

Chapter 3

Physiological Significance of Oxidative Stress and Anti-oxidative System



Saddam Hussain, Azhar Rasul, Ghulam Hussain, Majeeda Rasheed, Maria Manan, Komal Riaz, Saba Riaz, Muhammad Asif Khalil, Ayesha Sadiqa, and Sevki Adem

3.1 Introduction

Two valence electrons have parallel spins in each of their two anti-bonding orbitals in molecular oxygen. This spin restriction allows it to accept a pair of electrons from a donor. A redox reaction is a fundamental metabolic activity in living organisms [1]. The movement of a single electron may result in the formation of free radicals and other issues [2]. Free radicals generally show a high level of reactivity. These radicals are extremely unstable and reactive with other chemicals. Guyton de Morveau coined the term “radical” in 1786, and later, Gay-Lussac, Berzelius, and Liebig used it to refer to unaltered atomic groups in numerous substances [3].

S. Hussain · A. Rasul (✉) · K. Riaz · S. Riaz · M. A. Khalil · A. Sadiqa
Department of Zoology, Faculty of Life Sciences, Government College University,
Faisalabad, Pakistan
e-mail: azharrasul@gcuf.edu.pk

G. Hussain
Department of Physiology, Government College University Faisalabad, Faisalabad, Pakistan

M. Rasheed
Department of Life Sciences, Khwaja Fareed University of Engineering and Information
Technology, Rahim Yar Khan, Pakistan

M. Manan
Department of Pharmacology, Faculty of Pharmaceutical Sciences, Government College
University, Faisalabad, Pakistan

S. Adem
Department of Biochemistry, Çankırı Karatekin Üniversitesi, Çankırı, Turkey

Free radicals not only take part in pathogenic processes but are also essential for many physiological activities of living organisms, such as healthy aging [4]. Lipid peroxidation was reported to have both negative and positive consequences [5]. Free radicals can cause numerous diseases in humans by damaging lipids, proteins, and DNA. ROS and RNS are responsible for cellular damage by substituting macromolecules [6]. There are numerous antioxidants, both natural and artificial. Endogenous antioxidants are characterized as enzymatic or non-enzymatic [7].

3.2 Roots of Oxidative Stress

An imbalance in the production of reactive oxygen species results in the oxidative stress and capacity of an organism's antioxidative defense mechanisms to lessen the harm due to oxidants. As a byproduct of normal aerobic metabolism, ROS may provide a fundamental health concern when the amount increases in response to stress [8]. The mitochondrion is a primary organelle which is taking part in the production of ROS. ATP is produced by numerous processes including the electron transport chain. Only one or two electron of oxygen are reduced instead of four electrons during this process, which is responsible for the formation of O_2 or H_2O_2 , which then changes into other ROS [9]. Free radicals may be created by both internal and external processes. Infection, inflammation, ischemia, immune cell activation, cancer, mental stress, and aging contribute to endogenous free radical formation [10].

Numerous studies show that excessive macronutrient intakes might increase oxidative stress. An excessive amount of high caloric intake will increase the number of substrates entering mitochondrial respiration. As a consequence, the number of contributed electrons to the electron transport chain will be surged [11]. When superoxide concentrations cross a certain point, extra electrons may gather at complex III and donate more electrons to molecular oxygen [12].

ROS generation is fundamentally dependent on enzymatic and non-enzymatic processes. Superoxide radical is produced by xanthine oxidase, peroxidases, and NADPH oxidase [10]. The sole class of enzymes with the specific purpose of producing ROS is the NADPH oxidases, which differentiates it from other enzymes producing ROS as the byproduct of their activity [13]. Free radicals may also be created by non-enzymatic processes like oxygen's interactions with organic materials or the radiation that is exposed to cells. Non-enzymatic free radicals production may also take place during mitochondrial respiration [14].

3.2.1 Endogenous Sources of ROS Production

Different cellular organelles with high oxygen consumption rates, including the endoplasmic reticulum, mitochondria, and peroxisomes are examples of endogenous generators of ROS.

3.2.1.1 Production of Oxidative Stress in Mitochondria

Mitochondria generate the majority of the intracellular ROS. Oxidizing radicals are generated at complex-I and complex-III in oxidative phosphorylation [15]. Alongside cytochrome c oxidase, monoamine oxidase, glycerol phosphate dehydrogenase, α -ketoglutarate dehydrogenase, and p66shc also take part in ROS generation within mitochondria [16].

3.2.1.2 Generation of Oxidative Stress in Peroxisomes

The respiratory pathway in peroxisomes involves the transport of electrons from different metabolites to O_2 , which ultimately causes the generation of hydrogen peroxides [17]. β -oxidation of fatty acids produces hydrogen peroxide in peroxisomes. OH^\cdot , H_2O_2 , and O^{2-} are also produced in peroxisome [18].

3.2.1.3 Generation of Oxidative Stress in the Endoplasmic Reticulum

Diamine oxidase, cytochrome b5, and cytochrome P-450 play role in ROS production [19]. Ero1p is a thiol oxidase that leads to the production of H_2O_2 [20]. Auto oxidation of the prostaglandin synthesis, immune cell activation, adrenaline, cytochrome P-450, phagocytic cells, flavin mononucleotide (FMNH₂), flavin adenine dinucleotide (FADH₂), inflammation, anxiety, mental stress [21], infection, excessive exercise, aging, ischemia, and cancer are other endogenous sources of ROS [19].

3.2.2 Production of Oxidative Stress by Exogenous Sources

Various synthetic products are causing oxidative stress directly or via producing by-products. Some of the major exogenous sources for the generation of oxidative stress are given below (Fig. 3.1).

I. Smoke-generated oxidative stress

Smoke from cigarettes comprises a variety of extremely unstable free radicals that increase the generation of ROS and RNS and cause oxidative stress [22]. Lung

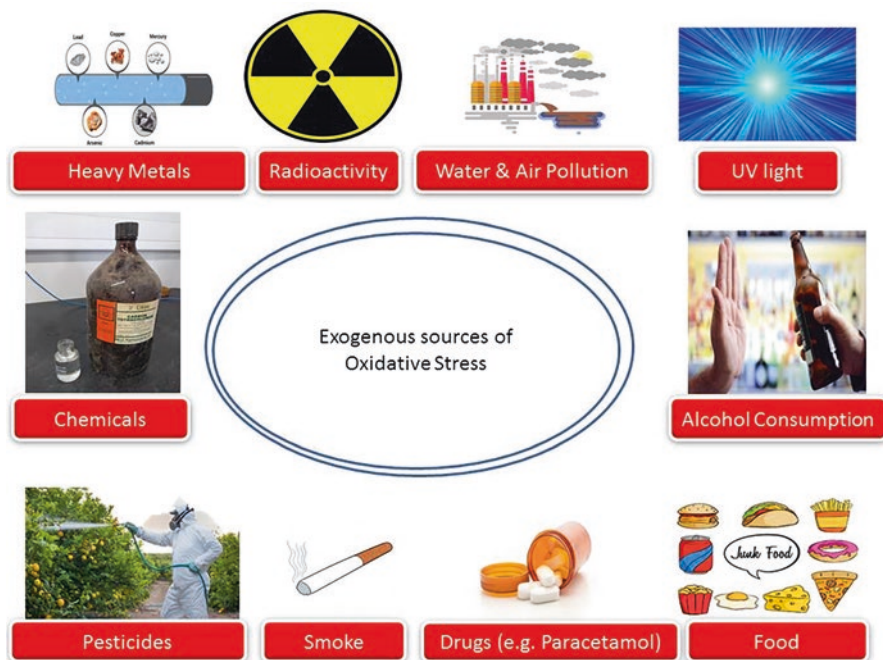


Fig. 3.1 Exogenous sources of oxidative stress

inflammatory cells (macrophages, epithelium, and neutrophils) are affected by cigarette smoke, due to the activation of NADPH oxidase 2, which produces superoxide radicals [23].

II. Ultra-violet generated oxidative stress

There are two ways that UV light might harm cellular components. The first method involves the cell and its constituent parts directly absorbing incoming light. This results in the production of an excited state of the molecules following chemical reactions. The second mechanism is photosensitization. Incoming radiation is absorbed by photosensitizers such as bilirubin. As a result, the sensitizers are excited to triple states [24].

III. Other exogenous sources of oxidative stress

Other factors like air and water pollution are involved in the production of oxidative stress in the body. Radiations and radioactivity also take part in the production of oxidative stress. Drugs like halothane, bleomycin, paracetamol, doxorubicin, and metronidazole have a record of generation of oxidants. Industrial solvents, pesticides, chemicals like carbon tetrachloride, transition metals, heavy metals, alcohol consumption, and cooking (smoked meat, fat, and junk foods) are also recorded as the sources of oxidative stress [25].

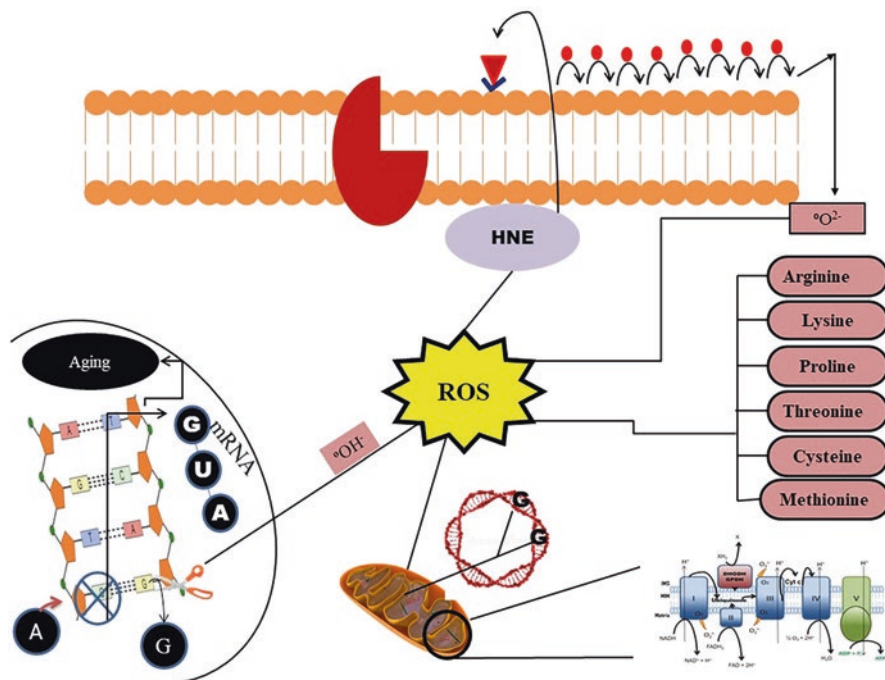


Fig. 3.2 Involvement of ROS in the pathophysiology of cell (a) ROS produces lipid peroxides in the cell membrane, inducing lipid peroxidation chain reaction or the generation of aldehydes such as 4-Hydroxy-2-nominal (HNE), that are detrimental to cellular activities via Ca^{2+} signaling and thus cause diseases such as inflammation [26], (b) Proteins are primary targets of ROS with reversible or irreversible modifications to the amino acid residues like Cys, Met, Arg and Tyr [26], (c) ROS are produced by electron transport chain in mitochondria to a large extent. ROS are generated via single electron leakage in the following situations: (i) during normal ETC function, at complex-I and complex-III; (ii) during conditions of high NADH/NAD^+ ratio and low electron transport chain activity, (iii) during conditions of a high pool of reduced ubiquinone and transmembrane H^+ gradient, at complex I and (iv) during hypoxic conditions, at complex III [27], (d) The oxidative modifications of guanine base is one of the most common forms of DNA damage. Nuclear DNA is far less susceptible to ROS than mitochondrial DNA, which contributes to age-related mitochondrial malfunction. The fact that guanine is quickly oxidized could have important physiological consequences [26].

3.3 Molecular Targets of Free Radicals

Increased generation of RNS and ROS and decreased antioxidant defense result in nitrosative and oxidative stress. Major components of cells (mitochondria, plasma membrane, and DNA molecule) are damaged as shown in Fig. 3.2, leading to multiple disorders [14].

3.4 Role of Oxidative Stress in Health Illness

Oxidative stress is related to the emergence of many acute and chronic ailments in addition to speeding up aging and generating acute illnesses. The impact of oxidative stress on hypertension, Alzheimer's disease, and some malignancies will be covered in this chapter.

3.4.1 *Oxidative Stress and Hypertension*

The intricate and prevalent cardiovascular risk factor is hypertension [28], which is responsible for morbidity and mortality worldwide [29]. Hypertension is linked with inflammatory processes but is not confirmed whether inflammation is the consequence or cause of hypertension [30]. Tissue damage and remodeling in hypertension ensure its central role in hypertension and its side effects [31].

3.4.1.1 Sources of ROS

Research on ROS sources in hypertension is extensive. These are NADPH oxidase, uncoupled eNOS, xanthine oxidases, and mitochondria [32].

3.4.1.2 Oxidative Stress as a Mediator of Hypertension

In the year of 1991, Nakazono and his colleagues described that the blood pressure of spontaneously hypertensive rats (SHR) was reduced by intravenous injection of a fusion protein composed of human Cu/ Zn SOD and COOH terminal basic peptides with enhanced attraction for heparan sulfate. This result indicated that oxidative stress could be a mediator of hypertension in SHR. Additionally, they discovered that the xanthine oxidase inhibitor oxypurinol decreased the blood pressure in male SHR, correlating hypertension in male SHR to oxidative stress [33].

After 5 years of Nakazono's findings, Rajagopalan and his colleagues reported that administering large doses of angiotensin to rats raised their blood pressure and increased vascular superoxide, which was mediated by NADPH oxidase [34]. Superoxide levels were unaffected by norepinephrine, which raised blood pressure to comparable levels. Vascular dysfunction and constriction were eliminated when researchers administered a liposome-encapsulated superoxide dismutase [35]. These researchers later demonstrated that superoxide probably degraded vascular NO to raise blood pressure [36].

Superoxide binds to the NO generated by endothelial NO synthase (eNOS), forming peroxynitrite. This decreases NO bioavailability which results in vasoconstriction. Furthermore, in the presence of ROS, the eNOS cofactor,

tetrahydrobiopterin (BH4) is converted to dihydrobiopterin leading eNOS to synthesize superoxide [37]. The instability was only partially reversed by the addition of BH4. These researchers hypothesized that peroxynitrite can inactivate eNOS by oxidizing BH4, as well as by damaging the enzyme's heme/heme core [38]. Antioxidants such as vitamins E and C; tempol, apocynin, allopurinol, N-acetylcysteine, and BH4 reduced depression according to a study performed in male animals [39].

3.4.2 Oxidative Stress and Alzheimer's Disease

Clinical symptoms of Alzheimer's disease include a gradual decline in memory and cognitive abilities and severe dementia. Over the next few decades, people with Alzheimer's disease are expected to rise upto 15 million from the present number of over 4 million [40, 41]. When hyperphosphorylated tau protein aggregates bind to Fe³⁺, neurofibrillary tangles are produced [42]. The amyloid-peptide may form a chelation complex with transition metal ions, which then catalyzes the production of H₂O₂ and the poisonous OH radical [43]. In AD patients, there is significant lipid peroxidation, which could lead to neuronal loss by a variety of pathways, gathered with impaired activity of glucose transporters, ion pumps, and glutamate transporters. Patients with AD have been found to have additional oxidative protein damage indicators like 3-nitrotyrosine and protein carbonyls [44].

3.4.3 Oxidative Stress and Cancer

Cancer ranks among the main causes of mortality in people. Free radicals alter DNA chemically in many ways, make them potentially mutagenic, and contribute to the development of cancer [45, 46]. Cancer cells exhibit increased levels of oxidative stress due to the activation of the oncogenes and loss of tumor suppressors [47]. ROS changes the gene expression and growth signals, which leads to cancer cell proliferation [48].

3.4.3.1 Colorectal Cancer (CRC)

CRC is one of the important types of cancer with 608,000 fatalities per year [49]. ROS from internal and external sources are continually exposed to the gastrointestinal system, especially the colon, and rectum [50]. Epithelial cells are sites where colon cancer begins to develop. These cells have high metabolic rate and divide quickly [51]. This exposure eventually leads to a disrupted intestinal metabolic equilibrium that results in cancer [52].

3.4.3.2 Breast Cancer

ROS damages the breast epithelium which results in hyperplasia of epithelium, breast cancer, and fibroblast proliferation [53]. Thymidine phosphorylase produces oxygen radicals in the carcinoma cell when proteins are quickly glycosylated. It can be overexpressed in a majority of breast cancer which might cause oxidative stress [54].

3.4.3.3 Prostate Cancer

Cellular growth of prostate cancer is caused by ROS production [55]. Prostate cancer first appears when the protein NADPH oxidase 1 (Nox1) is overexpressed. ROS and Nox1 levels are noticeably greater in prostate cancer [56].

3.4.3.4 Lung Cancer

Among the main global causes of cancer mortality in males, lung cancer has been increasing at a steady rate in recent decades. Approximately 30% of all cancer deaths are caused by lung cancer. Lung inflammation and cancer are two conditions that oxidative stress contributes to significantly [57]. The significant environmental risk factor for lung cancer is cigarette smoking. The particulate matter from cigarette smoke is a complicated combination of several stable ROS and carcinogens with very long half-lives [49].

3.5 Antioxidants and Classification of Antioxidants

Antioxidants may be synthetic or natural. The natural antioxidant system has two categories, enzymatic antioxidants, and non-enzymatic antioxidants as shown in Fig. 3.3 [58]. Free radicals may be stabilized or inactivated by antioxidant enzymes before they damage cellular components. Synthetic antioxidants are chemically prepared substances [58]. Natural antioxidants are further divided into two categories. They may be endogenous and exogenous antioxidants [59]. Exogenous are those antioxidants that we take through food and supplements that are high in antioxidants [60].

Examples of exogenous antioxidants include vitamins, minerals, carotenoids, beta carotene, lycopene, lutein, zeaxanthin, organic sulfur compounds, allium, allyl sulfide, indoles, uric acid, glutathione and polyphenols which are phenolic acids and flavonoids. Flavonoids may be anthocyanidins cyanidin, pelargonidin, isoflavonoids, genistein, flavonols, catechin, EGCG, flavonols quercetin kaempferol, and flavanones. Endogenous antioxidants are the primary defense system including glutathione peroxidase, superoxide dismutase, catalase, and the secondary defense system which includes glucose-6 phosphate dehydrogenase and glutathione reductase.

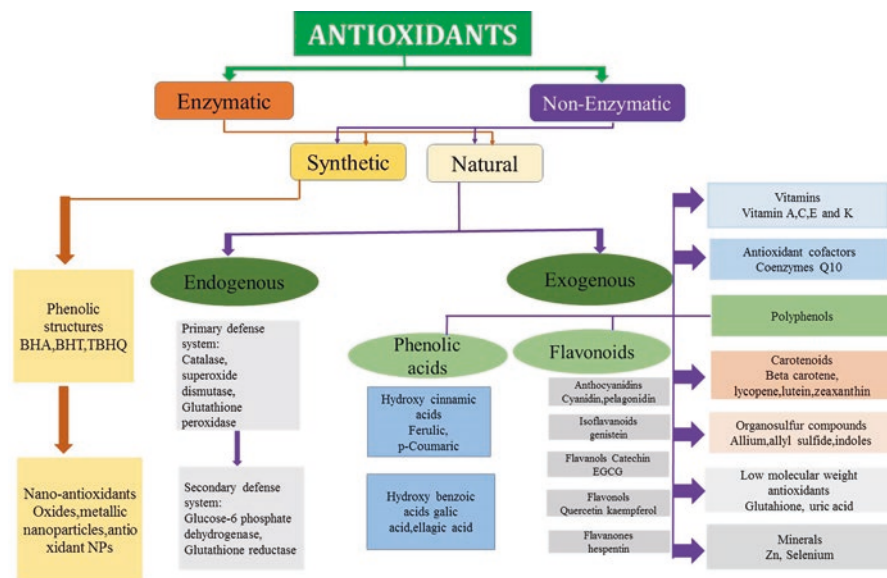


Fig. 3.3 Classification of antioxidants based on enzymatic and non-enzymatic categories

Synthetic antioxidants are also categorized as enzymatic and non-enzymatic antioxidants. They are phenolic structures, nano-antioxidants, oxides, and metallic nanoparticles [61].

3.6 Sources of Antioxidants

Antioxidants are found in natural foods and can also be synthesized. Antioxidants are mostly found in plants [62]. Phenolic structures are endogenous. A brief description of sources of antioxidants is elaborated in Fig. 3.4. We get phenolic structures from apples, grapes, pomace, pomegranate, berries, oranges, tomatoes, olive oil, coffee, and tea. Exogenous may be polyphenols, minerals, carotenoids, vitamins, and organosulfur compounds [63].

Polyphenols are found in spices, berries, nuts, herbs, cocoa powder, flaxseeds, olives, vegetables, coffee, and tea. Polyphenols may trigger apoptosis, inhibit tumor development and increase cell survival since they are prooxidants and antioxidants. However, polyphenols' biological impacts could go well beyond just reducing oxidative stress [64]. Minerals are found in meat, dairy foods, cereals, fish, nuts milk, fruits, and vegetables [65]. Other sources of antioxidants are vitamins which are found in potatoes, citrus fruits, red and green peppers, strawberries, green leafy vegetables, blueberries, blackberries, carrots, and kale [66]. Carotenoids are also the type of antioxidants that are found in spinach, yams, cantaloupe, kale, watermelon, tomatoes, bell peppers, and carrots [67]. Organosulfur compounds are found in



Fig. 3.4 Natural sources of antioxidants enriched with phenolic compounds, polyphenols, minerals, vitamins, carotenoids, and organosulphur compounds

cabbage, broccoli, cauliflower, brussels sprouts, garlic, onion, meat, eggs, and fish [68]. Additionally, there are excellent sources of certain particular antioxidants, such as the allium sulfur compounds found in garlic, onions, and leeks [69]. Anthocyanins are found in berries, grapes, and eggplant [70]. Beta carotene is found in apricots, pumpkins, carrots, mangoes, parsley, and spinach [71]. Flavonoids are found in different fruits, onions, tea, green tea, and apples [72].

3.7 Mechanism of Action of Antioxidants

Reactive intermediates are produced both endogenously and exogenously. Concerning the mechanism of antioxidants, there are five basic ways by which antioxidants work namely (1) radical-scavenging mechanisms (2) H^\bullet species donation, (3) oxidant enzyme inhibition, (4) metal chelation, and lastly (5) repair of damaged cell components [73]. Several physical, chemical, and enzymatic factors promote oxidative reactions that result in the loss of an electron from the outermost shell of a given substance [74, 75]. This series of damage is prevented when there are enough antioxidants present in the body through the five mechanisms which are illustrated in Fig. 3.5. The first one employs the free radical scavenging mechanism thus interrupting the chain reactions by inhibiting further oxidation Fig. 3.5 Part 1. The second way of the antioxidant system involves the donation of H^0 species to unstable molecules thus producing a more stable radical which does not contribute to further propagation and is stable comparatively to Fig. 3.5 part 2 [76–87]. The

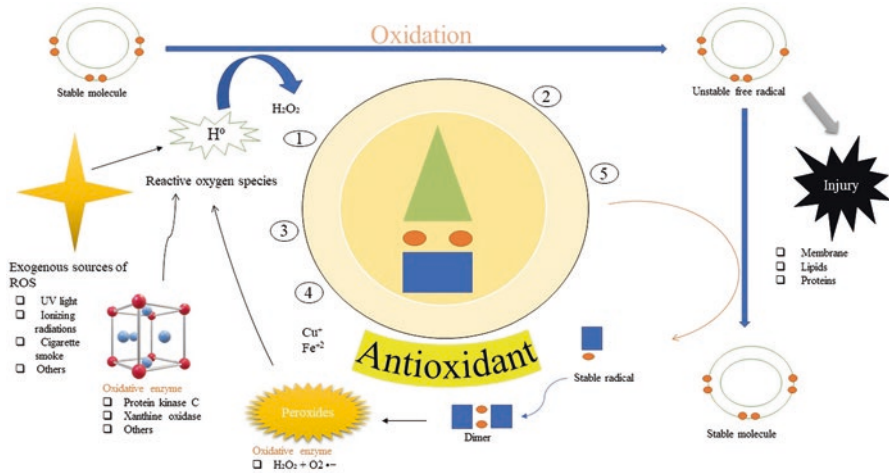


Fig. 3.5 Mechanism of action of antioxidants

third path of the antioxidant system involves the inhibition or deactivation of oxidative enzymes Fig. 3.5 part 03 [88–90]. The fourth mechanism refers to the chelation of different metals such as Fe^{2+} which results in the production of highly aggressive HO^\bullet radical which, in turn, prevents metal-induced free radical formation Fig. 3.5 part 4 [91–95]. The last mechanism, the fifth one, employs the repairing of damaged components of the cell such as proteins, membrane, lipids, and deoxyribonucleic acid (DNA) [83, 96–99]. Depending upon the structure and nature of the antioxidant agents, the said mechanisms may act alone or in association with one another [74, 100, 101].

3.8 Role of Antioxidants in the Treatment of Different Diseases

Antioxidants play a major role in the treatment of different ailments by scavenging free radicals and eliminating them from the body through different processes. Some of them are enlisted below;

3.8.1 Antioxidants and Hypertension

Hypertension is an important cardiovascular issue that contributes to almost half of the prevalent coronary heart diseases and associated disorders like chronic kidney diseases (CKD) [102–107]. In addition, hypertension ranks third among the list of six major factors which cause global diseases [108].

Antioxidant treatment appears to be an effective method for reestablishing a healthy equilibrium between oxidants and antioxidants in hypertensive patients. Antioxidants have promising potential to relieve hypertension in animal models. In spontaneously hypertensive rats (SHR), NO viability was enhanced and blood pressure was lowered after oral administration of lazaroid, the ROS scavenger medicine [109]. Similar results were seen when N-acetylcysteine (NAC), another antioxidant, was used to treat high blood pressure. NAC reduced blood pressure by preventing ROS production and increasing NOS activity [110]. A similar pattern was observed in SHR given the xanthine oxidase inhibitor allopurinol [111]. The bioavailability of nitric oxide was greatly increased, and the treatment blunted the progressive and time-dependent rise in systolic blood pressure [112].

3.8.1.1 Anti-hypertensive Drugs with Antioxidant Properties

Several molecules with anti-hypertensive and antioxidant properties have been discovered so far. Among these celiprolol, nebivolol, propranolol, and carvedilol got major focus [113]. Tissue lipid peroxidation and oxidative stress are both decreased by propranolol [113, 114]. Patients with heart failure can benefit from carvedilol's free radical scavenging properties which decrease lipid peroxidation [113, 115]. However, not all beta-blockers have these antioxidant properties; for example, atenolol has been demonstrated to possess no affect on ROS generation in lining cells [116].

3.8.2 Antioxidants and Aging

Aging is a universal, inevitable, biological phenomenon affecting almost all living organisms from multicellular to unicellular life [117–119]. When we talk about the process behind the oxidative stress associated with aging, we can't find clear data despite the presence of many different hypotheses, most probably elevated levels of RONS, a process that inhibits the proliferation that results due to damage during replication [120].

Several antioxidants are available which have anti-aging properties such as retinoids [121–123], vitamin C [124–129], tea extracts [130–132], grapes seed extracts [133], peptides, and hydroxy acids have anti-aging character. The interesting thing is that almost all of these are antioxidants [134–144].

3.8.3 Antioxidants and Cancer

Antioxidants have the ability to avoid harmful and sometimes carcinogenic effects. Mice that have been exposed to carcinogens or have lost tumor suppressor genes got benefit from many isoforms of glutathione S-transferases (GSTs) which work

together to keep the liver, skin, and colon cancer-free [145–147]. Glutathione Peroxidases (GPXs) can also protect against carcinogen and ROS-induced malignancies initiation in a variety of animals. In colon cancer mouse models, GPX3 inhibits tumor initiation [32]. Similarly, animals with reduced SOD2 expression, either alone or in combination with GPX1 loss, exhibited higher DNA damage and tumor incidence [148, 149].

Catechins, especially epigallocatechin-3-gallate (EGCG), are abundant in green tea (*Camellia sinensis*). Animal studies on carcinogenesis have revealed that EGCG and green tea can reduce tumor growth. Polyphenols found in tea are potent radical scavengers due to the presence of dihydroxy and trihydroxy groups. NRF2-antioxidant response element-dependent upregulation of glutamate cysteine ligase, glutamyl transferase, and heme oxygenase-1 gene expression in EGCG-treated mice [150]. Berberine has been shown to suppress the growth of a wide variety of cancers by binding to oligonucleotides, stabilizing DNA triplexes or G-quadruplexes, and blocking the enzymes telomerase and topoisomerase. Berberine can scavenge reactive oxygen species (ROS), inhibit lipid peroxidation, and decrease metal ion concentrations associated with lipid peroxidation [151].

3.9 Conclusion and Future Perspectives

Oxidative stress arises when the balance between the rate at which oxygen-reactive species are produced and accumulated in cells and tissues and the rate at which the body can eliminate them is disturbed. Mainly ROS is generated as a byproduct of normal cellular reactions. ROS production that is necessarily produced at a limited level is easily diminished, but certain chemicals, drugs, and other sources become responsible for high ROS production. Oxidative stress has a vital role in different diseases including cancer. Antioxidants are substances that counteract oxidative stress. Although a lot of research work regarding the mechanism of product and action of ROS was discovered, more investigations should be done to find out a link between disease and ROS level, food and antioxidant production, and the role of ROS in normal cellular activities. There should be educational seminars and public awareness campaigns that emphasize the importance of antioxidants and encourage antioxidants-enriched diets.

References

1. Gulcin İ (2020) Antioxidants and antioxidant methods: an updated overview. Arch Toxicol 94(3):651–715. <https://doi.org/10.1007/s00204-020-02689-3>
2. Singh A, Kukreti R, Saso L, Kukreti S (2019) Oxidative stress: a key modulator in neurodegenerative diseases. Molecules 24(8). <https://doi.org/10.3390/molecules24081583>
3. Di Meo S, Venditti P (2020) Evolution of the knowledge of free radicals and other oxidants. Oxidative Med Cell Longev 2020:9829176. <https://doi.org/10.1155/2020/9829176>

4. Linnane AW, Kios M, Vitetta L (2007) The essential requirement for superoxide radical and nitric oxide formation for normal physiological function and healthy aging. *Mitochondrion* 7(1-2):1–5. <https://doi.org/10.1016/j.mito.2006.11.009>
5. Niki E (2009) Lipid peroxidation: physiological levels and dual biological effects. *Free Radic Biol Med* 47(5):469–484. <https://doi.org/10.1016/j.freeradbiomed.2009.05.032>
6. Circu ML, Aw TY (2010) Reactive oxygen species, cellular redox systems, and apoptosis. *Free Radic Biol Med* 48(6):749–762. <https://doi.org/10.1016/j.freeradbiomed.2009.12.022>
7. Neha K, Haider MR, Pathak A, Yar MS (2019) Medicinal prospects of antioxidants: a review. *Eur J Med Chem* 178:687–704. <https://doi.org/10.1016/j.ejmech.2019.06.010>
8. Rahal A, Kumar A, Singh V, Yadav B, Tiwari R, Chakraborty S, Dhama K (2014) Oxidative stress, prooxidants, and antioxidants: the interplay. *Biomed Res Int* 2014:761264. <https://doi.org/10.1155/2014/761264>
9. Inoue M, Sato EF, Nishikawa M, Park AM, Kira Y, Imada I, Utsumi K (2003) Mitochondrial generation of reactive oxygen species and its role in aerobic life. *Curr Med Chem* 10(23):2495–2505. <https://doi.org/10.2174/0929867033456477>
10. Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, Squadrito F, Altavilla D, Bitto A (2017) Oxidative stress: harms and benefits for human health. *Oxidative Med Cell Longev* 2017:8416763. <https://doi.org/10.1155/2017/8416763>
11. Teodoro JS, Duarte FV, Gomes AP, Varela AT, Peixoto FM, Rolo AP, Palmeira CM (2013) Berberine reverts hepatic mitochondrial dysfunction in high-fat fed rats: a possible role for SirT3 activation. *Mitochondrion* 13(6):637–646. <https://doi.org/10.1016/j.mito.2013.09.002>
12. Brownlee M (2001) Biochemistry and molecular cell biology of diabetic complications. *Nature* 414(6865):813–820. <https://doi.org/10.1038/414813a>
13. Selemidis S, Sobey CG, Wingler K, Schmidt HH, Drummond GR (2008) NADPH oxidases in the vasculature: molecular features, roles in disease and pharmacological inhibition. *Pharmacol Ther* 120(3):254–291. <https://doi.org/10.1016/j.pharmthera.2008.08.005>
14. Dröge W (2002) Free radicals in the physiological control of cell function. *Physiol Rev* 82(1):47–95. <https://doi.org/10.1152/physrev.00018.2001>
15. Finkel T, Holbrook NJ (2000) Oxidants, oxidative stress and the biology of ageing. *Nature* 408(6809):239–247. <https://doi.org/10.1038/35041687>
16. Starkov AA (2008) The role of mitochondria in reactive oxygen species metabolism and signaling. *Ann N Y Acad Sci* 1147:37–52. <https://doi.org/10.1196/annals.1427.015>
17. De Duve C, Baudhuin P (1966) Peroxisomes (microbodies and related particles). *Physiol Rev* 46(2):323–357. <https://doi.org/10.1152/physrev.1966.46.2.323>
18. Schrader M, Fahimi HD (2006) Peroxisomes and oxidative stress. *Biochim Biophys Acta* 1763(12):1755–1766. <https://doi.org/10.1016/j.bbamcr.2006.09.006>
19. Cheeseman KH, Slater TF (1993) An introduction to free radical biochemistry. *Br Med Bull* 49(3):481–493. <https://doi.org/10.1093/oxfordjournals.bmb.a072625>
20. Gross E, Sevier CS, Heldman N, Vitu E, Bentzur M, Kaiser CA, Thorpe C, Fass D (2006) Generating disulfides enzymatically: reaction products and electron acceptors of the endoplasmic reticulum thiol oxidase Ero1p. *Proc Natl Acad Sci U S A* 103(2):299–304. <https://doi.org/10.1073/pnas.0506448103>
21. Wang L, Muxin G, Nishida H, Shirakawa C, Sato S, Konishi T (2007) Psychological stress-induced oxidative stress as a model of sub-healthy condition and the effect of TCM. *Evid Based Complement Alternat Med* 4(2):195–202. <https://doi.org/10.1093/ecam/nel080>
22. Padmavathi P, Raghu PS, Reddy VD, Bulle S, Marthadu SB, Maturu P, Varadacharyulu N (2018) Chronic cigarette smoking-induced oxidative/nitrosative stress in human erythrocytes and platelets. *Mol Cell Toxicol* 14(1):27–34
23. Gloire G, Legrand-Poels S, Piette J (2006) NF-kappaB activation by reactive oxygen species: fifteen years later. *Biochem Pharmacol* 72(11):1493–1505. <https://doi.org/10.1016/j.bcp.2006.04.011>
24. de Jager TL, Cockrell AE, Du Plessis SS (2017) Ultraviolet light induced generation of reactive oxygen species. *Adv Exp Med Biol* 996:15–23. https://doi.org/10.1007/978-3-319-56017-5_2

25. Pham-Huy LA, He H, Pham-Huy C (2008) Free radicals, antioxidants in disease and health. *Int J Biomed Sci* 4(2):89–96
26. Radak Z, Zhao Z, Goto S, Koltai E (2011) Age-associated neurodegeneration and oxidative damage to lipids, proteins and DNA. *Mol Asp Med* 32(4-6):305–315. <https://doi.org/10.1016/j.mam.2011.10.010>
27. Payen VL, Zampieri LX, Porporato PE, Sonveaux P (2019) Pro- and antitumor effects of mitochondrial reactive oxygen species. *Cancer Metastasis Rev* 38(1-2):189–203. <https://doi.org/10.1007/s10555-019-09789-2>
28. Ventura HO, Taler SJ, Strobeck JE (2005) Hypertension as a hemodynamic disease: the role of impedance cardiography in diagnostic, prognostic, and therapeutic decision making. *Am J Hypertens* 18(S2):26S–43S
29. Kearney PM, Whelton M, Reynolds K, Muntner P, Whelton PK, He J (2005) Global burden of hypertension: analysis of worldwide data. *Lancet* 365(9455):217–223
30. Agita A, Alsagaff MT (2017) Inflammation, immunity, and hypertension. *Acta Medica Indonesiana* 49(2):158
31. Castro MM, Tanus-Santos JE (2013) Inhibition of matrix metalloproteinases (MMPs) as a potential strategy to ameliorate hypertension-induced cardiovascular alterations. *Curr Drug Targets* 14(3):335–343
32. Pinheiro LC, Oliveira-Paula GH (2020) Sources and effects of oxidative stress in hypertension. *Curr Hypertens Rev* 16(3):166–180
33. Nakazono K, Watanabe N, Matsuno K, Sasaki J, Sato T, Inoue M (1991) Does superoxide underlie the pathogenesis of hypertension? *Proc Natl Acad Sci* 88(22):10045–10048
34. Rajagopalan S, Kurz S, Münzel T, Tarpey M, Freeman BA, Griendling KK, Harrison DG (1996) Angiotensin II-mediated hypertension in the rat increases vascular superoxide production via membrane NADH/NADPH oxidase activation. Contribution to alterations of vasomotor tone. *J Clin Invest* 97(8):1916–1923
35. Ojeda NB, Hennington BS, Williamson DT, Hill ML, Betson NE, Sartori-Valinotti JC, Reckelhoff JF, Royals TP, Alexander BT (2012) Oxidative stress contributes to sex differences in blood pressure in adult growth-restricted offspring. *Hypertension* 60(1):114–122
36. Laursen JB, Rajagopalan S, Galis Z, Tarpey M, Freeman BA, Harrison DG (1997) Role of superoxide in angiotensin II-induced but not catecholamine-induced hypertension. *Circulation* 95(3):588–593
37. Chen D-D, Chen L-Y, Xie J-B, Shu C, Yang T, Zhou S, Yuan HF, Chen A (2014) Tetrahydrobiopterin regulation of eNOS redox function. *Curr Pharm Des* 20(22):3554–3562
38. Chen W, Druhan LJ, Chen C-A, Hemann C, Chen Y-R, Berka V, Tsai A-L, Zweier JL (2010) Peroxynitrite induces destruction of the tetrahydrobiopterin and heme in endothelial nitric oxide synthase: transition from reversible to irreversible enzyme inhibition. *Biochemistry* 49(14):3129–3137
39. Montezano AC, Dulak-Lis M, Tsiropoulou S, Harvey A, Briones AM, Touyz RM (2015) Oxidative stress and human hypertension: vascular mechanisms, biomarkers, and novel therapies. *Can J Cardiol* 31(5):631–641
40. Katzman R, Saitoh T (1991) Advances in Alzheimer's disease. *FASEB J* 5(3):278–286
41. Salmon DP, Thomas R, Pay M, Booth A, Hofstetter C, Thal L, Katzman R (2002) Alzheimer's disease can be accurately diagnosed in very mildly impaired individuals. *Neurology* 59(7):1022–1028
42. Chang Y, Kong Q, Shan X, Tian G, Ilieva H, Cleveland DW, Rothstein JD, Borchelt DR, Wong PC, Lin C-IG (2008) Messenger RNA oxidation occurs early in disease pathogenesis and promotes motor neuron degeneration in ALS. *PLoS One* 3(8):e2849
43. Uttara B, Singh AV, Zamboni P, Mahajan R (2009) Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. *Curr Neuropharmacol* 7(1):65–74
44. Butterfield DA, Perluigi M, Sultana R (2006) Oxidative stress in Alzheimer's disease brain: new insights from redox proteomics. *Eur J Pharmacol* 545(1):39–50

45. Goldstein BD, Witz G (1990) Free radicals and carcinogenesis. *Free Radic Res Commun* 11(1-3):3–10
46. Dreher D, Junod AF (1996) Role of oxygen free radicals in cancer development. *Eur J Cancer* 32(1):30–38
47. Cairns R, Harris I, McCracken S, Mak T Cancer cell metabolism. In: *Cold Spring Harbor symposia on quantitative biology*, 2011. Cold Spring Harbor Laboratory Press, pp 299–311
48. Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J (2007) Free radicals and antioxidants in normal physiological functions and human disease. *Int J Biochem Cell Biol* 39(1):44–84
49. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D (2011) Global cancer statistics. *CA Cancer J Clin* 61(2):69–90
50. Blau S, Rubinstein A, Bass P, Singaram C, Kohen R (1999) Differences in the reducing power along the rat GI tract: lower antioxidant capacity of the colon. *Mol Cell Biochem* 194(1):185–191
51. Foksinski M, Rozalski R, Guz J, Ruszkowska B, Sztukowska P, Piwowarski M, Klungland A, Olinski R (2004) Urinary excretion of DNA repair products correlates with metabolic rates as well as with maximum life spans of different mammalian species. *Free Radic Biol Med* 37(9):1449–1454
52. Guz J, Foksinski M, Siomek A, Gackowski D, Rozalski R, Dziaman T, Szpila A, Olinski R (2008) The relationship between 8-oxo-7, 8-dihydro-2'-deoxyguanosine level and extent of cytosine methylation in leukocytes DNA of healthy subjects and in patients with colon adenomas and carcinomas. *Mutat Res/Fundament Mol Mech Mutagen* 640(1-2):170–173
53. Murrell T (1991) Epidemiological and biochemical support for a theory on the cause and prevention of breast cancer. *Med Hypotheses* 36(4):389–396
54. Brown NS, Jones A, Fujiyama C, Harris AL, Bicknell R (2000) Thymidine phosphorylase induces carcinoma cell oxidative stress and promotes secretion of angiogenic factors. *Cancer Res* 60(22):6298–6302
55. Arnold RS, He J, Remo A, Ritsick D, Yin-Goen Q, Lambeth JD, Datta MW, Young AN, Petros JA (2007) Nox1 expression determines cellular reactive oxygen and modulates c-fos-induced growth factor, interleukin-8, and Cav-1. *Am J Pathol* 171(6):2021–2032
56. Lim SD, Sun C, Lambeth JD, Marshall F, Amin M, Chung L, Petros JA, Arnold RS (2005) Increased Nox1 and hydrogen peroxide in prostate cancer. *Prostate* 62(2):200–207
57. Azad N, Rojanasakul Y, Vallyathan V (2008) Inflammation and lung cancer: roles of reactive oxygen/nitrogen species. *J Toxicol Environ Health Part B* 11(1):1–15
58. Atta EM, Mohamed NH, Silaev AAA (2017) Antioxidants: An overview on the natural and synthetic types. *Eur Chem Bull* 6(8):365–375
59. Moussa Z, Judeh Z, Ahmed SA (2019) Nonenzymatic exogenous and endogenous antioxidants. *Free Rad Med Biol*:1–22
60. Admassu S, Kebede M (2019) Application of antioxidants in food processing industry: Options to improve the extraction yields and market value of natural products. *Adv Food Technol Nutr Sci* 5:38–49
61. Bouayed J, Bohn T (2010) Exogenous antioxidants – double-edged swords in cellular redox state: Health beneficial effects at physiologic doses versus deleterious effects at high doses. *Oxidative Med Cell Longev* 3(4):228–237. <https://doi.org/10.4161/oxim.3.4.12858>
62. Shahidi F (2015) *Handbook of antioxidants for food preservation*. Woodhead Publishing
63. Tan BL, Norhaizan ME, Liew WP, Sulaiman Rahman H (2018) Antioxidant and oxidative stress: a mutual interplay in age-related diseases. *Front Pharmacol* 9:1162. <https://doi.org/10.3389/fphar.2018.01162>
64. Scalbert A, Johnson IT, Saltmarsh M (2005) Polyphenols: antioxidants and beyond. *Am J Clin Nutr* 81 (1 Suppl):215s–217s. <https://doi.org/10.1093/ajcn/81.1.215S>
65. Gammeren DV (2008) *Vitamins and minerals. Essentials of sports nutrition and supplements*. Springer, In, pp 313–328
66. Hands ES (1990) *Food finder: food sources of vitamins and minerals*. Esha Research

67. Xavier AA, Pérez-Gálvez A (2016) Carotenoids as a source of antioxidants in the diet. *Subcell Biochem* 79:359–375. https://doi.org/10.1007/978-3-319-39126-7_14
68. Petropoulos S, Di Gioia F, Ntatsi G (2017) Vegetable organosulfur compounds and their health promoting effects. *Curr Pharm Des* 23(19):2850–2875. <https://doi.org/10.2174/1381612823666170111100531>
69. Nicastro HL, Ross SA, Milner JA (2015) Garlic and onions: their cancer prevention properties. *Cancer Prev Res (Phila)* 8(3):181–189. <https://doi.org/10.1158/1940-6207.Capr-14-0172>
70. Netzel M, Strass G, Janssen M, Bitsch I, Bitsch R (2001) Bioactive anthocyanins detected in human urine after ingestion of blackcurrant juice. *J Environ Pathol Toxicol Oncol* 20(2):89–95
71. Arts IC, van De Putte B, Hollman PC (2000) Catechin contents of foods commonly consumed in The Netherlands. 2. Tea, wine, fruit juices, and chocolate milk. *J Agric Food Chem* 48(5):1752–1757. <https://doi.org/10.1021/jf000026+>
72. Panche AN, Diwan AD, Chandra SR (2016) Flavonoids: an overview. *J Nutr Sci* 5:e47. <https://doi.org/10.1017/jns.2016.41>
73. Bhattacharyya A, Chattopadhyay R, Mitra S, Crowe SE (2014) Oxidative stress: an essential factor in the pathogenesis of gastrointestinal mucosal diseases. *Physiol Rev* 94(2):329–354. <https://doi.org/10.1152/physrev.00040.2012>
74. Fridovich I (1989) Superoxide dismutases. An adaptation to a paramagnetic gas. *J Biol Chem* 264(14):7761–7764
75. Battino M, Ferreiro MS, Gallardo I, Newman HN, Bullon P (2002) The antioxidant capacity of saliva. *J Clin Periodontol* 29(3):189–194. <https://doi.org/10.1034/j.1600-051x.2002.290301x.x>
76. Farghaly TA, Al-Hussain SA, Zaki ME, Asghar BH, Muhammad ZA (2022) Synthesis of spiropyrazoles under organic and nonorganic catalysis. *Curr Org Chem* 26(9):834–856
77. Piatkivskiy A, Lau JK-C, Berden G, Oomens J, Hopkinson AC, Siu KM, Ryzhov V (2019) Hydrogen atom transfer in the radical cations of tryptophan-containing peptides AW and WA studied by mass spectrometry, infrared multiple-photon dissociation spectroscopy, and theoretical calculations. *Eur J Mass Spectromet* 25(1):112–121
78. Guo Y, Baschieri A, Mollica F, Valgimigli L, Cedrowski J, Litwinienko G, Amorati R (2021) Hydrogen atom transfer from HOO(·) to ortho-quinones explains the antioxidant activity of polydopamine. *Angew Chem Int Ed Eng* 60(28):15220–15224. <https://doi.org/10.1002/anie.202101033>
79. Galeotti M, Trasatti C, Sisti S, Salamone M, Bietti M (2022) Factors governing reactivity and selectivity in hydrogen atom transfer from C(sp³)-H bonds of nitrogen-containing heterocycles to the cumyloxyl radical. *J Org Chem* 87(11):7456–7463. <https://doi.org/10.1021/acs.joc.2c00955>
80. Lesslie M, Lau JK, Lawler JT, Siu KW, Oomens J, Berden G, Hopkinson AC, Ryzhov V (2016) Alkali-metal-ion-assisted hydrogen atom transfer in the homocysteine radical. *Chemistry* 22(7):2243–2246. <https://doi.org/10.1002/chem.201504631>
81. León EI, Martín Á, Montes AS, Pérez-Martín I, Del Sol RM, Suárez E (2021) 1,5-hydrogen atom transfer/Surzur-Tanner rearrangement: a radical cascade approach for the synthesis of 1,6-dioxaspiro[4.5]decane and 6,8-dioxabicyclo[3.2.1]octane scaffolds in carbohydrate systems. *J Org Chem* 86(21):14508–14552. <https://doi.org/10.1021/acs.joc.1c01376>
82. Zheng YZ, Deng G, Zhang YC (2021) Multiple free radical scavenging reactions of auronones. *Phytochemistry* 190:112853. <https://doi.org/10.1016/j.phytochem.2021.112853>
83. Dalla Tiezza M, Hamlin TA, Bickelhaupt FM, Orian L (2021) Radical scavenging potential of the phenothiazine scaffold: a computational analysis. *ChemMedChem* 16(24):3763–3771. <https://doi.org/10.1002/cmdc.202100546>
84. Boulebd H (2021) Modeling the peroxy radical scavenging behavior of carnosic acid: mechanism, kinetics, and effects of physiological environments. *Phytochemistry* 192:112950. <https://doi.org/10.1016/j.phytochem.2021.112950>

85. Vo QV, Hoa NT, Nam PC, Quang DT, Mechler A (2020) In silico evaluation of the radical scavenging mechanism of mactanamide. *ACS Omega* 5(37):24106–24110. <https://doi.org/10.1021/acsomega.0c03646>
86. Alisi IO, Uzairu A, Abechi SE (2020) Free radical scavenging mechanism of 1,3,4-oxadiazole derivatives: thermodynamics of O-H and N-H bond cleavage. *Heliyon* 6(3):e03683. <https://doi.org/10.1016/j.heliyon.2020.e03683>
87. Miro P, Marin ML, Miranda MA (2016) Radical-mediated dehydrogenation of bile acids by means of hydrogen atom transfer to triplet carbonyls. *Org Biomol Chem* 14(9):2679–2683. <https://doi.org/10.1039/c5ob02561c>
88. Herman A, Herman AP (2013) Caffeine's mechanisms of action and its cosmetic use. *Skin Pharmacol Physiol* 26(1):8–14. <https://doi.org/10.1159/000343174>
89. Ranganatha VL, Begum AB, Naveen P, Zameer F, Hegdekatte R, Khanum SA (2014) Synthesis, xanthine oxidase inhibition, and antioxidant screening of benzophenone tagged thiazolidinone analogs. *Arch Pharm (Weinheim)* 347(8):589–598. <https://doi.org/10.1002/ardp.201400058>
90. Mohamed Isa SSP, Ablat A, Mohamad J (2018) The antioxidant and xanthine oxidase inhibitory activity of plumeria rubra flowers. *Molecules* 23(2). <https://doi.org/10.3390/molecules23020400>
91. Simunkova M, Alwasel SH, Alhazza IM, Jomova K, Kollar V, Rusko M, Valko M (2019) Management of oxidative stress and other pathologies in Alzheimer's disease. *Arch Toxicol* 93(9):2491–2513. <https://doi.org/10.1007/s00204-019-02538-y>
92. Patrick L (2002) Mercury toxicity and antioxidants: Part 1: role of glutathione and alpha-lipoic acid in the treatment of mercury toxicity. *Altern Med Rev* 7(6):456–471
93. Parvin MS, Chlebek J, Hošťálková A, Catapano MC, Lomozová Z, Macáková K, Mladěnka P (2022) Interactions of isoquinoline alkaloids with transition metals iron and copper. *Molecules* 27(19). <https://doi.org/10.3390/molecules27196429>
94. Filipický T, Říha M, Macáková K, Anzenbacherová E, Karlíčková J, Mladěnka P (2015) Antioxidant effects of coumarins include direct radical scavenging, metal chelation and inhibition of ROS-producing enzymes. *Curr Top Med Chem* 15(5):415–431. <https://doi.org/10.2174/1568026615666150206152233>
95. Ajisaka K, Oyanagi Y, Miyazaki T, Suzuki Y (2016) Effect of the chelation of metal cation on the antioxidant activity of chondroitin sulfates. *Biosci Biotechnol Biochem* 80(6):1179–1185. <https://doi.org/10.1080/09168451.2016.1141036>
96. Lin TK, Zhong L, Santiago JL (2017) Anti-inflammatory and skin barrier repair effects of topical application of some plant oils. *Int J Mol Sci* 19(1). <https://doi.org/10.3390/ijms19010070>
97. de Oliveira SE, Batista R (2017) Ferulic acid and naturally occurring compounds bearing a feruloyl moiety: a review on their structures, occurrence, and potential health benefits. *Compr Rev Food Sci Food Saf* 16(4):580–616. <https://doi.org/10.1111/1541-4337.12266>
98. Deng L, Du C, Song P, Chen T, Rui S, Armstrong DG, Deng W (2021) The role of oxidative stress and antioxidants in diabetic wound healing. *Oxidative Med Cell Longev* 2021:8852759. <https://doi.org/10.1155/2021/8852759>
99. Xu Z, Han S, Gu Z, Wu J (2020) Advances and impact of antioxidant hydrogel in chronic wound healing. *Adv Healthc Mater* 9(5):e1901502. <https://doi.org/10.1002/adhm.201901502>
100. Dai J, Mumper RJ (2010) Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules* 15(10):7313–7352. <https://doi.org/10.3390/molecules15107313>
101. Pfister R, Heider K, Illgen B, Beglinger R (1990) Trichospirura leptostoma: a possible cause of wasting disease in the marmoset. *Z Versuchstierkd* 33(4):157–161
102. LT FMAAA, Anderson HR, Bhutta ZA, Biryukov S et al (2016) Global, regional, and national comparative risk assessment of 79 behavioural, environmental and occupational, and metabolic risks or clusters of risks, 1990–2015: a systematic analysis for the global burden of disease study 2015. *Lancet* 388:1659–1724

103. Colafella KMM, Denton KM (2018) Sex-specific differences in hypertension and associated cardiovascular disease. *Nat Rev Nephrol* 14(3):185–201. <https://doi.org/10.1038/nrneph.2017.189>
104. Di Raimondo D, Buscemi S, Musiari G, Rizzo G, Pirera E, Corleo D, Pinto A, Tuttolomondo A (2021) Ketogenic diet, physical activity, and hypertension – a narrative review. *Nutrients* 13(8). <https://doi.org/10.3390/nu13082567>
105. Aidietis A, Laucevicius A, Marinskis G (2007) Hypertension and cardiac arrhythmias. *Curr Pharm Des* 13(25):2545–2555. <https://doi.org/10.2174/138161207781663037>
106. Ferdinand DP, Nedunchezian S, Ferdinand KC (2020) Hypertension in African Americans: Advances in community outreach and public health approaches. *Prog Cardiovasc Dis* 63(1):40–45. <https://doi.org/10.1016/j.pcad.2019.12.005>
107. Hermida RC, Crespo JJ, Domínguez-Sardiña M, Otero A, Moyá A, Ríos MT, Sineiro E, Castiñeira MC, Callejas PA, Pousa L, Salgado JL, Durán C, Sánchez JJ, Fernández JR, Mojon A, Ayala DE (2020) Bedtime hypertension treatment improves cardiovascular risk reduction: the hygia chronotherapy trial. *Eur Heart J* 41(48):4565–4576. <https://doi.org/10.1093/eurheartj/ehz754>
108. Ondimu DO, Kikuvu GM, Otieno WN (2019) Risk factors for hypertension among young adults (18-35) years attending in Tenwek Mission Hospital, Bomet County, Kenya in 2018. *Pan Afr Med J* 33:210. doi:<https://doi.org/10.11604/pamj.2019.33.210.18407>
109. Vaziri N (2000) Ni Z, Oveisi F, and Trnavsky-Hobbs DL. Effect of antioxidant therapy on blood pressure and NO synthase expression in hypertensive rats. *Hypertension* 36:957–964
110. Ahmad KA, Yuan Yuan D, Nawaz W, Ze H, Zhuo CX, Talal B, Taleb A, Mais E, Qilong D (2017) Antioxidant therapy for management of oxidative stress induced hypertension. *Free Radic Res* 51(4):428–438
111. Mazzali M, Hughes J, Kim YG, Jefferson JA, Kang DH, Gordon KL, Lan HY, Kivlighn S, Johnson RJ (2001) Elevated uric acid increases blood pressure in the rat by a novel crystal-independent mechanism. *Hypertension* 38:1101–1106
112. Greig JA, Shirley R, Graham D, Denby L, Dominiczak AF, Work LM, Baker AH (2010) Vascular-targeting antioxidant therapy in a model of hypertension and stroke. *J Cardiovasc Pharmacol* 56(6):642–650
113. Yao E-H, Fukuda N, Matsumoto T, Katakawa M, Yamamoto C, Han Y, Ueno T, Kobayashi N, Matsumoto K (2008) Effects of the antioxidative β -blocker celiprolol on endothelial progenitor cells in hypertensive rats. *Am J Hypertens* 21(9):1062–1068
114. Mak IT, Weglicki WB (1988) Protection by beta-blocking agents against free radical-mediated sarcolemmal lipid peroxidation. *Circ Res* 63(1):262–266
115. Kukin ML, Kalman J, Charney RH, Levy DK, Buchholz-Varley C, Ocampo ON, Eng C (1999) Prospective, randomized comparison of effect of long-term treatment with metoprolol or carvedilol on symptoms, exercise, ejection fraction, and oxidative stress in heart failure. *Circulation* 99(20):2645–2651
116. Pasini AF, Garbin U, Nava MC, Stranieri C, Davoli A, Sawamura T, Cascio VL, Cominacini L (2005) Nebivolol decreases oxidative stress in essential hypertensive patients and increases nitric oxide by reducing its oxidative inactivation. *J Hypertens* 23(3):589–596
117. Lin SJ, Austriaco N (2014) Aging and cell death in the other yeasts, *Schizosaccharomyces pombe* and *Candida albicans*. *FEMS Yeast Res* 14(1):119–135. <https://doi.org/10.1111/1567-1364.12113>
118. Książek K (2010) Let's stop overlooking bacterial aging. *Biogerontology* 11(6):717–723. <https://doi.org/10.1007/s10522-010-9278-3>
119. Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, Gargiulo G, Testa G, Cacciatore F, Bonaduce D, Abete P (2018) Oxidative stress, aging, and diseases. *Clin Interv Aging* 13:757–772. <https://doi.org/10.2147/cia.S158513>
120. Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, Gargiulo G, Testa G, Cacciatore F, Bonaduce D et al (2018) Oxidative stress, aging, and diseases. *Clin Interv Aging* 13:757–772

121. Sadgrove NJ, Simmonds MSJ (2021) Topical and nutricosmetic products for healthy hair and dermal antiaging using “dual-acting” (2 for 1) plant-based peptides, hormones, and cannabinoids. *FASEB Bioadv* 3(8):601–610. <https://doi.org/10.1096/fba.2021-00022>
122. Milosheska D, Roškar R (2022) Use of retinoids in topical antiaging treatments: a focused review of clinical evidence for conventional and nanoformulations. *Adv Ther* 39(12):5351–5375. <https://doi.org/10.1007/s12325-022-02319-7>
123. Berry K, Hallock K, Lam C (2022) Photoaging and topical rejuvenation. *Facial Plast Surg Clin North Am* 30(3):291–300. <https://doi.org/10.1016/j.fsc.2022.03.003>
124. Mumtaz S, Ali S, Tahir HM, Kazmi SAR, Shakir HA, Mughal TA, Mumtaz S, Summer M, Farooq MA (2021) Aging and its treatment with vitamin C: a comprehensive mechanistic review. *Mol Biol Rep* 48(12):8141–8153. <https://doi.org/10.1007/s11033-021-06781-4>
125. Tonnard P, Verpaele A, Carvas M (2020) Fat grafting for facial rejuvenation with nanofat grafts. *Clin Plast Surg* 47(1):53–62. <https://doi.org/10.1016/j.cps.2019.08.006>
126. Masaki H (2010) Role of antioxidants in the skin: anti-aging effects. *J Dermatol Sci* 58(2):85–90. <https://doi.org/10.1016/j.jdermsci.2010.03.003>
127. Boo YC (2022) Ascorbic acid (Vitamin C) as a cosmeceutical to increase dermal collagen for skin antiaging purposes: emerging combination therapies. *Antioxidants (Basel)* 11(9). <https://doi.org/10.3390/antiox11091663>
128. Kamel NS, Gammack J, Cepeda O, Flaherty JH (2006) Antioxidants and hormones as antiaging therapies: high hopes, disappointing results. *Cleve Clin J Med* 73 (12):1049-1056, 1058. <https://doi.org/10.3949/ccjm.73.12.1049>
129. Gao Y, Wei Y, Wang Y, Gao F, Chen Z (2017) *Lycium Barbarum*: a traditional Chinese herb and a promising anti-aging agent. *Aging Dis* 8(6):778–791. <https://doi.org/10.14336/ad.2017.0725>
130. Zou H, Ye H, Kamaraj R, Zhang T, Zhang J, Pavak P (2021) A review on pharmacological activities and synergistic effect of quercetin with small molecule agents. *Phytomedicine* 92:153736. <https://doi.org/10.1016/j.phymed.2021.153736>
131. Si H, Lai CQ, Liu D (2021) Dietary epicatechin, a novel anti-aging bioactive small molecule. *Curr Med Chem* 28(1):3–18. <https://doi.org/10.2174/0929867327666191230104958>
132. Ide K, Matsuoka N, Yamada H, Furushima D, Kawakami K (2018) Effects of tea catechins on Alzheimer’s disease: recent updates and perspectives. *Molecules* 23(9). <https://doi.org/10.3390/molecules23092357>
133. Kim H, Eliuk S, Deshane J, Meleth S, Sanderson T, Pinner A, Robinson G, Wilson L, Kirk M, Barnes S (2007) 2D gel proteomics: an approach to study age-related differences in protein abundance or isoform complexity in biological samples. *Methods Mol Biol* 371:349–391. https://doi.org/10.1007/978-1-59745-361-5_24
134. Milani A, Basirnejad M, Shahbazi S, Bolhassani A (2017) Carotenoids: biochemistry, pharmacology and treatment. *Br J Pharmacol* 174(11):1290–1324. <https://doi.org/10.1111/bph.13625>
135. Sunder S (2019) Relevant topical skin care products for prevention and treatment of aging skin. *Facial Plast Surg Clin North Am* 27(3):413–418. <https://doi.org/10.1016/j.fsc.2019.04.007>
136. Pullar JM, Carr AC, Vissers MCM (2017) The roles of Vitamin C in skin health. *Nutrients* 9(8). <https://doi.org/10.3390/nu9080866>
137. Carr AC, Maggini S (2017) Vitamin C and immune function. *Nutrients* 9(11). <https://doi.org/10.3390/nu9111211>
138. Shenoy N, Creagan E, Witzig T, Levine M (2018) Ascorbic acid in cancer treatment: let the phoenix fly. *Cancer Cell* 34(5):700–706. <https://doi.org/10.1016/j.ccell.2018.07.014>
139. Milani GP, Macchi M, Guz-Mark A (2021) Vitamin C in the treatment of COVID-19. *Nutrients* 13(4). <https://doi.org/10.3390/nu13041172>
140. Moritz B, Schmitz AE, Rodrigues ALS, Dafre AL, Cunha MP (2020) The role of vitamin C in stress-related disorders. *J Nutr Biochem* 85:108459. <https://doi.org/10.1016/j.jnutbio.2020.108459>

141. Njus D, Kelley PM, Tu YJ, Schlegel HB (2020) Ascorbic acid: the chemistry underlying its antioxidant properties. *Free Radic Biol Med* 159:37–43. <https://doi.org/10.1016/j.freeradbiomed.2020.07.013>
142. Naveed M, Hejazi V, Abbas M, Kamboh AA, Khan GJ, Shumzaid M, Ahmad F, Babazadeh D, FangFang X, Modarresi-Ghazani F, WenHua L, XiaoHui Z (2018) Chlorogenic acid (CGA): a pharmacological review and call for further research. *Biomed Pharmacother* 97:67–74. <https://doi.org/10.1016/j.biopha.2017.10.064>
143. Mlcek J, Jurikova T, Skrovankova S, Sochor J (2016) Quercetin and its anti-allergic immune response. *Molecules* 21(5). <https://doi.org/10.3390/molecules21050623>
144. Saric S, Sivamani RK (2016) Polyphenols and sunburn. *Int J Mol Sci* 17(9). <https://doi.org/10.3390/ijms17091521>
145. Ritchie KJ, Walsh S, Sansom OJ, Henderson CJ, Wolf CR (2009) Markedly enhanced colon tumorigenesis in Apc(Min) mice lacking glutathione S-transferase Pi. *Proc Natl Acad Sci U S A* 106(49):20859–20864. <https://doi.org/10.1073/pnas.0911351106>
146. Henderson CJ, Ritchie KJ, McLaren A, Chakravarty P, Wolf CR (2011) Increased skin papilloma formation in mice lacking glutathione transferase GSTP. *Cancer Res* 71(22):7048–7060
147. Abel EL, Angel JM, Riggs PK, Langfield L, Lo H-H, Person MD, Awasthi YC, Wang L-E, Strom SS, Wei Q (2010) Evidence that Gsta4 modifies susceptibility to skin tumor development in mice and humans. *J Natl Cancer Inst* 102(21):1663–1675
148. Zhang Y, Ikeno Y, Qi W, Chaudhuri A, Li Y, Bokov A, Thorpe SR, Baynes JW, Epstein C, Richardson A, Van Remmen H (2009) Mice deficient in both Mn superoxide dismutase and glutathione peroxidase-1 have increased oxidative damage and a greater incidence of pathology but no reduction in longevity. *J Gerontol A Biol Sci Med Sci* 64(12):1212–1220. <https://doi.org/10.1093/gerona/glp132>
149. Van Remmen H, Ikeno Y, Hamilton M, Pahlavani M, Wolf N, Thorpe SR, Alderson NL, Baynes JW, Epstein CJ, Huang TT, Nelson J, Strong R, Richardson A (2003) Life-long reduction in MnSOD activity results in increased DNA damage and higher incidence of cancer but does not accelerate aging. *Physiol Genomics* 16(1):29–37. <https://doi.org/10.1152/physiolgenomics.00122.2003>
150. Shen G, Xu C, Hu R, Jain MR, Nair S, Lin W, Yang CS, Chan JY, Kong AN (2005) Comparison of (-)-epigallocatechin-3-gallate elicited liver and small intestine gene expression profiles between C57BL/6J mice and C57BL/6J/Nrf2 (-/-) mice. *Pharm Res* 22(11):1805–1820. <https://doi.org/10.1007/s11095-005-7546-8>
151. Saeidnia S, Abdollahi M (2013) Antioxidants: friends or foe in prevention or treatment of cancer: the debate of the century. *Toxicol Appl Pharmacol* 271(1):49–63. <https://doi.org/10.1016/j.taap.2013.05.004>