

Chapter 4

Nano and Microencapsulation of Foods, Vitamins and Minerals



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Abbreviations

AAO	Aryl-alcohol oxidase
BHA	Butylated hydroxyanisole
BHT	Butylated hydroxytoluene
CD	Cyclodextrin
CMC	Critical micellar concentration
CPO	Chloroperoxidase
CTAB	Cationic surfactant cetyl trimethyl ammonium bromide
DFS	Dual fortified salt
EPC	Egg phosphatidylcholine
FeCl ₃	Ferric chloride
FeSO ₄	Ferrous sulphate
FFAs	Free fatty acids
FIP WP	Ferri polyphosphate whey protein
GA	Gum Arabic
GIT	Gastrointestinal tract
IDFA	International Dairy Foods Association
L-Ca	Encapsulated calcium in liposome
LMFS	Large microencapsulated FeSO ₄
MD	Maltodextrin
Mm	Micrometer
MRI	Magnetic resonance imaging
nm	Nanometer
SDS	Sodium dodecyl sulfate
SLNs	Solid-liquid nanoparticles
SMFS	Small microencapsulated FeSO ₄

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TBA	Thiobarbituric acid
TBARS	TBA reactive substance
TFS	Triple fortified salt
USDA	United State Department of Agriculture
WPI	Whey protein isolate

4.1 Introduction

Nanoparticles delivery approach has obtained constant recognition from the food industry in the last few decades for applications that range from the slow-release vitamin and mineral formulations and micronutrients encapsulated and stabilized for nutritional supplement utilization. Because of the current growth and prominence of “niche” subgroups of useful ingredients, the colloidal delivery approach (dealing mainly with the micro and nanoencapsulations) has been considered as a useful resource for resolving multiple difficulties associated with food fortification. Encapsulation can be defined as either layering a droplet, gas, or solid particle center with a solid polymeric matrix or integrating active components into polymer matrices to reduce the interaction of the encapsulated substances with the surrounding environment [1]. Colors, flavors, preservatives, and other bioactive components are added to packaged foods to improve their aesthetic and shelf life. Furthermore, fortified and value-added items frequently require specific health-promoting bioactives, such as micronutrients and nutraceuticals. There are numerous challenges in food fortification; among which, durability, matrix compatibility, and organoleptic changes are common difficulties [2]. Fortification is a difficult process due to difficulties in product processing and issues related to in vivo bioavailability, controlled release, and bio accessibility. Nano- and microencapsulation can address some or all of these problems. The gastro-intestinal (GI) tract stability of the encapsulated materials could also be improved to enable controlled drug release at suitable GI targets [3]. The present chapter discusses a variety of colloidal systems (micro and nano) and their applications in vitamin and mineral encapsulation and delivery.

4.2 Foods and Nutraceuticals Vitamins

4.2.1 *Vitamins: Dietary and Biological Needs*

Probiotics should include vitamins necessary for human growth and development (like niacin and choline). The advantages of nutraceuticals are more than simply preventing illness. Oil and water-soluble vitamins function as antioxidants and hormone regulators. Vitamin B-complex is beneficial for vision, immunity, bones, teeth, and skin. Vitamin E improves cognitive function.

4.2.2 *Vitamins: Stability and Formulation Concerns*

Vitamins encounter a number of challenges, including oxidative stability. Other additives may also impede water-soluble vitamins, impacting texture, appearance, and storage. Vitamin A (retinoids): When exposed to air and heat, trace metal ions Fe^{2+} and Cu^{2+} increase the oxidation sensitivity of vitamin A.

Thiamine, a heat and UV-sensitive vitamin B, is a potent Maillard reactive due to its chemistry. Baking, pasteurizing, or boiling thiamine-fortified meals can reduce their content by up to 50%. B2 vitamin: Riboflavin is a brilliant yellow pigment soluble in water. Visible spectrum light below 500–520 nm can degrade riboflavin. **Vitamin B3:** Niacin is highly stable when exposed to heat, light, air, and alkalis. Niacin can be destabilized by other micronutrients, including minerals.

Vitamin B5: Pantothenic acid decomposes when heated under slightly acidic to neutral conditions but stays stable within the pH range of 5–7. It is used as calcium and sodium salts to enhance stability in the gastrointestinal pH. **Vitamin B6:** Pyridoxine is heat stable, but not pyridoxal or pyridoxamine. **Vitamin B6** is degraded by oxidation, UV radiation, and alkaline conditions. **Vitamin B7** appears to be reasonably stable; however, heat treatment causes relatively minor losses. It is stable in air at neutral and acidic PH, but susceptible to alkaline pH. **Vitamin B9:** Folic acid is a reddish, tasteless, and odorless vitamin, which is water soluble at 12.5 mg/mL. Light, air, acid, and alkali deactivate it. **Vitamin B12:** In addition to being heat stable, light, oxygen, acid, and alkali cause vitamin B12 to lose its activity. **Vitamin C:** Ascorbic acid is easily degraded by metals such as copper and iron during production and storage. In neutral or alkaline conditions, it is oxidized by air. Metals, particularly copper and iron, and enzymes expedite vitamin C breakdown (particularly enzymes containing copper or iron, such as ascorbic acid oxidase). **Vitamin D:** D2 and D3 are crystalline compounds white to yellow in color. They are insoluble in vegetable oils and water but can be dissolved in organic solvents. Light, oxygen, and acid quickly dissociate both these vitamins, especially in solution and powder forms. However, they tend to isomerize in oily solutions. The crystalline compounds have good thermal stability. **Vitamin E:** Food processing and storage reduce vitamin E levels due to light, oxygen, and heat. Some foods can lose half their flavor after just 2 weeks. Vegetable oils are destroyed by frying. During storage, tocopherol esters (acetate and succinate of tocopherol) resist oxidation, which makes them useful in nutritional supplements. **Vitamin K:** Light and alkaline substances decompose K1 into a yellow-oily liquid. Vitamin K2 is a crystalline substance that is resistant to heat and reducing agents but susceptible to acid, alkali, light, and oxidizing agents.

4.3 The Encapsulation of Vitamins (Nano and Micro) Colloidal Form

4.3.1 *Dispersions of Solids in Liquids*

4.3.1.1 Microparticles

In recent years, microencapsulation has become more popular in pharmaceuticals. It has been found that functional ingredients can be delivered in microstructures of size between 1 and 1000 μm . In contrast to microspheres, the microcapsule core is enclosed in a well-formed polymer shell. Encapsulated vitamins are protected from the external environment. Vitamin C is a well-studied model vitamin for encapsulation. Vitamin C powder is stable but not in the solution form as light and oxygen readily destroy vitamin C in the solution. Trace metal ions and formulation can also affect vitamin C stability. Thus, processing can easily degrade vitamin C, causing discoloration. Vitamin C can also interact with other substances, changing the product's flavor or color. Microencapsulation has been widely used for the stabilization and processing of vitamin C in food items. A wall-forming material, ethyl cellulose, and carnauba wax were used to study the characteristics of ascorbic acid and four different encapsulation methods. Carnauba wax releases slower than ethyl cellulose. Core-shell microcapsules encapsulating vitamin C (2–5 μm) was prepared using a new interfacial/emulsion reaction. Folic acid was encapsulated in alginate and pectin microparticles [4]. Depending on the experiment, the microparticles were of 300–650 nm in size. Less trapping efficiency means less folic acid release. Microencapsulation has also been utilized for light-sensitive vitamins like A and D. Gelatin-acacia coacervates contain vitamin A palmitate. Acacia was mixed with vitamin A palmitate in maize oil, and the pH was adjusted to make microcapsules. This research focused on the effects of hydrocolloid mixing ratio, hardening agent concentration, and drying technique. An average diameter of 25 μm and encapsulation efficiency of 83.43% were achieved. Alginate and chitosan were used by Albertini et al. to create a double-layer microcapsule containing vitamin A palmitate [10]. Encapsulation increased vitamin A palmitate storage stability compared to the pure form. A novel sort of microcapsule was recently reported by solidifying heated emulsion droplets. Tocopherol, a liposoluble vitamin, was effectively encapsulated into microcapsules. Sunflower oil and water were emulsified at 85 °C, and then diluted in cold water to create micro-capsule-size particles. The sudden change in temperature led the oil droplets to freeze. Making microcapsules with a core-shell structure is as simple as adding calcium chloride to the cold water phase and stirring. Firstly, α -tocopherol was dissolved in the oil phase, and then encapsulation was accomplished as previously described. The antioxidant tests showed that encapsulation has no influence on α -tocopherol antioxidant activity [5].

4.3.1.2 Polymeric Nanoparticles

The size of colloidal particles is in the sub-micron range. This category comprises polymer and solid lipid nanoparticulate systems. Adding vitamins and minerals to a “clear” food beverage that makes fat-based foods taste better without changing their overall taste, or creating a bioavailable formula relies on tiny particles that are easily dissolved. Vitamins encapsulated in colloidal particles can be used in products like fortified milk, juices, and cooking oils. Liposoluble actives can be encapsulated by antisolvents, or nanoprecipitation, and co-precipitating them with hydrophilic polymers makes it convenient to load them in colloidal particles [6]. This approach has been used to encapsulate liposoluble vitamins like A, D, E, and K. Vitamin E (tocopherol) was co-precipitated with wheat gluten vegetable protein fractions (gliadins). They were formed as colloidal particles with an encapsulation effectiveness of 77% or higher and dual release features. The anti-solvent technique works well with zein, corn protein rich in proline. Zein interacts with particles composed of charged biopolymers, such as caseinate and chitosan. Submicron nanoparticles were synthesized using the ionic zein and α -tocopherol with hydrophobic interactions [7]. These particles were poly dispersible and ranged from 300–1000 nm in size. The zein-chitosan combination generated substantially smaller and homogenous sized particles (400–500 nm). When chitosan was added to the preparation process, it improved the control of the encapsulated vitamin’s release. A liposoluble vitamin, cholecalciferol, was coated with zein-carboxymethyl chitosan nanoparticles [8].

Phase separation was used to produce monodispersed nanoparticles as small as 120 nm. The controlled release of vitamin D3 from nanoparticle increased its photostability. These solid lipid nanoparticles (SLNs) have been employed to deliver lipophilic bioactives for food applications. The initial stage of making an oil-soluble vitamin loaded SLNs is dispersion in the lipid phase. For high melting lipid carriers, dispersion of vitamins in lipid carriers takes place at high temperature (about 80 °C). Aqueous surfactant solution is subsequently mixed with the melt, resulting in the spontaneous production of SLNs [9]. SLNs are made from glyceryl behenate and tripalmitate encapsulated vitamin A and its palmitate. Factors affecting size and shape were assessed, such as manufacturing process, surfactant system, and lipid type, which have an influence on experimental outcomes. The interplay between these variables affects the internal and external architecture of SLNs, and therefore the distribution of vitamins inside SLNs (in the lipid matrix or in the outer shell).

4.3.2 *Liquid-in-Liquid Dispersions*

4.3.2.1 Emulsions, Microemulsions and Nanoemulsions

In food science, emulsions are the most explored colloidal system, forming the basis of a wide range of edible goods, like ice creams, sauces, spreads, whipped creams, etc.

A mixture is made up of dispersed and continuous liquids. The use of surface active amphiphilic emulsifiers stabilizes the internal phase. Emulsions oil/water (o/w) or water/oil (w/o) are commonly used in food science and technology. For margarines and butter, a continuous phase of solid fat gives the finished product a structural appearance. This is because the water continuous phase is less viscous. However, changing the internal phase can affect emulsion rheology in general, it is either gelled or increases the proportion of internal phase (or both). Double or multiple emulsions generate an o/w emulsion, which is further emulsified into an oil phase or water phase [10]. Size and stability of droplets can also be considered when classifying emulsions. Emulsions having droplet sizes greater than 1 μm are termed as macroemulsions. The droplets of size 1 μm /1000 nm or smaller fall. When homogenization is performed, high-pressure or high-shear homogenizers are used, which might produce nanoemulsions or mini emulsions. The droplet size of microemulsions is typically smaller than 100 nanometers and they are formed spontaneously within liquids. Surfactant and co-surfactant synergy reduces interfacial tension, resulting in spontaneous production [11].

Despite their popularity, macroscale emulsions are thermodynamically unstable and tend to cream and coalesce with time. Emulsions are frequently employed to encapsulate and supply both liposoluble and water-soluble vitamins. This enhances water dispersibility and bioavailability while preventing oil-soluble vitamins from deterioration in the product condition and the human body condition. Numerous studies have used the most active form of vitamin E that is α -tocopherol emulsion, which is food-grade [12]. It was determined that the distribution of α -tocopherol in macroemulsion systems may be enhanced by emulsions of α -tocopherol using a pseudo phase model for thermodynamically stable microemulsions. As a result of α -tocopherol's surfactant-like properties (polar phenolic headgroup with hydrocarbon tail), they identified it mostly in the interfacial region. High pressure homogenization encapsulated α -tocopherol in o/w nanoemulsions with droplet sizes typically smaller than 500 nm [13]. The protein coating the tocopherol-filled fat droplets protected them from oxidation. Vitamin E-loaded nanoemulsions were recently described. This novel nanoemulsion contains vitamin E with >99% efficiency and particles are on average 78 nm in size. Microemulsion consisting of water, vitamin E, ethyl butyrate, EL-35, and ethanol was also studied. A regulated vitamin E released from microemulsions was shown having droplet size of 20 nm. Vitamins A and D can also be encapsulated in emulsions. Encapsulated trans retinoic acid/retinol is required in a variety of emulsion processes [14]. Besides high loading, encapsulation of vitamin A must address retention of loaded vitamins. Researchers employed an oil gelling agent, and coated oil droplets with solid fat as an alternative to oil to enhance vitamin retention in the oil phase thus encapsulating the vitamin. By means of a high-pressure homogenizer, 10% oil/90% water emulsion was created with an average particle size of 220 nm.

4.3.3 *Self-Assembled Colloidal Dispersions*

An organized structure or pattern is produced when pre-existing, disordered components interact. In an aqueous medium, an amphiphilic compound generates self-assembled colloids. The transport of hydrophobic and hydrophilic bioactive can be achieved by microparticles, procolloids, lipid bilayers, liquid crystal phases, and mesophases [15].

4.3.3.1 **Micelles**

Molecules containing lipids are micelles that form spheres in aqueous solutions. Fatty acids have an amphiphilic nature, which contain both water-loving (polar head groups) and water-hating sites, causing micelle formation (the long hydrophobic chain). A micelle's polar head group creates a surface, and the nonpolar hydrophobic tail group is located inside and away from water. Single hydrocarbon chains make micelle-forming fatty acids round, reducing steric hindrance. Micelles are too bulky for the two hydrophobic chains in the fatty acids found in glycolipids and phospholipids and they prefer "lipid bilayers" [16]. Micelles arise spontaneously due to hydrophobic interactions between molecules. From simple casein micelles in milk to sophisticated digesting dietary mixed micelles, fat-soluble vitamins (A, D, E, and K) enter the bloodstream via GIT. Micelles also act as mechanisms for transporting fat-soluble vitamin from the digestive tract to the blood. Externally manufactured micelles include lipid-soluble vitamins [17]. In order to supply lipid-soluble vitamin D₂, casein micelles from milk were used. Nano delivery systems such as casein micelles are found in milk to stabilize and transport minerals like calcium and vitamin D₂ which are encapsulated in 155 nm sized particles. In addition to stabilizing the encapsulated vitamins, the assembled micelles also protect them from photochemical destruction. In a recent study, an ultra-high-pressure homogenization was employed to encapsulate vitamin D₃ into reassembled casein micelles. A double-blind placebo-controlled clinical research in 87 human volunteers reported that vitamin D₃ infused in reassembled casein micelles was highly bioavailable. The vitamin-encapsulated micelles formed large aggregates after they interacted with bile. Unlike empty micelles, the encapsulated vitamins influenced aggregation. As polymeric micelles cannot afford vitamin K absorption, the researchers concluded that free bile acid is essential for bioavailability. Micellar systems have also been used to analyze that vitamin C is water-soluble, in addition to liposoluble vitamins. In a surfactant solution, the degree of ascorbic acid oxidation depends on the concentration of surfactant. Critical micellar concentration (CMC) is the concentration at which ascorbic acid oxidation is at its peak, which is then declined by the surfactant concentration in both SDS and CTAB.

4.3.3.2 Liposomes

Liposomes in aqueous solutions are self-assembled vesicles made up of lipids and lipid-like molecules. Liposomes may contain both hydrophilic and hydrophobic actives. Their internal pH may be altered, allowing them to contain otherwise unstable chemicals. Liposomes can be made in many forms and sizes. Phospholipids can be organized to produce numerous or single vesicles. The dairy industry uses vitamins A, D, and E so that vitamins are stabilized and delivered to enhance nutritional quality. Banville et al. [18] found that liposome-encapsulated vitamin D recovered 62% more vitamin D than vitamin D produced commercially (43%) and cream contains 41% solubilized vitamin D. Vitamins were protected from degradation by liposomal encapsulation. The liposome approach protects water-soluble vitamins and has been studied showing beneficial effects on water-soluble vitamins (vitamin C). Soy phosphatidylcholine liposomes protected heat-labile vitamin C [19]. They were also evaluated with orange juice. Some researchers have found that poly phosphatidyl choline (60:40) liposomes can improve the storage stability of vitamin C. Encapsulated water-soluble and liposoluble vitamins are shown in Fig. 4.1.

4.3.3.3 Procolloidal System

Incorporating an aqueous phase dilutes a procolloidal system. Water-soluble colloids are formed when these concentrated systems are diluted with liquid crystal phases that form microphases when diluted. A fine oil-in-water emulsion is formed when oil, a surfactant, a co-surfactant, and a co-solvent are gently mixed together. Spontaneously produced aqueous microemulsions (50–1000 nm droplet size) are developed in the GI tract after ingestion of soft or firm gelatin capsules. When a surfactant interacts with a co-surfactant, spontaneous dispersion is produced on the interface, and as a result of co-surfactant assistance, the surfactant film formed has a low interfacial energy and a low bending stress. The incorporation and encapsulation of vitamin E in procolloidal systems, especially those that self-emulsify, have been extensively studied, due to its powerful nutraceutical and pharmaceutical properties [20]. Self-emulsifying and self-micro emulsifying vitamin E systems notably enhance bioavailability. Self-emulsifying systems increased tocotrienol bioavailability by two to three times in six healthy individuals who participated in the study. Food products have been extensively studied for water- and liposoluble vitamins delivered through liposomes. However, mass-production of these systems in a reproducible manner remains unsolved. The use of proliposomes is one of the methods for scaling up liposome production. Proliposomes are phospholipid particles formulated with functional ingredients, which are

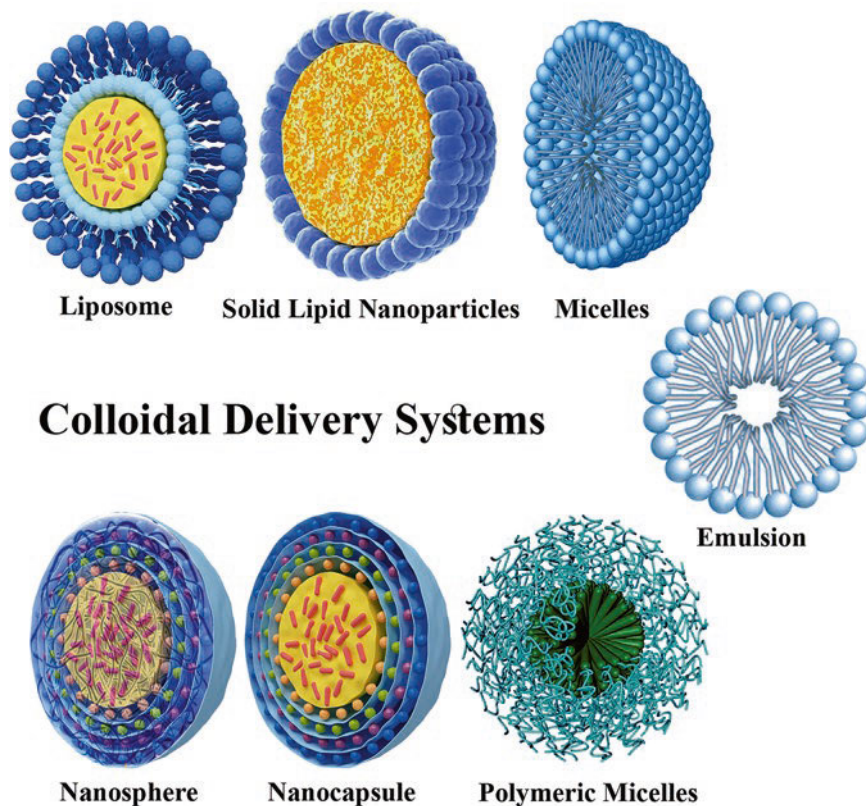


Fig. 4.1 Illustrations of colloidal delivery systems for water- and liposoluble vitamins

soluble in water and when hydrated above the transition temperature and agitated form liposomes. The addition of beta-carotene to a proliposomal formulation containing hydrogenated soy phosphatidylcholine improved chemical stability. The shelf life was reported to be 60 days when refrigerated [21]. Moreover, proliposomal formulations of glyceryl dioleate-PEG12 and glyceryl dipalmitate-PEG23 at 35% and 20% weight, respectively, were effectively combined with Vitamins D and E.

4.3.4 Dry Matrix Encapsulation

Industrial feasibility and scale-up include spray-drying, extruding, coating with pans, and using fluidized beds. These methods can easily produce microcapsules with a consistent particle size distribution.

4.3.4.1 Spray-Drying

The food industry's most popular encapsulation method is spray-drying. The process is cheap, easy, and produces consistent particles. Spray-drying is thought to be cheaper than most encapsulation processes. There are usually two steps in spray-drying: Firstly, a coating or core material solution is prepared, and then the solvent is evaporated, which is termed as dispersion. Secondly, a layer of coating material is applied over the core material or a coating of polymer matrix containing a core material is distributed. Food manufacturers have used spray-drying encapsulation since the late 1950s to protect flavor oils from deterioration and to better handle liquids. However, one drawback of spray-drying is the requirement of good water solubility of the coating material. The availability of coating materials is limited due to their water-based nature; therefore, several biopolymers have been used to coat water-soluble vitamins (vitamin C) such as gum Arabic, pea protein, and chitosan [22]. A microcapsule of less than 10 nanometers in size was produced using pea protein, chitosan, and gum Arabic, while rice starch was used to produce microcapsules of 5–40 μm . We found that microencapsulating vitamin C microcapsules containing gum Arabic and starch improved its oxidative stability, and that microcapsules of vitamin C prepared using pea protein and chitosan showed controlled release. The same group used pea protein concentrate liposoluble vitamins (α -tocopherol) for encapsulation [23].

4.3.4.2 Extrusion (in a Glassy Matrix)

Extrusion-based microencapsulation is a relatively newer method of drying in contrast to spray-drying, which is performed by forcing a series of dies, molten polymer and core material into a dehydrating liquid. Discrete microcapsules, drying the material, and pulverizing have also been accomplished with other materials such as gelatin, sodium alginate, carrageenan, gum acacia, fats and fatty acids, waxes, etc.; however, with this method, the coating material completely encapsulates the core material, providing excellent stability against oxidative and other chemical degradation. Maltodextrin and gelatin were encapsulated in a glassy matrix containing tocopherol in a 3:1:1 ratio [24]. The glassy matrix also protected the encapsulated α -tocopherol from oxidation, while encapsulating carotene in amorphous trehalose delayed degradation time.

4.3.5 *Vitamins Encapsulation in Cyclodextrins*

Cyclodextrins are cyclic oligomers formed by enzyme conversion of starch. The D-glucopyranose unit has six, seven, or eight (-CD) units joined together by α -(1,4) bonds. Their structure allows them to form molecular complexes with

many hydrophobic molecules. Encapsulation of hydrophobic drugs protects volatile aromatic compounds from degradation. The delivery of functional ingredients and bioactives, including vitamins, is the major advantage of CDs [25]. A CD's structure resembles a cage with a hydrophobic cavity. Hydrophobic compound inclusion complex is formed when less polar guest molecules replace the polar hydrophobic cavity, which is lined with water molecules. Though the exact cause of the inclusion complex formation is unknown, the following mechanism is thought to be involved: A polar interaction between a guest molecule and the hydrophobic cavity releases molecules of cavity i.e., water, and bulk water replaces the energetically disfavored polar interactions between the water molecules in the cavity and the included water. True inclusion complexes can only be formed by lipid soluble vitamins in cyclodextrins. Thus, the creation of inclusion complexes to encapsulate liposoluble vitamins has been studied extensively. The retinoid-CD complexes were made in aqueous alcohol at ambient temperature. Many studies have shown that hydroxyl propyl-CD and -CD inclusion complexes increase the solubility of vitamin A. Other studies have shown that CD complexation stabilizes vitamin A by inhibiting photoisomerization and photodegradation of retinoids [26]. Beyond retinoids-CD complexes, most studied vitamin is vitamin D as CD complex. The low water solubility of vitamin D affects its biological activity. Vitamin D and CD inclusion complexes can be made using ethanol as a solvent. A study on the stability of vitamin D after cyclodextrin complexation has also been reported. Menadione is another liposoluble vitamin (or vitamin K3). Benzene is incorporated into the cavity of a CD, while the quinine moiety is inserted outside the cavity, thus improving its aqueous solubility [27].

4.4 The Food Industry and Encapsulated Minerals

Encapsulated minerals can be integrated into the formulations of food and medicine to provide unique benefits and sensory features to the physically and chemically improved functional products. Encapsulated mineral salts have advantages such as reduced discoloration, reduced taste and smell by masking-off flavor, production and storage processes that control the release of mineral components, and improved taste and smell. Encapsulation of minerals into dairy products and table salt has been studied extensively during the last two decades. $C_4H_8FeN_2O_4$, $NH_4Fe(SO_4)_2$, and $C_6H_{10}FeO_6$ are some examples of electrolytic-Fe (Table 4.1) [28]. Soy-yogurt and soy milk are fortified with encapsulated tricalcium phosphate ($Ca_3(PO_4)_2$) and calcium citrate ($Ca_3(C_6H_5O_7)_2$) salts. KI and KIO_3 are the most common encapsulated iodine fortification salts, while $C_4H_2FeO_4$ and $FeSO_4$ are the most common encapsulated Fe fortification salts.

Table 4.1 Bioavailability, toxicity, and technological aspects of several mineral nanoparticles

Mineral	Shape/size	Technical aspects	Bioavailability/toxicology aspect	References
Se	Spherical/10–50 nm	Eliminates six food-borne pathogens from growing and forming biofilms	No toxicity to <i>Artemia</i> larvae at 100 mg/mL	[29]
Se	Spherical/80 nm	When heat is applied to Se-NPs, their thermo-stability varies with their size: Smaller Se-NPs are more resistant to transforming into nanorods than larger ones	Se-deficient mice are less likely to experience phase II enzyme induction	[30]
Se	Spherical/~36	Comparison of dietary supplements with Se and Se-NPs	At supranutritional levels, induce phase II enzymes without affecting their ability to up-regulate seleno-enzymes	[31]
Se	Spherical/20-30 nm	Introducing elemental Se-NPs to lambs through probiotic bacteria resulted in enriching lamb meat with Se (1) as an indirect method of improving the nutritional value and functionality of meat products.	The activity of glutathione peroxidase (GPx), thioredoxin reductase (TrxR), and glutathione-S-transferase (GST) is equally increased by Se-NPs	[32]
Iron oxide (Fe ₂ O ₃)	NR/<50 nm	The application of food with quercetin and Fe ₂ O ₃ nanoparticles	Cytotoxicity induced by Fe ₂ O ₃ -NPs and apoptosis and quercetin (50* μ mol/L) protects against them	[33]

(continued)

Table 4.1 (continued)

Mineral	Shape/size	Technical aspects	Bioavailability/toxicology aspect	References
Magnesium oxide(MgO)	Hexagonal/10–50 nm	Combination of Mg-NPs and nisin in milk, resulting in a leak in the bacterial cell membrane, leads to death. A protocol developed for reducing pasteurizing milk temperature and controlling pathogens required MgO-NP levels	MgO-NPs may cause DNA damage and cell death. They cause oxidative damage in consumer products	[34]
Zinc oxide(ZnO)	Spherical/10–30 nm	This engineered film of Ca alginate loaded with ZnO-NPs exhibited antimicrobial activity against two pathogens (<i>S. typhimurium</i> and <i>S. aureus</i>) 10 days at 8 °C produced zero log drop in poultry meat	In adult male Wistar rats, ZnO-NPs more than 50 mg/kg significantly altered hepatic enzymes, oxidative stress, and renal tissue	[35]
Ferric phosphate (FePo ₄)	Spherical/10.7 nm	Food and beverage formulations can be fortified with stable nanosized FePO ₄ colloid additives	FePO ₄ -NPs are soluble and bioavailable. A nanoscale coating (e.g., FePO ₄) might optimize Fe uptake and bioavailability	[36]

(continued)

Table 4.1 (continued)

Mineral	Shape/size	Technical aspects	Bioavailability/toxicology aspect	References
Calcium phosphate($\text{Ca}_3(\text{PO}_4)_2$)	Spherical/<100 nm	$\text{Ca}_3(\text{PO}_4)_2$ -NPs have great potential industry of poultry, particularly in feed management and waste minimization	$\text{Ca}_3(\text{PO}_4)_2$ -NPs are 200% bioavailable in broiler chickens	[37]
Calcium carbonate and citrate $\text{CaCO}_3/\text{Ca}_3(\text{C}_6\text{H}_5\text{O}_7)_2$	NR/ ≤ 50 nm	Due to its high MIC in broth, CaCO_3 -NP has proven to be useful antimicrobial agent, which can also be used in food and agriculture industries	Studies reveal that taking calcium carbonate or citrate NPs is more effective than administering micro-NPs to enhance serum calcium level in order to maintain bone mineral density	[38]

4.4.1 Dairy Products Fortified with Encapsulated Minerals

4.4.1.1 Milk

Despite being one of the world's most vital foods, milk is low in Fe. Thus, fortifying milk with encapsulated iron might be beneficial. Increasing Fe intake, for example, FeSO_4 microencapsulated in milk and its bioavailability was studied using a lecithin liposome method. The fortified milk's Fe bioavailability did not change after 6 months of storage and heat treatment. A similar bioavailability rate of Fe to high-bioavailable FeSO_4 absorption was also recorded. Coatings made from PGMS were used to microencapsulate Fe to fortify milk. While just 3% of Fe was released in vitro (pH 6.0), increasing pH from 5.0 to 8.0 dramatically boosted the rate of Fe release from 12.3 to 95.7%. The sensory study found no significant variations in most sensory features between the control and microencapsulated-Fe samples after 3 days of storage [39]. Moreover, the unencapsulated Fe sample had greater TBA than the encapsulated Fe sample. It was discovered that free Fe-fortified milk had a higher TBA value. Supplemented TBA of milk with high microencapsulated-Fe loadings was not altered. Microencapsulation slowed down lipid oxidation by 60%, while masking milk's metallic taste. However, comparing astringency and bitterness to control milk, microencapsulated Fe milk produced similar results. The fortification ability of Fe microcapsules in milk and the factors that affect the physical,

chemical, and sensory properties of the final product were investigated. Fortification of low-level milk with Fe microcapsules (0.1–0.3% w/v) did not increase TBA levels. It was found that 0.1% (w/v) Fe-microcapsule powder was optimal for producing fortified milk. Researchers observed that microencapsulated FeSO_4 in a powdered milk diet had better Fe bioavailability than the control diet. One study selected three Fe microcapsules fortified with EE from four distinct techniques (62.97%–74.85%) [40]. To compare the organoleptic ratings of Fe-fortified milk with the control milk, they used modified starch and sodium alginate microcapsules (10 mg/L Fe). GA, MD, and modified starch were mixed and vaporized to create microcapsules of Fe with an average size of 15.54 μm [41]. When compared to fortified milk containing Fe microcapsules, panelists preferred salt-fortified milk over microcapsule-fortified milk. In addition, the microcapsules of Fe in fortified milk had better bioavailability than those in control and Fe salt enriched milk. The sensory qualities of microcapsules of Fe from three Fe salts were also studied. All treatments had the same appearance and flavor. The flavor of the control and $\text{C}_6\text{H}_{10}\text{FeO}_6$ enriched milks did not differ significantly. This product was the finest option for fortifying pasteurized liquid milk. In comparison to cow milk (120 mg/100 g), soy milk contains a lower level of calcium (12 mg/100 g) [42]. As a result, many researchers have tried to improve the nutritional value of soy milk by adding calcium. It was discovered that adding Ca salts ($\text{Ca}_3(\text{PO}_4)_2$ and $\text{Ca}_3(\text{C}_6\text{H}_5\text{O}_7)_2$) to soy beverage was ineffective due to unfavorable Ca protein interactions, coagulation, and precipitation. Using a lecithin liposome structure could fortify soy milk in a better way. Researchers added Ca to soy milk (110 mg Ca/100 g soy milk) to match the level to that in cow's milk. The fortified soy milk was stable and had high Ca bioavailability for 1 week at 4 °C. $\text{Ca}_3(\text{PO}_4)_2$ in the soy milk was 2000 mg per liter, and to prevent Ca protein interaction and enhance the stability of soy milk, researchers added 30 g/L potassium citrate to samples. However, combining $\text{Ca}_3(\text{PO}_4)_2$ and $\text{C}_6\text{H}_5\text{K}_3\text{O}_7$ reduced soy milk stability.

4.4.1.2 Yogurt

The main consumers of yogurt are children, menstruating, pregnant or nursing women, and teenagers. FeSO_4 and a microencapsulated compound of Fe whey protein were added to yogurt for 7 days at -70 °C to assess lipid oxidation and sensory properties. The three oxidation rates (TBA and peroxide) did not change significantly. Iron-enriched yogurt tasted rusted and metallic. The sensory panel favored the Fe whey protein complex microencapsulated yogurt since the overall flavor and quality were better [43]. In recent years, researchers have enriched probiotic yogurt with *Lactobacillus acidophilus* and iron microcapsules derived from whey protein complexes. Compared to unfortified yogurt, fortified yogurt had less oxidized flavors. TBA levels were elevated due to interactions among iron and casein proteins, prooxidant activity of oxygen species, and stimulated lipid oxidation. In oxidative conditions, free fatty acids get accumulated. Three commercial Fe-salt microcapsules were added to yogurt to improve its organoleptic quality. Both the control and

FeSO₄ microcapsule samples passed the sensory test after 7 days of storage. C₄H₈FeN₂O₄ and C₆H₁₀FeO₆ yogurts were clearly distinguished from the control. TBA and peroxide levels were the highest in C₄H₈FeN₂O₄ yogurt, whereas the control and C₆H₁₀FeO₆ supplemented samples showed the lowest TBA and peroxide level. Microcapsules of FeSO₄ were found to be useful and ideal for fortifying yogurt. On 0, 1, 2, and 3 weeks, the starter bacteria count did not differ between the control and Fe-fortification samples. The quantity of these bacteria decreased with extended storage in both control and Fe-fortified yogurt samples. The added Fe had no impact on yogurt bacterial viability. Free Fe enriched yogurts had lower TBA values than encapsulated Fe enriched yogurt [44]. After 21 days, there were significant variations in astringent, oxidized, and overall acceptability assessments. To improve Fe-bioavailability, it was advised to add 80 mg Fe-WP/L to yogurt without affecting its appearance or organoleptic properties. This yogurt drink had Fe micro-encapsulated in it. Unencapsulated Fe was added, pH was decreased and titratable acidity was increased, while the microbial count of the yogurt was found to be unaffected by unencapsulated Fe. TBA kinetics in encapsulated therapies was also noted. Unencapsulated Fe changed the astringency and bitterness of yogurt drink, whereas encapsulated Fe concealed Fe taste and flavor. FeSO₄ 7H₂O, 12 mg Fe/L, and calcium (Ca₃C₆H₅O₇)₂, 600 mg Ca/L, enhanced the stability of soy yogurt. The investigation was conducted for 28 days at 10 °C. Although the fortified sample had low viscosity, extending the storage period had no significant effect on this parameter. Moreover, there were no significant variations in all sensory characteristics as well as acidity levels during the storage. Fe-fortified yogurt stored for 14 days exhibited both physicochemical and sensory qualities. The yogurts were fortified with Cold-set WPI gel powder encapsulating Fe-encapsulated in FeSO₄ solution (20–60 mg Fe/kg). Compared to the unfortified control sample, WPI-Fe particles fortified yogurt (up to 60 mg of Fe/kg) exhibited identical qualitative characteristics (color and flavor). A physicochemical and sensory evaluation demonstrated that samples fortified with WPI-Fe may retain quality criteria for 2 weeks regardless of the Fe concentration employed [45]. A Fe-enriched usual yogurt (B225 g) was likewise reported to give up to 60% of a woman's daily Fe requirement, with no evident color or flavor differences from the control yogurt. This feat would be unachievable with even minimal levels of Fe fortification in yogurt. To maximize the effects of three independent factors on EE and Hunter Lab color features, an application of a second-order polynomial model using the RSM-central composite design was used. The optimal pH, WPI content, and Fe concentration for pre-paring WPI-Fe particles were 7.0, 6.8%, and 18.8 mM, respectively. This is because the particles were released into the intestinal environment at a high rate (95%), WPI-Fe particles were generated under optimal conditions, and a site-specific network was an ideal delivery option of Fe (Pancreatin was added, pH 7.5). However, it was found that iron encapsulated in only 28% of powders could be released in the stomach environment (pepsin, pH 1.2).

4.4.1.3 Cheese

Cheese, like milk, is low in Fe. Most research on Fe encapsulation in cheese has been focused on Cheddar. According to IDFA (2015), 28.5% of all cheese consumed in the US was cheddar. Researchers reported that FeCl_3 , Fe casein, FIP WP, and Fe WP complex constitute the Fe content of cheddar cheese. The study used FeSO_4 as a Fe source to fortify cheese, which showed a somewhat higher TBA level than unfortified cheese. TBA value and oxidized off-taste had no correlation with Fe level. Another link was between off-taste or flavor of the oxidized cheese and TBA value. Aged cheese had no effect on TBA levels, oxidized off-taste, or cheese flavor scores. The greatest bioavailability of Fe was 85% for FeCl_3 , 71% for Fe casein, 73% for FIP-WP, and 72% for Fe WP. Feeding equivalent fortified Fe for 10 and 14 days showed similar basal Fe level, while the bioavailability sources were 5%, 4%, 6%, and 3% ($P \leq 0.05$) [46]. Nonetheless, FeSO_4 had 85% maximum and 5% basal bioavailability, and it accelerated lipid oxidation. In addition to WP FeCl_3 , researchers employed the same fortification method as before. FeSO_4 produced oxidation and off flavors in aged cheddar. Sensory perception was better in samples having Fe WP complexes. The chemical and sensory qualities of cheddar cheese were examined with encapsulated-Fe. Encapsulated cheese ripened slower than unencapsulated cheese. During ripening, compounds with short-chain FFAs and neutral volatiles were generated insignificantly. The samples with maximum encapsulated Fe had the most bitterness, astringency, and sourness. However, it did not influence the quality of cheddar cheese, whether LMFS (large microencapsulated FeSO_4 , 700, 1000 m) or SMFS (small microencapsulated FeSO_4 , 220, 422 m) enriched cheddar cheese [47]. After 90 days, LMFS (66%) and SMFS (91%) recovered Fe. No significant differences in lipid oxidation rate or chemical makeup (for example, dry matter, fat, ash, magnesium, zinc, and calcium) were observed. Microencapsulating FeSO_4 in cheddar cheese did not overpower the flavor, odor, or color of Fe. Less SMFS-fortified cheddar cheese was found to have better Fe retention and sensory attributes. Mozzarella cheese was fortified with ferric chloride, WP, and casein sodium iron salts (25–50 mg Fe/kg). Mozzarella cheese with 50 mg Fe/kg exhibited an off-odor, metallic or oxidized flavor, and more unattractive qualities. Consumers despised all mozzarella due to metallic and off flavors. Cheddar cheese was fortified with FeCl_3 and Fe protein components, indicating that it is not possible to generally apply Fe fortification to every other cheese. To plain feta cheese, 80 mg/kg of compounds containing iron (FeSO_4 , FeCl_3 , and microencapsulated FeSO_4) was added. The chemistry and FFAs of the control and Fe or vitamin C samples were not different. Fe-supplemented cheese compared to unsupplemented cheese revealed a significant difference in Fe TBA and organoleptic ratings, which were lowest in feta cheese containing microencapsulated iron (80 mg) and vitamin C (150 mg) [48].

4.4.2 Fortification of Salt by Encapsulating Iron and Iodine

4.4.2.1 Dual Fortified Salt

Since Fe compounds have a greater stability than iodine compounds, encapsulating iodine to provide a protective partition was explored. $C_4H_2FeO_4$ or $FeSO_4$ (1 g/kg Fe) was blended with FBC to make DFS from iodine (KI and KIO_3) microcapsules. Later, the stability of several fortifiers was studied in relation to temperature and relative humidity during storage. A good result was obtained for encapsulating iodine into dextrin, indicating a wall-to-core ratio of 1:200. After 30 days of storage at 40 °C and 60% relative humidity, DFS was enhanced with $FeSO_4$ and KI lost 100% of its iodine concentration, whereas KIO_3 lost 93% at 40 °C and 60% relative humidity; DFS retained 79.7% iodine and KIO_3 retained 80.9% iodine. $C_4H_2FeO_4$ and KI microcapsules demonstrated the greatest iodine retention (98.4%, 101.9%) at temperature of 40 °C and a relative humidity of 60% for 12 months. Despite improvements in iodine chemical stability, fine microcapsules generated by the technique prevented encapsulation [49]. Also, iodizing salt with KIO_3 is expensive. Condensing Fe-compounds in salt formations, iodine and Fe DFS were created and tested for storage stability in highland (Nairobi) and coastal (Mombasa) cities in Kenya. Nairobi retained 87.3% Fe and only 92% iodine in encapsulated DFS with a KI and $C_4H_2FeO_4$ premix, whereas Mombasa retained 92% Fe and 90% iodine. DFS encapsulated in $C_4H_2FeO_4$ resisted color fluctuation and increased bioavailability. It was found that encapsulating Fe in DFS reduced unpleasant sensory characteristics and iodine loss, while maintaining considerable bioavailability. Researchers studied 19 Fe-containing compounds (e.g., fumarate, sulfate, pyrophosphate, and elemental Fe) bound to indigenous Moroccan and Cote d'Ivoire salts. After 6 months of storage, samples were examined for iodine concentration and color. According to the findings, encapsulation did not prevent most chemicals from ionic losses and undesired organoleptic alterations. Iron oxide (B2.5 and 0.5 m) particles were used to enhance salt. In the presence of Fe ($C_4H_2FeO_4$) and iodine (KI and KIO_3), the stability of table DFS was measured in the mountains and coasts of Kenya. Also, high iodine (90%) was preserved after 3 months of storage. Encapsulated $C_4H_2FeO_4$ was combined with salt-encapsulated-KI or iodate salt to make DFS. Because of the significant retention of Fe21, a little quantity of Fe21 was oxidized to Fe31 (17%). During distribution and retail, both iodine and Fe21 were preserved, making DFS with iodine and encapsulated $C_4H_2FeO_4$ extremely stable for 3 months. In vitro and in vivo studies have been focused on Fe bioavailability in iodine DFS and encapsulating $C_4H_2FeO_4$; initially, researchers made a $C_4H_2FeO_4$ premix for salt integration by agglomerating $C_4H_2FeO_4$ into salt-size particles and then encapsulation was performed [50]. The in vitro bioavailability of various DFS and encapsulated $C_4H_2FeO_4$ premixes generated by numerous techniques was then examined. The materials and techniques used in encapsulating $C_4H_2FeO_4$ marginally affected the in vitro Fe bioavailability. The researchers discovered that adding TiO_2 to the coating materials as

a masking agent effectively concealed the reddish-brown color of $C_4H_2FeO_4$ and improved overall sensory acceptance and iodine stability. Bioavailability of $C_4H_2FeO_4$ encapsulated in a lipid structure was determined indicating that $FeSO_4$ was 95% bioavailable compared to $FeSO_4$ in rats. Extrusion-based encapsulation of Fe ($C_4H_2FeO_4$) enhanced the iodine stability in iodized salt. The optimal formulation contained $C_4H_2FeO_4$ (10%), binders (30%), TiO_2 (25%), and Methocel, a water-soluble polymer (10%). Uncoated Fe particles, on the other hand, lost 50% of their DFS iodine content during extrusion. The usual salt distribution mechanism will be used to supply iodine and iron to DFS. Li et al. [51] investigated whether it was possible to encapsulate $C_4H_2FeO_4$ in DFS using the cold-forming extrusion process. Extrudable dough containing substantial levels of $C_4H_2FeO_4$ was thus possible using grain flours [51]. Additionally, all extruded Fe particles (300,700 nm) were analyzed indicating high in vitro Fe digestibility. Yadava et al. [52] enhanced masking colors and coating polymers phases utilizing extrusion agglomeration to prepare a fortification of salt with Fe premix. After mixing iodine and Fe21, stability was tested at 35 °C and 60% relative humidity for 90 days after Fe premix with iodized salt. Water-soluble polymeric coatings showed a good efficacy to retain micronutrients at 10% encapsulation capacity. They also had great bioavailability and pleasing sensory characteristics. To add encapsulated Fe ($C_4H_2FeO_4$) to coarse iodized salt, Romita et al. [53] used spray drying. A 10% w/v suspension was employed, and 1.2% of the suspension was dissolved using biopolymers for 6 months. DFS was monitored in iodine stability at room temperature (20% RH) and under harsh conditions (40 °C and 60% RH). Sixty percentage of unencapsulated samples exhibiting reduced iodine stability was not surprising.

4.4.2.2 Triple Fortified Salt

A formulation containing iron, iodine, and vitamin A, three essential micronutrients in a single food product is a low-cost method of fortifying foods. Strong metabolic interactions among these components can treble salt fortification. A mixture of micronized $Fe_4O_{21}P_6$, KIO_3 , and retinyl palmitate, was incorporated into HPF to create TFS with 30 g of I, 2 mg of iron, and 60 g of vitamin A in each gram of salt. After 6 months of storage, the color stable TFS lost iodine and vitamin A (B12 15%) [54]. At 10 months, the TFS group had much less vitamin A and Fe anemia deficit than the iodized salt group. In addition to $Fe_4O_{21}P_6$, KI, and retinyl palmitate (vitamin A), HPF nano capsules were also used to formulate TFS. TFS's color changes during storage were acceptable, as was the loss of iodine, which was similar to that in iodized salt. Retinyl palmitate was found to be highly stable, with only 12% loss after 6 months. The overall sensory tolerability of TFS and iodized salt was not found to be changed.

4.5 Iron Encapsulation in Cereals and Bakery Products

FeSO₄ was incorporated into mixers that mix baking flour, and a fine-sized white powder with freely flowing characteristics was obtained which could be stored without quality loss for 6 months [55]. Noodles encapsulated with Fe (B5 mg Fe per serving) got comparable sensory scores to those containing inorganic fortification agents such as ferric sodium ethylenediamine tetraacetate (NaFeEDTA) and other ingredients. A three-month storage at ambient temperature did not influence physical (color) or oxidative (peroxide value) parameters. Biofortified wheat flours were made by Biebinger et al. with HPF wall material [56]. The prepared microcapsules may be able to overcome undesirable sensory changes and Fe shortage in biscuits made with wheat flour. Average iodine loss during spray-cooling was B25%, but no measurable iodine losses occurred during baking. Fortified wheat-based biscuits were compared with non-fortified controls in a feeding trial randomized with Kuwaiti young women. Biscuits that were fortified with Fe microcapsules and those that were unfortified were the same in taste and color. However, the fortified ones lowered the prevalence of Fe-deficiency by 50%. The fortified samples group had higher serum ferritin and urine iodine levels. The encapsulated FeSO₄ Fe absorption was measured. Iron deficiency in Thai women was overcome by consuming foods based on wheat reinforced with unencapsulated FeSO₄, electrolytic Fe, or elemental Fe reduced by hydrogen. After 20 weeks, about 11% of the prescribed Fe dosage was absorbed. Encapsulation could not impact Fe bioavailability. Low-extraction wheat flours can be supplemented with FeSO₄ or C₄H₂FeO₄ encapsulated (less than 0.8% ash) at the same amounts as without encapsulation [57]. In terms of quality parameters, Fe (Fe₄O₂₁P₆, FeSO₄, NaFeEDTA, and decreased Fe) and gluten have significant effects. Both elements also influenced the sensory attributes such as odor and the color of wheat bread. The color, stiffness of the crust, cell count, odor, and metallic taste of the crust were found to significantly differ in unfortified gluten-free loaves than Fe-fortified loaves. The encapsulation also failed to protect Fe from oxidation, since the color, taste, and texture of breads fortified with FeSO₄ varied from those of unfortified breads. Children in Sao Paulo childcare centers previously rated bread supplements encapsulated with FeSO₄ as acceptable as reported by Souto et al. [58]. Breads that were fortified with Fe were less acceptable to children than those that were not. In young children, fortified bread may offer some protection against Fe deficiency anemia. Using a paste made from rice flour, iron compounds (0.5 and 1.0 g Fe/100 g), and 25% water, Moretti et al. [59] produced a replica of rice grains. In order to achieve a final Fe content of 5 mg per 100 g of rice, Fe components (1:100 or 1:200) were combined with rice grains. For this purpose, dispersible micronized Fe₄O₂₁P₆ (B0.5 m) was cold extruded, and reagent-grade Fe₄O₂₁P₆, Fe₄O₂₁P₆ (B2.5 and B20 m), FeSO₄ encapsulated in a liposome and HPF, as well as rice grains containing FeSO₄ (positive control) were utilized [60]. The rice seeds were simulated and dried overnight to a 10% water content. Uncooked

and cooked rice grains had very different colors due to all Fe components except for micronized $\text{Fe}_4\text{O}_{21}\text{P}_6$ (0.5–20 μm). The enriched rice seeds with $\text{Fe}_4\text{O}_{21}\text{P}_6$ and elemental Fe had the lowest Fe loss (2%) during the 30 min pre-washing at 30 °C. Researchers in India hope to develop Rice seeds fortified with iron and outstanding sensory properties and enhanced bioavailability rates using micronized $\text{Fe}_4\text{O}_{21}\text{P}_6$. Li and colleagues created Ultra Rice with $\text{C}_4\text{H}_2\text{FeO}_4$, mixtures of micronized $\text{Fe}_{41}\text{O}_{21}\text{P}_6$, FeNaEDTA, thiamine, and other antioxidants. Fe was not bioavailable or stable when added to Ultra Rice. Rice seeds that were encapsulated with $\text{Fe}_4\text{O}_{21}\text{P}_6$ exhibited an acceptable creamy-yellow color, but 50% less thiamine [61]. Unencapsulated $\text{C}_4\text{H}_2\text{FeO}_4$ exhibited a reddish-brown tint. By combining FeNaEDTA with encapsulated $\text{C}_4\text{H}_2\text{FeO}_4$, *in vitro* bioavailability was improved. Using colloids or particles of $\text{Fe}_4\text{O}_{21}\text{P}_6$ could reduce thiamine losses while maintaining physical and organoleptic properties for 32 weeks at 60% RH and 40 °C. Dietary thiamine loss was minimal; however, oxidative rancidity was observed. BHA, BHT, hydrophilic citric acid, and sodium hexa meta phosphate were found to be the most effective storage forms of uncoated and encapsulated Fe containing free-radical scavenging constituents. Women who consumed cooked Ultra rice containing micronized $\text{Fe}_4\text{O}_{21}\text{P}_6$ daily reduced their anemia by 80% and Fe deficiency by 29%. Beinert, Velasquez-Melendez, Pessoa, and Greiner also reported that the consumption of Ultra Rice encapsulated with micronized $\text{Fe}_4\text{O}_{21}\text{P}_6$ (3.14 μm) reduced Fe deficiency and anemia in Brazilian youngsters (6–24 months) [62].

4.6 Minerals Encapsulation in Other Foods Fortification

The effects of injecting L-Ca with EPC into rabbits before slaughter were studied to determine its impact on meat aging. An infusion of L-Ca into rabbit meat could lower meat's ageing without contaminating it or causing any physical trauma. Water in oil emulsion was prepared using an emulsifier such as Tween 60 to make a water-in-corn-oil emulsion, indicating that the TBARS generation was low, while the Fe-EE (99.75%) was high [63]. To assess the effect of Fe on fish oil stability, fish oil-based emulsion was created and combined with droplets of the initial emulsion. TBA values increased when Fe interacted with fish oil and activated oxidation processes. A spray drying process was conducted to encapsulate natural green pigments from Pandan leaves with three coatings (GA, MS, and MD). For powders encapsulated using Ms. wall material, SEM micro-graphs showed spherical and smooth particles, while powders were generated by the encapsulants. MD and GA surface shrinking Greenness, total chlorophyll, and antioxidants were determined to be best in Zn chlorophyll microcapsules, which comprised of 30% Ms. The produced powder has a predicted half-life (462 days) longer than GA (330 days) and MD (330 days) powders. Fe microparticles were studied in black beans in 2011, which contains 5–10 mg encapsulated Fe per spoon [64]. Using samples

supplemented with FeSO_4 micro-encapsules could help prevent or control anemia. Peach and apple juice were used to determine Fe status. $\text{Fe}_4\text{O}_2\text{P}_6$ was the core and lecithin was the wall of spray-dried Fe microcapsules.

4.7 Application of Nanoparticles in Minerals

In recent years, scientists have become interested in reducing mineral inclusion levels and enhancing mineral absorption through nanoparticles. Mineral NPs are generally spherical and under 100 nm in size. They are frequently employed in agriculture, food, and pharmaceutical industries. Natural mineral nanoparticles (NMNs) have numerous industrial and medical applications. Se-NPs are the most commonly used NPs in the food industry to inhibit bacterial growth and biofilm formation. Melatonin, PEG, adenosine triphosphate, polysaccharides, and epigallocatechin-3-gallate are employed to make Se-NPs with minimal toxicity [65]. A nutritional supplement containing Se-NPs may help improve the body's function and bio accessibility, as Se offers several benefits for the body's health, including enhancing the immune system and reducing cancer cell proliferation. Meal supplementation with Se-NPs lowered bioavailability in Se-deficient mice. We are also aware of the antibacterial properties of inorganic nanometal oxides such as magnesia, zirconium, and calcium oxide (CaO) NPs. In order to kill bacteria, metal oxide nanoparticles produce reactive oxygen species. Surface superoxide and H_2O_2 can cause intracellular leakage and cell death. In vitro studies of MgO -NPs conducted on human liver (HepG2), kidney (NRK-52E), gut (Caco-2), and lung (A549) cell lines indicated oxidative stress, DNA damage, and cell death, according to the USDA (21CFR184.1431) [66]. Thus, their use in consumer goods should be monitored for safety reasons. Several studies have shown the efficacy of MgO -NPs in treating various ailments. In U87 human astrocytoma, MgO -NPs were less cytotoxic than ZnO and TiO_2 NPs. Boubeta et al. [67] employed MRI agents made of Fe/ MgO nanoshells. Iron/ MgO -NPs mediated magnetic hyperthermia in cancer therapy. Nano cryosurgery for cancers and Iron oxide (Fe_2O_3) nanoparticles are biocompatible and less toxic than other metals [67]. A scientist reported the use of Fe_2O_3 -NPs as mineral supplements, controlled release medicinal and nutraceutical biomaterials, and colorants in cosmetics. The USDA (21CFR182.8991) also considers ZnO , one of the five Zn compounds, to be safe. By adding antimicrobial ZnO -NPs to active packaging films, especially for meat formulas, we investigated ways to prevent food-borne infections [68]. It was discovered that these NPs are not only antimicrobial but also a Zn source for numerous meals. Although they possess antibacterial properties, $\text{Ca}_3(\text{PO}_4)_2^-$, CaCO_3^- , and $\text{Ca}_3(\text{C}_6\text{H}_5\text{O}_7)_2^-$ -NPs have also shown promise in agriculture, poultry, and food processing. In addition to improving serum Ca levels and bone mineral density, Ca-based nanoparticles are highly bioavailable in the body.

4.8 Conclusion

In summary, the information provided herein can help in the encapsulation of difficult-to-formulate liposoluble vitamins, reactive water-soluble vitamins, and chemically sensitive water-soluble and liposoluble vitamins. Inventors select encapsulation techniques and micro/nanostructures based on resource availability, material qualities, process economics, and end-use applications. These techniques should also be safe to use, have a long shelf life, and require no dangerous chemicals or toxic solvents in their production. Recent research has focused on how to provide active substances like micronutrients in foods. In this perspective, most food fortification and nutraceutical R&D should concentrate on nano- and microencapsulation. To avoid bottlenecks, the hunt for food-grade protective coatings, shorter development times, better understanding of encapsulating techniques, and better understanding of in vivo behavior of encapsulates should be continued.

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