



Factors Affecting the Clearance and Biodistribution of Polymeric Nanoparticles

14

Komal Parmar, Jayvadan Patel,
and Yashwant Pathak

Contents

1	Introduction	261
2	Pharmacokinetic Functions	262
3	Factors Affecting Biodistribution and Clearance of Nanoparticles	264
4	Conclusion	268
	References	268

Abstract

Nanoparticles are promising drug delivery for various therapeutic applications. Pharmacokinetics is important to study the in vivo fate of nanoparticles. Biodistribution and clearance are the important parameters of pharmacokinetics to be considered. Impact of various characteristics of polymeric nanoparticles affects biodistribution and clearance of nanoparticles. The chapter focuses on four important characteristics of polymeric nanoparticles affecting their biodistribution and clearance.

K. Parmar (✉)
ROFEL, Shri G.M. Bilakhia College of Pharmacy,
Vapi, Gujarat, India

J. Patel
Nootan Pharmacy College, Faculty of Pharmacy,
Sankalchand Patel University,
Visnagar, Gujarat, India

Y. Pathak
College of Pharmacy, University of South Florida
Health, Tampa, FL, USA

Keywords

Clearance · Biodistribution · Polymeric
nanoparticles

1 Introduction

Nanodelivery systems are a comparatively new but quickly emerging field in which nanoscale materials are used as diagnostic tools or to administer therapeutic medicines to precise targets in a controlled mode [30, 57, 82, 84]. Nanoparticles can be turned into intelligent devices, encapsulating medicinal and imaging chemicals while also having stealth properties, by manipulating their size, surface features, and composition [69]. They are intended to alter the biodistribution and pharmacokinetics of the drugs, allowing for a higher dose to be delivered to a targeted disease tissue, in order to improve the therapeutic efficacy and render reduced toxicity. Many different materials and shapes of nanoparticles have been

produced for use in disease therapy, and many of them have shown to be effective [59, 61, 78]. Currently, there are a number of nanopharmaceutical products in the market [10, 19, 29, 31, 34, 42, 43]. When using nanoparticles, several factors must be considered: distribution efficiency, therapy effects, and clearance [53, 66, 90, 95]. Clinical uses for nanoparticles with great efficacy and good biosafety are on the horizon. Therapeutic effectiveness of nanoparticles is inextricably tied to pharmacological and toxicological characteristics. The most significant elements for establishing a high therapeutical index and related clinical performance are drug target residence, maximal tolerated dose, and selectivity [7, 40, 49]. Optimizing drug pharmacokinetic qualities to improve therapeutic effects and prevent adverse effects is an important feature of nanoparticle formulation design, which involves considering not just the characteristics of nanoparticles but also pharmacokinetic characteristics. The interaction between body systems and nanoparticles is responsible for all therapy outcomes.

The shape and polymer content in the core and periphery of polymeric nanoparticles characterize them. Drug is either adsorbed on the surface or encapsulated inside the core part of the polymeric nanoparticles. Delivery formulation governs the release of drug either to be controlled, sustained, or triggered release [3]. Furthermore, the surface of the polymeric nanoparticles can be attached with functional groups to obtain certain added characteristics such as prolonged systemic residence time, minimal non-specific distribution, and/or target specific cell or tissue, for example, coating the surface of the nanoparticles with polyethylene glycol to prolong the systemic circulation of the particulate system. PEGylation of nanoparticles defends the surface of the nanoparticles from protein absorption which leads to aggregation, opsonization, and phagocytosis, thereby providing extended systemic retention [52, 89]. The resultant elimination is due to phagocytosis by the monomolecular phagocyte system. Because of the large number of phagocytic cells in the liver and spleen, the majority of opsonized particles are removed by a receptor-mediated mechanism in less than a few minutes,

or they are expelled. Thus, numerous approaches are explored by the investigators to help retain the nanoparticles in the systemic circulation, so that drug delivery system can deliver the drug for prolonged period of time with specific distribution [9, 12, 83]. The effects of physiological tissue deficits and polymeric nanoparticle physicochemical characteristics on clearance and biodistribution will be discussed in this chapter to consider probable means for their advancement.

2 Pharmacokinetic Functions

Investigators have explored various factors such as physicochemical properties, administration route, dosing, and coating, affecting distribution and clearance of nanoparticles. However, all these variables are dependent on the physiological environment. Optimization of such variables after understanding of pharmacokinetics of body will help to obtain the successful drug delivery using polymeric nanoparticles.

Briefly, once the nanoparticles are introduced into systemic circulation, they are distributed to various tissues and organs where they are encountered with physical and biological challenges that may change their properties and affect their deposition and are concurrently cleared later [85] (Fig. 14.1). The interactions between nanoparticles and each organ are unique. In vivo, the major clearance process for polymeric nanoparticles is reticular endothelial system (RES) also termed as mononuclear phagocyte system (MPS) [13, 75]. Phagocytosis of polymeric nanoparticles is usually initiated by opsonization process. Opsonization occurs when opsonins, a heterogeneous group of proteins or protein fragments including C3, C4, and C5, immunoglobulins, fibronectin, and apolipoproteins, are deposited on the surface of nanoparticles and interact with a variety of surface receptors on RES cells, including complement, Fc, and fibronectin receptors [4, 71]. Proteins other than opsonins also get attached on the surface of nanoparticles forming a corona which further enables the scavenger receptors to categorize [80]. This adds an alternative way by which RES clears polymeric nanoparticles. Once

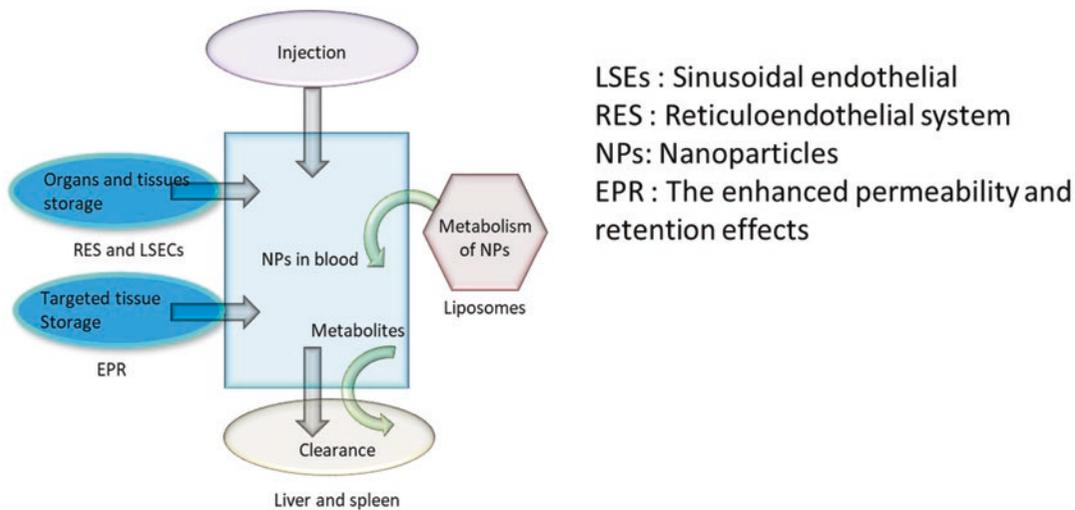


Fig. 14.1 Schematic diagram illustrating biodistribution and clearance of polymeric nanoparticles

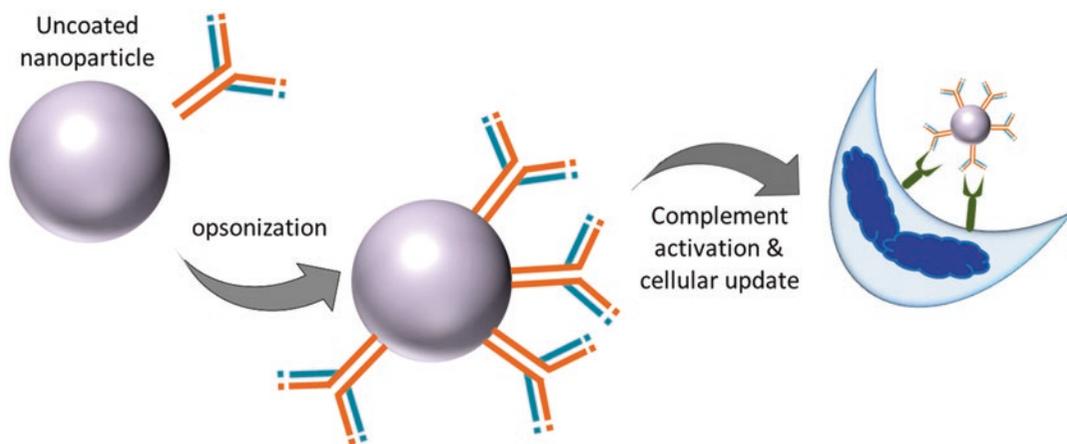


Fig. 14.2 Opsonization and uptake of uncoated polymer nanoparticles

opsonized and transported from the systemic circulation, a polymeric nanoparticle is usually localized in one of the MPS organs, mainly the liver and spleen [62] (Fig. 14.2). There are numerous such biological barriers naturally designed to safeguard the human body from foreign material. Among these barriers are the immune system's cellular and humoral arms, as well as mucosal barriers. Nanoparticles must overcome such constraints in order to reach their desired target. Nanoparticles are exceptionally well adapted to overcoming these limitations due

to their nanoscale size and ability to surface functionalize to encompass desired properties.

Important nutrients, oxygen, and other molecules are transported throughout the body via blood vessels. Circulatory system plays an important role in continuous transportation of materials in the body. The endothelium of the blood vessels has been classified as continuous, fenestrated, or discontinuous (sinusoidal), depending on the arrangement of cells. Arteries and vessels of the brain, lungs, skin, and heart have continuous endothelium. Fenestrated endo-

thelium is found in capillaries of exocrine and endocrine glands, gastric and intestinal mucosa, choroid plexus, glomeruli, and a subpopulation of renal tubules (fenestrae of approximately 70 nm in diameter). Discontinuous endothelium is found in certain sinusoidal vascular beds, most particularly the liver with fenestrations of 100–200 nm in diameter [1]. Blood vessel endothelial cells can react to the physiological conditions, culminating in angiogenic activity. The creation of new blood vessels is known as angiogenesis. Endothelial cells, which line the inside walls of blood arteries, migrate, proliferate, and differentiate during this process. Chemical impulses in the body influence the process of angiogenesis. Angiogenesis results in a weak lymphatic drainage system and a faulty hyper-vasculature during tumor growth [60]. These openings help the passive movement of nanoparticles to target tumors through the enhanced permeability and retention effect (EPR) [41], wherein the macromolecules or nanoparticles accumulate and diffuse into tumor tissue releasing the therapeutic drug locally [5, 6, 26].

3 Factors Affecting Biodistribution and Clearance of Nanoparticles

Over the last few years, research investigators have developed various types of nanoparticles with exclusive functions and characteristics for targeting purpose. Characteristics such as different therapeutic or imaging functions, special drug loading and release competences, particle sizes, type of materials, different surface charges, hydrophilic or hydrophobic properties, biodegradability, biocompatibility, and different molecular-targeting capabilities aid the nanoparticles to achieve the desired performance. However, among all the listed properties of nanoparticles, only four factors are considered to be critical for their biodistribution and clearance, namely, particle size, shape, surface charge, and surface modification. Table 14.1 demonstrates various polymeric nanoparticles and the factors associated with their pharmacokinetics. Figure 14.3 illustrates various forms of polymeric nanoparticles.

Table 14.1 Factors affecting pharmacokinetics of various polymeric nanoparticles

Polymeric nanoparticles	Pharmacokinetics	Factor	Reference
PLGA/polyvinyl acid nanospheres	Hepatic uptake	Particle size: 200 nm	Di Mascolo et al. [23]
PLAcore/PVAshell nanoparticles	Hepatic uptake	Particle size: 273.1 nm	Canup et al. [8]
Chitosan	Hepatic uptake	Particle size: 210–279 nm	Xiao et al. [88]
Polyethylenimine nanoparticles	Hepatic uptake	Particle size: 150–200 nm	Iranpur Mobarakeh et al. [39]
Polymeric nanoparticle	Lung and spleen uptake	Shape: Rod	Kolhar et al. [46]
Copolymer of poly[(ethylene glycol) methyl ether methacrylate] and poly(glycidyl methacrylate) nanoparticles	RES uptake	Shape: Cylinder Particle size: 35–1200 nm	Müllner et al. [58]
Glyceryl monostearate nanoparticles	Spleen uptake	Particle size: 350–500 nm	Patil et al. [64]
Carboxylated polystyrene nanoparticles	Lungs uptake	Surface modification: ICAM-antibody-coating	Anselmo et al. [2]
Phosphatidylcholine/cholesterol liposomes	Liver uptake	Surface charge	Levchenko et al. [50]
PLGA nanoparticles	Tumor uptake	Surface modification with chitosan or Eudragit® RS 100	Kırımlioğlu and Görgülü [44]
Polystyrene nanoparticles	M1 macrophage uptake reduced	Surface modification with PEG	Qie et al. [68]
PEG-b-PLA nanoparticles	Tumor uptake enhanced	Positive charge	Wang et al. [81]

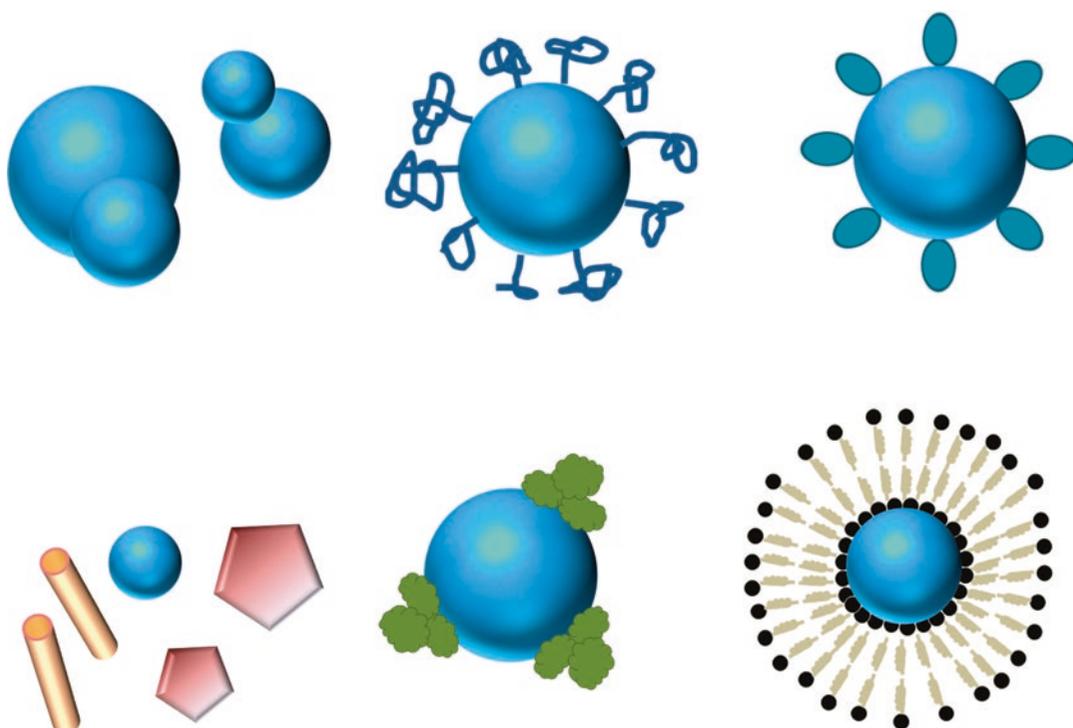


Fig. 14.3 Various shapes and surface modifications of nanoparticles

3.1 Particle Size

It is obvious, based on physiological characteristics including hepatic filtration, tissue extravasation, tissue diffusion, and kidney excretion, that the size of the nanoparticles has a significant impact on their distribution and clearance. The systemic life of employed nanoparticles in blood is determined by not only the organs' clearance efficiency, their ability to target tumors using the EPR effect, and the degradation duration in the blood, but also the particles' physicochemical features. Thus, as one of the most important attributes of a nanoparticle, size should be addressed first when tailoring drug loading and other therapeutic characteristics. Furthermore, polymeric nanoparticles' aggregation ability in tumors may be reduced if they are removed too quickly, yet too long retention in the body might contribute to greater toxicity. As a result, a nanoparticle size optimization is necessary to improve the applications of nanoparticles.

Circulation half-lives, extravasation through leaky vasculature, and macrophage uptake are all driven by size and have discrete cut-off size ranges. Various researchers have investigated optimized nanoparticle size for systemic retention resulting in desired therapeutic efficacy. From the studies, it is concluded that particle size less than 6 nm are easily filtered out through the kidney [16, 54]. In one study, *in vivo* biodistribution results of PLGA nanoparticles with consistent composition and particle size 160 nm showed intrahepatic delivery and therapeutic efficacy [48]. In another study, the *in vivo* spleen administration of the degradable poly(amine-co-ester) nanoparticles encapsulating siRNA protein with a size of 240–300 nm allowed up to 60% Nogo-B protein suppression [17]. Particles in the micrometer range have been demonstrated to efficiently aggregate within pulmonary capillaries, potentially providing a special benefit when targeting among the most major intersections of metastatic malignancy [32]. Pore sizes in leaky tumor vasculatures have been observed to range between

380 and 780 nm [35, 93]. Thus, due to the EPR effect, only nanoparticles smaller than 600 nm can be employed. In one study, PLGA nanoparticles encapsulating curcumin were investigated for cellular uptake by cervical cancer cells. Results demonstrated that polymeric curcumin nanoparticles of 132 nm in size were targeted to P-glycoprotein on the cell surface membrane of KB-V1 cells [67]. Drug carriers' size has a significant impact on their *in vivo* circulation time. It is found that as particle size increases, *in vivo* circulation time also increases [65]. This does not, however, imply that an unlimited increase in particle size leads to an infinitely long *in vivo* circulation duration. If merely considering the size difference in nanoparticle clearance, nanoparticles with diameters between 100 and 200 nm are more acceptable for utilization since they have a longer blood circulation and a lower rate of MPS absorption [47, 86, 94]. However, the size of nanoparticles is determined by the desired properties of nanoparticles and their application significance. For instance, a few smaller nanoparticles have been developed by some investigators because of their ease of clearance, which reduces the risk of chronic toxicity, or their high infiltration and retaining behavior, which improves the targeting outcome [20].

3.2 Particle Shape

The spherical shape of nanoparticles is the most frequent since it has the fewest dimensions and is the easiest to make. Diverse forms of nanoparticles have been found to have a major impact on their biodistribution and clearance, giving specific roles in medical applications. Wire [38], sphere [25], ellipsoid [21], rod [18], cylinder [37], sheet [94], cube [56], needle [45], and cluster-like [11] nanoparticle forms have been identified. The effect of shape of nanoparticles can be traced to a variety of factors. For instance, it is observed that nanoparticles with irregular shapes were more likely to settle in the spleen [22].

Various investigators have reported selective uptake of spherical nanoparticles by MPS over

rod-shaped counterparts [14, 15]. Li and co-researchers reported that cellular uptake of nanoparticles was in the sequence of sphere > cube > rod > disk in an *in vitro* cell uptake examination of various shaped PEGylated nanoparticles. This is likely owing to the ease of folding the cell membrane around the particles [51]. The oblate form of particles helps them circulate in the bloodstream because macrophages have lower uptake [72]. Intravenously injected filamentous micelles lasted ten times longer in circulation than spherical polymersomes, lasting up to 1 week [27]. Rod-like camptothecin-conjugated PEGylated dendrimers [99] showed faster cell uptake *in vitro* and longer circulation half-life along with high tumor uptake *in vivo* as compared to nanospheres. In one study, the renal system efficiently cleared single-walled carbon nanotubes with rod lengths ranging from 100 to 500 nm and diameters of 0.8–1.2 nm; driven by flow-induced orientation, the rods' long-axis went readily through the glomerular capillary fenestrations [70].

3.3 Surface Charge

Remarkably, certain nanoparticles' surface physicochemical characteristics can alter when used *in vivo*, modifying biodistribution and clearance. It has been proven that a polymeric nanoparticle's physicochemical attributes, such as surface charge and functional groups, can influence its uptake by phagocytic cells. For instance, when compared to neutral or negatively charged formulations, positively charged nanoparticles have a higher rate of cell uptake. In a study, less negatively charged rhodamine B-carboxymethyl chitosan-grafted nanoparticles and more positively charged rhodamine B-chitosan hydrochloride-grafted nanoparticles were tended to be more efficiently internalized by both L02 and SMMC-7721 cells, suggesting that surface charge played an important role in cellular uptake of polymeric nanoparticles [33]. In most situations, the nanoparticle surface contributes the driving forces for cellular internalization (electrostatic, hydrophobic, and hydrophilic (polar)

forces) and determines the uptake pathway. Nanoparticles with a positively charged surface are projected to have a high nonspecific internalization rate because of their effective binding to negatively charged groups on the cell surface and tend to have a short half-life in the bloodstream [63, 79, 94, 95]. Furthermore, an excess of positive charge on the complexes can result in nonspecific binding and absorption by cells that are not targeted. For complexes containing a targeting ligand, a weakly positively charged surface is desirable for receptor-mediated endocytosis selective binding. In a study, an amphoteric hyaluronic acid derivative with polyethylenimine chains for gene delivery overcome the disadvantages of polyethylenimine as gene carrier including the cytotoxicity caused by excess of positive charge, non-specific interaction and aggregation in the blood, and non-target gene delivery [92]. In another study, effect of surface charge on the cellular uptake and in vivo fate of PEG-oligocholeic acid-based micellar nanoparticles was reported. After opsonization in fresh mouse serum, RAW 264.7 murine macrophages took up nanoparticles with a high surface charge, whether positive or negative. In vivo biodistribution experiments

revealed that strongly positively or negatively charged nanoparticles had very high liver absorption, which is likely owing to active phagocytosis by macrophages in the liver [87].

3.4 Surface Modification

Due to the ease of detection by the RES, most uncoated nanoparticles can be swiftly removed from blood circulation when utilized in vivo, resulting in a significant loss in targeting ability [76]. Surface functionalization of nanoparticles mainly comprises PEG, the negative carboxyl ($-\text{COOH}$) group, neutral functional groups like hydroxyl ($-\text{OH}$) groups, and the positive amine ($-\text{NH}_2$) group. The increase in ($-\text{NH}_2$) resulted in a higher positive surface charge, which increased the cellular uptake of nanoparticles [55]. Rate of blood clearance of non-PEGylated nanoparticles is found to be more than PEGylated nanoparticles. PEGylation provides stealth properties to the nanoparticles surface protecting further from uptake by MPS (Fig. 14.4) [24, 91]. In one investigation, in vivo study in ICR mice showed polyethylene glycol and heparin (PEG/

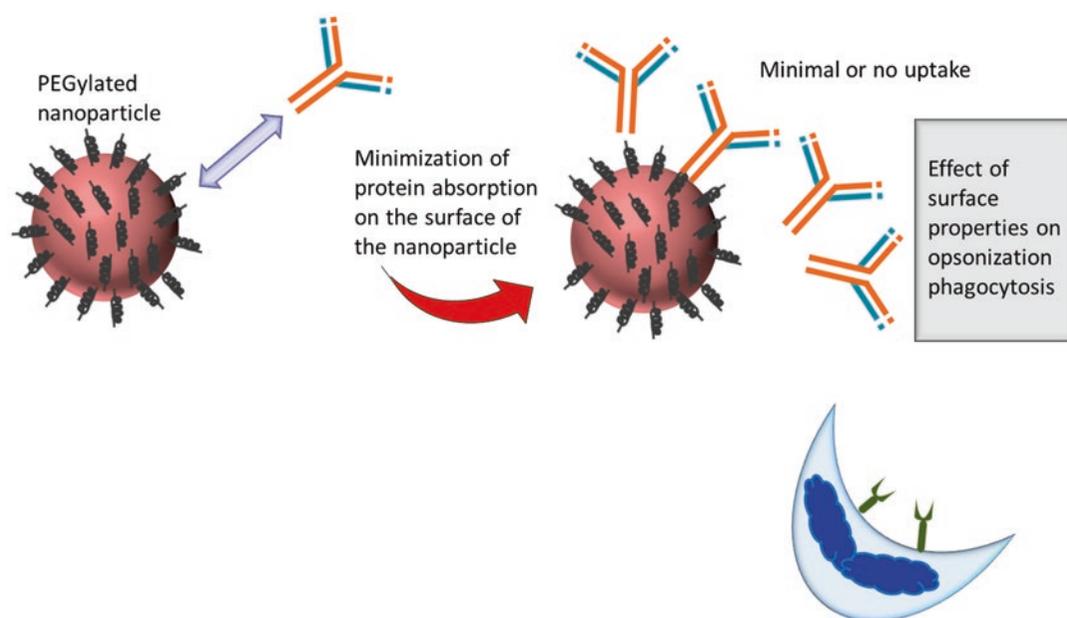


Fig. 14.4 Effect of PEGylation on nanoparticle pharmacokinetics

HEP) coating increased the blood circulation half-life of lipid polymer hybrid nanoparticles (LPHNPs) from 0.3 to 72.6 h. Moreover, PEG/HEP LPHNPs exhibited dramatically reduced liver accumulation when compared to LPHNPs [73]. In a similar study, researchers had developed polyethylene glycol and human serum albumin coating of nanoparticles carrying resveratrol for pancreatic tumor therapy. The surface-modified nanoparticles demonstrated prolonged blood circulation approximately 5.43-fold [28]. Doxorubicin-loaded nanoparticles based on polyethylene glycol-conjugated chitosan oligosaccharide-arachidic acid were explored for potential application to leukemia therapy. Results illustrated higher uptake of the conjugated nanoparticles by K562 cells with slower in vivo clearance rate, subsequently extending the blood circulation [77].

4 Conclusion

The development of a complete concept of nanoparticle pharmacokinetics in order to recognize their distribution and clearance is influenced by physiological factors and nanoparticle component such as particle size, shape, surface charge, and modification. Various investigators have been studying these characteristics since long time. Because the interactions between nanoparticles and the body are so complicated and variable, it is important to understand and follow past knowledge and principles for nanoparticle design.

References

- Aird WC. Phenotypic heterogeneity of the endothelium: I. Structure, function, and mechanisms. *Circ Res.* 2007;100(2):158–73. <https://doi.org/10.1161/01.RES.0000255691.76142.4a>.
- Anselmo AC, Kumar S, Gupta V, Pearce AM, Ragusa A, Muzykantov V, Mitragotri S. Exploiting shape, cellular-hitchhiking and antibodies to target nanoparticles to lung endothelium: synergy between physical, chemical and biological approaches. *Biomaterials.* 2015;68:1–8. <https://doi.org/10.1016/j.biomaterials.2015.07.043>.
- Begines B, Ortiz T, Pérez-Aranda M, Martínez G, Merinero M, Argüelles-Arias F, Alcudia A. Polymeric nanoparticles for drug delivery: recent developments and future prospects. *Nanomaterials (Basel).* 2020;10(7):1403. <https://doi.org/10.3390/nano10071403>.
- Behzadi S, Serpooshan V, Tao W, Hamaly MA, Alkawareek MY, Dreaden EC, Brown D, Alkilany AM, Farokhzad OC, Mahmoudi M. Cellular uptake of nanoparticles: journey inside the cell. *Chem Soc Rev.* 2017;46(14):4218–44. <https://doi.org/10.1039/c6cs00636a>.
- Bertrand N, Wu J, Xu X, Kamaly N, Farokhzad OC. Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. *Adv Drug Deliv Rev.* 2014;66:2–25. <https://doi.org/10.1016/j.addr.2013.11.009>.
- Bort G, Lux F, Dufort S, Crémillieux Y, Verry C, Tillement O. EPR-mediated tumor targeting using ultrasmall-hybrid nanoparticles: from animal to human with theranostic AGuIX nanoparticles. *Theranostics.* 2020;10(3):1319–31. <https://doi.org/10.7150/thno.37543>.
- Bottino DC, Patel M, Kadakia E, Zhou J, Patel C, Neuwirth R, Iartchouk N, Brake R, Venkatakrishnan K, Chakravarty A. Dose optimization for anticancer drug combinations: maximizing therapeutic index via clinical exposure-toxicity/preclinical exposure-efficacy modeling. *Clin Cancer Res.* 2019;25(22):6633–43. <https://doi.org/10.1158/1078-0432.CCR-18-3882>.
- Canup BS, Song H, Le Ngo V, Meng X, Denning TL, Garg P, Laroui H. CD98 siRNA-loaded nanoparticles decrease hepatic steatosis in mice. *Dig Liver Dis.* 2017;49(2):188–96. <https://doi.org/10.1016/j.dld.2016.11.008>.
- Chambers E, Mitragotri S. Prolonged circulation of large polymeric nanoparticles by non-covalent adsorption on erythrocytes. *J Control Release.* 2004;100(1):111–9. <https://doi.org/10.1016/j.jconrel.2004.08.005>.
- Chao Y, Makale M, Karmali PP, Sharikov Y, Tsigelny I, Merkulov S, Kesari S, Wrasidlo W, Ruoslahti E, Simberg D. Recognition of dextran-superparamagnetic iron oxide nanoparticle conjugates (Feridex) via macrophage scavenger receptor charged domains. *Bioconjug Chem.* 2012;23(5):1003–9. <https://doi.org/10.1021/bc200685a>.
- Chen K, Liao S, Guo S, Zheng X, Wang B, Duan Z, Zhang H, Gong Q, Luo K. Multistimuli-responsive PEGylated polymeric bioconjugate-based nano-aggregate for cancer therapy. *Chem Eng J.* 2020;391:123543. <https://doi.org/10.1016/j.cej.2019.123543>.
- Chen S, Zhong Y, Fan W, Xiang J, Wang G, Zhou Q, Wang J, Geng Y, Sun R, Zhang Z, Piao Y, Wang J, Zhuo J, Cong H, Jiang H, Ling J, Li Z, Yang D, Yao X, Xu X, Zhou Z, Tang J, Shen Y. Enhanced tumour penetration and prolonged circulation in blood of polyzwitterion-drug conjugates with cell-membrane

- affinity. *Nat Biomed Eng.* 2021;5(9):1019–37. <https://doi.org/10.1038/s41551-021-00701-4>.
13. Chenthamara D, Subramaniam S, Ramakrishnan SG, Krishnaswamy S, Essa MM, Lin FH, Qoronfleh MW. Therapeutic efficacy of nanoparticles and routes of administration. *Biomater Res.* 2019;23:20. <https://doi.org/10.1186/s40824-019-0166-x>.
 14. Chithrani BD, Chan WC. Elucidating the mechanism of cellular uptake and removal of protein-coated gold nanoparticles of different sizes and shapes. *Nano Lett.* 2007;7(6):1542–50. <https://doi.org/10.1021/nl070363y>.
 15. Chithrani BD, Ghazani AA, Chan WC. Determining the size and shape dependence of gold nanoparticle uptake into mammalian cells. *Nano Lett.* 2006;6(4):662–8. <https://doi.org/10.1021/nl052396o>.
 16. Choi HS, Liu W, Misra P, Tanaka E, Zimmer JP, Itty Ipe B, Bawendi MG, Frangioni JV. Renal clearance of quantum dots. *Nat Biotechnol.* 2007;25(10):1165–70. <https://doi.org/10.1038/nbt1340>.
 17. Cui J, Piotrowski-Daspit AS, Zhang J, Shao M, Braccaglia LG, Utsumi T, Seo YE, DiRito J, Song E, Wu C, Inada A, Tietjen GT, Pober JS, Iwakiri Y, Saltzman WM. Poly(amine-co-ester) nanoparticles for effective Nogo-B knockdown in the liver. *J Control Release.* 2019;304:259–67. <https://doi.org/10.1016/j.jconrel.2019.04.044>.
 18. Darwish WMA, Bayoumi NA. Gold nanorod-loaded (PLGA-PEG) nanocapsules as near-infrared controlled release model of anticancer therapeutics. *Lasers Med Sci.* 2020;35(8):1729–40. <https://doi.org/10.1007/s10103-020-02964-w>.
 19. de Freitas CSM, Soares AN. Efficacy of Leuproreline acetate (Eligard®) in daily practice in Brazil: a retrospective study with depot formulations in patients with prostate cancer. *Int Braz J Urol.* 2020;46(3):383–9. <https://doi.org/10.1590/S1677-5538.IBJU.2019.0212>.
 20. Dehaini D, Fang RH, Luk BT, Pang Z, Hu CM, Kroll AV, Yu CL, Gao W, Zhang L. Ultra-small lipid-polymer hybrid nanoparticles for tumor-penetrating drug delivery. *Nanoscale.* 2016;8(30):14411–9. <https://doi.org/10.1039/c6nr04091h>.
 21. Desai P, Venkataramanan A, Schneider R, Jaiswal MK, Carrow JK, Purwada A, Singh A, Gaharwar AK. Self-assembled, ellipsoidal polymeric nanoparticles for intracellular delivery of therapeutics. *J Biomed Mater Res A.* 2018;106(7):2048–58. <https://doi.org/10.1002/jbm.a.36400>.
 22. Devarajan PV, Jindal AB, Patil RR, Mulla F, Gaikwad RV, Samad A. Particle shape: a new design parameter for passive targeting in splenotropic drug delivery. *J Pharm Sci.* 2010;99(6):2576–81. <https://doi.org/10.1002/jps.22052>.
 23. Di Mascolo D, Lyon CJ, Aryal S, Ramirez MR, Wang J, Candeloro P, Guindani M, Hsueh WA, Decuzzi P. Rosiglitazone-loaded nanospheres for modulating macrophage-specific inflammation in obesity. *J Control Release.* 2013;170(3):460–8. <https://doi.org/10.1016/j.jconrel.2013.06.012>.
 24. Essa S, Rabanel JM, Hildgen P. Characterization of rhodamine loaded PEG-g-PLA nanoparticles (NPs): effect of poly(ethylene glycol) grafting density. *Int J Pharm.* 2011;411(1–2):178–87. <https://doi.org/10.1016/j.ijpharm.2011.02.039>.
 25. Evans CW, Latter MJ, Ho D, Peerzade SAMA, Clemons TD, Fitzgerald M, Dunlop SA, Iyer KS. Multimodal and multifunctional stealth polymer nanospheres for sustained drug delivery. *New J Chem.* 2012;36:1457–62. <https://doi.org/10.1039/C2NJ40016B>.
 26. Ge Z, Liu S. Functional block copolymer assemblies responsive to tumor and intracellular micro-environments for site-specific drug delivery and enhanced imaging performance. *Chem Soc Rev.* 2013;42(17):7289–325. <https://doi.org/10.1039/c3cs60048c>.
 27. Geng Y, Dalhaimer P, Cai S, Tsai R, Tewari M, Minko T, Discher DE. Shape effects of filaments versus spherical particles in flow and drug delivery. *Nat Nanotechnol.* 2007;2(4):249–55. <https://doi.org/10.1038/nnano.2007.70>.
 28. Geng T, Zhao X, Ma M, Zhu G, Yin L. Resveratrol-loaded albumin nanoparticles with prolonged blood circulation and improved biocompatibility for highly effective targeted pancreatic tumor therapy. *Nanoscale Res Lett.* 2017;12(1):437. <https://doi.org/10.1186/s11671-017-2206-6>.
 29. Goel N, Stephens S. Certolizumab pegol. *MAbs.* 2010;2(2):137–47. <https://doi.org/10.4161/mabs.2.2.11271>.
 30. Gonzalez-Valdivieso J, Girotti A, Muñoz R, Rodriguez-Cabello JC, Arias FJ. Self-assembling ELR-based nanoparticles as smart drug-delivery systems modulating cellular growth via Akt. *Biomacromolecules.* 2019;20(5):1996–2007. <https://doi.org/10.1021/acs.biomac.9b00206>.
 31. Gordon EM, Hall FL. REXIN-G, a targeted genetic medicine for cancer. *Expert Opin Biol Ther.* 2010;10(5):819–32. <https://doi.org/10.1517/14712598.2010.481666>.
 32. Harsha NS, Rani RHS. Drug targeting to lungs by way of microspheres. *Arch Pharm Res.* 2006;29:598–604. <https://doi.org/10.1007/BF02969272>.
 33. He C, Hu Y, Yin L, Tang C, Yin C. Effects of particle size and surface charge on cellular uptake and biodistribution of polymeric nanoparticles. *Biomaterials.* 2010;31(13):3657–66. <https://doi.org/10.1016/j.biomaterials.2010.01.065>.
 34. Herzog C, Hartmann K, Künzi V, Kürsteiner O, Mischler R, Lazar H, Glück R. Eleven years of Inflflex V-a virosomal adjuvanted influenza vaccine. *Vaccine.* 2009;27(33):4381–7. <https://doi.org/10.1016/j.vaccine.2009.05.029>.
 35. Hobbs SK, Monsky WL, Yuan F, Roberts WG, Griffith L, Torchilin VP, Jain RK. Regulation of transport pathways in tumor vessels: role of tumor type and microenvironment. *Proc Natl Acad Sci U S A.* 1998;95(8):4607–12. <https://doi.org/10.1073/pnas.95.8.4607>.

36. Hu J, Li HY, Williams GR, Yang HH, Tao L, Zhu LM. Electrospun poly(N-isopropylacrylamide)/ethyl cellulose nanofibers as thermoresponsive drug delivery systems. *J Pharm Sci.* 2016;105(3):1104–12. [https://doi.org/10.1016/S0022-3549\(15\)00191-4](https://doi.org/10.1016/S0022-3549(15)00191-4).
37. Hubbe H, Mendes E, Boukany PE. Polymeric nanowires for diagnostic applications. *Micromachines (Basel).* 2019;10(4):225. <https://doi.org/10.3390/mi10040225>.
38. Iranpur Mobarakeh V, Modarressi MH, Rahimi P, Bolhassani A, Arefian E, Atyabi F, Vahabpour R. Optimization of chitosan nanoparticles as an anti-HIV siRNA delivery vehicle. *Int J Biol Macromol.* 2019;129:305–15. <https://doi.org/10.1016/j.ijbiomac.2019.02.036>.
39. Juretić D, Golemac A, Strand DE, Chung K, Ilić N, Goić-Barišić I, Pellay FX. The spectrum of design solutions for improving the activity-selectivity product of peptide antibiotics against multidrug-resistant bacteria and prostate cancer PC-3 cells. *Molecules.* 2020;25(15):3526. <https://doi.org/10.3390/molecules25153526>.
40. Kalyane D, Raval N, Maheshwari R, Tambe V, Kalia K, Tekade RK. Employment of enhanced permeability and retention effect (EPR): nanoparticle-based precision tools for targeting of therapeutic and diagnostic agent in cancer. *Mater Sci Eng C Mater Biol Appl.* 2019;98:1252–76. <https://doi.org/10.1016/j.msec.2019.01.066>.
41. Kanaparthy A, Kukura S, Slenkovich N, AlGhamdi F, Shafy SZ, Hakim M, Tobias JD. Perioperative Administration of Emend® (Aprepitant) at a Tertiary Care Children's Hospital: a 12-month survey. *Clin Pharmacol.* 2019;11:155–60. <https://doi.org/10.2147/CPAA.S221736>.
42. Kim JY, Do YR, Song HS, Cho YY, Ryoo HM, Bae SH, Kim JG, Chae YS, Kang BW, Baek JH, Kim MK, Lee KH, Park K. Multicenter Phase II Clinical Trial of Genexol-PM® with gemcitabine in advanced biliary tract cancer. *Anticancer Res.* 2017;37(3):1467–73. <https://doi.org/10.21873/anticancerres.11471>.
43. Kırmıoğlu GY, Görgülü S. Surface modification of PLGA nanoparticles with chitosan or Eudragit® RS 100: characterization, prolonged release, cytotoxicity, and enhanced antimicrobial activity. *J Drug Deliv Sci Technol.* 2021;61:102145. <https://doi.org/10.1016/j.jddst.2020.102145>.
44. Kolhar P, Doshi N, Mitragotri S. Polymer nanoneedle-mediated intracellular drug delivery. *Small.* 2011;7(14):2094–100. <https://doi.org/10.1002/sml.201100497>.
45. Kolhar P, Anselmo AC, Gupta V, Pant K, Prabhakarandian B, Ruoslahti E, Mitragotri S. Using shape effects to target antibody-coated nanoparticles to lung and brain endothelium. *Proc Natl Acad Sci U S A.* 2013;110(26):10753–8. <https://doi.org/10.1073/pnas.1308345110>.
46. Kulkarni SA, Feng SS. Effects of particle size and surface modification on cellular uptake and biodistribution of polymeric nanoparticles for drug delivery. *Pharm Res.* 2013;30(10):2512–22. <https://doi.org/10.1007/s11095-012-0958-3>.
47. Kurniawan DW, Jajoriya AK, Dhawan G, Mishra D, Argemi J, Bataller R, Storm G, Mishra DP, Prakash J, Bansal R. Therapeutic inhibition of spleen tyrosine kinase in inflammatory macrophages using PLGA nanoparticles for the treatment of non-alcoholic steatohepatitis. *J Control Release.* 2018;288:227–38. <https://doi.org/10.1016/j.jconrel.2018.09.004>.
48. Lee KSS, Yang J, Niu J, Ng CJ, Wagner KM, Dong H, Kodani SD, Wan D, Morisseau C, Hammock BD. Drug-target residence time affects in vivo target occupancy through multiple pathways. *ACS Cent Sci.* 2019;5(9):1614–24. <https://doi.org/10.1021/acscentsci.9b00770>.
49. Levchenko TS, Rammohan R, Lukyanov AN, Whiteman KR, Torchilin VP. Liposome clearance in mice: the effect of a separate and combined presence of surface charge and polymer coating. *Int J Pharm.* 2002;240(1–2):95–102. [https://doi.org/10.1016/S0378-5173\(02\)00129-1](https://doi.org/10.1016/S0378-5173(02)00129-1).
50. Li Y, Kroger M, Liu WK. Shape effect in cellular uptake of PEGylated nanoparticles: comparison between sphere, rod, cube and disk. *Nanoscale.* 2015;7:16631–46. <https://doi.org/10.1039/C5NR02970H>.
51. Li M, Jiang S, Simon J, Paßlick D, Frey ML, Wagner M, Mailänder V, Crespy D, Landfester K. Brush conformation of polyethylene glycol determines the stealth effect of nanocarriers in the low protein adsorption regime. *Nano Lett.* 2021;21(4):1591–8. <https://doi.org/10.1021/acs.nanolett.0c03756>.
52. Lin YS, Hurley KR, Haynes CL. Critical considerations in the biomedical use of mesoporous silica nanoparticles. *J Phys Chem Lett.* 2012;3(3):364–74. <https://doi.org/10.1021/jz2013837>.
53. Longmire M, Choyke PL, Kobayashi H. Clearance properties of nano-sized particles and molecules as imaging agents: considerations and caveats. *Nanomedicine (Lond).* 2008;3(5):703–17. <https://doi.org/10.2217/17435889.3.5.703>.
54. Lorenz MR, Holzapfel V, Musyanovych A, Nothelfer K, Walther P, Frank H, Landfester K, Schrezenmeier H, Mailänder V. Uptake of functionalized, fluorescent-labeled polymeric particles in different cell lines and stem cells. *Biomaterials.* 2006;27(14):2820–8. <https://doi.org/10.1016/j.biomaterials.2005.12.022>.
55. Margulis K, Zhang X, Joubert LM, Bruening K, Tassone CJ, Zare RN, Waymouth RM. Formation of polymeric nanocubes by self-assembly and crystallization of dithiolane-containing triblock copolymers. *Angew Chem Int Ed Engl.* 2017;56(51):16357–62. <https://doi.org/10.1002/anie.201709564>.
56. Mitchell MJ, Billingsley MM, Haley RM, Wechsler ME, Peppas NA, Langer R. Engineering precision nanoparticles for drug delivery. *Nat Rev Drug Discov.* 2021;20(2):101–24. <https://doi.org/10.1038/s41573-020-0090-8>.
57. Müllner M, Dodds SJ, Nguyen TH, Senyschyn D, Porter CJ, Boyd BJ, Caruso F. Size and rigidity of cylindrical polymer brushes dictate long circulating

- properties in vivo. *ACS Nano*. 2015;9(2):1294–304. <https://doi.org/10.1021/nn505125f>.
58. Nima ZA, Alwbari AM, Dantuluri V, Hamzah RN, Sra N, Motwani P, Arnaoutakis K, Levy RA, Bohliq AF, Nedosekin D, Zharov VP, Makhoul I, Biris AS. Targeting nano drug delivery to cancer cells using tunable, multi-layer, silver-decorated gold nanorods. *J Appl Toxicol*. 2017;37(12):1370–8. <https://doi.org/10.1002/jat.3495>.
59. Nishida N, Yano H, Nishida T, Kamura T, Kojiro M. Angiogenesis in cancer. *Vasc Health Risk Manag*. 2006;2(3):213–9. <https://doi.org/10.2147/vhrm.2006.2.3.213>.
60. Numata M, Grinkova YV, Mitchell JR, Chu HW, Sligar SG, Voelker DR. Nanodiscs as a therapeutic delivery agent: inhibition of respiratory syncytial virus infection in the lung. *Int J Nanomedicine*. 2013;8:1417–27. <https://doi.org/10.2147/IJN.S39888>.
61. Owens DE 3rd, Peppas NA. Opsonization, biodistribution, and pharmacokinetics of polymeric nanoparticles. *Int J Pharm*. 2006;307(1):93–102. <https://doi.org/10.1016/j.ijpharm.2005.10.010>.
62. Panariti A, Miserocchi G, Rivolta I. The effect of nanoparticle uptake on cellular behavior: disrupting or enabling functions? *Nanotechnol Sci Appl*. 2012;5:87–100. <https://doi.org/10.2147/NSA.S25515>.
63. Patil RR, Gaikwad RV, Samad A, Devarajan PV. Role of lipids in enhancing splenic uptake of polymer-lipid (LIPOMER) nanoparticles. *J Biomed Nano*. 2008;4(3):359–66. <https://doi.org/10.1166/jbn.2008.320>.
64. Perrault SD, Walkey C, Jennings T, Fischer HC, Chan WC. Mediating tumor targeting efficiency of nanoparticles through design. *Nano Lett*. 2009;9(5):1909–15. <https://doi.org/10.1021/nl900031y>.
65. Prabhakar U, Maeda H, Jain RK, Sevick-Muraca EM, Zamboni W, Farokhzad OC, Barry ST, Gabizon A, Grodzinski P, Blakey DC. Challenges and key considerations of the enhanced permeability and retention effect for nanomedicine drug delivery in oncology. *Cancer Res*. 2013;73(8):2412–7. <https://doi.org/10.1158/0008-5472.CAN-12-4561>.
66. Punfa W, Yodkeeree S, Pitchakarn P, Ampasavate C, Limtrakul P. Enhancement of cellular uptake and cytotoxicity of curcumin-loaded PLGA nanoparticles by conjugation with anti-P-glycoprotein in drug resistance cancer cells. *Acta Pharmacol Sin*. 2012;33(6):823–31. <https://doi.org/10.1038/aps.2012.34>.
67. Qie Y, Yuan H, von Roemeling CA, Chen Y, Liu X, Shih KD, Knight JA, Tun HW, Wharen RE, Jiang W, Kim BY. Corrigendum: surface modification of nanoparticles enables selective evasion of phagocytic clearance by distinct macrophage phenotypes. *Sci Rep*. 2016;6:30663. <https://doi.org/10.1038/srep30663>.
68. Rizvi SAA, Saleh AM. Applications of nanoparticle systems in drug delivery technology. *Saudi Pharm J*. 2018;26(1):64–70. <https://doi.org/10.1016/j.jsps.2017.10.012>.
69. Ruggiero A, Villa CH, Bander E, Rey DA, Bergkvist M, Batt CA, Manova-Todorova K, Deen WM, Scheinberg DA, McDevitt MR. Paradoxical glomerular filtration of carbon nanotubes. *Proc Natl Acad Sci U S A*. 2010;107(27):12369–74. <https://doi.org/10.1073/pnas.0913667107>.
70. Salmaso S, Caliceti P. Stealth properties to improve therapeutic efficacy of drug nanocarriers. *J Drug Deliv*. 2013;2013:374252. <https://doi.org/10.1155/2013/374252>.
71. Sharma G, Valenta DT, Altman Y, Harvey S, Xie H, Mitragotri S, Smith JW. Polymer particle shape independently influences binding and internalization by macrophages. *J Control Release*. 2010;147(3):408–12. <https://doi.org/10.1016/j.jconrel.2010.07.116>.
72. Sheng Y, Chang L, Kuang T, Hu J. PEG/heparin-decorated lipid-polymer hybrid nanoparticles for long-circulating drug delivery. *RSC Adv*. 2016;6:23279–87. <https://doi.org/10.1039/C5RA26215A>.
73. Song G, Petschauer JS, Madden AJ, Zamboni WC. Nanoparticles and the mononuclear phagocyte system: pharmacokinetics and applications for inflammatory diseases. *Curr Rheumatol Rev*. 2014;10(1):22–34. <https://doi.org/10.2174/1573403x10666140914160554>.
74. Suk JS, Xu Q, Kim N, Hanes J, Ensign LM. PEGylation as a strategy for improving nanoparticle-based drug and gene delivery. *Adv Drug Deliv Rev*. 2016;99(Pt A):28–51. <https://doi.org/10.1016/j.addr.2015.09.012>.
75. Termsarasab U, Yoon IS, Park JH, Moon HT, Cho HJ, Kim DD. Polyethylene glycol-modified arachidyl chitosan-based nanoparticles for prolonged blood circulation of doxorubicin. *Int J Pharm*. 2014;464(1–2):127–34. <https://doi.org/10.1016/j.ijpharm.2014.01.015>.
76. Torres-Martinez EJ, Cornejo Bravo JM, Serrano Medina A, Pérez González GL, Villarreal Gómez LJ. A summary of electrospun nanofibers as drug delivery system: drugs loaded and biopolymers used as matrices. *Curr Drug Deliv*. 2018;15(10):1360–74. <https://doi.org/10.2174/1567201815666180723114326>.
77. Verma A, Stellacci F. Effect of surface properties on nanoparticle-cell interactions. *Small*. 2010;6(1):12–21. <https://doi.org/10.1002/sml.200901158>.
78. Walkey CD, Olsen JB, Guo H, Emili A, Chan WC. Nanoparticle size and surface chemistry determine serum protein adsorption and macrophage uptake. *J Am Chem Soc*. 2012;134(4):2139–47. <https://doi.org/10.1021/ja2084338>.
79. Wang HX, Zuo ZQ, Du JZ, Wang YC, Sun R, Cao ZT, Ye XD, Wang JL, Leong KW, Wang J. Surface charge critically affects tumor penetration and therapeutic efficacy of cancer nanomedicines. *NanoToday*. 2016;11(2):133–44. <https://doi.org/10.1016/j.nantod.2016.04.008>.
80. Wang J, Hu X, Xiang D. Nanoparticle drug delivery systems: an excellent carrier for tumor peptide vaccines. *Drug Deliv*. 2018;25(1):1319–27. <https://doi.org/10.1080/10717544.2018.1477857>.

81. Wang Y, Zhou C, Ding Y, Liu M, Tai Z, Jin Q, Yang Y, Li Z, Yang M, Gong W, Gao C. Red blood cell-hitchhiking chitosan nanoparticles for prolonged blood circulation time of vitamin K1. *Int J Pharm.* 2021;592:120084. <https://doi.org/10.1016/j.ijpharm.2020.120084>.
82. Wen R, Umeano AC, Kou Y, Xu J, Farooqi AA. Nanoparticle systems for cancer vaccine. *Nanomedicine (Lond).* 2019;14(5):627–48. <https://doi.org/10.2217/nmm-2018-0147>.
83. Wilhelm S, Tavares AJ, Dai Q, Ohta S, Audet J, Dvorak HF, Chan WCW. Analysis of nanoparticle delivery to tumours. *Nat Rev Mater.* 2016;1:16014. <https://doi.org/10.1038/natrevmats.2016.14>.
84. Wisse E, Jacobs F, Topal B, Frederik P, De Geest B. The size of endothelial fenestrae in human liver sinusoids: implications for hepatocyte-directed gene transfer. *Gene Ther.* 2008;15(17):1193–9. <https://doi.org/10.1038/gt.2008.60>.
85. Xiao K, Li Y, Luo J, Lee JS, Xiao W, Gonik AM, Agarwal RG, Lam KS. The effect of surface charge on in vivo biodistribution of PEG-oligocholic acid based micellar nanoparticles. *Biomaterials.* 2011;32(13):3435–46. <https://doi.org/10.1016/j.biomaterials.2011.01.021>.
86. Xiao B, Ma P, Ma L, Chen Q, Si X, Walter L, Merlin D. Effects of tripolyphosphate on cellular uptake and RNA interference efficiency of chitosan-based nanoparticles in Raw 264.7 macrophages. *J Colloid Interface Sci.* 2017;490:520–8. <https://doi.org/10.1016/j.jcis.2016.11.088>.
87. Yadav D, Dewangan H. PEGYLATION: an important approach for novel drug delivery system. *J Biomater Sci Polym Ed.* 2021;32(2):266–80. <https://doi.org/10.1080/09205063.2020.1825304>.
88. Yamashita F, Hashida M. Pharmacokinetic considerations for targeted drug delivery. *Adv Drug Deliv Rev.* 2013;65(1):139–47. <https://doi.org/10.1016/j.addr.2012.11.006>.
89. Yang Q, Jones SW, Parker CL, Zamboni WC, Bear JE, Lai SK. Evading immune cell uptake and clearance requires PEG grafting at densities substantially exceeding the minimum for brush conformation. *Mol Pharm.* 2014;11(4):1250–8. <https://doi.org/10.1021/mp400703d>.
90. Yao J, Fan Y, Du R, Zhou J, Lu Y, Wang W, Ren J, Sun X. Amphoteric hyaluronic acid derivative for targeting gene delivery. *Biomaterials.* 2010;31(35):9357–65. <https://doi.org/10.1016/j.biomaterials.2010.08.043>.
91. Yuan F, Dellian M, Fukumura D, Leunig M, Berk DA, Torchilin VP, Jain RK. Vascular permeability in a human tumor xenograft: molecular size dependence and cutoff size. *Cancer Res.* 1995;55(17):3752–6.
92. Yue ZG, Wei W, Lv PP, Yue H, Wang LY, Su ZG, Ma GH. Surface charge affects cellular uptake and intracellular trafficking of chitosan-based nanoparticles. *Biomacromolecules.* 2011;12(7):2440–6. <https://doi.org/10.1021/bm101482r>.
93. Zhang J, Liu J, Zhao Y, Wang G, Zhou F. Plasma and cellular pharmacokinetic considerations for the development and optimization of antitumor block copolymer micelles. *Expert Opin Drug Deliv.* 2015;12(2):263–81. <https://doi.org/10.1517/17425247.2014.945417>.
94. Zhang YN, Poon W, Tavares AJ, McGilvray ID, Chan WCW. Nanoparticle-liver interactions: cellular uptake and hepatobiliary elimination. *J Control Release.* 2016;240:332–48. <https://doi.org/10.1016/j.jconrel.2016.01.020>.
95. Zhao F, Zhao Y, Liu Y, Chang X, Chen C, Zhao Y. Cellular uptake, intracellular trafficking, and cytotoxicity of nanomaterials. *Small.* 2011;7(10):1322–37. <https://doi.org/10.1002/sml.201100001>.
96. Zhao Y, Wei C, Chen X, Liu J, Yu Q, Liu Y, Liu J. Drug delivery system based on near-infrared light-responsive molybdenum disulfide nanosheets controls the high-efficiency release of dexamethasone to inhibit inflammation and treat osteoarthritis. *ACS Appl Mater Interfaces.* 2019;11(12):11587–601. <https://doi.org/10.1021/acsami.8b20372>.
97. Zhou Z, Ma X, Jin E, Tang J, Sui M, Shen Y, Van Kirk EA, Murdoch WJ, Radosz M. Linear-dendritic drug conjugates forming long-circulating nanorods for cancer-drug delivery. *Biomaterials.* 2013;34:5722–35. <https://doi.org/10.1016/j.biomaterials.2013.04.012>.