Metal–Microbe Interaction and Bioremediation

S. R. Joshi, Debajit Kalita, Rakshak Kumar, Macmillan Nongkhlaw and Pynskhem Bok Swer

Abstract Bioremediation of heavy metals and radionuclides has shown immense promise as an alternative approach for cleaning up, especially the biosphere component of the environment. The ubiquitous nature of microbes has made them the pioneers in any bioremediation approaches. Understanding the working mechanism of these microorganisms either independently, or as a community in relation to their interaction with heavy metal will enlighten and improve the use of bioremediation for environmental cleanup. The discovery of plants that have metal accumulation potential has opened fresh avenues for bioremediation. Plant-based bioremediation is still at a nascent stage, but it has attracted considerable attention in recent years. A concerted approach of using plants and microbes for bioremediation is another strategy that might work efficiently as both can complement each other through various interactions. Moreover, genetic engineering can be used to improve the bioremediation capacity of both plants and microbes and further improve their capacity in bioremediation of heavy metals and radionuclide.

Keywords Bioremediation · Microbes · Heavy metals · Radionuclides

Contents

S. R. Joshi (⊠) · D. Kalita · R. Kumar · M. Nongkhlaw · P. B. Swer Department of Biotechnology and Bioinformatics, Microbiology Laboratory, North Eastern Hill University, Shillong 793022, Meghalaya, India e-mail: srjoshi2006@yahoo.co.in

1 Introduction

The continuous growth of human population has resulted in the increasing demand for basic life-supporting commodities worldwide. There has also been a concomitant increase in the industrialization process to meet the basic human demands. Furthermore, this has led to the increase in energy demands that are partly met by nuclear energy. Nuclear energy is an important alternative energy source which accounts for 17 % of the overall electricity of the world. As a consequence, mining for radioactive elements like uranium is crucial, which has led to the large amounts of toxic chemicals being released into the environment either deliberately or accidentally. These toxic contaminants include radionuclides like uranium and heavy metals such as lead, cadmium arsenic, and mercury. Heavy metals like copper, zinc, cobalt, and iron are essential micronutrients for both plants and microorganisms, but if present at higher concentration, they can impair various metabolic processes. Other metals like lead, cadmium, mercury, and uranium have no known biological functions and are toxic even at lower concentrations. Inorganic contaminants like heavy metals and radionuclide are not degradable and tend to accumulate in the environment for a long time (Sarma et al. [2004](#page-14-0); Renshaw et al. [2005\)](#page-14-0). Several technologies have been developed to address the problem of heavy metal and radionuclide contamination as they pose serious ecological and health hazards. To compensate the higher cost involved in the newer technologies, alternative methods have been explored to address this issue.

Natural habitats harbor abundant and diverse type of microorganisms which can be explored for bioremediation processes. However, there is a need for the identification and characterization of the microbial strains that reveal high metal accumulation capacity and specificity. Uranium-tolerant bacteria have been isolated from various uranium-mining sites and deposits (Kumar et al. [2011,](#page-13-0) [2013a;](#page-13-0) Sarma et al. [2012,](#page-14-0) [2013\)](#page-14-0). Bacteria from uranium deposits have been characterized for their metal tolerance and uranium binding (Kumar et al. [2011](#page-13-0), [2013a;](#page-13-0) Sarma et al. [2012,](#page-14-0) [2013](#page-14-0)). Understanding and exploring the microbe–metal interaction have resulted in an upsurge in the research interest with their importance in various high-throughput biotechnological applications such as biosensor, biofuel cells, and most promisingly in microbe-mediated nanomaterial synthesis (Devi and Joshi [2012\)](#page-11-0). Identification of the microbial ligands/cellular processes involved in metal sequestration has lead to the development of engineered organisms with various cell surface displays that facilitate their applications in industrial catalysis, biosorption, bioremediation, biofuel, and biosensor technology (Mandal et al. [2006\)](#page-13-0). Metal nanocrystal synthesis through microbial process is another very promising aspect with importance in metal bioremediation and synthesis of nanoparticles for diverse applications.

2 Metal-Tolerant Microbes

A plethora of microorganisms capable of efficiently degrading toxic compounds in the environment have either been isolated or engineered. One of the prominent bacteria which exhibits extreme radiation resistance is Deinococcus radiodurans. It was first discovered in 1956 by Arthur W. Anderson while trying to sterilize meat by irradiating it with gamma radiation (Anderson et al. [1956](#page-11-0)). For many years, D. radiodurans has fascinated biologists by its extraordinary resistance to ionizing radiation. This bacterium, as a polyploid, can withstand radiation exposures of up to \sim 17,000 gray (Gy) (Daly et al. [1995,](#page-11-0) [1996](#page-11-0), [1997\)](#page-11-0). D. radiodurans also dominate the arid environments over other less resilient species by their capacity to regrow after rehydration. Besides UV and IR, D. radiodurans is resistant to lethal and mutagenic agents like mitomycin C (Sweet and Moseley [1976\)](#page-15-0), an antibiotic known to cross-link DNA.

Genome sequence of the highly IR-resistant *D. radiodurans* reveals the same number and types of DNA repair proteins as the IR-sensitive bacteria (Makarova et al. [2001,](#page-13-0) [2007](#page-13-0)). Following irradiation with lethal doses of IR, radiation-resistant organisms suffer from similar levels of genomic damage as the sensitive ones. This is due to the ability of the resistant strains to survive the formation of hundreds of IR-induced double-strand breaks (DSBs) per genome (Argueso et al. [2008;](#page-11-0) Gladyshev and Meselson [2008](#page-12-0)). Studies have shown that a highly IR-sensitive mutant of D. radiodurans that contains a mutated DNA polymerase I gene (polA) was fully restored by expression of the corresponding gene of the IR-sensitive Escherichia coli (Gutman et al. [1994\)](#page-12-0) The enzymes that mediate DNA repair in *D. radiodurans* are, therefore, probably not unique. Surprisingly, the mechanisms of IR resistance in Deinococcus spp. remain unclear along with their unique ability to resist desiccation (Cox and Battista [2005\)](#page-11-0) and UV radiation (Gutman et al. [1994;](#page-12-0) Minton [1994\)](#page-13-0).

D. radiodurans has been studied for its ability to detoxify or immobilize metallic pollutants. This was made possible by genetic engineering to obtain radiation-resistant organisms that can simultaneously detoxify metals. The merA gene encodes mercuric ion reductase that reduces highly toxic, thiol-reactive mercuric ion, $Hg(II)$, to much less toxic and volatile $Hg(0)$ (Schottel [1978;](#page-15-0) Summers [1986\)](#page-15-0). The merA locus from E. coli was cloned into D. radiodurans to confer both metal resistance and metal-remediating capabilities. In highly irradiating environments, cells that expressed the merA protein were better protected from the effects of $Hg(II)$ than the wild-type counterparts. Cr(VI), a potent carcinogen, is another heavy metal commonly found in radioactive waste sites. In this case as well, genetically engineered D. radiodurans capable of reducing Cr(VI) to Cr(III) was preferentially used for rendering it non-mutagenic and non-carcinogenic (Brim et al. [2006\)](#page-11-0). In general, the ability of a microorganism to resist the toxic effect of metals is frequently associated with its ability to transform those metals into less toxic chemical states.

The limitations associated with *D. radiodurans* for in situ bioremediation of nuclear waste sites are the requirements to genetically engineer the bacterium to

acquire toxic metal resistance and bioremediating capabilities. The culturing of this bacterium also requires a constant supply of carbon nutrient source and is therefore prone to contamination. The highly radiation-resistant eukaryote counterpart of D. radiodurans is found in the form of a microalga, Coccomyxa actinabiotis (Rivasseau et al. [2013\)](#page-14-0). It can withstand ionizing radiation doses up to 20,000 Gy with half the population able to survive at 10,000 Gy. Metabolic activity of the cell is marginally affected by radiation doses up to 10,000 Gy and a complete recovery of cellular functioning within a few days. This unique microalga also has the capacity to fix radionuclides such as 238 U, 137 Cs, 110 Ag, 60 Co, 54 Mn, 65 Zn, and 14 C via metabolically inactive and active processes even in highly radioactive environments. The main advantage of using photosynthetic organisms is the minimal requirement for energy and culture media, which directly alleviates the problem of bacterial contamination. This newly discovered organism therefore offers great potential for the bioremediation of highly toxic radioactive wastes. Detailed studies with respect to elucidating its metabolic activity and its capability to remediate radionuclides are not only necessary but also inevitable.

Sulfate-reducing bacteria (SRB) are nonpathogenic anaerobic prokaryotes known for their non-photosynthetic activity to generate ATP through electrontransfer-coupled phosphorylation. During this process, SRB uses sulfate as the terminal electron acceptor for respiration of hydrogen to produce sulfide. The sulfide produced is highly reactive and toxic, and therefore, SRB are able to cause severe corrosion of metals in a water system by producing enzymes which can accelerate the reduction of sulfate compounds to hyrdrogen sulfide (Little [1998;](#page-13-0) Beaton [2007\)](#page-11-0). SRB are used in several in situ technologies like in acid mine drainage (AMD). Extensive mining activity is responsible for changing the basic property of water. When pyrite-containing rocks come into contact with surface water or groundwater, under oxidizing conditions, these rocks produce sulfuric acid and dissolved iron. This acidic water in turn dissolves other metals contained in the rock, resulting in low pH, metal-bearing water known as AMD, or acid rock drainage (ARD). Carbonate minerals may neutralize the acidity and bring the pH to approximately 7.0 to give rise to neutral mine drainage. SRB are used in AMD treatment with the purpose of producing sulfides (for metal sulfide precipitation) and generating alkalinity at the same time.

Microbe-mediated sulfate reduction coupled with organic matter (represented by CH2O) oxidation forms the chemical basis of SRB remediation

$$
2CH_2O(aq) + SO4^{-2} + H^+ \rightarrow H_2S + 2HCO_3
$$

It also involves the chemical reaction of metal (Me) precipitation:

$$
H_2S + Me^{+2} \rightarrow MeS + 2H^+
$$

Precipitation of cadmium, copper, iron, lead, mercury, nickel, and zinc is facilitated by the formation of respective metal sulfides. In addition, arsenic, antimony, and molybdenum form more complex sulfide minerals (Figueroa [2005\)](#page-12-0). Co-precipitation with other metal sulfides can also be achieved for metals such as manganese, iron, nickel, copper, zinc, cadmium, mercury, and lead (Figueroa [2005\)](#page-12-0). Other SRB species are known to reduce certain metals to a more insoluble form, like reduction of uranium (VI) to uranium (IV) (Spear et al. [2000](#page-15-0)). Sulfate reduction by SRB also consumes acidity, as a result of which there is an increase in the pH. The above precipitation reactions for metal hydroxides are facilitated by increasing the pH (Gadd [2004\)](#page-12-0).

SRB are known to have a major negative economic impact on the petroleum industry because of their involvement in biocorrosion of ferrous metals in anaerobic environments (Hamilton [2003\)](#page-12-0). Hydrocarbons in petroleum (e.g., benzene, toluene, ethylbenzene, xylenes, naphthalene, phenanthrene, and alkanes) may also serve as electron donors in the normal metabolism of SRB resulting in sulfide production. This biogenic sulfide production typically results in metal corrosion and reservoir souring. In addition, it can also result in the acidulation and plugging of petroleum reservoirs and biocorrosion of metal surfaces of pipelines and tanks (Nemati et al. [2001](#page-14-0)). Due to the explosive nature of the sulfide, this may pose a risk at high concentrations. The accumulation of SRB biomass thus causes a reduction in the oil recovery (Muyzer and Stams [2008](#page-14-0)). The need to control or inhibit the growth of SRB in petroleum industries is usually achieved by biocide dosage (Korenblum et al. [2010](#page-13-0)). Though effective, the inherent problems associated with biocides are the occurrence of antimicrobial resistance (Stewart and Costerton [2001;](#page-15-0) Fraise [2002\)](#page-12-0), the residual concentration, toxicity, and persistence of biocides in industrial effluents. Alternative strategies for SRB control are therefore of great interest to the petroleum industry (Korenblum et al. [2013\)](#page-13-0).

Remediation of radionuclide or radioactive wastes through microbial processes is an emerging field of research. It has been suggested by current researches that improper treatment approaches can lead to negative impacts on environment and biodiversity, which may even increase distribution of radioactive materials (e.g., wind-aided transport of plutonium-contaminated soil) (Whicker et al. [2004\)](#page-16-0). Microbial consortium is a biological tool widely used for the remediation of pollutants, consisting of several species of microorganisms in the form of bioflocculant. Biofilms produced by microbes, which exist predominantly in natural environments (\sim 99 %) (Costerton et al. [1995\)](#page-11-0), have the capability to immobilize metals. The different mechanisms adopted by biofilms to immobilize metals or radionuclides are as follows: (1) biosorption to cell components or extracellular polymeric substances (EPS), (2) bioaccumulation, (3) precipitation by reaction with inorganic ligands such as phosphate, and (4) microbial reduction of soluble metal to insoluble form (Gorby and Lovley [1992;](#page-12-0) Merroun et al. [2003](#page-13-0); Renninger et al. [2004](#page-14-0)). Microbial activity can influence the release of radionuclides by altering bulk pore water chemistry (especially pH and redox reaction), by producing organic complexing ligands or by direct accumulation onto or into cells (West et al. [2002](#page-15-0)). Microbes can also cause corrosion and hence potentially affect the longevity of the metal waste containers in a repository (Stroes-Gascoyne et al. [2007\)](#page-15-0).

3 Bioremediation: Plant–Microbe Interaction Perspective

The concept of using plants to clean up the environment has generated considerable interest in the last few decades. With the discovery of some plant that has high metal-accumulating capacity (hyperaccumulators) such as *Thlaspi caerules*cence and Alyssum murales, cleaning up of metal-contaminated sites using these plants seems like a promising strategy. Metal accumulation in plant biomass constitutes a subclass of phytoremediation called phytoextraction (Raskin and Eansley [2000\)](#page-14-0). Approximately 400 plants have been identified so far which have potential for phytoextraction. Plants species such as Thlaspi sp and Alyssum spp. from the family Brassicaceae, Viola calaminaria and Astragalus racemosus from Violaceae and Leguminosae have been found to accumulate high concentration of heavy metals and radionuclides (Negri and Hinchman [2000](#page-14-0); Reeves and Baker [2000\)](#page-14-0). Metal-accumulating phenotype in plants is a complex mechanism that requires a concerted effort of tolerance, translocation, and sequestration of targeted metal/s (Hall [2002;](#page-12-0) Eapen and D'Souza [2005](#page-11-0)). Understanding the working mechanism of these areas will help enable or improve the metal-accumulating property in hyperaccumulator or even non-accumulating plants. There have been efforts to understand the mechanism of metal accumulation in potential candidates like T. caerulescence and other members of Brassicaceae (Baker et al. [1994;](#page-11-0) Kramer et al. [1996;](#page-13-0) Salt et al. [1999](#page-14-0); Bert et al. [2000](#page-11-0); Zhao et al. [2002;](#page-16-0) Milner and Kochian [2008](#page-13-0)). However, naturally occurring hyperaccumulators lack certain qualities such as large biomass, fast growth, and habitat incompatibility which limit their use in phytoremediation (Eapen and D'Souza [2005;](#page-11-0) Kotrba et al. [2009\)](#page-13-0). The success of hyperaccumulation as a mean of cleaning up metal contaminated soil relies on the ability of these plants to tolerate high concentrations and wider metal resistant properties, possessing efficient transport mechanism for metal uptake to accumulation in deep or wide spread roots and aerial portions of the plant that can be easily harvested (Eapen and D'Souza [2005;](#page-11-0) Kotrba et al. [2009\)](#page-13-0).

Another important aspect of phytoremediation is the relationship of plants with microorganisms existing in the rhizosphere or within the plants itself (endophytes) (Glick [2003](#page-12-0), [2010;](#page-12-0) Kavamura and Esposito [2010](#page-12-0); Ma et al. [2011](#page-13-0)). Plant–microbe interactions are well-known relationships which have been studied thoroughly in laboratory and field studies. Hence, there is no surprise that plants and microbes existing in metal-contaminated site also use this relationship to thrive in stressful environment (Tokala et al. [2002;](#page-15-0) Gray and Smith [2005](#page-12-0)). Microorganisms like bacteria are specialists in dealing with metals as they have existed together long before any other higher life forms. Hence, microbes are better adapted and possess well-organized mechanism to deal with the presence or invasion of toxic metals. The various strategies that bacteria utilize in order to negate the presence or increase concentrations of metals include the efflux of metal by different transporters (Nucifora et al. [1989;](#page-14-0) Solioz and Odermatt [1995\)](#page-15-0), complexation inside the cell (Silver [1996](#page-15-0); Robinson [2008](#page-14-0)), bioprecipitation, and reduction to a less toxic state (Bosecker [1997\)](#page-11-0) (Fig. [1](#page-6-0)). Bacteria existing in metal-contaminated site can

Fig. 1 Diagrammatic representation of the mechanisms of metal/radionuclide–microbes interaction (adapted from Geissler [2007](#page-12-0))

influence the physicochemical properties of their habitat including metal(s) in more than one ways.

Bacterial communities have been known to solubilize or precipitate metals in soil, which in turn affects the bioavailability of metals. The bacterial capacity to change the availability of metal in soil can affect plants either by solubilizing the metals so that plant can easily absorb the metals or by precipitating the metals, affecting the survival of plants. Besides changing the bioavailability of metal(s), bacteria can influence the efficiency of phytoextraction by nitrogen fixation and secreting plant growth-promoting hormones such as IAA, siderophore, etc. (Glick [2010;](#page-12-0) Ma et al. [2011\)](#page-13-0). Plants on the other hand secrete nutrients such as amino acids, sugars, and other metabolites from the roots, which nourish different bacterial species in the rhizosphere.

4 Metagenomics of Metal-Contaminated Sites

The existence of plant–microbe interactions in metal-contaminated site is irrefutable, and various studies have been carried out to understand this relationship (Sriprang et al. [2003;](#page-15-0) Kuiper et al. [2004](#page-13-0); Wu et al. [2006](#page-16-0)). Moreover, the success of phytoremediation does not depend on plant alone but also on its interaction with the microorganisms in the rhizosphere (Whiting et al. [2001](#page-16-0)). Understanding the microbial communities existing in metal-contaminated sites is a prerequisite for understanding this relationship. Culturable bacteria have been studied in metal- and

radionuclide-contaminated site where diverse groups of bacteria have been isolated and identified (Roane and Kellogg [1996](#page-14-0); Selenska-Pobell et al. [2001a,](#page-15-0) [b;](#page-15-0) Shelobolina et al. [2004;](#page-15-0) Nedelkova et al. [2007](#page-14-0); Islam and Sar [2011](#page-12-0); Kumar et al. [2013a](#page-13-0)). However, culture-dependent methods do not give the true diversity and types of microorganism as \sim 99 % of bacteria cannot be cultured in laboratory (Pace [1997](#page-14-0); Torsvik and Øvreås [2002](#page-15-0)). With the improvement in metagenomic studies, total bacterial communities existing in metal- and radionuclide-contaminated sites have been explored (Selenska-Pobell et al. [2001a,](#page-15-0) [b](#page-15-0); Satchanska et al. [2004;](#page-15-0) Islam and Sar [2011;](#page-12-0) Kumar et al. [2013a](#page-13-0)). This method allows a better understanding on the existing relationship of these bacterial communities with the plant species in that habitat. Comparative metagenomics have been carried out to study the effects of heavy metals on the diversity of bacterial communities. Bacterial communities' structure in uranium deposits has also been studied using both culture-dependent and culture-independent techniques to obtain baseline knowledge on the bacterial communities prior to any mining activities (Kumar et al. [2013b\)](#page-13-0). Similarly, bacterial communities have been explored in uranium-mining sites using both culture-dependent and culture-independent methods. The rhizosphere tends to harbor diverse group of microorganisms as compared to the bulk soil. Hence, a comparative study of bacterial species between the rhizosphere and the bulk soil provides an idea about different bacterial species that contribute to the well-being of the plant(s). Bacterial species in the rhizosphere of hyperaccumulators plants such as T. caerulescence, A. murales, etc., have been studied and identified (Gremion et al. [2003](#page-12-0)). Besides the rhizosphere of hyperaccumulators, bacterial communities in the rhizosphere of pioneer plants in metal-contaminated sites have also been explored (Navarro-Noya et al. [2010\)](#page-14-0). Metagenomics has also been used to compare the effects of different metals on the diversity of bacterial communities (Sobolev and Begonia [2008](#page-15-0); Gołębiewski et al. [2013\)](#page-12-0). Besides understanding the community structure, metagenomics has also been in use to understand the different metabolism, evolution, and adaptation of microbial communities in different habitats. Functional metabolic markers such as nirS, nirK, dsrAB, amoA, pmoA, etc., have been used to understand the dominant metabolic activities in radionuclide contamination sites (Hemme et al. [2010\)](#page-12-0). Similarly, genes encoding for metal resistance were found to be prevalent and disseminated among those microorganism in metal-contaminated sites (Coombs and Barkay [2004](#page-11-0); Martinez et al. [2006;](#page-13-0) Nongkhlaw et al. [2012\)](#page-14-0) An in-depth and comprehensive study on the microbial communities and its function is very important before any bioremediation practice is established, and metagenomics is an important approach to achieve that goal.

5 Transgenic Metal-Tolerant Plants

The inherent property of some plants to accumulate metal(s) in their biomass has generated and opened new scope for heavy metal remediation. Plants with the capacity to accumulate 50–500 times metal in their biomass as compared to their counterpart growing in the same habitat are termed as hyperaccumulators and considered potent candidates for use in phytoremediation.

The success of hyperaccumulation as a means of cleaning up metal contaminated soil depends on various factors like metal tolerance capability, efficient transport mechanism, high biomass in aerial portion of the plant etc. So far, such ideal plants have not been discovered or do not exist. However, genetic engineering can be used to optimize the existing potential of hyperaccumulators or non-accumulators by introducing new traits from other sources.

With the increased knowledge in plant genetics procedures and metal-tolerant plants with established genetic makeup, B. juncea, Helianthus annuus, and Nicotiana glaucum appear good candidates for genetic engineering for the purpose of phytoextraction (Eapen and D'Souza [2005](#page-11-0); Kotrba et al. [2009\)](#page-13-0). Understanding the mechanism that involves in metal accumulation is very important before genetic manipulation. The ability of plants to accumulate metal(s) in their biomass involves well-organized mechanism that includes metal uptake from the soil, translocation to target compartments, and sequestration. Genetically engineered plants for phytoremediation usually involve the introduction of metal-tolerating genes from other plants, but genes from human, animal, fungi, and bacteria have also been used to improve metal accumulation (Eapen and D'Souza [2005;](#page-11-0) Kotrba et al. [2009\)](#page-13-0). Genes that encode metal sequestration factors such as metallothioneins (Misra and Gedamu [1998](#page-13-0); Evans et al. [1992](#page-12-0)) and phytochelatins (Zhu et al. [1999a](#page-16-0), [b;](#page-16-0) Harada et al. [2001](#page-12-0)) from other sources have been introduced in plants. Constitutive expression of genes encoding metallothioneins from mouse, human, and Chinese hamster in Nicotiana tabaccum, Brassica oleracea, and Arabidopsis thaliana showed increased $Cd²⁺$ tolerance but reduced metal accumulation in shoots. Similarly, genes from bacteria and fungi have been cloned and expressed in plants cells for obtaining metal-tolerant phenotype (Table [1\)](#page-9-0). Expression of yeast $CUPI$ increased Cd^{2+} tolerance in B. oleracea, but there is no increased accumulation. However, expression of yeast CUP1 increased $Cu⁺$ but not $Cd²⁺$ accumulation in leaves of N. tabaccum. A more promising Cd^{2+} accumulation in transgenic line was seen with recombinant HisCUP. Improved Hg^{2+} accumulation was also seen in A. *thaliana* transgenic expressing bacterial Hg^{2+} binding protein, MerP. In addition to the improvement of metal accumulation trait, plants also need to combat the oxidative stress known to be induced by heavy metals. Hence Glutathione synthesis genes from bacteria and fungi have been introduced and overexpressed in plants with the aim of increasing metal tolerance and sequestration (reviewed in Eapen and D'Souza [2005;](#page-11-0) Kotrba et al. [2009\)](#page-13-0). Maintenance of metal homeostasis carried out by various metal-transporting proteins is another important aspect for metal tolerance and accumulation. Similarly, metal transporters from bacteria and fungi have been cloned and expressed in plants. Zinc/Lead/Cadmium metal-transporting P_{IB} -ATPase (ZntA) from *E. coli* has been successfully expressed in *A. thaliana* (reviewed in Eapen and D'Souza [2005;](#page-11-0) Kotrba et al. [2009\)](#page-13-0). Similarly, mercury-resistant genes such as merA, merB, and merC have been successfully cloned in plant species

Table 1 Examples of transcenic plants expressing bacterial/fungal genes Table 1 Examples of transgenic plants expressing bacterial/fungal genes resulting in increased tolerance and accumulation of mercury by the transformed plant. Very few reports are available on the use of metal transporters from bacterial origin for making metal-tolerant transgenic plants except for those mentioned above.

Negri and Hinchman ([2000\)](#page-14-0) have reported the use of the plants for treatment of ${}^{3}H$, U, Pu, ${}^{137}Cs$, and ${}^{90}Sr$. Phytoextraction removes radionuclides from soil without destroying the soil structure having limited impact on soil fertility for the treatment of large areas of low-level contamination, and its success depends on the bioavailability of radionuclides in soil, on the rate of uptake by plant roots and transportation efficiency of the vascular system of plants (Slavik Dushenkov [2003\)](#page-11-0).

6 Conclusion

Microbial bioremediation is the process by which microbes degrade or transform hazardous organic compounds into non-toxic products. Since the plethora of microorganisms teeming in nature are not capable of degrading all toxic compounds, especially xenobiotics, the use of genetically modified organisms forms an indispensable part of bioremediation approaches with the advancement in genetic manipulation. Although genetically engineered microbes are quite promising, their implementation for in situ bioremediation still requires additional routes for developing safe steps to environmental cleanup. One of the major challenges is to optimize conditions and procedures for sustained and effective bioremediation in the presence of toxic metals and organic compounds. Conditions are created to enhance microbial activity for in situ biostimulation or bioaugmentation which may disrupt the natural microbiota. Various issues are to be dealt with to enhance the metabolic activity while maintaining the required growth conditions such as pH, temperature, the levels of contaminants and nutrients, etc. Due to the complex nature of interactions between microorganisms and radionuclides, it is far from easy to understand the wide range of environments these organisms inhabit. To study the molecular mechanisms and identify novel genes, proteins, and enzymes involved in the bioremediation of radionuclides necessitates the study toward the structural and functional interactions between proteins and other metabolites. Therefore, identification of potential genes and proteins involved in the metabolism of radionuclides can be achieved by advanced genomics and proteomics techniques. With the recent advances in next-generation sequencing, genomics, and proteomics, it has become possible to check for the expression of proteins and enzymes of interest with the potential for radionuclide resistance. Genome-wide transcriptome analysis can further provide detailed insight into better understanding of the metabolic pathways and the physiology of the microorganisms.

References

- Anderson A, Nordan H, Cain R, Parrish G, Duggan D (1956) Studies on a radioresistant micrococcus I. Isolation, morphology, cultural characteristics, and resistance to gamma radiation. Food Technol 10:575–578
- Argueso JL, Westmoreland J, Mieczkowski PA, Petes TD, Resnick MA (2008) Double-strand breaks associated with repetitive DNA can reshape the genome. Proc Natl Acad Sci USA 105:11845–11850
- Baker AJM, Reeves RD, Hajar ASM (1994) Heavy metal accumulation and tolerance in British populations of the metallophyte Thlaspi caerulescens J. & C. Presl (Brassicaceae). New Phytol 127:61–68
- Beaton E (2007) Understanding Mic in process water systems: Recent findings on its control. Corrosion Conference Nashville, Tennessee, 11–15 March, 2007
- Bennett LE, Burkhead JL, Hale KL, Terry N, Pilon P, Pilon-Smits EAH (2003) Analysis of transgenic Indian mustard plants for phytoremediation of metal-contaminated mine tailings. J Environ Qual 32:432–440
- Bert V, MacNair MR, Delaguerie P, Saumitou-Laprade P, Petit D (2000) Zinc tolerance and accumulation in metallicolous and nonmetallicolous populations of Arabidopsis halleri (Brassicaceae). New Phytol 146:225–233
- Bosecker K (1997) Bioleaching: metal solubilization by microorganisms. FEMS Microbiol 20:591–604
- Brim H, Osborne JP, Kostandarithes HM, Fredrickson JK, Wackett LP, Daly MJ (2006) Deinococcus radiodurans engineered for complete toluene degradation facilitates Cr(VI) reduction. Microbiology 152:2469–2477
- Coombs JM, Barkay T (2004) Molecular evidence for the evolution of metal homeostasis genes by lateral gene transfer in bacteria from the deep terrestrial subsurface. Appl Environ Microbiol 70:1698–1707
- Costerton JW, Lewandowski Z, Caldwell DE, Korber DR, Lappin-Scott HM (1995) Microbial biofilms. Annu Rev Microbiol 49:711–745
- Cox MM, Battista JR (2005) Deinococcus radiodurans—the consummate survivor. Nat Rev Microbiol 3:882–892
- Daly MJ, Minton KW (1995) Interchromosomal recombination in the extremely radioresistant bacterium Deinococcus radiodurans. J Bacteriol 177:5495–5505
- Daly MJ, Minton KW (1996) An alternative pathway of recombination of chromosomal fragments precedes recA-dependent recombination in the radioresistant bacterium Deinococcus radiodurans. J Bacteriol 178:4461–4471
- Daly MJ, Minton KW (1997) Recombination between a resident plasmid and the chromosome following irradiation of the radioresistant bacterium *Deinococcus radiodurans*. Gene 187:225–229
- De la Fuente JM, Ramirez-Rodriguez Y, Cabrera-Ponce JL, Herrera Estrella L (1997) Aluminium tolerance in transgenic plants by alteration of citrate synthesis. Science 276:1566–1568
- Devi LS, Joshi SR (2012) Antimicrobial and synergistic effects of silver nanoparticles synthesized using soil fungi of high altitudes of Eastern Himalaya. Mycobiology 40:27–34
- Dhankher OP, Li Y, Rosen BP, Shi J, Salt D, Senecoff JF, Sashti NA, Meagher RB (2002) Engineering tolerance and hyperaccumulation of arsenic in plants by combining arsenate reductase and γ -glutamylcysteine synthetase expression. Nat Biotechnol 20:1140–1145
- Dixit P, Singh S, Ramachandran V, Kiranmayi P, Eapen S (2010) Expression of Neurospora crassa zinc transporter gene in transgenic Nicotiana tabacum enhances plant zinc accumulation without co-transport of cadmium. Plant Cell Environ 33:1697–1707
- Dushenkov S (2003) Trends in phytoremediation of radionuclides. Plant Soil 249:167–75
- Eapen S, D'Souza S (2005) Prospects of genetic engineering of plants for phytoremediation of toxic metals. Biotechnol Adv 23:97–114
- Evans KM, Gatehouse JA, Lindsay WP, Shi J, Tommey AM, Robinson NJ (1992) Expression of the pea metallothionein like gene Ps MTA in Escherichia coli and Arabidopsis thaliana and analysis of trace metal ion accumulation: Implications of Ps MTA function. Plant Mol Biol 20:1019–1028
- Figueroa L (2005) Microbial ecology of anaerobic biosystems treating mining influenced waters. In: Mine water treatment technology conference, Pittsburgh, PA. 15–18 Aug 2005
- Fraise AP (2002) Biocide abuse and antimicrobial resistance—a cause for concern? J Antimicrob Chemother 49:11–12
- Gadd G (2004) Microbial influence on metal mobility and application for bioremediation. Geoderma 122:109–119
- Geissler A (2007) Prokaryotic microorganisms in uranium mining waste piles and their interactions with uranium and other heavy metals. PhD thesis, TU Bergakademie Freiberg, Freiberg, Germany
- Gladyshev E, Meselson M (2008) Extreme resistance of bdelloid rotifers to ionizing radiation. Proc Natl Acad Sci USA 105:5139–5144
- Glick BR (2003) Phytoremediation: synergistic use of plants and bacteria to clean up the environment. Biotechnol Adv 21:383–393
- Glick BR (2010) Using soil bacteria to facilitate phytoremediation. Biotechnol Adv 21:367–374
- Gołębiewski M, Deja-Sikora E, Cichosz M, Tretyn A, Wróbel B (2013) 16S rDNA Pyrosequencing analysis of bacterial community in heavy metals polluted soils. Microb Ecol. doi:[10.1007/](http://dx.doi.org/10.1007/s00248-013-0344-7) [s00248-013-0344-7](http://dx.doi.org/10.1007/s00248-013-0344-7)
- Gorby YA, Lovley DR (1992) Enzymatic U precipitation. Environ Sci Technol 26:205–207
- Gray EJ, Smith DL (2005) Intracellular and extracellular PGPR: commonalities and distinctions in the plant–bacterium signaling processes. Soil Biol Biochem 37:395–412
- Gremion F, Chatzinotas A, Harms H (2003) Comparative 16S rDNA and 16S rRNA sequence analysis indicates that actinobacteria might be a dominant part of the metabolically active bacteria in heavy metal contaminated bulk and rhizosphere soil. Environ Microbiol 5:896–907
- Grichko VP, Filby B, Glick BR (2000) Increased ability of transgenic plants expressing the bacterial enzyme ACC deaminase to accumulate Cd Co, Cu, Ni, Pb and zinc. J Biotechnol 81:45–53
- Guo J, Dai X, Xu W, Ma M (2008) Overexpressing GSH1 and AsPCS1 simultaneously increases the tolerance and accumulation of cadmium and arsenic in *Arabidopsis thaliana*. Chemosphere 72:1020–1026
- Gutman PD, Fuchs P, Minton KW (1994) Restoration of the DNA damage resistance of Deinococcus radiodurans DNA polymerase mutants by Escherichia coli DNA polymerase I and Klenow fragment. Mutat Res 314:87–97
- Hall JL (2002) Cellular mechanisms for heavy metal detoxification and tolerance. J Exp Bot 53:1–11
- Hamilton WA (2003) Microbial influenced corrosion as a model system for the study of metal microbe interactions: a unifying electron transfer hypothesis. Biofouling 19:65–76
- Harada E, Choi YE, Tsuchisaka A, Obata H, Sano H (2001) Transgenic tobacco plants expressing a rice cysteine synthase gene are tolerant to toxic levels of cadmium. J Plant Physiol 158:655–661
- Hemme CL, Deng Y, Gentry TJ, Fields MW et al (2010) Metagenomic insights into evolution of a heavy metal-contaminated groundwater microbial community.ISME J 4:660–672
- Hasegawa I, Terada E, Sunairi M, Wakita H, Shinmachi F, Noguchi A (1997) Genetic improvement of heavy metal tolerance in plants by transfer of the yeast metallothionein gene (CUPI). Plant Soil 196:277–281
- Islam E, Sar P (2011) Culture-dependent and -independent molecular analysis of the bacterial community within uranium ore. J Basic Microbiol 51:1–13
- Kavamura VN, Esposito E (2010) Biotechnological strategies applied to the decontamination of soils polluted with heavy metals. Biotechnol Adv 28:61–69
- Kiyono M, Oka Y, Sone Y, Tanaka M, Nakamura R, Sato MH, Pan-Hou H, Sakabe K, Inoue K (2012) Expression of the bacterial heavy metal transporter MerC fused with a plant SNARE,

SYP121, in Arabidopsis thaliana increases cadmium accumulation and tolerance. Planta 235:841–850

- Korenblum E, Valoni E, Penna M, Seldin L (2010) Bacterial diversity in water injection systems of Brazilian offshore oil platforms. Appl Microbiol Biotechnol 85:791–800
- Korenblum E, de Vasconcelos Goulart FR, de Almeida Rodrigues I , Abreu F, Lins U, Alves PB, Blank AF, Valoni E, Sebastián GV, Alviano DS, Alviano CS, Seldin L (2013) Antimicrobial action and anti-corrosion effect against sulfate reducing bacteria by lemongrass (Cymbopogon citratus) essential oil and its major component, the citral. AMB Express 3:44
- Kotrba P, Najmanova J, Macek T, Ruml T, Mackova M (2009) Genetically modified plants in phytoremediation of heavy metal and metalloid soil and sediment pollution. Biotechnol Adv 27: 799–810
- Kramer U, Cotter-Howells JD, Charnock JM, Baker AJM, Smith JAC (1996) Free histidine as a metal chelator in plants that accumulate nickel. Nature 379:635–638
- Kuiper I, Lagendijk EL, Bloemberg GV, Lugtenberg BJ (2004) Rhizoremediation: a beneficial plant-microbe interaction. Mol Plant-Microbe Interact 17:6–15
- Kumar R, Acharya C, Joshi SR (2011) Isolation and analyses of uranium tolerant Serratia marcescens strains and their utilization for aerobic uranium U(VI) bioadsorption. J Microbiol 49:568–574
- Kumar R, Nongkhlaw M, Acharya C, Joshi SR (2013a) Uranium (U)-tolerant bacterial diversity from U ore deposits of Domiasiat in North-East India and their prospective utilization in bioremediation. Microbes Environ 28:33–41
- Kumar R, Nongkhlaw M, Acharya C, Joshi SR (2013b) Soil bacterial metagenomic analysis from uranium ore deposit of Domiasiat in Northeast India. Curr Sci 105(4):495–499
- Lee J, Bae H, Jeong J, Lee JY, Yang YY, Hwang I, Martinoia E, Lee Y (2003) Functional expression of heavy metal transporter in *Arabidopsis* enhances resistance to and decreases uptake of heavy metals. Plant Physiol 133:589–596
- Little B, Wagner P, Hart K, Ray R, Lavoie D, Nealson K, Aguilar C (1998) The role of biomineralization in microbiologically influenced corrosion. Biodegradation 9:1–10
- Ma Y, Prasad MNV, Rajkumar M, Freitas H (2011) Plant growth promoting rhizobacteria and endophytes accelerate phytoremediation of metalliferous soils. Biotechnol Adv 29:248–258
- Makarova KS, Aravind L, Wolf YI, Tatusov RL, Minton KW, Koonin EV, Daly MJ (2001) Genome of the extremely radiation resistant bacterium *Deinococcus radiodurans* viewed from the perspective of comparative genomics. Microbiol Mol Biol Rev 65:44–79
- Makarova KS, Omelchenko MV, Gaidamakova EK, Matrosova VY, Vasilenko A, Zhai M, Lapidus A, Copeland A, Kim E, Land M, Mavrommatis K, Pitluck S, Richardson PM, Detter C, Brettin T, Saunders E, Lai B, Ravel B, Kemner KM, Wolf YI, Sorokin A, Gerasimova AV, Gelfand MS, Fredrickson JK, Koonin EV, Daly MJ (2007) Deinococcus geothermalis: the pool of extreme radiation resistance genes shrinks. PLoS One 9:e955
- Mandal D, Bolander ME, Mukhopadhyay D, Sarkar G, Mukherjee P (2006) The use of microorganisms for the formation of metal nanoparticles and their application. Appl Microbiol Biotechnol 69:485–492
- Martinez RJ, Wang Y, Raimondo MA, Coombs JM, Barkay T, Sobecky PA (2006) Horizontal gene transfer of PIB-type ATPases among bacteria isolated from radionuclide- and metalcontaminated subsurface soils. Appl Environ Microbiol 72:3111–3118
- Merroun M, Henning C, Rossberg A, Reich T, Selenska-Pobell S (2003) Characterization of U(VI)—acidithiobacillus ferroxidans complexes using EXAFS, transmission electron microscopy, and energy-dispersive X-ray analysis. Radiochim Acta 91:583–592
- Milner MJ, Kochian LV (2008) Investigating heavy-metal hyperaccumulation using *Thlaspi* caerulescens as a model system. Ann Bot 102:3–13
- Minton KW (1994) DNA repair in the extremely radioresistant bacterium Deinococcus radiodurans. Mol Microbiol 13:9–15
- Misra S, Gedamu L (1998) Heavy metal tolerant transgenic Brassica napus L and Nicotiana tabacum L plants. Theor Appl Genet 78:16–18
- Muyzer G, Stams AJM (2008) The ecology and biotechnology of sulphate reducing bacteria. Nat Rev Microbiol 6:441–454
- Navarro-Noya YE, Jan-Roblero J, González-Chávez MC, Hernández-Gama R, Hernández-Rodríguez C (2010) Bacterial communities associated with the rhizosphere of pioneer plants (Bahia xylopoda and Viguiera linearis) growing on heavy metals-contaminated soils. Antonie van Leeuwenhoek 97: 335–349
- Nedelkova M, Merroun ML, Rossberg A, Hennig C, Selenska-Pobell S (2007) Microbacterium isolates from the vicinity of a radioactive waste depository and their interactions with uranium. FEMS Microbiol Ecol 59:694–705
- Negri MC, Hinchman RR (2000) The use of plants for the treatment of radionuclides. In: Raskin I, Ensley ED (eds) Phytoremediation of toxic metals: using plants to clean up the environment. Wiley, New York
- Nemati M, Jenneman GE, Voordouw G (2001) Mechanistic study of microbial control of hydrogen sulfide production in oil reservoirs. Biotechnol Bioeng 74:424–434
- Nongkhlaw M, Kumar R, Acharya C, Joshi SR (2012) Occurrence of horizontal gene transfer of PIB-type ATPase genes among bacteria isolated from an uranium rich deposit of Domiasiat in North East India. PloS One 7(10):e48199. doi:[10.1371/journal.pone.0048199](http://dx.doi.org/10.1371/journal.pone.0048199)
- Nucifora G, Chu L, Misra TK, Silver S (1989) Cadmium resistance from Staphylococcus aureus plasmid pI258 cadA gene results from a cadmium-efflux ATPase. Proc Natl Acad Sci 86:3544–3548
- Pace NR (1997) A molecular view of microbial diversity and the biosphere. Science 276:734–740
- Raskin I, Ensley BD (2000) Phytoremediation of toxic metals: using plants to clean the environment 2002. Wiley, New York
- Reeves RD, Baker AJH (2000) Metal accumulating plants. In: Raskin I, Ensley ED (eds) Phytoremediation of toxic metals: using plants to clean up the environment. Wiley, New York
- Renninger N, Knopp R, Nitsche H, Clark DS, Keasling JD (2004) Uranyl precipitation by Pseudomonas aeruginosa via controlled polyphosphate metabolism. Appl Environ Microbiol 70:7404–7412
- Renshaw JC, Butchins LJ, Livens FR, May I, Charnock JM, LIoyd JR (2005) Bioreduction of uranium: environmental implications of a pentavalent intermediate. Environ Sci Technol 39:5657–5660
- Rivasseau C, Farhi E, Atteia A, Coute A, Gromova M, Saint Cyr DDG, Boisson A-M, Ferret A-S, Compagnon E, Bligny R (2013) An extremely radioresistant green eukaryote for radionuclide bio decontamination in the nuclear industry. Energy Environ Sci 6:1230–1239
- Roane TM, Kellogg ST (1996) Characterization of bacterial communities in heavy metal contaminated soils. Can J Microbiol 42:593–603
- Robinson NJ (2008) A bacterial copper metallothionein. Nat Chem Biol 4:582–583
- Salt DE, Prince RC, Baker AJM, Raskin I, Pickering IJ (1999) Zinc ligands in the metal hyperaccumulator Thlaspi caerulescens as determined using X-ray absorption spectroscopy. Environ Sci Technol 33:713– 7
- Samuelsen AI, Martin RC, Mok DWS, Machteld CM (1998) Expression of the yeast FRE genes in transgenic tobacco. Plant Physiol 118:51–58
- Sarma PM, Bhattacharya D, Krishnan S, Lal B (2004) Degradation of polycyclic aromatic hydrocarbons by a newly discovered enteric bacterium, Leclercia adecarboxylata. Appl Environ Microbiol 77:3163–3166
- Sarma B, Acharya C, Joshi SR (2012) Plant growth promoting and metal bioadsorption activity of metal tolerant Pseudomonas aeruginosa isolate characterized from uranium ore deposit. B Biol Sci, Proc Natl Acad Sci, India, Sect. doi[:10.1007/s40011-012-0136-8](http://dx.doi.org/10.1007/s40011-012-0136-8)
- Sarma B, Acharya C, Joshi SR (2013) Characterization of metal tolerant Serratia spp. isolates from sediments of uranium ore deposit of Domiasiat in Northeast India. Proc Natl Acad Sci India Sect B Biol Sci. doi[:10.1007/s40011-013-0236-0](http://dx.doi.org/10.1007/s40011-013-0236-0)
- Sasaki Y, Hayakawa T, Inoue C, Miyazaki A, Silver S, Kusano T (2006) Generation of mercuryhyperaccumulating plants through transgenic expression of the bacterial mercury membrane transport protein MerC. Transgenic Res 15:615–625
- Satchanska GE, Golovinsky E, Selenska-Pobell S (2004) Bacterial diversity in a soil sample from uranium mining waste pile as estimated via. a culture-independent 16SrDNA approach. Comptes Rendus de l'Academie Bulgare des Sciences 57:75–82
- Schottel JL (1978) The mercuric and organomercurial detoxifying enzymes from a plasmidbearing strain of Esherichia coli. J Biol Chem 253:4341–4349
- Selenska-Pobell S, Kampf G, Flemming K, Radeva G, Satchanska G (2001a) Bacterial diversity in soil samples from two uranium waste piles as determined by rep-APD, RISA and 16S rDNA retrieval. Anton Leeuw Int J G 79:149–161
- Selenska-Pobell S, Kampf G, Flemming K, Radeva G, Satchanska G (2001b) Bacterial diversity in soil samples from two uranium waste piles as determined by rep-APD, RISA and 16S rDNA retrieval. Anton Leeuw Int J G 79:149–161
- Shelobolina E, Sullivan S, Neill KO, Nevin K, Lovley D (2004) Isolation, characterization, and U(VI)-reducing potential of a facultatively anaerobic, acid resistant bacterium from low-pH, nitrate- and U(VI)-contaminated subsurface sediment and description of Salmonella subterranaea sp. nov. Appl Environ Microbiol 70:2959–2965
- Silver S (1996) Bacterial resistance to toxic metal ions—a review. Gene179:9–19
- Singh S, Korripally P, Vancheeswaran R, Eapen S (2011) Transgenic Nicotiana tabacum plants expressing a fungal copper transporter gene show enhanced acquisition of copper. Plant Cell Rep 30:1929–1938
- Sobolev D, Begonia MFT (2008) Effects of heavy metal contamination upon soil microbes: lead-induced changes in general and denitrifying microbial communities as evidenced by molecular markers. Int J Environ Res Public Health 5(5):450–456
- Solioz M, Odermatt A (1995) Copper and silver transport by CopB-ATPase in membrane vesicles of Enterococcus hirae. J Biol Chem 270:9217–9221
- Song WY, Sohn EJ, Martinoia E, Lee YJ, Yang YY, Jasinski M, Forestier C, Hwang I, Lee Y (2003) Engineering tolerance and accumulation of lead and cadmium in transgenic plants. Nat Biotechnol 21:914–919
- Spear JR, Figueroa LA, Honeyman BD (2000) Modeling the removal of uranium U(VI) from aqueous solution in the presence of sulfate reducing bacteria. Environ Sci Technol 66: 3711–3721
- Sriprang R, Hayashi M, Ono H, Takagi M, Hirata K, Murooka Y (2003) Enhanced accumulation of Cd2 by a *Mesorhizobium* sp. transformed with a gene from *Arabidopsis thaliana* coding for phytochelatin synthase. Appl Environ Microbiol 69:1791–1796
- Stewart PS, Costerton JW (2001) Antibiotic resistance of bacteria in biofilms. Lancet 358: 135–138
- Stroes-Gascoyne S, Schippers A, Schwyn B, Poulain S, Sergeant C, Simonoff M, Le Marrec C, Altmann S, Nagaoka T, Mauclaire L, McKenzie J, Daumas S, Vinsot A, Beaucaire C, Matray JM (2007) Microbial community analysis of Opalinus clay drill core samples from the Mont Terri underground research laboratory, Switzerland. Geomicrobiol J 24:1–17
- Summers AO (1986) Organization, expression, and evolution of genes for mercury resistance. Annu Rev Microbiol 40:607–634
- Sweet DM, Moseley BE (1976) The resistance of *Micrococcus radiodurans* to killing and mutation by agents which damage DNA. Mutat Res 34:175–186
- Thomas JC, Davies EC, Malick FK, Endreszi C, Williams CR, Abbas M (2003) Yeast metallothionein in transgenic tobacco promotes copper uptake from contaminated soils. Biotechnol Prog 19: 273–280
- Tokala RK, Strap JL, Jung CM, Crawford DL, Salove H, Deobald LA, et al (2002) Novel plantmicrobe rhizosphere interaction involving S. lydicus WYEC108 and the pea plant (Pisum sativum). Appl Environ Microbiol 68:2161–2171
- Torsvik V, Øvreås L (2002) Microbial diversity and function in soil: From genes to ecosystems. Curr Opin Microbiol 5:240–245
- West JM, Mckinley IG, Stroes-Gascoyne S (2002) Microbial effects on waste repository materials. In: Keith-Roach M, Livens F (eds) Interactions of microorganisms with radionuclides. Elsevier Sciences, Oxford
- Whicker FW, Hinton TG, MacDonell MM, Pinder JE III, Habegger LJ (2004) Avoiding destructive remediation at DOE sites. Science 303:1615–1616
- Whiting SN, de Souza MP, Terry N (2001) Rhizosphere bacteria mobilize Zn for hyperaccumulation by Thlapsi caerulescens. Environ Sci Technol 35:3144–3150
- Wu CH, Wood TK, Mulchandani A, Chen W (2006) Engineering plant-microbe symbiosis for rhizoremediation of heavy metals. Appl Environ Microbiol 72:1129–1134
- Zhao FJ, Hamon RE, Lombi E, McLaughlin MJ, McGrath SP (2002) Characteristics of cadmium uptake in two contrasting ecotypes of the hyperaccumulator Thlaspi caerulescens. J Exp Bot 53:535–543
- Zhu YL, Pilon-Smits EA, Jouanin L, Terry N (1999a) Overexpression of glutathione synthetase in Indian mustard enhances cadmium accumulation and tolerance. Plant Physiol 119:73–80
- Zhu YL, Pilon-Smits EA, Tarun AS, Weber SU, Jouanin L, Terry N (1999b) Cadmium tolerance and accumulation in Indian mustard is enhanced by overexpressing γ -glutamylcysteine synthetase. Plant Physiol 121:1169–1178