



Central Nervous System Neoplasms in Microgravity

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Introduction

Since humans first inhabited the International Space Station (ISS) over two decades ago, the duration of space missions per astronaut have been limited to 1–12 months. Astronauts in low Earth orbit are also partially protected from galactic cosmic radiation (GCR) due to the Earth's magnetic field. The next steps in human exploration will include long-duration missions beyond low Earth orbit (LEO) and higher concerns for harmful effects of space radiation. Prolonged exposure to microgravity may also alter the central nervous system at the cytoarchitectural level [1], and it has been suggested that microgravity may even inhibit proliferation [2] of malignant glioma. Therefore, with the aim to protect astronaut's health and exploit space environment conditions to potentially develop on-ground countermeasures, the need to understand the behavior of the central nervous system (CNS) in space has emerged. The dismal knowledge regarding the characteristics of the combined effects of microgravity and space radiations arises interest in researchers, especially

regarding the long-term risk to develop neurodegenerative diseases and cancer. Recent progression in aerospace medicine and research opened several questions regarding the behavior of the neoplasms in microgravity and under ionizing radiations and a potential treatment window [3, 4].

Central nervous system (CNS) neoplasms are solid tumors arising from the brain, meninges, or spinal cord with different prognosis dependent on their location and histology.

CNS neoplasms are considered to be rare but lethal tumors as they account around 30% of cancer deaths in children and young adults [5]. The 2016 World Health Organization (WHO) classification categorizes CNS neoplasms based on histogenesis and molecular parameters to help aid in identification and prognostication [6]. The grading of some CNS tumor according to this classification is consultable in Fig. 8.1.

The treatment and prognosis of CNS neoplasms vary according to the location, severity of symptoms, and the type of neoplasm. In adults, the majority of primary CNS tumors are malignant in nature and treated with a combination of surgery, radiation, and chemotherapy depending [7]. Unfortunately, even after initial disease control, the majority of malignant tumors progress and the mortality rate remains high [8].

In this chapter, two main areas of cancer research in the spaceflight environment will be covered: (1) Central Nervous System malignancies tumorigenesis and (2) tumor suppression. We will analyze the molecular bases of tumorigenesis in terrestrial gravity and hypothesize on the tumorigenesis in microgravity with close attention to the contribution of radiation. We will further evaluate the tumor suppressive characteristics of microgravity in the treatment of CNS tumors. The chapter aims to review the currently available literature regarding these arguments and to identify the role of microgravity on CNS neoplasms behavior.

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WHO grades of select CNS tumours			
Diffuse astrocytic and oligodendroglial tumours			
Diffuse astrocytoma, IDH-mutant	II	Desmoplastic infantile astrocytoma and ganglioglioma	I
Anaplastic astrocytoma, IDH-mutant	III	Papillary glioneuronal tumour	I
Glioblastoma, IDH-wildtype	IV	Rosette-forming glioneuronal tumour	I
Glioblastoma, IDH-mutant	IV	Central neurocytoma	II
Diffuse midline glioma, H3K27M-mutant	IV	Extraventricular neurocytoma	II
Oligodendroglioma, IDH-mutant and 1p/19q-codeleted	II	Cerebellar liponeurocytoma	II
Anaplastic oligodendroglioma, IDH-mutant and 1p/19q-codeleted	III		
Other astrocytic tumours		Tumours of the pineal region	
Pilocytic astrocytoma	I	Pineocytoma	I
Subependymal giant cell astrocytoma	I	Pineal parenchymal tumour of intermediate differentiation	II or III
Pleomorphic xanthoastrocytoma	II	Pineoblastoma	IV
Anaplastic pleomorphic xanthoastrocytoma	III	Papillary tumour of the pineal region	II or III
Ependymal tumours		Embryonal tumours	
Subependymoma	I	Medulloblastoma(all subtypes)	IV
Myxopapillary ependymoma	I	Embryonal tumour with multilayered rosettes, C19MC-altered	IV
Ependymoma	II	Medulloepithelioma	IV
Ependymoma, <i>RELA</i> fusion-positive	II or III	CNS embryonal tumour, NOS	IV
Anaplastic ependymoma	III	Atypical teratoid/rhabdoid tumour	IV
Other gliomas		CNS embryonal tumour with rhabdoid features	IV
Angiocentric glioma	I	Tumours of the cranial and paraspinal nerves	
Chordoid glioma of third ventricle	II	Schwannoma	I
Choroid plexus tumours		Neurofibroma	I
Choroid plexus papilloma	I	Perineurioma	I
Atypical choroid plexus papilloma	II	Malignant peripheral nerve sheath tumour (MPNST)	II, III or IV
Choroid plexus carcinoma	III	Meningiomas	
Neuronal and mixed neuronal-glioma tumours		Meningioma	I
Dysembryoplastic neuroepithelial tumour	I	Atypical meningioma	II
Gangliocytoma	I	Anaplastic (malignant) meningioma	III
Ganglioglioma	I	Mesenchymal, non-meningothelial tumours	
Anaplastic ganglioglioma	III	Solitary fibrous tumour/ haemangiopericytoma	I, II or III
Dysplastic gangliocytoma of cerebellum (Lhermitte-Duclos)	I	Haemangioblastoma	I
		Tumours of the sellar region	
		Craniopharyngioma	I
		Granular cell tumour	I
		Pituitaryoma	I
		Spindle cell oncocytoma	I

Fig. 8.1 Grading of central nervous system tumours (2016 WHO). Reprinted from [6] with permission

Environment

Microgravity

The space environment is hazardous, outlined by high vacuum, extreme radiation of galactic and solar origin, and extreme temperatures [9]. Perhaps the most obvious unique influence on the pathophysiology of CNS neoplasms during spaceflight is that of microgravity. Microgravity or zero-g is used to describe the condition of weightlessness experienced during spaceflight [10]. The term does not necessarily refer to a reduced level of gravity in an absolute sense, but to the lack of counteracting inertial g-forces or any other forces than gravity. Microgravity is expressed as a fraction of g, where g is the gravitational acceleration at Earth's surface, on average 9.81 m/s². This should not be confused with gravitational field. At ISS altitude, the gravitational field is around 90% of that on the Earth's surface. However, the ISS orbits the Earth in a constant free fall and with almost negligible air resistance. The inertial g-forces on ISS are therefore virtu-

ally absent and equivalent to micro-fractions (10⁻⁶) of the normal force exerted on an individual on the surface of the Earth due to gravity. The lunar gravity of 0.16 g and Martian gravity of 0.38 g induce less gravitational forces than the Earth's gravity at ISS altitude, but the astronauts based on the surface of Moon or Mars are still exposed to higher gravitational load as compared to the microgravity environment experienced during orbit.

From now onwards, we will refer to the spaceflight gravitational environment as microgravity [11].

Space Radiations

The next largest hazard to take in consideration is that of ionizing radiations. Ionizing radiations are particles with a sufficient amount of energy which can totally discard an electron from its orbit, consequently generating a more positively charged atom. On the other hand, the non-ionizing radiation (Low energy) does not have adequate energy to separate

electrons. There are three naturally occurring types of ionizing space radiations; galactic cosmic rays (GCRs) originating beyond the solar system, localized trapped particle belts of electrons and protons (ERBs)—known also as Van Allen radiation belts—and solar particle events (SPEs) [12]. The Galactic cosmic rays (GCRs) are an isotropic flux of charged particles originating from sources beyond the solar system with unidentified origins which can penetrate through a typical spacecraft or an astronaut [13]. When the particles strike the spacecraft, hadronic cascades are also initiated and result in secondary particles. The average GCR absorbed per day in a mission to Mars has estimated to be around 1.75–3.0 mSv/day [14, 15]. The Van Allen radiation belts or ERBs are two zones confining the Earth in which energetic charged particles are trapped due to the Earth's magnetic field [16], and the planetary magnetic field varies among other planets in our Solar System [17, 18]. The majority of the inner Van Allen Belt is located beyond the ISS orbit and protects the station from incoming particles. However, the South Atlantic Anomaly (SAA) is an area where the inner belt dips closer to Earth and expose ISS to large amounts of radiation. The Solar Particle Events (SPEs) are made mostly of protons with a high-value flux representing a risk for astronaut health, however, in contrary to the GCRs, they are feasible of defense [19]. The SPEs are shielded by the Earth magnetic field, so they are of greater concern for planetary and interplanetary missions. Very large SPEs are rare, but challenges in prediction of their occurrence may impose significant operational constraints or radiation risk to the crew.

To grasp the impact of ionizing radiations, it is crucial to distinguish the Low Earth Orbit (LEO) and the interplanetary space beyond LEO. In this section, the suborbital flights are not considered, as they operate at a low altitude avoiding the ERBs. On the ISS (LEO), the astronauts are partially protected from SPEs and GCRs due to the magnetosphere [20]. While in deep space, missions do not benefit from the protection against planet atmospheres against SPEs or GCRs. Consequently, SPEs and GCR will stumble the spacecraft with fluxes in a position-dependent manner. The average radiation dose-equivalent rate is around 4.3.84 mSv/day, three times higher than in LEO [14].

Lack of a strong global magnetic field and the thin atmosphere on Mars result in only minimal protection from radiation on the surface of Mars. According to the MSL-Rad workshop data, Mars surface and ISS radiation dose rates are similar, 0.213 mGy/day and 0.240 mGy/day, respectively [21]. The current maximal exposure of an astronaut to radiation, according to NASA indications, is set to 3% of the risk of exposure-induced death (REID) cancer fatality with a 95% confidence interval (C.I.) [22].

CNS Neoplasms Overview

The term central nervous system neoplasms refer to a group of heterogeneous benign and malignant tumors [23] which ranges from an extremely invasive and nearly untreatable Glioblastoma multiforme to a non-invasive and treatable pilocytic astrocytoma. The CNS tumors can either be primary or secondary. They are the most common solid tumors in children in the USA, and responsible for approximately 15–20% of all childhood cancers. They are the leading cause of death in children between 0 and 14 years [24–26]. CNS tumors are estimated to occur with an incidence rate of 23.8 per 100,000 people in adults, and they account for 2% of all cancers [24].

The classification of this group of tumors has always been challenging, and it has been under constant revision and update since Bailey and Cushing's publication in 1926 [27].

Currently, the WHO 2016 Classification—in comparison to the WHO 2007 classification based exclusively on histogenesis [28]—categorizes CNS tumors into four grades also basing on molecular markers and genetic factors [6]. Moreover, new changes to the classifications of diffuse glioma have been suggested by the cIMPACT (the Consortium to Inform Molecular and Practical Approaches to CNS Tumor Taxonomy) with an upcoming fifth edition of WHO Classification in the current year 2021 [29, 30].

The criteria of WHO Classification are: (1) anaplasia, (2) mitotic activity, (3) endothelial cell proliferation, and (4) necrosis [31]. Thus, the grading of a CNS tumor is based on these criteria.

Grade I tumors do not meet any criteria, they are benign and slow growing tumors with a good prognosis, i.e., Juvenile Pilocytic Astrocytoma. On the contrary, Grade II tumors fulfill the criterion of anaplasia. They are either malignant or non-malignant slow growing tumors with the potential to recur as higher-grade tumors. For instance, diffuse astrocytoma falls in the Grade II category. Anaplasia and mitotic activity are the two criteria met by III grade tumors like anaplastic astrocytoma. They are malignant tumors that can progress to higher-graded tumors. Grade IV tumors, such as glioblastoma multiforme (GBM), meet all three or four of the above-mentioned criteria. They have a rapid reproducing rate and they are considered to be aggressive malignant tumors [32]. These CNS Grades are predicted to be switched into Arabic numeral nomenclature according to the WHO fifth edition preview [29]. The grading of some selected tumors is consultable in Fig. 8.1.

Glioma, a category of malignant brain tumors that includes high-grade glioma or glioblastoma and low-grade gliomas, is the most common histological form of primary

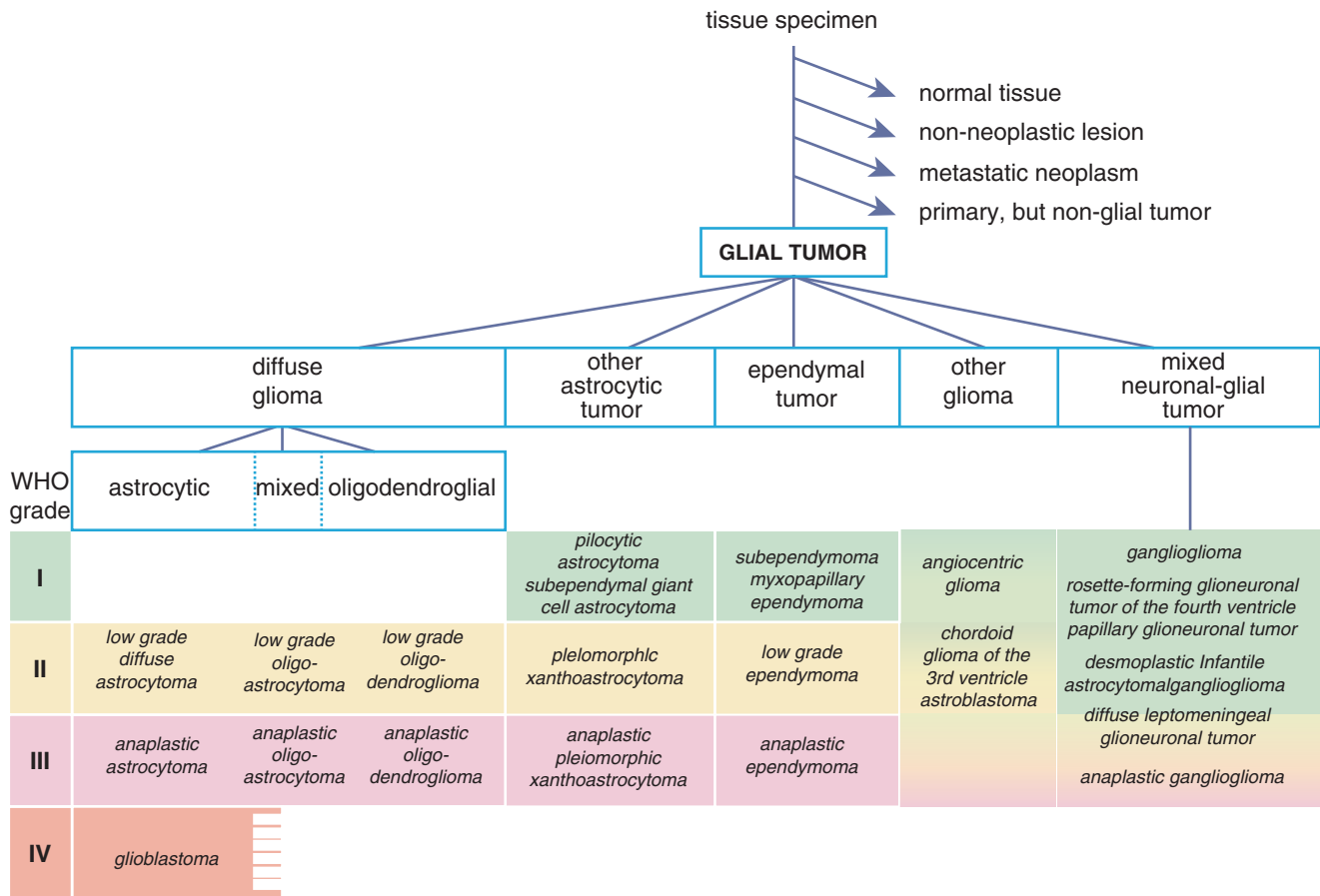


Fig. 8.2 Decision tree for histologic diagnosis of glial and neuronal-glia central nervous system neoplasms. Reprinted with permission from [34]

CNS cancer, therefore, this chapter will be focusing mainly on their classification and description [33]. Gliomas originate from progenitor glial cells or stem cells, and they mirror the glial characteristics after undergoing neoplastic transformation. There are several kinds of glial tumors, i.e. diffuse glioma, other astrocytic glioma, ependymal tumor, other glioma, and mixed neuronal-glia tumor (Fig. 8.2) [34].

Diffuse gliomas account for the vast majority of glial neoplasms in adults. They are defined by diffusive infiltration growth and tumor cell migration into the CNS parenchyma over large distances. The WHO grade II and grade III astrocytic tumors, the grade II and III oligodendrogliomas, the grade IV glioblastomas, and the associated diffuse gliomas of childhood are all classified as diffuse gliomas. The main molecular markers employed in the diagnosis of glioma are isocitrate dehydrogenase (IDH) mutation, chromosomal arm 1p19q deletion, O⁶-methylguanine-DNA methyltransferase (MGMT) promoter methylation, telomerase reverse transcriptase (TERT) promoter mutation, alpha-thalassemia retardation syndrome linked (ATRX) mutation or loss of nuclear expression, and tumor protein p53 (Tp53) [6]. Other markers introduced by the cIMPACT are H3 K27M mutation

and H3.3 G34 mutation, EGFR amplification, CDKN2A homozygous deletion, and +7/-10 genotype [35].

Briefly, IDH marker is at the core of differential diagnosis between glioma and gliosis, and in astrocytoma, oligodendrogloma, and even in 10% glioblastoma, it is positive. Mutation in both IDH 1 and 2, known as IDH Mutant, while the negativity to both forms of IDH is referred to as IDH wild type [36]. The 1p/19q co-deletion, on the other hand, is correlated with the diagnosis of Grade II and Grade III (anaplastic) oligodendrogloma. It plays an important role also in the prognostication of the outcome, and it is linked to procarbazine–lomustine–vincristine (PCV) chemotherapy sensitivity [37]. Basing on these two markers, diffuse gliomas have been classified into diffuse astrocytic tumors IDH-wildtype, diffuse astrocytic tumors IDH-mutant and oligodendroglial tumors IDH-mutant and 1p/19q-codeleted. Where it is not possible to conduct proper molecular testing, the tumor falls into the category of not otherwise specified (NOS).

In gliomas, TERT mutations in the promoter region (C228T and C250T) predict poor survival and radiotherapy resistance, especially in glioblastoma and oligodendroglia-

oma [38]. On the other hand, the methylation of the *O*⁶-methylguanine-DNA methyltransferase (MGMT) gene promoter acts as a positive prognostic factor in GBM patients [39].

Grade II, diffuse astrocytomas, known also as low-grade infiltrative astrocytomas, can be IDH mutant or wild-type [40]. It affects mainly young adults with a mean age of 35 years. In almost 40% of the cases this grade II CNS tumor presents with seizure, and depending on the location and size of the lesion, it can induce focal neurological disfunctions [41, 42]. According to the cIMPACT-NOW recommendations, Astrocytoma IDH-mutant WHO grade II, would be graded as Astrocytoma, IDH-mutant, WHO grade 2, characterized as well-differentiated, lacking histologic features of anaplasia and with low or absent mitotic activity. Microvascular proliferation, necrosis, and CDKN2A/B homozygous deletions are absent [43]. A safe total resection and radiographic follow-up are indicated in this type of gliomas [44].

Grade III, anaplastic astrocytoma (AA), is a rapidly growing, diffusely infiltrating tumor with a median age of onset around 41 years [28]. It can also be IDH mutant or wild-type. Depending on the location of the tumor, the clinical manifestation is mutable. The symptoms include focal or generalized neurological deficits, headaches, visual and sensory impairment, strength loss, and gait disturbances; seizures are less common in anaplastic astrocytomas in comparison to low-grade gliomas [45]. The new recommendations characterize AA IDH-mutant WHO grade III glioma as an Astrocytoma, IDH-mutant, WHO grade 3 that manifests focal or dispersed anaplasia in concomitancy of significant mitotic activity. Microvascular proliferation, necrosis, and CDKN2A/B homozygous deletions are absent [43, 46]. Where necessary and regardless of the mutational status of the IDH gene, the first therapeutic strategy in the treatment of AA is a maximal safe surgical resection along with radiographic follow-up and chemotherapy as per Stupp protocol [47].

The previously classified oligodendroglioma, IDH-mutant, and 1p/19q-codeleted, WHO grade II would be remain oligodendroglioma, IDH-mutant, and 1p/19q-codeleted, WHO grade 2 in the new recommendations [43]. Oligodendrogliomas constitutes around 5% of primary brain tumors and in most of the cases the symptoms are non-specific such as headache. Seizure is experienced in around 35–85% of the cases. Surgical therapy, chemotherapy, and radiation therapy are the main treatments of oligodendroglioma [48].

Glioblastoma (GB), a grade IV glioma, is one of the most aggressive brain tumors, with an estimated survival time of just 15 months after diagnosis [6]. This tumor can be either primary—in case of a de-novo development—or secondary, progressing from a low-grade glioma [49]. The former is denominated in the 2016 WHO Classification as IDH wild-

type, while the latter as IDH-mutant for its various pathways of progression [50]. Both forms of GB have the same characterization, such as necrosis, pleomorphism, and vascularization. Early relapse is caused by its high resistance to radiotherapy and chemotherapy, as well as incomplete surgery due to diffuse invasion of the guerrilla cells [50, 51]. The main symptoms referred by the patients are headache—due to high intracranial pressure—seizure, cognitive impairment, and nausea. The therapeutical indications consist in a safe total resection followed by radiation and temozolomide. This type of tumors is very hard to treat, therefore, various clinical trials and studies are in progress. The main sword of Damocles in the treating of GB is represented by the heterogeneity dictated by the glioblastoma stem cells (GSCs) which were first described in 2003 [52]. These GSCs, as “the apex of a dynamic network,” are renominated for their two key features being self-renewal and differentiation [53]. The heterogeneity which consists in the unpredictability of cancer cells’ subtypes across individual tumors, de facto, seems to limit the efficacy of selective targeting of oncogenic pathways and of tumor microenvironment [54]. According to the new recommendations, glioblastoma, IDH-mutant, WHO grade IV should be renominated as Astrocytoma, IDH-mutant, WHO grade 4, and the previous wild-type grade II diffuse and anaplastic astrocytomas, as well as glioblastoma, IDH-wildtype, WHO grade IV are suggested to be classified as glioblastoma, IDH-wildtype, WHO grade 4 for their poor outcome predicted by TERT, EGFR and +7/–10 genotype [43]. The glioblastoma, IDH-wildtype, WHO grade IV, can moreover be classified as Diffuse hemispheric glioma, H3.3 G34-mutant, WHO grade 4 in the presence of a missense mutation in the Histone H3.3 protein, codon 34.

Methodology of Studying CNS Neoplastic Behavior in Space

Real microgravity studies are expensive and rare; thus, on-ground simulated microgravity (SMG) is more prevalent. Several contemporary devices are used to simulate microgravity such as random positioning machine (RPM), rotating wall vessel (RWV), and fast rotating clinostats (Fig. 8.3) [55, 56].

The main tools utilized in the studies taken in consideration in this chapter are: the 2D clinostat system and the 3D clinostat system. The 2D clinostat system is a 3-dimensional rotational device which rotates around 1 (2D) axis (Fig. 8.3). Essentially, it is a rotating device which prevents the biological system from achieving a sustained gravitational acceleration vector [57, 58]. A random positioning machine (RPM), known also as 3D clinostat, is a simulator based on the principle of vector averaged gravity [59]. This simulator is often compared to the 2D clinostat although it has several differ-

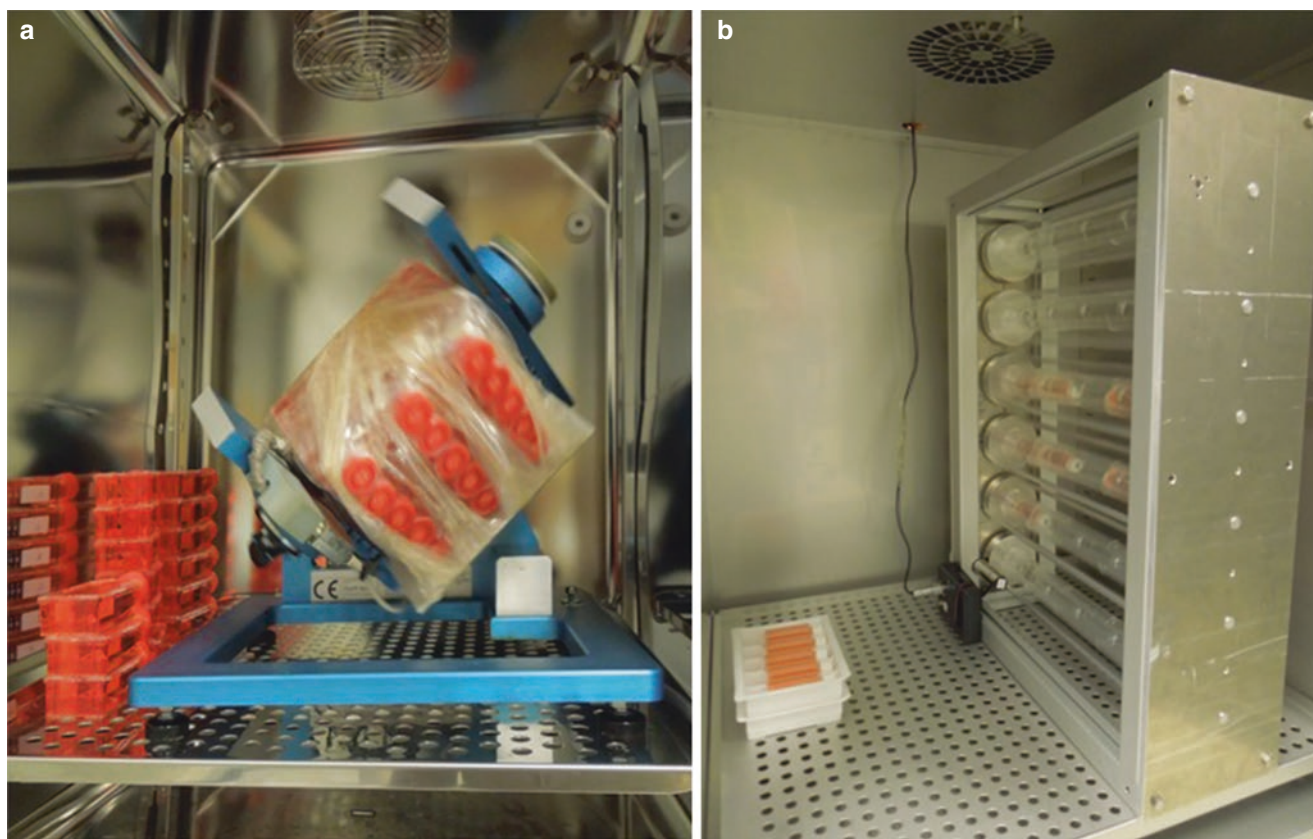


Fig. 8.3 Random positioning machine (a) and 2-D Clinostat (b). Copyright: © 2015 Svejgaard et al. This is an open access article distributed under the terms of the Creative Commons Attribution License,

which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited

ences. The 3D clinostat presents two axes of rotation and a major swimming velocity [55]. The RWV, instead, is a bioreactor of 5–20 cm diameter with liquid-filled container that rotates around a horizontal axis at 10–20 rpm [60].

Most of the studies are made *in vitro* on 2D or 3D cultures. 2D cultures are well known to be simple, with a low time of culture formation, and low-cost maintenance but they have the disadvantage of not portraying faithfully the natural structures of tumors. Further disadvantages lay in the diverse phenotype loss, lack of representation of the cell-cell and cell-extracellular interactions and a monolayer composition which translates into an unlimited metabolic resource on contrary to the *in vivo* cells [61]. The other limitation of 2D cultures, as reported by Birgersdotter et al., is the change in gene expression and splicing, topology and the cellular biochemistry [62]. On the other hand, 3D cultures are beloved for the faithful imitation of *in vivo* tissues and organs, and for the recreation of proper interactions of cell–cell and cell-extracellular environment [61]. Also, the morphology, the phenotype and cellular reproduction phases, as well as the molecular mechanisms, are preserved in 3D cultures in contrast to the 2D cultures [62, 63]. The

limitations of 3D cultures reside in a longer time of culture formation, in a worse performance in culture quality and in expensive costs [64, 65].

The analysis part of each studies varies depending on the type of cells and the objective of studies.

The ideal method to observe the behavior of CNS cancer in space would be to have *in vivo* models analyzed in space. In Cancer and Health research in space, a conference paper, it was intended to examine human GBM derived cancer stem cells (GSCs) in space. The GSCs would be inoculated into mice brain and subsequently, 12 healthy mice, as control, and 12 mice with GBM, sent on the ISS, whereas 24 mice would be maintained in on-ground laboratories under observation for the corresponding on-ground experiments.

Procedures involving animals and their care would be conducted in accordance with the national and international guidelines of the National Institutes of Health Guide (NIH) for the care and use of laboratory animals. On board the ISS, mice will be kept in special cages, previously used by Japan Aerospace Exploration Agency (JAXA) [66], and they would be monitored 24/7 with internal cams. Cages will be equipped with automatized systems to provide food and water, hygiene

and adjust sleep/wake cycles. At the end of the mission, mice would be examined with behavioral tests through our specifically projected maze to evaluate their cognitive abilities and scanned with MRI to rate the volumetric variation in dimension and vascularization of the tumor mass. Furthermore, tumor mass would be explanted and study at morphological, cellular, molecular, and genetic levels. The core point of this project consists on the possibility to study cancer models *in vivo*, rather than *in vitro*, on the ISS. Unfortunately, for economic restrictions the project is in stand-by. Recently, Larose et al. published a paper on their intention to analyze tumors in space. They intend to observe by 2025 human organoids on the Chinese space station [67].

CNS Neoplasms Behavior in Space

Carcinogenesis in Space

Carcinogenesis in space may differ in comparison to the carcinogenesis on-ground. Thus far, no experiments have been conducted in high linear energy transfer (LET) due to obvious limitations, and our knowledge regarding carcinogenesis derives from studies made in low LET. Consequently, there are no sufficient studies on CNS neoplasms carcinogenesis stages in space. Most of the studies are done on survivors of the atomic bomb from Japan.

Initiation under HZE ions—high energy nuclei originating from GCRs or SPEs [68]—differs from the initiation process described earlier. It produces cluster damage in DNA helix strands, causing multiple lesions, instead of non-clustered lesions [3, 69]. These radiation-induced lesions translate into a genomic instability due to the activation of multiple pathways affecting different carcinogenesis stages.

Promotion and progression stages are also different due to the duplex promoter and initiator role of radiation [70]. As reported by Hanahan and Weinberg, the above-mentioned genetic instability promotes the carcinogenesis through several mechanisms, such as evasion of apoptosis, self-sufficiency in growth signals, insensitivity to anti-growth signals, and limitless replicative potential [71]. Other factors deemed to contribute to the carcinogenesis are the extracellular matrix remodeling, persistent inflammation, and oxidative damage [22, 72]. According to Cucinotta et al. fatal cancer risks are less than 10% at upper 95% of confidence interval, deducing that the risk model is accurate. In the hypothesis that the risk model is incorrect, the percentage of fatality could approach 20% with significant life loss [73].

Nevertheless, the risk assessment of carcinogenesis in space, and in particular of central nervous carcinogenesis, is so far, not possible due to a lack of sufficient data and studies. As a matter of fact, mechanisms and inter-species variations are still poorly understood.

Tumor Suppression in Space: A Dual Theory

In the past couple of decades, particular attention has been given to the cancerous cell behavior in microgravity. Sahebi et al. defined microgravity as a dual edge sword as it is still not clear the exact function of microgravity in relation to cancer [74].

It is hypothesized that microgravity impacts the cancerous cells by repressing survival signaling pathways and inducing apoptosis. De facto, the role of microgravity in cellular viability and apoptosis is demonstrated by the inhibition or downregulation of BCL-2 and Bnip3 anti-apoptotic proteins, and by the enhancement of Bax, p53, Caspase -3, 7,8, and PARP pro-apoptotic proteins [74–76]. Another characteristic of microgravity consists in the ability of preventing the formation of spherical colonies and cell proliferation due to a downregulation of ATM/ATR and CDK1/2 proteins which prevents the transition from the cellular phase S to G2 [74, 75, 77]. A key factor is also the induction of early alterations of cytoskeleton, of extracellular matrix (ECM) and focal adhesions [78–80], as a matter of fact, many studies reported a spheroid formation in some types of cancers [81]. Spheroid cultures are cell clusters organized in 3D which alters some signaling pathways and gives a greater differentiation potential in microgravity to stem cells [82]. Furthermore, it seems that this remodeling of cytoskeleton and ECM is an adaptive response to microgravity [74] and that actin microfilament structures are sensitive to microgravity leading to an alteration of signal transduction [83].

Concerning the CNS neoplasms, several changes in cell viability, proliferation, and apoptosis were studied in the past few decades. Especially, U251MG glioma cell line and U87 cells were analyzed in simulated microgravity in a time-dependent manner. U251 cells and U87 derive both from a malignant glioblastoma tumor but with different phenotypes and variances in nicotinamide nucleotide metabolic process regulation, RNA splicing, glycolysis, and purine metabolism [84, 85].

Deng et al. reported a time-dependent inhibition of U251 cell viability by SMG as well as a blockage of cell cycle in G2/M phases. Additionally, an upregulation of cleaved caspase 3 and 9, and a downregulation of BCL-2 and BNIP-3 were evident after a Western blot analysis [86]. Similarly, Zhao et al. identified an upregulation of p21 and a downregulation of Insulin-like growth factor binding protein-2 (IGFBP-2) (Fig. 8.4) [87]. An upregulation of p21 translates into a major tumor suppression, hence to an increased apoptosis. The Insulin-like growth factor binding protein-2, instead, is one of the over-expressed factors and a biomarker in high-grade glioma. A downregulation of IGFBP-2 is linked to an inhibition of glioma cells proliferation [88]. Additionally to the changes in proliferation, apoptosis, and morphology, SMG effects also the migration and the inva-

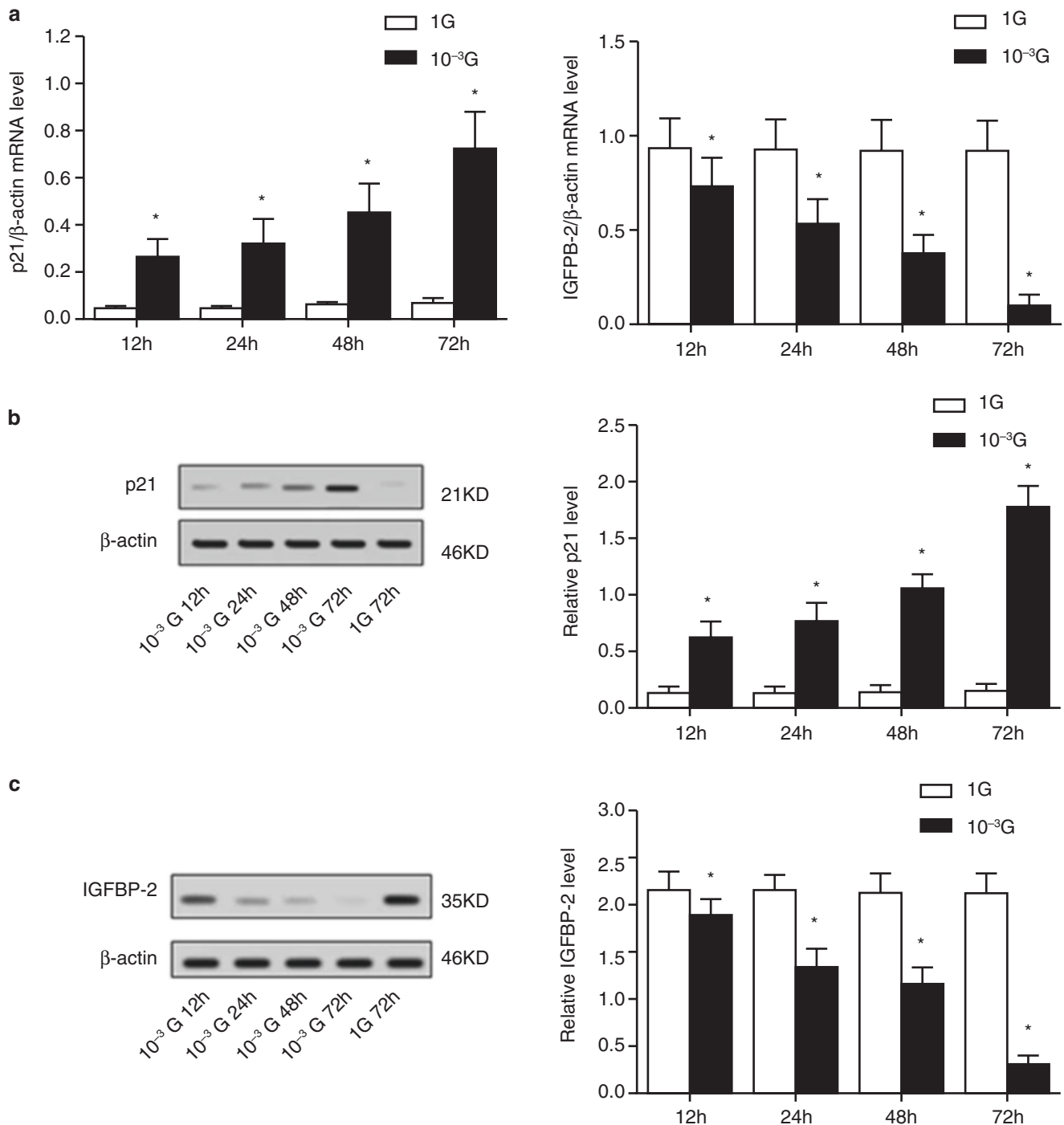


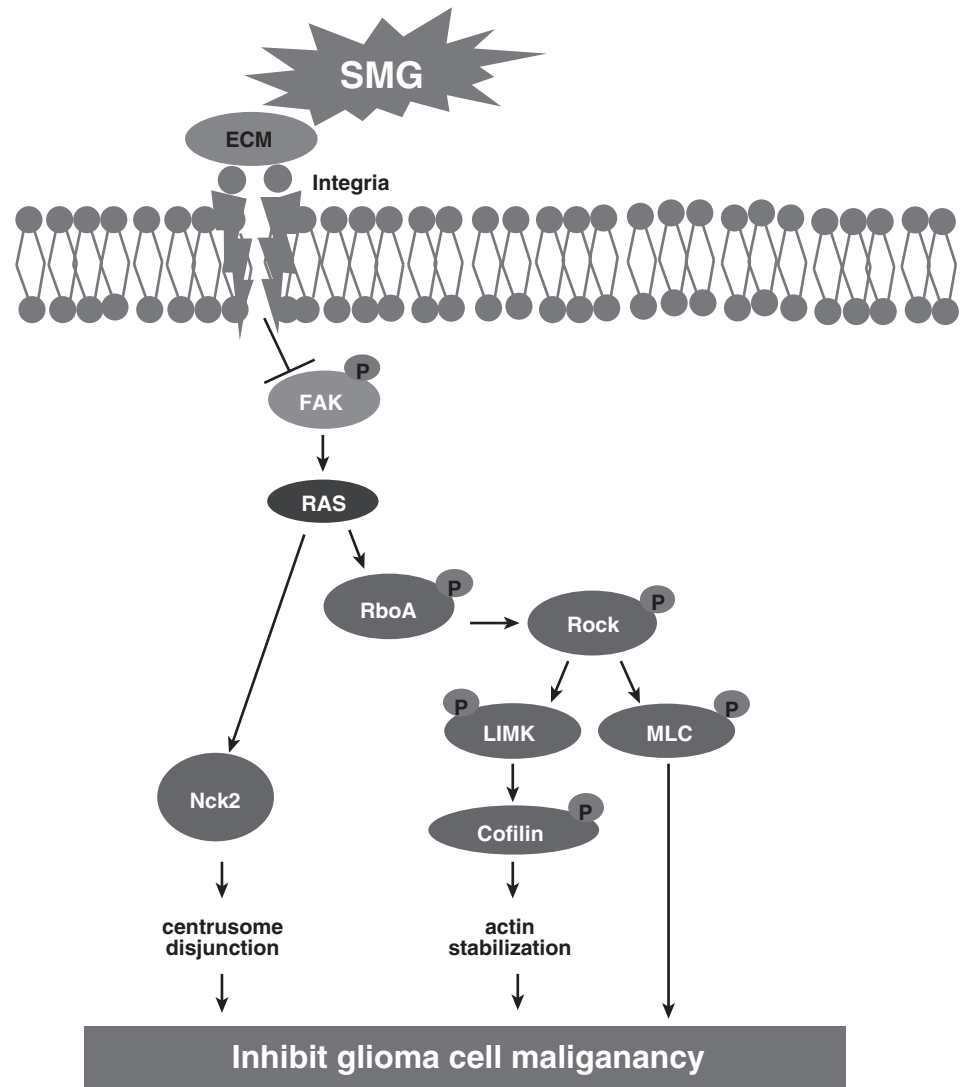
Fig. 8.4 Expression of p21 and of Insulin-like growth factor binding protein-2 (IGFBP-2) in simulated microgravity vs. normal gravity. (a) The bar graph shows the ratios of p21 (left panel) and IGFBP-2 (right panel) mRNA relative to the amount of β-actin mRNA. The west-

ern blot images of p21 and IGFBP-2 are, respectively, shown in (b, left panel) and (c, left panel). The right panels represent the densitometric analysis of the data. β-Actin was used as a loading control. Reprinted and cited with permission from the authors [87]

tion of U87 and U251 cells [86, 89]. In fact, SMG was associated to an inhibition of focal adhesion kinase (FAK), and to a reduced RhoA/Rock signaling and Nek2 expression which transposes into a decreased viability and migration of U251 glioma cells [86] (Fig. 8.5). Focal adhesion kinase (FAK) is

an integrin-based focal adhesion tyrosine kinase and it has a crucial role in the regulation of cytoskeletal networking and cellular signaling [90]. Moreover, FAK appears to be overexpressed in highly invasive tumors and it is interconnected with RhoA/Rock pathway which regulates the cytoskeleton

Fig. 8.5 Systematic diagram of the signaling pathways affected by SMG in glioma cell via FAK activation. Reprinted with permission of the authors [86]



and morphology [91]. By the inhibition of FAK by SMG, also GTP-RhoA gets inhibited with a consequent arrest of Rock, LIMK, MLC, and Cofilin phosphorylation which along with the Nek-2 inhibition results into an inhibited glioma cell malignancy [86]. Likewise, the invasion and migration potentials of U87 cells were found to be effected by SMG, through an inhibition of store-operated Ca^{2+} entry (SOCE) and a subsequent downregulation of Orai1, a cell membrane pore structure, and expression [92, 93].

Overall, majority of the studies in simulated microgravity suggests an inhibition in the glioma cell malignancy which could contribute to the development of therapeutical possibilities. Conversely, ionizing radiations are deemed to enhance the carcinogenesis [94–96]. As a matter of fact, according to Hanahan and Weinberg, the space radiations could lead to DNA damage with subsequent mutations and genomic changes, and to epigenetic changes, i.e., methylation, altered replication or inflammatory responses. These could potentially lead to a genetic instability which could

trigger mechanisms that could lead to an incremented carcinogenesis (Fig. 8.6) [71].

Another important aspect to take in consideration while analyzing the effects of microgravity on tumor suppression is the enhanced sensitizing of cancer stem cells (CSCs) to chemotherapeutic agents [97]. Kelly et al. performed experiments on CSC in a hydrofocusing bioreactor (HBR) and in the rotary cell culture system (RCCS). The HBR is constituted of a 50 mL fluid-filled sphere that rotates at a set speed to furnish a particular hydrofocusing capability that, in the absence of gas bubbles, permits for a low-shear culture conditions in which cells can grow in simulated microgravity [98]. The RCCS is also 50 mL horizontally rotating culture vessel that decreases the shear and turbulence caused by traditional stirred bioreactors, reducing mechanical cell damage, and simulating microgravity [99]. The result of the study indicates that potentially, basing on the core concept of elimination of cancer stem cells which are reputed to be the responsible of tumor recurrence after

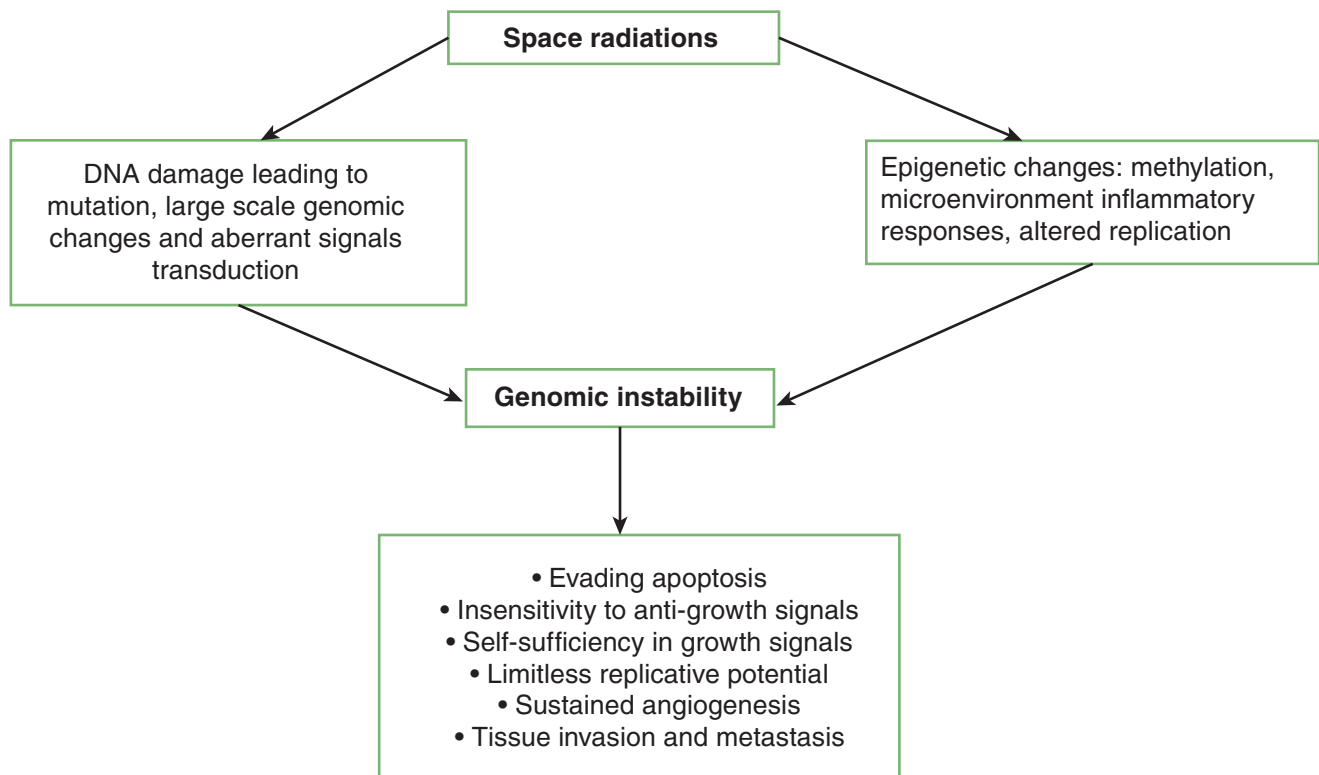


Fig. 8.6 The hallmarks of cancer and possible mechanisms of radiation damage that lead to these changes observed in all human tumors. Readapted from Hanahan and Weinberg, 2000

invasive therapies, it might be possible to develop an anti-cancer therapy through microgravity. As a matter of fact, in T98G, U87MG cell lines CD133 (+) in HBF appear to be more sensitive to chemotherapeutics in comparison to RCCS and normal gravity. Not many studies have been conducted on CD133—positive stem cells, even if they could be an ideal target for further therapy implementations [100]. Also, Takeda M et al. reported an increased chemosensitivity to cisplatin in microgravity in GBM cells, suggesting microgravity might serve as an expectable role of protection for GBM patients [2]. These results were confirmed also by Yuan et al. in their study as well [101]. Unfortunately, studies on other solid CNS tumors in microgravity is lacking. No other relevant studies were found after researching PubMed, Cochrane or Google Scholar databases. This constitutes a limit in the understanding of CNS neoplasms behavior in space.

As the radiation has been largely considered as an initiator of cancer through induction of DNA mutation, and on the other hand given the potential tumor suppressive and sensitizing to chemotherapeutic characteristics of microgravity, the questions is whether to protect or to expose. Thus, given the dichotomic outcomes predicted, reported in literature, the main question would be if the astronauts are subject to incremented risk of malignant CNS tumors following prolonged space missions, or if exposure of patients with CNS

neoplasms to the space environment will result in tumor suppression? Will tumor suppression effect of microgravity balance the potential carcinogenesis mediated by ionizing radiations? To answer to this dilemma, further studies are needed to analyze the combined effects of microgravity and ionizing radiations in pharmaceutically treated, non-treated, and control subjects.

History of Literature

Several studies have addressed the effects of spaceflight on CNS [1, 102, 103], but, hitherto, no clear view has been obtained regarding the behavior of CNS neoplasms or generally, of cancer, in space.

Already in 2001, Cucinotta et al. have expressed their perplexity regarding the lack of knowledge and the uncertainty deriving from it in matter of cancer behavior and risk assessment [104]. In late 90s, several researches addressed the possibility of a higher cancer induction due to high-LET radiations in space compared to the normal X-rays (Low-LET radiations) with the consequence of a permanent damage to the CNS independently from the site and typology of tumor [105, 106]. Uncertainty remains the key word also today regarding the risk assessment of carcinogenesis in space. As discussed earlier the effect of radiations depends also on the type of radiation taken

in consideration, the amount of radiation on LEO (ISS) or Mars surface is different to the one in deeper space [14, 107]. In addition to the studies related to radiation and cancer risk assessment, further studies have been made on the role of microgravity. As discussed in the tumor suppression part, up to day, it is possible that simulated microgravity inhibits the malignancy of high-grade central nervous system cancers [86, 87]. Whereas, it still remains unclear the combined effects of radiations and microgravity in space, placing a shadow on the results and the future of long-term space colonization.

Literature Review Methodology

Concerning the CNS Neoplasm tumor-suppression part, a literature review has been conducted by May 2021. PRISMA guidelines were the point of reference for the literature review. PubMed was the database of reference and several keywords were utilized. Keywords employed were: “Central nervous system neoplasm AND microgravity”; “Central nervous system neoplasm AND spaceflight”; “Solid tumor AND Microgravity”; “Cancer stem cell AND microgravity”; “Glioblastoma AND microgravity”; “Glioblastoma AND spaceflight”; “Cancer Stem Cell AND spaceflight”; “Tumor suppression AND spaceflight”; “Tumor suppression AND microgravity.”

Foreign language literature was excluded. Zotero software was used to manage citations, abstracts, and documents.

The search strategy returned 124 references. Of these 26 were eliminated as duplicates, and a further 82 were excluded at the title and abstract screening stage. The remaining 16 papers were included for full-text screening. Of these 16 papers 7 were included in the study. An additional study was included through citation searching (Fig. 8.7).

Limitations

Several limitations have to be addressed in this chapter. As discussed earlier, a clear view of the CNS neoplasm behavior is not currently available. This may be due to financial or ethical concerns.

Despite extensive space radiation research, significant uncertainties remain in predicting the biological consequences for humans as terrestrial simulations differ from an actual spaceflight environment [108]. As reported by Chancellor et al., it is arduous to properly simulate the spectrum of energies, ion species, concentrations, and dose rates found in the space radiation world.

Additionally, the information available for extrapolating radiation risk concerning the spaceflight is restricted by several factors such as limitations in the emulation of the radiation environment, and choice of surrogate animal model. Also the impossibility to delivery of sufficient complexity, rate, and magnitude of doses can be considered as important limitations [108].

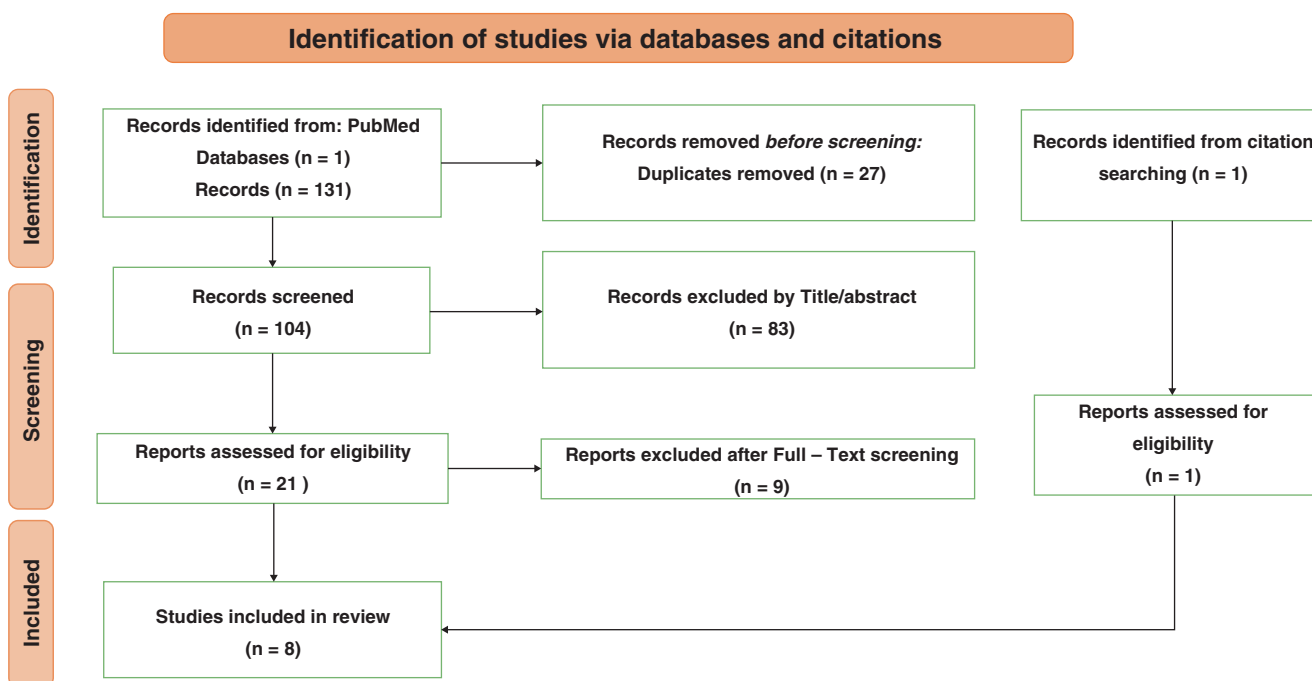


Fig. 8.7 Literature review methodology: Readapted from PRISMA 2020 flowchart

Furthermore, observations in studies conducted in a simulated microgravity environment on Earth may differ from real microgravity. The principle of clinostats and random positioning machines is to vary the gravity between -1 g and $+1\text{ g}$ with the purpose of achieving an average of 0 g over time [109]. Hydrostatic gradients are still present, even if the vector varies. This is different from a real and sustained microgravity.

Conclusion

Heretofore, the precise behavior of central nervous neoplasms in space is dismal. Currently, experiment results in simulated microgravity seem to be auspicious for a possible usage of microgravity as a tool for therapies. The role of microgravity in space is hypothesized to be akin to the simulated microgravity. The correlation between microgravity effects and space radiations remains obscure, given the speculated propension of a carcinogenesis enhancement under ionizing radiations. Several questions remain still open. With the current data, it would be preliminary to declare that extended missions would increase CNS cancer risk, and it is not possible to assert that microgravity could lead to tumor suppression. The dual sword theory remains pertinent. The CubeSat to study Solar Particles (CuSP) spacecraft, on board of Artemis, might reveal further information in terms of space radiation, widening the understanding, thereby, also of the related cancer risks. Further researches are needed to clarify the aspects and questions raised in this chapter, also in the light of the information which will be obtained from the upcoming missions.

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