William Charles Kreisl

17

Introduction

Microglia are immunocompetent cells in the central nervous system with various physiological functions, such as synaptic pruning during neural development, providing trophic support, and clearing neuronal debris. In response to cellular injury, microglia undergo morphological changes, with reduction in the size and number of their processes, and accompanying shift toward pro-infammatory activities [\[1](#page-4-0)]. These include release of cytokines, activation of complement, and generation of reactive oxygen species. Chronic, dysfunctional infammatory microglial activity, possibly in response to aggregation of misfolded proteins, has been considered a pathogenic contributor to neurodegenerative disease.

PET imaging of microglial activation, targeting proteins expressed by microglia, allows in vivo quantifcation of neuroinfammation. Unlike measurement of soluble infammatory biomarkers found in cerebrospinal fuid, such as sTREM2 or YKL-40, PET imaging allows spatial information regarding neuroinfammatory changes in the brain.

The most commonly employed microglial target for PET imaging of neuroinfammation is the 18 kDa translocator protein (TSPO). TSPO is a mitochondrial protein (coded by nuclear DNA, not mitochondrial DNA) with proposed roles in cholesterol transport, steroid synthesis, and heme metabolism. Alternative pathways may compensate for loss of TSPO function, as null knockout mice have been shown to be viable and fertile [[2\]](#page-4-1). TSPO is expressed in several tissues, including peripheral monocytes, heart, lung, kidneys, adrenals, gonads, and vascular epithelium. In the brain parenchyma, TSPO is mainly expressed by microglia but also by astrocytes [\[3](#page-4-2)]. Under physiological conditions, TSPO expression is low in the brain. However, in response to cellular injury, the density of TSPO increases. Whether this increase in TSPO density is due to increased TSPO expression per microglia, or increased number of microglia by migration to the site of injury, may depend on the species and mechanism of injury involved [\[4](#page-4-3)]. Recruitment of peripheral myeloid cells into the brain may also contribute to TSPO density [\[5](#page-4-4)]. The exact role of TSPO in neuroinfammation is not well established, but evidence suggests this protein may infuence mitochondrial reactivity, release of reactive oxygen species, and activation of the inflammasome [[6\]](#page-4-5).

PET Radiotracers

The frst PET radioligand developed for TSPO was 11C-*(R)*-PK 11195, an isoquinoline carboxamide. Studies have demonstrated increased 11C-*(R)*-PK 11195 signal in neurodegenerative disorders such as Alzheimer's disease, frontotemporal dementia, Huntington's disease, and parkinsonian disorders (see [\[7](#page-4-6)] for review). Increased 11C-*(R)*-PK 11195 binding has also been shown in patients with multiple sclerosis, HIV infection, stroke, and epilepsy. However, ${}^{11}C$ - (R) -PK 11195 has the disadvantage of low ratio of specifc-to-nonspecifc binding, resulting in low signal-to-noise on imaging [\[8](#page-4-7)].

Second-generation TSPO radioligands with improved ratio of specifc-to-nonspecifc binding include 11C-PBR28, 11C-DPA-713, 18F-DPA-714, 11C-DAA1106, and 18F-FEPPA. A shared limitation of second generation radioligands is their sensitivity to a common polymorphism on the *TSPO* gene that variably affects the binding affnity [[9\]](#page-4-8). Therefore, individuals who have two gene copies of the rare allele (lowaffnity binders) have lower binding than those who only express the common allele (high-affnity binders), while those heterozygous have intermediate binding since they express TSPO with both high- and low-affnity binding sites (mixed affnity binders). In Caucasians and African-Americans, the allelic frequency of the polymorphism is about 30%, resulting in approximately 9% of the population

Microglial Activation and Neuroinflammation

W. C. Kreisl (\boxtimes)

The Taub Institute for Research on Alzheimer's Disease and the Aging Brain, Columbia University Irving Medical Center, New York, NY, USA e-mail[: wck2107@cumc.columbia.edu](mailto:wck2107@cumc.columbia.edu)

[©] The Author(s), under exclusive license to Springer Nature Switzerland AG 2022 191

A. M. Franceschi, D. Franceschi (eds.), *Hybrid PET/MR Neuroimaging*, [https://doi.org/10.1007/978-3-030-82367-2_17](https://doi.org/10.1007/978-3-030-82367-2_17#DOI)

being low-affnity binders. Exclusion of low-affnity binders (identifed by genetic analysis or binding assays performed on peripheral blood cells), and statistical correction for variance in binding caused by presence or absence of one copy of the rare allele, has allowed use of second-generation TSPO radioligands in clinical PET studies [[10\]](#page-4-9).

A recently developed "third-generation" TSPO radioligand, 11C-ER176, has reduced sensitivity to the *TSPO* polymorphism, allowing inclusion of low-affnity binders but still requiring statistical correction of variance created by the presence of the polymorphism $[11]$. ¹¹C-ER176 also has higher signal-to-noise than some second-generation radioligands and does not generate brain-penetrant radiometabolites $[12]$ $[12]$ $[12]$. ¹⁸F-GE-180 has demonstrated high signal-to-noise in rodent studies [[13](#page-4-12)]. However, its poor brain uptake in clinical studies has raised concerns about the ability of 18F-GE-180 to effectively measure TSPO density in humans [[14](#page-4-13), [15\]](#page-4-14).

While TSPO is the most studied target for infammatory PET radioligands, other proteins have been proposed. Monoamine oxidase B (MAO-B) is an enzyme expressed by astrocytes and serotonin-releasing neurons. MAO-B not only is involved in monoamine metabolism but also has prooxidative functions by generating hydrogen peroxide. Increased MAO-B expression has been demonstrated in reactive astrocytosis, similar to the elevation seen in glial fbrillary acidic protein [[16\]](#page-4-15). Radioligands used to target MAO-B include ¹¹C-deuterium-L-deprenyl, ¹¹C-SL25.1188 [\[17](#page-4-16)], and ¹⁸F-SMBT-1 [[18\]](#page-4-17).

Radioligands for several other infammatory targets are currently under development (Table [17.1](#page-1-0)). These targets include cyclooxygenase 1 (COX-1), colony stimulating factor 1 receptor (CSF1R), and the purinergic P2X7 receptor. Each is predominantly expressed by microglia in the brain. However, to date, radioligands for these novel targets are in early developmental stages and have not yet been well validated in human disease models.

Image Interpretation

TSPO is diffusely expressed throughout the brain, and therefore, no true reference region exists. Accordingly, accurate quantifcation of TSPO density in the brain requires kinetic modeling using the metabolite-corrected arterial input function. While this method allows absolute measurement of TSPO, the need for arterial sampling and plasma analysis is not practical for clinical use. Alternative methods that obviate the need for arterial catheterization have therefore been developed; however, these come with trade-offs such as underestimation bias. The use of a "pseudo-reference" region, one where pathological expression of TSPO is expected to be low or absent (e.g., the cerebellum in patients with Alzheimer's disease), has been used to measure relative binding, similar to the standard uptake value ratio approach commonly applied to amyloid and tau PET imaging [\[19](#page-4-18)]. Similar strategies have been used for kinetic modeling of PET data, which provides more robust quantifcation, either using an a priori pseudo-reference region or a supervised cluster analysis, in which voxels that behave as if they are devoid of specifc binding are identifed and grouped to create a data-driven reference region [[20\]](#page-4-19). Currently, no consen-

Target protein	Cell expression in the brain	Role in inflammation	Current radioligands
18 kDa translocator protein (TSPO)	Microglia Astrocytes Also expressed by peripheral monocytes	Possible role in mitochondrial reactivity and activation of the inflammasome	${}^{11}C-(R)$ -PK 11195 11 C-PBR28 11 C-DPA-713 11 C-DAA1106 11 C-ER176 18 F-FEPPA 18 F-DPA-714 ${}^{18}F$ -GE180
Monoamine oxidase B (MAO-B)	Astrocytes Serotonin-releasing neurons	Generation of hydrogen peroxide	11 C-deuterium-L- deprenyl 11 C-SL25.1188 18 F-SMBT-1
Cyclooxygenase 1	Microglia	Facilitates pro-inflammatory prostaglandin upregulation	11 C-PS13
Colony stimulating factor 1 receptor $(CSF1-R)$	Microglia Also expressed by peripheral macrophages	Controls the activation and survival of macrophages and microglia	11 C-CPPC
Purinergic P2X7 receptor (P2X7R)	Microglia Oligodendrocytes Astrocytes Also expressed by peripheral macrophages	Inflammatory cytokine release	11 C-JNJ-54173717 ¹⁸ F-JNJ-64413739 11 C-SMW139

Table 17.1 Neuroinflammatory proteins with currently available PET radioligands

sus exists on which is the preferred method of quantifcation, as this may vary depending on which radioligand is used and for what clinical application.

Clinical Application

TSPO PET studies have been performed in several neurological and psychiatric diseases, often with disparate results, the latter likely resulting from differences in methodologies used across laboratories. The most consistent positive results have been demonstrated in patients with Alzheimer's disease, major depressive disorder, and traumatic brain injury.

Alzheimer's Disease

Several studies using the prototypical TSPO radioligand ${}^{11}C$ -*(R)*-PK 11195 have shown increased binding in patients with Alzheimer's disease than in age-matched control participants [[21–](#page-4-20)[23\]](#page-4-21). Most studies using second-generation radioligands, when correcting for the *TSPO* polymorphism, have shown similar results [[24,](#page-4-22) [25](#page-4-23)]. Studies have additionally shown that increased TSPO binding is associated with worse cognitive performance, cortical volume loss, and the extent of tau pathology (Fig. [17.1](#page-2-0)) [[26,](#page-4-24) [27\]](#page-4-25). While multiple longitudinal studies have shown overall increase in TSPO binding in patients with Alzheimer's disease over time [\[28,](#page-4-26) [29](#page-4-27)], results

from one study demonstrated an interval decrease in TSPO signal in patients with mild cognitive impairment [\[30](#page-4-28)]. A meta-analysis of 28 TSPO PET studies showed increased binding in mild cognitive impairment and Alzheimer's disease patients, particularly in neocortical regions, with inverse association between TSPO and Mini-Mental Status Exam score in the parietal lobe [\[31\]](#page-4-29).

A small number of PET studies have used non-TSPO radioligands to detect infammation in patients with Alzheimer's disease. One study showed increased MAO-B binding in patients with mild cognitive impairment, while decreased in patients with Alzheimer's disease, suggesting that increased astrocytosis might be limited to early clinical stages of the disease [\[32](#page-4-30)]. A study using arachidonic acid PET imaging showed increased radioligand turnover in Alzheimer's disease patients [\[33](#page-4-31)].

Major Depressive Disorder

Several studies have shown increased TSPO binding in patients with major depressive disorder, particularly in untreated patients during major depressive episodes (Fig. [17.2\)](#page-3-0) [[34](#page-4-32)[–39](#page-4-33)]. These include studies using ${}^{11}C$ -*(R)*-PK 11195 and second-generation radioligand studies using either ¹¹C-PBR28 or 18F-FEPPA. One study using 11C-PBR28 in nine patients with depression (eight untreated, one treated) and ten healthy control participants reported no difference in TSPO binding between groups [[40\]](#page-4-34). However, this negative study included

Fig. 17.1 Patterns of increased TSPO binding mirror those of neurodegeneration in patients with different clinical variants of Alzheimer's disease. Surface-based projection maps showing differences in 11C-PBR28 binding (left) and cortical thickness (right) among patients with posterior cortical atrophy (PCA) and amnestic

Alzheimer's disease (AD) and in healthy controls (HC). Voxels with threshold $P < 0.05$ correcting for family-wise error are displayed and color bars denote *T*-values. (Adapted from Kreisl et al., Distinct patterns of increased translocator protein in posterior cortical atrophy and amnestic Alzheimer's disease, Neurobiology of Aging, 2017)

Fig. 17.2 Statistical significance maps rendered on axial slices and semi-infated surface showing increase in 11C-PBR28 binding in unmedicated depressed patients compared to healthy controls. Only clusters with threshold $p < 0.05$ for cluster-wise multiple comparisons

are displayed. The color bar represents T values. (Adapted from Richards et al. PET radioligand binding to translocator protein (TSPO) is increased in unmedicated depressed subjects. EJNMMI Res. 2018; 8: 57)

fewer participants than the mentioned positive TSPO studies. Moreover, patients experiencing a major depressive episode who also had elevated ¹⁸F-FEPPA binding were more likely to experience reduction in mood symptoms after treatment with the anti-infammatory drug celecoxib [[41\]](#page-4-35).

Traumatic Brain Injury

Increased TSPO binding has been detected with both $¹¹C-(R)$ -PK 11195 and ¹¹C-PBR28 in participants with prior</sup> history of traumatic brain injury [\[42](#page-5-0), [43](#page-5-1)]. Two studies using ¹¹C-DPA-713 were performed in players from the National Football League, one that included active and recently retired football players and one that included former players 57–74 years of age. Both studies showed widespread increase in TSPO binding, with particularly high signal in the supramarginal gyrus [\[44](#page-5-2), [45\]](#page-5-3). However, elevated TSPO signal was not associated with differences in cognitive performance between NFL players and controls. Therefore, the clinical signifcance of these neuroinfammatory changes detected in active and retired professional football players remains to be seen.

PET/MRI

Clinical TSPO studies using simultaneous PET/MRI modalities are limited. However, those studies using TSPO PET/ MRI demonstrate both the feasibility of this technique in human research and potential advantages. For example, one

study performed dual PET/MRI using 18F-DPA-714 in nine patients with suspected primary angiitis of the central nervous system, a rare infammatory disorder that affects the cerebrovasculature and is associated with changes on MRI, such as multifocal ischemic and hemorrhagic lesions, vessel wall and leptomeningeal enhancement, and stenosis of the blood vessels [[46\]](#page-5-4). Two patients in this study showed focal increase in TSPO binding that extended beyond the areas of contrast enhancement or restricted diffusion on MRI and were confrmed to have primary angiitis of the central nervous system on biopsy. Three patients with primary angiitis of the central nervous system did not have increased 18F-DPA-714 binding despite MRI changes. However, these three were on immunosuppressive therapy for a longer period of time than the TSPO-positive patients, which may have affected the PET results. Four patients were determined to have strokes not associated with vasculitis, and those cases showed variable amounts of 18F-DPA-714 uptake, not extending beyond the area of infarction seen on MRI.

In another TSPO PET/MRI study, ¹¹C-PBR28 detected increased TSPO binding in veterans with Gulf War illness in several cortical regions, including precuneus, prefrontal, primary motor, and somatosensory cortices [\[47](#page-5-5)]. TSPO binding was not related to clinical symptoms or concentrations of peripheral infammatory markers.

In a PET/MRI study of patients with Huntington's disease, 11C-PBR28 binding was greater in patients than controls in the putamen and pallidum [[48\]](#page-5-6). Increased TSPO signal was also detectable at the single subject level, with some participants demonstrating binding in the thalamic and brain stem nuclei thought to be related to visual and motor function.

References

- 1. Wang M, et al. Macroglia-microglia interactions via TSPO signaling regulates microglial activation in the mouse retina. J Neurosci. 2014;34(10):3793–806.
- 2. Tu LN, et al. Peripheral benzodiazepine receptor/translocator protein global knock-out mice are viable with no effects on steroid hormone biosynthesis. J Biol Chem. 2014;289(40):27444–54.
- 3. Gui Y, et al. Characterization of the 18 kDa translocator protein (TSPO) expression in post-mortem normal and Alzheimer's disease brains. Brain Pathol. 2020;30(1):151–64.
- 4. Tournier BB, et al. Fluorescence-activated cell sorting to reveal the cell origin of radioligand binding. J Cereb Blood Flow Metab. 2020;40(6):1242–55.
- 5. Pannell M, et al. Imaging of translocator protein upregulation is selective for pro-infammatory polarized astrocytes and microglia. Glia. 2020;68(2):280–97.
- 6. Feng H, et al. TSPO ligands PK11195 and midazolam reduce NLRP3 infammasome activation and proinfammatory cytokine release in BV-2 cells. Front Cell Neurosci. 2020;14:544431.
- 7. Kreisl WC, et al. PET imaging of neuroinfammation in neurological disorders. Lancet Neurol. 2020;19(11):940–50.
- 8. Kobayashi M, et al. (11)C-DPA-713 has much greater specifc binding to translocator protein 18 kDa (TSPO) in human brain than (11)C-(R)-PK11195. J Cereb Blood Flow Metab. 2018;38(3):393–403.
- 9. Owen DR, et al. An 18-kDa translocator protein (TSPO) polymorphism explains differences in binding affnity of the PET radioligand PBR28. J Cereb Blood Flow Metab. 2012;32(1):1–5.
- 10. Kreisl WC, et al. A genetic polymorphism for translocator protein 18 kDa affects both in vitro and in vivo radioligand binding in human brain to this putative biomarker of neuroinfammation. J Cereb Blood Flow Metab. 2013;33(1):53–8.
- 11. Ikawa M, et al. 11C-ER176, a radioligand for 18-kDa translocator protein, has adequate sensitivity to robustly image all three affnity genotypes in human brain. J Nucl Med. 2017;58(2):320–5.
- 12. Fujita M, et al. Comparison of four (11)C-labeled PET ligands to quantify translocator protein 18 kDa (TSPO) in human brain: (R)- PK11195, PBR28, DPA-713, and ER176-based on recent publications that measured specifc-to-non-displaceable ratios. EJNMMI Res. 2017;7(1):84.
- 13. Boutin H, et al. 18F-GE-180: a novel TSPO radiotracer compared to 11C-R-PK11195 in a preclinical model of stroke. Eur J Nucl Med Mol Imaging. 2015;42(3):503–11.
- 14. Feeney C, et al. Kinetic analysis of the translocator protein positron emission tomography ligand [(18)F]GE-180 in the human brain. Eur J Nucl Med Mol Imaging. 2016;43(12):2201–10.
- 15. Zanotti-Fregonara P, et al. Head-to-head comparison of (11) C-PBR28 and (18)F-GE180 for quantifcation of the translocator protein in the human brain. J Nucl Med. 2018;59(8):1260–6.
- 16. Ekblom J, et al. Reactive gliosis and monoamine oxidase B. J Neural Transm Suppl. 1994;41:253–8.
- 17. Moriguchi S, et al. Monoamine oxidase B total distribution volume in the prefrontal cortex of major depressive disorder: an [11C] SL25.1188 positron emission tomography study. JAMA Psychiat. 2019;76(6):634–41.
- 18. Harada R, et al. (18)F-SMBT-1: a selective and reversible PET tracer for monoamine oxidase-B imaging. J Nucl Med. 2021;62(2):253–8.
- 19. Lyoo CH, et al. Cerebellum can serve as a pseudo-reference region in Alzheimer disease to detect Neuroinfammation measured with PET Radioligand binding to translocator protein. J Nucl Med. 2015;56(5):701–6.
- 20. Zanotti-Fregonara P, et al. Automatic extraction of a reference region for the noninvasive quantifcation of translocator protein in brain using (11)C-PBR28. J Nucl Med. 2019;60(7):978–84.
- 21. Edison P, et al. Microglia, amyloid, and cognition in Alzheimer's disease: an [11C](R)PK11195-PET and [11C]PIB-PET study. Neurobiol Dis. 2008;32(3):412–9.
- 22. Parbo P, et al. Brain infammation accompanies amyloid in the majority of mild cognitive impairment cases due to Alzheimer's disease. Brain. 2017;140(7):2002–11.
- 23. Okello A, et al. Microglial activation and amyloid deposition in mild cognitive impairment: a PET study. Neurology. 2009;72(1):56–62.
- 24. Kreisl WC, et al. In vivo radioligand binding to translocator protein correlates with severity of Alzheimer's disease. Brain. 2013;136(Pt 7):2228–38.
- 25. Suridjan I, et al. In-vivo imaging of grey and white matter neuroinfammation in Alzheimer's disease: a positron emission tomography study with a novel radioligand, [18F]-FEPPA. Mol Psychiatry. 2015;20(12):1579–87.
- 26. Zou J, et al. Microglial activation, but not tau pathology, is independently associated with amyloid positivity and memory impairment. Neurobiol Aging. 2020;85:11–21.
- 27. Dani M, et al. Microglial activation correlates in vivo with both tau and amyloid in Alzheimer's disease. Brain. 2018;141(9):2740–54.
- 28. Kreisl WC, et al. (11)C-PBR28 binding to translocator protein increases with progression of Alzheimer's disease. Neurobiol Aging. 2016;44:53–61.
- 29. Fan Z, et al. Longitudinal infuence of microglial activation and amyloid on neuronal function in Alzheimer's disease. Brain. 2015;138(Pt 12):3685–98.
- 30. Fan Z, et al. An early and late peak in microglial activation in Alzheimer's disease trajectory. Brain. 2017;140(3):792–803.
- 31. Bradburn S, Murgatroyd C, Ray N. Neuroinfammation in mild cognitive impairment and Alzheimer's disease: a meta-analysis. Ageing Res Rev. 2019;50:1–8.
- 32. Carter SF, et al. Evidence for astrocytosis in prodromal Alzheimer disease provided by 11C-deuterium-L-deprenyl: a multitracer PET paradigm combining 11C-Pittsburgh compound B and 18F-FDG. J Nucl Med. 2012;53(1):37–46.
- 33. Esposito G, et al. Imaging neuroinfammation in Alzheimer's disease with radiolabeled arachidonic acid and PET. J Nucl Med. 2008;49(9):1414–21.
- 34. Richards EM, et al. PET radioligand binding to translocator protein (TSPO) is increased in unmedicated depressed subjects. EJNMMI Res. 2018;8(1):57.
- 35. Setiawan E, et al. Association of translocator protein total distribution volume with duration of untreated major depressive disorder: a cross-sectional study. Lancet Psychiatry. 2018;5(4):339–47.
- 36. Holmes SE, et al. Elevated translocator protein in anterior cingulate in major depression and a role for infammation in suicidal thinking: a positron emission tomography study. Biol Psychiatry. 2018;83(1):61–9.
- 37. Li H, Sagar AP, Keri S. Translocator protein (18kDa TSPO) binding, a marker of microglia, is reduced in major depression during cognitive-behavioral therapy. Prog Neuro-Psychopharmacol Biol Psychiatry. 2018;83:1–7.
- 38. Li H, Sagar AP, Keri S. Microglial markers in the frontal cortex are related to cognitive dysfunctions in major depressive disorder. J Affect Disord. 2018;241:305–10.
- 39. Setiawan E, et al. Role of translocator protein density, a marker of neuroinfammation, in the brain during major depressive episodes. JAMA Psychiat. 2015;72(3):268–75.
- 40. Hannestad J, et al. The neuroinfammation marker translocator protein is not elevated in individuals with mild-to-moderate depression: a [(1)(1)C]PBR28 PET study. Brain Behav Immun. 2013;33:131–8.
- 41. Attwells S, et al. Translocator protein distribution volume predicts reduction of symptoms during open-label trial of celecoxib in major depressive disorder. Biol Psychiatry. 2020;88(8):649–56.
- 42. Scott G, et al. Minocycline reduces chronic microglial activation after brain trauma but increases neurodegeneration. Brain. 2018;141(2):459–71.
- 43. Folkersma H, et al. Widespread and prolonged increase in (R)- (11)C-PK11195 binding after traumatic brain injury. J Nucl Med. 2011;52(8):1235–9.
- 44. Coughlin JM, et al. Imaging of glial cell activation and white matter integrity in brains of active and recently retired National Football League Players. JAMA Neurol. 2017;74(1):67–74.
- 45. Coughlin JM, et al. Neuroinfammation and brain atrophy in former NFL players: an in vivo multimodal imaging pilot study. Neurobiol Dis. 2015;74:58–65.
- 46. Backhaus P, et al. Initial experience with [(18)F]DPA-714 TSPO-PET to image inflammation in primary angiitis of the central nervous system. Eur J Nucl Med Mol Imaging. 2020;47(9):2131–41.
- 47. Alshelh Z, et al. In-vivo imaging of neuroinfammation in veterans with gulf war illness. Brain Behav Immun. 2020;87:498–507.
- 48. Lois C, et al. Neuroinfammation in Huntington's disease: new insights with (11)C-PBR28 PET/MRI. ACS Chem Neurosci. 2018;9(11):2563–71.