Chapter 5 Environmentally Benign Synthesis of Metal Nanoparticles for Fertilizer Applications in Agriculture

Mohammad Enayet Hossain, Paramita Saha, and Achintya N. Bezbaruah

1 Introduction

The global population has been growing steadily over the past few centuries. According to the Food and Agriculture Organization of the United Nations (FAO, [2009\)](#page-21-0), the global population will grow by 2.3 billion, between 2009 and 2050, and to feed a world population of 9.1 billion people in 2050, food production will have to increase by 70%. However, the productivity of crops has been decreasing due to biotic and abiotic stresses, climate change, and lack of water. As a result, agricultural development is being severely affected worldwide (Vijayakumar et al., [2022\)](#page-25-0). As such, world agriculture is beset with a wide range of challenges, such as stagnating crop yields, low nutrient utilization effciency, declining soil organic matter, deficiencies of several nutrients, shrinking arable lands, less water availability, shortage of labor, etc. (Raliya et al., [2017](#page-23-0)). Moreover, with a declining rural labor force and increasing food and fber needs, agriculture is facing multiple challenges in the twenty-frst century, which include producing more food and fbers to feed a growing population, producing more feedstocks for a growing bioenergy market, contributing to the overall development of many agriculture-dependent developing countries, adopting more sustainable and efficient production methods, and adapting to climate change (FAO, [2009](#page-21-0)). To resolve these issues, farming communities have been using chemical fertilizers and pesticides and genetically modifed or disease-resistant crop varieties for the past fve decades (Chhipa, [2017\)](#page-20-0).

M. E. Hossain (⊠) · P. Saha

Department of Soil, Water, and Environment, University of Dhaka, Dhaka, Bangladesh e-mail: enayetswe@du.ac.bd

A. N. Bezbaruah Department of Civil, Construction, and Environmental Engineering, North Dakota State University, Fargo, ND, USA

[©] The Author(s), under exclusive license to Springer Nature Switzerland AG 2024 125 K. A. Abd-Elsalam, M. A. Alghuthaymi (eds.), *Nanofertilizers for Sustainable Agroecosystems*, Nanotechnology in the Life Sciences, [https://doi.org/10.1007/978-3-031-41329-2_5](https://doi.org/10.1007/978-3-031-41329-2_5#DOI)

Although the use of chemical fertilizers and pesticides signifcantly enhance food production, food quality and soil fertility are negatively impacted. Moreover, the application of fertilizers and pesticides is not effcient from the standpoint of economy. Most of the applied agrochemicals are lost via different processes such as leaching, mineralization, and bioconversion (Bollag et al., [1992](#page-20-1)). From an estimate, 40–70% nitrogen (N), 80–90% phosphorus (P), and 50–90% potassium (K) fertilizers were found to be either lost or fxed in soils, leading to economic losses (El-Saadony et al., [2021](#page-21-1)). Additionally, the overuse of pesticides and artifcial fertilizers has disrupted many ecosystems and created several health risks. Therefore, a different solution is required for precision farming and improving the existing circumstance. Nanotechnology is a key strategy for resolving this problem.

After biotechnology, nanotechnology is the ffth breakthrough technology of the century. It has demonstrated a broad range of applications in many felds, including agriculture, medicine, biology, physics, chemistry, electronics, energy, materials science, and environmental science (Chhipa, [2017](#page-20-0)). The Greek term "nano" signifes "one billionth of something." One nanometer is defned as one billionth of a meter. The science of nanotechnology focuses on creating and modifying materials with sizes between one and one hundred nanometers $(1-100 \text{ nm})$ (Vijayakumar et al., [2022\)](#page-25-0). With a focus on protecting soil and promoting environmental sustainability, nanotechnology is quickly becoming the essential enabling technology that helps boost agricultural output. The major drivers for motivating the scientifc community to concentrate on advancing the expansion of nano-agrotechnology are challenging climatic conditions and increased global food security (Sangeetha et al., [2017\)](#page-23-1). The improvement in nanotechnology has gained momentum through the innovation of nanoparticles (NPs). Surface area, pore size, particle shape, and reactivity are some of the distinct physical and chemical characteristics that defne nanoparticles (NPs). Because of their widespread use in the agricultural sector, NPs are also known as "magic bullets." Nanoparticles can be employed as nanofertilizers, nanopesticides, and nanoherbicides, which can help crops grow more productively, reduce the overuse of chemical fertilizers, and improve their ability to withstand biotic stress. They control plant growth and boost metabolic activity. Depending on the type and concentration employed, NPs may have a benefcial or detrimental impact on the growth and yield of different plant species (Goswami & Mathur, [2019\)](#page-21-2).

Site-directed delivery and controlled delivery of functional components are two features of nano-enabled agrochemicals that increase their efficiency and capacity for managing pests and illnesses. As a result, they present a fresh method of lowering the toxicity of agrochemicals to human health by minimizing their long-term consequences and reducing environmental pollution by lowering their volatilization, leaching, and drainage. Such nano-enabled agrochemicals improve crop nutrient uptake, solubility, and stability and also provide a workable alternative for managing pests and diseases (Rodrigues et al., [2017](#page-23-2); Duhan et al., [2017;](#page-21-3) Aranaz et al., [2010](#page-20-2); Sarkar et al., [2022\)](#page-24-0).

2 Synthesis of Metal Nanoparticles

For the creation and stabilization of metallic nanoparticles, a variety of physical and chemical techniques, including electrochemical changes, chemical reduction, and photochemical reduction, are frequently used. The choice of metallic nanoparticle preparation technique is crucial because processes used in nanoparticle synthesis, such as the kinetics of metal ions' interactions with reducing agents, the process by which stabilizing agents adhere to metal nanoparticles, and various experimental techniques, have a signifcant impact on the stability, physicochemical properties, and morphology (structure and size) of the nanoparticles (Jamkhande et al., [2019\)](#page-21-4). Metal nanoparticles can be produced using a variety of techniques. However, their synthesis can be roughly categorized into two approaches: (i) the top-down approach and (ii) the bottom-up approach.

2.1 Top-Down and Bottom-Up Approaches

2.1.1 Top-Down Approach

In the top-down method, bulk materials are split to create nanostructured materials. Top-down techniques include electro-explosion, mechanical milling, laser ablation, etching, and sputtering (Baig et al., [2021\)](#page-20-3). The major drawbacks of the top-down method include elevated levels of contaminants in the fnished product and poor control over the size and surface structure of the resultant NPs (Zulfiqar et al., [2019;](#page-25-1) Ndaba et al., [2022](#page-23-3)). Inadequacies in the surface structure indicate a significant disadvantage of the top-down method. Due to their high aspect ratio, these surface structure restrictions can have a considerable negative impact on the physical characteristics and surface chemistry of metallic NPs (Saratale et al.*,* [2018a](#page-24-1), [b](#page-24-2)).

2.1.2 Bottom-Up Approach

The bottom-up method entails creating NPs from much smaller units like atoms and molecules. This method involves common chemical processes along with biological processes. Since the procedure provides for better control of particle size and reduces the quantity of contaminants in the fnished product, NPs manufactured utilizing the bottom-up method are more homogeneous (Ndaba et al., [2022](#page-23-3)).

The fundamental distinction between the two approaches is the raw material used to prepare the nanoparticles. While atoms or molecules are the starting material in bottom-up approaches, top-down methods start with bulk material and use various physical, chemical, and mechanical processes to reduce the particle size to nanoparticles (Jamkhande et al., [2019\)](#page-21-4). These two methodologies primarily rely on diverse physical, chemical, and biological techniques. Most of the physical

Fig. 5.1 There are two main methods for synthesizing nanoparticles: top-down and bottom-up. (1) Top-down strategy: Using mechanical or chemical methods, the top-down strategy breaks down large materials into smaller nanoparticles. Starting with a huge piece of material, this method often includes shrinking it down to the desired nanoparticle size range using physical or chemical procedures. Top-down techniques include milling, lithography, and etching as examples. (2) Bottom-up strategy: In the bottom-up strategy, individual atoms or molecules are put together to create nanoparticles. In this method, the required nanoparticle structure is built up from individual atoms or molecules using chemical or physical processes. Chemical vapor deposition, sol–gel synthesis, and coprecipitation are a few examples of bottom-up techniques

approaches, along with some chemical ways, are included in the top-down strategy; meanwhile, the bottom-up approach primarily concentrates on chemical and biological processes to synthesize metal nanoparticles. Figure [5.1](#page-3-0) depicts the top-down and bottom-up approaches.

2.2 Physical, Chemical, and Biological Methods

2.2.1 Physical Methods

Top-down is a physical procedure dependent on material milling. This method's drawbacks include a lack of control over nanoparticle size and a higher level of contaminants. Mechanical milling, laser ablation, sputtering, and other typical physical processes are utilized to create metal nanoparticles.

Mechanical Milling

A feasible method for creating materials at the nanoscale from bulk materials is mechanical milling. It is a useful technique for creating mixtures of various phases and is useful in the creation of nanocomposites (Baig et al., [2021](#page-20-3)). It involves the

structural decomposition of coarser particles into smaller ones. In this technique, a container is flled with bulk powder and numerous large balls. With the aid of a high-speed spinning ball, high mechanical energy is imparted to bulk powder material. Various high-energy mills can be used for particle size reduction. According to Rajput [\(2015](#page-23-4)), these high-energy mills include:

- Attrition ball mill
- Planetary ball mill
- Vibrating ball mill
- Low-energy tumbling mill
- High-energy ball mill

In each of these methods, large, freely moving, high-energy balls can either fall freely and strike the powder or can roll down the surface of the chamber housing the bulk powder material in a succession of parallel layers. It is a commonly used technique for mechanical alloying to create amorphous alloys for a variety of uses, including metal–metal, transition metal–metalloid, and metal–carbon systems.

Laser Ablation

The laser irradiation employed in the laser ablation method causes the particle size to be reduced to the nanoscale. After being covered by a thin layer, the solid target material is exposed to pulsed laser irradiation. The most used lasers are copper vapor lasers, titanium-doped sapphire lasers, Nd: YAG (neodymium-doped yttrium aluminum garnet) lasers at 106 m output, and their harmonics. When a material is exposed to laser energy, it breaks down into tiny pieces called nanoparticles (Jamkhande et al., [2019\)](#page-21-4). This method is used to produce aluminum oxide (AI_2O_3) metal nanoparticles and other metalloid nanoparticles.

Sputtering

Ion sputtering is a technique that involves vaporizing a material by sputtering with a stream of ions from an inert gas. It involves bombarding solid surfaces with highenergy particles, such as plasma or gas, to create nanomaterials. Sputtering is believed to be a useful technique for creating thin nanomaterial flms (Baig et al., [2021\)](#page-20-3). It can be carried out in a variety of ways, including using radio-frequency diodes, magnetrons, and direct current (DC) diodes. Recently, employing magnetron sputtering of metal targets, this technique has been used to create nanoparticles from a variety of metals.

2.2.2 Chemical Methods

Sol–Gel Process

Compared to regular molecules or nanoparticles, colloidal particles are signifcantly bigger. However, colloids become bulky when mixed with a liquid, whereas nanoscale molecules always appear transparent. It involves the development of networks through the production of colloidal suspension (sol) and gelatin to create a network in a continuous liquid phase (gel). Metal alkoxide and alkoxysilane ions serve as the precursor to the synthesis of these colloids. Tetramethoxysilane (TMOS) and tetraethoxysilane (TEOS), which create silica gels, are the most often utilized. Alkoxides cannot be mixed with water. They are silica, aluminum, titanium, zirconium, and many more organometallic precursors. Alcohol is utilized as a mutual solvent. An initial homogeneous solution of one or more chosen alkoxides is used in the sol–gel procedure. These serve as organic precursors to materials like zirconia, titania, alumina, silica, and more. The catalyst controls pH and initiates the reaction. Four phases are involved in sol–gel formation: 1. hydrolysis, 2. condensation, 3. growth of particles, and 4. agglomeration of particles (Rajput, [2015](#page-23-4)).

Electrochemical Precipitation

This strategy uses an arrested precipitation mechanism to manage size. The fundamental strategy is to create and study the nanomaterial in situ, or in the same liquid media, to prevent physical changes and the accumulation of microscopic crystallites. Double-layer repulsion of crystallites utilizing nonaqueous solvents at lower temperatures for synthesis was used to control thermal coagulation and Oswald ripening. The synthesis involved constituent materials reacting with one another in an appropriate solvent. Prior to the precipitation reaction, the dopant is incorporated into the parent solution. A surfactant is employed to keep the produced particles apart. The resulting nanocrystals are centrifuged apart, cleaned, and vacuum dried. The dried material is then subjected to ultraviolet (UV) curing to see whether the surfactant capping coating on the nanocluster's surface could polymerize and provide real quantum confnement (Rajput, [2015\)](#page-23-4).

Vapor Deposition

A solid is deposited on a heated surface through a chemical reaction from the vapor or gas phase in a process known as chemical vapor deposition (CVD). In thermal CVD, a high temperature of more than 900 °C activates the process. An exhaust system, a deposition chamber, and a gas supply system make up a typical apparatus. Plasma at temperatures between 300 and 700 °C initiates the reaction in plasma CVD. Pyrolysis takes place in laser CVD when a heat-absorbing substrate is heated by a laser's thermal energy. Ultraviolet radiation that has enough photon energy to

break the chemical bond in the reactant molecules is used to trigger the chemical reaction in photo-laser CVD. This method involves photon activation of the reaction, and deposition takes place at room temperature. Nanocomposite powders can be synthesized using CVD (Rajput, [2015\)](#page-23-4).

2.2.3 Biological Methods

The biological method involves various biological entities such as microbes (bacteria, algae, fungi, viruses), plants, organic wastes, etc.

3 Why Environmentally Benign Synthesis of Metal Nanoparticles (NPs) Is Necessary

Although physical and chemical processes have been employed for decades to produce nanoparticles, there are still many issues with them. The basic drawbacks of physical procedures are (i) excessive production cost, (ii) consumption of large amounts of energy, and (iii) low manufacturing yield (Shedbalkar et al., [2014\)](#page-24-3).

According to Gahlawat and Choudhury ([2019\)](#page-21-5), chemical methods result in more uniform NPs in terms of size and shape, and the reduction step does not require as much energy. Therefore, the most preferred method of NP synthesis throughout the past decade has been chemical synthesis. However, chemical techniques of NP synthesis entail the use of toxic chemicals that are associated with cytotoxicity, carcinogenicity, and genotoxicity, contributing to the notion that such processes are environmentally hazardous.

In contrast, NPs produced by biological means are regarded as clean, safe, economical, and nontoxic when compared to conventional ways; as such, they are suggested as potential environmentally friendly substitutes for chemical and physical processes. Plants and microbes have the ability to gather and absorb metallic ions from their surroundings, making them suitable candidates for the synthesis of nanomaterials. Although a wide variety of biological entities are utilized in the production of NPs, plants, algae, fungi, yeast, bacteria, actinomycetes, and viruses are the most frequently used bioorganisms (Saratale et al.*,* [2018a](#page-24-1)).

3.1 Green Synthesis of Metal Nanoparticles

The biological synthesis of metal NPs has advanced signifcantly and is currently being developed as an alternative environmentally friendly procedure. The environmentally benign biological synthesis of NPs is commonly referred to as "green synthesis" or "green chemistry" processes. Using entire cells, metabolites, or extracts from plants and microbes as environmentally friendly raw materials, the

green synthesis of nanoparticles creates metallic nanoparticles. It has advantages over chemical and physical processes in that it is secure, straightforward, costeffective, reasonably reproducible, and it frequently produces more stable materials (Adelere & Lateef, [2016](#page-20-4)).

Plants and plant parts have been extensively used recently in the synthesis of numerous nanoparticles due to the rich biodiversity of plants and their potential secondary metabolites. Alkaloids, favonoids, saponins, steroids, tannins, and other benefcial natural chemicals are prevalent in plant extracts. These items can be made from a variety of plant parts, including leaves, stems, roots, shoots, fowers, barks, and seeds. In the bioreduction technique used to create metallic nanoparticles, they serve as reducing and stabilizing agents. Many greener nanoparticles, including cobalt, copper, silver, gold, palladium, platinum, zinc oxide, and magnetite, have been successfully synthesized using plants (Adelere & Lateef, [2016\)](#page-20-4).

A wide range of materials, including plants and plant products, algae, fungi, yeasts, bacteria, and viruses, can be used in the biological production of NPs. Precursors of noble metal salts are combined with biomaterials to begin the production of NPs. Proteins, alkaloids, favonoids, reducing sugars, polyphenols, and other substances are present in biomaterials and act as reducing and capping agents for the synthesis of NPs from their metal salt predecessors. The color shift of the colloidal solution can be used to visually check the reduction of the metal salt precursor to its subsequent NPs. In the recent past, several research documented the synthesis of Ag, Au, Cu, Pt, Cd, Pt, Pd, Ru, Rh, etc. utilizing different biological agents (Dikshit et al., [2021\)](#page-21-6). Figure [5.2](#page-7-0) describes the general steps in the biosynthesis of metal nanoparticles both using microorganisms and plant elements.

Fig. 5.2 A schematic representation of metallic nanoparticle biosynthesis. (Modifed from Kumari et al. [\(2020](#page-22-0)), Ndaba et al. [\(2022](#page-23-3)), and Dikshit et al. [\(2021](#page-21-6)))

3.2 Microbial Synthesis of Metal NPs

Various microorganisms are involved in the production of metal nanoparticles because of their properties. Among the microorganisms, bacteria, fungi, algae, and viruses are the most common. Bacteria and viruses can survive in various adverse environments, and, owing to their ease of culture and less production costs, they can be broadly used to produce MtNPs.

3.2.1 Bacteria-Mediated Synthesis of Nanoparticles

Diverse groups of bacteria were used to synthesize various metal nanoparticles. As they can grow faster and can adapt to different adverse environments, bacteria are used nowadays for production, although the mechanism is not yet fully understood.

A *Bacillus subtilis* EWP-46 cell-free extract was used for the reduction of nitrate in silver NP (AgNP) production. Several variables, including hydrogen ion concentration, temperature, silver ion $(Ag⁺ ion)$, and time, influenced the formation of AgNPs. More AgNPs were found to be produced when the conditions were held constant at pH 10.0, 60 $^{\circ}$ C, 1.0 mM Ag⁺ ion, and 720 min. AgNPs were tested against Gram-positive (*Staphylococcus aureus*) and Gram-negative (*Pseudomonas fuorescens*) bacteria to determine their primary inhibitory focus and least bactericidal convergence (Velmurugan et al., [2014](#page-24-4)).

In another study, *Bacillus licheniformis* cell-free extract (BLCFE)-coated silver nanoparticles were produced by the organism with an average particle size of 18–63 nm, and the synthesized nanoparticles resulted in disintegrated bioflm production of *Vibrio parahaemolyticus* (Shanthi et al., [2016](#page-24-5)).

Ghorbani ([2017\)](#page-21-7) used *Salmonella typhimurium* for the fast production of AgNPs. Table [5.1](#page-9-0) summarizes some of the past research studies that were conducted involving the green synthesis of metallic nanoparticles using bacteria.

3.2.2 Fungi-Mediated Synthesis of Metal Nanoparticles

For the biological synthesis of metal nanoparticles, several fungal families have been investigated, including *Alternaria*, *Amylomyces*, *Aspergillus*, *Bipolaris*, *Candida*, *Cladosporium*, *Colletotrichum*, *Coriolus*, *Cylindrocladium*, *Fusarium*, *Ganoderma*, *Helminthosporium*, *Humicola*, *Lecanicillium*, *Mucor*, *Neurospora*, *Penicillium*, *Pestalotiopsis*, and *Phanerochaete*. The *Aspergillus* and *Fusarium* fungus families have been the most thoroughly studied for the nanosynthesis of the following metals and their metal oxides: Au, Ag, Ti, Zn, Ce, Fe, Mg, P, and Pt (Chhipa, [2019](#page-21-8)).

Trichoderma reesei, among the *Trichoderma* species, is used for the mycosynthesis of AgNPs. The ability of this fungi to detoxify microclimates makes them eligible for the biosynthesis of nanoparticles. These AgNPs have antimicrobial or

| | Metallic | Size | | Cellular | |
|---|--------------------------------|-------------------|---------------------------------------|---------------|--------------------------------------|
| Bacteria | nanoparticles | (nm) | Morphology | location | References |
| Bacillus subtilis | Ag | $3 - 20$ | Spherical | ND | Alsamhary (2020) |
| Bacillus licheniformis | Ag | 40 | ND | ND | Kalishwaralal et al. (2008) |
| Pseudomonas stutzeri | Ag | 200 | Triangular | ND | Klaus et al. (1999) |
| Actinobacteria | Ag | 13.2 | Spherical | ND | Wypij et al. (2017) |
| Ochrobactrum anthropi | Ag | $38 - 85$ | Spherical | ND | Thomas et al. (2014) |
| Pantoea ananatis | Ag | $8.06 -$ 91.31 | Spherical | ND | Monowar et al. (2018) |
| Corynebacterium sp. SH09 | Ag | $10 - 15$ | ND | Intracellular | Narayanan and Sakthivel (2010) |
| Escherichia coli | Ag | 50 | Irregular | ND | Gurunathan et al. (2009) |
| Morganella sp. | Ag | 20 ± 5 | Spherical | Extracellular | Parikh et al. (2008) |
| Bacillus cereus | Ag | $4 - 5$ | Spherical | Intracellular | Babu and Gunasekaran (2009) |
| Bacillus licheniformis | Ag | 50 | Irregular | Intracellular | Kalimuthu et al. (2008) |
| Corynebacterium glutamicum | Ag | $5 - 50$ | Irregular | Extracellular | Sneha et al. (2010) |
| Lactobacillus sp. | Ti | $40 - 60$ | Spherical | Extracellular | Prasad et al. (2007) |
| Desulfobacteraceae | ZnS | $2 - 5$ | Spherical | Intracellular | Labrenz et al. (2000) |
| Desulfobacteraceae | ZnS | $2 - 5$ | Biofilm | ND | Labrenz et al. (2000) |
| Aquaspirillum magnetotacticum | Fe ₃ O ₄ | $40 - 50$ | Octahedral prism | Intracellular | Mann et al. (1984) |
| Magnetospirillum magnetotacticum | Fe ₃ O ₄ | 47.1 | Cuboctahedron | Intracellular | Philip (2009) |
| Magnetospirillum magnetotacticum $(MS-1)$ | Fe ₃ O ₄ | \sim 50 | Cuboctahedron | Intracellular | Lee et al. (2004) |
| Shewanella oneidensis | Fe ₃ O ₄ | $40 - 50$ | Rectangular, rhombic, hexagonal | ND | Suresh et al. (2011) |
| Lactobacillus acidophilus | Se | $2 - 15$ | Spherical | ND | Alam et al. (2020) |
| Lysinibacillus sp. $ZYM-1$ | Se | $100 -$ 200 | Cubic | ND | Che et al. (2017) |

Table 5.1 Metallic nanoparticles synthesized using bacteria and the size and morphology of the synthesized nanoparticles

Modifed from Dikshit et al. [\(2021](#page-21-6)) and Saratale et al. ([2018a,](#page-24-1) [b](#page-24-2)) *ND* not defned

antibacterial abilities, which work against Gram-positive and Gram-negative microorganisms like bacteria (Vahabi & Dorcheh, [2014](#page-24-9)).

The fungal strains of *Aspergillus favus* SP-3, *Trichoderma gamsii* SP-4, *Talaromyces favus* SP-5, and *Aspergillus oryzae* SP-6 were treated with silver nitrate to produce AgNPs in an experiment by Anand et al. ([2015\)](#page-20-9). The synthesized nanoparticles had an average size of 20–60 nm and had antimicrobial properties against both Gram-positive and Gram-negative bacteria.

An *Aspergillus terreus* fltrate was used for AgNP production in an experiment conducted by Li et al. [\(2011](#page-22-6)). The synthesized particle size ranged from 1 to 20 nm. NADH was present in the fungal fltrate, and it acted as a secondary metabolite to convert metal precursors to metal nanoparticles.

Pestalotiopsis longiseta was used for the extracellular production of AgNPs. The particle size ranged from 123 to 195 nm (Vardhana & Kathiravan, [2015](#page-24-10)). Table [5.2](#page-11-0) summarizes the names of the fungi that were used to produce MtNPs.

3.2.3 Algae-Mediated Synthesis of Nanoparticles

Spirogyra varians is utilized for the production of AgNPs and is considered the most feasible method. The produced nanoparticles can be effectively used as an antibacterial agent (Salari et al., [2016](#page-23-9)). Table [5.3](#page-12-0) summarizes the algae used for MtNP production.

3.3 Plant-Mediated Synthesis of Nanoparticles

Anogeissus latifolia, a protein-rich edible gum is used to produce AgNPs. The gum extracts are used to convert metal precursors to metal nanoparticles. The synthesized particles are size controlled and easy to handle. The gum encapsulates AgNPs and increases their effciency as the reaction time increases and it gets more time to get involved in various biological and antimicrobial activities (Kora et al., [2012](#page-22-7)) (Table [5.4\)](#page-13-0). Table [5.4](#page-13-0) summarizes the use of some plant extracts for the synthesis of various MtNPs.

4 Characterization of Metal Nanoparticles

The exploration of nanoparticles' uses, absorption, and toxicology depends heavily on their characterization. Nanoparticles are characterized using a variety of techniques depending on the matrix, analyte, concentration, complexity, and intrinsic qualities (Singh et al., [2021](#page-24-11)). The characterization of metal nanoparticles can be divided into two parts: (i) structural characterization and (ii) morphological

| | Metallic | | | Cellular | | |
|---------------------------------|--------------------------------|----------------------|------------------------|-------------------|--------------------------------|--|
| Fungi | nanoparticles | Size | Morphology | location | References | |
| Fusarium oxysporum | Ag | $5 - 50$ | ND | Extracellular | Senapati et al. (2004) | |
| Fusarium solani USM 3799 | Ag | 16.23 | Spherical | Extracellular | Ingle et al. (2009) | |
| Coriolus versicolor | Ag | 25–75 | Spherical | Extracellular | Sanghi and Verma (2009) | |
| Aspergillus niger | Ag | 20 | Spherical | Extracellular | Gade et al. (2008) | |
| Phoma glomerata | Ag | $60 - 80$ | Spherical | Extracellular | Birla et al. (2009) | |
| Penicillium brevicompactum | Ag | $58.35 \pm$ 17.88 | ND | Extracellular | Shaligram et al. (2009) | |
| Cladosporium cladosporioides | Ag | $10 - 100$ | Spherical | Extracellular | Balaji et al. (2009) | |
| Penicillium fellutanum | Ag | $5 - 25$ | Spherical | Extracellular | Kathiresan et al. (2009) | |
| Aspergillus fumigatus | Ag | $5 - 25$ | Spherical | Extra cellular | Bhainsa and D'souza (2006). | |
| Fusarium oxysporum | Ag | $5 - 15$ | Variable | ND | Mohammadian (2007) | |
| Fusarium semitectum | Ag | $10 - 60$ | Spherical | ND | Basavaraja et al. (2008) | |
| Verticillium sp. | Ag | $5 - 50$ | Spherical | ND | Senapati et al. (2004) | |
| Yeast strain MKY3 | Ag | $2 - 5$ | Hexagonal | Extracellular | Kowshik et al. (2002a) | |
| Yeast strain MKY3 | Ag | $9 - 25$ | Irregular | ND | Kowshik et al. (2002a) | |
| Fusarium oxysporum | Si | $5 - 15$ | Ouasi- spherical | Extracellular | Bansal et al. (2005) | |
| Fusarium oxysporum | Ti | $6 - 13$ | Spherical | Extracellular | Bansal et al. (2005) | |
| Fusarium oxysporum | Zr | $3 - 11$ | Quasi- spherical | Extracellular | Bansal et al. (2004) | |
| Fusarium oxysporum | TiO ₂ | $6 - 13$ | Spherical | ND | Bansal et al. (2005) | |
| Fusarium oxysporum | ZrO ₂ | $3 - 11$ | Spherical | ND | Bansal et al. (2004) | |
| Schizosaccharomyces pombe | CdS | $1 - 1.5$ | Wurtzite- hexagonal | Intracellular | Kowshik et al. (2002b) | |
| Yeast | CdS | 3.6 | Spherical | ND | Prasad and Jha (2010) | |
| Torulopsis sp. | PbS | $2 - 5$ | Spherical | Intracellular | Kowshik et al. (2002b) | |
| Yeast | Fe ₃ O ₄ | < 100 | Wormhole- like | ND | Zhou et al. (2009) | |
| Saccharomyces cerevisiae | s_{b2} ^o 3 | $2 - 10$ | Spherical | ND | Jha et al. (2009) | |

Table 5.2 Metallic nanoparticles synthesized using fungi and the size and morphology of the synthesized nanoparticles

Modifed from Saratale et al. ([2018a,](#page-24-1) [b](#page-24-2))

| | | Size of NPs | | |
|---------------------------------|------------|--------------------------|-----------------------------|--------------------------------------|
| Algal species | NPs | (nm) | Morphology | References |
| Cystophora | Αg | $50 - 100$ | Spherical | Prasad et al. (2013) |
| moniliformis | | | | |
| Caulerpa racemosa | Ag | $0.5 - 2.5$ | Spherical, triangular | Kathiraven et al. (2015) |
| Chaetomorpha linum | Ag | $03 - 44$ | Clusters | Nuraje et al. (2014) |
| Scenedesmus sp. | Αg | $15 - 20$ | Spherical, crystalline | Jena et al. (2014) |
| Gracilaria corticata | Αg | $18 - 46$ | Nanospheres | Kumar et al. (2012) |
| Leptolyngbya valderianum | Αg | $02 - 20$ | Spherical, intracellular | Roychoudhury and Pal (2014) |
| Pithophora oedogonia | Ag | $25 - 44$ | Cubical, hexagonal | Sinha et al. (2015) |
| Porphyra vietnamensis | Ag | 13 ± 03 | Spherical | Venkatpurwar and Pokharkar (2011) |
| Sargassum tenerrimum | Αg | 20 | Spherical | Kumar et al. (2012) |
| Sargassum wightii | Αg | $08 - 27$ | ND | Saratale et al. (2017) |
| Spirogyra varians | Ag | 35 | Quasi-spheres | Salari et al. (2016) |
| Ulva lactuca | Ag | $\overline{}$ | Spherical | Murugan et al. (2015) |
| Sargassum muticum | Ag | $43 - 79$ | Spherical | Madhiyazhagan et al. (2015) |
| Gelidium amansii | Αg | $27 - 54$ | Spherical | Pugazhendhi et al. (2018) |
| Laminaria japonica | Ag | 31 | Spherical to oval | Kim et al. (2018) |
| Chlorococcum sp. MM11 | Fe | $20 - 50$ | Spherical | Vigneshwaran et al. (2006) |
| Sargassum bovinum | Pd | $05 - 10$ | Octahedral | Momeni and Nabipour (2015) |

Table 5.3 Metallic nanoparticle synthesized using algae and the size and morphology of the synthesized nanoparticles

Modifed from Saratale et al. ([2018a,](#page-24-1) [b](#page-24-2))

characterization. Researchers mostly employ Fourier transform infrared (FTIR) spectroscopy, X-ray diffraction (XRD), and X-ray fuorescence (XRF) techniques for the structural characterization of nanomaterials. Scanning electron microscopy (SEM), transmission electron microscopy (TEM), and energy-dispersive X-ray spectroscopy (EDS) techniques can all be used to analyze the morphology of nanomaterials (Samaddar et al., [2018](#page-23-16)).

4.1 Structural Characterization

FTIR spectroscopy is used to characterize the vibrational modes of the precursors and synthesized nanoparticles. The presence of impurities in the fnal product can also be determined using an FTIR spectrogram. If the product's spectrogram shows

| NPs | Plants | Parts | Extractants | Precursors | Size (nm) |
|------------|-------------------------------------|--------------------|--------------------|---------------------------------------|-------------------------|
| AuNPs | Butea monosperma | Leaf | Water | HAuCl ₄ | $10 - 100$ |
| | Pelargonium graveolens | Leaf | Water | HAuCl ₄ | $20 - 40$ |
| | Salix alba | Leaf | Water | $HAuCl_4.3H2O$ | $50 - 80$ |
| | Guazuma ulmifolia L. | Bark | Water | HAuCl ₄ ·3H ₂ O | $20 - 25$ |
| | Nerium oleander | Bark | Methanol | HAuCl ₄ | $20 - 40$ |
| | Rubia cordifolia | Fruit | Ethanol | HAuCl ₄ | $5 - 20$ |
| | Litsea cubeba | Fruit | Water | $HAuCl_4.3H_2O$ | $8 - 18$ |
| | Piper longum | Fruit | Water | HAuCl ₄ | $20 - 200$ |
| | Hibiscus sabdariffa | Flower | Water | HAuCl ₄ .3H ₂ O | $15 - 45$ |
| | Coleus forskohlii | Root | Water | HAuCl ₄ | $5 - 18$ |
| | Stachys lavandulifolia | Overground part | Overground part | HAuCl ₄ | $34 - 80$ |
| AgNPs | Lotus garcinii | Leaf | Water | AgNO ₃ | $7 - 20$ |
| | Morinda citrifolia | Leaf | Methanol | AgNO ₃ | $10 - 100$ |
| | Prunus mume | Fruit | Water | AgNO ₃ | ~ 30 |
| | Eugenia stipitata McVaugh | Fruit | Water | AgNO ₃ | $15 - 45$ |
| | Aconitum toxicum Reichenb. | Root | 96% ethanol | AgNO ₃ | $53 - 67$ |
| | Catharanthus roseus | Bark | Water | AgNO ₃ | $1 - 26$ |
| CuNPs | Ocimum sanctum | Leaf | Water | CuSO ₄ ·5H ₂ O | $50 - 70$ |
| | Hibiscus rosa-sinensis | Flower | Water | $Cu(CH_3COO)_{2}·H_2O$ | $0.115-$ $1.1 \mu m$ |
| PtNPs | Costus speciosus | Leaf | 95% ethanol | Platinum 2,4-pentanedionate | $10 - 50$ |

Table 5.4 Metallic nanoparticles synthesized using plant extracts and the size of the synthesized nanoparticles

Modifed from Bao et al. [\(2021](#page-20-16))

peaks at a different level than the precursor's, then this indicates that there might be some impurities present in the fnal product. The features of MtNPs, including chemical concentration, surface chemistry, surface functional groups, and atomic organization and transmission, are measured using Fourier transform infrared (FTIR) spectroscopy.

The crystalline structure of synthetic nano-samples is investigated using X-ray diffraction (XRD).

The content of different particles contained in the produced nanomaterials is identifed using X-ray fuorescence (XRF) analysis. According to an experiment by Li et al. [\(2016](#page-22-17)), produced α -Fe₂O₃ contained various impurities such as 0.898% SiO_2 , 0.486% TiO₂, and 0.112% MgO.

4.2 Morphological Characterization

Scanning electron microscopy (SEM) is used to morphologically characterize metal nanoparticles. Since the application of metallic nanoparticles is largely dependent on the particle size and shape of the NPs, SEM is used to characterize the internal dispersion of the NPs.

Energy-dispersive X-ray spectroscopic analysis is performed alongside SEM imaging to investigate the distribution pattern of other metal species on synthesized NPs (Samaddar et al., [2018\)](#page-23-16). In an experiment by Fang et al. [\(2011](#page-21-17)), the EDS analysis showed that the amount of nickel (Ni) and Zn on zero-valent FeNPs was too low to be detected. Energy-dispersive X-ray spectroscopy (EDS) is typically used to analyze the elemental composition of MtNPs.

Transmission electron microscopic images are used to determine the particle size or pore size of NPs. Since the synthesis of nanoparticles is size-dependent, a change in temperature can cause alterations in NP size. Therefore, TEM images are used to identify any size change during the synthesis process (Samaddar et al., [2018\)](#page-23-16). The position, size, and shape of MtNPs can be seen using transmission electron microscopy (TEM), scanning electron microscopy (SEM), and atomic force microscopy (AFM).

UV–visible (UV-vis) spectroscopy is another method that is used for structural characterization. MtNPs are typically tested for stability and synthesis using UV– visible spectroscopy.

The size of the NPs at an extremely low level can be estimated using a dynamic light scattering (DLS)/zeta potential size analyzer. Zeta potential describes the surface condition of a nanoparticle and predicts its stability over time (Singh et al., [2021\)](#page-24-11). The size and surface charge of MtNPs is mostly assessed using the dynamic light scattering (DLS) method.

5 Use of Metallic Nanoparticles in Sustainable Agriculture

The primary issue with excessive and prolonged use of chemical fertilizers in agriculture is the decline in soil fertility, which ultimately has an impact on the output of agricultural goods. According to the literature, weeds cause 13% damage, plant infections cause 13% loss, and insect pests cause 14% loss globally. The loss value of crops has been calculated to be USD 2000 billion annually. As a result, it is crucial to increase crops that are resistant to pests and droughts to enhance crop productivity (Rai & Ingle, [2012](#page-23-17); Saratale et al.*,* [2018a\)](#page-24-1). Nanomaterials can be applied to soil systems as both nanofertilizers and nanopesticides. The term "nanofertilizers" refers to nanomaterials that are either nutrients themselves or act as carriers or additions for the nutrients (by, for example, combining with minerals) (macro- or micronutrients). Nutrients can also be enclosed within nanoparticles to create these types of fertilizers (Saleem & Zaidi, [2020\)](#page-23-18).

Nanomaterials that can provide one or more nutrients to plants to promote their development and production are known as nanofertilizers. One of the possible methods for boosting plant growth and productivity to meet the world's rising food demand is the use of nanoparticle fertilizers. The distribution of chemicals to the desired places is made possible by the nanofertilizers' extremely high sorption capacity, surface area, and regulated chemical release kinetics (Snehal & Lohani, [2018\)](#page-24-17). With higher nutrient use effciency, nanofertilizers can boost crop output and quality while lowering production costs, resulting in sustainable agriculture. Nanofertilizers are organic fertilizers or smart fertilizers that provide plants tiny but potent doses of nutrients. Encapsulating nanofertilizers can increase nutrient uptake, which eventually lowers nutrient loss, promotes healthy plant development, and enhances crop quality. By preventing nutrients from interacting with soil, water, air, and microbes, nanoformulations minimize the risk of environmental degradation by providing progressive and controlled release of nutrients to the target regions. The usage of MtNP-based nanofertilizers was found to have a great potential to boost crop productivity (Bahrulolum et al., [2021\)](#page-20-17). The effects of metal nanoparticles as nanofertilizers can be discussed in two pathways. The produced nanoparticles can be applied directly on soil or can be used through foliar application.

MtNPs, which include silver, gold, cadmium, copper, zinc, iron, and selenium (Se), have a variety of uses in agriculture, including promoting plant development, having antibacterial and antifungal effects, and acting as nanofertilizers and nanobiosensors (Bahrulolum et al., [2021\)](#page-20-17). Zn and Fe, two nanonutrients produced by biosynthesis, help plants cope with stress and avoid cell damage. Different microbially produced nanonutrients, such as those containing boron, iron, sulfur, molybdenum, magnesium, nitrogen, phosphorus, and potassium, improved grain yield by 12%–54% and dry matter yield by 18%–34% in crops of caulifower, capsicum, castor, cluster beans, chickpeas, maize, mung beans, pearl millet, rice, tomatoes, and wheat, depending on the nutrient ratios (Chhipa, [2019\)](#page-21-8).

Acid phosphatase, alkaline phosphatase, aryl sulfatase, cellulase, dehydrogenase, esterase, hemicellulase, lignase, and nitrate reductase had an increase in rhizosphere activity between 18% and 283%. Nutrient usage effciency was also enhanced by 3- to 20-fold by nanofertilizers. Smart nanonutrients' large surface area and targeted delivery could drastically reduce the amount of nanofertilizers used at the feld level from kilograms to milligrams, thus reducing fertilizer leaching and eutrophication (Chhipa, [2019\)](#page-21-8).

NPs' actual functionality is determined by how they interact with the substrate on which they have been used. The uptake and distribution of NPs in crops directly affect their actual infuence and impact (Singh et al., [2021\)](#page-24-11).

5.1 Silver Nanoparticles (AgNPs)

Silver ions make up a signifcant portion of these nanoparticles (10–20 nm in size). Inside living systems, these particles are highly active and effective. Various harmful microorganisms have been fought off using silver nanoparticles. Silver nanoparticles, for instance, have been used to treat spot blotch disease. They are also used to break seed dormancy, boost seed vigor index, and raise seedling fresh weight, among other things. Silver nanoparticles are simple to synthesize and safe for the environment. Research on the ability of plants to withstand stress brought on by silver nanoparticles is still ongoing (Singh et al., [2021](#page-24-11)).

5.2 Zinc Oxide Nanoparticles (ZnO NPs)

Due to the lower availability of zinc and its restricted amount in calcium carbonateenriched soils due to their alkaline pH, both soils and plants commonly suffer from zinc defciency. Zinc sulfate fertilizers are used as an alternative to lessen this. Despite this, plants still experience zinc defciency. Due to their tiny size and wide surface area, zinc oxide nanoparticles are the ideal solution to treat zinc deficiency since they are quickly absorbed by plants. The 100-nm size of zinc oxide nanoparticles makes them particularly effective. In order to synthesize zinc oxide nanoparticles, zinc sulfate heptahydrate is typically dissolved in water. Concomitantly, a bioactive extract is obtained from desirable living organisms such as plants and animals. Water or ethanol are used to prepare the extract. The two prepared solutions are then combined at the proper pH to produce the desired zinc oxide nanoparticles. These nanoparticles may offer a low-cost, environmentally safe, and long-lasting solution to a few plant-related issues, including pathogenic invasions and zinc shortages (Singh et al., [2021](#page-24-11)).

5.3 Titanium Dioxide Nanoparticles (TiO₂ NPs)

Titanium dioxide nanoparticles (TiO₂ NPs), which range in size from 5 to 30 nm, are widely produced and applied owing to their photocatalytic properties. Therefore, they are utilized in pigment formation. These nanoparticles help plants grow and photosynthesize more. Broad beans (a common leguminous crop) had their soil salinity reduced by titanium dioxide nanoparticles $(nTiO₂)$ (Abdel Latef et al., [2018\)](#page-20-18). Additionally, it is applied via roots or foliar spray at extremely low concentrations to promote plant growth, enzymatic activity, photosynthesis of chlorophyll content that promotes nutrient uptake, and stress tolerance and to increase crop yield and quality (Singh et al., [2021](#page-24-11)).

5.4 Iron Oxide Nanoparticles (Fe₂O₃ NPs)

Due to their ability to substitute conventionally ineffective Fe fertilizers, iron oxide nanoparticles ($Fe₂O₃$ NPs) are vital oxide nanomaterials that are extensively used in agriculture. They typically range in size from 10 to 20 nm and exhibit a unique sort

of magnetism. They can take on a variety of forms, including rods, spheres, cubes, self-oriented flowers, etc. Fe is naturally abundant mostly in the form of $Fe³⁺$, whereas plants and other living things can only take up Fe^{2+} (Singh et al., [2021](#page-24-11)). Fe mediates a few physiological responses in plants, including leghemoglobin generation in nodules, chlorophyll synthesis, redox reaction, respiration, etc. However, Fe must be applied in optimum amounts, as both defciency and excess are harmful to plants. Fe defciency is a common problem in various crops. According to Sánchez-Alcalá et al., *Arachis hypogaea* (peanut) is extremely susceptible to Fe defciency. As demonstrated by the peanut, soybean, and wheat crops, studies have shown that iron oxide nanoparticles have a good impact on plant development and productivity (Sánchez-Alcalá et al., [2014\)](#page-23-19).

5.5 Copper Nanoparticles (CuNPs)

Due to characteristics such as their extremely small size and high surface area to volume ratio as compared to materials formed from bigger particles, CuNPs perform better than bulk copper particles. CuNPs have several uses in agriculture due to their antifungal and antibacterial actions against Gram-positive and Gramnegative bacteria as well as harmful fungi. CuNPs show antifungal effcacy against plant pathogenic fungi such *Phytophthora infestans*, *Fusarium oxysporum*, *Fusarium culmorum*, and *Fusarium graminearum*. At doses under 100 ppm, they have also been shown to behave as germination promoters and growth stimulants in several plants. CuNPs have been synthesized thus far using a variety of chemical, physical, and green synthesis techniques in diverse quantities, confgurations, and morphologies (Bahrulolum et al., [2021](#page-20-17)).

5.6 Selenium Nanoparticles (SeNPs)

Most living things require selenium, which is present in soil, water, seeds, animals, and food. Se fertilizers must be added to the soil to enhance the Se content in plant nutrients, and Se levels in food must be balanced, as SeNPs boost the plant's capacity to suppress infections and activate antifungal characteristics. Se-balanced food processing is a quick procedure that aids in resolving the Se imbalance problem in agriculture. It is crucial to ensure the optimum amount of selenium in the soil, and fertilizers made of pure selenium compounds are employed to achieve this. However, Se fertilizers only last for one or a few harvests in rich topsoil, and, over a short time, inorganic Se compounds are washed away by rain into the infertile horizons beneath the soil. Organic Se compounds are not actively leached, but they are readily broken down after application. SeNPs have the beneft of not slowly leaching from the soil and not dissolving in water or aqueous solutions, making them excellent nanofertilizers (Bahrulolum et al., [2021\)](#page-20-17).

6 Other Uses

In addition to agriculture, nanoparticles are employed in plasmonics, optoelectronics, surface-enhanced Raman scattering (SERS), biological sensors, catalysts, sorbents, energy production, and DNA sequencing. The easy growth of fungi and their abundant production of reducing and stabilizing agents make mycosynthesis of nanomaterials an effcient method in industrial manufacturing. At the laboratory scale, new cutting-edge nanotechnology offers a superior answer to sustainable agriculture. Nanoparticle applications are still in their infancy, including those such as nanofertilizers, nanopesticides, nanofungicides, nanoherbicides, nanosensors for pathogen detection, and nanomaterials for pesticide sorption. In addition to this, nanoparticles utilized in the food packaging sector are also widely accepted. Many packaged food products have longer shelf lives owing to nanocoating and wrapping in wrappers that include nanomaterials (nanoflms) (Chhipa, [2019](#page-21-8)).

6.1 Nanopesticides

The widespread use of chemical pesticides is a major global concern since they can cause serious problems like biomagnifcation. Many organophosphate insecticides build up in the animal adipose tissue and affect the food chain. The creation and application of nanoparticles with an organic origin and pesticidal characteristics is a practical remedy for this issue (Singh et al., [2021](#page-24-11)).

With regard to fungi that cause fungal plant diseases, silver nanomaterials have shown antifungal activity against *Alternaria alternata*, *Botrytis cinerea*, *Bipolaris sorokiniana*, *Magnaporthe grisea*, *Sclerotium*, *Sclerotium cepivorum*, *Candida tropicalis*, *Candida parapsilosis*, *C. albicans*, *Colletotrichum gloeosporioides*, and *Raffaelea* sp. Similar to this, ZnO exhibits antifungal action against *Penicillium expansum*, *B. cinerea*, and *A. niger.* The cell wall is damaged by nanopesticide's interaction with fungus hyphae, which prevents conidial germination and fungal growth (Chhipa, [2019\)](#page-21-8).

In recent studies, a *Taraxacum offcinale* leaf extract has been used to produce AgNPs, which exhibited signifcant antibacterial action against two signifcant phytopathogens, *Xanthomonas axonopodis* and *Pseudomonas syringae*. A tetracyclinecontaining nanoformulation demonstrated increased antibacterial activity against phytopathogens. Additionally, it was discovered in this study that synthetic AgNPs had a stronger antibacterial impact than those sold in stores. In order to manage phytopathogens, these AgNPs may be used as a less expensive substitute for commercial pesticides (Saratale et al.*,* [2018a](#page-24-1)).

Cu-based nanoparticles (Cu/Cu₂O NPs, Cu₂O NPs, and CuO NPs) have been identifed as promising agro-fungicides and have been shown to be effective against a variety of phytopathogens, including *Fusarium* sp., *Phoma destructiva*, *Cochliobolus lunatus*, *A. alternata*, *F. oxysporum*, *Penicillium italicum*, *Penicillium*

Digitatum, and *Rhizoctonia solani*). Giannousi et al. ([2013\)](#page-21-18) reported the use of Cu2O NPs as agro-fungicides to control the important plant pathogen *Phytophthora infestans*. It is interesting to note that CuNPs cause oxidative stress, which can function differently depending on the species of fungus.

6.2 Nanosensors

In addition to the use of nanopesticides and nanofertilizers, nanosensors were created to identify plant diseases, plant hormones, soil moisture, and residual pesticides. Nanosensors are useful in providing real-time information on feld conditions and soil health. Chemical sensors based on carbon nanomaterials were created to detect pesticide residues in plants. Additionally, *Xanthomonas axonopodis*, the causative agent of the bacterial spot illness, was targeted for detection using nanoprobes. To fnd the pathogen, they combined anti-rabbit secondary antibodies with silica nanoparticles (Chhipa, [2019\)](#page-21-8).

7 Conclusions

Considering the growing population demand for food and the critical environmental issues, precision farming is an obvious solution. The application of metal nanoparticles is essential for achieving precision farming. Typical production processes of metal nanoparticles are fraught with a number of problems, including the use of hazardous solvents, the generation of toxic by-products products and the consumption of high energy. As a result, environmentally friendly production routes, such as "green synthesis," are being adopted more and more for metal nanoparticles. The green synthesis process has a variety of environmental benefts, including its effciency, cost-effectiveness, and eco-friendliness. The green synthesis process is likely to play an enormous role in achieving sustainable agriculture. Metal nanoparticles produced via green synthesis are not only eco-friendly but are also capable of being delivered to specifc sites over a long period of time. As a result, these nanoparticles are good candidates for slow-release fertilizers. Metal nanoparticles offer advantageous qualities that can be used to improve plant growth through both soil and foliar applications. High germination rates, high growth rates, etc. are only a few of the plant growth indices that are improved by the usage of metal nanoparticles. In addition to their use as fertilizers, metal nanoparticles can also be used as nanosensors, soil and water amendment tools, and other applications that can be crucial for achieving increased and sustainable agricultural output.

References

- Abdel Latef, A. A. H., Srivastava, A. K., El-sadek, M. S. A., Kordrostami, M., & Tran, L. S. P. (2018). Titanium dioxide nanoparticles improve growth and enhance tolerance of broad bean plants under saline soil conditions. *Land Degradation & Development, 29*(4), 1065–1073.
- Adelere, I. A., & Lateef, A. (2016). A novel approach to the green synthesis of metallic nanoparticles: The use of agro-wastes, enzymes, and pigments. *Nanotechnology Reviews, 5*(6), 567–587.
- Alam, H., Khatoon, N., Khan, M. A., Husain, S. A., Saravanan, M., & Sardar, M. (2020). Synthesis of selenium nanoparticles using probiotic bacteria *Lactobacillus acidophilus* and their enhanced antimicrobial activity against resistant bacteria. *Journal of Cluster Science, 31*(5), 1003–1011.
- Alsamhary, K. I. (2020). Eco-friendly synthesis of silver nanoparticles by Bacillus subtilis and their antibacterial activity. *Saudi Journal of Biological Sciences, 27*(8), 2185–2191.
- Anand, B. G., Thomas, C. N., Prakash, S., & Kumar, C. S. (2015). Biosynthesis of silver nanoparticles by marine sediment fungi for a dose dependent cytotoxicity against HEp2 cell lines. *Biocatalysis and Agricultural Biotechnology, 4*(2), 150–157.
- Aranaz, I., Harris, R., & Heras, A. (2010). Chitosan amphiphilic derivatives. Chemistry and applications. *Current Organic Chemistry, 14*(3), 308–330.
- Babu, M. G., & Gunasekaran, P. (2009). Production and structural characterization of crystalline silver nanoparticles from Bacillus cereus isolate. *Colloids and Surfaces B: Biointerfaces, 74*(1), 191–195.
- Bahrulolum, H., Nooraei, S., Javanshir, N., Tarrahimofrad, H., Mirbagheri, V. S., Easton, A. J., & Ahmadian, G. (2021). Green synthesis of metal nanoparticles using microorganisms and their application in the agrifood sector. *Journal of Nanobiotechnology, 19*(1), 1–26.
- Baig, N., Kammakakam, I., & Falath, W. (2021). Nanomaterials: A review of synthesis methods, properties, recent progress, and challenges. *Materials Advances, 2*(6), 1821–1871.
- Balaji, D. S., Basavaraja, S., Deshpande, R., Mahesh, D. B., Prabhakar, B. K., & Venkataraman, A. (2009). Extracellular biosynthesis of functionalized silver nanoparticles by strains of *Cladosporium cladosporioides* fungus. *Colloids and Surfaces B: Biointerfaces, 68*(1), 88–92.
- Bansal, V., Rautaray, D., Ahmad, A., & Sastry, M. (2004). Biosynthesis of zirconia nanoparticles using the fungus *Fusarium oxysporum*. *Journal of Materials Chemistry, 14*(22), 3303–3305.
- Bansal, V., Rautaray, D., Bharde, A., Ahire, K., Sanyal, A., Ahmad, A., & Sastry, M. (2005). Fungus-mediated biosynthesis of silica and titania particles. *Journal of Materials Chemistry, 15*(26), 2583–2589.
- Bao, Y., He, J., Song, K., Guo, J., Zhou, X., & Liu, S. (2021). Plant-extract-mediated synthesis of metal nanoparticles. *Journal of Chemistry, 2021*, 1–14.
- Basavaraja, S., Balaji, S. D., Lagashetty, A., Rajasab, A. H., & Venkataraman, A. (2008). Extracellular biosynthesis of silver nanoparticles using the fungus Fusarium semitectum. *Materials Research Bulletin, 43*(5), 1164–1170.
- Bhainsa, K. C., & D'souza, S. F. (2006). Extracellular biosynthesis of silver nanoparticles using the fungus Aspergillus fumigatus. *Colloids and Surfaces B: Biointerfaces, 47*(2), 160–164.
- Birla, S. S., Tiwari, V. V., Gade, A. K., Ingle, A. P., Yadav, A. P., & Rai, M. K. (2009). Fabrication of silver nanoparticles by *Phoma glomerata* and its combined effect against *Escherichia coli, Pseudomonas aeruginosa* and *Staphylococcus aureus*. *Letters in Applied Microbiology, 48*(2), 173–179.
- Bollag, J. M., Myers, C. J., & Minard, R. D. (1992). Biological and chemical interactions of pesticides with soil organic matter. *Science of the Total Environment, 123*, 205–217.
- Che, L., Dong, Y., Wu, M., Zhao, Y., Liu, L., & Zhou, H. (2017). Characterization of selenite reduction by *Lysinibacillus* sp. ZYM-1 and photocatalytic performance of biogenic selenium nanospheres. *ACS Sustainable Chemistry & Engineering, 5*(3), 2535–2543.
- Chhipa, H. (2017). Nanofertilizers and nanopesticides for agriculture. *Environmental Chemistry Letters, 15*(1), 15–22.
- Chhipa, H. (2019). Mycosynthesis of nanoparticles for smart agricultural practice: A green and eco-friendly approach. In *Green synthesis, characterization and applications of nanoparticles* (pp. 87–109). Elsevier.
- Dikshit, P. K., Kumar, J., Das, A. K., Sadhu, S., Sharma, S., Singh, S., et al. (2021). Green synthesis of metallic nanoparticles: Applications and limitations. *Catalysts, 11*(8), 902.
- Duhan, J. S., Kumar, R., Kumar, N., Kaur, P., Nehra, K., & Duhan, S. (2017). Nanotechnology: The new perspective in precision agriculture. *Biotechnology Reports, 15*, 11–23.
- El-Saadony, M. T., ALmoshadak, A. S., Shaf, M. E., Albaqami, N. M., Saad, A. M., El-Tahan, A. M., et al. (2021). Vital roles of sustainable nano-fertilizers in improving plant quality and quantity-an updated review. *Saudi Journal of Biological Sciences, 28*(12), 7349–7359.
- Fang, Z., Qiu, X., Chen, J., & Qiu, X. (2011). Degradation of the polybrominated diphenyl ethers by nanoscale zero-valent metallic particles prepared from steel pickling waste liquor. *Desalination, 267*(1), 34–41.
- Food and Agriculture Organization of the United Nations (FAO). (2009). *Global agriculture towards 2050*. FAO, [https://www.fao.org/fleadmin/templates/wsfs/docs/Issues_papers/](https://www.fao.org/fileadmin/templates/wsfs/docs/Issues_papers/HLEF2050_Global_Agriculture.pdf) [HLEF2050_Global_Agriculture.pdf](https://www.fao.org/fileadmin/templates/wsfs/docs/Issues_papers/HLEF2050_Global_Agriculture.pdf) (accessed November 25, 2022)
- Gade, A. K., Bonde, P. P., Ingle, A. P., Marcato, P. D., Duran, N., & Rai, M. K. (2008). Exploitation of *Aspergillus niger* for synthesis of silver nanoparticles. *Journal of Biobased Materials and Bioenergy, 2*(3), 243–247.
- Gahlawat, G., & Choudhury, A. R. (2019). A review on the biosynthesis of metal and metal salt nanoparticles by microbes. *RSC Advances, 9*(23), 12944–12967.
- Ghorbani, H. R. (2017). Biosynthesis of nanosilver particles using extract of Salmonella typhirium. *Arabian Journal of Chemistry, 10*, S1699–S1702.
- Giannousi, K., Avramidis, I., & Dendrinou-Samara, C. (2013). Synthesis, characterization and evaluation of copper-based nanoparticles as agrochemicals against Phytophthora infestans. *RSC Advances, 3*(44), 21743–21752.
- Goswami, P., & Mathur, J. (2019). Positive and negative effects of nanoparticles on plants and their applications in agriculture. *Plant Science Today, 6*(2), 232–242.
- Gurunathan, S., Kalishwaralal, K., Vaidyanathan, R., Venkataraman, D., Pandian, S. R. K., Muniyandi, J., et al. (2009). Biosynthesis, purifcation and characterization of silver nanoparticles using Escherichia coli. *Colloids and Surfaces B: Biointerfaces, 74*(1), 328–335.
- Ingle, A., Rai, M., Gade, A., & Bawaskar, M. (2009). Fusarium solani: A novel biological agent for the extracellular synthesis of silver nanoparticles. *Journal of Nanoparticle Research, 11*(8), 2079–2085.
- Jamkhande, P. G., Ghule, N. W., Bamer, A. H., & Kalaskar, M. G. (2019). Metal nanoparticles synthesis: An overview on methods of preparation, advantages and disadvantages, and applications. *Journal of Drug Delivery Science and Technology, 53*, 101174.
- Jena, J., Pradhan, N., Nayak, R. R., Dash, B. P., Sukla, L. B., Panda, P. K., & Mishra, B. K. (2014). Microalga Scenedesmus sp.: A potential low-cost green machine for silver nanoparticle synthesis. *Journal of Microbiology and Biotechnology, 24*(4), 522–533.
- Jha, A. K., Prasad, K., & Prasad, K. (2009). A green low-cost biosynthesis of Sb2O3 nanoparticles. *Biochemical Engineering Journal, 43*(3), 303–306.
- Kalimuthu, K., Babu, R. S., Venkataraman, D., Bilal, M., & Gurunathan, S. (2008). Biosynthesis of silver nanocrystals by Bacillus licheniformis. *Colloids and Surfaces B: Biointerfaces, 65*(1), 150–153.
- Kalishwaralal, K., Deepak, V., Ramkumarpandian, S., Nellaiah, H., & Sangiliyandi, G. (2008). Extracellular biosynthesis of silver nanoparticles by the culture supernatant of *Bacillus licheniformis*. *Materials Letters, 62*(29), 4411–4413.
- Kathiraven, T., Sundaramanickam, A., Shanmugam, N., & Balasubramanian, T. (2015). Green synthesis of silver nanoparticles using marine algae Caulerpa racemosa and their antibacterial activity against some human pathogens. *Applied Nanoscience, 5*(4), 499–504.
- Kathiresan, K., Manivannan, S., Nabeel, M. A., & Dhivya, B. (2009). Studies on silver nanoparticles synthesized by a marine fungus, *Penicillium fellutanum* isolated from coastal mangrove sediment. *Colloids and Surfaces B: Biointerfaces, 71*(1), 133–137.
- Kim, D. Y., Saratale, R. G., Shinde, S., Syed, A., Ameen, F., & Ghodake, G. (2018). Green synthesis of silver nanoparticles using Laminaria japonica extract: Characterization and seedling growth assessment. *Journal of Cleaner Production, 172*, 2910–2918.
- Klaus, T., Joerger, R., Olsson, E., & Granqvist, C. G. (1999). Silver-based crystalline nanoparticles, microbially fabricated. *Proceedings of the National Academy of Sciences, 96*(24), 13611–13614.
- Kora, A. J., Beedu, S. R., & Jayaraman, A. (2012). Size-controlled green synthesis of silver nanoparticles mediated by gum ghatti (Anogeissus latifolia) and its biological activity. *Organic and Medicinal Chemistry Letters, 2*(1), 1–10.
- Kowshik, M., Ashtaputre, S., Kharrazi, S., Vogel, W., Urban, J., Kulkarni, S. K., & Paknikar, K. M. (2002a). Extracellular synthesis of silver nanoparticles by a silver-tolerant yeast strain MKY3. *Nanotechnology, 14*(1), 95.
- Kowshik, M., Deshmukh, N., Vogel, W., Urban, J., Kulkarni, S. K., & Paknikar, K. M. (2002b). Microbial synthesis of semiconductor CdS nanoparticles, their characterization, and their use in the fabrication of an ideal diode. *Biotechnology and Bioengineering, 78*(5), 583–588.
- Kumar, P., Senthamil Selvi, S., Lakshmi Prabha, A., Prem Kumar, K., Ganeshkumar, R. S., & Govindaraju, M. (2012). Synthesis of silver nanoparticles from Sargassum tenerrimum and screening phytochemicals for its antibacterial activity. *Nano Biomedicine and Engineering, 4*(1), 12–16.
- Kumari, S., Tehri, N., Gahlaut, A., & Hooda, V. (2020). Actinomycetes mediated synthesis, characterization, and applications of metallic nanoparticles. *Inorganic and Nano-Metal Chemistry, 51*(10), 1386–1395.
- Labrenz, M., Druschel, G. K., Thomsen-Ebert, T., Gilbert, B., Welch, S. A., Kemner, K. M., et al. (2000). Formation of sphalerite (ZnS) deposits in natural bioflms of sulfate-reducing bacteria. *Science, 290*(5497), 1744–1747.
- Lee, H., Purdon, A. M., Chu, V., & Westervelt, R. M. (2004). Controlled assembly of magnetic nanoparticles from magnetotactic bacteria using microelectromagnets arrays. *Nano Letters, 4*(5), 995–998.
- Li, G., He, D., Qian, Y., Guan, B., Gao, S., Cui, Y., et al. (2011). Fungus-mediated green synthesis of silver nanoparticles using *Aspergillus terreus*. *International Journal of Molecular Sciences, 13*(1), 466–476.
- Li, X., Wang, C., Zeng, Y., Li, P., Xie, T., & Zhang, Y. (2016). Bacteria-assisted preparation of nano α-Fe2O3 red pigment powders from waste ferrous sulfate. *Journal of Hazardous Materials, 317*, 563–569.
- Madhiyazhagan, P., Murugan, K., Kumar, A. N., Nataraj, T., Dinesh, D., Panneerselvam, C., et al. (2015). *Sargassum muticum*-synthesized silver nanoparticles: An effective control tool against mosquito vectors and bacterial pathogens. *Parasitology Research, 114*(11), 4305–4317.
- Mann, S., Frankel, R. B., & Blakemore, R. P. (1984). Structure, morphology and crystal growth of bacterial magnetite. *Nature, 310*(5976), 405–407.
- Mohammadian, A. (2007). Fusarium oxysporum mediates photogeneration of silver nanoparticles. *Scientia Iranica, 14*(4).
- Momeni, S., & Nabipour, I. (2015). A simple green synthesis of palladium nanoparticles with Sargassum alga and their electrocatalytic activities towards hydrogen peroxide. *Applied Biochemistry and Biotechnology, 176*(7), 1937–1949.
- Monowar, T., Rahman, M. S., Bhore, S. J., Raju, G., & Sathasivam, K. V. (2018). Silver nanoparticles synthesized by using the endophytic bacterium *Pantoea ananatis* are promising antimicrobial agents against multidrug resistant bacteria. *Molecules, 23*(12), 3220.
- Murugan, K., Samidoss, C. M., Panneerselvam, C., Higuchi, A., Roni, M., Suresh, U., et al. (2015). Seaweed-synthesized silver nanoparticles: An eco-friendly tool in the fght against Plasmodium falciparum and its vector Anopheles stephensi? *Parasitology Research, 114*(11), 4087–4097.
- Narayanan, K. B., & Sakthivel, N. (2010). Biological synthesis of metal nanoparticles by microbes. *Advances in Colloid and Interface Science, 156*(1–2), 1–13.
- Ndaba, B., Roopnarain, A., Haripriya, R. A. M. A., & Maaza, M. (2022). Biosynthesized metallic nanoparticles as fertilizers: An emerging precision agriculture strategy. *Journal of Integrative Agriculture, 21*(5), 1225–1242.
- Nuraje, N., Lei, Y., & Belcher, A. (2014). Virus-templated visible spectrum active perovskite photocatalyst. *Catalysis Communications, 44*, 68–72.
- Parikh, R. Y., Singh, S., Prasad, B. L. V., Patole, M. S., Sastry, M., & Shouche, Y. S. (2008). Extracellular synthesis of crystalline silver nanoparticles and molecular evidence of silver resistance from *Morganella* sp.: Towards understanding biochemical synthesis mechanism. *Chembiochem, 9*(9), 1415–1422.
- Philip, D. (2009). Biosynthesis of Au, Ag and Au–Ag nanoparticles using edible mushroom extract. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy, 73*(2), 374–381.
- Prasad, K., & Jha, A. K. (2010). Biosynthesis of CdS nanoparticles: An improved green and rapid procedure. *Journal of Colloid and Interface Science, 342*(1), 68–72.
- Prasad, K., Jha, A. K., & Kulkarni, A. R. (2007). Lactobacillus assisted synthesis of titanium nanoparticles. *Nanoscale Research Letters, 2*(5), 248–250.
- Prasad, T. N., Kambala, V. S. R., & Naidu, R. (2013). Phyconanotechnology: Synthesis of silver nanoparticles using brown marine algae *Cystophora moniliformis* and their characterisation. *Journal of Applied Phycology, 25*(1), 177–182.
- Pugazhendhi, A., Prabakar, D., Jacob, J. M., Karuppusamy, I., & Saratale, R. G. (2018). Synthesis and characterization of silver nanoparticles using *Gelidium amansii* and its antimicrobial property against various pathogenic bacteria. *Microbial Pathogenesis, 114*, 41–45.
- Rai, M., & Ingle, A. (2012). Role of nanotechnology in agriculture with special reference to management of insect pests. *Applied Microbiology and Biotechnology, 94*(2), 287–293.
- Rajput, N. (2015). Methods of preparation of nanoparticles-a review. *International Journal of Advances in Engineering & Technology, 7*(6), 1806.
- Raliya, R., Saharan, V., Dimkpa, C., & Biswas, P. (2017). Nanofertilizer for precision and sustainable agriculture: Current state and future perspectives. *Journal of Agricultural and Food Chemistry, 66*(26), 6487–6503.
- Rodrigues, S. M., Demokritou, P., Dokoozlian, N., Hendren, C. O., Karn, B., Mauter, M. S., et al. (2017). Nanotechnology for sustainable food production: Promising opportunities and scientifc challenges. *Environmental Science: Nano, 4*(4), 767–781.
- Roychoudhury, P., & Pal, R. (2014). Synthesis and characterization of nanosilver—A blue green approach. *Indian Journal of Applied Research, 4*(1), 69–72.
- Salari, Z., Danafar, F., Dabaghi, S., & Ataei, S. A. (2016). Sustainable synthesis of silver nanoparticles using macroalgae *Spirogyra varians* and analysis of their antibacterial activity. *Journal of Saudi Chemical Society, 20*(4), 459–464.
- Saleem, H., & Zaidi, S. J. (2020). Recent developments in the application of nanomaterials in agroecosystems. *Nanomaterials, 10*(12), 2411.
- Samaddar, P., Ok, Y. S., Kim, K. H., Kwon, E. E., & Tsang, D. C. (2018). Synthesis of nanomaterials from various wastes and their new age applications. *Journal of Cleaner Production, 197*, 1190–1209.
- Sánchez-Alcalá, I., del Campillo, M. D. C., Barrón, V., & Torrent, J. (2014). Evaluation of prefooding effects on iron extractability and phytoavailability in highly calcareous soil in containers. *Journal of Plant Nutrition and Soil Science, 177*(2), 150–158.
- Sangeetha, J., Thangadurai, D., Hospet, R., Harish, E. R., Purushotham, P., Mujeeb, M. A., et al. (2017). Nanoagrotechnology for soil quality, crop performance and environmental management. In *Nanotechnology* (pp. 73–97). Springer.
- Sanghi, R., & Verma, P. (2009). Biomimetic synthesis and characterisation of protein capped silver nanoparticles. *Bioresource Technology, 100*(1), 501–504.
- Saratale, R. G., Kuppam, C., Mudhoo, A., Saratale, G. D., Periyasamy, S., Zhen, G., et al. (2017). Bioelectrochemical systems using microalgae–A concise research update. *Chemosphere, 177*, 35–43.
- Saratale, R. G., Karuppusamy, I., Saratale, G. D., Pugazhendhi, A., Kumar, G., Park, Y., et al. (2018a). A comprehensive review on green nanomaterials using biological systems: Recent perception and their future applications. *Colloids and Surfaces B: Biointerfaces, 170*, 20–35.
- Saratale, R. G., Saratale, G. D., Shin, H. S., Jacob, J. M., Pugazhendhi, A., Bhaisare, M., & Kumar, G. (2018b). New insights on the green synthesis of metallic nanoparticles using plant and waste biomaterials: Current knowledge, their agricultural and environmental applications. *Environmental Science and Pollution Research, 25*(11), 10164–10183.
- Sarkar, M. R., Rashid, M. H. O., Rahman, A., Kaf, M. A., Hosen, M. I., Rahman, M. S., & Khan, M. N. (2022). Recent advances in nanomaterials based sustainable agriculture: An overview. *Environmental Nanotechnology, Monitoring & Management, 18*, 100687.
- Senapati, S., Mandal, D., Ahmad, A., Khan, M. I., Sastry, M., & Kumar, R. (2004). Fungus mediated synthesis of silver nanoparticles: A novel biological approach. *Indian Journal of Physics, 78*, 101–105.
- Shaligram, N. S., Bule, M., Bhambure, R., Singhal, R. S., Singh, S. K., Szakacs, G., & Pandey, A. (2009). Biosynthesis of silver nanoparticles using aqueous extract from the compactin producing fungal strain. *Process Biochemistry, 44*(8), 939–943.
- Shanthi, S., Jayaseelan, B. D., Velusamy, P., Vijayakumar, S., Chih, C. T., & Vaseeharan, B. (2016). Biosynthesis of silver nanoparticles using a probiotic Bacillus licheniformis Dahb1 and their antibioflm activity and toxicity effects in *Ceriodaphnia cornuta*. *Microbial Pathogenesis, 93*, 70–77.
- Shedbalkar, U., Singh, R., Wadhwani, S., Gaidhani, S., & Chopade, B. A. (2014). Microbial synthesis of gold nanoparticles: Current status and future prospects. *Advances in Colloid and Interface Science, 209*, 40–48.
- Singh, R. P., Handa, R., & Manchanda, G. (2021). Nanoparticles in sustainable agriculture: An emerging opportunity. *Journal of Controlled Release, 329*, 1234–1248.
- Sinha, S. N., Paul, D., Halder, N., Sengupta, D., & Patra, S. K. (2015). Green synthesis of silver nanoparticles using fresh water green alga Pithophora oedogonia (Mont.) Wittrock and evaluation of their antibacterial activity. *Applied Nanoscience, 5*(6), 703–709.
- Sneha, K., Sathishkumar, M., Mao, J., Kwak, I. S., & Yun, Y. S. (2010). Corynebacterium glutamicum-mediated crystallization of silver ions through sorption and reduction processes. *Chemical Engineering Journal, 162*(3), 989–996.
- Snehal, S., & Lohani, P. (2018). Silica nanoparticles: Its green synthesis and importance in agriculture. *Journal of Pharmacognosy and Phytochemistry, 7*(5), 3383–3393.
- Suresh, A. K., Pelletier, D. A., Wang, W., Broich, M. L., Moon, J. W., Gu, B., et al. (2011). Biofabrication of discrete spherical gold nanoparticles using the metal-reducing bacterium *Shewanella oneidensis*. *Acta Biomaterialia, 7*(5), 2148–2152.
- Thomas, R., Janardhanan, A., Varghese, R. T., Soniya, E. V., Mathew, J., & Radhakrishnan, E. K. (2014). Antibacterial properties of silver nanoparticles synthesized by marine Ochrobactrum sp. *Brazilian Journal of Microbiology, 45*, 1221–1227.
- Vahabi, K., & Dorcheh, S. K. (2014). Biosynthesis of silver nano-particles by Trichoderma and its medical applications. In *Biotechnology and biology of Trichoderma* (pp. 393–404). Elsevier.
- Vardhana, J., & Kathiravan, G. (2015). Biosynthesis of silver nanoparticles by endophytic fungi *Pestaloptiopsis pauciseta* isolated from the leaves of *Psidium guajava* Linn. *International Journal of Pharmaceutical Sciences Review and Research, 31*(1), 29–31.
- Velmurugan, P., Iydroose, M., Mohideen, M. H. A. K., Mohan, T. S., Cho, M., & Oh, B. T. (2014). Biosynthesis of silver nanoparticles using *Bacillus subtilis* EWP-46 cell-free extract and evaluation of its antibacterial activity. *Bioprocess and Biosystems Engineering, 37*(8), 1527–1534.
- Venkatpurwar, V., & Pokharkar, V. (2011). Green synthesis of silver nanoparticles using marine polysaccharide: Study of in-vitro antibacterial activity. *Materials Letters, 65*(6), 999–1002.
- Vigneshwaran, N., Kathe, A. A., Varadarajan, P. V., Nachane, R. P., & Balasubramanya, R. H. (2006). Biomimetics of silver nanoparticles by white rot fungus, *Phaenerochaete chrysosporium*. *Colloids and Surfaces B: Biointerfaces, 53*(1), 55–59.
- Vijayakumar, M. D., Surendhar, G. J., Natrayan, L., Patil, P. P., Ram, P. M., & Paramasivam, P. (2022). Evolution and recent scenario of nanotechnology in agriculture and food industries. *Journal of Nanomaterials, 2022*, 1280411.
- Wypij, M., Golinska, P., Dahm, H., & Rai, M. (2017). Actinobacterial-mediated synthesis of silver nanoparticles and their activity against pathogenic bacteria. *IET Nanobiotechnology, 11*(3), 336–342.
- Zhou, W., He, W., Zhong, S., Wang, Y., Zhao, H., Li, Z., & Yan, S. (2009). Biosynthesis and magnetic properties of mesoporous Fe3O4 composites. *Journal of Magnetism and Magnetic Materials, 321*(8), 1025–1028.
- Zulfqar, F., Navarro, M., Ashraf, M., Akram, N. A., & Munné-Bosch, S. (2019). Nanofertilizer use for sustainable agriculture: Advantages and limitations. *Plant Science, 289*, 110270.