

Supercritical Fluid for Extraction and Isolation of Natural Compounds



K. Vidwathpriya, S. Sriranjani, P. K. Niharika, and N. V. Anil Kumar

Abstract Supercritical fluid (SCF) extraction has emerged as an effective and efficient method for separating important phytoconstituents. The extraction process is simple and environmentally friendly, generating minimal to no waste. This procedure offers various advantages over traditional extraction techniques. This chapter discusses the procedure, advantages, and different types of phytoconstituents isolated using supercritical fluids, with a preference for natural products.

Keywords Supercritical · Extraction · Phytoconstituents · Solvent · CO₂ · Dissolve · Oil · Alkaloids · Flavonoids · Terpenes

1 Introduction

Supercritical fluid (SCF) extraction is an analytical method to separate the analyte from the sample matrix using supercritical fluids as solvents (Hedrick et al. 1992). This technique is rapid, inexpensive, sustainable, and simple to execute, compared to the traditional Soxhlet extraction, where solvent costs are usually high, requiring several hours accompanied by an additional concentration step that aids pollution (Sapkale et al. 2010).

SCF extraction was initiated along with supercritical fluid chromatography in the late twentieth century for isolating forensically relevant compounds (Khaw et al. 2017). It later gained popularity when supercritical toluene was used mainly in the petroleum industry with many commercial interests.

Over the last few years, SCF extraction has gained recognition for its many established advantages, particularly supercritical carbon dioxide, because of its easy-to-use properties (2017). CO₂ has a near ambient critical temperature of 31 °C,

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allowing many biological materials and natural products to be processed around 35 °C without denaturation. It is being extensively used in decaffeination and power generation processes and is widely used to extract natural products, leaving no toxic residues behind (Khaw et al. 2017).

The advantage of supercritical CO₂ (ScCO₂) is that its extraction properties can be precisely varied with just minute changes in temperature and pressure. The properties can also be modified using solvents like ethanol (Camel 2001). Other than CO₂, various solvents are used to extract bioactive components from plants, namely, propane, DME, SF₂, and ethanol (Bizaj et al. 2021).

2 Methodology/Mechanism

A supercritical fluid is a substance whose thermodynamic properties are higher than the critical temperature and pressure of the source compound. The maximum temperature, beyond which the gaseous state of a substance cannot be liquified, irrespective of the amount of pressure applied, is called the critical temperature of the substance. Critical pressure is the minimum pressure required to condense a gaseous substance to a liquid at its critical temperature (Alekseev et al. 2020). For carbon dioxide, the critical temperature is 304.2 K and 73.0 atm.

In the supercritical region, a homogenous fluid materializes, which has unique physiochemical properties. In this region, the surface tension of the supercritical fluid is equal to zero, the dissolving and swelling capacity increases, and the viscosity decreases (Alekseev et al. 2020). The physiochemical properties can be modulated by changing the parameters of the supercritical state. The density of supercritical fluids changes with variations in pressure and temperature; a slight increase in pressure can cause a drastic increase in the density of the supercritical fluid, which in turn causes an increase in the solubility of the supercritical solvent. Once extraction is complete, solvent recovery is relatively simple due to the volatility of the supercritical fluid leaving behind the extracted analyte (Pourmortazavi et al. 2014). Such manipulations of the physiochemical properties make SCFs an excellent solvent for extraction due to their high selectivity, solubility, and extraction efficiency (Yousefi et al. 2019).

The setup (Fig. 1) for supercritical fluid extraction involves a pump, a pressurized compartment, and a collecting vessel. The solvent is commonly stored in a tank connected to a pressurized pump. The commonly used solvent is CO₂, pumped into the system as a liquid below 5 °C and at around 50 bars of pressure. The fluid is cooled to remain a liquid but heated to critical condition after pressurization. The pressure must be maintained in the extraction cell, and heating should be provided to counteract the cooling caused by the adiabatic expansion of the CO₂. Raw material from which the natural product is extracted is placed in the extraction cell, where pressure and temperature are controlled. The raw material is also pre-treated to modulate the moisture content and particle size for optimal extraction. The supercritical fluid is allowed to enter the pressurized extraction cell, where the natural

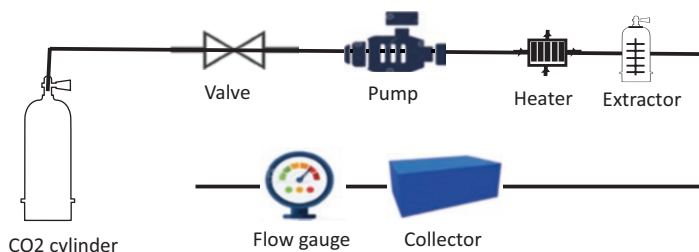


Fig. 1 Setup of ScCO₂ extraction

products to be extracted dissolve in the supercritical fluid based on its solubility, which in turn is dictated by its density and pressure. Once the extraction is completed, the fluid with the dissolved natural product is passed through a chamber with lesser pressure, reducing the fluid's dissolving power, and the natural product gets precipitated out. The depressurization of the supercritical CO₂ causes the fluid to become a gas and can be collected separately for further use (Sapkale et al. 2010).

Table 1 lists the chemicals/phytochemicals extracted from the species. Some of the chemicals listed in this table are generic in name, as the literature does not specify the individual compound. The table is arranged in alphabetical order of the species name.

Table 1 List of chemicals isolated from different species using ScCO₂

Sl no	Species name	Chemicals/phytochemicals	Ref
1	<i>Abelmoschus manihot</i>	Rutin, hyperin, isoquercetin, hibifolin, myricetin, quercetin-3'-O-glucoside, quercetin	Li et al. (2016a)
2	<i>Acacia dealbata</i>	Oxygenated triterpenes	Casas et al. (2021)
3	<i>Acacia dealbata</i>	Lupenyl acetate, lupenone, tetracosanoic acid, hexacosan-1-ol	Rodrigues et al. (2021)
4	<i>Acanthophoenix rubra</i>	Vitamin E	2018)
5	<i>Acanthus ilicifolius</i>	2-benzoxazolinone	Arumugam and Thiruganasambantham (2018)
6	<i>Acer nikoense</i>	Diarylheptanoids	Alberti et al. (2018)
7	<i>Alnus glutinosa</i>	β-Sitosterol, betulin, betulinic acid, lupeol	Felföldi-Gáva et al. (2012)
8	<i>Alnus hirsuta</i>	Diarylheptanoids	Alberti et al. (2018)
9	<i>Aloysia citrodora</i>	Phenylpropanoids, Flavonoids	Leyva-Jiménez et al. (2020)
10	<i>Alpinia blepharocalyx</i>	Diarylheptanoids	Alberti et al. (2018)
11	<i>Alpinia officinarum</i>	Diarylheptanoids	Alberti et al. (2018)

(continued)

Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
12	<i>Amaranthus cruentus</i>	Linoleic acid, decadieneal, linoleic acid propyl ester, 2,5-pentadecadiene-1-ol, 9-oxononanoic acid	Velikorodov et al. (2018)
13	<i>Ananas comosus</i>	Esters, ketones, alcohols, aldehydes, acids	Mohamad et al. (2019)
14	<i>Andrographis paniculata</i>	Rosmarinic acid, eurycomanone, andrographolide	Abd Aziz et al. (2021)
15	<i>Andrographis paniculata</i>	Andrographolide	Kumoro et al. (2019)
16	<i>Andrographis paniculata</i>	Andrographolides	Kumar et al. (2014)
17	<i>Annona muricata</i>	Flavonoids, Tannins, Phenolics, Phytate	Mesquita et al. (2021)
18	<i>Aquilaria malaccensis</i>	n-Hexadecanoic, 1H-Cycloprop[e]azulene, decahydro-1,1,7-trimethyl-4-methylene	Eissa et al. (2018)
19	<i>Artemisia annua</i>	Artemisinin	Baldino and Reverchon (2018)
20	<i>Ascophyllum nodosum</i> ,	Alginate, agar, carrageenan	Abdul Khalil et al. (2018)
21	<i>Azadirachta indica</i>	Terpinen-4-ol, 1,2,4-Trithiolane, 3,5-diethyl, allyl isopropyl sulphide, Cycloisolongifolene, á-Bisabolene, (-)-α-Panasinsen, Isocaryophyllene, trans-Sesquisabinene hydrate, 1-Naphthalenol	Swapna Sonale et al. (2018)
22	<i>Baccharis uncinella</i>	α-Pinene, β-pinene, limonene, (E)-caryophyllene, germacrene D, bicyclogermacrene, spathulenol, caryophyllene oxide	Minteguiaga et al. (2021)
23	<i>Betula platyphylla</i>	Diarylheptanoids	Alberti et al. (2018)
24	<i>Boswellia serrata</i>	α-Thujene, camphene, β-pinene, myrcene, limonene, m-cymene, cis-verbenol	Ayub et al. (2018)
25	<i>Brassica campestris</i>	Linolenic acid amide, linolenic acid glyceride, linolenic acid, palmitic acid	Li et al. (2016c)
26	<i>Brassica napus</i>	Phytosterols	Jafarian Asl et al. (2020)
27	<i>Bryonopsis laciniosa</i>	Linoleic acid, linolenic acid, β-sitosterol stigmasterol	Balkrishna et al. (2022)
28	<i>Calendula officinalis</i>	Bioactive pentacyclic triterpenes	Villanueva-Bermejo et al. (2019)
29	<i>Calluna vulgaris</i>	Bioactive pentacyclic triterpenes	Villanueva-Bermejo et al. (2019)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
30	<i>Camellia oleifera</i>	Palmitic acid, stearic acid, oleic acid, linoleic acid, α -tocopherol, β -carotene, squalene phytosterol, 3-hydroxytyrosol, benzoic acid, catechins, 4-hydroxybenzoic acid, chlorogenic acid	Fang et al. (2015)
31	<i>Cananga latifolia</i>	Phenolic acids, flavonoids, tannins, alkaloids	Chhouk et al. (2018)
32	<i>Cannabis sativa</i>	Tetrahydrocannabinol	Gallo-Molina et al. (2019)
33	<i>Cannabis sativa</i>	ω -6 linoleic acid, ω -3 α -linolenic acid	Devi and Khanam (2019b)
34	<i>Cannabis sativa</i>	Cannabidiol	Marzorati et al. (2020)
35	<i>Capsicum annum</i>	γ -Tocopherol	Cvetković et al. (2020)
36	<i>Capsicum chinense</i>	Rutin, vicenin-2	de Aguiar et al. (2019)
37	<i>Capsicum frutescens</i>	Capsaicinoids	de Aguiar et al. (2018)
38	<i>Carica papaya</i>	Oleic acid	Devi and Khanam (2019a)
39	<i>Catharanthus roseus</i>	Vincristine	Karimi and Raofie (2019)
40	<i>Chaenomeles japonica</i>	α -Tocopherol, β -tocopherol, γ -tocopherol	Górnaś et al. (2019)
41	<i>Chenopodium quinoa</i>	Tocopherol	Benito-Román et al. (2018)
42	<i>Cinnamomum cambodianum</i>	Phenolic acids, flavonoids, tannins, alkaloids	Chhouk et al. (2018)
43	<i>Cinnamomum verum</i>	Cinnamaldehyde, eugenol	Masghati and Ghoreishi (2018)
44	<i>Cinnamomum verum</i>	Eugenol, eugenol acetate	Khalil et al. (2017)
45	<i>Citrus grandis</i>	7-Methoxy-8-(2-oxo-3-methylbutyl) coumarin, (6E,8E,10E)-2,6,11,15-tetramethyl-2,6,8,10,14-hexadecapentaene, γ -sitosterol, hexadecanoic acid, (E,E)-2,4-decadienal, pentacosane	Gyawali et al. (2012a)
46	<i>Citrus grandis</i>	(Z)-9-Octadecenoic acid, limonene, α -Terpineol, (E,E)-2,4-decadienal, hexadecanoic acid, pentacosane, stigmasterol, γ -sitosterol	Gyawali et al. (2012b)
47	<i>Citrus hassaku</i>	(Z)-9-Octadecenoic acid, limonene, α -Terpineol, (E,E)-2,4-decadienal, hexadecanoic acid, pentacosane, stigmasterol, γ -sitosterol	Gyawali et al. (2012b)
48	<i>Citrus Iyo</i>	(Z)-9-Octadecenoic acid, limonene, α -Terpineol, (E,E)-2,4-decadienal, hexadecanoic acid, pentacosane, stigmasterol, γ -sitosterol	Gyawali et al. (2012b)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
49	<i>Citrus maxima</i>	Terpenes, terpenoids, aldehydes, alcohols, esters	Chen et al. (2018a)
50	<i>Citrus reticulata</i>	Nobiletin, 3,5,6,7,8,3',4'-heptamethoxyflavone, tangeretin	Long et al. (2019)
51	<i>Citrus sinensis</i>	α -Terpineol, D-Limonene, hesperidin	Barrales et al. (2018)
52	<i>Citrus sinensis</i>	Limonene, Hesperidin	Jokić et al. (2020)
53	<i>Colchicum speciosum</i>	Colchicine	Bayrak et al. (2019)
54	<i>Corallina officinalis</i>	Acyclic alkanes, branched alkanes, alkenes, organobromine compounds, organosulfur compounds, aromatic compounds, monoterpenes, sesquiterpenes, diterpenes, triterpene	Djapic (2018)
55	<i>Coriandrum sativum</i>	Linalool, camphor, linalool oxide, p-cymene, α -pinene, limonene, geranyl acetate	Choi and Lee (2018)
56	<i>Corylus avellana</i>	Diarylheptanoids	Alberti et al. (2018)
57	<i>Crocus sativus</i>	Crocetin sugar esters, picrocrocin, safranin	Kyriakoudi and Z. Tsimidou (2018)
58	<i>Crocus sativus</i>	Apocarotenoids, anthocyanins, flavonoids, anthocyanidins, phenolic compounds	Bakshi et al. (2022)
59	<i>Croton Polycarpus</i>	Flavanols, sesquiterpenoids	Aponte-Buitrago et al. (2017)
60	<i>Cucumis melo</i>	Linoleic acid, oleic acid, palmitic acid, stearic acid	Bouazzaoui et al. (2018)
61	<i>Cucurbita maxima</i>	Tocopherols	Rohman and Imawati (2020)
62	<i>Cucurbita pepo</i>	Desmosterol, campesterol, stigmasterol, β -sitosterol, spinasterol, Δ 7,22,25-stigmastatrienol, Δ 7-stigmastenol, Δ 7,25-stigmastadienol, Δ 7-avenasterol	Hrabovski et al. (2012)
63	<i>Cuminum cyminum</i>	Cumin aldehyde, γ -terpinene, β -pinene, β -Cumic aldehyde, α -phellandrene	Fang et al. (2018)
64	<i>Curcuma caesia</i>	Beta-elemene, curzerenone, boldenone, 2-cyclohexen-1-one, 4-ethynyl-4-hydroxy-3, 5, 5-trimethyl.	Chaturvedi et al. (2020)
65	<i>Curcuma longa</i>	Tumerone, ar-turmerone, curlone	Haiyee et al. (2016)
66	<i>Curcuma longa</i>	Turmeric oil	Priyanka and Khanam (2018)
67	<i>Cymbopogon citronella</i>	Essential oil	Wu et al. (2019)
68	<i>Cymbopogon winterianus</i>	Citronella oil	Salea et al. (2018)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
69	<i>Cynomorium coccineum</i>	Glucose, Fructose, Sucrose, Alanine, Asparagine, Glutamine Proline, Valine, Acetate, Citrate, Formate, Fumarate, Malate, Malonate, Succinate, Betaine, Choline	Attia et al. (2018)
70	<i>Cyphomandra Betacea</i>	Linoleic acid, oleic acid, palmitic acid, stearic acid, linolenic acid, palmitoleic acid, squalene, β -sitosterol, cycloartenol, dihydrolanosterol, sterols, γ -tocopherol	Dorado Achicanoy et al. (2018)
71	<i>Dalbergia ecastophyllum</i>	Artepillin C, p-coumaric acid	Machado et al. (2016)
72	<i>Daucus carota</i>	Carotenoids	Miękus et al. (2019)
73	<i>Derris elliptica</i>	Rotenoids	Baldino et al. (2018b)
74	<i>Descurainia sophia</i>	Sinapic acid	Hadinezhad et al. (2015)
75	<i>Dialium cochinchinense</i>	Phenolic acids, flavonoids, tannins, alkaloids	Chhouk et al. (2018)
76	<i>Dipteryx odorata</i>	Alcohols, carbonyl compounds, acids, esters, terpenes, terpenoids, lactones, aliphatic aromatic hydrocarbons	Bajer et al. (2018)
77	<i>Duguetia furfuracea</i>	Alloaromadendrene oxide-1, β -caryophyllene oxide,(+)-Spathulenol, Spathulenol,(–)(–) Caryophyllene oxide, Methyl eladiate, Aromadendrene oxide-2,Alloaromadendrene oxide-2,(–)-Spathulenol,Isoaromadendrene epoxide, 2-methylenecholestran-3-ol, α -tocoferol,Palmitic acid,3-Deoxyestradiol,2 Methyhexadecan-1-ol	Favareto et al. (2019)
78	<i>Echinacea purpurea</i>	Caftaric acid, cichoric acid, chlorogenic acid, cynarin, echinacoside	Konar et al. (2014)
79	<i>Eichhornia crassipes</i>	Stigmasterol, cholesterol, β -sitosterol	Martins et al. (2016)
80	<i>Elaeagnus angustifoli</i>	Linoleic acid, decadieneal, linoleic acid propyl ester, 2,5-pentadecadiene-1-ol, 9-oxononanoic acid	Velikorodov et al. (2018)
81	<i>Elaeagnus mollis</i>	Linoleic acid, oleic acid, palmitic acid	Mu et al. (2021)
82	<i>Elaeis guineensis</i>	Phenolics, flavonoids, carotenoids	Bezerra et al. (2018)
83	<i>Elaeis guineensis</i>	Phenolic compounds	Chan et al. (2018)
84	<i>Elaeis guineensis</i>	Vitamin E	Damrongwattanakool and Raviyan (2018)
85	<i>Elaeis guineensis</i>	Hexadecanoic acid, octadecanoic acid	Jaafar et al. (2011)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
86	<i>Elaeis guineensis</i>	α -Carotene, β -carotene	Carmona et al. (2018)
87	<i>Elettaria cardamomum</i>	1,8-cineol	Ghosh et al. (2015)
88	<i>Eremanthus erythropappus</i>	α -Bisabolol	Náthia-Neves et al. (2020)
89	<i>Eucalyptus globulus</i>	Quinolizidine alkaloids, β - Carotenes, Saponins, tannins, steroids, flavonoids	Abd Hamid et al. (2018)
90	<i>Eucommia ulmoides</i>	Linolenic acid	Zhang et al. (2018)
91	<i>Eugenia involucrata</i>	α -Tocopherol	Barzotto et al. (2019)
92	<i>Eurycoma longifolia</i>	Rosmarinic acid, eurycomanone, andrographolide	Abd Aziz et al. (2021)
93	<i>Ficus hirta</i>	Elemicin, Psoralen, Palmitic acid, Bergapten, Linolenic acid, Medicarpin, Retinoic Acid, Maackiain, Squalene	Deng et al. (2018)
94	<i>Foeniculum vulgare</i>	Sterols	Bettaieb Rebey et al. (2019)
95	<i>Furcraea selloa</i>	Saponins	Ramli et al. (2019)
96	<i>Ganoderma lucidum</i>	Oleic acid, palmitic acid, linoleic acid, Ergosta-7, 22-dien-3 β -ol, ergosterol	Li et al. (2016b)
97	<i>Garcinia mangostana</i>	Squalene, α -Cubebene	Hamid et al. (2013)
98	<i>Garcinia Mangostana</i>	α -Mangostin	Hamid et al. (2018)
99	<i>Gardenia angkorensis</i>	Phenolic acids, flavonoids, tannins, alkaloids	Chhouk et al. (2018)
100	<i>Glycine max</i>	Phytosterol, tocopherol	Han et al. (2016)
101	<i>Glycine max</i>	Polyene phosphatidyl choline	Jiang et al. (2016)
102	<i>Glycyrrhiza uralensis</i>	1-Methoxyerythrayssin II, 6,8-diprenylgenistein, gancaonin G, isoglycyrol, licorisoflavan C, licoricidin, licorisoflavan D, licorisoflavan E	Villinski et al. (2014)
103	<i>Haematococcus pluviialis</i>	Astaxanthin	Cheng et al. (2018)
104	<i>Haematococcus pluviialis</i>	Phorbol 12-myristate 13-acetate, doxycycline	Chou et al. (2016)
105	<i>Haematococcus pluviialis</i>	Astaxanthin, lutein, fatty acids	Di Sanzo et al. (2018)
106	<i>Hancornia speciosa</i>	Amyrin, lupeol, α -amyrin, β -carotene	Maia et al. (2018)
107	<i>Helianthus annuus</i>	Chlorogenic acid	Daraee et al. (2019)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
108	<i>Hippophae rhamnoides</i>	β -Sitosterol, α -tocopherol	Dienait� et al. (2021)
109	<i>Hippophae rhamnoides</i>	Zeaxanthin, β -carotene, lycopene, α -tocopherol, β -tocopherol, δ -tocopherol, β -sitosterol	Mihalcea et al. (2021)
110	<i>Humulus lupulus</i>	Xanthohumol, desmethylxanthohumol, bitter acids, phenolic compounds	Bizaj et al. (2021)
111	<i>Humulus lupulus</i>	Phenolic acids, ferulic acid, flavonoids, resveratrol, xanthohumol	Veiga et al. (2021)
112	<i>Hylocereus polyrhizus</i>	Linoleic acid	Abdullah et al. (2018)
113	<i>Ilex guayusa</i>	Caffeine, squalene, α -amyrin.	Cadena-Carrera et al. (2019)
114	<i>Inula racemose</i>	Alantolactone, isovalantolactone	Chi et al. (2016)
115	<i>Iris lactea</i>	Linoleic acid, oleic acid, docosahexaenoic acid.	Luan et al. (2020)
116	<i>Isatis tinctoria</i>	Isatin, tryptanthrin, deoxyvasicinone, isaindigotone, isaindigotidione, quinazolines, indolinone, benzodiazepine, glucoraphanin progoitrine, glucobrassicine, aromatic, aliphatic carboxylic acids	Hamburger (2002)
117	<i>Juniperus communis</i>	Sesquiterpene, diterpene alcohols, terpene oxides, ketones	Bogolitsyn et al. (2019)
118	<i>Laminaria digitata</i>	Alginate, agar, carrageenan	Abdul Khalil et al. (2018)
119	<i>Laminaria hyperborean</i>	Alginate, agar, carrageenan	Abdul Khalil et al. (2018)
120	<i>Larix sibirica</i>	Dehydroquercetin	Averyanova et al. (2018)
121	<i>Lavandula angustifolia</i>	Linalyl acetate	Gy�ri et al. (2019)
122	<i>Lavandula angustifolia</i>	Tannins, flavonols, anthocyanins	Tyskiewicz et al. (2019)
123	<i>Leucas cephalotes</i>	Oleanolic acid	Kaushik et al. (2021)
124	<i>Linum usitatissimum</i>	α -Linolenic acid, lignans, proteins, dietary fibers	Tang et al. (2021)
125	<i>Lippia graveolens</i>	Flavonoids	Arias et al. (2020)
126	<i>Lippia organoides</i>	Flavonoids	Arias et al. (2020)
127	<i>Lupinus luteus</i>	Apigenin, fisetin	Buszewski et al. (2019)
128	<i>Lycopodium clavatum</i>	Quinolizidine alkaloids, β -Carotenes, Saponins, tannins, steroids, flavonoids	Abd Hamid et al. (2018)
129	<i>Macrocystis pyrifera</i>	Alginate, agar, carrageenan	Abdul Khalil et al. (2018)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
130	<i>Mangifera indica</i>	Pectin, phenolic compounds, carotenoids (mainly all-trans- β -carotene), various vitamins	Sánchez-Camargo et al. (2019)
131	<i>Mangifera indica</i>	Mangiferin, isomangiferin, quercetin 3-O-galactoside, quercetin 3-O-glucoside, quercetin 3-O-xyloside, quercetin 3-O-arabinoside, quercetin, kaempferol	Meneses et al. (2015)
132	<i>Marrubium vulgare</i>	Marrubiin	Gavarić et al. (2021)
133	<i>Matricaria chamomilla</i>	Cycloalkane polyols	Al-Suod et al. (2019)
134	<i>Melaleuca cajuputi</i>	Caryophyllene, humulene	Kueh et al. (2018)
135	<i>Melissa officinalis</i>	Eugenol, geraniol, D-limonene, ortho-cresol	Zaid et al. (2020)
136	<i>Mitragyna speciosa</i>	Quinolizidine alkaloids, β -Carotenes, Saponins, tannins, steroids, flavonoids	Abd Hamid et al. (2018)
137	<i>Momordica cochinchinensis</i>	β -Carotene, lycopene	Kha et al. (2014)
138	<i>Moringa oleifera</i>	Quinolizidine alkaloids, β -Carotenes, Saponins, tannins, steroids, flavonoids	Abd Hamid et al. (2018)
139	<i>Moringa oleifera</i>	Gallic acid, vanillic acid, p-coumaric acid, catechin, 1-triacontanol, nonacosane, heptacosane, phytol, γ -tocopherol, α -tocopherol	da Silva et al. (2022)
140	<i>Morus nigra</i>	Phenolic acids, flavonoids	Nastić et al. (2018)
141	<i>Muricauda lutaonensis</i>	Zeaxanthin	Hameed et al. (2011)
142	<i>Musa paradisiaca</i>	Lupenone, methyl 2-hydroxy-2-(3-nitrophenyl)-2-(4-nitrophenyl)-acetate, pentacosane, 3,6,9-nonacosatriene, 10-hentriacontene, 7,23-dimethyltrtriacontane	Correa et al. (2016)
143	<i>Myrcia blanchetiana</i>	Myrciaine	de Cerqueira et al. (2013)
144	<i>Myrica rubra</i>	Diarylheptanoids	Alberti et al. (2018)
145	<i>Myrmecodia pendans</i>	Gallic acid, catechin, ferulic acid, caffeic acid, p-coumaric acid, quercetin, luteolin, kaempferol	Sanjaya et al. (2014)
146	<i>Myrtus communis</i>	Quinolizidine alkaloids, β - carotenes, Saponins, tannins, steroids, flavonoids	Abd Hamid et al. (2018)
147	<i>Narcissus poeticus</i>	Benzyl benzoate, benzyl linoleate, benzyl alcohol α -Terpineol, Limonene, (3E)-hexenol, heneicosanol, dihydroactinidiolide, 4,8,12,16-tetramethyl heptadecan-4-olide, heptanal, nonanal, (2E,4E)-decadienal, octadecanal	Baranauskienė and Venskutonis (2022)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
148	<i>Nelumbo nucifera</i>	Linoleic acid, decadieneal, linoleic acid propyl ester, 2,5-pentadecadiene-1-ol, 9-oxononanoic acid	Velikorodov et al. (2018)
149	<i>Nicotiana tabacum</i>	Nicotine, neophytadiene, 4,8,13-duvatriene-1,3-diol. Palmitic acid, stearic acid, oleic acid, linoleic acid, linolenic acid	Banožić et al. (2021)
150	<i>Nigella sativa</i>	Thymoquinone, thymol, p-cymene, chlorquinaldol, amylmetacresol, 2,4-dichlorobenzyl alcohol	Gawron et al. (2019)
151	<i>Ocimum basilicum</i>	1,8-cineole, linalool, eugenol, germacrene D, T-cadinol	Occhipinti et al. (2013)
152	<i>Ocimum basilicum</i>	Linalool, estragol	Győri et al. (2019)
153	<i>Ocimum sanctum</i>	Eugenol	Ghosh et al. (2013)
154	<i>Ocimum tenuiflorum</i>	Eugenol, eugenol acetate	Khalil et al. (2017)
155	<i>Odontonema strictum</i>	Flavonoids	Ouédraogo et al. (2018)
156	<i>Oenocarpus distichus</i>	Oleic acid, palmitic acid, linoleic acid	Cunha et al. (2019)
157	<i>Olea europaea</i>	Polyphenols	Trucillo et al. (2018)
158	<i>Olea europaea</i>	Oleuropein (OLE)	Baldino et al. (2018a)
159	<i>Olea europaea</i>	Oleuropein, luteolin-7-glucoside were the main phenolic antioxidants	Cejudo Bastante et al. (2018)
160	<i>Olea europaea</i>	Oleuropein	Uzel (2018)
161	<i>Olea europaea</i>	β -Cyclodextrin	Jaski et al. (2019)
162	<i>Opuntia ficus-indica</i>	Isorhamnetin-3-O-glucosyl-rhamnosyl-rhamnoside, isorhamnetin-3-O-glucosyl-rhamnosyl-pentoside, isorhamnetin-3-O-glucosyl-rhmanoside	Antunes-Ricardo et al. (2018)
163	<i>Orbignya phalerata</i>	Lauric acid, oleic acid, lauric acid	de Oliveira et al. (2019)
164	<i>Origanum majorana</i>	Cis-sabinene hydrate	Busatta et al. (2017)
165	<i>Origanum vulgare</i>	Cis-sabinene hydrate	Busatta et al. (2017)
166	<i>Origanum vulgare</i>	α -Linolenic acid, palmitic acid, oleic acid, linoleic acid, carvacrol, heneicosane, nonacosane, docosane, borneol, thymol	García-Pérez et al. (2019)
167	<i>Oroxylum indicum</i>	Phenolic acids, flavonoids, tannins, alkaloids	Chhouk et al. (2018)
168	<i>Orthosiphon aristatus</i>	Rosmarinic acid, eurycomanone, andrographolide	Abd Aziz et al. (2021)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
169	<i>Orthosiphon stamineus</i>	Sinensetin	Aziz et al. (2018)
170	<i>Orthosiphon stamineus</i>	Sinensetin, Isosinensetin, Rosmarinic Acid	Abdul Aziz et al. (2020)
171	<i>Parthenium argentatum</i>	Terpenoids, phenolics, alkaloids, sterols, fatty acids/triglycerides	Dehghanizadeh and Brewer (2020)
172	<i>Passiflora mucronate</i>	β -amyrin, β -sitosterol, stigmasterol, oleanolic acid	da Silva et al. (2020)
173	<i>Petroselinum crispum</i>	Apigenin	Saotome and Imai (2018)
174	<i>Physalis angulate</i>	Trinitrobenzenesulphonic acid	Almeida Jr. et al. (2017)
175	<i>Picea Abies</i>	Methyl dehydroabietate	Burčová et al. (2018)
176	<i>Picea abies</i>	Catechin, dihydroquercetin, astringin, isorhaptin	Ferrentino et al. (2021)
177	<i>Pimpinella anisum</i>	Sterols	Bettaieb Rebey et al. (2019)
178	<i>Piper amalago</i>	Pyrrolidine Alkaloid	Carrara et al. (2017)
179	<i>Piper betle</i>	Phenolic compounds	Pise et al. (2022)
180	<i>Piper betle</i>	Tannins, quercetin, eugenol, hydroxychavicol, chavibetol	Azahar et al. (2020)
181	<i>Piper hispidum</i>	Cinnamoyl pyrrolidine amides	Lima et al. (2020)
182	<i>Piper klotzschianum</i>	Germacrene D, pipercollosidine, 14-oxy- α -muuroleno, bicyclogermacrene, (E)-caryophyllene	Lima et al. (2019)
183	<i>Piper nigrum</i>	Eugenol, eugenol acetate	Khalil et al. (2017)
184	<i>Piper nigrum</i>	Piperine, piperlonguminine, piperanine, pipercollosine, dehydropiperonaline, piperonatinine, retrofractamide B, pellitorine, guineensine	Yu et al. (2022)
185	<i>Pistacia lentiscus</i>	α -Pinene, terpinene-4-ol	Aydi et al. (2020)
186	<i>Pistacia vera</i>	α -Pinene, β -myrcene, limonene-D, α -terpinolene	Demirkoz et al. (2018)
187	<i>Pleurotus ostreatus</i>	Heteropolysaccharides, β -glucans, α -glucans, oligosaccharides	Barbosa et al. (2020)
188	<i>Pongamia pinnata</i>	Oleic acid, arachidic acid, cis-10-pentadecenoic acid, stearic acid, cis-8,11,14-Eicosatrienoic acid, linolenic acid, gamma(γ)-linolenic acid, cis-11-Eicosenoic acid	Suryawanshi and Mohanty (2018)
189	<i>Populus balsamifera</i>	Pinostrobin, tectochrysin, pinocembrin, chrysin	Adekenov et al. (2020)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
190	<i>Prunus armeniaca</i>	Tocopherols, Amygdalin, Fatty Acids	Pavlović et al. (2018)
191	<i>Punica granatum</i>	Punicic acid, tocopherols, phytosterols, triterpenes, phospholipids, quercetin, epicatechin, catechins, delphinidin, pelargonidin, cyanidin, punicalagin, punicalin, gallic acids, caffeic acids, chlorogenic acids	El-Shamy and Farag (2021)
192	<i>Punica granatum</i>	Punicic acid, linoleic acid, oleic acids	Khoddami et al. (2014)
193	<i>Putranjiva roxburghii</i>	β -Sitosterol, oleic acid, linoleic acid	Balkrishna et al. (2021)
194	<i>Rhodiola rosea</i>	Salidroside, rhodioloside B, rhodioloside C, rhodiosin, luteolin, catechin, quercetin, quercitrin, herbacetin, sacranoside A, vimalin, dihydroquercetin, acacetin, mearnsetin, taxifolin-O-pentoside, tricetin trimethyl ether 7-O-hexosyl-hexoside, tricetin 7-O-glucuronyl-O-hexoside, tricetin O-pentoside, tricetin-O-dihexoside, eriodictyol-7-O-glucoside; flavan-3-ols: gallocatechin, hydroxycinnamic acid, caffeoylmalic acid, di-O-caffeoylquinic acid, esculetin, esculin, fraxin, lignans: hinokinin, pinoresinol, L-ascorbic acid, glucaric acid, palmitic acid, linolenic acid	Zakharenko et al. (2021)
195	<i>Rhus punjabensis</i>	Dihydrofisetin	Dong et al. (2020)
196	<i>Rosa canina</i>	Linoleic acid, linolenic acid, palmitic acid, stearic acid	Jahongir et al. (2019)
197	<i>Rosa damascene</i>	Citronellol, geraniol, nerol, nonadecane, nonadecene, heneicosane, heptadecane	Antonova et al. (2021)
198	<i>Roselle calyces</i>	Anthocyanins	Idham et al. (2021)
199	<i>Rosmarinus eriocalyx</i>	β -Amyrin, camphor, tetradecenoic acid, linolenic acid	Bendif et al. (2018c)
200	<i>Rosmarinus officinalis</i>	Carnosic acid, carnosol, rosmanol, genkwanin, cirsimaritin, homoplantagin, ursolic acid	Sharifi-Rad et al. (2020)
201	<i>Rosmarinus officinalis</i>	Carnosic acid, carnosol, methyl carnosate, rosmanol, rosmarinic acid. Moreover, carnosic acid, carnosol	Fornari et al. (2014)
202	<i>Rosmarinus officinalis</i>	Verbenone, cirsimaritin, salvigenin, carnosol, carnosic acid	Kuo et al. (2011)
203	<i>Rosmarinus officinalis</i>	Essential oils, phenolic compounds	Ali et al. (2019)
204	<i>Rosmarinus officinalis</i>	Palmitic acid, α -linolenic acid, linoleic acid, oleic acid, stearic acid, d-camphor, eicosane, 1,8-cineole, tetracosane, borneol, β -caryophyllene	García-Pérez et al. (2020)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
205	<i>Rubia tinctorum</i>	Alizarin, lucidin, rubiadin	Yekefallah and Raofie (2022)
206	<i>Rubus idaeus</i>	Fatty acids, tocopherols	Marić et al. (2020)
207	<i>Ruellia angustiflora</i>	Fatty acids, triterpenes, tetraterpenes, tocopherols, phytosterols	Pires et al. (2021)
208	<i>Saccharum officinarum</i>	Alcohols, esters, hydrocarbons, ketones, aldehydes	Ahmed Baloch et al. (2018)
209	<i>Saccharum officinarum</i>	Long-chain fatty alcohols, phytosterols	Albarelli et al. (2018)
210	<i>Salvia hispanica</i>	Linoleic acid, a-linolenic acid, tocopherols, polyphenols	Ixtaina et al. (2014)
211	<i>Salvia hispanica</i>	Squalene, sterols, tocopherols, polyphenols, carotenoids	Dąbrowski et al. (2018)
212	<i>Salvia officinalis</i>	1,8-cineole, α - β -thujone, camphor, α -humulene, viridiflorol, manool	Jokić et al. (2018)
213	<i>Salvia officinalis</i>	Carnosic acid, carnosol	Pavić et al. (2019)
214	<i>Salvia Rosmarinus</i>	α -Pinene	Allawzi et al. (2019)
215	<i>Salvia Rosmarinus</i>	Carnosic acid, rosmarinic acid, carotenoids, chlorophyll	Lefebvre et al. (2021)
216	<i>Salvia viridis</i>	Vanillin, Ethyl syringate, Syringaldehyde (3,5-Dimethoxy-4-hydroxybenzaldehyde), Antiarol (3,4,5-Trimethoxyphenol), Indole-4-carbaldehyde, Coumarin, Coniferyl aldehyde (4-Hydroxy-3-methoxycinnamaldehyde), N-(2-Phenylethyl)acetamide, Sinapyl aldehyde (3,5-Dimethoxy-4-hydroxycinnamaldehyde), Dimethoxy-trihydroxy(iso)flavone isomer 1, Dihydroxy-dimethoxy(iso)flavone, Dimethoxy-trihydroxy(iso)flavone isomer 2, Genkwanin, Dihydroxy-trimethoxy(iso)flavone, Hydroxy-trimethoxy(iso)flavone, Hydroxy-tetramethoxy(iso)flavone, 1-Oxomicrosteviol, Viroxocin, Apigenin-4',7-dimethyl ether (4',7-Dimethoxy-5-hydroxyflavone), 3-Oxomicrosteviol, Hexadecanedioic acid, Viridoquinone	Zengin et al. (2019)
217	<i>Sambucus nigra</i>	Quercetin, kaempferol, rutin	Anusha Siddiqui et al. (2022)
218	<i>Satureja montana</i>	Thymol, carvacrol, γ -terpinene, p-cymene	Damjanović-Vratnica et al. (2016)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
219	<i>Saururus chinensis</i>	Aurantiamide acetate, echinuline, (–)-(7R, 8R) -7-O-acetylpolysphorin, elemicin, isoelemicin, 1, 4-bis (3, 4-dimethoxyphenyl) 2, 3-dimethyl-1, 4-butanedione, saucerneol D, (2R) -3-(3', 4', 5'-trimethoxyphenyl) -1, 2-propanediol, grandisin, rel-(7R, 8R, 7'R, 8'R) 3', 4'- methylenedioxy-3, 4, 5, 5'-tetramethoxy-7, 7 -epoxylignan, zanthopyranone, (±) -eritro-1-(3, 4, 5-trimethoxy) -1, 2 -propanodiol, threo-3, 4, 5-trimethoxy-7-hydroxy-1'-allyl-3', 5'-dimethoxy-8. O. 4'-neolignan, (+) -(8R) -(2, 6 -dimethoxy-4-propenylphenoxy) -1-(3, 4, 5-trimethoxyphenyl) propan-1-one, meso-dihydroguaiaietic acid, (–) -galbacin, (–) - (7R, 8R) -7-O-acetylraphidecurisinol B	Chen et al. (2018b)
220	<i>Scenedesmus almeriensis</i>	Lutein	Mehariya et al. (2019)
221	<i>Schinus terebinthifolia</i>	Germacrene D, sabinene, β phellandrene, α- phellandrene	Andrade et al. (2017)
222	<i>Schinziophyton rautanenii</i>	Campesterol, stigmasterol, β-sitosterol, Δ5- avenasterol, 22-dihydrospinasterol, Δ7-avenasterol, lanosterol, Δ5,23-stigmastadienol, Δ7-campesterol, clerosterol, obtusifoliol, Δ 5,24(25)-stigmastadienol, α-amyrin, gramisterol, cycloeucalenol, cycloartenol, stigmasta-8,24-dienol-3-β-ol, 28-methylbtusifoliol, 24-methylenecycloartenol, citrostadienol, β-sitosterol, Δ5-avenasterol, campesterol.	Gwatidzo et al. (2014)
223	<i>Serenoa repens</i>	Fatty acids, beta-sitosterol, fatty alcohols	Bartolomé Ortega et al. (2017)
224	<i>Sesamum indicum</i>	Sesamin, sesaminol, sesamolinal	Hu et al. (2004)
225	<i>Sesamum indicum</i>	γ-Tocopherol, lignan	Shi et al. (2018)
226	<i>Sesamum indicum</i>	Sesamin, sesamolinal, tocopherols, linoleic acid, oleic acid	Buranachokpaisan et al. (2021)
227	<i>Sida rhombifolia</i>	Isoquercitin	Ferro et al. (2019)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
228	<i>Sideritis sipylea</i>	β -Caryophyllene, α -Humulene, 9-epi-(E)-Caryophyllene, Germacrene D, Bicyclogermacrene, cis-Sesquisabinene hydrate, Spathulenol, Caryophyllene oxide, Humulene epoxide II, (E)-Sesquilavandulyl acetate, Cyclopentadecanolide, Hexahydrofarnesyl acetone, (Z)-Lanceol acetate, Isopimara-9(11),15-diene, Totarene, Beyerene, Geranyl- α -terpinene, Geranyl-p-cymene, (Z,Z)-Geranyl linalool, Dolabradiene, Sclarene, (E,Z)-Geranyl linalool, (Z,E)-Geranyl linalool, 13-epi-Dolabradiene, 13-epi-Manool oxide, (E,E)-Geranyl linalool, Manool, 13-epi-Manool, Phytol, Abienol, Abieta-8(14),13(15)-diene, Sandaracopimarinal, Sclareol, 7- α -hydroxy-Manool, 3- α -hydroxy-Manool, Isopimarol, Sideridiol, n-Hexacosane, 7-Epicandicandiol, Siderol, n-Heptacosane, n-Octacosane, n-Nonacosane, Sidol, Sesquiterpene hydrocarbons, Oxygenated sesquiterpenes, Diterpene hydrocarbons, Oxygenated diterpenes, lkanes	Axiotis et al. (2020)
229	<i>Solanum lycopersicum</i>	Polyphenols, flavonoids, lycopenes, carotenoids	Haddadin and Haddadin (2015)
230	<i>Solanum lycopersicum</i>	Lycopene	Inakuma (2015)
231	<i>Solanum lycopersicum</i>	Lycopene, β -carotene	Cante et al. (2022)
232	<i>Solanum lycopersicum</i>	Lycopene	Reverchon et al. (2022)
233	<i>Solanum lycopersicum</i>	α -Tocopherol, γ -tocopherol, lycopene, β -carotene	Romano et al. (2020)
234	<i>Solanum viarum</i>	1,2-Benzenedicarboxylic acid, quinic acid, octadecenoic acid, solasodine	Confortin et al. (2019)
235	<i>Sophora flavescens</i>	Genistein	Han and Kang (2015)
236	<i>Sorbus aucuparia</i>	Linoleic acid, oleic acid, palmitic acid	Bobinaité et al. (2020)
237	<i>Sorghum bicolor</i>	Linoleic acid, decadieneal, linoleic acid propyl ester, 2,5-pentadecadiene-1-ol, 9-oxononanoic acid	Velikorodov et al. (2018)
238	<i>Spina gleditsiae</i>	Saponins	Liu (2018)
239	<i>Spinacia oleracea</i>	Lutein, chlorophyll	Derrien et al. (2018)
240	<i>Spinacia oleracea</i>	Phenolics	Lee et al. (2018)

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Table 1 (continued)

SI no	Species name	Chemicals/phytochemicals	Ref
241	<i>Stellera chamaejasme</i>	Hexanedioic acid, bis(2-ethylhexyl) ester, π sitosterol, 7-methyl-Z-tetradecen-1-ol acetate, 9-hexadecenoic acid-hexadecyl ester (Z), 1,2- benzenedicarboxylic acid-diisooctyl ester, (3 π 24Z) stigmasta-5,24(28)-dien-3-ol, stigmastan-3,5-diene, squalene	Bai et al. (2012)
242	<i>Stevia rebaudiana</i>	Polyphenols, chlorophylls, carotenoids	Bursac Kovačević et al. (2018)
243	<i>Sucupira branca</i>	Alpha-humulene, beta-caryophyllene, alpha-copaene, (-)-beta-elemene, (E)-germacrene D(-)-gamma-elemene, spathulenol	Chañi-Paucar et al. (2022)
244	<i>Swietenia mahagoni</i>	Linoleic acid	Hartati et al. (2018)
245	<i>Syzygium aromaticum</i>	Eugenol, eugenol acetate	Idowu et al. (2021)
246	<i>Syzygium aromaticum</i>	Eugenol, chavicol, n-pentacosane, hexacosanal, vitamin E	Frohlich et al. (2019)
247	<i>Syzygium aromaticum</i>	Eugenol, caryophyllene, eugenol acetate	Györi et al. (2019)
248	<i>Syzygium aromaticum</i>	Eugenyl acetate, β -caryophyllene, α -humulene	Haro-González et al. (2021)
249	<i>Syzygium campanulatum</i>	Flavanones, chalcone, triterpenoids	Memon et al. (2016)
250	<i>Tagetes erecta</i>	Lutein	Pal and Bhattacharjee (2018)
251	<i>Tanacetum parthenium</i>	Parthenolide, sudachitin, aceronin, nevadensin	Végh et al. (2018)
252	<i>Teucrium polium</i>	Germacrene D, β -eudesmol, shyobunol, δ -cadinene	Bendif et al. (2018b)
253	<i>Theobroma cacao</i>	Polyphenols, mainly procyanidins, flavan-3-ols	Hernández et al. (2019)
254	<i>Thymus mastichina</i>	Thymol, α -terpinene, p-cymene	Kessler et al. (2022)
255	<i>Thymus munbyanus</i>	Tocopherol	Bendif et al. (2018a)
256	<i>Thymus vulgaris</i>	Chlorophyll b, chlorophyll a	Hamdan and Daood (2011)
257	<i>Trachyspermum ammi</i>	Thymol, o-Cymene, γ -Terpinene, 2-methyl-5-(1-methylethyl)-phenol	Bhatt et al. (2018)
258	<i>Trifolium pratense</i>	Isoflavonoids (3-phenyl chromones), flavonoids (2-phenyl chromones)	Klejdus et al. (2005)
259	<i>Triticum Vulgare</i>	Tocopherol	Özcan and Ören (2019)

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Table 1 (continued)

Sl no	Species name	Chemicals/phytochemicals	Ref
260	<i>Vaccinium Meridionale</i>	Anthocyanins (ACNs)	Colorado et al. (2020)
261	<i>Vaccinium myrtillus</i>	Anthocyanins, flavonols, tocopherols. Polyunsaturated fatty acids, vitamin E.	Gustinelli et al. (2018)
262	<i>Viburnum opulus</i>	β -Sitosterol, α -tocopherol	Dienaitè et al. (2021)
263	<i>Viburnum opulus</i>	Phenolic acids, iridoids, quercetin, (epi) catechina, flavalignans, procyanidins, anthocyanins	Dienaitè et al. (2020)
264	<i>Viola surinamensis</i>	Steroids, terpenes, coumarins, phenolics	Cordeiro et al. (2019)
265	<i>Vitis vinifera</i>	1-Hexacosanol, 1-octacosanol, 1-triacontanol, α -tocopherol, β -sitosterol, β -amyirin	de Melo et al. (2020)
266	<i>Xanthium strumarium</i>	Linoleic acid, decadieneal, linoleic acid propyl ester, 2,5-pentadecadiene-1-ol, 9-oxononanoic acid	Velikorodov et al. (2018)
267	<i>Xinjiang jujube</i>	Quercetin-3-O-robinobioside, Rutin (Quercetin-3-O-rutinoside), Hyperoside (Quercetin-3-O- β -d-galactoside), Quercetin-3-O- β -d-glucoside, Kaempferol-3-O-robinobioside, Kaempferol-3-O-glucoside, Quercetin-3-O- β -l-arabinosyl-(1 \rightarrow 2)- α -l-rhamnoside, Quercetin-3-O- β -d-xylosyl-(1 \rightarrow 2)- α -l-rhamnoside.	Song et al. (2019)
268	<i>Zingiber officinale</i>	α -Zingiberene	de Souza Junior et al. (2020)
269	<i>Zingiber officinale</i>	6-Gingerol	Gan et al. (2016)

References

- 2017 Supercritical Fluid Extraction. Natural Food Flavors and Colorants
- Abd Aziz NA, Hasham R, Sarmidi MR, Suhaimi SH, Idris MKH (2021) A review on extraction techniques and therapeutic value of polar bioactives from Asian medicinal herbs: case study on *Orthosiphon aristatus*, *Eurycoma longifolia* and *Andrographis paniculata*. *Saudi Pharm J* 29:143–165
- Abd Hamid IA, Ismail N, Abd Rahman N (2018) Supercritical carbon dioxide extraction of selected herbal leaves: an overview. In: 3rd International Conference on Global Sustainability and Chemical Engineering, ICGSCE 2017
- Abdul Aziz AH, Putra NR, Kong H, che yunus, M. A. (2020) Supercritical carbon dioxide extraction of sinensetin, isosinensetin, and rosmarinic acid from *Orthosiphon stamineus* leaves: optimization and modeling. *Arab J Sci Eng* 45:7467–7476

- Abdul Khalil HPS, Lai TK, Tye YY, Rizal S, Chong EWN, Yap SW, Hamzah AA, Nurul Fazita MR, Paridah MT (2018) A review of extractions of seaweed hydrocolloids: properties and applications. *Express Polym Lett* 12:296–317
- Abdullah A, Gani SSA, Mokhtar NFM, Hin TYY, Haiyee ZA, Mustafa S (2018) Supercritical carbon dioxide extraction of red pitaya (*Hylocereus polyrhizus*) seeds: response surface optimization, fatty acid composition and physicochemical properties. *Malays Appl Biol* 47:39–46
- Adekenov SM, Baysarov GM, Khabarov IA, Polyakov VV (2020) Flavonoids of populus balsamifera L. Buds and methods for their isolation. *Khimiya Rastitel'nogo Syr'ya*:181–188
- Ahmed Baloch H, Nizamuddin S, Siddiqui MTH, Mubarak NM, Dumbre DK, Srinivasan MP, Griffin GJ (2018) Sub-supercritical liquefaction of sugarcane bagasse for production of bio-oil and char: effect of two solvents. *J Environ Chem Eng* 6:6589–6601
- AIP Conference Proceedings (2018) 4th International Conference on Engineering, Technology, and Industrial Application: Human-Dedicated Sustainable Product and Process Design: Materials, Resources, and Energy, ICETIA 2017
- Albarelli JQ, Santos DT, Meireles MAA (2018) Thermo-economic evaluation of a new approach to extract sugarcane wax integrated to a first and second generation biorefinery. *Biomass Bioenergy* 119:69–74
- Alberti Á, Riethmüller E, Béni S (2018) Characterization of diarylheptanoids: an emerging class of bioactive natural products. *J Pharm Biomed Anal* 147:13–34
- Alekseev ES, Alentiev AY, Belova AS, Bogdan VI, Bogdan TV, Bystrova AV, Gafarova ER, Golubeva EN, Grebenik EA, Gromov OI, Davankov VA (2020) Supercritical fluids in chemistry. *Russ Chem Rev* 89:1337–1427
- Ali A, Chua BL, Chow YH (2019) An insight into the extraction and fractionation technologies of the essential oils and bioactive compounds in *Rosmarinus officinalis* L.: past, present and future. *TrAC* 118:338–351
- Allawzi M, Allaboun H, Almasri A (2019) CO₂ supercritical extraction of essential oil of Jordanian rosemary. *J AOAC Int* 102:662–665
- Almeida LD Jr, Quaglio AEV, de Almeida Costa CAR, di Stasi LC (2017) Intestinal anti-inflammatory activity of ground cherry (*Physalis angulata* L.) standardized CO₂ phytopharmaceutical preparation. *World J Gastroenterol* 23:4369–4380
- Al-Suod H, Ratiu IA, Krakowska-Sieprawska A, Lahuta L, Górecki R, Buszewski B (2019) Supercritical fluid extraction in isolation of cyclitols and sugars from chamomile flowers. *J Sep Sci* 42:3243–3252
- Andrade KS, Poncelet D, Ferreira SRS (2017) Sustainable extraction and encapsulation of pink pepper oil. *J Food Eng* 204:38–45
- Antonova DV, Medarska YN, Stoyanova AS, Nenov NS, Slavov AM, Antonov LM (2021) Chemical profile and sensory evaluation of Bulgarian rose (*Rosa damascena* Mill.) aroma products, isolated by different techniques. *J Essent Oil Res* 33:171–181
- Antunes-Ricardo M, García-Cayuela T, Mendiola JA, Ibañez E, Gutiérrez-Urbe JA, Cano MP, Guajardo-Flores D (2018) Supercritical CO₂ enzyme hydrolysis as a pretreatment for the release of isorhamnetin conjugates from *Opuntia ficus-indica* (L.) Mill. *J Supercrit Fluids* 141:21–28
- Anusha Siddiqui S, Redha AA, Esmaeili Y, Mehdizadeh M (2022) Novel insights on extraction and encapsulation techniques of elderberry bioactive compounds. *Crit Rev Food Sci Nutr*:1
- Aponte-Buitrago R, Mayorga-Wandurraga H, Moreno-Murillo B (2017) Flavonols and sesquiterpenoids from outer bark and leaves of *Croton Polycarpus* Benth. (Euphorbiaceae). *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromaticas* 16:471–485
- Arias J, Mejía J, Córdoba Y, Martínez JR, Stashenko E, del Valle JM (2020) Optimization of flavonoids extraction from *Lippia graveolens* and *Lippia organoides* chemotypes with ethanol-modified supercritical CO₂ after steam distillation. *Ind Crop Prod* 146:112170
- Arumugam S, Thiruganasambantham R (2018) Standardized supercritical CO₂ extract of *acanthus ilicifolius* (Linn.) leaves inhibits the pro-inflammatory cytokine tumor necrosis factor-A

- in lipopolysaccharide-activated murine raw 264.7 macrophage cells. *Asian J Pharm Clin Res* 11:193–198
- Attia IB, Zucca P, Marincola FC, Piras A, Rosa A, Chaieb M, Rescigno A (2018) Chemical composition and antioxidant potential differences between *Cynomorium coccineum* L. growing in Italy and in Tunisia: effect of environmental stress. *Diversity* 10
- Averyanova EV, Shkolnikova MN, Tsyganok SN, Shakura VA (2018) Intensification of the process of ultrasonic extraction of dehydroquercetin from wood waste. In: 19th International Conference of Young Specialists on Micro/Nanotechnologies and Electron Devices, EDM 2018. 312–317
- Axiotis E, Petrakis EA, Halabalaki M, Mitakou S (2020) Phytochemical profile and biological activity of endemic *Sideritis siphylea* Boiss. In North Aegean Greek islands. *Molecules* 25
- Aydi A, Zibetti AW, Al-Khazaal AZ, Eladeb A, Adberraba M, Barth D (2020) Supercritical CO₂ extraction of extracted oil from *Pistacia lentiscus* L.: mathematical modeling, economic evaluation and scale-up. *Molecules* 25
- Ayub MA, Hanif MA, Sarfraz RA, Shahid M (2018) Biological activity of *boswellia serrata* roxb. Oleo gum resin essential oil: effects of extraction by supercritical carbon dioxide and traditional methods. *Int J Food Prop* 21:808–820
- Azahar NI, Mokhtar NM, Arifin MA (2020) Piper betle: a review on its bioactive compounds, pharmacological properties, and extraction process. In: 5th International Conference of Chemical Engineering and Industrial Biotechnology, ICCEIB 2020
- Aziz AHA, Yunus MAC, Yian LN, Idham Z, Rithwan F, Hadzri HM, Mustapha AN (2018) Enhancement and optimization of sinensetin extract from orthosiphon stamineus using supercritical carbon dioxide extraction. *Malaysian J Anal Sci* 22:867–876
- Bai XN, Cheng J, Liang W, Ma LQ, Liu YB, Shi GL, Wang YN (2012) Antifungal activity of extracts by supercritical carbon dioxide extraction from roots of *Stellera chamaejasme* L. and analysis of their constituents using GC-MS. In: 2011 International Conference on Information Technology and Agricultural Engineering, ICITAE 2011. Sanya
- Bajer T, Surmová S, Eisner A, Ventura K, Bajerová P (2018) Use of simultaneous distillation-extraction, supercritical fluid extraction and solid-phase microextraction for characterisation of the volatile profile of *Dipteryx odorata* (Aubl.) Willd. *Ind Crop Prod* 119:313–321
- Bakshi RA, Sodhi NS, Wani IA, Khan ZS, Dhillon B, Gani A (2022) Bioactive constituents of saffron plant: extraction, encapsulation and their food and pharmaceutical applications. *Appl Food Res* 2:100076
- Baldino L, Reverchon E (2018) *Artemisia annua* organic solvent extract, processed by supercritical CO₂. *J Chem Technol Biotechnol* 93:3171–3175
- Baldino L, Della Porta G, Osseo LS, Reverchon E, Adami R (2018a) Concentrated oleuropein powder from olive leaves using alcoholic extraction and supercritical CO₂ assisted extraction. *J Supercrit Fluids* 133:65–69
- Baldino L, Scognamiglio M, Reverchon E (2018b) Extraction of rotenoids from *Derris elliptica* using supercritical CO₂. *J Chem Technol Biotechnol* 93:3656–3660
- Balkrishna A, Nain P, Joshi M, Khandrika L, Varshney A (2021) Supercritical fluid extract of *putranjiva roxburghii* wall. Seeds mitigates fertility impairment in a zebrafish model. *Molecules* 26
- Balkrishna A, Nain P, Joshi M, Kumar B, Varshney A (2022) Super-critical fluid extract of *Bryonopsis laciniosa* (Shivlingi) seeds restores fertility in zebrafish models through revival of cytological and anatomical features. *J Ovarian Res* 15
- Banožić M, Gagić T, Čolnik M, Knez Ž, Škerget M, Jerković I, Jokić S (2021) Sequence of supercritical CO₂ extraction and subcritical H₂O extraction for the separation of tobacco waste into lipophilic and hydrophilic fractions. *Chem Eng Res Des* 169:103–115
- Baranauskienė R, Venskutonis PR (2022) upercritical CO₂ Extraction of *Narcissus poeticus* L. Flowers for the Isolation of Volatile Fragrance Compounds. *Molecules* 27
- Barbosa JR, S. Freitas MM, Oliveira LC, S. Martins LH, Almada-Vilhena AO, Oliveira RM, Pieczarka JC, B. Brasil DDS, Carvalho Junior RN (2020) Obtaining extracts rich in antioxidant

- polysaccharides from the edible mushroom *Pleurotus ostreatus* using binary system with hot water and supercritical CO₂. *Food Chem* 330:127173
- Barrales FM, Silveira P, Barbosa P, Ruviano AR, Paulino BN, Pastore GM, Macedo GA, Martínez J (2018) Recovery of phenolic compounds from citrus by-products using pressurized liquids — an application to orange peel. *Food Bioprod Process* 112:9–21
- Bartolomé Ortega A, Cano Calvo A, Szekely E, Škerget M, Knez Ž (2017) Supercritical fluid extraction from Saw Palmetto berries at a pressure range between 300 bar and 450 bar. *J Supercrit Fluids* 120:132–139
- Barzotto ILM, Santos KA, da Silva EA, Sene AC, da Silva NS, Vieira L (2019) Supercritical extraction of *Eugenia involucrata* leaves: influence of operating conditions on yield and A-tocopherol content. *J Supercrit Fluids* 143:55–63
- Bayrak S, Sökmen M, Aytaç E, Sökmen A (2019) Conventional and supercritical fluid extraction (SFE) of colchicine from *Colchicum speciosum*. *Ind Crop Prod* 128:80–84
- Bendif H, Adouni K, Miara MD, Baranauskienė R, Kraujalis P, Venskutonis PR, Nabavi SM, Maggi F (2018a) Essential oils (EOs), pressurized liquid extracts (PLE) and carbon dioxide supercritical fluid extracts (SFE-CO₂) from Algerian thymus *munbyanus* as valuable sources of antioxidants to be used on an industrial level. *Food Chem* 260:289–298
- Bendif H, Lazali M, Souilah N, Miara MD, Kazernavičiūtė R, Baranauskienė R, Venskutonis PR, Maggi F (2018b) Supercritical CO₂ extracts and essential oils from *Teucrium polium* L. growing in Algeria: chemical composition and antioxidant activity. *J Essent Oil Res* 30:488–497
- Bendif H, Miara MD, Kalboussi Z, Grauzdytė D, Povilaitis D, Venskutonis PR, Maggi F (2018c) Supercritical CO₂ extraction of *Rosmarinus eriocalyx* growing in Algeria: chemical composition and antioxidant activity of extracts and their solid plant materials. *Ind Crop Prod* 111:768–774
- Benito-Román O, Rodríguez-Perrino M, Sanz MT, Melgosa R, Beltrán S (2018) Supercritical carbon dioxide extraction of quinoa oil: study of the influence of process parameters on the extraction yield and oil quality. *J Supercrit Fluids* 139:62–71
- Bettaieb Rebey I, Bourgou S, Detry P, Wannes WA, Kenny T, Ksouri R, Sellami IH, Fauconnier ML (eds) (2019) Green extraction of fennel and Anise edible oils using bio-based solvent and supercritical fluid: assessment of chemical composition, antioxidant property, and oxidative stability. *Food Bioprocess Technol* 12:1798–1807
- Bezerra FWF, Costa WAD, Oliveira MSD, Aguiar Andrade EHD, Carvalho RNDJ (2018) Transesterification of palm pressed-fibers (*Elaeis guineensis* Jacq.) oil by supercritical fluid carbon dioxide with entrainer ethanol. *J Supercrit Fluids* 136:136–143
- Bhatt V, Mahesh Kumar M, Periyar Selvam S (2018) Antimicrobial effect of ajwain seed ethanolic extract against food borne pathogenic bacteria. *Int Food Res J* 25:908–912
- Bizaj K, Škerget M, Košir IJ, Knez Ž (2021) Sub- and supercritical extraction of slovenian hops (*Humulus lupulus* l.) aurora variety using different solvents. *Plan Theory* 10
- Bobinaitė R, Kraujalis P, Tamkutė L, Urbonavičienė D, Viškelis P, Venskutonis PR (2020) Recovery of bioactive substances from rowanberry pomace by consecutive extraction with supercritical carbon dioxide and pressurized solvents. *J Ind Eng Chem* 85:152–160
- Bogolitsyn K, Krasikova A, Gusakova M, Ivakhnov A, Gravitis J (2019) Selective extraction of terpenoid compounds of *Juniperus communis* L. wood in the medium of a binary solvent (supercritical CO₂ with modifier). *Phytochem Anal* 30:609–616
- Bouazzaoui N, Bouajila J, Camy S, Mulengi JK, Condore JS (2018) Fatty acid composition, cytotoxicity and anti-inflammatory evaluation of melon (*Cucumis melo* L. *Inodorus*) seed oil extracted by supercritical carbon dioxide. *Sep Sci Technol (Philadelphia)* 53:2622–2627
- Buranachokpaisan K, Muangrat R, Chalermchat Y (2021) Supercritical CO₂ extraction of residual oil from pressed sesame seed cake: optimization and its physicochemical properties. *J Food Process Preserv* 45
- Burčová Z, Kreps F, Greifová M, Jablonský M, Ház A, Schmidt Š, Šurina I (2018) Antibacterial and antifungal activity of phytosterols and methyl dehydroabietate of Norway spruce bark extracts. *J Biotechnol* 282:18–24

- Bursać Kovačević D, Maras M, Barba FJ, Granato D, Roohinejad S, Mallikarjunan K, Montesano D, Lorenzo JM, Putnik P (2018) Innovative technologies for the recovery of phytochemicals from *Stevia rebaudiana* Bertoni leaves: A review. *Food Chem* 268:513–521
- Busatta C, Barbosa J, Cardoso RI, Paroul N, Rodrigues M, Oliveira D, Oliveira JV, Cansian RL (2017) Chemical profiles of essential oils of marjoram (*Origanum majorana*) and oregano (*Origanum vulgare*) obtained by hydrodistillation and supercritical CO₂. *J Essent Oil Res* 29:367–374
- Buszewski B, Rafińska K, Cvetanović A, Walczak J, Krakowska A, Rudnicka J, Zeković Z (2019) Phytochemical analysis and biological activity of *Lupinus luteus* seeds extracts obtained by supercritical fluid extraction. *Phytochem Lett* 30:338–348
- Cadena-Carrera S, Tramontin DP, Bella Cruz A, Bella Cruz RC, Müller JM, Hense H (2019) Biological activity of extracts from guayusa leaves (*Ilex guayusa* Loes.) obtained by supercritical CO₂ and ethanol as cosolvent. *J Supercrit Fluids* 152:104543
- Camel V (2001) Recent extraction techniques for solid matrices—supercritical fluid extraction, pressurized fluid extraction and microwave-assisted extraction: their potential and pitfalls. *Analyst* 126:1182–1193
- Cante RC, Gallo M, Variiale L, Garella I, Nigro R (2022) Recovery of carotenoids from tomato pomace using a hydrofluorocarbon solvent in sub-critical conditions. *Appl Sci (Switzerland)* 12
- Carmona PAO, Garcia LC, Ribeiro JAA, Valadares LF, Marçal AF, de França LF, Mendonça S (2018) Effect of solids content and spray-drying operating conditions on the carotenoids microencapsulation from pressed palm fiber oil extracted with supercritical CO₂. *Food Bioprocess Technol* 11:1703–1718
- Carrara VS, Filho LC, Garcia VAS, Faiões VS, Cunha-Júnior EF, Torres-Santos EC, Cortez DAG (2017) Supercritical fluid extraction of pyrrolidine alkaloid from leaves of *Piper amalago* L. *Evid-based Complement Alternat Med* 2017
- Casas MP, López-Hortas L, Díaz-Reinoso B, Moure A, Domínguez H (2021) Supercritical CO₂ extracts from *Acacia dealbata* flowers. *J Supercrit Fluids* 173:105223
- Cejudo Bastante C, Casas Cardoso L, Fernández Ponce MT, Mantell Serrano C, Martínez de la Ossa-Fernández EJ (2018) Characterization of olive leaf extract polyphenols loaded by supercritical solvent impregnation into PET/PP food packaging films. *J Supercrit Fluids* 140:196–206
- Chan YH, Yusup S, Quitain AT, Chai YH, Uemura Y, Loh SK (2018) Extraction of palm kernel shell derived pyrolysis oil by supercritical carbon dioxide: evaluation and modeling of phenol solubility. *Biomass Bioenergy* 116:106–112
- Chañi-Paucar LO, Johner JCF, Zabot GL, Meireles MAA (2022) Technical and economic evaluation of supercritical CO₂ extraction of oil from *Sucupira branca* seeds. *J Supercrit Fluids* 181:105494
- Chaturvedi M, Rani R, Sharma D, Yadav JP (2020) Effect of temperature and pressure on antimycobacterial activity of *Curcuma caesia* extract by supercritical fluid extraction method. *Int J Mycobacteriol* 9:296–302
- Chen F, Sun H, Ni H, Hong P, Li L, Yang Y, Jiang Z (2018a) Pummelo essential oil: extraction, volatiles, storage, and application. *J Food Sci Technol (China)* 36:1–10
- Chen HJ, Chen JW, Li X (2018b) Chemical constituents from the rhizome of *Saururus chinensis*. *Chin Pharm J* 53:340–345
- Cheng X, Qi Z, Burdyny T, Kong T, Sinton D (2018) Low pressure supercritical CO₂ extraction of astaxanthin from *Haematococcus pluvialis* demonstrated on a microfluidic chip. *Bioresour Technol* 250:481–485
- Chhouk K, Kanda H, Goto M (2018) Efficacy of supercritical carbon dioxide integrated hydrothermal extraction of Khmer medicinal plants with potential pharmaceutical activity. *J Environ Chem Eng* 6:2944–2956
- Chi XF, Yue HL, Zhao XH, Hu FZ (2016) Obtaining alantolactone and isoalantolactone from *Inula racemosa* Hook.f. by optimized supercritical fluid extraction. *Ind Crop Prod* 79:63–69
- Choi SA, Lee HS (2018) Insecticidal activities of *Russia coriander* oils and these constituents against *Sitophilus oryzae* and *Sitophilus zeamais*. *J Appl Biol Chem* 61:239–243

- Chou HY, Lee C, Pan JL, Wen ZH, Huang SH, Lan CWJ, Liu WT, Hour TC, Hseu YC, Hwang BH, Cheng KC, Wang HMD (2016) Enriched astaxanthin extract from *haematococcus pluvialis* augments growth factor secretions to increase cell proliferation and induces MMP1 degradation to enhance collagen production in human dermal fibroblasts. *Int J Mol Sci* 17
- Colorado D, Fernandez M, Orozco J, Lopera Y, Muñoz DL, Acín S, Balcazar N (2020) Metabolic activity of anthocyanin extracts loaded into non-ionic niosomes in diet-induced obese mice. *Pharm Res* 37
- Confortin TC, Todero I, Luft L, Teixeira AL, Mazutti MA, Zabot GL, Tres MV (2019) Valorization of solanum viarum dunal by extracting bioactive compounds from roots and fruits using ultrasound and supercritical CO₂. *Braz J Chem Eng* 36:1689–1702
- Cordeiro RM, de S. e Silva AP, Pinto RH, da Costa WA, da Silva SH, de Souza Pinheiro WB, Arruda MS, Carvalho Junior RN (2019) Supercritical CO₂ extraction of ucuúba (*Virola surinamensis*) seed oil: global yield, kinetic data, fatty acid profile, and antimicrobial activities. *Chem Eng Commun* 206:86–97
- Correa M, Mesomo MC, Pianowski KE, Torres YR, Corazza ML (2016) Extraction of inflorescences of *Musa paradisiaca* L. using supercritical CO₂ and compressed propane. *J Supercrit Fluids* 113:128–135
- Cunha VM, da Silva MP, de Sousa SH, do Nascimento Bezerra P, Menezes EG, da Silva NJ, da Silva Banna DA, Araújo ME, de Carvalho Junior RN (2019) Bacaba-de-leque (*Oenocarpus distichus* Mart.) oil extraction using supercritical CO₂ and bioactive compounds determination in the residual pulp. *J Supercrit Fluids* 144:81–90
- Cvetković T, Ranilović J, Gajari D, Tomić-Obrdalj H, Šubarić D, Moslavac T, Cikoš AM, Jokić S (2020) Podravka and slavonska varieties of pepper seeds (*Capsicum annuum* L.) as a new source of highly nutritional edible oil. *Foods* 9:1262
- da Silva IC, Oliveira PF, Barbosa GM, Wessjohann LA, Cardozo-Filho L, Holandino C, Muzitano MF, Leal IC (2020) Passiflora mucronata leaves extracts obtained from different methodologies: A phytochemical study based on cytotoxic and apoptosis activities of triterpenes and phytosterols constituents. *Brazilian J Pharm Sci* 56
- da Silva M, Trancoso J, Tormen L, Bombardelli MM, Corazza ML, Bainy EM (2022) Extraction of compounds from *Moringa oleifera* leaves using supercritical CO₂ plus ethanol as a cosolvent. *J Food Process Eng* 45
- Dąbrowski G, Konopka I, Czaplicki S (2018) Supercritical CO₂ extraction in chia oils production: impact of process duration and CO-solvent addition. *Food Sci Biotechnol* 27:677–686
- Damjanović-Vratnica B, Perović S, LU T, Santos R (2016) Effect of matrix pretreatment on the supercritical CO₂ extraction of satireja montana essential oil. *Chem Ind Chem Eng Q* 22:201–209
- Damrongwattanakool N, Raviyan P (2018) Enrichment of vitamin e in palm fatty acid distillate using sequential-cooling urea–Fatty acid complexation. *Songklanakarin J Sci Technol* 40:1175–1180
- Daraee A, Ghoreishi SM, Hedayati A (2019) Supercritical CO₂ extraction of chlorogenic acid from sunflower (*Helianthus annuus*) seed kernels: modeling and optimization by response surface methodology. *J Supercrit Fluids* 144:19–27
- de Aguiar AC, Osorio-Tobón JF, Silva LP, Barbero GF, Martinez J (2018) Economic analysis of oleoresin production from malagueta peppers (*Capsicum frutescens*) by supercritical fluid extraction. *J Supercrit Fluids* 133:86–93
- de Aguiar AC, da Fonseca Machado AP, Angolini CF, de Moraes DR, Baseggio AM, Eberlin MN, Junior MR, Martinez J (2019) Sequential high-pressure extraction to obtain capsinoids and phenolic compounds from biquinho pepper (*Capsicum chinense*). *J Supercrit Fluids* 150:112–121
- de Cerqueira MD, de Souza-Neta LC, Guedes ML, Rivelino R, Cruz FG (2013) Myrciaine, a new nicotinic ester from *Myrcia blanchetiana* (Myrtaceae). *Tetrahedron Lett* 54:1421–1423
- de Melo MM, Carius B, Simões MM, Portugal I, Saraiva J, Silva CM (2020) Supercritical CO₂ extraction of *V. vinifera* leaves: influence of cosolvents and particle size on removal kinetics and selectivity to target compounds. *J Supercrit Fluids* 165:104959

- de Oliveira NA, Mazzali MR, Fukumasu H, Goncalves CB, de Oliveira AL (2019) Composition and physical properties of babassu seed (*Orbignya phalerata*) oil obtained by supercritical CO₂ extraction. *J Supercrit Fluids* 150:21–29
- de Souza Junior ET, Siqueira LM, Almeida RN, Lucas AM, Silva CG, Cassel E, Vargas RM (2020) Comparison of different extraction techniques of zingiber officinale essential oil. *Braz Arch Biol Technol* 63
- Dehghanizadeh M, Brewer CE (2020) Guayule resin: Chemistry, extraction, and applications. 2020 ASABE Annual International Meeting
- Demirköz AB, Karakaş M, Bayramoğlu P, Üner M (2018) Analysis of volatile flavour components by dynamic headspace analysis/gas chromatography-mass spectrometry in roasted pistachio extracts using supercritical carbon dioxide extraction and sensory analysis. *Int J Food Prop* 21:972–981
- Deng SB, Chen JP, Chen YZ, Yu CQ, Yang Y, Wu SH, Chen CZ (2018) Chemical composition analysis of extracts from *Ficus Hirta* using supercritical fluid. In: 2018 International Conference on Computer Information and Automation Engineering, ICCIAE 2018
- Derrien M, Aghabaranjad M, Gosselin A, Desjardins Y, Angers P, Boumghar Y (2018) Optimization of supercritical carbon dioxide extraction of lutein and chlorophyll from spinach by-products using response surface methodology. *LWT* 93:79–87
- Devi V, Khanam S (2019a) Development of generalized and simplified models for supercritical fluid extraction: case study of papaya (*Carica papaya*) seed oil. *Chem Eng Res Des* 150:341–358
- Devi V, Khanam S (2019b) Study of ω -6 linoleic and ω -3 α -linolenic acids of hemp (*Cannabis sativa*) seed oil extracted by supercritical CO₂ extraction: CCD optimization. *J Environ Chem Eng* 7:1–10
- Di Sanzo GD, Mehariya S, Martino M, Larocca V, Casella P, Chianese S, Musmarra D, Balducci R, Molino A (2018) Supercritical carbon dioxide extraction of astaxanthin, lutein, and fatty acids from *haematococcus pluvialis* microalgae. *Mar Drugs* 16
- Dienaitė L., Pukalskienė, M., Pereira, C. V., Matias, A. A. & Venskutonis, P. R. 2020. Valorization of european cranberry bush (*viburnum opulus* L.) berry pomace extracts isolated with pressurized ethanol and water by assessing their phytochemical composition, antioxidant, and antiproliferative activities. *Foods*, 9
- Dienaitė L, Baranauskienė R, Rimantas Venskutonis P (2021) Lipophilic extracts isolated from European cranberry bush (*Viburnum opulus*) and sea buckthorn (*Hippophae rhamnoides*) berry pomace by supercritical CO₂ – promising bioactive ingredients for foods and nutraceuticals. *Food Chem* 348:129047
- Djapic N (2018) *Corallina officinalis* chemical compounds obtained by supercritical fluid extraction. *AAAC Bioflux* 11:422–428
- Dong A, Li W, Wang H, Zou Y (2020) Extraction of Dihydrofisetin from *Rhus punjabensis* var. *sinica* Rehd. Fruit by supercritical CO₂ and its antioxidant activity in vitro. *Chem Ind Forest Prod* 40:108–114
- Dorado Achicanoy D, Hurtado Benavides A, Martínez-Correa HA (2018) Study of supercritical CO₂ extraction of tamarillo (*Cyphomandra Betacea*) seed oil containing high added value compounds. *Electrophoresis* 39:1917–1925
- Eissa M, Hashim YZHY, Zainurin NAA (eds) (2018) *Aquilaria malaccensis* leaf as an alternative source of antiinflammatory compounds. *Int J Adv Sci Eng Inform Technol* 8:1625–1632
- El-Shamy S, Farag MA (2021) Novel trends in extraction and optimization methods of bioactives recovery from pomegranate fruit biowastes: valorization purposes for industrial applications. *Food Chem* 365:130465
- Fang X, Du M, Luo F, Jin Y (2015) Physicochemical properties and lipid composition of camellia seed oil (*camellia oleifera* abel.) extracted using different methods. *Food Sci Technol Res* 21:779–785
- Fang L, Wang X, Guo L, Liu Q (2018) Antioxidant, anti-microbial properties and chemical composition of cumin essential oils extracted by three methods. *Open Chem* 16:291–297

- Favareto R, Teixeira MB, Soares FAL, Belisário CM, Cabral JF, da Silva EA, Moia TA, Cardozo-Filho L (eds) (2019) Extraction of bioactive compounds of leaves of *duguetia furfuracea* (annonaceae) using green and organic solvents. *Braz J Chem Eng* 36:549–556
- Felföldi-Gáva A, Szarka S, Simándi B, Blazics B, Simon B, Kéry A (2012) Supercritical fluid extraction of *Alnus glutinosa* (L.) Gaertn. *J Supercrit Fluids* 61:55–61
- Ferrentino G, Haman N, Morozova K, Tonon G, Scampicchio M (2021) Phenolic compounds extracted from spruce (*Picea abies*) by supercritical carbon dioxide as antimicrobial agents against gram-positive bacteria assessed by isothermal calorimetry. *J Therm Anal Calorim* 145:3093–3103
- Ferro DM, Mazzutti S, Vitali L, Müller CM, Ferreira SR (2019) Integrated extraction approach to increase the recovery of antioxidant compounds from *Sida rhombifolia* leaves. *J Supercrit Fluids* 149:10–19
- Fornari T, de Molina AR, Reglero G (2014) Phenolic diterpenes from rosemary as enhancement agents of usual chemotherapeutic drugs for colorectal cancer therapy. *New Developments in Terpenes Research*
- Frohlich PC, Santos KA, Palú F, Cardozo-Filho L, da Silva C, da Silva EA (2019) Evaluation of the effects of temperature and pressure on the extraction of eugenol from clove (*Syzygium aromaticum*) leaves using supercritical CO₂. *J Supercrit Fluids* 143:313–320
- Gallo-Molina AC, Castro-Vargas HI, Garzón-Méndez WF, Ramírez JA, Monroy ZJ, King JW (2019) Extraction, isolation and purification of tetrahydrocannabinol from the *Cannabis sativa* L. plant using supercritical fluid extraction and solid phase extraction. *J Supercrit Fluids* 146:208–216
- Gan Z, Liang Z, Chen X, Wen X, Wang Y, Li M, Ni Y (2016) Separation and preparation of 6-gingerol from molecular distillation residue of Yunnan ginger rhizomes by high-speed counter-current chromatography and the antioxidant activity of ginger oils in vitro. *J Chromatogr B Anal Technol Biomed Life Sci* 1011:99–107
- García-Pérez JS, Cuéllar-Bermúdez SP, de la Cruz-Quiroz R, Arévalo-Gallegos A, Esquivel-Hernandez DA, Rodríguez-Rodríguez J, García-García R, Iqbal HM, Parra-Saldivar R (2019) Supercritical CO₂-based tailor made valorization of *Origanum vulgare* L extracts: A green approach to extract high-value compounds with applied perspectives. *J Environ Manag* 232:796–802
- García-Pérez JS, Cuéllar-Bermúdez SP, Arévalo-Gallegos A, Salinas-Salazar C, Rodríguez-Rodríguez J, de la Cruz-Quiroz R, Iqbal HM, Parra-Saldivar R (2020) Influence of supercritical CO₂ extraction on fatty acids profile, volatile compounds and bioactivities from *Rosmarinus officinalis*. *Waste Biomass Valorization* 11:1527–1537
- Gavarić A, Vidović S, Aladić K, Jokić S, Vladić J (2021) Supercritical CO₂ extraction of *Marrubium vulgare*: intensification of marrubiin. *RSC Adv* 11:9067–9075
- Gawron G, Krzyczkowski W, Lemke K, Ołdak A, Kadziński L, Banecki B (2019) *Nigella sativa* seed extract applicability in preparations against methicillin-resistant *Staphylococcus aureus* and effects on human dermal fibroblasts viability. *J Ethnopharmacol* 244:112135
- Ghosh S, Chatterjee D, Das S, Bhattacharjee P (2013) Supercritical carbon dioxide extraction of eugenol-rich fraction from *Ocimum sanctum* Linn and a comparative evaluation with other extraction techniques: process optimization and phytochemical characterization. *Ind Crop Prod* 47:78–85
- Ghosh S, Bhattacharjee P, Das S (2015) 1,8-cineol-rich cardamom seed (*Elettaria cardamomum*) extracts using green technologies and conventional extractions: process analysis, phytochemical characterization, and food application. *Sep Sci Technol (Philadelphia)* 50:1974–1985
- Górnaś P, Siger A, Rudzińska M, Grygier A, Marszałkiewicz S, Ying Q, Sobieszczkańska N, Segliņa D (2019) Impact of the extraction technique and genotype on the oil yield and composition of lipophilic compounds in the oil recovered from Japanese Quince (*Chaenomeles japonica*) seeds. *Eur J Lipid Sci Technol* 121:1800262

- Gustinelli G, Eliasson L, Svelander C, Alminger M, Ahrné L (2018) Supercritical CO₂ extraction of bilberry (*Vaccinium myrtillus* L.) seed oil: fatty acid composition and antioxidant activity. *J Supercrit Fluids* 135:91–97
- Gwatidzo L, Botha BM, Mccrindle RI, Combrinck S (eds) (2014) Extraction and identification of phytosterols in manketti (*Schinziophyton rautanenii*) nut oil. *JAOCS* 91:783–794
- Gyawali R, Jeon DH, Moon J, Kim H, Song YW, Hyun HB, Jeong D, Cho SK (2012a) Chemical composition and antiproliferative activity of supercritical extract of *Citrus grandis* (L.) Osbeck fruits from Korea. *J Essent Oil-Bear Plants* 15:915–925
- Gyawali R, Moon JY, Jeon DH, Kim HJ, Song YW, Hyun HB, Kang TH, Moon KS, Jeong S, Kim JC, Ahn KS, Cho SK (2012b) Chemical composition and antiproliferative activity of supercritical CO₂ extracts from citrus fruits. *Food Sci Technol Res* 18:813–823
- Györi E, Varga A, Fábán I, Lázár I (eds) (2019) Supercritical CO₂ extraction and selective adsorption of aroma materials of selected spice plants in functionalized silica aerogels. *J Supercrit Fluids* 148:16–23
- Haddadin MSY, Haddadin JS (2015) Lycopene extraction from tomato pomace with supercritical carbon dioxide: effect of pressures, temperatures and CO₂ flow rates and evaluation of antioxidant activity and stability of lycopene. *Pak J Nutr* 14:942–956
- Hadinezhad M, Rowland O, Hosseinian F (2015) The fatty acid profile and phenolic composition of *Descurainia sophia* seeds extracted by supercritical CO₂. *JAOCS* 92:1379–1390
- Haiyee ZA, Mohd Shah SH, Ismail K, Hashim N, Wan Ismail WI (2016) Quality parameters of *Curcuma longa* L. extracts by supercritical fluid extraction (SFE) and ultrasonic assisted extraction (UAE). *Malaysian J Anal Sci* 20:626–632
- Hamburger M (2002) *Isatis tinctoria* - from the rediscovery of an ancient medicinal plant towards a novel anti-inflammatory phytopharmaceutical. *Phytochem Rev* 1:333–344
- Hamdan S, Daood HG (2011) Changes in the chlorophyll and carotenoid content and composition of ground thyme leaves as a function of supercritical carbon dioxide and subcritical propane extraction. *Acta Aliment* 40:8–18
- Hameed A, Arun AB, Ho HP, Chang CMJ, Rekha PD, Lee MR, Singh S, Young CC (2011) Supercritical carbon dioxide micronization of zeaxanthin from moderately thermophilic bacteria *muricauda lutaonensis* CC-HSB-11T. *J Agric Food Chem* 59:4119–4124
- Hamid IAA, Mustapa AN, Ismail N, Abdullah Z (2013) Solubility prediction of mangosteen peel oil in Supercritical Carbon Dioxide using Neural Network. In: 2013 IEEE Business Engineering and Industrial Applications Colloquium, BEIAC 2013, Langkawi. pp. 91–96
- Hamid MA, Bakar NA, Park CS, Ramli F, Wan WR (2018) Optimisation of alpha mangostin extraction using supercritical CO₂ from *garcinia mangostana*. *Chem Eng Trans* 63:577–582
- Han CN, Kang CH (2015) Extraction of genistein from *sophora flavescens* with supercritical carbon dioxide. *Korean Chem Eng Res* 53:445–449
- Han JH, Wu QF, Xu B, Zhou SL, Ding F (2016) Quality characteristics of soybean germ oil obtained by innovative subcritical butane experimental equipment. *Qual Assur Safety Crops Foods* 8:369–377
- Haro-González JN, Castillo-Herrera GA, Martínez-Velázquez M, Espinosa-Andrews H (2021) Clove essential oil (*Syzygium aromaticum* l. myrtaceae): extraction, chemical composition, food applications, and essential bioactivity for human health. *Molecules* 26
- Hartati, Salleh LM, Pagarra H, Rachmawaty (2018) Response Surfaces of Linoleic Acid of *Swietenia Mahagoni* in Supercritical Carbon Dioxide. In: 2nd International Conference on Statistics, Mathematics, Teaching, and Research 2017, ICSMTR 2017, 2018
- Hedrick JL, Mulcahey LJ, Taylor LT (1992) Supercritical fluid extraction. *Mikrochim Acta* 108:115–132
- Hernández SMP, Estévez JJ, Giraldo L JL, Méndez CJM (2019) Supercritical extraction of bioactive compounds from cocoa husk: study of the main parameters. *Revista Facultad de Ingenieria*:95–105

- Hrabovski N, Sinadinović-Fišer S, Nikolovski B, Sovilj M, Borota O (2012) Phytosterols in pumpkin seed oil extracted by organic solvents and supercritical CO₂. *Eur J Lipid Sci Technol* 114:1204–1211
- Hu Q, Xu J, Chen S, Yang F (2004) Antioxidant activity of extracts of black sesame seed (*Sesamum indicum* L.) by supercritical carbon dioxide extraction. *J Agric Food Chem* 52:943–947
- Idham Z, Putra NR, Aziz AHA, Zaini AS, Rasidek NAM, Mili N, Yunus MAC (eds) (2021) Improvement of extraction and stability of anthocyanins, the natural red pigment from roselle calyces using supercritical carbon dioxide extraction. *J CO₂ Util* 56:101839
- Idowu S, Adekoya AE, Igiehon OO, Idowu AT (2021) Clove (*Syzygium aromaticum*) spices: a review on their bioactivities, current use, and potential application in dairy products. *J Food Measure Charact* 15:3419–3435
- Inakuma T (2015) Study of carotenoid activity in vegetables : application to food development. *Nippon Shokuhin Kagaku Kogaku Kaishi* 62:263–273
- Ixtaina VY, Nolasco SM, Tomás MC (2014) Characterization of Argentinean chia seed oil obtained by different processes: a multivariate study. In: *Seed oil: biological properties, health benefits and commercial applications*
- Jaafar SNBS, Haimer E, Liebner F, Böhmdorfer S, Potthast A, Rosenau T (2011) Empty palm fruit bunches-A CO₂-based biorefinery concept. *J Biobased Mater Bioenergy* 5:225–233
- Jafarian Asl P, Niazmand R, Jahani M (2020) Theoretical and experimental assessment of supercritical CO₂ in the extraction of phytosterols from rapeseed oil deodorizer distillates. *J Food Eng* 269:109748
- Jahongir H, Miansong Z, Amankeldi I, Yu Z, Changheng L (2019) The influence of particle size on supercritical extraction of dog rose (*Rosa canina*) seed oil. *J King Saud Univ* 31:140–143
- Jaski JM, Barão CE, Moraes Lião L, da Silva Pinto V, Zanoelo EF, Cardozo-Filho L (2019) β -Cyclodextrin complexation of extracts of olive leaves obtained by pressurized liquid extraction. *Ind Crop Prod* 129:662–672
- Jiang Q, Liu W, Li X, Zhang T, Wang Y, Liu X (2016) Detection of related substances in polyene phosphatidyl choline extracted from soybean and in its commercial capsule by comprehensive supercritical fluid chromatography with mass spectrometry compared with HPLC with evaporative light scattering detection. *J Sep Sci* 39:350–357
- Jokić S, Molnar M, Jakovljević M, Aladić K, Jerković I (2018) Optimization of supercritical CO₂ extraction of *Salvia officinalis* L. leaves targeted on oxygenated monoterpenes, A-humulene, viridiflorol and manool. *J Supercrit Fluids* 133:253–262
- Jokić S, Molnar M, Cikoš AM, Jakovljević M, Šafranko S, Jerković I (2020) Separation of selected bioactive compounds from orange peel using the sequence of supercritical CO₂ extraction and ultrasound solvent extraction: optimization of limonene and hesperidin content. *Sep Sci Technol (Philadelphia)* 55:2799–2811
- Karimi M, Raofie F (2019) Micronization of vincristine extracted from *Catharanthus roseus* by expansion of supercritical fluid solution. *J Supercrit Fluids* 146:172–179
- Kaushik S, Dar L, Kaushik S, Yadav JP (2021) Anti-dengue activity of super critical extract and isolated oleanolic acid of *Leucas cephalotes* using in vitro and in silico approach. *BMC Complement Med Ther* 21
- Kessler JC, Vieira VA, Martins IM, Manrique YA, Afonso A, Ferreira P, Mandim F, Ferreira ICFR, Barros L, Rodrigues AE, Dias MM (2022) Obtaining aromatic extracts from Portuguese thymus *mastichina* L. by hydrodistillation and supercritical fluid extraction with CO₂ as potential Flavouring additives for food applications. *Molecules* 27
- Kha TC, Phan-Tài H, Nguyen MH (2014) Effects of pre-treatments on the yield and carotenoid content of Gac oil using supercritical carbon dioxide extraction. *J Food Eng* 120:44–49
- Khalil AA, Rahmal UU, Khan MR, Sahar A, Mehmood T, Khan M (2017) Essential oil eugenol: Sources, extraction techniques and nutraceutical perspectives. *RSC Adv* 7:32669–32681
- Khaw KY, Parat MO, Shaw PN, Falconer JR (2017) Solvent supercritical fluid technologies to extract bioactive compounds from natural sources: a review. *Molecules* 22

- Khoddami A, Man YBC, Roberts TH (2014) Physico-chemical properties and fatty acid profile of seed oils from pomegranate (*Punica granatum L.*) extracted by cold pressing. *Eur J Lipid Sci Technol* 116:553–562
- Klejdus B, Lojková L, Lapčík O, Koblovská R, Moravcová J, Kubáň V (2005) Supercritical fluid extraction of isoflavones from biological samples with ultra-fast high-performance liquid chromatography/mass spectrometry. *J Sep Sci* 28:1334–1346
- Konar N, Dalabasmaz S, Poyrazoglu ES, Artik N, Colak A (2014) The determination of the caffeic acid derivatives of *Echinacea purpurea* aerial parts under various extraction conditions by supercritical fluid extraction (SFE). *J Supercrit Fluids* 89:128–136
- Kueh BWB, Yusup S, Osman N (2018) Supercritical carbon dioxide extraction of *Melaleuca cajuputi* leaves for herbicides allelopathy: optimization and kinetics modelling. *J CO₂ Util* 24:220–227
- Kumar S, Dhanani T, Shah S (2014) Extraction of three bioactive diterpenoids from *andrographis paniculata*: effect of the extraction techniques on extract composition and quantification of three andrographolides using high-performance liquid chromatography. *J Chromatogr Sci* 52:1043–1050
- Kumoro AC, Hasan M, Singh H (2019) Extraction of Andrographolide from *Andrographis paniculata* dried leaves using supercritical CO₂ and ethanol mixture. *Ind Eng Chem Res* 58:742–751
- Kuo CF, Su JD, Chiu CH, Peng CC, Chang CH, Sung TY, Huang SH, Lee WC, Chyau CC (2011) Anti-inflammatory effects of supercritical carbon dioxide extract and its isolated carnosic acid from *rosmarinus officinalis* leaves. *J Agric Food Chem* 59:3674–3685
- Kyriakoudi A, Z. Tsimidou M (2018) Latest advances in the extraction and determination of saffron apocarotenoids. *Electrophoresis* 39:1846–1859
- Lee YS, Park KY, Ji SH, Jo GS, Lee SK (2018) Effect of harvest seasons and extraction methods on the nutritional and functional components of Seomcho (*Spinacia oleracea L.*). *Korean J Food Preserv* 25:682–688
- Lefebvre T, Destandau E, Lesellier E (2021) Sequential extraction of carnosic acid, rosmarinic acid and pigments (carotenoids and chlorophylls) from Rosemary by online supercritical fluid extraction-supercritical fluid chromatography. *J Chromatogr A* 1639:461709
- Leyva-Jiménez FJ, Lozano-Sánchez J, Fernández-Ochoa Á, Cádiz-Gurrea MDLL, Arraéz-Román D, Segura-Carretero A (2020) Optimized extraction of phenylpropanoids and flavonoids from lemon verbena leaves by supercritical fluid system using response surface methodology. *Foods* 9
- Li J, Zhang J, Wang M (2016a) Extraction of flavonoids from the flowers of *abelmoschus manihot (L.) medic* by modified supercritical CO₂ extraction and determination of antioxidant and anti-adipogenic activity. *Molecules* 21
- Li J, Zhang X, Liu Y (2016b) Supercritical carbon dioxide extraction of *Ganoderma lucidum* spore lipids. *LWT* 70:16–23
- Li L, Wu Y, Yuan SW, Yang J, Yang YF, Ding G, Xiao W (2016c) Extraction and purification process of total fatty acid in *Brassica campestris* pollen. *Zhongguo Zhong yao za zhi* 41:226–232
- Lima RN, Ribeiro AS, Cardozo-Filho L, Vedoy D, Alves PB (2019) Extraction from leaves of *piper klotzschianum* using supercritical carbon dioxide and co-solvents. *J Supercrit Fluids* 147:205–212
- Lima RN, Santos ADC, Ribeiro AS, Cardozo-Filho L, Freitas LS, Barison A, Costa EV, Alves PB (2020) Selective amides extraction and biological activity from *Piper hispidum* leaves using the supercritical extraction. *J Supercrit Fluids* 157:104712
- Liu X (2018) Optimization of Supercritical CO₂ Fluid Extraction Conditions of Saponins from *Spina Gleditsiae*. In: 2018 International Conference on Advanced Electronic Materials, Computers and Materials Engineering, AEMCME 2018
- Long T, Lv X, Xu Y, Yang G, Xu LY, Li S (2019) Supercritical fluid CO₂ extraction of three poly-methoxyflavones from *Citri reticulatae* pericarpium and subsequent preparative separation by continuous high-speed counter-current chromatography. *J Chromatogr B Anal Technol Biomed Life Sci* 1124:284–289

- Luan ZJ, Li PP, Li D, Meng XP, Sun J (2020) Optimization of supercritical-CO₂ extraction of *Iris lactea* seed oil: component analysis and antioxidant activity of the oil. *Ind Crop Prod* 152:112553
- Machado BAS, Silva RPD, Barreto GDA, Costa SS, da Silva DF, Brandão HN, da Rocha JLC, Dellagostin OA, Henriques JAP, Umsza-Guez MA, Padilha FF (2016) Chemical composition and biological activity of extracts obtained by supercritical extraction and ethanolic extraction of brown, green and red propolis derived from different geographic regions in Brazil. *PLoS ONE* 11
- Maia JD, Ávila CRD, Mezzomo N, Lanza M (2018) Evaluation of bioactive extracts of mangaba (*Hancornia speciosa*) using low and high pressure processes. *J Supercrit Fluids* 135:198–210
- Marić B, Abramović B, Ilić N, Krulj J, Kojić J, Perović J, Bodroža-Solarov M, Teslić N (2020) Valorization of red raspberry (*Rubus idaeus* L.) seeds as a source of health beneficial compounds: extraction by different methods. *J Food Process Preserv* 44
- Martins PF, de Melo MMR, Sarmento P, Silva CM (2016) Supercritical fluid extraction of sterols from *Eichhornia crassipes* biomass using pure and modified carbon dioxide. Enhancement of stigmasteryl yield and extract concentration. *J Supercrit Fluids* 107:441–449
- Marzorati S, Friscione D, Picchi E, Verotta L (2020) Cannabidiol from inflorescences of *Cannabis sativa* L.: green extraction and purification processes. *Ind Crop Prod* 155:112816
- Masghati S, Ghoreishi SM (2018) Supercritical CO₂ extraction of cinnamaldehyde and eugenol from cinnamon bark: optimization of operating conditions via response surface methodology. *J Supercrit Fluids* 140:62–71
- Mehariya S, Iovine A, Di Sanzo G, Larocca V, Martino M, Leone G, Casella P, Karatza D, Marino T, Musmarra D, Molino A (2019) Supercritical fluid extraction of lutein from *scenedesmus almeriensis*. *Molecules* 24
- Memon AH, Hamil MSR, Laghari M, Rithwan F, Zhari S, Saeed MAA, Ismail Z, Majid AMSA (2016) A comparative study of conventional and supercritical fluid extraction methods for the recovery of secondary metabolites from *Syzygium campanulatum* Korth. *J Zhejiang Univ Sci B* 17:683–691
- Meneses MA, Caputo G, Scognamiglio M, Reverchon E, Adami R (2015) Antioxidant phenolic compounds recovery from *Mangifera indica* L. by-products by supercritical antisolvent extraction. *J Food Eng* 163:45–53
- Mesquita PC, Rodrigues LGG, Mazzutti S, da Silva M, Vitali L, Lanza M (2021) Intensified green-based extraction process as a circular economy approach to recover bioactive compounds from soursop seeds (*Annona muricata* L.). *Food Chem X*:12
- Miękus N, Iqbal A, Marszałek K, Puchalski C, Świergiel A (2019) Green chemistry extractions of carotenoids from *daucus carota* L.-Supercritical carbon dioxide and enzyme-assisted methods. *Molecules* 24
- Mihalcea L, Turturică M, Cucolea EI, Dănilă GM, Dumitrașcu L, Coman G, Constantin OE, Grigore-Gurgu L, Stănciuc N (2021) CO₂ supercritical fluid extraction of oleoresins from sea buckthorn pomace: evidence of advanced bioactive profile and selected functionality. *Antioxidants* 10
- Minteguiaga M, Catalán CAN, Cassel E, Dellacassa E (2021) The “other Vassoura oil” and volatile fractions from *Baccharis uncinella* DC. (Asteraceae) as potential sources for flavor and fragrance industry. In: Volatile oils: production, composition and uses
- Mohamad N, Ramli N, Abd-Aziz S, Ibrahim MF (2019) Comparison of hydro-distillation, hydro-distillation with enzyme-assisted and supercritical fluid for the extraction of essential oil from pineapple peels. *3 Biotech* 9
- Mu J, Wu G, Chen Z, Brennan CS, Tran K, Dilrukshi HNN, Shi C, Zhen H, Hui X (2021) Identification of the fatty acids profiles in supercritical CO₂ fluid and Soxhlet extraction of Samara oil from different cultivars of *Elaeagnus mollis* Diels seeds. *J Food Compos Anal* 101:103982
- Nastić N, Borrás-Linares I, Lozano-Sánchez J, Švarc-Gajić J, Segura-Carretero A (2018) Optimization of the extraction of phytochemicals from black mulberry (*Morus nigra* L.) leaves. *J Ind Eng Chem* 68:282–292

- Náthia-Neves G, Vardanega R, Martinez Urango AC, Meireles MAA (2020) Supercritical CO₂ extraction of α -bisabolol from different parts of candeia wood (*Eremanthus erythropappus*). *J Supercrit Fluids* 166:105026
- Occhipinti A, Capuzzo A, Bossi S, Milanese C, Maffei ME (2013) Comparative analysis of supercritical CO₂ extracts and essential oils from an *Ocimum basilicum* chemotype particularly rich in T-cadinol. *J Essent Oil Res* 25:272–277
- Ouédraogo JCW, Dicko C, Kini FB, Bonzi-Coulibaly YL, Dey ES (2018) Enhanced extraction of flavonoids from *Odontonema strictum* leaves with antioxidant activity using supercritical carbon dioxide fluid combined with ethanol. *J Supercrit Fluids* 131:66–71
- Özcan MM, Ören D (2019) Comparative of physico-chemical properties of wheat germ oil extracted with cold press and supercritical CO₂ extraction. *Iran J Chem Chem Eng* 38:167–174
- Pal S, Bhattacharjee P (2018) Spray dried powder of lutein-rich supercritical carbon dioxide extract of gamma-irradiated marigold flowers: process optimization, characterization and food application. *Powder Technol* 327:512–523
- Pavić V, Jakovljević M, Molnar M, Jokić S (2019) Extraction of carnosic acid and carnosol from sage (*Salvia officinalis* L.) leaves by supercritical fluid extraction and their antioxidant and antibacterial activity. *Plan Theory* 8
- Pavlović N, Vidović S, Vladić J, Popović L, Moslavac T, Jakobović S, Jokić S (2018) Recovery of tocopherols, amygdalin, and fatty acids from apricot kernel oil: cold pressing versus supercritical carbon dioxide. *Eur J Lipid Sci Technol* 120
- Pires FB, Dolwitsch CB, Ugalde GA, Menezes BB, Fontana MEZ, Rieffeld RC, Sagrillo MR, Essi L, Mazutti MA, da Rosa MB, Pizzutti IR (2021) Chemical study, antioxidant activity, and genotoxicity and cytotoxicity evaluation of *Ruellia angustiflora*. *Nat Prod Res* 35:5317–5322
- Pise VH, Shirkole SS, Thorat BN (2022) Visualization of oil cells and preservation during drying of betel leaf (*Piper betel*) using hot-stage microscopy. *Dry Technol* 40:2494
- Pourmortazavi SM, Rahimi-Nasrabadi M, Hajimirsadeghi SS (2014) Supercritical fluid technology in analytical chemistry - review. *Curr Anal Chem* 10:3–28
- Priyanka, Khanam S (2018) Influence of operating parameters on supercritical fluid extraction of essential oil from turmeric root. *J Clean Prod* 188:816–824
- Ramli NH, Yusup S, Quitain AT, Johari K, Kueh BWB (2019) Optimization of saponin extracts using microwave-assisted extraction as a sustainable biopesticide to reduce *Pomacea canalicularata* population in paddy cultivation. *Sustain Chem Pharm* 11:23–35
- Reverchon E, Scognamiglio M, Baldino L (2022) Lycopene extract from tomato concentrate and its co-precipitation with PVP using hybrid supercritical processes. *J CO₂ Util* 64:102157
- Rodrigues VH, De Melo MMR, Portugal I, Silva CM (2021) Lupane-type triterpenoids from *Acacia dealbata* bark extracted by different methods. *Ind Crop Prod* 170:113734
- Rohman A, Irnawati (2020) Pumpkin (*Cucurbita maxima*) seed oil: chemical composition, antioxidant activities and its authentication analysis. *Food Res* 4:578–584
- Romano R, Aiello A, Pizzolongo F, Rispoli A, De Luca L, Masi P (2020) Characterisation of oleoresins extracted from tomato waste by liquid and supercritical carbon dioxide. *Int J Food Sci Technol* 55:3334–3342
- Salea R, Hiendrawan S, Subroto E, Veriansyah B, Tjandrawinata RR (2018) Supercritical carbon dioxide extraction of citronella oil from *Cymbopogon winterianus* using taguchi orthogonal array design. *Int J Appl Pharm* 10:147–151
- Sánchez-Camargo ADP, Gutiérrez LF, Vargas SM, Martínez-Correa HA, Parada-Alfonso F, Narváez-Cuenca CE (2019) Valorisation of mango peel: proximate composition, supercritical fluid extraction of carotenoids, and application as an antioxidant additive for an edible oil. *J Supercrit Fluids* 152:104574
- Sanjaya RE, Tedjo YY, Kurniawan A, Ju YH, Ayucitra A, Ismadji S (2014) Investigation on supercritical CO₂ extraction of phenolic-phytochemicals from an epiphytic plant tuber (*Myrmecodia pendans*). *J CO₂ Util* 6:26–33
- Saotome Y, Imai M (2018) Supercritical carbon dioxide extraction of apigenin from parsley leaves pre-treated to maximize yield. *Food Sci Technol Res* 24:63–73

- Sapkale GN, Patil SM, Surwase US, Bhatbhave PK (2010) Supercritical fluid extraction - a review. *Int J Chem Sci* 8:729–743
- Sharifi-Rad J, Ezzat SM, El Bishbishy MH, Mnayer D, Sharopov F, Kiliç CS, Neagu M, Constantin C, Sharifi-Rad M, Atanassova M, Nicola S, Pignata G, Salehi B, Fokou PVT, Martins N (2020) Rosmarinus plants: key farm concepts towards food applications. *Phytother Res* 34:1474–1518
- Shi LK, Zheng L, Liu RJ, Chang M, Jin QZ, Wang XG (2018) Chemical characterization, oxidative stability, and in vitro antioxidant capacity of sesame oils extracted by supercritical and subcritical techniques and conventional methods: a comparative study using chemometrics. *Eur J Lipid Sci Technol* 120:1700326
- Song L, Liu P, Yan Y, Huang Y, Bai B, Hou X, Zhang L (2019) Supercritical CO₂ fluid extraction of flavonoid compounds from Xinjiang jujube (*Ziziphus jujuba* Mill.) leaves and associated biological activities and flavonoid compositions. *Ind Crop Prod* 139:111508
- Suryawanshi B, Mohanty B (2018) Modeling and optimization: supercritical CO₂ extraction of *Pongamia pinnata* (L.) seed oil. *J Environ Chem Eng* 6:2660–2673
- Swapna Sonale R, Ramalakshmi K, Udaya Sankar K (2018) Characterization of neem (*Azadirachta indica* A. Juss) seed volatile compounds obtained by supercritical carbon dioxide process. *J Food Sci Technol* 55:1444–1454
- Tang ZX, Ying RF, Lv BF, Yang LH, Xu Z, Yan LQ, Bu JZ, Wei YS (2021) Flaxseed oil: extraction, health benefits and products. *Qual Assur Safety Crops Foods* 13:1–19
- Trucillo P, Campardelli R, Aliakbarian B, Perego P, Reverchon E (2018) Supercritical assisted process for the encapsulation of olive pomace extract into liposomes. *J Supercrit Fluids* 135:152–159
- Tyskiewicz K, Konkol M, Rójs E (2019) Supercritical carbon dioxide (scCO₂) extraction of phenolic compounds from lavender (*Lavandula angustifolia*) flowers: A box-Behnken experimental optimization. *Molecules* 24
- Uzel RA (2018) Effect of extraction method and extraction solvent on recovery of phenolic compounds from olive leaves in Kemalpaşa-İzmir (Turkey): Oleuropein recovery as a case example. *Sep Sci Technol (Philadelphia)* 53:1531–1539
- Végh K, Riethmüller E, Hosszú L, Darcsi A, Müller J, Alberti Á, Tóth A, Béni S, Könczöl Á, Balogh GT, Kéry Á (2018) Three newly identified lipophilic flavonoids in *Tanacetum parthenium* supercritical fluid extract penetrating the Blood-Brain Barrier. *J Pharm Biomed Anal* 149:488–493
- Veiga BA, Hamerski F, Clausen MP, Errico M, De Paula Scheer A, Corazza ML (2021) Compressed fluids extraction methods, yields, antioxidant activities, total phenolics and flavonoids content for Brazilian Mantiueira hops. *J Supercrit Fluids* 170:105155
- Velikorodov AV, Kovalev VB, Nosachev SB, Tyrkov AG, Morozova LV (2018) Fatty-oxygen composition of seeds oils of some wild-growing and cultivated plants of the Astrakhan Region Obtained by the supercritical fluid extraction method. *Khimiya Rastitel'nogo Syr'ya*:153–158
- Villanueva-Bermejo D, Vázquez E, Villalva M, Santoyo S, Fornari T, Reglero G, García-Risco MR (2019) Simultaneous supercritical fluid extraction of heather (*Calluna vulgaris* L.) and marigold (*Calendula officinalis* L.) and anti-inflammatory activity of the extracts. *Appl Sci (Switzerland)* 9
- Villinski JR, Bergeron C, Cannistra JC, Gloer JB, Coleman CM, Ferreira D, Azelmat J, Grenier D, Gafner S (2014) Pyrano-isoflavans from *Glycyrrhiza uralensis* with antibacterial activity against *Streptococcus mutans* and *Porphyromonas gingivalis*. *J Nat Prod* 77:521–526
- Wu H, Li J, Jia Y, Xiao Z, Li P, Xie Y, Zhang A, Liu R, Ren Z, Zhao M, Zeng C, Li C (2019) Essential oil extracted from *Cymbopogon citroneola* leaves by supercritical carbon dioxide: antioxidant and antimicrobial activities. *J Anal Methods Chem* 2019
- Yekefallah M, Raofie F (2022) Production of herbal nanocolloids from *Rubia tinctorum* L. roots by rapid expansion from supercritical solution into suspension system. *Ind Crop Prod* 176:114286
- Yousefi M, Rahimi-Nasrabadi M, Pourmortazavi SM, Wysokowski M, Jesionowski T, Ehrlich H, Mirsadeghi S (2019) Supercritical fluid extraction of essential oils. *TrAC* 118:182–193

- Yu L, Hu X, Xu R, Ba Y, Chen X, Wang X, Cao B, Wu X (2022) Amide alkaloids characterization and neuroprotective properties of *Piper nigrum* L.: A comparative study with fruits, pericarp, stalks and leaves. *Food Chem* 368:130832
- Zaid R, Mouhouche F, Canela-Garayoa R, Chacón NMO (2020) Supercritical fluid extraction of Algerian *Melissa officinalis* L. 1753 (Lamiaceae) and its biological activity against two species of the genus *Chaitophorus* (Homoptera-Aphididae). *Arch Phytopathol Plant Protect* 53:940–953
- Zakharenko AM, Razgonova MP, Pikula KS, Golokhvast KS (2021) Simultaneous determination of 78 compounds of *Rhodiola rosea* extract by supercritical CO₂-extraction and HPLC-ESI-MS/MS spectrometry. *Biochem Res Int* 2021
- Zengin G, Mahomoodally F, Picot-Allain C, Diuzheva A, Jekó J, Cziáky Z, Cvetanović A, Aktumsek A, Zeković Z, Rengasamy KRR (2019) Metabolomic profile of *Salvia viridis* L. root extracts using HPLC–MS/MS technique and their pharmacological properties: A comparative study. *Ind Crop Prod* 131:266–280
- Zhang ZS, Liu YL, Che LM (2018) Optimization of supercritical carbon dioxide extraction of *eucommia ulmoides* seed oil and quality evaluation of the oil. *J Oleo Sci* 67:255–263