Chapter 17 Biotechnological Aspects of Soil Decontamination

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Abstract Soils have been subjected to several contaminants that vary in concentration and composition. Soil pollution causes significant damage to the environment and human health as a result of their mobility and solubility. Significant progress has been made in regulating soil pollution, with a parallel development of methodologies for soil assessment and remediation. The selection of most appropriate soil and sediment remediation method depends on the site characteristics, concentration, type of pollutants to be removed, and the end use of the contaminated medium. This chapter provides the developing biotechnological aspects of soil decontamination. The study also reviews other available remediation options, which includes physical, chemical, and thermal technologies. All these technologies may be used in conjunction with one another to reduce the contamination to an acceptable level, and may offer potential technical solution to most soil pollution.

17.1 Introduction

Globally, the increasing human population, industrial revolution, and the number of anthropogenic inefficiencies with unplanned growth of urban system have been putting an intense pressure on the consumption of natural resources, thus threatening human health and the environment. Over time, the quantities of these

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nonrecyclable toxic by-products are at the levels that present an unacceptable risk to the sensitive biosphere (Sheoran et al. 2008; Sikdar et al. 1998)

Historically, soil pollution has been of very little concern, and its contamination is a relatively recent issue when, actually, it began a long time ago, particularly after the industrial revolution, even though only recently has mankind become aware of its dimension, persistence, and harmful effects (Castelo-Grande et al. 2010). Pollutants can be built up in the soil directly or indirectly from several sources, such as industrial emissions, mining and smelting of metalliferous ore, electroplating energy and fuel production, and sludge dumping. A wide range of inorganic and organic compounds cause contamination, and these include heavy metals, combustible and putrescible substances, hazardous wastes, explosives, and petroleum products. There has been increasing concern over the last few decades regarding organic pollutants, which includes PCBs such as dioxin, polycyclic aromatic hydrocarbons (PAHs) such as benzoapyrene, nitroaromatics such as trinitrotoluene (TNT), and linear halogenated hydrocarbons such as trichloroethylene (TCE). Major components of inorganic contaminants are heavy metals (Ghosh and Singh 2005). Heavy metals cannot be destroyed biologically and are present in soils as free metal ions, soluble metal complexes (sequestered to ligands), exchangeable metal ions, organically bound metals, precipitated or insoluble compounds such as oxides, carbonates, and hydroxides, or they may form part of the structure of silicate materials (indigenous soil content) (Davies et al. 2001).

Contaminated soils are the target of several thousands of pollutants that vary in their compositions and concentration. Elevated concentration of these pollutants impair the functioning of human population and pose significant risk to the development of flora and fauna (Scullion 2006). Contaminated soil fails to support crop growth and negatively affects its development because of interference of phytotoxic contaminants with metabolic processes and sometimes also leading to plant death (Hoffmann 1983). Besides this, soil toxicity disrupts biological cycling of nutrients and also affects the hydrosphere compromising with the quality of drinking water resources, and threatening the aquatic ecosystem (Bilek 2004). Human beings are also at risk from polluted soils, thus the magnitude of pollution in our soils calls for immediate action (Friberg et al. 1986; Knasmuller et al. 1998; Nathanail and Earl 2001). When contaminant concentrations in soil are too high for natural biodegradation to occur, cleanup action is warranted (Sikdar et al. 1998). Thus, in response to a growing need to address this environmental contamination, many remediation technologies have been developed to treat soil contaminated by various pollutants, including in situ (treating relatively undisturbed soils) and ex situ (treating excavated soils) methods either onsite or in designated soil treatment facilities. In situ has the advantage of minimal disruption to activities on site or on adjacent land. Ex situ approaches generally offer greater scope for managing conditions to optimize treatment efficiency and for controlling potential spread of pollutants. In situ methods are favored over the ex situ techniques due to their low cost and reduced impact on the ecosystem (Reed et al. 1992).

Biological, physical, chemical, and other technologies can be used in conjunction with one another to reduce the contamination to a safe and acceptable level (RAAG 2000). Physical methods employs soil washing, encapsulation, and solidification; precipitation and ion exchange are chemical treatments, and for the biological treatment plants are used. Even though many technologies are available for the decontamination of polluted sites, the selection depends on contaminant and site-by-site basis, regulatory requirements, costs, and time constraints. Since most remediation techniques are site-specific, the selection of appropriate technology is often a difficult, but extremely important step in the successful remediation of a contaminated site. Therefore, the successful decontamination of a contaminated site depends on proper selection of the methodology, its design, and adjustment of the remediation technology's operations based on properties of the contaminants and soils, and on the performance of the whole system (USEPA 1998; Khan et al. 2004 and Pazos et al. 2010).

This chapter provides the developing biotechnological aspects of soil decontamination, and also reviews various other physical, chemical, electro-remediation or electro-reclamation techniques. It also outlines the types of waste and media in which the technology could be successfully applied so that wide-scale implementation and commercialization of the technique may be recommended on a global basis.

17.2 Physical Techniques

Decontamination of soil relies on an understanding of the physical behavior of the pollutants in the site specific environment. Physical remediation methods are most effective in coarser textured soils, although fracturing of finer textured soils may extend their applicability, and for pollutants that are more soluble or volatile. This treatment can enhance the effectiveness of biological degradation of contaminates or indirectly cause their destruction (Scullion 2006).

17.2.1 Off-Site Management

The most common traditional remediation technique is off-site management. The contaminated soil is taken for burial at land fill sites. This method of remediation merely shifts the contamination problem else where. Additionally, there are hazards associated with the transport of contaminated soil and migration of contaminant from landfill into adjacent environment (Williams 1988).

17.2.2 Isolation and Containment

Contaminants can be isolated and contained, to prevent further movement, to reduce the permeability of the waste to less than 1×10^{-7} m/s (as required by the USEPA), and to increase the strength or bearing capacity of the waste (USEPA 1994). This technique consists of the use of barriers that inhibit the migration of contaminants to the neighboring uncontaminated site. Physical barriers made of steel, cement, bentonite, and grout walls can be used for capping, vertical, and horizontal containment. Capping is a site specific proven technology, which uses synthetic membranes to reduce water infiltration. Horizontal barriers restrict the downward movement of metal contaminant within the soil, whereas vertical barrier reduces the migration from one site to another. These barriers are made of slurry walls, grout, or geomembrane curtains, and sheet pile walls. It is the least expensive approach, but leaves the contaminant in place without treatment. The selection of each technology is site-specific. They are beneficial where the area of contaminant is shallow but large. More research is required to match reactive media with contaminants, model life time performance, optimize retention times, and develop methods for regeneration of reactive media.

In terms of risk management, these above mentioned approaches aimed to control the pathway linking hazard and receptor without treating the source of the hazard. Remediation practices emphasized containment rather than treatment.

17.2.3 Solidification/Stabilization

Solidification process is a nondestructive physical method to immobilize the contaminants by encapsulating them in a solid of high structural integrity, while stabilization includes chemical reactions to reduce contaminant mobility. It is also known as waste fixation through both physical and chemical means. Some variants like liquid monomers that polymerize, pozzolans, bitumen, fly-ash, asphalt, and cement are injected to encapsulate the soils. Capping or jacketing or complete coating of the contaminated sediment with sandy material, such as clean sediment, sand, or gravel, which decreases the direct contact area between the water and the contaminated sediment, is what is done under encapsulation (Peng et al. 2009). Two ways of encapsulation are (i) microencapsulation (ii) macroencapsulation. Port land cement, pozzolans, or lime/hydrated lime, and organic polymers may be used for microencapsulation, whereas concrete, organic materials (polythene, polyesters, etc.), sulfur cement, etc. can be used for macroencapsulation.

Some researchers have reported that a good cap thickness was approximately 50 cm; and through capping the sediment by sands materials, the heavy metal concentration in water could reduce to 80 %. The cost of implementing this technology is dependent on the lithology of the site and the depth of the contaminant. As the depth of contamination increases, so does the cost (Khan et al. 2004). The stabilization converts the contaminants into less soluble immobilized and less

toxic form by mixing soil or waste with chemical binders, such as cement, sulfide, and phosphate binders, polyester resins, or polysiloxane compounds to create a slurry, paste or other semi-liquid state, and is allowed time to cure into solid form (Wang et al. 2012). Additives through which solidification is achieved are either cement-based, pozzolon-based, the thermoplastic methods, the organic polymerization methods, the encapsulation method, and organophillic clay-based (Wang et al. 2012). Among these methods, cement-based solidification/stabilization is of increasing importance as option for remediating contaminated sites because of its low material and equipment cost.

This technique is suitable for contaminant in shallow depths and of large volume, and is not suitable for metals, which are not highly soluble, and do not form hydroxides, such as arsenic, chromium, and mercury. In situ solidification/stabilization techniques are preferred since labor and energy costs are lower, but site conditions, such as bedrock, large boulders, clay, and oily patches may cause mixing problems (Mulligan et al. 2001).

17.2.4 Vapor Extraction and Air Sparging

Vapor extraction and air sparging techniques are based on the manipulation of pollutant distribution between liquid and vapor phases. These treatments promote the volatilization of pollutants (e.g., benzene, toluene, ethyl benzene, xylene, and chlorobenzenes) in unsaturated and saturated zones. Extracted gases or vapors may be adsorbed onto activated carbon or treated (e.g., by oxidation). The effectiveness of vapor extraction systems may be extended to semi-volatile pollutants by injection of heated air or heating by microwave/radiowave, and rates of extraction may be improved by increasing air flow rates to a point when mass transfer limits volatilization (George et al. 1992; Park et al. 2005). Air sparging also known as in situ air stripping or in situ volatilization induces partitioning of dissolved and free-phase contaminants into the vapor phase, and increases in dissolved oxygen can stimulate aerobic degradation. Benzene removal by air sparging has been shown by Adams and Reddy (2003). Both these approaches are less well suited to fine textured soils because of restricted rates of movement in the mobile phases and increased distances over which volatile organic contaminants have to diffuse through an aqueous phase. Also treatment rates are slower in soils with higher organic contents (Gomez-Lahoz et al. 1995).

17.2.5 Vitrification

Vitrification of molten glass is another method of solidification/stabilization process requiring thermal energy. It uses heat of up to > 1,000 > 1,600 °C - > 2,000 °C to destroy organic pollutants by pyrolysis and immobilize most of the

pollutants in organic (Gavrilesseu et al. 2009). It is a two stage process in which pollutants are desorbed at lower (<600 °C) temperature and then combusted. Vitrification is mainly used to remediate soils contaminated with heavy metals mixed with radioactive elements (Wang et al. 2012). It involves insertion of electrodes into the soil, which must be able to carry a current, and then to solidify, as it cools. Full scale applications exist for arsenic, lead, and chromium contaminated soils. Vitrification is expensive and suitable for shallow contamination and also toxic gases can be produced during the process, but is applicable to mixed wastes where few technologies are available (Mulligan et al. 2001).

The in situ or ex situ (where the soil is excavated and treated) vitrification consists in the insertion of graphite electrodes into the soil creating a high electric current, such that the released heat provokes the fusion of the soil matrix (Castelo-Grande et al. 2005). This leads to the formation of vitrified end product into which the contaminants are incorporated and subsequently immobilized. During this process, the majority of contaminants initially present in the soil are volatilized reducing their concentration in the soil and the waste, while the remainder are converted into a chemically inert, stable glass, and crystalline product. The vitrification can be performed by three different processes namely electrical process, thermal process, and plasma process. Precisely, electrical process makes use of application of electrical energy through graphite electrodes inserted into ground, whereas thermal process requires an external heat source and a typical reactor (Dermatas and Meng 2003); and moreover in plasma process, electrical discharges are used to achieve temperatures up to 5,000 °C .The advantage of this method is that the volume of waste can be reduced with long-term stability but it is a costly method (Suthersan 1997).

17.2.6 Mechanical Separation

This involves the size selection process to remove larger, cleaner particles from the smaller, and more polluted ones. Characterization in terms of particle size and contaminant level in each fraction is the most important parameter in determining the suitability of this process. They include hydrocyclones, which separate the larger particles greater than 10–20 μ m by centrifugal force from the smaller particles, fluidized separation removes smaller particles at the top (Less than 50 μ m) in countercurrent overflow in a vertical column, by gravimetric settling and flotation, which is based on different surface characteristics of contaminated particles, magnetic separation, used to separate these from ferrous metals. These methods have been used in mineral ore processing.

17.2.7 Pyrometallurgical Separation

Pyrometallurgical processes use high temperature furnace to volatilize metals in contaminated soil. Temperatures of 200–700 °C are used to evaporate the contaminant. After volatilization, metals are then recovered or immobilized. These methods are most applicable to mercury since it is easily converted to its metallic form at high temperature. Other valuable metals such as gold and platinum can also be recovered from low soil concentration. This type of treatment is usually performed off-site due to lack of mobile units, and is applicable to highly contaminated soils (5–20 %) where metal recovery is profitable. Prior to pyrometal-lurgical separation the soil must be concentrated by physical or soil washing.

17.2.8 Soil Washing

Soil decontamination can also be carried out by in situ washing of soil. It is a physical separation technique, which consists of extraction of contaminants by suspending them in watery solutions, i.e., by dissolution. The main principle of soil washing is a selective classification of highly contaminated pollutants followed by the solid or liquid phase separation of the remaining suspension (Bradl and Xenidis 2005). It may consist of excavation, fragmentation, separation in different grain sizes, washing of the different fraction, and their disposal (Castelo-Grande et al. 2010). This technique is often considered as a pretreatment for the reduction of the toxic contents of the contaminated soil, and to be treated by another technology. The physical process of soil washing involves two principal steps:

Mechanical/Physical sorting Physical separation may include screening followed by density or gravity separation. Mechanical screens and hydrocyclones are often used to separate the soils into various size fractions. The bulk oversize material consists of clean or slightly contaminated cobbles and stones, and may undergo a water rinse before being returned to the site as fill. The slit and clay fraction generally contains the highest concentration of the contaminants, and is usually treated by the solidification/stabilization techniques to immobilize the contaminants prior to land filling. The remaining fine and coarse sands can be further treated using density/gravity separation process to separate high density aggregates and metal fragments. Magnetic contents are removed manually with the help of magnets.

Wash water treatment Fresh water and cleaned process water are added to the soil. From the suspension, cleaned soils fractions are separated and contaminants are further proceeded for further treatment.

For the process of soil washing the knowledge of particle size dependent pollutant distribution is of vital significance. It is one of the few permanent treatment alternatives to remove metal contaminants from the soils. This technology is particularly cost-effective and well established in mineral industry.

17.3 Chemical Techniques

A range of chemical processes have been applied to soil to destroy or convert pollutants into less toxic forms, to extract them, or to immobilize them. Wood (2001) suggested that chemical treatments can be highly specific for some pollutants, e.g., PCBs and halogenated alkanes.

17.3.1 Oxidation-Reduction Reaction

Chemical treatment by reductive as well as oxidative mechanisms may be used to detoxify or decrease the mobility of metal contaminants (Evanko and Dzombak 1997). This is commonly used for waste water treatment. Oxidation reactions detoxify, precipitate, or solubilize metals and involve addition of potassium permanganate, hydrogen peroxide, and hypochlorite, or chlorine gas. Neutralization reactions are performed to adjust the pH of acidic or basic soils (lime). Reduction reactions are induced through the addition of alkali metals, such as sodium, sulfur dioxide, sulfite salts, and ferrous sulfate. Sometimes chemical treatment is used to pretreat the soil for solidification or other treatments. These reactions are, however, not specific and, therefore, there is a risk of converting other metals into more toxic or mobile forms. Arsenic is most applicable for chemical oxidation since As (V) is less toxic than As (III). Hg, Pb, Se, and Ag are also applicable for reduction. These chemical treatments can be performed in situ by injection into ground water, but have the potential to introduce further contamination.

Over the last two decades, Fenton treatment has emerged as a viable remediation technology for PAH-contaminated soils. Several reviews on various Fentonbased treatments for contaminated soils have been published (Cravotto et al. 2005). In these works, PAHs have been grouped as hydrophobic or semi-volatile contaminants. PAHs are known to be toxic, mutagenic, carcinogenic, and teratrogenic, most commonly found at sites contaminated with coal tar and creosote, especially as the heritage from the manufactured gas plants (MGP) and wood treatment facilities of the last few centuries. Effective decontamination of the soil can be achieved by using advanced oxidation process (AOPs), which is based on Fenton's reaction (hydrogen peroxide catalysed by iron), involving solely one or a combination of physical, chemical, biological, and thermal processes .Faster and more efficient degradation of recalcitrant compounds such as PAHs can be achieved using AOPs (Cravotto et al. 2007).

17.3.2 Immobilization

Immobilization approaches are favored where pollution covers an extensive area, and where the main targets to be protected are water resources and plants. Immobilization can be achieved by complexing the contaminants or through increasing the soil pH by various amendments including addition of liming materials, phosphate compounds, and biosoilds (Alloway and Jackson 1991). Increased pH decreases the solubility of heavy metals like Cd, Cu, Ni, and Zn in soil. Although the risk of potential exposure to plants is reduced, their concentration remains unchanged. Mechanisms include increasing metal adsorption through higher surface charge, formation of insoluble metal complexes, precipitation, and redox reactions leading to immobile valency form. Basta et al. (2001) found that a range of soil amendments, especially alkaline biosolids, reduced the extractability and phytotoxicity of smelter waste polluted with Cd, Pb, or Zn.

17.3.3 Soil Washing (with Solvents)

Soil contaminated with metal pollutants can be decontaminated by two treatment methodologies:

- That leaves the metal in the soil such as solidification/stabilization and vitrification, which immobilizes the contaminants, thus limiting their movement.
- That removes the heavy metals from the soils. Technology such as soil washing, in situ soil flushing transfer the contaminants to liquid phase by desorption and solubilization.

Soil washing for metals after physical treatment processes is then washed with solvents on the basis of their ability to solubilize specific contaminants and to transform them into nonhazardous material and also on their environmental impacts (Feng et al. 2001; Chu and Chan 2003; Khan et al. 2004). Soil washing usually employs wash solutions, such as acids, bases, chelating agents, reducing agents, or other additives as the extracting agents. Thus, heavy metals can be removed from soils using various agents added to the soil. This can be done in reactors or as heap leaching. These agents are inorganic acids such as sulfuric and hydrochloric acids (pH less than 2), organic acids including acetic and citric acids (pH not less than 4), chelating agents such as ethylenediaminetetracetic acid (EDTA) and nitrilotriacetic acid (NTA), and the various combinations of the abovementioned .The cleaned soil can then be returned to the original site. Both organics and metals are removed. The effectiveness of this treatment approach can be high for hydrophilic pollutants such as aniline and phenols (Rajput et al. 1994). Metal removing efficiencies during soil washing depends on the soil particle size, metal characteristics, extractant chemistry, and processing conditions. pH plays a very important role in metal extraction from soils (Peters 1999). Limited

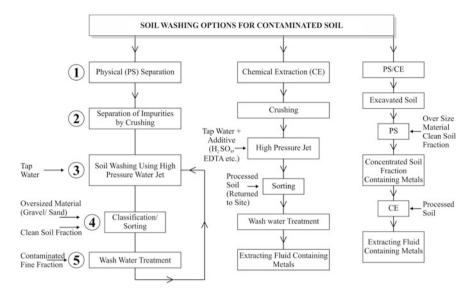


Fig. 17.1 Schematic diagram of soil washing

experience with such technique has demonstrated that they create new problems, e.g., increase in mobility (and bioavailable) of the remaining metals in the decontaminated substratum, redistribution of pollution to other remaining fractions of the process (resins, sludges, etc.) (Woelders 1998; Dermont et al. 2008) (Fig. 17.1).

17.3.4 Soil Flushing

This is in situ innovative remediation technology in which water is used with or without additives as flushing solutions to solubilize contaminants in soil to an area where they can be removed, with various methods of infiltration, such as infiltration basins, injection wells, and an infiltration trench. Various additives include organic or inorganic acids, sodium hydroxide, which can dissolve organic soil matter, water soluble solvents such as methanol, displacement of toxic cations with nontoxic cations, complexing agents such as EDTA, acids in combination with complexing agents or oxidizing/reducing agents. Once the water is pumped from soil, it must be extracted and then treated to remove the metals in wastewater treatment facilities or reused in the flushing process (Fig. 17.2). Significant removal of chromium was achieved. Levels of chromium were reduced to 18 from 2,000 mg/l (USEPA 1996). Urlings (1990) decrease the Cd content 90 % of the soil from 10 to less than 1 mg/kg with dilute hydrochloric acid (pH 3). Since

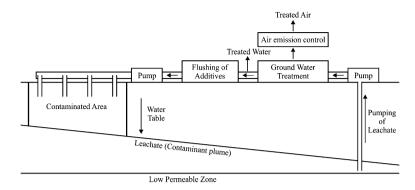


Fig. 17.2 Schematic representation of soil flushing

flushing is conducted in situ, it reduces the need for excavation, handling or transportation of hazardous substances (Khan et al. 2004). More demonstrations are needed in this methodology, in addition to developing more understanding into the mechanisms for solution, metal recovery, and use of nontoxic additives.

17.3.5 Dechlorination

Sometimes, reduction reaction can contribute to the decontamination of polluted soils. Chemical-reductive dechlorination is one such technique. Dechlorination, also known as dehalogenation, is a chemical technique based on the loss of halogen atoms (i.e., atoms of chlorine, fluorine, bromine, and iodine) from the halogenated organic molecules. Thus, converting toxic compounds into less toxic substances. These substances are frequently soluble in water, thus enhancing their separation from the soil. This technique applicable to polychlorinated biphenyls (PCBs), halogenated semivolatile volatile organic compounds, and pesticides (Castelo-Grande et al. 2010).

17.4 Biological Techniques

These techniques use living organisms in order to transform or degrade contaminants into less toxic form or remove the toxic contaminants of the soil (Kavamura and Esposito 2010). Microorganisms, soil invertebrates, and plants have all been exploited as potential agents of soil bioremediation, although most treatments have been based on microbial activity. The strategies can be applied in both in situ or ex situ ways depending on the contaminant and soil characteristics. Although biological solutions to inorganic pollution exist, the main emphasis has been on the treatment of organic compounds.

17.4.1 Biodegradation of Soil Pollutants

Degradation of soil pollutants mostly involves consortia of microorganisms, and may be achieved using in situ, on-site, or bioreactor approaches. On-site processes or bioreactors are likely to involve solid phase approaches such as landfarming (spreading of excavated contaminated soils in a thin layer usually combined with cultivation and/or nutrient inputs) or more "engineered" solutions compositing or the use of bio-piles (piles of polluted soils constructed to facilitate aeration and addition of nutrients). It can be applied to soils contaminated by crude oil, and also efficient in degradation of polycyclic hydrocarbons (PAHs). The management of the treatment environment is easiest with on site process bioreactors and difficult with in situ approaches, with bio-piles and compost techniques intermediate in this respect. Biodegradation technique is effective on a range of organic constituents. Bioaugmentation (addition of cultured microorganisms with the capacity to degrade target contaminants) or biostimulation (addition of nutrients to increase indigenous biomass or of substrates to promote cometabolism) of soil microbial populations may provide a means of accelerating pollutant degradation (Singer et al. 2005). There are many environmental factors that limit microbial biodegradation of soil pollutants including low temperature, restricted activity under anaerobic conditions, low levels of available nutrients or co-substrates, and limited bioavailability of pollutants. Although soil animals are not thought to have a significant direct role in the biological degradation of contaminants, their activities may stimulate microorganisms and improve the soil environment for microbial degradation (Haimi 2000).

Soils on many polluted sites are physically degraded and macro-fauna such as earthworms can improve these conditions (Scullion and Malik 2000). Recently, interest in the role of earthworm as "bioreactors" for degradation of contaminant such as trinitrotoluene have been reported. Biodegradation rate is controlled by microbial catabolic capacity, (Renoux et al. 2000). The presence of high concentration of heavy metals may inhibit the microbial growth and also the volatile components tend to evaporate rather than biodegrade. Thus, in most of these treatments, there will be some loss to atmosphere through volatilization and some physical or chemical stabilization of pollutants. Bioreactor treatments are increasingly favored as the microbes may also lead to formation of biofilm, which might accumulate organic pollutants for subsequent degradation of high molecular weight PAHs and heavy metals (Sheoran et al. 2010).

17.4.2 Bioleaching

It is a process of recovery of metals by some microorganisms capable of dissolving them from the environment. It is an effective alternative to chemical extraction processes. Microorganisms like *Thiobacillus ferrooxidans* and *T. thiooxidans*

bacteria under aerobic and acidic conditions (pH 4) at temperature between 15 and 55 °C, depending on strain promote the microbiological leaching of metals such as copper, silver, uranium, and zinc by the oxidation followed by electron transfer to oxygen inducing the metal solubilization (Gadd 2004; Kavamura and Esposito 2010). Leaching can be performed directly by oxidation of metal sulfides to produce sulfuric acid, which then can desorb the metals on the soil by substitution of protons. Indirect leaching involves conversion of Fe^{2+} to Fe^{3+} , which in turn oxidizes sulfur minerals to Fe^{2+} producing acidity. Several options are available for bioleaching including heap leaching, bioslurry reactors, and in situ processes. Anoxic sediments are more suitable for treatment since the bacteria can solubilize the metal compounds without substantially decreasing the pH. Copper, zinc, uranium, and gold have been removed by *Thiobacillus* species in biohydrometallurgical processes (Karavaiko et al. 1988). Another fungus Aspergillus niger, which can produce citric and gluconic acids has a potential for remediation of metal contaminated soil. They can act as acids (pH 3.5) and chelating agents for the removal of metals such as copper from oxide mining residues (Mulligan et al. 1999b). Mercury and cadmium can be oxidized, while arsenic and iron can be reduced by microorganism. Cr (VI) can be oxidized to Cr (III) that is less mobile and toxic. Bacteria such as *Bacillus subtilis* and sulfate reducing bacteria in the presence of sulfur can perform this reaction.

17.4.3 Biosorption

Biosorption is the process of binding of metals to cell surfaces (Vidali 2001). It is a biological treatment method, which involves the adsorption of metals into biomass of algal or bacterial cells that can be dead or alive. With the use of *Trichoderma reesei* adsorption and desorption of cadmium and copper were analyzed by Kim et al. (2003). Pb(II) and Cd (II) were recovered employing biomass of *Amanita rubescens* from aqueous solution using the ability of the macrofungus by Sari and Tuzen (2009). If large scale, inexpensive production techniques for the biomass are developed, this heavy metal treatment is promising (Hazardous waste consultant 1996). This method is only applicable for low concentrations of metals in water. Therefore, the cells could potentially be placed in permeable barriers for adsorption of metals in ground water.

17.4.4 Biodegradable Biosurfactants

Surfactants are a class of natural chemicals, which are amphiphilic in nature, that promote the solubilization and emulsification of various types of organic and inorganic contaminants. Bio-surfactants are produced by bacteria and yeast. Biodegradable biosurfactants (surfactin, rhamnolipid and sophorolipid) are used to remove heavy metals from an oil contaminated soil (Mulligan et al. 1999a). The first two agents are produced by bacteria, while the last is produced by yeast. Biosurfactants are also able to remove metals from the sediments. Caustic surfactant could be used for removing the organically associated metals, while foam surfactant can be employed for extracting metals bound to carbonates and oxides (Dermont et al. 2008; Wang and Catherine 2004). Since these agents are biode-gradable, they are able to enhance hydrocarbon removal, and can potentially be produced in situ, as they have a great potential for soil washing and soil flushing. This technology yet needs to be performed on commercial scale.

17.4.5 Bioventing

This technique can be applied in situ with a source of oxygen necessary to initiate the processes. It injects air into the contaminated media necessary to run the process at a rate designed to maximize in situ biodegradation. It eliminates the offgassing of volatilized contaminants to the atmosphere (Khan et al. 2004). The simulation of decontamination of several organic pollutants including variety of petroleum refinery products by bioventing was performed by Sui et al. (2007). Unlike biosparging, which involves pumping air and nutrients into the saturated zone, bioventing pumps the air only into the unsaturated or vadose zone (USEPA 1998). Bioventing also degrades less volatile organic contaminants and, because a reduced volume of air is required, it allows for the treatment of less permeable soils. During the application of bioventing in volatile solute transportation in soils, volatilization has an important role in the first day followed by biodegradation after this period (Suko et al. 2006). Baker and Moore (2000) have reported the optimized performance and effectiveness of in situ bioventing. Diele et al. (2002) have discussed numerical models and their applications in bioventing system design and operation. Any aerobic degradable substance can be treated by bioventing and ultimately leading to biodegradation. Bioventing is most successful on mid-weight petroleum products like diesel since lighter products tend to volatilize quickly, while the heavier products generally take longer time to biodegrade. If the contaminant has to be cleaned to a level lower than 0.1 ppm or if total petroleum hydrocarbon (TPH) has to be reduced to greater than 95 %, bioventing at that site may not be effective, and some other method may be needed for this saturated zone (USEPA 1998).

17.4.6 Phytoremediation

In the past few years, green plants have shown several response patterns to the presence of potentially toxic concentrations of heavy metal ions. Most are sensitive even to very low concentrations; others have developed resistance and tolerance and accumulate toxic metals within roots and above-ground tissues, such as shoot, flower, stem, and leaves, etc. (Barcelo and Poschenrieder 2003). Such extraordinary ability of plants to accumulate heavy metals is described as hyperaccumulators. Hyperaccumulators are able to accumulate Zn concentration higher than 1 % and Cu, Pb, and Ni higher than 0.1 % of the tissue weight. The current criterion used to define a hyperaccumulator is a plant that can accumulate metal to a concentration that is 100 times greater than "normal" plants growing in the same environment (Brooks 1977; Reeves and Brooks 1983). This particular capacity to accumulate and tolerate large metal concentrations has opened up the possibility to use for remediation of polluted soils and waters. The use of hyperaccumulator plants to remove, destroy, or sequester hazardous toxic heavy metals is termed phytoremediation (Schnoor 1997).

Hyperaccumulator plants such as *Thlaspi*, *Utrica*, *Chenopodium*, *Polygonum* sachalase, and *Alyssim* have the capability to accumulate cadmium, lead, nickel, and zinc (Baker et al. 1991). In addition to hyperaccumulator, plants such as trees (Poplar) and grasses (Vetiver) are now being actively evaluated though, their metal bioconcentrating capability is well below that of hyperaccumulator plants (Burken and Schnoor 1998; Sebastiani et al. 2004). Decontamination of soil from radio-active contaminants by phytoremediation has been reported (Van denhove 2013). Two novel approaches, the use of agrobacterium transformed plant roots and mycelia cultures of fungi have been reported as research tool in the study of remediation of contaminated soil by Wenzel et al. (1999).

Phytoremediation is often also referred as botanical bioremediation or green remediation (Chaney et al. 1997). It involves the repeated cropping of plants on heavy metal contaminated soils until the soils metal concentrations have reached acceptable levels. After each cropping, the plant biomass is removed from the area and may be washed to reduce its volume where upon it can be stored in an appropriate area that does not pose a risk to the environment (Raskin et al. 1994; Chaney et al. 1997). Currently phytoremediation is used for treating many classes of contaminants including elemental (heavy metals and radionuclides) as well as organic pollutants (PCBs, PAHs, nitroaromatics) (Cunningham et al. 1996; Dushenkov 2003). This is most applicable to shallow soils with low levels of contamination. The main disadvantage is that longer times are required compared to other methods. Phytoremediation can be classified according to the mechanism and nature of contaminant (Table 17.1).

17.4.6.1 Phytoextraction

It is the biological approach to remove the contamination primarily from soil and isolate it, without destroying the soil structure and fertility. It is also called phytoaccumulation. It is the process that involves the uptake and translocation of heavy metals by roots into the above ground portions of "hyperaccumulator" plants (Brown et al. 1994) (Table 17.2).

No.	Process	Mechanism	Contaminant
1.	Phytostabilisation	Complexation	Inorganics
2.	Phytoextraction	Hyperaccumulation	Inorganics
3.	Phytovolatilization	Volatilization by leaves	Organics/inorganics
4.	Phytodegradation/ phytotransformation	Degradation in plant	Organics
5.	Rhizofilteration	Rhizosphere accumulation	Organics/inorganics
6.	Rhizodegradation	Rhizosphere degradation	Organics

 Table 17.1
 Phytoremediation—on the basis of mechanism and nature of contaminant (Sheoran et al. 2012)

In phytoextraction practice, metal accumulating plants are seeded or transplanted into metal polluted soil, and are cultivated using established agricultural practices. The roots of established plants absorb metal elements from the soil and translocate them to the above-ground shoots where they accumulate. After sufficient plant growth and metal accumulation, the above-ground portions of the plant are harvested and removed, resulting in the permanent removal of metal from the site. Following harvesting of pollutant-enriched plants the weight and volume of contaminated material can be further reduced by washing or compositing. Metal enriched plants can be disposed of as hazardous material or, if economically feasible, used for metal recovery (Cunningham and Ow 1996). Phytoextraction of metals for commercial gain is called phytomining (Au, Tl, Ni) (Chaney et al. 1998). This approach is suitable to remove most metals (such as Pb, Cd, Ni, Cu, Cr) and excess nutrient from contaminated soils. Examples of plant species (Table 17.2) used are plants belonging to Brassicaceae family, such as *Thlaspi* sp., Brassica sp., (Kumar et al. 1995a, b), and Alyssum sp. (Kramer et al. 1996). Radioactive contaminants have also been decontaminated by phytoextraction (Van denhove 2013) (Fig. 17.3).

There are two basic strategies of phytoextraction namely continuous phytoextraction and chelate assisted or induced phytoextraction. Continuous phytoextraction is the removal of metals, which depends on the natural ability of the plant to extract extraordinarily metal concentration from metal contaminated soil. Natural hyperaccumulators have the ability to solubilize readily available metals from the soil matrix, efficiently absorb them into the root, and translocate them to the shoot and storage in a non-phytotoxic form in the aerial portions (Pollard et al. 2002). Some of the natural metal accumulating plants secrete metal chelating compounds such as "phytosiderophores" (mugenic acid) to the rhizosphere and some secrete organic acids (citric, malic, and oxalic acid), which act as metal chelators and decrease the rhizosphere pH, thus increase the bioavailability of metals that are tightly bound to the soil and help to carry them into plant tissues (Kinnersely 1993; Nascimento and Xing 2006). The mycorrhizal fungi associated to rhizosphere also play important role in metal (Chen et al. 2005; Khan 2005; Abou-Shanab et al. 2006).

Metal	Plant species	References	
Cadmium	Chamomilla recutita	Kral'ova and Masarovicova (2003)	
	Helianthus annus	Fenus and MacNeil (2003)	
	Arabidopsis halleri	Macnair (2002)	
	Brassica juncea	Kumar et al. (1995a), Salt et al. 1995b, Ebbs and Kochian (1998), Huang et al. (1997)	
	Thlaspi caerulescens	Escarre et al. (2000), Lombi et al. (2001), Basic et al. (2006), Keller et al. (2006)	
	Salsola kali	De la Rosa et al. (2004)	
	Hypericum perforatum	Kral'ova and Masarovicova (2003)	
	Medicago sativa	Drazic et al. (2006)	
	Zea mays		
Copper	Commelina communis	Tang et al. (1997)	
	B. juncea	Ebbs and Kochian (1998)	
	Ipomea alpina	Malaisse et al. (1979), Baker and Walker (1990)	
	Erica andevalensis	Asensi et al. (1999)	
	Elsholtzia splendens	Jiang et al. (2002)	
	Pelargonium species	Krishnaraj et al. (1999)	
	Silene vulgaris	Song et al. (2004)	
	Hirschfeldia incana		
	Haumaniastrum katangense, Crepidorhopalon perennis, Acalypha cupricola	Faucon et al. (2007)	
Chromium	Helianthus annus	Davis et al. (2001)	
	Brassica juncea	Kumar et al. (1995a), Huang et al. (1997), Han et al. (2004)	
	Convolvulus arvensis	Gardea-Torresdey et al. (2004a, b)	
	Pelaronium species	Krishnaraj et al. (1999)	
	Prosopis species	Aldrich et al. (2003)	
	Salsola kali	Gardea-Torresdey et al. (2005)	
	Sutera fodina, Dicoma niccolifera, Leptospermum scoparium, Genipa americana		
	Typha spp.	Barbosa et al. (2007)	
	Amaranthus viridis	Dong et al. (2007)	
	Miscanthus	Zou et al. (2006)	
	Oryza sativa	Arduini et al. (2006),	
	Convonvulus arvensis	Bhattacharyya et al. (2005)	
	Leucaea leucocephalla	Gardea-Torresdey et al. (2004a, b)	
	Willows (Salix sp.)	Rout et al. (1999)	
	Loilium perenne	Yu and Gu (2008)	

 Table 17.2
 Some of the plants with potential for phytoextraction of various metals (Sheoran et al. 2011, 2012)

(continued)

Metal	Plant species	References	
		Vernay et al. (2007)	
Mercury	Eichhornia crassipes	Riddle et al. (2002)	
Nickel	Psyshotria douarrei	Davis et al. (2001)	
	Brassicae juncea	Ebbs and Kochian (1998)	
	Thlaspi goesingense	Reeves and Baker (1984)	
	Streptanthus polygaloides	Reeves et al. (1981)	
	Alyssum bertoloni	Minguzzi and Vergnano (1948)	
	Berkheya codii	Robinson et al. (1997a)	
	Alyssum murale	Robinson et al. (1997b)	
		Bani et al. (2007), Chaney et al. (2008)	
	Alyssum narkgrafii	Vinterhalter and Vinterhalter (2005)	
		Perrier et al. (2004)	
	Sebertia acuminate Phyllanthus species, Euphorbia helenae, Leucocroton flavicans, L. linearifolius	Berazain et al. (2007a, b)	
Lead	Dittrichia viscose	Melendo et al. (2002)	
	B. pekinesis, B.campetris, B. carinata, B. juncea, B. napus, B. nigra, Helianthus annus, Pisum sativum		
	Thlaspi rotundifolium Zea mays	Blaylock et al. (1997), Ebbs and Kochian (1998)	
	Sesbania drummondii	Reeves and Brooks (1983)	
	Pelargonium species	Huang and Cunningham (1996)	
	Vetiveria zizaniodes	Sahi et al. (2002)	
	Pelargonium crispum	Krishnaraj et al. (1999)	
	Helianthus annus, Triticum aestivum L., Trifolium respens L.	Chen et al. (2000), Boonyapookana et al. (2005), Krishnaraj et al. (2000	
	Vicia faba	Yang et al. (1996)	
	B. juncea	Srivastava et al. (2005)	
	Hemidesmus indicus	Liu et al. (2000), Clemente et al. (2005)	
		Chandra Sekhar et al. (2005)	
Zinc	Sedum alfredii	Long et al. (2002), Yang et al. (2002	
	B. juncea, B.napus, B.rapa, Hordeum vulgare, Avena sativa,	Ebbs and Kochian (1997), (1998),	
	Arabidopsis halleri	Dahmani-Muller et al. (2000),	
	Viola calaminaria, Thlaspi calaminare	McGrath et al. (2006)	
		Baumann (1885)	
	Thlaspi careulescens	Baker and Walker (1990), Li et al.	
	Polygonium aviculare	(2006)	
		Gonzalez and Gonzalez-Chavez (2006)	
		(continued	

Table 17.2 (continued)

Metal	Plant species	References	
Selenium	B.napus, Festuca arundianacea, Hibiscus cannabinus		
	Astragalus racemose	Banuelos et al. (1997)	
	Sinapis arvensis	Rosenfeld and Beath (1964)	
	Astragallus bisulcatus	Hambuckers et al. (2008)	
	Grindelia squarosa, Stangeria pinnata	Smrkolj et al. (2007)	
	Larrea tridentate, Salvia roemeriana	Goodson et al. (2003)	
	Dryopteris fern, Pteris genera	Cruiz-Jimenez et al. (2005)	
	Typha spp.	Srivastava et al. (2005)	
		Pollard et al. (2007)	
Uranium	B. chinensis, B. juncea, B. narinosa, Amaranthus species	Huang et al. (1998)	
	Uncinia leptostachya, Coprosma arborea	Whiting et al. (2004), Chang et al. (2005)	
	Picea mariana	Babula et al. (2008)	
Thallium	Iberis intermedia, Biscutela laevigata	Anderson et al. (1999), Leblanc et al. (1999)	
	Zea mays, B. napus		
	Hirschfeldia incana, Diplotaxis catholica	Madejon et al. (2007)	
	Lolium perenne, B. napus, Phaseolus vulgaris		
	B. oleracea acephala,	Makridis et al. (1996)	
	Iberis intermedia	Al-Nazar et al. (2005)	
Cobalt	Haumaniastrum roberti	Brooks (1977)	
	Haumaniastrum katangense, Crepidorhopalon perennis, Acalypha cupricola	Faucon et al. (2007)	
	Anisopapus chinesis		
Arsenic	B. junceae	Pickering et al. (2000)	
	Pteris vittata(Brake fern)	Ma et al. (2001), Caille et al. (2004)	
	Eleocharis spp.,	Flores-Tavizon et al. (2003)	
	Pityrogramma calomelanos	Francesconi et al. (2002)	
	Pteris cretica, Pteris longifolia, Pteris umbrosa	Zhao et al. (2002)	
Gold	B. junceae,	Anderson et al. (1999)	
	B. codii, Chicory	Lamb et al. (2001), Msuya et al.	
	C. linearis	(2000)	
		Gardea-Torresdey et al. (2005)	
	1	(continued)	

Table 17.2 (continued)

(continued)

Metal	Plant species	References
Silver	B. juncea, Medicago sativa	Harris and Bali (2008)
		Borovicka et al. (2007)
	Amanita strobiliformis	
Manganese	Macadamia neurophylla	Brooks (1997)
	Phytolacca acinosa	Xue et al. (2004)
Platinum	Sinapis alba	Alt et al. (1998), (Kologziej et al. 2007), Babula et al. (2007)
	Lolium perenne	

Table 17.2 (continued)

Table 17.3 Some of the plants with potential for phytostabilization of various metals

Plant species	Metals	References
Agrostis tenuis, Festuca rubra L.	Pb, Zn, Cu	Smith and Bradshaw (1992)
Sesbania rostrata	Pb, Zn	Yang et al. (1997)
Cynodon dactylon and Festuca rubra, Typha latifolia, Phragmites australis	Pb, Zn, Cu	Wong, (1982), Ye et al. (1997, 1998)
Paspalum notatum, C. dactylcon, Imperata cyclindrica	Pb, Zn	Shu et al. (2000)
Lolium italicum, Festuca arundinaceae	Pb, Zn	Rizzi et al. (2004)
Hyparrhenia hirta, Zygophyllum fabago	Pb, Zn, Cu	Conesa et al. (2006)
Horedeum vulgare, Lupinus angustifolius, Secale cereale	As	Mains et al. (2006a, b)
B. juncea	Cd Zn, Cu, Mn, Fe, Pb, Cd	Bolan et al. (2003), Clemente et al. (2003, 2006)
Anthyllss vulneraria, Festuca arvernensis, Koeleria vallesiana, Armeria arenaria	Zn, Cd, Pb	Frerot et al. (2006)
H. hirta, Z. fabago	Pb, Zn, Cu	Conesa et al. (2006)

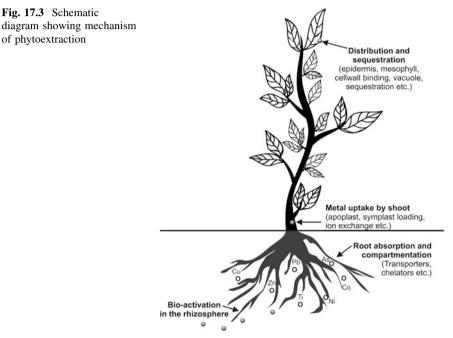
17.4.6.2 Phytostabilization

Phytostabilization, also known as phytorestoration, is a plant-based innovative remediation technique that stabilizes wastes and prevents exposure pathways via wind and water erosion; provides hydraulic control, which suppresses the vertical migration of contaminants into ground water, and physically and chemically immobilizes contaminants by root sorption and by chemical fixation with various soil amendments (Cunningham et al. 1995; Salt et al. 1995a; Berti and Cunningham 2000). It may also serve as an interim strategy to reduce risk at sites where complications delay the selection of the most appropriate technique for the site (Fig. 17.4 and Table 17.3).

of phytoextraction

Plant species	Metals	References
Arabidopsis thaliana	Hg	Rugh et al. (1996)
Liriondendron tulipifera		Rugh et al. (1998)
Nicotiana tabacum		
Astragalus racemosus	Se	Evans et al. (1968)
Brassica juncea		Pilon-Smits (2005)
Salicornia bigelowii		Pilon-Smits et al. (1999)
Typha latifolia		

Table 17.4 Some of the plants with potential for phytovolatilization of Hg and Se



For phytostabilization, the normal practice is to choose drought-resistant, fast growing crops or fodder, which can grow in metal contaminated and nutrient deficient soils. Plant should also be poor translocator of metal contaminants to above-ground plant tissues that could be consumed by humans or animals, the lack of appreciable metals in shoot tissues also eliminates the necessity of treating harvested shoot residue as hazardous waste. Metal tolerant plant species (Table 17.4) immobilize heavy metals through adsorption and accumulation by roots, absorption on to roots, or precipitation within the rhizosphere (Flathman and Lanza 1998). Phytostabilization also involves soil amendments (organic matter) to promote the formation of insoluble metal complexes that reduce biological availability and plant uptake, thus preventing metals from entering the food chain (Berti and Cunningham 2000).

Fig. 17.4 Schematic diagram showing mechanism of phytostabilization

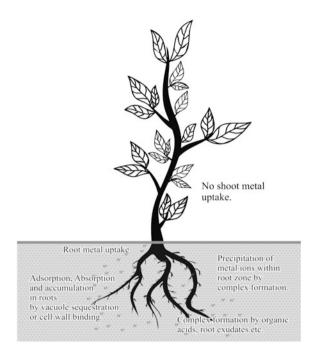


 Table 17.5
 Some of the plants with potential for rhizofilteration of various metals

Metals	Plant species	References
As, Cd, Cr, Ni, Pb, Zn	Eichnornia crassipes Populus sp., Thlaspi sp.	Zhu et al. (1999), Mangabeira et al. (2004), Salt et al. (1995a, b)
Fe, Cu, Cr	Lemna minor, Azolla pinnata	Jain et al. (1989)
Cr	Hydrocotyle umballata	Yong-pisanphop et al. (2005)
Cr	Bacopa monnieri	Mangabeira et al. (2004),
Pb, Cu, U, Sr, Cs, Co Zn	Helianthus annus	Dushenkov et al. (1995)
Cr	Spirodela polyrhiza	Appenroth et al. (2000)
Pb, Ni, Cd, Zn, Cd, Cr, Cu	Brassica juncea	Raskin et al. (1997)
Pb	Hemidesmus indicus	Sekhar et al. (2004)

17.4.6.3 Phytovolatilization

This involves use of naturally occurring or genetically modified plants that are capable of absorbing elemental forms of metal contaminants of subgroups II, V, and VI of the periodic table, such as As, Hg, and Se from the soil, and biologically converting them to gaseous species in the plant and releasing them into the

Enzymes	Plants known to produce enzymatic activity	Application
Dehalogenases	Hybrid poplar (<i>populus</i> spp.), algae (various spp.), Parrot feather (<i>Myriophyllum aquaticum</i>)	Dehalogenates chlorinated solvents
Laccase	Stonewort (<i>Nitella spp.</i>), parrot- feather (<i>Myriophyllum aquaticum</i>)	Cleaves aromatic ring after TNT is reduced to triaminotoluene
Nitrilase	Willow (Salix spp.)	Cleaves cyanide groups from aromatic rings
Nitroreductase	Hybrid poplar (<i>Populus spp.</i>), Stonewort (<i>Nitella spp.</i>), parrot feather (<i>Myriophyllum aquaticum</i>)	Reduces nitro groups on explosives and other nitroaromatic compounds, and removes nitrogen from ring structures
Peroxidase	Horseradish (<i>Armoracia rusticana</i> P.Gaertner, Meyer & Scherb)	Degradation of phenols (mainly used in wastewater treatment)
Phosphatase	Giant duckweed (Spirodela polyrhiza)	Cleaves phosphate groups from large organophosphate pesticides

 Table 17.6
 Plant enzymes that have a role in transforming organic compounds (Susarla et al.2002)

atmosphere (LeDuc et al. 2004). The well-known example of genetic manipulation is the transfer and expression of a modified *E. coli* Hg^{2+} reductase gene (*merA9pe*) in transgenic *Arabidopsis thaliana* plants (Rugh et al. 1996). Bacteria possessing *merA* are capable of converting highly toxic, Hg^{2+} to less toxic elemental Hg. Thus, expression of *merA* in transgenic plants helps the removal of elemental Hg as vapors through natural mechanisms of respiration. Rugh et al. (1998) also examined the ability of yellow poplar (L*iriondendron tulipifera*) tissue cultures and plantlets to express modified mercuric reductase (*merA*) gene constructs (Table 17.4).

Phytovolatization potentially offers a low cost alternative for Se removal from soil and water. During the process of Se volatilization, plants metabolize various inorganic species of Se [e.g., selenate, selenite, and Se-Met (Met)] into a gaseous form dimethyl selenide, the major volatile form of Se, is more than 600 times less toxic than inorganic forms (Evans et al. 1968; Berken et al. 2002; Neumann et al. 2003). Terry et al. (1992) reported that members of Brassicaceae are capable of releasing up to 40 g Se/ha/day as various gaseous compounds. Indian mustard (Brassica juncea) has a high rate of Se accumulation and volatilization, and a fast growth rate, making it a promising species for Se remediation (Pilon-Smits 2005). Some aquatic plants, such as cattail (Typha latifolia L.), have potential for Se phytoremediation (Pilon-Smits et al. 1999). Volatilization of arsenic (As) as dimethylarsenite has also been postulated as a resistance mechanism in marine algae (Salt et al. 1995a, b). Phytovolatilization has been successful in tritium (3H), a radioisotope of hydrogen; it is decayed to stable helium with a half-life of about 12 years reported by Dushenkov (2003). This remediation method has the added benefits of minimal site disturbance, less erosion, and no need to dispose of contaminated plant material (Heaton et al. 1998). However, phytovolatilization should be avoided for sites near population centers and at places with unique meteorological conditions that promote the rapid deposition of volatile compounds. Hence, the consequences of releasing the metals to the atmosphere need to be considered carefully before adopting this method as a remediation tool (Suko et al. 2006; Padmavathiamma and Loretta 2007).

17.4.6.4 Rhizofilteration/Phytofilteration

Plant assisted technique, which involves the use of both terrestrial and aquatic plants, to absorb, concentrate, and precipitate contaminants in the aqueous system has low contaminant concentration in their roots or seedlings (blastofilteration) (Dushenkov et al. 1995; Prasad and Frietas 2003). Mechanisms involved in rhizofilteration include chemisorption, complexation, ion exchange, micro precipitation, hydroxide condensation onto the biosurface, and surface adsorption (Gardea-Torresdey et al. 2004a, b). Root exudates and changes in rhizosphere pH also may cause metals to precipitate onto root surfaces. As they become saturated with metal contaminants, roots or whole plants are harvested for disposal (Flathman and Lanza 1998). Rhizofilteration can partially treat industrial discharge, agricultural runoff, or acid mine drainage. It can be used for Pb, Cr, Cd, Ca, Cu, Ni, and Zn, excess nutrients, and radionuclides (U, Cs, Sr), which are primarily retained with the roots (Ensley 2000) (Table 17.5).

17.4.6.5 Phytodegradation

It involves the breakdown of organics to simpler molecules that are incorporated into the plant tissues. Plant contains enzymes or enzyme cofactors that can breakdown and convert ammunition wastes, chlorinated solvents such as trichloroethylene and other herbicides (Newman and Reynolds 2004). Various plant species that can degrade aromatic rings in the absence of microorganisms have been described by Dec and Bollag (1994) and Singh and Jain (2003). Polychlorinated biphenyls (PCBs) have been metabolized by sterile plant tissues. Phenols have been degraded by plants, such as horseradish, potato (Solanum tuberosum), and white radish (Raphanus sativus) that contain peroxidases (Roper et al. 1996). Poplar trees (*populus* species) are capable of transforming trichloroethylene in soil and ground water (Newman et al. 1997). The enzymes are usually dehalogenases (transformation of chlorinated compounds), peroxidases (transformation of phenolic compounds), nitroreductases (transformation of explosives and other nitrated compounds), nitrilase (transformation of cyanated aromatic compounds), and phosphatases (transformation of organophosphates pesticides (Boyajian and Carriera 1997) (Table 17.6).

17.4.6.6 Rhizodegradation

Rhizodegradation is a biological treatment of a contaminant by enhanced bacterial and fungal activity in the rhizosphere of certain vascular plants. The rhizosphere is a zone of increased microbial density and activity at the root surface, and was described originally for legumes. Plants and microorganisms often have symbiotic relationships making the root zone or rhizosphere an area of very active microbial activity (bacteria and fungi) (Kirk et al. 2005). Plant litter and root exudates provide nutrients such as nitrate and phosphate that reduce or eliminate the need for costly fertilizer additives. Plant roots penetrate the soil, providing zones of aeration and stimulate aerobic biodegradation (Anderson et al. 1993).

Many plant molecules released by root dies and this exudation resembles common contaminants chemically and can be used as co-substrates. For example, phenolic substances released by plants have been found to stimulate the growth of PCB degrading bacteria (Fletcher and Hegde 1995). Recent studies have described enhanced degradation of pentachlorophenol in the rhizosphere of wheat grass (*Agropyron cristatum*), increased initial mineralization of surfactants in soil-plant cores, and enhanced degradation of TCE in soils collected from the rhizospheres (Knabel and Vestal 1992; Ferro et al. 1994).

17.4.7 Biochar

It is environmental friendly, carbon rich, fine grained, and porous substance, which is produced by thermal decomposition of several kinds of biomass under oxygenlimited conditions and at a relatively low temperatures, and have the capability of moisture and nutrients retention (Tang et al. 2013). It can also mitigate climate change by sequestrating C from atmosphere into the soil (Marris 2006) and also improve soil properties and enhance recycle of agricultural and forestry waste (Luo et al. 2011), and also microbial activity (Lehman et al. 2011). Amendment by adding biochar in the soil with poor fertility can improve the crop yield. It has been reported by various studies that biochar acts as an efficient sorbent of various organic and inorganic contaminants because of its increased surface area and special structure. Biochar can also be used for heavy metal removal from contaminated soils. Several kinds of organic waste like animal manure, woodchips, and crop waste can serve as source materials of biochar, thus showing a relationship among biochar, waste recycle, and soil decontamination.

Surface adsorbtion and partition of pollutant molecules in the micropores of biochar enhances the bioremediation process .However, from different point of view, further studies on the safety uses of biochar is needed to be carried out (Beesley et al. 2011).

17.5 Special Techniques

17.5.1 Electrokinetic Enhanced Phytoremediation

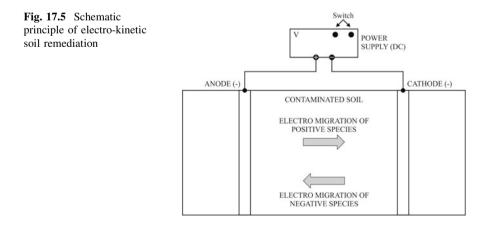
Electro-kinetic remediation is an environmental restoration technique, which involves energy application of a low DC current or a low potential gradient, in the order of mA/cm², to the two electrodes that are inserted into the sediments and encompass the contaminated zone. The application of the electric potential causes the ions to move to their respective electrodes designated as cathode and anode for subsequent removal out of the contaminated soils. It is especially designed for the in situ remediation of the contaminated soils (Cameselle et al. 2013). The electro-kinetic technology has been researched for over last two decades for the decontamination of soil. It helps in removal of heavy metals, recalcitrant, and hydrophobic organic contaminants.

Cameselle et al. (2013) reported the development of new coupled technology of electro-kinetic enhanced phytoremediation. This technology may lead to more effective and efficient remedial strategy as compared to the sequential use of these individual technologies (Fig. 17.5).

Basically, the coupled phytoremediation–electrokinetic technology consists of the application of a low intensity electric field to the contaminated soil in the vicinity of growing plants. The electric field may enhance the removal of the contaminants by increasing the bioavailability of the contaminants by desorption and transport of contaminants, even over short distances (Cameselle et al. 2013).

Variables that affect the coupled technology are: the use of AC or DC current, voltage level and mode of voltage application (continuous or periodic), soil pH evolution, and the addition of facilitating agents to enhance the mobility and bioavailability of the contaminants.

In the coupled phytoremediation-EK technology, the removal or degradation of the contaminants is performed by the plants; where as the electric field enhances the plant activity by increasing the bioavailability of the contaminants. Since the



electric field efficiency drives increased amount of soluble heavy metals toward plant roots, which results in stress conditions for the plants, hyperaccumulator plant with a rapid growth period are considered the best candidates for use in combination with EK technique .Phytoremediation can be applied after EKR to remove residual concentration of contaminants, and to achieve cleaner soil (Wan et al. 2012). The coupled EK–phytoremediation technology has showed very promising results for the restoration of heavy metal contaminated soils, and may lead to more effective and efficient remedial strategy as compared to the sequential use of these technologies (Cameselle et al. 2013).

17.6 Conclusion and Future Scope

Soil contamination is a global concern and disrupts the health of the biosphere in numerous ways; as a result reduces the capacity of the soil to meet the needs of future generation. There is an urgent need to develop an effective and affordable technological solution. There are physical, chemical, and biological methods available for such remediation, but effective remediation of polluted soils requires accurate information on the distribution and behavior of contaminants as they interact with soil and broader environment. The selection of each technique is site specific. On site with a range of organic and inorganic pollutants present, combination of different treatment approaches may offer the best prospect for effective remediation. Physical treatment process is an inexpensive comparison to chemical treatment, but most methods of physical treatments remove pollutants from the complex polluted form for further treatment or disposal. Chemical process converts the pollutant into less toxic form, or to extract them, or to immobilize them. This is highly scientific and technical process, and requires expert manpower with technological resources. Chemical additives increase the remediation cost and in situ application also increases the chances of leaching of pollutant to the other uncontaminated area and ground water. More field demonstrations are required to match reactive media with contaminants, model lifetime performance, optimize retention times, and develop methods for regeneration of reactive media. Biological methods include microorganisms (bacteria), soil invertebrates, and plant. Various field of research are needed to optimize the efficiency of biological methodology including identification of microorganisms capable of promoting their degradation, and better systems for delivering microbes, and nutrients to pollutants. Phytoremediation is environment friendly and cost-effective emerging new technology for remediation of low to moderate area of contamination as well as have important role in ecology restoration. The majority of the research for phytoremediation has been conducted in laboratories under relatively controlled conditions for short period of time.

Finally, to optimize the ecologic and economic efficiencies, it must be recognized that we need to focus more on ecological engineering approach, which is more sustainable. Assessing remediation progress and efficiency is also important, especially because pollutants are rarely completely destroyed or removed from polluted soils and evaluation of remediation has focused on the extent to which they achieve acceptable reductions in the risks posed by pollutants. Although it is clear that remediation of environment contamination is important, the need of the hour is to shift the focus from remediation to prevention for sustainable future.

References

- Abou-Shanab RAI, Angle JS, Chaney RL (2006) Bacterial inoculants affecting nickel uptake by *Alyssum murale* from low, moderate and high Ni soils. Soil Biol Biochem 38:2882–2889
- Adams JA, Reddy KR (2003) Extent of benzene biodegradation in saturated soil column during air sparging. Ground Water Moint Remediat 23:85–94
- Aldrich MV, Gardea-Torresdey JL, Peralta-Videa JR, Parsons JG (2003) Uptake and reduction of Cr(VI) to Cr(III) by mesquite (*Prosopis* spp.): Chromate-plant interaction in hydroponics and solid media studied using XAS. Environ Sci Technol 37:1859
- Alloway BJ, Jackson AP (1991) The behaviour of heavy metals in sewage-sludge amended soils. Sci Total Environ 100:151–176
- Al-Nazar H, Kaschl A, Schulz R, Romheld V (2005) Effects of thallium fractions in the soil and pollution origin in thallium uptake by hyperaccumulator plants: a key factor for assessment of phytoextraction. Int J Phytorem 7:55–67
- Alt F, Messerschmidt J, Weber G (1998) Investigation of low molecular weight platinum species in grass. Anal Chim Acta 359:65–70
- Anderson CWN, Brooks RR, Chiarucci A, LaCoste CJ, Leblanc M, Robinson BH, Simcock R, Stewart RB (1999) Phytomining for nickel, thallium and gold. J Geochem Explor 67:407–415
- Anderson TA, Guthrie EA, Walton BT (1993) Bioremediation in the rhizosphere. Environ Sci Technol 27:2630–2636
- Appenroth KJ, Bischoff M, Gabrys H, Stoeckel J, Walckzak T (2000) Kinetics of chromium (V) formation and reduction in fronds of duckweed *Spirodela polyrhiza*-a low frequency EPR study. J Inorg Biochem 78:235–242
- Arduini I, Masoni A, Ercoli L (2006) Effects of high chromium applications on *Miscanthus* during the period of maximum growth. Environ Exp Bot 58:234–243
- Asensi A, Bennet F, Brooks R, Robinson B, Stewart R (1999) Copper uptake studies on *Erica andevalensis*, a metal-tolerant plant from Southwestern Spain. Commun Soil Sci Plant Anal 30(11, 12):1615–1624
- Babula P, Supalkova V, Adam V, Havel L, Beklova M, Sladky Z, Kizek R (2007) An influence of cisplatin on the cell culture of *Nicotiana tabacum* BY-2. Plant Soil Environ 53:350–354
- Babula P, Vojtech A, Radka O, Zehnalek J, Havel L, Kizek R (2008) Uncommon heavy metals, metalloids and their plant toxicity: a review. Environ Chem Lett 6:189–213
- Baker AJM, Reeves RD, McGrath SP (1991) In situ decontamination of heavy metal polluted soils using crops of metal-accumulating plants—a feasibility study. In: Hinchee RE, Olfenbuttel RF (eds) In situ bioreclamation. Butterworth-Heinemann Publishers, Stoneham, pp 539–544
- Baker AJM, Walker PL (1990) Ecophysiology of metal uptake by tolerant plants. In: Shaw AJ (ed) Heavy metal tolerance in plants: evolutionary aspects. CRC Press, Boca Raton, pp 155–177
- Baker RS, Moore AT (2000) Optimizing the effectiveness of in situ bioventing. Pollut Eng 32(7):44-47
- Bani A, Echevarria G, Sulce S, Morel JL, Mullai A (2007) In situ phytoextraction of Ni by a native population of *Alyssum murale* on an ultramafic site (Albania). Plant Soil 293:79–89

- Banuelos GS, Airua HA, WU L, Guo X, Akohouy S, Zambrowski S (1997) Selenium induced growth reduction in *Brassicae landraces* considered for phytoremediation. Ecotoxicol Environ Saf 36:282–287
- Barbosa RMT, deAlmeida A-AF, Mielke MS, Longuercio LL, mangabeira PAO, Gomes FP (2007) A physiological analysis of *Genipa americana* L.: a potential phytoremediator tree for chromium polluted watersheds. Environ Exp Bot 61:264–271
- Barcelo J, Poschenrieder C (2003) Phytoremediation: principles and perspectives. Contrib Sci 2(3):333–344
- Basic N, Salamin C, Keller N, Galland N, Besnard G (2006) Cadmium hyperaccumulation and genetic differentiation of *Thlaspi caerulescens* populations. Biochem Syst Ecol 34:667–677
- Basta NT, Gradwohl R, Sneyhen KL, Schroder JL (2001) Chemical iimmobilization of lead, zinc, and cadmium in smelter-contaminated soils using biosolids and rock phosphate. J Environ Qual 30:1222–1230
- Baumann A (1885) Das verhalten von zinksalzen gegen pflanzen und im boden. Landwirtscha Verss 3:1–53
- Beesley L, Moreno-Jimenez E, Gomez-Eyles JL, Harris E, Robinson B, Sizmur T (2011) A review of biochars' potential role in the remediation, revegetation and restoration of contaminated soils. Environ Pollut 159:3269–3282
- Berazain R, de la Fuente V, Rufo L, Rodriguez Nuria, Amils R, Diez-Garretas B, Sanchez-Mata D, Asensi A (2007a) Nickel localization in tissues of different hyperaccumulator species of euphorbiaceae from ultramafic areas of Cuba. Plant Soil 293:99–106
- Berazain R, de la Fuente V, Sanchez-Mata D, Rufo L, Rodriguez N, Amils R (2007b) Nickel localization on tissues of hyperaccumulator species of *Phyllanthus* L. (Euphorbiaceae) from ultramafic areas of Cuba. Biol Trace Elem Res 115:67–86
- Berken A, Mulholland MM, LeDuc DL, Terry N (2002) Genetic engineering of plants to enhance selenium phytoremediation. Crit Rev Plant Sci 21:567–582
- Berti WR, Cunningham SD (2000) Phytostabilization of metals. In: Raskin I, Ensley BD (eds) Phytoremediation of toxic metals—using plants to clean up the environment. Wiley, New York, pp 71–88
- Bhattacharyya P, Chakraborty A, Chakrabarti K, Tripathy S, Powell MA (2005) Chromium uptake by rice and accumulation in soil amended with municipal solid waste compost. Chemosphere 60:1481–1486
- Bilek F (2004) Prediction of ground water quality affected by acid mine drainage to accompany in situ remediation. Appl Earth Sci 113:B31–B42
- Blaylock MJ, Salt DE, Dushenkhov S, Zakharova O, Gussman C, Kapulnik Y, Ensley BD, Raskin I (1997) Enhanced accumulation of Pb in Indian mustard by soil-applied chelating agents. Environ Sci Technol 31:860–865
- Bolan NS, Adriano DC, Naidu R (2003) Role of phosphorus in immobilization and bioavailability of heavy metals in the soil-plant system. Rev Environ Contam Toxicol 177:1–44
- Boonyapookana B, Parkplan P, Techapinyawat S, DeLaune RD, Jugsujinda A (2005) Phytoaccumulation of lead by sunflower (*Helianthus annus*), tobacco (*Nicotiana tabaccum*), and (*Vetiveria zizanioides*). J Environ Sci Heal A 40:117–137
- Borovicka J, Randa Z, Jelinek E, Kotrba P, Dunn CE (2007) Hyperaccumulation of silver by *Amanita strobiliformis* and related species of the section lepidella. Mycol Res 111(11):1339–1344
- Boyajian G, Carriera LH (1997) Phytoremediation : a clean transition from laboratory to marketplace. Natur Biotechnol 15:127–128
- Bradl H, Xenidis A (2005) Remediation techniques. In: Bradl HB (ed) Heavy metals in environment, pp 165–261. Elsevier Ltd, London
- Brooks RR (1977) Copper and cobalt uptake by Haumanniastrum species. Plant Soil 48:541-544
- Brooks RR (1997) Plants that hyperaccumulate heavy metals. C.A.B. International, Wallingford, pp 88–105

- Brown SL, Chaney RL, Angle JS, Baker AJM (1994) Phytoremediation potential of *Thlaspi* caerulescens and *Bladder campion* for zinc and cadmium-contaminated soil. J Environ Qual 23:1151–1157
- Burken JG, Schnoor JL (1998) Predictive relationship for uptake of organic contaminants by hybrid poplar trees. Environ Sci Technol 32:3379–3385
- Caille N, Swanwick S, Zhao FJ, McGrath SP (2004) Arsenic hyperaccumulation by *Pteris vittata* from arsenic contaminated soils and the effect of liming and phosphate fertilization. Environ Pollut 132:113–120
- Cameselle C, Chirakkara RA, Reddy KR (2013) Electrokinetic-enhanced photoremediation of soils: status and opportunities. Chemosphere 93:626–636
- Castelo-Grande T, Augusto PA, Monteiro P, Estevej AM, Barbosa D (2010) Remediation of soil contaminated by pesticides: a review. Int J Environ Anal Chem 90(3):438–467
- Castelo-Grande T, Augusto PA, Barbosa D (2005) Removal of pesticides from soil by superficial extraction- a preliminary study. Chem Eng J 111:167–171
- Chandra Sekhar K, Kamala CT, Chary NS, Balaram V, Garcia G (2005) Potential of *Hemidesmus indicus* for phytoextraction of lead from industrially contaminated soils. Chemosphere 58:507–514
- Chaney RL, Angle JS, Baker AJM, Li JM (1998) Method for phytomining of nickel, cobalt, and other metal from soil. U.S. Patent # 5, 711, 784
- Chaney RL, Malik M, Li YM, Brown SL, Brewer EP, Angle JS, Baker AJM (1997) Phytoremediation of soil metal. Curr Opin Biotechnol 8:279–284
- Chaney RL, Chen KY, Li Y-M, Angle JS, Baker AJM (2008) Effects of calcium on nickel tolerance and accumulation in *Alyssum* species and cabbage grown in nutrient solution. Plant Soil. doi:10.1007/s11104-008-9664-7
- Chang P, Kim K-W, Yoshida S, Kim S-Y (2005) Uranium accumulation of crop plants enhanced by citric acid. Environ Geochem Health 27:529–538
- Chen BD, Jakobsen I, Roos P, Borggaard OK, Zhu YG (2005) Mycorrhiza and root hairs enhance acquisition of Phosphorus and uranium from phosphate rock but mycorrhiza decreases root to shoot uranium transfer. New Phytol 165:591–598
- Chen HM, Zheng CR, Tu C, Shen ZG (2000) Chemical methods and phytoremediation of soil contaminated with heavy metals. Chemosphere 41:229–234
- Chu W, Chan KH (2003) The mechanism of the surfactant-aided soil washing system for hydrophobic and partial hydrophobic organics. Sci Total Envir 307(1-3):83–92
- Clemente R, Almela C, Bernal PM (2006) A remediation strategy based on active phytoremediation followed by natural attenuation in a soil contaminated by pyrite waste. Environ Pollut 143(3):397–406
- Clemente R, Walker DJ, Bernal MP (2005) Uptake of heavy metals and as by *B. juncea* grown in contaminated soil in Aznalcollar (Spain): the effect of soil amendments. Environ Pollut 138:46–58
- Clemente R, Walker JD, Roig A, Bernal PM (2003) Heavy metal bioavailability in a soil affected by mineral sulphides contamination following the mine spillage at Aznalcollar (Spain). Biodegradation 14:199–205
- Conesa MH, Faz A, Arnaldos R (2006) Initial Studies for the phytostabilization of a mine tailing from the Cartagena–La Union Mining District (SE Spain). Chemosphere 66(1):38–44
- Cravotto G, Carlo, S, Tumiatti V, Roggero C, Bremner HD (2005) Degradation of persistent organic pollutants by Fenton's reagent facilitated by microwave or high intensity ultrasound. Environ Technol 26:721–724
- Cravotto G, Carlo, S, Tumiatti V, Roggero CM (2007) Decontamination of soil containing POPs by the combined action of solid Fenton-like reagents and microwaves. Chemosphere 69:1326–1329
- Cruiz- Jimenez G, Peralta-Videa JR, de la Rosa G, Meitzner G, Parsons JG, Gardea-Torresdey JL (2005) Effect of sulfate on selenium uptake and chemical speciation in *Convolvulus arvensis* L. Environ Chem 2:100–107

- Cunningham SD, Anderson TA, Schwab P, Hsu FC (1996) Phytoremediation of soils contaminated with organic pollutants. Adv Agron 56:55-114
- Cunningham SD, Berti WR, Huang JW (1995) Phytoremediation of contaminated soils. Trends Biotechnol 13:393–397
- Cunningham SD, Ow DW (1996) Promises and prospects of phytoremediation. Plant Physiol 110(3):715-719
- Dahmani-Muller H, van Oort F, Gélie B, Balabane M (2000) Strategies of heavy metal uptake by three plant species growing near a metal smelter. Environ Pollut 109:231–238
- Davies FT Jr, Puryear JD, Newton RJ, Egilla JN, Saraivag JA (2001) Mycorrhizal fungi enhance accumulation and tolerance of chromium in sunflower (Helianthus annuus). Plant Physiol 158:777–786
- Davis MA, Pritchard SG, Boyd RS, Prior SA (2001) Developmental and induced responses of nickel- based and organic defenses of the nickel-hyperaccumulating shrub *Psichotria douarrei*. New Phytol 150:49–58
- De la Rosa G, Peralta-Videa JR, Montes M, Parsons JG, Cano-Aguilera I, Gardea-Torresdey JL (2004) Cadmium uptake and translocation in tumbleweed (*Salsola kali*), a potential Cd-hyperaccumulator desert plant species: ICP/OES and XAS studies. Chemosphere 55:1159–1168
- Dec J, Bollag JM (1994) Use of plant material for the decontamination of water polluted with phenols. Biotech Bioeng 44:1132–1139
- Dermatas D, Meng X (2003) Utilisation of fly ash for stabilisation/solidification of heavy metal contaminated soils. Eng Geo 70(3–4):377–394
- Dermont G, Bergeron M, Mercier G, Richer-Lafleche M (2008) Soil washing for metal removal: a review of physical/chemical technologies and field application. J Hazard Mater 152:1–31
- Diele F, Notarnicola F, Sgura I (2002) Uniform air velocity field for a bioventing system design: some numerical results. Int J Eng Sci 40(11):1199–1210
- Do Nascimento CWA, Xing B (2006) Phytoextraction a review on enhanced metal availability and plant accumulation. Scienta Agricola (Piracicaba, Brazil) 3(3):299–311
- Dong J, Wu FB, Huang RG, Zang GP (2007) A chromium-tolerant plant growing in crcontaminated land. Int J Phytoremediation 9:167–179
- Drazic G, Mihalovic N, Lolic M (2006) Cadmium concentration in *Medicago sativa* seedlings treated with salicylic acid. Biol Plant 50:239–244
- Dushenkov V, Kumar PBAN, Motto H, Raskin I (1995) Rhizofiltration: the use of plants to remove heavy metals from aqueous streams. Environ Sci Technol 29:1239–1245
- Dushenkov D (2003) Trends in phytoremediation of radionuclides. Plant Soil 249:167-175
- Ebbs SD, Kochian LV (1997) Toxicity of zinc and copper to *Brassica* species: implications for phytoremediation. J Environ Qual 26:776–781
- Ebbs SD, Kochian LV (1998) Phytoextraction of zinc by oat (Avena sativa), barley (Hordeum vulgare), and Indian mustard (Brassica juncea). Environ Sci Technol 32:802–806
- Ensley BD (2000) Rational for use of phytoremediation. In: Raskin I, Ensley BD (eds) Phytoremediation of toxic metals: using plants to clean- up the environment. Wiley, New York, pp 3–12
- Escarre J, Lefebre C, Gruber W, Leblanc M, Lipart J, Riviere Y, Delay B (2000) Zinc and cadmium hyperaccumulation by *Thlaspi caerulescens* from metalliferous and non metalliferous sites in the miditerranean area: implications for phytoremediation. New Phytol 145:429–437
- Evanko CR, Dzombak DA (1997) Remediation of metals-contaminated soils and ground water, technology evaluation report, TE-97-01. Ground Water Remediation Technologies Analysis Center, Pittsburgh, P.A
- Evans CS, Asher C, Johnson CM (1968) Isolation of dimethyl diselenide and other volatile selenium compounds from *Astragalus racemosus* (Pursh.). Aust J Biol Sci 21:13–20
- Faucon M-P, Shutcha N, Meerts P (2007) Revisiting copper and cobalt concentrations in supposed hyperaccumulators from SC Africa: influence of washing and metal concentrations in soil. Plant Soil 301:29–36

- Feng D, Lorenzen L, Aldrich C, Mare PW (2001) Ex- situ diesel contaminated soil washing with mechanical methods. Miner Eng 14(9):1093–1100
- Fenus TJ, MacNeil JH (2003) Hyperaccumulation of cadmium by *Helianthus annuus*. In: Abstract of the proceedings of the 225th ACS national meeting, New Orleans, LA, pp 23–27
- Ferro AM, Sims RC, Bugbee B (1994) Hycrest crested wheatgrass accelerates the degradation of pentachlorophenol in soil. J Environ Qual 23:272–279
- Flathman PE, Lanza GR (1998) Phytoremediation: current views on an emerging green technology. J Soil Contam 7(4):415–432
- Fletcher JS, Hegde RS (1995) Release of phenols by perennial plant roots and their potential importance in bioremediation. Chemosphere 31:3009–3016
- Flores-Tavizon E, Alarcon-Herrera MT, Gonzalez Elizondo S, Olguin EJ (2003) Arsenic tolerating plants from mine sites and hot springs in the semi arid region of Chihuahua Mexico. Acta Biotechnol 23:113–119
- Francesconi K, Visoottiviseth P, Sridokchan W, Goessler W (2002) Arsenic species in an arsenic hyperaccumulating fern, *Pityrogramma calomelanos*: a potential phytoremediater of arseniccontaminated soils. Sci Total Environ 284:27–35
- Frerot H, Lefèbvre C, Gruber W, Collin C, Dos Santos A, Escarre J (2006) Specific interactions between local metallicolous plants improve the phytostabilization of mine soils. Plant Soil 282:53–65
- Friberg L, Nordberg GF, Vouk VB (1986) Handbook on the toxicology of metals, 2nd edn. Elsevier, Amsterdam
- Gadd GM (2004) Microbial influence on metal mobility and application for bioremediation. Geoderma 122:109–119
- Gardea-Torresdey JL, de la Rosa G, Peralta-Videa JR (2004a) Use of phytofiltration technologies in the removal of heavy metals: a review. Pure Appl Chem 76(4):801–813
- Gardea-Torresdey JL, Peralta-Videa JR, de La Rosa G, Parsons JG (2005) Phytoremediation of heavy metals and study of the metal coordination by x-ray absorption spectroscopy. Coord Chem Rev 249:1797–1810
- Gardea-Torresdey JL, Peralta-Videa JR, Montes M, deLa Rosa G, Corral-Diaz B (2004b) Bioaccumulation of cadmium, chromium and copper by *Convolvulus arvensis* L.: impact on plant growth and uptake of nutritional elements. Bioresour Technol 92(3):229–235
- Gavrilescu M, Pavel LV, Cretescu I (2009) Characterization and remediation of soils contaminated with uranium. J Hazard Mater 163:475–510
- George CE, Lightsey GR, Jun I, Fan JY (1992) Soil decontamination via microwave and radiofrequency covolatilisation. Environ Prog 11:216–219
- Ghosh M, Singh SP (2005) A review on phytoremediation of heavy metals and utilization of it's by product. Appl Ecol Environ Res 3(1):1–18
- Gomez-Lahoz C, Rodriguez-Maroto JM, Wilson DJ (1995) Soil clean up by insitu aeration XXII. Impact of natural soil organic matter on clean up rates. Sep Sci Technol 30:659–682
- Gonzalez RC, Gonzalez-Chavez MCA (2006) Metal accumulation in wild plants surrounding mining wastes. Environ Pollut 144(1):84–92
- Goodson CC, Parker DR, Amrhein C, Zhang Y (2003) Soil selenium uptake and root system development in plant taxa differing in Se- accumulating capability. New Phytol 159:391–401
- Haimi J (2000) Decomposer animals and bioremediation of soils. Environ Pollut 107:233-238
- Hambuckers A, Dotreppe O, Hornick JL, Istasse L, Dufrasne I (2008) Soil applied selenium effects on tissue selenium concentrations in cultivated and adventitious grassland and pasture plant species. Soil Sci Plant Anal 39:800–811
- Han FXX, Sridhar BBM, Monts DL, Su Y (2004) Phytoavailability and toxicity of trivalent and hexavalent chromium to *B. juncea*. New Phytol 162:489–499
- Harris AT, Bali R (2008) On the formation and extent of uptake of silver nanoparticles by live plants. J Nanopart Res 10:691–695
- Hazardous Waste Consultant (1996) Remediating Soil and sediment contaminated with heavy metals, Nov/Dec. Elsevier science, Netherlands

- Heaton ACP, Rugh CL, Wang N, Meagher RB (1998) Phytoremediation of mercury and methyl mercury polluted soils using genetically engineered plants. J Soil Contam 74:497–510
- Hoffmann G (1983) Relationships between critical levels of pollutants in soils, fodder, and crops, (In German.). Landwirtsch Forsch Sonderh 39:130–152
- Huang JW, Blaylock MJ, Kapulnik Y, Ensley BD (1998) Phytoremediation of uranium contaminated soils: role of organic acids in triggering uranium hyperaccumulation in plants. Environ Sci Technol 32(13):2004–2008
- Huang JW, Chen J, Berti WB, Cunningham SD (1997) Phytoremediation of lead-contaminated soils: role of synthetic chelates in lead phytoextraction. Environ Sci Technol 31:800–805
- Huang JW, Cunningham SD (1996) Lead phytoextraction: species variation in lead uptake and ttranslocation. New Phytol 134:75–84
- Jain SK, Vasudevan P, Jha NK (1989) Removal of some heavy metals from polluted water by aquatic plants: studies on duckweed and water velvet. Biol Wastes 28(2):115
- Jiang LY, Shi WY, Yang XE, Fu CX, Chen WG (2002) Hyperaccumulators in mining area. Chinese J Appl Ecol 13(7):906–908
- Karavaiko GI, Rossi G, Agates AD, Groudev SN, Avakyan ZA (1988) Biogeotechnology of metals: manual. Center for International Projects GKNT, Moscow
- Kavamura VN, Esposito E (2010) Biotechnological strategies applied to the decontamination of soils polluted with heavy metals. Biotech Adv 28:61–69
- Keller C, Diallo C, Cosio N, Basic N, Galland N (2006) Cadmium tolerance and hyperaccumulation by *Thlaspi caerulescens* populations grown in hydroponics are related to plant uptake characteristics in the field function. Plant Biol 33:673–684
- Khan FI, Husain T, Hejazi R (2004) An overview and analysis of site remediation technologies. J Environ Mgmt 71:95–255
- Khan AG (2005) Role of soil microbes in the rhizospheres of plants growing on trace metal contaminated soils in phytoremediation. J Trace Elem Med Biol 18:355–364
- Kim SK, Park CB, Koo YM, Yun HS (2003) Biosorption of cadmium and copper ions by *Trichoderma reesei* RUT C30. J Ind Eng Chem 9:403–406
- Kinnersely AM (1993) The role of phytochelates in plant growth and productivity. Plant Growth Regul 12:207–217
- Kirk I, Klironomos I, Lee H, Trevors JT (2005) The effects of perennial ryegrass and alfalfa on microbial abundance and diversity in petroleum contaminated soil. Environ Pollut 133:455–465
- Knabel DB, Vestal JR (1992) Effects of intact rhizosphere microbial communities on the mineralization of surfactants in surface soils. Can J Microbiol 38:643–653
- Knasmuller S, Gottmann E, Steinkellner H, Fomin A, Pickl C, Paschke A, God R, Kundi M (1998) Detection of genotoxic effects of heavy metal contaminated soils with plant aioassay. Mutat Res 420:37–48
- Kologziej M, Baranowska I, Matyja A (2007) Determination of platinum in plant samples by voltammetric analysis. Electro-analysis 19:1585–1589
- Kral'ova K, Masarovicova E (2003) *Hypericum perforatum* L. and *Chamomilla recutita* (L.) rausch.—accumulators of some toxic metals. Pharmazie 58(5):359–359
- Kramer U, Cotter-Howells JD, Charnock JM, Baker AJM, Smith JAC (1996) Free histidine as a metal chelator in plants that accumulate nickl. Nature 379:635–638
- Krishnaraj S, Dan TV, Saxena PK (2000) A fragment solution to soil remediation. Int J Phytorem 2:117–132
- Krishnaraj S, Saxena PK, Perras MR, Michel R (1999) Method of using *Pelargonium* species as hyperaccumulators for remediating contaminated soil. PCT/CA9801027 Int Appl 1–20
- Kumar PBAN, Motto H, Raskin I (1995a) Rhizofiltration: the use of plants to remove heavy metals from aqueous streams. Environ Sci Technol 29(5):1239–1245
- Kumar PBAN, Dushenkov V, Motto H, Raskin I (1995b) Phytoextraction –the use of plants to remove heavy metals from soils. Environ Sci Technol 29:1232–1238
- Lamb AE, Anderson CWN, Haverkamp RG (2001) The induced accumulation of gold in the plants *Brassica juncea*, *Berkheya codii* and *Chicory*. Chem New Zealand 65(2):34–36

- Leblanc M, Robinson BH, Petit D, Deram A, Brooks RR (1999) The phytomining and environmental significance of hyperaccumulation of thallium by *Iberis intermedia* from southern France. Econ Geol 94:109–114
- LeDuc DL, Tarun AS, Montes-Bayon M, Meija J, Malit MF, Wu CP, Abdel Samie M, Chiang CY, Tagmount A, DeSouza M, Neuhierl B, Bock A, Caruso J, Terry N (2004) Overexpression of selenocysteine methyltransferase in *Arabidopsis* and Indian mustard increases selenium tolerance and accumulation. Plant Physiol 135:377–383
- Lehmann J, Rillig MC, Thies J, Masiello CA, Hockaday WC, Crowley D (2011) Biochar effects on soil biota—a review. Soil Biol Biochem 43:1812–1836
- Li Y-M, Chaney RL, Reeves RD, Angle JS, Baker AJM (2006) *Thlaspi caerulescens* sub species for Cd and Zn recovery. US Patent No.7049, 492. Date issued-23 May
- Liu D, Jiang W, Liu C, Xin C, How W (2000) Uptake and accumulation of lead by roots, hypocotyls and shoots of Indian mustard (*Brassica juncea* L.). Bioresour Technol 71:273–277
- Lombi E, Zhao FJ, Dunham SJ, McGrath SP (2001) Phytoremediation of heavy metalcontaminated soils: natural hyperaccumulation versus chemically enhanced phytoextraction. J Environ Qual 30:1919–1926
- Long XX, Yang XE, Ye ZQ, Ni WZ, Shi WY (2002) Differences of uptake of and accumulation of zinc in four species of *Sedum*. Acta Botanica Sinica 44:152–157
- Luo L, Lou LP, Cui XY, Wu BB, Hou J, Xun B, Xu XH, Chen YX (2011) Sorption and desorption of pentachlorophenol to black carbon of three different origins. J Hazard Mater 185:639–646
- Ma JF, Ryan PR, Delhaize E (2001) Aluminium tolerance in plants and the complexing role of organic acids. Trends Plant Sci 6:273–278
- Macnair MR (2002) Within and between population genetic variations for zinc accumulation in *Arabidopsis halleri*. New Phytol 155(1):9–66
- Madejon P, Murillo JM, Maranon T, Lepp NW (2007) Factors affecting accumulation of thallium and other trace elements in two wild Brassicaceae spontaneously growing on soils contaminated by tailings dam waste. Chemosphere 67:20–28
- Mains D, Craw D, Rufaut CG, Smith CMS (2006a) Phytostabilization of gold mine tailings, New Zealand. Part 1: plant establishment in alkaline saline substrate. Int J Phytorem 8(2):131–147
- Mains D, Craw D, Rufaut CG, Smith CMS (2006b) Phytostabilization of gold mine tailings from New Zea-land. Part 2: experimental evaluation of arsenic mobiliza- tion during revegetation. Int J Phytorem 8(2):163–183
- Makridis C, Pateras D, Amberger A (1996) Thallium pollution risk to food chain from cement plant. Fresenius Environ Bull 5:643–648
- Malaisse F, Gregoire J, Morrison RS, Reeves RD (1979) Copper and cobalt in vegetation of Fungurume, Shaba Province, Zaire. Oikos 33:472–478
- Mangabeira PAO, Labejof L, Lamperti A, deAlmeida AAF, Oliveira AH, Escaig F, Severo MIG (2004) Accumulation of chromium in roots tissues of *Eichhornia crassipes* (Mart.) Solms. In Cachoeira river-Brazil. Appl Surf Sci 231(232):497–501
- Marris E (2006) Putting the carbon back: black is the New Green. Nature 442:624-626
- McGrath SP, Lombi E, Gray CW, Caille N, Dunham SJ, Zhao FJ (2006) Field evaluation of Cd and Zn phytoextraction potential by the hyperaccumulators *Thlaspi caerulescens* and *Arabidopsis halleri*. Environ Pollut 141:115–125
- Melendo M, Benítez E, Nogales R (2002) Assessment of the feasibility of endogeneous Mediterranean species for phytoremediation of Pb-contaminated areas. Fresenius Environ Bull 11:1105–1109
- Minguzzi C, Vergnano O (1948) II cotenuto di nichel nelle ceneri di *Alyssum bertolonii*. Atti Soc Tosc Sci Nat 55:49–74
- Msuya FA, Brooks RR, Anderson CWN (2000) Chemically-induced uptake of gold by root crops: its significance for phytomining. Gold Bull 33(4):134–137
- Mulligan CN, Yong RN, Gibbs BF (1999a) On the use of biosurfactants for the removal of heavy metals from oil-contaminated soil. Environ Prog 18(1):50–54

- Mulligan CN, Yong RN, Gibbs BF (2001) Remediation technologies for metal-contaminated soils and groundwater: an evaluation. Eng Geol 60:193–207
- Mulligan CN, Galvez-Cloutier R, Renaud N (1999b) Biological leaching of copper mine residues by Aspergillus niger. Presented at AMERICANA 1999, Pan-American Environment Trade Show and Conference, Montreal, Canada, pp 24–26
- Nathanail CP, Earl N (2001) Human health risk assessment: guidelines values and magic numbers. In: Hester RE, Harrison RM (eds) Assessment and reclamation of contaminated land. Royal Society of Chemistry, Cambridge, pp 85–102
- Neumann PM, DeSouza MP, Pickering IJ, Terry N (2003) Rapid microalgal metabolism of selenate to volatile dimethylselenide. Plant Cell Environ 26:897–905
- Newman LA, Reynolds CM (2004) Phytodegradation of organic compounds. Curr Opin Biotechnol 15:225–230
- Newman LA, Strand SE, Choe N, Duffy J, Ekuan G (1997) Uptake and biotransformation of trichloroethylene by hybrid poplars. Environ Sci Technol 31:1062–1067
- Padmavathiamma PK, Loretta YM (2007) Phytoremediation technology: hyperaccumulation metals in plants. Water Air Soil Pollut 184:105–126
- Park G, Shin HS, Ko SO (2005) A laboratory and pilot study of thermally enhanced soil vapor extraction method for the removal of semi-volatile organic contaminants. J Environ Sci Health Part Am 40:881–897
- Pazos M, Rosales E, Alcantara T, Gomez J, Sanaroman MA (2010) decontamination of soils containing PAHs by electroremediation, a review. J Hazard Mater 177:1–11
- Peng JF, Song YH, Yuan P, Cui XY, Qui GL (2009) The remediation of heavy metals contaminated sediments. J Hazard Mater 161:633–640
- Perrier N, Colin F, Jaffre T, Ambrosi JP, Rose J, Bottero JY (2004) Nickel speciation in Sebertia acuminate, a plant growing on a lateritic soil of New Caledonia. CR Geosci 336:567–577
- Peters RW (1999) Chelant extraction of heavy metals from contaminated Soils. J Hazard Mater 66:151–210
- Pickering IJ, Prince RC, George MJ, Smith RD, George GN, Salt DE (2000) Reduction and coordination of arsenic in Indian mustard. Plant Physiol 122:1171–1177
- Pilon-Smits E (2005) Phytoremediation. Annu Rev Plant Biol 56:15-39
- Pilon-Smits EAH, Hwang S, Lytle CM, Zhu Y, Tai JC, Bravo RC, Chen Y, Leustek T, Terry N (1999) Overexpression of ATP sulfurylase in Indian mustard leads to increased selenate uptake, reduction, and tolerance. Plant Physiol 119:123–132
- Pollard AJ, Powell KD, Harper FA, Smith JAC (2002) The genetic basis of metal hyperaccumulation in plants. Crit Rev Plant Sci 21(6):539–566
- Pollard J, Cizdziel J, Stave K, Reid M (2007) Selenium concentrations in water and plant tissues of a newly formed arid wetland in Las Vegas. Nevada Env Monit Assess 135:447–457
- Prasad VMN, Frietas HMO (2003) Metal hyperaccumulation in plants—biodiversity prospecting for phytoremediation technology. Electron J Biotechnol 6(3):285–321
- Rajput VS, Higgins AJ, Singley ME (1994) Cleaning of excavated soil contaminated with hazardous organic compounds by washing. Water Environ Res 66:819–827
- Raskin I, Kumar PBAN, Dushenkov S, Salt DE (1994) Bioconcentration of heavy metals by plants. Curr Opin Biotechnol 5:285–290
- Raskin I, Smith RD, Salt DE (1997) Phytoremediation of metals: using plants to remove pollutants from environment. Curr Opin Biotechnol 8:221–226
- Reed DT, Tasker IR, Cunnane JC, Vandegrift GF (1992) In: Vandgrift GF, Reed DT, Tasker IR (eds) Environmental remediation removing organic and metal ion pollutants. Am Chem Soc, Washington DC, pp 1–9
- Reeves RD, Brooks R (1983) Hyperaccumulation of lead and zinc by two metallophytes from a mining area in Central Europe. Environ Pollut 31:277–287
- Reeves RD, Baker AJM (1984) Studies on metal uptake by plants from serpentine and non serpentine populations of *Thlaspi goesingense* Halacsy (Crucifera). New Phytol 98:191–204

- Reeves RD, Brooks RR, Macfarlane RM (1981) Nickel uptake by Californian *Streptanthus* and *Caulanthus* with particular reference to the hyperaccumulator *S. polygaloides* Gray (Brassicaceae). Am J Bot 68:708–712
- Renoux AY, Sarrazin M, Hawari J, Sunahara GI (2000) Transformation of 2,4,6-trinitrotoluene in soil in the presence of the earthworm eisenia andrei. Environ Toxicol Chem 19:1473–1480
- Riddle SG, Tran HH, Dewitt JG, Andrews JC (2002) Field, laboratory, and x-ray absorption spectroscopic studies of mercury accumulation by water hyacinths. Environ Sci Technol 36:1965
- Rizzi L, Petruzzelli G, Poggio G, Vigna G (2004) Soil physical changes and plant availability of zn and pb in a treatability test of phytostabilization. Chemosphere 57(9):1039–1046
- Robinson BH, Brooks RR, Howes AW, Kirkma JH, Gregg PEH (1997a) The potential of the high- biomass nickel hyperaccumulator *Berkheya coddii* for phytoremediation and phytomining. J Geochem Explor 60:115–126
- Robinson BH, Chiarucci A, Brooks RR, Petit D, Kirkman JH, Gregg PEH, De Dominicis V (1997b) The nickel hyperaccumulator plant *Alyssum bertolonii* as a potential agent for phytoremediation and phytomining of nickel. J Geochem Explor 59:75–86
- Roper JC, Dec J, Bollag J (1996) Using minced horseradish roots for the treatment of polluted waters. J Environ Qual 25:1242–1247
- Rosenfeld I, Beath OA (1964) Selenium- geobotany, biochemistry, toxicity and nutrition. Academic Press, New York
- Rout GR, Samantary S, Das P (1999) Chromium, nickel and zinc tolerance in *Leucaena leucocephala* (K8). Silvae Genet 48:151–157
- Rugh CL, Senecoff JF, Meagher RB, Merkle SA (1998) Development of transgenic yellow poplar for mercury phtoremediation. Nature Biotechnol 16:925–928
- Rugh CL, Wilde HD, Stack NM, Thompson DM, Summers AO, Meagher RB (1996) Mercuric ion reduction and resistance in transgenic *Arabidopsis thaliana* plants expressing a modified bacterial merA gene. Proc Natl Acad Sci USA 93:3182–3187
- Sahi SV, Bryant NL, Sharma NC, Singh SR (2002) Characterization of a lead hyperaccumulator shrub, *Sesbania drummondii*. Environ Sci Technol 36(21):4676–4680
- Salt DE, Blaylock M, Kumar PBAN, Dushenkov V, Ensley BD, Chet I, Raskin I (1995a) Phytoremediation: a novel strategy for the removal of toxic metals from the environment using plants. Biotechnology 13:468–475
- Salt DE, Prince RC, Pickering IJ, Raskin I (1995b) Mechanism of cadmium mobility and accumulation in Indian mustard. Plant Physiol 109:1426–1433
- Sari A, Tuzen M (2009) Kinetic and equilibrium studies of biosorption of Pb(II) and Cd(II) from aqueous solution by macrofungus (Amanita rubescens) biomass. J Hazard Mater 164:1004–1011
- Schnoor JL (1997) Phytoremediation: technical and organisatoric issues, key factors. Ground-Water Remediation Technologies Analysis Center, Pittsburgh
- Scullion J (2006) Remediating polluted soils. Naturwissenschaften 93:51-65
- Scullion J, Malik A (2000) Earthworm effects on aggregate stability, organic matter composition and disposition, and their relationships. Soil Biol Biochem 32:119–126
- Sebastiani L, Scebba F, Tognetti R (2004) Heavy metal accumulation and growth responses in poplar clones Eridano (*Populus deltoides x maximowiczii*) and I-214 (*P. x euramericana*) exposed to industrial waste. Environ Exp Bot 52:79–88
- Sekhar KC, Kamala CT, Chary NS, Sastry ARK, Rao TN, Vairamani M (2004) Removal of lead from aqueous solutions using an immobilized biomaterial derived from a plant biomass. J Hazard Mater 108:111–117
- Sheoran V, Sheoran AS, Poonia P (2008) Remediation techniques for contaminated soil. Environ Enging Managt J 7(4):379–387
- Sheoran AS, Sheoran V, Choudhary RP (2010) Bioremediation of acid-rock drainage by sulphate-reducing prokaryotes: a review. Miner Eng 23(14):1073–1100
- Sheoran V, Sheoran AS, Poonia P (2011) Role of hyperaccumulators in phytoextraction of metals from contaminated mining sites: a review. Crit Rev Environ Sci Technol 41:168–214

- Sheoran V, Sheoran AS, Poonia P (2012) Phytoremediation technologies for the reclamation of organic and inorganic polluted soils and water: a review. Environ Res J 6(4/5):1–23
- Shu WS, Lan CY, Zhang ZQ, Wong MH (2000) Use of vetiver and other three grasses for revegetation of Pb/Zn Mine tailings at Lechang, Guangdong Province: field experiment. In: 2nd international vetiver conference, Bangkok, Thailand
- Sikdar SK, Grosse D, Rogut I (1998) Membrane technologies for remediating contaminated soils: a critical review. J Membrane Sci 151:75–85
- Singer AC, van der Gast CJ, Thompson IP (2005) Perspectives and vision for strain selection in bioaugmentation. Trends Biotech 23:74–77
- Singh OV, Jain RK (2003) Phytoremediation of toxic aromatic pollutants from soil. Appl Microbiol Biotechnol 63:128–135
- Smith RAH, Bradshaw AD (1992) Stabilisation of toxic mine wastes by the use of tolerant plant populations. Trans Inst Min Metall Sect A 81:230–237
- Smrkolj P, Osvald M, Osvald J (2007) Selenium uptake and species distribution in *Seliniumaseolus vulgaris* seeds obtained by two different cultivations. Eur Food Res Technol 225:233–237
- Song SQ, Zhou X, Wu H, Zhou YZ (2004) Application of municipal garbage compost on revegetation of tin tailings dams. Rural Eco-Environ 20(2):59–61
- Srivastava M, Ma LQ, Contruva JA (2005) Uptake and distribution of selenium in different fern species. Int. J Phytorem 7:33–42
- Sui H, Li X, Jiang B, Huang G (2007) Simulation of remediation of multiple organic contaminats system by bioventing. Huagong Xuebo (Chinese Edition). 58:1025–1031
- Suko T, Fujikawa T, Miyazaki T (2006) Transport phenomena of volatile solute in soil during bioventing technology. J ASTM Int 3:374–379
- Susarla S, Medina VF, McCutcheon SC (2002) Phytoremediation: an ecological solution to organic chemical contamination. Ecol Engineer 18:647–658
- Suthersan SS (1997) Remediation engineering: design concepts. Lewis Publishers, Boca Raton
- Tang J, Zhu W, Kookana R, Arata K (2013) Characteristics of biochar and its applications in remediation of contaminated soils. J Biosci Bioeng 116(6):653–659
- Tang SR, Huang CY, Zhu ZX (1997) *Commelina communis* L.: copper hyperaccumulator found in Anhui Province of China. Pedosphere 7(3):207–210
- Terry N, Carlson C, Raab TK, Zayed A (1992) Rates of selenium volatilization among crop species. J Environ Qual 21:341–344
- Urlings LGCM (1990) In situ cadmium removal-full scale remedial action of contaminated soil. In: International symposium on hazardous waste treatment: treatment of contaminated soil, air, waste association and US.EPA Risk education laboratory, Cincinnati, Ohio, 5–8 Feb 1990
- USEPA (1998) Bioventing. Office of the Underground Storage Tank U.S. Environmental Protection Agency, Publication EPA, 510-B-95-007
- USEPA (1994) Selection of control technologies for remediation of soil contaminated with arsenic, cadmium, chromium, lead or mercury. Revised Draft Engineering Bulletin, Jan 31
- USEPA (1996) Engineering bulletin: technology alternatives for the remediation of soils contaminated with arsenic, cadmium, chromium, mercury and lead. U.S. Environmental Protection Agency, Office Of Emergency And Remedial Response, Cincinnati, OH
- Van denhove H (2013) Phytoremediation options for radioactively contaminated sites evaluated. Ann Nucl Energy 62:596–606
- Vernay P, Gauthier-Moussard C, Hitmi A (2007) Interaction of bioaccumulation of heavy metal chromium with water relation, mineral nutrition and photosynthesis in developed leaves of *Lolium perenne* L. Chemosphere 68:1563–1575
- Vidali M (2001) Bioremediation, An overview. Pure Appl Chem 73:1163-1172
- Vinterhalter B, Vinterhalter D (2005) Nickel hyperaccumulation in shoot cultures of Alyssum narkgrafii. Biol Plant 49:121–124
- Wan QF, Deng DC, Bai Y, Xia CQ (2012) Phytoremediation and electrokinetic remediation of uranium contaminated soils: a review. He-Huaxue yu Fangshe Huaxue. J Nucl Radiochem 34:148–156

- Wang J, Feng X, Anderson CWN, Xing Y, Shang L (2012) Remediation of mercury contaminated sites- a review. J Hazard Mater 221–222:1–18
- Wang S, Catherine NM (2004) An evaluation of surfactant technology in remediation of ctaminated soil. Chemosphere 57:1079–1089
- Wenzel WW, Adrino DC, Salt D, Smith R (1999) Phytoremediation: a plant-microbe-based remediation system. In: Adrino DC, Bollag JM, Frankenberger WT, Sims RC (eds) Bioremediation of contaminated soil, pp 456–508. Agronomy Monograph no. 37, Madison
- Whiting SN, Reeves RD, Richards D, Johnson MS, Cooke JA, Malaisse F, Paton A, Smith JAC, Angle JS, Chaney RL, Ginocchio R, Jaffre T, Johns R, Mcintyre T, Wojcik M, Tukiendorf A (2005) Cadmium uptake, localization and detoxification in *Zea mays*. Biol Plant 49:237–245
- Williams GM (1988) Integrated studies into ground water pollution by hazardus wastes. In: Gronow JR, Scho-field AN, Jain RK (eds) Land disposal of hazardous waste, engineering and environmental issues. Horwood Ltd. Chichester
- Woelders J (1998) Sanirengsmogelijkheden Cadmium: In situ Reiniging Van Cadmium houdede zandground. In Cadmium: Vooromen impact en sanering. Lisec, Genk, Belgium
- Wong MH (1982) Metal co-tolerance to copper, lead and zinc in *Festuca rubra*. Environ Res 29:42–47
- Wood P (2001) Remediation methods for contaminated land. In: Hester RE, Harrison RM (eds) Assessment and reclamation of contaminated land. Issues in environmental science and technology. pp 115–139. Royal Society f Chemistry, Cambridge
- Xue SG, Chen YX, Reeves RD, Lin Q, Fernando DR (2004) Mangenese uptake and accumulation by the hyperaccumulator plant *Phytolacca acinosa Roxb*. (Phytolaccaeae). Environ Pollut 131:393–399
- Yang X, Baligar DC, Martens DC, Clark RB (1996) Plant tolerance to nickel toxicity: I. Influx, transport, and accumulation of nickel in four species. J Plant Nutr 19:73–85
- Yang XE, Long XX, Ni WZ (2002) Physiological and molecular mechanisms of heavy metal uptake by hyperaccumulting plants. Plant Nutr Fertilizer Sci 8(1):8–15
- Yang ZY, Yuan JG, Xin GR, Chang HT, Wong MH (1997) Germination, growth and nodulation of *Sesbania rostrata* grown in Pb/Zn mine tailings. Environ Manage 21:617–622
- Ye ZH, Baker AJM, Wong MH, Willis AJ (1997) Zinc, lead and cadmium tolerance, uptake and accumulation by *Typha latifolia*. New Phytol 136:469–480
- Ye ZH, Wong MH, Baker AJM, Willis AJ (1998) Comparison of biomass and metal uptake between two populations of *Phragmites australis* grown in flooded and dry conditions. Ann Bot 80:363–370
- Yong-pisanphop J, Kruatrachue M, Pokethitiyook P (2005) Toxicity and accumulation of lead and chromium in *Hydrocotyle umbellate*. J Environ Biol 26:79–89
- Yu X-Z, Gu J-D (2008) The role of EDTA in Phytoextraction of hexavalent and trivalent chromium by two willow trees. Ecotoxicology 17:143–152
- Zhao FJ, Dunham SJ, McGrath SP (2002) Arsenic hyperaccumulation by different fern species. New Phytol 156:27–31
- Zhu YL, Zayed AM, Quian JH, De Souza M, Terry N (1999) Phytoaccumulation of trace elements by wetland plants: II. Water hyacinth. J Environ Qual 28:339–344
- Zou JH, Wang M, Jiang WS, Liu DH (2006) Chromium accumulation and its effect on other mineral elements in amaranthus viridis L. Acta Biol Crac Ser Bot 48:7–12