# The Ovary

Debra L. Friedman

### Contents

| 13.1   | Pathophysiology   | 253                      |
|--|---|--------------------------|
| 13.1.1   | Normal Organ Development  | 253                      |
| 13.1.2   | Organ Damage Induced by   |                          |
|  | Cytotoxic Therapy   | 254                      |
| 13.1.3   | Cytotoxic Effects of Radiotherapy   | 254                      |
| 13.1.4   | Cytotoxic Effects of Chemotherapy   | 255                      |
| 13.2   | Clinical Manifestations   | 255                      |
| 13.2.1   | Effects of Radiotherapy on  |                          |
|  | Ovarian Function  | 255                      |
| 13.2.2   | Effects of Chemotherapy on  |                          |
|  | Ovarian Function  | 256                      |
| 13.2.3   | Effects of Radiotherapy and   |                          |
|  | Chemotherapy on Reproductive  |                          |
|  | Outcomes  | 257                      |
| 13.3   | Detection and Screening   | 258                      |
|  |   |                          |
| 13.4   | Management of Established Problems  | 260                      |
| 13.4<br>13.4.1                                 | Management of Established Problems<br>Prevention Strategies   | 260<br>260               |
|  | 0   |                          |
| 13.4.1   | Prevention Strategies   | 260                      |
| 13.4.1<br>13.4.2                               | Prevention Strategies<br>Management of Delayed Puberty  | 260<br>261               |
| 13.4.1<br>13.4.2<br>13.4.3                     | Prevention Strategies<br>Management of Delayed Puberty<br>Management of Infertility   | 260<br>261<br>261        |
| 13.4.1<br>13.4.2<br>13.4.3<br>13.4.4<br>13.4.5 | Prevention Strategies<br>Management of Delayed Puberty<br>Management of Infertility<br>Management of Pregnancy and Delivery | 260<br>261<br>261<br>262 |

D.L. Friedman, MD

Division of Pediatric Hematology/Oncology, Vanderbilt University School of Medicine, 2220 Pierce Avenue 397 PRB, Nashville, TN 37232-6310, USA e-mail: debra.l.friedman@vanderbilt.edu

### 13.1 Pathophysiology

#### 13.1.1 Normal Organ Development

Hormonal function and potential for fertility are synchronous in females, as the ovary both produces oocytes and secretes steroid hormones. Prepubertal females possess their lifetime supply of oocytes with no new oogonia formed after birth. Active mitosis of oogonia occurs during fetal life, reaching a peak of six to seven million by 20 weeks of gestation and then rapidly declining to one to two million at birth. At the onset of puberty, only 300,000 remain [1]. The cortices of the ovaries harbor the follicles within connective tissue. These follicles arise from the germinal epithelium, which covers the free surface of the ovary. Through involution, atresia, and, to a much lesser extent, ovulation, the follicles disappear entirely at menopause.

At initiation of puberty, there is a surge in the production of gonadotropin-releasing hormone (GnRH) by the hypothalamus. GnRH then stimulates release of the gonadotropins by the pituitary gland: follicle-stimulating hormone (FSH), responsible for follicular maturation, and luteinizing hormone (LH), responsible for ovarian luteinization. With menarche, the menstrual cycle occurs approximately every 28 days. Each cycle is marked by an estrogen-dependent midcycle surge of FSH and LH. After ovulation, the corpus luteum forms and produces progesterone,

<sup>©</sup> Springer International Publishing 2015

C.L. Schwartz et al. (eds.), Survivors of Childhood and Adolescent Cancer: A Multidisciplinary Approach, Pediatric Oncology, DOI 10.1007/978-3-319-16435-9\_13

estradiol, and 17-hydroxyprogesterone, as well as the endometrial changes required for implantation of a fetus. In the absence of fertilization, there is no chorionic gonadotropin from a conceptus; the corpus luteum becomes exhausted and progesterone and estrogen falls. At this time, FSH increases and the endometrium sloughs, resulting in menstruation. The normal premenopausal ovary contains degenerating ova and follicles in varying stages of maturity. Another ovarian hormone now being investigated in the field of reproductive endocrinology is anti-Mullerian hormone (AMH) thought to be the most sensitive indicator of ovarian reserve [2].

Ovarian hormones also have critical physiologic effects on other organs and bodily processes, including the stimulation of libido, the maturation and function of the breasts and vagina, bone mineralization, and the integrity of the cardiovascular system. With depletion of oocytes by radiotherapy, chemotherapy, or normal senescence, the ovaries undergo atresia. As a result, menstruation and estrogen production cease, and menopause ensues.

# 13.1.2 Organ Damage Induced by Cytotoxic Therapy

Radiotherapy and chemotherapy each may cause transitory or permanent effects on hormonal function, reproductive capacity, and sexual function. Primary ovarian failure, impaired development of secondary sexual characteristics, menstrual irregularities, including oligomenorrhea and amenorrhea, or premature menopause may occur. The menopausal state, when it occurs prematurely, is associated with the same physical symptoms as are seen with normal aging, including hot flashes, loss of libido, and osteoporosis [3, 4]. Such effects are not simply physically bothersome to survivors, but adversely impact their quality of life [5]. The specific effects are dependent on the ovarian dose of radiation and the chemotherapeutic agents and their doses. They also depend on the developmental status of the patient at the age of treatment.

## 13.1.3 Cytotoxic Effects of Radiotherapy

Radiation causes a decrease in the number of ovarian follicles, impaired follicular maturation, cortical fibrosis and atrophy, generalized hypoplasia, and hyalinization of the capsule. Females treated prior to puberty have a greater number of ova than do older women. Thus, ovarian function is more likely to be preserved after radiotherapy in prepubertal females, compared with postpubertal females [6]. The dose of radiation that will ablate ovarian function depends on the patient's age and, by implication, stage of sexual development, but overall modeling suggests that the dose of radiation required to destroy 50 % of immature oocytes is <2 Gy [7].

Several investigators have provided information regarding the dose of radiotherapy that results in sterility in women of varying ages. Wallace and colleagues reported on 19 adult females treated in childhood with whole abdominal radiotherapy to a total dose of 30 Gy. Using the assumption that the number of oocytes within the ovary declines exponentially by atresia from approximately 2,000,000 at birth to approximately 2,000 at menopause, they were able to estimate that the LD50 (radiation causing ablation in 50 % of patients) for the human oocyte is not greater than 4 Gy [8]. Ash's summary of clinical information on radiation to the human ovary is shown in Table 13.1. Menopause was induced by a dose of 12-15 Gy in women under 40 years of age, whereas women over 40 years of age required only 4-7 Gy for the same clinical effect. Permanent sterility occurred in 60 % of females 15-40 years of age receiving 5-6 Gy [9]. When one considers doses to the ovary after single fractions, temporary sterility can occur with ovarian doses of 1.7-6.4 Gy and permanent sterility after doses of 3.2–10 Gy [10]. Whole

| Minimum<br>ovarian dose<br>(GY) | Effect  |
|---------------------------------|---|
| 0.6                             | None  |
| 1.5                             | No deleterious effect in most young<br>women. Some risk of sterilization<br>especially in women aged >40  |
| 2.5–5.0                         | Variable. Aged 15–40 years: about<br>60 % sterilized permanently, some with<br>temporary amenorrhea. Aged >40:<br>usually 100 % permanently sterile |
| 5–8                             | Variable. Aged 15–40 years: about<br>70 % sterilized permanently; of the<br>remainder, some temporary<br>amenorrhea                                 |
| >8                              | 100 % permanently sterilized  |

 Table 13.1
 Effect of fractionated ovarian X-irradiation

 on ovarian function in women of reproductive age irradiated for malignant or nonmalignant disease

Modified from [7]

No attempt has been made to allow for variation in mode of fractionation

abdomen doses of 20–30 Gy are associated with primary or premature secondary ovarian failure in young females [8, 11].

# 13.1.4 Cytotoxic Effects of Chemotherapy

The effects of chemotherapy on ovarian function are both agent- and dose-dependent, and this effect may be additive to that resulting from abdominopelvic radiotherapy. Alkylating agents affect the resting oocyte in a dose-dependent, cell cycle-independent manner. Thecal cells and ova are depleted, as are the primordial follicles, resulting in arrest of follicular maturation and decreased estrogen secretion. Again, as was the case with radiotherapy, the effects are more pronounced in postpubertal as compared with prepubertal females, due to the fact that postpubertal females have fewer remaining viable oocytes. The effects worsen with age, as the normal aging process is accompanied by an ongoing depletion of oocytes. Risks of menstrual irregularity,

ovarian failure, and infertility increase with age at treatments. Conversely, younger females can tolerate higher doses of alkylating agents without impairment of fertility, compared with adult females [12–17].

## 13.2 Clinical Manifestations

# 13.2.1 Effects of Radiotherapy on Ovarian Function

The clinical relationship between ovarian failure and the dose of radiation to the ovary is well illustrated by Stillman's study of 182 girls treated at less than 17 years of age with 12-15 Gy of abdominal radiotherapy. Overall, primary ovarian failure occurred in 22 girls (12 %). However, ovarian failure was noted in 68 % of the girls whose ovaries received the full irradiation dose, but in only 14 % of those who had at least one ovary at the edge of the abdominal treatment volume (estimated dose 0.9-10 Gy, with a mean of 2.9 Gy). Conversely, none of 34 girls who received an estimated ovarian dose of 0.5–1.5 Gy (mean: 0.54 Gy) to at least one ovary outside the direct treatment volume had ovarian failure. Covariate and multivariate revealed that the location of the ovaries relative to radiation treatment fields was the only risk factor for ovarian failure [18].

In considering the risk of ovarian failure related to radiotherapy, other fields than the abdomen and pelvis must be considered. Direct or scattered irradiation from the spinal component of craniospinal radiotherapy may also produce ovarian damage [8, 19]. With the expanded use of hematopoietic stem cell transplantation in pediatric oncology, it is important to recall that total body irradiation (TBI) utilized in the conditioning regimen is commonly associated with primary ovarian failure or premature menopause, with prevalence rates as high as 90–100 %. Fraction size is of importance as well as the age of the patient at the time of radiotherapy [20–24].

# 13.2.2 Effects of Chemotherapy on Ovarian Function

The dose–response relationship of alkylating agents, and the effect of age, is a recurring theme in studies of fertility following chemotherapy. Amenorrhea and ovarian failure occur more commonly in adult women treated with cyclo-phosphamide and other alkylating agents than with adolescents, with prepubertal females tolerating cumulative cyclophosphamide doses as high as  $25 \text{ g/m}^2$  [13, 25]. In examining protocols with common chemotherapy, 86 % of women >24–30 years have been shown to have ovarian failure, compared with 28–31 % of younger women [3, 25].

It is clear that the sterilizing effects of all alkylating agents are not equal. Mechlorethamine and procarbazine together are perhaps the most damaging of the alkylating agents. These chemotherapy agents were used in the past together for the treatment of Hodgkin lymphoma, often in combination with radiotherapy, resulting in impaired fertility, among other adverse long-term effects [26]. Newer risk-adapted protocols for Hodgkin lymphoma have been developed to avoid mechlorethamine or procarbazine and to limit cumulative doses of other gonadotoxic alkylating agents, without negatively impacting the efficacy of the chemotherapy regimens [27–30].

Newer studies have also been designed to collect long-term follow-up data, and investigators are starting to collect data on the impact of these changes.

In recent years, ifosfamide, a congener of cyclophosphamide, has been used for a variety of solid tumors and lymphoma. The effects of ifosfamide on reproductive function are only beginning to be evaluated. A case report of successful pregnancies in two young women treated with high-dose ifosfamide and cyclophosphamide at Memorial Sloan Kettering was reported in 2001 [31]. In 2008, a small case series, which included 13 females treated for sarcoma with ifosfamide as the only alkylating agent, there was no primary ovarian failure reported. However, AMH levels were lower than an age-matched reference group, suggesting risk for early menopause [32].

Due to improved survivorship from childhood cancer noted as early as the 1970s–1980s, large cohorts of female survivors have reached the third and fourth decades of life, where the risk for infertility and premature menopause has been examined. In this era of treatment for these cohorts, the use of both radiotherapy and chemotherapy together was common, and thus it is not possible to fully separate the effects of the two modalities of therapy.

Two large studies of these survivors demonstrated elevated risks for infertility and premature menopause [11, 33]. A study of 2,498 female survivors, treated between 1945 and 1975, showed a 7 % deficit in fertility, compared with siblings. Between ages 21 and 25 years, survivors had a risk of premature menopause four times greater than that of siblings. Treatment-related risk factors included radiotherapy alone (RR=3.7), alkylating agents alone (RR=9.2), or a combination of both (RR=27). By age 31, 42 % of these women had reached menopause, compared with 5 % of siblings [11]. In a study of 719 survivors treated between 1964 and 1988, 15.5 % of women were unable to conceive. Women treated with abdominopelvic radiotherapy alone had a fertility deficit of 23 %, compared with those treated with surgery. As with the previous study, the risk of infertility and premature menopause increased with increasing dose of abdominopelvic radiotherapy and amount of alkylating agent [33].

Several studies have been conducted within the Childhood Cancer Survivor Study (CCSS), a cohort of survivors treated between 1970 and 1987. In an analysis of 3,390 female survivors, acute ovarian failure was self-reported in 215 (6.3 %). Risk factors in multivariate analysis included increased dose of ovarian radiation, exposure to cyclophosphamide in those 13-20 years of age, and any exposure to procarbazine [34]. Premature menopause was also noted in 8 % of participants studied compared to 0.8 % in a sibling cohort. Risk factors for premature menopause were higher-attained age, increased dose of radiotherapy to the ovaries, increased alkylating agent exposure as determined by the alkylating agent score, and a diagnosis of Hodgkin lymphoma [35]. In another CCSS analysis of 5,149 female participants and 1,441 female siblings, the relative risk for survivors ever being pregnant was 0.81 compared to the siblings. Risk factors in multivariate analysis included ovarian/uterine radiation dose of >5 Gy, hypothalamic/pituitary dose of  $\geq$ 30 Gy, and alkylating agent dose score of 3 or 4 [36].

Similar to what has been done with conventional chemoradiotherapy protocols, transplant conditioning protocols without TBI are being utilized to avoid some of the associated adverse long-term sequelae. The use of high-dose cyclophosphamide without TBI or other alkylating agents is associated with a lower risk of ovarian failure than conditioning regimens with TBI or multiple alkylating agents. In a study by Sanders, 100 % of women (n = 15) younger than age 26 and three of nine older than age 26 who were treated with 200 mg/kg cyclophosphamide recovered normal gonadotropin levels and menstruation posttransplantation [37]. However, many transplant protocols use high doses of alkylating agents together, most commonly busulfan and cyclophosphamide, which are associated with similar degrees of ovarian failure in females as protocols containing TBI [38].

# 13.2.3 Effects of Radiotherapy and Chemotherapy on Reproductive Outcomes

Many survivors of childhood cancer previously treated with cytotoxic therapy will remain fertile, and, therefore, pregnancy outcomes and the risk of cancer or genetic disease in offspring must be addressed. Young women who have been exposed to radiotherapy below the diaphragm are also at risk of impaired uterine development, which can adversely affect pregnancy outcomes, often resulting in premature labor and low-birth-weight infants. The magnitude of the risk is related to the radiotherapy field, total dose, and fractionation schedule. Female longterm survivors treated with total body irradiation and marrow transplantation are at risk for impaired uterine growth and blood flow, and, if pregnancy is achieved, for early pregnancy loss and premature labor. Despite standard hormone replacement, the uterus of the childhood cancer survivor may be impaired in its development and measure only 40 % of normal adult size, the ultimate uterine volume correlating with the age at which radiotherapy was received [7, 20].

With more childhood cancer survivors retaining fertility, pregnancy outcome data is now available. Of 4,029 pregnancies occurring among 1,915 women followed in the Childhood Cancer Survivor Study (CCSS), there were 63 % live births, 1 % stillbirths, 15 % miscarriages, 17 % abortions, and 3 % unknown or in gestation. Risk of miscarriage was 3.6-fold higher in women treated with craniospinal radiotherapy and 1.7-fold higher in those treated with pelvic radiotherapy. Chemotherapy exposure alone did not increase the risk of miscarriage. Compared with siblings, however, survivors were less likely to have live births and more likely to have medical abortions and low-birth-weight babies [39]. In an updated analysis of this cohort, it was noted that offspring of women who receive uterine radiation doses of >5 Gy were more likely to be small for gestational age [35]. In another analysis from the CCSS, Signorello also found that uterine and ovarian radiation doses of >10 Gy increased risk of stillbirth or neonatal death, and furthermore, for girls treated prior to menarche, uterine or ovarian doses as low as 1.0-2.49 Gy increased the risk of stillbirth or neonatal death [40].

In a Danish population-based cohort, in analysis of 34,000 pregnancies, which included 1,479 pregnancies of childhood cancer survivors, there were no significant differences noted in the proportions of live births, stillbirths, or all kinds of abortions combined between survivors and women without cancer. However, survivors had a 23 % increased risk for spontaneous abortion, with ovarian and uterine radiotherapy as the major significant risk factor [41].

In a Finish population-based cohort, in an analysis of 3,501 and 16,908 children of female cancer patients and siblings, respectively, the risk of stillbirth or early neonatal death was not significantly increased among offspring of cancer survivors as compared to offspring of siblings [42]. In a case-cohort study conducted involving 472 Danish survivors of childhood and adolescent cancer and their 1,037 pregnancies, no statistically significant associations were found between genetic disease in children and parental treatment with alkylating drugs or preconception radiation doses to the testes in male and ovaries

in female cancer survivors. A statistically significant association between abdominopelvic irradiation and malformations, stillbirths, and neonatal deaths was not seen in the children of female survivors overall or in the children of mothers receiving high uterine doses [43].

In the National Wilms Tumor Study, records were obtained for 427 pregnancies of >20 weeks duration. In this group, there were 409 single and 12 twin live births. Early or threatened labor, malposition of the fetus, lower-birth-weight (<2,500 g), and premature delivery (<36 weeks) were more frequent among women who had received flank radiotherapy, in a dose-dependent manner [44].

Preservation of fertility and successful pregnancies may occur following HSCT. Sanders and colleagues evaluated pregnancy outcomes in a group of females treated with bone marrow transplant. Among 116 treated before puberty and 23 treated after the onset of puberty who retained ovarian function, 32 (28 %) and 9 (30 %), respectively, became pregnant. Of the 32 pregnancies in those treated with TBI, 16 resulted in early termination, compared with a 21 % prevalence of early termination in those treated with cyclophosphamide alone. There were no pregnancies among the women treated with busulfan and cyclophosphamide [37].

For childhood cancer survivors who have offspring, there is the concern about congenital anomalies, genetic disease, or risk of cancer in the offspring. In the report from the National Wilms Tumor Group, congenital anomalies were marginally increased in the offspring of females who had received flank radiotherapy [44]. However, this risk was not observed in a study of 247 offspring of 148 cancer survivors treated at a single institution [45] or in several larger cohort studies. In a study that compared a group of 2,198 offspring from adult survivors treated for childhood cancer between 1945 and 1975 with a group of 4,544 offspring from sibling controls, there were no differences in the proportion of offspring with cytogenetic syndromes, single-gene defects, or simple malformations. There was no association of type of childhood cancer treatment used and the occurrence of genetic disease in the offspring [46]. In the CCSS, among the 1,915 female survivors who reported 4,029 pregnancies, there was no increased risk of offspring with simple malformations, cytogenetic syndromes, singlegene defects, or congenital malformations [35]. In a subsequent analysis from the CCSS, among children of 1,627 female cancer survivors, there was no increased risk for congenital anomalies and no increase conferred from ovarian radiation or alkylating exposure [47].

Similar results were reported in a study of 5,847 offspring of survivors of childhood cancers treated in five Scandinavian countries. In the absence of a hereditary cancer syndrome (such as hereditary retinoblastoma), there was no increased risk of cancer [48]. In an updated analysis from Finland among 26,331 children of pediatric and young adult cancer survivors and 58,155 children of siblings, there was no increased risk of cancer in the offspring of the cancer survivors in the absence of a known cancer predisposition syndrome [49].

Further follow-ups are needed to determine whether patterns of cancer or genetic disease in offspring change with changes in cancer treatments, further elapsed time, and studies of greater numbers of offspring.

#### 13.3 Detection and Screening

All prepubertal females who are treated with potentially gonadal toxic radiotherapy or chemotherapy should be rigorously assessed for appropriate progression through puberty. The average age for menarche is 12.7 years  $\pm$  1.0 year [50]. An evaluation should include a complete history, a physical examination that includes an assessment of sexual development and pubertal milestones (Tables 13.2 and 13.3) and selected laboratory studies (Table 13.4), as summarized in Table 13.5. Reduced ovarian volume and low inhibin B and anti-Mullerian hormone concentrations in survivors with regular menses may be markers of incipient ovarian failure [2, 7]. In conjunction with the evaluation of gonadal effects, attention must be paid to growth. Cranial radiotherapy confers significant risk for growth hormone deficiency. Once 
 Table 13.2
 Tanner staging (pubertal milestones) for breast development [40]

| Stage   | Age<br>(mean ±<br>SD, years) |
|---|------------------------------|
| I. Preadolescent. Only papilla is elevated  |                              |
| II. Breast and papilla are elevated as small mound. Areolar diameter is enlarged                                      | $10.0 \pm 1.0$               |
| III. Areola and papilla project to form a secondary mound above the level of the breast                               | 11.9±1.0                     |
| IV. There is projection only of papilla<br>because of recession of the areola to the<br>general contour of the breast | 12.9±1.2                     |

 Table 13.3
 Tanner staging (pubertal milestones) for pubic hair growth [40]

| Stage   | Age<br>(mean ±<br>SD,<br>years) |
|---|---------------------------------|
| I. Preadolescent vellus over pubis is no<br>further developed than that over anterior<br>abdominal wall (i.e., no pubic hair)                                       |                                 |
| II. There is sparse growth of long, slightly<br>pigmented, downy hair, straight or only<br>slightly curled, appearing chiefly along the<br>labia                    | 11.2±1.1                        |
| III. Hair is considerably darker, coarser, and<br>more curled. Hair spreads sparsely over<br>pubic junction   | 11.9±1.1                        |
| IV. Hair is now adult in type but area<br>covered by it is still considerably smaller<br>than in most adults. There is no spread to<br>medial surface of the thighs | 12.6±1.1                        |

patients have reached full sexual maturity, linear growth will stop. Linear and sexual development must, therefore, be monitored simultaneously (see the chapter on "Neuroendocrine Late Effects" for further details). Patients who received radiotherapy to the central nervous system or the neck are also at risk for thyroid dysfunction that can negatively impact gonadal function and linear growth. Even after successful progression through puberty, it is important to monitor the frequency and characteristics of menstrual periods, due to risk for premature menopause. Females with ovarian failure, either primary or secondary, should undergo assessments for impaired bone mineral density. Calcium intake, weight-bearing exercise, a history

| Laboratory assessment for ovarian function |   |  |
|--|---|--|
| Testing                                    | Treatment exposure  | Time and<br>frequency of<br>evaluations  |
| LH, FSH,<br>estradiol                      | Alkylating agents   | Baseline at<br>11 years of age or<br>older, and then<br>yearly   |
|  | Abdominopelvic,<br>cranial, or total body<br>radiotherapy | Assessment also<br>of whether the<br>following are<br>present: delayed<br>puberty, irregular<br>menses or<br>amenorrhea,<br>clinical signs or<br>symptoms of<br>estrogen<br>deficiency |
| Free T4,<br>TSH                            | Neck, cranial, or total<br>body radiotherapy              | Yearly<br>Assessment also<br>of presence of<br>signs or<br>symptoms of<br>thyroid<br>dysfunction   |

 Table 13.4
 Laboratory assessment for ovarian function

Also see Sect. 13.2

of fractures, and a family history of osteopenia/ osteoporosis should be evaluated. The determination of bone mineral density, using dualenergy X-ray absorptiometry (DXA) scan, and comparison of results with the well-established adult normative values, is indicated for all adult females. Screening in children is less defined. Several different measurement techniques and standards have been applied, but none has been well validated in large pediatric populations (much less in pediatric oncology patients). However, some monitoring is indicated, and trends over time may be of greater value than a single DXA scan. The Children's Oncology Group has published guidelines (www.survivorshipguidelines.org) [51] that are helping in determining surveillance for adverse long-term ovarian or ovarian-associated outcomes, as reviewed by Metzger and colleagues [52].

Pediatric endocrinologists and reproductive endocrinologists/gynecologists are essential consultants in the monitoring, prevention, and management of ovarian late effects in childhood cancer survivors.

| History   | Physical examination   |
|---|--|
| Doses and types of<br>chemotherapy agents<br>received   | Height, weight, and height velocity  |
| Doses and fields of radiotherapy  | Complete examination of all<br>organ systems, with<br>particular attention to<br>pubertal status and thyroid<br>gland                            |
| Surgical history,<br>especially for patients<br>with CNS and GU<br>tumors<br>Patient and maternal | Gynecologic examination in<br>postpubertal females as<br>indicated by treatment<br>history, sexual activity, and<br>overall developmental status |
| history of menarche and<br>thelarche  | overall developmental status   |
| Menstrual periods –<br>timing and tempo   |  |
| Symptoms of estrogen<br>deficiency (hot flashes,<br>dry skin, leg cramps,<br>reduced libido)      |  |
| Parental heights  |  |
| Family history of<br>infertility, pregnancy,<br>labor complications,<br>assisted fertilization    |  |

 Table 13.5
 Pertinent history and physical examination

## 13.4 Management of Established Problems

#### 13.4.1 Prevention Strategies

Reduction in the dose or use of alkylating agents and abdominopelvic radiotherapy is the most effective means of preserving ovarian function and promoting positive reproductive outcomes. There are, however, many instances where cytotoxic and gonadal toxic chemotherapy and radiotherapy are still required for long-term cure. As a result, additional strategies need to be employed to minimize adverse long-term outcomes. To shield the ovaries from direct irradiation during abdominal or pelvic radiotherapy, an oophoropexy may be performed if it is possible to move the ovaries to a location that can be safely shielded without jeopardizing the patient for tumor recurrence. Typically, with abdominal radiotherapy for Hodgkin lymphoma that targets lymph nodes, the ovaries are moved to a midline position in front of or behind the uterus. For pelvic radiotherapy, they may be moved laterally to the iliac wings. This may also be helpful for young girls or adolescents undergoing cranial spinal radiotherapy for brain tumors using historic radiation techniques, though current approaches with intensity modulation allow ovarian sparing. However, if such techniques are not possible, then the ovaries should be marked by the surgeon with clips that can later by identified by a simulator film. Central pelvic blocking at the time of "inverted Y" field will prevent direct irradiation, although scatter dose and transmitted dose will be inevitable. Medial or lateral transposition of the ovaries results in ovarian doses of 8-10 % and 4-5 %, respectively, of the pelvic dose [53]. For most patients, this will be compatible with the preservation of fertility, although there may be temporary amenorrhea.

Because dividing cells are more sensitive to the cytotoxic effects of alkylating agents than are cells at rest, it has been hypothesized that inhibition of the pituitary-gonadal axis by gonadotropin-releasing hormone (GnRH) agonists may protect the ovarian germinal epithelium from the cytotoxic effects of chemotherapy. In a mixed teenage and young adult group of women treated for lymphoma, leukemia, or autoimmune disease, Blumenfeld and colleagues [54, 55] reported a significant benefit in the concomitant use of GnRH agonist treatment with cytotoxic chemotherapy. Pereya and colleagues evaluated the role of GnRH analogs with respect to the prevention of early-onset ovarian insufficiency following chemotherapy in adolescent females. Their study compared prepubertal females treated with GnRH analogs prior to chemotherapy with a control group of prepubertal patients who were not given GnRH analogs. Pereya and colleagues found that GnRH analog treatment before and during chemotherapy might enhance ovarian function and preserve adolescent fertility [56]. However, as reviewed in the literature, and in two randomized prospective studies, it is not clear that there is a benefit, and thus, this is currently not considered a standard of care [57-60].

Progress in reproductive endocrinology has resulted in the availability of several potential options for preserving or permitting fertility in females about to receive potentially toxic chemotherapy or radiotherapy. In pre- and postpubertal females, cryopreservation of ovarian cortical tissue and enzymatically extracted follicles, with the in vitro maturation of prenatal follicles, is of potential clinical use. There is now feasibility of doing this at time of diagnosis in young females diagnosed with cancer, and thus, this is an area of ongoing important research [61–68]. Another option available to the postpubertal female is the stimulation of ovaries with exogenous gonadotropins and the retrieval of mature oocytes for cryopreservation and later in vitro fertilization. These interventions, however, may not be readily available to the pediatric and adolescent patient, and the necessary delay in cancer therapy for ovarian stimulation may then be impractical. Oocyte cryopreservation may be useful in the survivorship population where there is concern over decreasing ovarian reserve. All such approaches harbor the risk that malignant cells will be present in the specimen and reintroduced in the patient at a later date. Those with hematologic or gonadal tumors would be at greatest risk for this eventuality. However, success rates are increasing with newer technologies and further research is ongoing [68]. Standards for best practice in the cryopreservation of gonadal tissue remain to be defined. Should offspring result as a consequence of these assisted fertility techniques, it would be imperative to evaluate the risk of chromosomal and other congenital disorders, which have been reported following assisted reproductive techniques [69, 70].

A critical component to prevention is health counseling for females at risk. For females treated during the prepubertal period, parents should be counseled regarding the risk of primary ovarian failure. Normal gonadal development should be reviewed with recommendations for monitoring of growth and development. Reproductive counseling should be made cautiously and preferably, in conjunction with a specialist in reproductive endocrinology. The effects on the female gonadal system from radiotherapy and chemotherapy may demonstrate significant interindividual variation, even with identical exposures at identical ages. Postpubertal females who have normal menstrual function should be appropriate contraception counseled about should they currently not wish to conceive a child, and they should also be made aware of their potential risk for premature menopause. Not inconsequential for young adults is the impact of ovarian failure or impending failure in sexual drive or libido, an effect that may be treatable if addressed. Risks for osteopenia and osteoporosis also must be addressed. Appropriate calcium intake, avoidance of substances that interfere with bone deposition and appropriate weight-bearing exercise should be encouraged to maintain skeletal health.

## 13.4.2 Management of Delayed Puberty

Female patients exposed to gonadal toxic therapies during the prepubertal period and who are not progressing appropriately through puberty should be promptly referred to a pediatric endocrinologist for further evaluation and treatment. The use of hormonal replacement therapy for induction and progression of puberty must be closely monitored together with skeletal growth, as the two processes are closely linked. Generally, the recommendation will be to initiate a regimen of hormone replacement such as estrogen, which is now available in a variety of doses and modes of administration. Gonadotropins, gonadotropin agonists or antagonists, progesterone, and growth hormone may also be part of the treatment regimen.

#### 13.4.3 Management of Infertility

Postpubertal patients at risk for infertility should be referred to a reproductive endocrinologist to discuss assisted fertility techniques that may be appropriate. These specialists can also monitor fertility status and assist survivors with reproductive decisions.

# 13.4.4 Management of Pregnancy and Delivery

While many childhood cancer survivors may have no prenatal or perinatal complications, others may be at risk and should be managed appropriately by obstetricians and perinatologists. Patients treated with abdominopelvic radiotherapy are at risk for spontaneous abortion, premature labor and delivery, and, compared with controls, small for gestational stage neonates. Those treated with anthracyclines at doses  $>300 \text{ mg/m}^2$  or at lower doses, when combined with thoracic radiotherapy, or those women treated with high doses of thoracic radiotherapy (>35 Gy) without anthracyclines, may be at risk for cardiac complications, which may manifest during pregnancy (especially during the third trimester and during delivery). Similarly, women previously treated with bleomycin, carmustine, or busulfan, with higher doses of thoracic radiotherapy, may be at risk for pulmonary fibrosis or decreased diffusing capacity, and this may result in complications during pregnancy and delivery (see the chapters on "Heart and Lung Late Effects" for further details).

## 13.4.5 Management of Premature Menopause

Female survivors who develop premature menopause should be referred to a gynecologist for management and consideration for hormone replacement therapy. The decision to proceed with hormone replacement therapy, and the form that it should take, involves a careful evaluation of many competing healthcare factors, a subject that is beyond the scope of this chapter. However, it is imperative that patients be managed by a team of physicians who are well versed in this area and can assist in carefully weighing the risks and benefits of various hormonal replacement strategies.

#### Summary

Both chemotherapy and radiotherapy can affect ovarian function in female survivors of childhood cancer. The effects are varied and dependent on the chemotherapeutic agents and doses, radiotherapy doses, techniques, volumes and fields, and the age and pubertal status of the female. There is also considerable individual variation, the reasons for which remain largely unknown. Problems may include primary ovarian failure, reduced libido, pregnancy complications, and premature menopause. Preventive strategies remain limited. Avoidance or reduction in the dose of gonadal toxic therapies should be attempted where possible. Where this is not possible, advances in reproductive medicine may ultimately allow for ovarian cryopreservation and similar techniques. Survivors should receive health counseling about risks, annual physical examinations with attention paid to endocrine and reproductive function, close monitoring of gonadal function, and referral to pediatric endocrinologists, reproductive endocrinologists, gynecologists, and perinatologists as indicated. In survivors who do become pregnant, the majority will have favorable pregnancy outcomes with healthy offspring, and it does not appear that the offspring will have an increased risk of cancer (in the absence of a known heritable syndrome or congenital anomalies).

Much of what we have learned about gonadal function in female childhood cancer survivors is based on patients treated in the 1960s through the middle 1980s. During the past 20 years, there has been increased awareness of the adverse gonadal effects, and where possible, therapies have been altered to limit these effects. This period has also resulted in the increased use of more doseintensive chemotherapy regimens and greater use of myeloablative hematopoietic stem cell transplants. Survivorship has increased and, as a result, there are now large cohorts of adult young women treated with more contemporary therapy who will require close follow-up of their gonadal status. It is only with continued follow-up that we will be able to fully appreciate the impact of relatively recent changes in therapy. Challenges still face young females being treated for cancer today with respect to gonadal function. Therefore, it is incumbent upon pediatric oncologists and reproductive specialists to develop better preventive strategies. In addition, there is more to be learned about the interindividual differences in gonadal effects that are seen, despite very similar treatment exposures. The role of genetic predisposition and inherent chemotherapy (or radiotherapy) sensitivity has yet to be studied with respect to most adverse long-term outcomes (including ovarian function) for childhood cancer survivors.

#### References

- Palter S, Olive D (2002) Reproductive physiology. In: Berek J, Adams Hillard P, Adashi E (eds) Novak's gynecology. Lippincott Williams and Wilkins, Philadelphia, pp 163–164
- Lie Fong S, Laven JS, Hakvoort-Cammel FG, Schipper I, Visser JA, Themmen AP, de Jong FH, van den Heuvel-Eibrink MM (2009) Assessment of ovarian reserve in adult childhood cancer survivors using anti-Mullerian hormone. Hum Reprod 24(4):982–990
- Chapman RM, Sutcliffe SB, Malpas JS (1979) Cytotoxic induced ovarian failure in women with Hodgkin's disease. JAMA 242:1877–1881
- Shalet SM (1996) Endocrine sequelae of cancer therapy. Eur J Endocrinol 135:135–143
- Zebrack BJ, Casillas J, Nohr L, Adams H, Zeltzer LK (2004) Fertility issues for young adult survivors of childhood cancer. Psychooncology 13(10):689–699
- Friedman DL, Constine LS (2011) Late effects of cancer treatment. In: Haperin EC, Constine LS, Tarbell NJ, Kun LE (eds) Pediatric radiation oncology. Lippincott Williams and Wilkins, Philadelphia, pp 353–396
- Critchley HO, Wallace WH (2005) Impact of cancer treatment on uterine function. J Natl Cancer Inst Monogr 34:64–68
- Wallace WH, Shalet SM, Hendry JH, Morris-Jones PH, Gattamaneni HR (1989) Ovarian failure following abdominal irradiation in childhood: the radiosensitivity of the human oocyte. Br J Radiol 62:995–998
- 9. Ash P (1980) The influence of radiation on fertility in man. Br J Radiol 53:271–278
- Lushbaugh CC, Casarett GW (1976) The effects of gonadal irradiation in clinical radiation therapy: a review. Cancer 37(Suppl 2):1111–1125
- Byrne J, Mulvihill JJ, Myers MH, Connelly RR, Naughton MD, Krauss MR, Steinhorn SC, Hassinger DD, Austin DF, Bragg K (1987) Effects of treatment on fertility in long-term survivors of childhood or adolescent cancer. N Engl J Med 317:1315–1321
- Nicosia SV, Matus-Ridley M, Meadows AT (1985) Gonadal effects of cancer therapy in girls. Cancer 55:2364–2372
- Damewood MD, Grochow LB (1986) Prospects for fertility after chemotherapy or radiation for neoplastic disease. Fertil Steril 45:443–459

- Bath LE, Hamish W, Wallace B, Critchley HO (2002) Late effects of the treatment of childhood cancer on the female reproductive system and the potential for fertility preservation. BJOG Int J Obstet Gynaecol 109:107–114
- Hill M, Milan S, Cunningham D, Mansi J, Smith I, Catovsky D, Gore M, Zulian G, Selby P, Horwich A et al (1995) Evaluation of the efficacy of the VEEP regimen in adult Hodgkin's disease with assessment of gonadal and cardiac toxicity. J Clin Oncol 13:387– 395, (see comments). Comment in: J Clin Oncol 1995,13:1283–1284
- Mayer EI, Dopfer RE, Klingebiel T, Scheel-Walter H, Ranke MB, Niethammer D (1999) Longitudinal gonadal function after bone marrow transplantation for acute lymphoblastic leukemia during childhood. Pediatr Transplant 3:38–44
- Thomson AB, Critchley HO, Kelnar CJ, Wallace WH (2002) Late reproductive sequelae following treatment of childhood cancer and options for fertility preservation. Best Pract Res Clin Endocrinol Metab 16:311–334
- Stillman RJ, Schinfeld JS, Schiff I, Gelber RD, Greenberger J, Larson M, Jaffe N, Li FP (1981) Ovarian failure in long-term survivors of childhood malignancy. Am J Obstet Gynecol 139:62–66
- Halperin EC (1993) Concerning the inferior portion of the spinal radiotherapy field for malignancies that disseminate via the cerebrospinal fluid. Int J Radiat Oncol Biol Phys 26:357–362
- Critchley HO, Bath LE, Wallace WH (2002) Radiation damage to the uterus – review of the effects of treatment of childhood cancer. Hum Fertil (Camb) 5:61–66
- Sklar C, Boulad F, Small T, Kernan N (2001) Endocrine complications of pediatric stem cell transplantation. Front Biosci 6:G17–G22
- 22. Ogilvy-Stuart AL, Clark DJ, Wallace WH, Gibson BE, Stevens RF, Shalet SM, Donaldson MD (1992) Endocrine deficit after fractionated total body irradiation. Arch Dis Child 67:1107–1110
- Sklar CA (1991) Growth and pubertal development in survivors of childhood cancer. Pediatrician 18:53–60
- Howell S, Shalet S (1998) Gonadal damage from chemotherapy and radiotherapy. Endocrinol Metab Clin N Am 27:927–943
- 25. Kreuser ED, Xiros N, Hetzel WD, Heimpel H (1987) Reproductive and endocrine gonadal capacity in patients treated with COPP chemotherapy for Hodgkin's disease. J Cancer Res Clin Oncol 113: 260–266
- Friedman DL, Constine LS (2006) Late effects of treatment for Hodgkin lymphoma. J Natl Compr Cancer Netw 4(3):249–257
- 27. Schwartz CL, Constine LS, Villaluna D, London WB, Hutchison RE, Sposto R, Lipshultz SE, Turner CS, de Alarcon PA, Chauvenet A (2009) A risk-adapted, response-based approach using ABVE-PC for children and adolescents with intermediate- and high-risk Hodgkin lymphoma: the results of P9425. Blood 114(10):2051–2059, PMCID: 2744567

- 28. Mauz-Korholz C, Hasenclever D, Dorffel W, Ruschke K, Pelz T, Voigt A, Stiefel M, Winkler M, Vilser C, Dieckmann K, Karlen J, Bergstrasser E, Fossa A, Mann G, Hummel M, Klapper W, Stein H, Vordermark D, Kluge R, Korholz D (2010) Procarbazine-free OEPA-COPDAC chemotherapy in boys and standard OPPA-COPP in girls have comparable effectiveness in pediatric Hodgkin's lymphoma: the GPOH-HD-2002 study. J Clin Oncol 28(23): 3680–3686
- Metzger ML, Weinstein HJ, Hudson MM, Billett AL, Larsen EC, Friedmann A, Howard SC, Donaldson SS, Krasin MJ, Kun LE, Marcus KJ, Yock TI, Tarbell N, Billups CA, Wu J, Link MP (2012) Association between radiotherapy vs no radiotherapy based on early response to VAMP chemotherapy and survival among children with favorable-risk Hodgkin lymphoma. JAMA 307(24):2609–2616, PMCID: 3526806
- 30. Wolden SL, Chen L, Kelly KM, Herzog P, Gilchrist GS, Thomson J, Sposto R, Kadin ME, Hutchinson RJ, Nachman J (2012) Long-term results of CCG 5942: a randomized comparison of chemotherapy with and without radiotherapy for children with Hodgkin's lymphoma – a report from the Children's Oncology Group. J Clin Oncol 30(26):3174–3180, PMCID: 3434976
- 31. Sharon N, Neumann Y, Kenet G, Schachter J, Rechavi G, Toren A (2001) Successful pregnancy after highdose cyclophosphamide and ifosfamide treatment in two postpubertal women. Pediatr Hematol Oncol 18(4):247–252
- Williams D, Crofton PM, Levitt G (2008) Does ifosfamide affect gonadal function? Pediatr Blood Cancer 50(2):347–351
- 33. Chiarelli AM, Marrett LD, Darlington G (1999) Early menopause and infertility in females after treatment for childhood cancer diagnosed in 1964–1988 in Ontario, Canada. Am J Epidemiol 150:245–254
- 34. Chemaitilly W, Mertens AC, Mitby P, Whitton J, Stovall M, Yasui Y, Robison LL, Sklar CA (2006) Acute ovarian failure in the childhood cancer survivor study. J Clin Endocrinol Metab 91(5):1723–1728
- 35. Green DM, Sklar CA, Boice JD Jr, Mulvihill JJ, Whitton JA, Stovall M, Yasui Y (2009) Ovarian failure and reproductive outcomes after childhood cancer treatment: results from the Childhood Cancer Survivor Study. J Clin Oncol 27(14):2374–2381, PMCID: 2677923
- 36. Green DM, Kawashima T, Stovall M, Leisenring W, Sklar CA, Mertens AC, Donaldson SS, Byrne J, Robison LL (2009) Fertility of female survivors of childhood cancer: a report from the childhood cancer survivor study. J Clin Oncol 27(16):2677–2685, PMCID: 2690392
- 37. Sanders JE, Buckner CD, Leonard JM, Sullivan KM, Witherspoon RP, Deeg HJ, Storb R, Thomas ED (1983) Late effects on gonadal function of cyclophosphamide, total-body irradiation, and marrow transplantation. Transplantation 36:252–255

- 38. Sanders JE, Hawley J, Levy W, Gooley T, Buckner CD, Deeg HJ, Doney K, Storb R, Sullivan K, Witherspoon R, Appelbaum FR (1996) Pregnancies following high-dose cyclophosphamide with or without high-dose busulfan or total-body irradiation and bone marrow transplantation. Blood 87:3045–3052
- 39. Green DM, Whitton JA, Stovall M, Mertens AC, Donaldson SS, Ruymann FB, Pendergrass TW, Robison LL (2002) Pregnancy outcome of female survivors of childhood cancer: a report from the Childhood Cancer Survivor Study. Am J Obstet Gynecol 187:1070–1080
- Signorello LB, Mulvihill JJ, Green DM, Munro HM, Stovall M, Weathers RE, Mertens AC, Whitton JA, Robison LL, Boice JD Jr (2010) Stillbirth and neonatal death in relation to radiation exposure before conception: a retrospective cohort study. Lancet 376(9741):624–630, PMCID: 3008402
- 41. Winther JF, Boice JD Jr, Svendsen AL, Frederiksen K, Stovall M, Olsen JH (2008) Spontaneous abortion in a Danish population-based cohort of childhood cancer survivors. J Clin Oncol 26(26):4340–4346, PMCID: 2653117
- 42. Madanat-Harjuoja LM, Lahteenmaki PM, Dyba T, Gissler M, Boice JD Jr, Malila N (2013) Stillbirth, early death and neonatal morbidity among offspring of female cancer survivors. Acta Oncol 52(6):1152–1159
- 43. Winther JF, Olsen JH, Wu H, Shyr Y, Mulvihill JJ, Stovall M, Nielsen A, Schmiegelow M, Boice JD Jr (2012) Genetic disease in the children of Danish survivors of childhood and adolescent cancer. J Clin Oncol 30(1):27–33, PMCID: 3255559
- 44. Green DM, Peabody EM, Nan B, Peterson S, Kalapurakal JA, Breslow NE (2002) Pregnancy outcome after treatment for Wilms' tumor: a report from the National Wilms' Tumor Study Group. J Clin Oncol 20:2506–2513
- 45. Green DM, Fiorello A, Zevon MA, Hall B, Seigelstein N (1997) Birth defects and childhood cancer in offspring of survivors of childhood cancer. Arch Pediatr Adolesc Med 151:379–383
- 46. Byrne J, Rasmussen SA, Steinhorn SC, Connelly RR, Myers MH, Lynch CF, Flannery J, Austin DF, Holmes FF, Holmes GE, Strong LC, Mulvihill JJ (1998) Genetic disease in offspring of long-term survivors of childhood and adolescent cancer (comment). Am J Hum Genet 62:45–52
- 47. Signorello LB, Mulvihill JJ, Green DM, Munro HM, Stovall M, Weathers RE, Mertens AC, Whitton JA, Robison LL, Boice JD Jr (2012) Congenital anomalies in the children of cancer survivors: a report from the childhood cancer survivor study. J Clin Oncol 30(3):239–245, PMCID: 3269950
- 48. Sankila R, Olsen JH, Anderson H, Garwicz S, Glattre E, Hertz H, Langmark F, Lanning M, Moller T, Tulinius H (1998) Risk of cancer among offspring of childhood-cancer survivors. Association of the Nordic Cancer Registries and the Nordic Society of Paediatric

Haematology and Oncology. N Engl J Med 338: 1339–1344

- Madanat-Harjuoja LM, Malila N, Lahteenmaki P, Pukkala E, Mulvihill JJ, Boice JD Jr, Sankila R (2010) Risk of cancer among children of cancer patients – a nationwide study in Finland. Int J Cancer 126(5):1196–1205, PMCID: 2801768
- Rosenfield RL (2002) Puberty in the female. In: Sperling MA (ed) Pediatric endocrinology. Saunders, Philadelphia
- 51. Children's Oncology Group (2008) Long-term follow-up guidelines for survivors of childhood, adolescent and young adult cancers, version 3.0. Children's Oncology Group, Arcadia, Available on-line: www. survivorshipguidelines.org
- 52. Metzger ML, Meacham LR, Patterson B, Casillas JS, Constine LS, Hijiya N, Kenney LB, Leonard M, Lockart BA, Likes W, Green DM (2013) Female reproductive health after childhood, adolescent, and young adult cancers: guidelines for the assessment and management of female reproductive complications. J Clin Oncol 31(9):1239–1247
- 53. Haie-Meder C, Mlika-Cabanne N, Michel G, Briot E, Gerbaulet A, Lhomme C, Cosset JM, Sarrazin D, Flamant F, Hayat M (1993) Radiotherapy after ovarian transposition: ovarian function and fertility preservation. Int J Radiat Oncol Biol Phys 25: 419–424
- Blumenfeld Z, Dann E, Avivi I, Epelbaum R, Rowe JM (2002) Fertility after treatment for Hodgkin's disease. Ann Oncol 13(Suppl 1):138–147
- 55. Blumenfeld Z, Avivi I, Linn S, Epelbaum R, Ben-Shahar M, Haim N (1996) Prevention of irreversible chemotherapy-induced ovarian damage in young women with lymphoma by a gonadotrophin-releasing hormone agonist in parallel to chemotherapy. Hum Reprod 11:1620–1626
- 56. Pereyra Pacheco B, Mendez Ribas JM, Milone G, Fernandez I, Kvicala R, Mila T, di Noto A, Contreras Ortiz O, Pavlovsky S (2001) Use of GnRH analogs for functional protection of the ovary and preservation of fertility during cancer treatment in adolescents: a preliminary report. Gynecol Oncol 81:391–397
- Oktay K, Oktem O (2009) Fertility preservation medicine: a new field in the care of young cancer survivors. Pediatr Blood Cancer 53(2):267–273
- Oktem O, Oktay K (2007) Quantitative assessment of the impact of chemotherapy on ovarian follicle reserve and stromal function. Cancer 110(10):2222–2229
- 59. Oktay K, Sonmezer M, Oktem O, Fox K, Emons G, Bang H (2007) Absence of conclusive evidence for the safety and efficacy of gonadotropin-releasing hormone analogue treatment in protecting against

chemotherapy-induced gonadal injury. Oncologist 12(9):1055–1066

- 60. Waxman JH, Ahmed R, Smith D, Wrigley PF, Gregory W, Shalet S, Crowther D, Rees LH, Besser GM, Malpas JS et al (1987) Failure to preserve fertility in patients with Hodgkin's disease. Cancer Chemother Pharmacol 19(2):159–162
- 61. Gracia CR, Chang J, Kondapalli L, Prewitt M, Carlson CA, Mattei P, Jeffers S, Ginsberg JP (2012) Ovarian tissue cryopreservation for fertility preservation in cancer patients: successful establishment and feasibility of a multidisciplinary collaboration. J Assist Reprod Genet 29(6):495–502. doi:10.1007/ s10815-012-9753-7. Epub 2012
- 62. Donnez J, Dolmans MM, Pellicer A et al (2013) Restoration of ovarian activity and pregnancy after transplantation of cryopreserved ovarian tissue: a review of 60 cases of reimplantation. Fertil Steril 99(6):p15031513
- Newton H (1998) The cryopreservation of ovarian tissue as a strategy for preserving the fertility of cancer patients. Hum Reprod Update 4:237–247
- Oktay K, Karlikaya G (2000) Ovarian function after transplantation of frozen, banked autologous ovarian tissue. N Engl J Med 342(25):1919
- 65. Donnez J, Dolmans MM, Demylle D, Jadoul P, Pirard C, Squifflet J, Martinez-Madrid B, van Langendonckt A (2004) Livebirth after orthotopic transplantation of cryopreserved ovarian tissue. Lancet 364(9443): 1405–1410
- 66. Oktay K, Buyuk E, Veeck L, Zaninovic N, Xu K, Takeuchi T, Opsahl M, Rosenwaks Z (2004) Embryo development after heterotopic transplantation of cryopreserved ovarian tissue. Lancet 363(9412):837–840
- 67. Meirow D, Levron J, Eldar-Geva T, Hardan I, Fridman E, Zalel Y, Schiff E, Dor J (2005) Pregnancy after transplantation of cryopreserved ovarian tissue in a patient with ovarian failure after chemotherapy. N Engl J Med 353(3):318–321
- Oktay K, Cil AP, Bang H (2006) Efficiency of oocyte cryopreservation: a meta-analysis. Fertil Steril 86(1): 70–80
- 69. Davies MJ, Moore VM, Willson KJ, Van Essen P, Priest K, Scott H, Haan EA, Chan A (2012) Reproductive technologies and the risk of birth defects. N Engl J Med 366(19):1803–1813
- 70. Farhi A, Reichman B, Boyko V, Mashiach S, Hourvitz A, Margalioth EJ, Levran D, Calderon I, Orvieto R, Ellenbogen A, Meyerovitch J, Ron-El R, Lerner-Geva L (2013) Congenital malformations in infants conceived following assisted reproductive technology in comparison with spontaneously conceived infants. J Matern Fetal Neonatal 26(12):1171–1179