**Environmental Challenges and Solutions** *Series Editor:* Robert J. Cabin

# Telma Encarnação Alberto Canelas Pais *Editors*

# Marine Organisms: A Solution to Environmental Pollution?

Uses in Bioremediation and in Biorefinerv

# **Environmental Challenges and Solutions**

#### Series Editor

Robert J. Cabin, Brevard College, Brevard, NC, USA

The Environmental Challenges and Solutions series aims to improve our understanding of the Earth's most important environmental challenges, and how we might more effectively solve or at least mitigate these challenges. Books in this series focus on environmental challenges and solutions in particular geographic regions ranging from small to large spatial scales. These books provide multidisciplinary (technical, socioeconomic, political, etc.) analyses of their environmental challenges and the effectiveness of past and present efforts to address them. They conclude by offering holistic recommendations for more effectively solving these challenges now and into the future.

We are welcoming proposals specifically on the following topics:

- Environmental sciences and social aspects
- Environmental sciences and human conflicts
- Boosting de-carbonization of energy system to face climate and economic emergency
- Climate emergency and health: sources, science-based analysis and global solutions
- Indoor and outdoor chemicals, environmental quality and public health in a changing world
- Demographic, climate and environmental change: evidence and recommendation for global sustainability
- Technological, industrial and corporate change for supporting environmental and social sustainability
- Nanotechnology applied to water decontamination/purification processes
- Sorbents and other materials in environmental protection
- Raw materials supply and circular economy
- Strategies to boost clean mobility

All books are written in a concise and readable style, making them suitable for both specialists and non-specialists starting at first year graduate level. Proposals for the book series can be sent to the Publishing Editor, Éva Lörinczi eva. loerinczi@springer.com. Telma Encarnação • Alberto Canelas Pais Editors

# Marine Organisms: A Solution to Environmental Pollution?

Uses in Bioremediation and in Biorefinery



*Editors* Telma Encarnação Coimbra Chemistry Centre (CQC-IMS), Department of Chemistry, Faculty of Sciences and Technology University of Coimbra Coimbra, Portugal

Alberto Canelas Pais Coimbra Chemistry Centre (CQC-IMS), Department of Chemistry, Faculty of Sciences and Technology University of Coimbra Coimbra, Portugal

ISSN 2214-2827 ISSN 2214-2835 (electronic) Environmental Challenges and Solutions ISBN 978-3-031-17225-0 ISBN 978-3-031-17226-7 (eBook) https://doi.org/10.1007/978-3-031-17226-7

 $\ensuremath{\mathbb C}$  The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Switzerland AG The registered company address is: Gewerbestrasse 11, 6330 Cham, Switzerland

### Foreword

I am delighted to provide a Foreword to this book *Marine organisms: a solution to environmental pollution?*, edited by Telma Encarnação and Alberto Canelas Pais, with contributions from a number of other well respected workers in the field.

Our contemporary society, with its ready access to the convenience of fast communications, mass transportation, good illumination, home entertainment, etc. has developed over the last three Centuries through advances in science and technology. This has involved a series of industrial revolutions, based on increasing mechanisation, automation and mass production, with the involvement more recently of computer control, artificial intelligence and robotics. This has led to the demand for new materials, such as polymers, alloys and advanced ceramics to replace the traditional ones, such as wood and stone. The supply of the new materials, and their precursors, has been one of the factors responsible for the development of the chemical industry, with its consequent impact on the environment.

Chemicals are vital in almost all areas of our life, including medicine, agriculture, food and drinks, textiles, construction, packaging, personal care and domestic care products. Although most of these chemicals are beneficial for society, uncontrolled or inappropriate use of them can also have serious negative consequences for the environment. High priority must be given as a major goal to minimising such chemical pollution through the development of cyclic systems, or control or elimination of chemical pollutants by using some technique to transform them into innocuous species. The ideal scenario would be complete mineralisation, where pollutants are transformed into water, carbon dioxide and other small molecules.

Possible methodologies for pollutant treatment involve the use of chemical oxidants, such as hydrogen peroxide, photochemistry (using ultraviolet or visible light), frequently with a catalyst such as titanium dioxide, high energy radiation, and the use of various biological organisms. This book focuses on the latter approach using marine-based microorganisms.

Care must be taken in the treatment process that any reaction intermediates are not more toxic than the original pollutants. For detailed study of pollutant degradation, identification of all species involved and study of their effect on the viability of microorganisms is desirable.

It is sometimes convenient to distinguish between synthetic chemicals, such as aspirin, and naturally occurring ones, like water and oxygen. However, the fact that a substance occurs in nature does not necessarily mean that it is innocuous. Carbon dioxide is a colourless gas, that is essential for processes such as respiration and photosynthesis, However, its presence above a certain concentration can prove fatal. It is also implicated as a greenhouse gas in global warming.

Environmental pollution can be described in terms of the bulk phase involved (water, air, soil...), type of pollutant (pesticides, pharmaceuticals...), chemical category (polycyclic aromatic hydrocarbons (PAHs), heavy metals, polychlorinated compounds...) or by chemical structure (carbon monoxide...). Methods of pollutant treatment frequently show selectivity in terms of chemical structure, and in the case of water treatment may depend on parameters such as pH, salinity and temperature.

The choice of treatment method depends on a variety of factors. Economic parameters are of particular importance, and the use of plants and microorganisms is particularly attractive in this respect. In addition to their Green credentials, since they do not normally require the use of any expensive chemicals, biological processes frequently produce biomass, which is a value added product that can be used as a feedstock for a variety of materials, including pharmaceuticals, biopolymers, fertilisers and biofuels, and may be a source of novel chemicals. This makes such systems particularly attractive, and economically viable, for the treatment of environmental wastes. Their potential is further enhanced by the number and diversity of species available. Although until recently the use of marine sources for bioremediation has been limited, they are attracting increasing interest for various applications in biotechnology. In part this is because they are rich in a variety of interesting species, with a number of actual and potential applications.

In this book, *Marine organisms: a solution to environmental pollution?*, Telma Encarnação, Alberto Canelas País and co-authors present an excellent overview of the potential of marine based species such as microalgae, fungi, bacteria, yeasts and sponges in bioremediation and in the preparation of biobased materials. It makes an important contribution to environmental technology and pollution control, and its multidisciplinary approach makes it attractive to research workers in a wide variety of areas, including biotechnology, microbiology, chemistry, and environmental sciences.

It is well structured and provides an excellent introduction to this rapidly developing field. A succinct introductory chapter is followed by 11 chapters covering various aspects of bioremediation, treatment of specific pollutants, and biomass valorisation using different marine based microorganisms. The book concludes with a chapter on environmental management. A comprehensive and up-to-date bibliography provides rapid access to the current research in this area.

This book fills a gap in the literature on the use of marine microorganisms in the treatment of pollutants. It also acts as a valuable source of information on the application of such systems in the production of value-added biomass, makes an excellent contribution to the fields of biotechnology and bioremediation, and

provides valuable information on how these species can be used for sustainable bioharvesting of novel molecules. These may have interesting properties for applications such as pharmaceuticals, cosmetics, and fine chemicals.

In conclusion, the book, *Marine organisms: a solution to environmental pollution?*, makes an important contribution to the literature and provides an excellent testimony to the authors well recognised scholarly insight in this area.

Coimbra Chemistry Centre (CQC-IMS), Department of Chemistry, University of Coimbra, Coimbra, Portugal Hugh D. Burrows

# Contents

1	Introduction: Environmental Pollution and Biotechnological	1
	Telma Encarnação, Maria da Graça Campos, and Artur Mateus	1
2	Bioremediation Using Microalgae and Cyanobacteria and Biomass Valorisation	5
3	Removal of Heavy Metals and Organic Pollutants by Marine Microalgae	29
4	Algal-Bacterial Consortiums, from Fundamental Interactions to Environmental Applications	65
5	<b>Biodegradation of Environmental Pollutants by Marine Yeasts</b> Danouche Mohammed, El Arroussi Hicham, and El Ghachtouli Naima	79
6	Sustainable Direct Digital Manufacturing Using MarineResourcesLuís André R. Marques, Sara Biscaia, Anabela Massano,Rafael M. Tavares, and Artur Mateus	93

7	Exploiting Marine Fungi in the Removal of Hazardous Pollutantsand Biomass ValorisationDushyant R. Dudhagara, Bhumi M. Javia, and Anjana K. Vala	117
8	Marine Bacteria for Bioremediation Paolo Stincone, Robson Andreazza, Carolina Faccio Demarco, Thays França Afonso, and Adriano Brandelli	147
9	Marine Bacteria for Biofertilizers Poonam Singh, Kaleemunnisa FNU, and Telma Encarnação	189
10	Marine Sponges for Bioremediation Purposes and for SecondaryMetabolites ProductionIsmail Marzuki and Khairun Nisaa	205
11	Genetic Modification: A Gateway to Stimulate the Industrial Production of Biofuels	237
12	<b>"Omics" Technologies in Biodegradation Processes</b> Sevcan Aydin and Mahmut Çalışkan	261
13	Conclusion: Environmental Protection—Our Common         Responsibility         Alberto A. C. C. Pais and Telma Encarnação	267

## **Chapter 1 Introduction: Environmental Pollution and Biotechnological Solutions**



Telma Encarnação, Maria da Graça Campos, and Artur Mateus

**Abstract** Progress, wealth, comfort, increased productivity and economic growth are some of the repercussions that emerged from the Industrial Revolutions, providing greater benefits and opportunities. But these advances also brought us great challenges, bringing uncertainties for the future. Our social organization, economic models and lifestyles are altering ecosystems and Earth's patterns, causing environmental degradation, and shaping the face of the Earth. But the same revolutions that brought us challenges can provide us with opportunities to restore the environment. And biotechnology can unlock potential solutions for a more sustainable and resilient future.

**Keywords** Industrial Revolutions · Environmental pollutants · Biotechnology · Bioremediation · Circular economy · Industry 5.0

The Industrial Revolution marked significant and remarkable milestones in human history. They spurred faster progress in several areas that benefit human life and led to economic, political, and societal changes, changing life in unforeseen ways. The First Industrial Revolution began in Great Britain after the 1750s with the introduction of hydraulic power and steam engine, which led to the mechanization of

M. da Graça Campos CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

A. Mateus Centre for Rapid and Sustainable Product Development, Polytechnic Institute of Leiria, Marinha Grande, Portugal

T. Encarnação (⊠)

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

Centre for Rapid and Sustainable Product Development, Polytechnic Institute of Leiria, Marinha Grande, Portugal

PTScience Avenida do Atlântico, Lisbon, Portugal e-mail: tencarnacao@qui.uc.pt

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_1

agriculture, manufacturing, and transportation. In 1870, the Second Industrial revolution started with the generation of electricity, shortly after its discovery, incorporating electromotive force in the industry, enabling mass production for the first time. A century later, in the 1970s, the Third Industrial Revolution started with the invention of the transistor and the first microchip; the advances in computer technology after the Second World War triggered further development in the automation of the production process. Since then, a digital revolution has been disrupting traditional industries. A technological fusion of physical, chemical, digital, and biological dimensions is transforming industry, economies, jobs, and society itself, leading us to a New Era, the Fourth Industrial Revolution or Industry 4.0. Although no consensually accepted definition exists yet, the Industry 4.0 concept can be described as the advent of cyber-physical systems (CPSs), the Internet of Things (IoT), and services (MinHwa Lee et al. 2018) which have a significant impact on the efficiency of the production processes and in product development. The COVID-19 pandemic forced a rapid transition to digital technologies, even in sectors that were particularly resistant to the digital transition. The development of the Industry 4.0 technologies such as the Internet, Big Data (BD), blockchain, IoT, Additive Manufacturing (AM), virtual and enhanced reality, and Artificial Intelligence (AI) will probably lead us to radical changes, more human-centered, more sustainable, and more resilient: Industry 5.0. Without a doubt, Industry 5.0 (European Commission 2021) will significantly transform the manufacturing processes and services sectors where collaborative interactions between humans, machines, and systems materialize.

Despite the fact that social inequality may be found in all societies, the industrialized world brought us comfort and wealth in our lives. But this comes with a high price to pay: with all the improvements, a rapid urbanization brought significant challenges to the cities that suffered an increased pressure, such as growing population, lack of potable water, and residues management; a high consumption increased the release of pollutants into the environment. Carbon dioxide, dioxins, phthalates, bisphenols, pharmaceuticals metabolites, pesticides, flame retardants, and so many thousands of chemicals are released into the environment every day and have a dramatic impact on human health and wildlife. Although some damages could become irreversible in a very short time, we still have the opportunity to reverse the trend. We must boost science, technology, and innovation to implement biobased solutions to benefit from the progress that industrial revolutions brought us, reducing the environmental footprint. To achieve that, the entire planet needs to be united in a Green Revolution: civil society, governments, companies, and industries, organizations, all segments of society.

We are living a crisis for resources that will push forward the transition to new and renewable raw materials. Allying this with the emergency of the pollutants mitigation, we will be able to create a structure that allows a sustainable evolution of the planet, both in terms of well-being for humans and the environment.

Among all the tasks that should be done in a very short time, bioremediation, which has already started using plants and microorganisms, allow the production of biomass to be used in different feedstock sources for many industries applications:

for instance, bioplastics, fertilizers, new pharmaceuticals compounds, building blocks, energy, and many other exciting solutions, which unlock new sustainable and eco-friendly possibilities. These challenges also create enthusiastic economic opportunities. These economic outputs should be improved by countries with credible, efficient, and concrete policies that enhance the advancement of sustainable technologies. Science should provide the tools and robust evidence to support decisions before these are implemented into legislation for an ecological transition toward sustainable development.

This book aims to contribute to this fascinating process toward the next industrial revolution. The following chapters will comprehensively discuss bioremediation technologies, to reduce environmental pollution while producing value-added biomass that meets the need for new and better sustainable materials. Various marine microorganisms, such as sponges, microalgae, fungi, bacteria, yeasts, and consortiums of different microorganisms, are all able to biodegrade several families of compounds; for instance, carbon dioxide, PAHs, heavy metals, petroleum sludge, naphthalene and pyrene, pharmaceuticals, persistent organic pollutants. Tailored solutions will be discussed with the primary goal of contributing to zero carbon and zero pollution, and groundbreakingly scientific and technological advances will be critically approached. Legislation and regulatory requirements are also an essential topic of discussion and debate since the release into the market of new developed biobased products can be hindered by legal requirements.

Some Industry 4.0 technologies, such as additive manufacturing and Artificial Intelligence, will be debated in the approached context. The massification application of technologies that result from industry digitization plays and will play an increasingly important role in the development of intelligent solutions. Production and treatment systems will be increasingly monitored and controlled in real time based on machine learning. Waste treatment plants, as well as collection systems, present an increasing application of Digital Twins concepts. Digitization is and should be extended to electromechanical systems for the collection and treatment of waste and water to the point that, in real-time, there is such a collection of data that allows the existence of a digital twin, if possible autonomous and controlled by machine learning (Vitorino et al. 2019). The digital definition of systems (more common on electromechanical systems) must also be followed by the digital definition of products and every tangible material. The increasing of understanding on transforming processes and the increasing of optimization, monitoring and control, allow, the spatial and temporal definition of molecular organization, applied (for example) to additive manufacturing. In this way, this depth increasing of digital control is a precursor to a molecular-level definition of meso parts (from molecules to parts). The products of the future will be designed at the molecular level and implemented with digital, spatial, and temporal control (da Silva et al. 2022).

Can we glimpse the future? Can we glimpse the next industrial revolution, Industry 6.0? It is predicted that possible new technologies may include quantum computing and nanotechnologies. But what about changes in our values and our perceptions? Changes in how we socially relate, work, and connect with Nature for many years separated? Perhaps we can come to say: "Industry 6.0 significantly transformed the world where collaborative interactions between humans, machines, and Nature materialize."

#### References

- European Commission, D.-G. f. R. a. I. (2021) "Industry 5.0 : human-centric, sustainable and resilient." Publications Office
- MinHwa Lee JJY, Pyka A, Won DK, Kodama F, Schiuma G, Park HS, Jeon J, Park KB, Jung KH, Yan M-R, Lee SY, Zhao X (2018) How to respond to the fourth industrial revolution, or the second information technology revolution? Dynamic new combinations between technology, market, and society through open innovation. J Open Innovation Technol Market Complexity 4: 21
- da Silva DP, Pinheiro J, Abdulghani S, Lorger CK, Martinez JC, Solano E, Mateus A, Pascoal-Faria P, Mitchell GR (2022) Changing the paradigm-controlling polymer morphology during 3D printing defines properties. Polymers 14:9
- Vitorino J, Ribeiro E, Silva R, Santos C, Carreira P, Mitchell GR, Mateus A (2019) Industry 4.0 digital twin applied to direct digital manufacturing. AMM 890:54–60

## **Chapter 2 Bioremediation Using Microalgae and Cyanobacteria and Biomass Valorisation**



Telma Encarnação, Pedro Ramos, Danouche Mohammed, Joe McDonald, Marco Lizzul, Nadia Nicolau, Maria da Graça Campos, and Abílio J. F. N. Sobral

**Abstract** Microalgae and cyanobacteria are photosynthetic microorganisms that can be used to bioremediate anthropogenic pollutants from air, water and soil. These organisms can remediate several anthropogenic pollutants, such as carbon dioxide, nitrates and phosphates, heavy metals, pharmaceuticals, pesticides and persistent organic pollutants. The biomass generated in this process can be used as a feedstock source for the production of a multitude of valuable biobased products and applications. Polymers, resins, binders, lubricants, and coatings are some of the promising examples. This chapter provides an overview of the entire process: bioremediation using microalgae and production of value-added products, based

P. Ramos · N. Nicolau PTScience Avenida do Atlântico, Lisbon, Portugal

D. Mohammed Green Biotechnology Center, Moroccan Foundation for Advanced Science, Innovation and Research (MAScIR), Rabat, Morocco

Microbial Biotechnology and Bioactive Molecules Laboratory, Sciences and Technologies Faculty, Sidi Mohamed Ben Abdellah University, Fez, Morocco

Departement of Chemical & Biochemical Sciences-Green Process Engineering (CBS-GPE), Mohammed VI Polytechnic University (UM6P), Benguerir, Morocco

J. McDonald · M. Lizzul Varicon Aqua Solutions Ltd, Varicon Aqua Solutions Ltd, Worcester, UK

M. da Graça Campos · A. J. F. N. Sobral CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

T. Encarnação (🖂)

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

PTScience Avenida do Atlântico, Lisbon, Portugal

Centre for Rapid and Sustainable Product Development, Polytechnic Institute of Leiria, Marinha Grande, Portugal e-mail: tencarnacao@qui.uc.pt

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023 T. Encarnação, A. Canelas Pais (eds.), *Marine Organisms: A Solution to Environmental Pollution?*, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_2

on a biorefinery concept, focusing on circular economy and sustainability. Essential aspects of legislation and regulations are also approached.

**Keywords** Biorefinery · Microalgae · Bioremediation · Biomass · Biofuels · Wastewater · Lipids · Biobased · Circular economy

#### 2.1 Introduction

Microalgae is a diverse group of prokaryotic and eukaryotic photosynthetic microorganisms living in different environments. Microalgae include prokaryotic bluegreen algae (cyanobacteria) and eukaryotic microalgae (diatoms and green algae). These organisms use light energy and carbon dioxide as carbon source and, through metabolic processes, convert them into various biopolymers such as proteins, nucleic acids, lipids, and polysaccharides, releasing molecular oxygen in the process. They produce a wide range of compounds, some of which have potential commercial value. Depending on species, microalgae and cyanobacteria produce different biomolecules, and, depending on the growth conditions, these biomolecules might vary in composition and concentration. This versatility is reflected in the remarkable number of products and applications that can be developed.

Microalgae and cyanobacteria have been cultivated for decades, but only a limited number of species have been commercially used, mainly for pigments production and food and feed supplements. The most commercially used species are *Chlorella*, *Spirulina*, *Dunaliella*, *Haematococcus*, *Nannochloropsis*, *tetraselmis* and *Isochrisis*.

Despite their versatility, resilience, and potentialities, a consolidated and accepted microalgae cultivation system is not yet implemented worldwide. Many factors contribute to this reality: the many decades of established and implemented fossil economy, the confidence from investors, the initial costs of the investment, the downstream processing challenges, and the acceptance and willingness of all the stakeholders (political, industry, civil society). In recent years, the number of microalgae production units has increased and represents a significant step towards a sustainable resource for the future. These aspects will be further discussed in the following sections of this chapter.

#### 2.2 Brief History

The importance of microalgae in wastewater treatment was recognised several decades ago. At the beginning of the twentieth century, the first studies focused on nitrogen/phosphorus uptake alongside oxygenation potential. Early studies on wastewater treatment using algal pond systems were developed in the mid-1950s; these systems were used to study algal growth and photosynthetic oxygen production to assist with bacterial degradation of Biological Oxygen Demand (BOD). In the

1960s, researchers began to focus on nutrient removal from sewage and wastewater, such as nitrates and phosphates. At the beginning of the 1970s, a 300  $m^2$  pilot plant was built and operated for algae wastewater treatment (Borowitzka 2013) and

heavy metals, such as cadmium, chromium and mercury, from wastewaters. The concern with environmental degradation by chemicals is not new. Several reports from the late 1960s and during the decade of 1970s state the problem of persistent pesticides in the environment and the impact on human health. And in this period, many reports were published to address the issue of their presence in underground and superficial waters, mainly using microalgae in the process. A notable a paper published in 1976 reported the removal of two herbicides (amitrole and atrazine), from water, by four microalgae species (*Chlorella pyrenoidosa, Scenedesmus quadricauta, Chlamydomonas reinhardtii, and Euglena gracilis*). During the decade of 1980s, many studies on the uptake of nitrogen and phosphorus from wastewater by microalgae were undertaken. The studies on this subject have been constant over many decades (PierreChevalier 1985, Hammouda et al. 1995, Shi et al. 2007, Eroglu et al. 2012, Ferrando and Matamoros 2020, Castellanos-Estupinan et al. 2022).

semicontinuous cultures of microalgae began to be studied for their ability to remove

Much progress has been made in understanding many aspects of microalgal biology and physiology, nutrition, and cultivation conditions. Less research has focused on the uptake and removal of emerging contaminants. But one of the significant limitations of an implemented microalgae system, limiting further expansion, is the high cost of investment and cost-effective harvesting of biomass and extraction of compounds (downstream process).

In the last three decades, researchers have studied harvesting and extraction techniques involving physical (centrifugation, sedimentation, filtration, flotation, etc.) and chemical processes (e.g. flocculation). However, these are energy-demanding processes, and some chemical processes use toxic chemicals (Hoang et al. 2022). Academy and public research institutions, companies, and government agencies have funded research projects focused on the field of microalgae technology. Nevertheless, it still requires much knowledge and innovation, particularly in the engineering field applied to the downstream processing steps.

The history of each field of science is of great importance since one can analyse what was done by the peers in previous decades, what remained to be studied, what failed, and why it was not moved forward. In the case of the removal of pollutants from the environment, although there was some level of concern in previous decades, it was only more recently that public opinion, governments, world organisations, and companies are more willing to protect the environment by restoration and with more ecological alternatives.

#### 2.3 Microalgae Bioremediation: An Effective Approach Towards Environment Restoration

Globally, the emissions of greenhouse gas of which, carbon dioxide ( $CO_2$ ), methane ( $CH_4$ ) and nitrous oxide ( $N_2O$ ), as well as surface and groundwater contamination by organic and metal pollutants, are the major global environmental issues that have captured the attention of scientists, environmentalists and legislators (Sundarrajan et al. 2019). Various physical-chemical methods have thus been proposed. While they are effective, such methods require expensive chemicals and energy-intensive equipment, rendering the treatment costs high and limiting their large-scale application (Danouche et al. 2021).

#### 2.3.1 Application of Microalgae in CO<sub>2</sub> Mitigation

Sequestration of CO<sub>2</sub> can be achieved by using physicochemical and/or biological approaches. Currently, the main abiotic approaches employed for the mitigation of  $CO_2$  include physicochemical adsorption (Song et al. 2019), direct injection into the deep ocean, old coal mines, oil wells, geological formations such as saline aquifers, and CO<sub>2</sub> mineral carbonation. However, some drawbacks are associated with these approaches, for instance, the control of the physicochemical adsorption process is generally difficult, and the sorbent materials are generally expensive and non-renewable. Additionally, the injection of CO<sub>2</sub> into geological formations presents significant challenges in terms of space requirements and potential leakage over time (Zeng et al. 2011). Naturally, microalgae capture photons from the sun's energy to convert CO<sub>2</sub> dissolved in water (as free CO<sub>2</sub>, bicarbonate (HCO<sub>3</sub><sup>-</sup>), carbonate  $(CO_3^{2-})$ , and carbonic acid  $(H_2CO_3)$ ) to produce organic molecules through the process of photosynthesis (Eze et al. 2018; Fu et al. 2019). As reported by Zhou et al. (2017), microalgae have a high growth rate and a photoautotrophic efficiency 10-50 times higher than that of terrestrial plants, which makes the capture of  $CO_2$  using microalgae as one of the most promising approaches (An et al. 2021). A number of Chlorophyceae species have exhibited a high capacity for  $CO_2$ sequestration. The most studied species for CO<sub>2</sub> fixation from flue gases belong to Chlorella genus, such as C. vulgaris, C. fusca, C. sorokiniana, C. pyrenoidosa and C. kessleri (Kong et al. 2021). The detailed mechanisms of CO<sub>2</sub> fixation and nitrogen assimilation in microalgae are described in Fig. 2.1.

#### 2.3.2 Application of Microalgae in Wastewater Treatment

Phycoremediation, or the use of microalgae for the remediation of wastewater or contaminated aquatic ecosystems by organic or metallic pollutants, has recently



Fig. 2.1 Mechanisms of CO<sub>2</sub> fixation and nitrogen assimilation in microalgae

emerged as a promising, efficient, economical and environmentally friendly strategy compared to other physicochemical processes (Danouche et al. 2020; Singh et al. 2021a, b). The use of wastewater as a culture medium for microalgae is an innovative concept particularly suited to tertiary wastewater treatment. Indeed, microalgae can assimilate a wide range of inorganic contaminants as well as some organic pollutants, besides their capacity to accumulate heavy metals (HMs).

Phycoremediation of Inorganic Pollutants In wastewater treatment systems, microalgae can use several inorganic pollutants during their growth, such as nitrogen, phosphates, chlorides, sulfates and other inorganic pollutants, and may have a major role as intermediates in metabolic activity (Abdel-Raouf et al. 2012). Several studies have highlighted the possibility of using wastewater as a source of nutrients for the cultivation of microalgae, allowing both the elimination of the pollution load and the production of biomass at low cost (Fal et al. 2021). On the other hand, the ability of microalgae to eliminate and detoxify HMs is the result of adaptation mechanisms developed over centuries of evolution in contaminated environments (Ubando et al. 2021). These mechanisms are subdivided into metabolism-dependent and metabolism-independent pathways. Extracellular biosorption of HMs refers to a physicochemical property of the microalgae cell surface that binds to HMs ions independently to the cellular metabolism. However, HMs biosorption into extracellular polymeric substances (EPS) formed by microalgae under conditions of stress is a metabolism-dependent process (Naveed et al. 2019). It has been reported that the biosorption efficiency varies depending on the genus and the species of microalgae (Kumar et al. 2015a, b). For instance, the growth of C. sorokiniana and S. obliguus in media contaminated with Pb(II), Cd(II), Cu(II) and Cr(VI) was significantly



Fig. 2.2 Intracellular and extracellular mitigation pathways for HMs using microalgae

different (Danouche et al. 2020). This can be attributed to the physiology of the strain, in particular, the cell wall composition (Baudelet et al. 2017). In contrast, bioaccumulation is a metabolism-dependent mechanism. It consists of an intracellular accumulation of HMs into the cytosolic compartment through passive and/or active transport across the cell membranes (Chojnacka 2010). According to Pérez-Rama et al. (2002), the bioaccumulation of Cd(II) using *Tetraselmis suecica* was a biphasic process, assisted in the first phase by an adsorption to proteins or polysaccharides, followed by an energy-dependent accumulation to the cytosol. The intracellular mitigation of toxic HMs may involve the chelation by metallothioneins (Balzano et al. 2020), phytochelatins (Gómez-Jacinto et al. 2015), poly-phosphates (Wang and Dei 2006), the compartmentalisation in the vacuole (Shanab et al. 2012), chloroplast (Hanikenne et al. 2009) and mitochondria (Mendoza-co et al. 2005) or the biotransformation via an enzymatic reaction such the biotransformation of Cr (VI) to Cr(III) by strains of C. vulgaris through an enzymatic reaction catalysed by the chromate reductase (Lee et al. 2017; Yen et al. 2017). Figure 2.2 depicts the intracellular and extracellular mitigation pathways for HMs using microalgae.

**Phycoremediation of Organic Pollutants** Although microalgae are classified as autophotrophic organisms, some species have a heterotrophic metabolism, and under certain conditions, some microalgae strains are able to grow in mixotrophic mode. This trophic particularity allows microalgae cells to use the carbons contained in organic pollutants and to ensure the bioremediation of contaminated aquatic ecosystems (Zhou et al. 2017). It has been reported that many microalgae strains have the capability of removing a range of organic pollutants such as polycyclic aromatic hydrocarbon (Semple et al. 1999), synthetic dyes (Bhardwaj and Bharadvaja 2021), pharmaceuticals and personal care products (Hena et al. 2021), pesticides and other emerging contaminants (Maryjoseph and Ketheesan 2020).

Based on the above considerations, we can infer that the benefits of phycoremediation technology are to allow both  $CO_2$  capture through photosynthesis and to remove nutrients and xenobiotics from wastewater. Thus, the resulting biomass can be used as a raw material for several valuable products depending on their composition and the type of pollutant to which it has been exposed. For

example, it can be used for the production of biofuels, animal feed, fertilisers, pharmaceuticals, biosurfactants, proteins, pigments and many other valuable products, that can be extracted from microalgae.

Microalgae production could be integrated into a biorefinery to achieve greater economic potential.

#### 2.4 Microalgae Biomass: Valorisation within a Biorefinery Concept

Like a traditional petroleum refinery, a biorefinery converts feedstock into energy and several chemicals. The process entails different technologies and can be applied for processing different raw materials. There are several types of biorefineries based on biobased feedstock or waste source, end-products and conversion technologies. Potential organic feedstock sources include corn, potato, cellulosic biomass, forestry, agricultural waste, food waste and algae. The variety of possible organic raw materials implies a rich diversity of potential chemicals. Bioethanol, biogas, lignin, secondary metabolites, carbohydrates, lipids and proteins are some of the products that can be obtained using the biorefinery concept (Espinoza Pérez et al. 2017). To obtain this variety, different conversion processing technologies are required. Biorefinery conversion techniques can involve different separation technologies such as thermochemical conversion, chemical conversion and biochemical conversion (Sankaran et al. 2018).

There are three different phases of development of a biorefinery depending upon feedstock and products. Phase I converts a single raw material into one main product using a fixed process. The Phase II biorefinery also processes a single raw material but is capable of producing various products with diverse processing technologies. The biorefinery processes using single raw material sources can lead to food competition, land use issues and environmental impact (Espinoza Pérez et al. 2017; Sankaran et al. 2018). Phase III biorefinery uses a mixture of biomass from different sources, such as whole-crop, lignocellulose and microalgae, that allows the production of many biobased industrial products using different processing technologies. Phase III, the most advanced form of biorefinery, is also an engineering challenge due to the complexity involved. Some of these constraints are related to product separation and purification.

Among the various biomass sources, microalgae are a very promising and remarkable feedstock for the biorefinery process; they do not compete with food, do not require arable land, and can be used for air and water cleaning processes. Large-scale microalgae production requires a high initial investment, especially for the installation of photobioreactors, and production costs; these include high-power consumption, artificial light illumination, the  $CO_2$  feed, the cultivation medium, and nitrates and phosphates. To save water and decrease production costs, microalgae can be used in wastewater treatment stations; they can also be integrated into different industrial production units such as cement, paper, textile, tannery and dairy. An integrated system requires the full use of waste and exhaust gases.

The biorefinery process consists of the separation of different fractions, such as lipids, minerals, carbohydrates and secondary metabolites. That separation process should not cause damage to the other fractions. Microalgae are rich in lipids, but during the separation process is possible to obtain many other products that can be also transformed into value-added products (pigments, polyunsaturated fatty acids (PUFAs), toxins, and polysaccharides) or may be used to produce bioplastics, for instance. Microalgae biorefinery is divided into several stages which can be categorised into upstream and downstream processes. The upstream processing is determined by the strain selection, carbon dioxide supply, light source and intensity, and nutrients, such as nitrogen and phosphorus sources.

#### 2.4.1 Upstream Production Systems

The production of microalgal biomass, and derived products, is highly dependent on the cultivation production systems. The choice of production system is particularly important for bioremediation applications, as large-scale algae production requires high initial capital investment. Broadly speaking, there are two main types of production system for microalgae; the open pond system or closed systems (Fig. 2.3).



Fig. 2.3 Microalgae cultivation systems

#### 2.4.1.1 Open Systems

Open ponds are systems that allow large-scale production for commercial purposes. Open ponds have various sizes, shapes and types of turbulence. Their construction is often dictated by local conditions and available materials and can be constructed of plastic, bricks, concrete, or adobe. Open ponds include lakes and lagoons, raceway ponds, paddle-wheel-driven open raceway ponds, circular ponds and Inclined and cascaded systems. Open systems generally present some shortcomings in relation to control over process parameters. These systems offer little or no guarantee on the control of the operational variables (temperature and incident light intensity). The contamination by other microorganisms that may occur during the process and the low efficiency of  $CO_2$  utilisation due to lack of agitation of the flow and poor gas exchange in the culture medium compromise the overall algal growth rates.

- *Natural ponds*. It is a naturally selective system, the type of species and strain is closely linked to the soil and climatic conditions of the region, generally being a low-cost, monospecies that can be grown almost all year round. In these systems, the risk of contamination is very high due to its open characteristics.
- *Raceway ponds (also known as high-rate algal ponds).* These are open-air extensive cultivation systems, in the form of a racetrack, shallow, with mixing undertaken by a paddlewheel that distributes nutrients homogeneously to the microalgae. Generally, they are built with low-cost materials, cement, clay and white plastic to facilitate light capture.
- *Circular ponds*. Since these systems present an inefficient configuration compared with raceway ponds, they are rarely used for commercial purposes. However, in some countries, such as Japan, Taiwan and Indonesia, this process is widely used to produce biomass.
- *Inclined and cascaded systems*. In these systems, the turbulence is generated by gravity and the crop moves from the top to the bottom of an inclined suspended surface. This process is particularly interesting because the flow is highly turbulent, and the thin culture layers improve light absorption, and produce a greater concentration of cells. However, this system has a high evaporation rate, increased sedimentation of the cells under low turbulence mixing regimes and a high energy consumption compared to other pond systems.

The major disadvantages of open systems are essentially the high evaporation rates, the difficulties of temperature control and the high risk of contamination. In general, open cultivation systems produce less biomass per unit area compared to closed photobioreactors.

Although research to develop cultivation systems for microalgae has focused more on closed cultivation systems, today's large-scale industries rely more on open systems for economic reasons; they are cheaper to maintain, easier to operate and require less energy. However, only a small number of algae species can be successfully cultivated in outdoor systems due to contamination, which directly compromises productivity. Due to the several problems associated with open culture systems, there is a particular interest in closed photobioreactors which, offer more efficiency and more advantages over open culture systems. These advantages include better results in terms of efficiency in photosynthesis; greater capacity for  $CO_2$  removal; versatility in terms of the culture medium; production is not seasonal; the absence or reduction of contaminations; ease of monitoring operational variables such as temperature, pH and  $CO_2$ ; greater incidence of light on the culture medium. The photobioreactors can be exposed to sunlight or artificial light, the latter offers better control over the process variables.

#### 2.4.1.2 Closed Systems

Closed photobioreactor systems strongly limit any direct exchange of contaminants into the cultivation medium. The algae and cultivation medium flows within the transparent walls of the reactor to reach the cultured cells. Photobioreactors are classified according to their mode of operation and design. Some examples include tubular vertical and horizontal, shaking tank photobioreactors, helical tubular, flat plate photobioreactors, and photobioreactors mixed by air "airlifts".

Although photobioreactors have limitation of contamination as the main advantage, this may not be completely achievable, except in some designs specifically developed for this purpose. They are also more expensive than open systems.

- Vertical Tubular Photobioreactors. Since the first invention of photobioreactors in 1950, several models have been developed. The construction cost of this system is very high; however, its maintenance and monitoring are very economical. The vertical stacked system presents a higher concentration of cells and a higher productivity thanks to the gentle and controlled agitation of the mixture which is obtained through the injection of compressed air. Furthermore, vertical tubular photobioreactors are compatible with the majority of microalgae species. The disadvantage of this system is that it has a lower efficiency of sunlight incidence, an issue that can easily be circumvented by applying artificial lighting.
- *Horizontal tubular photobioreactors*. Horizontal tubular photobioreactors were the best solution to solve the problem of sunlight incidence in vertical tubular systems. The horizontal orientation of the tubes increases the capacity to absorb the incidence of light. However, this system also has some difficulties in the O<sub>2</sub> removal and in the CO<sub>2</sub> supply, also the increase of the light absorption capacity implies the growth of the installation area of the tubes.
- *Flat photobioreactors.* These are systems with large, transparent panels, which can be made of glass or polycarbonate, arranged vertically or at an inclined angle. The light intensity is easily controlled due to the possibility of directing the panels in different directions and at different angles. This system has received special attention from the scientific community, which considers it to be a promising system thanks to the light conversion efficiency and productivity per unit area.

• Airlift photobioreactors. These are cylindrical tubes with two interconnecting zones, one of which is called a "riser", where the gas mixture is spread, while the other zone, the "downcomer", does not receive the gas. This system stands out for having high levels of mass transfer. The pneumatic agitation caused by the injection of  $CO_2$  at the bottom of the tank increases the velocity of the medium circulation, which in turn increases the efficiency of the microalgae growth.

#### 2.4.2 Downstream Processing

After the cultivation stage, the upstream processing ends, and the downstream processing begins. Typically, downstream processing involves several stages: harvesting of biomass through centrifugation, filtration or flocculation, drying, product extraction, purification and conversion processes. These operation units account for 50–60% of the total costs in the microalgae multi-product biorefinery, with harvesting accounting for at least 20% of these costs ('t Lam et al. 2018; Xu et al. 2020). Microalgae cultures are dilute suspensions, typically varying from 0.5 to 5 g L<sup>-1</sup>, depending on species, making harvesting a challenge. Common harvesting and dewatering methods encompass centrifugation, filtration, sedimentation, flotation and flocculation. The latter include chemical, biological and electroflocculation.

After the dewatering process, the microalgae biomass will be subjected to drying processes such as spray drying, solar drying, convective and freeze drying (Chen et al. 2015).

The extraction of high-value products such as lipids, pigments, secondary metabolites, and others can be performed using either wet or dry biomass. The extraction processes vary depending on the products. Some microalgae species can have thick and multilayered walls, silicified, and wall-bound exopolysacharides membranes implying a reduced extraction efficiency. Therefore, appropriated pre-treatment methods are often required before extraction takes place. Cell disruption can be achieved through soaking, maceration, bead-beating, sonification, alkaline lysis, etc. (Catherine Dupré et al. 2020).

After the pre-treatment steps, the biomass can be converted into several products. Extraction methods include conventional solvent extraction, supercritical fluid extraction, enzyme extraction, subcritical water extraction, among others (Catherine Dupré et al. 2020).

#### 2.4.2.1 Production of High-Value Products and Applications

A variety of products and compounds can be extracted and isolated from microalgae biomass: biopolymers, lipids, bioactive compounds, proteins and carbohydrates, which can be applied in a multitude of applications (Table 2.1). The currently existing applications on the market focus on food, feed, nutraceuticals, cosmetics

		Potential industrial products and	Bioremediation/	
Marine species	Compounds	applications	Use	Reference
Chlamydomonas reinhardtii	Spidroin	Biomedical	Cd, Cu, Al and Zn	João Vitor Dutra Molino et al. (2016), Ibuot et al. (2017)
Chaetoceros sp.	Lipids, carotenoids	Nutraceutical, pharmaceuticals, cosmetics, paint, and paper indus- tries, and aquaculture	Bioremediation nitrites, phospho- rus, lead	Parvin Molazadeh et al. (2015), Singh et al. (2021a), Tiwari (2021)
Chlorella marina	Lipids, carotenoids	Biodiesel, cos- metics and phar- maceutical industries, feed (colourants and additives), and the healthcare sector	Bioremediation of nitrate, nitrite, ammonia, phos- phorus, silicate, chromium, lead, zinc, copper, cad- mium, and sul- phide, tannery wastewater	Muthukumar et al. (2012), Adam et al. (2015), Kumar et al. (2015a), Cezare-Gomes et al. (2019), Singaram (2022)
Chlorella sp.	РНВ	Biomedical, package applications	Bioremediation of nitrogen, ammo- nia, heavy metals	Da Silva et al. (2018)
Dunaliella salina	Lipids, caroten- oids, phytosterols	Biodiesel, nutra- ceutical industry, aquaculture	Bioremediation of nitrate, silicate, chromium and sul- phide, tannery wastewater	Francavilla et al. (2010), Adam et al. (2015), Cesário et al. (2018), Singaram (2022)
Isochrysis galbana	Lipids, carbohy- drates, proteins, carotenoids, polyunsaturated fatty acids	Biofuels, food and nutraceutical industry, cosmetics	Biodegradation of phenol, bioremedi- ation of nitrate, silicate, chromium and sulphide	Wang et al. (2019), Koutra et al. (2018), Gomez- Loredo et al. (2016), Ruiz- Dominguez et al. (2020), Adam et al. (2015)
Isochrysis sp.	Lipids, carbohy- drates, proteins, carotenoids, polyunsaturated fatty acids, alkenones	Nutraceutical industry, aqua- culture, fuels, polymers, phase change materials,	Bioremediation of nitrites and phosphorus	O'Neil et al. (2021), Singh et al. (2021a)

Table 2.1 Species of marine microalgae, their products and potential applications

(continued)

Marine species	Compounds	Potential industrial products and applications	Bioremediation/ biodegradation Use	Reference
		sunscreens and cosmetics		
Nannochloropsis sp.	Lipids, carbohy- drates, proteins, carotenoids, polyunsaturated fatty acids	Biodiesel, aquaculture	Bioremediation of nitrate, silicate, chromium and sul- phide, imidacloprid, paracetamol, ibu- profen, olanzapine. Tan- nery wastewater	Adam et al. (2015), Encarnação et al. (2020), Encarnação et al. (2021), Singaram (2022)
Rhodomonas sp.	Lipids, carbohydrates	Feed for aquaculture	Bioremediation of para-xylene	Cesario et al. (2018), Li et al. (2020)
Scenedesmus sp.	РНВ	Biomedical, package applications	Bioremediation of nitrogen, phospho- rus, heavy metals	García et al. (2020)
Spirulina sp.	РНВ	Biomedical, package applications	Bioremediation of nitrogen, phospho- rus, ammonia, heavy metals	Da Silva et al. (2018), Selvaraj et al. (2021)
Thalassiosira sp.	Lipids	Biodiesel	Tannery wastewater	Singaram (2022)
Tetraselmis sp.	Lipids, carotenoids	Aquaculture, nutraceuticals, cosmetics	Bioremediation of nitrate, silicate, chromium and sulphide	Adam et al. (2015), Schuler et al. (2020)

 Table 2.1 (continued)

and pharmaceuticals with several companies commercialising different microalgae products. Few are focused on the chemical industry, and the potentialities are vast and could extend into new areas.

Biopolymers, such as polyhydroxy butyrate (PHB) have been identified in some marine and freshwater microalgae species. They are used in the food industry, pharmaceutical industry, environmental remediation and medical devices. Polyhydroxyalkanoates (PHAs) are biodegradable biopolyesters produced by microorganisms, including microalgae. With properties like those of polyethylene and polypropylene, they can be processed similarly to fossil-based thermoplastics, including injection moulding, extrusion and blow moulding. PHAs are generally produced by heterotrophic bacteria, natural or artificially modified bacteria, such as *Cupriavidus necator*, recombinant *Escherichia coli.*, *Ralstonia* sp., *Halomonas* sp., among several others {Khatami, 2021 #1}(Khatami et al. 2021); it is known that at least 75 distinct genera synthesise PHAs (Reddy et al. 2003). The intracellular levels accumulated can reach 90% of the cell's dry weight under conditions of nutrient stress (Reddy et al. 2003). At present, more than 160 different monomer units have been identified, and their molecular weight ranges from 50,000 to a million Da (Taguchi and Matsumoto 2021; Vermeer et al. 2022). The variation in the composition of the monomeric units implies a diversity of chemical and physical properties. Microalgae are an alternative to the production of bacterial PHA. PHAs such as PHB can be stored by microalgae and cyanobacteria as reserves of energy material in response to nutritional stress. Excess carbon and nutrient depletion growth conditions lead to the production of these polyesters by these microorganisms. Microalgae species *Chlorella* sp., *Scenedesmus* sp., and *Spirulina* sp. are three species capable of producing PHB (Da Silva et al. 2018; García et al. 2020; Selvaraj et al. 2021). Large quantities of carbon source, such as glucose, are required to produce PHAs, which represents 50% of the total costs of production (Costa et al. 2019). This limitation can be overcome using waste feedstock from industrial waste streams.

Spider silk proteins, such as spider fibroins or spidroins, are another interesting biomaterial that can be obtained from the cultivation of microalgae. The recombinant spider silk proteins have similar properties to those of natural spider silk. This biomaterial has extraordinary properties, such as toughness, strength, elasticity, and biocompatibility, exceeding those of other natural and synthetic materials such as steel, and textile fibres, including Kevlar-like super fibres. Potential applications include biomedical (scaffolds and tissue engineering), hydrogel formation, constructing fibres and electronics.

Genes encoding recombinant spidroin have been expressed in *Chlamydomonas reinhardtii*. (João Vitor Dutra Molino et al. 2016).

Microalgae lipids provide a potential and attractive alternative to crude oil and a source of building blocks for many interesting chemicals. Many microalgae species can produce large amounts of lipids depending on culture parameters, such as light intensity and nitrates concentration. Several reports on lipid production indicate a production range of up to 80% of dry weight (Encarnação et al. 2018, Hess et al. 2018). These microalgal oils are a great source of feedstock chemicals. Lubricants (Farfan-Cabrera et al. 2022), resins (Hidalgo et al. 2019), additives, polyols and polyurethanes (Peyrton et al. 2020), coatings (Decostanzi et al. 2018), plasticisers and surfactants (Pleissner et al. 2015) are some of the potential products.

Regarding biofuel applications, the development has been at a lab and pilot scale. The conversion of microalgae lipids to biodiesel is performed by a transesterification process in which triglycerides react with monoalcohol (methanol or ethanol) in the presence of acid or alkali catalyst to produce fatty acid methyl esters (FAME) and glycerol as a side product. The two most imposing barriers and limitations to the scale-up of this technology to produce biofuels are the downstream processing costs and the bioprocessing.

Companies, industries, and governments should invest in the circular economy, stimulating the transition for a biobased and green economy. The development has been supported by several National, European and International projects. The European Commission has launched ambitious programmes boosting these transitions (Table 2.2).

				Total cost
Acronym	Project title	Programme	Date	(Private + Public)
ALGAECEUTICALS	Development of microalgae-based nat- ural UV Sunscreens and Proteins as cosmeceuticals and nutraceuticals	H2020 EU	2018–2023	€ 1,129,500
ALGFUEL	Biodiesel production from microalgae	FP7-EU	2011–2013	153,917
BISIGODOS	High value-added chemicals and BIoreSIns from alGae biorefineries produced from CO2 provided by industrial emissions	FP7-EU	2013–2017	€ 5,605,438,85
DEMA	Direct Ethanol from MicroAlgae	FP7-EU	2012–2017	€ 6,388,935,04
D-FACTORY	The microalgae biorefinery	FP7-KBBE	2013–2017	€ 10,074,870,03
INTERCOME	International commercialisation of innovative products based on Microalgae	H2020 EU	2016–2018	€ 2,426,437,75
FUEL4ME	Sustainable biofuel from algae	FP7-EU	2013–2016	€ 5,369,514,10
MAGNIFICENT	Microalgae as a green source for nutritional ingredients for food/ feed and ingredients for cosmetics by cost- effective new technologies	H2020 EU	2017–2021	€ 5,685,015,41
SOLENALGAE	Algae biomass: Unlocking new uses as food, feed and fuel	ERC	2016–2021	€ 1,441,875
SUNBIOPATH	Microalgae engineering—Greener biomass and biofuel production	FP7-KBBE	2010–2013	€ 4,366,894,60

 Table 2.2
 Selection of European projects with focus on microalgae. Information related to funded projects can be found on cordis.europa.eu

#### 2.5 Legislation Framework and Regulations/Policy and Legal Framework

Biobased products are non-food goods, fundamental in a Circular Bioeconomy, which can help to reduce  $CO_2$  and pollution (EuropeanCommission n.d.). They are derived from renewable raw materials. When these are obtained from microalgae biomass used in cleaning processes, as the present chapter discusses, special attention to the legislation applicable to them should be verified. Despite the importance attributed to those products by the Regulatory Entities, and some progress has been made, there is still a long way ahead for better clarification of the rules. For instance, the Legislation for Biobased products in the EU and USA should guarantee standards and measurements which allow the industries to make them available in terms of sustainability. Depending on the type of biobased products, details in quality control still need to ensure better products and to achieve high acceptance by the Market. It would be ideal that the EU and USA, among others, develop harmonised standards for such products to reduce trade barriers and foster a more broad market. However, until now, limited research has been conducted on biobased products derived from microalgae used in bioremediation, and this could drive some constraints on the companies due to some toxic contaminants not yet well evaluated or unknown.

Before we go further in detail on the legislation available, we must highlight some considerations. Regarding biobased materials from microalgae, it is important to identify the products produced and what will be the targeting industry. Thus, it is possible to organise and orient the application of the available legislation. Until now, the well-established bioproducts obtained from algae (macroalgae) were alginates, agar and carrageenans (phycocolloid production), almost exclusively from seaweeds (macroalgae brown and red) and other biopolymers, such as starch, cellulose, chitin and PHA (biodegradable plastic). It is consensual that in these last years, the growth in the production of microalgae introduced a new perspective in the biotransformation and production of different new biomaterials, which require a more detailed look at the possibilities. Moreover, as referred to above in this chapter, the production of high-value products include biopolymers, carbohydrates, lipids, phycobilins, pigments, proteins, polyhydroxyalkanoates, and many more compounds. Resins, coatings, binders, and bioplastics are the main industrial applications as well biofertilisers, food and feed, pharmaceuticals, cosmetics and products for personal care. Even bioplastics which can be converted to produce high-value final products such as medical equipment, prosthetics and scaffolds, will add a new perspective market for these biobased products. Therefore, the legislation available should reflect clear information to be followed, if different from the biobased obtained from plants, for instance. Other interesting microalgae products include the bioactive oligosaccharides (extracted by enzymatic methods), biopigments for food supplements (carotenoids and xanthophylls) or for paints, and cosmetic bioactive compounds for skincare (ex. Porphyridium cruentum extract) and  $\gamma$ -linoleic acid or alguronic acid from Chlorella extract. For all of the cited products, the legislation

used is the same that should be followed for other similar products from different sources. Fertilisers or soil improvers, feed, energy or fuels, soap, and building or packaging materials are some of the possible examples.

Nevertheless, the main concern for biobased materials production for various applications requires intensive regulatory work in order to protect human health and the environment from harmful fractions of waste, especially if the start material is microalgae previously used for wastewater bioremediation. The legal maximum of residues should be cautious, especially because, in certain cases, some of them are unknown. Despite this, the legislation for Agro-food, Cosmetics and Pharmaceutics have already a list of contaminants that should be avoided, and when admitted, the low amounts of them need to be reevaluated cycling, face to the evolution of data provided by science.

Nevertheless, the companies, when preparing a submission to obtain clearance of the material, should evaluate, for instance, which are the appropriate food simulants to be used to estimate the potential for migration and found to prove to authorities that the substance is stable for an intended application that involves a specific type of food, cosmetic or a pharmaceutical purpose. Generally speaking, it may be necessary to demonstrate the suitable purity of a product with respect to the potential presence of possible contaminants such as, for instance, algal biotoxins and mycotoxins, toxic organic compounds such as dioxins and polychlorinated biphenyls, or inorganic compounds already regulated in various materials as human and veterinary medicine residues, heavy metals, nitrates and pesticides.

For instance, a scientific risk assessment carried out by the Scientific Committee for Consumer Safety (SCCS) in Regulation (EC) No. 1223/2009, in order to address potential risks for human health, lays down a system of restrictions and bans on the use of certain substances in cosmetics based classified as carcinogenic, mutagenic or toxic for the reproduction (CMR), of category 1a, 1b or 2, under Part 3 of Annex VI to Regulation (EC) No. 1272/2008. Substances classified as endocrine disruptors are only banned or restricted automatically in cosmetics if they are also classified as CMR. Actually, some substances classified as endocrine disruptors derived from plants are still found in certain cosmetic products, but this issue should be reviewed for the safety of consumers once scientists continue to link endocrine-disrupting chemicals to various diseases and disorders such as cancer, infertility and obesity.

The legislation here referred to below is representative of the most detailed data that should be consulted before submitting for Market Authorization in EU and in the USA biobased products obtained from microalgae used in the bioremediation of wastewater from different sources (Tables 2.3 and 2.4).

In summary, eco-innovation is a priority on the international agenda, and, despite this, all the regulation available is mainly for biobased products other than microalgae derivatives. The tailored production of microalgae biomass and the respective bio-products still have barriers and constraints, mainly in the understanding of the levels and types of contaminants that sometimes could be under-evaluated in the current legislation. Better and more active involvement of the companies in a green intervention will contribute to a stable policy framework with greater harmonisation and coordination around the world, together with a simplification of

Compared in formulation	1. the set of the second
General information	https://eur-lex.europa.eu/
	http://data.europa.eu/growth/sectors/biotechnology/bio
	hased_products_en
	https://www.biobasedconsultancy.com/en/database
	https://www.orobasedeonsultaney.com/en/edutabase
	enabling-technologies en)
Conversion of waste streams into	https://www.bbi_europe.eu/
value-added products	https://ec.europa.eu/growth/tools-databases/ein-
	rawmaterials/en
Materials and articles intended to	https://ec.europa.eu/food/safety/chemical_safety/food_
come into contact with food	contact_materials/legislation_en
	Plastics Regulation, (EU) No. 10/2011, for all multi-layer
	food contact materials (FCM). This regulation includes
	monomers and other substances and additives (other than
	colourants) and some polymer production aids that are
	permissible. For new monomers or additives produced, it
	is possible to start a petition to the European Food Safety
	Authority (EFSA), which will review and issue a formation on the sefective of the substance and if enproved
	will be included in the Plastics Perulation's positive list
	through an amendment to the regulation
	All ECMs in the EU must comply with the safety criteria
	specified in Framework Regulation (EC) No. 1935/2004:
	"The principle underlying this Regulation is that any
	material or article intended to come into contact directly
	or indirectly with food must be sufficiently inert to pre-
	clude substances from being transferred to food in quan-
	tities large enough to endanger human health or to bring
	about an unacceptable change in the composition of the
	food or a deterioration in its organoleptic properties".
	All FCMs must also comply with the Regulation,
	(EC) No. 2023/2006, on good manufacturing practice for
	materials and articles intended to come into contact
	with food.
	Bioplastics from biobased products from algae are
	required to comply with the same regulations with respect
	to tood and safety as fossil fuel-based plastics, and some
	concerns related to end-of-life issues remain on the actual
	agenda.
Fertilising products	Regulation (EU) 2019/1009 of the European Parliament
	and of the Council of 5 June 2019 laying down rules on
	the making available on the Market of EU fertilising
	products and amending Regulations (EC) No 1069/2009
	and $(EC)$ No 110//2009 and repeating Kegulation
	2018/REV/1
Pharmaceutics	All products used should be pharmaceutical grade and. in
	general, fulfil the guidelines required for specific
	purposes.
	· · · · · · · · · · · · · · · · · · ·

 Table 2.3
 Lead Market for biobased products from microalgae in Europe legislation

General information	Any substance, the intended use of which is rea- sonably expected to become a component of food must be authorised for such use by the US Food and Drug Administration (FDA) through a food additive regulation, and all the substances must be recognised as safe.
Food-contact polymers/plastic materials and articles intended to contact food	Food additives are listed in the Code of Federal Regulations (C.F.R.), Part 177, "Indirect Food Additives: Polymers", and food packaging material intended to come in contact with food must comply with FDA's Good Manufacturing Practices (GMP). This implies that "additives may only be used in an amount necessary to achieve their function or pur- pose and may not contain impurities at levels suffi- ciently high as to result in the adulteration of food". In US regulations, the Plastics Regulation includes limits on co-reactants or use levels for starting materials, temperature restrictions, specification of single versus repeated use, and food types for spe- cific substances. www.food.gov.uk/sites/default/files/media/docu ment/bio-based-materials-for-use-in-food-contact- applications.pdf

Table 2.4 Lead Market for biobased products from microalgae in US legislation

complex national regulations, which provides more transparency for relevant stakeholders in knowing how sustainable are their products.

Also, national and international programmes and research funding will push forward the development and innovation of these technologies. Moreover, the fact that the alternative products are considered green and biobased is not guaranteed that they have not a similar negative impact on the environment and human health as their conventional counterparts. Therefore, more research is needed to mitigate the potential impacts.

As a final note, the biorefinery concept can transform the linear economy through a biobased circular economy which is an integrated concept that envisages the cascade use of biomass from different sources.

Acknowledgements The authors acknowledge the Fundação para a Ciência e a Tecnologia (FCT) through the project PTDC/BTA-GES/2740/2020\_NABIA. The Coimbra Chemistry Centre (CQC) is supported by the FCT through the projects UIDB/00313/2020 and UIDP/00313/2020. CDRSP is financed by national funds through the FCT/MCTES (UIDB/00481/2020 & UIDP/00481/2020). We are grateful for funding from PTScience which is supported through the programmes CENTRO-05-4740-FSE-001526 and FEDER.

#### References

- Abdel-Raouf N, Al-Homaidan AA, Ibraheem IBM (2012) Microalgae and wastewater treatment. Saudi J Biol Sci 19:257–275. https://doi.org/10.1016/j.sjbs.2012.04.005
- Adam S, Santhanam KPP, Kumar SD, Prabhavathi P (2015) Bioremediation of tannery wastewater using immobilized marine microalga Tetraselmis. J Mar Biol Oceanogr 4:1
- An Y, Nan M, Chern S, Ilankoon IMSK (2021) Analysis of direct and indirect quantification methods of CO 2 fixation via microalgae cultivation in photobioreactors: a critical review. Renew Sust Energ Rev 137:110579. https://doi.org/10.1016/j.rser.2020.110579
- Balzano S, Sardo A, Blasio M, Chahine TB, Anno FD, Sansone C, Brunet C (2020) Microalgal Metallothioneins and Phytochelatins and their potential use in bioremediation. Front Microbiol 11:1–16. https://doi.org/10.3389/fmicb.2020.00517
- Baudelet PH, Ricochon G, Linder M, Muniglia L (2017) A new insight into cell walls of Chlorophyta. Algal Res 25:333–371. https://doi.org/10.1016/j.algal.2017.04.008
- Bhardwaj D, Bharadvaja N (2021) Phycoremediation of effluents containing dyes and its prospects for value-added products: a review of opportunities. J Water Process Eng 41:102080. https://doi. org/10.1016/j.jwpe.2021.102080
- Borowitzka MA (2013) In: M. A. B. a. N. R (ed) Energy from microalgae: a short history. Springer Science, Moheimani
- Castellanos-Estupinan MA, Carrillo-Botello AM, Rozo-Granados LS, Becerra-Moreno D, Garcia-Martinez JB, Urbina-Suarez NA, Lopez-Barrera GL, Barajas-Solano AF, Bryan SJ, Zuorro A (2022) Removal of nutrients and pesticides from agricultural runoff using microalgae and cyanobacteria. Water 14:4
- Catherine Dupré HDB, Campos MG, Delattre C, Encarnação T, Fauchon M, Gaignard C, Hellio C, Ito J, Laroche C, Legrand J, Michaud P, Pais AACC, Pierre G, Serive B, Watanabe MM (2020) Microalgal biomass of industrial interest: methods of characterization. Handbook on characterization of biomass, biowaste and related by-products. C. e. a. Dupré, Springer, Cham
- Cesário MT, Da Fonseca MMR, Marques MM, De Almeida MCMD (2018) Marine algal carbohydrates as carbon sources for the production of biochemicals and biomaterials. Biotechnol Adv 36(3):798–817
- Cesario TM, da Fonseca MMR, Marques MM, de Almeida MCMD (2018) Marine algal carbohydrates as carbon sources for the production of biochemicals and biomaterials. Biotechnol Adv 36(3):798–817
- Cezare-Gomes EA, Mejia-da-Silva LDC, Perez-Mora LS, Matsudo MC, Ferreira-Camargo LS, Singh AK, de Carvalho JCM (2019) Potential of microalgae carotenoids for industrial application. Appl Biochem Biotechnol 188(3):602–634
- Chen C-L, Chang J-S, Lee D-J (2015) Dewatering and drying methods for microalgae. Dry Technol 33(4):443–454
- Chojnacka K (2010) Biosorption and bioaccumulation-the prospects for practical applications. Environ Int 36:299–307. https://doi.org/10.1016/j.envint.2009.12.001
- Costa SS, Miranda AL, de Morais MG, Costa JAV, Druzian JI (2019) Microalgae as source of polyhydroxyalkanoates (PHAs) a review. Int J Biol Macromol 131:536–547
- Da Silva CK, Costa JAV, De Morais MG (2018) Polyhydroxybutyrate (PHB) synthesis by Spirulina sp. LEB 18 using biopolymer extraction waste. Appl Biochem Biotechnol 185(3): 822–833
- Danouche M, El Arroussi H, Bahafid W, El Ghachtouli N, El Arroussi H, Bahafid W, An NEG (2021) An overview of the biosorption mechanism for the bioremediation of synthetic dyes using yeast cells. Environ Technol Rev 10:58–76. https://doi.org/10.1080/21622515.2020. 1869839
- Danouche M, El Ghachtouli N, El Baouchi A, El Arroussi H (2020) Heavy metals phycoremediation using tolerant green microalgae: enzymatic and non-enzymatic antioxidant systems for the management of oxidative stress. J Environ Chem Eng 8:104460. https://doi.org/ 10.1016/j.jece.2020.104460

- Decostanzi M, Lomege J, Ecochard Y, Mora AS, Negrell C, Caillol S (2018) Fatty acid-based cross-linkable polymethacrylate coatings. Prog Org Coat 124:147–157
- Encarnação T, Arranja CT, Cova TFGG, Pais AACC, Campos MG, Sobral AJFN, Burrows HD (2018) Monitoring oil production for biobased feedstock in the microalga Nannochloropsis sp.: a novel method combining the BODIPY BD-C12 fluorescent probe and simple image processing. J Appl Phycol 30:2273–2285
- Encarnacao T, Palito C, Pais A, Valente AJM, Burrows HD (2020) Removal of pharmaceuticals from water by free and imobilised microalgae. Molecules 25:16
- Encarnacao T, Santos D, Ferreira S, Valente AJM, Pereira JC, Campos MG, Burrows HD, Pais A (2021) Removal of imidacloprid from water by microalgae Nannochloropsis sp. and its determination by a validated RP-HPLC method. Bull Environ Contam Toxicol 107(1):131–139
- Eroglu E, Agarwal V, Bradshaw M, Chen XJ, Smith SM, Raston CL, Iyer KS (2012) Nitrate removal from liquid effluents using microalgae immobilized on chitosan nanofiber mats. Green Chem 14(10):2682–2685
- Espinoza Pérez AT, Camargo M, Narváez Rincón PC, Alfaro Marchant M (2017) Key challenges and requirements for sustainable and industrialized biorefinery supply chain design and management: a bibliographic analysis. Renew Sust Energ Rev 69:350–359
- European Commission (n.d.) Communication from the Commission to the European Parliament, the Council, the European Economic and Social Committee and the Committee of the Regions. A new Circular Economy Action Plan For a cleaner and more competitive Europe COM/2020/ 98 final. https://ec.europa.eu/growth/industry/sustainability/circular-economy\_en
- Eze VC, Velasquez-orta SB, Hernández-garcía A, Monje-ramírez I, Orta-ledesma MT (2018) Kinetic modelling of microalgae cultivation for wastewater treatment and carbon dioxide sequestration. Algal Res 32:131–141. https://doi.org/10.1016/j.algal.2018.03.015
- Fal S, Benhima R, El Mernissi N, Kasmi Y, Smouni A, El Arroussi H (2021) Microalgae as promising source for integrated wastewater treatment and biodiesel production. Int J Phytoremediation 1–13. https://doi.org/10.1080/15226514.2021.1920572
- Farfan-Cabrera LI, Franco-Morgado M, Gonzalez-Sanchez A, Perez-Gonzalez J, Marin-Santibanez BM (2022) Microalgae biomass as a new potential source of sustainable green lubricants. Molecules 27:4
- Ferrando L, Matamoros V (2020) Attenuation of nitrates, antibiotics and pesticides from groundwater using immobilised microalgae-based systems. Sci Total Environ 703:134740
- Francavilla M, Trotta P, Luque R (2010) Phytosterols from Dunaliella tertiolecta and Dunaliella salina: a potentially novel industrial application. Bioresour Technol 101(11):4144–4150
- Fu J, Huang Y, Liao Q, Xia A, Fu Q, Zhu X (2019) Bioresource technology photo-bioreactor design for microalgae: a review from the aspect of CO<sub>2</sub> transfer and conversion. Bioresour Technol 292:121947. https://doi.org/10.1016/j.biortech.2019.121947
- García G, Sosa-Hernández JE, Rodas-Zuluaga LI, Castillo-Zacarías C, Iqbal H, Parra-Saldívar R (2020) Accumulation of PHA in the microalgae Scenedesmus sp. under nutrient-deficient conditions. Polymers 13(1):131
- Gómez-Jacinto V, García-Barrera T, Gómez-Ariza JL, Garbayo-Nores I, Vílchez-Lobato C (2015) Elucidation of the defence mechanism in microalgae *Chlorella sorokiniana* under mercury exposure. Identification of Hg-phytochelatins. Chem Biol Interact 238:82–90. https://doi.org/ 10.1016/j.cbi.2015.06.013
- Gomez-Loredo A, Benavides J, Rito-Palomares M (2016) Growth kinetics and fucoxanthin production of Phaeodactylum tricornutum and Isochrysis galbana cultures at different light and agitation conditions. J Appl Phycol 28(2):849–860
- Hammouda O, Gaber A, Abdel-Raouf N (1995) Microalgae and wastewater treatment. Ecotoxicol Environ Saf 31:205–210
- Hanikenne M, Merchant SS, Hamel P (2009) Transition metal nutrition: a balance between deficiency and toxicity, the chlamydomonas sourcebook. 333–399. https://doi.org/10.1016/ B978-0-12-370873-1.00018-6

- Hena S, Gutierrez L, Crou J (2021) Removal of pharmaceutical and personal care products (PPCPs) from wastewater using microalgae : a review. J Hazard Mater 403. https://doi.org/10.1016/j. jhazmat.2020.124041
- Hess SK, Lepetit B, Kroth PG, Mecking S (2018) Production of chemicals from microalgae lipids status and perspectives. Eur J Lipid Sci Technol 120(1):1700152
- Hidalgo P, Navia R, Hunter R, Gonzalez ME, Echeverria A (2019) Development of novel bio-based epoxides from microalgae Nannochloropsis gaditana lipids. Compos Part B-Eng 166:653–662
- Hoang AT, Sirohi R, Pandey A, Nizetic S, Lam SS, Chen WH, Luque R, Thomas S, Arici M, Pham VV (2022) Biofuel production from microalgae: challenges and chances. Phytochem Rev
- Ibuot A, Dean AP, Mcintosh OA, Pittman JK (2017) Metal bioremediation by CrMTP4 overexpressing Chlamydomonas reinhardtii in comparison to natural wastewater-tolerant microalgae strains. Algal Res 24:89–96
- João Vitor Dutra Molino TLA, Ferreira-Camargo LS, Croce MA, Tanaka A, Buson FX, de Freitas Ribeiro P, Campos-Salazar AB, Antonio EP, Maizel AS, Siratuti VM, Costa C, Włodarczyk SR, de Souza Lima R, Mello FN, Mayfield SP, Carvalho JCM (2016) Chimeric spider silk production in microalgae: a modular bionanomateria. Res Ideas Outcomes 2:e9342
- Khatami K, Perez-Zabaleta M, Cetecioglu Z (2021) Pure cultures for synthetic culture development: next level municipal waste treatment for polyhydroxyalkanoates production. J Environ Manag 305:114337
- Kong W, Shen B, Lyu H, Kong J, Ma J (2021) Review on carbon dioxide fixation coupled with nutrients removal from wastewater by microalgae. J Clean Prod 292:125975. https://doi.org/10. 1016/j.jclepro.2021.125975
- Koutra E, Economou CN, Tsafrakidou P, Kornaros M (2018) Bio-based products from microalgae cultivated in digestates. Trends Biotechnol 36(8):819–833
- Kumar D, Santhanam SP, Jayalakshmi T, Nandakumar R, Ananth S, Devi AS, Prasath BB (2015a) Excessive nutrients and heavy metals removal from diverse wastewaters using marine microalga Chlorella marina (Butcher). Indian J Geo-Mar Sci 44(1):97–103
- Kumar KS, Dahms HU, Won EJ, Lee JS, Shin KH (2015b) Microalgae—a promising tool for heavy metal remediation. Ecotoxicol Environ Saf 113:329–352. https://doi.org/10.1016/j.ecoenv. 2014.12.019
- Lee L, Hsu CY, Yen HW (2017) The effects of hydraulic retention time (HRT) on chromium (VI) reduction using autotrophic cultivation of *Chlorella vulgaris*. Bioprocess Biosyst Eng 40: 1725–1731. https://doi.org/10.1007/s00449-017-1827-6
- Li H, Li H, Meng F, Zhang B, Lin Y, Wu J, Wang G, Zheng Y (2020) The biodegradation of Paraxylene in seawater by a newly isolated oceanic microalga Rhodomonas sp. JZB-2. J Water Process Eng 36:101311
- Maryjoseph S, Ketheesan B (2020) Microalgae based wastewater treatment for the removal of emerging contaminants: a review of challenges and opportunities. Case Stud Chem Environ Eng 2:100046. https://doi.org/10.1016/j.cscee.2020.100046
- Mendoza-co D, Loza-tavera H, Herna A, Moreno-sa R (2005) Sulfur assimilation and glutathione metabolism under cadmium stress in yeast, protists and plants. FEMS Microbiol Rev 29:653– 671. https://doi.org/10.1016/j.femsre.2004.09.004
- Muthukumar ASE, Ajithkumar TT, Kumaresan S, Balasubramanian T (2012) Biodiesel production from marine microalgae chlorella marinaand Nannochloropsissalina. J Pet Technol Altern Fuels 3:58–62
- Naveed S, Li C, Lu X, Chen S, Yin B, Zhang C, Ge Y (2019) Microalgal extracellular polymeric substances and their interactions with metal(loid)s: a review. Crit Rev Environ Sci Technol 49: 1769–1802. https://doi.org/10.1080/10643389.2019.1583052
- O'Neil GW, Gale AC, Nelson RK, Dhaliwal HK, Reddy CM (2021) Unusual shorter-chain C35 and C36 alkenones from commercially grown Isochrysis sp. Microalgae. J Am Oil Chem Soc 98: 757–768
- Parvin Molazadeh NK, Rahimi MR, Nasiri A (2015) Adsorption of lead by microalgae Chaetoceros Sp. and Chlorella Sp. from aqueous solution. J Commun Health Res 4(2):114–127
- Pérez-Rama M, Abalde Alonso J, Herrero López C, Torres Vaamonde E (2002) Cadmium removal by living cells of the marine microalga *Tetraselmis suecica*. Bioresour Technol 84:265–270. https://doi.org/10.1016/S0960-8524(02)00045-7
- Peyrton J, Chambaretaud C, Sarbu A, Averous L (2020) Biobased polyurethane foams based on new polyol architectures from microalgae oil. ACS Sustain Chem Eng 8(32):12187–12196
- PierreChevalier JLN (1985) Wastewater nutrient removal with microalgae immobilized in carrageenan. Enzym Microb Technol 7:621–624
- Pleissner D, Lau KY, Zhang CW, Lin CSK (2015) Plasticizer and surfactant formation from foodwaste and algal biomass-derived lipids. ChemSusChem 8(10):1686–1691
- Reddy CS, Ghai R, Rashmi, Kalia VC (2003) Polyhydroxyalkanoates: an overview. Bioresour Technol 87(2):137–146
- Ruiz-Dominguez MC, Cerezal P, Salinas F, Medina E, Renato-Castro G (2020) Application of box-Behnken design and desirability function for green prospection of bioactive compounds from Isochrysis galbana. Appl Sci-Basel 10:8
- Sankaran R, Show PL, Nagarajan D, Chang JS (2018) Exploitation and biorefinery of microalgae. Waste Bioref Potential Perspect:571–601
- Schuler LM, Santos T, Pereira H, Duarte P, Gangadhar KN, Florindo C, Schulze PSC, Barreira L, Varela JCS (2020) Improved production of lutein and beta-carotene by thermal and light intensity upshifts in the marine microalga Tetraselmis sp. CTP4. Algal Res-Biomass Biofuels and Bioproducts 45
- Selvaraj K, Vishvanathan N, Dhandapani R (2021) Screening, optimization and characterization of poly hydroxy butyrate from fresh water microalgal isolates. Int J Biobased Plast 3(1):139–162
- Semple KT, Cain RB, Schmidt S (1999) Biodegradation of aromatic compounds by microalgae. FEMS Microbiol Lett 170:291–300. https://doi.org/10.1111/j.1574-6968.1999.tb13386.x
- Shanab S, Essa A, Shalaby E (2012) Bioremoval capacity of three heavy metals by some microalgae species (Egyptian isolates). Plant Signal Behav 7. https://doi.org/10.4161/psb.19173
- Shi J, Podola B, Melkonian M (2007) Removal of nitrogen and phosphorus from wastewater using microalgae immobilized on twin layers: an experimental study. J Appl Phycol 19(5):417–423
- Singaram VNJ (2022) Enhanced biodiesel production by optimizing growth conditions of Chlorella marina in tannery wastewater. Fuel 316:123431
- Singh DV, Bhat RA, Upadhyay AK, Singh R, Singh DP (2021b) Microalgae in aquatic environs: a sustainable approach for remediation of heavy metals and emerging contaminants. Environ Technol Innov 21:101340. https://doi.org/10.1016/j.eti.2020.101340
- Singh PK, Bhattacharjya R, Saxena A, Mishra B, Tiwari A (2021a) Utilization of wastewater as nutrient media and biomass valorization in marine Chrysophytes- Chaetoceros and Isochrysis. Energy Convers Manage-X 10:100062
- Song C, Liu Q, Qi Y, Chen G, Song Y, Kansha Y (2019) Absorption-microalgae hybrid CO<sub>2</sub> capture and biotransformation strategy-a review. Int J Greenhouse Gas Control 88:109–117. https://doi.org/10.1016/j.ijggc.2019.06.002
- Sundarrajan P, Panchamoorthy K, Greetham D, Joseph A (2019) A review on cleaner production of biofuel feedstock from integrated CO<sub>2</sub> sequestration and wastewater treatment system. J Clean Prod 210:445–458. https://doi.org/10.1016/j.jclepro.2018.11.010
- 't Lam, G. P., M. H. Vermue, M. H. M. Eppink, R. H. Wijffels and C. van den Berg (2018). "Multiproduct microalgae biorefineries: from concept towards reality." Trends Biotechnol 36(2): 216–227
- Taguchi S, Matsumoto K (2021) Evolution of polyhydroxyalkanoate synthesizing systems toward a sustainable plastic industry. Polym J 53(1):67–79
- Tiwari BMA (2021) Cultivation of Anabena variabilis, Synechococcus elongatus, Spirulina platensis for the production of C-phycocyanin, C-phycocrythrin and Thalassiosira, Skeletonema, Chaetoceros for fucoxanthin. Syst Microbiol Biomanufacturing 1:356–361
- Ubando AT, Africa ADM, Maniquiz-Redillas MC, Culaba AB, Chen WH, Chang JS (2021) Microalgal biosorption of heavy metals: a comprehensive bibliometric review. J Hazard Mater 402:123431. https://doi.org/10.1016/j.jhazmat.2020.123431

- Vermeer CM, Bons LJ, Kleerebezem R (2022) Production of a newly discovered PHA family member with an isobutyrate-fed enrichment culture. Appl Microbiol Biotechnol
- Wang W, Dei RCH (2006) Metal stoichiometry in predicting Cd and Cu toxicity to a freshwater green alga *Chlamydomonas reinhardtii*. Environ Pollut 142:303–312. https://doi.org/10.1016/j. envpol.2005.10.005
- Wang YJ, Meng FP, Li H, Zhao SL, Liu QQ, Lin YF, Wang GS, Wu JY (2019) Biodegradation of phenol by Isochrysis galbana screened from eight species of marine microalgae: growth kinetic models, enzyme analysis and biodegradation pathway. J Appl Phycol 31(1):445–455
- Xu Z, Wang H, Cheng P, Chang T, Chen P, Zhou C, Ruan R (2020) Development of integrated culture systems and harvesting methods for improved algal biomass productivity and wastewater resource recovery—a review. Sci Total Environ 746:141039
- Yen H, Chen P, Hsu C, Lee L (2017) The use of autotrophic *Chlorella vulgaris* in chromium (VI) reduction under different reduction conditions. J Taiwan Inst Chem Eng 74:1–6. https://doi. org/10.1016/j.jtice.2016.08.017
- Zeng X, Danquah MK, Dong X, Lu Y (2011) Microalgae bioengineering: from CO<sub>2</sub> fixation to biofuel production. Renew Sust Energ Rev 15:3252–3260. https://doi.org/10.1016/j.rser.2011. 04.014
- Zhou W, Wang J, Chen P, Ji C, Kang Q, Lu B, Li K, Liu J, Ruan R (2017) Bio-mitigation of carbon dioxide using microalgal systems: advances and perspectives. Renew Sust Energ Rev 76:1163– 1175. https://doi.org/10.1016/j.rser.2017.03.065

# **Chapter 3 Removal of Heavy Metals and Organic Pollutants by Marine Microalgae**



Jagannathan Umamaheswari, Ricky Rajamanickam, Sowmya Vilvanathan, Subramanian Shanthakumar, Kadiyala Venkateswarlu, Sudharsanam Abinandan, and Mallavarapu Megharaj

Abstract Marine environment is a predominant player in the overall ecosystem functioning with almost half of oxygen evolution into the atmosphere through the photosynthetic activity of plankton communities. Anthropogenic activities cause pollution at an enhanced pace and pose a major threat to the biological cyclings in the marine ecosystem. Pollutants such as heavy metals and organic compounds in the marine environment are a serious concern as they are associated with complex challenges. Marine microalgae are promising candidates in remediating inorganic and organic pollutants due to their versatile metabolic mechanisms. The present chapter provides a comprehensive understanding of the response of marine microalgae in the removal of heavy metals and organic pollutants. Initially, we present the importance of microalgae and the sources of heavy metals and organic pollutants that reach the marine environment besides highlighting the merits and demerits of the conventional and biological treatment systems used for the removal of these pollutants. Finally, we provide a general perspective on the implication of

M. Megharaj (🖂)

J. Umamaheswari · R. Rajamanickam · S. Vilvanathan · S. Shanthakumar

Department of Environmental and Water Resources Engineering, School of Civil and Chemical Engineering, VIT University, Vellore, India

K. Venkateswarlu Formerly Department of Microbiology, Sri Krishnadevaraya University, Anantapuramu, India

S. Abinandan

Global Centre for Environmental Remediation (GCER), College of Engineering, Science and Environment, University of Newcastle, Callaghan, NSW, Australia

Global Centre for Environmental Remediation (GCER), College of Engineering, Science and Environment, University of Newcastle, Callaghan, NSW, Australia

CRC CARE, Newcastle University LPO, Callaghan, NSW, Australia e-mail: megh.mallavarapu@newcastle.edu.au

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_3

marine microalgae and the associated mechanisms in the removal of heavy metals and organic pollutants.

Keywords Marine microalgae · Heavy metals · Organic pollutants · Bioremediation

## 3.1 Introduction

The environment is the global ecological life-supporting system that has been affected in complex and accelerating ways because of pervasive and profound human activities. The past few decades witnessed rapid industrial development, population growth, economic wealth, and urbanization, which ultimately disturb the very processes and components of the nature. Marine ecosystems are at serious risk due to the elevated levels of pollutants discharged from industrial and domestic activities (Bergmann et al. 2015; Nelms et al. 2017). The impact of these pollutants on coastal zone has been significantly greater in estuaries due to their residence time than in inland rivers (Saldarriaga-Hernandez et al. 2020). In fact, the occurrence of both heavy metals and organic pollutants in the marine environment is of significant ecological concern. International scientific experts on marine protection define marine pollution as "chemicals introduced by human activities either directly or indirectly into the marine environment affecting the biota and impairment of water quality" (Kuppusamy et al. 2020). Fish inhabiting polluted waters was reported to accumulate metals in the tissues and the accumulation depends on various biotic and abiotic factors (Zeitoun and Mehana 2014). The heavy metals tend to be widely distributed in liver, kidney, and other tissues and potentially get transferred to humans as they are at the top of the food web (Gabriel et al. 2006). Oils are the major sources of organic contaminants released into the marine environment either during processing or accidentally from drilling, production, and storage (Kuppusamy et al. 2020). Consequently, the deteriorated health of the oceans around the world impacted the social and economic status and prompted to bring international options for safer and healthier marine systems (Gelcich et al. 2014).

Several environmental agencies recognized the severity of these pollutants and proposed various policies in reducing the risk toward marine biota. For instance, a list of priority pollutants that should be universally avoided has been prepared as they can cause shorter or longer effects in any ecosystem (Grip 2017; Beiras 2018). The United States Environmental Protection Agency recently updated the priority pollutant list in the Clean Water Act, which includes several heavy metals, organic contaminants such as dyes, phenols, organophosphates, etc. (USEPA 2014). The United Nations Convention on Law of Sea proposed major duties for member states to investigate potential threats in the marine environment (Stelzenmüller et al. 2018). European Commission endorsed the marine strategy framework directive with an aim to manage Europe-bound seas to gain a healthy state following an ecosystem-based approach (Borja et al. 2013; Danovaro et al. 2016). Marine environments contain several biotas together with marine microalgae that serve as the primary producers and can also be used as sensitive bioindicators (Torres et al. 2008). This

chapter highlights the implication of marine microalgae in the removal of heavy metals and organic pollutants.

# 3.2 Marine Microalgae—An Overview

Marine environments are inhabited by assemblages of several organisms (Tragin and Vaulot 2018). The diversity of marine plankters based on their size is presented in Fig. 3.1. They are easily distinguished based on the nutrition mode: autotrophic organisms, referred to as phytoplankton (microalgae), and grazing organisms, called zooplankton. In addition, marine microalgae are the major primary producers in the marine environment that use solar energy for  $CO_2$  uptake, thus contributing to ocean carbon sink (Huang et al. 2017). These microalgae are generally divided into two



Fig. 3.1 Diversity and classification of marine plankters based on their sizes

lineages such as green and red, with the former being originated by primary and the latter from the secondary or tertiary endosymbiotic process (Nakayama et al. 1998). Chlorophyta is the major algal group in marine waters representing the green lineage, whereas the protists and dinoflagellates fall within the red lineage. Chlorophyta encompasses prasinophytes and chlorophytes, where the abundance is dominated later with Ulvophyceae, Trebouxiophyceae, and Chlorophyceae, all known as the UTC Clade (Leliaert et al. 2012; Fučíková et al. 2014). Chlorophyta consists of chloroplasts surrounded by two membranes with chlorophyll b as the major pigment. Parsinophytes comprise eight lineages of different taxonomic levels, and the numbers increase based on the environmental sequences and novel cultures (Tragin and Vaulot 2018). Chlorophyceae alone comprises two thousand species and are well known for several biotechnological applications (Barra et al. 2014). For example, microalgal biomass is reported to yield several primary metabolites such as carotenoids, proteins, lipids, and polyunsaturated fatty acids (Becker 2004: Guedes et al. 2011; Sharma et al. 2012; Christaki et al. 2013). Due to their biomass productivity and surface ratio, microalgae also play a crucial role in biogeochemical cycling of pollutants in marine waters (Van Gestel and Van Brummelen 1996). For example, the cell wall composition of microalgae is reported to have greater capacity for the metal-binding that can be transferred to food chain through grazing (Wang et al. 1998).

Due to the abundance of microalgae in waters, they have been overwhelmingly considered as sensitive bioindicators to monitor pollutants in the marine environment (Levine 1984; Whitton and Kelly 1995; Ali et al. 1999; Volterra and Conti 2000). While thoroughly reviewing the toxic profile of marine algae, Torres et al. (2008) proposed that the widespread abundance of microalgae in the marine environment can be used for seasonal evaluation or the effect of time change in the ecosystem in response to heavy metals and organic pollutants. Owing to the presence of these pollutants, microalgae tend to respond through physiological changes. Reports indicate that among other marine plankton, diatoms are severely affected by pollutants than green microalgae (Harrison et al. 1986; González et al. 2009). In addition, the green marine microalgae are reported to often dominate the bloom of natural population in the marine environment, particularly at increased pollution levels (Bonin et al. 1986; Folgar et al. 2009). Microalgae are known to respond to pollutants through two mechanisms: accumulation and sorption, and they also synthesize phytochelatins that are responsible for metal detoxification (Gekeler et al. 1988; Folgar et al. 2009). Furthermore, the antioxidants and innate enzymes have been shown to detoxify organic pollutants (Sunda et al. 2002; Stahl and Sies 2003; Sharma et al. 2012). Despite their versatile biochemical mechanisms, a detailed understanding of metal and organic pollutant removal by marine microalgae is very limited. The following sections present a comprehensive overview on the role of marine microalgae in the removal of heavy metals and organic pollutants.

# **3.3** Pollution in the Marine Environment—Sources of Heavy Metals and Organics

Anthropogenic sources of heavy metals and organic pollutants in the marine ecosystem generally result from the direct discharge of wastes, water runoff, and airborne pollutants (Leprovost 2001). These pollutants are carried from inland through sewage, dredged spoil, rainwater, and domestic and industrial waste discharged into coastal waterbodies through estuaries that enter the oceans (Wu et al. 2001; Adeniji et al. 2017). Hydrocarbon pollution is one of the great threats to the marine environment, with estimates of discharge accounting for 1-8million tons per year (National Research Council Committee on Oil in the Sea, 2003). In addition, around 25,000 ship cargo with 18.5 million barrels of oil per day navigates through gulf waters which can potentially result in minor accidental spills causing a threat to the marine environment (Chitrakar et al. 2019). These crude oil spills affect marine organisms by limiting gas exchange and reducing light penetration (González et al. 2009). Moreover, crude oil spills release several organic pollutants such as benzene, toluene, xylene, and aromatic hydrocarbons that can accumulate in marine biota and sediments, thus acting as a sink affecting the ecosystems (Kachel 2008).

The marine environment is also reported to receive copious and stable inputs of pyrogenic hydrocarbons from coal and oil combustion as well as other organic products such as wood (Ravindra et al. 2008; Page et al. 1999). The predominant source of heavy metals in the marine environment is the industrial effluents discharged into the ocean either through runoff or improper disposal. Three types of heavy metals that cause major environmental problems include toxic metals such as cadmium (Cd), lead (Pb), copper (Cu), zinc (Zn), nickel (Ni), cobalt (Co), etc., precious metals like silver (Ag), gold (Au), palladium (Pd), platinum (Pt), etc., and radionuclides such as uranium (U), thorium (Th), radium (Ra), etc. (Wang and Chen 2009). Human exposure to heavy metals has dramatically risen because of an exponential increase in their use in several industrial, domestic, agricultural and technological applications. Other potential anthropogenic sources of heavy metal pollution are industrial effluents, acid mine drainage associated with mining operations, and coal-based and nuclear power plants. Various industries produce and discharge different heavy metals at varying concentrations into the environment; few of them include electroplating, metallurgy, surface finishing industries, energy and fuel production, iron and steel manufacturing, lead-acid battery manufacturing, fertilizer and pesticide industry, electrolysis, electro-osmosis, microelectronics, leather manufacturing, electrical appliance manufacturing, photography, etc. (Ahmaruzzaman 2011). Natural phenomena such as weathering of rocks and volcanic eruptions also significantly contribute to heavy metal pollution.

# 3.4 Removal of Heavy Metals and Organic Pollutants by Marine Microalgae

A comparison of conventional remediation techniques such as chemical precipitation, ion exchange, membrane filtration, electrochemical treatment, coagulation, and flocculation with those of bioremediation approaches, in terms of their merits and demerits (Table 3.1), clearly indicates that remediation of the polluted sites following the conventional engineering approaches is challenging both technically and economically. Also, bioremediation that involves the capabilities of microorganisms in the removal of pollutants is the most promising, relatively efficient, and costeffective technology. The following sections particularly deal with the innate capabilities of microalgae in the removal of heavy metals and organic pollutants from marine environments.

### 3.4.1 Removal of Heavy Metals by Marine Microalgae

Abundant occurrence of metals in the environment leads to their increased concentration in the organisms over time. Bioavailability most often refers to the availability of contaminants, such as heavy metals or organic pollutants, in an ecosystem. Frequently, it is also used to determine the potential risk of pollutants toward nontarget organisms in any system. Bioavailability in the environment primarily involves physical, chemical, and biological processes. Contaminants or pollutants may be present in varying forms: (i) associated with soil and or sediment particles (bound form), (ii) released from liquid and or gaseous phases (release form), and (iii) associated with living organisms (attached form). A contaminant enters a liquid or gaseous phase once it is released from the bound phase. During this stage contaminant transport will take place through advection, diffusion, and dispersion, which result in the movement of contaminant molecules in the medium (liquid or gas) and thereby reassociation of contaminant or return to the bound state (soil). Meanwhile, the contaminants are carried to the surface of the living organisms (Fig. 3.1). Similar processes occur in the medium and eventually the contaminant reaches the living organisms and enters their tissues through cell membrane. Thus, contaminant transport is an important component of its bioavailability. The contaminants after their entry into the cells are metabolized and/or excreted, causing adverse or toxic effects to living organisms (Fig. 3.2).

#### **3.4.1.1** Biosorption of Metals

Biosorption is the process of removing sorbet (metal ions) from the solvent (water) using biological material called a biosorbent. Marine microalgae have recently gained attention for the development of biosorbent materials due to their high

		UNIT LECTIFICATES TOT LIFE LETITOVAL OF		lianus
Treatment technology	Treatment process	Heavy metal treatment	Organic contaminant treatment	References
Physico-chemica	l treatment methods			
Physical separation	The process involves: Mechanical screening, Gravity	<i>Merits:</i> Large-size heavy metals can be	<i>Merits</i> : Organic contaminants that are	Marino et al. (1997), Dermont et al. (2008), Gupta et al. (2012)
	separator, Magnetic separation, Electrostatic separation, and	easily segregated. Easy maintenance and operation.	clogged onto solid wastes such as wood, plastic, etc. can be	
	Attrition scrubbing		removed easily. Gravity separation reduces 50–60% of the suspended solids.	
		Demerits:	Demerits:	
		Removal efficiency depends	Regular maintenance is	
		upon the particle size distribu-	required.	
		tion, soil-clay matrix, moisture	Screens are clogged frequently.	
		content, metal magnetic proper-		
		ties, and pollutant hydrophobic properties.		
Conventional	Chemical precipitation	Merits:	Merits:	Liao and Randtke (1986),
chemical		Heavy metals are precipitated	Effective in removing polymeric	Matlock et al. (2002),
treatment		and removed as sludge.	electrolytes such as carboxyl,	Wingenfelder et al. (2005),
		Low cost of precipitant.	phenol, phosphoryl, and sulfonyl	Özverdi and Erdem (2006),
		Simplicity.	groups.	Chen et al. (2018b)
		pH can be controlled.		
		Demerits:	Demerits:	
		Removal efficiency depends	Usage of large proportions of	
		upon the pH, temperature, and	chemicals for precipitations	
		charge of heavy metal ions.	produces a large amount of	
		Metal hydroxides solubility	sludge.	
		depends upon the pH conditions	Handling of sludge and usage of	
		(8.0–11.0).	cnemical is cosuly.	

(continued)

Table 3.1 (contin	ued)			
Treatment technology	Treatment process	Heavy metal treatment	Organic contaminant treatment	References
		Toxic sludge. Metal precipitation is slow with poor settling. Sulfide precipitation results in the production of toxic H <sub>2</sub> S gas.	pH-dependent. Weakly adsorbing organic pol- lutants can retard precipitation.	
	Coagulation and flocculation	<i>Merits:</i> Sludge settling is faster. Easy dewatering. Sludge stability. Heavy metal concentrations less than 100 mg $L^{-1}$ can be treated. <i>Demerits:</i> The use of coagulants increases settling time. High chemical consumption. A large volume of sludge pro- duction with toxic properties. pH-dependent (11.0–11.5).	Merits: Higher molecular weight organic contaminants are eliminated easily. Demerits: Removes only selected group of organic contaminants. Produces large quantity of sludge. Molecular weights less than 500 are not effectively removed.	Randtke (1988), Ayoub et al. (2001), Shen et al. (2015), Sakhi et al. (2019)
	Flotation	Merits: Mecovery of heavy metals is selective. Low sludge generation. High separation efficiency. Better removal of small particles. Fewer energy requirements.	Merits: Merits: Simple in operation. Ability to treat a wide range of organic contaminants. Recovery of organic contami- nants is possible. Surfactant cost is low.	Hu et al. (2002), Matis et al. (2004), Salmani et al. (2013)

	Space requirement is low.	Effective in treating volatile	
	Less operating cost.	organic contaminants.	
	Demerits:	Demerits:	
	Removal efficiency is dependent	Surfactant needs to be selected	
	on interfacial chemistry and	based on the organic contami-	
	aggregation effectiveness.	nants present in the wastewater.	
 Adsorption	Merits:	Merits:	Anirudhan and Sreekumari
	Low-cost adsorbents can be used	A very effective option for ter-	(2011), Burakov et al. (2018),
	(agricultural waste such as coco-	tiary treatment.	Alalwan et al. (2020)
	nut coir pith, orange peel pith,	Various organic pollutants can	
	plant-derived biomass).	be handled.	
	Sludge production is very less.	Synthetic organic contaminants	
	Metal recovery is possible.	are effectively treated.	
	Higher removal efficiency.		
	Demerits:	Demerits:	
	Difficulty in adsorbent regenera-	Regeneration of adsorbent is	
	tion	difficult.	
	Activated charcoal is efficient in	Disposal of adsorbent is costly.	
	treating the pollutant but costly.		
	Efficiency depends upon the		
	adsorbents.		
Ion-exchange	Merits:	Merits:	Bolto et al. (2002), Dabrowski
	Ion-exchange resins are soluble	Operation is simple.	et al. (2004), Bashir et al. (2019)
	in water.	Installation is easy.	
	Valuable heavy metals can be	Usage of anion exchange resins	
	recovered.	effectively removes natural	
	Cost-effective.	organic matter.	
	Ease of operation.		
			(continued)

Table 3.1 (contin	ued)			
Treatment technology	Treatment process	Heavy metal treatment	Organic contaminant treatment	References
		Very effective in treating low-concentration heavy metal ions.		
		<i>Demerits:</i> Highly pH sensitive. Matrix gets easily fouled.	Demertits: Treatment efficiency depends upon the resins and type of organic contaminants in the	
			wastewater. Loss of capacity of resins with time.	
Electrochemical	The process involves: Electro-deposition, Electro-	<i>Merits:</i> Ease of operation.	<i>Merits</i> : Both soluble and insoluble	Chen (2004), Cheng et al. (2007), Sun et al. (2009), Tran
	coagulation, Electro-flotation, and Electro-oxidation	In situ generation of coagulant. Sludge production is verv less.	organic contaminants can be degraded.	et al. (2017)
		Can be used as an alternative to	Carboxyl functional group pol-	
		conventional coagulation treat-	lutants can be treated effectively.	
		ment. Very high efficiency of the metal		
		removal process.		
		Demerits:	Demerits:	
		A high proportion of chemicals are required for cosmilation and	Chlorinated organics create sec-	
		precipitation.	Energy consumption is high.	
		A solid waste disposal problem.	Compound-specific removal.	
		The time of precipitation		
		depends upon the metal ions.		
		Poor settling.		
		It is not a cost-effective		
		treatment.		

38

And Corper         Merins:         Current et au.         Separated.         Current et au.         Curent au.         Curent au. <t< th=""><th></th><th>Electrodialysis</th><th>Merits: Highly concentrated heavy metal</th><th><i>Merits:</i> Metal-organic complexes can be</th><th>Kongsricharoern and Polprasert (1996), Aydin et al. (2019),</th></t<>		Electrodialysis	Merits: Highly concentrated heavy metal	<i>Merits:</i> Metal-organic complexes can be	Kongsricharoern and Polprasert (1996), Aydin et al. (2019),
Image: constraint of the second of			tous can be treated. Suitable for metal ion concentra- tion below $20 \text{ mg L}^{-1}$ . Chromium and copper metals	separated. Reclamation of treated water.	OUTIEN EI AI. (2020)
Image: Construction of the construction of Nacl is low.         Demecrits: Correston problem.         Demecrits: Constantiants induce Heavy metal concentration ingher than 1000 mg L <sup>-1</sup> cannot be effectively treated.         Demecrits: Concentration of Nacl is low.           Energy consumption is high.         Demecrits: Deriodic maintenance.         Demecrits: Concentration of Nacl is low.         Demecrits: Energy consumption is high.           e         Utra-filtration (UF)         Demecrits: Deriodic maintenance.         Demecrits: Concentration of Nacl is low.           e         Utra-filtration (MEUT)         Merins: Demecrits:         Merins: Use of chemicals is avoided.         Landabun-Aguine et al. (2010). If Zefatavy and Multi- Now then the Nome than 90% removal efficiency for pld 5-9.5 can be treated.         No hazardous by-products.         Coll.01, Izervi (2019)           Polymer-enhanced ultra- filtration (PEUF)         Definer et al. (2010). IZ Zefatavy and Multi- Polymer-enhanced ultra- filt space requirement for metals:         Merins: Demecrits:         Demecrits: Demecrits:           Demecrits:         Demecrits:         Demecrits:         Demecrits:         Demecrits:			ions can be recovered.		
Image: Conservent protect     Earcy state procentration     Fourteent of the efficient only when the beefficient only when the beefficient antitenance.       Indent team     Deerfficient only when the concentration is high.     Periodic maintenance.       Interaint methods     Ditra-filtration UF), and     Merits:       Interaint methods     Arris:     Ditra-filtration UF), and       Intracher     Merits:     Merits:       Polymer-enhanced ultra-filtra-     More than 90% removal effi-     Usage of chemicals is avoided.       Intracher     More than 90% removal effi-     Usage of chemicals is avoided.       Polymer-enhanced ultra-filtra-     Intra-filtra-     Dimercial and burn-spaine et al.       Intration (PEUF)     p15 -9,5 can be handled.     wastewater.       Polymer-enhanced ultra-     Intration (PEUF)     p15 -9,5 can be handled.       Recovery of heavy metals.     Demerits:     Demerits:       Demerits:     Demerits:     Demerits:       Demerits:     Demerits:     Merits:       und sufficiency depends     wastewater.     Merits:       Demerits:     Demerits:     Merits:       Demerits:     Demerits:     Demerits:       Demerits:     Demerits:     Demerits:       Dop of the ph     Merits cost is high.       Demerits:     Demerits:     Demerits:       D			Demerits:	Demerits: Organic contaminante induca	
Periodic maintenance.     Very efficient only when the be effectively treated.     concentration of NaCl is low.       Periodic maintenance.     Deration is not simple.     Energy consumption is high.       Periodic maintenance.     Deration is not simple.     Energy consumption is high.       Periodic maintenance.     Ditra-filtration (UF), and Micellar-enhanced ultra-filtra- tion (MEUF)     Menins:     Molimari et al. (2008).       Polymer-enhanced ultra- filtration (PEUF)     Menins:     Menins:     Molimari et al. (2008).       Polymer-enhanced ultra- filtration (PEUF)     Disconcentration with Biltration (PEUF)     Molimari et al. (2008).       Periodic maintenance.     Menins:     Disconcentration with Biltration officiency for pH 5-9.5 can be handled.     Menins:       Periodic maintenance     Dometration with Biltration (PEUF)     Disconcentration with Biltration officiency for ph 15-9.5 can be handled.     Menins:       Periodic maintenance     Disconcentration with Biltration (PEUF)     Disconcentration with Biltration officiency depends     Menins:       Peneritis:     Demeritis:     Demeritis:     Demeritis:     Demeritis:       Peneritis:     Demeritis:     Demeritis:     Beneritis:       Peneritis:     Demeritis:     Demeritis:     Beneritis:       Peneritis:     Demeritis:     Demeritis:     Beneritis:       Only efficient when the PH     Distration cost is high.			Heavy metal concentration	fouling.	
Internation of NaCl is low.     Deficition is not simple.     Energy consumption is high.       Periodic maintenance.     Energy consumption is high.     Deration is not simple.       Intra-filtration (UF), and Micellar-enhanced ultra-filtra- tion (MEUF)     Merins:     Merins:       Micellar-enhanced ultra-filtra- tion (MEUF)     Merins:     Merins:       Polymer-enhanced ultra-filtra- tion (MEUF)     Merins:     Merins:       Noiner-enhanced ultra- filtration (PEUF)     Ion comentation with High separation efficiency for Polymer-enhanced ultra- filtration (PEUF)     Molimari et al. (2008), High separation efficiency for PH 5-9.5 can be handled.       Noiner-enhanced ultra- filtration (PEUF)     IO-112 mg L <sup>-1</sup> can be treated. PH 5-9.5 can be handled.     No hazardous by-products. PH 5-9.5 can be handled.       Noiner-enhanced ultra- filtration (PEUF)     IO-112 mg L <sup>-1</sup> can be treated. PH 5-9.5 can be handled.     No mazadous by-products. PH 400 metals.       Demeritis:     Demeritis:     Membrane replacement. Membrane replacement.     Membrane easily fouled by nat- ural organic contaminants.       Demeritis:     Demeritis:     Demeritis:     Membrane replacement. Membrane replacement.       Upto of polymer used in treating officets PEUF.     Operating cost is high.       Only efficient when the pH     Only efficient when the pH			higher than 1000 mg L <sup>-1</sup> cannot	Very efficient only when the	
Iteratment methods     Energy consumption is high.       e     Ultra-filtration (UF), and Micellar-enhanced ultra-filtra- tion (MEUF)     Merits:     Molimari et al. (2008), Usage of chemicals is avoided.       Polymer-enhanced ultra-filtra- tion (MEUF)     More than 90% removal effi- bolymer-enhanced ultra-filtra- ciency     No hazardous by-products.     Molimari et al. (2008), No hazardous by-products.       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     No hazardous by-products.     Molimari et al. (2019)       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     No hazardous by-products.     gan (2011), Isawi (2019)       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     wastewater.     gan (2011), Isawi (2019)       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     wastewater.     gan (2011), Isawi (2019)       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     wastewater.     gan (2011), Isawi (2019)       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     wastewater.     gan (2011), Isawi (2019)       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     wastewater.     gan (2011), Isawi (2019)       Polymer-enhanced ultra- filtration (PEUF)     10-112 mg L <sup>-1</sup> can be treated.     wastewater.     gan (2011), Isawi (2019)       Polymer-enhanced ultra- filtration (			be effectively treated.	concentration of NaCl is low.	
Irreatment methods     Merins:     Merins:       e     Ultra-filtration (UF), and Micellar-enhanced ultra-filtra- tion (MEUF)     Merins:     Molinari et al. (2008), Usage of chemicals is avoided.       ioon (MEUF)     More than 90% removal effi- tion (MEUF)     No hazardous by-products.     Molinari et al. (2008), Landaburu-Aguirre et al.       ioon (MEUF)     Dons concentration with filtration (PEUF)     No hazardous by-products.     Molinari et al. (2008), Complexes in the pH 5-9.5 can be handled.       Recovery of heavy metals.     Dons L-1 can be treated.     wastewater.     (2011), Isawi (2019)       Recovery of heavy metals.     Demerits:     Demerits:     (2011), Isawi (2019)       Recovery of heavy metals.     Demerits:     Membrane casily fouled by nat- ural organic complexes in the wastewater.     Molinari et al. (2008), El Zeftavy and Muli- heavy metals.       Demerits:     Demerits:     Demerits:     Demerits:     (2011), Isawi (2019)       Recovery of heavy metals.     Demerits:     Demerits:     Demerits:       Demerits:     Demerits:     Demerits:     Demerits:       Optorating cost is high.     Optorating cost is high.     Demerits:       Optorating cost is high.     Optorating cost is high.     Deperating cost is high.			Periodic maintenance. Operation is not simple.	Energy consumption is high.	
metrix:         Metrix:         Metrix:         Molimari et al. (2008), Landaburu-Aguirre et al.           Micellar-enhanced ultra-filtra- tion (MEUF)         More than 90% removal effi- bolymer-enhanced ultra-filtra- tion (MEUF)         Molimari et al. (2008), Landaburu-Aguirre et al.           Polymer-enhanced ultra- filtration (PEUF)         Ions concentration with Domeration with 10-112 mg L <sup>-1</sup> can be treated, 10-112 mg L <sup>-1</sup> can be treated, mailer space requirement for installation.         Molimari et al. (2019), Regovery of heavy metals.           Demerits:         Demerits:         Demerits:         Molimari et al. (2019)           Merrix:         No hazardous by-products.         2010), El Zeftawy and Muli- tigan (2011), Isawi (2019)           Merrix:         Demerits:         Demerits:         Demerits:           Membrane replacement.         Newstewater.         Newstewater.         Newstewater.           Membrane replacement.         Membrane easily fouled by nat- ureal organic contaminants.         Membrane easily fouled by nat- ureal organic contaminants.           Demerits:         Demerits:         Demerits:         Demerits:           Operating cost is high.         Operating cost is high.         Demerits:	d treatme	nt methods			
Micellar-enhanced ultra-filtra- tion (MEUF)More than 90% removal effi- tion (MEUF)Usage of chemicals is avoided. Usage of chemicals is avoided.Landaburu-Aguirre et al.Polymer-enhanced ultra- tion (MEUF)Ions concentration with tight separation efficiency for organic complexes in the Smaller space requirement for installation.No hazardous by-products. tight separation efficiency for wastewater.Landaburu-Aguirre et al.Polymer-enhanced ultra- filtration (PEUF)Ions concentration with tight separation efficiency for voganic complexes in the wastewater.Landaburu-Aguirre et al. (2010), El Zeftawy and Mulli- gam (2011), Isawi (2019)Polymer-enhanced ultra- filtration (PEUF)Ion-112 mg L <sup>-1</sup> can be treated. wastewater.No hazardous by-products. wastewater.Landaburu-Aguirre et al. (2010), El Zeftawy and Mulli- morganic complexes in the wastewater.Polymer-enhanced ultra- filtration (PEUF)Denerits: Membrane replacement. Membrane replacement.Demerits: Membrane cost is high.Col11, Isawi (2019) gam (2011), Isawi (2019)Pererits: moval efficiency depends upon the concentration of metals of polymer used in treating effects PEUF.Demerits: The operating cost is high.Landaburu-Aguirre et al. (2011), Isawi (2019)Operating cost is high.Operating cost is high.Demerits: metalDemerits: metalOnly efficient when the pHDemerits: metalDemerits: metalDemerits: metal	e	Ultra-filtration (UF), and	Merits:	Merits:	Molinari et al. (2008),
tion (MEUF)ciencyNo hazardous by-products.(2010), El Zeftawy and Mulli- Ins concentration with Ins concentration with Ins concentration with Delymer-enhanced ultra- installation.No hazardous by-products. Ins enconcentration with High separation efficiency for organic complexes in the wastewater.No hazardous by-products. (2010), El Zeftawy and Mulli- gan (2011), Isawi (2019)Polymer-enhanced ultra- installation.IO-112 mg L <sup>-1</sup> can be treated. wastewater.No hazardous by-products. magnic complexes in the wastewater.(2010), El Zeftawy and Mulli- gan (2011), Isawi (2019)Demerins: mistallation.Demerins: Membrane requirement for installation.Demerins: matewater.Demerins: matewater.Demerins: monthe concentration of metals upon the concentration and type of polymer used in treating effects PEUF.Demerins: metal concentration and type of polymer used in treating offects PEUF.Demerins: metal concentration and type of polymer used in treating offects PEUF.Operating cost is high.		Micellar-enhanced ultra-filtra-	More than 90% removal effi-	Usage of chemicals is avoided.	Landaburu-Aguirre et al.
Polymer-enhanced ultration (PEUF)Ions concentration with filtration (PEUF)High separation efficiency for organic complexes in the organic complexes in the organic complexes in the matter space requirement for installation.High separation efficiency for organic complexes in the organic complexes in the matter space requirement for installation.High separation efficiency for gan (2011), Isawi (2019)Polymer-enhanced installation.IO-112 mg L <sup>-1</sup> can be treated. wastewater.High separation efficiency for organic complexes in the wastewater.gan (2011), Isawi (2019)Demerits: metallation.Demerits: Membrane replacement. Membrane replacement. Membrane replacement. Membrane for MEUF.Demerits: Membrane contaminants. The operating cost is high.gan (2011), Isawi (2019)Poterating of polymer used in treating of type of polymer used in treating officient when the pHDemerits: Membrane the pHgan (2011), Isawi (2019)		tion (MEUF)	ciency	No hazardous by-products.	(2010), El Zeftawy and Mulli-
filtration (PEUF)10–112 mg L <sup>-1</sup> can be treated. pH 5–9.5 can be handled.organic complexes in the wastewater.PH 5-9.5 can be handled.wastewater.Smaller space requirement for installation.wastewater.Smaller space requirement for installation.wastewater.Becovery of heavy metals.bemerits:Demerits:Demerits:Demerits:Membrane replacement.Membrane replacement.Membrane easily fouled by nat- ural organic contaminants.Heavy metal concentration of metals type of polymer used in treating effects PEUF.The operating cost is high.Operating cost is high.Operating cost is high.Only efficient when the pHOnly efficient when the pH		Polymer-enhanced ultra-	Ions concentration with	High separation efficiency for	gan (2011), Isawi (2019)
pH 5-9.5 can be handled.wastewater.Smaller space requirement for installation.wastewater.Smaller space requirement for installation.wastewater.Recovery of heavy metals.Demerits:Recovery of heavy metals.Demerits:Removal efficiency depends upon the concentration of metalsDemerits:Membrane replacement.Membrane easily fouled by nat- ural organic contaminants.Heavy metal concentration of metals and surfactants for MEUF.The operating cost is high.Gorating cost is high.Operating cost is high.Only efficient when the pHOnly efficient when the pH		filtration (PEUF)	$10-112 \text{ mg L}^{-1}$ can be treated.	organic complexes in the	
Smaller space requirement for installation.Smaller space requirement for installation.Recovery of heavy metals.Recovery of heavy metals.Recovery of heavy metals.Demerits:Rembrane replacement.Demerits:Membrane replacement.Membrane easily fouled by nat- ural organic contaminants.Upon the concentration of metalsural organic contaminants.Iteavy metal concentration and type of polymer used in treating effects PEUF.The operating cost is high.Only efficient when the pHOnly efficient when the pH			pH 5–9.5 can be handled.	wastewater.	
installation.Recovery of heavy metals.Recovery of heavy metals.Recovery of heavy metals.Demerits:Demerits:Membrane replacement.Membrane replacement.Membrane replacement.Membrane easily fouled by nat-Npon the concentration of metalsupon the concentration of metalsand surfactants for MEUF.Heavy metal concentration andtype of polymer used in treatingeffects PEUF.Operating cost is high.Only efficient when the pH			Smaller space requirement for		
Recovery of heavy metals.Demerits:Demerits:Demerits:Membrane replacement.Membrane replacement.Membrane replacement.Membrane replacement.Membrane replacement.Membrane replacement.Membrane resits fouled by nat-Npon the concentration of metalsupon the concentration of metalsand surfactants for MEUF.Heavy metal concentration andtype of polymer used in treatingeffects PEUF.Operating cost is high.Only efficient when the pH			installation.		
Demerits:Demerits:Membrane replacement.Demerits:Membrane replacement.Membrane easily fouled by nat-Removal efficiency dependsural organic contaminants.upon the concentration of metalsThe operating cost is high.and surfactants for MEUF.The operating cost is high.Heavy metal concentration and type of polymer used in treating effects PEUF.Pemerits:Only efficient when the pHOnly efficient when the pH			Recovery of heavy metals.		
Membrane replacement.Membrane easily fouled by nat- Removal efficiency depends upon the concentration of metals upon the concentration of metals and surfactants for MEUF.Membrane easily fouled by nat- ural organic contaminants.Removal efficiency depends upon the concentration of metals and surfactants for MEUF.Membrane easily fouled by nat- ural organic contaminants.Removal efficiency depends upon the concentration and type of polymer used in treating effects PEUF.Membrane easily fouled by nat- ural organic contaminants.Operating cost is high.Operating cost is high.Only efficient when the pHOnly efficient when the pH			Demerits:	Demerits:	
Removal efficiency depends upon the concentration of metals upon the concentration of metalsural organic contaminants.upon the concentration of metals and surfactants for MEUF.The operating cost is high.Heavy metal concentration and type of polymer used in treating effects PEUF.The operating cost is high.Operating cost is high.Only efficient when the pH			Membrane replacement.	Membrane easily fouled by nat-	
upon the concentration of metals The operating cost is high. and surfactants for MEUF. Heavy metal concentration and type of polymer used in treating effects PEUF. Operating cost is high.			Removal efficiency depends	ural organic contaminants.	
and surfactants for MEUF. Heavy metal concentration and type of polymer used in treating effects PEUF. Operating cost is high. Only efficient when the pH			upon the concentration of metals	The operating cost is high.	
Heavy metal concentration and type of polymer used in treating effects PEUF. Operating cost is high. Only efficient when the pH			and surfactants for MEUF.		
type of polymer used in treating effects PEUF. Operating cost is high. Only efficient when the pH			Heavy metal concentration and		
Operating cost is high. Only efficient when the pH			type of polymer used in treating		
Only efficient when the pH			Onerating cost is high		
-			Only efficient when the pH		

Table 3.1 (contin	ned)			
Treatment technology	Treatment process	Heavy metal treatment	Organic contaminant treatment	References
		condition of the wastewater is above 6.		
	Complexation-ultra-filtration	Merits:	Merits:	Trivunac and Stevanovic
	1	High separation selectivity.	Simultaneous removal of organic	(2006), Siyanytsya et al. (2008),
		Water-soluble polymer ligands.	pollutants and heavy metal com-	Borbély and Nagy (2009),
		Lower energy requirements.	plexes.	Garba et al. (2019)
		Higher bonding selectivity.	Lower energy requirement.	
		mensed	nonto ves u ace organic contanti- nants effectively	
		The complexing agent can be	The binding capacity of polv-	
		easily regenerated and reused.	meric complexing agents.	
		Demerits:	Demerits:	
		Efficiency depends upon com-	Ultrafiltration efficiency	
		plexation agent.	decreases when organic pollut-	
		Condition of membrane deter-	ant complex increases.	
		mines the metal removal process.		
		Membrane fouling.		
		The presence of salt affects the		
		membrane surface charge reduc-		
		ing the efficiency.		
	Reverse osmosis (RO)	Merits:	Merits:	Ipek (2005), Yangali-
		Many types of ions, as well as	Trace organic pollutants are eas-	Quintanilla et al. (2011), Rodri-
		bacteria, can be removed.	ily removed.	gues Pires da Silva et al. (2016)
		Membrane pore size is less than	97% removal efficiency for ionic	
		2 nm.	contaminants.	
		90-99% removal efficiency.	Membrane availability is	
			plentiful.	
		Demerits:	Demerits:	
		Separation efficiency depends	Membrane fouling.	
		upon the concentration, pressure,		

Table 3.1 (contin	ued)			
Treatment technology	Treatment process	Heavy metal treatment	Organic contaminant treatment	References
		depends upon the current density and pH. Oxidant concentration generated is relatively low.	Wide ranges of organic con- taminants cannot be treated. pH-sensitive.	
	Photocatalytic oxidation	<i>Merits:</i> Widely used in the removal of heavy metal complexes. UV radiation can be used in the disinfection process as well. Solar energy can be utilized.	Merits: Effective polishing step. Taste and odor-causing organic pollutants such as pesticides, geosmin, and methyl tert-butyl ether are effectively oxidized.	Antoniadis et al. (2010), Chen et al. (2017), Brillas (2020)
		Demerits: Depends upon the characteristics of metal ions. Efficient UV illumination in large-scale treatment is inefficient.	Demertis: Bulk organic content and high turbidity requires a high UV dosage. Not cost-effective.	
	Ozonation	Merits: High removal efficiency. Improvement of wastewater biodegradability.	Merits: Lower ozone dose requirement. Very efficient. Aromatic compounds, alkenes, and certain pesticides are effec- tively removed by ozone.	Clark et al. (1988), Xia and Hu (2018), Honarmandrad et al. (2020)
		Demerits: The low solubility of ozone in water. Energy consumption is high to produce ozone. Low oxidation efficiency.	<i>Demerits:</i> Ozone solubility depends upon pH. Alkanes are treated poorly. Expensive treatment technology.	

42

	Discharge plasma oxidation	<i>Merits</i> : High reactivity and can be oper- ated without chemical reagents.	<i>Merits:</i> Very effective in treating pesti- cides 90% removal in 5 minutes. High concentration of pollutants	Magureanu et al. (2018), Wang et al. (2017), Liu et al. (2020)
		<i>Demerits:</i> High energy consumption and operating cost. Handling of this technology is complex.	can be treated. <i>Demerits:</i> Degrading efficiency depends upon the molecular structure of the target pollutants. Unstable organic peroxides are	
Biological treatm	nent methods		TOTILICA.	
Activated sludge	The process involves: Bacteria, Aeration, and Agitation	<i>Merits:</i> Sludge is re-circulated back con- tinually. Metals like iron and lead are efficiently removed by the process. <i>Demerits:</i> Toxic sludge and disposal of sludge is problematic. Large area and proper maintenance.	Merits: Acid and lipophilic compounds are biodegradable and treated effectively. Biodegradation is the major removal mechanism. Demerits: High toxic contaminants cannot be treated. The concentration of pollutants should be low enough to support bacterial life. Trace organic compounds are poorly treated by activated sludge.	Oliver and Cosgrove (1974), Pagnanelli et al. (2009), Karpińska and Kotowska (2019)
Trickling filters	The process involves: Aeration, Coarse media (stone or plastic), and Micro- organisms	<i>Merits:</i> Various types of filter media (gravel media or plastic media) are available for effective	<i>Merits:</i> Surfactants polluted water are effectively treated. Energy consumption is less.	Dermou et al. (2007), Ziolko et al. (2009), Katam et al. (2020)
			-	(continued)

ble 3.1 (contin	(ned)			
atment mology	Treatment process	Heavy metal treatment	Organic contaminant treatment	References
		treatment. The bacterial population can be easily maintained and controlled. Very effective. Economical process for indus- trial wastewater treatment. The use of chemicals is avoided.	Biofilm culture (algal-bacteria consortia) enhances the removal efficiency of the filters. Organic matters are easily biodegraded.	
		<i>Demerits:</i> High probability of clogging. High capital cost. Microbial community is affected by toxic metals. Not effective as activated sludge process in treating heavy metals.	Demerits: Longer retention time is needed. Toxic contaminants affect the micro-organism population.	
ds	The process involves: Constructed shallow basins, and Micro-organisms	<i>Merits:</i> Various types of heavy metal ions can be treated. Very cheap and easy to operate.	<i>Merits:</i> A low concentration of organic contaminants is removed effec- tively. 60% removal efficiency for trace organic pollutants such as caf- feine and naproxen.	Polprasert and Champratheep (1989), Jasper et al. (2013), Gad and Abdalla (2017), Qaderi et al. (2019)
		<i>Demerits:</i> Slow and inefficient in removing heavy metal ions. Depends upon the interaction of micro-organisms, sunlight, and oxygen. Water source is one of the	Demertits: Time-consuming process. Wide range of organic contami- nants cannot be treated.	

		limiting factors. Not suitable for all geographic areas only for arid and semi-arid zones.		
Biosorption	The process involves: Adsorption, and Precipitation	Merits: Simple in operation and high efficiency. Agricultural by-products can be used as adsorbents (rice husk, pecan shells, maize cob, rice straw, rice husk, coconut shell). Low cost of adsorbents and regeneration of bio sorbent. Usage of dead biomass reduces the need for nutrient supplication. <i>Demerits:</i> Removal efficiency depends completely upon the cellulose, lignin, carbohydrate, and silica properties present in the adsor- bents. A sudden change in pH and temperature affects the efficiency of the bio sorbent.	Merits: Simple and cheap compared to advanced treatment technolo- gies. Ability to treat multicomponent organic contaminants. High selectivity of treatment. Less sludge is generated. Less sludge is generated. Demerits: Disposal of biowaste (spent bio sorbent). Putrefaction of biomass under moist conditions. Limited regeneration capacity of bio sorbents.	Chaukura et al. (2016), Ramírez Calderón et al. (2020)
Biopolymers	Biopolymer adsorbent	<i>Merits:</i> Widely used in industries capable of reducing metal ions concentration to parts per billion. Low cost and higher removal	<i>Merits:</i> Biopolymers such as chitin and chitosan are renewable and bio- degradable. Very effective in the treatment of	Vidal and Moraes (2019); Zia et al. (2020)
				(continued)

	~			
Treatment technology	Treatment process	Heavy metal treatment	Organic contaminant treatment	References
		efficiency. Abundant availability and bio- degradability of biopolymers.	phenolic compounds and pesti- cides (80–90% removal).	
		Demerits: Rapid degradability of biopolymers.	<i>Demerits:</i> The selection of biopolymers is based upon the pollutant load in the wastewater. Removal efficiency depends upon the ionic strength of the organic pollutant, pH, contact time, and temperature.	

 Table 3.1 (continued)



Fig. 3.2 Fate of the contaminants after their entry into the cell system

sorption capacity and availability in seas and oceans. Due to the presence of alkaline metal ions in the composition of algal cell walls the heavy metal ions in the water can be easily treated through a simple ion-exchange process. These sorbents have the metal-sequestering property that can be used to reduce the concentration of heavy metal ions in the solvent from parts per million (ppm) to parts per billion (ppb) level. Biosorption capacity determines the number of metal ions that microalgae can bind on the surface, and it is denoted by  $q_{max}$ . Brinza et al. (2007) reviewed the biosorption capacity of some marine microalgal species involving the commonly detected heavy metals found in the wastewater, as shown in Table 3.2.

	$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$						
Marine microalga							
	Pb	Cd	Ni	Zn	Cu		
Chlorella sp.	0.46	0.44	0.31	0.18	0.55		
Chlorococcum sp.	0.23	0.15	0.27	0.21	0.36		
Cyclotella cryptica	0.42	-	0.14	0.1	0.33		
Spirogyra sp.	0.49	0.27	0.12	0.23	0.53		
Lyngbya taylorii	0.84	0.32	0.43	0.37	-		
Microcystis aeruginosa	0.35	-	0.21	0.23	0.37		
Scendesmus sp.	0.45	0.11	-	0.35	0.22		

 Table 3.2
 The maximum heavy metal biosorption capacity of some marine microalgal species (based on data from Brinza et al. 2007)

#### **Biosorption mechanism**



Fig. 3.3 Biosorption of pollutants in marine microalgae

Biosorption is an extracellular process that is carried out in the cell membrane in which the algal biomass binds the heavy metals in the cell wall. The algal cell wall is composed of polysaccharides that contain sulfate. Imidazole, phosphate, hydroxyl, amine, and amino functional groups act as a binding site for the heavy metals to be adsorbed. The biosorption mechanism (Fig. 3.3) can be divided into metabolism-dependent biosorption in which transportation of the pollutant across the cell membrane takes place, followed by intercellular accumulation or detoxification. Metabolism-independent mechanisms involve ion exchange, complexation, chelation, and precipitation process. For example, the cell wall of microalgae is composed of polysaccharides, lipids, and proteins that provide many functional groups capable of attracting both anionic and cationic heavy metal ions exchanging them with the

functional groups present in the cell wall. While investigating the mechanism for removing Cr<sup>3+</sup>, Cd<sup>2+</sup>, and Cu<sup>2+</sup> by *Spirulina*, Chojnacka et al. (2005) found that hydroxyl, carboxyl, and phosphate functional groups were involved in the removal of the metal ions by the ionic-exchange process. Similarly, a microalgal strain, Tetraselmis marina AC16-MESO, could remove Cu (90%), Fe (100%), and Mn (50%) after 72-h incubation period, mostly by complexation of metal ions onto functional groups at the cell surface (Cameron et al. 2018). In fact, complexation mechanism is the result of electrostatic attraction between heavy metal ions and organic molecules present on the cell which act as ligands. The complex formation between the metal ion and ligand is due to the covalent bonds. The functional group (phosphonate, carboxyl, and amine) present in the cell wall of *Chlorella miniate* removed  $Cr^{3+}$  by the complexation process (Han et al. 2006). Organic acids such as citric, fumaric, lactic, oxalic and gluonic have been found to chelate metal ions resulting in the formation of metallo-organic complexes. Chelation is the advanced form of complexation mechanism in which the metal ion would bond with a ligand in many positions at the same time with higher stability. Chlamydomonas reinhardtii removed  $Hg^{2+}$  by direct chelation mechanism in which glutathione not only adsorbed the metal ion but also reduced the toxicity of the pollutant in water (Perales-Vela et al. 2006).

Two marine algae, *Chlorella* sp. and *Phormidium* sp., exposed to tannery wastewaters removed Cr concentration by 81 and 90%, respectively, at the end of 15 days incubation period as revealed by metabolic mechanism (Das et al. 2018). When the metal ion solubility decreases, the bioavailability is reduced, resulting in the mechanism of precipitation. Upon exposure to the heavy metal-polluted medium, the algal biomass favored precipitation that was based on pH of the medium. If pH of the medium increases, the active sites on the cell wall attract heavy metal ions. Cu, Ag, and Pb ions were removed by the alga, *Tertaselmis suecica*, by the mechanism of precipitation due to the presence of phosphates on the cellular surface (Ballan-Dufrançais et al. 1991). While growing *Chlorella* sp. in seawater-based medium, nearly 4% of Cd supplemented was found precipitated due to an increase in pH to 8 besides 67% accumulation and 25% adsorption of the metal (Matsunaga et al. 1999).

#### 3.4.1.2 Factors Influencing Biosorption of Heavy Metals

**Biotic Factors** 

**Algal Species** Marine microalgae can be classified into three broad categories based on the composition of pigment color in green algae (Chlorophyta), red algae (Rhodophyta), brown algae (Phaeophyta) (Davis et al. 2003). Romera et al. (2007) summarized the biosorption capacity of algae related to some heavy metals as indicated in Table 3.3. Brown algae have a higher sorption capacity than red and green algae due to their high alginate content and the presence of functional groups in the structure. But the use of brown and red marine algae has a major drawback due

Phylum	Cadmium	Nickel	Zinc	Copper	Lead
Chlorophyta	0.60	0.51	0.37	0.50	0.80
Rhodophyta	0.20	0.27	-	-	0.65
Phaeophyta	0.90	0.84	0.67	1.01	1.23

**Table 3.3** Average heavy metal sorption capacity  $(q_{\text{max}})$ , in mmol g<sup>-1</sup>, of different algae (data based on Romera et al. 2007)

to the presence of certain organic compounds such as alginate. Also, the pigments generate secondary pollutants and reduce the biosorption capacity. In green algae the secondary pollutant generation is insignificant but their biosorption capacity is lower as compared to brown and red algae. While studying the impact of biotic factors on Cu adsorption capacity in marine microalgae, Levy et al. (2007) observed that *Dunaliella tertiolecta* was least sensitive than *Minutocellus polymorphus* and was depended on uptake rates across cell membrane rather than the taxonomic status and cell size.

**Concentration of Biomass** In the biosorption process, the removal efficiency depends upon the biomass concentration because of the greater availability of binding sites on the cell surface. Increased biomass concentration enhances the removal percentage of heavy metals. An increase in biomass concentration of *Ulva fasciata* from 0.5 to 4 g L<sup>-1</sup> resulted in the improvement of Pb removal efficiency in the range of 42–75%, while the removal efficiency of Cd increased from 43 to 73% with the increase in biomass concentration from 0.5 to 6 g L<sup>-1</sup> (Nessim et al. 2011). Increased biomass concentration often reduces the biosorbent capacity of microalgae because of the reduction in intercellular distance and cell agglomeration. Kaparapu and Prasad (2018) observed higher biosorption of Cd(II) in *Nannochloropsis oculata* with biomass concentration probably due to the partial biomass aggregation that results in surface area reduction.

**Tolerance** Algal species are known to grow, adapt, and tolerate hazardous environmental conditions. Heavy metal tolerance in algae depends upon the algal species. However, members of Chlorophyceae are generally known to tolerate  $Cu^{2+}$ ,  $Zn^{2+}$ , and  $Cd^{2+}$ . Strains of *Chlorella* sp. isolated from mercury-contaminated sites tolerated higher Hg<sup>2+</sup> concentration than the isolates from uncontaminated habitats (Gaur and Rai 2001). Pérez-Rama et al. (2010) observed 87% of Cd accumulation in a marine microalga, *T. sueccia*, and was related to phytochelatin synthesis. Folgar et al. (2009) reported that *Dunaliella salina* was tolerant to higher concentrations of Cd due to the intracellular metal-binding ligands.

**Surface Area to Volume Ratio** The ratio of surface area to volume in microalgae influences the sequestration of heavy metals in the solution. The take-up nutrients, in terms of per biomass, are faster in microalgae than macroalgae because of the size, growth, metabolism, and biochemical composition (Hein et al. 1995). Khoshmanesh et al. (1997) reported that the uptake of Cd was similar in an algal species having different sizes. The microalgal culture with a specific surface area of 2.20 m<sup>2</sup> mg<sup>-1</sup>

cells showed higher uptake of Cd ions as compared to the culture with a specific surface area of 0.98  $m^2 mg^{-1}$  cells.

#### Abiotic Factors

**pH** Biosorption of heavy metals in solution depends upon pH conditions due to the functional groups that dissociate at certain pH levels in the algal biomass. The maximum sorption of heavy metal ions by the marine microalgal biomass was obtained at a pH range between 4.0 and 6.0. The observed percentage removal efficiency for Cr, Cd, As, Pb, and Hg at pH 6.0 were 98.30, 92.50, 96, 92.20, and 80, respectively (Kumar et al. 2020; Leong and Chang 2020). When the pH value is lower than 6.0, the hydrogen ion concentration does not compete with the metal ions, and during adsorption of heavy metals no vacant active sites are created in the algal biomass (Gupta et al. 2011). If the pH value is greater than 6.0, the metal species are hydrolysed and are no longer available for the biosorption process (Romera et al. 2007). Kaparapu and Prasad (2018) reported that the biosorption of Cd(II) at pH > 7was reduced in a marine microalga, Nannochloropsis sp., and at pH 2-4 there was a competition between metal ions and metal-binding sites located on algal cell surface. The reduced Cd biosorption at higher pH was attributed to the maximum immobilization of positive charges. When the initial pH was maintained at 7.8, the cells of T. suecica were metabolically active and increased in cell number from 30 to  $40 \text{ mg g}^{-1}$  within 48 h, suggesting that the live cells are more suitable for biosorption than dead cells (Pérez-Rama et al. 2010).

**Temperature** Temperature plays a vital role in the biosorption process as it influences the process in both positive and negative ways depending upon the range in temperature (Khambhaty et al. 2009). The solubility of metal ions was found to be higher at elevated temperatures, but an increase in temperature decreases the biosorption capacity of the biomass. The maximum biosorption of Cu<sup>2+</sup> ions attained at 37 °C was 90% in *Spirulina* species but the biosorption capacity was reduced to 82% at 60 °C and then gradually decreased with further increase in temperature (Al-Homaidan et al. 2014). The biosorption efficiency of *N. oculata* biomass increased with contact time up to 90 min and remained constant (Kaparapu and Prasad 2018).

**Contact Time** The efficiency of biosorption process depends upon the contact time between the algal biomass and the heavy metal ion. It was observed that the optimum contact time for the maximum adsorption of 80–90% of Cu, As, Cd, Cr, Pb, and Hg by various marine algal species was within 60–90 min (Al-Homaidan et al. 2014; Leong and Chang 2020). Initially, many active sites are available on the cell surface for the adsorption of heavy metals, and there will be a reduction in active sites with time, resulting in a gradual decline in the removal capacity of biomass that requires regeneration of algal biomass. The amount of biosorbed Cd in *D. salina* biomass was greater after 24-h contact time and was subsequently reduced due to the enhanced sorption onto the cellular surface (Folgar et al. 2009). It has been reported that

biosorption yield of Cd(II) decreased with increased temperature at an optimal contact time due to the following reasons: relative increase in leaching tendency of ions from solid phase to bulk phase, and weakness of active sites for biosorption in the sorbed phase (Kaparapu and Prasad 2018).

#### 3.4.1.3 Desorption of Heavy Metals and Biomass Regeneration

Desorption of heavy metals is the process of recovering valuable metal ions from the algal biomass by adding eluent. The eluent restores the biosorbent to its original state for the reuse of biomass in the process. Mineral acids, complexing agents, and organic acids are used as the eluents as they are non-damaging to the sorbent, and they ensure the metal-binding capacity of microalgae. Desorption of  $Cr^{3+}$ ,  $Cd^{2+}$ , and  $Cu^{2+}$  from biomass of *Spirulina* sp. by nitric acid resulted in 98% removal of the metal ions (Chojnacka et al. 2005). In fact, *Chlorella vulgaris* remains unaffected even after five cycles of biomass regeneration using 0.1 M EDTA as eluent to recover Cd metal ions, and the adsorption capacity loss was less the 5.8% (Kumar et al. 2018). Both HCl and EDTA are the most used eluents for desorbing algal biosorbents. However, HCl decreases biosorption capacity of algal biomass after every wash, and the use of EDTA is not eco-friendly as it dissolves alginate upon every use which can lead to secondary pollution. Therefore, it is essential to screen the desorbing agents for efficient metal ion recovery.

#### 3.4.1.4 Heavy Metal Detoxification by Marine Microalgae

The ability of microalgae to adapt and survive in habitats contaminated with heavy metals and organic pollutants depends on genetic adaptation which enables them to develop defence mechanisms to resist and adapt the harsh environmental conditions (Nayaka et al. 2017). This mechanism of defence allows microalgae to develop some tolerance and resistance toward the pollutant that can detoxify the pollutants inside the cell. The defence mechanism involves the production of short-chained polypeptides such as phytochelatins (PCs) and metallothioneins (MTs) that are abundant in sulfhydryl and carboxyl groups and can bind to the pollutants (Cobbett and Goldsbrough 2002). The bound pollutant further moves in for internal detoxification process which involves conjugation of the pollutant with the polypeptides and further compartmentalization of the pollutants by transporting them into the vacuoles (Qin et al. 2006). Folgar et al. (2009) reported that metal complexing ligands in D. salina were rich in cystine although most of the known are GSH and PCs. They observed that levels of cystine synthesis led to maximum Cd accumulation intracellularly. In another study, D. salina was shown to be resistant to As which exhibited higher levels of lipid peroxidation with a differential expression of 65 proteins involved in energy metabolism, protein synthesis and folding, ROS scavenging, and amino acid synthesis (Ge et al. 2016). Wang et al. (2017) reported variation in thiols such as cysteine, glutathione, and PCs in D. salina exposed to arsenite and



Fig. 3.4 Biotransformation mechanism for heavy metal detoxification in microalgae. MMA, Monomethylarsonic acid; DMA, Dimethylarsinic acid (Source: National Research Council 2003)

demonstrated that transformation of arsenite-induced several PCs initially and later decreased under various phosphate regimes. The synthesis of PCs varied with As (V) and As(III) which affected GSH levels, suggesting that the conversion of GSH to PCs is essential for arsenite mitigation (Wang et al. 2017). The biochemical mechanisms involved in heavy metal detoxification by microalgae (National Research Council 2003) are presented in Fig. 3.4. Sathasivam and Ki (2019) observed higher levels of phytoene synthase (PSY), phytoene desaturase (PDS), and  $\beta$ -lycopene cyclase (LCY-B) in *T. suecica* exposed to copper.

#### 3.4.2 Removal of Organic Pollutants by Marine Microalgae

Human attempts to produce various organic compounds to protect many lives and support economic advantages significantly resulted in acute and chronic toxicity of some of these chemical substances making the biota deteriorate rapidly (Adeola 2004). Although these organic compounds are susceptible to degradation at a very slow process, they tend to persist in the environment or accumulate inside the biota (Subashchandrabose et al. 2013). Organic pollutants that are widely distributed in marine environments and prone to biodegradation by marine microalgae include phenolics, pesticides, persistent organic pollutants (POPs), and hydrocarbons (Dsikowitzky et al. 2011).

#### 3.4.2.1 Pesticides

Pesticides including insecticides, fungicides, and herbicides are often detected in marine waters due to the urban or agriculture runoff causing serious threats to the marine biota. Atrazine sensitivity, in terms of 96-h growth inhibition, for the estuarine phytoplankter, D. tertiolecta, in nutrient-replete media was 159.16 µg L<sup>-</sup> and was influenced by the duration and nutrient-limited conditions (Flood et al. 2018). Chen and Jiang (2011) reported enhanced catalase activity in D. salina when exposed to trichlorfon and dimehypo at lower concentrations of 0.025 g  $L^{-1}$  and  $0.0005 \text{ g L}^{-1}$ , respectively. In a toxicity study involving treatment of *D. salina* with dimethylphenol and dinitroaniline, Zhu and Jiang (2009) observed that the  $EC_{50}$ values were significantly higher when exposed to a single pesticide compared to their combination. However, increased concentrations led to significant inhibition in the growth of the microalga that was attributed to the effect on osmosis of cell membrane allowing toxicants to react with internal parts and damage membrane lipids. Thakkar et al. (2013) exposed D. tertiolecta and a brown tide alga, Aureococcus anophagefferens, to various concentrations of metachlor and observed a significant increase in cell size with glutathione production as detoxification mechanism. Although 40-50% of sublethal concentration of tributylin (TBT) could be removed by N. oculata and Dunaliella parava during 2-6 days of incubation, the former microalga adsorbed most of the added anti-fouling agent while the latter degraded it to mono-butyltin and di-butyltin (Taha et al. 2009). DeLorenzo and Serrano (2003) determined the toxicity of atrazine, chlorpyrifos, and chlorothalonil individually and as mixtures on D. tertiolecta and observed that atrazine and chlorothalonil concentrations at 25 and 33  $\mu g \ L^{-1}$  decreased growth rate, while chlorpyrifos was toxic only at >400  $\mu$ g L<sup>-1</sup>. In another study, the effect of herbicides such as diuron, irgarol, atrazine, and ametryn was tested toward D. tertiolecta in four different scenarios of increased temperature and salinity and reported that increasing temperature reduced growth but enhanced the contents of chlorophyll and starch and lipids (DeLorenzo et al. 2013).

#### 3.4.2.2 Hydrocarbons

Water soluble fraction of crude oil containing mono- and diaromatic hydrocarbons affected *D. tertiolecta* within 24 h though photosynthesis impairment and cell division inhibition occurred. Despite the well-known tolerance of *Dunaliella* species, the exponential phase measured in terms of photosynthesis was reduced while lag phase showed growth inhibition, suggesting that duration of exposure influenced the overall growth (Siron et al. 1991). Fabregas et al. (1984) reported stimulation in the growth of *T. suecica* upon exposure to low hydrocarbon concentrations in crude oil whereas the dispersant did not exhibit any selective toxicity. Dunstan et al. (1975) observed that low concentration (10 mg L<sup>-1</sup>) of oil had no effect on the growth of *D. tertiolecta*. Similarly, low concentration (0.05%) of light diesel and an oil

dispersant (0.005%), either alone or in combination stimulated the growth of *Chlorella salina* and impaired respiration (Chan and Chiu 1985). Photosynthesis in *D. tertiolecta* exposed to oil samples from tanker spill was significantly affected within 60 min, while survival of the cells was slightly affected (Carrera-Martinez et al. 2011). Jiang et al. (2002) exposed microalgal strains to four PAHs, viz., toluene, naphthalene, 2-methylnapthalene, and phenanthrene, and reported that *C. vulgaris* and *Platymonas subcordiformis* were least sensitive compared to other tested species.

Exposure of *Chlorella salina* to phenanthrene significantly increased the toxicity with an EC<sub>50</sub> value that ranged from 1.893 to 0.23 mg  $L^{-1}$ , and a decrease in pH from 9 to 6 was also significantly toxic suggesting that the acidification of sweater greatly influenced the effect of organic compounds (Chen et al. 2018a). Bretherton et al. (2018) observed that marine alga, D. tertiolecta, was resistant to oil and dispersant and referred to it as "robust" because chlorophyll was not affected during lag phase and was followed by biomass accumulation. Moreover, short-term exposure of *D. tertiolecta* to petroleum and diesel oil impacted the growth and photosynthetic performance and reported to recover during long-term incubation (Romero-lopez et al. 2012). Recently, Salinas-Whittaker et al. (2020) observed that D. tertiolecta exposed to water-soluble fraction (WSF) from fuel oil/diesel mixture increased physiological and biochemical response in unsaturated acyl chain of fatty acid suggesting the uptake of hydrocarbons. Mohammady et al. (2005) exposed Nannochloropsis salina to various concentrations (0-100%) of diesel fuel oil aqueous extract and observed a decrease in cell bioavailability leading to cell division and enhanced membrane permeability. Both the limitation of carbon and hormesis phenomenon, as evaluated by stable isotope analysis, were prevalent in Platymonas helgolandica when it was treated with water accommodated fraction of fuel oil (Liu et al. 2020). Dissolved crude oil at lower concentration (20 mg  $L^{-1}$ ) stimulated the growth of Dicrateria sp. but growth was inhibited with increased exposure time. However, consortia of marine microalgae involving Dicrateria sp. on biotreated seawater showed enhanced cell density that ranged from  $4.0 \times 10^5$  to  $1.7 \times 10^6$  cells mL<sup>-1</sup>. Chao et al. (2012) reported that four fuel oils, viz., F120, F180, F380, and F20 were toxic to a marine alga, *Chlorella* sp., due to the concentration of several PAHs. Hing et al. (2011) demonstrated that C. salina was able to tolerate diesel concentrations at steady state and was only affected when the concentration exceeded 170 mg  $L^{-1}$ . Very recently, Marques et al. (2021) reported that N. oculata was able to grow in petroleum-contaminated water exhibiting a PAH removal efficiency of 94%. In particular, the percentage removal of several organic compounds such as naphthalene, benzopyrene, and acenaphthylene was 89–99% due to their intracellular biodegradation by oxidoreductase enzymes.

#### 3.4.2.3 Other Organic Compounds

Phenol is an organic compound that results from the transformation of aromatic compounds via degradation, oxidation, and synthesis. Besides being enriched in coal

tar, phenol is also produced as a by-product from several industrial processes as well as during organic matter decomposition (Michalowicz and Duda 2007). Mofeed and Abdel-Aal (2015) found that exposure of *D. salina* to various concentrations (50- $200 \,\mu\text{mol}\,\text{L}^{-1}$ ) of phenol significantly affected antioxidant enzyme activities. Phenol at a concentration of 72 mg  $L^{-1}$  led to programmed cell death in marine microalgae by inducing changes in ultrastructure with shrinkage of the nucleolus and vacuole enlargement (Duan et al. 2017). During treatment of real refinery wastewater containing phenol and its derivatives such as o-cresol and p-cresol, marine alga, *Nannochloropsis* sp., removed >80% of both the cresols as compared to freshwater Chlorella sp. (Surkatti and Al-Zuhair 2018). The biodegradation was reported to occur in two steps: split in methyl group resulting in its conversion to methanol and further breakdown of phenol produced as an intermediate (Papazi et al. 2012). Bisphenol A, with production estimates of approximately two million tons, is well distributed in the environment and known for its endocrine disruption potential (Burridge 2003). While reporting the first toxicity data of chlorophenols on D. tertiolecta, Ertürk and Saçan (2012) reported that toxicity of chlorophenols decreased between 48 and 96 h due to the increase in pH of the medium or acclimation response of the marine microalga to the toxicants. Regardless of the exposure time, the toxicity was greater with an increasing number of chlorine atoms, while ortho-substituted chlorophenol was lesser than meta and para congeners (Ertürk and Saçan 2012). POPs are widely distributed due to domestic and industrial activities that are reported to reach marine environments (ter Schure et al. 2004; Lema et al. 2007). Polybrominated diphenyl ethers (PBDEs), the flame retardants, enhanced oxidative stress in D. salina with increased activities of superoxide dismutase, catalase, and glutathione reductase and decreased glutathione peroxidase activity (Zhao et al. 2017). Similarly, exposure of D. salina to dibutyl phthalate at 100 mg  $L^{-1}$  decreased glutathione peroxidase and superoxide dismutase (Wei et al. 2021).

#### 3.5 Conclusions

Besides highlighting the advantages of the use of marine microalgae for the removal of heavy metals and organic pollutants, we presented the inherent drawbacks of the conventional treatment processes. Marine microalgae respond to heavy metals in several ways such as biosorption and bioaccumulation; however, there is a very clear paucity of data on organic contaminant removal and the associated mechanisms. Furthermore, it is very clear that marine microalgae can offer sustainable approach in the treatment of heavy metals and organic pollutants for safer marine ecosystem and biomass production from microalgae after detoxification. Thus, this chapter presents the overall understanding of the potential of marine microalgae in the removal of heavy metals and organic pollutants.

#### References

- Adeniji AO, Okoh OO, Okoh AI (2017) Petroleum hydrocarbon profiles of water and sediment of Algoa Bay, Eastern Cape, South Africa. Int J Environ Res Public Health 14:1263
- Adeola FO (2004) Boon or bane? The environmental and health impacts of persistent organic pollutants (POPs). Hum Ecol Rev 11:27–35
- Ahmaruzzaman M (2011) Industrial wastes as low-cost potential adsorbents for the treatment of wastewater laden with heavy metals. Adv Colloid Interface Sci 166:36–59
- Alalwan HA, Kadhom MA, Alminshid AH (2020) Removal of heavy metals from wastewater using agricultural byproducts. J Water Supply Res Technol AQUA 69:99–112
- Al-Homaidan AA, Al-Houri HJ, Al-Hazzani AA et al (2014) Biosorption of copper ions from aqueous solutions by *Spirulina platensis* biomass. Arab J Chem 7:57–62
- Ali MB, Tripathi RD, Rai UN, Pal A, Siugh SP (1999) Physico-chemical characteristics and pollution level of Lake Nainital (UP, India): role of macrophytes and phytoplankton in biomonitoring and phytoremediation of toxic metal ions. Chemosphere 39:2171–2182
- Anawar HM, Ahmed G (2019) Combined electrochemical-advanced oxidation and enzymatic process for treatment of wastewater containing emerging organic contaminants. In: Mishra AK, Anawar HMD, Drouiche NBT-E, NC in W (eds) Emerging and nanomaterial contaminants in wastewater: advanced treatment technologies. Elsevier, pp 277–307
- Anirudhan TS, Sreekumari SS (2011) Adsorptive removal of heavy metal ions from industrial effluents using activated carbon derived from waste coconut buttons. J Environ Sci 23:1989–1998
- Antoniadis A, Takavakoglou V, Zalidis G et al (2010) Municipal wastewater treatment by sequential combination of photocatalytic oxidation with constructed wetlands. Catal Today 151:114– 118
- Aydin MI, Yuzer B, Hasancebi B, Selcuk H (2019) Application of electrodialysis membrane process to recovery sulfuric acid and wastewater in the chalcopyrite mining industry. Desalin Water Treat 172:206–211
- Ayoub GM, Semerjian L, Acra A et al (2001) Heavy metal removal by coagulation with seawater liquid bittern. J Environ Eng 127:196–207
- Ballan-Dufrançais C, Marcaillou C, Amiard-Triquet C (1991) Response of the phytoplanctonic alga *Tetraselmis suecica* to copper and silver exposure: vesicular metal bioaccumulation and lack of starch bodies. Biol Cell 72:103–112
- Barra L, Chandrasekaran R, Corato F, Brunet C (2014) The challenge of ecophysiological biodiversity for biotechnological applications of marine microalgae. Mar Drugs 12:1641–1675
- Bashir A, Malik LA, Ahad S et al (2019) Removal of heavy metal ions from aqueous system by ion-exchange and biosorption methods. Environ Chem Lett 17:729–754
- Bautista P, Mohedano AF, Casas JA et al (2008) An overview of the application of Fenton oxidation to industrial wastewaters treatment. J Chem Technol Biotechnol 83:1323–1338
- Becker W (2004) Microalgae in human and animal nutrition. In: Richmond A (ed) Handbook of microalgal culture. Blackwell, Oxford, UK, pp 312–351
- Beiras R (2018) Marine pollution: sources, fate and effects of pollutants in coastal ecosystems. Chapter 11. Vol. 5, 187–204
- Bergmann M, Gutow L, Klages M (2015) Marine anthropogenic litter. Springer Nature, p 447
- Bolto B, Dixon D, Eldridge R et al (2002) Removal of natural organic matter by ion exchange. Water Res 36:5057–5065
- Bonin DJ, Droop MR, Maestrini SY, Bonin MC (1986) Physiological features of six micro–algae to be used as indicators of seawater quality. Cryptogamie Algol 7:23–83
- Borbély G, Nagy E (2009) Removal of zinc and nickel ions by complexation-membrane filtration process from industrial wastewater. Desalination 240:218–226
- Borja A, Elliott M, Andersen JH et al (2013) Good environmental status of marine ecosystems: what is it and how do we know when we have attained it? Mar Pollut Bull 76:16–27

- Bretherton L, Williams A, Genzer J et al (2018) Physiological response of 10 phytoplankton species exposed to Macondo oil and the dispersant, Corexit. J Phycol 54:317–328
- Brillas E (2020) A review on the photoelectro-Fenton process as efficient electrochemical advanced oxidation for wastewater remediation. Treatment with UV light, sunlight, and coupling with conventional and other photo-assisted advanced technologies. Chemosphere 250:126198
- Brinza L, Dring MJ, Gavrilescu M (2007) Marine micro and macro algal species as biosorbents for heavy metals. Environ Eng Manag J 6:237–251
- Burakov AE, Galunin EV, Burakova IV et al (2018) Adsorption of heavy metals on conventional and nanostructured materials for wastewater treatment purposes: a review. Ecotoxicol Environ Saf 148:702–712
- Burridge E (2003) Bisphenol a: product profile. Eur Chem News April 14-20:17
- Cameron H, Mata MT, Riquelme C (2018) The effect of heavy metals on the viability of *Tetraselmis marina* AC16-MESO and an evaluation of the potential use of this microalga in bioremediation. PeerJ 25:e5295
- Carrera-Martinez D, Mateos-Sanz A, Lopez-Rodas V, Costas E (2011) Adaptation of microalgae to a gradient of continuous petroleum contamination. Aquat Toxicol 101:342–350
- Chan KY, Chiu SY (1985) The effects of diesel oil and oil dispersants on growth, photosynthesis, and respiration of *Chlorella salina*. Arch Environ Contam Toxicol 14:325–331
- Chao M, Shen X, Lun F, Shen A, Yuan Q (2012) Toxicity of fuel oil water accommodated fractions on two marine microalgae, *Skeletonema costatum* and Chlorella spp. Bull Environ Contam Toxicol 88:712–716
- Chaukura N, Gwenzi W, Tavengwa N, Manyuchi MM (2016) Biosorbents for the removal of synthetic organics and emerging pollutants: opportunities and challenges for developing countries. Environ Dev 19:84–89
- Chen G (2004) Electrochemical technologies in wastewater treatment. Sep Pur Tech 38:11-41
- Chen H, Jiang J-G (2011) Toxic effects of chemical pesticides (trichlorfon and dimehypo) on *Dunaliella salina*. Chemosphere 84:664–670
- Chen H, Zhang Z, Tian F et al (2018a) The effect of pH on the acute toxicity of phenanthrene in a marine microalgae *Chlorella salina*. Sci Rep 8:17577
- Chen Q, Yao Y, Li X et al (2018b) Comparison of heavy metal removals from aqueous solutions by chemical precipitation and characteristics of precipitates. J Water Process Eng 26:289–300
- Chen Y, Zhao X, Guan W et al (2017) Photoelectrocatalytic oxidation of metal-EDTA and recovery of metals by electrodeposition with a rotating cathode. Chem Eng J 324:74–82
- Cheng H, Xu W, Liu J et al (2007) Pretreatment of wastewater from triazine manufacturing by coagulation, electrolysis, and internal microelectrolysis. J Hazard Mater 146:385–392
- Chitrakar P, Baawain MS, Sana A, Al-Mamun A (2019) Current status of marine pollution and mitigation strategies in arid region: a detailed review. Ocean Sci J 54:317–348
- Chojnacka K, Chojnacki A, Górecka H (2005) Biosorption of Cr<sup>3+</sup>, Cd<sup>2+</sup> and Cu<sup>2+</sup> ions by bluegreen algae *Spirulina* sp.: kinetics, equilibrium and the mechanism of the process. Chemosphere 59:75–84
- Christaki E, Bonos E, Giannenas I, Florou-Paneri P (2013) Functional properties of carotenoids originating from algae. J Sci Food Agric 93:5–11
- Clark RM, Fronk CA, Lykins BW (1988) Removing organic contaminants from groundwater. Environ Sci Technol 22:1126–1130
- Cobbett C, Goldsbrough P (2002) Phytochelatins and metallothioneins: roles in heavy metal detoxification and homeostasis. Annu Rev Plant Biol 53:159–182
- Dabrowski A, Hubicki Z, Podkościelny P, Robens E (2004) Selective removal of the heavy metal ions from waters and industrial wastewaters by ion-exchange method. Chemosphere 56:91–106
- Danovaro R, Carugati L, Berzano M (2016) Implementing and innovating marine monitoring approaches for assessing marine environmental status. Front Mar Sci 3:213
- Das C, Ramaiah N, Pereira E, Naseera K (2018) Efficient bioremediation of tannery wastewater by monostrains and consortium of marine *Chlorella* sp. and *Phormidium* sp. Int J Phytoremediation 20:284–292

- Davis TA, Volesky B, Mucci A (2003) A review of the biochemistry of heavy metal biosorption by brown algae. Water Res 37:4311–4330
- DeLorenzo ME, Danese LE, Baird TD (2013) Influence of increasing temperature and salinity on herbicide toxicity in estuarine phytoplankton. Environ Toxicol 28:359–371
- DeLorenzo ME, Serrano L (2003) Individual and mixture toxicity of three pesticides; atrazine, chlorpyrifos, and chlorothalonil to the marine phytoplankton species *Dunaliella tertiolecta*. J Environ Sci Health B 38:529–538
- Dermont G, Bergeron M, Mercier G, Richer-Laflèche M (2008) Metal-contaminated soils: remediation practices and treatment technologies. Pract Period Hazardous, Toxic, Radioact Waste Manag 12:188–209
- Dermou E, Velissariou A, Xenos D, Vayenas DV (2007) Biological removal of hexavalent chromium in trickling filters operating with different filter media types. Desalination 211: 156–163
- Dsikowitzky L, Nordhaus I, Jenner-sikowitzky L et al (2011) Anthropogenic organic contaminants in water, sediments and benthic organisms of the mangrove—fringed Segara Anakan Lagoon, Java Indonesia. Mar Pollut Bull 62:851–862
- Duan W, Meng F, Lin Y, Wang G (2017) Toxicological effects of phenol on four marine microalgae. Environ Toxicol Pharmacol 52:170–176
- Dunstan WM, Atkinson LP, Natoli J (1975) Stimulation and inhibition of phytoplankton growth by low molecular weight hydrocarbons. Mar Biol 31:305–310
- El Zeftawy MAM, Mulligan CN (2011) Use of rhamnolipid to remove heavy metals from wastewater by micellar-enhanced ultrafiltration (MEUF). Sep Purif Technol 77:120–127
- Ertürk MD, Saçan MT (2012) First toxicity data of chlorophenols on marine alga *Dunaliella tertiolecta*: correlation of marine algal toxicity with hydrophobicity and interspecies toxicity relationships. Environ Toxicol Chem 31:1113–1120
- Fabregas J, Herrero C, Veiga M (1984) Effect of oil and dispersant on growth and chlorophyll a content of the marine microalga *Tetraselmis suecica*. Appl Environ Microbiol 47:445–447
- Flood S, Burkholder J, Cope G (2018) Assessment of atrazine toxicity to the estuarine phytoplankter, *Dunaliella tertiolecta* (Chlorophyta), under varying nutrient conditions. Environ Sci Pollut Res 25:11409–11423
- Folgar S, Torres E, Pérez-Rama M et al (2009) *Dunaliella salina* as marine microalga highly tolerant to but a poor remover of cadmium. J Hazard Mater 165:486–493
- Fučíková K et al (2014) New phylogenetic hypotheses for the core Chlorophyta based on chloroplast sequence data. Front Ecol Evol 2:63
- Gabriel OM, Rita O, Clifford A, Kennedy O (2006) Heavy metal pollution of fish of Qua-Iboe River Estuary: possible implications for neurotoxicity. Int J Toxicol 3:1–6
- Gad AAM, Abdalla AMA (2017) Fate of heavy metals and nutrients in waste stabilization ponds in arid zones. JES J Eng Sci 45:1–16
- Garba MD, Usman M, Mazumder MAJ et al (2019) Complexing agents for metal removal using ultrafiltration membranes: a review. Environ Chem Lett 17:1195–1208
- Gaur JP, Rai LC (2001) Heavy metal tolerance in algae. In: Rai LC, Gaur JP (eds) Algal adaptation to environmental stresses. Springer, Berlin Heidelberg, Berlin, Heidelberg, pp 363–388
- Ge Y, Ning Z, Wang Y et al (2016) Quantitative proteomic analysis of *Dunaliella salina* upon acute arsenate exposure. Chemosphere 145:112–118
- Gekeler W, Grill E, Winnnacker EL, Zenk MH (1988) Algae sequester heavy metals via synthesis of phytochelatin complexes. Arch Microbiol 150:197–202
- Gelcich S, Buckley P, Pinnegar JK et al (2014) Public awareness, concerns, and priorities about anthropogenic impacts on marine environments. Proc Natl Acad Sci 111:15042–15047
- González J, Figueiras FG, Aranguren-Gassis M et al (2009) Effect of a simulated oil spill on natural assemblages of marine phytoplankton enclosed in microcosms. Estuar Coast Shelf Sci 83:265–276
- Grip K (2017) International marine environmental governance: a review. Ambio 46:413–427

- Guedes AC, Amaro HM, Malcata FX (2011) Microalgae as sources of carotenoids. Mar Drugs 9: 625–644
- Gupta VK, Ali I, Saleh TA et al (2012) Chemical treatment technologies for waste-water recycling—an overview. RSC Adv 2:6380–6388
- Gupta VK, Jain R, Saleh TA et al (2011) Equilibrium and thermodynamic studies on the removal and recovery of safranine-T dye from industrial effluents. Sep Sci Technol 46:839–846
- Gurreri L, Tamburini A, Cipollina A, Micale G (2020) Electrodialysis applications in wastewater treatment for environmental protection and resources recovery: a systematic review on progress and perspectives. Membranes 10:1–93
- Han X, Wong YS, Tam NFY (2006) Surface complexation mechanism and modeling in Cr(III) biosorption by a microalgal isolate, *Chlorella miniata*. J Colloid Interface Sci 303:365–371
- Harrison PJ, Cochlan WP, Acreman JC, Parsons TR et al (1986) The effect of crude oil and Corexit 9527 on marine phytoplankton in an experimental enclosure. Mar Environ Res 18:93–109
- Hein M, Pedersen MF, Sand Jensen K (1995) Size-dependent nitrogen uptake in micro- and macroalgae. Mar Ecol Prog Ser 118:247-254
- Hing LS, Ford T, Finch P, Crane M, Morritt D (2011) Laboratory stimulation of oil–spill effects on marine phytoplankton. Aquat Toxicol 103:32–37
- Honarmandrad Z, Javid N, Malakootian M (2020) Efficiency of ozonation process with calcium peroxide in removing heavy metals (Pb, Cu, Zn, Ni, Cd) from aqueous solutions. SN Appl Sci 2: 703
- Hu L, Adeyiga AA, Miamee E (2002) Removal of organic chemicals from wastewater by surfactant separation. United States
- Huang Y, Yang X, Lan L, Zhan J, Luo H, Jiang L (2017) Studies on the diversity of marine microalgae and the strains with high polysaccharides, lipids and proteins along Zhanjiang coastal areas. Act Hydrobiol Sinica 41:1080–1090
- Ipek U (2005) Removal of Ni(II) and Zn(II) from an aqueous solution by reverse osmosis. Desalination 174:161–169
- Isawi H (2019) Evaluating the performance of different nano-enhanced ultrafiltration membranes for the removal of organic pollutants from wastewater. J Water Process Eng 31:100833
- Jasper JT, Nguyen MT, Jones ZL et al (2013) Unit process wetlands for removal of trace organic contaminants and pathogens from municipal wastewater effluents. Environ Eng Sci 30:421–436
- Jiang Y, Zhihong W, Xiurong H, Lei Z, Xiulin W (2002) Toxicity of polycyclic aromatic hydrocarbons (PAHs) to marine algae. Mar Sci 26:46–50
- Kabdaşli I, Arslan T, Arslan-Alaton I et al (2010) Organic matter and heavy metal removals from complexed metal plating effluent by the combined electrocoagulation/Fenton process. Water Sci Technol 61:2617–2624
- Kachel MJ (2008) Threats to the marine environment: pollution and physical damage. Particularly Sensitive Sea Areas: The IMO's Role in Protecting Vulnerable Marine Areas, 23–36
- Kaparapu J, Prasad MK (2018) Equilibrium, kinetics and thermodynamic studies of cadmium (II) biosorption on *Nannochloropsis oculata*. Appl Water Sci 8:1–9
- Karpińska J, Kotowska U (2019) Removal of organic pollution in the water environment. Water (Switzerland) 11
- Katam K, Shimizu T, Soda S, Bhattacharyya D (2020) Performance evaluation of two trickling filters removing LAS and caffeine from wastewater: light reactor (algal-bacterial consortium) vs dark reactor (bacterial consortium). Sci Total Environ 707:135987
- Khambhaty Y, Mody K, Basha S, Jha B (2009) Biosorption of Cr(VI) onto marine *Aspergillus niger*: experimental studies and pseudo-second order kinetics. World J Microbiol Biotechnol 25: 1413–1421
- Khoshmanesh A, Lawson F, Prince IG (1997) Cell surface area as a major parameter in the uptake of cadmium by unicellular green microalgae. Chem Eng J Biochem Eng J 65:13–19
- Kongsricharoern N, Polprasert C (1996) Chromium removal by a bipolar electro-chemical precipitation process. Water Sci Technol 34:109–116

- Kumar M, Singh AK, Sikandar M (2018) Study of sorption and desorption of cd (II) from aqueous solution using isolated green algae *Chlorella vulgaris*. Appl Water Sci 8:225
- Kumar M, Singh AK, Sikandar M (2020) Biosorption of hg (II) from aqueous solution using algal biomass: kinetics and isotherm studies. Heliyon 6:e03321
- Kuppusamy S, Maddela NR, Megharaj M, Venkateswarlu K (2020) Fate of total petroleum hydrocarbons in the environment. In: Total Petroleum Hydrocarbons. Springer, Cham, pp 57–77
- Landaburu-Aguirre J, Pongrácz E, Perämäki P, Keiski RL (2010) Micellar-enhanced ultrafiltration for the removal of cadmium and zinc: use of response surface methodology to improve understanding of process performance and optimisation. J Hazard Mater 180:524–534
- Leliaert F et al (2012) Phylogeny and molecular evolution of the green algae. CRC Crit Rev Plant Sci 31:1–46
- Lema SC, Schultz I, Scholz N, Incardona J, Swanson P (2007) Neural defects and cardiac arrhythmia in fish larvae following embryonic exposure to 2,29,4,49-tetrabromodiphenyl ether (PBDE–47). Aquat Toxicol 82:296–307
- Leong YK, Chang JS (2020) Bioremediation of heavy metals using microalgae: recent advances and mechanisms. Bioresour Technol 303:122886
- LeProvost I (2001) Environmental impact of the offshore oil and gas industry. J Environ Assess Policy Manage 3:173–175
- Levine HG (1984) The use of seaweeds for monitoring coastal waters. In: Shubert EL (ed) Algae as Ecological Indicators. Academic Press, London, pp 189–210
- Levy JL, Stauber JL, Jolley DF (2007) Sensitivity of marine microalgae to copper: the effect of biotic factors on copper adsorption and toxicity. Sci Total Environ 387:141–154
- Liao MY, Randtke SJ (1986) Predicting the removal of soluble organic contaminants by lime softening. Water Res 20:27–35
- Liu Y, Liu Y, Liu Y, Zhao X, Wang G (2020) Effect of water accommodated fractions of fuel oil on fixed carbon and nitrogen by microalgae: implication by stable isotope analysis. Ecotoxicol Environ Safe 195:110488
- Magro C, Mateus EP, Paz-Garcia JM, Ribeiro AB (2020) Emerging organic contaminants in wastewater: understanding electrochemical reactors for triclosan and its by-products degradation. Chemosphere 247:125758
- Magureanu M, Bradu C, Parvulescu VI (2018) Plasma processes for the treatment of water contaminated with harmful organic compounds. J Phys D Appl Phys 51:313002
- Marino MA, Brica RM, Neale CN (1997) Heavy metal soil remediation: the effects of attrition scrubbing on a wet gravity concentration process. Environ Prog 16:208–214
- Marques IM, Oliveira ACV, de Oliveira OMC, Sales EA, Moreira ÍTA (2021) A photobioreactor using *Nannochloropsis oculata* marine microalgae for removal of polycyclic aromatic hydrocarbons and sorption of metals in produced water. Chemosphere 281:130775
- Matis KA, Zouboulis AI, Gallios GP et al (2004) Application of flotation for the separation of metal-loaded zeolites. Chemosphere 55:65–72
- Matlock MM, Howerton BS, Atwood DA (2002) Chemical precipitation of heavy metals from acid mine drainage. Water Res 36:4757–4764
- Matsunaga T, Takeyama H, Nakao T, Yamazawa A (1999) Screening of marine microalgae for bioremediation of cadmium-polluted seawater. J Biotechnol 70:33–38
- Michalowicz J, Duda W (2007) Phenols-sources and toxicity. Pol J Environ Stud 16:347-362
- Mofeed JM, Abdel-Aal EI (2015) Effect of phenol on some antioxidant enzymes in effect of phenol on some antioxidant enzymes in the marine microalga *Dunaliella salina*. J Environ Sci 44:185– 196
- Mohammady NGED, Chen YC, Mohammad RF (2005) Physiological responses of the eustigmatophycean *Nannochloropsis salina* to aqueous diesel fuel pollution. Oceanologia 47: 75–92
- Molinari R, Poerio T, Argurio P (2008) Selective separation of copper(II) and nickel(II) from aqueous media using the complexation-ultrafiltration process. Chemosphere 70:341–348

- Nakayama T et al (1998) The basal position of scaly green flagellates among the green algae (Chlorophyta) is revealed by analyses of nuclear-encoded SSU rRNA sequences. Protist 149: 367–380
- National Research Council (US) Committee on Oil in the Sea (2003) Oil in the Sea III: Inputs, Fates, and Effects. Washington, DC: National Academies Press
- Nayaka S, Toppo K, Verma S (2017) Adaptation in algae to environmental stress and ecological conditions. In: Shukla V, Kumar S, Kumar N (eds) Plant adaptation strategies in changing environment. Springer Singapore, Singapore, pp 103–115
- Nelms SE, Coombes C, Foster LC, Galloway TS, Godley BJ, Lindeque PK, Witt MJ (2017) Marine anthropogenic litter on British beaches: a 10-year nationwide assessment using citizen science data. Sci Total Environ 579:399–1409
- Nessim RB, Bassiouny AR, Zaki HR et al (2011) Biosorption of lead and cadmium using marine algae. Chem Ecol 27:579–594
- Nguyen MK, Tran VS, Pham TT et al (2021) Fenton/ozone-based oxidation and coagulation processes for removing metals (Cu, Ni)-EDTA from plating wastewater. J Water Process Eng 39:101836
- Oliver BG, Cosgrove EG (1974) The efficiency of heavy metal removal by a conventional activated sludge treatment plant. Water Res 8:869–874
- Özverdi A, Erdem M (2006) Cu<sup>2+</sup>, Cd<sup>2+</sup> and Pb<sup>2+</sup> adsorption from aqueous solutions by pyrite and synthetic iron sulphide. J Hazard Mater 137:626–632
- Page DS, Boehm PD, Douglas GS et al (1999) Pyrogenic polycyclic aromatic hydrocarbons in sediments record past human activity: a case study in Prince William sound, Alaska. Mar Pollut Bull 38:247–260
- Pagnanelli F, Mainelli S, Bornoroni L et al (2009) Mechanisms of heavy-metal removal by activated sludge. Chemosphere 75:1028–1034
- Papazi K, Kotzabasis AK, Kotzabasis K (2012) Bioenergetic strategy for the biodegradation of pcresol by the unicellular green alga Scenedesmus obliquus. PLoS One 7:e51852
- Perales-Vela HV, Peña-Castro JM, Cañizares-Villanueva RO (2006) Heavy metal detoxification in eukaryotic microalgae. Chemosphere 64:1–10
- Pérez M, Torrades F, García-Hortal JA et al (2002) Removal of organic contaminants in paper pulp treatment effluents under Fenton and photo-Fenton conditions. Appl Catal B Environ 36:63–74
- Pérez-Rama M, Torres E, Suárez C, Herrero C, Abalde J (2010) Sorption isotherm studies of Cd (II) ions using living cells of the marine microalga *Tetraselmis suecica* (Kylin) Butch. J Environ Manage 91:2045–2050
- Polprasert C, Charnpratheep K (1989) Heavy metal removal in attached-growth waste stabilization ponds. Water Res 23:625–631
- Qaderi F, Sayahzadeh AH, Azizpour F, Vosughi P (2019) Efficiency modeling of serial stabilization ponds in treatment of phenolic wastewater by response surface methodology. Int J Environ Sci Technol 16:4193–4202
- Qin J, Rosen BP, Zhang Y et al (2006) Arsenic detoxification and evolution of trimethylarsine gas by a microbial arsenite S-adenosylmethionine methyltransferase. Proc Natl Acad Sci U S A 103: 2075–2080
- Ramírez Calderón OA, Abdeldayem OM, Pugazhendhi A, Rene ER (2020) Current updates and perspectives of biosorption technology: an alternative for the removal of heavy metals from wastewater. Curr Pollut Rep 6:8–27
- Randtke SJ (1988) Organic contaminant removal by coagulation and related process combinations. J Am Water Works Assoc 80:40–56
- Ravindra K, Sokhi R, Van Grieken R (2008) Atmospheric polycyclic aromatic hydrocarbons: source attribution, emission factors and regulation. Atmos Environ 42:2895–2921
- Rodrigues Pires da Silva J, Merçon F, Guimarães Costa CM, Radoman Benjo D (2016) Application of reverse osmosis process associated with EDTA complexation for nickel and copper removal from wastewater. Desalin Water Treat 57:19466–19474
- Romera E, González F, Ballester A et al (2007) Comparative study of biosorption of heavy metals using different types of algae. Bioresour Technol 98:3344–3353
- Romero-Lopez J, Lopez-Rodas V, Costas E (2012) Estimating the capability of microalgae to physiological acclimatization and genetic adaptation to petroleum and diesel oil contamination. Aquat Toxicol 124–125:227–237
- Sakhi D, Rakhila Y, Elmchaouri A et al (2019) Optimization of coagulation flocculation process for the removal of heavy metals from real textile wastewater. In: Ezziyyani M (ed) Advances in intelligent systems and computing. Springer International Publishing, Cham, pp 257–266
- Saldarriaga-Hernandez S, Hernandez-Vargas G, Iqbal HM, Barcelo D, Parra-Saldívar R (2020) Bioremediation potential of *Sargassum* sp. biomass to tackle pollution in coastal ecosystems: circular economy approach. Sci Total Environ 715:136978
- Salinas-Whittaker S, Gómez-Gutiérrez CM, Cordero-Esquivel B, Luque PA, Guerra-Rivas G (2020) Effects of the water-soluble fraction of the mixture fuel oil/diesel on the microalgae *Dunaliella tertiolecta* through growth. Environ Sci Pollut Res 27:35148–35160
- Salmani MH, Davoodi M, Ehrampoush MH et al (2013) Removal of cadmium (II) from simulated wastewater by ion flotation technique. Iran J Environ Health Sci Eng 10:16
- Sathasivam R, Ki JS (2019) Differential transcriptional responses of carotenoid biosynthesis genes in the marine green alga *Tetraselmis suecica* exposed to redox and non-redox active metals. Mol Biol Rep 46:1167–1179
- ter Schure AFH, Larsen P, Agrell C (2004) Atmospheric transport of polybrominated diphenyl ethers and polybrominated biphenyls to the Baltic Sea. Environ Sci Technol 38:1282–1287
- Sharma KK, Schuhmann H, Schenk PM (2012) High lipid induction in microalgae for biodiesel production. Energies 5:1532–1553
- Shen LC, Nguyen XT, Hankins NP (2015) Removal of heavy metal ions from dilute aqueous solutions by polymer-surfactant aggregates: a novel effluent treatment process. Sep Purif Technol 152:101–107
- Siron R, Giusti G, Berland B, Morales-Loo R, Pelletier E (1991) Water-soluble petroleum compounds: chemical aspects and effects on the growth of microalgae. Sci Total Environ 104:211–227
- Siyanytsya V, Kochkodan V, Goncharuk V (2008) Natural organic matter removal from water by complexation-ultrafiltration. Desalination 223:91–96
- Song Y, Sun T, Cang L et al (2019) Migration and transformation of cu(II)-EDTA during electrodialysis accompanied by an electrochemical process with different compartment designs. Electrochim Acta 295:605–614
- Stahl W, Sies H (2003) Antioxidant activity of carotenoids. Mol Asp Med 24:345-351
- Stelzenmüller V, Coll M, Mazaris AD, Giakoumi S et al (2018) A risk-based approach to cumulative effect assessments for marine management. Sci Total Environ 612:1132–1140
- Subashchandrabose SR, Ramakrishnan B, Megharaj M, Venkateswarlu K, Naidu R (2013) Mixotrophic cyanobacteria and microalgae as distinctive biological agents for organic pollutant degradation. Environ Int 51:59–72
- Sun L, Miznikov E, Wang L, Adin A (2009) Nickel removal from wastewater by electroflocculation-filtration hybridization. Desalination 249:832–836
- Sunda W, Kieber DJ, Kiene RP, Huntsman S (2002) An antioxidant function for DMSP and DMS in marine algae. Nature 418:317–320
- Surkatti R, Al-Zuhair S (2018) Effect of cresols treatment by microalgae on the cells composition. J Water Process Eng 26:250–256
- Taha HM, Said HA, Abbas NH, Khaleafa AFM (2009) Biosorption and biodegradation of the antifouling compound tributyltin (TBT) by microalgae. Amer Eur J Sci Res 4:1–6
- Thakkar M, Randhawa V, Wei L (2013) Comparative responses of two species of marine phytoplankton to metolachlor exposure. Aquat Toxicol 126:198–206
- Torres MA, Barros MP, Campos SC, Pinto E, Rajamani S, Sayre RT, Colepicolo P (2008) Biochemical biomarkers in algae and marine pollution: a review. Ecotoxicol Environ Safe 71: 1–15

- Tragin M, Vaulot D (2018) Green microalgae in marine coastal waters: the ocean sampling day (OSD) dataset. Sci Rep 8:1–12
- Tran TK, Chiu KF, Lin CY, Leu HJ (2017) Electrochemical treatment of wastewater: selectivity of the heavy metals removal process. Int J Hydrog Energy 42:27741–27748
- Trivunac K, Stevanovic S (2006) Removal of heavy metal ions from water by complexationassisted ultrafiltration. Chemosphere 64:486–491
- USEPA (2014) Toxic and Priority Pollutants under the Clean Water Act. (p. Appendix A to Part 423–126 Priority Pollutants). p. Appendix A to Part 423–126 Priority Pollutants. https://www.epa.gov/eg/toxic-and-priority-pollutants-under-cleanwater-act (accessed: January 7, 2020)
- Van Gestel CA, Van Brummelen TC (1996) Incorporation of the biomarker concept in ecotoxicology calls for a redefinition of terms. Ecotoxicology 5:217–225
- Vidal RRL, Moraes JS (2019) Removal of organic pollutants from wastewater using chitosan: a literature review. Int J Environ Sci Technol 16:1741–1754
- Volterra L, Conti ME (2000) Algae as biomarkers, bioaccumulators and toxin producers. Int J Environ Pollut 13:92–125
- Wang J, Chen C (2009) Biosorbents for heavy metals removal and their future. Biotechnol Adv 27: 195–226
- Wang TC, Weissman JC, Ramesh G et al (1998) Heavy metal binding and removal by *Phormidium*. Bull Environ Contam Toxicol 60:739–744
- Wang Y, Zhang C, Zheng Y, Ge Y (2017) Phytochelatin synthesis in *Dunaliella salina* induced by arsenite and arsenate under various phosphate regimes. Ecotoxicol Environ Safe 136:150–160
- Wei C, Wang S, Hu X et al (2021) Exposure to dibutyl phthalate induced the growth inhibition and oxidative injury in *Dunaliella salina*. IOP Conference Series: Earth and Environmental Science 804:042038
- Whitton BA, Kelly MG (1995) Use of algae and other plants for monitoring rivers. Aust J Ecol 20: 45–56
- Wingenfelder U, Hansen C, Furrer G, Schulin R (2005) Removal of heavy metals from mine waters by natural zeolites. Environ Sci Technol 39:4606–4613
- Wu Y, Zhang J, Mi TZ, Li B (2001) Occurrence of n-alkanes and polycyclic aromatic hydrocarbons in the core sediments of the Yellow Sea. Mar Chem 76:1–15
- Xia Z, Hu L (2018) Treatment of organics contaminated wastewater by ozone micro-nano-bubbles. Water (Switzerland):11
- Yangali-Quintanilla V, Maeng SK, Fujioka T et al (2011) Nanofiltration vs. reverse osmosis for the removal of emerging organic contaminants in water reuse. Desalin Water Treat 34:50–56
- Zeitoun MM, Mehana EE (2014) Impact of water pollution with heavy metals on fish health: overview and updates. Global Vet 12:219–231
- Zhao Y, Wang Y, Li Y, Santschi PH, Quigg A (2017) Response of photosynthesis and the antioxidant defense system of two microalgal species (*Alexandrium minutum* and *Dunaliella salina*) to the toxicity of BDE–47. Mar Pollut Bull 124:459–469
- Zhu YH, Jiang JG (2009) Combined toxic effects of typical mutagens–dimethylphenol, tribromethane and dinitroaniline, on unicellular green algae *Dunaliella salina*. J Food Safe 29:1–13
- Zia Z, Hartland A, Mucalo MR (2020) Use of low-cost biopolymers and biopolymeric composite systems for heavy metal removal from water. Int J Environ Sci Technol 17:4389–4406
- Ziolko D, Hala D, Lester JN, Scrimshaw MD (2009) The effectiveness of conventional trickling filter treatment plants at reducing concentrations of copper in wastewaters. Sci Total Environ 407:6235–6241

# Chapter 4 Algal-Bacterial Consortiums, from Fundamental Interactions to Environmental Applications



#### Ignacio de Godos Crespo, Alfonso García Álvaro, César Ruíz Palomar, Félix Gaspar Gonzalo Ibrahim, and Raúl Muñoz Torre

**Abstract** Microalgae cultures offers solutions for pollution control at water reuse. The ability to nutrient and heavy metal up-take along with disinfection is a considerable more sustainable solution than conventional wastewater treatment processes based on mechanical aeration. Since microalgae have evolved together with bacteria in natural aquatic environments a whole range of interactions take place between both organisms. Ecology descriptions and population characterizations of the phycosphere are key elements for the design of microalgae-based units addressed to pollution control and for the understanding of the biochemical transformations that take place during cultivation. Negative interactions occur as consequence of resources competence or parasitism. Mutualism have the basis on substrate exchange and release of promotors. These and other interactions take place during the environmental applications such as wastewater treatment, gas treatment, or water reuse affecting the final performance.

Keywords Algal-bacterial processes  $\cdot$  Photosynthetic biodegradation  $\cdot$  Ecological interactions  $\cdot$  Bioremediation  $\cdot$  Biomass

#### R. M. Torre

I. de Godos Crespo ( $\boxtimes$ ) · A. G. Álvaro · C. R. Palomar · F. G. G. Ibrahim Department of Chemical Engineering and Environmental Technology, School of Industrial Engineering, University of Valladolid, Valladolid, Spain

School of Forestry, Agronomic and Bioenergy Industry Engineering (EiFAB), University of Valladolid, Soria, Spain

University of Valladolid, Institute for Sustainable Processes, Valladolid, Spain e-mail: ignacio.godos@uva.es

Department of Chemical Engineering and Environmental Technology, School of Industrial Engineering, University of Valladolid, Valladolid, Spain

University of Valladolid, Institute for Sustainable Processes, Valladolid, Spain

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_4

## 4.1 Microalgae Biomass Culture

Microalgae cultures are a highly productive pathway to convert solar energy and inorganic substrates (CO<sub>2</sub>, water and ions) in high value biomass and oxygen. The process of algae production and processing has been done for millennia in different parts of the globe. For instance, the cyanobacteria Nostoc was produced in China and Arthrospira in Chad and Mexico (Hamed 2018). Presently, microalgae have several applications apart of human and animal nutrition: cosmetics and the production of high value molecules (essential fatty acids, pigments, stable isotope biochemicals) (Pulz and Gross 2004). Proteins, lipids, sugars, minerals and vitamins and other biocomponents are contained in microalgae biomass. For decades, commercialization of Haematococcus is taking place in Japan and Israel while Chlorella is produced in Germany and Portugal. The ability to accumulate lipids of algae biomass has been extensively explored as source of biodiesel for decades, although implementation at industrial scale has not been materialized (Grima et al. 2003). Carbon dioxide fixation with intense algae cultivation has been researched using flue gas and other polluted gas streams (de Godos et al. 2014a). A summary of the microalgae culture applications is depicted in Fig. 4.1.



Fig. 4.1 Applications microalgae biomass

Microalgae production process can be summarized in chemical reaction as follows:

$$CO_2 + 0.148 NO_3 + 0.014 H_2 SO_4 + 0.012 H_2 PO_4 + 0.751 H_2 0$$
  

$$\rightarrow CH_{1.715}O_{0.427}N_{0.148}S_{0.014}P_{0.012} + 1.437 O_2$$
(4.1)

Therefore, carbon, nitrogen, and other elements are assimilated from culture broth and while oxygen is released along with the generation of biomass. This primary metabolism of microalgae is highly efficient in wastewater treatment applications (de Godos et al. 2017). The oxygen released by polycultures of different species of algae provides biological oxidation of organic matter while the high rates nitrogen and phosphorous assimilation results in high quality effluents. This ability to remove pollution from water has been proposed 60 years ago when wastewater treatment systems began to be implemented in western countries (Oswald and Ramani 1978). However, bacteria based systems consisted in aeration tanks such as activated sludge have been much more successful solution with millions of installations worldwide. Recent concern about sustainability have attracted interest toward systems based in carbon and nutrient recovery. In this sense, microalgae based systems offers assimilation of key elements (nitrogen, phosphorus) and neutral or even negative greenhouse gases emissions. Since the process is driven by sunlight, electricity consumption during the process is considerably lower than aeration systems. Beside this, the possible synergies between environmental applications and other algae uses, such energy production, carbon dioxide capture or bioproducts is attracting more interest around this technology (Murry et al. 2019).

Wrongly termed microalgae cultures are rather a co-culture of microalgae and bacteria in most of cases (except for axenic cultures of algae performed in conditions of microbiological isolation). The interactions between algae and bacteria could affect positively or negatively the biomass production and biochemical transformations that take place. A wider range of culture conditions can be found since microalgae grow in illuminated water bodies and wet surfaces all over the world. As consequence of the photosynthesis, microalgae create a particular environment with high concentrations of dissolved oxygen and elevated pH, particularly during central hours of the day when the light intensity reach its maximum (Arbib et al. 2017a). Organic materials are released as consequence of physiologic activity and algae cells decay. This particular environment presents appropriate conditions for the development of a wide range of organisms. In this sense, bacteria, which exhibits an extensive variety of metabolic routes from photo and chemoautotrophic to heterotrophic growth using different organic substrates, grow and interact with microalgae in natural and artificial environments (Rosenberg 2013). These interactions are still not well studied from a biotechnological point of view. At this point it must be notice that only a few thousand microalgae strains are kept in collections, only few hundred are researched for their possible application and just a handful are cultivated in industrial quantities, while more than 50.000 species are believed to exist. In case of environmental applications such as wastewater treatment, the knowledge about these interactions is even more limited since cultures are not usually studied form the microbiological point of view (Ferrero et al. 2012). Microbiology characterizations and ecology studies have been barely explored. Recent progress in molecular identifications and the interdisciplinary approach of the research projects have shed light on the importance and impact of the symbiotic bacteria in the microalgae growth (Barreiro-Vescovo et al. 2021). This chapter reviews the ecological interactions between algae and bacteria and their importance in microalgae environmental applications.

#### 4.2 Evolution of Microalgae and Bacteria

Half of the global net primary productivity is produced by microalgae in water bodies, with oceans as the major habitat of these organisms (Parlevliet and Moheimani 2014). These photosynthetic organisms beside with cyanobacteria grow in the illuminated region of the aquatic habitat and receive the name of phytoplankton. Bacterioplankton is the term applied to bacteria living in the same habitat. The metabolic activity of both communities impacts the global geocycles of oxygen, carbon, nitrogen, phosphorous, and other elements. The very intimate interactions between both groups of microorganisms have shaped the actual atmosphere and geosphere composition. Three thousand eight hundred million years ago, when the atmosphere was anoxic and mainly formed by N<sub>2</sub>, CO<sub>2</sub>, NH<sub>3</sub>, and CH<sub>4</sub>, simple living organisms have appeared (Sleep 2010). Prokaryotes existing in hydrothermal vents using chemoautotroph reactions are believed to be the first living organisms according to the reconstruction of the ancestral sequences. Photoautotrophic organisms arose 3500 million year ago, first without oxygen production and later with reactions that produce oxygen and final by-product. Photoxygenic organisms transformed the atmosphere increasing the levels of O<sub>2</sub> until the actual values of 20,9% (v/v) (Rasmussen et al. 2008). Cyanobacteria were responsible for this oxygenation environment and later to the formation of the photosynthetic eukaryotes, such microalgae and plants, through the process named primary endosymbiosis. This hypothesis established that heterotrophic eukaryotic organisms engulfed cyanobacterium and engaged it as an organelle. Consequently, eukaryote organisms acquired photosynthetic metabolism (Yoon et al. 2004). Although bacteria were proposed as the early host cell which received the cyanobacterium, most recent studies are suggesting that an archaea-type organism was responsible for this evolution event. Phylogenic studies indicate that three different lineages appear as a consequence of endosymbiosis: red algae, green algae, and glaucophytes (Ramanan et al. 2016). Subsequent endosymbiosis events, involving bacteria, archaea, protest, and cyanobacteria, resulted in a diversification of the phototrophic organism. More recent studies indicate that apart from the endosymbiotic events, horizontal gene transfer for each group of organism took and take place and facilitate the adaptation of an organism to the different environments (Schönknecht et al. 2013).

## 4.3 Ecological Interactions Between Organisms

Algae are the primary producers while bacteria present decomposing and producing activity depending on the substrate and light availability. Therefore most of the ecological interactions can be found between both organisms and most of them are present in the planktonic habitat (Ramanan et al. 2016). Occurrence of these interactions during microalgae culture in environmental applications affects positively when microalgae biomass production is enhanced resulting in high rates of pollutant up-take. However, negative interactions can negatively impact algae growth rate limiting the treatment capacity. At this point it must be stressed that microalgae growth rate (and biomass productivity) is mainly impacted by external factors such as temperature, light intensity, reactor configuration, and hydrodynamic factors (Fig. 4.2). Since bacteria are simultaneously co-cultured with the microalgae, and also affected by these external variables, microbial interactions can often go unnoticed or their effects may be attributed to the external variables. In this context, the improvement of culture techniques must take into consideration all the possible interactions: mutualism, parasitism, commensalism, and others.

Mutualisms interactions, where both parts take advantage of the coexistence, have been described between microalgae and bacteria in natural environments or lab-controlled experiments. These interactions can be facultative, obligate, or



Fig. 4.2 Diagram of the main factors affecting microalgae cultures



Fig. 4.3 Interactions between microalgae and bacteria

opportunistic. The main mutualistic interaction is the aforementioned gas exchange where microalgae can provide the oxygen that heterotrophic bacteria require for the breakdown of the organic materials, while bacteria concomitantly release the carbon dioxide, nitrogen, and phosphorus needed by microalgae during photosynthesis (Fig. 4.3). This is of important relevance in eutrophic environments with high concentrations of organic matter and it has been extensively used for the treatment of polluted water streams such as domestic, industrial, or livestock wastewaters. However, the mutualistic interactions are not limited to the inorganic components exchange. Exchange of organic compounds has been described in both directions. Vitamin B12 is produced by some bacteria and assimilated by microalgae. In some cases, microalgae even repress the expression of genes associated to Vitamin B12 synthesis when symbiotic bacteria provide this supplement (Croft et al. 2005). Mutualistic interactions have been described between green algae and bacteria that are also responsible for the very well-known growth promotion of plants. That is the case of Rhyzombium, Mesorhizobium and Azospirillum, which enhance microalgae growth and the other way around (Luz et al. 2004). Although some cyanobacteria are capable of atmospheric nitrogen fixation, green algae do not present this metabolic route. Rhyzobium provides organic nitrogen in oligotrophic ecosystems where this element is limiting the primary production. Besides this, this interaction is not limited to nitrogen since carbon can be also exchanged concomitantly. Organic compounds are supplied to the bacteria (Rhyzobium or others) while inorganic carbon as CO<sub>2</sub> or HCO<sub>3</sub><sup>-</sup> is supplied in return. These bacteria promote algae growth by the release of compounds such as indol acetic acid. It is worth noting that similar interactions have been also described between multicellular algae (Dao et al. 2018).

Negative interactions have also been described between microalgae and bacteria (parasitism interaction). Algicidal activity of some bacteria, for instance genera *Alteromonas*, *Pseudomonas*, *Bacillus*, *Dietzia*, *Janibacter*, has attracted the attention of its potential use as a controlling agent of algal or cyanobacteria blooms, also named Harmful algal blooms (Coyne et al. 2022; Croci et al. 2006). These natural phenomena occur across the world and considerably affect the aquaculture production systems placed in coastal waters. The occurrence of algal blooms has increased over the last decades likely due to two main factors: increasing pollution of water bodies and global warming. Species of dinoflagellates cause severe physiological and biochemical damage to fish. Algae toxins are accumulated by bivalves and human intoxication has been described after mollusk consumption. Worldwide, these algae toxins are responsible for more than 60,000 cases of intoxication per year, and the human mortality rate reaches a value of 1.5%.

Occurrence of bacteria with the capacity to kill microalgae during harmful blooms is being studied and proposed as a control system. Some bacteria species present physiological and biochemical mechanisms to attack microalgae. These mechanisms are similar to the existing in bacteria attacking higher plants. Chitinases, glucosidases, cellulases, and other enzymes found in the disruption of plant cell walls have been reported in phycosphere bacteria. It is worth mentioning that this parasitic interaction and similar enzymatic mechanism has been documented in other organisms such as marine fungi and mollusk (Nikolaeva et al. 1999). Algae cell decay results in the release of intracellular materials that are used as nutrients by bacteria and fungi. Consequently, competition for existing nutrients with microalgae results in reduced growth rates of phototrophs and outcompeting their existence in the environment. Enzymes present in these organisms (bacteria, mollusk, and fungi) are useful for many applications in algal and industrial biotechnology (Dahiya et al. 2006). Development of these industrial applications is based in the knowledge of the interactions and the study of the ecology of species succession, parasitism, and competition. Parasitic bacteria can easily be found attached to the algal cell wall or in the associated mats contributing degradation of the cell wall. Some marine bacteria living in association with microalgae are able to produce substances that block the sodium channels in algae resulting in algae decay. In the case of microalgae cultures used for environmental purposes, predatory events have been described on several occasions but the organisms involved are rather protozoa or animals than bacteria (Martínez et al. 2021).

Negative interactions between microorganisms are also based on competition. Substrate competition between microalgae and bacteria has been described in natural environments and bioreactors. While heterotrophic bacteria create a mutual interaction based on exchange of inorganic (CO<sub>2</sub>,  $NH_4^+$ ,  $PO_4^{3-}$ ) and organic substrates, autotrophic bacteria could be characterized by competitive interactions. In this sense, ammonia-oxidizing bacteria use the same substrates than microalgae, inorganic carbon, and ammonia, for their primary metabolism. As consequence, the

coexistence of nitrifiers and microalgae is hampered (de Godos et al. 2014b). In case of natural environments, a wide range of conditions can be found and the presence of both groups of microorganisms has been reported. In the case of artificial environments, such as photobioreactors treating wastewater, conditions are created for maximizing microalgae productivity. For instance, the design of photobioreactors provides elevated irradiance inside the culture media, creating an environment similar to surface of water bodies. That is the case of high-rate algae ponds treating wastewater of different natures: domestic, agro-industrial, or livestock. Limitation of ammonia-oxidizing bacteria has been reported under these conditions (de Godos et al. 2010). It is important to highlight that this competitive phenomenon impacts the performance of wastewater treatment facilities positively or negatively depending on the bioprocess configuration. In this sense, most of the photobioreactors treating wastewater are designed for a maximum ammonia assimilation by microalgae (Arbib et al. 2017b). Although microalgae can use nitrate, nitrite, and ammonia as nitrogen sources, the last one is assimilated in first place since oxidized forms involved more energy investment by cells. Therefore, ammonia oxidation can result in the accumulation of  $NO_2^-$  and  $NO_3^-$  and consequently effluents will not achieve the discharge limits (De Godos et al. 2016). However, other reactor configurations based on nitrification and denitrification rely on high rates of nitrification (de Godos et al. 2014b). In that case, sufficient inorganic carbon must be present in the influent.

# 4.4 Environmental Applications of the Microalgae-Bacteria Consortia

Effluents resulting from various human activities such as farming, households and industrial production contains pollutants that can potentially endanger ecosystems and reduce possibilities of water reuse. If not treated properly this leads to water ecosystem destruction and involved human health risks (UN-Water 2021). Conventional water treatment are focused in organic matter, phosphorus and nitrogen removal (Jenkins 2014). Besides this, organic and inorganic substances including important amounts of elements such as sulfur, arsenic, chlorine, magnesium, calcium, and metals could be present in wastewater (Metcalf and Eddy 2003). Pathogenic organisms present in wastewater include a wide variety of microorganisms, including protozoa, viruses, and bacteria (Chambonniere et al. 2021), which are the origin of waterborne diseases like cholera, hepatitis, typhoid, tuberculosis, and dysentery. These diseases cause more than one million deaths yearly (UN-Water 2021). Both organic and inorganic components of polluted effluents, such as nitrogen and phosphorous, and carbonaceous compounds, are assimilated into microalgae biomass and thus, the concentrations of these compounds are reduced reaching the discharge limits (Barreiro-Vescovo et al. 2020). The main advantage of using microalgae-bacteria consortia instead of only bacteria systems is energy and equipment saving involved in oxygen supply. Besides this, microalgae utilize any nitrogen source present in the wastewater: organic molecules, ammonium, nitrite, and nitrates (Taylor et al. 2012). Unlike, conventional water treatment facilities based on activated sludge, microalgae assimilate the nitrogen allowing for reutilization. Moreover, algae treatment units comprise primary and secondary treatment in one basin (normally a shallow pond mixed). Therefore, no transition between multiple operational conditions is required in order to remove inorganic nitrogen and phosphorous, simplifying the facilities and reducing the energy consumption of the treatment due to pumps and water elevation (de Godos et al. 2017).

Microalgae-based treatment is not limited to organic matter and nutrient removal. In fact, heavy metals and emerging contaminants like pharmaceutical and personal care products are efficiently removed during algae cultivation (López-Serna et al., 2022; de Godos et al. 2010). Heavy metals present a high binding affinity with the cell wall of microorganisms (including algae and bacteria) (Hwang et al., 2016). Metals that are usually found in domestic wastewater: include mercury, copper, zinc, nickel, arsenic, lead, cadmium, chromium, and manganese (Metcalf and Eddy 2003). Presence of these elements can be caused by oxidative damage in microorganisms cells by the stimulation of production of reactive oxygen species inside cells (Shahid et al., 2020). Experiments conducted by Shahid et al. (2020) demonstrate that microalgae exposed to high concentrations of copper presented higher levels of lipid peroxidation as a consequence of reactive oxygen species formation and a decrease in the level of pigments such as chlorophyll-a, chlorophyllb, and carotenoids. Metal and heavy metal elimination mechanisms taking place during algae-bacteria cultures include: absorption, detoxification by the low-weight proteins called metallothioneins which induce complexation by chelates and polysaccharides and alkaline precipitation due to high pH values. Active and passive mechanisms have an effect on metal ions uptake by cells. The passive uptake or biosorption consists in the intermolecular binding that takes place when the cellular structure entraps heavy metal ions at binding sites (Kumar et al., 2015). Metals ions are bonded to sulfate, carboxyl, and phosphate groups existing in cell polysaccharides, On the other hand, during active uptake, heavy metals are included in metabolic cell pathways. While active sorption is limited to living cells, passive removal includes the elimination of non-living biomass. At this point it must be stressed that dead biomass (of algae and bacteria) can account for an important proportion of total biomass existing in wastewater treatment process). During the first stage, metal cations undergo adsorption onto the cell surface and the active transportation of metal species inside the cells takes place in the second phase. Transportation is a complex and irreversible process that requires energy consumption. Active process of metal uptake involves the following steps: transport across the cell membrane, formation of complexes, cation exchange, physical adsorption, and precipitation metabolism of the cell. During the cation exchange process, metal ions that are bounded to the cell surface can be replaced with another cation present in solution of similar charge. Complexation involved the formation of secondary compounds formed as a result of the linkage between organic molecules and metal ions acting as ligands. Electrostatic interactions and covalent bonds stabilize these

complexes. It is worth mentioning that the precipitation of metals occurs in both active (dependent of metabolism) and passive processes.

Disinfection or pathogen removal is a key process in water reutilization. In this sense, algae systems provide a tertiary treatment since pathogenic bacteria face harsh conditions inside cultures (Neil 2014). Microalgae cultivation is normally carried out in shallow ponds specifically designed to support a high algal productivity by optimizing sunlight exposition of the culture broth. However, elevated biomass concentration increases light attenuation and reduces the significance of the light decay and inactivation mechanisms of bacteria a virus. Nevertheless, the negative impact of light reduction may be counteracted by the intense mixing, provided by paddle wheels or similar, that enables intermittent light exposure of moving cells toward the surface (Chambonniere et al. 2021). In addition, the high microalgae photosynthesis rates increase the magnitude of diurnal pH peaks (values higher than 10) and dissolved oxygen concentration, which can easily double the saturation levels. These conditions enhance sunlight-mediated pathogen deaths. Presence of toxic algae metabolites has been suggested; however, limited information is available in this topic.

## 4.5 Communities in the Phycosphere

Associated bacteria and other organisms growing in environments dominated by microalgae is defined as phycosphere. In some cases, specific microbiota has been described in cultures treating effluents or cultures devoted to biomass production. Other studies have found wider distributed organisms community found in the association. At this point, it must be stressed that techniques employed in microbial population determination finally impacts the amount of organism detected. In this sense, first studies based on characterization by selective media cultivation introduce a significant bias in population determination (Ferrero et al. 2012). Molecular tools techniques developed at the end of the last century have emerged as the fundamental basis of microbial determination and therefore microbial ecology research. These techniques have provided new insights that have helped improve the understanding of biological processes and consequently the design and operation of bioreactors (Manoylov 2014). In contrast to the high biodiversity observed in bacterial communities associated with rhizospheres of higher plants, phycosphere is characterized by a significantly less diverse microbial community. Bacterial communities are dominated by the phyla: Bacteroidetes and Proteobacterial, classes alpha, beta, and epsilon. Archea species have not been detected in the reported studies (Sapp et al. 2008). Important metabolic features impacting the treatment performance have been detected: chemoautotrophs involved in organic matter oxidation, chemoautotrophs responsible for ammonia or nitrate oxidation, predatory organisms that could impact the concentration of pathogen. Recently the characterization followed by metatranscriptome analysis, study of the genes activated, have indicated activity in genes that are involved in the interaction of bacteria with the microalgae. These genes are related to infection-related secretion pathways, biosynthesis of exoenzymes, modifying important metabolic pathways such as the synthesis of carbohydrates, lipids, and flagella.

#### 4.6 Conclusions

Human activities deeply impact the Earth's equilibrium between organisms and geosphere, atmosphere and hydrosphere. Alteration of biogeocyles as a consequence of resource exploitation has been reported for many of the elements of the periodic table. In this scenario application of nature-based solutions with assimilatory capacity of key elements present in polluted effluents is becoming a necessity. Microalgae cultures present the ability to up-take some of the elements which present elevated concentrations in polluted wastewaters, such as nitrogen and phosphorous, but also more dangerous elements and compounds such as heavy metals and emerging pollutants. Since microalgae and bacteria have undergone a parallel evolution that has shaped the inferences between them, positive and negative, culture of microalgae with environmental purposes must be an account with a solid knowledge of the possible interactions and the consequences for the stability. In this sense population characterizations and ecological analysis should be considered in technical operations and scientific research.

# References

- Arbib Z, de Godos Crespo I, Corona EL, Rogalla F (2017a) Understanding the biological activity of high rate algae ponds through the calculation of oxygen balances. Appl Microbiol Biotechnol 101:12. https://doi.org/10.1007/s00253-017-8235-3
- Arbib Z, de Godos I, Ruiz J, Perales JA (2017b) Optimization of pilot high rate algal ponds for simultaneous nutrient removal and lipids production. Sci Total Environ 589. https://doi.org/10. 1016/j.scitotenv.2017.02.206
- Barreiro-Vescovo S, González-Fernández C, Ballesteros M, de Godos I (2020) Activity determination of an algal-bacterial consortium developed during wastewater treatment based on oxygen evolution. J Water Process Eng 36. https://doi.org/10.1016/j.jwpe.2020.101278
- Barreiro-Vescovo S, González-Fernández C, de Godos I (2021) Characterization of communities in a microalgae-bacteria system treating domestic wastewater reveals dominance of phototrophic and pigmented bacteria. Algal Res 59(March). https://doi.org/10.1016/j.algal.2021.102447
- Chambonniere P, Bronlund J, Guieysse B (2021) Pathogen removal in high-rate algae pond: state of the art and opportunities. J Appl Phycol 33(3):1501–1511. https://doi.org/10.1007/s10811-020-02354-3
- Coyne KJ, Wang Y, Johnson G (2022) Algicidal bacteria: a review of current knowledge and applications to control harmful algal blooms. Front Microbiol 13(April):1–23. https://doi.org/ 10.3389/fmicb.2022.871177
- Croci L, Cozzi L, Suffredini E, Benzi M, and Romagna EE (2006) Characterization of microalgae and associated bacteria collected from shellfish harvesting areas Characterization of microalgae

and associated bacteria collected from shellfish harvesting areas. no. November 2017, doi: https://doi.org/10.1016/j.hal.2005.08.001

- Croft MT, Lawrence AD, Raux-Deery E, Warren MJ, Smith AG (2005) Algae acquire vitamin B12 through a symbiotic relationship with bacteria. Nature 438(7064):90–93. https://doi.org/10. 1038/nature04056
- Dahiya N, Tewari R, Hoondal GS (2006) Biotechnological aspects of chitinolytic enzymes: a review. Appl Microbiol Biotechnol 71(6):773–782. https://doi.org/10.1007/s00253-005-0183-7
- Dao G, Wu G, Wang X, Zhang T, Zhan X (2018) Enhanced microalgae growth through stimulated secretion of indole acetic acid by symbiotic bacteria. Algal Res 33(June):345–351. https://doi. org/10.1016/j.algal.2018.06.006
- de Godos I, Arbib Z, Lara E, Cano R (2017) Wastewater treatment in algal systems. Innovative wastewater treatment & resource recovery technologies: impacts on energy, economy and environment. IWA Publishing
- De Godos I, Arbib Z, Lara E, Rogalla F (2016) Evaluation of high rate algae ponds for treatment of anaerobically digested wastewater : effect of CO 2 addition and modification of dilution rate Madrid , Spain. Bioresour Technol. https://doi.org/10.1016/j.biortech.2016.08.056
- Ferrero EM, De Godos I, Rodríguez EM, García-encina PA (2012) Molecular characterization of bacterial communities in algal—bacterial photobioreactors treating piggery wastewaters. 40: 121–130. https://doi.org/10.1016/j.ecoleng.2011.10.001
- de Godos I et al (2010) A comparative evaluation of microalgae for the degradation of piggery wastewater under photosynthetic oxygenation. Bioresour Technol 101(14):5150–5158. https:// doi.org/10.1016/j.biortech.2010.02.010
- de Godos I et al (2014a) Evaluation of carbon dioxide mass transfer in raceway reactors for microalgae culture using flue gases. Bioresour Technol 153. https://doi.org/10.1016/j.biortech. 2013.11.087
- de Godos I et al (2014b) Assessing carbon and nitrogen removal in a novel anoxic-aerobic cyanobacterial-bacterial photobioreactor configuration with enhanced biomass sedimentation. Water Res 61. https://doi.org/10.1016/j.watres.2014.04.050
- Grima EM, Acie FG, Medina AR, Chisti Y (2003) Recovery of microalgal biomass and metabolites : process options and economics. 20:491–515
- Hamed I (2018) The evolution and versatility of microalgal biotechnology : a review," no. January,doi: https://doi.org/10.1111/1541-4337.12227
- Hwang JH, Church J, Lee SJ, Park J, Lee WH (2016) Use of microalgae for advanced wastewater treatment and sustainable bioenergy generation. Environ Eng Sci 33(11):882–897. https://doi. org/10.1089/ees.2016.0132
- Jenkins D (2014) Activated sludge—100 years and counting. Water Intell Online 13. https://doi. org/10.2166/9781780404943
- Kumar KS, Hans-uwe D, Eun-ji W, Jae-seong L, Kyung-hoon S (2015) Ecotoxicology and environmental safety microalgae – a promising tool for heavy metal remediation. Ecotoxicol Environ Saf 113:329–352. https://doi.org/10.1016/j.ecoenv.2014.12.019
- López-Serna R, Bolado S, Irusta R, Jiménez JJ (2022) Determination of veterinary drugs in microalgae biomass from photobioreactors fed with piggery wastewater. Chemosphere 287. https://doi.org/10.1016/j.chemosphere.2021.132076
- Luz E, Hernandez J, Morey T, Bashan Y (2004) Microalgae growth-promoting bacteria as " helpers " for microalgae : a novel approach for removing ammonium and phosphorus from municipal wastewater. 38:466–474. https://doi.org/10.1016/j.watres.2003.09.022
- Manoylov KM (2014) Review taxonomic identification of algae (morphological and molecular): species concepts, methodologies, and their implications for ecological. 424:409–424. https://doi.org/10.1111/jpy.12183
- Martínez C, Pessi BA, Bernard O (2021) Dynamics and productivity of microalgae in presence of predators. IFAC-Papers OnLine 54(3):673–678. https://doi.org/10.1016/j.ifacol.2021.08.319
- Metcalf I, Eddy (2003) Wastewater engineering : treatment and reuse. Fourth edition/revised by George Tchobanoglous, Franklin L. Burton, H. David Stensel. McGraw-Hill, Boston. ©2003

- Murry MA, Murinda SE, Huang S, Mark A (2019) Bioconversion of agricultural wastes from the livestock industry for biofuel and feed production. Elsevier Inc.
- Neil A and B. B. V Sc (2014) "Comparing the performance of a High Rate Algal Pond with a Waste Stabilisation Pond in rural South Australia"
- Nikolaeva EV, Usov AI, Sinitsyn AP, Tambiev AH (1999) Degradation of agarophytic red algal cell wall components by new crude enzyme preparations. J Appl Phycol 11(4):385–389. https:// doi.org/10.1023/A:1008170722877
- Oswald WJ and Ramani RA (1978) An integrated high rate pond-algae harvesting system by Dodd JC and Anderson JC, pp. 713–726. S. H. B. T.-E. I. C. on W. P. R. JENKINS, Ed. Pergamon, pp. C-55-C–59
- Parlevliet D, Moheimani NR (2014) Efficient conversion of solar energy to biomass and electricity. Aquat Biosyst 10(1):1–9. https://doi.org/10.1186/2046-9063-10-4
- Pulz O, Gross W (2004) Valuable products from biotechnology of microalgae. Appl Microbiol Biotechnol 65(6):635–648. https://doi.org/10.1007/s00253-004-1647-x
- Ramanan R, Kim B, Cho D, Oh H, Kim H (2016) Algae—bacteria interactions : evolution , ecology and emerging applications. Biotechnol Adv 34(1):14–29. https://doi.org/10.1016/j.biotechadv. 2015.12.003
- Rasmussen B, Fletcher IR, Brocks JJ, Kilburn MR (2008) Reassessing the first appearance of eukaryotes and cyanobacteria. Nature 455(7216):1101–1104. https://doi.org/10.1038/ nature07381
- Rosenberg E (2013) The prokaryotes: alphaproteobacteria and betaproteobacteria. In: Rosenberg E, DeLong EF, Lory S, Stackebrandt E, Thompson F (eds). Springer Heidelberg, Berlin. https://doi.org/10.1007/978-3-642-30197-1
- Sapp M, Gerdts ÆG, and Wellinger ÆM (2008) Consuming algal products : trophic interactions of bacteria and a diatom species determined by RNA stable isotope probing. pp. 283–287, https:// doi.org/10.1007/s10152-008-0110-2
- Schönknecht G et al (2013) Gene transfer from bacteria and archaea facilitated evolution of an extremophilic eukaryote. Science (80- ) 339(6124):1207–1210. https://doi.org/10.1126/science. 1231707
- Shahid M, Xu Z, Shahid Z, Nawaz M (2020) Cultivating microalgae in wastewater for biomass production, pollutant removal, and atmospheric carbon mitigation; a review. Sci. Total Environ. 704:135303. https://doi.org/10.1016/j.scitotenv.2019.135303
- Sleep NH (2010) The Hadean-Archaean Environment. pp. 1-15
- Taylor P, Ruiz J, Arbib Z, and Barragán J (2012) Photobiotreatment model (PhBT): a kinetic model for microalgae biomass growth and nutrient removal in wastewater. no. October, pp. 37–41, https://doi.org/10.1080/09593330.2012.724451
- UN-Water (2021) "Summary Progress Update 2021 : SDG 6—water and sanitation for all," UN-Water Integr. Monit. Initiat., pp. 1–58, [Online]. https://www.unwater.org/new-data-onglobal-progress-towards-ensuring-water-and-sanitation-for-all-by-2030/
- Yoon HS, Hackett JD, Ciniglia C, Pinto G, Bhattacharya D (2004) A molecular timeline for the origin of photosynthetic eukaryotes. Mol Biol Evol 21(5):809–818. https://doi.org/10.1093/ molbev/msh075

# **Chapter 5 Biodegradation of Environmental Pollutants by Marine Yeasts**



#### Danouche Mohammed, El Arroussi Hicham, and El Ghachtouli Naima

**Abstract** Organic contaminants are among the main pollutants of ecosystems because of their presence in domestic, agricultural, or industrial effluents. Indeed, many organic xenobiotics such as aromatic hydrocarbons, pesticides, synthetic dyes, etc., are not easily biodegradable in the environment and can therefore accumulate in ecosystems causing various toxic symptoms in exposed organisms, including humans. Yeast-assisted biological treatment has emerged as a promising new strategy for the biodegradation of such hazardous contaminants. Firstly, this chapter provides an overview of the applications of yeast in the biodegradation of organic contaminants. Subsequently, synthetic dyes were chosen as a model of organic pollutants to highlight the enzymes involved in their biodegradation process using various yeast strains. Indeed, the main oxidases involved are laccase, tyrosinase, lignin peroxidase, and manganese peroxidase. While the main reductases are Azoreductase, NADH-DCIP reductase, and malachite green reductase. The last section highlights the effects of physicochemical conditions on the effectiveness of mycoremediation.

E. A. Hicham

AgroBioScience (AgBS), Mohammed VI Polytechnic University (UM6P), Ben Guerir, Morocco

E. G. Naima

Microbial Biotechnology and Bioactive Molecules Laboratory, Sciences and Technologies Faculty, Sidi Mohamed Ben Abdellah University, Fez, Morocco

D. Mohammed (🖂)

Green Biotechnology Center, Moroccan Foundation for Advanced Science, Innovation and Research (MAScIR), Rabat, Morocco

Microbial Biotechnology and Bioactive Molecules Laboratory, Sciences and Technologies Faculty, Sidi Mohamed Ben Abdellah University, Fez, Morocco

Departement of Chemical & Biochemical Sciences-Green Process Engineering (CBS-GPE), Mohammed VI Polytechnic University (UM6P), Benguerir, Morocco e-mail: mohammed.danouche@usmba.ac.ma

Green Biotechnology Center, Moroccan Foundation for Advanced Science, Innovation and Research (MAScIR), Rabat, Morocco

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023 T. Encarnação, A. Canelas Pais (eds.), *Marine Organisms: A Solution to Environmental Pollution?*, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_5

Keywords Yeast · Pollutants · Biodegradation · Oxidases · Reductases

#### 5.1 Introduction

Yeasts are a group of polyphyletic fungi composed of basidiomycetes and ascomycetes species that have the particularity of living in a single-celled state. The environmental role that yeasts play is identical to that of other fungi. In fact, they act as saprophytes that transform plant and animal organic matter into biomass and its by-products (Kutty and Philip 2008). These ecological properties have been exploited by man since antiquity and continue to be developed even in the present day in a range of applications such as the fermentation processes (Beer, Wine, Sake, Soy Sauce), in food and feed ingredients (enzymes, flavors, pigments, amino acids, organic acids), the biocatalysis (pharmaceuticals, chiral chemical intermediates, and biotransformation), the biocontrol (food and feed safety, crop protection, and probiotics), as well as in fundamental research in biology and biomedical (Molecular biology, pathway engineering, systems biology mechanisms, drug metabolism and resistance, etc.) (Johnson and Echavarri-erasun 2011).

In addition to these applications, the exploitation of this ecological principle of yeasts in the biodegradation of different organic materials has prompted scientists to evaluate their capacity in the biodegradation of different carbon-based xenobiotics such as organic solvents, humic substances, phenolic compounds, petroleum, surfactants, pesticides, pharmaceuticals, and dyes, etc. (Aksu 2005). Numerous studies have reported the ability of yeast to biodegrade a variety of hazardous contaminants, including aromatic hydrocarbons (Deeba et al. 2018), phenol compounds (Filipowicz et al. 2020), pesticides (Han et al. 2019; Isia et al. 2019), fungicides (Kucharska et al. 2020), insecticide (Chen et al. 2012) and herbicides (States and States 2011) and synthetic dyes (Danouche et al. 2021c, 2022).

In this chapter, we first reviewed existing studies dealing with the topic of biodegradation of organic pollutants using halotolerant yeast strains. Then, we approached a comprehensive analysis of the enzymatic process involved by various yeast strains in the biodegradation of synthetic dyes as a model of organic pollution. Finally, we discussed the involvement of physicochemical factors in enhancing the mycoremediation capacity of yeast species.

#### 5.2 Biodegradation of Organic Pollutants by Yeast

Biodegradation process is defined as an energy-dependent mechanism by which organic substances are decomposed into simpler and smaller by-products through the action of various enzymes (Kaushik and Malik 2009). This bioprocess is called mineralization when the products of the biodegradation are more straightforward elements, such as  $H_2O$ ,  $CO_2$ ,  $NH_3$ ,  $CH_4$ ,  $H_2S$ , or  $PO_3$ . This same process is defined

as biotransformation when the organic compounds are not completely mineralized (Danouche et al. 2021b). Numerous halophilic microorganisms belonging to bacteria, fungi, and microalgae have shown the ability to decompose a wide variety of organic hazardous substances under high salt conditions (Castillo-carvajal et al. 2014). In recent years, the biotechnological application of fungi (mycoremediation) has become a model example for bioremoval of organic contaminants. It has been reported that various species of fungal could be used for the biodegradation of organic chemicals including aromatic and aliphatic hydrocarbons, industrial dyes, and other organic contaminants, released into the aquatic environment from various industrial and agricultural sectors (Aleu and Collado 2009; Sen et al. 2016). Despite the advantages of yeast strains over other species of the fungal kingdom, including rapid growth, high plasticity, and the ability to adapt to adverse growth conditions (Jafari et al. 2014; Sen et al. 2016), only a few studies have been conducted on the use of halotolerant yeast strains for the biodegradation of organic pollutants.

Aliphatic and Aromatic Hydrocarbons The yeast species described as hydrocarbon degraders are mostly from the genera of *Yarrowia, Candida, Pichia, Debaryomyces, Sporidiobolus, Metschnikowia, Lodderomyces, Rhodosporidium, Leucosporidium, Rhodotorula, Stephanoascus, Sporobolomyces, Trichosporon, and Cryptococcus* (Csutak et al. 2010; Kumari and Abraham 2011; Jain and Bajpai 2012; Gargouri et al. 2015; Deeba et al. 2018). Recently, Hashem et al. (2018) reported that other yeast strains of *Meyerozyma guilliermondii KKUY-0214, Yamadazyma mexicana KKUY-0160, R. taiwanensis KKUY-0162, P. kluyveri KKUY- 0163, R. ingeniosa KKUY-0170, and C. pseudointermedia KKUY-0192 were approved for their ability to degrade both aromatic and aliphatic hydrocarbon.* 

**Phenolic Compounds** Pollution with phenolic compounds can occur in the soil as well as in water bodies, due to their presence in discharges from industrial, agricultural, or domestic activities (Anku et al. 2017). Species of the genus *Candida* are documented as yeast strains with the highest capacity to decompose a diverse range of phenolic compounds. For example, a yeast strain of *C. rugopelliculosa* was reported to be able to decompose various phenolic compounds such as phenol, bisphenol A, nonylphenol, 4-methylphenol, 4-ethylphenol, 4-tert-butylphenol, 4-tert-OP, 4-tert-, and isooctane (Huang et al. 2017). The phenol was also reported to be degraded by other strains of *C. tropicalis* (Gong et al. 2021), *C. tropicalis* PHB5 (Basak et al. 2019), *C. subhashii* A011, *C. oregonensis* B021, *Schizoblastosporion starkeyi-henricii* L012 (Filipowicz et al. 2020), and *R. kratochvilovae* HIMPA1 (Patel et al. 2017).

**Pesticides, Fungicides, Insecticide and Herbicides** Various agricultural practices lead to the release of organic contaminants into the soil and surface or ground water. Selected yeast strains can be used as biodegradation agents for these xenobiotics. The pesticides like diazinon or pendimethalin can be biodegraded with *Saccharomyces cerevisiae* (Ehrampoush et al. 2017) or *Clavispora lusitaniae* (Han et al. 2019) respectively. The biodegradation of fungicides such as propiconazole was illustrated through the use of yeast strains of *Aureobasidium pullulans, Rhodotorula glutinis, and Cryptococcus* sp. (Kucharska et al. 2020). Regarding the insecticide,

Chen et al. (2012) showed that *C. pelliculosa* was efficient in the biodegradation of Bifenthrin. Also, *C. xestobii* was documented to have a high biodegradation capacity of Metolachlor and Alachlor herbicides (States and States 2011).

**Synthetic Dyes** Biodegradation of synthetic dyes by yeast has also been documented in the literature using different yeast strains (Danouche et al. 2021b). The most commonly used yeast for the degradation of synthetic dyes are strains belonging to the phylum Ascomycetes, such as *Saccharomyces, Candida,* and *Pichia* species. While only a few studies have involved basidiomycetous yeast strains, namely *Trichosporon* and *Pseudozyma* (Pajot et al. 2014). In the following section, we will focus on synthetic dyes as a model of organic pollutants because of their different chemical characteristics that make them resistant to biodegradation in natural ecosystems, as well as because of their toxicity toward exposed organisms, including humans (Danouche et al. 2021a). In the remainder of this chapter, an in-depth review of the enzymatic mechanisms involved in the biodegradation of these chemicals by yeast is presented in detail.

# 5.3 Yeast Enzymes Implications in the Biodegradation of Synthetic Dyes

Synthetic dyes can be degraded enzymatically by yeast cells in either the extracellular or in the intracellular compartment. The most studied enzymes for the biodegradation of dyes by yeast are the oxidases, which are the class of enzymes that use oxygen ( $O_2$ ) as an electron acceptor to catalyze the redox reaction, generating H<sub>2</sub>O or H<sub>2</sub>O<sub>2</sub> as products. They contain a metal or a Flavin-type coenzyme on the active site (Phale et al. 2019). In addition, it has been shown that some reductases are also involved by some yeast strains. The reductases indicated for the biodegradation of synthetic dyes are Azoreductase, NADH-DCIP reductase, and Malachite green reductase (Danouche et al. 2021b).

Laccase (Lac: EC 1.10.3.2) Synthetic dyes biocatalysis with Lac can be achieved by direct biodegradation of the dye molecule by a nonspecific radical mechanism. This enzymatic pathway has the advantage of avoiding the formation of toxic by-products, such as aromatic amines, which are usually obtained as a result of specific cleavage of the azo bond of various dyes via reductases or chemical catalytic processes (Dave et al. 2015). Several studies have documented the involvement of Lac in the biodegradation of the synthetic dye by yeast strains of *Sterigmatomyces halophilus* SSA-1575 (Al-Tohamy et al. 2020), *Galactomyces geotrichum* GG (Guo et al. 2019), *Cyberlindnera fabianii* (Danouche et al. 2021c), *T. akiyoshidainum* HP2023 (Martorell et al. 2017a), *T. multisporum*, and *T. laibachii* (Pajot et al. 2007).

*Tyrosinase* (*Tyr: E.C. 1.14.18.1*): referred also to as monophenol monooxygenase or polyphenol oxidase. It is an oxidase with copper coenzyme, which can be employed for the detoxification of wastewater containing phenol or other organic pollutants (Kim and Uyama 2005). The catalytic reaction of the dyes

with Tyr occurs in two successive steps, a first catalyzing reaction is the o-hydroxylation of the monophenols to the corresponding catechols (monophenolase activity), next a second oxidation of monophenols to the corresponding o-quinones (diphenolase activity) (Duckworth and Coleman 1970). On their involvement in the biodegradation of synthetic dyes by yeast cells, it has been identified in only a few yeast species. (Danouche et al. 2021c) reported their involvement in the biodegradation of the azo dye Acid Red 14 with a yeast strain of C. fabianii, in addition to other yeast strains including G. geotrichum MTCC (Waghmode et al. 2012a, b), S. cerevisiae MTCC 463 (Jadhav et al. 2007), C. krusei strains (Charumathi and Das 2011), Candida sp. MM 4035, T. porosum MM 4037, C. satwnus MM 4034, Barnettozyma californica MM 4018 (Martorell et al. 2012).

Lignin Peroxidase (LiP: EC 1.11.1.14) LiP is an extracellular enzyme whose enzymatic substrate is nonspecific, this particularity confers it the capacity to degrade various aromatic phenolic and non-phenolic compounds (Chowdhary et al. 2018). For example, the biodegradation of sulfonated azo dye with LiP can be accomplished in two consecutive one-electron oxidations of the oxidized LiP forms by  $H_2O_2$  in the phenolic ring, where the corresponding carbonium ion bearing the azo bond contributes to the formation of quinone and phenyldiazine by nucleophilic attack by  $H_2O$ . The phenyldiazine product is then oxidized by  $O_2$  to a phenyl radical and the azo bond is removed as N<sub>2</sub>, and then the phenyl radical extracts hydrogen from its surroundings to produce a stable aromatic compound (Chivukula et al. 1995). The catalytic activity of LiP was investigated during the biodegradation of various synthetic dyes by basidiomycota yeast strains of T. laibachii and T. multisporum (Pajot et al. 2007), as well as, by ascomycota yeast strains of S. halophilus SSA-1575 (Al-Tohamy et al. 2020), G. geotrichum (Guo et al. 2019), P. occidentalis (Song et al. 2018a), S. cerevisiae (Jadhav et al. 2007), C. krusei (Charumathi and Das 2011), Diutina rugosa (Bankole et al. 2017), and C. samutprakarnensis (Song et al. 2018b).

**Manganese Peroxidase (EC 1.11.1.13)** MnP is a substrate-specific oxidase that oxidizes  $Mn^{2+}$  to  $Mn^{3+}$  from the surface of the enzyme and subsequently oxidizes phenolic substrates such as model lignin compounds or other organic contaminants (Zhou et al. 2013). Yeast used MnP for the biodegradation of synthetic dyes as well, it was revealed in yeast strains of *C. fabianii* (Danouche et al. 2021c), *P. occidentalis* (Song et al. 2018a), *D. polymorphus, C. tropicalis* (Yang et al. 2008), *T. multisporum*, and *T. laibachii* (Pajot et al. 2007).

Azoreductase (AzoR: EC 1.7.1.6): are a class of enzymes that catalyze the reduction reaction such as the reduction of azo bonds (-N=N-) of azo dyes and nitroaromatic and azoic drugs (Misal and Gawai 2018). The AzoR can be classified according to their structures, or according to flavin dependence. The flavin-dependent class of AzoR can also be divided based on their coenzymes like NADH, NADPH (Saratale et al. 2011; Solís et al. 2012). The involvement of AzoR in dye biodegradation by yeast has been reported in some research employing

yeast strains of *Issatchenkia occidentalis* (Ramalho et al. 2004), *C. fabianii* (Danouche et al. 2021c), *C. krusei* (Charumathi and Das 2011), *S. cerevisiae* MTCC 463 (Jadhav et al. 2007), and *T. beigelii* NCIM-3326 (Saratale et al. 2009a).

**NADH-Preferring 2,6-Dichloroindophenol Reductase** (*NADH-DCIP: EC* **1.6.99.3**) NADH-DCIP reductase is an oxidoreductase that reduces 2,6-dichloroindo-phenol (DCIP) with NADH as an electron donor (Nishiya and Yamamoto 2007). Some studies have shown an increase in the NADH-DCIP reductase activity during the biodegradation of various azo dyes with strains of *P. occidentalis* G<sub>1</sub> (Song et al. 2018a), *C. samutprakarnensis* (Song et al. 2018b), *D. rugosa* (Bankole et al. 2017), *P. kudriavzevii* CR-Y103 (Rosu et al. 2018), and *T. beigelii* NCIM-3326 (Saratale et al. 2009a).

**Malachite Green Reductase** NADH is used as an electron donor by this reductase to transform malachite green into leucomalachite green. It has been first reported by Jadhav and Govindwar (2006) that in *S. cerevisiae* MTCC 463 was used for the biodegradation of green malachite. Next, Jadhav et al. (2008b) demonstrated their implication in the biodegradation of methyl red by *G. geotrichum* MTCC 1360. Also, Charumathi and Das (2011) reported the increase of MG-reductase activity in *C. krusei* used for the biodegradation of Basic Violet 3, as well as, in *S. cerevisiae* used for the biodegradation of Malachite green (Biradar et al. 2016).

Regardless of the mechanisms involved, the performance of the microorganisms in the bioremediation of organic pollutants can be influenced by various environmental factors, including nutrients, pH, temperature, etc. It is therefore critical to emphasize the impact of these parameters on the yeast's ability to eliminate such contaminants.

#### 5.4 Factors Controlling Mycoremediation Performance

Yeast cells are sensitive to the environmental conditions in which they grow. Determining the impact of these physiochemical factors on the efficiency of the removal of synthetic dyes is therefore crucial in order to make the mycoremediation process faster, more efficient, and more practical for large-scale applications. There are two ways to perform this optimization, either using a single-factor optimization approach or based on statistical methods of optimization by the design of the experiment (Gönen and Aksu 2009; Mahmoud 2016).

**Carbon and Nitrogen Sources** The effect of carbon and nitrogen sources on the bioaccumulation or the biodegradation capacity of dye by yeast strains has been the subject of several studies. It has been reported that at constant sucrose content, the concentrations of both Remazol Black B and Remazol Blue dyes inhibited the growth of *C. tropicalis*, with constant dye concentration, the growth efficiency and the bioaccumulation capacity increased with sucrose concentration up to 15 g L<sup>-1</sup> (Aksu and Dönmez 2005). This combined effect was also analyzed using

a statistical approach of response surface methodology. Okur et al. (2014) found that the optimal values for dye uptake by C. tropicalis correspond to sugar concentration of 5.1 g  $L^{-1}$  and a dye concentration of 499 mg  $L^{-1}$  of initial dye concentration. Also, Gönen and Aksu (2009) found that the optimum combination predicted by response surface methodology (RSM) confirmed that C. utilis was able to bioaccumulate Remazol turquoise blue-G with a maximum uptake yield of 82.0% in 15 g  $L^{-1}$  sucrose and 50 mg  $L^{-1}$  dye concentration. Additionally, (Das et al. 2010) confirmed that P. fermentans MTCC 189 was able to accumulate Basic violet 3 up to 69.8% in 10 mg  $L^{-1}$  of dye-containing medium and 24 g  $L^{-1}$  sugar extracted from sugarcane bagasse through RSM analysis. Concerning the carbon sources effect on the biodegradation efficiency of dye by yeast cells, it has been reported that the addition of carbon sources, especially glucose at a certain level, stimulates the biodegradation of dyes (Chang et al. 2000; Waghmode et al. 2011). Indeed, glucose is an essential element in several mechanisms, it provides an energy source for yeast growth, it regenerates the redox mediators NADH and FADH, and also it serves as a substrate for the production of  $H_2O_2$ , which acts in turn as a co-substrate for MnP and LiP (Swamy and Ramsay 1999; Jafari et al. 2014). In the same way, the supplementation of the medium with nitrogen sources such as peptone, yeast extract, urea, or others favors the regeneration of NADH which is used as an electron donor for the reduction of azo dyes by the different enzymes (Bras et al. 2001).

**Dye Concentration** The initial dye concentration can significantly influence their removal efficiency. A higher dye concentration gradually decreases the percentage of their decolorization. This can be attributed to the toxicity of the dyes toward the yeast cells, or to a lower biomass production making the decolorization operation inefficient (Saratale et al. 2011). It has been reported by Das et al. (2010) that the bioaccumulation of Acid Blue 93, Direct Red 28, and Basic Violet 3 by *P. fermentans* MTCC 189 decreased as the initial concentration of these dyes increased from 10 to 30 mg L<sup>-1</sup>. Likewise, Dönmez (2002) and Aksu (2003) described that the increase in the initial dye concentration inhibited the growth and caused a long lag period of *S. cerevisiae and C. tropicalis*. Other studies indicate that the presence of dyes at high concentrations may inhibit azoreductase activity during enzymatic biodegradation of the synthetic dye, due to the binding of dye molecules to the active site of enzymes (Jadhav et al. 2008a; Saratale et al. 2009a).

**Temperature** Temperature is one of the crucial factors that influence the metabolic pathways, the enzymatic activity, and the physicochemical interaction of dye molecules with the cell wall. According to the available literature, no study has been devoted to the question of the effect of temperature on the bioaccumulation of dyes using yeast cells. Therefore, it is very important to consider this factor as a research question for future studies. Regarding the effect of temperature on the ability of yeast to biodegrade dyes, many studies have been conducted on the activation energy of the involved enzymes (Chequer et al. 2013; Miranda et al. 2013). The performance of decolorization increased with increasing temperature to the optimum temperature, then a reduction in activity occurs at higher temperatures (Tan et al. 2013, 2014, 2016). The decrease in the ability of yeast strains to remove the dye molecules can be

attributed to the denaturation of the enzymes involved or to the resulting loss of cell viability (Saratale et al. 2009b).

pH pH of solutions is one of the most influential factors on the ability of yeast cells to bioremediate various organic pollutants, including synthetic dyes, it can modify the physicochemical properties of the dye molecules, as well as the physicochemical properties of the yeast surface where the cell-dye molecule interaction initially occurs (Fu and Viraraghavan 2002). At low pH, the surface of yeast cells becomes protonated with a positive charge, which promotes the binding of anionic dyes. On the other hand, at higher pH values, the cell surface acquires a negative charge, resulting in the electrostatic attraction of cationic dyes (Charumathi and Das 2012). The pH may have an inhibitory effect on the transport of dye molecules across the cell membrane, which is considered the first stage of the intracellular biodegradation or bioaccumulation process (Khan et al. 2013). In addition, the initial pH can affect the physiology of the yeast cells and the enzymatic activity. Most yeast strains show better decolorization efficiency under neutral or acidic conditions. The optimal value for the bioaccumulation of Remazol Blue, Reactive Red, and Reactive Black by the yeast strain C. tropicalis (Dönmez and Aksu 2002) as well as for Remazol Red RB, Remazol Black B, and Remazol Blue by S. Cerevisiae (Aksu 2003) was observed was 3.0. Likewise, Das et al. (2010) investigated that the maximum bioaccumulation rate of Basic Violet3, Direct Red 28, and Acid Blue 93 by growing cells of P. fermentans MTCC 189 was at pH 5.0.

**Shaking** Agitation allows a uniform distribution of oxygen and nutrients in the medium. It also facilitates the exchange of gases produced during the fermentation process of dyes by yeast cells (Yu and Wen 2005; Pajot et al. 2007; Yang et al. 2008; Martorell et al. 2017b). Differing opinions have been expressed regarding the effect of oxygen on the decolorization process, with some researchers considering that during the dye reduction reaction by the reductase, oxygen behaves as a stronger electron acceptor than the dye molecule, thus preventing the azo dye reduction reaction (Kalyani et al. 2008). On the other hand, other research indicates that oxidizing enzymes, such as laccases, require oxygen to oxidize aromatic molecules, including dyes (Thurston 1994). Therefore, it is necessary to optimize the level of agitation that regulates the concentration of dissolved oxygen in the medium throughout the yeast decolorization process to ensure an effective treatment.

#### 5.5 Conclusions

On the basis of the research discussed in this chapter, it is appropriate to consider mycoremediation as a cost-effective, eco-friendly, and efficient approach for the biodegradation of organic contaminants. However, an efficient mycoremediation process requires the optimization of physicochemical conditions, notably the initial concentration of the pollutant, the supply of nitrogen or carbon sources, as well as growth conditions such as pH, temperature, and agitation. On the other hand, the effectiveness of this kind of bio-processes should not be limited to emphasizing the degradation of the considered pollutant molecule; instead, it requires characterization and evaluation of the toxicity of the obtained by-products, since they can have more harmful effects than the original molecule. Lastly, we strongly recommend the application of this emerging biotechnology in wastewater treatment at pilot or large scale in order to prove its potential application.

#### References

- Aksu Z (2003) Reactive dye bioaccumulation by *Saccharomyces cerevisiae*. Process Biochem 38: 1437–1444. https://doi.org/10.1016/S0032-9592(03)00034-7
- Aksu Z (2005) Application of biosorption for the removal of organic pollutants: a review. Process Biochem 40:997–1026. https://doi.org/10.1016/j.procbio.2004.04.008
- Aksu Z, Dönmez G (2005) Combined effects of molasses sucrose and reactive dye on the growth and dye bioaccumulation properties of *Candida tropicalis*. Process Biochem 40:2443–2454. https://doi.org/10.1016/j.procbio.2004.09.013
- Aleu J, Collado IG (2009) Pollutants biodegradation by fungi. Curr Org Chem 13:1194. https://doi. org/10.2174/138527209788921774
- Al-Tohamy R, Kenawy ER, Sun J, Ali SS (2020) Performance of a newly isolated salt-tolerant yeast strain *Sterigmatomyces halophilus* SSA-1575 for azo dye decolorization and detoxification. Front Microbiol 11:1–19. https://doi.org/10.3389/fmicb.2020.01163
- Anku WW, Mamo MA, Mamo MA, Penny PW, Govender PP (2017) Phenolic compounds compounds in in water : water : sources, phenolic toxicity and treatment methods toxicity and treatment methods, in: Phenolic compounds—natural sources, importance and applications. pp. 419–443. https://doi.org/10.5772/66927
- Bankole PO, Adekunle AA, Obidi OF, Olukanni OD, Govindwar SP (2017) Degradation of indigo dye by a newly isolated yeast, *Diutina rugosa* from dye wastewater polluted soil. J Environ Chem Eng 5:4639–4648. https://doi.org/10.1016/j.jece.2017.08.050
- Basak B, Jeon B, Kurade MB, Saratale GD, Bhunia B, Chatterjee PK, Dey A (2019) Biodegradation of high concentration phenol using sugarcane bagasse immobilized *Candida tropicalis* PHB5 in a packed-bed column reactor. Ecotoxicol Environ Saf 180:317–325. https://doi.org/10.1016/j. ecoenv.2019.05.020
- Biradar SP, Rane NR, Patil TS, Khandare RV, Govindwar SP, Pawar PK (2016) Herbal augmentation enhances malachite green biodegradation efficacy of Saccharomyces cerevisiae. Biol 71: 475–483. https://doi.org/10.1515/biolog-2016-0069
- Bras R, Ferra MIA, Pinheir HM, Goncalves IC (2001) Batch tests for assessing decolourisation of azo dyes by methanogenic and mixed cultures. J Biotechnol 89:155. https://doi.org/10.1016/ s0168-1656(01)00312-1
- Castillo-carvajal LC, Sanz-martín JL, Barragán-huerta BE (2014) Biodegradation of organic pollutants in saline wastewater by halophilic microorganisms : a review. Environ Sci Pollut Res. https://doi.org/10.1007/s11356-014-3036-z
- Chang J, Kuo T, Chao Y, Ho J, Lin P (2000) Azo dye decolorization with a mutant *Escherichia coli* strain. Biotechnol Lett 22:807–812. https://doi.org/10.1023/A:1005624707777
- Charumathi D, Das N (2011) Biodegradation of Basic Violet 3 by Candida krusei isolated from textile wastewater. Biodegradation 22:1169–1180. https://doi.org/10.1007/s10532-011-9472-2
- Charumathi D, Das N (2012) Packed bed column studies for the removal of synthetic dyes from textile wastewater using immobilised dead *C. tropicalis*. Desalination 285:22–30. https://doi. org/10.1016/j.desal.2011.09.023

- Chen S, Luo J, Hu M, Geng P, Zhang Y (2012) Microbial detoxification of bifenthrin by a novel yeast and its potential for contaminated soils treatment. PLoS One 7:30862. https://doi.org/10. 1371/journal.pone.0030862
- Chequer FD, de Oliveira GAR, Ferraz EA, Cardoso JC, Zanoni MB, de Oliveira DP (2013) Textile dyes: dyeing process and environmental impact, in: In eco-friendly textile dyeing and finishing. IntechOpen. 268. https://doi.org/10.5772/3436
- Chivukula M, Spadaro JT, Renganathan V (1995) Lignin peroxidase-catalyzed oxidation of sulfonated azo dyes generates novel Sulfophenyl hydroperoxides. Biochemistry 34:7765– 7772. https://doi.org/10.1021/bi00023a024
- Chowdhary P, More N, Yadav A, Bharagava RN (2018) Ligninolytic enzymes: an introduction and applications in the food industry, enzymes in food biotechnology: production, applications, and future prospects. Elsevier Inc. https://doi.org/10.1016/B978-0-12-813280-7.00012-8
- Csutak O, Stoica I, Ghindea R, Tanase AM, Vassu T (2010) Insights on yeast bioremediation processes. Rom Biotechnol Lett 15:5066–5071
- Danouche M, El Arroussi H, Bahafid W, El Ghachtouli N, El Arroussi H, Bahafid W, An NEG (2021a) An overview of the biosorption mechanism for the bioremediation of synthetic dyes using yeast cells. Environ Technol Rev 10:58–76. https://doi.org/10.1080/21622515.2020. 1869839
- Danouche M, El Arroussi H, El N (2021b) Mycoremediation of synthetic dyes by yeast cells: a sustainable biodegradation approach. Environ Sustain 1–18. https://doi.org/10.1007/s42398-020-00150-w
- Danouche M, Ferioun M, Bahafid W, El Ghachtouli N (2021c) Mycoremediation of azo dyes using *Cyberlindnera fabianii* yeast strain: application of designs of experiments for decolorization optimization. Water Environ Res:1–15. https://doi.org/10.1002/wer.1499
- Danouche M, El Arroussi H, El Ghachtouli N (2022) Bioremoval of Acid Red 14 dye by Wickerhamomyces anomalus biomass: kinetic and thermodynamic study, characterization of physicochemical interactions, and statistical optimization of the biosorption process. Biomass Conv Bioref. https://doi.org/10.1007/s13399-022-02711-x
- Das D, Charumathi D, Das N (2010) Combined effects of sugarcane bagasse extract and synthetic dyes on the growth and bioaccumulation properties of *Pichia fermentans* MTCC 189. J Hazard Mater 183:497–505. https://doi.org/10.1016/j.jhazmat.2010.07.051
- Dave SR, Patel TL, Tipre DR (2015) Bacterial degradation of azo dye containing wastes. In: Microbial degradation of synthetic dyes in wastewaters, environmental science and engineering. Cham, pp 57–83. https://doi.org/10.1007/978-3-319-10942-8\_3
- Deeba F, Pruthi V, Negi YS (2018) Aromatic hydrocarbon biodegradation activates neutral lipid biosynthesis in oleaginous yeast. Bioresour Technol 255:273–280. https://doi.org/10.1016/j. biortech.2018.01.096
- Dönmez G (2002) Bioaccumulation of the reactive textile dyes by *Candida tropicalis* growing in molasses medium. Enzym Microb Technol 30:363–366. https://doi.org/10.1016/S0141-0229 (01)00511-7
- Dönmez G, Aksu Z (2002) Removal of chromium(VI) from saline wastewaters by *Dunaliella* species. Process Biochem 38:751–762. https://doi.org/10.1016/S0032-9592(02)00204-2
- Duckworth HW, Coleman JE (1970) Physicochemical and kinetic properties of mushroom tyrosinase. J Biol Chem 245:1613–1625. https://doi.org/10.1016/S0031-9422(00)82247-5
- Ehrampoush MH, Sadeghi A, Ghaneian MT, Bonyadi Z (2017) Optimization of diazinon biodegradation from aqueous solutions by *Saccharomyces cerevisiae* using response surface methodology. AMB Express 1–6. https://doi.org/10.1186/s13568-017-0366-5
- Filipowicz N, Momotko M, Boczkaj G, Cieśliński H (2020) Determination of phenol biodegradation pathways in three psychrotolerant. Enzyme Microb Technol J 141:0141–0229. https://doi. org/10.1016/j.enzmictec.2020.109663
- Fu Y, Viraraghavan T (2002) Removal of Congo Red from an aqueous solution by fungus *Aspergillus niger*. Adv Environ Res 7:239–247. https://doi.org/10.1016/S1093-0191(01) 00123-X

- Gargouri B, Mhiri N, Karray F, Aloui F, Sayadi S (2015) Isolation and characterization of hydrocarbon-degrading yeast strains from petroleum contaminated industrial wastewater. Biomed Res Int 2015:11. https://doi.org/10.1155/2015/929424
- Gönen F, Aksu Z (2009) Predictive expressions of growth and Remazol Turquoise Blue-G reactive dye bioaccumulation properties of *Candida utilis*. Enzym Microb Technol 45:15–21. https://doi. org/10.1016/j.enzmictec.2009.03.006
- Gong Y, Ding P, Xu M, Zhang C, Xing K, Qin S (2021) Biodegradation of phenol by a halotolerant versatile yeast *Candida tropicalis* SDP-1 in wastewater and soil under high salinity conditions. J Environ Manag 289:112525. https://doi.org/10.1016/j.jenvman.2021.112525
- Guo G, Tian F, Zhao Y, Tang M, Liu W, Liu C, Xue S, Kong W, Sun Y, Wang S (2019) Aerobic decolorization and detoxification of Acid Scarlet GR by a newly isolated salt-tolerant yeast strain *Galactomyces geotrichum* GG. Int Biodeterior Biodegrad 145:104818. https://doi.org/10. 1016/j.ibiod.2019.104818
- Han Y, Tang Z, Bao H, Wu D, Deng X (2019) Degradation of pendimethalin by the yeast YC2 and determination of its two main metabolites <sup>†</sup>. RSC Adv 9:491–497. https://doi.org/10.1039/ c8ra07872f
- Hashem M, Alamri SA, Al-zomyh SSAA, Alrumman SA (2018) Biodegradation and detoxi fi cation of aliphatic and aromatic hydrocarbons by new yeast strains. Ecotoxicol Environ Saf 151: 28–34. https://doi.org/10.1016/j.ecoenv.2017.12.064
- Huang S, Lin C, Kirschner R (2017) Biodegradation of the endocrine disrupter 4-tert-octylphenol by the yeast strain *Candida rugopelliculosa* RRKY5 via phenolic ring hydroxylation and alkyl chain oxidation pathways Ranjith. Bioresour Technol 226:55–64. https://doi.org/10.1016/j. biortech.2016.11.129
- Isia I, Hadibarata T, Sari AA, Al Khulaifi MM (2019) Potential use of a pathogenic yeast *Pichia kluyveri* FM012 for degradation of dichlorodiphenyltrichloroethane (DDT). Water Air Soil Pollut 230:1–11
- Jadhav JP, Govindwar SP (2006) Biotransformation of malachite green by *Saccharomyces* cerevisiae MTCC 463. Yeast 23:315–323. https://doi.org/10.1002/yea.1356
- Jadhav JP, Parshetti GK, Kalme SD, Govindwar SP (2007) Decolourization of azo dye methyl red by Saccharomyces cerevisiae MTCC 463. Chemosphere 68:394–400. https://doi.org/10.1016/j. chemosphere.2006.12.087
- Jadhav SU, Jadhav MU, Kagalkar AN, Govindwar SP (2008a) Decolorization of brilliant blue G dye mediated by degradation of the microbial consortium of *Galactomyces geotrichum* and *Bacillus* sp. J Chinese Inst Chem Eng 39:563–570. https://doi.org/10.1016/j.jcjce.2008.06.003
- Jadhav SU, Kalme SD, Govindwar SP (2008b) Biodegradation of Methyl red by Galactomyces geotrichum MTCC 1360. Int Biodeterior Biodegrad 62:135–142. https://doi.org/10.1016/j. ibiod.2007.12.010
- Jafari N, Soudi MR, Kasra-Kermanshahi R (2014) Biodegradation perspectives of azo dyes by yeasts. Microbiology 83:484–497. https://doi.org/10.1134/s0026261714050130
- Jain PK, Bajpai V (2012) Biotechnology of bioremediation—a review. Int J Environ 3:535–549. https://doi.org/10.6088/ijes.2012030131053
- Johnson EA, Echavarri-erasun C (2011) Yeast biotechnology. In: The yeasts. Elsevier B.V, pp 21-44. https://doi.org/10.1016/B978-0-444-52149-1.00003-3
- Kalyani DC, Patil PS, Jadhav JP, Govindwar SP (2008) Biodegradation of reactive textile dye red BLI by an isolated bacterium *Pseudomonas* sp. SUK1. Bioresour Technol 99:4635–4641. https://doi.org/10.1016/j.biortech.2007.06.058
- Kaushik P, Malik A (2009) Fungal dye decolourization: recent advances and future potential. Environ Int 35:127–141. https://doi.org/10.1016/j.envint.2008.05.010
- Khan R, Bhawana P, Fulekar MH (2013) Microbial decolorization and degradation of synthetic dyes: a review. Rev Environ Sci Biotechnol 12:75–97. https://doi.org/10.1007/ s11157-012-9287-6
- Kim YJ, Uyama H (2005) Tyrosinase inhibitors from natural and synthetic sources: structure, inhibition mechanism and perspective for the future. Cell Mol Life Sci 62:1707–1723. https:// doi.org/10.1007/s00018-005-5054-y

- Kucharska K, Wachowska U, Czaplicki S (2020) Wheat phyllosphere yeasts degrade propiconazole. BMC Microbiol 20:1–14. https://doi.org/10.1186/s12866-020-01885-6
- Kumari M, Abraham J (2011) Biodegradation of diesel oil using yeast *Rhodosporidium toruloides*. Res J Environ Toxicol 5:369–377. https://doi.org/10.3923/rjet.2011.369.377
- Kutty SN, Philip R (2008) Marine yeasts-a review. Yeast 465-483. https://doi.org/10.1002/yea
- Mahmoud MS (2016) Decolorization of certain reactive dye from aqueous solution using Baker's yeast (*Saccharomyces cerevisiae*) strain. HBRC J 12:88–98. https://doi.org/10.1016/j.hbrcj. 2014.07.005
- Martorell MM, Pajot HF, Ahmed PM, de Figueroa LIC (2017a) Biodecoloration of Reactive Black 5 by the methylotrophic yeast *Candida boidinii* MM 4035. J Environ Sci (China) 53:78–87. https://doi.org/10.1016/j.jes.2016.01.033
- Martorell MM, Pajot HF, de Figueroa LIC (2012) Dye-decolourizing yeasts isolated from Las Yungas rainforest. Dye assimilation and removal used as selection criteria. Int Biodeterior Biodegrad 66:25–32. https://doi.org/10.1016/j.ibiod.2011.10.005
- Martorell MM, Pajot HF, Figueroa LICD (2017b) Biological degradation of Reactive Black 5 dye by yeast *Trichosporon akiyoshidainum*. J Environ Chem Eng 5:5987–5993. https://doi.org/10. 1016/j.jece.2017.11.012
- Miranda RDCMD, Gomes EDB, Pereira N, Marin-Morales MA, Machado KMG, de Gusmão NB (2013) Biotreatment of textile effluent in static bioreactor by *Curvularia lunata* URM 6179 and Phanerochaete chrysosporium URM 6181. Bioresour Technol 142:361–367. https://doi.org/10. 1016/j.biortech.2013.05.066
- Misal SA, Gawai KR (2018) Azoreductase: a key player of xenobiotic metabolism. Bioresour Bioprocess. https://doi.org/10.1186/s40643-018-0206-8
- Nishiya Y, Yamamoto Y (2007) Characterization of a NADH:dichloroindophenol oxidoreductase from *Bacillus subtilis*. Biosci Biotechnol Biochem 71:611–614. https://doi.org/10.1271/bbb. 60548
- Okur M, Saraçoglu N, Aksu Z (2014) Use of response surface methodology for the bioaccumulation of Violet 90 metal-complex dye by *Candida tropicalis*. Turkish J Eng Environ Sci 38:217–230. https://doi.org/10.3906/muh-1409-2
- Pajot HF, de Figueroa LIC, Fariña JI (2007) Dye-decolorizing activity in isolated yeasts from the ecoregion of Las Yungas (Tucumán, Argentina). Enzym Microb Technol 40:1503–1511. https://doi.org/10.1016/j.enzmictec.2006.10.038
- Pajot HF, Martorell MM, de Figueroa LIC (2014) Ecology of dye decolorizing yeasts. In: Alvarez A, Polti MA (eds) Bioremediation in Latin America: current research and perspectives. Springer, pp 223–240. https://doi.org/10.1007/978-3-319-05738-5\_14
- Patel A, Sartaj K, Arora N, Pruthi V, Pruthi PA (2017) Biodegradation of phenol via meta cleavage pathway triggers de novo TAG biosynthesis pathway in oleaginous yeast. J Hazard Mater 340: 47–56. https://doi.org/10.1016/j.jhazmat.2017.07.013
- Phale PS, Sharma A, Gautam K (2019) Microbial degradation of xenobiotics like aromatic pollutants from the terrestrial environments, Pharmaceuticals and Personal Care Products: Waste Management and Treatment Technology Emerging Contaminants and Micro Pollutants. Elsevier Inc. https://doi.org/10.1016/B978-0-12-816189-0.00011-1
- Ramalho PA, Cardoso MH, Ramalho MT (2004) Characterization of azo reduction activity in a novel ascomycete yeast strain characterization of azo reduction activity in a novel ascomycete yeast strain. Appl Environ Microbiol 70:2279–2288. https://doi.org/10.1128/AEM.70.4.2279
- Rosu CM, Avadanei M, Gherghel D, Mihasan M, Mihai C, Trifan A, Miron A, Vochita G (2018) Biodegradation and detoxification efficiency of azo-dye reactive Orange 16 by *Pichia kudriavzevii* CR-Y103. Water Air Soil Pollut 229. https://doi.org/10.1007/s11270-017-3668-y
- Saratale RG, Saratale GD, Chang JS, Govindwar SP (2009a) Decolorization and biodegradation of textile dye Navy blue HER by *Trichosporon beigelii* NCIM-3326. J Hazard Mater 166:1421– 1428. https://doi.org/10.1016/j.jhazmat.2008.12.068
- Saratale RG, Saratale GD, Chang JS, Govindwar SP (2009b) Ecofriendly degradation of sulfonated diazo dye C.I. Reactive Green 19A using *Micrococcus glutamicus* NCIM-2168. Bioresour Technol 100:3897–3905. https://doi.org/10.1016/j.biortech.2009.03.051

- Saratale RG, Saratale GD, Chang JS, Govindwar SP (2011) Bacterial decolorization and degradation of azo dyes: a review. J Taiwan Inst Chem Eng 42:138–157. https://doi.org/10.1016/j.jtice. 2010.06.006
- Sen SK, Raut S, Bandyopadhyay P, Raut S (2016) Fungal decolouration and degradation of azo dyes: a review. Fungal Biol Rev 30:112–133. https://doi.org/10.1016/j.fbr.2016.06.003
- Solís M, Solís A, Pérez HI, Manjarrez N, Flores M (2012) Microbial decolouration of azo dyes: a review. Process Biochem 47:1723–1748. https://doi.org/10.1016/j.procbio.2012.08.014
- Song L, Shao Y, Ning S, Tan L (2018a) Performance of a newly isolated salt-tolerant yeast strain *Pichia occidentalis* G1 for degrading and detoxifying azo dyes. Bioresour Technol 233:21–29. https://doi.org/10.1016/j.biortech.2017.02.065
- Song Z, Song L, Shao Y, Tan L (2018b) Degradation and detoxification of azo dyes by a salttolerant yeast *Cyberlindnera samutprakarnensis* S4 under high-salt conditions. World J Microbiol Biotechnol 34. https://doi.org/10.1007/s11274-018-2515-7
- States U, States U (2011) Biodegradation and mineralization of metolachlor and alachlor by *Candida xestobii*. J Agric Food Chem 619–627. https://doi.org/10.1021/jf103508w
- Swamy J, Ramsay JA (1999) Effects of glucose and NH 4+ concentrations on sequential dye decoloration by *Trametes versicolor*. Enzym Microb Technol 25:278–284. https://doi.org/10. 1016/S0141-0229(99)00058-7
- Tan L, He M, Song L, Fu X, Shi S (2016) Bioresource technology aerobic decolorization, degradation and detoxification of azo dyes by a newly isolated salt-tolerant yeast *Scheffersomyces spartinae* TLHS-SF1. Bioresour Technol 203:287–294. https://doi.org/10. 1016/j.biortech.2015.12.058
- Tan L, Li H, Ning S, Xu B (2014) Aerobic decolorization and degradation of azo dyes by suspended growing cells and immobilized cells of a newly isolated yeast *Magnusiomyces ingens* LH-F1. Bioresour Technol 158:321–328. https://doi.org/10.1016/j.biortech.2014.02.063
- Tan L, Ning S, Zhang X, Shi S (2013) Aerobic decolorization and degradation of azo dyes by growing cells of a newly isolated yeast *Candida tropicalis* TL-F1. Bioresour Technol 138:307– 313. https://doi.org/10.1016/j.biortech.2013.03.183
- Thurston CF (1994) The structure and function of fungal laccases. Microbiology 140:19-26
- Waghmode TR, Kurade MB, Govindwar SP (2011) Time dependent degradation of mixture of structurally different azo and non azo dyes by using *Galactomyces geotrichum* MTCC 1360. Int Biodeterior Biodegrad 65:479–486. https://doi.org/10.1016/j.ibiod.2011.01.010
- Waghmode TR, Kurade MB, Kabra AN, Govindwar SP (2012a) Degradation of remazol red dye by *Galactomyces geotrichum* MTCC 1360 leading to increased iron uptake in Sorghum vulgare and Phaseolus mungo from soil. Biotechnol Bioprocess Eng 17:117–126. https://doi.org/10. 1007/s12257-011-0307-0
- Waghmode TR, Kurade MB, Kabra AN, Govindwar SP (2012b) Biodegradation of Rubine GFL by Galactomyces geotrichum MTCC 1360 and subsequent toxicological analysis by using cytotoxicity, genotoxicity and oxidative stress studies. Microbiol (United Kingdom) 158:2344– 2352. https://doi.org/10.1099/mic.0.060467-0
- Yang Q, Tao L, Yang M, Zhang H (2008) Effects of glucose on the decolorization of Reactive Black 5 by yeast isolates. J Environ Sci 20:105–108. https://doi.org/10.1016/S1001-0742(08) 60016-9
- Yu Z, Wen X (2005) Screening and identification of yeasts for decolorizing synthetic dyes in industrial wastewater. Int Biodeterior Biodegrad 56:109–114. https://doi.org/10.1016/j.ibiod. 2005.05.006
- Zhou XW, Cong WR, Su KQ, Zhang YM (2013) Ligninolytic enzymes from Ganoderma spp: current status and potential applications. Crit Rev Microbiol 39:416–426. https://doi.org/10. 3109/1040841X.2012.722606

# **Chapter 6 Sustainable Direct Digital Manufacturing Using Marine Resources**



Luís André R. Marques, Sara Biscaia, Anabela Massano, Rafael M. Tavares, and Artur Mateus

**Abstract** For over 60 years, global apparent food fish consumption has increased considerably generating a large volume of preconsumer and postconsumer residues, which consists mainly of shells and bones. Usually, fish by-products are used directly as feed in aquaculture or fertilizers. However, other applications have been gathering attention recently, such as the production of biofuel and biogas, pharmaceuticals, cosmetics and many others. Nowadays, this biowaste represents a promising source of biomaterials from marine discarded materials and this approach is very attractive due to their abundant availability, accessibility and low-cost source.

Direct Digital Manufacturing (DDM) is currently a main subject in the manufacturing industry placing many advantages, such as a high degree of geometric freedom for design and reduction of material waste, when compared to conventional manufacturing techniques. Therefore, DDM is seen as an energy-efficient technology. The use of biobased and biodegradable polymers in DDM technologies has been an emerging field in recent years, mainly in biomedical areas, due to the increasing interest in sustainable products and solutions, instead of using limited resources such as fuel-based polymers.

Thus, the valorization of bioactive compounds from fish by-products is of great interest due to their high market value, also can satisfy the demand of low-cost biomaterials, reduce marine pollution and can be used as alternative materials for DDM technologies. The reuse of these waste resources to produce biomaterials through sustainable processes can be a way to create new companies and job opportunities.

**Keywords** Fish waste · Biomaterials · Direct Digital Manufacturing · Fused Filament Fabrication · Direct-Ink Writing · Inkjet 3D Printing · Stereolithography

e-mail: anabela.p.massano@ipleiria.pt

L. A. R. Marques · S. Biscaia · A. Massano  $(\boxtimes)$  · R. M. Tavares · A. Mateus Centre for Rapid and Sustainable Product Development, Polytechnic Institute of Leiria, Marinha Grande, Portugal

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_6

## Abbreviations

3D	Three-Dimensional
ABS	Acrylic Butadiene Styrene
AM	Additive Manufacturing
ASTM	American Society for Testing and Materials
CAD	Computer-Aided Design
CIJ	Continuous Inkjet
DDM	Direct Digital Manufacturing
DIW	Direct-Ink Writing
DOD	Drop on Demand
FDM	Fused Deposition Modelling
FFF	Fused Filament Fabrication
GelMA	Gelatin-methacryloyl
HA	Hydroxyapatite
ICT	Information and Communication Technology
ISO	International Standard Organisation
PEGDA	Polyethylene Glycol Diacrylate
PET	Polyethylene Terephthalate
PHA	Polyhydroxyalkanoate
PHB	Polyhydroxybutyrate
PLA	Polylactic Acid
SL	Stereolithography
SLA	Stereolithography Apparatus
β-TCP	Tricalcium phosphate

# 6.1 The Use of Fish Waste as 3D Printed Biomaterials

Annually, millions of fish waste are produced worldwide, despite the percentage of food losses and waste varies mostly according to culture (Yan and Chen 2015; de la Caba et al. 2019). The European Market Observatory for Fisheries and Aquaculture Products (EUMOFA) reported that in 2018 the total world catches and aquaculture production was of 97 million tonnes and 115 million tonnes, respectively (Fig. 6.1).

On the other hand, for over 60 years, global apparent food fish consumption has increased considerably. In the period 1961–2017, the average annual growth rate of total food fish consumption was 3.1%. Annual per capita fish consumption varies from 9 kg to more than 24 kg due to the influence of cultural, economic and geographical factors, including the proximity and access to fish landings and aquaculture facilities (Fig. 6.2) (FAO 2020).

The consumption of fish generates a large volume of preconsumer and postconsumer residue, the largest proportion of which consists of shells and bones, which could correspond to 50-70% of the product content (de la Caba et al. 2019).

Continent	Volume (1.000 tonnes)	% catches	% aquaculture
Asia	155.005	32%	68%
Americas	24.462	84%	16%
Europe	18.183	83%	17%
Africa	12.408	81%	19%
Oceania	1.634	87%	13%
Total	211.692	46%	54%



**Fig. 6.1** World catches and aquaculture production by continent in 2018 (adapted from European Market Observatory for Fisheries and Aquaculture Products (EUMOFA) (2020))

Region/economic grouping	Total food fish consumption (million tonnes live weight equivalent)	Per capita food fish consumption (kg/year)
World	152.9	20.3
World (excluding China)	97.7	16.0
Africa	12.4	9.9
North America	8.1	22.4
Latin America and the Caribbean	6.7	10.5
Asia	108.7	24.1
Europe	16.1	21.6
Oceania	1.0	24.2
Developed countries	31.0	24.4
Least developed countries	12.4	12.6
Other developing countries	109.5	20.7
Low-income food-deficit countries	23.3	9.3

Fig. 6.2 Total and per capita apparent fish consumption, 2017 (adapted from FAO (2020))

Considering the most consumed species in the world in 2018, around 34% of them present external shells (i.e. squids, clams, mussels, shrimps) and the remaining have bones in their composition (i.e. salmon, tunas, colds) (Fig. 6.3) (FAO 2020; Nisticò 2017).

Global food loss and waste is a serious issue concerning the focus of Sustainable development goal target 12.3, which aims at halving wastage by 2030 (FAO 2020). The current management of fish waste is the disposal in public waters or onto landfills. These can cause noxious odours as a consequence of the decay of remaining organic matter or of the microbial decomposition of salts into gases, and consequently, this causes the appearance of animals, such as insects, that feed of organic matter (Chierighini et al. 2011; Hamester et al. 2012). Regarding bivalve



**Fig. 6.3** Share of main groups of species in fish trade in terms of value, 2018 (adapted from FAO (2020))

shells waste, the major problem of discharge into the sea is the residue accumulation in the ground, which over the years causes the increase of sediments, a harmful factor for cultivation promoting the decrease in oxygen in the water, hindering the growth of microalgae, which are mainly responsible for the nutrition of some bivalves, which hinders their growth in their natural habitat (Chierighini et al. 2011; Shavandi et al. 2018).

Another scenario for fish waste, is their use as feed or raw material since the correct waste management must involve trying to avoid or reduce its production and minimize the risk to human health and the environment (Yao et al. 2014). The reduction of waste production can be done through the reuse of products and, when this is not possible, recycling should always be used for another type of recovery, using only waste disposal as the last alternative (Monteiro 2014). Typically, fish by-products are used directly as feed for aquaculture, silage or fertilizers (FAO 2020;

Nisticò 2017). However, in the last decades, other applications have been gaining attention, since presently can be treated with more efficiency as a result of improved processing technologies (FAO 2020). Fish by-products have also been used in the production of biofuel and biogas, dietetic products, pharmaceuticals, natural pigments, cosmetics, alternatives to plastic, and constituents in other industrial processes. Despite this, more research and innovation are still needed in this area, this biowaste needs to be valued through the development of economically sustainable routes in order to eliminate the related environmental problems (Nisticò 2017; Yao et al. 2014).

Fish bones are an excellent source of collagen, gelatine and calcium and other minerals such as phosphorus (Jafari et al. 2020; Maschmeyer et al. 2020; Terzioğlu et al. 2018). Calcium phosphates present in fish bone have been the focus of many researchers due to its potential to take the advantage of producing a high-quality bioengineering material for several applications such as drug delivery, tissue engineering and environmental remediation (FAO 2020; Terzioğlu et al. 2018). Crustaceans and bivalves also offer several applications for their by-products, which not only benefit the economy of industrial processors through the recovery of bioactive molecules from the waste but also help to reduce environmental pollution caused by the slow natural degradation rate of their shells (FAO 2020; Nirmal et al. 2020). Chitin, a polysaccharide extracted from crustacean shell waste, has gained much attention due to their non-toxicity and variety of bioactivities (Chierighini et al. 2011; Shavandi et al. 2018; Borić et al. 2020). Its derivative, chitosan, has been successfully 3D printed using extrusion-based processes for bone tissue engineering (Sanchez-Rexach et al. 2020). The shells of bivalves, such as mussels and ovsters, can be turned into calcium carbonate or calcium oxide, two highly versatile chemical compounds with wide biomedical applications (FAO 2020; Ismail et al. 2021).

Altogether, represent promising sources of biomaterials from marine discarded materials and this approach is very attractive due to their abundant availability, less processing time, low-cost source, easy accessibility and simple way of handling (Govindharaj et al. 2019). In addition, the reuse of these waste resources to produce biomaterials through sustainable processes can be a way to create new companies and job opportunities. So, the valorisation of bioactive compounds from these by-products is of great interest for their high market value, beyond can satisfy the demand of low-cost biomaterials for commercialization, can promote cleaner production, reduce marine pollution and can be used as alternative materials for DDM technologies (Govindharaj et al. 2019; Caruso et al. 2020).

#### 6.1.1 Biomaterials from Fish Waste

#### 6.1.1.1 Calcium Carbonate

Calcium carbonate  $(CaCO_3)$  is an exceptional material, especially in industrial and biomedical fields. Compared with other inorganic materials, CaCO<sub>3</sub> has shown



Fig. 6.4 Natural sources of HA and TCP (adapted from Terzioğlu et al. (2018))

promising potential for biomedical applications because of its biocompatibility, biodegradability and high mechanical strength (Barhoum et al. 2015a; Sambudi et al. 2015). Furthermore, CaCO<sub>3</sub> particles can be used as filler materials (Abdolmohammadi et al. 2012; Barhoum et al. 2015b), coatings (Barhoum et al. 2014), pharmaceuticals (Saveleva et al. 2018; Mohd Abd Ghafar et al. 2017) and bone tissue engineering (Luo et al. 2018; Didekhani et al. 2020). CaCO<sub>3</sub> can have mainly four polymorphs: calcite, vaterite, aragonite and amorphous CaCO<sub>3</sub> and can be extracted by mining or chemically synthesized in the laboratory (Mohd Abd Ghafar et al. 2017; Hoque et al. 2013). Calcite is the thermodynamically stable phase of CaCO<sub>3</sub> and vaterite and aragonite are the metastable phases of CaCO<sub>3</sub> (Ma et al. 2016). The amorphous phase is unstable, it rapidly crystallizes near ambient temperature and pressure within a few seconds to a few minutes and acts as a seed for crystal growth of the other polymorphs (Mohd Abd Ghafar et al. 2017; Boyjoo et al. 2014). Calcite, vaterite and aragonite have typical morphologies/shapes respectively rhombohedral, spherical and needle-like (Boyjoo et al. 2014).

Fish bones are rich in CaCO<sub>3</sub>, making it an alternative source of low-cost CaCO<sub>3</sub> for the synthesis of calcium phosphate bioceramic used in bone regeneration (Corrêa and Holanda 2019). Various fish bones have been used as a starting material to produce HA and  $\beta$ -TCP as listed in Fig. 6.4.

Regarding a traditional method to prepare calcium phosphates from fish bones, the thermal calcination method concerns some aspects such as bones source,



extraction method, calcination temperature and time that can influence the final properties of calcium phosphates, particularly, the morphology, purity degree, particle size and distribution, surface properties and Ca:P ratio (Terzioğlu et al. 2018; Naga et al. 2015; Piccirillo et al. 2013) (Fig. 6.5). Alternative methods have been studied to produce calcium phosphates, such as alkaline hydrolysis (Venkatesan et al. 2015), hydrothermal (Goto and Sasaki 2016) and laser ablation (Terzioğlu et al. 2018; Boutinguiza et al. 2007).

Hydroxyapatite (HA) has been prepared from fishbones using an alkaline hydrolysis method. This biomaterial, shows excellent results in bone remineralization processes, with regard to osteoblast cell proliferation (Shavandi et al. 2018). Côrrea and Holanda (2019) synthetized HA from fish bone waste by wet precipitation method and demonstrate that this approach has great potential for producing nanopowder of biphasic calcium phosphate, particularly for the regeneration of damaged bone tissue in orthopaedics. Another study by Naga et al. (Naga et al. 2015) aiming to obtain hydroxyapatite powder from fish bone skeletons by thermal treatment and preparation of highly porous 3D ceramic scaffolds by polymeric sponge method revealed the mechanical properties of scaffolds very near to the strength of trabecular bone.

Regarding the extraction of calcium carbonate from shellfish, studies revealed the chemical composition of various seashells in 92–99% of this mineral with about 5% of organic matter. The dry shell weight of oysters, mussels, molluscs, cockles and

Oxides	Oyster	Hard-shelled mussel	Clam	Short-necked clam
CaO	51.06	53.70	53.92	53.58
MgO	0.51	0.33	0.22	0.20
SiO <sub>2</sub>	2.00	0.20	0.46	0.66
Al <sub>2</sub> O <sub>3</sub>	0.50	0.13	0.20	0.40
Fe <sub>2</sub> O <sub>3</sub>	0.20	0.03	0.04	0.04
P <sub>2</sub> O <sub>5</sub>	0.18			
K <sub>2</sub> O	0.06			
SrO	0.09			
SO <sub>3</sub>	0.60			
Na <sub>2</sub> O	0.58			
TiO <sub>2</sub>	0.02			
Mn <sub>2</sub> O <sub>3</sub>	0.02			
lg. loss (%) <sup>a</sup>	44.16	45.61	45.16	45.12

Table 6.1 Chemical compositions of shells (adapted from Yao et al. (2014))

<sup>a</sup>Including CO and organic materials lost by heating

scallops ranges from 0.01 g to 58.33 g or approximately 52–80% of the whole animal. From a chemical point of view, calcing or pyrolyzing could convert calcium carbonate into calcium oxide, so it is possible to use shell waste as an alternative source to obtain these compounds (Yao et al. 2014). Several studies with the purpose of extracting calcium carbonate from different seashell types reported calcium oxide contents rather than calcium carbonate contents (Table 6.1). The results revealed that the removal of organic material from seashells may be difficult and heating to high temperatures results in forms with a higher degree of purity (Owuamanam and Cree 2020).

Islam et al. (Islam et al. 2013) reported a study using an environmentally friendly method for the synthesis of calcium carbonate nanoparticles from cockle shells and demonstrate that this extraction method has great potential in industry for the large-scale production of calcium carbonate nanoparticles for biomedical applications. Luo et al. (Luo et al. 2018) prepared polycaprolactone and oyster shell powder scaffolds by additive manufacturing using a 3D printing system and demonstrated no significant cytotoxicity effect of the prepared scaffolds towards MG-63 cells and favourable biocompatibility for bone tissue engineering.

#### 6.1.1.2 Chitin and its Derivatives

Chitin can be found in many organisms, such as fungi, plankton and the exoskeletons of insects and crustaceans.

The main marine sources of chitin and the commercial production of chitosan especially rely on chitin from crab, shrimp, prawn and krill species (Raghavankutty et al. 2018).


Fig. 6.6 Waste input and output from Chitosan Industries (adapted from Mathew et al. (2021))

Chitosan is composed of N-acetyl-d-glucosamine and d-glucosamine units and is polycationic in nature (Raghavankutty et al. 2018). Currently, the deacetylated form of chitin, chitosan and their derivatives has also been developed for a broad spectrum of applications, such as industrial chemistry, such as cosmetics, textiles, water treatment and biomedicine (Yao et al. 2014). Films, hydrogels, microspheres and nanoparticles are the major forms of chitosan in biomedical and pharmaceutical applications (Raghavankutty et al. 2018). Biocompatibility, biodegradability, antimicrobial activity, wound healing, and anti-tumour activity are some of their properties that contribute to its inevitable importance in fields already mentioned (Raghavankutty et al. 2018).

The industrial-scale production of chitin and chitosan require harsh chemicals like HCl for demineralization (to remove calcium carbonate) and NaOH (to remove the protein) (Mathew et al. 2021). The extraction of chitin from crustaceans requires approximately 6.3 kg of HCl and 1.8 kg NaOH, which turns the extraction very expensive. Furthermore, approximately 1.4 tonnes of water are required to completely remove the chemicals from the crustacean wastes, as well as high temperatures for longer durations (Mathew et al. 2021). Figure 6.6 explains the input and outputs derived from the chitosan factory.

Chitosan hydrogels have been successfully 3D printed using extrusion-based processes for engineering bone tissue. Inorganic molecules have also been mixed with chitosan for the purpose of improving its bioactivity to resemble bone (Sanchez-Rexach et al. 2020).

#### 6.1.1.3 Collagen and Gelatin

Collagen is the major structural protein of fish skin and bones, representing 30% of the total protein content, and it is the main component of the extracellular matrix (ECM) in connective tissues (skins, tendons, ligaments and bones) (Jafari et al. 2020). Collagen is classified according to its structural features; there are at least 28 types of collagen and in fish is possible to obtain types I and II. Collagen and gelatin are derived from the same macromolecules, being gelatin a partially hydrolysed form of collagen in a denaturated state (Caruso et al. 2020). The use of collagen from marine sources has attracted increasing attention due to their cost-effective process, high collagen content, biocompatibility similar to conventional bovine and porcine collagen and high absorption by the human body (Jafari et al. 2020).

Collagen and gelatin have a wide range of applications in food, cosmetic and pharmaceutical industries. In biomedical applications, they also have been explored as drug and gene carriers, bone-filling materials and wound dressings (Shavandi et al. 2018; Jafari et al. 2020).

Regarding the various extraction methods used to prepare collagen from marine sources, the conventional protocol includes the highlighted steps shown in Fig. 6.7. The type of fish collagen must be considered to use more adequate extraction parameters, such as the extraction medium pH and the temperature of the process, since they can influence the final properties of the extracted collagen (Shavandi et al. 2018; Raghavankutty et al. 2018).

Collagen can be used in several forms, such as injectable solutions and dispersions, that can be processed further using Direct Digital Manufacturing technologies. Govindharaj et al. (Govindharaj et al. 2019) reported the extraction of collagen from marine eel skin as a potential blue biomaterial and incorporated into alginate hydrogel to fabricate scaffolds using extrusion-based 3D printing technology. Sanz et al. (Sanz et al. 2021) described the extraction, characterization and methacrylation



Fig. 6.7 Collagen extraction procedure from fish by-products (adapted from Jafari et al. (2020))

of Red Snapper collagen for 3D Coaxial Printing of neural and skeletal muscle cell cultures as a model for neuromuscular junction formation.

# 6.2 Direct Digital Manufacturing: General Overview

Direct Digital Manufacturing (DDM) is currently a main subject in manufacturing industry. Therefore, it is important to discuss the evolution of this term. Additive Manufacturing (AM) is a continuously growing technology seen as a viable and reliable solution to process a wide range of different materials. AM is defined by the International Standard ISO/ASTM 52900:2021 as "the process of joining materials to make parts from 3D model data, usually layer upon layer, as opposed to subtractive and formative manufacturing methodologies" (ISO/ASTM52900:2021 (en) 2021).

AM techniques, commonly also referred to as 3D printing, were introduced in the market in the late 1980s, with the development of stereolithography (SLA) process by Charles "Chuck" W. Hull (Hull 1986). Since then and until now, with a constant increase in interest in this area, many technological advances have been developed. Therefore, the term AM is also evolving to DDM, referring to the interconnection of AM technologies and information and communication technology (ICT), such as computer software and communication through a network, making possible the creation and production of final products and end-use components. This technology introduces a change in the production paradigm, since the products will be derived from a 3D CAD model and could be produced closer or at the customer location utilizing an AM equipment (Chen et al. 2015).

According to the International Standard ISO/ASTM 52900:2021 (ISO/ASTM52900:2021(en) 2021), DDM technologies are divided into seven different categories (Fig. 6.8), such as:

 Material Extrusion – a process in which material is selectively dispensed through a nozzle or orifice.



**Fig. 6.8** Classification of additive manufacturing processes according to ISO/ASTM 52900:2021 (adapted from Dilberoglu et al. (2017))

- VAT Photopolymerization a process in which liquid photopolymer in a vat is selectively cured by light-activated polymerization.
- Material Jetting a process in which droplets of build material are selectively deposited.
- Binder Jetting a process in which a liquid bonding agent is selectively deposited to join powder materials.
- Powder Bed Fusion a process in which thermal energy selectively fuses regions of a powder bed.
- Directed Energy Deposition a process in which focused thermal energy is used to fuse materials by melting as they are being deposited.
- Sheet Lamination a process in which sheets of material are bonded to form a part.

DDM offers many advantages, such as a high degree of geometric freedom for design, reduction of material waste, reduction of tooling, time-to-market and, therefore, costs, when compared to conventional manufacturing techniques. The high flexibility of these technologies in terms of geometry, by building parts layer by layer, enables the production of complex and light functional parts, with associate weight and material savings. Thus, DDM is gaining recognition as an energy-efficient technology. The reduction or complete elimination in the need for tooling means that the cost of producing a single part through DDM is approximately the same despite the amount of parts needed. This makes DDM suitable for the production of smaller batch sizes when compared to conventional processes like injection moulding, also being an amazing alternative to produce personalized and high-quality products that respond to an individual need (Gibson et al. 2015). For these reasons, DDM is seen by the industry as a disruptive technology that will enable the growth of new products and new business models, with applications in various areas (e.g. aerospace, automotive, biomedical, food industry).

# 6.2.1 DDM for Biopolymers Production

DDM technologies using biobased and biodegradable polymers have been an emerging field in the last few years, due to the increasing interest in developing sustainable products and solutions, instead of using limited resources such as fuelbased polymers (Jiang and Zhang 2017). However, DDM has not yet achieved its full potential to be used as a regular production technology, due to limitations related with the availability of materials, their sustainability and their lack of processability concerning the DDM processes and equipment requirements (Liu et al. 2019).

In the last few years, the number of articles published regarding biopolymer DDM increased from less than 5 articles in 2013 to more than 1000 in 2020 (Santoni et al. 2022), which shows the effort on exploring natural and renewable biopolymers to achieve a recycling and sustainable economy. Biomass from residuals of renewable resources, such as woody or marine, is seen as a thriving alternative to fossil

resources. By using the biorefinery concept it is possible to apply biomass conversion processes and extract different components (e.g. collagen, chitosan, alginate) and then incorporate them into innovative materials for specific 3D printing techniques (Li et al. 2021).

The most promising DDM technologies for industrial biopolymers production are extrusion-based processes—Fused Filament Fabrication (FFF) and Direct-Ink Writing (DIW), Inkjet 3D printing—Drop on Demand (DOD) and Laser-assisted 3D printing—Stereolithography (SLA).

#### 6.2.1.1 Fused Filament Fabrication

Fused Filament Fabrication (FFF) is a material extrusion AM process in which a thermoplastic filament is melted and forced through a nozzle to the build platform, layer by layer (Fig. 6.9). Scott Crump, founder of Stratasys Inc., developed this technology in 1992 under the term of Fused Deposition Modelling (FDM<sup>TM</sup>) (Scott Crump 1992). Since FDM<sup>TM</sup> is the term trademarked by Stratasys Inc., FFF is commonly used to refer to the same process. FFF is the most used DDM technology for polymer AM due to the simplicity and low cost of the process when compared to other techniques. FFF is commonly used for printing polymeric materials such as polylactic acid (PLA), polyethylene terephthalate (PET), acrylic butadiene styrene (ABS), among others. The polymeric filament is heated above the melting temperature and rapidly hardens after being deposited in the build platform, which allows



Fig. 6.9 Schematic diagram of a typical Fused Filament Fabrication (FFF) printer

the adhesion of the layer immediately deposited after on it. In addition, it is also required to make sure that the filaments produced have the appropriate modulus and flexural strength so they can be spooled and pushed by the drive gear and the idle pulley (Ngo et al. 2018).

FFF was first developed to produce prototypes from plastic material. However, nowadays it is also used to fabricate 3D parts with biopolymers with applications in biomedical fields. A few studies reported that FFF is a capable technology in the production of bioproducts and scaffolds for tissue engineering using polyhydroxyalkanoates (PHAs) (Ausejo et al. 2018; Wu et al. 2017; Wu 2018; Rydz et al. 2020) and polyhydroxybutyrate (PHB) biopolymers (Kontárová et al. 2020; Duan et al. 2011; Giubilini et al. 2020). PHA and PHB are biodegradable and environmentally friendly polymers that can be extracted from marine microbes and organisms.

The main challenges regarding the application of biopolymers in FFF are related with the clogging of the printing nozzle, associated with the fragility of the filaments or with the changes of viscosity caused by the filler content (Mazzanti et al. 2019). One of the solutions is to combine the principles of FFF technology and use a different mechanism to push the material through the heated nozzle, through a material extrusion principle but without the need of preparing filaments.

### 6.2.1.2 Direct-Ink Writing

Direct-Ink Writing (DIW), like FFF, is a material extrusion AM process. In DIW, a viscoelastic ink is forced through the printing nozzle onto a build platform under controlled flow rates (Fig. 6.10), in order to form 3D fibre structures at ambient temperature with a predefined pattern, layer by layer (Saadi et al. 2022). The extrusion of the ink can be done by different extrusion mechanism, such as pneumatic-based, piston-based or screw-based extrusion, which make this process suitable for working with a huge range of fluids with different viscosity values (Murphy and Atala 2014).

Typically, the inks used in DIW are in the form of pastes or gels, which normally includes organic, colloidal, nanoparticle-filled and sol-gel inks. Since DIW allows the production of 3D parts directly from pastes and gels, instead of using heat in the extrusion process like in FFF, the inks need to go through a solidification process, such as evaporation, gelation or solvent-driven steps. Therefore, it is important that DIW materials match rigorous requirements in terms of rheology in order to flow with ease and recover after the deposition. Their formulations should behave as non-Newtonian fluids, usually referred to as yield stress fluids, which means that they are able to flow or deform indefinitely when submitted to a specific stress (Gibson et al. 2015; Rocha et al. 2020).

DIW is a promising technology in the production of biopolymers for biomedical areas, for example in tissue engineering. Several studies have been performed using alginate, a polysaccharide derived from seaweed with biocompatibility and non-toxic properties, to produce lattice 3D structures successfully (Bendtsen et al.



Fig. 6.10 Schematic diagram of Direct-Ink Writing (DIW) process

2017; Möller et al. 2017; Wu et al. 2016; Chawla et al. 2020; Choe et al. 2022). Other study evaluated the printability of biopolymers with collagen, an important protein that can be extracted from fish bones, with applications in biomedical or drug discovery (Zhao et al. 2015), and also the production of scaffolds for tissue regeneration using chitosan, a biocompatible polysaccharide with wound healing and anti-fungal effects that comes from the skeleton of shellfish (Demirtaş et al. 2017; Elviri et al. 2017).

### 6.2.1.3 Inkjet 3D Printing

Inkjet 3D Printing is a low-pressure and low-temperature process that involves the deposition of ink droplets on a platform, layer by layer (Fig. 6.11). The printing material is extruded through a small nozzle within a print head, with each individual layer cured between consecutive depositions (using an infrared or ultraviolet light)



Fig. 6.11 Schematic diagram of a Drop on Demand (DOD) inkjet process

(Gibson et al. 2015). Inkjet 3D Printing is able to produce a wide range of materials, such as polymers, dielectric and nano-conductive nanoparticles. This technology can be performed using two different mechanisms, namely Drop on Demand (DOD) and Continuous Inkjet (CIJ) printing. In the CIJ process, a continuous liquid ink flows through the nozzle into a droplet format. In DOD inkjet printing, the most common and more cost-effective when compared to CIJ, tiny droplets are deposited in the build substrate due to the action generated by thermal or piezoelectric actuators (Guo et al. 2017). In thermal DOD printing, heat is produced to create vapour bubbles that force the ejection of the material droplets. On the other hand, in piezoelectric DOD printing, an electric signal is applied to a piezoelectric material that generates acoustic pulses, which forces the ejection of the ink droplets (Derby 2010).

Besides the high printing speed and cost-effectiveness, this process provides higher precision and quality of production because the actuator mechanism allows to control the droplet uniformity and size. Therefore, DOD inkjet printing is gathering a lot of attention as a disruptive process in biomaterials production (Guo et al. 2017).

Köpf et al. reported that the use of agarose, a polysaccharide commonly extracted from red seaweed that is used in molecular biology for the separation of large molecules, combined with collagen allows to successfully print 3D complex structures that can be used in disease and drug discovery (Köpf et al. 2016). Another study showed that is possible to 3D print pre-vascularised tissue replacements by DOD inkjet printing using Gelatin-methacryloyl (GelMA) and collagen hydrogel solutions (Stratesteffen et al. 2017).

#### 6.2.1.4 Stereolithography

Stereolithography Apparatus (SLA) or usually Stereolithography (SL) was the first patented (1986) and commercialized (1988) AM process, developed by Charles W. Hull, founder of Stratasys Inc. (Hull 1986). In SL, a UV laser is used to selectively polymerize the photocurable resin contained in a vat (Fig. 6.12). This process starts with the scanning of the photosensitivity resin surface and after the first layer is complete, the build platform shifts down and another laser scanning is performed to the next layer, and so on until the 3D solid part is finished. After the print is completed, the part is submersed in a solvent, such as isopropyl alcohol, to remove any excess of resin and the support structures, which are usually necessary to ensure the success of the print and avoid deformations, can be removed manually. After that, and depending on the desired application, the final part can be submitted to a post-curing step to enhance material properties, where temperature and UV light are combined (Gibson et al. 2015).

The materials used in SLA are usually thermosetting photopolymer resins. This means that the photopolymerization process is irreversible and it is not possible to



Fig. 6.12 Schematic diagram of a stereolithography (SLA) process

heat and convert the printed SLA parts back to their original liquid form. SLA technique is capable of produce fine and complex structures with high dimensional accuracy and good surface properties with minimal stepping effect, when compared to the material extrusion AM processes. Due to the characteristics of SLA process, especially attention is being paid to the bioprinting area (Grigoryan et al. 2021).

The development of calcium phosphate suspensions for SLA was studied and reported as a viable solution for producing 3D components (Goutagny et al. 2021). Calcium phosphate is a mineral extracted from marine fish bones with good biocompatibility and osteoconductivity. Gyroid scaffolds were successfully produced by SLA using calcium phosphate nanoparticles, with great potential for bone repair applications (Ullah et al. 2021). 3D hybrid scaffolds with ear shape were also produced by SLA using a hybrid biocompatible resin with chitosan and polyethylene glycol diacrylate (PEGDA), a synthetic polymer. Long-term cell viability and spreading were observed, confirming the possibility of printing chitosan by SLA and providing cell-adhesive properties to the scaffolds, suitable for repairing complex geometries (Morris et al. 2016).

# 6.3 Current Challenges and Future Perspectives

In the last few years was made incredible progress in DDM of biopolymers, mainly in biomedical areas, with many process improvements and advances in feedstock materials. As a raw material, biopolymers represent an ideal option to be used in AM processes, combining their biodegradable, non-toxic and environment-friendly properties with the possibility of producing complex 3D porous structures.

However, the feedstock formulation is still the major challenge to overcome for the large adoption and large-scale production by the industry using DDM. Questions related to feedstock printability, sustainability and functionality need further attention to match and achieve the cost and eco-effectiveness inherent to DDM technologies. Properties such as material degradation, compatibility with living cells and mechanical strength and stiffness are the main barriers in today's biopolymer 3D printing. The development of biobased materials for DDM processes will be a growing trend, with an increasing focus on discovering sustainable and environment-friendly feedstocks.

Therefore, DDM shows a great potential for the development of natural-delivered biopolymer 3D structures, with many biocomponents available in marine environments or residues, the largest renewable resource on the planet.

### References

Abdolmohammadi S, Siyamak S, Ibrahim NA, Wan Yunus WMZ, Ab Rahman MZ, Azizi S, Fatehi A (2012) Enhancement of mechanical and thermal properties of polycaprolactone/chitosan

blend by calcium carbonate nanoparticles. Int J Mol Sci 13:4508–4522. https://doi.org/10.3390/ ijms13044508

- Ausejo JG et al (2018) Three-dimensional printing of PLA and PLA/PHA dumbbell-shaped specimens of crisscross and transverse patterns as promising materials in emerging application areas: prediction study. Polym Degrad Stab 156:100–110. https://doi.org/10.1016/j. polymdegradstab.2018.08.008
- Barhoum A, Rahier H, Abou-Zaied RE, Rehan M, Dufour T, Hill G, Dufresne A (2014) Effect of cationic and anionic surfactants on the application of calcium carbonate nanoparticles in paper coating. ACS Appl Mater Interfaces 6:2734–2744. https://doi.org/10.1021/am405278j
- Barhoum A, Van Assche G, Makhlouf ASH, Terryn H, Baert K, Delplancke MP, El-Sheikh SM, Rahier H (2015a) A green, simple chemical route for the synthesis of pure nanocalcite crystals. Cryst Growth Des 15:573–580. https://doi.org/10.1021/cg501121t
- Barhoum A, Van Lokeren L, Rahier H, Dufresne A, Van Assche G (2015b) Roles of in situ surface modification in controlling the growth and crystallization of CaCO3 nanoparticles, and their dispersion in polymeric materials. J Mater Sci 50:7908–7918. https://doi.org/10.1007/s10853-015-9327-z
- Bendtsen ST, Quinnell SP, Wei M (2017) Development of a novel alginatepolyvinyl alcoholhydroxyapatite hydrogel for 3D bioprinting bone tissue engineered scaffolds. J Biomed Mater Res A 105(5):1457–1468. https://doi.org/10.1002/jbm.a.36036
- Borić M, Vicente FA, Jurković DL, Novak U, Likozar B (2020) Chitin isolation from crustacean waste using a hybrid demineralization/DBD plasma process. Carbohydr Polym J 246. https:// doi.org/10.1016/j.carbpol.2020.116648
- Boutinguiza M, Lusquinos F, Comesana R, Riveiro A, Quintero F, Pou J (2007) Production of microscale particles from fish bone by gas flow assisted laser ablation. Appl Surf Sci 254:1264– 1267. https://doi.org/10.1016/j.apsusc.2007.08.015
- Boyjoo Y, Pareek VK, Liu J (2014) Synthesis of micro and nano-sized calcium carbonate particles and their applications. J Mater Chem A 2:14270–14288. https://doi.org/10.1039/c4ta02070g
- de la Caba K, Guerrero P, Trung TS, Cruz-Romero M, Kerry JP, Fluhr J, Maurer M, Kruijssen F, Albalat A, Bunting S, Burt S, Little D, Newton R (2019) From seafood waste to active seafood packaging: an emerging opportunity of the circular economy. J Clean Prod 208:86–98. https:// doi.org/10.1016/j.jclepro.2018.09.164
- Caruso G, Floris R, Serangeli C, Di Paola L (2020) Fishery wastes as a yet undiscovered treasure from the sea: biomolecules sources, extraction methods and valorization. Mar Drugs 18. https:// doi.org/10.3390/md18120622
- Chawla D, Kaur T, Joshi A, Singh N (2020) 3D bioprinted alginate-gelatin based scaffolds for soft tissue engineering. Int J Biol Macromol 141:560–567. https://doi.org/10.1016/j.ijbiomac.2019. 12.127
- Chen D et al (2015) Direct digital manufacturing: definition, evolution, and sustainability implications. J Clean Prod 107:615–625. https://doi.org/10.1016/j.jclepro.2015.05.009
- Chierighini D, Bridi R, da Rocha AA, Lapa KR (2011) Possibilidades do Uso das Conchas de Moluscos, in: 3rd Int. Work. Adv. Clean. Prod., São Paulo Brazil
- Choe G, Lee M, Oh S, Seok JM, Kim J, Im S, Park SA, Lee JY (2022) Three-dimensional bioprinting of mesenchymal stem cells using an osteoinductive bioink containing alginate and BMP-2-loaded PLGA nanoparticles for bone tissue engineering. Biomater Adv 136. https://doi. org/10.1016/j.bioadv.2022.212789
- Corrêa THA, Holanda JNF (2019) Fish bone as a source of raw material for synthesis of calcium phosphate. Mater Res 22:1–5. https://doi.org/10.1590/1980-5373-MR-2019-0486Fish
- Demirtaş TT, Irmak G, Gümüşderelioğlu M (2017) A bioprintable form of chitosan hydrogel for bone tissue engineering. Biofabrication 9:3. https://doi.org/10.1088/1758-5090/aa7b1d
- Derby B (2010) Inkjet printing of functional and structural materials: fluid property requirements, feature stability, and resolution. Annu Rev Mater Res 40(1):395–414. https://doi.org/10.1146/ annurev-matsci-070909-104502

- Didekhani R, Sohrabi MR, Soleimani M, Seyedjafari E, Hanaee-Ahvaz H (2020) Incorporating PCL nanofibers with oyster shell to improve osteogenic differentiation of mesenchymal stem cells. Polym Bull 77:701–715. https://doi.org/10.1007/s00289-019-02750-x
- Dilberoglu UM, Gharehpapagh B, Yaman U, Dolen M (2017) The role of additive manufacturing in the era of industry 4.0. Procedia Manuf 11:545–554. https://doi.org/10.1016/j.promfg.2017. 07.148
- Duan B, Cheung WL, Wang M (2011) Optimized fabrication of Ca–P/PHBV nanocomposite scaffolds via selective laser sintering for bone tissue engineering. Biofabrication 3(1). https:// doi.org/10.1088/1758-5082/3/1/015001
- Elviri L, Foresti R, Bergonzi C, Zimetti F, Marchi C, Bianchera A, Bettini R (2017) Highly defined 3D printed chitosan scaffolds featuring improved cell growth. Biomed Mater 12:4. https://doi. org/10.1088/1764-605X/aa7692
- European Market Observatory for Fisheries and Aquaculture Products (EUMOFA), The EU Fish Market (2020) https://doi.org/10.2771/664425
- FAO (2020) The state of world fisheries and aquaculture 2020, sustainability in action. https://doi. org/10.4060/ca9229en
- Gibson I, Rosen D, Stucker B (2015) Additive manufacturing technologies—3D printing, rapid prototyping, and direct digital manufacturing, *Springer*. https://doi.org/10.1007/978-1-4939-2113-3
- Giubilini A et al (2020) 3D-printing nanocellulose-poly (3-hydroxybutyrate-co-3hydroxyhexanoate) biodegradable composites by fused deposition modeling. ACS Sustain Chem Eng 8(27):10292–10302. https://doi.org/10.1021/acssuschemeng.0c03385
- Goto T, Sasaki K (2016) Synthesis of morphologically controlled hydroxyapatite from fi sh bone by urea-assisted hydrothermal treatment and its Sr 2 + sorption capacity. Powder Technol 292:314–322. https://doi.org/10.1016/j.powtec.2016.01.041
- Goutagny C, Hocquet S, Hautcoeur D, Lasgorceix M, Somers N, Leriche A (2021) Development of calcium phosphate suspensions suitable for the stereolithography process. Open Ceramics 7. https://doi.org/10.1016/j.oceram.2021.100167
- Govindharaj M, Roopavath UK, Rath SN (2019) Valorization of discarded Marina eel fish skin for collagen extraction as 3d printable blue biomaterial for tissue engineering. J Clean Prod 412–419
- Grigoryan B, Sazer DW, Avila A, Albritton JL, Padhye A, Ta AH, Greenfield PT, Gibbons DL, Miller JS (2021) Development, characterization, and applications of multi-material stereolithography bioprinting. Sci Rep 11:3171. https://doi.org/10.1038/s41598-021-82102-w
- Guo Y, Patanwala Huseini S, Bognet B, Ma Anson WK (2017) Inkjet and inkjet-based 3D printing: connecting fluid properties and printing performance. Rapid Prototyp J 23(3):562–576. https:// doi.org/10.1108/RPJ-05-2016-0076
- Hamester MRR, Balzer PS, Becker D (2012) Characterization of calcium carbonate obtained from oyster and mussel shells and incorporation in polypropylene. Mater Res 15:204–208. https://doi. org/10.1590/S1516-14392012005000014
- Hoque ME, Shehryar M, Islam KMN (2013) Processing and characterization of cockle shell calcium carbonate (CaCO3) bioceramic for potential application in bone tissue engineering. J Mater Sci Eng 02:2–6. https://doi.org/10.4172/2169-0022.1000132
- Hull CW (1986) Apparatus for production of three-dimensional objects by stereolitography, US Patent US4575330A
- Islam KN, Bakar MZBA, Ali ME, Bin Hussein MZ, Noordin MM, Loqman MY, Miah G, Wahid H, Hashim U (2013) A novel method for the synthesis of calcium carbonate (aragonite) nanoparticles from cockle shells. Powder Technol 235:70–75. https://doi.org/10.1016/j. powtec.2012.09.041
- Ismail R, Fitriyana DF, Santosa YI, Nugroho S, Hakim AJ, Al Mulqi MS, Jamari J, Bayuseno AP (2021) The potential use of green mussel (Perna Viridis) shells for synthetic calcium carbonate polymorphs in biomaterials. J Cryst Growth 572:126282. https://doi.org/10.1016/j.jcrysgro. 2021.126282

- ISO/ASTM52900:2021(E) (2021) Additive Manufacturing General Principles Fundamentals and vocabulary
- Jafari H, Lista A, Siekapen MM, Gha P (2020) Fish collagen: extraction, characterization, and applications for biomaterials engineering. Polymers (Basel) 12:1–36. https://doi.org/10.3390/ polym12102230
- Jiang L, Zhang J (2017) Biodegradable and biobased polymers. In: Applied plastics engineering handbook. Elsevier, pp 127–143
- Kontárová S et al (2020) Printability, mechanical and thermal properties of poly (3hydroxybutyrate)-poly(lactic acid)-plasticizer blends for three-dimensional (3D) printing, materials. 13:21. https://doi.org/10.3390/ma13214736
- Köpf M, DFD C, Blaeser A, Sen KS, Fischer H (2016) A tailored three-dimensionally printable agarose–collagen blend allows encapsulation, spreading, and attachment of human umbilical artery smooth muscle cells. Biofabrication 8(2):025011. https://doi.org/10.1088/1758-5090/8/2/ 025011
- Li N, Qiao D, Zhao S, Lin Q, Zhang B, Xie F (2021) 3D printing to innovate biopolymers materials for demanding applications: a review. Mater Today Chem 20. https://doi.org/10.1016/j.mtchem. 2021.100459
- Liu J, Sun L, Xu W, Wang Q, Yu S, Sun J (2019) Current advances and future perspectives of 3D printing natural-derived biopolymers. Carbohydr Polym 207:297–316. https://doi.org/10.1016/ j.jcarbpol.2018.11.077
- Luo W, Zhang S, Lan Y, Huang C, Wang C, Lai X, Chen H, Ao N (2018) 3D printed porous polycaprolactone/oyster shell powder (PCL/OSP) scaffolds for bone tissue engineering. Mater Res Express 29. https://doi.org/10.1088/2053-1591/aab916
- Ma MG, Liu S, Fu LH (2016) Calcium carbonate and cellulose/calcium carbonate composites: synthesis, characterization, and biomedical applications. Mater Sci Forum 875:24–44. https:// doi.org/10.4028/www.scientific.net/MSF.875.24
- Maschmeyer T, Luque R, Selva M (2020) Upgrading of marine (fish and crustaceans) biowaste for high added-value molecules and bio(nano)-materials. Chem Soc Rev 49:4527–4563. https://doi.org/10.1039/c9cs00653b
- Mathew GM, Sukumaran RK, Sindhu R, Pandey A (2021) Green remediation of the potential hazardous shellfish wastes generated from the processing industries and their bioprospecting. Environ Technol Innovation 24. https://doi.org/10.1016/j.eti.2021.101979
- Mazzanti V, Malagutti L, Mollica F (2019) FDM 3D printing of polymers containing natural fillers: a review of their mechanical properties. Polymers 11:1094. https://doi.org/10.3390/ polym11071094
- Mohd Abd Ghafar SL, Hussein MZ, Rukayadi Y, Zakaria MZAB (2017) Surface-functionalized cockle shell-based calcium carbonate aragonite polymorph as a drug nanocarrier. Nanotechnol Sci Appl 10:79–94. https://doi.org/10.2147/NSA.S120868
- Möller T, Amoroso M, Hägg D, Brantsing C, Rotter N, Apelgren P, Gatenholm P (2017) In vivo chondrogenesis in 3D bioprinted human cell-laden hydrogel constructs, plastic and reconstructive surgery. Global Open 5(2):1227. https://doi.org/10.1097/GOX.000000000001227
- Monteiro MJM (2014) Caracterização dos resíduos e subprodutos de origem animal da atividade de venda de produtos da pesca e respetivos sistemas de gestão nos mercados municipais de Lisboa. Universidade de Lisboa
- Morris VB, Nimbalkar S, Younesi M, McClellan P, Akkus O (2016) Mechanical properties, cytocompatibility and manufacturability of chitosan: PEGDA hybrid-gel scaffolds by stereolithography. Ann Biomed Eng 45(1):286–296. https://doi.org/10.1007/s10439-016-1643-1
- Murphy SV, Atala A (2014) 3D bioprinting of tissues and organs. Nat Biotechnol 32(8):773–785. https://doi.org/10.1038/nbt.2958
- Naga SM, El-maghraby HF, Mahmoud EM, Talaat MS, Ibrhim AM (2015) Preparation and characterization of highly porous ceramic scaffolds based on thermally treated fi sh bone. CERI 41:15010–15016. https://doi.org/10.1016/j.ceramint.2015.08.057

- Ngo TD, Kashani A, Imbalzano G, Nguyen KTQ, Hui D (2018) Additive manufacturing (3D printing): a review of amterials, methods, applications and challenges. Composites Part B. https://doi.org/10.1016/j.compositesb.2018.02.012
- Nirmal NP, Santivarangkna C, Rajput MS, Benjakul S (2020) Trends in shrimp processing waste utilization: an industrial prospective. Trends Food Sci Technol 103:20–35. https://doi.org/10. 1016/j.tifs.2020.07.001
- Nisticò R (2017) Aquatic-derived biomaterials for a sustainable future: a European opportunity. Resources 6:1–15. https://doi.org/10.3390/resources6040065
- Owuamanam S, Cree D (2020) Progress of bio-calcium carbonate waste eggshell and seashell fillers in polymer composites : a review. J Compos Sci 4. https://doi.org/10.3390/jcs4020070
- Piccirillo C, Silva MF, Pullar RC, Braga I, Jorge R, Pintado MME, Castro PML (2013) Extraction and characterisation of apatite- and tricalcium phosphate-based materials from cod fish bones. Mater Sci Eng C 33:103–110. https://doi.org/10.1016/j.msec.2012.08.014
- Raghavankutty M, Jose GM, Sulaiman M, Kurup GM (2018) Evaluating the biocompatibility of marine-derived chitosan–collagen polymeric blends for biomedical applications. J Bioact Compat Polym 33:439–455. https://doi.org/10.1177/0883911517747892
- Rocha VG, Saiz E, Tirichenko IS, García-Tuñón E (2020) Direct ink writing advances in multimaterial structures for a sustainable future. J Mater Chem A 8(31):15646–15657. https://doi.org/ 10.1039/D0TA04181E
- Rydz J et al (2020) Three-dimensional printed PLA and PLA/PHA dumbbell-shaped specimens: material defects and their impact on degradation behavior. Materials 13(8):2005. https://doi.org/ 10.3390/ma13082005
- Saadi MASR et al (2022) Direct ink writing: a 3D printing technology for diverse materials. Adv Mater 21. https://doi.org/10.1002/adma.202108855
- Sambudi NS, Sathyamurthy M, Lee GM, Bin Park S (2015) Electrospun chitosan/poly(vinyl alcohol) reinforced with CaCO3 nanoparticles with enhanced mechanical properties and biocompatibility for cartilage tissue engineering. Compos Sci Technol 106:76–84. https://doi.org/ 10.1016/j.compscitech.2014.11.003
- Sanchez-Rexach E, Johnston TG, Jehanno C, Sardon H, Nelson A (2020) Sustainable materials and chemical processes for additive manufacturing. Chem Mater 32:7105–7119. https://doi.org/10. 1021/acs.chemmater.0c02008
- Santoni S, Gugliandolo SG, Sponchioni M, Moscatelli D, Colosimo BM (2022) 3D bioprinting: current status and trends—a guide to the literature practice. Bio-Des Manuf 5:14–42. https://doi.org/10.1007/s42242-021-00165-0
- Sanz B, Sanchez AA, Tangey B, Gilmore K, Yue Z, Liu X, Wallace G (2021) Light cross-linkable marine collagen for coaxial printing of a 3D model of neuromuscular junction formation. Biomedicine 9. https://doi.org/10.3390/biomedicines9010016
- Saveleva MS, Ivanov AN, Kurtukova MO, Atkin VS, Ivanova AG, Lyubun GP, Martyukova AV, Cherevko EI, Sargsyan AK, Fedonnikov AS, Norkin IA, Skirtach AG, Gorin DA, Parakhonskiy BV (2018) Hybrid PCL/CaCO3 scaffolds with capabilities of carrying biologically active molecules: synthesis, loading and in vivo applications. Mater Sci Eng C 85:57–67. https://doi. org/10.1016/j.msec.2017.12.019
- Scott Crump S (1992) Apparatus and method for creating three-dimensional objects, US 5121329
- Shavandi A, Hou Y, Carne A, McConnell M, Bekhit AEDA (2018) Marine waste utilization as a source of functional and health compounds. Adv Food Nutr Res 87:187–254. https://doi.org/10. 1016/bs.afnr.2018.08.001
- Stratesteffen H, Köpf M, Kreimendahl F, Blaeser A, Jockenhoevel S, Fischer H (2017) GelMAcollagen blends enable drop-on-demand 3D printability and promote angiogenesis. Biofabrication 9(4):045002. https://doi.org/10.1088/1758-5090/aa857c
- Terzioğlu P, Öğüt H, Kalemtaş A (2018) Natural calcium phosphates from fi sh bones and their potential biomedical applications. Mater Sci Eng C 91:899–911. https://doi.org/10.1016/j.msec. 2018.06.010

- Ullah I, Cao L, Cui W, Xu Q, Yang R, Tang K-I, Zhang X (2021) Stereolithography printing of bone scaffolds using biofunctional calcium phosphate nanoparticles. J Mater Sci Technol 88: 99–108. https://doi.org/10.1016/j.jmst.2021.01.062
- Venkatesan J, Lowe B, Manivasagan P, Kang K-H, Chalisserry EP, Anil S, Kim DG, Kim S-K (2015) Isolation and characterization of nano-hydroxyapatite from Salmon fish bone. Materials (Basel):5426–5439. https://doi.org/10.3390/ma8085253
- Wu C-S (2018) Characterization, functionality and application of siliceous sponge spicules additive-based manufacturing biopolymer composites. Addit Manuf 22:13–20. https://doi.org/ 10.1016/j.addma.2018.04.034
- Wu C-S, Liao H-T, Cai Y-X (2017) Characterisation, biodegradability and application of palm fibre-reinforced polyhydroxyalkanoate composites. Polym Degrad Stab 140:55–63. https://doi. org/10.1016/j.polymdegradstab.2017.04.016
- Wu Z, Su X, Xu Y, Kong B, Sun W, Mi S (2016) Bioprinting three-dimensional cell-laden tissue constructs with controllable degradation. Sci Rep 6:24474. https://doi.org/10.1038/srep24474 Yan N, Chen X (2015) Don't waste seafood waste. Nature 524:155–157
- Yao Z, Xia M, Li H, Chen T, Ye Y, Zheng H (2014) Bivalve shell: not an abundant useless waste but a functional and versatile biomaterial. Crit Rev Environ Sci Technol 44:2502–2530. https:// doi.org/10.1080/10643389.2013.829763
- Zhao Y, Li Y, Mao S, Sun W, Yao R (2015) The influence of printing parameters on cell survival rate and printability in microextrusion-based 3D cell printing technology. Biofabrication 7:4. https://doi.org/10.1088/1758-5090/7/4/045002

# **Chapter 7 Exploiting Marine Fungi in the Removal of Hazardous Pollutants and Biomass Valorisation**



### Dushyant R. Dudhagara, Bhumi M. Javia, and Anjana K. Vala

Abstract The environment and human health are now seriously threatened by pollution. Organic contaminants have a long half-life in the environment and possess hydrophobic, mutagenic, and cytotoxic properties. Hence, they are a big challenge to the health of environment. Due to their hydrophobic and persistent character, non-biodegradable and recalcitrant chemicals have an adverse effect on terrestrial and aquatic ecosystems. According to current pollution management methods, biodegradation and bioremediation using various microorganisms is an efficient, reliable, and eco-friendly approach to combat the pollutants. Marine-derived fungi play a significant role in the remediation of organic pollutants because their unique morphological and physiological properties including survival in the extreme conditions and a diverse metabolic capability. This chapter highlights the role and mechanisms of marine-derived fungi in removal of various pollutants as polyaromatic hydrocarbons (PAHs), heavy metals, dyes, and the biomass valorization. Fungi can oxidize PAHs, alkanes, and other complex hydrocarbons using various intracellular and extracellular enzymatic machineries including monooxygenases and lignin-modifying enzymes. Marine-derived fungi act as biosorbents to remove heavy metal contaminants through active and passive mechanisms. Both living and non-living fungal biomass can be used to detoxify and degrade dyes from the contaminated environment. Furthermore, fungal biomass valorization facilitates the sustainable development of value-added products such as biofuels, enzymes, amino acids, organic acids, alcohol, pigments etc. Thus, the chapter emphasizes to understand the mechanisms of marine-derived fungi in degradation of organic pollutant, metabolic pathways, and enzymes responsible for degradation of organic compounds which would help to develop the future

e-mail: akv@mkbhavuni.edu.in

D. R. Dudhagara · B. M. Javia

Department of Life Sciences, Bhakta Kavi Narsinh Mehta University, Junagadh, India

A. K. Vala (🖂)

Department of Life Sciences, Maharaja Krishnakumarsinhji Bhavnagar University, Bhavnagar, India

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_7

mycoremediation policies and production of value-added products for sustainable future development.

**Keywords** Organic pollutants · Bioremediation · Biomass valorization · Valueadded products · Sustainable development

# 7.1 Introduction

Pollution is a matter of grave concern in this age. The condition of environment directly influences the quality of life in the ecosystem on earth. Industrialization and growing affluence in the developed world along with population explosion and rapid development in the developing countries has resulted in accelerated environmental degradation on a large-scale. Major sources of pollutants include industrial effluents, injudicious use of fertilizers, insecticides, pesticides, mining activities, sewage sludge, etc. Pollutants can be divided into two major types: biodegradable and non-biodegradable pollutants. Non-biodegradable pollutants cause hazardous effect on environment. Non-biodegradable pollutant includes heavy metals, pesticides, polyaromatic compounds and radionuclear material etc. (Peng et al. 2008). Many conventional physico-chemical methods of treatment/removal of these compounds, though effective, are not feasible for application on a large scale. Hence, there is need to develop treatments that can minimize or even eliminate such pollutants from environment. In recent years, the application of microorganisms which degrade or convert hazardous pollutants to less toxic compounds have become popular. Fungi proved to have high potential in the degradation of high molecular weight compounds and therefore are used widely to remediate environmental pollution (Akcil et al. 2015; Deshmukh et al. 2016; Varjani 2017).

Fungi are eukaryotic, chemoheterotrophic, parasitic or saprophytic, unicellular or multicellular filamentous organisms that include molds, yeasts, and mushrooms. The kingdom Fungi includes eight phyla. Fungi are found in variety of habitats like soil, fresh, and marine waters (Anastasi et al. 2013). Fungi produce secondary metabolites, enzymes, biosurfactants, and polysaccharides and applied in bioremediation of pollutants. Fungi have been efficiently used to treat water samples contaminated with micropollutants (Badia-Fabregat et al. 2015). Marine fungi are able to sustain high saline conditions and extreme pH this trait provides biological advantage over terrestrial fungi (Thatoi et al. 2013; Singh et al. 2019).

Marine fungi are found in oceans and colonize different niches. They act as parasites, saprobes, or symbionts and associated with organisms (Wang et al. 2012). Marine fungi can be isolated from different samples such as sediment, seawater, mangrove detritus, decaying wood, seaweeds, and invertebrates (Pang et al. 2016). Marine fungi have capacity to produce different pharmacological metabolites (Imhoff 2016). Marine fungi are source of novel antibacterial, antiviral, anticancer, antiplasmodial, and anti-inflammatory compounds (Bovio et al. 2019), enzymes, biosurfactant (Cicatiello et al. 2016; Nicoletti and Andolfi 2018) and are also useful in the bioremediation of pollutants (Bovio et al. 2017). Marine fungi tolerate high

concentrations of heavy metals such as copper and lead (Gazem and Nazareth 2013). Role of marine fungi in heavy metals, dyes, and hydrocarbons degradation has been well documented. Furthermore, enzymes from marine fungi can be used for paper, pulp, textile, leather, biofuel industries, food and beverages, for animal feed, for pharmaceutical, cosmetic, and environmental applications (Damare et al. 2012; Bonugli-Santos et al. 2015; Deshmukh et al. 2016).

It has been reported that marine fungi have capacity to bioremediate highly recalcitrant pollutants. Bioremediation is a process in which living organisms degrade or convert harmful organic contaminants to less toxic compounds. Mycoremediation is the process in which fungi are used for bioremediation. Mycoremediation is eco-friendly and effective method to combat increasing pollution of soil and water (Arun et al. 2008). Fungi are ideal candidates for remediation of pollutants due to their unique traits including ability to withstand fluctuation in temperature and pH, heavy metal resistance, high surface area to volume ratio, mycelial growth, hyphal network, and extracellular ligninolytic enzymes (Khan et al. 2019). Fungi produce variety of intracellular as well as extracellular enzymes including peroxidase and cytochrome P450, respectively, for detoxification and biodegradation of pollutants (Durairaj et al. 2015). The diversity of habitats and ability for secreting multitude of enzymes makes fungi potential candidates for bioremediation at various locations (Divya et al. 2014).

In this chapter, the role of marine fungi in degrading various recalcitrant, persistent, and harmful pollutants like polycyclicaromatic hydrocarbons (PAHs), heavy metals, dyes, and mechanisms behind the mycoremediation of these pollutants are summarized with process of biomass valorization. An attempt is made to understand how the process of degradation can be accelerated and the future strategy to overcome the existing limitations is discussed.

# 7.2 Hydrocarbon Degradation by Marine Fungi

Hydrocarbon contamination from petrochemical industry is the major environmental problem faced by humanity. Leakage and accidental spillage of petroleum products are to the tune of 2,00,000–6,00,000 metric tons per year (Kvenvolden and Cooper 2003; Das and Chandran 2011). These organic pollutants are carcinogenic and neurotoxic which cause the harmful effects on animal and plant.

Bioremediation or biodegradation by application of natural populations of microorganisms is one of the mechanisms by which hydrocarbon and other pollutants can be removed from the environment (Ulrici 2000). Biodegradation of petroleum hydrocarbons is a complex process which depends on the environment and the amount of the pollutants present in the site. Petroleum hydrocarbons are divided into four classes: saturates, aromatics, the asphaltenes (ketones, phenols, porphyrins, esters and fatty acids), and the resins (sulfoxides, amides, pyridines, carbazole, and quinolones). Hydrocarbons differ in their degradation susceptibility to microbial attack. The susceptibility of hydrocarbons to microbial degradation is linear alkanes > branched alkanes > small aromatics > cyclic alkanes (Ulrici 2000). Some compounds such as high molecular weight containing polycyclic aromatic hydrocarbon may be difficult to be degraded (Atlas 1995) but many fungal species are capable to degrade this recalcitrant hydrocarbon-containing pollutants. It has been observed that fungi present in polluted environment have developed adaptive mechanisms by which they are able to utilize hydrocarbons as the sole source of carbon (Dacco et al. 2020).

Chaillan et al. (2004) isolated fungi *Amorphotheca*, *Talaromyces*, *Neosartorya*, and *Graphium* from the soil contaminated by petroleum and have higher efficiency in the degradation of petroleum hydrocarbons. Some terrestrial fungi *Aspergillus*, *Penicillium*, and *Cephalosporium* are reported as idea candidate for the degradation of crude oil (Singh 2006; Das and Chandran 2011). The different hydrocarbon degrading pathways and their mechanisms operated by fungi are mentioned below.

### 7.2.1 Degradation Process of Alkane

Alkane is a saturated hydrocarbon with all single bonds in its structure and cycloalkane is a saturated hydrocarbon with several carbon rings in its structure. Alkane degradation can be catalyzed by some enzymes such as oxidase, dehydrogenase and converted into fatty acids, and followed by acetyl coA and which can be further mineralized as  $CO_2$  and  $H_2O$  (Singh 2006). It has been observed that in the process of Alkane degradation many enzymes played an important role such as alkane monooxygenase, fatty alcohol dehydrogenase, fatty aldehyde dehydrogenase, etc. Biological degradation of alkane is a subterminal oxidation process. In the primary step, cycloalkane is oxidized into alcohols by n-alkane monooxygenase type of oxidizing enzymes. Then, alcohols are converted into fatty acid. Cyclohexane is converted into corresponding compound, followed by cyclohexanol, cyclohexanone, and fatty acids. At last, the compound is mineralized and form  $CO_2$  and  $H_2O$  along with fungal biomass production as the end product (Fig. 7.1) (Dacco et al. 2020).

# 7.2.2 Degradation Process of Polycyclic Aromatic Hydrocarbons (PAHs)

Polycyclic aromatic hydrocarbons (PAH) are widely distributed in the environment and may persist for extended period of time. PAHs are composed of two or more fused benzene rings and are formed during combustion of organic molecules (Haritash and Kaushik 2009). Polycyclic aromatic hydrocarbons have carcinogenic, mutagenic, and teratogenic properties. Forest, oil seep, volcanic eruption and



Fig. 7.1 Degradation shows the terminal, diterminal, and subterminal pathways for the n-alkane (modified from Dacco et al. 2020)

exudates from trees are natural sources of PAHs. Anthropogenic sources include fossil fuel burning, coal tar, wood, garbage, lubricating oil, municipal waste incineration and petroleum spills, etc., are the major source of pollutants (Kaushik and Haritash 2006). Most important and ecologically damaging components of pollution are the PAHs and cause depletion of the ozone layer and affect Earth's heat balance, adding acidic air pollutants to atmosphere and reduce visibility (Chauhan et al. 2000).

According to Yumoto et al. (2002), PAHs can be degraded by enzymes and catalyzed into glycol or catechol, then decomposed into succinic acid or acetyl coA. In degradation pathway, PAHs are gradually degraded into epoxide, trans diol, phenol, and trans dihydro 2 phenol by monooxygenase enzyme of yeast. In another study of Mills et al. (2004), PAHs can be degraded by dioxygenase into cis dihydro 2 phenol, epoxide, cis diol etc. The final metabolites in both pathways are  $co_2$  and water. The ligninolytic and monooxygenase system of cytochrome P-450 may be involved in polycyclic aromatic hydrocarbon degradation by filamentous fungi. Induction of the monooxygenase before application in degradation of hydrocarbon could result in enhanced removal of PAHs (Haritash and Kaushik 2009). Fungal cytochrome P-450 Monooxygenase from white rot fungi *Phenerochaete* 

*chrysosporium* have capacity to oxidize pollutants like aliphatic hydrocarbons, crude oil, n-alkane, polyaromatic hydrocarbons alkylphenol, etc.

PAHs degradation depends on solubility, number of benzene rings, the species and number of substituent species, the properties of heterocyclic atom of PAHs. Asphalt is very difficult to be degraded by biodegradation due to its most complicated structure. Many researches indicated that the PAH can be degraded only in aerobic conditions. While it has been observed that PAH can be degraded in anaerobic condition as well like sulfate reduction, denitrification, or methanogenic fermentation. Though rate of aerobic degradation of PAHs is higher than the rate of anaerobic degradation. (Meckenstock Rainer 2004).

Ligninolytic fungi are capable of oxidizing PAH by non-specific extracellular enzymatic complexes, normally used to depolymerize lignin. These lignindegrading enzymes include laccase, lignin peroxidase, manganese peroxidase, etc. (Peng et al. 2008). A novel PAH metabolic pathway in fungi involves hydroxylation by cytochrome P-450 monooxygenase enzyme through a sequence of reactions mammalian metabolism (Capotorti et al. 2004). similar to In many non-ligninolytic fungi, this pathway occurs to effectively degrade hydrocarbons (Ravelet et al. 2000). Many researchers have studied that purified fungal laccase enzyme can be used for the oxidation of PAH. Laccase enzyme of T. versicolor, C. hirsutus, P. ostreatus, and Coriolopsis gallica was the most studied in the fungi. It has been reported that activity of enzyme in T. versicolor fungi is 29 times higher than other microorganisms (Margot et al. 2013). For example, T. versicolor laccase, in combination with 1-hyrdoxybenzotriazole (HBT), was capable to oxidize two PAHs, acenaphthene and acenaphthylene; Laccasewithout mediator oxidized about 35% of the acenaphthene and only 3% of acenaphthylene. The end products obtained after incubation were 1,2-acenaphthenedione and 1,8-naphthalic acid anhydride (Johannes et al. 1998).

Kirk and Gordon (1988) explained that 14 strains of obligate marine fungi belonging to genus, *Varicosporina, Dendryphiella, Lulworthia*, and *Corollospora* species can grow using alkenes and alkanes as a sole source of carbon and mineralized into n-hexadecane. The study has shown that the 14 lignicolous and arenicolous strains utilized pristine, 1-hexadecene and some degree of tetradecane as a sole source of carbon. Raikar et al. (2001) isolated*Thraustochytrids* fungi from several oil spills polluted sites in Goa and they were capable of degrading tar-balls added to peptone broth and degradation was observed up to 30% in 7 days as estimated by gas chromatography and gravimetry. *A.sclerotiorum* showed 99.7% pyrene 2 mg in 30 mL and 76.6% benzo pyrene 1 mg in 30 mL degradation after 8 and 16 days, respectively (Passarini et al. 2011). Two non- identified marine-derived fungi were able to remove phenanthrene from a media by adsorption through fungal mycelium. Fungus *Aspergillus* sp. BAP14 isolated from marine sediment showed degradation of benzopyrene and removed approx 30% BaP after 3 days (Raghukumar et al. 2006; Damare et al. 2012).

In the degradation process of aromatic hydrocarbons, aromatic hydrocarbon is oxidized by oxidase into dihydrodiol. Then, the dihydrodiol is degraded into o-dihydroxybenzene. Dihydroxybenzene is degraded by following two processes,



Fig. 7.2 Degradation of polycyclic aromatic hydrocarbons by fungi

which are ortho position ring opening and meta position ring opening reaction. Then, these compounds are oxidized into long chain compounds and gradually metabolized into acetyl coA. Degradation process by microorganisms such as bacteria and fungi are comparatively different from each other. In bacteria, aromatic hydrocarbon is oxidized by two oxygen atoms and converted into cis-dihydrodiol. While in fungi, aromatic hydrocarbon is oxidized and converted into trans-dihydrodiol (Fig. 7.2) (Xue et al. 2015; Kadri et al. 2017).

### 7.3 Heavy Metal Removal by Marine Fungi

Heavy metals are considered as one of the most hazardous pollutants having a specific density of more than 5 gm/cm<sup>3</sup>. Heavy toxic metals are directly or indirectly released into environment and as a result, annual worldwide release of heavy metals reached 13,50,000 tons of zinc, 9,39,000 tons of copper, 22,000 tons of cadmium and 7,83,000 tons of lead (Singh et al. 2003). Metals like iron, zinc, copper, and manganese are essential for biological process. While mercury, cadmium, and nickel have no physicological role but these metals can cause harmful disorders at high concentration (Lenin et al. 2014). High concentration of heavy metals can cause serious environmental as well as health problems. Unlike organic contaminants, pollutants from heavy metals are non-biodegradable these and cause bioaccumulation in food chain. Conventional physico-chemical treatment technologies become less effective and more expensive when metal concentrations are in the higher range (1–100 ppm) (Dermont et al. 2008).

Marine fungi can tolerate high concentration of heavy metals and its their interaction to metal ions can be used to remove heavy metal pollutants from environment (Lopez Errasquín and Vázquez 2003). Marine fungi can remove toxic metals from the environment by adsorption as well as their metabolic activities (Davis et al. 2003). Living as well as dead fungal biomass has been recognized for the removal of heavy metals through absorption (Bishnoi and Garima 2005). Fungi can be used as a biosorbent for the removal of heavy metals with excellent metal uptake and recovery (Fu et al. 2012). Rehman et al. (2008) isolated yeast *Lodderomyces elongisporus* from metal-contaminated site and found to tolerate various heavy metals. Damare et al. 2012 stated that the fungus *Thraustochytrids* from shallow water hydrothermal vents have efficiency to withstand high concentration of heavy metals. Majeau et al. (2010) reported that psychrophilic fungi, *Cryptococcus* sp. found in deep-sea sediments have capability to tolerate high concentration of heavy metals such as ZnSO<sub>4</sub>, CuSO<sub>4</sub>, Pb (CH<sub>3</sub>COO)<sub>2</sub> and CdCl<sub>2</sub> up to 100 mg/L.

Vala et al. (2004) while searching for new sources of marine fungi for the removal and tolerance of heavy metal confirmed two seaweeds associated fungi *Aspergillus flavus* and *A. niger* for their tolerance potential against hexavalent chromium. Both the confirmed fungi had remarkable chromium tolerance and removal capability. Chromium removal (mg/g dry wt) was noticed to increase with increasing chromium concentrations.

Khambhaty et al. (2009) isolated three marine-derived aspergilli viz. *Aspergillus niger*, *A. wentii* and *A. terreus* from Gujarat coastal area and were examined for their hexavalent chromium (Cr(VI))removal capacity. Out of the three, *A. niger* was monitored as the most potential candidate for Chromium removal. Complete analyses of biosorption and sorption capability discovered 117.33 mg/g adsorption by *A. niger* under optimized conditions and sorption efficiency was noticed to be 100%. Biosorption process was observed to be endothermic. On the basis of FTIR analysis, amino, methylene, hydroxyl, and phosphorous groups were involved in binding of chromium to fungal biomass.

Taboski et al. (2005) evaluated the toxicity level of Cadmium (Cd) and lead (Pb) to two fungal species *Corollospora lacera* and *Monodictys pelagica* isolated from the marine environment by exploring their growth rate and biomass. Biosorption of metals was also checked. Growth rate of fungi was not affected by lead, though, high cadmium concentration reduced the growth rate of fungi, particularly, *M. pelagica*. About 93% of extracellular lead segregation by *C. lacera* was observed. *M. Pelagica* accumulated about 60 mg/g Cd and about 6 mg/gPb. About 7 mg/g Cd and up to 250 mg/g Pb was accumulated by *C. lacera*.

Khambhaty et al. (2009) studied dead fungal biomass of four marine *Aspergillus* species for Hg(II) biosorption and noticed *Aspergillus niger* as the most efficient Hg (II) biosorbent. Dead biomass of *A. niger* showed 40.53 mg/g Hg(II) removal under optimized conditions. Assessment of possible cell-metal ion interaction disclosed involvement of hydroxyl(–OH) and amino (NH<sub>2</sub>) groups present on the cell surface in Hg(II) biosorption.

El-Kassas and El-Taher (2009) isolated hexavalent chromium (Cr(VI)) tolerant strain of *Trichoderma viride* from water samples of the Mediterranean Sea. The fungus could remove 4.66 mg/g Cr(VI). On the basis of transmission electron microscopic (TEM) analysis, it was observed that accumulation of chromium by the fungus did not affect its mycelial and conidial structures. Mendoza et al.(2010) studied that two marine fungal strains of *Dendryphiella salina* were observed to absorb 80–92% Hg<sup>2+</sup> from the liquid media. Strain Den32 had higher absorption

		Initial Metal Ion Concentration	Sorption capacity	
Fungal Biosorbent	Metal	(mg/L)	(mg/g)	References
Aspergillus Niger	Cu	100	15.6	Dursun et al. (2003)
	Pb	100	34.6	
	Cr	50	6.6	
	(VI)			
Botrytis cinerea	Pb	350	107.1	Akar and Tunali (2005)
Phanerochaete	Cu	100	88.16	Iqbal and Edyvean (2004)
chrysosporium	Pb	100	68.73	
	Zn	100	39.62	
Pleurotus platypus	Ag	200	46.7	Das et al. (2010)
Rhizopus oryzae	Cu	100	34	Fu et al. (2012)

 Table 7.1
 Metal biosorption by marine fungal biosorbents (Ayangbenro and Babalola 2017)

efficiency than strain Den35. The study disclosed the potential application of both the strains for bioremediation of mercury, mainly through biosorption.

Vala and Upadhyay (2008) isolated arsenic tolerating *Aspergillus* sp. from coastal waters of Bhavnagar, Gulf of Khambhat, West coast of India, disclosed the fungus to tolerate supplied 100mg/L As (III) or As(V). Hydride generation atomic absorption spectrometric (HGAAS) analysis revealed higher removal of As (V) than As(III). Energy Dispersive X-ray spectroscopic (EDX) data further confirmed the presence of arsenic in fungal biomass.

Vala (2010) explored removal and tolerance of arsenic by *Aspergillus candidus* isolated from coastal waters of Bhavnagar, Gulf of Khambhat, West coast of India. The fungus showed tolerance for the trivalent and pentavalent forms of arsenic (25 and 50 mg/L). Maximum arsenic removal (mg/g) by the fungus was observed on third day. Vala (2010) also suggested that facultative marine fungus *A. candidus* was one of the most promising fungus for bioremediation of arsenic.

Vala (2010) and Vala et al. (2011) reported that *Aspergillus flavus* and *A. niger*, facultative marine fungi, have tolerance and removal capability for arsenic. Vala (2010) has reviewed *A. niger* as potential biosorbent. This perception was supported by marine-derived fungus *A. niger*. Vala and Patel (2011) explained that heat-killed biomass of marine-derived *A. niger was* studied for its As(III) biosorption capacity, it was noted to remove more than 90% of provided As(III) concentrations. Highest biosorption was found108.083 mg/g at the concentration 600 mg/L. (Table 7.1 depicts the sorption capacity of fungi for various metal ions).

Vala and Sutariya (2012) explored the amount of arsenic tolerance and removal efficacy of two facultative marine fungi *A. flavus* and *Rhizopus* sp. Upon exposure to 25 mg/L and 50 mg/L sodium arsenite (As (III)), both the fungi showed arsenic tolerance and accumulation. A little better accumulation was observed by *Rhizopus* sp. Increase in accumulation was observed with increasing concentration representing higher complexation rates between arsenic and arsenic complexing group on the fungal biomass.

Yeasts from the marine environment have been less studied for heavy metal removal. Strains of *Yarrowia lipolytica* have been reported as potential hexavalent chromium remediators by several workers (Rao et al. 2013; Imandi et al. 2014). Likewise, marine yeast *Rhodotorula rubra* has been exploited for arsenic metabolism (Cullen and Reimer 1989; Maher and Butler 1988). Though, arsenic remediation by marine yeasts has not been attended much attention in the recent past (Vala and Dave 2017). Abe et al. (2001) isolated thirteen yeast strains from deep-sea sediment samples of Japan Trench. Among them, *Cryptococcus* sp. was observed to have the maximum tolerance for  $Cu^{2+}$ . The authors also suggested the importance of enzyme superoxide dismutase (SOD) to resist high  $Cu^{2+}$  stress.

Deep-sea psychrotolerant yeast isolates *Cryptococcus* sp., when grown in presence of various concentrations of heavy metal salts viz.  $CdCl_2$ ,  $CuSO_4$ , Pb  $(CH_3COO)_2$ , and  $ZnSO_4$ , demonstrated remarkable growth in the presence of 100 mg/lmetal concentrations. Tolerance to these metals showed by the isolate was comparatively higher than other deep-sea and terrestrial yeasts. Modification in the cell morphology was observed in presence of heavy metals. The yeast can remove 30–90% of the provided heavy metals. The authors recommended the *Cryptococcus* sp. as a potential candidate for bioremediation of heavy metal-contaminated sites. The authors postulated the metal-tolerant property and characteristics of the yeast for the contribution to its ecological role and adaptations in extreme environments (Singh et al. 2013).

Oyetibo et al. (2015) isolated both resting and growing cells of mercury-resistant *Yarrowia* spp. from estuarine sediments polluted with mercury. The resting cells of yeast strain were recommended to be applicable as a reusable bioadsorbent, whereas the growing cells were recommended to be more suitable as efficient mercury bioreduction and volatilization agent.

Srivastava and Thakur (2006) reported efficiency of Aspergillus sp. for the removal of chromium in tannery wastewater. 85% chromium can be removed at 6 pH in a bioreactor by using synthetic medium, compared to 65% removal from the effluent. Lakkireddy and Kues (2017) studiedCoprinopsis atramentaria for its ability to accumulate 76% cadmium at concentration 1 mg/L and 94.7% of lead at concentration 800 mg/L. Study of Park et al. (2005) suggested that dead fungal biomass of A. niger, Rhizopus oryzae, Saccharomyces cerevisiae, and Penicilliumchrysogenum can be used to convert toxic metal Cr (VI) into less toxic or non-toxic Cr (III). It has been reported that catalase enzyme provides heavy metal tolerance capacity to fungi such as lead, copper, zinc, cadmium, etc. It has been observed A. niger, Rhizopus, and Penicillium fungi produce high amount catalase in the presence of heavy metals such as  $Pb^{2+}$ ,  $Cu^{2+}$  (Thippeswamy et al. 2014). Aspergillus foetidus has capacity to tolerate lead (Pb) concentration up to 200 mg/ L and can produce antioxidative enzymes including catalase for detoxifying  $H_2O_2$ and malondialdehyde. Aspergillus spp. Have oxidative stress tolerance for heavy metals like zinc and Copper. (Chakraborty et al. 2013; Mitra et al. 2014; Deshmukh et al. 2016).

Luna et al. (2016) stated that *Candida sphaerica* can produce biosurfactants with the removal of 95%, 90%, and 79% for Fe, Zn, and Pb, respectively. According to

Mulligan et al. (2001) biosurfactants have been widely used in recent years because of their low toxicity, biodegradability, and diversity. Surfactin, rhamnolipid, and sophorolipid can be used for the removal of copper and zinc. A single wash of 0.5% rhamnolipid can remove 65% copper and 18% zinc, while 4% sophorolipid can remove 25% copper and 60% zinc. Chatterjee et al. (2012) reported that *Hansenula polymorpha, S. cerevisiae, Yarrowia lipolytica, Rhodotorula pilimanae, Pichia guilliermondii*, and *Rhodotorula mucilage* can be used to convert toxic Cr (VI) to less toxic Cr (III).

Over the past few years, many bioremediation technologies have been applied all over the world to solve the problem of contaminated environment. Many research and review articles on these technologies for bioremediating heavy metals are available (Khan et al. 2004). Though, there are several gaps in the understanding of heavy metals bioremediation specifically because of the great complexity of soil chemistry. Therefore, expansive and site-specific research is still required to bring out the optimum performance from the technologies of fungal remediation.

# 7.3.1 Mechanisms of Mycoremediation for the Removal of Heavy Metals

Mycoremediation of heavy metals involves interaction between fungi and metal. Bioremediation process includes adsorption, precipitation, oxidation, reductions, and complexation reactions. The chemical reactions between microorganisms and metals can be categorized into six different processes: intracellular accumulation, cell wall-associated metals, extracellular mobilization or immobilization of metals, metal siderophore interactions, extracellular polymer-metals interaction with transformation and volatilization of metals (Davies and Bennett 1983; Siddiquee et al. 2015).

Gadd (2007) demonstrated that Fungi can use three possible strategies for toxic metal removal: (1) Active metal bioaccumulation in fungal cell and storage in vacuoles and/or passive metals bioabsorption on fungal wall; (2) Metalmobilization/transformation/immobilization in the external environments, due to metabolites and secondary organic acids production; (3) metal exclusion.

Gadd (2007) also mentioned that fungi can restrict entry of toxic metal into cells by these three mechanisms: (1) reduced metal uptake and/or increased metal efflux; (2) metal immobilization, e.g., cell wall adsorption, extracellular precipitation of secondary neoformed minerals (e.g., oxalates); (3) extracellular metal sequestration by, e.g., exopolysaccharides and other extracellular metabolites. Five different mechanisms for the heavy metal removal in fungi are mentioned below; (Fig. 7.3 depicts the mechanisms for the removal of heavy metals).



Fig. 7.3 Mechanisms of mycoremediation for the removal of heavy metals (modified from Siddiquee et al. 2015)

### 7.3.1.1 Mobilization of Metals

Mobilization of metals by fungi takes place due to the production and secretion of fungal products like citric acid, a metal ions chelator and oxalic acid that can interact with metal ions to form insoluble oxalate, which can be resulted from the dissolution of primary metals containing phosphate. These organic acids produced by fungi can increase the metal solubility by acidification and formation of metal-complex structure (Siddiquee et al. 2015).

### 7.3.1.2 Biosorption to Cell Wall

Fungal cell wall is the protective layer and barrier that controls uptake of toxic metals into the cell and the first cellular components that interact with metal. Heavy metal affects fungal growth and metabolism results in uptake of metals through chemisorption processes (Kapoor and Viraraghavan 1997), which includes adsorption coupled reduction process, ion exchange, precipitation, crystallization, and electrostatic interaction (Pundir et al. 2016). *Aspergillus niger* strains have been found better in biosorption capabilities of heavy metals such as Cu<sup>2+</sup>, Zn<sup>2+</sup> and Ni<sup>+2</sup> at 4 to 6 pH (Siddiquee et al. 2015).

#### 7.3.1.3 Metal Uptake and Translocation through Cell Membrane

Transporters are located on the cell wall of fungi and they are responsible to uptake essential metals. Carriers may consist of all the metabolically-coupled and H<sup>+</sup> gradient-driven transporter system (Siddiquee et al. 2015). Transportation of heavy metals into the cell from the extracellular environment using active or passive

transport mechanism through the cell membrane requires for the removal of heavy metal ions from the environment. Transportation system requires energy for the removal of heavy metals from the aqueous solution (Veglio and Beolchini 1997).

#### 7.3.1.4 Intracellular Metal Immobilization

Intracellular metal immobilization includes two processes that are vacuoles compartmentation and complexation by cytoplasmic protein, called metallothioneins and phytochelatins (Siddiquee et al. 2015). Fungal vacuole plays important roles in molecular degradation, storage of metabolites, regulation of cytosolic concentrations of metal ions and detoxifies potentially toxic metal ions. Metal-tolerant fungi can survive due to their abilities of intracellular chelation, for example, metallothioneins, phytochelatins, and metal sequestration within vacuoles (Liu and Culotta 1999). Metallothioneins is a metal-binding protein that can modulate the intracellular concentrations and bind both the essential metals such as Cu and Zn and inessential metals such as Cd.

### 7.3.1.5 Metal Transformations

In fungi, biotransformation of metal occurs through chemical reactions such as oxidation, reduction, methylation, and dealkylation. These reactions convert metal ion into non or less toxic form. Chemisorption involves bond formation in chemical group (hydroxyl, amine, phosphoryl, thiol, etc.) present on fungal cell wall or on the surface and substrate to be adsorbed. Chemisorption excludes dependency on metabolic reaction within fungal cell (Bhainsa and D'Souza 2009) and forms strong bonding than ion exchange (Sheoran and Sheoran 2006). Metals may transfer to other parts of the fungi mycelium and plant symbionts by cytosplasmic vesicles and vacuoles. Condensation of heavy metal concentration in the absence of water molecules known as precipitation that helps in removal of contaminants in bulk amount (Gibert et al. 2005; Siddiquee et al. 2015).

# 7.4 Dye Degradation

Dyes are synthetic chemicals and recalcitrant in nature. More than 1,00,000 commercial dyes including acidic, basic, reactive, azo, and anthraquinone-based dyes are produced every year (Campos et al. 2001). Synthetic dyes are widely used in textile dyeing, color photography, paper printing, food, pharmaceutical, cosmetic, and leather industries. Among various industries, the textile dying industries discharge large amount of wastewater effluent after dyeing process. More than  $7 \times 10^5$  metric tons of dyes are produced worldwide yearly (Supaka et al. 2004). The amount of dyes that does not bind to the fibers, enters into wastewater during textile processing (Rai et al. 2005). It has been estimated that 2,80,000 tons of textile dyes are discharged in textile industrial effluents every year worldwide (Jin et al. 2007). Many dyes are visible in water at concentration as low as 1 mg/L (Sandhya 2010). Synthetic dyes can cause environmental pollution and serious health-risk factors due to large-scale production and extensive application (Forgacs et al. 2004).

Based on the chemical structure of the chromophoric group, dyes are classified as azo, triphenylmethane, anthraquinone, polymeric, and heterocyclic dyes. The versatile triphenylmethane and azo dyes account for most textile dyes (Yang et al. 2009). Azo dyes are characterized by the presence of one or more azo bonds [-N=N-] with aromatic ring. Different substitutions on aromatic nucleus give structurally different and versatile group of compounds which makes them recalcitrant and xenobiotic compound (Khan et al. 2013; Jain et al. 2012). Many dyes contain known carcinogens such as benzidine and other aromatic carcinogens (Singh 2006b).

In aquatic environment, dyes can interfere with photosynthetic activities of aquatic flora, diffusion of gases and badly affect food source of aquatic organisms and are of human health concern also. Dye forms thin layer over the surface of a water and thus decreases the amount of dissolved oxygen in the water, therefore adversely affects the aquatic flora and fauna. Dye-containing effluent increases biochemical oxygen demand of the contaminated water (Ciullini et al. 2008; Annuar et al. 2009; Ali 2010). Thus, nowadays degradation of dye is major point of concern.

Due to complex chemical nature, most of the synthetic dyes are highly resistant to degradation (Lin et al. 2010). Physical and chemical treatment methods such as precipitation, coagulation, adsorption, flocculation, flotation, electrochemical destruction, and mineralization and decolorization process have some disadvantages such as cost, time, and release of residues. All these techniques are minimizing the toxicity level but not neutralizing the toxicity. To replace these techniques, biodegradation can be used to completely degrade the dyes (Pandey et al. 2007).

Dye-contaminated industrial effluent is usually treated by physico-chemical processes include membrane filtration, precipitation, ion exchange, flocculation, flotation, ozonation, electro flotation, irradiation, and adsorption using activated carbon or by using bacteria, fungi, algae, plant biomass or other biological material (Robinson et al. 2001). Both living as well as dead cells are used for biosorption. Biodegradation is the most efficient method to remove dyes from industrial effluent and is energy-dependent process that involves the breakdown of dye into various byproducts by action of various enzymes such as laccase, azo reductase, peroxidase, and hydrogenase (Fu and Viraraghavan 2001).

Decolorization of the dye occurs when the chromophoric center of the dye is cleaved (Kaushik and Malik 2009). In biosorption process, the original structure of the dye remains intact and not degraded into fragments. Biosorption plays important role in the decolorization of dye by living fungi (Fu and Viraraghavan 2001). Several microorganisms, including bacteria, fungi, yeasts, and algae, can decolorize and completely mineralize many azo dyes under certain environmental conditions (Pandey et al. 2007). Both live as well as dead fungal biomasses can be utilized to remove dyes from the contaminated ecosystem.

# 7.4.1 Types of Toxic Dyes

During the manufacture and processing of textiles many different chemical reagents, such as acids, bases, water softeners, salts, and organic solvents dyes are utilized. From the 12 classes of chromogenic groups, azo dyes are largest group of synthetic colorants and the most common synthetic dyes released into environment (Zhao and Hardin 2007) followed by the anthraquinone type. They are widely used in the textile, food, pharmaceutical, cosmetics, plastics, paint, ink, photographic, and paper industries. Different types of dyes include azo, direct, acidic, basic and anthraquinone etc., are mentioned below with its structure and molecular weight in Table 7.2. (Pande et al. 2019).

### 7.4.2 Dye Decolorization and Degradation by Marine Fungi

Marine-derived fungi, due to their adaptability to extreme conditions are better suitable in treatment of colored effluents than their counterparts in the terrestrial environment (Raghukumar 2004; Bonugli-Santos et al. 2015; Vala and Dave 2017). Fungi growing under marine conditions have adapted to grow under saline and alkaline conditions since the pH of seawater ranges from 7.5 to 8.2. *Alternaria tenuissima* (El Aty et al. 2017), *Cerrena unicolor* (D'Souza-Ticlo et al. 2009), *Aspergillus niger* (Lu et al. 2016; Joshi et al. 2012), *Flavodon flavus, Penicillium janthinellum* (Wang et al. 2015), *Peniophora* sp. (Bonugli-Santos et al. 2015), *Tinctoporellus* sp. (CBMAI 1061), *Marasmiellus* sp. (CBMAI 1062), and *Peniophora* sp. (CBMAI 1063) (Bonugli-Santos et al. 2012) *Phialophora* sp. (MF 6), *Penicillium* sp. (MF 49), and *Cladosporium* sp. (Torres et al. 2011) are some examples of potential dye degrading fungi.

It has been reported that marine-derived fungi have potential for the decolorization of textile effluents and synthetic dyes such as Congo red, Brilliant green and RBBR (Raghukumar 2004, Raghukumar et al. 2008; D'Souza et al. 2006). According to Arun et al. (2008) the lignolytic extracellular enzymes produced by filamentous fungi have great relevance in bioremediation of toxic dyes. Bartlett (1971) illustrated that Some dyes are used as indicators for production of lignolytic enzymes and also play a very important role in bioremediation of lignin-based derivatives in colored industrial pollutants such as paper and pulp mills, textile mills, tanneries, and molasses-based distilleries.

Baccar et al. (2011) explained that White rot fungi such as *Trametes versicolor*, *Ganoderma lucidum*, and *Irpex lacteus* were evaluated for decolorization of Tannery Dye Black Dycem TTO and suggested adsorption and biodegradation as a key mechanism for removal of dye. Aksu et al. (2007) analyzed that *Trichoderma versicolor* biomass can be utilized for biosorption of Remazol Black B reactive dye. Laccase produced from Marine fungi can decolourize and mineralize high concentrations of pollutants (Vishwanath et al. 2014). *Myceliopthora thermophila* 

Table 7.2 Chemical structures of toxic dytempoint	es used in the textile industry (modified from Pande et al. 2019)	
Types of dyes	Structure	M.Wt.
Azo dye	OH N	248.28
Direct yellow 4	HO HO N N N N N N N N N N N N N N N N N	624.55
Direct blue 98	SO <sub>3</sub> Na N OH N HO N N N N N N N N N N N N N N N	928.79
Acid dye	Ho No So <sub>3</sub> Na So	400.38



(*Ascomycetes*) have capacity for decolourization of flexographic inks in presence of mediators (Fillat et al. 2012; Deshmukh et al. 2016).

Young and Yu (1997) stated that Binding of dyes to the fungal hyphae, physical adsorption and enzymatic degradation by extracellular and intracellular enzymes are major mechanisms for the dye degradation (Young and Yu 1997). White rot fungi produce lignin peroxidase, manganese peroxidase and laccase that degrades many aromatic compounds due to their non-specific enzyme systems (Robinson et al. 2001; Madhavi et al. 2007). The predominantly reported enzymes for dye degradation are azoreductase, laccases, lignin peroxidase, manganese peroxidase, and hydroxylases. Azoreductase and Laccase have been shown to degenerate azo dyes (Rodrigue et al. 1999).

Chivukula and Renganathan (1995) stated that laccase enzymes can degrade the azo dye through a non-specific free radical mechanism to form phenolic compounds and prevent the formation of toxic aromatic amines. Manganese peroxidase has been reported as the main enzyme involved in dye decolorization by fungus *Phanerochaete chrysosporium* (Chagas and Durrant 2001). In fungus *Bjerkandera adusta*, lignin peroxidase has been reported as important enzyme in dye degradation (Robinson et al. 2001).

Dwivedi and Singh Tomar (2018) explained that *A.allhabadii* and *A. sulphureus* have higher decolorization capacity up to 95.13% and 93.01%, while *A. niger* has little lesser 83% decolorization capacity. Namdhari et al. (2012) explained that decolorization of azo and anthraquinonic dyes can be achieved by brown rot fungi such as *Coprinus micaceus, Fomtopsispinicola,* and *Gloeophyllum odoratum.* Saranraj et al. (2010) isolated *Aserpgillus niger, Aspergillus flavus, Aspergillus fumigatus, Trichoderma viride, Fusarium oxysporum, Penicillium chrysogenum* and Mucor sp. that are responsible for the degradation of a wide range of textile dyes.

Huanga et al. (2016) modified *Aspergillus versicolor* by using cetyl trimethyl ammonium bromide (CTAB) to enhance the fungal biosorption of Reactive Black 5 at various physiochemical conditions. Basidiomycetes fungi have been reported to decolourize dye by adsorption to the mycelial surface and further metabolic breakdown by both batch mode and continuous mode. *Schizophyllum commune* has found to be more efficient than *Lenzites eximia* for the treatment of azo dyes and textile dye industry effluent, (Selvam and Shanmuga Priya 2012). Yesilada (1995) analyzed that *Coriolus versicolour* and *Funalia trogii*are responsible for decolorization of crystal violet dye. Versatile peroxidase and lignin peroxidase have ability to oxidize non-phenolic aromatic compounds, reactive Black B dye and low redox potential was observed as seen from oxidation of phenolic substrates (Karigar and Rao 2011; Deshmukh et al. 2016).

Ollikka et al. (1993) described that isozymes of lignin peroxidase found in *Phanerochaete chrysosporium* has capability for decolourization of synthetic dye. *Phanerochaetechrysosporium* has been observed to degrade many dyes such as polymeric dyes, azo dyes, heterocyclic dyes, and crystal violet dyes. The fungus *P. Chrysosporium* can aerobically degrade three azo dyes includes congo red, orange II, and tropaeolin (Cripps et al. 1990).

Bonugli-Santos et al. (2015) studied that the ascomycetes and basidiomycetes can decolorize 30 to 60% of azo dye-containing effluent and 33 to 80% decolorization of mixture of 8 reactive dyes under saline conditions. Bucher et al. (2004) illustrated that decolorization of azure B is possible due to the production of ligninolytic peroxidase by fungi Rhizophila marina, Bathyascus grandisporus, Verruculina enalia, and Cryptovalsa halosarceicola. It has been found that Penicillium citrinum CBMAI 853 is the most efficient fungus that decolorizes RBBR (100%) after 12 days, A. sulphurous CBMAI 849 (95%), Cladosporium cladosporioides CBMAI 857 (93%), and Trichoderma sp. CBMAI 852 (89%) (Da Silva et al. 2008). Molitoris et al. (2000) isolated filamentous halophilic fungi Gymnoscella marismortui from the Dead Sea which is responsible for the decolorization of synthetic dye belonging to 4 different groups. Lalitha et al. (2011) explained that marine Aspergillus flavus has capacity for the bioremediation of synthetic, color photographic and paper mill dyes and can remove 80% and 90% synthetic dyes and 100% of color. It has been observed that sponge-derived basidiomycetes fungi have ability to decolorize textile dyes in solid medium under both saline as well as non-saline condition (Bonugli-Santos et al. 2012; Vala et al. 2018).

Raghukumar et al. (2008) explained that whole cell immobilization of marinederived fungi *Penicillium janthinellum* P1 and *Pestalotiopsis* sp. J63 showed the decolorization of Azure B dye. 70% decolorization of MSW in five days has been observed when marine-derived fungus NIOCC #312 is Immobilized on polyurethane foam (PUF) cubes. It has been seen that *Basidiomycetes* fungus, *Flavodon flavus* can decolorize synthetic dyes such as Congo red, Remazol brilliant blue R, Poly-B, and Poly-R. This fungus efficiently decolorized pigments in the molasses spent wash and could also reduce the total phenolic and COD up to 50% and toxicity completely. Verma et al. (2012) described that rapid decolorization and detoxification of anthraquinone dye Reactive Blue 4 can be achieved by enzymatic oxidation and sorption of degraded products on marine-derived fungal biomass. *Saagaromyces ratnagiriensis*, a non-white rot obligate marine ascomycete fungus has capability to decolorize effluent of paper mill (Sarma 2018).

### 7.5 **Biomass Valorization**

Rapid exploration of Earth's resources has been steadily increased, giving rise to depletion of resources and rapid generation of waste product (Ferreira 2015). Nowadays, the best way to reduce the waste generation in large amount is their application for the production of value-added products. Valorization is the approach that facilitates sustainable development by value-added products from the waste (Kumari et al. 2018). Waste valorization is the process of converting waste materials into more useful products such as chemicals, reusable materials and fuels (Arancon et al. 2013).

Fungi are heterotrophic and obtain sustenance by hydrolyzing complex material and convert into simple form by taking up and utilizing substance for their biosynthesis and energy production (Hanson 2008). Filamentous fungi exist in a myriad of environments and have capacity to process complex and diverse substrates include starch or lignocellulosic polymers such as cellulose, hemicellulose, and lignin (Maity 2015). The biomass and byproducts produced by fungi during waste treatment are better valorized than bacteria. Fungi provide valuable enzymes as well as proteins. Several value-added products such as biofuels and biochemicals are produced by industrial cultivation of fungi using industrial waste as substrates.

It has been reported that processing of waste from dairy, sugarcane, tanning, oil, cotton, bioethanol, agro, marine, and poultry industries can be utilized as an attractive alternative source of low-cost organics and nutrients to valorize waste into fungal products with associated waste treatment. (Ali et al. 2020; Mahari et al. 2020: Gaur et al. 2020: Patel et al. 2017: Koutra et al. 2018). Compared to conventional physico-chemical processes, biological processes by using microorganisms including bacteria, fungi, and yeasts are offers a promising technique to produce biofuel while decolorizing recalcitrant synthetic dyes and lignin (Ali et al. 2019, 2020; Kiayia et al. 2019). Waste produced from the agro-industrial could be used as a sugar-based carbon source that can be either used alone or supplemented with various expensive nutrients like yeast extract for the production value-added products such as biodiesel, bioplastic, and exopolysaccharides at laboratory scale and pilot scale. Several value-added products that have been produced from wastes include biofuels like bioethanol and biohydrogen, short-chain organic acids, building-blocks, including 2, 3-butanediol, 1, 3-propanediol, and succinic acid, polymers like bioplastics, i.e., polyhydroxyalkanoates (Koutinas et al. 2014). The major waste generated in paper pulp industry is cellulose-based fibers that can be further treated for the production of useful products like fabric and paper.

Palmqvist and Hahn-Hägerdal (2000) explained that waste generated in pulp and paper industry is spent sulphite liquor (SSL) that can be used for the fabrication of phenolic compounds mainly aromatics syringic, gallic, and vanillic acids. According to Alexandri et al. (2016) the SSL can also be utilized as a raw material for the production of single-cell protein, bioethanol, bioplastics, bacterial cellulose and other valuable products. Mukherjee et al. (2015) stated that food waste can be product utilized for the production of another value-added called hydroxymethylfurfural (HMF), which could be utilized as the precursor of medicines, polymers, resins, solvents, and biofuels.

Similarly, lignocellulosic waste biomass has been used for the production of phytosterols, polypropylene, acrylic acid and esters (Bardhan et al. 2015). The main enzymes identified for lignin degradation include lignin peroxidase (LiP), manganese peroxidase (MnP), and the copper-containing phenoloxidase known as laccase. These ligninolytic enzymes have already been demonstrated utility in the food industry, pulp and paper industry, textile industry, and as biocatalysts (Jaqueline et al. 2010). Studies on utilization of lignin and/or lignin-like dyes by oleaginous yeasts hold much promise for achieving overall efficiency and sustainable utilization of lignocellulosic biomass and textile azo dyes for biofuel production (Ali et al. 2021).

White rot fungi, especially *Phanerochaete*, *Trametes*, *Bjerkandera*, and *Pleurotus* genera have ability to produce several lignocellulolytic extracellular enzymes. In general, the conversion of lignocellulosic biomass includes a pretreatment step for removal of protective lignin seal surrounding structural polysaccharides, followed by enzymatic hydrolysis and subsequent fermentation of released soluble sugars. Thus, the reducing sugars from hydrolysis of cellulose and hemicellulose fractions could be metabolized by other microorganisms producing value-added compounds such as alcohols, flavonoids, organic acids, and phenolics (Sánchez 2009; Mateo and Maicas 2015).

According to the study of pandey et al. (2015) *Aspergillus* spp. is involved in the production of many value-added products including enzymes such as amylase, protease, lipase, phytase, lactase, and catalase. *Trichoderma* spp. can be used for production of cellulose and xylanase. Moreover, *Aspergillus* spp. are responsible for a major fraction of commercial production of organic acids including citric acid, gluconic acid, and itaconic acid and they are also potential sources of malic and oxalic acid. Chitosan is used for production of superabsorbents can be obtained via hydrolysis of chitin from the cell walls of *Aspergillus* spp. and these ascomycetes can be used for production of keratinase hydrolysates (Pandey et al. 2015; Zamani 2010).

*Monascus* spp. have been important sources of pigments for the food industry and together with *Aspergillus* spp., *Fusarium* spp. and *Neurospora* spp. have been a source of different human food products (Ferreira 2015). However, unicellular ascomycetes, that is, yeasts such as *Saccharomyces* spp., *Pichia* spp., and *Yarrowia* spp. have also been reported to be potential sources of organic acids (such as a-ketoglutaric acid, lactic acid, malic acid and pyruvic acid), polysaccharides such as glucan, proteins like collagen, polyunsaturated fatty acids, sterols (e.g., squalene) and lipids (e.g., ceramides). Marine fungi can potentially play an important role as a bio catalyst in waste biorefineries due to their ability to produce enzymes that can break down these recalcitrant structures. By using filamentous ascomycetes, their biomass, normally rich in proteins and lipids, can represent another value-added product of the biorefinery (Ferreira 2015; Ferreira et al. 2016).

# 7.6 Conclusions and Future Prospectives

Fungi are considered as natural decomposers which can significantly reduce and degrade various recalcitrant, persistent, and toxic pollutants like hydrocarbons, heavy metals, and dyes. Most of the studies show the role of various extracellular ligninolytic enzymes and cytochrome 450 in the degradation of these pollutants. However, in most cases, the underlying mechanism of the mycoremediation of these harmful pollutants is elusive and needs further research. Mycoremediation can be augmented by adding carbon sources at polluted sites and providing optimum condition to increase degradation process. Naturally present community of microbes acts in concert with the fungi to decompose the harmful contaminants. White rot
fungi are extremely effective in decomposing toxic aromatic pollutants, heavy metals and dyes etc. Further studies could be helpful in understanding the mechanism and optimizing the process of mycoremediation. Benefit is offered that land that is contaminated and unfit for agriculture could be both restored and made to yield a nutritious food crop. Biomass valorization is the process which can convert waste materials into more useful products such as chemicals, reusable materials and fuels. This chapter will help to expand our understanding for the fungi from marine environment as potential candidates for biomass valorization and mycoremediation of hazardous pollutants that would be important for economical, ecological, and legal reasons as well.

## References

- Abe F, Miura T, Nagahama T, Inoue A, Usami R, Horikoshi K (2001) Isolation of a highly copper tolerant yeast, Cryptococcus sp., from the Japan trench and the induction of superoxide dismutase activity by Cu2+. Biotechnol Lett 23:2027–2034
- Akar T, Tunali S (2005) Biosorption performance of Botrytis cinerea fungal by-products for removal of Cd (II) and Cu (II) ions from aqueous solutions. Miner Eng 18(11):1099–1109
- Akcil A, Erust C, Ozdemiroglu S, Fonti V, Beolchini F (2015) A review of approaches and techniques used in aquatic contaminated sediments: metal removal and stabilization by chemical and biotechnological processes. J Clean Prod 86:24–36
- Aksu Z, Kilic NK, Ertugrul S, Donmez G (2007) Inhibitory effects of chromium (VI) and Remazol black B on chromium(VI) and dyestuff removals by *Trametesversicolor*. Enzym Microb Technol 40:1167–1174
- Alexandri M, Papapostolou H, Vlysidis A, Gardeli C, Komaitis M, Papanikolaou S, Koutinas AA (2016) Extraction of phenolic compounds and succinic acid production from spent sulphite liquor, vol 91. J Chem Technol Biotechnol, p 2751
- Ali H (2010) Biodegradation of synthetic dyes–a review. Water Air Soil Pollut 213(1–4):251–273. https://doi.org/10.1007/s11270-010-0382-4
- Ali H, Khan E, Ilahi I (2019) Environmental chemistry and ecotoxicology of hazardous heavy metals: environmental persistence, toxicity, and bioaccumulation. J Chem 2019:6730305
- Ali SS, Al-Tohamy R, Xie RM, El-Sheekh M, Sun J (2020) Construction of a new lipase- and xylanase-producing oleaginous yeast consortium capable of reactive azo dye degradation and detoxification. Bioresour Technol 313:123631
- Ali SS, Sun J, Koutra E, El-Zawawy N, Elsamahy T, El-Shetehy M (2021) Construction of a novel cold-adapted oleaginous yeast consortium valued for textile azo dye wastewater processing and biorefinery. Fuel 285:119050
- Anastasi A, Tigini V, Varese GC (2013) The bioremediation potential of different ecophysiological groups of fungi. In: Goltapeh EM et al (eds) Fungi as bioremediators. Soil Biol, vol 32, pp 29–49
- Annuar M, Adnan S, Vikineswary S, Chisti Y (2009) Kinetics and energetics of azo dye decolorization by *Pycnoporus sanguineus*. Water Air Soil Pollut 202:179–188
- Arancon RAD, Lin CSK, Chan KM, Kwan TH, Luque R (2013) Advances on waste valorization: new horizons for a more sustainable society. Energy Sci Eng 1(2):53–71
- Arun A, Raja PP, Arthi R, Ananthi M, Kumar KS, Eyini M (2008) Polycyclic aromatic hydrocarbons (PAHs) biodegradation by basidiomycetes fungi, *pseudomonas* isolate, and their cocultures: comparative *in vivo* and *in silico* approach. Appl Biochem Biotechnol 151:132–142
- Atlas RM (1995) Petroleum biodegradation and oil spill bioremediation. Mar Pollut Bull 31:178– 182

- Ayangbenro AS, Babalola OO (2017) A new strategy for heavy metal polluted environments: a review of microbial biosorbents. Int J Environ Res Public Health 14(1):94
- Baccar R, Blanquez P, Bouzid J, Feki M, Attiya H, Sarra M (2011) Decolorization of a tannery dye: from fungal screening to bioreactor application. Biochem Eng J 56:84–189
- Badia-Fabregat M, Lucas D, Gros M, Rodríguez-Mozaz S, Barceló D, Caminal G, Vicent T (2015) Identification of some factors affecting pharmaceutical active compounds (PhACs) removal in real wastewater. Case study of fungal treatment of reverse osmosis concentrate. J Hazard Mater 11(283):663–671
- Bardhan SK, Gupta S, Gorman ME, Haider MA (2015) Biorenewable chemicals: feedstocks, technologies and the conflict with food production. Renew Sust Energy Rev 51:506–520
- Bartlett RE (1971) Public health engineering-Design in Metric Waste-Water treatment. Applied Science Publishers, London
- Bhainsa KC, D'Souza SF (2009) Thorium biosorption by *aspergillus fumigatus*, a filamentous fungal biomass. J Hazard Mater 165:670–676
- Bishnoi NR, Garima A (2005) Fungus-an alternative for bioremediation of heavy metal containing wastewater: a review. J Sci Ind Res 64:93–100
- Bonugli-Santos RC, Durrant LR, Sette LD (2012) The production of ligninolytic enzymes by marinederived basidiomycetes and their biotechnological potential in the biodegradation of recalcitrant pollutants and the treatment of textile effluents. Water Air Soil Pollut 223(5): 2333–2345
- Bonugli-Santos RC, dos Santos Vasconcelos MR, Passarini MRZ, Vieira GAL, Lopes VCP, Mainardi PH, dos Santos JA, de Azevedo DL, Otero IVR, da Silva Yoshida AM, Feitosa VA, Pessoa A Jr, Sette LD (2015) Marine-derived fungi: diversity of enzymes and biotechnological applications. Front Microbiol 6:269
- Bovio E, Fauchon M, Toueix Y, Mehiri M, Varese GC, Hellio C (2019) The sponge-associated fungus Eurotium chevalieri MUT 2316 and its bioactive molecules: potential applications in the field of antifouling. Mar Biotechnol 21(6):743–752
- Bovio E, Gnavi G, Prigione V, Spina F, Denaro R, Yakimov M, Calogero R, Crisafi F, Varese GC (2017) The culturable mycobiota of a Mediterranean marine site after an oil spill: isolation, identification and potential application in bioremediation. Sci Total Environ 576:310–318
- Bucher VVC, Hyde KD, Pointing SB, Reddy CA (2004) Production of wood decay enzymes mass loss and lignin solubilization in wood by marine *ascomycetes* and their anamorphs. Fungal Divers 15:1–14
- Campos R, Kandelbauer A, Robra KH, Cavaco-Paulo A, Gübitz GM (2001) Indigo degradation with purified laccases from *Trametes hirsuta* and *Sclerotium rolfsii*. J Biotechnol 89(2\_3): 131\_139
- Capotorti G, Digianvincenzo P, Cesti P, Bernardi A, Guglielmetti G (2004) Pyrene and benzo (a) pyrene metabolism by an *aspergillus terreus* strain isolated from a polycylic aromatic hydrocarbons polluted soil. Biodegradation 15:79–85
- Chagas EP, Durrant LR (2001) Decolorization of azo dyes by *Phanerochaetechrysosporium* and *Pleurotussajorcaju*. Enzym Microb Technol 29(8–9):473–477
- Chaillan F, Le Flèche A, Bury E, Phantavong YH, Grimont P, Saliot A, Oudot J (2004) Identification and biodegradation potential of tropical aerobic hydrocarbon-degrading microorganisms. Res Microbiol 155(7):587–595
- Chakraborty S, Mukherjee A, Das TK (2013) Biochemical characterization of a lead-tolerant strain of *aspergillus foetidus*: an implication of bioremediation of lead from liquid media. Int Biodeterior Biodegradation 84:134–142
- Chatterjee S, Chatterjee NC, Dutta S (2012) Bioreduction of chromium (VI) to chromium (III) by a novel yeast strain *Rhodotorula mucilaginosa* (MTCC 9315). Afr J Biotechnol 11(83): 14920–14929
- Chauhan A, Samanta SK, Jain RK (2000) Degradation of 4-nitrocatechol by Burkholderia cepacia: a plasmid-encoded novel pathway. J Appl Microbiol 88(5):764\_772

- Chivukula M, Renganathan V (1995) Phenolic azo dye oxidation by laccase from *Pyricularia oryzae*. Appl Environ Microbiol 61(12):4374–4377
- Cicatiello P, Gravagnuolo AM, Gnavi G, Varese GC, Giardina P (2016) Marine fungi as source of new hydrophobins. Int J Biol Macromol 92:1229–1233
- Ciullini I, Tilli S, Scozzafava A, Briganti F (2008) Fungal laccase, cellobiose dehydrogenase, and chemical mediators: combined actions for the decolorization of different classes of textile dyes. Bioresour Technol 99:7003–7010
- Cripps C, Bumpus JA, Aust SD (1990) Biodegradation of azo and heterocyclic dyes by *Phanerocheate chrysosporium*. Appl Environ Microbiol 56:1114–1118
- Cullen WR, Reimer KJ (1989) Arsenic speciation in the environment. Chem Rev 89(4):713-764
- D'Souza-Ticlo D, Sharma D, Raghukumar C (2009) A thermostable metal-tolerant laccase with bioremediation potential from a marine-derived fungus. Mar Biotechnol 11:725–737
- Da Silva M, Passarini MRZ, Bonugli RC, Sette LD (2008) Cnidarian derived filamentous fungi from Brazil: isolation, characterization and RBBR decolourisation screening. Environ Technol 29:1331–1339
- Dacco C, Girometta C, Asemoloye MD, Carpani G, Picco AM, Tosi S (2020) Key fungal degradation patterns, enzymes and their applications for the removal of aliphatic hydrocarbons in polluted soils: a review. Int Biodeterior Biodegradation 147:104866
- Damare S, Singh P, Raghukumar S (2012) Biotechnology of marine fungi. Prog Mol Subcell Biol 53:277–297
- Das D, Das N, Mathew L (2010) Kinetics, equilibrium and thermodynamic studies on biosorption of Ag (I) from aqueous solution by macrofungusPleurotus platypus. J Hazard Mater 184(1–3): 765–774
- Das N, Chandran P (2011) Microbial degradation of petroleum hydrocarbon contaminants: an overview. Biotechnol Res Int 2011:1–13
- Davies DJA, Bennett BG (1983) Exposure commitment assessment of environment pollutants, vol 3. MARC (Monitoring and Assessment Research Centre) Report Number 30. MARC Publication, London
- Davis TA, Volesky B, Mucci A (2003) A review of biochemistry of heavy metal biosorption by brown algae. Water Res 37:4311–4330
- Dermont G, Bergeron M, Mercier G, Richer-Lafleche M (2008) Soil washing for metal removal: a review of physical/chemical technologies and field applications. J Hazard Mater 152:1–31
- Deshmukh R, Khardenavis AA, Purohit HJ (2016) Diverse metabolic capacities of fungi for bioremediation. Indian J Microbiol 56(3):247–264. https://doi.org/10.1007/s12088-016-0584-6
- Divya LM, Prasanth GK, Sadasivan C (2014) Potential of the salt-tolerant laccase-producing strain *Trichoderma viride* Pers. NFCCI-2745 from an estuary in the bioremediation of phenol-polluted environments. J Basic Microbiol 54(6):542–547
- D'Souza DT, Tiwari R, Sah AK, Raghukumar C (2006) Enhanced production of laccase by a marine fungus during treatment of colored effluents and synthetic dyes. Enzym Microb Technol 38(3–4):504–511
- Durairaj P, Malla S, Nadarajan SP et al (2015) Fungal cytochrome P450 monooxygenases of *fusarium oxysporum* for the synthesis of x-hydroxy fatty acids in engineered *Saccharomyces cerevisiae*. Microb Cell Factories 14:45. https://doi.org/10.1186/s12934-015-0228-2
- Dursun AY, Uslu G, Cuci Y, Aksu Z (2003) Bioaccumulation of copper (II), lead (II) and chromium (VI) by growing Aspergillus niger. Process Biochem 38(12):1647–1651
- Dwivedi P, Singh Tomar R (2018) Bioremediation of textile effluent for degradation and decolourization of synthetic dyes: a review. Int J Curr Res Life Sci 7(04):1948–1951
- El Aty AAA, Mostafa FA, Hassan ME, Hamed ER, Esawy MA (2017) Covalent immobilization of *Alternaria tenuissima* KM651985 laccase and some applied aspects. Biocatal Agricult Biotechnol 9:74–81
- El-Kassas HY, El-Taher EM (2009) Optimization of batch process parameters by response surface methodology for mycoremediation of chrome-VI by a chromium resistant strain of marine *Trichoderma viride*. Am-Eurasian J Agric Environ Sci 5:676–681

- Ferreira JA (2015) Integration of filamentous fungi in ethanol dry-mill biorefinery (Ph.D. thesis). Swedish Centre for Resource Recovery. University of Borås, Borås, Sweden
- Ferreira JA, Mahboubi A, Lennartsson PR, Taherzadeh MJ (2016) Waste biorefineries using filamentous ascomycetes fungi: present status and future prospects. Bioresour Technol 215: 334–345
- Fillat U, Prieto A, Camararo S, Martinez AT, Martinez MJ (2012) Biodeinking of flexographic inks by fungal laccase using synthetic and natural mediators. Biochem Eng J 67:97–103
- Forgacs E, Cserhati T, Oros G (2004) Removal of synthetic dyes from wastewaters: a review. Environ Int 30:953–971
- Fu Y, Viraraghavan T (2001) Fungal Decolorization of dye wastewaters: a review. Bioresour Technol 79:251–262
- Fu YQ, Li S, Zhu HY, Jiang R, Yin LF (2012) Biosorption of copper (II) from aqueous solution by mycelial pellets of Rhizopus oryzae. Afr J Biotechnol 11(6):1403–1411
- Gadd GM (2007) Presidential address geomycology: Biogeochemical transformations of rocks, minerals, metals and radionuclides by fungi, bioweathering and bioremediation. Mycol Res 3:3– 49
- Gaur VK, Sharma P, Sirohi R, Awasthi MK, C-Gilles D, Pandey A (2020) Assessing the impact of industrial waste on environment and mitigation strategies: a comprehensive review. J Hazard Mater 123019:123019
- Gazem MAH, Nazareth S (2013) Sorption of lead and copper from an aqueous phase system by marine-derived *aspergillus* species. Ann Microbiol 63:503–511
- Gibert O, Pablo J, Cortina JL, Ayora C (2005) Municipal compost-based mixture for acid mine drainage bioremediation: metal retention mechanisms. Appl Geochem 20:1648–1657
- Hanson JR (2008) RSo chemistry. Chemistry of fungi. Royal Society of Chemistry
- Haritash AK, Kaushik CP (2009) Biodegradation aspects of polycyclic aromatic hydrocarbons (PAHs): a review. J Hazard Mater 169:1–15. https://doi.org/10.1016/j.jhazmat.2009.03.137
- Huanga J, Liua D, Lub J, Wanga H, Weia X, Liua J (2016) Biosorption of reactive black 5 by modified *aspergillus versicolor* biomass: kinetics, capacity and mechanism studies. Colloids Surf A Physicochem Eng Asp 492:242–248
- Imandi SB, Chinthala R, Saka S, Vechalapu RR, Nalla KK (2014) Optimization of chromium biosorption in aqueous solution by marine yeast biomass of Yarrowialipolytica using Doehlert experimental design. Afr J Biotechnol 13(12)
- Imhoff JF (2016) Natural products from marine fungi—still an underrepresented resource. Mar Drugs 14:19
- Iqbal M, Edyvean RGJ (2004) Biosorption of lead, copper and zinc ions on loofa sponge immobilized biomass of Phanerochaetechrysosporium. Miner Eng 17(2):217–223
- Jain K, Shah V, Chapla D, Madamwar D (2012) Decolorization and degradation of azo dye– reactive violet 5R by an acclimatized indigenous bacterial mixed cultures-SB4 isolated from anthropogenic dye contaminated soil. J Hazard Mater 213–214:378–386
- Jaqueline M, Maciel M, Castro e Silva A, Ribeiro H (2010) Industrial and biotechnological applications of ligninolytic enzymes of the basidiomycota: a review. Electron. J Biotechnol 13(6)
- Jin X, Liu G, Xu Z, Yao W (2007) Decolorization of a dye industry effluent by aspergillus funigatus XC6. Appl Microbiol Biotechnol 74:239–243
- Johannes C, Majcherczyk A, Hüttermann A (1998) Oxidation of acenaphthene and acenaphthylene by laccase of Trametes versicolor in a laccase-mediator system. J Biotechnol 61:151–156
- Joshi HV, Vala AK, Bhatt PN, Anand N (2012) Treatment of CETP effluent by a seaweed associated *aspergillus Niger* isolate. In: Sahoo D, Kaushik BD (eds) Algal biotechnology and environment. I.K. International Publishing House, New Delhi, pp 193–197
- Kadri T, Rouissi T, Kaur Brar S, Cledon M, Sarma S, Verma M (2017) Biodegradation of polycyclic aromatic hydrocarbons (PAHs) by fungal enzymes: a review. J Environ Sci 51:52–74
- Kapoor A, Viraraghavan T (1997) Biosorption of heavy metals on aspergillus Niger: effect of pretreatment. Bioresour Technol 63:109–113

- Karigar CS, Rao SS (2011) Role of microbial enzymes in the bioremediation of pollutants: a review, vol 2011. Enzyme Res, p 1
- Kaushik CP, Haritash AK (2006) Polycyclic aromatic hydrocarbons (PAHs) and environmental health. Our Earth 3:1–7
- Kaushik P, Malik A (2009) Fungal dye decolorization: recent advances and future potential. Environ Int 35:127–141
- Khambhaty Y, Mody K, Basha S, Jha B (2009) Biosorption of Cr (VI) onto marine aspergillus Niger: experimental studies and pseudo-second order kinetics. World J Microbiol Biotechnol 25:1413
- Khan I, Aftab M, Shakir S, Ali M, Qayyum S, Rehman MU, Touseef I (2019) Mycoremediation of heavy metal (cd and Cr)–polluted soil through indigenous metallotolerant fungal isolates. Environ Monit Assess 191(9)
- Khan R, Bhawana P, Fulekar MH (2013) Microbial decolorization and degradation of synthetic dyes. Rev Environ Sci Biotechnol 12(1):75–97. https://doi.org/10.1007/s11157-012-9287-6
- Khan FI, Husain T, Hejazi R (2004) An overview and analysis of site remediation technologies. J Environ Manag 71:95–122
- Kiayia Z, Lotfabad TB, Heidarinasab A, Shahcheraghi F (2019) Microbial degradation of azo dye carmoisine in aqueous medium using Saccharomyces cerevisiae ATCC 9763. J Hazard Mater 373:608–619
- Kirk PW, Gordon AS (1988) Hydrocarbon degradation by filamentous marine higher fungi. Mycologia 80:776–782
- Koutinas AA, Vlysidis A, Pleissner D, Kopsahelis N, Garcia IL, Kookos IK, Lin CSK (2014) Valorization of industrial waste and by-product streams via fermentation for the production of chemicals and biopolymers. Chem Soc Rev 43:2587–2627
- Koutra E, Grammatikopoulos G, Kornaros M (2018) Selection of microalgae intended for valorization of digestate from agro-waste mixtures. Waste Manag 73:123–129
- Kumari M, Ghosh P, Thakur IS (2018) Application of microbes in remediation of hazardous wastes: a review. Energy Environ Sustain:223–241. https://doi.org/10.1007/978-981-10-7485-1\_11
- Kvenvolden KA, Cooper CK (2003) Natural seepage of crude oil into the marine environment. Geo-Mar Lett 23:140–146
- Lakkireddy K, Kues U (2017) Bulk isolation of basidiospores from wild mushrooms by electrostatic attraction with low risk of microbial contaminations. AMB Express 7(1):28
- Lalitha P, Ngeswara Rao Reddy N, Arunalakshmi K (2011) Decolorization of synthetic dyes by *aspergillus flavus*. Biorem J 15(2):121–132
- Lenin M, Kumar MS, Mycin TR (2014) Effect of sago factory effluent on seed germination and seedling growth of gingelly (Sesamumindicum L.) varieties. Int J Life Sci Bioteh Pharma Res 3(1):151–160
- Lin J, Zhang X, Li Z, Lei L (2010) Biodegradation of Reactive blue 13 in a two-stage anaerobic/ aerobic fluidized beds system with a Pseudomonas sp. isolate. Bioresour Technol 101(1):34–40
- Liu XF, Culotta VC (1999) Mutational analysis of *Saccharomyces cerevisiae* Smf1p, a member of the Nramp family of metal transporters. J Mol Biol 289:885–891
- Lopez Errasquín E, Vázquez C (2003) Tolerance and uptake of heavy metals by Trichoderma atroviride isolated from sludge. Chemosphere 50:137–143
- Lu T, Zhang Q, Yao S (2016) Efficient decolorization of dye-containing wastewater using mycelial pellets formed of marine-derived *aspergillus Niger*. Chin J Chem Eng A 25:330
- Luna JM, Rufino RD, Sarubbo LA (2016) Biosurfactant from *Candida sphaerica* UCP0995 exhibiting heavy metal remediation properties. Process Saf Environ Prot 102:558–566
- Madhavi S, Revankar S, Lele S (2007) Synthetic dye decolorization by white rot fungus, Ganoderma sp. WR-1. Bioresour Technol 984:775–780
- Mahari WAW, Peng W, Nam WL, Yang H, Lee XY, Lee YK, Liew RK, Ma NL, Mohammad A, Sonne C, Van Le Q (2020) A review on valorization of oyster mushroom and waste generated in the mushroom cultivation industry. J Hazard Mater 400:123156

- Maher W, Butler E (1988) Arsenic in the marine environment. Appl Organomet Chem 2(3): 191–214
- Maity SK (2015) Opportunities, recent trends and challenges of integrated biorefinery: part I. Renew Sust Energ Rev 43:1427–1445
- Majeau J-A, Brar SK, Tyagi RD (2010) Laccases for removal of recalcitrant and emerging pollutants. Bioresour Technol 101:2331e2350
- Margot J, Bennati-Granier C, Maillard J, Bla'nquez P, Barry DA, Holliger C (2013) Bacterial versus fungal laccase: potential for micropollutant degradation. AMB Express 3:63. https://doi. org/10.1186/2191-0855-3-63
- Mateo JJ, Maicas S (2015) Valorization of winery and oil mill wastes by microbial technologies. Food Res Int 73:13–25
- Meckenstock Rainer U (2004) Anaerobic degradation of polycyclic aromatic hydrocarbons. FEMS Microbiol Ecol 49(1):27–36
- Mills MA, Bonner JS, Page CA et al (2004) Evaluation of bioremediation strategies of a controlled oil release in a wetland. Mar Pollut Bull 49:425–435
- Mitra A, Roy D, Roy P, Bor AM, Sarkar Mitra AK (2014) Sustainability of *aspergillus* spp. in metal enriched substrate aiming towards increasing bioremediation potential. World J Pharm Pharm Sci 3:864–878
- Molitoris HP, Buchalo AS, Kurchenko I, Nevo E, Rawal BS, Wasser SP, Oren A (2000) Physiological diversity of the first filamentous fungi isolated from the hypersaline Dead Sea. In: Hyde KD, Ho WH, Pointing SB (eds) Aquatic mycology across the Millenium, vol 5, pp 55–70Fungal Diversity
- Mukherjee A, Dumont MJ, Raghavan V (2015) Sustainable production of hydroxymethylfurfural and levulinic acid: challenges and opportunities. Biomass Bioenergy 72:143–183
- Mulligan CN, Yong RN, Gibbs BF (2001) Remediation technologies for metal-contaminated soils and groundwater: an evaluation. Eng Geol 60(1–4):193–207. https://doi.org/10.1016/S0013-7952(00)00101-0
- Namdhari BS, Rohilla SK, Salar RK, Gahlawat SK, Bansal P, Saran AK (2012) Decolorization of reactive blue MR, using *aspergillus* species isolated from textile waste water. ISCA J Biological Sci 1(1):24–29
- Nicoletti R, Andolfi A (2018) The marine-derived filamentous fungi in biotechnology, grand challenges in biology and biotechnology. https://doi.org/10.1007/978-3-319-69075-9\_4
- Ollikka P, Alhonmaki K, Leppanen V, Glumoff T, Raijola T, Suominen I (1993) Decolorization of azo, triphenyl methane, heterocyclic and polymeric dyes by ligni-peroxidase isozymes from *Phanerochaetechrysosporium*. Appl Environ Microbiol 59:4010±4016
- Oyetibo GO, Ishola ST, Ikeda-Ohtsubo W, Miyauchi K, Ilori MO, Endo G (2015) Mercury bioremoval by *Yarrowia* strains isolated from sediments of mercury-polluted estuarine water. Appl Microbiol Biotechnol 99:3651–3657
- Palmqvist E, Hahn-Hägerdal B (2000) Fermentation of lignocellulosic hydrolysates. I: inhibition and detoxification. Bioresour Technol 74:17–24
- Pande V, Pandey SC, Joshi T, Sati D, Gangola S, Kumar S, Samant M (2019) Biodegradation of toxic dyes: a comparative study of enzyme action in a microbial system. In: Smart bioremediation technologies. Academic Press, pp 255–287
- Pandey A, Höfer R, Larroche C, Taherzadeh MJ, Nampoothiri M (2015) Industrial biorefineries and white biotechnology. Elsevier, USA
- Pandey A, Singh P, Iyengar L (2007) Bacterial decolorization and degradation of azo dyes. Int Biodeterior Biodegrad 59:73–84
- Pang K-L, Overy DP, Jones EBG, Calado MDL, Burgaud G, Walker AK, Johnson JA, Kerr RG, Cha H-J, Bills GF (2016) 'Marine fungi' and 'marine-derived fungi' in natural product chemistry research: toward a new consensual definition. Fungal Biol Rev 30:163–175
- Park D, Yun YS, Jo JH, Park JM (2005) Mechanism of hexavalent chromium removal by dead fungal biomass of Aspergillusniger. Water Res 39(4):533–540

- Passarini MZR, Rodrigues MVN, da Silva M, Sette LD (2011) Marine-derived filamentous fungi and their potential application for polycyclic aromatic hydrocarbon bioremediation. Mar Poll Bull 62:364–370
- Patel A, Sartaj K, Arora N, Pruthi V, Pruthi PA (2017) Biodegradation of phenol via meta cleavage pathway triggers de novo TAG biosynthesis pathway in oleaginous yeast. J Hazard Mater 340: 47–56
- Peng R, Ai-Sheng X, Yong X, Xiao-Yan F, Feng G, Wei Z, Yong-Sheng T, Quan-Hong Y (2008) Microbial biodegradation of polyaromatic hydrocarbons. FEMS Microbiol Rev 32:927–955
- Pundir R, Chary GHVC, Dastidar MG (2016) Application of Taguchi method for optimizing the process parameters for the removal of copper and nickel by growing *aspergillus* sp. Water Resour Res
- Raghukumar C (2004) Marine fungi and their enzymes of decolorization of colored effluents. In: Ramaiah N (ed) Marine microbiology: facets and opportunities. National Institute of Oceanagraphy, Goa, India, pp 145–158
- Raghukumar C, D'Souza-Ticlo D, Verma AK (2008) Treatment of colored effluents with lignindegrading enzymes: an emerging role of marine-derived fungi. Crit Rev Microbiol 34(3–4): 189–206
- Raghukumar C, Shailaja MS, Parameswaran PS, Singh SK (2006) Removal of polycyclic aromatic hydrocarbons from aqueous media by the marine fungus NIOCC#312:involvement of lignindegrading enzymes and exopolysaccharides. Indian J Mar Sci 35:373–379
- Rai H, Bhattacharya M, Singh J, Bansal TK, Vats P, Banerjee UC (2005) Removal of dyes from the effluent of textile and dyestuff manufacturing industry: a review of emerging techniques with reference to biological treatment. Crit Rev Environ Sci Technol 35:219–238
- Raikar MT, Raghukumar S, Vani V, David JJ, Chandramohan D (2001) *Thraustochytrid protists* degrade hydrocarbons. Ind J Marine Sci 30:139–145
- Rao AR, Baskaran V, Sarada R, Ravishankar GA (2013) In vivo bioavailability and antioxidant activity of carotenoids from microalgal biomass—A repeated dose study. Food Res Int 54(1): 711–717
- Ravelet C, Krivobok S, Sage L, Steiman R (2000) Biodegradation of pyrene by sediment fungi. Chemosphere 40:557–563
- Rehman A, Farooq H, Hasnain H (2008) Biosorption of copper by yeast, Lodderomyces elongisporus, isolated from industrial effluents: its potential use in wastewater treatment. J Basic Microbiol 48:195–201
- Robinson T, McMullan G, Marchant R, Nigam P (2001) Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. Bioresour Technol 77:247–255
- Rodrigue E, Picard MA, Vazquez Duhal TR (1999) Industrial dye decolorization by laccases from lignolytic fungi. Curr Microbiol 38(27):32
- Sánchez C (2009) Lignocellulosic residues: biodegradation and bioconversion by fungi. Biotechnol Adv 27:185–194
- Sandhya S (2010) Biodegradation of azo dyes under anaerobic condition: role of azoreductase. In: Biodegradation of azo dyes, pp 39–57
- Saranraj P, Sumathi V, Reetha D, Stella D (2010) Fungal decolorization of direct azo dyes and biodegradation of textile dye effluent. J Ecobiotechnol 2(7):12–16
- Sarma VV (2018) Obligate marine fungi and bioremediation, pp 307–323. https://doi.org/10.1007/ 978-3-319-77386-5
- Selvam K, Shanmuga Priya M (2012) Biological treatment of azo dyes and textile industry effluent by newly isolated white rot fungi *Schizophyllumcommune* and *Lenziteseximia*. Int J Environ Sci 2:4
- Sheoran AS, Sheoran V (2006) Heavy metal removal mechanism of acid mine drainage in wetlands: a critical review. Miner Eng 19:105–116
- Siddiquee S, Rovina K, Al Azad S (2015) Heavy metal contaminants removal from wastewater using the potential filamentous fungi biomass: a review. J Microbiol Biochem Tech 07(06)

Singh H (2006) Mycoremediation: fungal bioremediation. John Wiley & Sons, New Jersey

- Singh H (2006b) Fungal decolorization and degradation of dyes. In: Singh H (ed) Mycoremediation: fungal bioremediation, pp 420–483
- Singh OV, Labana S, Pandey G, Budhiraja R, Jain RK (2003) Phytoremediation: an overview of metallic ion decontamination from soil. Appl Microbiol Biotechnol 61(5):405–412
- Singh P, Raghukumar C, Parvatkar RR, Mascarenhas-Pereira MBL (2013) Heavy metal tolerance in the *psychrotolerant Cryptococcus* sp. isolated from deep-sea sediments of the central Indian Basin. Yeast 30:93–101
- Singh RK, Tripathi R, Ranjan A, Srivastava AK (2019) Fungi as potential candidates for bioremediation. In: Abatement of environmental pollutants, pp 177–191
- Srivastava S, Thakur IS (2006) Isolation and process parameter optimization of *aspergillus* sp. for removal of chromium from tannery effluent. Bioresour Technol 97(10):1167–1173
- Supaka NJ, Somsak DKD, Pierre MLS (2004) Microbial decolorization of reactive azo dyes in a sequential anaerobic–aerobic system. Chem Eng J 99:169–176
- Taboski MAS, Rand TG, Piorko A (2005) Lead and cadmium uptake in the marine fungi Corollospora lacera and Monodictys pelagica. FEMS Microbiol Ecol 53:445–453
- Thatoi H, Behera B, Mishra R (2013) Ecological role and biotechnological potential of mangrove fungi : a review. Mycology : An Int J Fungal Biol 4:37–41
- Thippeswamy B, Shivakumar CK, Krishnappa M (2014) Studies on heavy metals detoxification biomarkers in fungal consortia. Caribb J Sci Technol 2:496–502
- Torres JMO, Cardenas CV, Moron LS, Guzman APA, dela Cruz TEE (2011) Dye decolorization activities of marine-derived fungi isolated from Manila Bay and Calatagan Bay. Philipp J Sci 140(2):133–143
- Ulrici W (2000) Contaminant soil areas, different countries and contaminant monitoring of contaminants. In: Rehm HJ, Reed G (eds) Environmental process II. Soil decontamination biotechnology, vol 11, pp 5–42
- Vala AK (2010) Tolerance and removal of arsenic by a facultative marine fungus Aspergillus candidus. Bioresour Technol 101(7):2565–2567
- Vala AK, Anand N, Bhatt PN, Joshi HV (2004) Tolerance and accumulation of hexavalent chromium by two seaweed associated aspergilli. Mar Pollut Bull 48:983–985
- Vala AK, Dave BP (2017) Marine-derived fungi: prospective candidates for bioremediation. In: Prasad R (ed) Mycoremediation and environmental sustainability. Springer, Cham, pp 17–37
- Vala AK, Sachaniya B, Dave BP (2018) Marine-derived fungi: promising candidates for enhanced bioremediation. Nanotechnology in the. Life Sci:281–300. https://doi.org/10.1007/978-3-030-02369-0\_12
- Vala AK, Patel RJ (2011) Biosorption of trivalent arsenic by facultative marine Aspergillus niger. In: Bioremediation: biotechnology, engineering and environmental management, pp 459–464
- Vala AK, Sutariya V (2012) Trivalent arsenic tolerance and accumulation in two facultative marine fungi. Jundishapur J Microbiol 5:542–545
- Vala AK, Sutariya V, Upadhyay RV (2011) Investigations on trivalent arsenic tolerance and removal potential of a facultative marine Aspergillus niger. Environ Prog Sustain Energy 30(4):586–588
- Vala AK, Upadhyay RV (2008) On the tolerance and accumulation of arsenic by facultative marine Aspergillussp. Res. J Biotechnol:366–368
- Varjani SJ (2017) Microbial degradation of petroleum hydrocarbons. Bioresour Technol 223:277– 286
- Veglio F, Beolchini F (1997) Removal of metals by biosorption: a review. Hydrometallurgy 44(3): 301–316
- Verma AK, Raghukumar C, Parvatkar RR, Naik CG (2012) A rapid two-step bioremediation of the anthraquinone dye, reactive blue 4 by a marine-derived fungus. Water Air Soil Pollut 223:3499– 3509

- Vishwanath B, Rajesh B, Janardhan A, Kumar AP, Narasimha G (2014) Fungal laccases and their applications in bioremediation. Enzyme Res 2014:1
- Wang G, Wang X, Liu X, Li Q (2012) Diversity and biogeochemical function of planktonic fungi in the ocean. In: Raghukumar C (ed) Biology of marine fungi. Springer, Berlin, pp 71–88. https:// doi.org/10.1007/978-3-642-23342-5\_4
- Wang MX, Zhang QL, Yao SJ (2015) A novel biosorbent formed of marine-derived *Penicillium janthinellum* mycelial pellets for removing dyes from dye-containing wastewater. Chem Eng J 259:837–844
- Xue J, Yu Y, Bai Y, Wang L, Wu Y (2015) Marine oil-degrading microorganisms and biodegradation process of petroleum hydrocarbon in marine environments: a review. Curr Microbiol 71(2):220–228
- Yang XQ, Zhao XX, Liu CY, Zheng Y, Qian SJ (2009) Decolorization of azo, triphenylmethane and anthraquinone dyes by a newly isolated Trametes sp. SQ01 and its laccase. Process Biochem 44:1185–1189
- Yesilada O (1995) Decolorization of crystal violet by fungi. World J Microbiol Biotechnol 11:601– 602
- Young L, Yu JT (1997) Ligninase-catalysed decolorization of synthetic dyes. Water Res 31:1187– 1193
- Yumoto I, Nakamura A, Iwata H et al (2002) A novel, facultatively psychrophilic alkaliphile that grows on hydrocarbons. Int J Syst Evol Microbiol 52:85–90
- Zamani A (2010) Superabsorbent polymers from the Cell Wall of Zygomycetes fungi. (Ph.D. thesis). Department of Chemical and Biological Engineering, Chalmers University of Technology, Gothenburg, Sweden
- Zhao X, Hardin I (2007) HPLC and spectrophotometric analysis of biodegradation of azo dyes by *Pleurotus ostreatus*. Dyes Pigments 73:322–325

# **Chapter 8 Marine Bacteria for Bioremediation**



147

Paolo Stincone, Robson Andreazza, Carolina Faccio Demarco, Thays França Afonso, and Adriano Brandelli

Abstract Marine pollution has been increasing over the years and can impact directly living organisms. The continued pollution of soil and fresh water by agriculture, industrial and urban activities frequently reaches the rivers and the ocean by run over polluting from these environments. Bioremediation is an eco-friendly technique that can immobilize, reduce damage, inactivate or remove contaminants using living organisms or their structures or products for cleaning up the environment. The technique used for bioremediation depends on the type of contaminant, including structure, oxidation stage, complexation form. Bacteria are recognized as important agents in bioremediation processes, including removal of heavy metals, biodegradation of polyaromatic and halogenated hydrocarbons, petroleum and diesel, and biodegradation of plastics. Marine bacteria present a great diversity of metabolic activities and their potential for bioremediation is still poorly exploited.

Keywords Marine pollution  $\cdot$  Biodegradation  $\cdot$  Biotechnology  $\cdot$  Bacterial metabolism  $\cdot$  Green technology

# 8.1 Introduction

Environmental pollution has becoming gradually a more complex and severe issue, generating great concern worldwide. By definition, environmental pollution refers to changes in physicochemical and biological components to such an extent that adversely affects the environmental process and quality of life. The unbalance

P. Stincone  $\cdot$  A. Brandelli ( $\boxtimes$ )

Laboratório de Bioquímica e Microbiologia Aplicada, Departamento de Ciência de Alimentos, Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil e-mail: abrand@ufrgs.br

R. Andreazza · C. F. Demarco · T. F. Afonso Centro de Engenharias, Universidade Federal de Pelotas, Pelotas, Brazil

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_8

between use of natural resources and its restoring capacity can lead to damage in different spheres, including water, soil, and air (Krishna et al. 2017; Li and Zhou 2020).

Regarding water pollution, the pollutants reaching aquatic environment can be classified as point and non-point origin, being the point source normally from an identifiable effluent discharge and the non-point source being widely distributed over an area. The release of untreated effluents into the environment as a result of different activities in industry is listed as one of the causes of water contamination, along with domestic sewage discharge (Singh et al. 2020). The contaminants with great concern in water can be both inorganic and organic: heavy metals, persistent organic pollutants (POPs), emerging pollutants, pesticides, herbicides, among others (Jeevanantham et al. 2019).

Amongst the effects of water pollution, the nutrient in excess causes eutrophication of aquatic environment. It can be highlighted by the oxygen reduction and photosynthesis damage due to growth of aquatic weed blocking the light entrance in watercourses (Bougarne et al. 2019). This eutrophic condition affects the natural cycle, being responsible for death of several species. Another important impact is the bioaccumulation of contaminants in trophic chain, leading to toxicity to human health (Ali et al. 2019).

When mentioning air pollution, the source of emission and specific climatic conditions from a location make the air pollution to be a complex combination of several particulates and gases (Leni et al. 2020). Among the pollutants affecting air quality, sulfur dioxide, ozone, carbon monoxide, and volatile organic compounds (VOCs) are emphasized. The effects of long-term exposure to air pollution in human health include cardiovascular diseases/increased mortality, along with pulmonary insufficiency and asthma (Manisalidis et al. 2020).

On the other hand, among the sources of soil pollution from anthropogenic activities, it can be mentioned the release of untreated effluent and solid wastes from industrial sector, along with accidental spill and agricultural application of pesticides, herbicides, and fertilizers. The main consequences of soil pollution are related with security in food production, along with the surface runoff carrying contaminants that end up reaching the aquatic environments (Cachada et al. 2018). This consequence poses a serious threat to population, considering the presence and accumulation of several contaminants in water, such as emerging contaminants, heavy metals, and pathological agents, potentially causing contagious diseases and toxicity effects on human health (Zulkifli et al. 2018).

The study and attempt to recover these affected areas and control the environment pollution is increasingly growing, considering the importance for population health and ecosystem equilibrium. The recovery of degraded areas can be done through several different conventional techniques, including the physicochemical methods. However, the use of eco-friendly alternatives, as the bioremediation, is gaining attention and presents cost benefits, in addition to being a more sustainable approach. In this sense, the bioremediation stands out as a technique that is based on the ability of microorganisms in reducing the pollution level through their metabolism (Verma and Kuila 2019).

In the case of heavy metal remediation, for example, some conventional techniques include the membrane filtration, chemical precipitation, coagulationfiltration, electrochemical treatment, solid-liquid separation, among others (Rahman and Singh 2020). The use of bioremediation instead of the aforementioned methods aims to overcome the disadvantage of operation cost and generation of secondary pollution as sludge formation, also being able to remove concentrations in levels that could not be removed by other practices (Leong and Chang 2020).

The marine environment presents a wide variation in its characteristics, changing the temperature, precipitation regimes, pH, salinity, pressure, currents, and prevailing winds. Considering this great environmental variability, marine organisms present great potential of adaptation, thus resisting to several adverse conditions when applied to bioremediation (Dash et al. 2013; Theerachat et al. 2018).

In addition, the marine environment is well known by possessing an enormous diversity of organisms, as yeasts, algae, archaea, protozoa, fungi, microalgae, eubacteria, cyanobacteria, and actinomycetes (Beygmoradi and Homaei 2017). Besides are being studied for bioremediation purposes, some marine organisms have already important potential uses in industry such as the production of biosurfactants, enzymes, and antibiotics (Tipre et al. 2020).

The importance of eco-friendly solutions to mitigate the environment pollution is in evidence, since they are a less aggressive alternative, considering that can be made in situ and without changing great amounts of soil from one location to another (physical changes in landscape), in addition with the fact that microorganisms or plants are used to reduce the contamination level. This alternative presents a high acceptability by the population, besides the fact that it reduces the costs for decontamination (Azubuike et al. 2020).

The understanding of gene pathways and degradation mechanisms is extremely important in discovering organisms that can enhance even more the bioremediation process of a degraded area. Thus, the genetic investigation allows the screening of bacteria with amplified potential of pollution control, along with the screening of genes and increasing the expression of enzymes responsible for degrading the contaminants. In addition, the genetically engineered bacteria are also being studied considering the advances in biotechnological and microbiological areas, where these organisms can be able to enhance the efficiency of remediation of several contaminants (Liu et al. 2019).

Thus, this chapter aims to elucidate the bioremediation principles and general aspects, along with presenting the related techniques, as bioaugmentation, bioleaching, composting, among others, showing the most recent information available. The chapter also focus on illustrating the importance of bacteria for bioremediation of different contaminants, as the removal of heavy metals, biodegradation of polyaromatic and halogenated hydrocarbons, biodegradation of petroleum and diesel, and biodegradation of plastics. Another important point of this chapter is bringing up the potential of marine organisms in bioremediation, along with the innovative use of engineering bacteria for bioremediation. The focus is also to demonstrate the great advances and benefits of application of this eco-friendly technique to remediate polluted environments.

# 8.2 Bioremediation

The bioremediation approach is based on the use of metabolic systems of microorganisms or plants to clean up the environment. The microorganisms that are used to perform the breakdown or transformation of the pollutant can be naturally existing thus named indigenous microorganisms—or from other location, intentionally added to the remediation area. The bioremediating may involve several mechanisms executed by the decontaminating agents (bacteria, fungi, or plants), such as degradation, immobilization, and alteration of contaminant, along with its entire removal from the environment (Abatenh et al. 2017).

The bioremediation can be classified into two different types: ex situ and in situ. The in situ treatment is expected to be less expensive and more easily applicable, since there is no need for moving large amounts of soil from one location to another. However, the design and installation of equipment to improve the microbial activities should be considered, which can increase the operation costs (Azubuike et al. 2016). In this technique, the microorganisms are placed in the area, along with oxygen adjustment through aeration and nutrient supply, permitting the surveillance and efficient removal of the target contaminant. It is widely used to remediate petroleum contaminated areas. On the other hand, the ex situ application is based on the removal of soil or water from the contaminated area in order to decontaminate and thereafter, the material is bring back to the original location. The ex situ is known to be more efficient and faster than the in situ method and can be applied in two most common ways, the slurry-phase in bioreactors, and the solid-phase, which includes landfarming, biopiles, and composting techniques (Kumar et al. 2018a, b, c).

The efficiency of bioremediation is directly related with environmental parameters as the pH, moisture content, and temperature. These aspects have direct influence on microbial growth and decontamination rate in such a way that studies in this area aims to enhance the efficiency of the technique through the understanding of the influence of these parameters on bioremediation mechanisms. The researchers intend to make the microorganisms capable to remove contaminants in field conditions, in addition to controlled simulations as laboratory conditions (Sharma et al. 2018a, b).

One of the factors most influencing the bioremediation is the availability of nutrients, since it impacts the growth of microorganisms, reflecting directly in the biodegradation rate and effectiveness. Mainly N, C, and P are essential nutrients for bacterial activities. The adequate C:N ratio is essential in order to guarantee the complete degradation of the target compounds, increasing the metabolic activity of microorganisms (Huang et al. 2018).

The temperature is another parameter widely affecting the bioremediation process, since it determines the survival of organisms and the inactivation/activation of their metabolism. The enzyme activities that participate in degradation of contaminants present a specific optimum temperature, where it reaches the maximum, thus declining when this value is changed. Moreover, this physical parameter is responsible for directly influencing physiological proprieties of microorganisms (Hong et al. 2020).

Regarding the concentration of oxygen, it influences the degradation condition of contaminants, promoting the aerobic or anaerobic condition. The necessity of aeration aims to increase the oxygen level, since the element is the main electron acceptor for aerobic bioremediation. The pH of environment widely affects the bioremediation process since there are values for optimum growth and removal of contaminants by the microbial metabolism. Any slight alteration from this value can change metabolic processes and affect the removal. The moisture content also affects directly the bioremediation, since each microorganism presents a requirement for its growth and development, and it can change the availability and solubility of contaminants (Kumar et al. 2018a, b, c).

Some additional factors may be considered when the bioremediation is intended to be applied to an area, according to Das and Dash et al. (2014). Firstly, it is related with levels, toxicity, mobility, and degradation property of the contaminants. Thus, it is highly recommended that the location should be scanned and properly characterized, aiming the determination of the real extent of the contamination and probable transport to other areas. Regarding the degradability of the contaminants, this property is essential in determining the type of microorganism able to perform the environmental removal and the time for remediating the area. Some compounds present slow degradation rate due to its high molecular weight, presence of complex ring structures or chlorine in its composition, affecting directly the time for achieving the expected results.

Secondly, it is necessary to verify the vicinity areas in order to prevent the contamination in achieving population and/or environmental receptors. In addition, in order to select the best bioremediation technique for the location, the future use of the bioremediation site might be considered, along with planning the most adequate monitoring of physicochemical and biological parameters (Das and Dash 2014).

Among the inorganic contaminants to which bioremediation can be applied for removal, heavy metals stand out. The bioremediation of heavy metals has widely been studied, as showed by the research of Talukdar et al. (2020), where a consortium using two fungal isolates, namely *Aspergillus fumigatus* and *Aspergillus flavus* was reported as presenting bioremediation potential of highly toxic heavy metals Cr (VI) and Cd (II). This high affinity of fungal consortium reveals great ability for uses in environments contaminated with heavy metals.

The microorganisms that can perform efficient bioremediation are widely listed in the literature, with different contaminants being removed from environment with this technique. Some bacterial genera such as *Pseudomonas*, *Rhodococcus*, *Sphingomonas*, and *Achromobacter* have been extensively used in bioremediation since they present ability of converting pollutants to non-toxic compounds (Liu et al. 2019).

Organic contaminants are also widely studied in research area. The petroleum hydrocarbons have successfully been bioremediated, for example, in a study using vermicomposting process bioaugmentating with indigenous bacterial consortium isolated from petroleum oily sludge, composed of *Acinetobacter radioresistens* and *Enterobacter hormaechei* (Koolivand et al. 2020).

In addition, among these several organisms that are able to remove several contaminants from environment, the marine organisms are extremely important and are receiving attention of studies, considering the great potential for uses in bioremediation technique (Mohanrasu et al. 2020). The bioremediation of plastics, more specifically low-density polyethylene (LDPE), was reported as efficient by the marine bacteria *Microbulbifer hydrolyticus*. The degradation of LDPE by the strain was evidenced by the modification on material surface seen with scanning electron microscopy and the changes in functional groups identified by infrared spectroscopy analysis. The discovery of a novel microorganism able to degrade polyethylene highlights the relevance for application of this bacteria in bioremediation of other contaminants with further studies (Li et al. 2020).

The advantages of application of bioremediation are related with the possibility of remediation on site, thus reducing the physical disruption of the contaminated area and need for transportation of large amounts of contaminated soil from one location to another. Besides that, the residues generated in treatment are harmless compared to other techniques, being mainly water, carbon dioxide, and cell biomass. Additional benefits that can be cited are the cost effectiveness and popular acceptance, since it is a sustainable and eco-friendly approach not generating chemical hazardous waste and thus not demanding disposal of products. The transformation of contaminants into harmless products allows the technique to avoid the future liability related with treatment and disposal of material (Kumar et al. 2018a, b, c).

On the other hand, the disadvantages of bioremediation are related with limitations regarding the biodegradation of some pollutants, that can be incomplete or even undegradable. The use of microorganisms requires the adequate selection of species with affinity with the target pollutant, besides the environment conditions and nutrient supply that must be satisfactory for the efficient bioremoval. Another important point is the time demanded for the organisms to perform the bioremediation, which can be longer than other techniques (Morillo et al. 2020).

The combination of several contaminants also represents a challenge to microorganisms to overcome and perform bioremediation. In addition to type of contaminants, they can occur in distinct phases, as liquid, solid, or gas. The studies in this area aim to select the most suitable organism for performing the contaminant removal; and another difficulty is going from pilot-scale to real conditions, with different factors affecting the microbial metabolism (Abatenh et al. 2017).

Another important type of bioremediation process is through enzyme activity. Enzymes are biological macromolecules that can enhance several chemical reactions, enhancing the reaction rate by lowering the activation energy of specific substrate molecules. There are several enzymes that can act in biodegradation, being the oxygenases that catalyze the oxidation of aromatic compounds, an important example (Gao et al. 2021). Another example are the carboxylesterases that catalyze the hydrolysis of carboxyl ester bonds, a functional group widely present in synthetic pesticides like organophosphates (Zhang et al. 2020). Finally, the genetic engineering applied to enzymatic bioremediation aims to increase the half-life,

activities, and stability of enzymes, along with production of recombinant types (Sharma et al. 2018a, b).

The future prospects in bioremediation consists in overcome the costs and the need of isolation and characterization of microorganism and enzymes for bioprospecting new organisms. In this sense, the techniques of metaproteomics, metagenomics, and biomolecular engineering can be applied for identification and characterization of organisms naturally found in different environments, besides enhancing the biodegradation ability (Gu et al. 2021). More specifically, metagenomics involves the understanding of genetic material from environmental samples. The technique aims to identify new bacteria or genes encoding for specific enzymes able to biodegrade contaminants (Datta et al. 2020). On the other hand, metaproteomics studies the proteins involved in physiological responses caused by pollutants in the organisms, aiming the identification of new metabolic pathways (Zakaria et al. 2021). Lastly, the biomolecular engineering includes the use of engineering biomolecules as nuclei acids and proteins, with biomolecular processes aiming improvement of bioremediation process (Pandey et al. 2018).

# 8.3 Bioremediation Techniques

Different bioremediation techniques have been used as management tools for recovering and remediating contaminated environments. In the process of removing environmental contaminants, bioremediation uses biological agents, mainly microorganisms, to eliminate, reduce, contain, or transform dangerous substances into less toxic or non-toxic ones (EPA 2001, 2002; Saxena and Bharagava 2016; Bharagava et al. 2019). Bioremediation is classified into two main types, in situ bioremediation and ex situ bioremediation. Different in situ and ex situ bioremediation methods are discussed in the following sections.

# 8.3.1 In Situ Bioremediation

In situ bioremediation involves the treatment of harmful substances at the contamination site (Kumar et al. 2018a), with no need to excavate, remove, or transport toxic waste from its original position (Enerijiofi 2021). Pollution is eliminated or mitigated directly where it occurs or at the site of contamination. It is an ecologically correct and low-cost technique, with the possibility of treating large volumes of contaminated material (Mishra et al. 2020).

Different techniques can be employed for in situ bioremediation, such as bioattenuation, bioventing, bioaugmentation, biostimulation, biosparging, bioslurping, biofilters, bioleaching and biomining, phytoremediation and mycoremediation.

#### 8.3.1.1 Bioattenuation

This bioremediation technique is also known as natural attenuation, which involves the passive and unattended remediation of a contaminated area without the need for human interference. Although the bioattenuation process involves the absence of any external forces, regular monitoring is necessary for the process to be sustainable and successful (Azubuike et al. 2016; Vásquez-Murrieta et al. 2016; Enerijiofi 2021).

Bioattenuation includes physical, chemical, and biological processes, which cause a reduction in the concentration of pollutants, mass, or toxicity (Vásquez-Murrietaet al. 2016). Anaerobic and aerobic microbial processes in the treatment of recalcitrant and biodegradable pollutants are also involved (Enerijiofi 2021). This technique is applicable to contaminated sites with a low concentration of contaminants, where other remediation techniques do not apply (Vásquez-Murrieta et al. 2016; Ossai et al. 2020).

### 8.3.1.2 Bioventing

In the process of degradation using the bioventing technique, nutrients and air (oxygen) are supplied in a controlled manner through wells to the contaminated site (i.e., soil) to stimulate microbial growth and proliferation (Enerijiofi 2021). This technique uses reduced airflow rates, providing only the amount of oxygen needed for biodegradative processes, reducing volatilization and release of pollutants into the atmosphere to lower levels (Atlas and Philp 2005).

Biodegradation involves the supply of oxygen and nutrients through aqueous solutions in contaminated soils, allowing the stimulation of native bacteria, thus facilitating the decomposition of biogenic pollutants (Tyagi and Kumar 2021). This technique is applied for removing pollutants in the depths of the surface (vadose zone), biodegradation of petroleum hydrocarbons, reducing volatile compounds (VOCs) rising to the surface, spilled light oil, absorbed fuel residues, non-chlorinated solvents and pesticides (Höhener and Ponsin 2014; Kumar and Gunasundari 2018; Olu-arotiowa et al. 2019).

Some factors must be considered before applying the bioventing technique, such as adequate concentration of pre-existing microorganisms; airflow conditions; the soil pH (6 to 8); hot temperature and water table height—when the water table is close to the surface, then the bioventing is not effective. Other conditions for the ineffectiveness of bioventing are low temperatures and moisture content in extremes levels, low or high (Kumar and Gunasundari 2018).

#### 8.3.1.3 Bioaugmentation

Bioaugmentation involves adding native microbial and exogenous microbial cultures, microbes or genetically modified communities with specific catabolic activity, at the contamination site, to increase the rate of biodegradation (Andreolli et al. 2015; Nwankwegu and Onwosi 2017; Poi et al. 2017; Enerijiofi 2021).

At bioaugmentation, active agents are characterized by the ability to degrade the contaminant and their resilience to the prevailing environmental conditions, as well as its genetic stability and ability to survive under competition (Gentry et al. 2004).

Bioaugmentation can be applied for treating wastewater and contaminated soil. It is used in the degradation process of substances such as polyvinyl compounds, PAHs (pyrene and benzo[a]pyrene, pesticides, insecticides, trinitrotoluene (TNT), aromatic and chlorinated hydrocarbons) (Kumar et al. 2018b; Olu-arotiowa et al. 2019; Sarkar et al. 2020). It has also been widely used in the treatment of medium and long chain alkanes, xenobiotic compounds, water and soils contaminated with polycyclic and monocyclic aromatic hydrocarbons and others (Akinde et al. 2012; Pal et al. 2017; Varjani 2017).

#### 8.3.1.4 Biostimulation

Biostimulation is applied in the treatment of pollutants such as metals and hydrocarbons (Ossai et al. 2020). It consists of the addition of materials (biosurfactants, bulking agents, biopolymers, nutrient correctives, and slow-release fertilizers) to stimulate the growth of indigenous microorganisms capable of degrading various compounds (Adams et al. 2015; Agarry and Latinwo 2015; Lim et al. 2016; Wu et al. 2016).

Substrates containing micronutrients (copper, chlorine, iron, magnesium, manganese, silicon, sodium, and zinc), macronutrients (phosphorus, potassium, and nitrogen), and organic nutrients are essential to improve the degradative capacity of foreign or indigenous microorganisms, and as an example, the concentration and the types of nutrients interfere with the biodegradation of polycyclic aromatic hydrocarbons (PAHs). The ability to deliver nutrients, C:N:P ratio 30:5:1 for equilibrium growth, presence of target microorganisms, ability to stimulate target microorganisms, are requirements for biostimulation (Hazen 2010; Ossai et al. 2020).

#### 8.3.1.5 Biosparging

In the biosparging technique, air (oxygen) and nutrients are injected into the saturated zone under pressure to stimulate the biological activities of indigenous microorganisms to degrade contaminants, thus increasing the concentration of oxygen in ground water (Azubuike et al. 2016; Enerijiofi 2021). This technique is similar to bioventing (Kumar et al. 2018b), being used to reduce the concentration of the contaminant dissolved in ground water or adsorbed to the soil (within the capillary fringe above the water table).

Factors such as pollutant degradability and soil permeability interfere with the effectiveness of the technique (Philp and Atlas 2005; Kumar et al. 2018a), as well as

predicting the direction of the injected airflow can limit its application, along with a high rate of airflow to achieve the volatilization of the pollutant and promote degradation (Ossai et al. 2020).

Biosparging is widely used in the treatment of aquifers contaminated with medium-weight petroleum hydrocarbons (jet and diesel fuels) and other compounds (xylene, benzene, ethylbenzene, and toluene) (Kumar and Gunasundari 2018; Kumar et al. 2018a).

#### 8.3.1.6 Bioslurping

Bioslurping or multiphase extraction is used to remove the free product that is floating on the water table. It combines vacuum-enhanced free product recovery with bioventing. Thus, vacuum-enhanced free product recovery extracts Light Non-Aqueous Phase Liquid (LNAPLs) from the capillary fringe and groundwater (Kumar and Gunasundari 2018; Mishra et al. 2020).

Bioslurping combines several processes, extracting steam from the soil, vacuum pumping, bioventilation, to remove contaminants from groundwater and soil, using an indirect supply of oxygen and stimulating microbial biodegradation (Vidali 2001). The low permeability of the soil reduces the rate of oxygen transfer by limiting microbial activity, which limits the use of the technique.

As bioventing is used in the bioremediation of contaminated soil/water, a combination of techniques (bioventing and bioslurping) can be designed, mainly to control the release of gas from soil and groundwater. Petroleum hydrocarbon compounds and LNAPLs are treated using bioslurping and bioventing. This technique is economical, and applicable to places with deep water table (Kumar and Gunasundari 2018; Mishra et al. 2020).

### 8.3.1.7 Biofilters

Biofilters are used to degrade pollutants (gaseous) in industrial air emissions. Columns embedded with microbes are applied to eliminate gaseous pollutants (Boopathy 2000), which means that the community of microorganisms that grow on a solid surface is able to immobilize or degrade the pollutant. Thus, the air containing toxic pollutants when passing through the biofilter is adsorbed on biofilms, incorporated into microbes, degrading the pollutant. Several materials are applied as biofilters, among them, peat, soil, agricultural residues (husks), and others. Biofilters are used to remove terpenes, mercaptans, dimethyl sulfides, hydrogen sulfide, ethylbenzene, sulfur gases, and other compounds from the air (Boopathy 2000; Kumar et al. 2018b).

#### 8.3.1.8 Bioleaching and Biomining

The solubilization of sulfide ore metals or solid residues in aqueous solutions using live microorganisms is known as bioleaching or biomining (Shukla et al. 2017). Bioleaching occurs at the mineral–microorganism interface. Most of the microorganisms that participate in bioleaching grow attached to the surfaces of sulfide ores, as natural biofilms (Senel and Hanay 2017; Shukla et al. 2017). Bioleaching can be applied to recover metals. As it involves the use of microorganisms, the following factors must be considered in its application: (a) the microbial ability to resist toxicity and the heterogeneity of the waste; (b) type of microorganism and its growth rate; (c) inoculum concentration and cell genotype; (d) physical–chemical conditions of the pollutant and microorganisms; (e) nutrients required by microorganisms (Priya and Hait 2017; Minimol et al. 2020).

On an industrial scale, the process of bioleaching can be carried out through bioreactors, considering the behavior and principles of bioleaching of a specific microbe. In this sense, the bioleaching process can be employed by continuous or batch operation mode.

When the microorganism and the material containing the metal are simultaneously added and incubated for solubilization, the technique is named one-step bioleaching (Yang et al. 2008; Rasoulnia and Mousavi 2016). Thus, when using this technique, microbial growth can be inhibited by the presence of metals leached in solution, which reduces the leaching rate. Microbes can take longer to acclimatize in the medium containing leachate metals, so the latency phase would be longer; leachate metals can be adsorbed, accumulated, or used in cells, thus reducing metal recovery (Jagannath et al. 2017).

When the addition of metal-containing solids after the pre-cultured microorganism reaches the late exponential growth phase, it is named two-stage bioleaching (Pradhan and Kumar 2012; Işıldar et al. 2016). In this process, the inhibitory effect of metals on growth is reduced, but, even so, it can lead to adsorption and accumulation of metals leached in cells, which can reduce the recovery capacity, and reduce the time in the growth delay phase (first stage) (Minimol et al. 2020).

The average spent bioleaching is the dissolution of metals from solids in the cellfree supernatant that contains extracellular proteins and secondary metabolites (Ilyas et al. 2013; Natarajan and Ting 2013). Thus, in this process, extracellular proteins and metabolites, which tend to be secreted only in the presence of metals, would not be available for any leaching action. However, the inhibitory or toxic effect of metals on microbial growth can be prevented. As for the time required for microbial growth, this may be shorter during the production of the medium, as the latency phase period is shorter and the growth rate in the absence of metal-containing residues is higher (Minimol et al. 2020).

To select which type of bioleaching process will be applied, requirements such as metal-microbe interaction involved, inhibitory nature of the metals, and type of microbes chosen must be considered.

Bioleaching is a low-cost and simple technology that can be applied in the removal of mineral compounds and low-quality ore metals, as well as in the treatment of industrial effluents. The technique is applied in biomineral processing, because the process of bioleaching metals by microbes has the possibility of recovering metals from mineral resources not available by conventional mining. Thus, microorganisms act as biocatalysts in the conversion of metallic compounds to their water-soluble forms in the process of bioleaching (Varjani et al. 2018).

#### 8.3.1.9 Phytoremediation

Phytoremediation (from Greek "phyto," plant and Latin "remedium," cure) refers to the use of plants to contain, reduce, confiscate, accumulate, extract, eradicate, and/or decontaminate organic and inorganic contaminants from soils, sediments, surface, and groundwater (Sharma et al. 2018a, b; Enerijiofi 2021; Mishra 2021; Saxena et al. 2021).

It is one of the methods in situ, ecologically correct, it is a process considered inactive, cheap, with low specialization. The main contaminants that can be removed by phytoremediation are solvents, crude oil, hydrocarbons, pesticides, metals, explosives, and landfill leachate (Wang et al. 2017).

The phytoremediation process can be broadly classified based on fundamental processes, type of contaminant, and applicability (Table 8.1).

Thus, in phytoremediation of organics, the plant metabolism contributes to the reduction of contaminants through the breaking, transformation, volatilization, or stabilization of contaminating compounds in water and soil. In this way, phytodegradation breaks down the organic compounds absorbed by the plant, into simpler molecules and incorporates them into its tissues. Plants contain enzymes like oxygenases, dehalogenases, and reductases that can convert and degrade chlorinated solvents, ammunition residues, and others (Ghosh and Singh 2005; Malode et al. 2013; Mishra 2021; Saxena et al. 2021).

#### 8.3.1.10 Mycoremediation

Due to the ability to produce and secrete extracellular enzymes, such as laccases, polyphenol oxidases, lignin peroxidases, which can break down cellulose and lignin, fungi are used to degrade several environmental recalcitrant pollutants (Singh 2006; Jang et al. 2009). Thus, mycoremediation uses fungal mycelia to carry out the biodegradative process, which converts contaminants in less toxic or non-toxic forms, remediating contaminated soils and groundwater (Gadd 2001; Singh 2006; Anderson and Juday 2016; Ali et al. 2017; Kumar et al. 2018c; Enerjijofi 2021).

Fungi are often more tolerant to pollutants and their hyphae are able to penetrate the soil and reach the pollutant much faster than other microbes (Reddy and Mathew 2002; Harms et al. 2011). They end up being more efficient in bioremediation than other microorganisms. White rot fungus has been used for biotransformation of

Technique	Description
Phytodegradation	This technique involves the absorption, storage, and decomposition of contaminants by the activity of proteins and enzymes produced by plant tissues and microorganisms present in the rhizosphere (Enerijiofi 2021; Saxena et al. 2021).
Phytoextraction	In this method, plants accumulate and transport pollutants in aerial part and produce a mass of plants containing pollutants that are later transported for recycling and/or disposal (Mishra 2021; Saxena et al. 2021).
Phytofiltration	It uses the absorption capacity and the large surface area of the entire plant to remove the pollutant, for example, heavy metals from wastewater (Malode et al. 2013). The mechanism is similar to phytoextraction and aquatic plants and terrestrial plants can be used in phytofiltration.
Rhizofiltration	In rhizofiltration there is a mutual relationship between the plant roots and rhizosphere microorganisms creating an environment where pollutants are broken down through metabolic activities and secretion of proteins and enzymes from the plant's root system (Malode et al. 2013; Enerijiofi 2021; Mishra 2021; Saxena et al. 2021).
Rhizodegradation or Phytostimulation	Contaminants are degraded by microbial activity in the rhizosphere. It is a slower process than phytodegradation. In this technique, microorganisms consume and digest organic substances such as fuels and solvents (Malode et al. 2013; Saxena et al. 2021).
Phytostabilization	The pollutant in the soil or water is immobilized by absorption or precipitation by the plant roots, reducing their mobility by forming a stable vegetable mass with the pollutant, thus preventing the entry of the pollutant into the environment (Mishra 2021; Saxena et al. 2021).
Phytotransformation	The technique involves the process by which the plant absorbs and transforms highly toxic organic contaminants, from a contaminated site (soil, sediment, and water body) into less toxic forms (Mishra 2021; Saxena et al. 2021).
Phytovolatilization	The technique uses plants to remove volatile pollutants from the environment. Plants absorb pollutants and transfer them to gaseous substances, which are then released into the atmosphere (Malode et al. 2013). Volatile organic compounds are the best candidates for this process to be treated with the aid of evapotranspiration (Mishra 2021).

 Table 8.1
 Types of phytoremediation

pesticides, degradation of petroleum hydrocarbons and lignocellulolytic wastes in the pulp and paper industry. Some common types of macroscopic fungi used in mycoremediation are described in Table 8.2.

Mycoremediation can be effective in breaking down some chlorinated compounds and petroleum hydrocarbons, as well as being, capable of stimulating native microbes and enzymes in situ, and bioaccumulating heavy metals, reducing contamination (CRC 2018). It also employs three types of mechanisms (mycosorption, mycodegradation, and mycoenzymes) for the removal of organic compounds

Type of fungi	Target contaminants
Button mushrooms	Cadmium
Elm oyster	Dioxins, wood preservatives
King Stropharia	E. coli and other biological contaminants
Pearl oyster	Cadmium, mercury, dioxins, polycyclic aromatic hydrocarbons (PAHs); polychlorinated biphenyls (PCBs)
Phoenix oyster	Cadmium, copper, mercury
Shaggy mane	Arsenic, cadmium, mercury
Shitake	PAHs, PCBs, pentachlorophenol
Turkey tail	Organophosphates, PAHs, mercury

Table 8.2 Common fungi used in mycoremediation. Source: CRC (2018)

(Noman et al. 2019). Some parameters must be considered when applying the mycoremediation technique, such as soil pH, temperature, and availability (or lack) of oxygen. Mycoremediation is a sustainable, low-cost, low-maintenance technique, presenting fast results (usually completed in a short time (weeks/months)) (CRC 2018).

# 8.3.2 Ex Situ Bioremediation

Ex situ bioremediation involves the collection/transport (excavation or pumping) of polluting substances from the place of origin to a controlled and designed area, where their decontamination will occur through physical–chemical or biological methods (Varjani et al. 2018). Decontamination involves the treatment of the pollutant after it is transported from the area of occurrence to the treatment area, which facilitates microbial degradation. This removal of the pollutant from the place of origin to the treatment area increases operational costs (Azubuike et al. 2016).

Different techniques can be applied for ex situ bioremediation, such as landfarming, composting, biopiling/biopiles or windrows, bioreactors. These techniques are used efficiently in the treatment of contaminated soil or water (Ghangrekar et al. 2020). Ex situ bioremediation techniques can be easier to control and faster, in addition to the possibility of treating a wide range of contaminants and soil types, in comparison with in situ techniques.

When the treatment of contaminated surface or underground water occurs through the extraction of contaminated water from the place of origin to the place of distant treatment, its remediation is known as the "pumping and treatment system." This system can take years to treat the contaminated area, because the complete discharge of the pollutant requires that a large volume of water be treated continuously over a long period of time (Varjani et al. 2018).

#### 8.3.2.1 Landfarming

Landfarming is a biodegradation technique consisting of excavating and extracting contaminated soil and spreading it over a thin area (the soil layer support fixed above the ground) and periodically plowing the soil to stimulate aerobic biodegradation of pollutants by indigenous microbes (Senel and Hanay 2017; Yadav et al. 2017; Mishra et al. 2020).

Landfarming can be applied only for the treatment of 10–35 cm of soil surface (Kumar et al. 2018a) and excavation of contaminants is not recommended if the polluted soil is less than one meter deep (Kumar and Gunasundari 2018). The application of landfarming is advantageous because it has reduced maintenance and monitoring costs, it is simpler and more economical, however, the main disadvantage of this process is the limitation to treat 10–35 cm of top soil (Williams 2006; Yadav et al. 2017; Mishra et al. 2020).

In the landfarming process, nutrients, microorganisms, and moisture can be added to the soil. By providing moisture to the soil through seasonal spraying of water, it forms a barrier around the contaminated soil, controlling erosion and minimizing the formation of dust when plowing the soil to maintain aeration (Kumar and Gunasundari 2018). Plastic or clay liners can also be used on site, before laying contaminated soil, to prevent the leaching of contaminants into groundwater (Senel and Hanay 2017).

Thus, the treatment of contaminated soil occurs through biodegradation combined with aeration and light photooxidation (Senel and Hanay 2017). This type of treatment is more efficient in areas with hot and humid climate, and in a sunny place. Therefore, during periods with colder temperatures (winter), when snow covers the soil, there is a reduction in the biodegradation process (Mishra et al. 2020).

Although land cultivation is considered an ex situ bioremediation technique, in some cases it applies to in situ bioremediation. This technique can be applied in the treatment of sites contaminated by PCBs, aliphatic hydrocarbons, and polycyclic aromatics (Silva-Castro et al. 2012; Kumar et al. 2018b).

#### 8.3.2.2 Composting

Composting is a technique that involves mixing polluted soils with bulking agents, for example, harmless organic additives (organic manure, wood chips, and agricultural waste such as plant waste) (Kumar et al. 2018b). The additives favor the growth, development, and proliferation of microbial populations (bacteria) capable of degrading the pollutants found in the soil to be treated (Adam et al. 2017). Thus, the pollutants will be degraded and converted into a stabilized final product, which can be applied as a soil conditioner to remedy soil contaminated with organic compounds or be safely discarded in the environment (Cai et al. 2017; Saum et al. 2018; Ren et al. 2018; Kumar et al. 2018b). In the composting process, parameters like the addition of appropriate microbial consortia and nutrients, cultivation, irrigation, as well as bulks materials in the form of organic residues to improve bioremediation, must be considered (Prakash et al. 2015). This process is conducted outdoors and can be carried out by means of composting in open and static windrow systems. In static windrow systems, the piles are aerated by a forced air system, while in open windrow systems, the compost is loaded in elongated stacks (Cunningham and Philip 2000; Girma 2015).

In order to properly compost soils contaminated with dangerous compounds, for example, petroleum hydrocarbons, the composition needs to occur in a thermophilic phase of 50–65 °C. During the decomposition process of organic materials present in the compost, there is an increase in temperature due to heat generated by microbial activities (Prakash et al. 2015; Ossai et al. 2020). Thus, the aerobic process is viable because the heat produced during the biodegradation reactions with the release of oxidative energy can lead to a substantial increase in temperature (Adam et al. 2017). The efficiency of degradation and the growth of microorganisms must be monitored under the level of humidity, aeration, and temperature.

The composting technique can be used to treat explosives (HMX, TNT, and RDX), hydrocarbons, dangerous chemicals, and others (Kumar et al. 2018b).

#### 8.3.2.3 Biopiles or Windrows

Biopiling or stacking technique is a combination of methods of agriculture (landfarming) and composting, in anaerobic cells designed with an irrigation and nutrient system, vacuum pumps and blowers, leachate collection system for bioremediation of polluting components adsorbed to sediments and soil (Wu and Coulon 2015; Benyahia and Embaby 2016; Kim et al. 2018; Mishra et al. 2020).

The technique involves stacking a contaminated excavated soil, which can be placed in piles (biopiles) or rows (windrows) and subsequent soil correction with nutrients, biostimulation, and forced aeration to increase microbial degradation activities (Azubuike et al. 2016; CRC 2018).

As a hybrid system, biopiling is a favorable system to stimulate microbial growth (anaerobic microorganisms and indigenous aerobes). This system increases aerobic creosote catabolism by inoculating air in piles of contaminated soils (Kumar et al. 2018a) and is suitable for the treatment of large volumes of contaminated sediments and soils in a limited space and in addition to remediating contamination in extreme environments (Whelan et al. 2015).

Biopiles can have heights in the range of three to ten feet (about 1 to 3 m) and can be applied in the treatment of surface contaminants in order to control the physical losses of the contaminants by volatilization and leaching (Azubuike et al. 2016; Kumar et al. 2018a). It can also be used to treat soils contaminated with low molecular weight pollutants and petroleum hydrocarbons (phenols, PAH, sand BTEX). However, biopiling can limit the volatilization of low molecular weight compounds (Dias et al. 2015; Ossai et al. 2020). Biopiles involve some limitations, when compared to other ex situ bioremediation techniques, as they require much more space and when there is an extreme heating of the air, it leads to excessive drying of the soil to be treated, therefore occurring an interruption of microbial activities (Sanscartier et al. 2009).

### 8.3.2.4 Bioreactors

A bioreactor involves a controlled container to create a three-phase mixture condition (solid, liquid, and gaseous) to increase the rate of degradation of the water-soluble and soil-bound contaminant. Unrefined materials are transformed into specific non-toxic products due to a series of different biological reactions. Thus, bioreactors are used to degrade contaminants under controlled conditions. They are designed containment systems (reactors or aqueous sludges) that are applied for the treatment of polluted solid materials (sediment, soil, and sludge) and water (Vidali 2001).

Bioreactors are used to treat water or soil contaminated with volatile organic pollutants (toluene, xylene, ethylbenzene, and benzene) (Azubuike et al. 2016). Its use in the biodegradation process is much more efficient, faster, and beneficial than in other techniques, because in these systems, bioremediation occurs in a controlled way. Therefore, parameters such as pH, temperature, amount of nutrients, humidity, bioavailability of pollutants, mass transfer, and others are monitored to improve the biological reactions used in the biodegradation of pollutants, increasing efficiency in the bioremediation process (Mohan et al. 2004; Azubuike et al. 2016; Kumar et al. 2018a).

The main disadvantage of using bioreactors is due to the fact that it spends extra safety measures in transporting the pollutant from the place of origin to the treatment site, as well as treating high volumes of contaminants, which increases the cost of applying the technique (Philp and Atlas 2005; Kumar et al. 2018a). Another disadvantage is that contaminated soil requires pre-treatment, or the contaminant should be removed from the soil through physical extraction or soil washing, before being placed in a bioreactor (EPA 2002; Mishra et al. 2020).

### 8.4 Importance of Bacteria for Bioremediation

Bioremediation has been considered as the remediation of pulled sites employing microbes, and in this context, bacteria play a fundamental role (Azubuike et al. 2016). As discussed above, all the bioremediation approaches have been classified as in situ or ex situ depending on the polluted material sites. Bioremediation is mainly a prospective method for in situ subsurface remediation with a reduced cost and production of secondary pollution compared with other environmental remediation techniques, these are the motivations for the consideration as eco-friendly and cost-effective approach (Wei et al. 2012; Igiri et al. 2018). Furthermore, two main

strategies have been employed to improve the effectiveness of in situ bioremediation, specifically, bioaugmentation that comports the addition of pre-grown microbial cultures to enhance the degradation of unwanted compounds; whereas biostimulation includes the injection of nutrients and other supplementary components to the native microbial population inducing the propagation at a hastened rate (Tyagi et al. 2011). Thus, for the bioremediation the selection of the appropriate strategy to treat a polluted site has become pivotal, three basic principles have to be considered: the pollutant tractability for biological transformation to less toxic products, the physical accessibility of the contaminant to microorganisms, and also the opportunity for optimization of biological activity (Dua et al. 2002). In this context, the wide metabolic diversity of bacteria is crucial for bioremediation of many different contaminated sites.

## 8.4.1 Removal of Heavy Metals

Microbial activities strongly influence metal speciation and transport in the environment. In bacteria a number of specific resistance mechanisms, indeed, including active efflux and sequestration or transformation to other chemical species become functional at concentrations above the homeostatic or non-toxic levels (Silver 1998).

For this reason, different organisms exhibit diverse responses to toxic ions, which confer upon them a certain range of metal tolerance. A family of metal-chelating proteins named metallothioneins (MTs) represent a typical mechanism for regulating intracellular metal ions. Bacterial MTs were observed to confer resistance to Zn (II) and Cd (II) and have been considered as the main mechanism of tolerance to metals in the bacterial world (Robinson et al. 2001; Blindauer 2011). These mechanisms of resistance to metal ions are often plasmid-borne, which facilitates dispersion from cell to cell. A common example of heavy metal resistance was observed in *Ralstonia metallidurans* CH34 previously classified as *Alcaligenes eutrophus* CH34 (Goris et al. 2001).

The sulfate-reducing bacteria (SRB) represent another important process involved in the bacterial resistance to heavy metals. They have significant ecological functions in anaerobic conditions for complete mineralization of organic carbon particularly in marine sediments. Furthermore, these bacteria play an important role in bioremediation of contaminated sites (Jørgensen 1982; Ayangbenro et al. 2018). Sulfate can be biologically reduced to hydrogen sulfide by SRB with significant ecological functions, but also play an important role in bioremediation of contaminated sites. SRB can react with the dissolved heavy metal ions and transform them into highly stable metals sulfides, which are usually are more stable than the hydroxides produced by chemical treatment. In addition, metal sulfides can be recycled and reused (Jalali and Baldwin 2000; Kiran et al. 2017; Li et al. 2018).

Finally, another class of bacteria named acidophilic chemolithotrophs shown efficient growth at high metal concentrations, these are iron and sulfur-oxidizing bacteria. The metal resistance of these microbes is an adaptation to very acidic environments, where metal solubility is high including the thiobacillus group of bacteria. Interestingly, when the growth of *Thiobacillus ferrooxidans* is dependent on Fe (II), this bacterium is highly resistant to Al, Cu, Co, Ni, Mn, and Zn (0.1-0.3 M), although it remains quite sensitive to other metal ions (Hutchins et al. 1986).

Different strategies have been contemplated to improve the bioremediation of heavy metals (Kocur et al. 2016; Liu et al. 2018). In this context, microbes represented an optimal strategy, as they have various mechanisms in response to heavy metals that can be applied for bioremediation of various contaminated sites (Kushwaha et al. 2018; Cornu et al. 2017; Ahemad 2019). In fact, as the role of bacteria and plants in biotransformation of heavy metals into non-toxic forms is well documented, the study of these biological elements at the molecular level permits to detect the mechanism of metal accumulation opening to biotechnological implications for bioremediation of metal-contaminated sites (Dixit et al. 2015). However, for the microbial side the success of the bioremediation action in pulled places depends mainly upon the kind of microorganism and the contaminants involved in the process (Kapahi and Sachdeva 2019). The combination of different biological strategies could lead to an improvement in bioremediation performance. Recently the triple treatment combining bacteria, plants, and invertebrate-like earthworms showed best yields in terms of removal rates and soil health improvements (Urionabarrenetxea et al. 2021).

# 8.4.2 Biodegradation of Polyaromatic and Halogenated Hydrocarbons

For the persistent organic pollutants, different bioremediation approaches need some improvements due to the long period of biological remediation determined by low degradation rates and the insufficient energy source (Atlas and Hazen 2011). Now-adays petroleum derivatives and organic waste are among the main environmental contaminants. Recently, the bacteria–fungi associations have been improved for the oil bioremediation of soils, making the process more robust against environmental changes that can perturb the bioremediation by bacteria (Quintella et al. 2019). To overcome the process velocity problem on the bioremediation of organic pollutants, the association of plants and bacteria, such as endophytic bacteria and rhizosphere bacteria, has been employed to contribute on the biodegradation of toxic organic compounds in contaminated soil with the potential improving of the bioremoval process (Divya and Deepak Kumar 2011; Gkorezis et al. 2016; Arslan et al. 2017).

Halogenated hydrocarbons are synthesized as industrially valuable materials, or produced as a by-product during chemical reactions, or being released as a result of burning of municipal waste; these are ubiquitous in our environment and several bacteria have been isolated and reported acting on their degradation (Akram et al. 2021). The potential of the nitrification process, where ammonia is oxidized to

nitrate, has been proposed for in situ bioremediation of halogenated compounds. In this process, ammonia-oxidizing bacteria play a key role starting the degradation of many halogenated hydrocarbons (Sayavedra-Soto et al. 2010).

Some microbes have evolved enzymes and metabolic pathways to detoxify and utilize halogenated aromatic compounds as their sole carbon sources. The degradation pathways can be simplified into three stages: an initial debranching process to remove additional moieties attached to the aromatic ring, an intermediate pathway dealing with halide removal and activation of aromatic rings by incorporation of oxygen (in the case of aerobes), and a final stage of ring-cleavage reactions to convert aromatic into aliphatic molecules which can be further converted into common metabolites of central metabolic pathways such as pyruvate, acetyl CoA, succinate, oxaloacetate, and acetaldehyde (Pimviriyakul et al. 2020). Among bacteria, *Pseudomonas* becomes a paradigmatic bacterial genus for the catabolism of aromatic compounds and for the bioremediation of toxic pollutants and the valorization of aromatic compounds present in biowaste (Nogales et al. 2017).

### 8.4.3 Biodegradation of Petroleum and Diesel

Organic pollutants such as petroleum-derived hydrocarbons are bio-transformed in presence of optimal ecological factors and the necessary nutrients by microbial metabolic activities. Most of the cases the biodegradation takes longer than traditional remediation methods, however the complete degradation is often accomplished. Hydrocarbon biodegradation in soil is determined by a variable number of abiotic and biotic factors, such as the pH, temperature, oxygen availability and nutrient content, and survival of hydrocarbon-degrading bacteria including the bioavailability of pollutants to microbial attack (Koshlaf and Ball 2017). To improve the degradation of oily sludge the combination of bioaugmentation and biostimulation treatments has been carried out with positive results (Varjani et al. 2020). Biodegradation of the petroleum hydrocarbons is a complex process that depends also on the nature and the amount of the hydrocarbons present into the contaminate place (Colwell et al. 1977). Bioremediation techniques based on free cell cultures have been largely used to remove compounds from contaminated areas. However, a recent promising technique to be employed in harsh environments included the immobilization of microbial cell systems, presenting the main advantages of possible reutilization of microorganisms and more tolerance to temperature and pH changes (Partovinia and Rasekh 2018). Besides the important chemicalphysical factors affecting diesel biodegradation, the bacterial species have been reported as the major factor that affect the biodegradation performance. Specifically-isolated bacteria seem to be a very promising approach to remediate diesel-contaminated environments (Imron et al. 2020). The screening of bacterial strains isolated from diesel-contaminated soils permitted to discriminate biosurfactant (BS) production and biofilm formation abilities as fundamental characteristics of the selected bacterial strains (Balseiro-Romero et al. 2017). The bacterial bioremediation technologies as well as the fungi and algae bioremediation strategies may not be enough to remediate hydrocarbon-contaminated sites. In addition, in this case, it was comprehensively illustrated that the integration of remediating techniques can improve the degradation of petroleum hydrocarbons with an improving removal efficiency (Naeem and Qazi 2020).

# 8.4.4 Biodegradation of Plastics

The countless utilization of plastics is the result of their desirable characteristics, including light weight. durability, corrosion resistance, and low price (Raziyafathima et al. 2016). Consequently, the plastic pollution represents a significant environmental concern as a result of the persistence and potential adverse effects on biota. A number of scientific articles showed how bacteria can work on plastic material degradation, describing how microbial characteristics (e.g., biofilm organization) and environmental factors can affect the plastic biodegradation by bacteria (Yuan et al. 2020). Bacterial groups including Gram-positive and Gramnegative strains have been reported acting on plastic degradation (Razivafathima et al. 2016). Microbial biodegradation of plastics includes the polymers conversion into monomers, due to bacteria were not able to introduce into the cytoplasmic compartment some polymers, so they must first be depolymerized to smaller monomers through physical and natural processes (Swift 1997). Among the natural processes of depolymerization, microbial enzymes were observed acting on polymers before the monomer formation, absorption into microbial cells and completely biodegraded (Goldberg 1995). Strains of the genus *Bacillus* were detected as one of the main components of the North Pacific Gyre plastic fouling community (Carson et al. 2013). Characterization of the extracellular hydrolase enzymes secreted by Bacillus strains included lipase, carboxymethyl cellulase, xylanase, chitinase, and proteases with different levels of activity (Dang et al. 2018). The enzymes that are known to degrade plastic polymers usually belong to the class "hydrolases." Enzymes belonging to this class are involved in a catalytic reaction that causes a breakdown of chemical bonds of its substrate in the presence of water. Cutinase, lipase, and PETase (an esterase) are some of the most common enzymes associated with the degradation of plastics according to recent research developed in this area (Kaushal et al. 2021). All these enzymes act on the plastic polymer in a similar manner, causing hydrolytic cleavage of the long carbon chains and then assimilating these smaller subunits into the microbial cell for further enzymatic degradation and release of metabolic products.

# 8.5 Potential of Marine Bacteria for Bioremediation

Many of the marine pollutants are the result of direct or secondary human activities. Some of these substances are considered biodegradable, but others unfortunately not (Vikas and Dwarakish 2015). Marine bacteria possess a wide variety of bioremediation potentials, which are beneficial from both environmental and economic point of view (Amidei 1997). Diverse marine bacteria have been reported to have bioremediation potential and they can be utilized in the bioremediation process of heavy metals, hydrocarbons, and many other recalcitrant compounds (Fig. 8.1). The utilization of these microbes permits the in situ bioremediation including the direct use of microorganisms in any adverse conditions and possibly without any genetic manipulation (Dash et al. 2013).

It has been already estimated that in 2025, the oceans will contain more than 25 million metric tons of plastic litter (Jambeck et al. 2015). Plastics are considered the main concern in marine environment representing that more than 60% of all floating debris and the amounts increase each year. Chemical–physical factors start the plastic degradation processes forming polymer fragments susceptible to biodeg-radation (Gewert et al. 2015). Micro-nano plastics enter into the agro-ecosystem through the wildlife arriving into the human body following the food chain; the processes through which the plastic enter into the human body include ingestion or inhalation, causing various problems as consequence.



Fig. 8.1 Bioremediation potential of marine microorganisms. Bacteria isolated from different marine environments have been characterized for their ability for bioremoval and bioconversion of heavy metals and biodegradation of polyaromatic hydrocarbons, halogenated hydrocarbons, petroleum and derivatives and plastics

The microbial utilization of the plastic carbon content as energy source with complete degradation/removal of these components has been described as an efficient bioremediation method (Tiwari et al. 2020). Marine bacteria such as Bacillus cereus, Bacillus sphaericus, Vibrio furnissii, and Brevundimonas vesicularis shown capability to degrade nylon fibers in mineral salt medium at 35 °C and pH of 7.5 under submerged enrichment conditions with the polymer as the sole carbon source (Sudhakar et al. 2007). Efficient degradation of conventional high-density polyethylene (HDPE) can be mostly achieved by plastic-degrading bacterial isolates belonging to the genus *Bacillus* spp. and *Pseudomonas* spp. (Devi et al. 2019). In another research, the marine bacteria Brevibacillus borstelensis showed HDPE degrading activity (Mohanrasu et al. 2018). Moreover, other plastic-associated marine bacteria groups and families such as the families Erythrobacteraceae and Rhodobacteraceae (Alphaproteobacteria), Flavobacteriaceae (Bacteroidetes), and the phylum cyanobacteria (such as the Phormidium genus) have been frequently observed (Roager and Sonnenschein 2019).

Marine bacteria interaction with eukaryotic microorganisms such as diatoms was reported to be able forming aggregation called marine snow, which bring organic carbon and nutrients to the sea floor (Gärdes et al. 2011). In this context, biofilm formation by marine bacteria could became pivotal for the plastic particle elimination or reduction by the oceanic water column, increasing the speed by which this plastic makes its way to the sea floor. The presence of plastic nanoparticles significantly influenced the formation of biofilms both positively and negatively in a species-specific manner (Okshevsky et al. 2020). Deeper studies on these bacteria together with bacterial engineering approaches may help on this important marine environmental concern. The study on plastic-degrading microorganisms focused also on their searching in deep-sea sediments where temperature decreases below 4 °C (Ravenschlag et al. 1999) and it was possible to isolate microorganisms adapted to cold with unique characteristics (Urbanek et al. 2018). In this interesting environmental context, studies were conducted and detected bacteria with plastic degradation properties, including *Pseudomonas* spp., *Shewanella* sp., *Moritella* sp., Psychrobacter sp., Alcanivorax sp., Tenacibaculum sp. (Sekiguchi et al. 2009, 2011a, b;) Vibrio sp., (Raghul et al. 2014), Clonostachys rosea, and Rhodococcus sp. (Urbanek et al. 2017).

In marine environment oil pollution represents another important concern, it can occur from either catastrophic accidents (shipping disasters or pipeline failures) or natural oil seepages and biota. Various technologies have been developed to contrast these events and one of the reliable and eco-friendly was considered certainly the bioremediation with the objective to minimize the impact on the environment (Catania et al. 2015). When spilled in water, oil cause immediate and long-term repercussion. For this reason, clean-up methods and remediation were contemplated as fundamental to preventing the intoxication and death of a large number of living organisms. In this context, biotechnology applications also play a fundamental role (De Almeida et al. 2016). For petroleum pollution in seawater physicochemical methods alone were earlier defined not enough to contrast the problem (Prince 1997). However, the petroleum hydrocarbons being not soluble in water, it is

difficult to achieve the bioavailability for bioremediation as consequence surfactants are employed to improve the mobility of oil reinforcing the biodegradation process. Thus, an improved hydrocarbon solubility and the emulsification of hydrocarbon– water mixtures permit a better ability of oil-degrading bacteria to use hydrocarbons (Bao et al. 2012). The importance of the microbial biosurfactant production and its use were defined promising for the efficacy, low toxicity, and biodegradability working on the increasing biodegradation and solubilization process of insoluble compounds at extreme environmental conditions including at the common hydrocarbon derivatives, heavy metals, pesticides, and organic/inorganic contaminants (Deepika et al. 2021).

*Bacillus* genus was characterized as one of the main groups of bacteria able to synthesize biosurfactants as surfactin among the other families of cyclic lipopeptides (Jacques 2011). Moreover, *Bacillus* genus was detected as one of the main studied marine bacteria groups mainly for their easy cultivation in microbiological laboratories (Stincone and Brandelli 2020). Isolation of bacterial strains from seawater contaminated with petroleum derivatives permitted to discover promising biosurfactant-producing isolates belonging to the genus *Bacillus*, indicating their possible use in the bioremediation of marine environments (Durval et al. 2019). Microorganisms producing biosurfactants are efficient accelerators of the hydrocarbon biodegradation process by enhancing their bioavailability and facilitating their degradation by bacteria (Xu et al. 2020).

Another important group of bacteria include those acting on hydrocarbon biodegradation. In this group, a total of 79 genera of marine bacteria were identified and among these the following were the most representative: Achromobacter, Acinetobacter, Alcaligenes, Actinomycetes, Archrobacter, Bacillus, Cycloclasticus, Corvneforms, Chromobacterium, Flavobacterium, Micrococcus, Microbacterium, Mycobacterium, Nocardia, Pseudomonas, Sarcina, Serratia, Streptomyces, Vibrio, Xanthomonas (Xue et al. 2015). Features of microorganisms isolated from extreme environments can be employed for enhanced bioremediation of oil hydrocarbons, especially under aerobic conditions in moderate to high salinity conditions. In this condition, strains belonging to various genera have been shown to degrade hydrocarbons but in particular, members of the genera Halomonas, Alcanivorax, and Marinobacter dominate the literature (Fathepure 2014). The presence of both groups of bacteria, petroleum-degrading bacteria and biosurfactant-producing bacteria, improved the petroleum-degrading rate (Shi et al. 2019). Similarly, polycyclic aromatic hydrocarbons (PAHs) are widespread in marine ecosystems and they are usually resulting from human activities or even natural sources. PAHs adhere to sediments leading to accumulation in coastal and deep sediments and the microbial assemblages play a pivotal role on their degradation (Duran and Cravo-Laureau 2016). The metagenomic analysis of a contaminated coastal sediment allowed to detect three main phyla (Proteobacteria, Firmicutes, and Bacteroidetes) accounting approximately  $\geq 93.0\%$  of the total microbial community (Lee et al. 2018). Bacterial species like Cycloclasticus spirillensus, Lutibacterium anuloederans, and Neptunomonas naphthovorans have also been utilized in enhanced biodegradation of PAHs in marine environment (Hedlund et al. 1999; Chung and King 2001). Moreover, degradation of PAHs carried out by marine bacteria through mineralization process was reported by *Achromobacter denitrificans, Bacillus cereus, Corynebacterium renale, Cyclotrophicus* sp., *Moraxella* sp., *Mycobacterium* sp., *Burkholderia cepacia, Pseudomonas fluorescens, Pseudomonas paucimobilis, Pseudomonas putida, Brevundimonas vesicularis, Comamonas testosteroni, Rhodococcus* sp., *Streptomyces* sp., and *Vibrio* sp. (Samanta et al. 2002). Recently it was possible to identify *Mycobacterium* and *Sphingomonads* spp. as bioaugmentation and genetic bioaugmentation targets, respectively, due to their positive associations with PAHs and their high abundance and species diversity in contaminated soils (Redfern et al. 2019). Moreover, the marine *Stenotrophomonas acidaminiphila* NCW-702 in biofilm organization was able to degrade PAHs more efficiently as compared to planktonic cells, these findings supporting the efficacy of biofilms over planktonic culture in bioremediation applications (Mangwani et al. 2014).

Industrialization and urbanization activities have increased the flow of toxic metal ions to the marine environments, being harmful when present above critical values. Bacteria from metal polluted habitats are suitable candidates for heavy metal remediation, maintaining the sustainability as they were more capable for quick adjustment to the changing environmental factors such pH, salinity, temperature (Dash et al. 2013; Mohapatra et al. 2017). Bacteria own several mechanisms to act on tolerating and bioremediating high concentration of toxic heavy metals such as: precipitation as phosphates, sulfides, and carbonates through the biomineralization, volatilization via methylation/ethylation/reduction through the biotransformation process, ATP mediated efflux systems, intracellular bioaccumulation, biosorption and sequestration through Extracellular Polymeric Substances (EPS) (Naik et al. 2012).

An important process is represented by the biomineralization that happens continuously on the deep-sea minerals forming polymetallic nodules. In fact, free-living and biofilm-forming bacteria provide the matrix for manganese deposition, and cobalt-rich crusts (Wang and Müller 2009). The marine bacterium Idiomarina loihiensis MAH1, growing in environmentally relevant concentrations of uranium, was capable to form uranyl phosphate mineral phases, structurally resembling metaautunite [Ca(UO<sub>2</sub>)<sub>2</sub>(PO<sub>4</sub>)<sub>2</sub> 2-6H<sub>2</sub>O] precipitated at the bacterial cell surfaces (Morcillo et al. 2014). In a similar way, Shewanella sp. strain PV-4, from the microbial mat at a hydrothermal vent of Loihi Seamount in the Pacific Ocean, has been characterized with emphases on metal reduction including Fe (III), Co (III), Cr (VI), Mn (IV), and U (VI) as electron acceptors while using lactate, formate, pyruvate, or hydrogen as electron donor, showing the possibility to be exploited for bioreduction or immobilization of many toxic metals (Roh et al. 2006). More recently, the marine heterotrophic *Pseudoalteromonas* sp. MT33b with strong resistance to Cd, showed as major strategy to eliminate Cd stress the formation of insoluble CdS precipitates and massive biofilm. These characteristics open to possible utilization of bacterium from extreme environments for bioremediation purposes (Ma and Sun 2021).

Bioremediation realized by the marine bacteria Pseudomonas aeruginosa CH07 (NRRL B-30604) through the biotransformation process includes enzymatic reduction of toxic mercury (inorganic and organic) to volatile elemental mercury (Hg) in a two-step reaction. The bacterial resistance to Hg has shown activity against other highly toxic heavy metals (De et al. 2006). In this context, most of the Hg-resistant bacteria have shown the mer operon mechanisms located either on transposons, plasmids, or bacterial chromosomes (Dash et al. 2014). Bacteria hosting the genes merT and merP, indeed, encode for two transporters, which are responsible for the transport of Hg (II) into the cytoplasm and the subsequent action of mercuric ion reductase encoded by *merA* reduces Hg (II) to less toxic  $Hg^0$ , which diffuses out of the cell through cell membrane (Dash and Das 2012). It was observed that Hg-resistant marine bacteria shown potential for Cd and Pb detoxification, these bacteria were identified as Alcaligenes faecalis, Bacillus pumilus, Bacillus sp., Pseudomonas aeruginosa, and Brevibacterium iodinum. They were not able to produce any byproducts, and highly efficient even at low metal concentrations (De et al. 2008).

Common inhabitant of the marine environment, *Vibrio harveyi* was reported to possess the potential for bioaccumulation of Cd (Abd-Elnaby et al. 2011), while a consortium of marine bacteria was able to remove Hg by the same mode of action (Von Canstein et al. 2002). More recently, this strategy was detected in culturable marine deep sediment bacteria part of the phyla Firmicutes and Actinobacteria bioaccumulating heavy metals within the cells and/or in EPS (Jroundi et al. 2020). Marine isolates of *Streptomycetes*, phylogenetically inserted into the actinobacteria phylum, show unique growth characteristics including the ability to form spores and mycelia, and relatively rapid colonization of substrates, acting as suitable agents for bioremediation of metals and organic compounds in polluted soil and water (Timková et al. 2018).

Another important process included as bioremediation process of heavy metals carried out by marine bacteria was the absorption, in this case the bacteria together with other marine microbes secrete extracellular polymeric substances (ESP) to facilitate the attachment on surfaces and this led to the formation of structured biofilm communities (Decho and Gutierrez 2017). Metal cations form complexes with EPS resulting in metal immobilization generally promoted by electrostatic interactions between metal and negatively charged components of EPS biopolymers. However, the enzymatic activities in EPS also can assist detoxification of heavy metals (Pal and Paul 2008). The marine environment includes multi-cellular organisms as the sponges able to host huge microbial communities including bacteria, representing an important source of new bacteria with bioremediation capabilities. The sponge-associated Antarctic bacteria Winogradskyella sp. strains CAL384 and CAL396, Colwellia sp. strain GW185, and Shewanella sp. strain CAL606 were studied for their production of EPS with possible application on heavy metals absorption (Caruso et al. 2018a; Giudice et al. 2020). The marine bacteria Enterobacter cloacae showed EPS chelating properties with respect to Cd (65%) followed by Cu (20%) and Co (8%) at high heavy metal concentration (Iyer et al. 2005). Interestingly the EPS production by the seawater *Pseudoalteromonas*  sp. MER144 under optimal conditions at different concentrations of Hg and Cd revealed a modulate production of EPS when the heavy metal concentrations increased (Caruso et al. 2018b). Combination of strategies for removal of heavy metals has been shown by marine bacteria as *Rhodobium marinum* and *Rhodobacter sphaeroides* specifically for those involving either bioabsorption or biotransformation (Panwichian et al. 2011). Also, the marine *Bacillus thuringiensis* PW-05 demonstrated its capacity to resist at 50 ppm of Hg as HgCl<sub>2</sub> as well as higher concentrations of CdCl<sub>2</sub>, ZnSO<sub>4</sub>, PbNO<sub>3</sub>, and Na<sub>2</sub>HAsO<sub>4</sub>. Atomic absorption spectroscopy revealed that the isolate can volatilize >90% of inorganic Hg. Moreover, it showed biofilm formation in the presence of 50 ppm HgCl<sub>2</sub> and also the possibility to produce EPS under same conditions (Dash et al. 2014).

Another bacterial strategy for an eco-friendly, time saving, inexpensive with easily scaled up for large-scale synthesis to employ in heavy metal bioremediation is the synthesis of metallic nanoparticles either intracellularly or extracellularly (Manivasagan et al. 2016). Marine microorganisms can easily adapt themselves to extreme environmental conditions, it is very essential to explore marine bioresource for the green synthesis of different types of metallic nanoparticles. A strain of *Stenotrophomonas*, isolated from the Mandapam coast, Bay of Bengal in India, was employed for the biosynthesis of Au and Ag nanoparticles by extracellular secretion (Malhotra et al. 2013). In this context, the bacterium *Saccharophagus degradans* was investigated for the synthesis of MnO nanoparticles (Salunke et al. 2015). Microbial extracellular polymeric substances EPS act as capping and stabilizing agents for the biosynthesis of CdS nanoparticles by the marine bacterium *Pseudomonas aeruginosa* JP-11 and its comparison with chemical method were more efficient showing a higher Cd removal efficiency (Raj et al. 2016).

### 8.6 Engineering Bacteria for Bioremediation

Studies carried out to understand the interactions between xenobiotics and microorganisms and on the fate, survival, and activities of microorganisms in the environment have to intersect with the investigation on biochemical and genetic engineering aspects (Pieper and Reineke 2000). Natural microbes have already shown considerable ability to remove many environmental pollutants with no external intervention. The onset of genetic engineering in the 1980s allowed the possibility of rational design of bacteria to catabolize specific compounds with reduced or no possibilities to be removed (Dvořák et al. 2017). Once identified the genes and their genome location in bacteria that promote bioremediation, the objective becomes modify and incorporate them into a suitable host to be used as a bioremediation agent, in this context at the beginning usually was the *E. coli* (Zylstra et al. 1989). The indispensable incorporation of a gene marker (to discriminate the genetic modification) conferring antibiotic resistance was the simplest way of screening in producing genetically modified bacteria, representing an important concern (Dale et al. 2002). However, during these decades, since the discovery of recombinant DNA
technology many challenges have been encountered in constructing genetically engineered bacteria intended for environmental release and most of them have been resolved.

In general, the literature still counts very few cases where the use of genetically engineered bacteria has been confirmed to be more efficient than natural microorganisms in removal of recalcitrant compounds under natural (in situ) conditions (De Lorenzo 2009). Although the important steps achieved on the field of genetically modified bacteria, deeper studies should be realized to face ethical responsibilities before using such novel strategies for bioremediation (Perpetuo et al. 2011). Among the main concerns necessary to be considered, the ecological impacts of including genetically modified organisms into the environment become fundamental (Rajakaruna and Robinson 2016). Into the field of the engineered bacteria, for example, become necessary the detection and enumeration of these modified microorganisms in complex samples (Widada et al. 2002). Thus, a deeper approach may permit to overcome both environmental concerns and regulatory constraints that have limited the in situ application of genetically engineered microorganisms. Considering that the main objective of this limitation is to avoid the risk of a possible uncontrolled survival/dispersal of engineered microorganisms or recombinant plasmids into the environment, some strategies have been suggested, including the induction of suicide microbes once their mission is completed (Paul et al. 2005; Ezezika and Singer 2010). Furthermore, the integration of microbiological, biological, and ecological acquaintance accompanied by field engineering designs was considered features for effective application of in situ bioremediation of polluted sites by recombinant bacteria (Liu et al. 2019).

Systems biology studies, based on top-down and bottom-up large-scale "omics" tools and mathematical modeling methods, have become pivotal on the engineered bacteria build (Park et al. 2017). The enormous amount of genomics, transcriptomics, proteomics, and metabolomics information, coming from systems biology studies, allow to integrate the data for a more holistic understanding. In this context, novel data mining and analytics approaches, including artificial intelligence can provide breakthroughs where traditional low-throughput experiment-alone methods cannot easily achieve (Helmy et al. 2020). Both integration and mapping of systems biology and metabolic engineering tools and techniques can help to achieve the environmental bioremediation objective, concretely these may permit to pass from a theoretical stage to a practical stage (Dangi et al. 2019).

Various laboratory studies have evaluated engineered bacteria for possible use in bioremediation of contaminated sites. Bacteria genetically modified have been evaluated, for example, to neutralize heavy metals and transform them into less toxic forms (Paliwal et al. 2012; Pratush et al. 2018), to improve the degradation of plastic debris (Kumari and Chaudhary 2020), for petroleum hydrocarbons degradation (Naeem and Qazi 2020), and also acting on the degradation of PAHs (Haritash 2020). Once the metabolic pathways are properly understood, the genetic manipulation could permit the development of both transgenic plants and microbes that may work twice efficiently in direction of the bioremediation of contaminated sites of heavy metal and organic pollutants (Ojuederie and Babalola 2017). Some examples

report that engineered bacteria may be a viable technology for Hg bioremediation from liquid matrices, employing transgenic bacteria expressing metallothionein (mt-1) and polyphosphate kinase (ppk) genes. It was possible to achieve high Hg resistance and accumulation while reduced Hg volatilization was observed. Specifically, the reduction of the Hg volatility represents an important advantage comparing the reduction at the volatile elementary form (Hg<sup>0</sup>) by simple natural bacteria expressing *mer* genes (Ruiz et al. 2011).

Recent advances to face regulatory hurdles and environmental concerns come from engineered bacteria applied to clean up oil spills, showing high efficiency for possible in situ application. The study included the genetic vector detection among environmental bacteria of the contaminated site treated with genetically modified bacteria. A crude oil soil remediation employing *E. coli* transporting three enzymes (almA, xylE, p450cam) resulted in the degradation of 60–99% of target hydrocarbon substrates, from the vector detection it was possible to notice that *E. coli* cells died after five days, while a variety of bacteria received and carried the vector for over 60 days after inoculation. However, it disappeared when the carbon source was removed providing minimal ecosystem disturbance (French et al. 2020).

Some studies have been conducted with bacteria showing the ability to work under ionizing radiation, including marine strains of *Deinococcus* spp., considered for its extremely radio-resistant capacity. This bacterial group has shown DNA repairing proficiency permitting their use in a number of strategies for bioremediation of radioactive waste, collaborating with those capacities such as desiccation, temperature, and metal tolerance (Suresh et al. 2004; Ferreira et al. 1997). For these peculiar characteristics, it has become an organism of choice for bacterial engineering (Brim et al. 2000, 2003). The lyophilized recombinant *Deinococcus radiodurans* genetically modified for PhoN expression showed PhoN activity and uranium precipitation ability as well as other metals like cadmium. PhoN promote the liberation of inorganic phosphate from a suitable substrate molecule like  $\beta$ -glycerophosphate, causing the precipitation of metals as cell-bound metal phosphates and facilitates their easy removal from aqueous solution (Misra et al. 2012).

An important chemical groundwater pollutant is 1,2,3-trichloropropane (TCP) used as degreasing agent with high water solubility that causes large spreading of this recalcitrant component in the environment. Currently, no natural microorganisms are known to be able to mineralize TCP. This necessitates isolation and remediation measures, which may be based on (accelerated) in situ treatment or pump-and-treat methodologies. TCP was chosen as a target for constructing bacteria that use it as a growth substrate since some structurally similar compounds are biodegradable. This was indeed achieved by a combination of protein- and metabolic engineering. In this case, the use of genetically modified bacteria still awaits full-scale application, while successful full-scale application has been achieved for the similar 1,2-dichloroethane by isolating bacteria degrading this compounds and bioremediation activities were carried out with ex situ procedures (Janssen and Stucki 2020).

## 8.7 Concluding Remarks

Environmental pollution has been widely spread around the world, and the pollution is cumulative, once naturally, it is impossible the auto depuration of all contaminants released in the environment and accumulation in some parts such as in marine water and sediments. Marine pollution is affecting all kinds of systems such as bacterial communities, corals, fish, and other living organisms, even the humans once the ocean has been feeding the human beings since the humanity began. In this scenario, bioremediation became for helping and has efficient strategies for each kind of pollution. Refinery wastes are common contaminants widespread released in the environment and studied in bioremediation. Marine bacteria have been used as efficient biological agents for bioremediation of contaminants as plastics, petroleum, hydrocarbons, and heavy metals. Biodegradation combined with other strategies such as bioaugmentation, biosurfactant production, and biotransformation is very powerful for bioremediation of marine environments with very good clean-up results. Thus, additional studies and applications need to be performed and are necessary to improve bioremediation technologies using marine bacteria.

Acknowledgments The authors thank the financial support of CNPq, FAPERGS, and CAPES.

### References

- Abatenh E, Gizaw B, Tsegaye Z, Wassie M (2017) The role of microorganisms in bioremediation: a review. Open J Environ Biol 2:38–46
- Abd-Elnaby H, Abou-Elela GM, El-Sersy NA (2011) Cadmium resisting bacteria in Alexandria Eastern Harbor (Egypt) and optimization of cadmium bioaccumulation by *Vibrio harveyi*. African J Biotechnol 10:3412–3423
- Adam IKU, Duarte AM, Pathmanathan J, Miltner A, Brüls T, Kästner M (2017) Microbial communities in pyrene amended soil compost mixture and fertilized soil. AMB Expr 7:7
- Adams GO, Fufeyin PT, Okoro SE, Ehinomen I (2015) Bioremediation, biostimulation and bioaugmention: a review. Int J Environ Bioremed Biodegrad 3:28–39
- Agarry S, Latinwo GK (2015) Biodegradation of diesel oil in soil and its enhancement by application of bioventing and amendment with brewery waste effluents as biostimulatingbioaugmentation agents. J Ecol Eng 16:82–91
- Ahemad M (2019) Remediation of metalliferous soils through the heavy metal resistant plant growth promoting bacteria: paradigms and prospects. Arab J Chem 12:1365–1377
- Akinde SB, Iwuozor CC, Obire O (2012) Alkane degradative potentials of bacteria isolated from the deep Atlantic Ocean of the Gulf of Guinea. J Bioremed Biodegr 3:2–6
- Akram MS, Rashid N, Basheer S (2021) Physiological and molecular basis of plants tolerance to linear halogenated hydrocarbons. In: Hasanuzzaman M, Prasad NMV (eds) Handbook of bioremediation. Academic Press, New York, pp 591–602
- Ali A, Guo D, Mahar A, Wang P, Shen F, Li R, Zhang Z (2017) Mycoremediation of potentially toxic trace elements a biological tool for soil clean up: a review. Pedosphere 27:205–222
- Ali H, Khan E, Ilahi I (2019) Environmental chemistry and ecotoxicology of hazardous heavy metals: environmental persistence, toxicity, and bioaccumulation. J Chem 2019:6730305
- Amidei R (1997) Marine bacteria: a better cleaner-upper? Calif Agric 51:47-48

- Anderson C, Juday G (2016) Mycoremediation of petroleum: a literature review. J Environ Sci Eng A5:397–405
- Andreolli M, Lampis S, Brignoli P, Vallini G (2015) Bioaugmentation and biostimulation as strategies for the bioremediation of a burned woodland soil contaminated by toxic hydrocarbons: a comparative study. J Environ Manag 153C:121–130
- Arslan M, Imran A, Khan QM, Afzal M (2017) Plant–bacteria partnerships for the remediation of persistent organic pollutants. Environ Sci Pollut Res 24:4322–4336
- Atlas RM, Hazen TC (2011) Oil biodegradation and bioremediation: a tale of the two worst spills in US history. Environ Sci Technol 45:6709–6715
- Atlas RM, Philp J (2005) Bioremediation. Applied microbial solutions for real-world environmental cleanup. ASM Press, Washington, p 370
- Ayangbenro AS, Olanrewaju OS, Babalola OO (2018) Sulfate-reducing bacteria as an effective tool for sustainable acid mine bioremediation. Front Microbiol 9:1986
- Azubuike CC, Chikere CB, Okpokwasili GC (2016) Bioremediation techniques -classification based on site of application: principles, advantages, limitations and prospects. World J Microbiol Biotechnol 32:180
- Azubuike CC, Chikere CB, Okpokwasili GC (2020) Bioremediation: an eco-friendly sustainable technology for environmental management. In: Saxena G, Bharagava R (eds) Bioremediation of industrial waste for environmental safety. Springer, Singapore, pp 19–39
- Balseiro-Romero M, Gkorezis P, Kidd PS, Van Hamme J, Weyens N, Monterroso C, Vangronsveld J (2017) Characterization and degradation potential of diesel-degrading bacterial strains for application in bioremediation. Int J Phytoremediation 19:955–963
- Bao MT, Wang LN, Sun PY, Cao LX, Zou J, Li YM (2012) Biodegradation of crude oil using an efficient microbial consortium in a simulated marine environment. Mar Pollut Bull 64:1177– 1185
- Benyahia F, Embaby AS (2016) Bioremediation of crude oil contaminated desert soil: effect of biostimulation, bioaugmentation and bioavailability in biopile treatment systems. Int J Environ Res Public Health 13:219
- Beygmoradi A, Homaei A (2017) Marine microbes as a valuable resource for brand new industrial biocatalysts. Biocatal Agric Biotechnol 11:131–152
- Bharagava RN, Purchase D, Saxena G, Mulla I (2019) Applications of metagenomics in microbial bioremediation of pollutants: from genomics to environmental cleanup. In: Das S, Dash HR (eds) Microbial diversity in the genomic era. Elsevier, New York, pp 459–477
- Blindauer CA (2011) Bacterial metallothioneins: past, present, and questions for the future. J Biol Inorg Chem 16:1011–1024
- Boopathy R (2000) Factors limiting bioremediation technologies. Bioresour Technol 74:63-67
- Bougarne L, Abbou MB, El Haji M, Bouka H (2019) Consequences of surface water eutrophication: remedy and environmental interest. Mater Today: Proceed 13:654–662
- Brim H, McFarlan SC, Fredrickson JK, Minton KW, Zhai M, Wackett LP, Daly MJ (2000) Engineering *Deinococcus radiodurans* for metal remediation in radioactive mixed waste environments. Nat Biotechnol 18:85–90
- Brim H, Venkateswaran A, Kostandarithes HM, Fredrickson JK, Daly MJ (2003) Engineering *Deinococcus geothermalis* for bioremediation of high-temperature radioactive waste environments. Appl Environ Microbiol 69:4575–4582
- Cachada A, Rocha-Santos T, Duarte AC (2018) Soil and pollution: an introduction to the main issues. In: Duarte AC, Cachada A, Rocha-Santos T (eds) Soil Pollution. Academic Press, New York, pp 1–28
- Cai D, Yang X, Wang S, Chao Y, Morel JL, Qiu R (2017) Effects of dissolved organic matter derived from forest leaf litter on biodegradation of phenanthrene in aqueous phase. J Hazard Mater 324:516–525
- Carson HS, Nerheim MS, Carroll KA, Eriksen M (2013) The plastic-associated microorganisms of the North Pacific gyre. Mar Pollut Bull 75:126–132

- Caruso C, Rizzo C, Mangano S, Poli A, Di Donato P, Finore I, Nucolaus B, Di Marco B, Michaud L, Giudice AL (2018a) Production and biotechnological potential of extracellular polymeric substances from sponge-associated Antarctic bacteria. Appl Environ Microbiol 84: e01624–e01617
- Caruso C, Rizzo C, Mangano S, Poli A, Di Donato P, Nicolaus B, Di Marco B, Michaud L, Giudice AL (2018b) Extracellular polymeric substances with metal adsorption capacity produced by *Pseudoalteromonas* sp. MER144 from Antarctic seawater. Environ Sci Pollut Res 25:4667– 4677
- Catania V, Santisi S, Signa G, Vizzini S, Mazzola A, Cappello S, Yakimov MM, Quatrini P (2015) Intrinsic bioremediation potential of a chronically polluted marine coastal area. Mar Pollut Bull 99:138–149
- Chung WK, King GM (2001) Isolation, characterization, and polyaromatic hydrocarbon degradation potential of aerobic bacteria from marine macrofaunal burrow sediments and description of *Lutibacterium anuloederans* gen. Nov., sp. nov., and *Cycloclasticus spirillensus* sp. nov. Appl Environ Microbiol 67:5585–5592
- Colwell RR, Walker JD, Cooney JJ (1977) Ecological aspects of microbial degradation of petroleum in the marine environment. Crit Rev Microbiol 5:423–445
- Cornu JY, Huguenot D, Jézéquel K, Lollier M, Lebeau T (2017) Bioremediation of coppercontaminated soils by bacteria. World J Microbiol Biotechnol 33:26
- CRC (2018) Contamination assessment and remediation of the environment national remediation framework technology guide: bioremediation Version 0.1. https://www.crccare.com/files/dmfile/CTechguide\_Bioremediation\_Rev0.pdf
- Cunningham CJ, Philip JC (2000) Comparison of bioaugmentation and biostimulation in ex situ treatment of diesel contaminated soil. Land Contam Reclamat 8:261–269
- Dale PJ, Clarke B, Fontes EM (2002) Potential for the environmental impact of transgenic crops. Nat Biotechnol 20:567–574
- Dang TCH, Nguyen DT, Thai H, Nguyen TC, Tran TTH, Le VH, Nguyen VH, Tran XB, Pham TPT, Nguyen TG (2018) Plastic degradation by thermophilic *bacillus* sp. BCBT21 isolated from composting agricultural residual in Vietnam. Adv Nat Sci Nanosci Nanotechnol 9:015014
- Dangi AK, Sharma B, Hill RT, Shukla P (2019) Bioremediation through microbes: systems biology and metabolic engineering approach. Crit Rev Biotechnol 39:79–98
- Das S, Dash HR (2014) Microbial bioremediation: a potential tool for restoration of contaminated areas. In: Das S (ed) Microbial biodegradation and bioremediation. Elsevier, New York, pp 1–21
- Dash HR, Das S (2012) Bioremediation of mercury and importance of bacterial *mer* genes. Int Biodeter Biodegrad 75:207–213
- Dash HR, Mangwani N, Chakraborty J, Kumari S, Das S (2013) Marine bacteria: potential candidates for enhanced bioremediation. Appl Microbiol Biotechnol 97:561–571
- Dash HR, Mangwani N, Das S (2014) Characterization and potential application in mercury bioremediation of highly mercury-resistant marine bacterium *bacillus thuringiensis* PW-05. Environ Sci Pollut Res 21:2642–2653
- Datta S, Rajnish KN, Samuel MS, Pugazlendhi A, Selvarajan E (2020) Metagenomic applications in microbial diversity, bioremediation, pollution monitoring, enzyme and drug discovery. A review Environ Chem Lett 18:1229–1241
- De Almeida DG, Soares Da Silva RDCF, Luna JM, Rufino RD, Santos VA, Banat IM, Sarubbo LA (2016) Biosurfactants: promising molecules for petroleum biotechnology advances. Front Microbiol 7:1718
- De Lorenzo V (2009) Recombinant bacteria for environmental release: what went wrong and what we have learnt from it. Clin Microbiol Infect 15:63–65
- De J, Ramaiah N, Vardanyan L (2008) Detoxification of toxic heavy metals by marine bacteria highly resistant to mercury. Mar Biotechnol 10:471–477
- De J, Sarkar A, Ramaiah N (2006) Bioremediation of toxic substances by mercury resistant marine bacteria. Ecotoxicology 15:385–389

- Decho AW, Gutierrez T (2017) Microbial extracellular polymeric substances (EPSs) in ocean systems. Front Microbiol 8:922
- Deepika KV, Sheela GM, Bramhachari PV (2021) Marine microbial biosurfactants: ecological and environmental applications. In: Gothandam KM, Ranjan S, Dasgupta N, Lichtfouse E (eds) Environmental biotechnology, vol 3. Springer, Cham, pp 221–232
- Devi RS, Ramya R, Kannan K, Antony AR, Kannan VR (2019) Investigation of biodegradation potentials of high density polyethylene degrading marine bacteria isolated from the coastal regions of Tamil Nadu, India. Mar Pollut Bull 138:549–560
- Dias RL, Ruberto L, Calabró A, Balbo AL, Del Panno MT, MacCormack WP (2015) Hydrocarbon removal and bacterial community structure in on-site biostimulated biopile systems designed for bioremediation of diesel-contaminated Antarctic soil. Polar Biol 38:677–687
- Divya B, Deepak Kumar M (2011) Plant–microbe interaction with enhanced bioremediation. Res J Biotechnol 6:4
- Dixit R, Malaviya D, Pandiyan K, Singh UB, Sahu A, Shukla R, Singh BP, Rai JP, Sharma PK, Lade H, Paul D (2015) Bioremediation of heavy metals from soil and aquatic environment: an overview of principles and criteria of fundamental processes. Sustainability 7:2189–2212
- Dua M, Singh A, Sethunathan N, Johri A (2002) Biotechnology and bioremediation: successes and limitations. Appl Microbiol Biotechnol 59:143–152
- Duran R, Cravo-Laureau C (2016) Role of environmental factors and microorganisms in determining the fate of polycyclic aromatic hydrocarbons in the marine environment. FEMS Microbiol Rev 40:814–830
- Durval IJB, Resende AHM, Figueiredo MA, Luna JM, Rufino RD, Sarubbo LA (2019) Studies on biosurfactants produced using *Bacillus cereus* isolated from seawater with biotechnological potential for marine oil-spill bioremediation. J Surfactant Deterg 22:349–363
- Dvořák P, Nikel PI, Damborský J, De Lorenzo V (2017) Bioremediation 3.0: engineering pollutantremoving bacteria in the times of systemic biology. Biotechnol Adv 35:845–866
- Enerijiofi KE (2021) Bioremediation of environmental contaminants: a sustainable alternative to environmental management. In: Saxena G, Kumar V, Shah MP (eds) Bioremediation for environmental sustainability. Elsevier, New York, pp 461–480
- EPA (2001) Remediation case studies. Federal Remediation Technology Roundtable. Report No. 542-F-01-032. US Environmental Protection Agency, Washington
- EPA (2002) Handbook on in situ treatment of hazardous waste contaminated soils. US Environmental Protection Agency, Washington
- Ezezika OC, Singer PA (2010) Genetically engineered oil-eating microbes for bioremediation: prospects and regulatory challenges. Technol Soc 32:331–335
- Fathepure BZ (2014) Recent studies in microbial degradation of petroleum hydrocarbons in hypersaline environments. Front Microbiol 5:173
- Ferreira AC, Nobre MF, Rainey FA, Silva MT, Wait R, Burghardt J, Chung AP, Da Costa MS (1997) *Deinococcus geothermalis* sp. nov. and *Deinococcus murrayi* sp. nov., two extremely radiation-resistant and slightly thermophilic species from hot springs. Int J Syst Evol Microbiol 47:939–947
- French KE, Zhou Z, Terry N (2020) Horizontal 'gene drives' harness indigenous bacteria for bioremediation. Sci Rep 10:15091
- Gadd GM (2001) Fungi in Bioremediation. Cambridge University Press, Cambridge, p 481
- Gao J, Ming J, Xu M, Fu X, Duan LF, Xu CC, Gao Y, Xue J-L, Xiao XF (2021) Isolation and characterization of a high-efficiency marine diesel oil-degrading bacterium. Pet Sci 18:641–653
- Gärdes A, Iversen MH, Grossart HP, Passow U, Ullrich MS (2011) Diatom-associated bacteria are required for aggregation of *Thalassiosira weissflogii*. ISME J 5:436–445
- Gentry T, Rensing C, Pepper IAN (2004) New approaches for bioaugmentation as a remediation technology. Crit Rev Environ Sci Technol 34:447–494
- Gewert B, Plassmann MM, MacLeod M (2015) Pathways for degradation of plastic polymers floating in the marine environment. Environ Sci: Proc Impacts 17:1513–1521

- Ghangrekar MM, Sathe SM, Chakraborty I (2020) In Situ bioremediation techniques for the removal of emerging contaminants and heavy metals using hybrid microbial electrochemical technologies. In: Shah MP, Rodriguez-Couto S, Sengör S (eds) Emerging technologies in environmental bioremediation. Elsevier, New York, pp 233–256
- Ghosh M, Singh S (2005) A review on phytoremediation of heavy metals and utilization of its byproducts. Appl Ecol Environ Res 3:1–18
- Girma G (2015) Microbial bioremediation of some heavy metals in soils: an updated review. Egyptian Acad J Biol Sci 7:29–45
- Giudice AL, Poli A, Finore I, Rizzo C (2020) Peculiarities of extracellular polymeric substances produced by Antarctic bacteria and their possible applications. Appl Microbiol Biotechnol 104: 2923–2934
- Gkorezis P, Daghio M, Franzetti A, Van Hamme JD, Sillen W, Vangronsveld J (2016) The interaction between plants and bacteria in the remediation of petroleum hydrocarbons: an environmental perspective. Front Microbiol 7:1836
- Goldberg D (1995) A review of the biodegradability and utility of poly (caprolactone). J Environ Polym Degrad 3:61–67
- Goris J, De Vos P, Coenye T, Hoste B, Janssens D, Brim H, Diels L, Mergeay M, Kersters K, Vandamme P (2001) Classification of metal-resistant bacteria from industrial biotopes as *Ralstonia campinensis* sp. nov., *Ralstonia metallidurans* sp. nov. and *Ralstonia basilensis* Steinle et al. 1998 emend. Int J Syst Evol Microbiol 51:1773–1782
- Gu W, Li X, Li Q, Hou Y, Zheng M, Li Y (2021) Combined remediation of polychlorinated naphthalene-contaminated soil under multiple scenarios: an integrated method of genetic engineering and environmental remediation technology. J Hazard Mater 405:124139
- Haritash AK (2020) A comprehensive review of metabolic and genomic aspects of PAH-degradation. Arch Microbiol 202:2033–2058
- Harms H, Schlosser D, Wick LY (2011) Untapped potential: exploiting fungi in bioremediation of hazardous chemicals. Nat Rev Microbiol 9:177–192
- Hazen TC (2010) Biostimulation. In: Timmis KN, McGenity T, van der Meer JR, de Lorenzo V (eds) Handbook of hydrocarbon and lipid microbiology. Springer, Berlin, pp 4517–4530
- Hedlund BP, Geiselbrecht AD, Bair TJ, Staley JT (1999) Polycyclic aromatic hydrocarbon degradation by a new marine bacterium, *Neptunomonas naphthovorans* gen. Nov., sp. nov. Appl Environ Microbiol 65:251–259
- Helmy M, Smith D, Selvarajoo K (2020) Systems biology approaches integrated with artificial intelligence for optimized food-focused metabolic engineering. Metab Eng Commun 11:e00149
- Höhener P, Ponsin V (2014) In situ vadose zone bioremediation. Curr Op Biotechnol 27:1-7
- Hong X, Zhao Y, Zhuang R, Liu J, Guo G, Chen J, Yao Y (2020) Bioremediation of tetracycline antibiotics-contaminated soil by bioaugmentation. RSC Adv 10:33086–33102
- Huang Y, Xiao L, Li F, Xiao M, Lin D, Long X, Wu Z (2018) Microbial degradation of pesticide residues and an emphasis on the degradation of cypermethrin and 3-phenoxy benzoic acid: a review. Molecules 23:2313
- Hutchins SR, Davidson MS, Brierley JA, Brierley CL (1986) Microorganisms in reclamation of metals. Annu Rev Microbiol 40:311–336
- Igiri BE, Okoduwa SI, Idoko GO, Akabuogu EP, Adeyi AO, Ejiogu IK (2018) Toxicity and bioremediation of heavy metals contaminated ecosystem from tannery wastewater: a review. J Toxicol 2018:2568038
- Ilyas S, Chi RA, Lee JC (2013) Fungal bioleaching of metals from mine tailing. Min Process Extr Met Rev 34:185–194
- Imron MF, Kurniawan SB, Ismail NI, Abdullah SRS (2020) Future challenges in diesel biodegradation by bacteria isolates: a review. J Clean Prod 251:119716
- Işıldar A, Van de Vossenberg J, Rene ER, Van Hullebusch ED, Lens PNL (2016) Two-step bioleaching of copper and gold from discarded printed circuit boards (PCB). Waste Manag 57:149–157

- Iyer A, Mody K, Jha B (2005) Biosorption of heavy metals by a marine bacterium. Mar Pollut Bull 50:340–343
- Jacques P (2011) Surfactin and other lipopeptides from *bacillus* spp. In: Soberón-Chávez G (ed) Biosurfactants. Springer, Heidelberg, pp 57–91
- Jagannath A, Vidya SK, Saidutta MB (2017) Bioleaching of copper from electronic waste using Acinetobacter sp. Cr B2 in a pulsed plate column operated in batch and sequential batch mode. J Environ Chem Eng 5:1599–1607
- Jalali K, Baldwin SA (2000) The role of sulphate reducing bacteria in copper removal from aqueous sulphate solutions. Water Res 34:797–806
- Jambeck JR, Geyer R, Wilcox C, Siegler TR, Perryman M, Andrady A, Narayan R, Law KL (2015) Plastic waste inputs from land into the ocean. Science 347:768–771
- Jang KY, Cho SM, Seok SJ, Kong WS, Kim GH, Sung JM (2009) Screening of biodegradable function of indigenous lingo-degrading mushroom using dyes. Mycobiology 4:53–61
- Janssen DB, Stucki G (2020) Perspectives of genetically engineered microbes for groundwater bioremediation. Environ Sci Process Impacts 22:487–499
- Jeevanantham S, Saravanan A, Hemavathy RV, Kumar PS, Yaashikaa PR, Yuvaraj D (2019) Removal of toxic pollutants from water environment by phytoremediation: a survey on application and future prospects. Environ Technol Innov 13:264–276
- Jørgensen BB (1982) Mineralization of organic matter in the sea bed the role of sulphate reduction. Nature 296:643–645
- Jroundi F, Martinez-Ruiz F, Merroun ML, Gonzalez-Muñoz MT (2020) Exploring bacterial community composition in Mediterranean deep-sea sediments and their role in heavy metal accumulation. Sci Total Environ 712:135660
- Kapahi M, Sachdeva S (2019) Bioremediation options for heavy metal pollution. J Health Pollut 9: 191203
- Kaushal J, Khatri M, Arya SK (2021) Recent insights into enzymatic degradation of plastics prevalent in the environment: a mini review. Clean Eng Technol 2:1000083
- Kim J, Lee AH, Chang W (2018) Enhanced bioremediation of nutrient-amended petroleum hydrocarbon contaminated soil over a cold climate winter: the rate and extent of hydrocarbon biodegradation and microbial response in a pilot-scale biopile subjected to natural seasonal freeze-thaw temperatures. Sci Total Environ 612:903–913
- Kiran MG, Pakshirajan K, Das G (2017) Heavy metal removal from multicomponent system by sulfate reducing bacteria: mechanism and cell surface characterization. J Hazard Mater 324:62– 70
- Kocur CM, Lomheim L, Molenda O, Weber KP, Austrins LM, Sleep BE, Boparai HK, Edwards EA, O'Carroll DM (2016) Long-term field study of microbial community and dechlorinating activity following carboxymethyl cellulose-stabilized nanoscale zero-valent iron injection. Environ Sci Technol 50:7658–7670
- Koolivand A, Saeedi R, Coulon F, Kumar V, Villaseñor J, Asghari F, Hesampoor F (2020) Bioremediation of petroleum hydrocarbons by vermicomposting process bioaugmentated with indigenous bacterial consortium isolated from petroleum oily sludge. Ecotoxicol Environ Saf 198:110645
- Koshlaf E, Ball AS (2017) Soil bioremediation approaches for petroleum hydrocarbon polluted environments. AIMS Microbiol 3:25
- Krishna IM, Manickam V, Shah A, Davergave N (2017) Environmental management: science and engineering for industry. Butterworth-Heinemann, Oxford, p 664
- Kumar PS, Gunasundari E (2018) Bioremediation of heavy metals. In: Varjani SJ, Agarwal AK, Gnansounou E, Gurunathan B (eds) Bioremediation: applications for environmental protection and management. Springer, Singapore, pp 165–196
- Kumar V, Shahi SK, Singh S (2018a) Bioremediation: an eco-sustainable approach for restoration of contaminated sites. In: Singh J, Sharma D, Kumar G, Sharma N (eds) Microbial bioprospecting for sustainable development. Springer, Singapore, pp 115–136

- Kumar NM, Muthukumaran C, Sharmila G, Gurunathan B (2018b) Genetically modified organisms and its impact on the enhancement of bioremediation. In: Varjani SJ, Agarwal AK, Gnansounou E, Gurunathan B (eds) Bioremediation: applications for environmental protection and management. Springer, Singapore, pp 56–76
- Kumar A, Tripti PMNV, Maiti SK, Favas PJC (2018c) Mycoremediation of mine site rehabilitation. In: Prasad MNV, Favas PJC, Maiti SK (eds) Bio-Geotechnologies of mine site rehabilitation. Elsevier, New York, pp 233–260
- Kumari A, Chaudhary DR (2020) Engineered microbes and evolving plastic bioremediation technology. In: Pandey VC, Singh V (eds) Bioremediation of pollutants. Elsevier, New York, pp 417–443
- Kushwaha A, Hans N, Kumar S, Rani R (2018) A critical review on speciation, mobilization and toxicity of lead in soil-microbe-plant system and bioremediation strategies. Ecotoxicol Environ Saf 147:1035–1045
- Lee DW, Lee H, Lee AH, Kwon BO, Khim JS, Yim UH, Kim BS, Kim JJ (2018) Microbial community composition and PAHs removal potential of indigenous bacteria in oil contaminated sediment of Taean coast, Korea. Environ Pollut 234:503–512
- Leni Z, Künzi L, Geiser M (2020) Air pollution causing oxidative stress. Curr Op Toxicol 20:1-8
- Leong YK, Chang JS (2020) Bioremediation of heavy metals using microalgae: recent advances and mechanisms. Bioresour Technol 303:122886
- Li F, Zhou T (2020) Effects of objective and subjective environmental pollution on Well-being in urban China: a structural equation model approach. Soc Sci Med 249:112859
- Li X, Lan SM, Zhu ZP, Zhang C, Zeng GM, Liu YG, Cao WC, Song B, Yang H, Wang SF, Wu SH (2018) The bioenergetics mechanisms and applications of sulfate-reducing bacteria in remediation of pollutants in drainage: a review. Ecotoxicol Environ Saf 158:162–170
- Li Z, Wei R, Gao M, Ren Y, Yu B, Nie K, Xu H, Liu L (2020) Biodegradation of low-density polyethylene by *Microbulbiferhydrolyticus* IRE-31. J Environ Manag 263:110402
- Lim MW, Lau EV, Poh PE (2016) A comprehensive guide of remediation technologies for oil contaminated soil–present works and future directions. Mar Pollut Bull 109:14–45
- Liu F, Zhang G, Liu S, Fu Z, Chen J, Ma C (2018) Bioremoval of arsenic and antimony from wastewater by a mixed culture of sulfate-reducing bacteria using lactate and ethanol as carbon sources. Int Biodeter Biodegrad 126:152–159
- Liu L, Bilal M, Duan X, Iqbal HM (2019) Mitigation of environmental pollution by genetically engineered bacteria–current challenges and future perspectives. Sci Total Environ 667:444–454
- Ma N, Sun C (2021) Cadmium sulfide nanoparticle biomineralization and biofilm formation mediate cadmium resistance of the deep-sea bacterium *Pseudoalteromonas* sp. MT33b. Environ Microbiol Rep 13:325–336
- Malhotra A, Dolma K, Kaur N, Rathore YS, Mayilraj S, Choudhury AR (2013) Biosynthesis of gold and silver nanoparticles using a novel marine strain of *Stenotrophomonas*. Bioresour Technol 142:727–731
- Malode SN, Nayana S, Shirbhate H (2013) A review on phytoremediation: a novel strategy for the removal of toxic metals and contaminants from the environment by using plants. Bionano Frontier 6:74–79
- Mangwani N, Shukla SK, Kumari S, Rao TS, Das S (2014) Characterization of *Stenotrophomonas* acidaminiphila NCW-702 biofilm for implication in the degradation of polycyclic aromatic hydrocarbons. J Appl Microbiol 117:1012–1024
- Manisalidis I, Stavropoulou E, Stavropoulos A, Bezirtzoglou E (2020) Environmental and health impacts of air pollution: a review. Front Public Health 8:14
- Manivasagan P, Nam SY, Oh J (2016) Marine microorganisms as potential biofactories for synthesis of metallic nanoparticles. Crit Rev Microbiol 42:1007–1019
- Minimol M, Vidya SK, Saidutta MB (2020) Process engineering aspects in bioleaching of metals from electronic waste. In: Jerold M, Arockiasamy S, Sivasubramanian V (eds) Bioprocess engineering for bioremediation, valorization and management techniques. Springer, Cham, pp 27–44

- Mishra A, Mishra SP, Arshi A, Agarwal A, Dwivedi SK (2020) Plant-microbe interactions for bioremediation and phytoremediation of environmental pollutants and agroecosystem development. In: Bharagava R, Saxena G (eds) Bioremediation of industrial waste for environmental safety: volume II, Biological agents and methods for industrial waste management. Springer, Singapore, pp 415–436
- Mishra A (2021) Phytoremediation of heavy metal-contaminated soils: recent advances, challenges, and future prospects. In: Saxena G, Kumar V, Shah MP (eds) Bioremediation for environmental sustainability toxicity, mechanisms of contaminants degradation, detoxification, and challenges. Elsevier, New York, pp 29–45
- Misra CS, Appukuttan D, Kantamreddi VSS, Rao AS, Apte SK (2012) Recombinant D. radiodurans cells for bioremediation of heavy metals from acidic/neutral aqueous wastes. Bioengineered 3:44–48
- Mohan SV, Sirisha K, Rao NC, Sarma PN, Reddy SJ (2004) Degradation of chlorpyrifos contaminated soil by bioslurry reactor operated in sequencing batch mode: bioprocess monitoring. J Hazard Mater 116:39–48
- Mohanrasu K, Premnath N, Prakash GS, Sudhakar M, Boobalan T, Arun A (2018) Exploring multi potential uses of marine bacteria; an integrated approach for PHB production, PAHs and polyethylene biodegradation. J Photochem Photobiol B Biol 185:55–65
- Mohanrasu K, Rao RGR, Raja R, Arun A (2020) Bioremediation process by marine microorganisms. In: Kim SK (ed) Encyclopedia of marine biotechnology, vol 4. Wiley, Hoboken, pp 2211–2228
- Mohapatra RK, Parhi PK, Patra JK, Panda CR, Thatoi HN (2017) Biodetoxification of toxic heavy metals by marine metal resistant bacteria–a novel approach for bioremediation of the polluted saline environment. In: Patra J, Vishnuprasad C, Das G (eds) Microbial biotechnology. Springer, Singapore, pp 343–376
- Morcillo F, González-Muñoz MT, Reitz T, Romero-González ME, Arias JM, Merroun ML (2014) Biosorption and biomineralization of U (VI) by the marine bacterium *Idiomarina loihiensis* MAH1: effect of background electrolyte and pH. PLoS One 9:e91305
- Morillo E, Madrid F, Lara-Moreno A, Villaverde J (2020) Soil bioremediation by cyclodextrins. A review Int J Pharm 591:119943
- Naeem U, Qazi MA (2020) Leading edges in bioremediation technologies for removal of petroleum hydrocarbons. Environ Sci Pollut Res 27:27370–27382
- Naik MM, Pandey A, Dubey SK (2012) Bioremediation of metals mediated by marine bacteria. In: Satyanarayana T, Johri B, Prakash A (eds) Microorganisms in environmental management. Springer, Dordrecht, pp 665–682
- Natarajan G, Ting YP (2013) Two-step bioleaching and spent medium leaching of gold from electronic scrap material using *Chromobacterium violaceum*. Adv Mater Res 825:270–273
- Nogales J, García JL, Díaz E (2017) Degradation of aromatic compounds in *pseudomonas*: a systems biology view. In: Rojo F (ed) Aerobic utilization of hydrocarbons, oils and lipids. Handbook of Hydrocarbon and Lipid Microbiology, Springer, Cham, pp 1–49
- Noman E, Al-Gheethi A, Mohamed RMSR, Talip BA (2019) Myco-remediation of xenobiotic organic compounds for a sustainable environment: a critical review. Top Curr Chem 377:17–41
- Nwankwegu AS, Onwosi CO (2017) Bioremediation of gasoline contaminated agricultural soil by bioaugmentation. Environ Technol Innov 7:1–11
- Ojuederie OB, Babalola OO (2017) Microbial and plant-assisted bioremediation of heavy metal polluted environments: a review. Int J Environ Res Public Health 14:1504
- Okshevsky M, Gautier E, Farner JM, Schreiber L, Tufenkji N (2020) Biofilm formation by marine bacteria is impacted by concentration and surface functionalization of polystyrene nanoparticles in a species-specific manner. Environ Microbiol Rep 12:203–213
- Olu-arotiowa OA, Ajani AO, Aremu MO, Agarry SE (2019) Bioremediation of atrazine herbicide contaminated soil using different bioremediation strategies. J Appl Sci Environ Manag 23:99– 109

- Ossai IC, Ahmed A, Hassan A, Hamid FS (2020) Remediation of soil and water contaminated with petroleum hydrocarbon: a review. Environ Technol Innov 17:100526
- Pal A, Paul AK (2008) Microbial extracellular polymeric substances: central elements in heavy metal bioremediation. Indian J Microbiol 48:49–64
- Pal S, Kundu A, Banerjee TD, Mohapatra B, Roy A, Manna R, Kazy SK (2017) Genome analysis of crude oil degrading *Franconibacter pulveris* strain DJ34 revealed its genetic basis for hydrocarbon degradation and survival in oil contaminated environment. Genomics 109:374–382
- Paliwal V, Puranik S, Purohit HJ (2012) Integrated perspective for effective bioremediation. Appl Biochem Biotechnol 166:903–924
- Pandey P, Pathak H, Dave S (2018) Metabolic engineering and future prospects in bioremediation: a minireview. Int J Res Anal Rev 5:664–668
- Panwichian S, Kantachote D, Wittayaweerasak B, Mallavarapu M (2011) Removal of heavy metals by exopolymeric substances produced by resistant purple nonsulfur bacteria isolated from contaminated shrimp ponds. Electron J Biotechnol 14:2
- Park BG, Kim M, Kim J, Yoo H, Kim BG (2017) Systems biology for understanding and engineering of heterotrophic oleaginous microorganisms. Biotechnol J 12:1600104
- Partovinia A, Rasekh B (2018) Review of the immobilized microbial cell systems for bioremediation of petroleum hydrocarbons polluted environments. Crit Rev Environ Sci Technol 48:1–38
- Paul D, Pandey G, Jain RK (2005) Suicidal genetically engineered microorganisms for bioremediation: need and perspectives. BioEssays 27:563–573
- Perpetuo EA, Souza CB, Nascimento CAO (2011) Engineering bacteria for bioremediation. In: Capri A (ed) Progress in molecular and environmental bioengineering-from analysis and modeling to technology applications. IntechOpen, London. https://doi.org/10.5772/19546
- Philp JC, Atlas RM (2005) Bioremediation of contaminated soils and aquifers. In: Atlas RM, Philip JC (eds) Bioremediation: applied microbial solutions for real-world environmental cleanup. ASM Press, Washington, pp 139–236
- Pieper DH, Reineke W (2000) Engineering bacteria for bioremediation. Curr Op Biotechnol 11: 262–270
- Pimviriyakul P, Wongnate T, Tinikul R, Chaiyen P (2020) Microbial degradation of halogenated aromatics: mechanisms and enzymatic reactions. Microb Biotechnol 13:67–86
- Poi G, Aburto-Medina A, Mok PC, Ball AS, Shahsavari E (2017) Large-scale bioaugmentation of soil contaminated with petroleum hydrocarbons using a mixed microbial consortium. Ecol Eng 102:64–71
- Pradhan JK, Kumar S (2012) Metals bioleaching from electronic waste by *Chromobacterium* violaceum and pseudomonads sp. Waste Manag Res 30:1151–1159
- Prakash V, Saxena S, Sharma A, Singh S, Singh SK (2015) Treatment of oil sludge contamination by composting. J Bioremed Biodegr 6:284
- Pratush A, Kumar A, Hu Z (2018) Adverse effect of heavy metals (as, Pb, hg, and Cr) on health and their bioremediation strategies: a review. Int Microbiol 21:97–106
- Prince RC (1997) Bioremediation of marine oil spills. Trends Biotechnol 15:158-160
- Priya A, Hait S (2017) Comparative assessment of metallurgical recovery of metals from electronic waste with special emphasis on bioleaching. Environ Sci Pollut Res 24:6989–7008
- Quintella CM, Mata AM, Lima LC (2019) Overview of bioremediation with technology assessment and emphasis on fungal bioremediation of oil contaminated soils. J Environ Manag 241:156– 166
- Raghul SS, Bhat SG, Chandrasekaran M, Francis V, Thachil ET (2014) Biodegradation of polyvinyl alcohol-low linear density polyethylene-blended plastic film by consortium of marine benthic vibrios. Int J Environ Sci Technol 11:1827–1834
- Rahman Z, Singh VP (2020) Bioremediation of toxic heavy metals (THMs) contaminated sites: concepts, applications and challenges. Environ Sci Pollut Res 27:27563–27581
- Raj R, Dalei K, Chakraborty J, Das S (2016) Extracellular polymeric substances of a marine bacterium mediated synthesis of CdS nanoparticles for removal of cadmium from aqueous solution. J Colloid Interface Sci 462:166–175

- Rajakaruna SS, Robinson TAW (2016) Application of recombinant DNA technology (genetically modified organisms) to the advancement of agriculture, medicine, bioremediation and biotechnology industries. J Appl Biotechnol Bioeng 1:78–80
- Rasoulnia P, Mousavi SM (2016) Maximization of organic acids production by *aspergillus Niger* in a bubble column bioreactor for V and Ni recovery enhancement from power plant residual ash in spent-medium bioleaching experiments. Bioresour Technol 216:729–736
- Ravenschlag K, Sahm K, Pernthaler J, Amann R (1999) High bacterial diversity in permanently cold marine sediments. Appl Environ Microbiol 65:3982–3989
- Raziyafathima M, Praseetha PK, Rimal Isaac RS (2016) Microbial degradation of plastic waste: a review. Chem Biol Sci 4:231–242
- Reddy C, Mathew Z (2002) Bioremediation potential of white rot fungi. In: Gadd GM (ed) Fungi in Bioremediation. Cambridge University Press, Cambridge, pp 52–78
- Redfern LK, Gardner CM, Hodzic E, Ferguson PL, Hsu-Kim H, Gunsch CK (2019) A new framework for approaching precision bioremediation of PAH contaminated soils. J Hazard Mater 378:120859
- Ren X, Zeng G, Tang L, Wang J, Wang J, Deng Y, Liu Y, Peng B (2018) The potential impact on the biodegradation of organic pollutants from composting technology for soil remediation. Waste Manag 72:138–149
- Roager L, Sonnenschein EC (2019) Bacterial candidates for colonization and degradation of marine plastic debris. Environ Science Technol 53:11636–11643
- Robinson NJ, Whitehall SK, Cavet JS (2001) Microbial metallothioneins. Adv Microb Physiol 44: 183–213
- Roh Y, Gao H, Vali H, Kennedy DW, Yang ZK, Gao W, Dohnalkova AC, Stapleton RD, Moon JW, Phelps TJ, Fredrickson JK, Zhou J (2006) Metal reduction and iron biomineralization by a psychrotolerant Fe (III)-reducing bacterium, *Shewanella* sp. strain PV-4. Appl Environ Microbiol 72:3236–3244
- Ruiz ON, Alvarez D, Gonzalez-Ruiz G, Torres C (2011) Characterization of mercury bioremediation by transgenic bacteria expressing metallothionein and polyphosphate kinase. BMC Biotechnol 11:82
- Salunke BK, Sawant SS, Lee S-I, Kim BS (2015) Comparative study of MnO<sub>2</sub> nanoparticle synthesis by marine bacterium *Saccharophagus degradans* and yeast *Saccharomyces cerevisiae*. Appl Microbiol Biotechnol 99:5419–5427
- Samanta SK, Singh OV, Jain RK (2002) Polycyclic aromatic hydrocarbons: environmental pollution and bioremediation. Trends Biotechnol 20:243–248
- Sanscartier D, Zeeb B, Koch I, Reimer K (2009) Bioremediation of diesel contaminated soil by heated and humidified biopile system in cold climates. Cold Regions Sci Technol 55:167–173
- Sarkar J, Roy A, Asr P, Kazy SK (2020) Accelerated bioremediation of petroleum refinery sludge through biostimulation and bioaugmentation of native microbiome. In: Shah MP, Rodriguez-Couto S, Sengör S (eds) Emerging Technologies in Environmental Bioremediation. Elsevier, New York, pp 23–66
- Saum L, Jiménez MB, Crowley D (2018) Influence of biochar and compost on phytoremediation of oil-contaminated soil. Int J Phytoremediation 20:54–60
- Saxena G, Bharagava RN (2016) Ram Chandra: advances in biodegradation and bioremediation of industrial waste. Clean Techn Environ Policy 18:979–980
- Saxena G, Kishor R, Bharagava RN, Das P, Gupta PK, Kumar N (2021) Emerging green technologies for biological treatment of leather tannery chemicals and wastewater. In: Saxena G, Kumar V, Shah MP (eds) Bioremediation for environmental sustainability toxicity, mechanisms of contaminants degradation, detoxification, and challenges. Elsevier, New York, pp 435–460
- Sayavedra-Soto LA, Gvakharia B, Bottomley PJ, Arp DJ, Dolan ME (2010) Nitrification and degradation of halogenated hydrocarbons–a tenuous balance for ammonia-oxidizing bacteria. Appl Microbiol Biotechnology 86:435–444

- Sekiguchi T, Ebisui A, Nomura K, Watanabe T, Enoki M, Kanehiro H (2009) Biodegradation of several fibers submerged in deep sea water and isolation of biodegradable plastic degrading bacteria from deep ocean water. Nippon Suisan Gakkaishi 75:1011–1018
- Sekiguchi T, Saika A, Nomura K, Watanabe T, Watanabe T, Fujimoto Y, Enoki M, Sato T, Kato C, Kanehiro H (2011b) Biodegradation of aliphatic polyesters soaked in deep seawaters and isolation of poly (ε-caprolactone)-degrading bacteria. Polym Degrad Stability 96:1397–1403
- Sekiguchi T, Sato T, Enoki M, Kanehiro H, Uematsu K, Kato C (2011a) Isolation and characterization of biodegradable plastic degrading bacteria from deep-sea environments. JAMSTEC Rep Res Dev 11:33–41
- Senel UG, Hanay O (2017) Metal-microbe interactions and microbial bioremediation of toxic metals. In: Uslu GS, Hanay O (eds) Handbook of metal-microbe interactions and bioremediation. Taylor & Francis, New York, pp 99–110
- Sharma B, Dangi AK, Shukla P (2018a) Contemporary enzyme based technologies for bioremediation: a review. J Environ Manag 210:10–22
- Sharma R, Bhardwaj R, Gautam V, Bali S, Kaur R, Kaur P, Shama M, Kumar V, Sharma A, Sonia TAK, Vig AP, Ohri P (2018b) Phytoremediation in waste management: Hyperaccumulation diversity and techniques. In: Hasanuzzaman M, Nahar K, Fujita M (eds) Plants under metal and metalloid stress. Springer, Singapore, pp 277–302
- Shi K, Xue J, Xiao X, Qiao Y, Wu Y, Gao Y (2019) Mechanism of degrading petroleum hydrocarbons by compound marine petroleum-degrading bacteria: surface adsorption, cell uptake, and biodegradation. Energy Fuel 33:11373–11379
- Shukla KS, Mangwani N, Karley D, Rao TS (2017) Bacterial biofilms and genetic regulation for metal detoxification. In: Das S, Dash HR (eds) Handbook of metal-microbe interactions and bioremediation. Taylor & Francis, New York, pp 317–332
- Silva-Castro GA, Uad I, Gónzalez-López J, Fandiño CG, Toledo FL, Calvo C (2012) Application of selected microbial consortia combined with inorganic and oleophilic fertilizers to recuperate oil-polluted soil using landfarming technology. Clean Techn Environ Policy 14:719–726
- Silver S (1998) Genes for all metals-a bacterial view of the periodic table. J Ind Microbiol Biotechnol 20:1–12
- Singh H (2006) Mycoremediation: fungal bioremediation. Wiley-Interscience, New York, NY, USA
- Singh J, Yadav P, Pal AK, Mishra V (2020) Water pollutants: origin and status. In: Pooja D, Kumar P, Singh P, Patil S (eds) Sensors in water pollutants monitoring: role of material. Springer, Singapore, pp 5–20
- Stincone P, Brandelli A (2020) Marine bacteria as source of antimicrobial compounds. Crit Rev Biotechnol 40:306–319
- Sudhakar M, Priyadarshini C, Doble M, Murthy PS, Venkatesan R (2007) Marine bacteria mediated degradation of nylon 66 and 6. Int Biodeter Biodegrad 60:144–151
- Suresh K, Reddy GSN, Sengupta S, Shivaji S (2004) Deinococcus indicus sp. nov., an arsenicresistant bacterium from an aquifer in West Bengal, India. Int J Syst Evol Microbiol 54:457–461
- Swift G (1997) Non-medical biodegradable polymers: environmentally degradable polymers. In: Domb AJ, Kost J, Wiseman DM (eds) Handbook of biodegradable polymers. Hardwood Academic, Amsterdam, pp 473–511
- Talukdar D, Jasrotia T, Sharma R, Jaglan S, Kumar R, Vats R, Kumar R, Mahnashi M, Umar A (2020) Evaluation of novel indigenous fungal consortium for enhanced bioremediation of heavy metals from contaminated sites. Environ Technol Innov 20:101050
- Theerachat M, Guieysse D, Morel S, Remaud-Siméon M, Chulalaksananukul W (2018) Laccases from marine organisms and their applications in the biodegradation of toxic and environmental pollutants: a review. Appl Biochem Biotechnol 187:583–611
- Timková I, Sedláková-Kaduková J, Pristaš P (2018) Biosorption and bioaccumulation abilities of actinomycetes/streptomycetes isolated from metal contaminated sites. Separations 5:54

- Tipre DR, Purohit MS, Dave SR (2020) Marine bacteria–a treasure house of valuable products and functions. In: Niche M (ed) Applications in pharmaceutical sciences. Springer, Singapore, pp 415–436
- Tiwari N, Santhiya D, Sharma JG (2020) Microbial remediation of micro-nano plastics: current knowledge and future trends. Environ Pollut 265:115044
- Tyagi B, Kumar N (2021) Bioremediation: principles and applications in environmental management. In: Bioremediation for environmental sustainability toxicity, mechanisms of contaminants degradation, detoxification, and challenges. Elsevier, New York, pp 3–28
- Tyagi M, da Fonseca MMR, de Carvalho CC (2011) Bioaugmentation and biostimulation strategies to improve the effectiveness of bioremediation processes. Biodegradation 22:231–241
- Urbanek AK, Rymowicz W, Mirończuk AM (2018) Degradation of plastics and plastic-degrading bacteria in cold marine habitats. Appl Microbiol Biotechnol 102:7669–7678
- Urbanek AK, Rymowicz W, Strzelecki MC, Kociuba W, Franczak Ł, Mirończuk AM (2017) Isolation and characterization of Arctic microorganisms decomposing bioplastics. AMB Expr 7:148
- Urionabarrenetxea E, Garcia-Velasco N, Anza M, Artetxe U, Lacalle R, Garbisu C, Becerril T, Soto M (2021) Application of in situ bioremediation strategies in soils amended with sewage sludges. Sci Total Environ 766:144099
- Varjani SJ (2017) Microbial degradation of petroleum hydrocarbons. Bioresour Technol 223:277– 286
- Varjani SJ, Agarwal AK, Gnansounou E, Gurunathan B (2018) Introduction to environmental protection and management. In Varjani SJ, Agarwal AK, Gnansounou E, Gurunathan B (Ed) Bioremediation: applications for environmental protection and management. Springer Nature, pp. 1–6
- Varjani S, Upasani VN, Pandey A (2020) Bioremediation of oily sludge polluted soil employing a novel strain of Pseudomonas aeruginosa and phytotoxicity of petroleum hydrocarbons for seed germination. Sci Total Environ 737:139766
- Vásquez-Murrieta MS, Hernández-Hernández OJ, Cruz-Maya JA, Cancino-Díaz JC, Jan-Roblero J (2016) Approaches for removal of PAHs in soils: bioaugmentation, biostimulation and bioattenuation. In: Larramendy ML, Soloneski S (eds) Soil contamination–current consequences and further solutions. IntechOpen, London. https://doi.org/10.5772/64682
- Verma S, Kuila A (2019) Bioremediation of heavy metals by microbial process. Environ Technol Innov 14:100369
- Vidali M (2001) Bioremediation: an overview. Pure Appl Chem 73:1163–1172
- Vikas M, Dwarakish GS (2015) Coastal pollution: a review. Aquat Proc 4:381-388
- Von Canstein H, Kelly S, Li Y, Wagner-Döbler I (2002) Species diversity improves the efficiency of mercury-reducing biofilms under changing environmental conditions. Appl Environ Microbiol 68:2829–2837
- Wang L, Ji B, Hu Y, Liu R, Sun W (2017) A review on in situ phytoremediation of mine tailings. Chemosphere 184:594–600
- Wang X, Müller WE (2009) Marine biominerals: perspectives and challenges for polymetallic nodules and crusts. Trends Biotechnol 27:375–383
- Wei YT, Wu SC, Yang SW, Che CH, Lien HL, Huang DH (2012) Biodegradable surfactant stabilized nanoscale zero-valent iron for in situ treatment of vinyl chloride and 1,2-dichloroethane. J Hazard Mater 211:373–380
- Whelan MJ, Coulon F, Hince G, Rayner J, McWatters R, Spedding T, Snape I (2015) Fate and transport of petroleum hydrocarbons in engineered biopiles in polar regions. Chemosphere 131: 232–240
- Widada J, Nojiri H, Omori T (2002) Recent developments in molecular techniques for identification and monitoring of xenobiotic-degrading bacteria and their catabolic genes in bioremediation. Appl Microbiol Biotechnol 60:45–59
- Williams J (2006) Bioremediation of contaminated soils: a comparison of in situ and ex situ techniques. Available at: http://home.eng.iastate.edu/Btge/ce421-521/jera.pdf

- Wu G, Coulon F (2015) Protocol for biopile construction treating contaminated soils with petroleum hydrocarbons. In: McGenity T, Timmis K, Nogales B (eds) Hydrocarbon and lipid microbiology protocols. Springer Protocols Handbooks, Springer, Berlin, pp 181–194
- Wu M, Dick WA, Li W, Wang X, Yang Q, Wang T, Xu L, Zhang M, Chen L (2016) Bioaugmentation and biostimulation of hydrocarbon degradation and the microbial community in a petroleum contaminated soil. Int Biodeter Biodegrad 107:158–164
- Xu M, Fu X, Gao Y, Duan L, Xu C, Sun W, Li Y, Meng X, Xiao X (2020) Characterization of a biosurfactant-producing bacteria isolated from marine environment: surface activity, chemical characterization and biodegradation. J Environ Chem Eng 8:104277
- Xue J, Yu Y, Bai Y, Wang L, Wu Y (2015) Marine oil-degrading microorganisms and biodegradation process of petroleum hydrocarbon in marine environments: a review. Curr Microbiol 71: 220–228
- Yadav A, Chowdhary P, Kaithwas G, Bharagava RN (2017) Toxic metals in the environment threats on ecosystem and bioremediation approaches. In: Das S, Dash HR (eds) Handbook of metal-microbe interactions and bioremediation. Taylor & Francis, New York, pp 127–142
- Yang J, Wang Q, Wang Q, Wu T (2008) Comparisons of one-step and two-step bioleaching for heavy metals removed from municipal solid waste incineration fly ash. Environ Eng Sci 25:783– 789
- Yuan J, Ma J, Sun Y, Zhou T, Zhao Y, Yu F (2020) Microbial degradation and other environmental aspects of microplastics/plastics. Sci Total Environ 715:136968
- Zakaria NN, Convey P, Gomez-Fuentes C, Zulkharnain A, Sabri S, Shaharuddin NA, Ahmad SA (2021) Oil bioremediation in the marine environment of Antarctica: a review and bibliometric keyword cluster analysis. Microorganisms 9:419
- Zhang H, Yuan X, Xiong T, Wang H, Jiang L (2020) Bioremediation of co-contaminated soil with heavy metals and pesticides: influence factors, mechanisms and evaluation methods. Chem Eng J 398:125657
- Zulkifli SN, Rahim HA, Lau WJ (2018) Detection of contaminants in water supply: a review on state-of-the-art monitoring technologies and their applications. Sensors Actuators B Chem 255: 2657–2689
- Zylstra GJ, Wackett LP, Gibson DT (1989) Trichloroethylene degradation by *Escherichia coli* containing the cloned *pseudomonas putida* F1 toluene dioxygenase genes. Appl Environ Microbiol 55:3162–3166

# **Chapter 9 Marine Bacteria for Biofertilizers**



Poonam Singh, Kaleemunnisa FNU, and Telma Encarnação

**Abstract** The fertilizer industry is growing with time and demand. The production of most used nitrogenous fertilizer utilizes methane supplies and sums up to 3% of worldwide greenhouse emissions. Chemical fertilizers have created a large number of environmental dead zones with low oxygens where life forms cannot grow. With a decline in soil quality, the yield of crops go and have gone from minimum to none. Stress caused by abiotic and biotic factors is the constraint that needs to be overcome as it affects the productivity of the crops. Improvement of crop growth under stress depends on unexplored tools of nature. One of such tools is microbe, which are still unexplored and found in enormous quantity. Microbes that produce a variety of metabolites either stimulate the nutrient formation or increase the uptake of nutrients in plants it is symbiotically associated with. The term microbial biofertilizers is associated with the formulations of sufficient densities of strains in active or inactive form of microorganisms, that can be used in rhizospheres for plant growth. The microbes used can be either a single strain or in combination with different specific beneficial effects.

P. Singh (🖂)

T. Encarnação

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

PTScience Avenida do Atlântico, Lisbon, Portugal

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

K. FNU

Seidenberg School of CSIS, Pace University, New York, NY, USA

Centre for Rapid and Sustainable Product Development, Polytechnic Institute of Leiria, Marinha Grande, Portugal

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_9



An overview of responses of bacteria as biofertilizer on biochemical properties of the soil

**Keywords** Biofertilizers · Bacteria · Nutrients uptake · Regulations · Machine learning · Artificial intelligence

# 9.1 Introduction

Modern agriculture requires expeditious yet responsible measures. The increasing crop productivity for meeting the demand of the growing population has been possible due to synthetic chemical fertilizers and pesticides. The practices followed by farmers have been changing throughout the years with a negligible focus on the problems that are impacting the environment (Mishra et al. 2012; Swapna 2013). This is causing many alarming situations that are going to impact the earth and living

beings not only in the present but also in near future. However, we cannot deny the fact that food security is an important cause and we need to fulfill it. The continuous climate change with increasing temperature and  $CO_2$  levels is a threat to the agriculture (Spiertz 2009).

Living organisms that can deliver nutrients to soil, plant or can help in the germination increasing the biomass of the root is termed as biofertilizer. The use of microbial metabolic ability for degradation/removal of environmental pollutants provides an economic and safe alternative compared to other physicochemical methodologies. Microbes have an ability to alter the microbiota of the soil and living forms around the particular area where they are used as biofertilizer (Hafeez et al. 2006). These microbes might be found in different sources, the most common ones are bacteria, fungi, and cyanobacteria (blue-green algae). These microbes have enzymes that perform the nutrient conversions maintaining the nitrogen and phosphorus balance and hence keeping the soil constitution in equilibrium (Ritika and Utpal 2014).

## 9.1.1 Marine Organisms as Biofertilizer

The majorities of microalgae are capable to fix the atmospheric nitrogen and are effectively used as biofertilizers. Microalgae or microphytes are microscopic algae invisible to the naked eye. They are phytoplankton typically found in freshwater and marine systems, containing all the essential nutrients needed for plant growth. Symbiotic marine origin form is not only environmentally friendly but also cost-effective (Ravikumar et al. 2005). In general, the molecules produced from marine sources possess no harm to the environment and provide a foundation for organic biomass production. The main factors affecting marine diazotroph distribution in the ocean are oxygen, light, temperature, inorganic N forms, phosphorus (P), Fe, and organic matter (Fu et al. 2014).

#### 9.2 Literature Review

Conversion of molecular nitrogen into ammonia and other nitrogenous compound that can enrich the soil is primarily done by Diazotrophs that include *Rhizobia*, *Frankia*, and *Azospirillum*. These bacteria provide phytohormones to the host and confer resistance against pathogens. The exchange of signals occurs through diffusion in symbiosis pathway. The soil bacteria are mostly studied in the rhizobia–legume and legume–arbuscular mycorrhiza symbioses (Pankievicz et al. 2021). The evolutionary link has been recognized between these two symbioses.

The ability of rhizobia to fix the nitrogen in the root nodule and maintaining the nutrient uptake during stress conditions, makes it interesting for the crops like soyabean reducing the production cost and reducing adverse environmental impacts.

The impact caused by the microbes can be determined by their ability to promote plant growth in any kind of stress condition. The soil bacteria are known to promote plant growth by providing ammonia, enhancing the uptake of indole-3-acetic acid (IAA), gibberellic acids, cytokinin, plant hormones (Grobelak et al. 2015).

The deep ocean is an unsolved mystery and has organisms that have essential application in different industries and one of them is as natural fertilizers. One of the important evidence to identify nitrogen-fixing bacteria is found in their DNA fingerprints. That contains the gene specifically encoding the protein responsible for nitrogen fixation (Bianchi 2011).

Cyanobacteria is a biological unit to convert solar energy to chemical energy and contributing to the atmospheric oxygen. The capture of photon and by cyanobacteria is a source to generate biofuels and chemicals using sunlight and the greenhouse gas CO<sub>2</sub>. The mechanism of how cyanobacteria work as a bio fertilizer in different parts of a plant is shown in Fig. 9.1. Synechocystis sp. PCC 6803 is one of the most extensively studied species in research due to its similarity with the plant chloroplasts. The carbon metabolism under variety of conditions makes it suitable for the stress adaption in plants and studying stress using this cyanobacteria species as a model. The genetic modification in Synechocystis 6803 has ensured that the species can produce diverse types of chemicals and work as a photosynthetic host (Ouiroz-Arita et al. 2019). In absence of nitrogenase it cannot fix  $N_2$  but by producing cyanophycin. Cyanophycin is a polypeptide containing multiple arginine and aspartate residues. However, it is able to store ammonium nitrogen inside of the cell. This generates a possibility to use it as a bio fertilizer by removing the nitrogen from the wastewater. A study done by Krasikov and other researchers found that nitrogen production protein is upregulated, photosynthetic carbon fixation is done in a limited manner, and cells hardly have any enzymatic activity under reduced nitrogen conditions in Synechocystis 6803. Thiobacillus a widely distributed bacterium



**Fig. 9.1** Depiction of working mechanism of cyanobacteria as biofertilizer in the growth of the plant: (1) Colonization of leaf by cyanobacteria. (2) Colonization in the root hair, and root surface. (3) Co-inoculation with 2,4-dichlorophenoxy acetic acid (2,4-D) (synthetic auxin) and Nostoc spp. increases para-nodule formation and nitrogen fixation (Lee and Ryu 2021)

produces nitrous oxide and is found in both soil and water sources. *Thiobacillus* is an autotroph capable of using both sulfur and iron(II) as an electron donor (Hedrich et al. 2011; Krasikov et al. 2012). Canola, rapeseed plant when inoculated with Iron along with *Thiobacillus* sp., along with the application of Zn and Fe micronutrient, showed higher content of zinc and iron in seeds. The yield and oil content in canola was enhanced too (Jashni et al. 2017). Green beans had an increase in their productivity when *Thiobacillus*, sulfur application was used with zinc. The mechanism thought is reduction of the pH soil by providing sulfur, which results in bigger food yield. The maize and wheat production was increased by applying *Thiobacillus thiooxidans* when used with iron and zinc (Davaran et al. 2016; Visser 1997).

*Bacillus cereus* UW85 and *Pseudomonas protegens* CHA0, the known biocontrol agents, when used together can aim at different pathogenic fungi, producing metabolites against other unwanted growth in plants. The bacteria found on land soil convert N<sub>2</sub> into organic nutrients like ammonium (NH<sup>4+</sup>) and nitrate (NO<sup>3-</sup>) that can be utilized by plants. In marine infrastructure, cyanobacteria, blue-green in color are abundantly found, fixing nitrogen. Approximately 90% of natural nitrogen fixation is performed via these organisms also known as diazotrophs. NO<sub>3</sub><sup>-</sup> or O<sub>2</sub> and temperature are important to understand the wider latitudinal distribution of different diazotrophs in the ocean tropical surface waters and tropical marine waters (Caulier et al. 2018).

Synechocystis has carboxysome that concentrates inorganic carbon for biomass growth. It is a special organelle which encapsulates Ribulose-1,5-bisphosphate carboxylase (RuBisCO), this enzyme helps with  $CO_2$  fixation. Inorganic polyphosphate can be stored by *Synechocystis* 6803, photosynthetic host for the production of diverse types of chemicals. *phoU* and *sphU* are the phosphate regulator mutant genes that enhance cyanobacterial phosphorus uptake. Mineral fertilizers that contain macronutrients such as nitrogen, phosphorus, and potassium when added increase the crop yield (Tabita 1999).

#### 9.3 Understanding the Mechanisms of Nutrient Uptake

Biofertilizer, Nitragin of *Rhizobium* sp. (Singh and Singh 2019) was used in 1895 by Nobbe and Hiltner. Mycorrhizal fungi inoculants had a positive impact on plant growth promotion (PGP), studied in the 1950s (Koide and Mosse 2004). It has been a great effort to analyze and understand the mechanism of nitrogen fixation via microbes. The difference in location came up as a predominant factor. For example, if the bacteria are growing on coastal or open ocean, deep or shallow, warm or cold water, it makes a variation in rate and order of fixation. N<sub>2</sub> in the rhizosphere and bulk soil can be fixed into atmospheric N<sub>2</sub> in cereal crops such as wheat, maize, rice, and corn by heterotrophic free-living diazotrophic species such as Azotobacter, Azospirillum, and cyanobacteria. In other crops, N availability has been enhanced by free-living diazotrophs (Zeffa et al. 2019).

#### 9.3.1 Nitrogen Fixation by the Bacteria

Only a small number of marine bacteria can convert dissolved  $N_2$  gas into bioavailable ammonia (NH<sub>3</sub>) and known as N<sub>2</sub> fixers. This is an intensely energy-requiring process carried out by different kinds of cyanobacteria species. Trichodesmium, Oscillatoria, Lyngby, Aphanizomenon, Nodularia Richelia, Calothrix Crocosphaera watsonii, Marine diazotrophs, heterotrophic bacteria, phototrophic bacteria, and *Cyanothece* are the species with nitrogenase complex, which catalyzes N2 fixation. Protein bound by molybdenum and iron constitutes nitrogenase. Oxygen destroys nitrogenase, to cope up with this phenomenon cyanobacteria generate specialized N<sub>2</sub>-fixing cells as a protection mechanism. Heterocyst cells are formed that are deficient in oxygen. Time delays are another strategy used against oxidation by proteobacterial diazotrophs and Trichodesmium and heterocyst-forming cyanobacteria (Pajares et al. 2019).

# 9.3.2 Phosphorus: Solubilization and Mineralization by the Bacteria

Phosphorus is one of the main limiting elements and second most important macronutrient for biomass production found in phosphate and sedimentary rock in the marine environment. *Pseudomonas aeruginosa* from marine origin, was reported to the produce pyoverdin type of siderophores, when tested for additional metabolite production, was found to solubilize phosphate. Marine bacteria are capable of solubilizing inorganic phosphorus from insoluble compounds to be assimilated by plants. Marine systems receive phosphorous minerals by continental weathering, however the weathering depends on environmental factors. That includes how old the earth is, erosion conditions, atmospheric composition, phosphorus content in rock, microaggregate fractions from the soil, and biological response. Achromobacter can be found in marine sources. It mineralizes phosphate through production of extracellular enzymes (Kalayu 2019; Rane et al. 2008).

#### 9.3.3 Potassium: Solubilization by the Bacteria

Potassium a vital macronutrient, required in enzyme activation, protein synthesis, and photosynthesis. It is absorbed from the root and helps with the movement of the water, nutrients, and carbohydrates in most agronomic crops. Potassium-solubilizing bacteria minimize health hazards that are resulted by the use of chemical fertilizers. *Serratia marcescens* can exhibit plant growth promoting molecules hence a potential PGPB with diverse activities tested by Sindu et al. The processes like cell osmotic regulation and enzyme activation require potassium, the availability depends on the

quantity in the soil (Janice and Carmen 2007). Potassium is present in the soil in three different forms, in readily unavailable, slowly available, and readily available. The minerals present in soil (feldspar and biotite) have the potassium mostly in readily unavailable form, clay has slowly available, non-exchangeable form to the plants. Water-soluble potassium is readily available and exchangeable present in soil (Teotia et al. 2016).

#### 9.3.4 Oxidation of Sulfur by Bacteria

Sulfur an essential nutrient for the production of proteins, glutathione, chloroplast membrane lipids, coenzymes, and vitamins is mostly found in organic form. The inorganic sulfur that is in the form of sulfate has been depleted recently as a consequence of many environmental factors. Element sulfur based fertilizers are present in the market, but for plants uptake it needs to be oxidized to sulfate. Bacteria are capable of metabolizing sulfur and its compounds (Tang et al. 2009). Hydrogen sulfide (H2S), sulfur, and thiosulfate (S2O32–) are oxidized by bacteria to convert it to sulfate. Thiobacillus, found in marine environment, oxidizes sulfur. The sulfate produced has several advantages when it goes deep into the ground it produces sulfuric acid dissolving metal, concrete, and steel to soluble monomeric and oligomeric compounds. Hence, improving the quality of the soil. Phreagena soyoae is one of the intracellular symbiotic sulfur-oxidizing bacteria (Lin et al. 2018). Purple sulfur bacteria Thiospirillum are strict anaerobes and convert sulfide to sulfur and then sulfate (Pfennig 1975).

#### 9.3.5 Micronutrients Solubilization by Bacteria

Micronutrients are essential for the growth and enzymatic reactions in plants. Processes like photosynthesis, respiration, water oxidation, and oxidative stress protection (Castro et al. 2018) are dependent on the intake of iron, zinc, copper, silicon, cobalt, nickel. If the soil is deficient in micronutrients, it will affect the production of the crop. Silicate classified as a beneficial nutrient by the Association of American Plant Food Control Officials, is found in soil but not readily available. Silicate solubilizing bacteria found in soil, water, marine sediments, and silicate minerals exhibiting their unique property can solubilize the silicate and make it accessible in the form of monosilicic acid for plants (Vasanthi et al. 2018). *Aeromonas, Rhizobium, Enterobacter*, and *Bacillus* are the most studied species.

Iron sequestration is performed by siderophores that possess high affinity and selectivity for Fe(III). They help the plants to survive in iron deficient environment. Siderophores work during many physiological processes such as oxidative phosphorylation and nucleic acid biosynthesis. Insoluble Fe(III) is transported to microbial cells by membrane receptors, after that reduction of Fe (III) to Fe (II) occurs by

redox processes. *Marinobacter* sp. DS40M6 produces the suit of marinobactin siderophores (Martinez et al. 2003). *Synechococcus sp.* and *Vibrio cyclitrophicus* 1F53 have been found to have peptidic hydroxamate siderophore that has an ability to chelate metals via the presence of two oxygen atoms (Chen et al. 2019).

#### 9.4 Regulations

The environmental impact of traditionally used agricultural techniques has moderately led European union to put some regulations on it. The regulatory framework has encouraged better use of microorganisms to improve agricultural practices according to their taxonomic classification and usefulness.

Marine forms maximize the use of the waste and hence play a role in the recycling of the nutrients.

The plethora of marine sources present on this earth remains majorly unexplored giving a subsequent scope for research of the potential application of the extracted components. New advancements have made it possible to investigate the compounds and facilitate the expedition to analyze new components from marine life. Figure 9.2



Fig. 9.2 Nitrogen cycle taking place through marine bacteria (White et al. 2020)

is the interpretation of nitrogen cycle in which conversion of nitrogen takes place in different forms via marine bacteria in the ocean and circulating through different ecosystems.

The use of waste from the production of biomass to produce molecules encourages the circular economy. The use of biofertilizers from bacteria should not cause any damage to the environment or human health. To ensure the safety, The European authorities have developed the recent new policy content of the EU Circular Economy Fertilising Products Regulation (EU 2019/1009) on biofertilizer in order to protect humans, plants, animals, and the environment. To trade the fertilizers the products must comply with strict regulations and affix the CE mark. The producers must ensure and demonstrate the labeling requirements. The EU regulation directs the product to be divided into function categories that will determine the specific requirements of quality and safety to be used for intended purposes. There are certain control criteria involved for different categories of the product before making it available to the market. The harmonized quality standards introduce new limit values for contaminants to trade the fertilizers keeping the health attributes into account (Schmidt and Haccius 2020; Saha et al. 2017).

The biofertilizer has to guarantee reduction of soil and environmental risk and provide a high level of soil protection this is regulated with a limit for cadmium that is <1.5 mg/kg. The low cadmium content allows producers to obtain improved level of soil production. The natural source boosts the circular economy and also the use of recycled bio wastes. The use of bacteria as a source of biofertilizers also emphasizes on green innovation reducing the need to use other sources of nutrients for the growth of the plants. The structured framework ensuring the detailed guide-lines to use the biofertilizer from bacteria encourages the farmers to use this abundantly available natural source hence avoiding toxic contaminants and elements. The presence of this alternative bacteria based fertilizer in accordance with the market if applied all over the world would remove all the costs in terms of clearance from the national borders. It will persist the homogenous protection of environment and health of the living beings as well (Calvo et al. 2014; Das et al. 2019).

# 9.5 Limitations

Extensive and long-term application of biofertilizers can cause technological, infrastructural, financial, environmental, human resources constraints. The use of biofertilizers may result in accumulation of salts, nutrients, and heavy metals. This could cause detrimental effects on plant growth. There are very few places where the extraction and purification of the marine bacteria is done; this unawareness can affect the quality of the crop and ultimately impact human health.

Variable concentrations of nutrients in different species and high implementation costs are something to be taken care of otherwise it makes the entire process too expensive. Even after having the desired qualities in a bacterial species it is possible that low transfer of micro- and macronutrients can cause nutritional deficiency. In comparison to chemical fertilizers large volumes of biofertilizers from bacteria are required for land application, this might be because the process is not yet standardized it is difficult to know exact contents of nutrients (Carvajal-Munoz and Carmona-Garcia 2012).

#### 9.6 Future Outlook

Machine Learning(ML) concept gives computers the ability to think and helps us to solve many problems. ML is a subset of artificial intelligence focused on using algorithms that learn and improve without being explicitly programmed to do so. ML is about learning from existing data to make predictions about the future. It is based on creating models from input datasets for data-driven decision-making. ML uses large datasets to identify (infer) patterns and make decisions (predictions). Automated decision-making is what makes ML so appealing. You can teach a system from a dataset and let the system act by itself to predict future. Currently many researchers' study concluded that there are enormous resources available in the extensive marine environment. These sources play a vital role in the development of renewable sources.

With the data available on the level of nutrients present in the marine bacteria, we can analyze the growth of plant and with the help of Artificial Intelligence algorithms we can predict the growth based on the nutrients content and also classify the nutrients (Larrañaga et al. 2006).

The process of machine learning is as follows (Fig. 9.3):

Machine learning is a method of data analysis, which is a sub section of Artificial Intelligence that automates analytical model building. Using these algorithms that iteratively learn from data, machine learning finds hidden patterns without being explicitly programmed.

These features of ML can be used on the data collected on bacteria from marine source for fertilization to classify the type of species, classification can be done on



Fig. 9.3 Flowchart of machine learning process

both binary and multivariate, which can help in deciding what kind of crops grows faster with different strain of bacteria. And we can predict the rate of growth at which these fertilizers are helping in achieving cost effectiveness, level of non-toxic achieved and also to check the level of contamination of the ground water. Also, you can check the subspecies of bacteria growing in different weather, temperature, and area (Fig. 9.4).

At the most fundamental level, machine learning is categorized into two main types: supervised learning and unsupervised learning. Supervised learning involves modeling the relationship between measured features of data and some label associated with the data; once this model is determined, it can be used to apply labels to new, unknown data. This is further subdivided into classification tasks and regression tasks: in classification, the labels are discrete categories, while in regression, the labels are continuous quantities. Supervised learning used in applications where historical data predicts likely future events (Goodfellow et al. 2016) (Fig. 9.5).

In supervised learning, an algorithm learns from labeled data. This labeled data is used for training algorithms. The algorithm receives a set of inputs along with outputs. An algorithm learns by comparing its actual output with correct outputs to find the errors. And then it modifies the model accordingly. Once the model is trained, we can test it by new input/data (Fig. 9.6).

There are more than two classes available for classification with the labels for a multi-class classifier. Decision tree is an example of multi-class classification.

Plant growth promoting bacteria (PGPB) contain all the essential nutrients needed for plant growth. The majorities of PGPB are capable to fix the atmospheric nitrogen and are effectively used as biofertilizers. PGPB have a tendency to harbor genetic factor for antibiotic and metal resistance. Through antibiotic resistance property acts as the intrinsic property which attributes to the presence of multidrug efflux pumps, which are involved in performing metabolic processes in bacteria. With the data, available on the level of nutrients present in the PGPB, we can analyze the growth of





Fig. 9.6 Diagram showing how the data contributes in understanding of a data



plant and with the help of Artificial Intelligence algorithms we can predict the growth based on the nutrients content and classify the nutrients (Fig. 9.7).

Data comes without labels and the task is to find similar data points in the dataset in order to identify any underlying hierarchical structure. Consider a dataset that contains purchase preferences from Amazon users, these users will likely form a cluster that behave in the similar way. They may have similar purchase behavior, they may spend same amount of money, or buy similar type of items. You can think of these different groups as different communities with different preferences. Once these communities are identified you can describe each data point which is each user in terms of the community it belongs to gaining deeper understanding of the data, and also reducing the number of data points that we have to deal with (Larrañaga et al. 2006; Goodfellow et al. 2016) (Fig. 9.8).

# 9.7 Conclusion

Excessive use of hazardous chemical, pesticides used as fertilizers is posing high risk for imminent agriculture. The alternative to these hazardous chemicals can be biofertilizers, which can provide sustainable agriculture as biofertilizers helps in increasing crop yielding, maintaining, and enduring soil fertility. This can help in meeting the food consumption demand globally. Microbes can help in enhancing the immunity, growth, of crops, by aiding the essential nutrients required in growth of a crops such as nitrogen, phosphorous, potassium, zinc, and silica, to dissolve which are present in the form of insolubilized. The advantage of using biofertilizers is that they are cost-effective, non-toxic, contribute for the reduction of contamination of the ground water, as well as eco-friendly.

Currently, many research studies concluded that there are enormous resources available in the extensive marine environment. These sources play a vital role in the development of renewable sources. Marine scientists aim to evaluate the factor that controls the reservoir of nitrogen in the ocean and its influence on the fertility of the soil. Using a wide range of different techniques causes uncertainty in the estimation of nitrogen fixation rates as they are not comparable among ecosystems.

The global biofertilizer market is set to reach to 3.9 billion in 2025 (Mitter et al. 2021). Even though biofertilizer are considered safe for the environment, they have the drawback of short shelf life and survival of the strains in different environments. In case the necessary requirements are fulfilled during storage and transportation, the entire process is very costly, and mass production is very tricky too. There is no proper guidance for farmers regarding the recommended use.

There is no easy way to deal with the coming challenges but the bio resources that can be exploited with undiscovered marine diversity constituted with culturable microbes might help to overcome them. Especially when Machine Learning can process and analyze the existing and ongoing data making the task organized and simpler.

Not only environmentalists but economists are also concerned with the increase in the cost of crop production due to the application of expensive chemical fertilizers. Availability of nutrients, if maintained and recovered by the ocean-bacteria, can solubilize the minerals in the soil. They can enhance and make the nutrients available for the growth of the plant by  $N_2$ -fixing, phosphate-/potassium hence working as biofertilizers.

Biofertilization requires less chemicals in the soil and might be effective in its decontamination of pollutants; therefore, can reduce the environmental impacts associated with agriculture activities.

Acknowledgements The authors acknowledge the Fundação para a Ciência e a Tecnologia (FCT) through the project PTDC/BTA-GES/2740/2020\_NABIA. The Coimbra Chemistry Centre (CQC) is supported by the FCT through the projects UIDB/00313/2020 and UIDP/00313/2020. CDRSP is financed by national funds through the FCT/MCTES (UIDB/00481/2020 & UIDP/00481/2020). We are grateful for funding from PTScience which is supported through the programs CENTRO-05-4740-FSE-001526 and FEDER.

#### References

- Bianchi TS (2011) The role of terrestrially derived organic carbon in the coastal ocean: a changing paradigm and the priming effect. Proc Natl Acad Sci 108(49):19473–19481
- Calvo P, Nelson L, Kloepper JW (2014) Agricultural uses of plant biostimulants. Plant and Soil: an International Journal on Plant-Soil Relationships 383:3–41
- Carvajal-Munoz JS, Carmona-Garcia CE (2012) Benefits and limitations of biofertilization in agricultural practices. Livest Res Rural Dev 24:3
- Castro PH, Lilay GH, Assunção AGL (2018) Regulation of micronutrient homeostasis and deficiency response in plants, pp 1–15
- Caulier S, Colau G, Liepin M, Desoignies N, Legreve A, Bragard C, Caulier S, Mahillon J (2018) Versatile antagonistic activities of soil-borne bacillus spp. and pseudomonas spp. against Phytophthora infestans and other potato pathogens. Front Microbiol 9
- Chen J, Guo Y, Lu Y, Wang B, Sun J, Zhang H, Wang H (2019) Chemistry and biology of Siderophores from marine microbes. Mar Drugs 17(10):562
- Das P, Khan S, Chaudhary AK, Abdul Quadir M, Thaher MI, Al-Jabri H (2019) Potential applications of algae-based bio-fertilizer
- Davaran HE, Mirshekari B, Ardakani MR, Farahvash F, Rejali F (2016) Optimizing phosphorus use in sustainable maize cropping via mycorrhizal inoculation. J Plant Nutr 39(9):1348–1356
- Fu FX, Yu E, Garcia NS, Gale J, Luo Y, Webb EA, Hutchins DA (2014) Differing responses of marine N2 fixers to warming and consequences for future diazotroph community structure. Aquat Microb Ecol 72(1):33–46
- Goodfellow I, Bengio Y, Courville A (2016) Deep learning. MIT Press. http://www. deeplearningbook
- Grobelak A, Napora A, Kacprzak M (2015) Using plant growth-promoting rhizobacteria (PGPR) to improve plant growth. Ecol Eng 84:22–28
- Hafeez FY, Yasmin S, Ariani D, ur-Rahman M, Zafar Y, Malik KA (2006) Plant growth-promoting bacteria as biofertilizer. Agron Sustain Dev 26(2):143–150
- Hedrich S, Schlomann M, Johnson DB (2011) The iron-oxidizing proteobacteria. Microbiology Reading 157:1551–1564
- Janice V, Carmen SFB (2007) Growth and development of Mentha piperita L. in nutrient solution as affected by rates of potassium. Braz Arch Biol Technol 50(3):379–384
- Jashni R, Fateh E, Aynehband A (2017) Effect of Thiobacillus and Nitrocara biological fertilizers and foliar application of zinc and iron on some qualitative characteristic and remobilization of rapeseed (Brassica napus L.). Journal of Plant Productions (Agronomy, Breeding and Horticulture) 40(1):1–14
- Kalayu G (2019) Phosphate solubilizing microorganisms: promising approach as biofertilizers. Int J Agronomy 2019:1–7
- Koide RT, Mosse B (2004) A history of research on arbuscular mycorrhiza. Mycorrhiza 14(3):145
- Krasikov V, Aguirre WE, Dekker HL, Huisman J, Matthijs HCP (2012) Time-series resolution of gradual nitrogen starvation and its impact on photosynthesis in the cyanobacterium Synechocystis PCC 6803. Physiol Plant 145(3):426–439
- Larrañaga P, Calvo B, Santana R, Bielza C, Galdiano J, Inza I, Lozano JA, Armañanzas R, Santafé G, Pérez A, Robles V (2006) Machine learning in bioinformatics. Brief Bioinform 7(1):86–112
- Lee S-M, Ryu C-M (2021) Algae as new kids in the beneficial plant microbiome. Front Plant Sci 12: 599742
- Lin S, Hao T, Guo G, Chen G, Mackey HR, Hao T, van Loosdrecht MCM, Chen G (2018) Biological sulfur oxidation in wastewater treatment: A review of emerging opportunities. Water Res 143:399–415
- Martinez JS, Carter-Franklin JN, Mann EL, Martin JD, Haygood MG, Butler A (2003) Structure and membrane affinity of a suite of amphiphilic siderophores produced by a marine bacterium. Proc Natl Acad Sci U S A 100(7):3754–3759

- Mishra DJ, Singh R, Mishra UK, Shahi SK (2012) Role of bio-fertilizer in organic agriculture: A review. Res J Recent Sci 2:39–41
- Mitter EK, Tosi M, Obregón D, Dunfield KE, Germida JJ (2021) Rethinking crop nutrition in times of modern microbiology: innovative biofertilizer technologies. Front Sustainable Food Sys 5
- Pajares S, Ramos R, Ramos R (2019) Processes and microorganisms involved in the marine nitrogen cycle: knowledge and gaps. Front Marine Sci 6
- Pankievicz VCS, Do AFP, Ané JM, Stacey G (2021) Diazotrophic bacteria and their mechanisms to interact and benefit cereals. Molecular Plant-Microbe Interactions: Mpmi 34(5):491–498
- Pfennig N (1975) The phototrophic bacteria and their role in the sulfur cycle. Plant Soil 43:1-16
- Quiroz-Arita C, Sheehan JJ, Baral NR, Hughes A, Peers G, Hodgson B, Sharvelle S, Bradley TH (2019) A cyanobacterial Sidestream nutrient removal process and its life cycle implications. Bioenergy Res 12(1):217–228
- Rane MR, Sarode PD, Chaudhari BL, Chincholkar SB (2008) Exploring antagonistic metabolites of established biocontrol agent of marine origin. Appl Biochem Biotechnol 151(2–3):665–675
- Ravikumar S, Kathiresan K, Ignatiammal STM, Babu SM, Shanthy S (2005) Nitrogen-fixing azotobacters from mangrove habitat and their utility as marine biofertilizers. J Exp Mar Biol Ecol 312(1):5
- Ritika B, Utpal D (2014) Biofertilizer, a way towards organic agriculture: a review. Afr J Microbiol Res 8(24):2332–2343
- Saha B, Saha S, Das A, Bhattacharyya PK, Basak N, Sinha AK, Poddar P (2017) Biological nitrogen fixation for sustainable agriculture
- Schmidt H, Haccius M (2020) Analysis of regulation (EU) 2018/848 on organic food: new rules for farming, processing, and organic controls. EFFL 15(1):2–17
- Singh SP, Singh MK (2019) Mycorrhiza in sustainable crop production
- Spiertz JHJ (2009) Nitrogen. A Review, Sustainable Agriculture and Food Security
- Swapna LA (2013) Development of bio-fertilizers and its future perspective. Sch Acad J Pharm 2(4):327–332
- Tabita FR (1999) Microbial ribulose 1,5-bisphosphate carboxylase/oxygenase: A different perspective. Photosynth Res 60(1):1–28
- Tang K, Baskaran V, Nemati M (2009) Bacteria of the Sulphur cycle: an overview of microbiology, biokinetics and their role in petroleum and mining industries. Biochem Eng J 44(1):73–94
- Teotia P, Kumar V, Kumar M, Shrivastava N, Varma A (2016) Rhizosphere microbes: potassium Solubilization and crop productivity–present and future aspects
- Vasanthi N, Raj SA, Saleena LM (2018) Silica Solubilization potential of certain bacterial species in the presence of different silicate minerals. SILICON 10(2):267–275
- Visser JM (1997) Sulfur compound oxidation and sulfur production by Thiobacillus sp. W5. Antonie van Leeuwenhoek 72(2):127–134
- White AE, Granger J, Selden C, Gradoville MR, Potts L, Bourbonnais A, Fulweiler RW, Knapp AN, Mohr W, Moisander PH, Tobias CR, Caffin M, Wilson ST, Benavides M, Bonnet S, Mulholland MR, Chang BX (2020) A critical review of the <sup>15</sup>N<sub>2</sub> tracer method to measure diazotrophic production in pelagic ecosystems, vol 18. Limnol Oceanogr Methods, pp 129–147
- Zeffa DM, Perini LJ, Silva MB, Scapim CA, Oliveira ALM, Amaral JAT, Pardha-Saradhi P (2019) Azospirillum brasilense promotes increases in growth and nitrogen use efficiency of maize genotypes. PLoS One 14:4

# **Chapter 10 Marine Sponges for Bioremediation Purposes and for Secondary Metabolites Production**



205

Ismail Marzuki 🗈 and Khairun Nisaa

Abstract Environmental bioremediation is necessary to maintain the balance of the ecosystem to remain friendly and supports the continuity of life in the future. Comprehensive screening of marine sponges, symbiotic bacteria and secondary metabolites produced has been carried out. The activity begins with the identification and characterization of the morphology and histology of sponges. Furthermore, the analysis of phenotype and genotype of symbiotic bacteria continued by exploring the function of several types of bacteria in the biodegradation method of PAHs and bio-adsorption for several kinds of heavy metals. These activities include analyzing secondary metabolite components produced by sponges with specific characteristics and specific behaviour of enzymes in enzymatic reaction mechanisms in several environmental improvement uses. Based on the screening results, it is known that there are 11 types of marine sponges from Kodingareng Keke Island, which are in symbiosis with eight varieties of bacteria from the Bacillus sp., Pseudomonas sp. and Acinetobacter sp. groups. These bacteria can biodegrade PAHs, especially against petroleum sludge, naphthalene and pyrene. Also found 12 types of symbiotic sponge bacteria with the ability to bio-adsorb heavy metals, especially Cr (III), Cr (VI), Mn (II), Mn (VII), Pb, Hg, As, Cu and Ni. Adsorption varies. The interesting part of the results of the bacterial symbiotic test was that three types of symbiotic sponge bacteria were found, which have dual functions as biodegradators of PAHs and bio-adsorbents of heavy metals. The sponges included Acinetobacter calcoaceticus strain PHCDB14, Bacillus pumilus strain GLB197 and Pseudomonas stutzeri strain SLG510A3–8. Therefore, this type of sea sponge is recommended for population propagation through the transplant method.

I. Marzuki (🖂)

K. Nisaa

Department of Chemical Engineering, Fajar University, Makassar, South Sulawesi, Indonesia e-mail: ismailmz@unifa.ac.id

Research Center for Fishery National Research and Innovation Agency, Cibinong, West Java, Indonesia

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_10

Keywords Bioremediation  $\cdot$  Biodegradation  $\cdot$  Bio-adsorption  $\cdot$  Sponge microsymbiont

#### **10.1** Introduction

Exploration of marine microorganisms to overcome environmental pollution is essential to maintain and maintain the existence of the earth as the best and most comfortable dwelling for every living creature, more specifically in the marine environment, which is an area where various types of ecosystems of life and activities of animals to maintain themselves exist, on earth. It is realized that the actions and dynamics of human life contribute the most in producing various environmental problems today. Environmental pollution in multiple parts of the earth, the largest, is caused by human activities in maintaining their lives, directly and indirectly harming environmental sustainability (Dadrasnia et al. 2020). Human initiatives and efforts to save the environment are not merely desires that are free from interests and without purpose but are indirect efforts to maintain their existence on earth as living beings who want to exist and live for all time (Idah et al. 2018).

Human efforts to save the environment are carried out in various ways. These methods are, for example, by conducting research, experiments and trials both on a micro-scale and on a broad scale, whether carried out in the laboratory or directly carried out in the field. These activities become a necessity of life that humans must carry out because humans are most responsible and have an interest in maintaining the environment to remain a friendly place. Seeing that the living environment of living beings is not simple, comprehensive studies and studies are needed so that the environment remains sustainable throughout time so that the natural balance and dynamics of the ecosystem continue to run naturally (Marzuki et al. 2020a). Actual examples of human activities that contribute to the contaminant component for the environment can be seen in the petroleum mining industry, which produces various types of fossil fuels to meet the needs of life (Lu et al. 2019). There are three main activity stages of petroleum production, namely the stages of production through the processing industry, distribution and use of the product. The processing stage turns out that petroleum exploitation activities always produce by-products in the form of oil sludge or sludge. The distribution stage, which generally uses sea transportation in the form of tankers and distribution pipes, has the potential for incidents of distribution pipe leaks, tanker fires, collisions, sinking ships and other potential incidents that result in oil spills and pollute the environment (Medic et al. 2020). The stage of using this fuel to run production machines, cars, motorcycles and other work equipment that uses fossil fuels produces various types of combustion gases, all of which put a burden on the earth or the potential for environmental pollution (Rua et al. 2018).

Petroleum sludge is composed of two main components, namely hydrocarbon components and heavy metals. Hydrocarbon components contain several sub-components: total solid residue, fixed residue, volatile solid residue and sludge moisture content. These components are suspected of having hazardous and toxic materials such as polycyclic aromatic hydrocarbons (PAHs), both simple ones such as benzene to heavy ones such as benzo(a)pyrene (Marzuki et al. 2021c). Several types of heavy metals are often found in petroleum sludge, such as lead (Pb), nickel (Ni), copper (Cu), arsenic (As), chromium (Cr), aluminium (Al) and other types of heavy metals. Based on the characteristics of the components of PAHs and heavy metals that make up petroleum sludge, we can state that these components are a severe threat to the air, water and soil environment, so they must be watched out for and prevented from contaminating the environment (Marzuki et al. 2020b; Lajayer et al. 2019). We can find pAHs components in the form of gaseous residues combined with air, liquid, solid and semi-solid. Heavy metal components also include toxic elements to plants, animals and humans (Marzuki et al. 2020c). Generally in the form of particulates that can mix with air, associate with water and soil. Observing the characteristics of PAHs and heavy metals that make up petroleum sludge, We can say that these components are a severe threat to the air, water and soil environment, so they must be watched out for and prevented from contaminating the environment (Yogaswara 2017).

The sea area is a giant container, is the last shelter of the processes that occur in nature. The ocean is also a place for recovery to take home to a balanced state that occurs naturally due to shifts in a balance due to the dynamics of living things that appear on the earth's surface. Pollution that occurs in the atmosphere by heavy metal particulates and other contaminants will eventually end up in the oceans. The same is true for soil contamination by PAHs, heavy metals, microplastics and other components. Pollutants over time, the occurrence of rain that flows following low areas and finally empties into the sea will eventually empty into the ocean (Ziarati et al. 2019a, 2019b; Tereza et al. 2018).

The sea has protected itself to recover the presence of foreign contaminants through natural processes in which dynamics occur, traversed by adaptation between components until the balance is achieved. The problems that exist are contaminants that enter the sea body with the characteristics: (1) large volume, (2) various types, (3) toxic, (4) varying or even non-biodegradable decomposition time. The characteristics of these contaminants are variables that contribute dominantly to the recovery that occurs in the sea. These four variables tend to increase over time so that at a particular time, they pass the tolerance threshold for recovery that happens at sea. This condition causes the capacity and ability, performance and degradability of materials in the sea to decrease. The range of balance that can provide continuity of life is getting wider (Marzuki et al. 2021d).

The effect of hazardous materials in an area creates a chain pressure on various aspects of life, not only on the habitat of a population but also harms environmental productivity in providing support for long-term survival. Of course, this condition cannot be left alone. Herefore, we need solutions and concrete steps to reduce the volume and types of contaminants. In addition, we also need efforts to increase the carrying capacity of the environment for the lives of living things, especially humans, so that they remain in a balanced position between the needs of humans and living things with the availability and production of all conditions that can be provided by the environment (Parhamfar et al. 2020). Environmental management is

determined by technological advances and environmental engineering, either by physical, chemical or biological methods. Aspects of utilizing the wealth of natural materials, both plants and animals, including microorganisms, have a decisive role in environmental management (Orani et al. 2018).

Various types of living things, including plants, animals and microorganisms, can potentially use pollution management and environmental management, which are available and widely distributed both inland areas and in the aquatic environment. Several groups of microorganisms such as fungi, fungi and bacteria are commonly used in environmental improvements to overcome and control the toxic nature of pollutant contaminants. The hydrocarbon, heavy metal and microplastic pollutant components are most often found accumulating in waters, especially the sea. In the marine environment, it is also found that many types of microorganisms have the potential to be used as materials in improving environmental quality (de Kluijver et al. 2021). Bioremediation methods to enhance the quality of the environment contaminated with toxic components using microorganisms have been widely applied. These activities are carried out both on a small scale in the laboratory, as well as on a medium scale and continuously by the company internally, especially in managing the waste produced, so that it meets the requirements for disposal to the accessible environment (Knobloch et al. 2018).

Marine sponge is a marine biota used as a biomonitoring material to determine heavy metal pollution in the marine environment. Determination of the pollution level is carried out by slicing certain body parts of the sponge as a sample. Then preparation is carried out so that the type and content of heavy metals in the sponge can be measured using ICP and or SSA instruments (Akinde and Iwuozor 2012). The measurement data obtained can be used as a basis for determining the level of heavy metal contamination in the waters of the sponge habitat. This example shows that sea sponges are bioindicators of heavy metal pollution. The results showed that several types of sponges could survive in aquatic environments contaminated with heavy metals, presumably due to the ability of these sponges to symbiotic with certain types of bacteria and force them to produce substances that behave enzymes. The substance is then spread on the surface of the sponge body as a mask to protect itself from the stress of heavy metal toxicity (Marzuki et al. 2020b).

Further exploration of sponges, especially their way of life, growth, breeding, self-defence against predators, and their adaptability to the presence of contaminants, is an interesting study to analyze because it holds many secrets of knowledge that need to be learned to be solved. This study also includes a unique feature of sponges: they can associate and symbiotically with several types of microorganisms. In addition, sponges also can nourish the hydrocarbon components. Sponges also play a role in forming parts of metabolically active substances by linking to the ability of environmental bioremediation through biodegradation of hydrocarbon components and bio-adsorption of heavy metals (Schmittmann et al. 2020).

# **10.2** Potential and Contribution of Marine Sponges in Environmental Bioremediation

Sponges are one of the marine biotas widely used as biomaterials to evaluate and analyze the presence of harmful contaminants in the marine environment. The role of sponges as biomonitoring and bioindicators of the quality of the marine environment is interesting to be studied more systematically. Many aspects need to be analyzed against marine sponges, starting from the way of life, nutrition, growth and symbiotic ability with microorganisms concerning the function of biomonitoring and bioindicators of pollutants and the level of pollution that occurs in the marine environment. A lot of evidence shows that some types of marine sponges can survive and even breed in environments contaminated with toxic wastes, such as hydrocarbon compounds, especially PAHs, heavy metals and microplastics (Marzuki et al. 2021a). The phenomenon of adaptation of sponges to the environment polluted with toxic components gives several assumptions: first, the body of the sponge can carry out the function of detoxifying contaminants; second, sponges can produce mucus substances that are spread on the body surface to protect the penetration of toxic materials; third, marine sponges Auletta sp., Clathria (Thalysias) reinwardtii, Callyspongia aerizusa are biomonitoring and bioindicators of heavy metal pollution levels (Marzuki et al. 2021b); Fourth, several types of marine sponge symbiotic bacteria of the Bacillus group (Bacillus licheniformis strain ATCC 9789, Bacillus sp. partial AB353F) and Pseudomonas (Pseudomonas stutzeri RCH2, Pseudomonas stutzeri strain SLG510A3-8) were able to degrade hydrocarbon components (Marzuki et al. 2020a; Orani et al., 2020a; Orani et al. 2018).

The world's population of sponges is estimated at 15,000 species scattered in seas and lakes, but only 46.67% or  $\pm$ 7000 species have been reported, and only 830 species have been isolated and characterized. Sponges are living organisms with a reasonably old civilization that has existed since  $\pm$ 600 million years ago. Sponges can associate with many different microorganisms, including cyanobacteria, heterotrophic bacteria and unicellular algae (Campana et al. 2021a). Sponges are primitive multicellular animals (metazoans) without natural tissues, unique ways of life, capturing food by filter feeders. Sponges are sponges that reproduce both asexually and sexually. Asexual reproduction occurs by the formation of internal buds or gemmules (Maldonado et al. 2021). Sponges can produce and contain more active compounds than compounds produced by land plants. The facts attached to sponges are an essential and logical argument as multi-functional animals in the role of biota upholding the balance of the ecosystem (Schuster et al. 2018).

Bioindicators of heavy metal pollution in the sea can be determined by analyzing the growth and development of sponges (Siahaya et al. 2013). Several research results show that marine sponges contaminated with several heavy metals do not experience growth and development disorders (Melawaty et al. 2014). These conditions indicate that marine sponges can adapt to the presence of heavy metal contaminants in their habitat. The method of determining heavy metal contaminants in the marine environment is carried out by analyzing the types of heavy metals in the sponge's

body. Meanwhile, the concentration level of heavy metal contamination in the marine environment is carried out using appropriate instruments, for example, analysis using Atomic Absorption Spectrophotometer (SSA). It can also be carried out using Inductively Coupled Plasma, both optical emission type (ICP-OES) and mass spectrometry type (ICP-MS) (Ulli et al. 2016).

Sea sponge, one of the biomaterials for monitoring heavy metal pollution in the marine environment, is an essential contributor to marine natural materials in maintaining the balance and sustainability of the growth and development of ecosystems in the sea. Sponges also act as biodegradators of contaminants containing hydrocarbon components. Sponges also act as biodegradators of contaminants containing hydrocarbon components. Symbiotic bacteria of marine sponges play a role in the biodegradation of polycyclic aromatic hydrocarbons (PAHs) (Marzuki et al. 2021c, d). Pyrene, anthracene, phenanthrene and naphthalene are PAHs that symbiotic sponge bacteria can degrade. Types of PAHs-degrading sponge symbiont bacteria include Bacillus licheniformis strain ATCC 9789, a sponge symbiont of Auletta Sp; Acinetobacter calcoaceticus strain PHCDB14, symbiont Callyspongia aerizusa; Bacillus Sp isolated from Neopetrosia Sp; Bacillus pumilus strain GLB197 isolate sponge Niphates Sp; and Pseudomonas stutzeri strain SLG510A3-8 isolate the sponge Hyrtios erectus (Marzuki et al. 2020b). Bacteria Bacillus sp. AB353f partial symbiosis of Neopetrosia sp. Bacillus cohnii strain DSM 6307 symbiont Niphates sp. Several types of sponges suspected to have potential symbiotic bacteria in the biodegradation of PAHs based on the results of identification and morphological characterization include Petrosia (Strongylo Phora) corticata, Clathria (Thalysias) reinwardtii, Callyspongia sp., Coelocarteria singaporensis, Callyspongia (Cladocalina) vaginalis and Callyspongia (Cladocalina) vaginalis. Marzuki et al. 2020a). It was further explained that this type of sponge was thought to have potential symbiotic bacteria through histology of the sponge and analysis of the phenotype and genotype of the symbiotic bacteria. Researchers obtained all the types of sponges mentioned above, about 11 species from three different islands, namely Kodingareng Keke Island, Barrang Caddi Island and Langkawi Island, the administrative area of the Makassar City Government, South Sulawesi Province, Indonesia. These islands are included in the cluster of the Spermonde Archipelago (Marzuki et al. 2021d: Knobloch et al. 2018).

Sponges have an external morphological structure influenced by several general factors, both physical, chemical and biological habitats. Sponges that grow in different habitats have varying growth structures. Types of sponges that live in less stable, shallow, turbid and high waves water environments tend to have creeping growth and are generally shorter. In contrast, the same type of sponges that live in a more stable environment, such as protected growth areas, calm currents and deep waters, tend to grow taller, upright, more symmetrical and have a more extensive body posture (Marzuki et al. 2016; Laport et al. 2009).

The presence of predators, polluting contaminants and competition of sponges with other biota is thought to affect the morphology of sponge growth. The presence of predators such as echinoderms, prosobranchia, opisthobranchia and other types of predators affects the morphology of sponge growth, even forcing sponges to evolve
body structure as a form of adaptation to avoid these predators. Several types of sponges have body structures to drill as a form of disguise and transformation to predatory threats (Yang et al. 2019).

Sponges that live in marine waters that contain coral and are overgrown with algae trigger a competition, where the sponge has a high chance of winning the match if all three are in a relatively deep environment and lack light. Still, the body structure of the growth of such sponges is generally angular (Costa et al. 2020). Sponges that live in marine waters contaminated with hydrocarbons, both aliphatic and aromatic groups, still have a high potential to survive and grow and develop because of their high adaptability to adapt through two mechanisms. First, the nutritional ability of sponges by eating matter. Organic suspended in water flow in body cavities and is sprayed back out. Second, Sponges can have a symbiotic relationship with certain microorganisms, especially bacteria that can selectively and specifically produce enzymes that behave in the form of mucus spread on the sponge body's surface. The function of the mucus is to protect and detoxify the toxic properties of PAHs (Kamaruddin et al. 2021). Sea sponges are also thought to have the ability to survive in areas contaminated with microplastics and produce mucus that can re-glue cracked concrete. Some of the benefits of this marine biota are that sponges deserve the nickname multi-functional biota and have been named green chemistry biota (Obire et al. 2020; Parama Cita et al. 2017).

The presence of predators, polluting contaminants and competition of sponges with other biota is thought to affect the morphology of sponge growth. The presence of predators such as echinoderms, prosobranchia, opisthobranchia and other types of predators affects the morphology of sponge growth, even forcing sponges to evolve body structure as a form of adaptation to avoid these predators. Several types of sponges have body structures to drill as a form of disguise and transformation to predatory threats (Yang et al. 2019).

The internal structure of the marine sponge body at the cellular level is found in several parts such as oscula, surface granules, skeleton and spicules with varying skeletons from each type of sponge. A variety of skeletons are influenced by the growth environment and dynamics of life experienced by the sponge in its growth period. The growth dynamics of sea sponges are influenced by their growth habitat conditions, especially currents, depth, wave height, exposure to sunlight, nutrients, predators and contaminants. These factors also affect the structure and anatomy of cells. The histology of marine sponges based on the catalogue (Krishnamoorthy et al. 1983) is as per Fig. 10.1a–f, for the marine sponge *Niphates sp.*, belongs to the family Niphatidae (Duchassaing de Fonbressin and Michelotti 1864).

The interaction mechanism of sponges with bacteria provides a place or sponge host for certain bacteria to carry out cell growth and division activities. Then there is interaction and adaptation to the new environment. If there are interfering components, then the host organism is stimulated to produce active substances or synthesize bioactive compounds as secondary metabolites. The active substance is delivered for self-protection and maintains environmental balance (Campana et al. 2021a, 2021b). The completion and arrangement of substances or chemical components of the active substance are natural. The type of active substance formed is



**Fig. 10.1** (a) Growth from Sligthy globular sponge, with big size oscula. (b) Consistency Slippery surface sponge, covered by mullike slime. Inelastic and brittle body sponge. (c) Surface granular sponge surface. (d) Skeleton Spicule skeleton with echinating spicule. (e) Skeleton tract Paucispicular tract skeleton with high fibre. (f) Spicule Small megasclera oxea (magnification 40x)



Fig. 10.1 (continued)

influenced by the kind of food and the interfering components in the sponge growth process. Generally, sponge nutrition is rich in new microorganisms with potential pharmacological activity so that sponges can carry out the growth process well, even in extreme environments. On the other hand, microorganisms or bacteria benefit from living on a sponge, i.e. bacteria are protected from the pressure of waves and currents. The interaction between sponges and bacteria occurs as a form of commensalism symbiosis, in which bioactive compounds are produced (Marzuki et al. 2020a).

Sponge symbiont bacteria have a significant influence on the formation of bioactive substances. This influence can be seen in the role of bacteria as the leading supplier of energy needed by sponges. Screening of symbiotic sponge bacteria was carried out by inoculating bacteria associated with the sponge on NA media overgrown with the test bacteria. This qualitative method can only determine whether the bacteria can inhibit the test bacteria E. coli and S. Aureus without knowing the effectiveness of these bacteria (Gunathilake 2018). It is estimated that 40% of the biomass of some types of sponges is composed of bacterial communities. Cyanobacteria symbiosis can make up one-third of the total mass of living tissue in some types of sponges. The energy needs of some types of sponges are supplied from 48% to 80% of microorganisms (Knobloch et al. 2018).

The bioremediation function of marine sponges on the environment is thought to be played by the active substances in the sponge's body. Sponges produce these chemical components to defend themselves against potential growth disorders. The characters, types and specific properties of these secondary metabolites are formed according to the dynamics of life experienced by sponges during their growth period. These metabolic substances are thought to have several functions and benefits, both in utilization studies in the medical field and environmental bioremediation processes (Obire et al. 2020). Its environmental remediation function is primarily for the application and function of biodegradation of PAHs and heavy metal bio-absorption, including the potential to overcome the presence of microplastics in the environment (Marzuki et al. 2021b). The researcher can achieve knowledge of the role and function of these sponges in the biodegradation of PAHs, bio-adsorption of heavy metals and the absorption potential of microplastic components by conducting a series of studies and analyses. The assessment and analysis activity begins with observing the sponge's growth environment, screening the morphology and histology of potential sponges for environmental remediation functions. The next activity is the isolation of symbiotic bacteria, characterization through phenotypic and genotypic analysis of symbiotic bacteria, to conducting micro-scale tests and field experiments to determine the performance of symbiotic sponge bacteria in the biodegradation of PAHs and bio-adsorption of heavy metals (Marzuki 2020; Rua et al. 2018).

## **10.3** Search for Potential Sponges for Bioremediation Functions in Polluted Environments

The method of determining potential sponges for the biodegradation function of PAHs was carried out by isolating potential sponge symbiotic microorganisms. The purpose of potential sponges is to select specific types of sponges that meet the following criteria: first, selecting sponges that are suspected of living in areas

contaminated with hydrocarbons; second, the surface of the sponge body is covered by mucus or enzyme behaviour substances or at least dark-coloured sponges; third. The colour of the sponge is generally less bright. The selected sponges were then isolated to obtain bacterial isolates. Isolation of marine sponge symbiotic bacteria using a simple method, as shown in Experiment 10.1.

#### **Experiment 10.1: Isolation of Marine Sponge Symbiotic Bacteria**

The selected potential marine sponges were cleaned by spraying the surface using sterile seawater with a ratio of 1 cm2: 5 mL of sterile seawater. The mesohyl sections were  $\pm 1 \times 1$  cm in size, crushed and diluted with sterile Phosphate Buffer Saline (PBS) in a ratio of 1:1. Isolate the bacteria on the outer surface using a sterile swab, then wipe in one direction on the outer surface of the sponge. The sterile swab was put into a dilution tube containing sterile PBS and vortexed. The results of the dilution were spread into a petri dish that already contained Seawater Complete (SWC) media, incubated at 26°C for 24–36 h, observed colony growth, bacterial morphology. Selected colonies, separated using a round needle, were purified using the same medium to obtain pure isolates. Purifying bacterial symbionts using the direct plating method was performed by scratching one ose of colonies in a zig-zag direction on a petri dish containing 100% marine-agar media. Incubation temperature 30°C, 1–2 days. Re-scratching on 100% marine agar until a single colony was obtained (Marzuki et al. 2021a).

Marine sponge symbiotic bacteria biodegraded PAHs contaminants by integrating one type (Naphthalene) with a potential bacterial suspension. The complete procedure for biodegradation of marine sponge symbiont bacteria against PAHs is presented according to the method in Experiment 10.2.

## Experiment 10.2: Biodegradation of PAHs Using Marine Sponge Symbiotic Bacteria

Determination of potential bacteria is done through preliminary tests. Bacteria that pass the initial test are bacteria that show growth activity on hydrocarboncontaminated media. Propagation of selected bacterial cells by culture. The bacterial suspension was made and determined the number of cells. Entered 25 mL of bacterial suspension in several degradation reactors (vials), adapted for 1 × 24 h, put 5 mL of contaminant PAHs (naphthalene) with a concentration of 1000 ppm, shaker incubator 200 rpm. Observations and measurements of media degradation parameters were carried out every 2 days for 30 days. Determination of the ability and performance of biodegradation of symbiotic bacteria were analyzed using Gas Chromatography–Mass Spectroscopy (GC–MS) instruments. Biodegradation products were analyzed using Fourier Transform – Infrared Spectroscopy (FT–IR) for serial interaction periods, such as 3, 6, 9, 12, 15, 18, 21, 24, 27 and 30 (Lu et al. 2019).

Parameters of PAHs biodegradation by symbiotic sponge bacteria consisted of optical density, pH, temperature, an abundance of gas bubbles and fermentation odour. Optical density was determined using a UV-Vis spectrometer on a mask. For each serial interaction period in multiples of 2 days. Suppose an increase in the optical density (OD) value for the first few days of interaction. In that case, it



Fig. 10.2 The rate of increase in the value of OD (bacterial cell growth rate) based on the interaction time in days between a sponge symbiotic bacterial suspension and 10,000 ppm naph-thalene. Source: Marzuki et al. 2021d)

indicates a bacterial activity in the degradation medium contaminated with PAHs until it enters the saturation period marked by no increase in the OD value in the next few days. The rate of increase in OD value can be identified with the growth rate of bacterial isolates isolated from four different types of marine sponges, namely *Hyrtios erectus* (SpA.B2), *Clathria (Thalysias) reinwardtii*) (SpB.B2), *Niphates sp.* (SpC.B2), and *Callyspongia sp.* (SpD.B2), respectively, interacting with 10,000 ppm naphthalene and 10,000 pyrenes for 15 days (Marzuki et al. 2021d), according to Figs. 10.2 and 10.3.



**Fig. 10.3** The rate of increase in the value of OD (bacterial cell growth rate) based on the interaction time in days between a sponge symbiotic bacterial suspension and 10,000 ppm pyrene. Source: Marzuki et al. 2021d)

The increase in OD value in the reactor containing PAHs contaminants is directly proportional to interaction time. The rise in OD value indicated an activity of isolates of sponge symbiont bacteria containing PAH. These conditions showed that the bacteria were able to adapt to media containing PAHs contaminants, although the OD values of each isolate were different. The sequence of growth of symbiotic sponge bacterial cells in naphthalene-contaminated media (Fig. 10.2), SpC.B2 SpD. B2 SpB.B2 SpA.B2, while the growth rate of bacterial cells in pyrene-contaminated media (Fig. 10.3) in the order SpD.B2 SpC.B2 SpB.B2 SpA.B2.

Changes in the pH of the degradation medium towards a lower value or tend to be more acidic indicate that there is bacterial biodegradation activity on the hydrocarbon component substrate. The temperature of the degradation media tends to increase generally in the range of  $\pm 0.5$ –2.3 0C after the interaction lasts for several days if there is biodegradation activity in the reactor (Bendouz et al. 2017).

The gas bubbles in the reactor are relatively increased as a sign that there is biodegradation activity. The abundance of gas bubbles formed is directly proportional to the level of biodegradation. Gas bubbles are formed and generally increase after the interaction runs for a few days. The abundance decreases as the biodegradation process in the reactor weaken, indicating a decline in the number of bacterial cells that carry out the degradation process (Marzuki et al. 2020a). Gas bubbles formed as a sign that there are simple organic compounds in the reactor in methane gas,  $CO_2$ ,  $NO_2$  and other gases resulting from biodegradation or decomposition/ rehabilitation of PAHs components. The smell of fermentation is also part of the biodegradation parameter as an indicator. The degradation that occurs results from the work of bacteria through the mechanism of degradation of enzymatic reactions produced by biodegradative bacteria (Onwosi et al. 2017).

The speciality of bacteria in the PAHs degradation method is the ability of bacteria to destroy the hydrocarbon structure and turn it into an energy source for the activity and survival of bacteria. The biodegradation performance of symbiotic sponge bacteria against PAHs was determined using GC–MS. The data obtained on the chromatogram show the components of the biodegradation product (Marzuki et al. 2020c). The chromatogram data is in the form of component abundance peaks and the potential for new peaks to appear. In general, the abundance of the tested PAHs components as substrates will decrease with the emergence of new peaks. The higher the performance of bacterial biodegradation, the lower the peak height (abundance) of the PAHs test components. The lower the PAHs, the lower the concentration of the degradation components. Figure 10.4 shows an example of a GC–MS chromatogram resulting from the biodegradation of Bacillus SP strain AB353f in symbiosis with the Neopetrosia Sp sponge against naphthalene compounds.

The biodegradation process of PAHs (naphthalene) substrate at 1-day interaction was not seen (Fig. 10.4a). This biodegradation process began to appear after 10 days of interaction between Bacillus sp. strain AB353f and naphthalene (PAHs substrate). An indication between Bacillus sp. strain AB353f and PAHs substrate is indicated by the decreasing peak height of naphthalene in the graph (Fig. 10.4b). Decrease of peak showed that the abundance of the substrate had decreased, meaning that there



**Fig. 10.4** Chromatogram of naphthalene biodegradation of Bacillus SP strain AB353f based on interaction time. (a). Interaction time 1 day; (b) The interaction time is 10 days, and C. the interaction time is 20 days (Marzuki et al. 2020c)

was a decrease in the concentration of PAHs along with the emergence of two new peaks resulting from biodegradation in the form of simple organic compounds. The new peak increased to three after the interaction period lasted 20 days (Fig. 10.4c). marked by the decreasing peak height of naphthalene which means that the biodegradation process continues. The interaction period in this experiment was up to 30 days. Still, it appears that during the interaction period above 20 days, there was no addition of new peaks, and the peak height of PAHs tended to stagnate. This condition indicates that the biodegradation process is no longer running. It could be because the bacterial cells as biodegradators died due to poisoning by the toxic nature of PAHs or because the bacterial cells could not withstand the increase in the acidic properties of the media in the reactor. The increase in the acidic properties of the reactor media occurred, presumably because components were resulting from the biodegradation of simple organic compounds from the carboxylic acid group (Marzuki et al. 2021c). The results of the GC-MS measurement also provide data on the level of biodegradation. After calculating, the result indicated that the biodegradation rate was in the range of 26% - 46% for using one type of symbiotic sponge bacteria against one type of PAHs (Costa et al. 2020).

Several ways can improve the biodegradation performance of bacteria against PAHs. For example, it increases the number of bacterial cells in the reactor, prolonging the life of bacterial cells by providing nutrients in adding nitrogen, phosphorus and potassium in the degradation reactor. In addition, during the interaction period, oxygen is supplied to prevent an increase in the acidity of the degradation media or the use of several types of bacteria (a consortium of bacteria), which can degrade hydrocarbon components. The use of consortium bacteria to increase the level of biodegradation does not necessarily increase the capacity of these bacteria to degrade, presumably due to competition between bacteria in obtaining energy supplies from the PAH components that have been destroyed. However, the biodegradation of PAHs with consortium bacteria is currently being carried out because of the ease of multiplying bacterial cells and can be carried out very quickly (Bendouz et al. 2017).

Many collections of bacteria that can biodegrade PAHs have been carried out, especially bacteria from the Bacillus and Pseudomonas groups. Both groups of these bacteria can be isolated from several sources, such as soil contaminated with PAHs, seawater contaminated with hydrocarbons, sponges and several other marine biotas.

A collection of PAH-degrading bacteria is known as hydrocarbonoclastic bacteria (Lu et al. 2019).

The mechanism of bacterial biodegradation of hydrocarbon components, both aliphatic and aromatic groups, in principle, is through the oxidation pathway to form alcohol. Alcohol products become aldehydes and finally produce carboxylic acids and a small portion of esters before entering the -oxidation cycle and producing energy for biodegradation activity—next cycle. The biodegradation process also produces by-products in simple organic compounds, such as CO2, methane gas, NO and other gases, depending on the degraded hydrocarbon component. The biodegradation of PAHs is relatively the same as aliphatic components (Medic et al. 2020). The striking difference for the aromatic hydrocarbon component at the stage after the oxidation reaction is that it is preceded by the destruction stage of the aromatic molecular structure so that the biodegradation of PAHs components by suitable specific bacteria takes longer. The resulting degradation rate is generally much smaller than in the degradation of aliphatic hydrocarbon components (Fang et al. 2020). A simple description of the biodegradation mechanism of aliphatic components is presented in Fig. 10.5.

Specific degradation pathways of Bacillus subtilis strain BAB-1684 sponge symbiont Callyspongia sp. against the hydrocarbon components of petroleum sludge waste after 20 days of contact are presented in Fig. 10.6.

Based on the illustration of the biodegradation of hydrocarbon components by the symbiotic marine sponge bacteria (Figs. 10.5 and 10.6), some steps allow the biodegradation process to stop completely. The biodegradation process stopped entirely because the bacterial cells experienced mass death. The biodegradation process can stop completely at the stage of changing the biodegradation products of aldehydes or ketones into carboxylic acids. The formation of carboxylic compounds can cause the interaction medium to become acidic so that bacterial cells have the potential to experience sudden death because they cannot survive at a certain acidity level in the degradation medium (Marzuki et al. 2021a; Yogaswara 2017). This stage is known as the reaction rate stop stage, or the biodegradation process stops completely.

The contribution of marine biota, especially sponges, in maintaining the balance of the marine ecosystem is substantial. The contribution of marine life, especially sponges, in maintaining the balance of the marine ecosystem is enormous. Suppose an in-depth review is carried out on the formation of secondary metabolite



Fig. 10.5 Simple mechanism of biodegradation of aliphatic hydrocarbon components



Fig. 10.6 Estimation of a simple path of biodegradation of aliphatic hydrocarbon components of petroleum sludge by the bacterium Bacillus subtilis strain BAB-1684 symbiosis of sponge Callyspongia sp

components of sponges and the effect of these substances on their biodegradability, bio-adsorption and activity of sponge bioactive substances against pathogenic bacteria and fungi. In addition, the active substance of sea sponges is thought to have the potential as primary and secondary raw materials for certain drugs to treat a disease (Shareef et al. 2016). Studies on sponges, symbiotic microorganisms and components of secondary metabolites are interesting to do more comprehensively, including studies related to the competition for the growth of sponges versus other biotas such as corals and algae if all three are in the same area (Schuster et al. 2018).

A simple study of the morphology and histology of marine sponges of the Niphates sp. species obtained from the sea waters around Kodingareng Keke Island has been described in point 10.2 above. Part of the characterization of sponges is to identify and characterize the symbiotic sponge bacteria that are thought to have biodegradation performance against PAHs potentially. Phenotype and genotype characterization of symbiotic sponge bacteria was carried out by taking samples of four isolates of sponge bacteria *Petrosia (Strongylo Phora) corticata* (Sp. 1), Auletta sp. (Sp. 2), *Neopetrosia sp.* (Sp. 3) and *Callyspongia aerizusa* (Sp. 4). (Marzuki et al. 2020a, 2020b, 2020c, Bioflux), are presented in Table 10.1.

The urease test results (Table 10.1) on two isolates (Sp. 1 and 2) showed positive results, meaning that both isolates were able to hydrolyze urea. This ability means that the isolate can produce and possess the enzyme urease. On the other hand, the isolates (Sp. 3 and 4) were negative, meaning that both isolates could not hydrolyze urea or did not have urease enzyme. V-P reagent test isolates (Sp. 1–3) showed a positive reaction, meaning that there were components in the three isolates capable of carrying out the fermentation reaction. In the methyl red test (R-Mr), the four

		Sponge bacterial symbiont			
Biochemical reagents	Media	Sp. 1	Sp. 2	Sp. 3	Sp. 4
Starch hydrolysis	Starch agar	Base	Base	Base	Base
Casein hydrolysis	Milk agar	Acid	Acid	Acid	Acid
Gelatin hydrolysis	Gelatins	-	-	-	-
Nitrate reduction	Nitrate broth	-	-	-	-
Indole	Tryptone broth	-	-	-	-
H <sub>2</sub> S	H <sub>2</sub> S	-	-	-	-
Reagent methyl red	R-Mr broth	+	+	+	+
Reagent- Voges Proskauer	R-VP broth	+	+	+	-
Citrates	Citrate	+	-	+	-
Urease	Urea broth	+	+	-	-
Glucose	Glucose broth	-	-	-	-
Lactose	Lactose broth	+	+	+	+
Sukrose	Sucrose broth	-	-	-	-
Mannitol	Mannitol broth	-	-	-	-
Catalase	Nutrient agar (NA) slant		+	+	-

 Table 10.1
 Phenotype characterization of sponge symbiont bacteria Callyspongia sp, Biochemical Test method

Note: + (reaction); - (no reaction)

Source: Marzuki et al. (2020a)

isolates showed a positive response, indicating that the isolates could produce acid in glucose fermentation (Marzuki et al. 2015). The catalase test on all isolates resulted in a positive reaction (Sp. 1–3 isolates). The isolates had catalase enzymes that could degrade hydrogen peroxide (H2O2), while the fermentation test used glucose reagent did not show any reaction activity. The lactose test showed that the four isolates gave a positive reaction. The sucrose test resulted in a negative response, meaning that the bacteria that grew in the fermented liquid media were acidic components (Bibi et al. 2016). The results of characterization through gram staining and biochemical tests showed that the two microsymbiont isolates of four different types of marine sponges contained enzymes and could ferment and process carbon from their environment. With the six criteria described above, it is suspected that three isolates (Sp. 1, Sp. 2, Sp. 3) have the potential and ability to degrade, especially to aliphatic hydrocarbon components. Still, it is necessary to test and follow up on analysis further to degrade aromatic hydrocarbons (Khabouchi et al. 2020).

The genotype identification of four isolates from four different marine sponges was carried out to see the pair structure and nitrogen base-pair composition through the Basic Local Alignment Search Tool (BLAST), PCR method. The results of the isolate sequencing were opened through the bioedit programme. The sample bacterial DNA sequences were entered into the BLAST (Basic Local Alignment Search Tool) programme (http://blast.ncbi.nlm.nih.gov/Blast.cgi), the sequences were identified with the DNA database. GenBank on the site (Kadhim et al. 2013; Cowden 1976). The results of the alignment of the sample sequences with the GenBank sequences showed a high similarity of the homologous series, which can be seen in the table as presented in Table 10.2.

Symbiont code	Sample Sequence	Sequence Gen Bank	Quantity (%)	Difference (%)	Species
Sp. 1	17-972 (955)	608.723-609.690 (967)	944/955 (98.85)	4/955 (0.42)	Pseudomonas stutzeri RCH2
Sp. 2	11-985 (974)	524.589–525.563 (974)	956/974 (98.15)	14/974 (0.01)	Bacillus licheniformis strain ATCC9789
Sp. 3	15-975 (960)	574.123-575.089 (966)	932/960 (97.49)	16/960 (1.66)	Bacillus sp. AB353F.Partial
Sp. 4	21-934 (913)	574.323-575.258 (935)	906/935 (96.90)	12/935 (1.28)	Acinotobacter calcoaceticus strain PHKDB14
Source: Marzuki e	t al. (2020a)				

CR method
a, PC
bacteria
Sponge
Symbiont
of
results
LAST
BI
10.2
Table

I. Marzuki and K. Nisaa



Fig. 10.7 Representative neighbour-joining tree reconstructed with the 16S rRNA sequences of marine sponge symbiont bacteria (Marzuki et al. 2021d)

The results of genetic analysis using PCR found each sponge symbiotic isolate (Table 10.2). There are two types of bacteria with different strains. Each of which is isolates from sponges Auletta sp. (Sp. 2), Neopetrosia sp. (Sp. 3), one group of Pseudomonas, isolates of sponges Auletta sp. (Sp. 2) and one group of Acinetobacter. Which is an isolate from sponge Callyspongia aerizusa (Sp. 4). Based on the results of biochemical tests (Table 10.1) by observing the reaction results of each test medium combined with the results of the identification of the 16S rRNA gene molecule by PCR method (Table 10.2). It is indicated that the four species can carry out chemical reactions to break down hydrocarbon molecules (Maldonado et al. 2021).

Characterization and phylogenetic identification of symbiotic sponge bacteria according to the reconstructed marine sponge symbiotic bacterial phylogenetic tree against 16S rRNA bacteria, three species were well resolved supported by moderate bootstrap values (Fig. 10.7). The first clade, isolates of sponges SpA.B2, SpB.B2, SpD.B2, live in groups with *Pseudomonas stutzeri* and *Pseudomonas songnenensis* with 97–99% homology. The second clade, consisting of SpA. B1/B2, which is positioned as a child clade of the Pseudomonas species. The third clade, SpC isolates. B1 grouped with Bacillus isolates reached 96%.

## 10.4 Development of the Function of Symbiotic Sponge Bacteria Through Heavy Metal Bio-Adsorption Method

The application of marine sponge symbiotic bacteria in bioremediation of heavy metal-contaminated environments is being developed. Several studies have shown that several types of bacteria can bio-adsorb heavy metals. Heavy metals such as Zn, Pb, and Cd can be absorbed using certain bacteria (Konkolewska et al. 2020); Pb, Ni, Cd and Cr (Alimardan et al. 2016); Cd and Pb (Alaboudi et al. 2018). Even several reports were showing that several types of heavy metals were found in marine sponges, for example, Pb, Cd, Zn, Fe and Cr (Wibowo et al. 2019; Siahaya et al. 2013; Melawaty et al. 2014), where the sponge showed a reasonable growth rate. Pseudomonas bacteria isolated from marine sponges can adsorb heavy metals Cr and Mn (Marzuki 2020).

Almost all types of bacteria that breed in areas exposed to heavy metals can carry out the function of bio-adsorption. The problem that exists for the application of bacteria in the bioremediation of environmental heavy metal contaminants is how to identify and obtain bacteria that are capable and have high performance in the bio-adsorption of heavy metals, including types of heavy metals that can be absorbed. This condition demands an assessment to find out and obtain data on the type of bacteria, the level of absorption and the types of heavy metals that bacteria can adsorb. A data bank on this matter is needed to provide convenience and quick handling of areas exposed to heavy metal, both in the aquatic environment and on land or land, especially ex-mining grounds for land function recovery, for agricultural activities. Alimardan et al. 2016).

Several types of bacteria isolated from sea sponges were stated to be able to absorb several types of heavy metals, including *Bacillus sp.* strain AB353f against heavy metals Cr and Mn (Marzuki et al. 2020b; Marzuki 2020), *Bacillus pumilus* strain GLB197 *and Pseudomonas stutzeri* strain SLG510A3-8 against heavy metals Cr(VI) and Cd(II) (Marzuki 2020). Types of bacterial isolates without known species that are symbiotic sponges have been reported to absorb several types of heavy metals. It is including *Petrosia (Strongylo Phore) Corticata, Neopetrosia sp., Callyspongia aerizusa, Niphates Sp, Hyrtios erectus* and *Auletta sp.* to heavy metal Nickel (Ni), Copper (Cu), Lead (Pb), Chromium (Cr) and Mercury (Hg) (Angela and Marzuki, 2021; Wibowo et al. 2019).

The general method of tracing bacteria has the function of bio-adsorption of heavy metals: first, identification of marine sponges that live in waters exposed to heavy metals. Second, sampling several sponges and isolate the associated bacteria. Third, the isolates obtained were tested for their bio-adsorption ability on certain heavy metals by placing some bacterial colonies on media engineered to be contaminated with several types of heavy metals, incubating and observing the growth activity of the isolates. Fourth, potential isolates from possible test results were identified and phenotype and genotype characterization. Fifth, make stock, catalogue of isolates related to types of heavy metals that can be absorbed (Gebregewergis 2020).

Researchers have carried out trials of stock bacteria that have been characterized against certain metals to see their bio-adsorption ability and performance in an engineered environment contaminated with several heavy metals in known concentrations. During the contact process between bacterial suspensions and heavy metals in an engineering environment, bacterial cell growth was observed by measuring the optical density of the interaction medium based on the contact time in days.

# Experiment 10.3: Bio-adsorption of Heavy Metals Using Marine Sponge Symbiotic Bacteria

The Bio-adsorption ability of marine sponge symbiotic bacteria to heavy metal as a sewage contaminant can be determined by several procedures. First, waste is made contaminated with heavy metals, for example, Cr (III), Cr (VI), Mn (II) and Mn (VII), each with a concentration of 250 ppm and a volume of 1000 mL. Second, determine isolates of marine sponge symbiotic bacteria with bio-adsorption potential. For example, Bacillus licheniformis strain ATCC 9789 (BS) and Acinetobacter calcoaceticus strain PHKD B14 (AC), both types of bacteria was cultured on NA media, then incubated  $1 \times 24$  O'clock. Each culture was suspended in a solution of 1,000 mL of physiological 0.9% NaCl solution. Prepare 10 reactors, each filled with 100 mL of isolate suspension, adapted for  $1 \times 24$  h in a new environment, (3) each reactor is added 10 mL of heavy metal contaminants, such as Cr (III) or Cr (VI). The reactor is in the Shaker incubator at 150 rpm. Observation of bacterial cell growth (optical density) was carried out every 3 days. The Observation used a spectrophotometer and determined the level of bio-adsorption of heavy metals by measuring media absorption using AAS, then plotted in Eqs. (10.1) and (10.2) to determine the capacity and efficiency of bio-adsorption (Marzuki 2020).

$$Q = \frac{C1 - C2}{C1} x V \tag{10.1}$$

$$\% E = \frac{C1 - C2}{C1} \times 100\%; \tag{10.2}$$

Based on Experiment 10.3 that has been carried out, it was obtained an overview of the growth of BC and AC bacterial cells in media contaminated with Cr (III), Cr (VI), Mn (II) and Mn (VII).

The experimental results showed that the growth of BS and AC bacterial cells was more dominant in media exposed to Cr(III) than in media contaminated with Cr (VI) (Fig. 10.8). The growth of BS cells in Cr(III) and Cr(VI) contaminated media was maximum during the first 3 days of contact and relatively low after 6 days of contact onwards (Fig. 10.8a). The growth of AC bacteria cells in media exposed to Cr (III) and Cr (VI) was relatively the same as BS bacteria. Only the first 3 and 6 days of contact in Cr (III) media were much more dominant than in Cr (VI) media which sloped from childhood. First 3 days of contact (Fig. 10.8b). The difference in the growth rate of bacterial cells in these two types of media is influenced by the ability





The treatment was the same, and the types of symbiotic bacteria were also the same. Still, Mn (II) and Mn (VII) replaced the types of heavy metal contaminants, showing that the growth of BS and AC bacteria cells did not change significantly. The growth of BS bacteria in Mn (II) media was more dominant in a wider range than in Mn (VII) media. However, the maximum growth of both types of bacteria occurred during the first 3 days of contact (Fig. 10.9a). While the dominant growth of AC bacteria cells in Mn (II) media happened at a more extended contact period, namely on days 3-6 compared to Mn (VII) media and the following contact period, the growth of AC bacterial cells in Mn (II) and Mn (VII) media was relatively the same and slopes (Fig. 10.9b) (Marzuki 2020).

in Cr(III) and Cr



The bio-adsorption capacity was calculated using Eq. (10.1), while the bio-adsorption efficiency was determined using eq. 2, based on absorption data measured using Atomic Absorption Spectrophotometer (AAS). The bio-adsorption capacity of BS and AC bacteria to heavy metal contaminants Cr (VI) was relatively the same, as was the bio-adsorption efficiency shown by the two types of bacteria (Fig. 10.10a, b). Still, in the same picture, the curved line of bio-adsorption capacity (blue colour) does not coincide with the curve line of bio-adsorption efficiency (orange). These conditions indicate that the bio-adsorption performance of BS and AC bacteria against toxic heavy metals Cr (VI) does not reach 50%; Cr (VI) contaminants were chosen because the toxicity level is higher than Cr (III) (Marzuki 2020).

The bio-adsorption capacity of BS and AC bacteria against heavy metal contaminants Mn (II) was relatively the same, including the bio-adsorption efficiency



Fig. 10.10 (a) Bio-adsorption capacity and efficiency of BS bacteria against Cr(VI) contaminants. (b) AC bacteria bio-adsorption capacity and efficiency against Cr(VI) contaminants

performance shown by the two types of bacteria (Fig. 10.11a, b). What is different from the bio-adsorption version of these two types of symbiotic sponge bacteria is the performance of the bio-adsorption capacity shown against heavy metal contaminants Mn (II) exceeds 50%. The two types of bacteria used as bio-adsorbents for heavy metals are Bacteria Bacillus sp. AB353f partial (BS) is an isolate of the sponge *Neopetrosia sp.*, while the bacterium *Acinetobacter calcoaceticus* strain PHCDB14 (AC) is an isolate of the marine sponge *Callyspongia aerizusa* (Marzuki et al. 2021b).

The performance and level of bio-adsorption of heavy metals by symbiotic bacteria of marine sponges have not shown maximum achievement in environmental remediation. So it is necessary to make efforts and developments to obtain bio-adsorption results of heavy metals bio-adsorption capacity reaching 75–90%. The approach that can be taken to improve the bio-adsorption performance of heavy metals is by tracing the types of bacteria that have high bio-adsorption performance against heavy metals. A consortium of bacterial cells that can bio-adsorb heavy



**Fig. 10.11** (a) Bio-adsorption capacity and efficiency of BS bacteria against Mn(II) contaminant. (b) AC bacteria bio-adsorption capacity and efficiency against Mn(II) contaminants

metals; waste engineering and modelling interactions between bacterial suspensions and heavy metal contaminants, interaction time, bacterial nutrient supply; and other modifications aimed at improving the bio-adsorption performance of heavy metals by bacteria (Orani et al. 2018).

Efforts to improve the bio-adsorption performance of sponge symbiotic bacterial cells against heavy metals, especially the use of consortium bacteria, are currently underway by our research team. But, the researcher cannot publish the results and achievements of the bio-adsorption obtained because they are still in the experimental stage and process. Nevertheless, we offer recommendations for the method that we are currently running and the detailed procedure in Experiment 10.4. An overview of the development of heavy metal bio-adsorption methods using several types of heavy metal bio-adsorbent bacteria for utilization in waste exposed to several kinds of heavy metals aims to improve performance maximal bio-adsorption and bioremediation on various types of heavy metal contaminants in parallel (Sobrinho et al. 2020).

## Experiment 10.4: Application of Marine Sponge Symbiotic Bacteria Consortium Against Heavy Metal Extreme Waste

The heavy metal bio-adsorption procedure using a consortium of sponge symbiotic bacteria isolates was carried out in several stages. (1) Manufacture of artificial waste contaminated with several types of heavy metals (Pb, Hg, As, Cr and Cu). Each type of heavy metal made a solution of each concentration of 1000 ppm as much as 1000 mL. The heavy metal solution is mixed in a sizeable homogenized portion (extreme waste). (2) Determine five types of potential sponge symbiotic bacterial isolates, the cells of each selected isolate were propagated by culture and incubated for 1×24 h. The cultured isolates were suspended in a 1000 mL solution containing physiological 0.9% NaCl. (3) The five types of mixed isolate suspension in one large container, homogenized (consortium bacteria). (4) Prepare ten reactors, each filled with 500 mL of bacterial suspension of the consortium, adapted for 1 x 24 hours in an incubator. (5) Each reactor is put in 100 mL of extreme waste and a shaker incubator at 200 rpm. (6) Observations and measurements of bacterial cell growth were carried out every 2 days, and measurements of the absorption of each type of heavy metal were to determine the bio-adsorption achievement of the consortium bacteria based on the contact period.

Other developments related to the exploration of the dual function of sponge symbiotic bacterial cells in the biodegradation of PAHs as well as the bio-adsorption function of heavy metals can be carried out by carrying out a similar procedure in Experiment 10.4 above, but with differences in the type of waste and the bacterial consortium used. The engineering waste is made to be super extreme. In addition to containing PAHs contaminants (the type and concentration of PAHs are known), the waste also includes several types of heavy metals (the type and concentration of heavy metals are known). This super extreme waste is homogenized (Bertollino 2019). This extreme super waste modification resembles petroleum waste, which contains several types of heavy metals and containing PAHs. The bacterial consortium was made by having  $\pm$  six types of symbiotic sponge bacteria, a combination of three types of bacteria that can biodegrade and three types of bacteria with the ability to bio-adsorb heavy metals. The selected sponge symbiotic bacterial cells were multiplied by the culture method, made a suspension, mixed in one container, homogenized, and adapted first. This dual-function bacterial consortium suspension is ready to have interacted with super extreme waste (Wibowo et al. 2019).

The parameters observed for this method include the optical density of bacterial cell growth, changes in pH values, shifts in interaction temperature values, whether there is the formation of gas bubbles in the interaction medium, and whether there is the formation of a fermentation odour. At the same time, the instrumentation recommended for use is GC–MS, FT–IR, and SSA. GC–MS is an instrument in determining the biodegradation process by looking at the abundance of PAHs, the types of simple organic compounds as biodegradation products. FT–IR is an instrument to assess the kind of biodegradation product, whether the alcohol group is an aldehyde, ketode, carboxylic acid and other organic components. In comparison, SSA is an instrument to determine the bio-adsorption capacity that occurs by

determining the uptake of each heavy metal present in the extreme super waste and compared with the initial concentration (Ziarati et al. 2019b).

Bioremediation methods to change and produce friendly environmental quality are the dream of every citizen on this earth. Still, this dream is sometimes difficult to realize because, directly or indirectly, at the same time, we produce by-products, by-products and wastes that are pressing environment so that the environment is massively under continuous pressure and at its natural balance point (Sabdono and Radjasa 2008). Nature provides biomaterials that have a role and function in environmental bioremediation, one of which is sea sponges that can form a symbiosis with various types of bacteria. Symbiotic sponge bacteria have a biodegradation function against PAH components. It includes a heavy metal bio-adsorption function and a strong suspicion that several types of symbiotic sponge bacteria have dual biodegradation and bio-adsorption functions. A collection of several types of symbiotic marine sponge bacteria that have the same role in both the biodegradation of PAHs and the bio-adsorption of heavy metals is called a bacteria consortium (Parhamfar et al. 2020). The term for the type of bacteria with the ability to biodegrade hydrocarbons is known as carbonoclastic bacteria. The type of bacteria that has the capacity for bio-adsorption of heavy metals is metallo clastic bacteria. The term for the symbiotic bacteria of marine sponges can biodegrade hydrocarbons and bio-adsorption of heavy metals called metallo-carbonoclastic bacteria (Marzuki et al. 2021b).

The role and contribution of marine sponges in environmental bioremediation are evident. Including the content of secondary metabolite components possessed by marine sponges have an essential role in biodegradation and bio-adsorption, including the potential utilization of sponge secondary metabolites as biomaterials in overcoming exposure to bioplastics and primary materials in the manufacture of drugs and various other functional potentials. The potentials inherent in sponges, so that marine sponges are recommended for screening to explore potential and develop utilization for the common good of humans and the environment (Saputra et al. 2019). One of the recommendations for marine sponge population is those development efforts to maintain and increase the sponge population in their habitats. Keeping the population of marine sponges can be carried out through the transplant method. Especially the type of marine sponge has been confirmed to have symbiosis against several bacteria in bioremediation and sponges that have been identified as capable of producing chemical components bioactive (Bibi et al. 2016).

## References

- Akinde SB, Iwuozor CC (2012) Alkane degradative potentials of bacteria isolated from the deep Atlantic Ocean of the Gulf of Guinea. J Bioremed Biodegr 03(01):1–6. https://doi.org/10.4172/ 2155-6199.1000135
- Alaboudi KA, Ahmed B, Brodie G (2018) Annals of agricultural sciences phytoremediation of Pb and cd contaminated soils by using sun fl ower (Helianthus annuus) plant. Ann Agric Sci 63(1): 123–127. https://doi.org/10.1016/j.aoas.2018.05.007

- Alimardan M, Ziarati P, Moghadam RJ (2016) Adsorption of Heavy Metal Ions from Contaminated Soil by B. integerrima Barberry Adsorption of Heavy Metal Ions from Contaminated Soil by B. integerrima Barberry. Biomed Pharmacol J 9(1):1–7. https://doi.org/10.13005/bpj/924
- Bendouz M, Dionne J, Tran LH, Coudert L, Mercier G, Blais JF (2017) Polycyclic aromatic hydrocarbon oxidation from concentrates issued from an attrition process of polluted soil using the Fenton reagent and permanganate. Water Air Soil Pollut 228(3):114–127. https:// doi.org/10.1007/s11270-017-3292-x
- Bertollino, M. (2019). Evaluation of in vitro anti sponges against denaturation of evaluation Inflanmmatoryactivity of five selected marine sponges, (December)
- Bibi F, Faheem M, Azhar EI, Yasir M, Alvi SA, Kamal MA et al (2016) Bacteria from marine sponges : A source of new drugs bacteria from marine sponges : A source of new drugs. Curr Drug Metab 17(October):1–6. https://doi.org/10.2174/13892002176661610130906
- Campana S, Busch K, Hentschel U, Muyzer G, de Goeij JM (2021b) DNA -stable isotope probing DNA-stable isotope probing (DNA-SIP) identifies marine sponge-associated bacteria actively utilizing dissolved organic matter (DOM). Environ Microbiol 00(00):1–16. https://doi.org/10. 1111/1462-2920.15642
- Campana S, Hudspith M, Lankes D, de Kluijver A, Demey C, Schoorl J et al (2021a) Processing of naturally sourced macroalgal- and coral-dissolved organic matter (DOM) by high and low microbial abundance encrusting sponges. Front Mar Sci 8(May):1–14. https://doi.org/10. 3389/fmars.2021.640583
- Costa G, Violi B, Bavestrello G, Pansini M, Bertolino M (2020) Aplysina aerophoba (Nardo, 1833) (Porifera, Demospongiae): an unexpected miniaturised growth form from the tidal zone of Mediterranean caves: morphology and DNA barcoding. Eur Zoolog J 87(1):73–81. https:// doi.org/10.1080/24750263.2020.1720833
- Cowden WH (1976) Factors controlling sponge distribution in the gulf of Mexicomand the resulting zonation. Aspec of sponge biology. A subsidiary of Harcourt brace. In: Ecology. JovanovichPublishers. Academic press. New, London, pp 261–276
- Dadrasnia A, Maikudi Usman M, Tzin Lim K, Farahiyah FH, Binti Mohd Rodzhan NS, Abdul Karim SH, Ismail S (2020) Bio-enhancement of petroleum hydrocarbon polluted soil using newly isolated bacteria. Polycycl Aromat Compd 40(2):484–493. https://doi.org/10.1080/10406638.2018.1454966
- de Kluijver A, Nierop KGJ, Morganti TM, Bart MC, Slaby BM, Hanz U et al (2021) Bacterial precursors and unsaturated long-chain fatty acids are biomarkers of North-Atlantic deep-sea demosponges. PLoS One 16:1–18. https://doi.org/10.1371/journal.pone.0241095
- Duchassaing de Fonbressin P, Michelotti G (1864) Spongiaires de la mer Caraibe. Natuurkundige Verhandelingen van de Hollandsche Maatschappij Der Wetenschappen Te Haarlem 21(2): 1–124
- Fang H, Shi Y, Zhou M, Niu Q (2020) Influence of n-hexadecane and naphthalene on anaerobic digestion: kinetic simulation, DOM variation and microbial community assessment. IOP Conference Series: Earth and Environ Sci 555(1):012038. https://doi.org/10.1088/1755-1315/555/1/ 012038
- Gebregewergis A (2020) Levels of selected metals in white teff grain samples collected from there different areas of Ethiopia by using microwave plasma atomic emission spectroscopy (MP-AES). Int J Novel Res Phys Chem Mathemat 7(1):13–24
- Gunathilake V (2018) Evaluation of in vitro anti sponges against denaturation of protein. Int J Curr Res 10(4):68349–68353
- Idah T, Sheer S, Feldstein T, Yahel R, Huchon D, Ilan M (2018) Shedding light on an East-Mediterranean mesophotic sponge ground community and the regional sponge fauna. Mediterr Mar Sci 19(1):549–565
- Kadhim R, Maajeed H, Munim G (2013) Sequence similarity for identification of locally isolate Geobacillus stearothermophilus according to lipase gene sequence. Full Length Research Paper Sequence similarity for identification of locally isolate Geobacillus stearothermophilus according to l. Sci Park J 2(2):42–50

- Kamaruddin M, Marzuki I, Burhan A, Ahmad R (2021) Screening acetylcholinesterase inhibitors from marine-derived actinomycetes by simple chromatography. In: The 1st International Conference on Biotechnology and Food Sciences IOP Conf. Series: Earth and Environmental Science, vol 679, p 012011. https://doi.org/10.1088/1755-1315/679/1/012011
- Khabouchi I, Khadhar S, Chaouachi D, Chekirbene A, Doumenq P (2020) Study of organic pollution in superficial sediments of Meliane river catchment area: aliphatic and polycyclic aromatic hydrocarbons study of organic pollution in superficial sediments of Meliane river catchment area: aliphatic and polycyclic aromatic hydrocarbons. Environ Monit Assess 192(5): 282–290. https://doi.org/10.1007/s10661-020-8213-6
- Knobloch S, Jóhannsson R, Marteinsson V (2018) Bacterial diversity in the marine sponge Halichondria panicea from Icelandic waters and host-specificity of its dominant symbiont "candidatus Halichondribacter symbioticus". FEMS Microbiol Ecol 95(1):1–13. https://doi. org/10.1093/femsec/fiy220
- Konkolewska A, Piechalak A, Ciszewska L, Antos-krzemi N, Skrzypczak T (2020) Combined use of companion planting and PGPR for the assisted phytoextraction of trace metals (Zn, Pb, Cd). Environ Sci Pollut Res 27(12):13809–13825
- Krishnamoorthy U, Sniffen CJ, Stern MD, Van Soest PJ (1983) Evaluation of a mathematical model of rumen digestion and an in vitro simulation of rumen proteolysis to estimate the rumenundegraded nitrogen content of feedstuffs. Br J Nutr 50(3):555–568. https://doi.org/10.1079/ bjn19830127
- Lajayer BA, Khadem N, Maghsoodi MR, Ghorbanpour M (2019) Phytoextraction of heavy metals from contaminated soil, water, atmosphere using ornamental plants: mechanisms and efficiency improvement strategies. Environ Sci Pollut Res 26(9):8468–8484. https://doi.org/10.1007/ s11356-019-04241-y
- Laport M, Santos O, Muricy G (2009) Marine sponges: potential sources of new antimicrobial drugs. Curr Pharm Biotechnol 10(1):86–105. https://doi.org/10.2174/138920109787048625
- Lu C, Hong Y, Liu J, Gao Y, Ma Z, Yang B et al (2019) A PAH-degrading bacterial community enriched with contaminated agricultural soil and its utility for microbial bioremediation  $\stackrel{\wedge}{\sim}$ . Environ Pollut 251(Mei):773–782. https://doi.org/10.1016/j.envpol.2019.05.044
- Maldonado M, López-Acosta M, Busch K, Slaby BM, Bayer K, Beazley L et al (2021) A microbial nitrogen engine modulated by Bacteriosyncytia in Hexactinellid sponges: ecological implications for Deep-Sea communities. Front Mar Sci 8(March):1–35. https://doi.org/10.3389/fmars. 2021.638505
- Marzuki I, Sinardi S, Pratama I, Chaerul M, Paserangi I, Mudyawati M, Asaf R (2021c) Performance of sea sponges micro symbionts as a biomaterial in biodegradation naphthalene waste of modified. In: *The 5th international seminar on sustainable urban development; IOP conference series: earth and environmental science*, vol 737, p 012016. https://doi.org/10.1088/1755-1315/737/1/012016
- Marzuki I (2020) The bio-adsorption pattern bacteria symbiont sponge marine against contaminants chromium and manganese in the waste modification of laboratory scale. Indonesia Chimica Acta 13(1):1–9. https://doi.org/10.20956/ica.v13i1.9972
- Marzuki I, Ali MY, Syarif HU, Gusty S, Daris L, Nisaa K (2021d) Investigation of Biodegradable Bacteria as Bio indicators of the Presence of PAHs Contaminants in Marine Waters in the Marine Tourism Area of Makassar City. In: *The 6th International Conference on Tropical and Coastal Region Eco*, p 012006. https://doi.org/10.1088/1755-1315/750/1/012006
- Marzuki I, Chaerul M, Erniati A, Paserangi I (2020a) Biodegradation of aliphatic waste components of oil sludge used micro symbiont of Sponge Niphates sp. Biodegradation of aliphatic waste components of oil sludge used micro symbiont of Sponge Niphates sp. ICMS 429(1):012056. https://doi.org/10.1088/1755-1315/429/1/012056
- Marzuki I, Daris L, Nisaa K, Emelda A (2020b) The power of biodegradation and bio-adsorption of bacteria symbiont sponges sea on waste contaminated of polycyclic aromatic hydrocarbons and heavy metals. *IOP Conference Series: Earth and Environmental Science* 584:012013. https:// doi.org/10.1088/1755-1315/584/1/012013

- Marzuki I, Daris L, Yunus S, Riana AD (2020c) Selection and characterization of potential bacteria for polycyclic aromatic biodegradation of hydrocarbons in sea sponges from Spermonde Islands, Indonesia. *AACL Bioflux* 13(6):3493–3506
- Marzuki I, Kamaruddin M, Ahmad R (2021a) Identification of marine sponges-symbiotic bacteria and their application in degrading polycyclic aromatic hydrocarbons. Biodiversitas 22(3): 1481–1488. https://doi.org/10.13057/biodiv/d220352
- Marzuki I, Noor A, La Nafie N, Djide MN (2015) Sponge role in alleviating oil pollution through sludge reduction, a preliminary approach. Int J Appl Chem 11(4):427–441
- Marzuki I, Noor A, La Nafie N, Djide MN (2016) Morphological and phenotype analysis of microsymbiont and biomass marine sponge from melawai beach, Balikpapan, east kalimantan. Int J Marina Chimic Acta 17(1):8–15
- Marzuki I, Pratama I, Ismail HE, Paserangi I, Kamaruddin M, Chaerul M, Ahmad R (2021b) The identification and distribution components of polycyclic aromatic hydrocarbon contaminants at the port of Paotere, Makassar, South Sulawesi. In: *The 1st International Conference on Biotechnology and Food Sciences IOP Conf. Series: Earth and Environmental Science IOP Publishing*, vol 679, p 012017. https://doi.org/10.1088/1755-1315/679/1/012017
- Medic A, Ljesevic M, Inui H, Stojanovic K, Karadzic I, Be'skoski V, Koji I (2020) Efficient biodegradation of petroleum n-alkanes and polycyclic aromatic hydrocarbons by polyextremophilic Pseudomonas aeruginosa san ai with multidegradative capacity. Royal Soc Chem Adv 10(April):14060–14070. https://doi.org/10.1039/C9RA10371F
- Melawaty L, Noor A, Harlim T, de Voogd N (2014) Essential metal Zn in sponge Callyspongia aerizusa from Spermonde archipelago. Adv Biol Chem 04(01):86–90. https://doi.org/10.4236/ abc.2014.41012
- Obire O, Aleruchi O, Wemedo S (2020) Fungi in biodegradation of polycyclic aromatic hydrocarbons in oilfield wastewater. Acta scientific. Microbiology 3(4):220–224. https://doi.org/10. 31080/asmi.2020.03.0572
- Onwosi CO, Odibo FJC, Enebechi CK, Nwankwegu S, Ikele AI, Okeh OC et al (2017) Soil and sediment contamination : an international bioremediation of diesel-contaminated soil by composting with locally generated bulking agents bioremediation of diesel-contaminated soil by composting. Soil Sediment Contam 0(0):1–19. https://doi.org/10.1080/15320383.2017. 1348337
- Orani AM, Barats A, Vassileva E, Thomas OP (2018) Marine sponges as a powerful tool for trace elements biomonitoring studies in coastal environment. Mar Pollut Bull 131(April):633–645. https://doi.org/10.1016/j.marpolbul.2018.04.073
- Parama Cita Y, Kamal Muzaki F, Radjasa OK, Sudarmono P (2017) Screening of antimicrobial activity of sponges extract from Pasir Putih, East Java (Indonesia). J Marine Sci Res Develop 07(05):1000237. https://doi.org/10.4172/2155-9910.1000237
- Parhamfar M, Abtahi H, Godini K, Saeedi R, Sartaj M, Villaseñor J et al (2020) Biodegradation of heavy oily sludge by a two-step inoculation composting process using synergistic effect of indigenous isolated bacteria. Process Biochem 91(September):223–230. https://doi.org/10. 1016/j.procbio.2019.12.014
- Rua, C. P. J., Oliveira, L. S. De, Froes, A., Tschoeke, D. A., Soares, A. C., Leomil, L., ... Thompson, F. L. (2018). Microbial and functional biodiversity patterns in sponges that accumulate Bromopyrrole alkaloids suggest horizontal gene transfer of Halogenase genes. *Microbial* ecology journal-springer science+business media, (march)
- Sabdono A, Radjasa OK (2008) Microbial symbionts in marine sponges: marine natural product factory. J Coastal Develop 11(2):57–61
- Saputra MT, Sadarun B, Rahmadani, Subhan (2019) Relationship between conditions of live coral cover with abudance of chaetodontidae fishes in lalanu water, Soropia District, Konawe District. Sapa Laut 4(2):53–60
- Schmittmann L, Jahn MT, Pita L, Hentschel U (2020) Decoding cellular dialogues between sponges, bacteria, and phages. In: *Cellular dialogues in the holobiont*, pp 49–63. https://doi. org/10.1201/9780429277375-4

- Schuster A, Cárdenas P, Pisera A, Pomponi SA, Kelly M, Wörheide G, Erpenbeck D (2018) Seven new deep-water Tetractinellida (Porifera: Demospongiae) from the Galápagos Islands - morphological descriptions and DNA barcodes. Zool J Linnean Soc 184(2):273–303. https://doi. org/10.1093/zoolinnean/zlx110
- Shareef HK, Muhammaed HJ, Hussein HM, Hameed IH (2016) Antibacterial effect of ginger (Zingiber officinale) roscoe and bioactive chemical analysis using gas chromatography mass Spectrum antibacterial effect of ginger (Zingiber officinale) roscoe and bioactive chemical analysis using gas chromatography mas. Orient J Chem 32(2):817–837. https://doi.org/10. 13005/ojc/320207
- Siahaya, N., Noor, A., Sukamto, N., & de Voogd, N. (2013). A preliminary effort to assign sponge <i>Callispongia sp</i>) as trace metal biomonitor for Pb, Cd, Zn, and Cr, an environmental perspective in Hative gulf waters Ambon. Adv Biol Chem, 03(06), 549–552. https://doi.org/10. 4236/abc.2013.36062
- Sobrinho ANMBD, Andrade AFMD, Lima ESA, Zonta E, Magalhães MOL (2020) Metals Phytoextraction by Cordia africana from soils contaminated with oil drilling waste. Floresta eAmbiente 27(1):1–8
- Tereza Č, Wiesnerová L, Praus L, Jablonský I, Koudela M, Tlusto P (2018) ScienceDirect comparing the removal of polycyclic aromatic hydrocarbons in soil after different bioremediation approaches in relation to the extracellular enzyme activities. J Environ Sci 76:1–10. https:// doi.org/10.1016/j.jes.2018.05.007
- Ulli H, Noor A, Mandey FW, Sapar A (2016) Isolation, identification, and bioactivity test of non polar compounds on n-hexane extract of Haliclona (Reniera) fascigera from Samalona Island-Spermonde archipelago. Marina Chimica Acta 17(2):32–41
- Wibowo N, Nurcahyo R, Gabriel DS (2019) Sponge iron plant feasibility Studi in Kalimantan, Indonesia. ARPN J Eng Appl Sci 14(23):4013–4020
- Yang Q, Franco CMM, Lin HW, Zhang W (2019) Untapped sponge microbiomes: structure specificity at host order and family levels. FEMS Microbiol Ecol 95(9). https://doi.org/10. 1093/femsec/fiz136
- Yogaswara D (2017) Concentrations of PAHs (Polycyclicaromatic Hydrocarbons) Pollutant in Sediment of The Banten Bay. Bull Marine Geol 32(2):61–66. https://doi.org/10.32693/bomg. 32.2.2017.377
- Ziarati P, Mirmohammad Makki F, Vambol S, Vambol V (2019a) Determination of toxic metals content iranian and Italian flavoured olive oil. Acta Technol Agric 22(2):64–69. https://doi.org/ 10.2478/ata-2019-0012
- Ziarati P, Vambol V, Vambol S (2019b) Use of inductively coupled plasma optical emission spectrometry detection in determination of arsenic bioaccumulation in Trifolium pratense L. from contaminated soil. Ecol Questions 31(1):1. https://doi.org/10.12775/eq.2020.003

## **Chapter 11 Genetic Modification: A Gateway to Stimulate the Industrial Production of Biofuels**



## Poonam Singh, Kaleemunnisa FNU, and Telma Encarnação

Abstract Recent years have seen an explosion in the use of advanced biotechnology techniques in academic and industrial activities to modulate microorganism pathways for the production of fuels or chemicals. Synthetic biology is adopted for biofuel production, and it needs scientific evidence to support the fundamentals and risk assessments. Biofuel is derived from biomass of a plant, animal waste, or microalgae. These materials can be replenished after some time; hence they are a renewable source of energy. Risk assessment is considered significant to maintain and comply with regulatory frameworks existing around the world. The use of scientific tools such as enzymes and microbes itself needs review and approval. Risk profiles are done for the toxicity, infectivity, and strategies. Recent years have seen an exciting increase in developing strategies for the use of advanced biotechnology techniques to enhance the productivity of existing biosynthetic pathways in microbes by cutting off the competing pathways. The biomass is pretreated to speed up the process of obtaining biofuel. The mutant, after genetically modifying the enzymes, produces cellulases and hemicellulases in higher levels. Functional analysis can confirm the changes in several transcription regulatory elements. Generally, successful engineering is demonstrated with an enhanced supply of amino acids.

**Keywords** Biofuels · Genetic engineering · Lignocellulose · Enzymes · Fungus · Artificial intelligence

P. Singh (🖂)

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

K. FNU

T. Encarnação

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

PTScience Avenida do Atlântico, Lisbon, Portugal

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_11

Seidenberg School of CSIS, Pace University, New York, NY, USA

Centre for Rapid and Sustainable Product Development, Polytechnic Institute of Leiria, Marinha Grande, Portugal

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

## 11.1 Introduction

The geographical production of fuel from fossils is a gradual process; an alternative is to obtain the fuel using biomass or living beings, which is termed biofuel. The times have changed, and there is a constantly increasing need for petroleum (Lazarus and Van 2018). The natural sources of the fuel industry are not present in all the countries. Fuel exhaustion has become a significant problem putting the countries unrest and dependent on buying fuel from the enriched countries (Pfaltzgraff and Clark 2014). The readily available biomass that is used directly to convert to biofuel is recovered from wood and grass. Biofuels produce heat energy which generates electricity after being run in a generator. No corner in the world is left to be negatively impacted by the effects of petroleum extraction, refining, transportation, and use. Therefore, biofuels are increasingly getting attention as an alternative source of energy (Nehring 2009; Zou et al. 2016).

## 11.1.1 Biofuels Basic Definition

Biofuel exists in all forms such as solid, liquid, and gas. Liquid and gaseous forms are easier to transport and deliver. Biofuel term is used for ethanol, biodiesel, green diesel, and biogas (Masjuki et al. 2012). The biofuel production is broadly characterized into first-generation, derived from sugars, starches, and oils, and the crops versus second-generation biofuels extracted from lignocellulosic biomass sources. Recently, the effort has been started to derive biofuels from microalgae and cyanobacteria, termed third-generation production (Rodionova et al. 2017); and fourth-generation biofuels which include the genetic modification of the microorganisms for the enhancement of biohydrogen production processes.

It is no secret that biofuels are preferred and viable substitutes over fossil fuels. However, biofuels such as ethanol create a net energy loss when compared to petrol. If we consider the food-based crops, they must be used for feeding the enormous population than used for fuel. As the demand to produce bio crops increases with the demand for organic consumables, soil erosion, deforestation, fertilizer run-off, and salinity are some of the major issues (Larson and United Nations Conference on Trade and Development 2008; Cheng and Timilsina 2011).

## 11.1.2 Why the Need for Biofuel?

The need for alternative and efficient methods has made researchers find new ways to produce biofuel. Sugar and sugar-derived ethanol is making a significant contribution to satisfying the need at the moment. Sugar fermentation is used for transportation fuel. Starch-derived ethanol fills the energy supply by mainly using corn grain production. After the conversion of corn grain into ethanol, its burning causes an emission that impacts greenhouse gas emission but not in a net increase in atmospheric carbon dioxide (Lal 2005). The countries are announcing policies and goals to produce their own biofuel and be less dependent on foreign oil. Britain, the USA, and Canada are encouraging to grow primarily biofuel crops and establish cellulosic ethanol refineries major biofuel centers to produce a million gallons of cellulosic ethanol per year (Sims et al. 2010).

#### 11.1.2.1 Bioethanol

Petroleum demand has put much industrial unease in the countries. Ethanol is readily biodegradable, but its use, just like petroleum, produces air and water-borne pollutants. Feedstock production of the crops to get bioethanol reduces the greenhouse gases like carbon dioxide from the environment, which is being used for photosynthesis (Lima et al. 2012). As the cellulosic matter is present in an abundant amount, it is more fitted than starch and sugar for ethanol because of its limited supply. Food supplies rely on starch-based crops such as sugar cane or corn that need specific climatic conditions to grow; hence, cellulose biomass that is not dependent on weather conditions is convenient to produce bioethanol in most countries (Sarkar et al. 2012). Ethanol produced from cellulose-based material has the potential to replace petroleum (In Marcel 2015).

#### 11.1.2.2 Biodiesel

The second most demanding biofuel is biodiesel, which is obtained from soybean, palm oil, and fat of cooking oil. Algae and cyanobacteria have the potential to account for a large amount of fuel per unit area. Biodiesel is used in combination with petroleum and is widely accepted in European countries.

In this chapter, an oversimplified view of the production of biofuels through various resources, their economic analysis, and possible genetic manipulation of the crops to overcome the existing challenges are presented.

## **11.2 Biofuel from Different Sources**

#### 11.2.1 Biofuels from Lignocellulose Biomass

Lignocellulose biomass is universal and widespread renewable biomaterial on our planet. The bioconversion of lignocellulose comes as a feasible strategy when juxtaposed with the other alternative energies. Three components that are rigidly packed with each other in lignocellulose are cellulose, constituting 30–50% part of it, whereas hemicelluloses 20–40%, and lignin 20–30%. Due to the compact

structure, degradation into fermentable sugars and further conversion into fuels and other value-added materials become difficult (Menon and Rao 2012; Sharma et al. 2017).

Cellulose is linked with  $\beta$ , 1–4 linkage of glucose. Cellulase enzymes can break it down into glucose. Cellulose is not found in pure form in nature but with hemicelluloses and lignin, which act as a physical barrier for cellulase to access cellulose (Volynets and Dahman 2011). Cellulosic ethanol is associated with a high cost of bioreactors; the breakdown of lignin and its removal is required to access cellulose biomass by cellulases to obtain the biofuel from a plant source. The use of xylanases with cellulases is more effective as xylanases hydrolyze hemicelluloses and make cellulose available for biomass degradation. This co-acting releases more fermentable sugars from the biomass (Hu et al. 2011); this method is economically more expensive in comparison to ethanol from corn. Genetic engineering offers a substitution to minimize the cost production of cellulosic ethanol. The first approach can be an integration of cell wall degrading enzymes cellulases and hemicellulases in the crop instead of adding them directly in the bioreactor. Secondly, the amount of lignin can be configured, and the pretreatment process can be avoided. Most importantly, the maximization of polysaccharides can boost cellulosic biofuel production (Hu and Catchmark 2011).

### 11.2.1.1 The Cellulosic Ethanol Production Process

Lignocellulosic biomass harvested from the feedstock crop is transported to a refinery where it is stored. Either this biomass is treated with extreme heat or with chemicals to remove the lignin by breaking it down into the intermediates. The separation of the solid and liquid components is done by filtration, distillation, evaporation, and chromatography. After the enzymatic hydroxylation using bacteria or fungi, it is ready for the conversion to cellulosic ethanol. After the separation of sugar is done, pure ethanol is obtained (Liu et al. 2019; Zheng et al. 2009).

#### 11.2.1.2 Factors Affecting the Production from Lignocellulose Biomass

Cellulosic ethanol production changes with country, region, agriculture, economy, and politics. These complex factors depend on the type of crop produced, its demand, and the transportation fuel used in that area. The support from the community and government regarding breeding strategy is also paramount for conversion to cellulosic ethanol as it gives a high amount of cellulosic biomass. C4 photosynthetic pathway, perennial growth, water usage efficiency, and segregated underground storage nutrients are the prototypical features of a non-consumable cellulose crop (Huang et al. 2009). These might include silver grass, switchgrass, and woody crops. Other edible crops are rice, corn, and sugarcane. The plant cell wall is the source of lignocellulosic biomass, and it determines the structural configuration of the plant; a representation is shown in Fig. 11.1. The various combinations of glucose sugars are



Fig. 11.1 A depiction of the biomass treatment process and genetic engineering for biofuel production

converted to ethanol (Xiros et al. 2013). The cell wall has crosslinked glycan in cellulose, and it is classified in consonance with the type of crosslinks. There are two types: Type I walls have the same amounts of glucan and xyloglucan embedded in a matrix of pectin in dicotyledonous plants; whereas Type II has glucuronoarabinoxylans and lacks pectin and structural proteins in cereals and grasses (Fig. 11.2).

Cellulose, hemicellulose, and pectin are the polysaccharides present in plant primary cell walls; the hydrolysis of their fermentable sugars provides bioethanol production. In trees, the secondary cell wall has three layers, distinguished based on different arrangements of cellulose microfibrils, with only the outermost layer containing the helices. The secondary cell wall of a plant has cellulose, hemicellulose, and lignin, mostly in which cellulose is embedded in lignin in the form of microfibrils (Huang et al. 2009; Sainz 2009; Kenney and Idaho National Laboratory (INL) 2007).

## 11.2.2 Biofuels from Enzymes

Aerobic and anaerobic microorganisms both produce enzymes cellulose-degrading enzymes. Bacteria and anaerobic fungi produce cellulosomes. It is a complex of cellulolytic enzymes associated with their cell wall. The secretion of cellulases is either free or cell surface-bound (Binod et al. 2019). With the unraveling of new places and areas, cell wall deconstructing enzymes have been and being isolated and



Fig. 11.2 Cell wall primary structure (above) and secondary structure (below) with cellulose microfibrils, hemicellulose, pectin, lignin, and soluble proteins (Sticklen 2008)

characterized from a variety of samples in order to be used in pretreatment investigation. The objective is to achieve higher resistance to the conversion temperatures and a range of pHs. Enzymes cellulases and hemicellulases convert the polysaccharides into fermentable sugars through enzymatic hydrolysis (Himmel et al. 2007).

## 11.2.2.1 Cellulases

Filamentous fungi produce extracellular cellulases. Due to the ability to produce extracellular cellulose, fungi have been highly researched for cellulase production for a long time. Both fungi and bacteria can degrade biomass; hence the biological method of producing sugars with enzymes is an eco-friendly and accomplishable method (Wilson 2009).

The maximum operating efficiency of the enzymes and their complex composition are always unrepresentative of each other. *Trichoderma viride* and *T. reesei* are amongst the excessively studied microbes (Schuster and Schmoll 2010).

Cellulases are not a single but a group of enzymes composed of endoglucanase and exoglucanases, it also includes cellobiohydrolases and  $\beta$ -glucosidase. Their topologies include  $\beta$ -sheet proteins,  $\beta/\alpha$ -barrels, and  $\alpha$ -helical protein. Lignin prevents cell wall hydrolysis by not let enzymes accessing polysaccharides and acting as a barrier. Enzyme production by microbes exposes cellulose to be broken down by cellulases (Schuster and Schmoll 2010). The microbes have the ability to synthesize different functional enzymes effectuating fermentable sugar for biofuels. Cellulases have many applications in industries, and the production of secondgeneration bioethanol is amongst one of them, it is a low-cost material obtained from lignocellulose. The most abundant renewable biomass; bioethanol production does not create any food insecurity, when lignocellulose is used, hence avoiding any food crops before harvesting. Biological pretreatment concerns with the ligninolytic potential of certain microorganisms that can reduce the recalcitrant nature and cleave it via hydrolytic enzymes. The addition of a molecule with two glucose units into the cultures can increase the cellulase expression. Differential hybridization in many studies showed that gene regulation occurs at transcriptional level (Siqueira et al. 2020; Srivastava et al. 2015).

Abstraction of expression of the glycolytic phosphoglycerate kinase gene, *pgk1*, is the common metabolism observed in cultured media of cellulose. Identification of the functional regions in the promoters of the cellulase genes or the regulatory proteins involved is still a highly debatable topic (Da and Srikrishnan 2012).

#### 11.2.2.2 Factors Affecting the Production from Cellulases

Currently, there are two important factors restricting the production of cellulosic ethanol. The first is the production of strain-specific cellulases, and the other is non-appliance of existing commercial hemicellulases that can increase the output of multiple carbon fermentable sugars. Bioconversion of the complex lignocellulosic material to simple sugars is a complicated process. Genetically modified thermophilic bacteria is the envision for the future as it may lead to enhanced cellulase production through recombinant technology. Isolation of cellulase gene from thermophilic bacteria and its expression in suitable hosts via recombinant technology might enhance cellulase production. It could be done through a classical approach, whole-genome isolation, whole metagenome isolation, or a pre-study with bioinformatics (Verma et al. 2021; Kuhad et al. 2011).

## 11.2.3 Induction of Cellulase Expression

Plant-based material or cellulose is used in the media to promote high cellulase expression. Expression of cellulose is promoted in cellulose, lactose, and cellobiose that are poor carbon sources rather than in glucose and glycerol (Amore et al. 2013). Cellulase expression is thought to be induced using natural compound sophorose, but the status of cellobiose stays controversial as it needs to be fed in a controlled manner. Cellulase expression might be interfered by the type of nitrogen source that is used. Most of the natural carbon sources used to study cellulase expression offer a competitive growth to other microbes like fungus (Ilmén and Saloheimo 1997).

## **11.3** Literature Review

#### 11.3.1 Fungus as the Source

When the expression of the cellulases of filamentous fungus Trichoderma reesei QM9414 was studied with genes encoding enzymes cellobiohydrolases and endoglucanases, the steady-state mRNA for cellobiohydrolases was highly expressed. It is also being concluded that cellobiose as a carbon source as an inducer does not show immediate effect and seems to vary depending on the culture conditions, in a study where cellobiose promoted cellulase transcription to a moderate level when compared to that of lactose (Ilmén and Saloheimo 1997). Accumulation of glucose in the culture medium might result in glucose repression of cellulase transcription. The inducing power very much depends on the ratio between carbon source, formation of glucose, and their uptake from the medium. Most of the natural carbon sources used to study cellulase expression offer a competitive growth to other microbes like fungus. Sorbitol and glycerol generally have a neutral effect; it neither promote nor inhibit expression. Glycerol and sorbitol without affecting the fungal growth show a cellulase gene induction in cultures with 1-2 mM sophorose. To understand what is the source of inducing compounds and if they are released from cellulose, the studies have been performed only with glucose and no inducer as a carbon source. If the amount of glucose in a media is subdued, the level of mRNA is found to have a difference in fully induced and repressed states in an actively growing fungus. Without having to add an inducer and still able to cellulase expression after glucose depletion, it can be crucial to keep the fungus alive under starvation conditions (Ilmén and Saloheimo 1997; Margolles-clark et al. 1997).

Aerobic, anaerobic bacteria, and fungi are the models to study cellulolytic enzyme systems. Fungi such as *Trichoderma reese*i, *Penicillium spp., Aspergillus niger*, and basidiomycetes secrete extracellular cellulolytic enzymes. Higher fungi also have oxidative systems and can be capable of degrading lignocelluloses through their ligninolytic enzymes.

Fungi are studied for genetic modification reason being their capacity to produce large amounts of extracellular cellulases. Three mainly synergistically acting enzymes in cellulases are cellobiohydrolase/exoglucanase, endo- $\beta$ -1,4-glucanase, and (c)  $\beta$ -glucosidase. The action of endoglucanase is expedited by lytic polysaccharide monooxygenases. Non-hydrolytic proteins accelerate the action of endoglucanase. Optimization of physical and nutritional parameters can be done by engineering the cellulases, it is also the way to enhance cellulose production. Strain improvement can be approached via random mutagenesis and site-specific mutagenesis in cellulases. One of the most explored fungal strains for commercial cellulase production is *T. reesei* RUT-C30, followed by *Penicillium sp., Aspergillus sp., Myceliophthora*, and *Humicola sp.* (Srivastava et al. 2020).

A study concluded that in a growing fungus, the regulation of cellulase expression depends on glucose repression (Amore et al. 2013). This effect is reversed after the glucose is impoverished and derepression of cellulose occurs with no other inducer present. The use of a mutant can support new ways for biodiesel production; some mutant studies support the theory that separate regulation could exist for different fungal cellulolytic enzymes. This suggests that there is a high possibility that the cellulose enzymes are coordinately expressed. It is always to be mentioned that biologically relevant mRNA levels are not easy to be detected and site-specific proteins might be the ones governing glucose repression or the cellulase expression (Mach and Zeilinger 2003).

#### 11.3.2 Algae as the Source

Photosynthetic algae, both micro and macro, are thought to have the potential to be turned as a possible biofuel resource. Microalgae have the competence to store triacylglycerol and fat that can be turned into biodiesel and ethanol. Due to a high lipid profile, it is believed that crude oil deposits have been created by microalgae over a period of time. Therefore, scientists have a huge interest in understanding and exhilarating the productivity of algae to produce biofuels. What makes microalgae even more interesting is the fact that they are an attractive source of fuel that intake carbon dioxide and keep the environment low on carbon. They grow on marginal land hence are not a competition to terrestrial crops and flourish in waste or saltwater. It means algae does not compete with the resources of food-based crops, which is a problem with lignocellulose-based biofuels. Metabolic pathways of microalgae can significantly be manipulated to produce a greater quantity and a variety of biofuels (Demirbas 2010).

Algae efficiently use photosynthesis to obtain important oils and biomass from carbon dioxide. These oils can be later transformed into feedstocks to produce biofuels, Omega-3 fatty acid oils, and feed for animals feed. Productivity of algae can give an estimate about the approximation of area to fulfill the requirements in a particular country. The protein that comes as a byproduct of fuel production from algae might serve as a very useful food source for protein and other useful products.



Fig. 11.3 Representation of the structure of microalgae cell wall membrane

The synergistically designed facility takes the edge off liquid fuel hence relaxing the consequences of biofuels production on the land, mitigating global warming, and promoting biofuel production to relieve energy shortage (Demirbas 2011). Microalgae can be aggregated by adjusting the pH; with these adjustment properties of the cell surface, biomass concentration can be modified as per the requirement. After the culture of microalgae, the medium can be recycled, reducing the cost and environmental pollution (Fig. 11.3). The high-pH-induced flocculation method showed an effect on biodiesel production by showing changes in the lipid extraction process and fatty acid profiles of marine microalgae (Castrillo et al. 2013; Liu et al. 2013a).

Nonetheless, and contrary to microalgae, seaweed is a renewable feedstock, and there are some potential concerns and impacts on ecosystems. Polymers in seaweed are mixed sugars, and depolymerization of seaweed polysaccharides is relatively easy, but the conversion of such sugars to biofuels is not an easy task. Excess of seaweed farming can alter natural habitats leading to nutrient depletion and reduction in biodiversity (Kraan 2013).

Economically profitable biofuel production from seaweed is acquired by an efficient conversion of mixed sugars in seaweed hydrolysates. Henceforth, metabolic engineering can be used during the fermentation of sugars. Red marine algae have galactose (up to 23%) as major sugar compound in the hydrolysate Ceylon moss (Wei et al. 2013). Galactose fermentation by engineering *S. cerevisiae* is the eminent way to produce ethanol. The wild-type yeast *S. cerevisiae* is capable of galactose fermentation by controlling the ethanol yield and productivity. However, ethanol production rate and galactose yield are not high when compared to glucose. Also, glucose represses the utilization of galactose by stringent transcriptional repression of GAL genes that are responsible to code enzymes for galactose metabolism.

Hence, excess consumption of glucose and galactose in red seaweed hydrolysates might reduce overall ethanol productivity, which is thought to be improved by metabolic engineering (Johnston et al. 1994).

## 11.4 Genetic Gateway to Obtain Biofuels

Genetic engineering enables microbe to produce a high number of metabolites. The inherent complexity of the organisms ranges from simple protein structures to folded and globular protein with a variety of medicinal properties. The genetic manipulation requires a preamble of whole-genome sequence to understand and select the desired sites for genetic alterations (Peralta-Yahya et al. 2012). A few examples of such moderation in the gene of the different microbes are mentioned in Table 11.1.

Organism	Mutant/gene	Modification	Origin	Reference
T. reesei Qm 6a	RUT-C30	There was an increase by 20 times in cellulase secretion	Fungal	(Peterson and Nevalainen 2012)
Fusarium oxysporum	NTG-19	Cellulolytic activity was 80% more than its parent strain	Fungal	(Kuhad et al. 1994)
Aspergillus nidulans	creAd3	D-glucose metabolism was seen to be improved	Fungal	(Van et al. 1995)
Aspergillus niger DSM 26641	<i>A. niger</i> DSM 28712	β-1,4-endoxylanase activity was found to be enhanced by 82%	Fungal	(Ottenheim et al. 2015)
Cellulomonas flavigena	(M4, M9, M11, and M12)	Xylanolytic activities were enhanced	Bacterial	(Mayorga- Reyes and Ponce-Noyola 1998)
<i>Escherichia coli</i> strains	cydC-D86G, cydC-D86V	Biofuels and bi-products under ionic liquid stress were higher in the concentration	Bacterial	(Eng et al. 2018)
Chlorella vulgaris	SDEC-3 M	The mutant is supposed to benefit $CO_2$ biofixation from industrial exhaust gas	Algal	(Qi et al. 2016)
Chlorella minutissima (CM)	CM7	Monounsaturated fatty acids showed an elevation	Algal	(Mehtani et al. 2017)
Saccharomyces cerevisiae	Saccharomyces cerevisiae UAF-1	Ethanol production was improved by 12.0% with aeration	Yeast	(Abbas et al. 2017)

Table 11.1 Improvement of the biofuel production by genetically altering the organisms
### 11.4.1 Principle

Targeted strain engineering aims at a considerable amount of transformants by homologous integration or deletion of the expression cassette. Nevertheless, introns in the genes and glycosylation convolution cause low-efficiency gene targeting.

Filamentous fungi have the tendency to bring about post-translational modification. The modification includes attachment to a functional group of another molecule by a glycosyl donor and synthesis of organosulfur compounds. They also secrete metabolites that have the ability to expand on a cheaper substrate that makes them suitable for industrial applications. They almost make a good choice to be used as a host for recombinant DNA except for the challenges on non-homologous recombination. By inactivating the double-strand breaks in the DNA pathway this challenge can be resolved (Dellomonaco et al. 2010).

### 11.4.2 Some Examples from the Previous Studies

Genetic induction or end-product inhibition in microbial cells produces a higher amount of cellulases. Catabolite repression or end-product inhibition in a mutant of B. pumilus resulted in a four times higher yield when compared to Trichoderma reesei (Kotchoni et al. 2003). RUT-C30 obtained through mutation in T. reesei Qm 6a at Rutgers University showed a 20 times increase in cellulase secretion. Most of the available studies state enhancements in cellulase production by mutation without mentioning the changes that might have occurred at genetic level (Peterson and Nevalainen 2012). Penicillium decumbens is used in China for the industrial production of lignocellulolytic enzymes. When a comparative genomics analysis by Liu et al. (2013b) was made with the phylogenetically similar species *Penicillium* chrysogenum it was found that the cell wall degradation has advanced with P. decumbens. The reason suggested was its strong cellulolytic ability due to more genes involved in cell wall degradation than cellular metabolism, that happens in a medium with cellulose as a carbon source. It has made the lignocellulolytic enzyme system in *P. decumbens* became variegated with hemicellulases and proteins in the cellulose binding site (Liu et al. 2013b). Genetic engineering seems a very efficient method for gene expression by regulating the promoters and can be achieved with minimum changes in the genetic content. Mutagenesis is done to bring the expected changes in the DNA sequences of a gene with specific primers, termed as site directed mutagenesis. The changes are incorporated in a genome by homologous recombination using amino acid sequence primers. However, the sequence in a genome and the site to target are not easy to identify. This process of improving the properties of a protein by alteration in its amino acid sequence increases the secretion of cellulase. Serinine and threonine on the surface of xylanase in A. niger BCC14405 were replaced with arginines (Sriprang et al. 2006). The modified enzyme had increased activity than the wild-type strain. As the enzyme activity

also increased half-life of the mutant stability was simultaneously raised. To obtain a hyperthermostability, *Thermotoga maritima* cel5A endoglucanase, when subjected to site directed mutagenesis and CBM engineering, demonstrated 10% higher activity at one site when compared with the native cel5A (Arumugam et al. 2008).

Aspergillus and T. reesei are used to express genes from different origins, improving cellulose production. Fungal and bacterial cellulose or carbohydrate binding domains were used from T. reesei and Clostridium stercorarium xylanase A to be integrated with cel5A. Avicel was used to observe hydrolytic activity in which engineered carbohydrate bonding molecule from both species showed better activity. The activity was linked to binding ability, which was checked via immune gold labeling assay. Mutagenesis of D232A in fungus Macrophomina was used to generate an engineered form for the production of an enzyme with novel substrate requirements. The substrate size of the engineered one was found to be higher than the wild-type 5 b-1,4-endoglucanase but with an equivalent activity on cellohexaose (Druzhinina et al. 2017; Hilden and Johansson 2004). The modified endoglucanase can be used to get complexed carbohydrates by a double decomposition reaction with water present. A study by Liu et al. (2013b) involved Penicillium oxalicum mutant JU-A10-T gene with the wild strain 112–2. It has high cellulolytic ability on the processing of decayed organic matter, comparison of whole-genome sequencing, transcriptomes, and secretomes was done. The study revealed that a new lignocellulose-degrading enzyme has emerged (Liu et al. 2013b; Wang and Jones 1997).

#### 11.5 Strategies for Genetic Modification of Microorganisms

Creation, selection, or improvement of strains of desired microorganisms to direct the well-suited output rely on the microbial strains that will be used to catalyze the biosynthesis of the desired compound. Biotechnologies develop befitting strains for fuel by synthetic process.

The imposition of these regulations concerns with the underlying risk and their assessments by the government, taking into consideration proper planning and management. Modification of microorganisms for fuels such as n-butanol, isobutanol, mixtures of alkanes or lipids is evident in number or researches (Keasling et al. 2009; Bhatia et al. 2017).

This is done by overexpression, directed evolution, and codon optimization of key endogenous enzymes to increase the yield of the targeted product (Mythili et al. 2016; Jiménez-Díaz et al. 2017).

The genetic modification deals with the introduction of two or more genes encoding heterologous enzymes to create entirely new biosynthetic pathways or enabling new enzymatic activities of different feedstocks as energy sources. It might also knock out genes encoding enzymes in competing pathways and augment the flow of carbon into a desired pathway. The potential risks are unique to each type of microorganisms, such as in algae; it is possible to have effects on native populations in order to create or intensify and create a dangerous mutant (Jang et al. 2012; Mary 2011).

To obtain a befitting product, the use of various both prokaryotic and eukaryotic species and strains is common in industrial production. Some of the prevalent and focused microorganisms are yeast fungal and bacterial strains such as *Saccharomyces cerevisiae, Aspergillus* and *Trichoderma, Lactobacillus,* and strains of *Escherichia coli.* The existing strains always have a scope of improvement to have more productivity with genetic engineering for valuable properties (Rubin 2008).

The indication toward the improvisation of carbon fixation enhances pathway in proteins like RuBisCO or alteration in lipid synthesis in algae can help both with the environmental pollution reduction and biodiesel production. To ensure the safety conduct and industrial use of genetically modified organisms, they need to have appropriate risk assessment tools. The methods selected must express, enhance transporter proteins, maximize the carbon flow, and remove toxins and harmful compounds. The change in metabolism should emphasize easier cell lysis and making existing pathways more applicable for commercial purposes. For industrial purposes, fermentation of the microbes must be conducted in a protected environment and must prohibit any exposure or accidental release of the microorganism. Any inherent exposure of the genetically engineered organism that is to be produced commercially. Its exposure into the environment should be assessed for all possible hazards (Jiménez-Díaz et al. 2017; Chen and Dou 2016).

# **11.6 Regulations for Genetically Modifying Plants** and Microbes for Biofuel Production

To fulfill the current demand, the production of renewable fuels or bio-based chemicals is carried out with genetically modified microorganisms. The production includes microalgae, fungus, plants, and cyanobacteria. Researching and finding new biological methods of manufacturing renewable fuels from petrochemical feedstocks foresees a great potential toward more sustainable industrial activities. Without much speculation, genetically modified organisms exhibit certain advantages over other microbiological methods to obtain biofuel. Some of the factors of risk are summarized in Fig. 11.4.

Genetically modified organisms offer remodeled productivity, less operational costs, a variety of feedstocks, and most importantly, substantial carbon footprints. Genetically modified microorganisms (GMM) in most countries require regulatory guidelines before they could enter the market. The government has certain regulations when it comes to genetic engineering of plants and animals. In the late 1980s, risks caused by potential genetically engineered microorganisms to the environment were already getting huge attention. Early scientific reviews laid the foundation for regulatory risk assessments of proposed field tests.



Fig. 11.4 Factors that can influence the risk associated with genetically modifying an organism

EPA and USDA regulations govern the use of modified organisms for the production of fuels or chemicals in the US EPA works under the Toxic Substances Control Act (Glass 2015; Wozniak-Karczewska et al. 2019; Wozniak et al. 2012). The aim of these regulations is to declare it to the agency before merchandising any genetically modified microorganisms.

Ecologists have recommended that engineered strains can perform like non-engineered strains when they are introduced into new environments. This gives a fundamental estimation to predict and monitor the behavior using appropriate risk assessment tools (Viebahn and Chappin 2018). However, we cannot deny the fact that scientific concerns about the potential environmental effects of microorganisms with new traits are reasonable. Tribal Energy Resource Agreements and United States Environmental Protection agency ensure risks are minimized, and it is scalable to establish means of genetic manipulation. The choice of the reactor, its design, and features govern the choice of the starting culture and product.

In the 1990s, there was a standard that was set for the development of biodiesel in order to promote the use of alkyl esters-based biodiesel in pure form or as blends in automotive fuels. The standards vary in the USA and Europe. For the first time in 1999, the American Society for Testing and Materials (ASTM) adopted a provisional specification PS121 for biodiesel. ASTM D6751 was approved in 2002 for middle distillate fuels. ASTM D6751, 2012 onward defines two grades of biodiesel: grade 2-B and grade 1-B. The grades have a strict policy on monoglycerides and cold soak filterability derived from vegetable oils and animal fats. Two automotive standards for biodiesel/diesel fuel are ASTM D975 to allow up to 5% biodiesel to be blended into the fuel, and ASTM D7467, for biodiesel blends from B6 to B20.

In October 2003, the standard for biodiesel EN 14214 was accomplished in Europe for unblended FAME diesel fuel and some biodiesel blends. These standards

set the stepping stone in international standards and became the starting point for biodiesel specifications developed in other countries. Low-level blends are categorized by EN 590. For fatty acid methyl esters fused in diesel engines, EN 14214 makes the regulations.

A category B100 could be used unblended in a diesel engine or blended with diesel fuel to produce a blend conforming EN 590. The changes to cover heating oil applications were inducted by EN14214:2012 to cover blends up to B10. Mono-glycerides content was also considered as a separate class. EN 590 covers biodiesel/ diesel fuel blends up to B7. The version released in 2004 introduced blending up to 5% of fatty acid methyl ester (FAME) in diesel fuel, which was increased to 7% in 2009. The European biodiesel specification, EN 14214, a European indication that applies only to mono-alkyl esters made with *methanol*, specifies that ester content should be 96.5%, and no additives other than fatty acids can be added. Increased oxidation stability, reducing the sulfated ash limit to 0.005% from 0.02%, and limiting blends to B5 maximum are some of the guidelines for B100 used to make biodiesel/diesel fuel blends used all over the world (ACEA 2009; ASTM 2002; Tasios et al. 2013).

The work carried out in a stepwise and responsible manner can respond to the needs of developing novel sources of energy worldwide. Reducing carbon emissions and avoiding any harmful environmental impacts is the considerable factor that will control, manage, and constrain the genetic engineering of an organism for biofuel production.

Certain uses of GMM could be subjected to FDA regulations because of the production of foods, pharmaceuticals, or other products. However, a standard alternative for companies working with nonpathogenic microorganisms and obtaining ethanol, can be utilization of the excess and waste from the biomass in animal feed.

In the USA, animal feed ingredients are monitored by a non-profit organization that is called Association of American Feed Control Officials. They define if the ingredients are fit for animal feed and pet food.

#### 11.7 Future Outlook

# 11.7.1 Artificial Intelligence Can Help in Genetic Modification of Biofuels

As we struggle to find out a way to restore diminishing fossil fuel resources, the scientific community is working hard to find alternative sources of fuels; one such form is biofuel, through plantations of certain plants, which takes up to 90 days to grow and be ready processing the biofuel. The goal of the scientific community is to design a plant by a genetic modification that has the ability to produce a large amount of biofuel in less time.

At the molecular level, in a biofuel plant, a gene is responsible for the synthesis of the triglycerides (hydrocarbons). It is necessary to carry out research to identify this gene and isolate it (Rao and Pingali 2008), which might take time for the human eye to get the sequence of the gene. By leveraging Artificial Intelligence, it can be done quicker and with limited sources through AI Machine learning and Deep learning models.

Artificial Intelligence applications in biotechnology include analysis required in modification of genetics, drug screening, predictive modeling. These problems have been solved, which are difficult for humans to solve in a short period of time. AI now exists in every field of study, from recognizing a pattern, forecasting, DNA sequencing of tens of thousands of genes, and plays a crucial role in biotechnology (Klyuchko 2017).

#### 11.7.2 Machine Learning in Action

With the sophisticated machine learning models, we can achieve different clinical trial datasets, enable simulated screening, and analyze vast amounts of data. The basic concept connecting machine learning with artificial intelligence and deep learning is represented in Fig. 11.5. Apart from savings on clinical trial costs, with ML models, we can also gain exclusive insights and feed them back into the process. Machine Learning (ML) concept gives computers the ability to think and helps us to solve many problems. Machine learning is the part of artificial intelligence focused on algorithms which has the ability to learn from experience, and when exposed to new data, its accuracy is measured without explicit programming (Oliveira 2019; Kim et al. 2020).

The ML Algorithms take data as an input, and the output is predicted data or actions. The algorithms improve as they are exposed to more data.





As the Dataset grows, the performance of the ML model degrades and may tend to drop its accuracy. Hence, it will be difficult to just work with ML alone. The reasons are

• High dimensions:

When we have a large number of inputs and outputs, which are nothing but high dimensions, ML is not useful.

• Crucial AI problems:

Machine learning cannot solve natural language processing, image recognition problems due to their huge data amounts.

• Feature extraction:

For complex problems, such as object recognition, handwriting recognition, etc., ML will face a big challenge (Jordan and Mitchell 2015).

## 11.7.3 Deep Learning Methods

Deep learning is an approach of machine learning that has been designed based on the knowledge of the human brain and neurons, a simple depiction in Fig. 11.6. In recent years, deep learning has seen tremendous growth in its popularity and usefulness in the field of biotechnology (Sugomori et al. 2017).

Deep learning is a subset of machine learning methods based on artificial neural networks with multiple layers representation learning in different dimensions, which can be understood with Fig. 11.7.

Machine Learning can be supervised, semi-supervised, or unsupervised. The difference is in supervised learning, we provide labeled data, whereas unsupervised learning machine comes up with its own patterns without labels.

Deep learning algorithms use the concept of multilayer perceptron's. Data is filtered through multiple layers, with each consecutive layer using the output from the previous one to inform its results (Ardabili et al. 2020; Aghbashlo et al. 2021).

Deep learning models will achieve its accuracy level and can provide more data to process. If there is not enough data, then we can apply different deep learning techniques like data augmentation to generate more data for training our model, the more training we provide, algorithm will learn to perform better when unexposed





Fig. 11.7 Schematic representation of deep learning methods

data is tested, deep learning models learn from previous results to enhance their capability to make correlations and connections.

- DL algorithms are capable to focus on important features, and this can be achieved through minimal coding.
- DL models are capable to solve high dimensions.
- The main idea behind Deep Learning is to build learning algorithms that mimic biological neurons neural network system.
- Deep Learning is implemented through Neural Network.

Like many other disruptive technologies in Biotechnology, AI is generating much anticipation with Deep Learning's convolutional neural networks, which can work with minimum data as well. Also, a transfer learning model which gained information from a previously trained data can be applied to a new set of data, which can give more accurate results as it has seen certain features of data (Mosavi et al. 2020).

As per American physicist, Kaku and 3M Company (2011), we could be witnessing the next transition from transistors based on silicon to transistors based on atoms. In a decade, Moore's Law will slow down and computer power will level up, and we are going to many such transitions, one of them could be Quantum Computing.

This new field of Quantum Computing is now on the leading edge of computing. World's leading organizations like IBM, Google, etc., are working toward Quantum Supremacy.

As per MIT "One of the goals of quantum computation and quantum information is to develop tools which sharpen our intuition about quantum mechanics and make its predictions more transparent to human minds." The development of Quantum Machine Algorithms has begun for the genetic sample classification. Unlike classical computing, which is built on either 1 or 0, whereas quantum computing has both the positions at once, that is 1 and 0, and is based on Quantum mechanics (Li et al. 2001; Roe 1998).

There is a future of biotechnology when combined with quantum computing in deriving molecular level properties to modify the genetics of biofuels.

Acknowledgements The authors acknowledge the Fundação para a Ciência e a Tecnologia (FCT) through the project PTDC/BTA-GES/2740/2020\_NABIA. The Coimbra Chemistry Centre (CQC) is supported by the FCT through the projects UIDB/00313/2020 and UIDP/00313/2020. CDRSP is financed by national funds through the FCT/MCTES (UIDB/00481/2020 & UIDP/00481/2020). We are grateful for funding from PTScience which is supported through the programs CENTRO-05-4740-FSE-001526 and FEDER.

# References

- Abbas M, Arshad M, Hussain T, Iqbal M (2017) Enhanced ethanol production at commercial scale from molasses using high gravity technology by mutant S. cerevisiae. Braz J Microbiol 48(3): 403–409
- ACEA (2009) Biodiesel guidelines. European Automobile Manufacturers Association, Brussels
- Aghbashlo M, Peng W, Tabatabaei M, Kalogirou SA, Soltanian S, Hosseinzadeh-Bandbafha H, Mahian O, Lam SS (2021) Machine learning technology in biodiesel research: a review. Prog Energy Combust Sci 85:100904
- Amore A, Giacobbe S, Faraco V (2013) Regulation of Cellulase and Hemicellulase gene expression in fungi. Curr Genomics 14(4):230–249
- Ardabili S, Mosavi A, Mosavi A, Varkonyi-Koczy AR (2020) Systematic review of deep learning and machine learning models in biofuels research. Lecture Notes in Networks and Systems 101: 19–32
- Arumugam MS, Gon WS, Lee DS, Bae HJ (2008) Site-directed mutagenesis and CBM engineering of Cel5A (Thermotoga maritima). FEMS Microbiol Lett 287(2):205–211
- ASTM (2002) Standard specification for biodiesel fuel (B100) blend stock for distillate fuels. American Society for Testing and Materials, pp D6751–D6702
- Bhatia SK, Kim S-H, Yoon J-J, Yang Y-H (2017) Current status and strategies for second generation biofuel production using microbial systems. Energy Convers Manag 148:1142–1156
- Binod P, Gnansounou E, Sindhu R, Pandey A (2019) Enzymes for second generation biofuels: recent developments and future perspectives. Bioresour Technol Rep 5:317–325
- Castrillo M, Lucas-Salas LM, Rodriguez-Gil C, Martinez D (2013) High pH-induced flocculationsedimentation and effect of supernatant reuse on growth rate and lipid productivity of Scenedesmus obliquus and Chlorella vulgaris. Bioresour Technol 128:324–329
- Chen R, Dou J (2016) Biofuels and bio-based chemicals from lignocellulose: metabolic engineering strategies in strain development. Biotechnol Lett 38(2):213–221
- Cheng JJ, Timilsina GR (2011) Status and barriers of advanced biofuel technologies: a review. Renew Energy 36(12):3541–3549
- Da SNA, Srikrishnan S (2012) Introduction and expression of genes for metabolic engineering applications in Saccharomyces cerevisiae. FEMS Yeast Res 12(2):197–214
- Dellomonaco C, Fava F, Gonzalez R, SpringerLink (Online service) (2010) The path to next generation biofuels: successes and challenges in the era of synthetic biology. Microb Cell Factories 9(1):1–15
- Demirbas A (2010) Use of algae as biofuel sources. Energy Convers Manag 51(12):2738-2749

- Demirbas MF (2011) Biofuels from algae for sustainable development. Appl Energy 88(10): 3473–3480
- Druzhinina IS, Kubicek CP, Averous L, Blank LM, O'Connor K, Diaz E, Prieto A, Zimmermann W (January 01, 2017) Genetic engineering of Trichoderma reesei cellulases and their production. Microb Biotechnol 10:1485
- Eng T, Demling P, Herbert RA, Chen Y, Benites V, Martin J, Lipzen A, SpringerLink (Online service) (2018) Restoration of biofuel production levels and increased tolerance under ionic liquid stress is enabled by a mutation in the essential Escherichia coli gene cydC. Microbial Cell Factories 17(1):1–16
- Glass DJ (2015) Government regulation of the uses of genetically modified algae and other microorganisms in biofuel and bio-based chemical production. In: Algal biorefineries. Springer, Cham, pp 23–60
- Hilden L, Johansson G (2004) Recent developments on cellulases and carbohydrate-binding modules with cellulose affinity. Biotechnol Lett 26(22):1683–1693
- Himmel ME, Ding SY, Johnson DK, Adney WS, Nimlos MR, Brady JW, Foust TD (2007) Biomass recalcitrance: engineering plants and enzymes for biofuels production. Science (New York) 315(5813):804–807
- Hu W, Chen S, Yang Z, Liu L, Wang H (2011) Flexible electrically conductive nanocomposite membrane based on bacterial cellulose and polyaniline. J Phys Chem B 115(26):8453–8457
- Hu Y, Catchmark JM (2011) Integration of cellulases into bacterial cellulose: toward bioabsorbable cellulose composites. J Biomed Mater Res Part B Appl Biomater 1:114–123
- Huang H-J, Ramaswamy S, Al-Dajani W, Tschirner U, Cairneross RA (2009) Effect of biomass species and plant size on cellulosic ethanol: a comparative process and economic analysis. Biomass Bioenergy 33(2):234–246
- Ilmén M, Saloheimo A, Onnela ML, Penttilä ME (1997) Regulation of cellulase gene expression in the filamentous fungus Trichoderma reesei. Appl Environl Microbiol 63(4):1298–1306
- In Marcel, K. (2015). Biodiesel handbook
- Jang Y-S, Park JM, Choi S, Choi YJ, Seung DY, Cho JH, Lee SY (2012) Engineering of microorganisms for the production of biofuels and perspectives based on systems metabolic engineering approaches. Biotechnol Adv 30(5):989–1000
- Jiménez-Díaz L, Caballero A, Pérez-Hernández N, Segura A (2017) Microbial alkane production for jet fuel industry: motivation, state of the art and perspectives. Microb Biotechnol 10(1): 103–124
- Johnston M, Flick JS, Pexton T (1994) Multiple mechanisms provide rapid and stringent glucose repression of GAL gene expression in Saccharomyces cerevisiae. Mol Cell Biol. 14:3834
- Jordan MI, Mitchell TM (2015) Machine learning: trends, perspectives, and prospects. Science 349(6245):255-260
- Kaku M, 3M Company (2011) Visions: how science will revolutionize the 21st century. In: Place of publication not identified. Knopf Doubleday Publishing Group
- Keasling JD, Chou H, Kuk LS, Ham TS, Soon LT, Physical Biosciences Division (2009) Metabolic engineering of microorganisms for biofuels production: from bugs to synthetic biology to fuels. Lawrence Berkeley National Laboratory, Berkeley, CA
- Kenney K, Idaho National Laboratory (INL) (2007) Cellulosic biomass feedstocks and logistics for ethanol production. Biofuels Bioprod Biorefin 1:3
- Kim GB, Kim WJ, Kim HU, Lee SY (2020) Machine learning applications in systems metabolic engineering. Curr Opin Biotechnol 64:1–9
- Klyuchko OM (2017) Application of artificial neural networks method in biotechnology. Biotechnologia Acta 10(4):5–13
- Kotchoni OS, Shonukan OO, Gachomo WE (2003) Bacillus pumilus BpCRI 6, a promising candidate for cellulase production under conditions of catabolite repression. Afr J Biotechnol 2(6):140–146. Academic Journals (Kenya)
- Kraan S (2013) Mass-cultivation of carbohydrate rich macroalgae, a possible solution for sustainable biofuel production. Mitigat Adaptat Strateg Global Change 18(1):27–46

- Kuhad RC, Gupta R, Singh A (2011) Microbial cellulases and their industrial applications. Enzyme Res 2011:1
- Kuhad RC, Kumar M, Singh A (1994) A hypercellulolytic mutant of fusarium oxysporum. Lett Appl Microbiol 19(5):397–400
- Lal R (2005) World crop residues production and implications of its use as a biofuel. Environ Int 31(4):575–584
- Larson ED, United Nations Conference on Trade and Development (2008) Biofuel production technologies: status, prospects and implications for trade and development. United Nations Conference on Trade and Development, New York
- Lazarus M, Van AH (2018) Fossil fuel supply and climate policy: exploring the road less taken. Clim Chang 150:1–13
- Li S-S, Long G-L, Bai F-S, Feng S-L, Zheng H-Z (2001) Quantum Computing. Proc Natl Acad Sci U S A 98(21):11847–11848
- Lima MAP, Natalense APP, Institute for New Technologies (Maastricht, Netherlands) (2012) Bioethanol. In: Place of publication not identified. INTECH
- Liu C-G, Xiao Y, Xia X-X, Zhao X-Q, Peng L, Srinophakun P, Bai F-W (2019) Cellulosic ethanol production: Progress, challenges and strategies for solutions. Biotechnol Adv 37(3):491–504
- Liu X, Saydah B, Eranki P, Colosi LM, Greg MB, Rhodes J, Clarens AF (2013a) Pilot-scale data provide enhanced estimates of the life cycle energy and emissions profile of algae biofuels produced via hydrothermal liquefaction. Bioresour Technol 148:163–171
- Liu G, Zhang L, Wei X, Zou G, Qin Y, Ma L, Li J, Qu Y (2013b) Genomic and secretomic analyses reveal unique features of the lignocellulolytic enzyme system of Penicillium decumbens. PLoS One 8:2
- Mach R, Zeilinger S (2003) Regulation of gene expression in industrial fungi: Trichoderma. Appl Microbiol Biotechnol 60(5):515–522
- Margolles-clark E, Ihnen M, Penttilä M (1997) Expression patterns of ten hemicellulase genes of the filamentous fungus Trichoderma reesei on various carbon sources. J Biotechnol 57(1): 167–179
- Mary JD (2011) Engineering microbes for tolerance to next-generation biofuels. Biotechnol Biofuels 4(1):32
- Masjuki HH, Md AK, International Conference on Thermal Engineering (2012) An overview of biofuel as a renewable energy source: development and challenges. Proc Eng 56:39–53
- Mayorga-Reyes L, Ponce-Noyola T (1998) Isolation of a Hyperxylanolytic Cellulomonas Flavigena mutant growing on continuous culture on sugar cane bagasse. Biotechnol Lett 20(5):443–446
- Mehtani J, Arora N, Patel A, Jain P, Pruthi PA, Poluri KM, Pruthi V (2017) Augmented lipid accumulation in ethyl methyl sulphonate mutants of oleaginous microalga for biodiesel production. Bioresour Technol 242:121–127
- Menon V, Rao M (2012) Trends in bioconversion of lignocellulose: biofuels, platform chemicals & biorefinery concept. Prog Energy Combust Sci 38(4):522–550
- Mosavi A, Mosavi A, Ardabili S, Varkonyi-Koczy AR (2020) List of deep learning models. Lecture Notes in Networks and Systems 101:202–214
- Mythili S, Rao A, Sathiavelu A (2016) Genetic engineering in BioButanol production and tolerance. Braz Arch Biol Technol 59(0). https://doi.org/10.1590/1678-4324-2016150612
- Nehring R (2009) Traversing the mountaintop: world fossil fuel production to 2050. Philos Transact Biol Sci 364(1532):3067–3079
- Oliveira AL (2019) Biotechnology, big data and artificial intelligence. Biotechnol J 14:8
- Ottenheim C, Werner KA, Zimmermann W, Wu JC (2015) Improved endoxylanase production and colony morphology of aspergillus Niger DSM 26641 by γ-ray induced mutagenesis. Biochem Eng J 94:9–14
- Peralta-Yahya PP, Zhang F, del Cardayre SB, Keasling JD (2012) Microbial engineering for the production of advanced biofuels. Nature 488(7411):320–328

- Peterson R, Nevalainen H (2012) Trichoderma reesei RUT-C30 thirty years of strain improvement. Microbiology 158(1):58–68
- Pfaltzgraff LA, Clark JH (2014) Green chemistry, biorefineries and second generation strategies for re-use of waste: an overview. Adv Biorefine 2014:3–33
- Qi F, Pei H, Hu W, Mu R, Zhang S (2016) Characterization of a microalgal mutant for CO2 biofixation and biofuel production. Energy Convers Manag 122:344–349
- Rao P, Pingali S (2008) Sustainable energy generation. In: 2008 IEEE International Symposium on Electronics and the Environment, pp 1–4
- Rodionova MV, Poudyal RS, Tiwari I, Voloshin RA, Zharmukhamedov SK, Nam HG, Zayadan BK, Allakhverdiev SI (2017) Biofuel production: challenges and opportunities. Int J Hydrog Energy 42(12):8450–8461
- Roe P (1998) Explorations in Quantum Computing. Aiaa Journal 36(11):2152
- Rubin EM (2008) Genomics of cellulosic biofuels. Nature London 454(7206):841-845
- Sainz MB (2009) Commercial cellulosic ethanol: the role of plant-expressed enzymes. In Vitro Cellular Developl Biol Plant 45(3):314–329
- Sarkar N, Ghosh SK, Bannerjee S, Aikat K (2012) Bioethanol production from agricultural wastes: an overview. Renew Energy 37(1):19–27
- Schuster A, Schmoll M (2010) Biology and biotechnology of Trichoderma. Appl Microbiol Biotechnol 87(3):787–799
- Sharma HK, Xu C, Qin W, SpringerLink (Online service) (2017) Biological pretreatment of lignocellulosic biomass for biofuels and bioproducts: an overview. Waste Biomass Valorizat 10(2):235–251
- Sims RE, Mabee W, Saddler JN, Taylor M (2010) An overview of second generation biofuel technologies. Bioresour Technol 101(6):1570–1580
- Siqueira JGW, Rodrigues C, Vandenberghe LPDS, Woiciechowski AL, Soccol CR (2020) Current advances in on-site cellulase production and application on lignocellulosic biomass conversion to biofuels: a review. Biomass Bioenergy 132:105419
- Sriprang R, Asano K, Gobsuk J, Tanapongpipat S, Champreda V, Eurwilaichitr L (2006) Improvement of thermostability of fungal xylanase by using site-directed mutagenesis. J Biotechnol 126(4):454–462
- Srivastava N, Mishra PK, Upadhyay SN (2020) Industrial enzymes for biofuels production: recent updates and future trends. Elsevier
- Srivastava N, Srivastava M, Mishra PK, Singh P, Ramteke PW (2015) Application of Cellulases in biofuels industries: an overview. J Biofuels Bioenergy 1(1):55
- Sticklen MB (2008) Plant genetic engineering for biofuel production: towards affordable cellulosic ethanol. Nat Rev Genet 9(6):433–443
- Sugomori Y, Kaluza B, Soares FM, Souza AMF (2017) Deep learning. Nature 521(7553):436-444
- Tasios N, Apostolaki E, Capros P, De VA (2013) Analyzing the bio-energy supply system in the context of the 20-20-20 targets and the 2050 decarbonization targets in the EU. Biofuels Bioprod Biorefin 7(2):126–146
- Van VP, Ruijter GJ, Visser J (1995) An extreme creA mutation in aspergillus nidulans has severe effects on D-glucose utilization. Microbiology 141:2301–2306
- Verma N, Kumar V, Bansal MC (2021) Valorization of waste biomass in fermentative production of Cellulases: a review. Waste Biomass Valoriz 12(2):613–640
- Viebahn P, Chappin EJL (2018) Scrutinising the gap between the expected and actual deployment of carbon capture and storage: a bibliometric analysis. Energies 11(9):2319
- Volynets B, Dahman Y (2011) Assessment of pretreatments and enzymatic hydrolysis of wheat straw as a sugar source for bioprocess industry. Int J Energy Environ 2:427–446
- Wang H, Jones RW (1997) Site-directed mutagenesis of a fungal –1,4-endoglucanase increases the minimum size required for the substrate. Appl Microbiol Biotechnol 48(2):225–231
- Wei N, Quarterman J, Jin Y-S (2013) Marine macroalgae: an untapped resource for producing fuels and chemicals. Trends Biotechnol 31(2):70–77
- Wilson DB (2009) Cellulases and biofuels. Curr Opin Biotechnol 20(3):295-299

- Wozniak CA, McClung G, Gagliardi J, Segal M, Matthews K (2012) Regulation of genetically engineered microorganisms under FIFRA. FFDCA and TSCA:57–94
- Wozniak-Karczewska M, Lisiecki P, Lawniczak C, Bialas W, Owsianiak M, Piotrowska-Cyplik A, Gutierrez T (2019) Effect of bioaugmentation on long-term biodegradation of diesel/biodiesel blends in soil microcosms. Sci Total Environ 671:948–958
- Xiros C, Topakas E, Christakopoulos P (2013) Hydrolysis and fermentation for cellulosic ethanol production. Wiley Interdiscipl Rev Energy Environ 2(6):633–654
- Zheng Y, Pan Z, Zhang R (2009) Overview of biomass pretreatment for cellulosic ethanol production. Int J Agric Biol Eng 2(3):51-68
- Zou C, Zhao Q, Zhang G, Xiong B (2016) Energy revolution: from a fossil energy era to a new energy era. Nat Gas Ind B 3(1):1-11

# Chapter 12 "Omics" Technologies in Biodegradation Processes



Sevcan Aydin and Mahmut Çalışkan

**Abstract** The considerable environmental impact of pollution due to human activities is driving the development of new decontamination and clean-up methods. Hereby, the interactions between the various microbial communities at polluted sites is receiving increased research attention, with the novel omics approaches, opening up new ways to study bioremediation pathways and their mechanisms. This has allowed innovation in the field of new bioremediation as an effective way to combat pollution. In particular, the omics approaches offer great potential to predict the metabolic processes of microbes in environments experiencing pollution. The highperformance analyses provided by these approaches can, amongst others, help track novel organisms for use in bioremediation and offer new, high-quality insights into those molecular pathways that are critical to biodegradation. In short, through multiomics approaches, the field of bioremediation will benefit from the establishment of new theory-based methodologies to mitigate pollution.

**Keywords** Bioremediation · Biodegradation · Microorganisms · Metagenomics · Genetic analysis · Horizontal gene transfer

# 12.1 Introduction

Due to population growth and the accompanying increase in industrial activities, pollution is rising across the world. The growth of industrialization in particular has led to manufacturing processes that produce and incorporate various chemicals for use in high-tech products, including heavy metals, hydrocarbons, and xenobiotics. These substances are not only toxic but also persist in the environment, and thus have a significant adverse effect on ecosystems and organisms. However, there are

e-mail: sevcan.aydin@istanbul.edu.tr

S. Aydin (🖂) · M. Çalışkan

Division of Biotechnology, Biology Department, Faculty of Science, Istanbul University, Istanbul, Turkey

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_12

several natural methods that can address these imbalances, with microorganisms representing an effective natural tool with which to remove toxic compounds from contaminated sites. Microbial-mediated biodegradation therefore has considerable potential to naturally restore polluted ecosystems (Shahi et al. 2016a; Aydin et al. 2017; Nwankwo et al. 2021; Sales et al. 2021).

The microbial ecology of an environment, whether pristine or anthropogenically affected, can be assessed using culture-based methods. However, as many of the identified microorganisms of interest to research are not culturable in a laboratory, there remain several challenges in their study. The research trying to culture such microbial communities has so far only been successful with no more than 1% of any given sample of prokaryotes. Moreover, there remains a gap in the knowledge of which factors influence the microbial communities of polluted environments, e.g. their metabolism, growth, or dynamics. Of the abovementioned omics approaches, metagenomics offers a way to conduct genome-level studies of microbial communities, providing substantial insights into the so-called uncultured microbiota (Aydin et al. 2015). The other omics approaches, such as genomics, transcriptomics, proteomics, and metabolomics, have recently also begun to offer an array of advanced techniques designed to treat pollutants in a way that causes no further damage to the environment (Aydin et al. 2022; Malik et al. 2021; Wright et al. 2021).

This chapter sets out to explore to what extent omics techniques can be applied to monitor the processes of biodegradation. In this context, it proposes using a community systems biology approach, integrating the abovementioned multi-omics with bioinformatics as well as simulation tools. The aim hereby is to enhance the predictive ability of models regarding the persistence of toxic chemicals in environmental settings, which should be taken into account and integrated into a tiered assessment strategy regarding the persistence of toxic chemicals. The findings of the research should offer an enhanced understanding of how the composition of the microbial community relates to the catabolic potential and environmental conditions as well as the properties of the chemical under investigation.

# 12.2 Omics Approaches for Monitoring Biodegradation Processes

Pollution continues to be both a tenacious problem and a significant threat to the health of humans and the environment. While numerous strategies have been applied to address this issue, it is still a challenge, even in terms of simple monitoring, and environmental contamination continues on a global scale. The severity of the issue calls for novel pollution mitigation strategies to ensure environmental and human health, with bioremediation being one of the most promising to emerge in recent years. Bioremediation using microorganisms is receiving increasing research interest, with studies striving to identify ways to effectively restore environments affected

by pollution (Shahi et al. 2016b). Microorganisms present themselves as particularly suitable as they can exist in a variety of environments, even degraded ones, by producing metabolites that have the ability to degrade or even transform contaminants, thereby allowing polluted sites to be naturally restored (Kour et al. 2021). The design of effective strategies hereby depends on modern omics technologies, including metagenomics, proteomics, and transcriptomics, which enable the microbiota's diversity and ecology to be studied to facilitate their implementation in the context of environmental monitoring as well as bioremediation (Aydin 2016).

Strategies based on bioremediation or biotransformation aim to exploit the natural diverse catabolic abilities of microorganisms regarding the degradation, accumulation, or transformation of multitudinous environmentally harmful compounds, such as pharmaceutical substances, heavy metals and polyaromatic hydrocarbons (PAHs). Recent developments in next-generation sequencing (NGS) have enabled key microorganisms to be genomically, metagenomically, and bioinformatically analyzed, allowing hitherto incomprehensible biodegradative pathways to be understood (Shahi et al. 2016c; Tancsics et al. 2021). Similarly, horizontal gene transfer (HGT) is crucial to the processes of microbial biodegradation in that it affects the activities of the microbiota. Specifically, bioremediation relies on functional genes to be transferred between key members of the microbial community. As the availability of macronutrients, among various other abiotic factors, influence HGT in polluted settings, the microbiota's capacity for biodegradation is dissimilar between sites. Hence, a thorough knowledge of which mobile elements and functional genes lead to HGT would further aid the potential for the bioremediation of a polluted area. In addition, as the biodegradation abilities of the microbial community dictate to what extent HGT can occur in a contaminated site, assessing HGT can be employed to monitor bioremediation to evaluate the biodegradation process (Shahi et al. 2017).

The prior research has evidenced a significant correlation between HGT and the existence of functional genes (French et al. 2020). For instance, in their investigation into the horizontal transfer of the alkB, nah, and phnAc genes in the biostimulation of soils contaminated by petroleum, Shahi et al. (2016b) found that the alkB and phnAc genes transferred when nutrients were highly available. They also demonstrated that HGT was positively linked to nutrient content, with an increase in the carbon to nitrogen ratio from 100:5 to 100:15 correlating with increased rates of HGT. Building on this, Shahi et al. (2016a, b, c) showed that the evaluation of the microbial population and the presence of functional genes could be used to assess the efficiency of existing bioremediation efforts. In this study, they were successful in showing that HGT could serve as a suitable monitoring approach to measure the effectiveness of bioremediation for soils that had been contaminated by petroleum. They also found that this method could further be utilized to assess contaminated sites' potential for biodegradation.

## 12.3 Conclusion

The combined effects of globalization leading to increased industrialization, population growth, and consumption-based lifestyles have caused non-degradable pollutants to persist in the environment, adversely affecting human health. Due to their environmentally and cost-effective characteristics, methods based on bioremediation are opening up new avenues to mitigate these pollutants beyond conventional physicochemical treatments. Almost all polluted environments contain microbial strains that are capable of degrading the contaminants specific to those environments, with the population density increasing with the increasing presence of such contaminants. As the metabolic effectiveness of the microbiota relies on HGT between microorganisms, choosing the right gene for the degradation process and then monitoring how this gene is horizontally transferred comprise a feasible strategy to assess the effectiveness of bioremediation. The bioremediation potential of specific sites can hereby also be assessed. The necessary information on the genes as well as their genomic structure, metabolic and biological pathways, functions, and evolution can be gathered through the omics approaches. Moreover, a functional characterization of the relevant genes, proteins, coding regions, and products of the metabolism should be conducted to quantify the relevant biological processes of the microbiota. Recent developments in the omics fields, especially genomics, proteomics, metabolomics, transcriptomics, and interactomics, thus provide an array of technical techniques and expertise in this respect.

Acknowledgement The authors are grateful for the financial support from TUBITAK (Turkish Association of Science and Technology) (No: 116Y096).

## References

- Aydin S (2016) Microbial sequencing methods for monitoring of anaerobic treatment of antibiotics to optimize performance and prevent system failure. Appl Microbiol Biotechnol 100(12): 5313–5321
- Aydin S, Ince B, Ince O (2015) Development of antibiotic resistance genes in microbial communities during long-term operation of anaerobic reactors in the treatment of pharmaceutical wastewater. Water Res 83:337–344
- Aydin S, Karaçay HA, Shahi A, Gökçe S, Ince B, Ince O (2017) Aerobic and anaerobic fungal metabolism and omics insights for increasing polycyclic aromatic hydrocarbons biodegradation. Fungal Biol Rev 31(2):61–72
- Aydin S, Ünlü İD, Arabacı DN, Duru ÖA (2022) Evaluating the effect of microalga Haematococcus pluvialis bioaugmentation on aerobic membrane bioreactor in terms of performance, membrane fouling and microbial community structure. Sci Total Environ 807:149908
- French KE, Zhou Z, Terry N (2020) Horizontal 'gene drives' harness indigenous bacteria for bioremediation. Sci Rep 10(1):1–11
- Kour D, Kaur T, Devi R, Yadav A, Singh M, Joshi D et al (2021) Beneficial microbiomes for bioremediation of diverse contaminated environments for environmental sustainability: present status and future challenges. Environ Sci Pollut Res 28:1–23

- Malik G, Arora R, Chaturvedi R, Paul MS (2021) Implementation of genetic engineering and novel omics approaches to enhance bioremediation: a focused review. Bull Environ Contam Toxicol 108:1–8
- Nwankwo W, Adetunji CO, Ukhurebor KE, Panpatte DG, Makinde AS, Hefft DI (2021) Recent advances in application of microbial enzymes for biodegradation of waste and hazardous waste material. In: Microbial rejuvenation of polluted environment, pp 35–56
- Sales JCS, Santos AG, de Castro AM, Coelho MAZ (2021) A critical view on the technology readiness level (TRL) of microbial plastics biodegradation. World J Microbiol Biotechnol 37(7): 1–15
- Shahi A, Aydin S, Ince B, Ince O (2016a) The effects of white-rot fungi Trametes versicolor and Bjerkandera adusta on microbial community structure and functional genes during the bioaugmentation process following biostimulation practice of petroleum contaminated soil. Int Biodeterior Biodegradation 114:67–74
- Shahi A, Aydin S, Ince B, Ince O (2016b) Evaluation of microbial population and functional genes during the bioremediation of petroleum-contaminated soil as an effective monitoring approach. Ecotoxicol Environ Saf 125:153–160
- Shahi A, Aydin S, Ince B, Ince O (2016c) Reconstruction of bacterial community structure and variation for enhanced petroleum hydrocarbons degradation through biostimulation of oil contaminated soil. Chem Eng J 306:60–66
- Shahi A, Ince B, Aydin S, Ince O (2017) Assessment of the horizontal transfer of functional genes as a suitable approach for evaluation of the bioremediation potential of petroleum-contaminated sites: a mini-review. Appl Microbiol Biotechnol 101(11):4341–4348
- Tancsics A, Li M, Chrzanowski L (2021) New insights into the biodegradation of organic contaminants in subsurface ecosystems: approaches and achievements of the multiomics era. Front Microbiol 12:946
- Wright RJ, Bosch R, Langille MG, Gibson MI, Christie-Oleza JA (2021) A multi-OMIC characterisation of biodegradation and microbial community succession within the PET plastisphere. Microbiome 9(1):1–22

# Chapter 13 Conclusion: Environmental Protection— Our Common Responsibility



Alberto A. C. C. Pais and Telma Encarnação

**Abstract** Environmental pollution is increasing globally and, together with climate change, is a priority on the environmental, political, business, and scientific agendas. Air, land, and water pollution have an impact on all ecosystems and our lives and can jeopardize our future and future generations.

The importance of policies on public awareness and perception is recognized and can have an effective role in the protection of the environment. Policymakers, companies and industries, civil society, scientists, all sectors of society should be involved for the same purpose; coordinated efforts at an international level are needed to tackle all the challenges planet Earth face.

Therefore, it is crucial to stimulate the discourse, narrative, and debate about environmental pollution and degradation and mitigation strategies.

**Keywords** Environment · Marine pollution · Civil society · Policies · Public awareness · Climate challenge

Approximately 80% of marine pollution originates in dry land. To address this mighty problem, a decisive coordinated policy is needed, and the UN Environment Programme contributes by hosting a specific initiative, the Global Programme of Action for the Protection of the Marine Environment from Land-based Activities (GPA). After the Manila Declaration of 2012, GPA targeted three priority pollution sources: marine litter, nutrient management, and wastewater through global

A. A. C. C. Pais

T. Encarnação (🖂)

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

Centre for Rapid and Sustainable Product Development, Polytechnic Institute of Leiria, Marinha Grande, Portugal

PTScience Avenida do Atlântico, Lisbon, Portugal e-mail: tencarnacao@qui.uc.pt

to Environmental Pollution?, Environmental Challenges and Solutions, https://doi.org/10.1007/978-3-031-17226-7\_13

CQC-IMS, Department of Chemistry, University of Coimbra, Coimbra, Portugal

<sup>©</sup> The Author(s), under exclusive license to Springer Nature Switzerland AG 2023

T. Encarnação, A. Canelas Pais (eds.), Marine Organisms: A Solution

voluntary multi-stakeholder partnerships of governments, intergovernmental agencies, academia, the private sector, and civil society.

Various other international organizations of different types have promoted the development of actions and strategies to achieve sustainability and avoid degradation in marine environments. Typically, such strategies rely on information from expert groups about the causes of degradation and the policy options portfolio to tackle them. These strategies rarely consider trustworthy data on public awareness, their concerns, and priorities. Recent results (Gelcich et al. 2014) show that the level of concern regarding marine impacts is closely associated with the level of public acquired information, with pollution and overfishing being two areas prioritized by the public for policy development. Results also suggest that the overall public understand human generated impacts on marine and are highly concerned about ocean pollution, overfishing, and ocean acidification. Promoting further public awareness, concerns, and priorities can enable scientists and funders to understand how the public relates to marine environments and make the much-needed correspondence between policy and public demand (Gelcich et al. 2014).

Naturally, the vastly comprehensive nature of the social involvement necessary to tackle this massive scale problem implies taking into consideration the importance of awareness raising campaigns. It has been shown (Latinopoulos et al. 2018) that, when environmental knowledge is limited, public information campaigns can be effectively used to help individuals make more informed choices. Also, public information campaigns have a positive effect on the willing to pay (WTP) estimates.

A further problem is that situations are heterogenous, in terms of gravity and characteristics. The way they are seen by the public also varies. This was recently assessed (Brouwer et al. 2017) in terms of the social costs of both marine debris washed ashore and litter left behind by beach visitors along different European coasts. The social costs were estimated based on public perception. Previous studies focusing on the valuation of beach recreation did not make a distinction between pollution sources. This latter distinction is considered important in view of the fact that a large share of the litter originates from beach visitors and requires another type of coastal zone policy intervention than diffuse pollution washed ashore. Assessing how responsible beachgoers feel for the presence of beach litter they partly leave behind themselves and to what extent they are willing to pay for the clean-up of this litter compared to litter washed ashore provides important information for priority setting in coastal policy and management.

Naturally, objective means for marine environment monitoring are also required (Xu et al. 2019). During the past two decades, advanced information and communication technologies have been applied to the development of various marine environment monitoring systems. Among others, the Internet of Things (IoT) has been playing an important role in this area. New technologies including advanced Big Data analytics and their applications in this area have also been introduced. These types of data must be associated to relevant metrics (Roberts et al. 2018) so that evolution of contaminated, endangered, or protected sites is recorded. A new set of metrics has been recently suggested focusing on marine biodiversity in protected

sites. This type of approach also permits to compare networks of protected sites, so as to assess the local and overall level of protection.

It cannot, however, be forgotten that the threats posed by different sources of pollutants still persist upon marine environments, and that these pollutants can be detected even in remote polar regions. A further threat arises from climate change causing sea level rise and shifts and changes in flora and fauna of certain sea areas. Thus, policy prompting environmental awareness must be based on a global perspective in this vastly interconnected Earth.

### References

- Brouwer R, Hadzhiyska D, Ioakeimidis C, Ouderdorp H (2017) The social costs of marine litter along European coasts. Ocean Coastal Manag 138:38–49
- Gelcich S, Buckley P, Pinnegar JK, Chilvers J, Lorenzoni I, Terry G, Guerrero M, Castilla JC, Valdebenito A, Duarte CM (2014) Public awareness, concerns, and priorities about anthropogenic impacts on marine environments. PNAS 111:15042–15047
- Latinopoulos D, Mentis C, Bithas K (2018) The impact of a public information campaign on preferences for marine environmental protection. The case of plastic waste. Mar Pollut Bull 131: 151–162
- Roberts KE, Valkan RS, Cook CN (2018) Measuring progress in marine protection: a new set of metrics to evaluate the strength of marine protected area networks. Biol Conserv 219:20–27
- Xu G, Shi Y, Sun X, Shen W (2019) Internet of Things in marine environment monitoring: a review. Sensors 19:1711–1731