# Chapter 1 Nanotechnologies in Aquatic Disease Diagnosis and Drug Delivery



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# Abbreviations

ALP	Alkaline phosphatase
CNTs	Carbon nanotubes
DDS	Drug delivery system
DNA	Deoxyribonucleic acid
EMA	European Medicine Agency
FAT	Fluorescence antibody test
GCRC	Grass carp reovirus
GO	Graphene oxide
HK	Head kidney
IFAT	Indirect fluorescence antibody test
IHC	Immunohistochemistry
IHNV	Infectious hematopoietic necrosis virus
IM	Intramuscular
IP	Intraperitoneal
IPNV	Infectious pancreatic necrosis virus
ISAV	Infectious salmon anaemia virus
KHV	Koi herpes virus
LCDV	Lymphocystis disease virus
LPS	Lipopolysaccharides
NP's	Nanoparticles
PCR	Polymerase chain reaction
PLA	Poly lactic acid
PLGA	Poly (lactic-co-glycolic acid)
RAPD	Random amplification of polymorphic DNA
rcb-PCR	Reverse cross blot PCR
RNA	Ribonucleic acid
ROS	Reactive oxygen species

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V. K. Arivarasan et al. (eds.), *Nanotechnological Approaches to the Advancement of Innovations in Aquaculture*, Nanotechnology in the Life Sciences, https://doi.org/10.1007/978-3-031-15519-2\_1

RT-PCR	Reverse transcriptase polymerase chain reaction
SLN	Solid lipid nanoparticles
TLRs	Toll-like receptors
VHSV	Viral haemorrhagic septicaemia virus
WSSV	White spot syndrome virus

# 1.1 Introduction

Aquaculture represents the fastest-growing domain with prolific growth potential. It serves as the source of easily digestible protein, healthy fat, and a cache of various essential micronutrients for billions across the globe. Aquaculture provides a potential platform for rural employment and livelihood development, possesses high export earning potential, and can support national GDP through income generation. Recent years have seen development in the production of farmed fin and shellfish that has increased rapidly due to enhanced, intensive aquaculture practices (FAO 2018) and demand. Due to the increase in intensive farming systems, the need for wild seed stocks as well as fish supplies has declined (Naylor et al. 2006). A report from the Food and Agriculture Organization shows that the average annual consumption of fish increased by 3.2% globally from 1961 to 2016. In 2015, of the total animal protein consumed globally, fish food dominated at 17% and supplied almost 20% of the average consumption of animal protein for 3.2 billion populations (FAO 2018). However, environmental factors like excessive accumulation of nutrients result in the eutrophication of ponds due to intensive aquaculture practices, which are being looked at as a block in aquaculture growth dynamics (Herbeck et al. 2013). Along with eutrophication, climate change also poses a serious threat to aquaculture due to the temperature rise, increased methane and carbon dioxide emissions, etc. (FAO 2018). An increase in water temperature will have an impact on plankton communities, the survival of larvae and juveniles, and the reproductive ability of fishes. It has been also observed that contaminated water is being introduced into reservoirs and fish culture systems (Patel et al. 2019). Moreover, detecting pollution and toxic levels in water is a tedious and time-consuming process (Altenburger et al. 2019). In extensive aquaculture practices, nutritional deficiency is common among juvenile and broodfish populations, and health management is also another challenging problem. Depleting environmental quality and the impact of climate change have led to a surge in infectious diseases in aquaculture. Diseases like fin and gill rot and epizootic ulcerative syndrome (EUS) are still prevalent with no possible solutions and become incurable with antibiotics.

A white spot disease outbreak has inhibited the otherwise profitable *Penaeus monodon* culture system (Zhang et al. 2016). It has been estimated that the global shrimp industry has faced a loss of US\$10 billion from white spot disease syndrome and the infectious myonecrotic virus alone since 1990. Meanwhile, the \$15 billion global ornamental fish business also faced a problem with antibiotic resistance, which created uncertainty in its therapeutic practices. Conventional drug delivery

models have low efficacy and bioavailability in aquatic ecosystems. Besides these, traditional disease detection methods are also not feasible in the long run.

At this point, there is a need for some innovative technological interventions to get rid of these problems in aquaculture. Nanotechnology has emerged as a promising solution, leading to new possibilities by developing and applying materials at nanoscale dimensions (1-100 nm) with unique properties, paving the way for novel therapeutic as well as diagnostic applications (Matteucci et al. 2018; Mohamed et al. 2015). The application of nanotechnology has become a part of human life with multiple value-added products (Stern et al. 2016; Vinay et al. 2018). With recent technologies, nanoparticles are being prepared by different physical and chemical methods. The main commercial nanoparticles include silver, gold, oxides of zinc, iron, calcium, titanium, and manganese, as well as carbon nanotubes (CNTs), mesoporous silica nanoparticles, quantum dots (Khan et al. 2019; Stone et al. 2010), etc. With complex three-dimension technology, 'Second-generation' nanomaterials are now being synthesized with highly functional surfaces (Sarkar et al. 2022; Jeevanandam et al. 2018; Ju-Nam and Lead 2008). Even the impact of toxic pollutants on aquatic species can be intervened by using nanotechnology. Besides conventional toxicological assays, scientists are also using cellular cytotoxicity, apoptosis as well as bioinformatics-based interactive tools to narrate nanotoxicity and its remediation by applying green nanoparticles in bacteria and fish models (Kumari et al. 2017; Husain et al. 2021; Verma et al. 2017, 2021). Hence, the efficient application of nanotechnology can be addressed for maximizing aquaculture productivity and a schematic diagram is presented in this regard as Fig. 1.1.

### 1.2 Fish Disease and Immune System

The disease is a primary cause of fish mortality, especially when fishes are in the larval stage. There are two types of fish diseases according to their infectious pattern, viz. pathogenic diseases and non-pathogenic diseases (Arfat et al. 2014; Vevers and Jha 2008). Different infectious diseases are the main cause of death that might be due to bacterial, viral, or parasitic infection, and sometimes pathogens cause high levels of toxicity. It is mainly related to poor water quality management, inadequate feed supply, etc. A high-nutrition diet will have a positive effect on fish health. In this way, nanoparticles serve as a main source of transportation as well as supplementation of materials across the membrane. Non-infectious diseases are mainly caused by gas bubble diseases due to extensive aeration, nutritional diseases due to deficiency of nutrients such as vitamins and minerals, disorders caused by pollutants in the form of agricultural and industrial wastes, and neo-plastic and genetic anomalies resulting in abnormal growth which makes organs lose their structure and function (Rajkumar et al. 2016; Gao et al. 2017; Vinay et al. 2018; Bacchetta et al. 2017; Verma et al. 2020).

The fish immune system plays an important role in the protection against pathogens. Under the infectious stage, the immune system can detect pathogens

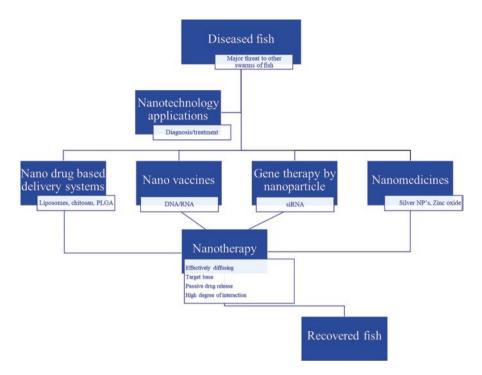


Fig. 1.1 Application of nanotechnology in aquaculture

(Firdaus-Nawi and Saad 2016). The immune system of fish is composed of two main components: innate and adaptive immunities (Firdaus-Nawi and Saad 2016). Innate immunity is nonspecific and serves as the primary defence against pathogen invasion, while adaptive immunity is much more specific to a particular pathogen. Innate immunity is composed of nonspecific cellular and humoral elements. The nonspecific cellular portion consists of toll-like receptors (TLRs), macrophages, neutrophils, eosinophils, and nonspecific cytotoxic cells, while the nonspecific humoral portion includes lysozyme, complement cells, interferons, C-reactive proteins, transferrins, and lectins, where they work together to prevent pathogen invasion at the primary stage of infection (Firdaus-Nawi and Saad 2016). On the other hand, the adaptive immune system comprises highly specific systemic cells and mechanisms that are divided into two major components: humoral and cellular (Firdaus-Nawi and Saad 2016). Three forms of antibodies, IgM, IgD, and IgT, are essential components of humoral immunity which operate on invading extracellular diseases. The cytotoxic T-lymphocyte cells are a significant component of cellular immunity that often destroys invading bacterial, viral, or parasite-infected cells and intracellular cells (Firdaus-Nawi and Saad 2016). Both innate and adaptive immune systems work together efficiently to protect the body from any diseases, whereas innate immunity responds to invading pathogens by identifying line-encoded molecules of the germ.

# **1.3** Key Factors to Improve Disease Diagnosis and Vaccine Development

The major underlying factors for improving disease diagnosis and vaccine development are the continued significant losses to the industry caused by pathogens. Bacterial diseases cause substantial economic inflation in the aquaculture industry, and although antibiotics and chemotherapeutics are extensively used to control disease outbreaks, there is increasing concern about their use because of drug residues in food, the development of antimicrobial drug resistance, and their detrimental effect on aquatic microbial ecosystems and populations (Thompson and Adams 2004).

# 1.4 Limitations of Current Diagnostic Methods

Most of the current techniques for the detection of pathogens and the diagnosis of diseases are reliable. On the other hand, it is difficult to identify certain pathogens, and some of the methods developed may be too complicated to apply or infer. Conventional pathogen isolation and characterization techniques, combined with histopathology, remain the methods of choice for the diagnosis of many diseases. However, these traditional methods are costly, labour-intensive, slow, and time-consuming, and might not always lead to a definitive diagnosis being made. The rapid progress made in biotechnology since the 1990s has enabled the development and improvement of a wide range of immunodiagnostic and molecular techniques (Cunningham 2004; Adams and Thompson 2006, 2008), and reagents and commercial kits have become more widely available. These rapid methods both complement and enhance the traditional methods of disease diagnosis.

### **1.5** Advances in Methods of Disease Diagnosis

Disease diagnosis is currently made using a variety of methods, as reviewed by Adams and Thompson (2008). Traditional bacteriological methods, where the pathogen is isolated and identified biochemically (e.g. using API strips), and observation of histological sections from diseased fish are widely used. Rapid methods that specifically identify the pathogen using antibodies (immunodiagnostics) or by amplifying specific sequences of DNA or RNA using polymerase chain reaction (PCR) (i.e. molecular diagnostics) are also being used in many laboratories. In some cases, recent advances in molecular diagnostics have completely replaced other methodologies.

Immunodiagnostic methods such as immunohistochemistry (IHC), the fluorescence antibody test (FAT), and the indirect fluorescence antibody test (IFAT) enable rapid and specific detection of pathogens in tissue samples without the need to first isolate the pathogen. The use of molecular technologies for the detection of fish bacterial pathogens is rapidly increasing, and a vast array of methods has already been developed (Karunasagar et al. 1997; Cunningham 2004; Adams and Thompson 2006, 2008; Wilson and Carson 2003). Molecular methods generally have the highest sensitivity and are particularly useful for detecting microorganisms that are present in low copy numbers or those that are unculturable. The PCR is the best-known method, although there are many different kits, including nested PCR, random amplification of polymorphic DNA (RAPD), reverse transcriptase-PCR (RT-PCR), reverse cross blot PCR (rcb-PCR), and RT-PCR enzyme hybridization assay (Puttinaowarat et al. 2000; Wilson and Carson 2003; Cunningham 2004).

# 1.6 Nanotechnology Tool for Enhancing Aquaculture Operations and Fishing System

For any aquaculture practice, the pond is the core unit, and its professional management is the primary criterion for developing a successful aquaculture system. To design a perfect pond structure, multiple construction materials, as well as fabrication items, are required. Moreover, water supply and management of quality water are of primary importance. Sometimes, natural water bodies like the extensions of reservoirs and lakes, commonly called creeks, can be used for cage or open culture systems. The use of nanoparticles like silver, zinc oxide, copper oxide, titanium oxide, etc. can be mixed and coated with fabrication materials to increase the shelf life of the pond. Carbon nanotubes are approximately a hundred times stronger than steel and could be employed as an additive in boat and gear making or in designing cages and nets. ULVAC Inc., a corporate leader in vacuum equipment manufacturing, has designed a fishing lure coated with several hundred-nanometre-thick polyimide films that have a natural shining effect. Using this lure, better fishing practices can be achieved (Handy 2012).

## **1.7** Nanotechnology in Fish Disease Control

Disease occurrence is one of the major perils in intensive aquaculture practices, (Toranzo et al. 2005), and management of pond health is also important for the prevention and curing of fish species from pathogenic infestation. Nanotechnology can contribute significantly through novel methods as well as restructuring conventional technology (Kaul et al. 2018).

### **1.8** Nanomaterials as a Diagnostic Tool

In the modern era, aquaculture scientists are looking for an easy and reliable method for early detection and routine diagnostics of important fish diseases. An antibodybased, highly sensitive immunodiagnostics protocol incorporating nanoscale gold with alkaline phosphatase (ALP) conjugated secondary antibody was developed to measure titre against white spot syndrome virus (WSSV) in shrimp (Thiruppathiraja et al. 2011). Moreover, it has been reported that immuno-targeted gold nanoparticles are being conjugated with antibodies targeted to a particular biomolecule of interest, such as immunoglobulin G-capped gold nanoparticles, that bind specifically to antibodies produced against *Staphylococcus pyogenes* and *S. aureus*, among others (Roy et al. 2012).

Similarly, nanosensors also serve as an effective and easy method to identify pathogens. Nano biosensor systems are currently being developed for detecting very low concentrations not only of parasites, bacteria, and viruses but also of polluting elements in the water (Chen et al. 2016). Different nanosensors are being used to detect important aquaculture viruses like aquabirnavirus, salmonid alphavirus, and betanodavirus (Crane and Hyatt 2011). Currently, graphene oxide (GO) is used for designing electrochemical biosensors due to its unique chemical constitution and biocompatibility. A simple, sensitive, real-time, and rapid detection method of white spot syndrome virus (WSSV) in shrimp samples was demonstrated by applying a GO-based electrochemical immunosensor. The detection limit was found to be very low, at  $1.36 \times 10^{-3}$  copies  $\mu L^{-1}$ . This is a unique and alternative qualitative and quantitative method, unlike PCR amplification-based detection techniques (Natarajan et al. 2017). This is particularly important in outbreaks at commercial aquaculture systems since conventional methods take too much time to identify the etiological agent causes, delaying the treatment to control the pathogen and creating an important economic impact. In this regard, nanotechnology holds the key to overcoming the potential threat through early detection and eradication of pathogens. Currently, nanosensors can detect a wide range of pathogens, be it bacteria, viruses, or parasites. For instance, electrical nanosensors are feasible to detect single virus particles (Patolsky et al. 2004). Nanobiosensors are also used for fishpond cleaning and stock inspections, such as those based on carbon nanotubes, which are highly sensitive for the detection of traces of pathogens like viruses, parasites, and bacteria, as well as heavy metals, in both food and water (Husain et al. 2021).

### **1.9** Nanomaterials as Nanomedicine

Nanomedicine, another rapidly growing area in nanotechnology, has a wide opportunity to use the intrinsic properties of different forms of nanoparticles for improving fish health. The antimicrobial and prophylactic actions of nanomaterials like nanosilver and zinc oxide nanoparticles are already exploited to reduce the pathogenic load in the aquaculture system (Siddiqi et al. 2018). This unique phenomenon is nonspecific, universal, and widely applicable. Antibacterial properties of nanoparticles such as titanium dioxide and copper oxide are being investigated as a potential nanomedicine for fish. Graphene appears as a commercially attractive, cheap, and renewable nanomaterial when, in its oxidized form, it is easy to process and dispersible in water (Brisebois and Siaj 2019). Graphene oxide (GO) exhibited an inhibitory effect against important aquatic pathogens like *S. aureus*, *P. aeruginosa*, *E. coli*, and *Vibrio harveyi*, both individually and in combination with metal/ metal oxide and polymeric nanoparticles. Graphene oxide causes mechanical enfolding followed by cell membrane damage and lysis upon interacting with pathogens (Kumar et al. 2019).

# 1.10 Phyto-Nanocomposites as Nanomedicines

Different herbal and phytoextracts are currently being used as potential drugs in treating fish diseases. Different nanoparticles are being synthesized using medicinal plant and herbal extracts at optimized hydrodynamic conditions, and a composite of the phyto-nanoformulation is delivered as drugs with synergistic effects on improving fish health. Mahanty et al. (2013a, b) described some methods of phytoextract-synthesized nanosilver composite showing effective inhibition against the important fish pathogen, *Aeromonas hydrophila*.

# **1.11** Nanomaterials as Drug Delivery Systems (DDS) and Therapeutic Tools

Nano-delivery of drugs is attributed to novel properties like the sustained release, regulation, and control of size, shape, dispersity, and surface charge of targeted materials; location-specific, multi-route delivery processes; and regulated degradability of nanocarriers (Patra et al. 2018).

# **1.12** Nanovaccine Delivery

Nanotechnology, in conjunction with biotechnology, has made tremendous progress in the biomedicine field (Zhao et al. 2014) and has expanded its portfolio in the field of vaccinology, thereby giving rise to a new scientific field called nanovaccinology (Mamo and Poland 2012; Zhao et al. 2014). Vaccination plays a pivotal role in large-scale, commercial pisciculture and has been a major factor in the success of salmon and trout cultivation (FAO 2018) and partially in Indian major carp farming.

In the last 20 years, fish vaccinations have become a highly recognized, costeffective method for preventing certain pathogenic infections in aquaculture (Assefa and Abunna 2018; Vinitnantharat et al. 1999). Most of the vaccines produced are kept and preserved in liquid form at low temperatures and are usually injected through blood networks due to their short life span. These bottlenecks have reduced the chances of vaccines being widely used, particularly in certain finfish and shellfish. 'Nanovaccine' is an emerging mass vaccination method in aquaculture (Assefa and Abunna 2018). The USDA demonstrated ultrasound-driven tools for mass vaccination of fish. Short strands of DNA-loaded nanocapsules are placed in the fish pond, where they adsorb onto the surface of fishes. Then ultrasound is triggered to rupture the capsules for releasing the DNA eliciting an immunological response. This technology has been examined on rainbow trout and commercialized by 'Clear Springs Foods', USA (Mongillo 2007). Similarly, oral delivery of vaccines and targeted release of active ingredients for vaccination will reduce the cost of fish farming (Aravena et al. 2013).

DNA nanovaccines, comprising short strands of DNA within nanocapsules, have been used in the aquaculture industry to induce an immune response in fishes. Iron nanoparticles have been shown to accelerate fish development when combined with drug delivery via programmed release (Hussain et al. 2021). For instance, Bhattacharyya et al. (2015) successfully showed nanoencapsulated vaccines against the bacterium *Listonella anguillarum* in Asian carp and also in rainbow trout (*Oncorhynchus mykiss*), as shown in studies by (Mongillo 2007; Ogunkalu 2019; Zheng et al. 2016).

# **1.13** Nanoparticle-Based Gene Therapy

Gene therapy is another well-practiced method used in the aquaculture sector (Xiang 2015). In contemporary times, the European Medicines Agency (EMA) has recommended the use of 'Clynav', a DNA vaccine for protecting Atlantic salmon against acute infections. In another report, an efficient carrier vehicle was designed using chitosan-dextran sulfate and silica nanoparticles to deliver dsRNA into *Penaeus monodon* post-larvae for silencing the Monodon baculovirus (MBV) structural gene, p74. This carrier system exhibited a significant survival rate (86.63%) after successful gene silencing in *P. monodon* (Ramesh Kumar et al. 2016).

# 1.14 Different Nanocarriers for Drug Delivery

#### Chitosan

Chitosan is a processed by-product of the shrimp industry and has been developed as a well-known polymer-based nanocarrier. Chitosan nanoparticles were orally delivered to black tiger shrimp (*Penaeus monodon*) loaded with a DNA construct comprising the VP28 gene of white spot syndrome virus (WSSV). A significant survivability rate was monitored in WSSV-challenged shrimp compared to the 100% mortality in the control group (Rajeshkumar et al. 2009). Chitosan microspheres exhibited positive trends as an orally delivered plasmid vaccine carrier for immunization in Japanese flounder (Paralichthys olivaceus) (Tian et al. 2008). Many studies have successfully implemented encapsulation and delivery by chitosan-based systems in aquaculture, viz. treatment methods have been developed against Vibrio parahaemolyticus in the blackhead seabream (Acanthopagrus schlegelii) (Li et al. 2013), Philasterides dicentrarchi in the turbot (S. maximus) (Leon-Rodriguez et al. 2013), and Vibrio anguillarum in Asian sea bass (Lates cal*carifer*) (Kumar et al. 2008). Both DNA and RNA have been successfully encapsulated and delivered using this system. Studies by Ferosekahn et al. (2014) showed dietary RNA has been used in rohu (Labeo rohita), as well as inactive particles of the viral hemorrhagic septicemia virus (VHSV) in Japanese flounder (Paralichthys olivaceus), as reported by Kole et al. (2019). Recently, Kitiyodom et al. (2019) reported a chitosan-coated mucoadhesive nanovaccine with enhanced efficacy by immersion vaccination in tilapia (Oreochromis sp.) against columnaris disease. Additionally, chitosan-coated membrane vesicles (cMVs) derived from the pathogen Piscirickettsia salmonis were injected into adult zebrafish (Danio rerio), which conferred significant protection with an increased survival rate and induced an increased immune response (Tandberg et al. 2018).

# 1.15 Liposomes

Another commonly applied product is liposomes, which are artificially designed nanostructures comprising a lipid bilayer and have natural properties for application as carriers for drugs, vaccines, etc. It is nontoxic, biodegradable, and nonimmunogenic, and several nanoliposomal products are approved by the FDA. Moreover, it serves as an efficient delivery system, as it possesses the capacity to control and target the release of hydrophobic and hydrophilic compounds that are easily permeable through cell membranes. Nanoliposomal encapsulated Omega-3 fatty acids synthesized from seafood and fish oil are used in food fortification for efficient delivery and protection (Hadian 2016). It is also used to deliver important vitamins like folic acid via a dermal route that paved the way for delivering nutraceuticals (Kapoor et al. 2018). Liposome-encapsulated Aeromonas salmonicida antigen exhibited effective immunization in carp (Cyprinus carpio) (Irie et al. 2005). Similarly, an immunostimulant cocktail was applied as a nonspecific nanocarrier of the vaccine to multiple fish species. A liposomal lipopolysaccharide (LPS)-dsRNA cocktail was observed entering into the hepatocytes of zebrafish and trout macrophage plasma membranes, triggering pro-inflammatory and antiviral reactivity (Ruyra et al. 2013). Positive results were recorded after encapsulating curcumin into liposomes, prepared from natural sources (salmon lecithin), and examining them under primary cortical neuron culture (Hasan et al. 2018).

# 1.16 Poly (Lactic-Co-Glycolic) Acid (PLGA)

Polymeric nanoparticles of poly (lactic-co-glycolic) acid (PLGA) are examples of well-established drug delivery vehicles having adjuvant properties. It was found that an antigenic (TNPLPH) and immunostimulant ( $\beta$ -glucan) candidate encapsulated within the PLGA nanoparticles had the potentiality to enhance the expression of pro-inflammatory markers in salmon (Fredriksen et al. 2011). PLGA nanoparticles have also been monitored as a potential carrier of DNA vaccine for oral immunization against the lymphocystis disease virus, as evaluated in Japanese flounder (Paralichthys olivaceus) (Tian and Yu 2011). Surfactant-free PLGA nanoparticles displayed optimal biocompatibility and targeted dendritic cells in adult zebrafish after mucosal administration. This experiment opens a powerful platform for mucosal vaccine delivery in aquaculture (Resseguier et al. 2017). Shah and Mraz (2019) showed the use of encapsulation and delivery of different compounds in fish. For example, a DNA vaccine encapsulated in PLGA showed an improved immunological response against lymphocystis (Tian and Yu 2011). Additionally, Behera et al. (2010) showed the potential of PLGA encapsulated antigens from Aeromonas hydrophila, used as a vaccine in Rohu, to produce an enhanced immune-stimulatory and antibody response, both at 21 and 42 days post-immunization. Similarly, Yun et al. (2017) synthesized PLGA microparticles to carry A. hydrophila cells that were previously killed by formalin treatment as a way to deliver antigens to pond loaches (Misgurnus anguillicaudatus) and common carps (C. carpio). Upon being challenged with A. hydrophila, the fishes were treated with PLGA-A. hydrophila, which showed a higher survival rate with increased innate and adaptive immune responses than the group treated just with the inactivated A. hydrophila, which could potentially induce higher and more lasting immune responses than the antigens alone. Additionally, mass vaccination can be achieved using nanocapsules resistant to digestion and degradation. Both oral administration and site-specific release of the active agent will reduce the effort and cost related to disease management, leading to more sustainable practices (Rather et al. 2011).

# 1.17 Solid Lipid Nanoparticles

Solid lipid nanoparticles (SLNs) are investigated due to their potential topical delivery and controlled release in the skin. Pandita et al. (2011) investigated the potential applications of solid lipid nanoparticles (SLNs) in improving the oral bioavailability of paclitaxel, a hydrophobic drug. Experiments were conducted to evaluate the delivery of solid lipid nanoparticles (SLN) encapsulating potential antioxidant glutathione (GSH) on immunocompetent fish cells of a seawater teleost, the gilthead seabream (*S. aurata L.*). It was observed that the phagocytic function of HK (head kidney) leucocytes incubated with encapsulated glutathione was significantly enhanced. A better *in vitro* antioxidant efficacy of SLN was also monitored (Trapania

et al. 2016). Fin rot, dropsy, black and white spot disease, gill rot, columnaris, anchor worms, hole in the head, and *Argulus* infestations are some of the important external fish diseases that can be administered for recovery.

# 1.18 Microbial Disinfection

Nanoparticles synthesized from silver, titanium, and copper, among others, have been used for disease prevention and treatment. They have different modes of action against bacteria, and the most important function is acting on the cell membrane and cell wall by adhering to the cell wall through electrostatic interaction and disrupting the stability of the membrane. They can also disrupt the ion transport chain by interacting with ions and ion channels (Fajardo et al. 2022). It also generates doublestrand breaks in the DNA, interfering with the ribosome assembly and enzymatic activity via electrostatic interactions. They are also known to trigger a higher oxidative stress state by increasing the number of reactive oxygen species (ROS) that can damage proteins, lipids, other cellular contents, and DNA. Colloidal silver nanoparticles are one of the most widely used nanotechnology products against a wide spectrum of pathogens, including viruses, parasites, fungi, and bacteria. Silver is considered to be one of the most effective nanomaterials among the oligo-dynamic metallic nanoparticles due to its wide-range effects on different microbial species, different application forms, crystallographic structure, high surface exposure compared to volume, and compatibility with several compounds, making it a go-to use candidate for any treatment (Nangmenyi and Economy 2014). It triggers the oxidation of DNA and proteins with highly damaging effects, thus exerting antimicrobial activity. For example, silver nanoparticles are effective against methicillin-resistant S. aureus (Jeong et al. 2005). Eliminating microbes using visible light photocatalysts mediated by nanoparticles of metal oxides, as well as nano-porous fibres and foams, were also found to be effective (Li et al. 2014). Their effect is not limited to microbial elimination but also includes the elimination of organic pollutants from the pharmaceutical and cosmetics industries (Chena and Yadab 2011). Titanium dioxide nanoparticles, for example, have a strong anti-bacterial activity and are capable of destroying fish pathogens in vitro (Cheng et al. 2009), and their actions have been confirmed as a strong immune modulator of fish neutrophil function (Jovanovic 2011). NanoCheck, a commercially available nanotechnology product that is currently used for cleaning fishponds with lanthanum-based particles (40 nm in size), functions by inhibiting algal growth by absorbing phosphates from the water (Rather et al. 2011). Korni and Khalil (2017) showed that ginger-derived nanoparticles control the infection by motile Aeromonas septicaemia in the Asian carp fingerlings. Rather et al. (2017) reported that silver nanoparticles synthesized by Azadirachta indica (neem) showed antibacterial and immunomodulatory activity in mrigal (Cirrhinus mrigala) fingerlings when challenged with A. hydrophila. More recently, Erdem et al. (2018) demonstrated the antibacterial efficacy of silver nanoparticles synthesized by Aeromonas sobria against A. hydrophila. This raised the possibility to improve sanitary management through the use of more ecofriendly and economically based antimicrobial agents (Shah and Mraz 2019). Recent studies by Kepiro et al. (2020) explain the conceptual design of protein pseudocapsids exerting broad-spectrum antimicrobial activity. In contrast to conventional antibiotics, these pseudocapsids are highly effective against diverse bacterial strains. This method can eliminate antibiotic-resistant bacteria *in vivo* without causing toxicity. Pseudocapsids, which are nothing but an icosahedral structure that is polymorphic in size, but not in shape, are available in both D and L epimeric forms. It has to be noted that pseudocapsids can cause fast and irreversible harm to bacterial cells.

### **1.19** Conclusions and Future Prospects

There are some significant disadvantages to the classical methods of treatment and remedy that have been observed. But those classical approaches have given a path for modern technology, without which science can't flourish. One such field is nanotechnology, which has given rise to numerous options with its wide field of application. Nowadays, disease in the aquaculture sector is bringing huge losses, and methods are being developed for rapid diagnosis and treatment. One such method is nanovaccination, which is a new attempt to enhance the immunogenicity of vaccines using nanoparticles as carriers and/or adjuvants. The immune system can very well be triggered because of the similar scale (size) between the nanoparticles and the pathogens, resulting in triggered cellular and humoral immunity responses. Thus, using nanotechnology in aquaculture has become a comprehensive tool for solving a lot of problems, not only in disease diagnosis and treatment but also in water quality control, fish nutrition, environmental management, etc. Currently, various nanotechnological applications have been implemented to improve the aquaculture industry, which could play an important role in the development and sustainability of this industry in the future. Nowadays, there are many potential applications for nanomaterials in the fisheries and aquaculture industries. Some of the most promising areas in this field are applications related to fish health management, nanoscale ingredient incorporation, the use of nanotechnology in aquaculture feeds and food packaging, as well as applications linked to value-added products, stress reduction, and health management. Sustainable development of nanotechnology in the fisheries and aquaculture industries will require a comprehensive assessment of its potential negative impacts; therefore, a careful analysis of the life cycle and shelf life of new nanomaterials, combined with an assessment of potential health and environmental risks, including exposure, release, and deposition, must be performed. However, considering the negative sides of this method, we should not refrain from trying to implement nanotechnological applications in the aquaculture sector, which should be linked to careful monitoring and controlled use, thus favouring efforts to minimize risks and maximize benefits. In the future, nanoparticles can be used in the detection of different diseases caused by bacteria as well as viruses in fishes (Tables 1.1 and 1.2).

Type of nanoparticles	Merits	Demerits	
Polymeric nanoparticles	Better immunogenicity can be obtained by easy modification of surface properties, biodegradable and targeted antigen delivery	Low aqueous solubility and synthesis require the use of organic solvents, low antigen loading, premature release of antigens, insufficient antigen protection	
Inorganic nanoparticles	Easy to modify, less chances of premature release, and better protection of adsorbed antigens	Low aqueous solubility and low biodegradability	
Nanoliposomes	Possess intrinsic adjuvant properties, accommodate both hydrophilic and lipophilic antigens and relatively stable in gastrointestinal fluids when modified	Low mucus penetration, limited antigen loading, and poor gastrointestinal stability of naked liposomes	
Virus-like particles (VLP's)	Possess self-adjuvant properties, mimic original virus and high gastrointestinal stability	Lack of reproducibility	
Nanoemulsions Possess self-adjuvant properties, encapsulate both hydrophilic and lipophilic antigens		Premature release of antigens and poor gastrointestinal stability	

 Table 1.1
 Merits and demerits of nanoparticles

 Table 1.2
 Progress in nanoparticle-based fish vaccine delivery

Nanoparticle	Pathogen	Fish species	Mode of administration	Vaccine formulations with nanoparticles	References
Chitosan	VHSV	Danio rerio	IP	NPrgpG pICrgpG	Kavaliauskis et al. (2016)
Chitosan	ISAV	Salmo salar	Oral	NP-V/ NP-Ad+NP-V	Aravena et al. (2013)
Chitosan/ TPP	Nodavirus	Lates calcarifer	Oral	pFNCPE42-CS/TPP	Vimal et al. (2014)
Chitosan	Vibrio anguillarum (Listonella)	Lates calcarifer	Oral	Chitosan- pVAOMP38	Rajesh et al. (2008)
PLGA	Aeromonas hydrophila	Labeo rohita	Oral	Np-rOmpW (HiAg) 8 µg/g Np-rOmpW (LoAg) 4 µg/g	Dubey et al. (2016)
PLGA, PLA	Aeromonas hydrophila	Labeo rohita	IP	PLGA-Omp	Rauta and Nayak 2015
PLGA	IPNV	Salmo salar	IP	PLGA nanoparticle-TA PLGA nanoparticle-PT	Munangandu et al. (2012)

(continued)

Nanoparticle	Pathogen	Fish species	Mode of administration	Vaccine formulations with nanoparticles	References
PLGA	IHNV	Oncorhynchus mykiss	Oral	High dose PLGA(10WPV) Low dose PLGA- pCDNA-G high dose PLGA-pCDNA-G high dose PLGA	Adomako et al. (2012)
PLGA, PLA	_	Salmo salar	IP	PLGA-NP50L PLGA-NP50H PLGA-NP75H PLA-NP100L	Fredriksen and Grip (2012)
PLGA	LCDV	Paralichthys olivaceus	Oral	pEGFP-N2-MCP PLGA PLGA- pEGFP-N2- MCP	Tian and Yu (2011)
PLGA	-	Salmo saliar	IP	NP NP/TNP-LPH, NP/β glucan, NP/ TNP-LPH/β glucan	Fredriksen et al. (2011)
Liposome	Vibrio harveyi	Epinephelus bruneus	IP	Liposome-V. harveyi	Harikrishnan et al. (2012)
Liposome	Aeromonas salmonicida	Cyprinus carpio	Oral	Liposome-T1031	Irie et al. (2005)
Carbon nanotubes	GCRV	Ctenopharyngon idellus	IM Bath	SWCNTs- pEGFP-vp5 1 µg SWCNTs- pEGFP-vp5 2.5 µg SWCNTs- pEGFP-vp5 5 µg SWCNTs- pEGFP-vp5 1 mgL-1 SWCNTspEGFP-vp5 10 mg L-1 SWCNTs- pEGFP-vp5 20 mg L-1	Wang et al. (2015)

Table 1.2 (continued)

# References

- Adams A, Thompson KD (2006) Biotechnology offers revolution to fish health management. Trends Biotechnol 24:201–205
- Adams A, Thompson KD (2008) Recent applications of biotechnology to novel diagnostics for aquatic animals. Rev Sci Tech Off Int Epiz 27:197–209
- Adomako M, St-Hilaire S, Zheng Y, Eley J, Marcum RD (2012) Oral DNA vaccination of rainbow trout, Oncorhynchus mykiss (Walbaum), against infectious haematopoietic necrosis virus using PLGA [Poly (D, L-Lactic- Co-Glycolic Acid)] nanoparticles. J Fish Dis 35(3):203–214
- Altenburger R, Brack W, Burgess RM, Busch W, Escher BI, Focks A et al (2019) Future water quality monitoring: improving the balance between exposure and toxicity assessments of realworld pollutant mixtures. Environ Sci Eur 31:1–17

- Aravena AR, Sandino AM, Spencer E (2013) Nanoparticles and microparticles of polymers and polysaccharides to administer fish vaccines. Biol Res 46(4):407–419
- Arfat YA, Benjakul S, Prodpran T, Sumpavapol P, Songtipya P (2014) Properties and antimicrobial activity of fish protein isolate/fish skin gelatin film containing basil leaf essential oil and zinc oxide nanoparticles. Food Hydrocoll 41:265–273
- Assefa A, Abunna F (2018) Maintenance of fish health in aquaculture: review of epidemiological approaches for prevention and control of infectious disease of fish. Vet Med Int 5432497. https://doi.org/10.1155/2018/5432497
- Bacchetta C, Ale A, Simoniello MF, Gervasio S, Davico C, Rossi AS et al (2017) Genotoxicity and oxidative stress in fish after a short-term exposure to silver nanoparticles. Ecol Indic 76:230–239
- Behera T, Nanda PK, Mohanty C, Mohapatra D, Swain P, Das BK, Routray P, Mishra BK, Sahoo SK (2010) Parenteral immunization of fish, *Labeo rohita* with Poly D, L-lactide-co-glycolic acid (PLGA) encapsulated antigen microparticles promotes innate and adaptive immune responses. Fish Shellfish Immunol 28(2):320–325. https://doi.org/10.1016/j.fsi.2009.11.009
- Bhattacharyya A, Reddy J, Hasan MM, Adeyemi MM, Marye RR (2015) Nanotechnology-a unique future technology in aquaculture for the food security. Int J Bioassays 4(7):4115–4126
- Brisebois P, Siaj M (2019) Harvesting graphene oxide-years: 1859 to 2019: a review of its structure, synthesis, properties and exfoliation. J Mater Chem C. https://doi.org/10.1039/c9tc03251g
- Chen B, Zou L, Wu Z, Sun M (2016) The application of quantum dots in aquaculture pollution detection. Toxicol Environ Chem 98(3–4):385–394. https://doi.org/10.1080/0277224 8.2015.1123482
- Chena H, Yadab R (2011) Nanotechnologies in agriculture: new tools for sustainable development. Trends Food Sci Technol 22(11):585–594. https://doi.org/10.1016/j.tifs.2011.09.00
- Cheng TC, Yao KS, Yeh N, Chang CI, Hsu HC, Chien YT, Chang CY (2009) Visible light activated bactericidal effect of TiO2/Fe3O4 magnetic particles on fish pathogens. Surf Coat Technol 204(6–7):1141–1144. https://doi.org/10.1016/j.surfcoat.2009.06.050
- Crane M, Hyatt A (2011) Viruses of fish: an overview of significant pathogens. Virus 3:2025-2046
- Cunningham CO (2004) Use of molecular diagnostic tests in disease control: making the leap from laboratory to fi eld application. In: Leung K-Y (ed) Current trends in the study of bacterial and viral fish and shrimp diseases, molecular aspects of fish and marine biology, vol 3. World Scientific, Singapore, pp 292–312
- Dubey S, Avadhani K, Mutalik S, Sivadasan SM, Maiti B (2016) Aeromonas hydrophila OmpW PLGA nanoparticle oral vaccine shows a dose-dependent protective immunity in rohu (Labeo rohita). Vaccine 4(2):21
- Erdem B, Dayangac A, Kıray E, Duygu D (2018) Biosynthesis of silver nanoparticles from *Aeromonas sobria* and antibacterial activity against fish pathogens. Int J Environ Sci Technol 16(9):1–6. https://doi.org/10.1007/s13762-018-1944-z
- Fajardo C, Martinez G, Rodriguez J Blasco JM Mancera B Thomas MDD (2022). Nanotechnology in aquaculture: applications, perspectives and regulatory challenges. Aquac Fish 7(2): 185–200. ISSN 2468-550X. https://doi.org/10.1016/j.aaf.2021.12.006
- FAO (2018) The state of world fisheries and aquaculture 2018 meeting the sustainable development goals. Food & Agriculture Organization, Rome. Retrieved from http://www.fao.org/3/ i9540en/i9540en.pdf
- Ferosekhan S, Gupta S, Singh A, Rather M, Kumari R, Kothari D, Pal KA, Jadhao SB (2014) RNAloaded chitosan nanoparticles for enhanced growth, immunostimulation and disease resistance in fish. Curr Nanosci 10(3):453–464. https://doi.org/10.2174/1573413710666140115220300
- Firdaus-Nawi M, Saad M (2016) Major components of fish immunity: a review. Pertanika J Trop Agric Sci 39(4):393–420
- Fredriksen BN, Grip J (2012) PLGA/PLA micro-and nanoparticle formulations serve as antigen depots and induce elevated humoral responses after immunization of Atlantic salmon (Salmo salar L.). Vaccine 30(3):656–667

- Fredriksen BN, Saevareid K, McAuley L, Lane ME, Bogwald J et al (2011) Early immune responses in Atlantic salmon (*Salmo salar* L.) after immunization with PLGA nanoparticles loaded with a model antigen and β-glucan. Vaccine 29(46):8338–8349
- Gao J, Lin, Wei A, Sepulveda (2017) Protein corona analysis of silver nanoparticles exposed to fish plasma. Environ Sci Technol Lett 4(5):174–179
- Hadian Z (2016) A review of nanoliposomal delivery system for stabilization of bioactive Omega-3 fatty acids. Electron Phys. https://doi.org/10.19082/1776
- Handy RD (2012) FSBI briefing paper: nanotechnology in fisheries and aquaculture. Fisheries Society of the British Isles
- Harikrishnan R, Kim JS, Balasundaram C, Heo MS (2012) Vaccination effect of liposomes entrapped whole cell bacterial vaccine on immune response and disease protection in Epinephelus bruneus against Vibrio harveyi. Aquaculture 342–343:69–74
- Hasan M, Latifi S, Kahn CJF, Tamayol A, Habibey R, Passeri E, Linder M, Arab-Tehrany E (2018) The positive role of curcumin-loaded salmon nanoliposomes on the culture of primary cortical neurons. Mar Drugs 16:218
- Herbeck LS, Unger D, Wu Y, Jennerjahn TC (2013) Effluent, nutrient and organic matter export from shrimp and fish ponds causing eutrophication in coastal and back-reef waters of NE Hainan, tropical China. Cont Shelf Res 57:92–104
- Husain S, Verma SK, Yasin D, Hemlataa, Rizvi MMA, Fatma T (2021) Facile green bio- fabricated silver nanoparticles from microchaete infer dose-dependent antioxidant and anti-proliferative activity to mediate cellular apoptosis. Bioorg Chem 107:104535
- Irie T, Watarai S, Iwasaki T, Kodama H (2005) Protection against experimental Aeromonas salmonicida infection in carp by oral immunisation with bacterial antigen entrapped liposomes. Fish Shellfish Immunol 18(3):235–242. https://doi.org/10.1016/j.fsi.2004.07.006
- Jeevanandam J, Barhoum A, Chan YS, Dufresne A, Danquah MK (2018) Review on nanoparticles and nanostructured materials: history, sources, toxicity and regulations. Beilstein J Nanotechnol 9:1050–1074
- Jeong SH, Hwang YH, Yi SC (2005) Antibacterial properties of padded PP/PE nonwovens incorporating nano-sized silver colloids. J Mater Sci 40(20):5413–5418. https://doi.org/10.1007/ s10853-005-4340-2
- Jovanovic B (2011) Immunotoxicology of titanium dioxide and hydroxylated fullerenes engineered nanoparticles in fish models. Graduate theses and dissertations Iowa State University Capstones, Theses and Dissertations: 1–176
- Ju-Nam Y, Lead JR (2008) Manufactured nanoparticles: an overview of their chemistry, interactions and potential environmental implications. Sci Total Environ 400:396–414
- Kapoor MS, D'Souza A, Aibani N, Nair SS, Sandbhor P, Banerjee R (2018) Stable liposome in cosmetic platforms for transdermal folic acid delivery for fortification and treatment of micronutrient deficiencies. Sci Rep 8:1–12
- Karunasagar I, Nayak BB, Karunasagar I (1997) Rapid detection of Vibrio parahaemolyticus from fi sh by polymerase chain reaction. In: Flegel TW, MacRae LH (eds) Diseases in Asian aquaculture III. Asian Fisheries Society, Manila, pp 119–122
- Kaul S, Gulati N, Verma D, Mukherjee S, Nagaich U (2018) Role of nanotechnology in cosmeceuticals: a review of recent advances. J Pharma 3420204. https://doi.org/10.1155/2018/3420204
- Kavaliauskis A, Arnemo M, Speth M, Lagos L, Rishovd AL et al (2016) Protective effect of recombinant VHSV-G vaccine using poly (I:C) loaded nanoparticles as an adjuvant in zebrafish (Danio rerio) infection model. Dev Comp Immunol 61:248–257
- Kepiro IE, Marzuoli I, Hammond K, Ba X, Lewis H, Shaw M, Gunnoo SB, De Santis E, Kapinska U, Pagliara S, Holmes MA, Lorenz CD, Hoogenboom BW, Fraternali F, Ryadnov MG (2020) Engineering chirally blind protein pseudocapsids into antibacterial persisters. ACS Nano 14(2):1609–1622. https://doi.org/10.1021/acsnano.9b06814
- Khan I, Saeed K, Khan I (2019) Nanoparticles: properties, applications and toxicities. Arab J Chem 12:908–931

- Kitiyodom S, Yata T, Yostawornkul J, Kaewmalun S, Nittayasut N, Suktham K, Surassmo S, Namdee K, Rodkhum C, Pirarat N (2019) Enhanced efficacy of immersion vaccination in tilapia against columnaris disease by chitosan-coated "pathogen-like" mucoadhesive nanovaccines. Fish Shellfish Immunol 95:213–219. https://doi.org/10.1016/j.fsi.2019.09.064
- Kole S, Qadiri SSN, Shin SM, Kim WS, Lee J, Jung SJ (2019) Nanoencapsulation of inactivatedviral vaccine using chitosan nanoparticles: evaluation of its protective efficacy and immune modulatory effects in olive flounder (*Paralichthys olivaceus*) against viral haemorrhagic septicaemia virus (VHSV) infection. Fish Shellfish Immunol 91:136–147. https://doi.org/10.1016/j. fsi.2019.05.017
- Korni FMM, Khalil F (2017) Effect of ginger and its nanoparticles on growth performance, cognition capability, immunity and prevention of motile Aeromonas septicaemia in Cyprinus carpio fingerlings. Aquac Nutr 23(6):1492–1499. https://doi.org/10.1111/anu.12526
- Kumar SR, Ahmed VI, Parameswaran V, Sudhakaran R, Babu VS, Hameed AS (2008) Potential use of chitosan nanoparticles for oral delivery of DNA vaccine in Asian sea bass (*Lates calcarifer*) to protect from *vibrio* (*Listonella*) anguillarum. Fish Shellfish Immunol 25(1–2):47–56. https://doi.org/10.1016/j.fsi.2007.12.004
- Kumar P, Huo P, Zhang R, Liu B (2019) Antibacterial properties of graphene-based nanomaterials. Nano. https://doi.org/10.3390/nano9050737
- Kumari P, Panda PM, Jha E, Kumari K, Nisha K, Mallick MA, Verma SK (2017) Mechanistic insight to ROS and apoptosis regulated cytotoxicity inferred by green synthesized CuO nanoparticles from *Calotropis gigantea* to embryonic zebrafish. Sci Rep 7:1–17
- Leon-Rodriguez L, Luzardo-Alvarez A, Blanco-Mendez J, Lamas J, Leiro J (2013) Biodegradable microparticles covalently linked to surface antigens of the scuticociliate parasite P. dicentrarchi promote innate immune responses in vitro. Fish Shellfish Immunol 34(1):236–243. https://doi. org/10.1016/j.fsi.2012.10.029
- Li L, Lin SL, Deng L, Liu ZG (2013) Potential use of chitosan nanoparticles for oral delivery of DNA vaccine in black seabream Acanthopagrus schlegelii Bleeker to protect from Vibrio parahaemolyticus. J Fish Dis 36(12):987–995. https://doi.org/10.1111/jfd.12032
- Li Q, Wu P, Shang JK (2014) Nanostructured visible-light photocatalysts for water purification. In: Street A, Sustich R, Duncan J, Savage N (eds) Nanotechnology applications for clean water: solutions for improving water quality. William Andrew, Norwich, pp 297–317. https://doi. org/10.1016/B978-1-4557-3116-9.00019-6
- Mahanty A, Bosu R, Panda P, Netam SP, Sarkar B (2013a) Microwave assisted rapid combinatorial synthesis of silver nanoparticles using *E.coli* culture supernatant. Int J Pharma Biosci 4:1030–1035
- Mahanty A, Mishra S, Bosu R, Maurya UK, Netam SP, Sarakar B (2013b) Phytoextractssynthesized silver nanoparticles inhibit bacterial fish pathogen Aeromonas hydrophila. Ind J Microbiol 53:438–446
- Mamo T, Poland GA (2012) Nanovaccinology: the next generation of vaccines meets 21st century materials science and engineering. Vaccine 30:6609–6611
- Matteucci F, Giannantonio R, Calabi F, Agostiano A, Gigli G, Rossi M (2018) Deployment and exploitation of nanotechnology nanomaterials and nanomedicine. In: AIP conference proceedings. https://doi.org/10.1063/1.5047755
- Mohamed S, Mona S, Magdy EM, Mansour EM (2015) Recent progress in applications of nanoparticles in fish medicine: a review. Nanomed Nanotechnol Biol Med. https://doi.org/10.1016/j. nano.2015.11.005
- Mongillo JF (2007) Nanotechnology 101. Greenwood Press, Westport
- Munangandu HM, Fredriksen BN, Mutoloki S, Brudeseth B, Kuo TY et al (2012) Comparison of vaccine efficacy for different antigen delivery systems for infectious pancreatic necrosis virus vaccines in Atlantic salmon (Salmo salar L.) in a cohabitation challenge model. Vaccine 30(27):4007–4016

- Nangmenyi G, Economy J (2014) Nanometallic particles for oligodynamic microbial disinfection. In: Street A, Sustich R, Duncan J, Savage N (eds) Nanotechnology applications for clean water: solutions for improving water quality. William Andrew, Norwich, pp 283–295
- Natarajan A, Shalini Devi KS, Raja S, Senthil Kumar A (2017) An elegant analysis of white spot syndrome virus using a graphene oxide/methylene blue based electrochemical immunosensor platform. Sci Rep. https://doi.org/10.1038/srep46169
- Naylor RL, Goldburg RJ, Primavera JH, Kautsky N, Beveridge MCM, Clay J et al (2006) Effect of aquaculture on world fish supplies. Nature 405:1017–1024
- Ogunkalu OA (2019) Utilization of nanotechnology in aquaculture and seafood sectors. EJFST 19:26–33
- Pandita D, Ahuja A, Lather V, Benjamin B, Dutta T, Velpandian T (2011) Development of lipidbased nanoparticles for enhancing the oral bioavailability of Paclitaxel. AAPS Pharm Sci Technol 12:712–722
- Patel M, Kumar R, Kishor K, Mlsna T, Pittman CU, Mohan D (2019) Pharmaceuticals of emerging concern in aquatic systems: chemistry, occurrence, effects, and removal methods. Chem Rev 119:3510–3673
- Patolsky F, Zheng GF, Hayden O, Lakadamyali M, Zhuang XW, Lieber CM (2004) Electrical detection of single viruses. Proc Natl Acad Sci 101:14017–14022. https://doi.org/10.1073/ pnas.0406159101
- Patra JK, Das G, Fraceto LF, Campos E, Rodriguez-Torres M, Acosta-Torres LS et al (2018) Nano based drug delivery systems: recent developments and future prospects. J Nanobiotechnol 16:71. https://doi.org/10.1186/s12951-018-0392-8
- Puttinaowarat S, Thompson KD, Adams A (2000) Mycobacteriosis: detection and identification of aquatic mycobacterium species. Fish Vet J 5:6–21
- Rajesh SK, Ahmed VPI, Parameswaran V, Sudhakaran R, Babu VS et al (2008) Potential of chitosan nanoparticles for oral delivery of DNA vaccine in Asian sea bass (*Lates calcarifer*) to protect from vibrio (Listonella) anguillarum. Fish Shellfish Immunol 25(1–2):47–56
- Rajeshkumar S, Venkatesan C, Sarathi M, Sarathbabu V, Thomas J, Anver Basha K, Hameed ASS (2009) Oral delivery of DNA construct using chitosan nanoparticles to protect the shrimp from white spot syndrome virus (WSSV). Fish Shellfish Immunol 26:429–437. https://doi. org/10.1016/j.fsi.2009.01.003
- Rajkumar KS, Kanipandian N, Ramasamy T (2016) Toxicity assessment on haemotology, biochemical and histopathological alterations of silver nanoparticles-exposed freshwater fish Labeo rohita. Appl Nanosci 6(1):19–29
- Ramesh Kumar D, Elumalai R, Raichur AM, Sanjuktha M, Rajan JJ, Alavandi SV, Vijayan KK, Poornima M, Santiago TC (2016) Development of antiviral gene therapy for monodon baculovirus using dsRNA loaded chitosan-dextran sulfate nanocapsule delivery system in Penaeus monodon post-larvae. Antivir Res 131:124–130
- Rather MA, Sharma R, Aklakur M, Ahmad S, Kumar N, Khan M, Ramya VL (2011) Nanotechnology: a novel tool for aquaculture and fisheries development. A prospective minireview. Fish Aquac J FAJ 16
- Rather MA, Bhat IA, Sharma N, Gora A, Ganie PA, Sharma R (2017) Synthesis and characterization of Azadirachta indica constructed silver nanoparticles and their immunomodulatory activity in fish. Aquac Res 48:3742–3754
- Rauta PR, Nayak B (2015) Parenteral immunization of PLA/PLGA nanoparticle encapsulating outer membrane protein (Omp) from *Aeromonas hydrophila*. Evaluation of immunostimulatory action in *Labeo rohita* (rohu). Fish Shellfish Immunol 44(1):287–294
- Resseguier J, Delaune E, Coolen AL, Levraud JP, Boudinot P, Guellec DL, Verrier B (2017) Specific and efficient uptake of surfactant-free poly (Lactic Acid) nanovaccine vehicles by mucosal dendritic cells in adult zebrafish after bath immersion. Front Immunol. https://doi. org/10.3389/fimmu.2017.00190
- Roy S, Kumar V, Barman D, Mandal S (2012) Nanotechnology and its application in fisheries and aquaculture. AQUA Int 20(6):2–3

- Ruyra A, Cano-Sarabia M, MacKenzie SA, Maspoch D, Roher N (2013) A novel liposome-based nanocarrier loaded with an LPS-dsRNA cocktail for fish innate immune system stimulation. PLoS One 8:e76338
- Sarkar B, Mahanty A, Gupta SK, Choudhury AR, Daware A, Bhattacharjee S (2022) Nanotechnology: a next-generation tool for sustainable aquaculture. Aquaculture 546:737330
- Shah BR, Mraz J (2019) Advances in nanotechnology for sustainable aquaculture and fisheries. Rev Aquac 1–18. https://doi.org/10.1111/raq.12356
- Siddiqi KS, Husen A, Rao R (2018) A review on biosynthesis of silver nanoparticles and their biocidal properties. J Nanobiotechnol 16:14
- Stern ST, Martinez MN, Stevens DM (2016) When is it important to measure unbound drug in evaluating nanomedicine pharmacokinetics? Drug Metabol Disposal 44:1934–1939
- Stone V, Nowack B, Baun A, van den Brink N, von der Kammer F, Dusinska M, Handy R, Hankin S, Hassellov M, Joner E, Fernandes TF (2010) Nanomaterials for environmental studies: classification, reference material issues, and strategies for physico-chemical characterisation. Sci Total Environ 408:1745–1754. https://doi.org/10.1016/j.scitotenv.2009.10.035
- Tandberg J, Lagos L, Ropstad E, Smistad G, Hiorth M, Winther-Larsen HC (2018) The use of chitosan-coated membrane vesicles for immunization against salmonid rickettsial septicemia in an adult zebrafish model. Zebrafish 15(4):372–381. https://doi.org/10.1089/zeb.2017.1556
- Thiruppathiraja C, Kumar S, Murugan V, Adaikkappan P, Sankaran K, Alagar M (2011) An enhanced immuno-dot blot assay for the detection of white spot syndrome virus in shrimp using antibody conjugated gold nanoparticles probe. Aquaculture 318:262–267
- Thompson KD, Adams A (2004) Current trends in immunotherapy and vaccine development for bacterial diseases of fish. In: Leung K-Y (ed) Current trends in the study of bacterial and viral fish and shrimp diseases, Molecular aspects of fish and marine biology, vol 3. World Scientific, Singapore, pp 313–362
- Tian J, Yu J (2011) Poly (lactic-co-glycolic acid) nanoparticles as candidate DNA vaccine carrier for oral immunization of Japanese flounder (*Paralichthys olivaceus*) against lymphocystis disease virus. Fish Shellfish Immunol 30:109–117. https://doi.org/10.1016/j.fsi.2010.09.016
- Tian J, Yu J, Sun X (2008) Chitosan microspheres as candidate plasmid vaccine carrier for oral immunisation of Japanese flounder (*Paralichthys olivaceus*). Vet Immunol Immunopath 126:220–229
- Toranzo AE, Magarinos TB, Romalde JL (2005) A review of the main bacterial fish diseases in mariculture systems. Aquaculture 246:37–61
- Trapania A, Tripodob G, Mandracchiaa D, Cioffic N, Ditarantoc N, Cerezuela R, Esteband MA (2016) Glutathione loaded solid lipid nanoparticles: preparation and *in vitro* evaluation as delivery systems of the antioxidant peptide to immunocompetent fish cells. J Cell Biotechnol 2:1–14
- Verma SK, Jha E, Sahoo B, Panda PK, Thirumurugan A, Parashare SKS, Suar M (2017) Mechanistic insight into the rapid one-step facile biofabrication of antibacterial silver nanoparticles from bacterial release and their biogenicity and concentration-dependent in vitro cytotoxicity to colon cells. RSC Adv 7:40034–40045
- Verma SK, Nisha K, Panda PK, Patel P, Kumari P, Mallick MA, Sarkar B, Das B (2020) Green synthesized MgO nanoparticles infer biocompatibility by reducing in vivo molecular nanotoxicity in embryonic zebrafish through arginine interaction elicited apoptosis. Sci Total Environ 713:136521
- Verma SK, Panda PK, Kumari P, Patel P, Arunima A, Jha E, Husain S, Prakash R, Hergenroder R, Mishra YK, Ahuja R, Varma RS, Suar M (2021) Determining factors for the nanobiocompatibility of cobalt oxide nanoparticles: proximal discrepancy in intrinsic atomic interactions at differential vicinage. Green Chem 23:3439–3458
- Vevers WF, Jha AN (2008) Genotoxic and cytotoxic potential of titanium dioxide (TiO2) nanoparticles on fish cells in vitro. Ecotoxicology 17(5):410–420
- Vimal S, Majeed AS, Nambi KSN, Madan N, Farook MA et al (2014) Delivery of DNA vaccine using chitosan-tripolyphosphate (CS/TPP) nanoparticles in Asian sea bass. *Lates calcarifer* (Bloch,1790) for protection against nodavirus infection. Aquaculture 420–421:240–246

- Vinay TN, Bhat S, Choudhury TG, Paria A, Jung MH, Kallappa GS, Jung SJ (2018) Recent advances in application of nanoparticles in fish vaccine delivery. Rev Fish Sci Aquacult 26:29–41
- Vinitnantharat S, Gravningen K, Greger E (1999) Fish vaccines. Adv Vet Med 41:539-550
- Wang Y, Liu GL, Li DL, Ling F, Zhu B et al (2015) The protective immunity against grass carp reovirus in grass carp induced by a DNA vaccination using single-walled carbon nanotubes as delivery vehicles. Fish Shellfish Immunol 47(2):732–742
- Wilson T, Carson J (2003) Development of sensitive, high-throughput one-tube RT-PCR-enzyme hybridisation assay to detect selected bacterial fish pathogens. Dis Aquatic Org 54:127–134
- Xiang J (2015) Recent major advances of biotechnology and sustainable aquaculture in China. Curr Biotechnol 4:296–310
- Yun S, Jun JW, Giri SS, Kim HJ, Chi C, Kim SG, Kim SW, Park SC (2017) Efficacy of PLGA microparticle-encapsulated formalin-killed Aeromonas hydrophila cells as a single-shot vaccine against A. hydrophila infection. Vaccine 35(32):3959–3965. https://doi.org/10.1016/j. vaccine.2017.06.005
- Zhang JS, Li ZJ, Wen GL, Wang YL, Luo L, Zhang HJ, Dong HB (2016) Relationship between white spot syndrome virus (WSSV) loads and characterizations of water quality in *Litopenaeus* vannamei culture ponds during the tropical storm. Iran J Vet Res 17:210–214
- Zhao L, Seth A, Wibowo N, Zhao C-X, Mitter N, Yu C, Anton PJ, Middelberg APJ (2014) Nanoparticle vaccines. Vaccine 32(3):327–337. https://doi.org/10.1016/j.vaccine.2013.11.069
- Zheng F, Liu H, Sun X, Zhang Y, Zhang B (2016) Development of oral DNA vaccine based on chitosan nanoparticles for the immunization against reddish body iridovirus in turbots (Scophthalmus maximus). Aquaculture 452:263–271