan AM (2011) Common gene rearrangements in prostate cancer. J Clin Oncol 29:3659–3668
- 49. Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y et al (2007) Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. Nature 448:561–566
- 50. Campbell PJ, Stephens PJ, Pleasance ED, O'Meara S, Li H et al (2008) Identification of somatically acquired rearrangements in cancer using genome-wide massively parallel pairedend sequencing. Nat Genet 40:722–729
- 51. Maher CA, Kumar-Sinha C, Cao X, Kalyana-Sundaram S, Han B et al (2009) Transcriptome sequencing to detect gene fusions in cancer. Nature 458:97–101
- 52. Stephens PJ, McBride DJ, Lin M-L, Varela I, Pleasance ED et al (2009) Complex landscapes of somatic rearrangement in human breast cancer genomes. Nature 462:1005–1010
- 53. Robinson DR, Kalyana-Sundaram S, Wu Y-M, Shankar S, Cao X et al (2011) Functionally recurrent rearrangements of the MAST kinase and Notch gene families in breast cancer. Nat Med 17:1646–1651
- 54. Asmann YW, Necela BM, Kalari KR, Hossain A, Baker TR et al (2012) Detection of redundant fusion transcripts as biomarkers or disease-specific therapeutic targets in breast cancer. Cancer Res 72:1921–1928
- 55. Hammerman PS, Hayes DN, Wilkerson MD, Schultz N, Bose R et al (2012) Comprehensive genomic characterization of squamous cell lung cancers. Nature 489:519–525
- 56. Lapuk AV, Wu C, Wyatt AW, McPherson A, McConeghy BJ et al (2012) From sequence to molecular pathology, and a mechanism driving the neuroendocrine phenotype in prostate cancer. J Pathol 227:286–297
- 57. Kandoth C, Schultz N, Cherniack AD, Akbani R, Liu Y et al (2013) Integrated genomic characterization of endometrial carcinoma. Nature 497:67–73
- 58. Kandoth C, McLellan MD, Vandin F, Ye K, Niu B et al (2013) Mutational landscape and significance across 12 major cancer types. Nature 402:333–341
- 59. Weinstein JN, Collisson EA, Mills GB, Mills Shaw KR, Ozenberger BA et al (2013) The cancer genome atlas pan-cancer analysis project. Nat Genet 45:1113–1120