

Chapter 3

Gas Exchange



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The main function of the respiratory system is to remove CO_2 from and add O_2 to systemic venous blood brought to the lung. Tissue demands of O_2 supply and CO_2 removal requires the process of matching perfusion and alveolar ventilation (V_A), diffusion of gases across the alveolar capillary membrane, O_2 delivery (DO_2) and consumption (VO_2). These processes are schematically represented in (Fig. 3.1).

3.1 Alveolar Gas Equation

The total pressure of atmosphere (P_{ATM}) at sea level is 760 torr or mm Hg. P_{ATM} is also sometimes expressed in kilopascal unit. 1 kilopascal is approximately 7.5 torr. P_{ATM} decreases progressively at higher altitude (Table 3.1). The total atmospheric pressure is the sum of pressures exerted by each of its component gases. With increasing altitude, P_{ATM} decreases while the fraction of O_2 (FiO_2) remains constant. At temperature of 37°C (98.6°F), and 100% humidity, water vapor exerts pressure of 47 torr regardless of the altitude. Alveolar air is 100% humidified, therefore the inspired gas is also assumed to be fully saturated with water. To subtract the contribution of water vapor, 47 torr is subtracted from the atmospheric pressure to account for the pressure exerted by gases alone. Our atmosphere contains 20.93% ($\approx 21\%$) oxygen at any altitude. Thus, the fraction of atmosphere

The original version of this chapter was revised with correct equation at Figure 3.6. The correction to this chapter can be found at https://doi.org/10.1007/978-3-030-83738-9_14

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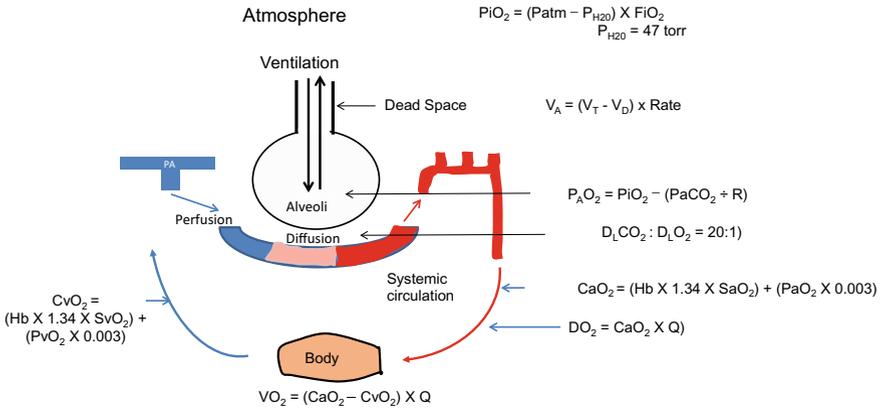


Fig. 3.1 Various factors involved in respiration; Atmospheric composition, Ventilation, Diffusion, Perfusion, Oxygen delivery (DO_2), Oxygen consumption (VO_2), CO_2 production (VCO_2)

Table.3.1 Relationship of barometric pressure and partial pressure of inspired air (PiO_2) when 100% humidified at different altitudes

Altitudes (feet)	P_{ATM} (Torr)	$(P_{ATM}-47[P_{H_2O}])$ Torr	$O_2\%$	PiO_2 (Torr)
0	760	713	20.93	149
600	747	700	20.93	147
5000	632	585	20.93	123
10,000	523	476	20.93	100
15,000	429	382	20.93	80
18,000 ^a	380	333	20.93	70
20,000	349	302	20.93	63
25,000	282	235	20.93	49
30,000	225	178	20.93	37

^aHighest Village. Modified from Comroe JH. Physiology of Respiration, Year Book Medical Publishers, 2nd ED, Chicago, USA 1974

comprising of oxygen (FiO_2) is 0.21. Partial pressure of oxygen in inspired gas (PiO_2) is calculated as:

$$PiO_2 = (P_{atm} - 47) \times FiO_2$$

At sea level, $PiO_2 = (760-47) \times 0.21 = 149$ torr. When breathing 40% O_2 at sea level, $PiO_2 = (760 - 47) \times 0.4 = 285$ torr. At higher altitudes, breathing the same FiO_2 results in a lower PiO_2 . In Denver (altitude 5,000 feet, $P_{ATM} = 632$ mmHg) for example, breathing FiO_2 of 0.21 will result in $PiO_2 = (632 - 47) \times 0.21 = 123$ torr and at FiO_2 of 0.4, it will be $(632 - 47) \times 0.4 = 234$ torr.

3.2 Oxygenation and Ventilation

The amount of air moved in and out of the lungs every minute (V_T x respiratory rate) is termed minute volume. Part of the inspired V_T occupies conducting airways (anatomic dead space) which does not contribute to gas exchange. Still another part of V_T enters alveoli that are not sufficiently perfused (alveolar dead space). Total dead space (V_{Dtot}) is the sum of anatomic dead space (V_{Danat}) and alveolar dead space (V_{Dalv}). Alveolar ventilation (V_A) is calculated as:

$$\dot{V}_A = [V_T - (V_{D_{anat}} + V_{D_{alv}})] \times RR$$

where RR is the respiratory rate (Fig. 3.2).

Although dead space is often looked at as moving in bulk, in reality the gas moves at a higher velocity in the center compared to the periphery where frictional resistance slows it down. Thus, alveolar ventilation may be higher than expected because of asymmetric velocity of the inspired gas compared to uniform velocity in a bulk flow model. The relationship of V_T and V_{Dtot} is calculated as:

$$\frac{V_D}{V_T} = \frac{(P_{A_{CO_2}} - P_{E_{CO_2}})}{P_{A_{CO_2}}}$$

where, $P_{E_{CO_2}}$ is mixed expired PCO_2 . $P_{A_{CO_2}}$ is assumed to be same as $P_{a_{CO_2}}$ since there is no A-a CO_2 gradient. To calculate V_{Dalv} , $P_{ET_{CO_2}}$ is used to replace mixed $P_{E_{CO_2}}$.

$$V_{Dalv}/V_T = (P_{a_{CO_2}} - P_{ET_{CO_2}}) \div P_{a_{CO_2}}$$

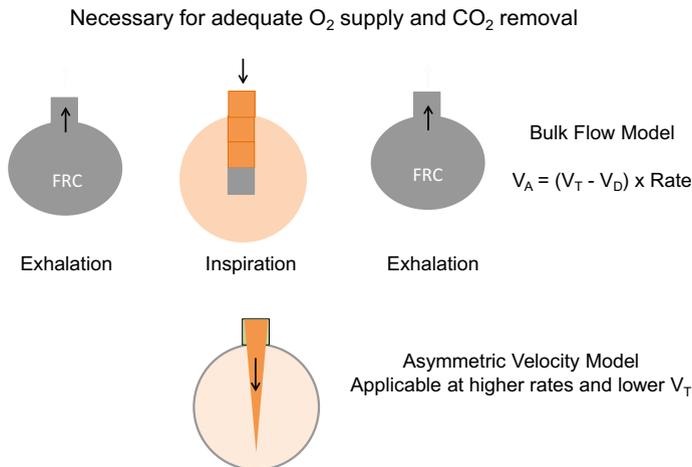


Fig. 3.2 Alveolar ventilation with bulk flow model and asymmetric velocity model

In normal lungs, $P_{ET}CO_2$ should be close to $PaCO_2$ and thus $V_{D}alv$ should be negligible. Increasing difference in $PaCO_2$ and $P_{ET}CO_2$ is indicative of increasing $V_{D}alv$. Increased $V_{D}alv$ is encountered when pulmonary perfusion is insufficient to match ventilation such as in decreased cardiac output, pulmonary embolism, hypovolemia and excessive PEEP. V_A is inversely proportional to $PaCO_2$. The relationship between V_A and $PaCO_2$ is hyperbolic but at the bedside, in the ranges of $PaCO_2$ commonly seen, it can be assumed to be linear. Therefore, when V_A is doubled, $PaCO_2$ is halved. Conversely when V_A is halved, $PaCO_2$ is doubled. With minor changes during the respiratory cycle the total pressure of all gases in alveoli is very similar to the total pressure of inspired gas. Alveolar gas composition depends on partial pressure of gases in the inspired gas, $PaCO_2$ (assumed to be same as alveolar PCO_2) and respiratory quotient (R). The simplified alveolar air equation is used to calculate the alveolar PO_2 ($P_{A}O_2$) as follows:

$$P_{A}O_2 = P_iO_2 - \left(\frac{P_aCO_2}{R} \right)$$

For practical purposes, R is assumed to be 0.8. According to the alveolar air equation, for a given P_iO_2 , a rise in $PaCO_2$ of 10 torr will result in a decrease in $P_{A}O_2$ by $10 \div 0.8 = 10 \times 1.25 = 12.5$ torr. Thus, pure hypoventilation will decrease $P_{A}O_2$ with increase in $PaCO_2$ by a factor of 1.25. In a normal person, P_iO_2 is about 150 torr. With a $PaCO_2$ of 40 torr, the $P_{A}O_2$ will be $150 - (40 \times 1.25)$ torr or $150 - 50 = 100$ torr. An increase in FiO_2 and therefore the P_iO_2 will raise $P_{A}O_2$ without affecting $PaCO_2$. Dangerous level of hypercarbia may coexist without hypoxia in hypoventilating patients who are breathing supplemental O_2 .

The alveolar gas is exchanged with the systemic venous (pulmonary arterial) blood through the process of diffusion which is influenced by the alveolar capillary barrier and the time available for equilibration. The "arterialized" blood is returned via pulmonary venous circulation to the heart to be pumped through the systemic arterial circulation. Diffusion in the gas phase (within the alveoli) is inversely proportional to the square root of the molecular weight of a gas molecule. Diffusion into the liquid phase (pulmonary capillary blood) is directly proportional to the solubility of a gas molecule. Considering the respective molecular weights and solubility, CO_2 is about 20 times more diffusible than O_2 . In health, the diffusion for both O_2 and CO_2 is complete by the time pulmonary capillary blood is no longer in contact with the alveolar gas. Clinically significant diffusion barrier manifests as hypoxia without impairing CO_2 elimination. Increasing FiO_2 and increasing alveolar-capillary O_2 gradient will improve oxygenation to some extent. However, even 100% O_2 is only about 5 times more concentrated than room air (as far as O_2 is concerned) while CO_2 is 20 times more diffusible than O_2 . In other words, before hypercarbia to develop solely because of a diffusion gradient, life will be incompatible in presence of severe hypoxemia even while breathing 100% oxygen. Presence of hypercarbia suggests additional factors such as alveolar hypoventilation and ventilation/perfusion mismatch. Alveolar-arterial oxygen (A-a O_2) gradient is often utilized to monitor for oxygenation defects in impairment of diffusion and V/Q mismatch.

3.3 Distribution of Ventilation

Alveoli are perfused with systemic venous (pulmonary arterial) blood which gets arterialized after diffusion is complete. Pulmonary venous (systemic arterial) blood should have the same PO_2 and PCO_2 as in the alveolar gas. However, the arterial blood gas composition is different from alveolar gases even in normal individuals because alveolar ventilation (V) and perfusion (Q) are not matched uniformly. Some alveoli receive more ventilation compared to perfusion (high V/Q ratio, dead space ventilation units) while some receive perfusion in excess of ventilation (low V/Q ratio, shunt perfusion units). Ventilation perfusion (V/Q) relationships in a normal lung are easily understood by consideration of West Zones (Fig. 3.5)

Because of the lung recoil and gravitational force, the intrapleural pressure is more negative in non-dependent parts (upper lobes in upright position) of the lung compared to the dependent parts (lower lobes in upright position). At FRC, the alveoli in non-dependent areas of the lung are therefore at a more horizontal part of the pressure volume curve (more distended or aerated but less compliant) whereas the alveoli in the dependent areas are at a more vertical portion of the lung (less distended or aerated but more compliant). For the same change in intrapleural

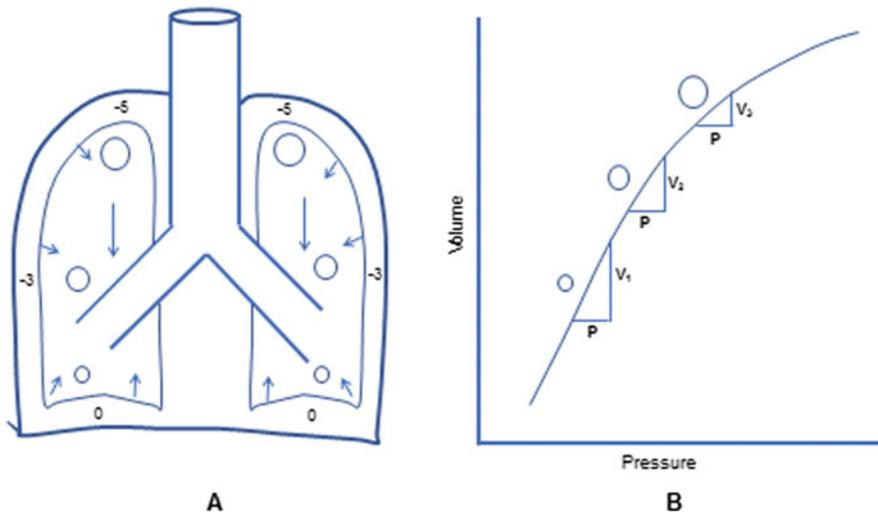
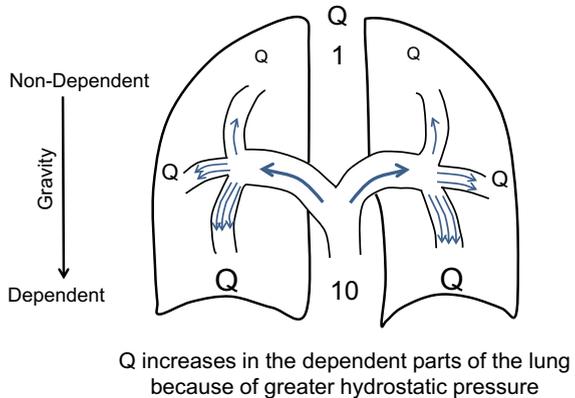


Fig. 3.3 a Illustrative representation of intrapleural pressures at FRC. Because of the lung recoil pressure and gravitational force, the intrapleural pressure is more negative in the non-dependent (upper lobes in upright position) portions of the lung compared to the dependent (lower lobes in upright position) portions. b Alveoli subjected to greater intrapleural pressure are therefore more distended (aerated) but less compliant. Alveoli that are surrounded by less negative pressure are less aerated but more compliant. For an identical change in inflation pressure, the change in volume is greater in dependent regions of the lung compared to non-dependent regions

Fig. 3.4 Distribution of pulmonary perfusion is gravity dependent with dependent areas of the lung getting a larger share of the blood flow



pressure during inspiration, the dependent alveoli will receive greater portion of ventilation. At FRC, ventilation increases from non-dependent to dependent areas of the lung (Fig. 3.3).

3.4 Distribution of Perfusion

The distribution of perfusion is very much position dependent. In an upright individual, perfusion is greatest in the lower lobes (Fig. 3.4). In a supine position in which most critically ill patients are cared for, greater proportion of perfusion as well as ventilation are distributed to posterior regions of the lung.

3.5 Distribution of Ventilation and Perfusion

Because of the higher hydrostatic force aided by gravity, perfusion also increases from non-dependent portions to dependent portions. However, the increase in perfusion is considerably greater than the increase in ventilation. V/Q ratios favor ventilation in the non-dependent areas and they favor perfusion in the dependent areas. Although both ventilation and perfusion increase from dependent to non-dependent areas of the lung, the increase in perfusion is considerably greater than the increase in ventilation. While ventilation increases a little over 3 folds, the increase in perfusion may be as much as tenfold. The $V:Q$ ratio is about 3:1 in the non-dependent and about 0.6:1 in the dependent parts of the lung. Thus, the V/Q ratios favor dead space ventilation in non-dependent parts and venous admixture in the dependent parts (Fig. 3.5). In situations such as ARDS, the dependent areas are more affected with capillary leakage. These areas receive less ventilation while still receiving larger share of perfusion. This leads to marked $V:Q$ mismatch and

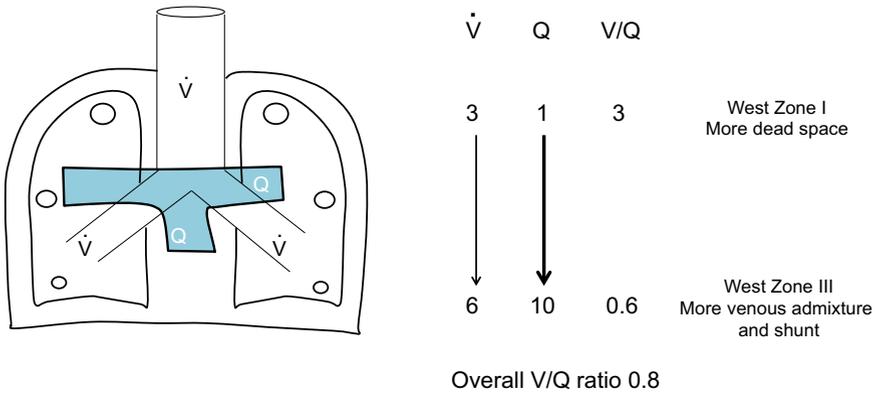


Fig. 3.5 Regional ventilation, perfusion and V/Q Ratios: Both ventilation and perfusion increase from non-dependent to dependent parts of the lung. The increase in perfusion is disproportionately greater than increase in ventilation resulting in V:Q ratios approximately 3:1 in non-dependent areas and 0.6:1 in dependent areas

hypoxemia. Caring such patients in prone position, shifts the perfusion to anterior portions of the lung which are less affected and better ventilated.

3.6 Regional V:Q Relationships (West Zones)

Alveolar pressure (P_A), pulmonary capillary arterial pressure (P_a) and pulmonary capillary venous pressure (P_v) determine the type of V:Q relationship and they form the basis of West zones described by John West. In West zone I, P_A is $> P_a$ which is $> P_v$. The alveolar pressure tamponades the pulmonary blood flow in Zone I. Ventilation is in excess of what is required to fully arterialize the pulmonary arterial blood amounting to wasted or dead space ventilation. Zone I is accentuated in hypovolemia, low cardiac output, pulmonary hypertension, excessive PEEP and pulmonary embolism. The overall V_D/V_T ratio is increased in Zone I. In West zone II, P_a is $> P_A$ which is $> P_v$. Ventilation and perfusion are better matched in the sense the pulmonary arterial blood is adequately arterialized with appropriate amount of ventilation. In West zone III, both P_a and P_v exceed P_A resulting in a relationship of $P_a > P_v > P_A$. In these areas ventilation is insufficient to fully arterialize the pulmonary arterial blood thereby leading to venous admixture or right to left shunting. Zone III is increased in pulmonary edema, fluid overload and atelectasis. When using prone positioning in ARDS, the clinician attempts to improve V:Q matching by converting Zone III to Zone II type of V:Q relationship (Fig. 3.6).

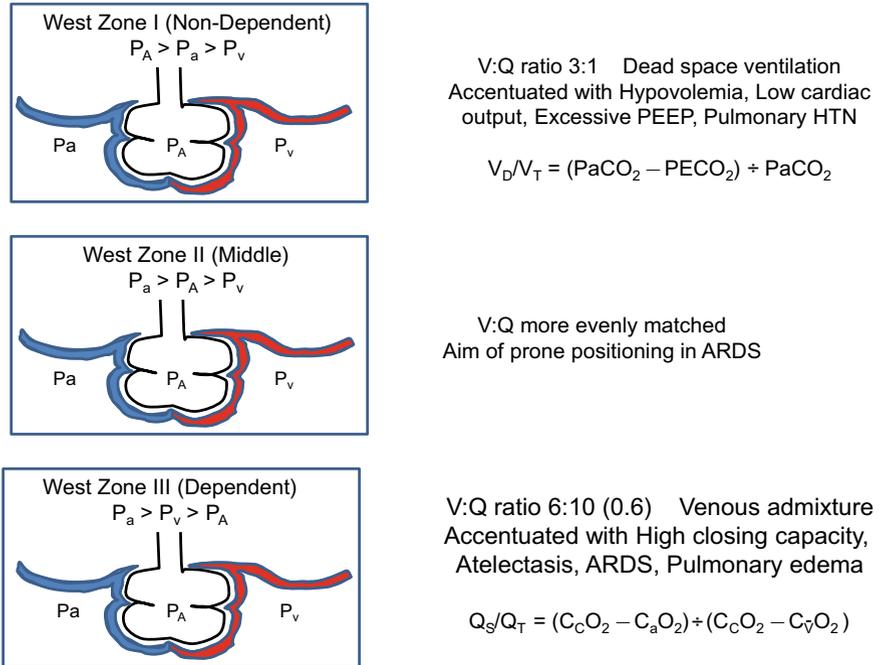


Fig. 3.6 Ventilation–Perfusion Relationship (West zones). Zone I is characterized by high V:Q ratios whereas Zone III has lower V:Q ratios

3.7 Ventilation (V) and Perfusion (Q) Mismatch

There are 3 prototypical V:Q relationships (Fig. 3.7). The first one is when ventilation is inadequate to fully arterialize the blood flowing past the hypoventilating alveolus. The end-result is a part (or all) of pulmonary capillary blood to remain variably deoxygenated and mix with the arterialized blood from other adequately ventilating lung segments. In this situation the arterial PO_2 (PaO_2) will be less than alveolar PO_2 (P_AO_2) as calculated by the alveolar air equation. The blood gas abnormality is referred to as venous admixture or intrapulmonary right to left shunt. The second V:Q relationship is when ventilation is appropriate for the given perfusion. In this situation, the pulmonary capillary blood is fully arterialized and PaO_2 and P_AO_2 are equal. The third V:Q relationship characterizes areas with decreased perfusion compared to ventilation. The part of the atmospheric air enters and leaves alveoli without contributing to the gas exchange. This type of V/Q mismatch is termed dead-space ventilation. In such situations P_ACO_2 is markedly less than $PaCO_2$.

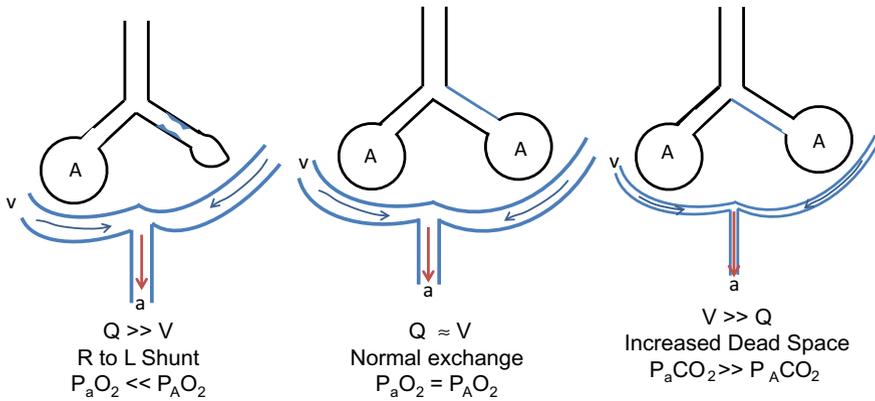


Fig. 3.7 Three types of V:Q relationship may exist: Normal V/Q, Shunt units and Dead space units

3.8 O₂ Transport and Utilization

The amount of oxygen carried by the arterial blood is in two forms: (i) in a dissolved state and (ii) combined with hemoglobin. The amount of dissolved oxygen is linearly related to PO_2 . For every 100 torr PO_2 , there is 0.3 mL oxygen dissolved in 100 mL of solvent. Dissolved O_2 serves two important functions. In this form O_2 is immediately available for tissue uptake. Dissolved oxygen also determines the extent to which hemoglobin is saturated by O_2 . However this amount in and of itself, is hardly sufficient to satisfy tissue O_2 demands (Fig. 3.8).

The O_2 consumption for a healthy adult is approximately 250 mL/min. At a normal PaO_2 of 100 torr, there is 3 mL/L of dissolved O_2 in blood. If all of O_2 in blood was in the state of dissolved oxygen, it would require 83 L/min of cardiac output even if all of it were to be utilized by the body. The way hemoglobin associates and dissociates with O_2 constitutes an efficient means of transporting and utilizing oxygen. Each gram of hemoglobin when 100% saturated with O_2 , carries 1.34 ml of oxygen. Thus, 15 G of hemoglobin per 100 mL of blood can carry approximately 20 ml of O_2 when nearly fully saturated with oxygen. This amounts to 200 mL of O_2 in 1000 mL of blood. With cardiac output of 5L per minute, an adult delivers 200 X 5 or 1000 mL of oxygen to the tissues every minute. With a resting O_2 consumption of 250 mL/min, as much as 750 mL (75% HbO_2 saturation) can still be returned back to the heart in mixed venous blood. Normally, arterial blood HbO_2 saturation is nearly complete much before we attain a normal PaO_2 because of the dissociation curve. For example, at PO_2 of 100 torr; hemoglobin-oxygen saturation is 97.5%, an increase of only 3.5% compared to the saturation at PO_2 of 70 torr where it is 94%. Relatively little O_2 can be added at higher PO_2 s (Fig. 3.9).

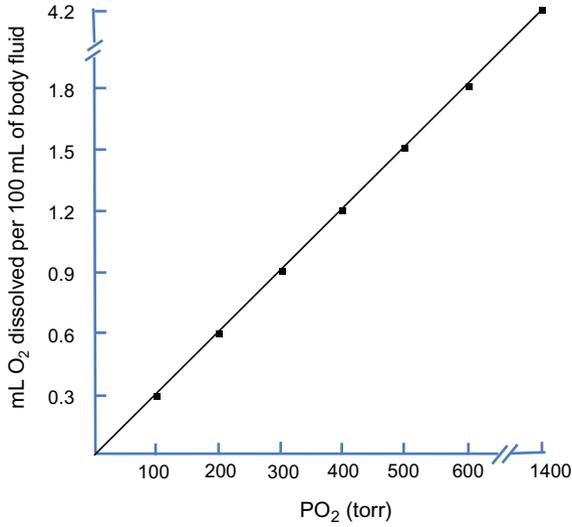


Fig. 3.8 Relationship of dissolved O₂ and PO₂ is linear. For every 100 torr, 0.3 ml of O₂ is in a dissolved state in a solution. Dissolved O₂ = (PO₂ in torr × 0.003)/mL of body fluid

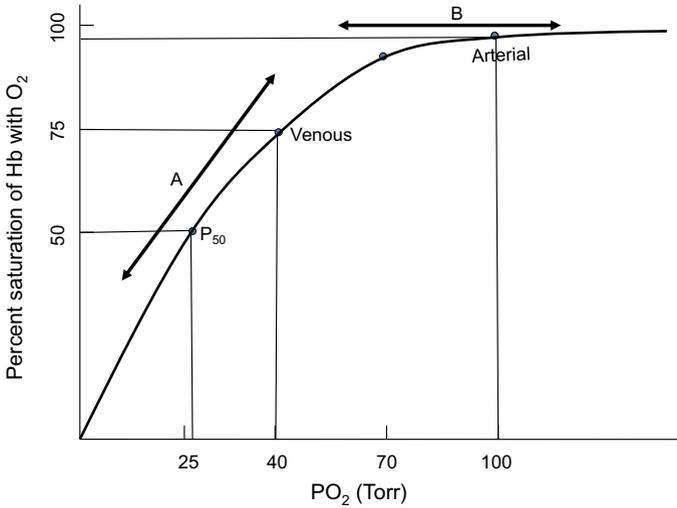


Fig. 3.9 Relationship of Hb-O₂ saturation and PO₂. In part of the curve labeled A, the change in the amount of Hb-O₂ saturation is considerably greater for a given change in PO₂ compared to the part labeled B. P₅₀ refers to PO₂ at which Hb is 50% saturated

The hemoglobin molecule contains 4 heme chains each with an iron molecule in a reduced, ferrous (Fe^{++}) state, and 4 globin chains. The spatial arrangement of the heme chains, Fe^{++} and globin chains is necessary for O_2 to bind reversibly with the heme part of the hemoglobin molecule. If the iron molecule is in an oxidized to a ferric state (Fe^{+++}), methemoglobin is formed which is incapable of binding with O_2 . Carbon monoxide (CO) reversibly binds with hemoglobin at the same sites as O_2 but with about 210 time greater affinity. Thus presence of mere 0.1% atmospheric CO will result in 50% carboxyhemoglobin (COHb) and 50% HbO_2 in arterial blood when breathing room air (21% O_2)! Both methemoglobinemia and CO poisoning can lead to life-threatening decline in blood O_2 content despite adequate PaO_2 . 2,3-DPG, a product of RBC anaerobic glycolysis, plays an important part in association and dissociation of hemoglobin and oxygen. It binds with deoxyhemoglobin much more efficiently than with oxyhemoglobin. At lower PO_2 levels such as would occur in the tissues, 2,3-DPG facilitates O_2 to dissociate from the hemoglobin and make itself available for aerobic metabolism. At higher PO_2 levels such as would occur in the lung, oxygen binds more readily with hemoglobin not allowing 2,3-DPG to bind with hemoglobin. At high altitudes and in patients with anemia and other hypoxic conditions, 2,3-DPG concentration is increased allowing for greater release of O_2 to the tissues. Fetal hemoglobin has less affinity to 2,3-DPG and therefore it binds to O_2 more readily.

Several factors influence the shape of HbO_2 dissociation curve. A shift to the left or a decrease in P_{50} (PO_2 at which HbO_2 saturation is 50%) is a characteristic of fetal hemoglobin, hypothermia, alkalosis and a decrease in 2,3-DPG. On the other hand, a shift to the right or an increase in P_{50} is observed in hyperthermia, acidosis and an increase in 2,3-DPG. A shift to the right facilitates release of O_2 at the tissue level. (Fig. 3.10).

Following equations represent the relationship of arterial O_2 content (CaO_2), venous O_2 content (CvO_2), Cardiac output (CO), Oxygen delivery (DO_2), and Oxygen consumption (VO_2).

$$\text{CaO}_2 = [(\text{Hb} \times 1.34 \times \text{SaO}_2\%) + (\text{PaO}_2 \times 0.003)] \times 10$$

where CaO_2 is Oxygen content of blood in mL/L, Hb is Hemoglobin concentration in G/dL, SaO_2 is the Arterial Oxygen Saturation, PaO_2 is the Arterial Oxygen Tension in mmHg. Please note that the multiplication by 10 is to convert O_2 content/100 mL to O_2 content/1000 mL or 1 L.

Similarly, the oxygen content of mixed venous blood can be calculated as follows:

$$\text{CvO}_2 = [(\text{Hgb} \times 1.34 \times \text{SvO}_2\%) + (\text{PvO}_2 \times 0.003)] \times 10$$

Delivery of oxygen to the tissues by the circulatory system is estimated using the formula:

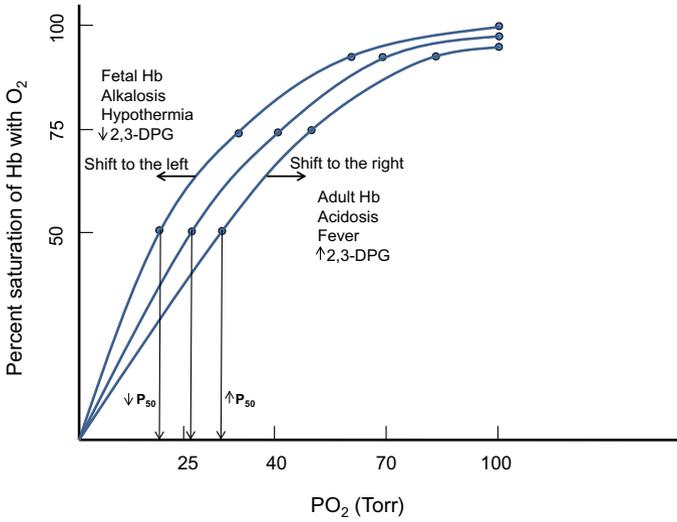


Fig. 3.10 HbO₂ dissociation curves shift to the left (increased O₂ affinity) or to the right (decreased O₂ affinity) under certain clinically encountered situations

$$\dot{D}O_2 = CaO_2 \times C.O.$$

where $\dot{D}O_2$ is the delivery of oxygen in mL/min, CaO_2 is the arterial oxygen content in mL/L, and C.O. is the cardiac output in L/min.

Oxygen consumption by the tissues is measured in mL of oxygen per minute and is expressed as:

$$\dot{V}O_2 = (CaO_2 - C\bar{v}O_2) \times C.O.$$

where $\dot{V}O_2$ is oxygen consumption per minute, CaO_2 is the arterial oxygen content, $C\bar{v}O_2$ is the mixed venous oxygen content, and C.O. is the cardiac output.

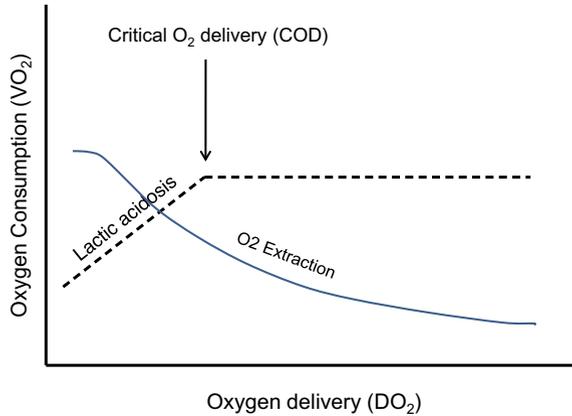
The ratio of oxygen consumption to oxygen delivery is called Oxygen Extraction. It is the fraction of the oxygen delivery that is consumed by the tissues. It is calculated as follows:

$$Oxygen\ Extraction(O_{2Extr}) = \frac{\dot{V}O_2}{\dot{D}O_2}$$

Since Cardiac Output is in both the numerator and the denominator, oxygen extraction can be simplified as follows (Fig. 3.11):

$$Oxygen\ Extraction(O_{2Extr}) = \frac{(CaO_2 - C\bar{v}O_2)}{CaO_2}$$

Fig. 3.11 As DO_2 decreases, VO_2 remains constant over a wide range by increasing O_2 extraction to maintain aerobic metabolism. Fall in DO_2 beyond a certain point (critical O_2 delivery) results in anaerobic metabolism and lactic acidosis despite continued increase in O_2 extraction



Normal resting adult has DO_2 of approximately 1 L/min and VO_2 of 250 mL/min. Thus 75% of delivered O_2 is returned back to heart in the mixed venous blood. CvO_2 is a reflection of the DO_2 - VO_2 relationship. For sake of convenience, SvO_2 is substituted for CvO_2 since most of the oxygen content is accounted for by hemoglobin saturated with O_2 . The relationship between VO_2 , DO_2 and O_2 extraction is shown in Fig. 3.11. When DO_2 is decreased, VO_2 is initially kept constant to maintain aerobic metabolism by increasing O_2 extraction. A decrease in DO_2 below a certain level results in a decreased VO_2 despite increased O_2 extraction. DO_2 below which increased O_2 extraction does not satisfy aerobic metabolic demand of the tissues is termed critical O_2 delivery (COD). When DO_2 falls below COD, anaerobic metabolism begins with accumulation of lactic acid. Provided VO_2 and CaO_2 remain unchanged, SvO_2 is an indication of adequacy of CO to maintain aerobic metabolism. Declining SVO_2 is suggestive of decreasing cardiac output.

3.9 Abnormalities of Gas Exchange

As outlined in the preceding discussion, several factors determine gas exchange at the alveolar capillary junction. Analysis of arterial blood gases provides both diagnostic clues as well as therapeutic approach in management of respiratory disorders. The challenge to the clinician is that arterial sample is often not available and a capillary blood sample has to be relied on in many circumstances. Also, a precise FiO_2 is usually not available in many patients. The clinician has to rely on many assumptions and clinical experience.

There are four main types of abnormalities in gas exchange. These are (a) alveolar hypoventilation (b) ventilation-perfusion (V-Q) mismatch (c) diffusion defects and (d) absolute right to left shunt (Table 3.2). In many patients more than one disorder may be present. For example, a patient with alveolar hypoventilation may also have a component of V-Q mismatch and a patient with a diffusion defect

may become exhausted and develop hypoventilation. In such a situation, the clinician must determine the major component of gas exchange abnormality to plan a targeted intervention.

Alveolar hypoventilation results when sufficient air is not moved in and out of the alveoli. There are 3 major types of clinical situations which manifest as alveolar hypoventilation. These are: airway obstruction above the carina (choanal atresia,

Table.3.2 Interpretation of arterial blood gas (ABG) values

Lesion	Effect	Typical ABG
<ul style="list-style-type: none"> * Central (above the carina) airway obstruction * Depressed respiratory center * Ineffective neuromuscular function 	Uniform alveolar hypoventilation	<ul style="list-style-type: none"> * Early increase in PaCO₂ * Proportionate decrease in PO₂ depending on air equation * Response to supplemental oxygen: Excellent
Intrapulmonary airway obstruction	Venous admixture V/Q mismatch	<ul style="list-style-type: none"> * Mild: ↓ PCO₂, ↓ PO₂ * Moderate: “Normal PCO₂ ↓↓PO₂ * Severe: ↑ ↑PCO₂ ↓↓↓ PO₂ * Response to supplemental oxygen: Good
Alveolar-Interstitial pathology	V/Q mismatch, venous admixture, R to L shunt Diffusion defect	<ul style="list-style-type: none"> * Early decrease in PO₂ depending on severity * Normal or low PCO₂ * ↑ PCO₂ if fatigue occurs * Response to supplemental oxygen: Fair to poor
Extrapulmonary right to left shunt	Systemic venous blood bypasses alveoli, absolute R to L shunt	<ul style="list-style-type: none"> * Hypoxemia depending on magnitude of the shunt * Response to supplemental oxygen: Very poor

subglottic stenosis, vascular ring etc.), weakness of muscles of respiration (Guillain-Barré syndrome, myasthenia gravis, diaphragmatic paralysis etc.) and depressed respiratory center (CNS depressants, congenital central hypoventilation syndrome, brain stem dysfunction etc.). Airway obstruction below the carina may also manifest with predominant alveolar ventilation if the obstruction is relatively uniform such as in bronchiolitis obliterans. Alveolar ventilation is inversely proportional to PaCO_2 ; a certain percentage decline in alveolar ventilation $[(V_t - V_d) \times \text{rate}]$ will lead to an increase in PaCO_2 by a similar percentage. The hallmark of alveolar hypoventilation is elevated PaCO_2 and a proportionate decline in PAO_2 as determined by alveolar air equation;

For $\text{FiO}_2 < 1$, the equation can be simplified to:

$$\text{PAO}_2 = \text{PiO}_2 - \frac{\text{PACO}_2}{R}$$

For bedside calculations, PACO_2 is substituted for PaCO_2 . Thus for a given PiO_2 , PAO_2 will fall only by the rise in $\text{PaCO}_2 \div R$. Since R is assumed to be 0.8, the fall in PAO_2 will be approximately by rise in $\text{PaCO}_2 \times 1.25$. In the absence of significant parenchymal disease and intrapulmonary shunting, administration of supplemental O_2 will increase PiO_2 and readily reverse hypoxemia despite persistent hypercarbia.

In intrapulmonary airway obstruction (asthma, bronchiolitis, aspiration), the obstruction is not uniform in nature. Some areas are more obstructed than others while some still are relatively unaffected resulting in multiple areas having different extent of ventilation; some are hypoventilated while others are hyperventilated. Pulmonary capillary blood coming from hypoventilated areas has higher PaCO_2 and a lower PaO_2 , whereas that coming from hyperventilated areas has lower PaCO_2 and higher PaO_2 . A lower PaCO_2 can compensate for the higher PaCO_2 because the Hb-CO_2 dissociation curve is relatively linear. An equal amount of blood with PaCO_2 of 30 torr mixing with PaCO_2 of 50 torr will result in a PaCO_2 of 40 torr. A higher PaO_2 however cannot compensate for a lower PaO_2 in the presence of desaturated hemoglobin because of the shape of the HbO_2 dissociation curve. It is the % HbO_2 saturation that averages out since far more O_2 is responsible for combining with Hb than the dissolved O_2 reflecting the PO_2 . For example, an equal amount of blood with PaO_2 of 25 torr and HbO_2 saturation of 50% mixing with PaO_2 of 110 torr and HbO_2 saturation of near 100% will result in HbO_2 saturation of 75% and PaO_2 of 40 torr. The blood gas abnormality in such situations is referred to as V-Q mismatch, venous admixture or partial right to left intrapulmonary shunting. In mild disease, the hyperventilated areas predominate outnumbering the hypoventilated ones. The end result is hypocarbia and respiratory alkalosis. An elevated PaO_2 in the hyperventilated areas however cannot compensate for the low PaO_2 in hyperventilated area resulting in mild hypoxemia. With increasing severity, more areas become hypoventilated resulting in normalization of PaCO_2 (crossover point) with a further progressive decline in PaO_2 . A normal or slightly elevated PaCO_2 in intrapulmonary airway obstruction raises concern for

impending respiratory failure. As the disease severity increases, more and more lung units are hypoventilated, resulting in hypercarbia, respiratory acidosis and hypoxemia. Supplemental O_2 is effective if it is able to reach the hypoventilated alveoli.

In alveolar and interstitial pathology (ARDS, interstitial pneumonia, pulmonary edema), arterial blood gas values reflect intrapulmonary right to left shunting and diffusion barrier. Systemic venous blood flows across unventilated alveoli without getting oxygenated. The diffusion impediment is 20 times greater for O_2 than for CO_2 . Hypoxemia developing early and getting progressively severe is a hall mark of such diseases. Most patients develop hyperventilation manifesting hypocarbia. An increase in $PaCO_2$ is observed only after muscle fatigue and exhaustion ensue. Response to supplemental O_2 , while life-saving, may not be as robust as in other respiratory pathophysiologic alterations. In severe situations, hypoxemia may become resistant to O_2 therapy.

In conditions where systemic venous blood completely bypasses the alveolar-capillary bed (cyanotic heart disease, pulmonary arteriovenous fistula etc.), hypoxemia is the predominant feature as a fixed amount of deoxygenated blood mixes with oxygenated blood. Supplemental oxygen does not increase PaO_2 since the deoxygenated shunted blood has no chance of getting in contact with alveolar gas.

3.10 Regulation of Respiration

Blood gas homeostasis to suit the body's requirements is maintained by a complex interaction of controllers, sensors and effectors (Fig. 3.12). The central respiratory controller is represented by a group of neurons in the CNS that receives information from sensors and sends motor impulses to muscles of respiration which serve the function of the effectors. The most important effector is the diaphragm which is aided by the intercostal, abdominal and neck muscles as accessories when needed. The effectors target the lungs to adjust alveolar ventilation and control pH, $PaCO_2$ and PaO_2 . The entire respiratory regulatory mechanism undergoes maturational changes from the neonatal to adult life. It is also subject to modifications by the sleep states, disease processes, pharmacologic agents and acclimatization to the environment.

3.10.1 Central Respiratory Controller

Two functionally and anatomically distinct group of neurons located in the CNS control the process of respiration: voluntary and automatic.

Voluntary control of respiration resides in the cerebral cortex and limbic fore-brain areas. Major sensory inputs consist of smell, vision, emotions, pain, touch

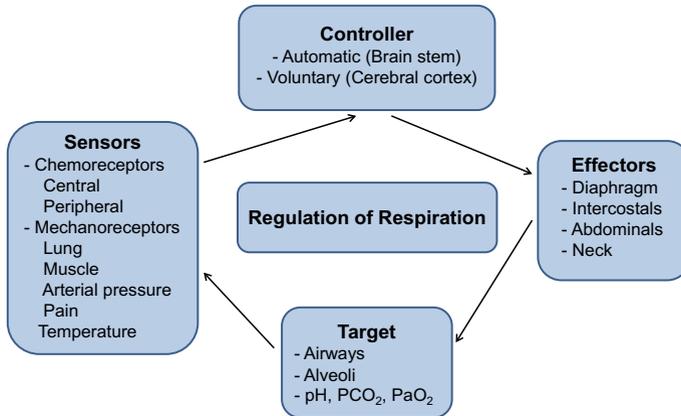


Fig. 3.12 Control of respiration

etc., and motor impulses are sent to the effectors through corticobulbar and corticospinal tracts. Voluntary control of respiration requires a certain level of consciousness, and is important for protection against aspiration and inhalation of noxious gases. Patient with toxic/metabolic/infectious/traumatic encephalopathies and pharmacologic sedation may lose voluntary control of respiration depending on the extent of CNS dysfunction.

Automatic control of respiration is located in the brainstem. Neuronal circuits, referred to as central pattern generators (CPGs) spontaneously generate rhythmic motor output without requiring conscious input, and are responsible for breathing, swallowing and chewing. CPGs responsible for breathing are located in pons and medulla. A group of neurons located in lower pons constitutes the apneustic center which is responsible for prolonged inspiratory effort interrupted by brief periods of expiratory activity. Another group of neurons in the upper pons termed pneumotaxic center, is involved in inhibiting the activity of CPGs. The role of apneustic and pneumotaxic centers is to fine-tune the rhythmic respiratory activity of CPGs. Global CNS depression from any cause can manifest as slow and shallow respirations, hypoventilation and respiratory acidosis. Similarly, localized CNS lesions are manifested by specific patterns of abnormal ventilation.

3.10.2 Sensors

Multiple mechanisms exist that can sense abnormalities of gas exchange, acid–base imbalance and respiratory system dysfunction, and send that information to the central respiratory controller to modify the breathing pattern. These mechanisms exist in the form of sensory nerve endings termed chemoreceptors and mechanoreceptors depending on the type of stimulus that is being sensed.

Chemoreceptors are further classified as central or peripheral depending on their location.

Central chemoreceptors are located within the CNS. They reside over a wide area that includes posterior hypothalamus, cerebellum, locus ceruleus, raphe and brain stem. They sense a change in chemical composition of the body fluid they are exposed to. Central chemoreceptors respond to the chemical changes in the extracellular fluid (ECF) of the brain represented by cerebrospinal fluid (CSF). The ventilatory response is predominantly due to a change in the H^+ concentration (pH) of the brain ECF. The brain ECF and blood are separated by the blood brain barrier which is relatively impermeable to H^+ and HCO_3^- but freely permeable to PCO_2 . A rise in $PaCO_2$ is quickly reflected in a rapid rise in the CSF PCO_2 . The consequent fall in CSF pH is sensed by the central chemoreceptors which then send excitatory impulses to the controller resulting in increased ventilation via the effectors. CSF pH in normal conditions is slightly acidic, around 7.32. With its lower protein level and absence of hemoglobin, CSF also has much less buffering capacity compared to that of the blood. Consequently, for an equivalent change in $PaCO_2$, the change in CSF pH is much more pronounced than that in the blood. In disease states characterized by chronically elevated $PaCO_2$, the CSF pH tends to normalize as HCO_3^- eventually equilibrates across the BBB. Patients with chronically elevated $PaCO_2$ therefore have a relatively normal CSF pH and they do not have the same ventilatory response that is observed with acute hypercarbia.

Peripheral chemoreceptors are clusters of cells referred to as carotid bodies just above the bifurcation of the common carotid and external carotid arteries, and aortic bodies above and below the aortic arch. Carotid bodies are far more powerful sensors than aortic bodies. The cells comprising carotid and aortic bodies have a very high metabolic rate as well as blood flow to meet their metabolic demands. The main stimulus for peripheral chemoreceptors is hypoxia. Decrease in PaO_2 (SaO_2), blood flow (low cardiac output), and impaired O_2 utilization (cyanide poisoning) classically described as hypoxemic hypoxia, stagnant hypoxia and histotoxic hypoxia respectively, are potent stimulators of peripheral chemoreceptors. Anemia and dyshemoglobinemias do not stimulate peripheral chemoreceptors as long as PaO_2 and cardiac output are adequate. This is because the dissolved oxygen in the blood in form of PaO_2 and high blood flow easily satisfy the exceptionally high O_2 requirement of the chemoreceptors. Relationship of PaO_2 and stimulation of peripheral chemoreceptors is non-linear. Chemoreceptor stimulation begins at PaO_2 below 500 torr and a small increase in ventilation occurs incrementally until PaO_2 reaches 100 torr. The response time for peripheral chemoreceptor is much faster than central chemoreceptor stimulation. Even during normal respiration, carotid bodies response rate is fast enough to alter their discharges sensing small cyclic changes in PaO_2 during inspiration and exhalation. At PaO_2 less than ~ 50 torr, carotid body stimulation increases exponentially. Subjective feeling of dyspnea from pure hypoxia alone does not occur until PaO_2 falls below ~ 50 torr (SaO_2 below $\sim 85\%$). Peripheral chemoreceptors account for almost all the hyperventilation response to hypoxia. They also respond to PCO_2 but the increase in alveolar ventilation per torr PCO_2 is much less than that from central

Table.3.3 Characteristics of central and peripheral chemoreceptor stimulation

	Central chemoreception	Peripheral chemoreception
Site	Central Nervous System	Carotid and Aortic bodies
Primary Stimulus	CSF pH (PaCO ₂)	Hypoxia
Response to hypoxia	None or depressed	Marked stimulation
Response to acutely PaCO ₂	+ + +	+
Response time	Slow	Fast
Acclimatization	Readily occurs	Does not occur easily
Sedation/anesthesia	Easily depressed	Not easily depressed

chemoreceptor stimulation. Adaptation to stimulus (Increased PaCO₂) occurs in days for central chemoreceptors but hypoxic stimulation of peripheral chemoreceptors persists for a long time, even for life, for people living at high altitudes as reflected by their lower PaCO₂ levels. The difference in central and peripheral chemoreception is presented in Table 3.3.

In certain disease states such as asthma a blunted response to hypoxia is well documented. In recent SARS-CoV-2 pandemic, similar observations of “happy hypoxia” have been reported describing patients with minimal respiratory distress in spite of considerable arterial O₂ desaturation. Pure peripheral chemoreceptor stimulation results in bradycardia. In most situations with acute hypoxia, tachycardia develops because of action of muscles of respirations causing lung inflation. Two situations where bradycardia is a pronounced effect of sole peripheral chemoreceptor stimulation are: hypoxic patients with neuromuscular blockade and neuromyopathy, and in intrauterine life. In chronic hypercarbic states, the central chemoreceptors have adapted to elevated PaCO₂ and the respirations are maintained mainly by the hypoxic drive from the peripheral chemoreceptors. Administration of high concentration of O₂ can potentially abolish the hypoxic peripheral chemoreception resulting in hypoventilation, respiratory acidosis and CO₂ narcosis. Care must be taken in delivering supplemental O₂ to such patients to avoid serious hypoventilation.

Mechanoreceptors

Stretch receptors located within the airway smooth muscles, are stimulated by lung inflation. They are important in adjusting the respiratory rate in health and disease and minimizing work of breathing. In diseases with decreased lung compliance, alveoli fill up quickly thus the transpulmonary pressure is transmitted quickly to the airways stretch receptors resulting in inhibition of inspiration. The breathing pattern is rapid and shallow. In diseases of increased resistance, alveoli take longer time to fill up thus delaying stimulation of the stretch receptors and thus resulting in deeper and slower respiration.

Muscle receptors are located in diaphragm and intercostal muscles. Stretching of these muscles is sensed by the muscle spindle to control the strength of contraction.

Excessive distortion of diaphragm and intercostal muscles inhibits inspiratory activity.

J receptors are located in the alveolar walls close to the pulmonary capillaries. Pulmonary capillary engorgement and interstitial and alveolar fluid collection activate J receptors resulting in rapid, shallow respiration and dyspnea.

Irritant receptors are located in between the epithelial cells in the airway mucous membrane throughout the respiratory tract. They are stimulated by particulate matter and noxious gases as well as cold air. Their stimulation results in bronchoconstriction and cough.

Arterial baroreceptors located in aortic arch and carotid sinus, respond to arterial blood pressure. Hypotension results in tachypnea and hyperpnea.

3.10.3 Effectors

The most important effector of respiration is the diaphragm. Intercostals and abdominal muscles are recruited when additional increase in ventilation is necessary. Sternocleidomastoids and paraspinal muscles may be called upon to additional contribution to the respiratory effort. Developmental changes significantly influence the ability of the diaphragm to sustain large elastic work load and resist fatigue. As compared to the skeletal muscles which contain primarily fast twitch, fatigable type 2 × and 2b fibers, the predominant muscle fiber types in the diaphragm are the fatigue resistant slow twitch type 1 and intermediate fatigue resistant fast twitch type 2a fibers. The type 1 fibers have slower shortening velocities than fast type 2a fibers but are highly fatigue resistant due to their lower ATP consumption and their reliance almost exclusively on aerobic metabolism. Type 2a fibers on the other hand are fast-twitch, less oxidative and are more prone to fatigue. This combination of fiber types allows for good fatigue resistance and increased force generation when necessary. Diaphragms of newborns and infants have a lower muscle mass when indexed for body size. In addition, they have lower percentages of fatigue resistant type I fibers. The diaphragm of preterm infants contains only 10% type I fibers. This increases to 25% in term neonates and 55% in children greater than 2 years of age. These developmental differences predispose neonates and infants to respiratory muscle fatigue and respiratory failure.

Suggested Readings

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