Chapter 10 Stimuli Responsive Carriers: Magnetically, Thermally and pH Assisted Drug Delivery

Eameema Muntimadugu, Anjali Jain, and Wahid Khan

Abbreviations

10.1 Introduction

 Clinical application of most of the drugs is limited by their side effects in spite of their beneficial action. There has been a long time desire to achieve selective delivery of bioactives to target areas in the body in order to maximize therapeutic potential and minimize side-effects. For achieving better therapeutic application,

 Department of Pharmaceutics , National Institute of Pharmaceutical Education and Research (NIPER), Hyderabad, India e-mail: [wahid@niperhyd.ac.in](mailto: wahid@niperhyd.ac.in)

© Controlled Release Society 2015 341

P.V. Devarajan, S. Jain (eds.), *Targeted Drug Delivery: Concepts and Design*, Advances in Delivery Science and Technology, DOI 10.1007/978-3-319-11355-5_10

E. Muntimadugu • A. Jain • W. Khan (\boxtimes)

nanocarriers are considered for target-specific delivery of drugs to various sites in the body in order to improve the therapeutic efficacy, while minimizing undesirable side effects [1]. Nanocarriers possess in vivo longevity and specific capability of extravasation through the endothelium of inflammatory tissues (the so-called enhanced permeability and retention effect), whereas their functionalisation with biologically active ligands facilitates the targeting of specific cells. However, the translation of both the enhanced permeability and retention effect and ligand recognition into the clinic still remains questionable. This may be, to a certain extent a consequence of the stochastic nature of ligand–receptor interactions and of difficulties in the control of the release of the drug from targeting nanocarriers. One alternative involves on-demand processes (also termed "switch on/off"), which in principle allow for tailored release profiles with excellent spatial, temporal and dosage control. On-demand drug delivery is becoming feasible through the design of stimuli responsive systems that recognize their microenvironment and react in a dynamic way, mimicking the responsiveness of living organisms [2]. The concept of stimuliresponsive drug delivery was first suggested in the late 1970s with the use of thermosensitive liposomes for the local release of drugs through hyperthermia. Since then, and particularly in the past decade a great deal of research has been carried out on stimuli-responsive materials for drug delivery, especially concerning their design and applications as nanocarriers [3].

 Stimuli-responsive nanocarriers are specialised nano-sized active delivery vehicles that evolve with an external signal and are equipped with "load-and-release" modalities within their constituting units. The central operating principle of these carriers lies in the fact that a specific cellular/extracellular stimulus of chemical, biochemical, or physical origin can modify the structural composition/conformation of the nanocarriers, thereby promoting release of the active species to specific biological environment. The observed changes are mainly decomposition, isomerisation, polymerisation, activation of supramolecular aggregation among many others. The general concept of triggered release can be divided mainly into two major modes according to the nature of the interaction between the bioactive molecule and the nanocarriers. In the complexation approach, where the bioactive agent is entrapped within the nanocarrier, the release can be triggered by structural change within the carrier scaffold (i.e. carrier degradation, cleavage of shell, charging of functional groups), while in the nanocarrier-conjugate approach; the mechanism of release involves the splitting of the linker between the carrier and the bioactive agent. The external stimuli which bring about these changes are numerous and cross related. These advanced nanocarriers thus become an active participant in the therapeutic landscape, rather than an inert carrier molecule [4].

10.2 Classification

Stimuli that trigger drug release from the nanocarriers can be broadly classified with respect to the biological systems as either endogenous (physiological, pathological, and patho-chemical conditions) or exogenous (physical) (Fig. 10.1).

Fig. 10.1 Classification of stimuli involved in responsive drug delivery systems

Endogenous stimuli of chemical and biochemical origin include cellular pH-shift, redox, and ionic microenvironment of the specific tissues, enzyme over-expression in certain pathological states, host–guest recognitions, and antigen–antibody interactions. Physical stimuli that can be applied externally to bring about a triggered release of active guest may involve temperature, light, mechanical pressure, and strength of magnetic or electrical fields. This chapter throws light on pH, thermally and magnetic field assisted drug delivery either alone or as dual responsive systems.

10.3 pH-Responsive Carriers

 Among the common stimuli, pH-responsiveness is the most frequently used. These carriers respond to pH gradients within the microenvironments of organs, tissues and cell organelles to achieve drug release at desired site. Certain tumours and inflamed tissue have a slightly lower pH values (between pH 5.4 and 7.4) than homeostatic conditions (pH 7.4). Furthermore, there exists a lower intracellular pH in endosomes and lysosomes. As such particles internalised through endocytosis will experience a pH gradient from neutral $(pH ~ 7.4)$ in extracellular environment, to acidic (pH \sim 6.2) in early endosome and more acidic (pH \sim 5.0) in lysosome [5, 6]. Moreover, members of the ATP-binding cassette (ABC) efflux pump superfamily, such as P-glycoprotein (P-gp)/ABCB1, MDR-associated protein/ABCC, and breast cancer resistance protein/ABCG2, play important roles in drug kinetics including absorption, distribution, metabolism and excretion, which limits the accumulation of drugs inside the cells and results in drug resistance [\[7](#page-19-0)]. pH responsive carriers are expected to provide fast intracellular drug release and make the intracellular drug

concentration to reach a sufficiently high level to exceed the efflux capacity of drug transporters and the threshold concentration to kill the MDR tumour cells.

 There are several general approaches of such systems that undergo chemical transitions around the critical pH range of $5-7$ [8]. One approach is to introduce "titratable" or "protonizable" chemical groups such as amines and carboxylic acids into the components assembling the nanocarriers. The systems containing amines or carboxylic groups with different chemical structures and pKa values could change their physical and chemical properties, such as swelling ratio or solubility in response to local pH level $[9]$. Another approach is to incorporate acid-labile linkages directly to attach drugs covalently to the vectors or into the main-chains of the polymers constructing the carriers. The pH sensitive bonds are cleaved at acidic pH, accompanied by dissolution of carriers and release of drugs. The third approach is to incorporate carbon dioxide $(CO₂)$ -generating ingredient for inducing $CO₂$ gas in acidic environment and leading to disintegration of the vehicles $[10]$.

10.3.1 Delivery Systems

 pH-responsive systems are mainly designed and reported as dendrimer, liposomes, nanoparticles and nanofibres (Fig. 10.2 and Table 10.1). Dendrimers are highly branched oligomers or polymers characterised by three structural features: (1) the central core from which the polymeric branches emanate, (2) the nature of the repeating unit which determines the microenvironment of the interior and thus the solubilisation ability of the dendrimer and (3) the nature and number of the terminal functional groups, mainly responsible for the behavior of dendrimers in solution. Pistolis et al. Developed pyrene loaded poly(propyleneimine) dendrimers for pH dependent release of pyrene. The release was increased up to tenfold by decreasing the pH to 2–4 $[11]$. Dual acting pH and thermosensitive dendrimer with a shell of poly $(N, N$ -dimethylaminoethyl methacrylate) were also developed $[12]$. Similarly Yuan et al., reported pH-sensitiveness and cellular targeting dendrimer to provide the advantage of thermo regulated targeting. This system contained $poly(L-glutamic)$ acid) dendrimers with a polyhedral oligomeric silsesquioxane nanocubic core. Doxorubicin was attached via pH-sensitive hydrazine bonds and biotin was used as targeting moiety [13].

 As another delivery system, pH-sensitive liposomes are designed to undergo acid-triggered destabilisation. For this, first generation pH-sensitive liposomes which are based on the cone-shaped lipid dioleoylphosphatidylethanolamine and later, serum-resistant pH-sensitive liposome formulations containing egg phosphatidylcholine and cholesteryl hemisuccinate are developed. These liposomes exhibited excellent stability at pH 7.4 and underwent rapid destabilisation upon acidification $[14, 15]$ $[14, 15]$ $[14, 15]$.

 For polymeric delivery, few common examples of pH sensitive polymers are poly(methacrylic acid)s, poly(vinylpyridine)s, poly(vinylimmidazole)s.

 Fig. 10.2 Stimuli responsive carriers

Moreover, efforts are being made to develop new co-polymers with pH responsive properties. Giacomellietal. Synthesised poly(ethyleneoxide)-b-poly(glycerolmonomethacrylate)-Indomethacin conjugates. This polymer–drug conjugate self assembled into micelle in water. The release of Indomethacin (IND) was governed by intrinsic molecular characteristics of free-IND (aqueous dissociation behavior) and pH-sensitivity of ester linkages in the conjugates. At neutral pH, the ester bond linkages were stable which promoted diffusion of free-IND out of the carrier, whereas acidic pH facilitates sustained release with slow kinetics $[16]$.

 pH sensitive poly(methacrylic acid and methacrylate) nanoparticles were designed to improve oral bioavailability of cyclosporin [17, 18]. Borchert et al. observed the pH-induced release of hydrophilic dyes from poly(2-vinylpyridine-bethylene oxide) block copolymer vesicles. At pH less than 5, protonation and dissolution of the poly-2-vinylpyridine blocks facilitated the release of dye from vesicles [19]. Similarly, gold-decorated shape-persistent, pH-responsive polymersomes were prepared by the self-assembly of a novel poly(ethyleneoxide)-blockpoly[2-(diethylamino)ethylmethacrylate-stat-3-(trimethoxysilyl)-propylmethacrylate], copolymer. These pH-sensitive blocks are located in the membrane walls, while the hydrophilic PEO chains forms the corona [20].

Table 10.1 Polymers and systems used for different stimuli responsive carriers **Table 10.1** Polymers and systems used for different stimuli responsive carriers

10.3.2 Applications

10.3.2.1 Anti Cancer Therapy

 pH sensitive liposomes made of dioleoylphosphatidylethanolamine (DOPE) and oleic acid or DOPE and 1,2-dipalmitoylsuccinylglycerol explored multiple possibilities to treat cancer. Problems associated with less circulation half-life, stability in blood have been resolved $[21]$. Recently, liposomal delivery system modified with pH-responsive cell penetrating peptide TH (TH-Lip) has been reported. TH was found to be a wonderful pH responsive ligand as the cell penetrating capacity of TH concealed during the blood circulation and in normal tissues at neutral pH. However, when TH-Lip reached the tumour, low pH at these sites promoted protonation of TH and the surface charge of TH-Lip converted from negative to positive thus promoted enhanced cellular and tumour spheroid uptake [22].

10.3.2.2 Antibacterial Therapy

 Bacterial infections are generally characterised by very low pH values because of anaerobic fermentation and subsequent inflammation. In this regard, systemic antibiotic therapy was achieved by incorporating an ionisable polyhistidine segment in a block copolymer to make PLGA-b-polyhistidine-b-PEG triblock copolymer nanoparticles. A charge switch at the sites of localised acidity promoted interactions with the negatively charged bacterial wall, and led to increased nanoparticle uptake in both Gram-positive and Gram-negative bacteria [23].

10.3.2.3 Intracellular Trafficking

 The usefulness of pH-sensitive liposomes has been well exhibited in a wide variety of applications, these include the transport of fluorescent probes to estimate the efficacy of different liposome compositions and to explain the mechanisms involved in intracellular trafficking, the intracellular transport of antigens, targeting intracellular pathways involved in processing and presentation of antigens and enhancing the immune response to tumour cells [24].

10.3.2.4 Oral Bioavailability Improvement

 Because of the broad range of pH found throughout the gastrointestinal tract, pHresponsive systems for oral drug delivery have been designed to protect drugs from the harsh conditions found in the gastric cavity and to improve their absorption in the intestine $[25]$. For instance, poly(methacrylic acid)-based copolymers have been used as pH-sensitive coatings at the surface of porous silica nanoparticles, as well as to prepare copolymer micelles able to disassemble at the intestinal pH $[26]$.

This charge-reversal approach was also applied to multi stimuli responsive nanocarriers to achieve drug release at neutral pH by taking advantage of electrostatic interactions, and to chitosan nanoparticles for gastric or intestinal delivery [\[27](#page-19-0)]. Similarly it was reported, pH sensitive oly(methacrylic acid and methacrylate) nanoparticles improved the oral bioavailability of cyclosporin [18].

10.3.2.5 DNA Therapeutics

Gene therapy is most widely explored field in biomedical research and increasing interest in stimuli responsive carriers to deliver DNA therapeutics can open multiple opportunities in this area. pH senstive liposomes were developed to deliver plasmid DNA into mammalian cell lines [28].

10.4 Thermoresponsive Carriers

 Thermoresponsive carriers are usually governed by a nonlinear sharp change in the properties of at least one component of the nanocarrier materials with temperature (Fig. [10.2](#page-4-0) and Table [10.1](#page-5-0)). Such a sharp response triggers the release of the drug following a variation in the surrounding temperature. Ideally, thermosensitive nanocarriers should retain their load at body temperature $(\sim 37 \degree C)$, and rapidly deliver the drug within a locally heated tumour $(\sim40-42$ °C) to counteract rapid bloodpassage time and washout from the tumour $[2]$. The use of temperature as a signal has been justified by the fact that the actual body temperature often deviates from the physiological value (37 \degree C) in the presence of pathogens or pyrogens. This deviation can be a useful stimulus to activate release of therapeutic agents from various temperature-responsive drug delivery systems for diseases accompanied by fever. Drug-delivery systems responsive to temperature utilize various polymer properties, including the thermally reversible transition of polymer molecules, swelling change of networks, glass transition and crystalline melting [29].

 Thermoresponsive polymers utilize subtle changes in temperature to trigger macroscopic changes in material properties. Polymers that possess a lower critical solution temperature (LCST) typically undergo a sol-gel phase transition when heated above their LCST, whereas polymers that become soluble upon heating are said to possess an upper critical solution temperature (UCST) [6]. Both systems can be exploited for drug delivery purposes. LCST copolymers can simply be mixed with drug as a liquid suspension at room temperature and delivered via minimally invasive injection techniques directly to hard-to-access target tissues within the body. Heating to physiologic temperature drives a sol-gel phase transition, which entraps the infused drug within a solid depot and can provide sustained release of therapeutic concentrations of drug directly at the site of interest [30]. Drug-releasing polymer systems possessing a UCST may employ temperatureinduced swelling or scaffold destabilisation to rapidly release drug at a target site [[31 \]](#page-20-0). Localised heating (tumour tissues) or the application of an externally applied

 stimulus (ultrasound, infrared laser and so on) may be utilised to induce the local destabilisation of a UCST drug-releasing copolymer scaffold to produce targeted release [32, [33](#page-20-0)].

Typical LCST polymers are based on *N*-isopropylacrylamide (NIPAM), *N,Ndiethylacrylamide* (DEA), methylvinylether and *N* -vinylcaprolactam (NVCL) as monomers. Some example of these categories are Poly(*N*-vinylcaprolactam) [34], Poly(*N*-isopropylacrylamide) [35], Poly(*N*,*N*-ethylmethylacrylamide) [36], Poly(*Nethylacrylamide*) [37], Poly(*N,N*-diethylacrylamide) [38]. A typical UCST system is based on a combination of acrylamide and acrylic acid $[6]$. Thermoresponsiveness can also occur on a brief temperature decrease (also called cold shock or cryotherapy). In this case, a thermally reversible swelling or de-swelling of the nanocarrier leads to free diffusion of the encapsulated drugs as a consequence of increased porosity [39]. Thermosensitive amphiphilic polymers generally have temperatureresponsive hydrophilic segments and a suitable hydrophobic segment. NIPAM and its random copolymers are the most intensively investigated temperature-sensitive hydrophilic segments [40]. Block copolymers of PEG as a hydrophilic block and NIPAM or poly(*N*-isopropylacrylamide)-co-*N*-(2-hydroxypropyl) methacrylamidedilactate as a thermosensitive block are able to self-assemble in water into temperature- responsive nanocarriers above the LCST of the thermosensitive block [41]. The hydrophobic segments, poly (*L*-lactide), cholic acid, alkyl, and poly (γ- benzyl L -glutamate) have also been used in diblock polymers with the temperature- sensitive polyacrylamide derivatives being the hydrophilic segments.

10.4.1 Delivery Systems

 Generally used thermoresponsive carriers are liposomes, or polymer micelles, nanoparticles and nanofibres $[42-48]$. For liposomes, thermoresponsiveness usually arises from a phase transition of the constituent lipids and the associated conformational variations in the lipid bilayer $[49]$. In vivo, heat is generally applied by using temperature-controlled water sacks, radiofrequency oscillators or miniature annularphased array microwave applicators. Liposome-embedded hydrogels have been widely used for controlled drug release. Liu et al. embedded egg phosphatidylcholine liposome into a poly(*N*-isopropylacrylamide) (pNIPAM) hydrogel via chemical cross-linking. It was found that the confinement of the network and the hydrophobic interactions between the liposome and pNIPAM modulated the integrity of the liposome and the release profile of the encapsulated content, such as calcein $[50]$.

 Polymeric micelles have been explored for temperature induced release of actives for drug and gene delivery. The temperature-sensitive property is possessed by the outer shell of the polymeric micelles and the drug molecules are incorporated into the hydrophobic inner core [1]. Co-polymer of pNIPAM and poly(acrylic acid) with LCST 33 °C had shown potential to be developed as novel injectable drug delivery system due to rapid sol to gel conversion upon subcutaneous injection [51].

 Chang et al. developed a block co-polymer from pNIPAM and poly(methyl methacrylate). They prepared prednisone acetate loaded uncross-linked micelles and cross-linked micelles with newly developed block co-polymer. LCST of uncross-linked and cross-linked micelles were 31.0 and 40.8 °C respectively. Uncross-linked micelles showed a rapid drug release near to 30 °C while crosslinked (SCL) micelles displayed negligible release up to 37 °C which increased rapidly above 40 °C [52]. Thermoresponsive, self-assembling polymersomes of poly(*N* -[3-aminopropyl] methacrylamide hydrochloride) and pNIPAM were also synthesised and used for similar applications [53].

Vitamin B-12 loaded nanofibres of pNIPAM/poly(ethylene oxide) (PEO) blend were also able to program drug release with the variation of temperature. Fibres containing higher ratios of pNIPAM displayed rapid release below LCST while the prolonged release was observed at 37 °C [54]. Stover and coworkers developed thermoresponsive, biodegradable linear-dendritic nanoparticles for targeted and sustained release of a pro-apoptotic drug ceramide (C6). These nanoparticles showed preferential uptake into human MDA-MB-231 breast adenocarcinoma cells at temperature above the LCST (37 °C) and sustained release of C6 up to 1 month in vitro $[55]$.

10.4.2 Applications

10.4.2.1 Cancer Therapy

 Thermoresponsive drug delivery is among the most investigated stimuli-responsive strategies, and has been widely explored in oncology. Qin et al. prepared thermoresponsive, doxorubicin-containing PEG–pNIPAM based polymersomes. Temperature induced transition facilitated self-assembly of polymer into vesicles at temperatures above 32 °C. Temperature-controlled release was determined by incorporating a hydrophobic fluorescent dye into their membrane. These vesicles destabilised, or ruptured upon local cooling with either ice or penetrating cryoprobes [56]. For the treatment of gastric cancer, linoleic acid-coupled Pluronic F-127 (Plu-CLA) based thermoresponsive hydrogel loaded with docetaxel were developed. Docetaxel–Plu-CLA showed excellent anti-tumour activity, induced apoptosis and significantly reduced the number of peritoneal metastatic nodules than docetaxel alone [57].

10.4.2.2 Anti-Adhesive

 Another application of thermosensitive polymers to inhibit ischemia-induced postoperative peritoneal adhesion was highlighted by Wu and coworkers. The PCL–PEG–PCL, developed for this purpose exhibited rapid micelle formation at 10 °C and sol to gel conversion at body temperature. They were found to be well tolerated, less toxic and therapeutically more effective as compared to control

group. Emergence of a layer of neo-mesothelial cells on the injured tissues after micelle treatment provides a strong evidence in the support of its anti-adhesion activity [58].

10.4.2.3 Temperature-Responsive Surfaces

 Temperature-responsive cell culture surfaces of pNIPAM with the ability to alter its surface hydrophobicity in response to temperature were developed. The so developed cell culture surfaces facilitated cell adhesion and proliferation at 37 °C while released spread cultured cells below 32 °C without any need of trypsin. Further, pre-coating of these surfaces with fibronectin improved spreading of less adhesive cultured hepatocytes [59]. Liao et al. developed NIPAM-based thermoresponsive polyelectrolyte multilayer films as culture substrates to support hMSC expansion. These film were via layer-by-layer adsorption of thermoresponsive polymer and positively charged allylamine hydrochloride, or negatively charged styrene sulfonic acid. Surface charge was found to alter ECM structure and subsequently cellular response for the surface. The positively charged surfaces resulted improved cell adhesion and growth compared to control surfaces [60].

10.4.2.4 Diabetes Mellitus

 For treatment of diabetes mellitus, pancreatic islet cells were harvested on laminin- 5 coated temperature-responsive dishes functional activity of the islet cell sheets was confirmed by histological examination and Insulin secretion assay prior to in vivo transplantation $[61]$. Thermoreversible hydrogel composed of poly(lactic acid-co-glycolic acid)-poly(ethylene glycol)-poly(lactic acid-co-glycolic acid) (PLGA-PEG-PLGA) triblock copolymers loaded with exenatide were also evaluated to treat diabetes. Polymer decreased the degradation of the polypeptide. Further, the problems of loading of water soluble peptide and sustaining the release of peptide were solved by synergistic effect of zinc acetate, PEG, and sucrose [62].

10.4.2.5 Ocular Therapy

Thermosensitive poly(*N*-isopropylacrylamide)–chitosan (pNIPAM–CS) solution loaded with timolol maleate was investigated for ocular application due to its in situ gel-forming properties. Polymer showed lower critical solution temperature of 32 °C, which was close to the surface temperature of the eye. Ocular pharmacokinetic analysis on rabbit eye showed higher Cmax and AUC as compared to conventional eye drop solution. Moreover, pNIPAM–CS solution showed reduced in vitro cytotoxicity and higher capacity to reduce the intra-ocular pressure as compared to conventional solution [63]

10.4.2.6 CNS Disorders

Thermo-gelling injectable nanogels amphiphilically modified chitosan were reported for delivery ethosuximide. In vivo studies suggested prominent therapeutic effect of ethosuximide loaded nanogels by suppressing spike wave discharges in Long Evan rat model [64].

10.5 Magnetically Responsive Carriers

A magnetic field-responsive nanocarrier generally involves paramagnetic or superparamagnetic materials either embedded into a polymeric scaffold forming liposo-mal, micellar, or supramolecular aggregates (Fig. [10.2](#page-4-0) and Table [10.1](#page-5-0)). The versatile intrinsic properties of magnetic particles enable their use in numerous medical applications, such as: localisation of therapy, where magnetic carriers, associated with drugs, nucleic acids or loaded within cells can be directed or guided by means of a magnetic field gradient towards certain biological targets; magnetic fluid hyperthermia, where selective thermal ablation of tumours is achieved through heating of tumour-localised magnetic particles exposed to a high frequency magnetic field; tissue engineering, where particles can be used in remote actuation for control of cellular behaviour enabling development of functional tissue or to provide means for a patterned cell assembly and facilitated seeding of tissue engineered scaffold with functional cells; and MRI, where magnetic particles are used as contrast agents $[65]$.

 Magnetic systems for magnetic targeting that have been proposed or employed so far fall into two main classes. In one class, magnets external to the body provide both the field to magnetize the carrier and field gradients for targeting $[66, 67]$. However, the use of external magnets imposes serious limitations in targeting deep tissues as their field strength and field gradient decrease exponentially with the distance from the surface. The other class is based on a combination of external magnets and magnets (or magnetizable devices) implanted local to the target region. In the second class of systems, the external magnet would typically provide the magnetizing field for the carrier, while the local magnet (or magnetisable implant) will provide the largest possible field gradients for targeting. The second type of magnet system can be of potential use for targeting deep tissues, including blood vessels where magnetizable implants can be placed $[68-70]$. The effective use of magnetically responsive nanocarriers for biomedical applications such as targeted drug delivery depends on a number of factors related to the size and magnetism of the biocompatible nanoparticles. Parameters such as the physicochemical properties of the drug-loaded nanocarriers, field strength and geometry, depth of the target tissue, rate of blood flow, and vascular supply play a role in determining the effectiveness of this method of drug delivery $[71, 72]$.

 Iron oxides with core/shell structures are the most widely used as sources of magnetic materials. Iron oxides have several crystalline polymorphs known as α -Fe₂O₃ (hematite), β -Fe₂O₃, γ -Fe₂O₃ (maghemite), ε -Fe₂O₃, Fe₃O₄ (magnetite) and some others (amorphous and high pressure forms). Nevertheless, only maghemite and magnetite found the greatest interest of bio-applications [\[73](#page-22-0)]. Readily, carbonyl iron, which is well-known material with a unique form of elemental iron because of its small particle size, was also used as magnetic core [[74 \]](#page-22-0). In some reports, pure metals, such as Fe and Co were chosen as a magnetic material because they have several advantages over iron oxides, e.g. better magnetic properties, high saturation magnetisation, and high specific loss of power $[75, 76]$.

 Functionalisation of magnetically responsive carriers with amino group, silica, polymer, various surfactants or other organic compounds is usually provided in order to achieve better physical and chemical properties. Moreover, the core/shell structures of nanocarriers have the advantages of good dispersion, high stability against oxidation and appreciable amount of drug can be loaded to the polymer shell [\[77](#page-22-0)]. Lecommandoux et al. developed magnetic nanocomposites of polypeptidebased diblock copolymers of polybutadiene-block-poly(glutamic acid) in combination with hydrophobically modified γ -Fe₂O₃ nanoparticles [78]. Furthermore, lots of functional groups from polymers on the surface can be used for further functionalisation to get various properties [79]. It is favoured that magnetically responsive carriers retain sufficient hydrophilicity and, with coating, do not exceed 100 nm in size to avoid rapid clearance by reticuloendothelial system. It was found that surface functionalisation also plays the key role in nanoparticle toxicity [80].

10.5.1 Delivery Systems

 Candidate nanosystems for such a therapeutic approach are core–shell nanoparticles (a magnetic core made of magnetite coated with silica or polymer) $[81, 82]$ $[81, 82]$ $[81, 82]$, magnetoliposomes (maghemite nanocrystals encapsulated in liposomes) [\[83 ,](#page-22-0) [84 \]](#page-22-0) and porous metallic nanocapsules [85] (Fig. [10.2](#page-4-0)). A novel nanocarrier, containing functionalised magnetite (Fe_3O_4) core that was conjugated with drug via acid-labile hydrazone-bond and encapsulated by the thermosensitive chitosan-g-poly(N-isopropylacrylamide-co-*N*, *N*-dimethylacrylamide) was reported. Polymer exhibited a LCST of 38 °C below which the drug release response was appreciably low $[86]$. Antibody-conjugated magnetoliposomes for targeting cancer cells were also reported [87].

10.5.2 Applications

Magnetically responsive carriers are getting significant attention in the field of theranostics. Theranostics is the fusion of therapeutics and diagnostics to design individualised pharmacotherapy. Paramagnetic nanoparticles were initially used as contrast agents for magnetic resonance imaging (MRI) later on surface modification of these nanoparticles introduced various functions for the nanoparticles to be used for both gene delivery and MR imaging. The combination based on the nanoparticles allows non-invasive monitoring of in vivo gene delivery with MRI and delivery of therapeutic genes [\[88](#page-23-0) , [89 \]](#page-23-0). Such magnetofection experiments were generally performed using nanoassemblies with cationic coatings to condense nucleic acids, which resulted in higher transfection efficiencies under a permanent magnetic field. These technique led to improved effectiveness in the transfection of siRNA in vitro and/or in vivo when directed against prostate $[90]$ and breast cancers $[91]$, as well as in the gene transfer to oligodendrocyte precursors for neural repair [92].

10.6 Recent Advancements

10.6.1 Dual and Multi Stimuli Responsive Carriers

In an effort to further fine tune drug release and augment therapeutic efficacy of nanoparticulate drugs, sophisticated polymeric nanoparticles that respond to dual and multi-stimuli such as pH/temperature, pH/redox, pH/magnetic field, temperature/reduction, double pH, pH and diols, temperature/magnetic field, temperature/ enzyme, temperature/pH/redox, temperature/pH/magnetic, pH/redox/magnetic, temperature/redox/guest molecules and temperature/pH/guest molecules have been aggressively pursued (Fig. 10.2) [93–96]. It should be noted that the responses take place either simultaneously at the same location or in a sequential manner in different settings and/or compartments. These dual and multi-stimuli responsive polymeric nanoparticles might on one hand offer unprecedented control over drug delivery and release leading to superior in vitro and/or in vivo anti-cancer potency, and on the other hand also facilitate nanoparticles preparation and loading of drugs under mild conditions $[97]$. These two and more stimuli are combined in order to: (1) facilitate preparation of nanoparticles under mild conditions through application of an external stimulus such as temperature and pH; (2) trigger drug release via application of an external stimulus such as magnetic field, ultrasonic, light and temperature; (3) trigger drug release or reverse deshielding of nanoparticles thereby enhancing tumour cell uptake of nanoparticulate drugs in a mildly acidic tumour microenvironment; and/or (4) boost intracellular drug release in tumour cells under endo/lysosomal pH and/or cytosolic reductive conditions.

 Shim et al. developed a polymer containing sulfamethazine as the pH-responsive component, and $poly(\varepsilon$ -caprolactone-co-lactide) (PCLA) in a triblock with PEG, PCLA–PEG–PCLA, as the thermosensitive moiety [98]. By controlling precise ratios between the two parts the co-polymer showed a reversible sol–gel–sol transition phase. At room temperature and pH 8 the polymer remained in a solution state, when the environment was altered to 37 \degree C and pH 7.4, i.e. normal physiological conditions, there was a rapid phase transition to a gel state. Once the gel was formed it remained stable and degraded over time without changing local pH levels [98, 99]. Despite the advantageous versatility of these systems, they often appear as too

Table 10.2 Clinical trials for thermal and magnetically responsive carriers **Table 10.2** Clinical trials for thermal and magnetically responsive carriers

c omplicated and many still remain as proofs of concept. To ascertain the viability of these strategies, evidence of the regulation of the response to each stimulus would be needed both in vitro and in vivo.

10.6.2 Breathing Vesicles

The breathing, in this context, can be defined as a highly reversible vesicle volume change by a factor of approximately 7, which was accompanied by diffusion of s pecies into and out of the vesicles with a relaxation time of approximately 1 min. A three-layered vesicle system with pH-induced "breathing" feature was designed with triblock copolymer poly(ethylene oxide)-block-polystyrene-block-poly (2- diethylaminoethylmethacrylate). Self-assembly into vesicles was observed at a pH of 10.4. As the pH decreased, both the vesicle size and the thickness of all three layers increased. Progressive swelling of the middle layer with a decrease in pH below 6 induced cracking of the two outer layers and also a sharp increase of the vesicle size and the wall thickness. When pH reached up to 3.4, the vesicle size was found to be increased by a factor of 1.9 and the wall showed a cracked surface. These changes between pH 10.4 and 3.4 were highly reversible with the relaxation time of 1 min with marked repeatedly. The change in the wall structure dramatically helped to increase the wall permeability to water along with rate of proton diffusion from practically zero to extremely rapid $[100]$.

Similarly, CO₂-responsive breathing vesicles were synthesised with block polymer poly(ethylene oxide)-poly(N-amidino)dodecyl acrylamide (PEO-b-PAD). PEO-b-PAD self-assembled in to micelle like structure in which the PEO portion formed hydrophilic exterior and the PAD portion formed hydrophobic interior. The amidine group in copolymer transformed into a charged amidinium species upon reaction with $CO₂$ which reverted back to its original form upon exposure to argon (Ar) . It was confirmed by much larger intact vesicles with a diameter of 205.25 nm and strikingly increases volume by 83.5 $%$ after CO₂ treatment for 20 min. These vesicles shrunk back to their initial size in the presence of Ar [\[101](#page-23-0)].

10.7 Challenges

 Despite their responsive feature and versatility for drug delivery, thermoresponsive systems have some unanswered questions too. Co-polymeric systems provided ease to modulate the system properties but they are facing the problem of biocompatibility, biodegradability, reproducibility and tailored drug release as per requirement of site of action $[51, 99]$ $[51, 99]$ $[51, 99]$. Development of amphiphilic poly(asparagine) based polymers, Poly[α/β-(DL-aspartate isopropylamide)-co-(succinimide)] PCL-PEG copolymer had solved the biodegradability issue up to certain extent still, more in vivo studies are needed to ascertain their complete safety [58, 102-104].

10.8 Clinical Status

 Nanocarriers that are responsive to exogenous stimuli (temperature and magnetic field) have reached clinical stage as they are more promising. Endogenous triggers are indeed difficult to control because they may vary from one patient to another (such as the pH of a tumour or the presence of reducing agents in the blood circulation). Table [10.2](#page-15-0) gives information regarding responsive carriers that are in different phases of clinical trials.

10.9 Conclusions

 With greater understanding of physiological differences between normal and disease tissues and advances in material design, there is an opportunity to develop nanocarrier systems for target-specific drug delivery that will respond to local stimuli. They can shift the paradigm in the delivery of medicine and diagnostics. This chapter explained the role of pH, temperature and magnetic field responsive nanocarriers for targeted-drug delivery. However, compared to the amount of research being done in the field, relatively few medical nanotechnologies have made it to the market. Clear demonstrations of biocompatibility and including biodegradable components will make these materials even more attractive for in vivo applications. Furthermore, development and implementation of scalable, cost-effective fabrication techniques will help promote clinical translation. Together, intelligent materials and nanocarriers provide a versatile toolbox that we believe will revolutionize the future of modern medicine.

References

- 1. Ganta S, Devalapally H, Shahiwala A, Amiji M (2008) A review of stimuli-responsive nanocarriers for drug and gene delivery. J Control Release 126:187–204
- 2. Mura S, Nicolas J, Couvreur P (2013) Stimuli-responsive nanocarriers for drug delivery. Nat Mater 12:991–1003
- 3. Yatvin MB, Weinstein JN, Dennis WH, Blumenthal R (1978) Design of liposomes for enhanced local release of drugs by hyperthermia. Science 202:1290–1293
- 4. Fleige E, Quadir MA, Haag R (2012) Stimuli-responsive polymeric nanocarriers for the controlled transport of active compounds: concepts and applications. Adv Drug Deliv Rev 64:866–884
- 5. Mellman I, Fuchs R, Helenius A (1986) Acidification of the endocytic and exocytic pathways. Annu Rev Biochem 55:663–700
- 6. Schmaljohann D (2006) Thermo-and pH-responsive polymers in drug delivery. Adv Drug Deliv Rev 58:1655–1670
- 7. Yin Q, Shen J, Zhang Z, Yu H, Li Y (2013) Reversal of multidrug resistance by stimuliresponsive drug delivery systems for therapy of tumor. Adv Drug Deliv Rev 65:1699–1715
- 8. Tang R, Ji W, Panus D, Palumbo RN, Wang C (2011) Block copolymer micelles with acidlabile ortho ester side-chains: synthesis, characterization, and enhanced drug delivery to human glioma cells. J Control Release 151:18–27
- 9. Gao W, Chan JM, Farokhzad OC (2010) pH-responsive nanoparticles for drug delivery. Mol Pharm 7:1913–1920
- 10. Zhang L, Guo R, Yang M, Jiang X, Liu B (2007) Thermo and pH dual-responsive materials for controllable oil/water separation. Adv Mater 19:2988–2992
- 11. Pistolis G, Malliaris A, Tsiourvas D, Paleos CM (1999) Poly(propyleneimine) dendrimers as pH-sensitive controlled-release systems. Chem Eur J 5:1440–1444
- 12. Hui H, Xiao dong F, Zhong lin C (2005) Thermo-and pH-sensitive dendrimer derivatives with a shell of poly (N, N-dimethylaminoethyl methacrylate) and study of their controlled drug release behavior. Polymer 46:9514–9522
- 13. Yuan H, Luo K, Lai Y, Pu Y, He B, Wang G, Wu Y, Gu Z (2010) A novel poly (L-glutamic acid) dendrimer based drug delivery system with both pH-sensitive and targeting functions. Mol Pharm 7:953–962
- 14. Sudimack JJ, Guo W, Tjarks W, Lee RJ (2002) A novel pH-sensitive liposome formulation containing oleyl alcohol. Biochim Biophys Acta 1564:31–37
- 15. Torchilin VP, Zhou F, Huang L (1993) pH-sensitive liposomes. J Liposome Res 3:201–255
- 16. Giacomelli C, Schmidt V, Borsali R (2007) Nanocontainers formed by self-assembly of poly (ethylene oxide)-b-poly (glycerol monomethacrylate)-drug conjugates. Macromolecules 40:2148–2157
- 17. Dai J, Nagai T, Wang X, Zhang T, Meng M, Zhang Q (2004) pH-sensitive nanoparticles for improving the oral bioavailability of cyclosporine A. Int J Pharm 280:229–240
- 18. Wang XQ, Dai JD, Chen Z, Zhang T, Xia GM, Nagai T, Zhang Q (2004) Bioavailability and pharmacokinetics of cyclosporine A-loaded pH-sensitive nanoparticles for oral administration. J Control Release 97:421–429
- 19. Borchert U, Lipprandt U, Bilang M, Kimpfler A, Rank A, Peschka-Suss R, Schubert R, Lindner P, Forster S (2006) pH-induced release from P2VP-PEO block copolymer vesicles. Langmuir 22:5843–5847
- 20. Du J, Armes SP (2005) pH-responsive vesicles based on a hydrolytically self-cross-linkable copolymer. J Am Chem Soc 127:12800–12801
- 21. Hong MS, Lim SJ, Oh YK, Kim CK (2002) pH-sensitive, serum-stable and long-circulating liposomes as a new drug delivery system. J Pharm Pharmacol 54:51–58
- 22. Zhang Q, Tang J, Fu L, Ran R, Liu Y, Yuan M, He Q (2013) A pH-responsive helical cell penetrating peptide-mediated liposomal delivery system. Biomaterials 34:7980–7993
- 23. Radovic-Moreno AF, Lu TK, Puscasu VA, Yoon CJ, Langer R, Farokhzad OC (2012) Surface charge-switching polymeric nanoparticles for bacterial cell wall-targeted delivery of antibiotics. ACS Nano 6:4279–4287
- 24. Karanth H, Murthy RSR (2007) pH-Sensitive liposomes-principle and application in cancer therapy. J Pharm Pharmacol 59:469–483
- 25. Wang X-Q, Zhang Q (2012) pH-sensitive polymeric nanoparticles to improve oral bioavailability of peptide/protein drugs and poorly water-soluble drugs. Eur J Pharm Biopharm 82:219–229
- 26. Qu W, Li Y, Hovgaard L, Li S, Dai W, Wang J, Zhang X, Zhang Q (2011) A silica-based pHsensitive nanomatrix system improves the oral absorption and efficacy of incretin hormone glucagon-like peptide-1. Int J Nanomedicine 7:4983–4994
- 27. Yang YQ, Zheng LS, Guo XD, Qian Y, Zhang LJ (2010) pH-sensitive micelles self- assembled from amphiphilic copolymer brush for delivery of poorly water-soluble drugs. Biomacromolecules 12:116–122
- 28. Legendre J-Y, Szoka FC Jr (1992) Delivery of plasmid DNA into mammalian cell lines using pH-sensitive liposomes: comparison with cationic liposomes. Pharm Res 9: 1235–1242
- 29. Bajpai AK, Shukla SK, Bhanu S, Kankane S (2008) Responsive polymers in controlled drug delivery. Prog Polym Sci 33:1088–1118
- 30. Li Z, Guan J (2011) Thermosensitive hydrogels for drug delivery. Expert Opin Drug Deliv 8:991–1007
- 31. Hoogenboom R, Lambermont-Thijs HML, Jochems MJHC, Hoeppener S, Guerlain C, Fustin C-A, Gohy J-F, Schubert US (2009) A schizophrenic gradient copolymer: switching and reversing poly (2-oxazoline) micelles based on UCST and subtle solvent changes. Soft Matter 5:3590–3592
- 32. Hirsch LR, Stafford RJ, Bankson JA, Sershen SR, Rivera B, Price RE, Hazle JD, Halas NJ, West JL (2003) Nanoshell-mediated near-infrared thermal therapy of tumors under magnetic resonance guidance. Proc Natl Acad Sci 100:13549–13554
- 33. Rapoport N (2007) Physical stimuli-responsive polymeric micelles for anti-cancer drug delivery. Prog Polym Sci 32:962–990
- 34. Laukkanen A, Valtola L, Winnik FM, Tenhu H (2004) Formation of colloidally stable phase separated poly (N-vinylcaprolactam) in water: a study by dynamic light scattering, microcalorimetry, and pressure perturbation calorimetry. Macromolecules 37:2268–2274
- 35. Schild HG, Tirrell DA (1990) Microcalorimetric detection of lower critical solution temperatures in aqueous polymer solutions. J Phys Chem 94:4352–4356
- 36. Cao Y, Zhu XX, Luo J, Liu H (2007) Effects of substitution groups on the RAFT polymerization of N-alkylacrylamides in the preparation of thermosensitive block copolymers. Macromolecules 40:6481–6488
- 37. Liu HY, Zhu XX (1999) Lower critical solution temperatures of N-substituted acrylamide copolymers in aqueous solutions. Polymer 40:6985–6990
- 38. Geever LM, Lyons JG, Higginbotham CL (2011) Photopolymerisation and characterisation of negative temperature sensitive hydrogels based on N, N-diethylacrylamide. J Mater Sci 46:509–517
- 39. Lee SH, Choi SH, Kim SH, Park TG (2008) Thermally sensitive cationic polymer nanocapsules for specific cytosolic delivery and efficient gene silencing of siRNA: swelling induced physical disruption of endosome by cold shock. J Control Release 125:25–32
- 40. Yoshida R, Uchida K, Kaneko Y, Sakai K, Kikuchi A, Sakurai Y, Okano T (1995) Comb-type grafted hydrogels with rapid deswelling response to temperature changes. Nature 374:240–242
- 41. Shenoy DB, Amiji MM (2005) Poly(ethylene oxide)-modified poly(ε -caprolactone) nanoparticles for targeted delivery of tamoxifen in breast cancer. Int J Pharm 293: 261–270
- 42. Abulateefeh SR, Spain SG, Thurecht KJ, Aylott JW, Chan WC, Garnett MC, Alexander C (2013) Enhanced uptake of nanoparticle drug carriers via a thermoresponsive shell enhances cytotoxicity in a cancer cell line. Biomater Sci 1:434–442
- 43. Cammas S, Suzuki K, Sone C, Sakurai Y, Kataoka K, Okano T (1997) Thermo-responsive polymer nanoparticles with a core-shell micelle structure as site-specific drug carriers. J Control Release 48:157–164
- 44. Chen M, Dong M, Havelund R, Regina VR, Meyer RL, Besenbacher F, Kingshott P (2010) Thermo-responsive core-sheath electrospun nanofibers from poly (n-isopropylacrylamide)/ polycaprolactone blends. Chem Mater 22:4214–4221
- 45. Kryuchkov VA, Daigle JC, Skupov KM, Claverie JP, Winnik FM (2010) Amphiphilic polyethylenes leading to surfactant-free thermoresponsive nanoparticles. J Am Chem Soc 132:15573–15579
- 46. McDaniel JR, Dewhirst MW, Chilkoti A (2013) Actively targeting solid tumours with thermoresponsive drug delivery systems that respond to mild hyperthermia. Int J Hyperth 29:501–510
- 47. Okuzaki H, Kobayashi K, Yan H (2009) Thermo-responsive nanofiber mats. Macromolecules 42:5916–5918
- 48. Wang C, Flynn NT, Langer R (2004) Controlled structure and properties of thermoresponsive nanoparticle-hydrogel composites. Adv Mater 16:1074–1079
- 49. Culver HR, Daily AM, Khademhosseini A, Peppas NA (2014) Intelligent recognitive systems in nanomedicine. Curr Opin Chem Eng 4:105–113
- 50. Liu Y, Li Z, Liang D (2012) Behaviors of liposomes in a thermo-responsive poly (N-isopropylacrylamide) hydrogel. Soft Matter 8:4517–4523
- 51. Hu Z, Xia X (2004) Hydrogel nanoparticle dispersions with inverse thermoreversible gelation. Adv Mater 16:305–309
- 52. Chang C, Wei H, Wu DQ, Yang B, Chen N, Cheng SX, Zhang XZ, Zhuo RX (2011) Thermoresponsive shell cross-linked PMMA-b-P(NIPAAm-co-NAS) micelles for drug delivery. Int J Pharm 420:333–340
- 53. Li Y, Lokitz BS, McCormick CL (2006) Thermally responsive vesicles and their structural "locking" through polyelectrolyte complex formation. Angew Chem Int Ed 45:5792–5795
- 54. Song F, Wang X-L, Wang Y-Z (2011) Poly (N-isopropylacrylamide)/poly (ethylene oxide) blend nanofibrous scaffolds: Thermo-responsive carrier for controlled drug release. Colloids Surf B: Biointerfaces 88:749–754
- 55. Stover TC, Kim YS, Lowe TL, Kester M (2008) Thermoresponsive and biodegradable lineardendritic nanoparticles for targeted and sustained release of a pro-apoptotic drug. Biomaterials 29:359–369
- 56. Qin S, Geng Y, Discher DE, Yang S (2006) Temperature-controlled assembly and release from polymer vesicles of poly (ethylene oxide)-block-poly (N-isopropylacrylamide). Adv Mater 18:2905–2909
- 57. Bae WK, Park MS, Lee JH, Hwang JE, Shim HJ, Cho SH, Kim DE, Ko HM, Cho CS, Park IK (2013) Docetaxel-loaded thermoresponsive conjugated linoleic acid-incorporated poloxamer hydrogel for the suppression of peritoneal metastasis of gastric cancer. Biomaterials 34:1433–1441
- 58. Wu Q, Li L, Wang N, Gao X, Wang B, Liu X, Qian Z, Wei Y, Gong C (2014) Biodegradable and thermosensitive micelles inhibit ischemia-induced postoperative peritoneal adhesion. Int J Nanomedicine 9:727–734
- 59. Yamato M, Konno C, Kushida A, Hirose M, Utsumi M, Kikuchi A, Okano T (2000) Release of adsorbed fibronectin from temperature-responsive culture surfaces requires cellular activity. Biomaterials 21:981–986
- 60. Liao T, Moussallem MD, Kim J, Schlenoff JB, Ma T (2010) N-isopropylacrylamide-based thermoresponsive polyelectrolyte multilayer films for human mesenchymal stem cell expansion. Biotechnol Prog 26:1705–1713
- 61. Shimizu H, Ohashi K, Utoh R, Ise K, Gotoh M, Yamato M, Okano T (2009) Bioengineering of a functional sheet of islet cells for the treatment of diabetes mellitus. Biomaterials 30:5943–5949
- 62. Li K, Yu L, Liu X, Chen C, Chen Q, Ding J (2013) A long-acting formulation of a polypeptide drug exenatide in treatment of diabetes using an injectable block copolymer hydrogel. Biomaterials 34:2834–2842
- 63. Cao Y, Zhang C, Shen W, Cheng Z, Yu LL, Ping Q (2007) Poly(N-isopropylacrylamide) chitosan as thermosensitive in situ gel-forming system for ocular drug delivery. J Control Release 120:186–194
- 64. Hsiao MH, Larsson M, Larsson A, Evenbratt H, Chen YY, Chen YY, Liu DM (2012) Design and characterization of a novel amphiphilic chitosan nanocapsule-based thermo-gelling biogel with sustained in vivo release of the hydrophilic anti-epilepsy drug ethosuximide. J Control Release 161:942–948
- 65. Sensenig R, Sapir Y, MacDonald C, Cohen S, Polyak B (2012) Magnetic nanoparticle-based approaches to locally target therapy and enhance tissue regeneration in vivo. Nanomedicine 7:1425–1442
- 66. Alexiou C, Schmid RJ, Jurgons R, Kremer M, Wanner G, Bergemann C, Huenges E, Nawroth T, Arnold W, Parak FG (2006) Targeting cancer cells: magnetic nanoparticles as drug carriers. Eur Biophys J 35:446–450
- 67. Lubbe AS, Alexiou C, Bergemann C (2001) Clinical applications of magnetic drug targeting. J Surg Res 95:200–206
- 68. Chorny M, Fishbein I, Yellen BB, Alferiev IS, Bakay M, Ganta S, Adamo R, Amiji M, Friedman G, Levy RJ (2010) Targeting stents with local delivery of paclitaxel-loaded magnetic nanoparticles using uniform fields. Proc Natl Acad Sci 107:8346–8351
- 69. Pislaru SV, Harbuzariu A, Agarwal G, Witt T, Gulati R, Sandhu NP, Mueske C, Kalra M, Simari RD, Sandhu GS (2006) Magnetic forces enable rapid endothelialization of synthetic vascular grafts. Circulation 114:I-314–I-318
- 70. Yellen BB, Forbes ZG, Halverson DS, Fridman G, Barbee KA, Chorny M, Levy R, Friedman G (2005) Targeted drug delivery to magnetic implants for therapeutic applications. J Magn Magn Mater 293:647–654
- 71. Chomoucka J, Drbohlavova J, Huska D, Adam V, Kizek R, Hubalek J (2010) Magnetic nanoparticles and targeted drug delivering. Pharmacol Res 62:144–149
- 72. Sun C, Lee JSH, Zhang M (2008) Magnetic nanoparticles in MR imaging and drug delivery. Adv Drug Deliv Rev 60:1252–1265
- 73. Tucek J, Zboril R, Petridis D (2006) Maghemite nanoparticles by view of Mössbauer spectroscopy. J Nanosci Nanotechnol 6:926–947
- 74. Reshmi G, Mohan Kumar P, Malathi M (2009) Preparation, characterization and dielectric studies on carbonyl iron/cellulose acetate hydrogen phthalate core/shell nanoparticles for drug delivery applications. Int J Pharm 365:131–135
- 75. Arias J, Loez-Viota M, Lopez-Viota J, Delgado AV (2009) Development of iron/ethylcellulose (core/shell) nanoparticles loaded with diclofenac sodium for arthritis treatment. Int J Pharm 382:270–276
- 76. Pouponneau P, Leroux J-C, Martel S (2009) Magnetic nanoparticles encapsulated into biodegradable microparticles steered with an upgraded magnetic resonance imaging system for tumor chemoembolization. Biomaterials 30:6327–6332
- 77. Hu FX, Neoh KG, Kang ET (2006) Synthesis and in vitro anti-cancer evaluation of tamoxifenloaded magnetite/PLLA composite nanoparticles. Biomaterials 27:5725–5733
- 78. Lecommandoux S, Sandre O, Chécot F, Rodriguez-Hernandez J, Perzynski R (2005) Magnetic nanocomposite micelles and vesicles. Adv Mater 17:712–718
- 79. Parvin S, Matsui J, Sato E, Miyashita T (2007) Side-chain effect on Langmuir and Langmuir– Blodgett film properties of poly(N-alkylmethacrylamide)-coated magnetic nanoparticle. J Colloid Interface Sci 313:128–134
- 80. Shubayev VI, Pisanic Ii TR, Jin S (2009) Magnetic nanoparticles for theragnostics. Adv Drug Deliv Rev 61:467–477
- 81. Hua M-Y, Liu H-L, Yang H-W, Chen P-Y, Tsai R-Y, Huang C-Y, Tseng I, Lyu L-A, Ma C-C, Tang H-J (2011) The effectiveness of a magnetic nanoparticle-based delivery system for BCNU in the treatment of gliomas. Biomaterials 32:516–527
- 82. Zhang L, Wang T, Yang L, Liu C, Wang C, Liu H, Wang YA, Su Z (2012) General route to multifunctional uniform yolk/mesoporous silica shell nanocapsules: a platform for simultaneous cancer-targeted imaging and magnetically guided drug delivery. Chemistry 18: 12512–12521
- 83. De Cuyper M, Joniau M (1988) Magnetoliposomes. Eur Biophys J 15:311–319
- 84. Plassat V, Wilhelm C, Marsaud V, Manager C, Gazeau F, Renoir JM, Lesieur S (2011) Antiestrogen- loaded superparamagnetic liposomes for intracellular magnetic targeting and treatment of breast cancer tumors. Adv Funct Mater 21:83–92
- 85. Zhang F, Braun GB, Pallaoro A, Zhang Y, Shi Y, Cui D, Moskovits M, Zhao D, Stucky GD (2012) Mesoporous multifunctional upconversion luminescent and magnetic "nanorattle" materials for targeted chemotherapy. Nano Lett 12:61–67
- 86. Yuan Q, Venkatasubramanian R, Hein S, Misra RDK (2008) A stimulus-responsive magnetic nanoparticle drug carrier: magnetite encapsulated by chitosan-grafted-copolymer. Acta Biomater 4:1024–1037
- 87. Shinkai M, Suzuki M, Iijima S, Kobayashi T (1995) Antibody-conjugated magnetoliposomes for targeting cancer cells and their application in hyperthermia. Biotechnol Appl Biochem 21:125–137
- 88. Ma X, Zhao Y, Liang XJ (2011) Theranostic nanoparticles engineered for clinic and pharmaceutics. Acc Chem Res 44:1114-1122
- 89. Wu X, Hu J, Zhou L, Mao Y, Yang B, Gao L, Xie R, Xu F, Zhang D, Liu J (2008) In vivo tracking of superparamagnetic iron oxide nanoparticle-labeled mesenchymal stem cell tropism to malignant gliomas using magnetic resonance imaging. J Neurosurg 108:320–329
- 90. Park JW, Bae KH, Kim C, Park TG (2010) Clustered magnetite nanocrystals cross-linked with PEI for efficient siRNA delivery. Biomacromolecules 12:457-465
- 91. Prijic S, Prosen L, Cemazar M, Scancar J, Romih R, Lavrencak J, Bregar VB, Coer A, Krzan M, Znidarsic A (2012) Surface modified magnetic nanoparticles for immuno-gene therapy of murine mammary adenocarcinoma. Biomaterials 33:4379–4391
- 92. Jenkins SI, Pickard MR, Granger N, Chari DM (2011) Magnetic nanoparticle-mediated gene transfer to oligodendrocyte precursor cell transplant populations is enhanced by magnetofection strategies. ACS Nano 5:6527–6538
- 93. Chiang WH, Ho VT, Huang WC, Huang YF, Chern CS, Chiu HC (2012) Dual stimuliresponsive polymeric hollow nanogels designed as carriers for intracellular triggered drug release. Langmuir 28:15056–15064
- 94. Jeong B, Gutowska A (2002) Lessons from nature: stimuli-responsive polymers and their biomedical applications. Trends Biotechnol 20:305–311
- 95. Morimoto N, Qiu X-P, Winnik FM, Akiyoshi K (2008) Dual stimuli-responsive nanogels by self-assembly of polysaccharides lightly grafted with thiol-terminated poly (N-isopropylacrylamide) chains. Macromolecules 41:5985–5987
- 96. Pan Y-J, Chen Y-Y, Wang D-R, Wei C, Guo J, Lu D-R, Chu C-C, Wang C-C (2012) Redox/pH dual stimuli-responsive biodegradable nanohydrogels with varying responses to dithiothreitol and glutathione for controlled drug release. Biomaterials 33:6570–6579
- 97. Cheng R, Meng F, Deng C, Klok H-A, Zhong Z (2013) Dual and multi-stimuli responsive polymeric nanoparticles for programmed site-specific drug delivery. Biomaterials 34: 3647–3657
- 98. Shim WS, Yoo JS, Bae YH, Lee DS (2005) Novel injectable pH and temperature sensitive block copolymer hydrogel. Biomacromolecules 6:2930–2934
- 99. Chan A, Orme RP, Fricker RA, Roach P (2005) Remote and local control of stimuli responsive materials for therapeutic applications. Adv Drug Deliv Rev 65:497–514
- 100. Yu S, Azzam T, Rouiller I, Eisenberg A (2009) "Breathing" vesicles. J Am Chem Soc 131:10557–10566
- 101. Yan Q, Zhou R, Fu C, Zhang H, Yin Y, Yuan J (2011) CO2-responsive polymeric vesicles that breathe. Angew Chem 123:5025–5029
- 102. Tanimoto F, Kitamura Y, Ono T, Yoshizawa H (2010) A versatile biodegradable polymer with a thermo-reversible/irreversible transition. ACS Appl Mater Interfaces 2:606–610
- 103. Watanabe E, Boutis GS, Sato H, Sekine S, Asakura T (2014) NMR studies of thermoresponsive behavior of an amphiphilic poly (asparagine) derivative in water. Polymer 55:278–286
- 104. Watanabe E, Tomoshige N, Uyama H (2007) New biodegradable and thermoresponsive polymers based on amphiphilic poly (asparagine) derivatives. Macromolecular symposia. Wiley Online Library. pp 509–514
- 105. McCarthy JR (2009) The future of theranostic nanoagents. Nanomedicine 4:693–695
- 106. Checot F, Lecommandoux S, Klok HA, Gnanou Y (2003) From supramolecular polymersomes to stimuli-responsive nano-capsules based on poly (diene-b-peptide) diblock copolymers. Eur Phys J E 10:25–35
- 107. Kukula H, Schlaad H, Antonietti M, Forster S (2002) The formation of polymer vesicles or "Peptosomes" by polybutadiene-block-poly (L-glutamate)s in dilute aqueous solution. J Am Chem Soc 124:1658–1663
- 108. Bellomo EG, Wyrsta MD, Pakstis L, Pochan DJ, Deming TJ (2004) Stimuli-responsive polypeptide vesicles by conformation-specific assembly. Nat Mater 3:244–248
- 109. Lomas H, Canton I, MacNeil S, Du J, Armes SP, Ryan AJ, Lewis AL, Battaglia G (2007) Biomimetic pH sensitive polymersomes for efficient DNA encapsulation and delivery. Adv Mater 19:4238–4243
- 110. Checot F, Rodriguez Hernandez J, Gnanou Y, Lecommandoux S (2006) Responsive micelles and vesicles based on polypeptide diblock copolymers. Polymer Adv Tech 17:782–785
- 111. Chen X, Ding X, Zheng Z, Peng Y (2006) Thermosensitive cross-linked polymer vesicles for controlled release system. New J Chem 30:577–582
- 112. Hales M, Barner Kowollik C, Davis TP, Stenzel MH (2004) Shell-cross-linked vesicles synthesized from block copolymers of poly (D, L-lactide) and poly (N-isopropyl acrylamide) as thermoresponsive nanocontainers. Langmuir 20:10809–10817
- 113. Hastings CL, Kelly HM, Murphy MJ, Barry FP, O Brien FJ, Duffy GP (2012) Development of a thermoresponsive chitosan gel combined with human mesenchymal stem cells and desferrioxamine as a multimodal pro-angiogenic therapeutic for the treatment of critical limb ischaemia. J Control Release 161:73–80
- 114. Licciardi M, Amato G, Cappelli A, Paolino M, Giuliani G, Belmonte B, Guarnotta C, Pitarresi G, Giammona G (2012) Evaluation of thermoresponsive properties and biocompatibility of polybenzofulvene aggregates for leuprolide delivery. Int J Pharm 438:279–286
- 115. Krack M, Hohenberg H, Kornowski A, Lindner P, Weller H, Forster S (2008) Nanoparticleloaded magnetophoretic vesicles. J Am Chem Soc 130:7315–7320