

IL-17A Family, Receptors, Proinflammatory Effects, and Production

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Abstract IL-17A is a proinflammatory cytokine with critical effects on many cell types. It is part of a larger family of cytokines with important roles in protection against pathogens. The IL-17A receptors are IL-17RA and IL-17RC. They are expressed on most cell types. IL-17A, in synergy with other cytokines such as TNF- α , induces the production of cytokines, chemokines, and mediators of tissue destruction in several cell types. The identification of IL-17A and, later on, of Th17 cells has modified the established Th1 and Th2 paradigm, led to the definition of a new CD3⁺ CD4⁺ effector T cell subset, and introduced a new paradigm to explain the origin of several autoimmune events. However, this paradigm shift tended also to identify the effects of IL-17A with those of Th17 cells and vice versa. This view might be insufficient to explain the role of IL-17A in infections and autoimmune models. IL-17A is in fact produced by several other cell types involved in host defense, autoimmunity, and inflammation, and they might also be involved in IL-17A induced pathology.

Keywords IL-17A • IL17F • IL-17RA • IL17RC • Th17 cell

1 IL-17A and Other IL17 Family Members

In 1993, Rouvier et al. [1] described the cloning of a rodent cDNA sequence, termed CTLA8, from an activated T cell hybridoma using a subtractive hybridization approach and reported the sequence of the corresponding protein. This sequence

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had homology to an open reading frame encoded within a T cell-tropic *c*-herpesvirus, Herpesvirus Saimiri, and they suggested that it might be a novel cytokine [1]. Later on, the protein described was found to be a rat protein [2]. The human homologue was reported in 1995 and was mainly produced by activated CD4⁺ T cells, induced the secretion of IL-6 and IL-8 in human foreskin fibroblasts, and enhanced the expression of ICAM-1 [2]. This molecule was named IL-17. The mouse homolog was reported a year later [3, 4] and, in one case, was cloned from mouse NKT cells [3].

Human IL-17A is a glycoprotein containing 155 amino acid residues and shares 63 % amino acid identity with murine IL-17A (147 amino acids). Both human and mouse IL-17A are secreted as disulfide-linked homodimers. Now, the IL-17 family includes seven members (IL-17 or IL-17A, IL-17B, IL-17C, IL-17D, IL-17E or IL-25, and IL-17F plus the viral homologue ORF13 or vIL-17). In the mouse system, IL-17F has 45 % amino acid homology to IL-17A, followed by IL-17C (24 %), IL-17B (21 %), IL-17D (16 %), and IL-17E (16 %) [5]. IL-17A and IL-17F are secreted by overlapping populations of T cells and form both homodimers and heterodimers [6, 7]. IL-17B, IL-17C, and IL-17E differ substantially from IL-17A and IL-17F in the N terminus, having longer extensions. Furthermore, IL-17B is secreted as a noncovalent dimer [8]. Common features of the IL-17 family are a highly conserved carboxyl terminus and five spatially conserved cysteine residues. Four of these cysteines form a cystine knot fold. Transforming growth factor (TGF)- β , bone morphogenic protein, and nerve growth factor superfamilies show structural similarities but have an additional disulfide bond [8].

The genes for IL-17A and IL-17F are located on human chromosome 6p12.2, while the genes of the other IL-17 family members are located on different chromosomes: IL-17B on Chr 5q32, IL-17C on Chr 16q24.3, IL-17D on Chr 13q12.111, and IL-25 on Chr 14q11.2. Polymorphisms in IL-17A (rs2275913) gene have been weakly associated with rheumatoid arthritis [9], while polymorphism in IL-17F (rs763780) gene has been associated with a protective role in asthma [10]. Polymorphism at the 197A (IL-17A) and 7488T (IL-17F) alleles may influence the susceptibility to and pathophysiological features of ulcerative colitis [11]. In the Chinese population, rs2275913 and rs763780 are associated with immune diseases and infectious diseases, including asthma, Vogt–Koyanagi–Harada syndrome, bacterial bronchiolitis, and neuromyelitis optica [12–14].

2 The Receptors

When the mouse and the human IL-17 receptors (IL-17RA) were cloned, IL-17RA was found to be ubiquitously expressed and unique. It consisted of a single transmembrane receptor with an unusually long cytoplasmic tail [15, 16]. Now, five different receptors have been described, IL-17RA, IL-17RB, IL-17RC, IL-17RD, and IL-17RE [17]. The IL-17R family contains conserved structural characteristics such as extracellular fibronectin III-like domains and cytoplasmic similar expression to fibroblast growth factor, IL-17R, and Toll-IL-1R family

(SEFIR) domains. IL-17A and IL-17F signal through a receptor complex composed of IL-17RA and IL-17RC [18]. In humans IL-17RA binds with higher affinity to IL-17A than to IL-17F, whereas IL-17RC associates with both IL-17A and IL-17F [18, 19]. Crystal structure of the IL-17F-IL-17RA interaction reveals the presence of a loop of IL-17RA which interacts with the deep groove at the interface of IL-17F homodimer [18]. IL-17RD has also been associated with the IL-17R complex and may contribute to IL-17A signaling [20]. IL-17RB is the receptor chain for IL-25 and IL-17B [21, 22]. The receptor for IL-17C appears to be a heterodimeric complex formed by IL-17RA (low-affinity interaction) and IL-17RE (high-affinity interaction) [23–25].

In conclusion, IL-17RA is shared among the members of the IL-17 family, while the specificity of the interaction mainly resides in the second subunit of the heterodimeric receptor complex. IL-17RC is common to IL-17A and IL-17F homo- and heterodimers; IL-17RB is specific for IL-25 and IL-17B, IL-17RE for IL-17C, and IL-17RD may be part of the IL-17A receptor complex [23–25]. Since IL-17RA is ubiquitously expressed, also the tissue specificity of the responses resides in the second receptor chain which is differently expressed in various tissues.

3 IL-17A as a Proinflammatory Cytokine

Several reports suggest that IL-17A has a critical role in the protection of the organism against bacteria and fungi due to the ability of IL-17A to recruit neutrophils to the infected organs/tissues [26, 27]. IL-17A and IL-17F can induce the production of proinflammatory cytokines (IL-1, IL-6, TNF- α , G-CSF, and GM-CSF), chemokines (CXCL1, CXCL5, IL-8, CCL2, and CCL7), antimicrobial peptides (defensins, lipocalin2, and S100 proteins), and matrix metalloproteinases (MMP1, MMP3, and MMP13) from different cell types. IL-17A also controls the expression of intercellular cell adhesion molecule 1 (ICAM-1) in keratinocytes as well as iNOS and cyclooxygenase-2 in chondrocytes. IL-17F is a weaker inducer of proinflammatory cytokine but seems to be produced by a wider range of cell types, including innate immune cells and epithelial cells. Not unexpectedly, IL-17A and IL-17F act in concert with other cytokines such as TNF- α , with which they strongly synergize in *in vitro* cellular assays and *in vivo*. It is expected that this characteristic is shared by the other IL-17 family members. Moreover, IL-17A contributes to germinal center formation and class switch recombination by its action on B cells [28, 29].

In summary, IL-17A, acting in synergy with other cytokines, contributes to modulate and control the inflammatory response. Some events, e.g., neutrophil recruitment, are indirectly mediated through the activity of IL-17A on cells which secrete chemotactic factors for neutrophils. Therefore, the mediators, secreted by activated cells, modulate the final outcome of the response to IL-17A.

4 IL-17A-Producing Cells

In 2000, Infante-Duarte et al. [30] recognized that IL-17A cannot be categorized as either a Th1 or a Th2 cytokine, but a major advance in IL-17 research came with the recognition of a novel T cell lineage producing IL-17A and/or IL-17F [31, 32]. This finding modified the established Th1-Th2 paradigm, led to the definition of the CD3+ CD4+ Th17 cell subset, and introduced a new paradigm to explain the origin of several autoimmune events.

In the mouse, IL-17A and/or IL-17F are however produced by several other cell types involved in host defense, autoimmunity, and inflammation, including subpopulations of CD8+ T cells (Tc17), $\gamma\delta$ T cells [33], invariant natural killer (iNKT) cells [34], lymphoid tissue inducers [35], neutrophils [36], and macrophages [37]. In addition, IL-17A is produced by intestinal Paneth cells in an IL-23 independent fashion being one of the causes of a systemic inflammatory response syndrome [38], and IL-17F mRNA is expressed in colonic epithelial cells [39]. In particular, Jensen et al. [33] found that $\gamma\delta$ T cells are uniquely suited for the initial IL-17A response, which often is elicited without a clear antigen exposure. They also found evidence for two distinct functional subsets of $\gamma\delta$ T cells (T $\gamma\delta$ -17s and T $\gamma\delta$ -IFN- γ s).

Yoshiga et al. [40] demonstrated that also iNKT cells synthesize and release IL-17A, accelerate IL-17 production by Th17 cells, and play a role in the development of collagen-induced arthritis (CIA). Moreover, they found that iNKT cells can be induced to secrete IL-17A through two pathways, one involving the direct activation of the TCR by glycolipid antigen and the other the IL-23-IL-23R signaling pathway [40]. It appears therefore that IL-17A is produced by different subsets of T cells, which are involved in both innate and adaptive immunity, and it is well suited to participate to both early and late phases of the immune response.

In addition, in IL-10- or IL-10R-deficient mice, even macrophages stimulated with lipopolysaccharide appear to produce high levels of IL-17A and IL-22 [41]. Addition of exogenous IL-10 abolishes IL-17A production both in Th17 cells and in macrophages [41], again suggesting that IL-17A production is highly regulated.

In humans, IL-17A has been detected in several T cell subsets, such as naive, central memory, and effector memory CD4+ IL-17+ T cells [42], NKT-like cells [43], macrophages [44], astrocytes [45], oligodendroglia [45], mast cells [46], neutrophils [47], and also a human myeloma cell line [48].

The large number of cell types able to secrete IL-17A and IL-17F, the migratory potential of the cells secreting these cytokines, the ubiquitous distribution of the receptor, and the synergism with other proinflammatory cytokines allow IL-17A and IL-17F to exert effects in multiple pathological conditions [26, 49]. In addition, Th17 cells act in close contact with their target cell, and IL-17A might have a short range of action and act in synergy with other cytokines secreted by Th-17 cells [50]. The role of other family members might be more restricted due to a more limited expression and tissue distribution of their receptors.

A pathological role for IL-17A in several autoimmune models of disease, such as experimental autoimmune encephalomyelitis (EAE) and arthritis, has been documented [5], and increased production of IL-17A has been reported in various

human autoimmune and allergic diseases, such as rheumatoid arthritis [51], multiple sclerosis [45], psoriasis [52], and asthma [53]. All these findings suggest a pathogenic role in several human diseases which has been confirmed by the clinical efficacy of IL-17A neutralization in different disease [54].

References

1. Rouvier E, Luciani M-F, Mattei MG, Denizot F, Golstein P (1993) CTLA-8, cloned from an activated T cell, bearing AU-rich messenger RNA instability sequences, and homologous to a herpesvirus saimiri gene. *J Immunol* 150:5445–5456
2. Yao Z, Painter SL, Fanslow WC, Ulrich D, Macduff BM, Spriggs MK, Armitage RJ (1995) Cutting edge: human IL-17: a novel cytokine derived from T cells. *J Immunol* 155:5483–5486
3. Kennedy J, Rossi DL, Zurawski SM, Vega F Jr, Kastelein RA, Wagner JL, Hannum CH, Zlotnik A (1996) Mouse IL-17: a cytokine preferentially expressed by alpha beta TCR + CD4-CD8-T cells. *J Interferon Cytokine Res* 16:611–617
4. Yao Z, Timour M, Painter S, Fanslow W, Spriggs M (1996) Complete nucleotide sequence of the mouse CTLA8 gene. *Gene* 168:223–225
5. Weaver CT, Hatton RD, Mangan PR, Harrington LE (2007) IL-17 family cytokines and the expanding diversity of effector T cell lineages. *Annu Rev Immunol* 25:821–852
6. Liang SC, Long AJ, Bennett F, Whitters MJ, Karim R, Collins M, Goldman SJ, Dunussi-Joannopoulos K, Williams CMM, Wright JF et al (2007) An IL-17F/A heterodimer protein is produced by mouse Th17 cells and induces airway neutrophil recruitment. *J Immunol* 179:7791–7799
7. Wright JF, Guo Y, Quazi A, Luxenberg DP, Bennett F, Ross JF, Qiu Y, Whitters MJ, Tomkinson KN, Dunussi-Joannopoulos K et al (2007) Identification of an interleukin 17F/17A heterodimer in activated human CD4+ T cells. *J Biol Chem* 282:13447–13455
8. Hymowitz SG, Filvaroff EH, Yin JP, Lee J, Cai L, Risser P, Maruoka M, Mao W, Foster J, Kelley RF et al (2001) IL-17s adopt a cystine knot fold: structure and activity of a novel cytokine, IL-17F, and implications for receptor binding. *EMBO J* 20:5332–5341
9. Nordang GB, Viken MK, Hollis-Moffatt JE, Merriman TR, Førre ØT, Helgetveit K, Kvien TK, Lie BA (2009) Association analysis of the interleukin 17A gene in Caucasian rheumatoid arthritis patients from Norway and New Zealand. *Rheumatology (Oxford)* 48:367–370
10. Kawaguchi M, Takahashi D, Hizawa N, Suzuki S, Matsukura S, Kokubu F, Maeda Y, Fukui Y, Konno S, Huang SK et al (2006) IL-17F sequence variant (His161Arg) is associated with protection against asthma and antagonizes wild-type IL-17F activity. *J Allergy Clin Immunol* 117:795–801
11. Arisawa T, Tahara T, Shibata T, Nagasaka M, Nakamura M, Kamiya Y, Fujita H, Nakamura M, Yoshioka D, Arima Y et al (2008) The influence of polymorphisms of interleukin-17A and interleukin-17F genes on the susceptibility to ulcerative colitis. *J Clin Immunol* 28:44–49
12. Chen J, Deng Y, Zhao J, Luo Z, Peng W, Yang J, Ren L, Wang L, Fu Z, Yang X et al (2010) The polymorphism of IL-17G-152A was associated with childhood asthma and bacterial colonization of the hypopharynx in bronchiolitis. *J Clin Immunol* 30:539–545
13. Shu Q, Yang P, Hou S, Li F, Chen Y, Du L, Jiang Z (2010) Interleukin-17 gene polymorphism is associated with Vogt–Koyanagi–Harada syndrome but not with Behçet’s disease in a Chinese Han population. *Hum Immunol* 71:988–991
14. Wang H, Zhong X, Wang K, Qiu W, Li J, Dai Y, Hu X (2012) Interleukin 17 gene polymorphism is associated with anti-aquaporin 4 antibody-positive neuromyelitis optica in the Southern Han Chinese—a case control study. *J Neurol Sci* 314:26–28

15. Yao Z, Fanslow WC, Seldin MF, Rousseau A-M, Painter SL, Comeau MR, Cohen JI, Spriggs MK (1995) Herpesvirus saimiri encodes a new cytokine, IL-17, which binds to a novel cytokine receptor. *Immunity* 3:811–821
16. Yao Z, Spriggs MK, Derry JMJ, Strockbine L, Park LS, VandenBos T, Zappone JD, Painter SL, Armitage RJ (1997) Molecular characterization of the human interleukin-17 receptor. *Cytokine* 9:794–800
17. Shen F, Gaffen SL (2008) Structure–function relationships in the IL-17 receptor: implications for signal transduction and therapy. *Cytokine* 41:92–104
18. Ely LK, Fischer S, Garcia KC (2009) Structural basis of receptor sharing by interleukin 17 cytokines. *Nat Immunol* 10:1245–1251
19. Kuestner RE, Taft DW, Haran A, Brandt CS, Brender T, Lum K, Harder B, Okada S, Ostrander CD, Kreindler JL et al (2007) Identification of the IL-17 receptor related molecule IL-17RC as the receptor for IL-17F. *J Immunol* 179:5462–5473
20. Rong Z, Wang A, Li Z, Ren Y, Cheng L, Li Y, Wang Y, Ren F, Zhang X, Hu J et al (2009) IL-17RD (Sef or IL-17RLM) interacts with IL-17 receptor and mediates IL-17 signaling. *Cell Res* 19:208–215
21. Shi Y, Ullrich SJ, Zhang J, Connolly K, Grzegorzewski KJ, Barber MC, Wang W, Wathen K, Hodge V, Fisher CL et al (2000) A novel cytokine receptor–ligand pair. Identification, molecular characterization, and in vivo immunomodulatory activity. *J Biol Chem* 275:19167–19176
22. Rickel EA, Siegel LA, Yoon BR, Rottman JB, Kugler DG, Swart DA, Anders PM, Tocker JE, Comeau MR, Budelsky AL (2008) Identification of functional roles for both IL-17RB and IL-17RA in mediating IL-25-induced activities. *J Immunol* 181:4299–4310
23. Chang SH, Reynolds JM, Pappu BP, Chen G, Martinez GJ, Dong C (2011) Interleukin-17C promotes Th17 cell responses and autoimmune disease via interleukin-17 receptor E. *Immunity* 35:611–621
24. Ramirez-Carrozzi V, Sambandam A, Luis E, Lin Z, Jeet S, Lesch J, Hackney J, Kim J, Zhou M, Lai J et al (2011) IL-17C regulates the innate immune function of epithelial cells in an autocrine manner. *Nat Immunol* 12:1159–1166
25. Song X, Zhu S, Shi P, Liu Y, Shi Y, Levin SD, Qian Y (2011) IL-17RE is the functional receptor for IL-17C and mediates mucosal immunity to infection with intestinal pathogens. *Nat Immunol* 12:1151–1158
26. Ye P, Rodriguez FH, Kanaly S, Stocking KL, Schurr J, Schwarzenberger P, Oliver P, Huang W, Zhang P, Zhang J et al (2001) Requirement of interleukin 17 receptor signaling for lung CXC chemokine and granulocyte colony-stimulating factor expression, neutrophil recruitment, and host defense. *J Exp Med* 194:519–527
27. Ye P, Garvey PB, Zhang P, Nelson S, Bagby G, Summer WR, Schwarzenberger P, Shellito JE, Kolls JK (2001) Interleukin-17 and lung host defense against *Klebsiella pneumoniae* infection. *Am J Respir Cell Mol Biol* 25:335–340
28. Mitsdoerffer M, Lee Y, Jaeger A, Kim HJ, Korn T, Kolls JK, Cantor H, Bettelli E, Kuchroo VK (2010) Proinflammatory T helper type 17 cells are effective B-cell helpers. *Proc Natl Acad Sci USA* 107:14292–14297
29. Wu HJ, Ivanov II, Darce J, Hattori K, Shima T, Umesaki Y, Littman DR, Benoist C, Mathis D (2010) Gut-residing segmented filamentous bacteria drive autoimmune arthritis via T helper 17 cells. *Immunity* 32:815–827
30. Infante-Duarte C, Horton HF, Byrne MC, Kamradt T (2000) Microbial lipopeptides induce the production of IL-17 in Th cells. *J Immunol* 165:6107–6115
31. Harrington LE, Hatton RD, Mangan PR, Turner H, Murphy TL, Murphy KM, Weaver CT (2005) Interleukin 17-producing CD4+ effector T cells develop via a lineage distinct from the T helper type 1 and 2 lineages. *Nat Immunol* 6:1123–1132
32. Park H, Li Z, Yang XO, Chang SH, Nurieva R, Wang YH, Wang Y, Hood L, Zhu Z, Tian Q et al (2005) A distinct lineage of CD4 T cells regulates tissue inflammation by producing interleukin 17. *Nat Immunol* 6:1133–1141

33. Jensen KD, Su X, Shin S, Li L, Youssef S, Yamasaki S, Steinman L, Saito T, Locksley RM, Davis MM et al (2008) Thymic selection determines gammadelta T cell effector fate: antigen-naive cells make interleukin-17 and antigen-experienced cells make interferon gamma. *Immunity* 29:90–100
34. Michel ML, Keller AC, Paget C, Fujio M, Trottein F, Savage PB, Wong CH, Schneider E, Dy M, Leite-de-Moraes MC (2007) Identification of an IL-17-producing NK1.1(neg) iNKT cell population involved in airway neutrophilia. *J Exp Med* 204:995–1001
35. Cua DJ, Tato CM (2010) Innate IL-17-producing cells: the sentinels of the immune system. *Nat Rev Immunol* 10:479–489
36. Ferretti S, Bonneau O, Dubois GR, Jones CE, Trifileff A (2003) IL-17, produced by lymphocytes and neutrophils, is necessary for lipopolysaccharide-induced airway neutrophilia: IL-15 as a possible trigger. *J Immunol* 170:2106–2112
37. Song C, Luo L, Lei Z, Li B, Liang Z, Liu G, Li D, Zhang G, Huang B, Feng ZH (2008) IL-17-producing alveolar macrophages mediate allergic lung inflammation related to asthma. *J Immunol* 181:6117–6124
38. Takahashi N, Vanlaere I, de Rycke R, Cauwels A, Joosten LA, Lubberts E, van den Berg WB, Libert C (2008) IL-17 produced by Paneth cells drives TNF-induced shock. *J Exp Med* 205:1755–1761
39. Ishigame H, Kakuta S, Nagai T, Kadoki M, Nambu A, Komiyama Y, Fujikado N, Tanahashi Y, Akitsu A, Kotaki H et al (2009) Differential roles of interleukin-17A and -17F in host defense against mucoc epithelial bacterial infection and allergic responses. *Immunity* 30:108–119
40. Yoshiga Y, Goto D, Segawa S, Ohnishi Y, Matsumoto I, Ito S, Tsutsumi A, Taniguchi M, Sumida T (2008) Invariant NKT cells produce IL-17 through IL-23-dependent and -independent pathways with potential modulation of Th17 response in collagen-induced arthritis. *Int J Mol Med* 22:369–374
41. Gu Y, Yang J, Ouyang X, Liu W, Li H, Yang J, Bromberg J, Chen SH, Mayer L, Unkeless JC, Xiong H (2008) Interleukin 10 suppresses Th17 cytokines secreted by macrophages and T cells. *Eur J Immunol* 38:1807–1813
42. Jandus C, Bioley G, Rivals JP, Dudler J, Speiser D, Romero P (2008) Increased numbers of circulating polyfunctional Th17 memory cells in patients with seronegative spondylarthritides. *Arthritis Rheum* 58:2307–2317
43. Cosmi L, De Palma R, Santarlasci V, Maggi L, Capone M, Frosali F, Rodolico G, Querci V, Abbate G, Angeli R et al (2008) Human interleukin 17-producing cells originate from a CD161+ CD4+ T cell precursor. *J Exp Med* 205:1903–1916
44. Fujino S, Andoh A, Bamba S, Ogawa A, Hata K, Araki Y, Bamba T, Fujiyama Y (2003) Increased expression of interleukin 17 in inflammatory bowel disease. *Gut* 52:65–70
45. Tzartos JS, Friese MA, Craner M, Palace J, Newcombe J, Esiri MM, Fugger L (2007) Interleukin-17 production in CNS-infiltrating T-cells and glial cells is associated with active disease in multiple sclerosis. *Am J Pathol* 172:146–155
46. Hueber AJ, Asquith DL, Miller AM, Reilly J, Kerr S, Leipe J, Melendez AJ, McInnes IB (2010) Mast cells express IL-17A in rheumatoid arthritis synovium. *J Immunol* 184:3336–3340
47. Lin AM, Rubin CJ, Khandpur R, Wang JY, Riblett M, Yalavarthi S, Villanueva EC, Shah P, Kaplan MJ, Bruce AT (2011) Mast cells and neutrophils release IL-17 through extracellular trap formation in psoriasis. *J Immunol* 187:490–500
48. Zhou L, Peng S, Duan J, Zhou J, Wang L, Wang J (1998) A human B cell line AF10 expressing HIL-17. *Biochem Mol Biol Int* 45:1113–1119
49. Iwakura Y, Ishigame H, Saijo S, Nakae S (2011) Functional specialization of interleukin-17 family members. *Immunity* 34:149–162
50. Pöllinger B, Junt T, Metzler B, Walker UA, Tyndall A, Allard C, Bay S, Keller R, Raulf F, Di Padova F et al (2011) Th17 cells, not IL-17+ $\gamma\delta$ T cells, drive arthritic bone destruction in mice and humans. *J Immunol* 186:2602–2612

51. Ziolkowska M, Koc A, Luszczkiewicz G, Ksiezopolska-Pietrzak K, Klimczak E, Chwalinska-Sadowska H, Maslinski W (2000) High levels of IL-17 in rheumatoid arthritis patients: IL-15 triggers in vitro IL-17 production via cyclosporin A-sensitive mechanism. *J Immunol* 164:2832–2838
52. Pène J, Chevalier S, Preisser L, Vénéreau E, Guilleux MH, Ghannam S, Molès JP, Danger Y, Ravon E, Lesaux S et al (2008) Chronically inflamed human tissues are infiltrated by highly differentiated Th17 lymphocytes. *J Immunol* 180:7423–7430
53. Molet S, Hamid Q, Davoine F, Nutku E, Taha R, Page N, Olivenstein R, Elias J, Chakir J (2001) IL-17 is increased in asthmatic airways and induces human bronchial fibroblasts to produce cytokines. *J Allergy Clin Immunol* 108:430–438
54. Hueber W, Patel DD, Dryja T, Wright AM, Koroleva I, Bruin G, Antoni C, Draelos Z, Gold MH, Durez P et al (2010) Effects of AIN457, a fully human antibody to interleukin-17A, on psoriasis, rheumatoid arthritis, and uveitis. *Sci Transl Med* 2:52ra72