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Editors



Infection in Knee Replacement



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Foreword

It is an honor to write a preface for an excellent international text on Infection in Total Knee Arthroplasty (TKA). While knee arthroplasty can be traced back to 1863 when membrane interposition, including pig bladders, was used to resurface the arthritic knee joint. The modern history of TKA surface replacements began circa 1970 [1]. The first internationally recorded and widely used surface replacement designs were the Polycentric (Gunston 1969) and the Geomedic (aka Geometric 1970 USA). These anterior (ACL) and posterior cruciate (PCL) retention designs laid the foundation for an international frenzy to develop the science of non-hinged knee arthroplasty.

It is interesting to remember that the polycentric prosthesis was initially two independent femoral and tibial components which laid the groundwork for the now popular uni-compartment replacements. While the focus of the early designs was on the retention of both the ACL and PCL, John Insall and others at the Hospital for Special Surgery in New York felt that an entire ACL and PCL cruciate sacrificing prosthesis, with the first patellar replacement (Total Condylar 1974) [2], would be a better alternative. Within a few years, this cruciate-sacrificing design was modified and improved by adding the design change of an intercondylar femoral “cam” and tibial post to enhance the patient’s range of motion and in addition, intercondylar coronal and sagittal stability. This Insall Burstein Posterior Stabilized Prosthesis of 1977 was modified further in the following decades and in 2021, the cruciate-substituting design is currently used in more than 50% of all total knee procedures in the USA.

In the first edition of *Surgery of the Knee*, 1984 [3], there were 26 chapters on all aspects of knee diseases requiring arthroplasty but no chapters on infection. In fact, to the best of our knowledge, the first significant text dealing with the infected Knee Arthroplasty was in 1992, *The Knee* [1]. Well, time has certainly brought about huge changes as evidenced by the abundance of articles, texts, and conferences devoted to the infected knee arthroplasty since 1984. This book, *Infection in Knee Replacement*, is a most welcome international update.

It is important to recognize that for the first 2–3 decades in the development of knee arthroplasty the patient was very different from our present population of surgical candidates. Circa 1970–1995, the primary pathological pathway for the development of severe destructive disease was inflammatory arthritis, primarily rheumatoid. These patients typically had severe biplanar deformities with similar complicating pathology often present in the

ipsilateral and/or contralateral hip, foot, and ankle. Due to the massive amounts of steroids required for the treatment of these patients, their skin was frequently very attenuated and ulcerated, and wound closure often required skin grafting, muscle transfers, and tissue expanders [4]. The patient's deficient immune system often led to the necessity for multiple types of antibiotics with subsequent problems. Thank God for the DMARDS!

The worldwide variable operating theaters presented, in retrospect, very difficult sterility issues. These rooms were almost never dedicated to "clean cases." Sterilization procedures likewise had little resemblance to today's approaches. The surgical procedures were additionally often compromised by limited prosthetic sizes resulting in unnecessarily tight or excessive joint spaces as well as inadequate fixation which resulted in non-physiological component motion and instability. Hematomas developed and infection often followed.

This new text *Infection in Knee Replacement* is a compendium of the tremendous improvements in the treatment of one of the most dreadful complications in Knee Arthroplasty. The science attendant to the treatment of infection in Knee Replacement is extensively detailed by all the contributors in this book. Prosthetic joint infection (PJI) is, and unfortunately always will be, a very challenging therapeutic problem. It is especially apparent that when we consider that in the USA alone, the present estimate of annual knee replacements is 600,000 cases [5]. In 2030, there is a growth projection of 673% for a total number of 4,638,000 cases per annum (USA). In the world, the present projection of knee replacement cases is 1,234,000. Using the same rate of growth to 2030, the annual number of cases would be 10,234,520 in 2030. With infection rates presently ranging from 1 to 2% (primary and revision), we might anticipate upwards of 200,000 infected knee replacements per year in 2030. These are daunting numbers and require strict and updated attention to detail, in order to minimize the problem as best we can. The editors of this book accomplish this goal.

This book, *Infection in Knee Replacement*, is thorough in evaluating all the parameters that must be constantly evaluated and reassessed in minimizing TKR infections. Risk factors and prevention are a start. The patient's health must be maximized prior to surgery. Just giving prophylactic antibiotics is not enough. Each patient is somewhat different and these accomplished authors address that issue.

The diagnosis of an infected TKA is not necessarily as easy as one would expect. Precise periprosthetic joint infection (PJI) diagnosis has proven to be difficult throughout the years. The Musculoskeletal Infection Society (MSIS) has taken a worldwide leadership role in developing criteria required to establish a PJI. These criteria are identified clearly and further augmented by several critical points including a detailed and accepted identification of the pathogen. These exhaustive chapters are written by international experts who, in the past, helped to develop and currently continue to reassess the multiple complex criteria required for a PJI diagnosis. Treatment modalities from innovative pharmacological strategies are constantly changing for the better and the authors thoroughly explain the rationale.

In this text, all surgical options from Debridement, Antibiotics, and Implant Retention (DAIR), staged procedures, the use of static or dynamic spacers, arthrodesis, and unfortunately the extremely rare amputation are discussed in detail. This is a comprehensive book that will thoroughly educate the surgeon, medical physicians, and paraprofessionals who are all so crucial to the successful treatment of the PJI. The patient will be the winner thanks to the efforts of all these authors and editors who have done a tremendous service by advancing our knowledge in the prevention and treatment of the PJI.

Congratulations to the Editors, Umile Giuseppe Longo, Nicolaas C. Budhiparama, Sébastien Lustig, Roland Becker, and Joao Espregueira-Mendes, for invaluable, current, and an international perspective on a prevalent and difficult problem.

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Foreword

Knee replacement has evolved over decades to become one of the most successful interventions in orthopedic surgery. Many of the challenges that have been encountered over this time in the development of knee replacement have been addressed to a great extent. Improvements in implant materials have seen a reduction in revision surgery due to polyethylene wear, and implant design has continued to evolve to provide surgeons with consistent, reliable designs to address different severities and patterns of arthritis. New technologies such as computer navigation and robotics have allowed surgeons to perform surgery very precisely and reduce the risk of malalignment, and both cemented and cementless fixation are very reliable, with the risk of revision for loosening due to fixation failure very low. Multiple national joint registries have also assisted in informing surgeons and industry in a very objective, independent fashion about the many variables in implant design and technique that may influence outcome, particularly the risk of revision.

Prosthetic joint infection, however, remains a major challenge that is yet to be resolved and continues to be a major contributor to prosthetic revision and patient dissatisfaction. The exponential growth in the number of knee replacements performed annually across the world, coupled with the emergence of increasing numbers of resistant strains of bacteria, means that prosthetic joint infection remains a major burden for the health care community and a major challenge for orthopedic surgeons. There is a clear concern, with increasing antibiotic resistance, that infection will continue to increase in prevalence as a cause of knee replacement failure and will compromise the gains made in other areas of knee replacement technology. There is understandably great concern that, while all other areas of knee replacement surgery continue to advance, the morbidity associated with prosthetic knee joint infection may actually increase.

Infection accounts for nearly one quarter of all first revisions on the Australian National Registry, has a high rate of second revision, and is responsible for one quarter of all second revisions. Over one million knee replacements are now performed worldwide, with approximately 600,000 in the USA alone. A very conservative estimate of an infection incidence of 1% would result in 60,000 knee replacement infections requiring treatment annually in the USA alone. Prevention of primary infection is therefore of paramount importance and ideally could be achieved in over 99% of patients, reducing this burden. However, even with a reduced incidence of infection, there will still be a considerable number of infected prostheses requiring

management, and surgeons therefore require clear guidelines for managing prosthetic infection. Early, accurate diagnosis and the application of timely, evidence-based management for each individual scenario are critical to optimize the outcome for these patients and to reduce the likelihood of second revision.

The timely publication of this excellent, comprehensive book on *Infection in Knee Replacement* provides a valuable resource for the orthopedic surgeon performing knee replacement surgery, clearly informing about the latest evidence for prevention, diagnosis, and management of prosthetic joint infection. Each chapter has been methodically researched to provide an evidence-based approach, coupled with the wisdom of the experience of orthopedic surgeons renowned for their expertise in this area. The result is a reference that surgeons can turn to for the most current and valid information to guide their practice in this area. I am honored to write this foreword, and I commend the editors and their team of authors for this fine work, which is a significant contribution to the ongoing fight against prosthetic knee joint infection.

David A. Parker
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Preface

The number of total knee arthroplasty (TKA) has dramatically increased over the last few years. Due to the increasing number of primary surgery, we are facing an increase in revision surgery at the same time. The infection rate after TKA is currently about 0.7%. Infection in TKA, regardless of the early or late stage, is the most devastating situation for both patients and orthopedic surgeons. Due to the aging population, comorbidities are more likely and thus increase the risk of infection. We are dealing with a new microorganism often multiresistant and difficult to treat.

Patients require long-lasting treatment and sometimes weeks of hospitalization. The treatment of infected TKA is one of the most challenging works for the Orthopedic Surgeon. It requires aggressive treatment but often surgeons the management of knee infections requires aggressive treatments, and surgeons are usually scared for the complications is sometimes very difficult. Every painful knee after TKA should be considered for being infected until the tentative diagnosis is rejected. Finding new methods that can accurately detect an early infection is essential for rapid diagnosis and a therapeutic algorithm design. With new technologies, it is now possible to achieve a timely diagnosis and appropriate treatment, with clinical outcomes previously only imaginable.

The treatment of infected TKA is complex and requires a good team work, including the infection specialist, microbiologist, plastic surgeon, internist, and sometimes even the psychologist in order to provide the best care for these patients.

This book aims to collect the most up-to-date information regarding the optimal management of infected TKA, with the guidance of experts from all over the world.

We hope that readers will find the data they need, improving patient treatment management and allowing patients to recover “normal” knee function.

Rome, Italy
Jakarta, Indonesia
Lyon, France
Brandenburg, Germany
Braga, Portugal

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Nicolaas C. Budhiparama
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Part I

Introduction



Epidemiology and Socioeconomic Impact of Infections in Knee Replacement

1

Laura Risi Ambrogioni, Calogero Di Naro,
Vincenzo Candela, Carlo Casciaro,
Umile Giuseppe Longo, and Vincenzo Denaro

1.1 Introduction

Periprosthetic joint infection (PJI) after total joint replacement is the most serious and feared complication [1]. While more commonly observed in the postoperative period, PJIs may occur throughout life with negative physical and psychological repercussions on the patient, leading to potentially fatal consequences [2–4]. A recent meta-analysis based on a cohort of 20,719 patients who received a two-stage revision for PJI after TKA observed that the 1-year mortality rate was 4.33% and the 5-year mortality rate was 21.64% [5]. Recent studies demonstrate that PJI is the third most common indication for revision hip arthroplasty (14.7% of cases) and the leading cause for failure of total knee arthroplasty (TKA) (25.2% of cases) [6–8]. Boddapati et al. conducted a study of the American College of Surgeons National Surgical Quality Improvement Program from 2005 to 2015. During this study period, 162,981 primary TKAs and 12,780 revision TKAs were recorded, of which 17.2% were performed as a result of PJI. Apart from being the primary cause of TKA revision, PJIs are more frequently responsible for non-household resignations, readmissions and length of stay com-

pared to other non-infectious causes of TKA revision [9]. Male gender, advanced age, high body mass index, comorbidities and immunosuppression, prolonged operative time (>90') and tourniquet time (>60') have been identified as significant contributors to the development of PJI following total joint arthroplasty. Genetic predisposition has not been excluded with possible risk factor for the onset of PJIs [8, 10, 11].

Significant rise in the number of total joint arthroplasty procedures has been recorded in the USA over the years [1]. The frequency of joint replacement is estimated to grow in all countries, implying an increased risk of PJIs [12–14]. Approximately 3.5 million primary hip and knee arthroplasties will be performed in the USA alone by the year 2030 [15–18]. Similar forecasts are expected for England and Wales [19]. Moreover, future projections indicate that PJIs, compared to other causes of failure, will account for 60% of all revisions over the next two decades [20].

The treatment of PJIs commonly comprises one or two stages of surgical revision. The first approach consists of a single major surgical procedure whereby the prosthesis and infected tissues are removed while a second implant is replaced. Conversely, in the two-stage revision treatment, the replacement of a new prosthesis is performed in a second surgery. Between these two procedures, antibiotic-impregnated cement spacer is administered. Although the latter

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treatment option is preferred, it is more expensive compared to one-stage revision [21]. The use of antibiotics has been implemented in the surgical procedure to reduce the rate of PJIs. In particular, a retrospective study conducted on 15,972 US veterans showed that the use of antibiotic-laden bone cement in primary TKA is related to a lower revision rate for PJI [22, 23]. As *S. Aureus* is the primary bacterial pathogen involved in the development of PJIs, a retrospective study on 400 patients showed that a decolonization protocol based on mupirocin nasal ointment and chlorhexidine soap reduced the onset of PJI [24].

Conversely, a national joint registry study conducted on 258 consecutive primary hip and knee arthroplasties showed that the use of gentamycin-loaded bone cement at primary surgery could increase Staphylococcal resistance to gentamycin and methicillin at revision for PJI [25]. Recent evidence has suggested a possible role of anaesthesia in PJI development. In particular, it was observed that out of 3909 procedures, of which 42% were performed under general anaesthesia and 58% under spinal anaesthesia, early PJI occurred more frequently in the general anaesthesia group [26].

Therefore, it appears crucial to define the optimal prevention and treatment protocols for PJIs to reduce the socioeconomic burden across all countries.

1.2 Epidemiology and Socioeconomic Burden of PJIs in TKA

The incidence and prevalence rates of PJIs after TKA remain controversial. To date, most of the data available on PJIs are reported from monocentric studies or studies conducted in few centres [3, 27–29]. Moreover, the sample population is often small and limited to a geographical region that may not be representative of the general population, since a considerable influence of geographic variability on the risk of PJI development has been described [28].

The advent of National Joint Registry data has opened up the potential for dataset linkage, vastly

expanding the number of cases available to design prophylactic and therapeutic management strategies [30]. To date, comparisons can be conducted across five nation-wide total joint registries: the Swedish Knee Arthroplasty Register [27], the New Zealand Joint Registry [3], the National Joint Registry of England, Wales, Northern Ireland, and the Isle of Man [31], the Australian Orthopaedic Association National Joint Replacement Registry [32] and the National Inpatient Sample (NIS) database (USA) [7].

The oldest nation-wide registry study was conducted by Knutson et al. in Sweden from 1976 to 1992 on 30,003 knees, showing a steady increase in the number of TKA [27]. The incidence and prevalence rates reported in this study were calculated over a relatively distant study period. Due to the lengthening of the average life expectancy, the improvement in diagnosis and the growth in surgical procedures, it is not possible to calculate statistical inference by generalizing these rates to the overall population.

Koh et al. conducted a retrospective study on the New Zealand Joint Registry from 2000 to 2015. PJI was the most common reason for TKA revision in patients with an average age of 65 years, accounting for up to 45% of all causes of failure. The peak of incidence was observed during the first 2 years (1%), while it diminished to less than 0.2% after 5 years from the primary intervention [3, 33]. The time after surgery in which complications occurred has been widely noted. The observation that the peak of PJIs was found in the first 2 years in this national registry study is following the evidence available in the literature that ascribes 18–27% of early revisions to PJIs.

Lenguerrand et al. recorded revision knee replacements due to PJI from the National Joint Registry of England, Wales, Northern Ireland and the Isle of Man during the study period 2003–2014. The three postoperative months were identified as the most sensitive period for PJI development after TKA. Interestingly, a significantly higher rate of PJIs was observed after aseptic knee revision compared to primary knee replacement [31].

Ackerman et al. obtained data of TKA from the Australian Orthopaedic Association National

Joint Replacement Registry during 2003–2013. 350,994 TKA procedures were performed on 279,453 patients in 10 years. Even though a continuous growth in procedure rates was observed for all age groups, a significant peak was recorded in patients between 40 and 69 years of age. Furthermore, it has been reported that the costs associated with the procedure were almost doubled in 2013 compared to 2003 [32].

Kamath et al. identified the TKA revision procedures due to PJIs from the National Inpatient Sample (NIS) database (USA) during 2005–2010. The highest number of revisions for PJI was performed in white patients aged 65–69 years. The average length of stay following the revision was 7.5 days (3.5 days longer than an uninfected procedure) with a total cost of \$25,692 per patient. As found in the Australian registry study, the annual cost of a TKA revision has nearly doubled from \$320 million in 2005 to \$566 million in 2010 [7].

Although one or more organisms may be responsible for PJI, the bacterial infections are the most widespread. Fungal PJI after TKA is rare and is estimated to account for nearly 1% of all PJIs. The bacteria most frequently isolated is *Staphylococcus Aureus* accounting for 72% of all cases, whereas Gram-negative bacteria of the substantial amount ranging from 5 to 23% [19]. *S. Aureus* is a commensal bacterium of the human body that colonizes the anterior nares and extra nasal skin surfaces in approximately one-third of the population. It has been shown that being a carrier of *S. Aureus* is a risk factor for the development of PJIs. Conversely, streptococcal infections are responsible for 4–16% of all PJI and related to a poor outcome. They generally occur through haematogenic spread from infected sites such as oral cavity, heart valves, skin and soft tissue, intestinal or genitourinary tract. Despite the high sensitivity to antibiotics, total eradication is difficult in the presence of a foreign body such as a prosthesis. However, long-term oral treatment with antibiotics has been associated with a significantly better outcome in PJIs management [34].

Moreover, wide variability in the distribution of methicillin-resistant *S. aureus* (MRSA) strains has been described, ranging from 13% in Europe

to 48% in the USA [35]. Furthermore, it was observed that MRSA infections acquired in the community increased compared to those developed in the hospital. Regardless of how the microorganism is received, the PJI caused by MRSA had the highest failure rates. Besides, the increased length of stay, readmissions and reduced joint function have been associated with PJIs caused by MRSA compared to methicillin-sensitive PJIs, resulting in additional costs related to the management of methicillin-resistant PJIs [17]. The treatment of PJI does not depend on pathogen sensitivity to antibiotic therapy. However, whether the aetiological agent is atypical or not detected at culture, then a consult with an infectious disease specialist should be considered. Current guidelines provide that systemic therapy should not be discontinued until infection remission, normalization of serum markers and culture negation at least 2 weeks after cessation of antimicrobials. If laboratory data remain positive, consultation with an infectious disease specialist is required. Parvizi et al. have shown that the additional cost for the treatment of methicillin-resistant cases compared to sensitive cases was about \$20,000 [17]. Generally, it has been estimated that compared to PJI caused by microorganisms sensitive to antibiotic therapy, the costs of PJI caused by MRSA increase by about 60%. Preventive *S. Aureus* decolonization in patients undergoing TKA could be a therapeutic option to reduce the risk of PJI. According to recent evidence, the risk of PJI may also depend on the type of prosthetic material, in particular, polymethylmethacrylate cement. A cost analysis should be performed to evaluate the potential benefits of adding antibiotic-loaded polymethylmethacrylate to the primary TKA protocol in advance [36].

1.3 The Psychological Impact of PJIs

Quality of life and fear of the disease in patients with PJIs are comparable to those of oncology patients [37]. Routine screening should be conducted to identify affected patients early for appropriate treatment, improving long-term outcomes.

Patients with PJI should be followed by psychologists to maintain long-term quality of life [37]. PJIs are life-changing complications for patients. Loss of joint function, prolonged hospitalization and follow-up visits impact not only on the patient's life but also on those around him. To date, few studies on the psychological effects of PJIs are available. The inability to fulfil daily life activities in which the identity of the person is determined, the increasing dependence on relatives, depression and anxiety, as well as uncertainty about the future are the main psychological consequences of PJIs [38, 39]. The patient's psychology may affect the perception and management of pain that could affect the outcome. Recent evidence suggests that patients who received a preoperative education for TKA have a statistically reduced hospital stay of almost 2 days [40].

1.4 Conclusion

PJI after TKA is the most common, severe and feared complication with a 1-year mortality rate of 4.33% and a 5-year mortality rate of 21.64%. Several modifiable and unmodifiable risk factors have been identified, but their treatment has not yet been implemented in PJIs prevention protocols. Given the relentless increase in TKA procedures all over the world, more than half of the TKA revisions in the next two decades will be due to the occurrence of PJIs. The primary pathogen responsible for the development of PJIs is *S. Aureus*, while the family of gram-negative bacteria determines up to 20% of PJIs. Due to the spread of antibiotic resistance of *S. Aureus*, ranging from 13% in Europe to 48% in the USA, the management of PJIs is more challenging. The economic burden on national health systems for total joint replacement interventions is significant and continuously increasing. PJIs are responsible for economic overburdening due to non-household resignations, readmissions, length of stay and, eventually, drug therapy of methicillin-resistant infections. However, PJIs cannot be defined only as postoperative complications. They represent real aggression to the patients'

self-awareness and the course of their lives. The need to find preventive strategies for PJIs can no longer be postponed since the socioeconomic implications of the PJI after TKA remain overwhelming.

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Etiology and Pathogenesis of Knee Replacement Infections

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2.1 Introduction

Prosthetic joint infection (PJI) is the most dramatic complication after arthroplasty [1, 2]. Its relative incidence ranged from 1 to 5%, but can reach 50% depending on the patient comorbidity, the smoking status, and the number of prior procedures [1–6]. With the aging of the population, the absolute number of arthroplasty never stops to increase during the last decades, resulting in an ever increasing absolute number of PJI [7]. PJI is considered to be one of the most difficult-to-treat bacterial diseases, with a significant morbidity, cost, risk of relapse, and loss of function [1–5].

S. aureus is one the most frequent pathogen involved in PJI and is particularly associated with persistence and relapse [8–11]. In the USA, the annual cost of infected revisions to hospitals increased from \$320 million to \$566 million from 2001 to 2009 and was projected to exceed \$1.62 billion in 2020 [7].

As a consequence, prevent, diagnose, and treat PJI accordingly are essential. For that purpose, a better understanding of its etiology and pathogenesis is the first step.

We propose in this chapter to describe the different ways that have the bacteria to contaminate the prosthesis. It is of crucial importance, as depending on the age of the infection, the bacteria could have the opportunity to develop different mechanisms of persistence. We also plan to describe the variability of the microbiological epidemiology, especially depending on the time to PJI-onset, in the context of worldwide spread of antimicrobial resistance. Finally, we will decipher the different mechanisms involved in the bacterial persistence, such as the biofilm production and the intracellular persistence in bone cells, to finally conclude on what can bring the best knowledge of the pathophysiology in the management of PJI.

2.2 The Different Ways to Infect a Prosthetic Joint

To reach the prosthesis surface, bacteria use three main ways: (a) the inoculation could occur at the time of surgery, or during any invasive procedure

that concerns the prosthetic joint; (b) the inoculation could take as origin a contiguous infection from a nearby site that spreads gradually until the prosthesis; (c) the inoculation could occur during a bacteremia, with hematogenous seeding from a separate infectious site that could be clinically obvious or occult (Fig. 2.1) [1, 2, 12].

2.2.1 Direct Inoculation During Invasive Procedures

It is largely considered that a PJI occurring in the year following the implantation is generally associated with an intraoperative inoculation. This inoculation may come from fallout of aerosolized bacteria or from direct contamination of the operating site by bacteria on the instruments, the gloves, or the patient's own skin [1, 12]. Despite considerable progress in preoperative and intraoperative antiseptic measures, completely sterilizing the skin at the operating site remains illusory [13, 14]. It was found that skin of the incisional edges was recolonized with bacteria within 30–180 min after the antiseptic procedures [13, 14]. In addition, airborne bacteria can never be completely eliminated from operating rooms since microorganisms could not be eradicated from the skin, hair, nose, and mouth of patients and staff of the operating room [14, 15]. This inoculation could occur at the time of implantation and also during revision with a gradual increased risk depending on the number of procedures and the implant's surface. In fact, the “perioperative” inoculation also includes inoculation that could also occur in the days following the surgery, especially as the scar is not fully waterproofed and could be soiled.

2.2.2 Inoculation by Contiguous Spread

A superficial post-operative infection of the scar could occur few days after the surgery, due to the recolonization of the skin by bacteria that can then gain depth step by step, until the articular space through the tissues that are incompletely

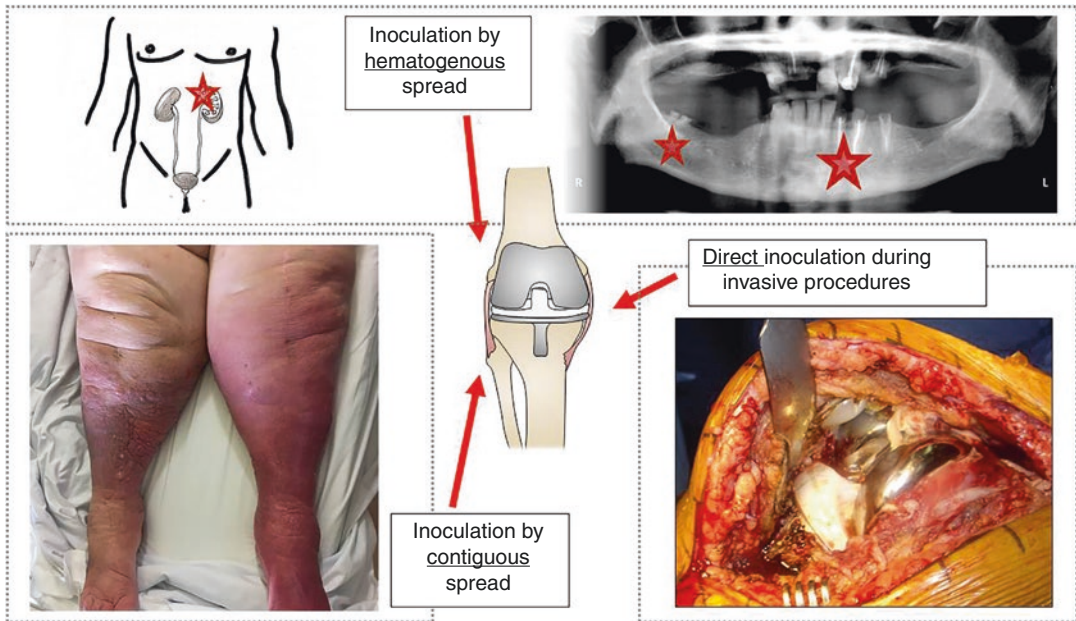


Fig. 2.1 The three different ways to infect a knee prosthesis: The most frequent way of inoculation of a knee prosthesis is a direct inoculation during invasive procedure, such as surgery, despite all the preventive measures that are usually implemented to reduce this risk. PJI could

also occur by contiguous spread of a bacterial disease, which reaches the prosthesis as during erysipelas. Finally, an acute or chronic focus of infection could be responsible for clinically obvious or occult bacteremia with secondary seeding to the prosthetic joint

healed. This is the second mechanism of inoculation that results from the contiguous spread of an adjacent infection [1, 2]. This mechanism could also occur years after the prosthesis implantation, if the patient experienced an erysipela or another skin and soft tissue infection of the index limb [16]. The typical clinical situation is the patient with a knee prosthesis, who develops extensive erysipelas (Fig. 2.1).

2.2.3 Inoculation by Hematogenous Spread

The final way of prosthesis bacterial inoculation concerns inoculation by hematogenous spread that occurs usually years after prosthesis implantation [12, 17–20]. Bacteria could enter the bloodstream from a distant infectious focus or during transient bacteremia of dental, urogenital, or gastrointestinal origin (Fig. 2.1). Few studies focused on the rates of PJI in patients with a prosthetic joint after bacteremia. This risk seems

to be extremely variable (6–40%), depending on the clinical presentation and the pathogen involved. *S. aureus* seems to be clearly associated with a high risk of hematogenous contamination of a joint prosthesis, in comparison with other bacteria [12, 17–20]. Even if the risk of PJI from a dental site of infection is considered to be low, this risk exists as: (a) occult bacteremia with odontogenic bacteria could occur in patients with chronic dental foci of infection, during surgical dental procedures, but even also during tooth brushing [21]; (b) some patients with chronic dental foci of infection can develop PJI several years after the prosthesis implantation due to typical odontogenic bacteria [22]. However, several studies concluded that given routine dental antibiotic prophylaxis prior to dental procedures for non-infected causes does not reduce the rate of PJI [23–26]. Patients with clear suppurative dental infection have to receive antimicrobial therapy for the local infection, but it has not been demonstrated if it can prevent a hematogenous location on the prosthesis. In the

absence of evidence supporting the role of antimicrobial prophylaxis for patients with a prosthetic joint undergoing dental procedure for a non-infectious cause, the International Consensus Meeting (ICM2018) and Dutch guidelines recommended that prophylaxis should be reserved to patients with extensive comorbidities in whom the probability of developing PJI is higher or those with complex reconstructive procedure in whom development of PJI may have more dire consequences [27–29].

2.3 Epidemiology

Microbiological epidemiology depends on time and way of infection. Staphylococci, that include *S. aureus* and also coagulase-negative staphylococci, are the most common pathogen involved in PJI, as they are responsible for about 60% of all PJI [30–34]. Other gram-positive pathogens (streptococci, enterococci) represent about 10–20% of PJI and gram-negative rods, 5–20% [30–34]. Beyond this global distribution, there are some important differences that have to be pointed out.

2.3.1 Knee vs. Hip Location

Most of the available data show results for pooled microbiology from knee and hip arthroplasties. However, some differences have been reported in large cohort studies. For example, the rate of *S. aureus* infection seems to be higher in the knee location, in comparison with the hip. On the contrary, a lower rate of *C. acnes* is described in prosthetic-knee infections. Finally, polymicrobial infection seems to be more represented in the hip location. These differences might be explained by a diversity of the microbial flora, which can vary according to the sites and according to the kind of surgical approach [35, 36].

2.3.2 Antimicrobial Resistance

Antimicrobial resistance is considered as a worldwide health issue, as it is considered as a

slow motion tsunami [37]. Antimicrobial resistance is particularly a key issue in the management of patients with PJI, as it impacts the antimicrobial prophylaxis, the type of antimicrobial used for the treatment (mainly intravenous), the cost, and the outcome [8, 38–43]. Antimicrobial resistance differs a lot according to the geographic area. For example, methicillin-resistant *S. aureus* represent almost 50% of the strains in the USA, while 12% in Europe (but with large differences between West vs. East, as vs. North vs. South countries) and has a range between 2 and 5% to 39% according to different countries in Asia [40, 44, 45]. Even if methicillin resistance in coagulase-negative staphylococci is less studied, the global rate of methicillin resistance in staphylococci is significant worldwide, making the prescription of a broad spectrum anti-gram positive agent (vancomycin, teicoplanin, daptomycin, or linezolid) essential as empirical antimicrobial therapy [30–32, 34, 40]. However, as these antibiotics are not active on gram-negative rods, they have to be combined with another antibiotic (usually a broad spectrum β -lactam) to complete the spectrum of activity. Resistance in gram-negative rods is clearly emergent, especially concerning Enterobacteriaceae, *P. aeruginosa*, and *Acinetobacter spp.* Indeed, multidrug- and extensively drug-resistance are increasing rapidly worldwide in such species (especially extended spectrum β -lactamases or carbapenemase in *E. coli* and *K. pneumoniae* and extensively drug resistance in *P. aeruginosa*), and these bacteria are currently not covered by usual antimicrobial prophylaxis [40, 42, 43].

2.3.3 Time to PJI-Onset

In fact, the PJI bacterial epidemiology mainly depends on the time and way of infection. For that purpose, multiple classifications have been previously described and used. The most common one classifies PJI according to the time between the prosthesis implantation and the symptoms occurrence [4, 46]:

- “Early,” if the PJI occurred in the first 3 months after arthroplasty;
- “Delayed,” if the PJI occurred between 3 months and 1 year (some suggest 2 years) after arthroplasty;
- “Late,” if the PJI occurred after 1 year (some suggest 2 years) after arthroplasty.
- Nevertheless, it’s more clinically relevant to classify the PJI according to the time of symptoms duration. In this way, a PJI can be called as [1, 47]:
- “Post-operative,” when symptom occurs within a month following the surgery. Usually, the inoculation suspected way is per-operative or due to healing trouble;
- “Acute” when symptoms last for less than 3 weeks;
- “Chronic,” when symptoms last for more than 3 weeks.

This latter classification guides the physician in the PJI management: an acute PJI can be managed with a “debridement antibiotic and implant retention” (DAIR) procedure, while a chronic one needs a prosthesis replacement. However, this classification is independent of the time of prosthesis implantation. For example, in a PJI occurring by a hematogenous way, clinical picture will be “acute,” with frequently fever, pain, and clinical signs of septic arthritis and will be managed by a DAIR strategy, even if it occurs over a year after the prosthesis implantation. That makes sense, especially as the inoculation of the bacteria on the prosthesis is recent, and as it did not usually have significant time to develop the mechanisms of persistence like the biofilm (see below). The primary bacterial focus of infection that secondary spreads to the prosthesis is most often evident clinically, and blood cultures are usually positive, especially if the primary source of infection comes from a urinary tract infection or a catheter-related infection. However, some patients develop acute clinical presentation, late after the prosthesis implantation, but there is clinically no detectable focus of primary infection. These latter patients with “late acute” PJI probably have potentially and inoculation at the time of surgery that was asymptomatic, with set up of

microbial persistence and dormant mechanism (see the pathophysiology of PJI below), until sudden appearance of clinical symptoms a long time after the implantation. Staphylococci, that are associated with a low success rate in this clinical setting, seemed to be particularly involved in such clinical presentation [11]. In clinical practice, the physician has to be stubborn in the research for the pathogen inoculation way in, and when he cannot find it, which is more frequent than we would like, he has to ask himself the question: Is it an acute presentation of an acute PJI? Or an acute presentation of a suddenly woke up of an old sleeping pathogen, with dispersal of the biofilm? Answering this question is crucial for “late acute” PJI, as if the inoculation occurred at the time of surgery, the treatment would be prosthesis exchange, or DAIR followed by suppressive antimicrobial therapy, as DAIR is not able to eradicate the biofilm. We developed in our center an algorithm, called CRIOAc Lyon’s PJI treatment algorithm, that combines clinical symptoms, the delay from the implantation, and a potential prosthesis loosening to integrate the suspected pathophysiology and the time of inoculation in the patient’s management (Fig. 2.2).

2.3.4 Main Pathogens Involved

S. aureus: Because combining virulent and persistence factors, *S. aureus* is one of the most involved pathogens in PJI. From the virulence results frequently acute presentations with fever, pain, and clinical signs of septic arthritis, while the persistence factors concern adherence proteins, intracellular persistence, and production of biofilm (see below). With this pathogen, it is noteworthy that the PJI can occur at any time from the implantation. The most well-known side of *S. aureus* is the virulent presentation, with an over-representation in post-operative and acute PJI [1, 2, 8, 34, 40, 42, 48–50]. However, it has also been described in chronic PJI, with indolent clinical signs of infection, probably by setting up only mechanisms of persistence in vivo [51].

Coagulase-negative staphylococci: The most represented pathogen of this group is *S. epider-*

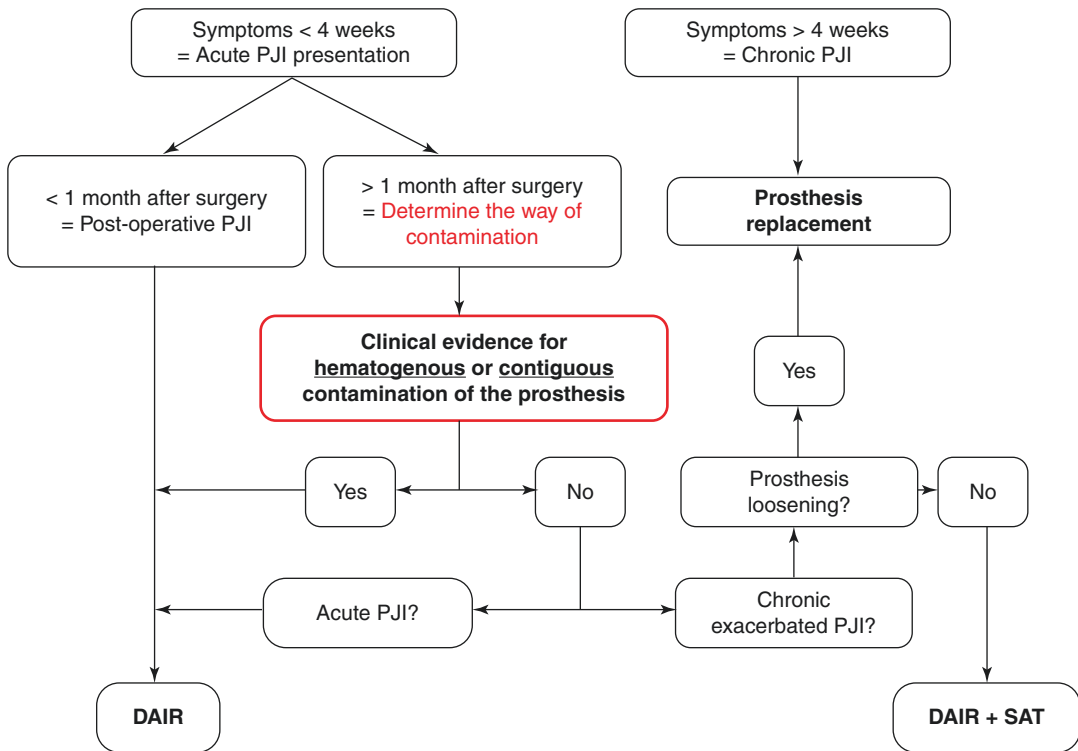


Fig. 2.2 Pathophysiology-based CRIOAc Lyon’s PJI treatment algorithm: The clinical management of patients with acute post-operative PJI, or with chronic PJI with prosthesis loosening is clear: A “debridement antibiotics and implant retention” (DAIR) procedure or a prosthesis exchange has to be, respectively, done. The clinical inves-

tigation, by determining the mechanism of prosthesis inoculation, is essential in patients with acute symptoms but occurring >1 month after the implantation. Depending on the clinical presentation, the appropriate medicosurgical could be a DAIR, a prosthesis exchange, or a DAIR followed by suppressive antimicrobial therapy (SAT)

midis, that is a commensal bacterial of human skin. Coagulase-negative staphylococci are classically considered as low-virulent pathogens with insidious clinical presentations and pain that is the main if not the only one symptom in chronic PJI. But they can also be responsible, and especially *S. lugdunensis*, of early-onset PJI with acute clinical picture. *S. lugdunensis* is particular coagulase-negative staphylococci that acts in many points with *S. aureus* and can also lead to invasive infections. Main differences are observed concerning the antimicrobial resistance profile. Indeed, a methicillin resistance rate around 50–60% is described for coagulase-negative staphylococci, but *S. lugdunensis* is reported to be almost all the time methicillin susceptible [34, 40, 44, 52, 53]. Moreover, 50–75% of the *S. lugdunensis* strains do even not produce any

β -lactamase, making it in those cases, susceptible to penicillin G [53, 54].

Other gram-positive pathogens: streptococci and enterococci. Those two species together represent about 10–20% of PJI, regardless of the time or way of infection. Streptococci are more represented in acute PJI, and more frequently in polymicrobial infections if occurring in a post-operative or early timing. They are in contrary more responsible for monomicrobial infection when involved in acute hematogenous PJI. Streptococci represent a wide range of pathogens with multiple human habitats: oral streptococci, digestive streptococci, or cutaneous streptococci, with here again the need to look for the way in (neoplasia, dental abscess, post-digestive procedures, etc). Enterococci are considered as less virulent agents, with a digestive

location, and can be involved in acute, or chronic presentation, but are considered to be difficult to treat [34, 35, 40].

Gram-negative rods: This group involved various species, with very different virulent characteristics. By order of frequency, *E. coli* and *P. aeruginosa* are the main encountered pathogens. They are mainly involved in acute PJI, whether in post-operative/early-onset or in late-onset by a hematogenous way [34, 40, 41, 55]. Enterobacteriaceae are usually not associated with chronic PJI, probably due to a less capacity, in comparison with staphylococci, to survive and set up mechanisms of persistence in vivo. *P. aeruginosa* must be separately classified. It is a non-fermenting pathogen, not very frequently involved in PJI but with persistence and adhesive factors, who makes it very difficult to treat. Mainly found in early PJI, it can also sometimes be responsible for chronic PJI, occurring lately [34].

Anaerobes: *C. acnes* is the most common pathogen in this group, even if less involved in TKA infection than in hip or shoulder prosthesis [36, 56]. *C. acnes*, a commensal bacterium of the human skin, is a typically low-virulent pathogens, responsible for indolent and chronic infection, which can become symptomatic years after the inoculation and the bacterial inoculation. Because they have a slow growth which usually needed an extended incubation time or at least a supplemented growth culture media, they are often under-diagnosed and involved in many failure management [56–58]. Other anaerobes could be involved, especially anaerobes from the gastrointestinal tracts, and they are frequently associated with other pathogens, leading to polymicrobial PJI [30, 34, 42, 51].

Polymicrobial PJI: It occurs in 15% of all the PJI. They are more frequently found in post-operative and early PJI, especially in knee PJI where difficulties of coverage with skin necrosis occurred after surgery. Some authors reported that multiple pathogens are more frequent in hip PJI in comparison with the knee PJI, probably as the hip location, in comparison with the knee, is closer to the perineum [30, 34, 42, 51]. Another reported risk factor for polymicrobial infection is

obesity, with a polymicrobial infection rate reaching 60% in this specific population [59].

Negative cultures: Sometimes, besides history, clinical presentation, and per-operative findings which make the physician convinced of the PJI, cultures will remain sterile. This is often the case when a prior antimicrobial therapy is used before surgery. This can also be due to very fastidious growth bacteria. That reminds us the importance of the free antimicrobial therapy period before surgery, classically 15 days for patients with chronic PJI, and the need of multiple microbiologic samples with extended cultures (14 days). In those cases, pathology and molecular microbiology could be very helpful [60, 61].

2.4 The Pathophysiology of Chronic PJI

The pathogenesis of PJI involves interactions between the bacteria, the implant, and the host's immune system [62]. Very few numbers of microbes are needed to infect the prosthesis. Such organisms firstly adhere to the prosthesis surface at the bone–implant interface (stem) and/or into the joint cavity. In the latter, microbes frequently replicate themselves as planktonic bacteria that are bacteria in “optimal” environmental conditions to growth (i.e. with a lot of nutrients), leading to recruitment of polymorphonuclear cells (PMNs), and clinical signs of septic arthritis (Fig. 2.3). PMNs are major actors of inflammation that try to control the bacterial multiplication and could result in the formation of pus that is composed by bacterial and PMNs remnants. At the surface of the implant, most of the bacteria have the ability to modify their phenotype and to develop biofilm, after adhering to the surface. Once the biofilm is made, it is inseparable from the implant surface and tolerant to the immune system. It is indeed quite impossible for PMNs to eradicate the biofilm, and other components of the immune system cannot penetrate the biofilm that mainly contain dormant bacterial cells, with low replication process. Different types of biofilm exist, which form themselves at

different speeds, depending on the pathogen involved in the PJI. Bacteria can persist for decades in biofilm, and the interaction between the surface of the biofilm and the host cells could lead to prosthesis loosening, by persistent local activation of immune cells. Intracellular penetration and survival are another mechanism of persistence that particular pathogen could combine with biofilm formation.

2.4.1 Biofilm in TKA Infection

PJI is often described as typical biofilm-associated infections, especially the chronic and persistent ones. Biofilm is “a protected mode of growth that allows survival in a hostile environment” as defined by Bill Costerton, a pioneer in biofilm research [63]. Biofilm is defined as a bacterial community which is metabolically heterogeneous and embedded in a self-produced extracellular matrix, a kind of glue that defini-

tively attached the bacterial community to the prosthesis. To note, all bacteria incriminated in PJI can virtually form biofilm. However, the ability to form biofilm in vivo, in patients with an arthroplasty, can vary from a strain to another in the same species. Biofilm formation is classically described as a succession of several stages that are mostly conserved in all the bacterial species: (a) attachment, (b) accumulation/maturation, and (c) detachment/dispersal (Fig. 2.3) [64, 65].

2.4.2 Attachment

First, free bacteria (called planktonic bacteria, with usual multiplication process) attach to abiotic (metallic or polyethylene components of the prosthesis) or biotic (soft tissues, bone, or prosthesis surfaces covered by host proteins), into the joint or at the bone/implant interface. Bacteria use their adhesin to adhere to the surfaces. In *S. aureus*, initial attachment is made through pro-

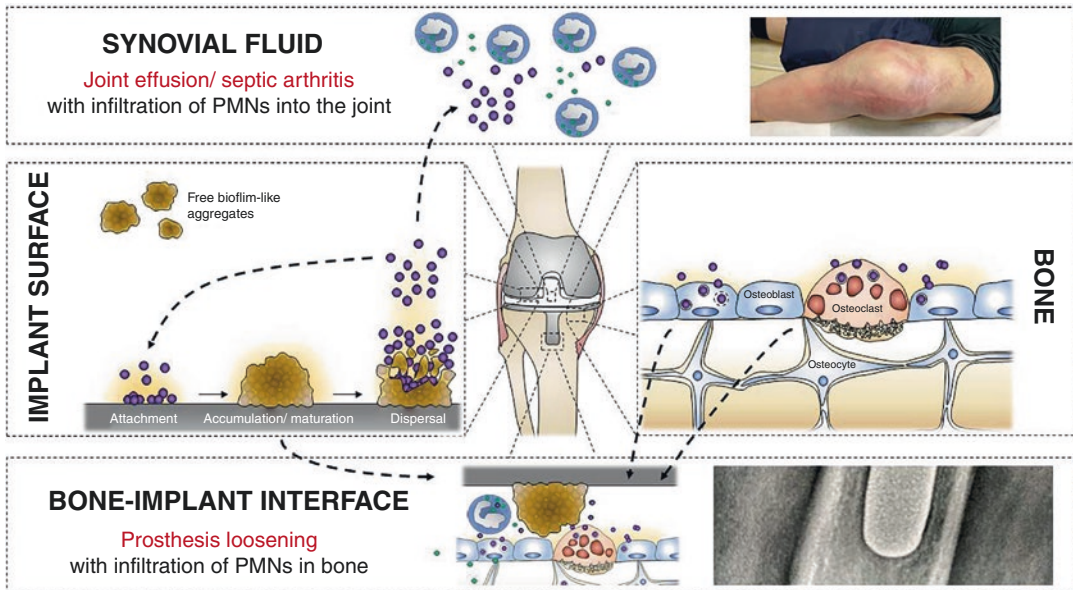


Fig. 2.3 Pathophysiology of chronic prosthetic-knee infection with set up of antimicrobial persistence mechanisms and host response: At the implant surface, the bacteria firstly adhere and then produce a biofilm that can release planktonic bacteria, especially in the synovial fluid, leading to a joint effusion with infiltration of polymorphonuclear cells (PMNs) into the joint synovial fluid. The sec-

ond mechanism of persistence, mainly described for *S. aureus*, concerns the intracellular persistence in bone cells such as osteoblast and osteoclasts, with activation of osteoclasts and induction of osteolysis. Finally, both mechanisms of persistence (biofilm and intracellular persistence) lead to inflammation and infiltration of PMNs at the bone-implant interface, leading to prosthesis loosening

teins belonging to the group of microbial surface components recognizing adhesive matrix molecules (MSCRAMMs). These adhesins can attach to various host matrix components such as collagen, fibronectin, or fibrinogen that quickly coat the surface of the prosthesis [65]. Similar mechanisms, involving adhesins that can bind to host matrix components, were described in *S. epidermidis* and *S. lugdunensis* [66, 67]. In *S. epidermidis*, the giant extracellular matrix-binding protein (Embp) mediates attachment to fibronectin [68]. Concerning primary attachment to abiotic surfaces (nude surface of the prosthesis components), it may be due to electrostatic or hydrophobic interactions with the help of autolysin (AtlA in *S. aureus*, AtlE in *S. epidermidis*, and AtlL in *S. lugdunensis*) that can induce modifications in bacterial surface hydrophobicity [65, 67, 69]. In *P. aeruginosa*, the flagellum, type IV pili and surface adhesion are required for biofilm attachment [70].

2.4.3 Accumulation/Maturation

The second stage called “accumulation/maturation” is characterized by the formation of intercellular bindings and production of the extracellular biofilm matrix. This stage leads to the development of the typical biofilm architecture. The principle components of the biofilm matrix are polysaccharides, proteins, lipids, and extracellular DNA. In staphylococci, the polysaccharide intercellular adhesin (PIA) coded by the operon *icaADBC* was the first described molecule of the biofilm matrix. It favors cell aggregation in *S. aureus* and *S. epidermidis* [71, 72]. Its importance in biofilm accumulation is species and strain dependent. It was notably reported for *S. aureus* that biofilm formation by methicillin-susceptible *S. aureus* (MSSA) are more dependent to PIA than the one by methicillin-resistant *S. aureus* (MRSA) [73]. Moreover, Frank and Patel observed that PIA is not a major component of extracellular matrix in biofilms formed by *S. lugdunensis* [74]. Proteins and extracellular DNA (eDNA) are also components of extracellular matrix of staphylococcal biofilms [65, 67, 75].

These two types of molecules come from the bacteria themselves. Indeed, lysis of bacteria allows the release of eDNA and cytoplasmic proteins [76, 77]. Proteins and eDNA can interact together to form a kind of network that keeps biofilm tight [78]. Interaction between PIA and eDNA has also been reported in *S. aureus* [79]. To note, proteins that are involved in primary attachment are also involved in biofilm accumulation, such as fibronectin-binding proteins (FnBP) in *S. aureus* or Embp in *S. epidermidis* [68, 80]. In *P. aeruginosa*, the matrix is composed of at least three types of exopolysaccharides (PIs, Pel, and alginate) and contains eDNA [81].

2.4.4 Detachment/Dispersal

The last stage is the dispersal of the biofilm, allowing the propagation of the infection to other surfaces or tissues. This mechanism could be involved in “late acute” PJI, in patients without evidence of hematogenous spreading of a distant site of infection. In *S. aureus*, dispersal is controlled by quorum sensing through the activity of the accessory gene regulator *agr* [65]. It promotes the degradation of the protein inside the biofilms through the production of proteases such as metalloprotease aureolysin and serine proteases splABCDEF [82]. Phenol soluble modulins (PSM), also controlled by *agr*, can play a role in biofilm dispersal. These toxins, notably known for their cytotoxic effects [83, 84], could disrupt biofilm structure thanks to their surfactant properties [85]. Similar properties of PSM in biofilm dispersal have been reported for *S. epidermidis* [86]. A role in biofilm stabilization was also suggested for PSMs through the formation of amyloid fibers but this possibility is debated [87–90].

To note, it is important to keep in mind that most of the current data about biofilm were obtained from in vitro models that are not representative of in vivo biofilms found in chronic infections. Indeed, in vivo biofilms are smaller, they are not shaped in mushroom-like structures, incorporate host components, and are continuously challenged by the host immune system [91].

Two major properties, tolerance to antibiotics and the hijacking of host immunity, confer to biofilm-associated infection its “difficult-to-treat” character.

The tolerance to antibiotics corresponds to a transient loss of susceptibility to antibiotics that can be restored when bacteria in biofilms switch back to a planktonic phenotype. In opposite to resistance, there is no impact on the minimal inhibitory concentration (MIC) [92]. This property is mostly due to the metabolic status of bacteria inside the biofilm. Indeed, they are deprived of oxygen and nutrient and lower their metabolic activity to survive in these conditions. In consequence, bacteria in biofilm are less susceptible to antibiotics as antibiotics are mostly efficient on metabolically active bacteria [93]. To eradicate biofilms, antibiotic concentrations have to be 10 to 1000-fold superior to the MIC determined for planktonic bacteria. This tolerance property is shared by most of the bacterial species. Regarding antibiotics that can be used for treating biofilm-related PJIs, rifampicin is considered as the “anti-biofilm” antibiotics [94].

Another important property of biofilm is its ability to interact with the immune cells and to hijack the normal immune response. In the course of an infection, the first responders are the PMNs. Even if PMNs can attack and phagocyte biofilms by staphylococci, meaning that biofilms are not protected from immune cells, it is reduced compared to what is observed with planktonic bacteria [95–97]. Moreover, PMNs can favor the bone resorption through the release of IL-8 and the activation of osteoclasts [98]. PMNs infiltrations in the synovial fluid or in bone tissue at the interface of the prosthesis are major pathology criteria for a PJI. Biofilm can also interact with macrophages and hijacking their inflammatory properties. Instead of eliciting pro-inflammatory properties with high phagocytosis and the production of pro-inflammatory cytokines and chemokines (defined as M1 phenotype), macrophages interacting with biofilm display a low capacity of phagocytosis and gene patterns relative to anti-inflammatory properties, revealing a polarization to M2 phenotype [99, 100]. Macrophage infiltration is how-

ever non-specific, as mechanical prosthesis loosening could also be associated with such infiltration, as these cells participated in the phagocytic process of the released microparticles [62]. Biofilm-related PJIs are also associated with the presence of myeloid-derived suppressor cells (MDSC). These immature cells are involved in the regulation of inflammation and the immunosuppression. They inhibit the T cell proliferation and prevent the pro-inflammatory activity of macrophages, especially through the release of IL-10 [101, 102]. MDSC infiltrates were also observed in periprosthetic tissues from PJI patients whereas they were not observed in tissues from patients with aseptic loosening [103]. The presence and role of T lymphocytes in biofilm-associated PJI are still unclear. The few studies on this subject reported a reduced T cell proliferation (in accordance with the high presence of MDSCs) with a pro-inflammatory Th1/Th17 profile [104]. Finally, infiltration of plasma cells could participate in the immune response in patients with PJI. Plasma cells are the final step in maturation of B cell line, they are part of adaptive immunity, and classically, it has been described that acute inflammation can evolve into prolonged subacute and chronic inflammation when the initial cause of inflammation persists in tissues [105–107]. Few data are available about the description of plasma cell infiltration in patients with PJI, but such infiltration has to be investigated as a potential pathology marker of chronic PJI [108, 109].

Regarding the localization of biofilm in knee prosthesis environment, it can first attach and develop onto the metallic, ceramic, or polyethylene components of the biofilm. Studying the components from total hip prosthesis, Lass et al. reported that a higher bacterial load was present on the polyethylene liner than on the metal components [104]. Similar results were observed for biofilm by *S. aureus*, *S. epidermidis*, *E. coli*, *K. pneumoniae*, and *P. aeruginosa* [110]. Biofilm can also form biofilm-like aggregates in synovial fluid for *S. aureus* and *S. epidermidis* [111, 112]. The formation of these aggregates is mostly due to the presence of fibrin in synovial fluid. However, surprisingly, in vitro experiments

showed that synovial fluid reduced the ability of biofilm-like aggregates to adhere to various surfaces [113]. Abscesses are sometimes described as a type of biofilm [114]. However, this definition is controversial. Indeed, Cheng et al. strictly affirmed that abscess formation “should not be mistaken for biofilm growth” [115]. Finally, invasion of osteocyte-lacuno canalicular network by bacteria was also proposed as a new type of biofilm [114, 116]. It was reported that *S. aureus* can invade the lacuna left empty by osteocytes after their death [117]. However, even if this new mechanism must be taken into account to fully understand PJI pathogenesis, formation of extracellular matrix was not investigated in this case, questioning the “biofilm” appellation.

2.4.5 Bacterial Interactions with Bone Cells

Despite no existing data specifically regarding knee prosthesis infection, the ability of bacteria to invade and persist within bone cells has been proposed as another mechanism of host immune system subversion, constituting a reservoir that might lead to infection chronicity and relapse (Fig. 2.3) [118, 119]. The interaction of *S. aureus* with osteoblast—the non-professional phagocytes specialized in bone apposition—has been extensively described in vitro, mostly in gentamicin- or lysostaphin-protection assays. The internalization of *S. aureus* within osteoblasts is mainly driven by the interaction of staphylococcal fibronectin-binding proteins (FnBP) with host fibronectin that acts as a bridge with the cellular $\alpha 5 \beta 1$ integrin to prompted bacterial endocytosis by an active cellular process akin to phagocytosis [120–122]. After cell invasion, *S. aureus* can persist within vacuoles or escape to the cytosol, through complex mechanisms (Fig. 2.3). First, staphylococcal membrane-damaging factors and toxins, including phenol soluble modulins, are involved in vesicular escape and cytotoxicity [83, 123]. Second, *S. aureus* has been shown to subvert the autophagy process, a highly conserved eukaryote cellular process allowing cellular component destruction and recycling. This mechanism is also

part of the innate immune response, leading to the formation of a double-membrane compartment that encapsulates intracellular bacteria (phagosome) that merges with lysosome to create a phagolysosome, a digestive hybrid organelle with an acidic environment base of intracellular bacteria eradication [124, 125]. *S. aureus* has been shown to activate autophagosome formation while inhibiting the autophagic flux and especially the fusion of autophagosomes with lysosomes, then promoting its intracellular persistence [126, 127]. Finally, the intracellular conversion to a slow-growing phenotype called small-colony variants (SCVs) facilitates intracellular survival and resistance to antimicrobial therapy [128, 129]. The impact of *S. aureus* persistence within osteoblast in vivo has not been well established and is still controversial. Some electronic microscopy observations revealed the presence of *S. aureus* within bone cells during the course of chronic osteomyelitis [130]. Additionally, some cohort studies showed a correlation between infection chronicity and the ability of *S. aureus* to invade osteoblasts [83, 131].

The ability of other bacteria to invade and persist within bone cells has been less investigated. In a study screening 15 different coagulase-negative staphylococci species, only *S. pseudintermedius* appeared to be able to invade osteoblastic cells in a comparable way to *S. aureus*, but is not a common etiological agent of BJI [132]. *S. delphini*, a recently described invasive staphylococci species, also seem to be able to invade osteoblasts, following a FnBP-like protein pathway [133]. This mechanism seems less important for *S. epidermidis*, the most commonly involved coagulase-negative staphylococci in PJI, even if this point is still controversial [134–136]. Streptococcal invasion of eukaryotic host cells has been assessed for *S. pyogenes* [137]. Turning especially to bone cells, interaction of oral streptococci with osteoblasts has been studied, for a better understanding of osteo-dental pathology: *S. gordonii* is internalized in osteoblast via a process similar to *S. aureus*, triggering an inflammatory response that promotes bone resorption, but its intracellular survival is shorter than *S. aureus* [138]. Invasion of human cell has been described

as a major feature for Enterobacteriaceae pathogeny in other clinical settings, including internalization in intestinal epithelial cells to cross the intestinal barrier during *Salmonella* infections [139–141]. However, few authors studied the interaction of Enterobacteriaceae with bone cells. Using clinical isolates responsible for PJI, Crémet et al. showed that *E. coli* was unable to invade osteoblastic cells in gentamicin protection assay and elicited a high cytotoxicity [142]. Regarding non-fermenting gram-negative bacteria, *P. aeruginosa* invades various epithelial cells (as proven by electronic microscopy and gentamicin protection assays) via complex mechanisms, probably dependent of the type of infected cells [143–146]. However, the invasive ability of *P. aeruginosa* within osteoblasts remains to be confirmed [142]. Among anaerobic bacteria, *C. acnes* is probably the most frequent and better-characterized pathogen in bone and joint infection—especially device-associated chronic infections—and has been shown to be able to invade osteoblasts [147]. Finally, corynebacteria, mostly implicated in knee infections after major trauma, displayed strain-dependent capacities of osteoblastic internalization, via a fibronectin-dependent pathway similar to *S. aureus*, which seems implicated in chronicity [148].

The interaction of *S. aureus* with osteoclasts has also been studied, leading to bone resorption and consequently potentially involved in prosthesis loosening (Fig. 2.3). In particular, an in vitro model of osteoclasts infection at different maturation stages provided insight into this complex mechanism [149]. The infection of osteoclast precursors inhibits osteoclastogenesis but leads to pro-inflammatory cytokine secretion that enhances bone resorption by mature osteoclasts. Conversely, infection of mature osteoclasts directly increases their bone resorption ability. In addition, the infection of surrounding osteoblasts can lead to a direct decreased bone formation, and a secretion of RANK-L that stimulates osteoclast activities [118]. With the exception of *C. acnes*, for which the inhibitory effect on osteoclastogenesis has been suggested in one study [147], there is no data regarding the interactions of other bacterial species with osteoclasts.

Unlike the “antibiofilm” activity of antimicrobials, their ability of eradicating the intracellular bacterial reservoir is currently not taken into account in the choice of treatment strategies for PJI. However, intraosteoblastic persisting *S. aureus* demonstrate heterogeneous antimicrobial susceptibility [150]. This intracellular activity is difficult to predict and relies on: (a) intracellular penetrations of the molecules and their distribution in the subcellular location of *S. aureus*; (b) intracellular bacterial wall modifications [151] and reduced metabolism [152]; and (c) drug inactivation by the acidic pH of intracellular organelles [153, 154]. Consequently, some antimicrobials with a low cellular accumulation can have a surprisingly high activity due to local chemical conditions, such as β -lactams of which activity is even restored intracellularly against methicillin-resistant staphylococci [153]. In final, the most active molecules against intraosteoblastic *S. aureus* remain clindamycin, fluoroquinolones, and rifamycins [150, 155]. The intraosteoblastic activity of antimicrobials against other bacterial species implicated in PJI is unknown.

2.5 Conclusion

Understanding the etiology and pathogenesis of knee replacement infections is crucial to prevent, diagnose, and treat such catastrophic complication. Physicians have to be aware about the different ways that has a bacteria to contaminate the prosthesis, to precise the date of inoculation in each individual patient. Moreover, depending on different factors, the bacterial epidemiology, and the resistance profile of the bacteria involved are heterogenous, leading to discuss the empirical antimicrobial also individually. Finally, the different involved bacteria have various capacities to persist in vivo. The two main mechanisms of persistence are the biofilm and the intracellular persistence, especially for *S. aureus* in osteoblast and osteoclast cells. If these mechanisms have been set up in a patient with PJI, eradication is considered as impossible, and the patient has to be managed by prosthesis exchange. However, a

conservative approach is sometimes performed, especially in patients with revision prosthesis without loosening, but these patients have to receive a suppressive antimicrobial therapy to keep the bacteria asleep and prevent biofilm growing, bacterial multiplication, and prosthesis loosening. The pathophysiology of the infection directly affects the clinical management of PJI, and bacterial mechanisms of persistence have to be targeted by innovative therapeutic approaches.

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Part II

**Biomaterials in Artificial Joint
Replacements**

The Role of the Surface on Bacteria-Implant Interactions

3

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3.1 Introduction

TKA prostheses postoperative infection (periprosthetic be infection, PJI) is the most terrible complications, the most serious in TKA can have disastrous consequences [1–4]. As the population ages and the demand for TKA continues to rise, the incidence of PJI is also increasing. The main reason for treatment failure is the formation of bacterial biofilm on the surface of the implant and the adhesion of bacterial biofilm in the surrounding tissues and bones. Biofilm is a barrier to protect bacterial cells and has many unique properties leading to antibiotic resistance [4]. The first step in clinical PJI treatment is to find the pathogenic microorganisms. However, in clinical microbial culture, the result of culture test is negative because the bacteria are wrapped in biofilm, which makes microbial diagnosis difficult [5]. Moreover, because biofilms can protect pathogenic bacteria from antibiotics and host defense, PJI after total knee replacement (TKA) is difficult to treat. This chapter will elaborate the formation mechanism of the implant biofilm and the mechanism of antibiotic resistance in the biofilm as well as the detection and treatment of the biofilm.

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3.2 Mechanism of Implant Biofilm Formation

Biofilm studies of microorganisms are getting more and more attention. Biofilms are common in natural industry and clinical environment, but it is difficult to eradicate them. The main difficulty of PJI treatment after KTA is the biofilm formation of pathogenic bacteria. Bacterial cells in the surface of the orthopaedic material have very high affinity, the most commonly used material in modern orthopedic surgery including titanium (and its alloys), cobalt chromium stainless steel, and various polymers, including ultra-high molecular weight polyethylene (UHMWPE) silicone polyether ketone all kinds of ceramic and hydroxyapatite and polymethyl methacrylate (PMMA) cement are vulnerable to biofilm formation of bacteria to colonize [6]. Bacterial biofilms have been reported to form after placement for 16 h [7]. The National Institutes of Health estimates that about 65 percent of infections are caused by bacterial biofilms, biofilm pathogens include gram-positive bacteria (*Staphylococcus aureus*, *Staphylococcus epidermococcus*, *Streptococcus enterococcus*) or gram-negative bacteria (*E. coli*, *K. pneumoniae*, *Pseudomonas aeruginosa*) [8]. *Propionibacterium acneum* is a gram-positive facultative anaerobe and a conditioned pathogen capable of forming biofilms [9]. The most common biofilm bacteria in PJI in TKA are *Staphylococcus aureus*, especially

Staphylococcus aureus and *Staphylococcus epidermidis*, which account for nearly 50–60% of culture infections [10]. Zimmerli et al. [11] reported that it only takes 100 colony forming units to cause PJI. Once the bacteria adhesion together, they gathered in the community and produce extracellular polymer matrix (extra polysaccharide matrix, EPS), EPS by extracellular polysaccharide protein cell DNA (extracellular DNA, eDNA) lipid composition, called biofilms [12, 13]. Biofilm is adhered on the surface of the bacterial cell, it is composed of 10% of the cells and 90% of EPS, EPS provides the mechanical stability of the biofilm, mediated and surface adhesion, and form a tight three-dimensional polymer network, make the mutual connection and temporarily fixed biofilm cells, has a defense mechanism that can effectively protect bacteria against their antibiotic treatment [14, 15]. According to Gilbert et al. [16], biofilms have the ability to protect bacteria in the membrane against antibiotic treatment. Compared with planktonic bacteria, mature biofilms can tolerate antibiotics with a concentration 100–1000 times higher. This makes them harder to detect

and eradicate with conventional therapies, increasing antibiotic resistance [4]. Arciola et al. [17] summarized the process of biofilm formation in four steps: (1) free floating (plankton-cell) adhesion; (2) colony formation; (3) mature; (4) separation. The first step in the formation of a biofilm is for free-floating bacteria to attach to the surface of an object. *Staphylococcus epidermidis* (Fig. 3.1) and *Staphylococcus aureus* specifically express proteins at this stage of growth that strongly interact with the host extracellular matrix (ECM). These proteins are thought to be key to the bacteria's attachment to foreign bodies, as the ECM wraps them up when they enter the body. Surgery can lead to tissue destruction and trauma caused by partial produce extracellular matrix (ECM) protein host (such as the fibrous connections to the implant surface protein and collagen), the mechanism of extracellular protein deposition enhances bacterial colonization ability, let host bacteria on the surface of protein and protein matrix combination, more easily in the kind of anchor implant surface [13]. In addition, surgical trauma also leads to tissue ischemia and local immunosuppression, which further pro-

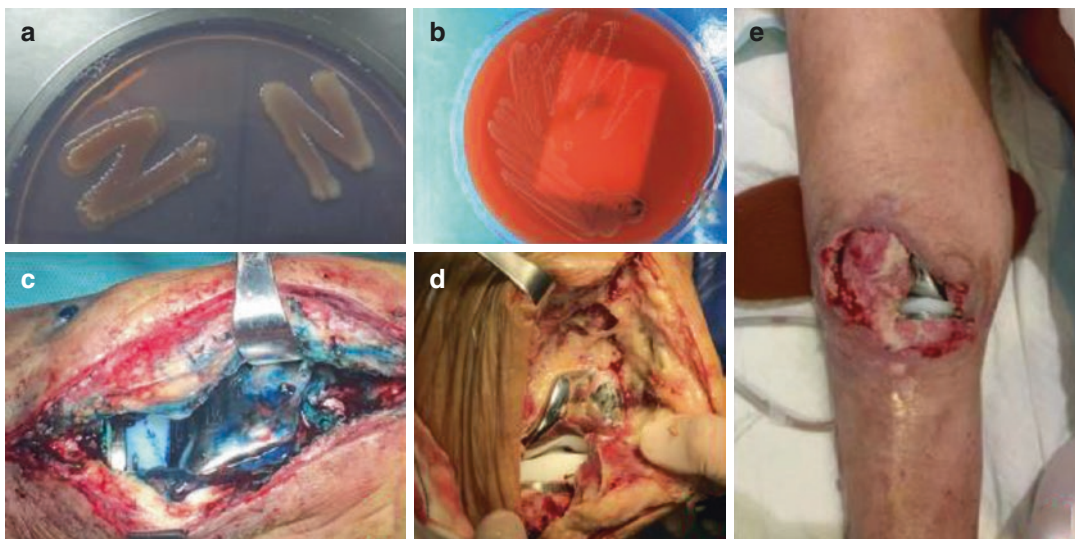


Fig. 3.1 (a) Observation of polysaccharide secretion by *Staphylococcus epidermidis* in PJI Congo red medium after TKA operation (the picture is a gift from doctor Chen peng of Guizhou Provincial People's Hospital), (b) culture observation of *Streptococcus agalactiae* of PJI

after TKA operation, (c) after TKA, chronic PJI infection was demonstrated by injection of sterile meilan through the sinus tract, (d) knee cavity abscess of PJI after TKA operation, (e) PJI prosthesis was exposed after TKA

notes the colonization of bacterial cells [18]. Extracellular DNA (eDNA) also accumulates during the attachment phase through bacterial autolysis. Although the role of extracellular DNA (eDNA) in biofilm formation is not fully understood, it is thought to contribute to the stability of biofilms and may be an intercellular adhesin. This is followed by irreversible attachment to the surface and loss of movement. The new plankton-bacteria can interact with the bacteria attached to the surface to form bacterial microcolonies [19]. In addition to population expansion, bacteria also produce extracellular compounds called self-inducers during the entire attachment growth phase [20, 21]. These automatic inducers act as signals between and within bacteria, specifically to convey the local density of bacterial populations [19]. With the aggregation of bacteria, the production of extracellular polymer matrix (EPS) was upregulated and embedded into the EPS matrix. In this screened environment, the biofilms begin to mature and thicken as bacterial cells multiply to form multilayer colonies [22]. In the mature stage of biofilm, bacteria are constantly multiplying. When the critical threshold is reached, the bacteria in the biofilm will undergo densi-related changes. This process is described as quorum sensing. Quorum sensing mechanism is the basic chemical signal means for bacterial population to communicate, coordinate, and cooperate [19]. Quorum sensing is a complex internal network that connects colonies in a biofilm microenvironment. Bacterial cells use quorum sensing to communicate with one another through cellular signals, exchanging nutrients and promoting resistance to antibiotics and avoiding the immune system's toxic genes [23]. The different stages of bacterial biofilm formation are closely related to quorum sensing. Besides the formation of biofilm, quorum sensing also regulates the production of bioluminescent sporogenic antibiotics and the secretion of virulence factors [24]. Biofilm separation requires a protease to separate the bacterial cells, helping the plankton-bacteria to release from the biofilm, spread again and cause further infection [4]. The maturation time of the biofilm of different bacteria was not consistent, and the

time of *Staphylococcus epidermidis* was 24 h [25, 26],

Staphylococcus aureus for 24 h [27, 28], *Pseudomonas aeruginosa* for 72 h [29]. At present, PJI infection can be divided into acute phase and chronic phase according to the maturity of biofilms. In acute PJI, microorganisms do not form biofilms, while in chronic PJI, microorganisms have formed mature biofilms. Therefore, the treatment methods for different infections are different [30].

3.3 Mechanisms of Antibiotic Resistance in Biofilms

Under the protection of the biofilm, the bacteria developed drug resistance by increasing the minimum antibiotic inhibition concentration required for infection control [16]. Lazăr [31] pointed out that resistance to antibiotics of biological membrane has four main mechanisms: (1) due to the existence of the biofilm matrix, antibiotics cannot infiltrate into the depths of the mature biofilm; (2) accumulation of high levels of antibiotic degrading enzymes; (3) in the depths of the biofilm, bacterial cells are experiencing nutrient restriction and entering a state of slow growth or starvation; slow-growing or non-growing bacterial cells are not sensitive to antimicrobial agents, which can be amplified by phenotypic variation or the presence of permanent cells. Moreover, biofilm bacteria can turn on stress response genes and convert to a more drug-resistant phenotype when exposed to environmental stress. (4) Genetic changes may be selected by different stress conditions, such as mutations and gene transfer may occur in the biofilm. Akanda et al. [4] proposed three main factors of antibiotic resistance mechanism in biofilms: (1) physical barrier against host defense and antibiotic penetration; (2) reduce the metabolic activity of bacterial cells; (3) quorum sensing promotes the communication of antibiotic resistance genes. In order for antibiotics to have a bactericidal effect on the bacteria in the biofilm in the treatment of PJI, the antibiotics used must be able to penetrate the EPS matrix. Singh, etc. [32] discussed the various antibiotics

on *Staphylococcus aureus* and epidermis *Staphylococcus aureus* biological membrane permeability, the results confirmed that the benzazole Westwood cefotaxime (beta lactam type) and vancomycin (sugar peptide) on *Staphylococcus aureus* and epidermis *Staphylococcus aureus* biofilm osmosis are decreased obviously, and aminoglycoside drug amikacin ciprofloxacin and fluoroquinolone drugs of osmosis are not affected.

Biofilm the physical structure of the antibiotic produced a concentration gradient, internal bacterial biofilm to accept less than planktonic bacteria antibiotic concentration, which resulted in increased the risk of resistance, but the bacteria in the biofilm mutations faster, this phenomenon is associated with increased oxidative stress in biofilm environment, this kind of oxidative stress are both endogenous and caused by the antibiotics [33]. Adaptive stress response in biofilms is an active process mediated by quorum sensing, which promotes the expression of genes and signaling molecules that contribute to antibiotic resistance, and the production of enzymes and effervescent pumps that put cells into a dormant state or inactivate antibiotics, depending on the strain [34]. Donlan et al. [35] proposed another hypothesis that antibiotic resistance was related to the weakened growth state of microorganisms in the biofilm and found that the deep biofilm microorganisms with nutrient deficiency showed lower metabolic activity and slower growth rate. Studies have shown a correlation between slow-growing microbes and reduced antibiotic sensitivity to the microbes. Antibiotics depend on interfering with cell metabolism to regulate their bactericidal action, so they need to actively proliferate cells to kill them [36]. In biofilms, groups of cells that remain after antibiotic treatment are called permanent cells and these permanent cells can regenerate after antibiotic treatment, making it very difficult to get rid of all the biofilms that are embedded in the infection [31, 37]. Permanent cells are highly antibiotic resistant, metabolically dormant non-dividing bacteria that acquire this phenotype in the presence of antibiotics; once the antibiotics are removed, they resume metabolic activity and because drug-resistant cells are a

source of recurrent chronic infections, they play an important role in the challenge of treating biofilms with antibiotics [36]. Environments such as pH metabolites and oxygen levels in the biofilm can alter the efficacy of antibiotics [38]. Ernest et al. [39] found that biofilms could survive slight changes in pH and could only partially eradicate the biofilms of *Staphylococcus aureus* in common topical adjuvant therapies (such as povidone-iodohypochlorite or hydrogen peroxide).

3.4 Microbiology of Periprosthetic Infection (PJI)

Infection control after total knee arthroplasty (TKA) is difficult, mainly because biofilm formation effectively protects pathogenic bacteria from antibiotics and host defense. For PJI biofilms, there is still a lack of adequate prevention, diagnosis, and treatment. There are new chemical and mechanical approaches to the treatment of biofilm infection, which will be important for the eradication of orthopedic infections in the future. Biofilms have been studied using topical and systemic antibiotics. Strategies for the prevention and treatment of biofilms include the use of surface coatings (including surface reserved antibiotic and metal oxide nanoparticle coatings) and the destruction of established biofilms by mechanical or pharmacological means. It has been reported that the most common pathogens of PJI in the USA are *Staphylococcus aureus* and *Staphylococcus epidermidis*, while the most common pathogens in Europe are thrombin negative *Staphylococcus aureus* and *Staphylococcus enterococcus*, followed by *Staphylococcus aureus* and *Staphylococcus enterococcus* [40]. Benito et al. [41] collected 2288 cases (hip and knee joint) for microbiological diagnosis in 15 years, and the result was that the gram-positive bacteria accounted for 78% (mainly *Staphylococcus*), gram-negative bacteria accounted for 28%, and anaerobe bacteria accounted for 7%. Methicillin-resistant *Staphylococcus aureus* (MRSA) is a tricky problem, leading to more complications in postoperative functional recovery and reoperation rates than other pathogens [42]. *M. tuberculosis* is

also a pathogen leading to artificial joint infection. In 2013, Kim et al. [43] reported a systematic evaluation of mycobacterium tuberculosis infection. Jakobs et al. [44] found that candida accounted for about 80% of fungal PJIs (36 out of 45 cases). PJI caused by mycoplasma is extremely rare in clinical practice. Qiu et al. [45] reported 1 case of mycoplasma infection after TKA.

3.5 Methods for the Detection of Biofilms

Recent studies have focused on the detection of the formation and destruction of biofilms. The improvement of the detection method of biofilm bacteria is usually based on the growth of culture. The presence of biofilm leads to negative culture of bacteria, which increases the difficulty of diagnosis. Parvizi et al. [46] proposed the latest diagnostic criteria for infection around joint prosthesis in 2018: the main criteria and diagnostic method for infection around joint prosthesis are the presence of positive culture or sinus tract for two or more times. Serum c-reactive protein (>10 mg/L), D-dimer (>860 living g/L), and serum erythrocyte sedimentation rate (>30 mm/h) were 2, 2, and 1, respectively. In addition, elevated leukocyte count in the fluid (>3109/L), leucocyte esterase (++) , leucocyte percentage (>80%), and syn c-reactive protein (>6.9 mg/L) scored 3, 3, 3, 2, and 1, respectively. Patients with a total score of 6 are considered to be infected, while patients with a score of 2–5 need intraoperative results to confirm the diagnosis. The results of positive histology, purulent, and single positive culture were 3, 3, and 2 points, respectively. Combined with preoperative scores, a total of 6 points were considered infected, 4–5 points were inconclusive, and 3 points were not infected. The diagnostic criteria were 97.7% sensitivity and 99.5% specificity as defined by the society for musculoskeletal infection (79.3%) and the international consensus meeting (86.9%). Despite the introduction of the new diagnostic criteria, there are still limitations. In some cases, patients who do not meet the diagnostic criteria may still be infected.

Therefore, other methods should be used to improve the diagnosis of PJI, especially those related to body recognition. PCR has been used to identify a variety of bacteria in biofilms [47] and has been shown to be more sensitive to tissue culture [48, 49]. Other methods for detecting biofilms include fluorescence in situ hybridization (FISH) and DNA microarrays [50–52]. FISH can identify bacteria in culture negative infection, reduce false positive by improving the recognition of environmental bacterial pollution, detect cross-reactivity with human tissues, and eliminate viable staining dead bacteria [52]. DNA microarrays, which can simultaneously evaluate the DNA of thousands of bacteria, are cheaper, faster, and more productive than PCR [52]. Biofilm imaging technology also includes confocal laser scanning microscopy [53] and scanning electron microscope. Confocal laser scanning microscopy can show live bacteria in biofilms and even culture of negative PJI, while scanning electron microscopy can see the aggregation of microbial cells [52]. However, limitations of these imaging techniques include cost of use and training requirements for obtaining the best images. Sonication refers to the use of ultrasound to degrade cell viruses, etc. It can also be used to detect PJI. It improves the sensitivity and specificity of culture to microbial detection, and even the samples obtained after antibiotic treatment can be used to detect bacteria [52].

3.6 The Treatment of PJI Biofilm After TKA

A number of new treatments are being developed, focusing on ways to improve bacterial clearance and destruction of biofilms, thereby reducing bacterial resistance to antibiotics and immune defenses. New antibiotic research can improve the penetration of bone and joint tissue, which may increase activity against biofilm bacteria. These antibiotics (such as tizolamide phosphate and oliban star) target gram-positive bacteria, including methicillin-resistant *Staphylococcus aureus* (MSSA), methicillin-resistant *Staphylococcus aureus* (MRSA),

vancomycin-resistant *Staphylococcus aureus*, and methicillin-resistant coagulase-negative *Staphylococcus* [54, 55]. In clinical biology studies, it has been found that some drugs used for tumor chemotherapy can effectively combat biofilm activity. For example, cisplatin 5-fluorouracil and mitomycin C have been reported to be able to remove bacteria embedded in the biofilm. These chemotherapeutic drugs may be the subject of research in the emerging field in the future [56]. Currently, two antibiotics with anti-biofilm activity are commonly used: rifampicin (inhibition of transcription) and meropenem (inhibition of cell wall biosynthesis) [57, 58].

Antibiotic bone cement can be used to prevent biofilm formation, which is an evidence-based placement method to prevent biofilm formation [59], but there is growing evidence that a strategy based on the surface properties of the implant (containing some specific metals) may play a role in preventing biofilm formation. The surface properties of the implant affect the ability of the bacteria to adhere to and form biofilms [60]. Basic scientific research suggests that current implant materials, such as vitamin E hybrid ultra-high molecular weight polyethylene (ve-pe) and ceramics, may offer a degree of protection against the formation of biofilms [61–64]. Many researchers are now trying to develop an anti-biofilm coating on the surface of the implant, including changing the surface shape of the implant material using the material's inherent antibacterial properties (such as when coated with silver or copper) and placing antibiotics on the surface of the implant [65]. The surface of the implant can be modified to reduce the risk of PJI, with the aim of achieving a bactericidal coating that does not inhibit bone growth, is biocompatible and durable. For example, silver oxide, titanium oxide, copper oxide, iodine, and other nanoparticles have bactericidal effects on gram-positive and negative-positive bacteria, and silver oxide particles have been shown to inhibit the formation of biofilms on the catheter. Secinti et al. [66] studied in the rabbit model and concluded that nanometer silver ion coated implants were as safe as uncoated titanium screws, and nanometer silver ion

coated implants could prevent the formation and infection of biofilms. Silver-containing hydroxyapatite (Ag-HA) coating can reduce the formation of MRSA biofilm both in vivo and in vitro, which may be an effective method to reduce implant-related infections. Silver-containing hydroxyapatite (Ag-HA) coating can reduce the formation of MRSA biofilm both in vivo and in vitro, which may be an effective method to reduce implant-related infections [67]. Immunotherapy, especially monoclonal antibodies, can be used as an alternative and adjunct to antibiotic therapy, and multiple targets of antibody therapy can be studied in the preclinical stage [68]. Bacteriophages are another potential therapy for biofilm-related bacteria. Bacteriophages are naturally occurring viruses that target and kill bacteria and can destroy the biofilm matrix, potentially affecting metabolically active bacteria and persistent cells because bacteriophages remain active at low temperatures and in low nutrient status [4].

3.7 Conclusions

The main reason for the difficulty in treating PJI after TKA is the formation of biofilm by pathogenic microorganisms. Biofilms prevent antibiotics and host defense, promote bacterial nutrition, allow interactions between bacteria (quorum sensing), and allow the spread of drug resistance. While acute PJI can be identified by clinical and planktonic bacterial cultures, chronic PJI is associated with the production of biofilms by pathogens, significantly reducing the ability to prevent diagnosis and treat infection. Current biofilm treatment strategies include antibiotics and surgery, and other innovative treatment strategies, including new antibiotics with improved permeability, drug Immunotherapy against persistent bacteria, antimicrobial peptide nanoparticles and bacteriophage ultrasound, are also being improved.

Although some of these strategies are still in the early stages of research, the results of future studies may revolutionize the prevention and treatment of PJI.


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In-Vitro and In-Vivo Models for the Study of Prosthetic Joint Infections

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4.1 In-Vitro Models of PJI

In-vitro, from the Latin meaning “within the glass”, refers to the study of processes taking place outside a living organism. This can take many forms and specific to the study of PJI involves assays developed to identify bacterial properties [1–3], biofilm formation [1, 4–9], and the response to antimicrobial agents [5, 10–16]. Moriarty et al. [17] outline several key areas for in-vitro modelling of PJI, including (1) bacterial species to be tested; (2) antimicrobial efficacy/activity; and (3) if the in-vitro study is deemed suitable to proceed to in-vivo testing.

4.1.1 Bacterial Adherence

Gram-positive bacteria demonstrate persistent adherence to prostheses, which has influenced prosthetic design. The adherence of *Staphylococcus epidermidis* strains to orthopaedic grade hydroxyapatite-coated stainless steel screws was examined in-vitro by Arciola et al.

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[18]. Coated prostheses had significantly lower bacterial adherence compared with uncoated metal prostheses. An earlier in-vitro study also concluded that *S. aureus* colonized both metal prostheses and ultra-high molecular weight polyethylene (UHMWPE) more rapidly than *S. epidermidis* [19]. It was also shown that *S. aureus* had an affinity to metal, whereas *S. epidermidis* preferentially colonized UHMWPE. As a potential therapeutic option, vitamin E impregnated UHMWPE is associated with reduced bacterial adherence of *S. aureus* and *S. epidermidis* strains [20], in addition to providing oxidative protection and lower wear rates [21–24].

4.1.2 Biofilm

The ability to form biofilm plays a crucial role in the pathogenesis of PJI, for both gram-positive and gram-negative bacteria [25, 26]. This has major implications for in-vitro studies, which have assessed the structural and protective qualities of the polymeric matrix, polysaccharides, proteins, and extracellular DNA which these bacteria generate and are imbedded within. Bacterial populations may harbour key gene loci implicated in biofilm formation, such as the *ica* gene encoding polysaccharide intracellular adhesion [27]. However, other researchers have shown that *ica* genes are not required [28], demonstrating the complexity and variability of bio-

film formation. Thomson et al. [3] described that *Pseudomonas aeruginosa* generated significantly increased biofilm compared to all *E. coli* strains tested, suggesting that this enhanced activity correlates with more aggressive in-vivo models of *P. aeruginosa* PJI. Investigators agree that biofilm is difficult to study in-vitro due to the protection against antibiotics that biofilm provides, along with bacterial species variation within biofilm and its propensity for prosthetic adherence.

4.1.3 Antimicrobial Efficacy

Adequate antimicrobial efficacy is crucial for controlling PJI, in terms of either definitive cure or long-term suppression. This is highlighted by the increasing incidence of difficult-to-treat infections, such as gram-negative PJI [29, 30] and methicillin resistant *Staphylococcus Aureus* (MRSA) [31]. Results of an in-vitro study utilizing a standardized medium, the Calgary Biofilm Device [32], demonstrated that Rifampicin and Tigecycline had low minimum biofilm eradication concentrations (MBEC), highlighting their antibiofilm activity against *S. aureus* and *S. epidermidis* which were isolated from patients diagnosed with PJI [33]. Similar results have been shown in other in-vitro studies [6], as well as in clinical evidence that supports the use of Rifampicin for PJI [31, 34].

In addition to systemic antibiotics, localized therapy using antibiotic-loaded cement spacers has demonstrated good efficacy in-vitro. Antibiotic elutions reach peak values within the first 2 or 3 days, inhibiting bacterial growth of *S. epidermidis* for 14–30 days [13]. Whilst there has been concern regarding the mechanical strength of antibiotic-loaded bone cement [35], a recent in-vitro study of a double layered spacer demonstrated beneficial biomechanical and drug-eluting properties [36]. Other constructs consisting of a vancomycin-loaded hydroxyapatite/poly amino acid scaffold [37] show consistent bactericidal effect on *S. aureus* and MRSA in-vitro, with good drug-eluting delivery over 38 days and also promote osteogenesis.

4.1.4 Modelling for Risk Factors

Some risk factors associated with PJI, including diabetes and obesity [38, 39], may also be studied in-vitro. It has been shown that elevated glucose levels aid the formation of biofilm [40] in an *E. faecalis* model. This may relate to findings that even perioperative hyperglycaemia in nondiabetic patients influences infection rates [41]. Whilst helpful, in-vitro studies have limited utility in this area, and these associations are potentially more relevant for in-vivo models.

4.1.5 Limitations

Whilst in-vitro models are effective for studying the cellular and biochemical responses to microorganisms and antibiotics, several elements are still lacking. Firstly, it is impossible to fully replicate in-vitro the complex bio-cellular environment that exists in an infected prosthetic joint. Secondly, bone metabolism and inflammatory responses occur in a dynamic weight bearing joint, which should be replicated experimentally. Thirdly, effectiveness of treatment needs to be examined in a living model which replicates as close as possible the biomechanical, immunological, and pharmacological dynamics in clinical PJI. Hence, in-vivo models in a controlled and ethical environment are required as the next step in the study of PJI pathogenesis and management.

4.2 In-Vivo Models of PJI

In-vivo models of prosthetic joint infection have developed significantly since Rodet's first experimental demonstrations in 1884 [42]. Rodet confirmed that osteomyelitis could be induced by inoculation of a "micrococcus" into rabbits, which formed localized infection at the femoral and tibial metaphysis. As a more recent guide, Carli et al. [43] described the ideal characteristics of a clinically representative model of PJI. In such a model: (1) biofilm can be formed on the prosthesis surface; (2) prosthesis materials should

be similar to clinical materials and create a similar intra-articular environment; (3) animals chosen should have similar musculoskeletal and immunological systems compared with humans; and (4) bacteria, biofilm, and host immune responses can be measured quantitatively.

4.2.1 Animal Characteristics

The majority of in-vivo implant and prosthetic joint related infections have been modelled in rabbits [44], in particular the New Zealand rabbit. The use of rabbits presents several advantages, including their docility, relatively low costs, reasonable size, and ease of handling. In their review, Bottagisio et al. [44] identified that rabbit tibias were used in 60.9% of in-vivo surgical sites and rabbit femurs in 27% of surgical sites. However, due to their bone fragility, rabbits have been shown to have a high incidence of postoperative fracture [45]. Studies are further limited by the substantial biomechanical and kinematic differences between the rabbit knee and the human knee. Despite these limitations, rabbits are still the most widely used animals for PJI models.

The second most common model is the murine model [46], typically C57BL/6 wildtype mice [3, 47–51]. Some studies euthanized mice between 3 and 6 weeks for harvesting of bone/joint tissue and implants for sonication [3, 51]. Mice occupy less space than rabbits, are easy to monitor and maintain, and can be bioengineered to emit fluorescent signals from immune cells [50, 52].

4.2.2 Prosthetic Designs

Various prosthetic designs have been utilized to model PJI. One of the first animal models of PJI, performed in 1976, utilized stainless steel particles infected with *S. aureus* into the suprapatellar bursa of rabbits [53]. Since then, multiple iterations of implants have been described.

Determining the relevance of various animal prosthesis trials to the human total knee arthroplasty (TKA) can be difficult. For example,

Kirschner-wires (K-wires) have limitations in that the prosthesis is a non-weight bearing, non-articulating, stainless steel construct, as opposed to the titanium alloy (Ti-6Al-4V) and cobalt-chromium-molybdenum alloy in human TKA. Secondly, whilst tibial plateau replacement has been shown to develop biofilm [48], it is unable to replicate the biomechanical dynamics of a total anatomical prosthesis. Finally, there is minimal consensus on the type of animal prosthesis to best mimic TKA PJI [46], and authors appreciate the impracticality and difficulty of miniaturizing total joint prostheses in animals [17].

Nevertheless, PJI studies have been performed on knee joint arthroplasty designs in rabbits as early as 1996 [54]. Much later, having identified a deficiency in prosthetic designs, Carli et al. [48] were the first to apply a three-dimensional printed tibial prosthesis with Ti-6Al-4V in a PJI murine model. Using this prosthesis, Carli et al. [47] revised the tibial replacement with a mouse-sized vancomycin eluting cement spacer, mimicking the first stage of a standard two-stage revision procedure for PJI in humans. Mice treated with antibiotic spacers had significantly lower inflammatory markers, had more preserved tibial bone, and had no intra-articular purulence. Retrieved spacers demonstrated lower bacterial counts compared with Ti-6Al-4V implants, although they did not have the same effect in periprosthetic tissue, suggesting that local antimicrobial activity was limited to the joint.

4.2.3 Gram-Positive Models

Given the leading causative group of microorganisms for PJI is gram-positive [29, 55, 56], studies have strongly focused on developing animal models to replicate these pathogens in-vivo [48, 50, 51, 57–60].

In order to study intraoperative contamination, rabbit models of early onset PJI have involved the injection of a high inoculum of *S. aureus* derived from an infected joint replacement into the joint space of a rabbit [54]. It was found that <102 colony forming units (CFU) of

S. aureus are necessary to establish infection in a rabbit hip hemiarthroplasty model, compared with 10⁴ CFU without a prosthetic implant. Infection remains within the joint space initially, then spreads to the adjacent metaphysis, with only the upper one-third of the metaphysis being involved at 3 weeks. Infection then progressed to involve the entire metaphysis of the periprosthetic bone. Other studies modelling haematogenous spread postulate that long bone osteomyelitis secondary to bacteraemia also begins in the metaphysis [42], then subsequently spreads to the implant. Furthermore, an in-vivo rabbit model of haematogenous spread demonstrated that lower levels of bacteraemia were needed to initiate PJI when inoculated in the immediate postoperative period compared to 3 weeks later [61].

In their mouse model of diabetes and implant related infection, Lovati et al. [62] demonstrated that diabetic mice showed severe infection resulting from *S. aureus* induced into the femur after an intramedullary pin implantation and an inability to respond to treatment with standard antibiotics alone. In their later work, Lovati et al. [63] showed that diabetic mice treated with a prostaglandin vasodilator in conjunction with antibiotics showed restrained signs of infection, pointing to a potential therapeutic combination in this at-risk group.

4.2.4 Gram-Negative Models

Gram-negative PJI was previously a rare complication, accounting for between 3 and 6% of all PJI [26, 64]. However, due to a rise in gram-negative PJI, between 15 and 36% [29, 30], a greater understanding of its pathogenesis will need to be developed. A model for the in-vivo experimentation of gram-negative PJI was established by Thompson et al. [3], which utilized an orthopaedic grade K-wire inserted into the femur of C57BL/6 mice with the implant protruding into the knee joint. Bacterial inoculation with either *P. aeruginosa* or *E. coli* was then injected into the joint before closure. It was found that 1 x 10⁴ Colony Forming Units (CFU) of *P. aeruginosa* were needed to achieve ade-

quate bioluminescence imaging signals, compared to 1 x 10⁵ CFU for *E. coli*. Furthermore, tissue and sonicated implants demonstrated greater bacterial growth of *P. aeruginosa* infected implants (67%), compared to *E. coli* infected implants (7%).

4.2.5 Biofilm Formation

Biofilm formation has been implicated as a major virulence mechanism of bacteria to adhere to tissue and protect the microorganism from antibiotics or the host immune system [65]. In a post-arthroplasty mouse model, Pribaz et al. [51] isolated four different strains of *S. aureus* and inoculated the knee joint of mice after implantation of stainless steel K-wire prostheses. All four strains demonstrated similar biofilm formation on scanning electron microscopy. In addition, these produced the same amount of infection induced inflammation as demonstrated by fluorescent neutrophil imaging. Thompson et al. [3] also utilized scanning electron microscopy for the detection of biofilm. They found implants infected with *P. aeruginosa* had more substantial biofilm formation on the intra-articular component of the implant, as well as adherent host immune cells, compared with *E. coli*. These results, together with the greater bacterial growth, suggest *P. aeruginosa* infection to be a more problematic disease.

4.2.6 Immune Reactions

Flow cytometry is commonly used to identify cell infiltrates into joint tissue. Whilst fewer pathogens are required to trigger infection in the presence of prosthetic implants, such implants induce substantial migration of cells including neutrophils and macrophages [3, 54]. Instead of activating phagocytosis, immune cells attempt to break down biofilm by releasing cytokines [66], reactive oxygen intermediates, and degradative enzymes. This process is complicated, however, by the depletion of local oxygen levels by bacterial communities, as well as restricted blood

perfusion [67]. Whilst most of these processes may be similar, it is important to recognize that inflammatory reactions in rodent models differ from humans; for example, no homolog of the human genes IL-26, CXCL8, and CXCR1 exists in mice. Furthermore, neutrophils are the predominant circulating leukocyte in humans, whilst lymphocytes circulate in higher ratios in mice [68].

4.2.7 Limitations

In-vivo studies identify many important aetiological and potential treatment factors for PJI, but still carry significant limitations. Studies in animals cannot translate directly to the periprosthetic environment in humans, due to differences in anatomy and biomechanical characteristics of animal bone, poorly reproducible prostheses, and other physiological, immunological, and genetic differences. Furthermore, the International Consensus on Orthopaedic Infection acknowledges that there is no established ideal prosthetic design for use in modelling PJI [46]. However, as with in-vitro models, the relevance and importance of in-vivo models will only strengthen with time and rigorous scientific application.

4.3 Conclusion

Orthopaedic surgeons and researchers are constantly evaluating the diagnostic tools and treatments for prosthetic joint infections [69, 70]. Although not without important limitations, in-vitro and in-vivo models will continue to be a relevant and growing research field to aid the management of PJI. Emerging technologies and advances, such as three-dimensional printing, manufacturing techniques, drug delivery systems, and gene specific therapies, will create exciting modelling opportunities for further study. The increase in total joint arthroplasties performed worldwide will drive orthopaedic surgeons to seek new and innovative ways to combat PJI. In a truly interdisciplinary field, in-vitro and in-vivo models of PJI will help solve unanswered

questions in the management of this complicated disease, which represents a significant clinical challenge and considerable burden for the entire orthopaedic community.

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Part III
Clinical Manifestation



General and Local Symptoms of Infection in Knee Replacement

5

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5.1 Introduction

Infection of periprosthetic joints is among the leading complications following total knee arthroplasty (TKA). The length and complexity of the treatment represent a physical, psychological and economic challenge for both the patient and the doctor. Despite the low incidence rate of infection after TKA, efforts to prevent infection and reduce the overall impact of periprosthetic joint infections (PJIs) seem justified by the increasing prevalence of TKA. In general, the prevention of PJI is based on the improvement of the host's defences, on the optimization of the conditions of the surgical wound and the minimization of microbial contamination throughout the entire treatment period [1]. The outcome of the treatment seems to be influenced by multiple variables such as the type of microorganism involved, the patient's comorbidities and the extent of soft tissue and bone involvement and the physician's expertise. For these reasons, the management of PJIs has become more rigorous and standardized in recent decades, mainly as regards the approach to the routine use of local antibiotics with high dosage and the necessary

deferment of reimplantation. Several risk factors relating to patients predispose to deep postoperative infection. Host factors include rheumatoid arthritis; skin ulcers; diabetes mellitus; past cancers; obesity; smoking habits; liver transplantation; HIV immunodeficiency virus seropositivity; open sky on the knee or a periarticular fracture; previous septic arthritis or adjacent osteomyelitis. Definitive diagnosis requires constant and considerable attention to the slightest clinical suspicion in both early and late infections. A meticulous anamnesis, the clinical examination, the study of imaging, arthrocentesis and haematological tests are all an integral part of the diagnostic process in the suspected infection. The timing of the clinical presentation is a crucial factor in the choice of the appropriate management strategy.

5.2 General Consideration

The different modalities of clinical presentation of the patient have been well described to define the most suitable management approach to PJI [2]. Postoperative infections diagnosed through the positivity of intraoperative culture tests during prosthetic revision are usually triggered by virulence-lowering microorganisms such as coagulase-negative staphylococci and Propionibacterium. A timely and precise diagnosis is necessary to avoid delays that can lead to

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diagnosing a prosthetic infection as chronic or late, which could instead be identified and treated in the acute phase. Pain is the main symptom of onset. Sustained wound secretion raises the highest suspicion of infection, which should be treated with arthrotomy, surgical toilet and flushing [3]. Culture tests for serous secretions are often challenging to portray and, therefore, they are not recommended. The empirical antibacterial therapy for persistent wound secretions should be prevented since it can merely relieve the clinical symptoms and can even delay the diagnosis, thus compromising the possibility of treating the infection without removing the prosthesis [4]. In the first few weeks after the surgical procedure, purposeful management of wound healing delays or marginal skin necrosis by the surgical toilet and closure by primary intention is desirable over empirical antibacterial treatment with long-term observation. This strategy may eventually result in the development of a profound infection [5]. An acute haematogenic infection typically occurs with unexpected onset of pain or tightness in a prosthesis that used to work well. An infectious outbreak elsewhere or a recent invasive procedure capable of triggering a battery should be evaluated as potential risk factors for the appearance of a haematogenic infection. The seriousness of symptoms such as pain, joint stiffness facilitates rapid diagnosis in these situations. Unfortunately, empirical antibiotic therapy is frequently prescribed for the appearance of painful symptoms without apparent cause in patients with prostheses, without attempting to make a precise diagnosis. This approach only serves to make subsequent attempts to detect a profound infection difficult. In the most infected TKA, the diagnosis is made in the subacute or chronic phase. The pain after surgery, prolonged secretion from the wound in the postoperative period, administration of antibiotic therapy for delayed healing and joint stiffness despite intense rehabilitation treatment are all elements that lay down for a deep infection. To date, PJI is defined as a variety of clinical symptoms and signs, tissue histological examination and culture examination. The certain diagnosis of infection is made if at least one of the following elements is found on

knee examination: two or more cultures from arthrocentesis or from deep tissues taken surgically are positive for the same microorganism; histopathological evaluation of intraarticular tissues detects alterations attributable to acute inflammation; frankly purulent secretion during the surgical procedure; the potential occurrence of a secreting fistula [6]. The identification and treatment of the microorganism responsible for the infection can ensure optimal results, where possible, and by implementing the consequent therapeutic strategies. Joint inspection, evaluation of knee function and measurement of range of motion are features that must be evaluated. Examination of the spine and ipsilateral hip is essential to avoid radicular or referred pain, respectively. Both neurological and vascular examination of the leg complete the clinical evaluation of the patient.

5.3 Clinical Examination

The patient who complains of persistent pain after TKA should receive an appropriate objective examination to assess the limb properly. In addition to normal physiological parameters such as vital signs, height, weight, a detailed examination of the skin for lesions, erythema and signs of infection such as heat, effusion, vascular changes and sinus tract drainage should be performed (Fig. 5.1). The presence of signs of wound dehiscence may indicate an infectious aetiology (Figs. 5.2 and 5.3). The evaluation of the gait is useful to highlight the presence of an antalgic thrust varus and valgus. Measurement of active and passive range of motion and the capability to actively sustain full extension without extensor delay must be included in the physical examination. The stability of the knee can be assessed with varus and valgus forces at 0° and 30°. In comparison, stability in the sagittal plane can be determined with posterior flexion tests at 60° and 90° flexion. Both the manual strength test and the assessment of muscle atrophy need to be observed. The knee has to be probed and evaluated for sensitivity with the iliotibial band and pes anserine bursa to exclude bursitis or flexion



Fig. 5.1 Periprosthetic joint infection of total knee arthroplasty; local signs of infection

instability. Because patellofemoral tracking is a common source of continuous pain after TKA, painful patellar crackling, patellar clunk syndrome, reduction in patellar size or patellar thickness, shortening of patellar tendon length, increased posterior femoral condylar offset, utilization of smaller femoral parts, thicker tibial polyethylene inserts and greater flexion of femoral constituents should be evaluated. These factors increase the contact of the quadriceps tendon with the upper aspect of the intercondylar box and, consequently, the risk of fibrosynovial proliferation increases. A neurovascular examination should be conducted to assess the quality and symmetry of peripheral pulses and the strength of both the quadriceps and the vastus medialis obliquus. The physical examination should con-



Fig. 5.2 Periprosthetic joint infection of total knee arthroplasty; wound dehiscence

clude with an evaluation of the spine, and other joints to exclude other extra-articular causes of pain, such as lumbar radiculopathy, referred pain from coxarthrosis and vascular claudication. In some patients, the replacement can be clearly visible through a skin hole (Fig. 5.4).

5.4 Presenting Symptoms and Clinical Assessment

When a patient reports a painful TKA, the clinical suspicion of PJI should always be investigated even if there are no apparent signs of infection, such as redness or swelling. The clinical onset of PJIs after TKA is often blurred and insidious. History of wound drainage, invasive procedures or dental procedures and comorbidities such as diabetes mellitus or immunosuppressive conditions increase the risk of PJIs and, therefore, meticulous evaluation [7, 8]. Although the progress in laboratory and imaging techniques may



Fig. 5.3 Periprosthetic joint infection of total knee arthroplasty; wound dehiscence

enhance the detection of PJI, the clinical presentation is still the basis for diagnosis. Stratification of the risk of the likelihood of infection is warranted in any patient presenting with a painful TKA [9]. The diagnosis of a PJI after TKA is critical because the treatment of infected patients is very different from the treatment of uncomplicated TKA. Faced with suspicion of PJI, a comprehensive and more thorough evaluation of the patient is necessary. Possible risk factors for infection are obesity, inflammatory arthritis, diabetes, malnutrition, early implant mobilization (<5 years) and early osteolysis (<5 years). The signs or symptoms most commonly complained of by patients with chronic PJI are fever, pain, joint effusion and periarticular erythema [10, 11]. Despite the clinical



Fig. 5.4 The replacement visible through a skin hole

presentation of the infection after TKA, to date, no study evaluates the role of physical examination for the diagnosis of PJI [12]. While aseptic prosthetic loosening pain increases with weight gain and decreases at rest, persistent and progressive pain at rest is one of the PJI's first symptoms. This pain occurs at the time of surgery and has been associated with superficial infection, leakage or healing problems of the wound. In contrast, an acute haematogenic infection can occur in a previously painless and well-functioning knee. The clinical picture of an infected prosthetic joint varies about the source of infection, the time required for the infection to develop and the viral load of the infecting organism. The classic presentation is that of a painful joint even at rest, hot and erythematous (Fig. 5.5). The most common symptom of prosthetic infections is pain, present in more than 90% of patients. As it is difficult to differentiate the pain caused by an aseptic mobilization from that of a prosthetic infection, this symptom alone has a low diagnostic prediction. Joint loosening pain occurs mainly with movement or load, while infection pain is less likely to be associated



Fig. 5.5 Local signs of infection with hot and erythematous skin

with joint loading, has a more constant duration and a tendency to increase over time. Other symptoms, such as fever, swelling and suppuration, occur to a lesser extent, and their frequency and intensity are related to the patient's age. In very elderly subjects or therapy with corticosteroids or other anti-inflammatories, the febrile response may be modest or completely absent, especially in late infections or supported by low virulent strains. Rarely, the patient in good general condition and with age < 60 years comes to the observation with manifest septicemia, high fever, hypotension and multi-organ dysfunction. The triad of pain at rest, fever and localized oedema takes on high predictive value and constitutes, even in the absence of microbiological confirmation, sufficient reason to establish empirical therapy. In the presence of suspected infection, an arthrocentesis must always be performed and, if the conditions require it, an empirical therapy

must be set up pending a crop assessment. Despite clinical presentation, treatment with empirical antibiotics for presumed cellulite or superficial infection is not recommended without a definite diagnosis. Antibiotic treatment can suppress clinical symptoms, decrease the ability to isolate the microorganism and delay diagnosis [13].

5.5 Wound Complications

Wound complications after TKA are dramatic and responsible for increased risk of PJI and other postoperative complications such as component resection, myofascial or fasciocutaneous flap reconstruction or amputation [14]. Wound healing may include three phases: inflammation, a fibroblastic proliferative phase and a wound maturation phase. The faster gains in wound strength occur during the first weeks of wound healing. In contrast, the maturation phase of the wound proceeds for several months as the collagen fibrils become increasingly structured and well-organized. There are several patient-specific, intraoperative and postoperative factors that can influence normal wound healing [14]. Intraoperative factors impacting the healing process include the site of the incision, soft tissue flap management and appropriate tissue handling.

Wound complications differ in prolonged postoperative drainage, superficial or full-thickness soft tissue necrosis. The increase in the severity of the infection occurs if the treatment of these complications is delayed. Although prolonged serous drainage is performed, this represents a difficult challenge after TKA. Initially, a chronically draining wound in the absence of erythema or purulence can be managed with local wound care, elevation and immobilization. Surgical debridement is required when drainage persists for more than 5–7 days. Persistent wound drainage is often due to the presence of a sizeable subcutaneous hematoma or intraarticular hemarthrosis. Hematomas increase the soft tissue tension and create an ideal medium for bacterial growth [15].

Small necrotic lesions with a diameter of less than 3 cm, usually located at the edges of the

wound can be treated with local wound care and delayed secondary closure. Careful surveillance of these small necrotic lesions is imperative. Superficial soft tissue necrosis usually demands surgical debridement. Necrotic lesions with a diameter greater than 3 cm usually demand split-thickness skin grafts, fasciocutaneous flap or myocardial flap coverage. It has been shown that closing the wound with the aid of vacuum can reduce oedema from the extravascular space, improve blood supply and wound granulation, suppress bacterial proliferation and reduce wound size. Vacuum wound closure systems may be useful to reduce wound size before any soft tissue covering procedures or may facilitate wound healing without additional surgery. In cases of full-thickness necrosis, the prosthetic components are usually exposed, and immediate debridement is required. After irrigation and debridement, secondary closure is rarely successful. Vascularized tissue transfer with fasciocutaneous, myocutaneous and myotendinous flaps have been described [5]. Consultation plastic surgeon can aid in determining appropriate flap coverage.

5.6 Conclusions

The clinical presentation can be hugely useful to guide the diagnosis, but PJI must be confirmed by clinical tests. Differences in pre-test probability can also greatly influence the post-test probability of patients with similar laboratory results. For example, two patients with elevated ESR and PCR, but no other elevation of serum or synovial marker may have a different probability of PJI based on differences in their clinical presentation. While the importance of pre-test probability is recognized by the American Academy of Orthopaedic Surgeons (AAOS), no attention is given to the clinical presentation of the patient.

Although clinical presentation in PJI currently plays a marginal role in diagnostic guidelines, fever and erythema around the joint are suggestive findings of PJI. The existing diagnostic criteria are based on both cultures and laboratory results to define PJI [8, 16]. The analysis of the patient's signs and symptoms with PJI is non-

invasive, simple and can substantially guide the diagnosis. Pain may be the only symptom of chronic infection (especially in cases of low virulence) and is a sufficient symptom to warrant further evaluation to exclude PJI. Compared to aseptic revisions, the presence of joint effusion appears to be significantly higher in patients with PJI [17]. The difficulties encountered in treating infections following TKA can be considerable, and the treatment must be carefully planned after appropriate clinical and diagnostic work-up.

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Part IV

Diagnosis



Laboratory Diagnosis of Periprosthetic Joint Infections

6

Graham S. Goh and Javad Parvizi

6.1 Introduction

Total knee arthroplasty (TKA) is one of the most common elective surgical procedures in the world. The volume of primary and revision TKA has risen dramatically and is projected to grow over the next decade [1]. Periprosthetic joint infection (PJI) is a rare but devastating complication after TKA, with an estimated risk of 0.5–2% following primary procedures [2]. Despite the low incidence of this complication, PJI is the most common indication for revision in the Medicare population [3] and the main cause of failure in modern total joint arthroplasty (TJA) [4, 5]. With over a million joint replacement procedures performed each year in the USA [6], the overall burden of PJI will also invariably increase. Despite global efforts to reduce the incidence of PJI, several international arthroplasty registries have shown that the infection burden has in fact increased over time [7]. Furthermore, as the prevalence of risk factors such as obesity and diabetes increases around the world [8, 9], some authors have also projected an increase in PJI rates in the near future [10]. This rare but devastating com-

plication is not only associated with a significantly increased risk of mortality and decreased quality of life [11, 12], but also poses a substantial economic burden to the healthcare system as costs can be up to four times higher than that of uninfected cases [13]. As the management of an infected knee arthroplasty is drastically different from aseptic cases, it is imperative that orthopedic surgeons definitively establish or rule out the diagnosis of PJI prior to revision surgery, helping patients avoid the increased morbidity and costs associated with this complication wherever possible.

The lack of a “gold standard” diagnostic test makes the diagnosis of PJI extremely challenging. Historically, there has been no standardized criteria or algorithm for the diagnosis of PJI, which led to the use of a wide variety of tests and procedures that were unnecessarily burdensome and costly for patients, often resulting in treatment delays or misdiagnosis. Recently, several evidence-based guidelines have been introduced to standardize the approach to a patient with a suspected PJI, including the American Academy of Orthopaedic Surgeons (AAOS) Clinical Practice Guidelines on Diagnosis of Periprosthetic Joint Infection [14] as well as the Proceedings of the 2018 International Consensus Meeting (ICM) on Periprosthetic Joint Infection (Fig. 6.1) [15]. These documents should be familiar to all orthopedic surgeons as well as other physicians who

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| Major criteria (at least one of the following) | Decision |
|--|----------|
| Two positive cultures of the same organism | Infected |
| Sinus tract with evidence of communication to the joint or visualization of the prosthesis | |

| Preoperative Diagnosis | Minor Criteria | | Score | Decision |
|------------------------|----------------|---|-------|---|
| | Serum | Elevated CRP <i>or</i> D-Dimer | 2 | ≥ 6 Infected 2-5 Possibly Infected* 0-1 Not Infected |
| | | Elevated ESR | 1 | |
| | Synovial | Elevated Synovial WBC <i>or</i> LE (++) | 3 | |
| | | Positive Alpha-defensin | 3 | |
| | | Elevated Synovial PMN % | 2 | |
| | | Elevated Synovial CRP | 1 | |

| Postoperative Diagnosis | *Inconclusive pre-op score <i>or</i> dry tap | | Score | Decision |
|-------------------------|--|--|-------|--|
| | Preoperative score | | - | ≥ 6 Infected 4-5 Inconclusive** ≤ 3 Not Infected |
| | Positive Histology | | 3 | |
| | Positive Purulence | | 3 | |
| | Positive Single Culture | | 2 | |

Fig. 6.1 Evidence-based criteria for the diagnosis of periprosthetic joint infections as recommended by the 2018 International Consensus Meeting (ICM). *For patients with inconclusive minor criteria, operative criteria can also be used to fulfill definition for PJI for PJI. **Consider fur-

ther molecular diagnostics such as Next-generation sequencing. (Reprinted with permission from “The 2018 Definition of Periprosthetic Hip and Knee Infection: An Evidence-Based and Validated Criteria.” The Journal of Arthroplasty. Elsevier; 2018;)

frequently encounter patients with joint replacements in their practice.

The general approach to diagnosing a PJI is twofold. First, the presence or absence of a joint infection must be confirmed; second, the infecting microorganism(s) must be isolated and its antimicrobial susceptibility elucidated. In addition to clinical findings from history and physical examination, the diagnosis of PJI often relies on laboratory results from peripheral blood and synovial fluid, microbiological evaluation, histological examination of periprosthetic tissue, intraoperative findings, and in some cases, radiographic evaluation [16–18]. In particular, isolating the causative microorganism from cultures of fluid or tissue within the joint remains the cornerstone for diagnosis and targeted antibiotic therapy, which has been shown to increase the chances of treatment success [19] and influence the prognosis of

patients with this condition [16]. This chapter reviews the laboratory tests available in an orthopedic surgeon’s armamentarium to diagnose PJI following knee replacement surgery.

6.2 Peripheral Blood Tests

Serum biomarkers are useful adjuncts in the diagnosis of PJI [16, 17], especially in the absence of major diagnostic criteria such as a communicating sinus tract or two positive cultures [15]. Biomarkers are measurable biological substances that are part of a physiological or pathological pathway or the pharmacological response to therapeutic interventions [20]. Given their high accessibility, peripheral blood tests are often first-line investigations for any patient with a suspected PJI.

6.2.1 Erythrocyte Sedimentation Rate and C-Reactive Protein

Erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) are two of the most well-researched serological biomarkers in the diagnosis of PJI. CRP is an acute-phase reactant that is produced by the liver in response to systemic infections. ESR is the rate at which red blood cells form a sediment at the bottom of a standardized tube, which is increased by the presence of fibrinogen and other clotting factors that are produced during inflammation. While non-specific for the diagnosis of localized infections due to their elevation in non-infectious inflamma-

tory conditions, these tests are often used as first-line screening tools for PJI due to their high sensitivities exceeding 90%, making them particularly valuable in ruling out PJI [21–23]. When both ESR and CRP are below their diagnostic thresholds of 30 mm/h and 10 mg/L, respectively, the negative likelihood ratio of PJI ranges from 0 to 0.06 [21]. As such, the use of these markers in the first step of the evaluation of a patient with suspected PJI has been endorsed by the 2018 ICM (Fig. 6.2) [24].

Despite the accessibility and utility of these markers, ESR and CRP may be falsely low or normal in cases when the infecting organism is slow-growing and may not elicit an adequate

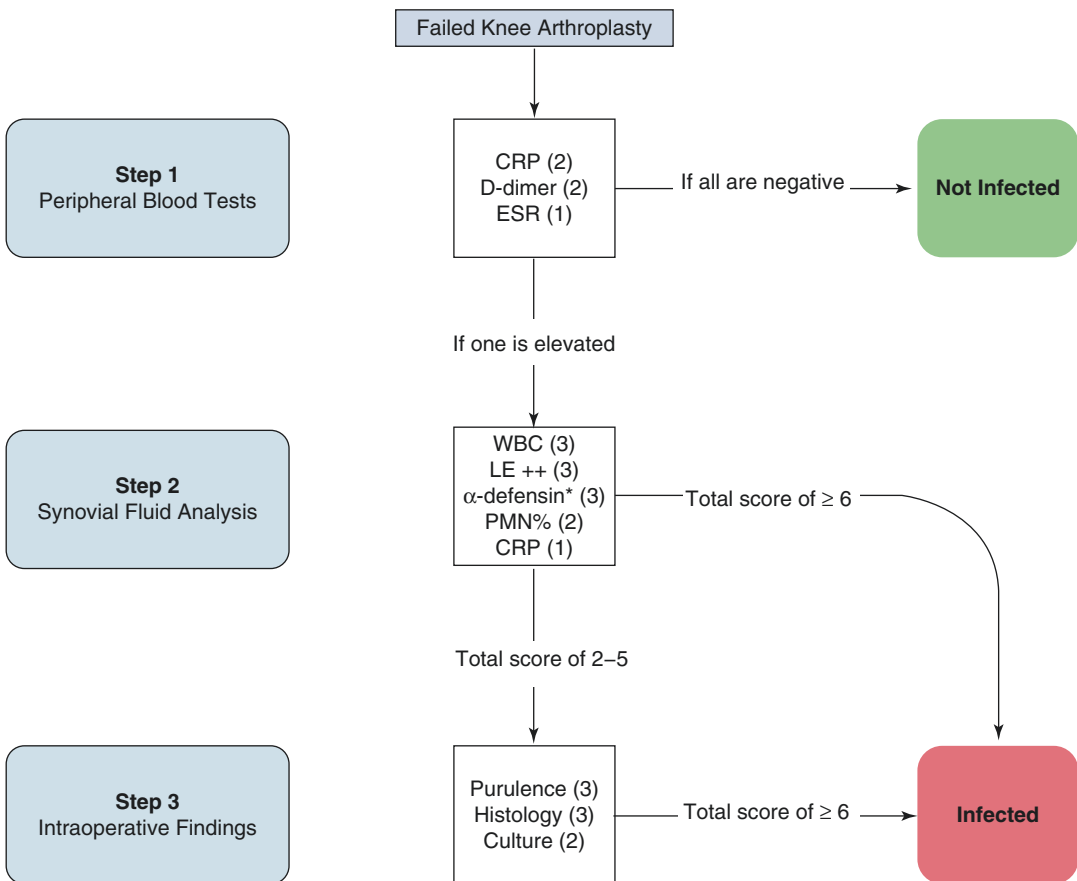


Fig. 6.2 Diagnostic algorithm to guide the selection of laboratory tests. *CRP* C-reactive protein, *ESR* erythrocyte sedimentation rate, *WBC* white blood cell count, *LE* leukocyte esterase, *PMN%* polymorphonuclear neutrophil percentage. Points are stated in parentheses. *Does not

need to be performed routinely. (Reprinted with permission from “Development and Validation of an Evidence-Based Algorithm for Diagnosing Periprosthetic Joint Infection” The Journal of Arthroplasty. Elsevier; 2019;)

inflammatory response [25–28]. Perez et al. found that ESR and CRP were not as accurate in diagnosing PJI caused by low-virulence organisms such as coagulase-negative staphylococci, *Bacillus* spp, *Corynebacterium* spp, and *Cutibacterium acnes* (previously known as *Propionibacterium acnes*) [27]. Similarly, Akgün et al. demonstrated that CRP was not a reliable screening marker for PJI and led to high false-negative rates especially in cases with a low-virulence organism [28]. It is also worth noting that these markers are not specific for infection-induced inflammation and may be elevated in other inflammatory states including autoimmune diseases and tissue damage from trauma or surgery. Conversely, the anti-inflammatory effect of systemic corticosteroids may also decrease their concentrations [29]. Although Cipriano et al. demonstrated that ESR and CRP had a similar diagnostic accuracy in patients with and without inflammatory arthritis [30], their limited specificity cautions against the isolated use of these markers to diagnose PJI in complex clinical situations.

6.2.2 D-Dimer

Similar to traditional inflammatory markers such as CRP and ESR, the measurement of serum D-dimer has gained attention as a valuable prognostic tool in patients with systemic sepsis and bacteremia [31, 32]. Systemic and local infections are known to stimulate fibrinolytic activity and coagulation abnormalities as part of the host inflammatory response [33, 34]. This in turn leads to an elevation in breakdown products of fibrinolysis such as D-dimer [35]. The increased fibrinolytic activity and creation of byproducts are hypothesized to trap infecting microorganism(s) and/or inflammatory cells to limit the extent of systemic damage. Joint inflammation or infection in particular has been shown to elicit a rise in D-dimer levels due to the high concentration of fibrin released by inflamed synovium [31, 32, 34], which breaks down into degradation products and increases the concentration of D-dimer in serum and synovial fluid

[36]. In an *in vivo* study of foals with septic arthritis, Ribera et al. found that the concentration of D-dimer in synovial fluid was markedly elevated, reinforcing the notion that D-dimer is a mediator of joint inflammation or infection [34]. It is thus not surprising that recent studies in arthroplasty literature have extrapolated this concept and identified D-dimer as a promising biomarker for the diagnosis of PJI [37]. In the initial study on 245 primary and revision arthroplasty patients, Shahi et al. found that serum D-dimer was more accurate in diagnosing PJI than ESR and CRP combined, with a sensitivity and specificity of 89% and 93% compared to 84% and 47%, respectively [37]. Importantly, the authors also showed that D-dimer was only elevated in 12% of patients with an infection in sites other than the prosthetic joint, in contrast to ESR and CRP, which were elevated in 100% and 84% of these patients, respectively. Another advantage cited by proponents of this novel biomarker is its utility in ascertaining the optimal timing for reimplantation in patients undergoing two-stage exchange arthroplasty for PJI. ESR and CRP are not reliable in this aspect due to their elevated levels in the postoperative period [38, 39], which has led several authors to conclude that these markers are poorly predictive of treatment failure at the time of reimplantation [40, 41]. On the contrary, D-dimer rises and falls more rapidly in the early postoperative period compared to ESR and CRP, returning to baseline levels by postoperative day 2 before reaching a second peak at postoperative week 2 [42]. Shahi et al. demonstrated that two of five patients who were free of infection as defined by the Musculoskeletal Infection Society (MSIS) criteria but had elevated D-dimer levels at the time of the second stage subsequently failed treatment for PJI [37], thus supporting the role of D-dimer as a prognostic marker for patients undergoing reimplantation. Practical considerations that further enhance the appeal of this novel biomarker are its relatively low cost and high accessibility in routine clinical practice [37]. These factors have culminated in the endorsement of serum D-dimer as an inexpensive and reliable test for the diagnostic workup of PJI [24].

Despite its utility, recent studies have questioned the usefulness of D-dimer for the diagnosis of PJI. Li et al. identified 565 patients who underwent revision surgery (95 PJI cases, 470 aseptic cases) and found that the area under the curve (AUC) for plasma D-dimer was 0.657, with an optimal threshold of 1250 ng/mL, sensitivity of 64%, and specificity of 65% [43]. The authors postulated that the use of plasma D-dimer instead of serum D-dimer as well as the predominantly Asian cohort could have accounted for the inconsistent findings when compared to the study by Shahi et al. [37]. Serum samples, unlike plasma ones, are obtained following the consumption of coagulation factors, which may hence alter D-dimer levels. Despite this methodological difference in measurement, current evidence suggests that plasma and serum D-dimer concentrations are very highly correlated and show a strong linear relationship [44, 45]. In another study evaluating plasma D-dimer, Xu et al. found that a threshold of 1.02 mg/L fibrinogen equivalent units (FEU) to discriminate between infected and non-infected revisions demonstrated poor sensitivity (68%) and specificity (51%) [46]. Similarly, Pannu et al. found that the threshold of 850 ng/mL led to a good sensitivity (96%) but poor specificity (32%) with an AUC of 0.742 [47], whereas a different cutoff point of 2300 ng/mL had a moderate sensitivity of 71% and improved specificity of 74%, although these values were still much lower than the values reported in the original study by Shahi et al. [37]. It is possible that the lack of consensus regarding the diagnostic threshold for D-dimer in the literature could have arisen from the inconsistencies in laboratory techniques used to measure this biomarker. The lack of standardization among the various assays has been a topic of dispute, with several authors questioning the clinical utility of D-dimer in view of the high variability in its measurement and reporting [48]. An additional reason could be that the proposed threshold by Shahi et al. may not be the optimal one, as evidenced from the aforementioned studies. While D-dimer is an inexpensive and accessible assay in routine practice that has the potential to detect PJI with a higher sensitivity and specific-

ity, additional studies with consistent laboratory measurement methods are needed to confirm its superior diagnostic performance over traditional tests and determine the optimal threshold value for this marker.

6.2.3 Fibrinogen

Similar to the rationale for the use of D-dimer in systemic and local infections [33, 34], other fibrinolytic markers such as fibrinogen and fibrinogen degradation product (FDP) have gained recognition for their accuracy in diagnosing PJI [43]. Fibrinogen is a glycoprotein found in human plasma that is converted into fibrin by thrombin for the formation of a fibrin-based blood clot in the final steps of the coagulation cascade [33]. It is also a positive acute-phase reactant that increases in concentration during inflammation. When evaluating the relationship between thrombosis and infection, Kirschenbaum et al. found that fibrinogen played an integral role in neutrophil adherence during systemic sepsis [49], while Horn et al. found that neutrophil alpha-defensins stimulated the production of fibrinogen and thrombospondin-1 amyloid-like structures to entrap infecting microorganisms [50]. Not surprisingly, the use of fibrinogen has also garnered attention from the arthroplasty community recently. In the study by Li et al., plasma fibrinogen was found to be a promising biomarker with the highest AUC compared to plasma D-dimer, ESR, or CRP [43]. When 4.01 g/L was used as the cutoff point, the sensitivity and specificity were 76% and 86%, respectively. In another study, Wu et al. noted that the AUC for fibrinogen was higher than that of FDP or D-dimer [51]. The authors determined the optimal threshold for fibrinogen to be 3.61 g/L, with 76% sensitivity and 86% specificity. Similar to this study, Xu et al. found that plasma fibrinogen was useful for diagnosing PJI as well as confirming the presence of persistent infection at the time of reimplantation [52]. These encouraging findings support the use of plasma fibrinogen in the evaluation of a patient with suspected PJI before revision surgery and at the time of reimplantation,

although further studies are needed to validate these findings in more diverse populations.

6.2.4 Interleukin-6

Interleukin-6 (IL-6) is pleiotropic cytokine produced by macrophages in response to tissue injury. It stimulates the production of acute-phase reactants such as CRP, regulates pyrexia by pituitary hormones, modulates bone resorption, promotes hematopoiesis, and induces plasma cell development [53]. Serum IL-6 has been established as a valuable inflammatory marker in association with sepsis, trauma, and major surgery [54, 55]. Given that IL-6 lies upstream of other markers, such as CRP, in the inflammatory cascade [56], it is postulated to be a more rapid and sensitive marker for the detection of PJI [57]. IL-6 exhibits a more rapid increase and return to baseline levels following TJA compared to ESR or CRP, often peaking within the first 6–12 h after surgery and returning to baseline within the first two to three postoperative days, leading authors to conclude that IL-6 may be a superior indicator of an early-stage immune response [58, 59]. Importantly, studies have shown that IL-6 is more elevated in patients with PJI compared to patients with aseptic loosening [60]. Berbari et al. found that the pooled sensitivity and specificity for IL-6 in three studies were 97% and 87%, respectively [61], and the authors concluded that serum IL-6 had the highest accuracy in diagnosing PJI when compared to ESR and CRP. In a more recent meta-analysis of eleven studies, Xie et al. reported a pooled sensitivity and specificity of 72% and 89%, respectively, although the poorer sensitivity was likely due to the inclusion of two studies on shoulder PJIs that had a higher proportion of low-virulence organisms such as *Cutibacterium acnes* [57]. Overall, current literature suggests that serum IL-6 may be a promising marker for the diagnosis of PJI with a relatively high diagnostic accuracy. A growing interest in the use of IL-6 has led to its incorporation into the latest clinical practice guidelines by the AAOS [14]. Notwithstanding, serum IL-6 has only been evaluated in a small number of studies

and the optimal threshold for this marker has yet to be determined. Current barriers to its use include the relatively high cost and technical skills required to run the analysis. As serum IL-6 assays become more widely available for clinical use, this biomarker could be used in combination with other routine markers like serum CRP, further enhancing their diagnostic yield as shown in previous studies [62, 63].

6.2.5 Procalcitonin

Procalcitonin (PCT) is a protein produced by thyroid parafollicular C-cells and lung neuroendocrine cells. Serum PCT levels are undetectable in healthy individuals without evidence of infection and are greatly elevated in bacterial infections, giving the biomarker a high diagnostic accuracy for the identification of systemic infection [64]. The utility of serum PCT for diagnosing PJI has been investigated in several studies [62, 65, 66]. In a meta-analysis of six studies on PCT, the pooled sensitivity and specificity were 53% and 92%, respectively, making this test suitable as a rule-in rather than a rule-out diagnostic tool [66]. Similarly, Boettner et al. investigated the serum levels of PCT, IL-6, tumor necrosis factor (TNF)- α , ESR, and CRP in 78 patients undergoing revision arthroplasty for sepsis and found that serum PCT was very specific (98%) but had a very low sensitivity (33%) [62]. Based on current evidence, serum PCT has a limited diagnostic value as a biomarker for PJI due to its low sensitivity and should not be routinely used in the workup of an infected knee replacement.

6.2.6 Novel Serological Markers

In recent years, a myriad of serological markers have been evaluated for their diagnostic potential. Some examples of these novel biomarkers include TNF- α [62, 63], lipopolysaccharide-binding protein (LBP) [63, 67], toll-like receptor (TLRs) [68], intercellular adhesion molecule-1 (ICAM-1) [69, 70], soluble urokinase plasminogen activation receptor (su-PAR) [71], and CD64

[72]. While a few of these markers have shown a high diagnostic accuracy, practical considerations such as the high cost and technical competency required to run these tests remain important barriers to their adoption into clinical practice. Consequently, further research to discover accurate and clinically relevant serum biomarkers for the diagnosis of PJI is necessary.

6.3 Synovial Fluid Analysis

Joint aspiration is a crucial step in the diagnostic workup of any patient with a suspected joint infection [15]. Although listed as the second step in the ICM diagnostic algorithm [73], this procedure is commonly performed in the office alongside the abovementioned blood tests. Synovial fluid biomarkers play an integral role in the diagnosis of PJI and have been incorporated into recent guidelines as minor diagnostic criteria [15]. In a comprehensive meta-analysis by Carli et al., synovial fluid tests were found to have a superior diagnostic accuracy compared to serum or tissue-based diagnostic tests [74]. The authors identified five synovial fluid tests (alpha-defensin laboratory-based or lateral flow test, CRP, leukocyte esterase strips, polymorphonuclear neutrophil percentage, and white blood cell count) that had the best fitted hierarchical summary receiver operating characteristic (HSROC) curves and highest diagnostic performance of all 17 tests analyzed, concluding that any aspiration should utilize one of these five tests whenever PJI is suspected. This section will focus on these five synovial fluid biomarkers as well as new emerging tests for the diagnosis of PJI.

6.3.1 White Blood Cell Count and Polymorphonuclear Neutrophil Percentage

White blood cell (WBC) count and polymorphonuclear neutrophil percentage (PMN%) are two important synovial fluid tests that have been validated as minor criteria in the current evidence-based definition of PJI [15]. Multiple studies

have evaluated the accuracy of these markers for the diagnosis of PJI [75–78]. Unlike other criteria, it is important to note that the diagnostic thresholds for these biomarkers vary based on the timing of infection. For acute PJI within 6 weeks postoperatively, a threshold of >10,000 cells/ μ L for synovial WBC count and >90% for synovial PMN% should be used to diagnose PJI, whereas for chronic PJI greater than 6 weeks postoperatively, a threshold of >3000 cells/ μ L and >80% should be used [15]. Using these cutoff points, Shahi et al. found a sensitivity and specificity of 86% and 83% for synovial WBC count, respectively, as well as a sensitivity and specificity of 86% and 81% for synovial PMN%, respectively [79].

When using these biomarkers, clinicians should be cognizant of certain clinical scenarios that may negatively influence the accuracy of results [80, 81]. As is the case with other tests, the type of microorganism and premature use of antimicrobials before joint aspiration have been shown to impact diagnostic thresholds [76, 82], although a concurrent diagnosis of inflammatory arthritis did not appear to do so [30]. In the setting of a traumatic aspiration, a validated formula that adjusts for the synovial red blood cell, serum red blood cell, and serum WBC counts should be used to calculate the corrected synovial WBC count [80]. Additionally, Kwon et al. noted that automated synovial fluid WBC count and PMN% could be unreliable in the context of a failed metal-on-metal implant or corrosion reaction [83]. As the presence of macrophages with phagocytosed metal or amorphous material, fragmented cells, or blood clots can lead to inaccuracies when performing an automated cell count [81], it has been recommended that such cases require a manual synovial WBC count to alert the surgeon of the possibility of a false-positive result [84]. Similarly, the inability to generate a WBC differential in this context should cast doubts on the reliability of the automated synovial fluid WBC count. In a previous study by Wyles et al., the authors found that of the 35 metal-on-metal THAs analyzed, 12 (34%) had a false-positive synovial fluid WBC count (>10,000 cells/IL) and the differential could not be generated in 16

(46%) samples [85]. While these caveats may be more relevant to hip corrosion, a recent study by Deirmengian et al. also found a high rate of false positivity using automated cell counters to analyze synovial fluid of hip and knee arthroplasties [86]. Although false-positive rates were higher in THAs (34%), the frequency (10%) and magnitude of false-positive automated synovial fluid WBC counts were also concerning for TKAs, with higher modified ICM scores and culture positivity confirming the accuracy of manual rather than automated WBC counts [86]. These findings highlight the need to verify the accuracy of positive automated synovial fluid WBC counts with manual counts, as well as the need to integrate other synovial fluid tests and minor diagnostic criteria to reconcile any inconsistencies.

6.3.2 Leukocyte Esterase

Leukocyte esterase is an enzyme secreted by activated neutrophils following their migration to the site of infection. It has traditionally been used in the diagnosis of urinary tract infections, although its diagnostic utility in the workup of PJI has also gained recognition recently [87]. Leukocyte esterase tests are readily available, point-of-care tests requiring the application of infected joint fluid onto colorimetric strips. Detection of the enzyme is then reflected as a color change on the test strip [87], providing almost immediate results and guiding intraoperative decision-making. Furthermore, leukocyte esterase testing is currently the simplest and cheapest test available with an estimated cost of 0.17 USD per test [88]. Despite its accessibility and ease of use, one major limitation is the potential for blood contamination within the fluid samples to interfere with the colorimetric changes of the test strip [89], although this may be overcome by centrifuging synovial fluid samples for 2–3 min prior to application [90]. Excellent diagnostic accuracy has been demonstrated in previous studies, with Wetters et al. reporting a sensitivity of 92.9–93.3% and specificity of 77.0–88.8% [89], and Tischler et al. reporting a sensitivity of 79% and specificity of 81% in 221 patients that fulfilled

MSIS criteria for PJI [91]. In a meta-analysis of five studies, Wyatt et al. also found a sensitivity and specificity of 81% (95% CI, 49–95%) and 97% (95% CI, 82–99%) using a (++) reading, respectively [88]. When considering (++) as the diagnostic threshold instead of (+), Carli et al. reported a higher specificity (97% vs. 84%) at the expense of a slightly lower sensitivity (93% vs. 96%) [74]. A third study compared its diagnostic accuracy with that of other laboratory tests and found the highest diagnostic odds ratio for the leukocyte esterase strip test (OR 30.06, 95% CI 17.8–50.7) [79].

6.3.3 Alpha-Defensin

Alpha-defensin is another synovial fluid biomarker that has a high accuracy when used in the diagnostic workup for PJI [92]. Defensins are naturally occurring antimicrobial peptides that are part of the innate immune response against most gram positive and negative bacteria, fungi, and enveloped viruses [93]. They are commonly secreted by neutrophils as well as certain macrophage cell lines, and their synthesis is induced by pro-inflammatory cytokines or microbiological products. While their precise antimicrobial mechanism has yet to be fully elucidated, alpha-defensins are generally believed to cause a disruption in pathogen membrane integrity, resulting in cell lysis [94, 95]. Current literature has demonstrated the utility of alpha-defensin as a diagnostic tool for PJI, with studies reporting a sensitivity and specificity of over 95% [96, 97]. Bingham et al. even suggested that the diagnostic accuracy of synovial fluid alpha-defensin assays exceeded that of all other available tests [98]. This was confirmed in a recent meta-analysis by Carli et al., which found that laboratory-based alpha-defensin tests and leukocyte esterase strips (++) had a near-perfect diagnostic performance with the best fitted HSROC curves compared to other tests [74]. However, it is important to distinguish between the two available methods to analyze alpha-defensin: (1) the laboratory-based alpha-defensin immunoassay, which is a quantitative test that takes up to 24 h to complete and

(2) the alpha-defensin lateral flow test, which is a standalone device that produces a qualitative binary result within approximately 10 min. In the aforementioned study by Carli et al., the lateral flow test kit had a lower pooled sensitivity of 82% compared to the laboratory-based test, although a high specificity was still maintained [74]. These findings were echoed by previous systematic reviews and meta-analyses on alpha-defensin [99–102]. Eriksson et al. found a lower overall diagnostic accuracy for the lateral flow test compared to the laboratory immunoassay (AUC 0.75 vs. 0.98), whereas no difference in specificity was found (90% vs. 96%; $p = 0.06$) [100]. Similarly, Suen et al. analyzed ten studies and reported a pooled sensitivity and specificity of 77% and 91% for the lateral flow test, which was lower than that of the immunoassay (sensitivity 95%, specificity 97%) [102]. Although a recent meta-analysis by Kuiper et al. questioned the difference in sensitivity [103], current evidence still suggests that the lateral flow test may be a more suitable test for ruling in rather than ruling out infection [100]. Notwithstanding, several advantages of the lateral flow device exist, such as the improved accessibility of a point-of-care test with a rapid response time, obviating the need to ship samples to a centralized laboratory for analysis as in the case of the laboratory-based immunoassay.

Specific clinical scenarios may influence the diagnostic performance of alpha-defensin. When comparing lateral flow test results of 109 cases with the 2013 ICM definition of PJI, Plate et al. found a higher false-positive rate in patients with a concurrent diagnosis of inflammatory arthritides including crystal deposition disease, concluding that an assessment for crystal deposition in synovial fluid aspirates should also be performed if this test is to be used for the diagnosis of PJI [104]. As is the case with synovial fluid WBC counts, corrosion reactions seen in failed metal-on-metal implants may lead to a false-positive rate of 31% when using the alpha-defensin test [105]. Nonetheless, alpha-defensin provides consistent accuracy irrespective of the infecting organism species or premature antibiotic administration [92, 106]. While the impres-

sive performance of the laboratory-based alpha-defensin immunoassay has led to its incorporation into 2018 update of the ICM criteria for the diagnosis of PJI [15], it is important to acknowledge that relatively fewer studies have evaluated this novel biomarker compared to the wealth of literature on routine diagnostic tests [74], highlighting the need for additional investigations in more objective settings. Moreover, given the high costs of alpha-defensin (approximately USD 760) relative to leukocyte esterase (approximately 0.17 USD), future research should aim to evaluate the cost-effectiveness of this novel biomarker for the diagnosis of PJI.

6.3.4 Synovial CRP

As serum CRP is a marker of systemic inflammation, it has a limited specificity for localized infections and the potential for false-negatives in the presence of low-virulence biofilm infections [107]. Consequently, several authors have postulated that synovial fluid CRP could be a more specific and sensitive diagnostic test for PJI [108]. CRP functions by activating the complement system to eradicate foreign or dying cells. As a result, its concentration is often higher at the focus of inflammation, as shown in a previous study using synovial CRP to differentiate inflammatory from non-inflammatory monoarthritis [109]. In the initial study proposing its use for the diagnosis of PJI, Parvizi et al. found a sensitivity of 85% and specificity of 95% when using an automated turbidimetric method to analyze synovial CRP [110]. Similarly, Plate et al. studied 171 hip and knee PJIs and found an optimal synovial CRP threshold of 2.9 mg/L, with a sensitivity of 91% and specificity of 82% [111]. Although one study questioned the utility of this synovial fluid biomarker [112], the accuracy of synovial CRP was confirmed in a recent meta-analysis of seven studies, which yielded a pooled sensitivity and specificity of 92% (95% CI, 86–96%) and 90% (95% CI, 87–93%), respectively [113]. Furthermore, the combined measurement of CRP with other synovial fluid biomarkers such as synovial fluid WBC count [114], alpha-defensin

[115], and IL-6 [116] has been shown to greatly improve its diagnostic accuracy, with one study even demonstrating a sensitivity of 97% and specificity of 100% [97]. As with other biomarkers, synovial CRP levels are highly dependent on the causative microorganism, as higher false-negative rates may be observed in the context of less-virulent pathogens [117].

6.3.5 Novel Synovial Fluid Markers

The introduction of alpha-defensin into uniformly accepted diagnostic criteria for PJI has spurred the investigation and development of novel synovial fluid biomarkers in recent years [15, 118]. While some markers such as synovial fluid PCT have failed to demonstrate accuracy in the diagnosis of PJI [118], other markers including synovial fluid calprotectin [119], D-lactate [120], adenosine deaminase [114], and CD64 index (granulocyte:lymphocyte ratio of CD64 expression) [121] have shown promising results. Calprotectin is a protein component of the cytoplasmic membrane in neutrophils that is released following neutrophil activation [122]. While traditionally used in the diagnosis of inflammatory bowel disease, recent studies have shown that this marker can be analyzed in synovial fluid to monitor treatment in inflammatory arthritis [123]. A recent pilot study found that synovial calprotectin had a 89% sensitivity and 90% specificity for diagnosing PJI in a cohort of 61 patients (19 PJIs and 42 aseptic revisions) [119]. In the same vein, synovial D-lactate has been suggested to be an accurate biomarker for PJI diagnosis [124]. As human cells possess L-lactate dehydrogenase and can only produce the L-rotatory isomer of lactate [125], D-lactate has been identified as a bacteria-specific metabolite that can be quantified in synovial fluid and used as a specific marker for joint infections [126]. In a prospective study of 71 PJIs diagnosed based on MSIS criteria, Karbysheva et al. found that synovial D-lactate had a sensitivity and specificity of 94% and 78%, respectively [120]. Coupled with practical advantages such as a rapid turnaround duration and low costs of performing this

test, the authors concluded that this biomarker could be a useful point-of-care screening tool for the diagnosis of PJI. Despite these encouraging results, current literature on these novel biomarkers are limited and require validation in future studies before their incorporation into diagnostic algorithms.

6.4 Frozen Section Histopathology

Periprosthetic tissue can be sent for histological evaluation to support the diagnosis of PJI. Specifically, pathologists can examine frozen tissue for neutrophilic infiltration that is suggestive of acute inflammation. One advantage of this diagnostic tool is the relatively short time needed to obtain the results from frozen section analysis, which can provide valuable information for the surgeon and guide intraoperative decision-making. Additionally, histological analysis is unlikely to be influenced by the administration of antibiotics preoperatively, which may be necessary in the rare cases that PJI is complicated by sepsis. On the contrary, technical expertise is needed to utilize this diagnostic tool reliably, as the result is highly operator-dependent and can vary based on the experience or technique of the pathologist reviewing the sample. Moreover, it has been suggested that less-virulent organisms including *C. acnes* and coagulase-negative staphylococci may not elicit a robust neutrophilic inflammatory response, thus increasing the risk of false-negative results in such cases [127]. In order to maximize the diagnostic yield of this test and reduce sampling error, it is important that surgeons sample the periprosthetic tissues that appear infected based on gross inspection intraoperatively. Traditional sites for periprosthetic tissue sampling include the joint pseudocapsule and periprosthetic interface membrane between the implant and adjacent bone. Based on the 2018 ICM definition of PJI, for frozen section histology to be positive, greater than five neutrophils per high-powered field must be identified in at least five separate microscopic fields under $\times 400$ magnification [15].

The diagnostic accuracy of frozen section histopathology was reviewed in a previous meta-analysis of 26 studies involving 3269 patients (796 culture-positive PJIs) [128]. When considering a diagnostic threshold of 5 PMNs per high-powered field, the authors reported a positive likelihood ratio of 10.3 (95% CI 6.3–16.6) and negative likelihood ratio of 0.24 (95% CI, 0.14–0.39), suggesting that intraoperative frozen sections are more useful for confirming the diagnosis of PJI but moderately accurate excluding this diagnosis due to low sensitivity. This was confirmed in a subsequent study utilizing the MSIS criteria as a reference standard for PJI, which reported a sensitivity of 74% and specificity of 94% based on 200 samples sent for frozen section [129]. As frozen section histology appears to be more reliable for ruling in compared to ruling out a diagnosis of PJI, surgeons should consider limiting the use of frozen section histology to patients categorized into the “inconclusive” group preoperatively based on the 2018 ICM criteria. Given the high costs and increased procedural time associated with collecting samples, processing them in the laboratory and performing histological examinations, this more conservative approach could lead to substantial cost savings for the patient.

6.5 Microorganism Identification

The second goal in the diagnosis of PJI consists of identifying the causative organism(s) and determining its antimicrobial susceptibility. While multiple synovial fluid biomarkers have demonstrated superior sensitivity (alpha-defensin, leukocyte esterase, synovial CRP, WBC count, and PMN%) compared to that of culture-based tests [74], synovial fluid and/or periprosthetic tissue cultures still play a major role in any diagnostic algorithm as it allows clinicians to identify the infecting microorganism and administer targeted antibiotic therapy, thereby maximizing the chance of treatment success [19]. Microbial identification also provides valuable prognostic information for patients and guides perioperative counseling [16]. According to the

2018 ICM definition of PJI, the isolation of the same pathogen from two separate tissue or fluid samples is diagnostic of PJI [15]. However, while multiple clinical guidelines on the appropriate surgical and laboratory techniques to maximize culture yield have been published [130], an estimated 7–12% of patients still have negative cultures despite clear clinical evidence of infection such as a draining sinus or a high synovial fluid WBC count [131–134], thus creating a diagnostic conundrum known as culture-negative PJI.

6.5.1 Synovial Fluid Cultures

Synovial fluid culture is an invaluable diagnostic tool as it offers surgeons the opportunity to identify the infecting microorganism(s) and determine its susceptibility prior to surgery. This knowledge can help to guide treatment decisions, especially in regard to the type of antibiotics to administer perioperatively and mix into the antibiotic-loaded polymethylmethacrylate (PMMA) spacer. Following joint aspiration, synovial fluid should be transported to the microbiology laboratory and inoculated onto solid or liquid media as soon as possible, as long transportation times can lead to higher false-negative rates [135]. If this is not feasible, aspirated fluid can also be inoculated into blood culture bottles in the procedure suite to decrease the risk of contamination and improve pathogen recovery while awaiting sample processing [135]. Although cultures are part of major and minor criteria in the recent ICM definition of PJI [15], it must be acknowledged that preoperative aspiration culture only has a moderate-to-high sensitivity for diagnosing PJI. In a meta-analysis of 34 studies with a total of 3332 patients, Qu et al. reported a pooled sensitivity and specificity of 72% (95% CI, 65–78%) and 95% (95% CI, 93–97%), respectively, with subgroup analyses showing a trend toward poorer diagnostic accuracy for hip aspirations compared to knee aspirations (sensitivity 70% vs. 78%; specificity, 94% vs. 96%) [136]. Similarly, the meta-analysis by Carli et al. concluded that this test had a poorer fitted HSROC curve and

lower pooled sensitivity compared with other synovial fluid biomarkers [74]. Notwithstanding, the potential to identify the causative pathogen preoperatively should not be discounted, and synovial fluid cultures should remain a part of the workup for any patient with a suspected prosthetic knee infection.

6.5.2 Intraoperative Tissue Cultures

Isolation of the same pathogen from two separate cultures is considered to be diagnostic for PJI based on the 2018 ICM definition, whereas a single positive culture may be considered a contaminant and should be reconciled with other minor criteria [15]. However, it is important to note that cultures not only help to support or confirm the diagnosis of PJI, but also provide guidance in antimicrobial selection. Although cultures have traditionally been used as the gold standard reference for assessing the accuracy of novel diagnostic tests, it is now recognized that up to 30% of PJI cases have negative cultures [24, 133, 137], owing to their lower sensitivity and inability to rule out PJI [138]. Conversely, as it is well established that a varying degree of clinically relevant PJI may be detected in some cases of presumed aseptic loosening based on positive intraoperative cultures [139], it is imperative that intraoperative cultures be taken regardless of the preoperative diagnosis [138]. To maximize culture yield, recommendations from the 2018 ICM state that at least three intraoperative samples should be sent for culture as this produced the highest negative predictive value to rule out infection without reducing the positive predictive value [138]. Samples should be taken from the areas of infection based on gross inspection, which should include synovial, femoral, and tibial tissue [138]. These should then be incubated for 5–14 days. For suspected culture-negative PJI cases or cases in which the organism is suspected to be less-virulent or fastidious (e.g. *Cutibacterium* species), a longer incubation time should be used [133]. Swab cultures should not be taken due to their low diagnostic yield [140].

6.5.3 Sonication Fluid Cultures

Current evidence suggests that low-intensity sonication of explanted prostheses is an effective means to disrupt biofilm on the prosthetic surface to increase the sensitivity of microbiological isolation compared to traditional sampling of synovial fluid or periprosthetic tissues [141–144]. Sonication may also improve culture yield by dislodging sessile organisms on explanted prostheses [145, 146]. Cultures of sonication fluid have demonstrated an improved sensitivity (78–97%) in microorganism identification without compromising specificity (81–99%) [142, 145, 147–149]. Trampuz et al. studied 331 patients and found a sensitivity of 79% for sonication fluid cultures, which was significantly greater than that of tissue cultures (61%) [142]. Interestingly, these findings persisted even in the presence of antimicrobial therapy within 14 days prior to surgery (75% vs. 45%). Similarly, Rothenberg et al. reported a higher sensitivity for sonication fluid cultures in MSIS-confirmed PJIs compared to tissue cultures (97% vs. 57%) with no difference in specificity [149], while Janz et al. showed that these parameters could be improved to 100% by separating components into multiple sonication fluid cultures [150]. In a meta-analysis of 12 studies evaluating sonication fluid cultures, Zhai et al. found a pooled sensitivity and specificity of 80% (95% CI, 0.74–0.84) and 95% (95% CI, 0.90–0.98), respectively [151]. Despite these promising results, some authors have suggested that the accuracy of sonication fluid cultures may vary based on the sonication technique used [152] as well as timing of PJI [153]. False-positive results have also been observed and attributed to contamination during the sonication process [150]. To overcome this limitation, most authors have recommended a diagnostic threshold of at least five colony-forming units (CFUs) for sonication fluid cultures [142, 149, 151]. In view of the overwhelming evidence demonstrating improved pathogen isolation with the use of sonication fluid cultures relative to traditional synovial fluid or tissue cultures, current guidelines support the use of sonication in every patient suspected of having a PJI [138].

6.5.4 Culture-Negative Infections

The isolation of an organism from microbiological cultures is not always possible despite clinical evidence confirming the presence of PJI—a phenomenon commonly referred to as culture-negative periprosthetic joint infections (CN-PJI) [131]. False-negative results not only preclude the selection of targeted antimicrobial therapy and lead to lower rates of treatment success [154], but also result in unnecessary anxiety for patients who may challenge the diagnosis of PJI due to an inability to isolate a pathogen [133]. Furthermore, empirical treatment of CN-PJI usually entails administering broad-spectrum or multiple antibiotics to cover the most common microorganisms according to epidemiological surveys, which may be less effective and increases the risk of adverse reactions or systemic toxicity. The prevalence of CN-PJI has been noted to be as high as 30% [24, 133, 137]. Possible reasons for negative cultures have been proposed, such as infection by fastidious pathogens, biofilm encapsulation, uncommon organisms (e.g. fungi or mycobacteria) that do not replicate on routine culture media, inadequate sampling, or transportation, as well as insufficient resuscitation in the laboratory [16, 130, 135, 155]. Nonetheless, the most important cause of failure to isolate an organism is the administration of antibiotics before obtaining samples from the infected joint [131, 133, 155, 156].

6.5.5 Antibiotics and Culture Yield

Sub-therapeutic or mistargeted antimicrobial treatment has been shown to induce a viable but non-culturable (VBNC) physiological state in many pathogens [157–161], rendering the results of these cultures falsely negative [131, 133]. While most pathogens are generally unable to cause infection in a VBNC state, these bacteria still retain their virulence and can cause infection after being resuscitated [162], likely accounting for the phenomenon of CN-PJI. Current evidence cautions against the use of antibiotics in the period leading up to revision arthroplasty [82,

131, 142, 156]. Trampuz et al. demonstrated that any administration of antibiotics in the 2 weeks before obtaining intraarticular cultures adversely influenced the sensitivity of cultures and was associated with a higher false-negative rate (55% vs. 23%) [142]. In another case-control study of 60 patients, Barbari et al. found that 53% of patients who had CN-PJI received antimicrobial therapy within 3 months before the diagnosis and 23% received the antimicrobial agent up to the time samples were taken from the infected joint [131]. Similarly, Malekzadeh et al. found that patients with CN-PJI were 4 times more likely to have received antimicrobial therapy in the preceding 3 months before diagnosis [156], while Shahi et al. reported that patients with antibiotic use before aspiration had a higher rate of CN-PJI compared to those without any antibiotic history [82]. Given these considerations, clinical practice guidelines from the AAOS have recommended against preemptive treatment before a thorough evaluation for PJI, advising clinicians to withhold antibiotic therapy for at least 2 weeks before intraoperative specimen collection to improve culture yield [163]. However, whether these recommendations can be applied uniformly to all suspected cases of PJI remains unknown. In particular, several authors have proposed that an even longer period without antimicrobial exposure may be required to culture certain fastidious organisms [142, 164–166]. Future research is needed to refine the present guidelines with regard to the effect of different antimicrobial agents on the culture yield of differing organisms, as well as to define the optimal antibiotic-free period before obtaining samples in patients with suspected PJI.

It is important to distinguish between therapeutic antibiotics (which often requires a prolonged course of treatment) and prophylactic antibiotics (which often comprises a single dose administered perioperatively [167]). While the abovementioned studies have demonstrated that antibiotic administration prior to identifying the causative pathogen increases the risk of false-negative cultures [156], the need to withhold pre-incision prophylactic antibiotics remains a controversial issue in orthopedic surgery [168–174]. Prophylactic antibiotics

were traditionally believed to interfere with culture yields from intraoperative samples, leading some investigators to advocate against their use in the context of revision arthroplasty for suspected PJI [164, 175, 176]. Although this practice appears logical, withholding prophylactic antibiotics may increase the risk of surgical site infection or systemic dissemination perioperatively. Moreover, recent evidence has largely refuted this belief [168–174]. In particular, two randomized controlled trials have demonstrated identical rates of positive intraoperative cultures [169] and concordant cultures [170] in patients who did or did not receive prophylactic antibiotics before incision. A large cohort study of 425 revision TKAs also reported no difference in the percentage of positive cultures (26% vs. 27%) as well as the species of bacteria cultured [174]. Given the large body of evidence suggesting that the practice of withholding prophylactic antibiotics to maximize culture yield may not be as critical as previously thought, the 2018 ICM recommended that perioperative antibiotic administration for revision TJA should not be routinely withheld, but should instead be guided by the degree of clinical suspicion for PJI and whether or not a causative organism has been isolated before surgery [130].

6.5.6 Molecular Tests

The overreliance on cultures as the gold standard for microorganism identification has led to the conundrum of CN-PJI. Molecular techniques to detect bacterial DNA present a unique opportunity to improve the accuracy of diagnosis for PJI, particularly in the setting of negative cultures [133]. Multiplex polymerase chain reaction (PCR)-based assays allow the detection of common microorganisms and their resistance genes, improving sensitivity and reducing the time to diagnosis compared with traditional cultures [177–179]. However, the requirement for specific primers often results in the failure to detect atypical or less common pathogens as well as resistance mechanisms [180, 181]. Another molecular technique currently available is 16S rRNA gene sequencing [178]. Unlike PCR-based assays, this

method allows the detection of a wider variety of bacterial species, prompting some authors to suggest that 16S rRNA sequencing may have a higher sensitivity compared to bacterial cultures and PCR-based techniques [178, 182, 183]. Primers used in this technique are specific for highly conserved sequences that are found in almost all bacteria, as well as variable regions in between them, thereby allowing the identification of a broad range of bacteria. However, major limitations of this method include the inability to detect antimicrobial resistance genes and polymicrobial infections, which can only be determined using high-throughput sequencing methods rather than traditional capillary-based ones [184]. More recently, metagenomic next generation sequencing (mNGS) was introduced to overcome the shortcomings of previous molecular tests. This high-throughput sequencing technique enables the detection of complete bacterial genomes, including unculturable, unsuspected, and non-viable organisms in the sample [185–188]. Resistance genes can also be simultaneously detected using this technique [187]. Direct sequencing of specimens improves the diagnostic yield compared to traditional cultures [186], as recent studies have shown that mNGS was able to detect new organisms in 16–44% of CN-PJI cases and 4–67% of culture-positive cases [185–189].

In addition to improved diagnostic accuracy, other advantages of molecular testing have proposed. Current evidence suggests that molecular methods for pathogen identification are unaffected by prior antibiotic administration [181, 190], overcoming the limitations of traditional cultures. This advantage may be clinically useful in the management of patients undergoing two-stage exchange arthroplasty. As it is often difficult to ascertain whether infection has been eradicated following a course of 4–6 weeks of systemic antibiotics in the interim stage, current practice often involves rechecking inflammatory markers such as ESR and CRP, although this has been shown to correlate poorly with the likelihood of residual infection at the time of reimplantation [40, 41, 191, 192]. Alternatively, synovial fluid cultures may be taken after an “antibiotic holiday” of 2 weeks

prior to reimplantation to improve diagnostic yield. In such cases, molecular testing not only circumvents the need for an “antibiotic holiday,” but also provides more sensitive diagnostic information that can guide clinical decisions such as the appropriateness and timing of reimplantation [193]. The utility of molecular methods may further extend to patients on chronic suppressive antibiotic therapy, providing a reliable method for monitoring bacterial load as well as the development of antimicrobial resistance. However, it is important to note that while the ability to detect bacterial DNA even after cell death from antimicrobial therapy may seem advantageous in these situations, this is in fact a double-edged sword, as these techniques cannot differentiate between active vs. eradicated infections [194, 195]. Previous studies have demonstrated that DNA can also be isolated from non-viable bacteria in sterile joints, especially in patients with inflammatory arthritis [196, 197]. Consequently, the importance of clinical correlation and adjunctive tests to support the diagnosis of PJI cannot be further emphasized [15]. Currently, high costs and complex laboratory workflows are the main obstacles hindering the adoption of molecular testing. As these methods become more cost-efficient over time, their speed of detection as well as improved sensitivity especially in the setting of prior antibiotic administration will allow clinicians to initiate targeted antimicrobial therapy at an earlier time, potentially improving the treatment outcomes for PJI in the future.

6.6 Conclusion

Infection following knee arthroplasty is a rare but devastating complication that not only increases the risk of mortality and diminishes the quality of life of orthopedic patients [11, 12], but also poses a substantial economic burden to the healthcare system [13]. Due to the vast differences in the management of aseptic failure and PJI, obtaining an early and accurate diagnosis remains paramount [132]. Despite the extraordinary efforts by the orthopedic community, the

diagnosis of PJI still poses a formidable challenge to every surgeon. No single test can confirm or rule out the diagnosis, hence current diagnostic criteria are based on clinical findings as well as a combination of laboratory tests described in this chapter. Over the past decade, a plethora of novel serological and synovial fluid biomarkers have emerged as highly accurate tools for diagnosing PJI, some of which have been included in the latest 2018 definition of PJI [15]. Notwithstanding, surgeons should be cognizant of the challenging clinical scenarios and subpopulations that may alter the diagnostic performance of these laboratory tests, including patient comorbidities, timing of infection, pathogen virulence, and premature antibiotic use. Furthermore, one of the most difficult challenges in the diagnosis of PJI is the isolation of the causative microorganism. The limitations of traditional microbiological cultures have been highlighted repeatedly in orthopedic literature, culminating in a new diagnostic conundrum known as CN-PJI. To this end, molecular tests hold much promise in pathogen identification, maintaining their diagnostic accuracy in a variety of clinical situations. However, further research is necessary to translate this new technology into routine practice and validate its clinical utility in enhancing patient care, controlling healthcare costs and improving antimicrobial stewardship.

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Microbiological Diagnosis of Knee Prosthesis Infections

7

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7.1 Introduction

Microbiological analyses are one of the cornerstones of the management of knee prosthesis infections (KPIs) as the culture and isolation of the pathogen is a major criterion for their diagnosis [1, 2]. Then, bacterial identification and antimicrobial susceptibility testing are required to adapt and/or optimize antimicrobial treatment.

As bacteria responsible for acute and chronic infections can be different, microbiological analysis must be carried out in order to identify a wide panel of pathogens combining culture protocols adapted to slow growing bacteria, but also mycobacteria and fungi, and molecular

approaches. Indeed, while bacteria causing acute infections are usually virulent and easy to grow pathogens (*Staphylococcus aureus* and beta-hemolytic streptococci, *Enterobacteriaceae*), the bacteriological diagnosis of chronic infections can be much more challenging. Bacteria involved in chronic infections are more diverse including low-grade pathogens corresponding to bacteria belonging to commensal skin flora (e.g., coagulase negative staphylococci, corynebacteria, *P. acnes*) [3]. Identification of such bacteria can trigger difficulties of distinction between contamination and true infection. The formation of biofilm, the presence of metabolic variants, named *small colony variants* (SCVs), and the

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low microbial inoculum in chronic infections can also affect the sensitivity of microbiological diagnosis [4, 5]. Finally, some bacteria can be very difficult to grow or can only be identified with molecular biology techniques (*Mycoplasma* spp., *Tropheryma whipplei*).

7.2 Types of Samples

General rules have to be applied to improve the yield of microbiological diagnosis of KPIs:

- To prevent false negative samples, it is recommended to respect a minimum of 15 days without any antibiotherapy (except in case of sepsis) before the samples, 1 month if rifampicin, fluoroquinolones, or cyclins.
- To prevent false positive samples, it is recommended to respect a strict surgical asepsis.

7.2.1 Preoperative Samples

When abscesses or fluid collections in the joint or in the soft tissue are present, aspiration can be performed, with ultrasound guidance if needed. Vials for aerobic and anaerobic blood cultures should be inoculated with aspirated fluid immediately to increase the sensitivity but an aliquot must always be kept for microscopic examinations, conventional cultures, and molecular analyses [6]. Preoperative aspiration culture has a moderate sensitivity but a very high specificity [7]. Ultrasound guided percutaneous biopsies can also be taken when aspiration is not feasible. Superficial samples from wounds or fistulae using a swab must be avoided since they are most often contaminated by cutaneous flora and their results correlate poorly with those obtained with deep samples, with the only exception of *S. aureus* [8].

7.2.2 Peri-operative Samples

Multiple samples must be collected, from different anatomical sites and, if possible, from sites which are macroscopically pathological, due to

the heterogeneous distribution of pathogenic bacteria inside the infected site and the possible presence of commensal bacteria. These samples can be fluid (e.g., pus, articular fluid), solid samples (e.g., granulomatous tissue, bone tissue, interposition tissue, and any suspicious tissue), or osteosynthesis material (e.g., screws, cement, rods). International guidelines recommend that ideally five or six periprosthetic samples be obtained, but recent studies suggest that this number could be reduced to four samples [2, 9, 10]. Culture positivity rates can vary depending on the sample type including joint fluid and tissue samples being more frequently positive than bone samples [10]. A low number (<3) of samples may lead to a lack of sensitivity of culture or a misinterpretation of a single positive culture and a higher number to an increased probability of contamination without evidence of improved sensitivity of the examination. It is recommended to use new sterile instruments for each tissue specimen to avoid cross-contamination. Sampling with a swab must be avoided as the culture sensitivity is low, compared with tissue samples [11].

In case of implant-associated infections, prostheses and other devices can also be sent to the laboratory combined with culture of periprosthetic tissue samples to increase the sensitivity in cases with strong suspicion of KPI without microbiological documentation or prior antibiotic administration, but must be processed specifically, using mechanical methods such as sonication to dislodge bacteria from the biofilm formed on the material [1, 12, 13]. Particular attention must be paid during the collection of prosthetic/biomaterial components to avoid contamination which should be transferred into a non-perforable, sterile, leak-proof container of suitable size. Results from sonicate fluid must be interpreted with caution and remains a subject on debate.

In addition, the taking of paired blood cultures routinely in case of fever or sepsis associated with KPI, of arthritis, and presence of a secondary infected site must be encouraged and never forgotten. If positive, these blood cultures can be useful to guide the diagnosis if culture of periprosthetic samples remains negative or confirm the pathogenic nature of bacteria isolated in such samples.

7.3 Bacteriological Analysis

Transport is an important pre-analysis step for these types of samples and should be organized involving the surgical, laboratory, and logistics departments. The different samples must be transferred at room temperature as quickly as possible, ideally within 2 h. If this deadline cannot be met, transport medium to keep fragile bacteria and anaerobes alive must be used. It is essential to mention for the clinical lab the date and time of sampling, the anatomical site, and clinical information (e.g., antibiotherapy, use of a prosthesis).

It is essential to particularly pay attention to the risk of contamination of these samples in the lab as commensal bacteria can be involved in KPIs. They must be handled in BSC-2 by a technician wearing a disposable overall and gloves (changed regularly) and using sterile equipment.

Solid samples (bone fragments or tissue) must be crushed for example using sterile glass beads before examination, in order to homogenate the specimens and to release bacteria [14]. Molecular grade water should be preferred to culture broth as a diluent so that further molecular analyses (particularly broad range PCR) are not compromised. Sonication of prosthetic devices has been proposed especially to release bacteria from biofilm [12, 13]. The use of a solution of dithiothreitol has been proposed as an alternative to bead mill processing of the samples [15].

The part of the sample which is not inoculated must be preserved by freezing ($-20\text{ }^{\circ}\text{C}$) until definitive diagnosis is reached in case additional tests are needed (e.g., testing for mycobacterium, fungi, molecular biology).

7.3.1 Microscopic Examination

Microscopic examination includes:

- Direct microscopic examination for synovial fluids (e.g., on Malassez cell) to search for micro-crystals (for differential diagnosis of chondrocalcinosis and acute articular gout) and quantify leukocytes, followed by May-

Grunwald-Giemsa staining for quantification of polymorphonuclear neutrophils (PMNs). A percentage of PMNs $>65\%$ or a leukocyte count of $>1.7 \times 10^3/\mu\text{L}$ is indicative of KPI [16].

- Gram's staining to test for bacteria. It has a high specificity but a very low sensitivity [17]. It is useful mainly in acute infections.

7.3.2 Culture

Culture methods include the use of solid agar plates as well as broth media for enrichment incubated at approximately $35 \pm 2\text{ }^{\circ}\text{C}$ in various atmospheres.

Given the bacterial epidemiology of the KPI, samples are classically inoculated at least onto:

- A blood agar incubated aerobically with early reading on D1–D2 and late on D5,
- A supplemented chocolate agar incubated in a 5% CO_2 atmosphere with early reading on D1–D2 and late reading on D5,
- A blood agar or Schaedler agar incubated in anaerobic conditions with early reading at D3–D5 and late reading D14,
- An enriched anaerobic liquid medium such as Schaedler or Rosenow's broth with regular reading up to D14 if necessary, in order to increase the culture sensitivity, notably for the culture of slow growing anaerobic bacteria such as *Cutibacterium acnes* [18].

Inoculation of periprosthetic tissue samples after being crushed directly into blood culture bottles has been reported to increase sensitivity of culture, reduce time to culture positivity, and allows to reduce costs associated with the use of multiple plates and broths used for multiple samples [9, 10].

Reading of plates must be attentive to the examination for different appearances of colonies, notably micro-colonies which could signal the presence of metabolic variants (SCV). An early positive culture in a solid medium does not exclude continued readings and complete incubation to look for additional bacteria of

slower growth, polymicrobial infections representing 10–15% of infections. In order to limit the risks of false positives linked to contamination of boxes during their iterative openings at early readings, some authors recommend doubling the inoculation of the anaerobic agar plates and chocolate agar plates. In this case, one plate is used for early reading and the second plate is only opened for late reading. All reading procedures, re-inoculation, and handling of medium should be systematically performed in a BSC-2.

Identification and antibiotic susceptibility testing (according to EUCAST/CLSI recommendations) must be performed on all isolates and different colony morphotypes (such as SCV) of the same bacterial species because they may show different antimicrobial susceptibility profiles.

Additional cultures for fungi and mycobacteria should be performed depending on the clinical context on specific request.

7.4 Molecular Biology

If culture remains the current gold-standard for diagnosis of KPIs, the pathogen is not identified in up to 40% of cases [19]. To face this situation and overcome these difficulties, molecular assays have been developed during the last decades. Compared to culture, PCR is theoretically more sensitive, faster, and not as affected by antibiotic treatment. Broad range 16S rRNA PCR or specific PCR targeting frequent KPI pathogens can be used. The former has the advantage to detect all bacteria but requires sequencing of the PCR product for bacterial identification, may be non-specific (due to background bacterial DNA in samples/reagents), and polymicrobial infection may be missed by sequencing. Reports of sensitivities and specificities of these PCR vary depending on the study and the samples types (synovial fluid, tissue samples, sonication fluids) [20–24]. Published studies mainly report the evaluation of broad range PCR assays but more recently, multiplex specific PCR have been developed but some of them do not include primers for the common PJI targets such as coagulase nega-

tive staphylococci and *C. acnes* [21, 22]. On the whole, reported data do not support superiority of these molecular assays compared with traditional culture methods except in case of treatment with antibiotics at the time of surgery or infection by fastidious bacteria [20–24]. Molecular biology methods complement conventional cultures without replacing them and should be used in case of strong suspicion of infection with negative cultures but not routinely because of their high cost.

More recently, the use of next generation sequencing in the diagnosis of KPIs has been evaluated as an alternative to PCR assays, not suited to detect polymicrobial infections in the case of broad range PCR or uncommon pathogens not included in panels of specific PCR assays [25–28]. Metagenomic sequencing offers the possibility of directly detecting all nucleic acids from a clinical sample, giving access in theory to more information than just bacterial identification by sequencing of the whole bacterial genome necessary for antimicrobial resistance prediction. However, samples obtained in the context of KPIs are challenging sample types as they couple low bacterial load with high levels of contaminating human cells affecting the sensitivity [25]. More studies exploring protocols allowing bacterial DNA enrichment are required to assess the performance of these techniques for bacterial identification and antimicrobial resistance prediction to justify their expensive cost compared to conventional culture or PCR assays [29, 30].

7.5 Interpretation

According to different international guidelines, a prosthesis infection is defined by the presence of one major diagnosis criteria including two positive periprosthetic cultures with phenotypically identical organisms or by a combination of three minor criteria including a single positive culture [1, 2, 31]. However, clinically, PKI may be present without meeting these criteria. Interpretation of the bacteriological results in the context of KPI is thus often complex. It must take into account the clinical context, as the bacterial

inoculum is lower in chronic than acute infections and in case of previous antimicrobial therapy, affecting performances of microbial diagnosis. Bacterial species identified should also be interpreted regarding the number of positive samples and the number of positive culture media. Growth of a virulent microorganism (e.g., *S. aureus*, haemolytic streptococci, *Enterobacteriaceae*, etc.) in a single specimen may also represent PKI while one culture that yielding a bacteria part of the normal skin flora may be indicative of a contamination (e.g., coagulase negative staphylococci, *C. acnes*) and should be evaluated in the context of other available evidence [2].

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Molecular Analysis and Histological Evaluation

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8.1 Introduction

Periprosthetic joint infection (PJI) is one of the most serious and debilitating complications suffered by patients after total joint arthroplasty. The reported incidence of PJI for total knee arthroplasty has ranged from 0.8 to 1.9% and for total hip arthroplasty from 0.3 to 1.7% [1]. When faced with a periprosthetic joint infection, early and accurate diagnosis continues to be critical to deliver successful targeted treatment to the patient. Currently, under clinical suspicion, a combination of radiological, serological, synovial, microbiological, and histological investigation is performed to assist in diagnosis and isolate the offending organism(s) [2]. Although considerable advances have been made in the diagnostic armamentarium available to the clinician, microbiological culture continues to be the gold standard to identify pathogens and their antimicrobial sensitivities.

Although valuable in a considerable subset of cases, problems remain with the use of culture in the isolation of pathogenic organisms in PJI. In

spite of strategies to optimize yield, such as an increasing number of samples, longer culture incubation period, and implant sonication, the sensitivity of culture is only 39–70% [3–6]. In addition, cultures yield a negative result in 7–50% of PJI cases [3, 7, 8]. These culture-negative PJI cases prevent the selection of targeted antimicrobial therapy for the patient and result in poorer outcomes. Studies suggest that patients with a culture-negative PJI have a 4.5 times greater rate of re-operation than those with positive cultures [9].

Several factors have been implicated in the decreased yield of culture for certain PJI patients. Historically, it has been proven difficult to isolate sessile organisms that reside within a biofilm in comparison to their traditional planktonic state. Prior antibiotic therapy can also decrease the yield of culture, with 53% of culture-negative PJI having had preceding antibiotic therapy in some studies [10]. Both bacteria in biofilm and those exposed to antibiotics have been shown to enter a viable but non-culturable state (VBNC), characterized by a loss of culturability on routine agar medium [11]. Certain fastidious organisms, including *Cutibacterium acnes*, *Brucella*, and *C. burnetii* can be difficult to isolate with standard culture methods even when not in a VBNC state and require specialized microbiological techniques to detect [12]. Finally, polymicrobial infections pose a unique challenge when

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using traditional culture methods, with a detection rate as low as 13–17% [13]. When culturing these infections, secondary bacteria can be hidden due to overgrowth by a rapidly growing primary species.

With these limitations of traditional microbiological culture in mind, there has been an increasing interest in alternative molecular diagnostic techniques to isolate pathological organisms in PJI, such as polymerase chain reaction (PCR), next-generation sequencing (NGS), and metagenomic sequencing. As these techniques gradually occupy an increased role in the diagnosis of PJI, it is important to understand the advantages and limitations of each technique and consider what their role should be within the diagnostic framework for PJI in the future.

8.2 Molecular Analysis

8.2.1 PCR

Techniques for molecular analysis have expanded rapidly in recent years and have increasing roles in the diagnosis of genetic disease, cancer, and infection. The simplest of these techniques is pathogen specific polymerase chain reaction (PCR), which uses a primer to detect a specific organism (e.g. *S. aureus*) or a group of closely related species (e.g. all staphylococcal species) (Fig. 8.1). This primer targets a known DNA sequence and amplifies this fragment, which can be from a particular gene or non-coding region of bacterial DNA. Originally, techniques such as gel electrophoresis were used for “end-point” detec-

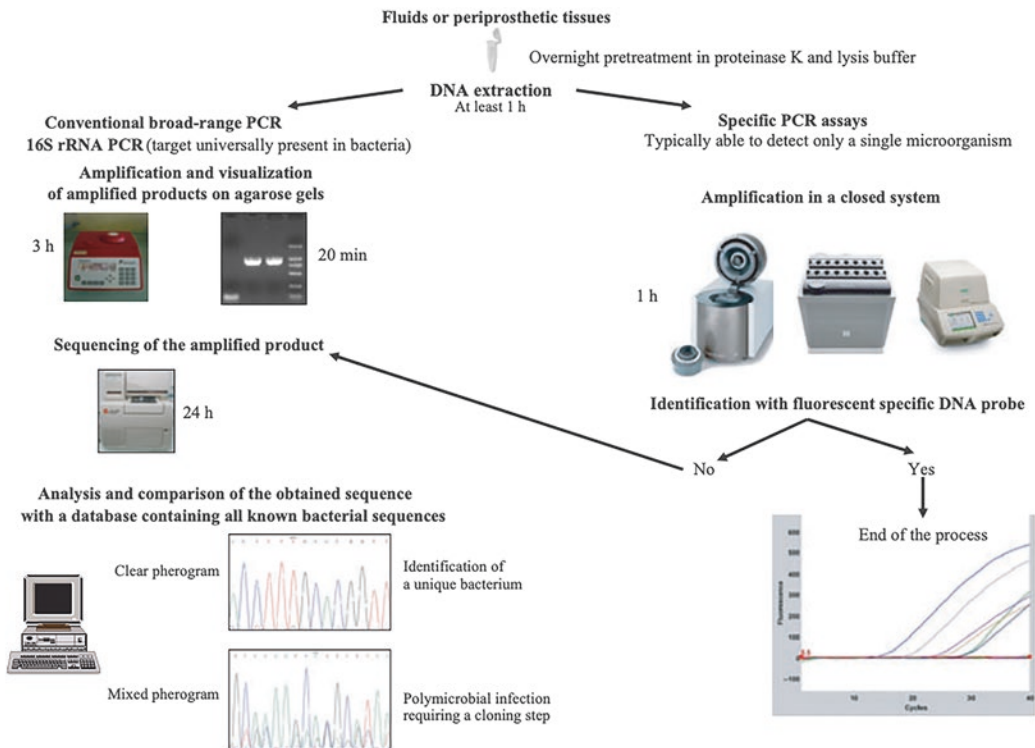


Fig. 8.1 Methodology of conventional broad-range PCR and specific PCR assays. (Reprinted from Lévy PY, Fenollar F. The role of molecular diagnostics in implant-

associated bone and joint infection. Clin Microbiol Infect. 2012 Sept;18:1168–1175, with permission from Elsevier)

tion (positive or negative) of this PCR product or “amplicon.” More recently, real-time PCR, where the PCR machine detects the amplicon in real time using fluorescent dyes, has been developed. This technique is both more sensitive than gel-based PCR and gives the user a quantitative result [14]. A primer can be designed that is unique for any organism and has the advantage of being extremely sensitive, potentially detecting even a single copy of the target DNA. Unfortunately, pathogen specific PCR has limited applicability in the diagnosis of PJI, due to its detection of only a single organism. Attempts to address this have been made with multiplex PCR assays, which use a group of specific primers to detect bacteria and fungi that are common in PJI. Unfortunately, these tests are labor intensive and may still overlook rare organisms. Studies have demonstrated that multiplex PCR does not outperform culture, with a sensitivity of 81% and a false positive rate of 88%, limiting its utility [15, 16].

Broad-range PCR provides the ability to find DNA from any bacteria. Most broad-range PCR techniques are based on the gene coding for the small subunit of the bacterial ribosome (16S rDNA) [14]. This gene contains highly conserved regions that are pan-bacterial, as well as more variable regions that are different among bacterial species. These highly conserved regions enable broad-range PCR assays to amplify DNA from any bacterial species, after which the specific identity of the bacterial species is subsequently determined by analyzing at least one variable region contained within the amplicon. Using traditional Sanger sequencing technology, direct analysis of the amplicon is only interpretable if a single sequence makes up greater than 70% of the total amplicon [14]. This makes interpretation difficult in the setting of a polymicrobial infection, which will result in mixed sequence data.

In addition to its inability to detect a polymicrobial infection, the main limitations of broad-range PCR relate to issues with contamination and sensitivity. As PCR can be an extremely sensitive assay, contamination from bacterial DNA introduced during the collection and handling of

the sample, as well as DNA present in PCR reagents can lead to false positive results. Development of techniques to remove extraneous DNA, such as UV irradiation or DNase treatment, have been partially successful in addressing this problem, but can actually reduce the activity of the *Taq* polymerase, thus considerably reducing the sensitivity of the PCR assay [17–19]. Reducing the number of PCR cycles can also avoid the effects of low-level reagent contamination, but results in a 100–1000 times lower sensitivity than pathogen specific PCR assays [14]. Newer methods, such as a PCR-based system with mass spectroscopic identification of pathogens (Ibis Biosciences T5000 biosensor system) have also been developed in an attempt to increase sensitivity. This technique appears to be less affected by prior antimicrobial therapy than traditional culture, with a sensitivity of 85.7% among subjects receiving antimicrobial therapy within 14–28 days of surgery [20]. A recent meta-analysis of all PCR studies found a pooled specificity of 94% and a pooled sensitivity of 76% [21].

8.2.2 NGS

In contrast to traditional Sanger sequencing, next-generation sequencing (NGS) parallelizes the sequencing process, so thousands of sequencing processes are able to occur in one reaction system concurrently [22]. This decreases both the time required and cost; NGS requires only 12 h to complete a workflow of whole genome sequencing, and multi-gene sequencing with NGS costs about the same as single-gene Sanger sequencing [23, 24]. In addition, fewer DNA or RNA samples are required [23]. In a recent meta-analysis, NGS sequencing by synthesis was demonstrated to have a similar specificity to Sanger sequencing at 96.3% vs. 96.7%, respectively [25].

Studies examining the efficacy of NGS for PJI demonstrate considerable promise. In a prospective study by Parvizi et al. examining 65 revision arthroplasties and 17 primary arthroplasties, NGS had a sensitivity of 90%, compared to culture with 60.7% [26]. NGS had a

88.2% concordance with culture and was able to detect a potential pathogen in 81.8% of culture-negative PJI. In 9 of the 11 culture-negative PJI cases, NGS detected a polymicrobial infection. Interestingly, NGS was positive in 35% of primary and 25% of aseptic revision cases, indicating a high false positive rate. In many of these cases, *C. acnes* was the predominant organism detected. Other organisms isolated were mostly microbiota, the significance of which is unclear in the context of our limited understanding of the joint microbiome. Further emerging data using NGS also indicates that a greater number of PJI cases may be polymicrobial at the DNA level than previously thought based on traditional culture. Data presented at the 2019 American Academy of Orthopedic Surgeons meeting suggests that 88.7% of PJI patients who subsequently failed during longitudinal follow-up with a new organism detected by culture had that infective organism isolated using NGS during the initial treatment resection arthroplasty [27].

A further evolution of NGS technology involves metagenomic shotgun sequencing. Instead of targeting a specific highly conserved region of interest, such as the 16S rDNA for bacteria or the internal transcribed spacer for fungi, shotgun metagenomics extracts and sequences of all nucleic acid in a sample both from the host and any microorganisms are present [28]. This data is then compared against comprehensive curated library databases containing all known pathogens [29, 30]. Using this “open read” technology, shotgun metagenomics is only limited by missing or incomplete taxonomic representation in databases, which may produce a false negative result. This technique can even detect organisms that are transcriptionally active (metatranscriptome), shedding light on potential antimicrobial resistance [31]. A study using shotgun metagenomic sequencing by Thoendel et al. was able to identify known pathogens in 94.8% of culture-positive PJI and new pathogens in 43.9% of culture-negative PJI. The rate of false positive detection of microorganisms from uninfected aseptic failure cases was in this study was 3.6%, and the authors noted that the presence of human

and contaminant microbial DNA continues to be a challenge when using this technique [28].

While it seems likely that NGS holds promise and will see an increasing role in the diagnosis of PJI in the future, there are important issues that remain to be determined regarding its clinical utility. Currently, NGS is a costly technology that requires highly specialized equipment, trained technicians, and bioinformatics expertise that is currently only available in a few institutions [27]. A recent cost analysis comparing traditional culture with NGS found NGS to be cost-effective when the pre-test probability of PJI is greater than 45.5% [32]. This reinforces the concept that NGS does not provide a definitive answer as to whether a patient has a PJI, but adds data that must be interpreted along with the clinical picture and other laboratory investigations that help determine the pre-test probability of a PJI. Further study is also required to determine the significance of the DNA signal that is identified by NGS. Due to its high sensitivity, it is currently unclear whether the polymicrobial signals detected by NGS represent pathologic entities, contaminants, or organisms natively present as a part of the natural microbiome of the joint. Moving forward, it will be critical to understand the clinical relevance of these polymicrobial metagenomic signals, in particular due to the antimicrobial stewardship implications of the broad-spectrum treatment that would be required if these signals are deemed to be pathologic.

8.2.3 Histological Evaluation

In cases where preoperative testing is equivocal, the histological study of periprosthetic tissue has long been an important tool used to confirm or rule out a PJI. The presence of a polymorphonuclear neutrophil (PMN) infiltrate has traditionally been considered indicative of septic implant failure. This is reflected in the inclusion of positive histology as an intraoperative finding in the 2018 Musculoskeletal Infection Society’s (MSIS) evidence-based definition for diagnosing PJI [33]. Within, the MSIS considers positive histology “greater than 5 neutrophils per high-power

field in 5 high-power fields observed from histologic analysis of periprosthetic tissue at $\times 400$ magnification.” In a meta-analysis of intraoperative frozen section histopathology in the diagnosis of PJI, this criterion was shown to be an accurate predictor of culture-positive PJI, with a likelihood ratio of 10.25 [34].

The MSIS criteria built on the work of Feldman et al., who, recognizing the difficulty of identifying a PMN infiltrate using standard hematoxylin-eosin staining, established five criteria to ensure the adequate analysis of specimens [35]. First, the tissue has to be pink-tan, and not simply white scar, to avoid analysis of dense fibrous tissue or fibrin. Second, at least two specific tissue samples are used in order to minimize the risk of sampling error. Third, the five most cellular areas in the tissue sample are chosen for evaluation. Fourth, all PMNs have to have defined cytoplasmic borders to be included. Debris that appears to be the result of nuclear fragmentation is excluded, as it cannot be categorized definitively as a PMN. Fifth, five separate fields are evaluated under high power magnification (HPF).

When obtaining samples, good surgical technique is important to ensure accurate results. To limit false positive results, tissue should be obtained using sharp dissection rather than cautery [36, 37]. Furthermore, the best sample to obtain for histological study is the periprosthetic membrane. A study by Bori et al. demonstrated that the proportion of patients with PJI that had a positive interface membrane was significantly higher than those with a positive pseudocapsule (83% vs. 42%, $p = 0.04$) [38]. This may be due either to the fact that fibrosis in the pseudocapsule hinders neutrophil infiltration, or that the largest bacterial biofilm is found at the implant–bone interface.

There are special circumstances in which the traditionally accepted histological cutoff of five PMNs per HPF in five HPFs may lead to inaccurate conclusions, either increasing the rate of false negative or false positive results. One such patient population are those who have been previously implanted with a cement spacer and have returned for reimplantation with a definitive prosthesis. Studies examining these patients have

concluded that histology has a low sensitivity, with one study showing histology was positive in only two of the seven patients who had positive cultures at the time of reimplantation and another showing a sensitivity of 25% (one of the four patients) [39, 40]. There are also two subsets of patients in which histology will produce a high false positive result: patients with underlying inflammatory disease and those receiving a prosthetic replacement in the setting of a periprosthetic fracture. A study by Kataoka et al. examined synovial tissue from 60 rheumatoid arthritis patients at the time of their total joint arthroplasty and found 16.6% (10 out of 60) had greater than five PMNs per HPF [41]. The authors postulated that there is a persistent neutrophil infiltration in the rheumatoid synovium due to the underlying active disease, and this common microscopic finding is not necessarily consistent with an infection. Another study by Muñoz-Mahamud et al., examining 11 patients undergoing arthroplasty due to periprosthetic fracture, found a 66.6% (four out of six cases) false positive rate using histology [42]. In these patients, it may be the case that a neutrophil infiltration can occur in the periprosthetic membrane secondary to inflammation caused by the fracture and injured surrounding vasculature and not solely due to an infectious process.

Although much of this work has been done with frozen section, as intraoperative histology is most useful to guide surgical decision making, it is important to note that the morphological identification of PMNs and their differentiation from other inflammatory components within periprosthetic tissue may be more difficult in frozen sections than in permanent paraffin sections [43]. Some authors, such as Stroh et al., report few differences between the results of frozen and paraffin sections, with a concordance of 97.7% (297 of 304 sections) and the difference not affecting the final outcome of any patient [44]. This is in contrast to a study by Tohtz et al., which found a 21.8% (14 of 64 cases) discrepancy between frozen and paraffin sections [37, 43, 44]. In 18.8% (12 patients) of cases, the frozen section diagnosis was unclear and permanent sections confirmed the diagnosis (8 patients had aseptic

loosening and 4 had septic loosening). In 3.2% (2 patients) of cases, the frozen section diagnosis was aseptic loosening and the permanent section diagnosis was septic loosening.

Even when examining paraffin sections, prosthetic wear particles and bone fragments, which are common in periprosthetic membranes, make the processing of tissue difficult and lead to artifacts or thick sections that complicate the identification of PMNs [45]. As a result, there is interest in developing more precise methods for the detection of PMNs, using molecular markers. In one study, by Morawietz et al., CD15 immunohistochemistry was used to identify PMNs, resulting in a sensitivity of 73% and a specificity of 95% in culture-positive PJI [46]. The authors

concluded that with their methods, only 23 PMNs in 10 HPFs are indicative of a PJI. Another study by Kashima et al. used chloroacetate esterase (CAE) enzyme histochemistry, which resulted in a sensitivity of 83% and a specificity of 96% in culture-positive PJI (Fig. 8.2) [43, 45, 46]. The authors noted that if the criterion for histological diagnosis of PJI was lowered to two PMNs per HPF, the sensitivity and specificity increased to 94% and 96%, respectively. In 17% (five out of 29) of the culture-positive PJI cases, the histology had between two and five PMNs per HPF, while in all culture-negative cases, there were fewer than two PMNs per HPF. The exception to this was two cases which were culture negative, but still met the MSIS criteria for PJI, both of

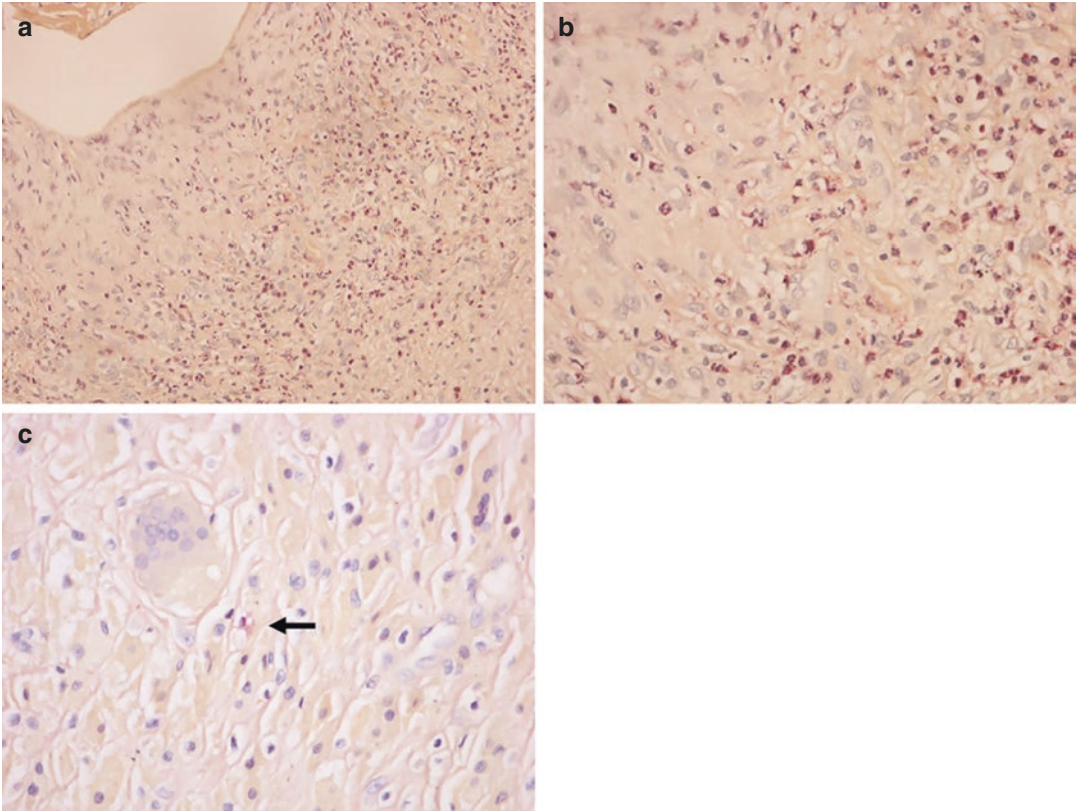


Fig. 8.2 CAE staining of paraffin sections of peri-implant tissues showing (a) low-power and (b) high-power views of a case of septic implant failure. There are numerous neutrophil polymorphs which show bright red cytoplasmic staining, whereas background foreign body macrophages are unstained. (c) Absence of CAE staining in a case of aseptic implant failure. Macrophages and a

macrophage polykaryon are unstained. The inflammatory infiltrate including a CAE-positive mast cell (arrow). (Reprinted from Kashima TG, Inagaki Y, Grammatopoulos G, Athanasou NA. Use of chloroacetate esterase staining for the histological diagnosis of prosthetic joint infection. *Virchows Arch.* 6 ed. 2015 Feb 17;466 (5):595–601, with permission from Springer)

which had between two and five PMNs per HPF. Both studies concluded that the MSIS criteria of 5 PMNs per HPF in 5 HPFs are too high and may miss some infections, in particular those caused by lower virulence organisms such as *C. acnes* [47].

Considerable work is now being done to help identify molecular biomarkers that are uniquely present in infected periprosthetic tissues. In the future, tissue biomarkers such as CD15 and certain toll-like receptors (TLR) may help us achieve a more reliable histological diagnosis of PJI [43].

8.3 Conclusions

Despite the progress in the study of PJI over the last 40 years, its diagnosis continues to be an elusive issue after total hip and knee arthroplasty. The current MSIS histological criteria for PJI remains the same as the one proposed by Mirra et al. in 1976 [43]. Increasing accuracy in the identification of bacteria and PMNs may assist clinicians in increasing the precision of their diagnosis by lowering currently established thresholds for PJI. The next step in this process may be examination not just on the cellular but also the molecular level.

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9.1 Introduction

Infection is one of the most debilitating complications following knee arthroplasty. A timely diagnosis of periprosthetic joint infection (PJI) is crucial for maintaining the joint function, avoid systemic sepsis, and limit the use of hospital and physician resources. Distinguishing PJI from aseptic mobilization is challenging but useful for the correct management of patients. Imaging can help in this purpose.

Imaging includes radiographs, resonance imaging (MR), computerized tomography (CT), ultrasound (US), and nuclear studies.

In addition to imaging, the AAOS study group recommended the erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) testing.

An X-ray could show periprosthetic radiolucency and the presence of intraarticular gas. MR imaging using metal artefact reduction techniques may detect osteolysis around the knee arthroplasty. CT scan may assist in distinguishing between septic and aseptic loosening. Nuclear techniques are used in patients with an uncertain diagnosis. They include labelled leukocyte imaging with bone or bone marrow scan, fluorodeoxyglucose-positron emission tomography

(FDG-PET), gallium, or labelled leukocyte imaging. Nowadays, nuclear medicine is valuable for joint arthroplasty assessment.

9.2 Ultrasound (US)

US is a useful imaging method to diagnose joint effusion and synovial hypertrophy, Baker's cysts, and synovitis. It is commonly used for guided arthrocentesis, aspiration of fluid around the joints, synovial and soft tissue biopsy, or abscess drainage [1, 2]. It can also be used for dynamic examination of the knee joint. Infected joint fluid often displays heterogeneous echoes with irregular hyperechoic synovial thickening and hypoechoic or non-echoic synovial fluid. Active synovial inflammation and the infection show strength or hyperaemia with colour-Doppler ultrasound. Advantages of US tests include accessibility, lack of ionizing radiation, no imaging contraindications, low cost, and a generally high tolerance for patients. However, US is operator dependent and has a constrained utility in the evaluation of osseous structures and surgical hardware.

9.3 Radiography

Radiography is usually the first imaging modality performed to assess a painful knee arthroplasty. It is used in the evaluation of total knee arthroplas-

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ties (TKAs) in both the immediate postoperative period and during the follow-up. The correct alignment of TKA, the type of implant used, the fixation interfaces, ligamentous laxity, and polyethylene wear are usually analysed through a weight-bearing X-ray. However, radiography lacks sensitivity and specificity in the assessment of PJI.

Radiographic signs of infected TKA, with low specificity, are swelling of soft tissue, periprosthetic lucency (Fig. 9.1), reflecting erosions (usually at the edges of the prosthesis) (Fig. 9.2), presence of intraarticular gas, and loosening of the elements.

Abnormalities like radiolucent lines are evaluated on AP views of the tibial component, on lateral views of the femoral component and skyline views of the patella. Changes in component position very reliably predict loosening. It is difficult to distinguish septic loosening from aseptic loosening. Septic loosening usually produces

extensive radiolucent zones and a periosteal reaction.

However, X-ray could show no abnormalities or nonspecific findings. These are periostitis, broad radiolucent lines, and focal osteolysis [3]. Li et al. showed that postoperative soft tissue gas on radiography is predictive of early PJI and is associated with a broader spectrum of microorganisms [1].

9.4 Resonance Imaging

MR imaging of the postoperative knee could be challenging. Knee arthroplasty produces significant susceptibility artefacts, which distort the appearance of the adjacent bony and soft tissue structures.

Cobalt, chrome, and molybdenum are usually associated with more extensive metal artefact compared with titanium or zirconium [2]. MR

Fig. 9.1 Periprosthetic lucency



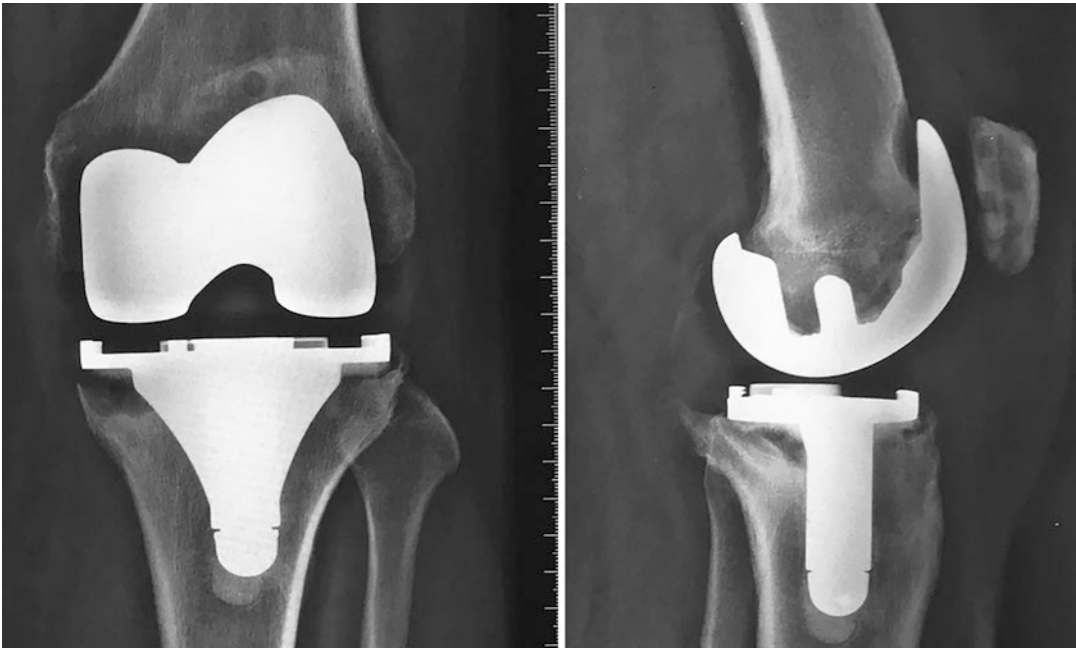


Fig. 9.2 Periprosthetic lucency

imaging using metal artefact reduction techniques, such as “section encoding for metal artefact correction (SEMAC)” and “multiacquisition variable-resonance image combination (MAVRIC)”, may detect osteolysis around the knee arthroplasty [4].

Several authors report lamellar and hyperintense synovitis during MR examination in patients with infected knee arthroplasty [5]. MR imaging can provide helpful information in patients with suspected periprosthetic joint infection. In acute infections, MR imaging can demonstrate the presence of a wound complication like a hematoma or abscess. The MR imaging demonstration of a sinus tract is diagnostic for joint infection.

9.5 Computerized Tomography Imaging

Computerized tomography (CT) scan may assist in distinguishing between septic and aseptic loosening [6]. Beam sclerosis artefacts limit the evaluation of adjacent bone and soft tissue structures,

so dual-energy CT can achieve metal suppression artefacts and is often used to detect radiographically hidden periosteal osteolysis. CT is better than MR imaging in isolating foreign bodies and gaseous lesions in the bone. CT imaging with enhanced contrast helps to outline soft tissue abscesses around the joints, synovitis, anatomical location, and the extent of soft tissue and bone infections [7]. MR and CT with intravenous contrast could detect soft tissue and intraosseous abscesses or active enhancing synovitis.

9.6 Single-Photon Emission Computed Tomography with Computerized Tomography (SPECT/CT)

Single-photon emission computed tomography (SPECT) is a **nuclear medicine tomographic** imaging technique using **gamma rays**. It may integrate and improves CT defects. Single-photon emission computed tomography with CT (SPECT/CT) provides information on the site of infection [8]. Several radiopharmaceuti-

cals have been used for SPECT/CT imaging. They include ^{99m}Tc -labelled diphosphonates, radiolabelled white blood cells with ^{111}In , ^{99m}Tc -hexamethylpropylene amine oxime, ^{67}Ga citrate, and other new tracers. Kim et al. [8] report that the sensitivity and specificity of SPECT/CT with ^{99m}Tc -hexamethylpropylene labelled leukocyte are both 93.3%. Graute et al. [9] underline that the SPECT/CT in addition to planar scintigraphy with ^{99m}Tc -labelled antigranulocyte antibodies for diagnosing and localizing low-grade joint infections has a sensitivity and specificity of 89% and 73%, respectively. Moreover, SPECT/CT provides an accurate anatomic localization of all positive areas, allowing a correct diagnosis of prosthesis versus soft tissue involvement.

9.7 Bone Scintigraphy/Three-Phase Bone Scan

Bone scintigraphy is a nuclear medicine procedure that uses small amounts of radioactive material to diagnose and assess the severity of a variety of bone diseases and conditions, including bone fractures, heterotopic ossifications, cancer, arthritis, aseptic loosening, and infection. It is the first method in nuclear medicine usually performed in cases of suspected PJI, and it is typically performed using ^{99m}Tc -methylene diphosphonate (^{99m}Tc -MDP), which accumulates on the surface of the mineral bone matrix.

Increased uptake on all three phases of the bone scan is indicative of infected TKA.

Bone scintigraphy can be positive as a standard postoperative appearance up to 2 years after TKA placement [10]. Moreover, osteolysis or synovitis from PE wear debris generally may simulate a PJI.

Bone scintigraphy is characterized by high sensitivity (95%) but low specificity for the diagnosis of PJI. Nevertheless, a negative three-phase Tc bone scan typically excludes infection [11–13].

9.8 Gallium-67 Citrate Scintigraphy

Gallium-67 (^{67}Ga) is a radionuclide. It was initially used to diagnose cancer [14]. ^{67}Ga accumulates in both septic and aseptic inflammation sites. The exact reason why ^{67}Ga accumulates in infection sites is still unclear. It is probably associated with the migration of lactoferrin, leukocytes, and siderophores produced by bacteria in inflammatory areas [12]. ^{67}Ga binds to lactoferrin and leukocytes in the plasma, and it migrates in inflammatory foci at high concentrations. However, Gallium-67 citrate scintigraphy has low specificity in the diagnosis of PJI, so it has been almost entirely replaced by other radiopharmaceuticals [10].

9.9 Indium-111 Leukocyte Scan

Indium-111 leukocyte scan is the most widely used for suspected PJI. Leukocyte labelling is usually performed with indium-111 (^{111}In) or ^{99m}Tc -hexamethylpropylene amine oxime (^{99m}Tc -HMPAO). Patient white blood cells are radiolabelled in vitro and then reinjected into the patients [10, 12]. Indium labelled leukocytes accumulate in areas of inflammation or infection or postoperative healing wounds. The evaluation of a negative indium scan is a strong predictor of the absence of infected TKA, but a positive indium scan has, unfortunately, a very limited value [12].

9.10 Dual-Isotope Imaging with In-111-Labelled Leukocytes and Tc-99m Sulphur Colloid Bone Marrow Scan

Dual-isotope imaging with In-111-labelled leukocytes and Tc-99m sulphur colloid bone marrow scan test is considered the most **vigorous** technique for detecting PJIs. However, it has some limitations. Patients leukocytes are labelled with In-111 and reinjected.

Simultaneously, Tc-99m sulphur colloid bone marrow scan (Tc-99m hexamethylpropyleneamineoxime [HMPAO]) is performed and delayed imaging after 24 h are acquired. The in vitro labelling process is challenging and requires the direct processing of blood products, which risks the contamination of the final product [15]. Sulphur colloid accumulates throughout the reticuloendothelial system, in the bone marrow, and in the liver and the spleen. In acute PJIs, chemotactic factors are secreted, so leukocytes migrate from the peripheral blood to the periprosthetic sites [12]. In chronic, long-standing infections, however, neutrophil recruitment is less evident. In PJIs an increased radiotracer uptake on both the three-phase bone scan and labelled leukocyte scan, in the same anatomical location, is found [11, 12]. A positive labelled leukocyte test has high sensitivity but low specificity. The conjunction of dual-isotope imaging with In-111-labelled leukocytes and Tc-99m sulphur colloid bone marrow scan test improves specificity and accuracy. Palestro et al. found that dual-isotope imaging with an In-111 and Tc-99m sulphur colloid bone marrow scan has 95% accuracy for the diagnosis of PJIs.

9.11 Labelled Leukocyte Scintigraphy with Antigranulocyte Antibody

99m Tc-antigranulocyte scintigraphy (AGS) is an alternative to autologous WBC scintigraphy in the detection of PJI. Leukocytes are labelled using monoclonal antibodies and antibody fragments against specific surface receptors on granulocytes [11]. Besilesomab and sulesomab are the monoclonal antibodies most commonly used. The sensitivity and specificity of besilesomab for PJI range from 67–91% to 57–75%, respectively [16]. AGS is a promising diagnostic tool, but, since the antibodies used are murine-derived, they could trigger a human anti-murine antibody (HAMA) response.

9.12 Positron Emission Tomography with 18F-Fluorodeoxyglucose (18F-FDG/PET)

Positron emission tomography (PET) is an imaging technique performed using a radioactive substance to visualize and measure metabolic processes in the body. PET with 18F-fluorodeoxyglucose (18F-FDG/PET) detects inflammatory cells with increased glucose uptake in infection sites [17]. This technique gives high-quality imaging with high spatial resolution, adequate capacity to determine anatomical location, and no need to manipulate blood products in vitro [18]. Fluorodeoxyglucose arrives in the cells via glucose transporters, where it is phosphorylated by hexokinase to 18F-2-18F FDG-6 phosphate. The uptake of fluorodeoxyglucose depends on the cellular metabolic rate, on the affinity, and on the number of glucose transporters, both of which are prevalent in the inflammatory cells. Scanning is performed 30–60 min after radiotracer injection.

Positron emission tomography with 18F-FDG can also be used to examine the potentially infected painful knee arthroplasty. High uptake at the prosthesis–bone interface indicates an infection. On the other side, an intermediate uptake suggests aseptic loosening. FDG uptake is usually estimated using SUV score [19].

Zhuang et al. [19] reported that 18F-FDG/PET has a sensitivity of 91%, a specificity of 72%, and an accuracy of 78% to assess total infected knee replacement.

Love et al. reported that 18F-FDG/PET has low specificity, so it can be used to exclude infection if the test is negative [11]. 18F-FDG/PET can quantify disease activity. It can be used to assess patients with infections during various phases of the disease [20].

Many authors have concluded from their results that 18F-FDG/PET is a promising tool to distinguish septic loosening from aseptic, with a sensitivity of approximately 80–100% and specificity of almost 90–100% [19]. More studies compare the value of FDG-PET with combined 111In-labelled leukocyte/99mTc-sulphur colloid

bone marrow imaging for diagnosing infection in knee prostheses. Compared to ^{111}In -labelled leukocyte/ $^{99\text{m}}\text{Tc}$ -sulphur colloid bone marrow imaging, positron emission tomography with ^{18}F -fluorodeoxyglucose has many advantages, including the availability, the presence of only one radiotracer injection, and the short execution time (less than 2 h) [15]. Moreover, positron emission tomography with ^{18}F -fluorodeoxyglucose provides superior spatial resolution compared to ^{111}In -labelled leukocyte/ $^{99\text{m}}\text{Tc}$ -sulphur colloid bone imaging [21].

9.13 Conclusion

In conjunction with the clinical examination and laboratory testing, radiologic imaging represents an important adjunct in the diagnosis of PJI. Radiologic imaging includes radiographs, MR, CT, US, and nuclear studies. MR and CT imaging are frequently used to evaluate the extent of disease and exclude other or additional pathologic and traumatic findings. With the presence of surgical hardware, both MR and CT imaging may be degraded by metal artefact that may be decreased with the utilization of metal suppression techniques. Various nuclear imaging studies are helpful in the diagnosis of PJI including SPECT/CT, Bone Scintigraphy, Gallium-67 Citrate Scintigraphy, Indium-111 Leukocyte Scan, Dual-Isotope Imaging with In-111-Labelled Leukocytes and Tc- $^{99\text{m}}$ Sulphur Colloid Bone Marrow Scan, Labelled Leukocyte Scintigraphy with Antigranulocyte Antibody and ^{18}F -FDG/PET.

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Definition of Periprosthetic Joint Infection

10

Elie Kozaily, Noam Shohat, and Javad Parvizi

Since periprosthetic joint infection (PJI) has emerged as a devastating complication after knee replacement, scientific societies have been mobilized to find valid criteria to define a PJI.

The first definition was proposed in 2011 by a group of experts convened by Musculoskeletal Infection Society (MSIS). Since then, many working groups outlined definition criteria for PJI. In fact, the Infection Disease Society of North America (IDSA) published a definition in 2013 then international experts met in Philadelphia for the International Consensus Meeting (ICM) in 2013 and later in 2018.

Accordingly, we walk you through the evolution of the PJI definition from its earliest version by MSIS 2011 to its latest 2018 version by ICM.

The MSIS 2011 criteria have been consistently used by clinicians and researchers. The working group describes two groups of patients who can be safely diagnosed with PJI by fulfilling one of the two major criteria (or both): The patients who present with a sinus tract communicating with the prosthesis and the patients in whom two separate

tissue or fluid samples—obtained from the affected prosthetic joint—put to evidence the same pathogen by culture. Otherwise, MSIS group recommends that four out of six minor criteria be present to diagnose a PJI. The minor criteria include: elevated serum erythrocyte sedimentation rate (ESR) and serum C-reactive protein (CRP), elevated synovial leukocyte count, elevated synovial neutrophil percentage, purulence in the affected joint, isolation of a microorganism in one culture of periprosthetic tissue or fluid, greater than five neutrophils per high-power field in five high-power fields on from histologic analysis of periprosthetic tissue [1].

Of note, the group acknowledged that PJI may be present even if fewer than four of these minor criteria are met. For instance, some of these criteria may be negative in low-grade infections.

The following points need to be considered when interpreting the MSIS criteria:

- Serum biomarkers thresholds for ESR and CRP were 30 mm/h and 10 mg/dL respectively.
- However, ESR and CRP interpretation can be challenging as their serum level depends on a myriad of patient-related factors (inflammatory arthritis, obesity) and the time from index joint replacement among others.
- Synovial fluid markers, white blood cells WBC and PMNs percentage, for knee chronic PJI were reported to range from 1100 to

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4000 cells/ μL and 64 to 69% respectively. In acute knee PJI, occurring 3 months or prior from index surgery, numbers were higher 20,000 cells/ μL and 89% for WBC and PMNs percentage respectively.

- Tissue samples sent for microbiology should be obtained from representative periprosthetic tissue or (synovial) fluid. At least three and no more than five samples should be sent for culture; gram stain and other tests may not be perfectly accurate for diagnosing a PJI. The isolated pathogen in the two samples should be confirmed identical based on its phenotype and anti-microbial susceptibility in vitro [*since genetic testing like polymerase chain reaction (PCR) or next-generation sequencing (NGS) are not routinely ordered*].
- Histology studies should take the clinical picture into consideration as the pathologist looks for at least five polymorphonuclears (PMNs) per high-power field (HPF) on surgical tissue examination.
- For instance, elevated neutrophil count may be present in a periprosthetic fracture or inflammatory arthritis. Other challenges include presence of foreign body macrophages that mimic neutrophils and neutrophils entrapped in superficial fibrin or adherent to endothelium or small veins as those should be disregarded.

In August 2013, hundreds of international experts gathered for the first consensus meeting in Philadelphia, USA. This International Consensus Meeting (ICM) endorsed the MSIS definition. The ICM added leukocyte esterase test as a minor criterion equivalent to elevated WBC count in synovial fluid and excluded the purulence surrounding the prosthesis from the minor criteria. Thus, out of the five minor criteria left, three should be present in order to diagnose PJI [2].

Furthermore, the ICM determined acceptable thresholds for minor criteria based on time from index arthroplasty, acute (within 90 days) vs. chronic infections. The thresholds in acute infections tend to be higher compared to chronic infections, respectively, serum CRP (100 vs. 10 mg/L), synovial fluid WBC (10,000 vs. 3000 cells/ μL), and PMN counts (90% vs. 80%). The criteria were similar for leukocyte esterase

and histological analysis in acute and chronic settings. Yet, no threshold was determined for ESR in acute infection compared to well-defined 30 mm/h in chronic infection [2].

In the same year 2013, the IDSA panelists followed the classical process for establishing guidelines, thereby commanding a strength for each of the criteria based on quality of evidence, changing the concept of major and minor criteria. The panel's definition included the presence of a single culture with virulent organism as a criterion for PJI diagnosis [3].

With new biomarkers surfacing along with traditional biomarkers' interpretation changing, the international experts met for the second edition of the ICM, in Philadelphia, USA in 2018 to refine and update PJI definition criteria in order to improve outcomes [4] (Table 10.1).

The ICM acknowledged that minor differences may exist between PJI after hip and knee replacement yet the proposed definition applies to both joints.

Although the definition did not reach a strong consensus with 68% of the experts agreeing, 28% disagreeing, and 4% abstaining, these recent ICM criteria have shown consistent validity. For instance, a higher sensitivity and a similar specificity compared to MSIS and previous ICM, 97.7% and 99.5% respectively. Similarly, thresholds for biomarkers have been chosen to optimize sensitivity in early stage PJI and specificity in advanced stage PJI.

Of note, chronicity of infection as well as invasiveness of the diagnostic tests is considered in this definition in order to set the ground for an algorithm-based approach. Relative weights for each of the findings and biomarkers have been put in place hence a scoring system, based on American Academy of Orthopedic Surgery (AAOS) guidelines.

The major benefit from these new criteria is the potential to establish a pre-operative diagnosis. In fact, in many cases, the diagnosis of PJI could not be definitive until the surgeon goes for a revision surgery and relies on peri-operative findings like intraoperative purulence or frozen section results. The role of joint aspiration pre-operatively has become pivotal, allowing more than 80% of cases to be diagnosed before going to revision diagnostic and therapeutic surgery.

Table 10.1 PJI definition criteria. Reprinted from Shohat N, Tan TL, Della Valle CJ, Calkins TE, George J, Higuera C, et al. Development and Validation of an Evidence-Based Algorithm for Diagnosing Periprosthetic Joint Infection. *J Arthroplasty* 2019 Nov;34 (11):2730–2736.e1, Copyright (2019), with permission from Elsevier

| Major criteria (at least one of the following) | Decision |
|--|----------|
| Two positive cultures of the same organism | Infected |
| Sinus tract with evidence of communication to the joint or visualization of the prosthesis | |

| Preoperative Diagnosis | Minor Criteria | | Score | Decision | |
|------------------------|----------------|---|-------|----------|---|
| | Serum | Elevated CRP <i>or</i> D-Dimer | 2 | | ≥6 Infected 2-5 Possibly Infected ^a 0-1 Not Infected |
| | | Elevated ESR | 1 | | |
| | Synovial | Elevated Synovial WBC <i>or</i> LE (++) | 3 | | |
| | | Positive Alpha-defensin | 3 | | |
| | | Elevated Synovial PMN % | 2 | | |
| | | Elevated Synovial CRP | 1 | | |

| Preoperative Diagnosis | *Inconclusive pre-op score <i>or</i> dry tap | | Score | Decision | |
|------------------------|--|--|-------|----------|---|
| | Preoperative score | | - | | ≥6 Infected 4-5 Inconclusive ^b ≤3 Not Infected |
| | Positive Histology | | 3 | | |
| | Positive Purulence | | 3 | | |
| | Positive Single Culture | | 2 | | |

^aFor patients with inconclusive minor criteria, operative criteria can also be used to fulfill definition for PJI

^bConsider further molecule diagnostics such as Next-generation sequencing

New biomarkers like serum D-dimer and synovial fluid alpha-defensin have been introduced.

In fact, a high serum D-dimer can be as relevant as a high CRP in chronic PJI; however, more studies are needed to validate serum D-dimer role in diagnosing acute PJI and PJI in general.

Even though synovial fluid alpha-defensin has been criticized as it is an expensive and not routinely ordered test, the experts still assert that the introduction of alpha-defensin aims to help specialists diagnose challenging cases and/or interpret and integrate this test's results, if available.

Of note, optimal thresholds for synovial fluid biomarkers in diagnosing chronic knee PJI were defined as higher than 3000 cells/ μ L for WBC and higher than 80% for PMNs [5].

Next-generation sequencing (NGS) can be valuable when the diagnosis is difficult to make, for example, when PJI is strongly suspected but serum

and/or synovial markers are within normal or the pathogen is not isolated by traditional culture.

Nonetheless, the working force behind the new criteria has reported several limitations and controversies. For instance, these criteria are mainly validated on chronic PJIs (at least 6 weeks from index joint replacement), Plus, in many instances like adverse local tissue reactions, inflammatory arthritis these criteria may not be applicable.

Like in previous recommendations, the experts reaffirm that patients can have a PJI but not meet the criteria and vice versa. Clinical judgment by the specialist should prevail to guide the management.

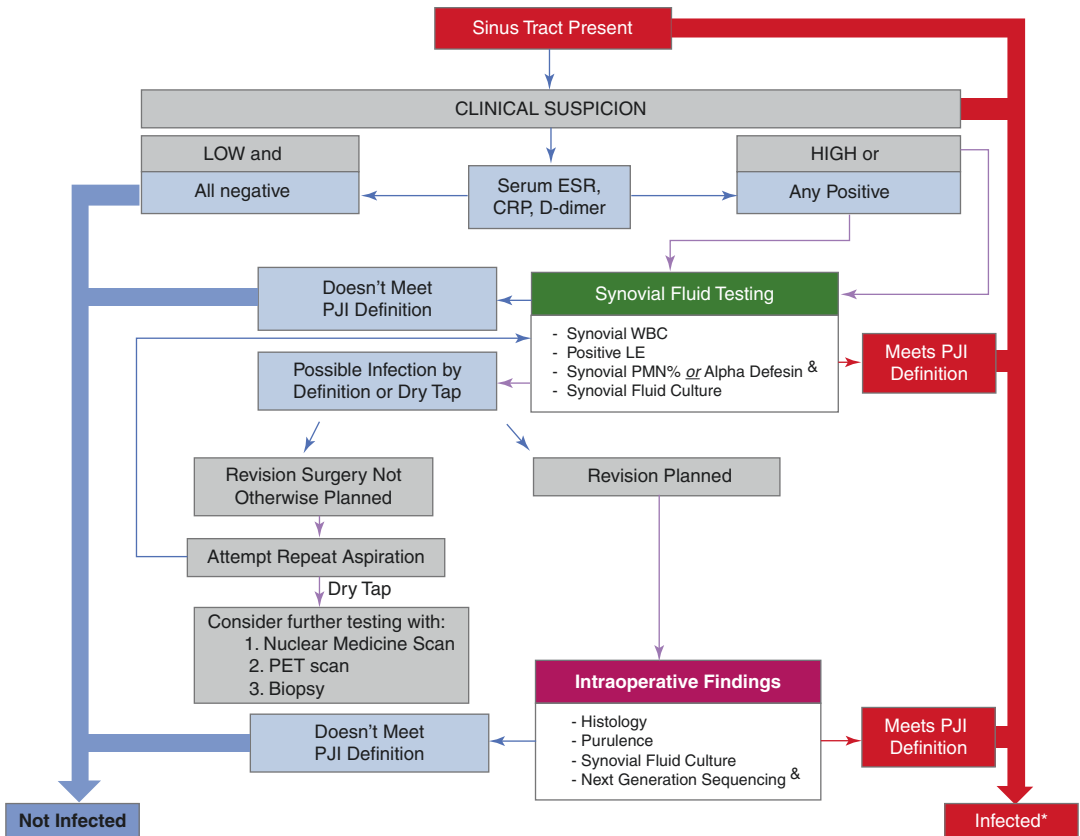
10.1 Algorithm for Diagnosis

The American Academy of Orthopaedic Surgeons (AAOS) provides guidelines in 2010 for the diagnosis of knee and hip PJI using serum and syno-

vial markers. Based on expert opinion and systematic review of the literature, this algorithm simplified a very confusing and challenging process. These guidelines were indorsed by the ICM on PJI in 2013 with slight modifications, and both algorithms have served physicians around the globe in daily clinical practice [6]. Developments in PJI diagnosis including new synovial and serum markers [7–12] has increased confusion among many surgeons who were unsure how to incorporate these tests into their practice and into the previously established guidelines.

In light of these developments, and the 2018 scoring system, Shohat et al. proposed an algorithm that would incorporate recent developments in the field [13]. The study relied on

previous AAOS and ICM guidelines to develop an evidence-based, validated diagnostic algorithm. Using data from three centers and machine learning analysis, a stepwise approach to diagnosing PJI was proposed. Step 1 included serum testing and clinical findings which are evident at the first patient encounter. Step 2 included synovial markers, and the final step, step 3 included intraoperative findings. The proposed algorithm relies on the 2108 PJI definition and was formally validated on an external cohort and demonstrated a high overall sensitivity (96.9%) and specificity (99.5%). Given the significant advantages of this algorithm, it has been introduced in the 2018 ICM on PJI and received a 73% agreement (super majority, strong consensus) (Fig. 10.1).



* At any time, 2 out of 3 of five cultures with the same organism or sinus tract are major criteria for infection & Does not need to be performed Routinely

Fig. 10.1 Algorithm for the diagnosis of PJI. (Reprinted from Shohat N, Tan TL, Della Valle CJ, Calkins TE, George J, Higuera C, et al. Development and Validation of an Evidence-Based Algorithm for Diagnosing

Periprosthetic Joint Infection. *J Arthroplasty* 2019 Nov;34(11):2730-2736.e1, Copyright (2019), with permission from Elsevier)

The first step in evaluating PJI should include serum testing for C-reactive protein, D-dimer, and erythrocyte sedimentation rate. If one of the three is elevated, physicians should proceed with a joint aspiration. However, the authors also noted that in 2.8% of PJI cases all three markers will be negative—and that emphasizes the importance of clinical findings and high clinical suspicion. Patients undergoing revision surgery less than 2 years from index surgery, those with more than 1 surgery on the same joint in the past, signs of erythema, tachycardia, reduced ROM, and serum PMN% above 70 should raise suspicion of PJI and in those cases the joint should be aspirated even if serum markers are negative.

The synovial fluid should be routinely investigated for white blood-cell count with differential and leukocyte esterase testing. Alpha-defensin received special attention as it is an expensive test and not routinely ordered. The authors found that alpha defensin does not add to the performance of the algorithm which implies it should not be ordered routinely. This step would rule in or out PJI in most patients and in those that a diagnosis cannot be reached, intraoperative findings will need to be taken into consideration. Intraoperative findings including purulence, histology, and next-generation sequencing (NGS) or a single positive culture can aid in these cases where the diagnosis has not been conclusive.

It is important to note that it is possible that the diagnosis of PJI may not be made even after reaching the third stage or may be inconclusive after obtaining synovial tests. These patients are often encountered in clinical practice and represent a real diagnostic challenge. Furthermore, it is important to note that the proposed algorithm and the definition of PJI may be inaccurate and require a modification in the tests utilized for the following conditions: adverse local tissue reactions, crystalline deposition arthropathy, inflammatory arthroplasty flares, and infection with slow growing organisms, such as *P. acnes*.

Overall the proposed algorithm has several advantages compared to previous published guidelines. The algorithm allows us to account for the interplay between the individual or combined diagnostic test results and their influence

on the probability for infection at different stages/times throughout the workup for infection. This interplay has a significant effect on the overall diagnostic performance [14–16]. It also allows clinicians to reach an educated conclusion before continuing forward to more invasive and costly tests. Furthermore, it minimizes the number of tests performed in each step and has the potential to reduce costs of unnecessary expensive tests, which are currently oftentimes performed.

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Differential Diagnosis of Periprosthetic Joint Infection

11

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11.1 Introduction

Pain is the most common symptom in PJI and is present in the majority of cases [1, 2]. In a recent study evaluating patients undergoing revision surgery for failed TKA due to PJI and aseptic causes [3], over 90% of patients in both groups complained of pain, making pain alone a very non-specific complaint. As so, any potential cause for pain following TKA should be included in the differential diagnosis of PJI. Causes for pain following TKA can be divided into those present immediately after surgery and those emerging after a period of time that surgery was considered successful (Table 11.1).

Common causes for persistent pain or new onset pain immediately after surgery should be first classified as extrinsic or intrinsic. Extrinsic causes of pain may be a result of poor indication

or dual pathology from sources outside the artificial joint, including the hip joint, spinal radiculopathy, vascular claudication, local bursitis or tendinopathy, and systemic conditions such as autoimmune diseases. Intrinsic causes of pain following surgery result from the artificial joint itself and include instability, malalignment, component mal-positioning and impingement, recurrent hemarthrosis, arthrofibrosis, and extensor mechanism problems.

When pain presents in an artificial joint that was previously pain free, wear, osteolysis, and aseptic loosening should be considered [4, 5]. Aseptic loosening is a frequent cause of pain and revision surgery [6]. It can be caused by various reasons with the end result being failure of the bond between an implant and bone in the absence of infection [7]. While usually occurring 10–20 years from index TKA, it can occur earlier due to patient characteristics and component material and positioning [8, 9]. Symptoms of aseptic causes of failure include pain, joint effusion, erythema, and restricted range of motion among others. These symptoms may present similarly to PJI, thus making it a difficult task to differentiate between these two different pathologies [10–13]. While at times, some of these aseptic causes for failure are overt, making it tempting to make a diagnosis of aseptic failure, PJI must always be ruled out before any revision surgery takes place as 12% of the so-called aseptic cases have an underlying PJI [14, 15].

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Table 11.1 Causes for pain following TKA

| Extra-articular | Intra-articular |
|---|---|
| Radiating: <ul style="list-style-type: none"> • Hip • Spine | Early onset: <ul style="list-style-type: none"> • Maltracking • Malalignment • Instability • Impingement • Clunk |
| Local: <ul style="list-style-type: none"> • Bursitis • Tendinitis | Late onset: <ul style="list-style-type: none"> • Wear • Osteolysis • Loosening |
| Vascular: <ul style="list-style-type: none"> • Claudication • DVT • Bleeding | |
| Systemic: <ul style="list-style-type: none"> • Inflammatory • Neuropathic | |

11.2 Serum Negative Infections

Serum screening tests (C-reactive protein [CRP] and erythrocyte sedimentation rate [ESR]) are usually the first step in the workup of a painful TKA as they are readily available and minimally invasive. While extremely useful, numerous studies have showed low sensitivity and specificity [16, 17]. Berbari et al. in a meta-analysis of 23 papers reported a pooled sensitivity of 75% (95% CI, 72–77%) for ESR and 88% (95% CI, 86–90%) for the CRP level. Pooled specificity for the same markers was 70% (95% CI, 68–72%) and 74% (95% CI) [18]. Kheir et al. calculated a false negative rate of 14.5% for ESR and 8.6% for CRP [17]. Parvizi et al. using machine learning found that 2.5% of patients with PJI will have negative serum screening workup, emphasizing the problem of relying solely on serum testing for screening [19]. This is especially true in infections caused by low-virulence organisms [20–22].

It is therefore fundamental to include clinical acumen in every investigation to ensure that diagnosis of PJI is not missed. In a recent paper investigating the efficacy of clinical findings in the workup of PJI, fever and erythema were found to be the most specific signs for diagnosing PJI with a positive likelihood ratio (LR) of 10.78 and 8.08, respectively. The authors concluded that clinical

presentation can and should be used to guide which future diagnostic tests should be ordered and in the interpretation of their results [12]. Parvizi et al. using random forest analysis found that the presence of fever, erythema, reduced range of motion, tachycardia, failure occurring less than 2 years from index arthroplasty, and a history of more than one surgery in the index joint are all important parameters that should raise suspicion for PJI and prompt aspiration of the joint even if serum markers are negative for infection [19].

11.3 Culture Negative and Slow Growing Organism Infections

Bacterial infections may be culture negative organisms or slow growing, which elicits a relatively subtle immune response, which makes establishing a diagnosis more challenging [17]. Culture negative PJI accounts for up to 40% of cases in some reports [23]. Kheir et al. performed a retrospective study on over 1000 revision arthroplasty cases (549 for PJI, 653 for aseptic causes) and compared commonly used serum and synovial marker cutoffs stratified based on the underlying organism causing infection [17]. Interestingly, they reported that culture negative and slow growing organisms such as coagulase-negative *Staphylococcus* had lower levels of ESR, CRP as well as synovial WBC and PMN%, thus are prone to being misdiagnosed as aseptic failure if not receiving special attention.

Taking these findings into consideration, all efforts should be taken to maximize the utility of culture growth. If a PJI is suspected, the use of empirical antimicrobials prior to fluid aspiration should be avoided to prevent sterilization prior to culture to prevent a false negative result [7, 24]. The American Academy of Orthopaedic Surgeons recommends against prescribing antimicrobials for 2 weeks for patients with suspected PJI until the infection is verified [25]. Fluid aspiration should be repeated and the aspirate incubated for 2–3 weeks on aerobic, anaerobic, fungal, and acid-fast bacilli cultures. One to 2 weeks is rec-

Table 11.2 Red flag patients

| Red flag patients | Presentation | Useful markers | Comments |
|-----------------------------|---|---|--|
| Slow growing organisms | Pain, effusion, erythema, reduced ROM | Serum and synovial markers using 2018 PJI definition molecular testing | In cases where a diagnosis is suspected and cannot be made using conventional methods, molecular testing should be ordered |
| Inflammatory arthritis | Pain, effusion, erythema, reduced ROM, fever | CRP, ESR, synovial WBC, synovial PMN, alpha defensin | Conventional thresholds should be adhered to, except in cases where a flare up is suspected |
| Crystalline deposit disease | Pain, effusion, erythema, reduced ROM, fever | Uric acid, calcium pyrophosphate, CRP, ESR, synovial fluid | Crystalline deposit may be present in small amounts which may be misclassified or missed |
| Hemarthrosis | Pain, effusion, erythema, reduced ROM | Serum PT, PTT, INR Joint aspiration | Joint aspiration will show sanguineous fluid containing red blood cells |
| Metal allergy | Pain, allergic dermatitis, prosthetic loosening | Eosinophils | Skin patch test, leukocyte migration inhibition test, lymphocyte transformation test may all be useful |

recommended for the growth of *P. acnes* and some coagulase-negative staphylococci [8, 15]. During surgery, three to five samples should be taken from the interphase and the pseudocapsule, and tissue should be sent for histopathological analysis.

Previous diagnostic criteria were specific but lacked sensitivity to diagnose less obvious infections. Recently, new diagnostic criteria have been developed to better diagnose these culture negative and low-grade infections [8, 26]. Parvizi et al. in their evidence-based definition for PJI have increased sensitivity for diagnosis to 97.7% (95% CI 94.7%–99.3%) compared to previous criteria showing sensitivity of 79.3–86.9%. Although they were able to capture a substantial amount of PJI patients, in 2.3% of the cohort a diagnosis was inconclusive which they define “gray area” patients, i.e. not clear if infected or not. These patients are often encountered in clinical practice and represent a real diagnostic challenge. Interestingly, all patients with an inconclusive diagnosis had negative cultures, and the authors propose they may benefit from molecular diagnostic testing.

In cases where a PJI diagnosis is still not confirmed, increasing evidence supports the use of genome sequencing [9–11, 26]. Using metagenomics shotgun sequencing, Thoendel et al. detected a wide range of PJI pathogens and suggested this method may aid in identifying the

infecting organism in culture negative PJI [27]. They were able to identify new potential pathogens in 43.9% (43/98) of culture negative PJIs. While the translation of sequencing data into a clinically useful instrument in the setting of PJI remains critically limited, it seems likely that genome sequencing may occupy an increased role for the diagnosis of infection in the future.

Red flag patients are reported in Table 11.2.

In certain populations, conventional workup and diagnostic criteria may be inaccurate. Due to the relative rarity of these patients and possible outliers, many studies exclude them from the analysis. Furthermore guidelines are established and rely on investigations of PJI in primarily patients without these underlying diseases [28].

11.3.1 Inflammatory Arthritis

Autoimmune and chronic diseases have similar clinical symptoms as PJI and obtaining a thorough patient history, including past medical history, is vital in determining the cause of joint failure. Rheumatoid arthritis (RA) has received the most focus as its complications are a common cause for joint arthroplasty [29]. Elevated serum CRP and ESR levels are diagnostic criteria for PJI and are also commonly used to aid in the diagnosis of RA. Thus, applying the same thresholds in RA patients as in the general population raises concern

over identification and false positive results. Yeganeh et al. and Shohat et al. examined whether different thresholds for commonly used serum and synovial markers should be used in patients with inflammatory arthritis, the majority with rheumatoid arthritis [29, 30]. Both studies concluded that conventional PJI thresholds for serum and synovial diagnostic markers are sufficient for differential diagnosis with respect to RA. Special attention should be given to patients presenting with an acute flare as these studies were performed on patients presenting in various stages of the disease activity.

11.3.2 Crystalline Deposit Diseases

Patients with a suspected PJI and a history of crystalline deposit diseases, such as gout and pseudogout, should have synovial fluid analysis performed before administration of antibiotics to avoid falsely negative cultures as infections may precipitate crystalline deposit disease as crystalline deposits may act as a nidus for infection [31, 32]. George et al. reported on 22 patients with crystalline deposition within the affected joint that fueled inflammation and presented with edema, erythema, fever, and pain within the joint; symptoms similar to that of a PJI [31]. As these two pathologies require different treatment, it is important for the physician to be aware of the proper diagnostic workup to proceed accordingly. Diagnostic studies of the aspirated fluid should include microscopic studies looking for uric acid crystals to detect gout and calcium pyrophosphate in suspected pseudogout. Overlapping manifestations of crystalline deposit disease and PJIs include elevated white blood cell count, ESR, and CRP. Synovial fluid aspiration should be examined for crystals, cultures should be taken, and uric acid levels measured. These crystalline deposit diseases are difficult to diagnose because of the small amount of crystals, which may be misclassified or missed entirely by physicians with limited clinical experience.

11.3.3 Hemarthrosis

Hemarthrosis occurs due to bleeding into the joint and may present similarly to PJI, including

decreased range of motion, edema, erythema, and pain [33]. Hemarthrosis should be suspected based on previous history of the patient and may occur from traumatic or atraumatic causes, such as coagulopathies and anticoagulant medication. Hemarthrosis can be diagnosed via imaging or aspiration of the joint. Joint aspiration will show sanguineous fluid containing red blood cells. Additional diagnostic studies should include cell differentials, gram stain, and culture to confirm the diagnosis.

11.3.4 Metal Allergy

While rare, metal hypersensitivity is also a potential cause of pain following TKA that may mimic PJI [34, 35]. The most common allergy-inducing metals are nickel (19.7–24.4%), cobalt (2–8.8%), and chromium (2.4–5.9%). Clinical symptoms of metal hypersensitivity include local or systemic allergic dermatitis, pain, and prosthetic loosening due to chronic aseptic inflammation [36]. With respect to standard laboratory examinations, the skin patch test, leukocyte migration inhibition test (LMIT), and lymphocyte transformation test (LTT) are the most popular diagnostic tests for metal hypersensitivity. However, none of the tests has been universally accepted and applied.

11.4 Conclusion

As the number of knee arthroplasties continues to increase each year, physicians will increasingly encounter unsatisfied patients and those with chronic pain and will need to recognize the cause of the pain and treat the patient accordingly. PJI is a severe complication following knee arthroplasty with several differential diagnoses that surgeons should be aware of. A working diagnosis of suspected PJI will be able to improve quality of care for patients through proper treatment. Future research should focus on cases where differences between PJI and aseptic cases are subtle in an effort to improve diagnosis and subsequent treatment.

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Part V

Treatment of Knee Replacement Infections



Systemic Antibiotic Therapy

12

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12.1 Introduction

Both systemic antibiotic therapy and surgery are crucial for the management of periprosthetic joint infections (PJI) of the knee [1]. Sometimes, antibiotic therapy is used as a standalone suppression therapy in patients where surgical intervention is impossible.

Delivery of systemic antibiotic agents can either be achieved orally or intravenously. While the intake of oral antibiotics generally is much more convenient and may even be managed in an outpatient setting, intravenous antibiotics usually exhibit a better bioavailability but, as a downside,

may require inpatient therapy and a thorough monitoring [2, 3].

Common goal of all systemic antibiotic treatments is to eradicate the cause of infection that should be identified prior to therapy induction (see Sect. 12.3) [4]. In case a bacterium is unknown, treatment with broad spectrum antibiotics should be considered at the beginning [5]. However, joint aspiration and biopsies are required first to ensure later optimization of any antibiotic treatment. The secondary goals of systemic antibiotic therapy are to deal with the biofilm, formed through bacterial colonization, e.g. with *Staphylococcus* spp. or *Streptococcus* spp. (see Sect. 12.1) or to prevent biofilm formation on newly implanted hardware in the first place [6].

Although being crucial to an effective antimicrobial therapy in cases of PJI, most systemic antibiotics do not reach an effective local concentration alone and thus require a synergistic use of a local antibiotics [7]. The importance of these locally delivered antibiotics (e.g. from cement spacers) will be discussed later (see Chap. 13). Quorum quenching, meaning the disruption of bacterial communication—the so-called quorum sensing—of certain species [8] as well as chemotherapeutic approaches [9] are technically not considered systemic antibiotic treatments in a narrower sense and will therefore be detailed later (see Chap. 13).

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12.2 Principles and Timing

As a basic principle, any antibiotic therapy should be delayed until culture specimens have been taken (see Sect. 12.3). Only this assures an increased probability to eventually start a focused therapy at a certain time point [10]. In contrast, some recent publications advocate to start with antibiotic treatment as soon as possible since culture yields may eventually be the same, but later recurrence rates of PJI are considerably lower [11]. However, systemic antibiotic treatment should always be planned in advance, taking into consideration all available diagnostic and clinical information of the given case. In an ideal scenario, antibiotic treatment is usually initiated in an interdisciplinary consensus between orthopedic surgeons, clinical microbiologists, and all other involved specialists [12].

In each case of TKA revision the need for an initial empirical therapy has to be weighed against its clinical implications. Especially in PJI cases accompanied by acute sepsis or positive systemic inflammatory response syndrome (SIRS) criteria, early systemic antibiotic treatment should not be delayed in order to improve clinical outcomes [13]. It may also be helpful to identify the most common infective agents in your hospital or department to enable an individualized empiric response, even differentiating between acute and chronic infections. Wherever possible, empirical antibiotic therapy should be converted into a focused therapy depending on culture results in the course of the treatment. While delayed systemic therapy applies for those cases with septic implant removal without previous cultures, systemic therapy in one-stage revision or mobile parts exchange procedures should usually be initiated 15–60 min before tourniquet inflation or skin incision [14, 15]. Dosage and duration of recommended antibiotic therapies are shown in Table 12.1.

In cases of retained hardware, biofilm-activity is a major requirement for an ideal systemic antibiotic treatment. Cases without hardware on the other hand mainly require an antibiotic therapy with preferable tissue penetration (e.g. to treat osteitis).

12.3 Debridement, Antibiotics, and Implant Retention (DAIR)

This treatment strategy may either involve an empiric therapy with an antibiotic agent of a much broader spectrum, in case the pathogen is still unknown, or a focused therapy, in case the pathogen has already been identified. Duration and dosages are documented in Table 12.1. In cases of DAIR, it is of crucial importance to also add biofilm-active substances (e.g. Rifampicin) to the antibiotic therapy regimen if they are needed (e.g. against *Staphylococcus* spp. or *Cutibacterium* spp.). Pathogens that belong to the difficult-to-treat (DTT) group, meaning those causative agents for which no biofilm-active antibiotics are available, can only be suppressed by prolongation of systemic antibiotic treatment [16]. In these cases, later suppressive therapy may be a valid option. Gram-negative bacteria should be treated with fluoroquinolones like Ciprofloxacin intravenously and even during a later sequential phase. Administration of Rifampicin should be avoided in cases of gram-negative infections [17].

Generally, an interdisciplinary approach together with a clinical microbiologist is advised to select the best possible biofilm-active agent in cases of multi-resistant bacteria strains [18]. All other causative agents are treated as described in paragraph 12.2 and in Table 12.1. In summary, the key to success of DAIR is a proven susceptibility of the identified pathogen to biofilm-active substances paired with a thorough surgical debridement.

12.4 One-Stage Exchange

The same principles as for DAIR also apply for the concept of one-stage exchanges. Even here, the main goal is a biofilm-active therapy. Duration and dosages of various substances are also documented in Table 12.1. However, no clear evidence exists for the ideal duration of intravenous and sequential therapy [16].

Table 12.1 Systemic antibiotic treatment: proposed substances, sequence, duration, and dosage

| Microorganism | Antibiotic substance | Dosage (route) | |
|---|---|---------------------------|---------------------|
| <i>Staphylococcus</i> spp. | | | |
| Methicillin/oxacillin-susceptible (with remaining hardware, DAIR) | Flucloxacillin <i>OR</i> | 3 × 4 g or 4 × 3 g (i.v.) | |
| | Cefazolin <i>OR</i> | 3 × 2 g (i.v.) | |
| | Daptomycin | 1 × 8–10 mg/kg (i.v.) | |
| | + Rifampicin ^a (each) | 2 × 0.45 g (p.o.) | |
| | <i>Followed by (depending on susceptibility testing)</i> | | |
| | Levofloxacin <i>OR</i> | 2 × 0.5 g (p.o.) | |
| | Doxycycline <i>OR</i> | 2 × 0.1 g (p.o.) | |
| | Cotrimoxazole | 3 × 0.96 mg (p.o.) | |
| | + Rifampicin ^a (each) | 2 × 0.45 mg (p.o.) | |
| | Methicillin/oxacillin-resistant (with remaining hardware, DAIR) | Daptomycin <i>OR</i> | 1 × 12 mg/kg (i.v.) |
| Vancomycin ^b | | 2 × 15 mg/kg (i.v.) | |
| + Rifampicin ^a (each) | | 2 × 0.45 g (p.o.) | |
| <i>Followed by (depending on susceptibility testing)</i> | | | |
| Levofloxacin <i>OR</i> | | 2 × 0.5 g (p.o.) | |
| Doxycycline <i>OR</i> | | 2 × 0.1 g (p.o.) | |
| Cotrimoxazole | | 3 × 0.96 mg (p.o.) | |
| + Rifampicin ^a (each) | | 2 × 0.45 mg (p.o.) | |
| Rifampicin-resistant | | Daptomycin <i>OR</i> | 1 × 10 mg/kg (i.v.) |
| | | Vancomycin ^b | 2 × 15 mg/kg (i.v.) |
| | <i>i.v. for 2–4 weeks, followed by life-long suppression therapy (Cotrimoxazole, Doxycycline) under certain circumstances</i> | | |
| <i>Streptococcus</i> spp. | | | |
| Penicillin-susceptible | Penicillin G <i>OR</i> | 4 × 5–10 M U (i.v.) | |
| | Ceftriaxone | 2 × 1–2 g (i.v.) | |
| | <i>i.v. for 2 weeks, followed by</i> | | |
| | Amoxicillin <i>OR</i> | 3 × 1 g (p.o.) | |
| | Levofloxacin | 2 × 0.5 g (p.o.) | |
| Penicillin-resistant | Vancomycin ^b <i>OR</i> | 2 × 15 mg/kg (i.v.) | |
| | Daptomycin | 1 × 10 mg/kg (i.v.) | |
| | <i>i.v. for 2 weeks, followed by</i> | | |
| | Levofloxacin <i>OR</i> | 2 × 0.5 g (p.o.) | |
| | Clindamycin | 3 × 0.6–0.9 g (p.o.) | |
| <i>Enterococcus</i> spp. | | | |
| Penicillin/Ampicillin-susceptible | Ampicillin | 3 × 5 g (i.v.) | |
| | + Gentamicin | 1 × 6–7 mg/kg (i.v.) | |
| | <i>i.v. for 2–3 weeks, followed by</i> | | |
| | Amoxicillin <i>OR</i> | 3 × 1 g (p.o.) | |
| | Linezolid (in cases of known allergies, max. 4 weeks) | 2 × 0.6 g (p.o.) | |
| Penicillin/Ampicillin-resistant | Daptomycin <i>OR</i> | 1 × 10–12 mg/kg (i.v.) | |
| | Vancomycin ^b | 2 × 15 mg/kg (i.v.) | |
| | + Gentamicin | 1 × 6–7 mg/kg (i.v.) | |
| | <i>i.v. for 2–4 weeks, followed by</i> | | |
| | Linezolid (max. 4 weeks) | 2 × 0.6 g (p.o.) | |

(continued)

Table 12.1 (continued)

| Microorganism | Antibiotic substance | Dosage (route) |
|---|--|--|
| Vancomycin-resistant (VRE) | Daptomycin <i>OR</i> | 1 × 10–12 mg/kg (i.v.) |
| | Linezolid (max. 4 weeks) | 2 × 600 mg (i.v.) |
| | <i>Followed by life-long suppression therapy under certain circumstances</i> | |
| <i>Enterobacterales</i> | | |
| Quinolone-susceptible | Ciprofloxacin | 2 × 750 mg (p.o.) |
| | + Ampicillin/Sulbactam <i>OR</i> | 4 × 3 g (i.v.) |
| | Piperacillin/Tazobactam | 4 × 4.5 g (i.v.) |
| <i>i.v. for 2 weeks</i> | | |
| Quinolone-resistant | Meropenem <i>OR</i> | 3 × 2 g (i.v.) |
| | Piperacillin/Tazobactam <i>OR</i> | 4 × 4.5 g (i.v.) |
| | Colistin | Loading dose 1 × 9 M U, then 3 × 3 M U (i.v.) <i>OR</i> 2 × 4.5 M U (i.v.) |
| | + Fosfomycin | 3 × 4–5 g (i.v.) |
| | <i>i.v. for 2 weeks, followed by sequential therapy in consensus with microbiologist</i> | |
| <i>Non-fermenters</i> | | |
| (e.g. <i>Pseudomonas aeruginosa</i> , <i>A. baumannii</i>) | Meropenem <i>OR</i> | 3 × 2 g (i.v.) |
| | Piperacillin/Tazobactam <i>OR</i> | 4 × 4.5 g (i.v.) |
| | Ceftazidime | 3 × 2 g (i.v.) |
| | + Tobramycin | 1 × 6–7 mg/kg (i.v.) |
| | <i>i.v. for 2 weeks, followed by (depending on susceptibility testing)</i> | |
| | Ciprofloxacin | 2 × 0.75 g (p.o.) |
| (e.g. <i>Pseudomonas aeruginosa</i> , <i>A. baumannii</i>) multi-resistant | <i>Adapted therapy in consensus with microbiologist</i> | |
| <i>Anaerobes</i> | | |
| <i>Cutibacterium acnes</i> | Penicillin G <i>OR</i> | 4 × 5–10 M U (i.v.) |
| | Clindamycin (in cases of known allergies) | 3 × 600 mg (i.v.) |
| | + Rifampicin ^a | 2 × 0.45 g (p.o.) |
| | <i>i.v. for 2 weeks, followed by</i> | |
| | Amoxicillin <i>OR</i> | 3 × 1 g (p.o.) |
| | Levofloxacin | 2 × 0.5 g (p.o.) |
| + Rifampicin ^a | | 2 × 0.45 g (p.o.) |
| <i>Gram-positive anaerobes</i> | | |
| Non <i>Cutibacterium acnes</i> | Ampicillin/Sulbactam <i>OR</i> | 4 × 3 g (i.v.) |
| | Piperacillin/Tazobactam <i>OR</i> | 4 × 4.5 g (i.v.) |
| | Moxifloxacin | 1 × 400 mg (p.o.) |
| | <i>i.v. for 2 weeks</i> | |
| <i>Gram-negative anaerobes</i> | | |
| | Ampicillin/Sulbactam <i>OR</i> | 4 × 3 g (i.v.) |
| | Piperacillin/Tazobactam <i>OR</i> | 4 × 4.5 g (i.v.) |
| | <i>i.v. for 2 weeks, followed by</i> | |
| | Metronidazole | 3 × 400 mg (p.o.) |
| <i>Candida</i> spp. | | |

Table 12.1 (continued)

| Microorganism | Antibiotic substance | Dosage (route) |
|-----------------------------|--|--|
| Fluconazole-susceptible | Caspofungin ^c OR | Loading dose 1 × 70 mg, then 1 × 50 mg (i.v.) |
| | Anidulafungin OR | Loading dose 1 × 200 mg, then 1 × 100 mg (i.v.) |
| | Fluconazole | 1 × 400 mg (p.o.) |
| | <i>Followed by life-long suppression therapy under certain circumstances</i> | |
| Fluconazole-resistant | Caspofungin ^c OR | Loading dose 1 × 70 mg, then 1 × 50 mg (i.v.) |
| | Anidulafungin OR | Loading dose 1 × 200 mg, Then 1 × 100 mg (i.v.) |
| | Voriconazole | 2 × 200 mg (p.o.) |
| <i>Culture-negative PJI</i> | | |
| | Ampicillin/Sulbactam | 4 × 3 g (i.v.) |
| | +/- Rifampicin ^a | 2 × 0.45 g (p.o.) |

DAIR debridement, antibiotics, and implant retention, *i.v.* intravenously, *p.o.* per os, VRE vancomycin-resistant Enterococcus; PJI periprosthetic joint infection. Comments: Generally, dose-adjustment according to regularly monitored renal and liver function as well as body weight is advised

^aBiofilm-active antibiotics (e.g. Rifampicin) should not be given over the course of a temporal cement spacer treatment or while surgical drains are still in situ. Patients aged 75 years and older should receive a reduced dosage of Rifampin with 2 × 0.30 g (p.o.)

^bVancomycin levels of concentration should be monitored closely every 2–3 days before administration of the next dose (low plasma-levels, ideal range 15–20 mg/L)

^cCaspofungin should be administered with a maintenance dose of 1 × 70 mg, if body weight exceeds 80 kg

12.5 Two-Stage Exchange

Any two-stage exchange procedure can be divided into two distinctive steps: (1) prosthesis removal and (2) prosthesis re-implantation. The first step is followed by a phase where the joint remains without an implant or with a spacer (e.g. bone cement). The main goal during this phase is an antibiotic therapy with a good tissue penetration to treat osteitis or osteomyelitis as well as soft tissue infections. Biofilm-active substances are unnecessary, even when the cement spacer remains in situ [19]. The surgical approach and subsequent strategies are explained in detail in Sect. 12.4. The duration between the first and second step is usually between 2 and 6 weeks, depending on the concept that was chosen. Drug holidays—a phase without systemic antibiotic

treatment in between—is no longer recommended, as their effect is not supported by the literature [16]. Following the second step there is a new prosthesis in situ yet again and thus the main goal then is to protect it against colonization with biofilm-active antibiotics. In cases of DTT a prolonged duration of systemic antibiotics or life-long suppressive therapy may be required.

12.6 Empiric Therapy

Main paradigm of an empiric systemic antibiotic therapy, especially in bacterial sepsis, is “hit early, hit hard.” This requires substances covering a broad spectrum of bacteria including *Staphylococcus* spp. (including methicillin-resistant strains) and certain gram-negative bacteria (*Enterobacterales*,

formerly *Enterobacteriaceae*). There should also be sufficient information about the pathogen-specific spectrum of germs of the department or region [16, 20].

In cases of acute sepsis or SIRS, joint aspiration and culture (in special small-volume sample media, e.g. pediatric blood culture flasks), native synovial fluid cell count (total leucocytes, total granulocytes, and leucocyte/granulocyte ratio) as well as two pairs of blood cultures are desirable before therapy initiation to eventually obtain a culture of the suspected causative agent [21]. Possible empiric treatment options include Vancomycin and a third or fourth generation cephalosporin or Piperacillin/Tazobactam (Table 12.1). Since therapeutic concentrations of cephalosporin in the knee joint can already be reached in standard dosage, no special adaption is required for an initial therapy [22]. One of the main causes of PJI with negative cultures is an early beginning of empiric treatment [23]. For this reason, individual patient risk must be calculated and weighed against a possibly unidentified causative agent during the course of further treatment. Empiric therapy should generally be administered intravenously [16, 24].

12.7 Focused Therapy

After gathering all available diagnostic information (see Sect. 12.3), initiation of a focused treatment can be discussed and planned. Ideally, histopathologic and microbiologic data, foremost culture results are the basis for an adequate antibiotic treatment using a single or in combined substances. The appropriate dosage but also the duration of therapy should be defined. Some antibiotics may require hematological testing on a regular basis in order to detect side effects. During this stage any already initiated empiric therapy should be revisited and adjusted. Antibiotic susceptibility profiles, which usually accompany extended microbiologic cultures, can help to identify the best possible treatment depending on the individual specifications of the causative agent [25, 26]. This tool becomes even more important in cases of mixed infections with

various bacteria in addition to further diagnostics like PCR (see Sect. 12.3).

An ideal systemic antibiotic agent will fulfill the following criteria [27]:

- Status and activity of the bacteria.
- Good tissue penetration of bone and other musculoskeletal tissues.
- High ratio of attainable local tissue concentration and minimal inhibitory concentration (MIC).
- Low rates of spontaneous resistance formation.
- Activity even against planktonic and biofilm-embedded bacteria (especially in cases of remaining hardware).
- Good tolerance and long-term tolerance for the patient.
- Suitability for sequential therapy (high oral bioavailability).

Moreover, the choice of any systemic antibiotic treatment also depends on the surgical approach [16, 28]. The principle of DAIR may require other substances and treatment durations than one-stage or two-stage revision of TKA [29, 30]. Focused or targeted therapy is usually started as a continuation of previous empiric therapy and should be administered intravenously for at least 2 weeks post-operatively (counting in the period of empiric therapy), before a conversion to oral administration is considered. Generally, a treatment period of 12 weeks is recommended for systemic antibiotic therapy starting from the time point of the index surgery and the first empiric antibiotic therapy. Intravenous antibiotics are given first during the perioperative period followed by oral therapy usually for 5–10 weeks [16]. The time frame between intravenous and oral antibiotic treatment will depend on the type of bacterium and the adequate antibiotic available for oral administration [31].

12.7.1 *Staphylococcus* spp.

Systemic antibiotics effective against methicillin-susceptible *S. aureus* (MSSA) are beta-lactam

antibiotics like Oxacillin, Cefazolin and Ceftriaxone (Table 12.1). In case of a known hypersensitivity (e.g. history of previous anaphylaxis) against beta-lactams, Vancomycin or Daptomycin may be chosen [18]. However, care has to be taken with regard to correct dosage monitoring due to their side effects and a possible risk of later re-infection [32, 33]. Infections caused by methicillin-resistant *S. aureus* (MRSA) may be treated with Vancomycin, Daptomycin or even Teicoplanin [16, 31]. This includes almost all coagulase-negative strains with the exception of *S. lugdunensis* which usually is methicillin-susceptible and can therefore be treated with a beta-lactam (Table 12.1).

Since most species of *Staphylococcus* are considered to be biofilm-forming, systemic antibiotic treatment should be supplemented by an adjunctive, biofilm-active antibiotic like Rifampicin [20]. Through inhibition of bacterial transcription in mural synthesis, Rifampicin breaks up biofilms and thus increases the anti-infective effect of the combination therapy agent in certain cases—especially in patients whose implants are retained during DAIR procedures or who undergo one-stage revisions [9]. Meropenem, an alternative to Rifampicin, instead inhibits transpeptidases to achieve a comparable effect of biofilm breakage [34]. Rifampicin monotherapy should be avoided, because it is associated with a high risk of early resistance induction due to point mutation [35]. In combination therapies, it is discussed that potentially Rifampicin should be administered with a certain delay to surgical debridement providing a non-oozing, dry wound, and initiation of systemic antibiotic therapy with another antibacterial substance [35]. Moreover, Rifampicin may induce Cytochrome P450 (CYP3A) and thus alter serum blood levels of other medications. In general, adjunctive, biofilm-active antibiotics should not be given over the course of a temporal cement spacer treatment or while surgical drains are still in situ [16].

Staphylococcus spp. unsusceptible to biofilm-active substances like Rifampicin belong to the DTT group of causative agents.

12.7.2 *Streptococcus* spp.

Streptococcus spp. should be treated with intravenous beta-lactam antibiotics like Penicillin or Ampicillin (Table 12.1). In the outpatient setting, an intravenous treatment with Ceftriaxone appears favorable due to the ease of dosing [36]. Again, in case of a known hypersensitivity against beta-lactams, Vancomycin is the alternative of choice given the previously named precautions [33].

12.7.3 *Enterococcus* spp.

Enterococcus spp. show a broad variety in their susceptibility patterns and therefore susceptibility testing appears to be mandatory in order to choose the right combination of substances. Intravenous monotherapy with a beta-lactam antibiotic like Ampicillin can be sufficient, if local delivery of a second antibiotic like gentamicin via the coating of an implant or cement spacer is ensured [37]. However, one should be aware about the limited time of antibiotics release. Ampicillin-resistant *Enterococcus* spp. should be treated with Vancomycin or Daptomycin (Table 12.1).

12.7.4 Gram-Negative Bacteria

Systemic substances with an activity against gram-negative bacteria like *Enterobacteriales* (*Escherichia coli*, *Klebsiella* spec., *Enterobacter* spec.) include beta-lactam antibiotics, carbapenems, and fluoroquinolones (Table 12.1). In cases of a suspected gram-negative infection, susceptibility testing is strongly advised to confirm the efficacy of the desired therapy agent [9, 16]. Since fluoroquinolones show a very high tissue penetration especially for bone, they should be considered primarily for oral treatment [16, 38].

Non-fermenters like *Pseudomonas aeruginosa* or *Acinetobacter* spp. represent difficult causative agents, since even radical surgical treatment sometimes is not able to eradicate infection. Proposed substances for systemic

antibiotic treatment include Ciprofloxacin, Levofloxacin, Ceftazidime, Piperacillin/Tazobactam, and Meropenem (Table 12.1). Ciprofloxacin-resistant gram-negative bacteria belong to the DTT group of causative agents [16].

12.7.5 Anaerobic Bacteria

Gram-positive anaerobic bacteria like *Cutibacterium acnes* (formerly *Propionibacterium acnes*) or *Peptostreptococcus* are more common in shoulder infections due to a proximity to the axilla but have also been reported to cause TKA infections [21, 39]. Suggested systemic substances are Penicillin or Ceftriaxone (Table 12.1). Gram-negative anaerobic bacteria like *Bacteroides* or *Fusobacterium* are usually treated by Ampicillin followed by Metronidazole [16].

12.7.6 Fungi

Fungi generally belong to the DTT group of causative agents [16] with *Candida* spp. making up for more than 80% of this group [40]. Infection with various fungi species are relatively rare but show a rising prevalence due to an increased rate of arthroplasty revision surgery [41]. They are often related to immunosuppression due to concomitant diseases or chronic treatment of medical conditions [40].

Systemic therapy options include Caspofungin, Anidulafungin, and Fluconazole (Table 12.1). Suppressive treatment with Fluconazole is recommended for more than 1 year in total, often even requiring more radical surgical approaches like total implant removal or even amputation due to recurrent infection.

12.8 Suppressive Therapy

In cases of PJI where surgical intervention is impossible (e.g. due to multimorbidity) or in DTT and recurrent infection despite numerous

surgical interventions, chronic suppressive oral antibiotic therapy may be warranted [26, 42]. Such an approach is generally not to be considered therapeutic, but rather symptomatic in order to suppress the infection. More than 80% of cases show a re-infection once the suppressive therapy is discontinued [43].

There is no clear guideline regarding dosage and duration of such a therapy regimen, so all decisions have to be made based on clinical rationale considering the underlying causative agent as well as the patient's individual situation. Besides the persistent suppression of infection, one major concern of a suppressive therapy is the side effects of chronic antibiotic intake (e.g. liver damage, kidney damage, general immunosuppression, resistance formation) as well as further soft tissue damage or bone loss [16]. Appropriate substances for suppressive therapy include Doxycycline and Trimethoprim/Sulfamethoxazole (Cotrimoxazole)—depending on susceptibility testing.

12.9 Conclusion

Multimodal management of PJI usually includes systemic antibiotic therapy as a major component. While an empiric intravenous therapy is usually started around the first surgical intervention, it has to be adapted to culture results throughout the course of treatment. A focused therapy with regard to the individual case should be continued for at least 2 weeks after the last surgical intervention before a conversion to an oral therapy is considered. In general, systemic antibiotic therapy has to be closely tailored to the chosen surgical approach. For this reason, there are no clear guidelines regarding timing and duration of such therapy since it has to be adapted to all concomitant interventions. Dosages of systemic antibiotics may vary with respect to culture results and susceptibility testing. In cases of DTT causative agents like multi-resistant bacteria strains or fungi, treatment can be complicated further, necessitating the use of additional interventions like more radical surgical approaches. Especially in cases of DAIR, one-stage revisions

or re-implantations, the use of biofilm-active antibiotics should be considered to prevent biofilm formation and protect the implant surface. For elderly or multimorbid patients, chronic suppressive oral antibiotic therapy may be an option.

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Local Delivery of Antibiotic and Antiseptic

13

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13.1 Introduction

Total knee arthroplasty (TKA) is considered one of the most cost-effective orthopedic surgical procedures with more than one million procedures performed every year [1]. Periprosthetic joint infection (PJI) is a rare but devastating complication associated with extensive economic, physical, and psychological costs. The number of THA and TKA performed every year has constantly grown during the last decades due to an active aging population [1]; however, PJI rates average between 0.5% and 2% in total joint arthroplasty (TJA) [2], and it is estimated that, in the USA alone, \$1.6 billion will be spent in 2020 on revision TJA for PJI [3]. PJI is considered the most frequent cause of reoperations within 2 years from the index surgery and the second overall cause of reoperations after TKA [4, 5]. Early PJI has been associated with preoperative bacterial infection of the patient, or intraopera-

tive bacterial contamination from surgical team, operative tools, and instruments [6]. Currently, the use of perioperative systemic antibiotics in total joint arthroplasty (TJA) is the only consensus recommendation by international authorities [4, 5]. In order to reduce the incidence of PJI, multiple prevention strategies have been progressively introduced in the preoperative, intraoperative and postoperative phases [7, 8], including operating room ventilation and temperature, body exhaust suits, preoperative patient optimization, perioperative skin preparation and wound management. However, their efficacy has to be proven yet [4, 5].

Despite the growing attention in preventing postoperative infection, the projected increased volume of TJAs ensures that this complication will be encountered with greater frequency in the future [1]. In this chapter, we discuss the use of antiseptic intraoperative irrigation before wound closure, local delivery of antibiotic through powder or beads, and implant coatings, developed in order to prevent and treat infections in primary and revision TKA.

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13.2 Antiseptic Intraoperative Lavage

Wound irrigation during TJA is a routine practice among orthopedic surgeons for preventing PJI. Multiple potential options have been

described including the use of 0.9% saline, castile soap, antibiotic solutions, and antiseptics like povidone-iodine, chlorhexidine gluconate, or hydrogen peroxide. However, no consensus has been reached yet due to a lack of evidence and a paucity of studies in the current literature. Intraoperative irrigation with antiseptic solutions could be considered a potential tool in reducing the risk of early PJI after TKA by preventing the formation of bacterial biofilm [9]. A 2014 focus group discussed evidence for standardization of surgical wound irrigation protocols and determined that, given a lack evidence-based science regarding this topic, they were unable to conclude on which solution, delivery method, or amount should be recommended [10]. However, the United States Center for Disease Control and Prevention guidelines for the prevention of surgical site infection (SSI) [4] and the World Health Organization (WHO) suggest the use of intraoperative irrigation with antiseptic solution considering its effectiveness in reducing the risk of SSI and deep infection [5]. Most of the available data in the current literature comes from in vitro studies, and only few clinical reports have been already published [11].

To date, there is no shared consensus regarding the usage of the best antiseptic solution in order to prevent and/or eradicate biofilm formation [11]. In this section we will review the clinical evidence available on intraoperative lavage with povidone-iodine and chlorhexidine gluconate solutions in TKA.

13.2.1 Povidone-Iodine Intraoperative Lavage

Intraoperative irrigation before wound closure with dilute povidone-iodine solution at different concentration has been associated with a decreased rate of postoperative infection in orthopedic (up to 1% dilution), urologic (1% dilution), cardiovascular (0.5% dilution), and general surgery (1% dilution) [12, 13]. Povidone-iodine is a stable chemical complex of polyvinylpyrrolidone (PVP) and elemental iodine (I) that progressively releases free iodine, a toxic element for microor-

ganisms [14, 15]. Hoekstra et al. [16], in an in vitro study, reported that diluted PVP-I was shown to be equivalent to, and in many cases better than, its competitors in the eradication of bacterial biofilm over 24 h and that was highly effective against *Pseudomonas aeruginosa*, *Candida albicans*, and *Methicillin-Resistant Staphylococcus aureus* (MRSA) at both 4 and 24 h. This highlighted its potential to be used as treatment of choice of highly exuding chronic biofilm-infected wounds. Similarly, Kanno et al. [17] reported that irrigation with 1% diluted PVP-I solution reduced bacterial count on contaminated wound's surface, especially when highly contaminated with *P. aeruginosa*. Despite its effectiveness toward a broad-spectrum of potentially infectious microorganisms, it is important to clarify that PVP-I solution may cause adverse reactions in human tissues. In an in vitro study it was reported that diluted PVP-I irrigation was found to be cytotoxic to bovine articular cartilage cells; however, Von Keudell et al. [18] found this effect to be much less evident in the 0.35% diluted solution in 1- to 3-min incubation periods. In addition, Kaysinger et al. [19] reported that diluted PVP-I solution at a concentration of 5% or greater was cytotoxic to embryo chick tibia and osteoblast cells.

However, only a few studies reported the outcomes of intraoperative irrigation with diluted PVP-I before wound closure in TKA [20–22]. Brown et al. [20] retrospectively analyzed 2250 primary TJA and compared the outcomes of 414 TKA and 274 total hip arthroplasty (THA) where 3 min irrigation with 0.35% dilute povidone-iodine solution was performed, followed by skin disinfection with 10% povidone-iodine solution prior to the final wound closure, with the outcomes of 1862 TJA where dilute povidone-iodine solution was not used (1232 TKA, 630 THA). The authors reported 18 early postoperative infections among the cases where irrigation with dilute PVP-I was not performed and 1 in the PVP-I irrigation group (0.97% and 0.15%, respectively; $p = 0.04$). The authors stated that a 3-min dilute PVP-I lavage combined with disinfection of the skin with 10% Betadine before surgical closure was associated

Table 13.1 Summary of orthopedic literature on the use of irrigation solutions containing povidone-iodine and/or chlorhexidine gluconate to prevent SSI

| Author | Joint | N | Comparison | Solution | Outcome/% | p-value | |
|-------------------------|--------------|-----------------------|------------------------|---------------------|--|-----------------------------|-------------|
| Brown et al. (2012) | Hip and knee | 274 THAs and 414 TKAs | 630 THAs and 1232 TKAs | 0.35% diluted PVP-I | Acute deep infection | 0.15% vs. 0.97 | 0.04 |
| Frish et al. (2017) | Knee and hip | 386 TJAs | 664 TJAs | 0.05% CHG solution | Surgical site infection/deep infection | 0.8% and 0.7%/1.2% vs. 0.8% | 0.913/0.534 |
| Hernandez et al. (2019) | Knee | 2410 TKAs | 3794 TKAs | 0.25% diluted PVP-I | Reoperation for infection | 0.8% vs. 0.5% | 0.525 |
| Calkins et al. (2019) | Knee and hip | 81 THAs and 153 TKAs | 79 THAs and 144 TKAs | 0.35% diluted PVP-I | Acute deep infection | 0.4% and 3.4% | 0.038 |

THA total hip arthroplasty, TKA total knee arthroplasty, PVP-I polyvinylpyrrolidone-iodine, SSI surgical site infection

with a significant reduction in the infection rate after primary TKA and THA. Hernandez et al. [21], in a register-based study, retrospectively analyzed 6204 primary TKA and reported the outcomes of 2410 TKA where a 0.25% diluted PVP-I irrigation was performed before wound closure, compared with 3794 TKA where the irrigation was performed with normal saline solution. The authors reported that no significant differences between the two groups were observed regarding reoperation rate at 3-months follow-up (0.8% vs. 0.3%; $p = 0.06$), and a higher reoperation rate was observed in the PVP-I group at 1-year follow-up (1.2% vs. 0.6%; $p = 0.03$). However, no differences were reported regarding reoperations due to infection after the application of propensity score. Calkins et al. [22] analyzed the outcomes of 478 patients who underwent aseptic revision TKA and THA and that were randomized to receive a 3-min dilute PVP-I lavage (0.35%) or normal saline lavage before surgical wound closure. Among them, 234 patients (153 knees, 81 hips) received normal saline lavage and 223 (144 knees, 79 hips) received dilute PVP-I lavage. Within 90 days postoperatively, the authors reported eight infections in the saline group and one in the PVP-I group (3.4% vs. 0.4%, $p = 0.038$) and no difference in wound complications between groups (1.3% vs. 0%, $p = 0.248$).

In addition, PVP-I is safe, inexpensive, simple to use, and it has a broad-spectrum bactericidal activity that includes MRSA [15, 23].

Finally, based on the current literature, dilute PVP-I irrigation before surgical wound closure in primary and revision TJA appears to be a simple, safe, and effective option to reduce the risk of acute postoperative PJI (Table 13.1).

13.2.2 Chlorhexidine Intraoperative Lavage

Chlorhexidine gluconate (CHG) is a widely used antiseptic agent and is present in a variety of preparations to prevent infection, including preoperative skin cleaning, surgical site preparation, intraoperative irrigation, CHG impregnated postoperative dressings, and hand antisepsis [24]. CHG has a broad-spectrum biocide bacteriostatic and bactericidal effect against Gram-Positive and Gram-Negative bacteria, and it has a faster onset of action than PVP-I [25]. It is a bactericidal agent, acting primarily disrupting the cell membrane [26, 27]. In addition, CHG has a particularly strong affinity for binding to skin and mucous membranes, theoretically enhancing the efficacy in prevention of SSI [28]. It has been previously stated in several animal studies that CHG could be safely used on wounds, and its potential use for wound lavage has been demonstrated by the studies on prevention of infection in humans [29]. In addition, it is considered effective in biofilm eradication when used to scrub an MRSA-coated titanium disc [30]. However, in an in vitro study, it has been reported

that the clinically used concentration of CHX (2%) permanently halts cell migration and significantly reduces survival of fibroblasts, myoblasts, and osteoblasts, regardless of the exposure duration [31]. Nevertheless, to date there is a paucity of literature regarding the safety of CHG as an intrawound irrigation agent and peri-incisional topical antiseptic.

Despite many studies have reported the efficacy of CHG in skin preparation before total knee arthroplasty [24, 32], there is currently only one study that reported the outcomes of intraoperative CHG lavage in TJAs. Frisch et al. [24] evaluated the effect of CHG intraoperative irrigation on infection rates following THA and TKA. Intraoperative irrigation was performed with 0.9% saline and periodic 0.05% CHG solution followed by a final 1-min soak in CHG with immediate closure afterward. The authors reported no significant differences in terms of SSI ($p = 0.913$) and deep infections at 1-year follow-up ($p = 0.534$) when compared 411 TKAs where intraoperative irrigation was performed with normal saline solution before wound closure, with 248 TKAs where intraoperative irrigation was performed with CHG solution. The authors suggested that intraoperative CHG during TJA had a comparable infection rate to different protocols using PVP-I in THA and 0.9% saline in TKA.

Intraoperative irrigation with diluted antiseptic solutions like PVP-I or CHG before wound closure may contribute to the prevention of the biofilm formation and reduce the incidence of early PJI in TKA. However, despite promising in vitro evidence, further in vivo studies are required to examine and optimize safety and efficacy when intraoperatively applied before wound closure (Table 13.1).

13.3 Antibiotic Local Delivery

The application of antimicrobial agents at the site of musculoskeletal infections has been widely documented, ranging from the direct intra-articular infusion of antibiotic after TKA [33] to the intrawound placement of antibiotic powder to

prevent infection in spinal surgery [34–40]. Due to a lack of clarification regarding the long-term efficacy of locally administered antibiotics, the combination of antibiotics with implantable materials has been progressively investigated in order to provide a predictable release profile [41].

13.3.1 Antibiotic Powder

Intrawound vancomycin powder (VP) was recently considered in orthopedics to decrease SSIs and subsequent deep infections for its capacity to provide a high local concentration of the antibiotic, maximizing local bactericidal effect while minimizing adverse systemic reactions. Previous reports showed that intrawound VP did not increase the rate of side effects [34–38] and that after local administration, serum vancomycin concentrations remained below toxic levels [35–37].

The use of VP in spinal surgery has been widely documented by several reports indicating a reduced rate of SSIs [34–37, 39, 40]; however, Ghobrial et al. [34] reported that wound complications, including seroma formation, were associated with intrawound VP after spinal surgery. Despite the promising results of VP in spinal surgery, information regarding its use in TJA are still not clear. In front of a lack of clinical data, pre-clinical results have been supported by Cavanaugh et al. [42] and Edelstein et al. [43], suggesting the effectiveness of intrawound VP on clearing *S. aureus* from contaminated femoral implants in in vivo rats investigations. However, numerous questions are still unanswered regarding this procedure in TJA, including clear information about seroma formation, bearing wear, nephrotoxicity, and ototoxicity. Third body wear remains a real concern in the setting of TJA. Although vancomycin is a soluble molecule, there is a paucity of information about its use outside of plasma and about its solubility and possibility to precipitate in other body fluids. In a closed space such as knee or hip joint, with a prosthetic implant, undissolved particulate of antibiotic could reach portions of the prosthetic implant and cause abnormal wear and potentially early failure.

Quadir et al. [44], in a biomechanical study, demonstrated that crystalline antibiotics do not alter wear rates in Cobalt-Chrome (Co-Cr) on ultra-high molecular weight polyethylene (UHMWPE) secondary to third body wear in simulated ten million cycles. However, the long-term effect on polyethylene wear is still unknown.

Otte et al. [45] retrospectively compared over a 2-year period the rate of early PJI in patients who underwent primary or revision THA and TKA with and without the use of intrawound VP. The authors reported a significant decrease in the early PJI rate in the revision settings (THA and TKA) when VP was used (0 of 134, 0%) compared to when VP was not used (7 of 180, 3.89%; $p = 0.0217$). Similarly, Patel et al. [46] retrospectively reviewed 460 primary THAs and TKAs and compared the early PJI rate in the VP group ($n = 348$) with the control group ($n = 112$). The authors reported a decreased overall infection rate (0.57% vs. 2.7%; $p = 0.031$) and PJI rate (0.29% vs. 2.7%; $p = 0.009$) in the VP group compared to the control group with a lower readmission rate due to infection (0.57% vs. 2.7%; $p = 0.031$). In addition, the authors determined a number needed to treat (NNT) of 47.5, suggesting that the cost to prevent 1 infection with the addition of intrawound vancomycin was \$816 (based on their institution costs) compared to the estimated average hospital cost per case in the USA of infected THA (\$30,329) and TKA (\$25,155) [47]. Matziolis et al. [48] retrospectively evaluated 8945 primary TJA and reported two infections among the TKA treated with intraoperative intrawound VP group (out of 650 TKAs, 0.4%) compared to 44 infections (out of 3471 TKAs, 1.3%; $p = 0.033$) in a control group. The differences noted among the two groups of patients who underwent THA did not achieve statistical significance; however, the infection rate in the control group was twofold greater compared to the VP group, and no wound complications were observed as a result of application of local vancomycin. Conversely, Dial et al. [38] retrospectively reviewed 265 consecutive THAs and despite a reduced deep infection rate when intrawound VP was used (0.7%) compared to the control group (5.5%, $p = 0.031$), the authors

reported an increased rate of sterile wound complications in the VP group (4.4% vs. 0%; $p = 0.030$). Similarly, Hanada et al. [6] prospectively evaluated 166 consecutive patients who underwent primary TKA or unicompartmental knee arthroplasty (UKA) and evaluated the efficacy and side effects of local intrawound VP. Despite a considerably high PJI rate in both groups (7.6% control group; 4.5% VP group), no significant difference was found among them (ns). However, operative wound complications were significantly more frequent in the VP group (11.8%) compared to the control group (4.3%) so that the authors did not recommend its use in the setting of primary TKA and UKA.

In conclusion, despite a paucity of available data in the current literature and contrasting opinion by authors, intrawound VP shows promising results in reducing early PJI, particularly if an increased rate of PJI is present due to the surgical procedure or the high-risk population involved. In addition, VP is a low-cost tool with an effective NNT (Table 13.2).

13.3.2 Nonabsorbable Polymethylmethacrylate (PMMA) Beads

Antibiotics have been used in combination with poly-methyl-methacrylate (PMMA) for decades [49], and it has been widely used as a fundamental tool in primary TKA in case of patients at high risk of infection or in revision TKA for PJI for cementing the implant components, as antibiotic-loaded articulated spacers or beads [50, 51]. The use of antibiotic-loaded cement has reported a significant reduction in infection rates in the settings of primary or revision THA [52] and TKA [53]. However, other studies reported limited clinical benefit of antibiotic-loaded cement in the treatment of PJI compared with systemic antibiotics and clinical data are not sufficient to support recommendations on dosages [54, 55]. In this paragraph we will discuss the role of antibiotic beads in TJA.

Gentamicin, vancomycin, and tobramycin-loaded PMMA beads are considered an effective

Table 13.2 Summary of orthopedic literature on the use of intrawound vancomycin powder and/or calcium sulfate beads to prevent SSI

| Author | Joint | N | Comparison | Local antibiotic | Outcome/% | p-value | |
|-------------------------|--------------|----------|------------------|-----------------------|-------------------------------------|-------------------------------------|-------------|
| Otte et al. (2017) | Knee and hip | 816 TJAs | 824 TJAs | Intrawound VP | Early PJI | 0% vs. 3.89% | 0.0217 |
| Flierl et al. (2017) | Hip and knee | 32 TJAs | No control group | Calcium sulfate beads | PJI | 48% | / |
| Dial et al. (2018) | Hip | 137 THAs | 128 THAs | Intrawound VP | PJI | 0.7% vs. 5.5% | 0.031 |
| Patel et al. (2018) | Knee and hip | 348 TJAs | 1122 TJAs | Intrawound VP | Overall infection rate/ PJI rate | 0.57% vs. 2.7%/0.29% vs. 2.7% | 0.031/0.009 |
| Lum et al. (2018) | Knee and hip | 56 TJAs | No control group | Calcium sulfate beads | PJI | 0% | / |
| Calanna et al. (2019) | Knee | 10 TKAs | No control group | Calcium sulfate beads | PJI | 20% | / |
| Gramlich et al. (2019) | Knee | 42 TKAs | No control group | Calcium sulfate beads | PJI | 26.2% | / |
| Hanada et al. (2019) | Knee | 92 TKAs | 90 TKAs | Intrawound VP | PJI | 4.5% vs. 7.6% | NS |
| Matziolis et al. (2020) | Knee | 650 TKAs | 3471 TKAs | Intrawound VP | PJI | 0.4% vs. 1.3% | 0.033 |

TKA total knee arthroplasty, *THA* total hip arthroplasty, *TJA* total joint arthroplasty, *VP* vancomycin powder, *PJI* periprosthetic joint infection, *SSI* surgical site infection, *NS* not significant

drug delivery system for local antibiotic therapy in bone and soft-tissue infections with subsequent antibiotic concentrations above the minimum inhibitory concentration (MIC) for the infecting organisms [56]. The release of the antibiotics from the PMMA beads is a diffusion process, as in all antibiotic-loaded bone cements [57]. However, due to the increased surface area of the many, relatively small beads, much more antibiotic is released compared to solid bone cement plugs. Usually, gentamicin-loaded beads are held in situ approximately 14 days, after which 20–70% of the total amount of gentamicin has been released into the body, so that the main antibiotic efficacy is immediately after implantation [58]. Despite the multiple advantages, once the antibiotics have eluted from the nonabsorbable cement, the surface becomes a foreign body that is potentially subjected to bacterial colonization and biofilm formation [59, 60]. Neut et al. [61] analyzed in an extensive laboratory procedure the gentamicin-loaded beads from 20 patients treated for PJI and reported the presence of bacteria on the beads in 18 of the 20 patients involved, and that 19 of 28 bacterial strains iso-

lated were gentamicin-resistant or highly resistant sub-populations. The authors suggested that despite their antibiotic release, the PMMA beads act as a biomaterial surface to which bacteria preferentially adhere, grow, and potentially develop antibiotic resistance. In a retrieval analysis of gentamicin-loaded beads left in situ for 5 years, it was reported that the gentamicin-release test revealed residual antibiotic release, and extensive microbiological sampling resulted in recovery of a gentamicin-resistant staphylococcal strain from the bead surface [62]. This case showed that even after 5 years, PMMA beads remained able to release subinhibitory concentration of antibiotics, approximately 0.4 mg of gentamicin per bead, stimulating the introduction of gentamicin-resistant strains. Finally, considering that every biomaterial left in the human body must be considered as a potential focus for infection [60], biodegradable beads are preferred as carriers for antibiotics as they do not show long-term release of subinhibitory antibiotic concentrations, do not require removal, and do not leave any biomaterial in situ to act as a potential focus for infection. In addition, in vitro results

have shown that tobramycin impregnated beads made of polycaprolactone, a bioresorbable polymer, have even superior antibiotic elution characteristics compared with PMMA beads, suggesting more effective antibiotic delivery vehicle [63].

13.3.3 Absorbable Calcium Sulfate Beads

Calcium sulfate (CS), ($\text{CaSO}_4 \cdot 1/2 \text{H}_2\text{O}$), also known as plaster of Paris, was introduced in orthopedic surgery in 1892 by Dreesman et al. as a filler for bone void [64]. Currently, antibiotic bone substitution materials/bone void fillers are based on biodegradable or resorbable materials such as polylactic acid, chitosan, or new combinations based on calcium sulfate [65]. Absorbable mineral-based bone cements, despite inferior mechanical characteristics compared to acrylic cements, provide multiple advantages regarding antibiotic delivery and infection control. Unlike PMMA beads, these materials do not have to be removed, they have the capacity to accommodate a wider range of antibiotics since there is little temperature increase during setting, and the antibiotics are slowly released meanwhile the material dissolves [59]. CS beads are suitable for application in the presence of infection, non-union, or bone loss, and as the beads are absorbed, CS releases 100% of the antibiotic load, resulting in superior elution characteristics and higher sustained antibiotic concentrations over several weeks [41].

CS is a well-studied, non-immunogenic, biocompatible bone void filler used in orthopedic applications since the nineteenth century and currently used as substitute for bone cement in multiple settings of orthopedic surgery [66–72]. To date, multiple studies reported the outcome of antibiotic-loaded CS beads in the treatment of chronic osteomyelitis of long bone [73, 74]. McKee et al. [75] reported that tobramycin-loaded CS beads were as effective as PMMA beads in the treatment of chronic osteomyelitis and infected non-unions. Despite promising results, occasionally, non-infectious inflammatory reactions have been observed after the

implantation of resorbable beads, suspected to be caused by calcium-rich fluid generated in the process of rapid graft resorption [76, 77]. Currently there are only a few studies that evaluated the outcomes of CS beads in TKA. Three studies reported outcomes after debridement, antibiotics, and implant retention (DAIR) procedure with antibiotic-loaded CS beads, and two studies where the beads were used in the setting of two-stage revisions for PJI [78–80]. A DAIR procedure is usually performed for acute infections without complicating factors such as significant comorbidity or implant loosening. Antibiotic-loaded CS beads are available in three commercial products, Stimulan® (Biocomposite Ltd., Staffordshire, England), OSTEOSET®-T (Wright Medical Technology Inc., Arlington, TN, USA), and Herafill® beads G (Heraeus Medical GmbH, Wehrheim, Germany).

Calanna et al. [81] described a modified surgical technique called debridement, antibiotic pearls, and retention of the implant (DAPRI). In order to reduce the risk of persistent infection, they performed a methylene-blue guided debridement, defined as “tumor like,” synovectomy, followed by Argon beam electrical stimulation of metallic surface, and by 4% dilute CHG irrigation. They added the CS beads loaded with vancomycin, tobramycin, and a third antibiotic based on preoperative antibiogram in the suprapatellar pouch, around the proximal tibia and the distal femur. The authors finally reported, at a mean follow-up of 24 months, that the procedure was considered a failure in 2 of 10 (20%) patients. Similarly, Flierl et al. [78] retrospectively evaluated at a mean follow-up of 13 months 32 patients (27 TKAs, 6 THAs) with acute hematogenous or acute postoperative PJI who underwent irrigation and debridement with implant retention and addition of antibiotic-impregnated CS beads. The authors reported an overall failure rate of 48%, in addition, acute hematogenous and acute postoperative PJI had similar failure rates of 47% and 50%, respectively ($p = 0.88$), suggesting that the addition of antibiotic-impregnated CS beads did not improve outcomes of DAIR in the setting of acute hematogenous or acute postoperative PJI. Kallala et al. [79] prospectively evaluated at

a mean follow-up of 35 months, the outcomes of 755 patients who underwent 456 revision TKAs and 299 revision THAs. The procedures included one-stage revisions, the first or second stage of two-stage revisions, and DAIR with the implantation of Stimulan beads. The first of stage of the two-stage revision included washout, debridement, components removal, and implantation of PMMA spacer and antibiotic-impregnated CS beads. The second stage included spacer removal and implantation of revision components followed by antibiotic-impregnated CS beads. The authors found no significant difference in beads volume between patient drainage groups ($p > 0.05$). In addition, they found an overall difference in the volume of the beads involved in variable types of complication, with a larger volume in the group with hypercalcemia compared to patients without complications ($p = 0.0014$). It is reported in the literature that wound drainage tends to occur more frequently in patients in whom a higher volume of beads had been used, with more subcutaneous placement and in those with a poor host grade, such as McPherson grade C [82]. Gramlich et al. [80] evaluated at a mean of 23-months follow-up the outcomes of 42 patients treated using a single-stage algorithm consisting of DAIR, followed by implantation of antibiotic-loaded beads chosen in accordance with an antibiogram (OSTEOSET-T® and Herafill-Gentamycin®). The authors reported that permanent remission was achieved in 73.8% of the cases, while 11.9% showed chronic PJI under implant retention, suggesting good outcomes of DAIR and antibiotic-loaded CS beads in patients with recurrent PJI where DAIR is typically considered inappropriate. Marczak et al. [83] evaluated the outcomes at mean 52-months follow-up of two groups formed by 28 consecutive patients who underwent two-stage revision TKA for PJI, one group received Herafill beads, while the control group did not. The authors reported no cases of reinfections in the study group, while five were seen in the control group. No other differences were observed between the two groups, and no side effects related to the use of Herafill were noted. Lum et al. [84] evaluated postoperative complications following the use of antibiotic-

loaded CS beads in 56 patients who underwent complex primary or revision hip or knee arthroplasty (26 knees and 30 hips). The authors reported one case (1.7%) of persistent wound drainage in a revision TKA that required subsequent surgical irrigation and a poly-exchange, and no postoperative infections were seen suggesting that CS beads may help to reduce postoperative wound complications and may be a safe adjunct tool in local antibiotic delivery.

The use of antibiotic-loaded beads for the treatment of PJI in TKA is reported in a limited number of studies in the current literature. Given the lack of evidence, the second International Consensus Meeting on PJI did not recommend the use of calcium sulfate/phosphate or PMMA beads, as local antibiotic carrier to prevent surgical site infection and PJI [85]. In addition, despite the encouraging clinical results in reducing the incidence of PJI, CS beads have been associated with multiple complications including hypercalcemia, persistent wound discharge, and heterotopic ossification [79, 86]. However, degradable, local antibiotics based on calcium sulfate could offer advantages and can be a reasonable addition to already established systems in the treatment of PJI (Table 13.2).

13.4 Coated Implant

Different implant-coating alternatives have been developed to reduce the risk of early PJI. The goal is to create a local environment favorable to the host and hostile to the microorganisms in order to reduce the bacterial adhesion to the implant and the subsequent biofilm formation. According to Romanò et al. [87], antibacterial coatings have been classified by their mechanism of action in passive surface, active surface finishing/modification, and perioperative antibacterial local carriers or coatings. The first one is based on preventing or reducing bacterial adhesion to implants through surface chemistry and/or structure modifications, without the use of any pharmacologically active substance. Examples of this approach include modified titanium dioxide surface or polymer coatings. The second one is

based on pharmacologically active pre-incorporated bactericidal agents, such as antibiotics, antiseptics, metal ions, or other organic and inorganic substances that are actively released from the implant in order to reduce bacterial adhesion. Examples of this approach are “contact killing” active surface with silver- or iodine-coated joint implants. The third one is based on local antibacterial carriers, or coatings, that are not built into the device, but rather are applied during surgery, prior to the insertion of the implant. Those carriers or coatings may have direct or synergistic antibacterial/anti-adhesive activity or may deliver high local concentrations of loaded antibiotics or antibacterial.

13.4.1 Silver-Coated Implant

Silver is considered a promising coating as it has a broad-spectrum of antibacterial activity against planktonic and sessile Gram-positive and Gram-negative bacteria, including multiresistant bacteria [88, 89]. The bactericidal ability of silver depends on the capacity of dissolved cations to interfere with bacterial cell membrane and bacterial metabolism. In addition, silver cations in an aqueous medium contribute to the formation of reactive oxygen species that potentially harm prokaryotic cells [87].

Calcium phosphates like hydroxyapatite containing silver have shown to reduce the bacterial adhesion against *S. epidermidis*, *P. aeruginosa*, and *S. aureus* when compared to surfaces without silver [90, 91]. In addition, silver deposited on surfaces such as titanium and stainless steel has shown a toxic effect toward bacterial pathogens within specified doses of silver [92–95]. Currently there are only few studies that reported the outcomes of patients that underwent surgeries with silver-coated implants [95–97]. Hardes et al. [95] prospectively evaluated over a 5-year period the infection rate in 51 patients with sarcoma of proximal femur or proximal tibia who received a silver-coated megaprosthesis compared with 74 patients who received an uncoated titanium megaprosthesis was implanted. The authors reported a substantial reduction of the infection

rate from 17.6% in the titanium group to 5.9% in the silver-coated group ($p = 0.062$). In a subsequent study, the same authors [96] assessed the infection rate in 98 patients with sarcoma or giant-cell tumor of the proximal tibia who underwent placement of a titanium uncoated ($n = 42$) or silver-coated ($n = 56$) megaprosthesis. The authors reported an infection rate of 16.7% in the titanium group compared to 8.9% in the silver-coated group ($p = 0.247$), resulting in 5-year survivorship of the implants of 90% and 84% in the silver and uncoated titanium group, respectively. Zajonz et al. [97] retrospectively evaluated the reinfection rate of 34 patients treated with modular mega-endoprosthesis after a cured bone infection of the lower limb (femur or tibia). The authors reported, over a median follow-up of 72 months, a reinfection rate of 40% in the silver-coated group (8 of 20) and of 57% in the non-silver-coated group (8 of 14). However, these results were not statistically significant due to the low number of cases. Wafa et al. [98] retrospectively evaluated the outcomes of silver-coated tumor prostheses in 85 patients compared with 85 matched control patients treated for primary reconstruction (30%), one-stage revision (47%), and two-stage revision for infection (23%). At a minimum follow-up of 12 months there was a significant reduction in the overall postoperative infection rate from 22.4% to 11.8% ($p = 0.03$) in favor of the silver-coated implant group.

Despite this broad clinical use, little is known about the stability of silver-coated alloys, their efficacy on biofilm formation, and the kinetics of release. The main concerns about the use of silver-coated implants are directed toward the toxicity of silver ions. The same effects on prokaryotic cells could apply to eukaryotic cells leading to toxicity on bone cells, and the silver ions released could produce adverse reactions by accumulating in further districts within the body [99]. In addition, silver has a wide range of bacterial targets [100, 101], including the respiratory chain, and has been shown to induce resistance in Gram-negative bacteria and toxicity in eukaryotic cells [102, 103]. Moreover, concern is expressed toward the incomplete protection of the implant, since some modular components of

the implants cannot be coated [87], and only a few implant designs are offered with silver-coating given a persistent relatively high cost of the technology when used outside oncology [104].

13.4.2 Iodine-Coated Implant

The use of PVP-I as an electrolyte was reported by Shirai et al. [105], in an in vitro study, and resulted in the formation of an adhesive porous anodic oxide with the antiseptic properties of iodine, suggesting the antibacterial attachment effect and cytocompatibility of iodine-supported implants. Similarly, Inoue et al. [106] showed that iodine-supported implants have a good antibacterial attachment effect in vivo and inhibit biofilm formation and growth by preventing initial bacterial attachment on the metal surface. Tsuchiya et al. [107] prospectively evaluated at a mean follow-up of 18 months 222 patients with postoperative infection or compromised status that were treated using a variety of iodine-supported titanium implants (spinal instrumentation, plates for osteosynthesis, pins, and wires). In 158 patients the iodine-supported implants were used to prevent infections while in 64 patients to treat active infections. The authors reported that acute infection developed in three tumor cases among the 158 patients on preventive therapy, and that infection was eradicated in all 64 infected patients suggesting that iodine-supported titanium implants can be very effective in preventing and treating infections after orthopedic surgery. Similarly, Shirai et al. [108] evaluated at a mean follow-up of 30 months 47 patients with malignant bone tumor or pyogenic arthritis treated with iodine-supported titanium megaprosthesis. The authors reported that only one patient (out of the 21) got infected and that in the 26 patients treated with one- or two-stage revision surgery, infection was eradicated without any additional surgery. In addition, Kabata et al. [109] retrospectively evaluated at a mean follow-up of 33 months the outcomes of a consecutive series of 30 hips including 13 primary THAs in compromised

immune system conditions or pyogenic arthritis, 14 revision THAs after PJI, and 3 conversions from hemiarthroplasty to THA in immunosuppressive conditions. The authors reported no signs of infection in any patient at the latest follow-up.

Finally, based on these findings, iodine-supported implants can be considered highly effective in preventing and treating postoperative infection while no adverse event has been reported to date. However, longer-term effects of local application of iodine coating and the effects on materials other than titanium have not been clarified yet and clinical trials are currently ongoing in order to confirm these preliminary results.

13.4.3 DAC® Hydrogel Coated Implant

Defensive Antibacterial Coating (DAC®) is a fast-resorbable hydrogel coating composed of covalently linked hyaluronan and poly-D,L-lactide (PLLA) (Novagenit Srl, Mezzolombardo, Italy) specifically designed to protect implanted biomaterials [110]. The rationale of this device is the capacity of hyaluronic acid to reduce biofilm formation on material surfaces exposed to bacterial contamination and affect different microbial species and, sometimes, different strains belonging to the same species [108, 109]. DAC® has been found to have a synergistic antibiofilm activity with various antibacterials and to be effectively manually spread onto the surface of various biomaterials commonly used in orthopedics, trauma, and dental surgery [111]. The adhesion density of *S. aureus* on titanium discs pre-treated with DAC® was significantly lower than adhesion on untreated controls at each time point. In particular, reductions of adhered bacteria equal to 86.8%, 80.4%, 74.6%, and 66.7% vs. untreated discs were observed after 15, 30, 60, and 120 min of incubation, respectively, while an increase of adhesion density during time was observed for both control and pre-treated discs [112]. In addition, DAC® hydrogel showed similar or superior in vitro activity, compared to gentamy-

cin and vancomycin, and a synergistic activity when used in combination with antibiotics providing a larger reduction of biofilm formation (approximately 75 to 80% in comparison with untreated controls) [113]. It has been reported the capacity of DAC[®] to entrap different antibacterial agents at concentrations ranging from 2% to 10% and then to slowly release them locally for up to 72 h at levels considerably higher than the minimum inhibitory concentration (MIC) [114]. Romanò et al. [115] evaluated 380 patients who underwent cementless or hybrid fixation primary ($n = 270$) or revision ($n = 110$) THA ($n = 298$) and TKA ($n = 82$) with and without the antibiotic-loaded DAC[®] coating, in a multicenter randomized prospective study, at a mean follow-up of 15 months. The authors reported 11 early surgical site infections in the non-coated group (6%) and only one in the coated group (0.6%, $p = 0.003$). No local or systemic side effects related to the DAC[®] hydrogel coating were observed, and no detectable interference with implant fixation was noted. Similarly, Malizos et al. [116], in a multicenter randomized controlled prospective study, evaluated at a mean follow-up of 18 months 256 patients who underwent osteosynthesis for a closed fracture and were randomly assigned to receive implants with antibiotic-loaded DAC[®] coating or without coating. The authors reported six surgical site infections in the coated group (4.6%), compared to none in the non-coated group ($p < 0.03$). No local or systemic side effects related to the DAC[®] hydrogel coating were observed, and no detectable interference with bone healing was noted. Recently, Capuano et al. [117] retrospectively evaluated at a mean follow-up of 29 months 22 patients treated with a one-stage revision for PJI using implants coated with an antibiotic-loaded DAC[®] hydrogel, and compared them with 22 matched controls treated with a two-stage revision procedure using non-coated implants. The authors reported, although in a relatively limited series of patients, a similar infection recurrence rate after one-stage exchange with DAC[®]-coated implants (9%) compared to two-stage revision

without coating (14%), with reduced overall hospitalization time and antibiotic treatment duration. Zagra et al. [118] retrospectively evaluated at a mean follow-up of 2.8 years 27 patients who underwent a two-stage revision THA for PJI, using cementless implants coated with the antibiotic-loaded DAC[®] hydrogel, and compared them with 27 matched controls, who underwent a two-stage cementless revision THA with non-coated implants. The authors reported no evidence of infection, implant loosening, or adverse events in the DAC[®]-coated group, compared to four cases of infection recurrence in the non-coated group ($p = 0.11$).

In conclusion, despite these encouraging results, longer-term data are required in order to evaluate the incidence of delayed or late PJIs. In fact, the fast resorption of the hydrogel protects from long-term side effect but may limit the protection of the implant from late, hematogenous infections.

13.5 Conclusions

In conclusion, implant-related infections have a pronounced social and economic impact with increased rates of morbidity and mortality after THA and TKA [119, 120]. According to the current literature these complications will become a growing burden to healthcare systems over the coming decades unless novel and effective measures are not taken to reduce the incidence of PJIs [1]. Despite the promising results of the newer technologies, only a few of them are currently available in orthopedic surgery. Some potentially effective solutions may be excluded from the daily practice due to cytotoxicity, immunoreactivity, or interference with bone healing and osseointegration. Conversely, other technologies safely tested in vitro and in vivo may not be able to be used on a large scale, due to biotechnological, economic, and regulatory issues. Finally, effort should be made in order to increase the awareness of healthcare providers and their patients regarding the newer technologies and their possible contribution to mitigate septic complication.

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Georgi P. Georgiev

14.1 Introduction

Nowadays, total knee arthroplasty (TKA) is performed after failure of non-operative treatment in cases of chronic pain in knee osteoarthritis and cases with significant knee deformities; usually, this procedure has significant success in pain relief and patient satisfaction [1]. However, following TKA, numerous complications have been reported in the literature; they include aseptic implant loosening with significant osteolysis, polyethylene wear, ligamentous laxity, periprosthetic fracture, arthrofibrosis, patellofemoral complications, and infections. Thus, revision surgery becomes necessary [2].

In revision TKA (rTKA), wound closure might be difficult. Precise anatomical knowledge of the knee and the appropriate surgical exposure could reduce the risk of complications and could dispose in good functional outcomes. The ideal approach ensures easy and straightforward joint exposure and facilitates the revision with minimum complication rates. Therefore, detailed knowledge of surgical approaches to the knee is essential. The surgeon should have various options in mind, even in cases when simple exci-

sion of previous scars ensures excellent visualization of the knee [3].

The preoperative planning of the surgical approach is crucial for rTKA. The correct approach depends on the selection of implants and allows for their optimal position and precise ligament balancing. After previous TKA, especially in an infected knee, subsequent approaches are impeded by scar formation in the deep tissue layers and the poor elasticity of the infected tissues. Existing debris and trauma of surrounding tissues due to instability additionally degrade tissue properties [4].

During rTKA, two essential rules should be borne in mind by the surgeon: safe approach and precise surgical technique during managing soft tissue flaps. At revision surgery, a medial parapatellar approach (MPA) with synovectomy, a quadriceps snip, a tibial tubercle osteotomy (TTO), a V-Y quadricepsplasty, a femoral peel (FP), and a medial epicondylar osteotomy (MEO) have been proposed [5]. Till now, no prospective randomized studies have presented and compared the results after different approaches for knee revision arthroplasty [4].

Detailed knowledge of the different surgical approaches described and discussed in this chapter, in the author's opinion, will be of help to knee surgeons in their works, as well as to the other colleagues who will perform this surgery in their future practice. The aim is to briefly summarize the characteristic anatomy of the area, discuss the

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possible complications and how to avoid them, and present the surgical approaches in detail. This information could be important in preventing damage to anatomical structures, especially in infected tissues, impeding surgical dissection.

14.2 Skin Incisions

The skin incision and approach should be considered to fully visualize the operated joint without excessive tension to the skin edges. However, before performing the incision, precise knowledge of the skin arterial blood supply, as well as the surrounding anatomical structures around the joint is mandatory. Detailed knowledge of anatomy will reduce the risk of possible iatrogenic damage during surgery and help avoid possible future complications.

The blood supply to the skin and the surrounding tissues around the knee is from the peripatellar arterial ring. This ring is formed by the supreme genicular, medial/lateral superior genicular, medial/lateral inferior genicular, and anterior tibial recurrent arteries [6]. The skin vascular supply is mainly ensured by subfascial arterioles that start from the medial side of the joint. Therefore, during the surgical approach, their protection without additional dissection is crucial. It should be pointed out that cutaneous blood supply may be disturbed after previous surgery, in rheumatoid or diabetic patients, after prolonged steroid/NSAIDs therapy, in cases of extra obesity, and older smokers [7].

A midline longitudinal incision is preferred because it better preserves the arterial supply of the skin. A large lateral flap should be avoided to minimize the complications in wound healing. This corresponds to the report of Johnson et al. [8], in which they established lower skin oxygenation of the lateral flap. Aso et al. [9], in midline skin incisions below 12 cm, did not establish any differences in oxygenation of the skin and pointed out that hypoxia was significant in the distal edges and might be due to excessive retraction during operation.

A straight longitudinal incision starts 6–12 cm proximally to the proximal border of

the patella, passes over its midpoint, and reaches the medial border of the tibial tuberosity [7]. As an alternative, the incision could make a gentle medial curve over the patella [10]. Of course, the extent of the incision reflects the surgery requirements. As mentioned above, the skin is supplied by perforating arteries running through the fascia; therefore, a soft-tissue flap should be developed deep into the fascia to avoid skin necrosis [7]. The incision is extended through the underlying tissues to ensure enough skin flaps superficial to the extensor mechanism. With a proper skin incision, the surgeon reduces skin retraction and the risk of postoperative necrosis [11].

In the case of a previous anterior scar in a proper position, it should be incorporated in the new approach. In cases of multiple previous incisions, the most lateral should be preferred with full-thickness flaps [12]. Khan et al. [13] recommend the most recent, longitudinal, and lateral skin incision to be used after a prior TKA [13]. Daines [14] advocates a midline approach if multiple scars existed from surgeries a long time ago. In extensive fibrosis, the extension of the incision into normal tissue helps in deep dissection. Previous transverse approaches need to be crossed perpendicularly, and a new incision with an angle of below 60° to a previous scar is not recommended [15]. Windsor et al. [15] considered that skin bridges need to be over 7 cm wide to reduce the risk of flap necrosis. If the surgeon needs another new approach, a safe distance between the incisions should be ensured. Thienpont [4] points out that a skin bridge of at least 2.5 cm to 8 cm is needed.

In summary, a previous skin incision should be used if possible, except in cases of direct medial, lateral, or transverse incisions [7]. In infected rTKA, the skin incision should be safe and anatomically considered. After limb draping, previous scars should be well visible [5, 16]. Firstly, the debridement starts with the excision of the previous cicatrix, and in the case of a fistula, it should also be included in the excised tissue. If any sinus tracts are present, they should be excised to the joint capsule together with radical synovectomy [17].

14.3 Medial Parapatellar Approach

The MPA has been accepted as the workhorse of rTKA [18]. In 1878, von Langenbeck [19] described this approach for the first time. In over 90% of revision knee arthroplasties, a medial parapatellar arthrotomy has been preferred [4]. Della Valle et al. [16] present that in 92% of patients, the MPA in rTKA gave an adequate view of the knee joint.

Usually, a longitudinal midline skin incision is used. The incision of the parapatellar retinaculum is extended proximally, just lateral to the medial border of the quadriceps tendon, with a 3- to 4-mm intact part of the tendon on the vastus medialis for better closure of the approach; distally, the incision is extended along the medial border of the patella and the patellar tendon, leaving enough soft tissues on the patella for later closure [7] (Fig. 14.1). As an alternative, the so-called *wandering resident's approach* could be used. In this technique, detachment of the distal part of the quadriceps tendon in an oblique proximal-lateral direction from its insertion to the patella could be performed [3] (Fig. 14.2). A medial parapatellar arthrotomy ensures lateral eversion of the patella and maximally preserves the lymphatic and nerve branches [12].

The advantages of MPA include an excellent view and an easy and safe performance. The reported disadvantages are disruption of the quadriceps mechanism and destabilized patella [19], injury of the superior lateral genicular artery, and injury of the infrapatellar branch of the saphenous nerve with a painful neuroma [20].

In rTKA, the MPA with an extensive synovectomy is usually performed. Excision of all adhesions from the suprapatellar pouch and the deep surface of the quadriceps tendon ensures better visualization of the joint; thereafter, with the knee extended, the medial and lateral gutters should be released. Elevation of a subperiosteal flap of the medial retinaculum and the deep medial collateral ligament down to the semimembranosus insertion allows for external rotation of the tibia and thus facilitates access to the knee [5]. The extensor apparatus should be mobilized

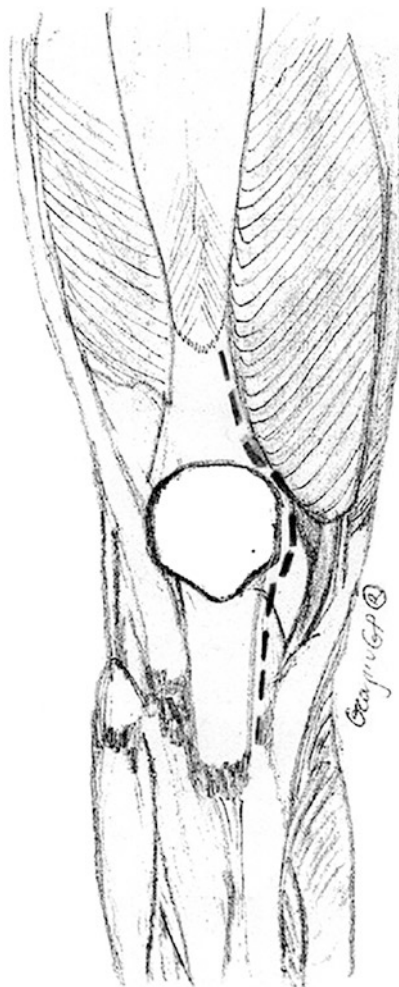


Fig. 14.1 Medial parapatellar approach

safely after releasing and excising the scar tissue between the patellar tendon and the anterolateral tibia, as well as adhesions on the lateral side of the patella to allow for patellar subluxation. This reduces the risk of patellar tendon injury [5]. Eversion of the patella is ensured by external rotation of the tibia together with knee flexion until anterior subluxation of the tibia occurs [13]. With severe adhesions and limited visualization, a lateral release should be performed for patellar mobilization. In cases of risk for avulsion of the tendon, a pin on the tendon insertion should be used [13]. Then, removal of the polyethylene inlay could be done [4]. Removing the polyethylene liner allows for better visualization and is

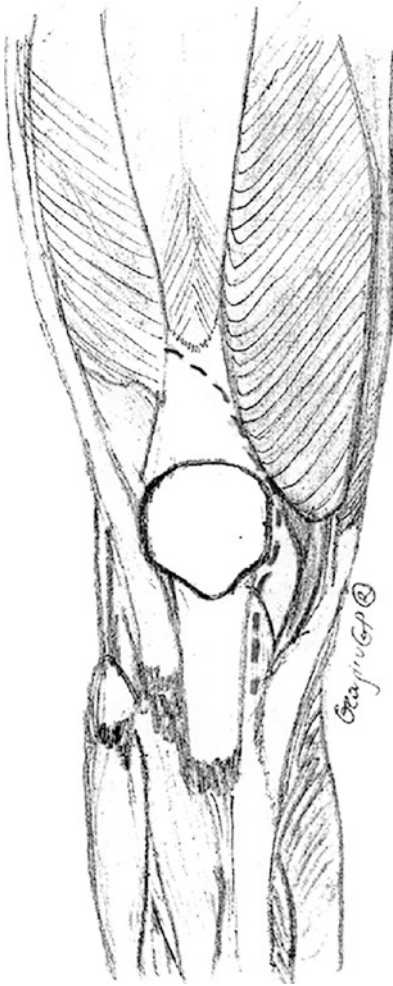


Fig. 14.2 The wandering resident's approach

followed by the removal of prosthesis components. After components removal, posterior synovectomy and posterior release could be made. Precise posterior release might prevent the elevation of the joint line [5].

14.4 Insall's Modification to Medial Parapatellar Approach

Due to the disruption of the extensor mechanism, instability, and damage to the patellar articular surface, a modification of MPA was proposed by Insall [21]. This technique ensures a more lateral

parapatellar arthrotomy and, in that way, allows for easier lateral subluxation or eversion of the patella [22].

In this approach, the division of the quadriceps tendon is 8–10 cm above the patella; the incision is prolonged over the medial one-third of the patella, thus detaching the medial patellofemoral ligament; the quadriceps expansion is sharply detached from the medial third of the bone till the medial part of the patella is clearly revealed; the incision is prolonged around the patella and over the medial one-third of the patellar tendon down to the tibial tuberosity [7, 22]. During wound closure, the medial retinaculum is sutured to the lateral two-thirds of the patella. Vaishya et al. [7], during suture of the extensor apparatus, applied three stitches between the medial retinaculum and the patella in the 90° flexed knee.

The disadvantages of the approach included injury to the infrapatellar branch of the saphenous nerve, patellar dislocation, subluxation, stress fractures, and fragmentation of the patella secondary to avascular necrosis [23, 24].

14.5 Lateral Approach

The lateral approach was published for the first time by Cameron and Fedorkow in 1882 [25]. Later, in 1991, Keblish [26] developed it for use in TKA in valgus knees and considered it as technically demanding. In cases of revision surgery, especially in infection, if a lateral arthrotomy has been previously performed, it should also be used in the subsequent approach; medial arthrotomy could provoke avascular osteonecrosis of the patella [13].

Anterior midline skin incision, a curvilinear midline skin incision, or a laterally placed anterior skin incision could be used [13, 26]. Usually, the skin incision starts around 5 cm proximally to the base of the patella and reaches the tibial tubercle. The incision is deepened through the subcutaneous tissue and the prepatellar bursa, and after reaching the lateral part of the patella, a parapatellar arthrotomy is started from the lateral side of the quadriceps tendon, passing over the

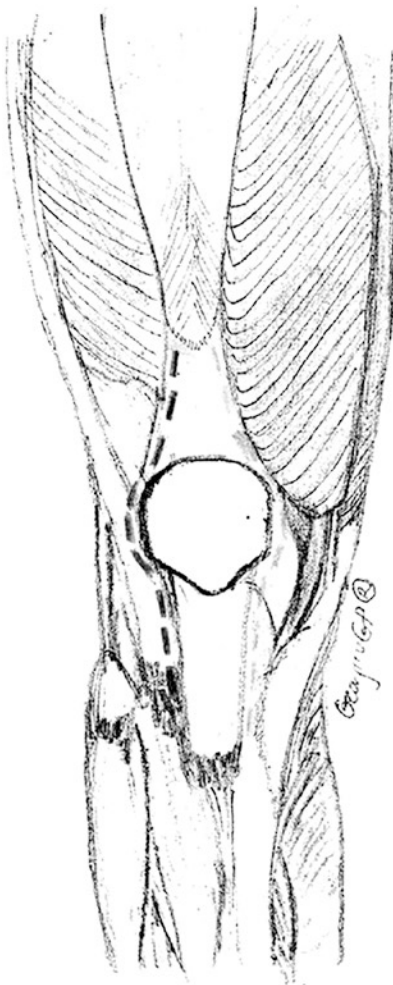


Fig. 14.3 Lateral parapatellar approach

lateral side of the patella to the anterior compartment fascia to the tibial tuberosity (Fig. 14.3). Then, medial eversion of the patella in extended knee is done; thereafter, knee flexion ensures exposure of the joint [7, 13]. Preserving a medial soft-tissue cuff ensures easier closure of the incision of the lateral retinaculum later [26].

14.6 Techniques for Exposure of Difficult TKA

rTKA is commonly performed using MPA. However, revision surgery is not an easy task and sometimes poses real challenges for sur-

geons. Scars from the previous operations make exposure to the joint difficult. Particular attention should be directed to the insertion site of the patellar tendon during eversion of the patella. In some difficult cases, the need for a more extensive exposure of the joint should be accepted [3–5, 7, 13, 27]. Different options for better exposure of the joint and easier removal of prosthetic components are clearly summarized and explained below. The author hopes this to be of use to revision surgeons and help them in their practice and other surgeons who will perform this surgery in the future.

14.6.1 Rectus Snip

In cases of limited exposure after MPA, a “quadriceps snip” or “rectus snip” is an option. Indications for a rectus snip include failure of adequate exposure after medial gutters debridement and medial release from the tibia together with debridement of the lateral gutters [13].

In 1983, Insall et al. [28] described the rectus snip for quadriceps lengthening, thus releasing the proximal tension of the quadriceps. In this technique, the proximal part of the medial parapatellar arthrotomy is prolonged obliquely and laterally at a 45°-angle to divide the tendon of the rectus femoris muscle from a distal-medial to proximal-lateral direction (Fig. 14.4). The underlying tendinous parts of the vasti muscles should also be cut. It should be pointed out that no detachment from the vastus lateralis should be performed [3–5, 7, 13, 27]. For anatomical repair of the tendon, Abdel and Della Valle [5] recommended putting two nonabsorbable sutures at the corners of where the quadriceps snip is started [5].

The advantages of this technique are its simplicity and effectiveness, ease of performance, no extensor lag, no need for modification of rehabilitation after surgery, protection of the lateral superior genicular artery, and the tendon of vastus lateralis [27, 29, 30]. Barrack et al. [31] and Garvin et al. [29] established no weakness in the strength of quadriceps after this technique.

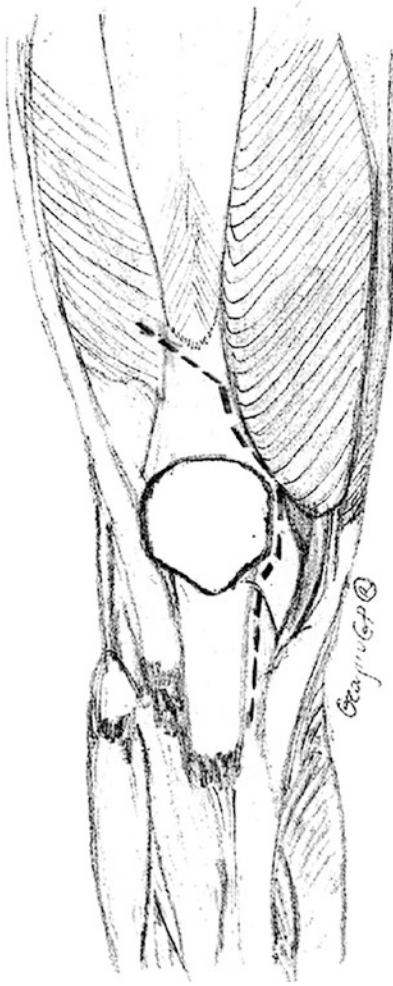


Fig. 14.4 The quadriceps snip

In summary, the rectus snip could be described simply as an oblique apical extension of the knee arthrotomy through the patellar tendon. This technique should not be performed through the muscle because it is difficult to repair, impeding the early postoperative rehabilitation [18].

14.6.2 V-Y Quadricepsplasty

A medial parapatellar retinacular incision can be extended into a V-Y quadricepsplasty procedure through the lateral patellar retinaculum if better exposure of the knee is needed in rTKA

[32]. Abdel and Della Valle [5] point out that a V-Y quadricepsplasty is rarely performed, usually in cases of shortening or contracture of the extensor apparatus when real lengthening is needed and also to facilitate the exposure. It could also be used in cases of degraded local skin conditions over the proximal tibia or with insufficient bone stock after TTO with poor capacity to heal. However, when the exposure is inadequate, and it seems to be due to contracture of the distal part of the extensor apparatus, a TTO is indicated [13].

Coonse and Adams [32] were the first to present a V-shaped turndown of the distal quadriceps for better visualization of the knee joint. Later, Insall used it during MPA; he extended the arthrotomy from the apex, in a distal and lateral direction at a 45° angle, through the vastus lateralis tendon down to the anterior fibers of the iliotibial band. In that way, the formed flap gives an easy approach to the knee [33] (Fig. 14.5). A V-shaped approach can be further changed into Y if the incision is extended proximally to the apex of the V [7].

In this technique, care should be taken to avoid injury of the superior lateral genicular artery. Moreover, excessive thinning of the peripatellar fat pad could lead to loss of blood supply to the patella. Cases of radiographic but asymptomatic patellar osteonecrosis after quadriceps turndown have been reported [34].

The closure of the V-Y quadricepsplasty should be performed at 30° of knee flexion [5]. Postoperatively, for 6 weeks brace protection, restriction of flexion, limited range of motion, and partial weight-bearing are mandatory [5, 35]. A V-Y quadricepsplasty could lead to an extensor lag after revision [5]. Scott and Siliski [36] presented their experience with this technique in 7 patients, and in 4 of them, transient extensor lag was established; in the other three, the lag was permanent. In contrast, Trousdale et al. [37] presented their experience with this technique in revision and primary TKA. They concluded that after V-Y quadricepsplasty, the patients had near to normal extension and moderate extensor weakness. These results were based on 9 revisions and 5 primary TKAs.

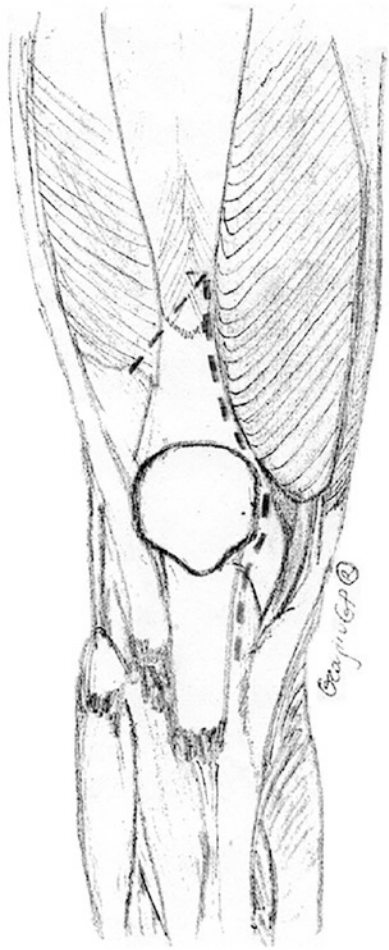


Fig. 14.5 V-Y quadricepsplasty

14.6.3 Tibial Tubercle Osteotomy

A TTO is performed in cases when other techniques cannot ensure adequate visibility of the knee. TTO allows for distal release of the extensor mechanism and is indicated in patients with stable tibial component or second re-implantation with excessive fibrotic tissue, in cases of extraction of a long-stemmed tibial component, in patients with excessive scarring at the anterolateral tibial area or when there is arthrofibrosis or patella baja [4, 5, 27]. A relative contraindication for TTO is the poor bone quality of the tibial tubercle, which impedes adequate fixation of the osteotomized fragment [38].

In 1983, Dolin reported a TTO for the first time [39]. Later, this technique was modified by Whiteside and Ohl [40]. The TTO is made in the coronal plane from the medial side of the tibia. The osteotomized fragment should be 7–10 cm long and should have a thickness of 1 cm proximally tapering to 5 mm distally [41] (Fig. 14.6). According to Tanzer and Burnett [27], the thickness of the osteotomized fragment should be 10–20 mm, because a smaller fragment could be fractured and would have a limited area for healing after fixation. Usually, the length of the fragment is 8–10 cm, but it depends on the needs of the surgical exposure. If the tibial tubercle will not be transferred, pre-drilling of the fragment before osteotomy is preferable. The TTO is made with the knee extended or slightly flexed [27]. After osteotomy, the fragment needs to be reflected laterally on an osteoperiosteal hinge. Preserving the attachments of the muscles of the anterior compartment is crucial for the fragment's vitality [5].

At the end of the procedure, the osteotomy needs to be reduced with the knee extended

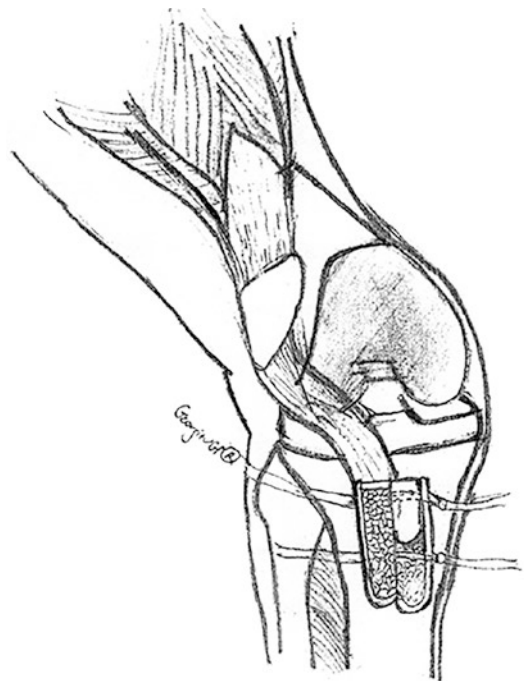


Fig. 14.6 Tibial tubercle osteotomy

and the fragment returned to its anatomical place. In cases of patella baja, the fragment could be transferred and reattached proximally [7]. In addition, autologous bone grafts could be placed around the fragment and in the surrounding free bony areas [27]. The fragment could be reattached by wire/cable fixation, screws, or a combination thereof. Usually, the wire fixation is performed by 2–4 stainless steel #16 g wires [27]. Della Valle et al. [5] prefer to fix the fragment with three 16-gauge wires [16].

According to Tanzer and Burnett [27], in larger fragments and good bone quality, fixation could be performed with two to four screws perpendicular to the cut surface, thus ensuring better compression. In stemmed tibial components, the screws need to be inserted obliquely to avoid the stem of the prosthesis. Usage of titanium screws is preferable, avoiding galvanic corrosion between stainless steel screws and titanium or cobalt-chrome stem.

After fixation of the fragment, the stability of fixation is tested by gentle flexion. This determines the postoperative active range of motion. Passive range of motion and high flexion attempts are not allowed in the first 12–16 weeks. A hinged brace locked in extension ensures weight-bearing [27]. According to Abdel and Della Valle [5], flexion is limited to $<90^\circ$ with a brace for 6 weeks. During mobilization, the brace is locked in extension. Active flexion is allowed, but an active extension and straight leg raises are avoided. Of course, the rehabilitation protocol with an increase of exercises depends on the stability of fixation. The brace could be removed at 12 weeks [27].

The advantages of TTO are excellent visibility of the knee, easy lateral eversion of the patella, sparing the attachment of the patellar tendon, easy and secure fixation of the fragment, and preservation of the blood supply to the extensor apparatus [27].

Different complications of TTO have been reported: non-union or delayed union, displacement of the fragment, iatrogenic fracture, hardware cutout, persistent anterior knee pain, infection, necrosis of the wound, excessive prom-

inence of the hardware, periprosthetic fracture, and restricted physiotherapy protocol [4, 13, 27, 31, 40, 41].

In summary, the TTO could be performed when other techniques have failed. Lower patient satisfaction after TTO was established [31].

14.6.4 Femoral Peel

As its name suggests, in FP, a full subperiosteal release of the distal femur is performed, the so-called femoral skeletonization. This technique violates the stability of the joint and needs to be used only in cases with restriction of flexion due to excessive scar tissue and failure of the other techniques. In rTKA with no excessive scar tissue, additional release of the surrounding knee structures will predispose to instability. In cases of excessive scar formation, this technique ensures stability despite the stripping of the ligaments and joint capsule [3, 13, 27].

In 1988, Windsor and Insall [42] used the FP in a severely ankylosed knee. During its performance, all soft tissues were subperiosteally released from the distal femur. In this technique, the medial collateral ligament and joint capsule are released, which causes knee instability but allows for excision of the fibrotic tissues that do not allow flexion. In case of inadequate release, the dissection is broadened with the detachment of the lateral collateral ligament and joint capsule, and thus real skeletonization of the femur is achieved; this could provoke devascularization of the distal part of the femur. All surrounding tissues should be detached with a scalpel or with electrocautery, as close to the bone as possible. The soft-tissue release should allow for full excision of the scar tissue from the posterior corner of the knee for better flexion. This technique could be extended with disinsertion of the origins of the gastrocnemius muscles. Finally, the surgeon does not reattach the ligaments and only closes the structures layer by layer in an extended knee [3, 13, 27].

With this technique, Lahav and Hofmann [43] had no cases of ruptures of the extensor apparatus or compromised extension of the knee.

The reported complications after femoral peel are iatrogenic vascular injury, tibiofemoral dislocations, infection, rupture of the patellar tendon, and periprosthetic fracture [44].

14.6.5 Medial Epicondylar Osteotomy

MEO is indicated in cases where flexion of the knee is blocked, but there is no excessive scarring of the surrounding structures. The choice between FP and MEO depends on the scarred tissue found during surgery [3, 13, 27].

In 1999, Engh [45] was the first to describe this technique for better visualization and correction of a varus deformity of the knee. The osteotomy is done with the knee flexed at 90°. With an osteotome, the bone cut is started laterally to the origin of the medial collateral ligament and finished above the insertion of the adductor magnus tendon, and thus the osteotomized fragment includes the medial epicondyle and the adductor tubercle (Fig. 14.7). A bone segment around 4 cm long and 1 cm wide is formed and hinged posteriorly, including the insertions of the medial collateral ligament and the tendons of the adductor magnus muscle. After flexion of the joint and eversion of the patella, the knee could be opened by external rotation and a valgus bend. At the end of the procedure, after implantation of the components, the bone segment is reattached by a minimum of three stitches with heavy nonabsorbable sutures, at 90° of knee flexion, or by a single screw. In some cases, this technique could be performed on the lateral epicondyle when lateral exposure is needed [3, 13, 27].

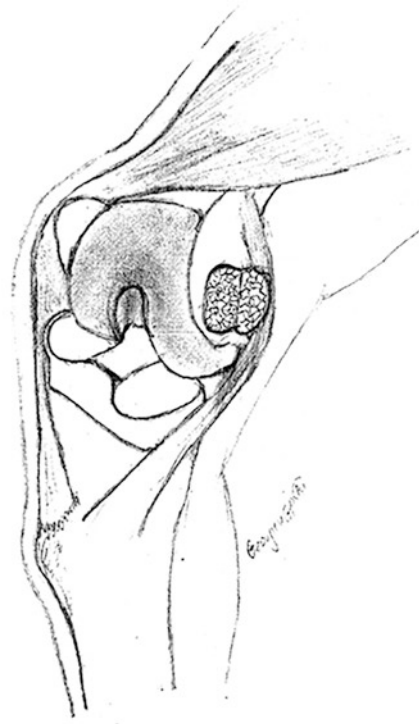


Fig. 14.7 Medial epicondylar osteotomy

options are essential. The aim of the surgical approach in rTKA should be to allow easy removal of components and implants and implantation of others without damaging the extensor apparatus or other surrounding structures. In cases where additional exposure is needed, a quadriceps snip could be performed. Rarely, in difficult revisions, TTO and V-Y quadricepsplasty are excellent options. Finally, the author would like to present the results and experience of Barrack et al. [31] after evaluation of 123 cases of rTKA from three centers. They concluded that the results after a standard MPA were the same with or without the quadriceps snip; patients with quadricepsplasty and a TTO had the same results, but worse than the standard approach. The results of patients with quadricepsplasty were significantly better in terms of a range of motion than the TTO group; patients after TTO had a lower degree of extension lag but a higher degree of difficulties with kneeling and stooping and a high rate of dissatisfaction with surgery.

14.7 Conclusion

Knowledge of the clinical anatomy of the knee is crucial for preparing and performing different options for better and atraumatic exposure of the knee. Apart from the gold standard, the MPA with synovectomy in rTKA, the knowledge of other approaches, and better visualization

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DAIR (Debridement, Antibiotics, and Implant Retention) for the Treatment of Periprosthetic Joint Infection of Knee

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15.1 Introduction

Prosthetic joint infection (PJI) is one of the most devastating complications following joint replacement. It occurs in approximately 1–2% of all joint replacement [1]. With increase in number of joint replacement procedures performed each year, it can be calculated that the number of PJI will also increase. Revisions for infected knee arthroplasties are complex, expensive, require more surgical time, longer hospitalization, and have higher risk of failure compared to aseptic revisions [1–3]. The primary goal of treatment is eradication of the infection. Maintenance

of a pain-free, functional joint is the secondary goal, which is also important [2, 4].

The surgical options include irrigation, debridement, antibiotics, and implant retention with or without polyethylene exchange (DAIR), one-stage or two-stage revision, resection arthroplasty, arthrodesis, and amputation [2, 5–8]. When patients are contraindicated to undergo DAIR treatment, either one stage or multiple stages revision surgery is the preferred option. Resection arthroplasty (without reimplantation), arthrodesis, and amputation remain valid options for difficult to treat and chronic PJI, and these treatment options very rarely have a role in acute PJI cases. Non-surgical medical treatment such as antibiotic suppression therapy should be reserved for patients who are unfit or contraindicated for surgery [1, 3].

DAIR (debridement, antibiotics, and implant retention) remains the treatment of choice for acute PJI. An irrigation and debridement procedure is not a new one and has been performed ever since infected knee arthroplasty cases emerged [9, 10]. The abbreviated term of “DAIR” itself has been first used in a publication by Byren et al. in 2009 [11]. Since then the procedure has been more popular and there have been increasing number of research and reports on the role of DAIR in the acute setting, with special attention given to the topic lately.

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15.2 Preoperative Considerations

15.2.1 Definition of PJI and Classification

During the past period, various definition criteria for PJI have been described by several organizations and societies. There is no uniform definition for periprosthetic joint infection (PJI). New diagnostic criteria are formulated and updated constantly [12]. The latest diagnostic criteria include the 2018 definition of periprosthetic hip and knee infection, the European Bone and Joint Infection Society (EBJIS) 2018, and the International Consensus Meeting (ICM) 2018 [13–15]. The 2018 definition of periprosthetic hip and knee infection is a scoring system which involves newer laboratory marker including *D-dimer*, *synovial alpha defensin*, *synovial CRP*, and *synovial leukocyte esterase (LE)* [13]. Those markers were not included in the previous diagnostic cri-

teria. These criteria have been proved to increase the diagnostic efficiency of PJI [16]. The scoring system is summarized in Fig. 15.1.

Understanding the classification of PJI is important as one of the factors to determine the appropriate treatment and ensure the best results. There are several classifications that have been proposed to define the onset of PJI [1]. Current guidelines from the ICM on PJI and the pro-implant foundation make a clear distinction between acute and chronic PJI [17, 18]:

1. Acute postoperative infection is considered to occur <4 weeks of the procedure,
2. Acute hematogenous infection is considered as <3 weeks of the development of symptoms.
3. Any infection which develops ≥ 4 weeks after the index surgery or symptoms duration of acute hematogenous ≥ 3 weeks considered as chronic PJI.

| Major criteria (at least one of the following) | | Decision |
|--|--|----------|
| Two positive cultures of the same organism | | Infected |
| Sinus tract with evidence of communication to the joint or visualization of the prosthesis | | |

| Preoperative Diagnosis | Minor Criteria | | Score | Decision | |
|------------------------|----------------|--|-------|----------|---|
| | Serum | Elevated CRP <i>or</i> D-Dimer | 2 | | ≥ 6 Infected 2-5 Possibly Infected ^a 0-1 Not Infected |
| | | Elevated ESR | 1 | | |
| | Synovial | Elevated synovial WBC count <i>or</i> LE | 3 | | |
| | | Positive alpha-defensin | 3 | | |
| | | Elevated synovial PMN (%) | 2 | | |
| | | Elevated synovial CRP | 1 | | |

| Intraoperative Diagnosis | Inconclusive pre-op score <i>or</i> dry tap ^a | | Score | Decision | |
|--------------------------|--|--|-------|----------|---|
| | Preoperative score | | - | | ≥ 6 Infected 4-5 Inconclusive ^b ≤ 3 Not Infected |
| | Positive histology | | 3 | | |
| | Positive purulence | | 3 | | |
| | Single positive culture | | 2 | | |

Fig. 15.1 The 2018 definition for periprosthetic joint infection (PJI). (From Parvizi et al. [13]. With kind permission from Elsevier)

15.2.2 Treatment Algorithm

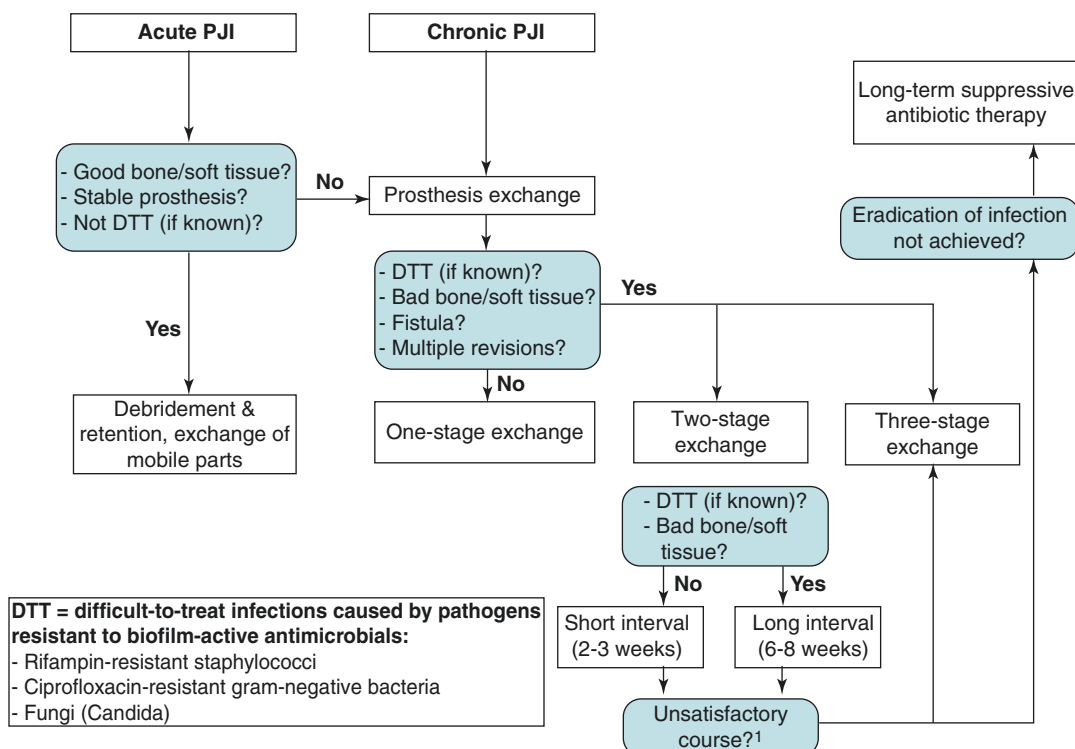
An early infection may be treated with aggressive debridement, antibiotics, exchange of modular parts, and retention of the fixed components, while late infection requires the removal of the components either in one- or two-stage fashion [7]. The fundamental aspects for a successful DAIR are related to tissue, stability of the prosthesis, and susceptibility of the organism. The management strategies of PJI are summarized in Fig. 15.2 [17].

15.2.3 Patients Selection for DAIR

Implant retention without infection is the ideal goal of treatment for an infected knee arthroplasty. If the conditions and criteria are met, DAIR treatment is preferable because it is less invasive, less technical demanding, has lower

morbidity, shorter hospitalization, better bone stock preservation, and lower economic burden [7, 19, 20]. However according to Koyonos et al., DAIR treatment is still a source of controversy among orthopedic surgeons because this procedure continues to be performed at relatively high rates despite an inability to consistently control infection, with rates of infection control ranging from 12% to 80% [19].

Microorganisms causing PJI are mainly *S. aureus* and coagulase-negative *Staphylococcus*, which account for up to more than half of the infections. Other microorganisms responsible include *Streptococcus* species, *Enterococcus* species, and Gram-negative bacteria [3, 4, 7, 21, 22]. Acute PJI is more often caused by *S. aureus* and *Streptococcus* species [1, 3]. *Coagulase-negative Staphylococci* are often associated with late chronic or clinically unapparent infection due to high biofilm production [3, 22]. Prolonged infection is associated with increased biofilm forma-



¹ Clinical signs of infection, elevated CRP, intra-operative pus, compromised tissue

Fig. 15.2 Treatment algorithm for PJI. (From Trampuz et al. [17]. With kind permission from PRO-IMPLANT Foundation)

tion and potential deep osteomyelitis [23]. The distinction between early and late PJI is based on the hypothesis that a biofilm was formed within 3 weeks on the surface of the components, thus necessitating their removal [6]. It is crucial to eradicate biofilm within a short time frame before it attaches to the implant [23]. Therefore, DAIR treatment shows better outcome in acute PJI compared to chronic infections [22, 23].

While tempting to perform DAIR on all PJI cases, the procedure is suitable only in selective cases and in recent years there is an emphasis on optimal patient selection. The wide range of success/failure between 10 and 90% emphasized this need. The decision to retain implants should be based on factors related to the host (comorbidities), the implant (stability), and infecting organism (virulence and ability to produce biofilm) [3, 6, 7, 24, 25]. Correct indications lead to higher rates of successful outcomes. Based on the 2018 ICM, the KLIC score is a recommended tool to predict treatment failure after DAIR procedure for early acute PJI (Table 15.1) [18, 26]. Patients with a score ≥ 7 are 2 times more likely to fail compared to those with a score < 7 [27]. Interestingly, patients with late acute infections show different characteristics and risk factors for failure. Recently the CRIME80 score was suggested as useful tool for patients stratification in cases of late acute infections (Table 15.2). The CRIME80 score ≥ 3 showed as independent predictor of failure for DAIR in late acute PJI [28].

The DAIR procedure is best performed if these criteria are met [1–4, 6, 8, 18, 22, 24]:

Table 15.1 KLIC score (from Tornero et al. [26]. With kind permission from Elsevier)

| Variable | Description | Score |
|----------|--|-------|
| K | Chronic renal failure (kidney) | 2 |
| L | Liver failure | 1.5 |
| I | Index surgery: | 1.5 |
| | Revision surgery Or prosthesis to treat femoral neck fracture | |
| C | Cemented prosthesis | 2 |
| C | C-reactive protein (CRP > 115 mg/L) | 2.5 |
| | Total | 9.5 |

Table 15.2 CRIME80-score, preoperative risk score for predicting DAIR failure in late acute periprosthetic joint infections. (From Wouthuyzen-Bakker M et al. [28]. With kind permission from Elsevier)

| Variable | Description | Score |
|----------|---|-------|
| C | COPD | 2 |
| | CRP > 150 mg/dL | 1 |
| R | Rheumatoid arthritis | 3 |
| I | Index surgery (prosthesis indicated for fracture) | 3 |
| M | Male gender | 1 |
| E | Exchange of mobile components | -1 |
| 80 | Age > 80 years | 2 |

1. Patients with an early acute PJI or acute haematogenous infection < 3 weeks.
2. Adequate skin coverage conditions.
3. Stable implant.
4. Identified definitive microorganisms, especially Gram-positive infection.
5. Availability of effective antimicrobial agent.
6. Patient with high risk of complication in more aggressive surgery.

The DAIR procedure has a higher rate of failure in below conditions; therefore, it should be contraindicated in [1, 3, 6, 8]:

1. Patients with risk factors for persistent or recurrent infection.
2. Poor local soft tissues conditions, especially in the presence of sinus.
3. Immunocompromised patients.
4. Resistant or unknown pathogens found in microbiology tests.
5. Polymicrobial infection.
6. Sepsis.
7. Prior failed procedures or debridements.
8. Late chronic infections.
9. Loose prosthesis.

Taking into account the complex interplay of factors associated with DAIR failure, the use of machine learning has been recently proposed for patient selection due to its ability to learn from continuous data input. By this means, machine learning models are able to process more complex data and make patient specific predictions. Recently, an algorithm based on

such technique was created and validate with promising results. Although such models still needs to be validated in external cohorts they have great potential to be used in daily practice by easily entering patient data in a computer-based software or phone application and may aid in clinical decision making and patient counseling [29].

15.3 Intraoperative Considerations

15.3.1 Surgical Technique

The components of DAIR include arthrotomy, extensive debridement and synovectomy, irrigation, retention of well-fixed implant, and exchange of modular components. The aggressive debridement of the periarticular tissues and the components aims to reduce the bioburden of the pathogens and to improve the efficiency of the patient's immune system and antibiotics against the surviving pathogens [6]. Debridement must be thorough and meticulous and all devitalized tissues must be excised. Various additional treatments have been used to improve local infection control and reduce biofilm, including the use of local antibiotics (e.g., antibiotic beads, sponges, and powder), chemical debridement and irrigation with various antiseptic agents (e.g., povidone iodine, chlorhexidine, peroxide, etc.), and physical treatment to the implants [18, 28, 30, 31].

Based on The International Consensus Group, we suggest the following protocol on how the DAIR procedure should be performed:

1. Debridement is not an emergency procedure in patients without sepsis. General conditions should be optimized prior to surgery [3, 18].
2. Acquire multiple tissue samples to identify the etiology of infection prior to surgery. Antibiotics should be withheld until representative samples are identified [3].
3. Adequate surgical exposure (preferred from the previous incision) to the infected area is

mandatory. Including excision of the skin fistula if it is present.

4. Obtain multiple intraoperative tissue culture for further isolation of causal microorganism. No less than 5 cultures should be obtained intraoperatively [6].
5. All non-bleeding soft or osseous tissues should be removed, including excision of sinuses and synovectomy [1, 3, 6].
6. The mobile component (PE liner/insert) must be removed to access all part of the joint, and exchange of the modular component is strongly recommended [1–3, 6, 18, 32].
7. All the components should be inspected for loosening and the interfaces of the components should be exposed [6].
8. Irrigation of the joint with copious amounts of irrigation solution (see irrigation protocol).
9. Switching to a clean patient setup prior to wound closure is also recommended to facilitate more sterile and uncontaminated wound [33].
10. A suction drain should be left in situ until there is minimal output. If drainage persists or if the infection fails to settle, then consideration has to be given to a further debridement procedure. Continuous closed irrigation has not shown any benefit compared to standard procedure with primary closure and in situ drain [3].

15.3.2 Irrigation Protocol

Several protocols for the irrigation have been proposed. The International Consensus Meeting 2018 strongly recommend 6–9 L of irrigation solution as minimum necessary volume [18]. The most commonly used to irrigate the joint is normal saline. Several authors believed that adding some chemical agent to the irrigation solutions could help in reducing pathogen load. Detergents, antiseptics, or even antibiotics have been proposed as adjuvant agent. Adding antibiotics including bacitracin, neomycin, polymyxin/neomycin, and gentamycin to the irrigation solution

has been shown to be no beneficial effect compared to saline alone [34, 35]. Recently, there has been increased interest in using chlorhexidine gluconate 0.05%. Studies by Smith et al. and Schwechter et al. showed chlorhexidine gluconate to be the most effective option at decreasing bacterial colony counts when compared to normal saline, povidone iodine, or castile soap [31, 36].

There is little consensus regarding use of low-pressure (<15 pounds per square inch) or high-pressure (>45 pounds per square inch) lavage. High-pressure lavage provides rapid and effective removal of necrotic tissues, but may cause tissue damage or penetration of bacteria into deeper soft-tissue layers [6, 37]. However, both low-pressure or high-pressure lavage can be used and no significant difference has been shown to exist in clinical practice [38].

15.3.3 Modular Component Exchange

Removing the modular components during DAIR (i.e., polyethylene liner/insert, femoral head) provides better access to the joint capsule for extensive debridement and synovectomy. Recent study by Hirsiger et al. showed that mobile component exchange doubled the probability for long-term remission [39]. They found the exchange of mobile parts was protective with hazard ratio 1.9 (95% confidence interval 1.2–2.9) in multivariate Cox regression analysis. These findings were supported by several previous studies. Lora-tamayo et al. showed that the exchange of removable components during debridement stands as an independent predictor of a favorable outcome with hazard ratio 0.6 [32]. In addition, Choi et al. reported that regardless of the causative organisms, lack of mobile component exchange resulted in poor outcome after DAIR treatment for PJI of the knee [9]. Another publications by Tsang et al. reviewed cohort studies published during 38 years period (1977–2015) on the results of DAIR for periprosthetic joint infection (PJI) of the hip. The success rate of DAIR with modular

component exchange was 73.9% (471/637 patients) compared with 60.7% (245/404 patients) in the non-modular component exchange group ($P < 0.0001$) [40]. The modular component exchange also became independent predictors of treatment success in a multicenter cohort study evaluating the outcome of DAIR in hip and knee PJIs caused by methicillin-resistant and methicillin-susceptible *S. aureus* [32]. Further, Grammatopoulos et al. reported a success rate of 93.3% when modular components were exchanged compared to 75.7% when modular components were retained in a series of acute PJI of the hips [41]. The rationale behind these findings might be associated with the evidence that bacterial load detected on polyethylene component is higher compared to the metal components of prostheses [42]. Thus, removing the polyethylene modular components will reduce the amount of bacterial load and biofilm in the infected joints.

15.3.4 Local Antibiotics Administration

Carriers for local antibiotic release include antibiotic loaded bone cement (polymethylmethacrylate, PMMA), beads, and dissolvable sponges. The rationale for using local antibiotic treatment is to achieve a high local concentration of antibiotic agents, thereby killing the causative microorganism, without the side-effects of high systemic concentrations [1]. Beads are usually loaded with gentamicin, but vancomycin and tobramycin are also used [1, 4]. Their use in DAIR treatment has been reported in a few studies, with relatively high success rates (75–83%) [43, 44].

Calcium sulfate beads have recently been reported as an alternative to PMMA beads. The advantages of calcium sulfate beads include a bioabsorbable material which excludes the need for additional surgery. It is also believed to have higher sustained concentration of local antibiotics and higher resistance to biofilm formation compared to PMMA beads. In contrast, the disadvantage of using antibiotic beads

includes decrease in local antibiotic concentration that occurs as soon as 24 h after implantation, possible colonization by bacteria and their capability of forming a foreign body on which a biofilm can develop after the antibiotic release (10–14 days) and potential for hypercalcemia [1, 22, 45, 46]. Unfortunately, its beneficial effect was still unpromising. Flierl et al. reported only 52% success rate with antibiotic-impregnated calcium sulfate beads in their retrospective report of 32 patients [45]. Calanna et al. modified surgical technique developed to enhance the classical irrigation and debridement procedure to improve the possibilities of retaining an infected total knee arthroplasty with the use of calcium sulfate beads [47]. This technique, debridement antibiotic pearls and retention of the implant (DAPRI), aims to remove the intra-articular biofilm allowing a higher and prolonged local antibiotic concentration using calcium sulfate beads. The combination of three different surgical techniques (methylene blue staining, argon beam electrical stimulation, and chlorhexidine gluconate brushing) might enhance the identification, disruption, and finally removal of the bacterial biofilm, which is mainly responsible for antibiotics and antibodies resistance. The DAPRI technique might represent a safe and more conservative treatment for acute and early hematogenous periprosthetic joint infection. They reported the success rate of infection eradication as high as 80% [47]. Recently, Gramlich et al. showed that the use of calcium based-antibiotic beads in combination DAIR has improved the 3-year infection-free survival compared to DAIR only which has re-infection rate until 81.8% in salvage procedure for chronic PJI of the knee [48].

Resorbable gentamicin-loaded sponges also have been used as local antibiotic in infection case after total hip arthroplasty, with success rates of 70% in a study reported by Kuiper et al. [49]. Another alternative for local antibiotic treatment is vancomycin powder. Riesgo et al. found that the combination of vancomycin powder and a dilute povidone iodine lavage to DAIR increases the success rates up to 83% [50].

15.4 Postoperative Consideration

15.4.1 Postoperative Antibiotics Regimen

The antimicrobial agent should have bactericidal action, even against slow-growth organisms or biofilm producers. Before starting any treatment, the susceptibility of the organism should be tested and alternative regimens should be discussed, given the growing levels of resistance [4]. A combination of rifampicin with quinolones has been used most often, with good results in vitro, in vivo, and in clinical trials [4, 11, 51]. Rifampin is thought to penetrate the biofilm and is recommended in all cases of Staphylococcal PJI treated with DAIR [1]. Higher success rates were found when rifampin was added with another antibiotic regimen [32, 51]. Options such as linezolid, sulfamethoxazole-trimethoprim, and minocycline are possible, although so far no clinical studies for validating their use have been published [5]. The best option is to discuss the best antimicrobial therapy for each case with the hospital infection control committee [3, 4].

Postoperative long-term combined intravenous (IV) antibiotic treatment of between 4 and 6 weeks followed by oral rifampin for 6 months is recommended [3, 6]. Byren et al. demonstrated that the infection-free survival rate after DAIR treatment was 82%, with a follow-up of 2.3 years. However, there is risk of recurrence following discontinuation of the antibiotics [11]. Some authors have suggested a combined protocol consisting of debridement, antibiotic treatment for >1 year, and implant retention (DAIR) for the treatment of PJI [6]. The risks of this happening are increased fourfold according to Byren et al. suggesting that this form of treatment did not eradicate the pathogen but postpones its reactivation [11]. However, there are also some studies that show short duration antibiotics could be used as effective as the long course administration. A study by Chaussade et al. showed no difference between 6 and 12 weeks of intravenous antibiotics with overall success rates of 69% [52]. A multicenter randomized clinical trial by Lora-Tamayo et al. suggests that 8 weeks of levofloxacin +

rifampicin therapy had similar outcomes to longer standard treatment (3–6 months) for acute PJI managed with DAIR [51]. A cohort studies in Leiden University by Scheper et al. also showed the outcome of acute PJI treated with DAIR and 5 days rifampicin was comparable to the outcomes of 3 months rifampicin combination therapy [53]. Recently, the international consensus meeting 2018 stated that a minimum of 6 weeks of antibiotic therapy seems to be sufficient in most cases of PJIs managed by DAIR-provided surgical treatment [18].

15.4.2 Factors Associated with Outcomes of Dair

The outcome of DAIR procedure varies between studies. The results between studies are highly variable due to many confounding variables such as: host condition, characteristics of the microorganism, implant state, operation history, type of surgery/procedure, surgeon's ability in the various series, lack of consistency in definition of acute infection, different failure criteria among studies, and the absence of randomized, controlled, prospective comparison studies [1, 3, 7, 20, 24]. Recent systematic review and meta-analysis by Kunutsor et al. showed that DAIR resulted in quite a wide range of infection control rate by 11.1–100% [54]. This was no better when compared to two-stage revision procedures which has success rate of 85–100% [7].

Outcome may be adversely affected by the time interval between the initial operation and the development of infection [43, 55]. The success rate of DAIR dropped to 40% when the infection started >6 weeks after the TKA [6]. Löwik et al. reported among 769 patients with acute PJI the treatment failure occurred in 38% (294/769) of the patients after DAIR. The treatment failure rate was almost similar between time intervals from index arthroplasty to DAIR: week 1–2 was 42% (95/226) failure rate, week 3–4 was 38% (143/378) failure rate, week 5–6 was 29% (29/100) failure rate, and week 7–12 was 42% (27/65) failure rate. They reported that DAIR could be viable option for acute PJI which pres-

ents more than 4 weeks after the index surgery as far as performed at least 1 weeks after the symptoms and modular component is exchanged [56]. Trebse et al. applied a DAIR protocol to a series of 24 patients with an 86% success rate over 3 years and defined that the factors for a good prognosis were the presence of a stable implant, absence of fistulas contiguous with the prosthetic component, and duration of symptoms less than 3 weeks [57]. Koyonos et al. performed DAIR in 136 patients and reported higher success rate in acute postoperative (31%) and acute hematogenous infections (44%) compared to chronic late infections (28%) [19]. Tsukayama et al., Segawa et al., Mont et al., Cobo et al., also reported a success rate of 57.3–80% when DAIR performed in early PJI (<4 weeks) [43, 55, 58, 59].

Recently some new evidences support the expansion of DAIR indication for a PJI which has symptoms >3–4 weeks. Zhang et al. in a small series of 24 patients with acute PJI, reported that 5 patients who had symptoms between 4 and 8 weeks have 100% success rate after DAIR treatment [60]. Lesens et al. also reported that failure after DAIR was not associated with time from index arthroplasty to debridement, nor with duration of symptoms (>3 weeks) in a retrospective series of 137 patients with early PJI [61]. Although further study is needed to prove these findings, as another recent evidence showed that late acute PJI still has lower success rate compared to early acute PJI especially when the causative agent is *Staphylococcus* spp [28, 62, 63].

Multiple studies have shown *S. aureus* infection to be a contributing factor for failure to eradicate infection due to more virulent nature than other microorganisms (possibly due to their biofilm production) [3, 7, 19, 20, 64, 65]. MRSA, in particular, has shown high failure rates with DAIR [1, 3]. Several studies reported low success rate of 0–45% when DAIR performed in MRSA infection [66–68]. Gram-negative organisms have shown a variable outcome in failure rates as compared to Gram-positive organisms [3, 65]. The DAIR procedure has also shown promising results in patients who are immunocompetent and with PJI caused by a low virulence organism, e.g., *Coagulase-negative staphylococci* [3, 19].

The international consensus meeting 2018 strongly agreed some factors which possibly associated with treatment success in acute PJIs treated with DAIR [18]:

- Exchanging the modular components during debridement.
- Performing a debridement within at least 7 days, but preferably as soon as possible, after the onset of symptoms.
- Adding rifampin to the antibiotic regimen, particularly when combined with a fluoroquinolone, in cases of susceptible *Staphylococci*.
- Treatment with fluoroquinolones in cases of susceptible Gram-negative bacilli.

The following factors also have been shown to be associated with treatment failure after DAIR in acute PJI [18]:

- Host-related factors: rheumatoid arthritis, old age, male sex, chronic renal failure, liver cirrhosis, and chronic obstructive pulmonary disease.
- Prosthesis indication: fracture as indication for the prosthesis, cemented prostheses, and revised prostheses.
- Clinical presentation representing the severity of the infection: a high C-reactive protein, a high bacterial inoculum, and the presence of bacteremia.
- Causative microorganisms: *S. aureus* and *Enterococci*.

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One-Stage Exchange Arthroplasty of the Infected Knee

16

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16.1 Introduction

Chronic periprosthetic joint infection (PJI) of the knee joint requires exchange arthroplasty. Worldwide, two-stage exchange arthroplasty has become the “golden standard.” In contrast, the ENDO-Klinik follows a distinct one-stage exchange for PJI in over 85% of all our infected cases according to the first implementation of mixing antibiotics into bone cement introduced by Prof. Buchholz in the 1970s.

From a global perspective, the concept of one-stage exchange arthroplasty has become increasingly popular in several specialized centers due to the potential functional benefits for the patients and a decreased burden to their national health-care system [1].

The one-stage exchange offers certain advantages, as mainly based on need for only one operative procedure, displayed in the chapter

“clinical results.” In order to fulfill a one-stage approach with its potential success, there are obligatory pre-, peri-, and postoperative requirements, which need to be meticulously respected. The following book chapter provides an evidence-based overview in regard to the key points of one-stage exchange arthroplasty of the infected knee.

16.2 Indications for One-Stage Exchange Arthroplasty

The germ has to be known for one-stage exchange arthroplasty based on microbiological diagnostics as well as a distinct patient specific plan for the topic and systemic antibiotic treatment by a multidisciplinary team (Table 16.1). Table 16.1 reveals the indications along with contraindications for the one-stage procedure. In a recent study by Citak et al. [2], risk factors for failure after one-stage exchange TKA in the management of PJI have been identified. According to the study results, the isolation of enterococci and streptococci had significantly higher risk for failure. However, similar results have also been found for the two-stage procedure [3, 4]. Therefore, further comparative studies are required to determine both germs as either indications or contraindications for the one-stage procedure.

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Table 16.1 The indications and contraindications for the one-stage procedure

| Indications | Contraindications |
|--|---|
| <ul style="list-style-type: none"> • PJI after TKA in which infection is proven, based on the ICM criteria 2018 • Late or chronic infection more than 30 days postoperatively • Hematogenous infection more than 30 days after onset of the symptoms • Known germ with known susceptibility based on microbiological diagnostics • Possibility of primary wound closure | <ul style="list-style-type: none"> • Culture-negative PJI • Non-availability of the required antibiotic • Systemic sepsis of the patient • Failure of 2 or more previous 1-stage procedures • Severe soft tissue infection spreading to the nerve-vessel bundle • Extensive soft tissue involvement that would prevent closure of the wound |

16.3 Endo-Klinik Diagnostic Protocol

Despite the fact that there are no specific symptoms for periprosthetic joint infections, it is recommended that every patient with the following criteria should be further tested to validate or exclude a PJI:

- patient's medical history (e.g. prolonged wound secretion, fever, wound healing disorders),
- painful total joint arthroplasty,
- loosening of the total joint arthroplasty within the first year,
- unspecific symptoms such as night sweat, fatigue, unwanted loss of weight,
- elevated inflammatory markers (e.g. serum C-reactive Protein, erythrocyte sedimentation rate),
- prior elective revision arthroplasty.

The joint aspiration is the most needed and relevant preoperative diagnostic test in any case of a planned one-stage exchange. Hereby, it is highly recommended to respect a prolonged microbiologic culture time of at least 14 days [5]. Furthermore, the antibiotics should be withheld

for 14 days prior to the aspiration. The joint aspiration should be performed under operating room conditions with sterile washing and draping. In order to avoid false negative results, local anesthetics or saline rinsing should not be administered.

Synchronous PJI is a rare, but serious complication with an incidence rate of 4% [6]. Therefore, joint aspiration is obligatory for exclusion of the suspicion of PJI after joint arthroplasty.

Several tests for diagnosing PJI are performed in order to differentiate between septic and aseptic failure. At our institution, besides serum CRP, synovial fluid is tested in the following descending order:

1. culture and susceptibility,
2. quantitative alpha defensin test,
3. leukocyte esterase test,
4. cell count,
5. polymorphonuclear leukocytes (PMN%).

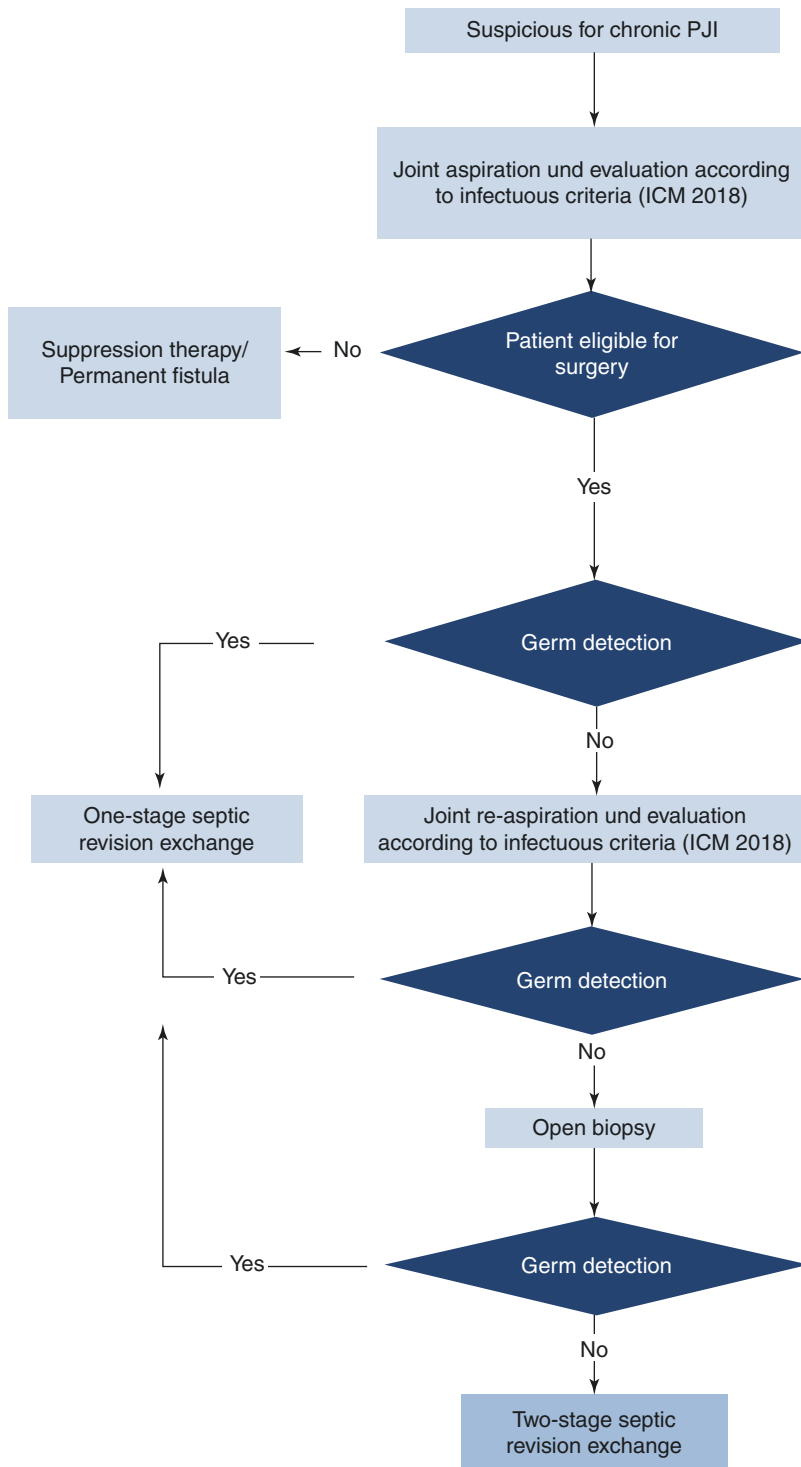
The verification of periprosthetic joint infection is made according to the ICM 2018 Criteria [7]. In case of a negative germ detection, a secondary control aspiration is recommended to eliminate suspicion for PJI. If the results of the second aspiration for germ identification remain negative, an open biopsy should be performed (Fig. 16.1).

16.4 Surgical Technique

16.4.1 Preoperative Planning and Surgical Approach

In every case, preoperative plain radiographs (anteroposterior and lateral views of the knee joint and tangential view of the patella) and anteroposterior standing long-leg radiographs are performed. The preoperative identification of the bacteria defines which antibiotic-loaded acrylic cement is required and is the main factor to perform the one-stage exchange arthroplasty.

Fig. 16.1 Flow chart of the Endo-Klinik Treatment Protocol of chronic PJI



16.4.2 Radical Debridement and Removal of All Hardware Materials

The patient is placed on the operating table in supine position. The skin is prepped four times with a propanol solution (Cutasept G, Bode Chemie, Hamburg, Germany) within at least 2 min of acting time. After disinfection, a standard knee draping with single use materials is performed.

First, the existing scar is excised (Fig. 16.2). If a fistula is present, a radical excision down to the joint capsule is necessary. Second, an extraarticular debridement of the joint capsule and the synovium is carried out. Third, the joint is opened and a radical debridement is performed, including a complete synovectomy (Figs. 16.3 and 16.4). Hereby, a radical excision needs to be done for all non-bleeding tissues and related bones. In addition, the radical soft tissue resection incorporates the debridement of collateral ligaments as well as the excision of each infected tissue around the patella region and in the patella surface. In order to assure an adequate debridement of the infected tissues/bone down to the viable tissue, the authors do not recommend utilizing the tourniquet during the debridement process.

After completing the radical debridement of the surrounding tissues, all hardware materials are removed. To increase the feasibility of the removal, the usage of adequate instruments is crucial. In this case, the implant–cement interface loosening of the femoral and tibial components is performed with either an oscillating saw (Fig. 16.5) or an osteotome (Fig. 16.6). Subsequently, a punch is utilized to remove the mobilized tibial and femoral components with direct blows. After hardware material removal, the cement has to be removed entirely. Afterwards, meticulous debridement of bone and soft-tissues is fundamental including all areas of the knee. All non-viable bone has to be removed (Fig. 16.7).

For the combined microbiological and histological evaluation, five samples of biopsy material are collected during the debridement from all relevant areas of the operation site and sent to the laboratory for further evaluation. After the last



Fig. 16.2 Intraoperative image displays the excision of the persisting scar

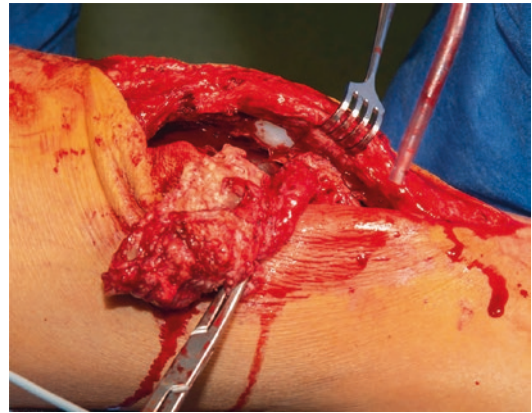


Fig. 16.3 Intraoperative situs showing the synovectomy



Fig. 16.4 The removal of the infected tissue



Fig. 16.5 The implant–cement interface loosening of the femoral component utilizing the oscillating saw



Fig. 16.6 Implant–cement interface loosening of the tibial component utilizing the osteotome

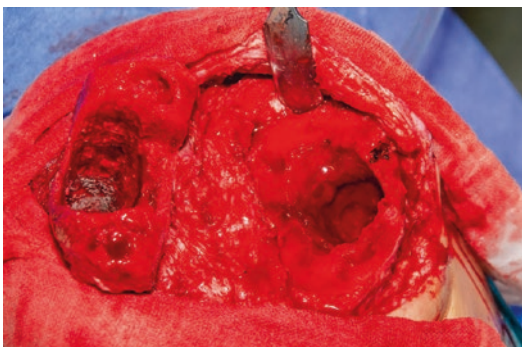


Fig. 16.7 The viable bone and soft tissue after radical debridement

biopsy that was taken and completed radical debridement, the irrigation is performed with pulsatile lavage with 0.02% polyhexanide solution (Lavasept, B. Braun, Melsungen, Germany) (Fig. 16.8). The polyhexanide-soaked swabs are placed over the wound area for at least 10 min



Fig. 16.8 The result of the radical debridement. Every infected tissue or bone and all foreign hardware materials have to be removed

before new operative setup is prepared. The new operative setup includes the re-draping of the surgical field, the change of light handles, suction tips, surgical gowns along with gloves.

16.4.3 Re-Implantation

After the last microbiological sample was taken, the systemic antibiotic therapy is started as recommended by the microbiologist. Then, the reconstruction of the joint is carried out with implantation of a cemented rotating hinge/full hinge knee implant (ENDO-Model, Waldemar Link, Hamburg, Germany) after preparing the tibia and femur with appropriate resection blocks. Antibiotic-loaded cement is utilized for both the fixation of the new implant and the reconstruction of bone defects. Instead of using allograft bone, we recommend to fill the defects either with polymethylmethacrylate (PMMA) bone cement (Copal, Heraeus Medical, Wehrheim, Germany) or trabecular metal cones (Fig. 16.9).

The preparation of antibiotic-loaded cement is followed after a strict protocol. In general, manufactured antibiotic bone cements are used, such as Copal G + C or Copal G + V (Heraeus Medical, Wehrheim, Germany). An admixture of antibiotics might be indicated, based on the preoperative microbiological findings. Finally, a primary soft tissue closure is accomplished, after the harden-

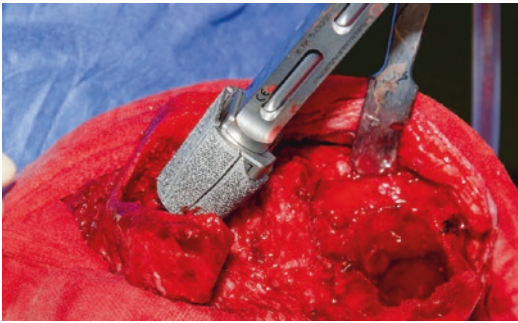


Fig. 16.9 Femoral bone defect filled with trabecular metal cone



Fig. 16.10 Implantation of the new prosthesis during the hardening process

ing process of the cement and irrigation (Fig. 16.10). Intraarticular suction drainage is strongly recommended.

16.4.4 Postoperative Course

Anteroposterior and lateral plain radiographs of the knee should be carried out instantly after surgery. Although the evidence about the optimal duration of intravenous antibiotic therapy remains scarce, we recommend to administer postoperative systemic antibiotic therapy for 14 days, based on the recommendations by the microbiologist [8]. On the day of surgery, the patient is mobilized with full weight bearing and support by crutches under intensive physiotherapy and sufficient analgesia. The wound drainage is taken off 48 h after surgery.

16.5 Clinical Results

There is a lack of studies on the outcomes after one-stage revision surgery [2, 9–20].

For instance, the reported eradication rates of one-stage revision technique vary between 73.1% and 100%, depending on the time of follow-up (Table 16.2). At our institution, the 10 year infection-free survival rate was 93% [20]. Other long-term studies with 10- or 12 years follow-up present success rates beyond 90% after one-stage revision arthroplasty [18, 21]. According to Citak et al. [2], the risk factors for failure after one-stage exchange TKA in the management of PJI have been identified. The top three causes for failure following one-stage knee exchange were with 51.6% recurrence of infection, followed by aseptic loosening with 40.7%, and finally patella complications with 3.3%. In this study, the mean time to failure after one-stage procedure was 25.2 months [2]. The top three risk factors for reinfection were the isolation of enterococcus, followed by failed one-stage exchange and persistent wound drainage. Isolation of streptococcus was also a significant risk factor for reinfection.

Table 16.2 Overview about the current literature of the one-stage septic knee exchange arthroplasty

| | N | Follow-up (in years) | Eradication rate (in %) |
|--------------------------|-----|----------------------|-------------------------|
| Freeman et al. [11] | 8 | 2.2 | 100 |
| von Foerster et al. [19] | 104 | Not reported | 73.1 |
| Goksan and Freeman [12] | 18 | 5 | 89 |
| Lu et al. [27] | 8 | 1.7 | 100 |
| Silva et al. [15] | 37 | 4 | 89.2 |
| Buechel et al. [10] | 21 | 10.2 | 90.9 |
| Sofer et al. [17] | 15 | 1.5 | 93.3 |
| Bauer et al. [9] | 30 | 4.3 | Not reported |
| Singer et al. [16] | 63 | 3 | 95 |
| Jenny et al. [14] | 47 | 2.75 | 87 |
| Haddad et al. [13] | 28 | 6.5 | 100 |
| Tibrewal et al. [18] | 50 | 10.5 | 98 |
| Zahar et al. [20] | 70 | 10 | 93 |

Streptococcus and enterococcus infections have also poor outcomes with the two-stage procedure [3, 4]. In a recently published review article, the one-stage exchange showed similar results in the eradication rate and functional outcomes, compared to the two-stage procedure, and offers the advantage of a unique surgical procedure [22]. Besides performing solely one operation, the one-stage exchange offers five main advantages:

1. higher quality of life [23],
2. lower morbidity/mortality rate [24],
3. lower in-hospital complications [25],
4. lower blood loss and allogeneic blood transfusion rates [26],
5. higher cost-effectiveness [23].

16.6 Conclusions

Based on the authors experience and the literature, the success of one-stage revision surgery is based on the following requirements: Well-defined intra-hospital infrastructure; meticulous preoperative aspiration regime; radical debridement including removal of all hardware materials; multidisciplinary approach (microbiologist, surgeon, etc.); adjusted local antibiotic-loaded bone cement; and postoperative adjusted systemic antibiotic therapy.

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Two-Stage Revision Arthroplasty for Periprosthetic Knee Infection

17

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17.1 Introduction

Periprosthetic joint infection (PJI) is one of the most catastrophic complications following joint arthroplasty. The complexity of treatment of PJI leads to dramatic physical, emotional, and financial costs. Despite advances in the prevention, diagnosis, and treatment of PJI, it remains the most commonly reported cause of early failure in total knee arthroplasty (TKA). The goal of PJI treatment is to eradicate the infection and restore a functional and stable joint [1]. Treatment of PJI includes surgical interventions such as irrigation and debridement, one-stage reimplantation, two-stage reimplantation, resection arthroplasty, or amputations. The choice of the treatment depends upon the type of infection and the type of the organism responsible, the general conditions of the patient and his life expectancy. Treatment of periprosthetic joint infections with a two-stage revision arthroplasty remains a widely used treatment strategy. The potential advantages of one-stage exchange arthroplasty are multiple, including a decrease in surgical morbidity and

mortality, earlier functional return, and lower costs. One-stage revision has a lower risk of mortality and morbidity compared to two-stage revision, which exposes the patient to the risks of an additional procedure. However, several authors demonstrated a reduced rate of recurrent infection after two-stage revision in comparison to one-stage revision [2–5]. The reinfection rate after two-stage revision is between 9 and 20% of cases [6]. Two-stage revision is the most used procedure for prosthetic joint infection treatment in North America [7].

17.2 History

Insall was the first to describe a two-stage reimplantation procedure for the management of infected total knee arthroplasties [8, 9]. Two-stage revision arthroplasty consists of removing all foreign materials from the joint, making an extensive debridement of periarticular tissues and inserting a static or articulating spacer.

The use of an impregnated antibiotic cement spacer block maintains the joint space, prevents retraction of the collateral ligaments, and provides a local antibiotic release. However, static spacers present several disadvantages, such as restriction of knee movement, tissue adherence formation, and quadriceps shortening. To overcome the problems of block spacers and to facilitate reimplantation surgery, articulating spacers

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were introduced. The implantation of the cement spacer is followed by systemic antibiotic therapy for an extended period. When the eradication of the infection is completed and the wound is healed, reimplantation could be considered. Patients report instability, pain, and limited function during this period, mainly if the non-articulating spacer is used. Reimplantation consists of removing the spacer, repeating debridement, and implanting revision arthroplasty components. There is a lack of high-quality evidence on the ideal type of spacer.

In our clinical practice, two-stage revision is the primary choice for the majority of PJI.

17.3 Timing for Reimplantation

There is not a consensus on the optimal timing for reimplantation [1]. The authors reported an interval between few weeks and several months [10–14]. The range of infection's eradication after two-stage revision is between 70 and 100%, without a clear correlation to the time of reimplantation [11–13, 15].

Several variables can be associated with unsuccessful eradication of infection following a two-stage revision knee procedure, including an increased duration between resection and reimplantation [16, 17]. On the other side, Babis et al. [18, 19] found a high rate of success with a mean 9-month interval in patients with multiresistant bacteria.

In our opinion, the timing for reimplantation must be decided based on clinical evaluations, wound healing, and serologic tests after a period of antibiotic therapy and subsequent antibiotic washout period. Our consideration finds its reason in the current literature. The most recent review about two-stage revision, realized by Tozun et al. in 2020, explains that a precise timing for the interval between the two stages still does not exist. In case of optimal local tissue conditions and quick time of recovery after the first stage, a short interval of 2–4 weeks should be considered. Conversely, when the culture in the first stage identifies a difficult-to-treat microorganisms, a longer interval of 4–6 weeks should

be preferred. Longer time intervals of over 8 weeks should be avoided as the antibiotic bone cement spacer misses its antibiotic concentration. The prolonged duration among resection and reimplantation seems to be associated with a greater risk of reinfection [20].

17.4 Non-Articulating Vs. Articulating Spacers

Periprosthetic joint infections could be managed with two-stage revision using non-articulating (Fig. 17.1) or articulating spacers (Fig. 17.2).

There is a lack of high-quality evidence on the ideal type of spacer [21–23]. Some authors reported the superiority of articulating spacers when compared to non-articulating spacers in terms of functional outcomes, time of hospital, and range of motion [24]. However, complications of spacers include fractures and dislocations. Surgeon-made articulating spacers are reported to have more risk of fracture when compared to preformed spacers.



Fig. 17.1 Non-articulating knee spacer



Fig. 17.2 Articulating knee spacer

Hofmann reported the use of the original, resterilized femoral component fixed with antibiotic-loaded cement and a new polyethylene insert cemented to the tibia for the knee with good outcomes (Fig. 17.3) [25].

Patients may be candidates for non-articulating spacers if they have major bone loss, ligamentous, or muscles injuries that cause a probability of dislocation or periprosthetic fractures or soft tissue defects. In these patients, a reduction in motion allows wound healing. On the other side, articulating spacers provide a better range of motion and less functional limitations but are indicated only in selected patients.

Emerson et al. [26] compared static block spacers with articulating spacers and reported an improvement in post-operative ROM with no significant difference in the reinfection rate.

However, antibiotic cement spacers should be used for a limited period: bacterial colonization of spacers can occur with increasing in situ time [27, 28].



Fig. 17.3 Hofmann reported the use of the original, resterilized femoral component fixed with antibiotic-loaded cement and a new polyethylene insert cemented to the tibia for the knee

17.5 Local Antibiotics

Local antibiotics added to cement have a higher concentration and duration in comparison to systemic antibiotic [29]. They should be tailored based on preoperative cultures and patients medical conditions, particularly renal function [27, 30]. If the infective organism is not isolated from preoperative cultures, a broad-spectrum empiric combination of antibiotics can be used [31, 32]. An ideal antibiotic should be safe, thermostable, hypoallergenic, water-soluble, with a high bacterial spectrum and available as sterile [33]. Antibiotics like gentamicin, vancomycin, ampicillin, clindamycin, and meropenem can be used as a combination based on organism susceptibility. Vancomycin is usually used for methicillin-resistant *Staphylococcus aureus* (MRSA) or methicillin-resistant *Staphylococcus epidermidis* (MRSE) [34], usually in combination with an aminoglycoside, such as gentamicin or tobramycin.

Third-generation cephalosporins, carbapenems, and monobactam antibiotics are used with success for susceptible gram-negative bacteria [35–38].

A rare complication of local antibiotic is the systemic toxicity as a result of elution of antibiotics from cement spacers. To prevent this complication renal clearance of the patients and viscosity of the cement must be checked. The optimal antibiotic dosage per 40-g bag of bone cement has not yet been determined. The reported doses range from 2 to 5 g for gentamicin, from 2.4 to 9.6 g for tobramycin, and from 3 to 9 g for vancomycin [39].

17.6 Systemic Antimicrobial Therapy

Systemic antimicrobial therapy should be tailored based on isolated bacteria and patients characteristics. Patients treated without bacteria isolation have 4.5 times increased risk of reinfection when compared to those patients where an organism was identified by culture [40, 41]. There is not a consensus on the optimal length of antibiotic treatment after resection arthroplasty. However, antibiotic therapy administrated for more than 6 weeks may increase the rate of antibiotic-related complications [42–44]. Excellent results are obtained with a combination of oral and intravenous antibiotic administration for 6 weeks or less [31, 45, 46]. Antimicrobial treatment is usually started with intravenous antibiotics to obtain the appropriate concentrations locally and after are switched to oral antibiotics.

17.7 Surgical Tips and Tricks

- During revision foreign materials, including cement, must be removed. These materials can act as a nidus for biofilm and persistence of infection [29, 47].
- Complete debridement of the joint and removal of all hardware is ideal during the surgical treatment.
- Is desirable to remove accessible heterotopic ossification if this procedure does not compromise future reconstruction.

- Allograft for management of bone defects during reimplantation seems not to increase the risk of reinfection [48, 49].
- The two-stage revision of unicompartmental knee arthroplasty requires the resection of all the compartments and of the fat pad.
- Soft tissue defects could be managed with a reconstructive flap at the time of explant or at the time of reimplantation. Medial gastrocnemius rotational flaps are usually used to manage soft tissue defects in knee arthroplasty revision. However, lateral gastrocnemius, latissimus dorsi, quadriceps, sartorius, and rectus abdominus could be alternatively used [50–52].
- During reimplantation cemented or cementless prosthesis could be alternatively used. No differences were demonstrated in terms of success rate of infection treatment. The choice between cemented or cementless components must be made based on classical factors such as bone quality or body mass. If cemented prostheses are used, consideration should be given to the addition of antibiotics active on the isolated bacteria [53].

17.8 Cement Spacer Exchange

Cement spacer exchange (Figs. 17.4 and 17.5) gives a new load of local antibiotics when the infection is not under control [54]. However, there is a lack of evidence on the benefit of this procedure. Indications for spacer exchange



Fig. 17.4 In situ non-articulating knee spacer

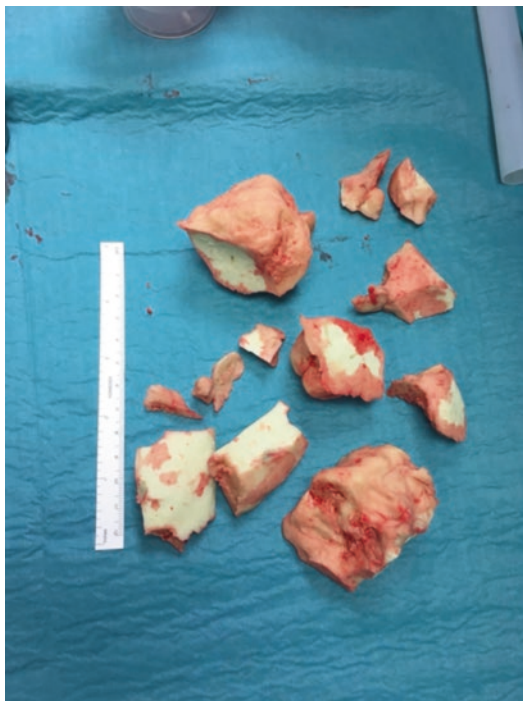


Fig. 17.5 Removal of the knee spacer

include persistent infection, wound-related problems, draining sinus, or mechanical complications such as spacer dislocation or fracture [55, 56].

17.9 Cement Spacer Irrigation and Debridement

Cement spacer irrigation and debridement is an alternative to cement spacer exchange to reduce the microbial bioburden. However, for this procedure, there is a lack of evidence on the practical benefits. Moreover, repeated cement spacer irrigation, without antibiotic spacer exchange, does not seem to have any evidence [57].

17.10 Infected Bilateral Knee Arthroplasties

Limited evidence is available to recommend a single-stage or two-stage revision procedure for infected bilateral knee arthroplasties [58–60]. Two-stage revision is made placing an antibiotic-

Table 17.1 Advantages and disadvantages of one-stage vs. two-stage PJI revision procedures

| | One-stage | Two-stage |
|---------------|---|--|
| Advantages | <ul style="list-style-type: none"> • Lower morbidity and higher functional outcomes • Higher quality of life • Lower recovery rates • Lower cost • Technically easier • Quicker rehabilitation | <ul style="list-style-type: none"> • Targeted micro-organism eradication • Lower rate of reinfection • Extensive debridement |
| Disadvantages | <ul style="list-style-type: none"> • Unable to direct antibiotic in cement to specific organism • Unable to observe response to antibiotic therapy • Higher reinfection rate • Only one debridement • Able to eradicate distant sites of infection | <ul style="list-style-type: none"> • Higher morbidity and mortality • High complexity of the surgical technique • Higher rate of recovery • Lower quality of life • Higher surgical risks • Higher cost • Slower rehabilitation |

impregnated cement spacer for at least 6–8 weeks before reimplantation. The authors reported that patients could wait for several days between each side reimplantation or perform a simultaneous bilateral revision surgery [58–60]. The decision to perform simultaneous bilateral revision surgery should consider several factors, such as the patient's comorbidities and functional status. Wolff et al. [61] demonstrated improved outcomes with a simultaneous two-staged revision when compared with irrigation, debridement, and prosthetic salvage. However, concerns exist about the morbidity of a two-stage revision and the immobility on both extremities during the antibiotic spacer period (Table 17.1).

17.11 Conclusion

Limited evidence is available on the superiority of two-stage over one-stage revision in terms of success, eradication of infection, or patient satis-

faction [2–4]. Future studies are necessary to delineate the superiority of a one- or two-stage revision approach.

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The Use of Static Spacers in Periprosthetic Knee Infections

18

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18.1 Introduction

The increasing number of total knee arthroplasty (TKA) being performed has led to a corresponding increase in the overall number of TKA infections. Periprosthetic knee infection is a severe and not infrequent complication, with an inci-

dence ranging from 0.4 to 2.5% for primary TKA and 4 to 8% for revision surgery.

The surgical treatment differs depending on the duration of the infection. The aim is to eradicate infection and maintain satisfactory knee function (range of motion, stability, no pain). For acute infection, prosthesis removal is not necessary and a simple DAIR (debridement, antibiotics, implant retention) should be performed in association with replacement of the polyethylene insert. For subacute or chronic infection, prosthetic replacement is necessary, and two methods of management can be discussed: single-stage or two-stage exchange arthroplasty.

Single-stage exchange arthroplasty involves implant removal with debridement, followed by reimplantation of a new prosthesis during the same operation. Although single-stage exchange knee arthroplasty is possible in certain specific cases, prosthetic replacement in two stages is currently considered as standard treatment. The indications of single-stage exchange are absence of systemic sepsis, minimal bone loss and soft tissue defects, absence of difficulties related to the skin, and preoperative isolation of a pathogenic organism which is sensitive to bactericidal treatment.

During two-stage exchange arthroplasty, the first stage is to remove all prosthetic materials with thorough debridement of the periprosthetic tissues. Multiple tissue samples are collected during the first stage debridement. An antibiotic-

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impregnated cement spacer is positioned in place of the TKA implants. Later, once the infection is controlled, prosthesis reimplantation is performed during the second stage. The optimal delay before the second surgery is still debated. The use of a cement spacer is practically systematic in the treatment of TKA infection, because it allows the preservation of sufficient joint space during the intermediate period without a prosthesis, which allows maintenance of the space for reimplantation of the new prosthesis during the second stage surgery. There are two types of spacer commonly used: static spacer or dynamic spacer. Both types of spacer have advantages and disadvantages. A good understanding of the spacer function and indications is critical for appropriate management of the two-stage exchange knee arthroplasty.

In this chapter, we will discuss the characteristics and use of a static spacer, the surgical technique, and outcomes using a static spacer.

18.2 General Spacer Properties

A spacer is a temporary piece of organic cement. After removal of the infected implant and tissue, the principle is to create a cement-based replacement prosthesis, shaping them manually or using moulds.

18.2.1 Mechanical Properties

The role of the spacer is to stabilize the femoro-tibial joint during the intermediate time between surgical stages, to prevent knee dislocation and avoid pain. Adequate knee stability during this period protects the periarticular soft tissue, such as the extensor mechanism and avoids additional tissue injuries. It also limits fibrosis filling the joint space and limits ligament and tendon retraction. Thus, using a spacer facilitates reimplantation surgery during the second stage. Without the use of a spacer, the knee ligaments significantly retract, possibly necessitating further bone resection, and therefore leg shortening, to create space for reimplantation of a new prosthesis or neces-

sitating ligament release and implantation of a highly constrained or hinged prosthesis.

18.2.2 Anti-Microbial Properties

Whilst the patient is receiving appropriate systemic antibiotic therapy, spacers are also delivering high doses of antibiotics directly within the knee. The local diffusion of antibiotics contained within the spacer facilitates the eradication of the microbes and limits the development of secondary infection. The antibiotics present in cement are usually aminoglycosides such as gentamycin or tobramycin and or glycopeptide such as vancomycin. The dose delivered locally is ten times greater than the critical minimum inhibitory concentration (CMI) for antibiotic activity. In addition, the spacer fills the femoro-tibial space and reduces the risk of secondary infection by limiting the volume of intra-articular dead space.

18.2.3 Two Types of Cement Spacer

Antibiotic-impregnated cement spacers are static or dynamic.

The static spacer consists of a single block of cement inserted between the femur and the tibia (Fig. 18.1). It is not articulated and constitutes a temporary knee arthrodesis keeping the knee in full extension. This temporary immobilization leads, among other things, to joint stiffness and exposure difficulties at the time of reimplantation.

As a result, dynamic spacers have been developed to solve these problems. The dynamic spacer consists of a femoral component articulated on a tibial baseplate. It is effectively a temporary prosthesis made out of cement. With a smooth and congruent interface, the articulated spacers are designed to allow knee range of motion. Thus, it allows passive mobilization of the knee during the intermediate phase. The dynamic spacer reduces the risk of muscular atrophy and retraction of the peripheral soft tissues.

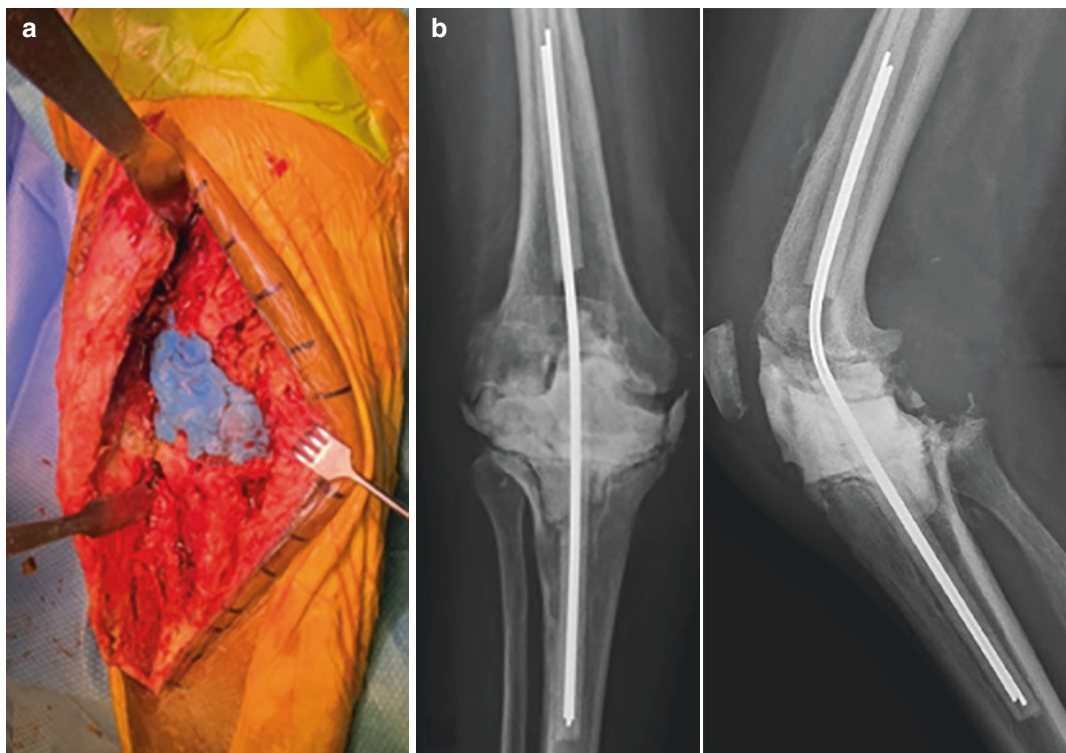


Fig. 18.1 (a) Perioperative photo of a static spacer with methylene blue denatured cement. (b) Postoperative radiograph after insertion of the static spacer reinforced with Kirschner wires

18.3 Indications for a Static Spacer in TKA Infections

The use of a spacer is indicated for subacute or chronic TKA infections requiring a two-stage revision. In the absence of contraindications, the dynamic spacer should be preferred, because it improves the knee function, as well as postoperative mobility and facilitates the exposure during the reimplantation.

The indications for a static spacer correspond to the contraindications of the dynamic spacer, specifically:

- Major bone loss, which is associated with a high risk of fracture, as well as a lack of fixation for a dynamic spacer (Figs. 18.2 and 18.3).
- An incompetence of the collateral ligaments or of the extensor mechanism, which can

cause femoro-tibial dislocation with a dynamic spacer (Fig. 18.3).

- A skin condition at high risk of complications, needing a limitation of flexion or even an immobilization of the knee to promote healing.

18.4 Surgical Technique

Knee exposure can be performed via a pre-existing scar or as per surgeon preference. After knee exposure, the level of the joint line is identified and measured relative to a drill hole which is made on the femur and the tibia at a safe distance from the joint level. The prosthesis is carefully explanted, and the surrounding contaminated tissues are excised. The femoral and tibial intramedullary canals are reamed and cleaned. After multiple tissue samples are taken, a thorough knee joint lavage is performed.

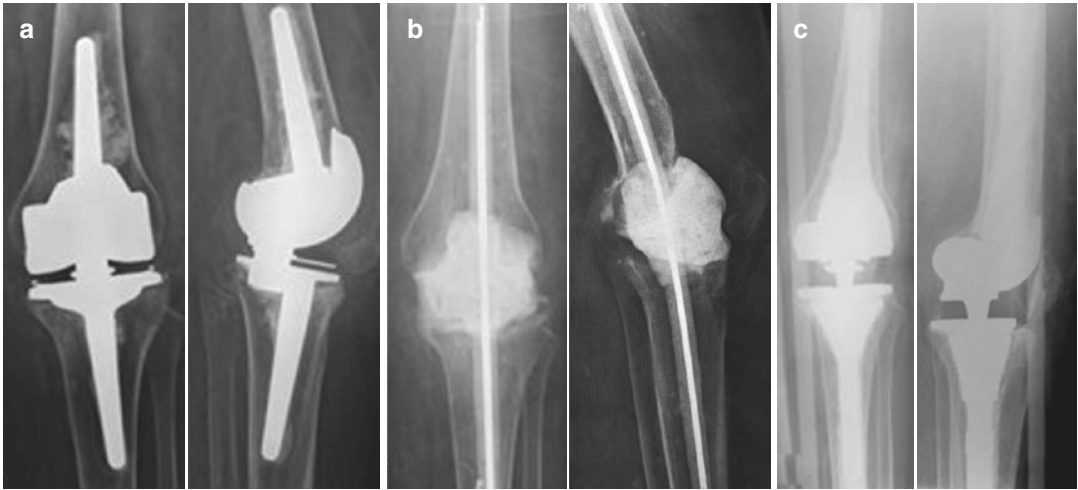


Fig. 18.2 Case report: A 57-year-old man had septic loosening of his revision TKA and chronic rupture of the quadriceps tendon. After removal of the TKA, there was major femoral and tibial bone loss. (a) Radiograph before revision showing loosening. (b) Radiograph after inser-

tion of the static spacer. (c) Radiograph after reimplantation of a hinge knee prosthesis, associated with reconstruction of extensor mechanism using the Hanssen technique

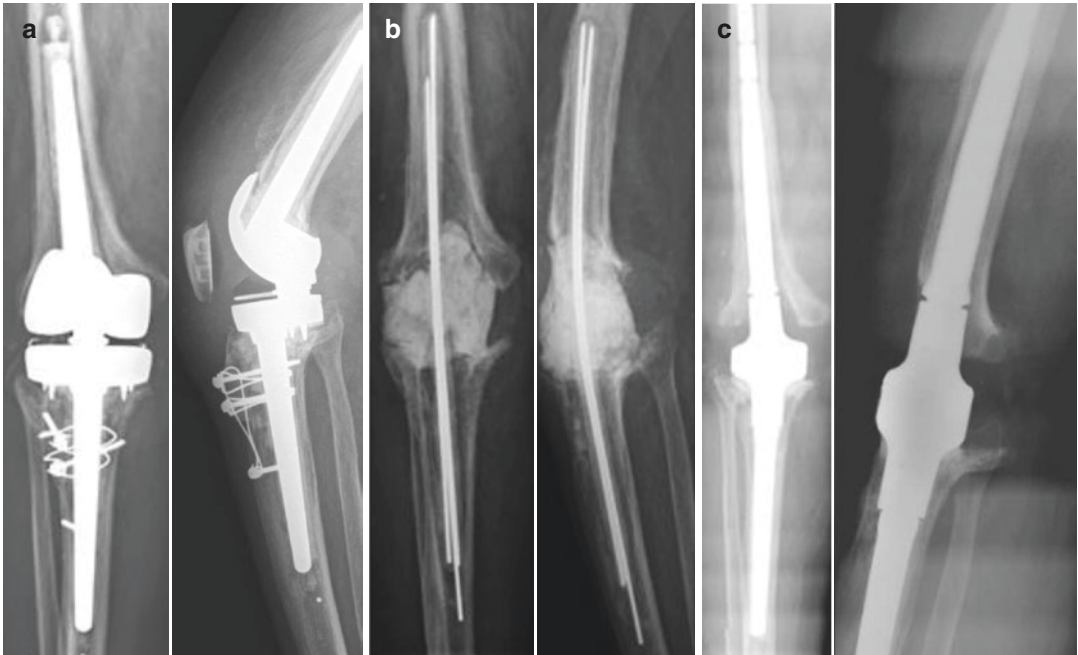


Fig. 18.3 Case report: A 69-year-old man had chronic sepsis of his revision TKA and rupture of the allograft extensor mechanism. (a) Radiograph before revision. (b) Radiograph after insertion of the static spacer with rein-

forcement Kirschner wires. (c) Radiograph after reimplantation of an arthrodesis prosthesis at the second stage revision

Static cement spacers must be reinforced with wires. Without reinforcement, the risk of spacer fracture is very high.

The first step is to create a rod of cement around Kirschner wires that will go into the intramedullary canal. The surgeon should use 3 or 4 wires of approximately 2 mm diameter and high-viscosity antibiotic cement. Manually, the surgeon fashions a rod of cement with the wires inside. The length must be long enough to have at least 6 cm of rod in each femoral and tibial canal, plus the length of the joint space. Once set, this rod, marked at its centre, is introduced back and forth into the femoral and tibial canals until the centre mark is at the midpoint of the joint space.

During the second step, high-viscosity antibiotic cement, with the addition of methylene blue, is prepared. This second cementation will fill the joint space, to maintain the native leg length. At 2 min after the mixing of the second cement mixture, the joint is opened with traction in leg extension, in order to fill any bone defects and the articular space. All of the knee joint space is filled with the second cementation. The size of the spacer should be appropriate, and not so large as to cause excessive skin tension during wound closure.

This second cementation stabilizes the construct and prevents spacer migration. The methylene blue is used to facilitate cement removal during the second stage surgery and causes poor fixation between bone and cement, which also aids removal. The joint capsule, subcutaneous tissues, and the skin are closed in layers.

Postoperatively, patients are kept in a brace without range of motion and locked in extension. Weight bearing is forbidden.

During the second stage surgery, the surgeon removes the cement spacer by breaking the spacer and removing the rod spacer (with wires inside). An osteotome and mallet can be used to carefully crack the fragment of the cement spacer. Another thorough debridement is performed prior to implantation of the new definitive prosthesis.

18.5 Results and Complications of Static Spacers (Table 18.1)

In the context of chronic TKA infections, several studies have compared infection management using articulated and static spacers. A meta-analysis published in 2017, including 10 studies, compared the effectiveness of static and dynamic spacers according to several criteria, specifically: rate of infection eradication, range of motion and functional scores, and soft tissue release during prosthetic reimplantation [1].

18.5.1 Rate of Infection Eradication

In a study of 81 static spacers and 34 dynamic spacers, Johnson et al. [2] found that the rate of infection eradication was 88% for the static spacer group and 82% for the dynamic spacer

Table 18.1 Literature review of the use of static and dynamic spacers during two-stage prosthesis exchanges

| | Date | Type of study | Number of static spacer | Infection eradication | Range of motion | Mean KSS function score | Mean HSS score | Lengthening of the femoral quadriceps | TTO |
|------------------------|------|---------------|-------------------------|-----------------------|-----------------|-------------------------|----------------|---------------------------------------|-----|
| Brunnekreef et al. [4] | 2013 | Retrospective | 9 | 100% | 73.8° | – | – | – | 55% |
| Chiang et al. [6] | 2011 | Prospective | 21 | 90% | 85° | – | 82 | 33% | – |
| Choi et al. [2] | 2012 | Retrospective | 33 | 67% | 97° | – | – | 18% | 57% |
| Emerson et al. [13] | 2002 | Retrospective | 26 | 92% | 93.7° | – | – | – | – |
| Fehring et al. [7] | 2000 | Retrospective | 25 | 88% | 98° | – | 83 | 8% | – |
| Freeman et al. [8] | 2007 | Retrospective | 28 | 89% | – | 45 | – | – | – |
| Hsu et al. [9] | 2006 | Retrospective | 7 | 85% | 78° | 57.8 | – | 28% | – |
| Johnson et al. [3] | 2012 | Retrospective | 81 | 82% | 95° | – | – | – | – |
| Jämsen et al. [14] | 2006 | Retrospective | 8 | 75% | 92° | 53 | – | – | – |
| Park et al. [5] | 2010 | Retrospective | 20 | 85% | 92° | 50 | 80 | 35% | 4% |

group. This rate was comparable in the two groups. Choi et al. [3] found lower, but comparable, infection eradication rates with 67% for the static spacer group and 71% for the dynamic spacer group. In a study by Brunnekreef et al. [4] 35 patients underwent two-stage revision surgery for chronic infection on TKA. The infection eradication rates were 100% for both the static and dynamic spacer groups.

Thus, the rate of eradication of infection using a static spacer is between 67% [2] and 100% [3]. There is no significant difference between static and dynamic spacers.

18.5.2 Range of Motion

Regarding range of motion, Park et al. [5] compared the clinical results of static and dynamic cement spacers for the treatment of infected TKA in 36 patients. They found a significant difference between groups: an average flexion at the last follow-up of 92° in the static spacer group versus 108° in the dynamic spacer group. In a study of 45 patients, Chiang et al. [6] reported similar results, with 85° of flexion in the static spacer group versus 113° in the dynamic spacer group.

In the literature review by Hai Ding et al. [1], the average flexion at the last follow-up is between 74° and 98°. Flexion was significantly lower after static spacer use compared to dynamic spacer use.

18.5.3 Knee Society Score (KSS) and Hospital for Special Surgery Knee Score (HSS)

Park et al. [5] and Freeman et al. [8] found an average KSS functional score of 50 and 45 points, respectively, in the static spacer group versus 76 and 70 points in the dynamic spacer group. Chiang et al. [6] and Park et al. [5], respectively, found an average HSS score of 82 and 80 points for the static spacer group against 90 and 87 points for the dynamic spacer group.

The functional scores at the last follow-up are comparable between different studies. These

scores are significantly lower in the static spacer groups compared to the dynamic spacer groups.

18.5.4 Rate of Surgical Soft Tissue Release

Several authors have sought to assess the retraction of peripheral soft tissues during prosthetic reimplantation and particularly the need to perform quadriceps tendon release or tibial tuberosity osteotomy (TTO).

In a study of 28 patients, Hsu et al. [9] performed two rectus femoris snips and one Y-plasty of the quadriceps tendon during prosthetic reimplantation. They found that 29% of patients in the static group required a more extensive approach compared to only 5% of patients in the articulated group. Choi et al. [2] found that a more extensive approach was more frequently required in the static spacer group than in the dynamic spacer group (5 rectus femoris snips, 1 Y-plasty of the quadriceps tendon and 19 TTO in the static spacer group versus 3 rectus femoris snips and 1 TTO in the dynamic spacer group).

Therefore, the use of articulated spacers facilitates the surgical exposure during the prosthetic reimplantation stage. The mobilization of the knee between the two surgeries avoids the retraction of the extensor mechanism and the articular capsule [10].

18.5.5 Complications

Johnson et al. [3] described complications requiring surgical revision due to dynamic spacers. Four of the 34 patients with dynamic spacers presented with mechanical failure and there were no failures of the 81 static spacers. Two patients with dynamic spacer failure who admitted to having resumed full weight bearing presented with fractures of the femoral component. The other patients presented with a dislocation of the femoral component and a subluxation of the tibial component with skin breakdown who needed flap coverage. In a study by Streulens et al. [11], the dynamic spacer dislocated and caused significant knee subluxation in 7% of the patients.

Wilson et al. [12] described a series of 3 complicated cases of anterior migration of the cement with partial or even total rupture of the patellar tendon following the implantation of dynamic spacers.

Thus, static spacers have less risk of complications than dynamic spacers (Figs. 18.4, 18.5, and 18.6).



Fig. 18.4 Case report: Radiographs of the knee of a 69-year-old man showing an anterior subluxation of the tibial cement spacer

18.6 Conclusion

Two-stage prosthetic replacement, with the use of a cement spacer during the intermediate phase, is currently considered as the gold standard treatment for chronic prosthetic knee infections.

During prosthetic reimplantation, static spacers are associated with retraction of peripheral soft tissue and greater difficulty in surgical exposure. This difficulty in exposure is related to the immobilization of the knee during the intermediate phase and may require an important soft tissue release. The use of a static spacer impacts the functional knee results of patients.

Articulated spacers allow limited knee mobilization between the two surgical stages and can facilitate the ease of prosthesis reimplantation during the second stage. However, the dynamic spacers are associated with a greater number of complications compared with static spacers, particularly in cases of improper use. When there are contraindications for the use of a dynamic spacer, a static cement spacer is preferred, such as when there is major bone loss, knee instability with collateral ligament or extensor mechanism incompetence, or when there is a precarious skin condition.

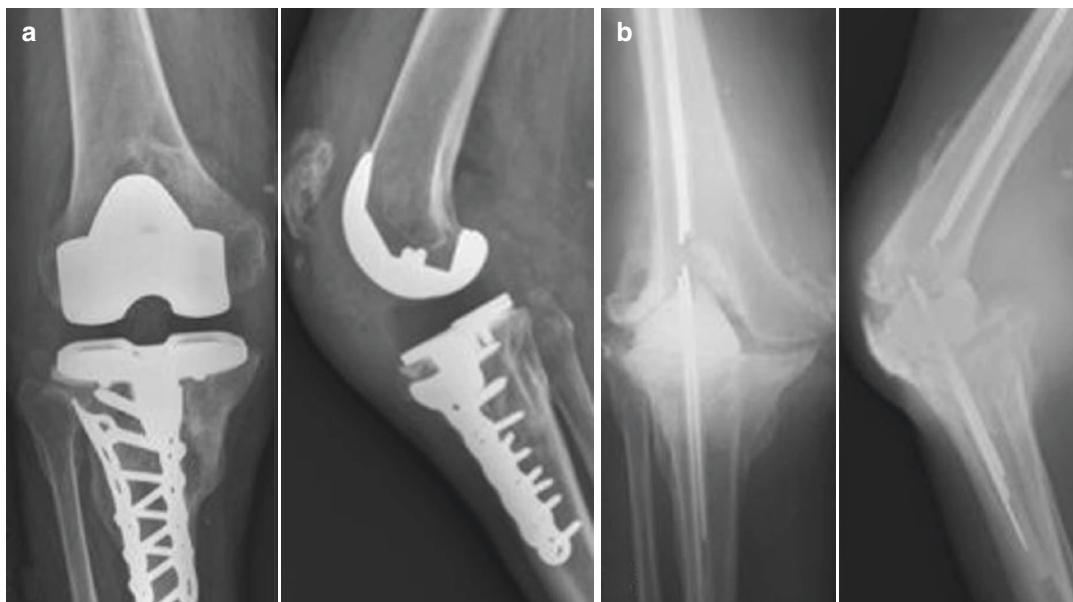


Fig. 18.5 Case report: A 69-year-old man had chronic sepsis of his TKA with chronic rupture of the patellar tendon. (a) Radiograph before revision. (b) Radiograph after the first stage revision showing a broken static cement spacer



Fig. 18.6 Case report: A 75-year-old man had chronic sepsis of his TKA with major femoral bone loss. (a) Radiograph before revision. (b) Radiograph showing a

broken massive static cement spacer. (c) Radiograph after reimplantation of rotating hinge distal femoral replacement prosthesis during the second stage revision

In order to minimize the risk of complications of spacers during the intermediate phase, the surgical technique and the indications of each type of spacer must be well known and understood.

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Dynamic (Mobile) Spacers in Infected Total Knee Arthroplasty

19

M. Enes Kayaalp and Roland Becker

19.1 Introduction

Two-stage exchange for infected knee arthroplasty is the most common method for prosthetic joint infection [1]. During the interim following removal of implants, the resection area is supported by implantation of spacers. There are two kinds of options: dynamic or static spacers.

Articulating spacers are called dynamic or mobile spacers, or simply articulating spacers. These spacers are assumed to provide some advantages over the static, i.e., non-articulating ones, in selected patients. This chapter will focus on the historical aspects and current scientific evidence on dynamic spacers.

The first reported two-stage treatment of prosthetic joint infection was in 1983 by Insall [2]. During the course of following clinical applications, it was revealed that the use of antibiotic-impregnated cement following removal of implants and until reimplantation significantly

increased the success rate of revision due to infection. Followingly, placement of antibiotic-impregnated cement rather than leaving the joint empty became a routine procedure. However, as experience gathered, it was revealed that static spacers caused unexpected bone loss due to migration of unstable spacer [3]. This led to introduction of dynamic spacers, which would theoretically decrease complications occurring when static spacers are used, such as bone loss, immobility-related problems, and adhesions around the operation site.

Currently, reliable scientific evidence in favor of neither type exists in the literature [4]. This is mainly caused by the multifactorial setting between cases and studies.

19.2 Types and Properties of Dynamic Spacers

Dynamic all cement spacers were shown to provide similar eradication rates as static spacers [1]. Lack of apparent disadvantages and quest for betterment in terms of range of motion and interim mobility by maintaining muscle activity lead to their widespread use. However, almost synchronously, different types were reported in the literature.

Dynamic spacers can be handmade by the operating surgeon, or re-sterilized components might be used during the interim. The first applied dynamic spacer was shaped antibiotic-

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impregnated cement by surgeon, either manually or using molds. This handmade facsimile of a knee replacement was followed by a molded prosthesis and commercially available pre-shaped and posterior stabilized designs [5].

Essentially, three types of dynamic spacers might be distinguished:

1. intraoperatively prepared cement spacers (either handmade or using molds) (Fig. 19.1),
2. re-sterilized components,
3. commercially available pre-shaped components (with or without metal or polyethylene components) [4, 6].

Regarding the contact area properties of articulating parts, one can also classify the dynamic spacers as:

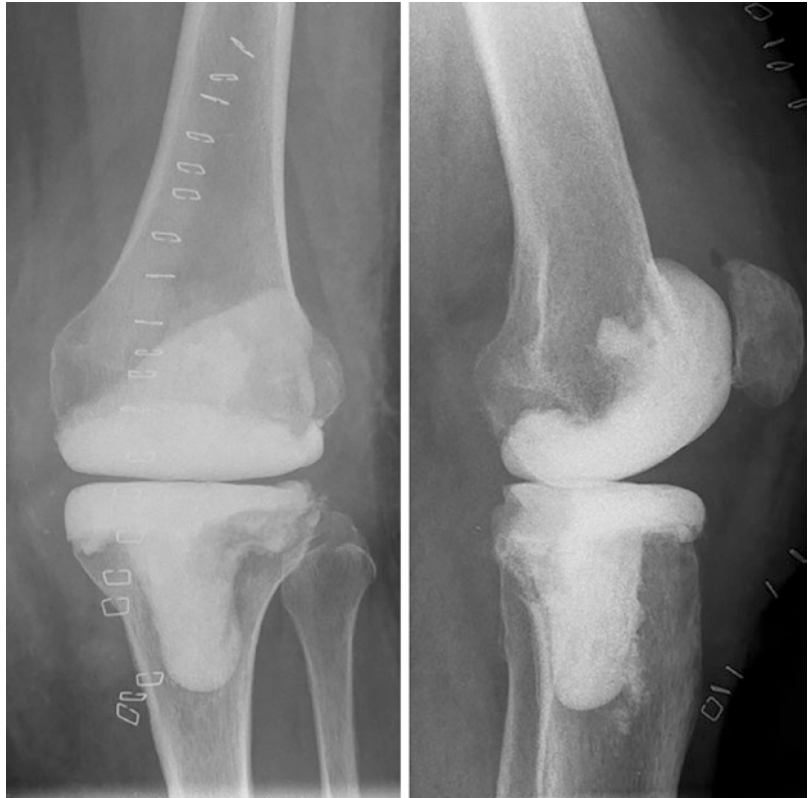
1. cement on cement
2. cement on polyethylene
3. metal-on-polyethylene.

Although variations exist regarding the preparation technique of handmade or molded spacers, there seems to be a consensus on using antibiotic-impregnated cement for their preparation [7].

Various authors reported good eradication rates of infection using dynamic cement spacers compared to static ones, wherefore the use of dynamic spacers became widespread [8–10].

Conversely, Hofmann et al. [11] used re-sterilized components in the interim, and many authors also reported successful results with this technique [1]. Chen et al. reported good results with autoclaved metal-on-cement spacer with satisfactory interim ROM without additional costs [12]. Regarding autoclaving of used components, potential legal implications and lack of standards were the primary concerns of operating surgeons, so this method lost interest. This method is now reserved for restricted conditions, where economic or logistic considerations do not allow for cement or commercially available spacers [13].

Fig. 19.1 Postoperative images of a patient implanted with handmade dynamic cement-on-cement spacer following stage one



As experience gathered, different authors proposed modified techniques. Akhtar et al. proposed using intramedullary pedestals. Their arguments consisted of obtaining a more stable spacer to prevent dynamic cement spacer-related complications such as fractures, dislocations, and malalignment, which were reportedly found at a level above expectations [7, 14].

Shen et al. proposed to use molded cement spacers created intraoperatively using prosthesis trials to overcome issues such as lack of proper antibiotic amount and type when commercially available spacers are used and shape and congruency related problems when hand-molded spacers are implanted. The authors reported promising results with their technique [15].

Functional demands and increasing expectations from patients, together with surgical tendency to ease and standardize the procedure, commercially available spacers found more recognition and area of use. As a result, the production of commercially available pre-shaped, ready to implant dynamic spacers, which include limited polyethylene and metal components, got more widespread [5].

One of the concerns about handmade cement spacers would be the mechanical fatigue strength of these spacers. Hand-molded spacers were compared with commercially available spacers in terms of mechanical strength [16]. The authors concluded that hand-molded spacers provide sufficient strength and that their use will be cost-effective in revision total knee arthroplasty. Similar functional results with handmade dynamic spacers compared to expensive, industrial spacers have been reported by others [17].

No difference in terms of eradication rate has been reported between the usage of a handmade spacer with 92.2% (84 reinfections) and an industry-made spacer with 90.5% [7].

However, Citak et al. observed that surgeon-made dynamic spacers were more likely to fracture than industry-made spacers despite having equivalent functional outcomes and infection eradication rates [18]. Similarly, other authors reported no increase in infection rates when industrial spacers with metal and polyethylene components were used [5].

An analysis of 1525 infected TKA cases showed no difference in functional outcome when manufactured versus surgeon-made dynamic spacers are used in the knee. The mean flexion at latest follow-up was tendentially higher with a mean of 102° using a handmade spacer compared to a mean of 90° using a manufactured spacer [7]. However, the clinical relevance of the difference in terms of range of motion remains questionable.

As of today, no conclusive evidence favors any of these methods one over another [10]. Prefomed spacers have the disadvantages of fixed antibiotic dose and type, and whereas using cement and antibiotics, surgeons can prepare hand-molded spacers intraoperatively. The disadvantages of this method are less congruent contact and more fragile cement due to mixture with antibiotics [4].

19.3 Advantages of Dynamic Spacers

The claimed advantages of dynamic spacers are various. However, as will be seen, there are also contradictory results within the literature. Due to the heterogeneity of studies and cases, a clear deduction in terms of advantages cannot be made with a high level of evidence.

Expert opinions and case series mostly report several advantages of dynamic spacers, consisting of maintaining joint motion, facilitating surgical exposure at the time of reimplantation, and an enhanced postoperative function [1, 5, 7, 13]. Conversely, selection bias and case-related restrictions and selections may hinder objective deductions on this issue.

The first and most important argument in favor of dynamic spacers is the comparable infection eradication rates as with the static spacers [6].

Dynamic spacers were shown to constitute a safe alternative to fixed spacers in two-stage revision for infected total knee arthroplasty, which equally preserves ligament balancing and has equal infection eradication rates. However, the authors did not observe a long-term improvement of the range of motion following reimplantation of the new joint [19].

Other authors also did not report any significant difference in ROM at a minimum of 2 years follow-up between dynamic and static spacers. Citak et al. showed that published studies reported a 96.4° of knee ROM after an average follow-up of 44.3 months using dynamic spacers, compared to 91.2° at an average follow-up of 52 months using static spacers. However, experts seem to favor dynamic spacers, despite the lack of significant difference [7].

In contrast, some studies found better ranges of motion not only during the interim but also following reimplantation of revision components after the usage of dynamic spacers. Park et al. compared 36 consecutive patients, 20 of whom received static and 16 received dynamic spacers. The authors concluded that dynamic spacers appear to provide a better range of motion and less functional limitations to the patients, wherefore they should be used whenever possible. The reported knee ROM at a mean follow-up time of 36 months following revision surgery was 92° (range 65–140°) in patients with a static spacer in the interim versus 108° (range 85–140°) in patients with a dynamic spacer [20].

Villanueva-Martinez et al. stated that a mobile and functional joint is a key factor related to a successful outcome with a two-stage reimplantation procedure. Moreover, they have the advantages of preserving bone stock, delivering high concentrations of antibiotics, facilitating patient comfort, and early hospital discharge [17]. The ease of reimplantation and ease of rehabilitation was the most important argument for some surgeons favoring dynamic spacers [5].

It has been stated that selection bias in the majority of studies comparing static and dynamic spacers exists, wherefore no clear conclusion can be drawn on the ideal type of spacer. However, the authors proposed to use dynamic spacers whenever possible, as a result of clinical evidence on dynamic spacers suggesting improved function, better satisfaction, reduced hospital stays, and better ROM [13].

A systemic review also showed no differences regarding infection control between static and dynamic spacers in the treatment of infected TKA [21].

19.4 Contraindication of Dynamic Spacers

Although there are no clear contraindications for the application of dynamic spacers, technical feasibility problems, lack of soft tissues or ligaments around the knee, or extensive bone deficiency are the major concerns that redirect surgeons to static spacers.

Pivec et al. draw attention to the differences between case series and questioned the quasi advantages of dynamic spacers over static ones due to the fact that more simple cases are mostly implanted with dynamic spacers for the interim, whereas complex cases with soft tissue or bone stock problems get static spacers. The authors also underlined the differences between former and modern static spacers, indicating a need for higher-level evidence for future clinical applications [9].

It was shown that as an expert preference, in patients with soft tissue compromise, surgeons tend to implant a static spacer to prevent motion and obtain a better healing environment for the soft tissue [7, 13].

19.5 Antibiotic Properties of Dynamic Spacers

Locally applied antibiotics far outweighs the concentration and effect of systemic antibiotics and are therefore preferred. This is done by mixing a powder form of antibiotic with the cement. There is also commercially available antibiotic-impregnated cement. However, the elution of antibiotics from the cement decreases over time, and bacterial colonization can occur on the spacers [22–25]. Supporting this argument, Nelson et al. showed that sonication of antibiotic spacers at the time of second stage operation predicted revision arthroplasty failures due to another infection [24]. Therefore, the interim length should be limited up to 6 weeks. After that time, the risk of colonization of the spacer becomes higher than its antibiotic effect itself.

For an antibiotic to be mixed into the cement, the first and most important requirement is to be

thermostable. Further desired properties of antibiotics are broad-spectrum, efficiency at low concentrations, and low risk of allergy [1]. Antibiotics are suggested to be mixed by hand in a bowl without a vacuum. Some fillers such as Xylitol, sugar alcohol, or Ancef were proposed to improve the elution of active antibiotics [7].

The most commonly used antibiotics are gentamicin, tobramycin, and vancomycin [1]. Antibiotics such as vancomycin, gentamicin, ampicillin, clindamycin, and meropenem can be used as a combination based on the causative organism and its susceptibility [26, 27].

The higher dose of antibiotics mixed with cement is a concerning issue. However, the commonly accepted limit of 5%–10% antibiotic for the whole cement mass is reliably exceeded in the clinical practice when spacers in two-stage revision are concerned [6]. Although it is a known fact that higher mixing rates decrease the fatigue strength of the cement, the desired antibiotic effect far outweighs this flaw. Even so, it must be remembered that the addition of more than 4.5 g of powder substantially weakens the cement [7].

Reported doses for selected antibiotics from various clinical studies are listed in Table 19.1. Although different antibiotics are recommended, the best choice of treatment depends on the antibiogram result obtained from the individual patient's samples prior to revision surgery.

Table 19.1 Reported doses of selected antibiotics from clinical studies

| Antibiotic type | Dose in g per 40 g cement |
|--|---------------------------|
| Vancomycin | 0.5–4 |
| Gentamicin | 0.25–4.8 |
| Tobramycin | 1–4.8 |
| Cefazolin (first generation cephalosporin) | 1–2 |
| Cefuroxime (second generation cephalosporin) | 1.5–2 |
| Ceftazidime (third generation cephalosporin) | 2 |
| Cefotaxime (fourth generation cephalosporin) | 2 |
| Ciprofloxacin | 0.2–3 |
| Clindamycin | 1–2 |
| Tazobactam | 0.5 |
| Meropenem | 0.5–4 |

Gentamycin and/or vancomycin are the most frequently used antibiotics [28]. Gentamycin is a bactericide against gram-negative coccus and vancomycin against gram-positive microorganisms.

Infection caused by methicillin-resistant staphylococcus aureus (MRSA) or methicillin-resistant staphylococcus epidermidis (MRSE) should be treated with vancomycin added to the cement. Infections with vancomycin-resistant enterococcus (VRE) or multidrug-resistant organisms should be treated with individual decision-making with consultation with infectious disease specialist [13].

Kuzyk et al. stated that they used 4 g of vancomycin and 4 g of ceftazidime per 40 g of cement when the infecting organism is unknown. The authors detail that they use three packs of 40 g cement bags to make their spacers [29].

Besides the antibiotic-loaded cement, appropriate IV antibiotic management is compulsory. A management plan should be discussed with the microbiologist and infectious disease specialist of the hospital.

Antifungal agents, such as amphotericin B and voriconazole, have also been reported to be mixed with cement [4].

19.6 Interim Period Length and Mobilization with Dynamic Spacers

19.6.1 Length of the Interim

Regarding the length of the interim, different approaches exist. A mainstream approach is to wait for 6 weeks with parenteral antibiotics until the reimplantation [30].

However, different studies report different intervals for reimplantation. Villanueva-Martinez et al. reported an average time of 14 (8–130) weeks [17]. Pitto et al. reported a shorter cement spacer period with mobile spacers than with static spacers (3.3 months vs. 4.2 months) [6]. Kuzyk et al. stated they prescribe intravenous antibiotics for a total of 6 to 8 weeks, ceasing antibiotics for 2 weeks before reoperation. The

authors prolong this treatment for patients, who are immunocompromised, whose soft tissue envelopes are poor, or who have a large draining sinus, or whose infecting organism is especially virulent (e.g., MRSA or MRSE). In these situations, the authors stated that the prolongation with continuous intravenous antibiotics exceeds for a total of 3 to 4 months before reimplantation and consider the use of oral antibiotics for 4 to 6 weeks after reimplantation [29].

Other authors reported a mean interval between stages for their entire cohort to be 15.5 weeks (3.6 to 96.7) using a commercially available dynamic spacer [5]. Another study reported even an interim time of 128.2 ± 80.8 days [31]. In contrast, Fink et al. reported reimplantation after 6 weeks. During this period, antibiotics were given intravenously, followed by 4 weeks of oral treatment [32]. Winkler et al. investigated the effect of interim length on reinfection rate and functional outcome. Taking 4 weeks as a cut-off value, the authors concluded that an interim length of less than 4 weeks had similar results in terms of controlling infection with those of more than 4 weeks of an interval. Moreover, patient inconvenience and care costs were found to be less in patients with a shorter interim. All patients in the study had received antimicrobial treatment for the first 7–10 days, vancomycin or daptomycin combined with ampicillin/sulbactam. Patients received i.v. antibiotics for 2 weeks following explantation and 1–2 weeks following reimplantation. After discharge, they were given oral antibiotics [33].

Pro-Implant Foundation from Germany (<https://pro-implant.org/>) proposed a treatment algorithm for patients undergoing a revision surgery due to an infection. The research group suggested that patients undergoing a two-stage revision with a short interim should be given a total of 3 weeks of i.v. antibiotics treatment, i.e., 2 weeks within the interval, followed by the revision surgery at the second week, and a week of i.v. antibiotics following reimplantation. The group proposed to use 2 weeks of i.v. antibiotics after explantation, followed by 4 weeks of oral antibiotics and reimplantation surgery at the sixth week, followed by a week of i.v. antibiotics, and

5 weeks of oral antibiotics in patients undergoing a two-stage revision with a long interim [34].

Although numerous suggestions exist, there is currently a lack of high-level evidence for the length of interim and antibiotic treatment.

19.6.2 Mobility During the Interim

One of the main advantages of dynamic spacers is the ability to maintain movement during the interim. However, different approaches exist in terms of weight-bearing due to the mechanical susceptibility of the cement spacer caused by mixing of antibiotics and inherently unstable mounting into the resected joint.

Some authors allow partial weight-bearing with the aid of crutches or a cane as tolerated and let patients wear an extension or hinge orthosis of the knee until soft tissue healing had occurred. If slight or mild laxity remained, the brace was worn until reimplantation, the authors stated [17].

Others use preformed cement spacer (InterSpace; Exactech) and wait for a short period of time to allow wound healing, followed by weight-bearing with one or two supports and active range of motion [4].

The limitation of hand-molded spacers in terms of mechanical strength has been highlighted [16]. The authors defined these spacers as potentially unstable and proposed to use an extension brace or orthosis to be worn during ambulation.

Fink et al. used dynamic cement spacers, and the patient was mobilized with crutches, and partial weight-bearing of 20 kg on the surgically treated leg was allowed [32].

19.7 Complications Related to the Use of Dynamic Spacers

Potential disadvantages of dynamic spacers were reported to be related to wound healing, cement fracture, and dislocation of the spacer [1].

Mobile spacers show cement abrasion causing fibrosis of the articular tissue already within

6 weeks [32]. An increase in immunomodulation of synovial tissue has also been reported [35]. Because of these findings, total synovectomy and extensive lavage are recommended at the second stage of surgery to decrease the number of particles and any retained bacteria.

Lanting et al. investigated dynamic spacer-related complications, more specifically coronal and sagittal subluxation, and their effect on postoperative outcome following the second stage of revision. They found that subluxated knees of more than one standard deviation from the mean in the sagittal plane had lower Knee Society Function Scores ($p=0.045$). The authors suggested that the operating surgeons who used dynamic spacers as part of a staged revision protocol should be aware that subluxation may affect on outcome following second stage revision [31]. The risk of subluxation might be reduced when both the femoral and tibial components are molded with a stem to gain stability [14].

According to an expert survey, a strong consensus exists not to revise or reduce dislocated dynamic antibiotic spacers, except for circumstances such as pressure against the skin with imminent necrosis/ulceration, resulting in severe, progressive loss of essential soft tissue or bone, neurovascular compromise, or notable pain and disability for the patient [13].

Chen et al. found less favorable results for static spacers, which resulted in a higher incidence of patella baja and decreased ROM [36].

19.8 Conclusion

Dynamic spacers are recommended whenever possible in two-stage revision for infected total knee arthroplasty. Although the evidence level is low, some benefits were reported favoring dynamic spacers, such as better ROM, shorter hospital stay, and better eventual outcome following the second stage revision.

Handmade cement spacers constitute an important option for these operations at a low cost, providing the possibility of tailoring antibiotic mixture with cement at the desired dose with the desired types, which should be culture and antibiogram specific, whenever possible.

Mobility and weight-bearing should be closely monitored, taking into account the added antibiotic dose to the cement, which would decrease the mechanical properties of the spacer.

The length of the interim period should be carefully planned, and it must be remembered that the elution of antibiotics decreases over time with an increased risk of bacterial colonization on the spacer.

Careful selection of patients to be implanted with dynamic spacers can decrease spacer-related complications and early address of spacer subluxations might be beneficial for better outcome following the second stage revision.

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20.1 Introduction

Knee arthrodesis is an option for limb salvage in complex periprosthetic joint infections for which revision knee arthroplasty cannot be considered. John Key first described compression knee arthrodesis in 1932 in the treatment for tuberculosis of the knee joint [1]. He used a turnbuckle applied across transtibial and transfemoral pins that could be tightened to maintain pressure across the arthrodesis site, with the addition of a circular plaster. He achieved union in 4 out of 5 patients, with the fifth dying of sepsis prior to union. Charnley [2] further expanded on this technique using special screw-clamps and wing-nuts to tighten across the construct until the Steinmann pins bent. His results showed successful arthrodesis in a total of 15 knees: 6 with old tuberculosis and 9 for osteoarthritis.

While knee arthrodesis usually provides good pain relief, it is associated with specific functional limitations and is not usually considered an attractive option for patients. It is therefore seen as a salvage procedure, and the decision as to when to proceed to a knee arthrodesis in the setting of a failed periprosthetic joint infection is a difficult one. Repeating a failed 2-stage revision procedure can lead to further bone loss and may compromise

the soft tissue envelope, both of which may cause difficulties with salvage procedures in the future. Kheir et al. [3] found that 38.4% of patients failed to have their infection controlled after a repeated 2-stage revision. Wu et al. [4] in a systematic review looked at the utility of different treatment options after failure of a 2-stage revision TKA for periprosthetic joint infection. The treatment options included repeat 2-stage revision, knee arthrodesis, above knee amputation, or suppressive antibiotics. Knee arthrodesis was found to be the intervention that was most likely to give the highest quality of life. Knee arthrodesis therefore does have a role, albeit a very limited one, in the salvage of the failed TKR when other reconstructive options are not viable.

20.2 Indications and Contraindications for Arthrodesis

Knee fusion was initially introduced as a treatment for septic arthritis, tuberculosis and poliomyelitis [1, 2, 5, 6]. Before the development of total knee arthroplasty, knee arthrodesis was also used to treat osteoarthritis and rheumatoid arthritis. These days the most common indication for knee arthrodesis is for failure of a total knee arthroplasty, for which revision is not an option due to presence of a persistent periprosthetic joint infection, extensor mechanism defect that is irreparable, or massive soft tissue or bone loss [7,

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8]. Persistence of periprosthetic joint infection in knees that have undergone implantation of an antibiotic spacer is reported in the literature as 9–12% [9]. Knee arthrodesis is also used outside of arthroplasty for treatment of extensive bone or tissue loss, weakness or loss of the extensor mechanism, for the treatment of tumours, and post traumatic arthritis.

Traditional contraindications are a contralateral knee fusion or an ipsilateral hip arthrodesis [10]. The ipsilateral hip and ankle should be supple and free from significant arthritis due to the increased load that will be transferred to these joints post arthrodesis.

20.3 Limitations of Knee Arthrodesis

Knee arthrodesis imparts several physical and psychological limitations to the patient. The inability to bend the knee can cause issues with sitting, climbing stairs and taking public transportation.

A study undertaken on healthy subjects to simulate knee arthrodesis using a brace assessed gait kinematics and kinetics affected by knee arthrodesis [11]. The results showed that compensations for knee immobilisation include (1) increased spinal movement of the lumbosacral spine, (2) increase in vertical excursion (hip-hiking) and transverse rotation of the pelvis on the involved side, (3) increase in extension of the contralateral hip, (4) increased peak flexion in the contralateral knee during swing phase, and (5) a decrease in plantarflexion of the ipsilateral ankle at toe off to help assist with foot clearance. This study looked at the effect of acclimation time, which did not alter the braced gait kinematics, and hence the authors felt the results support a model of longer-term knee rigidity. Marshall et al. [12] found similar results in their study on 2 postoperative knee arthrodesis patients who had achieved union.

Other studies have shown energy expenditure to be higher by 30% than for normal walking in experimental models of knee arthrodesis [13, 14]. In contrast, the increase in energy expenditure for a transfemoral amputation is higher at 50–60%.

20.4 Principles of Knee Arthrodesis

In order to improve the chances of successful knee arthrodesis and infection clearance, the host must be optimised preoperatively systemically. Infection needs to be controlled as much as possible, and the preoperative planning for the knee fusion should optimise available bone contact and required leg length.

20.4.1 Host Optimisation

The patient should be managed in a multidisciplinary team. Modifiable risk factors for infection that may contribute to problems with wound healing should be addressed, such as smoking, diabetic control, cessation of relevant medications, and the nutritional status of the patient. The periprosthetic infection should be managed with antibiotics while also ensuring any treatment of systemic or more widespread infection. If wound issues or soft tissue defects exist, then it is wise to consult a plastic surgeon.

20.4.2 Knee Fusion Position

The ideal position for knee fusion is in 5–7° of anatomical valgus and 10° of flexion, which shortens the limb to help with foot clearance during ambulation and is more practical for sitting than with the knee extended [14]. The aim for limb shortening is 1 cm; however, it is often significantly more than this when performed for failed total knee replacement due to bone loss.

20.4.3 One-Stage Versus Two-Stage Arthrodesis

Arthrodesis may be done as a single-stage procedure or a 2-stage procedure post failure for infection of a total knee replacement. A one-stage arthrodesis involves removal of total knee replacement prosthesis, thorough debridement and lavage, followed by completion of the knee

arthrodesis using the surgeon's preferred technique. Several studies have shown a one-stage procedure to increase the risk of a subsequent deep infection of the arthrodesis if internal implants are used [7, 13, 15]. However, this has been disputed by other studies that have shown a one-stage arthrodesis to be effective in low virulence organisms and in the absence of polymicrobial infections, when either an external fixator or intramedullary device is used [16–20].

A 2-stage arthrodesis involves removal of the total knee implants and thorough debridement and lavage of the joint, followed by application of an antibiotic-impregnated cement spacer and a period of intravenous antibiotics to allow the infection to clear prior to proceeding to the arthrodesis. At the second stage the spacer is removed, and the knee arthrodesis is then performed via the surgeon's preferred technique.

20.5 Techniques for Arthrodesis

Multiple techniques for arthrodesis of the knee have been described in the literature. The two more commonly used procedures are the external fixator and the fusion intramedullary nail.

20.5.1 External Fixator

External fixators include uniplanar and biplanar fixators as well as circular fixators. Circular frames are ideal in that multiplanar deformity correction as well as length can be adjusted through the frame. Circular frames also provide better stability to the fusion construct than the uniplanar and biplanar frames.

There are several advantages of using external fixators for arthrodesis. Blood loss is minimised as only small incisions are needed. Shortening of the limb can be achieved progressively with circular frames, in the setting of large bone defects, where immediate shortening may compromise soft tissues and neurovascular structures. Circular frames can also be used to perform a limb lengthening procedure concurrently with the knee arthrodesis, either through a separate site on the

tibia or femur. External fixation also minimises the presence of internal hardware, which is a potential advantage in cases of established infection.

Complications of external fixators include a high rate of pin-site infections, stress fractures through pin sites, and neurovascular injuries. Disadvantages to the technique, particularly for circular frames, is that it may not be a familiar procedure for many surgeons, and it requires specialist training to be performed successfully.

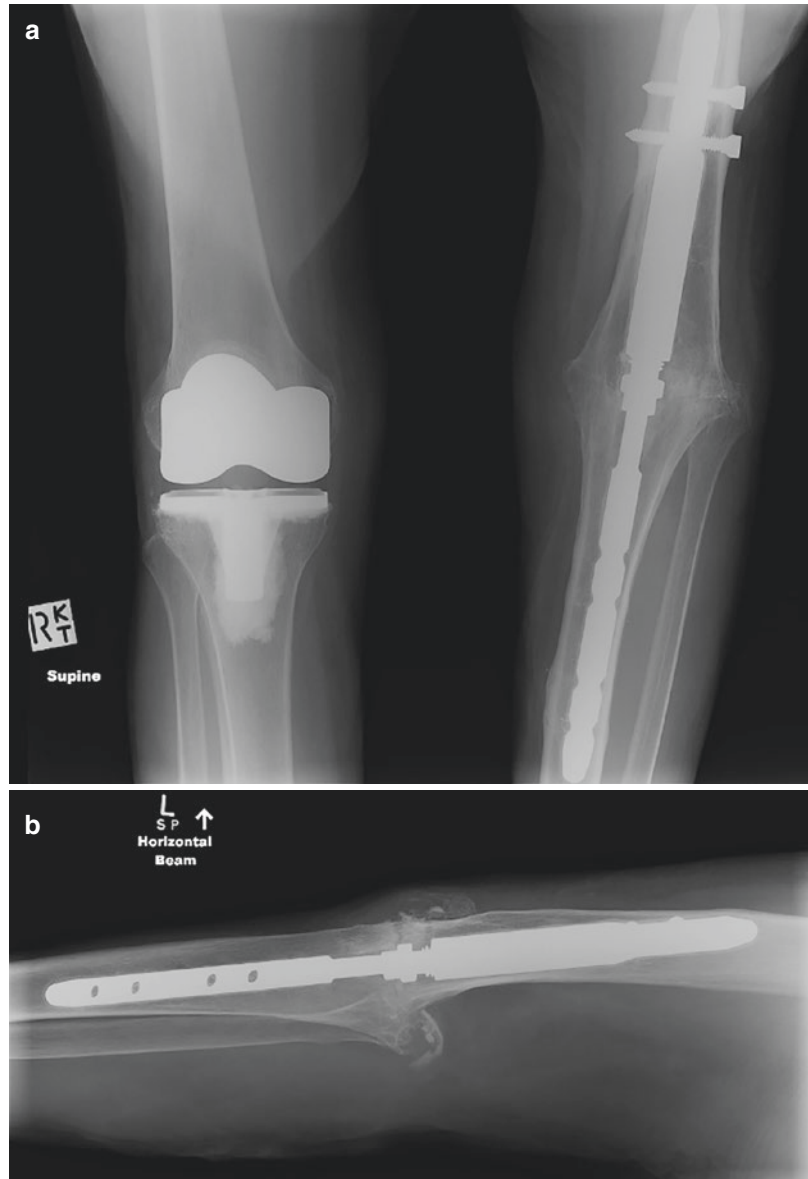
20.5.2 Fusion Nail

A variety of intramedullary nails (IMNs) have been used to achieve knee arthrodesis. Short nails are inserted through the knee joint and are either modular or non-modular. The modular nails have a coupling device that connects the two components, which are inserted into the femoral and tibial canals separately. The coupling device also allows for compression at the arthrodesis site. Short IMNs are ideal when there is an ipsilateral hip replacement.

Long IMNs are inserted through the piriformis fossa and interlocked both proximally and distally, following preparation of the knee fusion site via the knee incision. This technique can be more challenging to control the position of the knee fusion, mainly due to the involvement of the entire length of the femur in the fixation.

Disadvantages of IMNs are the increased operative time and increased perioperative blood loss. The position of fusion is limited by the relationship of the geometry of the nail with the patient's anatomy. The position achieved with an IMN is also fixed at the time of the procedure, unlike external fixation which can allow for ongoing adjustments. IMN may also not be able to be used when there is large deformity in the tibia or femur, or significant bone loss that would cause unacceptable substantial shortening of the limb. Newer technology with IMNs can however potentially facilitate limb lengthening at a further surgical procedure via "lengthening over the nail" or by exchange nailing to an adjustable intramedullary nail. An example of modular short intramedullary nail is reported in Fig. 20.1.

Fig. 20.1 AP radiograph (a) and lateral radiograph (b) of a modular short intramedullary nail (Witchita® Nail Stryker) placed for knee arthrodesis post periprosthetic joint infection with successful fusion



20.5.3 Combined Surgical Techniques

The use of a circular frame until the fusion site begins to consolidate, and then removing the frame and placing an intramedullary device is an example of a combined technique. The advantages of doing this are that no hardware is placed at the area of infection, which has the

theoretical advantage of increasing clearance rates for the prosthetic infection. Removal of the frame prior to complete fusion minimises the risk of pin-site infections from long-term frame use, while the intramedullary nail allows stability and protection of the fusion site to prevent against fusion site fracture. This is particularly useful when bone quality at the fusion site is poor [20].

20.5.4 Dealing with Massive Bone Defects

After prosthesis removal in infected total knee arthroplasty it is not uncommon to have large volumes of bone loss [21, 22]. If arthrodesis is performed at the residual bone ends, then a large leg length discrepancy is the outcome. In an attempt to minimise further surgery on patients who have multiple comorbidities, while maintaining leg length, several techniques have been described in the literature.

Vascularised fibular bone graft (VFBG) can be used to bridge the bone defect. This procedure must be done in a staged fashion for infected total knee arthroplasty so that the graft does not fail due to infection. Other disadvantages include the increase in operative time, donor site morbidity, and the graft itself offers little stability. Rasmussen et al. [23] used this technique in 13 patients, 4 of whom had a failed infected total knee replacement and achieved successful fusion in all patients.

Voss [24] described a technique using a long IMN combined with a cement spacer used to fill the bone defect at the knee site. Similar techniques have been reported using a short modular IMN and antibiotic-impregnated cement spacer [7, 25, 26]. Alt et al. [27] reported on the use of a modular IMN coated with silver and connected with a central spacer, also coated in silver, in an 86-year-old woman for treatment of failed infected total knee arthroplasty. The arthrodesis was done as a 2-stage procedure with cement spacer and cement coated rods placed in the first stage, and the modular IMN with central spacer all coated in silver. There was no recurrence of infection within 26 months follow-up.

Peterson et al. [28] described the use of trabecular metal cones (Zimmer, Biomet) together with a long IMN and autograft to bridge the massive bone loss during the knee arthrodesis for failure of infected total knee arthroplasty. In their series of 6 patients, 5 patients achieved a solid fusion, but one patient required an above knee amputation for a septic non-union.

20.6 Arthrodesis Outcomes

Rates of fusion following knee arthrodesis depend on the mode of fixation [29–33]. External fixation rates of fusion range from 50% to 99% [34], 88% to 100% for intramedullary nails [15], 90% to 95% for modular nails and 85% to 100% where a combined nail and external fixator is applied [15].

Fusion rate following conversion of total knee arthroplasty to arthrodesis is lower than in those knees where joint replacement has not been undertaken. This is theorised to be due to persistence of any infection, poorer bone stock and poorer bone apposition. Knutson et al. showed that after treating 91 failed total knee replacements with conversion to arthrodesis, 82 who had a form of external fixation and the remaining 9 with either intramedullary nail or plate osteosynthesis, only 50% achieved fusion [15]. Robinson et al. [9] showed in their study of 23 knees, a fusion rate of 87% using either intramedullary nail, external fixation or compression plating. Persistent infection has been shown to be high post knee arthrodesis [35–39]. In a study by Rohner et al. [39], the rate of persistent infection was 50%, with persistent infection resulting in patients either undergoing above knee amputation, exchange intramedullary nailing, or developing an established sinus overlying the fusion site [40].

Carr II et al. [40] compared outcomes of patients who underwent above knee amputation (AKA) with those who had a knee arthrodesis for the treatment of failed periprosthetic total knee arthroplasty. The arthrodesis cohort had significantly higher rates of postoperative infection as well as blood transfusion. The AKA patients had a higher rate of systemic complications and in-hospital mortality (3.7% vs. 2.1%). Rohner et al. reported a 50% rate of persistent infection after conversion of TKA to knee fusion [39]. AKA after failed infected TKA is also associated with infection rates reported at 20% [41]. Ambulation after AKA is dependent on a prosthesis, and in a study by Sierra et al. a prosthesis was only fitted in 9 out of 25 patients with limited ambulation achieved in 5 of those patients [41].

Patient reported outcome measures (PROMs) in patients converted from an infected total knee replacement to a knee arthrodesis have been shown to be comparable to patients who have undergone total knee arthroplasty [22]. This study on 8 patients, 7 of whom achieved a successful fusion, was assessed with the Japanese knee osteoarthritis measurement (JKOM) and the Knee Society score (KSS). The JKOM scored comparably to normative data for total knee arthroplasty patients, while the KSS, which has a significant component assessing range of motion, was not surprisingly worse when compared to normative data for total knee arthroplasty. Benson et al. [42] compared 9 patients who underwent a knee arthrodesis for failed total knee arthroplasty (8 for periprosthetic infection) to 9 total knee arthroplasty patients. The SF-36 score and arthritis impact measurement score (AIMS) were assessed, with the SF-36 score being similar between the 2 groups, while the AIMS was better in the TKA group. De Vil et al. also showed comparable SF-36 scores in physical functioning and role-emotional scores between knee arthrodesis and total knee arthroplasty cohorts, with the knee arthrodesis group having better scores for bodily pain and general health; however, the knee arthrodesis group scored worse on mental health and social functioning [35]. Therefore, although outcome scores are reasonable for knee arthrodesis, particularly secondary to good pain relief, they do not function as well as a successful total knee arthroplasty.

20.7 Arthrodesis Complications

Non-union is the most common complication of knee arthrodesis surgery [7]. Factors that affect this are persistence of infection, adequacy of bone stock, and deficiency in fixation construct or bony apposition. The treatment of the non-union is done via fracture principles. In an atrophic non-union, the host biology and any nutritional deficiency should be addressed, as well as optimisation

of control of conditions such as diabetes. Once the patient is optimised the non-union should be taken down and bone grafted with either iliac crest or the addition of a vascularised fibular graft. Hypertrophic non-unions can be treated by revision of fixation to increase rigidity and provide adequate stability to achieve union. Treatment of the infected non-union is difficult. Debridement of the non-union and revision fixation, with repeated cultures taken and antibiotics targeted at the infective organism may be performed as a single-stage or two-stage procedure. While intramedullary nails can be used as the fixation method, this is a situation for which there is a clear advantage of an external fixator to avoid the presence of hardware at the site of infection. Alternatively, if the surgeon believes that the chances of achieving union are unacceptably low, or a patient wouldn't tolerate multiple further procedures, then AKA would be indicated.

Leg length discrepancy of greater than 2 cm has been reported in several studies [37, 43, 44]. Other studies have suggested an average LLD of greater than 5 cm [21, 22]. A small LLD of less than 2 cm is of some advantage in achieving ground clearance during walking in the absence of knee flexion, and generally does not require treatment. If the LLD is between 2 and 5 cm, a shoe raise can be used. LLD greater than 5 cm are difficult to manage with a shoe raise as balance issues tend to occur, and so surgical intervention with distraction osteogenesis may be warranted. Techniques to achieve lengthening include lengthening over an IMN and exchange nailing to a lengthening nail [45].

Other complications include intraoperative fracture, persistent infection, additional superinfection, deep vein thrombosis, wound dehiscence and peroneal nerve palsy. Intraoperative fracture has been described to have an incidence of 6–12% [43, 44]. As previously discussed, persistence in infection is a problem when revising an infected TKA to a knee fusion. The rates reported in the literature for persistent infection range from 6–50% [16, 17, 19, 21, 44, 46–49]. Peroneal nerve palsy has an incidence in the literature of 6–12% [17, 43, 44].

20.8 Conclusion

Knee arthrodesis is a rarely used salvage procedure for failed infected total knee arthroplasty that has significant functional limitations for the patient. The benefits are that it may allow the patient to continue to ambulate independently on a pain-free, stable extremity, which is the main advantage in comparison to transfemoral amputation. Good results have been achieved with the use of IMN, both long and coupled devices, as well as with circular external fixators, and also when using a combined technique. The surgeon must choose whether to proceed with a one- or two-stage procedure and utilise the best surgical technique based on their expertise and a thorough assessment of the features of each individual patient.

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21.1 Introduction

Periprosthetic joint infection (PJI) is an important cause of failure after total knee replacement. Treatment of PJI aims to eradicate the infection, improve joint motion, improve patients' satisfaction and independence during the activity of daily living, and avoid medical and surgical complications. Treatment options include long-term suppressive antibiotics (in patients who are not suitable for surgery), debridement, antibiotics and implant retention (DAIR), one- or two-stage revision, resection arthroplasty, arthrodesis and amputation. However, there is not a univocal definition of success or failure after treatment of PJI [1, 2]. Volin et al. [3] define the success after two-stage revision as the absence of disease at the latest follow-up. Bradbury et al. [4] consider successful not only the clinical resolution of infection and lack of further surgery but also the clinical resolution of infection under suppressive oral antibiotics. Treatment response is defined as post-debridement period free from periprosthetic

joint infections relapse during the time of follow-up by Waagsbo et al. [5]. The absence of symptoms and signs of infection until the date of the last follow-up is the definition of success for Azzam et al. [2]. Success is considered as infection control with serum inflammatory markers (ESR and CRP) normalized and no clinical signs or symptoms of infection by Estes et al. [6]. Parvizi et al. [7] consider success the eradication of infection. Remission is defined by the absence of local or systemic signs of infection assessed during the most recent contact with the patient and lack of the need to reoperation or to administer antibiotic therapy directed to the first infected site from the end of treatment to the most recent contact by Senneville et al. [8]. The Delphi-based International Multidisciplinary Consensus definition of a successfully treated periprosthetic joint infections is infection eradication, no subsequent surgical intervention, no mortality related to periprosthetic joint infection. The Delphi-based International Multidisciplinary Consensus agrees on the definition of midterm follow-up defining the time of 5 or more years after the definitive surgery, and of long-term results determining the time of 10 or more years after surgery [9–11]. Summarizing, the outcomes reported are: infection control without antibiotic therapy; infection control with antibiotic treatment; aseptic revision longer than 1 year from initiation treatment; septic revision longer than 1 year from initiation of treatment; aseptic revision inferior or equal to

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1 year from initiation of treatment; septic revision inferior or equal to 1 year from initiation of treatment; amputation, resection arthroplasty, or fusion; retained spacer; death inferior or equal to 1 year from initiation of treatment; death longer than 1 year from initiation of treatment.

21.2 Debridement, Antibiotics and Implant Retention (DAIR)

DAIR consists of debridement, antibiotics and implant retention. Open DAIR is considered a less disruptive intervention that seeks to preserve a functional implant minimizing the significant morbidity of implant removal. DAIR approach is indicated in early postoperative periprosthetic joint infections and acute hematogenous periprosthetic joint infections, defined as symptoms existing for no longer than 4 weeks.

The infection control rate of DAIR ranges from 11.1% to 100% [12–14]. An infection control rate of 32.6% is reported in a review of 23 studies with 530 infected total knee arthroplasties using an open DAIR approach [15, 16]. The overall success rate after DAIR is reported to be 47% in a review of 28 studies involving 599 cases [13]. Infection control after DAIR is influenced by several factors, such as patient's age, type of infection, involved joint, and duration of antibiotic therapy [17]. Several studies report no differences in infection control rate comparing DAIR performed during the 3 weeks following the onset of symptoms or later [18, 19]. However, on the other side, other authors report that a longer duration between onset of symptoms and DAIR is associated with lower infection control [17, 20]. Several studies demonstrate a high treatment success rate when DAIR is performed within 1 week after the onset of symptoms [21–23].

Recommendations suggest that the DAIR approach is indicated in early postoperative periprosthetic joint infections and acute hematogenous periprosthetic joint infections. Chronic periprosthetic joint infections should be considered an absolute contraindication to perform a DAIR procedure [24]. Other contraindications are severe and

extensive infections, long duration of symptoms, any possibility to exchange the modular components, hard to eradicate causative microorganism [25, 26]. DAIR success is reported to be 73.9% in patients who undergo modular component exchange, compared to 60.7% in patients who don't do modular component exchange [25]. The great success obtained with modular component exchange is confirmed by several authors [21, 27]. Other factors associated with a poor outcome are rheumatoid arthritis, old age, male sex, chronic renal failure, liver cirrhosis, chronic obstructive pulmonary disease, fractures, especially in early acute periprosthetic joint infections, revision arthroplasty; high C-reactive protein, high bacterial inoculums, infections caused by *S. aureus* and Enterococci [28]. Patients with rheumatoid arthritis have a failure rate of DAIR of 74% in the case of late acute periprosthetic joint infections, versus 43% in patients without rheumatoid arthritis. Patients older than 80 years old with late acute periprosthetic joint infections have a significantly higher risk of failure of DAIR. Failure rates following DAIR for acute periprosthetic joint infections range from 20% to 70%, with higher failure seen in acute hematogenous periprosthetic joint infections. The failure rate at 2-year follow-up following DAIR for acute hematogenous infections is 52% [29]. Other authors report a success rate of 82.1% of DAIR for early postoperative infections and 57.1% for acute hematogenous infections [30]. On the other side, Bryan et al. report no significantly different outcomes between early postoperative infection versus acute hematogenous infection [30]. Factors associated with DAIR failure are the presence of a sinus, impaired immune response, short antibiotic therapy, and delayed DAIR [31].

In conclusion, the DAIR procedure has shown diminished morbidity and superior functional outcomes compared to one- or two-stage revision, reducing bone loss and soft tissue trauma. Despite its advantages, the outcome of DAIR must be balanced against the reduced eradication rates compared to more invasive revision surgery. Nevertheless, even if a failed previous DAIR could tend towards more formal one- or two-stage revision because of the possible increased physical or psychological impairment for the

patient with delay in definitive treatment, it does not appear to negatively impact eventual infection eradication [32]. Due to the lack of conclusive evidence, further large-scale prospective study or randomized controlled trials are required.

21.3 One-Stage Exchange Arthroplasty

One-stage exchange arthroplasty aims to decrease surgical morbidity and mortality of patients with periprosthetic joint infections, decrease economic costs, and increase patients' quality of life. One-stage revision procedures have a success rate between 75% and 95% [15, 33–45].

Good outcomes are observed in patients with strict selection criteria. A reinfection rate of 0% is documented in a series of 28 patients with infected knee arthroplasties at a minimum of 3 years follow-up [41]. Preoperative bacterial identification is required to delineate antibiotic therapy. Excellent results are reported in patients in which microbiological susceptibility is preoperatively known [41]. However, the lack of preoperative microbiological diagnosis is considered by several authors a relative, rather than absolute, contraindication for one-stage exchange arthroplasty [44, 46, 47].

Contraindications to one-stage exchange arthroplasty are failure of prior one-stage revision, unclear causative pathogen, lack of susceptibility to available antibiotics, extensive infections, systemic sepsis, gross tissue inflammation, and severe immunosuppression.

Infections caused by polymicrobial organisms, atypical and gram-negative organisms, methicillin-resistant *S. aureus* (MRSA), and methicillin-resistant *Staphylococcus epidermidis* (MRSE) are associated with a higher failure rate (44%) [48–50].

Confounding results are obtained by analyzing the impact of soft tissue defects and sinus tracts on outcomes. Jenny et al. [51] report a negative effect, with a reinfection rate of 27%. On the other side, in an earlier series of 47 patients, is documented an 87% infection-free survival period at 3 years even though 43% of patients presented a sinus tract [52]. For Raut et al., a fis-

tula is not an absolute contraindication to one-stage exchange arthroplasty [53].

Soft tissue debridement, removal of foreign material, and the use of antibiotic-loaded cement for reimplantation are recommended for success.

Two recent meta-analyses show equivalent reinfection (8.2%) comparing one-stage versus two-stage procedures for periprosthetic total knee infections [54]. Wolf et al. underline the superiority of a two-stage protocol in term of infection recurrence; on the other side, the same authors show the superiority of one-stage protocol in terms of quality of life [55]. One-stage mortality range from 4.4% to 11.4% [39, 55]. Loty et al. [56] report mortality of 4.4% analyzing 90 patients with a mean follow-up of 47 months. Miley et al. [57] found a mortality of 11% analyzing 100 patients with a mean follow-up of 48.5 months. Wolf et al. [55] after a Markov cohort simulation decision analysis report a mortality rate of 0.52% (3 of 576) for single-stage and 2.5% (8 of 321) for two-stage revision, based on 18 published papers. Haddad et al. have conducted a proper study on the analysis of functional outcomes after a one-stage revision procedure, considering patient's preoperative functions, evaluated by the Knee Society Score (KSS), and their postoperative status. This research has shown a statistically significant difference in the improvement of functional scores, supporting the one-stage procedure. The mean increase in KSS scores was +56 for one-stage and +45 for two-stage, which takes into account a patient's anatomical stability postoperatively, but also their self-reported functional status and pain levels [58]. These findings are, however, limited: future studies are necessary to delineate the superiority of a one- or two-stage revision approach.

21.4 Two-Stage Exchange Arthroplasty

Two-stage revision is the most used procedure for prosthetic joint infection treatment. It consists of removing all foreign material from the joint, making an extensive debridement of periarticular tissues and inserting a static or articulating spacer

in the joint. This surgical procedure is followed by antibiotic therapy for an extended period. Finally, reimplantation is made when the infection is eradicated.

The two-stage revision procedure is mostly used for patients with an unclear causative pathogen, bacteria unsusceptible to available antibiotics, patients with signs of systemic sepsis, and patients with extensive comorbidities.

Two-stage revision is traditionally considered the gold standard for the management of periprosthetic joint infections. However, compared to the one-stage revision, it exposes patients to the risks of an additional surgical procedure. The success of treatment with two-stage revision arthroplasty is between 70% and 100%. The reinfection rate after two-stage revision is between 9% and 20% of cases [54]. Citak et al. [59] reported superior functional outcomes with the use of articulating spacers when compared to static spacers. Articulating spacers are associated with a little time of hospitalization and improved range of motion. Kim et al. have shown the optimal functional outcomes of 20 patients treated by two-stage revision arthroplasty using an articulating spacer under the diagnosis of infected TKA, considering a follow-up period of about 22 months. The results obtained are described below. ROM increasing from 69.8° (range, 50° to 100°) before first stage surgery to 102.8° (range, 80° to 130°) following second stage surgery. KSKS (Knee Society knee score) increases from 33.8 points (range, 28 to 52 points) before first stage surgery to 85.3 points (range, 77 to 94 points) following second stage surgery. The mean KSFS (Knee Society function score) increases from 35.0 points (range, 20 to 55 points) before first stage surgery to 87.5 points (range, 70 to 100 points) following second stage surgery. Partial weight-bearing granted at 6 days after first stage surgery. There is no sign of infection recurrence in more than 90% of patients. No complications (e.g., medial collateral ligament tears and periprosthetic fractures) observed. These data confirm the advantage of the articulating spacer in terms of infection eradication and recovery of joint mobility and function [60]. An overall 15% complications rate, including reinfection rate, lower joint mobility, painful symptoms, [61], and

a 9.1% fractures rate are reported for static knee spacers [62]. No significant differences in terms of infection eradication and complications between articulating and non-articulating spacers for periprosthetic knee are found [63, 64]. Two-stage revision mortality ranges from 2.9% to 25.7% [65–70]. Chen et al. [65] report a mortality of 8.7% analyzing 57 patients with a mean follow-up of 67.2 months. Haddad et al. [66] report a mortality of 4.0% analyzing 50 patients with a mean follow-up of 5.8 years. Hsieh et al. [67], evaluating 99 patients at mean 43 months of follow-up, report attributable mortality of 3.0%. Romanò et al. [68] record mortality of 2.9% analyzing 102 patients with a mean follow-up of 48 months. Toulson et al. [69] report mortality of 25.7% analyzing 132 patients with a mean follow-up of 64.8 months. Finally, Ibrahim et al. report a mortality of 15.2% analyzing 125 patients with a mean follow-up of 5.8 years [70]. Currently, an interesting summarization about two-stage TKA revision complications rate and functional outcomes has been realized by Claassen et al. They reviewed study patient's charts including demographics, prior surgeries, comorbidities, incidence of persistent infection, and revisions. At the final follow-up examination, they evaluated patient's satisfaction, pain level, and disorders. A successful clinical outcome was defined as a functioning prosthesis without wound healing disorders, no sinus tracts, or other clinical evidence of a persistent infection, which resulted in about 86% of patients. Reimplantation of prosthesis was performed in 95% of patients; only three patients received a septic arthrodesis. Two-stage reimplantation has resulted in a success rate of 76.0%. Only one patient needed to be treated with knee amputation [71]. Due to the lack of conclusive evidence, further large-scale prospective study or randomized controlled trials are required.

21.5 Conclusion

There is not a univocal definition of success or failure after treatment of PJIs. Treatment of PJI aims to eradicate the infection, improve patients' satisfaction, and avoid medical and surgical com-

plications. The successful treatment of PJI depends largely on multiple factors, including the causing microorganisms, soft tissue and bone stock, host factors, prior treatments, and chronicity of infection. Treatment options for PJI include long-term suppressive antibiotics (in patients who are not suitable for surgery), DAIR, one- or two-stage revision, resection arthroplasty, arthrodesis and amputation. DAIR approach is indicated in early postoperative PJI and acute hematogenous PJI. Chronic PJI should be considered an absolute contraindication to perform a DAIR procedure. One-stage exchange arthroplasty has good outcomes in patients with strict selection criteria. Contraindications to one-stage exchange arthroplasty are failure of prior one-stage revision, unclear causative pathogen, lack of susceptibility to available antibiotics, extensive infections, systemic sepsis, gross tissue inflammation, and severe immunosuppression. Two-stage revision is the most used procedure for prosthetic joint infection treatment. The reinfection rate after two-stage revision is between 9% and 20% of cases. Undoubtedly, in the setting of chronic PJI, two-stage exchange arthroplasty is a safe and efficacious treatment. As widely already discussed, an absolute gold standard for the choice of PJI treatment doesn't still exist. However, we can assume that in carefully selected cases the DAIR protocol allows a non-invasive treatment of the patient with a low morbidity rate, even if the one-stage revision guarantees recovery times and superior functionality. The two-stage treatment remains the best procedure in terms of targeted and definitive eradication of the infection, despite elevated morbidity rates persist. However, the choice of the most suitable treatment must take into account the all patient's characteristics in their complexity.

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Complications

22

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22.1 Introduction

A prosthetic joint infection (PJI) following primary total knee arthroplasty (TKA) is a devastating complication for both the patient and the surgeon. Fortunately, PJI is relatively rare, with pooled international registry data suggesting a 1.03% risk following primary TKA [1]. The demand for TKA is projected to rise by 400% from the early 2000s to 2030 due to an aging and increasingly active population [2, 3]. This will inevitably result in an increased number of prosthetic joint infections and subsequent revision arthroplasties, which have higher complication rates and worse functional outcomes. Revision arthroplasty for PJI has a significantly higher risk of mortality compared to revision surgery for aseptic failure; there is evidence suggesting that the risk of 1-year mortality for PJI is comparable to a number of common cancers [4]. Considerable

efforts and resources have been expended in attempting to decrease the burden of PJI. Treatment requires a multidisciplinary approach with specialized microbiologists, physiotherapists, and revision arthroplasty specialists. There is increasing evidence that treatment of infected TKAs at specialized arthroplasty centers with high volume surgeons leads to improved outcomes [5–7].

Despite the improvements in preventing PJI, a proportion of patients will unfortunately, still develop a deep infection. Management of these patients ultimately involves both antibiotics and surgical intervention. Appropriate surgical intervention may consist of a DAIR (debridement, antibiotics, implant retention) procedure, single-stage revision, or two-stage revision depending on the organism and host factors. Revision TKA in the setting of infection is technically challenging and has a higher risk of complications compared to aseptic revisions [8].

Complications of infected total knee arthroplasty can arise during the procedure (intraoperative), early in the postoperative recovery (early), and later once recovery from the initial procedure is complete (late). It is imperative that surgeons recognize and appreciate the potential complications associated with revision TKA in the setting of infection. This will subsequently lead to a decreased risk of complications and an improved understanding of how to appropriately manage specific complications when they do occur.

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22.2 Intraoperative Complications

22.2.1 Surgical Exposure

Adequate surgical exposure and careful handling of the soft tissue envelope are critical for any revision arthroplasty procedure. Previous incisions should be employed in order to minimize wound complications and breakdown. In situations with multiple incisions, the most lateral incision should be used to preserve blood supply to the skin, ensuring that this allows access to the joint without creating large skin flaps. Sinus tracts should be excised to prevent ongoing and recurrent infection (Fig. 22.1), and this must be considered when planning incisions. Incisions should be of sufficient length to prevent any excess tension on the skin edges. Full-thickness fasciocutaneous flaps should be created to maintain vascular integrity. Any necrotic or infected subcutaneous tissue should be debrided to bleeding, healthy tissue.

Patients are often stiff with a thickened and inflamed soft tissue envelope. In the setting of infection, thorough and systematic debridement and complete synovectomy are necessary to eradicate infection and adequate exposure of the previous implants. Often after thorough debridement, adequate exposure of the implants is possible. However, extensile approaches may be required to prevent iatrogenic injury to the extensor mechanism. Surgeons should be prepared to perform a



Fig. 22.1 Sinus tract developed as a result of prosthetic joint infection

quadriceps snip, VY turndown, or tibial tubercle osteotomy if exposure is not adequate for implant removal or subsequent reconstruction.

22.2.2 Extensor Mechanism Rupture

Protection of the extensor mechanism is imperative when performing revision TKA for prosthetic joint infection; extensor mechanism injury or failure significantly worsens functional outcomes even with successful eradication of infection. The extensor mechanism can be disrupted as a result of infection or iatrogenic injury. This is a devastating complication that makes the restoration of knee function much more difficult. A study involving 60 patients with concomitant periprosthetic joint infection and extensor mechanism disruption has shown low success rates, regardless of treatment. Of these, 53 patients underwent extensor mechanism repair or replacement, with 41 of these considered failures, recurrent infection being the most common cause of failure [9].

Treatment of an extensor mechanism disruption typically involves a 2-stage revision with a subsequent extensor mechanism allograft or mesh reconstruction. Articulating spacers are preferred in order to allow for movement in the affected knee and facilitate recovery of knee function [10]. However, significant damage to the extensor mechanism or surrounding soft tissue envelope may necessitate the use of a static spacer [11, 12]. Spacers should be monitored for any impingement or translation of the extensor mechanism, and any problems should be appropriately addressed to prevent further complications. Extensor mechanism allograft reconstruction has shown good survivorship. A single-center study showed that 69% of knees retained their initial allograft at final follow-up. However, patients had consistently worse functional outcomes and high reoperation rates [13]. At the time of definitive revision TKA, it is crucial to ensure that the femoral and tibial components are well-positioned to optimize patellar tracking. Failure to do so may lead to early failure.

22.2.3 Increased Operative Time

Debridement, antibiotics, and implant retention (DAIR), one-stage, or two-stage revision procedures are the mainstay of surgical treatment options for periprosthetic knee joint infections. These operations are technically challenging and generally require increased operative time compared to primary TKA. Prolonged operating times have been shown to increase the risk of a surgical site infection, which in turn can lead to disastrous consequences for an already infected joint. It has been suggested that operative length over two and a half hours increases the risk of infection [14, 15]. As revision TKA procedures can often be time-consuming, these cases are subsequently at greater risk of infection compared to primary TKA. A nationwide study involving over 10,000 revision TKAs concluded that operative times had the greatest effect on length of stay in hospital postoperatively compared to other variables, including age, sex, and BMI [16]. There is growing evidence that treatment of prosthetic joint infections should be managed in specialized centers by experienced, high volume revision surgeons in order to minimize operative time and decrease complication rates [3, 5].

22.2.4 Intraoperative Fracture

An intraoperative fracture is a potential risk during revision knee arthroplasty. A North American study reported an intraoperative peri-prosthetic fracture incidence of 0.78% in 645 revision total knee replacements [17]. Surgeons should be conscious of the risk of fracture during explantation of previous components, preparation of the medullary canal, and insertion of stemmed revision components [18]. This risk of peri-prosthetic fracture during revision arthroplasty increases in the setting of infection due to poor bone quality and loss of bone stock. A study that analyzed 894 infected TKA patients treated with 2-stage revision demonstrated an intraoperative fracture rate of 2.3% [19]. Of these, 17% of fractures occurred during removal of components and 82% during

reimplantation. 56% of fractures were femoral fractures, 30% were tibial fractures, and 13% were patella fractures.

Fractures identified intraoperatively should be anatomically reduced and stabilized appropriately. Stemmed components should bypass the fracture site by a minimum of two cortical diameters. Fractures of the femoral and tibial condyles often require screw or plate fixation. The fracture should be sufficiently stable to allow an immediate range of motion postoperatively. However, weight-bearing may need to be delayed, which can further hinder the rehabilitation process [20]. An intraoperative fracture also significantly increases operative time; this may further increase the risk of complications, including reinfection.

22.2.5 Instability

The stability of the joint is at risk following a prosthetic knee infection. Surgeons must ensure that the knee is stable intraoperatively; it is often necessary to use revision components with an increased varus-valgus constraint or a rotating hinge design. The stability of the knee joint depends on intact collateral ligaments and balanced load distribution over the medial and lateral condyles. In order to successfully eradicate infection, aggressive debridement of the joint and surrounding soft tissue envelope is required. This may include important ligamentous and capsular structures, which subsequently would compromise stability. A semi-constrained or hinged implant is necessary after resection of the collateral ligaments following debridement [21].

It has been reported that the incidence of instability following revision knee surgery is 22% [22]. It is likely that the incidence of instability following revision TKA for PJI is significantly higher given the degree of necessary debridement. Instability can be divided into coronal and sagittal plane instability. Instability in extension results from an insufficient filling of the extension gap and can arise due to inadequate release of fixed deformity. A flexion instability comes as a result of a flexion gap greater than an extension gap. A femoral component that is

undersized or a steep tibial slope can give rise to this form of instability [23].

Consequently, during revision surgery for PJI, surgeons should endeavor to extract components with minimal bone loss, balance the knee ligaments, use stable components that restore the joint line, and also achieve good soft tissue coverage. Varying landmarks have been described in the literature to assist in the precise reproduction of the joint line in revision TKA procedures: 1.5–2 cm proximal to the fibular head, the “meniscal scar,” 2–2.5 cm distal to the lateral femoral epicondyle, 2.5–3 cm distal to the medial femoral epicondyle, 2 cm proximal to the tibial tubercle, and 2 cm below the inferior pole of the patella in extension [24–26]. Radiographs of the contralateral knee and then measuring the size of the resected prosthesis is often helpful. Careful preoperative planning and anticipation of potential instability are essential when undertaking revision TKA for infection.

22.2.6 Neurovascular Injury

The neurovascular structures surrounding the knee joint are at increased risk of iatrogenic injury during a revision procedure. The common peroneal nerve is the most commonly injured nerve during revision TKA procedures. While direct nerve injury is rare, compression, traction, and ischemia are the most common mechanisms of injury [27]. The incidence of common peroneal injury ranges from 0.58% to 1.8% in primary TKA [28]. Risk is increased in a preoperative valgus knee, fixed flexion of $>20^\circ$, previous laminectomy, and the use of a spinal anesthetic [29]. A prolonged tourniquet time is also associated with an increased risk of neural damage. This is especially important in revision procedures, which can often be more technically challenging and therefore longer operations. A retrospective study reported a 7.7% incidence of neural injury (common peroneal or tibial nerve) involving 1001 patients who underwent primary or revision

TKA with a tourniquet time of greater than 120 min [30].

Vascular injury, albeit less common than neural injury, can result in disastrous complications. Furthermore, the incidence of arterial damage in the revision TKA setting is greater than twice that of in primary TKAs (0.36% vs. 0.15%) [31]. A greater exposure is required during revision TKA for infection to enable adequate debridement of infected tissue, including the often thickened posterior capsule. The neurovascular bundle can be adherent to the posterior capsule, which increases the risk of injury during debridement. Vascular injury can be direct, for example, from a sharp object such as a scalpel or drill, or indirect, such as from the tourniquet or traction. Direct injury of the artery can result in a massive hemorrhage, which may be visualized intraoperatively or manifest as a significant drop in blood pressure. Surgeons should seek immediate vascular consultation for surgical treatment, a bypass graft, or direct repair of the artery. Indirect arterial injury, leading to ischemia, can easily be missed in the immediate postoperative phase due to a lack of bleeding and the masking of pain by analgesic agents commonly used postoperatively. These patients are at an increased risk of compartment syndrome, leading to fasciotomy, neural injury, and muscle necrosis [31]. Doppler ultrasound and an ankle-brachial pressure index (ABPI) should be immediately considered in patients suspected of having indirect arterial injury following revision TKA. An immediate vascular opinion and possible vascular studies should be requested if there is any clinical suspicion.

22.3 Early Complications

22.3.1 Venous Thromboembolism

Venous thromboembolism (VTE) is a potential early complication following revision TKA for PJI. Preoperative risk assessment and appropriate

use of both mechanical and pharmacological prophylaxis are of the utmost importance when treating these patients. The prevalence of proximal deep vein thrombosis (DVT) and distal DVT following primary TKA has been reported between 0% and 16% and 1% and 67%, respectively [32]. Moreover, the prevalence of symptomatic pulmonary embolism (PE) ranges from 1% to 1.9%, with fatal PE being reported as between 0.2% and 0.7% [32].

There is limited evidence on the prevalence of VTE in revision TKAs when compared to primary TKAs. A large multi-center study in Illinois involving 2986 revision TKA patients showed a reported DVT rate of 1.4% and PE rate of 1.6% [33]. Another study involving 645 revision TKAs reported rates of DVT and PE of 0.16% and 2.02%, respectively [17]. Careful soft tissue handling and minimization of excessive knee hyperflexion has been suggested to reduce venous stasis and ultimately decrease the risk of VTE [34].

Nonpharmacological prophylaxis includes using compressive stockings, intermittent pneumatic devices, and early mobilization with ankle exercises. Additionally, epidural anesthetics have a lower associated risk of DVT compared to a general anesthetic (4% and 9%, respectively) [35]. There is an ongoing debate as to the most effective form of pharmacological VTE prophylaxis. Low molecular weight heparin is thought to be more effective than warfarin in the prevention of symptomatic thrombosis [32, 36]. However, there is an increased risk of minor bleeding found with pharmacological prophylaxis [37], which can complicate revision surgery. The combined use of pharmacological and nonpharmacological VTE prophylaxis has been found to reduce the rate of VTE even further post TKA [38]. Surgeons should, therefore, consider a combined pharmacological and nonpharmacological approach to VTE prophylaxis for revision knee surgery for an infected joint, with careful, regular monitoring to assess for bleeding.

22.3.2 Hematoma and Wound Complications

Hematoma can develop postoperatively, resulting from increased bleeding from thorough debridement and pharmacological VTE prophylaxis. Hematoma can lead to persistent draining, wound complications, and dehiscence; this can increase the risk of recurrent infection [39]. A retrospective review study involving 17,784 patients who underwent TKA showed that the patients who underwent an evacuation of hematoma within 30 days of the procedure had a 5-year risk of deep infection of 6% compared to 0.8% for patients who did not have a reoperation [40]. Due to the extensive debridement of soft tissue in PJI revision cases, there is an increased risk of persistent venous bleeding. The use of drains, while controversial, may help decrease hematoma formation postoperatively.

Furthermore, the use of negative pressure wound therapy has also been shown to be beneficial in treating wounds treated for infection after TKA [41, 42]. Wound dehiscence is a devastating complication that can act as a gateway for microbes to enter the joint and increase the risk of recurrent infection. This is especially true in patients with comorbidities such as diabetes mellitus, obesity, hypertension, arteriosclerosis, neuropathy, and smoking. Therefore, it is crucial to ensure that medical treatments of chronic conditions are optimized to promote wound healing [43]. In the presence of wound dehiscence, plastic surgery consultation, and soft tissue coverage with a rotational or free muscle flap may be necessary for wound closure.

22.3.3 Recurrent Infection

Recurrent infection is another devastating early complication associated with the treatment of a PJI. Both surgical and patient factors can contribute to the risk of persistent or recurrent infection. A multidisciplinary approach

ensuring that patients have adequate surgical debridement, optimization of medical comorbidities, and appropriate antibiotics is essential in preventing recurrence. Despite this, a proportion of patients will still develop an ongoing infection. DAIR has a variable success rate, from 18% in treating PJI with MRSA to 100% in another arthroscopic study [44, 45]. A literature review on the efficacy of DAIR concluded that it could be an effective method of eradicating PJI. It is recommended to be carried out in the acute postoperative period, within 4 weeks of surgery, and that the procedure should be done in an open fashion rather than arthroscopically [46]. A systematic review article comparing the results of 687 patients who underwent a single-stage exchange arthroplasty to 1086 patients who underwent a two-stage exchange arthroplasty for chronic PJI showed that the eradication rate was 87.1% and 84.8%, respectively [47].

A retrospective study examining 548 patients that received a two-stage exchange arthroplasty for an infected hip or knee prosthesis showed that female gender, psychiatric illness, and heart disease all increased the likelihood of recurrent infection [48]. Moreover, an Italian study concluded that continuing with antibiotic therapy between the two stages of exchange arthroplasty, without an antibiotic-free “holiday period” reduced the prospect of a recurrence of PJI and provided better outcomes in immunocompromised patients [49].

22.4 Late Complications

22.4.1 Stiffness

Stiffness is a common complication after an infected knee PJI and revision surgery. A study conducted looking at the range of motion post revision TKA surgery showed that 4% of patients presented with stiffness, which was defined by a range of motion of $<90^\circ$ at the 3-month postoperative follow-up appointment [50]. A further German study which involved 867 primary TKAs and 176 revision TKAs found that 4.54% of pri-

mary TKAs and 5.11% of revision TKAs were deemed stiff ($<90^\circ$ flexion) [51].

Predicting stiffness after revision surgery for an infected implant is challenging. However, preoperative range of motion has been shown to be the greatest determinant [43, 44]. A shorter duration between primary and revision TKA has also been shown to increase the risk of recurrent stiffness [50]. A high BMI is a modifiable risk factor linked to postoperative stiffness in revision TKAs [52].

Conservative approaches, such as physical therapy, can be used initially. Early manipulation under anesthetic, within 3 months of the surgery, has proven successful in improving stiffness [53]. More invasive procedures such as open arthrolysis and revision surgery should be considered in more resistant cases [53].

22.4.2 Peri-Prosthetic Fracture

Peri-prosthetic fractures can occur both intraoperatively and postoperatively secondary to trauma. They are particularly difficult to treat in the setting of a periprosthetic joint infection. Goals of treatment include restoration of length, alignment, and rotation, stable fixation and/or reconstruction, and eradication of infection [54].

Peri-prosthetic distal femoral fractures around the femoral component of a TKA are extremely challenging to deal with. The Su classification divides fractures into 3 types (I–III) according to the location relative to the proximal aspect of the femoral component. Type I fractures are proximal to the femoral component, type II originates at the proximal aspect of the femoral component and extends proximally, and type III extends distal to the proximal border of the femoral component [55]. The Lewis and Rorabeck classification divides fracture into 3 types (I–III). Type I describes an undisplaced fracture with the intact prosthesis, type II is a displaced fracture with an intact prosthesis, and type III is a displaced or undisplaced fracture with loosening of the femoral component [56].

Tibial PPFs can be classified using the Felix classification [57]. This system divides fractures

around the tibial component into 4 types (I–IV). Type I represents a fracture of the tibial plateau, type II fractures are adjacent to the prosthetic stem, type III fractures are of the tibial shaft, distal to the tibial component and type IV fractures represent a fracture of the tibial tubercle. Additionally, peri-prosthetic fractures can be classified using the Unified classification system, which recognizes 6 types (A–F). Type A is an apophyseal fracture, B represents fractures at the bed of the implant, C are fractures clear of the implant, D is a fracture in between two prostheses, E is a fracture of each of two bones supporting one joint replacement, and F is a fracture articulating or facing an implant [58].

PPFs around a revision knee prosthesis can occur early or late postoperatively. The risk factors include poor bone stock, which is often seen in infected cases, multiple re-revisions, osteopenia, certain comorbidities, such as inflammatory arthritis and anterior cortical stress risers [59]. The risk of a PPF around the implant post-revision TKA is twice that of the risk post primary TKA [60, 61]. The Scottish registry showed that the incidence of a PPF was 0.6% and 1.7% in primary and revision TKAs, respectively [60]. The review also found that the only risks that significantly increased the likelihood of PPFs around a TKA were female gender, age over 70, and revision surgery.

The treatment of fractures around a revision component is much more challenging as orthodox treatment options for PPFs around a primary implant such as an intramedullary nail or a peri-articular locking plate are often unsuitable. Unstable fractures with a stable prosthesis require open reduction and internal fixation (ORIF). Displaced fractures with a loose component should be revised with a longer stem that bypasses the fracture site [57]. In the case of non-union due to comminution, bone loss or failure of ORIF around a PPF of the distal femur, a distal femoral replacement can be considered as a limb salvaging procedure [62].

Patella fractures post TKA are less common and can generally be managed non-operatively in the absence of extensor mechanism disruption and patella component instability [63]. If patella

component instability is present, treatment should be based on the bone stock available. If there is adequate bone stock, ORIF with or without component revision should be attempted; however, if bone stock is poor, partial or complete patellectomy may be an alternative [64].

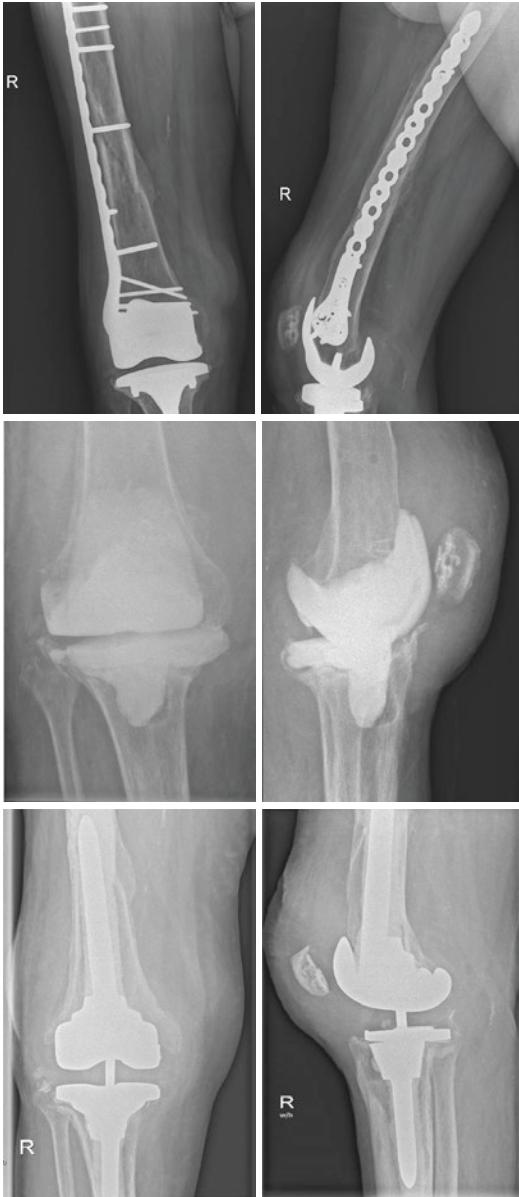
22.4.3 Stem Tip Pain

Stem tip pain is a common cause of discomfort expressed by patients following revision surgery. The poor bone stock that frequently coincides with PJI makes maintaining accurate component alignment difficult. A tibial stem extension is often employed to improve component fixation [65]. Pain in the region of the stem tip has been described in 14% of patients who had undergone revision TKA [66]. Also, stem design does not seem to influence the incidence of stem tip pain post revision [67]. Placement of a tibial plate in the region of the stem tip has been found to reduce the incidence of pain [48]; however, the use of additional metalwork in the treatment of PJI can increase risks further. For this reason, a fixation strategy that gains purchase in the metaphyseal regions of femur or tibia in conjunction with short, cemented stems may be a more successful strategy [68].

22.5 Conclusion

In conclusion, the mainstay of treatment for an infected knee prosthesis involves the eradication of the infection and the provision of a pain-free, stable and functioning knee joint that allows sufficient range of motion. The process of achieving this is often difficult and can seem interminable due to complexities that surround a PJI. Treatment must be tailored to patients on an individual and carefully planned basis in a specialized arthroplasty center where the input from a revision knee specialist, microbiologist, and specialist physiotherapists are available in order to achieve a satisfactory outcome and decrease the risks of further devastating complications [5].

Case Presentation



This 65-year-old patient sustained a distal femur peri-prosthetic fracture that was managed with an open reduction and internal fixation with a lateral locking plate. He subsequently developed a prosthetic joint infection and underwent a two-stage revision with an articulating spacer followed by a constrained revision prosthesis.

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Part VI

Current Evidence on Prevention of Knee Replacement Infections



Treatment of Periprosthetic Joint Infections with Resistant Organisms

23

Kevin A. Sonn and R. Michael Meneghini

23.1 Introduction

The successful eradication of periprosthetic joint infection (PJI) depends on various host factors, treatment modalities, and infection characteristics. Infections caused by antibiotic-resistant organisms have been increasing in recent years [1, 2]. Studies have clearly demonstrated the difficulty of treating PJI caused by organisms including methicillin-resistant *Staphylococcus epidermidis* (MRSE), methicillin-resistant *Staphylococcus aureus* (MRSA), and *enterococcus* [1, 3–6]. It is vital to understand the treatment ramifications of the various resistant organisms when treating PJI.

23.1.1 *Staphylococcus epidermidis*

Staphylococcus epidermidis (*S. epidermidis*) was previously thought of as an innocuous bacterial colonizer on human skin. However, it is now recognized as an opportunistic pathogen that is responsible for the greatest proportion of

infections on all indwelling medical devices [7]. *S. epidermidis* falls into the broader category of coagulase-negative staphylococci which causes 30–43% of all PJIs [8]. *S. epidermidis* first non-specifically binds to implanted prostheses, then subsequent biofilm formation occurs via a polysaccharide intercellular adhesin [9]. It is this ability to develop a strong glycocalyx that accounts for the difficulty of eradication of this low-virulent organism [1]. For these reasons, aggressive treatment of *S. epidermidis* is recommended (especially when methicillin resistance is encountered).

23.1.2 *Staphylococcus aureus*

Staphylococcus aureus (*S. aureus*) is a Gram-positive human commensal organism that has shown persistent nasal colonization in 20–25% of adults and intermittent colonization in up to 60% [10]. *S. aureus* infection causes 10–23% of all PJIs. *S. aureus* interacts with host fibronectin, fibrinogen, and collagen to cover a prosthesis immediately after implantation [8, 11]. A subcutaneous foreign body reduces the minimum infection causing inoculum with *S. aureus* more than 100,000×. The susceptibility to PJI caused by *S. aureus* combined with emerging and worsening resistance has increased recurrent infection rates [1]. Successful infection eradication of PJIs

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caused by methicillin-resistant *S. aureus* (MRSA) with debridement and implant retention (DAIR) is reported as low as 20% and is generally not recommended [12]. Even two-stage revision for MRSA infections has demonstrated low rates of infection eradication, thus highlighting the difficulty in managing this virulent and resistant organism [3].

23.1.3 *Enterococcus*

Enterococcus is a Gram-positive, facultative anaerobe which has been reported to cause 2–3% of all PJIs [4, 13]. El Helou et al. reported 94% success with two-stage exchange for enterococcal infections; however, 46% of their cohort were treated with definitive resection while only 34% underwent two-stage revision. Rasouli et al. achieved successful eradication of enterococcal PJIs in only 20% of cases treated with DAIR and only 44% treated with two-stage revision [4]. An additional challenge treating enterococcal PJIs occurs when the bacteria are resistant. Vancomycin-resistant *enterococcus* (VRE) infections remain exceptionally difficult to treat with reimplantation rather than salvage options such as definitive resection, fusion, or above-knee amputation [4, 14].

23.2 Debridement and Implant Retention

Debridement and implant retention (DAIR) is commonly utilized for the treatment of acute periprosthetic joint infections as discussed in previous sections. The success rates vary widely in the literature and largely depend on the infecting organism. Duque et al. report successful infection eradication with DAIR in only 20% of MRSA and 33.3% of *Pseudomonas aeruginosa* infections compared to 85.3% for all other bacteria [12]. Other authors have reported on similar difficulties and comparable failure rates when treating staphylococcal infections with DAIR

[15–17]. Chung et al. have recently reported on improved success of a planned two-stage DAIR [18]. In their protocol, the first stage consists of a thorough debridement with placement of antibiotic cement beads, while the second stage (occurring 5 days later) involves an additional debridement with exchange of modular components. They report successful infection eradication in 89.6% of TKAs (93.5% in primary TKAs), including overall successful treatment of 70% of MRSA infections [18].

The addition of rifampin to targeted intravenous (IV) antibiotic therapy is recommended for all cases of DAIR, especially those caused by staphylococcal species [19–24]. The successful results of adding rifampin are thought to result from its ability to penetrate biofilm when used in DAIR [8].

23.3 Two-Stage Revision

Two-stage revision remains the gold standard for treatment of chronic periprosthetic joint infection. Overall success rates between 80% and 100% are commonly quoted for infection eradication utilizing a two-stage approach [25–29]. However, when stratifying these results based on type of organism, treatment outcomes worsen with resistant bacteria. Kilgus et al. report 89% success treating methicillin-sensitive *S. aureus* (MSSA) compared to only 18% infection eradication of MRSA and MRSE infections utilizing a two-stage approach [3]. Mittal et al. found 24% reinfection rate when treating MRSA and MRSE in a two-staged fashion [1]. However, 14% were reinfected with new organisms rather than recurrence, therefore they recommend two-stage revision as a viable treatment option in this setting [1]. Rasouli et al. successfully eradicated enterococcal PJIs with two-stage revision in only 7 of 16 patients. Six patients were treated with definitive resection and 3 had either knee fusion or above-knee amputation [4].

Case Example

This is a 61-year-old male with a complex history starting with right total knee arthroplasty, subsequent revision for polyethylene wear, and then complete revision TKA. This was complicated by subsequent hematogenous MRSA PJI which was treated with a single-stage revision. Four years subsequent to that he was found to have an MSSA PJI which was treated with two-stage exchange which was complicated by a traumatic wound dehiscence. At this time he presented to our prac-

tice with a draining sinus and chronic extensor mechanism disruption with revision components in place (Fig. 23.1). He underwent resection and placement of a static antibiotic cement spacer with multiple intraoperative cultures demonstrating enterococcus. This required 2 additional repeat debridements with 1 spacer exchange before the infection was cleared and the knee was reimplanted (Fig. 23.2). At most recent follow-up 2 years after reimplantation, he demonstrated no evidence of infection and was off all antibiotics.

Fig. 23.1 Anteroposterior (AP) and lateral radiographs at presentation demonstrating revision components without evidence of implant loosening

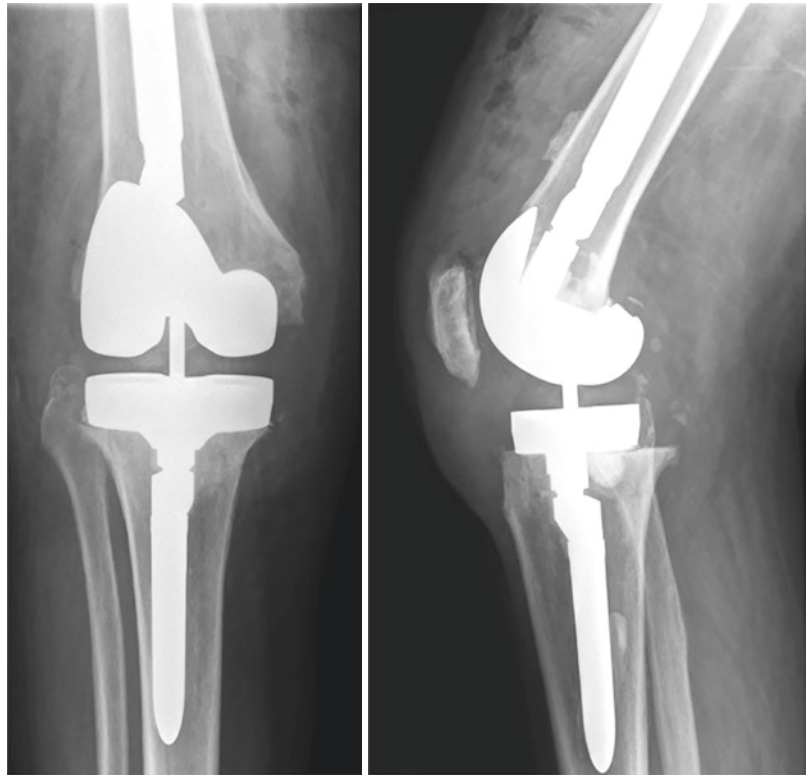
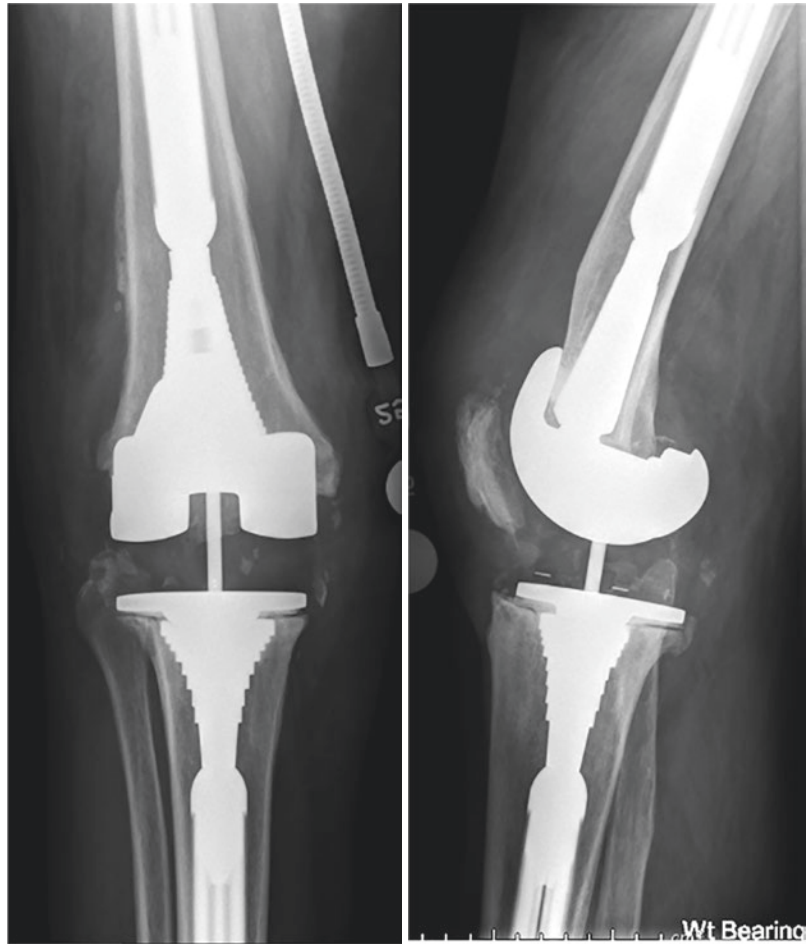


Fig. 23.2 Anteroposterior (AP) and lateral radiographs after reimplantation demonstrating sleeved revision components



23.4 Conclusion

Treatment of knee PJI with resistant organisms remains a challenge with high complication and reinfection rates. Two-stage revision is often the best approach to maximize chances of successful infection eradication. If DAIR is chosen, consideration should be given to performing a planned two-staged DAIR as described by Chung et al. [18]. Further, the addition of 6 months of oral rifampin to targeted IV antibiotic therapy when treating staphylococcal knee PJI with DAIR has demonstrated improved results. Regardless of treatment approach, infection recurrence remains high and salvage procedures such as fusion, definitive resection, and above-knee amputation are realistic outcomes despite best attempts to retain or reimplant prostheses.

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Medical Optimization of the Patient Prior to Surgery

24

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24.1 Introduction

Periprosthetic joint infection (PJI) after total knee arthroplasty (TKA) is a cause of major concern for the health systems and patients and remains the most common cause for early revision in practically every joint registry [1–3]. It is associated with an increased financial burden, inferior clinical outcomes and increased morbidity. Although the incidence of PJI is relatively low at 1.1–2.2% [4, 5], it adds significantly to the healthcare costs, as these patients have increased hospital stays, require readmissions and additional surgical procedures [6, 7]. It is estimated that the number of primary TKAs performed annually is likely to increase by 673% between 2005 and 2030 [8–10]. A similar increase is anticipated in the number of revision TKAs performed for PJI which will see additional financial strain on the already overburdened healthcare systems the world over [11]. Strategies to prevent the occurrence of PJI after TKA have clear benefits.

Many risk factors associated with the PJI after TKA have been identified and can broadly be categorized into preoperative, intraoperative and post-operative factors [12]. The mortality rate of a two-stage revision arthroplasty done for PJI

approaches 25–33% at 5 years [13, 14]. Surgeons should attempt to decrease this risk by managing the modifiable risk factors. In this chapter, we will present a summary of current practice and provide evidence for improved outcomes following the medical optimization of the patient prior to surgery.

24.2 Preoperative Risk Factors

24.2.1 Inflammatory Arthritis

Patients who suffer from inflammatory arthritis such as rheumatoid disease (RD), spondyloarthritis (SpA) including ankylosing spondylitis and psoriasis, and systemic lupus erythematosus (SLE) are at an increased risk of post-operative wound complications and PJI after TKA [15–18]. Many of these patients require chronic management with nonbiologic disease modifying anti-rheumatic medications (DMARDs), glucocorticoids, immunosuppressive medication or biologic agents, some of which puts them at higher risk of acquiring PJI. Consultation with the patients' rheumatologist to alter the medical management of inflammatory arthritis can help diminish wound complications without severely affecting the inflammatory process in other joints.

Rheumatoid disease has been reported as increasing the risk of PJI after TKA [15, 17–19]. The PJI rate in patients with rheumatoid arthritis

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was 1.6 times greater than those with osteoarthritis in a study by Schrama et al. [18] Ravi et al. reported 71,793 patients who had TKA, 4% whom had RD, and found an adjusted hazard ratio of 1.47 ($p = 0.03$) for increased rate of infection with the RD cohort having a rate of 1.26% and the OA group 0.84% [15]. Jämsen et al. [19] reviewed the Finnish Arthroplasty Register with a total of 43,149 primary and revision TKAs, and found an increased risk of PJI in seropositive rheumatoid disease with a hazard ratio of 1.7. This risk for PJI normalized to the same as the primary osteoarthritis cohort after 1-year post procedure.

Patients with psoriasis have been shown to have increased concentrations of bacteria on skin plaques than normal skin [20]. Routine perioperative preparation of these plaques has been found to be successful at sterilization of them with the use of an iodine and alcohol preparation [21]. Reports of PJI in patients with psoriasis are limited and mixed in outcomes for PJI. A deep infection rate of 16.6% and mild skin necrosis in 8.4% out of 24 patients who underwent TKA was reported in one study [22]. Another study of 50 patients undergoing primary TKA only had one (2.0%) deep infection at over 2 years post-surgery, and this patient also had alcoholic cirrhosis which may have contributed to this [23]. Both of these papers had low sample sizes and were retrospective in nature. Menon and Wroblewski [24] reviewed the results of Charnley low-friction hip arthroplasty patients with psoriasis and found a deep infection rate of 5.5% (3 patients); however, one of those patients also had rheumatoid disease.

The evidence for the perioperative management of antirheumatic medication is sparse [25–27]. In 2017, the American College of Rheumatology combined with American Association of Hip and Knee Surgeons to develop evidence-based guidelines on the preoperative management of antirheumatic medications [28]. Each recommendation was graded for strength of evidence with the advice, based on low- to moderate-quality evidence being to continue nonbiologic DMARDs such as methotrexate, sulfasalazine, hydroxychloroquine, leflunomide and

doxycycline through surgery. Biologic agents are recommended to be withheld prior to surgery, with the timing of surgery selected around the dosing regimen for that specific medication. These agents should be restarted once the wound shows evidence of healing, all sutures/staples are removed and there is no wound drainage. Tofacitinib should be withheld for at least 7 days prior to surgery based on non-surgical studies showing an increased incidence of generalized infection. Patients with severe SLE should continue their usual doses of methotrexate, mycophenolate mofetil, azathioprine, cyclosporine or tacrolimus. Patients' with non-severe SLE may have mycophenolate mofetil, azathioprine, cyclosporine or tacrolimus withheld for a week preceding surgery. Stress dosing of glucocorticoids was not recommended, rather the patients treated on glucocorticoids are advised to continue on their usual dose during the perioperative period. The Canadian Rheumatology Association recommendations from 2012 also suggest continuing methotrexate but withholding biologic agents [29].

24.2.1.1 Diabetes Mellitus

Diabetes mellitus is associated with an increased risk of PJI and wound complications in patients undergoing TKA [30–32]. HbA1c levels relate to control of blood glucose levels in the past 1–3 months. Tarabichi et al. showed that HbA1c > 7.7% is associated with increased infection risk of PJI, rather than the often quoted HbA1c of 7% [33]. In a paper by Stryker et al., a HbA1c of >6.7% was associated with an increased risk of wound complications; however, none of the 30 patients in their study developed a PJI [32]. Preoperative hyperglycaemia has also been shown to be associated with increased risk of PJI after TKA [31]. Jämsen et al. showed that PJI risk was more than doubled by the patient having diabetes, but also showed a trend toward a higher rate of PJI if the patient did not have a diagnosis of diabetes but had a preoperative glucose level of ≥ 6.9 mmol/L (124 mg/dL) in contrast to those with <6.9 mmol/L. [31] The infection rate after TKA in this study was 1.59% in the diabetic cohort vs. 0.66% in the non-diabetic group. In contrast to these studies,

Adams et al. [34] demonstrated no increased risk of PJI in patients with diabetes, regardless of HbA1c. Charstil et al. showed in their 2015 study that perioperative hyperglycaemia was associated with an increased risk of PJI, but HbA1c > 7% was not, with the risk increasing from a preoperative glucose level of ≥ 194 g/dL (10.6 mmol/L) [35]. Regardless of the discrepancy in the literature of the risk of PJI associated with HbA1c, it is advisable for diabetic control to be optimized prior to surgery with the assistance of an endocrinologist and diabetes educator.

24.2.1.2 Smoking

Smoking is a well-documented cause of PJI and wound healing issues after TKA in the literature [36–42]. Nicotine causes microvascular constriction and increases the level of carboxyhemoglobin, which further decreases the delivery of oxygen at the tissue level [36, 43]. Thus, nicotine is associated with decreased blood and oxygen supply at the microvascular level increasing the risk of wound healing problems and PJI after TKA. To further compound these issues, collagen synthesis is also impaired by nicotine [36, 43].

Duchman et al. found a higher rate of wound complications in a cohort of smokers at 1.8% vs. former smokers at 1.3% and non-smokers at 1.1% in a retrospective study on 78,191 patients who had undergone primary total hip or total knee arthroplasty [38]. Deep wound infections were present in a statistically greater number in the current smoker group at a rate of 0.7%. The increased incidence of surgical site infection in a smoking cohort undergoing THA or TKA was supported in Singh et al. in a study that had a 2.4% surgical site infection rate vs. 1.6% in life-long non-smokers and 1.7% in prior smokers [41]. Unfortunately this paper did not delineate between deep and superficial surgical site infections. A hazard ratio of 2.37 (95% CI 1.19, 4.72; $p = 0.01$) for developing a deep infection was found in current tobacco users in a separate publication by Singh et al. [37].

Recommendations for a smoke-free period vary from 4 to 8 weeks prior to surgery [39, 44]. There are few publications on the effectiveness of smoking cessation programmes prior to total joint

arthroplasty. Akhavan et al. conducted a study on 30 patients who were eligible for either total hip arthroplasty (THA) or TKA [45]. The patients were instructed that they needed to cease smoking prior to their surgery. They were given referrals for counselling, telephone support programmes and advised to see their general practitioner for nicotine replacement therapy (NRT). At 8 weeks they were reviewed in clinic and assessed for smoking cessation by an expired carbon monoxide breath test. Of the 70% of patients who passed this test, 62% quit “cold turkey”, 24% with NRT and 14% with outpatient treatment programmes. Importantly, 64% continued their smoking abstinence at 6 months post-operatively.

Compliance with smoking cessation can be tested by assessing the patients’ cotinine level in a blood test. Cotinine is a metabolite of nicotine and is found in the saliva, urine and blood of smokers. The half-life of cotinine is about 20 h and so a serum cotinine level of <10 ng/mL has been shown to reflect a patient’s compliance with non-smoking [39, 44].

24.2.1.3 Obesity

Obese patients appear to be at increased risk for deep joint infection following primary or revision total knee arthroplasty [46–49]. Potential causes included the longer surgical time, greater soft tissue dissection and the presence of thick layer of poorly vascularized subcutaneous tissue [50–52]. In addition, obesity is associated with the presence of comorbidities such as diabetes mellitus and immunosuppression and malnutrition [50–54]. Peterson et al. in 2016 showed an array of nutritional deficiencies in patients who were due to undergo bariatric surgery [53]. Of the 58 obese patients, 15.6% had hypoalbuminemia, 92.9% vitamin D deficiency and 36.2% iron deficiency.

Although many surgeons advocate for weight loss in patients with elevated BMI prior to surgery, the evidence on outcomes of patients who have lost weight prior to TKA is limited. Inacio et al. showed that patients that had a decrease of body weight by 5% had no difference in rate of surgical site infection than a cohort that did not lose weight [55] In contrast to this, Malinzak et al. demonstrated an increased risk of PJI by

approximately three times for obese patients with a BMI > 40 and by 21 times for patients with a BMI > 50 [56]. This study was supported by another study that showed increased PJI in patients weighing over 120 kg [57].

Bariatric surgery (a lap band or gastric bypass) may be performed to help morbidly obese patients lose weight; however, having had prior bariatric surgery does not seem to decrease the incidence of PJI, but might decrease the incidence of superficial surgical site infection [54, 58–60]. Bariatric surgery prior to TKA, as opposed to after TKA is advocated due to overall decrease in complications from TKA [58].

Prophylactic antibiotics should be dosed based on the patients' weight. Rondon et al. showed that in their cohort of patients 95.9% of patients who weighed over 120 kg were underdosed at time of primary total joint arthroplasties, and that his cohort also had a significant increase in incidence of PJI by 1 year than patients who weighed less than 120 kg. The current recommendation for cefazolin weight-based dosing protocols is to give 1 g if the patient weighs <60 kg, 2 g if the patient weighs 60–120 kg, and 3 g if the patient weighs >120 kg [61, 62].

24.2.1.4 Malnutrition

Recent studies have suggested malnutrition is associated with an increased risk of infection resulting in more screening for malnutrition. Tests for malnutrition include total leukocyte count <1500 mm³, serum albumin <3.5 g/dL and transferrin level <200 mg/dL [63]. The prevalence of malnutrition is higher than is often acknowledged and paradoxically, some obese patients requiring TKA tend to suffer from malnutrition despite large deposits of body fat.

Green et al. reported an increased risk of infection in malnourished patients who underwent THA or TKA [64]. Adequate preoperative nutritional status was defined as a total lymphocyte count of ≥ 1500 cells/mm³, and albumin level of ≥ 3.5 g/dL. Of the 217 patients included in the study, 57 had low preoperative lymphocyte counts and 4 low albumin levels, with 2 patients low with both parameters. Major wound complications were defined as either a superficial infection

(3.7%), deep infection (1.8%), or wound dehiscence (0.9%). The major wound complication cohort had significantly ($p = 0.002$) lower preoperative lymphocyte count than those who did not have a complication with a mean lymphocyte count of 1553 vs. 1995 cells/mm³. The albumin level trended lower in the major wound complication group but did not reach significance.

Jaberi et al. did a retrospective review of 300 patients who underwent a TKA or THA and who had persistent wound drainage [63]. Malnutrition as defined by total leukocyte count <1500 mm³, serum albumin <3.5 g/dL and transferrin level <200 mg/dL was associated with increased risk of developing an infection. Peersman et al. found an association with poor nutrition, obesity and diabetes mellitus and risk of PJI in a retrospective review of 97 PJI in 6489 TKAs, although this paper did not define what constituted poor nutrition [65].

Anthropometric parameters of malnutrition, specifically the triceps skinfold (TSF) was studied in 213 patients undergoing TKA, as well as biochemical measures of nutrition such as the total lymphocyte count and albumin levels [66]. This study found that none of the preoperative biochemical markers was associated with infection; however, there was a statistically significant inverse relationship between TSF and infection. Anthropometric parameters are thought to be a better measure of long-term nutrition than biochemical markers as biochemical markers can be affected by chronic and acute diseases such as renal failure, liver diseases, cancer, and conditions associated with stress and inflammation [66].

From this literature it could be suggested that preoperative management of arthroplasty patients should involve biochemical screening for malnutrition, and further management of any nutritional deficiencies in conjunction with advice from a dietician.

24.2.1.5 Urinary Tract Infection

Patients who are to undergo TKA should be screened for urinary tract infections (UTI). Wang et al. [67] recently showed in a meta-analysis of primary joint replacements that the relative risk for developing PJI when a patient had a periop-

erative UTI was 3.17. They also showed that the microorganism causing the UTI and PJI were the same in the same patient, supporting the notion that the PJI is caused by haematogenous spread from the genitourinary tract [67]. Pulido et al. [5]. showed an increased risk of PJI in patients with a urinary tract infection. In contrast to these studies, Schmitt et al. [68] showed that the post-operative UTI was associated with both SSI and PJI, but not preoperative UTI. Regardless of the controversy as to the risk of PJI from preoperative UTI, it seems low risk to at least instigate treatment for any preoperative UTI prior to proceeding to TKA.

24.2.1.6 Dental Procedures

The issue of dental surgery following TKA is controversial. The American Academy of Orthopaedic Surgeons (AAOS) and the American Dental Association released a report in 2012 that reviewed the literature around prophylactic antibiotics with dental procedures and joint replacements in situ. Recommendation one from this report is that “the practitioner might consider discontinuing the practice of routinely prescribing prophylactic antibiotics for patients with hip and knee prosthetic joint implants undergoing dental procedures” [69]. The Arthroplasty Society of Australia released a position statement in 2016 that echoes the recommendation from AAOS. The recommendation from this report is that “no routine antibiotic prophylaxis be given to patients with joint prostheses undergoing dental procedures” [70]. The second recommendation of this report is that patients who are immunocompromised or who have poor oral hygiene should be assessed on a case-by-case basis. A large study from Taiwan with a cohort of 255,568 patients who underwent THA or TKA were divided into dental and nondental cohorts based on whether the patients had undergone a dental procedure within the first 2 years following arthroplasty. There was no difference in the incidence of PJI between the cohorts, with a rate of 0.57% in the dental cohort and 0.61% in the nondental cohort [71]. Cost-effectiveness modelling has also been done to compare the benefits, harms and cost of prophylactic antibiotic dosing with dental proce-

dures, and has shown that routine antibiotic prophylaxis is not cost-effective for dental procedures, but might be in the higher risk population [72].

Common sense suggests that any dental issues should be dealt with prior to arthroplasty surgery. Vuorinen et al. showed that 29.4% of patients who underwent dental screening prior to joint arthroplasty did not pass dental clearance and proceeded to dental procedures prior to their arthroplasty [73]. Sonn et al., however, showed in a retrospective review that patients with poor oral hygiene do not have an increased risk of PJI, and so concluded that routine dental clearance was not necessary [74].

24.2.1.7 Hypothyroidism

Hypothyroidism has recently been shown to be associated with an increased risk of PJI [75, 76]. Buller et al. [75] found an odds ratio of 1.502 for developing a PJI in the first 90 days post TKA in patients with hypothyroidism. This study had 98,555 gender and age matched cohorts retrospectively reviewed from US Medicare patients who underwent a primary TKA. This study is supported by a paper by Tan et al. [76] who looked at 32,289 patients who underwent primary or revision TKA or THA and found hypothyroidism to be an independent risk factor for PJI with an adjusted odds ratio of 2.46 ($p < 0.0001$). Thyroid stimulating hormone (TSH) levels were also significantly higher in the patients who developed PJI [76]. We recommend consultation of an endocrinologist to optimize a patients’ serum thyroid stimulating hormone levels prior to surgery.

24.2.1.8 Preoperative Anaemia

Preoperative pathology is routinely recommended including haematology and biochemistry. Included in these studies should be evaluation of iron levels as well as haemoglobin. Anaemia is defined as haemoglobin level of less than 12 g/dL in women and less than 13 g/dL in men by the World Health Organization. The rate of preoperative anaemia in patients undergoing THA or TKA is 15–33% [77]. Preoperative anaemia has been associated with both an increased risk for devel-

oping a PJI, as well as failure of a debridement and polyethylene exchange (DAIR) in acute haematogenous PJI [77–80]. Allogenic blood transfusion has been shown to increase the risk of post-operative infections [81] and PJI [5]. Greenky et al. [77] found 19.6% of 15,222 patients undergoing a total joint arthroplasty (TJA) had anaemia, and the incidence of PJI in this group was 4.3% vs. 2% in the non-anaemic cohort. Allogenic blood transfusions were required in 44% of the anaemic cohort and 13.4% in the non-anaemic cohort [77]. Swenson et al. demonstrated that a preoperative haematocrit ≤ 32.1 had an odds ratio of 6.7% in predicting failure for open debridement and polyethylene exchange in THA or TKA with PJI from acute haematogenous spread [80]. Some centres prefer to optimize patients' level of haemoglobin and any iron deficiency prior to TKA.

24.2.1.9 Drug and Alcohol Abuse

Substance abuse disorder and alcohol abuse disorder have been shown to be independent risk factors for PJI [82]. In a study looking at 11,403 TKA, multivariate analyses showed the odds ratio was 19.419 for alcohol abuse disorder and 3.693 for substance abuse disorder and the development of PJI. When substance abuse disorder and depression were present in the same patient, the odds ratio for PJI increased to 13.639, a four-fold increase [82]. Bauer et al. [83] retrospectively reviewed 18 TKA performed in 12 patients who had a history of intravenous opioid abuse. An alarmingly high rate of PJI was found at 50%, nine knees in seven patients, with three knees requiring above knee amputation and a further three knees ending with arthrodesis [83].

24.2.1.10 Human Immunodeficiency Virus

The literature on whether a human immunodeficiency virus (HIV) positive patient has an increased risk of PJI is mixed. Parvizi et al. in a 2003 study showed an alarmingly high rate of post-operative complications, including six deep joint infections (29%) [84]. In this study, the average CD4 count was 239 cells/mm³ for patients who had a PJI compared to 523 cells/

mm³ in the overall study population. Only 3 patients out of the 15 in the study were on highly active antiretroviral therapy (HAART) at the time of arthroplasty [84]. A similarly high deep infection rate of 14.3% was found in a 2001 study in HIV positive patients, but the CD4 counts and whether the patients were on HAART were not disclosed [85]. A systematic review by Dimitriou et al. [86] of 6,516,186 TKA and THA revealed an elevated risk of complications but no difference in long-term survivorship of the implants. It is unfortunate that this review did not analyse infection risk but rather overall complication rate. Boylan et al. [87] showed that survivorship of TKR in a HIV cohort in comparison to a non-HIV cohort was no different, and importantly showed no deep infections in either cohort. These findings were supported by Roof et al. [88] PJI was found to be higher in patients who had both HIV and haemophilia than in patients who had HIV and no haemophilia with an odds ratio of 5.28 [89]. Issa et al. [90] showed that patients with HIV infection have TKA survivorship similar to that of patients who do not suffer from HIV infection. Patients in their study cohort had CD4 count >200 at the time of surgery and were on active treatment with two nucleotide reverse transcriptase inhibitors and one protease or integrase inhibitor which was continued in the perioperative and post-operative period along with the usual prophylactic antibiotic cover. It is possible that the earlier papers found a higher rate of PJI simply due to the lack of HAART treatment in their cohort of patients, as HAART only began to be used in 1997 [91]. Ensuring that a HIV positive patient has appropriate CD4 count prior to surgery may be a useful strategy to avoid post-operative complications particularly PJI after TKA in patients suffering from HIV infections.

24.2.1.11 Hepatitis C Infection

Patients with hepatitis C virus (HCV) have a higher risk of complications, including PJI, when undergoing TKA [92–95]. Schwarzkopf et al. [96] in their recent study showed that patients with hepatitis C infection should receive perioperative treatment for the infection to decrease the chance of PJI after TKA. This retrospective

review of patients with either cured HCV infection or untreated HCV infection showed that the untreated cohort had significantly higher infection rate (15.5% vs. 4.3% with odds ratio 4.1; $p = 0.03$) [96]. This study is supported by another two studies published in 2019 [97, 98]. Bedair et al. studied the effect of treated HCV compared to untreated HCV in THA and demonstrated the rate of PJI was 14.3% in patients who had untreated HCV and 0% in those who had been treated [97]. Bendich et al. did a similar study looking at treated compared to untreated HCV in patients undergoing either THA or TKA and showed an odds ratio of 3.30 for PJI in untreated patients at 90 days post-operative and 2.16 at 1 year post-operative [98]. These studies suggest that treatment of HCV should be undertaken prior to proceeding with TKA to decrease the risk of PJI.

24.2.1.12 Nasal colonization with *S. aureus*

Various studies have reported the benefits for screening patients for asymptomatic colonization with methicillin resistant (MRSA) or methicillin sensitive staphylococcus aureus (MSSA) organisms [99–106]. Anterior nares serve as the reservoir for staphylococcus organisms as well as the groin and axillae. Methicillin sensitive staphylococcus organisms are present in anterior nares of 20–30% of orthopaedic patients and methicillin resistant staphylococcus organisms are present in 2–6% of pre-op patients for TKA [103]. Surgical site infection after TJA is caused by MSSA or MRSA in >60% of patients [107]. Nasal decolonization of MSSA and MRSA can be done through twice daily application of mupirocin for 5 days prior to surgery [108]. Decolonizing patients who are positive with staphylococcus aureus helps to decrease the incidence of surgical site complications in total arthroplasty patients [99, 101, 102, 104–106]. Another added benefit for the preoperative screening is that it acts as a guide for choice of prophylactic antibiotic for surgery. If the patient is colonized with MSSA, routine antibiotics may be used. If a patient is colonized with MRSA, weight-based vancomycin is the appropriate choice [109]. Some centres

provide nasal decolonization empirically to all the preoperative patients undergoing TKA and do not rely on the results of the swab analyses. Patients may also be prescribed with betadine shower and chlorhexidine wipes or shower prior to surgery, which is an additional useful strategy if patients have skin or axillary/groin colonization with staphylococcus aureus.

24.2.1.13 Immunosuppression

Organ transplantation patients are known to require immunosuppression after receiving their transplantation and so are thought to be an at-risk group for developing PJI if they go on to require an arthroplasty. Palmisano et al. [110] did a retrospective review looking at patients who had had a solid organ transplantation and subsequently went on to have a THA or TKA. These patients were on different combinations of anti-rejection medications including azathioprine, mycophenolate, cyclosporine, prednisone and tacrolimus. There were three deep infections recorded out of the seven TKAs that were performed, giving a deep infection rate of 42.9% [110].

24.3 Conclusion

There are many potentially modifiable host risk factors which should be identified and addressed prior to TKA to minimize the risk of PJI. Screening should be performed 4–6 weeks prior to planned surgery to avoid the difficult decision for the clinician and the patient to postpone surgery in the anaesthetic bay. The time to identify a potentially modifiable risk factor is well prior to TKA. Involving the patient in the decision-making process by making the patient aware of the available options to optimize the risk factors prior to TKA is appropriate and to proceed with TKA when the patient is in as good a condition as is possible. Patients should be made aware that without optimization there is an increased chance of suffering from a PJI. The surgeon has an important role to identify the potentially modifiable risk factors of PJI prior to TKA and make every effort to optimize before surgery. A balanced clinical decision requires respect for the

patients' autonomy at the same time acting in their best interests to do no harm (Non-maleficence). This will minimize length of stay and additional surgical procedures for wound problems after joint arthroplasty and reduce readmission rates to avoid financial constraints on the healthcare providers.

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Antibiotic Prophylaxis in Primary and Revision Total Knee Arthroplasty

25

Francesco Giron

25.1 Introduction

Total knee arthroplasty (TKA) is one of the most successful surgical procedures for the treatment of end-stage arthritis of the knee and the number of primary TKAs will continue to increase because the arthritis is an age-related disease and the life expectancy is increased. It has been estimated that, by 2030, more than four million primary total hip arthroplasty (THA) and TKA procedures will be performed annually in the United States [1, 2]. Despite advances in surgical techniques and infection control efforts, surgical site infection (SSI) following TKA remains an unsolved catastrophic complication. Infection after TKA is one of the major problems that have not been solved during the last 30 years. It can be devastating, and although rarely causing death, infection is associated with increased morbidity and hospitalization. The deep infection rate after primary TKA has been reported between 0.86 and 2.5% [3–6]. If these rates remain constant, by 2030, the estimated number of deep infections following joint arthroplasty will be 40,000–80,000 per year. Along with additional risk for the patient, infection after TKA can bring on a huge financial burden. Patients with an infection

are twice as likely to die, twice as likely to spend time in an intensive care unit, and five times more likely to be readmitted after discharge [7].

Development of an infection depends on the number and virulence of the bacteria introduced into a wound, the host's ability to eliminate these bacteria, and the status or viability of the wound environment. Various measures can be adopted to prevent this devastating complication in TKA patients, including optimizing medical comorbidities and patient risk factors, managing the operating room environment (e.g., laminar flow, body exhaust suits, minimizing operating room traffic), using proper skin preparation, and carefully selecting and effectively using antibiotic prophylaxis [8–10].

The majority of early postoperative infections results from intraoperative contamination of the surgical site [11]. Even with a strict aseptic technique, bacterial contamination occurs in most if not all arthroplasty procedures [12].

Antimicrobial prophylaxis is considered beneficial for preventing surgical site infections in clean orthopedic surgery. It is probably the single most effective method for reducing the prevalence of postoperative wound infection. Prophylactic antibiotics have been described as antibiotics given for the purpose of preventing infection when infection is not present but the risk of postoperative infection is present [13]. The goal of antimicrobial prophylaxis is to achieve serum and tissue drug levels that exceed, for the duration of the operation, the minimum

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inhibitory concentration for the organisms likely to be encountered during the operation [14].

While the benefits of preventing surgical infections are apparent, one must also keep in mind the disadvantages of excess antimicrobial use. All infections cannot be prevented by the use of prophylactic antibiotics. Each patient has a unique set of immune defenses against, and risks of, infection. The goal of surgical prophylaxis is to decrease the bacterial load at the surgical site, not to sterilize the patient. Essentially, prophylaxis augments the host's natural immune defense mechanisms by increasing the amount of bacterial contamination needed to cause an infection [15].

Antimicrobial prophylaxis is considered helpful for preventing surgical site infections (SSIs) in orthopedic surgery [11]. The World Health Organization (WHO) [7] and the Centers for Disease Control and Prevention (CDC) [16] specified that microbial contamination during a surgical procedure is a precursor of a SSI. Use of prophylactic antibiotics with an antimicrobial spectrum that is effective against the pathogens likely to contaminate the procedure therefore is recommended for use in any clean surgical case. Conversely, the use of broad-spectrum antibiotics promotes the development of multi-drug-resistant organisms. Infections due to resistant organisms are associated with a worse clinical outcome for each individual patient. There must be a delicate balance between the use of antimicrobial agents to prevent infection and the overuse of antimicrobial agents, which is associated with the development of multi-drug-resistant organisms.

The effectiveness of prophylactic antibiotics administered shortly before skin incision to avoid microbial contamination during the procedure was established in the 1960s [17, 18], and it has been recommended in current guidelines for surgical prophylaxis. The ideal prophylactic antimicrobial agent should have excellent *in vitro* activity against bacteria, penetrate tissue well, have a relatively long serum half-life to provide coverage for the duration of the entire operative procedure, be relatively nontoxic, and be inexpensive.

The choice of antibiotics used as prophylaxis requires an understanding of the common micro-

organisms that cause surgical site infections associated with TKA.

25.2 Common Microorganism Involved in Surgical Site Infection

The identification of organisms most frequently involved in SSI is the most important factor for the appropriate decision on what could be the proper drug to be adopted in antibiotic prophylaxis before TKA. Early infections (within 1 year postoperatively) and infections in patients with persistent pain and discomfort since the index surgery are commonly thought to be caused by direct inoculation during the perioperative period, whereas late infections are thought to occur via hematogenous seeding of the prosthesis or through compromised local tissues [19].

Wound infections following clean surgical procedures are primarily caused by skin or exogenous airborne microorganisms since other reservoirs of bacteria, such as the gastrointestinal tract, are not entered. Numerous studies have documented that gram-positive organisms are the most common bacteria causing infections associated with joint arthroplasty.

The most frequent pathogenic organisms causing deep wound infections in clean orthopedic surgery are *methicillin-sensitive Staphylococcus aureus* (MSSA) and *coagulase-negative Staphylococci* (CoNS), such as *Staphylococcus Epidermidis* [11, 19]. CoNS in particular are recognized as the most common contaminant obtained in cultures taken from surgical sites other than MSSA [20], and generally are accepted as being one of the most resistant pathogens worldwide [21]. Other gram-positive organisms, including *Streptococcus* and *Enterococcus* species, can cause infections, as well. Furthermore, the increasing frequency of infection caused by organism such as *methicillin-resistant Staphylococcus aureus* (MRSA) and *vancomycin-resistant Enterococcus species*, which are generally resistant to more than one antibiotic, provides a dilemma with regard to prophylaxis and treatment. In some cases, infections

surrounding a joint replacement can be very difficult to eradicate due to the bacteria's ability to adhere to orthopedic implants and form a local biofilm [22]. This glycocalyx layer, which is formed on the surface of the orthopedic devices, creates a complex environment for the bacteria. This self-produced matrix of extracellular polymeric substance creates a favorable environment for bacterial replication, accelerates mutation rates, confers a relative resistance to host defenses, and impairs effective penetration of antibiotics [23]. Antibiotic treatment can suppress the symptoms of the infection, but eradication usually requires removal of the device and its associated glycocalyx layer.

Other possible sources of infection can be Gram-negative organism such as *Escherichia coli*, *Pseudomonas species*, and *Klebsiella species*. Gram-negative infections are less common and reportedly account for 10–20% of infections [24]. Approximately 20% of periprosthetic joint infections (PJIs) are polymicrobial [25].

All of these microorganisms can be part of normal skin flora. Therefore, direct infection from the patient's skin or airborne contamination from surgical team personnel and the operating room environment is the most probable route of infection. Hare and Thomas [26] described staphylococcal "dispersers" as people who are *Staphylococcus aureus* carriers and lose the organism in vast numbers. Ritter [27] also recognized the importance of the quantity of people in the operating room as a source of increased bacterial counts. Members of the surgical team who have direct contact with the sterile operating field have been linked to unusual outbreaks. Anesthesia personnel also may play a role in postsurgical infections. Although not directly involved in the operative field, they perform a variety of procedures leading up to the operation.

25.3 Patient's Individual Risk Factors

A critical component of preventing postoperative infection is assessing a patient's individual risk factors. While the significance of some risk fac-

tors remains controversial, a high body mass index (BMI), diabetes mellitus, malnutrition, preoperative anemia, cardiovascular disease, and immunosuppressive drugs are well documented as factors increasing infection risk [9, 28, 29]. In 2013, investigators at the Kaiser Permanente Orange County Department of Orthopedic Surgery reported the risk factors associated with SSIs, after analyzing over 56,000 knees [30]. After fully adjusting their model, they reported a patient with a BMI of over 35 was 1.47 times more likely to develop deep SSI, patients were 1.28 times more likely to develop an infection if they had diabetes mellitus and 3.23 times more likely to develop an infection if they had been previously diagnosed with posttraumatic arthritis. Other risk factors such as the male sex put patients at a hazard ratio of 1.89, and an American Society of Anesthesiologists (ASA) score of over 3 made a patient 1.65 times more likely to develop an infection. If a patient is noted to have multiple risk factors, proper counseling and measures should be taken to decrease their risk and increase their compliance with future instructions. To best prevent infection and minimize risk, preventative strategies should span across the preoperative, intraoperative, and postoperative stages [9]. For example, in the case of patients at increased risk of infection, the type of antibiotic and duration of the antibiotic prophylaxis must also be carefully evaluated [31, 32].

25.4 Properties of a Prophylactic Antibiotic

Bacteriostatic antibiotics limit the growth of bacteria predominantly by interrupting bacterial protein production or by inhibiting precursors in folic acid synthesis and DNA replication. These bacteriostatic agents inhibit the growth and reproduction of bacteria without killing them. Bactericidal antibiotics kill the bacteria. The beta-lactams accomplish this by inhibiting cell wall synthesis and inducing cytolysis [33]. Most of the prophylactic antibiotics used in orthopedic surgery are categorized as bactericidal. These include the penicillins, the cephalosporins, van-

comycin, and the aminoglycosides. Clindamycin, a lincosamide, is considered bacteriostatic. High concentrations of most bacteriostatic agents can be bactericidal, whereas low concentrations of bactericidal agents can be only bacteriostatic [34].

The most important consideration in choosing an antibiotic for prophylaxis is its spectrum of action. While the chosen antibiotic may not cover the entire spectrum of organisms that may be encountered, it must be active against the bacteria that commonly cause postoperative infection. Other factors to consider include the pharmacokinetics and pharmacodynamics of the drug. Specifically, the agent must have a half-life that covers the decisive interval (the first 2 h after incision or contamination with therapeutic tissue concentrations from the time of incision to wound closure). Failure to maintain tissue concentrations of the drug above the minimum inhibitory concentration increases the risk of wound infection [35]. Repeat doses of antibiotics may be necessary if the procedure is long, if multiple transfusions are needed, or if the antibiotic is cleared rapidly. The final consideration should be the cost associated with the use of the antibiotic, which should include the costs of drug monitoring, administration, repeat doses, adverse effects, and failure of prophylaxis (e.g., wound infection sequelae).

25.5 Antibiotic Selection and Dosage in Primary TKA

Systemic antibiotics are known to reduce the risk of perioperative and/or postoperative infection [11, 36, 37]. However, some previous studies had reported that systemic antibiotics may not prevent all postoperative infections [7, 38, 39]. Furthermore, conventional systemic dosages may not provide adequate tissue concentrations against more resistant organisms, such as coagulase-negative staphylococci. According to the Surgical Care Improvement Project (SCIP) Advisory Committee, part of a US initiative to reduce surgical morbidity and mortality by 25% by 2010, and the American Academy of

Orthopaedic Surgeons (AAOS), the preferred antimicrobials for patients undergoing total hip or knee arthroplasty are the cephalosporins, in particular cefazolin and cefuroxime [10, 15, 40, 41]. The cephalosporins have been the antibiotics of choice for both the prophylaxis and the treatment of orthopedic infections for at least three decades. Cefazolin has been extensively studied and its favorable activity against gram-positive organisms and its effectiveness against most clinically important aerobic gram-negative bacilli and nonbacteroid anaerobes have contributed to its widespread acceptance. In addition, cephalosporins have excellent distribution profiles in bone, synovium, muscle, and hematomas [42]. Studies have documented that minimum bactericidal concentrations for most non-methicillin-resistant *Staphylococcus aureus* organisms are achieved rapidly in these tissues [43].

Cefazolin is often dosed at 1 g for patients who weigh <80 kg or 2 g for patients who weigh >80 kg. In patients weighing >120 kg, a 3-g dose can be considered [44, 45] (Table 25.1). Cefuroxime is dosed at 1.5 g. It is recommended that, for extended operative times, cefazolin be readministered every 2–5 h; cefuroxime, every 3–4 h [11] (Table 25.1). Both of these cephalosporins are safe and have an effective spectrum of action against the most commonly encountered organisms, specifically gram-positive bacteria and 40% of gram-negative bacteria.

Anaphylactic reactions to cephalosporins are rare events, but they do occur and thus have led to the recommendation against their use in patients with known anaphylaxis to other beta-lactam antibiotics. Some of the more common reactions include skin rash (a rate of 1–5%), eosinophilia (3–10%), diarrhea (1–10%), and pseudomembranous colitis (<1%) [42].

Clindamycin and vancomycin are currently the preferred alternative antibiotics for people with an established allergy to a beta-lactam or with a contraindication to its use and at institutions with high rates of methicillin-resistant *Staphylococcus aureus* infection. Clindamycin has good bioavailability, and at 30 min after infusion has been shown to exceed the minimum inhibitory concentration for *Staphylococcus*

Table 25.1 Antibiotic dosage for routine prophylaxis in primary TKA

| Antibiotic | Dosage | Time before surgery/tourniquet | Redosing schedule (h) |
|-------------|--|--------------------------------|-----------------------|
| Cefazolin | 1 g (<80 kg body weight) 2 g (60–120 kg body weight) 3 g (>120 kg body weight) | Within 30–40 min | 2–5 |
| Cefuroxime | 1.5 g | Within 30–40 min | 3–4 |
| Clindamycin | 900 mg | Within 30–40 min | 3–6 |
| Vancomycin | 15 mg/kg (weight-based) | Within 60–90 min | 6–12 |
| Daptomycin | 6 mg/kg (weight-based) | Within 30–60 min | 24 |

aureus in both animal and human cortical bone samples [11]. The recommended dose of clindamycin is 600–900 mg and for extended operative time, every 3–6 h (Table 25.1). The most severe adverse effect of clindamycin is *C. difficile*-associated diarrhea (the most frequent cause of pseudomembranous colitis). Other side effects include the development of a rash, abdominal pain, cramps, and in high doses a metallic taste in the mouth.

Although clindamycin is effective against many MRSA species, vancomycin is a bactericidal agent that provides coverage against a greater percentage of MRSA species, making it a better choice to cover MRSA. Vancomycin is a large tricyclic glycopeptide molecule that has historically been the first line of treatment for *methicillin-resistant Staphylococcus aureus* infections [46]. The bactericidal action of vancomycin is a result of the inhibition of bacterial cell wall synthesis through the disruption of peptidoglycan biosynthesis. It is active against most gram-positive organisms including *Staphylococcus aureus*, *Staphylococcus epidermidis* (including heterogeneous methicillin-resistant strains), *streptococci*, *enterococci*, and *Clostridium*. Vancomycin lacks activity against *gram-negative bacteria*, fungi, or *mycobacteria*. Similar to cefazolin, vancomycin reaches high concentrations in bone, synovial tissue, and muscle within minutes after administration [47, 48].

Adverse reactions to vancomycin such as infusion-related pruritus and erythema can occur. Red man syndrome, a pruritic, erythematous rash on the upper trunk and face that is occasionally accompanied by hypotension, is associated with its rapid infusion and histamine release in approximately 5–13% of people [49]. This has led to the

recommendation that vancomycin be administered slowly, at a rate of 1 g over 60 min. The recommended dose, which is based on body mass, is 10–15 mg/kg, up to a limit of 1 g, in patients with normal renal function [15]. When vancomycin is used for prophylaxis, its infusion should begin 1–2 h before initiation of the operation (compared with within 1 h for cefazolin) to ensure that the entire dose is administered and adequate concentrations reach the tissues prior to the surgical incision [50]. For extended operative times, repeat administration is recommended in 6–12 h [11] (Table 25.1).

Nephrotoxicity and ototoxicity occur in <1% of patients, with nephrotoxicity being associated with concomitant aminoglycoside use. Other complications include hypersensitivity rash, reversible neutropenia, and drug fever. Daptomycin should be considered as an alternative for people with known anaphylactic or severe reactions to vancomycin [15] (Table 25.1).

Patient-specific factors should be considered with respect to vancomycin dosage. One report found that 69% of patients receiving vancomycin at the standard 1-g dose were being underdosed based on their actual weight [51, 52]. This suggests that, given the high rates of obesity in arthroplasty patients, a weight-based dose of 15 mg/kg should be used.

With regard to the selection and effectiveness of antibiotic prophylaxis, the clinician must consider whether the organism identified on cultures at SSI presentation was within the spectrum of the original prophylaxis administered at the time of the primary surgery. A recent study of 163 patients with PJI demonstrated that, in 63% of patients, the infections were caused by a bacterium that was resistant to the original prophylaxis.

laxis. MRSA was isolated from 26% of patients with cultures positive for infection [53].

Over the past decade, hospitals and emergency rooms have seen a changing pattern of infections caused by *Staphylococcus*. Usually resistant strains of *Staphylococcus* were reported in hospital settings and high-risk patient populations, such as intravenous drug users and people with chronic indwelling catheters. Recent articles have described an alarming upward trend in the prevalence of community-acquired methicillin-resistant *Staphylococcus aureus* strains in low-risk patients. One report from a large urban hospital in Chicago showed that the prevalence of community-acquired methicillin-resistant *Staphylococcus aureus* skin and soft-tissue infections increased 6.84-fold: from 24.0 cases per 100,000 people in 2000 to 164.2 cases per 100,000 people in 2005 [54]. Additional studies from large county institutions in Dallas and Atlanta have demonstrated similar trends of increasing prevalence of community-acquired methicillin-resistant *Staphylococcus aureus*, with the conclusion being that this is now the predominant organism in skin and soft-tissue infections [55].

Given the varying levels of antibiotic-resistant organisms present at institutions, it is important to customize antibiotics based on local trends. The use of a local and up-to-date antibiogram and consultation with an infectious disease specialist can help clinicians estimate the prevalence of antibiotic-resistant organisms, aiding selection of effective prophylactic agents. Given the increasing prevalence of MRSA, we must specifically address whether every arthroplasty patient should routinely receive vancomycin, either as a single medication or as a supplemental antibiotic. Current guidelines suggest that vancomycin is a reasonable choice of antibiotic for patients with a beta-lactam allergy, those known to be colonized with MRSA, and those at high risk of developing a MRSA infection (e.g., patients in regions with a high prevalence of MRSA, institutionalized patients, healthcare workers) [14, 45]. The 2013 Proceedings of the International Consensus on Periprosthetic Joint Infection broadly support the routine use of vancomycin in patients who

are known MRSA carriers, those with a known anaphylactic allergy to penicillin, or those at high risk of MRSA infection [40]. Additionally, the routine use of dual antibiotics is generally not supported [56].

Sewick et al. [38] compared dual prophylaxis with cefazolin and vancomycin versus cefazolin alone. In their retrospective analysis of 1828 primary THAs and TKAs, with 1-year follow-up, the authors found that the rates of infection with cefazolin and vancomycin versus cefazolin alone did not significantly differ (1.1% and 1.4%, respectively; $P = 0.636$). The prevalence of MRSA infections was significantly lower in the dual antibiotic group than in the cefazolin group (0.02% and 0.08%, respectively; $P < 0.05$). However, these infections were very rare in the cohort; therefore, the number needed to treat to prevent one MRSA infection was very high [43]. Tyllianakis et al. [57], in a prospective randomized study comparing cefuroxime to two antistaphylococcal agents (fusidic acid and vancomycin), for prophylaxis in total hip arthroplasty (THA) and total knee arthroplasty (TKA), investigated the incidence of SSI in an institute, where methicillin-resistant *Staphylococcus aureus* (MRSA) and methicillin-resistant *Staphylococcus epidermidis* (MRSE) prevalence exceeds 25% of orthopedic infections. Four hundred thirty-five patients, who were included in the study, were followed up for a mean time of 3.8 (2–5) years. The authors found that the use of alternative antibiotic agents (including vancomycin) was no better than cefuroxime alone in preventing SSIs. Wyles et al. [58], investigating 29,695 arthroplasties (22,705 patients) performed from 2004 to 2017 at the Mayo Clinic, in order to characterize antibiotic choices for perioperative TKA and THA prophylaxis, assess antibiotic allergy testing efficacy, and determine rates of prosthetic joint infection (PJI) based on perioperative antibiotic regimen, found that PJI rates were significantly higher when non-cefazolin antibiotics were used for perioperative TKA and THA prophylaxis. Given the low rate of true penicillin allergy positivity, and the readily modifiable risk factor that antibiotic choice provides, they also emphasized the role of perioperative testing and clearance for all patients presenting with penicillin and cephalosporin allergies.

Currently, the evidence to support the use of vancomycin for routine prophylaxis is controversial. Up to date, the American Academy of Orthopaedic Surgeons (AAOS) generally recommended vancomycin for patients with beta-lactam allergy, those with a known MRSA colonization, or those in institutions with a high prevalence of MRSA [59]. Moreover in a separate information statement, “The Use of Prophylactic Antibiotics in Orthopaedic Medicine and the Emergence of Vancomycin-Resistant Bacteria,” AAOS stated: “Vancomycin may be appropriate as a prophylactic antimicrobial for patients undergoing joint replacement at institutions that have identified a significant prevalence (e.g., >10–20%) of *methicillin-resistant S. aureus* (MRSA) and *S. epidermidis* among orthopedic patients” [60]. However based on the abovementioned papers that show no clear superiority of vancomycin compared to cephalosporins in reducing postoperative SSI rates after primary TKA, non-consensus actually exists on its routine prophylactic use also in institutions with a high prevalence of MRSA. Moreover, Song et al. [61], investigating the outcome of cefazolin prophylaxis in 1323 TKAs performed in a hospital with a high endemic rate of MRSA infection, found that antimicrobial prophylaxis using only cefazolin can maintain low SSI rates if other adequate infection management measures, such as the use of an appropriate antiseptic agent for surgical scrub and skin preparation, HEPA filter, laminar air flow, and traffic control, are employed, even where there is a high prevalence of MRSA infection.

The unwillingness to use vancomycin as a routine prophylactic agent can be referred to the limited number of antibiotics available to treat *methicillin-resistant Staphylococcus aureus* as well as when antimicrobial profiles did not support its use. In addition, the fear of promoting possible vancomycin-resistant strains of staphylococci and the emergence of vancomycin-resistant enterococci caused physicians to be appropriately cautious about its use. The use of oral vancomycin to treat pseudomembranous colitis contributed to the emergence of vancomycin-resistant enterococci [46]. The first staphylococci with reduced susceptibility to vancomycin were reported in Japan in 1997 [62].

These staphylococci, labeled “*vancomycin-intermediate Staphylococcus aureus*,” did not possess the resistance genes but had a reduced susceptibility to vancomycin. Since then, other strains with reduced susceptibility (heteroresistant vancomycin-intermediate *Staphylococcus aureus*) as well as resistant strains (vancomycin-resistant *Staphylococcus aureus*) have been identified but occur infrequently [63]. To help combat these resistant strains, new antibiotics that greatly expand the pharmacologic arsenal have been introduced. These newer antibiotics include linezolid, quinupristin/dalfopristin, daptomycin, and tigecycline. Whether a single preoperative parenteral dose of vancomycin is associated with increased vancomycin resistance or decreased vancomycin susceptibility has not been demonstrated. Conversely, prolonged exposure to antibiotics has been identified as a risk factor for promoting bacterial resistance [64].

Meehan et al. [15], in order to decrease all the risks related to the use of vancomycin as routine antibiotic prophylaxis in primary TKA, suggested to add a single preoperative dose of vancomycin along with the cefazolin to provide prophylaxis against these resistant organisms and the other common bacterial causes of infection in institutions with a high prevalence of MRSA infection.

25.6 Intraosseus Regional Prophylaxis

For antibiotic prophylaxis to be effective, the concentration of antibiotic in the tissues must exceed the minimum inhibitory concentration (MIC) of organisms that commonly cause infection for the period between skin incision and wound closure. *CoNS*, one of the most common causes of infection post-TKA, have relatively high MICs against cephalosporins. Conventional systemic dosing of prophylactic cephalosporins may therefore lead to inadequate tissue concentrations against these organisms [65]. Vancomycin has been proposed as an alternative [66]; however, it requires a prolonged administration time, can cause systemic toxicity, and risks promoting

further antibiotic resistance. Recently, the attention of the research has been focused on low-dose prophylactic antibiotic through intraosseous regional administration (IORA). This route of drug administration may diminish all the previous issues and in primary TKA achieves higher tissue concentrations than systemic administration by limiting distribution of the drug to the targeted limb [67].

IORA involves intraosseous injection of antibiotics through a specific cannula (Vidacare, San Antonio, TX, USA; FDA-approved) inserted into the proximal tibial bone after tourniquet inflation and before skin incision. Even in adults, intraosseous injection is equivalent to IV administration and is reliably successful in primary TKA, reaching concentrations 6–10 times higher than systemic administration [68–71]. In an animal model of TKA, IORA was also shown to provide more effective prophylaxis against PJI [72].

This approach seems to be very effective in case of obesity. Obesity is an important risk factor for PJI after TKA [73, 74], a devastating complication for the patient [2] and the healthcare system [67]. In a meta-analysis of 83,001 patients, obesity was associated with an odds ratio of 2.2 for superficial infections and 2.4 for deep infections [75]. Furthermore, registry data show a 7% increase in risk per unit of body mass index (BMI) above a threshold of 35 [76]. A number of potential mechanisms are implicated. Obese patients have disrupted microcirculation and macrocirculation [73, 77], decreased wound healing [73, 77], and impaired immune function [73, 78]. Surgically, they are associated with greater difficulty [79] and longer surgical time [80], prolonging exposure to microorganisms. The higher risk of PJI has led some authors to suggest refusing TKA in patients above a certain BMI threshold [81].

In the nonobese TKA population, IORA of prophylactic antibiotics provides tissue concentrations 5–8 times higher than systemic administration in TKA [70]. However, the physiology in the obese patient unpredictably alters pharmacokinetics for different drugs [82]. For vancomycin, there is a higher volume of distribution and a shorter elimination half-life in the morbidly

obese to nonobese individuals [83]. Thus, vancomycin requires total body weight-based dosing to achieve ideal target steady-state concentrations when given systemically [84]. The importance of higher dose of vancomycin is emphasized with bony infections as it displays poor bone penetration in animal models [85]. Chin et al. [86] in a randomized study comparing standard body weight-adjusted vancomycin prophylaxis versus low-dose vancomycin IORA in two groups of 11 obese patients found a statistically significant higher antibiotic concentration in bones of IORA patients.

Based on these assumptions an increased use of this approach should be considered providing higher tissue concentration of antibiotic prophylaxis than systemic administration. Moreover, IORA optimizes time of antibiotic administration and reduces the risk of systemic adverse effects, while providing high tissue antibiotic concentrations during TKA.

25.7 Timing and Duration of Action in Primary TKA

To be effective as an antimicrobial prophylaxis, the serum and tissue drug levels must be greater than the minimum inhibitory concentrations (MICs) for the target organisms for the period between skin incision and wound closure. Therefore, the goal should be to infuse antibiotics within adequate interval before incision or tourniquet inflation (whichever comes first). This allows an optimal antibiotic concentration at the surgical site when the procedure begins [14]. SSI risk increases incrementally with a longer interval between infusion and incision. Administration of antibiotics within 30 min before the incision was associated with a trend toward a lower risk of infection (1.6%) than administration 31–60 min (2.4%) before incision (odds ratio 1.74; 95% confidence interval, 0.98–3.04) [87]. If administered too rapidly, vancomycin can cause a histamine release, resulting in hypotension and a skin reaction called red man syndrome; therefore, infusion of vancomycin should take place over a longer period of time than that for other antibiot-

ics—60–120 min instead of the typical 30–60 min. Additionally, tissue penetration affects the varying infusion times for antibiotics. Cefazolin has a rapid tissue penetration into bone, synovium, and soft tissue [10]. Because slower tissue penetration vancomycin needs to be administered earlier. Moreover, while cefazolin viability is not affected by patient weight, vancomycin shows a substantial difference in trabecular bone concentrations with respect to patient weight with lower body mass index (BMI) achieving greater concentrations [88]. The only exception to these guidelines is in the setting of revision arthroplasty when preoperative cultures of aspiration are negative but there is a high index of suspicion for an infection. In these cases, prophylactic antibiotics should not be administered until deep intra-articular cultures are obtained. Once these cultures are obtained, then the antibiotics can be administered.

To maintain adequate serum concentrations, antibiotics should be redosed during longer surgeries (e.g., 4 h) and when there is increased blood loss (>2000 mL) and/or fluid resuscitation (>2000 mL) [89]. Finally, 24 h is considered the ideal duration for prophylactic antibiotic treatment. Many studies have failed to demonstrate any benefit associated with the use of antibiotics beyond 24 h in elective, clean surgical case [90–92]. In a study of the short-term use of antibiotic prophylaxis in patients undergoing THA and TKA procedures, Heydemann and Nelson [90] found no difference in infection rates between a 24-h and a 7-day dose of nafcillin or cefazolin. In a retrospective review of 1341 THA and TKA procedures, Williams and Gustilo [91] came to the same results in patients treated with either a 24-h or a 3-day course of cefazolin. The risks of excessive antimicrobial treatment, including toxicity and development of antibiotic-resistant organisms, have led to the recommendation for a 24-h course of antibiotics [53]. Limiting unnecessary antibiotic exposure can minimize adverse effects associated with overuse, such as *C. difficile* infection [93]. Hospital-associated *C. difficile* infections carry serious morbidity and result in extended hospital stays and increased costs of care [94].

25.8 Role of Screening for Methicillin-Resistant *Staphylococcus aureus* Carriers

There is increasing evidence that *S aureus* colonization is a risk factor for SSI. Patients undergoing orthopedic surgery are colonized with *S aureus* at rates similar to those of the general population and, in that undergoing total joint arthroplasty, nearly 20% are *S aureus* carriers [95–97]. In this scenario, prophylactic antimicrobials may be modified depending on the results of the screening test. Patients may be screened to determine whether they are colonized with drug-resistant bacteria. Nasal carriers have increased rates of skin colonization, which is important to note because the skin is directly exposed to the surgical field at incision. In those cases, attempts at eliminating these drug-resistant bacteria can be made. This approach has been used with success in The Netherlands and is thought to be a contributor to the fact that $\leq 1\%$ of *Staphylococcus aureus* isolates are methicillin-resistant there. At 49 hospitals in The Netherlands reporting to the European Antimicrobial Resistance Surveillance System during the years 1999 through 2004, only 58 (0.78%) of 7420 cultures were positive for methicillin-resistant *Staphylococcus aureus* isolates [98].

A universal program of *S aureus* screening and decolonization before high-risk orthopedic procedures (e.g., TJA, spine fusion) has been developed [96]. Patients' nares are cultured 7–10 days before the procedure. All patients are then given a prescription for nasal mupirocin for decolonization. On the day of surgery, patients are asked if they complied with the protocol. For those who report compliance, the cultures are checked and, if they test positive for MRSA, they receive vancomycin prophylaxis within 30 min of the skin incision and the typical preoperative preparation for their surgery. Patients whose cultures are negative receive the typical cephalosporin prophylaxis before the incision. If patients did not comply with the protocol and their cultures are negative or positive for methicillin-sensitive *S aureus* only, they also receive the typical cepha-

losporin or clindamycin prophylactic antibiotics before the incision. Patients who did not comply and have nasal cultures positive for MRSA receive preoperative vancomycin prophylaxis, are decolonized with mupirocin after surgery, and placed on isolation precautions after surgery until the decolonization protocol is complete. After adoption of this universal screening and decolonization protocol, the overall MRSA burden for the hospital was shown to decrease, and the overall deep infection rate associated with TJA was reduced from 1.45 to 1.28% after initiation of the protocol [97, 99]. However, this difference was not statistically significant and, to reach adequate power, 57,604 patients would be needed in each group.

As an alternative to obtaining preoperative cultures, polymerase chain reaction (PCR)-based testing has emerged as an effective tool for detecting MRSA colonization [95, 97, 100]. The accuracy of PCR-based tests for detection of *S aureus* has been validated in the literature and has been found to be sensitive, specific, and cost-effective [78, 101].

There is increasing evidence that *S aureus* carrier screening and decolonization have the ability to decrease perioperative infection rates; these procedures can be highly cost-effective and may improve outcomes [102]. It should be noted that the decolonization is not permanent, and patients who are decolonized have a significant risk of being recolonized [103]. This means that patients decolonized for a procedure will need to be rescreened if they undergo a second procedure or if their surgery is postponed. Additional studies are needed to determine the ideal screening and decolonization protocol and whether it is the decolonization process itself, administration of vancomycin for MRSA-colonized patients, or a combination of the two that is driving the trend of reduced infection rates.

25.9 Local Antibiotic Prophylaxis

Polymethyl methacrylate (PMMA) bone cement is commonly used for fixation of TKA components and its primary function is to transfer load

force from prosthesis to bone. Initial medical applications of PMMA were in dentistry beginning in 1940 and in orthopedic surgery with John Charnley's early work on THA between 1950 and 1960.

Bone cement has the capacity to release antibiotic molecules if any antibiotic is included in it, and these elution properties are improved as cement porosity is increased. In vitro studies have shown high local antibiotic concentration for many hours or few days after its use. Mixing antibiotics into bone cement allows for direct delivery of antibiotics to the implant and surgical site immediately following surgery. Buchholz et al. [104] were the first to report on the addition of aminoglycoside antibiotics to Palacos bone cement in a large series of exchange arthroplasties. The aminoglycosides are a class of antibiotics that can be used in a prophylactic fashion, being that they are administered locally rather than parenterally. They cause bacterial cell death by an intracellular mechanism, binding to a 30S subunit of the ribosome and thereby inhibiting protein synthesis. This practice is common and widely accepted in revision arthroplasty either in the creation of a spacer in the first stage of the procedure or as part of the cementing process in the replantation stage [105]. However, the use of antibiotic-loaded bone cement in primary TKA is controversial. Many authors have recommended the use of antibiotic-loaded bone cement (ALBC) in TKA for infection prophylaxis, but the evidence based on data from National Registries, randomized clinical trials and meta-analysis suggests a protective effect of ALBC against infection when used in hips, but not (or only mild) in knees. A possible explanation is that the quantity of locally delivered antibiotics after TKA is small.

There are some concerns about the routine use of ALBC in primary TKA as prophylaxis against infection. Firstly, there is a risk of hypersensitivity or toxicity even when the chance is highly improbable. Secondly, there is a reduction in the mechanical properties of the cement, but this can be probably neglected if the antibiotic is used in low doses, not more than 1 g per 40 g cement package. Another significant concern is the

increased economic cost, which could be overlooked if there were enough savings in treating fewer prosthetic infections. Finally, there is also a risk of selection of antibiotic-resistant strains of bacteria and this could be the main concern. If used, the choice of the antibiotic mixed in ALBC should consider microbiological aspects (broad antimicrobial spectrum and low rate of resistant bacteria), physical and chemical aspects (thermal stability, high water solubility), pharmacological characteristics (low risk to allergic reactions or toxicity), and economic aspects (not too expensive). Currently, the most commonly used antibiotics in ALBC are gentamicin, tobramycin, and vancomycin.

Several properties of bone cement are important to consider when creating an antibiotic-cement mixture [106, 107]. First, PMMA polymerization is an exothermic reaction; therefore, antibiotics must be heat stable. Second, the antibiotic itself must be water soluble to allow diffusion into the surrounding tissues. It must have a bactericidal effect at the tissue concentration and be released gradually over an extended period of time. Furthermore, the antibiotic must result in minimal local inflammatory or allergic reaction. Finally, composition of different bone cements differs and so the chance for release of antibiotics is not the same.

The mechanical and chemical stability of a variety of antibiotic-cement combinations has been studied. During the polymerization reaction of bone cement, there is an increase in temperature that causes the formation of air bubbles. Some of these bubbles escape from cement, but some others do not escape, causing some porosity in it. The final porosity of bone cement depends not only on the composition and method of manipulation, but also on the viscosity of the cement [108]. An increased cement porosity causes a decrease in the mechanical properties, but an increase in the capacity of the cement to release antibiotic molecules if any antibiotic is included in it. Historically, concerns about whether antibiotic loading decreases the strength of PMMA cement have been expressed. Lautenschlager et al. [109] showed that adding large doses of gentamicin (4.5 g per 40 g cement)

or liquid antibiotics caused a significant decrease in compressive strength to substandard levels. However, at the lower doses used for prophylaxis (2 g per 40 g cement), this change in strength is likely negligible [110, 111].

The initial release after exposure of ALBC to a fluid is mainly a surface phenomenon, while sustained release over the next days is a bulk diffusion phenomenon [108]. The elution of antibiotics from ALBC has been advocated to be effective for many days [43], but some other authors sustain that the process is sufficient for only few hours [112, 113]. Nevertheless, the hydrophobicity of the cement limits the antibiotic release at less than 10%, and most of this antibiotic is released during the first hours after surgery [108, 114, 115]. Three days after its use there is no effect of antibiotic in the ALBC in *in vitro* studies [114]. The elution can be improved by using liquid antibiotics instead of powder ones in the cement, but this choice creates a reduction in the compressive strength of the cement [116].

Aminoglycoside antibiotics (e.g., gentamicin, tobramycin) have favorable properties for this application [105]. Other antibiotics, including vancomycin, erythromycin, and colistin, have been used, as well.

Antibiotics contained in ALBC, though at low levels, are systemically absorbed and can potentially cause allergic reactions. Particular attention should be paid to an individual's antibiotic allergy history prior to implantation of any ALBC. The most frequently used antibiotics in ALBC are aminoglycosides (gentamicin and tobramycin), which very rarely cause allergic reactions. The possibility of an allergic reaction may become greater if other antibiotics such as cephalosporins are used [117].

There is an increasing concern in the emergence of drug-resistant organisms. No direct evidence links the development of bacterial resistance to the routine use of ALBC in primary arthroplasties and some authors do not believe that this risk is increased [118]. There is some evidence supporting the concern about antimicrobial resistance and the risk of selecting resistant mutants bacteria: *in vitro* studies show up to 8% of the antibiotic in ALBC is quickly released

after surgery, and thereafter there is a low-dose release, that may not be effective at fighting infection, but can cause antibiotic microbial resistance. Prolonged exposure to antibiotic at a dose concentration below the inhibitory one allows the development of mutational resistance in bacteria [108, 112]. Josefsson et al. [119] found that 88% of the infected patients who had received gentamicin-loaded cement in primary arthroplasty showed at least one gentamicin-resistant isolate. Aminoglycoside (gentamicin and tobramycin) resistance rate is higher if an antibiotic spacer is used in 2-stage revision arthroplasty [120], suggesting that the risk of selecting resistant mutants when using ALBC is real. In a large series of patients, Hansen et al. [121] found that the introduction of routine ALBC in TKA in a hospital did not cause any significant change in the infecting pathogen profile or any alarming increase in antibiotic resistance, but they recognized that the sample size of the infected cohort might not be big enough. Recently Wu et al. [122], analyzing the incidence of SSI and PJI in a group of 3152 patients who underwent TKA between 2009 and 2013, found that the incidence of SSI and deep-implant SSI was 1.52% and 0.79%, respectively. An optimal dose of systemic antibiotics adjusted by patients' body weight for prophylaxis and the use of ALBC were significant protective factors for SSI. Meanwhile, the use of ALBC also significantly decreased the risk of PJI ($P < 0.01$).

The United States Food and Drug Administration (FDA) has approved the use of premixed antibiotic bone cement (either gentamicin or tobramycin) for prophylaxis in a second-stage reimplantation following a previous infection at the site of an arthroplasty, but not as prophylaxis in routine primary arthroplasties. Outside the USA, the use of antibiotic cement for routine primary THA or TKA has been well studied. Large studies of data from the Scandinavian registry established the efficacy of antibiotic cement in THA [123, 124]. In a recent study of TKAs from the Canadian registry, Bohm et al. [125] analyzed a sample of 36,681 TKAs. In 45% of these procedures, antibiotic-loaded cement was used. No significant difference between the groups treated with

or without antibiotic-loaded cement was found with regard to 2-year revision rates for infection or any other cause. In a large, prospective, randomized controlled trial, Hinarejos et al. [126] examined the efficacy of antibiotic-loaded cement in reducing the incidence of infection following TKA. The authors randomized 2948 patients to TKA with standard cement or with erythromycin/colistin-containing cement. The authors reported comparable rates of deep infection in the two groups: a rate of 1.4% in the antibiotic group versus 1.35% in the standard cement group ($P = 0.96$). Similarly, Kleppel et al. [51] doing a systematic review did not find a statistically significant difference between ALBC and non-ALBC groups. Currently, no conclusive evidence exists regarding the efficacy of antibiotic-loaded cement in primary TKA.

Finally, the issue of cost is critical. The average cost of premixed antibiotic in PMMA is approximately \$300 per bag [8]. Illingworth et al. [8] reported that the cost of premixed antibiotic-loaded bone cement for 100 procedures (two bags per procedure) would be about \$60,000. This is similar to the cost of treating one prosthetic infection. Therefore, for routine antibiotic-loaded cement to be cost-effective, it would have to demonstrably prevent one infection for every 100 primary arthroplasty procedures. An absolute decrease in infection rate of 1% would be difficult to achieve in practice, given that the baseline infection rates are already low (1–2%). Gutowski et al. [127] performed a similar cost analysis for antibiotic-loaded bone cement used in TKA and found that there is likely a cost benefit with hand-mixed cement, with the average cost per infection prevented ranging from \$2112 to \$37,176. This is lower than the cost of a revision procedure. The cost of premixed cement for TKA was \$112,606 per infection spared. From a merely economic point of view, the use of ALBC might only be justified in high-risk groups of patients such as those having rheumatoid arthritis [128], immunodepression, morbid obesity [129–131], and diabetes [129, 132, 133], or patients with previous history of infection or fracture in the knee, and those having long surgeries [78, 91, 113, 162], groups where a much higher infection

rate than the average could be expected. Moreover, a recent study stated that the use of ALBC in primary TKA might not be justified even in the group of patients considered as high risk [134].

Currently, given the mixed results regarding its efficacy, no recommendation can be made regarding the routine use of antibiotic-loaded cement in primary arthroplasty. One common practice is to use it only in patients with a high risk of infection (e.g., patients with diabetes mellitus, morbid obesity, prior history of PJI). The 2013 Proceedings of the International Consensus on Periprosthetic Joint Infection echoes this recommendation, with 90% agreement on the statement that antibiotic-loaded PMMA should be used in elective arthroplasty in high-risk patients only [40].

25.10 Antibiotic Prophylaxis in Revision TKA

The use of a preoperative systemic antibiotic prophylaxis has been demonstrated to be effective in reducing the infection rate in primary TKA. However, no consensus exists on the efficacy of antibiotic prophylaxis in TKA revision surgery, mainly in case of PJI. To date there has not been any sort of systematic review of RCTs examining the effect of antibiotic prophylaxis solely on revision TKA. There have been systematic reviews and meta-analysis that have examined the effect of antibiotic prophylaxis on primary and revision TKA collectively, without separating them out [135, 136]. Several studies also recognize that the periprosthetic infection rate is 2–3 times higher in revision TKA than in primary TKA [137, 138]. Moreover, other studies have shown that patients are at a 9–13 times higher risk of infection in a revision TKA procedure than in a primary TKA [138, 139]. Further, the infection rates in revision TKA have also more than doubled from 1.4% in the 1991–1994 timeframe to 3.0% in the 2007–2010 timeframe [140]. Despite the statistically significant higher infection rates seen in revision TKA, surprisingly, the current strong consensus is that periop-

erative antibiotic prophylaxis should be the same for primary and uninfected revision TKA [121]. Moreover, preoperative antibiotics are sometimes withheld in patients undergoing revision arthroplasty, as there is concern that occult infection may be present and the administration of antibiotics might affect intraoperative culture results [16]. This would be important if true, as culture results are integral to the diagnosis of PJI and antibiotic sensitivities obtained from these cultures are critical for guiding subsequent antimicrobial therapy. However, recent papers [141, 142] show that preoperative antibiotics should not be withheld before revision TKA surgery because culture results were not affected by a single dose of prophylactic antibiotics.

As previously reported, the most common organisms for implant infection are *Staphylococcus aureus* (50–65%) and *Staphylococcus epidermidis* (25–30%) [143, 144]. However, in revision surgery there is also a constant risk of hospital acquired bacterial infections that are resistant to the antibiotics commonly used prophylactically in arthroplasty surgery [59]. These nosocomial infections include *C. difficile* and methicillin-resistant *Staphylococcus aureus* (MRSA). This is especially important in revision procedures as patients undergoing these procedures have an increased risk of developing bacterial infections due to their more advanced age and length of hospital stay, compared to primary TKA procedures.

In the USA, over the 2006–2012 period there has been a tremendous increase (35%) of TKA revision surgery. This increase in revision procedures in excess of the increase in the number of primary TKA implants is likely due to the prevalence of over seven million people living with THA and TKAs [145]. A main reason for revision has been due to infection, with approximately 35% of large joint implants (THA/TKA) being revised for this reason [146–148].

Based on these assumptions, new prospective randomized studies will have to be carried out to evaluate if it is necessary to adopt a different antibiotic prophylaxis strategy in case of aseptic revision of TKA. In those cases, probably the surgeon should carefully consider the age of the patient, the risk factors, the associated diseases, the dura-

tion and the complexity of the surgery, in order to customize the choice of antibiotics to be used for prophylaxis.

In case of PJI, the approach is different and more studies are available. A PJI typically develops in one of three ways: through perioperative colonization of the implant, hematogenous seeding caused by a bacteremia, or spread from an infection of the surrounding tissue [149]. Moreover, PJI can be classified in three categories based on time of occurrence. Early infections occur within 3 months after surgery. Delayed PJIs appear 3–24 months after implantation and late PJIs after 24 months [67]. Usually early and hematogenous PJIs are classified as acute infections, which often have an acute onset and are caused by virulent microorganisms [67].

The recommended treatment of an acute PJI is drainage, antibiotics, irrigation, and retention of the prosthesis (DAIR) [67, 150]. DAIR, for hip and knee prostheses, has a success rate of approximately 70% [150–152]. In the empirical phase, intravenous antibiotics are started blind after surgery until the causative microorganisms are determined in microbiological cultures [153, 154]. The importance of tailored antibiotic treatment during the targeted phase is well known [155]. However, far less literature is available on which antibiotic to use in the empirical phase.

25.11 Local Antibiotic Prophylaxis in Revision TKA

Revision TJA, even when performed for aseptic reasons, is well known to be associated with significantly higher infection rates as compared to primary procedures. Consequently, many authors advocate for the routine prophylactic use of ALBC in these surgeries. This is supported by a prospective pseudorandomized study of 189 first-time aseptic revision knee arthroplasties, which found a markedly lower deep infection rate at a mean of 89 months in patients whose components were fixed with cement containing 1 g of vancomycin per 40 g bag of plain cement (zero) compared with those who received plain bone cement (7%) [156]. A review of aseptic revision

cases performed between 2001 and 2012 and tracked through the US Kaiser Permanente Joint Replacement Registry found that the use of ALBC was associated with 50% reduction in all-cause re-revision [157]. Registry data confirm the widespread adoption of routine ALBC use in revision surgery. The Australian Joint Replacement Registry reported that 9 of the top 10 types of bone cement used in revision TJA in 2015 were ALBC, representing between 93 and 99% of all cemented component revisions [33]. In this context, the routine use of ALBC in cemented aseptic revision TJA can be considered the standard of care, conferring benefits in terms of both reduced infection rates and all-cause re-revision.

Most commercial preparations of ALBC are limited to aminoglycosides (gentamicin or tobramycin), which have been shown to have favorable bactericidal activity profile against both methicillin-susceptible and methicillin-resistant staphylococci within the joint space. Aminoglycoside resistance in staphylococci is a known issue, however, and increased resistance has been demonstrated after the use of high-dose ALBC in septic revision joint surgery [120]. Thus, there may be a theoretical benefit in adding low-dose vancomycin to ALBC in aseptic revision cases in which the patient had previously received aminoglycoside cement and/or is being treated in an institution with high rates of aminoglycoside resistance. However, this must be balanced against the risk of consequent selection of vancomycin-resistant organisms. At present, given the effectiveness of aminoglycoside-ALBC in reducing infection rates and lack of evidence supporting routine addition of vancomycin to aminoglycoside-ALBC, it is recommended that this is reserved for treatment of active infection.

The optimal dose ALBC in aseptic revision TKA remains controversial. Commercial preparations of ALBC using a base with favorable elution properties contain as little as 0.5 g of antibiotic (e.g., Palacos R + G; Zimmer Biomet), and these appear to be sufficient to confer benefits in reducing the risk of re-revision. Conversely, for surgeon-prepared ALBC, doses of ≥ 2 g of powdered antibiotic per 40 g bag of cement seem to be well tolerated in terms of mechanical characteristics.

In case of delayed and late PJI, ALBC is routinely used in two-stage septic revision surgery, with the purpose to leverage ALBC as a local drug delivery mechanism. Although historically periprosthetic infections were treated with resection arthroplasty and placement of antibiotic laden cement beads, contemporary approaches rely on ALBC-containing spacers to maintain joint space and function, while facilitating local antibiotic delivery. A number of different cement spacer designs are used, with the most marked difference being between static and dynamic spacers. In the latter category, these can be further subdivided based on whether the cement is preset or mixed and molded in the operating room; the bearing surfaces (e.g., cement-on-bone, cement-on-cement, metal-on-poly); whether they are molded or hand formed; and whether they use specific commercial products (e.g., preformed spacers, commercial molds) or regular off-the-shelf arthroplasty components. Although pros and cons exist to each spacer option, there is no consistent benefit of one design over another in terms of effectiveness in eradicating infection when principles of two-stage revision arthroplasty, such as the concomitant use of systemic antibiotics, are otherwise adhered to [40]. However, one disadvantage of preformed spacers is an inability to tailor the antibiotic regimen to the infecting organism.

It is generally accepted that high doses of antibiotics should be used in ALBC at the time of first-stage revision, with the goal of prolonged elution of antibiotics into the joint space and achievement of effective antimicrobial concentrations within the joint space and over a sustained period. Because these spacers are by definition temporary, strategies for optimal elution can be pursued without major concern about the potential adverse effect of mechanical properties of cement. In general, greater amounts of antibiotic will increase elution rates and length of time that effective intra-articular concentrations will be maintained. No high-quality data exist comparing clinical effectiveness of different antibiotic concentrations [158]. However, a general consensus exists that low-dose ALBC preparations of 1 g antibiotic per 40 g bag are insufficient

for therapeutic use, whereas doses >8 g per bag have been reported to adversely affect handling characteristics during spacer formation [159]. Effective infection control has been reported with doses as low as 1.2 g per bag, the most commonly reported doses in the literature ranging from 3.4 to 8.6 g of antibiotic per bag of cement [22]. Although isolated case reports of adverse events have been attributed to systemic antibiotic toxicity, these seem to be exceptionally rare and insufficient evidence exists to justify decreasing antibiotic concentrations.

Antibiotic elution rates are known to be the product of several factors that can be varied independent of the concentration of antibiotic used, including cement surface area and porosity. Several cement preparation strategies have been shown to increase the elution of antibiotics and should be used when forming spacers for first-stage revision procedures. These include high-speed hand-mixing (3+ cycles per second) under atmospheric pressure, adding powdered antibiotics after cement mixing is complete, not crushing antibiotic crystals, and not adding additional liquid monomer in an attempt to compensate for the increased volume of powder added [160–162]. It is worth noting that higher concentrations of powdered antibiotics increase cement porosity, further increasing their elution.

Selection of antibiotics for surgeon-mixed high-dose ALBC should be guided by the sensitivity profiles of the infecting organisms, while ensuring that they meet the prerequisite criteria for effective local activity (i.e., heat stable and water soluble). The most commonly used antibiotics include the aminoglycosides, such as gentamicin and tobramycin; vancomycin; and cephalosporins, such as cefazolin [22]. For susceptible organisms, the use of cefazolin either in place of or in combination with vancomycin may be advantageous because of its bactericidal activity and superior elution characteristics [163].

The use of at least two different classes of antibiotics is recommended. In vitro studies have demonstrated a synergistic effect of bi- and tri-antibiotic cement on elution rates, although evidence concerning a synergistic effect on bacterial growth is equivocal [163, 164]. However, inclu-

sion of more than one class of antibiotic maximizes the likelihood of effective local antimicrobial activity.

Particular care should be taken with antibiotic selection for ALBC in patients who have failed previous spacer implantation because aminoglycoside-reported resistance rates for isolates from these patients were between 1.7 and 2.5 times those from patients with first-time periprosthetic infections [120].

At the time of component reimplantation similar to aseptic revision procedures, the use of cemented fixation after periprosthetic infections is routine for knees. Given the evidence supporting the benefits of ALBC in aseptic revision surgery, little controversy exists surrounding its use at the time of definitive component reimplantation after periprosthetic infection when cemented fixation is used. Available strategies for cemented fixation include using a commercially prepared low-dose ALBC or a custom-mixed ALBC prepared in the operating room.

Commercially available low-dose ALBC in the United States is FDA approved specifically (and only) for use in second-stage revision. Custom-mixed ALBC provides theoretical advantages by allowing for tailoring of antibiotics based on local resistance patterns and/or sensitivities of organisms isolated from the infected joint, while providing elution characteristics similar to commercially prepared cement [165]. However, little guidance is available from the literature in terms of outcomes with either approach.

If a custom-mixed ALBC is used, given the previously described evidence concerning antibiotic concentration and mechanical properties of cement, it would seem reasonable to limit concentrations to no more than 2 g per 40 g bag [120, 156]. Similarly, liquid antibiotics should be avoided [109]. Consideration should be given to using more than one antibiotic, given the synergistic effects and broader antimicrobial activity spectrum that can be achieved [163, 164]. To maximize mechanical characteristics, the antibiotics should be added to the cement powder first, followed by the addition of the monomer and vacuum mixing [161].

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Preoperative Management: Staphylococcus aureus Decolonisation

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26.1 Introduction

Prosthetic joint infection is a catastrophic and an under-recognised complication of knee replacement. Infection can be acute or chronic and represents the most common indication for revision surgery in the first 2 years following implantation [1]. In the UK, the overall revision rate for infection following primary knee replacement, both total and unicompartmental, is 0.92 (95%CI 0.90–0.95) revisions per 1000 prosthesis years with lower revision rates for unicompartmental including patellofemoral replacement as compared to total knee replacement [1]. Management of prosthetic joint infection (PJI) comes at a significant cost to both the patient and healthcare system and despite a better understanding of PJI, and the risk factors for it, the incidence is increasing, in part through improved diagnosis [2, 3].

Reducing the incidence of PJI requires a multi-faceted, multi-disciplinary, approach. Prior to surgery, risk factors for PJI need to be screened for, with modifiable risk factors optimised. Once risk

factors for PJI have been optimised, then a patient may be booked for surgery and pre-, intra- and post-operative infection prevention packages implemented to further minimise the risk of this devastating complication. Medical optimisation of the patient prior to surgery has been covered in the last chapter. This chapter will focus on preoperative screening and decolonisation of *S. aureus* prior to knee replacement. It will cover the epidemiology of *S. aureus* colonisation, incidence of invasive infection in colonised and non-colonised individuals, methods of decolonisation and the outcomes of decolonisation prior to surgery. Finally, we will review the current international guidelines on decolonisation and outline our local approach.

26.2 Epidemiology of Staphylococcus aureus Colonisation

Colonisation with *S. aureus* is common and can be persistent or intermittent [4]. Around 20% of the population are persistently colonised with relatively high, yet typically asymptomatic, bacterial loads, whilst many more individuals are intermittently colonised, typically with lower bacterial loads [5]. The Danish Twin Study has identified that in the older adults genetics exhibits only a modest influence on risk of persistent nasal *S. aureus* colonisation, whereas male gender, non-smoking status, chronic skin condition, and living

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or working on a farm are associated with an increased risk [6, 7].

The primary site for *S. aureus* colonisation is the anterior nares, and colonisation at this location is predictive for colonisation at extra-nasal sites including the skin, throat, perineum, vagina and gastrointestinal tract [4]. Of those colonised at any site with *S. aureus* around 50% are colonised in the anterior nares thus to assess for colonisation status in addition to the anterior nares swabbing of extra-nasal sites should also be considered [4].

Nasal colonisation with *S. aureus* has been reported in around a quarter of patients undergoing joint arthroplasty [8, 9]. Whilst the majority of these are methicillin-sensitive *S. aureus* (MSSA) between 1 and 4% are methicillin-resistant *S. aureus* (MRSA) [8, 9]. Whilst there is some evidence that the overall prevalence of colonisation with *S. aureus* is decreasing over time, it has been reported that there has been a relative increase in the prevalence of MRSA nasal carriage [10].

26.3 Incidence of Invasive Infection in Colonised and Non-colonised Individuals

Persistent colonisation with *S. aureus* is associated with an increased risk of invasive infection compared to that seen in non-colonised individuals [4]. This is particularly true in individuals with regular healthcare contact and in those patients with indwelling devices, such as orthopaedic surgical patients, where a 3- to 11-fold increase in risk has been reported [11]. In hospitalised non-surgical, non-bacteraemic, patients with known *S. aureus* nasal colonisation, it has been reported that the risk of developing a *S. aureus* bacteraemia is three times higher (RR 3.0, 95%CI 2.0–4.7) than non-colonised individuals [12]. In colonised individuals who develop a bacteraemia genotyping has demonstrated that around 80% the isolates from the blood are clonally identical to those from the anterior nares [13, 14]. Data indicate that, compared to MSSA, the risk of invasive infection with MRSA colonisation is substantially higher; however, it is unclear

whether this increased risk is related to the relative virulence of the organism or whether this is due to the fact that those colonised with MRSA represent patients with greater medical comorbidity, broader antibiotic exposure and with longer hospital stays [14–16].

S. aureus is one of the most commonly isolated organisms in prosthetic joint infection and nasal carriage of *S. aureus* has been reported to be one of the most important risk factors for developing surgical site infection with *S. aureus* with surgical site infections noted to be higher in colonised as compared to non-colonised individuals [8, 17]. Based on these observations and work in other areas of healthcare decolonisation of patients colonised with *S. aureus*, both MSSA and MRSA, presents a potential opportunity to reduce the burden of surgical site infection following joint replacement.

26.4 Methods of Decolonisation

Decolonisation can be targeted at both nasal and extra-nasal sites and may be delivered selectively to patients that are known to be colonised with either MSSA or MRSA (or those at high risk of colonisation), or may be delivered universally to all patients undergoing joint replacement.

Nasal decolonisation has traditionally been performed using Mupirocin 2% ointment applied topically two to three times a day for 5 days to the inner surface of each nostril. Mupirocin is a topical antibacterial agent active against *S. aureus*, including MRSA. It is also active against other *Staphylococci*, *Streptococci* and gram-negative organisms such as *E. coli* and *H. influenzae*. Recently however, Mupirocin resistant *S. aureus* has reported to be increasing with the prevalence of Mupirocin resistant MSSA reported to be around 8% and Mupirocin resistant MRSA 14% [18]. Based on this, in combination with the need for multiple days treatment, which risks non-compliance, other methods of decolonisation have been trialled, including photo-disinfection as well as a single topical nasal treatment with either povidone iodine or chlorhexidine gluconate with promising results [19–23].

Extra-nasal decolonisation can be performed by skin washing prior to surgery which is intended to reduce the bacterial load and intuitively should involve the whole body such that, in addition to the surgical site, known extra-nasal colonisation sites are targeted. Skin washing can be performed using an antibacterial or antiseptic soap, povidone iodine or chlorhexidine gluconate. There remains uncertainty as to the effectiveness of preoperative skin washing, the optimal time to start cleaning and optimum agent to use [24–27]. In the UK, where skin washing is performed, chlorhexidine gluconate is probably the most common agent used as it has activity against many pathogens including MRSA.

Whilst decolonisation is successful in the majority of patients, it has been reported that up to twenty percent remain colonised with *S. aureus* despite treatment [28, 29]. The reasons for this remain unclear and whether this represents failure of treatment, potentially due to Mupirocin resistance, or non-compliance is uncertain. As such the optimal decolonisation regime remains unclear and whether to re-assess patients for colonisation following treatment remains controversial and further research is needed to clarify these areas [18, 19].

26.5 Outcomes of Decolonisation Prior to Surgery

The effect of *S. aureus* decolonisation on the risk of surgical site infection following hip and knee replacement has been the subject of several studies (Table 26.1). A recent meta-analysis found that *S. aureus* decolonisation significantly reduced the risk of overall surgical site infection (both superficial and deep infections, Odds Ratio 0.43 95%CI 0.31–0.59) as well as both superficial (Odds Ratio 0.43 95%CI 0.25–0.73) and deep prosthetic joint infections (Odds Ratio 0.40 95%CI 0.21–0.77) following hip and knee replacement surgery [40]. Whilst these results indicate that screening and decolonisation of *S. aureus* is associated with a reduction in prosthetic joint infection, it is important to acknowledge that these results represent the outcomes

seen in predominantly retrospective and non-randomised studies and as such does not take into account other improvements in practice that have been implemented to reduce prosthetic joint infection.

26.6 Current Guidelines

World Health Organisation

- *Nasal Decontamination.*
 - Does not specify about universal screening and decolonisation.
 - Topical Mupirocin for decolonisation of MRSA and/or MSSA colonised patients.
- *Skin Decontamination.*
 - Shower or bathe the night prior to surgery.
 - Consider cleansing with chlorhexidine gluconate body wash.

International Consensus Meeting

- *Nasal Decontamination.*
 - No definitive recommendation on screening and decolonisation.
 - No consensus for decolonisation method for colonised patients.
- *Skin Decontamination.*
 - Whole-body skin cleansing with chlorhexidine gluconate.
 - Start at least the night prior to surgery.

National Institute for Clinical Excellence

- *Nasal Decontamination.*
 - Consider topical Mupirocin where *S. aureus* is a likely cause of a surgical site infection.
- *Skin Decontamination.*
 - Shower or bathe the night prior to or day of surgery using soap.
 - Consider cleansing with chlorhexidine gluconate body wash.

Our institution has adopted a universal decolonisation policy. Our local practice is that all patients undergo nasal swab to assess for MRSA

Table 26.1 Incidence of surgical site infection before and after implementation of a decolonisation programme for *S. aureus* in patients undergoing hip and knee replacement

| Study | Screening | Intervention | Incidence surgical site infection | |
|----------------------|-----------------------------|--|-----------------------------------|--------------------|
| | | | Intervention | Control |
| Rao 2011 [27] | Nasal swab | N: Mupirocin 5d S: CHG 5d | 1.32% (17/1285) | 2.70% (20/741) |
| Sankar 2005 [30] | Nasal and extra-nasal swabs | N: Mupirocin/povidone iodine/triclosan | 0% (0/231) | 0.61% (1/164) |
| Hacek 2008 [31] | Nasal swab | N: Mupirocin 5d S: Nil | 1.21% (11/912) | 2.60% (14/583) |
| Hadley 2010 [32] | Nasal swab | N: Mupirocin 5d S: CHG 5d | 1.28% (21/1644) | 1.45% (6/414) |
| Kim 2010 [8] | Nasal swab | N: Mupirocin 5d S: CHG 5d | 0.19% (13/7019) | 0.45% (24/5293) |
| Gottschalk 2014 [33] | Nasal swab | N: Mupirocin 7d S: CHG 1d | 1.9% (2/108) | 12.9% (9/70) |
| Baratz 2015 [29] | Nasal swab | N: Mupirocin 5d S: CHG 5d | 0.79% (27/3434) | 1.07% (33/3080) |
| McDonald 2015 [34] | Nasal swab | N: Mupirocin 5d S: CHG 5d | 0.66% (2/305) | 1.8% (11/596) |
| Sporer 2016 [35] | Nasal swab | N: Mupirocin 5d S: CHG 5d | 0.34% (33/9690) | 1.11% (16/1443) |
| Hofmann 2017 [36] | No screening | N: Mupirocin 2d S: Nil | 0.74% (4/538) | 2.02% (10/496) |
| Stanbough 2017 [37] | No screening | N: Mupirocin 5d S: CHG 5d | 0.22% (5/2205) | 0.76% (15/1981) |
| Jeans 2018 [38] | Nasal & extra-nasal swabs | N: Mupirocin 5d S: Octenisan 5d | 1.41% (131/9318) | 1.92% (69/3593) |
| Pelfort 2019 [39] | Nasal swab | N: Mupirocin 5d S: CHG 5d | 1.24% (5/403) | 4.25% (17/400) |

CHG chlorhexidine gluconate, N nasal, S skin

colonisation 2 weeks before surgery followed by, in all patients, independent of *Staphylococcus* colonisation (both MSSA and MRSA), decolonisation treatment. The rationale for assessing for MRSA colonisation is that contact precautions are taken with all inpatients with a history of MRSA colonisation with these patients nursed in a side room and placed last on the surgical list.

Staphylococcus decolonisation treatment consists of a topical nasal chlorhexidine with neomycin cream (Naseptin, Alliance Pharmaceuticals Limited) together with a chlorhexidine skin wash. The protocol differs based on whether patients are MRSA colonised or not. In MRSA colonised patients, the topical nasal chlorhexidine with neomycin cream is applied three times daily for 5 days prior to surgery and the chlorhexidine wash (HiBiScrub 4%, Molnlycke Health Care Ltd) is used for a similar duration twice a day. In patients with an allergy to either chlorhex-

idine or neomycin or who have a peanut or soya allergy, Mupirocin (Bactroban 2% cream, GlaxoSmithKline UK Ltd) is used. In patients having an allergy to the chlorhexidine wash, Octenisan antimicrobial wash (Schulke) is used. In patients not MRSA colonised, independent of known MSSA colonisation, topical nasal chlorhexidine with neomycin cream is applied three times daily for 1 day prior to surgery and the chlorhexidine wash (HiBiScrub 4%, Molnlycke Health Care Ltd) is used the night before and on the morning of surgery only. Patients are not routinely swabbed to confirm decolonisation.

Our centre is part of the Quality Improvement in Surgical Teams (QIST) Collaborative model. This ongoing cluster-randomised trial led by Northumbria Healthcare NHS Foundation Trust, in partnership with the British Orthopaedic Association hopes to improve the evidence for

screening for and decolonisation of *S. aureus* with the results informing practice in the UK and worldwide [41].

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Intraoperative Prevention Strategies to Prevent Infection

27

Christopher Vertullo

27.1 Introduction

In modern knee replacement, surgical site infection (SSI) is now the most common reason for revision despite it being a largely avoidable complication, hence the infection prevention chapters of this book are probably the most pertinent [1]. This chapter deals with an important part of the optimisation of surgery outcomes, intraoperative management, and as such most of the principles discussed apply to all aspects of clean elective orthopaedic surgery.

The burden of infection in total knee replacement is devastating for the patient as it occurs usually in the early expected implant survivorship, with younger patients most at risk. As a result, SSI rates are increasingly being used as a quality indicator and comparison benchmark within and across healthcare facilities [2]. A multimodal methodology is compulsory in PJI prevention, and a meticulous approach is important with each component.

For this narrative review, it is assumed the patient will arrive in the operating theatre complex with any immune-compromise reversed, nutrition and body mass index optimised, normothermic, non-anaemic, decolonised and/or screened preoperatively for colonisation with

potential skin pathogens such as resistant and sensitive *Staph aureus* and other *Staph* species, with optimal blood sugars if the patient is a diabetic and cessation of tobacco products well prior. The details of preoperative optimisation will have been dealt with in preceding chapters.

One on the difficulties in making recommendations around infection prevention best-practice is the quality of the data and the strength of recommendations based around that data. The GRADE categories [3] (Grading of Recommendations Assessment, Development and Evaluation) allow clinicians to grade the quality or certainty of evidence and hence the strength of recommendations. The World Health Organisation's Global Guidelines for the Prevention of Surgical Site Infection uses the GRADE system to guide surgeons as to optimal technique to prevent SSI and is recommended reading; however, many of the recommendations are not arthroplasty specific.

27.2 Antibiotic Prophylaxis

The timing and type of preoperative antibiotics deserves very careful consideration to achieve adequate plasma levels of an appropriate prophylactic antibiotic for the typical local infecting organisms, typically an intravenous first-generation cephalosporin unless the patient is MRSA colonised, prior to the incision being

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made and as part of a pre-surgical checklist to avoid errors of omission [4]. Prophylaxis with cefazolin has the lowest risk of later infection compared to alternative antibiotics such as vancomycin with a lower risk of adverse events, hence true allergy screening is recommended [5].

Optimal timing of surgical antibiotic prophylaxis remains uncertain; however, strong evidence suggest administration after incision or tourniquet application increases risk of SSI, as does inadequate levels at the time of surgical closure [2].

Hence, timing should be prior to the incision being made, with reference to the half-life of the chosen antibiotic. If the interval since the initial antibiotic dose is greater than 4 h, further dosing during the procedure is appropriate. For primary TKR, second dosing would be unusual. Underdosing, particularly with vancomycin and in obese individuals is common. Readers are encouraged to review the previous chapter on optimal antibiotic prophylaxis.

The role of intraosseous antibiotics under tourniquet for primary and revision TKR has promising preclinical data [5–7], with 5–20 times plasma levels achieved compared to IV prophylaxis, but routine use in primary TKR remains uncertain [8]. In meta-analysis, no evidence exists that extended course of prophylaxis reduces infection risk [9, 10]; however, the overall GRADE of evidence of the available literature is low (high risk of bias, high risk of publication bias and low precision).

27.3 The Operating Room

A carefully monitored, robust quality assured process following clearly defined national and international standards is required to maintain the Operating Room (OR) infection prevention measures for appropriate environmental cleaning and waste disposal. The entire operating room must be cleaned daily [11], starting from the least soiled and moving to the most soiled areas, using techniques that prevent mists or aerosol creation. After each procedure, soiled and high touch areas require cleaning and disinfecting. Instrument decontamination, cleaning, disinfection and sterilisation must be stan-

darised to national and international standards [12]. A team approach between surgeons and nurses is vital to ensure that all equipment is visibly clean and sterile prior to commencing, especially loan equipment, and that the operating field is sterile and clearly defined. Theatre traffic must be reduced as much as possible to minimise aerosol creation. The long-standing recommendations against the use of laminar flow remains, with no evidence in 12 observational trials of it being of no benefit with greater cost [13]; however, this data has been criticised for the biases inherent in observation registry data [14].

27.4 Hand Preparation

Hand preparation should be undertaken by either hand-scrubbing with antimicrobial aqueous soap or hand-rubbing with alcohol-based hand rub complying with international standards, after an initial hand clean-up with soap prior to entering the OR [15]. Low GRADE evidence suggests alcohol-based hand-rubbing may be superior to antimicrobial aqueous hand-scrubbing, while aqueous chlorhexidine may be superior to iodine aqueous solutions if scrubbing [2].

27.5 Gloves

In a recent meta-analysis, evidence supporting reduced SSI with multiple glove changing remained weak [16], hence authors recommended gloves should be changed after draping, hourly, with any visible penetration and before handling implants [16].

27.6 Surgical Site Preparation

Prior to entering the OR, the surgical site should have been washed with either plain soap, chlorhexidine gluconate soap or chlorhexidine gluconate impregnated cloths, prior to the surgery [17]. It remains unclear of the optimal washing period pre-surgery; however, consensus currently suggests 3 days.

Moderate evidence suggest best-practice site preparation involves never shaving the skin as it increases the infection risk, keeping clipping to the minimum required and only using as a single use device outside the operating theatre [18]. The optimum timing of hair clipping prior to the surgery remains uncertain.

The WHO strongly recommends alcohol-based chlorhexidine gluconate preparation should be utilised with less risk of SSI when compared to alcohol-based povidone-iodine and aqueous solutions with alcoholic and aqueous povidone-iodine having similar SSI rates for all surgical cases based on meta-analysis [2]. The recent cluster-randomised ACAISA trial compared chlorhexidine alcohol versus iodine alcohol for surgical site skin preparation in an elective arthroplasty, finding results contrary to the WHO, with no difference for the primary outcome measure of SSI, but lower prosthetic joint infections. The differences may be attributable the ACASIA being arthroplasty specific. At this stage, it is recommended that prep solutions all contain 70% alcohol and either povidone-iodine or chlorhexidine until more data is available. It is important to recognise that alcoholic preparations are flammable and preparation technique should avoid saturated drapes and or pooling. A recent trial by Morrison et al. [19] suggested a repeated alcoholic iodine preparation just prior to iodine-impregnated incision drape application had a lower SSI than a single preparation; however, it should be recognised that a higher risk of intraoperative fire can occur with this technique [20]. There is no evidence that film-forming cyanoacrylate sealants such as InteguSeal reduce infection rates [21].

27.7 Intra-Articular Dilute Povidone-Iodine Lavage

The benefit of a dilute 500 ml, 0.35% povidone-iodine lavage for 3 min prior to closure in primary arthroplasty has mixed results with observational evidence [22–24], overall suggesting no benefit. In revision total knee arthroplasty, a recent randomised clinical trial [25] of 478 patients undergoing aseptic revision TKA and

THA had a lower infection rate with the dilute betadine lavage. The WHO recommends dilute povidone-iodine lavage for clean wounds [2].

27.8 Drapes and Gowns

Sterile impermeable reusable or single use drapes and gowns should be used, with no difference in SSI rates between the two [2, 26]. Conversely, despite theoretical claims of locking in dermal bacteria, plastic adhesive drapes do not reduce the risk of SSI [27] and may cause patient harm through allergy and skin damage. No evidence exists regarding changing gowns or drapes intraoperatively.

27.9 Perioperative Hyperoxygenation

The benefits of 80% fraction of inspired oxygen intraoperatively and post-operatively remain controversial. The WHO strongly recommends perioperative hyperoxygenation to those undergoing general anaesthesia with endotracheal intubation [2]; however, other recent meta-analysis question this recommendation, stating possibly increased mortality with no decrease in SSI [28]. No arthroplasty literature exists in this area.

27.10 Normothermia and Normovolemia Maintenance

Anaesthesia impairs patients' abilities to maintain body temperature, and heat loss is increased due to cool intravenous fluids and irrigation fluids. The WHO recommends active perioperative patient warming based on moderate evidence of reduced SSI in non-arthroplasty literature [2]. While many methods exist for active warming [29], forced air warming may be counterproductive due to aerosol creation.

With the use of tranexamic acid, significant hypovolemia in primary TKR would be atypical; however, in revision surgery normovolaemia needs to be maintained as some evidence in the

non-arthroplasty literature suggests lower SSI with a goal directed-fluid therapy [2].

27.11 Dressings, Drains, Sutures and Closure

A variety of antimicrobial sutures are currently available, with the most closely studied being Triclosan (5-chloro-2-(2,4-dichlorophenoxy) phenol), a broad-spectrum bactericidal agent used in a variety of applications including household soaps, which at a higher concentration is bactericidal, and at lower concentration is bacteriostatic [2]. The benefits of Triclosan sutures in arthroplasty remain uncertain, with the WHO recommending their use for all types of surgery; however, two recent clinical trials both failed to show a reduced risk of SSI [30, 31] with them. No data supports changing instruments for closure [2]. While the evidence is fragile, staples have a higher risk of superficial SSI [32, 33] than suture closure.

A recent Cochrane review suggested negative pressure wound therapy reduced the risk of SSI by approximately 33% with moderate evidence [34]; however, its routine use in knee arthroplasty remains uncertain compared to situations with questionable skin integrity. In recent Cochrane review [35], there was no evidence supporting the use of advanced dressing such as hydrocolloid, hydroactive, silver-containing (metallic or ionic) and polyhexamethylene biguanide (PHMB) dressings compared to standard dry absorbant dressings.

The role of surgical drains increasing SSI risk remains uncertain, with some authors finding increased risk [36] with their use, and other describing decreased risk [37]. If drains are used, no evidence supports lower SSI risk with early removal of the drain [2].

27.12 Surgical Hoods and Body Exhaust Suits

When surgical hoods are discussed, it is important to recognise between the two different systems that are available, the original cumbersome negative pressure body exhaust suits (BES) intro-

duced by Charnley, and later, the more portable positive pressure surgical helmet systems (SHS) [38]. BES that are characterised by bulky aspiration tubing and a negative intra-suit pressure, have clinical evidence in meta-analysis supporting their ability to reduce deep infection compared to standard surgical gowns [38].

SHS have been described as a “personal protection device”, are typically characterised by a fan on a helmet with a positive pressure within the suit, blowing air across the surgeon’s face and neck. In contrast to the BES, SHS have not been shown to reduce SSI [38] or wound contamination [39, 40] when compared to standard surgical gowns and in registry studies may increase rates [41]. While taping the gown glove interface does not alter contamination rate [40], authors have suggested using SHS solely as personal protective equipment and wearing a balaclava underneath to reduce bacterial load from the wearer’s face and neck [39].

27.13 Prosthesis Design and Antibiotic-Loaded Bone Cement

Despite some controversy around the benefits of antibiotic-loaded bone cement [42], a recent meta-analysis of nine randomised clinical trials using Cochrane methodology and prosthetic joint infection as the primary outcome measure suggested that it did reduce the risk of SSI in TKR [43]. The optimum antibiotics and dosing remain uncertain, with the two most common antibiotics utilised being vancomycin and aminoglycosides such as tobramycin and gentamycin. It should be noted that many randomised clinical trials in this area are underpowered to adequately investigate the primary outcome measure of prosthetic joint infection and observational studies and registry analysis are at risk of selection bias, where high risk patients receive antibiotic-loaded bone cement at higher rates than low risk patients. Future arthroplasty registry imbedded cluster-randomised trials would be a low-cost solution to providing more robust data in this area.

Recently, the interaction of prosthesis design and infection risk has been investigated in registry studies, with up to 100% greater revision for infection risk with posterior stabilised TKR compared to cruciate retaining and over 25% higher for non-cross linked polyethylene [1, 44]. As observational registry studies are at a risk of confounder bias, registry imbedded cluster-randomised trials would be beneficial.

27.14 Blood Management

The interaction of chemoprophylaxis with infection risk remains uncertain and is a complex area with competing risks of mortality and morbidity. Readers are encouraged to refer to the chapter dealing with thromboprophylaxis and haematoma. Some forms of chemoprophylaxis, particularly rivaroxaban [45], have been reported to increase infection rate in smaller observation studies; however, larger registry studies have not shown higher infection risk with direct oral anticoagulants when compared to aspirin [46]. Tranexamic acid has been reported to reduced infection risk [47, 48] in observational studies.

27.15 Post-operative Care

General principles of appropriate wound care are important to follow. Additionally, evidence suggests that post-operative recovery in either specialist elective surgery hospitals or “ring-fenced” elective areas within a non-elective hospital reduces infection risk [49].

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Dental Procedures After Joint Replacement

28

Kohei Nishitani and Shuichi Matsuda

28.1 Introduction

Prosthetic joint infection (PJI) is one of the most devastating problems following arthroplasty, and thus, orthopedic surgeons want to avoid it at all costs. Many orthopedic surgeons may consider that dental procedures may cause bacteremia, which is managed with antibiotics. Thus, antibiotics are better to be used during dental procedures in patients who have joint prostheses. In this chapter, we first describe the relationship between dental procedures and bacteremia. Next, we describe whether the dental procedure is a risk factor of PJI and whether antibiotic prophylaxis effectively inhibits PJI. We then provide an overview of the recent guidelines for dental procedures and prophylaxis for patients with a joint prosthesis. Finally, we discuss how this issue can be managed in practical setting.

28.2 Dental Procedure and Bacteremia

The oral cavity is one of the most common bacterial sites in the human body. The human oral microbiome comprises more than 2000 bacterial

taxa, including a large number of pathogens involved in the periodontal, respiratory, cardiovascular, and systemic diseases [1]. These bacteria can enter the bloodstream by dental procedures. Surprisingly, even daily oral care activities such as brushing and flossing as well as clinical procedures, such as scaling, planing, and oral surgical procedures may cause temporary bacteremia. For patients with healthy oral conditions, brushing is usually safe without causing bacteremia [2]. However, in patients with oral problems such as periodontitis, brushing is associated with an incidence of bacteremia in about 10 and 20% of patients [3–5], and after scaling and planing, in 13–75% of patients [3, 4, 6, 7]. More aggressive procedures, such as root-canal procedures and tooth extraction, cause bacteremia with a higher probability of 30–80% [8–11]. As many reports describe, non-invasive and invasive dental procedures have a risk of bacteremia, especially in patients who have poor oral conditions.

Antibiotics and topical antimicrobial prophylaxis effectively reduce the bacteremia caused by dental procedures. For example, Lockhart et al. randomized 290 patients into toothbrushing, single-tooth extraction with amoxicillin prophylaxis, or single-tooth extraction with identical placebo groups [10], and showed that the cumulative incidence of bacteria was 23%, 33%, and 60% for tooth brushing, extraction-oral amoxicillin, and extraction-placebo groups, respectively ($P < 0.0001$). Dios PD et al. randomized 220

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patients for dental extraction into four groups: a control group, an oral amoxicillin group, an oral clindamycin group, and an oral moxifloxacin group, and venous blood samples were collected from each patient at baseline, and various time points after dental extractions [12]. Results indicate the effectiveness of amoxicillin and moxifloxacin, showing 96%, 46%, 85%, and 57% bacteremia incidence rates at 30 s, and 20%, 4%, 22%, and 7% bacteremia incidence rates at 1 h, in control, amoxicillin, clindamycin, and moxifloxacin groups, respectively. Simple tooth extraction resulted in the second-highest median incidence of bacteremia and the highest median prevalence of bacteremia for all procedures, and the effectiveness of antibiotic prophylaxis has also been reported in numerous literature [13].

28.3 Dental Procedure and Prosthetic Joint Infection

There are many reports of PJI associated with dental procedures. It has been estimated that 6–13% of PJI cases are due to oral flora [14]. In the current review by Slullitel et al., nine studies focused on PJI diagnosis after dental procedures, in which total infections associated with a dental procedure ranged from 0 to 15.9% [15]. For example, Barbari et al. reported 35/339 (10.3%) PJI-related dental work [16]. Their report included organisms of potential oral or dental origin, such as *beta-hemolytic* ($n = 13$) and Viridans group *streptococci* ($n = 11$), *Peptostreptococcus* ($n = 5$), *Streptococcus*-like organisms ($n = 2$), *Abiotrophia/Granulicatella* species ($n = 2$), *Gemella* species ($n = 1$) and *Actinomyces* species ($n = 1$). In other reports, Uçkay et al. reported 3/71 (4.2%) dental works that were related to PJI caused by *Streptococcus oralis* ($n = 1$), *Streptococcus milleri* ($n = 1$), *Staphylococcus aureus* ($n = 1$) [17], and LaPorte et al. reported 3/52 (5.8%) PJI caused by *Streptococcus viridans* ($n = 2$), and *Peptostreptococcus* ($n = 1$) [18]. In recent years, there have been several case reports that show PJI in dental flora due to dental procedures [19–22]. Although many studies show the relationship

between organisms in dental flora and hematogenous PJI, a case-control study by Skaar et al. found no association between dental procedures and PJI [23]. In their report, 42 cases of PJI and 126 matched controls without PJI were analyzed. They reported that control participants were more likely to have undergone invasive dental procedures than case participants, although this result was not significant (hazard ratio = 0.78 [95% confidence interval [CI], 0.18–3.39]; odds ratio [OR] = 0.56 [95% CI, 0.18–1.74]). In a population-based cohort study using the Taiwan National Health Insurance Research Database, a dental cohort comprised of 57,066 patients who received dental treatment was compared with a 1:1 matched nondental cohort [24]. In their report, PJI occurred in 328 patients (0.57%) in the dental cohort and 348 patients (0.61%) in the nondental cohort, with no between-cohort difference in the 1-year cumulative incidence (0.6% in both, $P = 0.3$).

Although literatures describe that dental procedures cause bacteremia, and there are many reports of PJI by dental flora, there is much controversy regarding routine prophylaxis in patients with a history of joint replacement undergoing dental procedures. In a study of 1000 patients with 1112 joint replacements, the patients were advised not to take prophylactic antibiotics before any dental or surgical procedures. In this population, 284 infections developed in various organs, including the oral cavity, but none of these patients developed hematogenous infections [25]. In a case-control study by Barbari et al., 339 patients with total hip or knee infection and 339 controls undergoing total hip or knee replacement without infection during the same period were compared. Dental procedures are distinguished as high-risk (dental hygiene, mouth surgery, periodontal treatment, dental extraction, and therapy for dental abscess) and low-risk (restorative dentistry, dental filing, endodontic treatment, and fluoride treatment) dental procedures. As a result, they reported no increased risk of prosthetic hip or knee infection for patients who were undergoing high-risk (adjusted OR, 0.8; 95% CI, 0.4–1.6) or low-risk dental procedures (adjusted OR, 0.6; 95% CI, 0.4–1.1). Additionally, antibiotic prophylaxis in high-risk

(adjusted OR, 0.9; 95% CI, 0.5–1.6) or low-risk (adjusted OR, 1.2; 95% CI, 0.7–2.2) dental procedures did not decrease the risk of subsequent total hip or knee infection [16]. In a population-based cohort study using the Taiwan National Health Insurance Research Database, the dental cohort was further distinguished as an antibiotic ($n = 6513$) and nonantibiotic subcohorts ($n = 6513$) [24]. PJI occurred in 13 patients (0.2%) in the antibiotic subcohort and in 12 patients (0.18%) in the nonantibiotic subcohorts ($P = 0.8$). Multivariate-adjusted analyses confirmed that there were no association between PJI incidence and prophylactic antibiotics (adjusted hazard ratio, 1.03; 95% CI: 0.47–2.27).

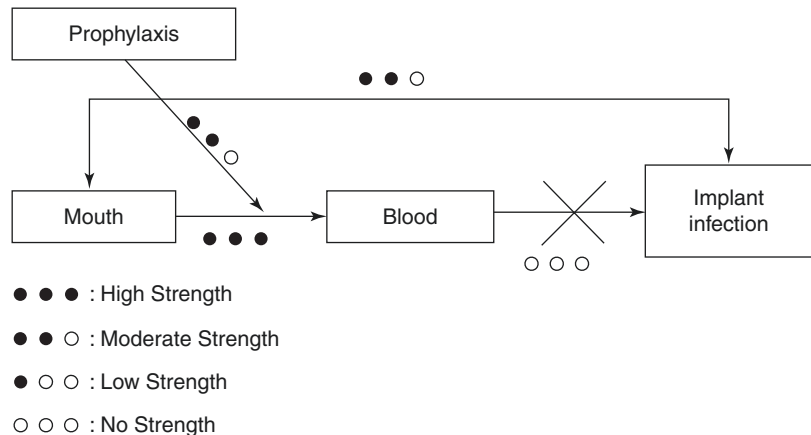
The time after arthroplasty may be a confounder for the risk of dental procedures. In an animal study, bacteremia caused hematogenous infection in a rabbit cemented stainless-steel implant in the early period; however, the rabbit became resistant to infection 3 weeks postoperatively [26]. In clinical studies, the timing after arthroplasty was related to hematogenous infection, which was higher during the first 2 years after arthroplasty [27]. A possible explanation for this might be that active local inflammation and osseointegration activity around components may lead to a higher blood flow to the prosthetic joint and the potential for organism seeding onto the implant surface [14, 15]. The number of bacteria in the blood flow also affected PJI. Zimmerli et al. found that a 10^2 *Staphylococcus aureus* colony forming unit (CFU) inoculum injected

into the region of the foreign material was required to induce infection in >95% of guinea pigs. Another report found that 10^4 – 10^6 CFU intravenous *Staphylococcus aureus* injections were required to cause endocarditis in rabbit [28, 29]. Although the dose-effect of the inoculation was evident in animal models, the magnitude of bacteremia required to cause clinically significant bacterial disease in humans is unknown.

28.4 Current Guidelines for Dental Procedures for Patients with Total Joint Prosthesis

Professional guidelines have provided evidence-based approaches regarding the relationship between oral procedures and PJI and antibiotic prophylaxis effectiveness. In the current guidelines published within the last 10 years, the American Academy of Orthopaedic Surgeons (AAOS) and the American Dental Association (ADA) released a new guideline in 2012. In this guideline, a vigorous literature review provides an overview of the evidence to explain the proposed association between dental procedures and orthopedic implant infection (Fig. 28.1) [13]. The guideline shows strong evidence between oral procedure and the occurrence of bacteremia, and moderate evidence strength between oral organisms and PJI. However, no evidence has described that oral procedures cause PJI via bacteremia.

Fig. 28.1 The strength of evidences among oral bacteria, bacteremia, and PJI. (From ADA and AAOS guideline in 2012 [13] with modification)



With this accumulation of evidence, this guideline provided three recommendations [13]: (1) The practitioner might consider discontinuing the practice of routinely prescribing prophylactic antibiotics for patients with hip and knee prosthetic joint implants undergoing dental procedures (Grade of Recommendation: Limited); (2) We are unable to recommend for or against the use of topical oral antimicrobials in patients with prosthetic joint implants or other orthopedic implants undergoing dental procedures (Grade of Recommendation: Inconclusive); (3) In the absence of reliable evidence linking poor oral health to prosthetic joint infection, it is the opinion of the workgroup that patients with prosthetic joint implants or other orthopedic implants maintain appropriate oral hygiene (Grade of Recommendation: Consensus). The 2014 ADA guidelines followed the AAOS and ADA 2012 guidelines. They conclude that evidence fails to demonstrate an association between dental procedures and PJI or any effectiveness for antibiotic prophylaxis, with a recommendation that in general, for patients with prosthetic joint implants, prophylactic antibiotics are not recommended prior to dental procedures to prevent PJI [30]. Although the two abovementioned guidelines did not recommend routinely using antibiotics for dental procedures, they also described that treatment decisions should be made in light of all circumstances presented by the patient. Treatments and procedures applicable to individual patient rely on mutual communication between patient, physician, dentist, and other healthcare practitioners [13]. This suggests that orthopedic surgeons, dentists, and patients should consider individual risk to decide the use of antibiotics for dental procedures.

In guidelines from an orthopedic surgeon's viewpoint, the first International Consensus Meeting (ICM) on Periprosthetic Joint Infection had some consensus statement on dental procedures for patients with PJI [31]. For the question "Should a patient with total joint arthroplasty (TJA) be given routine dental antibiotic prophylaxis?", the Consensus states "The use of dental antibiotic prophylaxis in patients with TJA should be individualized based on patient risk factors and the

complexity of the dental procedure to be performed." with a strong consensus (agree rate: 81%). Although there is no consensus that antibiotic prophylaxis before dental work can reduce PJI, most PJIs occur within the first 2 years postoperatively [18, 32]. Thus, this guideline concludes that using antibiotic prophylaxis for dental procedures after TJA to decrease the risk of bacteremia following dental procedures is justifiable to decrease the risk of sustaining a PJI within the first 2 years postoperatively [31]. However, in the second ICM consensus statement, for the question "What is the role of prophylactic antibiotics for invasive procedures (dental, gastrointestinal, urologic, etc.) in the presence of an arthroplasty to prevent subsequent PJI?", the recommendation stated that there is no role for routine prophylactic antibiotic administration prior to dental procedures (Level of Evidence: Limited, Weak Consensus) [33]. In the second ICM consensus statement, they also recommend that non-urgent invasive dental procedures, if possible, be delayed until the osseointegration of uncemented components is complete [33].

28.5 Practical Usage of Antibiotics for Patients with Prosthesis

Antibiotic prophylaxis against dental procedures should be addressed on a patient-by-patient basis, considering individual risk factors and the risk of the dental procedure. Who would be effectively protected by antibiotics before the dental procedure? Patients who were considered to be given antibiotics in the past guideline are likely to benefit most from antibiotics. In the advisory statement of ADA and AAOS in 2009 and first ICM consensus statement, high-risk patients included immunocompromised patients, patients with inflammatory arthropathy such as rheumatoid arthritis and systemic lupus erythematosus, immunosuppressed patients; patients with *Human Immunodeficiency Virus*, patients with previous joint infection, hemophilia, malnourishment, type 1 diabetes, or malignancy, and patients with mega prosthesis [34–38]. The following factors are determined by a dental care provider: high gingival score and gingival

index, high plaque score and plaque index, gum probing depth, and periodontitis [5, 39, 40]. Although the guidelines did not list any special situations, clinicians may consider antibiotic prophylaxis despite the lack of scientific evidence. To help clinicians make decisions regarding antibiotic prophylaxis for dental procedures, AAOS and ADA developed a tool for the appropriate use criteria (AUC) of antibiotics usage to assist orthopedic surgeons and dentists to aid their patients [41]. Clinicians can utilize the aforementioned AUC for the “Management of Patients with Orthopaedic Implants Undergoing Dental Procedures (2016)” in the ORTHO GUIDELINES website (<http://www.orthoguidelines.org/go/auc/>). By choosing an appropriate indication profile for planned dental procedure, immunocompromised status, diabetic glycemic control, history of periprosthetic or deep PJI that required an operation, and timing since joint replacement procedure, the clinicians can obtain recommendations if antibiotic prophylaxis is “rarely appropriate,” “maybe appropriate,” or is “appropriate.”

If prophylaxis was performed, what kind of antibiotics were suitable for the prophylaxis of dental procedures to reduce the risk of PJI? Because there is no evidence for the prophylaxis of PJI after dental procedures, antibiotics that effectively reduce bacteremia after dental procedures would be the clue for drug selection. The advisory statement of ADA and AAOS in 2003 provides the suggested antibiotic prophylaxis regimens, classified by patient type [42]. The suggested regimen is as follows: for patients who are not allergic to penicillin, 2 g of oral cephalexin, cephadrine, or amoxicillin 1 h prior to the dental procedure; for patients not allergic to penicillin and unable to take oral medications, 1 g of cefazolin or 2 g of ampicillin intramuscularly or intravenously 1 h prior to the dental procedure; for patients allergic to penicillin, 600 mg of clindamycin orally 1 h prior to the dental procedure; and for patients allergic to penicillin and unable to take oral medications, 600 mg of clindamycin intravenously 1 h prior to the dental procedure. No second doses are recommended for any of these dosing regimens. The first ICM consensus also referred to several antibiotics to

reduce the burden of bacteria released during dental procedures [31], with 2 g of amoxicillin, recommended to be administered a maximum of 1 h prior to the procedure [10, 12, 43, 44]. In the current concept review in 2014, Young et al. reported a relative decrease in bacteremia was decreased by antibiotics at 5 min postoperatively [14]. They also found a favorable bacterial reduction in oral amoxicillin (OR, 0.135; 95% CI, 0.097–0.187) and oral clindamycin (OR, 0.407; 95% CI, 0.223–0.725) vs. no antibiotic control.

Finally, physicians must be aware that the administration of antibiotics to individuals is not without its problems and may result in drug-related adverse effects such as swelling or itching, *C. difficile* colitis, and even more severe adverse effects such as anaphylaxis. The potential to cause the emergence of drug-resistant organisms is also considered. Young et al. provided an interesting analysis of the risk and benefit of antibiotic usage for dental procedures [14]. In their analyses, using 6–13% of PJI cases as being due to oral flora in approximately 140,000 PJIs out of 7,000,000 people with prosthetic joints, 8400 to 18,200 PJIs were secondary to oral bacteremia, and amoxicillin administered before the dental procedure may decrease PJI to 1746 and 3784 cases, respectively. Given a 2% antibiotic-related side effect incidence rate, if all 7,000,000 people with prosthetic joints had antibiotics, 140,000 side effects might have occurred. In their comment, 37–80 patients (140,000/[1746–3784]) would experience an adverse antibiotic effect for every PJI. The seriousness of PJI and antibiotic-related side effects cannot be the same. Therefore, this may be a somewhat radical calculation, but it is agreeable that the risk of abusing antibiotics cannot be overlooked. Therefore, carefully evaluating the patient-by-patient risk-benefit for antibiotic administration is mandatory for the dentist and orthopedic surgeon.

28.6 Conclusions

In conclusion, dental procedures cause bacteremia, and antibiotic prophylaxis reduces it. However, there is limited evidence that shows dental procedures are associated with PJI or antibiotic

prophylaxis before dental procedures reduce PJI. Thus, recent guidelines do not recommend the routine use of antibiotics for dental procedures. Hence, it is important for physicians and dentists to consider the individual patient risk and procedural risk to decide whether to use antibiotics before dental procedures, considering the side effects and potential drug resistance. The risk and benefit analysis of using antibiotics is better shared by the patient, dentist, and orthopedic surgeon.

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Hematoma and Thromboprophylaxis

29

Shinichiro Nakamura

29.1 Hematoma

Postoperative hematoma can be a reason for surgical site infection (SSI). The use of closed drainage has been advocated because there is less infection, postoperative pain, and swelling as well as better healing of the soft tissues and quicker mobilization of the extremities [1–3]. Kim et al. conducted a prospective study of 69 patients who had a primary simultaneous bilateral total knee arthroplasty (TKA) to assess the effect of postoperative suction drainage on infection and wound healing. The knees that had no drains had a higher incidence of drainage from the wound, had soaked dressings requiring dressing reinforcements, and had more ecchymosis and erythema around the wound. Although the incidence of infection in the two groups is not statistically different, the development of infection in two knees in which drains were not used suggests that suction drainage may reduce deep infection [1].

Recently conflicting results have been reported, and an increasing number of studies

have demonstrated no benefit to the use of closed drainage [4–6]. Li et al. conducted a prospective randomized, controlled trial in 100 patients to compare the postoperative use of wound drains with the use of no drains in patients who underwent unilateral primary TKA. The group treated without a drain needed comparatively less blood transfused. Differences in wound infection, incidence of deep vein thrombosis, and range of motion were not statistically significant [4].

Several systemic review and meta-analysis studies to assess the benefit and drawback of closed drainage were published [7, 8]. Si et al. reported that no significant differences in infection rate or blood loss were found between the closed drainage and nondrainage TKAs, and there was also no significant difference in hematoma formation, deep venous thrombosis, postoperative VAS score, or range of motion between the two groups [7]. Zhang et al. also reported no significant difference in total blood loss, hemoglobin drop, superficial wound infection, prosthetic joint infection, formation of deep vein thrombosis, duration of hospital stay, and range of movement [8].

The incidence of postoperative hematoma is decreasing with the use of local infiltration of anesthesia and tranexamic acid. The usage of a closed drainage in TKA will decrease in the future.

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29.2 Timing of Drain Removal

Closed suction drainage of wounds has been well established as a principle of management following joint arthroplasty, although the efficacy of this practice has been questioned recently. Drinkwater et al. conducted a prospective clinical trial, in which surgeons were asked to randomly allocate the time that the drains were left in situ after surgery [9]. The likelihood of bacterial colonization increased while wound drainage decreased with time. The proportion of drains contaminated after 24 h was significantly higher. The authors suggested that the optimal time to remove drains is 24 h after total joint arthroplasty. As fast-track program has been implemented in TKA, whether drainage tube could be removed early, and the ideal timing for removal after fast-track primary TKA has been a new topic. Zhang et al. evaluated the safety and feasibility of early removal of drainage tube in a prospective cohort study. A wound drainage tube was indwelled for 6, 12, and 18 h. There was no statistically significant difference in the volume of total and hidden blood loss among three groups, but as the time of drainage prolonged, total volume of drainage and dominant blood loss increased gradually. Early removal of wound drainage tube could drain the hematocoele and reduce the risk of infection, and it doesn't increase the sense of pain, inflammatory reaction, limb swelling, and total blood loss. It's safe and feasible to remove the drainage tube within 6–12 h after fast-track primary TKA [10].

There is no direct evidence to suggest that the use of surgical drains leads to an increase in the rate of subsequent SSI. The recommended time to remove drains is within 24 h because of higher contamination. The use of surgical drains leads to a higher volume of blood loss and an increased need for allogenic blood transfusion, which may indirectly increase the rate of SSI.

29.3 Tranexamic Acid (TXA)

Tranexamic acid (TXA) is an antifibrinolytic agent, which has become an integral component in postoperative blood management in orthope-

dic surgery [11, 12]. The published literature on TXA has dramatically expanded over the past several years. In a meta-analysis study, topical, intravenous (IV), and oral TXA formulations were all superior to placebo in terms of decreasing blood loss and risk of transfusion, and strong evidence supports the efficacy of TXA to decrease blood loss and the risk of transfusion after primary TKA [11]. Relatively large reductions in the mean difference of blood loss between 225 and 331 mL were observed in favor of TXA treatments compared with placebo.

Preoperative anemia is associated with development of subsequent postoperative periprosthetic infection, medical complication, and mortality [13, 14]. Greenky et al. defined anemia as hemoglobin 12 g/dL in women and hemoglobin 13 g/dL in men. An allogenic transfusion was received in 44% of anemic patients, compared with only 13.4% of nonanemic patients. Postoperative periprosthetic infection occurred more frequently in anemic patients at an incidence of 4.3% in anemic patients compared with 2% in nonanemic patients. Allogenic blood transfusions are also associated with infection and reoperation [15, 16]. Newman et al. showed that the rate of reoperations for suspected infection was higher among patients with perioperative allogenic exposure (1.67%) as compared with all others (0.72%, $p = 0.014$) [16]. Friedman et al. investigated the types of postoperative infection including lower or upper respiratory tract and lung infection, bone and joint infection, wound inflammation or infection, urinary tract infection, and other infections. The rates of any infection, lower or upper respiratory tract and lung infection, and wound inflammation or infection were significantly increased in patients receiving allogenic blood transfusion [15].

The direct effect of TXA on SSI has been unclear so far. Lacko aimed to analyze the effect of intravenous administration of TXA on reducing the risk of revision for acute and delayed periprosthetic joint infection. Cumulative revision rate of TKA was significantly lower in the TXA group (0.13% vs. 1.08%, $p = 0.043$). The use of TXA was shown as the significant protective factor [odds ratio (OR): 0.109; 95% confidence

interval (CI): 0.0128–0.929; $p = 0.043$] [17]. Further research should be conducted to examine whether TXA is effective for SSI. The administration of TXA potentially reduces the incidence of SSI by reducing postoperative anemia and the need for allogenic blood transfusion.

29.4 Thromboprophylaxis

29.4.1 Prevention of Venous Thromboembolism (VTE)

VTE is a serious complication following major orthopedic surgery. Several guidelines are suggested to reduce postoperative pulmonary embolism and deep vein thrombosis. The American Academy of Orthopaedic Surgeons (AAOS) guidelines on preventing venous thromboembolic disease in patients undergoing elective hip and knee arthroplasty, suggests the use of pharmacologic agents and/or mechanical compressive devices for the prevention of venous thromboembolism in patients undergoing elective hip or knee arthroplasty, and who are not at elevated risk beyond that of the surgery itself for venous thromboembolism or bleeding. In the absence of reliable evidence, it is the opinion of this work group that patients undergoing elective hip or knee arthroplasty, and who have also had a previous venous thromboembolism, receive pharmacologic prophylaxis and mechanical compressive devices [18].

In antithrombotic therapy and prevention of thrombosis, 9th ed: American College of Chest Physicians (ACCP) Evidence-Based Clinical Practice Guidelines, strategies for thromboprophylaxis after major orthopedic surgery are included. In patients undergoing TKA, use of one of the following is recommended for a minimum of 10–14 days rather than no antithrombotic prophylaxis: low-molecular-weight heparin (LMWH), fondaparinux, apixaban, dabigatran, rivaroxaban, low-dose unfractionated heparin (LDUH), adjusted-dose vitamin K antagonist (VKA), aspirin (all Grade 1B), or an intermittent pneumatic compression device (IPCD) (Grade 1C) [19].

The incidence of VTE after total hip arthroplasty (THA) or TKA is reduced by the use of

thromboprophylaxis. However, current evidence is unclear about which prophylactic strategy (or strategies) is/are optimal or suboptimal. Therefore, it is unable to recommend for or against specific prophylactics in these patients. In the absence of reliable evidence about how long to employ these prophylactic strategies, patients and physicians discuss the duration of prophylaxis.

29.5 Complication of Bleeding

The incidence of VTE after THA or TKA is reduced due to thromboprophylaxis. However, these medications have a number of limitations that impede their use, including increased bleeding risk. The potential for bleeding secondary to prophylaxis has been associated with prolonged recovery, infections, wound failure, and readmission. Therefore, the risk vs. benefit is a primary consideration when a provider chooses VTE prophylaxis in these patients.

Concerning bleeding, Lindquist et al. compared postoperative bleeding rates in patients receiving aspirin to patients who received enoxaparin or rivaroxaban after undergoing elective total joint arthroplasty [20]. Those who received aspirin or enoxaparin were less likely to experience any bleeding compared to those patients who received rivaroxaban ($P < 0.05$). There was also a lower rate of major bleeding in these groups. Suen et al. conducted systematic review of the surgical site bleeding complications of thromboprophylactic agents. LMWH increased the risk of surgical site bleeding compared with control, warfarin, and dabigatran and trended toward an increased risk compared with apixaban. The risk of surgical site bleeding was similar with LMWH and rivaroxaban [21].

29.6 Complication of Wound Complication

Wound-related complications following arthroplasty can cause restricted joint movement, reoperation, infection, and revision arthroplasty. Jameson evaluated the surgically relevant com-

plications of using either rivaroxaban or an LMWH as thromboprophylaxis, based on prospectively collected national data. The rivaroxaban group had a higher wound complication rate and a lower deep venous thrombosis rate; there were no differences in symptomatic pulmonary embolism or all-cause mortality [22]. Bloch et al. reported the impact of dabigatran on wound leakage. The use of dabigatran led to a significant increase in postoperative wound leakage (20% with dabigatran, 5% with a multimodal regimen; $p < 0.001$), which also resulted in an increased duration of hospital stay [23].

Garfinkel et al. conducted a retrospective review of a prospectively collected total joint arthroplasty registry to examine whether the choice of aspirin vs. factor Xa inhibitors for VTE prophylaxis is associated with differences in the rates of bleeding and wound complications in the early postoperative period. Six of 32 patients (18.7%) in the Xa inhibitor group had a postoperative bleeding/wound complication (4 delayed healing/blistering, 1 hematoma/excessive ecchymosis, and 1 readmission for cellulitis). There were no bleeding/wound complications in the aspirin group ($P < 0.03$). Factor Xa inhibitors were associated with a higher incidence of bleeding/wound complications in comparison with aspirin [24]. The choice of VTE prophylaxis should be based on the perceived risks of bleeding and wound complications compared to the risks of VTE in each patient.

29.7 Infection After Thromboprophylaxis

TKA is a relatively safe procedure, with <1% of these procedures complicated postoperatively by periprosthetic joint infection [25, 26]. Managing and/or eliminating risk factors that predispose a patient to periprosthetic joint infection is critically important. The use of certain agents to prevent deep vein thrombosis after arthroplasty has been linked to an increased risk of adverse effects including wound drainage and infection.

Chahal et al. measured the return to theatre rate for any cause related to wound complications

in patients undergoing total hip replacement and total knee replacement and compared these rates between patients on oral rivaroxaban 10 mg OD and subcutaneous enoxaparin 40 mg OD. In this retrospective cohort study, it was found that patients who received rivaroxaban were more than twice as likely to return to theatre for wound complications compared to patients receiving enoxaparin. Although not statistically significant, this increase is in line with previous studies. Infection rates increased from 0.9 to 1.9% after the introduction of rivaroxaban and microbiologically confirmed superficial infections rose from 1.3 to 3.1% after rivaroxaban was introduced. These rises were not statistically significant [27].

Brimmo et al. compared the early deep postoperative surgical site infection and subsequent reoperation rates in THA and TKA patients treated with either oral rivaroxaban or any other form of chemical thromboprophylaxis [28]. Patients were divided into two groups: the study group received rivaroxaban, whereas the control group received another form of chemical thromboprophylaxis for at least 2 weeks postoperative. There were no significant differences between groups regarding demographics, risk factors, or illness severity scores. Incidence of early deep SSI in the rivaroxaban group was higher than in the control group (2.5% vs. 0.2%; $P < 0.015$). The use of rivaroxaban for thromboprophylaxis led to a significantly increased incidence of deep SSI in a continuous series of patients.

Aspirin is a widely used antiplatelet drug. It prevents platelet aggregation by inhibiting the production of thromboxane A₂ by activated platelets [29]. AAOS has endorsed aspirin for VTE prevention after total joint arthroplasty (TJA) [30]. In 2012, ACCP evidence-based clinical practice guidelines (9th edition), for the first time, acknowledged the usage of aspirin for prophylaxis of pulmonary embolism (PE) after TJA (Grade IB recommendation) [19].

Raphael et al. compared the (1) overall frequency of symptomatic PE, (2) risk of symptomatic PE after propensity matching that adjusted for potentially confounding variables, and (3) other complications and length of stay before and after propensity matching in patients undergoing

TJA at our institution who received either aspirin or warfarin prophylaxis. The overall symptomatic PE rate was lower ($p < 0.001$) in patients receiving aspirin (0.14%) than in the patients receiving warfarin (1.07%). This difference did not change after matching. The aspirin group also had significantly fewer symptomatic DVTs and wound-related problems and shorter hospital stays, which did not change after matching [31].

Huang et al. compare the rates of periprosthetic joint infection (PJI) at our institution in patients receiving aspirin compared with warfarin for VTE prophylaxis following TJA. Incidence of PJI was significantly lower at 0.4% (8 of 1456 patients) in patients receiving aspirin as VTE prophylaxis compared to 1.5% (24 of 1700 patients) in patients receiving warfarin ($P < 0.001$). Rate of postoperative PE was also lower in the aspirin group at 0.1% (1 of 1456 patients) compared to 0.3% (5 of 1700 patients) in the warfarin group ($P < 0.001$). Multivariate analysis identified warfarin prophylaxis compared to aspirin as an independent risk factor for PJI following TJA ($P = 0.018$). Patients receiving aspirin prophylaxis have fewer wound-related complications following primary TJA, which theoretically explains its added benefits in reducing the incidence of SSI. The use of aspirin compared to warfarin for VTE prophylaxis provides adequate protection against postoperative VTE while reducing the risk of SSI following TJA [32].

In a majority of studies evaluating VTE prophylaxis in patients undergoing TJA, aspirin appears to result in a lower risk of SSI than anticoagulants (vitamin K antagonists, heparin-based products, factor Xa inhibitors, and direct thrombin inhibitors).

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30.1 Introduction

Polymethyl methacrylate (PMMA) or bone cement has been used since the late 1940s [1]. The first available reports are about its use for fracture fixation and even bone substitution in femoral and humeral head fractures [2, 3]. The mechanical properties of PMMA and the versatility it offers in terms of shape conformation as well as for substitution and fixation favored its introduction in the field of orthopedics. In 1964, John Charnley published the first study on cemented prosthesis. It is the one currently in use, albeit with some slight differences [4]. Then again, in subsequent decades some reports on its disadvantages like an allergic reaction, cardiac arrest, pulmonary embolism, among others, were also published [5–8].

The use of PMMA in orthopedic infections was first proposed by Buchholz in 1970. However, it was difficult to convince the orthopedic community that antibiotics could elute from a stony material such as PMMA [9–11]. Nevertheless, he continued with the use of antibiotic-loaded bone cement (ALBC) for the treatment of prosthetic infections and published promising results in 1981 [12]. Buchholz pioneered one-stage exchange by

using ALBC with two purposes, for fixation for a permanent implant and infection treatment in prosthetic joint infections (PJI).

The use of ALBC became popular and new indications such as prophylaxis were started in this decade [13]. Therefore, the purpose is not only for fixation of a permanent implant but also infection prophylaxis.

All the previously cited studies were about hip prosthesis. Ten years later, in 1983, John Insall described a new technique to treat PJI. It was denominated the two-stage exchange for knee PJI [14]. Although an ALBC spacer was not used in the interval, Borden and Wilde introduced the use of an ALBC spacer for the interim in TKA two-stage exchange a few years later [15, 16]. In this case, ALBC was used for void filling and space maintenance with a temporary implant along with infection treatment. In that decade, reports on one-stage exchange for TKA infections using ALBC were also published by groups distinct from the Buchholz group. However, the aims were the same, infection treatment and prosthesis fixation [17].

30.2 Characteristics

PMMA is a resistant plastic also known as acrylic. It has multiple uses such as headlights in vehicles, photo frames, tablecloths, and other household items like lamps. In medicine, it is

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used in the making of diagnostic tools, but its main use is as bone cement for dentist, orthopedic surgeons, and neurosurgeons.

The mechanical properties of bone cement are considerable, and it is rigid at room temperature. In contrast, it has low impact resistance and is sensitive to heat [18].

The composition of commercial acrylic bone cement differs in some modifications. It also comes with the addition of co-polymers of PMMA and some different co-monomers in the liquid. Bone cement powder predominantly contains PMMA in powder form that also carries radiopacifiers like barium sulfate or zirconium dioxide. In the case of ALBC, the antibiotic is usually incorporated into the powder phase [18].

The modifications of each brand make for the differences in mechanical properties as well as in the hydrophilic characteristics of the bone cement. The latter are crucial in ALBC as the elution of most antibiotics depends on how hydrophilic the cement is. The liberation of the antibiotic from the ALBC also depends on the porosity of the cement mantle and the total surface of the cement as the antibiotic only elutes from the outer surface, which can absorb fluid and then release it together with the antibiotics [19].

It is true that the characteristics of the powdered polymer and the liquid monomer influence antibiotic release, as stated before, but the opposite is also true. The addition of antibiotics affects its mechanical characteristics. Liquid antibiotics cause greater loss of compressive strength than powder preparations [20]. It has been found that the addition of antibiotics amounting to up to 10% of the weight of the PMMA barely affects mechanical strength [21, 22]. Other properties that can affect its mechanical characteristics are fatigue limit, fracture toughness, and the polymerization rate, which may differ between brands [22, 23].

30.3 Clinical Uses of the ALBC in the Knee Surgery

As previously stated, ALBC was first used for hip prosthesis but soon became popular among knee surgeons and its use spread worldwide. In fact,

contrary to hip prosthesis, total knee arthroplasty (TKA) is rarely uncemented nowadays. According to the Nordic registries, up to 90% of TKA are cemented [24].

Although the use of ALBC around the knee can be very different (osteomyelitis treatment, Masquelet technique, open fracture dead space management, etc.), this chapter will focus only the use of ALBC in the prosthetic field.

ALBC can be employed in cases of primary TKA in which the aim will be fixation and infection prevention. In cases of TKA revision, the surgeon can also use ALBC for fixation and infection prevention when dealing with aseptic revisions. It can also be used for fixation and infection treatment in cases of one-stage septic revision and two-stage septic revision with a short interval. In all those instances, the cement will be permanently left in the patient. Finally, ALBC can be utilized for dead space management and infection treatment, which is the case of temporary spacers in the first stage of two-stage septic revisions [25, 26].

30.4 ALBC in Primary TKA

The prevention of PJI is a major concern among orthopedic surgeons. One of the most important measures to reduce the risk of infection is intravenous antibiotic prophylaxis. It has shown an 81% reduction in the relative risk [27]. The rationale for the combination of both local and systemic prophylaxis is based on multiple factors. Among them, there is a broadening of the antimicrobial spectrum, an improved antimicrobials synergistic effect, pharmacokinetics optimization and its action as a local antimicrobial barrier [28].

Cephalosporins are the most frequent antibiotic prophylaxis used in orthopedic surgery and aminoglycosides are the predominant ones in ALBC. While the first group covers most Gram-positive bacteria, the second is effective against Gram-negative that may not be susceptible to first-generation cephalosporins [26]. Moreover, the synergistic effect of beta-lactams and aminoglycosides has been well-known for decades [29, 30].

From the pharmacokinetic point of view, ALBC provides a high local concentration of

antibiotic. That is something that is difficult to achieve when antibiotics are given intravenously. In that sense, Hendricks et al. found that the concentrations of gentamicin inside the gap between the bone and the prosthesis within the 2 h after surgery were about 1000 times higher than the minimal inhibitory concentration (MIC) for staphylococci [31]. This concentration may effectively decontaminate the prosthesis-related interfacial gap of any accidental contamination during surgery. Therefore, ALBC would act as a local barrier for accidental contamination.

For all the previously stated reasons, ALBC for PJI prevention in primary prosthesis is widely used in northern Europe. However, its use is not yet approved for that purpose in the United States. It is only approved for prophylaxis in revision cases in the USA [28, 32].

For local prophylaxis with ALBC, there is good evidence supporting its use for cemented hip arthroplasties [33–35]. In the case of TKA, the results from different studies are heterogeneous. Hinarejos et al., in one of the largest prospective randomized TRIAL with ALBC, found no differences between the ALBC and the plain cement group in terms of the infection rate [36]. On the other hand, Chiu did find a reduction of infection when cement loaded with cefuroxime was employed [37]. Additionally, data from arthroplasty registries seem to suggest that ALBC might reduce PJI after TKA [38, 39]. Aminoglycosides are the group of antibiotics most used in ALBC in the Finland, England, and Wales. They are the countries of origin of those studies. On the other hand, colistin, which has a limited spectrum against Gram-negative bacteria, and erythromycin, a bacteriostatic and with high rates of resistance to staphylococci were used in the study done by Hinarejos et al. [36, 38, 40]. This may be the reason for the different results obtained as the latter combination is not the optimal one.

In any case, what it is clear is that for fixation of a primary TKA along with prophylaxis, a low dose of antibiotic is advocated for to avoid mechanical property weakening. Aminoglycosides are the best antimicrobials to add (0.5 g of gentamicin per 40 g of PMMA or 1 g of tobramycin per 40 g of PMMA).

30.5 ALBC in Revision TKA

The reasons for TKA revision are varied. Nevertheless, the most important one that must be ruled out before surgery is infection. This is clear as the approach is completely different and not diagnosing PJI can make curing the infection more difficult in the long run. The big challenge in PJI is low-grade chronic infections as most of them are only accompanied by pain [41]. No other sign or symptom is seen in most of PJI caused by coagulase-negative staphylococci or *cutibacterium* spp. Therefore, it is crucial to use a pro-active and thorough diagnostic protocol to identify them and prevent unsuspected intraoperative positive cultures [42, 43]. In that sense, the criteria proposed by Zimmerli has been found to be more reliable in identifying PJI, specially low-grade infections [41, 43, 44].

Once PJI has been ruled out, aseptic revision is performed. In this case, ALBC has been clearly found to be superior to bone cement without antibiotics [45–48]. In that sense, the use of ALBC can be considered the gold standard for care in TKA revision [46]. There are several theories to explain these results. The most accepted is that some of those supposed aseptic revisions were actually low-grade infections that were not identified as previously stated [43, 49]. Some other reasons might be that the bone and soft tissues are of poor quality and less vascularized than in primary cases and therefore more prone to infection [50]. Moreover, the surgical duration of a revision is normally superior to a primary surgery and the blood loss is usually greater, which are factors that have been related to increased risk of infection [50–52].

As mentioned, ALBC in revision cases can be used for infection prevention in aseptic revisions. It is also used for infection prevention in second-stage reimplantation when there is a long interval between procedures in a two-stage exchange. In this case, infection has been cured after TKA removal and debridement and the interval with the knee spacer and systemic antibiotic treatment [25, 44, 53, 54]. In both cases, ALBC is used for prosthesis fixation and therefore its mechanical properties must be preserved.

To do so, a precise mixing technique that respects the prescribed mixing time and is done in vacuum is mandatory. Moreover, antibiotics must not exceed 10% of the weight of the PMMA to prevent weakening of the cement and the subsequent loss of strength [26]. The use of ALBC with a combination of two antibiotics is generally recommended in revision cases because of the synergistic effect and the broad spectrum of the combination. Moreover, studies seem to suggest there is better performance than with ALBC with gentamicin alone and it can also prevent resistant bacteria selection [34, 55].

In both aseptic revisions and the second stage of the two-stage septic revisions, a commercially available ALBC with two antibiotics is recommended for the previously stated reasons.

When dealing with one-stage exchange PJI revision, in which infection has not yet been cured, ALBC is used for the treatment of the infection as well as prosthesis fixation. The local activity of antibiotics along with prosthesis exchange, surgical debridement, and systemic antibiotics are used to cure infection [54, 56, 57]. The identification of the microorganism and its antibiotic susceptibility are crucial if this approach is used [57, 58]. Ideally, the microorganism is identified preoperatively, and the bone cement is loaded with antibiotics that are active against that microorganism. However, preoperative bacterial isolation is sometimes not possible. For those cases, a two-stage exchange with a short interval of approximately 2 weeks has been proposed [53, 59]. By doing so, the bacteria can be identified by means of intraoperative tissue cultures and prosthesis sonication. To do the implantation of the prosthesis in such a short interval, the bacteria must be susceptible to antibiofilm antibiotics [53, 60]. This interim and bacteria identification will also allow for the identification of the best ALBC antibiotic combination. It is important to mention that not all antibiotic combinations are commercially available. Therefore, an “off-label” hand-made mixing technique must be used in most of the cases [26]. Antibiotic mixing according to bacterial isolation can be seen in Table 30.1 [26, 58].

30.6 ALBC for Knee Spacers

Knee spacers are used in the two-stage septic revision technique. They can be either static or dynamic, self-made or preformed. Their differences are not the issue of the present chapter. Since they are used temporarily, high antibiotic doses can be used to treat infection regardless of the impact it can have on the mechanical properties [61]. It has been proven that antibiotic elution for the commonly used interim times is enough to eradicate infection [62]. Large amounts of antibiotic have been found when the spacer is analyzed after removal [63, 64]. However, it is worth knowing that if the spacer is left in place for a longer period (usually longer than 6–8 weeks), it may behave like a foreign body and biofilm may attach itself since antibiotic elution decreases [62, 65]. When using a self-made spacer, antibiotics for ALBC can be tailored to the preoperative cultures (when available) and the desired amount can be increased to even more than 10% of the weight of the PMMA [26, 61]. In addition, the mixing technique can be modified to obtain a “bad quality” cement. This means doing it without a vacuum to increase the number of bubbles which will explode afterwards eluting the antibiotics. Elution from the spacer also depends on the surface. A simple technique, making indentations with a scalpel, can increase the elution [66]. The legal issues around the off-label use of this mixing technique and manual addition of antibiotics mentioned in the previous point are less important here since the spacer is only a temporary device for a short period of time. The antibiotic combination in accordance with the microbiological results can be seen in Table 30.1 [26, 58].

30.7 Antibiotics for Bone Cement

Not all antibiotics are suitable for mixing with bone cement. The optimal characteristics of the antibiotics to be mixed with PMMA can be seen in Table 30.2 [61, 67].

Liquid antibiotics have a significant damaging effect on the mechanical properties of the bone cement, especially the compressive strength [68]. Some antibiotics, like rifampin, can impair

Table 30.1 Local antimicrobials in bone cement (PMMA) (additionally to systemic antimicrobial treatment)

| Situation | Antimicrobial (AM) | Fixation cement (prophylactic dose: per 40 g PMMA cement) | Spacer cement (therapeutic dose: per 40 g PMMA cement) |
|--|---|---|---|
| | | Black: industrially admixed AM Blue: manually admixed AM | |
| Standard situation • Susceptible or unknown pathogen(s) | Gentamicin + Clindamycin | 1 g 1 g | 1 g 1 g (+2 g vancomycin) |
| Special situations • <i>Staphylococcus</i> spp. (oxacillin-/methicillin-resistant) or <i>Enterococcus</i> spp. | Gentamicin + Vancomycin <i>or</i> Daptomycin | 0.5 g 2 g – | 0.5 g 2 g (+2 g ^a) 2 g |
| • Vancomycin-resistant enterococci (VRE) | Gentamicin + Linezolid <i>or</i> Daptomycin <i>or</i> Fosfomycin-sodium ^b | 0.5 g 1 g 2 g 1 g | 0.5–1 g 2 g 3 g 2 g |
| • Resistant gram-negative pathogens (e.g., <i>E. coli</i> , <i>Klebsiella</i> , <i>Enterobacter</i> , <i>Pseudomonas</i> spp.) | Gentamicin + Colistin ^c <i>or</i> Fosfomycin-sodium ^b <i>or</i> Meropenem <i>or</i> Ciprofloxacin | 0.5 g 2 g (=60 Mio E) 1 g 2 g 2 g | 0.5–1 g 4 g (=120 Mio E) 2 g 3 g ^d 3 g |
| • Yeasts (<i>Candida</i> spp.) or molds (e.g., <i>Aspergillus</i> spp.) | Gentamicin + Amphotericin B liposomal (Ambisome [®]) <i>or</i> Voriconazole | 0.5 g 0.1 g ^e 0.2 g | 0.5–1 g 0.2 g ^{a,c} 0.4 g ^d |

General considerations:

- When additional antimicrobials are admixed, industrially impregnated cements are preferred over plain cements (better mechanical properties and elution due to synergistic release)
- Antimicrobial susceptibility testing results are applicable for systemic antimicrobial application and might not be valid for local antimicrobial application due to high local concentrations and synergistic activity
- Side effects and interactions of local antimicrobials are rare. However, serum concentrations of vancomycin and gentamicin should be monitored in patients with kidney insufficiency and/or intravenous application
- Only use sterile antimicrobials in powder form. Liquid antimicrobials are not recommended due to inhomogeneous distribution in PMMA. Antibiotics that interfere with polymerization process (rifampin or metronidazole) or which are thermolabile or sensitive to oxidation (e.g., some beta lactams) should not be used
- Data on mechanical stability are not available for combinations of more than two antimicrobials. If possible, the total amount of antimicrobials should not exceed 10% of the PMMA powder weight (=4 g per 40 g)
- Recommendations are based on studies with PALACOS[®]/COPAL[®] PMMA cements and literature data. Elution data depend on the PMMA cement basis used
- Do not use vacuum mixing for preparation of spacer cement (higher porosity → better antimicrobial elution)

^aThese AM concentrations do not fulfill the mechanical ISO requirements for fixation cement

^bFosfomycin-sodium is preferred over fosfomycin-calcium due to better mechanical properties of PMMA

^cAvailable as colistin-sodium or colistin-sulfate (equal efficacy)

^dImproved efficacy and antimicrobial release in combination with gentamicin 1 g and clindamycin 1 g

^eLiterature is still controversial regarding minimal effective concentrations

cement polymerization and therefore hamper the curing process. The thermal stability of the antibiotic is also crucial as the exothermal reaction of the cement curing process may impair antibiotics like cloxacillin. Nevertheless, *in vitro* studies about the mechanical properties of ALBC are not always similar to *in vivo* results [69]. The brand of antibiotic added even has a different impact on the mechanical strength of the ALBC [22]. What

is clear is that the more antibiotic added, the more impairment of the mechanical characteristics.

As a general rule, low antibiotic doses (<2.5% PMMA weight) are used for PJI prevention. For fixation of revision cases (either septic or aseptic), medium doses of up to 7% of the weight of the PMMA are recommended (<10% in any case). For spacer purpose in PJI treatment, 10% or more is usually used [28].

Table 30.2 Ideal properties of antibiotics to be used in antibiotic-loaded bone cement

| |
|--|
| • Availability in powder form |
| • Wide antibacterial spectrum |
| • Bactericidal at low concentrations |
| • Elution from PMMA in high concentrations for prolonged periods |
| • Thermal stability |
| • Low or no risk of allergy or delayed hypersensitivity |
| • Low influence on the mechanical properties of the cement |
| • Low serum protein binding |

30.8 Types and Combination

Aminoglycosides are the most frequently used antibiotics worldwide. Gentamicin and tobramycin are mostly used for PJI prophylaxis (alone) and in combination for revision [67]. Commercially available bone cements loaded with two antibiotics are limited. The combination of gentamicin with clindamycin is recommended for aseptic revisions or PJI caused by anaerobes (*cutibacterium* spp.). The combination of gentamicin with vancomycin is recommended for septic revisions secondary to staphylococci (especially when dealing with MRSA) [26].

When it comes to self-made hand mixing in the operating theater, one must keep in mind that the cement characteristics on the brand's label are being modified and therefore an off-label use is being done. In that sense, there are studies that show that mechanical strength and antibiotic elution are worsened when the antibiotic is added manually [70, 71]. However, there are PJI cases that cannot be treated with the commercially available ALBC, for instance fungi PJI. For those cases, Table 30.1 summarizes the optimal antibiotic combination for PMMA.

30.9 Resistances

The selection of resistant bacteria is always a possibility when an antibiotic is used. Some factors that can favor this phenomenon are inadequate doses with a threshold under the MIC for the microorganism and a short antibiotic concen-

tration time above the MIC. In that sense, ALBC provides high elution of antibiotics that are clearly above the MIC of the bacteria causing PJI in the first hours [19]. However, the major concern is whether the ALBC can provide an antibiotic concentration over the MIC for a long period, specially in cases of coagulase-negative staphylococci. There has been a shift towards higher MICs in those cases in the recent years. These assumptions are particularly important in the case of ALBC with gentamicin used for PJI treatment in which a higher rate of gentamicin-resistant staphylococci has been found in the recent years [19, 72]. When aminoglycosides are used alone for prevention, resistance selection is not a concern since the high amount of local antibiotics are enough to eradicate intraoperative contamination that is caused by a small inoculum of bacteria.

In the cases of PJI, the combination of two or more antibiotics is crucial for the synergistic effect, to improve antibiotic elution and to broaden the treatment spectrum. The same explanation is true for primary cases in high-risk patients [34, 73, 74].

30.10 ALBC Costs

One of the major concerns with the use of ALBC is the added cost that it implies. In revision cases, there is no doubt relative to the use of ALBC. However, the issue of cost arises in primary cases when ALBC is used as a prophylactic measure. Most of the cost-effective studies on ALBC come from the USA. Therefore, there are some peculiarities. The diagnostic criteria for PJI proposed by American societies of infectious diseases and orthopedics is different from the one proposed by the European society. Furthermore, the costs for PJI treatment and ALBC are clearly different. Moreover, since the PJI rate in primary TKA is quite low, it would be difficult to take any preventive measure to reduce this low rate even further.

In that sense, Sanz-Ruiz studied those variations and found ALBC to be cost-effective when the PJI rate is 4% or greater [75].

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Arthroplasty After Septic Arthritis

31

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31.1 Introduction

Delayed or inadequate treatment of septic arthritis can rapidly result in irreversible joint destruction, systemic sepsis, and even death as it has a mortality rate of 9–11% [1]. This rate is considerably higher in elderly patients, the presence of multifocal disease, significant co-morbidity and treatment failure [2, 3].

A previous history of native joint infection presents unique challenges with risk of prosthetic joint infection (PJI) that is difficult to quantify [4–7]. While total knee arthroplasty (TKA) can provide a durable solution with excellent functional results, it is imperative that the treatment

strategy is patient specific, well planned, and performed with attention to detail in order to avoid potentially devastating and costly sequelae [8].

In this chapter we will briefly discuss the key aspects of the pathogenesis, explore the role of TKA in the context of previous knee infection, provide a synopsis of the evidence and experience for TKA and the treatment strategies utilized, and recommend a strategy for the evaluation and management of patients with post-infection arthropathy for whom knee arthroplasty is considered.

The focus will predominantly be on bacterial and mycobacterial knee infections.

31.2 Pathogenesis

The incidence of septic arthritis is bimodal, predominantly affecting young children and older adults. The spectrum of knee infections that may result in debilitating arthropathy includes intra-articular infections, with or without osteomyelitis, and can arise from a variety of systemic, local or iatrogenic causes, as highlighted in Table 31.1.

The most common pathogen is *S. aureus*, with *Staphylococcus epidermidis* and *Streptococci* less commonly implicated [1]. Other organisms are also associated with certain population groups, such as group A

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Table 31.1 Summary of causes of prior knee infection

| Origin of infection | Proportion (%) | | |
|---------------------|-------------------------|--------------------------|----------------------------|
| | Seo et al. [9] (N = 62) | Lee et al. [10] (N = 20) | Jerry et al. [11] (N = 65) |
| Post-operative | 50 | 35 | 49 |
| Hematogenous | 35 | 50 | 26 |
| IA injection | 10 | 0 | 25 |
| Miscellaneous | 5 | 15 | 17 |

Streptococcus and *Enterobacter* in children, *Salmonella* in patients with sickle-cell disease, methicillin-resistant *S. aureus* (MRSA) in intravenous drug users (IVDUs), mycobacterial and fungal infections in the context of immunocompromise [1, 12]. Although these associations may guide initial empiric therapy, pathogen identification is an important aspect of managing the initial infection as well as the potential long-term sequelae. A multi-disciplinary approach to these cases should be adopted. The increasing prevalence of drug-resistant organisms such as MRSA and methicillin-resistant coagulase negative staphylococcus (MRCNS) add to the complexity of treatment and necessitate input from a specialist in microbiology or infectious diseases. Figure 31.1 represents the individual and pooled proportion of organisms identified in patients undergoing TKA with a history of prior septic arthritis for the 3 largest published series [9, 11, 13]. This highlights the diversity of primary bacterial knee infections encountered prior to arthroplasty and demonstrates the groups of bacteria that show a tendency to persist or cause prosthetic joint infection, which are maybe different to those identified initially.

Staph aureus virulence factors have a significant role in the promotion of joint destruction in septic arthritis [14], and the presence of Pantan-Valentine leucocidin (PVL) has been associated with a higher incidence of fulminant infections, complications and treatment resistance [15].

Tuberculosis (TB) remains a significant cause of morbidity with an estimated worldwide prevalence greater than ten million [16]. Extra-

pulmonary musculoskeletal disease is reported in 1–3% of those cases and the knee joint is the most frequent musculoskeletal site after the spine and hip [17, 18].

31.3 Patterns of Arthropathy

Patient age influences the pattern of arthropathy that develops. In young children, reduced range of motion, growth disturbance and malalignment are the most frequently encountered sequelae [19], that may be followed by subsequent attempts at correction of deformity, leg length discrepancy and flexion contractures. Older adults may have significant comorbidities such as inflammatory arthropathy, diabetes, renal disease or pre-existent osteoarthritis [1].

Gächter described a classification system of arthroscopic findings of septic arthritis (Table 31.2) and demonstrated that more advanced stages (III and IV) are associated with multiple surgical procedures and treatment failure, and subsequent authors have correlated these stages with delayed treatment and adverse clinical outcomes [20–22].

In the case of TB infection, the radiographic appearance of the knee on presentation as described by Kerri and Martini (Table 31.3) is predictive of outcome, whereby the atrophic or arthritic types showing joint space narrowing and/or gross anatomic disorganisation are more likely to result in deformity, stiffness or even ankylosis [24, 25].

Primary TKA has been performed with a history of fungal infection, such as *Sporothrix schenckii* and *Candida Albicans*, but this is rare, and mostly limited to case reports [26, 27].

31.4 The Role of Arthroplasty in the Context of Previous Knee Infection

Historically, the presence or history of infection has been a contraindication to arthroplasty, and advanced, debilitating joint destruction in this

context has been managed with resection arthroplasty or arthrodesis if surgery is contemplated [28]. While these can control infection and relieve pain [29], they lead to a marked reduction in function [30]. With increasing experience in the management of prosthetic joint infections, the understanding of biofilm and virulence factors,

and a team approach, arthroplasty has become a viable option.

At present, the literature reporting on knee arthroplasty following septic arthritis is sparse and limited to case reports and case series. Sixteen studies have been identified reporting on >5 cases of knee arthroplasty for arthritis follow-

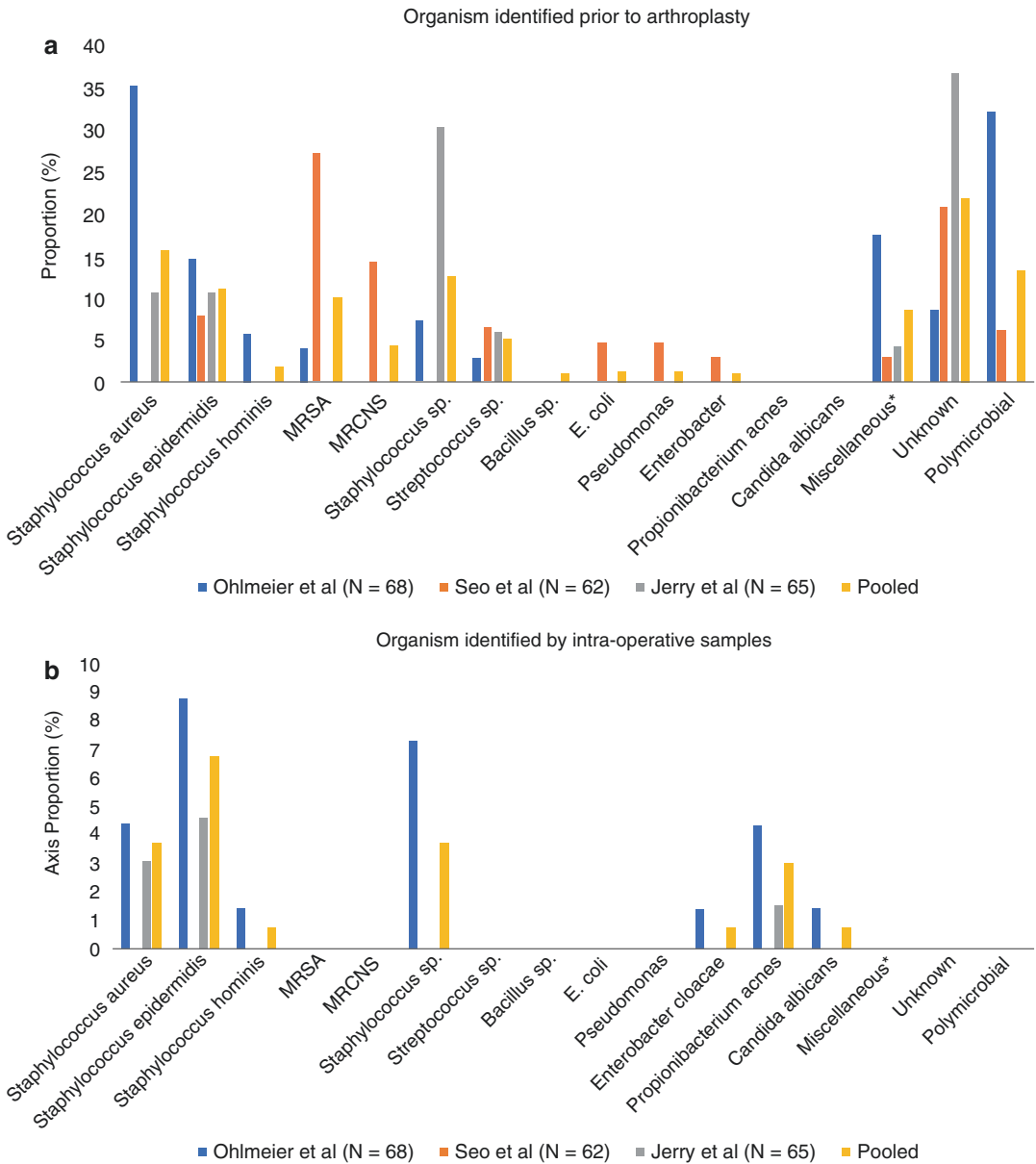


Fig. 31.1 Individual and pooled proportions of pathogens identified in patients undergoing TKA from Ohlmeier et al., Seo et al., and Jerry et al. MRCNS, methicillin-

resistant coagulase negative Staphylococci; MRSA, methicillin-resistant *S. aureus*

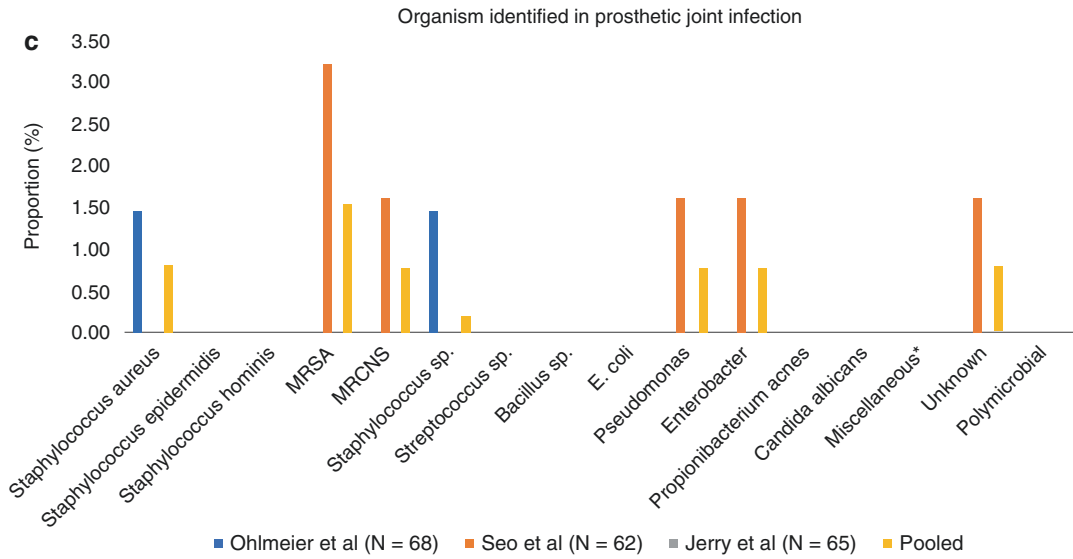


Fig. 31.1 (continued)

Table 31.2 Gächter grading of septic arthritis [23]

| Stage | Criteria |
|-------|---|
| I | Opacity of fluid, redness of the synovial membrane, possible petechial bleeding, no radiological alterations |
| II | Severe inflammation, fibrinous deposition, pus, no radiological alterations |
| III | Thickening of the synovial membrane, compartment formation (“sponge-like” arthroscopic view, especially in the suprapatellar pouch), no radiological alterations |
| IV | Aggressive pannus with infiltration of the cartilage, possibly undermining the cartilage, radiological signs of subchondral osteolysis, possible osseous erosions and cysts |

Table 31.3 Kerri and Martini radiographic classification of tuberculosis of the knee [25]

| Stage | Criteria |
|-------|---|
| 1 | No bone lesions. Localised osteoporosis |
| 2 | One or more erosions (or cavities) in the bone. Discrete diminution of the joint space |
| 3 | Involvement and destruction of the whole joint without gross anatomical disorganisation |
| 4 | Gross anatomical disorganisation |

ing bacterial or tuberculous infection of the native knee, summarised in Tables 31.4 and 31.5. Prior to the report of 68 patients by Ohlmeier et al. in 2019 [13], the largest published series of TKA

following knee infection was published in 1988 and included 65 patients (65 knees) [11]. Some authors have taken almost a decade to accumulate less than 20 cases [31, 32].

Pooling of data and comparison of results is challenging because of the marked heterogeneity in terms of age of onset of infection (childhood vs. adult), type of infection (joint with or without osteomyelitis), type of procedure performed (single- vs. 2-stage), time interval between index infection and arthroplasty, interval between stages, the pathogen, and the antimicrobial strategy. Nonetheless, these studies provide valuable insight into the unique challenges in this context.

There are two broad clinical scenarios related to knee infection in which the role of knee arthroplasty has been practised and studied:

1. The treatment of post-infective arthropathy.
2. The management of active or evolving knee infection that is resistant to standard medical and surgical treatment.

The latter stems from our evolving experience from the treatment of PJI, where many of the management principles have been extensively studied.

Table 31.4 Summary of studies reporting on arthroplasty following bacterial infection of the knee

| Study Author Journal Year | N | Type of infection | Years since infection Mean (range) | Follow-up in years Mean (range) | Antimicrobials | Staging | Persistent or re-infection rate % | Other complications | Outcome measures | | |
|---|---------|----------------------|--|--|--|---------|---|---|--|--|-----|
| | | | | | | | | | Survivorship | Knee scores | ROM |
| Ohlmeier et al. J Arthroplasty 2020 | 68 (68) | Bacterial | 9.6 (0–63) | 5 (1–9) | Drug (duration) 2 g vancomycin De-escalated to beta-lactam after 10.5 days (1–30) or culture specific ALBC (per 40 g) 2 g vancomycin + 1 g gentamicin ± 2 g meropenem Or 1 g clindamycin + 1 g gentamycin ± 2 g vancomycin | Single | 2.9% | Arthrofibrosis (1.5%) Aseptic loosening (1.5%) Wound healing (2.9%) Haematoma (2.9%) Nerve palsy (1.5%) Non-surgical (15%) | 97.1% at 5 years | OKS last FU 34.6 (8–48) KSS Last FU 14.9 (4–20) | |
| Xu et al. BMC MD 2019 | (19) | Bacterial | 4.9 ± 3.8 months between stages | 4.7 (2.2–10.8) | IV antibiotics ≥6 weeks after 1st stage and 5 days after 2nd stage ACCS (per 40 g) 4–6 g vancomycin + 2–4 g meropenem 2nd stage 1 g vancomycin per 40 g cement for implant | 2-stage | 16% | Not reported | Survivorship 1-year 94.7% 2-year 89.5% | NR | NR |

(continued)

Table 31.4 (continued)

| Study | N | Type of infection | Years since infection | Follow-up in years | Antimicrobials | Staging | Persistent or re-infection rate | Other complications | Outcome measures | | |
|---|------------------|-------------------|-----------------------|--------------------|--|---------|---------------------------------|---|---|---|-----|
| | | | | | | | | | Survivorship | Knee scores | ROM |
| Author Journal Year | Patients (knees) | | Mean (range) | Mean (range) | Drug (duration) | | % | | | | |
| Seo et al. J Arthroplasty 2014 | 62 (62) | Bacterial | 4.3 (0.3–22) | 6.1 (2–10.4) | ALBC 500 mg erythromycin + 240 mg colistin per 40 g cement for implant | Single | 9.7% | Wound healing problems (6.5%) MUA (4.8%) Superficial wound infection (1.6%) | UCLA Pre-op 3.3 (2–5) Last FU 6.8 (5–9) KSS Pre-op 58 (36–74) Last FU 86 (58–96) WOMAC Pre-op 76.9 (68–86) Last FU 34.5 (20–48) | ROM Pre-op 99° (50–140) Last FU 125° (90–140) | |

| | | | | | | | | | | | |
|-------------------------------------|------------------------------------|--|---|---------|---|---------|----|----|----|---|--|
| Shaikh et al CORR 2014 | 1st stage 15 2nd stage 13 | Bacterial (10) Fungal (2) TB (1) | Interval between 1st and 2nd stage 4 months (2–29 months) | 4 (2–7) | IV ABs minimum 2 weeks post 1st stage, based on pre-operative and intra- operative cultures Anti-tuberculous medication (12 m post TKA) ACCS (per 40 g) 4 g Vancomycin + 2 g Streptomycin ±400 mg Amphotericin B for fungal infections 2nd stage 1 g Vancomycin per 40 g cement for implant | 2-stage | 0% | NR | NR | KSS Pre-op 41 (26–73) Last FU 85 (46–93) WOMAC Pre-op 51 (40–65) Last FU 18 (11–31) VAS for pain Pre-op 66 (50–75) Last FU 18 (0–40) | ROM Pre-op 103° (60–155) Last FU 115° (75–150) |
| Chen et al. Orthopaedics 2013 | 22 | Any | Not reported | NR | NR | Single | NR | NR | NR | NR | NR |

(continued)

Table 31.4 (continued)

| Study | N | Patients (knees) | Type of infection | Years since infection | Follow-up in years | Antimicrobials | Staging | Persistent or re-infection rate | Other complications | Outcome measures | | |
|---|----|------------------|-------------------|--|--------------------------|---|---|---|---------------------|------------------|--|-----|
| | | | | | | | | | | Survivorship | Knee scores | ROM |
| Bauer et al. Orthop Traumatol Surg Res 2010 | 31 | | Bacterial | Mean (range) Resolved: 5 years (2–18) Evolutionary: for failed treatment of infection 2nd stage After mean 6w (4w–6m) when CRP stable for 2 weeks | Mean (range) 5 (2–13) | Drug (duration) Single stage: according to initial infection, continued until culture results 2-stage Dual antibiotic therapy for 93 days (45–180 days) Cement spacer without antibiotics | Single stage for 14 knees (Resolved septic arthritis) 2-stage for 17 knees (Evolutionary septic arthritis) | % 7% (single stage) 12% (2-stage) | Not reported | Survivorship | Knee scores KSS (single) Post-op 91 (75–100) KSS (2-stage) 83 (65–100) | ROM |

| | | | | | | | | | | |
|--|--|------------------------------|-------------------------------|-----------|---|---|------|---|--|--|
| Bae et al. JBJS Br 2005 | 32 (32) 20 ankylosed 12 partially ankylosed | Bacterial (14) TB (18) | 18.2 (1–51) | 10 (5–13) | IV cephalosporin immediately after toamiquet deflation, continued for 2 weeks 1 g cephalosporin per 40 g bone cement Anti-tuberculous medication for 6 months | Single | 3.1% | Flexion contractures (22%) Superficial infection (3%) Fracture (3%) Transient peroneal nerve palsy (3%) | HSS Knee score Pre-op 60 (26–87) for ankylosed knees 52 (29–66) for partially ankylosed knees Post-op 85 (64–94) for ankylosed knees 87 (69–97) for partially ankylosed knees | Mean post-op improvement in ROM 66° Post-op ROM 75° (30–115) for completely ankylosed knees Post-op ROM 99° (60–130) for partially ankylosed knees |
| Nazarian et al. J Arthroplasty 2003 | 14 (14) | Bacterial | 2.1 (3 weeks to 7.8 years) | 4.5 (2–7) | IV cefazolin, converted to oral cephalexin Or based on cultures vancomycin 1 g + tobramycin 2.4 g per 40 g cement | 2-staged with ACCS, 6-week interval | 0% | Patella subluxation (5%) Haematoma (5%) DVT (5%) Wound healing problem (5%) | Pre-op KSS 46 Last FU KSS 89 | ROM >110° in 50% No flexion contractures >5° |

(continued)

Table 31.4 (continued)

| Study | N | Patients (knees) | Type of infection | Years since infection | Follow-up in years | Antimicrobials | Staging | Persistent or re-infection rate | Other complications | Outcome measures | | |
|------------------------|---------|------------------|-------------------|--------------------------|-----------------------|---|--|--|--|---|--|-----|
| | | | | | | | | | | Survivorship | Knee scores | ROM |
| Lee et al. CORR 2002 | 19 (20) | | Bacterial | Mean (range) 23.3 (1–66) | Mean (range) 5 (2–11) | Drug (duration) Use of systemic antibiotics not reported 1 g Vancomycin or 1.2 g tobramycin per 40 g cement | Single (18) 2-staged (2) with ACCS and 6-week interval | % 5% | MUA (15%) Marginal skin necrosis (10%) Revision for aseptic loosening (5%) Haematoma (5%) Superficial wound infection (5%) | KSS Pre-op 39 (7–58) Last FU 91 (78–99) | ROM Pre-op 9 (0–40) to 86 (45–100) Post-op 1 (range 0–7) to 100 (85–110) | |
| Jerry et al. CORR 1988 | 65 (65) | | Bacterial | 17.6 (1–65) | 6.1 (2–15) | Semisynthetic penicillin or cephalosporin for 48–72 h ALBC not used | Single stage (65) | 7.7% overall 15% for bone and joint infection 4% for joint infection | Superficial infection (8%) Haematoma (6%) Fracture (5%) Extensor mechanism disruption (3%) | HSS knee score improved from 54 pre-op to 80 at last FU | ROM pre-op 6° (±9) to 90° (±23) pre-op, 4 (±10) to 87° (±32) post-op | |

ACCs antibiotic-containing cement spacer, ALBC antibiotic-loaded bone cement, DVT deep vein thrombosis, FU follow-up, HSS Hospital For Special Surgery, KSS Knee Society Score, MUA manipulation under anaesthesia, OKS Oxford Knee Score, ROM range of motion, UCLH University of California, Los Angeles

Table 31.5 Summary of studies reporting on arthroplasty following mycobacterial infections of the knee

| Study Author Journal Year | N | Patients (knees) | Years since infection | | Follow-up in years | | Antimicrobials Pre-TKA | Antimicrobials Post-TKA | Persistent or re-infection rate | Other complications | Outcome measures | |
|--|---------|---------------------|---|------------------|--|--|---|----------------------------|------------------------------------|--|--|-----|
| | | | Mean (range) | Mean (range) | Mean (range) | Mean (range) | | | | | Knee score | ROM |
| Zeng et al. Int Orthop 2016 | 9 (9) | | 8.7 (3–25) | 4.4 (2–7) | Staging Single (5) 2-stage (4) | Drug (duration) Isoniazid Rifampicin Ethambutol Pyrazinamide (≥3 months) | Drug (duration) Cefazolin 48 h Isoniazid Rifampicin Ethambutol Pyrazinamide (12 months) | % 0% | Nil | HSS Pre-op 44 (30–60) Last FU 82.7 (64–92) | Pre-op 56° (10–90) Last FU 94° (80–110) | |
| Habaxi et al. Eur Rev Med Pharmacol Sci 2014 | 10 (10) | | Active TB | 1.2 (0.5–2.3) | Single | Isoniazid Rifampicin Ethambutol Pyrazinamide (2–4 weeks pre-op) Topical Streptomycin during TKA | Not reported | 10% (1) | Nil | HSS Pre-op 25 ± 2 Last FU 87 ± 5 | Pre-op Not reported Last FU 95° ± 5° | |
| Öztürkmen et al. KSSTA 2013 | 12 | | Active TB 4 ± 1.5 months post diagnosis | 6.1 ± 1.8 | 2-stage | 1st stage: ACCS 2.4 g tobramycin + 2 g vancomycin per 40 g bone cement Interval: Rifampicin, isoniazid, pyrazinamide, ethambutol | After 2nd stage: Isoniazid Rifampicin Ethambutol Pyrazinamide (2 months) Isoniazid Rifampicin (min 10 months) | 0% | NR | NR | NR | |

(continued)

Table 31.5 (continued)

| Study | N | Years since infection | | Follow-up in years | | Antimicrobials Pre-TKA | Antimicrobials Post-TKA | Persistent or re-infection rate | Outcome measures | |
|----------------------------|---------|-----------------------|--------------|----------------------------|---|---|--|---|--|---|
| | | Mean (range) | Mean (range) | Mean (range) | ROM | | | | Knee score | Other complications |
| Su et al. CORR 1996 | 15 (16) | 2.1 (0.2–6) | 6.3 (3.4–11) | Staging Single | Drug (duration) 2–20 months (8) No treatment (8), undiagnosed TB | Drug (duration) Isoniazid Rifampicin Ethambutol Pyrazinamide Minimum 12 months | 31% (5) overall 50% (4/8) without pre-operative anti-TB drugs 12.5% (1/8) with pre-operative anti-TB drugs | Resection arthroplasty (6%) Revision for mechanical failure (6%) Periprosthetic fracture (6%) | KSS Pre-op 30.5 Last FU 83 KSS Function Pre-op 36 Last FU 74 | NR |
| Kim JBJS Am 1988 | 19 (22) | 1 (0.25–5) | 2.7 (2–4) | Single (21) 2-stage (1) | Isoniazid Rifampicin Ethambutol Pyridoxine Streptomycin 3 months (6 pts) 11–47 months (8 pts) No treatment (5 pts) | Isoniazid Rifampicin Ethambutol Pyridoxine Streptomycin All 5 drugs: 2 months isoniazid, rifampicin, ethambutol, and pyridoxine: 16 months | 16% (3) | NR | NR | NR |
| Eskola et al. JBJS Br 1988 | 6 (6) | 35 (4–66) | 6.3 (3–10) | Single | Rifampicin Isoniazid (2–3 weeks) (3 patients) No TB Rx (3 pts) Flucloxacillin (6 patients) | 3 weeks Rifampicin Isoniazid (3 patients) 5 days Flucloxacillin (6 patients) | 17% (1)—did not receive pre-op anti-TB Rx | Nil | HK Knee Score Pre-op 43 Last FU 25–60 flexion Last FU 80 (45–100) | Pre-op 80 (20–110) Last FU 67 (30–100) |

FU follow-up, HSS Hospital For Special Surgery, KSS Knee Society Score, OKS Oxford Knee Score, ROM range of motion, TB tuberculosis, UCLH University of California, Los Angeles

31.5 Challenges and Controversies in Evaluation and Management

31.5.1 Incomplete History

Patients with knee arthrosis may present several years after the original knee infection, and a detailed history of the original pathogen, sensitivities and treatment may be incomplete. Indeed, the interval between the infection and arthroplasty is as long as six decades in some reports [10, 11, 13]. The pre-operative evaluation, therefore, requires great attention to the history, clinical examination, and special investigations (inflammatory markers, imaging and tissue biopsies).

31.5.2 Comorbidities

Comorbidities such as diabetes, chronic renal disease and rheumatoid arthritis are not uncommon in adults with infection-related arthropathy. This adds complexity to perioperative management and may increase the risk associated with arthroplasty [10, 11, 33].

31.5.3 Joint Versus Bone and Joint Infection

The presence or a history of osteomyelitis have been shown to have a higher risk of PJI in comparison to isolated intra-articular infection of the knee, and has been proposed as a contraindication to arthroplasty [11]. However, the application of a 2-stage approach with radical debridement and cement spacer at the first stage has been the proposed method of mitigating this risk [10, 31–34].

31.5.4 Anatomical Challenges

Soft tissue scarring, decreased blood supply, difficult exposure and increased operative time have

been reported as surgical challenges [11]. This is especially pronounced in cases of complete or partial ankylosis, for which more extensile approach, capsular release and bony resection may be required to achieve adequate range of motion (ROM) [30]. Childhood infections may be associated with growth disturbances, malalignment and leg length discrepancy, which not only require careful pre-operative planning and implant selection, but may also require consideration of an extra-articular deformity correction.

31.5.5 Timing of Arthroplasty

Perhaps the most controversial challenge is to determine the appropriate timing of arthroplasty, for which there is more speculation than consensus.

Kim et al. [35] recommended an infection-free interval of at least 10 years prior to considering arthroplasty for the hip, based on their series of 170 total hip arthroplasty (THAs) for sequelae of childhood hip infections. In all but 1 patient (2 hips), the infection-free interval was >10 years. With a mean follow-up of 9.8 years, the only PJI in this cohort was documented in the patient with an infection-free interval of 7 years, although revision for aseptic loosening and osteolysis occurred in approximately 17% of patients.

Other authors have reported good outcomes with shorter intervals. Bauer et al. regarded patients with a minimum of 2 years free from infection as “resolved” or “quiescent”, for whom they performed single-stage TKA, and the patients who failed medical and surgical management of infection were treated with debridement, synovectomy and 2-stage TKA [34].

Two of the largest series reported by Seo et al. [9] (62 knees) and Ohlmeier et al. [13] (68 knees) described an infection-free interval of at least 2 years and 1 year respectively, showing a PJI rate of 9.7% and 2.9%, respectively.

The International Consensus on Orthopaedic Infections guidelines recommend, “in the absence of concrete evidence”, that arthroplasty is delayed “at least until completion of antibiotic treatment and resolution of clinical signs of infection but no

earlier than 3 months from the inciting event' [7]. Although there are no single accurate markers of resolution of infection, as demonstrated in a recent meta-analysis [36], the use of multiple tools is advocated.

Ohlmeier et al. defined resolution of infection as:

1. Absence of clinical signs and symptoms for acute infection or local inflammation.
2. No signs of active infection on plain radiographs.
3. Normal serum inflammatory markers (ESR, CRP, total leucocyte count).
4. 14-day culture negative results during routine microbiological analysis of synovial fluid taken pre-operatively
5. Minimum 1-year follow-up.

Validating the most accurate combination of metrics may be an area for future research.

31.5.6 Single Versus Staged Approach

Arthroplasty in the context of resolved or quiescent infection has been performed as a single or staged procedure. In patients with evolving active infection, a 2-staged approach is advocated, which essentially comprises of an aggressive debridement, antibiotic spacer, prolonged antibiotic therapy and then reconstruction once infection has resolved. These are different clinical scenarios, and only 2 studies of bacterial [10, 34] and 2 studies of tuberculous [37, 38] knee infections have reported on the use of both approaches. Either approach requires an experienced team and specialist microbiological support. The outcomes for these strategies are discussed in the next section.

31.5.7 Single-Stage Technique

A standard medial parapatellar approach with due respect to previous skin incisions is usually sufficient. Radical debridement, including a total

synovectomy, and antibiotic-loaded bone cement (ALBC) for the definitive implants is generally recommended prior to implantation, with multiple samples for culture and histology [9, 13].

31.5.8 Two-Staged Technique

Based on the experience from the staged management of PJI, this approach involves debridement of infected tissue, including any sinuses, synovectomy and debridement of affected bone [33, 39]. It is essential to do this patiently and thoroughly. Deep cultures are taken, and copious irrigation is performed. Some authors advocate the use of intra-operative frozen section [9, 10, 32, 33], although the value in this context is debated [40]. Various solutions have been utilised, including saline, antiseptic solution and antibiotic-containing solution, without clear evidence favouring any particular recipe [39].

The distal femoral and proximal tibial bone cuts can then be made. Here, the choice of instrumentation requires consideration. Extra-medullary referencing and navigation may avoid the potential for dissemination of microbials into the canal that may occur with intra-medullary referencing. Although both, extra- [33] and intra-medullary [31] referencing techniques, have been utilised.

To manage dead space, deliver antibiotics and prevent contractures, an antibiotic-containing cement spacer (ACCS) is introduced. This can be made with commercially available moulds or manufactured intra-operatively using a cement mould of trial implants [31, 41]. An articulating spacer promotes maintenance of ROM and improved function during the interval, and is preferred to a static spacer in the absence of soft tissue loss, gross instability or orthoplastic soft tissue reconstruction [39].

Perioperative antibiotic cover is followed by culture-specific antibiotics, which are continued for a minimum of 6 weeks, and ceased when clinical evaluation and inflammatory markers suggest resolution of infection [33, 34]. The second stage is planned after a 2-week antibiotic holiday to facilitate intra-operative evaluation and further

cultures—if the infection persists, then the first stage should be repeated [33, 34].

Patella resurfacing appears to be selective [30], routine [9, 33] or not performed, usually in the case of poor bone stock or compromised extensor mechanism [30].

31.5.9 Type, Duration and Mode of Delivery of Antibiotics

Although there is substantial variability in the antimicrobial protocols described (as indicated in Tables 31.4 and 31.5), it is important to adhere to the following principles.

Multimodal antibiotic delivery that is culture specific and based on the advice of a specialist microbiologist is recommended [13, 39]. Perioperative parenteral antibiotics may be given after deep cultures are taken and continued until clinical examination and inflammatory markers indicate resolution of infection. This usually takes at least 6 weeks, although oral equivalents with good bioavailability may be a suitable alternative.

The choice of antimicrobials for an ACCS is governed by cultures, and requires water soluble, heat stable agents, preferably in crystalline form (improved biomechanical strength of cement) [42] to be added to the polymethylmethacrylate (PMMA) powder at the time of cement mixing [39]. Since the load tolerance and endurance of the spacer does not need to match that of the cement used for definitive implants, up to 20% of the mass of the spacer can be composed of antibiotics [39]. Whether this is interpreted as 8 g per 40 g cement powder (48 g in total) or 10 g per 40 g cement powder (50 g in total) shall be left to the reader. The potential to cause adverse systemic sequelae, such as nephrotoxicity, may limit the amount of antibiotic added. Although this is rare [43], it may be more relevant in patients with renal or hepatic dysfunction.

Using 2 (or more) antibiotic agents in the ACCS is common practice and is supported by evidence for the synergistic antimicrobial effect (e.g., gentamicin or tobramycin combined with vancomycin), but the mechanisms for this and

optimal combinations are not well understood [42].

Examples of regimens utilised for ACCS per 40 g bone cement include:

- 4–6 g vancomycin + 2–4 g meropenem [32]
- 4 g vancomycin + 2 g streptomycin ± 400 mg amphotericin B for fungal infections [31]

Examples of ALBC regimens utilised for definitive implant per 40 g bone cement include:

- 2 g vancomycin + 1 g gentamicin ± 2 g meropenem or 1 g clindamycin + 1 g gentamicin ± 2 g vancomycin [13]
- 500 mg erythromycin + 240 mg colistin per 40 g cement for implant.

TB is not regarded as a biofilm producing organism, but the capacity to form granulomas and survive intra-cellularly mandates wide surgical debridement and prolonged antimicrobial therapy. Although the regimens utilised vary widely, some important principles should be highlighted.

Prevention of mycobacterial as well as bacterial infection is required during arthroplasty. ACCS containing 2.4 g tobramycin + 2 g vancomycin per 40 g bone cement [44] has been used, although there is little (if any) antimycobacterial activity with these agents (*refs*). Topical streptomycin, a second line antimycobacterial agent, has been used during definitive implantation [45].

The optimum duration of oral antimycobacterial chemotherapy is not known, but all studies reporting on TKA with a history of TB infection recommend a period of antimycobacterials pre-operatively, ranging from 2 weeks to 47 months [37, 45, 46]. This is supported by case series that have reported higher rates of post-TKA reactivation of TB infection in patients that did not receive pre-operative antimycobacterials in comparison to patients that did [28, 46, 47]. Post-operative regimens are typically 12–18 months, and longer if the clinical examination and inflammatory markers suggest ongoing TB activity [37, 38, 44, 47]. Generally 4 drugs are used initially, namely

isoniazid, rifampicin, ethambutol and pyrazinamide, with 1 study adding streptomycin as well [37]. While some authors recommend continuing isoniazid and rifampicin after the initial 2-month period [37, 44], others have recommended 4 drugs for the entire duration of therapy [38, 47].

No studies report on regimes for drug-resistant or multidrug-resistant TB. Treatment in this instance would be individualised and guided by specialist microbiological or infectious diseases recommendations.

A diagrammatic representation of the principles of the authors' preferred approach to arthroplasty following septic arthritis is summarised in Fig. 31.2.

31.6 Outcomes

Currently, there is no universally accepted, evidence-based definition of success for arthroplasty following septic arthritis [48]. Authors have used rates of recurrent infection, revision rates, range of motion and patient reported outcome measures (PROMs) as outcome measures, and/or based definitions of success on those used for the management of prosthetic joint infections [32]. Indeed, the definition of success may be specific for the individual and, therefore, the patients' perspective of their outcome is an essential element [48].

The International Consensus on Orthopaedic Infections [7] attempted to pool data from 9 studies to quantify the PJI rate for hip or knee arthroplasty following bacterial or mycobacterial septic arthritis. Overall, the PJI rate for 1300 total hip and knee arthroplasties was reported as approximately 6% (95% CI 4.24 to 7.94), with lower rates for childhood infection (2.18%, 95% CI 1.16 to 3.70) compared to adult onset (8.25%, 95% CI 6.48 to 10.55). TKA appears to have a slightly higher overall rate of PJI following bacterial or mycobacterial septic arthritis (8.26%, 95% CI 5.30 to 12.15), in comparison to total hip arthroplasty (5.2%, 95% CI 3.50 to 7.21).

Older age, high pre-operative CRP and drug-resistant organism were identified as risk factors for failure of 2-stage arthroplasty [32].

In their series of 65 TKAs, Jerry et al. [11] reported a PJI rate of 4% for patients with prior septic arthritis and 15% for patients with prior bone and joint infection and subsequently recommended avoiding TKA for patients where infection involved bone. Other authors have utilised a 2-stage approach for such cases.

The PJI rate varies from 2.9% to 9.7% [9, 13] for single-stage and 0% to 16% [31, 32] for 2-stage approaches, but the indications for each approach vary between studies and likely represent increasing complexity of the case and/or concomitant risk factors such as rheumatoid arthritis and immunosuppressive therapy as highlighted by Bauer et al. [34] and Lee et al. [10]. These are the only studies that have used both single- and two-staged approaches in their series. Bauer et al. reported 7% (1 case) PJI infection rate for single stage performed for "quiescent infections" and 12% (1 case) for 2-stage TKA performed for "evolutionary infected joints". The only PJI in the series by Lee et al. occurred in a patient undergoing 2-staged arthroplasty.

The largest series by Ohlmeier et al. [13] reported PJI free survivorship (Kaplan–Meier analysis) of 97.1% at a mean of 5 years (range 1–9, SD \pm 2.5 years). Of the 68 knees, 4 underwent re-operations (5.9%), 2 of which were revision for PJI (2.9%), 1 revision for aseptic loosening (1.5%), and 1 open arthrolysis for arthrofibrosis (1.5%). Complications were recorded in 15 patients (22%), including wound healing (2 patients, 2.9%), post-operative hematoma requiring an arthrocentesis (2 patients, 2.9%), and a temporary nerve palsy (1 patient, 1.5%). The remaining 10 patients experienced non-surgical complications such as pneumonia or electrolyte imbalance.

Most studies also report on PROMs, such as Knee Society Score (KSS), Oxford Knee Score (OKS), Western Ontario and McMaster University Index (WOMAC) and Hospital for Special Surgery (HSS), as well as ROM, Tables 31.4 and 31.5. Commonly, significant improvement in PROMs and ROM is reported between pre-operative and last follow-up measurements and highlights benefit in pain and function that

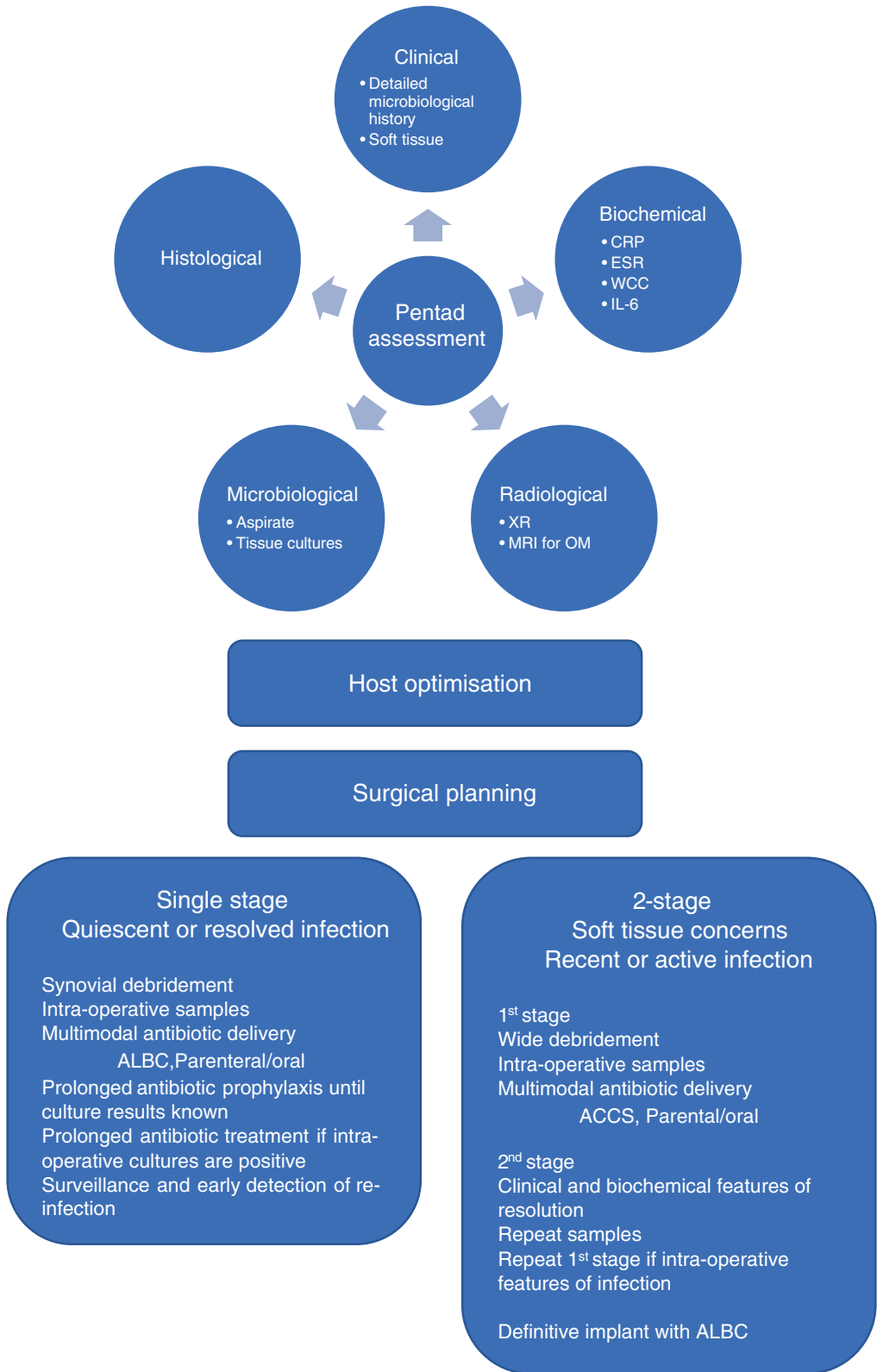


Fig. 31.2 Diagrammatic representation of authors' preferred approach

patients with infection-related knee arthrosis gain from reconstruction.

Reported results for TKA in the context of previous TB-related knee infections have similar variability between studies and approaches with PJI rates that range from 0 to 31%. However, the total number of patients (knees) being 71 (75) between 6 studies, challenges drawing clear conclusions. Despite the previous, it must be noted that low rates of reactivation and good clinical outcomes even in the treatment of active or recently diagnosed TB have been reported where antimycobacterial treatment is initiated prior to PJI in addition to prolonged post-operative therapy [37, 46, 47], and thorough surgical debridement is performed [44]. Additionally, reactivation of TB with a TKA in situ can be successfully treated with antimycobacterial therapy alone in many cases. Improvements in PROMs and ROM also support the notion that patients benefit greatly from reconstruction when indicated [28].

31.7 Conclusion

Successful eradication of native knee infection may require the excision of articular cartilage and subchondral bone. Alternatively, chronic or virulent acute infections may lead to extensive chondrolysis. Although arthroplasty has historically been viewed as being contraindicated in such situations, it is increasingly being used as an alternative to arthrodesis to improve patient functional outcomes. Although technically demanding and carrying greater risk, successful outcomes can be achieved by adherence to a thorough algorithm, emphasising the importance of contributions from a multi-disciplinary team.

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The Place of Antibiotic-Loaded Cement in TKA Infection

32

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and Jean Philippe Vivona

32.1 Introduction

Rate of infection after knee arthroplasty is depending on publications from 1–5% to 6% after TKA revision [1–5]. Primary TKA procedures are 2–3 times less susceptible to infection than TKA revisions [6, 7].

It has been reported in previous chapters.

After surgery, the inner bony tissue interacts with the biomaterial and forms a very thin biofilm as immune reaction toward the material [8]. If microorganisms reach the surface, they can adhere to it. Its persistence due to inflammation increases susceptibility to infection. And bacteria can themselves enhance the development of the biofilm. Surface roughness is of importance and cement is prone to bacteria adhesion [9, 10].

Conversely, the use of cement as a carrier for topical delivery for antibiotics has been initially described by Buchholz and Englebrecht using gentaline in Palacos resin [11]. It dramatically developed during last three decades in orthopedic field.

It is hypothesized that antibiotic-loaded cement (ABLC) help to treat local infections and

will result in lower infection rates during primary and/or revision TKA surgery.

32.1.1 Bacteriology

Most prosthetic joint infections (PJI) involve gram-positive organisms (*Staphylococcus aureus*, *Staphylococcus epidermidis*, or group B streptococcus) or gram-negative bacteria.

Chiu reported that those organisms identified through culture in revision infections are more virulent and less sensitive to antibiotics than those found in primary TKA infections [12].

To be suitable for use in bone cement antibiotics might be bactericidal for these bacteria with minimal risks of side effect especially allergy. It must be water-soluble and thermally stable during exothermic polymerization [13]. Consequently, the most commonly used antibiotics for ABLC (including spacers) reported in the literature are gentamicin, tobramycin, vancomycin, and cephalosporins.

Elution of antibiotic from cement used for spacers allows the local delivery of antibiotics toward the infected bone and soft tissue at high concentrations, much higher than can be achieved by intravenous or oral routes late 4 months locally after implantation [14–16]. It has been shown that at least 3.6 g of antibiotic per 40 g of acrylic cement is desirable for effective elution kinetics and sustained therapeutic levels of antibiotic [17].

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Doses as high as 6 to 8 g of antibiotic per 40 g cement, when ABLC is used in the form of beads or spacers, have been shown to be safe clinically [18].

32.1.2 Mechanics

Mixing antibiotic to cement allows elution of antibiotic but modifies mechanical properties. Compressive and tensile strengths of ABLC decrease with the quantity of antibiotic powder (maximum 2 g/40 g of cement). Using liquid or powder forms change ratio elution: liquid is more detrimental to mechanical properties [19]. Polymerization forms bubbles depending on cement and temperature: some escape from cement, others are sealed in. Antibiotic is released from the surface of the cement and from cracks and voids within the cement.

Gradual diffusion of antibiotics into surrounding tissues over time depends on cement porosity. This is related to the type of antibiotic, the composition of cement (viscosity) and preparation (e.g., vacuum mixing devices minimize porosity).

So, the mechanical and elution properties of commercially available premixed ABLC products seem superior to those of hand-mixed preparations [10, 20–22] but controversial [23].

At last, the potential risk of selection of antibiotic-resistant strains of bacteria was not confirmed [24].

32.1.3 Place of ABLC in the Spectrum of Prosthetic Joint Infection (PJI)

After TKA, deep PJI occur in an early to moderate time period after the operation, while none of the studies reported chronic deep infection to be the most common type of infection [13].

ABLC are useful in early PJI (local beads) and in late chronic PJI in one step or two steps (spacers) revision.

They are used too to prevent infection in primary TKA and revision of aseptic TKA.

32.2 ALBC in Primary TKA

To prevent infection in primary TKA is controversial using an ABLC depends on cultural practices. It is largely used in the UK, Nordic countries, and Australia.

Initially proposed in 2004 by Bourne [25] or Hanssen [26] to mimic Nordic registries which was effective to decrease the rate of PJI after total hip arthroplasty.

At the knee level, Jameson reporting 731,214 cases from the UK registry note less revision (aseptic or PJI) in comparison with plain cement at 10 years [27]; Jansen with the finish registry about 43,149 primary TKA as well [28].

But the other publications support the conclusion that ALBC could not prevent deep infection after primary TKA.

The results at 2 years follow-up after index surgery is a good threshold for analysis concerning infection as is linked to end of infection risk and mechanical changes for cement [29].

Bohm found no difference regarding the revision rate for infection nor any other cause (comorbidities included) [30].

Namba about 22,889 cases found no difference in the rate of deep infection between TKA with plain cement and those with ABLC, but a nonsignificant trend of higher proportion of aseptic loosening for the second group without risk factors [31].

Gandhi reached the same conclusion for 1625 TKA from a monocentric study [32].

A prospective randomized study by Hinarejos compared the rates of deep infection of ALBC versus plain cement for 2948 cases [33]. The use of erythromycin and colistin ALBC did not lead to a decrease in the rate of infection after primary TKA when systemic prophylactic antibiotics were used.

This is in accordance with other authors [34, 35], the result of the Australian Orthopaedic Association registry and two recent meta-analysis from Schiavone-Pani or Kleppel [13, 36, 37].

If some report did not found risk factors in large series [31, 38], numerous authors recommend the use of ABLC in patient at higher risk of infection: diabetic, immunocompromised, mor-

bidly obese, patients with previous history of fracture, contamination and/or infection of the knee [9, 10, 12, 29].

Lee includes thyroid, heart, or lung diseases [39].

Paucity of randomized clinical trials and the literature are not sufficient to confirm the reduction of the risk of infection with the use of ALBC in primary TKA.

It leads us to recommend with caution the use of ALBC only in high-risk patients.

In addition, culture from fluid or tissue at revision TKA can alter reliability of result when ABLC has been used at index surgery [40].

In primary TKA, the potential economic impact using ALBC is not valid [41, 42].

32.3 ALBC in Aseptic Revision of TKA

The occurrence of periprosthetic infection is 2 to 3 times higher in revision TKA than in primary THA/TKA [7].

The risk of infection following aseptic resurgery decreases by 6% at 89 months when using a vanco manually mixed ABLC [12]. In this level 1 prospective randomized study including 183 cases, local antibiotic delivery is very effective against infection with no infection in the ABLC group.

A retrospective study about 1154 re-TKA presents a high risk of failure (10%) and ABLC decreases by 50% the risk of all cause of re-revision [1].

The Kleppel's meta-analysis [13] reported two studies: at 62.5 months follow-up, the secondary infection rate is significantly lower with ALBC.

So the use of ALBC is clearly recommended in case of revision of TKA without sepsis. The effect of release of antibiotic is mainly at initial postoperative period [13].

Commercial gentamicin or tobramycin-LBC provide sufficient concentrations to be bactericidal even against methicillin-resistant organisms. The use of vancomycin should be considered in revisions following primary TKA in which gentamicin or tobramycin-loaded bone cement had been used because of the risk of gentamicin resistance [10].

32.4 Place of ABLC in Resurgery for PJI

32.4.1 Surgery in One Step

It concerns especially the early infection after primary TKA.

DAIR before 4 weeks can be effective in acute PJI. The procedure includes debridement, multiple bacteriological harvesting, change of PE. It is modified by addition of antibiotic (vancomycin) impregnated cement beads [8].

Small comparative studies [43–45] in combination with parenteral IV antibiotherapy can get 78% of healing in early cases. But with a higher risk of septic recurrence if the cement at index surgery was plain which is frequently the case [28].

32.4.2 Surgery in Two Steps

In case of late infection, the modified DAIR presents a high failure rate.

The treatment is first to implant a massive spacer to achieve high intra-articular concentrations of antibiotics while preserving joint space.

The final goal is to insert a new TKA once the infection has been healed.

The first step is to remove implants, *cement*, and debride the tissue. At this stage, the debridement is more important than the type of spacer or the additional antibiotic amount. Once the joint is cleaned, a spacer can be inserted.

The spacer is an ABLC which can be premolded by companies or molded peroperatively [46, 47].

Local antibiotic delivery via cement mixtures has been shown to achieve high local concentrations of antibiotic able to effectively treat the local bacterial burden [47].

But few premixed cements are available with a limited dose of antibiotic (1–2 g/40 g of cement). It needs to add antibiotic powder to reach high doses of antibiotics up to 8 g/40 mg.

The choice of antibiotic depends on individualized microbiological aspect (virulence of germ) and is often based on multiple preoperative punctures.

Surgeon creates his own antibiotic cement mixture peroperatively, producing a variable final

product [47]. He can also ask manufacturer to prepare a personalized spacer [48].

Vancomycin may be added to ABLC which already includes gentamicin or tobramycin [10]. Other antibiotic can be added in specific cases (e.g., *ceftazidime for Pseudomonas aeruginosa*) [49]. Cephalosporins are not effective against methicillin-resistant organisms.

The local treatment is always combined with parenteral antibiotherapy.

Spacer induces a very low risk of nephrotoxicity due to systemic absorption up to 8 weeks [50–53]. Some cases have been reported [53–56] to justify monitoring when inserted especially in patients with bad renal function.

Mechanically, high antibiotics dose does not influence the properties of cement as the spacer is used temporarily. However, virulence of the infection and delay to resurgery can influence quality of bone and soft tissue [46].

It can be static or mobile. Mobile is supposed to get a better post op mobility after re-implantation [57]. Described initially by Scott, Chang use former TKA (autoclaved peroperatively after explantation with high dose of ABLC) as a spacer before to change it later [43, 58].

Nozdo [47] compared 140 different mobile spacers (prefabricated, two separate cement spacers molded by surgeon, tibial spacer, and femoral autoclaved implant) without difference between them.

Struelens reported fractures, dislocations, and knee subluxation with mobile spacers [59].

Finally, the mobile spacers did not confirm their efficacy neither in terms of mobility [60] nor in mid-term functions [46, 61].

The second and final step is to remove the spacer once the infection is healed and to reimplant a new arthroplasty with medium dose ALBC.

- (a) beads with modified DAIR in early infection
- (b) spacer for re-TKA in two steps.

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32.5 Guidelines

Use of ALBC is recommended:

1. As prophylaxy (low dose of antibiotic)
 - (a) in primary TKA for high-risk patients only
 - (b) in TKA aseptic revision
2. As treatment in PJI (high dose of antibiotic)

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Operating Room Methods to Reduce Infection in Total Knee Arthroplasty

33

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33.1 Introduction

This chapter presents the current evidence-based measures to maintain a clean environment in the operating room, including the most recent guidelines, with the goal to minimize surgical site infections (SSI) after total knee arthroplasty (TKA).

SSI is one of the most common postoperative adverse events with associated patient morbidity and healthcare costs. Hospital-acquired infections (HAIs) are a leading cause of death in the United States, with an estimated 99,000 deaths occurring because of HAI [1]. Over 20% of HAIs are classified as SSI [2]. SSI is associated with increased length of stay, higher costs, and significant morbidity for the patient. In the era of cost control, reducing SSI is imperative for patient safety and optimal utilization of limited resources.

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Prevention efforts should target all surgical procedures, especially those in which the human and financial burden is most significant. In 2020, primary TKA will account for approximately 1.07 million arthroplasty procedures (primary and revision) performed in the United States, followed by approximately 500,000 total hip arthroplasties (THAs) [3]. Primary shoulder, elbow, and ankle arthroplasties are much less common. By 2030, prosthetic joint arthroplasties are projected to increase to 3.8 million procedures per year [4]. Infection is the most common indication for revision in TKA and the third most common in THA [5]. By 2030, the infection risk for TKA and THA is expected to increase from 2.2% to 6.8% and 6.5%, respectively [4, 6]. In addition, owing to increasing risk and the number of individuals undergoing prosthetic joint arthroplasty procedures, the total number of hip and knee prosthetic joint infections is projected to increase to 221,500 cases per year by 2030, at the cost of more than \$1.62 billion [4, 6].

The operating room represents the patient's environment most vulnerable to infection as the patient's physical barrier is weakened, and autoregulatory functions (e.g., control of body temperature) are disabled. Therefore, it is of paramount importance to reduce the patient's exposure to pathogens in the operating room. Surgical equipment, anesthesia, and operating room personnel impact the risk of SSI.

The following sections of the chapter will address in a systematic fashion how surgical equipment, anesthesia, and surgical personnel can reduce the risk of SSI in TKA.

33.2 Surgical Equipment Considerations

The term equipment is defined in this chapter as resources provided to the surgical staff to perform a TKA. This includes the operating room's airflow technology, body exhaust suits, adhesive drapes, and single-use instrumentation.

33.3 Operating Room Airflow Technology

In 1969 Sir John Charnley, a pioneer of THA, began to focus on preventing infection through air quality control [7]. He pioneered a purpose-built ultraclean laminar airflow (LAF) enclosure that functioned as a separate "room within a room." In combination with occlusive operating exhaust gowns, he reduced his periprosthetic joint infection rate from 9.5% to 0.5% [8]. As LAF gradually became a culturally accepted standard, further evidence supported its use [9]. This multi-center randomized controlled trial involved 8055 hip and knee replacements and compared LAF systems to conventional theaters and body exhaust suits to conventional clothing. Vertical LAF and body exhaust suits were associated with the lowest rate of PJI (0.1%) and the lowest bacterial air count (0.4 bacteria carrying particles per m³). When prophylactic antibiotic agents were used with LAF, the rate of PJI reduced from 3.4% to 0.3%. This study provided strong evidence supporting LAF systems. Further evaluation of 3175 arthroplasties revealed a reduced PJI rate after THA (2.0% to 1.2%) but an increased PJI rate after TKA (1.9% to 3.9%) with horizontal LAF [10]. A sub-analysis of confounding factors such as age, diagnosis, comorbidity, surgeon experience, and duration of surgery could not explain this difference.

More recently, there has been an increasing body of registry-based evidence that disputes the clinical efficacy of LAF. An analysis of 41,212 THAs and 20,554 TKAs from the German nosocomial infection surveillance system determined that hospitals with LAF systems had similar PJI rates to hospitals with conventional ventilation [11]. An interrogation of 51,485 THAs and 36,826 TKAs from the New Zealand Joint Registry found a substantially higher rate of early revision for PJI when procedures were performed with LAF compared with a conventional theater (0.15% vs. 0.06%) [12]. A more recent analysis of 91,585 THAs from the same registry observed nearly a twofold risk of revision for PJI within 6 months with LAF compared with conventional ventilation [13]. Data from the UK National Joint Registry has also been used to analyze the effect of LAF. In comparative series of 4915 THAs and 5928 TKAs, there was no substantial difference between LAF and conventional ventilation in the rate of SSI (0.92% vs. 1.14%, respectively) or revision for infection (0.53% vs. 0.45%, respectively) [14]. A meta-analysis of 196,819 hip or knee replacements revealed a higher risk of SSI with LAF (relative risks of 1.71 and 1.36 for THA and TKA, respectively) compared with conventional ventilation. A more recent meta-analysis of 12 studies consisting of 330,146 THAs and 134,368 TKAs showed no substantial difference in PJI risk for LAF compared with conventional ventilation (odds ratio 1.29 and 1.08 for THA and TKA, respectively) [15]. Based on these studies, the World Health Organization has now advised against the use of LAF to reduce the risk of SSI for patients undergoing arthroplasty surgery [16]. More recently, an International Consensus Meeting on PJI has also agreed that LAF is unnecessary for elective joint arthroplasty surgery [17].

33.4 Body Exhaust Suits

Body exhaust suits are commonly used during arthroplasty procedures. However, their role in reducing SSI and PJI is controversial. While negative-pressure body exhaust suits led to less

air contamination, less wound contamination, and fewer PJI, positive-pressure body exhaust suits or surgical helmet systems were not shown to reduce contamination or deep infection during arthroplasty [18]. A fundamental principle of negative-pressure exhaust suits uses aspiration tubing to create negative pressure inside the suit, which removes shed particles from the surgical field [19]. The application of negative-pressure body exhaust suits, combined with ultraclean operating rooms, has reduced the infection rate from 1.5% to 0.6% [9]. These results, published in 1982, have led to a widespread acceptance of negative-pressure body exhaust suits as a means of reducing SSIs and PJIs.

However, exhaust tubing is not practical during surgery [20]. Consequently, portable surgical helmet systems were introduced in the 1990s. The surgical helmet system creates a positive-pressure environment inside the gown by drawing air through the hood material, using the material as a filter, and blowing air across the surgeon's face and neck. In contrast to the negative-pressure body exhaust suits, the positive-pressure surgical helmet systems currently in use did not demonstrate a reduction in SSI or PJI and have been associated with a paradoxical increase in deep infection rates compared to standard sterile gowning [18, 21]. Despite being widely used, the current evidence does not support surgical helmet systems to reduce SSI or PJI [18].

33.5 Adhesive Drapes

Adhesive plastic drapes, either plain or saturated with an antimicrobial agent (mostly an iodophor), are used on the patient's skin after completing the surgical site preparation. The drape adheres to the skin, and the surgeon cuts through the skin and the drape itself [22]. Such a drape is theoretically believed to represent a mechanical and microbial barrier to reduce the microorganisms' migration from the skin to the operative site [23]. However, some reports have shown an increased recolonization of the skin following antiseptic preparation underneath adhesive drapes compared to no drapes [24].

A Cochrane review and its updates on the effect of adhesive drapes to reduce SSI concluded that there is no evidence that adhesive drapes reduce SSI [22]. No recommendation is available on using sterile disposable or reusable drapes and surgical gowns for SSI prevention.

Based on current evidence, the WHO suggests not to use adhesive drapes with or without antimicrobial properties to prevent SSI.

33.6 Single-Use Instruments

The introduction of single-use sterile prepackaged instrumentation is a strategy growing in interest to reduce cost, infection, and improve quality and efficiency. The risk of SSI could potentially be reduced through single-use instrumentation compared to traditional, reusable instruments. Reusable instrumentation could become contaminated following re-sterilization, before use in surgery, and may be associated with SSI [25–27]. No evidence is currently available demonstrating a reduction in SSI or PJI with single-use instruments. However, clinical data suggests a reduction in SSI as surgical field contamination is reduced when performing a TKA with single-use instruments in comparison to reusable instruments [28].

33.7 Anesthesia Considerations

Anesthesia staff has the means to modulate the risk of SSI and PJI in the operating room by administering either general or regional anesthesia, by controlling the patient's body temperature, tissue oxygenation, metabolism, and by maintaining a clean work environment to reduce contamination of the surgical field.

33.8 General Vs. Regional Anesthesia

Regional anesthesia in TKA is claimed to decrease the incidence of deep-vein thrombosis and pulmonary embolism and reduce intraopera-

tive bleeding, the need for transfusion, and the length of hospital stay [29–31]. However, spinal and epidural anesthesia and analgesia may cause hypotension, motor blockade, urinary retention, and pruritus [32]. Despite refinements made to reduce such complications, there is still the potential for inadvertent dural puncture and neurological injury, making these techniques less acceptable [33].

The risk of SSI is slightly reduced with regional vs. general anesthesia in patients undergoing TKA [34, 35]. However, none of the current guidelines recommend either for or against a specific mode of anesthesia.

33.9 Maintaining Normothermia

The body normally maintains its temperature between 36 °C and 38 °C by balancing heat production and heat loss to maintain a state of normothermia [36, 37]. These functions are controlled by the thermoregulatory systems in the central nervous system [37]. The body loses heat through radiation, conduction, evaporation, and convection. In surgery, this may occur via the normal process of heat moving away from the body (radiation), contact with cool operating surfaces (conduction), respiration (evaporation), and exposure to the airflow through the operating room (convection). The central nervous system is disrupted under general and regional anesthesia, and the body's thermoregulatory systems are unable to function appropriately; thus, both hypothermia, a core temperature < 36 °C, and hyperthermia, a core temperature > 38 °C, may occur during surgical procedures [37].

Hypothermia can lead to greater susceptibility to infections [38]. This effect can be considerable; a decrease of 1.9 °C in core temperature triples the relative risk of surgical periprosthetic infection and increases the hospitalization duration by 20%. Hypothermia increases patients' susceptibility to perioperative wound infections by causing vasoconstriction and impaired immunity [39]. The presence of sufficient intraopera-

tive hypothermia triggers thermoregulatory vasoconstriction, decreasing the partial pressure of oxygen in tissues, thus lowering resistance to infection [40, 41].

Although it is often assumed that hypothermia is a result of lengthy procedures, the greatest temperature drops may, in fact, occur before surgery and during induction of anesthesia [42]. The administration of inhaled gases (e.g., isoflurane, sevoflurane, or nitrous oxide), or the use of intravenous anesthetic induction agents (e.g., propofol or opioids), cause peripheral vasodilation and cause the transfer of core body heat to the periphery [43]. Once hypothermia occurs, it is difficult to correct efficiently [42]. The type of anesthesia, spinal vs. general, has no effect on thermoregulation in patients undergoing TKA [42]. The patient's body temperature before and during TKA can drop by 1.5 °C [42].

Maintaining normothermia during TKA is an important step for reducing the risk of SSI. The means of achieving this goal include the use of pre- and intraoperative warming devices and the administration of pre-warmed intravenous fluids [44]. However, the best warming device to ensure normothermia remains unknown. Concerns regarding the use of air warmer and the potential for contamination have been raised by a few authors, although this has not been proven [45, 46].

33.10 Perioperative Oxygenation

There is evidence that optimized blood flow to the surgical incision decreases SSI rates by avoiding hypothermia, hypoxia, and decreased perfusion [47]. This has also been demonstrated for tourniquet use [48, 49]. Because inadequate tissue oxygen tension impairs tissue repair and oxidative killing of surgical pathogens, supplemental oxygen is another important mechanism for SSI reduction [50]. The WHO recognizes the importance of sufficient oxygenation to the surgical incision and recommends 80% fractions of inspired oxygen concentration during surgery and in the postoperative recovery unit [51].

33.11 Glycemic Control

Between 8% and 22% of patients who undergo total joint replacement have diabetes, and about one-third have undiagnosed hyperglycemia [52, 53]. Diabetes, especially when uncontrolled, is a significant risk factor for SSI [54]. Even non-diabetic patients who develop hyperglycemia postoperatively have a significantly increased risk of SSI, with current recommendations that peri- and postoperative glucose levels are strictly monitored and maintained <180 mg/dL [55]. The identification of patients with diabetes or hyperglycemia and the implementation of strict perioperative glycemic control minimizes the risk of infection [56]. Maintaining a blood glucose level < 180 mg/dL might be important even after the patient is discharged. With the trend towards outpatient TKA, the glycemic control adds another task to the patient's daily routine of home recovery. Even non-diabetic patients, who are not familiar with blood sugar testing, should be trained with this task before discharge.

33.12 Maintaining a Clean Work Environment

The Society for Healthcare Epidemiology of America (SHEA) established guidelines for anesthesia personnel to maintain a clean work environment with the purpose to reduce SSI by microbial cross-transmission from the anesthesia work area in the intraoperative environment [57]: Hand hygiene ideally should be performed according to the WHO 5 Moments for Hand Hygiene. These 5 Moments of Hand Hygiene are:

1. before touching a patient,
2. before clean/aseptic procedures,
3. after body fluid exposure/risk,
4. after touching a patient,
5. after touching the patient's surroundings.

SHEA recommends that hand hygiene be performed at the minimum before aseptic tasks (e.g., inserting central venous catheters, inserting arte-

rial catheters, drawing medications, spiking IV bags); after removing gloves; when hands are soiled or contaminated (e.g., oropharyngeal secretions); before touching the contents of the anesthesia cart; and when entering and exiting the OR (even after removing gloves). These recommendations could mean that anesthesia providers have to perform hand hygiene up to 54 times per hour [58]. The current failure to perform hand hygiene for all anesthesia providers is 82% [58]. To facilitate hand hygiene, SHEA recommends positioning alcohol-based hand rub dispensers at the entrances to the operating room and near anesthesia providers inside the operating room.

To further reduce the risk of contamination in the OR, anesthesia providers should wear double gloves during airway management and remove the outer gloves immediately after airway manipulation. As soon as possible, providers should remove the inner gloves and perform hand hygiene.

33.13 Surgical Personnel

This section addresses methods the surgical personnel can apply to reduce the risk of wound contamination, SSI, and PJI. Intraoperative methods applied by the surgical team to reduce these risks are discussed in a different chapter.

33.14 Operating Room Traffic

Operating room (OR) traffic flow is linked to increased contamination rates [59]. It is hypothesized that OR personnel are the major contributors to OR contamination because of bacterial shedding, mainly from the skin, with shedding rates as high as 10,000 bacteria per minute [60–62]. The bacterial load in the OR is directly associated with the number of members present, with a 34-fold increase in bacterial counts with five OR personnel compared with an empty room [63, 64]. Also, increased personnel traffic implies more door openings, which is associated with increased bacterial counts [65]. Any obstacle in

the directional airflow path coupled with repeated door openings creates air turbulence that allows the mixing of filtered air with unclean air [10, 61, 66]. The average frequency of door openings during a primary arthroplasty is 0.65/min [67]. The most common OR personnel entering and leaving the OR are the circulating nurses and surgical equipment representatives. For almost 50% of the room entries, no identifiable purpose could be identified. Hence, most OR personnel traffic is extraneous and can be reduced. Ideally, the anticipated implants are in the OR prior to the start of the case, and the exchange of anesthesia and scrub technician personnel is limited.

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34.1 Introduction

34.1.1 Definition of Tourniquet

It is essentially an external device that applies pressure to a limb, causing constriction of blood flow, therefore creating better visualising of the surgical field and it reduces blood loss.

34.1.2 History of Tourniquet

The use of tourniquet dates back as far as the era of Alexandra The Great's military empire around the fourth century. It was not until 1718 when Jean-Louis Petit popularised its use in the field of surgery. As an anatomist as well as a surgeon, he made a respectable contribution to the advancement of Trauma and Orthopaedic surgery [1]. He made a screw device which by turning constricted the blood flow. Hence it was named tourniquet, which comes from the French word 'tourner = to

turn'. Over the centuries, his design evolved resulting in the different types of tourniquets that are in use today.

34.1.3 Tourniquet Use in Total Knee Arthroplasty (TKA)

The use of tourniquet in surgery has helped surgeons for centuries in minimising the amount of blood loss. Over the last several decades, the use of tourniquet has become controversial in lower limb arthroplasty. There is no conclusive evidence in the published literature, whether tourniquet should be routinely used in TKA. In orthopaedics currently, there are three schools of thought with regard to tourniquet usage; (1) use it for the entire case, (2) use in part (for cementing), (3) do not use it all. This chapter will cover essential topics regarding tourniquet usage and will include the relevant literature as well as focusing on infections.

34.1.4 Indications and Contraindications of Tourniquet Use in Surgery

Generally, it is indicated for procedures in which there is a likelihood of high blood loss or in delicate surgery in which the relevant anatomy needs to be identified during the approach.

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The main contraindication to tourniquet use is in patients with significant peripheral vascular disease. There are other situations in which tourniquet use may not be advised such as in some open injuries, intramedullary nailing and diabetic foot disease [2].

34.1.5 What Is Normal Tourniquet Pressure?

There is no universally agreed value to inflate the tourniquet pressure to although the vast majority would support using the lowest tourniquet pressure for the shortest period time if one has to use it. Several factors can influence the value to inflate above the systolic pressure. Some authors advocate inflating tourniquets to 2.5 times above the pre-anaesthetic resting systolic blood pressure and increasing further by 50 mmHg for obese patients [2]. A randomised control trial (RCT) compared two groups using tourniquet during TKA. One group had higher tourniquet pressure of 350 mmHg (set as standard), and the group had a lower tourniquet pressure set to 100mmHG above the patients' systolic pressure. The study consisted of 26 patients undergoing bilateral TKAs. They found using the lower tourniquet pressure (100 mmHg above the systolic) was as acceptable in controlling bleeding as the higher pressure (350 mmHg) group. Also, the lower tourniquet pressure group had less pain and faster recovery [3].

34.2 Tourniquet Use in TKA: Does It Influence the Risk of Infection?

34.2.1 Pathophysiology of Tourniquet Ischaemia, Wound Healing and Tourniquet Time

Healthy wound healing is dependent on adequate local cellular oxygenation, which in turn will reduce the risk of delayed wound healing or

infections [4]. Johnson in 1993 described it well in their study on wound healing following TKA. It explained the resultant wound ischaemia following the TKA. The wound edge oxygenation varied in the early postoperative period, and type of incision used (midline, medial, curved medial) had an influence on it [5]. Once a tourniquet is applied, around 1% of the normal circulation is perfusing the limb [5]. Furthermore, the paper pointed out that exsanguinating the limb prior to applying the tourniquet favoured any potentially opportunistic microbes present for two reasons. Firstly, once a tourniquet is applied, it leads to reduced circulatory defensive cells to kill off any bacteria that may be present. Secondly, if the antibiotics are not administered on time, it may result in inadequate therapeutic levels within the tissue. Possibly a combination of these two factors may contribute to the development of superficial wound healing problems or worse a deep infection.

There is a difference in skin edge perfusion with knee going into flexion in the early periods following TKA [6]. The lateral wound edge appears to have lower oxygen concentration compared to the medial edge wound [5] although this can be resolved by providing the patients with extra oxygen [7]. A large RCT found giving supplementary oxygen to patients undergoing colorectal resection halved their wound infection rate [8]. Given some of these factors, one might ask why to use the tourniquet if one's aim is to try and mitigate any risks of infection?

Tourniquet usage can result in ischaemia-related events. Clarke et al. looked at 31 patients undergoing TKA and divided them into groups: no tourniquet and two other groups in which tourniquet was inflated 125 mmHg or 250 mmHg above the mean anaesthetic systolic pressure. In this small series, they found all the groups had an element of critical hypoxia and this was most significant in the higher tourniquet pressure group. Also, in this group the duration of hypoxia persisted longer when compared to lower tourniquet pressure and the no tourniquet group [9].

A meta-analysis on the use of pneumatic tourniquet in TKA looked at 3 studies out of 13

RCTs that mentioned infection in their analysis. There were in total 223 patients and the study stated infection rate was higher in the tourniquet group than the control group, with RR 5.37 (95% confident interval (CI) 0.99–29.6) and *P*-value of 0.05. [10] Although at a glance, their results may appear significant; however, it must be interpreted with caution due to the small number of studies. It is difficult with certainty whether operative time or tourniquet is contributing to the development of prosthetic joint infection (PJI). In the published literature, some surgeons use tourniquet time and operative time almost interchangeably. Increased operative time will subsequently lead to the application of tourniquet for a more extended period which as a result of the prolonged surgery elevates the risk of PJI. For example, in a case-control study Blanco et al. reviewed retrospectively 132 patients records who had TKA in which of all the cases used tourniquet. The authors included in the study 66 TKAs with PJI and 66 TKAs without PJI as control. They concluded, prolonged operative >90 min and tourniquet time >60 min were the most relevant risk factor for PJI [11]. Carefully analysing their findings, one can conclude they found the risk of PJI increased after 60 min of operating time, further increasing fivefold every 15 min after 90 min of operative time. Table 34.1 below shows a summary of relevant studies of tourniquet use in TKA and infections.

34.3 Tourniquet Use on the Rate of Blood Loss, Operative Time and Deep Vein Thrombosis (DVT)

There is no substitute for excellent haemostasis during exposure and wound closure. Blood loss continues after the closure of the wound and this can be more than the blood loss in surgery. A recent meta-analysis of 11 RCTs involving 541 knees looked at tourniquet use and blood loss in TKA. It concluded tourniquet use in TKA

reduced blood loss during surgery as well as calculated blood loss and operative time. However, there was no difference in postoperative total blood loss or the need for blood transfusion [15].

This study also confirmed previous findings that the risk of having the thromboembolic event was fivefold higher with the use of tourniquet compared to without a tourniquet [15, 16]. Furthermore another two meta-analyses commented on the incidence of DVT being higher with the use of tourniquet, with similar fivefold increase as the previous studies [10, 17].

34.3.1 Tourniquet use in TKA: Pain and Range of Movement (ROM)

Previously published studies including RCTs (Table 34.1) suggested the use of tourniquet in TKA resulted in an increase in pain in the immediate period after surgery and it also affected the ability to straight leg raise. On the other hand, Deering's et al. published an informative systematic review and a meta-analysis last year which included 14 studies and 8 of them discussed pain [18]. These 8 studies consisted of 440 TKA, (221 TG vs. 219 NTG) and analysis showed no clinically important difference between the two groups 5.23 ± 1.94 cm vs. 3.78 ± 1.61 cm; standardised [STD] mean difference 0.88 mm; 95% confidence interval [CI], 0.54–1.23; $p < 0.001$. The authors used the Visual Analogue Scale (VAS) 0 mm to 100 mm and defined 20 mm on the VAS score as the minimum clinically important difference (MCID). Danoff et al. observational study of 139 total hip arthroplasty (THA) and 161 TKA demonstrated the MCID for worsening pain to be 23.6 for THA and 29.1 for TKA [19].

Deerings et al. found no clinically significant difference between TG and TNG on the range of movement patients achieved post-surgery. In addition, their results demonstrated no difference in the length of hospital stay between the two groups.

Table 34.1 Summary of RCTs looking at the correlation between Tourniquet use in TKA and infections

| Authors and year | LoE | Aim of the study | No. cases | TG | NTG | Conclusion | Comments |
|----------------------------------|-----|---|-----------|----|-----|---|---|
| Liu et al. (2017) [12] | I | Prospective RCT assessing the benefits of the use of a tourniquet in on limb in patients undergoing simultaneous bilateral TKAs | 52 | 25 | 27 | Use of tourniquet may save operative time but could also lead to increased postoperative pain, swelling, delayed straight leg raise and wound complications | The only 1 infection (deep) was in the TG |
| Vandenbussche et al. (2002) [13] | I | Prospective RCT looking at the effects of tourniquet use in TKA | 80 | 40 | 40 | There were no significant differences between the two groups in terms of operative time, complications and length of hospital stay. NT group had more blood loss, better ROM until day 5, less pain for the first 6 h post-op | No wound infections |
| Clarke et al. (2001) [9] | I | RCT looking whether thigh tourniquet influenced wound hypoxia and was the cause of delay | 31 | 21 | 10 | If a tourniquet is used, then it should be inflated at the lowest possible pressure to minimise wound complications | The only 1 infection (2 skin flaps) was in the TG. There were 3 skin flaps (×2 TG and 1× NTG) with delayed wound healing |
| Abdel-Salam et al. (1995) [14] | I | Prospective RCT looking at the effects of a tourniquet on wound healing, pain and muscle function | 80 | 40 | 40 | The complication rate was lower in the group with NT, better initial recovery and less pain | Tourniquet group had 5 wound infections vs. no cases in the NTG |

LoE level of evidence, TKA total knee arthroplasty, NTG no tourniquet group, TG tourniquet group

34.4 Conclusions

Tourniquet use in TKA may lead to an increase in some complications following surgery. These include the incidence of wound infections, tourniquet pain, thromboembolic events, neurovascular injury and stiffness. On the other hand, proponents of tourniquet use argue that there is less blood loss, a shorter operative time and no clinically significant difference in overall outcome compared to those who have TKA without a tourniquet.

The current published Level 1 studies do not conclusively support whether tourniquet should be used routinely or not—in particular concerning the incidence of infection. Infection is a serious complication with associated high morbidity

and mortality. PJI have worse 5-year survival rate than some cancers. It is known that prolonged surgery time is associated with infection and by virtue, some authors report prolonged tourniquet time leads to infection. Until we have adequately powered level 1 studies, the current status quo will continue. In our practice, we do not routinely use a tourniquet and neither do we support its use in TKA.

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Intraarticular Injection Prior to Joint Replacement and its Relationship to Prosthetic Joint Infection

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35.1 Introduction

The management of symptomatic osteoarthritis (OA) of the knee includes well-described treatment options such as oral analgesia, non-steroidal anti-inflammatory medications, braces and sleeves, physiotherapy particularly strengthening exercises, dietary/lifestyle changes, intraarticular (IA) injection of therapeutic substances and when these modalities have failed to adequately control symptoms, arthroplasty is appropriate [1–5].

IA injections of the knee are routinely performed both in the outpatient clinic and operating room setting [3]. It has been reported that approximately 30% to 50% of patients undergoing arthroplasty have had an IA injection in the ipsilateral knee during the year leading up to the surgical procedure [6, 7]. However, compared to other nonoperative treatment modalities intraarticular injection procedure is relatively more invasive. Hence several investigators have reported the risks and complications associated with this procedure [8, 9]. Local complications such as painful effusion [10], skin lesions [11], skin necrosis [12] and septic arthritis [13–15]

have been described following intraarticular injection.

Progression of symptoms despite the nonoperative treatment options is an indication for arthroplasty in the form of either a unicompartmental or total knee replacement (TKR) [2]. Outcomes of TKR may be influenced by some of the initial interventions used in the management of knee OA [16–18]. In this context, some investigators have evaluated the influence of preoperative IA injections on subsequent joint replacement [19–21].

This chapter discusses the risk of deep prosthetic joint infection (PJI) in TKR emanating from the practice of preoperative IA injections used in the management of knee OA and addresses some of the key questions encountered by clinicians in the decision-making process.

These include:

1. What are the common types of therapeutic substances used in IA injections of knee for the management of knee OA?
2. Which are the patient/clinician/procedure related factors that may influence the risk of PJI in TKR following IA injection in the preoperative period?
3. What is the current evidence to guide the safe surgical management when TKR is indicated in a patient with knee OA and previous history of ipsilateral IA injection?

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35.2 Types of Therapeutic Substances Used in Intraarticular Injection

35.2.1 Corticosteroids

Methylprednisolone or triamcinolone are the most commonly used corticosteroids for intraarticular injections. Corticosteroids are typically combined with local anaesthetic such as lidocaine or bupivacaine [22]. Glucocorticoids reduce the proinflammatory effects of arachidonic acid by acting directly on nuclear steroid receptors which alter the synthesis of mRNA and proteins leading to changes in T-cell and B-cell functions, decreases in the levels of cytokines and enzymes, and inhibition of phospholipase A2 [23]. Corticosteroids reduce inflammation by altering B- and T-cell function as well as stimulating hyaluronic acid synthesis [22].

More recent studies have demonstrated that such changes in the pericellular matrix in which the chondrocytes are situated and to which they are adapted can have significant effects on the chondrocytes in terms of gene expression with subsequent effect on the anabolic/catabolic homeostatic balance [24]. This alteration in normal chondrocyte function may tip the balance from an anabolic to catabolic state with subsequent degenerative change seen in osteoarthritis [25]. In summary, IA corticosteroids may be useful in treating synoviocyte-mediated inflammation, thereby providing some symptom relief but have a negative effect on chondrocyte function.

In their guidelines, the American Academy of Orthopaedic Surgeons (AAOS) noted that given the inconclusive evidence they were not able to recommend for or against the use of intraarticular corticosteroids to treat knee OA [26]. However, an intraarticular steroid injection remains the first step in management for many patients after initial presentation to a surgeon.

35.2.2 Hyaluronic Acid (HA)

Hyaluronic Acid (HA) is a complex high molecular weight polysaccharide that is an essential component of proteoglycans found in synovial

fluid [27]. It has a molecular weight in normal synovial fluid ranging from 6500 to 10,900 kDa [28]. In the normal adult knee, HA concentration ranges from 2 to 4 mg per ml and has a half-life of 20 hours [29]. It increases the viscosity of IA fluid and entangles between collagen fibres to trap water, providing increased compressive strength to articular cartilage [22, 30]. HA decreases inflammation by reducing oxidative stress and inhibiting macrophage phagocytosis [31].

In comparison to normal knee joint, the half-life of intrasynovial HA is reduced (11 to 12 h) in degenerative conditions such as knee OA [32]. Furthermore, HA has been demonstrated to undergo depolymerisation with reduced molecular weight ranging from 2700 to 4500 kDa [28].

Exogenous HA injections are known by the marketing term “viscosupplementation” to describe a combination of the presumed dual effects of adding viscous material to the joint and supplementing the supply of hyaluronic acid. Currently, the commercially available HA preparations are typically administered in single-dose regimens, but most require three to five weekly injections [22, 33]. These products vary in their molecular weight, method of production, half-lives and cost [33]. They are produced either from harvested rooster combs or via bacterial fermentation *in vitro* [34]. Higher molecular weight HA (above 6000 kDa) is suggested to have greater clinical efficacy, but the current literature is inconclusive [31, 32, 35, 36].

It must be noted that the AAOS guidelines do not recommend the use of HA injections for knee OA [26, 37]. However, the Federal Drug Administration (FDA) in the United States has approved their use in the treatment of knee OA as a medical device as opposed to pharmaceutical agents [38]. Furthermore, their use is approved only in the select cohort of patients where knee OA symptoms have progressed despite simple oral analgesics or nonpharmacologic therapy [22, 38].

35.2.3 Biological/Novel Agents

Several biological and novel agents have been introduced in the recent years in the management

of knee OA with variable outcomes [39–45]. These include:

35.2.3.1 Platelet-Rich Plasma (PRP)

The rationale for PRP injection in knee OA is the direct introduction and subsequent utilisation of platelet-derived growth factors stored in the alpha granules of platelets to stimulate the natural healing cascade with regeneration of tissue and mediation of the anti-inflammatory response [40, 44, 45].

Several commercial systems are available which enable PRP preparation from the patient's venous blood, which is centrifuged to separate the platelets from red and white blood cells [42, 45, 46]. Several investigators have evaluated the clinical effectiveness of PRP in the management of knee OA [46–48]. Recent evidence has suggested that IA administration of leukocyte poor PRP in patients with knee OA can result in significant improvement in Western Ontario and McMaster Universities Osteoarthritis Index (WOMAC) scores compared to HA or placebo [49–51]. However, other investigators have reported no significant improvement with PRP treatment [52, 53]. The current AAOS guidelines do not recommend for or against the use of PRP for knee OA [26].

35.2.3.2 Mesenchymal Stem Cells (MSC)

Mesenchymal stem cells (MSC) derived from bone marrow, adipose tissue and amnion have been described in the recent literature as biological modalities useful in the management of patients with knee OA [42, 43, 54, 55]. Whilst the literature regarding these treatment options is rapidly evolving, they are performed predominantly in specialist centres [55–57].

The mechanism(s) through which MSC work in knee OA remains an area of ongoing research [57, 58]. Some investigators have suggested that IA injections of MSC may act via a combination of direct differentiation into chondrocytes, expression of appropriate extracellular matrix (ECM) proteins, and secretion of growth factors and cytokines that suppress inflammatory cell activation and stimulate tissue repair [55–57].

35.2.3.3 Autologous Cell Free Preparations (ACS/APS)

Two common forms of autologous cell free preparations used in the management of knee OA include autologous conditioned serum (ACS) and autologous protein serum (APS) [39, 59]. Currently, several commercially available kits are utilised to prepare ACS/APS from whole-blood samples of patients [60–63]. The obtained blood sample is further conditioned by incubation with glass beads and centrifugation, leading to an increase in the production of IL-1Ra as well as multiple other cytokines and growth factors [63, 64]. Broadly, both ACS and APS are injectable solutions enriched in endogenous cytokines which may help to restore joint homeostasis preventing degenerative changes in cartilage and bone [39, 59].

35.3 Current Evidence

35.3.1 Literature Search and Databases

A literature search of all the available evidence was undertaken (March 2020) using the healthcare database website (<http://www.library.nhs.uk/hdas>). The databases searched were Medline, CINAHL, Embase and the Cochrane library. Medline, CINAHL and Embase search was performed using Boolean statements and the wildcard symbol (*). A review of the Cochrane database for relevant articles was performed. An adjunctive bibliography search was undertaken to identify additional relevant studies through review articles, Google scholar (<https://scholar.google.co.uk/>) and the grey literature database—OpenGrey [65].

35.3.2 Search Criteria

Corticosteroids: The search criteria included “knee* AND (replacement* OR arthroplasty*) AND (intraarticular* OR joint*) AND inject* AND (steroid* OR corticosteroid*)”.

Viscosupplementation: The search criteria included “knee* AND (replacement* OR arthroplasty*) AND (intraarticular* OR joint*) AND inject* AND (hyaluronic acid* OR viscosupplement*)”.

Biological/novel agents: The search criteria included “knee* AND (replacement* OR arthroplasty*) AND (intraarticular* OR joint*) AND inject* AND [(platelet rich plasma* OR mesenchymal stem cells* OR autologous conditioned serum* OR autologous protein solution*)]”.

35.3.3 Search Results

A brief survey of studies in the current literature is presented in Tables 35.1, 35.2 and 35.3.

Interestingly, despite the considerable amount of literature available regarding the biological/novel agents used in the management of knee OA, the current comprehensive search did not identify any study addressing the topic of PJI risk in TKR following their use in the preoperative period.

35.4 Pathogenesis of PJI in TKR Following IA Injection

Whilst the precise mechanism whereby an IA injection leads to PJI following TKR is unknown, there are a number of theories. It has been suggested that some of the depot preparations of corticosteroids may not dissolve completely and can remain trapped within the soft tissue areas of cystic degeneration [21]. These remnants may be reactivated during surgery and lead to risk of wound complications or PJI [21]. Other investigators have reported that skin bacteria may be introduced at the time of IA injection given the significant variations in the skin preparation and the aseptic technique used [66–69]. This view is supported by investigators who used the “spent needle” culture method and noted that despite alcohol skin preparation 14% to 28% needle tips showed evidence of organisms [70]. These organisms may remain dormant and later be activated by the surgery.

There is consensus that PJI in TKR patients is the result of a multiple factors [71–74]. However, given the paucity of high quality studies coupled with the conflicting conclusions arrived by the different investigators, there remains a lack of detailed understanding regarding the aetiopathological mechanisms leading to PJI in TKR patients who have received an IA injection in the preoperative period [7, 75, 76].

35.4.1 Corticosteroids

As evident from Table 35.1, the published studies evaluating the potential risk of PJI in TKR following an IA corticosteroid injection are varied. Furthermore, their retrospective design limits the level evidence with majority being level III/IV studies. This has been highlighted by the systematic reviews and meta-analyses on the topic [7, 75, 76]. The overall reported rate in the literature of PJI in TKR is relatively low at approximately 1% [71, 77, 78]. Hence it has been suggested that the minimum number of patients required per cohort to demonstrate a 50% increase in infection rates following IA corticosteroid injection is 2000 [79]. All the studies except that by Richardson et al. [80] do not satisfy this criteria.

In an initial study on this topic, Papavasiliou and colleagues investigated the association between preoperative corticosteroid injection and increased risk of PJI in TKR [66]. This retrospective cohort study divided the 144 patients into two groups; one group had received IA corticosteroids prior to TKR and a control group who had not. The authors reported three PJI cases in the study group. The time from last injection to surgery for the three infected cases was 8, 10 and 11 months. However, the timing of injection for those patients who did not develop infection was not stated. In contrast to the injection group, no infections were documented in the control group who had not received steroids prior to TKR ($p < 0.025$). The authors concluded that preoperative IA corticosteroid injection given within the 11-month period prior to TKR significantly increased the risk of deep infection. This study

Table 35.1 Survey of studies evaluating risk of PJI in TKR following preoperative corticosteroid injection

| Study | Study design | Number of patients/TKR | Steroid (type and dose) | Place of injection | Follow-up after TKR (months) | PJI | Time between injection and TKR (months) | Odds ratio (95% CI) and p-value |
|----------------------------|----------------------------|---|--|----------------------|------------------------------|-------------------|---|---|
| Richardson et al. (2019) | Retrospective cohort | 16,656 patients 6653 5569 4434 | NR | NR | 6 | 3.25% | ≤3 >3-6 >6-12 | 1.21 (1.04-1.40), p = 0.01 1.07 (0.90-1.27), p = 0.41 1.17 (0.98-1.40), p = 0.08 |
| Kokubun et al. (2017) | Retrospective cohort | 442 TKR | Methylprednisolone 80 mg or Triamcinolone 40 mg | NR | 51.5 | 3.0% | ≤3 ≤6 | 1.09 (0.52-2.23), p = 0.80 1.47 (0.71-3.04), p = 0.29 |
| Khanuja et al. (2016) | Retrospective cohort | 302 patients | Triamcinolone 40 mg | Clinic: 302 | 42 | 2.0% | 5 | 0.5 ^a (0.12-1.98), p = 0.5 |
| Desai et al. (2009) | Retrospective cohort | 90 TKR | Methylprednisolone 40 mg | Clinic: 30 OR: 60 | 12 | 0 | ≤12 | NR |
| Horne et al. (2008) | Retrospective case-control | 40 cases 352 control | NR | Clinic: 40 | NR | 1/40 ^b | 16 | 1.38 (0.55-3.31), p = NR |
| Joshy et al. (2006) | Retrospective cohort | 32 TKR | NR | NR | NR | NR | 46 | NR |
| Papavasiliou et al. (2006) | Retrospective cohort | 54 patients | Methylprednisolone 40 mg | Clinic: 54 | NR | 5.6% | 9.6 | NR |

PJI prosthetic joint infection, TKR total knee replacement, OR operating room, NR not reported, CI confidence interval

^aRelative risk of PJI due to corticosteroid injection

^bNumber of patients in each subgroup is not reported

Table 35.2 Survey of studies evaluating risk of PJI in TKR following preoperative viscosupplementation injection

| Study | Study design | Number of patients/TKR | HA (type and dose) | Place of injection | Follow-up after TKR (months) | PJI rate | Time between injection and TKR (months) | Odds ratio (95% CI) and <i>p</i> -value |
|--------------------------|----------------------|--------------------------------------|--|--------------------|------------------------------|----------|---|---|
| Richardson et al. (2019) | Retrospective cohort | 3249 patients 646 1113 1490 | NR | NR | 6 | 4.18% | ≤3 >3–6 >6–12 | 1.55(1.02–2.25), <i>p</i> = 0.02 0.84(0.55–1.23), <i>p</i> = 0.39 0.83(0.57–1.16), <i>p</i> = 0.28 |
| Kokubun et al. (2017) | Retrospective cohort | 442 TKR | Synvisc-One ^a Orthovisc ^a Gel-One ^a | NR | 51.5 | 3.0% | ≤3 ≤6 | 1.09(0.52–2.23), <i>p</i> = 0.80 1.47(0.71–3.04), <i>p</i> = 0.29 |

PJI prosthetic joint infection, TKR total knee replacement, HA hyaluronic acid, NR not reported, CI confidence interval

^aNumber of patients in each subgroup is not reported

collated data only from the hospital records and potentially could have missed those patients who received steroid injections from GPs and inadvertently including those patients in the control group introducing an obvious selection bias. Furthermore, the authors did not provide information regarding other patients who may have had IA corticosteroid injection closer to surgery than 8 months without developing infection.

Joshy et al. [81] performed a retrospective matched case-control study involving a group of 32 patients who had PJI following TKR comparing them with a similar number of TKR patients without infection. The authors reported that previous steroid injection was not a risk factor for PJI. However, the very small sample size renders this study significantly underpowered.

In 2008, Horne et al. conducted a similar retrospective matched case-control comparing 40 patients with PJI with 352 patients without PJI following TKR. Apart from reviewing hospital records, the authors assessed the number of IA corticosteroid injections performed in the community by the patient's GP and rheumatologists using questionnaires. Thus, the comparative groups were reduced to 28 (PJI) and 219 (without PJI). They reported that 32% patients in the con-

trol group and 39% in the study group had received a corticosteroid injection. The authors concluded that this was not significant (*p* = 0.44). This study has several inconsistencies on closer inspection. The study group was initially reported to include 29 patients in the methods but subsequently reported as only 28 patients in the results section. The follow-up period was limited to only 6 months post TKR. Furthermore, corticosteroid injection prior to TKA was identified via the questionnaire only, without attempts to screen orthopaedic, rheumatology or general practitioner (GP) records. Therefore, this study is potentially subject to significant recall bias.

Subsequently, Desai et al. [79] in 2009 reported the outcomes of 90 TKRs with preoperative history of corticosteroid injection and 1 year follow-up. Interestingly, 60 knees were injected in the operating room (OR) and the remainder were performed in the outpatient clinic. The authors compared the above patients with a cohort of 180 TKR from the same institution without a history of preoperative corticosteroid injection and reported no PJI in either group. Forty five knees in the study group received an injection within the 12 months prior to surgery. However, the authors did not state how close to

Table 35.3 Survey of studies investigating risk of PJI in TKR based on timing of preoperative intraarticular injection

| Study | Study design | Number of patients/TKR | Therapeutic substance | Place of injection | Follow-up after TKR (months) | PJI (%) | Time between injection and TKR (months) | Odds ratio (95% CI) and p-value |
|-------------------------|----------------------|------------------------|-----------------------|--------------------|------------------------------|---------|---|---------------------------------|
| Bedard et al. (2017) | Retrospective cohort | 29,603 patients | NR | NR | 6 | 4.6 | 1 | 1.29 (1.03–1.62), $p = 0.024$ |
| | | 1804 | | | | 4.4 | 2 | 1.23 (1.06–1.42), $p = 0.005$ |
| | | 5031 | | | | 4.4 | 3 | 1.23 (1.07–1.40), $p = 0.003$ |
| | | 5659 | | | | 4.6 | 4 | 1.28 (1.11–1.50), $p = 0.001$ |
| | | 4197 | | | | 4.6 | 5 | 1.30 (1.09–1.54), $p = 0.003$ |
| | | 3259 | | | | 5.2 | 6 | 1.46 (1.21–1.76), $p < 0.001$ |
| | | 2444 | | | | 3.8 | 7 | 1.06 (0.83–1.35), $p = 0.625$ |
| | | 1839 | | | | 3.5 | 8 | 0.96 (0.72–1.27), $p = 0.779$ |
| | | 1477 | | | | 4.4 | 9 | 1.23 (0.93–1.62), $p = 0.141$ |
| | | 1257 | | | | 3.4 | 10 | 0.95 (0.68–1.33), $p = 0.766$ |
| | | 1053 | | | | 3.4 | 11 | 0.95 (0.65–1.38), $p = 0.781$ |
| | | 850 | | | | 3.8 | 12 | 1.07 (0.73–1.56), $p = 0.741$ |
| Cancienne et al. (2015) | Retrospective cohort | 22,240 patients | NR | NR | 6 | 3.41 | ≤3 | 1.5 (1.2–1.8), $p < 0.0001$ |
| | | 5313 | | | | 2.48 | >3–6 | 1.1 (0.9–1.3), $p = 0.52$ |
| | | 8919 | | | | 2.24 | >6–12 | 1.0 (0.8–1.2), $p = 0.66$ |
| 8008 | | | | | | | | |

PJI prosthetic joint infection, TKR total knee replacement, NR not reported, CI confidence interval

the TKR procedure those injections were given and did not state the mean time between injection and TKR for the study group. It must be noted that the method of performing steroid injections in an operating room is not common. Hence, the findings of this study may not be applicable to all centres. Additionally, the authors did not investigate whether the patients in either group received corticosteroid injection in the community leading to a selection bias.

Khanuja and colleagues reported the outcomes of 302 patients who had received IA corticosteroid injection prior to undergoing TKR [21]. All the IA corticosteroid (triamcinolone 40 mg) injections were performed during office visits under aseptic precautions following skin preparation with alcoholic chlorhexidine solution. They compared the outcomes of the above group with a control group of patients matched for gender, age, body mass index (BMI) and American Society of Anaesthesiologists (ASA) status (1:1 matching). They observed that there was no significant difference between the two groups with respect to PJI (2%—injection cohort and 1%—non-injection cohort; risk ratio = 0.5, $p = 0.5$). Interestingly, the authors stated that whilst all patients in the non-injection control group were drawn based on records of a single institution, they were unable to rule out that this group may have included patients who received IA injection at other institutions.

In 2017, Kokubun et al. reported on the outcomes and complications of 442 TKRs with a mean follow-up of 51 months from a single surgeon series [82]. They reported that 13 patients (3%) had a PJI with 175 patients (40%) having received 4 or more injections whilst 267 patients (60%) patients had received 3 or fewer injections. The authors concluded that preoperative corticosteroid injections do not significantly influence the risk of PJI in subsequent TKR. The authors acknowledge the low numbers in their data and use the principle “a priori” power analysis using the above infection rate. Additionally, the details as to whether the injections were performed in hospital or community settings are missing. This study included both corticosteroid and hyaluronic acid injections. However, it must be noted that the authors do not report the number of patients

in each group, thereby limiting the interpretation of their results.

In their study, Richardson et al. used a large national database in the United States to identify 58,337 patients who had undergone primary TKR between 2007 to 2016 [80]. Using procedure codes, patients who had an IA injection in the 12 months leading up to TKR and underwent surgical procedures for PJI in the 6 months post TKR were reviewed. They had a cohort of 16,656 patients with history of IA corticosteroid injection in the preoperative period and further divided these patients based on the time between the IA injection and TKR (Table 35.1). There were 6653 (39.9%) patients who had had IA corticosteroid injection less than 3 months prior to their TKR with a PJI rate of 3.25% compared to 2.74% in patients without history of IA injection thereby representing a 19% increased risk. After controlling for age, sex and medical comorbidities, there was a significantly higher risk of PJI; the odds ratio was 1.21 (Table 35.1). The authors found that corticosteroid injections given more than 3 months prior to the TKR posed no added infection risk.

35.4.2 Hyaluronic Acid

Only two studies in the current literature have investigated the risk of PJI with TKR in patients receiving viscosupplementation treatment in the preoperative period [80, 82] (Table 35.2). Both studies are methodologically different with the data for the study by Richardson et al. [80] derived from large national database whilst the data for the study by Kokubun et al. [82] represents a single surgeon series.

The primary research question which Richardson and colleagues have investigated is the influence of time gap between viscosupplementation injections and TKR procedure and the associated risk of PJI. Of their total of 3249 patients, a cohort of 646 (19.9%) received HA injections less than 3 months before their TKR with a reported PJI rate of 4.18% compared to 2.74% in the non-injection group. This represented a 53% relatively higher risk of PJI (odds ratio, 1.55).

In their study, Kokubun et al. [82] investigated the influence of the number of injections along with time between viscosupplementation and TKR procedure on PJI. The authors reported an infection rate of 3% in their cohort. The authors concluded that viscosupplementation injections with hyaluronic acid including the timing or number of injections have no relationship with PJI in TKR. However, their data has patients who received IA injections of both corticosteroids and hyaluronic acid. Consequently, several limitations of this study become apparent including the lack of clarity regarding the number of patients who received viscosupplementation treatment and the time gap between the injections and TKR. Hence, the results from this study need to be interpreted carefully with due consideration to all the inherent limitations from a single surgeon series data set.

35.4.3 Biological/Novel Agents

Several studies have been published in the recent past describing the clinical outcomes of knee OA treatment with PRP [45, 46], MSC [43, 58], ACS [61, 83, 84] and APS [63, 85] amongst other biological/novel agents. However, there is no study in the published literature evaluating the risk of PJI in TKR following these treatment modalities. Nonetheless, it is useful to note that septic arthritis has been reported as either a complication of treatment or as an adverse event in the setting of experimental studies involving some of the aforementioned products [86, 87]. Theoretically, given the invasive nature of injections the potential for introduction of occult infection into the knee is present but due to the paucity of studies it is difficult to estimate the risk of PJI in TKR following IA injection with these products.

35.5 Timing of Injection

Two recent studies used information collated from large databases to investigate the influence of the time gap between IA injection in the ipsilateral knee and the risk of PJI in subsequent TKR [20, 88] (Table 35.3).

Cancienne et al. [88] investigated the outcomes of 22,240 patients with TKR and a history of IA injection in the preoperative period (upto 12 months). Their control group matched for age, gender, BMI, diabetes and smoking consisted of 13,650 TKR patients without a history of IA injection. They observed that in the subgroup of 5313 patients who received IA injection less than 3 months before TKR had a higher rate of PJI of 3.41% (odds ratio, 1.5; $p < 0.0001$). There was no significant difference in infection rates in patients who underwent TKA between 3–6 months or 6–12 months after ipsilateral knee injection compared to the control cohort.

In their 2017 retrospective cohort study, Bedard and colleagues [20] used the Humana Health Insurance database to obtain information on the outcomes of TKR patients. They identified 29,603 TKR patients who had had an IA injection in the ipsilateral knee at least 1 year before the TKR. The control group consisted of 54,081 TKR patients with no preoperative IA injection in the ipsilateral knee joint. Both groups were matched as per the Charlson comorbidity index [89]. In the group of TKR patients who had an ipsilateral injection in the preoperative period, they noted that the risk of PJI was higher compared to the control group (4.4% vs. 3.59%; odds ratio, 1.23; $p < 0.001$). Furthermore, the risk of a postoperative infection resulting in return to the operating room within 6 months after TKR was also higher for patients that received an IA injection before ipsilateral TKR than those that did not (1.49% vs. 1.04%; odds ratio, 1.4; $p < 0.001$).

Both the above studies concluded that IA injection prior to TKR was associated with not only a higher risk of PJI but in addition the risk was time-dependent.

35.6 Other Evidence and Guidelines

A review of literature pertaining to injections into the hip joint and subsequent arthroplasty has similar findings to the aforementioned studies [75, 90–93]. The working group which was part of International Consensus Meeting (ICM) noted that there was limited information in the current

literature to recommend the ideal timing for elective total ankle arthroplasty (TAA) after corticosteroid injection for the symptomatic native ankle joint [94]. They recommended that at least 3 months pass after corticosteroid injection and prior to performing TAA [94].

35.7 Conclusions

There is paucity of robust studies evaluating the adverse risk of PJI from the IA injection in subsequent TKR. This has contributed to the lack of consensus on this vital aspect of management of patients with symptomatic knee OA. There is global consensus that PJI in TKR patients is multifactorial with a complex interplay of risk factors. Whilst some risk factors are non-modifiable, factors such as IA injections and timing of TKR represent modifiable factors.

There is a need to continue further research on this topic to develop our understanding albeit with a combination of clinical studies, observational studies based on large databases, systematic reviews and meta-analysis of the available evidence.

IA injections continue to be a part of the management of patients with knee symptomatic OA. Hence these patients need to be appropriately counselled regarding the benefits and potential adverse effects including the risk of PJI in subsequent TKR. A minimum interval of 3 months between IA injection and TKR would be a safe approach to adopt until further evidence emerges to guide the management.

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