

Chapter 13

Application of Encapsulation Technology in the Agri-Food Sector



Marko Vinceković and Slaven Jurić

1 Introduction Into the Encapsulation Technology to the Agri-Food-Bio Sciences

Encapsulation might be defined as a process of substance (*internal phase, payload, or payload phase*) insertion into another substance (*membrane, shell, capsule, carrier material, external phase, wall, or matrix*) (Vinceković et al. 2021; Nedović et al. 2011). Throughout the encapsulation process, various sizes of particles can be produced, from a few nm (nanoparticles) to a few mm (microparticles) (Lengyel et al. 2019). The encapsulation technology was first introduced to the area of biotechnology to increase the efficiency of products. The developed technology became huge interest in the other areas like pharmaceuticals and cosmetics industry, as well as agricultural and food industry.

There are several advantages towards using the encapsulation process in agri-food-bio sciences: (i) easier handling (*e.g.* converting liquid ingredients into a powder form, which can be completely free of certain impurities with better rheological and sensory (smell/odor) properties), (ii) immobilization of encapsulated material for various production processes, (iii) better stability of encapsulated material during technological preparation and in the final product (*i.e.* significant reduction of volatiles evaporation, reduced degradation/decomposition and reduction of reaction with other ingredients in the complex matrix of the product), (iv) increase the safety and security (*e.g.* reduced flammability and explosive behavior of volatile compounds and easier handling), (v) improving visible and textural effects (visual signs) in the final product (cosmetic, food and agricultural industry), (vi) tuning the properties of encapsulated material (particle size distribution, structure, solubility in organic and inorganic solvents, color), (vii) time adjustment of the release of

M. Vinceković (✉) · S. Jurić

Department of Chemistry, University of Zagreb Faculty of Agriculture, Zagreb, Croatia

e-mail: mvincekovic@agr.hr

encapsulated material which may be activated under certain conditions or with the ingredients of a complex product system or by the action of external factors (Jurić et al. 2020a).

In addition to the above advantages, encapsulation processes have certain disadvantages, as (i) increasing the costs of upscaling the encapsulation process, (ii) the industrial production and/or supply chain process are complex, (iii) the final appearance of the product is not in line with customer expectations (visually or sensory does not meet customer criteria) products, and this is especially problematic in the food production sector, (iv) stability problems of prepared nano- and microparticle formulations during storage, transport and application in certain complex system products (Vinceković et al. 2021). Despite these shortcomings mentioned, encapsulation technology is increasingly being advanced and developed and continuously used as a process in the preparation of new products in the fields of agriculture, food technology, cosmetics, and nutraceuticals.

The encapsulation technology applies to the food industry as a useful tool to improve the delivery of bioactive compounds (*e.g.* antioxidants, minerals, vitamins, phytosterols, lutein, fatty acids, lycopene, esters, aromas, colors) and living cells (*e.g.* probiotics, yeast) in real food products (Jurić et al. 2021; Mrkonjić Fuka et al. 2021; Belščak-Cvitanović et al. 2017; Vos et al. 2010). Furthermore, there is an increasing trend towards using encapsulation technology for agricultural purposes to increase the viability and to control the delivery of living microorganisms into the field. These methods proved efficient and superior to the other formulations in terms of living organisms' protection against the harsh environment (Jurić et al. 2020c; Vinceković et al. 2016). The encapsulation process can be applied for the production of particles loaded with biological and chemical agents as an advanced tool for ecological and sustainable plant production. Encapsulation in biopolymer matrices has been recognized as an effective method for the controlled release of agents used for plant protection and nutrition (Jurić et al. 2019, 2020b). In the cosmetic industry encapsulation process have been proposed to increase the stability of the material, to protect it against degradation, and also to direct and control the release of encapsulated material used in cosmetic products (Casanova and Santos 2016).

The stability of encapsulated compounds mainly depends on a combination of environmental and chemical factors (*i.e.* pH, metal ions, light, high temperatures, enzymes, and oxygen) (Mahdavee Khazaei et al. 2014). In Table 13.1 we have outlined some of the recently used stabilization techniques and carriers for encapsulation of natural pigments. Usually, with regards to the encapsulation of various ingredients, research papers deal with the fabrication and development of new production methods but worryingly the research on the inclusion of encapsulated material into *e.g.* real food products is still scarce (Jurić et al. 2020a). Due to the stability issues under environmental conditions during product manufacturing and later storage, the incorporation of particles loaded with active ingredients into final products is still extremely challenging. Even though encapsulation is always advancing and represents an effective way to protect encapsulated material, incorporation of nano- and microparticles into products is still not investigated enough (Jurić et al. 2020a).

Table 13.1 Stabilization techniques and materials used for the protection of water and lipid-soluble natural pigments (Adapted from Jurić et al. 2020a, 2020b, 2020c)

Pigment	Co-pigment/Wall materials	Stabilization technique	Reference
Water soluble			
<i>Anthocyanins</i>	Dairy proteins	Complexation	Chung et al. (2015); He et al. (2016)
	Pectins	Complexation	Lin et al. (2016)
	Whey proteins and pectins	Complexation/physical entrapment	Arroyo-Maya et al. (2016)
	Gum arabic	Complexation	Chung et al. (2016a); Guan and Zhong (2015)
	β -cyclodextrins	Molecular inclusion	Howard et al. (2013); Fernandes et al. (2013)
	Green tea extracts	Complexation	Chung et al. (2016b)
	Ferric ion	Chelation	Tachibana et al. (2014)
	Stearic acid	Lyophilization	Cruz et al. (2015)
	Oleic acids	Lyophilization	Cruz et al. (2016)
	Different fatty acids	Lyophilization	Cruz et al. (2017, 2018); Luo et al. (2017); Yang et al. (2019)
	Montmorillonite	Hybridization	Kohno et al. (2009)
	Methoxyl pectin	Ionic gelation	de Moura et al. (2018)
	Sodium alginate	Ionic gelation	da Silva Carvalho et al. (2019)
	Polyethylene glycol (PEG)	Ionic gelation	Santos et al. (2013)
	Alginate	Ionic gelation	Belščak-Cvitanović et al. (2016)
	Pectin amide	Ionic gelation	Oidtmann et al. (2012)
	Whey protein isolate	Microemulsions	Oidtmann et al. (2012)
Maltodextrin, pectin amide	Spray drying	Oidtmann et al. (2012)	
Glycerol mono-oleate, soy Lecithin, maltodextrin, poloxamer 338	Spray drying	Ravanfar et al. (2018)	
Supercritical carbon dioxide	Liposomes	Zhao et al. (2017)	
<i>Betalains</i>	Sucrose	Co-crystallization	Karangutkar and Ananthanarayan (2020)
	Sodium alginate, sodium alginate-bovine serum	Ionic gelation	Otálora et al. (2016)
	Rapeseed oil, guar gum, xanthan gum	Double emulsions	Kaimainen et al. (2015)
Lipid Soluble			
<i>Carotenoids</i>	Whey protein concentrate, gum arabic	Spray drying	Chuyen et al. (2019)
<i>β-carotene</i>	Wheat gluten nanoparticles, wheat gluten nanoparticle-xanthan gum	Pickering emulsion	Fu et al. (2019)

(continued)

Table 13.1 (continued)

Pigment	Co-pigment/Wall materials	Stabilization technique	Reference
	Maltodextrin, gum arabic, chitosan, gelatin	Spray drying	Bonilla-Ahumada et al. (2018)
	Native and hydrolyzed Pinhao starches	Freeze-drying	da Silva Carvalho et al. (2019)
<i>Lycopene</i>	Gelatin, sucrose	Spray drying	Shu et al. (2006)
	Lecithin, α -tocopherol	Supercritical antisolvent co-precipitation (SAS)	Cheng et al. (2017)
<i>Lutein</i>	Gelatin, gum arabic	Coacervation	Qv et al. (2011)
<i>Chlorophylls</i>	Polycaprolactone	Microfluidic emulsification	Hsiao et al. (2020)
	Gum arabic, maltodextrin	Spray drying	Kang et al. (2019)
	Whey protein isolate	Spray drying	Zhang et al. (2020)

2 Classification of Next-Generation Biopolymer-Based Carriers as Sustainable Materials

Research is nowadays more focused on the investigation of alternative carriers such as biopolymers. Biodegradable polymers are suitable materials for the production of NPs because of their abundance, relative stability, and durability throughout the encapsulation processes. One of the most important advantages of encapsulation in biopolymeric particles is also high food compatibility and safety which is connected with the availability of polysaccharides, proteins, and lipids (Fathi et al. 2021). Biopolymeric carriers are generally easily prepared from natural biodegradable polymers. These types of materials are usually used because they are generally regarded as safe for the consumer and environment. Also, prepared particles have superior properties especially considering controlled and targeted release (Jana et al. 2020). Biopolymeric particles are usually spherical with some deviations. It is possible to distinguish two types of particles, the *reservoir* type, and the *matrix* type. The reservoir type (*capsule*) has a shell around the bioactive component (*filler*). These types can further be divided into a single-core/mono-core or a core-shell type. The release of the payload from reservoirs can be achieved via the application of force (pressure) or under specific conditions which are generally resulting in capsule breakage. Poly- or multiple-core type particles have several reservoir chambers loaded with encapsulated material in a single particle (Vinceković et al. 2021). In the matrix type of particles, the payload is usually dispersed over the biopolymer matrix carrier and it can be in the form of homogeneously dispersed small droplets or it can be adsorbed on the surface.

Currently, various materials of different origins and properties are being used for the encapsulation process either in solid, liquid, or gaseous forms. Materials used in the production of carrier systems can be divided into three groups as proteins, polysaccharides, and lipids (Fig. 13.1). Biopolymer carriers can be prepared in various

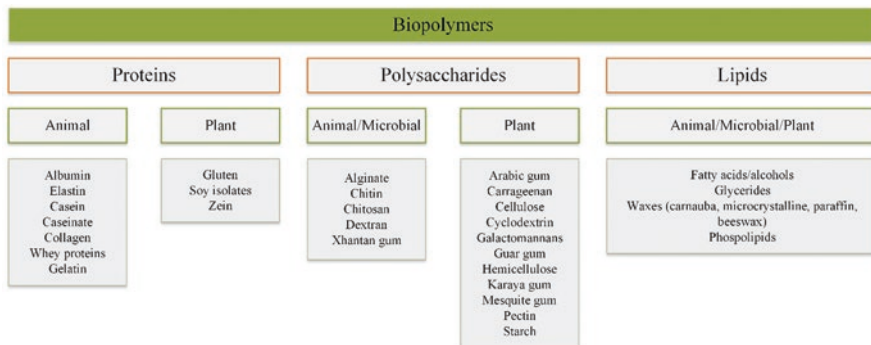


Fig. 13.1 Major natural biomolecules groups which are most often used in the preparation of nanosized carriers

shapes (films, spheres, irregular particles), or structures (compact or porous, amorphous or crystalline, rubbery or glassy) which depends mainly on the type of payload and application (Madene et al. 2006). Encapsulation in biopolymeric carrier systems is continuously developed and mostly advances to improve physicochemical, functional, and release properties while keeping in mind cost-effectiveness and the use of environmentally friendly material throughout the process (Lozano-Vazquez et al. 2015). The chosen material for the production depends on the purpose of encapsulation and final application. Generally, there are a couple of aims that are to be considered when choosing the right material for the encapsulation process: (i) the improvement of shelf life, (ii) type of encapsulation method, (iii) masking of taste or aroma, (iv) easier handling, (v) controlled and/or targeted release, (vi) improvement of appearance. Chosen materials for biopolymeric particle production are required to have several of the following requirements (Desai and Park 2005a). Furthermore, carriers should not react with a component which is to be encapsulated and should have good rheological properties and behavior even at very high concentration. Also, it should not release the encapsulated component during the storage or transport, provide maximal protection against environmental conditions, and should be completely solvent-free or from any other material used during encapsulation under drying or other desolvating conditions. Concerning the economical aspect, encapsulation material should be inexpensive, available in large quantities, and of constant quality (Desai and Park 2005b).

3 Modern Nanocarrier Systems

When considering the use of desired nanocarrier system, few basic points need to be considered. The main point is to take into the account type of used payload, consider its physicochemical stability, consider the overall sustainability of the production

process, and possible health risks (Fathi et al. 2021). There are carrier systems that can be considered to be used in agri-food sectors and these include:

- (i) *Nanofibers* are used due to their desirable properties for encapsulation of various payload materials. They are usually lightweight with small diameters and have controllable pore structures with a high surface-to-volume ratio (Vinceković et al. 2021).
- (ii) *Nanohydrogels* are nanosized networks of chemically/physically cross-linked polymers consisted of chains that are hydrophilic or amphiphilic. They are three-dimensional biocompatible materials with a large amount of water content. For delivery applications, few key properties are necessary: high water content/swellability, biocompatibility, and adjustable chemical/mechanical properties. Hydrogels can retain a large quantity of water or biological fluid without disturbing their basic polymeric chain structure. Hydrogels prepared from natural polymers have drawn huge attention due to their applications in pharmacy, agriculture, medicine, tissue engineering, cancer therapy, and drug delivery (Akram and Hussain 2017; Khoee and Asadi 2016).
- (iii) *Nanoemulsions* are kinetically stable liquid-in-liquid dispersions with droplet sizes in the range of 100 nm. They are characterized by high surface area per volume, robust stability, optically transparent appearance, and tunable rheology. Nanoemulsions are applicable in different areas from drug delivery, food production, agriculture, cosmetics, pharmaceuticals, to material synthesis (Gupta et al. 2016).
- (iv) *Nanostructured lipid carriers* are delivery systems composed of both solid and liquid lipids as core matrices. These types of carriers have advantages for drug therapy over conventional carriers. These include higher solubility, increased storage stability, better permeability, and bioavailability, decreased adverse effects, prolonged half-life, and tissue-targeted delivery (Nie et al. 2020).
- (v) *Bionanocrystals* are especially interesting due to their unique properties and have received considerable attention for the delivery of bioactive compounds. They are biocompatible, rigid, biodegradable, easy to modify, and are renewable (produced from food and agriculture waste) (Koshani and Madadlou 2018). For the food industry, especially are interesting starch, chitin, and cellulose nanocrystalline particles. They are considered promising contenders for the fabrication of reinforced, biodegradable carrier systems (Kasiri and Fathi 2018; Hao et al. 2018).

Despite many advantages to the nanocarrier systems, some problems are in the future to be overcome. These include (i) aggregation and adhesion of particles, (ii) special storage conditions and limited stability time of prepared formulations, (iii) difficulties in encapsulating some payloads of different hydrophilic properties in the same matrix, (iv) difficulties in regulating the polydispersity of particles (Vinceković et al. 2021).

To choose the optimal encapsulation method and the process of production of particles several things are needed to be considered. Mainly the type of material and encapsulated component because this is determinant when regarding pore size,

payload size, molecular weight, solubility in the carrier, the volumetric size of the carrier, and complex interactions between the payload and the carrier since this will govern the release mechanisms. Knowledge of this can significantly increase efficiency, loading capacity, and release properties which are the most important parameters (Panyam et al. 2004).

4 Encapsulation Technology in Agriculture – Present and Future

The encapsulation technology is widely used in different sectors from medicine, agriculture, food processing through the cosmetics and pharma industries. The scientific investigation in agricultural science technology and development in the last several years have concluded the huge necessity to set up a new type of microparticles (microspheres/microcapsules) as a delivery system of biological agents (fungi, bacteria, microalgae) and chemical agents (micro- and macronutrients, esters, peptides, amino acids, hormones, pesticides, etc.) (Vinceković et al. 2016; Jurić et al. 2020a, 2020b, 2020c; Slattery et al. 2019; Pereira et al. 2019; Rodríguez Nogales et al. 2020; Tsuji 2001).

One of the most important properties of the prepared microparticle formulations (microspheres/microcapsules) is the protection of active ingredients from external conditions and their decomposition and loss of activity in a particular environment. It can also improve their bioavailability and regulate the time release of ingredients over a longer period. Also, it brings the possibility of longer storage without loss of their activity. All setup properties depend on several important factors of encapsulation technology in agricultural application: (i) various types of wall materials and their concentration, (ii) encapsulation method/encapsulation process – microparticle production, (iii) pH and temperature, (iv) particle size (especially important for the method of application in agriculture (plant protection/nutrition)), (v) type and amount of encapsulated ingredients/additives and their interaction, as well as interaction with the carrier material. All these factors have a significant influence on the microparticle loading capacity, encapsulation efficiency, swelling degree, the strength of the membrane, and the type of release mechanism of bioactive components from microparticles (Li et al. 2019).

Present scientific investigation of microparticle formulations is focused on the preparation of a complex biopolymer-based network containing several bioactive components (synergistic effect). With this intensive research, new insights were gained connected to the complex processes and mechanisms of inter- and intramolecular interactions in biopolymer-based microparticles (microspheres/microcapsules). Inter- and intramolecular interactions are influencing the structural properties of microparticles loaded with active agents (biological and chemical) which in turn have an impact on their overall properties, especially on tuning their release mechanism from microparticle formulation in a specific environment. With this

knowledge, it is possible to prepare a new generation of microparticle formulations with the desired properties for different types of application in the agricultural production in the open field, greenhouse, hydroponics, or foliar approach of agroecological plant nutrition or protection and functional food production (higher level of plant metabolites) (Jurić et al. 2020a, 2020b, 2020c; Vinceković et al. 2019).

Besides the use of microparticles in agriculture, nanoparticles are also extensively utilized. Nanocapsules are vesicular systems consisted of a polymeric porous membrane that encapsulates an inner liquid core at the nanoscale. Some of the preferentially used nanoparticles are:

- Polymeric nanoparticles have superior biocompatibility and a minimal impact on non-targeted organisms. Polymeric types of nanomaterials are widely used in agriculture are polyethylene glycol, poly(epsilon-caprolactone), poly(lactide-co-glycolides), and poly(γ -glutamic acid) (Chand Mali et al. 2020; Clemente et al. 2014; Grillo et al. 2013; Ranganathan et al. 2019; Xu et al. 2013; Govender 1999).
- Silver nanoparticles are very effective against different phytopathogens (pesticide activity) with low toxicity and also in some cases, they are showing plant growth promotor properties. They are efficiently used for site-targeted delivery of important agrochemical products and diagnosis purpose tools in case of prior detection of plant diseases (Sadak 2019).
- Nano aluminosilicates, used as an effective pesticide (for different insects) in agriculture. They have very good properties: non-toxicity, biocompatibility, low costs, and environment-friendly nature (Singh et al. 2021; Mittal et al. 2020).
- Titanium dioxide nanoparticles (TiO_2) are one of the forms of titanium in the environment. TiO_2 nanoparticles are widely used for plant protection and environmental remediation because of their photoprotective and photocatalytic properties (Lyu et al. 2017).
- Carbon nanomaterials (graphene, graphene oxide, carbon dots, fullerenes, carbon nanotubes, fullerenes, carbon nanoparticles, and carbon nano-horns) have beneficial and stimulatory effects on plants *in vitro* or culture conditions. They are used to improve the seed germination process (Mukherjee et al. 2016; Husen and Siddiqi 2014).

Most nanotechnology products utilized in agriculture are used for plant protection and nutrition (nano herbicides, nano pesticides, nano fertilizers, and nanosystems for disease protection). All these systems explore the possible use of nanotechnology primarily in the process of controlled delivery of active ingredients that could be used as pesticides, herbicides, or fertilizers, but also secondary to improve the safety of the products which are applied in the process of plant protection and nutrition. Because of that, their group name is nanoagroparticles (Baker et al. 2017). In Table 13.2 we have presented examples of the application of colloidal delivery systems for essential oils in agriculture.

Viruses, fungi, and bacteria infections are causing huge economical losses in agricultural production. The preparation of nanomaterials enriched with certain components which are having specific antimicrobial properties against phytopathogenic fungi (*Colletotrichum gloeosporioides*, *Fusarium oxysporum*, *Fusarium*

Table 13.2 Examples of application of colloidal delivery systems for essential oils in agriculture (Adapted from Fathi et al. 2021)

Field of application	Essential oil delivery system	Preparation procedure	Claimed advantages
Organic farming	Carvacrol in alginate-whey protein biopolymeric particles	Emulsification and extrusion	Targeted release in chicken jejunum and ileum (Zhang et al. 2014)
Pest control	<i>Aegeratum conyzoides</i> , <i>Achillea fragrantissima</i> , and <i>Tagetes minuta</i> EOs nanoemulsions	High pressure homogenization	Higher toxicity against eggs and adults of beetle <i>Callosobruchus maculatus</i> than free oils (Nenaah et al. 2015)
	<i>Zanthoxylum rhoifolium</i> EO in biodegradable polycaprolactone nanospheres	Nanoprecipitation of the pre-formed polymer	Significantly higher reduction of <i>Bemisia tabaci</i> eggs and nymphs compared with control (Christofoli et al. 2015)
	<i>Carum copticum</i> EO in myristic acid-chitosan nanogels	Self-assembly	4–8-fold higher fumigant toxicity against <i>Sitophilus granarius</i> and <i>Tribolium confusum</i> than the free oil (Ziaee et al. 2014)
	Geranium and bergamot EOs in poly(ethylene glycol) nanoparticles	Self-assembly	Higher toxicity against <i>Tribolium castaneum</i> and <i>Rhizopertha dominica</i> than free oils (Werdin González et al. 2014)
	<i>Artemisia arborescens</i> EO in solid lipid nanoparticles	Hot high-pressure homogenization	Reduced volatility with respect to emulsions (Lai et al. 2006)
	<i>Lippia sidoides</i> EO in chitosan/cashew gum nanoparticles	Spray drying of nanoemulsion	Mortality rate of <i>St. aegypti</i> larvae correlates with EO loading (Abreu et al. 2012)
Pest luring	Geraniol in chitosan/gum arabic nanoparticles	Ionic gelation method	Improved EO stability and luring effect toward whitefly <i>Bemisia tabaci</i> (de Oliveira et al. 2018)
Repellant textile	Citronella EO in chitosan/gelatin microcapsules	Complex coacervation	Higher repellant effect and lasting protection from insects compared to textiles sprayed with EO in ethanol (Specos et al. 2010)

solani, *Dematophora necatrix*, etc.) can be used in the plant disease protection process. Prepared cobalt and nickel ferrite nanoparticles (CoFe_2O_4 and NiFe_2O_4) are successfully tested for antimycotic activity against three plant-pathogenic fungi: *Fusarium oxysporum*, *Colletotrichum gloeosporioides*, and *Dematophora necatrix* (Sharma et al. 2017). Copper nanoparticles with chitosan and celluloses showed antifungal and antibacterial properties against *Escherichia coli*, *Staphylococcus*

aureus, *Alternaria solani*, *Fusarium oxysporum*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Aspergillus niger*, *Aspergillus flavus*, and *Candida albicans*. Besides plant protection, copper nanoparticles are also used in plant nutrition (Rai et al. 2018). Copper is a micronutrient that can be found in high concentrations in chloroplasts. Almost 70% of the total Cu is found in chloroplasts. Cu has an important role in the process of synthesis of chlorophyll, other pigments and has a crucial role in the process of protein and carbohydrate metabolism (Mengel et al. 2001).

Despite the significant advantages of encapsulation technologies in the process of nano- and microparticle production, the preparation process still has several significant obstacles that must be addressed in the coming years to be able to achieve production in larger quantities:

- (a) insufficient number of methods used in the characterization of micro- and nanoparticle formulations,
- (b) the balance between biosafety and compatibility of wall materials,
- (c) various types of active ingredients release mechanisms,
- (d) stability during long-term storage at variable environmental conditions and temperatures.

Due to the abovementioned, it is necessary to conduct further research that will focus on increasing the stability of nano- and microcapsule formulations, control the uniformity of formulation sizes and release mechanisms of bioactive components in certain time intervals, testing their effectiveness on certain phytopathogenic fungi and bacteria and testing their action as new green formulations with 3 in 1 effects (plant protection, plant nutrition, and time-release mechanism). It can be concluded that from the above scientific research, technologies of encapsulation and production of nano- and microparticle formulations will more effectively promote the development in the agroecological agriculture and functional food production process.

5 Implementation of Encapsulated Material Into Final Food Products

There is a significant gap in the research with regards to the implementation of encapsulated natural pigments in real food products. Usually, with regards to this topic, research papers deal with bioactive compounds encapsulation procedures but the research on their inclusion in real food products is worryingly scarce and a couple of available examples are listed in Table 13.3.

It is important to observe the behavior of encapsulated bioactive compounds in food matrices and their influence on the sensory characteristics of food products. This would significantly advance the knowledge of ingredient behavior when considering implementation during food production.

Table 13.3 Examples of application of stabilized natural pigments in real food products (Adapted from Jurić et al. 2020a, 2020b, 2020c)

Encapsulated compounds	Compound donor	Stabilization method	Material	Functional food	References
Anthocyanin	Barberry (<i>Berberis vulgaris</i> L.)	Spray drying	Gum arabic, maltodextrin, gelatin	Jelly	Mahdavi et al. (2016)
	Black bean (<i>Phaseolus vulgaris</i> L.) coat	Molecular inclusion	β -cyclodextrin	Sport beverage	Aguilera et al. (2016)
	Grape (<i>Vitis vinifera</i> L.) skin	Spray drying	Maltodextrin	Apple puree	Lavelli et al. (2016)
	Sour cherry (<i>Prunus cerasus</i> L.) pomace extract	Freeze-drying	Whey and soy proteins	Cookies	Tumbas Šaponjac et al. (2016)
Betalains	Barbary fig (<i>Opuntia ficus-indica</i> L.)	Ionic gelation	Calcium alginate/Gelatin	Gummy candy	Otálora et al. (2019)
	Barbary fig (<i>Opuntia ficus-indica</i> L.)	Spray drying	Soluble fiber [(1–3) (1–4)- β -D-glucan	Extruded cereal	Ruiz-Gutiérrez et al. (2017)
	Beetroot (<i>Beta vulgaris</i> L.)	Freeze-drying	Maltodextrin	Chewing gum	Chranioti et al. (2015)
	Red beet (<i>Beta vulgaris</i> L.) extract diluted with dextrin	Thin-film hydration-sonication technique	Lecithin liposome	Gummy candy	Amjadi et al. (2018)
Carotenoids	Yellow bell pepper (<i>Capsicum annuum</i> L.)	Ultrasonic homogenization, kneading	β -cyclodextrin	Isotonic beverage	Lobo et al. (2018)
	Saffron (<i>Crocus sativus</i> L.)	Freeze-drying	Maltodextrin	Chewing gum	Chranioti et al. (2015)
Chlorophylls	Alfalfa (<i>Medicago sativa</i> L.)	Emulsification + Freeze-drying	Canola oil, glycerol monostearate, gelatin, agar	Gummy candy	Raei et al. (2017)

When considering the application of encapsulated ingredients into final food products the matrix can significantly affect its release behavior and particles physicochemical properties. Diffusion of ingredients may be affected by the presence of

proteins, carbohydrates, fatty acids, pH, water activity, packaging material, and trace metals. Furthermore, physicochemical stress factors like food processing, preservation, storage, food ingredients, etc. may start the degradation and collapse of the encapsulation system making it inefficient.

There are a couple of examples (Table 13.3) that have proven to have problems when the implementation of encapsulated bioactive compounds like anthocyanins into food products have negative effects on the shape, size, and uniformity of products (*e.g.* cookies, and other dough-based products). Furthermore, protein-based coatings (encapsulation systems) might induce significant changes in the structure of dough-based products (proteins absorb water resulting in increased hardness) while the presence of an additional particulate phase can increase the fragility of dough-based products (hindering the formation of a continuous starch network).

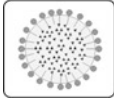

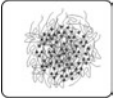


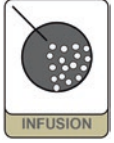
The main applications of encapsulated ingredients can be classified into four groups (Table 13.4). (1) Direct mixing with liquid foods or mixing with the food ingredients before food preparation; (2) washing the product surface with carrier systems in an aqueous dispersion; (3) infusion in porous food matrices; (4) coating with a biopolymeric layer incorporating the active ingredients delivery systems. Details about the strategies for the utilization of different colloidal systems for active ingredients like essential oils (EO) alongside the examples of application in food products are listed in Table 13.4 (Fathi et al. 2021).

Published research mainly deals with the fabrication procedures and the work on implementation into food products is scarce (even often contradicts the *in vitro* results). Thus it is important to understand the issues related to the application of encapsulated bioactive compounds (or other ingredients like microorganisms) into various food matrices. It is also necessary to investigate the behavior of carrier systems (*i.e.* protein-based) in complex food matrices alongside the influence on the sensory characteristics of final food products (Jurić et al. 2021).

6 Future Remarks

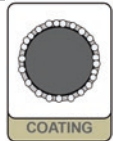
Encapsulation is becoming essential for the sustainable and economic development of various products, from agri-food to nutraceuticals. Respectively, it can resolve some problems regarding the stability of payload during industrial processing and storage. Even though still there is a significant gap when considering using this technology for a large scale production due to the limitations. Advancing the encapsulation methodology and technology these limitations could be minimized significantly. Advanced nanocarrier systems are becoming popular due to the low-cost materials necessary for their manufacturing especially when introduced to industrial-scale production.

Table 13.4 Strategies of the utilization of different colloidal systems for essential oils (EO) delivery, together with examples of application in food products (Adapted from Fathi et al. 2021)

	 Nanoemulsions	 Liposomes	 Biopolymeric nanoparticles
<i>Advantages</i>	<i>Active barrier with controlled release of antimicrobial, low amount needed (surface treatment), low impact on organoleptic properties</i>	<i>High efficiency of delivery through the biological membranes, additional loading of hydrophilic molecules</i>	<i>High food compatibility, several natural polysaccharides and proteins available</i>
<i>Disadvantages</i>	<i>Need for surfactant in formulation, high costs of nanoemulsion production</i>	<i>Limited loading of bioactives, high costs of phospholipids and of fabrication</i>	<i>Formation of particles in aqueous systems requires chemical/physical modification of hydrophilic polymers, or use of solvents for hydrophobic polymers</i>
 MIXING	Shelf life extension of milk and quality preservation by encapsulation of thyme EO (Ben Jemaa et al. 2017) Microbial stabilization of orange and pear juices by encapsulation of carvacrol (Donsi et al. 2011) Microbiological stabilization of chicken pâté by encapsulation of oregano EO (Moraes-Lovison et al. 2017)	Microbial stabilization of tofu by clove EO encapsulation (Cui et al. 2015)	Shelf life extension in bakery products by encapsulation of thyme EO by complex coacervation (Gonçalves et al. 2017)
 WASHING	Microbial stabilization of fresh lettuce by washing with oregano EO nanoemulsions (Bhargava et al. 2015) or of spinach leaves by carvacrol or eugenol nanoemulsions (Ruengvisesh et al. 2015)	–	–
 INFUSION	Enhancement of organoleptic properties and extension of the shelf life of trout fillets by infusion of nanoemulsions of rosemary, laurel and thyme EOs (Ozogul et al. 2017)	Preservation of minced beef by <i>Zataria multiflora</i> EO liposomes (Khosravi-Darani et al. 2016)	Reduction of lipid oxidation and microbial growth by infusion in meat patties of chitosan nanoparticles containing cinnamon EO (Ghaderi-Ghahfarokhi et al. 2017)

(continued)

Table 13.4 (continued)

	<p>Shelf life extension of rucola leaves coated with chitosan containing lemon EO nanoemulsions (Sessa et al. 2015)</p> <p>Microbial stabilization of green beans (Severino et al. 2014a, 2015) or broccoli florets (Severino et al. 2014b) by modified chitosan coatings containing citrus EO nanoemulsions</p> <p>Preservation of fresh-cut cheese by encapsulation of Oregano EO (Artiga-Artigas et al. 2017)</p> <p>Microbial stabilization of bread slice by an edible coating containing clove bud or oregano EO nanoemulsions (Otoni et al. 2014)</p>	<p>Shelf life extension and quality improvement of banana slices by a mucilage coating containing rosemary EO liposomes (Alikhani-Koupaei 2015)</p>	<p>Microbial stabilization of beef cutlet by spraying with a chitosan-myristic acid nanogels containing clove EO (Rajaei et al. 2017)</p> <p>Enhanced antimicrobial and antioxidant activity of cinnamon EO on pork by encapsulation in chitosan nanoparticles (Hu et al. 2015)</p>
---	--	---	---

References

- Abreu FOMS, Oliveira EF, Paula HCB, de Paula RCM (2012) Chitosan/cashew gum nanogels for essential oil encapsulation. *Carbohydr Polym* 89(4):1277–1282. <https://doi.org/10.1016/j.carbpol.2012.04.048>
- Aguilera Y, Mojica L, Rebollo-Hernanz M, Berhow M, De Mejía EG, Martín-Cabrejas MA (2016) Black bean coats: new source of anthocyanins stabilized by β -cyclodextrin copigmentation in a sport beverage. *Food Chem* 212(1):561–570. <https://doi.org/10.1016/j.foodchem.2016.06.022>
- Akram M, Hussain R (2017) Nanohydrogels: history, development, and applications in drug delivery. In: Jawaid M, Mohammad F (eds) *Nanocellulose and nanohydrogel matrices: biotechnological and biomedical applications*, Weinheim, Germany, pp 297–330. <https://doi.org/10.1002/9783527803835.ch11>
- Alikhani-Koupaei M (2015) Liposomal and edible coating as control release delivery systems for essential oils: comparison of application on storage life of fresh-cut banana. *Qual Assur Saf Crop Foods* 7(2):175–185. <https://doi.org/10.3920/QAS2013.0297>
- Amjadi S, Ghorbani M, Hamishehkar H, Roufegarinejad L (2018) Improvement in the stability of betanin by liposomal nanocarriers: its application in gummy candy as a food model. *Food Chem* 256(1):156–162. <https://doi.org/10.1016/j.foodchem.2018.02.114>
- Arroyo-Maya IJ, Campos-Terán J, Hernández-Arana A, McClements DJ (2016) Characterization of flavonoid-protein interactions using fluorescence spectroscopy: binding of pelargonidin to dairy proteins. *Food Chem* 213(1):431–439. <https://doi.org/10.1016/j.foodchem.2016.06.105>
- Artiga-Artigas M, Acevedo-Fani A, Martín-Belloso O (2017) Improving the shelf life of low-fat cut cheese using nanoemulsion-based edible coatings containing oregano essential oil and mandarin fiber. *Food Control* 76(1):1–12. <https://doi.org/10.1016/j.foodcont.2017.01.001>

- Baker S, Volova T, Prudnikova SV, Satish S, Prasad NMN (2017) Nanoagroparticles emerging trends and future prospect in modern agriculture system. *Environ Toxicol Pharmacol* 53(1):10–17. <https://doi.org/10.1016/j.etap.2017.04.012>
- Belščak-Cvitanović A, Bušić A, Barišić L, Vrsaljko D, Karlović S, Špoljarić I, Vojvodić A, Mršić G, Komes D (2016) Emulsion templated microencapsulation of dandelion (*Taraxacum officinale* L.) polyphenols and β -carotene by ionotropic gelation of alginate and pectin. *Food Hydrocoll* 57:139–152. <https://doi.org/10.1016/j.foodhyd.2016.01.020>
- Belščak-Cvitanović A, Jurić S, Đorđević V, Barišić L, Komes D, Ježek D, Bugarski B, Nedović V (2017) Chemometric evaluation of binary mixtures of alginate and polysaccharide biopolymers as carriers for microencapsulation of green tea polyphenols. *Int J Food Prop* 20(9):1971–1986. <https://doi.org/10.1080/10942912.2016.1225762>
- Ben Jemaa M, Falleh H, Neves MA, Isoda H, Nakajima M, Ksouri R (2017) Quality preservation of deliberately contaminated milk using thyme free and nanoemulsified essential oils. *Food Chem* 217(1):726–734. <https://doi.org/10.1016/j.foodchem.2016.09.030>
- Bhargava K, Conti DS, da Rocha SRP, Zhang Y (2015) Application of an oregano oil nanoemulsion to the control of foodborne bacteria on fresh lettuce. *Food Microbiol* 47(1):69–73. <https://doi.org/10.1016/j.fm.2014.11.007>
- Bonilla-Ahumada FDJ, Khandual S, Lugo-Cervantes EDC (2018) Microencapsulation of algal biomass (*Tetraselmis chuii*) by spray-drying using different encapsulation materials for better preservation of beta-carotene and antioxidant compounds. *Algal Res* 36(1):229–238. <https://doi.org/10.1016/j.algal.2018.10.006>
- Casanova F, Santos L (2016) Encapsulation of cosmetic active ingredients for topical application – a review. *J Microencapsul* 33(1):1–17. <https://doi.org/10.3109/02652048.2015.1115900>
- Chand Mali S, Raj S, Trivedi R (2020) Nanotechnology a novel approach to enhance crop productivity. *Biochem Biophys Rep* 24(1):e100821. <https://doi.org/10.1016/j.bbrep.2020.100821>
- Cheng YS, Lu PM, Huang CY, Wu JJ (2017) Encapsulation of lycopene with lecithin and α -tocopherol by supercritical antisolvent process for stability enhancement. *J Supercrit Fluids* 130(1):246–252. <https://doi.org/10.1016/j.supflu.2016.12.021>
- Chranioti C, Nikoloudaki A, Tzia C (2015) Saffron and beetroot extracts encapsulated in maltodextrin, gum Arabic, modified starch and chitosan: incorporation in a chewing gum system. *Carbohydr Polym* 127(1):252–263. <https://doi.org/10.1016/j.carbpol.2015.03.049>
- Christofoli M, Costa ECC, Bicalho KU, Domingues VC, Peixoto M, F., Alves, C. C. F., Araújo, W. L., Cazol, C. M. (2015) Insecticidal effect of nanoencapsulated essential oils from *Zanthoxylum rhoifolium* (Rutaceae) in *Bemisia tabaci* populations. *Ind Crop Prod* 70(1):301–308. <https://doi.org/10.1016/j.indcrop.2015.03.025>
- Chung C, Rojanasasithara T, Mutilangi W, McClements DJ (2015) Enhanced stability of anthocyanin-based color in model beverage systems through whey protein isolate complexation. *Food Res Int* 76(3):761–768. <https://doi.org/10.1016/j.foodres.2015.07.003>
- Chung C, Rojanasasithara T, Mutilangi W, McClements DJ (2016a) Enhancement of colour stability of anthocyanins in model beverages by gum arabic addition. *Food Chem* 201(1):14–22. <https://doi.org/10.1016/j.foodchem.2016.01.051>
- Chung C, Rojanasasithara T, Mutilangi W, McClements DJ (2016b) Stabilization of natural colors and nutraceuticals: inhibition of anthocyanin degradation in model beverages using polyphenols. *Food Chem* 212(1):596–603. <https://doi.org/10.1016/j.foodchem.2016.06.025>
- Chuyen HV, Roach PD, Golding JB, Parks SE, Nguyen MH (2019) Encapsulation of carotenoid-rich oil from Gac peel: optimisation of the encapsulating process using a spray drier and the storage stability of encapsulated powder. *Powder Technol* 344:373–379. <https://doi.org/10.1016/j.powtec.2018.12.012>
- Clemente Z, Grillo R, Jonsson M, Santos NZ, Feitosa LO, Lima R, Fraceto LF (2014) Ecotoxicological evaluation of poly(epsilon-caprolactone) nanocapsules containing triazine herbicides. *J Nanosci Nanotechnol* 14(7):4911–4917. <https://doi.org/10.1166/jnn.2014.8681>
- Cruz L, Benohoud M, Rayner CM, Mateus N, de Freitas V, Blackburn RS (2018) Selective enzymatic lipophilization of anthocyanin glucosides from blackcurrant (*Ribes nigrum* L.) skin

- extract and characterization of esterified anthocyanins. *Food Chem* 266(1):415–419. <https://doi.org/10.1016/j.foodchem.2018.06.024>
- Cruz L, Fernandes I, Guimarães M, De Freitas V, Mateus N (2016) Enzymatic synthesis, structural characterization and antioxidant capacity assessment of a new lipophilic malvidin-3-glucoside-oleic acid conjugate. *Food Funct* 7(6):2754–2762. <https://doi.org/10.1039/c6fo00466k>
- Cruz L, Fernandes VC, Araújo P, Mateus N, de Freitas V (2015) Synthesis, characterisation and antioxidant features of procyanidin B4 and malvidin-3-glucoside stearic acid derivatives. *Food Chem* 174:480–486. <https://doi.org/10.1016/j.foodchem.2014.11.062>
- Cruz L, Guimarães M, Araújo P, Évora A, De Freitas V, Mateus N (2017) Malvidin 3-glucoside-fatty acid conjugates: from hydrophilic toward novel lipophilic derivatives. *J Agric Food Chem* 65(31):6513–6518. <https://doi.org/10.1021/acs.jafc.6b05461>
- Cui H, Zhao C, Lin L (2015) The specific antibacterial activity of liposome-encapsulated Clove oil and its application in tofu. *Food Control* 56(1):128–134. <https://doi.org/10.1016/j.foodcont.2015.03.026>
- da Silva Carvalho AG, da Costa Machado MT, de Freitas Queiroz Barros HD, Cazarin CBB, Maróstica Junior MR, Hubinger MD (2019) Anthocyanins from jussara (*Euterpe edulis* Martius) extract carried by calcium alginate beads pre-prepared using ionic gelation. *Powder Technol* 345(1):283–291. <https://doi.org/10.1016/j.powtec.2019.01.016>
- de Moura SCSR, Berling CL, Germer SPM, Alvim ID, Hubinger MD (2018) Encapsulating anthocyanins from *Hibiscus sabdariffa* L. calyces by ionic gelation: pigment stability during storage of microparticles. *Food Chem* 241(1):317–327. <https://doi.org/10.1016/j.foodchem.2017.08.095>
- de Oliveira JL, Campos EVR, Pereira AES, Nunes LES, da Silva CCL, Pasquoto T, Lima R, Smaniotto G, Polaczyk RA, Fraceto LF (2018) Geraniol encapsulated in chitosan/gum Arabic nanoparticles: a promising system for pest management in sustainable agriculture. *J Agric Food Chem* 66(21):5325–5334. <https://doi.org/10.1021/acs.jafc.8b00331>
- Desai KGH, Park HJ (2005a) Recent developments in microencapsulation of food ingredients. *Dry Technol* 23(7):1361–1394. <https://doi.org/10.1081/DRT-200063478>
- Desai KGH, Park HJ (2005b) Encapsulation of Vitamin C in triphosphate cross-linked chitosan microspheres by spray drying. *J Microencapsul* 22(2):179–192. <https://doi.org/10.1080/02652040400026533>
- Donsì F, Annunziata M, Sessa M, Ferrari G (2011) Nanoencapsulation of essential oils to enhance their antimicrobial activity in foods. *LWT Food Sci Technol* 44(9):1908–1914. <https://doi.org/10.1016/j.lwt.2011.03.003>
- Fathi M, Vinceković M, Jurić S, Viskić M, Režek Jambrak A, Donsì F (2021) Food-grade colloidal systems for the delivery of essential oils. *Food Rev Int* 37(1):1–45. <https://doi.org/10.1080/087559129.2019.1687514>
- Fernandes A, Sousa A, Azevedo J, Mateus N, De Freitas V (2013) Effect of cyclodextrins on the thermodynamic and kinetic properties of cyanidin-3-O-glucoside. *Food Res Int* 51(2):748–755. <https://doi.org/10.1016/j.foodres.2013.01.037>
- Fu D, Deng S, McClements DJ, Zhou L, Zou L, Yi J, Liu C, Liu W (2019) Encapsulation of β -carotene in wheat gluten nanoparticle-xanthan gum-stabilized Pickering emulsions: enhancement of carotenoid stability and bioaccessibility. *Food Hydrocoll* 89(1):80–89. <https://doi.org/10.1016/j.foodhyd.2018.10.032>
- Ghaderi-Ghahfarokhi M, Barzegar M, Sahari MA, Ahmadi Gavlighi H, Gardini F (2017) Chitosan-cinnamon essential oil nano-formulation: as a novel additive for controlled release and shelf life extension of beef patties. *Int J Biol Macromol* 102(1):19–28. <https://doi.org/10.1016/j.ijbiomac.2017.04.002>
- Gonçalves ND, Pena F d L, Sartoratto A, Derlamelina C, Duarte MCT, Antunes AEC, Prata AP (2017) Encapsulated thyme (*Thymus vulgaris*) essential oil used as a natural preservative in bakery product. *Food Res Int* 96(1):154–160. <https://doi.org/10.1016/j.foodres.2017.03.006>
- Govender T, Stolnik S, Garnett MC, Illum L, Davis SS (1999) PLGA nanoparticles prepared by nanoprecipitation: drug loading and release studies of a water soluble drug. *J Control Release* 57(2):171–185. [https://doi.org/10.1016/s0168-3659\(98\)00116-3](https://doi.org/10.1016/s0168-3659(98)00116-3)

- Grillo R, Rosa AH, Fraceto LF (2013) Poly(ϵ -caprolactone) nanocapsules carrying the herbicide atrazine: effect of chitosan-coating agent on physico-chemical stability and herbicide release profile. *Int J Environ Sci Technol* 11(6):1691–1700. <https://doi.org/10.1007/s13762-013-0358-1>
- Guan Y, Zhong Q (2015) The improved thermal stability of anthocyanins at pH 5.0 by gum arabic. *LWT Food Sci Technol* 64(2):706–712. <https://doi.org/10.1016/j.lwt.2015.06.018>
- Gupta A, Eral HB, Hatton TA, Doyle PS (2016) Nanoemulsions: formation, properties and applications. *Soft Matter* 12(11):2826–2841. <https://doi.org/10.1039/c5sm02958a>
- Hao Y, Chen Y, Li Q, Gao Q (2018) Preparation of starch nanocrystals through enzymatic pretreatment from waxy potato starch. *Carbohydr Polym* 184(1):171–177. <https://doi.org/10.1016/j.carbpol.2017.12.042>
- He Z, Xu M, Zeng M, Qin F, Chen J (2016) Interactions of milk α - and β -casein with malvidin-3-O-glucoside and their effects on the stability of grape skin anthocyanin extracts. *Food Chem* 199(1):314–322. <https://doi.org/10.1016/j.foodchem.2015.12.035>
- Howard LR, Brownmiller C, Prior RL, Mauromoustakos A (2013) Improved stability of chokeberry juice anthocyanins by β -cyclodextrin addition and refrigeration. *J Agric Food Chem* 61(3):693–699. <https://doi.org/10.1021/jf3038314>
- Hsiao C-J, Lin J-F, Wen H-Y, Lin Y-M, Yang C-H, Huang K-S, Shaw J-F (2020) Enhancement of the stability of chlorophyll using chlorophyll-encapsulated polycaprolactone microparticles based on droplet microfluidics. *Food Chem* 306(1):e125300. <https://doi.org/10.1016/j.foodchem.2019.125300>
- Hu J, Wang X, Xiao Z, Bi W (2015) Effect of chitosan nanoparticles loaded with cinnamon essential oil on the quality of chilled pork. *LWT Food Sci Technol* 63(1):519–526. <https://doi.org/10.1016/j.lwt.2015.03.049>
- Husen A, Siddiqi KS (2014) Carbon and fullerene nanomaterials in plant system. *J Nanobiotechnol* 12(1):e16. <https://doi.org/10.1186/1477-3155-12-16>
- Jana P, Shyam M, Singh S, Jayaprakash V, Dev A (2020) Biodegradable polymers in drug delivery and oral vaccination. *Eur Polym J* 142(1):e110155. <https://doi.org/10.1016/j.eurpolymj.2020.110155>
- Jurić S, Đermić E, Topolovec-Pintarić S, Bedek M, Vinceković M (2019) Physicochemical properties and release characteristics of calcium alginate microspheres loaded with *Trichoderma viride* spores. *J Integr Agric* 18(11):2534–2548. [https://doi.org/10.1016/S2095-3119\(19\)62634-1](https://doi.org/10.1016/S2095-3119(19)62634-1)
- Jurić S, Jurić M, Król-Kilińska Ž, Vlahoviček-Kahlina K, Vinceković M, Dragović-Uzelac V, Donsi F (2020a) Sources, stability, encapsulation and application of natural pigments in foods. *Food Rev Int*, in press. <https://doi.org/10.1080/87559129.2020.1837862>
- Jurić S, Jurić M, Siddique MAB, Fathi M (2020b) Vegetable oils rich in polyunsaturated fatty acids: nanoencapsulation methods and stability enhancement. *Food Rev Int*, in press. <https://doi.org/10.1080/87559129.2020.1717524>
- Jurić S, Sopko K, Król-Kilinska Ž, Žutić I, Fabek Uher S, Đermić E, Topolovec-Pintarić S, Vinceković M (2020c) The enhancement of plant secondary metabolites contents in *Lactuca sativa* L. by encapsulated bioactive agents. *Sci Rep* 10(1):e3737. <https://doi.org/10.1038/s41598-020-60690-3>
- Jurić S, Tanuwidjaja I, Mrkonjić Fuka M, Vlahoviček-Kahlina K, Marijan M, Boras A, Udiković Kolić N, Vinceković M (2021) Encapsulation of two fermentation agents, *Lactobacillus sakei* and calcium ions in microspheres. *Colloids Surf B Biointerfaces* 197(1):e111387. <https://doi.org/10.1016/j.colsurfb.2020.111387>
- Kaimainen M, Marze S, Järvenpää E, Anton M, Huopalahti R (2015) Encapsulation of betalain into w/o/w double emulsion and release during invitro intestinal lipid digestion. *LWT Food Sci Technol* 60(2):899–904. <https://doi.org/10.1016/j.lwt.2014.10.016>
- Kang YR, Lee YK, Kim YJ, Chang YH (2019) Characterization and storage stability of chlorophylls microencapsulated in different combination of gum Arabic and maltodextrin. *Food Chem* 272(1):337–346. <https://doi.org/10.1016/j.foodchem.2018.08.063>

- Karangutkar AV, Ananthanarayan L (2020) Co-crystallization of *Basella rubra* extract with sucrose: characterization of co-crystals and evaluating the storage stability of betacyanin pigments. *J Food Eng* 271(1):e 109776. <https://doi.org/10.1016/j.jfoodeng.2019.109776>
- Kasiri N, Fathi M (2018) Production of cellulose nanocrystals from pistachio shells and their application for stabilizing pickering emulsions. *Int J Biol Macromol* 106(1):1023–1031. <https://doi.org/10.1016/j.ijbiomac.2017.08.112>
- Khoee S, Asadi H (2016) Nanogels: chemical approaches to preparation. In: Mishra MK (ed) *Encyclopedia of biomedical polymers and polymeric biomaterials*. Taylor and Francis, New York, NY, pp 5266–5293. <https://doi.org/10.1201/b19038-60>
- Khosravi-Darani K, Khoosfi ME, Hosseini H (2016) Encapsulation of *Zataria multiflora* Boiss. Essential oil in liposome: antibacterial activity against *E. Coli* O157:H7 in broth media and minced beef. *J Food Saf* 36(4):515–523. <https://doi.org/10.1111/jfs.12271>
- Kohno Y, Kinoshita R, Ikoma S, Yoda K, Shibata M, Matsushima R, Tomita Y, Maeda Y, Kobayashi K (2009) Stabilization of natural anthocyanin by intercalation into montmorillonite. *Appl Clay Sci* 42(3–4):519–523. <https://doi.org/10.1016/j.clay.2008.06.012>
- Koshani R, Madadlou AA (2018) A viewpoint on the gastrointestinal fate of cellulose nanocrystals. *Trends Food Sci Technol* 71(1):268–273. <https://doi.org/10.1016/j.tifs.2017.10.023>
- Lai F, Wissing SA, Müller RH, Fadda AM (2006) *Artemisia arborescens* L essential oil-loaded solid lipid nanoparticles for potential agricultural application: preparation and characterization. *AAPS PharmSciTech* 7(1):10–18. <https://doi.org/10.1208/pt070102>
- Lavelli V, Sri Harsha PSC, Spigno G (2016) Modelling the stability of maltodextrin-encapsulated grape skin phenolics used as a new ingredient in apple puree. *Food Chem* 209(1):323–331. <https://doi.org/10.1016/j.foodchem.2016.04.055>
- Lengyel M, Kállai-Szabó N, Antal V, Laki AJ, Antal I (2019) Microparticles, microspheres, and microcapsules for advanced drug delivery. *Sci Pharm* 87(3):e20. <https://doi.org/10.3390/scipharm87030020>
- Li T, Teng D, Mao R, Hao Y, Wang X, Wang J (2019) Recent progress in preparation and agricultural application of microcapsules. *J Biomed Mater Res* 107(10):2371–2385. <https://doi.org/10.1002/jbm.a.36739>
- Lin Z, Fischer J, Wicker L (2016) Intermolecular binding of blueberry pectin-rich fractions and anthocyanin. *Food Chem* 194(1):986–993. <https://doi.org/10.1016/j.foodchem.2015.08.113>
- Lobo FATF, Silva V, Domingues J, Rodrigues S, Costa V, Falcão D, de Lima Araújo KG (2018) Inclusion complexes of yellow bell pepper pigments with β -cyclodextrin: preparation, characterisation and application as food natural colorant. *J Sci Food Agric* 98(1):2665–2671. <https://doi.org/10.1002/jsfa.8760>
- Lozano-Vazquez G, Lobato-Calleros C, Escalona-Buendia H, Chavez G, Alvarez-Ramirez J, Vernon-Carter EJ (2015) Effect of the weight ratio of alginate-modified tapioca starch on the physicochemical properties and release kinetics of chlorogenic acid containing beads. *Food Hydrocoll* 48(1):301–311. <https://doi.org/10.1016/j.foodhyd.2015.02.032>
- Luo S-Z, Chen S-S, Pan L-H, Qin X-S, Zheng Z, Zhao Y-Y, Pang M, Jiang S-T (2017) Antioxidative capacity of crude camellia seed oil: impact of lipophilization products of blueberry anthocyanin. *Int J Food Prop* 20(2):1627–1636. <https://doi.org/10.1080/10942912.2017.1350974>
- Lyu S, Wei X, Chen J, Wang C, Wang X, Pan D (2017) Titanium as a beneficial element for crop production. *Front Plant Sci* 8(1):e597. <https://doi.org/10.3389/fpls.2017.00597>
- Madene A, Jacquot M, Scher J, Desobry S (2006) Flavour encapsulation and controlled release – a review. *Int J Food Sci Technol* 41(1):1–21. <https://doi.org/10.1111/j.1365-2621.2005.00980.x>
- Mahdavee Khazaei K, Jafari SM, Ghorbani M, Hemmati Kakhki A (2014) Application of maltodextrin and gum Arabic in microencapsulation of saffron Petal's anthocyanins and evaluating their storage stability and color. *Carbohydr Polym* 105(1):57–62. <https://doi.org/10.1016/j.carbpol.2014.01.042>
- Mahdavi SA, Jafari SM, Assadpour E, Ghorbani M (2016) Storage stability of encapsulated Barberry's anthocyanin and its application in jelly formulation. *J Food Eng* 181(1):59–66. <https://doi.org/10.1016/j.jfoodeng.2016.03.003>

- Mengel K, Kirkby EA, Kosegarten H, Appel T (2001) Soil copper. In: Principles of plant nutrition. Springer, Dordrecht, pp 599–611
- Mittal D, Kaur G, Singh P, Yadav K, Ali SA (2020) Nanoparticle-based sustainable agriculture and food science: recent advances and future outlook. *Front Nanotechnol* 2(1):e579954. <https://doi.org/10.3389/fnano.2020.579954>
- Moraes-Lovison M, Marostegan LFP, Peres MS, Menezes IF, Ghiraldi M, Rodrigues RAF, Fernandes AM, Pinho SC (2017) Nanoemulsions encapsulating oregano essential oil: production, stability, antibacterial activity and incorporation in chicken pâté. *LWT Food Sci Technol* 77(1):233–240. <https://doi.org/10.1016/j.lwt.2016.11.061>
- Mrkonjić Fuku M, Žgomba Maksimović A, Hulak N, Kos I, Marušić Radovčić N, Jurić S, Tanuwidjaja I, Karolyi D, Vinceković M (2021) The survival rate and efficiency of nonencapsulated and encapsulated native starter cultures to improve the quality of artisanal game meat sausages. *J Food Sci Technol* 58(1):710–719. <https://doi.org/10.1007/s13197-020-04587-z>
- Mukherjee A, Majumdar S, Servin AD, Pagano L, Dhankher OP, White JC (2016) Carbon nano-materials in agriculture: a critical review. *Front Plant Sci* 7(1):e172. <https://doi.org/10.3389/fpls.2016.00172>
- Nedović V, Kalusević A, Manojlović V, Lević S, Bugarski B (2011) An overview of encapsulation technologies for food applications. *Proced Food Sci* 1(1):1806–1815. <https://doi.org/10.1016/j.profoo.2011.09.265>
- Nenaah GE, Ibrahim SIA, Al-Assiuty BA (2015) Chemical composition, insecticidal activity and persistence of three Asteraceae essential oils and their nanoemulsions against *Callosobruchus maculatus* (F.). *J Stored Prod Res* 61(1):9–16. <https://doi.org/10.1016/j.jspr.2014.12.007>
- Nie X, Chen Z, Pang L, Wang L, Jiang H, Chen Y et al (2020) Oral nano drug delivery systems for the treatment of type 2 diabetes mellitus: an available administration strategy for anti-diabetic phytochemicals. *Int J Nanomedicine* 15(1):10215–10240. <https://doi.org/10.2147/ijn.s285134>
- Oidtmann J, Schantz M, Mäder K, Baum M, Berg S, Betz M, Kulozik U, Leick S, Rehage H, Schwarz K, Richling E (2012) Preparation and comparative release characteristics of three anthocyanin encapsulation systems. *J Agric Food Chem* 60(3):844–851. <https://doi.org/10.1021/jf2047515>
- Otálora MC, Carriazo JG, Iturriaga L, Osorio C, Nazareno MA (2016) Encapsulating betalains from *Opuntia ficus-indica* fruits by ionic gelation: pigment chemical stability during storage of beads. *Food Chem* 202(1):373–382. <https://doi.org/10.1016/j.foodchem.2016.01.115>
- Otálora MC, de Jesús Barbosa H, Perilla JE, Osorio C, Nazareno MA (2019) Encapsulated Betalains (*Opuntia Ficus-indica*) as natural colorants. Case study: gummy candies. *LWT Food Sci Technol* 103(1):222–227. <https://doi.org/10.1016/j.lwt.2018.12.074>
- Otoni CG, Pontes SFO, Medeiros EAA, Soares NDFF (2014) Edible films from methylcellulose and nanoemulsions of clove bud (*Syzygium aromaticum*) and oregano (*Origanum vulgare*) essential oils as shelf life extenders for sliced bread. *J Agric Food Chem* 62(22):5214–5219. <https://doi.org/10.1021/jf501055f>
- Ozogul Y, Yuvka İ, Ucar Y, Durmus M, Kösker AR, Öz M, Ozogul F (2017) Evaluation of effects of nanoemulsion based on herb essential oils (rosemary, laurel, thyme and sage) on sensory, chemical and microbiological quality of rainbow trout (*Oncorhynchus mykiss*) fillets during ice storage. *LWT Food Sci Technol* 75(1):677–684. <https://doi.org/10.1016/j.lwt.2016.10.009>
- Panyam J, Williams D, Dash A, Leslie-Pelecky D, Labhasetwar V (2004) Solid-state solubility influences encapsulation and release of hydrophobic drugs from PLGA/PLA nanoparticles. *J Pharm Sci* 93(7):1804–1814. <https://doi.org/10.1002/jps.20094>
- Pereira ADES, Oliveira HC, Fraceto LF (2019) Polymeric nanoparticles as an alternative for application of gibberellic acid in sustainable agriculture: a field study. *Sci Rep* 9(1):e7135. <https://doi.org/10.1038/s41598-019-43494-y>
- Qv X-Y, Zeng Z-P, Jiang J-G (2011) Preparation of lutein microencapsulation by complex coacervation method and its physicochemical properties and stability. *Food Hydrocoll* 25(6):1596–1603. <https://doi.org/10.1016/j.foodhyd.2011.01.006>

- Raei A, Yasini Ardakani SA, Daneshi M (2017) Microencapsulation of the green pigment of alfalfa and its applications on heated food. *J Food Process Eng* 40(5):e12529. <https://doi.org/10.1111/jfpe.12529>
- Rai M, Ingle AP, Pandit R, Paralikar P, Shende S, Gupta I, Biswas JK, da Silva SS (2018) Copper and copper nanoparticles: role in management of insect-pests and pathogenic microbes. *Nanotechnol Rev* 7(4):303–315. <https://doi.org/10.1515/ntrev-2018-0031>
- Rajaei A, Hadian M, Mohsenifar A, Rahmani-Cherati T, Tabatabaei M (2017) A coating based on clove essential oils encapsulated by chitosan-myristic acid nanogel efficiently enhanced the shelf-life of beef cutlet. *Food Packag Shelf Life* 14(1):137–145. <https://doi.org/10.1016/j.fpsl.2017.10.005>
- Ranganathan A, Manabe Y, Sugawara T, Hirata T, Shivanna N, Baskaran V (2019) Poly (D, L-lactide-co-glycolide)-phospholipid nanocarrier for efficient delivery of macular pigment lutein: absorption pharmacokinetics in mice and antiproliferative effect in Hep G2 cells. *Drug Deliv Transl Res* 9(1):178–191. <https://doi.org/10.1007/s13346-018-0590-9>
- Ravanfar R, Comunian TA, Abbaspourrad A (2018) Thermoresponsive, water-dispersible microcapsules with a lipid-polysaccharide shell to protect heat-sensitive colorants. *Food Hydrocoll* 81(1):419–428. <https://doi.org/10.1016/j.foodhyd.2018.03.030>
- Rodríguez Nogales JM, Simo G, Pérez Magarino S, Cano Mozo E, Fernández Fernández E, Ruiperez V, Vila Crespo J (2020) Evaluating the influence of simultaneous inoculation of SiO₂-alginate encapsulated bacteria and yeasts on volatiles, amino acids, biogenic amines and sensory profile of red wine with lysozyme addition. *Food Chem* 327(1):e126920. <https://doi.org/10.1016/j.foodchem.2020.126920>
- Ruengvisesh S, Loquercio A, Castell-Perez E, Taylor TM (2015) Inhibition of bacterial pathogens in medium and on spinach leaf surfaces using plant-derived antimicrobials loaded in surfactant micelles. *J Food Sci* 80(11):2522–2529. <https://doi.org/10.1111/1750-3841.13085>
- Ruiz-Gutiérrez MG, Amaya-Guerra CA, Quintero-Ramos A, Pérez-Carrillo E, Meléndez-Pizarro CO (2017) Use of red cactus pear (*Opuntia Ficus-indica*) encapsulated powder to pigment extruded cereal. *J Food Qual* 1:1–12. <https://doi.org/10.1155/2017/7262464>
- Sadak MS (2019) Impact of silver nanoparticles on plant growth, some biochemical aspects, and yield of fenugreek plant (*Trigonella foenum-graecum*). *Bull Natl Res Cent* 43(1):e38. <https://doi.org/10.1186/s42269-019-0077-y>
- Santos DT, Albarelli JQ, Beppu MM, Meireles MAA (2013) Stabilization of anthocyanin extract from jaboticaba skins by encapsulation using supercritical CO₂ as solvent. *Food Res Int* 50(2):617–624. <https://doi.org/10.1016/j.foodres.2011.04.019>
- Sessa M, Ferrari G, Donsì F (2015) Novel edible coating containing essential oil nanoemulsions to prolong the shelf life of vegetable products. *Chem Eng Trans* 43(1):55–60. <https://doi.org/10.3303/CET1543010>
- Severino R, Ferrari G, Vu KD, Donsì F, Salmieri S, Lacroix M (2015) Antimicrobial effects of modified chitosan based coating containing nanoemulsion of essential oils, modified atmosphere packaging and gamma irradiation against *Escherichia coli* O157:H7 and *Salmonella Typhimurium* on green beans. *Food Control* 50(1):215–222. <https://doi.org/10.1016/j.foodcont.2014.08.029>
- Severino R, Vu KD, Donsì F, Salmieri S, Ferrari G, Lacroix M (2014a) Antibacterial and physical effects of modified chitosan based-coating containing nanoemulsion of mandarin essential oil and three non-thermal treatments against *Listeria innocua* in green beans. *Int J Food Microbiol* 191(1):82–88. <https://doi.org/10.1016/j.ijfoodmicro.2014.09.007>
- Severino R, Vu KD, Donsì F, Salmieri S, Ferrari G, Lacroix M (2014b) Antimicrobial effects of different combined non-thermal treatments against *Listeria monocytogenes* in broccoli florets. *J Food Eng* 124(1):1–10. <https://doi.org/10.1016/j.jfoodeng.2013.09.026>
- Sharma P, Sharma A, Sharma M, Bhalla N, Estrela P, Jain A, Thakur A (2017) Nanomaterial fungicides: *In Vitro* and *In Vivo* antimycotic activity of cobalt and nickel nanoferrites on phytopathogenic fungi. *Glob Chall* 1(9):e1700041. <https://doi.org/10.1002/gch2.201700041>

- Shu B, Yu W, Zhao Y, Liu X (2006) Study on microencapsulation of lycopene by spray-drying. *J Food Eng* 76(4):664–669. <https://doi.org/10.1016/j.jfoodeng.2005.05.062>
- Singh H, Sharma A, Bhardwaj SK, Arya SK, Bhardwaj N, Khatri M (2021) Recent advances in the applications of nano-agrochemicals for sustainable agricultural development. *Environ Sci Process Impacts* 23(2):213–239. <https://doi.org/10.1039/D0EM00404A>
- Slattery M, Harper B, Harper S (2019) Pesticide encapsulation at the nanoscale drives changes to the hydrophobic partitioning and toxicity of an active ingredient. *Nano* 9(1):e81. <https://doi.org/10.3390/nano9010081>
- Specos MM, García JJ, Tornesello J, Marino P, Vecchia MD, Tesoriero MV, Hermida LG (2010) Microencapsulated citronella oil for mosquito repellent finishing of cotton textiles. *Trans R Soc Trop Med Hyg* 104(10):653–658. <https://doi.org/10.1016/j.trstmh.2010.06.004>
- Tachibana N, Kimura Y, Ohno T (2014) Examination of molecular mechanism for the enhanced thermal stability of anthocyanins by metal cations and polysaccharides. *Food Chem* 143(1):452–458. <https://doi.org/10.1016/j.foodchem.2013.08.017>
- Tsuji K (2001) Microencapsulation of pesticides and their improved handling safety. *J Microencapsul* 18(2):137–147. <https://doi.org/10.1080/026520401750063856>
- Tumbas Šaponjac V, Četković G, Čanadanović-Brunet J, Pajin B, Djilas S, Petrović J, Lončarević I, Stajčić S, Vulić J (2016) Sour cherry pomace extract encapsulated in whey and soy proteins: incorporation in cookies. *Food Chem* 207(1):27–33. <https://doi.org/10.1016/j.foodchem.2016.03.082>
- Vinceković M, Jalšenjak N, Topolovec-Pintarić S, Đermić E, Bujan M, Jurić S (2016) Encapsulation of biological and chemical agents for plant nutrition and protection: chitosan/alginate microcapsules loaded with copper cations and *Trichoderma viride*. *J Agric Food Chem* 64(43):8073–8083. <https://doi.org/10.1021/acs.jafc.6b02879>
- Vinceković M, Jurić S, Marijan M, Viskić M, Vlahoviček-Kahlina K, Maslov Bandić L (2021) Encapsulation of herb extracts (aromatic and medicinal herbs). In: Galanakis CM (ed) *Aromatic herbs in food: bioactive compounds, processing, and applications*. Academic Press (Elsevier), Cambridge, MA, pp 263–322. <https://doi.org/10.1016/B978-0-12-822716-9.00008-1>
- Vinceković M, Maslov Bandić L, Jurić S, Jalšenjak N, Čaić A, Živičnjak I, Đermić E, Karoglan M, Osrečak M, Topolovec-Pintarić S (2019) The Enhancement of bioactive potential in *Vitis vinifera* leaves by application of microspheres loaded with biological and chemical agents. *J Plant Nutr* 42(6):543–558. <https://doi.org/10.1080/01904167.2019.1568467>
- Vos P, Faas MM, Spasojević M, Sikkema J (2010) Review: encapsulation for preservation of functionality and targeted delivery of bioactive food components. *Int Dairy J* 20(4):292–302. <https://doi.org/10.1016/j.idairyj.2009.11.008>
- Werdin González JO, Gutiérrez MM, Ferrero AA, Fernández Band B (2014) Essential oils nano-formulations for stored-product pest control – characterization and biological properties. *Chemosphere* 100(1):130–138. <https://doi.org/10.1016/j.chemosphere.2013.11.056>
- Xu Z, Wan C, Xu X, Feng X, Xu H (2013) Effect of poly (γ -glutamic acid) on wheat productivity, nitrogen use efficiency and soil microbes. *J Soil Sci Plant Nutr* 13(3):744–755. <https://doi.org/10.4067/S0718-95162013005000059>
- Yang W, Kortensniemi M, Ma X, Zheng J, Yang B (2019) Enzymatic acylation of blackcurrant (*Ribes nigrum*) anthocyanins and evaluation of lipophilic properties and antioxidant capacity of derivatives. *Food Chem* 281(1):189–196. <https://doi.org/10.1016/j.foodchem.2018.12.111>
- Zhang Y, Gong J, Yu H, Guo Q, Defelice C, Hernandez M, Yin Y, Wang Q (2014) Alginate-whey protein dry powder optimized for target delivery of essential oils to the intestine of chickens. *Poult Sci* 93(10):2514–2525. <https://doi.org/10.3382/ps.2013-03843>
- Zhang Z-H, Peng H, Woo MW, Zeng X-A, Brennan M, Brennan CS (2020) Preparation and characterization of whey protein isolate-chlorophyll microcapsules by spray drying: effect of WPI ratios on the physicochemical and antioxidant properties. *J Food Eng* 267(1):e109729. <https://doi.org/10.1016/j.jfoodeng.2019.109729>

- Zhao L, Temelli F, Chen L (2017) Encapsulation of anthocyanin in liposomes using supercritical carbon dioxide: effects of anthocyanin and sterol concentrations. *J Funct Foods* 34(1):159–167. <https://doi.org/10.1016/j.jff.2017.04.021>
- Ziaee M, Moharramipour S, Mohsenifar A (2014) Toxicity of Carum copticum essential oil-loaded nanogel against *Sitophilus granarius* and *Tribolium confusum*. *J Appl Entomol* 138(10):763–771. <https://doi.org/10.1111/jen.12133>