

# Other 4-Methanolquinolines, Amyl Alcohols and Phentathrenes: Mefloquine, Lumefantrine and Halofantrine

Francois Nosten, Penelope A. Phillips-Howard, and Feiko O. ter Kuile

**Abstract** This chapter describes mefloquine, pyronaridine, halofantrine, piperavaquine and lumefantrine under the broader title of the 4-methanolquinolines, amyl alcohols and phentathrenes. We provide a brief resume of each drug, in terms of their chemical properties, formulae, pharmacokinetics, clinical indications for use, and their efficacy and safety. Recognizing the limited number of antimalarials available, and in the developmental pipeline, attention is focussed on describing the history of each drug and how their indications have evolved as data on safety in human populations accumulates over time, and how patterns of use have changed with growing multiple drug resistance. Their combined use with the artemisinin derivatives is briefly described and readers are recommended to consult other chapters in this book which more fully describe such combinations.

---

F. Nosten (✉)

Nuffield Department of Clinical Medicine, Centre for Tropical Medicine, Churchill Hospital, University of Oxford, Oxford OX3 7LJ, UK

Shoklo Malaria Research Unit, PO Box 46, 68/30 Baan Tung Road, Mae Sot, Tak Province 63110, Thailand

Mahidol University, Bangkok, Thailand

e-mail: [SMRU@tropmedres.ac](mailto:SMRU@tropmedres.ac)

P.A. Phillips-Howard

Centre for Public Health, Liverpool John Moores University, Liverpool, Henry Cotton Campus, 15–21 Webster Street, L3 2ET Liverpool, UK

e-mail: [p.phillips-howard@ljmu.ac.uk](mailto:p.phillips-howard@ljmu.ac.uk)

F.O. ter Kuile

Malaria Epidemiology Unit Child and Reproductive Health Group, Liverpool School of Tropical Medicine, Pembroke Place, Liverpool L3 5QA, UK

e-mail: [terkuile@liverpool.ac.uk](mailto:terkuile@liverpool.ac.uk)

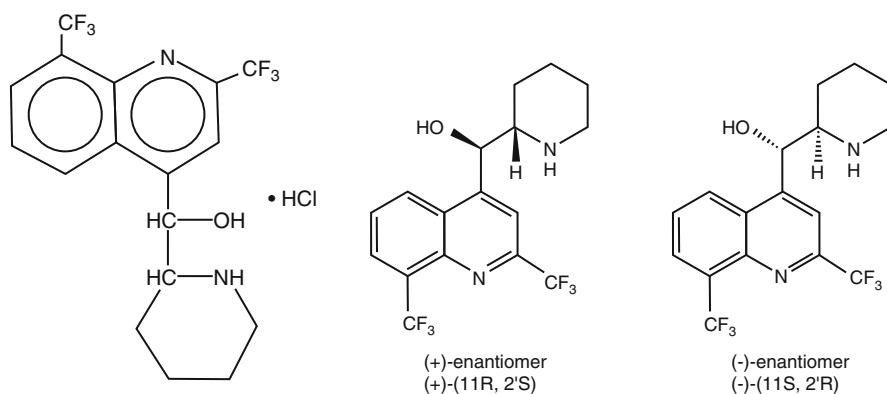
# 1 Mefloquine

## 1.1 Structure and Action

Mefloquine hydrochloride is a 4-quinolinemethol derivative synthesised as a structural analogue (2-aryl substituted chemical) of quinine. Its full chemical name is (R\*, S\*)-(±)-α-2-piperidinyl-2,8-bis (trifluoromethyl)-4-quinolinemethanol hydrochloride. Its formula is C<sub>17</sub>H<sub>16</sub>F<sub>6</sub>N<sub>2</sub>O (Fig. 1). Mefloquine was discovered by the Experimental Therapeutics Division of the Walter Reed Army Institute of Research (WRAIR) in the 1970s for chemoprophylaxis (250 mg weekly) and therapy (15–25 mg/kg) and was approved by the U.S. Food and Drug Administration in 1989.

Mefloquine is a blood schizonticide, active against the erythrocytic stages of *Plasmodium falciparum* and *P. vivax*, with no effect on the exoerythrocytic (hepatic) stages of the parasite, and with limited information of its effect on *P. ovale*, *P. malariae* and *P. knowlesi*. Studies indicate mefloquine interferes with the transport of haemoglobin and other substances from the host erythrocyte to the food vacuole of the malaria parasite, causing swelling and cytotoxicity [1]. Mefloquine strongly inhibits endocytosis in the D10 strain of *P. falciparum* using several lines of evidence: a reduction in haemoglobin levels in the parasite as assessed by Western blotting, decreased levels of accumulation of biotinylated dextran by the parasite in preloaded erythrocytes, significantly lower concentrations of fluorescent dextran in the food vacuole, and a reduced percentage of parasites with multiple transport vesicles [2].

Mefloquine is a chiral molecule; it has two asymmetric carbon atoms and exists in two racemic forms (erythro and threo), each of which is composed of a pair of optical isomers, i.e. (±)-erythro-enantiomers and the (±)-threo-epimers. Clinically, the racemic mixture of the erythro-enantiomers is used [3]. Unlike other antimalarial drugs such as chloroquine, halofantrine, and lumefantrine, there is stereoselectivity



**Fig. 1** Structure of mefloquine hydrochloride and its two enantiomers

in its antimalarial activity, with the (+)-isomer ~1.7 times more potent than the (–)-isomer in vitro [4, 5].

## 1.2 Pharmacokinetics

Mefloquine is moderately well absorbed orally and extensively distributed and is 98% bound to plasma proteins. Splitting 25 mg/kg mefloquine into 2 or 3 doses given 16–24 h apart reduces vomiting, improves oral bioavailability and the therapeutic response in the treatment of acute falciparum malaria [6]. Food increases its bioavailability by up to 40%. The parent compound is metabolized by the cytochrome P450 enzyme CYP3A4 to two major metabolites: carboxy- and hydroxyl-mefloquine, which are inactive against *P. falciparum*. Mefloquine is eliminated slowly and has a terminal elimination half-life of ~3 weeks in volunteers and 2 weeks in patients. Total clearance, which is essentially hepatic, is 30 ml/min in volunteers. A steady-state plasma concentration of 1,000–2,000 µg/l is reached after 7–10 weeks following weekly 250 mg prophylaxis and it is therefore recommended to start medication at least 2 weeks before travel. There are stereoselective differences in their pharmacokinetics and the ability of the mefloquine enantiomers to cause certain adverse effects [5]. In humans, the plasma concentration of the (–) enantiomer is approximately threefold higher than the (+) enantiomer, reflecting the stereoselectivity in the clearance and volume distribution [5, 7–11]. Co-administration with artemisinin does not appear to influence mefloquine enantiomer pharmacokinetics [12].

## 1.3 Clinical Use

One tablet of 250 mg mefloquine hydrochloride per week (adult dose; equivalent to 228 mg of the free base) has been used for prophylaxis in travellers, including for young children and pregnant women. The limited data available on the use of mefloquine in human pregnancy are reassuring and do not indicate an increased teratogenic risk [13]. A retrospective study of 208 women on the Thai–Burmese border treated with mefloquine found an increased risk of stillbirths [14], however, this finding was not confirmed in a large prospective trial of mefloquine prophylaxis in Malawian pregnant women [15] and remains unexplained. As a treatment, it is now mainly used in combination with artesunate, a water-soluble artemisinin derivative. It is available as a loose combination (as part of a blister pack containing both drugs), and as a new fixed-dose combination developed with support from the Drugs for Neglected Diseases Initiative (DNDi) and produced by Farmanguinhos/Fiocruz, Brazil and Cipla, India. The treatment dose is 25 mg/kg of mefloquine and 12 mg/kg of artesunate given as 8.3 and 4 mg/kg/day over 3 days, respectively. Tablets of the new fixed-dose combination come in adult and child “strengths”,

with several co-blistered formulations of the loose combinations made by different manufacturers. Mefloquine is also being explored for a new indication as intermittent preventive therapy (IPT) against malaria. Two trials evaluating the role of mefloquine as IPT in infants (IPTi) [16] and in pregnant women (IPTp) [17] found mefloquine to be very effective, but the low tolerability limited its acceptance for use as IPTi. Further IPTp studies are ongoing in five countries in Africa with the lower 15 mg/kg dose in pregnancy.

## 1.4 Resistance

Most experience with mefloquine as monotherapy and later in combination with the artemisinin derivatives has been gained from areas of multiple drug resistance in Southeast Asia, such as on the Thai–Burmese (Myanmar) border. Thailand was the first country to use mefloquine for first-line treatment of acute malaria. From 1985 to 1990, it was recommended in combination with sulfadoxine and pyrimethamine, as “MSP” in a fixed-dose combination, at a single dose of ~15/30/1.5 mg/kg, providing a 98% cure rate after its introduction in 1985, however, this dropped to <50% in children by 1990 [18]. Because of high levels of existing parasite resistance to SP, and lack of additional therapeutic efficacy over mefloquine alone, the SP component was dropped [18, 19] and replaced by mefloquine monotherapy; initially at a single dose of 15 mg/kg. High levels of treatment failure with this dose [18, 20] prompted 25 mg/kg dosing [21–23], split (750 and 500 mg, 16–24 h apart) to reduce vomiting [24]. Within 8 years, mefloquine monotherapy failure rates on the Thai–Burmese border reached 60% and, following extensive testing, the combination of mefloquine 25 mg/kg with artesunate 12 mg/kg given over 3 days (MAS<sub>3</sub>) became the new standard therapy [25]. This therapy, combined with early diagnosis and use of insecticide treated nets, reduced *P. falciparum* malaria incidence, and halted, and later reversed the progression of mefloquine resistance [26–28]. The combination offered a potential public health solution for multiple drug-resistant *P. falciparum*, and allowed time for the development of other new drugs [1].

In vivo resistance to mefloquine, mediated mainly by an increase in gene copy number and expression of the *P. falciparum* multi-drug resistance (MDR) gene-1 (*pfmdr1*), a gene encoding a parasite-transport protein [29, 30], has been confirmed. This has been reported on the borders of Thailand with Burma (Myanmar) and Cambodia, in the western provinces of Cambodia, the eastern states of Burma (Myanmar) and its border with China, along the Laos and Burma borders, the adjacent Thai–Cambodian border and in southern Vietnam. It is likely the initial deployment of low dose of mefloquine may have encouraged resistance. Theoretical evidence suggests that initial use of higher doses, preferably in combination with an artemisinin derivative, is less likely to lead to resistance [31].

## 1.5 Tolerability

Preclinical studies demonstrate mefloquine to be safe and effective [32], and extensive clinical experience to date supports this. Nevertheless, widespread deployment of mefloquine for treatment and prophylaxis has been hampered by concerns about its tolerability. Side effects following treatment are common; they are usually mild and restricted to dizziness/vertigo and gastro-intestinal disturbances [24]. Vomiting after mefloquine is a problem in young children, but can be mitigated by splitting the dose over 2 or 3 days, and by fever reduction [24, 33]. In older children and adults, mild neuro-psychiatric events (headache, dizziness, insomnia and vivid dreams) are reported in ~25% of patients treated with 25 mg/kg mefloquine.

Mefloquine is also associated with a self-limiting acute neuropsychiatric syndrome manifest by encephalopathy, convulsions or psychosis [34–36], apparent in international travellers taking 250 mg mefloquine each week for prophylaxis [35, 37]. Mefloquine is thus contraindicated for prophylaxis in patients with active depression, a recent history of depression, generalised anxiety disorder, psychosis, or schizophrenia or other major psychiatric disorders, or with a history of convulsions [38]. These contraindications are prevalent in 9–10% of the military [39] and civilian [40] populations presenting for malaria chemoprophylaxis [41], but are not documented in endemic populations. While the mechanism is not yet fully understood, neuropsychiatric events have been demonstrated to be associated with dose in humans [42, 43]. The rates in travellers are estimated to be 1:10,000 persons, equally frequent with chloroquine prophylaxis, but higher than in similar populations that used other forms of prophylaxis [35, 44]. The incidence following treatment doses is 1:1,000 in Asian patients [45], 1:200 in Caucasian or African patients with uncomplicated malaria and 1:20 in patients recovering from severe malaria [46]. Previous history of psychiatric illness or epilepsy is a risk factor. Females and individuals of low body mass index are also at apparent greater risk. Neuropsychiatric reactions are more common if mefloquine was used in the previous 2 months, and thus should not be used to treat recrudescing infections within 2 months of treatment.

Total (racemic) concentrations of mefloquine are ~30-fold higher in brain than in plasma [47]. In man, approximately threefold higher concentrations of the (–)-enantiomer is observed in plasma, and 1.5-fold higher in brain, but post-mortem studies demonstrated stereoselective brain penetration, greater for the (+)-enantiomer, with (–) and (+) concentrations at ~23- versus 56-fold higher in the brain's white matter compared with plasma (the reverse is found in rat models where the penetration of the (–)-enantiomer is greater than that of its antipode) [7, 47–49]. There is a growing body of evidence on the mechanisms of possible neurotoxicity (see reviews [50] and [41]). The high level of accumulation of mefloquine in brain tissue may be associated with direct neurotoxic damage and cell death, with binding to neuroreceptors and cholinesterases, inhibition of sarco (endo)plasmic reticulum  $\text{Ca}^{2+}$  ATPase (SERCA) activity and interference with

cellular  $\text{Ca}^{2+}$  homeostasis and reductions in central nervous system efflux in individuals possessing certain (human) MDR1 polymorphisms [50].

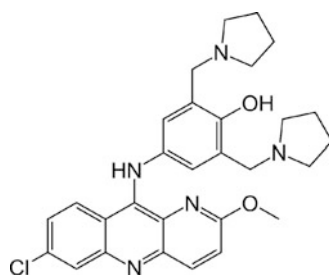
## 1.6 The Future

Using mefloquine as a scaffold, WRAIR has constructed a library of 200 potential next generation quinoline methanol compounds to identify leads that possess biological properties consistent with the target product profile for malaria chemoprophylaxis but less susceptible to passage across the blood–brain barrier (to reduce adverse neurological effects) [51, 52]. During a programme to examine the biochemical basis of side effects, investigators discovered that the (–)-(R,S)-enantiomer is a potent adenosine A2A receptor antagonist, resulting in a programme to develop novel adenosine A2A antagonists for the management of Parkinson's disease [53, 54]. Mefloquine is effective against JC virus and is reported to have successfully treated progressive multifocal leuko-encephalopathy (a progressive, usually fatal, demyelinating disease caused by the JC virus) [55, 56]. Mefloquine is undergoing in vitro and in vivo studies to evaluate its effectiveness for the treatment of helminth infections [57], including those caused by *Schistosoma* [58], *Clonorchis* and *Paragonimus* [59].

## 2 Pyronaridine

### 2.1 Structure and Action

Pyronaridine is a Mannich base with a pyronaridine nucleus synthesised from mepacrine (9 amino acridine). Its formula is  $\text{C}_{29}\text{H}_{32}\text{ClN}_5\text{O}_2$  (Fig. 2). It was synthesised in 1970 in China and is available as a free base and as a tetraphosphate, the salt used in current formulations. It was used in China as a monotherapy in the 1980s and 1990s but has now been developed as a combination therapy with



**Fig. 2** Structure of pyronaridine

artesunate by Shin Poong Pharmaceutical (Korea) and Medicines for Malaria Venture (MMV, Switzerland).

Pyronaridine is active against asexual forms of *Plasmodium* by forming complexes with ferriprotoporphyrin IX. Growth studies of *P. falciparum* K1 in culture demonstrate the ability of pyronaridine to inhibit in vitro  $\beta$ -haematin formation, to form a complex with a stoichiometry of 1:2, to enhance haematin-induced red blood cell lysis, and to inhibit glutathione-dependent degradation of haematin [60]. However, observations that pyronaridine exerted this mechanism of action in situ, based on showing antagonism of pyronaridine in combination with antimalarials (chloroquine, mefloquine, and quinine) that inhibit  $\beta$ -haematin formation, were equivocal. Interestingly pyronaridine is also active against young gametocytes (stage II and III) in vitro [61], although recent clinical studies did not detect a difference in gametocyte carriage following treatment with the fixed combination artesunate–pyronaridine when compared with artemether–lumefantrine [62]. The compound is also a poor substrate and an inhibitor of the Permeability glycoprotein (P-gp) ATP-dependant transporter, a product of the human multidrug resistance-1 (MDR1; ABCB1) gene that influences the passage of many drugs across epithelial barriers. The P-gp-mediated efflux could attenuate oral absorption of the drug when the luminal concentration falls; however, this is likely to play a minimal role in the initial absorption of the drug.

## 2.2 Pharmacokinetics

The pharmacokinetic properties of pyronaridine are not well characterised. The drug is readily absorbed from the small intestine following oral administration and is widely distributed in most tissues [63]. The peak value of the drug in the blood is reached at around 8 h post-administration, and it shows a poor permeability across the blood–brain barrier. Pyronaridine concentrates preferentially in infected red blood cells and its distribution and elimination are influenced by age and disease status. It is eliminated slowly, with the half-life currently estimated at 18 days in patients with malaria [64]. Improvements to the assay that measures blood concentrations are likely to reveal a longer half-life.

## 2.3 Resistance

Resistance is known to have developed in *P. falciparum* when the drug was used in China but the molecular mechanism is unknown. High recurrence rates were noted in early clinical trials in Thailand [65] and cross-resistance with chloroquine was suggested in vitro [66]. Pyronaridine is not used as a single agent anymore but only in combination with artesunate to prevent the emergence of de novo resistance.

### 3 Halofantrine

#### 3.1 Structure, Action and Resistance

Halofantrine belongs to the phenanthrene methanol group and was developed by WRAIR in the 1960s and then by Smith Klein, now GlaxoSmithKlein. It is a small chiral molecule and the chemical formula is  $C_{26}H_{30}Cl_2F_3NO$  (Fig. 3). The oral formulation is in tablets containing 250 mg of hydrochloride salt. Like other quinoline derivatives, the mode of action of halofantrine appears to be in the inhibition of the formation of  $\beta$ -haematin crystals but the precise mechanism of action is unclear. Recently it was shown that halofantrine forms complexes with ferriprotoporphyrin IX and that the inhibition of the haemozoin formation occurs principally at the lipid–aqueous interface, an environment more compatible with the crystal structure of halofantrine–ferriprotoporphyrin IX [67]. Halofantrine is poorly and erratically absorbed due to its low solubility. Absorption is enhanced by fat co-administration. The principal active metabolite is *N*-desbutyl-halofantrine. The terminal half-life in patients with malaria is  $\sim 4.7$  days, making it a relatively rapidly eliminated antimalarial when compared with other drugs of the same group. As halofantrine was withdrawn from use due to the discovery of halofantrine cardiotoxicity (see below), there are few studies on the development of resistance to this drug. Molecular studies have demonstrated that mutations in *pfmdr1* result in altered halofantrine transport, suggesting a role for this efflux transport mechanism in resistance to this drug [68]. Cross-resistance with mefloquine was shown in clinical studies in South East Asia [69] and it is likely that the drug shares common resistance mechanisms with mefloquine and lumefantrine due to their chemical similarities.

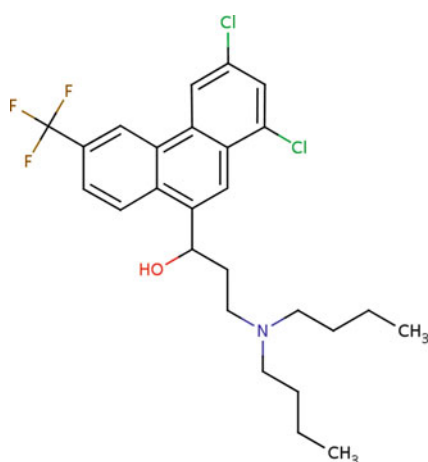


Fig. 3 Structure of halofantrine



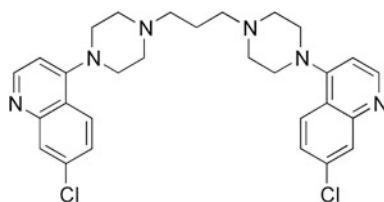
## 3.2 Tolerability

Initially, halofantrine looked like a promising drug for the treatment of uncomplicated falciparum infections caused by chloroquine-resistant parasites. It rapidly cleared parasites and was well tolerated. The first report of the cardiotoxicity of halofantrine in 1993 [70], came as a surprise since the drug had been developed in full compliance with GCP standards. Halofantrine and its principal quinidine-like metabolite have a Class III effect on cardiac repolarization [71]. It causes a dose-dependent blockade of the  $I_{kr}$  channel (through hERG) by binding to the open or inactivated state. This translates on an ECG to a marked prolongation of the QT interval and is more marked when halofantrine is given after mefloquine, probably as a result of the inhibition of the slow delayed rectifier potassium channel  $I_{ks}$  [72]. This QT prolongation, seen at therapeutic doses, increases the risk of the potentially fatal Torsade de Pointes and since the first report, several sudden deaths have been related to the drug. Because of this, halofantrine has been withdrawn in many countries and from international guidelines on the treatment of malaria.

## 4 Piperaquine

### 4.1 Structure and Action

Piperaquine is a bis-amino 4 quinoline synthesised more than 50 years ago by Rhone-Poulenc (France). It was abandoned and rediscovered in China by the Shanghai Research Institute of Pharmaceutical Industry. The drug was used on a large scale (140,000,000 doses) for prophylaxis and treatment of chloroquine resistant *P. falciparum* between 1978 and 1994, but resistance developed in the 1990s. Piperaquine is a lipophilic compound and its chemical formula is  $C_{29}H_{32}Cl_2N_6$  (Fig. 4). The full mechanism of action is unknown but piperaquine concentrates in the parasite food vacuole and inhibits the dimerisation of haematin by binding. Recent work by Warhurst and colleagues has shed more light on the possible mode of action [73]. The high activity of piperaquine against chloroquine-resistant falciparum could be explained by its high lipid accumulation ratio (LAR) leading to an increase in  $\beta$ -haematin inhibition in vacuolar lipids where the crystals of haemozoin are produced. The drug may also act by blocking efflux from the food



**Fig. 4** Structure of piperaquine

vacuole by hydrophobic interaction with the parasite chloroquine-resistance transporter, *pfcr1* [73].

## 4.2 Pharmacokinetics

Piperaquine is lipophilic and exhibits considerable inter-individual variability in pharmacokinetics. It accumulates preferentially in infected red blood cells and this affects the plasma/blood concentration ratio. Like chloroquine, it has a large apparent volume of distribution and a slow elimination. The terminal elimination half-life is probably longer than previously thought and could exceed 30 days [74]. This is valuable for ensuring a prolonged post-treatment prophylactic effect and when the drug is used for IPT. In children, studies have shown that, dose adjustment may be needed [75]. Likewise, pregnant women may have lower piperaquine exposure than non-pregnant women, and the dosage in pregnancy may also need to be adjusted. Piperaquine has two major metabolites: a carboxylic acid and a mono-*N*-oxidated piperaquine product.

## 4.3 Resistance

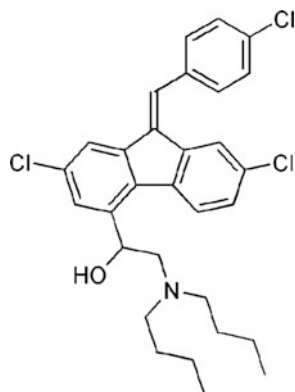
Resistance to piperaquine is known to have developed *in vivo* in China when it was used as monotherapy but there is no indication that it has spread elsewhere. There is no specific molecular marker of resistance to piperaquine and the role of the *P. falciparum* transport proteins *pfmdr1* and *pfcr1* remains unclear [76, 77]. In clinical use, piperaquine is now used in a fixed combination with dihydroartemisinin (DHA) developed by Holleypharm (China) and Sigma-Tau (Italy) in partnership with MMV (Switzerland). DHA-piperaquine is one of the most promising artemisinin-based combination therapies (ACTs) in the antimalarial armamentarium.

# 5 Lumefantrine

## 5.1 Structure and Action

Lumefantrine is a racemic 2,4,7,9-substituted fluorine derivative belonging to the arylamino-alcohol group of antimalarials with a molecular structure reminiscent of halofantrine. It was originally synthesised by the Academy of Military Sciences in Beijing (PRC) in the 1980s. Its chemical formula is  $C_{30}H_{32}Cl_3NO$  (Fig. 5). It is insoluble in water and the two enantiomers have equal antimalarial activities.

**Fig. 5** Structure of lumefantrine



Lumefantrine is active against *P. falciparum* and *P. vivax* asexual stages but not against pre-erythrocytic liver stages, including hypozoites, or against gametocytes. The mode of action of lumefantrine is not known precisely but by similarity of structure with the antimalarials of the same group, it is assumed that lumefantrine kills parasites by inhibiting the polymerisation of heme.

## 5.2 Pharmacokinetics

The pharmacokinetics of lumefantrine have been extensively described in various populations (European, Asian and African) in adults, children and also in pregnant women. The drug is slowly absorbed (time to peak concentrations is approximately 6 h) and metabolised to desbutyl-lumefantrine via CYP3A4 but largely eliminated as parent compound via the liver in faeces and urine. The absorption is dose limited, so the total daily dose must be given on two separate occasions in order to be absorbed, the first serious impediment to observance [78]. The terminal elimination half-life is approximately 4.5 days, reminiscent of halofantrine and much shorter than mefloquine or piperazine. Lumefantrine is highly lipophilic and has a low and variable bioavailability. This is a major contributor to the observed inter-individual variability in its kinetics. The relative fraction of the dose absorbed is also highly variable between patients and between doses. This is probably explained by the combined effects of illness on intestinal mobility, increased food intake with recovery and decreasing parasitaemia. Co-administration of food (or some fat) has a marked effect on the absorption of lumefantrine and this is the second major impediment to observance. Recent studies have shown that as little as 1.2 g of fat was needed for optimum lumefantrine absorption [71]. Unfortunately, this was not taken into account when the paediatric formulation of artemether–lumefantrine (Coartem) was developed. Initially, a 4-dose regimen of Coartem was recommended based on the initial Chinese trials. However this resulted in low cure rates in Thailand [79] but it helped in defining the lumefantrine exposure–cure

rate relationship. The most determinant factors of cure were found to be the initial parasite load and the Area Under the plasma concentration Curve (AUC) of lumefantrine. A useful surrogate of the AUC is the day 7 lumefantrine concentration. A plasma lumefantrine concentration of 280 ng/ml was found to be a useful discriminating cut-off to determine subsequent risk of recrudescence [80] and, in the absence of resistance, a day 7 concentration of 500 ng/ml would be expected to cure >90% of patients [81]. After too many years of delay, the 6-dose regimen became universally recommended. For pregnancy, however, this standard regimen is associated with lower plasma concentrations because of the increased volume of distribution and faster elimination, which will lead to treatment failures [82]. Modelling suggests that longer courses are needed to achieve lumefantrine concentrations comparable to that in non-pregnant patients [75].

### 5.3 Resistance

Resistance to lumefantrine can be readily obtained *in vitro* and in animal models. *In vitro*, single-nucleotide polymorphisms in the *pfmdr1* gene have been associated with increased IC<sub>50</sub> values for lumefantrine [83]. An increase in *pfmdr1* copy number also resulted in decreased *in vitro* susceptibility and increased risk of failure in patients receiving the 4-dose regimen [80]. Interestingly, resistance to lumefantrine in *P. falciparum* could be associated with the loss of chloroquine resistance (i.e. the loss of the *pfcr1* K76T mutation) [84]. Lumefantrine is not used as a single drug but only in combination with artemether (see Coartem). Its future depends on controlling the emergence of resistance to the artemisinin derivatives and/or to the emergence of resistance to lumefantrine itself or through cross resistance with mefloquine.

## References

1. Olliaro P (2001) Mode of action and mechanisms of resistance for antimalarial drugs. *Pharmacol Ther* 89:207–219
2. Hoppe HC, van Schalkwyk DA, Wiehart UI, Meredith SA, Egan J, Weber BW (2004) Antimalarial quinolines and artemisinin inhibit endocytosis in *Plasmodium falciparum*. *Antimicrob Agents Chemother* 48:2370–2378
3. Sweeney TR (1981) The present status of malaria chemotherapy: mefloquine, a novel antimalarial. *Med Res Rev* 1:281–301
4. Karle JM, Olmeda R, Gerena L, Milhous WK (1993) *Plasmodium falciparum*: role of absolute stereochemistry in the antimalarial activity of synthetic amino alcohol antimalarial agents. *Exp Parasitol* 76:345–351
5. Brocks DR, Mehvar R (2003) Stereoselectivity in the pharmacodynamics and pharmacokinetics of the chiral antimalarial drugs. *Clin Pharmacokinet* 42:1359–1382

6. Simpson JA, Price R, ter Kuile F, Teja-Isavatharm P, Nosten F, Chongsuphajaisiddhi T, Looareesuwan S, Aarons L, White NJ (1999) Population pharmacokinetics of mefloquine in patients with acute falciparum malaria. *Clin Pharmacol Ther* 66:472–484
7. Gimenez F, Pennie RA, Koren G, Crevoisier C, Wainer IW, Farinotti R (1994) Stereoselective pharmacokinetics of mefloquine in healthy Caucasians after multiple doses. *J Pharm Sci* 83:824–827
8. Martin C, Gimenez F, Bangchang KN, Karbwang J, Wainer IW, Farinotti R (1994) Whole blood concentrations of mefloquine enantiomers in healthy Thai volunteers. *Eur J Clin Pharmacol* 47:85–87
9. Hellgren U, Jastrebova J, Jerling M, Krysen B, Bergqvist Y (1996) Comparison between concentrations of racemic mefloquine, its separate enantiomers and the carboxylic acid metabolite in whole blood serum and plasma. *Eur J Clin Pharmacol* 51:171–173
10. Bourahla A, Martin C, Gimenez F, Singhasivanon V, Attanath P, Sabchearon A, Chongsuphajaisiddhi T, Farinotti R (1996) Stereoselective pharmacokinetics of mefloquine in young children. *Eur J Clin Pharmacol* 50:241–244
11. Kerb R, Fux R, Morike K, Kremsner PG, Gil JP, Gleiter CH, Schwab M (2009) Pharmacogenetics of antimalarial drugs: effect on metabolism and transport. *Lancet Infect Dis* 9:760–774
12. Svensson US, Alin H, Karlsson MO, Bergqvist Y, Ashton M (2002) Population pharmacokinetic and pharmacodynamic modelling of artemisinin and mefloquine enantiomers in patients with falciparum malaria. *Eur J Clin Pharmacol* 58:339–351
13. Phillips-Howard PA, Steffen R, Kerr L, Vanhauwere B, Schildknecht J, Fuchs E, Edwards R (1998) Safety of mefloquine and other antimalarial agents in the first trimester of pregnancy. *J Travel Med* 5:121–126
14. Nosten F, Vincent M, Simpson J, Yei P, Thwai KL, de Vries A, Chongsuphajaisiddhi T, White NJ (1999) The effects of mefloquine treatment in pregnancy. *Clin Infect Dis* 28:808–815
15. Steketee RW, Wirima JJ, Slutsker L, Roberts JM, Khoromana CO, Heymann DL, Breman JG (1996) Malaria parasite infection during pregnancy and at delivery in mother, placenta, and newborn: efficacy of chloroquine and mefloquine in rural Malawi. *Am J Trop Med Hyg* 55(1 Suppl):24–32
16. Gosling RD, Gesase S, Mosha JF, Carneiro I, Hashim R, Lemnge M, Mosha FW, Greenwood B, Chandramohan D (2009) Protective efficacy and safety of three antimalarial regimens for intermittent preventive treatment for malaria in infants: a randomised, double-blind, placebo-controlled trial. *Lancet* 374:1521–1532
17. Briand V, Bottero J, Noel H, Masse V, Cordel H, Guerra J, Kossou H, Fayomi B, Ayemonna P, Fievet N et al (2009) Intermittent treatment for the prevention of malaria during pregnancy in Benin: a randomized, open-label equivalence trial comparing sulfadoxine-pyrimethamine with mefloquine. *J Infect Dis* 200:991–1001
18. Nosten F, ter Kuile F, Chongsuphajaisiddhi T, Luxemburger C, Webster HK, Edstein M, Phaipun L, Thew KL, White NJ (1991) Mefloquine-resistant falciparum malaria on the Thai-Burmese border. *Lancet* 337:1140–1143
19. Nosten F, Price RN (1995) New antimalarials. A risk-benefit analysis. *Drug Saf* 12:264–273
20. White NJ (1992) Antimalarial drug resistance: the pace quickens. *J Antimicrob Chemother* 30:571–585
21. ter Kuile FO, Nosten F, Thieren M, Luxemburger C, Edstein MD, Chongsuphajaisiddhi T, Phaipun L, Webster HK, White NJ (1992) High-dose mefloquine in the treatment of multidrug-resistant falciparum malaria. *J Infect Dis* 166:1393–1400
22. Fontanet AL, Johnston DB, Walker AM, Rooney W, Thimasarn K, Sturchler D, Macdonald M, Hours M, Wirth DF (1993) High prevalence of mefloquine-resistant falciparum malaria in eastern Thailand. *Bull World Health Organ* 71:377–383
23. Fontanet AL, Johnston BD, Walker AM, Bergqvist Y, Hellgren U, Rooney W (1994) Falciparum malaria in eastern Thailand: a randomized trial of the efficacy of a single dose of mefloquine. *Bull World Health Organ* 72:73–78

24. ter Kuile FO, Nosten F, Luxemburger C, Kyle D, Teja-Isavatharm P, Phaipun L, Price R, Chongsuphajaisiddhi T, White NJ (1995) Mefloquine treatment of acute falciparum malaria: a prospective study of non-serious adverse effects in 3673 patients. *Bull World Health Organ* 73:631–642
25. Ter Kuile FO, Teja-Isavatharm P, Edstein MD, Keeratithakul D, Dolan G, Nosten F, Phaipun L, Webster HK, White NJ (1994) Comparison of capillary whole blood, venous whole blood, and plasma concentrations of mefloquine, halofantrine, and desbutyl-halofantrine measured by high-performance liquid chromatography. *Am J Trop Med Hyg* 51:778–784
26. Nosten F, van Vugt M, Price R, Luxemburger C, Thway KL, Brockman A, McGready R, ter Kuile F, Looareesuwan S, White NJ (2000) Effects of artesunate-mefloquine combination on incidence of *Plasmodium falciparum* malaria and mefloquine resistance in western Thailand: a prospective study. *Lancet* 356:297–302
27. Carrara VI, Sirlak S, Thonglairuam J, Rojanawatsirivet C, Proux S, Gilbos V, Brockman A, Ashley EA, McGready R, Krudsood S et al (2006) Deployment of early diagnosis and mefloquine-artesunate treatment of falciparum malaria in Thailand: the Tak Malaria Initiative. *PLoS Med* 3:e183
28. Carrara VI, Zwang J, Ashley EA, Price RN, Stepniewska K, Barends M, Brockman A, Anderson T, McGready R, Phaiphun L et al (2009) Changes in the treatment responses to artesunate-mefloquine on the northwestern border of Thailand during 13 years of continuous deployment. *PLoS One* 4:e4551
29. Price R, Robinson G, Brockman A, Cowman A, Krishna S (1997) Assessment of *pfmdr 1* gene copy number by tandem competitive polymerase chain reaction. *Mol Biochem Parasitol* 85:161–169
30. Price RN, Uhlemann AC, Brockman A, McGready R, Ashley E, Phaipun L, Patel R, Laing K, Looareesuwan S, White NJ et al (2004) Mefloquine resistance in *Plasmodium falciparum* and increased *pfmdr 1* gene copy number. *Lancet* 364:438–447
31. Simpson JA, Watkins ER, Price RN, Aarons L, Kyle DE, White NJ (2000) Mefloquine pharmacokinetic-pharmacodynamic models: implications for dosing and resistance. *Antimicrob Agents Chemother* 44:3414–3424
32. Palmer KJ, Holliday SM, Brogden RN (1993) Mefloquine: a review of its antimalarial activity, pharmacokinetic properties and therapeutic efficacy. *Drugs* 45:430–475
33. Ashley EA, McGready R, Hutagalung R, Phaiphun L, Slight T, Proux S, Thwai KL, Barends M, Looareesuwan S, White NJ et al (2005) A randomized, controlled study of a simple, once-daily regimen of dihydroartemisinin-piperaquine for the treatment of uncomplicated, multi-drug-resistant falciparum malaria. *Clin Infect Dis* 41:425–432
34. Bem JL, Kerr L, Stuerchler D (1992) Mefloquine prophylaxis: an overview of spontaneous reports of severe psychiatric reactions and convulsions. *J Trop Med Hyg* 95:167–179
35. Phillips-Howard PA, ter Kuile FO (1995) CNS adverse events associated with antimalarial agents. Fact or fiction? *Drug Saf* 12:370–383
36. WHO (1991) Review of central nervous system adverse events related to the antimalarial drug, mefloquine (1985–1990). World Health Organization, Geneva
37. Schlagenhauf P, Steffen R, Lobel H, Johnson R, Letz R, Tschopp A, Vranjes N, Bergqvist Y, Ericsson O, Hellgren U et al (1996) Mefloquine tolerability during chemoprophylaxis: focus on adverse event assessments, stereochemistry and compliance. *Trop Med Int Health* 1:485–494
38. Roche Laboratories Inc (2008) Lariam® (mefloquine hydrochloride) Complete Product Information. US Package Insert, Nutley, NJ
39. Nevin RL, Pietrusiak PP, Caci JB (2008) Prevalence of contraindications to mefloquine use among USA military personnel deployed to Afghanistan. *Malar J* 7:30
40. Hill DR (1991) Pre-travel health, immunization status, and demographics of travel to the developing world for individuals visiting a travel medicine service. *Am J Trop Med Hyg* 45:263–270

41. Nevin RL (2009) Epileptogenic potential of mefloquine chemoprophylaxis: a pathogenic hypothesis. *Malar J* 8:188
42. Taylor WR, White NJ (2004) Antimalarial drug toxicity: a review. *Drug Saf* 27:25–61
43. Baird JK (2005) Effectiveness of antimalarial drugs. *N Engl J Med* 352:1565–1577
44. Weinke T, Trautmann M, Held T, Weber G, Eichenlaub D, Fleischer K, Kern W, Pohle HD (1991) Neuropsychiatric side effects after the use of mefloquine. *Am J Trop Med Hyg* 45:86–91
45. Luxemburger C, Nosten F, ter Kuile F, Frejacques L, Chongsuphajaisiddhi T, White NJ (1991) Mefloquine for multidrug-resistant malaria. *Lancet* 338:1268
46. Nguyen TH, Day NP, Ly VC, Waller D, Mai NT, Bethell DB, Tran TH, White NJ (1996) Post-malaria neurological syndrome. *Lancet* 348:917–921
47. Pham YT, Nosten F, Farinotti R, White NJ, Gimenez F (1999) Cerebral uptake of mefloquine enantiomers in fatal cerebral malaria. *Int J Clin Pharmacol Ther* 37:58–61
48. Gimenez F, Gillotin C, Basco LK, Bouchaud O, Aubry AF, Wainer IW, Le Bras J, Farinotti R (1994) Plasma concentrations of the enantiomers of halofantrine and its main metabolite in malaria patients. *Eur J Clin Pharmacol* 46:561–562
49. Pham YT, Regina A, Farinotti R, Couraud P, Wainer IW, Roux F, Gimenez F (2000) Interactions of racemic mefloquine and its enantiomers with P-glycoprotein in an immortalised rat brain capillary endothelial cell line, GPNT. *Biochim Biophys Acta* 1524:212–219
50. Toovey S (2009) Mefloquine neurotoxicity: a literature review. *Travel Med Infect Dis* 7:2–6
51. Milner E, McCalmont W, Bhonsle J, Caridha D, Carroll D, Gardner S, Gerena L, Gettayacamin M, Lanteri C, Luong T et al (2010) Structure-activity relationships amongst 4-position quinoline methanol antimalarials that inhibit the growth of drug sensitive and resistant strains of *Plasmodium falciparum*. *Bioorg Med Chem Lett* 20:1347–1351
52. Milner E, McCalmont W, Bhonsle J, Caridha D, Cobar J, Gardner S, Gerena L, Goodine D, Lanteri C, Melendez V et al (2010) Anti-malarial activity of a non-piperidine library of next-generation quinoline methanols. *Malar J* 9:51
53. Weiss SM, Benwell K, Cliffe IA, Gillespie RJ, Knight AR, Lerpiniere J, Misra A, Pratt RM, Revell D, Upton R et al (2003) Discovery of nonxanthine adenosine A2A receptor antagonists for the treatment of Parkinson's disease. *Neurology* 61:S101–S106
54. Gillespie RJ, Adams DR, Bebbington D, Benwell K, Cliffe IA, Dawson CE, Dourish CT, Fletcher A, Gaur S, Giles PR et al (2008) Antagonists of the human adenosine A2A receptor. Part 1: Discovery and synthesis of thieno[3,2-d]pyrimidine-4-methanone derivatives. *Bioorg Med Chem Lett* 18:2916–2919
55. Gofton TE, Al-Khotani A, O'Farrell B, Ang LC, McLachlan RS (2011) Mefloquine in the treatment of progressive multifocal leukoencephalopathy. *J Neurol Neurosurg Psychiatry* 82:452–455
56. Brickelmaier M, Lugovskoy A, Kartikeyan R, Reviriego-Mendoza MM, Allaire N, Simon K, Frisque RJ, Gorelik L (2009) Identification and characterization of mefloquine efficacy against JC virus *in vitro*. *Antimicrob Agents Chemother* 53:1840–1849
57. Keiser J, Utzinger J (2010) The drugs we have and the drugs we need against major helminth infections. *Adv Parasitol* 73:197–230
58. Holtfreter MC, Loebermann M, Klammt S, Sombetzki M, Bodammer P, Riebold D, Kinzelbach R, Reisinger EC (2011) *Schistosoma mansoni*: schistosomicidal effect of mefloquine and primaquine *in vitro*. *Exp Parasitol* 127:270–276
59. Xiao SH, Xue J, Li-Li X, Zhang YN, Qiang HQ (2010) Effectiveness of mefloquine against *Clonorchis sinensis* in rats and *Paragonimus westermani* in dogs. *Parasitol Res* 107:1391–1397
60. Aparakkitanon S, Chapoomram S, Kuaha K, Chirachariyavej T, Wilairat P (2006) Targeting of hematins by the antimalarial pyronaridine. *Antimicrob Agents Chemother* 50:2197–2200
61. Chavalitshewinkoon-Petmitr P, Pongvilairat G, Aparakkitanon S, Wilairat P (2000) Gametocytocidal activity of pyronaridine and DNA topoisomerase II inhibitors against multi-drug-resistant *Plasmodium falciparum in vitro*. *Parasitol Int* 48:275–280

62. Tshefu AK, Gaye O, Kayentao K, Thompson R, Bhatt KM, Sesay SS, Bustos DG, Tjitra E, Bedu-Addo G, Borghini-Fuhrer I et al (2010) Efficacy and safety of a fixed-dose oral combination of pyronaridine-artesunate compared with artemether-lumefantrine in children and adults with uncomplicated *Plasmodium falciparum* malaria: a randomised non-inferiority trial. *Lancet* 375:1457–1467
63. Park SH, Pradeep K (2010) Absorption, distribution, excretion, and pharmacokinetics of 14 C-pyronaridine tetraphosphate in male and female Sprague-Dawley rats. *J Biomed Biotechnol* 2010:590707
64. Wattanavijitkul T. (2010) Population pharmacokinetics of pyronaridine in the treatment of malaria. Ph.D. dissertation, University of Iowa. <http://ir.uiowa.edu/etd/622>. Accessed 15 Apr 2011
65. Looareesuwan S, Kyle DE, Viravan C, Vanijanonta S, Wilairatana P, Wernsdorfer WH (1996) Clinical study of pyronaridine for the treatment of acute uncomplicated falciparum malaria in Thailand. *Am J Trop Med Hyg* 54:205–209
66. Elueze EI, Croft SL, Warhurst DC (1996) Activity of pyronaridine and mepacrine against twelve strains of *Plasmodium falciparum* *in vitro*. *J Antimicrob Chemother* 37:511–518
67. de Villiers KA, Marques HM, Egan TJ (2008) The crystal structure of halofantrine-ferriprotoporphyrin IX and the mechanism of action of arylmethanol antimalarials. *J Inorg Biochem* 102:1660–1667
68. Sanchez CP, Rotmann A, Stein WD, Lanzer M (2008) Polymorphisms within PfMDR1 alter the substrate specificity for anti-malarial drugs in *Plasmodium falciparum*. *Mol Microbiol* 70:786–798
69. ter Kuile FO, Dolan G, Nosten F, Edstein MD, Luxemburger C, Phaipun L, Chongsuphajaisiddhi T, Webster HK, White NJ (1993) Halofantrine versus mefloquine in treatment of multidrug-resistant falciparum malaria [see comments]. *Lancet* 341:1044–1049
70. Nosten F, ter Kuile F, Luxemburger C, Woodrow C, Kyle DE, Chongsuphajaisiddhi T, White NJ (1993) Cardiac effects of antimalarial treatment with halofantrine. *Lancet* 341:1054–1056
71. Ashley EA, Stepniewska K, Lindegardh N, Annerberg A, Kham A, Brockman A, Singhasivanon P, White NJ, Nosten F (2007) How much fat is necessary to optimize lumefantrine oral bioavailability? *Trop Med Int Health* 12:195–200
72. Tie H, Walker BD, Singleton CB, Valenzuela SM, Bursill JA, Wyse KR, Breit SN, Campbell TJ (2000) Inhibition of HERG potassium channels by the antimalarial agent halofantrine. *Br J Pharmacol* 130:1967–1975
73. Warhurst DC, Craig JC, Adagu IS, Guy RK, Madrid PB, Fivelman QL (2007) Activity of piperazine and other 4-aminoquinoline antiplasmodial drugs against chloroquine-sensitive and resistant blood-stages of *Plasmodium falciparum*. Role of beta-haematin inhibition and drug concentration in vacuolar water- and lipid-phases. *Biochem Pharmacol* 73:1910–1926
74. Tarning J, Lindegardh N, Annerberg A, Singtoroj T, Day NP, Ashton M, White NJ (2005) Pitfalls in estimating piperazine elimination. *Antimicrob Agents Chemother* 49:5127–5128
75. Tarning J, Ashley EA, Lindegardh N, Stepniewska K, Phaiphun L, Day NP, McGready R, Ashton M, Nosten F, White NJ (2008) Population pharmacokinetics of piperazine after two different treatment regimens with dihydroartemisinin-piperazine in patients with *Plasmodium falciparum* malaria in Thailand. *Antimicrob Agents Chemother* 52:1052–1061
76. Briolant S, Henry M, Oeuvray C, Amalvict R, Baret E, Didillon E, Rogier C, Pradines B (2010) Absence of association between piperazine *in vitro* responses and polymorphisms in the *pfert*, *pfmdr1*, *pfmnp*, and *pfhhe* genes in *Plasmodium falciparum*. *Antimicrob Agents Chemother* 54:3537–3544
77. Muangnoicharoen S, Johnson DJ, Looareesuwan S, Krudsood S, Ward SA (2009) Role of known molecular markers of resistance in the antimalarial potency of piperazine and dihydroartemisinin *in vitro*. *Antimicrob Agents Chemother* 53:1362–1366
78. Ashley EA, Stepniewska K, Lindegardh N, McGready R, Annerberg A, Hutagalung R, Singtoroj T, Hla G, Brockman A, Proux S et al (2007) Pharmacokinetic study of artemether-lumefantrine given once daily for the treatment of uncomplicated multidrug-resistant falciparum malaria. *Trop Med Int Health* 12:201–208



79. van Vugt M, Wilairatana P, Gemperli B, Gathmann I, Phaipun L, Brockman A, Luxemburger C, White NJ, Nosten F, Looareesuwan S (1999) Efficacy of six doses of artemether-lumefantrine (benflumetol) in multidrug-resistant *Plasmodium falciparum* malaria. *Am J Trop Med Hyg* 60:936–942
80. Price RN, Uhlemann AC, van Vugt M, Brockman A, Hutagalung R, Nair S, Nash D, Singhasivanon P, Anderson TJ, Krishna S et al (2006) Molecular and pharmacological determinants of the therapeutic response to artemether-lumefantrine in multidrug-resistant *Plasmodium falciparum* malaria. *Clin Infect Dis* 42:1570–1577
81. White NJ, van Vugt M, Ezzet F (1999) Clinical pharmacokinetics and pharmacodynamics and pharmacodynamics of artemether-lumefantrine. *Clin Pharmacokinet* 37:105–125
82. McGready R, Tan SO, Ashley EA, Pimanpanarak M, Viladpai-Nguen J, Phaiphun L, Wustefeld K, Barends M, Laochan N, Keereecharoen L et al (2008) A randomised controlled trial of artemether-lumefantrine versus artesunate for uncomplicated *Plasmodium falciparum* treatment in pregnancy. *PLoS Med* 5:e253
83. Anderson TJ, Nair S, Qin H, Singlam S, Brockman A, Paiphun L, Nosten F (2005) Are transporter genes other than the chloroquine resistance locus (*pfcr1*) and multidrug resistance gene (*pfmdr*) associated with antimalarial drug resistance? *Antimicrob Agents Chemother* 49:2180–2188
84. Johnson DJ, Fidock DA, Mungthin M, Lakshmanan V, Sidhu AB, Bray PG, Ward SA (2004) Evidence for a central role for PfCRT in conferring *Plasmodium falciparum* resistance to diverse antimalarial agents. *Mol Cell* 15:867–877