Pituitary Adenomas

The European Neuroendocrine Association's Young Researcher Committee Overview

Gianluca Tamagno Manuel D. Gahete *Editors*



Pituitary Adenomas

Gianluca Tamagno • Manuel D. Gahete Editors

Pituitary Adenomas

The European Neuroendocrine Association's Young Researcher Committee Overview





Editors Gianluca Tamagno Hermitage Medical Clinic Dublin, Ireland

Manuel D. Gahete Cell Biology, Physiology and Immunology University of Córdoba Cordoba, Spain

ISBN 978-3-030-90474-6 ISBN 978-3-030-90475-3 (eBook) https://doi.org/10.1007/978-3-030-90475-3

© Springer Nature Switzerland AG 2022

This work is subject to copyright. All rights are reserved by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Switzerland AG The registered company address is: Gewerbestrasse 11, 6330 Cham, Switzerland

Preface

Pituitary is a fascinating gland for endocrinologists, being the main center of the regulation of the rest of endocrine glands, in synergy with the hypothalamus. The bulk of scientific knowledge regarding pituitary physiology and dysfunctions, or diseases, increases year after year due to the efforts implemented by numerous clinical, translational, and basic researchers all over the world. Among the conditions possibly impacting on pituitary function, pituitary adenomas represent the largest group and the most frequent clinical scenario faced in pituitary clinics.

Considering the continuous changes and novelties arising in our understanding of the function of this master gland and the diseases that may affect that, together with the progressive optimization of the clinical care of the patients on the basis of the evolving scientific evidence and with the support of the international guidelines, we decided to build up a comprehensive and exhaustive overview of pituitary adenomas. As members of the European Neuroendocrine Association Young Researchers Committee (EYRC), we envisioned the delivery of a book focusing on the most relevant aspects of pituitary adenomas for the young endocrinologists, especially those who are active in the field of neuroendocrinology either from a clinical or a research point of view. With the priceless commitment of the authors of the chapters and the expert reviewers and endorsers of the same, all of them very well-known physicians and researchers in the field of neuroendocrinology, we have generated a book aiming to support the education and the scientific development of the young colleagues and pointing toward to help them in the management of their first pituitary adenoma patients with a straightforward and exhaustive collection of evidence-based data and expert advices.

Young physicians and researchers from worldwide scientific groups renowned in the field of pituitary diseases have greatly contributed as authors to the creation of this book. In other words, this work has been mostly done by young physicians and researchers for the benefit of other young physicians and researchers. The basic and translational aspects have been integrated with the clinical aspects arising from everyday practice, with the goal of representing a useful companion for every earlystage endocrinologist. Illustrative clinical scenarios and a "questions & answers" section have been included in order to translate the theoretical knowledge into the practice and for giving the readers the possibility of reviewing their clinical and research approaches. The combined enthusiasm and expertise of authors, reviewers, and editors, on behalf of EYRC, has finally shaped an original monothematic book on pituitary adenomas, which is at the same time solid and innovative, with an appropriate balance between the traditional knowledge and the most exciting innovations coming from recent or ongoing research.

Dublin, Ireland Cordoba, Spain Gianluca Tamagno Manuel D. Gahete

Contents

1	Anatomy of the Pituitary Gland1Nicolas Coronel-Restrepo, Luis V. Syro, Fabio Rotondo,1and Kalman Kovacs1
2	Physiology of the Pituitary Hormone Secretion21Antonio C. Fuentes-Fayos, Emilia Alors-Perez,21Juan M. Jiménez-Vacas, Vicente Herrero-Aguayo,21Prudencio Sáez-Martínez, Juan L. Lopez-Cánovas,21María C. Vázquez-Borrego, Justo P. Castaño, Rhonda D. Kineman,21Manuel D. Gahete, and Raúl M. Luque21
3	Pathogenesis of Pituitary Adenomas57Sicheng Tang, Adriana Albani, and Marily Theodoropoulou57
4	Genetics of Pituitary Adenomas
5	Acromegaly
6	Prolactinoma
7	Cushing's Disease
8	TSH-oma
9	Nonfunctioning Pituitary Adenoma
10	Clinical Case 1

11	Clinical Case 2
12	Clinical Case 3
13	Clinical Case 4
14	Questions and Answers. 325 Gianluca Tamagno and Manuel D. Gahete

Contributors

Nerea Aguirre-Moreno Department of Endocrinology, Hospital Universitario de la Princesa, Instituto de Investigación Princesa, Universidad Autónoma de Madrid. C/ Diego de León 62, Madrid, Spain

Adriana Albani Medizinische Klinik und Poliklinik IV, Ludwig Maximilian University Munich, Munich, Germany

Emilia Alors-Perez Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Maria Rosaria Ambrosio Section of Endocrinology and Internal Medicine, Department of Medical Sciences, University of Ferrara, Ferrara, Italy

Anna Aulinas Endocrinology/Medicine Department, Hospital de la Santa Creu i Sant Pau. Centro Investigación Biomédica en Red de Enfermedades Raras (CIBERER, Unidad 747), IIB-Sant Pau, ISCIII, Barcelona, Spain

Faculty of Medicine, University of Vic (UVIC/UCC), Vic, Barcelona, Spain

Renata S. Auriemma Dipartimento di Medicina Clinica e Chirurgia, Sezione di Endocrinologia, University Federico II of Naples, Naples, Italy

Anna Bogusławska Department of Endocrinology, Endocrine Oncology and Nuclear Medicine, Jagiellonian University Medical College, Cracow, Poland

Justo P. Castaño Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

R. Catalano Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

Philippe Chanson Université Paris-Saclay, Inserm, Physiologie et Physiopathologie Endocriniennes, Assistance Publique-Hôpitaux de Paris, Hôpital Bicêtre, Service d'Endocrinologie et des Maladies de la Reproduction, Centre de Référence des Maladies Rares de l'Hypophyse, Le Kremlin-Bicêtre, France

Annamaria Colao Dipartimento di Medicina Clinica e Chirurgia, Sezione di Endocrinologia, University Federico II of Naples, Naples, Italy

Nicolas Coronel-Restrepo Division of Endocrinology, Clinica Medellin – Grupo Quirónsalud, Medellin, Colombia

Francesca D'Ercole Section of Endocrinology and Internal Medicine, Department of Medical Sciences, University of Ferrara, Ferrara, Italy

Antonio C. Fuentes-Fayos Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Monica Gadelha Neuroendocrinology Research Center/Endocrinology Division – Medical School and Hospital Universitário Clementino Fraga Filho, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

Neuroendocrinology Division, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

Neuropathology and Molecular Genetics Laboratory, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

Rua Professor Rodolpho Paulo Rocco, 255, 9° andar - Setor 9F, Centro de Pesquisa em Neuroendocrinologia, Ilha do Fundão, Rio de Janeiro, Brazil

Irene Gagliardi Section of Endocrinology and Internal Medicine, Department of Medical Sciences, University of Ferrara, Ferrara, Italy

Manuel D. Gahete Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Cell Biology, Physiology and Immunology, University of Córdoba, Cordoba, Spain

E. Giardino Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

Aleksandra Gilis-Januszewska Department of Endocrinology, Endocrine Oncology and Nuclear Medicine, Jagiellonian University Medical College, Cracow, Poland

Ludovica F. S. Grasso Dipartimento di Medicina Clinica e Chirurgia, Sezione di Endocrinologia, University Federico II of Naples, Naples, Italy

Vicente Herrero-Aguayo Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Juan M. Jiménez-Vacas Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Gregory Kaltsas Endocrinology Unit, 1st Department of Propaedeutic and Internal Medicine, Medicine School, National and Kapodistrian University of Athens, Athens, Greece

Rhonda D. Kineman Section of Endocrinology, Diabetes, and Metabolism, Department of Medicine, University of Illinois at Chicago, Chicago, IL, USA

Research and Development Division, Jesse Brown Veterans Affairs Medical Center, Chicago, IL, USA

Márta Korbonits Centre for Endocrinology, William Harvey Research Institute, Barts and the London School of Medicine and Dentistry, Queen Mary University of London, London, UK

Kalman Kovacs Department of Laboratory Medicine, Division of Pathology and the Keenan Research Centre for Biomedical Science at the Li Ka Shing Knowledge Institute, St. Michael's Hospital, University of Toronto, Toronto, ON, Canada

Juan L. Lopez-Cánovas Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Raúl M. Luque Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Luigi Maione Université Paris-Saclay, Inserm, Physiologie et Physiopathologie Endocriniennes, Assistance Publique-Hôpitaux de Paris, Hôpital Bicêtre, Service d'Endocrinologie et des Maladies de la Reproduction, Centre de Référence des Maladies Rares de l'Hypophyse, Le Kremlin-Bicêtre, France

F. Mangili Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

G. Mantovani Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

Endocrinology Unit, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy

Monica Marazuela Department of Endocrinology, Hospital Universitario de la Princesa, Instituto de Investigación Princesa, Universidad Autónoma de Madrid. C/ Diego de León 62, Madrid, Spain

Maria Cristina De Martino Dipartimento di Medicina Clinica e Chirurgia, Sezione di Endocrinologia, University Federico II of Naples, Naples, Italy

E. Peverelli Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

Rosa Pirchio Dipartimento di Medicina Clinica e Chirurgia, Sezione di Endocrinologia, University Federico II of Naples, Naples, Italy

Rosario Pivonello Dipartimento di Medicina Clinica e Chirurgia, Sezione di Endocrinologia, University Federico II of Naples, Naples, Italy

Ana M. Ramos-Leví Department of Endocrinology, Hospital Universitario de la Princesa, Instituto de Investigación Princesa, Universidad Autónoma de Madrid. C/ Diego de León 62, Madrid, Spain

Fabio Rotondo Department of Laboratory Medicine, Division of Pathology and the Keenan Research Centre for Biomedical Science at the Li Ka Shing Knowledge Institute, St. Michael's Hospital, University of Toronto, Toronto, ON, Canada

Prudencio Sáez-Martínez Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Sharjeel Shaikh Department of Medicine, Wexford General Hospital – University College Dublin, Wexford, Ireland

Luis V. Syro Department of Neurosurgery, Hospital Pablo Tobon Uribe and Clinica Medellin – Grupo Quirónsalud, Medellin, Colombia

Gianluca Tamagno Hermitage Medical Clinic, Dublin, Ireland

Sicheng Tang Medizinische Klinik und Poliklinik IV, Ludwig Maximilian University Munich, Munich, Germany

Marily Theodoropoulou Medizinische Klinik und Poliklinik IV, Ludwig Maximilian University Munich, Munich, Germany

D. Treppiedi Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

Marina Tsoli Endocrinology Unit, 1st Department of Propaedeutic and Internal Medicine, Medicine School, National and Kapodistrian University of Athens, Athens, Greece

María C. Vázquez-Borrego Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain

Susan M. Webb Endocrinology/Medicine Department, Hospital de la Santa Creu i Sant Pau. Centro Investigación Biomédica en Red de Enfermedades Raras (CIBERER, Unidad 747), IIB-Sant Pau, ISCIII, Barcelona, Spain

Universitat Autònoma de Barcelona (UAB), Barcelona, Spain

Luiz Eduardo Wildemberg Neuroendocrinology Research Center/Endocrinology Division – Medical School and Hospital Universitário Clementino Fraga Filho, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

Neuroendocrinology Division, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

Neuropathology and Molecular Genetics Laboratory, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

Kartik Yadav Department of Medicine, Wexford General Hospital – University College Dublin, Wexford, Ireland

Maria Yavropoulou Endocrinology Unit, 1st Department of Propaedeutic and Internal Medicine, Medicine School, National and Kapodistrian University of Athens, Athens, Greece

Maria Chiara Zatelli Section of Endocrinology and Internal Medicine, Department of Medical Sciences, University of Ferrara, Ferrara, Italy



Anatomy of the Pituitary Gland

Nicolas Coronel-Restrepo, Luis V. Syro, Fabio Rotondo, and Kalman Kovacs

1.1 Anatomy

The pituitary gland or hypophysis is a small, bean-shaped gland located in the sella turcica or pituitary fossa, which is the largest bone depression in the superior surface of the sphenoid bone. The *tuberculum sellae*, a variable median elevation, forms the anterior boundary of the sella turcica; the posterior border is formed by the *dorsum sellae*, and its floor is the roof of the sphenoid sinus. Bone and dura mater protect the gland in its anterior, inferior, and posterior surfaces, whereas the lateral and the superior surfaces are only covered by dura (Fig. 1.1). The lateral walls of the sella have only one layer of dura, but a double-layered dura covers all other surfaces. Superiorly, the pituitary gland is covered by the *diaphragma sellae* (sellar diaphragm), a dural sheath with a medial opening which allows the passage of the pituitary stalk [1, 2]. The optic chiasm lies anterior to the pituitary stalk and, in approximately 70% of the cases, above the sellar diaphragm; it is *prefixed*, when it is

N. Coronel-Restrepo

Division of Endocrinology, Clinica Medellin – Grupo Quirónsalud, Medellin, Colombia

L. V. Syro (🖂)

F. Rotondo · K. Kovacs

1

The chapter has been endorsed by **Prof. Emmanuel Jouanneau**, emmanuel.jouanneau@ chu-lyon.fr, Service de Neurochirurgie, Hospices Civils de Lyon, Lyon, France

Department of Neurosurgery, Hospital Pablo Tobon Uribe and Clinica Medellin – Grupo Quirónsalud, Medellin, Colombia

Department of Laboratory Medicine, Division of Pathology and the Keenan Research Centre for Biomedical Science at the Li Ka Shing Knowledge Institute, St. Michael's Hospital, University of Toronto, Toronto, ON, Canada

[©] Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_1



Fig. 1.1 Sellar and parasellar region

situated posteriorly, above the dorsum sellae. Depending on the position of the optic chiasm—prefixed, normal, or postfixed—any lesion that extends upwards may cause different visual field defects [3].

The cavernous sinuses, containing the internal carotid artery, the oculomotor, trochlear, and abducens nerves, are located bilaterally to the pituitary gland [4] (Fig. 1.1). Hence, the medial wall of the cavernous sinuses forms the lateral border of the sella. The cavernous sinus is the only anatomic site in the body where an artery traverses completely through a large venous channel. The cavernous sinuses are connected in the midline by the anterior and posterior intercavernous sinuses; they drain into the superior and inferior petrosal sinuses and finally into the internal jugular veins. In some cases, below the pituitary gland, there exists an inferior intercavernous sinus or venous plexus that needs to be underlined as a potential source of bleeding during any surgical procedure. The anatomic variability in the size of the diaphragm opening, and the fact that the medial wall of the cavernous sinus has only one single layer of dura, may explain the growth pattern of some pituitary tumors.

The normal weight of the pituitary gland, in adults, is approximately 0.6 g and measures 13 mm (transverse), 10 mm (anteroposterior), and 6 mm (vertical). The pituitary gland changes according to different physiological stages. At puberty, or during pregnancy and postpartum, hypertrophy of the pituitary gland can be seen [5]. Since the pituitary gland is close to several vital neurovascular structures, any benign or malignant lesion may present with a broad clinical spectrum and diverse outcomes [6–8].

The human pituitary gland consists of two distinct lobes: the anterior lobe or adenohypophysis, and the posterior lobe or neurohypophysis, each one of different embryologic origin [9] (Fig. 1.2). The adenohypophysis develops from Rathke's pouch, an invagination of the ectodermal primitive oral cavity, whereas the neurohypophysis develops from a downward extension of the neural ectoderm in the floor



Fig. 1.2 Anatomy of the pituitary gland

of the diencephalon. The adenohypophysis consists of the *pars distalis*, the *pars intermedia*, and the *pars tuberalis*. The *pars distalis* (also known as pars anterior or pars glandularis) comprises 80% of the gland. The adult human pituitary gland has no anatomically distinct *pars intermedia*. It is made up of a small area of cells located at the junction of the two lobes, but it is morphologically rudimentary, and is incorporated into the *pars distalis*. The *pars tuberalis* is a minor upward extension of the adenohypophysis attached to the lower pituitary stalk. Histologically, the adenohypophysis comprises a central median wedge bordered by two lateral wings. The different hormone-producing cell types are distributed in an uneven, but specific manner. They are arranged within uniformly sized acini surrounded by a delicate reticulin fiber network, giving the pituitary its distinct architecture. The non-endocrine folliculo-stellate cells are localized in the center of the acini. The neurohypophysis is composed of nerve fibers that arise in the hypothalamic nuclei. It consists of three parts: the *medial eminence* or *infundibular process* (Fig. 1.2).

The vascular supply and blood flow to the pituitary gland play an important role in the regulation of the adenohypophysial hormone secretion [9-11]. They carry the stimulatory and inhibitory hypothalamic hormones from the medial eminence to the adenohypophysis (Fig. 1.3). The blood irrigation of the gland is supplied by the superior and inferior hypophysial arteries. The *superior hypophysial arteries* – often three or four on both sides, joining around the proximal part of the pituitary stalk – originate from the supraclinoid segment of the internal carotid arteries, or the posterior communicating arteries. They reach and encircle the medial eminence generating an external and an internal capillary plexus. The external plexus is composed of small arteries that surround the upper part of the pituitary stalk. The



Fig. 1.3 Blood circulation to the pituitary gland

internal plexus is formed by a mesh of capillaries that contribute to the formation of the gomitoli, specialized vascular structures composed of a central muscular artery surrounded by a dense spiral network of capillaries [9, 11]. The function of the gomitoli is not clear, but it appears that they regulate the blood flow, influencing the release of the hypothalamic hormones in the pituitary paracrine network. The *inferior hypophysial arteries* originate from the meningohypophysial trunk within the cavernous sinus. They anastomose with each other to form an arterial circle around the posterior lobe, providing most of the blood supply to the posterior lobe, and to the lower portion of the stalk.

The hypophysial portal system connects the hypothalamus with the anterior pituitary; it stems from the internal capillary plexus of the median eminence and stalk [11] (Fig. 1.3). The *long portal vessels* originate from the median eminence and superior pituitary stalk; they descend on the anterior surface of the pituitary stalk and provide 90% of the blood that nourishes the anterolateral pars distalis. The *short portal vessels* originate in the lower part of the pituitary stalk; they go to the central portion of the anterior lobe and provide the remaining 10% of blood supply to the anterolateral pars distalis. Aside from a minor direct arterial supply from the inferior hypophysial arteries, most of the anterior lobe circulation is venous, coming from the portal vessels. The blood supply of the posterior lobe is arterial, in contrast to that of the anterior lobe, which is venous, and is provided by the hypophysial portal system. This type of circulation has important clinical issues. If the pituitary stalk is damaged by a trauma that compromises the hypophysial portal system, the anterior pituitary can suffer a hemorrhagic infarct [12]. The fact that the anterior lobe receives minimal arterial supply can explain the infrequent metastases in that area [9].

The two most important pathways that link the hypothalamus to the pituitary are: the *tuberoinfundibular tract* and the *supraopticohypophysial tract* [13] (Fig. 1.3). The *tuberoinfundibular tract* arises from small neurons situated in the arcuate nucleus and the periventricular zone. The hypothalamic releasing hormones are carried by the axons down to the median eminence and to the infundibulum, where they are released into the hypophysial portal system. The *supraopticohypophysial tract* arises from large neurons located in the supraoptic and paraventricular nuclei. They synthesize oxytocin and vasopressin, which are transported and stored in specialized axon terminals (Herring bodies). When released, they enter the capillary plexus in the posterior pituitary and reach the general circulation via the hypophysial veins.

1.2 Histology

1.2.1 Cell Types of the Adenohypophysis

The endocrine cell types of the anterior pituitary gland develop from a common primordium. Cytodifferentiation occurs as a response to signaling pathways that emanate from distinct organizing centers. To develop and grow, each cell type requires additional specific transcription factors that regulate its cytodifferentiation and hormone production [14–16] (Table 1.1). Pituitary-specific transcription factor 1 (Pit-1) regulates the somatotroph, lactotroph, and thyrotroph cell types which produce growth hormone (GH), prolactin (PRL), and thyroid-stimulating hormone (TSH),

Transcription factor	Coexpression	Cell lineage	Hormone	Hypersecretion
Pit-1		Somatotrophs	GH	Acromegaly
	ER-α	Lactotrophs	PRL	Hyperprolactinemia
	GATA-2	Thyrotrophs	TSH	Hyperthyiroidism
Tpit		Corticotrophs	ACTH	Cushing disease
SF-1	ER- α, GATA-2,	Gonadotrophs	LH, FSH,	Most
	GATA-3		α-subunit	non-functional

 Table 1.1 Pituitary transcription factors, cell lineage, and hormone production in adenohypophysial cells

GH growth hormone, *PRL* prolactin, *TSH* thyroid-stimulating hormone, *ACTH* adrenocorticotropic hormone, *LH* luteinizing hormone, *FSH* follicle-stimulating hormone, *Pit-1* pituitaryspecific transcription factor 1, *Tpit* T-box transcription factor TBX19, *GATA-2* GATA binding protein 2, *GATA-3* GATA binding protein 3, *ER-* α estrogen receptor alpha, *SF-1* steroidogenic factor (Modified from Reference 35) respectively. T-box transcription factor TBX19 (Tpit) regulates the corticotroph cells which produce adrenocorticotropic hormone (ACTH). The expression of steroidogenic factor 1 (SF1), estrogen receptor alpha (ER- α), and Guanine-Adenine-Thymine-Adenine binding protein 2 (GATA-2) regulates the gonadotroph cell type development. Estrogen receptor alpha assists Pit-1 to boost PRL secretion while repressing the production of GH; consequently, lactotrophs require the co-expression of both Pit-1 and ER- α for their development and function. Thyrotroph cell development requires the co-expression of GATA-2 along with Pit-1. Besides their role in pituitary gland development, the pituitary transcription factors serve as diagnostic markers of cell lineage in the classification of pituitary tumors [17–19].

Somatotrophs (GH-producing cells) make up approximately 50% of the gland, occupying mainly the lateral wings. They show strong acidophilia and GH hormone immunoreactivity. Some somatotrophs also express PRL or the α -subunit of the glycoprotein hormones [20]. Lactotrophs (PRL-producing cells) account for 10-30% of the pituitary gland cell population. In men and nulliparous women, they may represent 10% of the cell population, whereas in multiparous or lactating women, they may account for approximately 30% of the cell population [21]. They are chromophobic or slightly acidophilic, show PRL immunoreactivity, and are irregularly scattered throughout the entire anterior pituitary; but most often, they accumulate in the posterolateral rim of the lateral wings. Corticotrophs (ACTHproducing cells) reside within the median wedge of the pituitary gland. They are basophilic, periodic acid-Schiff (PAS)-positive, and immunoreactive for ACTH, as well as for other pro-opiomelanocortin (POMC) peptides. They represent approximately 15-20% of the cell population. Thyrotrophs (TSH-producing cells) occupy the anterior one-third of the median wedge and represent approximately 5% of the gland. They are slightly basophilic and strongly immunoreactive for β -TSH and the α-subunit. Gonadotrophs (FSH- and LH-producing cells) account for 10% of the cells distributed throughout the pars distalis. They are immunoreactive for β-FSH, β -LH, and their α -subunit.

The hormonal function of the adenohypophysial cell types exhibits considerable flexibility, depending on the functional demand placed on them. Reversible transdifferentiation can occur between members of the Pit-1 group as it has been documented: GH cells to PRL cells during pregnancy, and GH cells to TSH cells in hypothyroidism [22]. Pituitary cell types differ not only in their function, structure, and hormone content, but also in their morphological response to functional stimulation or suppression.

Folliculo-stellate cells, a major group of cells localized in the parenchymal tissue of the anterior pituitary, make up to 10% of the cell population in the human adenohypophysis. They have no hormonal function, but various experimental in vitro and morphological studies have documented their importance [23, 24]. To date, the functional role of folliculo-stellate cells remains a mystery. Various theories published suggest that they could be pituitary progenitor or stem cells, but whether this is true in humans, remains to be elucidated.

1.3 Pathology

1.3.1 Pituitary Adenomas

Pituitary adenomas are the most common type of pituitary disorder and account for 15–20% of all intracranial tumors. They are monoclonal, benign lesions, and are produced by genetic and epigenetic alterations that influence either proto-oncogenes, or tumor suppressor genes. The initiating cause remains unclear, but inactivation or overexpression of cell cycle regulators may be enough to trigger pituitary tumorigenesis [6]. A tumor results when an induced change gives growth advantage to a previously normal cell among other surrounding cells. Once this happens, the tumor grows by selective pressures from microenvironmental ecology [25, 26]. Tumor growth can be regarded as a Darwinian, adaptive system that grows as an evolutionary process [27]. The limited resources of the microenvironment restrict its size and its progression. Hence, natural selection in tumors, as in organisms, takes place through competition for space and resources [25, 28–30]. While the majority of pituitary adenomas are sporadic, some of them are associated with familial syndromes, exhibiting different genetic background, variable phenotype, and diverse clinical behavior [31, 32].

Pituitary adenomas may be referred to as micro or macroadenomas (if they are smaller or larger than 10 mm in diameter, respectively) and, depending on their hormonal hypersecretion, as functional or non-functional [7]. Their growth pattern may be either expansive, as in a slowly growing mass exerting pressure on the pituitary gland and the sella, or infiltrative, as a tumor spreading into the surrounding bone, dura, or cavernous sinuses. Pituitary adenomas extending into the suprasellar space may compress the optic chiasm and cause visual disturbances, a frequent clinical manifestation of macroadenomas [33].

Using the hematoxylin & eosin (H&E) stain for examining cellular and tissue structure detail, cells making up pituitary adenomas may be classified as acidophilic, basophilic, or chromophobic; this tinctorial property is not completely related to their hormonal function. The next step is to demonstrate reticulin fibers to differentiate normal adenohypophysis from hyperplasia or adenoma. The normal acinar reticular pattern will be enlarged but intact in cases of hyperplasia and will be lost in adenomas (Fig. 1.4a). The use of immunohistochemical methods that employ monoclonal antibodies against pituitary hormones helps to characterize the different functional cell type(s) in the tumor (Table 1.2). The current World Health Organization (WHO) classification (2017) of pituitary adenomas recommends the use of pituitary cell lineages along with their hormone production [17, 18, 34, 35] (Table 1.3). Accordingly, the classification makes use of a wide range of monoclonal antibodies against pituitary hormones and pituitary transcription factors. Electron microscopy is currently utilized only in difficult to diagnose cases. A clinicopathological classification, which considers proliferation markers, invasion, and size, has been recommended [36] and, recognizing the variable behavior of pituitary adenomas, a proposal to replace the term pituitary adenoma to pituitary neuroendocrine tumor (PitNET) has also been made [37].



Fig. 1.4 Histologic features of pituitary pathology. (a) Light microphotograph showing the reticulin fiber network in both the normal pituitary gland and adenoma interface. The delicate normal acinar architecture is preserved in the normal gland (circle) but is totally broken down in the adenoma (asterisk). Stain: Collagen IV. Magnification: 10×. (b) Densely granulated somatotroph adenoma. Immunostaining for Cam5.2 demonstrates a strong, diffuse cytoplasmic pattern (arrow). Stain: Cam5.2 antibody. Magnification: 40×. (c) Sparsely granulated somatotroph adenoma. The fibrous bodies seen here are characteristic of this tumor type (arrow). Staining shows a strongly immunopositive perinuclear pattern as a result of the disruption and condensing of the cells keratin cytoskeleton. Stain: Cam5.2 antibody. Magnification: 40×. (d) Sparsely granulated somatotroph adenoma. Cells are chromophobic and pleomorphic on HE stain; in some of them, the fibrous bodies are apparent as spherical, paranuclear, unstained structures (arrows). Stain: Hematoxylin & eosin (HE). Magnification: 40x. (e) Non-tumorous pituitary gland in a patient with Cushing disease demonstrating the response of the non-tumoral corticotroph cells to glucocorticoid excess. The cytoplasm in these corticotroph cells undergoes an accumulation of keratin filaments which causes a glassy hyaline appearance (arrow). This is known as Crooke's hyaline change. Stain: Hematoxylin & eosin (HE). Magnification: 40×. (f) Light microphotograph showing the cytokeratin pattern seen in Crooke's cells. Massive accumulation of perinuclear cytokeratin, occurring in corticotrophs under the effect of glucocorticoid excess, is demonstrated as a strong ring-like pattern around the nucleus (arrow). Stain: Cam5.2 antibody. Magnification: 40×

Stain	Stain pattern	Indication
Hematoxylin-eosin (HE)	Acidophilic, basophilic	Ancillary stain
	or chromophobic	For initial classification of the tumor
Periodic acid–Schiff (PAS)	Cytoplasmic	To recognize corticotrophs and Crooke cells
Reticulin, Silver stain, Collagen IV	Acinar reticular pattern	To differentiate normal pituitary gland, hyperplasia, or tumor
Chromogranin, Synaptophysin	Secretory granules	Neuroendocrine markers
TTF-1	Nuclear	Positive in posterior pituitary gland and posterior pituitary tumors
GH, PRL, ACTH, TSH, FSH, LH, α-subunit	Cytoplasmic	To identify hormone secretion
Cam 5.2	Cytoplasmic Dot pattern Ring-like pattern	Positive in somatotrophs and corticotrophs To categorize pure somatotroph adenomas as densely or sparsely granulated To identify Crooke cells
Pit-1, Tpit, SF-1, ER-α, GATA-2, GATA-3	Nuclear	To identify cell lineage differentiation
E-cadherin	Membrane	Absent E-cadherin expression may be correlated to tumor invasiveness
Ki67, p53	Nuclear	Proliferation markers
SSTR2, SSTR5	Membrane	Predictive biomarkers for long-acting somatostatin analogs?
MGMT	Nuclear	To evaluate DNA repair pathway, DR Predictive biomarker for temozolomide treatment in aggressive pituitary adenomas and carcinomas
MSH2, MSH6, MLH1, PMS2	Nuclear	To evaluate integrity of DNA repair pathway, MMR
MPG	Nuclear	To evaluate integrity of DNA repair pathway, BER
VEGF	Cytoplasmic	To assess angiogenic growth and progression of tumor

 Table 1.2
 Morphological evaluation of pituitary adenomas

TTF-1 thyroid transcription factor 1, *GH* growth hormone, *PRL* prolactin, *TSH* thyroid-stimulating hormone, *FSH* follicle-stimulating hormone, *LH* luteinizing hormone, *ACTH* adrenocorticotropic hormone, *LMWK* low molecular weight keratin, *SSTR2* somatostatin receptor 2, *SSTR5* somatostatin receptor 5, *AIP* aryl-hydrocarbon receptor-interacting protein, *MGMT* O6-methylguanine DNA methyltransferase, *MSH2* MutS protein homolog 2, *MSH6* mutS homolog 6, *DNA* deoxyribonucleic acid, *DR* direct repair, *MMR* mismatch repair, *MPG* N-methylpurine DNA glycosylase, *BER* base excision repair, *VEGF* Vascular endothelial growth factor (Modified from reference 38)

1.3.1.1 Pit-1-Positive Tumors

Somatotroph Adenomas

Somatotroph adenomas arise from Pit-1 pituitary-cell lineage and express GH. Approximately 15% of surgically removed pituitary adenomas represent pure GH cell adenomas [38, 39]. Most of these tumors are associated with clinical signs of acromegaly, but silent forms have also been reported [40]. According to the density of GH-containing secretory granules in the cytoplasm of the cells, somatotroph

Tumor	Transcription factors	Hormones		
Pit-1-positive tumors				
Somatotroph adenomas				
Sparsely granulated	Pit-1	GH (weak)		
Densely granulated		GH, α-subunit		
Mammosomatotroph adenomas	Pit-1, ER-α	GH, PRL, α-subunit		
Mixed GH- PRL adenomas	Pit-1, ER-α	GH, PRL, α-subunit		
Plurihormonal GH-producing	Pit-1, ER-α	GH, PRL, α -subunit,		
adenomas		β-TSH		
Lactotroph adenomas				
Sparsely granulated	Pit-1, ER-α	PRL, α -subunit		
Densely granulated		PRL		
Acidophil stem cell adenomas		PRL, GH		
Thyrotroph adenomas	Pit-1, GATA-2	β-TSH, α-subunit		
Monomorphous plurihormonal				
adenomas				
Silent subtype 3	Pit-1	GH, PRL, β-TSH		
Tpit-positive tumors				
Corticotroph adenomas				
Densely granulated	Tpit	ACTH		
Sparsely granulated		ACTH		
Crooke cell adenoma		ACTH		
SF-1-positive tumors				
Gonadotroph adenomas	SF-1, ER-α, GATA-2, GATA-3	β-FSH, β-LH, α-subunit		
Hormone-positive				
Gonadotroph adenomas		None		
Hormone-negative				
Polymorphous plurihormonal adenomas				
Plurihormonal adenomas	Multiple	Multiple		
Transcription factor and hormone-negative adenomas				
Null cell adenomas	None	None		

 Table 1.3
 Classification of pituitary adenomas

GH growth hormone, *ACTH* adrenocorticotrophic hormone, *PRL* prolactin, *FSH* folliclestimulating hormone, *LH* luteinizing hormone, *TSH* thyroid-stimulating hormone, *Pit-1* pituitaryspecific transcription factor 1, *Tpit* T-box transcription factor TBX19, *GATA-2* GATA binding protein 2, *GATA-3* GATA binding protein 3, *ER-* α estrogen receptor alpha, *SF-1* steroidogenic factor 1 (Modified from reference 19)

adenomas can be classified as densely or sparsely granulated. A clear distinction between them is crucial for proper prognosis and treatment [41-43].

Densely granulated somatotroph adenomas are strongly acidophilic tumors with extensive immunoreactivity for GH and are often accompanied by α -subunit expression. Scattered immunopositivity for PRL and β -TSH is less frequent and not associated with hypersecretion. Immunostaining for Cam5.2 demonstrates a diffuse cytoplasmic pattern (Fig. 1.3b). Ultrastructurally, this adenoma type is comprised of cells that are like those of normal somatotrophs. A hallmark feature of densely granulated somatotroph adenomas is the large number of secretory granules dispersed throughout the cytoplasm. They occur with the same frequency in both sexes and display a slow, expansive growth, causing the typical "ballooning of the sella". They

may remain intrasellar for several years. Usually they have high expression of somatostatin receptors and respond with somatostatin analogs treatment [41].

Sparsely granulated somatotroph adenomas are usually chromophobic, with sparse and weak GH immunoreactivity. Nuclear pleomorphism may be evident, and the adenoma cells show spherical, paranuclear, unstained structures: the fibrous bodies. These fibrous bodies are strongly immunopositive for Cam5.2 in a dot pattern, characteristic of this tumor subtype (Fig. 1.3c, d). They also contain crescent-shaped cells scattered throughout the tumor, since the fiborous bodies can displace the nuclei. Sparsely granulated somatotroph adenomas, as opposed to the densely granulated type tumors, occur in younger patients. Multiple immunoreactivities for pituitary hormones are rarely noted. They tend to be macroadenomas at the time of diagnosis and are often aggressive, rapidly growing, and invasive [41]. Ultrastructurally, they show little similarity to the normal pituitary. They contain irregular-shaped nuclei, unevenly developed with varying amounts of rough endoplasmic reticulum present within the cell. Unlike the densely granulated somatotroph adenomas, this subtype contains few secretory granules which are much smaller in diameter.

Mixed somatotroph and lactotroph adenomas are made up of two distinct cell types. They are associated with acromegaly and different degrees of hyperprolactinemia. Mixed adenomas usually consist of densely granulated GH cells and sparsely granulated PRL cells. These acidophilic and chromophobic cells display immunoreactivity for GH and PRL. Ultrastructurally, two distinct features can be seen: morphology resembling that of the adenoma that produces only GH and one that produces only PRL.

Mammosomatotroph cell adenomas are monomorphous, consisting of one cell type displaying both GH and PRL immunoexpression, although immunopositivity is practically always stronger for GH. They are associated with acromegaly and hyperprolactinemia, showing biological behavior like that of densely granulated somatotroph adenomas. Ultrastructurally, these tumor cells contain both GH-containing and PRL-containing secretory granules, but they resemble adenoma cells found in densely granulated somatotroph adenomas. Mammosomatotroph adenomas contain two types of pleomorphic and heterogeneous secretory granules: large, irregular, or elongated granules; and, small to moderate-sized secretory granules that have a spherical or ovoid-shaped.

Lactotroph Adenomas

Lactotroph adenomas arise from the Pit-1 pituitary cell lineage and express mainly PRL. They are classified into three histological subtypes: sparsely granulated, densely granulated, and acidophil stem cell adenomas.

Sparsely granulated lactotroph adenomas are the most common adenoma type and they are associated with hyperprolactinemia. Lactotroph adenomas have a wide-ranging biological behavior from indolent to aggressive, and in men usually present with invasive radiological features. The PRL blood levels are generally proportional to the tumor size. Most lactotroph adenomas are chromophobic, PAS-negative cells with a diffuse or papillary growth pattern, and exhibit strong PRL immunopositivity with co-expression of Pit-1 and ER- α . Psammoma bodies and interstitial amyloid have been documented in some cases, while cellular and nuclear pleomorphism is rarely seen. Calcification within the adenoma can be found in circa 15% of all sparsely granulated lactotroph adenoma cases. Ultrastructurally, three noticeable features are: a rough endoplasmic reticulum, a prominent Golgi apparatus, and the lateral extrusion of the secretory granules—commonly known as misplaced exocytosis.

Densely granulated lactotroph adenomas are rare and have been considered aggressive tumors. Misplaced exocytosis is the hallmark of sparsely granulated lactotroph adenomas but is rarely shown in densely granulated lactotroph adenomas. Ultrastructurally, numerous ovoid/spherical large secretory granules are clearly noted, as well as, well-developed Golgi complexes but the rough endoplasmic reticulum is less abundant, as compared to the sparsely granulated lactotroph adenoma subtype.

Acidophil stem cell adenomas are monomorphous tumors with morphological signs of PRL and GH differentiation. They are rare and are associated with hyperprolactinemia, but the serum PRL levels may be disproportionately low for their size [44]. Elevation of GH levels and acromegaly are infrequent. They are chromophobic, with moderate to strong immunoreactivity for PRL. Immunopositivity for GH is weak or negative, and Cam5.2 reveals the dot-like positivity of fibrous bodies in some cells. Ultrastructure features include an increased number of giant mitochondria (oncocytic change), small secretory granules with lateral extrusion (PRL marker), and fibrous bodies (GH marker).

Although calcification is extremely rare in other adenoma types, about 10–15% of lactotroph adenomas may display varying degrees of calcification. Dopamine agonists produce remarkable morphological changes: the nucleus becomes hetero-chromatic and the cytoplasm displays marked shrinkage due to the loss of the hormone-producing apparatus. As a result, PRL immunoreactivity is reduced or lost. Although these morphological changes are reversible, some portions of the tumor may maintain their suppressed features when treatment is discontinued. Long-term treatment with dopamine agonists may also cause varying degrees of fibrosis and calcification which results in the presence of psammoma bodies.

Thyrotroph Adenomas

Thyrotroph adenomas express mainly TSH and arise from Pit-1 pituitary-cell lineage. They are associated either with hyperthyroidism or inappropriately elevated levels of TSH. In some cases, they occur in euthyroid subjects. They are often macroadenomas with a tendency to invade and their morphology exhibits surprising diversity. Cells making up thyrotroph adenomas are generally chromophobic and highly differentiated, comprising elongated polar cells that form pseudorosettes around vessels. In other cases, the pattern may be diffuse with considerable nuclear pleomorphism. Another variant is markedly fibrotic. Immunoreactivity for TSH is variable; it is often patchy or scattered, rarely extensive. Scattered cells may exhibit immunoreactivity for GH, PRL, and α -subunit. Both Pit-1 and GATA2 are expressed. In most cases, they are strongly positive for somatostatin receptor 2 (SSTR2) immunostaining. Ultrastructurally, these adenoma cells resemble normal thyrotroph cells: euchromatic nuclei, abundant rough endoplasmic reticulum, well-developed Golgi complexes, microtubules, and secretory granules located along the cell membrane.

Monomorphous Plurihormonal Adenomas

Plurihormonal adenomas produce more than one hormone. They can be monomorphous or plurimorphous, consisting of two or more different cell lineages. They include *plurihormonal Pit-1-positive adenomas* previously called silent subtype 3 adenomas. They are rare, but clinically significant due to their aggressive behavior. Plurihormonal Pit-1-positive adenomas can mimic lactotroph adenomas or, in some cases, may present with acromegaly. They are often acidophilic, may show mild PAS positivity, and form a diffuse or lobular pattern. Most tumor cells are immuno-negative for pituitary hormones, but some of them may demonstrate scattered minor positivity. They exhibit positive nuclear expression for Pit-1.

1.3.1.2 Tpit-Positive Tumors

Corticotroph Adenomas

Corticotroph adenomas arise from Tpit pituitary-cell lineage and express ACTH and other POMC-derived peptides. They are classified into three subtypes: densely granulated, sparsely granulated, and Crooke cell adenomas.

Densely granulated corticotroph adenomas are basophilic and PAS-positive, with a sinusoidal or diffuse pattern. Immunoreactivity can be demonstrated not only for ACTH, but also for other POMC peptides (β -endorphin, β -LPH, and CLIP). Perinuclear bundles of cytokeratin filaments are demonstrated with Cam5.2 as a cytoplasmic, perinuclear pattern. Most corticotroph tumors are small microadenomas causing Cushing's disease [42, 45]. The adenomas, often measuring only a few millimeters in diameter, may be too small to be detected by imaging or to be identified at surgery. In a few cases of milder hypercorticism, Cushing's disease is produced by large, aggressive, and invasive tumors. Histologically they exhibit variable, often weak, PAS positivity, and ACTH immunoreactivity. A few cases of aggressive macroadenomas also display immunoreactivity for LH and α -subunit. In cases of Nelson's syndrome, the morphological features are like those of densely granulated corticotroph adenomas in Cushing's disease, showing few or no cytokeratin filaments.

Sparsely granulated corticotroph adenomas are composed of slightly basophilic or chromophobic cells. They are PAS-positive with patchy immunopositivity for ACTH. In some cases, they are clinically silent without Cushing's disease.

The response of the non-tumoral corticotroph cells to high levels of glucocorticoids is an accumulation of cytokeratin filaments in the cytoplasm (Crooke's change), which causes a glassy hyaline appearance with a displacement of the ACTH-positive granules to the cell periphery (Fig. 1.3e). These "Crooke cells" are identified by using Cam5.2 immunostaining, which reveals a strong ring-like pattern around the nucleus (Fig. 1.3f). The identification of Crooke cells is of paramount importance in cases of persistent hypercortisolism after surgery, with only a non-tumoral pituitary gland identifiable in the resected tissue. The presence of Crooke cells in the non-tumoral pituitary gland confirms the previous hypercorticism and will dictate the next step of treatment. Therefore, Crooke hyalinization would be noted in: (1) non-tumoral corticotrophs adjacent to the lesion – in that case, the tumor was overlooked by the surgeon; (2) in ectopic ACTH/corticotrophinreleasing hormone (CRH) syndrome; (3) in patients with glucocorticoid secreting adrenocortical tumors; (4) in patients taking glucocorticoids (factitious Cushing syndrome). In cases of pseudo-Cushing, Crooke cells will not be present in the pituitary tissue [45].

Crooke cell adenomas are a rare type of corticotroph adenomas. They may cause Cushing's disease or may be endocrinologically silent [39]. Crooke hyalinization is not expected to develop in tumoral corticotroph cells, although a minority of them may present that alteration, identical to the Crooke cells seen in the adenohypophysis of patients with glucocorticoid excess. When this change affects more than 50% of the tumor's cells, it is diagnosed as a Crooke cell adenoma [46]. The reason why Crooke cell adenomas produce ACTH while simultaneously display Crooke hyaline changes due to increased glucocorticoid excess, is not well understood. These adenomas are usually invasive, may exhibit aggressive clinical behavior, and often recur with a low rate of success for a cure after reoperation and radiotherapy [46].

Silent corticotroph adenomas do not display functional activity and present as clinically non-functional adenomas [47]. There are two subtypes. Silent corticotroph adenomas subtype 1—densely granulated—which exhibit the same morphology of corticotroph adenomas associated with Cushing's disease. They display a high propensity for hemorrhage and may present with pituitary apoplexy. Silent corticotroph adenomas subtype 2—sparsely granulated—are chromophobic adenomas comprised of small cells, which exhibit only modest PAS positivity and scattered immunoreactivity for ACTH. No cytokeratin filaments are present with Cam5.2 [40, 47]. These two adenoma subtypes are probably derived from cells of the pars intermedia.

1.3.1.3 SF-1-Positive Tumors

Gonadotroph Adenomas

Gonadotroph adenomas arise from SF-1 pituitary-cell lineage and produce β -FSH, β -LH, and α -subunit. They can also express ER- α , GATA2, and GATA3. The morphology of gonadotroph adenomas is variable. Histology may reveal either polar cells forming pseudorosettes around vessels, or a diffuse pattern. Oncocytic change, exhibiting an excessive increase of number and volume density of mitochondria, is frequent. Immunoreactivity for FSH, LH, and α -subunit is variable and often patchy. In some cases, there is minimal immunoexpression of hormones and the diagnosis can only be confirmed by analyzing the transcription factors present in the adenoma.

1.3.1.4 Polymorphous Plurihormonal Adenomas

Plurihormonal Adenomas

Polymorphous plurihormonal adenomas are rare tumors, often with unique ultrastructure. The most common cell combinations making up the adenoma are GH-TSH-PRL or PRL-TSH, but other combinations of different cellular lineages have also been documented. The immunoexpression of different transcription factors will depend on the different cell lineages of the tumor.

1.3.1.5 Transcription Factor-Negative and Hormone-Negative Adenomas

Null Cell Adenomas

Null cell adenomas do not display evidence of cell lineage differentiation by pituitary transcription factors or pituitary hormones. These hormonally inactive adenomas account for approximately 1–2% of surgically removed tumors [48, 49]. They are chromophobic, and pseudorosette formation, a characteristic of glycoprotein hormone-producing tumors, may also occur. Null cell adenomas are immunonegative for adenohypophysial hormones and transcription factors. Accumulation of mitochondria can occur, and in these cases, they are referred to as pituitary oncocytomas. Characteristically, oncocytomas possess larger cells than null cell adenomas and may display acidophilia due to non-specific binding of acidic stains by mitochondria. The tumor cells are known as oncocytes. Nuclei may show signs of shrinkage, hyperchromatism, and clumping of chromosomes, a condition known as pyknosis. The pattern of immunoreactivity is equal to null cells or gonadotroph adenomas. Ultrastructurally, oncocytomas are like null cell adenomas except for the extensive accumulation of mitochondria seen in oncocytomas. All other organelles are poorly developed.

1.3.2 Pituitary Carcinomas

Pituitary carcinomas can be diagnosed only when a pituitary neoplasm gives rise to distant metastasis. They are rare, representing about 0.12–0.2% [50]. They usually present as invasive macroadenomas and appear like other pituitary adenomas. Currently, there are no accurate criteria to distinguish between an invasive adenoma and one that has the potential to produce metastases [51, 52]. Most pituitary carcinomas produce either PRL or ACTH. Pituitary carcinomas are not accompanied by specific histological features, and enhanced mitotic activity, or nuclear and cellular pleomorphism do not necessarily indicate malignancy. Neoplasms with bland features may give rise to metastasis. Carcinomas usually display a higher cell proliferation marker (Ki-67) than adenomas. Immunoreactivities of pituitary carcinomas follow the pattern of their non-malignant phenotype [52].

1.3.3 Aggressive Pituitary Adenomas

Clinically aggressive pituitary adenomas comprise a wide group of tumor types that do not respond well to therapy, are recurrent, and are generally associated with a poor prognostic outcome. About 10% of pituitary adenomas can have an aggressive clinical course [51]. Aggressive tumors can be either micro- or macro-adenomas, have the potential to invade surrounding structures, and have a higher rate of recurrence compared to their benign adenoma counterparts [53]. Some types of adenomas may suggest aggressiveness. According to WHO classification of pituitary adenomas (2017), sparsely granulated somatotroph adenomas, lactotroph adenomas in men, silent corticotroph adenomas, Crooke cell adenomas, and plurihormonal PIT-1-positive adenomas are considered as high risk (of recurrence) adenomas [17]. No single feature can provide a reliable assessment to predict aggressive behavior. To date, there is no way to diagnose well in advance an aggressive tumor; the most reliable and accepted immunohistochemical analysis is the assessment of the Ki-67 nuclear labeling. If the Ki-67 nuclear labeling index is more than 10%, the tumor may be classified as aggressive, although there is no agreement on this. Another histologic marker used to correlate tumor behavior is the p53 expression [52]. Immunohistochemical study of MGMT expression may be used as a prognostic marker for aggressive pituitary adenomas. Studies have shown that progression and recurrence of pituitary adenomas are often associated with loss of MGMT expression [54, 55].

1.4 Conclusion

The pituitary gland and the hypothalamus regulate the endocrine system. These two structures are closely related to each other, thanks to a specialized network of blood vessels and associated nerve endings. Pituitary adenomas comprise a broad spectrum of tumors; different structures around the pituitary gland will determine the possibility of their complete resection with surgery. A precise morphological characterization of pituitary adenomas will help to choose a personalized therapy.

References

- Campero A, Martins C, Yasuda A, Rhoton AL Jr. Microsurgical anatomy of the diaphragma sellae and its role in directing the pattern of growth of pituitary adenomas. Neurosurgery. 2008;62(3):717–23.; discussion -23. https://doi.org/10.1227/01.neu.0000317321.79106.37.
- Yasuda A, Campero A, Martins C, Rhoton AL Jr, Ribas GC. The medial wall of the cavernous sinus: microsurgical anatomy. Neurosurgery. 2004;55(1):179–89.; discussion 89-90. https:// doi.org/10.1227/01.neu.0000126953.59406.77.
- Griessenauer CJ, Raborn J, Mortazavi MM, Tubbs RS, Cohen-Gadol AA. Relationship between the pituitary stalk angle in prefixed, normal, and postfixed optic chiasmata: an anatomic study with microsurgical application. Acta Neurochir. 2014;156(1):147–51. https://doi. org/10.1007/s00701-013-1944-1.

- Cavallo LMSD, Villa A, Somma T, Cappabianca P. Anatomy of the sellar and parasellar region. In: Zada GM, Lopes MBS, Mukundan Jr S, Laws Jr ER, editors. Atlas of sellar and parasellar lesions. Switzerland: Springer International Publishing; 2016. p. 3–9.
- Bonneville JF. Normal pituitary gland and pregnancy. In: Bonneville JF, Bonneville F, Cattin F, Nagi S, editors. MRI of the pituitary gland. Switzerland: Springer International Publishing; 2016. p. 53–5.
- Melmed S. Pathogenesis of pituitary tumors. Nat Rev Endocrinol. 2011;7(5):257–66. https:// doi.org/10.1038/nrendo.2011.40.
- Melmed S. Pituitary-tumor endocrinopathies. N Engl J Med. 2020;382(10):937–50. https:// doi.org/10.1056/NEJMra1810772.
- Syro LV, Rotondo F, Moshkin O, Kovacs K. Nonpituitary sellar masses. In: Melmed S, editor. The pituitary. 4th ed. London: Academic Press; 2017. p. 631–41.
- Lopes SMB, Pernicone PJ, Scheithauer BW, Horvath E, Kovacs K. Pituitary and sellar region. In: Mills SE, editor. Histology for pathologists. Philadelphia: Lippincott Williams & Wilkins; 2012. p. 343–72.
- Le Tissier P, Fiordelisio Coll T, Mollard P. The processes of anterior pituitary hormone pulse generation. Endocrinology. 2018;159(10):3524–35. https://doi.org/10.1210/en.2018-00508.
- Le Tissier P, Campos P, Lafont C, Romanò N, Hodson DJ, Mollard P. An updated view of hypothalamic-vascular-pituitary unit function and plasticity. Nat Rev Endocrinol. 2017;13(5):257–67. https://doi.org/10.1038/nrendo.2016.193.
- 12. Sav A, Rotondo F, Syro LV, Serna CA, Kovacs K. Pituitary pathology in traumatic brain injury: a review. Pituitary. 2019; https://doi.org/10.1007/s11102-019-00958-8.
- Parent AD, Perkins E. The hypothalamus. In: Haines DE, Mihailoff GA, editors. Fundamental neuroscience for basic and clinical applications. 5th ed. Elsevier; 2018. p. 442–57.
- Scully KM, Rosenfeld MG. Pituitary development: regulatory codes in mammalian organogenesis. Science. 2002;295(5563):2231–5. https://doi.org/10.1126/science.1062736.
- Zhu X, Gleiberman AS, Rosenfeld MG. Molecular physiology of pituitary development: signaling and transcriptional networks. Physiol Rev. 2007;87(3):933–63. https://doi.org/10.1152/ physrev.00006.2006.
- Davis SW, Ellsworth BS, Peréz Millan MI, Gergics P, Schade V, Foyouzi N, et al. Pituitary gland development and disease: from stem cell to hormone production. Curr Top Dev Biol. 2013;106:1–47. https://doi.org/10.1016/b978-0-12-416021-7.00001-8.
- Osamura RY, Lopes MBS, Grossman A, Kontogeorgos G, Trouillas J. Tumours of the pituitary gland. Introduction. In: Lloyd RV, Osamura RY, Klöppel G, Rosai J, editors. WHO classification of tumours of endocrine organs. 4th ed. Lyon: IARC press; 2017. p. 13.
- Osamura RY, Grossman A, Korbonits M, Kovacs K, Lopes MBS, Matsuno A, Trouillas J. Pituitary adenoma. In: Lloyd RV, Osamura RY, Klöppel G, Rosai J, editors. WHO classification of tumours of endocrine organs. 4th ed. Lyon: IARC press; 2017. p. 14–8.
- Syro LV, Rotondo F, Ramirez A, Di Ieva A, Sav MA, Restrepo LM, et al. Progress in the diagnosis and classification of pituitary adenomas. Front Endocrinol (Lausanne). 2015;6:97. https://doi.org/10.3389/fendo.2015.00097.
- Murray JF. LTP. Anterior pituitary: somatotrophs (GH) and lactotrophs (PRL). In: Litwack G, editor. Hormonal Signaling in biology and medicine: comprehensive modern endocrinology. London: Academic Press Elsevier; 2019. p. 171–201.
- Heaney AP, Melmed S. Molecular targets in pituitary tumours. Nat Rev Cancer. 2004;4(4):285–95. https://doi.org/10.1038/nrc1320.
- Vidal S, Horvath E, Kovacs K, Cohen SM, Lloyd RV, Scheithauer BW. Transdifferentiation of somatotrophs to thyrotrophs in the pituitary of patients with protracted primary hypothyroidism. Virchows Arch. 2000;436(1):43–51.
- Allaerts W, Vankelecom H. History and perspectives of pituitary folliculo-stellate cell research. Eur J Endocrinol. 2005;153(1):1–12. https://doi.org/10.1530/eje.1.01949.
- Pires M, Tortosa F. Update on pituitary Folliculo-stellate cells. International archives of endocrinology. Clin Res. 2016;2 https://doi.org/10.23937/2572-407X.1510006.
- 25. Nowell PC. The clonal evolution of tumor cell populations. Science. 1976;194(4260):23-8.

- Greaves M, Maley CC. Clonal evolution in cancer. Nature. 2012;481(7381):306–13. https:// doi.org/10.1038/nature10762.
- Merlo LM, Pepper JW, Reid BJ, Maley CC. Cancer as an evolutionary and ecological process. Nat Rev Cancer. 2006;6(12):924–35. https://doi.org/10.1038/nrc2013.
- 28. Hanahan D, Weinberg RA. The hallmarks of cancer. Cell. 2000;100(1):57-70.
- Almendro V, Marusyk A, Polyak K. Cellular heterogeneity and molecular evolution in cancer. Annu Rev Pathol. 2013;8:277–302. https://doi.org/10.1146/annurev-pathol-020712-163923.
- Marusyk A, Polyak K. Tumor heterogeneity: causes and consequences. Biochim Biophys Acta. 2010;1805(1):105–17. https://doi.org/10.1016/j.bbcan.2009.11.002.
- 31. Caimari F, Korbonits M. Novel genetic causes of pituitary adenomas. Clin Cancer Res. 2016;22(20):5030–42. https://doi.org/10.1158/1078-0432.ccr-16-0452.
- Syro LV, Rotondo F, Kovacs K, Korbonits M. Clinicopathologic features of familial pituitary adenomas. Diagn Histopathol. 2016;22(3):85–91. https://doi.org/10.1016/j. mpdhp.2016.02.004.
- Syro LV, Builes CE, Di Ieva A, Sav A, Rotondo F, Kovacs K. Improving differential diagnosis of pituitary adenomas. Expert Rev Endocrinol Metab. 2014;9(4):377–86. https://doi.org/1 0.1586/17446651.2014.922412.
- Mete O, Lopes MB. Overview of the 2017 WHO classification of pituitary Tumors. Endocr Pathol. 2017; https://doi.org/10.1007/s12022-017-9498-z.
- Lopes MBS. The 2017 World Health Organization classification of tumors of the pituitary gland: a summary. Acta Neuropathol. 2017;134(4):521–35. https://doi.org/10.1007/ s00401-017-1769-8.
- 36. Trouillas J, Roy P, Sturm N, Dantony E, Cortet-Rudelli C, Viennet G, et al. A new prognostic clinicopathological classification of pituitary adenomas: a multicentric case-control study of 410 patients with 8 years post-operative follow-up. Acta Neuropathol. 2013;126(1):123–35. https://doi.org/10.1007/s00401-013-1084-y.
- 37. Asa SL, Casar-Borota O, Chanson P, Delgrange E, Earls P, Ezzat S, et al. From pituitary adenoma to pituitary neuroendocrine tumor (PitNET): an international pituitary pathology club proposal. Endocr Relat Cancer. 2017;24(4):C5–c8. https://doi.org/10.1530/erc-17-0004.
- Syro LV, Rotondo F, Serna CA, Ortiz LD, Kovacs K. Pathology of GH-producing pituitary adenomas and GH cell hyperplasia of the pituitary. Pituitary. 2016:1–9. https://doi.org/10.1007/ s11102-016-0748-8.
- Drummond J, Roncaroli F, Grossman AB, Korbonits M. Clinical and pathological aspects of silent pituitary adenomas. J Clin Endocrinol Metabol. 2018;104(7):2473–89. https://doi. org/10.1210/jc.2018-00688.
- Cooper O, Melmed S. Subclinical hyperfunctioning pituitary adenomas: the silent tumors. Best Pract Res Clin Endocrinol Metab. 2012;26(4):447–60. https://doi.org/10.1016/j. beem.2012.01.002.
- Cuevas-Ramos D, Carmichael JD, Cooper O, Bonert VS, Gertych A, Mamelak AN, et al. A structural and functional acromegaly classification. J Clin Endocrinol Metab. 2015;100:122–31. https://doi.org/10.1210/jc.2014-2468.
- 42. Syro LVRF, Serna CA, Restrepo LM, Kovacs K. Overproduction of hormones by pituitary tumors. In: Litwack G, editor. Hormonal signaling in biology and medicine: comprehensive modern endocrinology. London: Academic Press Elsevier; 2019. p. 655–65.
- Rotondo F, Kovacs K, Lloyd RV. The role of molecular pathology in the classification and clinicopathologic evaluation of pituitary neoplasms. Diagn Histopathol. 2018;24(3):95–103. https://doi.org/10.1016/j.mpdhp.2018.02.003.
- 44. Kleinschmidt-DeMasters BK, Lopes MBS. Histologic features of pituitary adenomas and sellar region masses. In: Perry A, Brat DJ, editors. Practical surgical neuropathology: a diagnostic approach. 2nd ed. Elsevier; 2018. p. 453–91.
- 45. Syro LV, Rotondo F, Cusimano MD, Di Ieva A, Horvath E, Restrepo LM, et al. Current status on histological classification in Cushing's disease. Pituitary. 2015;18(2):217–24. https://doi. org/10.1007/s11102-014-0619-0.

- Di Ieva A, Davidson JM, Syro LV, Rotondo F, Montoya JF, Horvath E, et al. Crooke's cell Tumors of the pituitary. Neurosurgery. 2015; https://doi.org/10.1227/NEU.00000000000657.
- 47. Ben-Shlomo A, Cooper O. Silent corticotroph adenomas. Pituitary. 2018;21(2):183–93. https://doi.org/10.1007/s11102-018-0864-8.
- Nishioka H, Inoshita N, Mete O, Asa SL, Hayashi K, Takeshita A, et al. The complementary role of transcription factors in the accurate diagnosis of clinically nonfunctioning pituitary adenomas. Endocr Pathol. 2015; https://doi.org/10.1007/s12022-015-9398-z.
- Nishioka H, Inoshita N. New WHO classification of pituitary adenomas (4th edition): assessment of pituitary transcription factors and the prognostic histological factors. Brain Tumor Pathol. 2018;35(2):57–61. https://doi.org/10.1007/s10014-017-0307-7.
- Saeger W, Ludecke DK, Buchfelder M, Fahlbusch R, Quabbe HJ, Petersenn S. Pathohistological classification of pituitary tumors: 10 years of experience with the German pituitary tumor registry. Eur J Endocrinol. 2007;156(2):203–16. https://doi.org/10.1530/eje.1.02326.
- Raverot G, Burman P, McCormack A, Heaney A, Petersenn S, Popovic V, et al. European Society of Endocrinology Clinical Practice Guidelines for the management of aggressive pituitary tumours and carcinomas. Eur J Endocrinol. 2018;178(1):G1–g24. https://doi.org/10.1530/ eje-17-0796.
- McCormack AI, Dekkers O, Petersenn S, Popovic V, Trouillas J, Raverot G, et al. Treatment of aggressive pituitary tumours and carcinomas: results of a European Society of Endocrinology (ESE) survey 2016. Eur J Endocrinol. 2018; https://doi.org/10.1530/eje-17-0933.
- Di Ieva A, Rotondo F, Syro LV, Cusimano MD, Kovacs K. Aggressive pituitary adenomas diagnosis and emerging treatments. Nat Rev Endocrinol. 2014;10(7):423–35. https://doi. org/10.1038/nrendo.2014.64.
- 54. Dai C, Sun B, Liu X, Bao X, Feng M, Yao Y, et al. O-6-Methylguanine-DNA methyltransferase expression is associated with pituitary adenoma tumor recurrence: a systematic meta-analysis. Oncotarget. 2017;8(12):19674–83. https://doi.org/10.18632/oncotarget.14936.
- Micko ASG, Wohrer A, Hoftberger R, Vila G, Marosi C, Knosp E, et al. MGMT and MSH6 immunoexpression for functioning pituitary macroadenomas. Pituitary. 2017;20(6):643–53. https://doi.org/10.1007/s11102-017-0829-3.

Check for updates

Physiology of the Pituitary Hormone Secretion

Antonio C. Fuentes-Fayos, Emilia Alors-Perez, Juan M. Jiménez-Vacas, Vicente Herrero-Aguayo, Prudencio Sáez-Martínez, Juan L. Lopez-Cánovas, María C. Vázquez-Borrego, Justo P. Castaño, Rhonda D. Kineman, Manuel D. Gahete, and Raúl M. Luque

Abbreviations

Acetylcholine
Adenylate cyclase
Adrenocorticotropic hormone
Angiotensin II
Antidiuretic hormone
Arginine-vasopressin
Brain-derived neurotrophic factor
Central nervous system

The chapter has been endorsed by **Dr Giampaolo Trivellin**, giampaolo.trivellin@humanitasresearch.it, Istituto Clinico Humanitas IRCCS, Rozzano – Milan, Italy

A. C. Fuentes-Fayos · E. Alors-Perez · J. M. Jiménez-Vacas · V. Herrero-Aguayo P. Sáez-Martínez · J. L. Lopez-Cánovas · M. C. Vázquez-Borrego · J. P. Castaño M. D. Gahete · R. M. Luque (\boxtimes)

Maimonides Biomedical Research Institute of Cordoba (IMIBIC), Cordoba, Spain

Department of Cell Biology, Physiology and Immunology, University of Cordoba, Cordoba, Spain

CIBER Physiopathology of Obesity and Nutrition (CIBERobn), Cordoba, Spain e-mail: b22fufaa@uco.es; b12alpee@uco.es; b12jivaj@uco.es; b22heagv@uco.es; b32samap@uco.es; b32locaj@uco.es; justo@uco.es; bc2gaorm@uco.es; raul.luque@uco.es

R. D. Kineman

Section of Endocrinology, Diabetes, and Metabolism, Department of Medicine, University of Illinois at Chicago, Chicago, IL, USA

Research and Development Division, Jesse Brown Veterans Affairs Medical Center, Chicago, IL, USA e-mail: kineman@uic.edu

PRL	Prolactin
РКА	Protein kinase A
РКС	Protein kinase C
PitNETs	Pituitary neuroendocrine tumors
RFRPs	RF-related peptides
5-HT	Serotonin
SST	Somatostatin
SSTR	Somatostatin Receptor
SRIF	Somatotropin-release inhibitory factor
SF1	Steroidogenic factor-1
TRH	Thyrotropin-releasing hormone
T4	Thyroxine
Т3	Triiodothyronine
TSH	Thyrotropin-stimulating hormone
VIP	Vasoactive intestinal peptide
VPAC1	Vasoactive intestinal polypeptide receptor 1

2.1 Introduction

The pituitary is considered one of the most important endocrine glands in the organism since it is responsible to modulate a series of critical biological processes and functions, by regulating the physiology of different tissues, organs, and other endocrine glands throughout the whole organism. Specifically, the pituitary participates in the control of multiple important patho-physiological functions such as growth, reproduction, metabolism, or stress response.

As extensively explained in Chap. 1, this complex gland is located at the base of the brain, wherein it displays a privileged location at the interphase between the central nervous system (CNS) and the rest of the organism. Indeed, this gland is strongly connected in an anatomical and physiological manner with one of the most relevant CNS areas, the hypothalamus. The pituitary is organized in two structurally and functionally distinct areas named adenohypophysis and neurohypophysis. The adenohypophysis (*adeno*, meaning "glandular") consists of three regions: *pars distalis, pars intermedia,* and *pars tuberalis.* The *pars distalis* (or *anterior pituitary gland*) occupies the major portion (70%) of the adenohypophysis. The *pars tuberalis* is a group of cells surrounding the pituitary stalk. The neurohypophysis, *pars nervosa* or posterior lobe, is not a glandular portion but represents a collection of axonal projections from the hypothalamus that serves as a site of secretion for neurohypophysial hormones.

In this context, the main aspects related to the physiology of the pituitary gland rely on the hormones that are produced and secreted from each of these different areas, which exhibit a plethora of pleiotropic functions to regulate the vast majority of endocrine glands, tissues, and organs in the body. In the anterior pituitary, there are five different specialized endocrine secretory cell types: somatotrophs, lactotrophs, corticotrophs, gonadotrophs, and thyrotrophs. These cells are responsible for the synthesis and secretion of growth hormone (GH), prolactin (PRL), adrenocorticotropic hormone (ACTH), luteinizing hormone (LH), and follicle-stimulating hormone (FSH), and thyrotropin-stimulating hormone (TSH), respectively [1]. The main cell type found in the *pars intermedia* is the melanotrope cell, which secretes melanocyte-stimulating hormone or MSH. However, the *pars intermedia* is typically involuted in adulthood, being insignificant or entirely absent [2]. In the case of the neurohypophysis, the axonal projections of neurons located at the supraoptic and paraventricular hypothalamic nuclei accumulate vesicles with arginine-vasopressin (AVP) and oxytocin (OT), which are released directly from the hypothalamus to the bloodstream [3].

Some of these hormones, once are released into the bloodstream, act directly on different target tissues. In fact, GH exerts modulatory actions on the growth of all tissues of the organism, and PRL is responsible for the development of glandular *acini* in the mammary glands and the regulation of human milk proteins synthesis. Similarly, AVP acts directly on kidney and arterioles increasing peripheral vascular resistance and raising arterial blood pressure, and OT directly regulates myoepithe-lial cells contraction of the mammary glands as well as of the uterus, controlling the contractions during lactation and childbirth. On the other hand, ACTH, FSH, LH, and TSH are trophic hormones that act by stimulating the production and release of hormones in specific peripheral endocrine glands such as the adrenal glands (ACTH), gonads (LH and FSH), and the thyroid gland (TSH) [1].

In any case, the synthesis and/or production of these hormones at the pituitary level is tightly controlled by a plethora of central and peripheral factors that finely interact to modulate the physiologic production of these hormones (Fig. 2.1). It is



Fig. 2.1 Representative diagram summarizing the main regulators involved in the modulation of the pituitary hormone secretion of different pituitary cells
now known that each pituitary cell type exhibits a particular pattern of receptors for different signals and that the appropriate sensing, transmission, and integration of all these inhibitory and stimulatory signals is crucial to adapt the circulating levels of these hormones to the physiological necessities of the organism.

2.2 Anterior Pituitary

2.2.1 Somatotrophs

Somatotrophs, which represent the most abundant secretory cell type in the anterior pituitary, are responsible for the synthesis and release of GH. Once secreted into the bloodstream, GH acts directly through the GH receptor (GHR), and/or indirectly, by stimulating the release of insulin-like growth factor 1 (IGF1) to promote linear growth in children [4]. In addition, GH has been shown to exhibit many metabolic actions including enhanced protein synthesis, lipolysis and lipid oxidation, retention of water, phosphate and sodium and acting as an insulin antagonist [5].

GH production begins early in fetal life and its levels vary during childhood, peaking during puberty and lowering in elderly. GH is secreted in a circadian pulsatile fashion, wherein the interplay between the hypothalamic hypo-physiotropic hormones GH-releasing hormone (GHRH) and its counteracting inhibitor somatostatin (SST) plays a crucial role. Indeed, the hypothalamic control of GH secretion in mammals has long been considered as a classic paradigm of the "dual control" system of pituitary hormone secretion mentioned above. However, the daily peaks of GH secretion also vary with age and are profoundly influenced by several factors including gender, sleep, obesity, nutrition, exercise, and metabolic disorders [6]. In fact, it is now accepted that the pulsatile secretion of GH can be also modulated by diverse factors residing in the pituitary or in other regions of the CNS, as well as by additional factors arriving from peripheral organs and tissues [7], as described below.

2.2.1.1 Growth Hormone Releasing Hormone (GHRH)

GHRH is a 44 amino acid amidated polypeptide produced and secreted by the terminals of neurons located in the hypothalamic arcuate nucleus [8]. GHRH leads to the stimulation of GH production, via the hypophyseal portal system, by increasing *GH* gene expression and inducing GH release by the somatotrophs. Indeed, GHRH has been long recognized as the main stimulator of GH secretion in humans and, together with somatostatin, the central modulators of GH pulsatile secretion. In addition, GHRH also acts as a growth-stimulatory factor of somatotroph cells inasmuch as GHRH absence produces anterior pituitary hypoplasia [9]. This hypothalamic hypo-physiotropic hormone acts through a seven trans-membrane G protein-coupled receptor (GPCR) named GHRHR (GH-releasing hormone receptor), which is expressed at high levels in somatotroph cells. GHRHR is crucial for the actions of GHRH and, indeed, mutations in this receptor have been largely reported to be associated with an impairment of the physiological development of pituitary somatotrophs [10], producing isolated GH deficiency, the most common pituitary hormone deficiency [11]. GHRH binding to GHRHR stimulates GH release by activating multiple signaling pathways, including adenylate cyclase (AC) and increasing cyclic adenosine monophosphate (cAMP) concentration, which leads to the increase of protein kinase A (PKA) activity followed by CREB activation [1, 12].

2.2.1.2 Ghrelin (GHRL)

Ghrelin, a 28 amino acid octanoylated peptide, was initially isolated from human and rat stomachs [13]. Subsequently, ghrelin expression was also found in multiple organs and tissues including the pituitary and hypothalamus [14]. Ghrelin can be found in two circulating isoforms, acylated (octanoylated) and non-acylated (which seems to lack endocrine actions but exerts non-endocrine actions) [15]. The acylated form is the endocrine functional form of ghrelin, which has been shown to act at the pituitary level but also at CNS level as an orexigenic factor. Particularly, ghrelin was rapidly recognized as a potent GH-releasing factor through its binding to the previously identified orphan GH-secretagogue receptor 1a (GHSR1a), producing a robust GH release, comparable or even higher to that exerted by the classical GH stimulator GHRH [16, 17]. The binding of acylated ghrelin to GHSR1a triggers the activation of multiple signal transduction pathways, including phospholipase C (PLC), protein kinase C (PKC), PKA, intracellular and extracellular Ca²⁺ or mitogen-activated protein kinases (MAPKs), resulting in a significant elevation of GH transcription (elicited by specific transcription factors) and GH secretion (as the result of Ca²⁺ mobilization, among other signals) [1, 18]. In addition, acylated ghrelin is able to induce an indirect, hypothalamic-mediated mechanism involving an increase in GHRH and a weak inhibition of SST neurons, promoting a complete program to stimulate GH release [19].

2.2.1.3 Somatostatin (SST)

SST or somatotropin-release inhibitory factor (SRIF) is a cyclic peptide derived from a 116 amino acid precursor that can generate two different cyclic variants by alternative post-translational processing, SST-14 and SST-28. Both peptides contain a disulfide bridge, have a cyclic structure as well as high affinity for SST receptor subtypes, and they are capable to inhibit GH release [20]. SST regulates different aspects of the endocrine system and controls neurotransmission and cell proliferation. Specifically, SST is a short half-life inhibitor of the secretion of several hormones including GH through the modulation of somatotroph cells. In particular, SST is considered the main negative regulator of GH secretion and its inhibitory function is conserved during vertebrate evolution. In mammals, SST is essential to establish and maintain pulsatility of GH secretion [21, 22]. In this sense, it has been proposed that in humans, the role of SST in the control of GH secretion seems to be mainly circumscribed to the adjustment of the magnitude of its basal and pulsatile release, while it may not be involved in the regulation of the generation of GH pulsatility [6]. Thus, although SST is undoubtedly the main inhibitory signal for GH secretion in all vertebrate groups, it is not the only one as it will be explained below, and its role can be more complex than it was initially envisioned [20]. SST exerts its actions via interaction with different GPCR subtypes (SSTR1-5 and their splicing variants, such as SST5TMD4 or SST5TMD5) [20]. SST binding to these receptors elicits the modulation of a constellation of downstream pathways, including AC, protein phosphatases, cGMP dependent protein kinases, and calcium and other ion channels [1], resulting in changes in intracellular calcium and cAMP concentrations, and the subsequent reduction in GH secretion [20].

2.2.1.4 Cortistatin (CST)

CST is a peptide with a strong structural and functional similarity with SST. CST is predominantly expressed in inhibitory neurons of the cerebral cortex but, as well as SST, it is distributed and expressed in a wide variety of tissues, including the pituitary gland [23, 24]. Human CST is produced from the 105 amino acid precortistatin, which can be cleaved into CST-17 and CST-29. CST-17, the mainly active peptide, shares a high homology with SST-14. This strong structural similarity explains their similar capacity to bind the same family of receptors (SSTRs) and to recruit the same downstream pathways [1, 25, 26]. However, CST is also able to bind other receptors such as GHSR1a [27, 28] or MRGX2 [29], which may explain some of the dissimilar actions exerted by CST such as the induction of slow wave sleep. In any case, at the pituitary level and specifically in somatotrophs, CST can inhibit GH release through the activation of SST receptors in young males and, indeed, CST and SST show equal inhibition of the GH release induced by GHRH, ghrelin, and synthetic analogs [30]. These effects of CST seem to be independent of GHSR1a binding, suggesting a predominant role of SST receptors on the known inhibitory actions of CST in this cell type [31].

2.2.1.5 Neuronostatin (NST)

NST is a 13 amino acid neuropeptide encoded by the somatostatin gene and derived from the alternative proteolytic processing of the same 116 amino acid precursor that gives rise to SST [32]. NST is known to exert important physiological and metabolic actions in diverse tissues. Conversely, the direct biological effect on anterior pituitary function, specifically in somatotrophs, has not been extensively explored. Recent studies have shown that NST functions seem to be associated with the inhibition of basal GH and LH release. However, this hormone seems to exert different actions from those shown by SST, especially related to the modulation of hormone (GH and LH) release and gene expression in this gland [33]. NST may exert its actions via alternative GPCRs, and, particularly. Through GPR107, which is a mechanism of action clearly different from that exhibited by SST and CST [34].

2.2.1.6 Kisspeptin (KISS1)

Kisspeptin is an amidated neurohormone encoded by the *KISS1* gene that is widely known for its role in the modulation of sexual development processes, such as puberty and ovulation, through the activation of gonadotropin-releasing hormone (GNRH) neurons excitability. The product of *KISS1* gene is a 145 amino acid precursor that can generate four possible modified peptides with different lengths, 54,

14, 13, or 10 amino acids. Kisspeptin peptides can bind and activate a GPCR named GPR54 or KISS1R [35]. All the kisspeptin peptides have the same efficacy and affinity for GPR54, despite the dissimilar length. This binding leads to the activation of multiple signaling pathways, including PLC, PKC, MAPK, and intracellular Ca²⁺ mobilization, to transduce the signal, resulting in an elevation in GH secretion driven by GH gene transcription activation and GH release induction [1, 36]. The expression of kisspeptin and its receptor has been found in several tissues, including the pituitary gland, suggesting a potential role of this neurohormone in the control of additional hormones. However, the potential action of kisspeptin on somatotroph function is still controversial. Indeed, *in vivo* studies have not found any effect in GH pulses after kisspeptin administration, while in vitro kisspeptins can induce a short stimulation of GH, although lower than induced by key GH stimulators [36, 37].

2.2.1.7 Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP)

PACAP is a member of the VIP/secretin/glucagon/GHRH/GIP superfamily of peptides. It is an amidated peptide with two molecular isoforms, PACAP38 and PACAP27. These peptides bind to the GPCR PACAP type-1, which stimulates AC and subsequently increases cAMP levels in target cells [38]. PACAP is highly conserved throughout evolution and it is a potent GH stimulator in different lower vertebrate species; however, in humans, GH secretion stimulated by PACAP is very weak compared with that produced by GHRH [39]. In general, in mammals, data concerning the PACAP effect on GH release are controversial. Some studies found a stimulatory effect of PACAP, while other reports indicate that PACAP has no effect on GH secretion [40]. Interestingly, PACAP and GHRH are encoded by two closely related genes [41]. In spite of being two highly related peptides, their role on GH regulation across vertebrate evolution seems to have diverged in mammals and, while GHRH is critical for GH release, PACAP seems to play only a secondary role in the regulation of the somatotropic axis [7].

2.2.1.8 Melatonin (MT)

MT or N-acetyl-5-methoxy tryptamine is a non-peptide hormone produced by the pineal gland [42]. MT is the main driver of circadian rhythms in humans, being secreted overnight. In fact, a close relationship between MT and pituitary secretions has been proposed since the secretion patterns of some pituitary hormones follow a circadian cycle [43]. In addition, the fact that the melatonin receptor MT1 is expressed in the pituitary gland reinforces this contention [44, 45]. However, although results obtained in primate models indicate that MT directly regulates somatotroph function [45] and that MT might substantially contribute to define the daily patterns of GH, the observations made in human are controversial. In human, the age of the individual and the administration route seem to be crucial for the effect of MT on GH release. Until puberty, there seems to be a negative correlation between MT and GH secretion, promoting a diurnal growth; however, this relation-ship seems to be the opposite in adulthood [46–48]. In contrast, other studies have

shown that MT may not influence GH release [49] or that this effect may be different in response to acute or chronic MT administration [50, 51]. In any case, the effects of MT on GH may be mediated by MT1 through the modulation of AC/PKA and extra-/intracellular calcium pathways requiring activation of the PLC route [1, 45].

2.2.1.9 Thyrotropin-Releasing Hormone (TRH)

TRH is a tripeptide (glu-his-pro-NH2) hormone primarily produced in the paraventricular nucleus of the hypothalamus, and mainly involved in the stimulation of TSH release from thyrotroph cells [7]. However, this hormone could also exert additional actions in the modulation of other pituitary secretions. Indeed, TRH has been demonstrated to stimulate GH release in different mammalian species [7]. In humans, TRH actions have been reported in cell cultures from acromegalic patients where TRH induced GH release, while this effect is less evident in normal subjects [52]. Although these data confirm the role of TRH controlling the normal physiology of GH-producing cells, they point out that TRH has lost part of its ability to act as a primary stimulus of the somatotropic axis during the evolution. In any case, TRH actions seem to be dependent on the calcium influx through L-type calcium channels, which may be associated with PKC activation [52].

2.2.1.10 Neuropeptide Y (NPY)

NPY is a 36 amino acid peptide that exhibits widespread physiological functions in the CNS and periphery, being preferentially expressed in neurons located at the hypothalamic arcuate nucleus [53]. NPY is able to bind multiple receptors that belong to the GPCR family as do other GH-release regulators [54]. In mammals, NPY seems to elicit a stimulatory effect on GH secretion in somatotroph cells [7]. However, in healthy humans, NPY administration did not alter GH release when administered to young men [55]. Therefore, more studies should be implemented in human in order to unveil the real potential of NPY as a GH regulator.

2.2.1.11 Other Systemic/Potential Regulators

Besides all the GH regulators mentioned above, there are additional hormones and molecules that could influence the production and normal release of GH, including leptin, resistin [56, 57], estrogens [58], glucocorticoids [59, 60], and opioids [61], which have demonstrated a role as positive modulators of GH release in somato-trophs. On the contrary, IGF1, insulin [60, 62, 63], thyroid hormones [64], obestatin [65], adiponectin [57], free fatty acids [66, 67], and endothelin [1] have been shown to have a role inhibiting GH production in somatotrophs. These additional regulators are also extremely important because they are able to modify the balance between the main inhibitory and stimulatory signals mentioned above. In general, these other systemic and potential regulators are considered weak modulators of GH release; however, they could play an important role in the pathophysiology of this gland, particularly in the somatotroph cells.

2.2.2 Lactotrophs

Lactotrophs are acidophilic cells that comprise about 20% of all cells in the anterior pituitary gland. These cells mainly produce prolactin (PRL), a peptide hormone that is primarily involved in the maturation of mammary glands and the secretion of milk in association with OT, estrogen, progesterone, glucocorticoids, and other factors. However, PRL has several additional effects in both sexes, including the regulation of metabolic processes, the immune system, and the pancreatic development [68]. PRL is secreted from lactotroph cells in response to eating, mating, estrogen treatment, ovulation, and/or nursing, among other stimuli. In particular, PRL is secreted in pulses in between these events by the fine interplay between different modulators as described below [68–70].

2.2.2.1 Dopamine (DA)

DA, a classical neurotransmitter produced by different areas of the brain, is the principal regulator involved in the inhibition of PRL expression and secretion. PRL regulation by DA is mainly mediated by its binding to dopamine receptor subtype-2 (DRD2), which is the predominant dopamine receptor in lactotrophs [71, 72]. Several studies have reported that the reduction of intracellular cAMP levels represents the main mechanism by which DA suppresses PRL expression, which can ultimately lead to a dysregulation of ion channel function and a desensitization of the GPCRs that control PRL release [70]. Importantly, due to the involvement of ion channels (Ca²⁺ signaling) in this process, PRL secretion is compromised just after few seconds of DA exposure [73]. Strikingly, a slight increase of PRL secretion in lactotrophs has been reported in response to especially low concentration of DA (<0.1 nM) [74], which has been explained by different theories (e.g., prevalence of the short DA receptor isoform [75], or involvement of the Gia3 protein [76], among others). However, the stimulatory effect on PRL of DA remains controversial while the inhibitory role of this hormone has been well established and deeply demonstrated.

2.2.2.2 Thyrotropin Releasing Hormone (TRH)

As indicated above, TRH is secreted by the hypothalamus and was originally named for its action on the modulation of TSH release from the pituitary. However, it was lately shown that it could also regulate other pituitary cells, such as lactotroph cells, wherein it plays a role as one of the main stimulators of PRL secretion [77]. In fact, it has been shown that the same concentration of TRH is able to induce TSH and PRL secretion in human pituitary cells, and that TRH blockade impedes PRL secretion [77]. In the context of hypothyroidism, the low level of thyroid hormones leads to a compensatory increase of TRH. This may lead to an excess of PRL causing hyperprolactinemia [78, 79]. In lactotroph cells, TRH binds and activates the PRL receptor (PRL-R) and recruits the Gq protein that stimulates inositol phospholipid production, which activates both the PKC pathway and the release of Ca²⁺ from different compartments [77, 80]. The activation of the PKC pathway may lead to MAPK activation via ERK, but it has been shown that this pathway may be also enhanced by TRH in a PKC-independent way [77, 80]. Additionally, it has been shown that TRH secretion is pulsatile and this can affect PRL secretion, in that PRL is more potently secreted when TRH pulse is more frequent [77]. Interestingly, DA inhibits the activation of the cAMP pathway, which is independent of TRH. However, DA may also inhibit the action of TRH in lactotroph cells, via Ca²⁺ release [81].

2.2.2.3 Ghrelin (GHRL)

As it has been described before, ghrelin is a hormone peptide of 28 amino acids whose main function is to increase GH secretion through its binding to the GHSR1a receptor. However, it has been demonstrated that ghrelin may also participate in the secretion of PRL as a stimulator factor [82]. Although the mechanisms underlying this event are not fully understood, it seems clear that ghrelin acts directly on lacto-troph cells, since it may produce PRL release in dispersed normal pituitary cells [45, 83]. However, this regulation is not under the control of the changes of menstrual cycle in women, being independent of the estrogens [84]. Moreover, ghrelin seems to be less important than TRH in the stimulation of PRL secretion because: (a) the effect of ghrelin is lower than TRH, and (b) the combination of ghrelin and TRH did not show an additive/synergic effect but produced the same effect than TRH alone [85].

2.2.2.4 Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP)

The action of PACAP in lactotrophs is closely related to the cAMP and inositol phosphate pathways, which seem to regulate PKA and MAPK, respectively, leading to the modulation of PRL expression [68]. Similar to that found for other modulators, PACAP has been linked to PRL regulation, but its role is controversial. First, it was thought that PACAP did not participate in the regulation of PRL release; however, further studies demonstrated that PACAP mainly increases PRL secretion in rat pituitary cells [86]. Specifically, in vitro studies demonstrated that both PACAP and its receptor PAC1R display a stimulatory effect on PRL secretion in GH3 cells [86]. Additionally, intravenous PACAP injection may regulate PRL secretion in mammals, including human [80]. Nevertheless, PACAP did not exert similar effects in female lactating rats with actively nursing pups [86]. Moreover, PACAP knockout mice exhibit significantly reduced levels of PRL [87]. Finally, PAC1R, which is expressed in the entire anterior pituitary, but mainly present in lactotroph cells, may act independently of PACAP in improving the effect of TRH. Similarly, PACAP also increases the expression of the TRH receptor, demonstrating a clear relationship between both regulatory axes [88].

2.2.2.5 Melatonin (MT)

MT is a crucial factor in the regulation of PRL secretion, by adjusting the pattern of PRL secretion to photoperiod and conferring a circadian profile [89, 90]. Indeed, MT seems to be fundamental to modulate PRL secretion in response to sleep, but, in addition, it seems that MT may also regulate the secretion of PRL in response to sleep-independent changes caused by bright light [91]. *In vitro* studies [45, 92] have suggested that MT exerts direct stimulatory effects on PRL release, which could

contribute to the nocturnal rise of this hormone observed in humans *in vivo* [93] in that the nocturnal rise in PRL release is preceded by an increase in MT [94]. Consistently, exogenous administration of MT can enhance the nocturnal pulsatile secretion of PRL in normally cycling women [94], further supporting the contention that MT can enhance PRL secretion in humans *in vivo*.

2.2.2.6 Somatostatin (SST)

SST has been shown to play an inhibitory role on PRL secretion, which is less pronounced than the effect exerted on GH secretion. Specifically, it has been reported that SST, but not NST, is able to reduce *in vitro* the spontaneous release of PRL [33, 95]. Consistently, an inhibition of basal and TRH-induced PRL release of pituitary cells from adult rats in response to SST has been described [96], although it has been suggested that regulation of PRL secretion by SST is estrogen-dependent [97]. Moreover, several studies in human PRL-secreting pituitary neuroendocrine tumors (PitNETs) have shown that SSTR1R- and SSTR5-specific agonists and pasireotide effectively inhibited PRL secretion (but not PRL expression) [98-102], while octreotide does not alter PRL secretion [102], likely due to the fact that SSTR5 is not highly expressed in this type of tumors [99]. Mechanistically, SST-dependent inhibition of PRL secretion could be explained by a modulation in Ca²⁺ signaling and a reduction in AC activity, inasmuch as SST fails to inhibit PRL release in the presence of the Ca²⁺ ionophore A23187 [103]. Therefore, SST inhibits PRL secretion through SSTR-dependent pathways, which appears to be receptor subtype-specific and sensitive to the presence of estrogens.

2.2.2.7 Cortistatin (CST)

The potential role of CST in the regulation of PRL secretion has been a matter of debate from its discovery. Initial studies have reported that CST, as well as SST, can inhibit PRL release from prolactinomas *in vivo* and *in vitro*, mainly due to the effect of its binding to SSTR5 [104, 105]. However, additional studies have also shown that CST can actually stimulate PRL release in rats [106] and mice [28]. Indeed, this latter study suggested that endogenous CST can indeed be considered a physiological stimulator of lactotroph function, inasmuch as circulating PRL levels are markedly suppressed in CST-KO mice [28].

2.2.2.8 Other Systemic/Potential Regulators

In addition to the classical regulators of PRL secretion described above, additional factors with the capacity to finely tune the concentration of PRL must be considered. This is the case of estrogens, which may increase PRL release. In fact, a positive regulation of PRL secretion by estradiol treatment has been reported in pituitary cell cultures from monkeys [107]. Additionally, it has been published that ET1, a peptide involved in the control of the blood pressure by blood vessel constriction, increased plasma PRL levels in men [108], likely due to modulation of Ca²⁺ signaling [109]. Moreover, different adipokines have been shown to play an important role regulating PRL secretion. In fact, both adiponectin and leptin directly increase PRL release from primary pituitary cell cultures derived from primate species by

modulation of Ca²⁺ and PLC/PKC signaling pathways [57]. On the other hand, besides DA, other inhibitors of PRL secretion have been identified, such as IGF1 [110] or GNRH-associated peptide (GAP), which were reported to be potent inhibitors of PRL release in primary cultures from human and rat pituitary tissues, respectively [111]. Therefore, there are many factors to be considered in the regulation of PRL expression and secretion that may act simultaneously and/or sequentially in lactotroph cells.

2.2.3 Gonadotrophs

Gonadotroph cells arise from the ventral pituitary and require the expression of steroidogenic factor-1 (SF1) and GATA2 for differentiation. These cells constitute 10% of all adenohypophysial cells and are spread out throughout the *pars distalis* and *pars tuberalis*. These cells produce two types of hormones known as gonado-tropins, FSH and LH, which are heterodimeric glycoproteins composed by a common α subunit (CGA) and a hormone-specific β subunit [112, 113]. Gonadotropins play an essential role in the normal mammalian sexual maturation and reproduction, acting at gonad levels by binding to GPCRs named FSHR and LHR, respectively [114]. As mentioned above regarding other anterior pituitary hormones, the precise control of gonadotrophs is also finely regulated by hypothalamic and peripheral signals [80], as described below.

2.2.3.1 Gonadotropin-Releasing Hormone (GNRH)

GNRH is a decapeptide produced by hypothalamic neurons and secreted into the hypophyseal portal circulation to directly act on the cells of the anterior pituitary gland [115]. GNRH is the main positive stimulator of FSH and LH release from the anterior pituitary. Likewise, GnRH-(1-5), a metabolite of GNRH, has been shown to indirectly enhance LH release via activation of kisspeptin neurons inducing GNRH expression and secretion [116, 117]. Indeed, this hormone is secreted in a pulsatile manner and the variations in its frequency and amplitude produce differential effects on the synthesis and release of gonadotropins [115, 118]. Interestingly, it has been found in animal models that FSH and LH release and expression are mainly stimulated by low and high GNRH pulse frequencies, respectively [115]. Moreover, the stimulatory effects of GNRH on gonadotropins have been also established in humans and non-human primate models [36, 119, 120]. In particular, GNRH treatment can directly produce an increase of FSH and LH secretion in vivo and in primary pituitary cell cultures from different non-human primate models [36, 120]. All these effects are mediated through the binding to the GNRH receptor (GNRHR) [115] and are exerted through the modulation of different signaling pathways including the NOS/NO/GC/cGMP pathway and extracellular Ca²⁺ mobilization, which result in the secretion of LH in baboons [36]. Additionally, other pathways such as PKC/MAPK and cAMP/PKA have been shown to be activated by GNRH in rodent models [115].

2.2.3.2 Ghrelin (GHRL)

In addition to its actions on somatotroph and lactotroph function, acylated ghrelin has been related with inhibitory actions in gonadotroph cells from rodent, ovine, and primate models, consequently modulating reproductive function [19]. Thus, ghrelin administration decreased LH pulse frequency, but not amplitude, in adult ovariectomized rhesus monkeys [121]. In the same line, ghrelin administration inhibited LH and FSH secretion in healthy men and women [19, 122, 123]. However, the effects of ghrelin on gonadotropins expression levels and the signaling pathways involved in these actions remain to be fully elucidated.

2.2.3.3 Somatostatin (SST)

Several reports have described that SST, besides its inhibitory actions on GH, TSH, PRL, and ACTH release, can also induce the inhibition of LH and FSH secretion in humans and non-human primate models [33, 124, 125]. Thus, somatostatin reduced LH, but not FSH secretion, after 4 and 24 h of incubation in primary pituitary cell cultures from baboons. Moreover, somatostatin treatment also reduced *LH*, but not *FSH* expression, after 24 h of incubation [33]. In the same line, somatostatin inhibited FSH and LH secretion in healthy humans and reduced LH response to GNRH in healthy women [124, 125]. Regarding the signaling pathways, the effects exerted by somatostatin on gonadotropins secretion/expression were mediated through AC/ PKA, MAPK, and extra-/intracellular calcium mobilization [33].

2.2.3.4 Kisspeptins (KISS1)

Kisspeptin-54 and kisspeptin-10 have been associated with an induction of FSH and LH levels in healthy humans, although these effects are less potent in comparison with the stimulatory actions of GNRH [119, 126]. Interestingly, kisspeptin-10 stimulated gonadotropins release in a different manner in women and men, showing a sexual dimorphism [127]. In line with these results, kisspeptin-10 infusion increased LH levels in rhesus monkeys [128], and also stimulated LH secretion/expression in primary pituitary cell cultures from baboons [36], being this stimulatory effect similar to that exerted by GNRH. In contrast, this peptide did not alter FSH secretion/ expression levels in the same non-human primate model [36]. The signaling pathways involved in the stimulation of LH synthesis and release by kisspeptin-10 were PLC, PKC, MAPK, intracellular calcium mobilization, mTOR, and PI3K pathways [36].

2.2.3.5 Melatonin (MT)

The information regarding the effect of MT on gonadotropins secretion/expression is not entirely consistent. Specifically, exogenous MT administration has been shown to cause different effects reducing but also increasing LH levels in humans [94, 129]. In this regard, long-term administration with MT has been shown to reduce basal blood LH levels, without altering FSH levels in humans [130]. In contrast, long-term or acute MT administration did not alter gonadotropins levels in healthy men [131, 132]. On the other hand, no changes in gonadotropins synthesis

and/or release were found in non-human primate models [92, 133]. Therefore, further studies are necessary to clarify the effect of MT on gonadotroph cells.

2.2.3.6 Gonadotropin-Inhibitory Hormone (GnIH)

GnIH is a hypothalamic peptide initially discovered in birds due to its inhibitory action on LH secretion [134]. The human GnIH peptides are called RF-related peptides (RFRPs) and have been considered as specific inhibitors of gonadotropin secretion in mammals, acting through the binding to GnIH receptor (GPR147) [135]. GnIH has been demonstrated to reduce LH secretion *in vitro* in avian, ovine, and bovine gonadotroph cells [134, 136, 137]. Moreover, a recent study has demonstrated the suppression of LH levels after exogenous GnIH administration in postmenopausal women [135]. Although further studies are required to better understand the function of GnIH in humans and the signaling pathways involved in its actions, results generated in a mouse gonadotroph cell line (LßT2) showed that the effects on gonadotropin secretion are mediated through the crosstalk inhibition of the AC/ cAMP/PKA/ERK pathway [138].

2.2.3.7 Neuropeptide-Y (NPY)

NPY is released to the hypophyseal portal circulation for transportation to the anterior pituitary gland where it modulates somatotroph cells function but also enhances LH release in response to GNRH [139]. Indeed, although NPY administration did not alter LH or FSH secretion in healthy men, the co-administration of NPY and GNRH produced a potentiation of LH and FSH release compared to GNRH alone [140]. However, in a different study, the administration of human NPY to the third ventricle of the brain in ovariectomized rhesus monkeys produced a striking reduction of LH secretion through the alteration of GNRH/LH secretory system, and these effects seemed to involve the mobilization of intracellular calcium [141]. Therefore, these contradictory results indicate the necessity of implementing further studies to undoubtedly define the role of NPY on gonadotropins release.

2.2.3.8 Other Systemic/Potential Regulators

Gonadotrophs function is also regulated by additional peripheral factors, in addition to the previously described above. Thus, short-term administration of glucocorticoids can reduce basal serum levels of LH and FSH in the follicular phase of healthy women [142], but not in healthy men [143]. On the same line, leptin has been demonstrated to directly increase FSH secretion *in vitro*, without altering LH secretion or *FSH/LH* expression, through the modulation of AC/PKA, PLC/PKC, PI3K, and extra-/intracellular calcium mobilization in two primate species [57]. Treatment with inhibins (A and B; glycoprotein hormones secreted by the granulosa and theca cells of the ovary and by the Sertoli cells of the testis [144]) can reduce FSH levels, with unclear effects on LH levels, in human fetal primary pituitary cell cultures [145]. Moreover, the reduction of inhibin B in older ovulatory women produces a monotropic FSH release [146]. In contrast, activins, glycoproteins belonging to the TGF- β superfamily, have been associated with a potent increase of FSH and LH

release in non-human primate models [147] and in human fetal primary pituitary cell cultures [145], mainly through the binding to different receptors (ActRII, ActRIIB, and ActIR/ALK4), and also producing the phosphorylation and translocation of SMAD proteins to the nucleus to finally regulate FSH/LH gene transcription [148, 149]. In addition, follistatin (FST; a monomeric polypeptide secreted from mature gonadal cells and considered a key regulator of activins [150]) has been associated with a decrease of basal and GNRH-stimulated LH and FSH levels in human fetal primary pituitary cell cultures, which might be due to the direct blockage of activin actions [145]. On the other hand, testosterone acts as a negative regulator of LH and FSH secretion in human and non-human primate models [151, 152]. Furthermore, the presence of endothelins has been detected in gonadotrophs cells, and its intravenous administration produced an increase of GNRH-stimulated LH and FSH levels in men probably mediated, at least in part, by calcium mobilization [109]. Likewise, the information about the effects of opioids at anterior pituitary level is quite contradictory; however, several reports have demonstrated a reduction on serum LH concentrations after a chronic and acute administration of opioids, and a reduction of FSH concentrations after a chronic administration, mainly acting through the μ -opioid receptor pathway [153–155]. Finally, NST has been recently related with an inhibition of basal, but not ghrelin or GNRH-stimulated, LH secretion in primary pituitary cell cultures from baboons [33]; however, this peptide did not alter LH mRNA expression levels or FSH secretion/expression levels. The signaling pathways involved in the effect of NST on LH secretion were AC/PKA, MAPK, and extra-/intracellular calcium mobilization [33].

2.2.4 Thyrotrophs

Thyrotrophs are the anterior pituitary cells responsible for producing the thyroidstimulating hormone (TSH) and their physiology is modulated by central and peripheral factors. TSH modulates the growth of the thyroid gland and the release of thyroid hormones by stimulating the thyroid follicular cells to release thyroxine (T4) and triiodothyronine (T3). The peptide hormone TSH has two different subunit, beta and alpha chains. Remarkably, TSH, LH, FSH, and the human chorionic gonadotropin (hCG) share the same alpha subunit but the beta chain is different and confers specificity and different capabilities. Indeed, FSH acts through the cAMP second messenger system and by the IP3 signaling cascade. These processes start when TSH actives TSHR (a GPCR). The final consequence of this process is the activation of the expression of a plethora of key proteins that influence many organs, promoting growth and bone maturation, increasing the basal metabolic rate and the basal cardiac output, and promoting the maturation of the central nervous system [156].

2.2.4.1 Thyrotropin-Releasing Hormone (TRH)

TRH, a tripeptide (glu-his-pro-NH2) hormone that is primarily produced in the paraventricular nucleus of the hypothalamus, has been long established and accepted

as the main regulator of TSH release. Specifically, TRH is produced by the hypophysiotropic TRHergic neurons, which integrate the information received and modulate the synthesis and release of TRH. This hormone is necessary to increase TSH release in the anterior pituitary, through the activation of TRH receptors, a family of calcium-mobilizing GPCRs that signal by elevating calcium levels and activating PKC [157]. In particular, TRH activates TRHR1 and the consequently increased IP3 levels mobilize, in turn, intracellular calcium levels, causing TSH release [157]. It seems that TRH can stimulate TSH alpha subunit expression via the PKC-MAPK pathway, while TRH-stimulated TSH beta subunit expression involves the calciumcalmodulin pathway. However, the expression of the TSH beta subunit by TRH may depends upon either the pituitary architecture, and/or paracrine factors, since it can be lost in cell culture [158].

2.2.4.2 Somatostatin (SST)

SST is one of the main inhibitors, together with DA, of TSH secretion in normal subjects [159]. Indeed, SST is synthesized in the hypothalamus and it is transported via the portal vessels of the pituitary stalk to the GH- and TSH-secreting cells. SST can inhibit TRH-induced TSH secretion in normal adult males [160]. Similarly, SST can suppress TSH pulse amplitude and frequency [161] and inhibits TSH levels in normal volunteers and in patients with primary hypothyroidism [162]. In addition, it has been recently reported that SST can directly inhibit TSH secretion in pituitary cell cultures from normal primates [33]. Consistently, other studies have shown that octreotide and lanreotide can reduce TSH secretion and normalize T3 and T4 levels in patients with pituitary TSH-secreting adenomas [163, 164]. These effects of SST inhibiting TSH release are dependent on its binding and activation of SSTR2 and SSTR5 receptors [165], which lead to the inhibition of AC and cAMP levels and the modulation of K⁺- and Ca²⁺-channels [166].

2.2.4.3 Dopamine (DA)

The inhibitory effect of DA on TSH release has been long recognized [167–169]. Indeed, DA rapidly decreases TSH subunit secretion, possibly as a result of the stimulation of DA receptors present on the surface of anterior pituitary cells [170–172]. Consistently, DA agonists, such as 1-dopa (L-3,4-dihydroxyphenylalanine) and bromocriptine, have been shown to decrease TSH secretion [173–175]. On the other hand, the administration of a therapeutic dose of metoclopramide, a DA antagonist, can enhance TSH secretion [173]. To exert these actions, DA and its agonists can act through a family of five different subtypes of DA receptors (DRD1–DRD5), which can act by classic mechanisms (AC, cAMP, etc.) or via GPCR-independent mechanisms like interactions with ion channels [176].

2.2.4.4 Other Systemic/Potential Regulators

In addition to these main TSH regulators, there are other molecules that can regulate the expression and release of TSH, including glucocorticoids, which are synthesized and released by the adrenal cortex [177]. Glucocorticoids can reduce T3 concentration in adults and preterm infants [173]. Although it is well known that

glucocorticoids activate cAMP/PKA or PKC signaling pathways and intracellular free calcium mobilization in rodents, the signaling pathways activated by glucocorticoids in humans or primates have yet to be fully elucidated [178]. Remarkably, thyroid hormones also exert a negative regulatory feedback on TSH production in that, when T3 and T4 levels are increased, they can suppress the synthesis and secretion of TSH [173]. However, the signaling pathways and mechanisms associated with the effects of thyroid hormones in humans and non-human primate pituitaries have not been fully identified yet [178]. Interestingly, a study analyzing the changes of pituitary hormones over a 24-hour period after a single subcutaneous dose of 40 micrograms/kg rhIGF1 demonstrated that IGF1 can reduce TSH levels in plasma from healthy volunteers [179]. Finally, three different studies have demonstrated that TSH can be elevated after opioids treatment [180–182]. Moreover, the use of opioids and their antagonists had profound effects in modifying the nocturnal pulses of TSH by altering the circadian rhythm of this hormone [178].

2.2.5 Corticotrophs

Corticotroph cells are mostly located in the anteromedial areas of the pituitary gland and constitute about 15–20% of all the anterior pituitary cells. These cells produce mainly ACTH, which is derived from the cleavage of the precursor hormone proopiomelanocortin (POMC) by the action of different prohormone convertase enzymes [183]. Corticotrophs are characteristically identified by their basophil staining, by their PAS-positivity due to the high glycoprotein content of the N-terminal glycopeptide of POMC, as well as by their ACTH immunopositivity. The main biological function of ACTH is to regulate the plasmatic levels of cortisol and androgens, by modulating their expression and secretion in adrenal cortex cells [55]. ACTH works through GPCRs located at the extracellular membranes on cells of the *zona fasciculata* and *zona reticularis* of the adrenal cortex. The activation of ACTH receptors leads to the stimulation of AC and, thus, the increase in cAMP production. The secretion of ACTH is influenced by diverse stimuli, as described below [183].

2.2.5.1 Corticotropin-Releasing Hormone (CRH)

CRH is a neuropeptide hormone that consists of 41 amino acids and it is secreted from the paraventricular nucleus of the hypothalamus [184]. It regulates neuroendocrine, sympathetic, and behavioral functions in response to stress. In particular, at the anterior pituitary level, CRH activates the synthesis and release of ACTH from corticotrophs, which, in turn, stimulates the secretion of glucocorticoid hormones (mainly cortisol in humans) from the adrenal cortex. In this way, CRH affects the response to stress, addiction, and depression, among others [185]. CRH acts via 2 distinct GPCRs, namely, CRHR1 and CRHR2. CRHR1 expression is prevalent in brain areas responsible for sensory and motor control, such as the cortical mantle, olfactory bulb, hippocampus, amygdala, hypothalamic nuclei, and cerebellum [186]. CRHR2 is predominant in subcortical regions, including the lateral septum, *stria terminalis*, or the ventromedial hypothalamic nucleus [185, 186]. In the anterior pituitary, CRHR1 mediates the release of ACTH in response to CRH through the recruitment of several intracellular effectors such as cAMP and protein kinases [184, 185].

2.2.5.2 Ghrelin (GHRL)

Different studies in humans and non-human primate models have demonstrated that ghrelin regulates ACTH release by anterior pituitary cells [187–189]. Indeed, ghrelin plays an important role in the activation of central pathways mediating stress-induced food reward behavior [190] and is involved in the response to acute stressors [191, 192] by the stimulation of hormones involved in the stress response, including ACTH, AVP, PRL, and cortisol [193]. The stimulation of ACTH release by ghrelin is exerted predominantly at the hypothalamic level through AVP stimulation and indirect activation of CRH neurons [194]. However, despite abundant GHSR1a expression in the anterior pituitary, the direct effects of ghrelin on pituitary ACTH synthesis and release are minor and not consistent across species, which may be explained by differences in the molecular regulation of downstream pathways [13, 187]. In addition, the stimulatory effect of GH secretagogues on corticotroph function is sensitive to glucocorticoid feedback, as the ACTH response to hexarelin, a synthetic GH secretagogue that activates GHSR1a, is suppressed by pre-treatment with dexamethasone [13].

2.2.5.3 Pituitary Adenylate Cyclase-Activating Polypeptide (PACAP)

PACAP has been shown to be involved in restraint stress-induced corticosterone release and concomitant expression of the genes involved in hypothalamic–pituitary–adrenal (HPA) axis activation. Indeed, it has been described that intravenous PACAP administration can regulate ACTH secretion in different mammalian species including humans [195]. Consistently, PACAP has been shown to be able to induce ACTH release from the corticotroph cell line AtT-20, which is suppressed by SST administration [196]. PACAP exerts its biological actions by coupling to different GPCRs classified into three groups based on their differential affinity for PACAP or vasoactive intestinal peptide (VIP). Thus, PACAP type 1 receptors (PAC1R) are more specific for PACAP while vasoactive intestinal polypeptide receptor 1 (VPAC1) and VPAC2 receptors present similar affinity for either PACAP isoforms or VIP [197]; however, the receptor subtypes mediating these effects on ACTH secretion have not been identified hitherto.

2.2.5.4 Vasopressin (AVP)

Vasopressin, also named as antidiuretic hormone (ADH), AVP or argipressin, is a hormone synthesized as a peptide prohormone in neurons of the hypothalamus and stored and released by the posterior pituitary gland. AVP is the major physiological regulator of renal water excretion and blood volume and is normally released in response to an increase in blood osmolality or decrease in blood volume [198, 199]. In addition, during acute stress, AVP expression and secretion rapidly increases similar to CRH and ACTH, and AVP contributes to the full ACTH response [200]. Indeed, AVP administration increases ACTH levels in healthy humans, wherein

AVP seems to enhance CRH-stimulated ACTH release [30]. In fact, it was reported that AVP from pituitary portal circulation is more important in altering ACTH levels than AVP derived from peripheral circulation. These effects seem to be mainly mediated by the V1b GPCRs, which are prominently expressed in corticotroph cells, and can modulate ACTH release [198].

2.2.5.5 Somatostatin (SST)

The role of SST in the regulation of ACTH secretion from pituitary corticotroph cells is still to be clearly clarified. Initial studies suggested that SST did not affect basal or CRH [201] or ghrelin [202] stimulated ACTH or cortisol levels in humans. However, more recent studies suggest that SST can regulate ACTH secretion, which is dependent on cortisol levels and cell milieu [203]. Moreover, it has been reported that SST knockout mice had elevated POMC expression and corticosterone levels suggesting that endogenous SST can inhibit pituitary-adrenal axis [187]. In line with this, inconsistent results have also been reported *in vitro*. Specifically, although SST did not affect basal or CRH-stimulated ACTH secretion in normal rat pituitary cells [204, 205], it inhibited CRH- and AVP-induced ACTH secretion in cultured pituitary cells derived from adrenalectomized rats and in serum-starved cultures [203]. In addition, it has been also reported that SST can inhibit ACTH release in primary pituitary cell cultures from mice [187] and primates [33, 65, 187]. These putative effects might involve the differential participation of SSTR2 and SSTR5 receptors [206–208].

2.2.5.6 Cortistatin (CST)

A limited number of studies indicate that CST may exert an important role in controlling ACTH levels. Indeed, these studies suggest that CST can directly modulate the function of different pituitary cell types, including corticotrophs, and these actions in humans and animal models are dose- and cell type-dependent and receptor-specific [165]. Moreover, it has been reported that endogenous CST may serve to suppress pituitary ACTH output in a gender-dependent fashion in that CST-KO female, but not male, mice had elevated circulating ACTH levels [28]. These effects may be, at least in part, direct, because the same study demonstrated that CST treatment inhibited ACTH release in primary pituitary cell cultures of normal mice and baboons [28].

2.2.5.7 Other Systemic/Potential Regulators

Besides the factors described above, there are other factors able to modulate ACTH secretion. As an example, the inhibition of ACTH release in response to opioids treatment has been reported in several studies [209–211], and this effect seems to be mediated through kappa-opioid receptors [212]. In the case of adipokines, it has been reported that while adiponectin decreases ACTH release, leptin and resistin increase ACTH release in primary cell cultures from non-human primates by modulating common (AC/PKA) and specific signaling pathways (PLC/PKC, Ca²⁺, mTOR) [57]. Moreover, obestatin has been reported to stimulate ACTH secretion in mice *in vitro* (primary cell cultures) and *in vivo* as well as in baboon primary

pituitary cell cultures [65]. Finally, as expected, glucocorticoids negatively feedback to suppress their own axis, therefore suppressing ACTH secretion *in vivo* and *in vitro* [213–215]. Thus, the actions of all of these factors, among others, should be considered when analyzing the regulation of ACTH secretion.

2.3 Posterior Pituitary

The posterior pituitary or *neurohypophysis* is a particular region of the pituitary gland with a different embryonic origin [216, 217]. The posterior pituitary is directly connected to the hypothalamus via a nerve tract (hypothalamo-hypophyseal nerve tract). Posterior pituitary can be structured in three parts: pars nervosa, infundibular stalk, and median eminence [217]. It is composed by neuronal projections (axons) of magnocellular neurosecretory cells (MNCs), which are responsible for OT and AVP secretion into the general circulation [216]. These hormones are transported in association with neurophysins proteins along the axons of these neurons to the nerve terminals within the posterior pituitary. Particularly, OT and AVP are synthesized by different populations of MNCs in the paraventricular and supraoptic nuclei of the hypothalamus [218], displaying important central and peripheral functions. OT is a nine amino acids peptide that binds different GPCRs (OT-Rs) distributed throughout the brain, although the molecular mechanism underlying their activation remains unknown [219]. The classical physiological roles of OT are the regulation of lactation, parturition, and reproductive behavior, being essential to stimulate uterine contraction and milk secretion [216]. As such, stimulation of vagal sensory afferents in the nipple by the act of suckling triggers reflex synchronized firing of oxytocin magnocellular neurons in the neurohypophysis, and corresponding pulsatile OT release [216]. In many mammals, there is also an increase in OT secretion and in uterine responsiveness to OT during parturition [220, 221]. OT also plays a key role within the brain to control behavior in social cognition and in fear conditioning, which is important in social anxiety as well as in other disorders with impaired social functioning [222]. AVP [223] is a nonapeptide that binds to at least three different GPCR subtypes (V1a, V1b, and V2). The most important action of AVP is its antidiuretic action on the collecting ducts of the kidney [199]. In fact, AVP maintains body fluid balance by keeping plasma osmolarity and controlling liquid kidney excretion and thirst [199]. AVP is secreted in response to increased osmolality, changes in blood volume or pressure, nausea, hypoglycemia, and nicotine [199, 224]. Indeed, the main stimulus for AVP release is dehydration, resulting in an increase in plasma osmolarity, but other stimuli also influence AVP secretion, mainly sodium, the reduction in circulating blood, stress situations [199, 225], or different hormones. In general terms, increased firing frequency of oxytocin and vasopressinergic neurons opens voltage-gated Ca2+ channels in their nerve terminals. This, in turn, leads to transient Ca2+ influx, fusion of the neurosecretory granules with the nerve terminal membrane, and release of the hormone and their neurophysins proteins into the systemic circulation in equimolar quantities.

Therefore, these two posterior pituitary hormones (OT and AVP) comprise a highly related and integrated system, but they exert clearly distinct actions and their synthesis and release is differentially regulated by central and peripheral stimuli. Some of these stimuli have been described above; however, the secretion of posterior pituitary hormones is influenced by additional factors, as described below.

2.3.1 Opioids

Endogenous opioid peptides (or endomorphins) are synthesized by different precursors of the neurohypophysis (mainly POMC, proenkephalin A and prodynorphin) and they act directly on specific receptors well distributed centrally and peripherally [226]. The effect of the opioid system and its receptors in posterior pituitary hormones is strongly complex and some controversial results have been described. It is well described that OT secretion is inhibited centrally by mu- and kappa-opioid agonists or directly at the posterior pituitary via kappa-opioid receptors [42, 226, 227]. However, kappa-opioid receptors also induce a suppression of AVP, increasing diuresis status, with the consequence of high sodium levels and an increase on OT and AVP [228]. Therefore, the exact role of the endogenous opioid system on OT release has to be further explored.

2.3.2 Glucocorticoids

In general, glucocorticoid administration is able to elevate plasma AVP concentration and improve the impaired water diuresis in adrenal insufficiency. However, the specific effect of cortisol on AVP release is still controversial in that it depends on the species studied. For example, there is no clear effect of cortisol in horse AVP release, but in contrast, in other species such as sheep or importantly in human, evidence obtained indicates that, under certain circumstances, cortisol could inhibit the release of AVP [229–231].

2.3.3 Angiotensin II (AII)

AII is a peptide hormone produced from angiotensin I through the removal of two C-terminal residues. AII has been shown to act as an endocrine, autocrine/paracrine, and intracrine hormone [232]. Indeed, immunoreactive AII has been localized in magnocellular cells of the supraoptic and paraventricular nuclei, which mostly contain AVP [233]. AII has been shown to increase aldosterone secretion, act on venous and arterial smooth muscle to cause vasoconstriction, and increase AVP production and secretion in different species [232, 234, 235].

2.3.4 Norepinephrine (NE)

NE, also known as noradrenaline (NA) or noradrenalin, is a catecholamine hormone that functions as a neurotransmitter in the sympathetic nervous system [236]. The infusion of norepinephrine increases both arterial blood pressure and left atrial pressure. These changes are capable of eliciting a reflex inhibition of AVP release, which can reduce plasma AVP. However, the inhibitory effects of the sino-aortic and cardiac reflexes on AVP release seem to be offset by the direct stimulatory effect of circulating NE [236].

2.3.5 Other Systemic/Potential Regulators

There are other intermediates that could be implicated in the secretion of posterior pituitary hormones, although their direct actions have to be still fully elucidated. The vasoactive intestinal polypeptide 1 or VIP1 seems to have an effect in neurosecretory nerve endings increasing OT and AVP plasma levels after an intracarotid infusion in cats [237]. Galanin, which is mainly produced in the small intestine and is susceptible to changes in fluid homeostasis, has been suggested as a putative modulator of neurohypophysial function and AVP secretion [238]. The relaxin hormone has been also related with an inhibition of OT and AVP release into plasma in rats, suggesting a possible role of this hormone in the neurohypophysis [239]. Exposure of neurohypophysis to interleukin-1 beta has been also associated with an increase of AVP release [240].

2.4 Pars Intermedia

The *pars intermedia* is a thin band of cells between the *pars distalis* and the neurohypophysis. The main functional elements of the *pars intermedia* are the melanotroph cells. Like the corticotrophs in the *pars distalis*, melanotrophs produce POMC; however, as a result of differential post-translational processing, the main end-products derived from POMC in these cells are α -MSH, β -MSH, and γ -MSH [241]. These peptides are associated with many physiological functions through the binding and activation of four of the five subtypes of melanocortin receptor (MC1R, MC3R, MC4R, and MC5R, excluding the ACTH-specific receptor MC2R), which are widely distributed throughout the body [242]. MC1R is present in melanocytes, keratinocytes, leukocytes, and adipocytes; MC3R is present in the CNS, kidney, testis, ovary, skeletal muscle, placenta, and mammary gland; MC4R is present in the CNS; and MC5R is present in exocrine glands, muscle, and CNS [242, 243]. The most representative physiological function of MSH peptides is the stimulation of melanocytes in the skin to promote the synthesis of melanin via MC1R activation. Moreover, other important functions associated with these elements include the modulation of energy homeostasis and natriuresis via MC3R, the regulation of synthesis and secretion of exocrine gland products via MC5R, the modulation of aldosterone release from the adrenal cortex, and the control of sodium metabolism and blood pressure [243, 244]. Most recently, α -MSH has been shown to exhibit an anorexigenic character and the capacity to regulate endothelial cell migration [245, 246]. All these receptors are GPCRs and exert their functions through AC and PKA activation [242]. Therefore, the secretion of MSH is influenced by diverse stimuli, as described below.

2.4.1 DA, NPY, and γ-Aminobutyric Acid

The control of the secretion of these peptides appears to be under tonic hypothalamic inhibition. In fact, and in line with this, melanotrophs can spontaneously secrete α -MSH when the connection between the hypothalamus and the *pars intermedia* is disrupted [247]. The specific molecular control of MSH release is well described in *Xenopus laevis*. Xenopus melanotroph cells are contacted by synaptic structures that secrete three types of neurotransmitters: DA, NPY, and γ -aminobutyric acid (GABA). All three transmitters inhibit the secretion of α -MSH but DA and NPY also inhibit POMC biosynthesis [248]. This inhibition is mediated by D₂-like, the GABA_B, and the NPYY1 receptor located in melanotrophs.

2.4.2 Other Systemic/Potential Regulators

Other classic neurotransmitters can induce MSH release *in vitro* including NE, serotonin (5-HT), CRH, TRH, brain-derived neurotrophic factor (BDNF), urocortin, mesotocin, and vasotocin [249]. In addition, alternative regulatory mechanisms have been described. It is known that factors produced by melanotrophs, such as acetylcholine (Ach) and BDNF, act in an autocrine way to stimulate MSH release [250]. Moreover, it has been described that the activation of calcium-sensing receptors [15] present in melanotrophs can also induce the production of MSH [251].

2.5 Conclusion

The pituitary gland is able to control a multiplicity of biological processes, including growth, reproduction, whole-body metabolism, and stress by the secretion of a variety of hormones, including GH, PRL, LH, FSH, TSH, ACTH, MSH, OT, and AVP. The synthesis and/or production of these hormones at the pituitary level is tightly controlled by a plethora of central and peripheral factors that finely interact to modulate the physiologic production of these hormones (Fig. 2.1). It is now known that each pituitary cell type exhibits a particular pattern of receptors for different signals and that the appropriate sensing, transmission, and integration of all these inhibitory and stimulating signals is crucial to adapt the circulating levels of these hormones to the physiological necessities of the organisms.

Funding Sources This work was funded by the Junta de Andalucía (BIO-0139, P20_00442), MINECO/MECD (FPU16/05059, FPU16/06190, FPU17/00263, PID2019-105564RB-100), and CIBERobn. CIBER is an initiative of Instituto de Salud Carlos III, Ministerio de Sanidad, Servicios Sociales e Igualdad, Spain.

Conflict of Interest The authors declare that they have no conflict of interest.

References

- 1. Vazquez-Borrego MC, et al. Multiple signaling pathways convey central and peripheral signals to regulate pituitary function: lessons from human and non-human primate models. Mol Cell Endocrinol. 2017;463:4.
- 2. Dores RM. The evolution of the pituitary \Rightarrow , in Reference Module in Biomedical Sciences. 2017.
- 3. Baylis PH. Posterior pituitary function in health and disease. Clin Endocrinol Metab. 1983;12(3):747–70.
- 4. Le Roith D, et al. The somatomedin hypothesis: 2001. Endocr Rev. 2001;22(1):53-74.
- Vijayakumar A, et al. Biological effects of growth hormone on carbohydrate and lipid metabolism. Growth Hormon IGF Res. 2010;20(1):1–7.
- Goldenberg N, Barkan A. Factors regulating growth hormone secretion in humans. Endocrinol Metab Clin N Am. 2007;36(1):37–55.
- 7. Gahete MD, et al. Understanding the multifactorial control of growth hormone release by somatotropes: lessons from comparative endocrinology. Ann N Y Acad Sci. 2009;1163:137–53.
- Grossman A, Savage MO, Besser GM. Growth hormone releasing hormone. Clin Endocrinol Metab. 1986;15(3):607–27.
- 9. Aguiar-Oliveira MH, et al. Hypothalamic abnormalities: growth failure due to defects of the GHRH receptor. Growth Hormon IGF Res. 2018;38:14–8.
- Corazzini V, Salvatori R. Molecular and clinical aspects of GHRH receptor mutations. Endocr Dev. 2013;24:106–17.
- Alatzoglou KS, Dattani MT. Genetic causes and treatment of isolated growth hormone deficiency-an update. Nat Rev Endocrinol. 2010;6(10):562–76.
- 12. Vitali E, et al. Cyclic adenosine 3'-5'-monophosphate (cAMP) exerts proliferative and antiproliferative effects in pituitary cells of different types by activating both cAMP-dependent protein kinase A (PKA) and exchange proteins directly activated by cAMP (Epac). Mol Cell Endocrinol. 2014;383(1–2):193–202.
- Kojima M, et al. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. Nature. 1999;402(6762):656–60.
- 14. Ueberberg B, et al. Expression of ghrelin and its receptor in human tissues. Horm Metab Res. 2009;41(11):814–21.
- Ghigo E, et al. Ghrelin: more than a natural GH secretagogue and/or an orexigenic factor. Clin Endocrinol. 2005;62(1):1–17.
- Hataya Y, et al. A low dose of ghrelin stimulates growth hormone (GH) release synergistically with GH-releasing hormone in humans. J Clin Endocrinol Metab. 2001;86(9):4552.
- 17. Kineman RD, Luque RM. Evidence that ghrelin is as potent as growth hormone (GH)releasing hormone (GHRH) in releasing GH from primary pituitary cell cultures of a

nonhuman primate (Papio anubis), acting through intracellular signaling pathways distinct from GHRH. Endocrinology. 2007;148(9):4440–9.

- Gahete MD, et al. A novel human ghrelin variant (In1-ghrelin) and ghrelin-O-acyltransferase are overexpressed in breast cancer: potential pathophysiological relevance. PLoS One. 2011;6(8):e23302.
- Motta G, et al. Ghrelin actions on somatotropic and gonadotropic function in humans. Prog Mol Biol Transl Sci. 2016;138:3–25.
- 20. O'Toole TJ and Sharma S. Physiology, somatostatin, in StatPearls. 2019: Treasure Island (FL).
- 21. Reisine T, Bell GI. Molecular biology of somatostatin receptors. Endocr Rev. 1995;16(4):427–42.
- 22. Lamberts SW. The role of somatostatin in the regulation of anterior pituitary hormone secretion and the use of its analogs in the treatment of human pituitary tumors. Endocr Rev. 1988;9(4):417–36.
- Dalm VA, et al. Distribution pattern of somatostatin and cortistatin mRNA in human central and peripheral tissues. Clin Endocrinol. 2004;60(5):625–9.
- Cordoba-Chacon J, et al. Cortistatin is a key factor regulating the sex-dependent response of the GH and stress axes to fasting in mice. Endocrinology. 2016;157(7):2810–23.
- de Lecea L, et al. A cortical neuropeptide with neuronal depressant and sleep-modulating properties. Nature. 1996;381(6579):242–5.
- Gahete MD, et al. Are somatostatin and cortistatin two siblings in regulating endocrine secretions? In vitro work ahead. Mol Cell Endocrinol. 2008;286(1–2):128–34.
- Deghenghi R, et al. Cortistatin, but not somatostatin, binds to growth hormone secretagogue (GHS) receptors of human pituitary gland. J Endocrinol Investig. 2001;24(1):RC1-3.
- Cordoba-Chacon J, et al. Cortistatin is not a somatostatin analogue but stimulates prolactin release and inhibits GH and ACTH in a gender-dependent fashion: potential role of ghrelin. Endocrinology. 2011;152(12):4800–12.
- Ibanez-Costa A, Luque RM, Castano JP. Cortistatin: a new link between the growth hormone/ prolactin axis, stress, and metabolism. Growth Hormon IGF Res. 2017;33:23–7.
- Broglio F, et al. Endocrine activities of cortistatin-14 and its interaction with GHRH and ghrelin in humans. J Clin Endocrinol Metab. 2002;87(8):3783–90.
- Prodam F, et al. Cortistatin-8, a synthetic cortistatin-derived ghrelin receptor ligand, does not modify the endocrine responses to acylated ghrelin or hexarelin in humans. Neuropeptides. 2008;42(1):89–93.
- Samson WK, et al. Neuronostatin encoded by the somatostatin gene regulates neuronal, cardiovascular, and metabolic functions. J Biol Chem. 2008;283(46):31949–59.
- Luque RM, Kineman RD. Neuronostatin exerts actions on pituitary that are unique from its sibling peptide somatostatin. J Endocrinol. 2018;237(3):217–27.
- Elrick MM, et al. Neuronostatin acts via GPR107 to increase cAMP-independent PKA phosphorylation and proglucagon mRNA accumulation in pancreatic alpha-cells. Am J Physiol Regul Integr Comp Physiol. 2016;310(2):R143–55.
- Ohtaki T, et al. Metastasis suppressor gene KiSS-1 encodes peptide ligand of a G-proteincoupled receptor. Nature. 2001;411(6837):613–7.
- 36. Luque RM, et al. Kisspeptin regulates gonadotroph and somatotroph function in nonhuman primate pituitary via common and distinct signaling mechanisms. Endocrinology. 2011;152(3):957–66.
- Gahete MD, et al. Role of the Kiss1/Kiss1r system in the regulation of pituitary cell function. Mol Cell Endocrinol. 2016;438:100–6.
- 38. Murakami Y, et al. Roles and mechanisms of action of pituitary adenylate cyclaseactivating polypeptide (PACAP) in growth hormone and prolactin secretion. Endocr J. 2001;48(2):123–32.
- 39. Peeters K, et al. Effects of pituitary adenylate cyclase-activating polypeptide (PACAP) on cAMP formation and growth hormone release from chicken anterior pituitary cells. Ann N Y Acad Sci. 1998;865:471–4.

- 40. Moody TW, et al. VIP and PACAP: recent insights into their functions/roles in physiology and disease from molecular and genetic studies. Curr Opin Endocrinol Diabetes Obes. 2011;18(1):61–7.
- 41. Lee LT, et al. Discovery of growth hormone-releasing hormones and receptors in nonmammalian vertebrates. Proc Natl Acad Sci U S A. 2007;104(7):2133–8.
- Zanos P, et al. Oxytocin and opioid addiction revisited: old drug, new applications. Br J Pharmacol. 2018;175(14):2809–24.
- Forsling ML, Wheeler MJ, Williams AJ. The effect of melatonin administration on pituitary hormone secretion in man. Clin Endocrinol. 1999;51(5):637–42.
- Dubocovich ML, Markowska M. Functional MT1 and MT2 melatonin receptors in mammals. Endocrine. 2005;27(2):101–10.
- 45. Ibáñez-Costa A, et al. Melatonin regulates somatotrope and lactotrope function through common and distinct signaling pathways in cultured primary pituitary cells from female primates. Endocrinology. 2015;156(3):1100–10.
- 46. Brandon DH, Holditch-Davis D, Belyea M. Preterm infants born at less than 31 weeks' gestation have improved growth in cycled light compared with continuous near darkness. J Pediatr. 2002;140(2):192–9.
- 47. Lisoni P, et al. Effect of an acute injection of melatonin on the basal secretion of hypophyseal hormones in prepubertal and pubertal healthy subjects. Acta Endocrinol. 1986;111(3):305–11.
- 48. Wright J, et al. The effects of exogenous melatonin on endocrine function in man. Clin Endocrinol. 1986;24(4):375–82.
- Kostoglou-Athanassiou I, et al. Neurohypophysial hormone and melatonin secretion over the natural and suppressed menstrual cycle in premenopausal women. Clin Endocrinol. 1998;49(2):209–16.
- 50. Smythe GA, Lazarus L. Growth hormone responses to melatonin in man. Science. 1974;184(4144):1373–4.
- Valcavi R, et al. Effect of oral administration of melatonin on GH responses to GRF 1–44 in normal subjects. Clin Endocrinol. 1987;26(4):453–8.
- Okinaga H, et al. Mechanisms of TRH-induced GH release (paradoxical response) in human somatotroph adenoma cells. Endocr J. 2005;52(6):763–7.
- Adams EF, et al. Neuropeptide Y directly inhibits growth hormone secretion by human pituitary somatotropic tumours. Acta Endocrinol. 1987;115(1):149–54.
- Pedrazzini T, Pralong F, Grouzmann E. Neuropeptide Y: the universal soldier. Cell Mol Life Sci. 2003;60(2):350–77.
- 55. Lim CT, Grossman A, Khoo B. Normal physiology of ACTH and GH release in the hypothalamus and anterior pituitary in man. South Dartmouth, MA: MDText.com, Inc.; 2000.
- Wilson ME, et al. Leptin administration increases nocturnal concentrations of luteinizing hormone and growth hormone in juvenile female rhesus monkeys. J Clin Endocrinol Metab. 2003;88(10):4874–83.
- 57. Sarmento-Cabral A, et al. Adipokines (leptin, adiponectin, resistin) differentially regulate all hormonal cell types in primary anterior pituitary cell cultures from two primate species. Sci Rep. 2017;7:43537.
- Copeland KC, et al. Estrogen stimulates growth hormone and somatomedin-C in castrate and intact female baboons. J Clin Endocrinol Metab. 1984;58(4):698–703.
- Burguera B, et al. Dual and selective actions of glucocorticoids upon basal and stimulated growth hormone release in man. Neuroendocrinology. 1990;51(1):51–8.
- Luque RM, et al. Cortistatin mimics somatostatin by inducing a dual, dose-dependent stimulatory and inhibitory effect on growth hormone secretion in somatotropes. J Mol Endocrinol. 2006;36(3):547–56.
- 61. Tomasi PA, et al. Opioid-receptor blockade blunts growth hormone (GH) secretion induced by GH-releasing hormone in the human male. Horm Metab Res. 1998;30(1):34–6.
- 62. Hartman ML, et al. A low dose euglycemic infusion of recombinant human insulin-like growth factor I rapidly suppresses fasting-enhanced pulsatile growth hormone secretion in humans. J Clin Invest. 1993;91(6):2453–62.

- 63. Gahete MD, et al. Elevated GH/IGF-I, due to somatotrope-specific loss of both IGF-I and insulin receptors, alters glucose homeostasis and insulin sensitivity in a diet-dependent manner. Endocrinology. 2011;152(12):4825–37.
- 64. Giustina A, Veldhuis JD. Pathophysiology of the neuroregulation of growth hormone secretion in experimental animals and the human. Endocr Rev. 1998;19(6):717–97.
- 65. Luque RM, et al. Obestatin plays an opposite role in the regulation of pituitary somatotrope and corticotrope function in female primates and male/female mice. Endocrinology. 2014;155(4):1407–17.
- Pombo M, et al. Regulation of growth hormone secretion by signals produced by the adipose tissue. J Endocrinol Investig. 1999;22(5 Suppl):22–6.
- Luque RM, et al. Examination of the direct effects of metabolic factors on somatotrope function in a non-human primate model, *Papio anubis*. J Mol Endocrinol. 2006;37(1):25–38.
- Freeman ME, et al. Prolactin: structure, function, and regulation of secretion. Physiol Rev. 2000;80(4):1523–631.
- 69. Gadelha MR, et al. Genetics of pituitary adenomas. Front Horm Res. 2013;41:111-40.
- Lamberts SW, Macleod RM. Regulation of prolactin secretion at the level of the lactotroph. Physiol Rev. 1990;70(2):279–318.
- Liu C, Kaeser PS. Mechanisms and regulation of dopamine release. Curr Opin Neurobiol. 2019;57:46–53.
- Mansour A, et al. Localization of dopamine D2 receptor mRNA and D1 and D2 receptor binding in the rat brain and pituitary: an in situ hybridization-receptor autoradiographic analysis. J Neurosci. 1990;10(8):2587–600.
- Ishibashi M, Yamaji T. Mechanism of the inhibitory action of dopamine and somatostatin on prolactin secretion from human lactotrophs in culture. J Clin Endocrinol Metab. 1985;60(3):599–606.
- Denef C, Manet D, Dewals R. Dopaminergic stimulation of prolactin release. Nature. 1980;285(5762):243–6.
- Chang A, Shin SH. Dopamine agonists both stimulate and inhibit prolactin release in GH4ZR7 cells. Eur J Endocrinol. 1999;141(4):387–95.
- Kineman RD, Gettys TW, Frawley LS. Paradoxical effects of dopamine (DA): Gi alpha 3 mediates DA inhibition of PRL release while masking its PRL-releasing activity. Endocrinology. 1994;135(2):790–3.
- Kanasaki H, et al. Role of thyrotropin-releasing hormone in prolactin-producing cell models. Neuropeptides. 2015;54:73–7.
- Ansari MS, Almalki MH. Primary hypothyroidism with markedly high prolactin. Front Endocrinol (Lausanne). 2016;7:35.
- Bahar A, et al. Hyperprolactinemia in association with subclinical hypothyroidism. Caspian J Intern Med. 2011;2(2):229–33.
- Vazquez-Borrego MC, et al. Multiple signaling pathways convey central and peripheral signals to regulate pituitary function: lessons from human and non-human primate models. Mol Cell Endocrinol. 2018;463:4–22.
- Ray KP, Wallis M. Studies of TRH-induced prolactin secretion and its inhibition by dopamine, using ovine pituitary cells. Mol Cell Endocrinol. 1984;36(1–2):131–9.
- 82. Muller TD, et al. Ghrelin. Mol Metab. 2015;4(6):437-60.
- Messini CI, et al. Effect of ghrelin and metoclopramide on prolactin secretion in normal women. J Endocrinol Investig. 2011;34(4):276–9.
- Messini CI, et al. Growth hormone and prolactin response to ghrelin during the normal menstrual cycle. Clin Endocrinol. 2009;71(3):383–7.
- 85. Messini CI, et al. Effect of ghrelin and thyrotropin-releasing hormone on prolactin secretion in normal women. Horm Metab Res. 2010;42(3):204–8.
- 86. Mijiddorj T, et al. Stimulatory effect of pituitary adenylate-cyclase activating polypeptide (PACAP) and its PACAP type I receptor (PAC1R) on prolactin synthesis in rat pituitary somatolactotroph GH3 cells. Mol Cell Endocrinol. 2011;339(1–2):172–9.

- 87. Isaac ER, Sherwood NM. Pituitary adenylate cyclase-activating polypeptide (PACAP) is important for embryo implantation in mice. Mol Cell Endocrinol. 2008;280(1–2):13–9.
- Oride A, Kanasaki H, Kyo S. Role of pituitary adenylate cyclase-activating polypeptide in modulating hypothalamic-pituitary system. Reprod Med Biol. 2018;17(3):234–41.
- Lincoln GA, Andersson H, Hazlerigg D. Clock genes and the long-term regulation of prolactin secretion: evidence for a photoperiod/circannual timer in the pars tuberalis. J Neuroendocrinol. 2003;15(4):390–7.
- Johnston JD, Skene DJ. 60 Years of neuroendocrinology: regulation of mammalian neuroendocrine physiology and rhythms by melatonin. J Endocrinol. 2015;226(2):T187–98.
- Bispink G, et al. Influence of melatonin on the sleep-independent component of prolactin secretion. J Pineal Res. 1990;8(2):97–106.
- Ibanez-Costa A, et al. Melatonin regulates somatotrope and lactotrope function through common and distinct signaling pathways in cultured primary pituitary cells from female primates. Endocrinology. 2015;156(3):1100–10.
- Forsling ML, Wheeler MJ, Williams AJ. The effect of melatonin administration on pituitary hormone secretion in man. Clin Endocrinol. 1999;51(5):637–42.
- Ninomiya T, et al. Effects of exogenous melatonin on pituitary hormones in humans. Clin Physiol. 2001;21(3):292–9.
- Vale W, et al. Effects of somatostatin on the secretion of thyrotropin and prolactin. Endocrinology. 1974;95(4):968–77.
- Drouin J, et al. Characteristics of the interaction between thyrotropin-releasing hormone and somatostatin for thyrotropin and prolactin release. Endocrinology. 1976;98(2):514–21.
- Gooren LJ, Harmsen-Louman W, van Kessel H. Somatostatin inhibits prolactin release from the lactotroph primed with oestrogen and cyproterone acetate in man. J Endocrinol. 1984;103(3):333–5.
- Borski RJ, Hyde GN, Fruchtman S. Signal transduction mechanisms mediating rapid, nongenomic effects of cortisol on prolactin release. Steroids. 2002;67(6):539–48.
- 99. Jaquet P, et al. Quantitative and functional expression of somatostatin receptor subtypes in human prolactinomas. J Clin Endocrinol Metab. 1999;84(9):3268–76.
- Fusco A, et al. Somatostatinergic ligands in dopamine-sensitive and -resistant prolactinomas. Eur J Endocrinol. 2008;158(5):595–603.
- 101. Gruszka A, Culler MD, Melmed S. Somatostatin analogs and chimeric somatostatin-dopamine molecules differentially regulate human growth hormone and prolactin gene expression and secretion in vitro. Mol Cell Endocrinol. 2012;362(1–2):104–9.
- 102. Hofland LJ, et al. The novel somatostatin analog SOM230 is a potent inhibitor of hormone release by growth hormone- and prolactin-secreting pituitary adenomas in vitro. J Clin Endocrinol Metab. 2004;89(4):1577–85.
- Rivas RJ, Nishioka RS, Bern HA. In vitro effects of somatostatin and urotensin II on prolactin and growth hormone secretion in tilapia, *Oreochromis mossambicus*. Gen Comp Endocrinol. 1986;63(2):245–51.
- 104. Rubinfeld H, et al. Cortistatin inhibits growth hormone release from human fetal and adenoma pituitary cells and prolactin secretion from cultured prolactinomas. J Clin Endocrinol Metab. 2006;91(6):2257–63.
- 105. Grottoli S, et al. Cortistatin-17 and somatostatin-14 display the same effects on growth hormone, prolactin, and insulin secretion in patients with acromegaly or prolactinoma. J Clin Endocrinol Metab. 2006;91(4):1595–9.
- Baranowska B, et al. Cortistatin and pituitary hormone secretion in rat. J Physiol Pharmacol. 2009;60(1):151–6.
- 107. Bethea CL, et al. The effect of simultaneous versus sequential estradiol and progesterone treatments on prolactin production in monkey pituitary cell cultures. Endocrinology. 1988;122(5):1786–800.
- 108. Kiefer F, et al. Comparison of the effects of endothelin-1 and -3 on secretion of pituitary hormones in healthy male volunteers. Exp Clin Endocrinol Diabetes. 2000;108(5):378–81.

- 109. Vierhapper H, et al. Effect of endothelin-1 in man—impact on basal and stimulated concentrations of luteinizing hormone, follicle-stimulating hormone, thyrotropin, growth hormone, corticotropin, and prolactin. Metabolism. 1993;42(7):902–6.
- 110. Goodyer CG, et al. Effect of insulin-like growth factors on human foetal, adult normal and tumour pituitary function in tissue culture. Acta Endocrinol. 1986;112(1):49–57.
- 111. Nikolics K, et al. A prolactin-inhibiting factor within the precursor for human gonadotropinreleasing hormone. Nature. 1985;316(6028):511–7.
- 112. Musumeci G, et al. A journey through the pituitary gland: development, structure and function, with emphasis on embryo-foetal and later development. Acta Histochem. 2015;117(4–5):355–66.
- 113. Asa SL, Ezzat S. The pathogenesis of pituitary tumors. Annu Rev Pathol. 2009;4:97-126.
- Ulloa-Aguirre A, Lira-Albarrán S. Clinical applications of gonadotropins in the male. Prog Mol Biol Transl Sci. 2016;143:121–74.
- 115. Stamatiades GA, Kaiser UB. Gonadotropin regulation by pulsatile GnRH: Signaling and gene expression. Mol Cell Endocrinol. 2018;463:131–41.
- 116. Ieda N, et al. GnRH(1-5), a metabolite of gonadotropin-releasing hormone, enhances luteinizing hormone release via activation of kisspeptin neurons in female rats. Endocr J. 2020;67(4):409–18.
- 117. Rudolph LM, et al. Peripheral and central mechanisms involved in the hormonal control of male and female reproduction. J Neuroendocrinol. 2016;28:7.
- 118. Krsmanovic LZ, et al. The hypothalamic GnRH pulse generator: multiple regulatory mechanisms. Trends Endocrinol Metab. 2009;20(8):402–8.
- 119. Jayasena CN, et al. Direct comparison of the effects of intravenous kisspeptin-10, kisspeptin-54 and GnRH on gonadotrophin secretion in healthy men. Hum Reprod. 2015;30(8):1934–41.
- 120. Weinbauer GF, Hankel P, Nieschlag E. Exogenous gonadotrophin-releasing hormone (GnRH) stimulates LH secretion in male monkeys (Macaca fascicularis) treated chronically with high doses of a GnRH antagonist. J Endocrinol. 1992;133(3):439–45.
- 121. Vulliemoz NR, et al. Decrease in luteinizing hormone pulse frequency during a five-hour peripheral ghrelin infusion in the ovariectomized rhesus monkey. J Clin Endocrinol Metab. 2004;89(11):5718–23.
- 122. Lanfranco F, et al. Acylated ghrelin inhibits spontaneous luteinizing hormone pulsatility and responsiveness to naloxone but not that to gonadotropin-releasing hormone in young men: evidence for a central inhibitory action of ghrelin on the gonadal axis. J Clin Endocrinol Metab. 2008;93(9):3633–9.
- 123. Kluge M, et al. Ghrelin suppresses secretion of luteinizing hormone (LH) and follicle-stimulating hormone (FSH) in women. J Clin Endocrinol Metab. 2012;97(3):E448–51.
- 124. Hadjidakis DJ, et al. Differences between somatostatin-28 and somatostatin-14 with respect to their biological effects in healthy humans and acromegalics. Clin Physiol Biochem. 1986;4(6):372–83.
- 125. Chiodera P, et al. Inhibition by somatostatin of LH-RH-induced LH release in normal menstruating women. Gynecol Obstet Investig. 1986;22(1):17–21.
- 126. Narayanaswamy S, et al. Subcutaneous infusion of kisspeptin-54 stimulates gonadotrophin release in women and the response correlates with basal oestradiol levels. Clin Endocrinol. 2016;84(6):939–45.
- 127. Jayasena CN, et al. The effects of kisspeptin-10 on reproductive hormone release show sexual dimorphism in humans. J Clin Endocrinol Metab. 2011;96(12):E1963–72.
- 128. Ramaswamy S, Gibbs RB, Plant TM. Studies of the localisation of kisspeptin within the pituitary of the rhesus monkey (Macaca mulatta) and the effect of kisspeptin on the release of non-gonadotropic pituitary hormones. J Neuroendocrinol. 2009;21(10):795–804.
- 129. Cagnacci A, Elliott JA, Yen SS. Amplification of pulsatile LH secretion by exogenous melatonin in women. J Clin Endocrinol Metab. 1991;73(1):210–2.
- 130. Nordlund JJ, Lerner AB. The effects of oral melatonin on skin color and on the release of pituitary hormones. J Clin Endocrinol Metab. 1977;45(4):768–74.

- 131. Lisoni P, et al. Effect of an acute injection of melatonin on the basal secretion of hypophyseal hormones in prepubertal and pubertal healthy subjects. Acta Endocrinol. 1986;111(3):305–11.
- 132. Luboshitzky R, et al. Long-term melatonin administration does not alter pituitary-gonadal hormone secretion in normal men. Hum Reprod. 2000;15(1):60–5.
- Chrousos GP, Brown T, Bercu BB. Pharmacologic effects of melatonin on hypothalamic-adenohypophyseal function in the nonhuman primate. Neuroendocrinology. 1982;34(5):343–6.
- 134. Tsutsui K, et al. A novel avian hypothalamic peptide inhibiting gonadotropin release. Biochem Biophys Res Commun. 2000;275(2):661–7.
- 135. George JT, et al. Effect of gonadotropin-inhibitory hormone on luteinizing hormone secretion in humans. Clin Endocrinol. 2017;86(5):731–8.
- 136. Clarke IJ, et al. Potent action of RFamide-related peptide-3 on pituitary gonadotropes indicative of a hypophysiotropic role in the negative regulation of gonadotropin secretion. Endocrinology. 2008;149(11):5811–21.
- 137. Kadokawa H, et al. Bovine C-terminal octapeptide of RFamide-related peptide-3 suppresses luteinizing hormone (LH) secretion from the pituitary as well as pulsatile LH secretion in bovines. Domest Anim Endocrinol. 2009;36(4):219–24.
- Son YL, et al. Gonadotropin-inhibitory hormone inhibits GnRH-induced gonadotropin subunit gene transcriptions by inhibiting AC/cAMP/PKA-dependent ERK pathway in LbetaT2 cells. Endocrinology. 2012;153(5):2332–43.
- Kalra SP, Crowley WR. Neuropeptide Y: a novel neuroendocrine peptide in the control of pituitary hormone secretion, and its relation to luteinizing hormone. Front Neuroendocrinol. 1992;13(1):1–46.
- 140. Watanobe H, et al. Neuropeptide Y potentiates the luteinizing hormone (LH) response to LH-releasing hormone in men. Biochem Biophys Res Commun. 1994;200(2):1111–7.
- 141. Kaynard AH, et al. Third-ventricular infusion of neuropeptide Y suppresses luteinizing hormone secretion in ovariectomized rhesus macaques. Endocrinology. 1990;127(5):2437–44.
- 142. Sowers JR, Rice BF, Blanchard S. Effect of dexamethsone on luteinizing hormone and follicle stimulating hormone responses to LHRH and to clomiphene in the follicular phase of women with normal menstrual cycles. Horm Metab Res. 1979;11(8):478–80.
- 143. Veldhuis JD, Lizarralde G, Iranmanesh A. Divergent effects of short term glucocorticoid excess on the gonadotropic and somatotropic axes in normal men. J Clin Endocrinol Metab. 1992;74(1):96–102.
- 144. Ying SY. Inhibins, activins, and follistatins: gonadal proteins modulating the secretion of follicle-stimulating hormone. Endocr Rev. 1988;9(2):267–93.
- 145. Blumenfeld Z, Ritter M. Inhibin, activin, and follistatin in human fetal pituitary and gonadal physiology. Ann N Y Acad Sci. 2001;943:34–48.
- 146. Hayes FJ, et al. Importance of inhibin B in the regulation of FSH secretion in the human male. J Clin Endocrinol Metab. 2001;86(11):5541–6.
- 147. Stouffer RL, et al. Human recombinant activin-A alters pituitary luteinizing hormone and follicle-stimulating hormone secretion, follicular development, and steroidogenesis, during the menstrual cycle in rhesus monkeys. J Clin Endocrinol Metab. 1993;77(1):241–8.
- Gregory SJ, Kaiser UB. Regulation of gonadotropins by inhibin and activin. Semin Reprod Med. 2004;22(3):253–67.
- 149. Bilezikjian LM, et al. Cell-type specific modulation of pituitary cells by activin, inhibin and follistatin. Mol Cell Endocrinol. 2012;359(1–2):43–52.
- 150. Welt CK, Crowley WF Jr. Activin: an endocrine or paracrine agent? Eur J Endocrinol. 1998;139(5):469–71.
- 151. Fingscheidt U, et al. Regulation of gonadotrophin secretion by inhibin, testosterone and gonadotrophin-releasing hormone in pituitary cell cultures of male monkeys. J Endocrinol. 1998;159(1):103–10.
- Matsumoto AM, Bremner WJ. Modulation of pulsatile gonadotropin secretion by testosterone in man. J Clin Endocrinol Metab. 1984;58(4):609–14.
- 153. Abs R, et al. Endocrine consequences of long-term intrathecal administration of opioids. J Clin Endocrinol Metab. 2000;85(6):2215–22.

- 154. Pende A, et al. Evaluation of the effects induced by four opiate drugs, with different affinities to opioid receptor subtypes, on anterior pituitary LH, TSH, PRL and GH secretion and on cortisol secretion in normal men. Biomed Pharmacother. 1986;40(5):178–82.
- 155. Mauras N, Rogol AD, Veldhuis JD. Appraising the instantaneous secretory rates of luteinizing hormone and testosterone in response to selective mu opiate receptor blockade in late pubertal boys. J Androl. 1987;8(4):203–9.
- 156. Pirahanchi YJI. Physiology, thyroid stimulating hormone (TSH). StatPearls, 2019.
- 157. Hinkle PM, Gehret AU, Jones BW. Desensitization, trafficking, and resensitization of the pituitary thyrotropin-releasing hormone receptor. Front Neurosci. 2012;6:180.
- Bargi-Souza P, Kucka M, Bjelobaba I, Tomić M, Janjic MM, Nunes MT, Stojilkovic SS. Loss of basal and TRH-stimulated Tshb expression in dispersed pituitary cells. Endocrinology. 2015;156(1):242–54.
- 159. Günther T, Tulipano G, Dournaud P, Bousquet C, Csaba Z, Kreienkamp HJ. International Union of Basic and Clinical Pharmacology. CV. Somatostatin receptors: structure, function, ligands, and new nomenclature. Pharmacol Rev. 2018;70(4):763–835.
- 160. Spoudeas HA, Matthews DR, Brook CG, Hindmarsh PC. The effect of changing somatostatin tone on the pituitary growth hormone and thyroid-stimulating hormone responses to their respective releasing factor stimuli. J Clin Endocrinol Metabol. 1992;75(2):453–8.
- 161. Samuels MHRE. Central hypothyroidism. Endocrinol Metab Clin N Am. 1992;21:903–19.
- 162. Reichlin S. Somatostatin. N Engl J Med. 1983;309:1495-501.
- 163. Gancel A, Vuillermet P, Legrand A, Catus F, Thomas F, Kuhn JM. Effects of a slow-release formulation of the new somatostatin analogue lanreotide in TSH-secreting pituitary adenomas. Clin Endocrinol. 1994;40:3.
- 164. Beck-Peccoz PPL. Medical management of thyrotropin-secreting pituitary adenomas. Pituitary. 2002;5(2):83-8.
- 165. Theodoropoulou M, Stalla GK. Somatostatin receptors: from signaling to clinical practice. Front Neuroendocrinol. 2013;34(3):228–52.
- 166. Diego Ferone FG, Arvigo M, Resmini E, Boschetti M, Teti C, Esposito D, Minuto F. The clinical-molecular interface of somatostatin, dopamine and their receptors in pituitary pathophysiology. J Mol Endocrinol. 2009;42(5):361–70.
- 167. Burrow GN, May PB, Spaulding SW, Donabedian RK. TRH and dopamine interactions affecting pituitary hormones secretion. J Clin Endocrinol Metab. 1977;45(1):65–72.
- 168. Scanlon MF, Weightman DR, Shale DJ, Mora B, Heath M, Snow MH, Lewis M, Hall R. Dopamine is a physiological regulator of thyrotrophin (TSH) secretion in normal man. Clin Endocrinol. 1979;10(1):7–15.
- 169. Rodriguez F, Jolin T. The role of somatostatin and/or dopamine in basal and TRH-stimulated TSH release in food-restricted rats. Eur J Endocrinol. 1991;125(2):186–91.
- Spaulding SW, Burrow GN, Donabedian R, van Woert M. L-DOPA suppression of thyrotropin-releasing hormone response in man. J Clin Endocrinol Metab. 1972;35:182–5.
- 171. Besses GS, Burrow GN, Spaulding SW, Donabedian RK. Dopamine infusion acutely inhibits the TSH and prolactin response to TRH. J Clin Endocrinol Metab. 1975;41:985–8.
- 172. Leebaw WF, Lee LA, Woolf PD. Dopamine affects basal and augmented pituitary hormone secretion. J Clin Endocrinol Metab. 1978;47:480–7.
- 173. Refetoff SFV, Rapoport B, Friesen HG. Interrelationships in the regulation of TSH and prolactin secretion in man: effects of L-dopa, TRH and thyroid hormone in various combinations. J Clin Endocrinol Metab. 1974;38:450–7.
- 174. Felt VNJ. Effect of bromocryptine on the secretion of thyrotropic hormone TSH, prolactin Pr, human growth hormone HGH, thyroxine T4 and triiodothyroxine T3 in hypothyroidism. Horm Metab Res. 1977;9:274–7.
- 175. Lee ECP, Rao H, et al. Effect of acute high dose dobutamine administration on serum thyrotrophin TSH. Clin Endocrinol Oxf. 1999;50:487–92.
- Melnikov M, et al. Dopaminergic therapeutics in multiple sclerosis: focus on Th17-cell functions. J Neuroimmune Pharmacol. 2019;15:37.

- 177. Savvidou OMM, Goumenos S, Flevas D, Papagelopoulos P, Moutsatsou P. Glucocorticoid signaling and osteoarthritis. Mol Cell Endocrinol. 2019;480:153–66.
- 178. Vazquez-Borrego MC, Gahete MD, Martínez-Fuentes AJ, Fuentes-Fayos AC, Castaño JP, Kineman RD, Luque RM. Multiple signaling pathways convey central and peripheral signals to regulate pituitary function: lessons from human and non-human primate models. Mol Cell Endocrinol. 2018;463:4–22.
- 179. Trainer PJ, Holly J, Medbak S, Rees LH, Besser GM. The effect of recombinant IGF-I on anterior pituitary function in healthy volunteers. Clin Endocrinol. 1994;41(6):801–7.
- Delitala G, Grossman A, Besser GM. The participation of hypothalamic dopamine in morphine-induced prolactin release in man. Clin Endocrinol. 1983;19(4):437–44.
- 181. Pende A. e.a., Evaluation of the effects induced by four opiate drugs, with different affinities to opioid receptor subtypes, on anterior pituitary LH, TSH, PRL and GH secretion and on cortisol secretion in normal men. Biomed Pharmacother. 1986;40(5):178–82.
- Roti E. e.a., *Dermorphin*, A new opioid peptide, stimulates thyrotropin secretion in normal subjects. J Endocrinol Investig. 1984;7(3):211–4.
- Drouin J. 60 Years of POMC: transcriptional and epigenetic regulation of POMC gene expression. J Mol Endocrinol. 2016;56(4):T99–T112.
- Grammatopoulos DK. Insights into mechanisms of corticotropin-releasing hormone receptor signal transduction. Br J Pharmacol. 2012;166(1):85–97.
- 185. Nezi M, Mastorakos G, Mouslech Z. Corticotropin releasing hormone and the immune/ inflammatory response. In: Feingold KR, et al., editors. Endotext. South Dartmouth (MA): Editors; 2000.
- Aguilera G, et al. Corticotropin releasing hormone receptors: two decades later. Peptides. 2004;25(3):319–29.
- Luque RM, et al. Evidence that endogenous SST inhibits ACTH and ghrelin expression by independent pathways. Am J Physiol Endocrinol Metab. 2006;291(2):E395–403.
- 188. Lanfranco F, et al. Ghrelin and anterior pituitary function. Front Horm Res. 2010;38:206–11.
- 189. Ibáñez-Costa A, et al. In1-ghrelin splicing variant is overexpressed in pituitary adenomas and increases their aggressive features. Sci Rep. 2015;5:8714.
- 190. Chuang JC, et al. Ghrelin mediates stress-induced food-reward behavior in mice. J Clin Invest. 2011;121(7):2684–92.
- 191. Maruna P, Gurlich R, Rosicka M. Ghrelin as an acute-phase reactant during postoperative stress response. Horm Metab Res. 2008;40(6):404–9.
- 192. Brzozowski T, et al. Exogenous and endogenous ghrelin in gastroprotection against stressinduced gastric damage. Regul Pept. 2004;120(1–3):39–51.
- Schmid DA, et al. Ghrelin stimulates appetite, imagination of food, GH, ACTH, and cortisol, but does not affect leptin in normal controls. Neuropsychopharmacology. 2005;30(6):1187–92.
- 194. Lengyel AM. Novel mechanisms of growth hormone regulation: growth hormone-releasing peptides and ghrelin. Braz J Med Biol Res. 2006;39(8):1003–11.
- 195. Kanasaki H, et al. Interactions between two different G protein-coupled receptors in reproductive hormone-producing cells: the role of PACAP and its receptor PAC1R. Int J Mol Sci. 2016;17:10.
- 196. Propato-Mussafiri R, et al. Pituitary adenylate cyclase-activating polypeptide releases 7B2, adrenocorticotrophin, growth hormone and prolactin from the mouse and rat clonal pituitary cell lines AtT-20 and GH3. J Endocrinol. 1992;132(1):107–13.
- 197. Hensen J, et al. Effects of incremental infusions of arginine vasopressin on adrenocorticotropin and cortisol secretion in man. J Clin Endocrinol Metab. 1988;66(4):668–71.
- 198. Antoni FA. Novel ligand specificity of pituitary vasopressin receptors in the rat. Neuroendocrinology. 1984;39(2):186–8.
- Bankir L, Bichet DG, Morgenthaler NG. Vasopressin: physiology, assessment and osmosensation. J Intern Med. 2017;282(4):284–97.
- Kovacs KJ, Sawchenko PE. Sequence of stress-induced alterations in indices of synaptic and transcriptional activation in parvocellular neurosecretory neurons. J Neurosci. 1996;16(1):262–73.

- 201. Stafford PJ, et al. The pituitary-adrenal response to CRF-41 is unaltered by intravenous somatostatin in normal subjects. Clin Endocrinol. 1989;30(6):661–6.
- 202. Broglio F, et al. Ghrelin secretion is inhibited by either somatostatin or cortistatin in humans. J Clin Endocrinol Metab. 2002;87(10):4829–32.
- Hofland LJ. Somatostatin and somatostatin receptors in Cushing's disease. Mol Cell Endocrinol. 2008;286(1–2):199–205.
- Brown TJ, Blaustein JD. 1-(o-Chlorophenyl)-1 (p-chlorophenyl)2,2,2-trichloroethane induces functional progestin receptors in the rat hypothalamus and pituitary gland. Endocrinology. 1984;115(6):2052–8.
- 205. Kraicer J, Gajewski TC, Moor BC. Release of pro-opiomelanocortin-derived peptides from the pars intermedia and pars distalis of the rat pituitary: effect of corticotrophin-releasing factor and somatostatin. Neuroendocrinology. 1985;41(5):363–73.
- 206. Shimon I, et al. Somatostatin receptor (SSTR) subtype-selective analogues differentially suppress in vitro growth hormone and prolactin in human pituitary adenomas. Novel potential therapy for functional pituitary tumors. J Clin Invest. 1997;100(9):2386–92.
- Weeke J, Hansen AP, Lundaek K. Inhibition by somatostatin of basal levels of serum thyrotropin (TSH) in normal men. J Clin Endocrinol Metab. 1975;41(1):168–71.
- 208. van der Hoek J, et al. Distinct functional properties of native somatostatin receptor subtype 5 compared with subtype 2 in the regulation of ACTH release by corticotroph tumor cells. Am J Physiol Endocrinol Metab. 2005;289(2):E278–87.
- 209. Allolio B, et al. Effect of oral morphine and naloxone on pituitary-adrenal response in man induced by human corticotropin-releasing hormone. Acta Endocrinol. 1987;114(4):509–14.
- Naber D, et al. Naloxone effects on beta-endorphin, cortisol, prolactin, growth hormone, HVA and MHPG in plasma of normal volunteers. Psychopharmacology. 1981;74(2):125–8.
- Geer EB, et al. Stimulation of the hypothalamic-pituitary-adrenal axis with the opioid antagonist nalmefene. Pituitary. 2005;8(2):115–22.
- 212. Pfeiffer A, et al. Effects of a kappa-opioid agonist on adrenocorticotropic and diuretic function in man. Horm Metab Res. 1986;18(12):842–8.
- Waltman C, et al. Spontaneous and glucocorticoid-inhibited adrenocorticotropic hormone and cortisol secretion are similar in healthy young and old men. J Clin Endocrinol Metab. 1991;73(3):495–502.
- 214. Arvat E, et al. Effects of dexamethasone and alprazolam, a benzodiazepine, on the stimulatory effect of hexarelin, a synthetic GHRP, on ACTH, cortisol and GH secretion in humans. Neuroendocrinology. 1998;67(5):310–6.
- 215. Roelfsema F, Aoun P, Veldhuis JD. Pulsatile cortisol feedback on ACTH secretion is mediated by the glucocorticoid receptor and modulated by gender. J Clin Endocrinol Metab. 2016;101(11):4094–102.
- 216. Brown CH. Magnocellular neurons and posterior pituitary function. Compr Physiol. 2016;6(4):1701-41.
- 217. Larkin S, Ansorge O. *Development and microscopic anatomy of the pituitary gland*. In: Feingold KR, et al., editors. *Endotext*. South Dartmouth, MA: Editors; 2000.
- Brown CH, et al. Physiological regulation of magnocellular neurosecretory cell activity: integration of intrinsic, local and afferent mechanisms. J Neuroendocrinol. 2013;25(8):678–710.
- 219. Kimura T, et al. Structure and expression of a human oxytocin receptor. Nature. 1992;356(6369):526–9.
- 220. Russell JA, Leng G. Sex, parturition and motherhood without oxytocin? J Endocrinol. 1998;157(3):343–59.
- 221. Dale HH. On some physiological actions of ergot. J Physiol. 1906;34(3):163-206.
- 222. Jones C, et al. Oxytocin and social functioning. Dialogues Clin Neurosci. 2017;19(2):193–201.
- 223. Mohr E, et al. Expression of the vasopressin and oxytocin genes in rats occurs in mutually exclusive sets of hypothalamic neurons. FEBS Lett. 1988;242(1):144–8.
- 224. Lozic M, et al. Vasopressin, central autonomic control and blood pressure regulation. Curr Hypertens Rep. 2018;20(2):11.

- 225. Robertson GL. The regulation of vasopressin function in health and disease. Recent Prog Horm Res. 1976;33:333–85.
- 226. Cesselin F. Opioid and anti-opioid peptides. Fundam Clin Pharmacol. 1995;9(5):409–33.
- Martin R, Voigt KH. Enkephalins co-exist with oxytocin and vasopressin in nerve terminals of rat neurohypophysis. Nature. 1981;289(5797):502–4.
- 228. Opioids, in LiverTox: Clinical and research information on drug-induced liver injury. 2012: Bethesda (MD).
- 229. Wood CE, Silbiger J. Does cortisol inhibit vasopressin secretion in sheep? Domest Anim Endocrinol. 1988;5(2):177–83.
- Travis RH, Share L. Vasopressin-renin-cortisol interrelations. Endocrinology. 1971;89(1):246–53.
- 231. Livesey JH, et al. The effects of cortisol, vasopressin (AVP), and corticotropin-releasing factor administration on pulsatile adrenocorticotropin, alpha-melanocyte-stimulating hormone, and AVP secretion in the pituitary venous effluent of the horse. Endocrinology. 1988;123(2):713–20.
- 232. Phillips PA, et al. Angiotensin II-induced thirst and vasopressin release in man. Clin Sci (Lond). 1985;68(6):669–74.
- 233. Kilcoyne MM, Hoffman DL, Zimmerman EA. Immunocytochemical localization of angiotensin II and vasopressin in rat hypothalamus: evidence for production in the same neuron. Clin Sci (Lond). 1980;59(Suppl 6):57s–60s.
- Brooks VL, Keil LC, Reid IA. Role of the renin-angiotensin system in the control of vasopressin secretion in conscious dogs. Circ Res. 1986;58(6):829–38.
- 235. Morton JJ, et al. The role of plasma osmolality, angiotensin II and dopamine in vasopressin release in man. Clin Endocrinol. 1985;23(2):129–38.
- Dluzen DE, Muraoka S, Landgraf R. Olfactory bulb norepinephrine depletion abolishes vasopressin and oxytocin preservation of social recognition responses in rats. Neurosci Lett. 1998;254(3):161–4.
- 237. Ottesen B, et al. Vasoactive intestinal peptide (VIP) stimulates oxytocin and vasopressin release from the neurohypophyis. Endocrinology. 1984;115(4):1648–50.
- 238. Koenig JI, et al. Potential involvement of galanin in the regulation of fluid homeostasis in the rat. Regul Pept. 1989;24(1):81–6.
- Dayanithi G, Cazalis M, Nordmann JJ. Relaxin affects the release of oxytocin and vasopressin from the neurohypophysis. Nature. 1987;325(6107):813–6.
- Christensen JD, Hansen EW, Fjalland B. Interleukin-1 beta stimulates the release of vasopressin from rat neurohypophysis. Eur J Pharmacol. 1989;171(2–3):233–5.
- Mains RE, Eipper BA. Synthesis and secretion of corticotropins, melanotropins, and endorphins by rat intermediate pituitary cells. J Biol Chem. 1979;16(254):7885–94.
- 242. Takahashi A. Melanocyte-stimulating hormone. In: Handbook of hormones; 2016. p. 120-e16B-7.
- 243. Ellacott KL, Cone RD. The role of the central melanocortin system in the regulation of food intake and energy homeostasis: lessons from mouse models. Philos Trans R Soc Lond Ser B Biol Sci. 2006;361(1471):1265–74.
- 244. Begriche K, et al. Genetic dissection of the functions of the melanocortin-3 receptor, a seventransmembrane G-protein-coupled receptor, suggests roles for central and peripheral receptors in energy homeostasis. J Biol Chem. 2011;286(47):40771–81.
- Baldini G, Phelan KD. The melanocortin pathway and control of appetite-progress and therapeutic implications. J Endocrinol. 2019;241(1):R1–R33.
- 246. Saporiti F, et al. Melanocortin-1 receptor positively regulates human artery endothelial cell migration. Cell Physiol Biochem. 2019;52(6):1339–60.
- Stojilkovic SS, Tabak J, Bertram R. Ion channels and signaling in the pituitary gland. Endocr Rev. 2010;31(6):845–915.
- 248. R Vázquez-Martínez R, Malagón MM, Castaño JP, Tonon MC, Vaudry H, Gracia-Navarro F. Amphibian melanotrope subpopulations respond differentially to hypothalamic Secretoinhibitors. Neuroendocrinology. 2001;73:426–34.

- 249. Roubos EW, Scheenen WJ, Jenks BG. Neuronal, neurohormonal, and autocrine control of Xenopus melanotrope cell activity. Ann NY Acad Sci. 2005;1040:172–83.
- 250. Kuribara M, et al. BDNF stimulates Ca2+ oscillation frequency in melanotrope cells of Xenopus laevis: contribution of IP3-receptor-mediated release of intracellular Ca2+ to gene expression. Gen Comp Endocrinol. 2010;169(2):123–9.
- 251. van den Hurk MJ, et al. Expression and characterization of the extracellular ca(2+)-sensing receptor in melanotrope cells of Xenopus laevis. Endocrinology. 2003;144(6):2524–33.



3

Pathogenesis of Pituitary Adenomas

Sicheng Tang, Adriana Albani, and Marily Theodoropoulou

3.1 Introduction

Pituitary tumors are in their majority sporadic (~95%). Since the early 1990s, X-chromosome inactivation studies on pituitary tumors of different histological backgrounds (gonadotroph, somatotroph, lactotroph, corticotroph) have established that pituitary tumors are monoclonal [1–4]. This entails that one genetic hit transforms a cell that gives rise to the whole tumor. Mutations in proto-oncogenes such as *CMYC*, *RAS*, and *BRAF* are rare in pituitary tumors [5–7] (reviewed in [8]). Germline genetic defects in tumors in familiar or syndromic setting are not in the scope of this chapter. Mutations in genes that are involved in endocrine neoplasia syndromes such as *MEN1* (multiple endocrine neoplasia 1), *PRKAR1A* (Carney complex), *CDKN1B* (MEN4), and *GPR101* (X-LAG) are extremely rare in sporadic pituitary tumors (reviewed in [9]).

As we will see in more detail below, the main genetic hotspots are found in GNAS and USP8 genes, mutated in ~40% and ~50% of sporadic somatotroph and corticotroph tumors respectively. The majority of pituitary tumors do not have known driver mutations. In contrast, numerous studies have reported posttranscriptional/posttranslational defects and aberrant signaling pathways that may trigger or contribute to pituitary tumorigenesis and progression. Furthermore, recent big data discoveries brought to our attention previously obscure putative mechanisms of pituitary tumor development and progression. This chapter will provide a brief

Medizinische Klinik und Poliklinik IV, Ludwig Maximilian University Munich, Munich, Germany

e-mail: s-stang@helios.med.uni-muenchen.de; Adriana.Albani@med.uni-muenchen.de; Marily.Theodoropoulou@med.uni-muenchen.de

© Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_3

The chapter has been endorsed by **Prof. Justo Castaño**, justo@uco.es, Instituto Maimónides de Investigación Biomédica de Córdoba, University of Córdoba, Córdoba, Spain

S. Tang · A. Albani · M. Theodoropoulou (🖂)

overview on the deregulated hypothalamic and peripheral signals and the signaling networks deregulated in sporadic pituitary tumors, before addressing the genetics of sporadic pituitary tumors.

3.2 Hypothalamic and Peripheral Feedback Regulatory Circuits

3.2.1 Hypothalamic Regulation

The trophic action of hypothalamic factors on pituitary cells suggested that aberrant hypothalamic regulation might trigger pituitary hyperplasia and subsequent tumor formation. Animal models offer evidence of a hypothalamic role in pituitary tumorigenesis. Growth hormone-releasing hormone (GHRH) transgenic mice develop somatotroph hyperplasia, while female dopamine receptor type 2 DRD2 knockout mice present with lactotroph hyperplasia and tumors [10–12]. However, clinical experience does not offer similar evidence in human patients: patients with ectopic GHRH or CRH producing neuroendocrine tumors rarely present with somatotroph or corticotroph hyperplasia and do not develop tumors [13]. In the early 1990s, the establishment of monoclonality pushed the hypothalamic hypothesis of pituitary tumorigenesis aside [1–4].

Receptors for the hypothalamic peptides GHRH (GHRHR), corticotrophinreleasing hormone (CRH; CRHR1), vasopressin (V3R, V1BR), thyrotrophin releasing hormone (TRH; TRHR1, TRHR2), and gonadotropin-releasing hormone receptor (GnRH; GNRHR, also abbreviated as LHRHR) are detected in varying degrees of abundance in the different human pituitary tumor subtypes [14-18]. Dopamine receptor type 2 (DRD2, also abbreviated as D2R or D2DR) is expressed in lactotroph as well as somatotroph, corticotroph, and nonfunctioning pituitary tumors [19]. DRD2 is alternatively spliced and in gonadotroph tumors, higher expression of the long over the short isoform has been associated with poor response to dopamine agonists [20]. Somatostatin receptors (SSTR1-5) are expressed in pituitary tumors in various combinations: somatotroph (mainly SSTR2), lactosomatotroph (SSTR2 and SSTR5), corticotroph (mainly SSTR5), and gonadotroph (mainly SSTR3). Reduced SSTR2 expression in sparsely granulated somatotroph tumors has been associated with worse prognosis [21]. Furthermore, somatotroph tumors expressing a truncated SSTR5 variant (SST₅TDM4), which does not respond to somatostatin analogs, present with more aggressive phenotype and have worse prognosis [22].

Constitutively activating mutations in the *GHRHR* are rare in somatotroph tumors [23, 24] and no activating *TRHR* and *GNRHR* mutations were reported in thyrotroph and gonadotroph tumors [25, 26]. Similarly, studies in corticotroph tumors did not reveal mutations in the coding regions of *CRHR1*, *V3R*, and *V1BR* [16, 27, 28]. Inactivating mutations were not reported in the *DRD2* gene encoding for dopamine receptor 2 in lactotroph tumors nor in the SSTR2 or SSTR5 genes in somatotroph tumors [29–33]. One exception is the report of a germline mutation in the *SSTR5* gene in a patient with acromegaly resistant to the somatostatin analog octreotide [34].

3.2.2 Negative Feedback Regulation

Pituitary function is tightly controlled by negative hormonal feedback loops from the periphery. Examples include the negative feedback of glucocorticoids on ACTH, thyroid hormones on TSH, and gonadal hormones on FSH/LH. Accordingly, compromised peripheral feedback may play a role in pituitary tumorigenesis through not only the uncontrolled hormone production, but also aberrant proliferation of the corresponding pituitary tumor cells. However, there is little clinical evidence supporting their tumorigenic role; thyrotroph (and in a lesser extend lactotroph) hyperplasia has been observed in few patients with primary hypothyroidism [35, 36]. Patients with primary hypogonadism or chronic sex steroid hormone treatment show no higher incidence of pituitary tumors in comparison to the general population [37–39].

Similar to the hypothalamic hypothesis, there is no circumstantial evidence that compromised peripheral negative feedback is initiating factor of pituitary tumorigenesis in humans. Genetic variations or mutations in the *NR3C1* gene are very rare in patients with Cushing's disease [40–42]. Similarly, no mutations were found in the genes encoding for TR β and rare mutations of unknown functional significance in the gene encoding for TR α [43].

On the other hand, transformed pituitary tumor cells may develop resistance to the negative feedback regulation and this would confer survival benefits to the tumor. In fact glucocorticoid resistance in corticotroph tumors is a hallmark of Cushing's disease [44]. As glucocorticoid receptor GR is not downregulated in corticotroph tumors [40–42], this glucocorticoid resistance was attributed to elevated expression of 11 β -hydroxysteroid dehydrogenases 2 (11 β -HSD2), which converts cortisol to cortisone thereby inactivating cortisol [45] and loss of Brahma-related gene 1 (BRG1), which is part of the GR repressor complex on the *POMC* promoter [46, 47]. Overexpression of the testicular orphan nuclear receptor 4 (TR4) may also promote glucocorticoid resistance as its interaction with GR abolishes GR's inhibitory effect on *POMC* transcription [48]. Finally, corticotroph tumors overexpress the GR chaperon HSP90 (heat shock protein 90) and this may impair the function of GR by preventing its maturation [49]. Indeed, the C-terminal HSP90 inhibitor silibinin was able to reverse glucocorticoid resistance and ameliorate the signs of Cushing's disease *in vitro* and *in vivo* [49].

3.3 Signaling and Cellular Processes

3.3.1 EGF & EGFR

Epidermal growth factor (EGF) is a prototype growth factor and the overexpression of its receptor EGFR is a hallmark of several human cancers. EGF is expressed in the pituitary gland where it may regulate pituitary function in an autocrine/paracrine manner (reviewed in [50]). It is noteworthy that in anterior pituitary cells, EGF alters endocrine cell function and hormone response, but is not mitogenic, choosing a cell differentiation role over its growth-promoting function [51].

EGFR expression was described in pituitary tumors with different studies reporting different prevalence depending on the method and—in case of immunohistochemical investigation—antibody used. Accordingly, EGFR overexpression was reported in clinically silent tumors compared to endocrinologically functional ones, in all subtypes of pituitary tumors, in invasive versus non-invasive, and in lactotroph and corticotroph tumors versus the nonfunctioning ones [52–56]. EGFR is predominantly expressed in corticotroph cells where EGF triggers replication and ACTH synthesis (reviewed in [57]). This is reflected by the fact that EGF triggers bromodeoxyuridine (BrdU) incorporation in dispersed corticotroph and lactotroph cells, but not in other pituitary cell subtypes [58]. The predominance of EGFR in corticotroph and lactotroph tumors indicates that the EGFR system may represent a putative therapeutic target [56, 59–61].

3.3.2 TGF-β

Members of the transforming growth factors β (TGF- β) family regulate cell differentiation, proliferation, extracellular matrix, and angiogenesis. The receptors of the family belong to the serine/threonine kinase receptor superfamily and transduce their signals via SMAD that upon phosphorylation translocate to the nucleus to regulate transcription. TGF- β 1 has a predominant role in lactotroph cells where it inhibits prolactin synthesis and proliferation [62]. Accordingly, its downregulation in lactotroph tumors was hypothesized to contribute to the development of these tumors. In fact, TGF- β 1 crosstalks with both estradiol and dopamine, both very important regulators of lactotroph function (reviewed in [63]). Moreover, differences in the expression of TGF- β 1 in males and females may explain gender differences in the incidence of prolactinomas [63, 64].

An important pituitary development member of the TGF- β family is the bone morphogenetic protein 4 (BMP4), which is required for the formation of Rathke's pouch (reviewed in [65]). BMP4 is overexpressed in lactotroph tumors and its crosstalk with estrogen receptor via Smad induces proliferation and PRL production, suggesting a crucial role in lactotroph tumorigenesis [66]. BMP4 expression in the pituitary gland is moderate where it is predominantly located in the corticotroph cells. BMP4 is downregulated in corticotroph tumors, where it inhibits proliferation and ACTH production *in vitro* and *in vivo* [67].

3.3.3 Cytokines

The pituitary gland expresses cytokines and their receptors, which regulate hormone synthesis in an autocrine-paracrine manner [50, 68]. Interleukin-1 (IL-1) is expressed in human pituitary tumors and rat anterior pituitary where it regulates growth and stimulates the secretion of anterior pituitary hormones, with the exception of prolactin [69, 70]. Similarly, IL-2 and its receptor are expressed in all
anterior pituitary cell subtypes, but IL-2 plays a particular role in corticotroph cells where it enhances ACTH synthesis [71–73].

The gp130 cytokine family is a heterogeneous group of cytokines (interleukin-6 (IL-6), IL-11, oncostatin M, ciliar neurotrophic factor (CNTF), leukemia inhibitory factor [74], and others) whose receptors have no intracellular signaling domain and thus use the gp130 membrane protein as a common signal transducer to activate the JAK-STAT signaling pathway [75]. Leukemia inhibitory factor [74] stimulates ACTH synthesis in synergy with CRH [76–78]. LIF overexpressing transgenic mice present with high ACTH and cortisol levels, reduced sensitivity to dexamethasone and cushingoid features [79] (reviewed in [80]). In contrast, the physiological HPA-mediated stress response is attenuated in LIF-deficient animals [81, 82].

IL-6 and its receptor are expressed in pituitary tumors where they correlate with tumor size and invasiveness and may trigger pituitary cell growth in autocrine/paracrine manner [83–87]. Inhibition of the signal transducer gp130, which mediates among the others the effects of IL-6, inhibits the growth of lactosomatotroph GH3 tumors in nude mice [88]. IL-6 stimulates ACTH, GH, and PRL secretion from corticotroph, somatotroph, and lactotroph tumors, respectively (reviewed in [75] [89]). Interestingly, IL-6 was detected in the secretome of tumor-associated fibroblast isolated from pituitary tumors, and its levels were significantly higher in invasive tumors [90]. As we will see in more detail below, there is evidence that IL-6 with its autocrine action contributes to pituitary senescence [91].

3.3.4 Hedgehog Pathway

Sonic hedgehog (Shh) signaling plays an important role in pituitary development [92]. Hedgehog proteins bind to Patched (PTCH) 1 and 2, releasing them from Smoothened (SMO) and enabling them to activate the GLI transcription factors. Shh expression in the adult pituitary gland concentrates mainly in corticotroph cells, where it triggers ACTH synthesis via a GLI1-CRHR1 crosstalk [93]. The majority of pituitary tumors express PTCH1 (mainly gonadotroph) and PTCH2 (mainly corticotroph, somatotroph, lactotroph), but not Shh itself, and Shh administration in dispersed anterior pituitary cells and human pituitary tumors increases ACTH, GH, and prolactin levels [94, 95]. Shh treatment in the murine corticotroph tumor cell line AtT-20 inhibited cell growth, which together with the loss of expression in human corticotroph tumors led to the hypothesis that it may act as a tumor suppressor [93, 94]. In contrast, studies in murine pituitary explants revealed increased proliferation of Sox2+ and Sox9+ stem cells after Shh stimulation [95].

3.3.5 Wnt Pathway

Wingless (Wnt) acts in paracrine manner to control cell proliferation, differentiation, and migration during embryogenesis. After binding a heterodimeric receptor complex, formed by Frizzled (Fzd) and LRP5/6 co-receptor protein, Wnt triggers the activation of three different intracellular molecular pathways: the β-catenin "canonical" pathway, the "calcium", and the "planar cell polarity pathway" [96]. Wnt4 and Wnt5 play an important role in pituitary cell differentiation [97]. WNT4 and its receptor FZD6 were shown to be overexpressed in somatotroph, lactotroph, and thyrotroph tumors, while the Wnt pathway inhibitor frizzled-related protein 2 (SFRP2) was shown to be downregulated [74, 98]. In corticotroph tumors, SFRP2 reduction correlated with tumor size and invasiveness and its overexpression suppressed proliferation and ACTH secretion in immortalized corticotroph tumor cells [99].

Regarding the downstream target of the canonical Wnt pathway, a study reported nuclear accumulation of β -catenin in 57% of pituitary tumors and not in the normal pituitary [100]. In contrast, other studies did not observe changes in β -catenin distribution in pituitary tumors [74, 98, 101].

3.3.6 Hippo Pathway

The Hippo pathway plays an important role in controlling stem cells proliferation and tumorigenesis. When the Hippo pathway is activated, mammalian sterile 20-like kinases 1 and 2 (MST1/2; ortholog of *Drosophila* Hippo) phosphorylate and activate the tumor suppressor homologs 1 and 2 (LATS1/2), which in turn phosphorylate and inactivate the transcriptional co-activators Yes-associated protein (YAP) and WW domain-containing transcription factor (TAZ/WWTR1) [102]. An inactive upstream pathway allows YAP/TAZ to translocate to the nucleus where they associate with TEAD transcription factors (TEA domain family members 1–4) eventually promoting stem cell proliferation and preventing apoptosis.

Mice deficient in *Lats1* develop hyperplasia of the anterior pituitary lobe, but show decreased secretion of pituitary hormones suggesting repressed endocrine differentiation [103]. Knocking down *Last1* in vitro leads to decreased anterior pituitary hormone synthesis and high YAP/TAZ expression was reported in null cell tumors, adamantinomatous, and papillary craniopharyngiomas, but not in hormone secreting pituitary tumors [104]. Human and mouse pituitary glands during embryonic and postnatal development show strong nuclear localization of YAP/TAZ exclusively in SOX2+ cells [104, 105]. As the Hippo pathway regulates SOX2+ pituitary stem cells (PSC) clonal expansion and specification, postnatal deletion of LATS kinases with subsequent upregulation of YAP/TAZ signaling may lead to loss of differentiation of SOX2+ PSC and uncontrolled clonal expansion, resulting in hormonally nonfunctioning pituitary tumors [106].

3.3.7 Cadherins

The pituitary gland expresses epithelial cadherin (E-cadherin), but reduced cytoplasmatic staining is observed in somatotroph, lactotroph, and hormonally nonfunctioning pituitary tumors [107–111]. No mutations in the *CDH1* gene or changes in transcript levels were reported for the majority of pituitary tumors. Instead, strong nuclear staining is observed, indicating aberrant nuclear translocation as a cause of E-cadherin loss from the cell membrane [112]. In somatotroph tumors, E-cadherin protein levels correlate with SSTR2 expression and response to somatostatin analogs [111]. Furthermore, reduced cytoplasmic E-cadherin correlated with tumor invasiveness and recurrence [113]. A study reported missense mutations in the gene encoding for cadherin-related 23 (*CDH23*) in 33% (4/12) of familial pituitary tumors and 12% (15/25) of sporadic pituitary tumors versus 0.8% (2/260) of controls carrying functional *CDH23* variants [114].

3.3.8 Cell Cycle

Cyclin-dependent kinases (CDKs) and their associated activators (cyclins) promote cell cycle transition and this is inhibited by CDK inhibitors [115]. Transgenic/knockout mouse models of genes involved in cell cycle regulation present with pituitary hyperplasia and tumors (reviewed in [116]).

The tumor suppressor RB encodes for retinoblastoma, a pocket protein that binds to E2 family (E2F) transcription factors preventing the transcription of genes important for DNA synthesis [117]. Heterozygous mice with disrupted Rb present with pituitary tumors [118]. However, human pituitary tumors do not present with *RB1* mutations, with the exception of extremely rare aggressive cases [119–121]. In contrast, *RB1* gene expression is downregulated in around 1/3 of pituitary tumors, possibly due to promoter hypermethylation [122]. An E2F1 target, the high mobility group protein A (HMGA), was reported to be highly expressed in lactotroph and somatotroph tumors [123, 124].

Retinoblastoma is hyperphosphorylated and inactivated by cyclin D-CDK4/6 and cyclin E/CDK2 complexes. Cyclin D1 is encoded by the *CCND1* gene and binds to CDK4. CDK4 deficiency is characterized by pituitary hypoplasia with the somatotroph and lactotroph axes being particularly affected [125, 126]. In fact, lactotroph cells in CDK4-deficient mice cannot proliferate in response to estrogen [125]. Cyclin D1 is overexpressed in nonfunctioning pituitary tumors [127]. Nuclear cyclin D1 immunoreactivity is more prominent in aggressive and nonfunctioning subtypes, and correlates positively with the Ki67 proliferation index and tumor size [74, 127, 128]. In contrast, cyclin D1 levels in corticotroph tumors are lower compared to nonfunctioning pituitary tumors [129]. Both invasive and non-invasive pituitary tumors may present with allelic imbalance in cyclin D1 (*CCND1*, chromosome 11q13), while certain *CCND1* genotypes are associated with tumor grade [130–132]. Cyclin D3 was found to be overexpressed in gonadotroph tumors [133].

Cyclin E is highly expressed, while protein (but not transcript) levels of its CDK inhibitor p27 are downregulated/lost predominantly in corticotroph tumors [127, 134]. Genetically engineered mice overexpressing cyclin E and haploinsufficient for p27 showed increased frequency, size, and proliferation index of pituitary tumors [135]. Corticotroph tumors show loss of the transcriptional regulator

Brahma-related gene 1 (BRG1) that inhibits cyclin E expression, with concomitant high cyclin E expression and p27 loss [136]. Pharmacological inhibition of CDK2/ cyclin E with roscovitine was shown to decrease ACTH synthesis in vitro and from corticotroph tumor xenografts *in vivo* [137]. Finally, cyclin A is highly expressed in recurrent pituitary tumors [138], but its expression in nonfunctioning pituitary adenomas did not correlate with invasion and proliferation [139].

CDK/cyclins are inhibited by CDK inhibitors belonging to the CIP/KIP (CDK Interacting Protein/Kinase Inhibitory Protein; representative members: p27, p21) and INK (INhibitors of CDK; representative members: p16, p18) families. As we have seen above the CDK2/Cyclin E inhibitor p27/Kip1 is downregulated or lost in the majority of corticotroph tumors [134]. Transcription of the CDKN1B gene that encodes for p27 is not altered and the observed downregulation is at the protein level. High EGFR levels were suggested to account for the reduced p27 in these tumors [56]. In addition, BRG1 decrease in corticotroph tumors leads to cyclin E accumulation, which in complex with CDK2 phosphorylates p27 and marks it for proteasomal degradation [136]. In line with this model, corticotroph tumors have relatively high phosphorylated p27 levels [140]. In contrast DKC1 (dyskeratosis congenital/dyskerin) that regulates p27 translation is not deregulated in the majority of pituitary tumors [141]. In addition to corticotroph tumors, p27 downregulation was also reported in other types of pituitary tumors where it correlates with invasiveness [142–144]. Mice with disrupted p27 show hyperplasia and tumors of the intermediate lobe and p27 downregulation sensitizes somatotroph cells to the proliferative action of GHRH excess in p27/GHRH transgenic mice [145-147]. Spontaneous mutation in the Cdkn1b gene in rats gives rise to a multiple endocrine neoplasia syndrome (MENX) that is characterized by focal non-secreting pituitary tumors [148].

Members of the INK family of CDK inhibitors like p16/INK4a and p18/INK4c are downregulated or lost in pituitary tumors [149–151]. Introducing p16 in murine pituitary tumor AtT-20 cells that do not express endogenous p16 arrested cells at G1 and reduced proliferation [152]. Loss of p16 expression is mainly observed in non-functioning (including silent corticotroph tumors), but not in hormone secreting such as somatotroph or corticotroph tumors [129, 153–155]. This may be due to promoter hypermethylation on the *CDKN2A* gene (that encodes for p16) that was frequently observed in gonadotroph tumors, but not as much in somatotroph or corticotroph tumors [156]. In addition to *CDKN2A*, hypermethylation was detected on *RB1* and *CDKN2B* (p15) gene promoters, with *RB1* and *CDKN2A* (p16) promoter methylations being mutually exclusive [157].

The CDK inhibitor p18 is downregulated in pituitary tumors and most prominently in corticotroph tumors [158]. Loss in the *CDKN2C* gene (that encodes for p18) was found in one-fourth of human pituitary tumors and *CDKN2C* promoter hypermethylation was reported [149, 153–155]. Concomitant loss of p18 and p27 in mice results in the development of large pituitary tumors suggesting a cooperation between these two cell cycle regulators [159].

The other member of the CIP/KIP family of CDK inhibitors is p21/CIP1. Mice deficient for both members of the CIP family, p21 and p27, present with pituitary tumors [160].

Decreased nuclear p21 immunoreactivity was described in all pituitary tumor types with the notable exception of sparsely granulated somatotroph tumors [161–163]. Lactotroph tumors were found to overexpress MIR-93, which targets the expression of *CDKN1A* that encodes for p21 [164].

3.3.9 PTTG

The pituitary tumor transforming gene (PTTG), also known as securing, regulates sister chromatid separation during cell division and its deregulation results in aneuploidy and chromosomal instability [165-169]. *Pttg* deletion *in vivo* induces p21 and disrupts the development of pituitary tumors in *Rb+/–* mice [170]. PTTG is expressed in most pituitary tumors, with higher expression positively correlating with tumor invasiveness and recurrence/regrowth [128, 171-175]. PTTG was hypothesized to promote pituitary tumorigenesis by deregulating RB1 and p21, via bFGF (FGF2) and VEGF and by activating the c-MYC oncogene [176-181]. PTTG stabilization by the RWD-containing sumoylation enhancer (RWDD3 or RSUME) may account for its abundance in pituitary tumors [182]. In somatotroph cells high PTTG expression is accompanied by increased p21 and other markers of senescence [170].

3.3.10 Senescence

Cellular senescence is an irreversible cell cycle arrest in G1 phase that occurs as a consequence of multiple stimuli, such as DNA damage, aneuploidy, oxidative stress, oncogene activation, and telomere shortening in aged tissues [183, 184]. Oncogene-induced senescence has been associated with activated p53/p21 and RB/p16 pathways [185]. Pituitary tumors show increased expression of senescence markers such as p53, p19, p21, cyclin D, and senescence-associated- β -galactosidase (SA- β -Gal), while senescence has been linked especially in somatotroph tumors [170, 186, 187]. As oncogene-induced senescence halts cell proliferation in the early stages of transformation; its presence may explain the usually benign nature of pituitary tumors (reviewed in [188]).

Senescent cells influence tumor microenvironment via secreted factors collectively referred to as senescence-associated secretory phenotype (SASP) (reviewed in [189]). The SASP cytokine IL-6 may contribute to pituitary senescence in an autocrine manner, since endogenous IL-6 inhibition decreased SA- β -gal in human pituitary tumors in vitro [91].

3.4 Genetics of Sporadic Pituitary Tumors

3.4.1 Chromosome Instability

Loss of heterozygosity (LOH) studies had revealed loss of the long arm of chromosome 11 (11q13) at the *MEN1* locus, short arm of chromosome 9, and the long arm of chromosome 10 (10q26), with LOH in 11q13, 13q, and 10q26 being more frequent in invasive pituitary tumors [190–195]. LOH at the *RB1* gene locus in the long arm of chromosome 13 (13q) was observed in invasive pituitary tumors and carcinomas [121, 196]. Pituitary tumors display gene copy number variations (CNV) with higher frequency in functioning compared to hormone inactive tumors [197– 204]. Lactotroph tumors were found to display the highest CNV compared to corticotroph and somatotroph, while gonadotroph tumors showed no CNV [205]. Corticotroph tumors from pediatric patients show high chromosomal instability in ~20% of cases, which correlated with aggressiveness [206]. Overall, chromosomal alterations depend on the secretion type with secreting tumors showing higher variations compared to silent (gonadotroph, silent corticotroph) types, while no correlation was found with tumor aggressiveness [204].

3.4.2 Mutations

Mutations in oncogenes and tumor suppressor genes frequently reported in other cancers are very rare in pituitary tumors. Recent next-generation sequencing efforts reveal that only two genes, *GNAS* and *USP8*, are mutated in >5% of sporadic pituitary tumors [201, 202, 204]. No other driver mutations were found and overall the number of mutations in pituitary tumors is relatively low [201, 202, 204, 207–210].

3.4.3 GNAS

The first genetic hotspot in sporadic pituitary tumors was identified in a hypothesisbased approach, since the receptors of the aforementioned hypothalamic peptides belong to the G protein-coupled receptor superfamily and are coupled to G proteins. In the mid-1980s, studies reported elevated adenosine cyclase (AC) activity and cAMP levels in one-third of somatotroph tumors that were not influenced by GHRH and cholera toxin, but were caused by activating mutations in the gene encoding for the stimulatory G alpha subunit (Gs α) [211–213]. Somatic mutations in the *GNAS* gene (previously referred to as *gsp* proto-oncogene) are found in ~40% of somatotroph tumors, but very rarely in corticotroph and other pituitary tumors [214]. Somatotroph hyperplasia and tumors are observed in the context of McCune– Albright syndrome, a multi-organ mosaicism disorder that is caused when somatic *GNAS1* mutations occur in post-zygotic state (reviewed in [215]). In contrast to *GNAS*, mutations in the genes encoding for G protein subunit alpha i2 (*GNAI2*; also known as *GIP2*) and Gq alpha subunit (*GNAQ*) are very rare in pituitary tumors [214].

3.4.4 USP8

Whole exome sequencing in corticotroph tumors led to the discovery of a single somatic mutational hotspot in the gene encoding for ubiquitin specific protease 8 (USP8) [216, 217]. Follow-up sequencing studies revealed that ~50% of cortico-troph tumors (including cases of corticotroph tumor progression after bilateral adrenalectomy also known as Nelson's syndrome) have somatic *USP8* mutations [216–221]. Notably, *USP8* mutations are found exclusively in corticotroph tumors and not in any other pituitary tumor subtype nor in ACTH-secreting ectopic neuro-endocrine tumors [222]. A germline USP8 mutation was identified in a female pediatric patient presenting with corticotroph tumor [223].

USP8 is a deubiquitinase that cleaves ubiquitin from target proteins, rescuing them from degradation. The mutational hotspot identified in corticotroph tumors is located in exon 14 and prevents the binding of 14-3-3 proteins, thereby enabling the cleavage to a highly catalytically active C-terminal fragment [216]. Overactivated USP8 mutants rescue ligand-bound EGFR from the lysosome and potentiate *POMC* expression and ACTH synthesis [216]. USP8 mutant corticotroph tumors have higher *POMC* levels [219]. USP8 mutants have less potent effect on cell growth in vitro, which is reflected by the relatively smaller size and lack of invasion of *USP8* mutant corticotroph tumors [216–218].

Overall, corticotroph tumors carrying USP8 mutations are usually smaller, occur mainly in female patients and may be hormonally more active, as reflected by higher basal and postoperative cortisol levels [218]. Patients with USP8 mutant tumors show worse postsurgical outcome as reflected by the lower prevalence of adrenal insufficiency after surgery [218]. Indeed, in these patients, the frequency of recurrence after postsurgical biochemical remission is higher and occurs earlier [221]. Interestingly, USP8 mutant corticotroph tumors display higher SSTR5 levels suggesting that they may respond favorably to pasireotide [204, 219, 224].

3.4.5 USP48

Whole exome sequencing in USP8-wild type corticotroph tumors revealed a second somatic mutational hotspot in the gene encoding for the USP48 deubiquitinase in about 10% of corticotroph tumors [225, 226]. USP48 is a deubiquitinase first identified in the rat brain as synUSP [227]. The pathogenic mutations are mainly concentrated on a single amino acid (Met415) and result in amino acid exchange of Met415Ile or Met415Val [226]. Mutant USP48 showed modest stimulatory action

on basal but potentiated CRH-induced ACTH synthesis [225]. One of the USP48 clients is Gli1 and knocking down Gli1 in corticotroph tumor cells abolished its stimulatory effect suggesting that mutant USP48 may use this mechanism to sensitize corticotroph tumor cells to the stimulatory action of CRH [93, 225].

3.4.6 SF3B1

Whole genome sequencing on lactotroph tumors identified a somatic mutational hotspot in the *SF3B1* gene encoding for splicing factor 3 subunit B1 [228]. The *SF3B1 R625H* hotspot mutation was found in ~20% of lactotroph tumors (but no other pituitary tumor type) and patients with *SF3B1* mutant tumors were more frequently male, had higher prolactin levels (relative to tumor size), and shorter progression-free survival; no statistically significant differences were found with tumor invasion and size [228]. SF3B1 is a component of the U2 small nuclear ribonucleoproteins (snRNP) complex that is involved in pre-mRNA splicing and one of SF3B1 splicing targets is *ESRRG* (Estrogen Related Receptor Gamma). The mutant SF3BP1 results in cryptic ESRRG, which has high affinity for the transcription factor Pit-1 that drives pituitary *PRL* gene expression, resulting in prolactin overproduction. Interestingly, inhibiting SF3B1 with pladienolide-B significantly reduced cell viability in pituitary tumors in vitro [229].

3.4.7 RAS-BRAF

Activating *RAS* mutations are rarely reported and only in highly invasive pituitary tumors [5–7]. The most common *BRAF* mutation V600E was very rarely described in nonfunctioning pituitary tumor [230, 231]. A recent study in an Asiatic cohort reported the *BRAF V600E* mutation in 16% of corticotroph tumors, but this was not replicated in a multicenter study that found the mutation in only one out of 94 corticotroph tumors [225, 226].

3.4.8 TP53

TP53 mutations were very rarely described in pituitary tumors and only exclusively in invasive pituitary tumors and carcinomas [232–235]. This extreme rarity of *TP53* mutations in pituitary tumors was challenged by recent next-generation sequencing studies that revealed pathogenic *TP53* mutations in ~30% of *USP8*-wild type corticotroph tumors, suggesting that they may be more frequent in certain tumor subcategories than previously thought [225, 236].

Mutations in *CDKN1B* (p27), *CDKN2A* (p16/INK4a), and *CDKN2C* (p18/INK4c) are rare in sporadic pituitary tumors [149–151, 163, 225, 237–241].

3.4.9 Other

The antiproliferative response to glucocorticoids was linked to the tumor suppressor CABLES1 (CDK5 and ABL1 enzyme substrate 1) [136]. Corticotroph tumors have low CABLES1 expression, and this may account for the decreased p27 protein levels observed in these tumors. Mutations in the *CABLES1* gene were described in four female patients (2 young adults and 2 children) with large corticotroph tumors [242].

3.4.10 Noncoding RNAs

Noncoding RNAs do not encode proteins and are classified into microRNAs (miR-NAs) and larger long noncoding RNAs (lncRNAs) (reviewed in [243]).

MicroRNAs (miRNAs) are 22 nucleotides long RNAs that are expressed at defined genomic loci and regulate gene expression posttranscriptionally by cleaving mRNAs or repressing their translation (reviewed in [244]). Pituitary tumors show aberrant expression of miRNAs (reviewed in [245]). The miRNA let-7 (that targets *HMGA2*) is downregulated in lactotroph, corticotroph, and gonadotroph, but not in somatotroph tumors, and correlates with high tumor grade [246]. Other "tumor suppressor" miRNAs such as miR-15a and 16-1 (that target cyclin D) are downregulated in lactotroph, somatotroph tumors, and their expression is negatively correlated with tumor size [247]. Corticotroph tumors also show downregulation in miR-15a and 16-1 as well as let-7a, miR-21, miR-141, miR-143, miR-145, and miR-150; no correlations were found with tumor size or remission after surgery, with the exception of miR-141 that correlated with higher remission rate [248]. Somatotroph tumors show downregulation of miRNAs (miR-34b, miR-326, miR-432, miR-548c-3p, miR-570, and miR-603) that target HMGA1/2 and E2F1 [124]. Nonfunctioning tumors have downregulated miR-149-3p, miR-130a-3p, and miR-370-3p compared to hormone-secreting pituitary tumors [249]. In contrast, miR-107 (that targets AIP) is upregulated in somatotroph and nonfunctioning pituitary tumors [250]. Somatotroph tumors also show upregulated miR-26b and miR-128 that regulate PTEN and subsequently the AKT survival pathway [251]. Nonfunctioning tumors show upregulation of miR-128a, miR-155, and miR-516a-3p (that target WEE1) and miR-135a, miR-140-5p, miR-582-3p, miR-582-5p, and miR-938 (that target *Smad3*) with expression levels correlating with tumor size [252, 253]. Corticotroph tumors have upregulated miR-122, miR-493 (target E2F1), and miR-26a (targets protein kinase C\delta) [254, 255].

In addition to miRNAs, large numbers of long (~200 nucleotide) noncoding RNAs (lncRNAs) are transcribed that regulate genome organization and mRNA stability and translation (reviewed in [256]). Pituitary tumors show aberrant expression of lncRNA that act in tumor-suppressing or growth-promoting manner (reviewed in [257]). The maternally expressed gene 3 (MEG3) lncRNA that

functions via p53 is lost in gonadotroph tumors and negatively correlates with invasion [258]. The lncRNA H19 is downregulated in lactotroph and other pituitary tumors and correlates negatively with tumor volume probably; H19 may suppress tumor growth via the mTOR-4E-BP1 pathway [259]. Lactotroph tumors also showed decreased expression of CLRN1-AS1, which regulates the WNT/b-catenin signaling cascade [260]. In contrast, Hox transcript antisense intergenic RNA (HOTAIR) was upregulated in nonfunctioning pituitary tumors and this correlated with invasive behavior [261]. Interestingly, a lncRNA, MIR205HG, was shown to regulate growth hormone and prolactin by regulating the transcriptional activity of Pit1 [262].

Circulating miRNAs were proposed as potential biomarkers in pituitary tumors [263]. Indeed, late postoperative plasma miR-143–3p levels were decreased in patients with gonadotroph tumors suggesting its usefulness as a biomarker of successful surgery [264].

3.5 Conclusion

Pituitary tumorigenesis is complex and cannot be attributed to a single factor. Mutations in oncogenes and tumor suppressor genes frequent in cancer are rarely found in pituitary tumors. Few driver mutations were identified mainly in the *GNAS* and *USP8* genes mutated in somatotroph and corticotroph tumors respectively. Additional mutational hotspots were found in the *USP48* in a smaller fraction of corticotroph tumors and in the *SF3B1* in a fraction of lactotroph tumors. Posttranscriptional/–translational alterations in cell cycle and growth factor signaling undoubtedly play an important role, but the causative genetic events remain in many cases obscure. The use of pangenomic techniques has shifted our attention to previously unconsidered processes and highlighted the fact that when it comes to pituitary tumors one size does not fit all and that different histological subtypes of pituitary tumors are distinct tumor entities each with their own pathogenetic mechanisms.

References

- Alexander JM, et al. Clinically nonfunctioning pituitary tumors are monoclonal in origin. J Clin Invest. 1990;86(1):336–40.
- 2. Herman V, et al. Clonal origin of pituitary adenomas. J Clin Endocrinol Metab. 1990;71(6):1427–33.
- Schulte HM, et al. Clonal composition of pituitary adenomas in patients with Cushing's disease: determination by X-chromosome inactivation analysis. J Clin Endocrinol Metab. 1991;73(6):1302–8.
- 4. Gicquel C, et al. Monoclonality of corticotroph macroadenomas in Cushing's disease. J Clin Endocrinol Metab. 1992;75(2):472–5.
- 5. Karga HJ, et al. Ras mutations in human pituitary tumors. J Clin Endocrinol Metab. 1992;74(4):914–9.

- Cai WY, et al. Ras mutations in human prolactinomas and pituitary carcinomas. J Clin Endocrinol Metab. 1994;78(1):89–93.
- 7. Pei L, et al. H-ras mutations in human pituitary carcinoma metastases. J Clin Endocrinol Metab. 1994;78(4):842–6.
- Lim CT, Korbonits M. Update on the clinicopathology of pituitary adenomas. Endocr Pract. 2018;24(5):473–88.
- Caimari F, Korbonits M. Novel genetic causes of pituitary adenomas. Clin Cancer Res. 2016;22(20):5030–42.
- Stefaneanu L, et al. Adenohypophysial changes in mice transgenic for human growth hormone-releasing factor: a histological, immunocytochemical, and electron microscopic investigation. Endocrinology. 1989;125(5):2710–8.
- Kelly MA, et al. Pituitary lactotroph hyperplasia and chronic hyperprolactinemia in dopamine D2 receptor-deficient mice. Neuron. 1997;19(1):103–13.
- Asa SL, et al. Pituitary lactotroph adenomas develop after prolonged lactotroph hyperplasia in dopamine D2 receptor-deficient mice. Endocrinology. 1999;140(11):5348–55.
- Sano T, Asa SL, Kovacs K. Growth hormone-releasing hormone-producing tumors: clinical, biochemical, and morphological manifestations. Endocr Rev. 1988;9(3):357–73.
- 14. Dieterich KD, et al. Mutation and expression analysis of corticotropin-releasing factor 1 receptor in adrenocorticotropin-secreting pituitary adenomas. J Clin Endocrinol Metab. 1998;83(9):3327–31.
- Sakai Y, et al. Desmopressin stimulation test for diagnosis of ACTH-dependent Cushing's syndrome. Endocr J. 1997;44(5):687–95.
- Dahia PL, et al. Vasopressin receptor expression and mutation analysis in corticotropinsecreting tumors. J Clin Endocrinol Metab. 1996;81(5):1768–71.
- 17. Wang FF, et al. Plasma corticotrophin response to desmopressin in patients with Cushing's disease correlates with the expression of vasopressin receptor 2, but not with that of vasopressin receptor 1 or 3, in their pituitary tumours. Clin Endocrinol. 2012;76(2):253–63.
- Alexander JM, Klibanski A. Gonadotropin-releasing hormone receptor mRNA expression by human pituitary tumors in vitro. J Clin Invest. 1994;93(6):2332–9.
- Wood DF, Johnston JM, Johnston DG. Dopamine, the dopamine D2 receptor and pituitary tumours. Clin Endocrinol. 1991;35(6):455–66.
- Renner U, et al. Heterogeneous dopamine D2 receptor subtype messenger ribonucleic acid expression in clinically nonfunctioning pituitary adenomas. J Clin Endocrinol Metab. 1998;83(4):1368–75.
- Fougner SL, et al. Adenoma granulation pattern correlates with clinical variables and effect of somatostatin analogue treatment in a large series of patients with acromegaly. Clin Endocrinol. 2012;76(1):96–102.
- Luque RM, et al. Truncated somatostatin receptor variant sst5TMD4 confers aggressive features (proliferation, invasion and reduced octreotide response) to somatotropinomas. Cancer Lett. 2015;359(2):299–306.
- Lee EJ, et al. Absence of constitutively activating mutations in the GHRH receptor in GH-producing pituitary tumors. J Clin Endocrinol Metab. 2001;86(8):3989–95.
- Salvatori R, et al. Absence of mutations in the growth hormone (GH)-releasing hormone receptor gene in GH-secreting pituitary adenomas. Clin Endocrinol. 2001;54(3):301–7.
- 25. Dong Q, et al. Screening of candidate oncogenes in human thyrotroph tumors: absence of activating mutations of the G alpha q, G alpha 11, G alpha s, or thyrotropin-releasing hormone receptor genes. J Clin Endocrinol Metab. 1996;81(3):1134–40.
- Chanson P, et al. Absence of activating mutations in the GnRH receptor gene in human pituitary gonadotroph adenomas. Eur J Endocrinol. 1998;139(2):157–60.
- 27. Luque RM, et al. A cellular and molecular basis for the selective desmopressin-induced ACTH release in Cushing disease patients: key role of AVPR1b receptor and potential therapeutic implications. J Clin Endocrinol Metab. 2013;98(10):4160–9.

- 28. de Keyzer Y, et al. Overexpression of vasopressin (V3) and corticotrophin-releasing hormone receptor genes in corticotroph tumours. Clin Endocrinol. 1998;49(4):475–82.
- 29. Friedman E, et al. Normal structural dopamine type 2 receptor gene in prolactin-secreting and other pituitary tumors. J Clin Endocrinol Metab. 1994;78(3):568–74.
- Greenman Y, Melmed S. Expression of three somatostatin receptor subtypes in pituitary adenomas: evidence for preferential SSTR5 expression in the mammosomatotroph lineage. J Clin Endocrinol Metab. 1994;79(3):724–9.
- Miller GM, et al. Somatostatin receptor subtype gene expression in pituitary adenomas. J Clin Endocrinol Metab. 1995;80(4):1386–92.
- 32. Petersenn S, et al. Absence of somatostatin receptor type 2 A mutations and gip oncogene in pituitary somatotroph adenomas. Clin Endocrinol. 2000;52(1):35–42.
- Corbetta S, et al. Somatostatin receptor subtype 2 and 5 in human GH-secreting pituitary adenomas: analysis of gene sequence and mRNA expression. Eur J Clin Investig. 2001;31(3):208–14.
- 34. Ballare E, et al. Mutation of somatostatin receptor type 5 in an acromegalic patient resistant to somatostatin analog treatment. J Clin Endocrinol Metab. 2001;86(8):3809–14.
- Scheithauer BW, et al. Pituitary gland in hypothyroidism. Histologic and immunocytologic study. Arch Pathol Lab Med. 1985;109(6):499–504.
- Du J, et al. Pituitary adenoma secondary to primary hypothyroidism: two case reports. Medicine (Baltimore). 2020;99(8):e19222.
- Wingrave SJ, Kay CR, Vessey MP. Oral contraceptives and pituitary adenomas. Br Med J. 1980;280(6215):685–6.
- Scheithauer BW, et al. Effects of estrogen on the human pituitary: a clinicopathologic study. Mayo Clin Proc. 1989;64(9):1077–84.
- 39. Nota NM, et al. The occurrence of benign brain tumours in transgender individuals during cross-sex hormone treatment. Brain. 2018;141(7):2047–54.
- 40. Antonini SR, et al. Glucocorticoid receptor gene polymorphisms in ACTH-secreting pituitary tumours. Clin Endocrinol. 2002;57(5):657–62.
- Dahia PL, et al. Expression of glucocorticoid receptor gene isoforms in corticotropinsecreting tumors. J Clin Endocrinol Metab. 1997;82(4):1088–93.
- 42. Karl M, et al. Cushing's disease preceded by generalized glucocorticoid resistance: clinical consequences of a novel, dominant-negative glucocorticoid receptor mutation. Proc Assoc Am Physicians. 1996;108(4):296–307.
- McCabe CJ, et al. Increased MEN1 mRNA expression in sporadic pituitary tumours. Clin Endocrinol. 1999;50(6):727–33.
- 44. Alexandraki KI, Grossman AB. Novel insights in the diagnosis of Cushing's syndrome. Neuroendocrinology. 2010;92(Suppl 1):35–43.
- 45. Korbonits M, et al. Expression of 11 beta-hydroxysteroid dehydrogenase isoenzymes in the human pituitary: induction of the type 2 enzyme in corticotropinomas and other pituitary tumors. J Clin Endocrinol Metab. 2001;86(6):2728–33.
- 46. Drouin J, Bilodeau S, Vallette S. Of old and new diseases: genetics of pituitary ACTH excess (Cushing) and deficiency. Clin Genet. 2007;72(3):175–82.
- Bilodeau S, et al. Role of Brg1 and HDAC2 in GR trans-repression of the pituitary POMC gene and misexpression in Cushing disease. Genes Dev. 2006;20(20):2871–86.
- Zhang D, Du L, Heaney AP. Testicular Receptor-4: novel regulator of glucocorticoid resistance. J Clin Endocrinol Metab. 2016;101(8):3123–33.
- 49. Riebold M, et al. A C-terminal HSP90 inhibitor restores glucocorticoid sensitivity and relieves a mouse allograft model of Cushing disease. Nat Med. 2015;21(3):276–80.
- Renner U, et al. Autocrine and paracrine roles of polypeptide growth factors, cytokines and vasogenic substances in normal and tumorous pituitary function and growth: a review. Eur J Endocrinol. 1996;135(5):515–32.
- Johnson LK, et al. Nuclear accumulation of epidermal growth factor in cultured rat pituitary cells. Nature. 1980;287(5780):340–3.

- Chaidarun SS, et al. Role of growth factors and estrogen as modulators of growth, differentiation, and expression of gonadotropin subunit genes in primary cultured sheep pituitary cells. Endocrinology. 1994;134(2):935–44.
- LeRiche VK, Asa SL, Ezzat S. Epidermal growth factor and its receptor (EGF-R) in human pituitary adenomas: EGF-R correlates with tumor aggressiveness. J Clin Endocrinol Metab. 1996;81(2):656–62.
- Jaffrain-Rea ML, et al. Epidermal growth factor binding sites in human pituitary macroadenomas. J Endocrinol. 1998;158(3):425–33.
- 55. Kontogeorgos G, et al. Localization of epidermal growth factor (EGF) and epidermal growth factor receptor (EGFr) in human pituitary adenomas and nontumorous pituitaries: an Immunocytochemical study. Endocr Pathol. 1996;7(1):63–70.
- Theodoropoulou M, et al. Expression of epidermal growth factor receptor in neoplastic pituitary cells: evidence for a role in corticotropinoma cells. J Endocrinol. 2004;183(2):385–94.
- Theodoropoulou M, et al. Decoding the genetic basis of Cushing's disease: USP8 in the spotlight. Eur J Endocrinol. 2015;173(4):M73–83.
- Honda J, et al. Identification of epidermal growth factor mRNA-expressing cells in the mouse anterior pituitary. Neuroendocrinology. 2000;71(3):155–62.
- 59. Fukuoka H, et al. EGFR as a therapeutic target for human, canine, and mouse ACTH-secreting pituitary adenomas. J Clin Invest. 2011;121(12):4712–21.
- Vlotides G, et al. Rat prolactinoma cell growth regulation by epidermal growth factor receptor ligands. Cancer Res. 2008;68(15):6377–86.
- Cooper O, et al. EGFR/ErbB2-targeting lapatinib therapy for aggressive prolactinomas. J Clin Endocrinol Metab. 2021;106(2):e917–25.
- Sarkar DK, Kim KH, Minami S. Transforming growth factor-beta 1 messenger RNA and protein expression in the pituitary gland: its action on prolactin secretion and lactotropic growth. Mol Endocrinol. 1992;6(11):1825–33.
- 63. Recouvreux MV, et al. The pituitary TGFbeta1 system as a novel target for the treatment of resistant prolactinomas. J Endocrinol. 2016;228(3):R73–83.
- 64. Faraoni EY, et al. Sex differences in the development of prolactinoma in mice overexpressing hCGbeta: role of TGFbeta1. J Endocrinol. 2017;232(3):535–46.
- 65. Kelberman D, et al. Genetic regulation of pituitary gland development in human and mouse. Endocr Rev. 2009;30(7):790–829.
- 66. Paez-Pereda M, et al. Involvement of bone morphogenetic protein 4 (BMP-4) in pituitary prolactinoma pathogenesis through a Smad/estrogen receptor crosstalk. Proc Natl Acad Sci U S A. 2003;100(3):1034–9.
- 67. Giacomini D, et al. Bone morphogenetic protein-4 inhibits corticotroph tumor cells: involvement in the retinoic acid inhibitory action. Endocrinology. 2006;147(1):247–56.
- 68. Arzt E, et al. Pathophysiological role of the cytokine network in the anterior pituitary gland. Front Neuroendocrinol. 1999;20(1):71–95.
- Bernton EW, et al. Release of multiple hormones by a direct action of interleukin-1 on pituitary cells. Science. 1987;238(4826):519–21.
- 70. Renner U, et al. Involvement of interleukin-1 and interleukin-1 receptor antagonist in rat pituitary cell growth regulation. Endocrinology. 1995;136(8):3186–93.
- Karanth S, McCann SM. Anterior pituitary hormone control by interleukin 2. Proc Natl Acad Sci U S A. 1991;88(7):2961–5.
- Karanth S, Marubayashi U, McCann SM. Influence of dopamine on the altered release of prolactin, luteinizing hormone, and follicle-stimulating hormone induced by interleukin-2 in vitro. Neuroendocrinology. 1992;56(6):871–80.
- Arzt E, et al. Interleukin-2 and interleukin-2 receptor expression in human corticotrophic adenoma and murine pituitary cell cultures. J Clin Invest. 1992;90(5):1944–51.
- Elston MS, et al. Wnt pathway inhibitors are strongly down-regulated in pituitary tumors. Endocrinology. 2008;149(3):1235–42.

- Renner U, et al. Intrapituitary expression and regulation of the gp130 cytokine interleukin-6 and its implication in pituitary physiology and pathophysiology. Ann N Y Acad Sci. 2009;1153:89–97.
- Wang Z, Ren SG, Melmed S. Hypothalamic and pituitary leukemia inhibitory factor gene expression in vivo: a novel endotoxin-inducible neuro-endocrine interface. Endocrinology. 1996;137(7):2947–53.
- Ray DW, Ren SG, Melmed S. Leukemia inhibitory factor (LIF) stimulates proopiomelanocortin (POMC) expression in a corticotroph cell line. Role of STAT pathway. J Clin Invest. 1996;97(8):1852–9.
- Bousquet C, Ray DW, Melmed S. A common pro-opiomelanocortin-binding element mediates leukemia inhibitory factor and corticotropin-releasing hormone transcriptional synergy. J Biol Chem. 1997;272(16):10551–7.
- Yano H, et al. Pituitary-directed leukemia inhibitory factor transgene causes Cushing's syndrome: neuro-immune-endocrine modulation of pituitary development. Mol Endocrinol. 1998;12(11):1708–20.
- Auernhammer CJ, Melmed S. Leukemia-inhibitory factor-neuroimmune modulator of endocrine function. Endocr Rev. 2000;21(3):313–45.
- Chesnokova V, Auernhammer CJ, Melmed S. Murine leukemia inhibitory factor gene disruption attenuates the hypothalamo-pituitary-adrenal axis stress response. Endocrinology. 1998;139(5):2209–16.
- Chesnokova V, Melmed S. Leukemia inhibitory factor mediates the hypothalamic pituitary adrenal axis response to inflammation. Endocrinology. 2000;141(11):4032–40.
- Jones TH, et al. Production of bioactive and immunoreactive interleukin-6 (IL-6) and expression of IL-6 messenger ribonucleic acid by human pituitary adenomas. J Clin Endocrinol Metab. 1994;78(1):180–7.
- Jones TH, et al. Interleukin-6 secreting human pituitary adenomas in vitro. J Clin Endocrinol Metab. 1991;73(1):207–9.
- Rezai AR, et al. Interleukin-6 and interleukin-6 receptor gene expression in pituitary tumors. J Neuro-Oncol. 1994;19(2):131–5.
- 86. Hanisch A, et al. Expression of members of the interleukin-6 family of cytokines and their receptors in human pituitary and pituitary adenomas. J Clin Endocrinol Metab. 2000;85(11):4411–4.
- 87. Borg SA, et al. Expression of interleukin-6 and its effects on growth of HP75 human pituitary tumor cells. J Clin Endocrinol Metab. 2003;88(10):4938–44.
- Castro CP, et al. Reduced expression of the cytokine transducer gp130 inhibits hormone secretion, cell growth, and tumor development of pituitary lactosomatotrophic GH3 cells. Endocrinology. 2003;144(2):693–700.
- Arzt E, et al. Interleukin involvement in anterior pituitary cell growth regulation: effects of IL-2 and IL-6. Endocrinology. 1993;132(1):459–67.
- Marques P, et al. Pituitary tumour fibroblast-derived cytokines influence tumour aggressiveness. Endocr Relat Cancer. 2019;26(12):853–65.
- 91. Sapochnik M, et al. Autocrine IL-6 mediates pituitary tumor senescence. Oncotarget. 2017;8(3):4690–702.
- 92. Treier M, et al. Hedgehog signaling is required for pituitary gland development. Development. 2001;128(3):377–86.
- Vila G, et al. Sonic hedgehog regulates CRH signal transduction in the adult pituitary. FASEB J. 2005;19(2):281–3.
- 94. Vila G, et al. Expression and function of sonic hedgehog pathway components in pituitary adenomas: evidence for a direct role in hormone secretion and cell proliferation. J Clin Endocrinol Metab. 2005;90(12):6687–94.
- 95. Pyczek J, et al. Hedgehog signaling activation induces stem cell proliferation and hormone release in the adult pituitary gland. Sci Rep. 2016;6:24928.
- 96. Clevers H, Nusse R. Wnt/beta-catenin signaling and disease. Cell. 2012;149(6):1192–205.

- 97. Treier M, et al. Multistep signaling requirements for pituitary organogenesis in vivo. Genes Dev. 1998;12(11):1691–704.
- 98. Miyakoshi T, et al. Expression of Wnt4 in human pituitary adenomas regulates activation of the beta-catenin-independent pathway. Endocr Pathol. 2008;19(4):261–73.
- Ren J, et al. Decreased expression of SFRP2 promotes development of the pituitary corticotroph adenoma by upregulating Wnt signaling. Int J Oncol. 2018;52(6):1934–46.
- Semba S, et al. Frequent nuclear accumulation of beta-catenin in pituitary adenoma. Cancer. 2001;91(1):42–8.
- 101. Formosa R, et al. Expression and clinical significance of Wnt players and survivin in pituitary tumours. Endocr Pathol. 2012;23(2):123–31.
- Zhao B, et al. Angiomotin is a novel hippo pathway component that inhibits YAP oncoprotein. Genes Dev. 2011;25(1):51–63.
- 103. St John MA, et al. Mice deficient of Lats1 develop soft-tissue sarcomas, ovarian tumours and pituitary dysfunction. Nat Genet. 1999;21(2):182–6.
- 104. Xekouki P, et al. Non-secreting pituitary tumours characterised by enhanced expression of YAP/TAZ. Endocr Relat Cancer. 2019;26(1):215–25.
- 105. Lodge EJ, et al. Expression analysis of the hippo Cascade indicates a role in pituitary stem cell development. Front Physiol. 2016;7:114.
- 106. Lodge EJ, et al. Homeostatic and tumourigenic activity of SOX2+ pituitary stem cells is controlled by the LATS/YAP/TAZ cascade. Elife. 2019:e43996.
- 107. Qian ZR, et al. Role of E-cadherin, alpha-, beta-, and gamma-catenins, and p120 (cell adhesion molecules) in prolactinoma behavior. Mod Pathol. 2002;15(12):1357–65.
- 108. Qian ZR, et al. Tumor-specific downregulation and methylation of the CDH13 (H-cadherin) and CDH1 (E-cadherin) genes correlate with aggressiveness of human pituitary adenomas. Mod Pathol. 2007;20(12):1269–77.
- Nishioka H, Haraoka J, Akada K. Fibrous bodies are associated with lower GH production and decreased expression of E-cadherin in GH-producing pituitary adenomas. Clin Endocrinol. 2003;59(6):768–72.
- 110. Yamada S, et al. A study of the correlation between morphological findings and biological activities in clinically nonfunctioning pituitary adenomas. Neurosurgery. 2007;61(3):580–4. discussion 584–5
- 111. Fougner SL, et al. The expression of E-cadherin in somatotroph pituitary adenomas is related to tumor size, invasiveness, and somatostatin analog response. J Clin Endocrinol Metab. 2010;95(5):2334–42.
- 112. Elston MS, et al. Nuclear accumulation of e-cadherin correlates with loss of cytoplasmic membrane staining and invasion in pituitary adenomas. J Clin Endocrinol Metab. 2009;94(4):1436–42.
- Zhou K, Jin H, Luo Y. Expression and significance of E-cadherin and beta-catenins in pituitary adenoma. Int J Surg Pathol. 2013;21(4):363–7.
- 114. Zhang Q, et al. Germline mutations in CDH23, encoding cadherin-related 23, are associated with both familial and sporadic pituitary adenomas. Am J Hum Genet. 2017;100(5):817–23.
- Malumbres M, Barbacid M. Cell cycle, CDKs and cancer: a changing paradigm. Nat Rev Cancer. 2009;9(3):153–66.
- Quereda V, Malumbres M. Cell cycle control of pituitary development and disease. J Mol Endocrinol. 2009;42(2):75–86.
- 117. Sherr CJ, McCormick F. The RB and p53 pathways in cancer. Cancer Cell. 2002;2(2):103–12.
- 118. Jacks T, et al. Effects of an Rb mutation in the mouse. Nature. 1992;359(6393):295–300.
- 119. Woloschak M, Roberts JL, Post KD. Loss of heterozygosity at the retinoblastoma locus in human pituitary tumors. Cancer. 1994;74(2):693–6.
- 120. Woloschak M, et al. Abundance and state of phosphorylation of the retinoblastoma gene product in human pituitary tumors. Int J Cancer. 1996;67(1):16–9.
- 121. Pei L, et al. Frequent loss of heterozygosity at the retinoblastoma susceptibility gene (RB) locus in aggressive pituitary tumors: evidence for a chromosome 13 tumor suppressor gene other than RB. Cancer Res. 1995;55(8):1613–6.

- 122. Simpson DJ, et al. Loss of pRb expression in pituitary adenomas is associated with methylation of the RB1 CpG island. Cancer Res. 2000;60(5):1211–6.
- 123. Massimi I, et al. The HMGA1 protoncogene frequently deregulated in cancer is a transcriptional target of E2F1. Mol Carcinog. 2013;52(7):526–34.
- 124. D'Angelo D, et al. Altered microRNA expression profile in human pituitary GH adenomas: down-regulation of miRNA targeting HMGA1, HMGA2, and E2F1. J Clin Endocrinol Metab. 2012;97(7):E1128–38.
- 125. Moons DS, et al. Pituitary hypoplasia and lactotroph dysfunction in mice deficient for cyclindependent kinase-4. Endocrinology. 2002;143(8):3001–8.
- 126. Gillam MP, et al. MEN1 tumorigenesis in the pituitary and pancreatic islet requires Cdk4 but not Cdk2. Oncogene. 2015;34(7):932–8.
- Jordan S, et al. Cyclin D and cyclin E expression in normal and adenomatous pituitary. Eur J Endocrinol. 2000;143(1):R1–6.
- 128. Gruppetta M, et al. Expression of cell cycle regulators and biomarkers of proliferation and regrowth in human pituitary adenomas. Pituitary. 2017;20(3):358–71.
- 129. Tani Y, et al. Upregulation of CDKN2A and suppression of cyclin D1 gene expressions in ACTH-secreting pituitary adenomas. Eur J Endocrinol. 2010;163(4):523–9.
- 130. Hibberts NA, et al. Analysis of cyclin D1 (CCND1) allelic imbalance and overexpression in sporadic human pituitary tumors. Clin Cancer Res. 1999;5(8):2133–9.
- 131. Simpson DJ, et al. Cyclin D1 (CCND1) genotype is associated with tumour grade in sporadic pituitary adenomas. Carcinogenesis. 2001;22(11):1801–7.
- 132. Cander S, et al. Effect of cyclin [corrected] D1 (CCND1) gene polymorphism on tumor formation and behavior in patients with prolactinoma. Gene. 2012;509(1):158–63.
- 133. Saeger W, Schreiber S, Ludecke DK. Cyclins D1 and D3 and topoisomerase II alpha in inactive pituitary adenomas. Endocr Pathol. 2001;12(1):39–47.
- 134. Lidhar K, et al. Low expression of the cell cycle inhibitor p27Kip1 in normal corticotroph cells, corticotroph tumors, and malignant pituitary tumors. J Clin Endocrinol Metab. 1999;84(10):3823–30.
- Roussel-Gervais A, et al. Cooperation between cyclin E and p27(Kip1) in pituitary tumorigenesis. Mol Endocrinol. 2010;24(9):1835–45.
- 136. Roussel-Gervais A, et al. The Cables1 gene in glucocorticoid regulation of pituitary corticotrope growth and cushing disease. J Clin Endocrinol Metab. 2016;101(2):513–22.
- 137. Liu NA, et al. Cyclin E-mediated human proopiomelanocortin regulation as a therapeutic target for cushing disease. J Clin Endocrinol Metab. 2015;100(7):2557–64.
- Nakabayashi H, Sunada I, Hara M. Immunohistochemical analyses of cell cycle-related proteins, apoptosis, and proliferation in pituitary adenomas. J Histochem Cytochem. 2001;49(9):1193–4.
- 139. Lamback EB, et al. Cyclin A in nonfunctioning pituitary adenomas. Endocrine. 2020;70(2):380-7.
- Korbonits M, et al. Expression of phosphorylated p27(Kip1) protein and Jun activation domainbinding protein 1 in human pituitary tumors. J Clin Endocrinol Metab. 2002;87(6):2635–43.
- 141. Bellodi C, et al. Loss of function of the tumor suppressor DKC1 perturbs p27 translation control and contributes to pituitary tumorigenesis. Cancer Res. 2010;70(14):6026–35.
- 142. Bamberger CM, et al. Reduced expression levels of the cell-cycle inhibitor p27Kip1 in human pituitary adenomas. Eur J Endocrinol. 1999;140(3):250–5.
- 143. Dong W, et al. P21(Waf1/Cip1) and p27(Kip1) are correlated with the development and invasion of prolactinoma. J Neuro-Oncol. 2018;136(3):485–94.
- 144. Zhao D, Tomono Y, Nose T. Expression of P27kip1 and Ki-67 in pituitary adenomas: an investigation of marker of adenoma invasiveness. Acta Neurochir. 1999;141(2):187–92.
- 145. Fero ML, et al. The murine gene p27Kip1 is haplo-insufficient for tumour suppression. Nature. 1998;396(6707):177–80.
- 146. Teixeira LT, et al. p27Kip1-deficient mice exhibit accelerated growth hormone-releasing hormone (GHRH)-induced somatotrope proliferation and adenoma formation. Oncogene. 2000;19(15):1875–84.

- 147. Nakayama K, et al. Mice lacking p27(Kip1) display increased body size, multiple organ hyperplasia, retinal dysplasia, and pituitary tumors. Cell. 1996;85(5):707–20.
- 148. Pellegata NS, et al. Germ-line mutations in p27Kip1 cause a multiple endocrine neoplasia syndrome in rats and humans. Proc Natl Acad Sci U S A. 2006;103(42):15558–63.
- Kirsch M, et al. Frequent loss of the CDKN2C (p18INK4c) gene product in pituitary adenomas. Genes Chromosomes Cancer. 2009;48(2):143–54.
- Hossain MG, et al. Expression of p18(INK4C) is down-regulated in human pituitary adenomas. Endocr Pathol. 2009;20(2):114–21.
- 151. Woloschak M, et al. Frequent loss of the P16INK4a gene product in human pituitary tumors. Cancer Res. 1996;56(11):2493–6.
- 152. Frost SJ, et al. Transfection of an inducible p16/CDKN2A construct mediates reversible growth inhibition and G1 arrest in the AtT20 pituitary tumor cell line. Mol Endocrinol. 1999;13(11):1801–10.
- 153. Simpson DJ, et al. Hypermethylation of the p16/CDKN2A/MTSI gene and loss of protein expression is associated with nonfunctional pituitary adenomas but not somatotrophinomas. Genes Chromosomes Cancer. 1999;24(4):328–36.
- 154. Ruebel KH, et al. Inactivation of the p16 gene in human pituitary nonfunctioning tumors by hypermethylation is more common in null cell adenomas. Endocr Pathol. 2001;12(3):281–9.
- 155. Machiavelli G, et al. Expression of p16(INK4A) gene in human pituitary tumours. Pituitary. 2008;11(1):71–5.
- Seemann N, et al. CDKN2A/p16 inactivation is related to pituitary adenoma type and size. J Pathol. 2001;193(4):491–7.
- 157. Yoshino A, et al. Promoter hypermethylation profile of cell cycle regulator genes in pituitary adenomas. J Neuro-Oncol. 2007;83(2):153–62.
- 158. Morris DG, et al. Differential gene expression in pituitary adenomas by oligonucleotide array analysis. Eur J Endocrinol. 2005;153(1):143–51.
- Franklin DS, et al. CDK inhibitors p18(INK4c) and p27(Kip1) mediate two separate pathways to collaboratively suppress pituitary tumorigenesis. Genes Dev. 1998;12(18):2899–911.
- 160. Garcia-Fernandez RA, et al. Combined loss of p21(waf1/cip1) and p27(kip1) enhances tumorigenesis in mice. Lab Investig. 2011;91(11):1634–42.
- 161. Manojlovic-Gacic E, et al. Oncogene-induced senescence in pituitary adenomas—an immunohistochemical study. Endocr Pathol. 2016;27(1):1–11.
- Neto AG, et al. Elevated expression of p21 (WAF1/Cip1) in hormonally active pituitary adenomas. Ann Diagn Pathol. 2005;9(1):6–10.
- 163. Ikeda H, Yoshimoto T, Shida N. Molecular analysis of p21 and p27 genes in human pituitary adenomas. Br J Cancer. 1997;76(9):1119–23.
- 164. Wu ZB, et al. MicroRNA expression profile of bromocriptine-resistant prolactinomas. Mol Cell Endocrinol. 2014;395(1–2):10–8.
- 165. Zou H, et al. Identification of a vertebrate sister-chromatid separation inhibitor involved in transformation and tumorigenesis. Science. 1999;285(5426):418–22.
- 166. Wang Z, Yu R, Melmed S. Mice lacking pituitary tumor transforming gene show testicular and splenic hypoplasia, thymic hyperplasia, thrombocytopenia, aberrant cell cycle progression, and premature centromere division. Mol Endocrinol. 2001;15(11):1870–9.
- 167. Chesnokova V, Melmed S. Pituitary senescence: the evolving role of Pttg. Mol Cell Endocrinol. 2010;326(1–2):55–9.
- Vlotides G, Eigler T, Melmed S. Pituitary tumor-transforming gene: physiology and implications for tumorigenesis. Endocr Rev. 2007;28(2):165–86.
- 169. Pei L, Melmed S. Isolation and characterization of a pituitary tumor-transforming gene (PTTG). Mol Endocrinol. 1997;11(4):433–41.
- 170. Chesnokova V, et al. p21(Cip1) restrains pituitary tumor growth. Proc Natl Acad Sci U S A. 2008;105(45):17498–503.
- 171. McCabe CJ, et al. Expression of pituitary tumour transforming gene (PTTG) and fibroblast growth factor-2 (FGF-2) in human pituitary adenomas: relationships to clinical tumour behaviour. Clin Endocrinol. 2003;58(2):141–50.

- 172. Saez C, et al. Hpttg is over-expressed in pituitary adenomas and other primary epithelial neoplasias. Oncogene. 1999;18(39):5473–6.
- 173. Zhang X, et al. Pituitary tumor transforming gene (PTTG) expression in pituitary adenomas. J Clin Endocrinol Metab. 1999;84(2):761–7.
- 174. Foltran RK, et al. Study of major genetic factors involved in pituitary tumorigenesis and their impact on clinical and biological characteristics of sporadic somatotropinomas and non-functioning pituitary adenomas. Braz J Med Biol Res. 2018;51(9):e7427.
- 175. Xiao JQ, et al. Correlations of pituitary tumor transforming gene expression with human pituitary adenomas: a meta-analysis. PLoS One. 2014;9(3):e90396.
- McCabe CJ, et al. Vascular endothelial growth factor, its receptor KDR/Flk-1, and pituitary tumor transforming gene in pituitary tumors. J Clin Endocrinol Metab. 2002;87(9):4238–44.
- 177. Hu N, et al. Heterozygous Rb-1 delta 20/+mice are predisposed to tumors of the pituitary gland with a nearly complete penetrance. Oncogene. 1994;9(4):1021–7.
- 178. Classon M, Harlow E. The retinoblastoma tumour suppressor in development and cancer. Nat Rev Cancer. 2002;2(12):910–7.
- 179. Chesnokova V, et al. Pituitary hypoplasia in Pttg-/- mice is protective for Rb+/- pituitary tumorigenesis. Mol Endocrinol. 2005;19(9):2371-9.
- 180. Heaney AP, et al. Early involvement of estrogen-induced pituitary tumor transforming gene and fibroblast growth factor expression in prolactinoma pathogenesis. Nat Med. 1999;5(11):1317–21.
- Pei L. Identification of c-myc as a down-stream target for pituitary tumor-transforming gene. J Biol Chem. 2001;276(11):8484–91.
- Fuertes M, et al. Protein stabilization by RSUME accounts for PTTG pituitary tumor abundance and oncogenicity. Endocr Relat Cancer. 2018;25(6):665–76.
- 183. Collado M, et al. Tumour biology: senescence in premalignant tumours. Nature. 2005;436(7051):642.
- 184. Serrano M, Blasco MA. Putting the stress on senescence. Curr Opin Cell Biol. 2001;13(6):748–53.
- 185. Herranz N, Gil J. Mechanisms and functions of cellular senescence. J Clin Invest. 2018;128(4):1238-46.
- Chesnokova V, et al. Senescence mediates pituitary hypoplasia and restrains pituitary tumor growth. Cancer Res. 2007;67(21):10564–72.
- Chesnokova V, et al. Growth hormone is a cellular senescence target in pituitary and nonpituitary cells. Proc Natl Acad Sci U S A. 2013;110(35):E3331–9.
- Arzt E, et al. Pituitary adenoma growth: a model for cellular senescence and cytokine action. Cell Cycle. 2009;8(5):677–8.
- 189. Carreno G, Guiho R, Martinez-Barbera JP. Cell senescence in neuropathology: a focus on neurodegeneration and tumours. Neuropathol Appl Neurobiol. 2021;47(3):359–78.
- Boggild MD, et al. Molecular genetic studies of sporadic pituitary tumors. J Clin Endocrinol Metab. 1994;78(2):387–92.
- Prezant TR, Levine J, Melmed S. Molecular characterization of the men1 tumor suppressor gene in sporadic pituitary tumors. J Clin Endocrinol Metab. 1998;83(4):1388–91.
- 192. Tanaka C, et al. Analysis of loss of heterozygosity on chromosome 11 and infrequent inactivation of the MEN1 gene in sporadic pituitary adenomas. J Clin Endocrinol Metab. 1998;83(8):2631–4.
- 193. Farrell WE, et al. Sequence analysis and transcript expression of the MEN1 gene in sporadic pituitary tumours. Br J Cancer. 1999;80(1–2):44–50.
- 194. Farrell WE, et al. Chromosome 9p deletions in invasive and noninvasive nonfunctional pituitary adenomas: the deleted region involves markers outside of the MTS1 and MTS2 genes. Cancer Res. 1997;57(13):2703–9.
- 195. Bates AS, et al. Allelic deletion in pituitary adenomas reflects aggressive biological activity and has potential value as a prognostic marker. J Clin Endocrinol Metab. 1997;82(3):818–24.

- 196. Simpson DJ, et al. Chromosome 13q deletion mapping in pituitary tumors: infrequent loss of the retinoblastoma susceptibility gene (RB1) locus despite loss of RB1 protein product in somatotrophinomas. Cancer Res. 1999;59(7):1562–6.
- 197. Harada K, et al. Cytogenetic alterations in pituitary adenomas detected by comparative genomic hybridization. Cancer Genet Cytogenet. 1999;112(1):38–41.
- 198. Szymas J, et al. Genomic instability in pituitary adenomas. Pituitary. 2002;5(4):211-9.
- 199. Pack SD, et al. Common genetic changes in hereditary and sporadic pituitary adenomas detected by comparative genomic hybridization. Genes Chromosomes Cancer. 2005;43(1):72–82.
- Bello MJ, et al. Chromosomal abnormalities in pituitary adenomas. Cancer Genet Cytogenet. 2001;124(1):76–9.
- 201. Bi WL, et al. Landscape of genomic alterations in pituitary adenomas. Clin Cancer Res. 2017;23(7):1841–51.
- 202. Song ZJ, et al. The genome-wide mutational landscape of pituitary adenomas. Cell Res. 2016;26(11):1255–9.
- 203. Salomon MP, et al. The epigenomic landscape of pituitary adenomas reveals specific alterations and differentiates among acromegaly, Cushing's disease and endocrine-inactive subtypes. Clin Cancer Res. 2018;24(17):4126–36.
- 204. Neou M, et al. Pangenomic classification of pituitary neuroendocrine tumors. Cancer Cell. 2020;37(1):123–34. e5
- Lasolle H, et al. Chromosomal instability in the prediction of pituitary neuroendocrine tumors prognosis. Acta Neuropathol Commun. 2020;8(1):190.
- 206. Tatsi C, Stratakis CA. The genetics of pituitary adenomas. J Clin Med. 2019;9:1.
- 207. Newey PJ, et al. Mutant prolactin receptor and familial hyperprolactinemia. N Engl J Med. 2013;369(21):2012–20.
- Ronchi CL, et al. Landscape of somatic mutations in sporadic GH-secreting pituitary adenomas. Eur J Endocrinol. 2016;174(3):363–72.
- Sapkota S, et al. Whole-exome sequencing study of thyrotropin-secreting pituitary adenomas. J Clin Endocrinol Metab. 2017;102(2):566–75.
- Valimaki N, et al. Genetic and epigenetic characterization of growth hormone-secreting pituitary tumors. Mol Cancer Res. 2019;17(12):2432–43.
- 211. Vallar L, Spada A, Giannattasio G. Altered Gs and adenylate cyclase activity in human GH-secreting pituitary adenomas. Nature. 1987;330(6148):566–8.
- 212. Landis CA, et al. GTPase inhibiting mutations activate the alpha chain of Gs and stimulate adenylyl cyclase in human pituitary tumours. Nature. 1989;340(6236):692–6.
- 213. Lyons J, et al. Two G protein oncogenes in human endocrine tumors. Science. 1990;249(4969):655–9.
- Lania A, Mantovani G, Spada A. G protein mutations in endocrine diseases. Eur J Endocrinol. 2001;145(5):543–59.
- 215. Weinstein LS, et al. Activating mutations of the stimulatory G protein in the McCune-Albright syndrome. N Engl J Med. 1991;325(24):1688–95.
- 216. Reincke M, et al. Mutations in the deubiquitinase gene USP8 cause Cushing's disease. Nat Genet. 2015;47(1):31–8.
- 217. Ma ZY, et al. Recurrent gain-of-function USP8 mutations in Cushing's disease. Cell Res. 2015;25(3):306–17.
- 218. Perez-Rivas LG, et al. The gene of the ubiquitin-specific protease 8 is frequently mutated in adenomas causing Cushing's disease. J Clin Endocrinol Metab. 2015;100(7):E997–1004.
- 219. Hayashi K, et al. The USP8 mutational status may predict drug susceptibility in corticotroph adenomas of Cushing's disease. Eur J Endocrinol. 2016;174(2):213–26.
- 220. Faucz FR, et al. Somatic USP8 gene mutations are a common cause of pediatric Cushing disease. J Clin Endocrinol Metab. 2017;102(8):2836–43.
- 221. Albani A, et al. The USP8 mutational status may predict long-term remission in patients with Cushing's disease. Clin Endocrinol. 2018;89:454–58.

- 222. Perez-Rivas LG, et al. Somatic USP8 mutations are frequent events in corticotroph tumor progression causing Nelson's tumor. Eur J Endocrinol. 2018;178(1):57–63.
- 223. Cohen M, et al. Germline USP8 mutation associated with pediatric Cushing disease and other clinical features: a new syndrome. J Clin Endocrinol Metab. 2019;104(10):4676–82.
- Castellnou S, et al. SST5 expression and USP8 mutation in functioning and silent corticotroph pituitary tumors. Endocr Connect. 2020;9:243–53.
- 225. Sbiera S, et al. Driver mutations in USP8 wild-type Cushing's disease. Neuro-Oncology. 2019;21(10):1273–83.
- 226. Chen J, et al. Identification of recurrent USP48 and BRAF mutations in Cushing's disease. Nat Commun. 2018;9(1):3171.
- 227. Tian QB, et al. A novel ubiquitin-specific protease, synUSP, is localized at the post-synaptic density and post-synaptic lipid raft. J Neurochem. 2003;87(3):665–75.
- 228. Li C, et al. Somatic SF3B1 hotspot mutation in prolactinomas. Nat Commun. 2020;11(1):2506.
- Vazquez-Borrego MC, et al. A somatostatin receptor Subtype-3 (SST3) peptide agonist shows antitumor effects in experimental models of nonfunctioning pituitary Tumors. Clin Cancer Res. 2020;26(4):957–69.
- Ewing I, et al. A mutation and expression analysis of the oncogene BRAF in pituitary adenomas. Clin Endocrinol. 2007;66(3):348–52.
- De Martino I, et al. B-RAF mutations are a rare event in pituitary adenomas. J Endocrinol Investig. 2007;30(1):RC1-3.
- 232. Herman V, et al. Molecular screening of pituitary adenomas for gene mutations and rearrangements. J Clin Endocrinol Metab. 1993;77(1):50–5.
- 233. Levy A, et al. p53 gene mutations in pituitary adenomas: rare events. Clin Endocrinol. 1994;41(6):809–14.
- 234. Tanizaki Y, et al. P53 gene mutations in pituitary carcinomas. Endocr Pathol. 2007;18(4):217–22.
- Kawashima ST, et al. P53 gene mutation in an atypical corticotroph adenoma with Cushing's disease. Clin Endocrinol. 2009;70(4):656–7.
- 236. Uzilov AV, et al. USP8 and TP53 drivers are associated with CNV in a Corticotroph adenoma cohort enriched for aggressive tumors. J Clin Endocrinol Metab. 2021;106(3):826–42.
- 237. Tanaka C, et al. Infrequent mutations of p27Kip1 gene and trisomy 12 in a subset of human pituitary adenomas. J Clin Endocrinol Metab. 1997;82(9):3141–7.
- 238. Dahia PL, et al. Mutation and expression analysis of the p27/kip1 gene in corticotrophinsecreting tumours. Oncogene. 1998;16(1):69–76.
- Takeuchi S, et al. Mutation and expression analysis of the cyclin-dependent kinase inhibitor gene p27/Kip1 in pituitary tumors. J Endocrinol. 1998;157(2):337–41.
- 240. Tichomirowa MA, et al. Cyclin-dependent kinase inhibitor 1B (CDKN1B) gene variants in AIP mutation-negative familial isolated pituitary adenoma kindreds. Endocr Relat Cancer. 2012;19(3):233–41.
- 241. Agarwal SK, et al. The MEN1 gene and pituitary tumours. Horm Res. 2009;71(Suppl 2):131–8.
- 242. Hernandez-Ramirez LC, et al. Loss-of-function mutations in the CABLES1 gene are a novel cause of Cushing's disease. Endocr Relat Cancer. 2017;24(8):379–92.
- 243. Nagano T, Fraser P. No-nonsense functions for long noncoding RNAs. Cell. 2011;145(2):178–81.
- 244. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell. 2004;116(2):281–97.
- 245. Li XH, et al. MicroRNAs in human pituitary adenomas. Int J Endocrinol. 2014;2014:435171.
- 246. Qian ZR, et al. Overexpression of HMGA2 relates to reduction of the let-7 and its relationship to clinicopathological features in pituitary adenomas. Mod Pathol. 2009;22(3):431–41.
- Bottoni A, et al. miR-15a and miR-16-1 down-regulation in pituitary adenomas. J Cell Physiol. 2005;204(1):280–5.
- Amaral FC, et al. MicroRNAs differentially expressed in ACTH-secreting pituitary tumors. J Clin Endocrinol Metab. 2009;94(1):320–3.

- Vicchio TM, et al. MicroRNAs expression in pituitary tumors: differences related to functional status, pathological features, and clinical behavior. J Endocrinol Investig. 2020;43(7):947–58.
- 250. Trivellin G, et al. MicroRNA miR-107 is overexpressed in pituitary adenomas and inhibits the expression of aryl hydrocarbon receptor-interacting protein in vitro. Am J Physiol Endocrinol Metab. 2012;303(6):E708–19.
- 251. Palumbo T, et al. Functional screen analysis reveals miR-26b and miR-128 as central regulators of pituitary somatomammotrophic tumor growth through activation of the PTEN-AKT pathway. Oncogene. 2013;32(13):1651–9.
- 252. Butz H, et al. Down-regulation of Wee1 kinase by a specific subset of microRNA in human sporadic pituitary adenomas. J Clin Endocrinol Metab. 2010;95(10):E181–91.
- 253. Butz H, et al. MicroRNA profile indicates downregulation of the TGFbeta pathway in sporadic non-functioning pituitary adenomas. Pituitary. 2011;14(2):112–24.
- Stilling G, et al. MicroRNA expression in ACTH-producing pituitary tumors: up-regulation of microRNA-122 and -493 in pituitary carcinomas. Endocrine. 2010;38(1):67–75.
- 255. Gentilin E, et al. miR-26a plays an important role in cell cycle regulation in ACTHsecreting pituitary adenomas by modulating protein kinase Cdelta. Endocrinology. 2013;154(5):1690–700.
- 256. Yao RW, Wang Y, Chen LL. Cellular functions of long noncoding RNAs. Nat Cell Biol. 2019;21(5):542–51.
- 257. Du Q, et al. Research progress on lncRNA functions and mechanisms in pituitary adenomas. Horm Metab Res. 2020;52(5):280–8.
- 258. Zhang X, et al. Maternally expressed gene 3 (MEG3) noncoding ribonucleic acid: isoform structure, expression, and functions. Endocrinology. 2010;151(3):939–47.
- Wu ZR, et al. Inhibition of mTORC1 by lncRNA H19 via disrupting 4E-BP1/raptor interaction in pituitary tumours. Nat Commun. 2018;9(1):4624.
- 260. Wang C, et al. FOXP1-induced lncRNA CLRN1-AS1 acts as a tumor suppressor in pituitary prolactinoma by repressing the autophagy via inactivating Wnt/beta-catenin signaling pathway. Cell Death Dis. 2019;10(7):499.
- 261. Li Z, et al. Expression of the long non-coding RNAs MEG3, HOTAIR, and MALAT-1 in non-functioning pituitary adenomas and their relationship to tumor behavior. Pituitary. 2015;18(1):42–7.
- 262. Du Q, et al. MIR205HG is a long noncoding RNA that regulates growth hormone and prolactin production in the anterior pituitary. Dev Cell. 2019;49(4):618–31. e5
- 263. Di Ieva A, et al. MicroRNAs as biomarkers in pituitary tumors. Neurosurgery. 2014;75(2):181–9. discussion 188-9
- 264. Nemeth K, et al. Comprehensive analysis of circulating microRNAs in plasma of patients with pituitary adenomas. J Clin Endocrinol Metab. 2019;jc.2018-02479.



Genetics of Pituitary Adenomas

Anna Bogusławska, Aleksandra Gilis-Januszewska, and Márta Korbonits

Abbreviations

3 Pa	Pituitary adenomas and paragangliomas or phaeochromocytomas		
ACTH	Adrenocorticotrophic hormone		
AHR	Aryl hydrocarbon receptor		
AIP	Aryl hydrocarbon receptor-interacting protein		
BRAF	Proto-oncogene B-Raf		
cAMP	Cyclic adenosine monophosphate		
CDH23	Cadherin-related 23		
CGH	Comparative genomic hybridisation		
EGFR	Epidermal growth factor receptor		
FIPA	Familial isolated pituitary adenomas		
GH	Growth hormone		
GHRH	Growth hormone-releasing hormone		
HSP90	Heat shock protein 90		
IGF-1	Insulin-like growth factor 1		

The chapter has been endorsed by **Prof. Antonio Pico**, antonio.pico@umh.es, Departamento de Endocrinología, Hospital General Universitario de Alicante, Alicante, Spain

A. Bogusławska · A. Gilis-Januszewska

Department of Endocrinology, Endocrine Oncology and Nuclear Medicine, Jagiellonian University Medical College, Cracow, Poland e-mail: aleksandra.gilis-januszewska@uj.edu.pl

M. Korbonits (🖂)

Centre for Endocrinology, William Harvey Research Institute, Barts and the London School of Medicine and Dentistry, Queen Mary University of London, London, UK e-mail: m.korbonits@qmul.ac.uk

[©] Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_4

MEN1	Multiple endocrine neoplasia type 1		
MEN2A	Multiple endocrine neoplasia type 2A		
MEN2B	Multiple endocrine neoplasia type 2B		
MEN4	Multiple endocrine neoplasia type 4		
MGMT	O6-methylguanine-DNA methyltransferase		
microRNA	Micro ribonucleic acid		
NF1	Neurofibromatosis type 1		
NFPA	Non-functioning pituitary adenoma		
PA	Pituitary adenoma		
PI3K	Phosphoinositide 3-kinases		
PIT-1	Pituitary-specific POU-class homeodomain transcription factor		
PitNET	Pituitary neuroendocrine tumour		
PKA	Protein kinase A		
POMC	Proopiomelanocortin		
PTEN	Phosphatase and tensin homolog		
SDH	Succinate dehydrogenase		
USP48	Ubiquitin-specific peptidase 48		
USP8	Ubiquitin-specific peptidase 8		

4.1 Introduction

Pituitary adenomas (PAs) or pituitary neuroendocrine tumours (PitNETs) are mostly benign tumours located in sella turcica, deriving from the anterior lobe of the pituitary [1, 2]. They are relatively common in the general population in autopsy and magnetic resonance imaging (MRI) studies, but in 0.1% of cases they are clinically relevant [3–8]. In the last few years, using modern scientific and molecular methods, several novel genetic mutations have been identified that can predispose to pituitary adenomas. Moreover, genetic testing is getting more easily available for patients in routine clinical practice worldwide.

Hereditary pituitary adenoma can manifest solely as an isolated tumour, without any other clinical manifestations, like familial isolated pituitary adenoma (FIPA). Pituitary adenoma can be also a part of a syndromic disorder such as Multiple endocrine neoplasia type 1 (MEN1), Multiple endocrine neoplasia type 4 (MEN4), Carney complex, McCune-Albright syndrome, pheochromocytoma-pituitary adenoma (3Pa) or DICER1. In some cases, pituitary adenomas in a familial setting are characterised by an earlier onset of the disease, more aggressive course and resistance to medical treatment [9].

In this chapter, we focus on clinical aspects of PAs with germline mutations. Genetic testing in family members could result in earlier recognition of the disease leading to more favourable outcomes. Careful selection of cases is key to keep patient anxiety and medical care costs under control [10]. Additionally, genetic results may influence family planning and reproductive choices, favouring the suggestion that genetic counselling should be widely available for PA patients [11].

4.2 Epidemiology

Pituitary adenomas are the third most common neoplasms of central nervous system after meningiomas and gliomas, representing about 15% of cases [12, 13]. Most of PAs develop sporadically, but in 5% they may occur in a familial setting. The incidence of pituitary adenomas rises with age, with a peak of diagnosis between 30 and 60 years old [14]. Many of them constitute incidental findings during imaging scans performed due to other reasons [15]. Although during childhood and adolescence pituitary tumours are very rare, the genetic background should be considered in almost every case as it is responsible in a higher proportion, especially among somatotrophinomas [16] where up to 46–49% of gigantism cases have identifiable genetic cause [16, 17]. In adults, however, the genetic cause is present only in a minority of patients. The most common pituitary adenomas in early childhood are corticotroph adenomas [18], while in adolescence, prolactinomas and somatotroph adenomas are most frequent [19, 20]. In adulthood, clinically non-functioning adenomas (NFPAs) has taken over the most frequently encountered pituitary adenomas from prolactinomas in terms of frequency, probably due to better and more frequently used imaging of the population [13, 21]. Non-functioning pituitary adenomas and prolactinomas are followed by somatotroph adenomas, corticotroph adenomas and thyrotroph adenomas [22].

4.3 Pathogenesis

Pituitary adenomas develop from the hormone-secreting anterior pituitary cell types, such as somatotroph, lactotroph, corticotroph, gonadotroph and rarely from thyrotroph cells or their precursors. Several different factors that lead to pituitary tumorigenesis have been described but in most cases, molecular pathogenesis and epigenetic mechanisms are still unknown [23–28]. The impact of environmental factors is also questionable. Some data suggest that in highly polluted areas the course of somatotrophinomas is more aggressive; further, independent studies are required to confirm these data [29–31].

PAs are predominantly benign tumours with clinical symptoms caused by hormonal hypersecretion or mass effects leading to visual field disturbances or hypopituitarism. In some cases, PAs can be locally aggressive with invasion of the surrounding sinuses and putting pressure on the cranial nerves. Progression of pituitary tumours to true malignancy with distant metastases is very rare, found in about 0.2% of all pituitary adenomas, but aggressive tumours occur more frequently, reported to account from 2.5% to 10% in surgical series [32] with often similar pathological characteristics [33]. Recently a change has been suggested to address adenohypophyseal tumours as pituitary neuroendocrine tumours (PitNETs). This suggestion has been met with some controversy. The International Pituitary Pathology Club suggests that the hormone-producing cells of the pituitary are a part of the neuroendocrine system and propose to replace the word 'adenoma' by 'neuroendocrine tumour' to highlight the similarity with other neuroendocrine neoplasms [34]. However, others note the risk of aligning pituitary adenohypophyseal tumours to other neuroendocrine tumours would mean unnecessary anxiety in patients and physicians less familiar with the disease [35–40]. Further discussion is welcome on this issue.

The 2017 WHO classification emphasises the diagnostic role of immunohistochemistry for adenohypohyseal hormones and their cell lineage-specific transcription factors [33, 34, 41]. Pangenomic classification of pituitary tumours supports the key role of transcription factors and showed association between POU1F1/PIT-1 lineage and DNA hypomethylation, the transcription of differentiation markers, transposable elements and chromosomal instability [42].

Sporadic PAs seem to be monoclonal in origin [43]. X-chromosome inactivation studies have shown that they arise from a single somatic pituitary cell, although recurrent tumours may show an evolved and slightly different clone from the original tumour [44]. Monoclonal character is supported by observed long-term hormonal remission after successful tumour resection without changes in the surrounding tissue [45–48]. In patients with germline mutations multiple separate adenomas may arise from the pituitary tissue, for example, in aryl hydrocarbon receptor-interacting protein (*AIP*) mutation-positive patients [49]. Pituitary hyperplasia can be characteristic in patients with germline mutations, such as Carney complex, McCune-Albrigth syndrome, X-inked acrogigantism (XLAG), and rarely has been seen in *AIP* cases as well [50]. Genetic alterations associated with PitNETs can be germline, mosaic or somatic (Fig. 4.1).

4.3.1 Somatic Mutation

In general, sporadic pituitary adenomas have lower somatic mutations rate than other malignant tumours. *GNAS* gene mutation is the first reported and it is the most frequent genetic cause of sporadic growth hormone (GH)-producing tumours. It can be found in up to 40% of cases. This modification of the alpha subunit of the G-protein leads to loss of the GTPase activity while retained adenylyl cyclase stimulation resulting in sustained cyclic adenosine monophosphate (cAMP) production and protein kinase A (PKA) activation. This results in an uncontrolled hormone production and cell proliferation [51]. Another common somatic mutation causing sporadic adreno-corticotrophic hormone (ACTH)-producing adenomas was found in gene encoding ubiquitin specific peptidase 8 (USP8) [52, 53]. It results in loss of deubiquitination of epidermal growth factor receptor (EGFR) and consequently increased EGFR recycling to the cell surface, which leads to proopiomelanocortin (POMC) gene transcription. The frequency of *USP8* mutation in corticotrophinomas ranges between 36% and 60% [52, 54, 55]. Several studies, including the recent comprehensive assessment, support the role of somatic *GNAS/USP8* mutation status in predicting treatment



Fig. 4.1 Genetic origin of pituitary adenomas (adapted from Caimari et al. 2016 [64]) Data on PIK3CA and complex 1 genes needs to be confirmed in further publications

response [42]. In 30% of all pituitary adenomas single nucleotide or gene mutations in the catalytic subunit of PI3K have been presented. The clinical behaviour of tumours harbouring this mutation might be more aggressive [56]. However, in Cowden Syndrome with germline mutation of PI3K-PTEN-AKT, the increased prevalence of pituitary adenomas has not been observed [57]. Recent studies reported novel somatic variants [42, 58–60] which could provide new insight into pituitary tumorigenesis. Somatic hotspot mutation in splicing factor 3 subunit B1 (*SF3B1*) has been found in 20% of prolactinomas [61], in a recent study, although not reported in [42]. This recurrent variant results in aberrant splicing of ESRRG, a member of the oestrogen receptor-related receptor family, leading to increase PIT-1 activation and abnormal prolactin secretion and lactotroph proliferation. Other reported novel somatic mutations are *NR3C1*, *BRAF*, *USP48 PABPC1*, and *TP53* mutations found in corticotroph pituitary tumour, *GRB10* mutation identified in somatotroph adenomas or *KIF5A* mutation associated with lactotroph adenomas [58–60, 62, 63].

4.3.2 Germline Mutations

Germline mutations are defined as any changes in germ cells that are inherited from the parents or arisen in the fertilised egg, therefore every single body cell is affected. Their prevalence is evaluated to be around 5% of all PAs [64]. Germline mutations

are responsible for syndromic diseases such as MEN1, MEN4, Carney complex, *SDH*-related familial PAs, DICER1 syndrome, NF1, or due to apparently isolated pituitary adenomas due to *AIP* mutation or *GPR101* duplication.

4.3.3 Mosaic Mutations

Mosaicism means that the patient's body consists of at least two different genotypes; one without any gene mutations and other which is a result of genetic changes. If the germ cells (i.e. sperm or ova) harbour an alternative clone, then germline mosaicism can occur with the possibility of affected children.

In pituitary adenomas, somatic mosaicism occurs only in about 1% of patients [64]; *GNAS* mutation in McCune–Albright syndrome, *GPR101* duplication in XLAG and recently *MEN1* mosaicism has also been described [65]. The phenotype in these cases is dependent on the cell type and the number of affected tissues.

Germline mosaicism is relatively rarely diagnosed among endocrinological diseases, but has been described, for example, in MEN1 syndrome [66] and Carney complex [67].

4.3.4 Epigenetic Mechanisms

In a significant proportion of cases, genetic background of pituitary tumorigenesis is still not fully understood. Recent studies found association between epigenetic mechanisms and clinicopathological features of pituitary tumours. Epigenetic alterations may occur at chromatin level such as DNA methylation and histone modification or via non-coding RNAs, microRNAs or proteomics, and we refer to a recent comprehensive review on this topic [68]. Moreover, the tumour microenvironment, including fibroblasts and immune cells, may also influence pituitary tumorigenesis [69–72].

4.4 Clinical Presentation

4.4.1 Local Symptoms

The growing pituitary tumour causes pressure on surrounding tissue and its effects depend on the size of the lesion. Generally, this 'mass effect' is frequently associated with macroadenomas (>10 mm).

Headaches – occur in up to 37.5–72% of cases [73, 74], but their clinical presentation is heterogeneous. Headaches can be chronic, and often the sign which leads to investigations and diagnosis. Stretching of the meninges is one of the proposed mechanisms of headaches, but majority of studies found no association between tumour size and the presence of cephalgia [75, 76]. Pituitary tumours may invade cavernous sinuses where the trigeminal nerves are located. In this case, headache might be characterised by a constant, uni- or bilateral frontal or midface pain [76]. Indirect mechanisms are also suggested, probably due to dysfunction of the pituitary axis. Patients with somatotroph adenomas and prolactinomas are more prone to headaches than other tumour types [77]. Release of cytokines or neuroendocrine hormones may play a role in the initiation of headaches [73, 74, 78]. Headaches could also be acute severe headaches (with or without associated previous chronic headache). Acute, severe pain accompanied by photophobia or stiff neck may represent pituitary apoplexy, which in some cases could require emergency surgery. *AIP* mutation-positive patients are especially susceptible to pituitary apoplexy, even with a childhood onset [79].

Visual field disturbances – Pituitary adenomas are the most common cause of the chiasmal syndrome. Clinical symptoms are present more frequently when the optic pathway is displaced more than 3 mm from its original position. Based on a recent study, true bitemporal hemianopsia occurs much rarely than other visual field disturbances; the most frequent is bilateral temporal visual field defect, either pure or mixed (43% of patients). If the compression of chiasma is asymmetric, it is likely to manifest with asymmetric visual field defect [80, 81].

Compression on healthy pituitary tissue – leads to a progressive hormonal pituitary insufficiency. Usually first appearing hormone deficiency involves GH axis, followed by gonadotroph, thyrotroph and corticotroph axis. Thus the first visible symptoms are those of hypogonadism, due to gonadal insufficiency or hyperprolactinemia caused by 'stalk effect' [82]. Special attention must be paid by physicians to the possibility of pituitary-adrenal axis failure which can be a life-threatening condition.

4.4.2 Hormonal Excess Symptoms

Hormone-secreting tumours and those with a genetic background arise more often in childhood and adolescence. Depending on the age of onset, symptomatology may be different.

4.4.2.1 GH Excess

Growth hormone (GH) is a peptide which stimulates the IGF-1 production in all the peripheral tissues, while circulating IGF-1 is mainly arising from the liver. When the overproduction of GH starts before epiphyseal closure, increased growth velocity resulting in tall stature can be the most prominent manifestation. The development of abnormal tall stature could be further aggravated by hypogonadism and therefore delayed epiphysis closure. The hypogonadism can result from (a) damaged LH/FSH secretion due to the tumour's mass effect, (b) prolactin secretion from the tumour itself inhibiting gonadotrophin secretion via kisspeptin or (c) the tumour pressing on the talk resulting in reduced dopamine input from the hypothalamus and increased prolactin levels. In adulthood, soft tissue swelling influences patient's physical appearance and leads to changes in facial features, hand, feet, nose, prognathism, macroglossia, sleep apnoea, skin tags and teeth separation. Common metabolic complications of GH-axis overactivation are insulin resistance, hyperglycaemia, hypertension and increased triglyceride level. Other features observed in patients with

acromegaly are arthropathy, carpal tunnel syndrome, organomegaly, thyroid goitre, colorectal polyps, sweating and many others [16, 17, 83–94]. Apart from somatotropinomas, cause of GH excess can be GHRH-secreting pancreas or bronchial neuroendocrine tumours, altered GH regulation in NF1-associated optic pathway gliomas, pituitary hyperplasia without a tumour and sometimes normal-sized pituitary gland (for example, in Carney complex, McCune-Albright syndrome or XLAG) and the extremely rare ectopic GH secreting tumours. Additionally, deficiency of the X-link immunoglobulin superfamily member 1 (IGSF1), associated with central hypothyroidism, hypoprolactinaemia, GH deficiency in childhood and macroorchidism, may result in adulthood in somatotrope neurosecretory hyperfunction in humans [95]. Adult patients harbouring *IGSF1* loss of function mutation present acromegalic facial features as well as organ consequences following GH excess but not tall stature.

4.4.2.2 ACTH Excess

Under normal conditions, corticotrophin-releasing hormone (CRH), produced in the hypothalamus, increases corticotroph cell POMC transcription and ACTH synthesis, resulting in adrenal cortex cortisol synthesis. Autonomous ACTH secretion from a tumour results in excess cortisol levels and impaired or lost negative feedback. Long-term exposure to cortisol leads to characteristic clinical signs of Cushing's syndrome. ACTH excess presents differently in children and adolescents than in adults. Pathognomonic features are short stature with subnormal growth velocity and unexplained obesity [96]. In adults, central obesity with proximal myopathy and moon face are characteristic. In physical examination physicians should pay attention to skin lesions like facial plethora, violaceus striae (unusual in children <7-year-old), acanthosis nigricans due to insulin resistance and signs of hyperandrogenism, such as hirsutism or acne. Metabolic complications present as hypertension, diabetes mellitus or glucose intolerance and dyslipidaemia. Amenorrhea, menstrual dysfunction, hypogonadism, osteoporosis, susceptibility to infections and psychological compliants like depression, anxiety and irritability are also common [19, 97–103].

4.4.2.3 Hyperprolactinaemia

Elevated prolactin level among patients with pituitary adenoma might be due to two main reasons (if not due to a medication-induced hyperprolactinaemia): prolactinsecreting tumour or a presence of large lesion which can cause a 'stalk effect' [82]. Hyperprolactinaemia inhibits periodic gonadotropin-releasing hormone (GnRH) secretion via a pathway including kisspeptin. In children and adolescents, it can cause delayed puberty and growth arrest. Galactorrhoea may occur at any age in both genders, but most common in adult females. In females after menarche, the most common symptom of hyperprolactinaemia are menstrual irregularities. In males, decreased libido and erectile problems might be the first symptoms, and hypogonadism can result in gynaecomastia. Generally, symptoms are more easily recognised in women. Diagnosis of hyperprolactinaemia is challenging among elderly male or postmenopausal female patients as they are not aware of symptoms of hypogonadism [104]. Hyperprolactinaemia-induced long-term hypogonadism decreases bone density and can lead to osteoporosis. In some studies, patients with prolactinoma and concomitant hypogonadism have a higher risk of vertebral fractures [105, 106]. They also might develop metabolic syndrome since hyperprolactinemia promotes weight gain and has adverse effect on glucose metabolism and lipid profile [107, 108].

4.4.2.4 TSH Excess

Elevated thyroid-stimulating hormone (TSH) levels due to pituitary adenoma are rare. Elevated thyroxine levels in the context of inappropriate normal or increased TSH levels are the key with independence of the presence of symptoms and signs of thyrotoxicosis. Indeed, in 90% of cases biochemical hyperthyroidism is present, but clinical symptoms are observed only in 67% of patients. In biochemically silent cases only tumour immunohistochemistry analysis helps to classify the type of tumour [109]. In 25–42% of patients co-secretion of other pituitary hormones is noted, often GH and/or prolactin [110, 111].

4.4.2.5 LH/FSH Excess

About 80–90% of gonadotroph adenoma are silent without any biochemical manifestation and symptoms appear due to the mass effect of the tumour [112]. In the minority of cases clinical complaints may manifest as precocious puberty in children, menstrual irregularities, enlarged ovaries, ovary hyperstimulation in women and enlarged testicles in male patients [113].

4.4.3 Clinical Features Suggesting Genetic Background

In all patients harbouring a pituitary adenoma, thorough family history, 4 generation family tree and detailed physical examination should be performed (Table 4.1). It helps with differential diagnosis and guides genetic testing.

4.4.4 Pituitary Adenomas Associated with Hereditary Diseases

4.4.4.1 Familial Isolated Pituitary Adenomas (FIPA)

Familial isolated pituitary adenoma is an inherited condition, defined as a presence of two or more cases of PA among family members without any other associated tumours [115]. The genetic background of FIPA families is heterogeneous. While the majority have no identifiable mutation, around 20% of cases harbour an *AIP* mutation, with rare cases and kindreds with *GPR101* duplications [116, 117].

AIP

AIP, located at chromosomal region 11q13.2, acts as a tumour suppressor gene in the pituitary. Being a co-chaperone, the AIP protein interacts with numerous other binding partners, including the heat shock protein family, nuclear or growth factor receptors and viral proteins. As a consequence, it influences a wide spectrum of molecular pathways [118, 119]. Patients usually inherit a loss-of-function mutations (majority resulting in truncated proteins while missense variants have shortened half-life [120]) and the second copy is lost in the tumour tissue as seen with loss of

heterozygosity (LOH) studies [121, 122]. Up to now, all cases had germline mutation (most inherited with one published de novo case [83]) with over 100 different gene variants described. The phenotype presents as a spectrum between youngonset aggressive disease and mild cases not needing treatment [79, 83, 115, 123, 124].

Around half of *AIP* mutation-positive families are homogenous with the vast majority having somatotrophinomas or somatolactotrophinomas [28, 124–126]. Among the heterogeneous family the most common combination is the co-existence

Organ	Manifestation	Gene and disease
Skin	angiofibromas (85%), collagenomas (70%), lipomas (30%)	MEN1
	mucosal fibromas (95-98%)	MEN2B (95-98%)
	lentiginosis (70%) cutaneous myxomas (50%)	Carney complex
	cafe au-lait spots	McCune-Albright syndrome, NF1*
	skin neurofibromas, crow syndrome	NF1*
Thyroid	goiter, thyroid tumour	DICER1, McCune-Albright syndrome
	medullary carcinoma	MEN2A (90%) MEN2B (90%), MEN4, NF1*
	thyroid tumours (75% benign, 25% malignant)	Carneycomplex
Parathyroid	Parathyroidadenoma	MEN1 (95%), MEN2 (up to 30%), MEN4, Carney complex, CDC73**
Chest abnormalities	cardiacmyxomas	Carneycomplex (up to 40%)
	pleuropulmonaryblastoma	DICER1 (70%)
Pancreatic tumour	neuroendocrinetumour	MEN1 (30-70%), MEN4, VHL, McCune-Albright
Renal manifestations	cystic nephroma	DICER1
	renaltumour	SDHx
Adrenal gland	PPNAD (up to 60%)	Carneycomplex
manifestations	Adrenal Cushing's syndrome	McCune-Albright syndrome
	phaeochromocytoma/paraganglioma	SDHx, MAX, MEN2A (40-50%), MEN2B (50%),
	adrenocorticaltumours	MEN1 (40%), MEN4
Gonadal abnormalities	precociouspuberty	McCune-Albright syndrome
	gonadaltumours (mostly calcifying Sertoli cell tumours)	Carneycomplex (up to 41%)
	Sertoli-Leydig cell tumour	DICER1
Bone manifestations	fibrous dysplasia	McCune-Albright syndrome
Central nervous system	Schwannomas	Carneycomplex (10%), NF1*
Eye abnormalities	optic pathwayglioma, Lish nodules	NF1*
	visual loss due to facial fibrous dysplasia	McCune-Albright syndrome

 Table 4.1
 The most common extra pituitary manifestations of hereditary systemic disorders associated with abnormal pituitary hormones

of acromegaly with prolactin-producing tumour (37% of patients) [79, 115, 121, 124]. Usually patients are diagnosed with macroadenoma and age at disease onset is before 30 years. Penetrance is around 20-23% [79, 124]. Genetic screening of *AIP* mutation should start at not later than 4 years of age.

GPR101 Duplication

The *GPR101* gene microduplication on chromosome Xq26.3 was recently found as an infant cause of gigantism, named X-linked acrogigantism (XLAG) [17, 117]. It is a very rare condition and less than 40 patients have been described so far [127]. Mostly *GRP101* mutation occurs sporadically but a few familial cases have also been found. Germline mutation was described only among female patients whereas somatic mutation has been identified in four male patients. In three families, motherto-son transmission with full penetrance [128] has been observed representing familial form of the disease. XLAG is characterised by excessive GH production before the age of 5 years, some acromegalic features (facial coarsening, prominent mandible, enlarged extremities) and increased appetite. Pituitary tumours are mostly macroadenomas with mixed GH and prolactin secretion. In about 25% of cases pituitary hyperplasia is observed. Preimplantation genetic diagnostic and prenatal *GPR101* testing for a pregnant patient with XLAG should be offered due to 50% chance of transmitting the mutation.

Other

The phenotypes of *AIP* and *GPR101* mutation-negative families are different. They have more varied tumour types, and the age of onset is similar to sporadic pituitary adenomas in the 4-fifth decade. Homogenous families represent about half of the kindreds with somatotrophinomas and prolactinoma families being the most common, while in the heterogeneous families acromegaly-prolactinoma and prolactinoma-NFPA combinations are the most common [79, 124]. Other genes with germline variants have been suggested as causing pituitary adenoma (*CDH23* [129] and *CABLES1* [130]), but more data are needed to confirm these results.

4.4.4.2 Multiple Endocrine Neoplasia Type 1 (MEN1)

Multiple endocrine neoplasia type 1 is a rare genetic disorder with an autosomal dominant inheritance. In most cases it is caused by an inactivating mutation in the *MEN1* gene, located on the 11q13 chromosome. This gene plays an important role as a tumour suppressor, and it encodes the protein menin, which is responsible for regulation of transcription, cell apoptosis and epigenetic changes [131–133]. Most of the *MEN1* mutations appear in a familial setting, but in 10% of cases are de novo and a few mosaic mutations have been observed. Somatic *MEN1* mutations are common in sporadic parathyroid tumours (1–18%) and pancreatic-gastrointestinal tumours (16–38%), but not in pituitary adenomas [134, 135]. The disease has a high (up to 95%) overall penetrance, increasing with age. In 17% of cases, MEN1-associated tumours manifest before the age of 21. Overall the prevalence of MEN1 is estimated to be 1:30,000 [131]. Characteristic main tumours are parathyroid adenoma, pancreatic neuroendocrine tumour, and pituitary adenoma. Some cutaneous



Fig. 4.2 Clinical presentation of skin changes in an MEN1 patient: (**a**) angiofibromas (**b**) lentigines and collagenomas. Courtesy of Prof. Alicja Hubalewska-Dydejczyk, Department of Endocrinology, Medical College, Jagiellonian University, Cracow, Poland

tumours, e.g. angiofibromas, collagenomas and café au lait macules have been associated with MEN1 (Fig. 4.2). The diagnosis of MEN1 is based on clinical, familial and genetic criteria: (1) a presence of two or more characteristic MEN1 tumours, (2) occurrence of one MEN1-associated tumour and first-degree family member with a clinical manifestation of MEN1, (3) asymptomatic patient with a germline *MEN1* mutation, who has not yet developed clinical, radiological or biochemical abnormalities [135].

Pituitary adenomas occur in 30% (range 15-50%) of patients with MEN1 syndrome, predominantly in the fourth decade of life, but various ages of onset have been noted in the literature (from 5 years to 90 years) [135–137]. The most common subtype is prolactinoma (60%), followed by NFPA (15%), somatotrophinoma (10%), coricotrophinomas (5%) and thyrotrophinomas (1%) [135, 138, 139], although recent studies have shown increased numbers of NFPA among asymptomatic patients as a result of clinical screening [140]. Pituitary adenoma as a first MEN1 manifestation appears more frequently in childhood and adolescence (21%) than among adults (15-20% of cases) [137]. PAs in symptomatic MEN1 patients are noted to be invasive macroadenomas with a low response rate to medical treatment compared to sporadic cases. Interestingly, tumours found by screening were mostly stable non-functioning microadenomas [140]. Pituitary adenomas associated with MEN1 have more frequently plurihormonal profiles and are more often characterised by the presence of multiple pituitary tumours within the same gland than sporadic cases [131, 139, 141, 142]. Genetic screening test should search for sequence variation or large deletions [131, 143]. It is suggested to start genetic screening at the age of 5 years.

4.4.4.3 Multiple Endocrine Neoplasia Type 4 (MEN 4)

Multiple endocrine neoplasia type 4 (MEN4) is a systemic disease, first described in 2006 [144], caused by *CDKN1B* gene mutation, with a phenotype similar to MEN1 syndrome. The *CDKN1B* gene, located on chromosome 12q13, encodes p27, which regulates the cell cycle and acts as a tumour suppressor. Genetic testing should be performed in the absence of *MEN1* mutation. The penetrance has not been determined due to the small number of identified patients. To date, only 20 cases

and eight pituitary tumours among adults have been noted [145]. In rare MEN1-like patients without identifiable germline *MEN1* mutations, other *CDKIs* pathological variants (p15 [*CDKN2B*, 1%], p18 [*CDKN2C*, 0.5%], p21 [*CDKN1A*, 0.5%]) have been found [146]. Moreover, 3 cases with *CDC73* gene mutation [147–149] (responsible for hyperparathyroidism-jaw tumour syndrome) and 2 cases with *CASR* mutation [147, 149] (causing familial hypocalciuric hypercalcemia) with MEN1-like phenotype have been described.

4.4.4.4 Multiple Endocrine Neoplasia Type 2 (MEN2)

MEN2 is divided into two subtypes: MEN2A and MEN2B. It is caused by *RET* mutation located on chromosome 10q11.21. The prominent features are medullary thyroid carcinoma, phaeochromocytoma and hyperparathyroidism for MEN2A or medullary thyroid carcinoma, pheochromocytoma, marfanoid feature, mucosal fibromas for MEN2B. An activating mutation of the *RET* gene results in increased stimulation of several protein kinase pathways. Pituitary tumour in MEN2 syndrome occurs very rarely; to date, only four patients with *RET* mutation and PA have been described [150–153]. It is unclear if these represent coincidence or there is a casual relationship.

4.4.4.5 Carney Complex

Carney complex is a rare, heterogeneous, systemic disorder with an autosomal dominant inheritance. It was first described in 1985 by Professor Carney as a "complex of myxomas, spotty pigmentation, and endocrine overactivity" [154]. Around 70% of cases occur in a familial setting, and the genetic mutation is caused by a germline inactivating mutation mainly in the *PRKAR1A* gene, located on 17q22–24 locus [155, 156]. This gene encodes the regulatory subunit type 1 α of protein kinase A. To date over 140 *PRKAR1A* mutations have been described, referred to as Carney complex 1. Another affected locus is noted on chromosome 2p16 (Carney complex 2 locus), but the gene responsible for this mutation has not been found. Recently, other PKA gene subunit including *PRKACB* on chromosome 1 [157] and *PRKACA* on chromosome 19 [158, 159] (associated with adrenal hyperplasia) have been described in patients with Carney complex.

Genotype-phenotype correlation has been observed in Carney complex patients. If the mutation involves exon deletions, then acromegaly, myxomas, lentigines and psammomatous melanotic schwannoma (PMS) develop more frequently [160]. Patients with large deletion develop Carney complex features earlier [156] and more severe [67]. The study with the largest number of Carney complex patients showed female predominance (63%) [161]. To confirm the Carney complex diagnosis, the occurrence of two major criteria is required or one main criterion concomitant with a known inactivating *PRKAR1A* mutation or affected first-degree relative [162]. Symptoms may manifest at every age, but the peak of Carney complex diagnosis is 20 years old [163]. Skin lesions can occur at birth, then the most common tumours in childhood and adolescence are cardiac and cutaneous myxomas (Fig. 4.3). Pigmented nodular adrenocortical disease (PPNAD) is the most frequent endocrinological dysfunction among Carney complex patients and usually manifests in the



Fig. 4.3 (**a**–**c**) Clinical presentation of a patient with Carney complex with *PRKAR1A* mutation: (**a**) 20-year-old adult male patient with scarfs after cardiac myxoma surgery and bilateral adrenalectomy due to PPNAD; (**b**) lentigines on the vermilion of the lips; (**c**) pigmented lesions in the left eye and lentigines on the face. (**d**, **e**) Radiological features of Carney complex (**d**) Cardiac myxoma in the left atrium*; (**e**) Vestibular nerve schwannoma. Courtesy of Prof. Alicja Hubalewska-Dydejczyk, Department of Endocrinology, Medical College, Jagiellonian University, Cracow, Poland. *Courtesy of Prof. Agnieszka Olszanecka, Department of Cardiology, Medical College, Jagiellonian University, Cracow, Poland

second or third decade of life as ACTH-independent Cushing Syndrome. Pituitary adenomas develop in 12% of cases, but in 75% of patients asymptomatic elevation of GH and IGF-1 appears due to pituitary hyperplasia [161]. As far as pituitary manifestations are concerned, no predominance among sexes has been noted. PA

may manifest as a single lesion or multifocal tumours, usually in the third decade of life [163]. Somatotrophinoma or somatotroph and/or lactotroph hyperplasia are the most common pituitary manifestations; prolactinomas are rarely observed [164]. In recent studies, two patients with *PRKAR1A* mutation and corticotrophinoma have been noted [165, 166]. Additionally, an occurrence of pituitary apoplexy with presumed GH adenoma in a 14-year-old Carney complex patient has been described [167].

4.4.4.6 McCune-Albright Syndrome

McCune-Albright syndrome is characterised by the occurrence of three main classic features: polycystic fibrous dysplasia, precocious puberty and café au lait spots (Fig. 4.4). This rare disorder with prevalence between 1/100.000 and 1/1.000.000 [168] is caused by an activating mutation of the *GNAS* gene, located on chromosome 20q13. It encodes the guanine-nucleotide-binding protein α -subunit, and its mutation results in constant activation of adenylate cyclase. The most frequent *GNAS* mutations in McCune-Albright syndrome are R201H or R201C [169–171]. They appear first at an early postzygotic stage and leads to somatic mosaicism. The phenotype of patients depends on affected tissues [172–174]. Theoretically, if the *GNAS* mutation involves germline cells the disease could be inherited, but probably those cells would be non-viable, as to date no familial case has been described. The most common pituitary disease related to McCune-Albright syndrome is a somatotroph adenoma (in one-third of cases) or pituitary hyperplasia [175]. PCR sequencing of samples of affected tissue or peripheral blood lymphocytes should be performed [176].

4.4.4.7 The Three P Association (3 Pa)

SDHx

Association of pituitary adenomas with paraganglioma/pheochromocytoma was first described in 1952 by Iversen [177]. Relatively recently the phaeochromocytoma/paraganglioma gene has been linked to a predisposition to pituitary tumorigenesis. The co-existence of the above tumours, also known as 'the three P association (3 Pa)', is a rare condition, caused by germline mutation of the SDHx gene which consists of several subunits (SDHA, -B, -C, -D or SDHA2F) [178–180]. To date, <100 cases have been described worldwide. Based on the available data, a PA related to SDHx mutation could manifest with different phenotypes within the same family (somatotrophinoma, prolactinoma, or NFPA). The prevalence of PitNETs in SDHx is very low (<1%). They usually present as macroadenomas and require a multimodal approach. In a histopathological analysis, PA related to SDHx presents with an interesting phenotype with intracytoplasmic vacuoles which can correspond to the presence of autophagic bodies [179]. Pituitary adenomas can be the first or only manifestation of SDHx mutation [181]. Most recently, pituitary carcinoma has been noted in a patient with a medical history of paraganglioma and SDHB mutation [182]. In addition, the association of pituitary tumours and phaeochromocytoma/paraganglioma appears also in other hereditary syndromes caused by MEN1, RET, VHL and MAX mutations [179].


Fig. 4.4 (**a–e**) Clinical presentation of a patient with McCune-Albright syndrome with tall stature: (**a**) deformity of the thorax and acromegalic facial features; (**b**) extensive and asymmetric fibrous dysplasia in the right ribs in bone scintigraphy; (**c**) acromegalic features in McCune-Albright syndrome: tall stature (192 cm), hand enlargement; (**d**) skeletal deformity in chest radiography; (**e**) fibrous dysplasia in CT scan. (**f–h**) Clinical presentation of an adult patient with McCune-Albright syndrome with precocious puberty and GH excess: (**f**) cafe au lait spots on the face, (**g**) fibrous dysplasia involving skull base, (**h**) skeletal deformity, cafe au lait spots. Courtesy of Prof. Wojciech Zgliczyński and Dr. Maria Stelmachowska-Banaś, Department of Endocrinology, Bielański Hospital, Warsaw, Poland

MYC-Associated Factor (MAX)

MYC-associated factor X plays an important role as a tumour suppressor gene. This germline mutation was first found in a familial pheochromocytoma and paraganglioma with aggressive behaviour. The *MAX* gene, located on chromosome 14q23.3, interacts with other parts of MAX-MLX network, and together they are responsible for integration of cellular signals and modulate gene expression [183]. The combination of phaeochromocytoma/paraganglioma related to *MAX* mutation with pituitary adenoma is extremely rare—to date, only single cases with somatotrophinoma and prolactinoma have been described [184, 185]. Patients with germline *MAX* mutation may also develop other systemic manifestations like renal oncocytoma or lung cancer [186]. Most of the *MAX* mutations occur in specific helix-loop-helix-leucin-zipper domain (bHLHZ) and casein kinase II phosphorylation sites. Loss of function mutation or deletion of *MAX* is associated with tumorigenesis involving neuroendocrine cells and correlated with metastatic potential [186, 187]. Assessing gene copy number profiling may have an important role in determining clinical risk [188].

4.4.4.8 Neurofibromatosis 1 (NF1)

Neurofibromatosis type 1 is an autosomal dominant systemic disorder caused by inactivating mutation of neurofibromin, predisposing to benign and malignant tumours with a prevalence estimated around 1: 2500–1: 3500 live births. The most common characteristic features are cutaneous neurofibromas, cafe-au-lait skin lesions, intertriginous freckling [189, 190], Lisch nodules and brain tumours, including the most common optic pathway glioma (Fig. 4.5). Based on a study with 64 children with NF1 and optic glioma, 10% of patients with NF1 and optic glioma have clinical features of GH excess with elevated GH and IGF1 levels. They had no visible pituitary lesion [191]. The cause of GH excess in patients with optic gliomas could be loss of hypothalamic somatostatinergic inhibition or abnormal GHRH secretion from the tumour. As optic pathway gliomas do not stain positive for GHRH, the loss of somatostatin drive is a more likely explanation [192, 193]. Somatostatin analogues are an effective treatment. Recently a 68-year-old patient with somatotrophinoma, hyperparathyroidism, follicular thyroid carcinoma associated with NF1 mutation and no MEN1 mutation has been described [194]. The pituitary tumour tissue showed no loss of the wild-type allele of the NF1 gene, but harboured a somatic GNAS R201C mutation. These data suggest that the rare case reports of PitNETs and NF1 could be coincidence of these relatively common diseases (PitNET 1:1000, NF1 1:2500–3500 of the general population) in the same patient.

4.4.4.9 DICER1 Syndrome

DICER1 syndrome is caused by a gene located on chromosome 14q32.13, which plays an important role in microRNA processing and DNA damaging. Dysregulation in the *DICER1* gene, which encodes ribonuclease, leads to tumorigenesis. Inheritance is of an autosomal dominant type with a variable penetrance. Almost every tumour develops a second, somatic *DICER1* mutation



Fig. 4.5 Clinical presentation of a patient with neurofibromatosis type 1: (**a**, **b**) multiple neurofibromas and cafe au lait spots; (**c**) Crowe's sign; (**d**) Lisch nodules and right orbital tumour. Courtesy of Prof. Alicja Hubalewska-Dydejczyk, Department of Endocrinology, Medical College, Jagiellonian University, Cracow, Poland

in the RNAse IIIa or IIIb domain [195]. DICER1 syndrome presents a high variability of aggressive tumours, the most frequent being the very malignant pleuropulmonary blastoma (PPB), which occurs in over 50% of cases. Multiple endocrine manifestation involving the thyroid, ovaries and pituitary gland may also occur. In animal models, *DICER1* mutations lead to anterior pituitary dysmorphology and hypoplasia. Pituitary blastoma has been described in a few cases. These tumours manifest before the age of 2 years and in 70% of cases, Cushing's disease is observed, and the mortality rate is very high (40%) by the second year of life [196, 197].

Recently one case of a 50-year-old woman with confirmed *DICER1* mutation who developed a microprolactinoma has been reported [198]. It is controversial if *DICER1* mutation could lead to a prolactin-producing adenoma or if it is only a coincidence. Confirmation of *DICER1* mutation requires identification of heterozygous germline *DICER1* variant.

4.5 Patients with Pituitary Adenomas

Patient assessment should start with careful history and family history (Fig. 4.6). Family tree is drawn for three generations. A negative family history does not exclude germline mutation due to de novo mutations or low penetrance diseases. Detailed physical examination, biochemical and imaging assessment can follow (Table 4.2 presenting specific features by type of pituitary adenoma).

4.5.1 Somatotroph Adenoma

In 2017, the WHO defined somatotroph adenoma as a tumour that expresses mainly GH and develops from PIT-1 cell lineage. The most common subtype is a densely granulated variant, followed by sparsely granulated, mammosomatotroph, mixed somatotroph-lactotroph and acidophil stem cell adenoma [41]. Furthermore, pangenomic classification of pituitary tumours revealed gonadotroph markers (SF1) expression in a subset of somatotroph tumours (*GNAS* wild type) [42]. GH immunoreactive tumours without clinical features occur in about 2% of cases and are classified as silent somatotroph adenomas [199].

GH-producing adenoma is the most common cause of acromegaly and gigantism, with a prevalence of 70–240 cases per million, rising with age; the peak of diagnosis is estimated to be 45 years old [200–202]. In childhood and adolescence somatotroph adenoma constitutes only 2% of all types of pituitary adenomas, but, in up to 46–49% genetic mutations are responsible for its occurrence [16, 17]. Additionally, at a younger age its behaviour is more aggressive: over 70% are macroadenomas and 30% are invasive.

IGF-1 blood measurement is used as a screening test for acromegaly. Usually normal concentrations exclude acromegaly and gigantism with high probability, while its elevation constitutes a reason for further diagnostics. The gold standard for diagnosis is the oral glucose GH suppression test [203]. Recent study showed that the main determinants of GH nadir after glucose load are BMI, sex and oestrogen in oral contraceptives. Moreover, GH nadir concentration was significantly lower among healthy subjects than cut-off values used in current acromegaly guidelines [204]. During adolescence physiological insulin resistance should be taken into consideration when interpreting GH and IGF-1 results. Clinical features of GH excess are very typical, but a delay in diagnosis remains considerable, amounting on average 5 years (but also delays of 15 or even 25 years have been reported) [201]. Prolactin concentration can be elevated in 16-27% of patients, due to co-secretion by the tumour cells or compression of the pituitary stalk [205, 206]. First-line treatment is surgical resection of the pituitary tumour. In patients with persistent disease following operation, adjuvant therapy is indicated and includes first-generation somatostatin analogues (SSA), pasireotide, pegvisomant, dopamine agonists and even reoperation or radiation [203].

XLAG (age of onset < 5 years) – In infant-onset somatotrophinoma, an X-linked acrogigantism diagnosis should be considered. This recently described disease is





Table 4.2 Description of specific features, mean age of diagnosis and genetic alteration in syndromes by type of pituitary adenoma

ype of PA	Syndrome	Genetic alteration	Mean age of diagnosis	Specific features
Somatotroph	X-linked acrogigantism, FIPA Patients	GPR101 duplication	first years of life (<5 years)	female predominance, pitultary hyperplasia or tumour, males can be mosaic or familial
	AIP mutation, FIPA Patients	AIP mutation	2 nd decade of life (<30 years)	male predominance, reduced SSTR 2 expression
	McCuneAlbright syndrome	mosaic GNAS mutation	2 rd decade of life	male predominance, pituitary hyperplasia, prolactincosecretion
	Carney complex	PRKAR1A, PRKACB	3 rd decade of life	no gender predominance hyperplasia (majority) or tumour
	Sporadic somatotrophinomas (GNAS mutation)	somatic GNAS mutation	3 ^d decade of life	smaller size, good response to medical treatment with SSA
	3Pa	SDHx mutation	single cases	macroadenomas
	MEN1	MEN1 mutation	4 th decade of life	2 nd most common MEN-1 related pituitary tumour, female predominance
	MEN4	CDKN1B mutation	single cases	single cases
Corticotroph Idenoma	DICER1 syndrome	DICER1 gene mutation	<2-year-old	pituitary blastoma, slight female predominance
	FIPA Patients	AIP mutation	2 nd decade of life (<30 years)	male predominance, no AIP positive case
	Carney complex	PRKAR1A mutation or PRKACB duplication	single cases	single cases, differential diagnosis of ACTH-independent PPNAD
	Sporadic corticotrophinomas USP8, USP48, BRAF mutation	somatic USP8, USP48, BRAF mutation	3rd decade of life (10 years earlier)	female predominance, smaller size, greater SSTR5 expression
	MEN1	MEN1 mutation	4 th decade of life	female predominance, differential diagnosis of ACTH-independent CS
	Sporadic corticotrophinomas CABLES1	CABLES1 mutation	single cases	Simplex cases (4 female patients with macro adenomas)
	MEN2	RET mutation	single cases	connection with RET gene is questionable
	MEN4	CDKN1B	single cases	single cases
.actotroph idenoma	AIP mutation, FIPA Patients	AIP mutation	2 nd decade of life (<30 years)	male predominance
	MEN1	MEN1 mutation	4 th decade of life	Most common adenoma type, female predominance
	3 Pa	MAX mutation, SDHx mutation	various age	aggressive behaviour
Thy rotroph Idenoma	AIP mutation, FIPA Patients	AIP mutation	4 th decade of life	mixed tumours, mostly GH+ TSH
	MEN1	MEN1 mutation	4th decade of life	single cases
	thyroid receptor beta	THRB mutation	single cases	single cases
4FPA	AIP mutation, FIPA Patients	AIP mutation	2 nd decade of life (<30 years)	male predominance
	MEN1	MEN1 mutation	4 th decade of life	female predominance, microadenomas during clinical screening

caused by duplication of the *GRP101* gene located at X26.3 and leads to overexpression of an orphan G protein-coupled receptor (GPCR) [207, 208]. This mutation can be germline or sporadic [17]. In all patients the most prominent feature is an accelerated growth velocity before 4 years of age, with the peak age of diagnosis around the first year. The XLAG prevalence is from 4.4% to 10% in gigantism cases [208], with a female predominance. These GH pituitary tumours are significantly more often macroadenomas with suprasellar extension and cavernous sinus invasion. The literature describes pituitary hyperplasia in around 25% cases. Hyperprolatinaemia can occur in up to 85% patients [17]. Genetic testing should be performed using standard comparative genomic hybridisation (CGH) array, but in negative cases with a suggestive phenotype, alternative methods such as CNV ddPCR for GPR101 or HD-aCGH should be considered. On suspicion of a mosaic XLAG mutation, analysis of affected tissue should be performed.

AIP mutation (age of onset usually < 30 years) – AIP is a well-known gene involved mainly in early-onset acromegaly and gigantism. AIP mutation may occur in 40% of gigantism patients [50, 79, 125]. Typically, clinical manifestations are observed in the second decade of life. Usually patients harbour macroadenomas, and age of the disease onset is before 30 years [79, 125, 209]. Males are slightly more represented, which could be due to ascertainment bias as males have taller stature and later puberty than females. Characteristic for AIP-related PA is a more aggressive behaviour than sporadic PA, specifically a larger tumour size (90%), suprasellar extension (50%), local invasiveness, invariably sparsely granulated cytokeratin pattern and higher risk of childhood-onset pituitary apoplexy. These features are responsible for an increased recurrence rate. AIP cases are mostly resistant to the first generation of somatostatin analogues and possibly somewhat better response to pasireotide [210]. Somatostatine receptor (SSTR) type 2 expression is not reduced in our studies [211], which is also supported by an animal model data [212]. Double adenomas [49] or pituitary hyperplasia [50, 213] has also been described.

MEN1 syndrome – Somatotrophinoma related to *MEN1* mutation occurs in about 10% of cases. In the absence of GPR101 duplication and *AIP* mutation, genetic testing for MEN1 could be undertaken. MEN1-associated somatotroph adenomas are mostly macroadenomas with local invasiveness and a poorer medical treatment response [135–137]. In MEN1 cases we specifically need to be aware of GHRH-secreting pancreas lesions causing GH excess and in children gigantism.

Somatic GNAS mutation – One of the most common genetic causes for isolated somatotroph adenomas is a sporadic activating guanine nucleotide-binding protein (*GNAS*) mutation. It results in a constitutively activated cAMP pathway [214]. These tumours contribute to higher GH and IGF-1 levels, but they are characterised by a smaller size and good response to SSA treatment in comparison to patients with *AIP* and *GPR101* mutations [84]. Furthermore, *GNAS*-mutated pituitary tumours have shown higher expression of dopamine receptor 2 in comparison to *GNAS* wild-type PAs [42]. They arise in older age than *AIP* affected. No association was observed between *GNAS* mutation and granulation pattern in histopathology

results [215, 216]. In aggressive sporadic somatotroph tumours resistant to SSA, novel therapeutic approaches including possibly RET pathways have been investigated. The use of sorafenib has been suggested as a potential treatment option in these cases [217].

McCune-Albright syndrome – Acromegaly is present in up to 26% of cases with the mean age of onset in the second decade of life. GH excess in McCune-Albright syndrome is significantly more frequent in males (75%). Concomitant hyperprolactinemia is present in 71–92% of patients. Hyperplasia is the most common pituitary manifestation [175].

Carney complex – Acromegaly is the main pituitary manifestation of Carney complex, usually preceded by other characteristic features, and can occur in up to 12% of cases, mostly in the third decade of life. Asymptomatic elevation of GH and IGF-1 is observed in over 75% of cases [161]. Somatotroph adenomas related to Careny complex are often multifocal with no predominance among sexes.

MEN 4 syndrome – In the absence of *MEN1* mutation in patients presenting MEN1-like features with somatotrophinoma, genetic testing for *CDKN1B* could be considered (to date only four cases of somatotroph adenomas have been described).

SDHx – The prevalence of somatotroph adenoma in *SDHx* mutation is very rare. A careful family history of pheochromocytoma and paraganglioma should be performed.

NF1 – Acromegaly and gigantism have been observed in about 10% of children with neurofibromatosis type 1 and optic pathway glioma without visible pituitary lesion [191]. GH excess should be considered as a relative common endocrine manifestation and frequent auxological evaluation should be performed.

DNMT3A – Acromegaly has also been described in other rare genetic disorders such as Tatton-Brown Rahman syndrome, caused by *DNMT3A* mutation [218], but the link between this gene and somatotroph adenoma has not been found.

4.5.2 Corticotroph Adenoma

Corticotrophinoma is defined as a tumour that presents immunostaining for ACTH and/or shows expression of the transcription factor T-PIT.

ACTH-producing tumours are usually microadenomas and lead to chronic hypercortisolaemia [219]. In children and adolescents, corticotrophinoma is the most common cause of Cushing syndrome (75–80% in comparison to 49–71% in adults) [220, 221]. The overall prevalence of Cushing disease (CD) is estimated to be 0.7 to 2.4 per million per year [219]. 20% of corticotrophinomas appear silently without biochemical and clinical manifestations. They tend to be macroadenomas and do not present local invasion but have a greater propensity for apoplexy [222]. The first choice of treatment is resection of corticotrophinomas. The second-line approach must be individualised to each patient; available options are repeated neurosurgery, radiotherapy, medical therapy and bilateral adrenalectomy [223].

FIPA – No patients have been identified with Cushing's disease with unequivocal pathogenic *AIP* variant. Among non-AIP FIPA families only 2.9% of patients have Cushing disease [115].

MEN1 – Corticotrophinoma appears in 5–10% of all pituitary tumours related to *MEN1* mutation. Rarely ACTH- producing adenomas can manifest as a first symptom of MEN1 [224]. In MEN1, due to adrenal Cushing's syndrome, differential diagnosis of hypercortisolaemia should be performed. In clinical management, calcium should be measured due to high penetrance of hyperparathyroidism among MEN1 patients.

DICER1 syndrome - Pituitary blastoma was first described in 2008 in a 13-month-old girl with ACTH-dependent Cushing disease and diabetes insipidus. Blastomas are primitive, malignant neoplasms with the possibility to metastasise, but for pituitary blastomas true metastasis has not been observed to date [195, 196]. The relationship between pituitary blastoma and DICER1 gene mutation was first noted in 2011 in a child presenting a pituitary blastoma with a positive familial history of DICER1 mutation. The histopathological appearance is typically characterised by rosettes, Rathke-like epithelial cells and gland-like cells with large secretory epithelium. Pituitary blastomas are a very rare manifestation of DICER1 syndrome with a low penetrance (<1% of cases), but represents a pathognomonic symptom [196]. Clinically, severe Cushing's disease with ophthalmoplegia in an infant strongly suggests pituitary blastoma. The mean age of PitB diagnosis is 8 months, with a range from 7 to 24 months. Almost half of the affected children die within months after diagnosis. Other systemic manifestations of DICER1 syndrome are pleuropulmonary blastoma, cystic nephroma, pineoblastoma, ovarian Sertoli-Leydig cell tumours and multinodular goitre [197, 225-227].

Germline *USP8* **mutation.** Recently a patient with heterozygous ubiquitinspecific protease 8 (*USP8*) germline mutation (c.2155 T > C, p. S719P) [228] has been described. Clinical presentation of the 16-year-old female with recurrent CD included developmental delay, dysmorphic features, ichthyosiform hyperkeratosis, chronic lung and kidney disease and cardiomyopathy. Further studies are required to investigate potential specific effects of USP8 in other organs. The CD with additional clinical symptoms might be a new genetic syndrome.

Somatic USP8 mutation is often identified in sporadic corticotroph adenomas. Two research groups identified somatic *USP8* point mutation in a short segment of the *USP8* gene in corticotrophinomas [52, 229]. USP8 removes ubiquitin from internalised epidermal growth factor receptor (EGFR), therefore protecting it from degradation. As a result of this gain-of-function mutation, EGFR recycling to the cell surface and stimulation of POMC production is increased. Normally, 14-3-3 protein regulates USP8 activity via binding to certain domain of the USB8 protein resulting in less cleavage and reduced activity. Mutations in the 14-3-3 binding area lead to increased USB8 cleavage and activity. *USP8* mutations are specific to corticotrophinomas. Corticotrophinomas with *USP8* mutation are common (36–63%), more frequent in women, smaller, and patients usually manifest 10 years earlier than wild-type corticotrophinomas in which the mean age of the diagnosis is 34 years [229]. They seem to have a higher recurrence rate and urinary free cortisol

level at the diagnosis. The prevalence of paediatric ACTH-producing adenomas harbouring *USP8* mutation is 30%, but recurrence of the disease is lower [230]. Identification of *USP8* mutation might also have an influence on the choice of medical treatment. Affected cells show a higher expression of SSTR type 5 and O6-methylguanine-DNA methyltransferase (MGMT) level, which suggests pasireotide as a first choice therapy [42, 231]. Additionally, EGFR inhibitors have already been studied in 2011 as a novel potential treatment option for Cushing's disease, before identification of somatic *USP8* mutation in corticotrophinomas. Recently in wild-type *USP8* corticotrophinomas [62] genetic testing showed new mutations: *USP48* (21.6% of cases) and *BRAF* (18.8% of individuals) mutation. Similar to USP8, they stimulate cells to produce POMC and are unique for ACTH-secreting pituitary tumours [229].

The CABLES 1 gene, located on chromosome 18q11.2, encodes a protein that regulates cell cycle progression and acts as a tumour suppressor [232]. It is expressed in the nucleus, prevents cells regulators such as DKN1A (P21), CDK5R1 (P35) and TP63 [130] from degradation, and interacts with TP53 and TP73 at the initiation of apoptosis. In 2016 somatic CABLES1 mutation was found in a few patients (four from 146) with sporadic Cushing's disease, but not in familial cases. All cases were female with macroadenoma – three presented extrasellar extension, and in three cases a second surgery was required to achieve biochemical control [130, 232]. Moreover, histopathology testing showed high Ki67 index. Further studies are needed to assess the prevalence of CABLES1 mutation among corticotrophinomas and characterise in detail their function in pituitary adenomas.

Carney complex – While the vast majority of patients with Carney complex develop adrenal Cushing's syndrome, now two cases have been described with pituitary Cushing's disease [165, 166]. The diagnosis can be challenging.

MEN2 syndrome – Corticotroph adenomas associated with MEN2 syndrome have been found in four cases (three patients with MEN2A and one paediatric patient with MEN2B) [150–153]. Whether these cases represent phenocopies or there is indeed a relationship between the RET gene and pituitary adenoma is unclear.

MEN4 syndrome – Corticotroph adenomas related to *CDKN1B* coding for p27 protein has been found in only two adult cases [145]. Recently the common *CDKN1B* rs2066827 polymorphism has been considered to play a role in cortico-troph adenoma tumorigenesis [233], but these data need to be confirmed independently.

4.5.3 Lactotroph Adenoma

Lactotroph adenoma is a pituitary tumour, derived from PIT-1 lineage and secretes prolactin. Prolactin can be produced together with GH in mixed somatotroph-lactotroph or mammosomatotroph adenoma. Prolactinomas express positive immunostaining for the oestrogen receptor- α (ER α), which could be considered as a prognostic factor. Lower ER α expression is associated with higher tumour

proliferation and it is more frequently observed in male patients [234]. Historically, prolactinoma was reported the most common tumour type of pituitary adenoma but recently, due to better availability of brain imaging techniques, detection of nonfunctional pituitary tumours increased. NFPAs are currently considered as the most prevalent. In most cases, lactotroph adenomas manifest after the age of 12 years as a microadenomas with the prevalence 40:100000, and the peak of diagnosis is between 20 and 50 years old. Females are more frequently and earlier diagnosed than males, with a ratio 10:1 [235, 236]. This imbalance between sexes might be due to reported complaints like menstrual irregularities. In male patients, lactotroph adenomas demonstrate more aggressive behaviour like larger tumour size, higher prolactin level, rapid growth, invasion into surrounding tissue and resistance to treatment. This difference between sexes might be due to galanin expression, oestrogen, vasoactive intestinal peptide (VIP) receptors or recently described somatic $SF3B1^{R625H}$ mutation [61]. The first line of treatment for prolactinomas is dopamine agonists. Surgery should be considered in cases unresponsive to pharmacological treatment. Genetic screening for AIP and MEN1 mutation should be considered.

FIPA – Lactotroph adenomas and somatotroph-lactotroph adenomas are the second most common PAs related to *AIP* mutation among FIPA, after somatotrophinomas [79, 124, 237, 238]. Patients harbouring *AIP* mutation are younger, with male predominance, higher prolactin level, greater tumour size and extrasellar extension in comparison to prolactinoma cases without genetic background. Resistance to dopamine agonist therapy is often reported. Feature and multiple neurosurgeries are required to achieve disease control [125], but there are too little data to determine the treatment response of *AIP* positive prolactinomas [79, 115, 124]. Prospective diagnosed *AIP*-related lactotroph adenomas among FIPA were similar to sporadic cases. They were microprolactinomas with no suprasellar extension or local invasion [79, 124]. *AIP* negative families have less number of affected members (in over 80% only two pituitary cases in comparison to three or more in *AIP* related FIPA) and microadenomas as well as macroadenomas have been observed [239].

MEN1 syndrome – Prolactinomas are the most common clinically presenting pituitary tumours in MEN1 patients and represent 50–60% of all pituitary adenomas. Clinical screening of MEN1 families revealed a similar prevalence of NFPA, although many of these are small lesions with no need of intervention [140]. MEN1-related prolactinomas are usually macroadenomas in children, with screening suggested to be started at the age of 5 years. Genetic testing for *MEN1* and *AIP* mutation should be considered for young patients with prolactinoma at the same time.

3 Pa – Paragangliomas and phaeochromocytomas can be associated with pituitary adenomas in families with *SDH* or *MAX* mutations with a very low penetrance, or rarely with MEN1 syndrome [178, 179, 184, 185, 240]. Prolactinomas and somatotrophinomas are the most frequent pituitary tumour types, often presenting as macroadenomas. Whether pituitary imaging and biochemical testing should be part of the paraganglioma surveillance in *SDH* or *MAX* mutation-positive families is currently under investigation. **Somatic** *SF3B1*R625H **mutations** – Using whole genome sequencing, recently a novel hotspot somatic mutation of splicing factor 3 subunit B1 (*SF3B1*) has been identified in 20% of prolactinomas [61]. The mutation is unique for prolactinsecreting pituitary tumours among PAs. Gender preference in affected prolactinomas has been observed with a male predominance of 24% v. 10.7% in females. The *SF3B1* variant impacts on aberrant splicing of one of the oestrogen-related receptor family members, oestrogen related receptor gamma. This promotes prolactin hypersecretion via a stronger affinity for PIT-1 and therefore greater transcriptional activation of prolactin. Additionally, this mutation affects cell proliferation and decreases apoptosis of lactotroph cells. Patients harbouring *SF3B1^{R625H}* mutation presented higher level of prolactin and significantly poorer treatment response in comparison to the wild-type group, whereas age at disease onset, tumour size and tumour invasion were not significantly different [61].

Prolactin receptor (PRLR) mutation – Previously this gene mutation was described in familial hyperprolactinaemia [241, 242]. Moreover, in animal investigations, PRLR knock-out mice develop lactotroph adenoma [243]. Recent studies have suggested that PRLR gene may play a role in sporadic prolactinomas in humans as well [244]; however, in another study, in a group of patients with sporadic lactotrophinomas, PRLR mutation has not been found [245].

4.5.4 Thyrotrope Adenoma

Thyrotrophinomas are rare and represent only 0.5–3% of all pituitary adenomas [246]. The incidence is around 0.26 per million per year, but in recent years a progressive increase in the recognition of thyrotroph adenomas has been observed, probably due to better imaging methods and heightened awareness among physicians [247]. The mean age at diagnosis is 45 years; there is no difference in sex ratio. Occurrence among paediatric patients is limited only to single case reports. Immunohistochemistry shows strong expression of nuclear staining for the acidophilic lineage transcription factor PIT-1 and GATA-2. In up to 55% of cases they can co-secrete other hormones, mostly GH or prolactin. At the time of diagnosis, TSHomas are more often invasive macroadenomas [110, 246]. In the diagnostic process, thyroid hormone resistance, euthyroid hyperthyroxinaemia, or other drug therapy intake (i.a. amiodaron) should be considered. The genetic background of the majority of thyrotroph adenomas remains undiscovered. TSHoma related to *AIP* mutation has been described in one case [125], while a metastatic TSHoma has been described in a 19-year-old MEN1 patient [248].

Some studies suggest that the mutation of thyroid receptor beta (TR β), which is responsible for thyroid hormone resistance, may cause predisposition to pituitary tumorigenesis; two cases with this mutation have been described in a patient harbouring TSHoma.

The first-line treatment for thyrotrophinomas is surgical resection. In patients with surgical failure, SSA have been found to be effective in normalising TSH secretion in 90% of cases [249].

4.5.5 Non-functioning Pituitary Adenoma/(Gonadotroph Adenoma and Null Cell Pituitary Adenoma)

Non-functioning pituitary adenomas (NFPAs) are lesions without clinically obvious excess secretion of a hormone. They represent up to 43% of all pituitary adenomas [21], with the manifestation usually in the fourth or fifth decade of life; among children, NFPAs are responsible for only 4–6% of pituitary tumours [19, 20]. Recently, some studies reported an increase of non-functioning pituitary adenoma as a result of better quality and better access to imagine techniques and are considered as the most prevalent [21].

The vast majority of NFPAs are gonadotroph adenomas with very few showing no expression of any hormones or their transcription factors, especially at the RNA level. Silent ACTH, GH, prolactin or TSH adenomas representing a significant minority of clinically non-functioning adenomas have also been described. Null cell adenomas are diagnosed if they do not stain for transcription factors for either of the linages (PIT-1, TPIT, SF1) nor for any of adenohypophyseal hormones [41, 109]. Surgery is the preferred first-line treatment, as most are diagnosed as macroadenomas. Cabergoline treatment has been shown to have stabilising effect in a subset of NFPAs [250].

AIP mutation – Among *AIP*-related pituitary adenomas, NFPA occur in about 10% of cases, but 60% of these are diagnosed as part of follow-up of *AIP* carriers,



Fig. 4.7 Suggested follow-up of family members of an *AIP* mutation carrier (adapted from Williams et al. 2014 [209])

similar to MEN1 cases, in mutation carriers and it is difficult to distinguish them from incidentalomas identified in patients without *AIP* mutation.

MEN1 syndrome – NFPA are the third most common pituitary adenoma in MEN1 patients. Most of them are macroadenomas, although e-clinical screening has allowed to find many small lesions (42%) with an indolent clinical course [140].

4.6 Follow-Up and Prognosis

Patients with confirmed pituitary adenoma with genetic background should be treated and followed according to current international guidelines for their tumour type and systemic disorder. To enable early detection and therefore a better prognosis of pituitary adenomas, family cascade screening should be performed [79, 125, 135, 209]. The following flowcharts present a suggested follow-up of *AIP* (Fig. 4.7 adapted from Williams et al. 2014 [209]) and *MEN1* (Fig. 4.8 from Thakker et al. 2014 [131]) mutation carrier.

Most pituitary adenomas related to genetic mutation are associated with more aggressive behaviour, i.e. extrasellar invasion, local invasiveness, greater tumour size and higher risk of pituitary apoplexy. Histopathological features may show elevated staining for Ki67 and reduced expression of SSTR. They result in a lower response rate to medical treatment with SSA and often a multimodal approach is required to achieve disease control.

4.7 Further Considerations on Genetic Testing

Genetics is becoming an increasingly important aspect of patient care. Genetic testing for a monogenic disease may result, in general, in three possible scenarios: (1) A pathogenic variant is identified. In this case, the mutation explains the disease and the patient can have further testing, if necessary, to identify other manifestations of the disease and can be treated and follow-up according to available guidelines. Furthermore, cascade testing of the family members can be initiated. (2) The genetic testing can be negative, in this case other genetic causes could be searched for. (3) The third possibility, and this is often the case, a variant of uncertain significance is identified in a particular gene known to be associated with the disease. In this case the disease cannot be attributed to this gene, and family testing is not recommended; however, patients can be later notified if the status of the particular variant has changed based on other patients identified, emerging experimental or genetic data. The genetic landscape is further complicated by the incomplete and variable penetrance of some of the genes. Whereas GPR101, PRKAR1, MEN1 and NF1 mutations result in almost 100% penetrance, only 20-30% of AIP-carrier patients present clinical signs of the disease and pituitary specific penetrance is less than 100% in most of the genes, except GPR101, with very low penetrance in DICER1 and SDHx. Identifying factors that play a role in gene penetrance would improve follow-up of asymptomatic patients and genetic counselling.



Fig. 4.8 Suggested follow-up of *MEN1* mutation carriers (from Thakker et al. 2014 [131]). *PIT* pituitary, *PANC* pancreas, *ADR* adrenal, *CAR* carcinoid, *PTH* parathyroid hormone, *PRL* prolactin, *IGF-1* insulin-growth-factor-1, *CgA* chromogranin A, and g–i, and gastro-intestinal gut-hormones

Another challenging aspect is false-negative genetic results. In case of strong clinical suspicion of genetic background without a genetic mutation confirmed in single analysis, physicians should use alternative molecular techniques, e.g. MLPA analysis should be performed in case of negative Sanger sequencing in tumour suppressor genes. Intronic mutations causing crypti splice sites and pseudoexons can be identified with cDNA sequencing. Some of these problems can be overcome with next-generation sequencing techniques. Moreover, due to somatic mosaicism, tissue testing could provide answer, such as in *GNAS* in McCune-Albright syndrome, *GRP101* in XLAG [251], and more recently in *MEN1* [66]. When inherited disease with complete penetrance occurs in more than one offspring, germline mosaicism of asymptomatic parents could be considered. Additionally, negative result for the most common genetic causes of a disease could initiate search for an alternative cause, such as cell cycle proteins in *MEN*-negative MEN1 syndrome patients.

4.8 Highlighted Definitions

XLAG – In infant-onset of somatotrophinoma, X-linked acrogigantism diagnosis should be considered. This recently described disease is caused by X26.3 microduplication located at the locus of the *GRP101* gene and leads to overexpression of an orphan G protein-coupled receptor (GPCR) in pituitary tissue.

FIPA – A familial/genetic condition associated with a pituitary adenoma and no other features of a syndrome known to be associated with pituitary adenomas.

MEN1 – An autosomal dominant genetic disorder caused by inactivating mutation in *MEN1* gene. Characteristic tumours are parathyroid adenoma (with prevalence of 90%), pancreatic neuroendocrine tumour, and pituitary adenoma, with other organs often affected such as skin fibromas and adrenal tumours.

McCune-Albright syndrome – A systemic disorder characterised by the occurrence of three classic features: polycystic fibrous dysplasia, precocious puberty, and café au lait spots, caused by an activating mutation of *GNAS* gene at an early postzygotic stage and leading to somatic mosaicism.

3 Pa – Association of pituitary adenoma with paraganglioma/pheochromocytoma ("the three P association") is low penetrance condition, caused by germline mutation of the *SDHx*, *MAX*, *MEN1*, and has been also described in cases with *RET* and *VHL* gene.

DICER1 – A systemic disorder associated with a very early onset of ACTHdependent Cushing's disease and pleuropulmonary blastoma, cystic nephroma, pineoblastoma, ovarian Sertoli–Leydig cell tumours and multinodular goitre. Pituitary blastoma is a very rare manifestation of DICER1 syndrome with a low penetrance (<1% of cases) but represents a pathognomonic symptom.

References

- 1. Melmed S. Pituitary tumors. Endocrinol Metab Clin N Am. 2015;44:1-9.
- Devnath S, Inoue K. An insight to pituitary folliculo-stellate cells. J Neuroendocrinol. 2008;20:687–91.
- Daly AF, Rixhon M, Adam C, Dempegioti A, Tichomirowa MA, Beckers A. High prevalence of pituitary adenomas: a cross-sectional study in the province of Liège, Belgium. J Clin Endocrinol Metab. 2006;91:4769–75.
- Fernandez A, Karavitaki N, Wass JAHH. Prevalence of pituitary adenomas: a communitybased, cross-sectional study in Banbury (Oxfordshire, UK). Clin Endocrinol. 2010;72:377–82.
- Wiegering V, Eyrich M, Rutkowski S, Wölfl M, Schlegel PG, Winkler B. TH1 predominance is associated with improved survival in pediatric medulloblastoma patients. Cancer Immunol Immunother. 2011;60:693–703.
- Fontana E, Gaillard R. Epidemiology of pituitary adenoma: results of the first Swiss study. Rev Med Suisse. 2009;5:2172–4.
- Gruppetta M, Mercieca C, Vassallo J. Prevalence and incidence of pituitary adenomas: a population based study in Malta. Pituitary. 2013;16:545–53.
- Raappana A, Koivukangas J, Pirilä T. 3D modeling-based surgical planning in transsphenoidal pituitary surgery—preliminary results. Acta Otolaryngol. 2008;128:1011–8.
- 9. Daly AF, Jaffrain-Rea ML, Ciccarelli A, et al. Clinical characterization of familial isolated pituitary adenomas. J Clin Endocrinol Metab. 2006;91:3316–23.

- Iacovazzo D, Hernández-Ramírez LC, Korbonits M. Sporadic pituitary adenomas: the role of germline mutations and recommendations for genetic screening. Expert Rev Endocrinol Metab. 2017;12:143–53.
- Lim CT, Korbonits M. Update on the clinicopathology of pituitary adenomas. Endocr Pract. 2018;24:473–88.
- Daly AF, Tichomirowa MA, Beckers A. The epidemiology and genetics of pituitary adenomas. Best Pract Res Clin Endocrinol Metab. 2009;23:543–54.
- 13. Ezzat S, Asa SL, Couldwell WT, Barr CE, Dodge WE, Vance ML, McCutcheon IE. The prevalence of pituitary adenomas: a systematic review. Cancer. 2004;101:613–9.
- McDowell BD, Wallace RB, Carnahan RM, Chrischilles EA, Lynch CF, Schlechte JA. Demographic differences in incidence for pituitary adenoma. Pituitary. 2011;14:23–30.
- Melmed S, Casanueva FF, Hoffman AR, Kleinberg DL, Montori VM, Schlechte JA, Wass JAH. Diagnosis and treatment of hyperprolactinemia: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2011;96:273–88.
- Rostomyan L, Daly AF, Petrossians P, et al. Clinical and genetic characterization of pituitary gigantism: an international collaborative study in 208 patients. Endocr Relat Cancer. 2015;22:745–57.
- 17. Iacovazzo D, Caswell R, Bunce B, et al. Germline or somatic GPR101 duplication leads to X-linked acrogigantism: a clinico-pathological and genetic study. Acta Neuropathol Commun. 2016;4:56.
- 18. Kunwar S, Wilson CB. Pediatric pituitary adenomas. J Clin Endocrinol Metab. 1999;84:4385–9.
- Guaraldi F, Storr HL, Ghizzoni L, Ghigo E, Savage MO. Paediatric pituitary adenomas: a decade of change. Horm Res Paediatr. 2014;81:145–55.
- Yamaguchi-Okada M, Inoshita N, Nishioka H, Fukuhara N, Yamada S. Clinicopathological analysis of nonfunctioning pituitary adenomas in patients younger than 25 years of age: clinical article. J Neurosurg Pediatr. 2012;9:511–6.
- Agustsson TT, Baldvinsdottir T, Jonasson JG, Olafsdottir E, Steinthorsdottir V, Sigurdsson G, Thorsson AV, Carroll PV, Korbonits M, Benediktsson R. The epidemiology of pituitary adenomas in Iceland, 1955-2012: a nationwide population-based study. Eur J Endocrinol. 2015;173:655–64.
- Aflorei ED, Korbonits M. Epidemiology and etiopathogenesis of pituitary adenomas. J Neuro-Oncol. 2014;117:379–94.
- 23. Hanahan D, Weinberg RA. The hallmarks of cancer. Cell. 2000;100:57-70.
- Korbonits M, Morris DG, Nanzer A, Kola B, Grossman AB. Role of regulatory factors in pituitary tumour formation. Front Horm Res. 2004;32:63–95.
- 25. Asa SL, Ezzat S. Genetics and proteomics of pituitary tumors. Endocrine. 2005;28:43-7.
- Yu R, Bonert V, Saporta I, Raffel LJ, Melmed S. Aryl hydrocarbon receptor interacting protein variants in sporadic pituitary adenomas. J Clin Endocrinol Metab. 2006;91:5126–9.
- 27. Fedele M, Palmieri D, Fusco A. HMGA2: a pituitary tumour subtype-specific oncogene? Mol Cell Endocrinol. 2010;326:19–24.
- Daly AF, Vanbellinghen JF, Sok KK, et al. Aryl hydrocarbon receptor-interacting protein gene mutations in familial isolated pituitary adenomas: analysis in 73 families. J Clin Endocrinol Metab. 2007;92:1891–6.
- 29. Cannavo S, Trimarchi F, Ferraù F. Acromegaly, genetic variants of the aryl hydrocarbon receptor pathway and environmental burden. Mol Cell Endocrinol. 2017;457:81–8.
- Cannavo S, Ragonese M, Puglisi S, et al. Acromegaly is more severe in patients with AHR or AIP gene variants living in highly polluted areas. J Clin Endocrinol Metab. 2016;101:1872–9.
- Thomson EM, Kumarathasan P, Calderón-Garcidueñas L, Vincent R. Air pollution alters brain and pituitary endothelin-1 and inducible nitric oxide synthase gene expression. Environ Res. 2007;105:224–33.
- 32. Raverot G, Burman P, McCormack A, Heaney A, Petersenn S, Popovic V, Trouillas J, Dekkers OM. European society of endocrinology clinical practice guidelines for the management of aggressive pituitary tumours and carcinomas. Eur J Endocrinol. 2018;178:G1–G24.

- Trouillas J, Burman P, McCormack A, Petersenn S, Popovic V, Dekkers O, Raverot G. Aggressive pituitary tumours and carcinomas: two sides of the same coin? Eur J Endocrinol. 2018;178:C7–9.
- Asa SL, Kucharczyk W, Ezzat S. Pituitary acromegaly: not one disease. Endocr Relat Cancer. 2017;24:C1–4.
- 35. Kaltsas GA, Besser GM, Grossman AB. The diagnosis and medical management of advanced neuroendocrine tumors. Endocr Rev. 2004;25:458–511.
- 36. Rindi G, Klimstra DS, Abedi-Ardekani B, et al. A common classification framework for neuroendocrine neoplasms: an International Agency for Research on Cancer (IARC) and World Health Organization (WHO) expert consensus proposal. Mod Pathol. 2018;31:1770–86.
- Ho KKY, Fleseriu M, Wass J, et al. The tale in evolution: clarity, consistency and consultation, not contradiction and confusion. Pituitary. 2020;23:5–6.
- Asa SL, Asioli S, Bozkurt S, et al. Pituitary neuroendocrine tumors (PitNETs): nomenclature evolution, not clinical revolution. Pituitary. 2020;23:322–5.
- Ho KKY, Fleseriu M, Wass J, et al. A tale of pituitary adenomas: to NET or not to NET: pituitary society position statement. Pituitary. 2019;22:569–73.
- Asa SL, Casar-Borota O, Chanson P, et al. From pituitary adenoma to pituitary neuroendocrine tumor (pitnet): an international pituitary pathology club proposal. Endocr Relat Cancer. 2017;24:C5–8.
- 41. Lopes MBS. The 2017 World Health Organization classification of tumors of the pituitary gland: a summary. Acta Neuropathol. 2017;134:521–35.
- Neou M, Villa C, Armignacco R, et al. Pangenomic classification of pituitary neuroendocrine tumors. Cancer Cell. 2020;37:123–134.e5.
- Herman V, Fagin J, Gonsky R, Kovacs K, Melmed S. Clonal origin of pituitary adenomas. J Clin Endocrinol Metab. 1990;71:1427–33.
- 44. Clayton RN, Pfeifer M, Atkinson AB, Belchetz P, Wass JAH, Kyrodimou E, Vanderpump M, Simpson D, Bicknell J, Farrell WE. Different patterns of allelic loss (loss of heterozygosity) in recurrent human pituitary tumors provide evidence for multiclonal origins. Clin Cancer Res. 2000;6:3973–82.
- 45. Zhou Y, Zhang X, Klibanski A. Genetic and epigenetic mutations of tumor suppressive genes in sporadic pituitary adenoma. Mol Cell Endocrinol. 2014;386:16–33.
- Alexander JM, Biller BM, Bikkal H, Zervas NT, Arnold A, Klibanski A. Clinically nonfunctioning pituitary tumors are monoclonal in origin. J Clin Invest. 1990;86:336–40.
- Biller BMK, Alexander JM, Zervas NT, Hedley-Whyte ET, Arnold A, Klibanski A. Clonal origins of adrenocorticotropin-secreting pituitary tissue in Cushing's disease. J Clin Endocrinol Metab. 1992;75:1303–9.
- Gicquel C, Le Bouc Y, Luton JP, Girard F, Bertagna X. Monoclonality of corticotroph macroadenomas in cushing's disease. J Clin Endocrinol Metab. 1992;75:472–5.
- 49. Marques P, Barry S, Ronaldson A, et al. Emergence of pituitary adenoma in a child during surveillance: clinical challenges and the family members' view in an AIP mutation-positive family. Int J Endocrinol. 2018;2018:8581626.
- Leontiou CA, Gueorguiev M, Van Der Spuy J, et al. The role of the aryl hydrocarbon receptorinteracting protein gene in familial and sporadic pituitary adenomas. J Clin Endocrinol Metab. 2008;93:2390–401.
- Peverelli E, Mantovani G, Lania AG, Spada A. cAMP in the pituitary: an old messenger for multiple signals. J Mol Endocrinol. 2013; https://doi.org/10.1530/JME-13-0172.
- Ma ZY, Song ZJ, Chen JH, et al. Recurrent gain-of-function USP8 mutations in Cushing's disease. Cell Res. 2015;25:306–17.
- Komada M. Mutations in the deubiquitinase gene USP8 cause Cushing's disease. Nat Genet. 2015;47:31–8.
- Ballmann C, Thiel A, Korah HE, et al. USP8 mutations in pituitary cushing adenomas—targeted analysis by next-generation sequencing. J Endocr Soc. 2018;2:266–78.

- 55. Pérez-Rivas LG, Theodoropoulou M, Puar TH, et al. Somatic USP8 mutations are frequent events in corticotroph tumor progression causing Nelson's tumor. Eur J Endocrinol. 2018;178:57–63.
- 56. Dworakowska D, Wlodek E, Leontiou CA, et al. Activation of RAF/MEK/ERK and PI3K/ AKT/mTOR pathways in pituitary adenomas and their effects on downstream effectors. Endocr Relat Cancer. 2009;16:1329–38.
- Orloff MS, He X, Peterson C, Chen F, Chen JL, Mester JL, Eng C. Germline PIK3CA and AKT1 mutations in cowden and cowden-like syndromes. Am J Hum Genet. 2013;92:76–80.
- 58. Salomon MP, Wang X, Marzese DM, et al. The epigenomic landscape of pituitary adenomas reveals specific alterations and differentiates among acromegaly, Cushing's disease and endocrine-inactive subtypes. Clin Cancer Res. 2018;24:4126–36.
- 59. Song ZJ, Reitman ZJ, Ma ZY, et al. The genome-wide mutational landscape of pituitary adenomas. Cell Res. 2016;26:1255–9.
- Bi WL, Greenwald NF, Ramkissoon SH, et al. Clinical identification of oncogenic drivers and copy-number alterations in pituitary tumors. Endocrinology. 2017;158:2284–91.
- Li C, Xie W, Rosenblum JS, et al. Somatic SF3B1 hotspot mutation in prolactinomas. Nat Commun. 2020; https://doi.org/10.1038/s41467-020-16052-8.
- 62. Chen J, Jian X, Deng S, et al. Identification of recurrent USP48 and BRAF mutations in Cushing's disease. Nat Commun. 2018;9:3171.
- Tanizaki Y, Jin L, Scheithauer BW, Kovacs K, Roncaroli F, Lloyd RV. P53 gene mutations in pituitary carcinomas. Endocr Pathol. 2007;18:217–22.
- Caimari F, Korbonits M. Novel genetic causes of pituitary adenomas. Clin Cancer Res. 2016;22:5030–42.
- Mauchlen R, Carty D, Talla M, Drummond R. Multiple endocrine neoplasia type 1 (MEN1) mosaicism caused by a c.124G>A variant in the MEN1 gene. Endocr Abstr. 2019; https://doi. org/10.1530/endoabs.65.cc4.
- 66. Beijers HJBH, Stikkelbroeck NML, Mensenkamp AR, Pfundt R, Van Der Luijt RB, Timmers HJLM, Hermus ARMM, Kempers MJE. Germline and somatic mosaicism in a family with multiple endocrine neoplasia type 1 (MEN1) syndrome. Eur J Endocrinol. 2019;180:K15–9.
- 67. Stelmachowska-Banas M, Zgliczynski W, Tutka P, Carney JA, Korbonits M. Fatal Carney complex in siblings due to de novo large gene deletion. J Clin Endocrinol Metab. 2017;102:3924–7.
- Srirangam Nadhamuni V, Korbonits M. Novel insights into pituitary tumorigenesis: genetic and epigenetic mechanisms 2020, https://doi.org/10.1210/endrev/bnaa006/5810899.
- 69. Marques P, Barry S, Carlsen E, et al. Chemokines modulate the tumour microenvironment in pituitary neuroendocrine tumours. Acta Neuropathol Commun. 2019;7:172.
- Marques P, Barry S, Carlsen E, et al. Pituitary tumour fibroblast-derived cytokines influence tumour aggressiveness. Endocr Relat Cancer. 2019;26:853–65.
- Marques P, Grossman AB, Korbonits M. The tumour microenvironment of pituitary neuroendocrine tumours. Front Neuroendocrinol. 2020;100852
- Ilie MD, Vasiljevic A, Raverot G, Bertolino P. The microenvironment of pituitary tumorsbiological and therapeutic implications. Cancers (Basel). 2019;11:1–22.
- Donovan LE, Welch MR. Headaches in patients with pituitary tumors: a clinical conundrum. Curr Pain Headache Rep. 2018; https://doi.org/10.1007/s11916-018-0709-1.
- Gondim JA, de Almeida JPC, de Albuquerque LAF, Schops M, Gomes E, Ferraz T. Headache associated with pituitary tumors. J Headache Pain. 2009;10:15–20.
- 75. Arafah BM, Prunty D, Ybarra J, Hlavin ML, Selman WR. The dominant role of increased intrasellar pressure in the pathogenesis of hypopituitarism, hyperprolactinemia, and head-aches in patients with pituitary adenomas. J Clin Endocrinol Metab. 2000;85:1789–93.
- Yu B, Ji N, Ma Y, Yang B, Kang P, Luo F. Clinical characteristics and risk factors for headache associated with non-functioning pituitary adenomas. Cephalalgia. 2017;37:348–55.
- Kreitschmann-Andermahr I, Siegel S, Weber Carneiro R, Maubach JM, Harbeck B, Brabant G. Headache and pituitary disease: a systematic review. Clin Endocrinol. 2013;79:760–9.

- Levy MJ, Matharu M, Goadsby PJ. Chronic headache and pituitary tumors. Curr Pain Headache Rep. 2008;12:74–8.
- Hernández-Ramírez LC, Gabrovska P, Dénes J, et al. Landscape of familial isolated and young-onset pituitary adenomas: prospective diagnosis in AIP mutation carriers. J Clin Endocrinol Metab. 2015;100:E1242–54.
- Lee IH, Miller NR, Zan E, Tavares F, Blitz AM, Sung H, Yousem DM, Boland MV. Visual defects in patients with pituitary adenomas: the myth of bitemporal hemianopsia. AJR Am J Roentgenol. 2015;205:W512–8.
- Huang WC, Lee LS. Visual field defects in patients with pituitary adenomas. Zhonghua Yi Xue Za Zhi (Taipei). 1997;60:245–51.
- Bergsneider M, Mirsadraei L, Yong WH, Salamon N, Linetsky M, Wang MB, McArthur DL, Heaney AP. The pituitary stalk effect: is it a passing phenomenon? J Neuro-Oncol. 2014;117:477–84.
- Ramírez-Rentería C, Hernández-Ramírez LC, Portocarrero-Ortiz L, et al. AIP mutations in young patients with acromegaly and the Tampico Giant: the Mexican experience. Endocrine. 2016;53:402–11.
- Matsumoto R, Izawa M, Fukuoka H, et al. Genetic and clinical characteristics of Japanese patients with sporadic somatotropinoma. Endocr J. 2016;63:953–63.
- Patt HP, Bothra N, Goel AH, Kasaliwal R, Lila AR, Bandgar TR, Shah NS. Pituitary gigantism—experience of a single center from Western India. Endocr Pract. 2015;21:621–8.
- Joshi K, Daly AF, Beckers A, Zacharin M. Resistant paediatric somatotropinomas due to AIP mutations: role of pegvisomant. Horm Res Paediatr. 2018;90:196–202.
- Takumi A, Tara LA, Lüdecke DK. Growth hormone-secreting pituitary adenomas in childhood and adolescence: features and results of transnasal surgery. Neurosurgery. 1999;45:1–10.
- Bergamaschi S, Ronchi CL, Giavoli C, Ferrante E, Verrua E, Ferrari DI, Lania A, Rusconi R, Spada A, Beck-Peccoz P. Eight-year follow-up of a child with a GH/prolactin-secreting adenoma: efficacy of pegvisomant therapy. Horm Res Paediatr. 2010;73:74–9.
- Bowden CL, Grunze H, Mullen J, Brecher M, Paulsson B, Jones M, Vågerö M, Svensson K. A randomized, double-blind, placebo-controlled efficacy and safety study of quetiapine or lithium as monotherapy for mania in bipolar disorder. J Clin Psychiatry. 2005;66:111–21.
- Ciresi A, Amato MC, Galluzzo A, Giordano C. Complete biochemical control and pituitary adenoma disappearance in a child with gigantism: efficacy of octreotide therapy. J Endocrinol Investig. 2011;34:162–3.
- Colao A, Pivonello R, Di Somma C, Tauchmanovà L, Savastano S, Lombardi G. Growth hormone excess with onset in adolescence: clinical appearance and long-term treatment outcome. Clin Endocrinol. 2007;66:714–22.
- Goldenberg N, Racine MS, Thomas P, Degnan B, Chandler W, Barkan A. Treatment of pituitary gigantism with the growth hormone receptor antagonist pegvisomant. J Clin Endocrinol Metab. 2008;93:2953–6.
- 93. Mete O, Gomez-Hernandez K, Kucharczyk W, Ridout R, Zadeh G, Gentili F, Ezzat S, Asa SL. Silent subtype 3 pituitary adenomas are not always silent and represent poorly differentiated monomorphous plurihormonal pit-1 lineage adenomas. Mod Pathol. 2016;29:131–42.
- Rostomyan L, Daly AF, Beckers A. Pituitary gigantism: causes and clinical characteristics. Ann Endocrinol (Paris). 2015;76:643–9.
- Joustra SD, Roelfsema F, Van Trotsenburg ASP, et al. IGSF1 deficiency results in human and murine somatotrope neurosecretory hyperfunction. J Clin Endocrinol Metab. 2020;105:70–84.
- Storr HL, Alexandraki KI, Martin L, et al. Comparisons in the epidemiology, diagnostic features and cure rate by transsphenoidal surgery between paediatric and adult-onset Cushing's disease. Eur J Endocrinol. 2011;164:667–74.
- Lonser RR, Wind JJ, Nieman LK, Weil RJ, DeVroom HL, Oldfield EH. Outcome of surgical treatment of 200 children with cushing's disease. J Clin Endocrinol Metab. 2013;98:892–901.
- Höybye C, Grenbäck E, Rähn T, Degerblad M, Thorén M, Hulting AL. Adrenocorticotropic hormone-producing pituitary tumors: 12- to 22-year follow-up after treatment with stereotactic radiosurgery. Neurosurgery. 2001;49:284–92.

- Chan LF, Storr HL, Grossman AB, Savage MO. Pediatric Cushing's syndrome: clinical features, diagnosis, and treatment. Arq Bras Endocrinol Metabol. 2007;51:1261–71.
- Magiakou MA, Smyrnaki P, Chrousos GP. Hypertension in Cushing's syndrome. Best Pract Res Clin Endocrinol Metab. 2006;20:467–82.
- 101. Magiakou MA, Manousaki D, Papadaki M, et al. The efficacy and safety of gonadotropinreleasing hormone analog treatment in childhood and adolescence: a single center, long-term follow-up study. J Clin Endocrinol Metab. 2010;95:109–17.
- 102. Dorn LD, Burgess ES, Dubbert B, Simpson SE, Friedman T, Kling M, Gold PW, Chrousos GP. Psychopathology in patients with endogenous Cushing's syndrome: "atypical" or melancholic features. Clin Endocrinol. 1995;43:433–42.
- 103. Dorn LD, Burgess ES, Friedman TC, Dubbert B, Gold PW, Chrousos GP. The longitudinal course of psychopathology in Cushing's syndrome after correction of hypercortisolism. J Clin Endocrinol Metab. 1997;82:912–9.
- Duskin-Bitan H, Shimon I. Prolactinomas in males: any differences? Pituitary. 2019; https:// doi.org/10.1007/s11102-019-01009-y.
- 105. Mazziotti G, Porcelli T, Mormando M, De Menis E, Bianchi A, Mejia C, Mancini T, De Marinis L, Giustina A. Vertebral fractures in males with prolactinoma. Endocrine. 2011;39:288–93.
- 106. Andereggen L, Frey J, Andres RH, El-Koussy M, Beck J, Seiler RW, Christ E. Long-term follow-up of primary medical versus surgical treatment of prolactinomas in men: effects on hyperprolactinemia, hypogonadism, and bone health. World Neurosurg. 2017;97:595–602.
- 107. Atmaca A, Bilgici B, Ecemis GC, Tuncel OK. Evaluation of body weight, insulin resistance, leptin and adiponectin levels in premenopausal women with hyperprolactinemia. Endocrine. 2012;44:756–61.
- Auriemma RS, De Alcubierre D, Pirchio R, Pivonello R, Colao A. The effects of hyperprolactinemia and its control on metabolic diseases. Expert Rev Endocrinol Metab. 2018;13:99–106.
- Drummond J, Roncaroli F, Grossman AB, Korbonits M. Clinical and pathological aspects of silent pituitary adenomas. J Clin Endocrinol Metab. 2019;104:2473–89.
- 110. Cyprich J, Donoho DA, Brunswick A, Hurth K, Carmichael JD, Weiss MH, Zada G. Surgical management of clinically silent thyrotropin pituitary adenomas: a single center series of 20 patients. J Clin Neurosci. 2019; https://doi.org/10.1016/j.jocn.2019.10.013.
- 111. Herguido NG, Fuentes ED, Venegas-Moreno E, et al. Surgical outcome and treatment of thyrotropin-secreting pituitary tumors in a tertiary referral center. World Neurosurg. 2019;130:e634–9.
- 112. Chaidarun SS, Klibanski A. Gonadotropinomas. Semin Reprod Med. 2002;20:339-48.
- Ntali G, Capatina C, Grossman A, Karavitaki N. Functioning gonadotroph adenomas. J Clin Endocrinol Metab. 2014;99:4423–33.
- 114. Nachtigall LB, Guarda FJ, Lines KE, et al. Clinical MEN-1 among a large cohort of patients with acromegaly. J Clin Endocrinol Metab. 2020;105:1–11.
- 115. Beckers A, Aaltonen LA, Daly AF, Karhu A. Familial isolated pituitary adenomas (FIPA) and the pituitary adenoma predisposition due to mutations in the aryl hydrocarbon receptor interacting protein (AIP) gene. Endocr Rev. 2013;34:239–77.
- Iacovazzo D, Korbonits M. Gigantism: X-linked acrogigantism and GPR101 mutations. Growth Hormon IGF Res. 2016;30–31:64–9.
- 117. Trivellin G, Daly AF, Faucz FR, et al. Supplementary data for Trivellin 2014_Gigantism and acromegaly due to Xq26 microduplications and GPR101 mutation. N Engl J Med. 2014; https://doi.org/10.1056/NEJMoa1408028.
- Trivellin G, Korbonits M. AIP interactors. J Endocrinol. 2011; https://doi.org/10.1530/ JOE-11-0054.
- 119. Stockinger B, Di MP, Gialitakis M, Duarte JH. The aryl hydrocarbon receptor: multitasking in the immune system. Annu Rev Immunol. 2014;32:403–32.
- 120. Hernández-Ramírez LC, Martucci F, Morgan RML, Trivellin G, Tilley D, Ramos-Guajardo N, Iacovazzo D, D'Acquisto F, Prodromou C, Korbonits M. Rapid proteasomal degradation of mutant proteins is the primary mechanism leading to tumorigenesis in patients with missense AIP mutations. J Clin Endocrinol Metab. 2016;101:3144–54.

- 121. Gadelha MR, Prezant TR, Une KN, Glick RP, Moskal SF, Vaisman M, Melmed S, Kineman RD, Frohman LA. Loss of heterozygosity on chromosome 11q13 in two families with acromegaly/gigantism is independent of mutations of the multiple endocrine neoplasia type I gene 1. J Clin Endocrinol Metab. 1999;84:249–56.
- 122. Vierimaa O, Georgitsi M, Lehtonen R, et al. Pituitary adenoma predisposition caused by germline mutations in the AIP gene. Science (80-). 2006;312:1228–30.
- 123. Stratakis CA, Tichomirowa MA, Boikos S, et al. The role of germline AIP, MEN1, PRKAR1A, CDKN1B and CDKN2C mutations in causing pituitary adenomas in a large cohort of children, adolescents, and patients with genetic syndromes. Clin Genet. 2010;78:457–63.
- 124. Marques P, Caimari F, Hernández-Ramírez LC, et al. Significant benefits of AIP testing and clinical screening in familial isolated and young-onset pituitary tumors. J Clin Endocrinol Metab. 2020; https://doi.org/10.1210/clinem/dgaa040.
- 125. Daly AF, Tichomirowa MA, Petrossians P, et al. Clinical characteristics and therapeutic responses in patients with germ-line AIP mutations and pituitary adenomas: an international collaborative study. J Clin Endocrinol Metab. 2010; https://doi.org/10.1210/jc.2009-2556.
- 126. Igreja S, Chahal HS, King P, et al. Characterization of aryl hydrocarbon receptor interacting protein (AIP) mutations in familial isolated pituitary adenoma families. Hum Mutat. 2010;31:950–60.
- 127. Vandeva S, Daly AF, Petrossians P, Zacharieva S, Beckers A. Somatic and germline mutations in the pathogenesis of pituitary adenomas. Eur J Endocrinol. 2019;181:R235–54.
- 128. Trivellin G, Hernández-Ramírez LC, Swan J, Stratakis CA. An orphan G-protein-coupled receptor causes human gigantism and/or acromegaly: molecular biology and clinical correlations. Best Pract Res Clin Endocrinol Metab. 2018;32:125–40.
- 129. Zhang Q, Peng C, Song J, et al. Germline mutations in CDH23, encoding cadherin-related 23, are associated with both familial and sporadic pituitary adenomas. Am J Hum Genet. 2017;100:817–23.
- Hernández-Ramírez LC, Gam R, Valdés N, et al. Loss-of-function mutations in the CABLES1 gene are a novel cause of Cushing's disease. Endocr Relat Cancer. 2017;24:379–92.
- 131. Thakker RV. Multiple endocrine neoplasia type 1 (MEN1) and type 4 (MEN4). Mol Cell Endocrinol. 2014;386:2–15.
- 132. Thakker RV. Genetics of parathyroid tumours. J Intern Med. 2016;280:574-83.
- 133. Luzi E, Marini F, Giusti F, Galli G, Cavalli L, Brandi ML. The negative feedback-loop between the Oncomir mir-24-1 and Menin modulates the men1 tumorigenesis by mimicking the "Knudson's second hit.". PLoS One. 2012; https://doi.org/10.1371/journal.pone.0039767.
- Thakker RV. Multiple endocrine neoplasia type 1 (MEN1). Best Pract Res Clin Endocrinol Metab. 2010;24:355–70.
- 135. Thakker RV, Newey PJ, Walls GV, Bilezikian J, Dralle H, Ebeling PR, Melmed S, Sakurai A, Tonelli F, Brandi ML. MEN1 2012 Guideline. J Clin Endocrinol Metab. 2012; https://doi.org/10.1210/jc.2012-1230.
- 136. Stratakis CA, Schussheim DH, Freedman SM, et al. Pituitary macroadenoma in a 5-yearold: an early expression of multiple endocrine neoplasia type 1 1. J Clin Endocrinol Metab. 2000;85:4776–80.
- 137. Goudet P, Dalac A, Le Bras M, et al. MEN1 disease occurring before 21 years old: a 160-patient cohort study from the Groupe d'étude des Tumeurs endocrines. J Clin Endocrinol Metab. 2015;100:1568–77.
- 138. Schernthaner-Reiter MH, Trivellin G, Stratakis CA. MEN1, MEN4, and Carney complex: pathology and molecular genetics. Neuroendocrinology. 2016;103:18–31.
- 139. Vergès B, Boureille F, Goudet P, Murat A, Beckers A, Sassolas G, Cougard P, Chambe B, Montvernay C, Calender A. Pituitary disease in MEN type 1 (MEN1): data from the France-Belgium MEN1 Multicenter Study. J Clin Endocrinol Metab. 2002;87:457–65.
- 140. De Laat JM, Dekkers OM, Pieterman CRC, et al. Long-term natural course of pituitary tumors in patients with MEN1: results from the Dutch MEN1 study group (DMSG). J Clin Endocrinol Metab. 2015;100:3288–96.

- 141. Cuny T, Pertuit M, Sahnoun-Fathallah M, et al. Genetic analysis in young patients with sporadic pituitary macroadenomas: besides AIP don't forget MEN1 genetic analysis. Eur J Endocrinol. 2013;168:533–41.
- 142. Trouillas J, Labat-Moleur F, Sturm N, et al. Pituitary tumors and hyperplasia in multiple endocrine neoplasia type 1 syndrome (MEN1): a case-control study in a series of 77 patients versus 2509 non-MEN1 patients. Am J Surg Pathol. 2008;32:534–43.
- 143. Cavaco BM, Domingues R, Bacelar MC, et al. Mutational analysis of Portuguese families with multiple endocrine neoplasia type 1 reveals large germline deletions. Clin Endocrinol. 2002;56:465–73.
- 144. Pellegata NS, Quintanilla-Martinez L, Siggelkow H, Samson E, Bink K, Höfler H, Fend F, Graw J, Atkinson MJ. Germ-line mutations in p27Kip1 cause a multiple endocrine neoplasia syndrome in rats and humans. Proc Natl Acad Sci U S A. 2006;103:15558–63.
- 145. Frederiksen A, Rossing M, Hermann P, Ejersted C, Thakker RV, Frost M. Clinical features of multiple endocrine neoplasia type 4: novel pathogenic variant and review of published cases. J Clin Endocrinol Metab. 2019;104:3637–46.
- 146. Agarwal SK, Mateo CM, Marx SJ. Rare germline mutations in cyclin-dependent kinase inhibitor genes in multiple endocrine neoplasia type 1 and related states. J Clin Endocrinol Metab. 2009;94:1826–34.
- 147. Turner JJO, Christie PT, Pearce SHS, Turnpenny PD, Thakker RV. Diagnostic challenges due to phenocopies: lessons from multiple endocrine neoplasia type1 (MEN1). Hum Mutat. 2010; https://doi.org/10.1002/humu.21170.
- 148. Lines KE, Nachtigall LB, Dichtel LE, et al. Multiple endocrine neoplasia type 1 (MEN1) phenocopy due to a P.Leu380Phe cell division cycle 23 (CDC73) mutation. Endocr Abstr. 2017; https://doi.org/10.1530/endoabs.50.p249.
- 149. Backman S, Bajic D, Crona J, Hellman P, Skogseid B, Stålberg P. Whole genome sequencing of apparently mutation-negative MEN1 patients. Eur J Endocrinol. 2020;182:35–45.
- 150. Saito T, Miura D, Taguchi M, Takeshita A, Miyakawa M, Takeuchi Y. Coincidence of multiple endocrine neoplasia type 2A with acromegaly. Am J Med Sci. 2010;340:329–31.
- 151. Heinlen JE, Buethe DD, Culkin DJ, Slobodov G. Multiple endocrine neoplasia 2a presenting with pheochromocytoma and pituitary macroadenoma. ISRN Oncol. 2011;2011:1–4.
- 152. Naziat A, Karavitaki N, Thakker R, Ansorge O, Sadler G, Gleeson F, Cranston T, McCormack A, Grossman AB, Shine B. Confusing genes: a patient with MEN2A and Cushing's disease. Clin Endocrinol. 2013;78:966–8.
- 153. Kasturi K, Fernandes L, Quezado M, Eid M, Marcus L, Chittiboina P, Rappaport M, Stratakis CA, Widemann B, Lodish M. Cushing disease in a patient with multiple endocrine neoplasia type 2B. J Clin Transl Endocrinol Case Rep. 2017;4:1–4.
- 154. Aidan Carney J, Gordon H, Carpenter PC, Vittal Shenoy B, Go VLW. The complex of myxomas, spotty pigmentation, and endocrine overactivity. Medicine (United States). 1985;64:270–83.
- 155. Bossis I, Stratakis CA. Minireview: PRKAR1A: normal and abnormal functions. Endocrinology. 2004;145:5452–8.
- 156. Salpea P, Horvath A, London E, et al. Deletions of the PRKAR1A locus at 17q24.2-q24.3 in Carney complex: genotype-phenotype correlations and implications for genetic testing. J Clin Endocrinol Metab. 2014;99:E183–8.
- 157. Forlino A, Vetro A, Garavelli L, Ciccone R, London E, Stratakis CA, Zuffardi O. PRKACB and Carney complex. N Engl J Med. 2014;370:1065–7.
- 158. Beuschlein F, Fassnacht M, Assié G, et al. Constitutive activation of PKA catalytic subunit in adrenal Cushing's syndrome. N Engl J Med. 2014;370:1019–28.
- 159. Forlino A, Vetro A, Garavelli L, Ciccone R, London E, Stratakis C a, Zuffardi O (2014) Genetic diagnosis in whole genome sequencing. N Engl J Med 1067–1069.
- Kamilaris CDC, Faucz FR, Voutetakis A, Stratakis CA. Carney complex. Exp Clin Endocrinol Diabetes. 2019;127:156–64.

- 161. Bertherat J, Horvath A, Groussin L, et al. Mutations in regulatory subunit type 1A of cyclic adenosine 5'-monophosphate-dependent protein kinase (PRKAR1A): phenotype analysis in 353 patients and 80 different genotypes. J Clin Endocrinol Metab. 2009;94:2085–91.
- 162. Stratakis CA, Kirschner LS, Carney JA. Clinical and molecular features of the Carney complex: diagnostic criteria and recommendations for patient evaluation. J Clin Endocrinol Metab. 2001;86:4041–6.
- 163. Boikos SA, Stratakis CA. Carney complex: the first 20 years. Curr Opin Oncol. 2007;19:24-9.
- 164. Correa R, Salpea P, Stratakis CA. Carney complex: an update. In: Eur J Endocrinol BioScientifica Ltd, 2015 pp. M85–M97.
- 165. Kiefer FW, Winhofer Y, Iacovazzo D, et al. PRKAR1A mutation causing pituitary-dependent Cushing disease in a patient with Carney complex. Eur J Endocrinol. 2017;177:K7–K12.
- 166. Hernández-Ramírez LC, Tatsi C, Lodish MB, et al. Corticotropinoma as a component of Carney complex. J Endocr Soc. 2017;1:918–25.
- 167. Naito Y, Mori J, Tazoe J, Tomida A, Yagyu S, Nakajima H, Iehara T, Tatsuzawa K, Mukai T, Hosoi H. Pituitary apoplexy after cardiac surgery in a 14-year-old girl with Carney complex: a case report. Endocr J. 2019; https://doi.org/10.1507/endocrj.ej19-0183.
- Dumitrescu CE, Collins MT. McCune-Albright syndrome. Orphanet J Rare Dis. 2008; https://doi.org/10.1186/1750-1172-3-12.
- Weinstein LS, Shenker A, Friedman E, Spiegel AM, Gejman PV, Merino MJ. Activating mutations of the stimulatory g protein in the mcCune–albright syndrome. N Engl J Med. 1991;325:1688–95.
- 170. Shenker A, Weinstein LS, Sweet DE, Spiegel AM. An activating Gs alpha mutation is present in fibrous dysplasia of bone in the McCune-Albright syndrome. J Clin Endocrinol Metab. 1994;79:750–5.
- 171. Schwindinger WF, Francomano CA, Levine MA. Identification of a mutation in the gene encoding the α subunit of the stimulatory G protein of adenylyl cyclase in McCune-Albright syndrome. Proc Natl Acad Sci U S A. 1992;89:5152–6.
- 172. Chanson P, Salenave S, Orcel P. McCune-Albright syndrome in adulthood. Pediatr Endocrinol Rev. 2007;4(Suppl 4):453–62.
- 173. Chanson P, Salenave S, Young J. Ovarian dysfunction by activating mutation of GS alpha: McCune-Albright syndrome as a model. Ann Endocrinol (Paris). 2010;71:210–3.
- 174. Boyce AM, Glover M, Kelly MH, et al. Optic neuropathy in McCune-Albright syndrome: effects of early diagnosis and treatment of growth hormone excess. J Clin Endocrinol Metab. 2013; https://doi.org/10.1210/jc.2012-2111.
- 175. Salenave S, Boyce AM, Collins MT, Chanson P. Acromegaly and mccune-albright syndrome. J Clin Endocrinol Metab. 2014;99:1955–69.
- 176. Romanet P, Philibert P, Fina F, Cuny T, Roche C, Ouafik L, Paris F, Reynaud R, Barlier A. Using digital droplet polymerase chain reaction to detect the mosaic GNAS mutations in whole blood DNA or circulating cell-free DNA in fibrous dysplasia and McCune-Albright syndrome. J Pediatr. 2019;205:281–285.e4.
- 177. Iversen K. Acromegaly associated with Phæochromocytoma. Acta Med Scand. 1952;142:1-5.
- 178. O'Toole SM, Dénes J, Robledo M, Stratakis CA, Korbonits M. The association of pituitary adenomas and phaeochromocytomas or paragangliomas. Endocr Relat Cancer. 2015;22:T105–22.
- 179. Dénes J, Swords F, Rattenberry E, et al. Heterogeneous genetic background of the association of pheochromocytoma/paraganglioma and pituitary adenoma: results from a large patient cohort. J Clin Endocrinol Metab. 2015;100:E531–41.
- Xekouki P, Szarek E, Bullova P, et al. Pituitary adenoma with paraganglioma/pheochromocytoma (3PAs) and succinate dehydrogenase defects in humans and mice. J Clin Endocrinol Metab. 2015;100:E710–9.
- 181. Maher M, Roncaroli F, Mendoza N, et al. A patient with a germline SDHB mutation presenting with an isolated pituitary macroprolactinoma. Endocrinol Diabetes Metab Case Rep. 2018; https://doi.org/10.1530/EDM-18-0078.

- Tufton N, Roncaroli F, Hadjidemetriou I, et al. Pituitary carcinoma in a patient with an SDHB mutation. Endocr Pathol. 2017;28:320–5.
- 183. Carroll PA, Freie BW, Mathsyaraja H, Eisenman RN. The MYC transcription factor network: balancing metabolism, proliferation and oncogenesis. Front Med. 2018;12:412–25.
- 184. Daly AF, Castermans E, Oudijk L, et al. Pheochromocytomas and pituitary adenomas in three patients with MAX exon deletions. Endocr Relat Cancer. 2018;25:L37–42.
- 185. Roszko KL, Blouch E, Blake M, Powers JF, Tischler AS, Hodin R, Sadow P, Lawson EA. Case report of a prolactinoma in a patient with a novel MAX mutation and bilateral pheochromocytomas. J Endocr Soc. 2017;1:1401–7.
- 186. Romero OA, Torres-Diz M, Pros E, et al. MAX inactivation in small cell lung cancer disrupts MYC-SWI/SNF programs and is synthetic lethal with BRG1. Cancer Discov. 2014;4:293–303.
- 187. Comino-Méndez I, Gracia-Aznárez FJ, Schiavi F, et al (2011) Exome sequencing identifies MAX mutations as a cause of hereditary pheochromocytoma. In: Nat. Genet. pp. 663–667.
- Smith JC, Sheltzer JM. Systematic identification of mutations and copy number alterations associated with cancer patient prognosis. elife. 2018; https://doi.org/10.7554/eLife.39217.
- Tadini G, Milani D, Menni F, Pezzani L, Sabatini C, Esposito S. Is it time to change the neurofibromatosis 1 diagnostic criteria? 2014; https://doi.org/10.1016/j.ejim.2014.04.004.
- Milani D, Pezzani L, Tadini G, Menni F, Esposito S. A multidisciplinary approach in neurofibromatosis 1. Lancet Neurol. 2015;14:29–30.
- 191. Cambiaso P, Galassi S, Palmiero M, et al. Growth hormone excess in children with neurofibromatosis type-1 and optic glioma. Am J Med Genet Part A. 2017;173:2353–8.
- 192. Josefson J, Listernick R, Fangusaro JR, Charrow J, Habiby R. Growth hormone excess in children with neurofibromatosis type 1-associated and sporadic optic pathway tumors. J Pediatr. 2011;158:433–6.
- Bizzarri C, Bottaro G. Endocrine implications of neurofibromatosis 1 in childhood. Horm Res Paediatr. 2015;83:232–41.
- 194. Hozumi K, Fukuoka H, Odake Y, et al. Acromegaly caused by a somatotroph adenoma in patient with neurofibromatosis type 1. Endocr J. 2019;66:853–7.
- 195. Sahakitrungruang T, Srichomthong C, Pornkunwilai S, Amornfa J, Shuangshoti S, Kulawonganunchai S, Suphapeetiporn K, Shotelersuk V. Germline and somatic DICER1 mutations in a pituitary blastoma causing infantile-onset Cushing's disease. J Clin Endocrinol Metab. 2014; https://doi.org/10.1210/jc.2014-1016.
- 196. De Kock L, Sabbaghian N, Plourde F, et al. Pituitary blastoma: a pathognomonic feature of germ-line DICER1 mutations. Acta Neuropathol. 2014;128:111–22.
- 197. Terzic T, Stewart CJ, De Kock L, Foulkes W, McCluggage WG, Shaw P, Clarke B. Sertolileydig cell tumors: examination of morphologic features and correlation with germline and somatic dicer 1 mutation. Lab Investig Conf 106th Annu Meet united states Can Acad Pathol USCAP 2017 United states 2017 97:312A.
- 198. Cotton E, Ray D (2018) DICER1 mutation and pituitary prolactinoma. 2018.
- 199. Chinezu L, Vasiljevic A, Trouillas J, Lapoirie M, Jouanneau E, Raverot G. Silent somatotroph tumour revisited from a study of 80 patients with and without acromegaly and a review of the literature. Eur J Endocrinol. 2017;176:195–201.
- 200. Ribeiro-Oliveira A, Barkan A. The changing face of acromegaly—advances in diagnosis and treatment. Nat Rev Endocrinol. 2012;8:605–11.
- Lavrentaki A, Paluzzi A, Wass JAH, Karavitaki N. Epidemiology of acromegaly: review of population studies. Pituitary. 2017;20:4–9.
- 202. Burton T, Le Nestour E, Neary M, Ludlam WH. Incidence and prevalence of acromegaly in a large US health plan database. Pituitary. 2016;19:262–7.
- Katznelson L, Laws ER, Melmed S, Molitch ME, Murad MH, Utz A, Wass JAH. Acromegaly: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2014;99:3933–51.
- Schilbach K, Gar C, Lechner A, et al. Determinants of the growth hormone nadir during oral glucose tolerance test in adults. Eur J Endocrinol. 2019;181:55–67.

- 205. Wang M, Mou C, Jiang M, Han L, Fan S, Huan C, Qu X, Han T, Qu Y, Xu G. The characteristics of acromegalic patients with hyperprolactinemia and the differences in patients with merely GH-secreting adenomas: clinical analysis of 279 cases. Eur J Endocrinol. 2012;166:797–802.
- Katznelson L. Approach to the patient with persistent acromegaly after pituitary surgery. J Clin Endocrinol Metab. 2010;95:4114–23.
- 207. Kamenický P, Bouligand J, Chanson P. Gigantism, acromegaly, and GPR101 mutations. N Engl J Med. 2015;372:1264–5.
- Beckers A, Lodish MB, Trivellin G, et al. X-linked acrogigantism syndrome: clinical profile and therapeutic responses. Endocr Relat Cancer. 2015;22:353–67.
- 209. Williams F, Hunter S, Bradley L, et al. Clinical experience in the screening and management of a large kindred with familial isolated pituitary adenoma due to an aryl hydrocarbon receptor interacting protein (AIP) mutation. J Clin Endocrinol Metab. 2014;99:1122–31.
- Daly AF, Rostomyan L, Betea D, et al. Aip-mutated acromegaly resistant to first-generation somatostatin analogs: long-term control with pasireotide Lar in two patients. Endocr Connect. 2019;8:367–77.
- 211. Chahal HS, Trivellin G, Leontiou CA, et al. Somatostatin analogs modulate AIP in somatotroph adenomas: the role of the ZAC1 pathway. J Clin Endocrinol Metab. 2012; https://doi. org/10.1210/jc.2012-1111.
- 212. Solomou A, Herincs M, Roncaroli F, Vignola ML, Gaston-Massuet CKM Investigating the role of AIP in mouse pituitary adenoma formation | SFEBES 2017.
- Villa C, Lagonigro MS, Magri F, et al. Hyperplasia-adenoma sequence in pituitary tumorigenesis related to aryl hydrocarbon receptor interacting protein gene mutation. Endocr Relat Cancer. 2011;18:347–56.
- 214. Vallar L, Spada A, Giannattasio G. Altered Gs and adenylate cyclase activity in human GH-secreting pituitary adenomas. Nature. 1987;330:566–8.
- 215. Bakhtiar Y, Hirano H, Arita K, et al. Relationship between cytokeratin staining patterns and clinico-pathological features in somatotropinomae. Eur J Endocrinol. 2010;163:531–9.
- 216. Mayr B, Buslei R, Theodoropoulou M, Stalla GK, Buchfelder M, Schöfl C. Molecular and functional properties of densely and sparsely granulated GH-producing pituitary adenomas. Eur J Endocrinol. 2013;169:391–400.
- 217. Chenlo M, Rodriguez-Gomez IA, Serramito R, et al. Unmasking a new prognostic marker and therapeutic target from the GDNF-RET/PIT1/p14ARF/p53 pathway in acromegaly. EBioMedicine. 2019;43:537–52.
- Hage C, Sabini E, Alsharhan H, Fahrner JA, Beckers A, Daly A, Salvatori R. Acromegaly in the setting of Tatton-Brown-Rahman Syndrome. Pituitary. 2019; https://doi.org/10.1007/ s11102-019-01019-w.
- Newell-Price J, Bertagna X, Grossman AB, Nieman LK. Cushing's syndrome. The Lancet, 2006;367(9522):1605–17. https://doi.org/10.1016/s0140-6736(06)68699-6).
- 220. Weber A, Trainer PJ, Grossman AB, Afshar F, Medbak S, Perry LA, Plowman PN, Rees LH, Besser GM, Savage MO. Investigation, management and therapeutic outcome in 12 cases of childhood and adolescent Cushing's syndrome. Clin Endocrinol. 1995;43:19–28.
- Magiakou A, Chrousos GP. Cushing's syndrome in children and adolescents: current diagnostic and therapeutic strategies. J Endocrinol Investig. 2002;25:181–94.
- Webb KM, Laurent JJ, Okonkwo DO, et al. Clinical characteristics of silent corticotrophic adenomas and creation of an internet-accessible database to facilitate their multi-institutional study. Neurosurgery. 2003;53:1076–85.
- 223. Nieman LK, Biller BMK, Findling JW, Murad MH, Newell-Price J, Savage MO, Tabarin A. Treatment of cushing's syndrome: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2015;100:2807–31.
- 224. Rix M, Hertel NT, Nielsen FC, Jacobsen BB, Hoejberg AS, Brixen K, Hangaard J, Kroustrup JP. Cushing's disease in childhood as the first manifestation of multiple endocrine neoplasia syndrome type 1. Eur J Endocrinol. 2004;151:709–15.

- Caimari F, Kumar AV, Kurzawinski T, Butler G, Sabbaghian N, Foulkes WD, Korbonits M. A novel DICER1 mutation in familial multinodular goitre. Clin Endocrinol. 2018;89:110–2.
- 226. Schultz KAP, Williams GM, Kamihara J, et al. Dicer1 and associated conditions: identification of at-risk individuals and recommended surveillance strategies. Clin Cancer Res. 2018;24:2251–61.
- 227. Apellaniz-Ruiz M, De Kock L, Sabbaghian N, Guaraldi F, Ghizzoni L, Beccuti G, Foulkes WD. Familial multinodular goiter and Sertoli-Leydig cell tumors associated with a large intragenic in-frame DICER1 deletion. Eur J Endocrinol. 2018;178:K11–9.
- 228. Cohen M, Persky R, Stegemann R, et al. Germline USP8 mutation associated with Pediatric Cushing disease and other clinical features: a new syndrome. J Clin Endocrinol Metab. 2019;104:4676–82.
- 229. Perez-Rivas LG, Theodoropoulou M, Ferraù F, et al. The gene of the ubiquitin-specific protease 8 is frequently mutated in adenomas causing cushing's disease. J Clin Endocrinol Metab. 2015;100:E997–E1004.
- 230. Faucz FR, Tirosh A, Tatsi C, et al. Somatic USP8 gene mutations are a common cause of pediatric Cushing disease. J Clin Endocrinol Metab. 2017;102:2836–43.
- 231. Hayashi K, Inoshita N, Kawaguchi K, et al. The USP8 mutational status may predict drug susceptibility in corticotroph adenomas of Cushing's disease. Eur J Endocrinol. 2016;174:213–26.
- 232. Roussel-Gervais A, Couture C, Langlais D, Takayasu S, Balsalobre A, Rueda BR, Zukerberg LR, Figarella-Branger D, Brue T, Drouin J. The Cables1 gene in glucocorticoid regulation of pituitary Corticotrope growth and Cushing disease. J Clin Endocrinol Metab. 2016;101:513–22.
- 233. Longuini VC, Lourenço DM, Sekiya T, et al. Association between the p27 rs2066827 variant and tumor multiplicity in patients harboring MEN1 germline mutations. Eur J Endocrinol. 2014;171:335–42.
- 234. Delgrange E, Vasiljevic A, Wierinckx A, François P, Jouanneau E, Raverot G, Trouillas J. Expression of estrogen receptor alpha is associated with prolactin pituitary tumor prognosis and supports the sex-related difference in tumor growth. Eur J Endocrinol. 2015;172:791–801.
- 235. Colao A, Di Sarno A, Cappabianca P, Briganti F, Pivonello R, Di Somma C, Faggiano A, Biondi B, Lombardi G. Gender diffrences in the prevalence, clinical features and response to cabergoline in hyperprolactinemia. Eur J Endocrinol. 2003;148:325–31.
- 236. Colao A. Pituitary tumours: the prolactinoma. Best Pract Res Clin Endocrinol Metab. 2009;23:575–96.
- 237. Araujo PB, Kasuki L, de Azeredo Lima CH, Ogino L, Camacho AHS, Chimelli L, Korbonits M, Gadelha MR. AIP mutations in brazilian patients with sporadic pituitary adenomas: a single-center evaluation. Endocr Connect. 2017;6:914–25.
- 238. Vandeva S, Tichomirowa MA, Zacharieva S, Daly AF, Beckers A. Genetic factors in the development of pituitary adenomas. Endocr Dev. 2009;17:121–33.
- Korbonits M, Kumar AV. AIP-related familial isolated pituitary adenomas. Seattle: University of Washington; 1993.
- 240. Xekouki P, Pacak K, Almeida M, et al. Succinate dehydrogenase (SDH) D subunit (SDHD) inactivation in a growth-hormone-producing pituitary tumor: a new association for SDH? J Clin Endocrinol Metab. 2012; https://doi.org/10.1210/jc.2011-1179.
- Newey PJ, Gorvin CM, Cleland SJ, et al. Mutant prolactin receptor and familial hyperprolactinemia. N Engl J Med. 2013;369:2012–20.
- Kobayashi T, Usui H, Tanaka H, Shozu M. Variant prolactin receptor in Agalactia and hyperprolactinemia. N Engl J Med. 2018;379:2230–6.
- 243. Schuff KG, Hentges ST, Kelly MA, Binart N, Kelly PA, Iuvone PM, Asa SL, Low MJ. Lack of prolactin receptor signaling in mice results in lactotroph proliferation and prolactinomas by dopamine-dependent and -independent mechanisms. J Clin Invest. 2002;110:973–81.
- 244. Gorvin CM, Newey PJ, Rogers A, et al. Association of prolactin receptor (PRLR) variants with prolactinomas. Hum Mol Genet. 2019;28:1023–37.

- 245. Bernard V, Bouilly J, Beau I, Broutin I, Chanson P, Young J, Binart N. Germline prolactin receptor mutation is not a major cause of sporadic prolactinoma in humans. Neuroendocrinology. 2016;103:738–45.
- Beck-Peccoz P, Giavoli C, Lania A. A 2019 update on TSH-secreting pituitary adenomas. J Endocrinol Investig. 2019;42:1401–6.
- 247. Önnestam L, Berinder K, Burman P, Dahlqvist P, Engström BE, Wahlberg J, Nyström HF. National incidence and prevalence of TSH-secreting pituitary adenomas in Sweden. J Clin Endocrinol Metab. 2013;98:626–35.
- 248. Scheithauer BW, Kovacs K, Nose V, Lombardero M, Osamura YR, Lloyd RV, Horvath E, Pagenstecher A, Bohl JE, Tews DS. Multiple endocrine neoplasia type 1-associated thyrotropin-producing pituitary carcinoma: report of a probable de novo example. Hum Pathol. 2009;40:270–8.
- Beck-Peccoz P, Lania A, Beckers A, Chatterjee K, Wemeau J-L. 2013 European Thyroid Association guidelines for the diagnosis and treatment of thyrotropin-secreting pituitary Tumors. Eur Thyroid J. 2013;2:76–82.
- Greenman Y, Cooper O, Yaish I, et al. Treatment of clinically nonfunctioning pituitary adenomas with dopamine agonists. Eur J Endocrinol. 2016;175:63–72.
- Rodd C, Millette M, Iacovazzo D, et al. Somatic GPR101 duplication causing X-linked acrogigantism (XLAG)—diagnosis and management. J Clin Endocrinol Metab. 2016;101:1927–30.



Acromegaly

5

Luigi Maione and Philippe Chanson

5.1 Introduction

Acromegaly is a typical rare multisystemic disease associated with progressive enlargement of some parts of the body, featuring somatic disfigurement mainly involving the face and extremities. The archetypal body changes and gigantism are renowned features (Figs. 5.1 and 5.2). Apart from body changes and disproportion, a number of multiorgan comorbidities are frequently associated with acromegaly. Most clinical repercussions originate from growth hormone (GH) and insulin-like growth factor-1 (IGF-I)-dependent organ overgrowth. In most cases, GH excess is due to a pituitary adenoma. Acromegaly itself and the related comorbidities might lead to premature death if not adequately treated.

5.2 Epidemiology

The prevalence of acromegaly has recently been estimated to be approximately 28 to 137 cases per million inhabitants [3, 4], challenging the historical figure of 40 to 70 cases per million [5]. A prevalence of approximately 1000 per million inhabitants was found in a German study based on screening with IGF-I measurement in

Université Paris-Saclay, Inserm, Physiologie et Physiopathologie Endocriniennes, Assistance Publique-Hôpitaux de Paris, Hôpital Bicêtre, Service d'Endocrinologie et des Maladies de la Reproduction, Centre de Référence des Maladies Rares de l'Hypophyse, Le Kremlin-Bicêtre, France

The chapter has been endorsed by **Prof. Vera Popovic**, popver@gmail.com, University of Belgrade, Belgrade, Serbia

L. Maione \cdot P. Chanson (\boxtimes)

e-mail: luigi.maione@aphp.fr; philippe.chanson@aphp.fr

[©] Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_5



Fig. 5.1 Schematic representation of signs and symptoms of acromegaly. Adapted from [1], with permission



Fig. 5.2 Disease evolution in a woman with acromegaly. Series of photographs taken over time showing the progressive changes in facial appearance. It is possible to presume that the first signs appeared between 1988 and 1990, 22 years before the diagnosis. In 1990, the patient had to enlarge her ring she bought 2 years before because it was too narrow (adapted from [2], with permission)

the general population [6]. Raappana estimated the annual incidence of acromegaly at 3.4 cases per million in Finland [7]. A recent survey analyzing all the studies providing epidemiological data refined the incidence to a range between 0.2 and 1.1 cases/100,000 [8].

Age at diagnosis typically falls within the fourth decade of life, following a quasi-Gaussian curve [9, 10]. However, although more rarely, acromegaly might be found in children and in the elderly.

The sex ratio has been found to be more or less constant across studies. A female to male ratio of 1.26 has been calculated by analyzing data concerning more than 16,000 patients across national registries [11]. Age at diagnosis is typically earlier in males than in females, and a clearly distinguishable sex-related dimorphic Gaussian curve is observed [9, 12, 13].

Owing to the insidious clinical onset and slow progression, acromegaly is often diagnosed late. Older series, in the 1980s, suggested a mean diagnostic latency of 3–10 years after onset, at an average age of approximately 40 years [8, 14–17].

Studies focusing on disease latency seem to show that diagnostic delay appears to be more or less constant throughout decades, without any improvement over time with an earlier diagnosis [9, 18].

5.3 Pathogenesis

In more than 95% of cases, acromegaly is secondary to GH hypersecretion by a benign monoclonal pituitary adenoma that develops from somatotroph cells [19–22]. Pituitary somatotroph adenomas are mostly isolated (sporadic). Rarely, they may develop in the frame of a genetic predisposing disease (Fig. 5.3a).

5.3.1 Somatotroph Pituitary Adenoma

Pure somatotroph pituitary adenomas (60%) are constituted by eosinophilic cells containing either densely or sparsely granulated (secretory granules) cell elements after immunolabeling [19]. In some adenomas, immunostaining reveals colocalization of free alpha-subunits [24]. Silent somatotroph adenomas do not determine clinical acromegaly. Diagnosis is almost exclusively made by tumor immunostaining. Nonetheless, some patients bearing these tumors may have supranormal circulating GH levels without overt clinical signs [25].

Most human somatotroph adenomas seem to be associated with the clonal expansion [26] of cells carrying a specific somatic mutation. However, as in other types of pituitary adenoma, it has proven difficult to isolate a single causative factor explaining pituitary tumorigenesis [27–29]. Mutations in stimulating G-protein [30, 295] have been identified in 35–55% of somatotroph adenomas, according to some series [30–32]. *gsp* mutations are able to inhibit GTPase activity and to lead to constitutive adenyl-cyclase activation [33]. Cell cycle disruption also seems to play an important role, as demonstrated in MEN1 or in patients with *CDKN1B* mutations (coding the



Fig. 5.3 Prevalence of somatotroph tumor forms and associated clinical genetic diseases. Panel **a**: Schematic representing the relative prevalence of sporadic pure or mixed somatotroph adenomas and associated mosaic/germline genetic diseases. Panel **b**: Characteristic clinical features of a patient with McCune Albright syndrome. Please note the typical café-au lait skin spots and X-rays and MRI hallmarks illustrating fibrous bone dysplasia. Panel **c**: Characteristic clinical features in a patient with acromegaly with multiple endocrine neoplasia type-1. Please note the pituitary tumor on a MRI T1W coronal post-gadolinium view, a pancreatic tumor revealed by echoendoscopy, and parathyroid hyperplasia/adenoma found on ^{99m}Tc -¹²³I subtraction parathyroid scintigraphy. Panel **d**: Characteristics of a patient with Carney complex, from [23], with permission. Panel **e**: Growth curve of a patient affected by X-LAG acro-gigantism

cyclin-dependent kinase inhibitor p27KIP1, a key regulator of the cell cycle) [29]. A number of other genes have been implicated in somatotroph tumorigenesis [29]. The function of the disrupted proteins deriving from mutated genes spans from oncogene/tumor suppression to cyclin/cell proliferation inhibition. The general belief is that a preexisting mutation within somatotrophs should be a predisposing factor for further cell proliferation and GH secretion. Premature senescence likely explains the persistence of a benign phenotype and the rarity of progression to carcinoma. Epigenetic mechanisms may also contribute to cell proliferation by silencing genes such as *CDKN2A*, encoding p16, a cell proliferation inhibitor. The sequence of events leading to somatotroph cell clonal expansion seems to be multifactorial [34, 35].

Cytogenetic studies show that somatotroph pituitary adenomas display substantial intertumor and intratumor DNA copy-number heterogeneity. Intriguingly, somatic *GNAS*-mutated adenomas have low copy number variations, whereas a higher heterogeneity is observed in *GNAS*-intact tumors [36].

5.3.2 Mixed Somatotroph Adenoma

The most frequent mixed adenomas coexpress GH and prolactin (PRL), accounting for 25% of cases. Histopathologists often distinguish between true mixed adenomas, containing either somatotroph or lactotroph cell types, and lactosomatotroph stem cells consisting of more mature monomorphic cells coexpressing GH and PRL [19]. Mixed GH- and TSH-secreting adenomas are rarer and are associated with acromegaly associated with hyperthyroidism by inappropriate secretion of thyroid-stimulating hormone [37, 38]. ACTH cosecretion in somatotroph adenomas is extremely rare.

5.3.3 Genetic Syndromes Associated with Acromegaly

Various genetic syndromes and diseases include acromegaly in a wider spectrum of clinical features. Despite representing a rare cause, the burden of related morbidities requires not neglecting them (Fig. 5.3a–e). This paragraph summarizes the main features of these diseases along with some peculiarities in acromegaly manifestation. In general terms, a germinal genetic disorder should be suspected in patients with early-onset acromegaly bearing other organ involvement. These diseases have also been reviewed in recent articles and book chapters [20, 39–42].

McCune-Albright syndrome (MAS) is a rare genetic disease associated with multiple fibrous bone dysplastic lesions, precocious puberty, "café-au-lait" spots, and multiorgan and soft-tissue tumors (Fig. 5.3b). Pathological features are related to postzygotic somatic mutations leading to constitutive activation of the Gs protein alpha subunit [43–45]. It is of note that this gene is also responsible for most sporadic pure GH adenomas, underlying the importance of stimulating G-protein in the somatotroph environment. In the case of diffuse germinal mutations (MAS), acromegaly is found in approximately 20% of patients [43, 44, 46]. A peculiarity of this form is a relative resistance to somatostatin analogs (approximately 30% of responders). The therapeutic approach is also markedly influenced by the coexistence of skull base dysplasia, making any neurosurgical approach challenging [45].

Acromegaly can also be associated with hyperparathyroidism, neuroendocrine tumors (e.g., gastrinoma, insulinoma, or a nonfunctional pancreatic tumor), adrenal and other endocrine or nonendocrine tumors in the frame of **multiple endocrine neoplasia** (**MEN**) **type 1**, which is related to menin (*MEN1*) germline mutations (Fig. 5.3c) [47, 48]. Pituitary adenomas are not enriched in the somatotroph lineage in MEN1 patients. Few cases of GHRH-secreting neuroendocrine tumors have been reported in patients with MEN1 (see the "Extrapituitary acromegaly" paragraph) [49, 50].

Mutations in the *CDKN1B* gene are responsible for a rarer and newer MEN syndrome, **multiple endocrine neoplasia type 4** (MEN4, initially known as MEN-X), which combines hyperparathyroidism, pituitary adenomas (including acromegaly), and other endocrine or nonendocrine tumors [51, 52]. Twenty cases have been published to date. Hyperparathyroidism seems to be the most prevalent disease. Nonetheless, acromegaly by a somatotroph adenoma is found in up to 20% of patients harboring a deleterious *CDKN1B* mutation. Furthermore, contrary to MEN1, pituitary adenomas in MEN4 seem to be more enriched in somatotroph adenomas.

When acromegaly is associated with bilateral pigmented micronodular adrenal hyperplasia (causing ACTH-independent hypercortisolism) or with typical cutaneous pigmentations or cardiac myxomas, the patient should be screened for the **Carney complex** (Fig. 5.3D). This genetic disease is related to a germline mutation of the regulatory subunit of protein kinase A (*PRKAR1A*), whose signaling cascade is also located downstream of the stimulating G-protein [53, 54]. A simplified schematic of the cyclic AMP-dependent signaling pathway with relevant targets for somatotroph tumorigenesis is provided in Fig. 5.4.

Acromegaly is also one of the features described in **familial isolated pituitary adenoma**, partly related to *AIP* germline mutations (aryl hydrocarbon receptor interacting protein) [55–57]. These mutations can also, albeit rarely, be found in some apparently sporadic cases of acromegaly, particularly in young patients [58–62].

GPR101 was the latest gene to be discovered in association with a very earlyonset form of **X-linked gigantism** or **X-LAG syndrome** [63]. Affected patients develop large or giant adenomas at a very young age. The pathophysiology is related to Xq26.3 microduplication involving the orphan G-protein-coupled receptor GPR101. However, germline GPR101 mutations are very rare in patients with sporadic pituitary adenomas, particularly in patients with gigantism or acromegaly [64]. Little is known about the pathophysiology and sequence of events leading to somatotroph tumorigenesis (Fig. 5.3E).

5.3.4 GH-Secreting Carcinomas

Somatotroph carcinomas are exceptional (fewer than 20 published cases). The presence of distant metastases is required to support the diagnosis of malignancy [39].

5.3.5 Extrapituitary Acromegaly

Extrapituitary acromegaly refers to growth hormone releasing hormone (GHRH) or GH hypersecretion other than from a pituitary adenoma [39].

GHRH hypersecretion could originate either from hypothalamic tumors, such as gangliocytomas, hamartomas, choristomas, and gliomas, or from the periphery. More often, GHRH hypersecretion comes from ectopic sources. The GHRH peptide



Fig. 5.4 The cyclic AMP-dependent signaling pathway in pituitary somatotroph cells as a model to understand different disease forms. GHRH induces a conformational change in the class II G protein-coupled receptor GHRHR. The Gs- α subunit exchanges GDP for GTP, which activates adenylyl cyclase (AC), converting ATP to cAMP. Elevated cAMP levels activate protein kinase A (PKA). PKA consists of a tetramer of two homo- or heterodimer regulatory subunits (R) and two catalytic subunits (C) responsible for the phosphorylation of several enzymes and transcription factors downstream [e.g., cAMP-response element-binding protein (CREB)]. *MCAS* McCune-Albright syndrome, *LOH* loss of heterozygosity, *MEN1* multiple endocrine neoplasia type-1, α , β , γ Gs-protein subunits, *AC* adenyl cyclase, *cAMP* cyclic AMP, *CREB* cyclic AMP response element-binding protein, a transcription factor, *PKA* protein kinase A, *R* the regulatory subunits of protein kinase A, *C* the catalytic subunit of protein kinase A

was indeed originally identified and cloned from large pancreatic tumors [65]. GHRH is expressed by several tissues. However, very large amounts are needed to induce pituitary somatotroph hyperplasia and clinical acromegaly. GHRH hypersecretion may derive from pancreatic cell tumors, small-cell lung cancers, bronchial and other-site carcinoids, adrenal adenomas, and pheochromocytomas. The delivery of high GHRH concentrations stimulates normal pituitary somatotrophs to become hyperplastic and to hypersecrete GH to produce acromegaly [50, 66, 67]. The diagnosis is established by measuring plasma GHRH and by identifying the source (a GHRH-staining neuroendocrine tumor) [68]. The prognosis largely depends on the characteristics of the underlying tumor [50].

GH can also be directly secreted by an ectopic somatotroph adenoma (located near the *sella turcica*, for example, in the sphenoidal sinus, petrous temporal bone,

nasopharyngeal cavity) or, in exceptional cases, by a peripheral tumor (pancreatic islet tumor or lymphoma) [69, 70].

5.4 Clinical Presentation

Acromegaly is generally suspected based on clinical signs and symptoms, which are important to recognize (Fig. 5.1) [2, 16, 17, 71–74].

5.4.1 The Dysmorphic Syndrome

In typical forms, patients present broadened extremities (hands and feet), widened, thickened and stubby fingers, and soft tissue thickening. When specifically asked, affected patients describe enlarged rings over the last years or the need to change shoe size. The facial aspect is somehow characteristic and includes a widened and thickened nose, prominent cheekbones, bulged forehead, thick lips, and marked facial lines (Figs. 5.1 and 5.2). The forehead and the overlying skin are thickened, sometimes leading to frontal bossing. The lower face is also affected, with several degrees of prognathism, maxillary widening, teeth separation, and, more rarely, jaw occlusion impairment. A useful step is to analyze comparatively ancient photographs. This could show a slow, insidious demarcation of the acrofacial syndrome spreading over several years (Fig. 5.2). Because of this slow progression, relatives and physicians may be unaware, and acromegaly may be diagnosed very late. Typically, after variable latency, the diagnosis is raised by a physician who has not seen the patient before [16, 17, 75]. A recent multicenter survey investigating medical practices of more than 3000 patients with acromegaly across several European countries reported that most diagnoses were reportedly made by an endocrinologist (45%), followed by general practitioner (17.5%), internist (13.2%), orthopedist (3.6%), neurologist (3.3%), ophthalmologist (2.3%), and the patient him or herself or one of their relatives (2.3%) [10]. It is therefore not uncommon that a patient could make the diagnosis him or herself by searching the Internet. In most cases, diagnosis is made owing to changes in the face or extremities; in some other cases, acromegaly is diagnosed not for its signs and symptoms but during the biochemical exploration of a pituitary adenoma.

5.4.2 Symptoms

Acromegaly can cause a broad variety of symptoms [16].

5.4.2.1 Skin Changes

Nearly 70% of patients have sweaty and oily skin. Skin thickening is due to glycosaminoglycan deposition and to increased collagen production by connective tissue. These changes may lead to hyperhidrosis and malodorous sweating. Facial
wrinkles, nasolabial folds, and heel pads are increased in thickness, and body hair may become coarsened. Skin tags are frequent and may be a marker of colonic polyps. Raynaud's disease is present in one-third of cases. In some cases, patients describe night-time malodorous sweating.

5.4.2.2 Bone Changes

In response to both GH and IGF-I, new periosteal bone formation leads to an increase in skeletal growth, especially at the level of the mandible (prognathism). Jaw thickening, tooth separation, frontal bossing, malocclusion, and nasal bone hypertrophy are the standard facial bony deformities in acromegaly.

Radiography shows a thickening of the cranial vault and protuberances, frontal internal hyperostosis, and condensation of the *sella turcica* walls with clinoid hypertrophy. Hypertrophy of the sinuses, especially the frontal sinuses, is also clearly visible. This, along with laryngeal hypertrophy, may explain why the voice tends to become deeper and acquires a sonorous resonance [76].

These changes are not only due to soft tissue hypertrophy and excessive growth of bone and cartilage but also to bone deformation. Indeed, radiographic findings are abnormal in half of these patients, showing distal tufting of the phalanges, widening of the base of the phalanges with osteophyte formation, enthesopathy (mineralization of ligamentous insertions), widening of the cortical bone diaphyses, and widening of joint spaces due to cartilage hypertrophy. Deformations can also affect the rest of the skeleton, and dorsal kyphosis with distortion of the rib cage may be observed in severe chronic forms, leading to the paradigmatic "punchinello" aspect, especially when GH hypersecretion begins prior to epiphysis closure.

Bony deformations also affect the spine, with upper dorsal kyphosis and compensatory lumbar hyperlordosis. Vertebral enlargement, widened intervertebral spaces, and osteophyte formation were also observed. The thorax is deformed by protuberance of the lower portion of the sternum and by elongation and divergence of the ribs due to overgrowth of the chondrocostal joints.

Imaging studies show diaphyseal cortical thickening of the long bones and widened joint spaces, sometimes with osteophytes.

Concerning mineral changes, bone remodeling is increased in acromegaly [77, 78]. Cortical bone thickens (as measured by the metacarpal index and histomorphometric parameters) and its porosity is diminished. Trabecular bone mass may be decreased, normal or increased. Measurement of spinal bone mass can give contradictory results, probably because acromegaly is often associated with other endocrine disorders interfering with bone mass. In general, bone mass is normal in the lumbar spine in patients with isolated acromegaly but could be decreased in patients having other related endocrinopathies impacting bone metabolism, such as hypogonadism or hyperparathyroidism. Despite similar bone mineral density values, a lower lumbar spine assessed by the trabecular bone score (TBS) technique was demonstrated in patients with acromegaly compared to controls, especially in hypogonadal patients and women [79]. In a study [80] independent of BMD, the prevalence of vertebral fractures was found to be higher in patients with acromegaly (57.5% vs 22.6%). Fractures were associated with higher serum IGF-I values, a longer duration of active disease, and a longer history of untreated hypogonadism. This higher prevalence of vertebral fractures persists despite biochemical control of acromegaly [81].

5.4.2.3 Rheumatologic Comorbidity

Peripheral Osteoarthritis

The topic has been reviewed in detail in [82, 83].

Peripheral joint symptoms are very frequent. Arthralgia and myalgia occur in 30–70% of patients. Among the sites, large joints such as the knees, shoulders, hands, wrists, and hips seem to be more affected. Acromegalic osteoarthritis develops within an average of 10 years after diagnosis.

Osteoarthritis develops in two stages. In the initial stage, the growth of joint cartilage and periarticular ligaments is stimulated, leading to enlargement and congestion of the interarticular space that limits mobility and induces joint pain; in the second stage, more degenerative changes of the joint geometry are observed: intra-articular microtrauma and exuberant repair reactions induce scars and subchondral resorption and osteophytotic development, leading to progressive joint deterioration. The pain is thus mainly mechanical, degenerative, and noninflammatory and often persists after treatment of acromegaly. However, rarely, some patients may present symptoms and signs of pseudoinflammatory osteoarthritis, which are dramatically relieved by the treatment of acromegaly. Joint mobility (especially of the shoulders) can be limited in the later stages of the disease. Joint effusion is rare, and synovial aspiration shows a generally degenerative picture with no evidence of inflammation; it may also reveal calcium microcrystals (associated chrondrocalcinosis).

Physical examination of the joints often provides little information. The abnormalities are generally minor despite subjective functional discomfort. The shoulders and hips may show a loss of mobility and function. In contrast, some patients have joint hyperlaxity. There was no correlation between the presence or severity of osteoarthropathy and the age of onset of acromegaly or the mean GH or IGF-I concentration at baseline or during follow-up. Osteoarthritis appears to be more frequent after 45 years of age.

Radiological studies show a widening of the joint spaces, reflecting hypertrophy of the hyaline cartilage, as well as the presence of osteophytes, bone proliferation at the attachment sites of tendons and ligaments, periarticular calcium deposition, and exostosis of the bone. The joint space subsequently diminishes due to destructive arthritis. Sonography shows a thickening of the cartilage in the shoulder, wrist, and knee joints, which is improved with treatment for acromegaly.

Osteoarthritis inexorably progresses in advanced stages of the disease. It is not influenced by successful treatment of acromegaly, with the exception of diffuse articular symptoms and some sites of pain [84]. Acromegalic osteoarthritis considerably impairs patients' quality of life [85–87].

Spinal Involvement

The estimated prevalence of spinal involvement is approximately 40–50% [88]. Backache is more frequent at the level of the lumbar spine than the cervical or dorsal segments. The pain is mainly mechanical in nature, but inflammatory features can occur in later stages (16%). Spinal involvement may be accompanied by nerve compression. Occasionally, bilateral intermittent claudication reveals lumbar spinal stenosis. Pain may also be related to an increased prevalence of vertebral fractures despite normal BMD [80, 89].

Radiological examination shows typical features, including ossification of the anterior and lateral surfaces of the vertebral bodies, contributing to enlargement of their anteroposterior diameter, as well as a biconcave vertebral aspect and scalloping of the vertebral bodies (exaggerated concavity of the posterior vertebral wall). The mechanism of these morphometric changes is poorly understood but may involve hypertrophy of the intraspinal soft tissues (ligamentous hypertrophy, epidural lipomatosis) or bone. In more severe cases, ossification of the anterior surface of the vertebral bodies can bridge the disk space and give an aspect of diffuse idiopathic skeletal hyperostosis. An increased number of vertebral fractures with wedge deformity and thoracic kyphosis is also more prevalent in patients with acromegaly than in the general population [90].

5.4.2.4 Neuropathies

Symptomatic carpal tunnel syndrome is frequent. Nerve conduction studies have shown that the vast majority of patients with acromegaly have subclinical abnormalities of nerve conduction. Magnetic resonance imaging (MRI) shows a higher amplitude and intensity of the median nerve signal in patients with symptomatic carpal tunnel syndrome compared to asymptomatic patients [91]. The mechanism appears to involve median nerve edema more than extrinsic compression due to an excess of connective tissue, bone or synovial hypertrophy or an increase in extracellular fluid within the carpal tunnel itself with Schwann cell demyelination. Nerve edema, which can also easily be evaluated with ultrasonography [92], improves when GH and IGF-I levels fall, suggesting that hormonal control is a key prerequisite for improving these patients' neurological status. Sometimes, however, carpal tunnel syndrome may persist.

Ulnar nerve neuropathy at the cubital tunnel is also frequent in patients with acromegaly [93] and improves with treatment of acromegaly.

Apart from mechanical/compressive effects on nerves, autonomic nervous system dysfunction is present in patients with acromegaly, as shown by assessing the heart rate variability indices (mean sinus heart rate, RR intervals) and reverses after effective treatment [94].

5.4.2.5 Psychologic Consequences

Self-esteem may diminish along with progressive facial and bodily disfigurement. Patients with acromegaly further exhibit impairment in body image distortion, disruption in interpersonal relations, and social withdrawal anxiety [95].

Patients reported more negative illness perceptions than patients with acute illness but more positive illness perceptions than patients with other chronic diseases [96].

Nonetheless, direct unstructured interviews reveal an association between the diagnostic delay and the doctor-patient encounter and the experience of this disease, which is often described as catastrophic, both before and after the diagnosis [97].

It is unclear whether reported depression, mood swings, and apathy result solely from these physical changes or whether they are intrinsic high GH exposure central effects.

Acromegaly carries a significant lifelong burden for the affected patient. When evaluating health-related quality of life by means of dedicated questionnaires (such as the ACROQoL), it is clear that values barely normalize despite disease remission or cure [98]. All the biological, environmental, and biopsychosocial aspects of this burden have been extensively covered in a recent review by Biermasz [99].

5.4.2.6 Cardiovascular Manifestations

Arterial Hypertension

Hypertension occurs in 20–50% of patients. Its prevalence increases with time after the onset of acromegaly, as well as with GH level and age. Several concomitant factors are likely to play a role in the pathogenesis of hypertension in acromegaly. The chronic expansion of extracellular fluid volume (hypervolemia) leads to fluid retention, with plasma volume being 10–40% above normal. At the kidney level, increased renal sodium reabsorption at the distal tubule level is generally observed [100]. Body fluid expansion is related to enhanced epithelial sodium channel (ENaC) activity [1, 101, 102]. Hypertension can also result from endothelial dysfunction [103]. Neither renin-angiotensin aldosterone nor the sympathetic system appears to be involved in the pathogenesis of hypertension in this setting. Other contributors to the onset and maintenance of hypertension in acromegaly are also the increase in peripheral vascular resistance, insulin resistance and diabetes, and the development of obstructive sleep apnea [104–106].

Cardiomyopathy

Cardiac disorders are a consistent feature. Many lines of evidence, especially from experimental studies, point to the existence of specific cardiac disorders in acromegaly that are completely independent of coronary involvement (currently found in only a minority of patients) or valve disorders, diabetes, or hypertension [107, 108].

The first step of acromegaly-related cardiomyopathy mainly consists of myocardial hypertrophy of the interventricular septum and left ventricular posterior wall. This condition is initially asymptomatic, at least at rest. The assessment of initial cardiomyopathy is generally performed by means of cardiac ultrasound examination or magnetic resonance imaging (MRI). Generally, left ventricle parameters are normal (concentric hypertrophy). Myocardial hypertrophy can occur in the absence of hypertension and even in young patients (<30 years), reflecting the impact of GH excess itself on the myocardium. Its prevalence is likely to be overestimated by echocardiography compared to MRI [109]. Hypertension further aggravates cardiac hypertrophy. Echocardiography and isotope studies show altered diastolic function (abnormal left and right ventricle filling) related to abnormal relaxation: parietal stiffness is, at least in part, probably linked to edematous infiltration of the ventricular wall [110] and perhaps also to a certain degree of fibrosis. Clinical symptoms such as dyspnea during exercise may be observed in patients who are asymptomatic at rest. Systolic function is normal if assessed by conventional methods. However, novel techniques such as two- or three-dimensional speckle-tracking echocardiography reveal, even at an earlier stage, an increased frequency of subclinical systolic impairment in active acromegaly [111, 112]. At later stages, hyperkinetic syndrome (increased cardiac index) is frequent.

Arrhythmias and/or conduction disorders may occur at any stage of acromegalic cardiomyopathy [113]. Their prevalence has long been underestimated in these patients. Ventricular premature complexes have been shown to frequently occur in patients with acromegaly. In one study, systematic 24 h Holter ECG recordings showed complex ventricular arrhythmias in 48% of patients compared to only 12% of controls [114]. Most of these arrhythmias are subclinical and persist despite successful treatment of acromegaly. Myocardial remodeling, hypertrophy, and fibrosis are all likely to play a role in their onset. However, recent studies did not confirm the high prevalence of dysrhythmias [115].

Congestive heart failure can occur if the cardiac disorders progress (if GH hypersecretion persists and, probably, if other risk factors such as diabetes, hypertension, and sleep apnea are also present). Functional signs first appear on effort before becoming permanent. At this stage, echocardiography shows variable degrees of cavity dilation. Fortunately, these severe forms are now far less frequent (prevalence 3%) [116].

A number of cardiovascular parameters improve during effective treatment of acromegaly, even if some changes appear to be irreversible in certain patients. In general, younger patients and patients with a relatively short history of acromegaly show better "recovery" (from diastolic disorders, myocardial hypertrophy, or systolic dysfunction) [71, 117]. In contrast, when dilated congestive heart failure occurs, cardiac function (especially systolic function) may show a short-term improvement [100], allowing some patients to survive or to avoid heart transplantation, but the longer-term prognosis is worse than that of patients with heart failure due to other causes (5-year mortality rate 37%) [116].

There is controversy surrounding the cardiovascular (ischemic) risk carried by patients with acromegaly [118]. An increased prevalence of hypertension, a history of diabetes mellitus and decreased levels of high-density lipoprotein, low-density lipoprotein, and total cholesterol were found in patients with acromegaly, leading to significantly higher Framingham risk scores than in controls [119]. Biomarkers of cardiovascular disease were also found to be altered in another study [120].

However, carotid atherosclerosis and carotid internal media thickening are not more extensive in patients with acromegaly than in nonacromegalic subjects [121, 122]. Importantly, no increase in the prevalence of coronary artery disease (assessed by different means, such as cardiovascular events, calcium scores, or myocardial scintigraphy) is found in patients with newly diagnosed acromegaly compared to the general population [123–127]. The reason for this apparent discordance between observed and expected coronary events is currently unclear. It has been suggested that the known atherogenic effects of hypertension, insulin resistance, and diabetes induced by GH excess are counterbalanced by some other cardioprotective factors, such as decreased endothelial and systemic inflammation [128–130].

Cardiac Valve Disease

Cardiac valve disorders are highly prevalent in patients with acromegaly and can, along with other cardiac abnormalities, also contribute to the onset or aggravation of heart disease in patients with acromegaly [131]. The risk of valve disease increases with time since onset [132]. Acromegaly-related cardiac valve abnormalities, which may be related to fibrotic changes, seem to persist after effective treatment of acromegaly [133]. Furthermore, no cabergoline-induced cardiac valve remodeling was observed.

5.4.2.7 Metabolic Complications

Physiologically, GH increases blood glucose levels, exerts a lipolytic effect, and promotes triglyceride hydrolysis into free fatty acids and glycerol.

GH excess leads to insulin resistance at the level of the liver or in the periphery, leading to fasting and stimulated hyperinsulinemia. The prevalence of type-2 diabetes mellitus (T2DM) in acromegalic patients is more or less constant across studies and ranges from 20% to 56%, depending on the series [71]. The weighted mean T2DM prevalence in individuals with acromegaly is approximately 27% when comparing data from 14 national registries [11]. As long as the compensatory increase in insulin secretion by pancreatic β cells counterbalances the reduction in insulin sensitivity, glucose tolerance remains normal. Impaired glucose tolerance occurs when insulin secretion is altered, followed by the onset of diabetes [134].

Acromegaly is associated with a decrease in fat mass (both visceral and subcutaneous) but an increase in intermuscular fat mass (which may contribute to insulin resistance) and lean body mass [135, 136]. A recent study found an increase in exercise-induced myokine irisin circulating levels in patients with acromegaly [137]. This increase was independent of the disease status. The consequences on either glucose metabolism or thermogenesis of these findings still need to be demonstrated.

Alterations in lipid metabolism are reported in 30–40% of patients with acromegaly. In uncontrolled disease, a typical lipid profile is found, characterized by increased levels of lipoprotein(a) and triglycerides and decreased levels of HDL cholesterol [138]. The course of lipid parameters (and other cardiovascular risk factors) may vary with the treatment modality after therapeutic control of acromegaly [139]. Hypercalciuria is frequent in patients with acromegaly and may be associated with an increased incidence of nephrolithiasis. It is related to an IGF-I-mediated and PTHindependent increase in calcitriol synthesis, which is responsible for both absorptive hypercalciuria and increased fasting plasma calcium linked to enhanced distal tubular calcium resorption [1, 140]. An increased prevalence of hyperparathyroidism is also observed in patients with acromegaly, either in the context of multiple endocrine neoplasia (see the above "5.3.3 Genetic Syndromes Associated with Acromegaly" section) or independently (phenocopy) as a usual sporadic hyperparathyroidism.

5.4.2.8 Respiratory Complications

Sleep apnea affects 60–80% of all patients with acromegaly at the diagnosis of acromegaly. Men seem to be affected more than women [141]. Sleep apnea is more likely to be sought in patients who snore (reported by 78% of patients with acromegaly) and in those with daytime sleepiness (51%) or morning fatigue and morning headache (16%). Sleep apnea may be a contributory factor in hypertension, cardiovascular disease, and even cognitive decline. In most cases, apnea is obstructive, but one-third of patients have central apnea. Obstructive apnea is linked to anatomical changes due to mandibular and maxillary growth, soft-tissue thickening (especially of the palate and uvula), and changes in the angles of the different bone segments, leading to hypercollapsibility of the posterior and lateral hypopharyngeal walls. Hypertrophy of the tongue also plays a role [142], as does hypertrophy of the submaxillary glands.

Changes in respiratory function are frequent but less well documented. Anatomical modifications of thoracic bones and cartilage (leading to profound changes in the geometry of the rib cage) and mechanical changes in thoracic elasticity and the inspiratory muscles can lead to ventilatory disorders. Respiratory muscle strength is also abnormal. Altered mechanical and energetic properties of some upper airway dilator muscles have recently been demonstrated [143]. The inspiratory time is shorter, and the breathing frequency may increase.

Patients with acromegaly often have an increase in their total lung capacity (81% of men and 56% of women), owing to an increase in alveolar volume. An obstruction is found in 20–30% of patients (small airway or upper airway narrowing). Subclinical hypoxemia may be present. No ventilation-perfusion mismatching has been demonstrated.

The apnea-hypopnea index improves during effective treatment of acromegaly, along with the obstructive apnea index and oximetry values [141, 142, 144]. However, while apnea can disappear in some patients whose acromegaly is cured, it may persist or even worsen (likely due at least in part to associated obesity [145]) in others who thus require nocturnal positive end expiratory pressure. The reevaluation of sleep apnea is thus useful even if patients are cured or well controlled after acromegaly treatment.

Vocal changes have been described in patients with acromegaly. A deepening of the voice and a low fundamental frequency are observed in the population with acromegaly [76]. Modifications of the laryngeal cords and muscles, as well as upper respiratory tract thickening, may be responsible for these findings. However, the clinical consequences and the phonetic handicap related to these changes are not currently known.

5.4.2.9 Pituitary and Sellar Mass Effects

Headache is a very common symptom. In contrast with nonfunctioning adenoma, headache may be present even in patients bearing a microadenoma, thus reflecting a multifactorial genesis other than a direct adenoma-related compressive/expansive effect. In large tumors (macroadenomas or giant adenomas), low visual acuity and visual field defects may be observed in cases of suprasellar progression. Compression of the normal pituitary may also lead to anterior pituitary deficiency, which must be explored clinically and biochemically. Diabetes insipidus is never associated with acromegaly, except after neurosurgery or in the context of pituitary apoplexy [146].

5.4.2.10 Neoplasia and Acromegaly

Through the GH- and IGF-I-related promotion of cellular proliferation and differentiation, neoplasm and cancer risks have always been a major issue when dealing with acromegaly. In vitro and in vivo studies have shown a direct effect of GH or IGF-I in mediating cell proliferation. Pharmacological blockade of these targets in some cases allowed tumor inhibition in cell and animal models [147]. Despite these data on molecular biology, the link between GH excess and cancer risk in acromegaly is still unclear [147]. Although cancer-related mortality varies across studies, it seems that an excess of cancer and related mortality is present in patients with acromegaly with uncontrolled disease [148]. An overall cancer prevalence of 10% (any type, any site) is found in national registries collecting real-life data [11]. There is also some controversy regarding the incidence of each individual cancer type in patients with acromegaly.

Figures of colorectal cancer relative risk compared with the general population, initially widely overestimated at 10-20, are probably only 2-3 as per novel estimates [149–153]. There are various potential biological mechanisms that could explain the increased risk of colonic cancer in acromegaly: direct effects of GH and/or IGF-I; hyperinsulinemia; increases in IGFBP-3, IGF-II, and IGFBP-2 levels; altered bile acid secretions and local immune response; increased large bowel length; and obesity [150, 153]. Some authors claim that epigenetic alterations predispose patients with acromegaly to cancer development [154]. As colonic cancer may be the consequence of colonic polyp degeneration, many studies have examined the prevalence of colon polyps in patients with acromegaly. Prospective studies show that up to 45% of patients with acromegaly have colonic polyps, which are adenomatous in 24% of cases [155] and can arise in all parts of the colon. The acromegaly-associated colonic lesions seem to exhibit some peculiarities, such as larger, multiple, and more dysplastic adenomatous polyps than in nonacromegaly patients [150]. There is no clear correlation between GH or IGF-I concentrations and the incidence of colonic polyps. Colonoscopy guidelines for patients with acromegaly are controversial. The British Society of Gastroenterology [156] recommends performing a colonoscopy in patients with acromegaly by the age of 40. A subsequent examination should depend on the findings at the original screening and on the disease activity: screening every 3 years

in patients with a previous adenoma or with elevated IGF-I and every 5–10 years in those without adenomatous/dysplastic polyps or those with only hyperplastic polyps. Some technical difficulties may be encountered in patients with acromegaly because of the increased colon length [153].

Goiter is found in a large proportion of patients with acromegaly. Thyroid nodules have been found in nearly 60–70% of patients [157]. Multinodular goiter is autonomous in 10–20% of patients, sometimes causing patent thyrotoxicosis. Although thyroid nodules are in most cases benign, the risk of thyroid cancer has been found to be higher than that in the general population (odds ratio, OR = 7.9, relative risk, RR = 7.6), with a prevalence of nearly 4%. These findings were confirmed by a recent Finnish study [158]. Nevertheless, contrary to colorectal cancer, most studies about thyroid cancer contain recruitment biases, and the real incidence of thyroid cancer in acromegaly is still a matter of debate [153, 157, 159]. As is the case for colonic cancer, a relative overestimation of thyroid cancer may arise because of increased physician awareness for these tumors, as well as the large use of ultrasonography during the screening of comorbidities in acromegaly.

Neoplasms of the breast, lung, prostate, skin, soft tissues, brain, bone, and lympho-hematopoietic system, initially described in association with acromegaly, do not seem to be overrepresented in these patients [160]. There is therefore remarkable agreement among all experts and reported guidelines, pointing out that surveillance in relation to these cancer sites should follow the same recommendations as for the general population [161].

It is currently acknowledged that, along with other cancers and neoplasms, the description of cancer occurrence is probably overestimated because of enhanced proactive screening. Modern imaging techniques may detect subclinical lesions and therefore affect the incidence rates. Other benign lesions may be found at a higher prevalence in patients with acromegaly. A higher incidence of meningioma has been found when analyzing encephalic MRI in patients with somatotroph adenomas versus those with other cell-type pituitary adenomas [162].

5.5 Diagnosis of Acromegaly

The diagnosis of acromegaly is suspected on clinical grounds and is confirmed by a typical biochemical profile [2, 74]. Clinical diagnosis is suggested by typical disfigurement due to progressive acral enlargement and modification of the facial appearance. In the case of very low progression or clinical incertitude, it is sometimes useful to assess the evolution by comparing serial photographs over several years (Fig. 5.2). Deep learning approaches are currently going to be tested to assist semiology [163]. It is of note that the regions of interest of these tools using aprioristic algorithms are primarily the same as those used by clinicians [163].

IGF-I (with reference to the age-adjusted normal range), the main GH-dependent growth factor, is the screening test recommended for acromegaly, with the diagnosis being confirmed by a nonsuppressive level of GH after an oral glucose load, OGTT [161].

5.5.1 GH and IGF-I

The introduction of international standards has minimized GH variability, which was mainly due to the use of polyclonal or monoclonal antibodies recognizing a mixture of different molecular forms. Manufacturers were recently advised to calibrate their GH assay kits with the international standard (IS) 98/574 [164].

The latest assays allow the limit of quantification to be as low as 0.05 μ g/l with an interassay coefficient of variation (CV) of <20% [164, 165].

In most cases, GH levels are elevated, both at baseline and after OGTT [166]. GH levels in the population with acromegaly are inversely correlated with age, in which the youngest patients have the most elevated serum GH concentrations, and with the maximal tumor diameter [10]. Previous recommendations consider a diagnosis of acromegaly if nadir GH levels are above $1 \mu g/l$ [74, 167]. However, a few patients with clear clinical signs of acromegaly and high IGF-I levels could have low GH output and can thus suppress GH levels to less than 1 µg/l during the OGTT. Thus, a more stringent criterion of a nadir GH at 0.4 µg/l after OGTT has been proposed [168] and is now increasingly recognized as the recommended threshold if a sensitive GH assay is used. This is in line with recent normative data in healthy subjects underlining the importance of sex, BMI, and the use of contraceptive (estroprogestative) pills in defining the threshold for GH under OGTT [169]. However, it must be emphasized that the last Endocrine Society guidelines continue to recommend the 1 µg/ml threshold rather than the $0.4 \,\mu g/l$ threshold, considering that in the United States, the use of ultrasensitive GH assays is not yet generalized [161]. A paradoxical increase in GH following OGTT is observed in approximately 10-30% of patients with acromegaly [170, 171].

For the IGF-I assay, the IS 02/254 WHO reference standard has recently become available. It is an ~97%-pure recombinant standard recommended for manufacturers [164, 165]. The IGF-I level increases in parallel to the log of the GH concentration and must be determined by using age-adjusted norms because levels fall with age. A multicenter cohort study comparing six IGF-I immunoassays in 911 healthy individuals showed good agreement at lower but not upper levels [172]. This variability, especially in upper levels, which are more interesting when evaluating the biologic control of disease, leads to a marked variability in each individual's IGF-I levels. Concordance between assay values in intraindividual patients with acromegaly was on average good (ranging from moderate to excellent) [173]. These differences in assay performances must be considered when evaluating disease control in subjects with acromegaly [173].

Similar to GH, IGF-I levels in patients with acromegaly are inversely correlated with age and with the maximal tumor diameter [10].

High IGF-I concentrations are also systematically found in other physiological states, such as pregnancy, puberty, and the postpubertal period. The concentration of IGFBP-3, the main IGF carrier protein, is usually increased in patients with acromegaly, but this marker offers little further diagnostic information in differential diagnosis.

GH and/or IGF-I measurements are of limited use for diagnosis (or treatment efficacy assessment) in patients with uncontrolled diabetes mellitus, chronic renal failure or pregnancy, and at the time of puberty.

Estradiol increases either basal GH or nadir GH levels after OGTT. This explains why GH is rarely inhibited by OGTT in women taking estrogen-containing pills. GH nadir concentrations are also significantly higher in lean and normal weight compared to overweight or obese subjects [169].

There are some individuals with a typical clinical picture of acromegaly but normal IGF-I and GH concentrations. This situation could correspond to two different situations: (1) spontaneously resolving real acromegaly, probably through necrosis or apoplexy of a previous GH-secreting pituitary adenoma; facial sequelae and disfigurement could have persisted despite the normalization of the somatotroph axis after spontaneous adenoma shrinkage; and (2) acromegaloid features may also be encountered in other diseases, such as severe insulin resistance, severe hypothyroidism, some forms of lipodystrophy, genodermatoses, or rarer overgrowth disease [174–176]. An extensive review focusing on various causes of pseudoacromegaly has recently been published [177].

Finally, some adenomas excised for mass effect or upon another surgical indication were revealed to derive from the somatotroph lineage only after histopathological examination [178]. In most of these cases, the somatotroph adenoma is silent, and no clinical signs of acromegaly are found. Nonetheless, subtle abnormalities revealing GH/IGF-I hypersecretion may be encountered [25, 179].

5.5.2 Neuroimaging

MRI is the imaging method of choice to detect a pituitary lesion. T1- and T2-weighted coronal and T1-weighted sagittal sections are routinely performed in diencephalic studies; gadolinium contrast usually shows a retardation in lesion enhancement demarcating the remaining hypophyseal tissue.

The majority of patients clearly have a pituitary macroadenoma (lesion above 10 mm). In patients with a sellar macroadenoma, once the diagnosis is established, before initiating treatment for acromegaly, patients must undergo a thorough workup focusing on tumor mass effects (headaches, changes in the visual field and acuity, MRI abnormalities) and anterior pituitary function.

Although the majority of somatotroph adenomas are large tumors, in recent decades, the prevalence of microadenomas has seemingly increased in patients with acromegaly. It has to be known whether this trend depends on an improved clinical skill to detect disease (and therefore smaller lesions) or an intrinsic biological characteristic of somatotroph adenomas.

GH-secreting pituitary adenomas can be hypo-, iso-, or hyperintense on T2-weighted MRI sequences. Some authors suggest that hypointense imaging on T2-weighted MRI predicts a better outcome after somatostatin analog treatment either in terms of biochemical profile or tumoral shrinkage [180].

When MRI is contraindicated, a skull base CT scan may still be used. In patients with macroadenoma, this technique may show the presence of the pituitary mass and various extents of enlargement of the sella turcica.

Novel tools such as ¹¹C-methionine positron emission tomography seem to detect small pituitary remnants, especially those with a high metabolic rate and hypersecretion [181]. This technique seems particularly promising in equivocal MRI images [182]. Nevertheless, the accuracy of this technique has yet to be extensively established, and ¹¹C-methionine PET sequences are not routinely indicated in assessing pituitary imaging. Moreover, this technique is available in very few centers where a cyclotron is available on site due to the very short half-life of the radionuclide.

5.5.3 Pituitary Assessment

Associated prolactin hypersecretion is present in up to 30% of cases and may be either functional, secondary to impairment of hypothalamic dopamine production or compression of the pituitary stalk by the tumor, or due to a mixed adenoma.

In patients with microadenoma, no other pituitary defect or sellar mass effect is expected.

5.5.4 Total Body Imaging

If, despite an overt disease, no image is found on MRI, an ectopic GHRH secretion must be suspected and appropriate imaging requested [50].

Some occult neuroendocrine tumors may require total body scans (CT scans, MRI) or functional imaging (Octreoscan®, F-DOPA, or DOTATOC) [183]. Biopsy may help prove the neuroendocrine nature of these neoplasms. Complete excision of the underlying tumor usually cures disease.

5.6 Management and Follow-Up

Management of acromegaly is multimodal and quite consensual across different American and European guidelines and clinical practices [74, 165, 184–188].

The main aim is to relieve symptoms, normalize or decrease GH/IGF-1 excess, remove or reduce pituitary tumors, and improve long-term morbidity and mortality [185, 189]. Recent epidemiological studies have helped to refine the definitions of "cure" and "good disease control", which are now far more precise: the GH concentration (in a random sample) must return to less than 1 μ g/l in the new sensitive assays that are now widely used (if the OGTT is used, the nadir needs to be less than 0.4 μ g/l) and the IGF-I level must return to normal according to sex and age [165, 187]. A stepwise therapeutic strategy based on surgery and/or radiotherapy and/or medical treatment allows these goals to be achieved.

5.6.1 Neurosurgery

Surgery is generally the first-line treatment. Tumor excision, usually by the transsphenoidal route, is the most rapid way of reducing GH and IGF-I concentrations in patients with acromegaly. Nevertheless, these levels normalize in only 40–70% of cases after surgery [11, 190–194]. The success rate depends on a range of features, such as the tumor size (microadenomas are more amenable to cure), the preoperative GH concentration (the success rate is higher when GH concentrations are <10 µg/l), and the surgeon's experience. Endoscopic techniques, now used in the majority of expert centers [195], though not improving the success rate, may attenuate local adverse effects [192].

Postoperative outcome in terms of symptom relief and biological disease control is generally assessed 3 months after surgery. When surgery fails to achieve disease control or when surgery is impossible or contraindicated, patients are offered radio-therapy and/or pharmacological treatments.

5.6.2 Radiating Techniques

Radiotherapy techniques have evolved over time, refining the techniques and number of sessions. Radiosurgery is a term used to define high-dose radiation delivery. It better applies to small targets and requires a single or few sittings. Fractionated radiotherapy refers to radiation therapy delivered at smaller doses but with multiple treatments (typically 25–30 sittings during 5–6 weeks). In order to minimize the dose to surrounding tissues, stereotactic localization is now used. Stereotactic radiosurgery (SRS) may use different radiating particles, such as photons (gamma knife, Linac, CyberKnife) or charged ions (protons). Stereotactic fractionated radiotherapy (SFR) is a hybrid form combining stereotactic localization with fractionated therapy administered by 3D-conformal radiation therapy, intensity-modulated radiation therapy, or proton radiation therapy [196].

Radiating techniques have consistently evolved over the last decade, from conventional radiotherapy to three-dimensional (3-D) conformal and stereotactic techniques. Technical improvements have been performed in all aspects of radiation treatment, including better imaging and 3-D planning, patient immobilization, sophisticated imaging systems for accurate patient repositioning and a more precise dose delivery, and reduction in normal surrounding brain structures exposed to high radiation doses [197].

In patients with somatotroph adenomas, normalization of GH/IGF-I levels occurs in approximately 40–60% of patients 5–10 years after treatment, with a 50% decline in GH and IGF-1 preradiation levels in approximately 2 and 5 years, respectively [190, 198, 199].

The choice between SFR and SRS, like for any other pituitary adenoma, in part depends on the size of the tumor and on its contiguity with the optic apparatus [200]. It is of note that the baseline GH concentrations predict treatment outcome

and the time-to-normalization of patients with high (>3–4-fold ULN) IGF-1 levels requiring up to 10 years to achieve biochemical control of disease [199, 201].

SRS provides more focused irradiation. In a French series of over 80 patients, the efficacy of gamma-knife irradiation was close to that of SFR [202]. In a recent meta-analysis, disease control (without complementary medical treatment) was achieved in 48–53% of cases after a mean follow-up of 4 years. The relatively larger figures by SRS are probably explained by the small size of tumors ($2.1 \pm 1.2 \text{ mL}$) [203]. Apart from disease control, the different SRS techniques give excellent results in controlling tumor growth, with >95% success according to different series [200, 202–204].

A recent review of the literature proved a similar rate of tumor control between stereotactic radiosurgery and SFR for patients with persistent active acromegaly after surgery and/or during medical therapy [205]. Tumors were stable or decreased in 93–100% of patients at 5–10 years, whereas endocrinological remission was achieved in 40–60% of patients at 5 years [205].

On the other hand, radiotherapy leads to variable degrees of anterior pituitary insufficiency in 50–100% of patients after 10–15 years, regardless of the technique. Complications such as radionecrosis and optic neuropathy are very rare. In contrast, the risk of stroke and cerebrovascular events may be increased, sometimes many years after irradiation [206]. When compared to patients not exposed to radiotherapy, stroke incidence appears to be increased from 1.7 to 2.8 times [200, 207]. Along with cortisol deficiency and inadequate hormonal substitution, these findings seem to account for the excess mortality in these patients [208]. The question of whether cerebrovascular risk may be lowered by newer radiating techniques is presently still unanswered.

5.6.3 Medical Treatment

5.6.3.1 Dopamine Agonists (DA)

Cabergoline appears to be the most effective among DA agents [209, 210]. In a meta-analysis of all published studies, IGF-I normalization was achieved in up to 34% of cases [211]. Multivariate analysis showed that the efficacy depended on the initial IGF-I concentration, the treatment duration, and the basal concentration of PRL (and, to a lesser degree, the dose of cabergoline) [211]. As in patients with hyperprolactinemia, cardiac valve disease does not seem to be increased in patients with acromegaly treated with cabergoline in the long term [133, 212].

Because of their dual origin, mixed lactotroph/somatotroph tumors are more likely to respond to DA [213]. Half of the patients with GH/PRL-secreting adenomas normalize their IGF-I levels, and 60% of those with macroadenomas display tumor shrinkage [209].

5.6.3.2 Somatostatin Receptor Ligands (SRLs)

SRLs suppress GH secretion by binding to somatostatin receptor subtypes (sst) sst2 and sst5, which are mainly present on somatotroph adenoma cells [214]. These



Fig. 5.5 Trends of disease control, treatment approach, and medical therapy over time in acromegaly—the example of the French Registry of Acromegaly. Panel **a**. Evolution of disease status across 4-year follow-up periods in the French acromegaly registry. Histograms indicate the percentages of patients and standard deviations; trt: medical treatment. Panel **b**. Distribution of treatment approaches in different follow-up periods. *MT* medical treatment, *RT* radiotherapy. Data are reported as percentages of patients. Panel **c**. Evolution of drug therapy in medically treated patients: somatostatin analogs (SSA), dopamine agonists (DA), GH receptor antagonist (GHRA). The histograms report the prescribed classes of treatment per patient according to different follow-up periods as the percentage of prescriptions. Note: Pegvisomant was approved for the treatment of acromegaly in France in late 2003. Adapted from [23], with permission

drugs have been demonstrated to exert either antisecretory or antitumoral effects (Fig. 5.5).

The first SRL to be marketed, octreotide (Sandostatin®), can be injected subcutaneously (SC), generally by the patient him/herself, at a dose of 100–200 μ g two or three times a day [215]. Sustained-release SRLs (lanreotide and octreotide LAR) progressively followed and had an impact on the market due to their comfort. The former requires deep SC injections every 28 days at variable doses (Somatuline® Autogel® 60, 90, or 120 mg). The latter is administered intramuscularly once a month (Octreotide LAR, Sandostatin® LAR 10–20 or 30 mg). The dose and frequency of injections may be initiated and adjusted depending on the GH/IGF-1 concentration.

These SRLs bear similar efficacy [216] in driving GH concentrations below $2 \mu g/l$ (60% to 70% of cases) and in normalizing IGF-1 levels (50–80%) [217, 218]. A recent meta-analysis has emphasized that control rates were highly variable from one study to the other. If clinical design characteristics had no statistically significant impact on efficacy determination, then later year of publication, study duration,

and prior somatostatin analog use were significant efficacy determinants for acromegaly trial outcomes. In that meta-analysis, overall achieved control rates were 56% for mean GH and 55% for IGF-1 normalization [219].

Several long-term studies have shown that the cure rate may improve over time [220–222].

In a handful of good responders, SRL injection frequency may be lengthened or even safely halted with no subsequent increase in GH/IGF-I concentrations [223, 224].

Tumor volume shrinks in a weighted mean of 37–51% of patients [225]. It seems that the reduction in tumor volume is larger when an SRL is used as first-line treatment [226]. When not shrinking, tumor volumes remain at least stable in the vast majority of cases [217].

SRLs may cause gastrointestinal disorders (abdominal bloating, nausea, diarrhea), which are generally transient. SRLs induce the occurrence of gallstones in 10–20% of cases that did not respond to ursodeoxycholic acid [227, 228]. Some practitioners prescribe pancreatic enzymes in SRL-related diarrhea. Changes in glucose metabolism are sometimes observed, including impaired glucose tolerance or even diabetes in patients who are overweight. In other cases, however, glucose tolerance improves following the reduction in insulin resistance due to the lowering of GH concentrations. Overall, according to recent meta-analyses, despite a decrease in fasting plasma insulin levels, no consistent changes in fasting glucose and HbA1c levels have been observed [229, 230].

Pasireotide (SOM230 or Signifor®) is a second-generation SRL compound that binds to sst1, 2, 3, and 5 with high affinity [231], which has been proven to be effective in controlling acromegaly [232–234]. When directly compared to octreotide LAR, pasireotide was able to control a higher proportion of patients (36% versus 21%) [232]. In a crossover study, 15% of noncontrolled patients under maximal octreotide doses responded to pasireotide [235].

Concerning the side effects, glucose metabolism abnormalities (diabetes or glucose intolerance) were far more frequent in patients receiving pasireotide than in those administered conventional SRLs [232, 233]. Gastrointestinal symptoms after pasireotide treatment seem to occur at a similar or slightly increased frequency [233].

This drug may be particularly interesting in patients with partial resistance to first-generation SRLs.

Among the factors believed to influence SRL effectiveness, there is the T2-weighted hypointense signal on MRI [236, 237], the presence of specific somatostatin receptor subtypes, and the aspect of densely granulated cells at histological examination [238].

5.6.3.3 GH-Receptor Antagonists

Pegvisomant (Somavert®) acts peripherally, blocking the effects of GH on its target organs by binding to GH receptors and by preventing their dimerization, GH signal transduction, and downstream activity, including IGF-I production [239]. As pegvisomant inhibits the action of GH but not its secretion, GH concentrations cannot be

used to evaluate treatment efficacy. IGF-I is used as a surrogate marker, together with clinical parameters. Pegvisomant is administered subcutaneously at a daily dose of 10–20 mg (sometimes more), with the dose being adapted to the hormone response (IGF-I normalization). Pegvisomant is highly effective, as IGF-I levels normalize in more than 90% of patients in the initial trials reported [240, 241]. In routine practice, the pegvisomant efficacy rate seems to be as low as 70% of cases, as shown by observational studies [242–247]. This treatment is reserved for patients in whom SRLs fail.

In a series of 304 patients in whom tumor volume was monitored for at least 3 years, an increase in tumor volume occurred in 9 cases within 8 months after commencing pegvisomant. This is likely related to rebound expansion after discontinuation of SRLs and/or to the natural history of aggressively growing pituitary tumors [248]; this latter situation may justify combination with an SRL to reduce tumor volume [249]. Tumor volume must therefore be monitored (by MRI) during this treatment. Available clinical data on pegvisomant concern a relatively small number of patients and relatively short treatment periods. Independent of disease control, pegvisomant improves glucose metabolism [250]. Adverse effects are limited to rare liver enzyme elevations, which are observed in between 2.5% and 3% of patients according to surveillance studies [242, 247]. Liver enzyme elevation generally normalizes either spontaneously or after treatment interruption. Exceptional cases of true hepatitis have been reported [246, 247]. Gilbert disease has been suggested as a risk factor for severe hepatitis [251, 252], but this was not confirmed by a recent Italian study [253].

SRL-pegvisomant combination therapy has also been developed [254]: a slow release formulation of the SRL is given once a month at the highest marketed dose (30 mg octreotide LAR or 120 mg lanreotide Autogel), and pegvisomant is injected once a week at escalating doses until the IGF-I level normalizes. IGF-I normalization was obtained in all patients with a median weekly pegvisomant dose of 60 mg [255]. This decreased dose requirement during combined therapy might be partially explained by an increase of approximately 20% in serum levels of pegvisomant [256]. Biochemical hepatic anomalies were quite frequent (although always transient) with this combination and appeared to be twice as common in patients with acromegaly with diabetes [257]. Compared with octreotide monotherapy, this combination appears to have a greater positive impact on quality of life for a given degree of IGF-I normalization [258]. This has raised the hypothesis of an extrahepatic effect of pegvisomant [259].

Cabergoline-pegvisomant combination therapy has also been proposed. In a multicenter, open-label, prospective clinical trial [260], the combination of cabergoline and low-dose pegvisomant (10 mg/day) was associated with a significant decrease in IGF-I levels compared with cabergoline alone, and 68% of patients achieved normalization. Then, when cabergoline was withdrawn and pegvisomant continued as monotherapy, only 26% of patients maintained normal IGF-I levels. The adjunction of cabergoline may be interesting when pegvisomant alone achieved minimally increased IGF-I levels [261].

5.6.4 Treatment Strategy

By analyzing large caseload series, several studies have evaluated medical practices and the evolution of treatment strategies over time [9, 11, 13]. The advantages, disadvantages, and costs of the different treatment options must be considered [262]. A marked evolution in clinical practice has been observed in recent decades (Fig. 5.5a–c).

An algorithm indicating a putative therapeutic strategy is proposed in Fig. 5.6. A surgical procedure is tried whenever possible. This depends on the availability of an



Fig. 5.6 Proposed algorithm of the treatment strategy for acromegaly. After confirmation of acromegaly, the first step is to establish the patient's eligibility for neurosurgery. In the absence of disease remission/cure after surgery, long-acting somatostatin receptor ligands (SRLs) are indicated. SRL doses and frequencies should be adapted and optimized, especially in partial responders (>50% decrease in growth hormone (GH) and/or insulin-like growth factor 1 (IGF-I)). In the case of mild IGF-I elevation (<2-2.5-fold of the adjusted value for sex and age of the upper limit of normal value (ULN)), the addition of cabergoline can be considered. If disease control is not achieved, patients should be switched to the second-generation SRL pasireotide if there is clinically relevant residual tumor on imaging and/or clinical concern of tumor growth. Patients with impaired glucose tolerance should be switched to the GH antagonist pegvisomant (PegV). Patients with impaired glucose tolerance and tumor concern could be treated with a combination of a firstgeneration SRL and PegV. Patients who remain uncontrolled despite this second-line medical therapy should be discussed by a multidisciplinary team and considered for a second surgical intervention, a radiating therapy or temozolomide (features of aggressiveness, high Ki-67, tumor progression). *: consider using cabergoline in place of first-generation SRL if IGF-I elevation is <2-2.5-fold the ULN and/or in case of mixed GH/prolactin-secreting tumor; **near-normal IGF-I is considered for IGF-I values <1.3 ULN

experienced neurosurgeon, on the feasibility, on the fact that there are no anesthesiology constraints, and on the patient's choice. Surgery has been chosen in nearly 80% of cases, considering the weighted mean of 19 studies across different countries encompassing more than 16,000 patients with acromegaly [11].

In the case of surgery failure to cure acromegaly, medical treatment with SRLs is preferred as an elective option. SRL therapy is not only indicated after surgical failure but can sometimes be used for first-line treatment, especially when severe comorbidities create a risk of perioperative complications. Thus, when heart failure or respiratory problems are associated with acromegaly [71, 141], it is preferable to prepare the patient for surgery by administering SRLs for a few months first. In some cases, when a large tumor extends outside the *sella* and is not completely extractable by surgery, SRLs can be administered in the hope of controlling GH hypersecretion and tumor growth, thus avoiding the need for surgery [226, 263–267]. First-line SRL treatment before surgery has been chosen in 0–52% of cases, according to different series [11]. In these cases, when clinical conditions improve, neurosurgery could be performed as second-line treatment. According to a meta-analysis, IGF-I more likely normalizes after second-line treatment than after first-line drug therapy [218].

There is some controversy surrounding the ability of preoperative SRL therapy to improve surgical outcome: some studies [268–274] indicate that, in some patients with isolated somatotroph macroadenomas, surgery provides better control of acromegaly when patients are pretreated with an SRL, while other studies showed no difference [275–278]. Yang et al., in their latest meta-analysis, showed that preoperative SRL treatment was able to improve short-term (OR 2.07, 95% CI 1.50–2.87, p < 0.00001) but not long-term biochemical control [279].

The overall ability to normalize IGF-I by a first-line full-dose SRL (either octreotide LAR or lanreotide) is approximately 50% in recent series and the latest metaanalyses [280].

In some selected patients, especially in those with moderately increased IGF-I levels (below 2–2.5-fold the upper limit of normal) or in those with elevated concomitant prolactin levels, cabergoline may be tried first. It has indeed been shown that cabergoline is particularly useful in patients with low IGF-I excess [211].

After a first surgical approach, when full-dosed SRL therapy fails to achieve remission, several options may be chosen, mainly according to the patient status, his/her willingness, and the severity of clinical/biochemical disease persistence:

- (a) In the case of a large tumor remnant, it may be of utmost interest to repeat surgery. The main aim, in this case, will not be to cure disease but to debulk and decrease the secretory mass before trying medical treatment again [281, 282].
- (b) In the case of partial response to SRLs, physicians may choose to further adapt the adjusted doses [283, 284], to combine with other agents such as cabergoline [211], to administer pasireotide, or to initiate pegvisomant. When analyzing current anti-GH/IGF-I medical choices in these cases, a striking similarity is found between Germany and France, two countries where drugs are similarly available [9, 13]. In second-line treatment (after surgery failure), 60% of

patients have been treated with SRLs alone, 10% by a combination of SRLs and DA, 10% by pegvisomant, 8.6% by DA alone, 6.8% by SRLs and pegvisomant, 2% by DA and pegvisomant, and 2.7% by tritherapy (SRLs+DA + pegvisomant). Notably, at the time of the survey, pasireotide was not yet available in these countries.

- (c) In the case of full persistence or disease that is still clinically and biochemically severe, pegvisomant should be rapidly tried. Increasing doses may be chosen to improve symptoms and to normalize IGF-I levels. If possible, surgical debulking may also be interesting to propose for reducing the dose of pegvisomant necessary to achieve normal IGF-I.
- (d) Radiotherapy is far less chosen as the first or second line. Studies evaluating clinical practices over time show a remarkable reduction in radiotherapy use between <2001 and > 2007 (Fig. 5.5b) [9]. However, radiotherapy may be particularly interesting when aiming at avoiding remnant enlargement and controlling GH secretion. In the case of a small remnant, gamma- or CyberKnife could be proposed. In the case of large tumors, fractionated radiotherapy may be proposed to avoid further tumor enlargement. Apart from clinical and biochemical issues, the cost of these long-term medical treatments should be weighed against the risks of radiotherapy. In any event, medical treatment will be necessary while waiting for the benefits of radiotherapy to emerge.

All these treatments must be reassessed on a yearly basis. After radiotherapy, if medical treatment is necessary while waiting for the effects of irradiation, regular withdrawal is necessary for assessing the persistence of active disease.

5.7 Prognosis

Several targeted studies and meta-analyses have been conducted to explore mortality in populations with acromegaly. The overall body of evidence globally shows an increased mortality rate in patients with acromegaly [11, 285, 286].

According to the earliest series published in the 1980s–1990s, approximately 60% of patients die from cardiovascular disease, 25% from respiratory complications, and 15% from cancer. If untreated, patients with acromegaly have been reported to die approximately 10 years earlier than healthy subjects [5]. Several studies have shown that cerebrovascular disorders are a frequent cause of death, especially among women, but these studies involved patients treated in various ways (craniotomy, radiotherapy), many years ago, and a deleterious effect of these treatments (especially radiotherapy) cannot be ruled out [287, 288]. Two recent meta-analyses [286, 289] showed a standardized mortality ratio (SMR), i.e., the ratio of observed mortality in the population with acromegaly to expected mortality in the general population, of 1.72 (IC 95%, 1.62–1.83). However, recent reports describe a reduction in this trend, with mortality rates that seem no longer higher than the general population [290]. Bolfi et al. compared the mortality rates across 26 studies according to the date of publication of the series [285]. They found that the mortality in acromegaly was increased from the 17 studies published before 2008, while no difference from the general population was found from the nine studies published after this date (SMR, 1.35; CI, 0.99–1.85). The posttreatment GH concentration is probably the best predictor of survival for all causes of death, independent of the type of complication. Thus, life expectancy outcomes can be stratified according to the posttreatment GH concentration: if GH secretion is controlled (<2 or 2.5 μ g/l, or IGF-I normalization), life expectancy merges with that of the matched general population [287, 290, 291].

In the meta-analysis published by Holdaway and colleagues, it has been shown that the prognosis of acromegaly has improved in the last 20 years. This improvement is probably due to better disease control through a multimodal treatment strategy or to better management of comorbidities through improved awareness by physicians. Concerning the former aspect, a multimodal strategy and/or various anti-GH drug combinations witnessed a more aggressive treatment of the disease [9, 13, 292]. Concerning the latter, it is believed that mortality in acromegaly largely depends on the numerous and severe comorbidities. High GH/IGF-I concentrations, arterial hypertension, and cardiomyopathy are factors of poor prognosis, while the duration of symptoms and other factors (diabetes, lipid disorders, and cancer) are less important. Quality of life is also altered in acromegaly and is improved by effective treatment [293].

Finally, it must be stressed that, with the current therapeutic strategy, the vast majority of patients with acromegaly achieve very good control of GH/IGF-I secretion and have no problems related to tumor growth. Up to two-thirds of patients have normal IGF-I at the last visit, and these numbers seem to have increased in the latest series [11].

Adverse effects are infrequent and minor, even in the very long term. The figure is broadly different from the disease management 20 or more years ago, before the advent of somatostatin analogs. In addition, the use of more stringent criteria to define cure, together with aggressive treatment of comorbidities, has significantly improved the outlook of patients with acromegaly [118, 294]. However, even cured or well-controlled patients may have invalidating sequelae, such as joint pain, deformities, and impaired quality of life [145].

5.8 Conclusion

Acromegaly is a rare disease characterized by excessive GH and IGF-I exposure. In addition to the typical dysmorphic syndrome, a number of highly morbid diseases are associated with acromegaly. The cardiovascular, respiratory, rheumatologic and metabolic consequences and an increased oncologic risk represent the real burden of acromegalic disease and determine the prognosis. A detailed workup of the various organs potentially involved in these complications is therefore recommended. Treatment is aimed at correcting (or preventing) tumor compression of neighboring tissues by excising the culprit lesion and at reducing GH and IGF-I levels to normal values. Selective pituitary sphenoidal surgery is often the first-line treatment. When surgery fails to correct GH/IGF-I hypersecretion, a multimodal therapeutic strategy is available, including several medical treatment classes (standard and novel somatostatin analogs, dopamine agonists, or the GH receptor antagonist pegvisomant). Radiotherapy is currently proposed as a third-line treatment. The prognosis of acromegaly has improved in recent years: ade-quate hormonal control is achieved in most cases, providing a life expectancy increasingly similar to that of the general population. It remains to be shown whether the criteria used to define control of the disease in terms of mortality also apply to optimal management of comorbidities.

5.9 Learning Points

- Acromegaly, even if clinicaly manifest is diagnosed after a long delay which makes the cure of the disease more difficult and aggravates comorbidities.
- Preoperative GH level (high GH output) is an important predictor of remission in acromegaly.
- Persistent elevation of biochemical markers (GH and IGF1) following surgery indicates uncontrolled disease.
- Medical therapy nowadays is an important part of the treatment of patients with acromegaly.
- Age, size of the tumor, GH levels on presentation, histopathological type, and the somatostatin receptor status of the tumor in acromegaly may predict response to medical therapy.
- Despite the availability of multiple treatment modalities (surgery, radiotherapy, somatostatin analogs first and second generation, GH receptor antagonists, etc.), acromegaly sometimes is a challenging condition to treat.

References

- Kamenicky P, Mazziotti G, Lombes M, Giustina A, Chanson P. Growth hormone, insulin-like growth factor-1, and the kidney: pathophysiological and clinical implications. Endocr Rev. 2014;35(2):234–81.
- 2. Chanson P, Salenave S, Kamenicky P. Acromegaly. Handb Clin Neurol. 2014;124:197-219.
- Daly AF, Rixhon M, Adam C, Dempegioti A, Tichomirowa MA, Beckers A. High prevalence of pituitary adenomas: a cross-sectional study in the province of Liege. Belgium J Clin Endocrinol Metab. 2006;91(12):4769–75.
- 4. Fernandez A, Karavitaki N, Wass JA. Prevalence of pituitary adenomas: a community-based, cross-sectional study in Banbury (Oxfordshire, UK). Clin Endocrinol. 2010;72(3):377–82.
- 5. Holdaway IM, Rajasoorya C. Epidemiology of acromegaly. Pituitary. 1999;2(1):29-41.
- Schneider HJ, Sievers C, Saller B, Wittchen HU, Stalla GK. High prevalence of biochemical acromegaly in primary care patients with elevated insulin-like growth factor-1 levels. Clin Endocrinol. 2008; in press (available on line)
- 7. Raappana A, Koivukangas J, Ebeling T, Pirila T. Incidence of pituitary adenomas in northern Finland in 1992–2007. J Clin Endocrinol Metab. 2010;95(9):4268–75.
- Lavrentaki A, Paluzzi A, Wass JA, Karavitaki N. Epidemiology of acromegaly: review of population studies. Pituitary. 2017;20(1):4–9.

- Maione L, Brue T, Beckers A, Delemer B, Petrossians P, Borson-Chazot F, et al. Changes in the management and comorbidities of acromegaly over three decades: the French Acromegaly Registry. Eur J Endocrinol. 2017;176(5):645–55.
- Petrossians P, Daly AF, Natchev E, Maione L, Blijdorp K, Sahnoun-Fathallah M, et al. Acromegaly at diagnosis in 3173 patients from the Liege acromegaly survey (LAS) database. Endocr Relat Cancer. 2017;24(10):505–18.
- 11. Maione L, Chanson P. National acromegaly registries. Best Pract Res Clin Endocrinol Metab. 2019;
- Bex M, Abs R, T'Sjoen G, Mockel J, Velkeniers B, Muermans K, et al. AcroBel the Belgian registry on acromegaly: a survey of the 'real-life' outcome in 418 acromegalic subjects. Eur J Endocrinol. 2007;157(4):399–409.
- Schofl C, Franz H, Grussendorf M, Honegger J, Jaursch-Hancke C, Mayr B, et al. Long-term outcome in patients with acromegaly: analysis of 1344 patients from the German Acromegaly Register. Eur J Endocrinol. 2013;168(1):39–47.
- Ezzat S, Forster MJ, Berchtold P, Redelmeier DA, Boerlin V, Harris AG. Acromegaly. Clinical and biochemical features in 500 patients. Medicine (Baltimore). 1994;73(5):233–40.
 Neharra ID. Acromegaly. Clin Endocrinol. 1087;26(4):481–512.
- 15. Nabarro JD. Acromegaly. Clin Endocrinol. 1987;26(4):481–512.
- Nachtigall L, Delgado A, Swearingen B, Lee H, Zerikly R, Klibanski A. Changing patterns in diagnosis and therapy of acromegaly over two decades. J Clin Endocrinol Metab. 2008;93(6):2035–41.
- Reid TJ, Post KD, Bruce JN, Nabi Kanibir M, Reyes-Vidal CM, Freda PU. Features at diagnosis of 324 patients with acromegaly did not change from 1981 to 2006: acromegaly remains under-recognized and under-diagnosed. Clin Endocrinol. 2010;72(2):203–8.
- Dal J, Feldt-Rasmussen U, Andersen M, Kristensen LO, Laurberg P, Pedersen L, et al. Acromegaly incidence, prevalence, complications and long-term prognosis: a nationwide cohort study. Eur J Endocrinol. 2016;175(3):181–90.
- 19. Asa SL, Ezzat S. The pathogenesis of pituitary tumours. Nat Rev Cancer. 2002;2(11):836–49.
- Dworakowska D, Korbonits M, Aylwin S, McGregor A, Grossman AB. The pathology of pituitary adenomas from a clinical perspective. Front Biosci (Schol Ed). 2011;3:105–16.
- 21. Heaney AP, Melmed S. Molecular targets in pituitary tumours. Nat Rev Cancer. 2004;4(4):285–95.
- 22. Melmed S. Pathogenesis of pituitary tumors. Nat Rev Endocrinol. 2011;7(5):257-66.
- 23. Ho KKY, On behalf of the 2007 GH Deficiency Consensus Workshop Participants. Consensus guidelines for the diagnosis and treatment of adults with GH deficiency II: a statement of the GH Research Society in association with the European Society for Pediatric Endocrinology, Lawson Wilkins Society, European Society of Endocrinology, Japan Endocrine Society, and Endocrine Society of Australia. Eur J Endocrinol. 2007;157(6):695–700. https://doi.org/10.1530/EJE-07-0631.
- Beck-Peccoz P, Bassetti M, Spada A, Medri G, Arosio M, Giannattasio G, et al. Glycoprotein hormone alpha-subunit response to growth hormone (GH)-releasing hormone in patients with active acromegaly. Evidence for alpha-subunit and GH coexistence in the same tumoral cell. J Clin Endocrinol Metab. 1985;61(3):541–6.
- Sakharova AA, Dimaraki EV, Chandler WF, Barkan AL. Clinically silent somatotropinomas may be biochemically active. J Clin Endocrinol Metab. 2005;90(4):2117–21.
- Herman V, Fagin J, Gonski R, Kovacs K, Melmed S. Clonal origin of pituitary adenomas. J Clin Endocrinol Metab. 1990;71:1427–30.
- Horvath A, Stratakis CA. Clinical and molecular genetics of acromegaly: MEN1, Carney complex, McCune-Albright syndrome, familial acromegaly and genetic defects in sporadic tumors. Rev Endocr Metab Disord. 2008;9(1):1–11.
- Lytras A, Tolis G. Growth hormone-secreting tumors: genetic aspects and data from animal models. Neuroendocrinology. 2006;83(3–4):166–78.
- 29. Melmed S. Acromegaly pathogenesis and treatment. J Clin Invest. 2009;119(11):3189–202.
- Bi WL, Horowitz P, Greenwald NF, Abedalthagafi M, Agarwalla PK, Gibson WJ, et al. Landscape of genomic alterations in pituitary adenomas. Clin Cancer Res. 2017;23(7):1841–51.

- Gadelha MR, Kasuki L, Korbonits M. The genetic background of acromegaly. Pituitary. 2017;20(1):10–21.
- Valimaki N, Demir H, Pitkanen E, Kaasinen E, Karppinen A, Kivipelto L, et al. Wholegenome sequencing of growth hormone (GH)-secreting pituitary adenomas. J Clin Endocrinol Metab. 2015;100(10):3918–27.
- 33. Vallar L, Spada A, Giannattasio G. Altered Gs and adenylate cyclase activity in human GH-secreting pituitary adenomas. Nature. 1987;330(6148):566–8.
- 34. Asa SL, Ezzat S. The cytogenesis and pathogenesis of pituitary adenomas. Endocr Rev. 1998;19(6):798–827.
- 35. Melmed S. Mechanisms for pituitary tumorigenesis: the plastic pituitary. J Clin Invest. 2003;112(11):1603–18.
- Hage M, Viengchareun S, Brunet E, Villa C, Pineau D, Bouligand J, et al. Genomic alterations and complex subclonal architecture in sporadic GH-secreting pituitary adenomas. J Clin Endocrinol Metab. 2018;103(5):1929–39.
- Beck-Peccoz P, Brucker-Davis F, Persani L, Smallridge RC, Weintraub BD. Thyrotropinsecreting pituitary adenomas. Endocr Rev. 1996;17(6):610–38.
- Socin HV, Chanson P, Delemer B, Tabarin A, Rohmer V, Mockel J, et al. The changing spectrum of TSH-secreting pituitary adenomas: diagnosis and management in 43 patients. Eur J Endocrinol. 2003;148(4):433–42.
- 39. Chanson P, Salenave S, Droumaguet C, Cazabat L, Galland F, Young J. Rare causes of acromegaly. In: Wass JA, editor. Acromegaly: a handbook of history, current therapy and future prospects. Bristol: Bioscientifica Ltd; 2009. p. 70–98.
- Daly AF, Tichomirowa MA, Beckers A. The epidemiology and genetics of pituitary adenomas. Best Pract Res Clin Endocrinol Metab. 2009;23(5):543–54.
- Gadelha MR, Trivellin G, Hernandez Ramirez LC, Korbonits M. Genetics of pituitary adenomas. Front Horm Res. 2013;41:111–40.
- 42. Lecoq AL, Kamenicky P, Guiochon-Mantel A, Chanson P. Genetic mutations in sporadic pituitary adenomas—what to screen for? Nat Rev Endocrinol. 2015;11(1):43–54.
- Chanson P, Salenave S, Orcel P. McCune-Albright syndrome in adulthood. Pediatr Endocr Rev. 2007;4(Suppl4):453–63.
- 44. Collins MT, Singer FR, Eugster E. McCune-Albright syndrome and the extraskeletal manifestations of fibrous dysplasia. Orphanet J Rare Dis. 2012;7(Suppl 1):S4.
- Salenave S, Boyce AM, Collins MT, Chanson P. Acromegaly and McCune-Albright syndrome. J Clin Endocrinol Metab. 2014;99(6):1955–69.
- 46. Vortmeyer AO, Glasker S, Mehta GU, Abu-Asab MS, Smith JH, Zhuang Z, et al. Somatic GNAS mutation causes widespread and diffuse pituitary disease in acromegalic patients with McCune-Albright syndrome. J Clin Endocrinol Metab. 2012;97(7):2404–13.
- Thakker RV, Newey PJ, Walls GV, Bilezikian J, Dralle H, Ebeling PR, et al. Clinical practice guidelines for multiple endocrine neoplasia type 1 (MEN1). J Clin Endocrinol Metab. 2012;97(9):2990–3011.
- Verges B, Boureille F, Goudet P, Murat A, Beckers A, Sassolas G, et al. Pituitary disease in MEN type 1 (MEN1): data from the France-Belgium MEN1 multicenter study. J Clin Endocrinol Metab. 2002;87(2):457–65.
- 49. Sano T, Yamasaki R, Saito H, Hirose T, Kudo E, Kameyama K, et al. Growth hormonereleasing hormone (GHRH)-secreting pancreatic tumor in a patient with multiple endocrine neoplasia type I. Am J Surg Pathol. 1987;11(10):810–9.
- Garby L, Caron P, Claustrat F, Chanson P, Tabarin A, Rohmer V, et al. Clinical characteristics and outcome of acromegaly induced by ectopic secretion of growth hormone-releasing hormone (GHRH): a French nationwide series of 21 cases. J Clin Endocrinol Metab. 2012;97(6):2093–104.
- Georgitsi M, Heliovaara E, Paschke R, Kumar AVK, Tischkowitz M, Vierimaa O, et al. Large genomic deletions in AIP in pituitary adenoma predisposition. J Clin Endocrinol Metab. 2008;93(10):4146–51. https://doi.org/10.1210/jc.2008-1003.

- 52. Pellegata NS, Quintanilla-Martinez L, Siggelkow H, Samson E, Bink K, Hofler H, et al. Germ-line mutations in p27Kip1 cause a multiple endocrine neoplasia syndrome in rats and humans. Proc Natl Acad Sci U S A. 2006;103(42):15558–63.
- 53. Bertherat J. Carney complex (CNC). Orphanet J Rare Dis. 2006;1:21.
- 54. Kirschner LS, Carney JA, Pack SD, Taymans SE, Giatzakis C, Cho YS, et al. Mutations of the gene encoding the protein kinase a type I-alpha regulatory subunit in patients with the Carney complex. Nat Genet. 2000;26(1):89–92.
- 55. Beckers A, Aaltonen LA, Daly AF, Karhu A. Familial isolated pituitary adenomas (FIPA) and the pituitary adenoma predisposition due to mutations in the aryl hydrocarbon receptor interacting protein (AIP) gene. Endocr Rev. 2013;34(2):239–77.
- Chahal HS, Stals K, Unterlander M, Balding DJ, Thomas MG, Kumar AV, et al. AIP mutation in pituitary adenomas in the 18th century and today. N Engl J Med. 2011;364(1):43–50.
- 57. Vierimaa O, Georgitsi M, Lehtonen R, Vahteristo P, Kokko A, Raitila A, et al. Pituitary adenoma predisposition caused by germline mutations in the AIP gene. Science. 2006;312(5777):1228–30.
- Barlier A, Vanbellinghen JF, Daly AF, Silvy M, Jaffrain-Rea ML, Trouillas J, et al. Mutations in the aryl hydrocarbon receptor interacting protein gene are not highly prevalent among subjects with sporadic pituitary adenomas. J Clin Endocrinol Metab. 2007;92(5):1952–5.
- 59. Cazabat L, Libe R, Perlemoine K, Rene-Corail F, Burnichon N, Gimenez-Roqueplo AP, et al. Germline inactivating mutations of the aryl hydrocarbon receptor-interacting protein gene in a large cohort of sporadic acromegaly: mutations are found in a subset of young patients with macroadenomas. Eur J Endocrinol. 2007;157(1):1–8.
- Cazabat L, Bouligand J, Chanson P. AIP mutation in pituitary adenomas. N Engl J Med. 2011;364(20):1973–4. author reply 4-5
- Cazabat L, Bouligand J, Salenave S, Bernier M, Gaillard S, Parker F, et al. Germline AIP mutations in apparently sporadic pituitary adenomas: prevalence in a prospective singlecenter cohort of 443 patients. J Clin Endocrinol Metab. 2012;97(4):E663–70.
- 62. Daly AF, Vanbellinghen JF, Khoo SK, Jaffrain-Rea ML, Naves LA, Guitelman MA, et al. Aryl hydrocarbon receptor-interacting protein gene mutations in familial isolated pituitary adenomas: analysis in 73 families. J Clin Endocrinol Metab. 2007;92(5):1891–6.
- Trivellin G, Daly AF, Faucz FR, Yuan B, Rostomyan L, Larco DO, et al. Gigantism and acromegaly due to Xq26 microduplications and GPR101 mutation. N Engl J Med. 2014;371(25):2363–74.
- Kamenicky P, Bouligand J, Chanson P. Gigantism, acromegaly, and GPR101 mutations. N Engl J Med. 2015;372(13):1264.
- Guillemin R, Brazeau P, Bohlen P, Esch F, Ling N, Wehrenberg WB. Growth hormonereleasing factor from a human pancreatic tumor that caused acromegaly. Science. 1982;218(4572):585–7.
- 66. Biermasz NR, Smit JW, Pereira AM, Frolich M, Romijn JA, Roelfsema F. Acromegaly caused by growth hormone-releasing hormone-producing tumors: long-term observational studies in three patients. Pituitary. 2007;10(3):237–49.
- 67. Gola M, Doga M, Bonadonna S, Mazziotti G, Vescovi PP, Giustina A. Neuroendocrine tumors secreting growth hormone-releasing hormone: pathophysiological and clinical aspects. Pituitary. 2006;9:221–9.
- 68. Thorner MO, Perryman RL, Cronin MJ, Rogol AD, Draznin M, Johanson A, et al. Somatotroph hyperplasia. Successful treatment of acromegaly by removal of a pancreatic islet tumor secreting a growth hormone-releasing factor. J Clin Invest. 1982;70(5):965–77.
- Beuschlein F, Strasburger CJ, Siegerstetter V, Moradpour D, Lichter P, Bidlingmaier M, et al. Acromegaly caused by secretion of growth hormone by a non-Hodgkin's lymphoma. N Engl J Med. 2000;342(25):1871–6.
- Melmed S, Ezrin C, Kovacs K, Goodman RS, Frohman LA. Acromegaly due to secretion of growth hormone by an ectopic pancreatic islet-cell tumor. N Engl J Med. 1985;312(1):9–17.
- Colao A, Ferone D, Marzullo P, Lombardi G. Systemic complications of acromegaly: epidemiology, pathogenesis, and management. Endocr Rev. 2004;25(1):102–52.

- Gadelha MR, Kasuki L, Lim DST, Fleseriu M. Systemic complications of acromegaly and the impact of the current treatment landscape: an update. Endocr Rev. 2019;40(1):268–332.
- 73. Molitch ME. Clinical manifestations of acromegaly. Endocrinol Metab Clin N Am. 1992;21(3):597–614.
- Ribeiro-Oliveira A Jr, Barkan A. The changing face of acromegaly—advances in diagnosis and treatment. Nat Rev Endocrinol. 2012;8(10):605–11.
- 75. Danzig J. Acromegaly. BMJ. 2007;335(7624):824-5.
- Bogazzi F, Nacci A, Campomori A, La Vela R, Rossi G, Lombardi M, et al. Analysis of voice in patients with untreated active acromegaly. J Endocrinol Investig. 2010;33(3):178–85.
- Giustina A, Mazziotti G, Canalis E. Growth hormone, insulin-like growth factors, and the skeleton. Endocr Rev. 2008;29(5):535–59. https://doi.org/10.1210/er.2007-0036.
- Ueland T, Fougner SL, Godang K, Schreiner T, Bollerslev J. Serum GH and IGF-I are significant determinants of bone turnover but not bone mineral density in active acromegaly: a prospective study of more than 70 consecutive patients. Eur J Endocrinol. 2006;155(5):709–15.
- Hong AR, Kim JH, Kim SW, Kim SY, Shin CS. Trabecular bone score as a skeletal fragility index in acromegaly patients. Osteoporos Int. 2016;27(3):1123–9.
- Mazziotti G, Bianchi A, Bonadonna S, Cimino V, Patelli I, Fusco A, et al. Prevalence of vertebral fractures in men with acromegaly. J Clin Endocrinol Metab. 2008;93(12):4649–55.
- Claessen KM, Kroon HM, Pereira AM, Appelman-Dijkstra NM, Verstegen MJ, Kloppenburg M, et al. Progression of vertebral fractures despite long-term biochemical control of acromegaly: a prospective follow-up study. J Clin Endocrinol Metab. 2013;98(12):4808–15.
- Kropf LL, Madeira M, Vieira Neto L, Gadelha MR, de Farias ML. Functional evaluation of the joints in acromegalic patients and associated factors. Clin Rheumatol. 2013;32(7):991–8.
- Liote F, Orcel P. Osteoarticular disorders of endocrine origin. Baillieres Best Pract Res Clin Rheumatol. 2000;14(2):251–76.
- 84. Wassenaar MJ, Biermasz NR, van Duinen N, van der Klaauw AA, Pereira AM, Roelfsema F, et al. High prevalence of arthropathy, according to the definitions of radiological and clinical osteoarthritis, in patients with long-term cure of acromegaly: a case-control study. Eur J Endocrinol. 2009;160(3):357–65.
- Claessen KM, Ramautar SR, Pereira AM, Romijn JA, Kroon HM, Kloppenburg M, et al. Increased clinical symptoms of acromegalic arthropathy in patients with long-term disease control: a prospective follow-up study. Pituitary. 2013;
- Miller A, Doll H, David J, Wass J. Impact of musculoskeletal disease on quality of life in long-standing acromegaly. Eur J Endocrinol. 2008;158(5):587–93.
- Wassenaar MJ, Biermasz NR, Kloppenburg M, van der Klaauw AA, Tiemensma J, Smit JW, et al. Clinical osteoarthritis predicts physical and psychological QoL in acromegaly patients. Growth Hormon IGF Res. 2010;20(3):226–33.
- Scarpa R, De Brasi D, Pivonello R, Marzullo P, Manguso F, Sodano A, et al. Acromegalic axial arthropathy: a clinical case-control study. J Clin Endocrinol Metab. 2004;89(2):598–603.
- Wassenaar MJ, Biermasz NR, Hamdy NA, Zillikens MC, van Meurs JB, Rivadeneira F, et al. High prevalence of vertebral fractures despite normal bone mineral density in patients with long-term controlled acromegaly. Eur J Endocrinol. 2011;164(4):475–83.
- 90. de Azevedo Oliveira B, Araujo B, Dos Santos TM, Ongaratti BR, Rech C, Ferreira NP, et al. The acromegalic spine: fractures, deformities and spinopelvic balance. Pituitary. 2019;22(6):601–6.
- Jenkins PJ, Sohaib SA, Akker S, Phillips RR, Spillane K, Wass JA, et al. The pathology of median neuropathy in acromegaly. Ann Intern Med. 2000;133(3):197–201.
- Tagliafico A, Resmini E, Nizzo R, Bianchi F, Minuto F, Ferone D, et al. Ultrasound measurement of median and ulnar nerve cross-sectional area in acromegaly. J Clin Endocrinol Metab. 2008;93(3):905–9.
- Tagliafico A, Resmini E, Nizzo R, Derchi LE, Minuto F, Giusti M, et al. The pathology of the ulnar nerve in acromegaly. Eur J Endocrinol. 2008;159(4):369–73.

- 94. Chemla D, Attal P, Maione L, Veyer AS, Mroue G, Baud D, et al. Impact of successful treatment of acromegaly on overnight heart rate variability and sleep apnea. J Clin Endocrinol Metab. 2014;99(8):2925–31.
- 95. Furman K, Ezzat S. Psychological features of acromegaly. Psychother Psychosom. 1998;67(3):147–53.
- 96. Tiemensma J, Kaptein AA, Pereira AM, Smit JW, Romijn JA, Biermasz NR. Affected illness perceptions and the association with impaired quality of life in patients with long-term remission of acromegaly. J Clin Endocrinol Metab. 2011;96(11):3550–8.
- Sibeoni J, Manolios E, Verneuil L, Chanson P, Revah-Levy A. Patients' perspectives on acromegaly diagnostic delay: a qualitative study. Eur J Endocrinol. 2019;180(6):339–52.
- 98. Andela CD, Niemeijer ND, Scharloo M, Tiemensma J, Kanagasabapathy S, Pereira AM, et al. Towards a better quality of life (QoL) for patients with pituitary diseases: results from a focus group study exploring QoL. Pituitary. 2015;18(1):86–100.
- Biermasz NR. The burden of disease for pituitary patients. Best Pract Res Clin Endocrinol Metab. 2019;33(2):101309.
- 100. Chanson P, Timsit J, Masquet C, Warnet A, Guillausseau PJ, Birman P, et al. Cardiovascular effects of the somatostatin analog octreotide in acromegaly. Ann Intern Med. 1990;113(12):921–5.
- 101. Kamenicky P, Viengchareun S, Blanchard A, Meduri G, Zizzari P, Imbert-Teboul M, et al. Epithelial sodium channel is a key mediator of growth hormone-induced sodium retention in acromegaly. Endocrinology. 2008;149(7):3294–305.
- 102. Kamenicky P, Blanchard A, Frank M, Salenave S, Letierce A, Azizi M, et al. Body fluid expansion in acromegaly is related to enhanced epithelial sodium channel (ENaC) activity. J Clin Endocrinol Metab. 2011;96(7):2127–35.
- 103. Maison P, Demolis P, Young J, Schaison G, Giudicelli JF, Chanson P. Vascular reactivity in acromegalic patients: preliminary evidence for regional endothelial dysfunction and increased sympathetic vasoconstriction. Clin Endocrinol. 2000;53(4):445–51.
- 104. Colao A, Baldelli R, Marzullo P, Ferretti E, Ferone D, Gargiulo P, et al. Systemic hypertension and impaired glucose tolerance are independently correlated to the severity of the acromegalic cardiomyopathy. J Clin Endocrinol Metab. 2000;85(1):193–9.
- 105. Jaffrain-Rea ML, Moroni C, Baldelli R, Battista C, Maffei P, Terzolo M, et al. Relationship between blood pressure and glucose tolerance in acromegaly. Clin Endocrinol. 2001;54(2):189–95.
- 106. Puglisi S, Terzolo M. Hypertension and acromegaly. Endocrinol Metab Clin N Am. 2019;48(4):779–93.
- 107. Clayton RN. Cardiovascular function in acromegaly. Endocr Rev. 2003;24(3):272-7.
- 108. Sacca L, Cittadini A, Fazio S. Growth hormone and the heart. Endocr Rev. 1994;15(5):555-73.
- 109. Dos Santos Silva CM, Lima GA, Volschan IC, Gottlieb I, Kasuki L, Neto LV, et al. Low risk of coronary artery disease in patients with acromegaly. Endocrine. 2015;50(3):749–55.
- 110. Gouya H, Vignaux O, Le Roux P, Chanson P, Bertherat J, Bertagna X, et al. Rapidly reversible myocardial edema in patients with acromegaly: assessment with ultrafast T2 mapping in a single-breath-hold MRI sequence. AJR Am J Roentgenol. 2008;190(6):1576–82.
- 111. Kormanyos A, Domsik P, Kalapos A, Valkusz Z, Lengyel C, Forster T, et al. Threedimensional speckle tracking echocardiography-derived left atrial deformation analysis in acromegaly (results from the MAGYAR-Path Study). Echocardiography. 2018;35(7):975–84.
- 112. Popielarz-Grygalewicz A, Gasior JS, Konwicka A, Grygalewicz P, Stelmachowska-Banas M, Zgliczynski W, et al. Heart in acromegaly: the echocardiographic characteristics of patients diagnosed with acromegaly in various stages of the disease. Int J Endocrinol. 2018;2018:6935054.
- 113. Sharma MD, Nguyen AV, Brown S, Robbins RJ. Cardiovascular disease in acromegaly. Methodist Debakey Cardiovasc J. 2017;13(2):64–7.
- 114. Kahaly G, Olshausen KV, Mohr-Kahaly S, Erbel R, Boor S, Beyer J, et al. Arrhythmia profile in acromegaly. Eur Heart J. 1992;13(1):51–6.

- 115. Warszawski L, Kasuki L, Sa R, Dos Santos Silva CM, Volschan I, Gottlieb I, et al. Low frequency of cardniac arrhythmias and lack of structural heart disease in medically-naive acromegaly patients: a prospective study at baseline and after 1 year of somatostatin analogs treatment. Pituitary. 2016;19(6):582–9.
- 116. Bihan H, Espinosa C, Valdes-Socin H, Salenave S, Young J, Levasseur S, et al. Long-term outcome of patients with acromegaly and congestive heart failure. J Clin Endocrinol Metab. 2004;89(11):5308–13.
- 117. Maison P, Tropeano AI, Macquin-Mavier I, Giustina A, Chanson P. Impact of somatostatin analogs on the heart in acromegaly: a metaanalysis. J Clin Endocrinol Metab. 2007;92(5):1743–7.
- 118. Maison P, Chanson P. Less is more risky? Growth hormone and insulin-like growth factor 1 levels and cardiovascular risk. Nat Clin Pract Endocrinol Metab. 2006;2(12):650–1.
- 119. Berg C, Petersenn S, Lahner H, Herrmann BL, Buchfelder M, Droste M, et al. Cardiovascular risk factors in patients with uncontrolled and long-term acromegaly: comparison with matched data from the general population and the effect of disease control. J Clin Endocrinol Metab. 2010;95(8):3648–56.
- 120. Boero L, Manavela M, Gomez Rosso L, Insua C, Berardi V, Fornari MC, et al. Alterations in biomarkers of cardiovascular disease (CVD) in active acromegaly. Clin Endocrinol. 2009;70(1):88–95.
- 121. Otsuki M, Kasayama S, Yamamoto H, Saito H, Sumitani S, Kouhara H, et al. Characterization of premature atherosclerosis of carotid arteries in acromegalic patients. Clin Endocrinol. 2001;54(6):791–6.
- 122. Paisley AN, Banerjee M, Rezai M, Schofield RE, Balakrishnannair S, Herbert A, et al. Changes in arterial stiffness but not carotid intimal thickness in acromegaly. J Clin Endocrinol Metab. 2011;96(5):1486–92.
- 123. Akutsu H, Kreutzer J, Wasmeier G, Ropers D, Rost C, Mohlig M, et al. Acromegaly per se does not increase the risk for coronary artery disease. Eur J Endocrinol. 2010;162(5):879–86.
- 124. Bogazzi F, Battolla L, Spinelli C, Rossi G, Gavioli S, Di Bello V, et al. Risk factors for development of coronary heart disease in patients with acromegaly: a five-year prospective study. J Clin Endocrinol Metab. 2007;92(11):4271–7.
- 125. Cannavo S, Almoto B, Cavalli G, Squadrito S, Romanello G, Vigo MT, et al. Acromegaly and coronary disease: an integrated evaluation of conventional coronary risk factors and coronary calcifications detected by computed tomography. J Clin Endocrinol Metab. 2006;91(10):3766–72.
- 126. Sardella C, Cappellani D, Urbani C, Manetti L, Marconcini G, Tomisti L, et al. Disease activity and lifestyle influence comorbidities and cardiovascular events in patients with acromegaly. Eur J Endocrinol. 2016;175(5):443–53.
- 127. Schofl C, Petroff D, Tonjes A, Grussendorf M, Droste M, Stalla G, et al. Incidence of myocardial infarction and stroke in acromegaly patients: results from the German Acromegaly Registry. Pituitary. 2017;20(6):635–42.
- 128. Parkinson C, Renehan AG, Ryder WD, O'Dwyer ST, Shalet SM, Trainer PJ. Gender and age influence the relationship between serum GH and IGF-I in patients with acromegaly. Clin Endocrinol. 2002;57(1):59–64.
- 129. Sesmilo G, Fairfield WP, Katznelson L, Pulaski K, Freda PU, Bonert V, et al. Cardiovascular risk factors in acromegaly before and after normalization of serum IGF-I levels with the GH antagonist pegvisomant. J Clin Endocrinol Metab. 2002;87(4):1692–9.
- 130. Verhelst J, Velkeniers B, Maiter D, Haentjens P, T'Sjoen G, Rietzschel E, et al. Active acromegaly is associated with decreased hs-CRP and NT-proBNP serum levels: insights from the Belgian registry of acromegaly. Eur J Endocrinol. 2013;168(2):177–84.
- 131. Colao A, Spinelli L, Marzullo P, Pivonello R, Petretta M, Di Somma C, et al. High prevalence of cardiac valve disease in acromegaly: an observational, analytical, case-control study. J Clin Endocrinol Metab. 2003;88(7):3196–201.

- 132. Pereira AM, van Thiel SW, Lindner JR, Roelfsema F, van der Wall EE, Morreau H, et al. Increased prevalence of regurgitant valvular heart disease in acromegaly. J Clin Endocrinol Metab. 2004;89(1):71–5.
- 133. Maione L, Garcia C, Bouchachi A, Kallel N, Maison P, Salenave S, et al. No evidence of a detrimental effect of cabergoline therapy on cardiac valves in patients with acromegaly. J Clin Endocrinol Metab. 2012;97(9):E1714–9.
- 134. Alexopoulou O, Bex M, Kamenicky P, Mvoula AB, Chanson P, Maiter D. Prevalence and risk factors of impaired glucose tolerance and diabetes mellitus at diagnosis of acromegaly: a study in 148 patients. Pituitary. 2014;17(1):81–9.
- 135. Freda PU, Shen W, Heymsfield SB, Reyes-Vidal CM, Geer EB, Bruce JN, et al. Lower visceral and subcutaneous but higher intermuscular adipose tissue depots in patients with growth hormone and insulin-like growth factor I excess due to acromegaly. J Clin Endocrinol Metab. 2008;93(6):2334–43.
- 136. Katznelson L. Alterations in body composition in acromegaly. Pituitary. 2009;12:136-42.
- 137. Calan M, Demirpence M. Increased circulating levels of irisin are associated with cardiovascular risk factors in subjects with acromegaly. Hormones (Athens). 2019;18:435.
- Mercado M, Ramirez-Renteria C. Metabolic complications of acromegaly. Front Horm Res. 2018;49:20–8.
- 139. Briet C, Ilie MD, Kuhn E, Maione L, Brailly-Tabard S, Salenave S, et al. Changes in metabolic parameters and cardiovascular risk factors after therapeutic control of acromegaly vary with the treatment modality. Data from the Bicetre cohort, and review of the literature. Endocrine. 2019;63(2):348–60.
- 140. Kamenicky P, Blanchard A, Gauci C, Salenave S, Letierce A, Lombes M, et al. Pathophysiology of renal calcium handling in acromegaly: what lies behind hypercalciuria? J Clin Endocrinol Metab. 2012;97(6):2124–33.
- 141. Attal P, Chanson P. Endocrine aspects of obstructive sleep apnea. J Clin Endocrinol Metab. 2010;95(2):483–95.
- 142. Herrmann BL, Wessendorf TE, Ajaj W, Kahlke S, Teschler H, Mann K. Effects of octreotide on sleep apnoea and tongue volume (magnetic resonance imaging) in patients with acromegaly. Eur J Endocrinol. 2004;151(3):309–15.
- 143. Attal P, Claes V, Bobin S, Chanson P, Kamenicky P, Zizzari P, et al. Growth hormone excess and sternohyoid muscle mechanics in rats. Eur Respir J. 2009;34(4):967–74.
- 144. Ip MSM, Tan KCB, Peh WCG, Lam KSL. Effect of Sandostatin® LAR® on sleep apneoa in acromegaly: correlation with computerized tomographic cephalometry and hormonal activity. Clin Endocrinol. 2001;55:477–83.
- 145. Annamalai AK, Webb A, Kandasamy N, Elkhawad M, Moir S, Khan F, et al. A comprehensive study of clinical, biochemical, radiological, vascular, cardiac, and sleep parameters in an unselected cohort of patients with acromegaly undergoing presurgical somatostatin receptor ligand therapy. J Clin Endocrinol Metab. 2013;98(3):1040–50.
- 146. Briet C, Salenave S, Bonneville JF, Laws ER, Chanson P. Pituitary apoplexy. Endocr Rev. 2015;36(6):622–45.
- 147. Boguszewski CL, Boguszewski M. Growth Hormone's links to cancer. Endocr Rev. 2019;40(2):558–74.
- 148. Dal J, Leisner MZ, Hermansen K, Farkas DK, Bengtsen M, Kistorp C, et al. Cancer incidence in patients with acromegaly: a cohort study and meta-analysis of the literature. J Clin Endocrinol Metab. 2018;103(6):2182–8.
- 149. Jenkins PJ, Besser M. Clinical perspective: acromegaly and cancer: a problem. J Clin Endocrinol Metab. 2001;86(7):2935–41.
- 150. Lois K, Bukowczan J, Perros P, Jones S, Gunn M, James RA. The role of colonoscopic screening in acromegaly revisited: review of current literature and practice guidelines. Pituitary. 2015;18(4):568–74.
- 151. Melmed S. Acromegaly and cancer: not a problem? J Clin Endocrinol Metab. 2001;86(7):2929–34.

- 152. Renehan AG, O'Connell J, O'Halloran D, Shanahan F, Potten CS, O'Dwyer ST, et al. Acromegaly and colorectal cancer: a comprehensive review of epidemiology, biological mechanisms, and clinical implications. Horm Metab Res. 2003;35(11–12):712–25.
- 153. Renehan AG, Brennan BM. Acromegaly, growth hormone and cancer risk. Best Pract Res Clin Endocrinol Metab. 2008;22(4):639–57.
- 154. Loeper S, Ezzat S. Acromegaly: re-thinking the cancer risk. Rev Endocr Metab Disord. 2008;9(1):41–58.
- 155. Delhougne B, Deneux C, Abs R, Chanson P, Fierens H, Laurent-Puig P, et al. The prevalence of colonic polyps in acromegaly : a prospective colonoscopic and pathological study in 103 patients. J Clin Endocrinol Metab. 1995;80:3223–6.
- 156. Cairns SR, Scholefield JH, Steele RJ, Dunlop MG, Thomas HJ, Evans GD, et al. Guidelines for colorectal cancer screening and surveillance in moderate and high risk groups (update from 2002). Gut. 2010;59(5):666–89.
- 157. Wolinski K, Czarnywojtek A, Ruchala M. Risk of thyroid nodular disease and thyroid cancer in patients with acromegaly—meta-analysis and systematic review. PLoS One. 2014;9(2):e88787.
- 158. Kauppinen-Makelin R, Sane T, Valimaki MJ, Markkanen H, Niskanen L, Ebeling T, et al. Increased cancer incidence in acromegaly—a nationwide survey. Clin Endocrinol. 2010;72(2):278–9.
- 159. dos Santos MC, Nascimento GC, Nascimento AG, Carvalho VC, Lopes MH, Montenegro R, et al. Thyroid cancer in patients with acromegaly: a case-control study. Pituitary. 2013;16(1):109–14.
- Boguszewski CL, Ayuk J. Management of endocrine disease: acromegaly and cancer: an old debate revisited. Eur J Endocrinol. 2016;175(4):R147–56.
- 161. Katznelson L, Laws ER Jr, Melmed S, Molitch ME, Murad MH, Utz A, et al. Acromegaly: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2014;99(11):3933–51.
- 162. Engelhardt J, Nunes ML, Pouchieu C, Ferriere A, San-Galli F, Gimbert E, et al. Increased incidence of intracranial meningiomas in patients with acromegaly. Neurosurgery. 2019;87:639.
- 163. Wei R, Jiang C, Gao J, Xu P, Zhang D, Sun Z, et al. Deep-learning approach to automatic identification of facial anomalies in endocrine disorders. Neuroendocrinology. 2019;110:328.
- 164. Clemmons DR. Consensus statement on the standardization and evaluation of growth hormone and insulin-like growth factor assays. Clin Chem. 2011;57(4):555–9.
- Giustina A, Chanson P, Bronstein MD, Klibanski A, Lamberts S, Casanueva FF, et al. A consensus on criteria for cure of acromegaly. J Clin Endocrinol Metab. 2010;95(7):3141–8.
- 166. Carmichael JD, Bonert VS, Mirocha JM, Melmed S. The utility of oral glucose tolerance testing for diagnosis and assessment of treatment outcomes in 166 patients with acromegaly. J Clin Endocrinol Metab. 2009;94(2):523–7.
- 167. Giustina A, Barkan A, Casanueva FF, Cavagnini F, Frohman L, Ho K, et al. Criteria for cure of acromegaly: a consensus statement. J Clin Endocrinol Metab. 2000;85(2):526–9.
- 168. Trainer PJ. Editorial: acromegaly–consensus, what consensus? J Clin Endocrinol Metab. 2002;87(8):3534–6.
- 169. Schilbach K, Gar C, Lechner A, Nicolay SS, Schwerdt L, Haenelt M, et al. Determinants of the growth hormone nadir during oral glucose tolerance test in adults. Eur J Endocrinol. 2019;181(1):55–67.
- 170. Scaroni C, Albiger N, Daniele A, Dassie F, Romualdi C, Vazza G, et al. Paradoxical GH increase during OGTT is associated with first-generation somatostatin analog responsiveness in acromegaly. J Clin Endocrinol Metab. 2019;104(3):856–62.
- 171. Hage M, Kamenicky P, Chanson P. Growth hormone response to Oral glucose load: from normal to pathological conditions. Neuroendocrinology. 2019;108(3):244–55.
- 172. Chanson P, Arnoux A, Mavromati M, Brailly-Tabard S, Massart C, Young J, et al. Reference values for IGF-I serum concentrations: comparison of six immunoassays. J Clin Endocrinol Metab. 2016;101(9):3450–8.
- 173. Mavromati M, Kuhn E, Agostini H, Brailly-Tabard S, Massart C, Piketty ML, et al. Classification of patients with GH disorders may vary according to the IGF-I assay. J Clin Endocrinol Metab. 2017;102(8):2844–52.

- 174. Chakraborty PP, Bhattacharjee R, Mukhopadhyay S, Chowdhury S. Pseudoacromegaly in pachydermoperiostosis. BMJ Case Rep. 2016;2016
- Chakraborty PP, Datta S, Mukhopadhyay S, Chowdhury S. Pseudoacromegaly in congenital generalised lipodystrophy (Berardinelli-Seip syndrome). BMJ Case Rep. 2016;2016 https:// doi.org/10.1136/bcr-2016-214493.
- 176. Dahlqvist P, Spencer R, Marques P, Dang MN, Glad CAM, Johannsson G, et al. Pseudoacromegaly: a differential diagnostic problem for acromegaly with a genetic solution. J Endocr Soc. 2017;1(8):1104–9.
- 177. Marques P, Korbonits M. Pseudoacromegaly. Front Neuroendocrinol. 2019;52:113-43.
- 178. Kovacs K, Lloyd R, Horvath E, Asa SL, Stefaneanu L, Killinger DW, et al. Silent somatotroph adenomas of the human pituitary. A morphologic study of three cases including immunocytochemistry, electron microscopy, in vitro examination, and in situ hybridization. Am J Pathol. 1989;134(2):345–53.
- 179. Dimaraki EV, Jaffe CA, DeMott-Friberg R, Chandler WF, Barkan AL. Acromegaly with apparently normal GH secretion: implications for diagnosis and follow-up. J Clin Endocrinol Metab. 2002;87(8):3537–42.
- 180. Potorac I, Petrossians P, Daly AF, Alexopoulou O, Borot S, Sahnoun-Fathallah M, et al. T2-weighted MRI signal predicts hormone and tumor responses to somatostatin analogs in acromegaly. Endocr Relat Cancer. 2016;23(11):871–81.
- 181. Rodriguez-Barcelo S, Gutierrez-Cardo A, Dominguez-Paez M, Medina-Imbroda J, Romero-Moreno L, Arraez-Sanchez M. Clinical usefulness of coregistered 11C-methionine positron emission tomography/3-T magnetic resonance imaging at the follow-up of acromegaly. World Neurosurg. 2014;82(3–4):468–73.
- 182. Feng Z, He D, Mao Z, Wang Z, Zhu Y, Zhang X, et al. Utility of 11C-methionine and 18F-FDG PET/CT in patients with functioning pituitary adenomas. Clin Nucl Med. 2016;41(3):e130–4.
- 183. Putzer D, Gabriel M, Kendler D, Henninger B, Knoflach M, Kroiss A, et al. Comparison of (68)Ga-DOTA-Tyr(3)-octreotide and (18)F-fluoro-L-dihydroxyphenylalanine positron emission tomography in neuroendocrine tumor patients. Q J Nucl Med Mol Imaging. 2010;54(1):68–75.
- Chanson P, Kamenicky P. Treatment of acromegaly: a critical analysis of the last ten years. Ann Endocrinol (Paris). 2012;73(2):99–106.
- Colao A, Grasso LFS, Giustina A, Melmed S, Chanson P, Pereira AM, et al. Acromegaly. Nat Rev Dis Primers. 2019;5(1):20.
- 186. Melmed S, Colao A, Barkan A, Molitch M, Grossman AB, Kleinberg D, et al. Guidelines for acromegaly management: an update. J Clin Endocrinol Metab. 2009;94(5):1509–17.
- Melmed S, Bronstein MD, Chanson P, Klibanski A, Casanueva FF, Wass JAH, et al. A consensus statement on acromegaly therapeutic outcomes. Nat Rev Endocrinol. 2018;14(9):552–61.
- Sherlock M, Woods C, Sheppard MC. Medical therapy in acromegaly. Nat Rev Endocrinol. 2011;7(5):291–300.
- Melmed S, Casanueva FF, Cavagnini F, Chanson P, Frohman L, Grossman A, et al. Guidelines for acromegaly management. J Clin Endocrinol Metab. 2002;87(9):4054–8.
- Biermasz NR, van Dulken H, Roelfsema F. Long-term follow-up results of postoperative radiotherapy in 36 patients with acromegaly. J Clin Endocrinol Metab. 2000;85(7):2476–82.
- 191. Fahlbusch R, Buchfelder M, Nomikos P. Pituitary surgery. In: Melmed S, editor. The pituitary. 2nd ed. Malden, MA: Blackwell Science Inc.; 2002. p. 405–18.
- 192. Jane JA Jr, Starke RM, Elzoghby MA, Reames DL, Payne SC, Thorner MO, et al. Endoscopic transsphenoidal surgery for acromegaly: remission using modern criteria, complications, and predictors of outcome. J Clin Endocrinol Metab. 2011;96(9):2732–40.
- 193. Nomikos P, Buchfelder M, Fahlbusch R. The outcome of surgery in 668 patients with acromegaly using current criteria of biochemical 'cure'. Eur J Endocrinol. 2005;152(3):379–87.
- 194. Swearingen B, Barker FG, Katznelson L, Biller BM, Grinspoon S, Klibanski A, et al. Longterm mortality after transphenoidal surgery and adjunctive therapy for acromegaly. J Clin Endocrinol Metab. 1998;83(10):3419–26.
- Cappabianca P, Cavallo LM, de Divitiis E. Endoscopic endonasal transsphenoidal surgery. Neurosurgery. 2004;55(4):933–40. discussion 40-1

- 196. Shih HA, Loeffler JS. Radiation therapy in acromegaly. Rev Endocr Metab Disord. 2008;9(1):59–65.
- 197. Minniti G, Flickinger J. The risk/benefit ratio of radiotherapy in pituitary tumors. Best Pract Res Clin Endocrinol Metab. 2019;33(2):101269.
- 198. Barrande G, Pittino-Lungo M, Coste J, Ponvert D, Bertagna X, Luton JP, et al. Hormonal and metabolic effects of radiotherapy in acromegaly: long-term results in 128 patients followed in a single center [in process citation]. J Clin Endocrinol Metab. 2000;85(10):3779–85.
- 199. Minniti G, Jaffrain-Rea ML, Osti M, Esposito V, Santoro A, Solda F, et al. The long-term efficacy of conventional radiotherapy in patients with GH-secreting pituitary adenomas. Clin Endocrinol. 2005;62(2):210–6.
- Loeffler JS, Shih HA. Radiation therapy in the management of pituitary adenomas. J Clin Endocrinol Metab. 2011;96(7):1992–2003.
- 201. Jenkins PJ, Bates P, Carson MN, Stewart PM, Wass JA. Conventional pituitary irradiation is effective in lowering serum growth hormone and insulin-like growth factor-I in patients with acromegaly. J Clin Endocrinol Metab. 2006;91(4):1239–45.
- 202. Castinetti F, Taieb D, Kuhn JM, Chanson P, Tamura M, Jaquet P, et al. Outcome of gamma knife radiosurgery in 82 patients with acromegaly: correlation with initial hypersecretion. J Clin Endocrinol Metab. 2005;90(8):4483–8.
- 203. Yang I, Kim W, De Salles A, Bergsneider M. A systematic analysis of disease control in acromegaly treated with radiosurgery. Neurosurg Focus. 2010;29(4):E13.
- Castinetti F, Nagai M, Morange I, Dufour H, Caron P, Chanson P, et al. Long-term results of stereotactic radiosurgery in secretory pituitary adenomas. J Clin Endocrinol Metab. 2009;94(9):3400–7.
- 205. Gheorghiu ML. Updates in outcomes of stereotactic radiation therapy in acromegaly. Pituitary. 2017;20(1):154-68.
- 206. Brada M, Burchell L, Ashley S, Traish D. The incidence of cerebrovascular accidents in patients with pituitary adenoma. Int J Radiat Oncol Biol Phys. 1999;45(3):693–8.
- 207. Minniti G, Scaringi C, Maurizi ER. Radiation techniques for acromegaly. Radiat Oncol. 2011;6(1):167.
- 208. Sherlock M, Ayuk J, Tomlinson JW, Toogood AA, Aragon-Alonso A, Sheppard MC, et al. Mortality in patients with pituitary disease. Endocr Rev. 2010;31(3):301–42.
- 209. Abs R, Verhelst J, Maiter D, Van Acker K, Nobels F, Coolens JL, et al. Cabergoline in the treatment of acromegaly: a study in 64 patients. J Clin Endocrinol Metab. 1998;83(2):374–8.
- 210. Newman CB. Medical therapy for acromegaly. Endocrinol Metab Clin N Am. 1999;28(1):171–90.
- Sandret L, Maison P, Chanson P. Place of cabergoline in acromegaly: a meta-analysis. J Clin Endocrinol Metab. 2011;96(5):1327–35.
- Valassi E, Klibanski A, Biller BM. Clinical review#: potential cardiac valve effects of dopamine agonists in hyperprolactinemia. J Clin Endocrinol Metab. 2010;95(3):1025–33.
- 213. Kuhn E, Chanson P. Cabergoline in acromegaly. Pituitary. 2017;20(1):121-8.
- 214. Schaer JC, Waser B, Mengod G, Reubi JC. Somatostatin receptor subtypes sst1, sst2, sst3 and sst5 expression in human pituitary, gastroentero-pancreatic and mammary tumors: comparison of mRNA analysis with receptor autoradiography. Int J Cancer. 1997;70(5):530–7.
- 215. Lamberts SW, van der Lely AJ, de Herder WW, Hofland LJ. Octreotide. N Engl J Med. 1996;334(4):246–54.
- Murray RD, Melmed S. A critical analysis of clinically available somatostatin analog formulations for therapy of acromegaly. J Clin Endocrinol Metab. 2008;93(8):2957–68.
- Bevan JS. Clinical review: the antitumoral effects of somatostatin analog therapy in acromegaly. J Clin Endocrinol Metab. 2005;90(3):1856–63.
- Freda PU, Katznelson L, van der Lely AJ, Reyes CM, Zhao S, Rabinowitz D. Long-acting somatostatin analog therapy of acromegaly: a meta-analysis. J Clin Endocrinol Metab. 2005;90(8):4465–73.
- Carmichael JD, Bonert VS, Nuno M, Ly D, Melmed S. Acromegaly clinical trial methodology impact on reported biochemical efficacy rates of somatostatin receptor ligand treatments: a meta-analysis. J Clin Endocrinol Metab. 2014;99(5):1825–33.

- Ayuk J, Stewart SE, Stewart PM, Sheppard MC. Long-term safety and efficacy of depot long-acting somatostatin analogs for the treatment of acromegaly. J Clin Endocrinol Metab. 2002;87(9):4142–6.
- 221. Cozzi R, Attanasio R, Montini M, Pagani G, Lasio G, Lodrini S, et al. Four-year treatment with octreotide-long-acting repeatable in 110 acromegalic patients: predictive value of short-term results? J Clin Endocrinol Metab. 2003;88(7):3090–8.
- 222. Maiza JC, Vezzosi D, Matta M, Donadille F, Loubes-Lacroix F, Cournot M, et al. Long-term (up to 18 years) effects on GH/IGF-1 hypersecretion and tumour size of primary somatostatin analogue (SSTa) therapy in patients with GH-secreting pituitary adenoma responsive to SSTa. Clin Endocrinol. 2007;67(2):282–9.
- 223. Ramirez C, Vargas G, Gonzalez B, Grossman A, Rabago J, Sosa E, et al. Discontinuation of octreotide LAR after long term, successful treatment of patients with acromegaly: is it worth trying? Eur J Endocrinol. 2012;166(1):21–6.
- 224. Ronchi CL, Rizzo E, Lania AG, Pivonello R, Grottoli S, Colao A, et al. Preliminary data on biochemical remission of acromegaly after somatostatin analogs withdrawal. Eur J Endocrinol. 2008;158(1):19–25.
- 225. Giustina A, Mazziotti G, Torri V, Spinello M, Floriani I, Melmed S. Meta-analysis on the effects of octreotide on tumor mass in acromegaly. PLoS One. 2012;7(5):e36411.
- 226. Colao A, Ferone D, Marzullo P, Cappabianca P, Cirillo S, Boerlin V, et al. Long-term effects of depot long-acting somatostatin analog octreotide on hormone levels and tumor mass in acromegaly. J Clin Endocrinol Metab. 2001;86(6):2779–86.
- 227. Attanasio R, Mainolfi A, Grimaldi F, Cozzi R, Montini M, Carzaniga C, et al. Somatostatin analogs and gallstones: a retrospective survey on a large series of acromegalic patients. J Endocrinol Investig. 2008;31(8):704–10.
- 228. Chanson P, Bertherat J, Beckers A, Bihan H, Brue T, Caron P, et al. French consensus on the management of acromegaly. Ann Endocrinol (Paris). 2009;70(2):92–106.
- Cozzolino A, Feola T, Simonelli I, Puliani G, Pozza C, Giannetta E, et al. Somatostatin analogs and glucose metabolism in acromegaly: a meta-analysis of prospective interventional studies. J Clin Endocrinol Metab. 2018;
- Mazziotti G, Floriani I, Bonadonna S, Torri V, Chanson P, Giustina A. Effects of somatostatin analogs on glucose homeostasis: a metaanalysis of acromegaly studies. J Clin Endocrinol Metab. 2009;94(5):1500–8.
- 231. Bruns C, Lewis I, Briner U, Meno-Tetang G, Weckbecker G. SOM230: a novel somatostatin peptidomimetic with broad somatotropin release inhibiting factor (SRIF) receptor binding and a unique antisecretory profile. Eur J Endocrinol. 2002;146(5):707–16.
- 232. Colao A, Bronstein MD, Freda P, Gu F, Shen CC, Gadelha M, et al. Pasireotide versus octreotide in acromegaly: a head-to-head superiority study. J Clin Endocrinol Metab. 2014;99(3):791–9.
- 233. Gadelha MR, Bronstein MD, Brue T, Coculescu M, Fleseriu M, Guitelman M, et al. Pasireotide versus continued treatment with octreotide or lanreotide in patients with inadequately controlled acromegaly (PAOLA): a randomised, phase 3 trial. Lancet Diabetes Endocrinol. 2014;2(11):875–84.
- 234. Petersenn S, Schopohl J, Barkan A, Mohideen P, Colao A, Abs R, et al. Pasireotide (SOM230) demonstrates efficacy and safety in patients with acromegaly: a randomized, multicenter, phase II trial. J Clin Endocrinol Metab. 2010;95(6):2781–9.
- 235. Bronstein MD, Fleseriu M, Neggers S, Colao A, Sheppard M, Gu F, et al. Switching patients with acromegaly from octreotide to pasireotide improves biochemical control: crossover extension to a randomized, double-blind, Phase III study. BMC Endocr Disord. 2016;16:16.
- 236. Heck A, Ringstad G, Fougner SL, Casar-Borota O, Nome T, Ramm-Pettersen J, et al. Intensity of pituitary adenoma on T2-weighted magnetic resonance imaging predicts the response to octreotide treatment in newly diagnosed acromegaly. Clin Endocrinol. 2012;77(1):72–8.
- 237. Puig-Domingo M, Resmini E, Gomez-Anson B, Nicolau J, Mora M, Palomera E, et al. Magnetic resonance imaging as a predictor of response to somatostatin analogs in acromegaly after surgical failure. J Clin Endocrinol Metab. 2010;95(11):4973–8.

- 238. Gatto F, Feelders RA, van der Pas R, Kros JM, Waaijers M, Sprij-Mooij D, et al. Immunoreactivity score using an anti-sst2A receptor monoclonal antibody strongly predicts the biochemical response to adjuvant treatment with somatostatin analogs in acromegaly. J Clin Endocrinol Metab. 2013;98(1):E66–71.
- Kopchick JJ, Parkinson C, Stevens EC, Trainer PJ. Growth hormone receptor antagonists: discovery, development, and use in patients with acromegaly. Endocr Rev. 2002;23(5):623–46.
- 240. Trainer PJ, Drake WM, Katznelson L, Freda PU, Herman-Bonert V, van der Lely AJ, et al. Treatment of acromegaly with the growth hormone-receptor antagonist pegvisomant. N Engl J Med. 2000;342(16):1171–7.
- 241. van der Lely AJ, Hutson RK, Trainer PJ, Besser GM, Barkan AL, Katznelson L, et al. Longterm treatment of acromegaly with pegvisomant, a growth hormone receptor antagonist. Lancet. 2001;358(9295):1754–9.
- 242. Buchfelder M, van der Lely AJ, Biller BMK, Webb SM, Brue T, Strasburger CJ, et al. Long-term treatment with pegvisomant: observations from 2090 acromegaly patients in ACROSTUDY. Eur J Endocrinol. 2018;179(6):419–27.
- 243. Colao A, Pivonello R, Auriemma RS, De Martino MC, Bidlingmaier M, Briganti F, et al. Efficacy of 12-month treatment with the GH receptor antagonist pegvisomant in patients with acromegaly resistant to long-term, high-dose somatostatin analog treatment: effect on IGF-I levels, tumor mass, hypertension and glucose tolerance. Eur J Endocrinol. 2006;154(3):467–77.
- 244. Marazuela M, Lucas T, Alvarez-Escola C, Puig-Domingo M, de la Torre NG, de Miguel-Novoa P, et al. Long-term treatment of acromegalic patients resistant to somatostatin analogues with the GH receptor antagonist pegvisomant: its efficacy in relation to gender and previous radiotherapy. Eur J Endocrinol. 2009;160(4):535–42.
- 245. Moore DJ, Adi Y, Connock MJ, Bayliss S. Clinical effectiveness and cost-effectiveness of pegvisomant for the treatment of acromegaly: a systematic review and economic evaluation. BMC Endocr Disord. 2009;9:20.
- 246. Schreiber I, Buchfelder M, Droste M, Forssmann K, Mann K, Saller B, et al. Treatment of acromegaly with the GH receptor antagonist pegvisomant in clinical practice: safety and efficacy evaluation from the German Pegvisomant observational study. Eur J Endocrinol. 2007;156(1):75–82.
- 247. van der Lely AJ, Biller BM, Brue T, Buchfelder M, Ghigo E, Gomez R, et al. Long-term safety of pegvisomant in patients with acromegaly: comprehensive review of 1288 subjects in ACROSTUDY. J Clin Endocrinol Metab. 2012;97(5):1589–97.
- 248. Jimenez C, Burman P, Abs R, Clemmons DR, Drake WM, Hutson KR, et al. Follow-up of pituitary tumor volume in patients with acromegaly treated with pegvisomant in clinical trials. Eur J Endocrinol. 2008;159(5):517–23.
- 249. van der Lely AJ, Muller A, Janssen JA, Davis RJ, Zib KA, Scarlett JA, et al. Control of tumor size and disease activity during cotreatment with octreotide and the growth hormone receptor antagonist pegvisomant in an acromegalic patient. J Clin Endocrinol Metab. 2001;86(2):478–81.
- 250. Feola T, Cozzolino A, Simonelli I, Sbardella E, Pozza C, Giannetta E, et al. Pegvisomant improves glucose metabolism in acromegaly: a meta-analysis of prospective interventional studies. J Clin Endocrinol Metab. 2019;104(7):2892–902.
- 251. Bernabeu I, Cameselle-Teijeiro J, Casanueva FF, Marazuela M. Pegvisomant-induced cholestatic hepatitis with jaundice in a patient with Gilbert's syndrome. Eur J Endocrinol. 2009;160(5):869–72.
- 252. Bernabeu I, Marazuela M, Lucas T, Loidi L, Alvarez-Escola C, Luque-Ramirez M, et al. Pegvisomant-induced liver injury is related to the UGT1A1*28 polymorphism of Gilbert's syndrome. J Clin Endocrinol Metab. 2010;95(5):2147–54.
- 253. Filopanti M, Barbieri AM, Mantovani G, Corbetta S, Gasco V, Ragonese M, et al. Role of UGT1A1 and ADH gene polymorphisms in pegvisomant-induced liver toxicity in acromegalic patients. Eur J Endocrinol. 2014;170(2):249–56.

- 254. Feenstra J, de Herder WW, ten Have SM, van den Beld AW, Feelders RA, Janssen JA, et al. Combined therapy with somatostatin analogues and weekly pegvisomant in active acromegaly. Lancet. 2005;365(9471):1644–6.
- 255. Neggers SJ, van Aken MO, Janssen JA, Feelders RA, de Herder WW, van der Lely AJ. Longterm efficacy and safety of combined treatment of somatostatin analogs and pegvisomant in acromegaly. J Clin Endocrinol Metab. 2007;92(12):4598–601.
- 256. Jorgensen JO, Feldt-Rasmussen U, Frystyk J, Chen JW, Kristensen LO, Hagen C, et al. Cotreatment of acromegaly with a somatostatin analog and a growth hormone receptor antagonist. J Clin Endocrinol Metab. 2005;90(10):5627–31.
- 257. Neggers SJ, de Herder WW, Janssen JA, Feelders RA, van der Lely AJ. Combined treatment for acromegaly with long-acting somatostatin analogs and pegvisomant: long-term safety for up to 4.5 years (median 2.2 years) of follow-up in 86 patients. Eur J Endocrinol. 2009;160(4):529–33.
- 258. Neggers SJCMM, van Aken MO, de Herder WW, Feelders RA, Janssen JAMJL, Badia X, et al. Quality of life in acromegalic patients during long-term somatostatin analog treatment with and without pegvisomant. J Clin Endocrinol Metab. 2008;93(10):3853–9.
- Neggers SJ, van der Lely AJ. Somatostatin analog and pegvisomant combination therapy for acromegaly. Nat Rev Endocrinol. 2009;5(10):546–52.
- 260. Higham CE, Atkinson AB, Aylwin S, Bidlingmaier M, Drake WM, Lewis A, et al. Effective combination treatment with cabergoline and low-dose pegvisomant in active acromegaly: a prospective clinical trial. J Clin Endocrinol Metab. 2012;97(4):1187–93.
- 261. Bernabeu I, Alvarez-Escola C, Paniagua AE, Lucas T, Pavon I, Cabezas-Agricola JM, et al. Pegvisomant and cabergoline combination therapy in acromegaly. Pituitary. 2013;16(1):101–8.
- 262. Leonart LP, Ferreira VL, Tonin FS, Fernandez-Llimos F, Pontarolo R. Medical treatments for acromegaly: a systematic review and network meta-analysis. Value Health. 2018;21(7):874–80.
- 263. Attanasio R, Baldelli R, Pivonello R, Grottoli S, Bocca L, Gasco V, et al. Lanreotide 60 mg, a new long-acting formulation: effectiveness in the chronic treatment of acromegaly. J Clin Endocrinol Metab. 2003;88(11):5258–65.
- 264. Bevan JS, Atkin SL, Atkinson AB, Bouloux PM, Hanna F, Harris PE, et al. Primary medical therapy for acromegaly: an open, prospective, multicenter study of the effects of subcutaneous and intramuscular slow- release octreotide on growth hormone, insulin-like growth factor-I, and tumor size. J Clin Endocrinol Metab. 2002;87(10):4554–63.
- 265. Caron PJ, Bevan JS, Petersenn S, Flanagan D, Tabarin A, Prevost G, et al. Tumor shrinkage with lanreotide Autogel 120 mg as primary therapy in acromegaly: results of a prospective multicenter clinical trial. J Clin Endocrinol Metab. 2014;99(4):1282–90.
- 266. Newman CB, Melmed S, George A, Torigian D, Duhaney M, Snyder P, et al. Octreotide as primary therapy for acromegaly. J Clin Endocrinol Metab. 1998;83(9):3034–40.
- 267. Sheppard MC. Primary medical therapy for acromegaly. Clin Endocrinol. 2003;58(4):387–99.
- 268. Abe T, Ludecke DK. Effects of preoperative octreotide treatment on different subtypes of 90 GH-secreting pituitary adenomas and outcome in one surgical centre. Eur J Endocrinol. 2001;145(2):137–45.
- Barkan AL, Lloyd RV, Chandler WF, Hatfield MK, Gebarski SS, Kelch RP, et al. Preoperative treatment of acromegaly with long-acting somatostatin analog SMS 201-995: shrinkage of invasive pituitary macroadenomas and improved surgical remission rate. J Clin Endocrinol Metab. 1988;67(5):1040–8.
- 270. Carlsen SM, Lund-Johansen M, Schreiner T, Aanderud S, Johannesen O, Svartberg J, et al. Preoperative octreotide treatment in newly diagnosed acromegalic patients with macroadenomas increases cure short-term postoperative rates: a prospective, randomized trial. J Clin Endocrinol Metab. 2008;93(8):2984–90.
- 271. Colao A, Merola B, Ferone D, Lombardi G. Acromegaly. J Clin Endocrinol Metab. 1997;82(9):2777–81.

- 272. Lucas T, Astorga R, Catala M. Preoperative lanreotide treatment for GH-secreting pituitary adenomas: effect on tumour volume and predictive factors of significant tumour shrinkage. Clin Endocrinol. 2003;58(4):471–81.
- 273. Mao ZG, Zhu YH, Tang HL, Wang DY, Zhou J, He DS, et al. Preoperative lanreotide treatment in acromegalic patients with macroadenomas increases short-term postoperative cure rates: a prospective, randomised trial. Eur J Endocrinol. 2010;162(4):661–6.
- Stevenaert A, Harris AG, Kovacs K, Beckers A. Presurgical octreotide treatment in acromegaly. Metabolism. 1992;41(9 Suppl 2):51–8.
- 275. Biermasz NR, van Dulken H, Roelfsema F. Direct postoperative and follow-up results of transsphenoidal surgery in 19 acromegalic patients pretreated with octreotide compared to those in untreated matched controls. J Clin Endocrinol Metab. 1999;84(10):3551–5.
- 276. Kristof RA, Stoffet-Wagner B, Klingmüller D, Schramm J. Does Octréotide treatment improve the surgical results of macroadenomas in acromegaly ? A randomised study. Acta Neurochir. 1999;141:399–405.
- 277. Losa M, Mortini P, Urbaz L, Ribotto P, Castrignano T, Giovanelli M. Presurgical treatment with somatostatin analogs in patients with acromegaly: effects on the remission and complication rates. J Neurosurg. 2006;104(6):899–906.
- 278. Pita-Gutierrez F, Pertega-Diaz S, Pita-Fernandez S, Pena L, Lugo G, Sangiao-Alvarellos S, et al. Place of preoperative treatment of acromegaly with somatostatin analog on surgical outcome: a systematic review and meta-analysis. PLoS One. 2013;8(4):e61523.
- 279. Yang C, Li G, Jiang S, Bao X, Wang R. Preoperative somatostatin analogues in patients with newly-diagnosed acromegaly: a systematic review and meta-analysis of comparative studies. Sci Rep. 2019;9(1):14070.
- 280. Carmichael JD, Broder MS, Cherepanov D, Chang E, Mamelak A, Said Q, et al. Long-term treatment outcomes of acromegaly patients presenting biochemically-uncontrolled at a tertiary pituitary center. BMC Endocr Disord. 2017;17(1):49.
- 281. Karavitaki N, Turner HE, Adams CB, Cudlip S, Byrne JV, Fazal-Sanderson V, et al. Surgical debulking of pituitary macroadenomas causing acromegaly improves control by lanreotide. Clin Endocrinol. 2008;68(6):970–5.
- 282. Petrossians P, Borges-Martins L, Espinoza C, Daly A, Betea D, Valdes-Socin H, et al. Gross total resection or debulking of pituitary adenomas improves hormonal control of acromegaly by somatostatin analogs. Eur J Endocrinol. 2005;152(1):61–6.
- 283. Giustina A, Bonadonna S, Bugari G, Colao A, Cozzi R, Cannavo S, et al. High-dose intramuscular octreotide in patients with acromegaly inadequately controlled on conventional somatostatin analogue therapy: a randomised controlled trial. Eur J Endocrinol. 2009;161(2):331–8.
- Giustina A, Mazziotti G, Cannavo S, Castello R, Arnaldi G, Bugari G, et al. High-dose and high-frequency lanreotide autogel in acromegaly: a randomized, multicenter study. J Clin Endocrinol Metab. 2017;102(7):2454–64.
- 285. Bolfi F, Neves AF, Boguszewski CL, Nunes-Nogueira VS. Mortality in acromegaly decreased in the last decade: a systematic review and meta-analysis. Eur J Endocrinol. 2018;179(1):59–71.
- Dekkers OM, Biermasz NR, Pereira AM, Romijn JA, Vandenbroucke JP. Mortality in acromegaly: a metaanalysis. J Clin Endocrinol Metab. 2008;93(1):61–7.
- Ayuk J, McGregor EJ, Mitchell RD, Gittoes NJ. Acute management of pituitary apoplexy surgery or conservative management? Clin Endocrinol. 2004;61(6):747–52.
- Kauppinen-Makelin R, Sane T, Reunanen A, Valimaki MJ, Niskanen L, Markkanen H, et al. A nationwide survey of mortality in acromegaly. J Clin Endocrinol Metab. 2005;90(7):4081–6.
- Holdaway IM, Bolland MJ, Gamble GD. A meta-analysis of the effect of lowering serum levels of GH and IGF-I on mortality in acromegaly. Eur J Endocrinol. 2008;159(2):89–95.
- 290. Mercado M, Gonzalez B, Vargas G, Ramirez C, de los Monteros AL, Sosa E, et al. Successful mortality reduction and control of comorbidities in patients with acromegaly followed at a highly specialized multidisciplinary clinic. J Clin Endocrinol Metab. 2014;99(12):4438–46.
- Holdaway IM, Rajasoorya RC, Gamble GD. Factors influencing mortality in acromegaly. J Clin Endocrinol Metab. 2004;89(2):667–74.
- 292. Chanson P, Maison P. Does attainment of target levels of growth hormone and insulinlike growth factor I improve acromegaly prognosis? Nat Clin Pract Endocrinol Metab. 2009;5(2):70–1.
- 293. Biermasz NR, van Thiel SW, Pereira AM, Hoftijzer HC, van Hemert AM, Smit JW, et al. Decreased quality of life in patients with acromegaly despite long-term cure of growth hormone excess. J Clin Endocrinol Metab. 2004;89(11):5369–76.
- 294. Arosio M, Reimondo G, Malchiodi E, Berchialla P, Borraccino A, De Marinis L, et al. Predictors of morbidity and mortality in acromegaly: an Italian survey. Eur J Endocrinol. 2012;167(2):189–98.
- 295. Claudia A, Landis Susan B, Masters Anna, Spada Ann M, Pace Henry R, Bourne Lucia, Vallar GTPase inhibiting mutations activate the α chain of Gs and stimulate adenylyl cyclase in human pituitary tumours. Nature 1989;340(6236):692–96 10.1038/340692a0.

Check for updates

Prolactinoma



Kartik Yadav, Sharjeel Shaikh, and Gianluca Tamagno

6.1 Introduction

Prolactin-secreting tumors of the pituitary gland are called prolactinomas. It is the most common secretory tumor of the pituitary gland accounting for up to 40% of total pituitary adenomas [1] and 44–67% of functioning pituitary adenomas [2]. Prolactinomas may cause a wide variety of symptoms either due to mass effect of the tumor or due to hypersecretion of prolactin. Based on the size of the tumor, prolactinomas can be classified as microprolactinoma (smaller than 10 mm), macroprolactinoma (10 mm or larger), or giant prolactinomas; however, the etiological diagnosis of prolactin levels above the reference range may be challenging since hyperprolactinemia is not always due to prolactinoma and other physiological or nonphysiological causes like pregnancy, drugs, and pituitary stalk effect due to locally expanding masses or infiltrative disorders should be considered in the differential diagnosis [3]. Moreover, macroprolactinemia, a laboratory finding due to the formation and the measurement of immunoglobulin–prolactin complexes, should be also ruled out [4].

K. Yadav · S. Shaikh

G. Tamagno (⊠) Hermitage Medical Clinic, Dublin, Ireland

The chapter has been endorsed by **Prof. Dominique Maiter**, dominique.maiter@uclouvain.be, Cliniques Universitaires Saint-Luc, UCL, Woluwe-Saint-Lambert, Belgium

Department of Medicine, Wexford General Hospital – University College Dublin, Wexford, Ireland

Most pituitary tumors occur sporadically. Prolactinomas arise from the monoclonal expansion of pituitary lactotrophs, which have undergone somatic mutation. Pituitary tumor-transforming gene overexpression and mutation of a receptor of fibroblast growth factor 4 have been found in pituitary adenoma mainly prolactinoma [5]. Some patients with prolactinomas are associated with a genetic disorder called multiple endocrine neoplasia type I (MEN-1). MEN-1 is an inherited condition most often characterized by the occurrence, or the predisposition of occurrence, of neuroendocrine tumors of the pancreas, parathyroid, and pituitary gland. Other syndromes or genetic predispositions have been identified as factors possibly leading to the development of a prolactinoma. For example, linkage to aryl hydrocarbon-interacting protein (AIP) gene mutation has been identified in some families with prolactinoma and childhood-onset pituitary adenomas [6].

In terms of anatomical behavior, a subset of prolactinomas, in particular macroprolactinomas, tend to grow with time and may require aggressive treatment to prevent complications. The growth rate varies with the individual and cannot be reliably predicted, depending on a number of factors. Careful monitoring of the clinical signs and symptoms coupled with pituitary gland imaging and with serial measurements of serum prolactin levels (i.e., to detect any major change in tumor behavior) remains the cornerstones of the follow-up for prolactinoma patients [7].

Dopamine agonists are the first-line treatment of most prolactinomas [8]. Surgical treatment, or radiotherapy, is reserved for patients who fail to respond to the medical treatment [9].

Patients with microprolactinoma generally have an excellent prognosis. In up to 95% of microprolactinoma patients, these pituitary tumors do not enlarge over a 4-to 6-year follow-up period [10]. These patients generally do well for extended periods on suppressive therapy with dopamine agonists, and tumor shrinkage is often achieved medically [10].

6.2 Epidemiology

The exact frequency with which prolactinomas occur in the general population is not established. However, a study of 81,449 inhabitants of an area of Oxfordshire, UK, determined the incidence of pituitary adenomas to be 77.6 cases per 100,000 population, with the majority of cases (57%, or 44.4 persons per 100,000 population) being prolactinoma [11].

Prolactinomas may account for up to 40% of all clinically pituitary adenomas, with a peak prevalence in women aged 25 to 34 years [1, 12]. Microprolactinoma is diagnosed more commonly in a female with a female-tomale ratio of 1:20, whereas macroprolactinoma is equally common in both genders [13]. Giant prolactinoma is a rare tumor that occurs mainly in males (about 85%) with an estimated frequency of 0.5–4.4% of all pituitary tumors in that patient population [14].

6.3 Pathophysiology

Prolactin secretion is under dual control from the hypothalamus, where dopamine serves as an inhibitory signal preventing prolactin secretion, while thyrotropinreleasing hormone somehow stimulates prolactin production and release [15]. Increased prolactin secretion can occur from a prolactin-producing adenoma or inflammation (hypophysitis). Also, conditions that result in impaired dopamine delivery or enhanced thyrotropin-releasing hormone signaling or both may result in increased prolactin release.

Some medications result in increased prolactin secretion through their antidopaminergic properties. Chest wall injury and breast stimulation serve as peripheral triggers of autonomic control, which impinge on central neurogenic pathways that attenuate dopamine release into the hypophyseal portal circulation. In some conditions, such as renal or hepatic insufficiency, prolactin is cleared less rapidly from the systemic circulation, which results in increased blood levels of prolactin.

Most cases of prolactinomas are sporadic, and no specific trigger for the development of the prolactin-secreting adenoma is identifiable. When compared to their sporadic counterpart, familial cases of prolactinoma are rare [16]. Most adenomas that secrete prolactin and cause hyperprolactinemia are comprised solely of lactotroph cells. However, combined prolactin and growth hormone (GH) secretion has been reported in 5% of all pituitary tumors, as these are comprised of both lactotroph and somatotroph cells, a co-secretion status, which is usually diagnosed simultaneously [17]. The combination of GH, PRL, and the alpha-subunit of the glycoprotein hormones is also possible, whereas other hormone combinations within a single tumor are extremely rare [18].

6.4 Clinical Presentation

The clinical manifestations of hyperprolactinemia are relatively few and usually easy to recognize. The clinical presentation of a prolactin-secreting adenoma may depend on the direct effect of high prolactin levels, hypogonadotropic hypogonadism occurring through the inhibitory effect of high prolactin concentrations on the hypothalamic gonadotropin-releasing hormone (GnRH) axis, or due to the mass effect of the prolactinoma.

In females, prolactinomas are most often smaller than in males, and therefore, the endocrine symptoms are more common than the mass-related effects, especially in premenopausal women. The classic symptoms of prolactinoma in women include oligomenorrhea or amenorrhea, galactorrhea, and infertility. Recent studies in female patients with hyperprolactinemia have found the prevalence of menstrual disorders as high as 85–90%, but galactorrhea is much less frequent 45–50% [19, 20]. The symptoms of hypogonadism due to hyperprolactinemia in premenopausal women correlate with the magnitude of the hyperprolactinemia. Very high prolactin levels may determine hypogonadism, ultimately leading to low estradiol levels. Low estrogens induce a series of issues, which may manifest as

amenorrhea, hot flushes, and vaginal dryness. A moderate degree of hyperprolactinemia can nevertheless cause amenorrhea or oligomenorrhea, while a mild degree of hyperprolactinemia may just affect progesterone secretion only and therefore determine a short luteal phase of the menstrual cycle [21]. In postmenopausal women, the clinical presentation is different due to the already established physiological hypogonadism. In this setting, prolactinomas most likely manifest with mass effect symptoms and the hormonal effects are very limited, if present at all (i.e., galactorrhea).

Males present with a macroprolactinoma more frequently than females [22], and this may impact on the clinical manifestations of the disease. Larger tumor size in males is not primarily related to diagnostic delay, but rather to gender-related differences in tumor behavior. Retrospective analyses reported higher Ki67 (marker that reflects tumor cell proliferation and growth) in surgically resected macroadenomas from males compared to similar tumors from females [23, 24]. In general, half of the men with prolactinoma present with symptoms due to mass effect of the pituitary adenoma and the other half present with symptoms of hypogonadism including loss of libido, erectile dysfunction, infertility, gynecomastia, osteopenia, or more rarely with galactorrhea [25]. The level of hyperprolactinemia roughly correlates with the symptoms [26]. Erectile dysfunction, on the other hand, appears to be mostly caused by mechanisms not directly related to hypogonadism, as it is more likely corrected by prolactin control via dopamine agonist administration rather than by testosterone replacement [27].

The mass effects of a prolactinoma depend on the size, the location, and the anatomical extension of the adenoma. Among the possible symptoms, which are common to all pituitary adenomas, headache is a well-recognized hallmark of pituitary tumors and may be disabling [28]. The reported incidence of headache in adenomatous pituitary disease ranges with tumor type from 33% to 72% and was reported to be particularly high in prolactinomas [28]. It was previously thought that headache in pituitary adenomas is related to the size of the adenoma via dural stretch due to expansion of the lesion within the sella turcica. These compressive effects in turn stimulate the afferent fibers innervating the dura mater causing pain [29]. Also, an adenoma invading the cavernous sinus can explain the occurrence of headache since the cavernous sinus contains the ophthalmic branch of the trigeminal nerve and internal carotid artery. However, there are cases of noninvasive microprolactinoma manifesting with a severe headache that quickly resolve with the administration of dopamine agonist, suggesting that pituitary tumor-associated headache may derive also from locally effective neurotransmitters or neuromodulators rather than from anatomical reasons only [28]. Various types of headaches can occur to patients with prolactinoma, including chronic and episodic migraine [30], short-lasting unilateral neuralgiform headache attacks with conjunctival injection and tearing [31], cluster headache [32], and trigeminal neuralgia [33]. Visual field deficits, blurred vision, and decreased visual acuity are most likely to occur in patients with a large prolactinoma, and these clinical manifestations are due to the direct mass effect of these lesions. The typical visual field defect, bitemporal hemianopia, is due to the anatomical compression of the optic chiasm, which contains the crossing nasal fibers of each optic nerve [34]. Nevertheless, the visual field defect depends on the relation between the optic chiasm and the adenoma itself. If the tumor is anterior to the optic chiasm or if the patient has an anatomical post-fixed chiasm, conditions such as central scotoma, arcuate scotoma, and monocular visual constriction can be noted [35].

Cranial nerve palsies are rare but occur especially with invasive tumors, including aggressive prolactinomas, or with pituitary apoplexy. Pituitary apoplexy although representing a more common complication of nonfunctioning pituitary adenomas than prolactinomas can happen in prolactinomas as well, especially in pregnant women who over-respond to dopamine or as an adverse effect of thyrotropin-releasing hormone (TRH) stimulation test. In its most dramatic presentation, pituitary apoplexy causes sudden onset of severe headache, diplopia due to pressure on the oculomotor nerves, and hypopituitarism with a possible lifethreatening outcome [36].

Hydrocephalus may rarely occur through several anatomical mechanisms; for example, the adenoma may grow upward and compress the anterior horn of the lateral ventricle. The most common directional growth is upward and backward invading the suprasellar area at an early stage, and large tumors may invaginate the floor of the third ventricle possibly obstructing the aqueduct of Sylvius. Also, the cisterna chiasmatica and the cisterna interpeduncularis can be encroached or obliterated and, as a consequence, may determine the occurrence of hydrocephalus [37]. Very rare though, possible manifestations of prolactinomas with extrasellar extension may also include impaired hearing, unilateral hemiparesis, temporal lobe epilepsy, or even dementia due to frontal lobe extension [38].

6.5 Diagnosis

The hallmark of prolactinoma is hyperprolactinemia. Assay-specific normal values are gender-dependent with a higher normal range in women. In general, a prolactin level below 25 μ g/L (530 mIU/L) is usually considered to be normal [39]. However, differences among laboratories and diagnostic kits exist and should be always carefully considered when comparing prolactin values. Diagnosis of hyperprolactinemia is determined by the measurement of basal prolactin, which roughly correlates with prolactinoma size. Ideally, serum sample should be obtained limiting as much stress as possible, while venipuncture is performed. Stimulatory or inhibitory dynamic tests of prolactin secretion following administration of TRH (stimulates), domperidone (stimulates), L-dopa (inhibits), or nomifensine (inhibits) are not superior to basal prolactin for the diagnosis of hyperprolactinemia and, moreover, may be risky for the patient [40]. Thus, no dynamic test is recommended for the diagnosis of hyperprolactinemia. Basal prolactin concentrations below 200 µg/L cannot discriminate prolactinoma-related hyperprolactinemia from physiological, pharmacological, or other secondary causes of hyperprolactinemia (Table 6.1). Grossly, higher prolactin levels tend to suggest an adenomatous prolactin hypersecretion, though this consideration cannot be taken as a definite rule.

Physiological	Hypothalamic– pituitary stalk damage	Pituitary diseases or dysfunctions	Systemic conditions	Pharmacological causes with (with examples)
Coitus Exercise Lactation Pregnancy Sleep Venepuncture	Infiltrative diseases Radiations Rathke's cyst Traumas with pituitary stalk section, including local surgery Renal cell carcinoma Gonadoblastoma Ovarian teratoma Perivascular epithelioid cell tumors	Acromegaly Idiopathic Local surgery Lymphocytic hypophysitis Macroadenoma with pituitary stalk compression Multiple hormones secreting adenoma	Chest wall trauma Surgery HZV Chronic renal failure Liver cirrhosis Cranial radiation Epilepsy PCOS	Anticonvulsants(valproate)AntipsychoticsTypical(prochlorperazine)Atypical (risperidone)AntidepressantsTricyclic antidepressants(amitriptyline)Selective serotoninreuptake inhibitors(sertraline)Monoamine oxidaseinhibitors (pargyline)Anti-H2 receptors(cimetidine)Antihypertensives(verapamil, labetalol)Prokinetic agents(metoclopramide)MiscellaneousEstrogens(17α-ethynylestradiol)Opiates (morphine)FenfluramineIsoniazid

 Table 6.1
 Possible causes of non-prolactinoma-dependent hyperprolactinemia

In the absence of evident secondary causes of hyperprolactinemia, in doubtful situations, or in cases with mass-related symptoms, imaging should be performed. Magnetic resonance imaging (MRI) of the pituitary is the radiological investigation of choice. A targeted MRI examination of the pituitary region includes sagittal (Fig. 6.1), and transverse and coronal (Fig. 6.2) views with accurate field of view. T1 pre- and post-contrast images as well as dynamic contrast-enhanced coronal images are critical for the identification of small microadenomas.

In general, it is recommended to perform MRI in every patient with any degree of hyperprolactinemia if secondary causes are excluded or very unlikely. If MRI is contraindicated, contrast CT with direct coronal scans can be done, although it is less informative due to poorly accurate definition of the surrounding vasculature, the optic chiasm, and the cavernous sinus. CT also has limited accuracy for small microadenomas.

Despite the often-straightforward clinical presentation and diagnostic pathway, there are several pitfalls when suspecting a prolactinoma. For example, patients with very high prolactin levels may display the so-called hook effect [41, 42]. This effect can occur when very high serum prolactin (i.e., above 5000 ng/mL) saturates both the capture and signal antibodies used in immunoradiometric and chemiluminescent assays, preventing the binding in a "sandwich." The result reflects into a

Fig. 6.1 Post-contrast sagittal T1 MRI showing a prolactinoma







lower than correct prolactin concentration, which is often only mildly or moderately elevated possibly misleading to a diagnosis of nonfunctioning adenoma. This laboratory artifact can be avoided by repeating the assay using a 1:100 dilution of serum [43, 44].

Another laboratory pitfall that may occur in the definition of the cause of hyperprolactinemia is represented by the presence of macroprolactin. Macroprolactin is native prolactin bound to immunoglobulin G and is usually 150–170 kDa in size compared with 23 kDa size of monomeric prolactin [45]. Macroprolactin causes a nonfunctionally active hyperprolactinemia through decreased prolactin clearance. This entity is not of clinical significance per se, but patients can be investigated and misdiagnosed with the obvious consequences of the same as if they were affected by hyperprolactinemia. The macroprolactinemia-induced misdiagnosis could be avoided by pretreating the serum with polyethylene glycol to precipitate the macroprolactin before performing the immunoassay for prolactin in the supernatant. Nowadays, this is standard practice for most laboratories in every patient with high prolactin.

The diagnostic approach in the suspicion of prolactinoma is summarized in Fig. 6.3.



Fig. 6.3 The diagnostic algorithm of hyperprolactinemia

6.6 Management

The objectives of the treatment of prolactinoma are to correct the biochemical consequences of prolactin hypersecretion and to reduce the tumor mass with preservation of normal pituitary function (Table 6.2). Asymptomatic eugonadal women with microprolactinomas do not represent an absolute indication for treatment as it is very unlikely for a microprolactinoma to grow significantly without increased serum prolactin levels [46]. As such, a number of patients with microadenomas verified by imaging may be monitored with serial prolactin measurements. Microprolactinoma with an increase in size mandates treatment as there is a tendency to become a macroadenoma, regardless of the symptoms and the prolactin levels. However, a significant increase in prolactin levels usually, though not always, reflects tumor growth. Women and men who are hypogonadal should be treated as hyperprolactinemia is associated with impairment of fertility and often also sexual functions. In hypogonadal women with microadenoma for which fertility is not a concern, the management options may include estrogen replacement therapy, providing that prolactin levels do not increase substantially and there is no evidence of tumor enlargement [47], or dopamine agonist, which remains the preferred therapeutic option anyway.

Macroprolactinomas are more likely to grow and become aggressive [23]. On the top of this, most macroprolactinomas are associated with prolactin elevation significant enough to elicit symptoms that would require treatment.

Dopamine agonists (DAs) are the first-line treatment for prolactinoma as they are proven to resolve the hyperprolactinemic symptoms in most cases and reduce the tumor size [8, 48]. DAs inhibit prolactin synthesis and secretion by binding to and activating dopamine D2 receptors on the pituitary lactotroph cell membrane. Dopamine and its agonists bind to D2 receptors that are functionally linked to membrane channels and G proteins and suppress the high intrinsic secretory activity of the pituitary lactotrophs. In addition to inhibiting PRL release by controlling calcium fluxes, DA activates several interacting intracellular signaling pathways and suppresses PRL gene expression and lactotroph proliferation [49]. It can also induce the apoptosis of lactotroph cells [50]. Several DAs are or have been available on the market upon prescription for the medical treatment of prolactinomas such as bromocriptine, cabergoline, pergolide, and quinagolide. Factors in favor of success of the treatment with a DA include continuous DA treatment for at least 2 years,

Table 6.2Indications for therapy inpatients with prolactinomas

Enlarging microadenoma Macroadenoma Infertility Amenorrhea or oligomenorrhea Acne and/or hirsutism Troublesome galactorrhea Gynecomastia Testosterone deficiency

MRI images of the pituitary gland

low-normal prolactin levels with the lowest dose of DA, and a reduction in tumor size of at least 50% with no evidence of cavernous sinus invasion. These patients require long-term follow-up with prolactin measurements every 3 months for the first year and annually thereafter, and review of the pituitary gland by MRI, yearly initially, or if prolactin increases above normal [51]. Among the DA, the use of cabergoline rather than bromocriptine seems to be characterized by higher chances of therapeutic success.

Bromocriptine was the first DA introduced to treat prolactinomas. Bromocriptine is an ergot derivative with D1 and D2 receptor agonist properties, has a short half-life of 3–4 h, and should be taken two or three times per day. Bromocriptine normalized serum prolactin in 78% and 72% of patients with microprolactinomas and macroprolactinomas, respectively [52]. It has been found to be able to restore ovulation in about 80% of women [53]. The majority of macroprolactinomas treated with bromocriptine show some degree of reduction in size as assessed by MRI [54].

Cabergoline is an ergot derivative, which is more selective for D2 receptors and has a long in vivo half-life, which permits once or twice weekly administration. The long duration of action stems from its slow elimination from the pituitary tissue due to its high-affinity binding to the dopamine receptors in the pituitary. It is highly effective in normalizing serum prolactin levels, reducing tumor mass, and restoring the gonadal function. Roughly, cabergoline treatment may normalize prolactin levels in 92% of patients with idiopathic hyperprolactinemia or microprolactinomas, 77% of patients with macroprolactinomas, and 86% of the overall study population [55]. The tumor size reduction rate has been estimated ranging between 50% and 100% [56]. Overall, cabergoline is superior to bromocriptine as it has greater affinity and selectivity for the pituitary dopamine D2 receptors and longer duration of action, presents a better tolerability profile, and achieves a higher rate of shrinkage of the prolactin-secreting adenoma [57]. Cabergoline is the first-choice drug for the management of patients with prolactinoma who require medical treatment.

Pergolide is a long-acting ergot derivative acting as an agonist of D1 and D2 receptors, and also 5-HT receptors. It has comparable efficacy and tolerance with bromocriptine, including tumor size reduction [58]. However, also because of concern regarding valvular abnormalities in patients with Parkinson disease taking high doses of pergolide, this drug is not used for the treatment of prolactinomas anymore.

Quinagolide is a non-ergot derivative DA with selective D2 receptor activity and relatively long duration of action. Its efficacy in normalizing prolactin levels and reducing tumor size is at least as good as that of bromocriptine and pergolide [56]. Furthermore, approximately 40% of patients resistant to bromocriptine respond to quinagolide [59]. This medication is not available in the USA, but available in some EU countries.

The principal side effects of DA are nausea, postural hypotension, and mental fogginess [60]. Less common side effects include nasal stuffiness, depression, Raynaud phenomenon, alcohol intolerance, and constipation. Nausea appears to be more common with bromocriptine than cabergoline. Symptoms of psychosis or exacerbation of pre-existing psychosis and impulse control disorders, such as pathological gambling, hypersexuality, or compulsive shopping or eating, have also been

shown to be associated with both bromocriptine and cabergoline. Ergot derivative DAs (cabergoline, bromocriptine, and pergolide) are potentially associated with an increased risk of dose-dependent valvular heart diseases [61]. However, no real concern for prolactinoma patients treated with a DA has been raised so far in the routine clinical practice [62] and the occurrence of DA-related valvular disease seems to basically occur in patients with Parkinson disease receiving a high cumulative dose of DA [62].

A subset of individuals with prolactinomas does not respond satisfactorily to DA. Prolactinomas exhibit varying degrees of responsiveness to DAs, ranging from the complete response at one end of the spectrum to total resistance at the other end. DA resistance is defined as failure to normalize prolactin levels and failure to decrease tumor size by 50% [63]. The factors determining the sensitivity of a prolactinoma to DA treatment are multiple, with the D2 receptor expression and sensitivity at the levels of the prolactinoma cells playing a key role. The estimated prevalence of DA resistance is approximately 24% for bromocriptine and 11% for cabergoline [56]. The management of drug-resistant prolactinomas includes several options. Any bromocriptine-resistant patient should be switched to cabergoline, which will likely normalize prolactin in 80% of patients. As long as adverse effects do not develop, dose escalation of cabergoline is reasonable, with the expectation that subsequent dose reduction will be possible, if and when appropriate. Echocardiographic monitoring is advised in patients taking high cabergoline dose because of the potential association with cardiac valvular fibrosis. Of note, complete resistance to cabergoline is infrequent [64]. Doses of cabergoline of up to 2.0 mg/week are usually effective in controlling prolactin secretion and reducing tumor size in most prolactinomas [65]. The clinical presentation, management, and outcome of patients that are not well controlled by the commonly used dose of DA-resistant patients are still largely undefined [65]. DA resistance is rarely encountered in microprolactinomas and is more frequent in cases of macroprolactinomas (3-6%). Often, it is characteristic of invasive tumors and associated with male gender [66]. Molecular mechanisms implicated in DA resistance likely encompass a diverse set of alterations, and genetic heterogeneity found among sparsely available surgical specimens complicates the assessment of individual factors responsible for aggressive behavior and drug resistance of prolactinomas [67].

Historically, surgical resection was the preferred treatment of prolactinoma, like for the other pituitary adenomas, before discovering the high efficacy of the DA in lowering prolactin levels, reducing prolactinoma size, and restoring gonadal function [44, 56]. Since DAs are so highly effective, recognized indications for surgery of prolactinoma have become rare, usually targeting specific situations where medical treatment is failing. Nowadays, surgical tumor resection is indeed indicated in the prolactinoma patients with severe intolerance or resistance to DA, in patients with acute complications such as apoplexy or cerebrospinal fluid leak, or in young patients with a likely resectable adenoma and who do not wish to take long-term medical treatment [44, 56, 68]. In the last decade, a number of studies have also raised the concern of an increased risk for cardiac valve disease with the prolonged use of DA with partial serotoninergic activity [61, 69]. However, this does not appear being a real issue in patients with prolactinomas, while it may be a serious concern in patients exposed to higher DA cumulative dose, like those affected by Parkinson's disease. Also, emerging evidence has been provided for a link between chronic DA therapy and impulsive behavior disturbances such as gambling, hyper-sexuality, or bulimia [70, 71]. Transsphenoidal approach represents the standard of surgical care for the prolactinomas requiring surgery, in particular for the over-whelming majority of macroprolactinomas. Craniotomy is reserved only for tumors requiring neurosurgery and that are inaccessible via the transsphenoidal approach. On the top of the invasiveness of the tumor, surgical outcomes are highly dependent upon the expertise and experience of the neurosurgeon, as well as the tumor size.

Radiotherapy, i.e., external beam radiation therapy or stereotactic radiosurgery, is only rarely used in patients with prolactinomas and should be reserved for those tumors that do not respond to DAs, recur or progress after surgery, are highly aggressive and/or malignant, or in cases where surgery cannot be an option [72]. Prolactinoma patients who recur after prior treatment with radiotherapy may also be potentially salvaged with radiosurgery. The most frequent long-term morbidity of radiotherapy is radiation-induced hypopituitarism, like for every kind of pituitary adenoma undergoing radiation [73].

Experimental data have demonstrated that different somatostatin receptor (SSTR) subtypes are expressed at various levels in prolactinomas, SSTR5 being the most important in the regulation of prolactin secretion [74, 75]. The most used somatostatin analogs, octreotide and lanreotide [76], which are the cornerstone of the medical treatment for the control of GH secretion in acromegaly, bind efficiently to SSTR2 and with lower affinity to SSTR5. Somatostatin suppresses in vitro prolactin secretion from prolactinoma cultures [77]. However, somatostatin and octreotide minimally alter serum prolactin levels in prolactinoma patients [78]. It appears that only SSTR5-selective agonists (BIM-23052, BIM-23268) can efficiently suppress prolactin release from adenoma cells [75]. SSTR5-specific agonists might potentially be effective in the treatment of PRL-secreting pituitary adenomas and could be tested in vivo in patients with dopamine agonist-resistant prolactinomas [75]. A few years ago, hybrid molecules such as dopastatin, a somatostatin, and dopamine receptor agonist with high binding affinity for SSTRs and D2DR have been developed. These molecules, when tested in culture studies from mixed GH/ PRL-secreting tumors partially responsive to the agonists taken individually, showed a greater efficacy in suppressing GH or PRL secretion [79]. Unfortunately, this therapeutic potential did not translate into a clinical effect due to in vivo pharmacological issues.

As a newcomer in the scenario of the medical treatment of hyperprolactinemia and prolactinomas, phytotherapy should be taken into consideration, and some phytotherapic agents have lately made their appearance among the potentially available therapeutic tools. For example, Vitex agnus-castus extracts, traditionally used for the treatment of menstrual disorders and premenstrual syndrome, have been used also for the treatment of hyperprolactinemia [80]. Good tolerability and substantial lack of significant side effects and drug interactions are the advantages of a trial of Vitex agnus-castus in patients willing to try an alternative approach for the management of their prolactinoma. However, no univocal evidence-based efficacy of Vitex agnus-castus extracts has been demonstrated yet.

6.7 Peculiar Aspects

6.7.1 Prolactinoma in Pregnancy

Prolactin physiologically increases during pregnancy and lactation, and therefore, it does not reliably reflect an increase in tumor size in pregnant prolactinoma patients and it cannot be useful for clinical assessment during pregnancy and lactation. Prolactinoma cells express estrogen receptors [81], and as a result of the increased estrogen level during pregnancy [44] (and eventually of cessation of previous DA treatment), there can be a substantial increase in the volume of the prolactinoma, with a progressive increase in serum prolactin due to lactotroph cell hyperplasia [82]. The main concern relates to the possible prolactinoma enlargement during pregnancy. The risk of tumor enlargement during pregnancy largely depends on adenoma size. Data in the literature indicate that although the average adenoma enlargement is only 3% for microprolactinomas, it can be as high as 32% for macroprolactinomas that were not previously operated on [56, 83].

In patients at risk of an increase in the size of their prolactinoma, definitive treatment by transsphenoidal surgery should be considered and ideally performed before attempting the pregnancy [84]. The treatment of pregnant women must be tailored to the individual patient, though it is generally advised against medical treatment throughout pregnancy and lactation in women with a microprolactinoma and in most women with macroprolactinoma not affecting the optic chiasm and presenting without any sign of invasiveness. On the other hand, in women with an invasive prolactinoma or with a macroprolactinoma abutting the optic chiasm or the cavernous sinus, continuation of the DA treatment may be preferred [83]. In the context of the very low risk of microprolactinoma enlargement during pregnancy, there is considerable evidence supporting the discontinuation of DA treatment once pregnancy is confirmed. In such a case, the patient should be told that the risk of enlargement of the adenoma during her pregnancy is indeed very small and medical treatment would not be required during her pregnancy. However, patients should be advised to report for urgent assessment in case of new symptoms such as severe headache or visual disturbances [44]. The patient should undergo baseline formal visual field testing at the time of diagnosis and should be followed clinically every 2-3 months during pregnancy. In case the patient becomes symptomatic with visual disturbance or progressive headaches, an MRI should be performed to assess changes in tumor size. Otherwise, no MRI followup is usually recommended during pregnancy. If substantial growth of the prolactinoma is clinically and radiologically evident, a suitable treatment option should be considered. The most straightforward option is represented by the DA treatment restoration. Traditionally, bromocriptine was the preferred option since there was a larger experience on the use of this molecule in such a specific setting

[44, 85]. However, in the last two decades, cabergoline has become the firstchoice DA because of its higher therapeutic ratio. Experience with both drugs shows no increase in spontaneous abortions, preterm deliveries, multiple births, or congenital malformations, compared to what is expected in the normal population [84, 86]. Some selected pregnant women with macroprolactinoma may require continuation of the DA throughout the pregnancy [44]. If an enlarging prolactinoma does not respond to DA therapy within 2–3 weeks from the restoration of the medical treatment, transsphenoidal surgery, preferably in the second trimester, should be considered. If the pregnancy is approaching the time of delivery, partum induction would be a reasonable option [87].

Treatment discontinuation is recommended at the time of delivery in women with microprolactinoma or non-compressive macroprolactinoma. For microprolactinomas, the risk of symptomatic tumor enlargement during pregnancy is very low (2–3%). It is higher for macroprolactinomas (20–30%) and careful follow-up is advised, including MRI without contrast injection if symptoms or visual disturbances develop. If a symptomatic tumor enlargement does occur, reinitiation of the dopamine agonist (BRC or CAB) is indicated rather than surgery. Breastfeeding has no harmful effect on tumor growth and DA treatment, if still needed, may be postponed as long as breastfeeding is desired. Finally, about 40% of women with a microprolactinoma or an intermediate size macroprolactinoma may be in prolonged remission after one or more pregnancies [86].

6.7.2 Giant Prolactinoma

Giant prolactinoma is defined as a prolactin-secreting adenoma with a diameter of 40 mm, at least. They usually present with massive extrasellar extension, baseline prolactin concentration of 1000 ng/mL or more, and no concomitant GH or ACTH secretion. They are much more frequent in young to middle-aged men than in women, with a male-to-female ratio of about 9:1 and a mean age around 40 years [38, 88, 89]. A giant prolactinomas have a challenging management as the therapeutic goals differ from the one of a smaller macroprolactinoma where normalization of prolactin levels, restoration of eugonadism, and reduction in tumor size are achievable targets. In giant prolactinomas, the priority of the treatment is often to obtain a rapid alleviation of the neurological symptoms, or of the risk of the same, and other complications due to encroachment upon or invasion of the prolactinoma into surrounding nervous and vascular structures. Furthermore, in longstanding tumors the reduction in the size may not reverse visual field defects or hypopituitarism. In this case, a reasonable goal may be represented just by the prevention of further growth of the prolactinoma and the replacement of the pituitary hormones, which eventually became insufficient, if indicated. DAs are the recommended initial modality of treatment of giant prolactinomas and surgery should be reserved for those patients who demonstrate inadequate responses to medical therapy [38, 90]. Even if surgery may be

necessary at some point with a debulking purpose, it may be curative only in exceptionally rare cases and reinitiating dopamine agonist is usually required for the treatment of persistent hyperprolactinemia, though possibly with higher chances of therapeutic response.

6.7.3 Malignant Prolactinomas

Malignant prolactinomas are very rare tumors, defined by the occurrence of distant cerebrospinal, meningeal, and/or systemic metastases [2]. They differ from invasive pituitary tumors, which remain contiguous with the primary tumor site. Malignant prolactinomas may occur at any age, even in children [91], but mostly develop in the fifth or sixth decade of life [92]. Their exact incidence is not precisely known, but overall, pituitary carcinomas account for only 0.1–0.2% of all pituitary tumors, and prolactinomas correspond to approximately one-third of these.

There are currently no reliable pathological markers, whereby the malignant potential of a prolactinoma can be predicted [2]. The diagnosis of an aggressive pituitary tumor should be considered in patients with a radiologically invasive tumor and/or with an unusually rapid tumor growth rate, or in those patients with a clinically relevant tumor growth despite optimal standard treatment (surgery, radiotherapy, and conventional medical treatment). MRI should be used for quantification of tumor dimensions, invasion, and growth [93]. Dopamine agonists are recommended standard medical treatment with maximally tolerated doses in order to control tumor growth [93]. Unfortunately, resistance to DA occurs very frequently in such clinical scenario, and once cerebrospinal or systemic metastases become obvious, the potentially effective therapeutic options are limited, and the treatment is mostly palliative. Chemotherapy including procarbazine, vincristine, cisplatinum, and etoposide has been used with little effect [94]. Temozolomide monotherapy should be used as first-line chemotherapy for aggressive pituitary tumors and pituitary carcinomas, following documented tumor growth [93, 95]. Temozolomide has been shown to reduce prolactin levels and control tumor growth if tumor specimens do not express methylguanine-DNA methyltransferase (MGMT) [96]. Surgery may be useful in debulking the lesion and relieving local compressive effects, if required. Surgery should be performed by a neurosurgeon with extensive experience in pituitary surgery [93]. The expected survival of patients with a diagnosis of malignant prolactinoma, which is made at the time of the detection of metastatic disease, is approximately 1 year [94].

6.7.4 Inherited Prolactinomas

Prolactinoma is most frequently a sporadic disease. However, it may occur also as part of an inherited syndrome. Roughly, inherited prolactinomas behave more aggressively and are larger than their sporadic counterpart. Prolactinomas occur in 15–25% of patients with MEN-1 and represent the most frequent pituitary adenoma type observed in the setting of this syndrome [92, 97]. The general treatment strategy for prolactinomas in patients with MEN-1 does not differ from that for sporadic prolactinomas, but more intensive pharmacologic therapy or the use of multiple therapeutic modalities may be required to achieve a fully satisfactory outcome [98].

Prolactinomas in MEN-4, an endocrine tumor syndrome more recently described, is caused by germline mutations in putative tumor suppressor gene CDKN1B and is characterized by primary hyperparathyroidism followed by pituitary adenoma [99].

Prolactinomas may also occur—although rarely—in Carney complex, wellcharacterized autosomal dominant disorder presents with a clinical picture of, or predisposition to, several types of tumors arising from a number of sites, including the pituitary, thyroid, testes, and ovaries [100].

AIP- related isolated familial pituitary adenoma is defined as the presence of an AIP germline pathogenic variant in an individual with a pituitary adenoma [101]. The most commonly occurring pituitary adenomas in this disorder are GH-secreting adenomas, followed by prolactinomas, mixed GH and prolactin co-secreting adenomas, and nonfunctioning pituitary adenomas [101].

As a matter of good practice, the presence of pituitary adenomas in the family of a patient with prolactinoma, like any other pituitary adenoma, should always be explored by anamnesis and in case of positive findings a more specific and targeted investigation of the possible familial issue should be carefully carried out.

6.8 Prognosis

The majority of patients with microprolactinomas have an excellent prognosis. These patients can be managed medically also for extended periods, if required, though often can be medically cured in a few years' time [12]. Macroprolactinomas, on the other hand, can grow over time and require more aggressive treatment, with lower success rate in curative terms, and definitely require a more careful follow-up than their sub-centimeter counterpart. The growth rate of macroprolactinomas is unpredictable, and thus, the patient must be closely followed up. The decision to taper medical therapy requires sound judgment because the tumor can grow in size if without treatment or after DA withdrawal [102].

Once prolactin levels have reached normal levels, or in some cases near-normal, they can be monitored every 3–6 months for the first year and then every 6–12 months thereafter. Macroadenoma tumor size can be monitored by serial yearly MRI scans, and once maximal size reduction has been documented and deemed stabilizing, further scans may not be necessary as long as prolactin levels are being monitored. Whether a follow-up MRI scan is necessary in patient with microadenomas is debatable, if prolactin levels are regularly monitored. It is extremely rare for a tumor to increase in size without evidence of a significant increase in prolactin levels. Visual field testing should be repeated until they normalize or remain stable and then do not need to be repeated [8].

In the PROLEARS study, patients with a microprolactinoma had no increased mortality, which is reassuring as this is the most common type of pituitary adenoma associated with hyperprolactinemia, while patients with hyperprolactinemia due to a macroprolactinoma had increased mortality [103]. There are insufficient data to explore the possible reasons for such increased risk of death, although premature mortality has been generally reported in patients with a pituitary macroadenoma. For example, other hormonal deficiencies as a result of hypopituitarism or excessive steroid replacement in the past may contribute to adverse health outcomes [104].

Acknowledgments We are grateful to Dr. Obada Yousif, consultant endocrinologist, Wexford General Hospital, and Dr. John Coffey, consultant radiologist, Wexford General Hospital, for their expert input and advice.

References

- Mindermann T, Wilson CB. Age-related and gender-related occurrence of pituitary adenomas. Clin Endocrinol. 1994;41(3):359–64.
- 2. Olarescu NC, Perez-Rivas LG, Gatto F, Cuny T, Tichomirowa MA, Tamagno G, et al. Aggressive and malignant prolactinomas. Neuroendocrinology. 2019;109(1):57–69.
- 3. Bronstein MD. Disorders of prolactin secretion and prolactinomas. 2nd ed. Philadelphia: Elsevier Saunders; 2010.
- Pałubska S, Adamiak-Godlewska A, Winkler I, Romanek-Piva K, Rechberger T, Gogacz M. Hyperprolactinaemia—a problem in patients from the reproductive period to the menopause. Menopausal Rev. 2017;1(16):1–7.
- Heaney AP, Horwitz GA, Wang Z, Singson R, Melmed S. Early involvement of estrogeninduced pituitary tumor transforming gene and fibroblast growth factor expression in prolactinoma pathogenesis. Nat Med. 1999;5(11):1317–21.
- Georgitsi M, De Menis E, Cannavò S, Mäkinen MJ, Tuppurainen K, Pauletto P, et al. Aryl hydrocarbon receptor interacting protein (AIP) gene mutation analysis in children and adolescents with sporadic pituitary adenomas. Clin Endocrinol. 2008;69(4):621–7.
- Vaneckova M, Seidl Z, Hana V, Jarkovska Z. Macroprolactinomas: retrospective follow up study in the MR imaging and correlation with clinical symptomatology. Neuro Endocrinol Lett. 2007;28(6):841–5.
- Molitch ME, Drummond J, Korbonits M. Prolactinoma Management. Nih.gov. MDText.com, Inc.; 2018.
- 9. Liu JK, Couldwell WT. Contemporary management of prolactinomas. Neurosurg Focus. 2004;16(4):E2.
- 10. Segu VB. Prolactinoma: practice essentials, pathophysiology, epidemiology. Medscape. com. 2019.
- Fernandez A, Karavitaki N, Wass JAH. Prevalence of pituitary adenomas: a community-based, cross-sectional study in Banbury (Oxfordshire, UK). Clin Endocrinol. 2010;72(3):377–82.
- 12. Yatavelli RKR, Bhusal K. Prolactinoma. Nih.gov. StatPearls Publishing; 2018.
- 13. Hoffmann A, Adelmann S, Lohle K, Claviez A, Müller HL. Pediatric prolactinoma: initial presentation, treatment, and long-term prognosis. Eur J Pediatr. 2017;177(1):125–32.
- Iglesias P, Bernal C, Villabona C, Castro JC, Arrieta F, Díez JJ. Prolactinomas in men: a multicentre and retrospective analysis of treatment outcome. Clin Endocrinol. 2012;77(2):281–7.
- 15. Al-Chalabi M, Alsalman I. Physiology, Prolactin. Nih.gov. StatPearls Publishing; 2019.

- Lecoq A-L, Kamenický P, Guiochon-Mantel A, Chanson P. Genetic mutations in sporadic pituitary adenomas—what to screen for? Nat Rev Endocrinol. 2014;11(1):43–54.
- 17. Kasantikul V, Shuangshoti S. Pituitary adenomas: immunohistochemical: study of 90 cases. J Med Assoc Thail. 1990;73(9):514–21.
- Kovacs K, Horvath E, Stefaneanu L, Bilbao J, Singer W, Muller PJ, et al. Pituitary adenoma producing growth hormone and adrenocorticotropin: a histological, immunocytochemical, electron microscopic, and in situ hybridization study. Case report. J Neurosurg. 1998;88(6):1111–5.
- 19. Kleinberg DL, Noel GL, Frantz AG. Galactorrhea: a study of 235 cases, including 48 with pituitary tumors. N Engl J Med. 1977;296(11):589–600.
- Clevenger CV, Zheng J, Jablonski EM, Galbaugh TL, Fang F. From bench to bedside: future potential for the translation of prolactin inhibitors as breast cancer therapeutics. J Mammary Gland Biol Neoplasia. 2008;13(1):147–56.
- Seppälä M, Ranta T, Hirvonen E. Hyperprolactinemia and luteal insufficiency. Lancet. 1976;307(7953):229–30.
- Nishioka H, Haraoka J, Akada K. Growth potential of prolactinomas in men: is it really different from women? Surg Neurol. 2003;59(5):386–90.
- Delgrange E, Trouillas J, Maiter D, Donckier J, Tourniaire J. Sex-related difference in the growth of Prolactinomas: a clinical and proliferation marker Study1. J Clin Endocrinol Metabol. 1997;82(7):2102–7.
- 24. Ramot Y, Rapoport MJ, Hagag P, Wysenbeek AJ. A study of the clinical differences between women and men with hyperprolactinemia. Gynecol Endocrinol. 1996;10(6):397–400.
- Carter JN, Tyson JE, Tolis G, Van Vliet S, Faiman C, Friesen HG. Prolactin-screening tumors and hypogonadism in 22 men. N Engl J Med. 1978;299(16):847–52.
- Tamagno G, Daly AF, Deprez M, Vroonen L, Andris C, Martin D, et al. Absence of hypogonadism in a male patient with a giant prolactinoma: a clinical paradox. Ann Endocrinol. 2008;69(1):47–52.
- 27. Zeitlin SI, Rajfer J. Hyperprolactinemia and erectile dysfunction. Rev Urol. 2000;2(1):39–42.
- Levy MJ, Jäger HR, Powell M, Matharu MS, Meeran K, Goadsby PJ. Pituitary volume and headache. Arch Neurol. 2004;61(5):721.
- McNaughton FL. The innervation of the intracranial blood vessels and dural sinuses. Proc Assoc Res Nervous Mental Dis. 1938;18:178–200.
- 30. Lee AHS. Pituitary adenoma causing classical migraine. Br J Neurosurg. 1990;4(4):347-8.
- Massiou H, Launay JM, Levy C, El Amrani M, Emperauger B, Bousser MG. SUNCT syndrome in two patients with prolactinomas and bromocriptine-induced attacks. Neurology. 2002;58(11):1698–9.
- Porta-Etessam J, Ramos-Carrasco A, Berbel-García A, Martínez-Salio A, Benito-León J. Clusterlike headache as first manifestation of a prolactinoma. Headache. 2001;41(7):723–5.
- Gazioğlu N, Tanriöver N, Tüzgen S. Pituitary tumour presenting with trigeminal neuralgia as an isolated symptom. Br J Neurosurg. 2000;14(6):579.
- Miller NR, Newman NJ, Biousse V, Kerrison JB. Walsh & Hoyt's clinical neuroophthalmology. 6th ed. Philadelphia: Lippincott Williams & Wilkins; 2005. p. 503–73.
- 35. Ho R-W, Huang H-M, Ho J-T. The influence of pituitary adenoma size on vision and visual outcomes after trans-sphenoidal adenectomy: a report of 78 cases. J Korean Neurosurg Soc. 2015;57(1):23.
- Capatina C, Inder W, Karavitaki N, Wass JAH. Management of endocrine disease: pituitary tumour apoplexy. Eur J Endocrinol. 2015;172(5):R179–90.
- Shenkin HA, Crowley JN. Hydrocephalus complicating pituitary adenoma. J Neurol Neurosurg Psychiatry. 1973;36(6):1063–8.
- Maiter D, Delgrange E. Therapy of endocrine disease: the challenges in managing giant prolactinomas. Eur J Endocrinol. 2014;170(6):R213–27.
- Casanueva FF, Molitch ME, Schlechte JA, Abs R, Bonert V, Bronstein MD, et al. Guidelines of the pituitary society for the diagnosis and management of prolactinomas. Clin Endocrinol. 2006;65(2):265–73.

- Mancini T, Casanueva FF, Giustina A. Hyperprolactinemia and prolactinomas. Endocrinol Metab Clin N Am. 2008;37(1):67–99.
- Delgrange E, de Hertogh R, Vankrieken L, Maiter D. Potential hook effect in prolactin assay in patients with giant prolactinoma. Clin Endocrinol. 1996;45(4):506–7.
- Vilar L, Vilar CF, Lyra R, Freitas MC. Pitfalls in the diagnostic evaluation of hyperprolactinemia. Neuroendocrinology. 2019;109(1):7–19.
- St-Jean E, Blain F, Comtois R. High prolactin levels may be missed by immunoradiometric assay in patients with macroprolactinomas. Clin Endocrinol. 1996;44(3):305–9.
- 44. Melmed S, Casanueva FF, Hoffman AR, Kleinberg DL, Montori VM, Schlechte JA, et al. Diagnosis and treatment of hyperprolactinemia: an Endocrine Society Clinical Practice Guideline. J Clin Endocrinol Metabol. 2011;96(2):273–88.
- Kavanagh-Wright L, Smith TP, Gibney J, McKenna TJ. Characterization of macroprolactin and assessment of markers of autoimmunity in macroprolactinaemic patients. Clin Endocrinol. 2009;70(4):599–605.
- Schlechte J, Dolan K, Sherman B, Chapler F, Luciano A. The natural history of untreated hyperprolactinemia: a prospective analysis. J Clin Endocrinol Metabol. 1989;68(2):412–8.
- 47. Testa G, Vegetti W, Motta T, Alagna F, Bianchedi D, Carlucci C, et al. Two-year treatment with oral contraceptives in hyperprolactinemic patients. Contraception. 1998;58(2):69–73.
- Auriemma RS, Pirchio R, De Alcubierre D, Pivonello R, Colao A. Dopamine agonists: from the 1970s to today. Neuroendocrinology. 2019;109(1):34–41.
- 49. Ben-Jonathan N, Hnasko R. Dopamine as a prolactin (PRL) inhibitor. Endocr Rev. 2001;22(6):724–63.
- Radl DB, Zárate S, Jaita G, Ferraris J, Zaldivar V, Eijo G, et al. Apoptosis of lactotrophs induced by D2 receptor activation is estrogen dependent. Neuroendocrinology. 2008;88(1):43–52.
- Dekkers OM, Lagro J, Burman P, Jørgensen JO, Romijn JA, Pereira AM. Recurrence of hyperprolactinemia after withdrawal of dopamine agonists: systematic review and metaanalysis. J Clin Endocrinol Metabol. 2010;95(1):43–51.
- Colao A, di Sarno A, Pivonello R, di Somma C, Lombardi G. Dopamine receptor agonists for treating prolactinomas. Expert Opin Investig Drugs. 2002;11(6):787–800.
- 53. Hamoda H, Khalaf Y, Carroll P. Hyperprolactinaemia and female reproductive function: what does the evidence say? Obstetr Gynaecol. 2012;14(2):81–6.
- Bevan JS, Webster J, Burke CW, Scanlon MF. Dopamine agonists and pituitary tumor shrinkage. Endocr Rev. 1992;13(2):220–40.
- Verhelst J, Abs R, Maiter D, van den Bruel A, Vandeweghe M, Velkeniers B, et al. Cabergoline in the treatment of hyperprolactinemia: a study in 455 patients. J Clin Endocrinol Metabol. 1999;84(7):2518–22.
- Gillam MP, Molitch ME, Lombardi G, Colao A. Advances in the treatment of prolactinomas. Endocr Rev. 2006;27(5):485–534.
- Webster J, Piscitelli G, Polli A, Ferrari CI, Ismail I, Scanlon MF. A comparison of cabergoline and bromocriptine in the treatment of hyperprolactinemic amenorrhea. Cabergoline Comparative Study Group. N Engl J Med. 1994;331(14):904–9.
- Lamberts SWJ, Quik RFP. A comparison of the efficacy and safety of pergolide and bromocriptine in the treatment of hyperprolactinemia. J Clin Endocrinol Metabol. 1991;72(3):635–41.
- Barlier A, Jaquet P. Quinagolide a valuable treatment option for hyperprolactinaemia. Eur J Endocrinol. 2006;154(2):187–95.
- Borovac JA. Side effects of a dopamine agonist therapy for Parkinson's disease: a minireview of clinical pharmacology. Yale J Biol Med. 2016;89(1):37–47.
- Schade R, Andersohn F, Suissa S, Haverkamp W, Garbe E. Dopamine agonists and the risk of cardiac-valve regurgitation. N Engl J Med. 2007;356(1):29–38.
- Cheung D, Heaney A. Dopamine agonists and valvular heart disease. Curr Opin Endocrinol Diabetes Obes. 2009;16(4):316–20.
- Brue T, Pellegrini I, Priou A, Morange I, Jaquet P. Prolactinomas and resistance to dopamine agonists. Horm Res. 1992;38(1–2):84–9.

- Maiter D. Management of dopamine agonist-resistant prolactinoma. Neuroendocrinology. 2019;109(1):42–50.
- 65. Vroonen L, Jaffrain-Rea M-L, Petrossians P, Tamagno G, Chanson P, Vilar L, et al. Prolactinomas resistant to standard doses of cabergoline: a multicenter study of 92 patients. Eur J Endocrinol. 2012;167(5):651–62.
- 66. Delgrange E, Daems T, Verhelst J, Abs R, Maiter D. Characterization of resistance to the prolactin-lowering effects of cabergoline in macroprolactinomas: a study in 122 patients. Eur J Endocrinol. 2009;160(5):747–52.
- 67. Melmed S. The pituitary. 4th ed. UK: Elsevier Science; 2016.
- Buchfelder M, Zhao Y, Schlaffer S-M. Surgery for prolactinomas to date. Neuroendocrinology. 2019;109(1):77–81.
- Zanettini R, Antonini A, Gatto G, Gentile R, Tesei S, Pezzoli G. Valvular heart disease and the use of dopamine agonists for Parkinson's disease. N Engl J Med. 2007;356(1):39–46.
- Ambermoon P, Carter A, Hall WD, Dissanayaka NNW, O'Sullivan JD. Impulse control disorders in patients with Parkinson's disease receiving dopamine replacement therapy: evidence and implications for the addictions field. Addiction. 2011;106(2):283–93.
- Barns Neurauter MP, Rickards H, Cavanna AE. The prevalence and clinical characteristics of pathological gambling in Parkinson's disease: an evidence-based review. Funct Neurol. 2010;25(1):9–13.
- Sheplan Olsen LJ, Robles Irizarry L, Chao ST, Weil RJ, Hamrahian AH, Hatipoglu B, et al. Radiotherapy for prolactin-secreting pituitary tumors. Pituitary. 2012;15(2):135–45.
- Loeffler JS, Shih HA. Radiation therapy in the management of pituitary adenomas. J Clin Endocrinol Metabol. 2011;96(7):1992–2003.
- 74. Jaquet P, Ouafik L, Saveanu A, Gunz G, Fina F, Dufour H, et al. Quantitative and functional expression of somatostatin receptor subtypes in human prolactinomas. J Clin Endocrinol Metabol. 1999;84(9):3268–76.
- 75. Shimon I, Yan X, Taylor JE, Weiss MH, Culler MD, Melmed S. Somatostatin receptor (SSTR) subtype-selective analogues differentially suppress in vitro growth hormone and prolactin in human pituitary adenomas. Novel potential therapy for functional pituitary tumors. J Clin Invest. 1997;100(9):2386–92.
- Caron P, Morange-Ramos I, Cogne M, Jaquet P. Three-year follow-up of acromegalic patients treated with intramuscular slow-release lanreotide. J Clin Endocrinol Metab. 1997;82(1):18–22.
- 77. Ishibashi M, Yamaji T. Mechanism of the inhibitory action of dopamine and somatostatin on prolactin secretion from human lactotrophs in culture. J Clin Endocrinol Metabol. 1985;60(3):599–606.
- Lamberts SW, Zweens M, Klijn JG, van Vroonhoven CC, Stefanko SZ, Del Pozo E. The sensitivity of growth hormone and prolactin secretion to the somatostatin analogue SMS 201-995 in patients with prolactinomas and acromegaly. Clin Endocrinol. 1986;25(2):201–12.
- 79. Jaquet P, Gunz G, Saveanu A, Dufour H, Taylor J, Dong J, et al. Efficacy of chimeric molecules directed towards multiple somatostatin and dopamine receptors on inhibition of GH and prolactin secretion from GH-secreting pituitary adenomas classified as partially responsive to somatostatin analog therapy. Eur J Endocrinol. 2005;153(1):135–41.
- Tamagno G, Burlacu MC, Daly AF, Beckers A. Vitex agnus castus might enrich the pharmacological armamentarium for medical treatment of prolactinoma. Eur J Obstet Gynecol Reprod Biol. 2007;135(1):139–40.
- Pichon MF, Bression D, Peillon F, Milgrom E. Estrogen receptors in human pituitary adenomas. J Clin Endocrinol Metabol. 1980;51(4):897–902.
- Huang W, Molitch ME. Pituitary tumors in pregnancy. Endocrinol Metab Clin N Am. 2019;48(3):569–81.
- Almalki MH, Alzahrani S, Alshahrani F, Alsherbeni S, Almoharib O, Aljohani N, et al. Managing prolactinomas during pregnancy. Front Endocrinol. 2015;6:85.
- Molitch ME. Prolactinoma in pregnancy. Best Pract Res Clin Endocrinol Metab. 2011;25(6):885–96.

- Kupersmith MJ. Visual loss in pregnant women with pituitary adenomas. Ann Intern Med. 1994;121(7):473.
- 86. Maiter D. Prolactinoma and pregnancy: from the wish of conception to lactation. Ann Endocrinol. 2016;77(2):128–34.
- Serri O, Chik CL, Ur E, Ezzat S. Diagnosis and management of hyperprolactinemia. Can Med Assoc J. 2003;169(6):575–81.
- Shrivastava RK, Arginteanu MS, King WA, Post KD. Giant prolactinomas: clinical management and long-term follow up. J Neurosurg. 2002;97(2):299–306.
- Moraes AB, Silva CMS, Vieira Neto L, Gadelha MR. Giant prolactinomas: the therapeutic approach. Clin Endocrinol. 2013;79(4):447–56.
- Shimon I. Giant prolactinomas. Neuroendocrinology. 2019;109(1):51–6.
- 91. Huang AP-H, Yang S-H, Yang C-C, Kuo M-F, Wu MZ, Tu Y-K. Malignant prolactinoma with craniospinal metastasis in a 12-year-old boy. J Neuro-Oncol. 2008;90(1):41–6.
- De Sousa SMC, McCormack AI. Aggressive pituitary tumors and pituitary carcinomas. Nih. gov. MDText.com, Inc.; 2018.
- 93. Raverot G, Burman P, McCormack A, Heaney A, Petersenn S, Popovic V, et al. European Society of Endocrinology Clinical Practice Guidelines for the management of aggressive pituitary tumours and carcinomas. Eur J Endocrinol. 2018;178(1):G1–24.
- Kaltsas GA, Nomikos P, Kontogeorgos G, Buchfelder M, Grossman AB. Diagnosis and management of pituitary carcinomas. J Clin Endocrinol Metabol. 2005;90(5):3089–99.
- Chentli F, Yaker F-A, Azzoug S, Belhimer F. Temozolomide: anti-tumor effect on giant, invasive and resistant pediatric prolactinoma. Indian J Endocrinol Metab. 2013;17(6):1136.
- McCormack AI, McDonald KL, Gill AJ, Clark SJ, Burt MG, Campbell KA, et al. Low O6-methylguanine-DNA methyltransferase (MGMT) expression and response to temozolomide in aggressive pituitary tumours. Clin Endocrinol. 2009;71(2):226–33.
- 97. Vergès B, Boureille F, Goudet P, Murat A, Beckers A, Sassolas G, et al. Pituitary disease in MEN type 1 (MEN1): data from the France-Belgium MEN1 multicenter study. J Clin Endocrinol Metabol. 2002;87(2):457–65.
- Thakker RV, Newey PJ, Walls GV, Bilezikian J, Dralle H, Ebeling PR, et al. Clinical practice guidelines for multiple endocrine neoplasia type 1 (MEN1). J Clin Endocrinol Metabol. 2012;97(9):2990–3011.
- 99. Pellegata NS, Quintanilla-Martinez L, Siggelkow H, Samson E, Bink K, Hofler H, et al. Germ-line mutations in p27Kip1 cause a multiple endocrine neoplasia syndrome in rats and humans. Proc Natl Acad Sci. 2006;103(42):15558–63.
- Stergiopoulos SG, Abu-Asab MS, Tsokos M, Stratakis CA. Pituitary pathology in carney complex patients. Pituitary. 2004;7(2):73–82.
- 101. Korbonits M, Kumar AV. AIP-related familial isolated pituitary adenomas. Seattle: Nih.gov. University of Washington; 2012.
- Rutkowski MJ, Aghi MK. Medical versus surgical treatment of prolactinomas: an analysis of treatment outcomes. Expert Rev Endocrinol Metab. 2018;13(1):25–33.
- 103. Soto-Pedre E, Newey PJ, Bevan JS, Leese GP. Morbidity and mortality in patients with hyperprolactinaemia: the PROLEARS study. Endocr Connect. 2017;6(8):580–8.
- 104. Jasim S, Alahdab F, Ahmed AT, Tamhane S, Prokop LJ, Nippoldt TB, et al. Mortality in adults with hypopituitarism: a systematic review and meta-analysis. Endocrine. 2017;56(1):33–42.



Cushing's Disease

Anna Aulinas and Susan M. Webb

7.1 Introduction

Cushing's syndrome (CS) constitutes a group of signs and symptoms due to extended and inappropriately high exposure to excess glucocorticoids (GCs). Iatrogenic corticosteroid administration is the most common cause of CS, and pituitary corticotroph adenoma (Cushing's disease; CD) is the most frequent cause of endogenous excessive GC secretion. CD is a rare and severe disease, and chronic exposure to high GC levels has been associated with increased multisystemic morbidity and mortality. Therefore, an early diagnosis and treatment are mandatory to avoid long-term complications. Although most of the related comorbidities improve after initial therapy of hypercortisolism, many of them are not completely reversible and a lifelong follow-up is necessary to control comorbidities and rule out potential recurrences. Therefore, prompt identification and treatment of hypercortisolism and its comorbidities are important to reduce mortality. This chapter addresses epidemiology, diagnoses, treatment, outcomes, and follow-up of Cushing's disease.

A. Aulinas (\boxtimes)

S. M. Webb

The chapter has been endorsed by **Prof. Maya Lodish**, Maya.Lodish@ucsf.edu, University of California, San Francisco, California, USA

Endocrinology/Medicine Department, Hospital de la Santa Creu i Sant Pau. Centro Investigación Biomédica en Red de Enfermedades Raras (CIBERER, Unidad 747), IIB-Sant Pau, ISCIII, Barcelona, Spain

Faculty of Medicine, University of Vic (UVIC/UCC), Vic, Barcelona, Spain e-mail: aaulinas@santpau.cat

Endocrinology/Medicine Department, Hospital de la Santa Creu i Sant Pau. Centro Investigación Biomédica en Red de Enfermedades Raras (CIBERER, Unidad 747), IIB-Sant Pau, ISCIII, Barcelona, Spain

Universitat Autònoma de Barcelona (UAB), Barcelona, Spain

[©] Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_7

7.2 Epidemiology

Cushing's syndrome is considered a rare disorder with an estimated incidence between 0.5 and 3.0/10⁶ people/year; however, accurate data on prevalence and its incidence in the general population are scarce [1]. Only countries with national registries or databases provide information on its incidence and prevalence (i.e., incidence ranged from 1.2 to 1.7/10⁶ inhabitants/year in Denmark [2]; standardized incidence rate (SIR) of 0.8/10⁵/year in Iceland [3]; and SIR of 0.07/10⁵/year in Malta [4]). The most frequent cause of endogenous hypercortisolism is an ACTHproducing pituitary adenoma, namely Cushing's disease (CD). The incidence according to an 11-year period population-based study in Denmark is 1.2–1.7/10⁶/ year for CD, followed by an incidence of $0.6/10^6$ /year for adrenal adenomas [2]. Other types of CS are extremely rare. Some data indicate a high proportion of subclinical CS in risk populations such as patients with uncontrolled type 2 diabetes or early-onset osteoporosis [5]. CS is more frequent in females with an estimated female:male ratio of 3:1 and median age at diagnosis of around 40 years [1, 6]. CS is associated with increased mortality (SMR of 3.7 (2.3-5.3)); mortality rates are increased during the first year after diagnosis and in those patients with persistent hypercortisolism after initial therapy [2]. Although mortality risk decreases with remission of hypercortisolism, it still remains increased compared to general population, with circulatory diseases being the most important cause of mortality [7, 8].

7.3 Pathogenesis

Table 7.1 summarizes the different causes of CS. In adult population, the most frequent cause of endogenous CS is an adrenocorticotropic (ACTH)-dependent CS; a corticotrope pituitary adenoma (CD) is the cause of about 70% of ACTH-dependent CS, followed by an ectopic (extrapituitary) ACTH-secreting tumor [1]. CD is characterized by an excessive production of ACTH due to an ACTH-producing pituitary adenoma (90% of which are microadenomas), which induces high levels of cortisol and androgens, often accompanied by diffuse adrenal hyperplasia [9].

The molecular pathophysiology of CD is not completely understood. However, the identification of some molecular genetic abnormalities has provided new insight into the pathogenesis of pituitary ACTH-producing adenomas. Most pituitary tumors are sporadic as a consequence of monoclonal expansion of a mutated cell. CD rarely occurs, although cases have been described, in the context of germline mutations like multiple endocrine neoplasia (MEN) type 1, aryl hydrocarbon receptor-interacting protein (AIP), and cyclin-dependent kinase inhibitor (CDKN1B) genes in patients with MEN1 or MEN4 features and in patients with succinate dehydrogenase subunit mutations [10]. In contrast, somatic mutations of MEN1 or AIP have not been found in sporadic pituitary adenomas or CD. Instead, very rare mutations of the GC receptor gene or proteins related to GC8 receptor functions have been found in CD [11]. Inactivation of the ubiquitin-specific peptidase 8 gene (USP8), present in 35–62% of the corticotroph adenomas, is responsible for the

Etiology	Prevalence	Features
ACTH-dependent	70-80%	
Cushing's disease (Corticotroph adenoma)	60–70%	Age peak around 40 years. F:M ratio 3:1. 50% tumors not visible on MRI. Very rare corticotroph hyperplasia
Ectopic source (ACTH, very rare CRH)	10–15%	Age peak benign 30–40 years. Malignant 50–60 years. Slightly more prevalent in males. Most frequent sources: Small cell lung carcinoma, neuroendocrine tumors of lung, thymus, pancreas
ACTH-independent	20-30%	
Adrenal adenoma	10-20%	Age peak 40-50 years. F:M ratio 6:1
Adrenal carcinoma	5–7%	Age peak <10, 50–60 years. F:M ratio 2:1. Mixed cortisol and androgen production is common
Macronodular adrenal hyperplasia (aberrant G-protein- coupled receptors, autocrine ACTH production, sporadic or familiar ARMC5)	<2%	Age peak 50–60 years. F:M ratio 2.5:1. Autocrine or paracrine ACTH might be produced and contribute to cortisol secretion (ACTH-independent classification might be modified in the future). Bilateral involvement
Micronodular adrenal hyperplasia (primary pigmented nodular adrenocortical disease, isolated or familial with Carney complex)	<1%	Age peak 10–30 years. F:M ratio 0.5:1 if <12 years and 2:1 if >12 years. Adrenal size often normal. Bilateral involvement
McCune Albright syndrome	Very rare	Infants (<6 months), F:M ratio 1:1. Internodular adrenal atrophy

Table 7.1 Causes and prevalence of endogenous Cushing's syndrome

^a*ACTH* adrenocorticotropic hormone, *CRH* corticotropin-releasing hormone, *F:M* female:male, *ARMC5* armadillo repeated containing 5 gene

increased expression of EGFR and ultimately ACTH synthesis. Epidermal growth factor receptor (EGFR) and pituitary-transforming gene (PTTG) can be overexpressed and play a causal role in the development of CD and defects in this pathway may also be a key to therapeutic targets. In animal models, a PTTG inhibitor (R-roscovitine) or an inhibitor of the EGFR inhibitor (gefitinib) inhibited corticotroph tumor growth and features of hypercortisolism [10], nevertheless, the translation of these findings in humans is still unknown.

7.4 Clinical Presentation

Manifestations of hypercortisolism are systemic and symptoms at clinical presentation may be unspecific and highly variable, and the diagnosis of the disease is often delayed up to several months or years. There is no pathognomonic sign or symptom. Table 7.2 summarizes the most frequent clinical presentation according to different series [1]. The most common features at diagnosis are weight gain, hypertension, skin abnormalities, myopathy, and menstrual irregularities. Hypertension and hirsutism present more frequently in ectopic CS than in other causes.

Features that best discriminate	Features also common in the general population and less
CS (higher specificity)	discriminatory of CS (lower specificity)
Decreasing growth in children	Weight gain or obesity (95%) (dorsocervical fat pad, facial
(100%)	fullness)
Facial plethora (90%)	Decreased libido (90%)
Easy bruising and skin atrophy	Menstrual abnormalities (80%)
(80%)	Hirsutism, acne, or female balding (75%)
Proximal myopathy and muscle	Depression, irritability, impaired memory (70%)
weakness (60%)	Early onset hypertension (75%), hyperlipidemia (70%),
Reddish purple striae (>1 cm	glucose intolerance (60%), osteoporosis (50%)
wide) (60%)	Kidney stones (50%)

Table 7.2 Clinical features and signs of Cushing's syndrome (CS) and overlapping conditions and prevalence (%)

Table 7.3 Physiological or pathological conditions associated with hypercortisolism in the absence of Cushing's syndrome (CS) features; however, if there is high clinical suspicion of CS, patients should be screened for hypercortisolism, especially in the pseudo-Cushing group

	Some clinical features of CS may be present
Unlikely to have clinical features of CS	(pseudo-Cushing)
Physical stress (pain, surgery)	Pregnancy
Malnutrition	Depression, bipolar disorder
Hypothalamic amenorrhea (anorexia	Morbid obesity
nervosa)	Alcohol abuse
Intense chronic physical exercise	Poorly controlled diabetes
Cortisol binding globulin excess in	Glucocorticoid resistance
serum	

Several signs and symptoms are more suggestive and specific of hypercortisolism due to catabolic effects of cortisol on skeletal muscles, skin, and connective tissue, including skin atrophy, easy bruising with no trauma, proximal muscle weakness, or unexplained osteoporosis. Increased and impaired distribution of fat depots is one of the most precocious signs, with abnormally increased fat in the trunk, face, and neck. Wide reddish-purple striae of more than 1 cm on the abdomen and limbs and proximal muscle weakness are characteristic features seen in CS. Lower bone mineral density and a high prevalence of fractures are common in CS. Hirsutism, menstrual irregularities, obesity, diabetes, and low libido may also be present, but are also frequent in the general population. Psychopathology (emotional lability, cognitive disturbances, anxiety, depression) is very common. Metabolic syndrome features (dyslipidemia, impaired glucose metabolism, hypertension) together with an hypercoagulability state lead to an increased cardiovascular risk.

Usually, progression of symptomatology is slow, except for ectopic ACTH CS due to a paraneoplastic manifestation of a malignant tumor, where clinical presentation is usually rapid with prominent myopathy, edema, hyperpigmentation, and hypokalemia.

Several physiological and pathological situations can produce an overactivity of the hypothalamic–pituitary–adrenal axis (Table 7.3). Hypercortisolism without signs or symptoms of CS might be found in physical stress (surgery, pain), anorexia

nervosa, malnutrition, cortisol-binding globulin elevation, or hypothalamic amenorrhea; hypercortisolism with mild signs/symptoms of CS might be seen in psychiatric disorders (depression, bipolar disorder), alcoholism, pregnancy, morbid obesity, or poor controlled diabetes mellitus.

7.5 Diagnosis

In this first step of diagnosing CS, hypercortisolism needs to be confirmed using screening tools. Once CS is diagnosed, the cause must be identified to determine the specific etiology in order to guide treatment decisions.

7.5.1 Establishing the Diagnosis of Cushing's Syndrome

Screening is recommended in individuals with a high risk of CS, after excluding current or recent use of any type of exogenous GC (oral, inhaled, topic, rectal, parenteral) and high dose of progestogens [12, 13]. In children with decreasing height percentile together with progressive obesity, suspicion should be raised for CS. In addition, in young adults with hypertension or osteopenia, in patients with adrenal incidentaloma, and in women with menstrual irregularities, screening for CS may be indicated, especially if patients have additional features of the syndrome. Some studies have revealed a higher prevalence of "occult CS" in patients with poorly controlled T2DM or early-onset osteoporosis; however, there is no strong evidence to support a routine screening for occult CS in these patients based on the data available currently [14–17].

Specificity and sensitivity of screening tools vary enormously across studies, and it is often necessary to perform several tests before reaching a diagnosis. Specificity is not optimal, and false positives may occur. Moreover, the use of screening tools and their variability vary across countries [18]. Importantly, it is important to recognize the impact of physiological stress on cortisol levels when interpreting results. Recommended screening tests are outlined in Fig. 7.1:

- 24-hour urinary free cortisol (UFC; at least two measurements) measures the cortisol that is not bound to cortisol-binding globulin (CBG) and thus filtered unchanged by the kidney. Therefore, UFC is not affected by medications or conditions that alter CBG. Concomitant measurement of urine creatinine helps to identify if collection is complete and to interpret collections with excessive volume (false positives with a 24-h diuresis of >5 l/day) [19]. False negatives may be seen in chronic kidney disease, if GFR < 60 ml/min and in cases with mild CS, in whom late-night salivary cortisol (LNSC) might be more useful [13, 20]. Importantly, the use of carbamazepine and fenofibrate or licorice can increase UFC levels.</p>
- 1 mg overnight dexamethasone suppression test (DST; alternatively, 2 mg/day for 48 h). In healthy individuals, the administration of supraphysiological GC



Fig. 7.1 Algorithm for testing patients when Cushing's syndrome (CS) is suspected and for the differential diagnoses of different etiologies once Cushing's syndrome is confirmed. *GC* glucocorticoids, *UFC* urinary free cortisol, *DST* dexamethasone suppression test, *LNSC* late-night salivary cortisol, *Dex-CRH* dexamethasone–corticotropin-releasing hormone test, *ACTH* adrenocorticotropic hormone, *CD* Cushing's disease, *BIPSS* bilateral inferior petrosal sinus sampling, *CT* computed tomography

 Table 7.4
 Drugs that may interfere with dexamethasone metabolism and alter dexamethasone suppression test (DST) results

Accelerate dexamethasone metabolism by	Impair dexamethasone metabolism by	
induction of CYP3A4 (causing false-positive	inhibition of CYP3A4 (causing false-	
results in DST)	negative results in DST)	
Carbamazepine Phenobarbital	Fluoxetine Ritonavir	
Phenytoin Primidone	Diltiazem Itraconazole	
Pioglitazone Rifampicin	Cimetidine	
Ethosuximide Rifapentine	Aprepitant/fosaprepitant	

doses suppresses ACTH and cortisol secretion, while in CD patients there is a failure of this suppression when low doses of dexamethasone are given. Variable absorption, use of concomitant drugs, and renal or liver failure may influence the results of DST. Some drugs enhance hepatic enzymatic clearance of dexamethasone, reducing plasma dexamethasone concentration, and ultimately causing false-positive results of the DST, as well as alcohol abuse (2 weeks of alcohol abstinence is recommended to reduce false-positive results). On the contrary, other agents impair dexamethasone metabolism, increasing plasma dexamethasone levels and causing false-positive results in DST (Table 7.4). Some authors advocate for simultaneous measurement of cortisol and dexamethasone concentrations to ensure appropriate plasma dexamethasone levels (>3.3 nmol/l (>0.13 μ g/dl)); however, dexamethasone measurement is not available in some countries [21]. The recommended serum cortisol cutoff for suppression is 50 nmol/l (1.8 μ g/dl) to maximize the sensitivity as a screening tool.

7 Cushing's Disease

- Late-night salivary cortisol (LNSC; two measurements) is a useful tool to assess the loss of circadian rhythmicity (the absence of LNSC nadir), a consistent abnormality in patients with CS. Salivary cortisol concentrations are in equilibrium with serum active free cortisol, there is a good correlation in cortisol concentrations between both specimens, and salivary cortisol levels are not affected by the rate of saliva production [22]. It is important to note that circadian rhythm might be altered in shift workers, smokers, patients with depression, or in those who are critically ill. Also, false-positive results are commonly seen in patients with obesity or diabetes [23].

To optimize sensitivity of screening tests, it is recommended to use the upper limit of the reference range for UFC and LNSC and a fasting cortisol lower than 50 nmol/L ($1.8 \mu g/dl$) following DST. Two abnormal screening tests are enough to confirm hypercortisolism in individuals with high pretest probability of having CS. For individuals with low pretest probability of having CS, or in those patients in whom cyclical CS is suspected, it is recommended to perform additional testing. In these cases, serial LNSC might be useful to follow the progression. Postponing additional testing to allow progression of biochemical and clinical features might be useful in some cases. Subjects with abnormal results in these tests should be referred to an endocrinologist for further testing [13]; performance and interpretation of subsequent testing require considerable expertise. Assay accuracy and normal ranges differ widely, so it is essential to interpret results in the context of the assay used and with appropriate normal ranges.

It is important to differentiate between endogenous pathological hypercortisolism (CS) and pseudo-Cushing states (alcoholism, depression, anorexia nervosa, obesity). Pseudo-Cushing seems to be mediated via increased hypothalamic secretion of CRH, instead of CRH suppression observed in CS. Commonly, pseudo-Cushing patients have biochemical hypercortisolism but minimum features of CS and no presence of tumor (Table 7.3).

UFC is recommended rather than DST in pregnancy, as well as in situations that can increase/decrease CBG (oral contraceptive pill use, critically ill, or nephrotic patients) or when concomitant use of antiepileptic drugs occurs, as these medications may enhance dexamethasone clearance. However, during pregnancy, only UFC values greater than 3 times the upper limit of normal can be considered for further testing for CS, since UFC excretion physiologically increases up to threefold during pregnancy. In contrast, DST rather than UFC is recommended in severe renal failure and adrenal incidentaloma for initial testing. If cyclical CS is suspected, LNSC or UFC rather than DST is recommended [13].

If further evaluation to rule out a possible non-Cushing's hypercortisolism is required, a *dexamethasone-suppressed CRH stimulation test* might be useful in specific situations. This test involves a 48-h 2 mg/d DST followed by the administration of CRH (1 μ g/kg iv) 2 hours after the last dose of dexamethasone is administered and cortisol measured 15 min later. However, the optimal cutoff for diagnosis needs further clarification and CRH is not available in some countries. The hypercortisolism of pseudo-Cushing's states is thought to be mediated through increased

hypothalamic secretion of CRH in the setting of an HPA axis that is otherwise appropriately restrained by negative feedback from cortisol. In contrast, the hypercortisolism of true CS suppresses hypothalamic CRH secretion and is less responsive to the negative feedback of exogenous CS. Therefore, in comparison with true CS, patients with pseudo-CS states show a greater suppression of cortisol production by exogenous GC and a diminished response to CRH injection [24]. Measurement of <u>midnight serum cortisol</u> while an individual is asleep (>50 nmol/l) provides high sensitivity for the diagnosis of CS, and it is useful to exclude CS when cortisol <50 nmol/l, but this test requires inpatient admission and this approach may not be possible in some clinical practice settings.

7.5.2 Establishing the Etiology of CS

The first step to localize the source of hypercortisolism is the measurement of plasma ACTH (at least twice), although most of the commercially available ACTH assays are not always accurate, especially in the low range. To avoid falsely low results, samples should be collected in an ice bath and processed immediately. Measurement of ACTH can be done at any time of the day because normal circadian rhythm is lost; however, peak values are higher in the morning. In a patient with endogenous hypercortisolism, ACTH concentrations <1.1 pmol/l (< 5 pg/ml) suggest an ACTH-independent origin (an adrenal cause), while ACTH concentrations >4.4 pmol/l (20 pg/ml) suggest an ACTH-dependent origin (mostly pituitary or less frequently an ectopic source). Intermediate values require further evaluation, with a CRH stimulation test that might unmask ACTH responsiveness, or else repeating test over time to either confirm or rule out the diagnosis. The concept that underlies the use of CRH stimulation in CS is that pituitary tumors (CD) usually respond to CRH with an increase in ACTH and cortisol levels, while ectopic ACTH-secreting tumors usually do not. A mean cortisol increase at 30 and 45 minutes of >20% and a mean ACTH increase at 15 and 30 min of >35% over their respective mean baseline values are thought to be consistent with CD [25, 26]. An increase greater than 50% in ACTH levels and/or 20% in cortisol levels following 100 µg of intravenous CRH administration is highly suggestive of CD (Fig. 7.1) [27].

No single best approach to test patients with ACTH-dependent CS exists. The evaluation of CS requires a systematic approach. After biochemical testing has confirmed a diagnosis of ACTH-dependent hypercortisolism, the source of ACTH excess has to be determined. The choice of diagnostic tests used to distinguish a pituitary vs ectopic source varies by institution and depends on the availability of IPSS, cost considerations, patient preference, and availability of CRH and sensitive MRI technology. The most accurate method to discriminate between pituitary and non-pituitary sources of ACTH is to evaluate the central-to-peripheral ACTH gradient via bilateral inferior petrosal sinus sampling (BIPSS). Corticotroph tumors have a clear setup in the petrosal samples, while ectopic ACTH-producing tumors do not. BIPSS is considered the gold standard test to differentiate between a pituitary and an ectopic source of ACTH (sensitivity and specificity of 95%), and it is recommended for

patients with pituitary lesions <6 mm or in those with discordant noninvasive tests [28, 29]. BIPSS is the best way to document a central-to-peripheral ACTH gradient in the blood draining the tumor. Despite the high sensitivity and specificity of BIPSS, false-positive results can occur in ectopic ACTH production with cyclical or mild hypercortisolism without suppression of normal corticotropes, and in CRH-producing tumors. It is recommended to perform BIPSS only in cases with documented hypercortisolism (at least twofold increase in UFC in the 6-8 weeks prior to BIPSS) to ensure that normal corticotroph cells are suppressed. Inability to cannulate veins or abnormal venous drainage might cause false-positive results. Concomitant measurement of prolactin can confirm successful catheterization. CRH is not available in some countries, and desmopressin administration has been shown to offer similar results to CRH stimulation in some studies. BIPSS has limited value in identifying intrapituitary localization of the tumor. If MRI is negative, the gradient of BIPSS might be helpful to choose the side for the initial surgical approach; however, if tumor is not found, the other side of the pituitary gland should be explored. Alternatively, bilateral internal jugular venous sampling has been proposed, since it is more simple and safe and it does not require specialized expertise; however, sensitivity is lower than that of BIPSS [30, 31].

Noninvasive tests to assess the etiology of CD:

- High-dose dexamethasone suppression test (HDDST), CRH, or desmopressin stimulation tests are noninvasive tests that might contribute to localization of the source of CS when tumor is not seen or is very small on an MRI, or when BIPSS is not available. The HDDST relies on the concept that pituitary corticotroph tumor cells retain sensitivity (albeit impaired) to glucocorticoids, but tumor cells in ectopic ACTH secretion (EAS) do not. HDDST (2 mg every 6 h for 48 h) is used to distinguish between pituitary CD and EAS. Accuracy of these tests is inferior to that of BIPSS. In general, CD adenomas maintain sensitivity to CRH or desmopressin stimulation and are resistant to negative feedback regulation by GC (HDDST), while malignant tumors (ectopic ACTH source) do not. However, some benign carcinoid tumors may respond equally to what is observed in CD. Similarly, some ectopic ACTH-secreting tumors might express vasopressin receptors and respond to desmopressin like CD tumors; in these cases, desmopressin test is not helpful in differentiating the source of ACTH. Actually, it is not recommended to use the desmopressin test routinely until more data validating the test become available [13]. Discordant results of these tests are reported in up to 60% of patients, BIPSS being the most recommended one.

Additional testing might be helpful to delineate the source of ACTH, as well as to evaluate comorbidities of CS. Chromogranin A, 5-hydroxy-indoleacetic acid, calcitonin, and gastrin might point to ectopic ACTH tumors. Metabolic alkalosis and hypokalemia as a result of severe hypercortisolism (UFC > 1500 μ g/24 h) are common in ectopic ACTH sources, but it can be also present in 10% of CD. Dehydroepiandrosterone sulfate is normal or increased in ACTH-dependent causes, but is decreased in cases of adrenal origin [32].

7.5.3 Localization of the ACTH Source

Once ACTH-dependent CS is confirmed and noninvasive tests or BIPSS suggest a pituitary origin, pituitary imaging is mandatory. T1 gadolinium contrast <u>MR1</u> identifies pituitary tumors in around 50% of patients with CD. Importantly, roughly 10% of healthy individuals have incidental pituitary lesions up to 6 mm; therefore, a lesion smaller than 6 mm does not always identify CD as a cause of CS (Fig. 7.1). Sensitivity of MRI can be improved up to 80% using more advanced techniques (spoiled gradient recalled acquisition or dynamic MRI sequences) [33]. A cranial CT scan might be requested by the surgeon since it provides better information on bone structures. It has recently been reported that pituitary adenomas not visible via MRI may be detectable by CRH-stimulated 18-F-fluoro-deoxy-glucose PET imaging [34].

7.6 Management

CD requires a multidisciplinary and individualized management strategy. The main goals of therapy are to reverse clinical features, normalize cortisol levels, minimalize morbidity, and reach long-term remission without recurrences. The best treatment options involve a multidisciplinary team, including an endocrinologist experienced in the management of CD. Surgery is considered the first therapeutic option. When surgery is not possible or non-curative, the choice for second-line therapy might depend on several factors, i.e., patient preferences, urgency to treat, location and size of remnant tumor, drug interactions and side effects, and cost and availability of medical therapies.

7.6.1 Surgical Treatment

Selective pituitary adenomectomy using transsphenoidal surgery (TSS; microscopic or endoscopic techniques) by an experienced neurosurgeon remains the first-line treatment for CD. If surgery is successful (pathological confirmation of the adenoma or biochemical demonstration of remission after resection), the patient is cured. Ideally, the whole pituitary tumor is removed and normal pituitary tissue is left. However, pituitary deficiencies as a consequence of surgery might be present after initial therapy and an interval of hypoadrenalism is common since normal corticotroph cells have been suppressed by longstanding hypercortisolism. Reasons for surgery failure include lack of experience of the surgeon, diffuse corticotroph hyperplasia (very rare), invasive tumor that cannot be resected, or adenomas arising in unusual sites (parasellar, pituitary stalk, neurohypophysis). On the contrary, a well-defined, noninvasive (to the cavernous sinus), and well-visualized tumor on MRI, histological confirmation of an ACTH-secreting tumor, low postoperative cortisol levels, and long-lasting adrenal insufficiency are favorable prognostic factors [35].

If surgery is not successful the first time, repeated TSS, medical therapy, and radiotherapy are potential second-line therapies. As a final resource, surgical or medical adrenalectomy can be performed if hypercortisolism is still uncontrolled. The choice of a second-line treatment must be discussed with the patient. Repeated surgery as soon as active and persistent hypercortisolism is confirmed might be undertaken especially if the pituitary tumor is visualized; nevertheless, overall successful rates are lower than for the first surgery and carries a high risk of pituitary insufficiencies [36].

Surgical complications are more likely to occur in macroadenomas or extensive pituitary exploration. Anterior pituitary deficiencies are observed in about 20 to 25% of the cases, as well as transient central diabetes insipidus. Permanent diabetes insipidus is less frequent. Symptomatic hyponatremia might occur between 1 and 10 days after surgery, with a maximum antidiuresis around day 5, in about 8% of the patients [37]. It is recommended to measure serum sodium several times during the first days after surgery. Other complications include venous thrombosis, hemorrhage, and infections [38]. Since a hypercoagulability state exists in CS, perioperative prophylaxis a few days prior to surgery is recommended [39]. Within 2 weeks of surgery, measurement of free T4 and prolactin may help to identify hypopituitarism, when compared to preoperative values.

7.6.2 Radiotherapy

Conventional fractionated photon beam radiotherapy (1.7-2 Gy daily for a total dose of 45 Gy over 6 weeks) or stereotactic radiosurgery (single-dose radiation, including gamma knife, linear accelerator, and proton beam) control hypercortisolemia in up to 80% of patients within 3-5 years [40], but results vary across series [41]. Radiotherapy is indicated in persistent CD after surgery and when local invasion precludes a surgical cure. Radiosurgery may provide a faster biochemical control and less risk of radiation damage to the surrounding structures than conventional radiation therapy, but there are no direct comparative studies. Since medical therapy is effective in normalizing cortisol, it is recommended before administering radiotherapy, since these agents will be required while waiting for the effects of radiation. Long-term follow-up is mandatory to detect both relapse and pituitary insufficiencies. The risk of a second neoplasia (most frequently meningiomas) is estimated in 2% of the patients at 20 years, but further studies are required to confirm these data [42]. Optic neuropathy and other cranial neuropathies may also occur (2%). Measuring UFC without concomitant glucocorticoid supplementation at 6-12 months after radiation therapy is necessary to assess if adrenal insufficiency (AI) has developed. In addition, patients should be counseled on symptoms of AI and to alert their physician if they develop symptoms so that more frequent testing may be undertaken. Diurnal rhythm is not necessarily achieved following radiotherapy, so LNSC is not a good tool to assess remission.

7.6.3 Bilateral Adrenalectomy

In general, bilateral adrenalectomy is considered the last treatment option: It provides definitive and immediate control of hypercortisolism, resulting in permanent hypocortisolism that requires lifelong GC and mineralocorticoid replacement therapy and careful education of the patient. Furthermore, regular pituitary MRI and assessment of hyperpigmentation and ACTH levels are mandatory because of the risk of developing Nelson's syndrome (corticotroph tumor progression) described in up to 25% of patients [43]. If remission is not achieved after the second surgery, the decision of pituitary radiotherapy or bilateral adrenalectomy must be individualized and requires consideration of the pituitary status after the second surgery. Another factor that needs to be taken into account includes patient tolerance to medical therapy while awaiting the effects of radiotherapy. Finally, women who wish to maintain fertility without the need for ovulation induction may opt for adrenalectomy instead of pituitary radiotherapy [44].

7.6.4 Medical Therapy

Medical therapy is indicated in preoperative patients with severe hypercortisolism and associated clinical manifestations, as well as in patients awaiting a response after radiotherapy, and/or when palliative treatment is needed. Hypoadrenalism may occur when treating with these agents; therefore, the possibility of adrenal insufficiency must be addressed if suspected and interrupting the medication may be indicated. Some of these agents affect CYP3A4 leading to significant drug–drug interactions; thus, reviewing all other medications taken by a patient is necessary before starting the treatment. Follow-up for individuals on medical therapy should include UFC and clinical features, aiming for normalization of both [45]. Medical treatments include steroidogenesis inhibitors, agents that modulate ACTH secretion or GC receptor antagonists (Table 7.5).

7.6.4.1 Steroidogenesis Inhibitors

Steroidogenesis inhibitors are effective at blocking the hypercortisolemia despite the fact that they do not treat the underlying tumor. The most commonly used steroidogenesis inhibitors include metyrapone, ketoconazole, mitotane, and etomidate.

– Metyrapone inhibits 11-β hydroxylase (conversion of 11-deoxycortisol to cortisol) and inhibits aldosterone biosynthesis with accumulation of aldosterone precursors that have a weak mineralocorticoid activity. It has a rapid onset of action, and it controls hypercortisolism in 50–75% of the patients with CS [46]. The risk of hirsutism and acne development due to accumulation of androgenic precursors may be metyrapone less desirable as initial therapy in females. Monitoring for edema, hypokalemia, and hypertension is recommended due to the accumulation of mineralocorticoid precursors.

Drug	Dose ^a	Adverse events (AE) and concerns/considerations			
Steroidogenesis inhibitors					
Ketoconazole	400–1600 mg/day every 6–12 h orally Quick onset of action	AE: Gastrointestinal, hepatotoxicity, male hypogonadism and gynecomastia Requires acid for biological activity (avoid use of proton-pump inhibitors) Approved by the European medicines agency (EMA) for the treatment of CS			
Metyrapone	500–4500 mg/day every 6–8 h orally Quick onset of action	AE: Gastrointestinal, hirsutism, hypertension, hypokalemia Accessibility variable across countries. Approved by the EMA for the treatment of CS			
Mitotane	3–5 g/day orally. Starting dose 250 mg/ day. Slow onset of action	AE: Gastrointestinal, gynecomastia, hepatotoxicity, neurotoxicity, teratogenic, hypercholesterolemia. Accumulates in body fat. Approved for the treatment of adrenal cancer			
Etomidate	0.1–0.3 mg/kg/h intravenously (bolus and titrate). Quick onset of action	AE: Gastrointestinal, myoclonus and pain at the injection site Requires monitoring in intensive care unit			
Osilodrostat (LCI699)	4–100 mg/day orally	AE: Gastrointestinal, headache, dizziness, arthralgia, hypokalemia Under investigation. Not approved yet			
Pituitary tumo	r-directed drugs				
Pasireotide	0.6–0.9 mg twice daily 10–20 mg/4 weeks intramuscular	AE: Diarrhea, cholelithiasis, hyperglycemia, transient increase LFTs Most successful when UFC < two-fold normal. Approved for patients with CD who are not surgical candidates or have failed surgery			
Cabergoline	1-7 mg/week orally	AE: Asthenia, dizziness, gastrointestinal, headache, potential risk of cardiac valvulopathy			
Retinoic acid	10–80 mg/day orally	AE: Arthralgia, dryness of mucosae, headache, gastrointestinal Under investigation, not approved yet			
Glucocorticoid	receptor antagonist				
Mifepristone (RU486)	300–1200 mg/day orally Difficult to titrate (no biomarker)	AE: Clinical adrenal insufficiency, endometrial hyperplasia, hypertension, edema, hypokalemia. Approved for patients with CD and diabetes mellitus or glucose intolerance who are not surgical candidates or have failed surgery			

Table 7.5 Medical therapy for Cushing's syndrome

^aThe lowest dose is recommended initially, unless UFC > 5xULN, starting dose must be doubled

- Ketoconazole is an imidazole derivative that inhibits steroidogenesis by blocking side-chain cleavage, and to a lesser degree 17,20-desmolase, and 11-β hydroxy-lase enzymes. It has a rapid onset of action and an acidic environment is required to maximize the absorption; therefore, concomitant use of proton-pump inhibitors is contraindicated. Efficacy reported varies across series; on average, it normalizes UFC levels in 60% of the patients with CD [47]. Liver function should be monitored; mild elevation of liver enzymes, up to threefold, is not a contraindication. Hepatotoxicity is usually mild and resolves after drug withdrawal. The

possibility of development of hypogonadism in men may favor the use of metyrapone as initial medical therapy in men. Recently, ketoconazole was withdrawn from the market due to hepatic dyscrasia for the treatment of fungal infections by the European Medicines Agency; however, it is indicated to control hypercortisolism.

Combination therapy with both ketoconazole and metyrapone can be used to control severe hypercortisolemia.

- Mitotane has a specific adrenolytic action and acts by inhibition of CYP11A1, P450 side-chain cleavage. Mitotane provides a long-term suppression of hyper-cortisolism in patients with CD. However, its onset of action is slow (weeks or months) and potential adverse events may be serious (gastrointestinal and neurological); careful monitoring of drug levels is required and only available in few centers [48]. However, monitoring circulating levels of mitotane is made available by the manufacturer in many countries. Mitotane increases CBG and plasma total cortisol levels; thus, biochemical monitoring relies on UFC or salivary cortisol. Hypoadrenalism is common and requires concomitant supplementation with higher doses of hydrocortisone than in other causes of hypo-adrenalism, because mitotane activates CYP3A4 and increases hydrocortisone clearance.
- Etomidate is an intravenous anesthetic and an imidazole derivative used when rapid control of cortisol levels is required, and oral agents cannot be taken. Etomidate may be useful in emergency settings with unmanageable symptoms (respiratory failure, psychosis). Monitoring cortisol levels every 4–6 h is required to titrate the infusion rate.
- Osilodrostat (LCI699), an oral 11 β-hydroxylase inhibitor, has recently been authorized for its use in Cushing's syndrome by the European Medicine Agency (January 2020). Osilodrostat was effective at lowering the levels of cortisol in at least 80% of the patients compared to placebo, and it was in general well-tolerated and more convenient since it can be given orally twice a day [49].
- Levoketoconazole (COR-003), a ketoconazole stereoisomer, is under investigation. Preliminary results show promising results in the treatment of CD patients; however, it is still being evaluated in multicenter trials [50].

7.6.4.2 Another Modality for Treating CS

Another modality for treating CS includes the class of medications that are *GC* receptor antagonists. One such drug is mifepristone (RU486), which is both a GC receptor antagonist and an anti-progestin. Few experiences with CD are reported, and the assessment of its efficacy without a biochemical marker makes its use challenging and restricts the ability to assess overtreatment or undertreatment. Cortisol levels remain unchanged or increased while on mifepristone and are not useful to guide efficacy of the therapy. Therefore, dose adjustment is based on clinical
symptoms, which have been shown to improve in a significant number of patients (diabetes in 60%, hypertension in 40%, and at least one of the following clinical parameters: weight, depression, quality of life (QoL), or clinical appearance in 87%) [51]. Mifepristone is a second-line therapy in CS relegated for patients with diabetes who failed surgery or those with CS who are not surgical candidates and have persistent disease. CORT125134 is a new selective GC receptor antagonist under investigation.

7.6.4.3 Tumor-Directed Therapeutic Agents

Tumor-directed therapeutic agents act directly on corticotroph cells, inhibiting ACTH production

- Dopamine D2 receptor agonists (bromocriptine, cabergoline) may be effective in a subset of CD patients. Usually, higher doses of cabergoline than the ones used for hyperprolactinemia are required (2–3 mg/week). Efficacy is limited; small studies showed normalization of UFC in 30–40% of patients [52]. Clinical trials are needed to evaluate combination therapy with cabergoline.
- Pasireotide (SOM230) is a somatostatin receptor (SST) agonist with higher and broader affinity for SST 1, 2, 3, and 5 than octreotide or lanreotide. It has been demonstrated to be useful as a tumor-directed medical therapy for CD, since corticotroph tumors have a high expression of SST5. It is approved for patients who have failed surgery or for those with contraindications to surgery. The main side effects of pasireotide include gastrointestinal upset and hyperglycemia. Long-acting pasireotide (monthly administration) has been developed, with a similar safety profile and efficacy to that of twice-daily formulations, normalizing UFC in about 40% of CD patients [53].

Potential new drugs are under investigation as a tumor-directed targets, such as retinoic acid, silibinin, and roscovitine, whose efficacy and safety are still unclear [54].

7.6.5 Perioperative Management

GC replacement before surgery for Cushing's syndrome is not necessary unless the patient has been treated with adrenal enzyme inhibitors. Regimens differ across centers, and no study comparisons between one approach and the other have been done. A typical regimen is the administration of GC during the 24 h prior to transsphenoidal surgery [55].

Almost all patients require GC replacement after transsphenoidal surgery for an ACTH-producing adenoma until the HPA axis is recovered [55]. Some patients may have a "GC withdrawal syndrome" despite the use of physiologic GC replacement therapy, and they should be warned that this is expected after "successful surgery." Occasionally transient supraphysiological doses of GC are needed [56].

7.7 Follow-Up

Initial remission rates are achieved in 80-90% of the patients with microadenomas and 60% of patients with macroadenomas with an experienced neurosurgeon. However, 20% of the patients present late recurrences even 10-15 years after initial therapy [2, 6]. Therefore, lifelong follow-up is mandatory for CD.

Measurement of morning serum cortisol during the first week after surgery is recommended to assess remission by withholding treatment with GCs at least 24 hours [44]. Hydrocortisone (<20 mg) 2–3 times daily is the preferred regimen of GC supplementation, since it has a shorter half-life and supraphysiological doses should be avoided to allow quicker recovery of HPA axis. Usually, GCs are maintained for a few days before withdrawal, being careful about the potential development of adrenal insufficiency. It is recommended to measure morning serum cortisol every 2-3 months, followed by an ACTH stimulation test if cortisol levels are >200 nmol/l (7.3 µg/dl). Alternatively, insulin-induced hypoglycemia can be done to assess recovery of the HPA axis within 6 weeks of surgery [45]. GC can be stopped if morning serum cortisol or cortisol response to ACTH stimulation test is >500 nmol/l (>18 µg/dl). If baseline cortisol is still <138 nmol/l (5 µg/dl), GC replacement should be continued while retesting the patient every 3-6 months. Recovery of the HPA axis may take up to 1 year after TSS. Once HPA axis function is recovered, assessment for possible recurrences should be performed annually or sooner if patients have clinical symptoms. Interestingly, LNSC seems to be one of the earliest biochemically detectable alterations of recurrence, preceding elevations of UFC [57].

There is no consensus on the criteria for "cure" after initial surgery for CD. Persistent elevations of UFC after the immediate postoperative period are indicative of lack of remission. While levels of cortisol at either end of the spectrum are more clear-cut, there are intermediate levels that may present a diagnostic dilemma. It is well accepted that serum cortisol levels <50 nmol/l (<1.8 µg/dl) or UFC levels below the normal range (<55 nmol/24 h, ideally <28 nmol/24 h) define remission and are associated with low recurrence rate of 10% at 10 years and high remission rates at long-term follow-up of 85–100%. On the contrary, persistent cortisol >138 nmol/l (>5 μ g/dl) for up to 6 weeks requires further evaluation [58]. In patients with intermediate postoperative values of serum plasma cortisol (between 50 and 138 nmol/l), management and follow-up should be individualized; a postoperative cortisol cutoff <138 nmol/l (5 µg/dl) is associated with long-term remission rates of between 65 and 80%. Sometimes cortisol levels decline gradually over the months following surgery, indicating a progressive necrosis of the remaining tumor cells. Therefore, it is important to confirm that levels reached a nadir, before additional testing and therapy are prescribed in patients with persisting hypercortisolism immediately after surgery [59]. Young individuals (<25 years) and patients with

macroadenomas are at a higher risk of recurrences [60]. Some patients might present a gradual decline of cortisol levels over the months following surgery, so followup is mandatory.

The true long-term recurrence rate is uncertain because of different criteria of remission and inadequate follow-up. The median interval to recurrence is about 40 months but varies across series and duration of follow-up. Postoperative serum cortisol levels or UFC might provide prognostic information. Long-term monitoring should be done annually for several years using LNSC, DST, and/or UFC or at any time that patients experience a return of symptomatology, to rule out recurrences. Documentation of recovery of a diurnal cortisol pattern might be helpful to support a remission state.

Testing for GH deficiency is advised at least 12 months after remission, because hypercortisolism affects GH axis [61] and a postoperative MRI scan is recommended beyond 3 months after surgery [45].

7.8 Prognosis

Although biochemical remission is associated with significant clinical improvement, some symptoms and comorbidities may not completely normalize [7]. Therefore, it is mandatory to monitor and provide additional treatment for cortisoldependent comorbidities throughout the life of a CS survivor (Fig. 7.2).



Fig. 7.2 Main clinical manifestation after remission of Cushing's syndrome

7.8.1 Mortality

Higher mortality, mainly due to cardiovascular and infectious diseases, is observed within the first year after diagnosis and initial therapy, and in those patients whose initial cure was not obtained. Before the development of steroidogenesis inhibitors and surgical techniques, prognosis was poor with a mean survival of 4 years [62]. In patients who have persistent mild hypercortisolism despite therapy, SMR is increased 3.8- to 5.0-fold compared with the general population [2, 8]. Although more recent studies show that mortality rates have been dramatically reduced after successful normalization of cortisol, mortality is still increased up to 2–3 times in most of the series, while others reported similar SMR to that of age-matched populations [60, 63, 64].

7.8.2 Cardiovascular Risk and Metabolic Syndrome

Cardiovascular risk and metabolic syndrome may remain abnormal after years of cortisol normalization. Hypertension, glucose, and lipid metabolism improve after normalization of hypercortisolism, but a higher risk of diabetes, dyslipidemia, and hypertension remains years after remission compared to a healthy population or patients with nonfunctioning pituitary adenomas [7, 65]. Moreover, persistence of central obesity contributes to insulin resistance and an increased inflammatory and hypercoagulable state with an unfavorable adipokine profile (increased levels of TNF-alpha, interleukin-6, and leptin) contributing to persistence of increased cardiovascular risk. An increased risk of myocardial infarction, greater prevalence of coronary calcifications, left ventricular dysfunction, and cerebrovascular disease have also been reported in survivors of CS [7, 63–66].

7.8.3 Psychopathology and Cognition

Even years after CD is cured, patients have an increased prevalence of psychopathology (anxiety disorders, major depression, maladaptive personality) compared to healthy individuals or patients treated for other types of pituitary diseases [7]. The use of screening tools in clinical practice to identify psychopathology promptly is recommended [67]. Cognitive impairment (short-term memory and attention deficits, as well as working memory and impaired decision making) is a common complaint after hypercortisolism [68]. Chronic hypercortisolism exposure causes structural changes in cerebral areas; specifically, structural brain abnormalities (more severe white matter lesions commonly associated with increased cardiovascular risk, smaller gray matter volumes in the anterior cingulate cortex, greater cerebellar volumes, hippocampal dysfunction) were found in patients in long-term remission of their CS compared to matched controls [69–72]. Fluoxetine has been suggested as a neuroprotectant and antidepressant for these patients, but no prospective studies are available yet [73]. Data on long-term recovery regarding cognitive impairment are scarce.

7.8.4 Bone Metabolism

Factors involved in higher risk of osteoporosis in CD are multiple: direct effects of GC on bone cells, GC-induced hypogonadism, hypopituitarism secondary to surgery, hydrocortisone replacement therapy, or reduced bone strain due to muscle atrophy [74]. Approximately 40% of CS patients present with fractures, particularly vertebral fractures. Although there is a decrease in bone mineral density (BMD) and increased fracture risk during hypercortisolism and before diagnosis, studies report BMD normalization after several years of cure [75, 76]. Meanwhile, in severe osteopenia, alendronate might be useful to induce a more rapid improvement of BMD or to prevent further bone loss in persistent postsurgical hypercortisolism [77]. Time of excess GC exposure and duration of GC replacement after surgery were found to be predictors of low BMD in CS. A detailed fracture assessment is recommended, evaluating BMD at the spine and hip, adequate calcium and vitamin D supplementation, and individualized long-term follow-up accordingly.

7.8.5 Quality of Life

Because of all comorbidities associated with CS despite cure, health-related quality of life is likely to be affected long term. Perceived QoL is impaired especially in patients with CD [7, 78], even independently of the disease control or adequately replaced hypopituitarism [2], compared to healthy individuals or other pituitary tumors. Although QoL improves in patients in remission, long-term residual impairment in physical and social functioning is commonly reported. Altogether, it has a significant impact, with social and economic consequences, since inability to return to work or sick days is common [73, 79].

7.8.6 Other Cortisol-Related Comorbidities

CS patients have a significant increased risk of venous thromboembolic events and impaired coagulation profile (activation of coagulation cascades and impaired fibrinolysis) compared to the general population, especially in the postoperative period [80]. Muscle myopathy may persist long-term after cure; aerobic and resistance exercises might be effective in attenuating GC-induced muscle atrophy. Increased rate of nephrolithiasis in both active and cured CD compared to healthy controls has been reported, but the pathogenic mechanisms in CS are not yet elucidated. Importantly, resolution of hypercortisolism is associated with new onset or exacerbation of pre-existing autoimmune or inflammatory diseases (psoriasis, rheumatoid arthritis, ulcerative colitis, etc.), likely due to the suppressive effects of hypercortisolism on the immune system [81]. The most common autoimmune disease is autoimmune thyroiditis; therefore, patients with positive antibodies should be followed after remission of hypercortisolism to identify precociously the onset of a hypothyroidism.

7.9 Summary and Conclusions

CD is a rare disorder characterized by chronic exposure to excess glucocorticoids leading to multisystemic comorbidities and increased mortality. Although clinical presentation is often nonspecific and hypercortisolism might not be suspected, a prompt diagnosis is mandatory to reduce long-term comorbidities due to hypercortisolism. Pituitary surgery can be curative in most of the cases but not all. Radiotherapy and medical therapy can be used as additional therapeutic options. Fortunately, new medical therapies are under development with promising results. A multidisciplinary and individualized approach is essential to choose the best treatment approach, minimize complications, and manage cortisol-related comorbidities, since some of them persist even after hormonal normalization. A lifelong surveillance is advisable to identify recurrences and treatment of persistent comorbidities.

References

- Valassi E, Santos A, Yaneva M, Toth M, Strasburger CJ, Chanson P, et al. The European Registry on Cushing's syndrome: 2-year experience. Baseline demographic and clinical characteristics. Eur J Endocrinol. 2011;165(3):383–92.
- Lindholm J, Juul S, Jorgensen JO, Astrup J, Bjerre P, Feldt-Rasmussen U, et al. Incidence and late prognosis of cushing's syndrome: a population-based study. J Clin Endocrinol Metab. 2001;86(1):117–23.
- Agustsson TT, Baldvinsdottir T, Jonasson JG, Olafsdottir E, Steinthorsdottir V, Sigurdsson G, et al. The epidemiology of pituitary adenomas in Iceland, 1955-2012: a nationwide populationbased study. Eur J Endocrinol. 2015;173(5):655–64.
- Gruppetta M, Mercieca C, Vassallo J. Prevalence and incidence of pituitary adenomas: a population based study in Malta. Pituitary. 2013;16(4):545–53.
- Leon-Justel A, Madrazo-Atutxa A, Alvarez-Rios AI, Infantes-Fontan R, Garcia-Arnes JA, Lillo-Munoz JA, et al. A probabilistic model for Cushing's syndrome screening in at-risk populations: a prospective multicenter study. J Clin Endocrinol Metab. 2016;101(10):3747–54.
- Steffensen C, Bak AM, Rubeck KZ, Jorgensen JO. Epidemiology of Cushing's syndrome. Neuroendocrinology. 2010;92(Suppl 1):1–5.
- 7. Feelders RA, Pulgar SJ, Kempel A, Pereira AM. The burden of Cushing's disease: clinical and health-related quality of life aspects. Eur J Endocrinol. 2012;167(3):311–26.
- 8. Etxabe J, Vazquez JA. Morbidity and mortality in Cushing's disease: an epidemiological approach. Clin Endocrinol. 1994;40(4):479–84.

- Sharma ST, Nieman LK, Feelders RA. Cushing's syndrome: epidemiology and developments in disease management. Clin Epidemiol. 2015;7:281–93.
- Lacroix A, Feelders RA, Stratakis CA, Nieman LK. Cushing's syndrome. Lancet. 2015;386(9996):913–27.
- Karl M, Lamberts SW, Koper JW, Katz DA, Huizenga NE, Kino T, et al. Cushing's disease preceded by generalized glucocorticoid resistance: clinical consequences of a novel, dominantnegative glucocorticoid receptor mutation. Proc Assoc Am Physicians. 1996;108(4):296–307.
- Mann M, Koller E, Murgo A, Malozowski S, Bacsanyi J, Leinung M. Glucocorticoid-like activity of megestrol. A summary of Food and Drug Administration experience and a review of the literature. Arch Intern Med. 1997;157(15):1651–6.
- Nieman LK, Biller BM, Findling JW, Newell-Price J, Savage MO, Stewart PM, et al. The diagnosis of Cushing's syndrome: an Endocrine Society Clinical Practice Guideline. J Clin Endocrinol Metab. 2008;93(5):1526–40.
- Tabarin A, Perez P. Pros and cons of screening for occult Cushing syndrome. Nat Rev Endocrinol. 2011;7(8):445–55.
- Catargi B, Rigalleau V, Poussin A, Ronci-Chaix N, Bex V, Vergnot V, et al. Occult Cushing's syndrome in type-2 diabetes. J Clin Endocrinol Metab. 2003;88(12):5808–13.
- Reimondo G, Pia A, Allasino B, Tassone F, Bovio S, Borretta G, et al. Screening of Cushing's syndrome in adult patients with newly diagnosed diabetes mellitus. Clin Endocrinol. 2007;67(2):225–9.
- Chiodini I, Vainicher CE, Morelli V, Palmieri S, Cairoli E, Salcuni AS, et al. Mechanisms in endocrinology: endogenous subclinical hypercortisolism and bone: a clinical review. Eur J Endocrinol. 2016;175(6):R265–R82.
- Valassi E, Franz H, Brue T, Feelders RA, Netea-Maier R, Tsagarakis S, et al. Diagnostic tests for Cushing's syndrome differ from published guidelines: data from ERCUSYN. Eur J Endocrinol. 2017;176(5):613–24.
- 19. Mericq MV, Cutler GB Jr. High fluid intake increases urine free cortisol excretion in normal subjects. J Clin Endocrinol Metab. 1998;83(2):682–4.
- Chan KC, Lit LC, Law EL, Tai MH, Yung CU, Chan MH, et al. Diminished urinary free cortisol excretion in patients with moderate and severe renal impairment. Clin Chem. 2004;50(4):757–9.
- Ueland GA, Methlie P, Kellmann R, Bjorgaas M, Asvold BO, Thorstensen K, et al. Simultaneous assay of cortisol and dexamethasone improved diagnostic accuracy of the dexamethasone suppression test. Eur J Endocrinol. 2017;176(6):705–13.
- Dorn LD, Lucke JF, Loucks TL, Berga SL. Salivary cortisol reflects serum cortisol: analysis of circadian profiles. Ann Clin Biochem. 2007;44(Pt 3):281–4.
- Liu H, Bravata DM, Cabaccan J, Raff H, Ryzen E. Elevated late-night salivary cortisol levels in elderly male type 2 diabetic veterans. Clin Endocrinol. 2005;63(6):642–9.
- Yanovski JA, Cutler GB Jr, Chrousos GP, Nieman LK. Corticotropin-releasing hormone stimulation following low-dose dexamethasone administration. A new test to distinguish Cushing's syndrome from pseudo-Cushing's states. JAMA. 1993;269(17):2232–8.
- Nieman LK, Oldfield EH, Wesley R, Chrousos GP, Loriaux DL, Cutler GB Jr. A simplified morning ovine corticotropin-releasing hormone stimulation test for the differential diagnosis of adrenocorticotropin-dependent Cushing's syndrome. J Clin Endocrinol Metab. 1993;77(5):1308–12.
- Chrousos GP, Schulte HM, Oldfield EH, Gold PW, Cutler GB Jr, Loriaux DL. The corticotropinreleasing factor stimulation test. An aid in the evaluation of patients with Cushing's syndrome. N Engl J Med. 1984;310(10):622–6.
- Arnaldi G, Angeli A, Atkinson AB, Bertagna X, Cavagnini F, Chrousos GP, et al. Diagnosis and complications of Cushing's syndrome: a consensus statement. J Clin Endocrinol Metab. 2003;88(12):5593–602.
- Nieman LK, Ilias I. Evaluation and treatment of Cushing's syndrome. Am J Med. 2005;118(12):1340–6.

- Newell-Price J, Bertagna X, Grossman AB, Nieman LK. Cushing's syndrome. Lancet. 2006;367(9522):1605–17.
- Swearingen B, Katznelson L, Miller K, Grinspoon S, Waltman A, Dorer DJ, et al. Diagnostic errors after inferior petrosal sinus sampling. J Clin Endocrinol Metab. 2004;89(8):3752–63.
- Colao A, Faggiano A, Pivonello R, Pecori Giraldi F, Cavagnini F, Lombardi G, et al. Inferior petrosal sinus sampling in the differential diagnosis of Cushing's syndrome: results of an Italian multicenter study. Eur J Endocrinol. 2001;144(5):499–507.
- 32. Hong AR, Kim JH, Hong ES, Kim IK, Park KS, Ahn CH, et al. Limited diagnostic utility of plasma adrenocorticotropic hormone for differentiation between adrenal Cushing syndrome and Cushing disease. Endocrinol Metab (Seoul). 2015;30(3):297–304.
- Grober Y, Grober H, Wintermark M, Jane JA, Oldfield EH. Comparison of MRI techniques for detecting microadenomas in Cushing's disease. J Neurosurg. 2018;128(4):1051–7.
- Boyle J, Patronas NJ, Smirniotopoulos J, Herscovitch P, Dieckman W, Millo C, et al. CRH stimulation improves (18)F-FDG-PET detection of pituitary adenomas in Cushing's disease. Endocrine. 2019;
- 35. Bochicchio D, Losa M, Buchfelder M. Factors influencing the immediate and late outcome of Cushing's disease treated by transsphenoidal surgery: a retrospective study by the European Cushing's Disease Survey Group. J Clin Endocrinol Metab. 1995;80(11):3114–20.
- 36. Hofmann BM, Hlavac M, Kreutzer J, Grabenbauer G, Fahlbusch R. Surgical treatment of recurrent Cushing's disease. Neurosurgery. 2006;58(6):1108–18. discussion -18
- 37. Adams JR, Blevins LS Jr, Allen GS, Verity DK, Devin JK. Disorders of water metabolism following transsphenoidal pituitary surgery: a single institution's experience. Pituitary. 2006;9(2):93–9.
- Prevedello DM, Pouratian N, Sherman J, Jane JA Jr, Vance ML, Lopes MB, et al. Management of Cushing's disease: outcome in patients with microadenoma detected on pituitary magnetic resonance imaging. J Neurosurg. 2008;109(4):751–9.
- 39. van der Pas R, de Bruin C, Leebeek FW, de Maat MP, Rijken DC, Pereira AM, et al. The hypercoagulable state in Cushing's disease is associated with increased levels of procoagulant factors and impaired fibrinolysis, but is not reversible after short-term biochemical remission induced by medical therapy. J Clin Endocrinol Metab. 2012;97(4):1303–10.
- 40. Petit JH, Biller BM, Yock TI, Swearingen B, Coen JJ, Chapman P, et al. Proton stereotactic radiotherapy for persistent adrenocorticotropin-producing adenomas. J Clin Endocrinol Metab. 2008;93(2):393–9.
- 41. Starke RM, Williams BJ, Vance ML, Sheehan JP. Radiation therapy and stereotactic radiosurgery for the treatment of Cushing's disease: an evidence-based review. Curr Opin Endocrinol Diabetes Obes. 2010;17(4):356–64.
- Minniti G, Traish D, Ashley S, Gonsalves A, Brada M. Risk of second brain tumor after conservative surgery and radiotherapy for pituitary adenoma: update after an additional 10 years. J Clin Endocrinol Metab. 2005;90(2):800–4.
- Assie G, Bahurel H, Coste J, Silvera S, Kujas M, Dugue MA, et al. Corticotroph tumor progression after adrenalectomy in Cushing's disease: a reappraisal of Nelson's syndrome. J Clin Endocrinol Metab. 2007;92(1):172–9.
- 44. Biller BM, Grossman AB, Stewart PM, Melmed S, Bertagna X, Bertherat J, et al. Treatment of adrenocorticotropin-dependent Cushing's syndrome: a consensus statement. J Clin Endocrinol Metab. 2008;93(7):2454–62.
- 45. Nieman LK, Biller BM, Findling JW, Murad MH, Newell-Price J, Savage MO, et al. Treatment of Cushing's syndrome: an Endocrine Society Clinical Practice Guideline. J Clin Endocrinol Metab. 2015;100(8):2807–31.
- 46. Ceccato F, Zilio M, Barbot M, Albiger N, Antonelli G, Plebani M, et al. Metyrapone treatment in Cushing's syndrome: a real-life study. Endocrine. 2018;62(3):701–11.
- 47. Castinetti F, Guignat L, Giraud P, Muller M, Kamenicky P, Drui D, et al. Ketoconazole in Cushing's disease: is it worth a try? J Clin Endocrinol Metab. 2014;99(5):1623–30.

- Baudry C, Coste J, Bou Khalil R, Silvera S, Guignat L, Guibourdenche J, et al. Efficiency and tolerance of mitotane in Cushing's disease in 76 patients from a single center. Eur J Endocrinol. 2012;167(4):473–81.
- 49. Pivonello R, Fleseriu M, Newell-Price J, Bertagna X, Findling J, Shimatsu A, et al. Efficacy and safety of osilodrostat in patients with Cushing's disease (LINC 3): a multicentre phase III study with a double-blind, randomised withdrawal phase. Lancet Diabetes Endocrinol. 2020;8(9):748–61.
- Feelders RA, Newell-Price J, Pivonello R, Nieman LK, Hofland LJ, Lacroix A. Advances in the medical treatment of Cushing's syndrome. Lancet Diabetes Endocrinol. 2019;7(4):300–12.
- 51. Fleseriu M, Biller BM, Findling JW, Molitch ME, Schteingart DE, Gross C, et al. Mifepristone, a glucocorticoid receptor antagonist, produces clinical and metabolic benefits in patients with Cushing's syndrome. J Clin Endocrinol Metab. 2012;97(6):2039–49.
- 52. Pivonello R, De Martino MC, Cappabianca P, De Leo M, Faggiano A, Lombardi G, et al. The medical treatment of Cushing's disease: effectiveness of chronic treatment with the dopamine agonist cabergoline in patients unsuccessfully treated by surgery. J Clin Endocrinol Metab. 2009;94(1):223–30.
- 53. Lacroix A, Gu F, Gallardo W, Pivonello R, Yu Y, Witek P, et al. Efficacy and safety of oncemonthly pasireotide in Cushing's disease: a 12 month clinical trial. Lancet Diabetes Endocrinol. 2018;6(1):17–26.
- Theodoropoulou M, Reincke M. Tumor-directed therapeutic targets in Cushing disease. J Clin Endocrinol Metab. 2019;104(3):925–33.
- 55. Prete A, Corsello SM, Salvatori R. Current best practice in the management of patients after pituitary surgery. Ther Adv Endocrinol Metab. 2017;8(3):33–48.
- 56. Hochberg Z, Pacak K, Chrousos GP. Endocrine withdrawal syndromes. Endocr Rev. 2003;24(4):523–38.
- 57. Danet-Lamasou M, Asselineau J, Perez P, Vivot A, Nunes ML, Loiseau H, et al. Accuracy of repeated measurements of late-night salivary cortisol to screen for early-stage recurrence of Cushing's disease following pituitary surgery. Clin Endocrinol. 2015;82(2):260–6.
- Hammer GD, Tyrrell JB, Lamborn KR, Applebury CB, Hannegan ET, Bell S, et al. Transsphenoidal microsurgery for Cushing's disease: initial outcome and long-term results. J Clin Endocrinol Metab. 2004;89(12):6348–57.
- Valassi E, Biller BM, Swearingen B, Pecori Giraldi F, Losa M, Mortini P, et al. Delayed remission after transphenoidal surgery in patients with Cushing's disease. J Clin Endocrinol Metab. 2010;95(2):601–10.
- Swearingen B, Biller BM, Barker FG 2nd, Katznelson L, Grinspoon S, Klibanski A, et al. Long-term mortality after transsphenoidal surgery for Cushing disease. Ann Intern Med. 1999;130(10):821–4.
- Tzanela M, Karavitaki N, Stylianidou C, Tsagarakis S, Thalassinos NC. Assessment of GH reserve before and after successful treatment of adult patients with Cushing's syndrome. Clin Endocrinol. 2004;60(3):309–14.
- 62. Plotz CM, Knowlton AI, Ragan C. The natural history of Cushing's syndrome. Am J Med. 1952;13(5):597–614.
- Dekkers OM, Horvath-Puho E, Jorgensen JO, Cannegieter SC, Ehrenstein V, Vandenbroucke JP, et al. Multisystem morbidity and mortality in Cushing's syndrome: a cohort study. J Clin Endocrinol Metab. 2013;98(6):2277–84.
- 64. Clayton RN, Raskauskiene D, Reulen RC, Jones PW. Mortality and morbidity in Cushing's disease over 50 years in Stoke-on-Trent, UK: audit and meta-analysis of literature. J Clin Endocrinol Metab. 2011;96(3):632–42.
- 65. Webb SM, Mo D, Lamberts SW, Melmed S, Cavagnini F, Pecori Giraldi F, et al. Metabolic, cardiovascular, and cerebrovascular outcomes in growth hormone-deficient subjects with previous cushing's disease or non-functioning pituitary adenoma. J Clin Endocrinol Metab. 2010;95(2):630–8.

- 66. Barahona MJ, Resmini E, Vilades D, Pons-Llado G, Leta R, Puig T, et al. Coronary artery disease detected by multislice computed tomography in patients after long-term cure of Cushing's syndrome. J Clin Endocrinol Metab. 2013;98(3):1093–9.
- Santos A, Resmini E, Pascual JC, Crespo I, Webb SM. Psychiatric symptoms in patients with Cushing's syndrome: prevalence. Diagn Manage Drugs. 2017;77(8):829–42.
- Tiemensma J, Kokshoorn NE, Biermasz NR, Keijser BJ, Wassenaar MJ, Middelkoop HA, et al. Subtle cognitive impairments in patients with long-term cure of Cushing's disease. J Clin Endocrinol Metab. 2010;95(6):2699–714.
- Santos A, Resmini E, Gomez-Anson B, Crespo I, Granell E, Valassi E, et al. Cardiovascular risk and white matter lesions after endocrine control of Cushing's syndrome. Eur J Endocrinol. 2015;173(6):765–75.
- 70. Andela CD, van der Werff SJ, Pannekoek JN, van den Berg SM, Meijer OC, van Buchem MA, et al. Smaller grey matter volumes in the anterior cingulate cortex and greater cerebellar volumes in patients with long-term remission of Cushing's disease: a case-control study. Eur J Endocrinol. 2013;169(6):811–9.
- Resmini E, Santos A, Gomez-Anson B, Lopez-Mourelo O, Pires P, Vives-Gilabert Y, et al. Hippocampal dysfunction in cured Cushing's syndrome patients, detected by (1) H-MRspectroscopy. Clin Endocrinol. 2013;79(5):700–7.
- Bourdeau I, Bard C, Noel B, Leclerc I, Cordeau MP, Belair M, et al. Loss of brain volume in endogenous Cushing's syndrome and its reversibility after correction of hypercortisolism. J Clin Endocrinol Metab. 2002;87(5):1949–54.
- Santos A, Crespo I, Aulinas A, Resmini E, Valassi E, Webb SM. Quality of life in Cushing's syndrome. Pituitary. 2015;18(2):195–200.
- 74. Toth M, Grossman A. Glucocorticoid-induced osteoporosis: lessons from Cushing's syndrome. Clin Endocrinol. 2013;79(1):1–11.
- 75. Kristo C, Jemtland R, Ueland T, Godang K, Bollerslev J. Restoration of the coupling process and normalization of bone mass following successful treatment of endogenous Cushing's syndrome: a prospective, long-term study. Eur J Endocrinol. 2006;154(1):109–18.
- Vestergaard P, Lindholm J, Jorgensen JO, Hagen C, Hoeck HC, Laurberg P, et al. Increased risk of osteoporotic fractures in patients with Cushing's syndrome. Eur J Endocrinol. 2002;146(1):51–6.
- 77. Mancini T, Doga M, Mazziotti G, Giustina A. Cushing's syndrome and bone. Pituitary. 2004;7(4):249–52.
- 78. Valassi E, Feelders R, Maiter D, Chanson P, Yaneva M, Reincke M, et al. Worse health-related quality of life at long-term follow-up in patients with Cushing's disease than patients with cortisol producing adenoma. Data from the ERCUSYN. Clin Endocrinol (Oxf). 2018;88(6):787–98.
- van Aken MO, Pereira AM, Biermasz NR, van Thiel SW, Hoftijzer HC, Smit JW, et al. Quality of life in patients after long-term biochemical cure of Cushing's disease. J Clin Endocrinol Metab. 2005;90(6):3279–86.
- Wagner J, Langlois F, Lim DST, McCartney S, Fleseriu M. Hypercoagulability and risk of venous thromboembolic events in endogenous Cushing's syndrome: a systematic metaanalysis. Front Endocrinol. 2018;9:805.
- Pivonello R, Isidori AM, De Martino MC, Newell-Price J, Biller BM, Colao A. Complications of Cushing's syndrome: state of the art. Lancet Diabetes Endocrinol. 2016;4(7):611–29.

TSH-oma



8

E. Peverelli, E. Giardino, D. Treppiedi, R. Catalano, F. Mangili, and G. Mantovani

8.1 Introduction

Thyrotropin (TSH)-secreting pituitary tumor (TSH-oma) is a rare disease. The excess of TSH secretion from tumoral cells, which are unresponsive to the negative feedback of thyroid hormones, leads to hyperstimulation of the thyroid with consequent hypersecretion of T4 and T3 [1–3]. Thus, TSH-omas can be classified as a form of "central hyperthyroidism."

In 1960, the first case of TSH-oma was documented by measuring serum TSH levels with a bioassay [4], and 10 years later, another case was proved by a RIA assay of TSH [5].

TSH-omas were typically diagnosed at the stage of invasive macroadenoma and were considered difficult to cure, but nowadays they are more often diagnosed at an earlier stage. Indeed, in the last years the routine use of ultrasensitive immunometric assays for TSH as first-line test of thyroid function has allowed to detect

The chapter has been endorsed by **Prof. Davide Carvalho**, davideccarvalho@gmail.com, Faculty of Medicine, University of Porto, Alameda Prof. Hernani Monteiro, Porto, Portugal

E. Peverelli · E. Giardino · D. Treppiedi · R. Catalano · F. Mangili

Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

G. Mantovani (⊠) Department of Clinical Sciences and Community Health, University of Milan, Milan, Italy

Endocrinology Unit, Fondazione IRCCS Ca' Granda Ospedale Maggiore Policlinico, Milan, Italy

e-mail: giovanna.mantovani@unimi.it

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_8

hyperthyroid patients with unsuppressed TSH secretion in the presence of high free thyroid hormone, improving the diagnostic workup. The recent introduction of the so-called TSH with reflex free T4 strategy (i.e., FT4 measurement only in the presence of abnormal TSH) fails to recognize both central hypo- and hyperthyroidism, thus leading to TSH-omas misdiagnosis.

Thyroid function tests that value characteristic of TSH-oma patients are also recorded in patients with resistance to thyroid hormones (RTHs), and signs and symptoms of hyperthyroidism are frequently found in the so-called pituitary RTH (PRTH), characterized by a more severe resistance to thyroid hormone action at the pituitary level than at the peripheral tissue level [6–8].

These rare entities represent a diagnostic and therapeutic challenge, because failure in distinguishing between the two disorders may result in improper treatment, while the correct identification of a TSH-oma may prevent the occurrence of both neurological and endocrinological complications, thus leading to a better cure rate.

8.2 Epidemiology

TSH-oma is a very rare disorder; it accounts for about 0.5-2% of all pituitary tumors. The prevalence in the general population is 1-2 cases per million [1, 3]. However, an increase in the number of reported cases was registered in the last decade, confirmed by data obtained from the Swedish Pituitary Registry [9]. This recent study demonstrated an increasing incidence of TSH-omas over time, with a national prevalence of 2.8 per one million inhabitants in 2010. The increased incidence of TSH-omas is principally due to a reasonable catch up effect resulting from improved diagnostic tools. The introduction of the ultrasensitive TSH assays in the late 1980s, which detects low TSH, allowed a distinction between primary hyperthyroidism (Graves' disease) and syndrome of inappropriate TSH secretion [1, 3, 6, 8].

TSH-omas has been reported at ages ranging from 8 to 84 years [3, 10, 11], with a peak of onset in the fifth/sixth decade of life. In contrast with other more common thyroid disorders, TSH-omas occur with equal frequency in men and women and occur mainly as sporadic forms; nevertheless, familial cases can be part of multiple endocrine neoplasia type 1 (MEN1) syndrome [12] and familial isolated pituitary adenoma (FIPA) with AIP mutation [13].

8.3 Pathogenesis

TSH-omas can be classified in functional and silent tumors. Classically, TSH-omas present with elevated thyroid hormones and elevated or inappropriately normal TSH levels. Patients with TSH-secreting adenomas present signs and symptoms of hyper-thyroidism, whereas silent thyrotropinomas are positive for β -TSH by immunohistochemistry without clinical or biochemical evidence of central hyperthyroidism [14, 15].

The presence of TSH beta subunit, either free or combined, has been confirmed in all tumor cells from every type of thyrotropinoma by immunostaining studies, with few exceptions [1, 3, 16–18].

In 70% of the cases, thyrotropinomas secrete TSH only, often accompanied by unbalanced hypersecretion of alpha-subunit of glycoprotein hormones (alpha-GSU) (Table 8.1). Interestingly, by double gold particle immunostaining, the existence of thyrotropinoma composed of two different cell types, one secreting alpha-GSU alone and another cosecreting alpha-GSU and the entire TSH molecules (mixed TSH/alpha-GSU tumors), has been documented [17]. In particular, the presence of a mixed TSH/alpha-GSU adenoma is suggested by the finding of an extremely high alpha-GSU/TSH molar ratio and/or by the observation of dissociated alpha-GSU and TSH responses to TRH [1, 19].

The remaining 30% of cases are classically considered mixed tumors since they cosecrete TSH and other anterior pituitary hormones (GH, PRL, and LH/FSH) (Table 8.1). In this regard, hypersecretion of GH and/or PRL is the most frequently found, possibly leading to acromegaly and/or amenorrhea/galactorrhea syndrome. The occurrence of these mixed tumors may rely on the expression of common transcription factors, such as Prop-1 and Pit-1, by thyrotroph, somatotroph, and lactotroph cells. On the contrary, no association with ACTH hypersecretion has been documented so far, probably due to the distant origin of corticotroph and thyrotroph lineages. Rare is the occurrence of mixed TSH/FSH/LH tumors [20].

It has to be taken into account that silent TSH-secreting tumors may give positive immunohistochemical results for one or more pituitary hormones without a

	Number of cases	% of total
Total TSH-secreting tumors	470	
Pure TSHsecreting tumors	332	70.6
TSH-secreting tumors associated with other pituitary hormones hypersecretion (mixed tumors)	138	29.4
Mixed TSH/GH-secreting tumors	83	17.7
Mixed TSH/PRL-secreting tumors	47	10.0
Mixed TSH/FSH/LH-secreting tumors	8	1.7

	Tab	e 8.1	Recorded	cases c	of	different	ty	pes o	of T	SH-	secreti	ng	tumors
--	-----	-------	----------	---------	----	-----------	----	-------	------	-----	---------	----	--------

correlation with its or their hypersecretion in vivo [17, 21, 22]. In addition, the coexistence of TSH-secreting tumors with Hashimoto's thyroiditis and hypothyroidism has been reported [2, 23].

When diagnosed, most thyrotropinomas have a diameter size >1 cm presenting invasive features in the surrounding structures (i.e., the dura mater and bone) with extrasellar extension in the supra- and/or parasellar direction found in the majority of cases [1, 16, 17, 18, 24, 25]. Interestingly, patients with intact thyroid display a significantly lower occurrence of invasive tumors with respect to those who underwent previous thyroid ablation by surgery or radioiodine [1, 3]. In these cases, the reduction in circulating thyroid hormone levels due to thyroid ablation may be the cause of feedback mechanism alterations, thus supporting tumor growth. Thyrotropinomas with a diameter <1 cm were usually found in less than 15% of the cases [25], but their prevalence is progressively increasing thanks to improved testing of thyroid function and awareness among endocrinologists. Indeed, 30% of thyrotropinomas have been found with a diameter <1 cm in the series recently published [7, 26].

As for majority of pituitary lesions, the molecular mechanisms leading to the formation of thyrotropinoma are not fully understood. TSH-secreting tumors are usually characterized by a very fibrous consistency [27]. This observation has been linked to enhanced basic fibroblast growth factor (bFGF) blood levels and specific transcript in the tissues removed from two patients with invasive mixed PRL/TSHsecreting tumors and displaying by marked fibrosis [28]. This finding suggests a possible autocrine role for bFGF in tumor development. According to X-chromosomal inactivation analysis, thyrotropinomas derive from the clonal expansion of a single cell initially transformed cell as most pituitary tumors [29]. The hypothesis of a transforming event favoring gain of cell proliferation followed by secondary mutations or alterations leading to tumor progression is supposed to be applicable to TSH-omas. Mutations of proto-oncogenes (Rb, MEN1), either oncogenes (Ras, protein kinase C, G-protein subunits, TRH receptor) or pituitary-specific genes, able to confer growth advantage to pituitary cells, have been screened extensively. However, to date, no mutations in these candidate genes have been found. In particular, none of the thyrotropinomas screened presented activating mutations of genes encoding for G-protein subunits, such as αs , αq , $\alpha 11$, or $\alpha i2$ [30]. On the contrary, GH-secreting tumors frequently present mutations in the oncogene gsp. Similarly, no mutations in the gene encoding for TRH receptor have been found in 9 and 3 thyrotropinomas, respectively [30, 31]. Since the transcription factor Pit-1 is a key regulator of cell differentiation and in PRL, GH, and thyrotropin gene expression, Pit-1 gene has been screened for mutations in 14 thyrotropinomas but found to be wild type [1]. However, Pit-1 resulted to be overexpressed in thyrotropinomas, similarly to what observed in GH-omas, although the biological meaning of this finding remains to be clarified [18, 20, 32].

Another candidate gene investigated is located on 11q13 and named MEN1. The MEN1 gene, encoding for menin, is linked to the multiple endocrine neoplasia type 1 (MEN1). About 3–30% sporadic pituitary tumors show loss of heterozygosity (LOH) at 11q13, which has been associated with the transition from the noninvasive

to the invasive phenotype. LOH on 11q13 has been found in 3 out 13 thyrotropinomas tested, but no MEN1 mutations were found [33]. Interestingly, hyperthyroidism due to thyrotropinomas has been reported in five cases within a familial setting of MEN1 syndrome [12].

As far as the loss of tumor suppressor genes is concerned, no loss of p53 was found in one thyrotropinoma studied, and analysis on retinoblastoma gene (Rb) is still lacking in thyrotropinomas. Mutations in the aryl hydrocarbon receptor-interacting protein (AIP) are involved in sporadic pituitary tumors, to note that AIP mutations were found in two patients with TSH-omas [13, 34].

Interestingly, recent whole-exome sequencing analysis is allowed to identify several candidate somatic mutations and variation in copy numbers in 12 sporadic TSH-secreting tumors [35]. Further studies in combination with epigenetic and transcriptomic approaches are needed to reveal the potential of such genetic lesions.

Thyroid hormone receptors (TRs) have been proposed as a potential candidate oncogenes and alterations in the feedback control mechanisms, as factors involved in the thyrotropinomas pathogenesis have been evaluated. Somatic mutations of thyroid hormone receptor beta (TRb) and aberrant alternative splicing of TRb2 mRNA encoding a TRb variant lacking T3-binding activity have been associated with impaired negative feedback on TSH secretion in some thyrotropinomas [36]. An aberrant expression of a thyroid hormone receptor β isoform (TR β 4) may partly cause the inappropriate secretion of TSH in thyrotropinomas [37].

In vitro studies on primary cultures from TSH-secreting tumors showed that these tumors express a large number of functioning receptors [3], although TSH response to TRH is usually lacking in vivo. Similarly, almost all TSH-omas express somatostatin receptors (SSTR) at variable ratio. Higher SSTR expression levels have been found in mixed GH/TSH-secreting tumor [38–40]. The inhibitory response to somatostatin analogs appears to be correctly mediated by SSTR in these tumors with resulted decrease in TSH secretion by neoplastic thyrotrophs [41–44]. Specific polymorphisms and LOH at the somatostatin receptor type 5 (SSTR5) gene locus seem to be associated with an aggressive phenotype and pharmacological resistance to somatostatin analogs, possibly because of absent somatostatin-induced inhibition of thyrotropin secretion [45]. In addition, the presence of dopamine receptors on the tumor thyrotrophs was the rationale for therapeutic trials with dopaminergic agonists, such as bromocriptine [38, 46, 47], with several studies showing a wide heterogeneity of TSH responses to dopaminergic agents, either in primary cultures or in vivo [48-50]. To date, no mutations in dopamine type 2 receptor (DRD2) have been found in TSH-omas [30, 31] and the effects of these inhibitory agents should be re-evaluated in light of the demonstration of the possible occurrence of SSTR5 and DRD2 heterodimerization [51].

Finally, the transformation of a TSH-oma into a carcinoma is an extremely rare event [52–54]. However, very high circulating levels of free alpha-subunit may be predictable of a progression toward a malignant behavior. Moreover, spontaneous and strong decrease in both TSH and alpha-GSU serum concentrations might indicate that the tumor is becoming less differentiated and correlates with invasive and metastatic features. In this regard, it has to be mentioned that, in a mouse model of

TSH-secreting tumor, the activation of phosphatidylinositol 3-kinase pathway favored pituitary growth that may induce transformation of the benign tumor into a carcinoma [55].

8.4 Clinical Presentation

Patients with TSH-omas usually present signs and symptoms of hyperthyroidism (Fig. 8.1). Many patients had been mistakenly diagnosed as having primary hyperthyroidism (Graves' disease), and about one third had inappropriate thyroidectomy and/or radioiodine treatment prior to the correct diagnosis. However, a true coexistence of Graves' disease and TSH-oma has been reported in a few cases [56, 57] and several untreated patients with TSH-oma were described as clinically euthyroid [11, 58]. The prevalence of circulating antithyroid autoantibodies (antithyroglobulin: Tg-Ab, and antithyroid peroxidase: TPO-Ab) is similar to that found in the general population, but some patients develop Graves' disease after pituitary surgery and few others present bilateral exophthalmos due to autoimmune thyroiditis [3, 59, 60] or unilateral exophthalmos due to orbital invasion by the pituitary tumor. It is worth



Fig. 8.1 Clinical manifestations in patients with TSH-omas. Based on previous thyroid ablation, patients have been divided into two groups. Patients usually present signs and symptoms of hyper-thyroidism. As other pituitary adenomas, TSH-omas present clinical features caused by mass pressure effects. Invasive tumors are seen in about half of the patients with previous thyroidectomy and in 1/4 of untreated patients. TSH strong stimulation causes uni- or multi-nodular goiter. The prevalence of circulating antithyroid autoantibodies (Abs) is similar to that found in the general population (modified from Beck-Peccoz et al., Endotext, 2019)

noting that patients with mixed TSH/GH adenomas may have hyperthyroid features overshadowed by those of acromegaly [61–64]. This underlines the importance of TSH and FT4 systematic measurement in patients with pituitary tumors.

In about 72% of cases, the TSH strong stimulation causes uni- or multi-nodular goiter; however, the progression toward functional autonomy seems to be rare [3, 65].

Moreover, in sporadic cases cardiotoxicosis with atrial fibrillation and cardiac failure have been reported [66–70]. Two patients with typical episodes of periodic paralysis [71, 72] or a high prevalence of radiological vertebral fracture [73] have also been described.

Other symptoms can be tachycardia, tremors, heat intolerance, asthenia, and irritability.

As other pituitary adenomas, TSH-omas may present clinical features caused by mass pressure effects. The tendency to the invasiveness and the frequent over-sellar extension found in these adenomas increase the probability that the clinical symptomatology linked to the mass effect is manifest. Typical features that may occur are as follows:

- 1. Alterations in the visual field in about 50% of patients, due to the compression of the optic chiasm and the possible subsequent involvement of the optic nerve [74].
- 2. Ophthalmoplegia or diplopia due to compression of the cranial nerves at the level of cavernous sinus.
- Hydrocephalus due to compression of the third ventricle and occlusion of foramen of Monroe.
- 4. Clear rhinorrhea due to erosion of the sellar floor with infiltration in the sphenoid sinus.

Furthermore, in 20–25% of patients frontal headache, continuous and resistant to analgesics can appear, caused by the distension of the sellar diaphragm that can be more rarely associated with other symptoms and signs of intracranial hypertension such as vomiting and edema of the papilla caused by the expanding intracranial mass.

Additionally, the compressive effect of adenomatous cells on surrounding adenopituitary cells can result in hypopituitarism in 25% of patients. The different sensitivity of adeno-pituitary cells to the compression effect establishes the progressive order according to which the secretory deficit occurs: first the GH-secreting cells, then the gonadotropin-secreting cells, then the TSH-secreting ones, and finally the corticotropic ones. Instead, the compression of the hypothalamic–pituitary peduncle is a cause of both the pseudo-hyperprolactinemia that is the consequence of dopamine-mediated inhibition on prolactin secretion and the diabetes insipidus due to the interruption of the axonic flow that carries the neurohypophysial hormones. Besides, dysfunction of the gonadal axis with menstrual disorders has been described mainly in mixed TSH/PRL adenomas and central hypogonadism, delayed puberty, and decreased libido in a number of male patients with TSH-oma and/or TSH/FSH adenomas [1, 49, 75, 67]. Finally, a recent publication has evidenced an estimated incidence of 4.8% of differentiated thyroid cancer (DTC) in patients who underwent surgery for TSH-oma, suggesting a possible role of TSH hypersecretion in the development of thyroid tumors [76].

8.5 Diagnosis

Patients harboring TSH-oma present with signs and symptoms of hyperthyroidism that usually are milder than expected owing to the levels of circulating FT4 and FT3, probably due to the long duration of the disease. From a biochemical point of view, the confirmed presence of elevated serum FT4/FT3 and measurable TSH levels (Fig. 8.2) is sufficient to exclude Graves' disease or other causes of primary hyperthyroidism. It is worth mentioning that a measurable TSH associated with high FT4/FT3 levels during levothyroxine replacement therapy may be caused by poor compliance or to the administration of levo-T4 (L-T4) before blood sampling.

A correct diagnosis of thyrotropinoma allows to avoid dramatic consequences, such as improper thyroid ablation (thyroidectomy or radioiodine) that may cause the pituitary tumor volume further expand. Since clinical features of hyperthyroidism can be overshadowed by those of acromegaly in case of mixed thyrotropin/ GH-secreting tumors, systematic measurement of thyrotropin and FT4 is recommended in patients who have pituitary tumor.

Measurements of different parameters are proposed as quantifying the degree of tissue hyperthyroidism [2]. An unbalanced hypersecretion of circulating free α -GSU levels and elevated α -GSU/thyrotropin molar ratio (Table 8.1) is found in 80% of patients who have thyrotropinomas [3]. Accordingly to recently published data, such α -GSU hypersecretion is a phenomenon correlated with progressive tumor volume increase [16, 17, 20] since serum α -GSU levels are almost always normal in microadenomas.



Fig. 8.2 Baseline serum TSH levels in TSH-omas from our case series. The biochemical hallmarks of TSH-omas are high serum free T3 and T4 concentrations while, as shown in the figure, TSH may be inappropriately normal or high

8.5.1 Dynamic Tests

TRH test and T3 suppression are recommended for the diagnosis of thyrotropinoma, although none of them is of clear-cut diagnostic value. Thus, the combination of both stimulatory and inhibitory tests increases the specificity and sensitivity of the diagnostic workup [3, 17, 20].

Classically, the T3 suppression test has been used to evaluate the presence of thyrotropinomas. The failure of a complete inhibition of TSH secretion after T3 suppression test (80–100 mg per day per 8–10 days) is typical of patients with thyrotropinoma (see Table 8.1). T3 suppression test is the most sensitive and specific test, particularly in patients who have had previous thyroid ablation [2, 77], whereas is contraindicated in elderly patients and in those with coronary heart disease.

As far as stimulatory tests are concerned, TRH injection (200 µg i.v.) does not increase either TSH or α -GSU levels in up to 85% of patients with thyrotropinoma [2]. Thus, this test has been used widely in the workup of thyrotropinomas. The finding of a discrepant response to TRH between thyrotropin and α -GSU (or GH in the case of acromegaly), however, is pathognomonic of pituitary tumors cosecreting thyrotropin and other hormones (Fig. 8.2) [16]. Interestingly, the administration of native somatostatin or its analogs (i.e., octreotide or lanreotide) induces a decrease in circulating TSH levels in normal and in the majority of the tumoral thyrotrophs [35, 36, 38, 53, 56, 65, 72], and this inhibitory response may predict the efficacy of long-term treatment with somatostatin analogs [78].

8.5.2 Differential Diagnosis

Primary hyperthyroidism in a hyperthyroid patient is ruled out by the presence of detectable TSH levels. However, in patients on L-T4 replacement for primary hypothyroidism, poor compliance is by far the most common cause of apparent inappropriate secretion of TSH (TSH still too high for the levels of the thyroid hormones). This underscores the importance of studying patients in steady state [79]. Inadequate measurement of TSH and peripheral thyroid hormones should always be taken into consideration. This may occur, for example, in medical therapy with amiodarone [79].

It is crucial to exclude the presence of methodological interferences due to the presence of circulating autoantibodies (e.g., against T3 and T4) or heterophilic antibodies (e.g., for TSH) that may give falsely elevated serum levels of thyrotropin or free thyroid hormones (i.e., rule out the presence of primary hyperthyroidism and the various forms of euthyroid hyperthyroxinemia) [2].

A thyrotropinoma must be suspected in the presence of neurologic signs and symptoms such as visual defects (25% of patients), headache (20%), and hypopituitarism (50%), all these being possible expressions of a tumor expansion [3, 17, 20]. Moreover, alteration of hypothalamic–pituitary–gonadal axis is frequent in case of pituitary tumors, with menstrual disorders present in all patients who have mixed thyrotropin/PRL tumors and in one third of those who have pure thyrotropinomas. Delayed puberty, central hypogonadism, and decreased libido are also found in men with thyrotropinomas or mixed thyrotropin/FSH tumors.

Once the diagnosis of central hyperthyroidism is confirmed, additional diagnostic steps have to be performed to differentiate thyrotropinoma from RTH, in particular PRTH [2, 3, 7, 16, 17, 78, 80] (Table 8.2). Liver (sex hormone-binding globulin [SHBG]) and bone (carboxyterminal cross-linked telopeptide of type I collagen [ICTP]) parameters are successfully used to differentiate hyperthyroid patients who have thyrotropinoma from those who have PRTH. Higher serum SHBG and ICTP levels are common in patients with thyrotropinoma, whereas they are in the normal range in patients who have RTH. No statistically significant differences in terms of sex, age, TSH levels, or free thyroid hormone concentrations have been observed between patients with thyrotropinoma and those with RTH [2]. As far as dynamic tests are concerned, typically TSH-omas do not respond to TRH stimulation and/or to T3 suppression tests. Moreover, elevated α -GSU concentrations or high α -GSU/ thyrotropin molar ratio and thyrotropin unresponsiveness to TRH stimulation or to T3 suppression tests, or both, favor the presence of a thyrotropinoma.

Another parameter that can be useful for the differential diagnosis is the evaluation of the sensitivity to long-acting somatostatin analogs [81]. More than 90% of

Parameter	TSH-omas	PRTH	Р
F/M	1.3	1.4	NS
Familial cases (%)	0	85	<0.0001
Thyrotropin mU/L	2.7 ± 0.6	2.1 ± 0.3	NS
FT4 pmol/L	40.0 ± 4.2	30.5 ± 2.6	NS
FT3 pmol/L	14.5 ± 1.4	12.7 ± 1.2	NS
SHBG nmol/L ^a	113 ± 17	60 ± 5	<0.0001
Presence of lesion at CT scan or MRI (%)	98	5	<0.0001
High α -GSU levels (%)	65	3	<0.0001
High α -GSU/thyrotropin m.r. (%)	81	2	<0.0001
Abnormal thyrotropin response to T3 suppression test (%) ^b	100	100	NS
Blunted thyrotropin response to TRH test (%)	94	4	<0.0001

Table 8.2 Differential diagnosis between TSH-oma and PRTH

TSH-oma are sensitive, and two or more administrations of analog are usually sufficient to induce significant decreases or normalization of circulating free thyroid hormone. These modifications have never been observed in PRTH patients (Table 8.2). Finally, TR β gene analysis may be useful in the differential diagnosis, as genomic TR β mutations have been detected in patients with RTH only [7].

8.5.3 Imaging Studies and Localization of the Tumor

Nuclear MRI currently represents the imaging strategy of choice for the visualization of a thyrotropinoma. High-resolution CT is the alternative approach in case MRI is contraindicated (e.g., in the presence of a pacemaker). Although the diagnosis of TSH-oma is strongly supported by the presence of a pituitary lesion at neuroradiological imaging, a pituitary lesion has been identified at MRI in about 20% of RTH, indicating that a pituitary incidentaloma and RTH may coexist [82]. The differential diagnosis with PRTH may be problematic when the pituitary adenoma is small in size or in the case of confusing lesions, such as empty sella, pituitary incidentalomas, or ectopic tumors [81]. Pituitary scintigraphy with radiolabeled octreotide (Octreoscan) has been used to detect thyrotropinomas expressing somatostatin receptors [83]. Although the specificity of this procedure is low, it has been useful in the recognition of nasopharyngeal mass in few patients with clinical and biochemical features of central hyperthyroidism [84, 85].

8.6 Management

As recommended by the guidelines published by the European Thyroid Association [80], the first-line therapy for TSH-omas is surgical resection by transsphenoidal or subfrontal adenomectomy, the choice of the route depending on the tumor volume, and its suprasellar extension and invasiveness. This procedure aims to removing neoplastic tissue and normalizing normal pituitary/thyroid function. The operation may be difficult as the tumor may present a marked fibrosis, possibly related to high expression of basic fibroblast growth factor [28], and local invasion involving the cavernous sinus, internal carotid artery, or optic chiasm. Particular attention has to be paid to presurgical preparation of the patient, particularly in the preanesthetic period [86]: Antithyroid drugs along with propranolol should be used aiming at restoration of euthyroidism. Presurgical treatment with somatostatin analogs (octreotide LAR, lanreotide autogel) might be effective in reducing TSH-oma size and normalizing circulating thyroid hormones levels [87]. It should be noted that this approach may cause TSH secretion from normal thyrotropes to be re-activated, leading to the loss of a useful parameter to evaluate the complete removal of the adenoma, which is undetectable TSH levels few days after successful surgery. Neurosurgical intervention may cause a partial or complete hypopituitarism. However, a case of thyroid storm after pituitary surgery was reported [88]. In case of failure of pituitary surgery and in the presence of life-threatening hyperthyroidism, total thyroidectomy or thyroid ablation with radioiodine is indicated [89]. According to the largest published series, pituitary surgery is effective in restoring euthyroidism in 75–83% of patients with TSH-omas [61, 90].

When the patients decline surgery or in case of surgical failure, radiotherapy and/ or medical treatment with somatostatin analogs should be considered [80].

In case of radiotherapy, the recommended dose is no less than 45 Gy fractionated at 2 Gy per day or 10–25 Gy in a single dose if a stereotactic gamma unit is available [80, 91]. This procedure manages in normalizing thyroid function in 37% of patients within 2–4 years [61].

Some patients require medical therapy in order to control hyperthyroidism, although earlier diagnosis has improved the surgical cure rate of TSH-omas. The medical treatment of TSH-omas is based on long-acting somatostatin analogs, such as octreotide or lanreotide [27, 41, 42, 80, 92–94]. Treatment with these analogs leads to a reduction in TSH and alpha-GSU secretion in almost all cases, with restoration of the euthyroid state in about 95% of patients. Somatostatin analogs are safe even though side effects, such as cholelithiasis and carbohydrate intolerance, may appear. They are safe even during pregnancy [26]. Octreotide treatment in pregnant women was reported to be effective in restoring euthyroidism in the mother and had no side effects on development and thyroid function of the fetuses [7, 26, 48]. Many papers suggest the use of somatostatin analogs as first-line therapy for patients with TSH-omas, particularly for invasive macroadenomas [95–98]. During somatostatin analog therapy, tumor shrinkage occurs in about 50% of patients and vision improvement is seen in 75% [61, 90, 99]. Very rapid shrinkage of the tumor has been described [100]. Dose should be tailored for each patient, depending on therapeutic response. Tolerance is usually very good, as gastrointestinal side effects are transient with long-acting analogs [38, 41, 42, 99, 101].

Resistance to somatostatin analogs treatment has been documented in a minority of cases. The presence of dopamine receptors in TSH-omas was the rationale for the use of dopaminergic agonists, such as bromocriptine and cabergoline. A heterogeneity of TSH responses to these drugs has been described with the best effects achieved in mixed PRL/TSH tumors [69, 102, 103].

8.7 Follow-Up

Few data on TSH-oma recurrence in patients considered cured after surgery or radiotherapy have been reported so far. However, recurrence of the adenoma seems to be an infrequent event, at least in the first years after successful surgery [64, 104]. In general, postoperatively, the patient should be evaluated clinically and biochemically 2 or 3 times during the first year and then once a year. Pituitary imaging should be performed every 2 or 3 years, but should be promptly done whenever an increase in TSH and thyroid hormone levels, or clinical symptoms occur. In the case of a persistent macroadenoma, close visual field follow-up is required, since visual function could be threatened. Emergency surgical decompression is not always able to reverse even a recent visual deficit.

8.8 Prognosis

The criteria of cure of patients operated or irradiated for TSH-omas have not been clearly defined, due to the rarity of the disease and the great heterogeneity of the methods used. Some of these criteria are inapplicable if patients underwent previous thyroid ablation (Table 8.3).

A positive prognostic event is the absence of neurological signs and symptoms, but lacks both sensitivity and specificity, as even an incomplete debulking of the tumor may cause visual field defects and headache disappearance. It is logical that cured patients have clinical and biochemical reversal of thyroid hyperfunction after withdrawal from antithyroid medications. Nevertheless, the presence of normal free thyroid hormone concentrations or normalization of parameters peripheral thyroid hormone action (SHBG, ICTP, etc.) does not attest the complete removal or destruction of tumoral cells, since transient clinical remission accompanied by normalization of thyroid function is possible [32, 64, 104–106]. The criteria of normalization of circulating TSH are not applicable to previously thyroidectomized patients and to the 26% of patients with normal basal values of TSH. In our practice, undetectable TSH levels 1 week after surgery indicate complete adenomectomy, provided that the patient was hyperthyroid and presurgical treatments were stopped before surgery [64]. Normalization of alpha-GSU and/or the alpha-GSU/TSH molar ratio is in general a good index for the evaluation of therapy efficacy [16, 64]. However, both parameters are characterized by less than optimal sensitivity, as they are normal in about 25% of patients with TSH-oma. The most sensitive and specific test to document the complete removal of the adenoma, in the absence of contraindication, is

Criteria	Comments
Remission from hyperthyroid manifestations	Clinical improvement may be transient
(clinical and biochemical)	No predictive value
Undetectable TSH 1 week after neurosurgery	Applicable to hyperthyroid patients that
	stopped treatments at least 10 days before
	surgery
	Good prognostic value
Normalization of circulating TSH levels	Not applicable to patients with normal
	TSH
	Poor predictive value
Normalization of free thyroid hormone levels	Biochemical remission may be transient
	Poor predictive value
Positive T3 suppression test with undetectable	Not applicable to elderly patients or in
TSH and no response to TRH (or central	those with cardiac diseases
hypothyroidism)	Optimal sensitivity/specificity and
	predictive value
Normalization of alpha-GSU levels and alpha-	Not applicable to patients with normal
GSU/TSH molar ratio	values before neurosurgery
	Lack of sensitivity
Disappearance of neurological manifestations	May be transient
(adenoma imaging, visual field defects, headache)	Poor predictive value

Table 8.3 Criteria for the evaluation of treatment outcome

the T3 suppression test [64]: Only patients in whom T3 administration completely inhibits basal and TRH-stimulated TSH secretion can be defined as cured.

References

- Beck-Peccoz P, Brucker-Davis F, Persani L, Smallridge RC, Weintraub BD. Thyrotropinsecreting pituitary tumors. Endocr Rev. 1996;17:610–38.
- Beck-Peccoz P, Persani L, Lania A. Thyrotropin-secreting pituitary adenomas. In: Feingold KR, Anawalt B, Boyce A, Chrousos G, Dungan K, Grossman A, Hershman JM, Kaltsas G, Koch C, Kopp P, Korbonits M, McLachlan R, Morley JE, New M, Perreault L, Purnell J, Rebar R, Singer F, Trence DL, Vinik A, Wilson DP, editors. Endotext. South Dartmouth, MA: MDText.com Inc; 2000.
- Beck-Peccoz P, Lania A, Persani L. TSH-producing adenomas. In: Jameson LJ, DeGroot LJ, editors. Endocrinology, adult and pediatric, vol. I. 7th ed. Philadelphia, PA: Sauderns Elsevier; 2015. p. 266–74.
- Jailer JW, Holub DA. Remission of Graves'' disease following radiotherapy of a pituitary neoplasm. Am J Med. 1960;28:497–500.
- Hamilton C, Adams LC, Maloof F. Hyperthyroidism due to thyrotropin-producing pituitary chromophobe adenoma. N Engl J Med. 1970;283:1077–80.
- Refetoff S, Dumitrescu AM. Impaired sensitivity to thyroid hormone: defects of transport, metabolism and action. 2010. https://www.thyroidmanager.org/chapter/ thyroid-hormone-resistance-syndromes/.
- Gurnell M, Visser TJ, Beck-Peccoz P, Chatterjee VKK. In: Jameson LJ, LJ DG, editors. Resistance to thyroid hormone: in endocrinology, adult and pediatric, vol. II. 7th ed. Philadelphia, PA: Sauderns Elsevier; 2015. p. 1648–65.
- Hannoush ZC, Weiss RE. Defects of thyroid hormone synthesis and action. Endocrinol Metab Clin N Am. 2017;46:375–88.
- Onnestam L, Berinder K, Burman P, Dahlqvist P, Engstrom BE, Wahlberg J, et al. National incidence and prevalence of TSH-secreting pituitary adenomas in Sweden. J Clin Endocrinol Metab. 2013;98(2):626–35.
- Nakayama Y, Jinguji S, Kumakura S, Nagasaki K, Natsumeda M, Yoneoka Y, et al. Thyroidstimulating hormone (thyrotropin)-secretion pituitary adenoma in an 8 year-old boy: case report. Pituitary. 2012;15:110–5.
- 11. Rabbiosi S, Peroni E, Tronconi GM, Chiumello G, Losa M, Weber G. Asymptomatic thyrotropin-secreting pituitary macroadenoma in a 13-year-old-girl: successful first-lin3e treatment with somatostatin analogs. Thyroid. 2012;22:1076–9.
- 12. Taylor TJ, Donlon SS, Bale AE, Smallridge RC, Francis TB, Christensen RS, et al. Treatment of a thyrotropinoma with octreotide-LAR in a patient with multiple endocrine neoplasia-1. Thyroid. 2000;10:1001–7.
- Daly AF, Tichomirowa MA, Petrossians P, Heliövaara E, Jaffrain-Rea ML, Barlier A, et al. Clinical characteristics and therapeutic responses in patients with germ-line AIP mutations and pituitary adenomas: an international collaborative study. J Clin Endocrinol Metab. 2010;95:E373–83.
- Polanco Santos C, Sandouk Z, Yogi-Morren D, Prayson R, Recinos P, Kennedy L, Hamrahian AH, Pantalone KM. TSH-staining pituitary adenomas: rare, silent, and plurihormonal. Endocr Pract. 2018;24(6):580–8.
- Azzalin A, Appin CL, Schniederjan MJ, Constantin T, Ritchie JC, Veledar E, Oyesiku NM, Ioachimescu AG. Comprehensive evaluation of thyrotropinomas: single-center 20-year experience. Pituitary. 2016;19:183–93.
- 16. Brucker-Davis F, Oldfield EH, Skarulis MC, Doppman JL, Weintraub BD. Thyrotropinsecreting pituitary tumors: diagnostic criteria, thyroid hormone sensitivity, and treatment

outcome in 25 patients followed at the National Institutes of Health. J Clin Endocrinol Metab. 1999;84:476–86.

- Bertholon-Grégoire M, Trouillas J, Guigard MP, Loras B, Tourniaire J. Mono- and plurihormonal thyrotropic pituitary adenomas: pathological, hormonal and clinical studies in 12 patients. Eur J Endocrinol. 1999;140:519–27.
- Sanno N, Teramoto A, Osamura RY. Thyrotropin-secreting pituitary adenomas. Clinical and biological heterogeneity and current treatment. J Neuro-Oncol. 2001;54:179–86.
- Terzolo M, Orlandi F, Bassetti M, Medri G, Paccotti P, Cortelazzi D, et al. Hyperthyroidism due to a pituitary adenoma composed of two different cell types, one secreting alpha-subunit alone and another cosecreting alpha-subunit and thyrotropin. J Clin Endocrinol Metab. 1991;72:415–21.
- Pereira BD, Raimundo L, Mete O, Oliveira A, Portugal J, Asa SL. Monomorphous plurihormonal pituitary adenoma of Pit-1 lineage in a giant adolescent with central hyperthyroidism. Endocr Pathol. 2016;27:25–33.
- Banerjee AK, Sharma BS, Kak VK. Clinically and biochemically silent thyrotroph adenoma with oncocytic change. Neurol India. 2000;48:374–7.
- Pawlikowski M, Pisarek H, Jaranowska M, Radek M, Winczyk K, Kunert-Radek J. "Silent" thyrotropin (TSH) expression in acromegaly and clinically non-functioning pituitary adenomas. Endokrynol Pol. 2016;67:515–8.
- Li J, Li J, Jiang S, Yu R, Yu Y. Case report of a pituitary thyrotropin-secreting macroadenoma with Hashimoto thyroiditis and infertility. Medicine (Baltimore). 2018;97:e9546.
- Kon YC, Loh KC, Tambyah JA, Lim LH, Marshall JC. Thyrotropin (TSH)-secreting pituitary macroadenoma with cavernous sinus invasion. Singap Med J. 2001;42:433–9.
- Socin HV, Chanson P, Delemer B, Tabarin A, Rohmer V, Mockel J, et al. The changing spectrum of TSH-secreting pituitary adenomas: diagnosis and management in 43 patients. Eur J Endocrinol. 2003;148:433–42.
- Blackhurst G, Strachan MW, Collie D, Gregor A, Statham PF, Seckl JE. The treatment of a thyrotropin-secreting pituitary macroadenoma with octreotide in twin pregnancy. Clin Endocrinol. 2002;57:401–4.
- Webster J, Peters JR, John R, Smith J, Chan V, Hall R, et al. Pituitary stone: two cases of densely calcified thyrotropin-secreting pituitary adenomas. Clin Endocrinol. 1994;40:137–43.
- Ezzat S, Horvath E, Kovacs K, Smyth HS, Singer W, Asa SL. Basic fibroblast growth factor expression by two prolactin and thyrotropin-producing pituitary adenomas. Endocr Pathol. 1995;6:125–34.
- Chien WM, Garrison K, Caufield E, Orthel J, Dill J, Fero ML. Differential gene expression of p27Kip1 and Rb knockout pituitary tumors associated with altered growth and angiogenesis. Cell Cycle. 2007;6:750–7.
- 30. Dong Q, Brucker-Davis F, Weintraub BD, Smallridge RC, Carr FE, Battey J, et al. Screening of candidate oncogenes in human thyrotroph tumors: absence of activating mutations of the Gαq, Gα11, Gαs, or thyrotropin-releasing hormone receptor genes. J Clin Endocrinol Metab. 1996;81:1134–40.
- Friedman E, Adams EF, Höög A, Gejman PV, Carson E, Larsson C, et al. Normal structural dopamine type 2 receptor gene in prolactin-secreting and other pituitary tumors. J Clin Endocrinol Metab. 1994;78:568–74.
- Beck-Peccoz P, Persani L, Calebiro D, Bonomi M, Mannavola D, Campi I. Syndromes of hormone resistance in the hypothalamic-pituitary-thyroid axis. Best Pract Res Clin Endocrinol Metab. 2006;20(4):529–46.
- Asteria C, Anagni M, Persani L, Beck-Peccoz P. Loss of heterozygosity of the MEN1 gene in a large series of TSH-secreting pituitary adenomas. J Endocrinol Investig. 2001;24:796–801.
- 34. Barlier A, Vanbellinghen JF, Daly AF, Silvy M, Jaffrain-Rea ML, Trouillas J, et al. Mutations in the aryl hydrocarbon receptor interacting protein gene are not highly prevalent among subjects with sporadic pituitary adenomas. J Clin Endocrinol Metab. 2007;92:1952–5.
- Sapkota S, Horiguchi K, Tosaka M, Yamada S, Yamada M. Whole-exome sequencing study of thyrotropin-secreting pituitary adenomas. J Clin Endocrinol Metab. 2017;102:566–75.

- Ando S, Sarlis NJ, Oldfield EH, Yen PM. Somatic mutation of TRbeta can cause a defect in negative regulation of TSH in a TSH-secreting pituitary tumor. J Clin Endocrinol Metab. 2001;86:5572–6.
- 37. Tagami T, Usui T, Shimatsu A, Beniko M, Yamamoto H, Moriyama K, et al. Aberrant expression of thyroid hormone receptor beta isoform may cause inappropriate secretion of TSH in a TSH-secreting pituitary adenoma. J Clin Endocrinol Metab. 2011;96:E948–52.
- Gatto F, Barbieri F, Gatti M, Wurth R, Schulz S, Ravetti JL, et al. Balance between somatostatin and D2 receptor expression drives TSH-secreting adenoma response to somatostatin analogues and dopastatins. Clin Endocrinol. 2012;76:407–14.
- Gancel A, Vuillermet P, Legrand A, Catus F, Thomas F, Kuhn JM. Effects of a slow-release formulation of the new somatostatin analogue lanreotide in TSH-secreting pituitary adenomas. Clin Endocrinol. 1994;40:421–8.
- 40. Bertherat J, Brue T, Enjalbert A, Gunz G, Rasolonjanahary R, Warnet A, et al. Somatostatin receptors on thyrotropin-secreting pituitary adenomas: comparison with the inhibitory effects of octreotide upon in vivo and in vitro hormonal secretions. J Clin Endocrinol Metab. 1992;75:540–6.
- 41. Kuhn JM, Arlot S, Lefebvre H, Caron P, Cortet-Rudelli C, Archambaud F, et al. Evaluation of the treatment of thyrotropin-secreting pituitary adenomas with a slow release formulation of the somatostatin analog lanreotide. J Clin Endocrinol Metab. 2000;85:1487–149.
- 42. Caron P, Arlot S, Bauters C, Chanson P, Kuhn JM, Pugeat M, et al. Efficacy of the long-acting octreotide formulation (octreotide-LAR) in patients with thyrotropin-secreting pituitary adenomas. J Clin Endocrinol Metab. 2001;86:2849–53.
- 43. Yoshihara A, Isozaki O, Hizuka N, Nozoe Y, Harada C, Ono M, et al. Expression of type 5 somatostatin receptor in TSH-secreting pituitary adenomas: a possible marker for predicting long-term response to octreotide therapy. Endocr J. 2007;54:133–8.
- 44. Gatto F, Barbieri F, Castelletti L, Arvigo M, Pattarozzi A, Annunziata F, et al. In vivo and in vitro response to octreotide LAR in a TSH-secreting adenoma: characterization of somatostatin receptor expression and role of subtype 5. Pituitary. 2011;14:141–7.
- 45. Filopanti M, Ballarè E, Lania AG, Bondioni S, Verga U, Locatelli M, et al. Loss of heterozygosity at the SS receptor type 5 locus in human GH- and TSH-secreting pituitary adenomas. J Endocrinol Investig. 2004;27:937–42.
- Wood DF, Johnston JM, Johnston DG. Dopamine, the dopamine D2 receptor and pituitary tumours. Clin Endocrinol. 1991;35:455–66.
- 47. Verhoeff NP, Bemelman FJ, Wiersinga WM, van Royen EA. Imaging of dopamine D2 and somatostatin receptors in vivo using single-photon emission tomography in a patient with a TSH/PRL-producing pituitary macroadenoma. Eur J Nucl Med. 1993;20:555–60.
- Foppiani L, Del Monte P, Ruelle A, Bandelloni R, Quilici P, Bernasconi D. TSH-secreting adenomas: rare pituitary tumors with multifaceted clinical and biological features. J Endocrinol Investig. 2007;30:603–9.
- Gesundheit N, Petrick P, Nissim M, Dahlberg PA, Doppman L, Emerson CH, et al. Thyrotropin-secreting pituitary adenomas: clinical and biochemical heterogeneity. Ann Intern Med. 1989;111:827–35.
- 50. Spada A, Bassetti M, Martino E, Giannattasio G, Beck-Peccoz P, Sartorio P, et al. In vitro studies on TSH secretion and adenylate cyclase activity in a human TSH-secreting pituitary adenoma. Effects of somatostatin and dopamine. J Endocrinol Investig. 1985;8:193–8.
- Rocheville M, Lange DC, Kumar U, Patel SC, Patel RC, Patel YC. Receptors for dopamine and somatostatin: formation of hetero-oligomers with enhanced functional activity. Science. 2000;288:154–7.
- 52. Brown RL, Muzzafar T, Wollman R, Weiss RE. A pituitary carcinoma secreting TSH and prolactin: a non-secreting adenoma gone awry. Eur J Endocrinol. 2006;154(5):639–43.
- Lee W, Cheung AS, Freilich R. TSH-secreting pituitary carcinoma with intrathecal drop metastases. Clin Endocrinol. 2012;76:604–6.
- Mixson AJ, Friedman TC, David AK, Feuerstein IM, Taubenberger JK, Colandrea JM, et al. Thyrotropin-secreting pituitary carcinoma. J Clin Endocrinol Metab. 1993;76:529–33.

- Lu C, Willingham MC, Furuya F, Cheng SY. Activation of phosphatidylinositol 3-kinase signaling promotes aberrant pituitary growth in a mouse model of thyroid-stimulating hormonesecreting pituitary tumors. Endocrinology. 2008;149:3339–45.
- Lee MT, Wang CY. Concomitant graves' hyperthyroidism with thyrotrophin-secreting pituitary adenoma. South Med J. 2010;103:347–9.
- Kamoun M, d'Herbomez M, Lemaire C, Fayard A, Desailloud R, Huglo D, et al. Coexistence of thyroid-stimulating hormone-secreting pituitary adenoma and Graves' hyperthyroidism. Eur Thyroid J. 2014;3:60–4.
- Lim EM, Bhagat CI, Walsh J. Asymptomatic thyrotropin-secreting pituitary microadenoma. Intern Med J. 2001;31:428–9.
- Nguyen HD, Galitz MS, Mai VQ, Clyde PW, Glister BC, Shakir MK. Management of coexisting thyrotropin/growth-hormone-secreting pituitary adenoma and papillary thyroid carcinoma: a therapeutic challenge. Thyroid. 2010;20:99–103.
- 60. Azukizawa M, Morimoto S, Miyai K, Miki T, Yabu Y, Amino N, et al. TSH-producing pituitary adenoma associated with Graves' disease. In: Stockigt JR, Nagataki S, editors. Thyroid research, vol. VIII. Canberra: Australian Academy of Sciences; 1980. p. 645–8.
- Malchiodi E, Profka E, Ferrante E, Sala E, Verrua E, Campi I, et al. Thyrotropin-secreting pituitary adenomas: outcome of pituitary surgery and irradiation. J Clin Endocrinol Metab. 2014;99:2069–76.
- 62. Beck-Peccoz P, Piscitelli G, Amr S, Ballabio M, Bassetti M, Giannattasio G, et al. Endocrine, biochemical, and morphological studies of a pituitary adenoma secreting growth hormone, thyrotropin (TSH), and alpha-subunit: evidence for secretion of TSH with increased bioactivity. J Clin Endocrinol Metab. 1986;62:704–11.
- 63. Johnston PC, Hamrahian AH, Prayson RA, Kennedy L, Weil RJ. Thyrotoxicosis with absence of clinical features of acromegaly in a TSH- and GH-secreting, invasive pituitary macroadenoma. Endocrinol Diabetes Metab Case Rep. 2015;2015:140070.
- 64. Losa M, Giovanelli M, Persani L, Mortini P, Faglia G, Beck-Peccoz P. Criteria of cure and follow-up of central hyperthyroidism due to thyrotropin-secreting pituitary adenomas. J Clin Endocrinol Metab. 1996;81:3086–90.
- 65. Abs R, Stevenaert A, Beckers A. Autonomously functioning thyroid nodules in a patient with a thyrotropin-secreting pituitary adenoma: possible cause-effect relationship. Eur J Endocrinol. 1994;131:355–8.
- 66. George JT, Thow JC, Matthews B, Pye MP, Jayagopal V. Atrial fibrillation associated with a thyroid stimulating hormone-secreting adenoma of the pituitary gland leading to a presentation of acute cardiac decompensation: a case report. J Med Case Rep. 2008;2:67.
- Lee JH, Park M, Park MJ, Jo YS. Massive pleural and pericardial effusion due to hypothyroidism in a patient with a surgically treated thyroid-stimulating hormone-producing pituitary adenoma. Acta Clin Belg. 2018;73:398–401.
- Fujio S, Ashari HM, Yamahata H, Moinuddin FM, Bohara M, et al. Thyroid storm induced by TSH-secreting pituitary adenoma: a case report. Endocr J. 2014;61:1131–6.
- Kao YH, Chang TJ, Huang TS. Thyrotropin-secreting pituitary tumor presenting with congestive heart failure and good response to dopaminergic agonist cabergoline. J Formos Med Assoc. 2013;112:721–4.
- Mousiolis AC, Rapti E, Grammatiki M, Yavropoulou M, Efstathiou M, Foroglou N, et al. Somatostatin analogue treatment of a TSH-secreting adenoma presenting with accelerated bone metabolism and a pericardial effusion: a case report. Medicine (Baltimore). 2016;95:e2358.
- Hsu FS, Tsai WS, Chau T, Chen HH, Chen YC, Lin SH. Thyrotropin-secreting pituitary adenoma presenting as hypokalemic periodic paralysis. Am J Med Sci. 2003;325:48–50.
- Pappa T, Papanastasiou L, Markou A, Androulakis I, Kontogeorgos G, Seretis A, et al. Thyrotoxic periodic paralysis as the first manifestation of a thyrotropin-secreting pituitary adenoma. Hormones (Athens). 2010;9:82–6.
- Frara S, Losa M, Doga M, Formenti AM, Mortini P, Mazziotti G, et al. High prevalence of radiological vertebral fractures in patients with TSH-secreting pituitary adenoma. J Endocr Soc. 2018;2:1089–99.

- Beck-Peccoz P, Giavoli C, Lania A. A 2019 update on TSH-secreting pituitary adenomas. J Endocrinol Investig. 2019;
- 75. Sy RA, Bernstein R, Chynn KI, Kourides IA. Reduction in size of a thyrotropin- and gonadotropin-secreting pituitary adenoma treated with octreotide acetate (somatostatin analogue). J Clin Endocrinol Metab. 1992;74:690–4.
- 76. Perticone F, Pigliaru F, Mariotti S, Deiana L, Furlani L, Mortini P, et al. Is the incidence of differentiated thyroid cancer increased in patients with thyrotropin-secreting adenomas? Report of three cases from a large consecutive series. Thyroid. 2015;25:417–24.
- Tjörnstrand A, Nyström HF. Diagnostic approach to TSH-producing pituitary adenoma. Eur J Endocrinol. 2017;177:R183–97.
- Mannavola D, Persani L, Vannucchi G, Zanardelli M, Fugazzola L, Verga U, et al. Different responses to chronic somatostatin analogues in patients with central hyperthyroidism. Clin Endocrinol. 2005;62:176–81.
- Lim CT, Korbonits M. Update on the clinicopathology of pituitary adenomas. Endocr Pract. 2018;24(5):473–88.
- Beck-Peccoz P, Lania A, Beckers A, Chatterjee K, Wemeau JL. European thyroid association guidelines for the diagnosis and treatment of thyrotropin-secreting pituitary tumors. Eur Thyroid J. 2013;2:76–82.
- Okuma H, Hashimoto K, Ohashi T, Mihara M, Minami I, Izumiyama H, et al. A case of TSH-secreting pituitary adenoma with cyclic fluctuations in serum TSH levels. Endocr J. 2018;65:737–46.
- 82. Sriphrapradang C, Srichomkwun P, Refetoff S, Mamanasiri S. A novel thyroid hormone receptor beta gene mutation (G251V) in a Thai patient with resistance to thyroid hormone coexisting with pituitary incidentaloma. Thyroid. 2016;26:1804–6.
- Koulouri O, Moran C, Halsall D, Chatterjee K, Gurnell M. Pitfalls in the measurement and interpretation of thyroid function tests. Best Pract Res Clin Endocrinol Metab. 2013;27:745–62.
- Schoenmakers N, Moran C, Campi I, Agostini M, Bacon O, Rajanayagam O, et al. A novel albumin gene mutation (R222I) in familial dysalbuminemic hyperthyroxinemia. J Clin Endocrinol Metab. 2014;99:E1381–6.
- Favresse J, Burlacu MC, Maiter D, Gruson D. Interferences with thyroid function immunoassays: clinical implications and detection algorithm. Endocr Rev. 2018;4339:830–50.
- Dyer MW, Gnagey A, Jones BT, Pula RD, Lanier WL, Atkinson JLD, et al. Perianesthetic management of patients with thyroid-stimulating hormone-secreting pituitary adenomas. J Neurosurg Anesthesiol. 2017 Jul;29(3):341–6.
- Fukuhara N, Horiguchi K, Nishioka H, Suzuki H, Takeshita A, et al. Short-term preoperative octreotide treatment for TSH-secreting pituitary adenoma. Endocr J. 2015;62:21–7.
- Page KA, Roehmholdt BF, Jablonski M, Mayerson AB. Development of thyroid storm after surgical resection of a thyrotropin-secreting pituitary adenoma. Endocr Pract. 2008;14:732–7.
- Daousi C, Foy PM, MacFarlane IA. Ablative thyroid treatment for thyrotoxicosis due to thyrotropin-producing pituitary tumours. J Neurol Neurosurg Psychiatry. 2007;78:93–5.
- 90. Yamada S, Fukuhara N, Horiguchi K, Yamaguchi-Okada M, Nishioka H, Takeshita A, et al. Clinicopathological characteristics and therapeutic outcomes in thyrotropin-secreting pituitary adenomas: a single-center study of 90 cases. J Neurosurg. 2014;121(6):1462–73.
- 91. Mouslech Z, Somali M, Sakali AK, Savopoulos C, Mastorakos G, Hatzitolios AI. TSHsecreting pituitary adenomas treated by gamma knife radiosurgery: our case experience and a review of the literature. Hormones (Athens). 2016;15(1):122–8.
- Prieto-Tenreiro A, Díaz-Guardiola P. Long term treatment of a thyrotropin-secreting microadenoma with somatostatin analogues. Arq Bras Endocrinol Metabol. 2010;54:502–6.
- Gruszka A, Zielinski GM, Kunert-Radek J. Preoperative long-acting octreotide treatment for invasive thyrotropin-secreting pituitary macroadenoma after previous radioiodine thyroid ablation. J Clin Neurosci. 2014;21:340–2.

- 94. Zhang CF, Liang D, Zhong LY. Efficacy of the long-acting octreotide formulation in patients with thyroid-stimulating hormone-secreting pituitary adenomas after incomplete surgery and octreotide treatment failure. Chin Med J. 2012;125:2758–63.
- Fliers E, van Furth WR, Bisschop PH. Cure of a thyrotrophin (TSH)-secreting pituitary adenoma by medical therapy. Clin Endocrinol. 2012;77:788–90.
- Neggers SJ, van der Lely AJ. Medical approach to pituitary tumors. Handb Clin Neurol. 2014;124:303–16.
- 97. Gatto F, Grasso LF, Nazzari E, Cuny T, Anania P, Di Somma C, et al. Clinical outcome and evidence of high rate post-surgical anterior hypopituitarism in a cohort of TSH-secreting adenoma patients: might somatostatin analogs have a role as first-line therapy? Pituitary. 2015;18:583–91.
- Rimareix F, Grunenwald S, Vezzosi D, Rivière LD, Bennet A, Caron P. Primary medical treatment of thyrotropin-secreting pituitary adenomas by first-generation somatostatin analogs: a case study of seven patients. Thyroid. 2015;25:877–82.
- Beck-Peccoz P, Persani L. Medical management of thyrotropin-secreting pituitary adenomas. Pituitary. 2002;5:83–8.
- Atkinson JL, Abboud CF, Lane JI. Dramatic volume reduction of a large GH/TSH secreting pituitary tumor with short term octreotide therapy. Pituitary. 2005;8(2):89–91.
- Chanson P, Weintraub BD, Harris AG. Octreotide therapy for thyroid stimulating-secreting pituitary adenomas. A follow-up of 52 patients. Ann Intern Med. 1993;119:236–40.
- Mulinda JR, Hasinski S, Rose LI. Successful therapy for a mixed thyrotropin-and prolactinsecreting pituitary macroadenoma with cabergoline. Endocr Pract. 1999;5:76–9.
- 103. Mouton F, Faivre-Defrance F, Cortet-Rudelli C, Assaker R, Soto-Ares G, Defoort-Dhellemmes S, et al. TSH-secreting adenoma improved with cabergoline. Ann Endocrinol (Paris). 2008;69:244–8.
- 104. Kirkman MA. The role of imaging in the development of neurosurgery. J Clin Neurosci. 2015;22(1):55–61.
- 105. van Varsseveld NC, Bisschop PH, Biermasz NR, Pereira AM, Fliers E, Drent ML. A longterm follow-up study of eighteen patients with thyrotrophin-secreting pituitary adenomas. Clin Endocrinol. 2014;80(3):395–402.
- 106. Nazato DM, Abucham J. Diagnosis and treatment of TSH-secreting adenomas: review of a longtime experience in a reference center. J Endocrinol Investig. 2018;41(4):447–54.



9

Nonfunctioning Pituitary Adenoma

Maria Yavropoulou, Marina Tsoli, and Gregory Kaltsas

9.1 Introduction

Pituitary adenomas are predominantly monoclonal benign tumors that arise from expansion of single mutated precursor cells that possess a certain proliferative advantage. Due to their neuroendocrine component, the term pituitary neuroendocrine tumors (PitNETs) has recently been introduced [1]. Pituitary adenomas comprise different histological subtypes, according to their specific adenohypophyseal hormonal immunostaining and expression of pituitary-specific transcription factors. A number can present with a distinct clinical syndrome secondary to the secretion of a peptidic hormone to the circulation (functioning PitNETs). Nonfunctioning pituitary adenomas (NFPAs), in contrast to their functioning counterparts, are defined as pituitary tumors that are not associated with a secretory phenotype. The majority of these neoplasms are benign and are following a relatively indolent course although a subset may present a more aggressive behavior with resistance to employed treatment and/or early recurrences, whereas a small subset may become truly malignant developing distant metastases [2].

NFPAs are diagnosed either due to symptoms related to mass effect and compression to nearby vital structures and/or the presence of anterior hormonal deficiencies or incidentally during imaging investigations performed for unrelated purposes (pituitary incidentalomas). In the presence of hormonal deficiencies,

The chapter has been endorsed by **Prof. Gerald Raverot**, gerald.raverot@chu-lyon.fr, Faculté de Médecine Lyon Est, Université Lyon, Lyon, France

M. Yavropoulou (🖂) · M. Tsoli · G. Kaltsas

Endocrinology Unit, 1st Department of Propaedeutic and Internal Medicine, Medicine School, National and Kapodistrian University of Athens, Athens, Greece e-mail: myavropoulou@med.uoa.gr

[©] Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_9

adequate replacement is needed, whereas surgical resection is undertaken in cases of neurological abnormalities or visual impairment and/or in the presence of rapid tumor growth. Alternatively, active surveillance can also be considered in the absence of such manifestations. Radiotherapy (RT) may also be considered as adjuvant treatment in aggressive tumors and in case of significant growth of a tumor remnant or relapsing adenoma, while temozolomide, which is an alkylating agent, has been currently recommended as a therapeutic option for aggressive neoplasms or pituitary carcinomas [2].

The pathogenesis of NFPAs still remains elusive, but it has been proposed that disruption of cell cycle regulators, as the initiating event, can induce pituitary tumorigenesis. Classic oncogene mutations are rarely encountered in pituitary tumors. Nevertheless, a considerable number of pituitary-specific cellular disruptions have been uncovered and affect oncogenes, transcription factors, tumor suppressors, cyclins, inactivating factors, or proliferation inhibitors. Unraveling the pathogenesis of NFPAs is a critical step in the quest of new subcellular treatment targets that will decrease morbidity and mortality [3].

In this chapter, we outline current knowledge about the pathophysiology and management of NFPAs.

9.2 Epidemiology

Nonfunctioning pituitary adenomas (NFPAs) represent approximately 30–40% [4] of anterior pituitary tumors, ranging from 14% to 54% in national-wide population studies [4–8]. They present with a peak occurrence between forth and eight decades, but data on gender predominance are inconsistent [9, 10].

Most of the NFPAs are sporadic but may also occur as a component of hereditary syndromes, such as multiple endocrine neoplasia type 1 (MEN1), and multiple endocrine neoplasia type 4 (MEN4), the Carney complex, and the familial isolated pituitary adenomas (FIPAs). Data from patients with MEN1 from a French and Belgian multicenter study demonstrated that among pituitary adenomas only 14.7% are NFPAs and half of them are invasive [11]. In contrast, results from the Dutch MEN1 Study Group have revealed a increased proportion of NFPAs among pituitary tumors in patients with MEN1 (approximately 42.3%) in a systematic presymptomatic screening [12].

NFPAs represent less than 20% of patients with FIPA and, in this context, are diagnosed at an average of 8 years earlier compared with their sporadic counterparts [13, 14]. Patients with Carney complex most commonly develop functioning somatotropinomas or less frequently prolactinomas, but cases with pituitary tumors and asymptomatic elevations of growth hormone (GH), insulin growth factor 1 (IGF-1), and prolactin (PRL) have also been described [1, 15, 16].

9.3 Histopathological Classification

In the 2017 WHO classification for endocrine tumors, the adenohypophyseal cell lineage designation of the secreting and nonfunctioning pituitary adenomas based on the immunohistochemical (IHC) profile is considered the golden standard for the new classification system [17, 18] (Table 9.1). Moreover, the introduction of transcription factors as a complementary diagnostic tool in the classification of NFPAs in order to confirm the specific pituitary cell linage differentiation has paved the way toward a more precise classification of NFPAs and accurate characterization of their biological behavior. Three of the pituitary-specific transcription factors are currently recommended in routine diagnostics of PAs: pituitary transcription factor 1 (Pit-1), steroidogenic factor 1 (SF-1), and T-box family member TBX19 (T-Pit).

Pit-1 plays a role in the differentiation of somatotroph, lactotroph, and thyrotroph cells and in the development of the respective tumors [19, 20]. SF-1 acts as transcription factor for the differentiation of gonadotroph cells and is expressed in gonadotropinomas [17, 20], and T-Pit is required for the

Subtype Hormone staining		Transcription factors				
Silent Somatotroph adenomas	GH, α-subunit	PIT-1				
Sparsely granulated	Diffuse and strong staining					
Densely granulated	Weak and; patchy staining					
Lactotroph adenomas	PRL	PT-1, ER a				
Sparsely granulated	Perinuclear staining					
Densely granulated	Diffuse staining					
Acidophilic stem cell Focal and variable staining of both PRL and GH						
Thyrotroph adenomas TSHβ, α-subunit		PIT-1, GATA2				
Silent Corticotroph adenomas	ACTH	T-PIT				
Type 1 (densely granulated)	Diffuse and strong staining					
Type 2 (Sparsely granulated)	Weak and patchy staining					
Crooke -cell	Peripheral staining					
GonadotrophFSHβ, LHβ, α-subunitAdenomas		SF1, GATA2, ERα				
Null cell adenomas	None	None				
PIT-1-positive adenomas (plurihormonal)	GH, PRL, TSHβ±, α-subunit	PIT-1				

 Table 9.1
 Classification of nonfunctioning pituitary adenomas based on hormone staining and nuclear expression of pituitary-specific transcription factors

Modified by Mete et al 2017 (17).

transcription of the precursor polypeptide pro-opiomelanocortin (POMC) to adrenocorticotropic hormone (ACTH) and promotes the differentiation of corticotroph cells [21, 22].

In addition, estrogen receptor- α (ER α) and guanine-adenine-thymine-adenine binding protein 2 (GATA-2) transcription factors are also associated with the differentiation of gonadotroph, lactotroph, and thyrotroph cells, and may be pathogenetically involved in the development of the respective tumors [19, 23–25].

9.3.1 Subtypes of Nonfunctioning Pituitary Adenomas

NFPAs comprise different histological subtypes, classified according to their immunostaining to different adenohypophyseal hormones and transcription factors.

The current WHO classification system recognizes NFPAs as variants of their functioning counterparts, with the exception of null cell PAs, which is the only sub-type without functioning counterpart [18, 26].

9.3.1.1 Nonfunctioning Silent Gonadotroph Adenomas

Silent gonadotroph adenomas (SGAs) comprise the vast majority (approximately 80%) of NFPAs and typically demonstrate, at least focally, immunostaining for beta-follicle-stimulating hormone (β -FSH), beta-luteinizing hormone (β -LH), and α -subunit [18]. Interestingly, it is the only type of NFPAs where the nonfunctioning variant predominates [27, 28].

Before the introduction of the SF-1, many silent gonadotroph adenomas that were LH/FSH immunonegative were mistakenly classified as null cell type pituitary adenomas [29, 30]. Although currently available anti-SF-1 antibodies exhibit several methodological difficulties, the SF-1 nuclear labeling is detected in a significant proportion of tumor cells, confirming the diagnosis in cases with sparse or no gonadotroph hormone expression [31]. The distinction between SGAs and true null cell adenomas is of significant clinical relevance since true null cell adenomas follow with a more aggressive biological behavior than SGAs [32]. In addition, low ER α nuclear expression in tumor cells of SGAs is considered a good predictive tool of aggressiveness in these tumors [33].

9.3.1.2 Nonfunctioning/Silent Corticotroph Adenomas

Silent corticotroph adenomas (SCAs) represent approximately 15–20% of NFPAs [34, 35], and it is second most common type after SGAs. The real proportion, however, is probably underestimated, since routine nuclear staining for the transcription factor T-Pit, which regulates corticotroph differentiation, is not widely available [36].

Despite the fact that clinical characteristics of Cushing's syndrome are not present by definition, SCAs can either present with normal cortisol secretion (totally silent) or slightly elevated ACTH levels (clinically silent) [37–40]. SCAs are usually macroadenomas associated with mass-related symptoms and cavernous sinus invasion and show a female preponderance [41]. Histologically, SCAs, similar to their functioning counterparts, can be further divided into two subtypes based on specific morphological and ultrastructural characteristics: type I densely granulated SCAs, which show strong ACTH immunoreactivity, and type 2 sparsely granulated SCAs, which are more common and demonstrate weak and focal ACTH staining. Both subtypes demonstrate a more aggressive biological behavior compared with their functioning counterparts [17]. In addition, POMC mRNA levels are higher in type I SCAs compared with type 2 SCAs, and similar to what is found in functioning corticotroph adenomas [42]. Crooke cell adenoma with a typical perinuclear ring-like accumulation of cytokeratin and relocation of ACTH positivity to the sub-membranous zone has been rarely documented in clinically silent corticotroph adenomas [43, 44] despite the absence of exposure to high circulating glucocorticoid levels.

In 2017 WHO Classification of Pituitary Tumors, SCAs are graded as "high-risk adenomas" due to their aggressive clinical behavior and high probability of recurrence. In a large cohort of patients with aggressive pituitary tumors and pituitary carcinomas, which investigated the efficacy of temozolomide as first-line chemo-therapeutic treatment, corticotroph tumors prevailed accounting for approximately 45% of the cohort. Interestingly, a high proportion of the initially silent corticotroph tumors (26%) of this cohort were evolved in ACTH-secreting tumors [45]. Other studies, however, comparing recurrence rates of SCAs with other NFPAs subtypes have provided conflicting results. In a recent meta-analysis, based on studies with mean follow-up of more than 5 years, SCAs show recurrence of approximately 31%, similar to what has been reported to other subtypes of NFPAs [46].

Transformation of SCAs into functioning corticotroph adenomas (and vice versa), although rare, has also been reported [39, 43, 47–49]. In a recent study with 15 years of follow-up, only 4% of ACTH-immunohistopositive adenomas showed signs of transformation based on histology (2.8% adenomas transformed from SCA to ACTH-secreting adenomas and 1.1% transformed from ACTH-secreting adenomas to SCA).

The underlying molecular mechanisms that drive the tumorigenesis of these unique pituitary tumors remain largely unknown. The vast majority of SCA cells, however, exhibit multiple dark lysosomes, fusion of secretory granules with lysosomes, and extensive disposal of granules by lysosomes and autophagy [50] on electron microscope suggesting that either lysosome dysfunction or abnormal ACTH synthesis leads to destroy of ACTH before it is secreted [50].

Another hypothesis on the pathogenesis of these tumors is related to an impaired post-translational processing of the pro-hormone POMC in the biologically active mature hormone ACTH that is dependent on the expression of pro-hormone convertase 1/3 (PC1/3). SCAs show a significant decrease in PC1/3 expression compared with corticotroph adenomas associated with ACTH-secreting adenomas [51, 52].

In general, SCAs are pituitary adenomas that are biologically and clinically distinct from both functioning corticotroph adenomas and other NFPAs. Approximately one-third of patients with SCA may develop new-onset hypopituitarism and show tumor recurrence, underlying the need for close monitoring of these patients [40]. As in most NFPAs, silent CAs are diagnosed relatively late when they have already formed invasively growing macroadenomas and, in rare cases, can transform to carcinomas [53, 54].

9.3.1.3 Nonfunctioning/Silent Somatotroph Adenomas

Silent somatotroph adenomas (SAs) represent approximately 2–4% of all pituitary adenomas in surgical series [55, 56].

Growth hormone immunostaining in silent SAs varies widely from very weak to strong positive but is usually less than somatotropinomas causing acromegaly [55, 57]. In cases with very low or absent GH staining nuclear expression of the transcription factor, Pit-1 is a valuable diagnostic tool, since all GH tumors express Pit-1 [56].

Patients with silent SAs usually present with normal GH and IGF-1 levels, but there have been few reports of "clinically silent" cases, with non-suppressible serum GH and elevated IGF-1 levels [58–61]. In addition, approximately 12% of silent SAs have been reported to progress in clinical acromegaly during long-term follow-up, particularly in women [62–64].

Similar to their functioning counterparts, silent SAs are classified as densely, sparsely, or intermediately granulated, based on the presence and pattern of the low molecular weight cytokeratin (CAM 5.2) staining. The sparsely granulated subtype is more frequently reported and is considered a more aggressive subtype, while more than 50% of the silent SAs (twofold higher than somatotropinomas causing acromegaly) are adenomas with mixed GH and prolactin (PRL) secretion [55, 62].

In addition, silent SAs exhibit distinct biological and epidemiological characteristics compared to their functioning tumors. They are usually larger, exhibit a more aggressive biological behavior, recur earlier and more frequently, and are more common in females, and at younger ages between 20 and 40 years [55, 62].

Both functioning and silent SAs express somatostatin receptors, 2 (SSTR2) and 5 (SSTR5), with SSTR2 expression being significantly lower in silent compared to functioning SAs [55]. In one series SSTR2 was expressed in all silent SAs reviewed, while in another one SSTR2 was expressed in approximately 50% of them [62]. Expression of SSTR5 was found similar between functioning and silent SAs. Nevertheless, silent SAs show a lower response to somatostatin analog therapy compared to their functioning counterparts [65].

The causes that lead to silence in these tumors remain unknown although several hypotheses have been suggested. Abnormalities downstream the Pit-1 signaling pathway [66], impaired synthesis or post-translational processing of GH leading to reduced circulating levels [30, 67], or synthesis of an immunoreactive form of GH without biologic activity has been proposed as the most plausible pathophysiological mechanisms in these tumors [48]. In addition, inhibition of GH release is highly unlike since cytoplasmic lysosome accumulation has not been described in silent SAs [57]. Mutations in the aryl hydrocarbon receptor-interacting protein (AIP) have also been reported in two cases of silent SAs [68]. Nevertheless, silent SAs usually present as giant tumors with higher recurrence rate and higher need for radiation therapy compared to other NFPAs, and thus, close follow-up is strongly indicated [56].

9.3.1.4 Nonfunctioning/Silent Thyrotroph Adenomas

Silent thyrotroph adenomas (STAs) are extremely rare but are reported more frequently compared to their functioning counterparts (thyroid-stimulating hormone, TSH-omas) associated with hyperthyroidism [17, 18]. STAs share many histopathological characteristics with TSH-omas and are almost indistinguishable on histology but display unique histopathological features compared to other types of pituitary adenomas. Both thyrotroph tumors present with TSH- β , and α -subunit immunohistochemical staining, express Pit-1 and GATA-2 transcription factors, and show significant membrane immunoreactivity for SSTR-2A and SSTR-5 [69]. Usually, they are larger and more invasive compared to TSH-omas causing hyperthyroidism [69], but they seem to behave similarly regarding treatment outcomes and recurrence rates [70]. A few cases of STAs evolving to functioning TSH-omas have also been described [33, 63].

9.3.1.5 Nonfunctioning/Silent Lactotroph Adenomas

Silent lactotroph adenomas (SLAs) are very rare in surgical series [29] although more frequently reported in autopsy series [71]. In addition to PRL-positive staining, SLAs also express Pit-1 and ER-a [17, 18]. Mono-hormonal SLAs are extremely rare and more often present as silent mixed somato-lactotroph adenomas in Pit-1-positive tumors [62].

As their functioning counterparts (PRL-omas), SLAs are classified into sparsely (with Golgi-like prolactin immunoreactivity) and densely (with diffuse cytoplasmic prolactin immunoreactivity) granulated subtypes [17, 18]. Despite that SLAs lack clinical symptomatology of hyperprolactinemia, high prolactin levels are reported due to stalk compression [71].

9.3.1.6 Null Cell Adenomas

Null cell adenomas (NCAs) are defined by the lack of immunohistochemical staining of any anterior pituitary hormone and pituitary-specific transcription factors [17, 18]. They present a very small proportion of all pituitary tumors [36], although their real frequency is probably overestimated due to methodological pitfalls in immunohistochemical protocols and lack of widely available and reliable antibodies to pituitary-specific transcription factors. The diagnosis of NCAs is usually made by exclusion and their differential diagnosis includes both adenohypophysial and nonadenohypophysial neuroendocrine tumors of the sellar region [72].

Data from a retrospective case series of 516 patients with NFPAs have shown that from the 23% of the tumors initially classified as NCAs by using only the classical pituitary hormone IHC, only 5% remained as true NCAs, when lineage-specific markers, such as Pit-1, SF-1, and T-Pit, were used [73].

9.3.1.7 Plurihormonal Pit-1-Positive

Pit-1-positive plurihormonal adenomas (previously named as silent subtype 3 adenomas) are a distinct entity, with reportedly aggressive behavior [17, 18]. In a single-center retrospective case series, their prevalence was less than 1% among resected pituitary tumors in a period of 13 years [74]. Histologically, these tumors
consist of large polygonal or spindle-shaped cells, with atypical nuclei, sometimes containing inclusions, that can be identified on routine HE stains or electron microscopy [17, 18, 75, 76].

They usually demonstrate immunoreactivity for GH, PRL, and TSH in different combinations, and despite their silent nature, approximately 30% of these tumors are associated with clinical signs of Pit-1 lineage hormones hypersecretion leading to hyperthyroidism, acromegaly, or hyperprolactinemia [77]. The correct diagnosis is very challenging in everyday clinical practice but of critical importance since these tumors are usually macroadenomas with aggressive biological behavior, high invasion rates to cavernous sinus and clivus, and high recurrence rates [75, 77].

9.3.2 Nonfunctioning Pituitary Carcinomas

Nonfunctioning pituitary carcinomas of gonadotroph or null cell type have rarely been reported, but their true prevalence is unknown since their metastases remain asymptomatic for many years [78, 79].

9.4 Pathophysiology

Despite recent advances in molecular biology and genetics, the pathophysiology of NFPAs is yet far from being fully elucidated. Many hypotheses, which mainly include genetic and epigenetic events, as well as hormonal stimulation, impaired intracellular signaling pathways, and microRNAs seem to finally converge in severely disrupted and uncontrolled tumor growth and proliferation of pituitary cells.

9.4.1 Genetics

Although the majority of NFPAs occur sporadically, recent advances in molecular genetics have identified an increased prevalence of inherited genetic susceptibility in PitNETs.

Genetic mutations in a single cell leading to overexpression of proto-oncogenes or inactivation of tumor suppressor genes, and functional or epigenetic alterations of transcription factors that regulate cell growth and differentiation are the most prominent events in the genetic background of pituitary tumorigenesis. Among the genes that have been implicated in the pathophysiology of pituitary adenomas, guanine nucleotide-binding protein, alpha-stimulating (GNAS), aryl hydrocarbon receptor-interacting protein (AIP), and pituitary tumor-transforming gene (PTTG) have been more frequently described in NFPAs [80, 81].

In a Brazilian cohort of patients with pituitary adenomas, GNAS mutations were found in much lower proportion in NFPAs compared to somatotropinomas (4.8% vs. 27%), whereas PTTG overexpression and AIP underexpression were reported in almost all cases without significant differences, however, between the NFPAs and

somatotropinomas [82]. Moreover, in this cohort neither GNAS mutations nor impaired expression of PTTG and AIP was associated with tumor clinical characteristics and treatment outcomes [82].

In the MEN1 syndrome caused by germline heterozygous mutations in the MEN1 gene, encoding for menin protein in chromosome 11q13, pituitary adenomas (that are present at approximately 15–50% of patients) coexist with parathyroid hyperplasia and neuroendocrine tumors of the gastro-enteropancreatic tract [83]. NFPAs are not the predominant type of Pit-NETs in patients with MEN1, but when they develop in this genetic background display a more aggressive behavior, similar to their functioning counterparts [84, 85].

Another genetic syndrome leading to the development of pituitary adenomas in association with other endocrine neoplasms is MEN4, caused by germline mutations in the CDKN1B gene, encoding for p27Kip1, a negative regulator of cell cycle progression. In patients with MEN4 that develop pituitary adenomas, NFPAs occur more frequently, [86] compared to MEN1. Whether CDKN1B mutations are also present in sporadic NFPAs is not yet known.

Carney complex is caused by inactivating mutations in the PRKAR1A gene that encodes for regulatory subunit type 1alpha of the cAMP-dependent protein kinase A (PKA) and acts as a tumor suppressor gene [1]. Almost half of the patients with Carney complex have germline-inactivating mutations in the PRKAR1A gene, and in those who develop pituitary tumors, the normal allele of PRKAR1A gene in the pituitary is lost. This loss of heterozygosity of the PRKAR1A gene leads to somato-mammotroph hyperplasia, which in turn may lead to additional genetic changes at the somatic level, and the formation of adenomas [1, 15, 16]. Patients with Carney complex develop acromegaly at a 10–12%, but clinically silent GH and IGF-1 elevations have also been described [16].

The role of PRKAR1A somatic mutations in the development of sporadic pituitary tumors is currently unknown [87].

Although NFPAs account frequently in familial isolated pituitary adenomas (FIPA) (14.5% of the patients), the role of AIP germline mutations in the pathophysiology of NFPAs has demonstrated conflict results [88–91] and therefore does not seem to significantly contribute in the genetic background of NFPA tumorigenesis.

Coexistence of pituitary adenomas and pheochromocytoma/paragangliomas is very uncommon, but a germline mutation in two of the genes encoding succinate dehydrogenase SDHC and SDHD has been reported in a 30-year-old man with a NFPA [92].

In general, data reported so far underline the significant contribution of epigenetic alterations and somatic mutations rather than a role for germline mutations in NFPA tumorigenesis.

As such, the tumor suppressor protein p16, a cyclin-dependent kinase inhibitor encoded by the CDKN2A gene, is frequently reported to be downregulated in NFPAs [93]. Similarly, significant downregulation in NFPAs is reported for the DNA damage-inducible gene 45 g (GADD45g), due to promoter methylation [94], and the maternally expressed gene 3 (MEG3), which acts as a tumor suppressor

gene [95]. Both GADD45g and MEG3 are regulated by p53-protein, which negatively regulates pituitary cell growth, pointing the fact that impaired control of cell proliferation plays a key role in NFPA tumorigenesis. Activating somatic mutations of the PIK3CA gene that encodes for the catalytic subunit of PI3-kinase IA, within the AKT signaling pathway [96], have also been described in NFPAs.

9.4.2 Epigenetics

Despite the advanced progress in the identification of recurrent somatic alterations and rare inherited variants, their frequencies among pituitary adenomas are low, suggesting that factors other than gene mutations may be linked with the pathogenesis of sporadic pituitary adenomas.

In recent years, epigenetics present a new discipline that attempts to explain significant differences in phenotypes among patients with the same disease, such as diabetes mellitus or cancer [97]. There are two main types of epigenetic modifications—DNA methylation and histone modifications.

Methylation, in particular, changes the activity of a DNA segment without changing the sequence and it usually acts to repress gene transcription.

DNA methylation analysis in a cohort of NFPAs and functioning adenomas revealed tumor-specific genome-wide patterns of DNA methylation and gene expression between different subtypes of pituitary adenomas. In particular, genomewide DNA methylation profile was very similar between silent and functioning corticotroph adenomas, whereas functioning SAs display higher levels of hypomethylated regions compared to functioning corticotroph adenomas and SGAs [98]. In addition, significant differences were noted in the methylated regions of SGAs (high proportion of hypermethylated DNA regions) and SCAs (higher proportion of hypomethylated DNA regions).

In contrast to the other epigenetic mechanisms that modulate gene transcription, microRNAs (miRs) are small noncoding single-stranded RNA molecules of approximately 22 nucleotides, acting at the post-transcriptional level and directly modulating gene expression of mRNA genes. MicroRNAs bind mRNAs through complementary base pairing, resulting in suppressed translation from the mRNAs or degradation of the mRNAs through the formation of an RNA-induced silencing complex (RISC). Although these molecules are reported to target both activating oncogenes and tumor suppressor genes, their exact role in pituitary tumorigenesis has not yet been clarified [99–104].

Several microRNAs have been found to be deregulated in NFPAs, compared to normal pituitary tissue (Table 9.2); however, a consistent molecular profile of specific microRNAs that could serve as novel biomarkers or as therapeutic molecular targets has not been identified. Nevertheless, a wide variety of target genes appear to be involved, with most being growth factors and regulators of cell cycle division. Interestingly, up to now research in microRNAs profile in pituitary adenomas has

MicroRNA	Differential Expression	Target Gene
miR-107(186)	Increase	AIP
miR-598/181d/191-3p/ 181b-5p/3676-5p/383(103)	Increase	-
miR-23b(187)	Decrease	HMGA2
miR-130b(187)	Decrease	CCNA2
miR-424/503(188)	Decrease	CDC25A
miR-135a/429/140-5p/ 582-3p/938/582-5p(189)	Increase	Smad3
miR-197/33b(189)	Increase	DLK1
miR-15/16/26a/196a2, Let- 7a(190)	Decrease	HMGA1/HMGA2
miR-133(191)	Decrease	FOXC1
miR-20a/17-5p(192)	Increase	PTEN/TIMP2
miR-106b(191, 192)	Increase	PTEN/TIMP2 PI3K/AKT
miR-124a(193)	Increase	-
miR-144/373/422b/202/520e/32/ 422a/181c/181b/520c/188/155/ 520 f/520b/182/10b/523/146a(193)	Increase	-
miR- 31/506/218/503/513/514(193)	Increase	-
miR-128a/516a-3p/15(194)	Increase	Wee1
miR-195(194)	Increase	
miR-20a/93(194)	Increase	
miR-524-5p(195)	Decrease	PBF

Table 9.2	Differential	expression	of microF	RNAs in	nonfunctioning	pituitary a	adenomas
	Differential	expression	or micror	VI 17 19 111	nomuneuoning	prununy	aucinomias

failed to demonstrate epigenetic alterations on specific genes that have been traditionally linked to pituitary tumorigenesis. Thus, it appears that genetic mutations and epigenetic alterations involve distinct target genes but may act synergistically in regulating pituitary tumorigenesis.

9.4.3 Intracellular Molecular Signaling Pathways

Over the past 30 years, studies of developmental biology have identified the role of evolutionarily highly conserved intracellular signaling pathways such as Wnt, and Notch in pituitary development and disease. Through these pathways, specific secreted proteins control differentiation and function of recipient cells in a paracrine and/or autocrine manner. Various components of the Notch and Wnt signaling have been found deregulated in NFPAs, while data on the role of Hh signaling are currently missing.

9.4.3.1 The Role of Notch Intracellular Signaling Pathway in the Pathophysiology of NFPAs

Notch signaling pathway regulates cell fate through lateral inhibition and formation of boundaries, both of which represent patterning processes of critical importance in the regulation of spacing of different cell types within tissues [105, 106]. There are four Notch receptors in mammals (Notch1, Notch2, Notch3, and Notch4) and five classic DSL (Delta/Serrate/Lag-2) ligands: Jagged1, Jagged 2, Delta-like 1, Delta-like 3, and Delta-like 4.

Notch receptors are single-pass transmembrane proteins composed of a functional extracellular domain (NECD), a transmembrane domain, and an intracellular domain (NICD). Notch ligands are also transmembrane proteins in the signalsending cells, and both ligands and receptors require a catalytic process to become active (Fig. 9.1).

Expression of Notch 3 and Jagged 1 protein has been found significantly elevated in NFPAs compared with normal pituitary tissue [107]. Gene microarrays and proteomic analyses have confirmed increased expression of Notch 3 gene and protein in human NFPAs [107, 108] but not in GH- and PRL-secreting adenomas. In addition delta-like 1 ligand is strongly downregulated in NFPAs and in PRL-secreting adenomas, while cyclin D1, one of the Notch target genes, is found overexpressed in NFPAs [109].

9.4.3.2 The Role of Wnt Intracellular Signaling Pathway in the Pathophysiology of NFPAs

Wnt forms a family of 19 highly conserved secreted signaling molecules, rich in cysteine, which play an important role in the development and function of multiple tissues [110]. The canonical Wnt signaling pathway involves the formation of a complex between extracellular Wnt proteins, and transmembrane co-receptors the



Frizzled and the LRP5 or LRP6 receptors [111]. When activated, the ligand–receptor complex inhibits the cytoplasmic degradation of beta-catenin, promotes its translocation in the nucleus, and activates the transcription of target genes [112] (Fig. 9.1).

In a mouse model of MEN1 syndrome, Wnt proteins (Wnt4 and Wnt9a) and receptors (Fz6 and LRP2) were found significantly downregulated in all tumors related to this syndrome, from pituitary, pancreas, parathyroids, ovary, and testes [113].

Nuclear accumulation of β -catenin was found increased in 57% of pituitary tumors, with the higher expression reported in tumors originated from cells of the Pit-1 lineage [114], but this result was not confirmed in others [115]. Wnt inhibitors were also studied in human pituitary adenomas. Wnt inhibitory factor-1 (WIF-1) is a secreted protein that binds to Wnt proteins and inhibits their activities. WIF-1 was found significantly downregulated on a microarray analysis in a series of both functioning and nonfunctioning pituitary adenomas when compared with healthy pituitary tissue. Collectively, recent studies in vitro, in vivo, and in human tissue point to a potential role of Wnt signaling in pituitary adenomas both functioning and nonfunctioning, but further work is warranted in order to elucidate the exact molecular mechanisms involved [115].

9.5 Clinical Presentation

The clinical presentation of NFPAs varies from completely asymptomatic to panhypopituitarism and manifestations attributed to mass effect to nearby structures. The absence of clinical symptoms related to hormonal hypersecretion is associated with a significant delay in diagnosis estimated approximately to 1.96 ± 2.9 years [9].

Headache is a common neurologic symptom attributed to an expanding sellar mass and is observed in 19–75% of patients [116]. Possible mechanisms related to headache include increased intrasellar pressure and starching of dural membrane or invasion of cavernous sinus and trigeminal nerve irritation [14, 117]. A retrospective study found that 20% of the patients with incidentally observed NFPAs reported that suffered from headache [118].

Visual impairment is observed in 58% of patients with NFPAs [119]. Suprasellar extension of the pituitary adenoma can cause pressure on the optic chiasm, typically resulting to bitemporal hemianopia. Initially, the patients may be unaware of the visual deficit, but in case of severe and long-term compression visual acuity may be affected and optic atrophy or papilledema may occur [120]. Older age is considered to be associated with delayed diagnosis of NFPAs in patients with visual defects [121]. Visual field loss may be uni-, bilateral, or central as well as complete or partial depending on the site and degree of nerve compression [120]. In a recent study of 103 patients presenting to a neuro-surgical unit with a pituitary adenoma, it was observed that although bitemporal visual field loss was the most common defect (41%), a significant proportion of patients had unilateral or altitudinal defects [122]. Several mechanisms are involved in the pathophysiology of visual impairment that

may initially be reversible but becomes irreversible in case of long or intense chiasm compression. Reversible mechanisms include axoplasmic flow disorder, conduction blocking, and demyelinization. Axonal fiber degeneration is associated with irreversible visual defects [120].

Ocular motor impairment associated with large pituitary adenomas is attributed to cavernous sinus invasion and compression of ocular motor nerves III (common ocular motor nerve), IV (trochlear nerve), and VI (abducens nerve) [120]. The clinical manifestations vary according to the compromised nerve. Before true ocular motor impairment develops, some patients report frequent episodes of diplopia. The common ocular motor nerve is the most frequently affected resulting to eyeball shift outward, ptosis, and mydriasis. Trochlear nerve involvement causes upward and slightly inward deviation of the eyeball, while nerve VI palsy is related to abduction deficit [120, 123, 124].

In some rare cases, large aggressive NFPAs may invade other intracranial structures such as the posterior and anterior midbrain, the third ventricle or the brain causing the dorsal midbrain syndrome, intracranial hypertension, and hydrocephalus or temporal epilepsy. Invasion of the sellar floor may result to cerebrospinal fluid rhinorrhea. These tumors also display early recurrence after the initial surgical resection and rapid local growth and tumor extension [2, 120, 125].

Hypopituitarism is a very common manifestation in patients with NFPAs. Multiple cases are diagnosed when central hypothyroidism is observed during routine thyroid testing or in male patients who present with decreased libido and erectile dysfunction due to secondary hypogonadism. Pituitary hormone deficiency is attributed to compression of the normal anterior pituitary cells and/or pituitary stalk, affecting the stimulation from the hypothalamic factors and the secretion of pituitary hormones. GH deficiency is the most commonly observed followed by gonadotropin, corticotropin, and thyrotropin insufficiency [14, 126, 127]. In addition, disconnection hyperprolactinemia may be observed due to pituitary stalk compression as it prevents dopamine from reaching the anterior pituitary and could account for the observed cases of hypogonadism in the absence of gonadotropin deficiency. In patients with NFPAs, serum prolactin levels are rarely higher than 2000 mU/L (95 ng/ml) in the absence of medications affecting the prolactin levels [71]. Diabetes insipidus (DI) is an uncommon finding in case of NFPA and when encountered another than an adenoma pathology should be considered [128].

Pituitary apoplexy is a life-threatening situation that is attributed to acute hemorrhage or ischemic infarction of pituitary and is characterized by severe headache of sudden onset, nausea, vomiting, neuro-ophthalmologic symptoms, and impaired consciousness level. Furthermore, it may cause partial or complete hypopituitarism, particularly hypocortisolemia that is observed in 70% of cases [129, 130]. The real risk of apoplexy in NFPAs is largely unknown. In a retrospective study of 485 patients with NFPAs, pituitary apoplexy was the first presentation in 8% of cases [131]. A systematic review and meta-analysis regarding the natural history of pituitary incidentalomas and NFPAs reported an incidence of apoplexy in macroadenomas of 1.1% per year [132].

9.6 Diagnostic Evaluation

The identification of NFPA is based on the identification of a sellar mass on computed tomography (CT) or magnetic resonance imaging (MRI) of the sella turcica. The increased availability of CT and MRI and recent progress on neuroimaging have resulted in increased recognition of sellar or parasellar lesions, randomly, during imaging investigations performed for other reasons. These lesions are termed as pituitary incidentalomas [133].

MRI of the sella turcica with gadolinium is considered the gold standard modality for the initial evaluation or follow-up of pituitary lesions, while CT may display some advantage in the estimation of bone structures or calcifications [134]. Pituitary adenomas appear hypo- or isointense to normal pituitary on T1-weighted images, while on T2-weighted images they are isointense to the white matter [135] (Fig. 9.2). After the administration of gadolinium, the pituitary adenomas display lowcontrast enhancement on T1-weighted images. There may be areas of necroses or hemorrhage that appear hyperintense on T1-weighted images without contrast. Microadenomas measuring less than 10 mm are typically small intrasellar lesions, while macroadenomas measuring above 10 mm may extend to the suprasellar cistern and compresses the optic chiasm or extend into the cavernous or sphenoid sinus [14, 135].

There are two systems of radiological classification of pituitary adenomas: Knosp's classification that divides pituitary adenomas into five categories according to parasellar extension and invasion of cavernous sinus (grades 0–IV) and Hardy's classification that comprises two subscales and estimates the sphenoid bone invasion (grades 0–IV) and the suprasellar extension of and adenoma (types A–E) [2, 136].

All patients with a pituitary incidentaloma or a clinically apparent NFPA should undergo a complete evaluation of anterior pituitary function as in many cases hormone hypersecretion or hypopituitarism may be subtle and slowly progressive



Fig. 9.2 T1 (**a**)-weighted and T2 (**b**)-weighted coronal MRI images showing a large pituitary lesion with suprasellar extension. The lesion causes elevation of the optic chiasm and contacts the right and left cavernous internal carotid artery

Table 9.3 Initial endocrine evaluation of clinically NFPAs

Evaluation for hormone hypersecretion

- Measure IGF-1
- Measure prolactin (in dilution in case of large macroadenomas)
- Screening for glucocorticoid excess in case of clinical suspicion (overnight dexamethasone suppression test, late-night salivary cortisol)

Evaluation for hypopituitarism

- Measure cortisol (09:00 a.m.), TSH, fT4, IGF-1, prolactin, FSH, LH, testosterone (males)
- A morning cortisol level < 3 μ g/dl is suggestive of adrenal insufficiency, while a cortisol level > 15 μ g/dl excludes the diagnosis. If cortisol level is between 3 μ g/dl and 15 μ g/dl, a corticotropin stimulation test should be performed
- In case, GH deficiency is suspected, and GH stimulation tests should be performed to exclude or confirm the diagnosis
- Gonadal function in premenopausal women is assessed through history and examination. Low gonadotropin levels in postmenopausal women suggest hypopituitarism, while in men exclude primary hypogonadism if testosterone levels are low

IGF-1 insulin-like growth factor 1, *TSH* thyroid-stimulating hormone, *fT4* free thyroxine, *FSH* follicle-stimulating hormone, *LH* luteinizing hormone, *GH* growth hormone

(Table 9.3). Screening tests should involve serum PRL, IGF-1, TSH, fT4, FSH, LH, and testosterone for males. Gonadal function in premenopausal women can be assessed by history and clinical examination, while in postmenopausal women low gonadotropin levels suggest the presence of hypogonadism. In addition, screening for cortisol excess should be based either on midnight salivary cortisol or on overnight dexamethasone suppression test [14, 137, 138]. In case of suspected GH or adrenal insufficiency, GH stimulation and corticotropin/insulin stimulation tests are recommended, respectively [14]. In case of adrenal or thyroid deficiency, proper replacement should be initiated.

Ophthalmologic investigation should include assessment of visual acuity, pupil, and fundus examination, ocular motor evaluation, and visual field assessment [120]. According to Endocrine Society, a formal visual field examination is recommended in all patients with a pituitary incidentaloma that abuts or compress the optic chiasm [138]. The Congress of Neurological Surgeons has recently introduced guidelines regarding the pretreatment ophthalmologic evaluation in patients with NFPAs [139]. Their approach aims at identifying early and asymptomatic visual deficits as well as at providing prognostic factors of recovery and facilitating the postsurgical follow-up. Automated static perimetry is recommended for the initial assessment of visual fields, while visual evoked potentials may be used in order to evaluate the optic nerve function. Patients of advanced age or with a long duration (>4 months) of vision deficit present a low probability of visual recover postoperatively. Optical coherence tomography (OCT) of the optic nerve head is not yet standard of practice but is usually performed in case of visual impairment for prognostic purposes, to estimate the chance of postoperative vision improvement [120, 139].

9.7 Differential Diagnosis

The differential diagnosis of a pituitary incidentaloma involves multiple entities arising from the sellar or parasellar region (Table 9.4). Pituitary adenomas and Rathke's cleft cysts account for up to 90% of all lesions [133]. Adenomas and meningiomas represent the most common solid tumors in the sellar region. Imaging characteristics provide valuable help for the differential diagnosis, while clinical and endocrine evaluation may also point to different diagnoses. It is important that diabetes insipidus is not a common feature of pituitary adenomas and may be associated with lesions of non-pituitary origin [140]. Furthermore, α -subunit of the gly-coprotein hormones may be elevated in 30% of patients with NFPAs, but normal values do not exclude the diagnosis of an NFPA and its measurement is not systematically recommended [141, 142]. The diagnosis of a truly malignant pituitary tumor (pituitary carcinoma) is based on the presence of craniospinal or systemic metastases [143].

In addition, it is very important to differentiate NFPA from a prolactinoma since sometimes the hormonal hypersecretion may not be clinically evident and the

 Table 9.4
 Differential diagnosis of sellar lesions

Pituitary tumors

- · Pituitary adenoma
- Pituitary carcinoma
- · Pituitary hyperplasia
- · Pituicytoma
- · Granular cell tumors
- **Other tumors**
- · Craniopharyngioma
- Meningioma
- Neurinoma
- · Germ cell tumor
- Teratoma
- Glioma
- Chordoma
- Lymphoma
- · Pituitary metastases

Cysts

- · Cyst of Rathke's pouch
- Arachnoid cyst
- · Epidermoid/dermoid cyst

Inflammatory/granulomatous lesions

- · Lymphocytic hypophysitis
- · Sarcoidosis
- · Langerhans cell histiocytosis
- Tuberculosis
- · Wegener's granulomatosis
- · Pituitary abscess

Vascular lesions

- Aneurysms
- · Cavernous angiomas

management of these two entities is significantly different. It has been observed that the incidence of hyperprolactinemia in histologically verified NFPAs is 25–65% [137]. PRL levels provide valuable help, while the hook effect may occasionally confound the diagnosis [144]. In a retrospective study, it has been observed that patients with NFPAs most often had PRL levels below 100 ng/ml, while levels >250 ng/ml were observed exclusively in patients with prolactinomas [145].

9.8 Treatment

9.8.1 Active Surveillance

Although few data are available regarding the treatment of asymptomatic NFPAs, the best strategy in these cases is considered the conservative follow-up. However, there is no established follow-up algorithm and guidelines are typically based on clinical experience [138, 146].

It is recommended that microadenomas are followed up in 6 months with MRI, and if there is no progression, MRI may be repeated at 2 years without visual or hormonal evaluation. Surveillance may be stopped if no progression is observed at 2 years. No surveillance is recommended for microadenomas with diameter lower than 5 mm [146]. In case of a macroadenoma that is not close to the optic chiasm, it is recommended follow-up with MRI 1 year after the initial diagnosis and if no progression is observed surveillance is recommended every 2 years. Visual field assessment is suggested if the tumor enlarges to abut or compress the optic chiasm during follow-up imaging. In case of a macroadenoma close to the optic chiasm, MRI should be performed at 6 months and annually thereafter. Visual assessment may be performed every 6 months [146].

In case of macroadenoma, it also recommended clinical and biochemical assessment of anterior pituitary function for hypopituitarism development 6 months after the diagnosis and annually afterward. Routine follow-up for hypopituitarism is not suggested for microadenomas whose clinical picture and MRI do not change over time [138, 146].

9.8.2 Surgery

Surgical treatment of NFPAs is recommended in patients with visual field deficits or other visual or neurological abnormalities associated with NFPAs abutting or compressing the optic nerves or chiasm and in case of pituitary apoplexy [138, 147]. In addition, Endocrine Society suggests that surgery should be considered in tumors that display significant growth or cause loss of endocrine function and in patients with persistent headache or planning to become pregnant and the NFPA is close to the optic chiasm [138]. Active surveillance may be a better approach for older patients as they display higher surgical intervention risk, while they also have a shorter lifetime probability of tumor enlargement.

Currently, the standard surgical technique is endoscopy or microscopy-assisted transsphenoidal surgery (TSS). Intraoperative MRI has been recently introduced as a mean to improve the surgical resection of the tumor, but its use remains controversial [148, 149].

TSS, when performed by an experienced surgeon, is a safe procedure associated with relatively low complication rates. A recent meta-analysis showed that total resection is achieved in 60–73% of NFPAs [150]. Immediate tumor volume decrease was observed in nearly all patients with a residual tumor rate of 10–36% [147]. Visual improvement is observed in 75–91% of patients, while 35–50% of cases display also hypopituitarism improvement [147]. It has been reported postoperative improvement in gonadal, thyroid, and adrenal axes in 64.9%, 71.9%. and 33.9% of cases, respectively [151]. Postoperative complications occur to less than 5% of cases and include cerebrospinal fluid leakage, meningitis, vision deterioration, persistent DI, or vascular injury. The mortality rate has also been demonstrated to be low (<1%) [152].

In the early postoperative period, the patients should be carefully monitored for water and sodium balance derangements. In approximately 4–20% of patients, a transient syndrome of inappropriate antidiuretic hormone secretion (SIADH) may be observed in the first 3–7 days postoperatively and should be properly treated that is related to intraoperative manipulations in the posterior pituitary or pituitary stalk [153–155]. DI may also develop in 18–31% of cases, is transient, and presents usually in the first 24–48 h [155].

Perioperative glucocorticoid treatment is frequently used to cover probable adrenal insufficiency development during the operation. Some centers interrupt glucocorticoid therapy 48 hours postoperatively, while others continue treatment until the evaluation of adrenal axis sufficiency [156, 157].

Assessment of pituitary function and visual field examination should be done 1–3 months postoperatively and treatment of probable hormone deficiencies introduced. A sellar MRI should be performed 3–6 months postoperatively to estimate tumor resection and to serve as baseline MRI during subsequent follow-up. If there is no residual adenoma, 10-year regrowth rate has been calculated to 0–6%. In case of residual adenoma, 10-year regrowth rate is 42–53% and 77–80% if the remnant is intrasellar or extrasellar, respectively [158–160].

9.8.3 Radiotherapy (RT)

In the past, RT was administered postoperatively in all patients to prevent recurrence or growth of residual tumor. However, today the role of RT as adjuvant treatment is under debate. Besides conventional radiotherapy, newer techniques are currently available such as fractionated stereotactic radiotherapy and Gamma Knife radiosurgery that aim to deliver high precision radiotherapy with lower complication rates [161].

Several studies have evaluated the efficacy of adjuvant RT on tumor growth or recurrence. A recent study has reported a 85–95% of tumor control at 5–10 years

postoperatively in patients with NFPAs [162]. Mean progression free survival (PFS) at 5 years postoperatively has been reported to be 95% after RT compared to 70% in patients not treated with RT. At 15 years postoperatively, mean PFS was calculated at 93% after RT compared to 33% if no RT was administered [163]. In addition, there are some reports of improved visual function in patients with visual impairment after surgery treated with RT [164]. However, there are no randomized controlled trials indicating the superiority of adjuvant postoperative RT compared to active surveillance, while the potential side effects of RT make the indication for RT debatable [165]. Furthermore, a recent meta-analysis reported that residual tumor growth occurs slowly as the tumor doubling time is 3.4 years, while no growth is observed during follow-up in 50–60% of patients [166].

Hence, there is no consensus regarding adjuvant treatment with RT after surgical resection of NFPAs. RT is considered at the time of disease progression during follow-up, while immediate postoperative treatment should be reserved for cases with significant tumor remnant and high risk of progression [147]. Furthermore, adjuvant RT should be considered for patients presenting with aggressive NFPAs, large tumors with suprasellar extension or cavernous sinus invasion or displaying aggressive histopathological characteristics such as labeling index Ki-67 > 3% or extensive immunostaining for p53 [165]. RT can also be used as primary treatment in cases when surgery is not feasible.

The most common side effect of RT is the high incidence of hypopituitarism. Five years after treatment with RT, the incidence of GH, gonadotropin, ACTH, and TSH deficiency has been calculated to 100%, 91%, 77%, and 42%, respectively [167]. Follow-up with assessment of anterior pituitary function is recommended every 6 months post-treatment with RT. Additional rare side effects include optic neuropathy, neurocognitive dysfunction, and a probably increased risk of secondary malignancies of the central nervous system [164, 168, 169]. Furthermore, various studies have shown an increased risk of cerebrovascular events in patients treated with RT [170, 171]. It has been observed that the relative risk of a first stroke in patients treated with RT compared to the general population was 4.1 [170]. The risk of these side effects can be reduced if limited radiation dose is administered.

9.8.4 Medical Therapy

The role of medical treatment of pituitary adenomas is not established. Based on observations that NFPAs express dopamine receptors and somatostatin receptors, multiple studies evaluated the efficacy of dopamine agonists and somatostatin analogs in patients with operated or nonoperated NFPAs [172–176]. Tumor control (shrinkage or stabilization) was observed in 87% of patients treated with dopamine agonists upon imaging detection of residual tumor, but tumor shrinkage was achieved in 29% of cases [176]. A recent case–control study evaluated the efficacy of long-acting somatostatin analog octreotide LAR in patients with residual tumors that displayed positive somatostatin receptor scintigraphy [177]. Tumor stabilization was observed in 81% of patients in the group that received treatment compared

with 47% in the control group. However, neither tumor shrinkage nor visual field and pituitary function improvement was observed in any patient after treatment with octreotide LAR.

Temozolomide (TMZ) is an alkylating agent currently approved for the treatment of brain gliomas and glioblastomas. Recently, the European Society of Endocrinology recommended the use of TMZ as first-line treatment of aggressive pituitary tumors and pituitary carcinomas [143]. The response rate has been observed to be approximately 42%, but tumor recurrence is common after cessation of treatment and 2-year PFS has been calculated to 47.7% [178–180]. In addition, there are some reports of disease stability achieved in patients with atypical adenomas or carcinoma treated with peptide receptor radionuclide therapy (PRRT) [181].

9.9 Quality of Life (QoL) and Mortality

Patients suffering from NFPA present morbidities associated with the tumor or with the treatment modalities offered. Multiple studies have observed that patients with NFPA display a mortality rate higher than the general population [182]. A series of 573 patients with NFPA reported a standardized mortality ratio (SMR) of 1.7 with the excess mortality attributed to respiratory and cardio/cerebrovascular deaths [183]. Another study of 2795 patients reported an SMR of 1.1, and mortality was associated with infectious and circulatory diseases [184]. Older age at diagnosis and high doses of glucocorticoid substitution therapy were identified as predictive factors of mortality [182].

Although it has been reported that following treatment for NFPA QoL displays significant improvement, the findings regarding QoL normalization remain inconclusive [185]. Multiple factors, including visual impairment, hypopituitarism, and the type of surgical technique, are associated with impaired QoL. Disease-specific questionnaires should be introduced in order to evaluate effectively and optimize QoL.

9.10 Conclusion

NFPAs are prevalent PitNETs that come to clinical attention either through the presence of compressive symptoms to surrounding tissue and/or anterior hormonal deficiencies or are discovered incidentally (Fig. 9.3). Although a number of genetic effects related to their pathogenesis have been identified their molecular pathogenesis has not been delineated as yet, the majority of these tumors follow a relatively indolent course; however, a subset may become more aggressive with early recurrences. Currently, existing biomarkers can help identify specific subtypes and predict tumoral behavior. Surgery remains the main treatment in the presence of symptoms related to compressive effects, whereas surveillance remains an option for incidentally discovered tumors. Radiotherapy is used for potentially aggressive and recurrent tumors and when needed systemic treatment with temozolomide may

(F Hyperprolactinemia Surgery (Pituitary stalk compression) Diplopia E 6 Local tumor expansion Radiotherap Visual defect (mass effect) Non-functioning Headache Pituitary aden Hypopituitarism Systemic Treatment GH, TSH, ACTH deficie ?

Fig. 9.3 Overview of clinical manifestations and management of nonfunctioning pituitary adenomas

be administered. Further studies are needed to help delineate their pathogenesis and provide specific tumor-directed treatment.

References

- Stratakis CA, Matyakhina L, Courkoutsakis N, Patronas N, Voutetakis A, Stergiopoulos S, et al. Pathology and molecular genetics of the pituitary gland in patients with the 'complex of spotty skin pigmentation, myxomas, endocrine overactivity and schwannomas' (Carney complex). Front Horm Res. 2004;32:253–64.
- Chatzellis E, Alexandraki KI, Androulakis II, Kaltsas G. Aggressive pituitary tumors. Neuroendocrinology. 2015;101(2):87–104.
- Aulinas A, Colom C, Ybarra J, Munoz F, Tresserras P, Resmini E, et al. Immediate and delayed postoperative morbidity in functional and non-functioning pituitary adenomas. Pituitary. 2012;15(3):380–5.
- 4. Melmed S. Pathogenesis of pituitary tumors. Nat Rev Endocrinol. 2011;7(5):257-66.
- Fernandez A, Karavitaki N, Wass JA. Prevalence of pituitary adenomas: a community-based, cross-sectional study in Banbury (Oxfordshire, UK). Clin Endocrinol. 2010;72(3):377–82.
- 6. Beckers A. Higher prevalence of clinically relevant pituitary adenomas confirmed. Clin Endocrinol. 2010;72(3):290–1.
- Gruppetta M, Mercieca C, Vassallo J. Prevalence and incidence of pituitary adenomas: a population based study in Malta. Pituitary. 2013;16(4):545–53.
- Tjornstrand A, Gunnarsson K, Evert M, Holmberg E, Ragnarsson O, Rosen T, et al. The incidence rate of pituitary adenomas in western Sweden for the period 2001–2011. Eur J Endocrinol. 2014;171(4):519–26.
- Drange MR, Fram NR, Herman-Bonert V, Melmed S. Pituitary tumor registry: a novel clinical resource. J Clin Endocrinol Metab. 2000;85(1):168–74.
- Robenshtok E, Benbassat CA, Hirsch D, Tzvetov G, Cohen ZR, Iraqi HM, et al. Clinical course and outcome of nonfunctioning pituitary adenomas in the elderly compared with younger age groups. Endocr Pract. 2014;20(2):159–64.
- Verges B, Boureille F, Goudet P, Murat A, Beckers A, Sassolas G, et al. Pituitary disease in MEN type 1 (MEN1): data from the France-Belgium MEN1 multicenter study. J Clin Endocrinol Metab. 2002;87(2):457–65.

- de Laat JM, Dekkers OM, Pieterman CR, Kluijfhout WP, Hermus AR, Pereira AM, et al. Long-term natural course of pituitary tumors in patients with MEN1: results from the DutchMEN1 study group (DMSG). J Clin Endocrinol Metab. 2015;100(9):3288–96.
- Daly AF, Jaffrain-Rea ML, Ciccarelli A, Valdes-Socin H, Rohmer V, Tamburrano G, et al. Clinical characterization of familial isolated pituitary adenomas. J Clin Endocrinol Metab. 2006;91(9):3316–23.
- Ntali G, Wass JA. Epidemiology, clinical presentation and diagnosis of non-functioning pituitary adenomas. Pituitary. 2018;21(2):111–8.
- 15. Stergiopoulos SG, Abu-Asab MS, Tsokos M, Stratakis CA. Pituitary pathology in carney complex patients. Pituitary. 2004;7(2):73–82.
- Correa R, Salpea P, Stratakis CA. Carney complex: an update. Eur J Endocrinol. 2015;173(4):M85–97.
- Mete O, Lopes MB. Overview of the 2017 WHO classification of pituitary tumors. Endocr Pathol. 2017;28(3):228–43.
- Lopes MBS. The 2017 World Health Organization classification of tumors of the pituitary gland: a summary. Acta Neuropathol. 2017;134(4):521–35.
- Asa SL, Puy LA, Lew AM, Sundmark VC, Elsholtz HP. Cell type-specific expression of the pituitary transcription activator pit-1 in the human pituitary and pituitary adenomas. J Clin Endocrinol Metab. 1993;77(5):1275–80.
- Asa SL, Bamberger AM, Cao B, Wong M, Parker KL, Ezzat S. The transcription activator steroidogenic factor-1 is preferentially expressed in the human pituitary gonadotroph. J Clin Endocrinol Metab. 1996;81(6):2165–70.
- Lamolet B, Pulichino AM, Lamonerie T, Gauthier Y, Brue T, Enjalbert A, et al. A pituitary cell-restricted T box factor, Tpit, activates POMC transcription in cooperation with Pitx homeoproteins. Cell. 2001;104(6):849–59.
- Pulichino AM, Vallette-Kasic S, Tsai JP, Couture C, Gauthier Y, Drouin J. Tpit determines alternate fates during pituitary cell differentiation. Genes Dev. 2003;17(6):738–47.
- Scully KM, Rosenfeld MG. Pituitary development: regulatory codes in mammalian organogenesis. Science. 2002;295(5563):2231–5.
- 24. Delgrange E, Vasiljevic A, Wierinckx A, Francois P, Jouanneau E, Raverot G, et al. Expression of estrogen receptor alpha is associated with prolactin pituitary tumor prognosis and supports the sex-related difference in tumor growth. Eur J Endocrinol. 2015;172(6):791–801.
- Friend KE, Chiou YK, Lopes MB, Laws ER Jr, Hughes KM, Shupnik MA. Estrogen receptor expression in human pituitary: correlation with immunohistochemistry in normal tissue, and immunohistochemistry and morphology in macroadenomas. J Clin Endocrinol Metab. 1994;78(6):1497–504.
- Manojlovic-Gacic E, Engstrom BE, Casar-Borota O. Histopathological classification of nonfunctioning pituitary neuroendocrine tumors. Pituitary. 2018;21(2):119–29.
- Mayson SE, Snyder PJ. Silent pituitary adenomas. Endocrinol Metab Clin N Am. 2015;44(1):79–87.
- Korbonits M, Carlsen E. Recent clinical and pathophysiological advances in non-functioning pituitary adenomas. Horm Res. 2009;71(Suppl 2):123–30.
- Saeger W, Ludecke DK, Buchfelder M, Fahlbusch R, Quabbe HJ, Petersenn S. Pathohistological classification of pituitary tumors: 10 years of experience with the German pituitary tumor registry. Eur J Endocrinol. 2007;156(2):203–16.
- Yamada S, Ohyama K, Taguchi M, Takeshita A, Morita K, Takano K, et al. A study of the correlation between morphological findings and biological activities in clinically nonfunctioning pituitary adenomas. Neurosurgery. 2007;61(3):580–4. discussion 4–5
- McDonald WC, Banerji N, McDonald KN, Ho B, Macias V, Kajdacsy-Balla A. Steroidogenic Factor 1, Pit-1, and adrenocorticotropic hormone: a rational starting place for the Immunohistochemical characterization of pituitary adenoma. Arch Pathol Lab Med. 2017;141(1):104–12.

- Balogun JA, Monsalves E, Juraschka K, Parvez K, Kucharczyk W, Mete O, et al. Null cell adenomas of the pituitary gland: an institutional review of their clinical imaging and behavioral characteristics. Endocr Pathol. 2015;26(1):63–70.
- 33. Oystese KA, Casar-Borota O, Normann KR, Zucknick M, Berg JP, Bollerslev J. Estrogen receptor alpha, a sex-dependent predictor of aggressiveness in nonfunctioning pituitary adenomas: SSTR and sex hormone receptor distribution in NFPA. J Clin Endocrinol Metab. 2017;102(9):3581–90.
- 34. Thodou E, Argyrakos T, Kontogeorgos G. Galectin-3 as a marker distinguishing functioning from silent corticotroph adenomas. Hormones (Athens). 2007;6(3):227–32.
- Ben-Shlomo A, Cooper O. Role of tyrosine kinase inhibitors in the treatment of pituitary tumours: from bench to bedside. Curr Opin Endocrinol Diabetes Obes. 2017;24(4):301–5.
- 36. Sjostedt E, Bollerslev J, Mulder J, Lindskog C, Ponten F, Casar-Borota O. A specific antibody to detect transcription factor T-pit: a reliable marker of corticotroph cell differentiation and a tool to improve the classification of pituitary neuroendocrine tumours. Acta Neuropathol. 2017;134(4):675–7.
- Horvath E, Kovacs K, Killinger DW, Smyth HS, Platts ME, Singer W. Silent corticotropic adenomas of the human pituitary gland: a histologic, immunocytologic, and ultrastructural study. Am J Pathol. 1980;98(3):617–38.
- Raverot G, Wierinckx A, Jouanneau E, Auger C, Borson-Chazot F, Lachuer J, et al. Clinical, hormonal and molecular characterization of pituitary ACTH adenomas without (silent corticotroph adenomas) and with Cushing's disease. Eur J Endocrinol. 2010;163(1):35–43.
- Righi A, Faustini-Fustini M, Morandi L, Monti V, Asioli S, Mazzatenta D, et al. The changing faces of corticotroph cell adenomas: the role of prohormone convertase 1/3. Endocrine. 2017;56(2):286–97.
- 40. Ben-Shlomo A, Cooper O. Silent corticotroph adenomas. Pituitary. 2018;21(2):183-93.
- Kim D, Ku CR, Park SH, Moon JH, Kim EH, Kim SH, et al. Clinical parameters to distinguish silent Corticotroph adenomas from other nonfunctioning pituitary adenomas. World Neurosurg. 2018;115:e464–e71.
- 42. Stefaneanu L, Kovacs K, Horvath E, Lloyd RV. In situ hybridization study of proopiomelanocortin (POMC) gene expression in human pituitary corticotrophs and their adenomas. Virchows Arch A Pathol Anat Histopathol. 1991;419(2):107–13.
- Scheithauer BW, Jaap AJ, Horvath E, Kovacs K, Lloyd RV, Meyer FB, et al. Clinically silent corticotroph tumors of the pituitary gland. Neurosurgery. 2000;47(3):723–9. discussion 9–30
- 44. Ioachimescu AG, Eiland L, Chhabra VS, Mastrogianakis GM, Schniederjan MJ, Brat D, et al. Silent corticotroph adenomas: Emory University cohort and comparison with ACTH-negative nonfunctioning pituitary adenomas. Neurosurgery. 2012;71(2):296–303. discussion 4
- 45. McCormack A, Dekkers OM, Petersenn S, Popovic V, Trouillas J, Raverot G, et al. Treatment of aggressive pituitary tumours and carcinomas: results of a European Society of Endocrinology (ESE) survey 2016. Eur J Endocrinol. 2018;178(3):265–76.
- 46. Fountas A, Lavrentaki A, Subramanian A, Toulis KA, Nirantharakumar K, Karavitaki N. Recurrence in silent corticotroph adenomas after primary treatment: a systematic review and meta-analysis. J Clin Endocrinol Metab. 2018;
- 47. Manoranjan B, Salehi F, Scheithauer BW, Rotondo F, Kovacs K, Cusimano MD. Estrogen receptors alpha and beta immunohistochemical expression: clinicopathological correlations in pituitary adenomas. Anticancer Res. 2010;30(7):2897–904.
- Webb KM, Laurent JJ, Okonkwo DO, Lopes MB, Vance ML, Laws ER Jr. Clinical characteristics of silent corticotrophic adenomas and creation of an internet-accessible database to facilitate their multi-institutional study. Neurosurgery. 2003;53(5):1076–84. discussion 84–5
- 49. Bradley KJ, Wass JA, Turner HE. Non-functioning pituitary adenomas with positive immunoreactivity for ACTH behave more aggressively than ACTH immunonegative tumours but do not recur more frequently. Clin Endocrinol. 2003;58(1):59–64.
- Kovacs K, Horvath E, Bayley TA, Hassaram ST, Ezrin C. Silent corticotroph cell adenoma with lysosomal accumulation and crinophagy. A distinct clinicopathologic entity. Am J Med. 1978;64(3):492–9.

- Tateno T, Izumiyama H, Doi M, Yoshimoto T, Shichiri M, Inoshita N, et al. Differential gene expression in ACTH -secreting and non-functioning pituitary tumors. Eur J Endocrinol. 2007;157(6):717–24.
- Tateno T, Izumiyama H, Doi M, Akashi T, Ohno K, Hirata Y. Defective expression of prohormone convertase 1/3 in silent corticotroph adenoma. Endocr J. 2007;54(5):777–82.
- Roncaroli F, Scheithauer BW, Young WF, Horvath E, Kovacs K, Kros JM, et al. Silent corticotroph carcinoma of the adenohypophysis: a report of five cases. Am J Surg Pathol. 2003;27(4):477–86.
- He L, Forbes JA, Carr K, Highfield Nickols H, Utz A, Moots P, et al. Response of silent corticotroph pituitary carcinoma to chemotherapy: case report. J Neurosurg Sci. 2016;60(2):272–80.
- 55. Chinezu L, Vasiljevic A, Trouillas J, Lapoirie M, Jouanneau E, Raverot G. Silent somatotroph tumour revisited from a study of 80 patients with and without acromegaly and a review of the literature. Eur J Endocrinol. 2017;176(2):195–201.
- 56. Langlois F, Woltjer R, Cetas JS, Fleseriu M. Silent somatotroph pituitary adenomas: an update. Pituitary. 2018;21(2):194–202.
- 57. Kovacs K, Lloyd R, Horvath E, Asa SL, Stefaneanu L, Killinger DW, et al. Silent somatotroph adenomas of the human pituitary. A morphologic study of three cases including immunocytochemistry, electron microscopy, in vitro examination, and in situ hybridization. Am J Pathol. 1989;134(2):345–53.
- Sidhaye A, Burger P, Rigamonti D, Salvatori R. Giant somatotrophinoma without acromegalic features: more "quiet" than "silent": case report. Neurosurgery. 2005;56(5):E1154. discussion E
- Wade AN, Baccon J, Grady MS, Judy KD, O'Rourke DM, Snyder PJ. Clinically silent somatotroph adenomas are common. Eur J Endocrinol. 2011;165(1):39–44.
- 60. Sakharova AA, Dimaraki EV, Chandler WF, Barkan AL. Clinically silent somatotropinomas may be biochemically active. J Clin Endocrinol Metab. 2005;90(4):2117–21.
- Pagesy P, Li JY, Kujas M, Peillon F, Delalande O, Visot A, et al. Apparently silent somatotroph adenomas. Pathol Res Pract. 1991;187(8):950–6.
- 62. Langlois F, Lim DST, Varlamov E, Yedinak CG, Cetas JS, McCartney S, et al. Clinical profile of silent growth hormone pituitary adenomas; higher recurrence rate compared to silent gonadotroph pituitary tumors, a large single center experience. Endocrine. 2017;58(3):528–34.
- 63. Daems T, Verhelst J, Michotte A, Abrams P, De Ridder D, Abs R. Modification of hormonal secretion in clinically silent pituitary adenomas. Pituitary. 2009;12(1):80–6.
- 64. Bengtsson D, Schroder HD, Andersen M, Maiter D, Berinder K, Feldt Rasmussen U, et al. Long-term outcome and MGMT as a predictive marker in 24 patients with atypical pituitary adenomas and pituitary carcinomas given treatment with temozolomide. J Clin Endocrinol Metab. 2015;100(4):1689–98.
- Naritaka H, Kameya T, Sato Y, Furuhata S, Otani M, Kawase T. Morphological characterization and subtyping of silent somatotroph adenomas. Pituitary. 1999;1(3–4):233–41.
- 66. Kobayashi I, Oka H, Naritaka H, Sato Y, Fujii K, Kameya T. Expression of Pit-1 and growth hormone-releasing hormone receptor mRNA in human pituitary adenomas: difference among functioning, silent, and other nonfunctioning adenomas. Endocr Pathol. 2002;13(2):83–98.
- 67. Trouillas J, Sassolas G, Loras B, Velkeniers B, Raccurt M, Chotard L, et al. Somatotropic adenomas without acromegaly. Pathol Res Pract. 1991;187(8):943–9.
- Villa C, Lagonigro MS, Magri F, Koziak M, Jaffrain-Rea ML, Brauner R, et al. Hyperplasiaadenoma sequence in pituitary tumorigenesis related to aryl hydrocarbon receptor interacting protein gene mutation. Endocr Relat Cancer. 2011;18(3):347–56.
- Wang EL, Qian ZR, Yamada S, Rahman MM, Inosita N, Kageji T, et al. Clinicopathological characterization of TSH-producing adenomas: special reference to TSH-immunoreactive but clinically non-functioning adenomas. Endocr Pathol. 2009;20(4):209–20.
- Kirkman MA, Jaunmuktane Z, Brandner S, Khan AA, Powell M, Baldeweg SE. Active and silent thyroid-stimulating hormone-expressing pituitary adenomas: presenting symptoms, treatment, outcomes, and recurrence. World Neurosurg. 2014;82(6):1224–31.

- 71. Karavitaki N, Thanabalasingham G, Shore HC, Trifanescu R, Ansorge O, Meston N, et al. Do the limits of serum prolactin in disconnection hyperprolactinaemia need re-definition? A study of 226 patients with histologically verified non-functioning pituitary macroadenoma. Clin Endocrinol. 2006;65(4):524–9.
- 72. Casar-Borota O, Botling J, Granberg D, Stigare J, Wikstrom J, Boldt HB, et al. Serotonin, ATRX, and DAXX expression in pituitary adenomas: markers in the differential diagnosis of neuroendocrine tumors of the Sellar region. Am J Surg Pathol. 2017;41(9):1238–46.
- Nishioka H, Inoshita N, Mete O, Asa SL, Hayashi K, Takeshita A, et al. The complementary role of transcription factors in the accurate diagnosis of clinically nonfunctioning pituitary adenomas. Endocr Pathol. 2015;26(4):349–55.
- 74. Erickson D, Scheithauer B, Atkinson J, Horvath E, Kovacs K, Lloyd RV, et al. Silent subtype 3 pituitary adenoma: a clinicopathologic analysis of the Mayo Clinic experience. Clin Endocrinol. 2009;71(1):92–9.
- Horvath E, Kovacs K, Smyth HS, Killinger DW, Scheithauer BW, Randall R, et al. A novel type of pituitary adenoma: morphological features and clinical correlations. J Clin Endocrinol Metab. 1988;66(6):1111–8.
- Horvath E, Kovacs K, Smyth HS, Cusimano M, Singer W. Silent adenoma subtype 3 of the pituitary—immunohistochemical and ultrastructural classification: a review of 29 cases. Ultrastruct Pathol. 2005;29(6):511–24.
- 77. Mete O, Gomez-Hernandez K, Kucharczyk W, Ridout R, Zadeh G, Gentili F, et al. Silent subtype 3 pituitary adenomas are not always silent and represent poorly differentiated monomorphous plurihormonal Pit-1 lineage adenomas. Mod Pathol. 2016;29(2):131–42.
- Farrell WE, Coll AP, Clayton RN, Harris PE. Corticotroph carcinoma presenting as a silent corticotroph adenoma. Pituitary. 2003;6(1):41–7.
- 79. Kaltsas GA, Nomikos P, Kontogeorgos G, Buchfelder M, Grossman AB. Clinical review: diagnosis and management of pituitary carcinomas. J Clin Endocrinol Metab. 2005;90(5):3089–99.
- Aflorei ED, Korbonits M. Epidemiology and etiopathogenesis of pituitary adenomas. J Neuro-Oncol. 2014;117(3):379–94.
- Jiang X, Zhang X. The molecular pathogenesis of pituitary adenomas: an update. Endocrinol Metab (Seoul). 2013;28(4):245–54.
- 82. Foltran RK, Amorim P, Duarte FH, Grande IPP, Freire A, Frassetto FP, et al. Study of major genetic factors involved in pituitary tumorigenesis and their impact on clinical and biological characteristics of sporadic somatotropinomas and non-functioning pituitary adenomas. Braz J Med Biol Res. 2018;51(9):e7427.
- Falchetti A. Genetics of multiple endocrine neoplasia type 1 syndrome: what's new and what's old. F1000Res. 2017;6
- Thakker RV, Newey PJ, Walls GV, Bilezikian J, Dralle H, Ebeling PR, et al. Clinical practice guidelines for multiple endocrine neoplasia type 1 (MEN1). J Clin Endocrinol Metab. 2012;97(9):2990–3011.
- Corbetta S, Pizzocaro A, Peracchi M, Beck-Peccoz P, Faglia G, Spada A. Multiple endocrine neoplasia type 1 in patients with recognized pituitary tumours of different types. Clin Endocrinol. 1997;47(5):507–12.
- Alrezk R, Hannah-Shmouni F, Stratakis CA. MEN4 and CDKN1B mutations: the latest of the MEN syndromes. Endocr Relat Cancer. 2017;24(10):T195–208.
- Kaltsas GA, Kola B, Borboli N, Morris DG, Gueorguiev M, Swords FM, et al. Sequence analysis of the PRKAR1A gene in sporadic somatotroph and other pituitary tumours. Clin Endocrinol. 2002;57(4):443–8.
- Beckers A, Aaltonen LA, Daly AF, Karhu A. Familial isolated pituitary adenomas (FIPA) and the pituitary adenoma predisposition due to mutations in the aryl hydrocarbon receptor interacting protein (AIP) gene. Endocr Rev. 2013;34(2):239–77.
- Daly AF, Beckers A. The role of AIP mutations in pituitary adenomas: 10 years on. Endocrine. 2017;55(2):333–5.

- Araujo PB, Kasuki L, de Azeredo Lima CH, Ogino L, Camacho AHS, Chimelli L, et al. AIP mutations in Brazilian patients with sporadic pituitary adenomas: a single-center evaluation. Endocr Connect. 2017;6(8):914–25.
- Hu Y, Yang J, Chang Y, Ma S, Qi J. SNPs in the aryl hydrocarbon receptor-interacting protein gene associated with sporadic non-functioning pituitary adenoma. Exp Ther Med. 2016;11(3):1142–6.
- Dwight T, Mann K, Benn DE, Robinson BG, McKelvie P, Gill AJ, et al. Familial SDHA mutation associated with pituitary adenoma and pheochromocytoma/paraganglioma. J Clin Endocrinol Metab. 2013;98(6):E1103–8.
- Simpson DJ, Bicknell JE, McNicol AM, Clayton RN, Farrell WE. Hypermethylation of the p16/CDKN2A/MTSI gene and loss of protein expression is associated with nonfunctional pituitary adenomas but not somatotrophinomas. Genes Chromosomes Cancer. 1999;24(4):328–36.
- 94. Zhang X, Sun H, Danila DC, Johnson SR, Zhou Y, Swearingen B, et al. Loss of expression of GADD45 gamma, a growth inhibitory gene, in human pituitary adenomas: implications for tumorigenesis. J Clin Endocrinol Metab. 2002;87(3):1262–7.
- Zhang X, Zhou Y, Mehta KR, Danila DC, Scolavino S, Johnson SR, et al. A pituitary-derived MEG3 isoform functions as a growth suppressor in tumor cells. J Clin Endocrinol Metab. 2003;88(11):5119–26.
- 96. Lin Y, Jiang X, Shen Y, Li M, Ma H, Xing M, et al. Frequent mutations and amplifications of the PIK3CA gene in pituitary tumors. Endocr Relat Cancer. 2009;16(1):301–10.
- 97. Esteller M. Cancer epigenetics for the 21st century: what's next? Genes Cancer. 2011;2(6):604–6.
- Salomon MP, Wang X, Marzese DM, Hsu SC, Nelson N, Zhang X, et al. The epigenomic landscape of pituitary adenomas reveals specific alterations and differentiates among acromegaly, Cushing's disease and endocrine-inactive subtypes. Clin Cancer Res. 2018;24(17):4126–36.
- Nemeth K, Darvasi O, Liko I, Szucs N, Czirjak S, Reiniger L, et al. Comprehensive analysis of circulating microRNAs in plasma of patients with pituitary adenomas. J Clin Endocrinol Metab. 2019;
- Li XH, Wang EL, Zhou HM, Yoshimoto K, Qian ZR. MicroRNAs in human pituitary adenomas. Int J Endocrinol. 2014;2014:435171.
- 101. He Z, Chen L, Hu X, Tang J, He L, Hu J, et al. Next-generation sequencing of microRNAs reveals a unique expression pattern in different types of pituitary adenomas. Endocr J. 2019;
- 102. Feng Y, Mao ZG, Wang X, Du Q, Jian M, Zhu D, et al. MicroRNAs and target genes in pituitary adenomas. Horm Metab Res. 2018;50(3):179–92.
- 103. Wu S, Gu Y, Huang Y, Wong TC, Ding H, Liu T, et al. Novel biomarkers for non-functioning invasive pituitary adenomas were identified by using analysis of microRNAs expression profile. Biochem Genet. 2017;55(3):253–67.
- 104. Bottoni A, Zatelli MC, Ferracin M, Tagliati F, Piccin D, Vignali C, et al. Identification of differentially expressed microRNAs by microarray: a possible role for microRNA genes in pituitary adenomas. J Cell Physiol. 2007;210(2):370–7.
- 105. Lewis J. Notch signalling and the control of cell fate choices in vertebrates. Semin Cell Dev Biol. 1998;9(6):583–9.
- 106. Bray S. Notch signalling in drosophila: three ways to use a pathway. Semin Cell Dev Biol. 1998;9(6):591–7.
- 107. Moreno CS, Evans CO, Zhan X, Okor M, Desiderio DM, Oyesiku NM. Novel molecular signaling and classification of human clinically nonfunctional pituitary adenomas identified by gene expression profiling and proteomic analyses. Cancer Res. 2005;65(22):10214–22.
- Desiderio DM, Zhan X. The human pituitary proteome: the characterization of differentially expressed proteins in an adenoma compared to a control. Cell Mol Biol (Noisy-le-Grand). 2003;49(5):689–712.
- 109. Jordan S, Lidhar K, Korbonits M, Lowe DG, Grossman AB. Cyclin D and cyclin E expression in normal and adenomatous pituitary. Eur J Endocrinol. 2000;143(1):R1–6.

- 110. Yavropoulou MP, Yovos JG. The role of the Wnt signaling pathway in osteoblast commitment and differentiation. Hormones (Athens). 2007;6(4):279–94.
- 111. Tamai K, Semenov M, Kato Y, Spokony R, Liu C, Katsuyama Y, et al. LDL-receptor-related proteins in Wnt signal transduction. Nature. 2000;407(6803):530–5.
- 112. Liu C, Li Y, Semenov M, Han C, Baeg GH, Tan Y, et al. Control of beta-catenin phosphorylation/degradation by a dual-kinase mechanism. Cell. 2002;108(6):837–47.
- 113. Mould AW, Duncan R, Serewko-Auret M, Loffler KA, Biondi C, Gartside M, et al. Global expression profiling of sex cord stromal tumors from Men1 heterozygous mice identifies altered TGF-beta signaling, decreased Gata6 and increased Csf1r expression. Int J Cancer. 2009;124(5):1122–32.
- 114. Semba S, Han SY, Ikeda H, Horii A. Frequent nuclear accumulation of beta-catenin in pituitary adenoma. Cancer. 2001;91(1):42–8.
- 115. Formosa R, Gruppetta M, Falzon S, Santillo G, DeGaetano J, Xuereb-Anastasi A, et al. Expression and clinical significance of Wnt players and survivin in pituitary tumours. Endocr Pathol. 2012;23(2):123–31.
- 116. Rizzoli P, Iuliano S, Weizenbaum E, Laws E. Headache in patients with pituitary lesions: a longitudinal cohort study. Neurosurgery. 2016;78(3):316–23.
- Greenman Y, Melmed S. Diagnosis and management of nonfunctioning pituitary tumors. Annu Rev Med. 1996;47:95–106.
- 118. Losa M, Donofrio CA, Barzaghi R, Mortini P. Presentation and surgical results of incidentally discovered nonfunctioning pituitary adenomas: evidence for a better outcome independently of other patients' characteristics. Eur J Endocrinol. 2013;169(6):735–42.
- Cohen AR, Cooper PR, Kupersmith MJ, Flamm ES, Ransohoff J. Visual recovery after transsphenoidal removal of pituitary adenomas. Neurosurgery. 1985;17(3):446–52.
- 120. Abouaf L, Vighetto A, Lebas M. Neuro-ophthalmologic exploration in non-functioning pituitary adenoma. Ann Endocrinol. 2015;76(3):210–9.
- Jahangiri A, Lamborn KR, Blevins L, Kunwar S, Aghi MK. Factors associated with delay to pituitary adenoma diagnosis in patients with visual loss. J Neurosurg. 2012;116(2):283–9.
- 122. Ogra S, Nichols AD, Stylli S, Kaye AH, Savino PJ, Danesh-Meyer HV. Visual acuity and pattern of visual field loss at presentation in pituitary adenoma. J Clin Neurosci. 2014;21(5):735–40.
- 123. Wray SH. Neuro-ophthalmologic manifestations of pituitary and parasellar lesions. Clin Neurosurg. 1977;24:86–117.
- 124. Kim SH, Lee KC, Kim SH. Cranial nerve palsies accompanying pituitary tumour. J Clin Neurosci. 2007;14(12):1158–62.
- 125. Landeiro JA, Fonseca EO, Monnerat AL, Taboada GF, Cabral GA, Antunes F. Nonfunctioning giant pituitary adenomas: invasiveness and recurrence. Surg Neurol Int. 2015;6:179.
- 126. Cury ML, Fernandes JC, Machado HR, Elias LL, Moreira AC, Castro M. Non-functioning pituitary adenomas: clinical feature, laboratorial and imaging assessment, therapeutic management and outcome. Arq Bras Endocrinol Metabol. 2009;53(1):31–9.
- Chen L, White WL, Spetzler RF, Xu B. A prospective study of nonfunctioning pituitary adenomas: presentation, management, and clinical outcome. J Neuro-Oncol. 2011;102(1):129–38.
- 128. Colao A, Cerbone G, Cappabianca P, Ferone D, Alfieri A, Di Salle F, et al. Effect of surgery and radiotherapy on visual and endocrine function in nonfunctioning pituitary adenomas. J Endocrinol Investig. 1998;21(5):284–90.
- 129. Randeva HS, Schoebel J, Byrne J, Esiri M, Adams CB, Wass JA. Classical pituitary apoplexy: clinical features, management and outcome. Clin Endocrinol. 1999;51(2):181–8.
- Ayuk J, McGregor EJ, Mitchell RD, Gittoes NJ. Acute management of pituitary apoplexy surgery or conservative management? Clin Endocrinol. 2004;61(6):747–52.
- 131. Vargas G, Gonzalez B, Ramirez C, Ferreira A, Espinosa E, Mendoza V, et al. Clinical characteristics and treatment outcome of 485 patients with nonfunctioning pituitary macroadenomas. Int J Endocrinol. 2015;2015:756069.

- 132. Fernandez-Balsells MM, Murad MH, Barwise A, Gallegos-Orozco JF, Paul A, Lane MA, et al. Natural history of nonfunctioning pituitary adenomas and incidentalomas: a systematic review and metaanalysis. J Clin Endocrinol Metab. 2011;96(4):905–12.
- 133. Vasilev V, Rostomyan L, Daly AF, Potorac I, Zacharieva S, Bonneville JF, et al. Management of endocrine disease: pituitary 'incidentaloma': neuroradiological assessment and differential diagnosis. Eur J Endocrinol. 2016;175(4):R171–84.
- 134. Orija IB, Weil RJ, Hamrahian AH. Pituitary incidentaloma. Best Pract Res Clin Endocrinol Metab. 2012;26(1):47–68.
- 135. Bonneville JF. Magnetic resonance imaging of pituitary tumors. Front Horm Res. 2016;45:97–120.
- 136. Knosp E, Steiner E, Kitz K, Matula C. Pituitary adenomas with invasion of the cavernous sinus space: a magnetic resonance imaging classification compared with surgical findings. Neurosurgery. 1993;33(4):610–7. discussion 7–8
- 137. Fleseriu M, Bodach ME, Tumialan LM, Bonert V, Oyesiku NM, Patil CG, et al. Congress of neurological surgeons systematic review and evidence-based guideline for pretreatment endocrine evaluation of patients with nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E527–9.
- Freda PU, Beckers AM, Katznelson L, Molitch ME, Montori VM, Post KD, et al. Pituitary incidentaloma: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2011;96(4):894–904.
- 139. Newman SA, Turbin RE, Bodach ME, Tumialan LM, Oyesiku NM, Litvack Z, et al. Congress of neurological surgeons systematic review and evidence-based guideline on pretreatment ophthalmology evaluation in patients with suspected nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E530–2.
- Kaltsas GA, Evanson J, Chrisoulidou A, Grossman AB. The diagnosis and management of parasellar tumours of the pituitary. Endocr Relat Cancer. 2008;15(4):885–903.
- 141. Samejima N, Yamada S, Takada K, Sano T, Ozawa Y, Shimizu T, et al. Serum alpha-subunit levels in patients with pituitary adenomas. Clin Endocrinol. 2001;54(4):479–84.
- 142. Chanson P, Raverot G, Castinetti F, Cortet-Rudelli C, Galland F, Salenave S, et al. Management of clinically non-functioning pituitary adenoma. Ann Endocrinol. 2015;76(3):239–47.
- 143. Raverot G, Burman P, McCormack A, Heaney A, Petersenn S, Popovic V, et al. European Society of Endocrinology Clinical Practice Guidelines for the management of aggressive pituitary tumours and carcinomas. Eur J Endocrinol. 2018;178(1):G1–G24.
- 144. Melmed S, Casanueva FF, Hoffman AR, Kleinberg DL, Montori VM, Schlechte JA, et al. Diagnosis and treatment of hyperprolactinemia: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab. 2011;96(2):273–88.
- 145. Hong JW, Lee MK, Kim SH, Lee EJ. Discrimination of prolactinoma from hyperprolactinemic non-functioning adenoma. Endocrine. 2010;37(1):140–7.
- 146. Galland F, Vantyghem MC, Cazabat L, Boulin A, Cotton F, Bonneville JF, et al. Management of nonfunctioning pituitary incidentaloma. Ann Endocrinol. 2015;76(3):191–200.
- 147. Lucas JW, Bodach ME, Tumialan LM, Oyesiku NM, Patil CG, Litvack Z, et al. Congress of neurological surgeons systematic review and evidence-based guideline on primary management of patients with nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E533–5.
- 148. Coburger J, Konig R, Seitz K, Bazner U, Wirtz CR, Hlavac M. Determining the utility of intraoperative magnetic resonance imaging for transsphenoidal surgery: a retrospective study. J Neurosurg. 2014;120(2):346–56.
- 149. Tandon V, Raheja A, Suri A, Chandra PS, Kale SS, Kumar R, et al. Randomized trial for superiority of high field strength intra-operative magnetic resonance imaging guided resection in pituitary surgery. J Clin Neurosci. 2017;37:96–103.
- 150. Yu SY, Du Q, Yao SY, Zhang KN, Wang J, Zhu Z, et al. Outcomes of endoscopic and microscopic transsphenoidal surgery on non-functioning pituitary adenomas: a systematic review and meta-analysis. J Cell Mol Med. 2018;22(3):2023–7.

- 151. Nomikos P, Ladar C, Fahlbusch R, Buchfelder M. Impact of primary surgery on pituitary function in patients with non-functioning pituitary adenomas—a study on 721 patients. Acta Neurochir. 2004;146(1):27–35.
- 152. Murad MH, Fernandez-Balsells MM, Barwise A, Gallegos-Orozco JF, Paul A, Lane MA, et al. Outcomes of surgical treatment for nonfunctioning pituitary adenomas: a systematic review and meta-analysis. Clin Endocrinol. 2010;73(6):777–91.
- 153. Burke WT, Cote DJ, Iuliano SI, Zaidi HA, Laws ER. A practical method for prevention of readmission for symptomatic hyponatremia following transsphenoidal surgery. Pituitary. 2018;21(1):25–31.
- 154. Hannon MJ, Thompson CJ. Neurosurgical Hyponatremia. J Clin Med. 2014;3(4):1084–104.
- 155. Hensen J, Henig A, Fahlbusch R, Meyer M, Boehnert M, Buchfelder M. Prevalence, predictors and patterns of postoperative polyuria and hyponatraemia in the immediate course after transsphenoidal surgery for pituitary adenomas. Clin Endocrinol. 1999;50(4):431–9.
- Inder WJ, Hunt PJ. Glucocorticoid replacement in pituitary surgery: guidelines for perioperative assessment and management. J Clin Endocrinol Metab. 2002;87(6):2745–50.
- 157. Fleseriu M, Hashim IA, Karavitaki N, Melmed S, Murad MH, Salvatori R, et al. Hormonal replacement in hypopituitarism in adults: an Endocrine Society Clinical Practice Guideline. J Clin Endocrinol Metab. 2016;101(11):3888–921.
- 158. O'Sullivan EP, Woods C, Glynn N, Behan LA, Crowley R, O'Kelly P, et al. The natural history of surgically treated but radiotherapy-naive nonfunctioning pituitary adenomas. Clin Endocrinol. 2009;71(5):709–14.
- 159. O'Reilly MW, Reulen RC, Gupta S, Thompson CA, Dineen R, Goulden EL, et al. ACTH and gonadotropin deficiencies predict mortality in patients treated for nonfunctioning pituitary adenoma: long-term follow-up of 519 patients in two large European centres. Clin Endocrinol. 2016;85(5):748–56.
- 160. Reddy R, Cudlip S, Byrne JV, Karavitaki N, Wass JA. Can we ever stop imaging in surgically treated and radiotherapy-naive patients with non-functioning pituitary adenoma? Eur J Endocrinol. 2011;165(5):739–44.
- 161. Li X, Li Y, Cao Y, Li P, Liang B, Sun J, et al. Safety and efficacy of fractionated stereotactic radiotherapy and stereotactic radiosurgery for treatment of pituitary adenomas: a systematic review and meta-analysis. J Neurol Sci. 2017;372:110–6.
- 162. Minniti G, Clarke E, Scaringi C, Enrici RM. Stereotactic radiotherapy and radiosurgery for non-functioning and secreting pituitary adenomas. Rep Pract Oncol Radiother. 2016;21(4):370–8.
- 163. Gittoes NJ, Bates AS, Tse W, Bullivant B, Sheppard MC, Clayton RN, et al. Radiotherapy for non-function pituitary tumours. Clin Endocrinol. 1998;48(3):331–7.
- 164. Brada M, Rajan B, Traish D, Ashley S, Holmes-Sellors PJ, Nussey S, et al. The long-term efficacy of conservative surgery and radiotherapy in the control of pituitary adenomas. Clin Endocrinol. 1993;38(6):571–8.
- Chanson P, Dormoy A, Dekkers O. Use of radiotherapy after pituitary surgery for nonfunctioning pituitary adenomas. Eur J Endocrinol. 2019;
- 166. Chen Y, Wang CD, Su ZP, Chen YX, Cai L, Zhuge QC, et al. Natural history of postoperative nonfunctioning pituitary adenomas: a systematic review and meta-analysis. Neuroendocrinology. 2012;96(4):333–42.
- Littley MD, Shalet SM, Beardwell CG, Ahmed SR, Applegate G, Sutton ML. Hypopituitarism following external radiotherapy for pituitary tumours in adults. QJ Med. 1989;70(262):145–60.
- 168. Tooze A, Gittoes NJ, Jones CA, Toogood AA. Neurocognitive consequences of surgery and radiotherapy for tumours of the pituitary. Clin Endocrinol. 2009;70(4):503–11.
- 169. Minniti G, Traish D, Ashley S, Gonsalves A, Brada M. Risk of second brain tumor after conservative surgery and radiotherapy for pituitary adenoma: update after an additional 10 years. J Clin Endocrinol Metab. 2005;90(2):800–4.
- 170. Brada M, Burchell L, Ashley S, Traish D. The incidence of cerebrovascular accidents in patients with pituitary adenoma. Int J Radiat Oncol Biol Phys. 1999;45(3):693–8.

- 171. Ayuk J. Does pituitary radiotherapy increase the risk of stroke and, if so, what preventative actions should be taken? Clin Endocrinol. 2012;76(3):328–31.
- 172. Garcia EC, Naves LA, Silva AO, de Castro LF, Casulari LA, Azevedo MF. Short-term treatment with cabergoline can lead to tumor shrinkage in patients with nonfunctioning pituitary adenomas. Pituitary. 2013;16(2):189–94.
- 173. Colao A, Di Somma C, Pivonello R, Faggiano A, Lombardi G, Savastano S. Medical therapy for clinically non-functioning pituitary adenomas. Endocr Relat Cancer. 2008;15(4):905–15.
- 174. Taboada GF, Luque RM, Bastos W, Guimaraes RF, Marcondes JB, Chimelli LM, et al. Quantitative analysis of somatostatin receptor subtype (SSTR1-5) gene expression levels in somatotropinomas and non-functioning pituitary adenomas. Eur J Endocrinol. 2007;156(1):65–74.
- 175. Lee M, Lupp A, Mendoza N, Martin N, Beschorner R, Honegger J, et al. SSTR3 is a putative target for the medical treatment of gonadotroph adenomas of the pituitary. Endocr Relat Cancer. 2015;22(1):111–9.
- 176. Greenman Y, Cooper O, Yaish I, Robenshtok E, Sagiv N, Jonas-Kimchi T, et al. Treatment of clinically nonfunctioning pituitary adenomas with dopamine agonists. Eur J Endocrinol. 2016;175(1):63–72.
- 177. Fusco A, Giampietro A, Bianchi A, Cimino V, Lugli F, Piacentini S, et al. Treatment with octreotide LAR in clinically non-functioning pituitary adenoma: results from a case-control study. Pituitary. 2012;15(4):571–8.
- 178. Halevy C, Whitelaw BC. How effective is temozolomide for treating pituitary tumours and when should it be used? Pituitary. 2017;20(2):261–6.
- 179. Lasolle H, Cortet C, Castinetti F, Cloix L, Caron P, Delemer B, et al. Temozolomide treatment can improve overall survival in aggressive pituitary tumors and pituitary carcinomas. Eur J Endocrinol. 2017;176(6):769–77.
- 180. Losa M, Bogazzi F, Cannavo S, Ceccato F, Curto L, De Marinis L, et al. Temozolomide therapy in patients with aggressive pituitary adenomas or carcinomas. J Neuro-Oncol. 2016;126(3):519–25.
- 181. Maclean J, Aldridge M, Bomanji J, Short S, Fersht N. Peptide receptor radionuclide therapy for aggressive atypical pituitary adenoma/carcinoma: variable clinical response in preliminary evaluation. Pituitary. 2014;17(6):530–8.
- Tampourlou M, Fountas A, Ntali G, Karavitaki N. Mortality in patients with non-functioning pituitary adenoma. Pituitary. 2018;21(2):203–7.
- Tomlinson JW, Holden N, Hills RK, Wheatley K, Clayton RN, Bates AS, et al. Association between premature mortality and hypopituitarism. West Midlands Prospective Hypopituitary Study Group. Lancet. 2001;357(9254):425–31.
- 184. Olsson DS, Nilsson AG, Bryngelsson IL, Trimpou P, Johannsson G, Andersson E. Excess mortality in women and young adults with nonfunctioning pituitary adenoma: a Swedish nationwide study. J Clin Endocrinol Metab. 2015;100(7):2651–8.
- 185. Andela CD, Lobatto DJ, Pereira AM, van Furth WR, Biermasz NR. How non-functioning pituitary adenomas can affect health-related quality of life: a conceptual model and literature review. Pituitary. 2018;21(2):208–16.
- 186. Trivellin G, Butz H, Delhove J, Igreja S, Chahal HS, Zivkovic V, et al. MicroRNA miR-107 is overexpressed in pituitary adenomas and inhibits the expression of aryl hydrocarbon receptor-interacting protein in vitro. Am J Physiol Endocrinol Metab. 2012;303(6):E708–19.
- 187. Leone V, Langella C, D'Angelo D, Mussnich P, Wierinckx A, Terracciano L, et al. Mir-23b and miR-130b expression is downregulated in pituitary adenomas. Mol Cell Endocrinol. 2014;390(1–2):1–7.
- Butz H, Nemeth K, Czenke D, Liko I, Czirjak S, Zivkovic V, et al. Systematic investigation of expression of G2/M transition genes reveals CDC25 alteration in nonfunctioning pituitary adenomas. Pathol Oncol Res. 2017;23(3):633–41.
- 189. Butz H, Liko I, Czirjak S, Igaz P, Korbonits M, Racz K, et al. MicroRNA profile indicates downregulation of the TGFbeta pathway in sporadic non-functioning pituitary adenomas. Pituitary. 2011;14(2):112–24.

- 190. Mussnich P, Raverot G, Jaffrain-Rea ML, Fraggetta F, Wierinckx A, Trouillas J, et al. Downregulation of miR-410 targeting the cyclin B1 gene plays a role in pituitary gonadotroph tumors. Cell Cycle. 2015;14(16):2590–7.
- 191. Wang DS, Zhang HQ, Zhang B, Yuan ZB, Yu ZK, Yang T, et al. miR-133 inhibits pituitary tumor cell migration and invasion via down-regulating FOXC1 expression. Genet Mol Res. 2016;15:1.
- 192. Wei Z, Zhou C, Liu M, Yao Y, Sun J, Xiao J, et al. MicroRNA involvement in a metastatic non-functioning pituitary carcinoma. Pituitary. 2015;18(5):710–21.
- 193. Liang S, Chen L, Huang H, Zhi D. The experimental study of miRNA in pituitary adenomas. Turk Neurosurg. 2013;23(6):721–7.
- 194. Butz H, Liko I, Czirjak S, Igaz P, Khan MM, Zivkovic V, et al. Down-regulation of Wee1 kinase by a specific subset of microRNA in human sporadic pituitary adenomas. J Clin Endocrinol Metab. 2010;95(10):E181–91.
- 195. Zhen W, Qiu D, Zhiyong C, Xin W, Mengyao J, Dimin Z, et al. MicroRNA-524-5p functions as a tumor suppressor in a human pituitary tumor-derived cell line. Horm Metab Res. 2017;49(7):550–7.

Check for updates

Clinical Case 1

10

Ludovica F. S. Grasso, Renata S. Auriemma, Maria Cristina De Martino, Rosa Pirchio, Rosario Pivonello, and Annamaria Colao

10.1 Case Report

A 34-year-old woman with a history of oligo-amenorrhea, acne, and hirsutism by age 28 was admitted to the Neuroendocrinology Clinical Center in September 2014. At age 29, polycystic ovary syndrome (PCOS) was diagnosed and the patient started contraceptive pill for 2 years. At age 32, due to primary infertility, the patient conceived after ovarian stimulation, and she safely delivered a healthy baby. No complications were reported during pregnancy and delivery. The patient reported galactorrhea even though she stopped breastfeeding, and she had persistent amenorrhea after the birth of her child. The patient reported a 9.5 kg weight gain in the last year despite diet and exercise, and increased sweating for the last 6 months. She presented with gradually worsening headache in the last 6 months. The headaches generally occurred three times per week. The patient's past medical history included thyroid nodule and uterine leiomyomata diagnosed 4 years earlier.

Because of the persistent galactorrhea and amenorrhea, the patient underwent hormonal evaluation, reporting a mildly elevated prolactin (PRL) levels (68 ng/mL, normal range 5–25 ng/mL), low levels of luteinizing hormone (LH, 1.2 IU/L, normal range 2.4–13 IU/L), follicle-stimulating hormone (FSH, 2.3 IU/L, normal range 3.5–13 IU/L), and estradiol (16 pg/mL, normal range 20–240 pg/mL). The patient was subsequently referred to the Neuroendocrinology Clinical Center for further evaluation.

The chapter has been endorsed by **Prof. Marek Bolanowski**, marek.bolanowski@umed.wroc.pl, Department of Endocrinology, Diabetes and Isotope Therapy, Medical University Wroclaw, Wroclaw, Poland

L. F. S. Grasso \cdot R. S. Auriemma \cdot M. C. De Martino \cdot R. Pirchio \cdot R. Pivonello A. Colao (\boxtimes)

Dipartimento di Medicina Clinica e Chirurgia, Sezione di Endocrinologia, University Federico II of Naples, Naples, Italy e-mail: colao@unina.it

[©] Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_10

At clinical evaluation, abnormal findings included obesity (body mass index 30.2 kg/m²), acne, hirsutism (modified Ferriman Gallwey score 16/36), and neck and axillary acanthosis. She did not spontaneously complain about acral changes. However, after specific questions, she recognized that her shoe size had increased from size 37 to size 39 during the last 3 years.

She denied significant changes in facial appearance, new spaces between her teeth, snoring, fatigue, joint pain, and paresthesia.

10.2 Differential Diagnosis

Considering the main clinical presentation of the patient, including secondary amenorrhea, galactorrhea, and headache, the prolactinoma and clinically nonfunctioning pituitary adenoma should be considered in the differential diagnosis [1, 2]. Moreover, considering the PCOS-like phenotype associated with acral changes, insulin-mediated pseudoacromegaly should be excluded.

Prolactinomas are the most common of the hormone-secreting pituitary tumors [1] representing approximately 40% of all pituitary tumors, and occur most frequently among women aged 20–50 years [1, 3, 4]. In women of reproductive age, usually the most relevant clinical manifestations are oligomenorrhea or secondary amenorrhea, galactorrhea, and infertility, followed by decreased libido and weight gain [3, 4]. In clinical practice, macroprolactinomas are less common than microprolactinomas and occur more often in men than in women [3]. Mass effects cause headache, hypopituitarism, and visual field defects. Prolactinomas are the most frequent cause of PRL excess even if several other causes should be excluded before the diagnosis is made [4]. Besides prolactinomas, hyperprolactinemia can be associated with a variety of causes, such as pregnancy, hypothyroidism, PCOS, renal insufficiency, and PRL-stimulating drugs, which need to be considered in the differential diagnosis [5]. Since there are multiple causes of hyperprolactinemia, other than prolactinoma, a careful medical history, clinical examination, and measurement of serum thyrotropin and creatinine are required [3].

Clinically nonfunctioning adenomas account for 15–54% of pituitary adenomas [1]. Most patients present with symptoms of mass effect due to their large size (67% are macroadenomas), whereas some patients are completely asymptomatic, detecting the pituitary adenoma as incidental findings (incidentalomas) on magnetic resonance imaging (MRI) or computed tomography scans were performed for other reasons [6]. All patients with clinically nonfunctioning macroadenomas, symptomatic or incidental, should be evaluated for hypopituitarism [1]. Hyperprolactinemia may occur in these patients because of pituitary stalk dysfunction [3, 6].

Pseudoacromegaly is an extremely rare condition characteristic of some patients with physical features resembling acromegaly, usually affecting the face and extremities, without abnormalities in the GH/IGF-I axis [7]. Due to the rarity and variability of these conditions, its correct diagnosis can be challenging [7]. Insulinmediated pseudoacromegaly is characterized by severe insulin resistance, acanthosis nigricans, and acromegaloid features in the absence of GH and IGF-I excess [7, 8]. Insulin-mediated pseudoacromegaly is associated with a selective post-receptor insulin signaling defect in which the insulin metabolic actions are impaired, but its mitogenic actions are preserved [7, 9]. Therefore, the metabolic actions of insulin are reduced resulting in hyperinsulinemia, while insulin mitogenic actions are preserved leading to acromegaloid features [7]. Insulin and IGF-I exhibit affinity for each other's receptor, and thus, high insulin levels resulting from insulin resistance may act on the type 1 IGF receptor [8]. Genetic abnormalities in the insulin receptor result in hyperinsulinemia, leading to diabetes mellitus and often acromegaloid features, a condition first described in 1976 by Kahn et al. [10]. In particular, the clinical presentation of insulin-mediated pseudoacromegaly may include face coarsening, frontal bossing, macroglossia, separated teeth, prognathism, large ears, acral enlargement, reduced subcutaneous fat on arms and legs, weight gain, acanthosis nigricans, skin tags, acne, hirsutism, hyperhidrosis, oligo-amenorrhea, and PCOS [7, 8, 11]. Adenomatous colonic polyps and multinodular goiter have been reported [8]. Clinical features of these patients overlap with those of acromegaly, but GH suppression on OGTT and IGF-I levels is generally normal. Moreover, these patients usually have increased LH levels and hyperandrogenism [7, 8].

10.3 Diagnostic Aspects

At the clinical evaluation, mild modification of nose, cheekbones, and lips was observed comparing previous photographs of the patient in order to detect the features of acromegaly. The evaluation of the pituitary function showed increased random GH (10.4 ng/mL) and IGF-I (874 ng/mL, normal range 80–290) levels. The oral glucose tolerance test (OGTT) was performed to confirm the diagnosis of acromegaly, resulting in GH nadir of 7.3 ng/mL. Other anterior pituitary hormone tests confirmed secondary hypogonadism (LH, 1.5 IU/L, FSH, 2.4 IU/L, estradiol, 16 pg/mL) and mildly elevated PRL levels (71 ng/mL). The MRI of the hypothalamus–pituitary region with gadolinium showed the presence of an intrasellar pituitary macroadenoma in the left side (14.5 \times 13 mm), with a mild suprasellar extension (Fig. 10.1). Visual field testing was normal.



Fig. 10.1 Pituitary MRI images at baseline (coronal (a, left) and sagittal (b, right))

1	U	5
Parameters	Baseline	3 months after surgery
IGF-I (ng/mL)	874	592
GH nadir(ng/mL)	7.3	2.4
LH (IU/L)	1.5	4.6
FSH (IU/L)	2.5	6.1
Estradiol (pg/mL)	16	81
PRL (ng/mL)	71	13
Cortisol (ng/mL)	103	103
TSH (µU/mL)	2	2.4
Free T4 (ng/dL)	0.98	1.07

Table 10.1 Patient profile before and 3 months after neurosurgery

The assessment of acromegaly complications showed biochemical hyperandrogenism and confirmed the presence of an isoechoic thyroid nodule of 9 mm of maximal diameter on the thyroid ultrasound, uterine leiomyomata, and ovarian cysts on the pelvic ultrasound. In line with the standard diagnostic criteria, acromegaly due to a GH-secreting pituitary macroadenoma was diagnosed. Patient's profile at baseline is shown in Table 10.1.

10.4 Treatment

The patient underwent first-line transsphenoidal surgery of the pituitary macroadenoma in November 2014. Histology of the excised specimen revealed moderate amounts of sparsely granular eosinophilic cytoplasm. Immunohistochemistry was positive for GH and negative for other pituitary hormones. Ki 67 index was <2%.

Her 3 months postoperative evaluation revealed elevated IGF-I levels (592 ng/ ml, normal range 80–290) and GH nadir during OGTT was 2.6 ng/mL, indicating persistent disease. Secondary hypogonadism resolved after surgery, and restoration of normal menstrual cycle was reported by the patient 2 months after surgery. Patient's profile 3 months after surgery is shown in Table 10.1. Postoperative MRI of the pituitary revealed millimetric parasellar area (4 mm in maximum diameter) in the left side of the pituitary (Fig. 10.2). Therefore, octreotide LAR treatment was initiated postoperatively, at the dose of 30 mg every 4 weeks. It was then increased stepwise, up to 40 mg every 4 weeks. The lowest IGF-I achieved during LAR treatment was 375 ng/mL (Fig. 10.3), indicating partial resistance to first-generation somatostatin analogs (SSA). LAR therapy was stopped in October 2015. Due to persistently uncontrolled acromegaly, despite high dose of LAR, pegvisomant (PEG) monotherapy was started, considering the small pituitary residual tumor. The dose of PEG was up-titrated up to 20 mg/day, achieving a disease control. Within a few months of PEG therapy, there was a significant clinical improvement.



Fig. 10.2 Pituitary MRI images 3 months after surgery (coronal (a, left) and sagittal (b, right))



10.5 Follow-Up

Within a month of starting PEG 20 mg/day, her IGF-I levels dropped to the normal range, remaining within the normal range for age over the follow-up, although tumor size remained unchanged. PEG was well tolerated. Patient's symptoms have significantly improved, and the patient reported normal menstrual cycle. At age 38, the patient spontaneously conceived. PEG therapy was discontinued after pregnancy confirmation, and the patient was regularly followed during pregnancy. She safely delivered a healthy baby; no complications were reported during pregnancy and delivery. The patient breastfed for 3 months, and then, PEG therapy was resumed.

10.6 Learning Points

- Although acromegaly manifests with distinct physical characteristics, diagnosis of the disease in its early stages can be difficult due to its insidious nature, meaning that neither the patient and their families nor their physicians may notice these changes [12].
- Acromegaly features develop insidiously over decades, often resulting in a delay
 of 7–10 years or more in diagnosis after the estimated onset of symptoms [13,
 14]. Among clinical features, female reproductive disorders, including menstrual
 abnormalities, galactorrhea, and decreased libido, are commonly complained in
 acromegaly [13, 14]. Particularly, women with acromegaly often present with
 menstrual irregularity, mainly represented by oligo-amenorrhea, associated with
 anovulation and infertility [15, 16].
- A direct action of GH and IGF-I excess on the pituitary–gonadal axis and the tumor mass effect per se have been proposed as potential mechanisms responsible for the occurrence of hyperprolactinemia and for the impairment in gonadotrophin secretion, leading to ovarian dysfunction and infertility [16–19]. Moreover, IGF-I excess has been found associated with overt PCOS or to a PCOS-like phenotype in 50% of acromegalic women [20].
- Evidence from literature has shown menstrual disturbances to occur in 40–80% of acromegalic women [16]. This wide variation in prevalence has been attributed mainly to the insidious onset of the disease and to the delay in diagnosis [18]. However, an earlier diagnosis and the availability of a wide spectrum of effective treatments for acromegaly could positively impact on the female fertility outcome in acromegaly [21].
- Treatment of acromegaly aims to normalize GH and IGF-I levels, control tumor mass, and decrease the risk of developing systemic comorbidities, thereby reducing mortality [13, 22, 23].
- Transsphenoidal adenomectomy remains a cornerstone treatment for GHsecreting pituitary tumors and is the treatment of choice except in those patients with high surgical risk, who refuse surgery or who have invasive, unresectable tumors [22, 23]. In patients with persistent disease despite surgical resection of the adenoma, medical therapy is recommended and first-generation SSA are the first-line medical therapy in most patients with acromegaly [22, 23].
- According to Endocrine Society Clinical Practice Guidelines [22], the GH receptor antagonist PEG is indicated as a second-line or third-line therapy, mostly in patients in whom surgery has failed or in those who show a poor response to first-line SSA.
- Currently, no medication is officially approved and recommended for acromegaly during pregnancy [24, 25]; however, based on a case-by-case analysis of patients the use of medical treatment during pregnancy should be weighed upon the risk-to-benefit ratio, balancing the risk of tumor enlargement, acromegaly symptoms, and maternal/fetal complications [24, 25]. Safety of PEG during pregnancy is yet to be clarified, since only few case reports in the literature have documented uneventful pregnancies following treatment with PEG [26–28].

Questions and Answers

In pregnant women with acromegaly:

- (a) Acromegaly medical therapy should be discontinued and administered only for patients with macroadenomas.
- (b) Acromegaly medical therapy should be discontinued and administered only for tumor and headache control.
- (c) Acromegaly medical therapy with somatostatin analogs and pegvisomant should be discontinued and dopamine agonist should be administered for tumor and headache control.
- (d) The dosage of acromegaly medical therapy should be reduced in all patients.

The GH receptor antagonist (PEG) is indicated:

- (a) As third-line therapy, after surgery in patients who show a poor response to first-line SSA.
- (b) Only in patients with persistent disease after surgery.
- (c) As a second- or third-line therapy, mostly in patients in whom surgery has failed or in those who show a poor response to first-line SSA.
- (d) Only in patients resistant to first-line therapy with conventional SSA.

First-line surgery:

- (a) Is indicated in all patients with visible pituitary adenoma.
- (b) Is the treatment of choice in the majority of patients with acromegaly except in those patients with high surgical risk or who refuse surgery.
- (c) Is the treatment of choice in the majority of patients with acromegaly except in those patients with invasive and unresectable tumors.
- (d) **b** + c.

Among clinical features of acromegaly:

- (a) Female reproductive disorders are commonly complained in acromegaly women, including menstrual abnormalities, galactorrhea, and decreased libido.
- (b) Up to 80% of women with acromegaly present with menstrual irregularity and hyperprolactinemia.
- (c) Menstrual disturbances occur in all young acromegalic women.
- (d) PCOS-like phenotype has been reported in the majority of acromegalic women.
- 1) In women with acromegaly onset during the fertile period:
 - (a) Menstrual cycle disorders do not improve despite disease control achieved with treatment.
 - (b) The most frequent alteration of the menstrual cycle is polymenorrhea.

- (c) Menstrual irregularities are often associated with anovulation and infertility.
- (d) Hyperprolactinemia and galactorrhea rarely occur.
- 2) PCOS-like phenotype in women with acromegaly:
 - (a) Is rarely found at diagnosis in young women.
 - (b) Is determined by the direct effect on the ovary of the IGF-1 excess and indirectly by the insulin resistance and hyperinsulinism.
 - (c) Differs from classical PCOS due to the absence of clinical and biochemical hyperandrogenism.
 - (d) Seldom is associated with infertility in these patients.
- 3) Follow-up of acromegaly during pregnancy requires:
 - (a) Monthly IGF-1 evaluation.
 - (b) MRI during the second trimester.
 - (c) Monitoring of GH but not IGF-1.
 - (d) Assessments of the possible growth of the adenoma using the visual field.

References

- 1. Molitch ME. Diagnosis and treatment of pituitary adenomas: a review. JAMA. 2017;317(5):516–24.
- Freda PU, Beckers AM, Katznelson L, Molitch ME, Montori VM, Post KD, Vance ML. Endocrine Society. Pituitary incidentaloma: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2011;96(4):894–904.
- Melmed S, Casanueva FF, Hoffman AR, Kleinberg DL, Montori VM, Schlechte JA, Wass JA. Endocrine Society. Diagnosis and treatment of hyperprolactinemia: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab. 2011;96(2):273–88.
- 4. Colao A. Pituitary tumours: the prolactinoma. Best Pract Res Clin Endocrinol Metab. 2009;23(5):575–96.
- 5. Klibanski A. Clinical practice. Prolactinomas. N Engl J Med. 2010;362(13):1219-26.
- Huang W, Molitch ME. Management of nonfunctioning pituitary adenomas (NFAs): observation. Pituitary. 2018;21(2):162–7.
- 7. Marques P, Korbonits M. Pseudoacromegaly. Front Neuroendocrinol. 2019;52:113-43.
- Sam AH, Tan T, Meeran K. Insulin-mediated "pseudoacromegaly". Hormones (Athens). 2011;10(2):156–61.
- Yaqub A, Yaqub N. Insulin-mediated pseudoacromegaly: a case report and review of the literature. W V Med J. 2008;104(5):12–5.
- Kahn CR, Flier JS, Bar RS, Archer JA, Gorden P, Martin MM, Roth J. The syndromes of insulin resistance and acanthosis nigricans. Insulin-receptor disorders in man. N Engl J Med. 1976;294(14):739–45.
- Flier JS, Moller DE, Moses AC, O'Rahilly S, Chaiken RL, Grigorescu F, Elahi D, Kahn BB, Weinreb JE, Eastman R. Insulin-mediated pseudoacromegaly: clinical and biochemical characterization of a syndrome of selective insulin resistance. J Clin Endocrinol Metab. 1993;76(6):1533–41.
- 12. Abreu A, Tovar AP, Castellanos R, Valenzuela A, Giraldo CM, Pinedo AC, Guerrero DP, Barrera CA, Franco HI, Ribeiro-Oliveira A Jr, Vilar L, Jallad RS, Duarte FG, Gadelha M, Boguszewski CL, Abucham J, Naves LA, Musolino NR, de Faria ME, Rossato C, Bronstein MD. Challenges in the diagnosis and management of acromegaly: a focus on comorbidities. Pituitary. 2016;19(4):448–57.

- Colao A, Grasso LFS, Giustina A, Melmed S, Chanson P, Pereira AM, Pivonello R. Acromegaly. Nat Rev Dis Primers. 2019;5(1):20.
- Colao A, Ferone D, Marzullo P, Lombardi G. Systemic complications of acromegaly: epidemiology, pathogenesis, and management. Endocr Rev. 2004;25(1):102–52.
- Colao A, Pivonello R, Di Somma C, Tauchmanovà L, Savastano S, Lombardi G. Growth hormone excess with onset in adolescence: clinical appearance and long-term treatment outcome. Clin Endocrinol. 2007;66(5):714–22.
- Kaltsas GA, Mukherjee JJ, Jenkins PJ, Satta MA, Islam N, Monson JP, Besser GM, Grossman AB. Menstrual irregularity in women with acromegaly. J Clin Endocrinol Metab. 1999;84(8):2731–5.
- Katznelson L, Kleinberg D, Vance ML, Stavrou S, Pulaski KJ, Schoenfeld DA, Hayden DL, Wright ME, Woodburn CJ, Klibanski A. Hypogonadism in patients with acromegaly: data from the multi-centre acromegaly registry pilot study. Clin Endocrinol. 2001;54(2):183–8.
- Grynberg M, Salenave S, Young J, Chanson P. Female gonadal function before and after treatment of acromegaly. J Clin Endocrinol Metab. 2010;95(10):4518–25. Epub 2010 Jul 21
- 19. Colao A, Lombardi G. Growth-hormone and prolactin excess. Lancet. 1998;352(9138):1455-61.
- Kaltsas GA, Androulakis II, Tziveriotis K, Papadogias D, Tsikini A, Makras P, Dimitriou K, Stathopoulou A, Piaditis G. Polycystic ovaries and the polycystic ovary syndrome phenotype in women with active acromegaly. Clin Endocrinol. 2007;67(6):917–22.
- Cheng S, Grasso L, Martinez-Orozco JA, Al-Agha R, Pivonello R, Colao A, Ezzat S. Pregnancy in acromegaly: experience from two referral centers and systematic review of the literature. Clin Endocrinol. 2012;76(2):264–71.
- Katznelson L, Laws ER Jr, Melmed S, Molitch ME, Murad MH, Utz A, Wass JA. Endocrine Society. Acromegaly: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2014;99(11):3933–51.
- Melmed S, Bronstein MD, Chanson P, Klibanski A, Casanueva FF, Wass JAH, Strasburger CJ, Luger A, Clemmons DR, Giustina A. A consensus statement on acromegaly therapeutic outcomes. Nat Rev Endocrinol. 2018;14(9):552–61.
- 24. Pivonello R, De Martino MC, Auriemma RS, Alviggi C, Grasso LF, Cozzolino A, De Leo M, De Placido G, Colao A, Lombardi G. Pituitary tumors and pregnancy: the interplay between a pathologic condition and a physiologic status. J Endocrinol Investig. 2014;37(2):99–112.
- Huang W, Molitch ME. Pituitary tumors in pregnancy. Endocrinol Metab Clin N Am. 2019;48(3):569–81.
- Brian SR, Bidlingmaier M, Wajnrajch MP, Weinzimer SA, Inzucchi SE. Treatment of acromegaly with pegvisomant during pregnancy: maternal and fetal effects. J Clin Endocrinol Metab. 2007;92(9):3374–7.
- Qureshi A, Kalu E, Ramanathan G, Bano G, Croucher C, Panahloo A. IVF/ICSI in a woman with active acromegaly: successful outcome following treatment with pegvisomant. J Assist Reprod Genet. 2006;23(11–12):439–42.
- van der Lely AJ, Gomez R, Heissler JF, Åkerblad AC, Jönsson P, Camacho-Hübner C, Kołtowska-Häggström M. Pregnancy in acromegaly patients treated with pegvisomant. Endocrine. 2015;49(3):769–73.



Clinical Case 2

11

Luiz Eduardo Wildemberg and Monica Gadelha

11.1 Case Report

A previously healthy 15-year-old female patient started to present irregular menses, weight gain, and edema in lower limbs and abdomen 4 months earlier. Later on, she noticed red striae in abdomen, and upper and lower limbs, increase in body hair in face and breasts, acne, mood instability, and fatigue. Previous history was

L. E. Wildemberg

Neuroendocrinology Division, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

Neuropathology and Molecular Genetics Laboratory, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

M. Gadelha (🖂)

Neuroendocrinology Division, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

Neuropathology and Molecular Genetics Laboratory, Instituto Estadual do Cérebro Paulo Niemeyer, Rio de Janeiro, Brazil

Rua Professor Rodolpho Paulo Rocco, 255, 9° andar - Setor 9F, Centro de Pesquisa em Neuroendocrinologia, Ilha do Fundão, Rio de Janeiro, Brazil e-mail: mgadelha@hucff.ufrj.br

The chapter has been endorsed by **Prof. Thierry Brue**, thierry.brue@mail.ap-hm.fr, Service d'Endocrinologie-Diabète-Maladies Métaboliques, CHU de Marseille - Hôpital de la Conception, Marseille, France

Neuroendocrinology Research Center/Endocrinology Division – Medical School and Hospital Universitário Clementino Fraga Filho, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

Neuroendocrinology Research Center/Endocrinology Division – Medical School and Hospital Universitário Clementino Fraga Filho, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil

unremarkable except for an isolated event of elevated blood pressure (150×90 mmHg). She denied the use of any medication.

At physical examination, she had a depressed mood, acne grade 3 in face and back, moon face and plethora, buffalo hump, and hirsutism (Ferriman–Gallwey score 16). Her height was 1.65 m and weight was 97.3 kg, with a BMI of 35.7 kg/m². Blood pressure (160/90 mmHg) and heart rate (122 bpm) were elevated. She had abdominal purple striae of approximately 2 cm.

At basal blood test, she had low potassium levels (3.1 mmol/L, NR 3.5–4.5), slightly elevated glycated hemoglobin (5.9%, NR < 5.7%), and elevated cortisol (45.8 mcg/L), and the rest of pituitary function was normal. A low-dose dexamethasone suppression test showed unsuppressed cortisol levels (22.9 mcg/L), and the free urinary cortisol (FUC) and salivary cortisol were very elevated, 720 mcg/24 h (NR 10–100) and 6.57 mcg/dL (NR < 0.35), respectively. So, the diagnosis of Cushing's syndrome was confirmed, and ACTH levels were elevated (89.9 pg/mL, NR < 46), in favor of ACTH-dependent Cushing's syndrome.

Due to the very rapid evolution of the case and to facilitate the diagnostic procedures, the patient was hospitalized. But during this period, she attempted suicide by throwing herself from the second floor. Luckily, she had no major injuries from this fall, but a chest computed tomography (CT) performed to investigate a thoracic pain after this episode revealed a 2.6 cm nodule in her left lung (Fig. 11.1).

A sella turcica magnetic resonance imaging (MRI) revealed a 0.4 cm lesion at right side of the adenohypophysis (Fig. 11.2). So, she was a patient with an ACTH-dependent Cushing's syndrome with lung and pituitary small lesions. Therefore, a simultaneous bilateral inferior petrosal sinus sampling (BIPSS) was realized, which confirmed central origin of the Cushing's syndrome (Table 11.1).

The patient was then submitted to microscopic transsphenoidal surgery. During surgical procedure, it was noted a liquefied material that was aspirated. Histopathological examination of the resected sample showed normal pituitary with Crooke's hyaline changes.

Fig. 11.1 Thorax computed tomography showing a 2.6 cm lesion in the left lung (white arrow)




Fig. 11.2 Pituitary magnetic resonance imaging coronal view in T1-weighted after gadolinium (a) and T2-weighted (b) sequences. A small 4 mm lesion with no contrast enhancement can be seen in the right side of the adenohypophysis in contact with left carotid artery. The lesion cannot be visualized in the T2 sequence

Time	Right	Periphery	Left
-5 min	672	56	5
0	6,4	52	45
3 min	889	46	53
5 min	696	48	50
10 min	1188	60	47

Table 11.1 ACTH levels during simultaneous bilateral inferior petrosal sinus sampling

Blood samples were collected minus 5 min and immediately before the intravenous administration of 10 mcg of desmopressin, and 3, 5, and 10 min after it. Samples were collected from both right and left inferior petrosal sinuses and from a peripheric site. ACTH levels are expressed in pg/ mL. Before desmopressin administration, central/periphery ratio was 12 and after it was 19.8, which confirms the central origin of the hypercortisolism

After surgery, the patient presented signs of adrenal insufficiency and was put on glucocorticoid treatment initially with hydrocortisone and later with prednisone. The patient lost weight and her mood was significantly improved, acne and hirsutism regressed, and regular menses returned. The patient used prednisone for 6 months, and after its withdrawal, a cortisol post-1 mg dexamethasone suppression test was 0.8 mcg/L and salivary cortisol was 0.31 mcg/dL and 0.29 mcg/dL. So, she had clinical and biochemical evidence of disease remission. Lung nodule was considered to be cicatricial and follow-up images revealed no increase.

Two years after surgery, she started to gain weight and presented irritability with fast progression. Laboratory examinations revealed disease relapse (post-1 mg test 20 mcg/L and FUC 1020 mcg/24 h). No lesion was observed in the pituitary MRI. She was reoperated transnasally, but no lesion was identified during pituitary exploration. After surgery, there was no improvement in the clinical signs of hyper-cortisolism, as well as in the laboratory examinations.

Due to the seriousness of the disease and the previous suicide attempt, bilateral adrenalectomy was performed. She is now stable under gluco- and mineralocorticoid substitutive therapy. Last ACTH levels were 165 pg/mL, and there was no tumor increase detected by MRI.

11.2 Differential Diagnosis

The main hypercortisolism etiology is external glucocorticoid use, so the first thing while dealing with a patient with clinical signs of hypercortisolism is to exclude exogenous corticoid use [1]. It is important to stress out that several dermatologic creams have glucocorticoids in their composition, so the use of these drugs must be extensively questioned. Moreover, inhaled glucocorticoids may also present systemic absorption leading to hypercortisolism in selected patients. Exogenous Cushing's syndrome may present a rapid course of development, similar to the present case. However, the patient denied chronic use of any medication, oral or topical, which excluded the hypothesis of exogenous Cushing's syndrome.

Endogenous Cushing's syndrome may be caused by excessive cortisol secretion due to adrenal tumor or hyperplasia or to excessive ACTH production. These Cushing's syndromes are of neoplastic origin and they must be distinguished from non-neoplastic hypercortisolism (NNH), also called pseudo-Cushing's syndrome (Table 11.2) [2]. NNH is usually associated with mild clinical and laboratory signs of hypercortisolism, which was not the case of our patient, making this diagnosis unlikely. Nevertheless, this was a teenage female patient with signs of androgen excess and menstrual irregularity, so the diagnosis of polycystic ovary syndrome (PCOS) must be considered [3]. The rapid course of the disease and the fact that she had previously regular menses make PCOS diagnosis very improbable. Psychiatric disorders are another important cause of NNH, and our patient had a depressive behavior. However, that started at the same time of other signs of hypercortisolism instead of being a previous condition, which may exclude depression as the cause of the clinical picture and makes it a part of the clinical picture of hypercortisolism. Other causes such as excessive alcohol intake, anorexia nervosa, and intense exercise were really unlikely [4].

After confirming endogenous hypercortisolism, it is necessary to differentiate between ACTH-dependent or ACTH-independent Cushing's syndrome [5]. The ACTH-independent cases are of adrenal origin, either caused by a cortisol-secreting

Table 11.2	Etiologies of	non-neoplastic	hypercortisolism
-------------------	---------------	----------------	------------------

Depression and psychiatric disorders
Alcohol abuse
Hyperinsulinemia (obesity, type 2 diabetes mellitus, polycystic ovary syndrome)
Anorexia nervosa
Chronic kidney disease
Intense chronic exercise
Multiple sclerosis
Glucocorticoid resistance

adrenal adenoma or adrenal hyperplasia. ACTH-dependent cases are caused mostly by a pituitary adenoma (corticotropinoma), called Cushing's disease, or less frequently by an ACTH-secreting tumor located in organs other than the pituitary (ectopic Cushing's syndrome—ECS). This differential diagnosis will be further detailed in the section about diagnostic aspects.

11.3 Diagnostic Aspects

The diagnosis of Cushing's syndrome is a challenge, from the clinical presentation to laboratory and imaging examinations. The clinical picture may vary from a very mild and insidious presentation to a rich and/or fast evolution of symptoms [6]. Cushing's syndrome clinical picture includes several signs and symptoms with higher or lower specificity. The most specific signs are easy bruising, facial plethora, purple striae >1 cm, proximal muscular atrophy, and weight gain with growth retardation in children [1]. Other signs that are frequently found in patients with Cushing's syndrome but also may be encountered in other diseases are acne, hirsutism, peripheral edema, central obesity, "buffalo hump," and supraclavicular fullness. Some symptoms that are also—and still more—nonspecific include depression, fatigue, weight gain, menstrual abnormalities, decreased libido, cognitive deficits, and irritability [6]. Cushing's syndrome is also associated with a diversity of comorbidities: hypertension, glucose metabolism imbalance, osteoporosis, kidney stones, predisposition to thromboembolic events, and infectious diseases [7].

Our patient had a very severe and typical presentation, so laboratory examinations were performed to confirm a diagnosis that had been clinically made. However, as mentioned above, some patients may present mild symptoms, not as obvious as our patient, making the clinical suspicious not as simple. With the progressive increase in obesity and diabetes prevalence in world population, an important question is as follows: Who to screen for Cushing's syndrome? According to Nieman LK [7], some characteristics should raise the suspicion and make clinicians investigate it further:

- disease progression, with accumulation and worsening of signs and symptoms overtime;
- new or unexplained changes in cognition, mood, or memory;
- features incompatible with patient's age or population cohort (such as osteoporosis in young patient or recurrent infections in immunocompetent individuals);
- changes in fat distribution that suggest Cushing's syndrome such as supraclavicular and temporal fat (not dorsal pad or central obesity, which can be found in other causes of obesity);
- growth impairment with weight gain in children.

If there is clinical suspicion of Cushing's syndrome and there is no exogenous use of glucocorticoids, the next step is to confirm the hypercortisolism. For that, we have three main examinations that can be used: dexamethasone suppression test, late-night salivary cortisol, and 24-hour urinary free cortisol (UFC). It is necessary to have two altered examinations out of the three [8]. Other tests that can be used as second-line examinations in selected patients are late-night serum cortisol, ovine CRH after longer low-dose dexamethasone suppression test, human CRH test, and desmopressin test [5]. Sensitivity and specificity of first-line tests can be found in Table 11.3.

Overnight, low-dose dexamethasone suppression test is performed by administration of 1 mg of dexamethasone between 11 p.m. and 12 p.m., with cortisol measurement the next day at 8 a.m. The test is considered positive for hypercortisolism if there is no cortisol suppression to 1.8 mcg/dL (i.e., 50 nmol/L) or lower. The physiopathology behind this test is that, in normal individuals, due to negative feedback in normal corticotrophs, the administration of dexamethasone leads to suppression of cortisol secretion, which does not occur in patients with either ACTH- or cortisol-producing tumors. Alternatively, a long low-dose dexamethasone suppression test can be performed. In this variant, dexamethasone 0.5 mg is administered every 6 h for 48 h and cortisol is measured after this period. The positive criterion is the same as post-1 mg test. Test can either start at 9 a.m., with last dexamethasone administration at 3 a.m. and cortisol collection at 9 a.m., or at 12 a.m., with last dexamethasone administration at 6 a.m. [5]. Several drugs may alter dexamethasone metabolism, either accelerating or reducing it, which may lead to false positive or false negative, respectively. Other causes of false positive are states that increase cortisol-binding globulin (CBG), such as estrogen use and pregnancy.

Late-night salivary cortisol evaluates the cortisol circadian rhythm. It is a test with high sensitivity and specificity, apart from being easier to perform than suppression tests and, especially, 24-h UFC. It measures free cortisol, so there is no influence of high CBG states. Causes of false positive include psychiatric disorders (depression, anxiety disorders, and obsessive–compulsive disorder), poorly controlled diabetes mellitus, gum disease, pregnancy, and alcoholism [9]. Patients should be oriented to not smoking in the day of the examination and to brush their teeth gently.

24-hour UFC is used both for diagnosis of hypercortisolism and for monitoring the disease. It measures the total cortisol production over 24 h. At least two or three samples should be collected consecutively or alternately to decrease false-negative results. Also, creatinine should always be measured to assure that sample was properly collected [5]. The performance of UFC in the detection of mild hypercortisolism is less than optimal, and salivary cortisol seems to be a better option in such cases [10].

Test	Reference value	Sensitivity	Specificity
Post-1 mg dexamethasone suppression test	<1.8 mcg/dL	>95%	80%
Long low-dose dexamethasone suppression test	<1.8 mcg/dL	92-100%	92-100%
Late-night salivary cortisol	2× the ULN	88-100%	92-100%
24-h urinary free cortisol	$3-4\times$ the ULN	90–98%	45-95%

 Table 11.3
 Sensitivity and specificity of first-line test for the detection of hypercortisolism

Adapted from reference 5

ULN: upper limit of normal range

After confirming hypercortisolism, ACTH levels should be measured in order to differentiate between ACTH-dependent or ACTH-independent causes. ACTH levels lower than 10 pg/mL indicate an adrenal cause for cortisol oversecretion, and an adrenal imaging (MRI or CT) should then be performed. ACTH higher than 20 pg/mL is compatible with ACTH-dependent Cushing's syndrome, and levels between 10 and 20 pg/mL are indeterminate and should be repeated [5]. In ACTH-dependent cases, the most frequent etiology is a corticotropinoma (90% of cases), so the next step is a pituitary MRI [8]. If an adenoma ≥ 6 mm is identified, the diagnosis of Cushing's disease is confirmed. If there is no visible adenoma or if it is smaller than 6 mm, the differential diagnosis between pituitary Cushing's syndrome and ectopic ACTH syndrome must be performed.

A BIPSS is the gold standard method to differentiate between these two causes of ACTH-dependent Cushing's syndrome. In this examination, blood samples are collected from both petrosal sinuses, by inserting a catheter through the femoral vein until the petrosal sinuses, and from a peripheral site. Samples are collected before and after stimulation with corticotropin-releasing hormone (CRH) or desmopressin [11]. A petrosal sinus/periphery gradient >2 before and >3 after stimuli indicates a central origin (CD).

In sites where a BIPSS cannot be performed, a high-dose dexamethasone suppression test can be used as an alternative. The rationale of using this test is that pituitary adenomas, although secrete ACTH autonomously, maintain some degree of suppression by cortisol, whereas non-pituitary tissues are unlikely to respond. It is performed by administration of dexamethasone 2.0 mg every 6 h for 48 h, and serum cortisol is evaluated immediately before and after dexamethasone use. A cortisol suppression >50%, and especially 80–90%, indicates that the etiology is a pituitary adenoma [8].

An algorithm for the diagnosis of Cushing's syndrome can be found in Fig. 11.3.

11.4 Treatment

Transsphenoidal surgery (TSS) is the treatment of choice for CD [12]. Success rates vary according to the size of the adenoma and vary from approximately 40% for macroadenomas to more than 70% for microadenomas [13]. A study evaluated 108 patients submitted to transsphenoidal surgery and found an overall remission rate of 69% [14]. In this series, endoscopic surgery results were slightly higher than microscopic surgery (71.4% vs 69.3%), but no statistical analysis was performed, probably because of the limited number of patients submitted to endoscopic surgery. Our patient was in remission after the first surgery. Evaluation of remission in the immediate postoperative period can be made by dosing cortisol levels in the first 7 days after surgery, especially in the first 2 days. Cortisol levels <5 mcg/L suggest remission. Cortisol levels were not evaluated in our patient since hydrocortisone was administered due to adrenal insufficiency symptoms, which is also a criterion of remission.



Fig. 11.3 Algorithm for the diagnosis of Cushing's syndrome

Although surgical success rate is high, especially for microadenomas, recurrence is an important problem in Cushing's disease. Recurrence rates vary from 15% to 66% within 5–10 years of a successful surgery, but can occur after up to 20 years or more [13, 15]. Our patient presented recurrence after 2 years of successful surgery.

In case of no remission or recurrence, options are a second surgery, medical treatment, bilateral adrenalectomy, or radiotherapy [13, 16]. In the previously cited study [14], efficacy of a second surgery was lower than first procedure, with remission rate of 50%. Burke and coll [17]. demonstrated remission in 60.8% of 51 patients submitted to a second surgery for treatment of recurrent disease. In the presented case, a second surgery was not successful, and the adenoma was not visualized during the procedure.

Due to the severity of the case, after this second surgical attempt, patient was submitted to bilateral adrenalectomy. The endocrine society guidelines for the treatment of Cushing's syndrome indicate bilateral adrenalectomy for occult or metastatic EAS or for patients with very severe ACTH-dependent disease who cannot be controlled by medical therapy, as a life-preserving emergency treatment [13]. It is also a viable option for young female patients with a desire to be pregnant, in order to avoid the possibility of hypopituitarism associated with repeated surgery and radiotherapy. The main adverse effect of this treatment is primary adrenal insufficiency with the need of life-long administration of gluco- and mineralocorticoids [18]. Another important side effect is the development of corticotroph tumor progression, classically described as Nelson's syndrome, that is supposed to occur due to the absence of negative feedback from the hypercortisolism in the corticotropinoma [19]. ACTH levels should be monitored every 3 months for the first year, every 6 months for 3 years, and then annually. A rise up to 200 pg/mL is expected after bilateral adrenalectomy. Pituitary MRI should be done 3 months after surgery, then every 6 months for 2 years, and finally annually. Criteria for Nelson's syndrome is tumor increase detected in an imaging examination, ACTH levels >500 pg/ mL with progressive increase >30% in three occasions and hyperpigmentation [20].

Another second-line treatment option is radiotherapy/radiosurgery. It was not a suitable treatment for our patient since it takes up to 2 years to reach goal and our patient had a severe disease, with risk of suicide, so it was not possible to wait for this long time. A study evaluated the efficacy of stereotactic radiosurgery in 18 patients [21]. The remission rates after 2 years of the procedure were 56% and after 5 years 77%, five patients needed additional therapy due to lack of biochemical control and tumor growth was detected in one patient. 24% of patients developed a new pituitary hormone deficit after radiosurgery.

Finally, medical treatment is also possible for the management of recurrent Cushing's disease, may be used drugs that act centrally inhibiting ACTH secretion (pasireotide, pasireotide LAR, cabergoline, temozolomide), that block adrenal cortisol secretion (ketoconazole, metyrapone, mitotane, etomidate), and that block glucocorticoid receptor (mifepristone) [22]. Dosage, efficacy, and safety of the drugs used for the treatment of Cushing's disease are summarized in Table 11.4. Osilodrostat and levoketoconazole, steroidogenesis blockers, are now in phase 3 trials with promising results [23]. Preclinical studies with tumor-directed therapeutic targets may help establish novel therapeutic options for the medical management of Cushing's disease [24].

11.5 Follow-Up

Patients with Cushing's disease have a standard mortality ratio (SMR) 2.5 times higher than general population [25]. This mortality was even higher for patients not in remission (6.9 times), but it is important to emphasize that even patients in remission had higher mortality (1.9 times). The main causes of death were myocardial and cerebrovascular infarction, but mortality due to infection, and respiratory and digestive diseases were also increased [25].

Due to this increased mortality, Endocrine Society Guidelines recommend treating specific comorbidities that are associated with Cushing's disease throughout patient's life. It is also recommended to monitor for disease relapse, since it can be detected up to 20 years after a successful treatment [13]. It is important to educate

Drug	Dosage	Efficacy	Safety
Pasireotide	0.3–0.9 mg SC bid	24-h UFC normalization in up to 26% of cases	Hyperglycemia common, gastrointestinal toxicity, gallstones
Pasireotide LAR	10–30 mg IM every 4 week	24-h UFC normalization in 41% of cases	Hyperglycemia common, gastrointestinal toxicity, gallstones
Cabergoline	0.5–7 mg PO week	24-h UFC normalization in up to 40% of cases	Nausea, orthostatic dizziness, nasal congestion, psychiatric manifestations
Temozolomide	150–200 mg/m ² / day PO for 5 days each month	Partial or complete tumor response in up to 80% of cases	Neutropenia, thrombocytopenia, gastrointestinal toxicity, hearing loss
Ketoconazole	200–600 mg PO bid–tid	24-h UFC normalization in 49% of cases	Gastrointestinal toxicity, liver enzymes increase, risk of hepatic failure
Metyrapone	250–1000 mg PO qid	24-h UFC normalization in 43%	Gastrointestinal symptoms, dizziness, hyperandrogenism, mineralocorticoid excess
Mitotane	0.5–3.0 g PO tid	24-h UFC normalization in up to 85%	Gastrointestinal, hepatic, metabolic, neurologic, adverse effects
Etomidate	0.03 mg/kg IV (bolus), followed by continuous infusion (0.1– 0.3 mg/kg/h)	Highly effective in the short term	Sedation, nausea, vomiting, myoclonus, dystonia
Mifepristone	300–1200 mg PO daily	Improvement in hyperglycemia in 60%; decrease in blood pressure in 38%; global clinical improvement in 87%	Hypoadrenalism, hypertension, hypokalemia, endometrial thickening/ vaginal bleeding

 Table 11.4
 Dosage, efficacy, and adverse effects of drugs used for the medical management of Cushing's disease

Adapted from reference 22

bid twice a day, *tid* thrice a day, *qd* once a day, *IM* intramuscular, *IV* intravenous, *SC* subcutaneous, *UFC* urinary free cortisol

patients and family members with respect to signs and symptoms of disease relapse, so it can be readily diagnosed. Monitoring of disease relapse can be made with either one of the three first-line examinations for the detection of hypercortisolism.

11.6 Learning Points

 Cushing's syndrome is a complex disease, with a myriad of clinical presentations that can vary from a rich clinical picture, which is readily diagnosed, to a mild presentation that needs a high suspicion in order to be sought.

- The diagnostic workup is also complex, involving a diversity of tests that may present false-positive or false-negative results, making it necessary to perform several examinations to reach a final conclusion.
- Treatment of Cushing's disease is multidisciplinary, involving endocrinologists, neurosurgeons, radiotherapists, and general surgeons. Recurrence after an initially successful treatment is not uncommon, so multimodal therapy is often necessary.
- Cushing's disease is associated with elevated mortality, even in patients in remission, so long-life monitoring of disease relapse and comorbidities is warranted.

Questions and Answers

- (1) Which is the main cause of hypercortisolism?
 - (A) Cushing's diseases.
 - (B) Adrenal adenoma.
 - (C) Ectopic ACTH-producing tumor-directed.
 - (D) Exogenous glucocorticoid use.
 - (E) None of the above.
- (2) Which of these signs are specific of Cushing's syndrome?
 - (A) Central obesity.
 - (B) Purple striae.
 - (C) Buffalo hump.
 - (D) Hirsutism.
 - (E) Acne.
- (3) Which comorbidity can be associated with Cushing's syndrome?
 - (A) Hypertension.
 - (B) Diabetes mellitus.
 - (C) Venous thromboembolism.
 - (D) Osteoporosis.
 - (E) All of the above.
- (4) Which one is a Cushing's syndrome differential diagnosis?
 - (A) Polycystic ovary syndromes.
 - (B) Depression.
 - (C) Anorexia nervosa.
 - (D) All of the above.
 - (E) None of the above.
- (5) In the initial investigation of Cushing's syndrome what examinations can be used?
 - (A) Dexamethasone suppression test.
 - (B) Basal 8 a.m. cortisol levels.

- (C) Late-night serum cortisol.
- (D) ACTH levels.
- (E) Magnetic resonance imaging.
- (6) After confirming hypercortisolism, which examination should be ordered next?
 - (A) Magnetic resonance imaging of the sella turcica.
 - (B) Abdominal computed tomography.
 - (C) ACTH levels.
 - (D) Simultaneous bilateral inferior petrosal sinus sampling.
 - (E) Desmopressin test.
- (7) Which gradient defines pituitary-dependent Cushing's syndrome in a simultaneous bilateral inferior petrosal sinus sampling?
 - (A) Petrosal sinus/periphery before stimuli >1.
 - (B) Periphery/petrosal sinus before stimuli >2.
 - (C) Petrosal sinus/periphery after stimuli >2.
 - (D) Periphery/petrosal sinus after stimuli >3.
 - (E) Petrosal sinus/periphery after stimuli >3.
- (8) What is the first-line treatment for Cushing's disease?
 - (A) Medical treatment.
 - (B) Transsphenoidal surgery.
 - (C) Adrenalectomy.
 - (D) Radiotherapy.
 - (E) All of the above.
- (9) Which drug is not used in the treatment of Cushing's disease?
 - (A) Octreotide.
 - (B) Pasireotide.
 - (C) Cabergoline.
 - (D) Ketoconazole.
 - (E) Metyrapone.
- (10) Which is the main cause of mortality in patients with Cushing's disease?
 - (A) Infectious diseases.
 - (B) Respiratory diseases.
 - (C) Cardiovascular diseases.
 - (D) Cancer.
 - (E) Digestive diseases.

Answers:

- 1. D
- 2. B
- 3. E
- 4. D

- 5. A
- 6. C
- 7. E
- 8. B
- 9. A
- 10. C

References

- 1. Debono M, Newell-Price JD. Cushing's syndrome: where and how to find it. Front Horm Res. 2016;46:15–27.
- Chabre O. The difficulties of pseudo-Cushing's syndrome (or "non-neoplastic hypercortisolism"). Ann Endocrinol (Paris). 2018;79(3):138–45.
- Brzana J, Yedinak CG, Hameed N, Plesiu A, McCartney S, Fleseriu M. Polycystic ovarian syndrome and Cushing's syndrome: a persistent diagnostic quandary. Eur J Obstet Gynecol Reprod Biol. 2014;175:145–8.
- Tirabassi G, Boscaro M, Arnaldi G. Harmful effects of functional hypercortisolism: a working hypothesis. Endocrine. 2014;46(3):370–86.
- Machado MC, Fragoso MC, Moreira AC, Boguszewski CL, Vieira L, Naves LA, et al. Recommendations of the neuroendocrinology Department of the Brazilian Society of endocrinology and metabolism for the diagnosis of Cushing's disease in Brazil. Arch Endocrinol Metab. 2016;60(3):267–86.
- 6. Guaraldi F, Salvatori R. Cushing syndrome: maybe not so uncommon of an endocrine disease. J Am Board Fam Med. 2012;25(2):199–208.
- Nieman LK. Diagnosis of Cushing's syndrome in the modern era. Endocrinol Metab Clin N Am. 2018;47(2):259–73.
- Nieman LK, Biller BM, Findling JW, Newell-Price J, Savage MO, Stewart PM, et al. The diagnosis of Cushing's syndrome: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab. 2008;93(5):1526–40.
- 9. Carroll T, Raff H, Findling JW. Late-night salivary cortisol measurement in the diagnosis of Cushing's syndrome. Nat Clin Pract Endocrinol Metab. 2008;4(6):344–50.
- Alexandraki KI, Grossman AB. Is urinary free cortisol of value in the diagnosis of Cushing's syndrome? Curr Opin Endocrinol Diabetes Obes. 2011;18(4):259–63.
- Deipolyi A, Bailin A, Hirsch JA, Walker TG, Oklu R. Bilateral inferior petrosal sinus sampling: experience in 327 patients. J Neurointerv Surg. 2017;9(2):196–9.
- Pivonello R, De Leo M, Cozzolino A, Colao A. The treatment of Cushing's disease. Endocr Rev. 2015;36(4):385–486.
- Nieman LK, Biller BM, Findling JW, Murad MH, Newell-Price J, Savage MO, et al. Treatment of Cushing's syndrome: an Endocrine Society clinical practice guideline. J Clin Endocrinol Metab. 2015;100(8):2807–31.
- Martínez Ortega AJ, Venegas-Moreno E, Dios E, Remón Ruíz PJ, Márquez Rivas FJ, Valdepeñas EC, et al. Surgical outcomes and comorbidities in Cushing disease: 30 years of experience in a referral Center. World Neurosurg. 2019;122:e436–e42.
- 15. Nadezhdina EY, Rebrova OY, Grigoriev AY, Ivaschenko OV, Azizyan VN, Melnichenko GA, et al. Prediction of recurrence and remission within 3 years in patients with Cushing disease after successful transnasal adenomectomy. Pituitary. 2019;22:574.
- 16. Tritos NA, Biller BMK. Current management of Cushing's disease. J Intern Med. 2019;286:526.
- Burke WT, Penn DL, Repetti CS, Iuliano S, Laws ER. Outcomes after repeat Transphenoidal surgery for recurrent Cushing disease: updated. Neurosurgery. 2019;85:E1030.
- Sarkis P, Rabilloud M, Lifante JC, Siamand A, Jouanneau E, Gay E, et al. Bilateral adrenalectomy in Cushing's disease: altered long-term quality of life compared to other treatment options. Ann Endocrinol (Paris). 2019;80(1):32–7.

- Reincke M, Ritzel K, O
 ßwald A, Berr C, Stalla G, Hallfeldt K, et al. A critical reappraisal of bilateral adrenalectomy for ACTH-dependent Cushing's syndrome. Eur J Endocrinol. 2015;173(4):M23–32.
- Patel J, Eloy JA, Liu JK. Nelson's syndrome: a review of the clinical manifestations, pathophysiology, and treatment strategies. Neurosurg Focus. 2015;38(2):E14.
- 21. Hughes JD, Young WF, Chang AY, Link MJ, Garces YI, Laack NN, et al. Radiosurgical Management of Patients with Persistent or recurrent Cushing disease after prior Transphenoidal surgery: a management algorithm based on a 25-year experience. Neurosurgery. 2019;
- 22. Tritos NA, Biller BMK. Medical management of Cushing disease. Neurosurg Clin N Am. 2019;30(4):499–508.
- Fleseriu M, Castinetti F. Updates on the role of adrenal steroidogenesis inhibitors in Cushing's syndrome: a focus on novel therapies. Pituitary. 2016;19(6):643–53.
- Theodoropoulou M, Reincke M. Tumor-directed therapeutic targets in Cushing disease. J Clin Endocrinol Metab. 2019;104(3):925–33.
- 25. Ragnarsson O, Olsson DS, Papakokkinou E, Chantzichristos D, Dahlqvist P, Segerstedt E, et al. Overall and disease-specific mortality in patients with Cushing disease: a Swedish Nationwide study. J Clin Endocrinol Metab. 2019;104(6):2375–84.



Clinical Case 3

12

Ana M. Ramos-Leví, Nerea Aguirre-Moreno, and Monica Marazuela

12.1 Case Report

A 57-year-old man with a past medical history of ulcerative colitis was referred for the evaluation of thyroid dysfunction concordant with secondary hyperthyroidism (TSH 6.44 uU/mL [reference levels 0.27–4.20] and FT4 5.07 [reference levels 0.93–1.70]). He recalled a 5-year history of similarly altered thyroid biochemical values that had never been previously considered. He did not smoke or drink alcohol, he denied intake of herbal or unknown products, his bowel habit was stable, and he had not undergone recent diagnostic tests with iodinated contrast media.

The patient acknowledged occasional tachycardia, anxiety, and sleeping difficulties, which he attributed to stress at work, and physical examination revealed normal blood pressure, a heart rate of 85 per minute, rapid pace of speech, mild diffuse goiter with no palpable nodules, and mild distal tremor and excess sweat in hands. Laboratory workup was consistent with secondary hyperthyroidism, but normality in the rest of the hormonal and biochemical parameters evaluated (pituitary hormones, testosterone, basal cortisol, IGF-I, ions, proteins, lipid, renal and hepatic profiles), and negative thyroid antibodies (anti-TPO, anti-Tg, and TRAb) (Table 12.1). Radioactive iodine uptake was diffusely elevated, suggesting global hyperfunction and thyroid ultrasound revealed a heterogeneous pattern. Pituitary magnetic resonance imaging (MRI) revealed an area of 14×5 mm on the left side of the pituitary, which was hypointense in T1-weighted images and isointense in

Department of Endocrinology, Hospital Universitario de la Princesa, Instituto de Investigación Princesa, Universidad Autónoma de Madrid. C/ Diego de León 62, Madrid, Spain

The chapter has been endorsed by **Prof. Diego Ferone**, ferone@unige.it, Endocrinology Unit, Department of Internal Medicine and Medical Specialties (DiMI), Center of Excellence for Biomedical Research (CEBR), IRCCS AOU San Martino-IST, University of Genoa, Genoa, Italy

A. M. Ramos-Leví · N. Aguirre-Moreno · M. Marazuela (🖂)

[©] Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_12

Clinical parameter (units)	Result	Reference range
FT4 ng/dL	3.77	0.93-1.77
TSH uU/mL	6.06	0.27-4.30
Total T3 ng/mL	2.5	0.8–2.1
LH mU/mL	3.34	1.26-10.50
FSH mU/mL	5.20	1.37-13.58
Prolactin ng/mL	14.65	2.58-20.00
Total testosterone ng/mL	9.01	2.21-7.15
SHBG nmol/L	81.8	14.5-48.4
Calculated free testosterone	377.2	228-720
pmol/L		
Basal GH ng/mL	0.44	0.2–2
IGF-I ng/mL	171.2	55–186
Basal cortisol ug/dL	19.2	2.3–19.4
ACTH pg/mL	41.49	5-60

 Table 12.1
 Patient's laboratory workup at the time of initial evaluation



Fig. 12.1 Pituitary magnetic resonance imaging (MRI) sections. Findings revealed a hypointense in T1-weighted images and isointense in T2-weighted images area of 14×5 mm on the left side of the pituitary, which extended to the left cavernous sinus and contacted the left internal carotid artery (indicated with small arrows). The rest of the pituitary was mildly displaced to the right, with mild stalk deviation. (a) Frontal T1-weighted after gadolinium; (b) frontal T2-weighted; and (c) sagittal T1-weighted after gadolinium at diagnosis. (d) Frontal T1-weighted after gadolinium; (e) frontal T2-weighted; and (f) sagittal T1-weighted after gadolinium after 6 months of medical treatment with long-acting laneotide

T2-weighted images, extended to the left cavernous sinus and contacted the left internal carotid artery. The rest of the pituitary was mildly displaced to the right, with mild stalk deviation (Fig. 12.1a–c). Neuro-ophthalmological evaluation, including optical coherence tomography, was unremarkable.

12.2 Differential Diagnosis

In the setting of this confirmed clinical setting of raised thyroid hormones with nonsuppressed TSH, and once potential confounding factors have been reasonably ruled out (intercurrent non-thyroidal illness, drug interactions, altered protein profile, and assay interference), genetic and acquired disorders of the hypothalamic– pituitary–thyroid axis function should be ruled out.

Specifically, in this case, the differential diagnosis of what has been termed the "syndrome of inappropriate secretion of TSH" would include the existence of a TSH-secreting pituitary adenoma (TSHoma) versus resistance to thyroid hormone syndrome (RTH) plus pituitary nonfunctioning adenoma [1] or RTH plus pituitary enlargement, for example, after thyroid ablative treatment [2]. In these settings, although central hyperthyroidism is present, the clinical picture usually helps in differentiating them, since patients with TSHoma frequently exhibit hyperthyroid symptoms, while RTH patients are generally euthyroid (so-called generalized RTH). However, in a minority of RTH patients, thyrotoxic features are also present, mainly in those considered to have predominant central or pituitary resistance [3, 4], and the coexistence of a pituitary incidentaloma may entail a blurred differential diagnosis [5].

The stability of the clinical and analytical scenario could lean the diagnostic suspicion toward the diagnosis of RTH. However, on the other side, the mildly elevated levels of SHBG, the increased radioactive iodine uptake, and the existence of a pituitary macroadenoma favored the existence of a TSHoma. Sequencing of exons 1 through 10 of the TR β gene was performed, which eventually resulted to be negative for the presence of specific known mutations. Sequencing of the TR α gene was not deemed necessary until other diagnostic tests had been performed [4, 6].

Regarding biochemical tests, serum TSH levels within the normal range are more frequently found in RTH, while elevated α -GSU concentrations and/or high α -GSU/TSH molar ratio are typical of TSHomas. However, no differences in terms of age, sex, TSH levels, or free thyroid hormone concentrations have been described as significant between patients with TSHoma and those with RTH [1]. In dynamic testing, the absence of TSH response to the T3 suppression test, combined with the absence of an increase in TSH or alpha-GSU after TRH administration, would be highly sensitive and specific for the diagnosis of TSHoma. However, dynamic testing may not always be recommended; for instance, the T3 suppression test is contraindicated in elderly patients or in those with coronary heart disease [1, 7, 8].

Dynamic testing or evaluation of alpha-GSU may not be readily available in all clinical settings. Thus, in the meantime, in our case, a pituitary indium-111 octreotide uptake (Octreoscan) was empirically performed [8], revealing a focal lesion of increased uptake of approximately 1.5 cm (Fig. 12.2), and a short-acting octreotide test was programmed [8]. Results of this dynamic test are shown in Table 12.2. We can observe how TSH and FT4 levels are normalized, PRL levels remained stable, and SHBG and total testosterone levels also decreased, with total testosterone remaining stable. Overall, this would imply a true effect of octreotide on TSH secretion, leading to a mild amelioration of the hyperthyroid status. The patient acknowl-edged a mild increase in his bowel habit with the first octreotide injections, which subsequently returned to normal, and, interestingly, referred a subjective impression



Fig. 12.2 Pituitary indium-111 octreotide uptake (Octreoscan) revealing a focal lesion of increased uptake of approximately 1.5 cm

Table 12.2 Results of the short-actin	ig octreotide test
---------------------------------------	--------------------

	Baseline	Result after 2 weeks of 50 ug	Reference
Clinical parameter (units)	result	octreotide every 8 h sc	range
FT4 ng/dL	3.89	0.98	0.93-1.77
TSH uU/mL	5.19	0.21	0.27-4.30
Prolactin ng/mL	13.91	10.11	2.58-20.00
Total testosterone ng/mL	11.13	5.46	2.21-7.15
SHBG nmol/L	119.8	47.0	14.5-48.4
Calculated free	342.5	321.0	228-720
testosterone pmol/L			
IGF-I ng/ml	177.9	85.03	55-186

of clinical improvement regarding his anxious symptoms. This combination of the use of somatostatin receptor imaging techniques (Octreoscan or Gallium-68-PET-DOTATOC scan) and the short-acting octreotide test reasonably ruled out the risk of pituitary incidentaloma (mainly clinically nonfunctioning pituitary adenoma), even though other classical thyroid hormone dynamic tests were not performed.

12.3 Diagnostic Aspects

The combined interpretation of clinical, analytical, and radiological findings favored the diagnosis of TSHoma. Specifically, the increased octreotide scintigraphy uptake where a pituitary adenoma was observed in the MRI [9, 10], and the significant decrease in hormonal values, normalization of SHBG levels, and subjective improvement of hyperthyroid symptoms (tachycardia, anxiety) following the octreotide test supported the existence of a TSH-secreting pituitary adenoma.

12.4 Treatment

The case was thoroughly revised by a multidisciplinary group of endocrinologist, radiologists, neurosurgeons, and radiotherapists. Because the lesion seemed to be in contact with the cavernous sinus and the patient was relatively reluctant to undergo surgery, medical treatment with long-acting somatostatin analogs was offered.

12.5 Follow-up

The patient was started on long-acting lanreotide 60 mg sc every 28 days [8, 11]. Initial follow-up laboratory workup 3 months after starting therapy revealed stabilization of thyroid function tests, as well as the rest of other hormonal and biochemical values: FT4 1.26 ng/dL, TSH 1.07 nuU/mL, total testosterone 6.29 ng/mL, SHBG 45.1 nmol/L, free testosterone 390.22 pmol/L, basal cortisol 17.7 ug/dL, and IGF-I 181.4 ng/mL (reference ranges as described in Tables 12.1 and 12.2). Follow-up pituitary MRI 6 months after revealed stability of the adenoma, with no change in its size or characteristics (Fig. 12.1d–f). Clinical, analytical, and radiological stabilization has endured over the 2-year follow-up that the patient has undergone up to now (Fig. 12.3).

12.6 Learning Points

 When a patient presents with high levels of circulating thyroid hormones and non-suppressed TSH, potential laboratory methodological interferences should be reasonably ruled out. Once the laboratory values have been confirmed, the differential diagnosis includes TSHoma and resistance to thyroid hormone syndrome (RTH).



Fig. 12.3 Timeline summarizing TSH and FT4 values during the patient's follow-up. Values are shown as TSH uU/mL and FT4 ng/dL. Shaded blue box represents normal reference values

- The clinical appearance of hyperthyroidism may be mild, progressive, and relatively well tolerated.
- Dynamic tests, such as T3 suppression test and TRH stimulation test, may not always be available. In these cases, an octreotide test may help as a diagnostic and therapeutic approach to the diagnosis of TSHoma.
- Although the first treatment of choice for TSHomas is surgery, medical treatment with long-acting somatostatin analogs may be considered when the patient is not an appropriate surgical candidate.

Questions and Answers

- 1. When should we consider the possible existence of a TSHoma?
 - Hyperthyroidism.
 - Increased free thyroid hormone levels with detectable TSH.
 - Pituitary tumors.
- 2. Which diagnostic tests can help in the differential diagnosis between TSHomas and thyroid hormone resistance (RTH)?
 - History and physical examination.
 - Elevated free T4 and total T4, with non-suppressed TSH.
 - T3 suppression test.
 - TRH stimulation test.
 - Octreotide test.
 - Alpha-subunit evaluation.

- Pituitary magnetic resonance imaging.
- Pituitary indium-111 octreotide uptake (Octreoscan).
- 3. What is the main treatment of TSHoma?
 - Surgery.
- 4. What medical approach can we use when surgery is not feasible?
 - Long-acting somatostatin analogs.

References

- Beck-Peccoz P, Lania A, Beckers A, Chatterjee K, Wemeau JL. 2013 European Thyroid Association Guidelines for the diagnosis and treatment of thyrotropin-secreting pituitary tumors. Eur Thyroid J. 2013;2:76–82.
- Marazuela M, Nattero L, Moure D, García-Polo I, Figueroa-Vega N, Guijarro C. Thyroid hormone resistance and pituitary enlargement after thyroid ablation in a woman on levothyroxine treatment. Thyroid. 2008;18:1119–23.
- Gurnell M, Visser TJ, Beck-Peccoz P, Chatterjee VKK. Resistance to thyroid hormone. In: Jameson LJ, LJ DG, editors. Endocrinology, adult and pediatric, vol. II. 6th ed. Philadelphia: Saunders Elsevier; 2010. p. 1745–59.
- 4. Bernal J. Thyroid hormone resistance syndromes. Endocrinol Nutr. 2011;58:185-96.
- Ramos-Leví AM, Moreno JC, Álvarez-Escolá C, Lacámara N, Montañez MC. Coexistence of thyroid hormone resistance syndrome, pituitary adenoma and Graves' disease. Endocrinol Nutr. 2016;63:139–41.
- Briet C, Bouhours-Nouet N, Illouz F, Prunier-Mirebeau D, Rodien P. TRα mutations in human. In: Plateroti M, Samarut J, editors. Thyroid hormone nuclear receptor. Methods in molecular biology, vol. 1801. New York, NY: Humana Press; 2018.
- Beck-Peccoz P, Persani L, Mannavola D, Campi I. Pituitary tumours: TSH-secreting adenomas. Best Pract Res Clin Endocrinol Metab. 2009;23:597–606.
- Beck-Peccoz P, Giavoli C, Lania A. A 2019 update on TSH-secreting pituitary adenomas. J Endocrinol Investig. 2019; https://doi.org/10.1007/s40618-019-01066-x.
- 9. Foppiani L, del Monte P, Ruelle A, et al. TSH-secreting adenomas: rare pituitary tumors with multifaceted clinical and biological features. J Endocrinol Invest. 2007;30:603–9.
- Losa M, Magnani P, Mortini P, et al. Indium-111 pentetreotide single-photon emission tomography in patients with TSH-secreting pituitary adenomas : correlation with the effect of a single administration of octreotide on serum TSH levels. Eur J Nuclear Med. 1997;24:728–31.
- Paniagua AE, Bernabeu I, Leskela S, Marazuela M. Lanreotide autogel-induced tumour shrinkage in thyrotropin-secreting pituitary macroadenomas. Clin Endocrinol. 2011;74:406–8.



Clinical Case 4

13

Francesca D'Ercole, Irene Gagliardi, Maria Rosaria Ambrosio, and Maria Chiara Zatelli

13.1 Case Report

We report the case of a male patient who was referred in 2003, at 59 years of age, for the incidental finding of a sellar mass, with partial sellar floor erosion, indentified after a computed tomography (CT) scan performed during investigation for sinusitis. Subsequently, the patient was submitted to sellar magnetic resonance imaging (MRI), showing a $29 \times 28 \times 20$ mm lesion with intra- and extrasellar expansion, suprasellar cistern engagement, hourglass morphology, homogeneous contrast enhancement, optic chiasm compression, and invasion of the sphenoid sinus and of the right cavernous sinus most likely consistent with a pituitary neuroendocrine tumor (Pit-NET) (Fig. 13.1, sections 1a and 1b).

The patient complained of frontal headache, otherwise he was in good clinical condition, in particular he did not refer visual or neurological deficits. An endocrine workup was requested, with the evaluation of pituitary function, which was normal. The patient was submitted to visual field (VF) evaluation that showed few points of altered sensitivity, bilaterally.

The chapter has been endorsed by **Prof. Alberto Pereira**, A.M.Pereira@lumc.nl, Center for Endocrine Tumors, Division of Endocrinology, Department of Medicine, Leiden University Medical Center, Leiden, Netherlands

Francesca D'Ercole and Irene Gagliardi equally contributed to this work.

F. D'Ercole · I. Gagliardi · M. R. Ambrosio · M. C. Zatelli (⊠) Section of Endocrinology and Internal Medicine, Department of Medical Sciences, University of Ferrara, Ferrara, Italy e-mail: ztlmch@unife.it



Fig. 13.1 (1a) Coronal and (1b) axial preoperative T1-weighted gadolinium contrast-enhanced MR image; (2a) coronal and (2b) axial postoperative T1-weighted gadolinium contrast-enhanced MR image; (3a) coronal and (3b) axial post-RT T1-weighted gadolinium contrast-enhanced MR image; (4a) coronal and (4b) axial T1-weighted gadolinium contrast-enhanced MR images at last follow-up

On the basis of the clinical and radiological findings, in February 2004 the patient was submitted to surgery by transsphenoidal adenomectomy (TSA) approach to remove the pituitary lesion. Histological examination disclosed a chromophobe cell adenoma with focal immunohistochemical reactivity for LH and FSH, consistent with a NFPA. Postoperative course was uneventful. Pituitary function assessed immediately after surgery and 2 months later was in the normal range. Three months after surgery, a control MRI showed a reduction in size of the sellar mass (maximum diameter ~ 20 mm); suprasellar cisterns and optic chiasm were no longer involved (Fig. 13.1, sections 2a and 2b). VF was normal and headache disappeared. Sellar MRI performed in 2005 (12 months after surgery) showed no significant changes and, due to the high risk of growth recurrence of the remnant tumor and the optimal distance from the chiasm after surgery, Gamma Knife stereotactic radiosurgery was performed in September 2005.

During the following year, the patient developed multiple pituitary hormone deficiencies, promptly replaced. The patient underwent annual follow-up with MRI, VF evaluation, and pituitary function assessment. Sellar MRI performed on March 2007 showed a ~ 20 mm mass with an irregular central area of liquid type signal, hypointense in T1, and hyperintense in T2 without contrast uptake, surrounded by parenchymatous tissue with contrast uptake. Pituitary stalk was clearly deviated to the left, and the optic chiasm had regular morphology (Fig. 13.1, sections 3a and 3b).

The last follow-up MRI, performed in September 2017, reported overlapping findings and pituitary function never recovered (Fig. 13.1, sections 4a and 4b). The patient reported physical well-being, but he considered his everyday quality of life (QoL) negatively affected by multiple pharmacological therapies including pituitary hormone replacement drugs.

13.2 Differential Diagnosis

Technological improvements in neuroradiology led to the identification of an increasing number of sellar/parasellar masses, often incidentally found during brain imaging performed for different diagnostic indications. Pit-NETs are the most common causes of sellar/parasellar masses, but other lesions could display similar clinical, endocrine, and radiological characteristics, indicating the need to consider them in the differential diagnosis. MRI is the gold standard radiological approach for sellar/parasellar region assessment, while CT scan could help to identify calcifications and bone remodeling [1, 2]. Pit-NET and Rathke's cleft cysts are the most common sellar/parasellar masses representing up to 90% of all lesions, followed by craniopharyngiomas (4.2–5.6%) and cystic malformations (2.9–5.2%) [2, 3] (Table 13.1).

Once the diagnosis of Pit-NET is established, it is well known that NFPAs occur more frequently as compared to functioning Pit-NET. The prevalence of NFPA varies between 60 and 100 cases per million inhabitants, with a bimodal peak incidence between the ages of 25–45 and 60–70 years. Less than 5% of Pit-NET occur as a component of hereditary syndromes, such as MEN1, MEN4, Carney complex, and familial isolated pituitary adenomas (FIPAs) [4].

13.3 Diagnostic Aspects

13.3.1 Clinical Presentation

NFPAs are benign Pit-NET not associated with clinical or biochemical evidence of hormonal hypersecretion. As a consequence, in many cases these lesions are found incidentally during radiological examinations (CT or MRI) performed for other reasons (i.e., pituitary incidentaloma). Moreover, the absence of endocrine symptoms often causes a diagnostic delay; as a consequence, NFPAs are frequently diagnosed when they are large enough to cause compressive effects on the surrounding structures. Besides, although usually benign, NFPAs represent a very heterogeneous group of tumors, with varying biological behaviors and clinical manifestations that range from completely asymptomatic lesions to rapidly growing neoplasms, causing mass effect, and neurological or endocrine symptoms. The most common neurological symptoms are visual defects and headache, usually leading to neuroradiological examination. The former occur due to optic chiasm compression; different types of alterations may arise depending on the degree and the site of compression: unilateral, bilateral, partial, or complete VF defects. The most typical VF alteration is bitemporal hemianopsia. Furthermore, lesions anterior to the optic chiasm can cause central scotoma, while posterior lesions can involve the optic tracts and lead to a homonymous hemianopsia. If compression is severe and prolonged, a decline in visual acuity may also develop. Optic atrophy and papilledema rarely occur [5].

			Current Contractions	Am 1 substance
		Definition	Clinical manifestations	MR characteristics
Normal anatomical variant	Small sella	Pituitary pseudo-enlargement in a relatively small sella (due to sphenoid sinus hyperpneumatization, thicker dorsum sellae, or small/narrow sella)		 Normal T1 and T2 signals Normal contrast enhancement
Pituitary hyperplasia	Physiologic sex- and age-dependent hyperplasia	Pituitary hyperplasia in young age or menopausal women Lactotrope hyperplasia in pregnant women		 Symmetrical pituitary enlargement and superior convexity Normal adeno- and neurohypophysis signals
	Pathologic hyperplasia	Thyrotrope hyperplasia Pituitary hyperplasia due to CRH or GHRH hypersecretion	Severe primary hypothyroidism Hyperprolactinemia and related symptoms Cushing or acromegaly disease McCune–Albrieht syndrome	Homogenous contrast enhancement
Anterior pituitary tumors	Pituitary adenoma Pituitary carcinoma	Deriving from adenohypophyseal cell proliferation	 Asymptomatic Tumor mass effects Hormonal hypersecretion symptoms 	 Intrasellar microadenomas (<1 cm): Lateralization inside the gland Possible sellar diaphragm deformation Pituitary stalk displacement Macroadenomas (>1 cm): Enlarged sella turcica Not intense contrast enhancement Possible areas of necrotic-hemorrhagic remodelin
Posterior pituitary tumors	Pituicytoma Granular cell tumors	Arising from neurohypophysis Arising from neurohypophysis or infundibulum	 Headache Hypopituitarism Diabetes insipidus (rare) 	 T1 isointensity Possible displacement of the normal adenohypophysis
Benign parasellar tumors	Craniopharyngioma	Epithelial tumors arising along the pathway of the craniopharyngeal duct. It can be predominantly solid, cystic, or mixed	 Headache Nausea/vomiting Visual impairment Hydrocephalus Endocrine dysfunction Hypothalamic dysfunction Papilledema Cranial nerve palsics Hydrocephalus 	 T1 isointensity/ipointensity and T2 hyperintensity of the solid part T1 hyperintensity of the cystic part Calcifications
	Ependymoma	Embryonal brain tumor	 Compression symptoms Endocrine dysfunction Diabetes insipidus Fever (lymphomas) 	

 Table 13.1
 Sellar mass differential diagnosis

ignant ors	Primary CNS lymphomas	Ansing primarily in the craniospinal axis	 Visual impairment Headache 	 Slightly 1.2 hypointensity Homogenous contrast enhancement
	Glioma (hypothalamic or optic pathway)	Arising from hypothalamic or optic pathway	 Endocrine dysfunction Dizziness, tinnitus, facial sensory deficits, ataxia, and hemiparesis (rare) 	 T1 hypointensity T2 hyperintensity
	Germinal cell tumors	Arising Around the third ventricle, followed by the suprasellar compartment and anterior hypothalamic regions		 T1 isointensity T2 isointensity/hyperintensity Homogeneous with a great contrast enhancement Rarely cystic Subrasellar infiltration
	Chondoma Chondromas Chondrosarcomas	Tumors that arise from the primary notochord Tumors that arise from cartilaginous remnants		 Bone erosion Heterogeneous mass with internal septations and heterogeneous enhancement Chordomas are typically midline lesions
	Metastases	Most common deriving from breast and lung cancers in women and men, respectively	 Rapid onset and progression of symptoms Masse effect symptom Endocrine dysfunction Diabetes insipidus Nerve palsies 	 Loss of posterior pituitary bright spot and thickening of the pituitary stalk Infiltration of the posterior pituitary and stalk Bone erosions Invasion of Sella diaphragm Invasion of Sella diaphragm Signal intensity varies depending on the primary tumor, but generally they present T1 hypointensity and T2 by veritheralty
	Plasmacytoma		Cranial neuropathies	 T1 and T2 hypointensity Bony destruction of the sella
				(continued)

Table 13.1 (cc	ontinued)			
		Definition	Clinical manifestations	MR characteristics
Malformative lesions	Rathke's cleft cyst	Originating from the remnant of the squamous epithelium of Rathke's pouch. Consist of a single layer of epithelial cells with mucoid, cellular, or serous components in the cyst fluid	 Usually asymptomatic Compressive symptoms Endocrine dysfunction Diabetes insipidus 	 Cystic lesions with variable signal intensity Same density of CSF, with low intensity on T1-weighted and high intensity on T2-weighted images Contrast enhancement is rare or confined to the cyst wall T2 hypointense intracystic nodules due to cholesterol
	Dermoid and epidermoid cyst	Epithelial elements originating from incomplete separation of the neuroectoderm from the cutaneous ectoderm	 Hydrocephalus Visual disturbance Hypopituitarism Diabetes insipidus Cranial nerve abnormalities 	 T1 and T2 hyperintensity (dermoid cysts) Radiologic findings similar to cerebrospinal fluid with no contrast enhancement (epidermoid cysts)
	Arachnoid cyst	Hemiation of the arachnoid diverticulum through the sellar diaphragm	 Increased intracranial pressure Hypopituitarism Visual impairment 	 Well-defined lesions T1 and T2 isointensity No contrast enhancement
	Hamartoma	Congenital heterotopias usually located within the tuber cinereum	Mass effect symptoms	 Pedunculated hypothalamic mass, isointense on MRI to gray matter. No contrast enhancement
Vascular lesions	Aneurysms Cavernous angiomas Cavernous sinus thrombosis			

Infective etiology Immune checkpoint inhibitor-induced

In addition, ophthalmoplegia may occur due to compression of nervous structures contained in the cavernous sinus, with the development of different clinical pictures depending on the involved cranial nerves, most frequently the oculomotor nerve. Diplopia could be caused by paralysis of the III or VI cranial nerve. V cranial nerve involvement with consequent trigeminal neuralgia is rare. Similarly, the prevalence of oculomotor abnormalities due to the involvement of cranial nerves is very low (<5%). Indeed, the presence of these symptoms occurring in the context of a large pituitary mass should prompt the suspicion of alternative diagnoses such as apoplexy, infiltrative disorders (sarcoidosis or histiocytosis), and pituitary metastasis from other primary tumors [4].

In the case of NFPA, visual defects appear to be the most frequently reported disorders. However, patients frequently seek medical examination several months or even several years after symptom onset, since in most cases the gradual onset of visual impairments typically remains unnoticed for patients for a long time. The main risk factor for a delayed diagnosis seems to be the age of the patient, who tends to accept the decline of visual function as part of the natural aging process [6]. On the contrary, a timely diagnosis is fundamental because there is a linear correlation between visual symptoms duration and permanent damage probability, persisting after debulking surgery.

Therefore, an ophthalmologic assessment is essential for all NFPA patients, and it should include:

- sensory evaluation: At least one visual acuity (VA) measurement and VF examination, preferably central static + peripheral kinetic. Anterior segment and fundus examinations are also essential for interpreting VA and VF data; optic coherence tomography (OCT) is helpful but not essential for determining visual prognosis;
- oculomotor evaluation: Diplopia should be ruled out on interview, checking the
 patient's nine gaze directions, which may be completed by orthoptic examination
 and Lancaster's test.

Headache is another very common symptom regardless of adenoma size. A recent prospective study estimated a 48.5% headache prevalence in NFPA [7]. Clinical presentation of this symptom may widely vary, and it is often difficult to distinguish from primary headache (such as migraine or tension headache). Several mechanisms have been proposed to explain its pathogenesis: an increase in intrasellar pressure, stretching of the dural membrane, and activation of the trigeminal pain pathway for tumors with cavernous sinus invasion [5, 8]. The latter in NFPA is considered a risk factor for headache, which has been shown to be homolateral to the site of invasion of the cavernous sinus [7].

At time of diagnosis of macro-NFPA, 60–85% of patients present at least one pituitary deficiency. The risk of hypopituitarism correlates with tumor volume. Indeed, mechanical compression of the normal anterior pituitary gland or pituitary stalk hampering stimulatory hypothalamic factor delivery can result in partial or complete hypopituitarism. Partial hypopituitarism prevalence ranges from 37% to 85%; panhypopituitarism, instead, is found in 6–29% of patients with a

NFPA. Growth hormone (GH) deficiency and central hypogonadism have been reported to be the most common defects detected in NFPA patients, followed by corticotropin and thyrotropin deficiencies. Available data are sparse for microade-nomas that can eventually lead to pituitary dysfunction [9].

Hyperprolactinemia is also a common finding in NFPA patients. Its prevalence in patients with histologically proven NFPA is 25–65% [10]. This "disconnection hyperprolactinemia" or nontumoral hyperprolactinemia is usually mild (<95 ng/mL) and is caused by pituitary stalk compression, which hampers dopamine delivery to the anterior pituitary (stalk effect) [11].

The presence of diabetes insipidus (DI) at the time of clinical presentation is very rare in NFPA [5].

Patients with NFPA may rarely present with pituitary apoplexy, an acute clinical syndrome characterized by sudden development of headache, vomiting, altered consciousness, visual abnormalities, and hypopituitarism in the context of hemorrhagic infarction of a pituitary adenoma. Pituitary apoplexy occurs in 5–20% of all pituitary tumors and appears to be more common in NFPA than in hormonally active lesions [12].

Rarer clinical manifestations are rhinorrhea, in the case of tumors that cause erosion of the sellar floor and extend inferiorly toward the sphenoid sinus, and occlusion of the internal carotid artery in the case of tumors extending to the parasellar region.

13.3.2 Hormonal Evaluation

Once a pituitary tumor is discovered, pituitary function should be tested looking either for a hyper- or hypopituitarism. All anterior pituitary axes should be evaluated, and when a hormonal hypersecretion is excluded, the tumor could be clinically defined as NFPA [3, 10].

However, hyperprolactinemia could be present at diagnosis due to pituitary stalk compression, and as outlined before, prolactin (PRL) levels are usually much lower as compared to "true" prolactinomas [13]. In addition, in the case of a large tumor with slightly elevated PRL levels and hyperprolactinemia symptoms, the diagnosis of macroprolactinoma should be taken into account due to the possibility of a "hook effect" in PRL level assessment. This laboratory artifact is due to high PRL levels that hinder the antibody–antigen–antibody sandwich complex formation. Serum dilution of the sample could avoid this bias. On the other hand, very high PRL levels in the absence of clinical hyperprolactinemia symptoms should prompt polyethylene glycol (PEG) precipitation to screen for macroprolactin interference [11, 14].

It is mandatory that all NFPA patients are investigated for pituitary hormone deficiencies, in order to promptly initiate hormone replacement therapy, especially in case of adrenal and thyroid insufficiency [10]. Very low 8–9 AM cortisol levels may be sufficient to diagnose central adrenal insufficiency; nevertheless, a dynamic test (Synacthen's test or insulin tolerance test, depending on general health status) is recommended. Stimulation tests are not required for the evaluation of other pituitary axes [11]. Low FT4 levels associated with low, inappropriately normal, or

mildly elevated TSH levels are consistent with central hypothyroidism. Hypogonadotropic hypogonadism is diagnosed by the presence of low serum testosterone levels associated with low or inappropriately normal LH/FSH levels, and hypogonadism symptoms in males. In premenopausal females presenting oligomenorrhea or amenorrhea, plasma FSH, LH, and E2 should be measured; low FSH and LH levels are consistent with central hypogonadism in post-menopausal women [15]. Finally, IGF1 concentration below gender- and age-specific lower limit of normal is consistent with GH deficiency, especially when three pituitary axis are compromised [11]. However, screening for GH deficiency is not recommended at diagnosis [9].

Regular monitoring of urine volume and serum sodium is necessary to identify hyponatremia and/or DI [13].

There is no evidence supporting the measurement of chromogranin A or routine genetic testing in patients with sporadic NFPA [10, 14].

13.3.3 Radiological Diagnosis

Gadolinium contrast MRI is the gold standard for sellar region evaluation since it provides high-resolution images of sellar contents and surrounding structures. On the other hand, CT is the preferred imaging technique for hospital emergencies or for surrounding bone structure evaluation. Thus, neuroradiological assessment of suspected NFPA is based on MRI, which should be performed according to specific criteria. In particular, MRI protocol should include:

- thin (≤3 mm) slices, high matrix, sagittal + coronal T1-weighted sequences with and without gadolinium injection;
- coronal 3D volume assessment with reconstruction;
- T2-weighted coronal slices;
- volume acquisition; otherwise, very thin slices are needed [9].

In T1-weighted images, adenomas can appear hypo- or isointense compared to nontumoral pituitary tissue with poor or absent gadolinium uptake. In T2-weighted images, adenomas appear isointense compared to the white matter. In the presence of tumor bleeding, such as pituitary apoplexy, hemorrhage appears as hyperintense in T1-weighted images without contrast. This characteristic hyperintensity may be absent in the early stage because hemorrhage is still in the form of deoxyhemoglobin. Hyperintensity of the optic chiasm on T2-weighted images can indicate a compromised visual function, which may persist even after prompt surgical removal of the pituitary adenoma [5].

The extension of the lesion should be determined:

- superiorly, toward the optic pathways;
- laterally, in the cavernous sinuses,
- inferiorly, in the sphenoidal sinus and posteriorly, toward the clivus.

The Knosp classification systems are currently mostly used for radiological classification, having a clinical and prognostic value. Knosp offered a grading system for predicting invasion of the cavernous sinus by pituitary macroadenomas based on MRI, describing tumor parasellar extension, which is a negative prognostic factor for surgical outcome. Briefly, the more laterally an adenoma grows and surrounds the internal carotid artery, the higher its grade level is, with grade 0 corresponding to an adenoma without any parasellar extension and grade 4 to the total encasement of the intracavernous carotid artery [13, 16]. There is insufficient evidence to indicate the use of MR spectroscopy, perfusion, positron emission tomography (PET), and single-photon emission computed tomography (PECT). Gradient echo and MR perfusion are promising tools to evaluate cavernous sinus invasion and tumor vascularity [17, 18], but need to be further confirmed.

13.4 Treatment

13.4.1 Surgery

According to recent guidelines issued by the Congress of Neurological Surgeons, surgery is the first-line treatment for symptomatic NFPA [18]. They state that the primary aim of surgery is relieving surrounding neural and vascular structures from pituitary tumor compression. Thus, visual abnormalities, optic nerves, or chiasm impairment, neurological deficit, and pituitary apoplexy with visual involvement are indicators for surgery. Even if a total tumor resection is ideal, an aggressive approach could compromise surrounding vital structures or damage healthy pituitary gland and its function [13, 19–21]. Indeed, pituitary surgery may also be indicated in the attempt to restore or preserve normal pituitary function in the presence of a large NFPA. However, surgery may not be effective in improving headache and/ or hypopituitarism. Thus, treatment decision should be individualized, based on clinical context, multidisciplinary team (MDT) discussion, and patient preference [20].

Prior to surgery, correcting adrenal insufficiency with glucocorticoid (GC) replacement therapy and/or hypothyroidism with L-thyroxine replacement therapy is mandatory. Furthermore, pretreatment neuro-ophthalmologic evaluation is recommended to define the preoperative baseline and the urgency of surgery and to provide prognostic factors for recovery [19, 20, 22].

To date, evidence is insufficient to define a primary treatment strategy for asymptomatic lesions [18] and a nonemergency surgery should be evaluated case by case. Factors that should be taken into account are (1) patient age; (2) NFPA natural history; (3) risk of VF impairment, correlated with the rate of tumor growth and proximity to the optic pathways; (4) risk of pituitary deficiency development; (5) risk of apoplexy; and (6) risks associated with transsphenoidal surgery. Thus, surgery could be evaluated in asymptomatic pituitary tumors with anatomic signs of impending visual loss and in young patients with macroadenomas considering the higher life-time probability of tumor growth [19, 20]. In other cases, a conservative approach

has been proposed [11]. In microadenomas, surgery is not indicated due to their slow progression [21]. Fernandez-Balsells et al. reported a greater tendency for tumor growth in macroadenomas in comparison with microadenomas (12.53 vs. 3.32 per 100 patient-years) [23].

TSA approach is the technique of choice for pituitary tumor removal and can be performed under endoscopic or microscopic visualization. No evidence confirms the superiority of one on the other, and the choice among them depends on surgeon's preference and skills. Microscopy offers a small vision and a limited field illumination with the risk of collision with other instruments. On the other hand, endoscopic approach decreases three-dimensional perspective by operating through a two-dimensional image but improves illumination and visualization of surgical field [19, 24]. The combined surgical strategy of TSA and transcranial approaches is recommended for invasive NFPA with significant suprasellar, frontal, or temporal extension. Surgery is defined as a safe and effective approach also in American Society of Anesthesiologists grade 1 to grade 3 elderly patients, as reported in recent guidelines [24]. Surgical indications are the same in older as compared to younger patients, but the higher comorbidity incidence in older patients should be taken into account. Thus, a careful anesthesia risk assessment is fundamental to guarantee the same mortality of the general population. Surgical approach should preferably be transsphenoidal, to lower the risk of complications. Some studies reported a longer hospitalization due to water and electrolyte disorders in older patients (10% vs. 6% of the general population), while other studies did not find any difference. Improvement of visual symptoms and pituitary deficiencies is comparable to that of younger patients. In young women with macroadenoma and no visual impairment, a pregnancy wish should be taken into account in the pituitary tumor management. Adenomectomy often recovers central hypogonadism due to disconnection hyperprolactinemia, but if hypogonadism is caused directly by tumor mass, it is effective only in about 30% of cases. Furthermore, lactotroph cell hyperplasia increases pituitary volume during pregnancy; therefore, any suprasellar extension toward optical structures must be taken into account. If the tumor mass is diagnosed during pregnancy, clinical, radiological (MRI without contrast enhancement), and ophthalmologic surveillance should be proposed [9, 20, 25].

Surgical treatment is a safe procedure with adequate preoperative counseling. Surgery may be effective in symptom relief and is associated with low morbidity and mortality [19]. In a recent meta-analysis, TSA approach in NFPA patients was associated with 1% mortality [26]. Complication incidence is lower in hospitals with skilled surgeons, occurring in $\leq 5\%$ of patients. The most common complications are cerebrospinal fluid leakage, fistula, meningitis, vascular injury, persistent DI, visual deterioration, temporary or permanent hypopituitarism, and sinonasal complications (crusting, septal perforation, epistaxis) [15, 19]. The more pituitary tumor is removed, the more symptoms relief is obtained. In a recent postsurgical series of 281 NFPA patients, 89.7% reported headache improvement and 70.1% achieved visual amelioration, while 5.1% had stable vision [19]. Visual recovery may be progressive and is probably correlated with duration and severity of visual field disorders [27]. Multiple retrospective studies showed improved visual function in 75–91% of surgically treated patients and improved hypopituitarism in 35–50% of patients. However, surgery itself may worsen hypopituitarism in 10% of patients [20].

Pituitary surgery allows a gross total resection in 60–73% of patients with NFPA [28], but recurrences occur in about 10–20% of completely resected tumors after 5–10 years. Moreover, when residual tumor is left after surgery, tumor growth rate could reach 40% and 50% at 5 and 10 years, respectively [17].

13.4.2 Radiotherapy

Radiation therapy (RT) is an effective treatment for patients with NFPA unsuccessfully treated by surgery, resulting in local control of 90–95% of patients at 5–10 years [29]. Several RT techniques are currently available, including fractionated conformal RT, stereotactic radiosurgery (SRS), and fractionated stereotactic RT.

Fractionated conformal RT delivers high-energy photons, with a total dose between 45 and 50 Gy, fractionated in 25 sessions of 1.8–2 Gy [9].

Stereotactic techniques, such as SRS or fractionated stereotactic RT, have been developed with the aim of delivering more localized irradiation and reducing long-term side effects, maximizing sparing of adjacent healthy structures. Stereotactic techniques represent the most commonly used radiation techniques in the treatment of NFPA [29]. Radiosurgery delivers radiation in a single session by using Gamma Knife and linear accelerator (LINAC). This type of RT is feasible only if the target volume is clearly defined, small (<2–3 cm on the long axis), and sufficiently remote from the optic pathways to ensure <8 Gy irradiation to the chiasm and optic nerves.

Fractionated stereotactic RT takes advantage of the ballistic precision and multiple beam entries of radiosurgery applied to healthy tissue radioprotection by fractioning. Hypofractionated stereotactic RT (in 3–5 fractions mainly, delivering a total dose of 20–30 Gy) may also be proposed [9].

The most frequent side effect of RT is the occurrence of anterior pituitary dysfunction, which develops in 5-40% of the cases and up to 80% of the patients over a 10-year period [30].

Since the optic chiasm is the most radiation-sensitive structure, another potential, but rare, side effect of RT is optic neuropathy, which presents by an acute vision loss. This complication has been attributed to microvascular obliteration of the visual tract and is directly related to the radiation dose and the proximity of the tumor to the optic chiasm; an advanced age and the concomitant presence of diabetes mellitus appear to increase the risk of visual side effects [31]. Vascular complications such as stroke have been reported with conventional external RT but not with fractionated stereotactic RT [32]. In the long term, older RT techniques have been reported to associate with the development of secondary brain tumors, such as gliomas, glioblastomas, sarcomas, or meningiomas, even 20 years after irradiation. However, this increased risk is rare and was not confirmed in all studies [31].

In general, side effects of risk are lowered by reducing the dose to healthy tissues. Therefore, thanks to the new stereotactic techniques side effects that are becoming less frequent.

The choice of RT technique is made according to adenoma size, proximity to the visual pathways, potential risk to healthy surrounding tissues, and local availability. If available, SRS is preferred when the adenoma maximal diameter is <3 cm and is distant at least 2 mm from the optic chiasm (Minniti 2016 Rep Pract Oncol Radiother) to allow a <8 Gy irradiation to the optic chiasm and nerves [33]. On the contrary for large lesions involving the optic apparatus, conventionally fractionated RT is advised [29].

The role of RT in the postoperative management of NFPA is still debated. Indeed, RT efficacy in reducing local tumor growth is indisputable, whatever the procedure used. Postoperative RT reduces NFPA recurrence rate from 40% in nonirradiated patients to <10% in those who have been irradiated [4]. On the other hand, the possible side effects must be taken into account. In addition, there are no randomized controlled trials showing the superiority of RT compared to a conservative approach after surgery for NFPA management. Therefore, at present, there is no consensus concerning the systematic use of RT in the postoperative period for patients with incompletely resected NFPA [34]. In most cases, regular surveillance can be proposed, postponing RT until recurrence is confirmed [9, 35].

Adjuvant RT may also be considered for patients who, at diagnosis, already present aggressive tumors, such as those invading parasellar structures or with high Ki-67 labeling index, a proliferative marker significantly associated with NFPA recurrence [36]. There is no consensus concerning the optimal timing for adjuvant radiotherapy. The risk of side effect development increases with longer time since RT, suggesting to postpone this intervention until tumor regrowth, since late RT seems as effective as early treatment in terms of local control [37].

In summary, systematic postoperative RT is not indicated following complete resection. In case of tumor remnant after initial surgery, the beneficial effects of RT need to be weighed against the other possible alternatives, such as wait and scan and reoperation, which is advised to be discussed within an experienced multidisciplinary pituitary team. However, RT and in particular stereotactic irradiation remain an important pillar in the management of patients with residual or recurrent NFPA, due to its efficacy in inhibiting tumor growth and reducing tumor volume [37].

Clearly, a short- and long-term assessment of the risk-benefit profile is always required, considering the patient's age, clinical history, and comorbidities. When considering patients already presenting with hypopituitarism, the occurrence of pituitary dysfunction after RT carries less concern, which may shift the benefit/risk ratio. The occurrence of optic neuropathy needs to be counterbalanced by the potential visual improvement that can be observed due to reduction in tumor volume.

Another type of RT approach is represented by proton beam radiotherapy, which takes advantage of the physical properties of protons, offering better dose distribution and normal tissue preservation as compared to photons. However, at present, the superiority of protons over photons in terms of long-term efficacy and toxicity remains unsubstantiated [29]. Furthermore, this therapeutic approach is not widely available.

There are several case reports describing the use of peptide receptor radionuclide therapy (PRRT) in Pit-NET. Some patients may respond to this treatment modality, but its overall efficacy cannot be established based on the small available clinical experience [38].

13.4.3 Medical Therapy

The recent Guidelines by the Congress of Neurological Surgeons state that there is no clinical evidence supporting medical therapy as primary treatment for NFPA. Indeed, dopamine agonists (DAs) and somatostatin analogs (SAs) demonstrated inconsistent tumor response rates (12–40% and 0–61%, respectively) [5]. However, several studies focused on a potential DA therapeutic role in the medical management of NFPA. Dopamine 2 receptors (DR2) are expressed on all anterior pituitary cell types, including NFPA cells [39-41]. Some authors demonstrated a positive effect of cabergoline on tumor shrinkage, which was evident after a 6-month treatment [42, 43]. Greenman et al. reported the largest series of postoperative NFPA patients with tumor remnants treated with DA. They compared the outcome between patients receiving DA therapy immediately after surgery and those who started therapy after tumor growth confirmation. Tumor control was achieved in 87% and 58% of cases, respectively. Thus, they suggested that DA therapy might avoid additional invasive treatment such as surgery and RT [44]. Young age, male gender, and a larger preoperative tumor size were associated with a lower response rate to DA treatment [38]. Cabergoline therapy is usually started with 0.5 mg weekly, increasing 0.5 mg each week until a maximum dose of 3 mg weekly [11]. In patients resistant to standard dose, higher doses did not prevent tumor progression. With regard to cardiac safety, reliable data on valvulopathy development in NFPA are not available. Periodic echocardiogram probably should be reserved only to patients taking >3 mg/week of cabergoline for prolonged periods [38, 44]. In conclusion, even if evidence is contrasting, recent data seem to suggest cabergoline as a safe and cheap alternative in cases where surgery is contraindicated, in patients refusing surgery and as adjuvant therapy in patients with postsurgical residual tumors. Further randomized controlled trials are needed to identify the optimal clinical scenario in which DA could be used [38, 39, 42]. NFPAs express also different somatostatin receptor (SSTR) subtypes, but their significance is unclear [39, 45]. Somatostatin receptor scintigraphy is nonpredictive of SA therapy response in NFPA. The significant antiproliferative and antisecretory effects of somatostatin on NFPA cells demonstrated in vitro were not confirmed in vivo. Further long-term well-designed studies are needed to better define SA treatment for NFPA tumor control [38]. To date, pasireotide demonstrated positive effects in vitro, but in vivo trials are still not available. Combination therapy with DA and SA did not show better efficacy [17].

Finally, temozolomide (TMZ) is an alkylating chemotherapeutic drug proposed by the European Society of Endocrinology Guidelines as first-line chemotherapy in aggressive pituitary tumors showing progression [46]. Low expression of O-methylguanine DNA methyltransferase (MGMT) correlates with a better response rate [11]. In progressive and regrowing NFPA, low MGMT expression was found, suggesting TMZ as an alternative treatment after conventional therapy failure [47].

13.5 Follow-up

Recent Guidelines and Expert's Consensus offer some recommendations about posttreatment NFPA follow-up, even if there is no high-grade level of evidence defining duration and frequency [9, 14, 48, 49]. NFPA recurrence rate was reported to be 15-66% for surgery alone and 2-28% for surgery followed by RT; therefore, a longterm radiologic follow-up of at least 10 years is reasonable [35, 50]. A postsurgical radiological tissue or tumor remnant and younger age are risk factors associated with recurrence. In the early postoperative period, signs and symptoms of water balance disorders (diabetes insipidus DI, syndrome of inappropriate ADH secretion [SIADH], and cerebral salt-wasting syndrome) and hypocortisolism should be promptly detected by measuring plasma electrolytes, urine specific gravity, water balance, osmolality, and early morning cortisol levels [13, 35, 48]. Central DI is a condition characterized by an inappropriate hypotonic polyuria (urine output >3 1/24 h and urine osmolality <300 mOsm/kg) associated with a high or normal serum sodium [48]. It may occur 12–48 hours after surgery and is generally due to a transient dysfunction of ADH-secreting neurons [13, 48], but it could persist in 2–7% of patients. Risks factors for permanent DI are young age, male gender, large intrasellar masses and postoperative CSF leaking, preoperative DI, and repeated pituitary surgery [48]. Sometimes (3-4% of cases), postsurgical water-electrolyte imbalance could present with a triphasic trend: DI occurs in the first 5-7 days, followed by an uncontrolled release of ADH (SIADH phase) lasting up to 2 weeks, and a final condition of permanent DI [13]. Immediate postsurgical DI should be treated with vasopressin with frequent retesting of response to avoid ADH administration during SIADH phase or after DI resolution [15, 48]. SIADH, instead, leads to a condition of euvolemic hyponatremia that should be treated in the first place with fluid intake restriction in the milder cases (Na 134-125 mmol/l), or with hypertonic saline or ADH receptor antagonists in more severe cases (Na < 125 mmol/l). Rapid decreases in serum Na and/or symptomatology (headaches, nausea, vomiting) also affect the action plan.

More rarely, a postsurgical hypovolemic hyponatremia may be due to a cerebral salt-wasting syndrome mediated by brain natriuretic peptide action [48]. As concerns the risk of secondary adrenal insufficiency, corticosteroid supplementation is recommended in the perioperative period in patients showing preoperative or immediate postoperative (day 2) hypocortisolemia [49]. Nevertheless, in clinical practice, the management of NFPA patients with normal preoperative HPA axis is still controversial. Prophylactic GC coverage is also controversial: Available data are not

clear concerning the efficacy of this approach in all patients, specifically in patients found to be adrenal insufficient after surgery [48, 51-53]. Since hypoadrenalism is a life-threatening condition, Prete et al. proposed hydrocortisone e.v. therapy to all patients in the first 24 h after pituitary surgery (50 mg every 6 h), rapidly tapering to standard oral replacement doses (15-20 mg/daily) during the following 48 hours, in the absence of complications. The same authors consider morning plasma cortisol levels (assessed before oral GC therapy) tested 1 week after surgery, as predictive of HPA axis integrity: Cortisol levels $<5 \mu g/dl$ are consistent with secondary adrenal insufficiency, whereas cortisol levels >15 µg/dl should rule out adrenal insufficiency. Intermediate values need to be confirmed with dynamic provocative tests after at least 6 weeks from surgery [48]. In case of persisting deficit, pituitary function should be assessed at least 3 months postoperatively, to screen for late recovery [9]. The insulin tolerance test (ITT) is considered the gold standard to evaluate HPA axis integrity. However, this test may cause serious side effects and is contraindicated in specific patients (older patients, patients with a history of epilepsy or ischemic heart disease). As an alternative, high-dose (250 μ g) or low-dose (1 μ g) Synacthen test can be proposed, but there is no evidence to date supporting one or the other dose. During the first year, after surgery regular evaluation of morning cortisol should be recommended [13]. Retesting of all pituitary axis (thyrotroph, gonadotroph, and somatotropic) should be completed 4-6 weeks after surgery, unless acute complications occur [15, 48]. If a preexisting deficiency is confirmed or it is newly diagnosed, pituitary function should be reassessed in order to adapt replacement therapy [9]. GH deficiency treatment should be evaluated case by case, also taking age into account, considering that, to date, there is no evidence supporting tumor remnant growth promotion or recurrence induction by rhGH replacement therapy in patients with NFPA treated exclusively by surgery. If pituitary function is conserved, re-evaluation should be performed only in case of recurrence/residual tumor progression or after adjuvant RT. In the latter case, pituitary function should be explored at least once yearly for at least 10 years [14].

Generally, first postoperative MRI is performed 3-6 months after surgery and the second one after 12 months. The acquired images are considered as reference for subsequent follow-up [9, 13, 49], which needs to be individualized. In the absence of residual tumor, MRI is suggested to be repeated annually for 5 years and then at 7, 10, and 15 years. If no clinical signs, residual disease, or suspect images are present, radiological follow-up may be terminated. In case of adenomatous remnant or suspect image, MRI is suggested to be repeated annually for 5 years and then every 2 or 3 years in the absence of progression [9]. In case of recurrence or tumor remnant growth, a multidisciplinary re-evaluation (endocrinologist, neuroradiologist, neurosurgeon, radiotherapist) should discuss tumor morphology and pathology characteristics, patient age, presence of hypopituitarism, and center experience, which are important issues in decision making. Considering patient preference, surveillance or adjuvant therapy may be proposed. Second surgery may be considered an option: (1) when a complete resection is possible; (2) in case of optic pathway compression; and (3) to ensure 3-5 mm safety margins between superior tumoral dome and optic pathway for complementary RT [9].
Ophthalmologic re-evaluation is advised 3 months after surgery and repeated every 6 months when a preoperative visual impairment is found. When the maximum visual function improvement is reached, assessment intervals may be lengthened. Ophthalmological follow-up can stop when first postoperative visit is negative for visual impairment, in the absence of suprasellar remnant threatening the optic pathways. In case of RT, prolonged annual follow-up is mandatory [9, 13].

13.6 Learning Points

- 1. Multimodal therapeutic approaches are needed in NFPA management.
- 2. Differential diagnosis should always be explored.
- 3. NFPA clinical presentation is highly variable, according to mass symptoms.
- 4. Hyperprolactinemia should be accurately evaluated due to possible laboratory bias.
- 5. Hypopituitarism should always be investigated, since it significantly influences further therapeutic management.
- 6. MRI represents the imaging gold standard, together with an expert radiologist.
- 7. Surgery is the first-line treatment for macroNFPA. Accurate patient preparation is mandatory.
- 8. The role of radiation therapy is still debated.
- 9. Post-radiotherapy hypopituitarism may develop after many years.
- 10. Medical approaches are still controversial.
- 11. Long-term follow-up by MRI imaging, endocrine workup, and ophthalmological evaluation is warranted.
- 12. Multidisciplinary team approach is necessary in order to achieve a correct diagnosis and plan an appropriate treatment.
- 13. Patient QoL must be always taken care of when considering NFPA management and follow-up.

Consent to Publish: All authors grant Springer specific permission to publish the work ("consent to publish").

Questions and Answers

Q: When should we suspect the presence of a nonfunctioning pituitary adenoma?

A: The absence of endocrine symptoms often delays a diagnosis, until mass effects reveal neurological symptoms, headache, visual disturbances, or secondary endocrine symptoms. The most common symptoms are visual defects and head-ache, possibly associated with hyperprolactinemia (and related hypogonadism), or pituitary deficiencies. Diabetes insipidus is rare and rhinorrhea even rarer.

Q: Which are the recommended tests to establish the diagnosis of a nonfunctioning pituitary adenoma?

A: Sellar MRI is the gold standard for diagnosis, interpreted by an expert radiologist. Hyperprolactinemia should be accurately evaluated to exclude the presence of a high-dose hook effect or macroprolactinemia.

Q: Which is the optimum treatment for nonfunctioning pituitary adenomas?

A: Transsphenoidal tumor resection is the first-line treatment for nonfunctioning pituitary macroadenomas (diameter > 1 cm), in order to relieve mass symptoms. Surgery may be associated with the development of hormonal deficiencies in the short term.

Q: After surgery, how is hypopituitarism diagnosed?

A: HPA axis should be assessed immediately after surgery. Pituitary function should be assessed within 4–6 weeks after surgery and at least 3 months postoperatively. The insulin tolerance test (ITT) is considered the gold standard to evaluate HPA axis integrity. However, this test is contraindicated in specific patients (older patients, patients with a history of epilepsy or ischemic heart disease). Synacthen test can be proposed as alternative. During the first year after surgery, regular evaluation of morning cortisol should be recommended. If a preexisting deficiency is confirmed or it is newly diagnosed, pituitary function should be reassessed in order to adapt replacement therapy. GH-deficit treatment should be performed only in cases of recurrence/residual tumor progression or after adjuvant RT.

Q: *How long is follow-up and monitoring of a nonfunctioning pituitary adenoma?* **A:** There is no firm evidence defining duration and frequency of follow-up for patients with nonfunctioning pituitary adenomas. Recurrence rate is significant (either after surgery alone or after surgery + RT); therefore, a long-term radiological and clinical follow-up of at least 10 years is advisable. In addition, hormonal deficiencies occurring after initial treatment should be supplemented and retested in order to detect pituitary function recovery, which, although rare, may occur.

References

- Molitch ME. Nonfunctioning pituitary tumors. In: Handbook of clinical neurology [internet]. Elsevier; 2014. p. 167–84. https://linkinghub.elsevier.com/retrieve/pii/ B9780444596024000125 [cited 2019 Dec 26].
- Vasilev V, Rostomyan L, Daly AF, Potorac I, Zacharieva S, Bonneville J-F, et al. Management of endocrine disease: Pituitary 'incidentaloma': neuroradiological assessment and differential diagnosis. Eur J Endocrinol. 2016;175(4):R171–84.
- Freda PU, Beckers AM, Katznelson L, Molitch ME, Montori VM, Post KD, et al. Pituitary incidentaloma: an endocrine society clinical practice guideline. J Clin Endocrinol Metab. 2011;96(4):894–904.
- Mercado M, Melgar V, Salame L, Cuenca D. Clinically non-functioning pituitary adenomas: pathogenic, diagnostic and therapeutic aspects. Endocrinol Diabetes Nutr. 2017;64(7):384–95.
- Ntali G, Wass JA. Epidemiology, clinical presentation and diagnosis of non-functioning pituitary adenomas. Pituitary. 2018;21(2):111–8.
- Jahangiri A, Lamborn KR, Blevins L, Kunwar S, Aghi MK. Factors associated with delay to pituitary adenoma diagnosis in patients with visual loss. J Neurosurg. 2012;116(2):283–9.

- 7. Yu B, Ji N, Ma Y, Yang B, Kang P, Luo F. Clinical characteristics and risk factors for headache associated with non-functioning pituitary adenomas. Cephalalgia. 2017;37(4):348–55.
- Greenman MDY, Melmed MDS. Diagnosis and management of nonfunctioning pituitary tumors. Annu Rev Med. 1996;47(1):95–106.
- 9. Chanson P, Raverot G, Castinetti F, Cortet-Rudelli C, Galland F, Salenave S. Management of clinically non-functioning pituitary adenoma. Ann Endocrinol. 2015;76(3):239–47.
- Fleseriu M, Bodach ME, Tumialan LM, Bonert V, Oyesiku NM, Patil CG, et al. Congress of Neurological Surgeons systematic review and evidence-based guideline for pretreatment endocrine evaluation of patients with nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E527–9.
- 11. Drummond JB, Ribeiro-Oliveira A, Soares BS. Non-functioning pituitary adenomas. In: Feingold KR, Anawalt B, Boyce A, Chrousos G, Dungan K, Grossman A, et al., editors. Endotext [internet]. South Dartmouth, MA: MDText.com, Inc.; 2000. [cited 2019 Dec 26]. http://www.ncbi.nlm.nih.gov/books/NBK534880/.
- Capatina C, Inder W, Karavitaki N, Wass JAH. Management of endocrine disease: pituitary tumour apoplexy. Eur J Endocrinol. 2015;172(5):R179–90.
- Esposito D, Olsson DS, Ragnarsson O, Buchfelder M, Skoglund T, Johannsson G. Nonfunctioning pituitary adenomas: indications for pituitary surgery and post-surgical management. Pituitary. 2019;22(4):422–34.
- Raverot G, Assié G, Cotton F, Cogne M, Boulin A, Dherbomez M, et al. Biological and radiological exploration and management of non-functioning pituitary adenoma. Ann Endocrinol. 2015;76(3):201–9.
- Fleseriu M, Hashim IA, Karavitaki N, Melmed S, Murad MH, Salvatori R, et al. Hormonal replacement in hypopituitarism in adults: an Endocrine Society Clinical Practice Guideline. J Clin Endocrinol Metab. 2016;101(11):3888–921.
- Knosp E, Steiner E, Kitz K, Matula C. Pituitary adenomas with invasion of the cavernous sinus space: a magnetic resonance imaging classification compared with surgical findings. Neurosurgery. 1993;33(4):610–8.
- Delgado-López PD, Pi-Barrio J, Dueñas-Polo MT, Pascual-Llorente M, Gordón-Bolaños MC. Recurrent non-functioning pituitary adenomas: a review on the new pathological classification, management guidelines and treatment options. Clin Transl Oncol. 2018;20(10):1233–45.
- Lucas JW, Bodach ME, Tumialan LM, Oyesiku NM, Patil CG, Litvack Z, et al. Congress of Neurological Surgeons systematic review and evidence-based guideline on primary management of patients with nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E533–5.
- 19. Penn DL, Burke WT, Laws ER. Management of non-functioning pituitary adenomas: surgery. Pituitary. 2018;21(2):145–53.
- Castinetti F, Dufour H, Gaillard S, Jouanneau E, Vasiljevic A, Villa C, et al. Non-functioning pituitary adenoma: when and how to operate? What pathologic criteria for typing? Ann Endocrinol. 2015;76(3):220–7.
- Dekkers OM, Hammer S, de Keizer RJW, Roelfsema F, Schutte PJ, Smit JWA, et al. The natural course of non-functioning pituitary macroadenomas. Eur J Endocrinol. 2007;156(2):217–24.
- 22. Newman SA, Turbin RE, Bodach ME, Tumialan LM, Oyesiku NM, Litvack Z, et al. Congress of Neurological Surgeons systematic review and evidence-based guideline on Pretreatment ophthalmology evaluation in patients with suspected nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E530–2.
- Fernández-Balsells MM, Murad MH, Barwise A, Gallegos-Orozco JF, Paul A, Lane MA, et al. Natural history of nonfunctioning pituitary adenomas and Incidentalomas: a systematic review and Metaanalysis. J Clin Endocrinol Metab. 2011;96(4):905–12.
- 24. Kuo JS, Barkhoudarian G, Farrell CJ, Bodach ME, Tumialan LM, Oyesiku NM, et al. Congress of Neurological Surgeons systematic review and evidence-based guideline on surgical techniques and technologies for the management of patients with nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E536–8.
- 25. Hong J, Ding X, Lu Y. Clinical analysis of 103 elderly patients with pituitary adenomas: Transsphenoidal surgery and follow-up. J Clin Neurosci. 2008;15(10):1091–5.

- 26. Murad MH, Fernández-Balsells MM, Barwise A, Gallegos-Orozco JF, Paul A, Lane MA, et al. Outcomes of surgical treatment for nonfunctioning pituitary adenomas: a systematic review and meta-analysis: surgical treatment for nonfunctioning pituitary adenomas. Clin Endocrinol. 2010;73(6):777–91.
- 27. Lithgow K, Batra R, Matthews T, Karavitaki N. Management of endocrine disease: visual morbidity in patients with pituitary adenoma. Eur J Endocrinol. 2019;181(5):R185–97.
- Yu S-Y, Du Q, Yao S-Y, Zhang K-N, Wang J, Zhu Z, et al. Outcomes of endoscopic and microscopic transphenoidal surgery on non-functioning pituitary adenomas: a systematic review and meta-analysis. J Cell Mol Med. 2018;22(3):2023–7.
- 29. Minniti G, Flickinger J, Tolu B, Paolini S. Management of nonfunctioning pituitary tumors: radiotherapy. Pituitary. 2018;21(2):154–61.
- Cortet-Rudelli C, Bonneville J-F, Borson-Chazot F, Clavier L, Coche Dequéant B, Desailloud R, et al. Post-surgical management of non-functioning pituitary adenoma. Ann Endocrinol. 2015;76(3):228–38.
- Ntali G, Karavitaki N. Efficacy and complications of pituitary irradiation. Endocrinol Metab Clin N Am. 2015;44(1):117–26.
- 32. Rim CH, Yang DS, Park YJ, Yoon WS, Lee JA, Kim CY. Radiotherapy for pituitary adenomas: long-term outcome and complications. Radiat Oncol J. 2011;29(3):156.
- Pomeraniec IJ, Dallapiazza RF, Xu Z, Jane JA, Sheehan JP. Early versus late gamma knife radiosurgery following transsphenoidal resection for nonfunctioning pituitary macroadenomas: a matched cohort study. J Neurosurg. 2016;125(1):202–12.
- Wildemberg LE, Glezer A, Bronstein MD, Gadelha MR. Apoplexy in nonfunctioning pituitary adenomas. Pituitary. 2018;21(2):138–44.
- 35. Tampourlou M, Ntali G, Ahmed S, Arlt W, Ayuk J, Byrne JV, et al. Outcome of nonfunctioning pituitary adenomas that regrow after primary treatment: a study from two large UK centers. J Clin Endocrinol Metab. 2017;102(6):1889–97.
- Minniti G, Clarke E, Scaringi C, Enrici RM. Stereotactic radiotherapy and radiosurgery for nonfunctioning and secreting pituitary adenomas. Rep Pract Oncol Radiother. 2016;21(4):370–8.
- Chanson P, Dormoy A, Dekkers OM. Use of radiotherapy after pituitary surgery for nonfunctioning pituitary adenomas. Eur J Endocrinol. 2019;181(1):D1–13.
- Even-Zohar N, Greenman Y. Management of NFAs: medical treatment. Pituitary. 2018;21(2):168–75.
- 39. Cooper O, Greenman Y. Dopamine agonists for pituitary adenomas. Front Endocrinol. 2018;9:469.
- 40. Pivonello R, Matrone C, Filippella M, Cavallo LM, Di Somma C, Cappabianca P, et al. Dopamine receptor expression and function in clinically nonfunctioning pituitary tumors: comparison with the effectiveness of cabergoline treatment. J Clin Endocrinol Metab. 2004;89(4):1674–83.
- 41. Gagliano T, Filieri C, Minoia M, Buratto M, Tagliati F, Ambrosio MR, et al. Cabergoline reduces cell viability in non functioning pituitary adenomas by inhibiting vascular endothelial growth factor secretion. Pituitary. 2013;16(1):91–100.
- 42. Vieira Neto L, Wildemberg LE, Moraes AB, Colli LM, Kasuki L, Marques NV, et al. Dopamine receptor subtype 2 expression profile in nonfunctioning pituitary adenomas and in vivo response to cabergoline therapy. Clin Endocrinol. 2015;82(5):739–46.
- 43. Garcia EC, Naves LA, Silva AO, de Castro LF, Casulari LA, Azevedo MF. Short-term treatment with cabergoline can lead to tumor shrinkage in patients with nonfunctioning pituitary adenomas. Pituitary. 2013;16(2):189–94.
- 44. Greenman Y, Cooper O, Yaish I, Robenshtok E, Sagiv N, Jonas-Kimchi T, et al. Treatment of clinically nonfunctioning pituitary adenomas with dopamine agonists. Eur J Endocrinol. 2016;175(1):63–72.
- 45. Zatelli MC, Piccin D, Vignali C, Tagliati F, Ambrosio MR, Bondanelli M, et al. Pasireotide, a multiple somatostatin receptor subtypes ligand, reduces cell viability in non-functioning pituitary adenomas by inhibiting vascular endothelial growth factor secretion. Endocr Relat Cancer. 2007;14(1):91–102.

- 46. Raverot G, Burman P, McCormack A, Heaney A, Petersenn S, Popovic V, et al. European Society of Endocrinology Clinical Practice Guidelines for the management of aggressive pituitary tumours and carcinomas. Eur J Endocrinol. 2018;178(1):G1–24.
- 47. Widhalm G, Wolfsberger S, Preusser M, Woehrer A, Kotter MR, Czech T, et al. O 6 -methylguanine DNA methyltransferase immunoexpression in nonfunctioning pituitary adenomas: are progressive tumors potential candidates for temozolomide treatment? Cancer. 2009;115(5):1070–80.
- Prete A, Corsello SM, Salvatori R. Current best practice in the management of patients after pituitary surgery. Ther Adv Endocrinol Metab. 2017;8(3):33–48.
- 49. Ziu M, Dunn IF, Hess C, Fleseriu M, Bodach ME, Tumialan LM, et al. Congress of Neurological Surgeons systematic review and evidence-based guideline on posttreatment follow-up evaluation of patients with nonfunctioning pituitary adenomas. Neurosurgery. 2016;79(4):E541–3.
- Reddy R, Cudlip S, Byrne JV, Karavitaki N, Wass JAH. Can we ever stop imaging in surgically treated and radiotherapy-naive patients with non-functioning pituitary adenoma? Eur J Endocrinol. 2011 Nov;165(5):739–44.
- Jia X, Pendharkar AV, Loftus P, Dodd RL, Chu O, Fraenkel M, et al. Utility of a glucocorticoid sparing strategy in the management of patients following transsphenoidal surgery. Endocr Pract. 2016;22(9):1033–9.
- 52. Tohti M, Li J, Zhou Y, Hu Y, Yu Z, Ma C. Is peri-operative steroid replacement therapy necessary for the pituitary adenomas treated with surgery? A systematic review and meta analysis. Atkin SL, editor. PLoS One. 2015;10(3):e0119621.
- Fridman-Bengtsson O, Höybye C, Porthén L, Stjärne P, Hulting A-L, Sunnergren O. Evaluation of different hydrocortisone treatment strategies in transsphenoidal pituitary surgery. Acta Neurochir. 2019;161(8):1715–21.



Questions and Answers

14

Gianluca Tamagno and Manuel D. Gahete

14.1 Questions

- 1. Why the secretion of pituitary hormones is controlled by a widely and complex landscape of central and peripheral modulators?
- Pituitary tumors are mainly: A. Sporadic
 B. Familiar
- 3. What gene mutation should be considered in a 3-year-old female patient with gigantism/acromegaly?
- 4. Which of the following sentences is true about somatotroph adenoma pathogenesis?
 - A. With the current genetic advances, a predisposing germline mutation is found in the majority of patients harboring somatotroph adenomas.
 - B. Despite advances in genetics, the majority of somatotroph adenomas have only somatic (i.e., nongermline) mutations.
 - C. With current histopathological staining techniques, the majority of somatotroph adenomas coexpress other pituitary markers, such as prolactin, glycoprotein alpha and beta subunits, and ACTH.
 - D. The pure or mixed GH-PRL somatotroph adenomas are polyclonal in origin, indicating pituitary hyperplasia as a pretumoral lesion.

G. Tamagno (🖂)

M. D. Gahete Cell Biology, Physiology and Immunology, University of Córdoba, Cordoba, Spain

© Springer Nature Switzerland AG 2022

G. Tamagno, M. D. Gahete (eds.), *Pituitary Adenomas*, https://doi.org/10.1007/978-3-030-90475-3_14

Hermitage Medical Clinic, Dublin, Ireland

5. What is the cutoff point to be defined as a macroprolactinoma?

- A. 2 cm
- B. 5 mm
- C. 4 cm
- D. 10 mm
- E. None of the above

6. Who should be tested for Cushing's syndrome?

7. Can all the GH-modulating factors be able to act synergistically?

8. Pituitary tumors are:

- A. Monoclonal
- B. Polyclonal

9. What gene mutation should be considered in a 10-month-old child with ACTH-dependent Cushing's syndrome?

- 10. What is the current evidence about cardiac ischemic disease in patients with acromegaly?
 - A. Acromegaly is associated with a decreased Framingham risk and a lower risk of myocardial infarction and cardiac ischemic disease than that of the general population.
 - B. Acromegaly is associated with an increased Framingham risk but no increase in myocardial infarction or cardiac ischemic disease compared to the general population.
 - C. Acromegaly is associated with increased Framingham risk and consequently with a higher risk of myocardial infarction and cardiac ischemic disease than that of the general population.
 - D. Despite a lower Framingham risk, patients with acromegaly have an increased risk of myocardial infarction and cardiac ischemic disease than the general population.

11. What pituitary cells do prolactinomas arise from?

- A. Somatotrophs
- B. Lactotrophs
- C. Gonadotrophs
- D. Thyrotrophs
- E. Corticotrophs

12. Once hypercortisolism is confirmed, how Cushing's disease is identified?

13. Does PRL exert a feedback regulatory loop to modulate its own expression controlling its regulators?

14. Pituitary tumorigenesis is triggered by:

- A. Aberrant hypothalamic factors causing hyperplasia
- B. Disrupted negative feedback from the periphery
- C. Overexpressed growth factors and cytokines
- D. Genetic hit (mutation) transforming a single cell

15. What are genetic causes of 3Pa?

16. Which of the following sentences concerning acromegaly and neoplasms are true?

- A. The prevalence of cancer in cohorts of patients with acromegaly is approximately 20%.
- B. Reinforced cancer screening must be adopted in patients with acromegaly, such as colonoscopy, cervical ultrasound, mammography, and frequent PSA samples.
- C. Following the latest evidence, concern for neoplasms in acromegaly is currently restrained to colorectal and thyroid cancer.
- D. The prevalence of certain cancer subtypes in individuals with acromegaly may be overestimated by systematic screening procedures and physician awareness.

17. What factors might affect basal prolactin value?

- A. Sertraline
- B. Exercise
- C. Pituitary stalk compression
- D. Venipuncture
- E. All the above

18. Which are the recommended initial tests to establish the diagnosis of hypercortisolism or Cushing's syndrome?

19. Is there any relationship between metabolic status and PRL secretion?

- 20. Mouse models of genes involved in cell cycle regulation present with pituitary hyperplasia and tumors; which of these genes are frequently mutated in human pituitary tumors?
 - A. *RB*
 - B. CCNE1 (cyclin E1)
 - C. *CDKN1B* (p27)
 - D. CDKN2C (p18)
 - E. All of the above
 - F. None of the above

21. What are the most common somatic mutations among sporadic pituitary adenomas?

22. Please identify the correct statement about radiating techniques in acromegaly

- A. Radiating techniques are considered third-line treatments because they have been roughly the same over the last decades.
- B. Radiating techniques are considered third-line treatment because of the improvement in surgical techniques over the last decades.
- C. Radiating techniques are considered third-line treatment because of the improvement in medical treatments over the last decades.
- D. Radiating techniques have consistently improved over the last decades and are now considered as first-line treatment along with other treatment approaches.

23. Which imaging modality is of choice in characterizing a prolactinoma?

- A. Ultrasound
- B. CT scan with contrast
- C. Nuclear medicine scans
- D. MRI scan
- E. None of the above

24. Which is the optimum treatment for CD?

25. Does the ghrelin/GHSR1a system modulate the same signaling pathways in all pituitary cells to control hormone secretion?

26. Which gene is mutated in ~40% of sporadic somatotroph tumors?

- A. PRKAR1A
- B. GNAS
- C. GNAI2
- D. GPR101
- E. AIP

27. Identify the correct statements about acromegaly management

- A. Stepwise treatment in acromegaly, according to the latest guidelines, indicates medical treatment as a first-line strategy, followed by selective pituitary surgery and radiotherapy.
- B. Mixed GH-prolactin adenomas may benefit from a first-line medical treatment by dopamine agonist, which may lead to IGF-I normalization and tumor shrinkage.
- C. According to the latest guidelines, the treatment approach in acromegaly is multimodal by delivering selective pituitary surgery, medical treatment, and radiotherapy simultaneously.
- D. First-line medical treatment is preferred in inoperable patients or in those with highly morbid concomitant diseases.

28. Which of the dopamine agonists has higher chances of therapeutic success for the treatment of prolactinomas?

- A. Bromocriptine
- B. Pergolide
- C. Cabergoline
- D. Octreotide
- E. Dopastatin

29. For how long is long-term follow-up and monitoring after remission required in Cushing's disease?

30. Has the administration of opioids had an effect on the OT system?

31. Mutations in the USP8 mutational hotspot can be found in:

- A. Most pituitary tumor types
- B. Only in ACTH-secreting tumors of both pituitary and non-pituitary origin
- C. Only in corticotroph tumors

32. What are the principal side effects of dopamine agonists?

- A. Nausea
- B. Postural hypotension
- C. Mental fogginess
- D. Compulsive eating
- E. All the above
- 33. Melanotrophs and corticotrophs can both synthesize POMC. However, the main end products derived from POMC are different in these cells. How can this phenomenon be explained?
- 34. How many genes are known to be mutated in >5% of sporadic pituitary tumors?
 - A. <5 genes
 - B. Between 5 and 10 genes
 - C. >10 genes

35. In which situations would surgical management be considered as an option in the management of prolactinomas?

- A. Dopamine agonist resistance
- B. Pituitary apoplexy
- C. Both answer A and answer B
- D. Old age
- E. Shrinking microadenoma

36. What is the prevalence of TSH-omas with respect to all pituitary tumors?

- A. 10%
- B. Below 3%
- C. 50%
- D. The same prevalence of GH-omas
- E. TSH-omas are the most frequent pituitary tumors

37. What pituitary cells do TSH-omas arise from?

- A. Somatotrophs
- B. Lactotrophs
- C. Thyrotropes
- D. Gonadotrophs
- E. Corticotropes

38. When diagnosed, most TSH-oma have:

- A. A diameter size <1 cm
- B. A diameter size <1 cm with invasive features
- C. A diameter size >1 cm
- D. A diameter size >1 cm with invasive features
- E. GH cosecretion

39. Which are the classical signs and symptoms of TSH-omas?

- A. Elevated thyroid hormones and elevated or inappropriately normal TSH levels
- B. If diagnosed as a silent adenoma, no clinical or biochemical evidence of central hyperthyroidism
- C. Mass pressure effects
- D. Answers a, b, and c
- E. None of the above

40. Which parameters are useful for the differential diagnosis between TSHoma and PRTH?

- A. SHBG and ICTP levels
- B. FT4 levels
- C. Abnormal thyreotropin response to T3 suppression test
- D. Female gender
- E. All the above

41. Which is the first-line treatment for TSH-omas?

- A. Radiotherapy
- B. Medical therapy with somatostatin analogs
- C. Medical therapy with dopamine analogs
- D. Levo-T4
- E. Pituitary surgery via transsphenoidal route

42. Which is the most sensitive and specific test to document the complete removal of a TSH-oma?

- A. Positive T3 suppression test
- B. Normalization of alpha-GSU levels and alpha-GSU/TSH ratio
- C. Normalization of circulating TSH levels
- D. Normalization of free thyroid hormone levels
- E. Both answers A and D

43. What percentage of anterior pituitary adenomas are nonfunctioning pituitary adenomas?

- A. Lower that 10%
- B. Approximately 50-60%
- C. Approximately 30-40%
- D. Approximately 85%
- E. None of the above

44. Which of the following hereditary syndromes may include nonfunctioning pituitary adenomas?

- A. Multiple endocrine neoplasia type 1 (MEN1) and type 2
- B. Von Hippel-Lindau syndrome
- C. Mc-Cune Albright syndrome
- D. Multiple endocrine neoplasia type 1 (MEN1) and type 4
- E. Paraganglioma syndrome type 1

45. Which of the following transcription factors are currently recommended in routine diagnostics of pituitary adenomas?

- A. Pituitary transcription factor 1 (Pit-1) and steroidogenic factor 1 (SF-1)
- B. T-cell factor/lymphoid enhancer factor (TCF/LEF) transcription factors
- C. Runt-related transcription factor 2 (RUNX2)
- D. Guanine-adenine-thymine-adenine binding protein 2 (GATA) transcription factors
- E. Thyroid transcription factor 1 (TTF1)

46. What is true for null cell adenomas?

- A. Are defined by the lack of immunohistochemical staining of any anterior pituitary hormone
- B. Represent the vast majority of nonfunctioning pituitary adenomas
- C. Are defined by the lack of immunohistochemical staining of any pituitaryspecific transcription factors
- D. A and C
- E. They usually present with aggressive behavior

47. Which of the genes that have been implicated in the pathophysiology of pituitary adenomas, have been more frequently described in nonfunctioning pituitary adenomas?

- A. Aryl hydrocarbon receptor-interacting protein (AIP) gene
- B. Menin 1 gene (MEN1)
- C. Neurofibromatosis type 1 (NF1) gene
- D. Rearranged during transfection (RET) gene
- E. None of the above

48. Which of the following symptoms are most frequently described in nonfunctioning pituitary adenomas

- A. Cerebrospinal fluid rhinorrhea
- B. Obstructive hydrocephalus
- C. Pituitary apoplexy
- D. Headache and visual impairment
- E. All of the above

49. Which of the following are considered criteria for the surgical treatment of nonfunctioning pituitary adenomas?

- A. Age above 50
- B. Patient that are planning to become pregnant
- C. Visual field deficits at presentation
- D. Tumors larger than 10 mm
- E. Extension within the sphenoid sinus

14.2 Answers

- The pituitary is a central gland in the organism that controls a plethora of biological processes important for growth, reproduction, metabolism, stress, etc.; therefore, pituitary cells act as metabolic sensors of the general status of the organisms in order to modulate pituitary secretions and maintain whole-body well-being. To do that, pituitary cells are able to sense and integrate multiple central and peripheral signals and respond by finely modulating pituitary secretions.
- 2. A.
- 3. In early-onset (<5-year-old) gigantism, X-linked acrogigantism should be strongly considered. McCune–Albright syndrome can also cause very young onset GH excess, and single case of *AIP* mutation-positive patient with age of onset of symptoms age 3 has also been described. Childhood-onset GH-secreting adenoma in MEN1 is very rare, but GHRH-secreting pancreas tumors have been described in children.
- 4. B.
- 5. D.

- 6. After excluding iatrogenic CS, it is recommended to test for CS patients if unusual features for age are present (osteoporosis, hypertension) or features suggestive of CS (easy bruising, proximal muscle weakness, reddish-purple striae), children with decreasing height percentile despite obesity, and patients with adrenal adenoma.
- 7. Somatotroph cells are able to sense different GH-modulating factors and integrate all these signals to adapt GH secretion to specific situations of needs. In this sense, it has been shown that GH-modulating factors do not always produce synergistic effects. This can be explained by the fact that many of these factors share several intracellular signaling pathways, which allows the somatotroph cells to integrate all the signals and coordinate their response to the extracellular milieu.
- 8. A.
- 9. In early-onset Cushing's disease (<2-year-old), *DICER1* mutation testing should be performed. Even if the penetrance of pituitary tumors is very low (1%), a pituitary blastoma could be a single and pathognomonic manifestation of DICER1 syndrome.
- 10. B.
- 11. B.
- 12. It is essential to measure ACTH (at least twice). Undetectable ACTH levels or <1.1 pmol/L suggest an adrenal etiology, and ACTH levels >4.4 pmol/L are suggestive of an ACTH-dependent CS. A pituitary MRI is mandatory to assess for a pituitary adenoma. If there is no visible tumor on the MRI or the lesion is <6mm, inferior petrosal sinus sampling is considered the gold standard test to identify a pituitary cause for CS. However, IPSS is not always available and noninvasive tests (high-dose DST, CRH, or desmopressin test) might be also useful.
- 13. It is well known that PRL may upregulate the expression of dopamine, its main negative regulator, triggering a negative feedback loop of its own production. Although this negative feedback loop has not been demonstrated for rest of the PRL regulators, different evidence suggests that this could be also the case for PACAP signaling and other potential factors.
- 14. D.
- 15. The association of Pit-NET and pheochromocytoma/paraganglioma is caused by germline mutation of *SDHx* and *MAX*. As MEN1 syndrome can be associated with pheochromocytoma, this must also be considered in 3Pa. Single cases of *RET* and *VHL* associations have been described, but it is unclear if these are coincidences or truly genetically related.
- 16. C.
- 17. E.
- 18. After excluding exogenous GC administration, it is recommended to use a combination of test with high diagnostic accuracy such as 24-h urinary free cortisol, late-night salivary cortisol, or 1 mg overnight dexamethasone suppression test. A second test should be repeated either UFC, LNSC, or DST to confirm hypercortisolism.

- 19. PRL secretion is strongly influenced by the metabolic status. The most obvious example is body weight. In fact, there is a clear association between obesity and PRL secretion since it has been reported that PRL levels are increased in plasma from obese patients as compared to those from individuals with normoweight. Indeed, these differences have been also observed between rapid gain weight women compared to stable weight women.
- 20. F.
- 21. Somatic *GNAS* mutation is one of the most common genetic causes for isolated somatotroph adenoma. Among sporadic corticotrophinomas, the most frequent cause is *USP8*.
- 22. C.
- 23. D.
- 24. Transsphenoidal selective tumor resection is the first-line treatment for CD. Successful tumor resection leads to low concentrations of ACTH and cortisol and therefore to remission. In case, transsphenoidal surgery cannot be done or is unsuccessful, medical therapy (steroidogenesis inhibitors, tumor-directed therapy, GC receptor antagonists), radiation, or bilateral adrenalectomy might be indicated.
- 25. Ghrelin/GHSR1a interaction has been shown to activate multiple signaling cascades. In the case of somatotroph and corticotroph cells, ghrelin can modulate phospholipase C (PLC), protein kinase C (PKC), PKA, intracellular and extracellular calcium, or MAPK pathways. However, the specific mechanisms involved in the regulation of gonadotrophs remain still unclear, although some reports have suggested that the ghrelin effects on gonadotropin secretion are mediated through nitric oxide, and calcium-dependent and cGMP-independent mechanism in rats.
- 26. B.
- 27. B, D.
- 28. C.
- 29. Cushing's disease patients should be follow-up life-long even if remission is obtained at the first therapy. Recurrences of the disease have been described even after 20 years of initial therapy. Although comorbidities improve after remission of hypercortisolism, most of them are not completely reversible; therefore, monitoring and treatment of glucocorticoid-related comorbidities are of great importance.
- 30. Chronic administration of morphine induced a significant decrease in OT immunoreactivity in the hippocampus, decreased OT mRNA levels within the SON, median eminence, and arcuate nucleus of the hypothalamus, and reduced

brain OT synthesis and plasma OT levels. This general downregulation of the oxytocinergic system following chronic opioid administration, in comparison with the acute stimulatory effects of opioid administration in different brain regions, may be a result of several neuroadaptive changes in the oxytocinergic system caused by chronic exposure to opioids.

- 31. C.
- 32. E.
- 33. Although melanotroph and corticotroph cells produce POMC, the main end product in corticotrophs is ACTH, while in melanotrophs are α -MSH, β -MSH, and γ -MSH. This phenomenon is the result of a differential posttranslational processing, inasmuch as these cell types express different patterns of proteolytic enzymes.
- 34. A.
- 35. C.
- 36. B.
- 37. C.
- 38. D.
- 39. D.
- 40. A.
- 41. E.
- 42. A.
- 43. C
- 44. D
- 45. A
- 46. D
- 47. A
- 48. D
- 49. C