

Chapter 4

Chemical Diversity and Ethnobotanical Survey of Indian Medicinal and Aromatic Plants Species



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Abstract Plants are the source of important drugs. Their secondary metabolite content is subject to environmental influences. Scientific evaluation of the chemical diversity of plants may be useful in exploring their medicinal as well as other uses. Still there are numerous medicinal plants for which no results either of ethnopharmacological uses or phytochemical studies could be found in the literature. Ethnobotany deals with the relationship between humans and plants. It has played an important role in the development of new drugs for centuries. Ethnobotany is attracting professionals from diverse academic backgrounds and interest. Ethnobotany may play an important role in securing sustainable supplies, and it can also be of use in the search for new medicinal recipe which could be used to treat diseases for which no standard therapy has been reported. Harnessing chemical diversity based on phytochemical research of species containing potentially active principles would be more relevant in context of ethnobotanical research.

Keywords Biodiversity · Secondary metabolites · Ethnomedicine · Traditional medicine

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4.1 Introduction

Secondary metabolites from plants are rich sources of bioactive compounds bringing forth many health beneficial effects in man and animals. Worldwide, medicinal plants have played a key role in providing health care. Since antiquity, medicinal plants have been employed in the treatment of a wide range of diseases and health conditions. Plants still continue to make important contributions both in providing lead molecules for further pharmaceutical research and as alternative sources of efficacious medication even in spite of the spectacular advances in the discovery of novel drugs that have occurred over the last few decades (Cragg et al. 1997; Shu 1998). Since time immemorial, plant products have been part of phytomedicines. Phytomedicines are composed of plant organs supplied either in natura (generally leaves, root or bark) or in processed form (typically liquid or powdered extracts). Commercial extracts normally contain the active principles of the plant material in crude or processed state, together with excipients, i.e. solvents, diluents or preservatives. The source and quality of the raw materials play a pivotal role in guaranteeing the quality and stability of the herbal preparations (Calixto 2000).

Scarcity coupled with strong demand on drugs has led to the cultivation of medicinal plants (Chopra et al. 1958). In an attempt to discover new drugs, multinational pharmaceutical companies typically spend an annual amount of US\$ 110 billion (Nair et al. 2014). A vast majority of medicinal plants have been recklessly exploited. Therefore it is an imperative to rationalize the use of important medicinal plants (Sultan et al. 2008).

Bioactive phytochemicals are naturally occurring compounds present in or derived from a plant (Hardy 2000). Bioactive compounds of plant origin are those secondary metabolites that possess desired health/ wellness benefits for man and/or animals. These metabolites are both chemically and taxonomically extremely diverse compounds with frequently obscure functions (Yadav and Agarwala 2011).

Phytochemicals may function as antioxidant (protect cells against oxidative damage), antiproliferative (interfere with replication of undesirable cancerous cell), carcinogen detoxifier, hypocholesterolemic, stimulant of enzymes and hormones, antibacterial and antiviral, anti-inflammatory, ligand to cell wall (some phytochemicals bind physically to human cell thereby preventing the adhesion of pathogens) and potential inhibitor of different actions affecting the initiation and progression of several pathogenic processes (Kaur and Das 2011).

Secondary metabolites are frequently called the vast “Chemical library” of biological systems. Most of the drugs, herbs, ethnomedicines, essential oils, perfumes and cosmetics derive from them. Cultivation is an important practice to conserve endangered medicinal plants growing in the wild and it works as a practical method to make available natural raw materials without affecting their actual habitat (IUCN 1993). A prerequisite for breeding is the study of genetic diversity of available plant germplasm (Bernáth 2002). Germplasm characterization is necessary to enhance

germplasm management and utilization. Genetic diversity is influenced by habitat types and the altitudinal range (Jugran et al. 2013, 2015). Diversity classification in germplasm collections is important for both plant breeding and germplasm collection. High diversity is an indicator of better adaptability of a population as a result of more fitness under rapidly changing environment. Wild plant species which can adapt easily in any conditions are always suitable for domestication or cultivation (Dhiman et al. 2020). The most important goals of any medicinal plant breeding program are to improve the morphological characteristics and increase the accumulation of biologically active substances. The quantitative and qualitative status of active constituents along with genetic diversity for a medicinal plant is the basis to devise conservation strategies and select right samples for maximum yields. Conservation strategies for populations should take into account both genetic diversity and chemical variation levels, especially in the case of populations having high differentiation to bioclimatic factors and the geographic location of populations (Nair et al. 2014).

Chemical constituents of MAP are the basis of their exploitation (Heywood 2002). Chemical markers are the group of chemical constituents derived from herbal/medicinal products. Chemical markers play an important role also in the quality control of herbal products and medicines. Chemical diversity plays an important role also in plant adaptation (Dhiman et al. 2020). Species, strains and geographical origin can be distinguished using chemical fingerprinting. It is imperative to identify elite plants/population based on their chemical attributes to ensure the quality of plant material (Jugran et al. 2015).

Association between the molecular markers and the phytochemical markers has been found to provide the best method of assessment of plant genetic diversity. This approach is used to screen and improve the gene pool of elite genotypes (Ray et al. 2019; Qaderi et al. 2019; Nair et al. 2014; Hennenke et al. 2016).

4.2 Chemical Diversity in Selected Threatened Indian Medicinal Plants

A species that has been described as species with small population is not presently endangered but is at risk. A species which is in danger of extinction throughout all or a significant portion of its range has been categorized as an endangered species. Threatened is a species that is likely to become endangered in the foreseeable future (IUCN 1978; Bryde 1979; Nayar and Sastry 1990). The chemical diversity in nine high value endangered medicinal plants of India is briefly described in terms of their major phytochemical principles. These species are not found in cultivation. They are collected from nature; therefore, their ecological study needs to be brought to the forefront, in addition to their *in-situ* conservation.

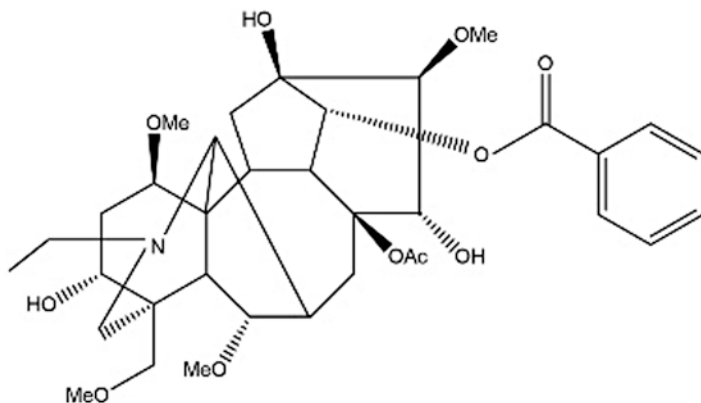


Fig. 4.1 Chemical structure of aconitine

4.2.1 *Aconitum heterophyllum*

Aconitum heterophyllum (Family: Ranunculaceae) is widely distributed across North Asia and North America. Worldwide about 300 species of *Aconitum* are found and 27 species, in India (Paramanick et al. 2017; Sharma et al. 1993). *A. heterophyllum* possesses potential immunomodulatory activity (Murayama and Hikino 1984; Weiner 1990). Content of aconitine (Fig. 4.1), an alkaloid, varies from species to species and also with place of origin (Prasad 2000; Hikino et al. 1983; Iwasa and Naruto 1996, Song et al. 1984). Variation of aconitine content in *A. chasmanthum* and *A. heterophyllum* from Kashmir Himalayas was reported by Jabeen et al. (2011). Aconitine content varied from 0.0310% to 0.0320%, 0.0014% to 0.0018% in *A. chasmanthum* and *A. heterophyllum*, respectively.

4.2.2 *Ephedera foliata*

Ephedera Linnaeus is a genus of about 40 species. Eight *Ephedera* species from India and adjoining regions were listed by Sahni (1990). Three additional species namely *E. kardangensis*, *E. khurickensis* and *E. sumlingensis* were also reported recently (Sharma and Singh 2015). *E. foliata* is a typical component of arid and semi arid regions of North-Western parts of India (Bhandari 1978). It is harvested on commercial basis in Gujarat (Gavali and Sharma 2004). However, over exploitation, extensive habitat destruction, very slow growth rate, poor regeneration, grazing and other anthropogenic pressure have caused tremendous reduction in its natural populations. It has now become a rare or endangered species from a vulnerable category (Kharin 2002; Joshi et al. 2013) and it is considered as a threatened

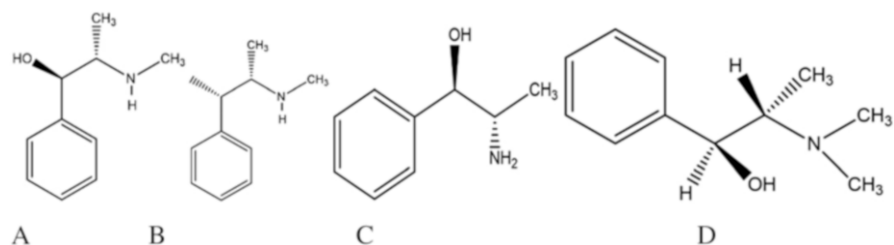


Fig. 4.2 Chemical structure of (a) (–)-ephedrine (b) (+)-pseudoephedrine (c) (–)-norephedrine (d) (+)-methylephedrine

species in India (Meena et al. 2019). Whole plant of *E. foliata* is used in fever, blood purification, asthma, dropsy, snake bite and as cardio tonic (Silori et al. 2005; Quattrocchi 2012). The major active principle in *Ephedra* is (–)-ephedrine and (+)-pseudoephedrine. Other minor alkaloids include (–)-norephedrine, (+)-methylephedrine (Fig. 4.2). Depending upon the species, the total alkaloid content in *Ephedra* can exceed 2% (Bruneton 1995; Chaudhary et al. 2020). The alkaloid content in Indian *Ephedra* ranged from 0.28–2.79% (Chauhan 1999; Polunin et al. 1987). Chaudhary et al. (2020) reported variation of metabolite content in *Ephedra* within same phytogeographical region of Kashmir Himalayas. The climatic conditions, physical and chemical property of soil (pH, soil moisture, macro-micro-nutrients, etc.) and other edaphic factors were attributed for the variation of metabolites content in *Ephedra* from same phytogeographical region.

4.2.3 *Malaxis acuminata*

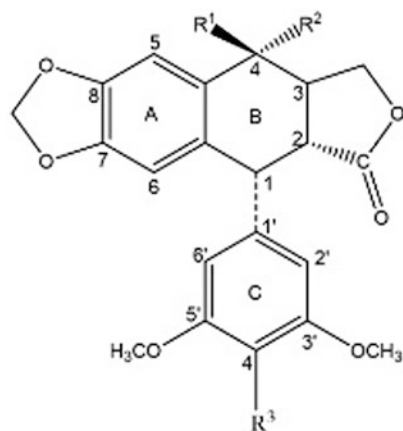
Malaxis acuminata is commonly known as “Jeevaka”. It is a small, perennial, pseudobulbous terrestrial orchid with pale yellowish-green to pinkish flowers in terminal racemes (Bose et al. 2017). The dried pseudobulbs are important ingredient of very reputed Ayurvedic drug “Ashtavarga” and also used in the preparation of polyherbal tonic “Chyavanprash” (Cheruvathur et al. 2010; Govindarajan et al. 2007). Due to the rapid loss of forest cover, jhum cultivation, etc., *M. acuminata* has become threatened in the nature. It has been listed in CITES (Conservation of International Trade of Endangered Species of Wild Fauna and Flora) Appendix II (Jalal 2012; Lohani et al. 2013). Secondary metabolite profiling in various parts of *M. acuminata* was reported by Bose et al. (2017). Presence of fatty acids, α -hydroxy acids, phenolic acids, sterols, amino acids, sugars and glycosides were reported using GC-MS analysis of methanolic extracts of leaves and stems of wild as well as *in vitro* plantlets of *M. acuminata*.

4.2.4 *Pterocarpus marsupium*

Pterocarpus marsupium is distributed in Central, Western and Southern regions of India (Devgun et al. 2009). It is listed as vulnerable plant in the INCN red data list (IUCN 2017). The Ayurvedic System of Medicine strongly recommends water stored in a tumbler made from hardwood of *P. marsupium* for effective diabetes control (Chopra et al. 1958; Jain 1968; Satyavathi et al. 1987). Mohankumar et al. (2012) reported that a high molecular constituent obtained from the fractionation of the aqueous extract of *P. marsupium* hardwood had potent insulinotropic and insulin like properties. The heartwood of *P. marsupium* is important source of pterostilbene. Other secondary metabolites such as epicatechin, pterocarpol, pterosupin, pterocarposide and marsuposides have also been reported from *P. marsupium* (Teixeira da Sliva et al. 2018).

4.2.5 *Podophyllum hexandrum*

The rhizomes of *Podophyllum hexandrum* are well known in medicine, as a source of podophyllin resin. Podophyllotoxin (Fig. 4.3) is the major lignan present in the resin and it is the starting material of etoposide. Vepesid, commercial name of etoposide, is an FDA approved anticancer drug used to treat testicular as well as lung



Compound	R ¹	R ²	R ³
1 DPT	H	H	OCH ₃
2 PPT	H	OH	OCH ₃
3 4'-DMEP	OH	H	OH

Fig. 4.3 Chemical structure of podophyllotoxin

cancer by inhibiting replication of cancer cells (Becker 2000; Henderson 2000; Jackson and Dewick 1984). Podophyllotoxin preparations are also commercially available to treat genital warts (Beutner 1996). The Indian *P. hexandrum* is superior to its American species *P. peltatum* in terms of its higher podophyllotoxin content (higher than 5%) in dried roots in comparison to only 0.25% of *P. peltatum* (Panda et al. 1992; Mishra et al. 2005). Sultan et al. (2008) reported high diversity in the concentration of marker compounds (podophyllotoxin β -D-glycoside and podophyllotoxin) in 36 individuals from 12 accession of *P. hexandrum*.

4.2.6 *Rauvolfia serpentina*

The genus *Rauvolfia* comprises of 80 species and is represented by five species namely *R. hookeri*, *R. micrantha*, *R. serpentina*, *R. verticillata* and *R. tetraphylla* (Bindu et al. 2014). *R. serpentina* has been designated as critically endangered in India and is included in Appendix II of CITES, which restricts its export (Singh et al. 2010). Population of *R. serpentina* declined more than 50% during the period of 1985–1995 due to loss of habitat and overexploitation (Bindu et al. 2014). Government of India has banned export of this species from the wild in order to prevent over exploitation of this species (Sukumaran and Raj 2008). Roots are being indiscriminately collected from the wild to meet the growing demands of the pharmaceutical industry and this has rendered listing the species “endangered”.

Indole alkaloids such as reserpine, reserpiline, rescinnamine, ajmaline, ajmalicine, rauwolfanine, serpentine, serpentinine and yohimbine, etc. have been reported from *Rauvolfia* species (Sahu 1983; Gao et al. 2012). Reserpine is the most prominent among these alkaloids (Nair et al. 2014). Reserpine (Fig. 4.4) has been known, documented and used to treat snakebites and insanity, however, the main use of the drug is as a sedative and hypnotic and for reducing blood pressure. The drug is now largely used in insanity and high blood pressure. It is more suitable for cases of mild anxiety or patients of chronic mental illness (Bleuler and Stoll 1955).

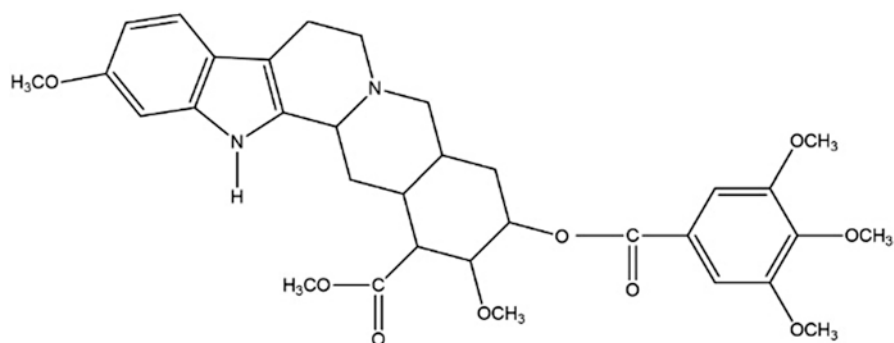


Fig. 4.4 Chemical structure of reserpine

Bindu et al. (2014) reported that reserpine content was highest for *R. tetraphylla* (450.7 µg/g dry wt.) among five *Rauvolfia* species from India. Reserpine content was comparatively low in the *R. serpentina* (254.8 µg/g dry wt.). Reserpine content in 10 population of *R. serpentina* ranged from 0.192 g/100 g to 1.312 g/100 g. *R. micrantha*, which is an endemic *Rauvolfia* species in India, had significantly higher reserpine (422.1 µg/g dry wt.) content (Bindu et al. 2014). Chemical synthesis of reserpine is economically still not feasible (Farooqi and Sreeramu 2001). Significantly higher reserpine content in *R. micrantha* makes this species a suitable candidate as a source of reserpine, replacing *R. serpentina* and *R. tetraphylla*.

4.2.7 *Rheum emodii*

The genus *Rheum* belonging to family *Polygonaceae* encompasses about 60 species. It is mainly distributed in the temperate and subtropical Asia (Anjen et al. 2003). Seven species from this genus have been reported from India (Hooker 1885). *Rheum emodii* is found at an elevation of 2000–3800 m in the temperate Himalayas from Kashmir to Sikkim (Zargar et al. 2011). Phytochemicals from *Rheum emodi* have been reported to possess antioxidant, antidiabetic, antimicrobial, antifungal, cytotoxic, hepatoprotective and nephroprotective activities. *R. emodii* has been used an ingredient in many herbal formulations used for treatment of various diseases, in particular for the regulation of blood fat, hepatitis and cancer (Zargar et al. 2011). Rhizome is the source of major phytochemicals from *R. emodii*. Free anthraquinones and their glycosides are the major phytochemicals from *R. emodii* (Fig. 4.5). Anthraquinone with carboxyl group (Rhein) and without carboxyl group including chrysophanol, aloe-emodin, emodin, physcion, chrysophanein and emodin glycoside and alkyl derivatives of anthraquinone namely 6- methyl rhein and 6-methyl aloe-emodin have also been reported from *R. emodii* (Malik et al. 2010; Singh et al. 2005). Anthrone C-glycoside derivatives, such as oxanthrone, ether (revandchinone-2) and revandchinone-3 have also been reported from *R. emodii* (Babu et al. 2003; Singh et al. 2005).

4.2.8 *Swertia chirata*

About 40 *Swertia* species are present in India. These species are randomly dispersed in the Western and Eastern Himalayan regions and Western Ghats. *Swertia* herb is used as a principal component in several commercial polyherbal formulations. *S. chirata* is known as the most well-known and elite species of *Swertia* (Kaur et al. 2019b). Amarogentin, swertiamarin (Fig. 4.6) and mangiferin are responsible for the therapeutic potential of *Swertia* (Kumar and Van Staden 2015). Kaur et al. (2019b) reported phytochemical diversity among 48 accessions of five *Swertia*

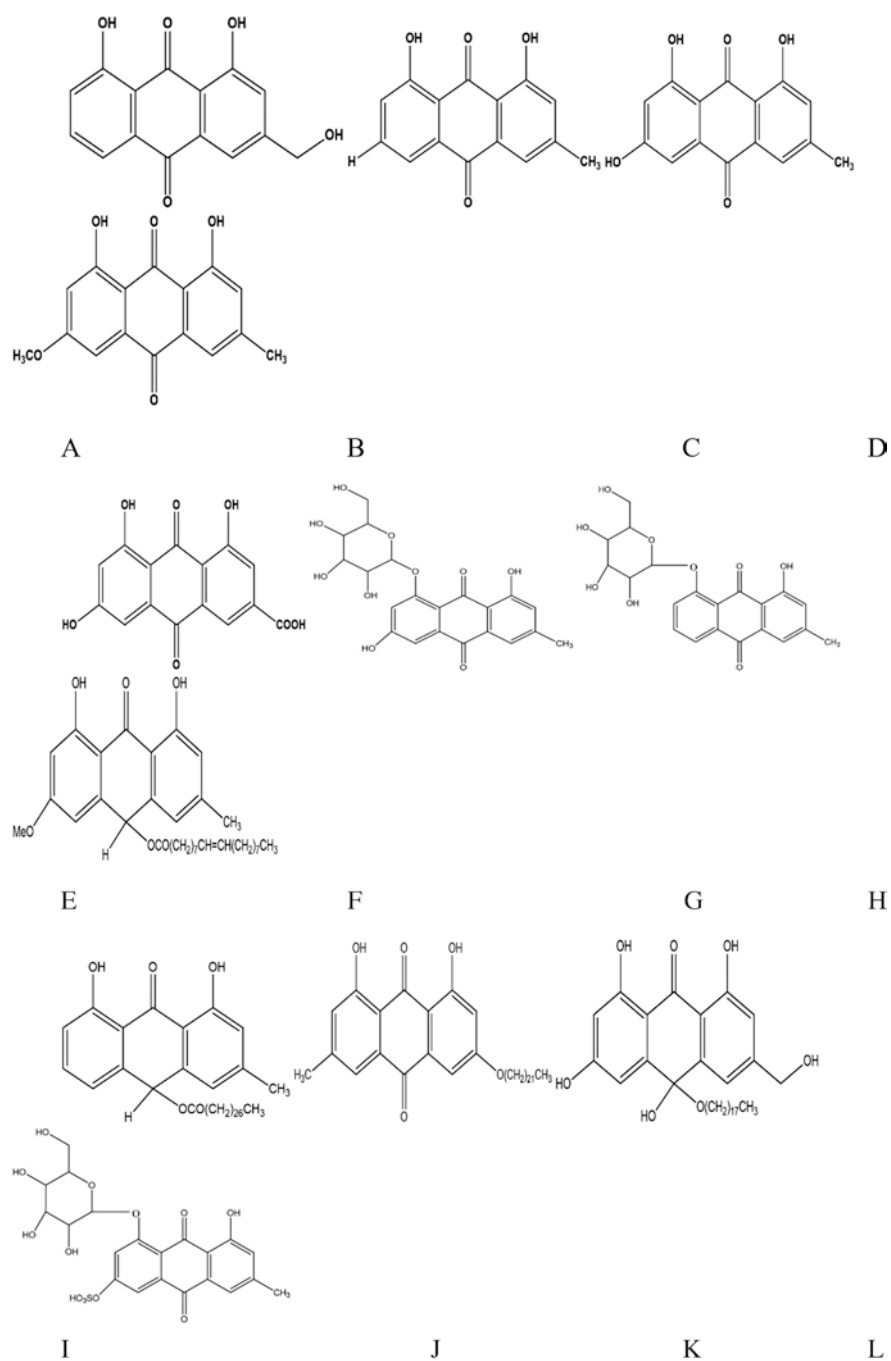


Fig. 4.5 Chemical structures of (a) aloemodin (b) crysofanol (c) emodin (d) physicon (e) rhein (f) emodin glucoside (g) crysofanol glucoside (h) revandchinone-I, (i) revandchinone-II, (j) revandchinone-III (k) revandchinone-IV and (l) sulfemodin-8-O-glucoside

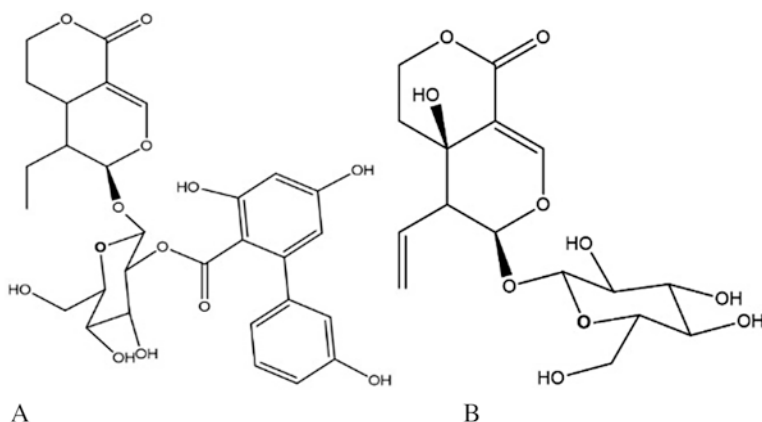


Fig. 4.6 Chemical structure of (a) amarogentin and (b) swertiamarin

species collected from Western Himalaya, India (*S. chirata*, *S. cordata*, *S. nervosa*, *S. paniculata* and *S. angustifolia*). Maximum yield of amarogentin ($0.75 \pm 0.20\%$) and swertiamarin ($6.68 \pm 0.10\%$) was found in *S. chirata* accessions and it was followed by *S. paniculata* accessions that had 0.66 ± 0.10 amarogentin and $5.76 \pm 0.03\%$ swertiamarin content. Also, substantial amount of secoiridoid glycosides were recorded in accessions of *S. angustifolia*. Similar results were reported in another study that represented high level of phytochemical diversity in the *Swertia* species/population on the basis of four triterpenoids namely oleanolic acid, ursolic acid, betulinic acid and lupeol (Kaur et al. 2019a).

4.2.9 *Valeriana jatamansi*

Valeriana jatamansi belonging to family *Valerinaceae*, is commonly known as the Indian Valerian of Tagar. *V. jatamansi* is used in both traditional and modern systems of medicine. Among the top selling herbal supplements, the drug valerian ranks at eighth place (Blumenthal 2001). *V. jatamansi* has a long history of uses as a medicine in the Rigveda, Charak Samhita and modern medicine (Jugran et al. 2019). It is used as an ingredient in the preparation of 39 Ayurvedic formulations (Prakash and Mehrotra 1991; Rawat and Vashishta 2011). Valepotriates and valerenic acid (Fig. 4.7) derived from the roots/rhizomes of *V. jatamansi* and other related species of the genus are considered to constitute the chemical fingerprint of these species. These two phytochemicals are used for assuring the quality of the plants (Singh et al. 2006, 2010; Jugran et al. 2015). Valerenic acid has been reported to possess sedative and antispasmodic properties (Houghton 1999). Content of valerenic acid in *V. jatamansi* has been explored (Singh et al. 2006, 2010; Jugran et al. 2015). Jugran et al. (2015) reported significant variation in valerenic acid content (%) in aerial and root portions of 25 population of *V. jatamansi*. It was in the

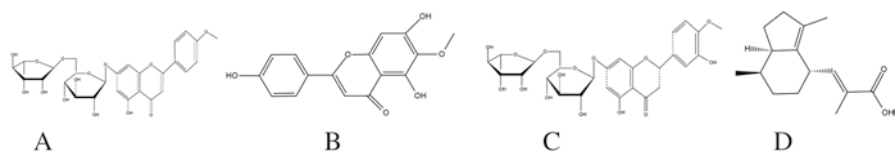


Fig. 4.7 Chemical structure of valepotriates (a) linarin (b) 6-methylapigenin (hispidulin) (c) hesperidin and valeric acid (d)

Table 4.1 Chemical diversity of nine selected Indian medicinal plants

Medicinal plant	Selected phytochemical markers	Reference
<i>Aconitum heterophyllum</i>	Aconitine	Jabeen et al. (2011)
<i>Ephedera foliata</i>	Alkaloid (ephedrine, (-)-ephedrine and (+)-pseudoephedrine, (-)-norephedrine, (+)-methylephedrine)	Bruneton (1995), Chaudhary et al. (2020)
<i>Malaxis acuminata</i>	Fatty acids, α -hydroxy acids, phenolic acids, sterols, amino acids, sugars and glycosides	Bose et al. (2017)
<i>Pterocarpus marsupium</i>	Pterostilbene, epicatechin, pterocarpol, pterosupin, pterocarposide and marsuposides	Teixeira da Sliva et al. (2018)
<i>Podophyllum hexandrum</i>	Podophyllotoxin	Sultan et al. (2008)
<i>Rauwolfia serpentina</i>	Reserpine, reserpiline, rescinnamine, ajmaline, ajmalacine, rauwolfinine, serpentine, serpentinine and yohimbine	Sahu (1983), Gao et al. (2012), Nair et al. (2014)
<i>Rheum emodi</i>	Anthraquinone, alkyl derivatives of anthraquinone, anthrone C-glucosides	Malik et al. (2010), Singh et al. (2005), Babu et al. (2003)
<i>Swertia species</i> (<i>S. chirata</i> , <i>S. cordata</i> , <i>S. nervosa</i> , <i>S. paniculata</i> and <i>S. angustifolia</i>)	Amarogentin, swertiamarin Oleanolic acid, ursolic acid, betulinic acid and lupeol	Kaur et al. (2019b) Kaur et al. (2019a)
<i>Valeriana jatamansi</i>	Valeric acid	Jugran et al. (2015)

range of 0.13 ± 0.01 to 0.57 ± 0.04 and 0.20 ± 0.03 to 1.70 ± 0.02 for aerial parts and roots, respectively.

The major phytochemicals present in the selected nine medicinal plants have been described in Table 4.1. Selected major phytochemical marker compounds may find use as reference for the purpose of certification of authentic materials, processing, quality control and value addition for post cultivation management. This information could be utilized to explore the cultivation prospects of these species in terms of technical and economical feasibility as well as marketability.

4.3 Ethnobotanical Research of MAP in India

4.3.1 Role of Ethnic Knowledge in Drug Discovery

Ethnobotanical knowledge is still transmitted from generation to generation chiefly by word of mouth. The botanical collections of early explorers and the later ethnobotany have played an important role in the development of new drugs for many centuries. In the middle of the last century interest in this approach had declined dramatically, but has risen again during last decade, when also new focuses have developed. Paul and Balick (1994) pointed out some important drugs discovered in different parts of the world, based on the ethnomedicinal knowledge (Table 4.2). Historically, much corporate drug discovery has depended on indigenous knowledge delivered to modern science through ethnobotany. Over 50% of modern prescription medicines have originally been discovered from plants and the reason behind that is that the plants were used in indigenous medicine and some common drugs were first used only on a local scale. In Europe, for example, aspirin was first isolated from *Filipendula ulmaria* because it had long been used in folk medicine. Another European folk cure that has become a drug was derived from *Digitalis purpurea*. The leaves of this plant were first used to treat congestive heart failure. Its active ingredients, digitoxin and digoxin have remained an important treatment for heart ailments. Balick and Cox (1996) showed that at least 89 plant-derived medicines used in the industrial world had originally been discovered by studying indigenous medicine. Among them, the best known is quinine, used in South America to treat fever. This has been the single most effective cure for malaria. Quinine comes from the bark of *Cinchona* trees that grow in the Andean region. More recently, the drugs vincristine and vinblastine were discovered in the rosy periwinkle (*Catharanthus roseus*) from Madagascar. When the Eli Lilly Company studied this plant, they found that the periwinkle had anti-cancer properties. Vincristine has given children with leukemia a likelihood of remission and vinblastine has cured many people with Hodgkin's disease. Native American peoples used the mayapple (*Podophyllum peltatum*) to treat warts. Two important drugs have been derived from

Table 4.2 Important drugs discovered on the basis of ethnomedicinal knowledge

Drug	Plant name	Medicinal use
Aspirin	<i>Filipendula ulmaria</i>	Reduces pain and inflammation
Codeine	<i>Papaver somniferum</i>	Eases pain and suppresses coughing
Ipecac	<i>Psychotria ipecacuanha</i>	Induces vomiting
Pilocarpine	<i>Pilocarpus jaborandi</i>	Reduces pressure in the eye
Pseudoephedrine	<i>Ephedra sinica</i>	Reduces nasal congestion
Quinine	<i>Cinchona pubescens</i>	Against malarial fever
Reserpine	<i>Rauwolfia serpentina</i>	Lowers blood pressure
Scopolamine	<i>Datura stramonium</i>	Eases motion sickness
Theophylline	<i>Camellia sinensis</i>	Open bronchial passages
Vinblastine	<i>Catharanthus roseus</i>	Combats Hodgkin's disease

it and are *teniposide* to treat bladder cancer and podophyllotoxin from which a powerful anti-tumor agent has been synthesized (Balick and Cox 1996).

To date, in India, based on ethnomedicinal knowledge, several drugs have been developed (Table 4.3). These are marketed by the pharmaceutical companies and research is going on in the development of drugs for some ailments including cardiovascular drugs from *Terminalia arjuna*; antidiabetic drugs from *Momordica charantia*, *Gymnema sylvestre* and *Andrographis paniculata*; antiprotozoal drugs from *Selaginella bryopteris*; anti malaria drugs from *Azadirachta indica*, *Ancistrocladus heyneanus*; for antileishmanial drugs *Diospyros spp.*, and *Plumbago spp.* (Bhutani and Gohil 2010).

4.3.2 Future Scope of Ethbotanical Research in India

Medicinal plant species still unknown from a phytochemical point of view, have been used to cure of ailments of different types. Ethnic population makes resort to traditional medicine because of difficult access to Western medicine as well as their high cost. These people use a wide range of plants therapeutically to maintain their health. There is great promise for new drug discovery based on traditional plant uses. Globally, 119 plant derived drugs from 90 plants are in use (Farnsworth and Morris 1985). Significantly, 77% of these were obtained as a result of examining the plants based on ethnomedical uses (Cordell 2000).

The value of ethnomedicine was recognized about six decades back in India with the pioneering work of Jain (1994). The Tropical Botanic Garden and Research Institute (TBGRI) conducted an ethnobotanical field study in the forests of south-west India in 1987. These forests are home to the Kani tribe, nomadic traditional

Table 4.3 Important drugs developed in India based on ethnomedicinal knowledge

Drug	Plant name	Medicinal use
Vasicine	<i>Adathoda zeylanica</i>	Bronchial disorder
Flavonoids	<i>Euphorbia prostrata</i>	Piles
Sennosides	<i>Cassia spp.</i>	Constipation
Baccosides	<i>Bacopa monnieri</i>	Memoray enhancer
Tylophorine	<i>Tylophora indica</i>	Bronchial disorder
Conessine	<i>Holarrhena antidysenterica</i>	Antiamoebic
Shatavarin	<i>Asparagus racemosus</i>	Tonic
Monoterpenes	<i>Ocimum sanctum</i>	Respiratory disease
Flavonoids	<i>Bauhinia variegata</i>	Diarrhoea, piles
Monoterpenes	<i>Cyperus rotundus</i>	Antibacterial
Boerhavinones	<i>Boerhavia diffusa</i>	Hepatoprotective
Anthocyanins	<i>Syzygium cumini</i>	Anti diabetic
Flavonoids	<i>Vitex negundo</i>	Anti inflammatory

collectors of non-timber forest products. The Kanis use a wild plant species for energy that they called arogyapacha, identified as *Trichopus zeylanicus* by the TBGRI. It provided a lead in the development of the drug “Jeevani” (giver of life) after the TBGRI transferred the manufacturing license to an Ayurvedic drug company in India. The TBGRI agreed to share 50% of the license fee and the 2% royalty on profits with the Kani (Anuradha 1998). Ethnobotany remains a fascinating and promising area of study for northeast India. The information about folk medicine of North-East India are still not gathered in systemic way or not documented in old literature, these are generally passed over generation to generation vocally. Multidisciplinary research and development work using the traditional folk medicinal plants based upon their traditional knowledge can provide deep motivation for identification of new pharmacophores. Newer approaches utilizing collaborative, multidisciplinary research on ethnomedicinal knowledge will help in near future in improving healthcare worldwide particularly from northeastern region of India. Some of the preliminary laboratory works carried out based on ethnobotanical knowledge in northeast India has been described here. Kar et al. 2005 carried out investigation based on traditional knowledge of Karbi and Hmar tribe of Assam on antimicrobial properties of extracts of *Curunga amara* Juss. against human pathogenic microorganisms. Tayung and Kar (2005) carried out investigation based on traditional knowledge of Monpa tribe of Arunachal Pradesh on antimicrobial activity of *Thalictrum javanicum* (Blume) root extracts against some human pathogens. Kalita et al. (2012) carried out antimicrobial study on *Paederia foetida* and *Hibiscus esculentus* which are generally used against stomach troubles, diarrhoea, hypertension, skin diseases, in urinary troubles and dental problem by the tribes of Assam. Similarly, *Hibiscus esculentus* extract was tested against the growth of *Staphylococcus aureus*. It possessed the potentiality against the growth of *E. coli*. Vijayakumar et al. (2012) carried out investigation on the *Illicium griffithii* fruit and seed from Arunachal Pradesh based on the ethnic knowledge. Phytochemical qualitative analysis revealed the presence of phenols, tannins, flavonoids, triterpenoids, steroids, alkaloids in the seeds. Ethyl acetate extract of fruits showed significant activity against *Staphylococcus aureus*, *Bacillus subtilis*, *Yersinia enterocolitica*, *Vibrio parahaemolyticus*, *Salmonella paratyphi*, *Xanthomonas oryzae* and *Pseudomonas aeruginosa*; methanol extract showed activity against *S. aureus*, *Bacillus subtilis* and *Xanthomonas oryzae*. Haripyaree et al. (2013) carried out microbial investigation *Mimosa pudica* L, *Vitex trifolia* Linn, *Leucas aspera* Spreng, *Centella asiatica* (L) Urban and *Plantago major* Linn against antimicrobial screening of six organisms viz., *Ceratocystis paradoxa*, *Aspergillus niger*, *Penicillium citrinum*, *Macrophomina phaseoli*, *Trichoderma viride* and *Rhizopus nigricans* in Manipur and reported that *M. pudica* showed highest antifungal activities against more than one microorganism. Methanol and hexane extract of *M. pudica* and *V. trifolia* showed moderate and strong activities against *C. paradoxa*. *C. asiatica* extract showed activity against *M. phaseoli*; *Leucas aspera* exhibited antifungal activity against pineapple fruit rotting fungus *C. paradoxa*. Kalita et al. (2012) carried out an investigation on local medicinal plants of Assam. Some of the research institutes of northeast India are doing research on drug formulation based on ethnic knowledge and few of them

are *Cajanus cajan* against jaundice, *Gomphostemma spp.* against malarial fever, *Terminalia spp.* against fungal diseases, *Oroxylum indicum* against cancer, *Dillenia indica* against diabetes. In addition to that some of research institute outside of northeast India collect plant sample from northeastern states and doing research in their laboratory. The above-mentioned species are some of the numerous examples only and there seems to be a possibility to explore more number of medicinal plants from this part of India for the development of useful drugs.

Existing rich ethnic heritage of India could be further explored through more dedicated ethnobotanical studies. It has also been gratifying that integrated forms of modern and traditional medicine have remained a part of reality. Information gathered from Ethnobotanical study would also be useful for conservation of traditional knowledge which is essentially required to save the cultural heritage of the natives. In this context confirmation of the therapeutical uses of the plants with scientific criteria and fostering phytochemical research on species containing potentially active principles would be more relevant for harnessing the value of ethnobotanical research carried out.

Silambarasan and Ayyanar (2015) carried out ethnobotanical studies in Eastern Ghats and recorded a total of 118 plants. Gairola et al. (2014) recorded a total of 948 plant taxa (923 angiosperms, 12 gymnosperms and 13 pteridophytes) belonging to 129 families, 509 genera, 937 species and 11 varieties from Jammu and Kashmir and Ladhakh for which no traditional medicinal use by indigenous communities of have been reported. Dey and De (2012) reported 56 plant species used against different types of gastrointestinal disorders like indigestion, stomach pain, vomiting tendency, constipation, piles, diarrhea, dysentery, cholera, loss of appetite, liver complaints, intestinal worms etc. Tetali et al. (2009) recorded 182 plants from Pune district of Maharashtra used by tribes and natives for different ailments. From these plants, 28 flowering plants were documented for diarrhoea. Amongst the 28 plants, antidiarrhoeal activity of five plants viz., *Caesalpinia sepiaria*, *Dioscorea pentaphylla*, *Launaea pinnatifida*, *Syzygium rubicundum* and *Ziziphus jujuba* has not been reported previously. Two species viz., *Ziziphus xylopyra* and *Syzygium rubicundum* are endemic to India. Bisht and Adhikari (2018) reported 70 medicinal plants from Uttarakhand which have been used against 31 ailments. Kaur et al. (2011) recorded 15 medicinal plant species used to treat leprosy, arthritis, nasal bleeding, ulcer etc. from Himachal Pradesh. Kaur et al. (2020) reported 51 plant species used to treat gastrointestinal disorder. Parul et al. (2017) recorded 18 medicinal plants from Harayana which have been used against digestive disorder.

4.4 Conclusions

Increasing demand on MAPs frequently comes along with the illegal overharvesting and unscrupulous collection practice of endangered plant species from the wild. Future strategies, regarding the conservation of the endangered plant species, are a major concern. Adaptation of some advanced plant biotechnological techniques,

namely micro-propagation, hairy root technologies and synthetic seed production may be useful in securing surplus supplies of such plant species to meet its future demand. Isolation of most effective compounds and development of analytical tools of various *in vitro* and *in vivo* studies may result numerous opportunities to further unravel the potential bioactivities of the species. Additionally, *in silico* molecular docking techniques may play an important role in the identification/design of the most effective molecules. These effective molecules may be synthesized from their analogues available in higher quantity to reduce the pressure on their natural habitats. Also, research on chemical diversity to identify the active constituents will open up opportunities to discover new chemotypes, as promising sources of drugs. There is a great scope for ethnobotanical studies as it points out to the species which most urgently should be studied scientifically. However, this approach in search of new pharmaceuticals is woefully underutilized.

References

- Anjen L, Bojian B, Grabovskaya-Borodina AE, Hong SP, McNeill J, Mosyakin SL, Ohba H, Park CW (2003) Polygonaceae. In: Wu ZY, Raven PH, Hong DY (eds) Flora of China, vol 5. Science Press/Missouri Botanical Garden Press, Beijing/St. Louis, pp 277–350
- Anuradha RV (1998) Sharing with the Kanis: a case study from Kerala, India, Secretariat to the Convention on Biological Diversity, Fourth Meeting of the COP, Bratislava, Slovakia
- Babu KS, Srinivas PV, Praveen B, Kishore KH, Murthy US, Rao JM (2003) Antimicrobial constituents from the rhizomes of *Rheum emodi*. *Phytochemistry* 62:203–207
- Balick MJ, Cox PA (1996) Plants, people, and culture: the science of ethnobotany, Scientific American Library series 60. Scientific American Library, New York, pp 2–3
- Becker H (2000) Mayapple's cancer –fighting precursor. *Agric Res* 48:9
- Bernáth J (2002) Strategies and recent achievements in selection of medicinal and aromatic plants. In: Bernáth J et al (eds) International conference on MAP., Acta Hort. ISHS, pp 115–128
- Beutner KR (1996) Podophyllotoxin in the treatment of genital warts. In: Eischman EP (ed) Sexually transmitted diseases. *Adv Treat*, vol 24, pp 122–132
- Bhandari MM (1978) Flora of the Indian desert. Scientific Publisher Jodhpur, p 439
- Bhutani KK, Gohil VM (2010) Natural products drug discovery research in India: status and appraisal. *Indian J Exp Biol* 48:199–207
- Bindu S, Rameshkumar KB, Kumar B, Singh A, Anilkumar C (2014) Distribution of reserpine in *Rauvolfia* species from India –HPTLC and LC-MS studies. *Ind Crop Prod* 62:430–436
- Bisht S, Adhikari BS (2018) Ethnobotanical study of traditional medicinal plants used by Banraji Community in Uttarakhand, West Himalaya. *J Ethnobiol Trad Med* 129:1426–1441
- Bleuler M, Stoll WA (1955) Clinical use of reserpine in psychiatry: comparison with chlorpromazine. *Ann N Y Acad Sci* 61:167–173
- Blumenthal M (2001) Herb sales down 15 percent sales in mainstream market. *Herbal Gram* 59:69
- Bose B, Choudhury H, Tandon P, Kumaria S (2017) Studies on secondary metabolite profiling, anti-inflammatory potential, *in vitro* photoprotective and skin-aging related enzyme inhibitory activities of *Malaxis acuminata*, a threatened orchid of nutraceutical importance. *J Photochem Photobiol B Biol* 173:686–695
- Bruneton J (1995) Pharmacognosy, phytochemistry, medicinal plants. Lavoisier Publishing, Paris
- Bryde MB (1979) Information needed to use the endangered species act for plant conservation, geographical data organization. Rare Plant Conservation, New York

- Calixto JB (2000) Efficacy, safety, quality control, marketing and regulatory guidelines for herbal medicines (phytotherapeutic agents). *Braz J Med Biol Res* 33:179–189
- Chaudhary MK, Misra A, Srivastava S (2020) Evaluation of ephedrine content and identification of elite chemotypes of *Ephedra gerardiana* (Wall.) from Kashmir Himalayas. *Proc Natl Acad Sci India Sect B Biol Sci* 90:833–841
- Chauhan NS (1999) Medicinal and aromatic plants of Himachal Pradesh. Indus Publishing, Delhi
- Cheruvathur MK, Abraham J, Mani B, Thomas TD (2010) Adventitious shoot induction from cultured intermodal explants of *Malaxis acuminata* (D. Don.) a valuable terrestrial medicinal orchid. *Plant Cell Tissue Organ Cult* 101:163–170
- Chopra RN, Chopra IC, Handa KL, Kapur LD (1958) Indigenous drugs of India. Academic Publishers, New Delhi, pp 30–36
- Cordell G (2000) Biodiversity and drug discovery: a symbiotic relationship. *Phytochemistry* 55:463–480
- Cragg GM, Newman DJ, Snader KM (1997) Natural products in drug discovery and development. *J Nat Prod* 60:52–60
- Devgun M, Nanda A, Ansari SH (2009) *Pterocarpus marsupium* Roxb. – a comprehensive review. *Phcog Rev* 3:359–363
- Dey A, De JN (2012) Ethnobotanical survey of Purulia district, West Bengal, India for medicinal plants used against gastrointestinal disorders. *J Ethnopharmacol* 143(1):68–80. <https://doi.org/10.1016/j.jep.2012.05.064>. Epub 2012 Jun 19. PMID: 22721882
- Dhiman B, Sharma P, Shivani and Pal, P.K. (2020) Biology, chemical diversity, agronomy, conservation and industrial importance of *Valeriana jatamansi*: a natural sedative. *J Appl Res Med Aromat Plants* 16:100243
- Farnsworth NR, Morris RW (1985) Higher plants- the sleeping giant of drug development. *Am J Pharm Educ* 49:46–52
- Farooqi AA, Sreeramu BS (2001) Cultivation of medicinal and aromatic crops. University Press Ltd., Hyderabad, pp 210–211
- Gairola S, Sharma J, Bedi YS (2014) A cross-cultural analysis of Jammu, Kashmir and Ladakh (India) medicinal plant use. *J Ethnopharmacol* 155(2):925–986. <https://doi.org/10.1016/j.jep.2014.06.029>. Epub 2014 Jun 19. PMID: 24952280
- Gao Y, Zhou DS, Kong LM, Hai P, Li Y, Wang F, Liu JK (2012) Rauvofe-traphyllines A-E new indole alkaloids from *Rauvolfia tetraphylla*. *Nat Product Bioprospect* 2:65–69
- Gavali D, Sharma D (2004) Traditional knowledge and biodiversity conservation in Gujarat. *Indian J Tradit Knowl* 3:51–58
- Govindarajan R, Singh DP, Rawat AKS (2007) High-performance liquid chromatography method for the quantification of phenolics in “Chyavanprash” a potent Ayurvedic drug. *J Pharm Biomed Anal* 43:527–532
- Hardy G (2000) Nutraceuticals and functional foods: introduction and meaning. *Nutrition* 16:688–698
- Hariyaree A, Guneshwor K, Damayanti M (2013) Antifungal and cytotoxic activities of five traditionally used Indian medicinal plants. *J Microbiol Biotechnol Food Sci* 2(4):2272–2278
- Henderson CW (2000) American mayapple yields anti-cancer extract. *Cancer Weekly* 8:24–25
- Hennicke F, Cheikh-Ali Z, Liebisch T, Macia-Vicente JG, Bode HB, Piepenbring M (2016) Distinguishing commercially grown *Ganoderma lucidum* from *Ganoderma lingzhi* from Europe and East Asia on the basis of morphology, molecular phylogeny and triterpenic acid profiles. *Phytochemistry* 127:29–37
- Heywood VH (2002) The conservation of genetic and chemical diversity in medicinal and aromatic plants. In: Sener (ed) Biodiversity: biomolecular aspects of biodiversity and innovative utilization. Kluwer Academic/Plenum Publishers, New York/Boston/Dordrecht, pp 13–22
- Hikino H, Murakami M, Konno C, Watanbe H (1983) Determination of aconitine alkaloids in *Aconitum* roots. *Planta Med* 48:67–71
- Hooker JD (1885) *Flora of British India*, vol V. L. Reeve and Co., London

- Houghton PJ (1999) Valerian the genus *Valeriana*. Harwood Academic Publishers, Abingdon, pp 16–18
- IUCN (1978) Plant red data book. Royal Botanic Garden, Kew
- IUCN (1993) Guidelines on the conservation of medicinal plants. The International Union for Conservation of Nature and Natural Resources (IUCN), Gland, p 38
- IUCN (2017) World Conservation Monitoring Centre 1998. *Pterocarpus marsupium*, IUCN red list threat. Species. version 2017.2
- Iwasa J, Naruto S (1996) Alkaloids from *Aconitum carmichaeli*. Yakugaku Zasshi 86:585–590
- Jabeen N, Rehman S, Bhat KA, Khuroo MA, Shawl AS (2011) Quantitative determination of aconite in *Aconitum chasmanthum* and *Aconitum heterophyllum* from Kashmir Himalayas using HPLC. J Pharm Res 4:2471–2473
- Jackson DE, Dewick PM (1984) Biosynthesis of *Podophyllum lignans*-II. Inter conversion of aryl-trialin lignan in *Podophyllum hexandrum*. Phytochemistry 23:1037–1042
- Jain SK (1968) Medicinal plants. National Book Trust, New Delhi
- Jain SK (1994) Ethnobotany and research on medicinal plants in India. Ciba Found Symp 185:153–164; discussion 164–8. PMID: 7736852
- Jalal JS (2012) Status, threat and conservation strategies for orchids of Western Himalaya, India. J Threatened Taxa 4:3401–3409
- Joshi PN, Soni HB, Sunderraj SFW, Joshua J (2013) Conservation and management strategies for threatened plant species of Kachchh Desert Island, Gujarat, India. IJE 2:45–59
- Jugran AK, Bhatt ID, Rawal RS, Nandi SK, Pande V (2013) Patterns of morphological and genetic diversity of *Valerianajatamansi* in different habitats and altitudinal range of West Himalaya, India. Flora 208:13–21
- Jugran AK, Bhatt ID, Rawal RS (2015) Identification of ISSR markers associated with valerenic acid antioxidant activity in *Valeriana jatamansi* Jones in West Himalaya. Mol Breed 35:1–14
- Jugran AK, Rawat S, Bhatt ID, Rawal RS (2019) *Valeriana jatamansi*: an herbaceous plant with multiple medicinal uses. Phytother Res 33:482–503
- Kalita D, Saikia JC, Sindagi AS, Anmol GK (2012) Antimicrobial activity of leaf extract of two medicinal plants of Bogora hills (Morigaon) Assam. The Bioscan 7(2):271–274
- Kar A, Gogoi DK, Rabha B, Gogoi HK (2005) *In vitro* evaluation of antimicrobial properties of stem with leaf extracts of *Curunga amara* Juss against human pathogenic microorganisms. Asian J Microbiol Biotech Environ Sci 7(4):759–762
- Kaur S, Das M (2011) Functional foods: an overview. Food Sci Biotechnol 20:861–875
- Kaur I, Sharma S, Lal S (2011) Ethnobotanical survey of medicinal plants used for different diseases in Mandi district, Himachal Pradesh. Int J Pharm Chem 1:1167–1171
- Kaur P, Gupta RC, Dey A, Pandey DK (2019a) Simultaneous quantification of oleanolic acid, ursolic acid, betulinic acid and lupeol in different populations of five *Swertia* species by using HPTLC-densitometry: comparison of different extraction methods and solvent selection. Ind Crop Prod 130:537–546
- Kaur P, Pandey DK, Gupta RC, Dey A (2019b) Assessment of genetic diversity among different population of five *Swertia* species by using molecular and phytochemical markers. Ind Crop Prod 138:115569
- Kaur K, Kaur A, Thakur A (2020) Use of medicinal plants in traditional health care practices: a case study in Talawandi Sabo, Bathinda District, Punjab (India). Curr Botany 11:75–86
- Kharin N (2002) Vegetation degradation in Central Asia under the impact of human activities. Springer, New York
- Kumar V, Van Staden J (2015) A review of *swertia chirayita* (Gentianaceae) as a traditional medicinal plant. Front Pharmacol 6:308
- Lohani N, Tewari LM, Joshi GC, Kumar R, Kishor K, Upreti BM (2013) Population assessment and threat categorization of endangered medicinal orchid *Malaxis acuminata* (D. Don.) from north-west Himalaya. Int J Conserv Sci 4:493–502

- Malik S, Sharma N, Sharma UK, Singh NP, Bhusan S, Sharma M, Sinha AK, Ahuja PS (2010) Qualitative and quantitative analysis of anthraquinone derivatives in rhizomes of tissue culture-raised *Rheum emodi* Wall. plants. *J Plant Physiol* 167:749–756
- Meena B, Singh N, Mahar KS, Sharma YK, Rana TS (2019) Molecular analysis of genetic diversity and population genetic structure in *Ephedra foliata*: an endemic and threatened plant species of arid and semi-arid regions of India. *Physiol Mol Biol Plants* 25:753–764
- Mishra N, Acharya R, Gupta A, Singh B, Kaul VK, Ahuja PS (2005) A simple microanalytical technique for determination of podophyllotoxin in *Podophyllum hexandrum* roots by RP-HPLC and RP-HPTLC. *Curr Sci* 88:1372–1373
- Mohankumar SK, O’Shea T, McFarlane JR (2012) Insulinotropic and insulin like effects of a high molecular weight aqueous extract of *Pterocarpus marsupium* Roxb. hardwood. *J Ethnopharmacol* 141:72–79
- Murayama M, Hikino H (1984) Stimulating actions on ribonucleic acid biosynthesis of aconites, diterpenic alkaloids of *Aconitum* root. *J Ethnopharmacol* 12:25–33
- Nair VD, Raj RPD, Panneerselvam R, Gopi R (2014) Assessment of diversity among populations of *Rauvolfia serpentina* Benth. Ex. Kurtz from Southern Western Ghats of India, based on chemical profiling, horticultural traits and RAPD analysis. *Fitoterapia* 92:46–60
- Nayar MP, Sastry ARK (1990) Red data book of Indian plants, vol I. II and III. Botanical Survey of India, Calcutta
- Panda AK, Bisaria VS, Misra S (1992) Alkaloid production by plant cell cultures of *Holarthraaentidy senteria*. Effect of precursor feeding and cultivation on stirred tank bioreactor. *Biotechnology. Bioengineering* 39:1042–1052
- Paramanick D, Panday R, Shukla SS, Sharma V (2017) Primary pharmacological and other important findings on the medicinal plants “*Aconitum heterophyllum*” (Aruna). *J Pharm* 20:89–92
- Parul RG, Lal M, Vashistha BD (2017) Ethnobotanical survey of traditional medicine practice to treat digestive disorder of Gurugram district, Harayana. *Int J Dev Res* 07(11):16623–16626
- Paul AC, Balick MJ (1994) The ethnobotanical approach to drug discovery. *Sci Am* 270(6):82–87
- Polunin O, Stainton A, Farrer A (1987) Concise flowers of the Himalaya. Oxford University Press, London
- Prakash V, Mehrotra BN (1991) Ethno-medicinal uses of some plants among Garos of Meghalaya. *Ethnobotany* 3:41–45
- Prasad P (2000) Impact of cultivation on active constituents on the medicinal plants *Podophyllum hexandrum* and *Aconitum heterophyllum* in Sikkim. *PGR Newsl* 124:33–35
- Qaderi A, Omidi M, Pour-Aboughadareh A, Poczai P, Shaghghi J, Mehrafarin A, Nohooji MG, Etminan A (2019) Molecular diversity and phytochemical variability in the Iranian poppy (*Papver bracteatum*): a baseline for conservation and utilization in future breeding programme. *Ind Crop Prod* 130:237–247
- Quattrocchi U (2012) CRC world dictionary of medicinal and poisonous plants: common names, scientific names, eponyms, synonyms and etymology. Taylor & Fransis, London, p 1578
- Rawat R, Vashishta DP (2011) Common Herbal Plant in Uttarakhand used in popular medicine preparation in Ayurveda. *Int J Pharamacogn Phytochem Res* 3:64–73
- Ray A, Jena S, Haldar T, Sahoo A, Kar B, Patnaik J, Ghosh B, Panda PC, Mahapatra N, Nayak S (2019) Population genetic structure and diversity analysis in *Hedychium coronarium* populations using morphological, phytochemical and molecular markers. *Ind Crop Prod* 132:118–133
- Sahni KC (1990) Gymnosperms of India and adjacent countries. Bishen Singh Mahendra Pal Singh, Dehradun, pp 139–140
- Sahu BN (1983) *Rauvolfias*. Chemistry and pharamacology, vol II. Today and Tomorrow Printers and Publishers, New Delhi
- Satyavathi GV, Gupta AK, Tondon N (1987) Medicinal plants of India. ICMR Publications, New Delhi
- Sharma P, Singh R (2015) A new species of *Ephedra* (Ephedraceae, Ephedrales) from India. *Phytotaxa* 218:189–192

- Sharma BD, Balakrishnan NP, Rao RR, Hajra PK (1993) Flora of India, vol 1. Botanical Survey of India, Calcutta, pp 3–23
- Shu YZ (1998) Recent natural products based drug development: a pharmaceutical industry perspective. *J Nat Prod* 61:1053–1071
- Silambarasan R, Ayyanar M (2015) An ethnobotanical study of medicinal plants in Palamalai region of Eastern Ghats, India. *J Ethnopharmacol* 172:162–178. <https://doi.org/10.1016/j.jep.2015.05.046>. Epub 2015 Jun 9. PMID: 26068426
- Silori CS, Dixit AM, Gupta L, Mistry N (2005) Observation on medicinal plant richness and associated conservation issues in district Kachchh, Gujarat. In: Trivedi PC (ed) Medicinal plants: utilization and conservation. Avishkar Publishers, Jaipur, p 154
- Singh SS, Pandey SC, Singh R, Agarwal SK (2005) 1,8 dihydroxy-antraquinone derivatives from rhizomes of *Rheum emodi* Wall. *Indian J Chem* 43B:1494–1496
- Singh N, Gupta AP, Singh B, Kaul VK (2006) Quantification of valerinic acid in *Valeriana jatamansi* and *Valeriana officinalis* by HPTLC. *Chromatographia* 63:209–213
- Singh RD, Gopichand, Meena RL, Sharma B, Singh B, Kaul VK, Ahuja PS (2010) Seasonal variation of bioactive components in *Valeriana jatamansi* from Himachal Pradesh, India. *Ind Crop Prod* 32:292–296
- Song WL, Chen DH, Wang LW, Xiango PG (1984) Chemical constitution and resource utilization of Dou Gen Wu Tou (*Aconitum karakolicum*). *Chin Tradit Herb Drug* 15:5–7
- Sukumaran S, Raj ADS (2008) Rare and endemic plants in the sacred groves of Kanyakumari district in Tamil Nadu. *Indian J For* 31:611–616
- Sultan P, Shawl AS, Ramteke PW, Kour A, Qazi PH (2008) Assessment of diversity in *Podophyllum hexandrum* genetic and phytochemical markers. *Sci Hortic* 115:398–408
- Tayung K, Kar A (2005) Antimicrobial activity of *Thalictrum javanicum* (Blume) root extracts against certain human pathogens. *J Curr Sci* 7(2):341–345
- Teixeira da Sliva JA, Kher MM, Soner D, Nataraj M (2018) Indian kino tree (*Pterocarpus marsupium*): propagation, micropropagation and biotechnology. *Environ Exp Biol* 16:1–8
- Tetali P, Waghchaure C, Daswani PG, Antia NH, Birdi T (2009) Ethnobotanical survey of anti-diarrhoeal plants of Parinche valley, Pune district, Maharashtra, India. *J Ethnopharmacol* 123(2):229–236. <https://doi.org/10.1016/j.jep.2009.03.013>. Epub 2009 Mar 24. PMID: 19429366
- Vijayakumar AD, Jeyaraj VB, Agastian P, Karunai RM, Ignacimuthu S (2012) Phytochemical analysis and *in vitro* antimicrobial activity of *Illicium griffithii* Hook. f. & Thoms extracts. *Asian Pac J Trop Dis* 2:190–199
- Weiner AM (1990) Weiners herbal. The guide to herbal medicine. Quantum Books, Mill Valley, pp 202–203
- Yadav RNS, Agarwala M (2011) Phytochemical analysis of some medicinal plants. *J Phytotherapy* 3:10–14
- Zargar BA, Masoodi MH, Ahmed B, Ganie SA (2011) Phytoconstituents and therapeutic uses of *Rheum emodi* wall. *Ex. Meissn. Food Chem* 128:585–589