



Diet and Micronutrients

Madhura Murittige Gopalakrishna and Roopa S. Rao

Contents

- 10.1 Introduction – 124**
- 10.2 Epidemiological Evidence on Diet and Nutrition – 124**
- 10.3 Role of Trace Elements in OSF – 125**
 - 10.3.1 Role of Copper – 125
 - 10.3.2 Role of Zinc – 125
 - 10.3.3 Role of Iron – 126
 - 10.3.4 Role of Selenium – 126
- 10.4 Role of Vitamins – 126**
- 10.5 Interventional Studies – 127**
- References – 127**

10.1 Introduction

Oral submucous fibrosis (OSF) is a potentially malignant disorder, primarily caused by areca nut consumption, and is characterized by fibrotic changes of oral and pharyngeal tissues. The etiopathogenesis of OSF is a complex orchestration involving a multitude of molecules and enzymes [1–6]. In this chapter, we discuss the role of diet, nutrition, and micronutrients as risk or protective factors in the causation of OSF.

WCRF/AICR defines “**Nutrition** as the set of integrated processes by which cells, tissues, organs, and indeed a whole organism acquire the energy and nutrients needed to function normally and have a normal structure” [7]

Nutrition plays a significant role in the growth, development, and functioning of an organism. Finally, all the energy requirements for body metabolism will be met by the nutritional status of the diet consumed. The so-called diet comprises essential nutrients that are to be mandatorily obtained from an external source, and some components will be converted into essential ingredients by the body from the consumed dietary portion. Further, the diet encompasses certain elements that are not essential nutrients but may influence the body’s metabolism. Such compounds are phytochemicals, fiber, caffeine, and others [7].

A healthy diet includes the following: (WHO Recommendation) [8, 9]

- Fruit, vegetables, legumes (e.g., lentils and beans), nuts, and whole grains (e.g., unprocessed maize, millet, oats, wheat, and brown rice)
- At least 400 g (i.e., five portions) of fruits and vegetables per day, excluding potatoes, sweet potatoes, cassava, and other starchy roots

A healthy diet comprises macro- and micronutrients. The **micronutrients** are needed by the body in smaller amounts, and they represent **vitamins, minerals, trace elements, and antioxidants**. These micronutrients have a great impact on the overall health of an individual. The micronutrients empower the body to produce hormones, enzymes, and other substances required for normal growth and development. The deficiency states of micronutrients may result in diseases affecting different parts of the body including oral tissues; further, these

deficiency states may also result in fatal and life-threatening conditions [10–13].

The mineral component of the micronutrients could be further typed as macrominerals and microminerals. The microminerals are the trace elements that exist in smaller amounts in natural and perturbed environments and play a significant role in various physiological and metabolic phenomena of the human body [14].

Learning Goals

- Epidemiological evidence on diet and nutrition in OSF
- Role of trace elements (Cu, Zn, Fe, Se) in OSF
- Role of vitamins in OSF
- Evidence from interventional studies in OSF

10.2 Epidemiological Evidence on Diet and Nutrition

An epidemiological (population-based case-control) study [15], was designed to assess the dietary factors in oral potentially malignant disorders (OPMDs) in Gujarat, India. The primary objective was to check the association of dietary components (antioxidants, vitamins, minerals, and fiber) with OPMDs of the oral cavity. A food frequency questionnaire (FFQ) was developed and tested to collect the dietary information to assess the exposure to various nutrients. Out of 5018 male subjects consuming tobacco, 318 exhibited OPMDs and qualified as cases. Age- and gender-matched healthy individuals without oral lesions were selected as the control group. The common OPMDs observed were OSF and oral leukoplakia. The FFQ was composed of questions on the frequency and quantity of 92 food items consumed, reflecting >95% of exposure to comprehensive energy, fiber, fat, minerals, and vitamins. Of all the dietary elements, the fiber component was observed to be significantly protective for oral submucous fibrosis with a 10% reduction in risk per g day ($P < 0.05$). The study revealed a strong linear protective effect (OR = 0.89) on a continuous scale (g d^{-1}), $P < 0.02$.

In another epidemiological study by Gupta et al. [16], the influence of dietary factors on OPMDs in a Kerala population (India) was assessed. A customized food frequency questionnaire (FFQ) was developed and validated for estimating the nutrient exposure in the target population. In a house-to-house survey, 5056 tobacco users were screened. Among this population, 226 people exhibited OPMDs, and were recruited as cases. Equal number of age- and gender-matched controls were selected for the control group. OSF was the

second common OPMD (next to oral leukoplakia) observed in the population. The confirmatory diagnosis of OSF was based on the presence of palpable fibrous bands. After adjusting for tobacco use, the intake of fruits, vegetables, and β -carotene showed an inverse relation to risk and an average reduction of about 10% per quartile of exposure. Zinc was shown to have a dose-response gradient and a larger effect in men. This study was undertaken to check the reduced risk of OPMDs in individuals who consume more fruits and vegetables; it confirmed the influence of these dietary factors in the development of OPMDs that had included both oral leukoplakia and oral submucous fibrosis [16].

The next section of this chapter highlights the role of dietary components in the disease process of OSF. Here, we discuss the role of trace elements, e.g., copper, zinc, iron, and selenium, in OSF and the role of vitamins in OSF.

10.3 Role of Trace Elements in OSF

10.3.1 Role of Copper

Copper (Cu) is the third most ample trace element in humans. Copper accounts for 75–100 mg of the total body. Copper is found almost in every tissue of the body, and the chief storage organs are the liver and brain, heart, kidney, and muscle. Further, copper is transported as ceruloplasmin into the plasma and excreted in the bile. In erythrocytes, 60% of the copper is found as copper-zinc metalloenzyme superoxide dismutase, and the remaining 40% is bound to other amino acids and proteins. Copper is a significant component of various enzymes involved in vital biological functions. Of significance in OSF is the synthesis of collagen and elastin as copper is a cofactor for the enzyme lysyl oxidase. Moreover, copper can act as an antioxidant protecting the tissues from oxidative stress, as well as a prooxidant causing damage to tissues [17–24].

The WHO Collaborating Centre research group based at King's College London first described raised copper levels in areca nut [25]. Copper dissolves in saliva and remains in the oral fluids for 30 min. This facilitates the uptake of copper by oral epithelial cells. The absorption of copper by oral mucosal keratinocytes is by a nonenzyme-dependent diffusion process, bound to metallothionein. With regard to areca nut chewing, there is raised level of copper seen in OSF patients. The higher copper levels upregulate the activity of lysyl oxidase causing more collagen production [26, 27]. In addition, when Cu is found in higher concentrations, there is a release of active oxygen species that further brings about oxidative damage to the cell [28, 29].

Several subsequent studies have shown elevated levels of Cu in the sera of OSF patients [29–34]. This has been attributed to the chewing of areca nut that is rich in Cu (302 nmol/g). The liver releases ceruloplasmin, a copper-carrying protein. The decreased catabolism of ceruloplasmin increases Cu levels in OSF patients. Further, the higher levels of Cu induce oxidative stress by Fenton and Haber-Weiss reaction. The serum copper levels in OSF patients show a gradual increase with advanced stages of the disease [30–35].

The role of Cu has been an interesting subject of investigation in carcinogenesis. High levels of copper within the cells generate hydroxyl radicals that can result in damage to the DNA and proteins. This may activate tumor necrosis factor-alpha and vascular endothelial growth factors. These factors are important for tumor growth and metastasis. During the malignant transformation of OSF, a four- to eightfold increase in the blood level of ceruloplasmin has been observed. Ceruloplasmin acts as a source of Cu ions, initiates LDL oxidation, and plays a role in the malignant transformation of OSF to carcinoma [36–42].

However, two studies have reported low levels of Cu in OSF patients when compared to healthy volunteers providing contradictory evidence. Varghese et al. speculated that the reduced Cu levels observed in the study could be attributed to the difference in laboratory methodologies employed and patient selection. Atomic absorption spectrophotometry was used to measure Cu levels, and the patients recruited for the study were not on any treatment in contrast to earlier studies where colorimeter was used and patients were on some form of treatment for OSF [43]. The study by Anuradha et al. also reported lower levels of Cu in OSF patients. In this study the cohort had poor dietary patterns and loss of appetite, suggesting reverse causation, also, the sample size was low [44]. However, many later studies have shown raised Cu levels in OSF patients, with atomic absorption spectrophotometry analysis [32, 45, 46].

10.3.2 Role of Zinc

Zinc (Zn) is the second most abundant transition metal in humans that appears in all enzyme systems. In blood plasma, Zn is transported by albumin and transferrin. Zn exhibits catalytic, regulatory, and structural roles in a biological system. Zn also shows antioxidant and antimicrobial properties.

An animal study (on rodents) has shown that Zn-deficient diet could result in change in the keratinization pattern (parakeratosis from orthokeratinization) of the oral mucosa.

Zn is a cofactor for the superoxide dismutase enzyme, and many studies have shown lower levels of Zn in OSF patients. This could be due to higher Cu levels and oxidative stress. Another interesting finding is lower levels of iron and higher serum levels of Zn in OSF patients. This is due to the common transporter molecule, transferrin, that carries both iron and Zn. Thus, OSF patients exhibiting lower iron levels would exhibit higher serum Zn levels.

The natural antioxidant of humans, superoxide dismutase, is a Cu–Zn protein complex that shows an anti-carcinogenic effect in OSF. Additionally, Zn reduces the activity of Cu-coupled lysyl oxidase and thus inhibits collagenic cross-linkage. Further, Zn promotes collagenic degradation via collagenases and matrix metalloproteinases. By interfering with the mucosal absorption of Cu, Zn shows an inverse relation with Cu. Higher Zn levels hamper Cu absorption as both the metals are absorbed through metallothioneins. Thus, the Cu:Zn ratio could be a reliable biomarker for assessing carcinogenesis. There is limited literature suggesting a carcinogenic effect of Zn [47–58].

The Zn level in body fluids (serum/plasma/saliva) of OSF patients has been evaluated in numerous studies. Of these, only four studies have not reported lower Zn levels in OSF patients. In one such study by Khanna et al., higher Zn levels were observed in OSF patients, but it was not statistically significant. The reason for elevated Zn levels was attributed to consumption of gutkha with higher Zn content [31, 33, 35, 44–46, 59–65].

The various effects of reduced Zn levels in OSF patients are the following:

- (i) Zn acts as a first-line defense against oxidative stress by forming cofactor Cu/Zn-superoxide dismutase enzyme [66].
- (ii) Zn is pivotal for the gene expression of metallothionein, which removes hydroxyl ions and confers protection against oxidative damage [67].
- (iii) Zn competes with other transition metals for binding sites, thereby reducing those metals from generating hydroxyl ions [67].
- (iv) Excessive cellular uptake of Zn for neutralization of free radicals [68].

Important attributes of Zn in preventing the development of malignancy [67]:

- (a) The tumor suppressor protein, p53, is Zn dependent and is involved in the repair of DNA.
- (b) The apoptotic regulating transcription factors, AP-1 and NF- κ B, show alterations with reduced cellular levels of Zn.

Lower levels of Zn may induce overexpression of COX-2 that promotes cell proliferation, prevents apoptosis, and therefore contributes to the malignant transformation of OSF [67].

10.3.3 Role of Iron

Iron (Fe) is the most abundant essential trace element in humans. Iron is an important component of heme and various enzymes in the body [69, 70]. The vital functions carried out by iron are the transport of oxygen, synthesis of DNA, energy metabolism, development and maintenance of oral mucosa. Iron deficiency leads to Plummer-Vinson's syndrome. Patients who have Plummer-Vinson's syndrome (sideropenic dysphagia) exhibit features of anemia; further glossitis, angular cheilitis, and koilonychia are noticed. These patients have greater risk of developing oral, postcricoidal, and esophageal carcinomas.

Further, the investigations on iron levels in OSF patients have revealed lower serum iron concentrations when compared to healthy controls [44, 60, 71–80].

The suggestions for diminished levels of iron in OSF patients could be due to:

- (i) Excessive use of iron for the hydroxylation of lysine and proline during collagen synthesis.
- (ii) The mechanical injury caused by areca nut chewing hampers intake of a nutritionally balanced diet.
- (iii) Vegetarian diet may predispose an individual to greater depletion of iron stores.

Ultimately, chronic iron deficiency in areca nut chewers is a factor that facilitates the development of OSF. Further, features of anemia have been noticed in the advanced stage of OSF [81, 82].

10.3.4 Role of Selenium

Selenium is yet another vital trace element that is an important constituent of antioxidant enzymes: glutathione peroxidase and thioredoxin reductase [83]. Lower serum levels of selenium have been reported in OSF patients when compared to normal individuals [84].

10.4 Role of Vitamins

In a case-control study (OSF, $n = 40$; control, $n = 25$), deficiency of vitamin B12 and iron was reported in OSF patients when compared to healthy volunteers. Further, the red cell indices such as packed cell volume, mean corpuscular volume, and mean corpuscular Hb were significantly reduced in OSF patients [85].

Another case-control study showed high frequencies of vitamin B12, folic acid deficiencies, and gastric parietal cell antibody positivity in OSF patients when compared to healthy individuals [86].

Shetty et al. in their study observed lower serum and salivary levels of ascorbic acid with progressive worsen-

ing of histopathological grading of OSF. The likely reason for lower levels of ascorbic acid is that it may have been used for the excessive synthesis of collagen during the progression of OSF [87].

Another study evaluated mean serum vitamin A and vitamin E levels in OSF patients. There was no statistically significant difference in these vitamin levels between OSF and control groups. It has been suggested that these vitamins are used by the tissues to combat oxidative stress generated due to the consumption of areca nut [88].

10.5 Interventional Studies

In an interventional study on OSF by Maher et al. [89], the beneficial clinical response to multiple micronutrients was evaluated in Karachi, Pakistan. Out of 169 OSF subjects recruited for the study, 117 compliant individuals were given daily oral micronutrient (vitamins and minerals) supplementations for 1–3 years. There was a significant improvement in symptoms, such as burning sensation, intolerance to spicy food, and restricted mouth opening. The interincisor distance increased in 48 (41%) and there was regression of concomitant lesions like oral leukoplakia and/or erythroplakia [89].

Summary

Points of clinical relevance:

Factors that promote fibrosis in Oral Submucous Fibrosis are as follows:

- Low dietary fibre
- Higher levels of copper
- Lower levels of Zinc, Iron, and Selenium
- Lower levels of Vitamins A, B, C, and E
- Dietary supplementation of the micronutrients has shown improvement in signs and symptoms of OSF

References

1. Pindborg JJ, Sirsat SM. Oral submucous fibrosis. *Oral Surg Oral Med Oral Pathol.* 1966;22(6):764–79. [https://doi.org/10.1016/0030-4220\(66\)90367-7](https://doi.org/10.1016/0030-4220(66)90367-7).
2. Sinor PN, Gupta PC, Murti PR, Bhonsle RB, Daftary DK, Mehta FS, et al. A case-control study of oral submucous fibrosis with special reference to the etiologic role of areca nut. *J Oral Pathol Med.* 1990;19(2):94–8.
3. Maher R, Lee AJ, Warnakulasuriya KA, Lewis JA, Johnson NW. Role of areca nut in the causation of oral submucous fibrosis: a case-control study in Pakistan. *J Oral Pathol Med.* 1994;23(2):65–9. <https://doi.org/10.1111/j.1600-0714.1994.tb00258.x>.
4. Murti PR, Bhonsle RB, Gupta PC, Daftary DK, Pindborg JJ, Mehta FS. Etiology of oral submucous fibrosis with special reference to the role of areca nut chewing. *J Oral Pathol Med.* 1995;24(4):145–52. <https://doi.org/10.1111/j.1600-0714.1995.tb01156.x>.
5. Tilakaratne WM, Klinikowski MF, Saku T, Peters TJ, Warnakulasuriya S. Oral submucous fibrosis: review on etiology and pathogenesis. *Oral Oncol.* 2006;42(6):561–8. <https://doi.org/10.1016/j.oraloncology.2005.08.005>.
6. Xu H, Lyu F, Song J, Xu Y, Jiang E, Shang Z, et al. Research achievements of oral submucous fibrosis: progress and prospect. *Biomed Res Int.* 2021.; Article ID 6631856; <https://doi.org/10.1155/2021/6631856>.
7. World Cancer Research Fund/American Institute of Cancer Research. Diet, nutrition, physical activity, and cancer: a global perspective. Continuous Update Project Expert Report 2018. www.dietandcancerreport.org
8. World Health Organization—Healthy Diet. <https://www.who.int/news-room/fact-sheets/detail/healthy-diet>, last accessed 28th Aug 2021
9. Diet, nutrition and the prevention of chronic diseases: report of a Joint WHO/FAO Expert Consultation. WHO Technical Report Series, No. 916. Geneva: World Health Organization; 2003.
10. World Health Organization—Health Topics. https://www.who.int/health-topics/micronutrients#tab=tab_1, last accessed 28th Aug 2021
11. Enwonwu CO, Phillips RS, Falkler WA Jr. Nutrition and oral infectious diseases: state of the science. *Compend Contin Educ Dent.* 2002;23(5):431–4.
12. Moynihan PJ. The role of diet and nutrition in the etiology and prevention of oral diseases. *Bull World Health Organ.* 2005;83(9):694–9.
13. Chen M, Andersen R, Barmes M, David E, Leclercq M, Lyttle H, et al. Comparing oral health care systems: a second international collaborative study. World Health Organization; 1997. <https://apps.who.int/iris/handle/10665/41976>
14. Bhattacharya PT, Misra SR, Hussain M. Nutritional aspects of essential trace elements in oral health and disease: an extensive review. *Scientifica (Cairo).* 2016;2016:5464373. <https://doi.org/10.1155/2016/5464373>.
15. Gupta PC, Hebert JR, Bhonsle RB, Sinor PN, Mehta H, Mehta FS. Dietary factors in oral leukoplakia and submucous fibrosis in a population-based case control study in Gujarat, India. *Oral Dis.* 1998;4:200–6.
16. Gupta PC, Hebert JR, Bhonsle RB, Murti PR, Mehta H, Mehta FS. Influence of dietary factors on oral precancerous lesion in a population-based case-control study in Kerala, India. *Cancer.* 1999;85(9):1885–93.
17. Adelstein SJ, Vallee BL. Copper metabolism in man. *N Engl J Med.* 1961;265:892–7. <https://doi.org/10.1056/NEJM196111022651806>.
18. Harris ED. Copper homeostasis: the role of cellular transporters. *Nutr Rev.* 2001;59(9):281–5. <https://doi.org/10.1111/j.1753-4887.2001.tb07017.x>.
19. Araya M, Pizarro F, Olivares M, Arredondo M, Gonzalez M, Mendez M. Understanding copper homeostasis in humans and copper effects on health. *Biol Res.* 2006;39(1):183–7. <https://doi.org/10.4067/S0716-97602006000100020>.
20. Bonham M, O'Connor J, Hannigan B, Strain J. The immune system as a physiological indicator of marginal copper status? *Brit J Nutr.* 2002;87(5):393–403. <https://doi.org/10.1079/BJN2002558>.
21. Rakel D. Integrative medicine. 2nd ed. Philadelphia, PA: Saunders Elsevier; 2007.
22. Davis CD. Low dietary copper increases fecal free radical production, fecal water alkaline phosphatase activity and cyto-

- toxicity in healthy men. *J Nutr.* 2003;133(2):522–7. <https://doi.org/10.1093/jn/133.2.522>.
23. Willis MS, Monaghan SA, Miller ML, McKenna RW, Perkins WD, Levinson BS, Bhushan V, Kroft SH. Zinc-induced copper deficiency: a report of three cases initially recognized on bone marrow examination. *Am J Clin Pathol.* 2005;123(1):125–31. <https://doi.org/10.1309/v6gvwy2qtyd5c5pj>.
 24. Osredkar J, Sustar N. Copper and zinc, biological role and significance of copper/zinc imbalance. *J Clin Toxicol.* 2011;3(2161):0495.
 25. Trivedy CR, Warnakulasuriya KA, Peters TJ, Senkus R, Hazarey VK, Johnson NW. Raised tissue copper levels in oral submucous fibrosis. *J Oral Pathol Med.* 2000;29(6):241–8. <https://doi.org/10.1034/j.1600-0714.2000.290601.x>.
 26. Trivedy C, Baldwin D, Warnakulasuriya S, Johnson N, Peters T. Copper content in Areca catechu (Betel nut) products and oral submucous fibrosis. *Lancet.* 1997;349(17):1447.
 27. Trivedy C, Warnakulasuriya KA, Hazarey VK, Tavassoli M, Sommer P, Johnson NW. The upregulation of lysyl oxidase in oral submucous fibrosis and squamous cell carcinoma. *J Oral Pathol Med.* 1999;28(6):246–51. <https://doi.org/10.1111/j.1600-0714.1999.tb02033.x>.
 28. Desai VD, Kumar MVS, Bathi RJ, Gaurav I, Sharma R. Molecular analysis of trace elements in oral submucous fibrosis and future perspectives. *Universal Res J Dent.* 2014;4(1):26–35. <https://doi.org/10.4103/2249-9725.127070>.
 29. Rajalalitha P, Vali S. Molecular pathogenesis of oral submucous fibrosis—a collagen metabolic disorder. *J Oral Pathol Med.* 2005;34(6):321–8. <https://doi.org/10.1111/j.1600-0714.2005.00325.x>.
 30. Theophanides T, Anastassopoulou J. Copper and carcinogenesis. *Crit Rev Oncol Hematol.* 2002;42(1):57–64. [https://doi.org/10.1016/s1040-8428\(02\)00007-0](https://doi.org/10.1016/s1040-8428(02)00007-0).
 31. Gaetke LM, Chow CK. Copper toxicity, oxidative stress, and antioxidant nutrients. *Toxicology.* 2003;189(1–2):147–63. [https://doi.org/10.1016/s0300-483x\(03\)00159-8](https://doi.org/10.1016/s0300-483x(03)00159-8).
 32. Shettar SS. Estimation of serum copper and zinc levels in patients with oral submucous fibrosis. *J Indian Acad Oral Med Radiol.* 2010;22(4):193–6.
 33. Tadakamadla J, Kumar S, Mamatha GP. Evaluation of serum copper and iron levels among oral submucous fibrosis patients. *Med Oral Patol Oral Cir Bucal.* 2011;16(7):e870–3. <https://doi.org/10.4317/medoral.17083>.
 34. Ayinampudi BK, Narsimhan M. Salivary copper and zinc levels in oral pre-malignant and malignant lesions. *J Oral Maxillofac Pathol.* 2012;16(2):178–82. <https://doi.org/10.4103/0973-029X.98452>.
 35. Rajendran R, Vasudevan DM, Vijayakumar T. Serum levels of iron and proteins in oral submucous fibrosis (OSMF). *Ann Dent.* 1990;49(2):23–25, 45.
 36. Hosthor SS, Mahesh P, Priya SA, Sharada P, Jyotsna M, Chitra S. Quantitative analysis of serum levels of trace elements in patients with oral submucous fibrosis and oral squamous cell carcinoma: a randomized cross-sectional study. *J Oral Maxillofac Pathol.* 2014;18(1):46–51. <https://doi.org/10.4103/0973-029X.131902>.
 37. Jani YV, Chaudhary AR, Dudhia BB, Bhatia PV, Soni NC, Patel PS. Evaluation of role of trace elements in oral submucous fibrosis patients: a study on Gujarati population. *J Oral Maxillofac Pathol.* 2017;21(3):455. https://doi.org/10.4103/jomfp.JOMFP_106_14.
 38. Sutton HC, Winterbourn CC. On the participation of higher oxidation states of iron and copper in Fenton reactions. *Free Radic Biol Med.* 1989;6(1):53–60. [https://doi.org/10.1016/0891-5849\(89\)90160-3](https://doi.org/10.1016/0891-5849(89)90160-3).
 39. Nasulewicz A, Mazur A, Opolski A. Role of copper in tumour angiogenesis—clinical implications. *J Trace Elem Med Biol.* 2004;18(1):1–8. <https://doi.org/10.1016/j.jtemb.2004.02.004>.
 40. Al-Rawi NH, Talabani NGA. Quantitative analysis of trace elements in saliva of oral cancer patients from Iraq. *J Coll Dent.* 2005;17(2):32–5.
 41. Mukhopadhyay CK, Mazumder B, Lindley PF, Fox PL. Identification of the prooxidant site of human ceruloplasmin: a model for oxidative damage by copper bound to protein surfaces. *Proc Natl Acad Sci U S A.* 1997;94(21):11546–51. <https://doi.org/10.1073/pnas.94.21.11546>.
 42. Jayadeep A, Raveendran Pillai K, Kannan S, Nalinakumari KR, Mathew B, Krishnan Nair M, Menon VP. Serum levels of copper, zinc, iron and ceruloplasmin in oral leukoplakia and squamous cell carcinoma. *J Exp Clin Cancer Res.* 1997;16(3):295–300.
 43. Varghese I, Sugathan CK, Balasubramoniyam G, Vijayakumar T. Serum copper and zinc levels in premalignant and malignant lesions of the oral cavity. *Oncology.* 1987;44(4):224–7. <https://doi.org/10.1159/000226482>.
 44. Anuradha CD, Devi CSS. Studies on the hematological profile and trace elements in oral submucous fibrosis. *J Clin Biochem Nutr.* 1995;19:9–17.
 45. Gupta RP, Rai K, Hemani DD, Gupta AK. Study of trace elements (copper & zinc) in oral submucous fibrosis. *Indian J Otolaryngol.* 1987;39:104–6.
 46. Khanna S, Udas AC, Kumar GK, Suvarna S, Karjodkar FR. Trace elements (copper, zinc, selenium and molybdenum) as markers in oral submucous fibrosis and oral squamous cell carcinoma. *J Trace Elem Med Biol.* 2013;27(4):307–11. <https://doi.org/10.1016/j.jtemb.2013.04.003>.
 47. Wapner RA. Protein nutrition and mineral absorption. Boca Raton, FL: CRC Press; 1990.
 48. Broadley MR, White PJ, Hammond JP, Zelko I, Lux A. Zinc in plants. *New Phytol.* 2007;173(4):677–702. <https://doi.org/10.1111/j.1469-8137.2007.01996.x>.
 49. Whitney EN, Rolfes SR. Understanding nutrition. 10th ed. Boston, MA: Thomson Learning; 2010.
 50. Sandstead HH. Understanding zinc: recent observations and interpretations. *J Lab Clin Med.* 1994;124(3):322–7.
 51. McCarthy TJ, Zeelie JJ, Krause DJ. The antimicrobial action of zinc ion/antioxidant combinations. *J Clin Pharmacy Therap.* 1992;17(1):51–4. <https://doi.org/10.1111/j.1365-2710.1992.tb01265.x>.
 52. Fabris N, Mocchegiani E. Zinc, human diseases and aging. *Aging Clin Exp Res.* 1995;7:77–93.
 53. Milbury PE, Richer AC. Understanding the antioxidant controversy: scrutinizing the “Fountain of Youth”. Greenwood Publishing Group; 2008.
 54. Das M, Das R. Need of education and awareness towards zinc supplementation: a review. *Int J Nutr Metab.* 2012;4(3):45–50. <https://doi.org/10.5897/IJNAM11.043>.
 55. Jayadeep A, Raveendran KP, Kannan S, Nalinakumari KR, Mathew B, Krishnan NM, et al. Serum levels of copper, zinc, iron, and ceruloplasmin in oral leukoplakia and squamous cell carcinoma. *J Exp Clin Can Res.* 1997;16(3):295–300.
 56. Ray JG, Ghosh R, Mallick D, Swain N, Gandhi P, Ram SS, Selvaraj S, Rathore A, Mathummal S, Chakraborty A. Correlation of trace elemental profiles in blood samples of Indian patients with leukoplakia and oral submucous fibrosis. *Biol Trace Elem Res.* 2011;144(1–3):295–305. <https://doi.org/10.1007/s12011-011-9091-0>.
 57. Desai V, Kumar M, Bathi R, Gaurav I, Sharma R. Molecular analysis of trace elements in oral submucous fibrosis and future perspectives. *Univ Res J Dent.* 2014;4(1):26–35. <https://doi.org/10.4103/2249-9725.127070>.

58. Mulware SJ. Trace elements and carcinogenicity: a subject in review. *3 Biotech.* 2013;3(2):85–96. <https://doi.org/10.1007/s13205-012-0072-6>.
59. Díez M, Arroyo M, Cerdán FJ, Muñoz M, Martín MA, Balibrea JL. Serum and tissue trace metal levels in lung cancer. *Oncology.* 1989;46(4):230–4. <https://doi.org/10.1159/000226722>.
60. Balpande AR, Sathawane RS. Estimation and comparative evaluation of serum iron, copper, zinc, and copper/zinc ratio in oral leukoplakia, submucous fibrosis and squamous cell carcinoma. *J Indian Acad Oral Med Radiol.* 2010;22(2):73–6.
61. More CB, Patel H. Trace elements in potentially oral malignant disorders and oral malignant lesion—a biochemical study. *Int J Oral Maxillofac Dis.* 2016;1(2):1–7.
62. Yadav A, Kumar L, Misra N, Deepak U, Shiv Kumar GC. Estimation of serum zinc, copper, and iron in the patients of oral submucous fibrosis. *Natl J Maxillofac Surg.* 2015;6(2):190–3. <https://doi.org/10.4103/0975-5950.183851>.
63. Srileka M. Copper and zinc level in oral submucosal fibrosis (OSMF) patients. *J Pharm Sci Res.* 2015;7(8):573–4.
64. Neethi H, Patil S, Rao RS. Estimation of serum copper and zinc levels in oral submucous fibrosis: an atomic absorption spectroscopic study. *J Contemp Dent Pract.* 2013;14(5):801–5. <https://doi.org/10.5005/jp-journals-10024-1406>.
65. Kode MA, Karjodkar FR. Estimation of the serum and the salivary trace elements in OSMF patients. *J Clin Diagn Res.* 2013;7(6):1215–8. <https://doi.org/10.7860/JCDR/2013/5207.3023>.
66. Klotz LO, Kröncke KD, Buchczyk DP, Sies H. Role of copper, zinc, selenium and tellurium in the cellular defense against oxidative and nitrosative stress. *J Nutr.* 2003;133(5 Suppl. 1):1448S–51S. <https://doi.org/10.1093/jn/133.5.1448S>.
67. Ho E. Zinc deficiency, DNA damage and cancer risk. *J Nutr Biochem.* 2004;15(10):572–8. <https://doi.org/10.1016/j.jnutbio.2004.07.005>.
68. Prasad AS. Zinc is an antioxidant and anti-inflammatory agent: its role in human health. *Front Nutr.* 2014;1:14. <https://doi.org/10.3389/fnut.2014.00014>.
69. Vasudevan DM, Sreekumari S. Textbook of biochemistry for medical students. 5th ed. New Delhi, India: Jaypee; 2007.
70. Satyanarayana U, Chakrapani U. Essentials of biochemistry, book and allied. 2nd ed. Kolkata, India: Elsevier; 2008.
71. Shetty SR, Babu S, Kumari S, Shetty P, Hegde S, Karikal A. Status of trace elements in saliva of oral precancer and oral cancer patients. *J Cancer Res Ther.* 2015;11(1):146–9. <https://doi.org/10.4103/0973-1482.137973>.
72. Okade AR, Hallikeri KS, Trivedi DJ. Salivary estimation of copper, iron, zinc, and manganese in oral submucous fibrosis patients: a case-control study. *Clin Can Invest J.* 2015;4(3):302–6. <https://doi.org/10.4103/2278-0513.156075>.
73. Tiwari R, David CM, Mahesh DR, Sambargi U, Rashmi KJ, Benakanal P. Assessment of serum copper, iron and immune complexes in potentially malignant disorders and oral cancer. *Braz Oral Res.* 2016;30(1):e101. <https://doi.org/10.1590/1807-3107BOR-2016.vol30.0101>.
74. Khanna SS, Karjodkar FR. Circulating immune complexes and trace elements (Copper, Iron and Selenium) as markers in oral precancer and cancer: a randomised, controlled clinical trial. *Head Face Med* 2006;2:33. doi: <https://doi.org/10.1186/1746-160X-2-33>.
75. Shetty SR, Babu S, Kumari S, Hegde S, Karikal A. Role of serum trace elements in oral precancer and oral cancer: a biochemical study. *J Can Res Treat.* 2013;1(1):1–3.
76. Guruprasad R, Nair PP, Singh M, Singh M, Singh M, Jain A. Serum vitamin c and iron levels in oral submucous fibrosis. *Indian J Dent.* 2014;5(2):81–5. <https://doi.org/10.4103/0975-962X.135266>.
77. Thakur M, Guttikonda VR. Estimation of hemoglobin, serum iron, total iron-binding capacity and serum ferritin levels in oral submucous fibrosis: a clinicopathological study. *J Oral Maxillofac Pathol.* 2017;21(1):30–5. https://doi.org/10.4103/jomfp.JOMFP_131_15.
78. Ganapathy KS, Gurudath S, Balikai B, Ballal S, Sujatha D. Role of iron deficiency in oral submucous fibrosis: an initiating or accelerating factor. *J Indian Acad Oral Med Radiol.* 2011;23(1):25–8. Retrieved from <https://www.proquest.com/scholarly-journals/role-iron-deficiency-oral-submucous-fibrosis/docview/868927327/se-2>
79. Novacek G. Plummer-Vinson syndrome. *Orphanet J Rare Dis.* 2006;1:36. <https://doi.org/10.1186/1750-1172-1-36>.
80. Lee J, Cho Y. Effect of ascorbic acid, silicon and iron on collagen synthesis in the human dermal fibroblast cell (HS27). *FASEB J.* 2008;22(S2):672. https://doi.org/10.1096/fasebj.22.2_supplement.672.
81. Pawlak R, Berger J, Hines I. Iron status of vegetarian adults: a review of literature. *Am J Lifestyle Med.* 2016;12(6):486–98. <https://doi.org/10.1177/1559827616682933>.
82. Mohammad H, Hadi NI, Younus S, Ahmed F, Younus N. Potentially significant biomarkers in oral submucous fibrosis. *Pakistan J Med Dent.* 2015;4(2):51–6.
83. Rayman MP. Selenium and human health. *Lancet.* 2012;379(9822):1256–68. [https://doi.org/10.1016/S0140-6736\(11\)61452-9](https://doi.org/10.1016/S0140-6736(11)61452-9).
84. Khanna S, Udas AC, Kumar GK, Suvarna S, Karjodkar FR. Trace elements (copper, zinc, selenium, and molybdenum) as markers in oral submucous fibrosis and oral squamous cell carcinoma. *J Trace Elem Med Biol.* 2013;27(4):307–11. <https://doi.org/10.1016/j.jtemb.2013.04.003>.
85. Patil DJ, Joshi M. Evaluation of hematological profile in oral submucous fibrosis: a cross-sectional study. *J Oral Maxillofac Pathol.* 2020;24:575.
86. Wang YP, Wu YC, Cheng SJ, Chen HM, Sun A, Chang JY. High frequencies of vitamin B12 and folic acid deficiencies and gastric parietal cell antibody positivity in oral submucous fibrosis patients. *J Formos Med Assoc.* 2015;114(9):813–9. <https://doi.org/10.1016/j.jfma.2015.05.011>.
87. Shetty SR, Babu S, Kumari S, Shetty P, Vijay R, Karikal A. Evaluation of micronutrient status in serum and saliva of oral submucous fibrosis patients: a clinicopathological study. *Indian J Med Paediatr Oncol.* 2012;33(4):224–6. <https://doi.org/10.4103/0971-5851.107087>.
88. Jain A, Kaur G, Ranjan R, Singh D, Porwal D, Shekhawat A. Estimation of serum level of Vitamin A and E in OSMF patients. *Int J Appl Dent Sci.* 2020;6(2):174–6.
89. Maher R, Aga P, Johnson NW, Sankaranarayanan R, Warnakulasuriya S. Evaluation of multiple micronutrient supplementation in the management of oral submucous fibrosis in Karachi. *Pakistan. Nutr Cancer.* 1997;27(1):41–7. <https://doi.org/10.1080/01635589709514499>.