Chapter 11 Contribution of Landraces in Wheat Breeding

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11.1 Introduction of Landraces

Agriculture is one of the oldest livelihood sources of mankind. Humans remained actively involved in the selection of favorable traits which resulted in signifcant changes in the phenotype and genotype of wild plants. In addition to man's

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© The Author(s), under exclusive license to Springer Nature 215 Switzerland AG 2021 N. Zencirci et al. (eds.), *Wheat Landraces*, [https://doi.org/10.1007/978-3-030-77388-5_11](https://doi.org/10.1007/978-3-030-77388-5_11#DOI)

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selection activities, environmental factors also played a signifcant contribution in the selection of various favorable traits suitable for man-made land and gardens. Combination of these activities resulted in the development of distinctive populations called "landraces" (Zeven [1998](#page-43-0)). Landraces are dynamic populations of cultivated plants having a historical background, genetically diverse and distinct identity, and good adaptation to local environment and that are associated with traditional farming systems (Villa et al. [2005\)](#page-41-0). Dwivedi et al. ([2016\)](#page-28-0) stated that landraces are heterogeneous populations of domesticated species having great adaptation to local environment and can serve as a source of genetic variations that can be very helpful to combat the current and new challenges for farming in changing environments. Landraces are found phenotypically diverse and less productive compared to their cultivated types (Mir et al. [2020\)](#page-35-0). However, regarding their quality attributes, landraces have been found highly nutritious compared to their cultivated ones (Azeez et al. [2018\)](#page-26-0). Landraces played a major role in plant breeding by providing novel genes for various agronomic, quality, mineral, biotic, and abiotic traits (Azeez et al. [2018;](#page-26-0) Lopes et al. [2015\)](#page-34-0). An impressive increase in yields per hectare was the result of the "Green Revolution" due the inclusion of high-yielding varieties (HYVs) having better response to inputs (Mir et al. [2020\)](#page-35-0). After the inclusion of these highyielding varieties, it was supposed that landraces will inevitably disappear with time (Frankel and Bennett [1970](#page-29-0); Zeven [1998](#page-43-0)). However, these breeding activities led to genetic erosion and emergence of various modern cultivars that are prone to various biotic and abiotic stresses. It is estimated that approximately 75% loss of genetic diversity is observed in the last 100 years (Hammer et al. [1996\)](#page-30-0). Globally, loss of genetic diversity is very alarming because it can be used to combat food scarcity problems in the long term. Therefore, it is very important to pay attention to collect, preserve, and grow these landraces as they guarantee the existence of variations that can be used for breeding of crops for the production of more quantity of food with high quality. Besides the inclusion of HYVs, landraces maintained their position by playing a key role in agricultural production, specifcally in those environments where commercial cultivars failed their competitive advantage (Casañas et al. [2017\)](#page-27-0).

11.2 Origin of Wheat Landraces

Wheat is one of the domesticated food crops cultivated in mild temperature and consumed as a staple crop by millions of people (Lodhi et al. [2020\)](#page-34-1). Domestication of wheat is considered a key reason behind increased human population, thereby participating in the emergence of the human civilization (Jaradat [2011\)](#page-31-0). Domestication of wild emmer (*Triticum dicoccoides*), which is the progenitor of all polyploid cultivated wheats, is considered an important event in the emergence of agriculture in Southwest Asia. Domestication of wild emmer occurs in the Fertile Crescent, and it acted as a prerequisite for the evolution of tetraploid durum and hexaploid bread wheat (Jaradat [2011\)](#page-31-0). However, the domestication and the

subsequent breeding activities drastically reduced wheat genetic diversity (Dvorak et al. [1998\)](#page-28-1).

Ancient farmers planted diverse assemblages of wheat genotypes (i.e., landraces) aiming to decrease the risk of crop failure and to improve food security because they had limited capacity to control the spatially heterogeneous and temporally unpredictable environment (Jaradat [2006;](#page-31-1) Peng et al., [2011a,](#page-37-0) [b\)](#page-37-1). This exercise resulted in the development of wheat landrace meta-populations and the evolution of farmers' seed systems through which they accessed and exchanged diverse genetic material. A meta-population structure can be defned as a group of subpopulations that is interconnected through gene fow and seed exchange and favors the evolution of diversity (Jaradat [2011\)](#page-31-0). It is believed that natural interference, human skills, and years of continuous cultivation resulted in great diversity in wheat genotypes (Lodhi et al. [2020\)](#page-34-1). Zeven ([2000\)](#page-43-1) stated that previously many farmers used wheat crops to develop new cultivars. Archaeological evidence are present revealing the cultivation of wheat in Iberian Peninsula, since the ffth millennium BC, and the development of wild wheats, traditional wheat varieties, and other crops happened in the Fertile Crescent (Diamond [2002](#page-28-2)).

Wheat landraces were developed from their older ones having the ability to grow in such conditions which are not feasible for the growth of the regular wheat (Witcombe et al. [1996\)](#page-42-0). Zeven [\(1999](#page-43-2)) stated that wheat landraces are crop varieties developed by farmers through human and natural selection and refect adaptation to local management practices and environmental conditions. Combination of both human and natural selection resulted in changes in the architecture of genotypes having better attributes like drought, salt, cold, or heat tolerance, quality traits, time to heading and maturity, and seed flling duration (Masood et al. [2005\)](#page-35-1). Due to genetically distinct plant populations, wheat landraces are conserved, and some specifc names were given by the traditional farmers in order to meet their environmental, cultural, social, and economic needs. Therefore, landraces are also known as farmer's varieties or folk varieties (Belay et al. [1995](#page-26-1)).

11.3 How Landraces Contributed in Wheat Breeding

Landraces played a signifcant role in wheat breeding by gaining focus from breeding community. Wheat landraces served as genetic resource for the development of climate-resilient cultivars with high yield (Abu-Zaitoun et al. [2018\)](#page-24-0). An increasing interest has been observed for the usage of landraces as source of nutritional traits and favor repertoire and landrace cultivation for niche markets (Roselló et al. [2018\)](#page-38-0). Wheat landraces contain higher genetic diversity compared to most modern wheat landraces, and this diversity includes their adaptation to environmental conditions according to the place of origin. Some countries used this characteristic in the development of frst improved cultivars through the selection of local landraces. For example, "Aragon 03" was the leading variety in Spain during the period 1960–1976. It was developed from indigenous landrace population "Catalan de Monte" (Gadea

[1958\)](#page-29-1) and showed high ability to drought resistance (Royo and Briceño-Félix [2011a](#page-38-1); [b](#page-39-0)). Similarly, "Turkey" (syn. "Turkey Red"), a hard red winter wheat having better adaption for cold regions, showed marvelous impact on wheat cultivation in the United States at the turn of the last century due to decreased winterkill, among other traits (Olmstead and Rhode [2002\)](#page-36-0). A Japanese landrace "Akakomugi," containing Rht8c and Ppd-D1, was used by Italian breeder Nazareno Strampelli to improve Italian wheat gene pool (Salvi et al. [2013\)](#page-39-1). The sensational varieties "Ardito and Mentana" developed from the crosses of Strampelli, including Akakomugi, became the basis of most of the new varieties developed in Mediterranean countries, South American countries, and several distant countries such as Russia and China. In Argentina, "Ardito" was used as parent to develop the variety Klein-33, which became the backbone of the former USSR breeding program, generating the variety Bezostaja-1 (Borojevic and Borojevic [2005](#page-26-2)). Contribution of landraces in wheat breeding for various traits is discussed comprehensively.

11.3.1 Role of Landraces in Adaptive Traits

Adaptive traits suited to target the environment have acted a decisive role during domestication and the spread out of domesticated wheat. Fitting fowering time to the current conditions in the target environments is presumably one of the main important factors during dispersal (Peng et al. [2011a;](#page-37-0) [b](#page-37-1); Royo et al. [2020\)](#page-38-2). The frst domesticated cereals/old landraces had most probably response to day length and cold temperatures like their wild relatives/progenitors. Motzo and Giunta [\(2007](#page-36-1)) hypothesized that old cultivars/landraces had the greatest day-length sensitivity and vernalization in comparison to intermediate and modern ones. However, novel adaptive traits for each target environment were naturally or artifcially selected during the domestication and spreading process from the Fertile Crescent to new agricultural areas (Kilian et al. [2009\)](#page-33-0). Especially other yield-related traits such as plant height, waxiness, number of spikes, and weight of spikes and grains were also coselected by ancient farmers, and many botanical variants have been developed in this process (Peng et al. [2011a;](#page-37-0) [b\)](#page-37-1). Wheat landraces arising from the migration from the Fertile Crescent to the other regions of the world had been grown extensively until the Green Revolution in the early 1970s (Harlan [1975](#page-30-1)). As a result of the Green Revolution, more productive semidwarf wheat cultivar shaving better response to inputs replaced the landraces/local populations which are generally identifed as tall, tended to lodge, sensitive to the foliar diseases, and low yielded (Reynolds and Borlaug [2006a](#page-38-3); [b;](#page-38-4) Lopes et al. [2015\)](#page-34-0). Nevertheless, their cultivation has continued in marginal environments and they currently support subsistence farming in many regions of the world (Newton et al. [2010](#page-36-2)).

The wide range adaptability of wheat is mainly based on three genetic groups such as vernalization (*Vrn*) genes, photoperiod (*Ppd*) genes, and genes controlling "narrow-sense earliness" or "earliness per se" (*Eps*). Vernalization, which is inducting fowering by exposure to cold, basically determines plant growth habit types as winter (strong vernalization requirement) and spring (no vernalization requirement). Vernalization in wheat has very allelic complex and previous studies have presented that *Vrn* allele combinations or frequencies with an adaptive value in target growing areas are varied geographically (Stelmakh [1990](#page-40-0); Damania et al. [1996;](#page-28-3) Iwaki et al. [2001;](#page-31-2) Zhang et al. [2008;](#page-43-3) Sun et al. [2009\)](#page-40-1). Kato and Yokoyama [\(1992](#page-32-0)) observed the main adaptive traits in 158 bread wheat landraces collected from various climatic regions including Asian and European countries, and they claimed that nearly half of the variation for observed traits was accounted by geographical differences of their origin centers. Kato et al. ([1997\)](#page-32-1) also studied geographical variation of wild emmer (*Triticum dicoccoides*) accessions for vernalization response and earliness in comparison to other tetraploid relatives such as cultivated emmer (*T. dicoccum*), durum wheat (*T. durum*), and *T. turgidum*. They concluded that spring growth habit in *T. dicoccoides* could have evolved from a winter type especially in temperate conditions.

Many studies presented that the vernalization requirement in wheat is considered to be genetically controlled by at least three loci, *Vrn-A1* (*Vrn-1*), *Vrn-B1* (*Vrn-2*), and *Vrn-D1* (*Vrn-3*), located in chromosomes 5A, 5B, and 5D, respectively (Pugsley [1971,](#page-37-2) [1972](#page-37-3); Law et al. [1976](#page-33-1); Galiba et al. [1995;](#page-29-2) Dubcovsky et al. [1998](#page-28-4); Yan et al. [2003\)](#page-43-4). While *Vrn-A1* has the major impact on transition from vegetative to generative phase, recessive mutants of *Vrn-B1* trigger fowering. While a dominant allele of any *Vrn* genes causes spring growth habit, wheats classifed as winter type must have recessive alleles at all *Vrn* loci (Turner et al. [2013\)](#page-40-2). On the other hand, photoperiodic response in wheat is primarily controlled by three major genes, *Ppd-D1* (*Ppd1*), *Ppd-B1* (*Ppd2*), and *Ppd-A1* (*Ppd3*), located in 2DS, 2BS, and 2AS chromosomes, respectively. It is known that *Ppd-D1* plays an important role in regulation of photoperiodic response. In addition, "earliness per se" or "narrow-sense earliness" is the difference in fowering times of genotypes whose vernalization and day-length requirements have been completed (Kato et al. [2001\)](#page-32-2). Earliness per se genes can also affect fowering time independently, but these genes have not been studied in detail because of major effects of vernalization and photoperiod genes on fowering time. Moreover, this trait is highly heritable and can be effectively used in breeding programs (Kato and Wada [1999](#page-32-3)). Many QTLs have been identifed for earliness per se in all three genomes with previous studies (Bullrich et al. [2002;](#page-27-1) Hanocq et al. [2004](#page-30-2); Kamran et al. [2013](#page-32-4)).

Previous studies with a marker-assisted selection approach have clarifed that landraces/accessions have a huge genetic diversity and very allelic complex for vernalization and photoperiod genes. Jiang et al. ([2010\)](#page-31-3) found that the frequencies of the dominant *Vrn* genes in 153 Chinese wheat landraces were 60.78% (*Vrn-D1*), 5.88% (*Vrn-A1a*), 5.23% (*Vrn-B1*), and 0 (*Vrn-B3*), respectively. Andeden et al. [\(2011](#page-25-0)) determined that Turkish wheat germplasm has mostly the dominant *Vrn-B1* allele followed by *Vrn-D1* and *Vrn-A1*. Derakhshan et al. ([2013\)](#page-28-5) reported that the frequencies of dominant *Vrn-D1* and *Vrn-B1* alleles in 395 Iranian wheat landraces were 67.35% and 38.48%, respectively. Manickavelu et al. ([2014\)](#page-35-2) characterized 400 wheat landraces genetically collected from different agroecological zones of Afghanistan for adaptive and other yield-related traits, and they reported that 53%

of all landraces were winter types, 43% had one or more dominant *Vrn* alleles, and 4% were either unknown or had *Vrn-A1c* – a rare spring allele. Guo et al. [\(2015](#page-30-3)) also studied distribution of the *Vrn-D1b* allele in Chinese wheat accessions and determined that the frequencies of *Vrn-D1a*, *Vrn-D1b*, and *Vrn-D1* alleles were 27.3, 20.6, and 52.1%, respectively, of 689 accessions. They also claimed that *Vrn-D1b* allele originated from Chinese landraces as a result of pedigree analysis. Goncharov ([1998\)](#page-29-3) claimed that there is a high rate of the *Vrn-D1* allele in countries near the equator in addition to Pakistan, Afghanistan, and China.

Other important genetic factors like dwarfng genes (*Rht*) are critical against environmental stresses to guarantee both adaptability and grain yield in addition to vernalization, photoperiod, and earliness. It is known that over 30 height-reducing genes have been identifed so far (McIntosh et al. [2013\)](#page-35-3). The major dwarfng genes "*Rht-B1*" and "*Rht-D1*" known as the *Reduced height* (*Rht*) loci were introduced during the "Green Revolution" that achieved to improve harvest index by reducing plant height. These genes are known as gibberellic acid (GA)-insensitive dwarfng genes and located on chromosomes 4BS and 4DS, respectively. Another important height-reducing gene is *Rht8* classifed as GA-sensitive. *Rht8* is located in chromosome 2D close to *Ppd-D1* and previous studies clarifed that *Rht8* and *Ppd-D1a* alleles are often derived together (Worland et al. [1998\)](#page-42-1), but *Ppd-D1a* has pleiotropic effects independently on plant height, grain yield, and yield-related traits (Börner et al. [2002](#page-26-3); Chebotar et al. [2013](#page-27-2); Zhang et al. [2019a,](#page-43-5) [b\)](#page-43-6). Zhang et al. [\(2006](#page-43-7)) determined that *Rht-B1b*, *Rht-D1b*, and *Rht8* are common in autumn-sown Chinese wheat germplasm, while the frequencies of alleles vary from between regions. Kolev et al. ([2011\)](#page-33-2) also reported the most frequent alleles as *Ppd-D1b*, *vrn-A1*, *vrn-B1*, *vrn-*D1, and *Rht-B1a* in Bulgarian germplasm including old cultivars and landraces. Rasheed et al. ([2016\)](#page-38-5) studied the allelic variation of economically important traits such as *Vrn*, *Ppd* and *Rht*, in 107 wheat landraces collected from different geographic zones of Pakistan. They determined that less than half of the landraces has *Ppd-D1a*, *Rht-B1b*, *Rht-D1b*, and spring-type alleles of *Vrn-A1* and *Vrn-D1*. The studies explained above highlight how these genes from landraces have geographically evolved in the target areas.

11.3.1.1 Success Stories of Wheat Landraces for Adaptive and Yield-Related Traits

There are many successful reports in the development of new wheat varieties with the use of landraces containing different dwarfng genes. A Japanese landrace "Akakomugi," containing *Rht8c* and *Ppd-D1*, was used by Italian breeder Nazareno Strampelli to improve Italian wheat gene pool (Salvi et al. [2013\)](#page-39-1). The crosses between Italian genotypes and Akakomugi resulted into the introgression of new alleles such as *Ppd-D1* and *Rht8c*. The sensational varieties "Ardito and Mentana" developed from the crosses of Strampelli, including Akakomugi, became the basis of most of the new varieties developed in Mediterranean countries, South American countries, and several distant countries such as Russia and China. In Argentina,

"Ardito" was used as parent to develop the variety Klein-33, which became the backbone of the former USSR breeding program, generating the variety Bezostaja-1 (Borojevic and Borojevic [2005\)](#page-26-2). Another variety, Frontana, derived from a cross with Mentana, was part of the pedigree of the varieties Penjamo 62, Yaqui 48, Lerma 50, Escobar, and Supremo. Similarly, many genotypes derived from Mentana were developed in breeding programs of Canada and Australia (Salvi et al. [2013](#page-39-1); Tadesse et al. [2016\)](#page-40-3).

A similar success story from the Nobel laureate Norman Borlaug in the mid-20th century was recorded with the Norin 10/Brevor cross containing *Rht-B1* and *Rht-D1*. The lineage of Norin-10, developed by a Japanese breeder G. Inazuka, is tracked back to a Japanese short-straw landrace "Shiro Daruma" containing *Rht-B1* and *Rht-D1* crossed with the American high-yielding varieties Fultz and later Turkey Red (Reitz and Salmon [1968](#page-38-6)). Norin 10-Brevor 14 cross was sent to N. Borlaug at the International Maize and Wheat Improvement Center (CIMMYT) in Mexico, and this cross and new crosses with Norin/Brevor 14 variants were tested for adaptation in tropical and subtropical climates in the center (Hedden [2003](#page-30-4)). Wheat varieties developed from semidwarf wheats developed by N. Borlaug and his colleagues in CIMMYT are grown in millions of hectares in many regions of the world.

The story of the Turkey Red brought to America is also very interesting. This bread wheat landrace, which was frstly grown in the USA around Kansas in the 1870s, was introduced to this region by German Mennonites who migrated from Crimea to the USA (Quisenberry and Reitz [1974;](#page-38-7) Smale [1996\)](#page-39-2). The landrace has thin stem, high plant height, tended to lodging, narrow and dark green leaves, resistance to harsh climate conditions, white grain, high biomass, resistance to rust diseases, and tolerance to other foliar diseases (Quisenberry and Reitz [1974;](#page-38-7) Lopes et al. [2015\)](#page-34-0). In addition, the landrace "Crimean," introduced at the same time as Turkey Red, was directly included into the Nebraska gene pool. The effects of these two landraces on wheat improvement were indirectly reported with previous studies (Ali et al. [2011](#page-25-1); Mengistu et al. [2012](#page-35-4)). Previous reports reported the investigation of major quantitative trait locus (QTL) related to grain yield on chromosome 3A originated from the cultivar "Wichita" which was obtained from these landraces.

Another important landrace "Chinese Spring" (CS) has also affected the wheat improvement and genetics in depth. This variety is known to be a Sichuan landrace, and Yen et al. ([1988\)](#page-43-8) claimed that CS is similar to a Sichuan white landrace "Chengdu-guang-tou" (CDGT) in terms of morphology, physiology, and cytogeneticsbased comparison. The similarity of these two landraces was also presented with RFLP profling by Ward et al. [\(1998](#page-42-2)). CDGT has still been used widely in Sichuan breeding programs because of its high tillering potential, high number of spikelets, and high level of foret fertility (Liu et al. [2018](#page-34-2)). In addition, the landrace has been widely used to develop wheat-rye translocation lines because of its ready crossability, and therefore, many cultivars and pre-/breeding lines have been developed using CS as parent both in China and many different regions of the world. However, the main important impact of CS is on genetics and molecular breeding of wheat in which CS (IWGSC RefSeq v2.0) was sequenced at genome and single chromosome

level and released the genomic data for public access [\(http://www.wheatgenome.](http://www.wheatgenome.org/News2/IWGSC-RefSeq-v2.0-now-available-at-URGI) [org/News2/IWGSC-RefSeq-v2.0-now-available-at-URGI\)](http://www.wheatgenome.org/News2/IWGSC-RefSeq-v2.0-now-available-at-URGI).

In addition to important wheat landraces mentioned above, several landraces with important adaptive and yield-related traits used by plant breeders in the early twentieth century have intensely been used in pedigrees of modern wheats such as Zeeuwse Witte in the Netherlands, Blount's Lambrigg and Purple Straw in Australia, Marquis and Red Fife in Canada, Kunduru in Turkey, Saragolla in Italy, and Turkey Red in USA, which actually originated from Turkey (Gökgöl [1935;](#page-29-4) Quisenberry and Reitz [1974;](#page-38-7) Ozberk et al. [2016;](#page-37-4) Alsaleh et al. [2016](#page-25-2)) and became a cornerstone of the early European and indirectly world breeding programs (Smale [1996;](#page-39-2) Braun et al. [2001\)](#page-27-3). In addition to these examples, wild progenitors/relatives and transition forms of wheat have formed the evolution and distribution of modern wheat landraces and (indirectly) cultivars. Especially, many unique alleles that provide resistance to different diseases and pests, including rust diseases, powdery mildew, *Septoria tritici* blotch, *Septoria nodorum* blotch, tan spot, cyst nematode, root knot nematode, Hessian fy, greenbug, Russian wheat aphid, wheat curl mite, and soilborne cereal mosaic virus, have been introgressed to modern wheat cultivars (Kishii [2019](#page-33-3)).

Introgression of new alleles from the locally adapted landraces to modern wheat cultivars should be one of the main breeding targets. Unfortunately, most of landraces have not still been identifed both genetically and agronomically. However, the effcient use of landraces in breeding programs requires understanding their genetic diversity and population structure. Baloch et al. ([2017\)](#page-26-4) evaluated the genetic diversity of 92 durum wheat landraces from the Central Fertile Crescent including Turkey and Syria with 39,568 DArT-seq and 20,661 SNP markers. As a result of the study, Turkish and Syrian landraces complexly clustered into three groups, and the results illustrated that farmer-mediated selection and lack of the commercial varieties might have concluded in the exchange of genetic materials between two neighboring regions. Soriano et al. ([2016\)](#page-39-3) classifed 172 durum wheat landraces, using molecular markers, into four genetic populations in relation to their geographic origin: eastern Mediterranean (EM), eastern Balkans and Turkey, western Balkans and Egypt, and western Mediterranean (WM). They determined that the genetic diversity among landraces increased during migration to West Mediterranean basin due to lower genetic diversity in the eastern Mediterranean population. Soriano et al. [\(2018](#page-39-4)) also support the theory with an association mapping study that 23 marker alleles in relation to important agronomic traits with different frequencies from east and west regions of Mediterranean basin were identifed. With a similar approach, Liu et al. ([2017a](#page-34-3); [b](#page-34-4); [c\)](#page-34-5) reported a genome-wide association study with 52,303 DArT-seq markers that 723 wheat landraces collected from ten different agroecological zones of China were investigated for 23 agronomic traits in six environments. As a result of the study, all landraces were classifed into fve clusters based on phenotypic data, and 25 candidate genes associated with signifcant markers were characterized.

Unveiling the genetic basis of yield-related traits in wheat landraces is vital to ensure global food security because of their higher genetic diversity, large number

of alleles, and potency of unique variants of alleles compared to modern wheat varieties. The advent of new technologies about sequencing, mapping, and other related technologies has been facilitating high-quality sequences of wheat and its relatives. The sequences will likely stimulate many new studies on evolution, genetics, and genomics of wheat, and accelerate characterization of novel genes controlling important adaptive and yield-related traits from landraces and wild relatives of wheat.

11.3.2 Role of Landraces in Abiotic Stress

Resistance to abiotic and biotic stresses, productivity, seed quality, seed mineral content, and many other traits will be future breeding aims to meet the world's rapidly increasing food demand. Availability of higher natural genetic diversity to increase selection efficiency is one of the most critical and significant objectives of breeding programs. The abiotic stress factors (salinity, heat, drought, etc.) adversely affect crop production and yield (Jaleel et al. [2009;](#page-31-4) Thakur et al. [2010;](#page-40-4) Mantri et al. [2012\)](#page-35-5). Traditional plant breeding is a long-term process that has been used effectively for many years, and molecular tools can be employed to overcome complications and to ensure the improvement of speed breeding strategies (Nadeem et al. [2018;](#page-36-3) Baloch et al. [2016](#page-26-5)). In this part, we discussed the role of landraces in different abiotic stress conditions such as salinity, heat, and drought to provide a signifcant resource for wheat breeders.

11.3.2.1 Wheat Landraces' Role in Salinity Tolerance

Salinity is a major feature that reduces crop production and affects nearly 1 billion hectares of land worldwide (Fageria et al. [2012](#page-28-6)). Therefore, developing crops providing a satisfactory amount of product in salty soils or different climatic conditions is important to meet the growing food demand. Screening of wheat germplasm for salt tolerance has been conducted by various researchers (Kumar et al. [2017;](#page-33-4) Arabbeigi et al., [2018](#page-25-3)). For example, Shahzad et al. ([2012\)](#page-39-5) evaluated wheat landrace genotypes using morphological and molecular markers for salinity tolerance at the vegetative stage. The authors proposed that accessions 10793 (Pakistan), 10790 (Pakistan), 10821 (Pakistan), and 11526 (Pakistan) are found salt-tolerant at 200 mM NaCl stress. At 250 mM NaCl stress, accession 11299 (Pakistan) was the most salt-tolerant followed by accessions 11335 (Pakistan), 11370 (Italy), and 11214 (Pakistan). Additionally, accessions 10790 (Pakistan), 10828 (Pakistan), 10823 (Pakistan), and 4098805 (4098805) performed better at both 200 and 250 mM NaCl stresses. In another study, Chaparzadeh et al. ([2014\)](#page-27-4) determined the effects of NaCl (control, 75, and 150 mM) on the plant leaves of 18 bread wheat (*Triticum aestivum* L.) landraces from the west area of the Urmia Saline Lake. While accessions 12194 (from Piranshahr), 11199 (from Urmia), and 11488 (from Salmas) were found as the most tolerant with combined salt tolerance indexes for all biochemical and physiological parameters, accessions 11479 (from Mahabad) and 11492 (from Urmia) were determined as the least tolerant. It was suggested that these parameters could be used together as powerful biomarkers to screen for salt-tolerant landraces using the cluster analysis method. Al-maskri et al. ([2014\)](#page-25-4) investigated specifc stem and leaf structural traits for water conservation. Based on the results of the study, cultivars/landraces were rated according to their degree of drought and salt tolerance as S-24 (from Pakistan) > J-305 (from Oman) > Sarraya (from Northern Asia, Africa, Middle East, Asia Minor) > Senain (from Oman) > Cooley (from Chile and Mongolia) > MH-97 (from Pakistan) > Missani (from the Mediterranean, Middle East Asia, and North Africa) > Hamira (from Oman) > Shwairaa (from Oman). Two of them (S-24 and J-305) are rated as highly tolerant, fve moderately tolerant (Sarraya, Senain, Cooley, MH-97, and Missani), and two sensitive (Hamira and Shwairaa). The recent advances in genomic information and technology have opened new horizons and foundations for genetic breeding of salt tolerance. Various QTL mapping studies for salt tolerance in wheat were conducted by Quarrie et al. [\(2005](#page-37-5)), Ma et al. [\(2007](#page-34-6)), Genc et al. [\(2010](#page-29-5)), Hussain et al. [\(2017](#page-31-5)), Shamaya et al. [\(2017](#page-39-6)), Ren et al. ([2018\)](#page-38-8), Devi et al. ([2019\)](#page-28-7), and Ilyas et al. [\(2020](#page-31-6)). On the other hand, Yu et al. [\(2020](#page-43-9)) analyzed in a GWAS using 307 wheat accessions including local landraces and exotic cultivars. Researchers found that some Chinese landraces such as Baihuamai, Youzimai, Beijing 10, Jimai 1, and Zaosui 30 displayed superior salt tolerance. According to kinship analysis, Chinese landraces revealed a source of rare favorable genetic variation. Moreover, many of these landraces have already adapted to the different environments in China (Liu et al. [2017c;](#page-34-5) Zhou et al. [2018\)](#page-43-10). In addition to these examples, wild relatives of wheat are also potential sources of important genetic materials such as salinity tolerance for wheat breeding. The use of wild relatives of *Triticum* species is one of the main breeding targets and may offer an opportunity to improve salinity tolerance by presenting availability to more variable germplasm (Shavrukov et al. [2009](#page-39-7)). For this content, researchers investigated the salinity tolerance of various accessions of *Aegilops tauschii*, and determined that the accessions studied are found similar to bread wheat. On the other hand, it was presented that accessions of *Aegilops tauschii* had a much lower Na+ ratio but higher K+/Na+ ratios in their leaves than did durum wheat (Gorham et al. [1987,](#page-30-5) [1990](#page-29-6)). Another important wild relative of wheat is jointed goatgrass, *Aegilops cylindrica* Host. $(2n = 4x = 28; CCDD)$ species, which was formed through amphidiploidization of a hybrid or hybrids between *Ae. tauschii Coss*. (2n = 2x = 14; DD) and *Ae. markgrafi* (Greuter) Hammer (2n = 2x = 14; CC). Farooq et al. [\(1989](#page-28-8)) screened *Ae. cylindrica* accessions obtained from inland Pakistan and oversea, and determined that some of salinity-tolerant accessions survived at 300 mM NaCl and 400 mM NaCl in treatments using Hoagland solution. Another researcher reviewed the use of wild relatives of wheat for salinity tolerance (Colmer et al. [2006\)](#page-27-5). Arabbeigi et al. [\(2014](#page-25-5)) evaluated the physiological response of the highly salinitytolerant *Ae. cylindrica* genotypes and the SSR and EST-SSR markers linked to the salinity tolerance. As a result of the study, ten most salinity-tolerant genotypes of *Ae. cylindrical* were identifed. In addition, Xgwm312, Xwmc170, Xgwm291, and Xgwm410 microsatellite markers produced a distinguished banding pattern in the ten most salinity-tolerant genotypes in the study. These markers can play important role in wheat breeding programs. Very recently, Ahmadi et al. [\(2020](#page-24-1)) investigated the domesticated and ancestral wheat genotypes, including *Ae. triuncialis*, *Ae. neglecta*, *Ae. umbellulata*, *Ae. caudata*, *Ae. speltoides*, *Ae. tauschii*, *T. boeoticum*, *T. durum*, *T. urartu*, and *T. aestivum*, under control and salinity stress to evaluate the mechanisms involved in salinity tolerance. It was found that two neglected (*Ae. triuncialis*) and ancestral (*Ae. tauschii*) wheat genotypes responded better to salinity tolerance than other genotypes. The studies explained above revealed that variation among the wild relatives and landraces of wheat is available for salinity tolerance, and they can be used to develop modern wheat cultivars in breeding studies.

11.3.2.2 Drought and Heat Stress Tolerance in Wheat

Drought and heat stress are important climatic factors that occur in almost all climatic areas of wheat-growing areas and cause a signifcant crop loss of up to 40% and 60% by drought and heat stresses in felds, respectively (Zampieri et al. [2017;](#page-43-11) Thirumalaikumar et al. [2018\)](#page-40-5). These factors affect crops at the physiological, morphological, and biochemical levels (Guo et al. [2020\)](#page-30-6); reduce photosynthesis (McKay et al. [2003\)](#page-35-6), cell turgor (Taiz and Zeiger [2006](#page-40-6)), and chlorophyll fuorescence with a critical reduction of the Fv/Fm ratio (Mohammed and Tarpley, [2009;](#page-36-4) Izanloo et al. [2008\)](#page-31-7), and impair cell division and elongation (Bal et al. [2010](#page-26-6)) in sensitive wheat lines compared with tolerant lines. Wheat yield is particularly sensitive to drought and heat stress factors that reduce spikelet productivity, individual grain weight, grain number, and grain flling time during the breeding season (Mahrookashani et al. [2017\)](#page-34-7). The lack of water is not invincible (Ballesta et al. [2019](#page-26-7)). The adverse effects of drought and heat factors can be overcome by using drought- and heatresistant cultivars (Van Oosten et al. [2016\)](#page-41-1). The global scenario consists of having a genetic balance of major/minor genes suitable key for these stress factors and developing stress-resistant varieties (Mujeeb-Kazi et al. [2009](#page-36-5)). Success in plant development commonly depends on the size of genetic variability and the extent to which the benefcial traits are inherited (Kahrizi et al. [2010](#page-32-5)). Information from the germplasm evaluation will be of great importance for drought- and heat-tolerant genotype selection (Okechukwu et al. [2016](#page-36-6)). Breeding wheat varieties that tolerate these stressors is currently a major challenge for wheat breeders (Mwadzingeni et al. [2016\)](#page-36-7). Exotic wheat landraces have been shown to be an excellent source of various genes and to function better under stressful conditions (Reynolds et al. [2007\)](#page-38-9). Various studies were conducted to evaluate genetic resources in terms of drought and heat resistance (Hede et al. [1999;](#page-30-7) Sareen et al. [2014;](#page-39-8) Pinto et al. [2017](#page-37-6); Al Khateeb et al. [2017](#page-25-6); Ullah et al. [2018;](#page-41-2) Korkut et al. [2019](#page-33-5)). Hede et al. [\(1999](#page-30-7)) used a group of 2255 accessions from a Mexican landrace collection in which three landrace accessions (CWI 60155, CWI 59788, and CWI 60391) were determined as having superior and stable leaf chlorophyll content in both environments in 1997. In a study conducted by Sareen et al. (2014) (2014) , six wheat genotypes (IC 28661, IC 57586, IC 78856, IC 28938B, IC 36761A, and IC 78869A) were identifed as tolerant to drought and heat stresses. Al-maskri et al. [\(2014](#page-25-4)) rated cultivars/landraces according to their degree of drought and salt tolerance as $S-24$ (from Pakistan) $> J-305$ (from Oman) > Sarraya (from Northern Asia, Africa, Middle East, Asia Minor) > Senain (from Oman) > Cooley (from Chile, Mongolia) > MH-97 (from Pakistan) > Missani (from Mediterranean, Middle East Asia, North Africa) > Hamira (from Oman) > Shwairaa (from Oman). Aktaş [\(2016](#page-25-7)) determined the most tolerant genotypes (SEN-DER genotypes G7, G10, landrace group genotype G11 (Sorık)) to be used to improve drought-tolerant varieties. Al Khateeb et al. ([2017\)](#page-25-6) used four wheat landraces collected from Jordan and indicated that Karak landrace may be selected as the most tolerant wheat capable of adapting to drought-prone environments. Chaichi et al. [\(2019](#page-27-6)) screened 123 Iranian wheat (*Triticum aestivum* L.) landraces (spring and winter genotype) for drought tolerance using morphological and physiological features. They determined L-82 and Marvdasht genotypes as droughttolerant and sensitive genotypes, respectively. Korkut et al. ([2019\)](#page-33-5) determined that some genotypes (Nota, Dropia, CIMMYT-HTN 2014/15-6, CIMMYT-HTN 2014/15-2, CIMMYT HTN 2014/15-10) could be evaluated as genitor(s)/ progenitor(s) in the wheat breeding programs for heat tolerance.

Landraces, wild relatives, and traditional varieties are potential reservoirs of novel alleles for improving abiotic stress tolerance (Karan and Subudhi [2012](#page-32-6)). In this context, a deeper understanding of the genetic mechanisms of drought and heat resistance is important to maintain and further develop the effciency of wheat breeding programs (Arriagada et al. [2017\)](#page-25-8). The initial genetic investigations of wheat under both drought and heat stress in controlled conditions were conducted in durum wheat and bread wheat by Aprile et al. [\(2013](#page-25-9)) and Qaseem et al. ([2018\)](#page-37-7), respectively. Merchuk-Ovnat et al. ([2016\)](#page-35-7) revealed that introgression of QTLs on chromosomes 1B and 2B of *T. turgidum* into *T. aestivum* can improve drought tolerance in domesticated wheat. B genome has been identifed carrying loci controlling water utilization efficiency, associated traits, and grain yield under water stress conditions (Mohammadi et al. [2012;](#page-35-8) Poersch-Bortolon et al. [2016\)](#page-37-8). In another study conducted by Touzy et al. ([2019\)](#page-40-7), a panel of 210 elite European wheat varieties in 35 feld trials was evaluated, and GWAS (genome-wide association study) was done with six characters in four different environment types to confirm 590 QTLs, some of which were specifc to the different water stress patterns. Schmidt et al. [\(2020](#page-39-9)) used 315 spring bread wheat accessions to evaluate in pots with semi-controlled environmental conditions that combined drought and heat stress in 2016 and 2017. Australian and Mexican varieties were rated as having great productivity potential under both stresses, which have been selected for their yield performance and made up about 70% of the spring wheat panels. Nearly one-ffth of the tolerant wheat came from varieties of various origins such as the Middle East, the USA, Central Africa, India, and Canada. In the study, QTLs were determined on all chromosomes, most of which were on chromosomes 3B, 5A, 5B, and 6B. Drought and heat stress factors, which together can lead to signifcant yield losses, have restricted wheat yields in various wheat-growing areas worldwide, and their combined impact could result in critical yield losses (Toreti et al. [2019\)](#page-40-8). Information about QTLs can help breeders to improve new cultivars tolerant to drought and heat stress in marginal environments in future global margins.

11.3.3 Role of Wheat Landraces in Quality Traits

11.3.3.1 Landraces for Biofortifcation

"Biofortifcation" or "biological fortifcation" is the process of improving the nutritional status of staple crops such as minerals, vitamins, and proteins through traditional breeding, modern biotechnological methods, and agronomic approaches (Garg et al. [2018;](#page-29-7) Yeken et al. [2018;](#page-43-12) Saini et al. [2020\)](#page-39-10). It is a long-term and sustainable approach, and a cost-effective way to overcome hidden hunger, which is a progressively severe universal challenge for humanity around the world (De Valença et al. [2017](#page-28-9)). In low-income countries, micronutrient defciencies have largely increased in the last decades. Zn and Fe defciencies in particular are a serious public health problem that negatively affects people's lifespan, health, and productivity (WHO [2009;](#page-42-3) Khan et al. [2008](#page-32-7)). People need cereals for their dietary requirements; hence, biofortifcation of cereals is important worldwide (Saini et al. [2020](#page-39-10)). Wheat is one of the world's most important crops for global food grain production, which was adversely affected by several biotic and abiotic stresses (Ozer et al. [2020\)](#page-37-9). Annual wheat production is expected to increase in the coming years depending on increases of population (Iizumi et al. [2017](#page-31-8)). Biofortifcation can be divided into two categories as agronomic biofortifcation and genetic biofortifcation (Saini et al. [2020\)](#page-39-10). The frst step of biofortifcation in food crops for plant breeders is to understand the current genetic diversity in germplasm collections (Baloch et al. [2014\)](#page-26-8). Wheat has a large number of wild relatives that can lead to its genetic development (Dempewolf et al. [2017;](#page-28-10) Ahmadi et al. [2018;](#page-24-2) Saini et al. [2020](#page-39-10)). The most frequently required mineral elements in the human diet can be obtained from genetic variations, which improve the levels of nutrients in crops (White and Broadley [2005;](#page-42-4) Bouis and Saltzman [2017\)](#page-26-9). Agronomical biofortifcation techniques include fertilizing crops with different fertilizers containing elements such as zinc, iron, and selenium, while genetic biofortifcation includes traditional and molecular breeding approaches. These techniques have the potential to increase the levels of these minerals in grains (Saini et al. [2020\)](#page-39-10). Monasterio and Graham ([2000\)](#page-35-9) claimed that iron and zinc concentrations especially in some bread wheat genotypes were negatively correlated with *Rht* genes. They also reported that the high-yielding wheat cultivars developed after Green Revolution contained less iron and zinc compared to old cultivars/landraces. Heidari et al. [\(2016](#page-30-8)) reported that landraces had higher Fe and Zn concentrations compared with commercial cultivars. Ram and Govindan [\(2020](#page-38-10)) clarifed that genetic diversity in wheat landraces and wild relatives provides novel alleles for genetic enhancement of Zn and Fe. Lyons et al. ([2005\)](#page-34-8) examined 665 wheats (ancestral and wild relatives, landrace accessions, and registered cultivars) in Australia and Mexico for Se concentration in grain. They found that Se

concentrations of grains changed between 5 and 720 microgr/kg. Khokhar et al. [\(2020](#page-32-8)) studied 245 bread wheat genotypes derived from crosses with landraces and the modern wheat cultivar Paragon to detect grain Zn concentration, and they reached promising results for high level of grain Zn where Zn concentration in wholegrain was positively correlated with Fe concentration and grain protein content. They claimed that landraces have a huge potential to increase the concentration of Zn in whole grain and flour of modern high-yielding bread wheat cultivars.

11.3.3.2 Landraces for Some Important Quality Traits

It is generally known that old landraces or cultivars have a huge diversity for some quality traits such as grain protein content, grain texture (hardness), and gluten strength and quality (glutenin and gliadin subunits) than modern wheat cultivars (Aguiriano et al. [2006;](#page-24-3) Moragues et al. [2006;](#page-36-8) Ruiz et al. [2012](#page-39-11)). The grain protein content (GPC) is a crucial trait in determining the quality of wheat (Veraverbeke and Delcour [2002](#page-41-3)), and modern wheat grains include inherently low protein levels. Hence, breeding for an increase in the protein levels of grain wheat is required to alleviate hunger and nutrient defciencies. However, the grain protein content was negatively related to grain yield (Blanco et al. [2006](#page-26-10); Iqbal et al. [2007;](#page-31-9) Klindworth et al. [2009\)](#page-33-6). Avivi ([1978\)](#page-25-10) claimed that wild emmer wheat (*T. turgidum* ssp. *dicoccoides*) can be a potential gene source to improve grain protein content in modern wheat. Joppa and Cantrell [\(1990](#page-31-10)) also studied this hypothesis that they crossed wild emmer wheat and durum wheat, and obtained substitution lines with high GPC. Joppa et al. [\(1997](#page-32-9)) reported that a QTL explained 66% of total variation in these substitution lines for GPC. The QTL was named as *Gpc-B1* (Distelfeld et al. [2004\)](#page-28-11), and Uauy et al. ([2006a](#page-41-4)) also positionally cloned the locus and renamed as *NAM-B1*. Hagenblad et al. [\(2012](#page-30-9)) studied 367 bread wheat germplasm with worldwide origin and determined that five accessions had wild-type *NAM-B1* allele where it confers high levels of protein and microelements. They also indicated that several accessions with wild-type *NAM-B1* were traced back to Fennoscandian origin. In addition to landraces, cultivated transitional forms of wheat such as einkorn (*T. monococcum* ssp. *monococcum*), emmer (*T. turgidum* ssp. *dicoccum*), and spelt (*T. aestivum* ssp. *spelta*) and wild relatives have the possibility to contain the wildtype *NAM-B1* allele. Uauy et al. (Uauy et al. [2006a](#page-41-4); [b](#page-41-5)) reported that wild emmer accessions and most of cultivated emmer accessions studied had wild-type *NAM-B1* allele. Asplund et al. [\(2010](#page-25-11)) also determined that only two spelts had a wild-type *NAM-B1* allele among 62 wheat germplasm displayed at the International Exhibition in London in 1862. It's likely that unique variants for grain protein content can be uncovered due to higher genetic diversity of landraces.

As another important trait, endosperm texture is mainly controlled by the *Hardness* (*Ha*) locus located in 5DS, and it's simply inherited despite the fact that softness is the dominant trait. The lipid binding proteins, puroindoline genes (*Pina-D1* and *Pinb-D1*), which are tightly linked to *Ha* locus, have been used to determine the differences between hard- and soft-textured wheats, and landraces

that originated from different geographic regions had different *Puroindoline* allele combinations. As an example of this situation, Ayala et al. ([2013\)](#page-26-11) studied 102 lines selected from 15 Mexican landraces and determined that while 16 lines had hard texture, 86 lines were soft-textured. Ten out of 16 lines had presence of both *Pina-D1* and *Pinb-D1* alleles. They concluded that the Mexican old landraces are potential sources for important quality traits to develop new wheat varieties with hard grain texture. Li et al. [\(2019](#page-34-9)) also studied 107 Chinese wheat cultivars and landraces in terms of diversity of *Puroindoline* genes and their association with kernel hardness. The most frequent combinations were *Pina-D1a/Pinb-D1a* and *PinaD1a/Pinb-D1b* with 39.3% and 34.6% ratios, respectively. They indicated that Chinese landraces had more allelic than do cultivars and are a valuable source of genetic variability in *Puroindoline* genes. Gluten strength and quality are other important quality traits of wheat. Many studies were conducted to determine the genetic variability of old durum wheat cultivars or landraces for glutenin and gliadin profles, which affected viscoelastic properties of dough, especially in Mediterranean basin (Melnikova et al. [2010;](#page-35-10) Xynias et al. [2011](#page-43-13); Ribeiro et al. [2011](#page-38-11); Ruiz et al. [2012;](#page-39-11) Janni et al. [2018\)](#page-31-11). Nazco et al. [\(2012](#page-36-9)) studied the variability of some quality traits such as protein content, SDS sedimentation, and yellow color index and gluten strength in 154 durum wheat landraces from 20 Mediterranean countries with 18 modern wheat cultivars. They determined that the largest variability for quality traits was observed in landraces from eastern Mediterranean basin followed by landraces from western Mediterranean basin, and identifed landraces could be used to improve quality traits especially for gluten strength and grain weight in durum wheat breeding programs. While *Glu-A1c* was the most frequent allele in almost all genetic materials studied for *Glu-A1* locus, but *Glu-A1a* was found at low frequency in Mediterranean basin (Mir Ali et al. [1999](#page-35-11); Moragues et al. [2006;](#page-36-8) Naghavi et al. [2009](#page-36-10)). In addition to *Glu-A1a*, *Glu-A1b*, and *Glu-A1VI*, encoding the subunits 2* and 2*** were determined at very low frequency. However, Henkrar et al. [\(2017](#page-30-10)) reported that in Moroccan genotypes, *Glu-A1a* and *Glu-A1b* were the predominant alleles. On the other hand, at the *Glu-B1* locus, there were more genetic variation between genotypes with *Glu-B1b*, *Glu-B1d*, and *Glu-B1e* alleles encoding the subunits 7+8, 6+8, and 20, respectively. Moreover, the variation varied geographically that while *Glu-B1d* allele was predominant in Algerian, Syrian, and Spanish germplasm (Mir Ali et al. [1999](#page-35-11); Moragues et al. [2006;](#page-36-8) Hamdi et al. [2010\)](#page-30-11), the allele was not present in Iranian landraces that they had more *Glu-B1a*, *Glu-B1e*, and *Glu-B1i* alleles (Naghavi et al. [2009](#page-36-10)). Similar genetic variation was determined for low molecular weight glutenin subunits (LMW-GS). Li et al. ([2009\)](#page-34-10) studied 615 Chinese wheat germplasm including 390 landraces and 225 varieties, for HMW-GS, LMW-GS, Zeleny sedimentation, volume, dough development time, stability time, and strength, and reported that genetic materials with good gluten strength and quality were identifed in landraces that did not contain wheat-rye translocation. Wheat-rye (the 1BL/1RS) translocation has been used widely in breeding programs because of its disease resistance genes especially for foliar diseases and increased grain yield in some environments, but it negatively affects bread-making quality of wheat at the same time (Zhao et al. [2012;](#page-43-14) Oak and Tamhankar [2017\)](#page-37-10).

On the other hand, the new technologies such as sequencing, mapping, and other related technologies have been recently used to reveal genetic diversity and novel variants/alleles among landraces related to quality traits of wheat. For instance, Giraldo et al. ([2016\)](#page-29-8) performed an association mapping study with 183 Spanish wheat landraces using 749 DArT markers for 18 agromorphological and grain quality traits including protein content, gluten strength, vitreousness, yellow color index, thousand kernel weight, and test weight. They identifed 85 stable MTAs (markertrait associations) with more than 10% explained phenotypic variation, and claimed that novel MTAs were identifed and can provide new information to understand genetic control of complex traits. Roselló et al. ([2018\)](#page-38-0) also performed an association mapping study with 165 durum wheat landraces from 21 Mediterranean countries using 1149 DArT markers. Landraces had generally higher GPC than modern ones in this study but lower gluten strength. In addition to this, while eastern landraces showed the highest yellow color index, Balkan landraces had the lowest test weight. They also identifed 15 meta-QTL (MQTL) for grain quality traits of wheat.

Various studies about landraces conducted in different countries have been briefy summarized and discussed above. We hope that improving the grain quality via agronomic/genetic biofortifcation and quality breeding studies and producing wheat genotypes with better quality will be beneficial to prevent hidden hunger and to live healthy. In this regard, collaboration among various specialists from public and private research institutes and universities can accelerate the improvement of wheat varieties with high bread- and pasta-making quality. This section will be helpful for wheat breeders, providing knowledge of the advancement made so far in wheat biofortifcation and quality.

11.4 Role of Landraces in Biotic Stress

There are many studies conducted to discover resistance properties of wheat landraces for different biotic stresses. Since in the wild the host and the pathogen have co-lived in mutual habitats for long periods of time, they co-evolved together. Thus, the sources of resistance can be found most often at these centers of origin, among the wild relatives and landraces of wheat (McIntosh et al. [1995](#page-35-12)). Pinpointing the resistance factors and genes in the genome and development of molecular markers to test their presence are of great importance.

11.4.1 Role of Wheat Landraces in Disease Resistance

11.4.1.1 Role of Wheat Landraces in Rust Diseases

11.4.1.1.1 Yellow Rust or Stripe Rust

Rust diseases of wheat are among most important and economically devastating diseases of wheat. Rust diseases of wheat consist of yellow (stripe) rust (YR) caused by *Puccinia striiformis* f. sp. *tritici*, leaf rust (LR) caused by *Puccinia triticina*, and stem rust (SR) caused by *Puccinia graminis* f. sp. *tritici* (Reynolds and Borlaug [2006a](#page-38-3); [b](#page-38-4)). Genes that confer resistance to the rust diseases are generally designated as *Yr*, *Lr*, and *Sr* for the effectiveness against yellow rust, leaf rust, and stem rust, respectively. Resistances against rust diseases are the most studied resistance properties in wheat landraces. To date, some genes against rust diseases have been identifed from landraces and wild relatives of wheat. Among them, *Sr2* gene, which provides resistance against stem rust, has been incorporated from an emmer wheat landrace (McIntosh et al. [1995](#page-35-12)). Race-nonspecifc resistance genes *Yr52*, *Yr56*, *Yr57*, and *Yr62*, which provide adult plant resistance (APR) against yellow rust, have been also incorporated from landraces (Mondal et al. [2016\)](#page-35-13).

Yellow rust or stripe rust is one of the most prevalent and devastating wheat foliar diseases worldwide (Kumar et al. [2016\)](#page-33-7). It is observed mostly on cool and moist regions and causes lower kernel quality and massive yield losses (Chen et al. [2013\)](#page-27-7). Recently, there are many studies on YR done by genome wide association studies (GWAS) using bread and durum wheat landraces (Tehseen et al. [2020](#page-40-9); Long et al. [2019;](#page-34-11) Liu et al. [2017a;](#page-34-3) [b;](#page-34-4) [c;](#page-34-5) Manickavelu et al. [2016\)](#page-35-14). Wu et al. [\(2016](#page-42-5)) used simple sequence repeats (SSR), sequence-related amplifed polymorphism (SRAP), and resistance gene analog polymorphism (RGAP) markers, Ma et al. [\(2015](#page-34-12)) used SSR and SRAP markers, while Wang et al. ([2010\)](#page-41-6) used SSR markers to fnd the source of resistance in a known resistant wheat landrace. Kandel et al. [\(2017](#page-32-10)) used microsatelite markers to pinpoint the resistance in the genome of known resistant wheat landrace. Wu et al. [\(2015](#page-42-6)) used molecular markers to screen wheat landraces to fnd a suppressor gene of the known resistance gene *Yr18*. Li et al. [\(2015](#page-33-8)) used DArTseq genotyping-by-sequencing (GBS) on 8416 Mexican Creole landrace wheats and found seven accessions from them with less than 20% disease severity after YR inoculation. Gessese et al. [\(2019](#page-29-9)) screened resistant landrace Aus27430 with 90K wheat SNP chip array by selective genotyping to locate a new resistance gene "*Yr81*." Yuan et al. [\(2018](#page-43-15)), Wang et al. [\(2019](#page-42-7)), and Liu et al. ([2020\)](#page-34-13) used also wheat SNP chip to locate resistance characteristics of wheat landraces. Bux et al. [\(2012](#page-27-8)) evaluated Pakistani wheat landraces phenotypically against the disease; on the other hand Akar et al. [\(2009](#page-24-4)) used durum wheat landraces from Turkey to evaluate their resistance against the yellow rust disease. Rola et al. ([2019\)](#page-38-12) have found two Lebanese wheat landraces that are resistant to different yellow rust pathogen races, including the devastating Warrior pathotype. Wamalwa et al. [\(2020](#page-41-7)) found that Kenyan Kenya Tai landrace shows resistance against many YR races. Mohammadi et al. ([2015\)](#page-35-15) screened 380 durum wheat landraces and found 46 accessions to be resistant against YR.

11.4.1.1.2 Leaf Rust

Leaf rust is one of the main wheat diseases seen worldwide, which can affect kernel weight and wheat biomass, causing major yield losses (Herrera-Foessel et al. [2006\)](#page-30-12). There are many studies done on leaf rust resistance. Qureshi et al. ([2018\)](#page-38-13) identifed a novel resistance gene "*Lr79*," from genotyping analysis of resistant durum wheat by using DArT-seq and 90K chip array, and also developed a Kompetitive Allele Specifc Polymerase (KASP) marker to locate the gene. Kolmer et al. ([2018\)](#page-33-9) used DArT-seq technology to genotype Uruguayan wheat landrace Americano 44. Qureshi et al. [\(2017](#page-38-14)) used DArT-seq markers to locate disease resistance in the genome of two wheat landraces from Portugal. Zhang et al. ([2019a](#page-43-5)), [b\)](#page-43-6) screened 46 Chinese wheat landraces for resistance against LR and used molecular markers to fnd out the presence of known resistance genes in those accessions. Akcura et al. [\(2017](#page-24-5)) used Turkish wheat landraces, while Riaz et al. [\(2017](#page-38-15)) used 136 wheat landraces from Vavilov Institute of Plant Genetic Resources in Russia to test against YR phenotypically. Andenow et al. [\(1997](#page-25-12)) used ten Ethiopian tetraploid wheat (*Triticum turgidum* L.) landraces and found some degree of resistance toward the YR disease.

11.4.1.1.3 Stem Rust

Stem rust is one of the major diseases of wheat which hinders with the nutrient fow to developing ears and result in shriveling of the grain and the breakage of the stem that can cause total yield loss (Roelfs et al. [1992](#page-38-16); Leonard and Szabo [2005](#page-33-10)). Studies on SR have been conducted by Babiker et al. [\(2015](#page-26-12)) and Zurn et al. [\(2014](#page-43-16)), which used quantitative trait loci (QTL) and linkage map, respectively, to locate the resistance region in the known resistant landrace against stem rust pathogen. Haile et al. [\(2013](#page-30-13)) used molecular markers for genotyping the Ethiopian durum wheat landraces. Newcomb et al. ([2013\)](#page-36-11) and Toor et al. ([2013\)](#page-40-10) have screened the landrace collection phenotypically against the SR disease and genotyped using molecular markers. Denbel and Badebo [\(2012](#page-28-12)) screened Ethiopian durum wheat landraces against SR race Ug99. On the other hand, Endresen et al. ([2011\)](#page-28-13) used ecogeographic data of landrace accessions to predict the resistance against SR according to climatic factors of their location of origin, while Bonman et al. ([2007\)](#page-26-13) studied the geographic origin or the resistant accessions. There are also studies conducted to fnd multiple rust resistance in wheat landraces. Studies which include resistance against all three rust diseases were conducted by DArT and molecular markers (Rahmatov et al. [2019;](#page-38-17) Bansal et al. [2013](#page-26-14)) by GWAS and resistance gene prediction (Kankwatsa et al. [2017;](#page-32-11) Pasam et al. [2017;](#page-37-11) Jordan et al. [2015;](#page-32-12) Daetwyler et al. [2014\)](#page-27-9). Kertho et al. [\(2015](#page-32-13)) studied YR and SR resistance traits with GWAS technique, Sthapit et al. [\(2014](#page-40-11)) used simple sequence repeat (SSR) markers to study YR

and SR resistance, and Aoun et al. [\(2019](#page-25-13)) used QTL in durum wheat to locate the resistance region against LR and SR in the known resistant durum wheat landrace.

11.4.1.2 Role of Wheat Landraces in Powdery Mildew (PM)

Powdery mildew (PM) is a foliar fungal disease caused by *Blumeria graminis* f. sp. *tritici*, an obligate biotrophic fungus that causes yield and quality loss in wheat grains (Newton et al. [2011](#page-36-12)). Chinese wheat landraces known for their PM resistance were screened by microsatelite markers (Xue et al. [2009,](#page-43-17) Huang et al. [2000](#page-31-12)), SSR markers (Qie et al. [2019](#page-37-12); Sun et al. [2018;](#page-40-12) Fu et al. [2017;](#page-29-10) Wang et al. [2015](#page-42-8); Xu et al. [2015;](#page-42-9) Fu et al. [2013](#page-29-11); Xue et al. [2012](#page-42-10)), and RNA-seq SNP markers (Li et al. [2020](#page-34-14), Xu et al. [2018](#page-42-11)) to locate genes in the plants' genome, responsible for the resistance trait. Li et al. [\(2018a;](#page-33-11) [b\)](#page-33-12) used SSR marker to pinpoint resistance in an Afghan wheat landrace. Tan et al. [\(2019](#page-40-13)) and Tan et al. ([2018\)](#page-40-14) used single Iranian and Afghan PM-resistant wheat landrace to defne new resistance genes "*Pm63*" and "*Pm59*," respectively, using SSR markers. Identifcation of germplasm strategy (FIGS) was used on wheat landraces in a study conducted by Wang et al. ([2015\)](#page-42-8), Bhullar et al. [\(2010](#page-26-15)), and Bhullar et al. ([2009\)](#page-26-16) to discover new alleles of powdery mildew resistance gene *Pm3*. Huang [\(1997](#page-31-13)) also used APR against powdery mildew found in the landrace accession k-15560, and monosomic and hybridological analyses were used to locate the gene (Peusha et al. [2002\)](#page-37-13). Amplifed fragment length polymorphism (AFLP) markers and microsatellite markers were used to locate Pm24 resistance gene in a Chinese spring wheat landrace. Li et al. ([2012\)](#page-34-15) used SSR markers to test the diversity of the single wheat landrace and its relation to the PM resistance. In their study, Li et al. ([2016a](#page-33-13); [b](#page-34-16)) used 1,297 landraces from 57 countries to screen for the PM resistance, and molecular markers were used to check the presence of known resistance genes. Hysing et al. [\(2007](#page-31-14)) screened 155 Nordic wheat landraces phenotypically and with molecular markers for resistance to PM.

11.4.1.3 Role of Wheat Landraces in Fusarium Head Blight (FHB)

Fusarium head blight (FHB) is caused by the fungal pathogen *Fusarium graminearum* Schwabe and has destructive effects on cereals and especially on wheat production all over the world. Moreover, the diseased plants become contaminated with mycotoxins which are poisonous to mammals (Cetin and Bullerman [2005;](#page-27-10) Goswami and Kistler [2004](#page-30-14)). Cai et al. [\(2019](#page-27-11)) used meta-analysis of previous QTL studies (MQTL) of fve wheat landraces to construct a consensus map, and they also developed 22 KASP markers to ease the MAS in breeding programs. Xiao et al. [\(2011](#page-42-12)) located a chromosomal region responsible for FHB resistance by fastneutron induced chromosome fragment deletion, causing the resistant wheat landrace to lose its resistance and become susceptible. Li et al. [\(2016a,](#page-33-13) [b\)](#page-34-16) used SSR and sequence-tagged site (STS) markers in 195 wheat accessions to fnd the presence of known resistance genes, whereas Wei et al. ([2005\)](#page-42-13) used microsatellite markers to

compare the difference between 20 resistant wheat landraces and 4 susceptible wheat lines. Xiao et al. ([2013\)](#page-42-14) used RNA sequencing to determine expression of a resistant wheat landrace during FHB infection. There are also studies where wheat landraces known for their resistance against Fusarium head blight have been screened with SSR markers to pinpoint the resistance source in the genome (Cai et al. [2016](#page-27-12); Zhang et al. [2012](#page-43-18); Li et al. [2011\)](#page-34-17). Talas et al. [\(2011](#page-40-15)) screened 68 Syrian durum wheat landraces and Yu et al. (2008) screened 94 wheat accessions to fnd new sources of resistance to FHB.

11.4.1.4 Role of Wheat Landraces in Septoria Tritici Blotch (STB)

Septoria tritici blotch (STB) is major foliar wheat disease caused by the fungal pathogen *Zymoseptoria tritici* previously known as *Mycosphaerella graminicola*. It is a major threat to wheat production globally, and it is the most damaging pathogen of wheat in Europe causing loss in chlorophyll, premature death of leaves, and reduction of grain production (O'Driscoll et al. [2014](#page-36-13); Ziv and Eyal [1977](#page-43-19)). Many European and Chinese landraces have been found to contain *Stb6* gene which provides resistance against STB (Chartrain et al. [2005a;](#page-27-13) [b](#page-27-14)). Kidane et al. ([2019\)](#page-32-14) used 318 Ethiopian wheat landraces for GWAS analysis and found four putative loci for STB resistance. Ouaja et al. ([2020\)](#page-36-14) screened 304 Tunisian wheat landraces, and Ghaneie et al. [\(2012](#page-29-12)) screened 45 tetraploid Iranian wheat landraces to test against STB disease phenotypically and found some promising accessions.

11.4.1.5 Role of Wheat Landraces in Tan Spot

Tan spot is caused by *Pyrenophora tritici-repentis* and is an important foliar wheat disease causing severe loss in the grain yield. The disease causes large-scale chlorosis and tan necrosis on leaves and grain shriveling (Maraite et al., [1997](#page-35-16), de Wolf et al. [1998](#page-28-14)). In their study, Gurung et al. ([2011\)](#page-30-15) assessed the resistance of 567 wheat landraces against *P. tritici-repentis* races 1 and 5 using DArT markers and developed association mapping.

11.4.1.6 Role of Wheat Landraces in Eyespot

Eyespot is caused by soilborne necrotrophic funguses *Oculimacula acuformis* and *Oculimacula yallundae*. The disease is seen in temperate areas and affects the stem base of the cereals including wheat, causing premature grain ripening and heavy crop losses (Crous et al. [2003,](#page-27-15) Fitt et al. [1990](#page-29-13), Scott and Hollins [1974\)](#page-39-12). Burt et al. [\(2014](#page-27-16)) screened all 1056 hexaploid wheat landraces of Watkins collection against both funguses and found two promising accessions with high level of resistance. They also genotyped the accessions that showed resistance to one or both funguses by SSR, STS, and QTL-linked markers.

11.4.1.7 Role of Wheat Landraces in Stagonospora Nodorum Blotch (SNB)

Stagonospora nodorum blotch (SNB) is caused by *Phaeosphaeria nodorum* and constitutes a serious disease of wheat worldwide (Eyal [1987\)](#page-28-15). SNB disease infects both leaves and glumes, subsequently causing decreased grain quality and yield losses (King et al. [1983](#page-33-14)). Adhikari et al. ([2011a,](#page-24-6) [b](#page-24-7)) evaluated 567 spring wheat landraces of different origin for resistance to SNB and used DArT markers to genotype and develop association map of the resistance traits.

11.4.1.8 Role of Wheat Landraces in Bacterial Leaf Streak (BLS)

Bacterial leaf streak (BLS) is caused by *Xanthomonas translucens* pv. *undulosa*, the most important wheat bacterial pathogen which can cause major outbreaks in the wheat felds under favorable conditions (Adhikari et al. [2011b](#page-24-7), Bragard et al. [1997\)](#page-27-17). Adhikari et al. ([2012\)](#page-24-8) screened 566 spring wheat landraces for resistance against BLS and used DArT markers to generate association mapping of the resistance regions. They found fve genomic regions which are associated with resistance to the BLS disease.

11.4.1.9 Role of Wheat Landraces in Spot Blotch (SB)

Spot blotch (SB) is caused by *Cochliobolus sativus* which is a fungal disease of wheat and barley, observed globally which results in severe yield losses (Kumar et al. [2002\)](#page-33-15). Adhikari et al. [\(2012](#page-24-8)) screened 566 spring wheat landraces also for resistance against SB and used DArT markers to create association mapping of the resistance regions. They found four genomic regions which are associated with resistance to the SB disease.

11.4.1.10 Role of Wheat Landraces in Common Bunt (CB)

Common bunt (CB) is caused by the fungal pathogen *Tilletia tritici* that causes signifcant yield losses in spring and winter wheat production worldwide (Goates and Peterson [1999](#page-29-14)). Bonman et al. ([2006\)](#page-26-17) investigated 10,759 wheat accessions for resistance against the common bunt disease. Accessions from Bakhtaran province in Iran showed the most resistance.

11.4.1.11 Role of Wheat Landraces in Dwarf Bunt (DB)

Dwarf bunt (DB) is caused by the fungus *Tilletia controversa* in winter wheat in regions where snow is persistent (Goates and Peterson [1999\)](#page-29-14). Bonman et al. [\(2006](#page-26-17)) studied 8167 wheat accessions against dwarf bunt resistance. Accessions from Hakkari province in Turkey showed the highest resistance against DB.

11.4.1.12 Role of Wheat Landraces in Wheat Blast (WB)

Wheat blast (WB) is a relatively new emerging disease (mid-1980s) caused by *Triticum* pathotype of *Pyricularia oryzae* fungus. It has immense impacts on wheat production (Inoue et al. [2017](#page-31-15)). Wang et al. [\(2018a,](#page-41-8) [b](#page-42-15)) evaluated 520 landraces of common wheat from different regions of the world for the resistance to Br48 isolate of the fungus and found a unique accession resistant to WB. The resistance was due to combination effect of two genes "*Rmg8*" and newly found "*RmgGR119*" gene.

11.4.2 Role of Wheat Landraces in Pest Resistance

11.4.2.1 Role of Wheat Landraces in Root Lesion Nematodes

Root lesion nematodes *Pratylenchus thornei* and *Pratylenchus neglectus* are the most common root lesion parasites that grow and develop in wheat roots, causing damage and substantial losses in wheat production (Nicol et al. [2002\)](#page-36-15). Thompson and Seymour [\(2011](#page-40-16)) analyzed the modes of inheritance of resistance to *P. thornei* in seven wheat accessions that showed resistance against the nematode. Schmidt et al. [\(2005](#page-39-13)) studied two resistant Middle Eastern wheat landraces with AFLP and microsatellite markers for QTL analysis of resistance to *P. thornei*. Thompson et al. ([2009\)](#page-40-17) screened 207 bread wheat and 102 durum wheat accessions from West Asia and North Africa for resistance against *P. thornei*. Among them, 13 bread wheat and 10 durum wheat showed signifcant resistance. Thompson et al. [\(2016](#page-40-18)) screened 78 Iranian wheat accessions for resistance against *P. thornei* and *P. neglectus*. Among them, 32 showed some degree of resistance to both nematodes.

11.4.2.2 Role of Wheat Landraces in Russian Wheat Aphid (RWA)

Russian wheat aphid (RWA) (*Diuraphis noxia*) is an important wheat pest indigenous to southern Russia and Mediterranean countries which have spread to all continents causing substantial damage to wheat felds (DuToit and Walters [1984;](#page-28-16) Hewitt et al. [1984\)](#page-30-16). Valdez et al. [\(2012](#page-41-9)) have evaluated a resistant Iranian wheat landrace using SSR markers to identify the location of resistance trait. It was found that the trait was due to dominant gene. Similarly, Li et al. ([2018a\)](#page-33-11) used an Iranian wheat

landrace known for its resistance to RWA to locate the trait in the genome using SSR markers.

11.4.2.3 Role of Wheat Landraces in Wheat Stem Sawfy (WSS)

Wheat stem sawfy (WSS), *Cephus cinctus* Norton, is a major pest insect of wheat observed in North America, with devastating consequences in wheat production (Michael et al. 1992). Mohammadi et al. ([2015\)](#page-35-15) evaluated the collection of 380 durum wheat landraces against WSS and found that 33 accessions showed resistance to the pest. Varella et al. ([2017\)](#page-41-10) screened 1409 accessions of wheat landraces collected from different regions to WSS. They found 204 accessions that have resistance to the disease. The resistant accessions were screened with KASP markers for QTL analysis. Varella et al. [\(2019](#page-41-11)) used four resistant wheat accessions and generated six recombinant inbred lines (RIL) with them and genotyped with 90K iSelect assay to fnd novel QTL related to WSS resistance.

11.4.2.4 Role of Wheat Landraces in Cereal Cyst Nematodes (CCN)

Cereal cyst nematodes (CCN) *(Heterodera* spp.*)* are a group of 12 known species with *H. avenae*, *H. flipjevi*, and *H. latipons* being the most important ones. The pest is observed in many regions of the world and causes major yield losses in cereals (Nicol et al. [2003](#page-36-16)). Yavuzaslanoglu et al. (2016) studied the response of 31 Iranian wheat landraces against *H. filipjevi* and found one resistant and five moderately resistant accessions.

11.4.2.5 Role of Wheat Landraces in Cereal Aphids

Cereal aphids cause important yield losses in wheat. There are 14 species of aphids that were observed causing damage to wheat. *Sitobion avenae*, *Rhopalosiphum maidis*, *R. padi*, and *Metopolophium dirhodum* are the most common of these (Popov et al. [1988](#page-37-14)). Amin et al. [\(2019](#page-25-14)) observed 114 wheat landraces for their resistance level against the disease and population dynamics of *R. padi*. They found promising accessions which can be used for breeding of resistant cultivars.

11.5 Landraces and the Future of Wheat Diversity

The world is confronting food scarcity problem due to rapid increase in population and climate change. Previous report showed 6–13% reduction in wheat yield for each °C rise in temperature. Continuously changing climate, extreme weather events, new pathogen strains, and pests further jeopardize linear productivity growth into the future (Mondal et al. [2016](#page-35-13)). It is believed that the world's population will cross the nine billion mark in 2050. By considering this factor, it is very important to increase wheat production by a rate of 1.6% (Lodhi et al. [2020](#page-34-1)). To feed the rapidly increasing world's population under changing climatic conditions, more pressure is put on agriculture to produce enough quantity of food. Therefore, it is very important to increase the wheat production to serve enough quantity of food. By considering these factors, it is very important to develop wheat cultivars having higher production and better adaptation to biotic and abiotic stresses (Khan et al. [2013\)](#page-32-15). These targets can be achieved by harnessing wheat genetic diversity. Previous studies explored the existence of higher genetic diversity in wheat landraces compared to its commercial cultivars (Lodhi et al. [2020;](#page-34-1) Jaradat [2011;](#page-31-0) Jaradat [2013\)](#page-31-16).

Genetic diversity present in wheat landraces has been successfully utilized for breeding perspectives. Wheat landraces possess a suffcient amount of diversity, including useful genes to adapt to stressful environments such as salinity, heat, and drought (Karagöz and Zencirci [2005](#page-32-16); Özkan et al. [2011\)](#page-37-15). The evaluation of genetic diversity in wheat landraces is important for the selection of the suitable landraces as donors of traits in breeding studies (Gurcan et al. [2017;](#page-30-17) Abbasov et al. [2018\)](#page-24-9). Landraces represent signifcantly broader genetic diversity than modern varieties (Azeez et al. [2018](#page-26-0)). For this reason, they can help to increase the genetic source of modern cultivars. However, for their utilization in breeding programs, it is very important that breeders should make crosses among elite lines having the highest likelihood of developing new varieties (Baenziger and DePauw [2009\)](#page-26-18). There is scarcity of information about the successful release of cultivars using wheat landraces. Gerek 79 which is a Turkish variety is developed through crosses with landraces (Smale and McBride [1996](#page-39-14)). One of the best examples of landraces serving as a source of novel genes is the identifcation of Rht dwarfng gene that was available through the Japanese variety "Norin 10" originating from a Japanese landrace Shiro Daruma (Reitz and Salmon [1968;](#page-38-6) Dreisigacker et al. [2005](#page-28-17)). Dr. Norman E. Borlaug utilized these genes to develop the high-yielding semidwarf wheat varieties that resulted in Green Revolution. Similarly, various wheat landraces served as a foundation in the wheat germplasm pool impotent like: "Cheyenne," a selection from landrace Crimea, founded the Nebraska wheat gene pool. Moreover, "Turkey Red" has been successfully used in winter wheat breeding in the US Great Plains (Lopes et al. [2015](#page-34-0)). Similarly, previous studies confrmed landrace diversity as a potential source for the breeding of grain yield and climate resilience, for example, the drought-tolerant variety "Aragon 03" was developed from a selection of a landrace population "Catalan de Monte" (Royo and Briceño-Félix [2011a;](#page-38-1) [b\)](#page-39-0). Vikram et al. [\(2016a;](#page-41-12) [b](#page-41-13)) stated that a group of Creole wheat landraces (the landraces introduced to Mexico from Europe) has better adaptation to various abiotic stresses including drought because of the presence of rare but benefcial alleles. Further, wheat landraces refected genetic diversity for various traits like 1000-kernel weight, biomass, and photosynthesis that can be used for cultivar development (Lopes et al. [2015\)](#page-34-0). Various studies have been conducted using wheat landraces as germplasm through molecular markers and explored their potential as a source of novel variations (Sansaloni et al. [2020](#page-39-15); Alipour et al. [2017](#page-25-15); Lopes et al. [2015](#page-34-0); Sofalian et al. [2008;](#page-39-16)

Alsaleh et al. [2015](#page-25-16); Jorgensen et al. [2017;](#page-32-17) Arystanbekkyzy et al. [2019;](#page-25-17) Dababat et al. [2020;](#page-27-18) Ozer et al. [2020](#page-37-9)). As is obvious from the above-provided information, there is a need to utilize wheat landrace diversity to develop climate-resilient cultivars having high yield. Similarly, some nonbreeding efforts that should be used to promote on-farm dynamic conservation and sustainable utilization of wheat landraces include the following:

- 1. Awareness should be raised in the farming community about their potential in changing climate.
- 2. Availability of wheat landrace seeds to the farmers.
- 3. Development of niche market for landrace products.
- 4. Involvement of wheat breeders, seed producers, farmers, and end-users, as stakeholders in wheat breeding activities to develop new cultivars (Newton et al. [2011\)](#page-36-12).

References

- Abbasov M, Akparov Z, Gross T, Babayeva S, Izzatullayeva V, Hajiyev E, Rustamov K, Gross P, Tekin M, Akar T, Chao S (2018) Genetic relationship of diploid wheat (*Triticum* spp.) species assessed by SSR markers. Genet Resour Crop Evol 65(5):1441–1453
- Abu-Zaitoun SY, Chandrasekhar K, Assili S, Shtaya MJ, Jamous RM, Mallah OB, Nashef K, Sela H, Distelfeld A, Alhajaj N, Ali-Shtayeh MS (2018) Unlocking the genetic diversity within a Middle-East panel of durum wheat landraces for adaptation to semi-arid climate. Agronomy 8(10):233
- Adhikari TB, Gurung S, Hansen JM, Jackson EW, Bonman JM (2012) Association mapping of quantitative trait loci in spring wheat landraces conferring resistance to bacterial leaf streak and spot blotch. Plant Genome 5(1):1–16
- Adhikari TB, Jackson EW, Gurung S, Hansen JM, Bonman JM (2011a) Association mapping of quantitative resistance to Phaeosphaeria nodorum in spring wheat landraces from the USDA National Small Grains Collection. Phytopathology 101(11):1301–1310
- Adhikari TB, Hansen JM, Gurung S, Bonman JM (2011b) Identifcation of new sources of resistance in winter wheat to multiple strains of Xanthomonas translucens pv. undulosa. Plant disease 95(5):582–588
- Aguiriano E, Ruiz M, Fite' R, Carrillo JM (2006) Analysis of genetic variability in a sample of the durum wheat (*Triticum durum* Desf.) Spanish collection based on gliadin markers. Genet Resour Crop Evol 53(8):1543–1552
- Ahmad M, Shahzad A, Iqbal M, Asif M, Hirani AH (2013) Morphological and molecular genetic variation in wheat for salinity tolerance at germination and early seedling stage. Aust J Crop Sci 7(1):66
- Ahmadi J, Pour-Aboughadareh A, Ourang SF, Mehrabi AA, Siddique KHM (2018) Wild relatives of wheat: Aegilops–Triticum accessions disclose differential antioxidative and physiological responses to water stress. Acta Physiol Plant 40:1–14
- Ahmadi J, Pour-Aboughadareh A, Ourang SF, Khalili P, Poczai P (2020) Unraveling salinity stress responses in ancestral and neglected wheat species at early growth stage: A baseline for utilization in future wheat improvement programs. Physiol Mol Biol Plants 1–13.
- Akar T, Mert Z, Yazar S, Sanal T, Avci M (2009) Sustainable use of winter Durum wheat landraces under Mediterranean conditions. Afr J Biotechnol 8(17)
- Akcura M, Kadir A, Hocaoglu O (2017) Biplot analysis of leaf rust resistance in pure lines selected from eastern Anatolian bread wheat landraces of turkey. Turkish J Field Crops 22(2):227–234
- Aktaş H (2016) Drought tolerance indices of selected landraces and bread wheat (Triticum aestivum L.) genotypes derived from synthetic wheats. Appl Ecol Environ Res 14(4):177–189
- Al Khateeb W, Schroeder D, Musallam I (2017) Phenotypic and molecular variation in drought tolerance of Jordanian durum wheat (Triticum durum Desf.) landraces. Physiol Mol Biol Plants 23(2):311–319
- Ali ML, Baenziger PS, Ajlouni ZA, Campbell BT, Gill KS, Eskridge KM, Mujeeb-Kazi A, Dweikat I (2011) Mapping QTLs for yield and agronomic traits on wheat chromosome 3A and a comparison of recombinant inbred chromosome line populations. Crop Sci 51:553–566
- Alipour H, Bihamta MR, Mohammadi V, Peyghambari SA, Bai G, Zhang G (2017) Genotypingby-sequencing (GBS) revealed molecular genetic diversity of Iranian wheat landraces and cultivars. Front Plant Sci 8:1293
- Al-maskri A, Hameed M, Ashraf M, Khan MM, Fatima S, Nawaz T, Batool R (2014) Structural features of some wheat (*Triticum spp*.) landraces/cultivars under drought and salt stress. Arid Land Res Manag 28(3):355–370
- Al-Naggar AMM, Abd El-Shaf MAE, El-Shal MH, Anany AH (2020) Evaluation of Egyptian wheat landraces (Triticum aestivum L.) for drought tolerance, agronomic, grain yield and quality traits. Plant Archives 20(Supplement 1):3487–3504
- Alsaleh A, Baloch FS, Derya M, Azrak M, Kilian B, Özkan H, Nachit M (2015) Genetic linkage map of Anatolian durum wheat derived from a cross of Kunduru-1149× Cham1. Plant Mol Biol Rep 33:209–220
- Alsaleh A, Baloch FS, Nachit M, Ozkan H (2016) Phenotypic and genotypic intra-diversity among Anatolian durum wheat "Kunduru" landraces. Biochem Syst Ecol 65:9–16
- Amin M, Mahmood K, Nazir N, Kassi AK, Ahmed S (2019) Population dynamics of wheat aphid on different landraces of wheat under feld conditions. Plant Protect 3(2):59–66
- Andeden EE, Yediay FE, Baloch FS, Shaaf S, Kilian B, Nachit M, Ozkan H (2011) Distribution of vernalization and photoperiod genes (Vrn-A1, Vrn-B1, Vrn-D1, Vrn-B3, Ppd-D1) in Turkish bread wheat cultivars and landraces. Cereal Res Commun 39:352–364
- Andenow Y, Hullukal M, Belay G (1997) Resistance and tolerance to leaf rust in Ethiopian tetraploid wheat landraces. Plant breeding 116(6):533–536
- Aoun M, Kolmer JA, Rouse MN, Elias EM, Breiland M, Bulbula WD, Chao S, Acevedo M (2019) Mapping of novel leaf rust and stem rust resistance genes in the Portuguese durum wheat landrace PI 192051. G3: Genes, Genomes. Genetics 9(8):2535–2547
- Aprile A, Havlickova L, Panna R, Mare C, Borrelli GM, Marone D, Perrotta C, Rampino P, De Bellis L, Curn V, Mastrangelo AM, Rizza F, Cattivelli L (2013) Different stress responsive strategies to drought and heat in two durum wheat cultivars with contrasting water use effciency. BMC Genomics 14:821–838
- Arabbeigi M, Arzani A, Majidi MM, Kiani R, Tabatabaei BES, Habibi F (2014) Salinity tolerance of Aegilops cylindrica genotypes collected from hyper-saline shores of Uremia Salt Lake using physiological traits and SSR markers. Acta Physiol Plant 36(8):2243–2251
- Arabbeigi M, Arzani A, Majidi MM, Sayed-Tabatabaei BE, Saha P (2018) Expression pattern of salt tolerance-related genes in Aegilops cylindrica. Physiol Mol Biol Plants 24(1):61–73
- Arriagada O, Mora F, Quitral Y, Del Pozo A (2017) Identifcation of QTL underlying agronomic, morphological and physiological traits in barley under rainfed conditions using SNP markers. Acta Sci Agron 39:321–329
- Arystanbekkyzy M, Nadeem MA, Aktas H, Yeken MZ, Zencirci N, Nawaz MA, Ali F, Haider MS, Tunç K, Chung G, Baloch FS (2019) Phylogenetic and taxonomic relationship of Turkish wild and cultivated emmer (Triticum turgidum ssp. dicoccoides) revealed by iPBSretrotransposons markers. Int J Agric Biol 21(1):155–163
- Asplund L, Hagenblad J, Leino MW (2010) Re-evaluating the history of the wheat domestication gene *NAM-B1* using historical plant material. J Archaeol Sci 37:2303–2307
- Avivi L (1978) High protein content in wild tetraploid *Triticum dicoccoides* Korn. In Ramanujam S (ed). In: Proceedings of the 5th international wheat genetics symposium, New Delhi, 23–28 Feb 1978. Indian Soc Genet Plant Breed, Indian Agric Res Inst, New Delhi, pp 372–380
- Ayala M, Guzmán AJB, Peña RJ (2013) Characterization of genetic diversity of puroindoline genes in Mexican wheat landraces. Euphytica 190(1):53–63
- Azeez MA, Adubi AO, Durodola FA (2018) Landraces and crop genetic improvement. In Rediscovery of landraces as a resource for the future. IntechOpen.
- Babiker EM, Gordon TC, Chao S, Newcomb M, Rouse MN, Jin Y, Wanyera B, Acevedo M, Brown-Guedira G, Williamson S, Bonman JM (2015) Mapping resistance to the Ug99 race group of the stem rust pathogen in a spring wheat landrace. Theor Appl Genet 128(4):605–612
- Baenziger PS, Depauw RM (2009) Wheat breeding: Procedures and strategies. In: Wheat science and trade. Wiley-Blackwell, Ames, pp 273–308
- Bal W, Kozlowski H, Robbins R, Pettit LD (2010) Improving drought tolerance by exogenous application of glycinebetaine and salicylic acid in sunfower. J Agron Crop Sci 194:193–199
- Blanco A, Simeone R, Gadaleta A (2006) Detection of QTLs for grain protein content in durum wheat. Theor Appl Genet 112:1195–1204
- Ballesta P, Mora F, Del Pozo A (2019) Association mapping of drought tolerance indices in wheat: QTL-rich regions on chromosome 4A. Sci Agri 77(2)
- Baloch FS, Alsaleh A, Andeden EE, Hatipoğlu R, Nachit M, Özkan H (2016) High levels of segregation distortion in the molecular linkage map of bread wheat representing the West Asia and North Africa region. Turk J Agric For 40(3):352–364
- Baloch FS, Alsaleh A, Shahid MQ, Çiftçi V, Aasim M, Nadeem MA, Aktaş H, Özkan H, Hatipoğlu R (2017) A whole genome DArTseq and SNP analysis for genetic diversity assessment in durum wheat from central fertile crescent. PLoS One 12(1):e0167821
- Baloch FS, Karaköy T, Demirbaş A, Toklu F, Özkan H, Hatipoğlu R (2014) Variation of some seed mineral contents in open pollinated faba bean (Vicia faba L.) landraces from Turkey. Turk J Agric For 38(5):591–602
- Bansal UK, Arief VN, DeLacy IH, Bariana HS (2013) Exploring wheat landraces for rust resistance using a single marker scan. Euphytica 194(2):219–233
- Belay G, Tesemma T, Bechere E, Mitiku D (1995) Natural and human selection for purple-grain tetraploid wheats in the Ethiopian highlands. Genet Resour Crop Evol 42:387–391
- Bhullar NK, Street K, Mackay M, Yahiaoui N, Keller B (2009) Unlocking wheat genetic resources for the molecular identifcation of previously undescribed functional alleles at the Pm3 resistance locus. Proc Natl Acad Sci U S A 106(23):9519–9524
- Bhullar NK, Zhang Z, Wicker T, Keller B (2010) Wheat gene bank accessions as a source of new alleles of the powdery mildew resistance gene Pm3: a large scale allele mining project. BMC Plant Biol 10(1):88
- Bonman JM, Bockelman HE, Goates BJ, Obert DE, McGuire PE, Qualset CO, Hijmans RJ (2006) Geographic distribution of common and dwarf bunt resistance in landraces of Triticum aestivum subsp. aestivum. Crop Sci 46(4):1622–1629
- Bonman JM, Bockelman HE, Jin Y, Hijmans RJ, Gironella AIN (2007) Geographic distribution of stem rust resistance in wheat landraces. Crop Sci 47(5):1955–1963
- Börner A, Schumann E, Fürste A, Cöster H, Leithold B, Röder M, Weber W (2002) Mapping of quantitative trait loci determining agronomic important characters in hexaploid wheat (Triticum aestivum L.). Theor Appl Genet 105(6-7):921–936
- Börner A, Worland AJ, Plaschke J, Schumann E, Law CN (1993) Pleiotropic effects of genes for reduced height (Rht) and day-length insensitivity (Ppd) on yield and its components for wheat grown in middle Europe. Plant Breed 111:204–216
- Borojevic K, Borojevic K (2005) Historic role of the wheat variety Akakomugi in Southern and Central European wheat breeding programs. Breed Sci 55:253–256
- Bouffer B (2014) Genetic and ecophysiological dissection of tolerance to drought and heat stress in bread wheat: from environmental characterization to QTL detection (Doctoral dissertation).
- Bouis HE, Saltzman A (2017) Improving nutrition through biofortifcation: a review of evidence from HarvestPlus, 2003 through 2016. Glob Food Sec 12:49–58
- Bragard C, Singer E, Alizadeh A, Vauterin L, Maraite H, Swings J (1997) Xanthomonas translucens from small grains: diversity and phytopathological relevance. Phytopathology 87(11):1111–1117
- Braun HJ, Zencirci N, Altay F, Atli A, Avci M, Eser V, Kambertay M, Payne TS (2001) Turkish wheat pool. In: Bonjean AP, Agnus WJ (eds) The world wheat book: A history of wheat breeding. Lavosier, Paris, pp 851–879
- Budak H, Shearman RC, Parmaksiz I, Gaussoin RE, Riordan TP, Dweikat I (2004) Molecular characterization of Buffalograss germplasm using sequence-related amplifed polymorphism markers. Theor Appl Genet 108:328–334
- Bullrich L, Appendino ML, Tranquilli G, Lewis S, Dubcovsky J (2002) Mapping of a thermosensitive earliness per se gene on Triticum monococcum chromosome 1Am. Theor Appl Genet 105:585–593
- Burt C, Griffe LL, Ridolfni AP, Orford S, Griffths S, Nicholson P (2014) Mining the Watkins collection of wheat landraces for novel sources of eyespot resistance. Plant Pathol 63(6):1241–1250
- Bux H, Ashraf M, Chen X (2012) Expression of high-temperature adult-plant (HTAP) resistance against stripe rust (Puccinia striiformis f. sp. tritici) in Pakistan wheat landraces. Can J Plant Pathol 34(1):68–74
- Cai J, Wang S, Li T, Zhang G, Bai G (2016) Multiple minor QTLs are responsible for Fusarium head blight resistance in Chinese wheat landrace Haiyanzhong. PLoS One 11(9):e0163292
- Cai J, Wang S, Su Z, Li T, Zhang X, Bai G (2019) Meta-analysis of QTL for Fusarium head blight resistance in Chinese wheat landraces. Crop J 7(6):784–798
- Casañas F, Simó J, Casals J, Prohens J (2017) Toward an evolved concept of landrace. Front Plant Sci 8:145
- Chaichi M, Sanjarian F, Razavi K, Gonzalez-Hernandez JL (2019) Phenotypic diversity among Iranian bread wheat landraces, as a screening tool for drought tolerance. Acta Physiol Plant 41(6):1–15
- Cetin Y, Bullerman LB (2005) Cytotoxicity of Fusarium mycotoxins to mammalian cell cultures as determined by the MTT bioassay. Food Chem Toxicol 43(5):755–764
- Chaparzadeh N, Aftabi Y, Dolati M, Mehrnejad F, Pessarakli M (2014) Salinity tolerance ranking of various wheat landraces from the west of the Urmia saline lake in Iran by using physiological parameters. J Plant Nutr 37(7):1025–1039
- Chartrain L, Berry ST, Brown JKM (2005a) Resistance of wheat line Kavkaz-K4500 L. 6. A. 4 to Septoria tritici blotch controlled by isolate-specifc resistance genes. Phytopathology 95(6):664–671
- Chartrain L, Brading PA, Brown JKM (2005b) Presence of the Stb6 gene for resistance to Septoria tritici blotch (Mycosphaerella graminicola) in cultivars used in wheat-breeding programmes worldwide. Plant Pathol 54(2):134–143
- Chebotar GO, Chebotar SV, Motsnyy II, Sivolap YM (2013) Clarifcation of the Rht8–PpdD1 gene linkage on the 2D chromosome of winter bread wheat. Cytol Genet 47:70–74
- Chen W, Wellings C, Chen X, Kang Z, Liu T (2014) Wheat stripe (yellow) rust caused by Puccinia striiformis f. sp. tritici. Mol Plant Pathol 15(5):433–446
- Chen P, You C, Hu Y, Chen S, Zhou B, Cao A, Wang X (2013) Radiation-induced translocations with reduced Haynaldia villosa chromatin at the Pm21 locus for powdery mildew resistance in wheat. Mol Breed 31(2):477–484
- Colmer TD, Flowers TJ, Munns R (2006) Use of wild relatives to improve salt tolerance in wheat. J Exp Bot 57:1059–1078
- Crous PW, Groenewald JE, Gams W (2003) Eyespot of cereals revisited: ITS phylogeny reveals new species relationships. Eur J Plant Pathol 109(8):841–850
- Dababat A, İmren M, Pridannikov M, Özer G, Zhapayev R, Mokrini F, Otemissova A, Yerimbetova A, Morgounov A (2020. Plant-parasitic nematodes on cereals in northern Kazakhstan. J Plant Dis Prot 1–9
- Daetwyler HD, Bansal UK, Bariana HS, Hayden MJ, Hayes BJ (2014) Genomic prediction for rust resistance in diverse wheat landraces. Theor Appl Genet 127(8):1795–1803
- Damania AB, Pecetti L, Qualset CO, Humeid BO (1996) Diversity and geographic distribution of adaptive traits in Triticum turgidum L. (durum group) wheat landraces from Turkey. Genet Resour Crop Evol 43:409–422
- De Valença AW, Bake A, Brouwer ID, Giller KE (2017) Agronomic biofortifcation of crops to fght hidden hunger in sub-Saharan Africa. Glob Food Sec 12:8–14
- De Wolf ED, Effertz RJ, Ali S, Francl LJ (1998) Vistas of tan spot research. Canadian journal of plant pathology= Revue Canadienne de phytopathologie
- Dempewolf H, Baute G, Anderson J, Kilian B, Smith C, Guarino L (2017) Past and future use of wild relatives in crop breeding. Crop Sci 57(3):1070–1082
- Denbel W, Badebo A (2012) Valuable sources of resistance in the Ethiopian durum wheat landraces to Ug33 and other stem rust races. Int. J. Agron. Plant Prod 3:191–195
- Derakhshan B, Mohammadi SA, Moghaddam M, Jalal Kamali MR (2013) Molecular characterization of vernalization genes in Iranian wheat landraces. Crop Breed J 3:11–14
- Devi R, Ram S, Rana V, Malik VK, Pande V, Singh GP (2019) QTL mapping for salt tolerance associated traits in wheat (*Triticum aestivum* L.). Euphytica 215(12):210
- Diamond J (2002) Evolution, consequences and future of plant and animal domestication. Nature 418:700–707
- Distelfeld A, Uauy C, Olmos S, Schlatter AR, Dubcovsky J, Fahima T (2004) Microcolinearity between a 2-cM region encompassing the grain protein content locus *Gpc-6B1* on wheat chromosome 6B and a 350-kb region on rice chromosome 2. Funct Integr Genomics 4:59–66
- Dreisigacker S, Zhang P, Warburton ML, Skovmand B, Hoisington D, Melchinger AE (2005) Genetic diversity among and within CIMMYT wheat landrace accessions investigated with SSRs and implications for plant genetic resources management. Crop Sci 45:653–661
- Dubcovsky J, Lijavetzky D, Appendino L, Tranquilli G (1998) Comparative RFLP mapping of Triticum monococcum genes controlling vernalization requirement. Theor Appl Genet 97:968–975
- DuToit F, Walters MC (1984) Damage assessment and economic threshold values for the chemical control of the Russian wheat aphid, Diuraphis noxia (Mordvilko) on winter wheat. Technical communication-South Africa, Department of Agriculture
- Dvorak J, Luo MC, Yang ZL, Zhang HB (1998) The structure of the Aegilops tauschii genepool and the evolution of hexaploid wheat. Theor Appl Genet 97:657–670
- Dvorak J, Noaman MM, Goyal S, Gorham J (1994) Enhancement of the salt tolerance of Triticum turgidum L by the Kna1 locus transferred from Triticum aestivum L. chromosome 4D by homoeologous recombination. Theor Appl Genet 87:872–877
- Dwivedi SL, Ceccarelli S, Blair MW, Upadhyaya HD, Are AK, Ortiz R (2016) Landrace germplasm for improving yield and abiotic stress adaptation. Trends Plant Sci 21:31–42
- ELshafei AA, Afah SA, Amer MA, El-enany MAM (2019) Validation of molecular markers linked with salinity tolerance in wheat (Triticum aestivum L.) grown on saline soil. Biosci Res 16(2):963–978
- Endresen DTF, Street K, Mackay M, Bari A, DePauw E (2011) Predictive association between biotic stress traits and eco-geographic data for wheat and barley landraces. Crop Sci 51(5):2036–2055
- Epstein E, Bloom AJ (2005) Mineral nutrition of plants: principles and perspectives. Sinauer, Sunderland
- Eyal Z (1987) The Septoria diseases of wheat: concepts and methods of disease management. CIMMYT
- Fageria NK, Stone LF, dos Santos AB (2012) Breeding for salinity tolerance. In plant breeding for abiotic stress tolerance. Springer, Berlin, pp 103–122
- Farooq S, Niazi M, Iqbal N, Shah TM (1989) Salt tolerance potential of wild resources of the tribe Triticeae II. Screening of species of the genus Aegilops. Plant and Soil 119:255–260
- Fei X, Wen-Wen Z, Xia-Yu D, Yi-Lin Z, Wan-Quan J (2009) Microsatellite mapping of a powdery mildew resistance gene in wheat landrace Xiaobaidong. Acta Agronomica Sinica 35(10):1806–1811
- Fitt BD, Goulds A, Hollins TW, Jones DR (1990) Strategies for control of eyespot (Pseudocercosporella herpotrichoides) in UK winter wheat and winter barley. Ann Appl Biol 117(2):473–486
- Flowers TJ (2004) Improving crop salt tolerance. J Exp Bot 55:307–319
- Frankel OH, Bennett E (1970) Genetic resources in plants-their exploration and conservation. In: Genetic resources in plants-their exploration and conservation. Distributed by Blackwell Scientifc, Oxford
- Fu B, Chen Y, Li N, Ma H, Kong Z, Zhang L, Jia H, Ma Z (2013) pmX: a recessive powdery mildew resistance gene at the Pm4 locus identifed in wheat landrace Xiaohongpi. Theor Appl Genet 126(4):913–921
- Fu B, Zhang Z, Zhang Q, Wu X, Wu J, Cai S (2017) Identifcation and mapping of a new powdery mildew resistance allele in the Chinese wheat landrace Hongyoumai. Molecular Breeding 37(11):133
- Gadea M (1958) Trigos cultivados en España y nuevas variedades recomendadas. Ministerio de Agricultura, Madrid
- Galiba G, Quarrie SA, Sutka J, Morgounov A, Snape JW (1995) RFLP mapping of the vernalization (Vrnl) and frost resistance (Frl) genes on chromosome 5A of wheat. Theor Appl Genet 90:1174–1179
- Garg AK, Kim JK, Owens TG, Ranwala AP, Do Choi Y, Kochian LV, Wu RJ (2002) Trehalose accumulation in rice plants confers high tolerance levels to different abiotic stresses. Proc Natl Acad Sci U S A 99:15898–15903
- Garg M, Sharma N, Sharma S, Kapoor P, Kumar A, Chunduri V, Arora P (2018) Biofortifed crops generated by breeding, agronomy, and transgenic approaches are improving lives of millions of people around the world. Front Nutr 5:12
- Genc Y, Oldach K, Verbyla AP, Lott G, Hassan M, Tester M, Wallwork H, McDonald GK (2010) Sodium exclusion QTL associated with improved seedling growth in bread wheat under salinity stress. Theor Appl Genet 121(5):877–894
- Genc Y, Verbyla AP, Torun AA, Cakmak I, Willsmore K, Wallwork H, McDonald GK (2009) Quantitative trait loci analysis of zinc efficiency and grain zinc concentration in wheat using whole genome average interval mapping. Plant and Soil 314:49
- Gessese M, Bariana H, Wong D, Hayden M, Bansal U (2019) Molecular mapping of stripe rust resistance gene Yr81 in a common wheat landrace Aus27430. Plant disease 103(6):1166–1171
- Ghaneie A, Mehrabi R, Safaie N, Abrinbana M, Saidi A, Aghaee M (2012) Genetic variation for resistance to septoria tritici blotch in Iranian tetraploid wheat landraces. Eur J Plant Pathol 132(2):191–202
- Giraldo P, Royo C, Gonzalez M, Carrillo JM, Ruiz M (2016) Genetic diversity and association mapping for agromorphological and grain quality traits of a structured collection of durum wheat landraces including subsp. *durum*, *turgidum* and *diccocon*. PLoS One 11(11):e0166577
- Goates BJ, Peterson GL (1999) Relationship between soilborne and seedborne inoculum density and the incidence of dwarf bunt of wheat. Plant Dis 83(9):819–824
- Gökgöl M (1935) Turkish wheats, vol I. Ministry of Agriculture, Yesilkoy Seed Breeding Institute Publications. No: 7, Devlet Press, Istanbul. (In Turkish), 436 pp.
- Goncharov NP (1998) Genetic resources of wheat related species: The Vrn genes controlling growth habit (spring vs. winter). Euphytica 100(1):371–376
- Gonzalez-Hernandez JL, Elias EM, Kianian SF (2004) Mapping genes for grain protein concentration and grain yield on chromosome 5B of Triticum turgidum (L.) var. dicoccoides. Euphytica 139:217–225
- Gorham J, Bristol A, Young EM, Wyn Jones RG, Kashour G (1990) Salt tolerance in the Triticeae: K/Na discrimination in barley. J Exp Bot 41:1095–1101
- Gorham J (1994) Salt tolerance in the Triticeae: K/Na discrimination in some perennial wheatgrasses and their amphiploids with wheat. J Exp Bot 45:441–447
- Gorham J, Bridges J, Dubcovsky J, Dvoák J, Hollington PA, Luo MC, Khan JA (1997) Genetic analysis and physiology of a trait for enhanced $K + Na +$ discrimination in wheat. New Phytol 137:109–116
- Gorham J, Hardy C, WynJones RG, Joppa LR, Law CN (1987) Chromosomal location of a K/Na discrimination character in the D genome of wheat. Theor Appl Genet 74:584–588
- Goswami RS, Kistler HC (2004) Heading for disaster: Fusarium graminearum on cereal crops. Mol Plant Pathol 5(6):515–525
- Graham RD, Welch RM, Bouis HE (2001) Addressing micronutrient malnutrition through enhancing the nutritional quality of staple foods: principles, perspectives and knowledge gaps. Adv Agron 70:77–142
- Guo X, Wang Y, Meng L, Liu H, Yang L, Zhou Y, Zhang H (2015) Distribution of the Vrn-D1b allele associated with facultative growth habit in Chinese wheat accessions. Euphytica 206:1–10
- Guo X, Xin Z, Yang T, Ma X, Zhang Y, Wang Z, Ren Y, Lin T (2020) Metabolomics Response for Drought Stress Tolerance in Chinese Wheat Genotypes (Triticum aestivum). Plan Theory 9(4):520
- Gurcan K, Demirel F, Tekin M, Demirel S, Akar T (2017) Molecular and agro-morphological characterization of ancient wheat landraces of Turkey. BMC Plant Biol 17(1):171
- Gurung S, Mamidi S, Bonman JM, Jackson EW, DelRio LE, Acevedo M, Mergoum M, Adhikari TB (2011) Identifcation of novel genomic regions associated with resistance to Pyrenophora tritici-repentis races 1 and 5 in spring wheat landraces using association analysis. Theor Appl Genet 123(6):1029
- Haile JK, Hammer K, Badebo A, Singh RP, Röder MS (2013) Haplotype analysis of molecular markers linked to stem rust resistance genes in Ethiopian improved durum wheat varieties and tetraploid wheat landraces. Genetic resources and crop evolution 60(3):853–864
- Hammer K, Knüpffer H, Xhuveli L, Perrino P (1996) Estimating genetic erosion in landraces two case studies. Genet Resour Crop Evol 43:329–336
- Hanocq E, Niarquin M, Heumez E, Rousset M, Le Gouis J (2004) Detection and mapping of QTL for earliness components in a bread wheat recombinant inbred lines population. Theor Appl Genet 110:106–115
- Hao Y, Velu G, Peña RJ, Singh S, Singh RP (2014) Genetic loci associated with high grain zinc concentration and pleiotropic effect on kernel weight in wheat (Triticum aestivum L.). Mol Breed 34:1893–1902
- Hagenblad J, Asplund L, Balfourier F, Ravel C, Leino MW (2012) Strong presence of the high grain protein content allele of *NAM-B1* in Fennoscandian wheat. Theor Appl Genet 125:1677–1686
- Hamdi O, Bellil I, Branlard G, Khelii D (2010) Genetic variation and geographical diversity for seed storage proteins of seventeen durum wheat populations collected in Algeria. Not Bot Horti Agrobo 38(2):22–32
- Harlan JR (1975) Our vanishing genetic resources. Science 188(4188):618–621
- Hedden P (2003) The genes of the Green Revolution. Trends Genet 19:5–9
- Hede AR, Skovmand B, Reynolds MP, Crossa J, Vilhelmsen AL, Stølen O (1999) Evaluating genetic diversity for heat tolerance traits in Mexican wheat landraces. Genet Resour Crop Evol 46(1):37–45
- Heidari B, Padash S, Dadkhodaie A (2016) Variations in micronutrients, bread quality and agronomic traits of wheat landrace varieties and commercial cultivars. Aust J Crop Sci 10:377–384
- Henkrar F, El-Haddoury J, Iraqi D, Bendaou N, Udupa SM (2017) Allelic variation at highmolecular weight and low-molecular weight glutenin subunit genes in Moroccan bread wheat and durum wheat cultivars.3. Biotech 7:287
- Herrera-Foessel S, Singh R, Huerta-Espino J, Crossa J, Yuen J, Djurle A (2006) Effect of leaf rust on grain yield and yield traits of durum wheats with race-specifc and slow-rusting resistance to leaf rust. Plant Dis 90:1065–1072
- Hewitt PH, Van Niekerk, GJJ, Walters MC, Kriel CF, Fouche A (1984) Aspects of the ecology of the Russian wheat aphid, Diuraphis noxia, in the Bloemfontein district. I. The colonization and

infestation of sown wheat, identifcation of summer hosts and cause of infestation symptoms. Technical Communication, Department of Agriculture, South Africa, (191), p. 3–13

- Huang XQ, Hsam SLK, Zeller FJ (1997) Identifcation of powdery mildew resistance genes in common wheat (Triticum aestivum L. em Thell.). IX. Cultivars, land races and breeding lines grown in China. Plant Breed 116:233–238
- Huang XQ, Hsam SLK, Zeller FJ, Wenzel G, Mohler V (2000) Molecular mapping of the wheat powdery mildew resistance gene Pm24 and marker validation for molecular breeding. Theor Appl Genet 101(3):407–414
- Hussain B, Lucas SJ, Ozturk L, Budak H (2017) Mapping QTLs conferring salt tolerance and micronutrient concentrations at seedling stage in wheat. Sci Rep 7(1):1–14
- Hysing SC, Merker A, Liljeroth E, Koebner RM, Zeller FJ, Hsam SL (2007) Powdery mildew resistance in 155 Nordic bread wheat cultivars and landraces. Hereditas 144(3):102–119
- Iizumi T, Furuya J, Shen Z, Kim W, Okada M, Fujimori S, Hasegawa T, Nishimori M (2017) Responses of crop yield growth to global temperature and socioeconomic changes. Sci Rep 7(1):1–10
- Ilyas N, Amjid MW, Saleem MA, Khan W, Wattoo FM, Rana RM, Rana HM, Zahid A, Shah GA, Anwar A, Ahmad MQ, Shaheen M, Riaz H, Ansari MJ (2020) Quantitative trait loci (QTL) mapping for physiological and biochemical attributes in a Pasban90/Frontana recombinant inbred lines (RILs) population of wheat (*Triticum aestivum*) under salt stress condition. Saudi J Biol Sci 27(1):341–351
- Inoue Y, Vy TT, Yoshida K, Asano H, Mitsuoka C, Asuke S, Anh VL, Cumagun CJR, Chuma I, Terauchi R, Kato K, Mitchell T, Valent B, Farman M, Yukio TY (2017) Evolution of the wheat blast fungus through functional losses in a host specifcity determinant. Science 357(6346):80–83
- Iqbal M, Navabi A, Salmon DF, Yang RC, Spaner D (2007) Simultaneous selection for early maturity, increased grain yield and elevated grain protein content in spring wheat. Plant Breed 126:244–250
- Iwaki K, Haruna S, Niwa T, Kato K (2001) Adaptation and ecological differentiation in wheat with special reference to geographical variation of growth habit and Vrn genotype. Plant Breed 120:107–114
- Izanloo A, Condon AG, Langridge P, Tester M, Schnurbusch T (2008) Different mechanisms of adaptation to cyclic water stress in two South Australian bread wheat cultivars. J Exp Bot 59(12):3327–3346
- Jalal A, Shah S, Filho MCMT, Khan A, Shah T, Ilyas M, Rosa PAL (2020) Agro-Biofortifcation of Zinc and Iron in Wheat Grains. Gesunde Pfanzen 72:227–236
- Jaleel CA, Manivannan P, Wahid A, Farooq M, Somasundaram R, Panneerselvam R (2009) Drought stress in plants: a review on morphological characteristics and pigments composition. Int J Agric Biol 11:100–105
- Jan SU, Jamil M, Alipour H, Bhatti MF, Gul A (2017) Analysis of salinity tolerance potential in synthetic hexaploid wheat. Pak J Bot 49(4):1269–1278
- Janni M, Cadonici S, Bonas U, Grasso A, Dahab AAD, Visioli G, Pignone D, Ceriotti A, Marmiroli N (2018) Gene-ecology of durum wheat HMW glutenin refects their diffusion from the center of origin. Sci Rep 8:16929
- Jaradat A (2006) Phenotypic divergence in the meta-population of the Hourani wheat landrace. J Food Agric Env 4:186–191
- Jaradat AA (2011) Wheat landraces: genetic resources for sustenance and sustainability. usda-ars, pp 1–20. [http://www.usmarc.usda.gov/SP2UserFiles/Place/36450000/products-wheat/AAJ](http://www.usmarc.usda.gov/SP2UserFiles/Place/36450000/products-wheat/AAJ-wheatlandraces.pdf)[wheatlandraces.pdf](http://www.usmarc.usda.gov/SP2UserFiles/Place/36450000/products-wheat/AAJ-wheatlandraces.pdf)
- Jaradat AA (2013) Wheat landraces: a mini review. Emir J Food Agric 25:20–29
- Jiang Y, Huang L, Hu Y (2010) Distribution of vernalization genes in Chinese wheat landraces and their relationship with winter hardness. Sci Agric Sin 43:2619–2632
- Joppa LR, Cantrell RG (1990) Chromosomal location of genes for grain protein content of wild tetraploid wheat. Crop Sci 30:1059–1064
- Joppa LR, Du C, Hart GE, Hareland GA (1997) Mapping gene(s) for grain protein in tetraploid wheat (*Triticum turgidum* L.) using a population of recombinant inbred chromosome lines. Crop Sci 37:1586–1589
- Jordan KW, Wang S, Lun Y, Gardiner LJ, MacLachlan R, Hucl P, Wiebe K, Wong D, Forrest KL, Sharpe AG, Sidebottom CHD, Hall N, Toomajian C, Close T, Dubcovsky J, Akhunova A, Talbert L, Bansal UK, Bariana HS, Hayden MJ, Pozniak C, Jeddeloh JA, Anthony Hall A, Akhunov E (2015) A haplotype map of allohexaploid wheat reveals distinct patterns of selection on homoeologous genomes. Genome Biol 16(1):1–18
- Jorgensen C, Luo MC, Ramasamy R, Dawson M, Gill BS, Korol AB, Distelfeld A, Dvorak J (2017) A high-density genetic map of wild emmer wheat from the Karaca Dağ region provides new evidence on the structure and evolution of wheat chromosomes. Front Plant Sci 8:1798
- Kahrizi D, Cheghamirza K, Kakaei M, Mohammadi R, Ebadi A (2010) Heritability and genetic gain of some morphophysiological variables of durum wheat (Triticum turgidum var. durum). Afr J Biotechnol 9(30):4687–4691
- Kamal NM, Goraf YSA, Mega R, Tsujimoto H (2018) Physiological response of wheat to chemical desiccants used to simulate post-anthesis drought stress. Agronomy 8(4):44
- Kamran A, Iqbal M, Navabi A, Randhawa HS, Pozniak C, Spaner D (2013) Earliness per QTLs and their interaction with photoperiod insensitive allele Ppd-D1a in Cutler x AC Barrie spring wheat population. Theor Appl Genet 126:1965–1976
- Kandel JS, Krishnan V, Jiwan D, Chen X, Skinner DZ, See DR (2017) Mapping genes for resistance to stripe rust in spring wheat landrace PI 480035. PloS one 12(5):e0177898
- Kankwatsa P, Singh D, Thomson PC, Babiker EM, Bonman JM, Newcomb M, Park RF (2017) Characterization and genome-wide association mapping of resistance to leaf rust, stem rust and stripe rust in a geographically diverse collection of spring wheat landraces. Mol Breed 37(9):113
- Karagöz A, Zencirci N (2005) Variation in wheat (Triticum spp.) landraces from different altitudes of three regions of Turkey. Genet Resour Crop Ev 52(6):775–785
- Karan R, Subudhi PK (2012) Approaches to increasing salt tolerance in crop plants. In: Abiotic stress responses in plants. Springer, New York, pp 63–88
- Kato K, Mori Y, Beiles A, Nevo E (1997) Geographical variation in heading traits in wild emmer wheat, Triticum dicoccoides. I. Variation in vernalization response and ecological differentiation. Theor Appl Genet 95:546–552
- Kato K, Taketa S, Ban T, Iriki N, Miura K (2001) The infuence of a spring habit gene, Vrn-D1, on heading time in wheat. Plant Breed 120:115–120
- Kato K, Wada T (1999) Genetic analysis and selection experiment for narrow-sense earliness in wheat by using segregating hybrid progenies. Breed Sci 49:233–238
- Kato K, Yokoyama H (1992) Geographical variation in heading characters among wheat landraces, Triticum aestivum L., and its implication for their adaptability. Theor Appl Genet 84:259–265
- Kertho A, Mamidi S, Bonman JM, McClean PE, Acevedo M (2015) Genome-wide association mapping for resistance to leaf and stripe rust in winter-habit hexaploid wheat landraces. PLoS One 10(6):e0129580
- Khan MA, Fuller MP, Baloch FS (2008) Effect of soil applied zinc sulphate on wheat *(Triticum aestivum* L.) grown on a calcareous soil in Pakistan. Cereal Res Commun 36:571–582
- Khan MH, Bukhari A, Dar ZA, Rizvi SM (2013) Status and strategies in breeding for rust resistance in wheat. Agr Sci 4:292
- Khokhar JS, King J, King IP, Young SD, Foulkes MJ, De Silva J et al (2020) Novel sources of variation in grain Zinc (Zn) concentration in bread wheat germplasm derived from Watkins landraces. PLoS One 15(2):e0229107
- Kidane YG, Gesesse CA, Hailemariam BN, Desta EA, Mengistu DK, Fadda C, Pe ME, Dell'Acqua M (2019) A large nested association mapping population for breeding and quantitative trait locus mapping in Ethiopian durum wheat. Plant Biotechnol J 17(7):1380–1393
- Kidane YG, Hailemariam BN, Mengistu DK, Fadda C, Pè ME, Dell'Acqua M (2017) Genomewide association study of Septoria tritici blotch resistance in Ethiopian durum wheat landraces. Front Plant Sci 8:1586
- Kilian B, Ozkan H, Pozzi C, Salamini F (2009) Domestication of the Triticeae in the Fertile Crescent. In: Feuillet C, Muehlbauer GJ (eds) Genetics and genomics of the Triticeae. USA pp, Springer-Verlag, New York, pp 81–119
- King JE, Cook RJ, Melville SC (1983) A review of Septoria diseases of wheat and barley. Ann Appl Biol 103(2):345–373
- Kishii M (2019) An update of recent use of Aegilops species in wheat breeding. Front Plant Sci 10:585
- Klindworth DL, Hareland GA, Elias EM, Faris JD, Chao S, Xu SS (2009) Agronomic and quality characteristics of two new sets of Langdon durum-wild emmer wheat chromosome substitution lines. J Cereal Sci 50:29–35
- Kolev S, Vassilev D, Kostov K, Todorovska E (2011) Allele variation in loci for adaptive response in Bulgarian wheat cultivars and landraces and its effect on heading date. Plant Genet Res 9:251–255
- Kolmer JA, Garvin DF, Hayden M, Spielmeyer W (2018) Adult plant leaf rust resistance derived from the wheat landrace cultivar Americano 44d is conditioned by interaction of three QTL. Euphytica 214(3):59
- Korkut ZK, Balkan A, Başer İ, Bilgin O (2019) Grain Yield and Some Physiological Traits Associated with Heat Tolerance in Bread Wheat (Triticum aestivum L.) Genotypes. J Agric Sci 25(3):391–400
- Kumar J, Jaiswal V, Kumar A, Kumar N, Mir RR, Kumar S, Dhariwala R, Tyagia S, Khandelwale M, Prabhub KV, Prasade R, Balyana HS, Guptaa PK (2011) Introgression of a major gene for high grain protein content in some Indian bread wheat cultivars. Field Crop Res 123:226–233
- Kumar J, Schäfer P, Hückelhoven R, Langen G, Baltruschat H, Stein E, Nagarajan S, Kogel KH (2002) Bipolaris sorokiniana, a cereal pathogen of global concern: cytological and molecular approaches towards better control. Mol Plant Pathol 3(4):185–195
- Kumar S, Archak S, Tyagi RK, Kumar J, Vikas VK, Jacob SR, Srinivasan K, Radhamani J, Parimalan R, Sivaswamy M, Tyagi S, Yadav M, Kumari J, Deepali SS, Bhagat I, Meeta M, Bains NS, Chowdhury AK, Saha BC, Bhattacharya PM, Kumari J, Singh MC, Gangwar OP, Prasad P, Bharadwaj SC, Gogoi R, Sharma JB, Kumar GMS, Saharan MS, Bag M, Roy A, Prasad TV, Sharma RK, Dutta M, Sharma I, Bansal KC (2016) Evaluation of 19,460 wheat accessions conserved in the Indian national genebank to identify new sources of resistance to rust and spot blotch diseases. PLoS One 12:e0175610
- Kumar S, Beena AS, Awana M, Singh A (2017) Physiological, biochemical, epigenetic and molecular analyses of wheat (Triticum aestivum) genotypes with contrasting salt tolerance. Front Plant Sci 8:1151
- Law CN, Worland AJ, Giorgi B (1976) The genetic control of ear-emergence time by chromosomes 5A and 5D of wheat. Heredity 36:49–58
- Leonard K, Szabo L (2005) Stem rust of small grains and grasses caused by Puccinia graminis. Mol Plant Pathol 6:99–111
- Li G, Carver BF, Cowger C, Bai G, Xu X (2018a) Pm223899, a new recessive powdery mildew resistance gene identifed in Afghanistan landrace PI 223899. Theor Appl Genet 131(12):2775–2783
- Li G, Xu X, Bai G, Carver BF, Hunger R, Bonman JM (2016a) Identifcation of novel powdery mildew resistance sources in wheat. Crop Sci 56(4):1817–1830
- Li G, Xu X, Carver BF, Guo P, Puterka G (2018b) Dn10, a new gene conferring resistance to Russian wheat aphid biotype 2 in Iranian wheat landrace PI 682675. Crop Sci 58(3):1219–1225
- Li H, Vikram P, Singh RP, Kilian A, Carling J, Song J, Burgueno-Ferreira JA, Bhavani S, Huerta-Espino J, Payne T, Sehgal D, Wenzl P, Singh S (2015) A high density GBS map of bread wheat and its application for dissecting complex disease resistance traits. BMC Genomics 16(1):1–15.
- Li T, Bai G, Wu S, Gu S (2011) Quantitative trait loci for resistance to fusarium head blight in a Chinese wheat landrace Haiyanzhong. Theor Appl Genet 122(8):1497–1502
- Li T, Zhang D, Zhou X, Bai G, Li L, Gu S (2016b) Fusarium head blight resistance loci in a stratifed population of wheat landraces and varieties. Euphytica 207(3):551–561
- Li XJ, Xu X, Yang XM, Li XQ, Liu WH, Gao AN, Li LH (2012) Genetic diversity of the wheat landrace Youzimai from different geographic regions investigated with morphological traits, seedling resistance to powdery mildew, gliadin and microsatellite markers. Cereal Res Commun 40(1):95–106
- Li X, Li Y, Zhang M, Yu X, Hu R, Chang J, Yang G, Wang Y, He G (2019) Diversity of *Puroindoline* genes and their association with kernel hardness in Chinese wheat cultivars and landraces. Mol Breed 39(4):1–13
- Li Y, Huang C, Sui X, Fan Q, Li G, Chu X (2009) Genetic variation of wheat glutenin subunits between landraces and varieties and their contributions to wheat quality improvement in China. Euphytica 169:159–168
- Li Y, Shi X, Hu J, Wu P, Qiu D, Qu Y, Xie J, Wu Q, Zhang H, Yang L, Liu H, Zhou Y, Liu Z, Li H (2020) Identifcation of a recessive gene PmQ conferring resistance to powdery mildew in wheat landrace Qingxinmai using BSR-Seq analysis. Plant Disease 104(3):743–751
- Liu D, Liu Y, Zhang W, Chen X, Zou C (2017a) Agronomic approach of zinc biofortifcation can increase zinc bioavailability in wheat four and thereby reduce zinc defciency in humans. Nutrients 9(5):465
- Liu D, Zhang L, Hao M, Ning S, Yuan Z, Dai S, Huang L, Wu B, Yan Z, Lan X, Zheng Y (2018) Wheat breeding in the hometown of Chinese Spring. Crop J 6:82–90
- Liu J, Huang L, Wang C, Liu Y, Yan Z, Wang Z, Xiang L, Zhong X, Gong F, Zheng Y, Liu D, Wu B (2019) Genome-Wide Association Study Reveals Novel Genomic Regions Associated With High Grain Protein Content in Wheat Lines Derived From Wild Emmer Wheat. Front Plant Sci 10:464
- Liu W, Maccaferri M, Rynearson S, Letta T, Zegeye H, Tuberosa R, Chen X, Pumphrey M (2017b) Novel sources of stripe rust resistance identifed by genome-wide association mapping in Ethiopian durum wheat (Triticum turgidum ssp. durum). Front Plant Sci 8:774.1
- Liu Y, Lin Y, Gao S, Li Z, Deng M, Chen G, Wei Y, Zheng Y (2017c) A genome-wide association study of 23 agronomic traits in Chinese wheat landraces. Plant J 91:861–873
- Liu Y, Qie Y, Li X, Wang M, Chen X (2020) Genome-Wide Mapping of Quantitative Trait Loci Conferring All-Stage and High-Temperature Adult-Plant Resistance to Stripe Rust in Spring Wheat Landrace PI 181410. Int J Mol Sci 21(2):478
- Lodhi SS, Maryam S, Rafque K, Shafque A, Yousaf ZA, Talha AM, Gul A, Amir R (2020) Overview of the prospective strategies for conservation of genomic diversity in wheat landraces. In: Climate change and food security with emphasis on wheat. Academic Press, London, pp 293–309
- Long L, Yao F, Yu C, Ye X, Cheng Y, Wang Y, Wu Y, Li J, Wang J, Jiang Q, Li W, Ma J, Liu Y, Deng M, Wei Y, Zheng Y, Chen G (2019) Genome-Wide association study for adult-plant resistance to stripe rust in Chinese wheat landraces (Triticum aestivum L.) from the yellow and huai river valleys. Front Plant Sci 10:596
- Lopes MS, El-Basyoni I, Baenziger PS, Singh S, Royo C, Ozbek K, Aktas H, Ozer E, Ozdemir F, Manickavelu A, Ban T, Vikram P (2015) Exploiting genetic diversity from landraces in wheat breeding for adaptation to climate change. J Exp Bot 66:3477–3486
- Lyons G, Ortiz-Monasterio I, Stangoulis J, Graham R (2005) Selenium concentration in wheat grain: Is there suffcient genotypic variation to use in breeding? Plant and Soil 269:369–380
- Ma D, Li Q, Tang M, Chao K, Li J, Wang B, Jing J (2015) Mapping of gene conferring adult-plant resistance to stripe rust in Chinese wheat landrace Baidatou. Mol Breed 35(8):157
- Ma L, Zhou E, Huo N, Zhou R, Wang G, Jia J (2007) Genetic analysis of salt tolerance in a recombinant inbred population of wheat (*Triticum aestivum* L.). Euphytica 153(1-2):109–117
- Mahrookashani A, Siebert S, Hüging H, Ewert F (2017) Independent and combined effects of high temperature and drought stress around anthesis on wheat. J Agron Crop Sci 203(6):453–463
- Manickavelu A, Joukhadar R, Jighly A, Lan C, Huerta-Espino J, Stanikzai AS, Kilian A, Singh RP, Ban T (2016) Genome wide association mapping of stripe rust resistance in Afghan wheat landraces. Plant Sci 252:222–229
- Manickavelu A, Niwa S, Ayumi K, Komatsu K, Naruoka Y, Ban T (2014) Molecular evaluation of Afghan wheat landraces. Plant Genet Resour-C 12:S31–S35
- Mantri N, Patade V, Penna S, Ford R, Pang E (2012) In: Ahmad P, Prasad MNV (eds) Abiotic stress responses in plants: metabolism, productivity and sustainability. Springer Science+Business Media, LLC, New York
- Maraite H, Di Zinno T, Longree H, Daumerie V, Duveiller E (1997) Fungi associated with foliar blight of wheat in warm areas. In Proceedings of the international workshop on helminthosporium diseases of wheat: Spot Blotch and Tan Spot, El Batán (pp. 293–300)
- Masood MS, Javaid A, Rabbani MA, Anwar R (2005) Phenotypic diversity and trait association in bread wheat (Triticum aestivum L.) landraces from Baluchistan, Pakistan. Pak J Bot 37:949
- McIntosh RA, Wellings CR, Park RF (1995) Wheat rusts: an atlas of resistance genes. CSIRO Publications, East Melbourne (Australia)
- McIntosh RA, Yamazaki Y, Dubcovsky J, Rogers J, Morris C, Appels R, Xia XC (2013) Catalogue of gene symbols for wheat.<https://shigen.nig.ac.jp/wheat/komugi/genes/download.jsp>
- McKay JK, Richards JH, Mitchell-Olds T (2003) Genetics of drought adaptation in Arabidopsis thaliana: I. Pleiotropy contributes to genetic correlations among ecological traits. Mol Ecol 12:1137–1151
- Melnikova NV, Ganeva GD, Popova ZG, Landjeva SP, Kudryavtsev AM (2010) Gliadins of Bulgarian durum wheat *Triticum durum* Desf. landraces: genetic diversity and geographical distribution. Genet Resour Crop Evol 57:587–595
- Mengistu N, Baenziger PS, Eskridge KM, Dweikat I, Wegulo SN, Gill KS, Mujeeb-Kazi A (2012) Validation of QTL for grain yield-related traits on wheat chromosome 3a using recombinant inbred chromosome lines. Crop Sci 52:1622–1632
- Merchuk-Ovnat L, BarakV FT, Ordon F, Lidzbarsky GA, Krugman T, Saranga Y (2016) Ancestral QTL alleles from wild emmer wheat improve drought resistance and productivity in modern wheat cultivars. Front Plant Sci 7:452
- Mir RA, Sharma A, Mahajan R (2020) Crop landraces: present threats and opportunities for conservation. In: Rediscovery of genetic and genomic resources for future food security 2020. Springer, Singapore, pp 335–349
- Mir Ali N, Arabi MIE, Al-Safadi B (1999) Frequencies of high and low molecular weight glutenin subunits in durum wheat grown in Syria. Cereal Res Commun 27:301–305
- Mishra VK, Gupta PK, Arun B, Vasistha NK, Vishwakarma MK, SinghYadav P, Kumar H, Joshiac AK (2015) Introgression of a gene for high grain protein content (Gpc-B1) into two leading cultivars of wheat in Eastern Gangetic Plains of India through marker assisted backcross breeding. J Plant Breed Crop Sci 7:292–300
- Mitrofanova OP, Khakimova AG (2017) New genetic resources in wheat breeding for increased grain protein content. Russ J Genet Appl Res 7(4):477–487
- Mohammadi R, Armion M, Kahrizi D, Amri A (2012) Effciency of screening techniques for evaluating durum wheat genotypes under mild drought conditions. Int J Plant Prod 4(1):11–24
- Mohammadi R, Sadeghzadeh B, Ahmadi H, Bahrami N, Amri A (2015) Field evaluation of durum wheat landraces for prevailing abiotic and biotic stresses in highland rainfed regions of Iran. The Crop Journal 3(5):423–433

Monasterio I, Graham RD (2000) Breeding for trace minerals in wheat. Food Nutr Bull 21:392–396

- Mondal S, Rutkoski JE, Velu G, Singh PK, Crespo-Herrera LA, Guzman C, Bhavani S, Lan C, He X, Singh RP (2016) Harnessing diversity in wheat to enhance grain yield, climate resilience, disease and insect pest resistance and nutrition through conventional and modern breeding approaches. Front Plant Sci 7:991
- Mora F, Castillo D, Lado B, Matus I, Poland J, Belzile F, Zitzewitz JV, del Pozo A (2015) Genomewide association mapping of agronomic traits and carbon isotope discrimination in a worldwide germplasm collection of spring wheat using SNP markers. Mol Breed 35(2):69
- Moragues M, Zarco-Hernández J, Moralejo MA, Royo C (2006) Genetic diversity of glutenin protein subunits composition in durum wheat landraces [*Triticum turgidum* ssp. turgidum convar. durum (Desf.) MacKey] from the Mediterranean basin. Genet Resour Crop Evol 53(5):993–1002
- Motzo R, Giunta F (2007) The effect of breeding on the phenology of Italian durum wheats: From landraces to modern cultivars. Eur J Agron 26:462–470
- Mohammed AR, Tarpley L (2009) High nighttime temperatures affect rice productivity through altered pollen germination and spikelet fertility. Agric For Meteorol 149(6–7):999–1008
- Mujeeb-Kazi A, Gul A, Ahmad I, Farooq M, Rauf Y, Riaz H (2009) Genetic resources for some wheat abiotic stress tolerances. In: Salinity and water stress. Springer, Dordrecht, pp 149–163
- Muqaddasi QH, Reif JC, Li Z, Basnet BR, Dreisigacker S, Röder MS (2017) Genome-wide association mapping and genome-wide prediction of anther extrusion in CIMMYT spring wheat. Euphytica 213(3):73
- Mwadzingeni L, Shimelis H, Tesfay S, Tsilo TJ (2016) Screening of bread wheat genotypes for drought tolerance using phenotypic and proline analyses. Front Plant Sci 7:1276
- Nadeem MA, Nawaz MA, Shahid MQ, Doğan Y, Comertpay G, Yıldız M, Hatipoğlu R, Ahmad F, Alsaleh A, Labhane N, Özkan H (2018) DNA molecular markers in plant breeding: current status and recent advancements in genomic selection and genome editing. Biotechnol Biotechnol Equip 32:261–285
- Naghavi MR, Monfared SR, Ahkami AH, Ombidbakhsh MA (2009) Genetic variation of durum wheat landraces and cultivars using morphological and protein markers. Proceedings of world academy of science. Eng Technol 37:73–75
- Nazco R, Villegas D, Ammar K, Pena RJ, Moragues M, Royo C (2012) Can Mediterranean durum wheat landraces contribute to improved grain quality attributes in modern cultivars? Euphytica 185(1):1–17
- Newcomb M, Acevedo M, Bockelman HE, Brown-Guedira G, Goates BJ, Jackson EW, Jin Y, Njau P, Rouse MN, Singh RD, Wanyera R, Bonman JM (2013) Field resistance to the Ug99 race group of the stem rust pathogen in spring wheat landraces. Plant disease 97(7):882–890
- Newton AC, Akar T, Baresel JP, Bebeli PJ, Bettencourt E, Bladenopoulos KV, Czembor JH, Fasoula DA, Katsiotis A, Koutis K, Koutsika-Sotiriou M, Kovacs G, Larsson H, Pinheiro de Carvalho MAA, Rubiales D, Russell J, Dos Santos TMM, Vaz Patto MC (2010) Cereal landraces for sustainable agriculture. Sustain Agric 2:147–186
- Newton AC, Johnson SN, Gregory PJ (2011) Implications of climate change for diseases, crop yields and food security. Euphytica 179:3–18
- Nicol J, Rivoal R, Taylor S, Zaharieva M (2003) Global importance of cyst (Heterodera spp.) and lesion nematodes (Pratylenchus spp.) on cereals: distribution, yield loss, use of host resistance and integration of molecular tools. Nematol Monogr Perspect 2:1–19
- Nicol JM, Rivoal R, Bolat N, Aktas H, Braun HJ, Mergoum M, Yildrim AF, Bagci A, Eleckcioglu IH, Yahyaoui A (2002) The frequency and diversity of the cyst and lesion nematode on wheat in the Turkish Central Anatolian Plateau. Nematology 4(2):272
- Norman A, Taylor J, Tanaka E, Telfer P, Edwards J, Martinant JP, Kuchel H (2017) Increased genomic prediction accuracy in wheat breeding using a large Australian panel. Theor Appl Genet 130(12):2543–2555
- O'Driscoll A, Kildea S, Doohan F, Spink J, Mullins E (2014) The wheat–Septoria confict: a new front opening up? Trends Plant Sci 19(9):602–610
- Okechukwu EC, Agbo CU, Uguru MI, Ogbonnaya FC (2016) Germplasm evaluation of heat tolerance in bread wheat in Tel Hadya, Syria. Chil J Agric Res 76(1):9–17
- Olmstead AL, Rhode PW (2002) The red queen and the hard reds: productivity grown in American wheat 1800–1940. J Econ Hist 62:929–966
- Ouaja M, Aouini L, Bahri B, Ferjaoui S, Medini M, Marcel TC, Hamza S (2020) Identifcation of valuable sources of resistance to Zymoseptoria tritici in the Tunisian durum wheat landraces. Eur J Plant Pathol 156(2):647–661
- Oak MD, Tamhankar SA (2017) 1BL/1RS translocation in durum wheat and its effect on end use quality traits. J Plant Biochem Biot 26(1):91–96
- Ozberk I, Atay S, Altay F, Cabi E, Ozkan H, Atli A (2016) The Wheat Atlas of Turkey. (World Wildlife Fund), Istanbul (in Turkish)
- Ozer G, Paulitz TC, Imren M, Alkan M, Muminjanov H, Dababat AA (2020) Identity and Pathogenicity of Fungi Associated with Crown and Root Rot of Dryland Winter Wheat in Azerbaijan. Plant Disease 104:2149–2157
- Özkan H, Brandolini A, Schäfer-Pregl R, Salamini F (2002) AFLP analysis of a collection of tetraploid wheats indicates the origin of emmer and hard wheat domestication in southeast Turkey. Mol Biol Evol 19(10):1797–1801
- Özkan H, Willcox G, Graner A, Salamini F, Kilian B (2011) Geographic distribution and domestication of wild emmer wheat (Triticum dicoccoides). Genet Resour Crop Evol 58(1):11–53
- Pasam RK, Bansal U, Daetwyler HD, Forrest KL, Wong D, Petkowski J, Willey N, Randhawa M, Chhetri M, Miah H, Tibbits J, Bariana H, Hayden MJ (2017) Detection and validation of genomic regions associated with resistance to rust diseases in a worldwide hexaploid wheat landrace collection using BayesR and mixed linear model approaches. Theor Appl Genet 130(4):777–793
- Peng JH, Sun D, Nevo E (2011a) Domestication evolution, genetics and genomics in wheat. Mol Breed 28:281–301
- Peng ZS, Li X, Yang ZJ, Liao ML (2011b) A new reduced height gene found in the tetraploid semidwarf wheat landrace Aiganfanmai. Genet Mol Res 10:2349–2357
- Peusha H, Lebedeva T, Prilinn O, Enno T (2002) Genetic analysis of durable powdery mildew resistance in a common wheat line. Hereditas 136:201–206
- Pinto RS, Molero G, Reynolds MP (2017) Identifcation of heat tolerant wheat lines showing genetic variation in leaf respiration and other physiological traits. Euphytica 213(3):76
- Poersch-Bortolon LB, Pereira JF, Nhani Junior A, Gonzáles HHS, Torres GAM, Consoli L, Arenhart RA, Bodanese-Zanettini MH, Margis-Pinheiro M (2016) Gene expression analysis reveals important pathways for drought response in leaves and roots of a wheat cultivar adapted to rainfed cropping in the Cerrado biome. Genet Mol Biol 39(4):629–645
- Popov C, Hondru N, Bărbulescu A, Vonica I, Mărgărit G (1988) Species of aphids attacking wheat and barley crops. Analele Institutului de Cercetări pentru Cereale și Plante Tehnice, Fundulea 56:379–384
- Pu Z, Pei Y, Yang J, Ma J, Li W, Liu D, Wang J, Wei Y, Zheng Y (2018) A QTL located on chromosome 3D enhances the selenium concentration of wheat grain by improving phytoavailability and root structure. Plant and Soil 425(1-2):287–296
- Pugsley AT (1971) A genetic analysis of the spring-winter habit of growth in wheat. Aust J Agr Res 22:21–23
- Pugsley AT (1972) Additional genes inhibiting winter habit in wheat. Euphytica 21:547–552
- Qaseem MF, Qureshi R, Muqaddasi QH, Shaheen H, Kousar R, Röder MS (2018) Genome-wide association mapping in bread wheat subjected to independent and combined high temperature and drought stress. PLoS One 13:6
- Qiang LI, Wang ZR, Ding LI, Wei JW, Qiao WC, Meng XH, Sun SI, Li HM, Zhao MH, Chen XM, Zhao FW (2018) Evaluation of a new method for quantifcation of heat tolerance in different wheat cultivars. J Integr Agric 17(4):786–795
- Qie Y, Sheng Y, Xu H, Jin Y, Ma F, Li L, Li X, An D (2019) Identifcation of a new powdery mildew resistance gene pmDHT at or closely linked to the Pm5 locus in the Chinese wheat landrace Dahongtou. Plant disease 103(10):2645–2651
- Quarrie SA, Steed A, Calestani C, Semikhodskii A, Lebreton C, Chinoy C, Steele N, Pljevljakusić D, Waterman E, Weyen J, Schondelmaier J, Habash DZ, Farmer P, Saker L, Clarkson DT, Abugalieva A, Yessimbekova M, Turuspekov Y, Abugalieva S, Tuberosa R, Sanguineti MC, Hollington PA, Aragués R, Royo A, Dodig D (2005) A high-density genetic map of hexaploid wheat (*Triticum aestivum* L.) from the cross Chinese Spring× SQ1 and its use to compare QTLs for grain yield across a range of environments. Theor Appl Genet 110(5):865–880
- Quisenberry KS, Reitz LP (1974) Turkey wheat: The cornerstone of an empire. Agric Hist 48:98–110
- Qureshi N, Bariana H, Kolmer JA, Miah H, Bansal U (2017) Genetic and molecular characterization of leaf rust resistance in two durum wheat landraces. Phytopathology 107(11):1381–1387
- Qureshi N, Bariana H, Kumran VV, Muruga S, Forrest KL, Hayden MJ, Bansal U (2018) A new leaf rust resistance gene Lr79 mapped in chromosome 3BL from the durum wheat landrace Aus26582. Theor Appl Genet 131(5):1091–1098
- Rahmatov M, Otambekova M, Muminjanov H, Rouse MN, Hovmøller MS, Nazari K, Steffenson BJ, Johansson E (2019) Characterization of stem, stripe and leaf rust resistance in Tajik bread wheat accessions. Euphytica 215(3):1–22
- Ram S, Govindan V (2020) Improving wheat nutritional quality through biofortifcation. In: Igrejas G, Ikeda TM, Guzman C (eds) Wheat quality for improving processing and human health. Springer, Switzerland, pp 205–224
- Rasheed A, Xia X, Mahmood T, Quraishi UM, Bux AAH, Mahmood Z, Mirza JI, Mujeeb-Kazi A, He Z (2016) Comparison of economically important loci in landraces and improved wheat cultivars from Pakistan. Crop Sci 56:287–301
- Rawson HM, Richards RA, Munns R (1988) An examination of selection criteria for salt tolerance in wheat, barley and triticale genotypes. Aust J Agr Res 39:759–772
- Reitz LP, Salmon SC (1968) Origin, history, and use of Norin 10 wheat. Crop Sci 8:686–689
- Ren Y, Xu Y, Teng W, Li B, Lin T (2018) QTLs for seedling traits under salinity stress in hexaploid wheat. Cienc Rural 48(3):e20170446
- Reynolds M, Dreccer F, Trethowan R (2007) Drought-adaptive traits derived from wheat wild relatives and landraces. J Exp Bot 58(2):177–186
- Reynolds MP, Borlaug NE (2006a) Applying innovations and new technologies for international collaborative wheat improvement. J Agr Sci-Cambridge 144:95
- Reynolds MP, Borlaug NE (2006b) Impacts of breeding on international collaborative wheat improvement. J Agr Sci-Cambridge 144:3–17
- Riaz A, Athiyannan N, Periyannan S, Afanasenko O, Mitrofanova O, Aitken EA, Lagudah E, Hickey LT (2017) Mining Vavilov's treasure chest of wheat diversity for adult plant resistance to Puccinia triticina. Plant disease 101(2):317–323
- Ribaut JM, Hoisington D (1998) Marker-assisted selection: new tools and strategies. Trends Plant Sci 3(6):236–239
- Ribeiro M, Carvalho C, Carnide V, Guedes-Pinto H, Igrejas G (2011) Towards allelic diversity in the storage proteins of old and currently growing tetraploid and hexaploid wheats in Portugal. Genet Resour Crop Evol 58:1051–1073
- Roelfs A, Huerto-Espino J, Marshall D (1992) Rust diseases of wheat: concepts and methods of disease management. CIMMYT, Mexico
- Rola EL, De Vallavieille-Pope C, Leconte M, Nazari K (2019) Diversity of genes for resistance to stripe rust in wheat elite lines, commercial varieties and landraces from Lebanon and Syria. Phytopathologia Mediterranea 58(3):607–627
- Roselló M, Royo C, Álvaro F, Villegas D, Nazco R, Soriano JM (2018) Pasta-making quality QTLome from Mediterranean durum wheat landraces. Front Plant Sci 9:1512
- Roshanzamir H, Kordenaeej A, Bostani A (2013) Mapping QTLs related to Zn and Fe concentrations in bread wheat (Triticum aestivum) grain using microsatellite markers. Iran J Genet Plant Breed 2:10–17
- Royo C, Briceño-Félix GA (2011a) Spanish wheat pool. In: Bojean AP, Angus WJ, van Ginkel M (eds) The world wheat book. A history of wheat breeding, vol 2. Lavoisier, Paris, pp 121–154
- Royo C, Dreisigacker S, Soriano JM, Lopes MS, Ammar K, Villegas D (2020) Allelic variation at the vernalization response (Vrn-1) and photoperiod sensitivity (Ppd-1) genes and their association with the development of durum wheat landraces and modern cultivars. Front Plant Sci 11:838
- Royo C, Briceño-Félix GA (2011b) Spanish wheat pool. In: Bojean AP, Angus WJ, van Ginkel M (eds) The world wheat book. A history of wheat breeding. Lavoisier Publishing, Paris, pp 121–154
- Ruiz M, Giraldo P, Royo C, Villegas D, Jose Aranzana M, Carrillo JM (2012) Diversity and genetic structure of a collection of spanish durum wheat landraces. Crop Sci 52(5):2262–2275
- Saini DK, Devi P, Kaushik P (2020) Advances in genomic interventions for wheat biofortifcation: a review. Agronomy 10(1):62
- Salvi S, Porfri O, Ceccarelli S (2013) Nazareno Strampelli, the 'Prophet' of the green revolution. J Agr Sci-Cambridge 151:1–5
- Sansaloni C, Franco J, Santos B, Percival-Alwyn L, Singh S, Petroli C, Campos J, Dreher K, Payne T, Marshall D, Kilian B (2020) Diversity analysis of 80,000 wheat accessions reveals consequences and opportunities of selection footprints. Nat Commun 11:1–2
- Sareen S, Tyagi BS, Sarial AK, Tiwari V, Sharma I (2014) Trait analysis, diversity, and genotype x environment interaction in some wheat landraces evaluated under drought and heat stress conditions. Chil J Agric Res 74(2):135–142
- Schmidt AL, McIntyre CL, Thompson J, Seymour NP, Liu CJ (2005) Quantitative trait loci for root lesion nematode (Pratylenchus thornei) resistance in Middle-Eastern landraces and their potential for introgression into Australian bread wheat. Aust J Agr Res 56(10):1059–1068
- Schmidt J, Tricker PJ, Eckermann P, Kalambettu P, Garcia M, Fleury DL (2020) Novel alleles for combined drought and heat stress tolerance in wheat. Front Plant Sci 10:1800
- Scott PR, Hollins TW (1974) Effects of eyespot on the yield of winter wheat. Ann Appl Biol 78(3):269–279
- Shahzad A, Ahmad M, Iqbal M, Ahmed I, Ali GM (2012) Evaluation of wheat landrace genotypes for salinity tolerance at vegetative stage by using morphological and molecular markers. Genet Mol Res 11(1):679–692
- Shamaya NJ, Shavrukov Y, Langridge P, Roy SJ, Tester M (2017) Genetics of Na+ exclusion and salinity tolerance in Afghani durum wheat landraces. BMC Plant Biol 17(1):209
- Sharma DK, Torp AM, Rosenqvist E, Ottosen CO, Andersen SB (2017) QTLs and potential candidate genes for heat stress tolerance identifed from the mapping populations specifcally segregating for Fv/Fm in wheat. Front Plant Sci 8:1668
- Shavrukov Y, Langridge P, Tester M (2009) Salinity tolerance and sodium exclusion in genus Triticum. Breed Sci 59(5):671–678
- Shi R, Li H, Tong Y, Jing R, Zhang F, Zou C (2008) Identifcation of quantitative trait locus of zinc and phosphorus density in wheat (Triticum aestivum L.) grain. Plant and Soil 306:95–104
- Singh R, Govindan V, Andersson MS (2017) Zinc-Biofortifed Wheat: Harnessing Genetic Diversity for Improved Nutritional Quality. Sci Br Biofortif Ser 1:1–4
- Smale M (1996) Understanding global trends in the use of wheat diversity and international fows of wheat genetic resources. CIMMYT, Mexico
- Smale, M. and McBride, T., 1996. Understanding global trends in the use of wheat diversity and international fows of wheat genetic resources: part 1. CIMMYT 1995/96 World Wheat Facts and Trends: Understanding Global Trends in the Use of Wheat Diversity and International Flows of Wheat Genetic Resources (No. Look under series title. CIMMYT.). Centro Internacional de Mejoramiento de Maiz y Trigo (CIMMYT), Mexico
- Sofalian O, Chaparzadeh N, Javanmard A, Hejazi MS (2008) Study the genetic diversity of wheat landraces from northwest of Iran based on ISSR molecular markers. Int J Agric Biol 10:466–468
- Soriano JM, Villegas D, Aranzana MJ, García del Moral LF, Royo C (2016) Genetic structure of modern durum wheat cultivars and Mediterranean landraces matches with their agronomic performance. PLoS One 11:e0160983
- Soriano JM, Villegas D, Sorrells MR, Royo C (2018) Durum wheat landraces from east and west regions of the Mediterranean basin are genetically distinct for yield components and phenology. Front Plant Sci 9:80
- Stelmakh AF (1990) Geographic distribution of Vrn-genes in landraces and improved varieties of spring bread wheat. Euphytica 45:113–118
- Sthapit J, Newcomb M, Bonman JM, Chen X, See DR (2014) Genetic diversity for stripe rust resistance in wheat landraces and identifcation of accessions with resistance to stem rust and stripe rust. Crop Sci 54(5):2131–2139
- Sun H, Hu J, Song W, Qiu D, Cui L, Wu P, Zhang H, Liu H, Li L, Qu Y, Li Y, Li T, Cheng W, Zhou Y, Liu Z, Li J, Li H (2018) Pm61: A recessive gene for resistance to powdery mildew in wheat landrace Xuxusanyuehuang identifed by comparative genomics analysis. Theor Appl Genet 131(10):2085–2097
- Sun QM, Zhou RH, Gao LF, Zhao GY, Jia JZ (2009) The characterization and geographical distribution of the genes responsible for vernalization requirement in Chinese bread wheat. J Integr Plant Biol 51:423–432
- Tadesse W, Amri A, Ogbonnaya FC, Sanchez-Garcia M, Sohail Q, Baum M (2016) Wheat. In: Mohar S, Upadhyaya H (eds) Genetic and genomic resources for grain cereals improvement. Academic Press, Oxford, UK, pp 81–124
- Taiz L, Zeiger E (2006) Plant Physiology, 4th edn. Sinauer Associates Inc Publishers, Sunderland
- Talas F, Longin F, Miedaner T (2011) Sources of resistance to Fusarium head blight within Syrian durum wheat landraces. Plant breeding 130(3):398–400
- Tan C, Li G, Cowger C, Carver BF, Xu X (2018) Characterization of Pm59, a novel powdery mildew resistance gene in Afghanistan wheat landrace PI 181356. Theor Appl Genet 131(5):1145–1152
- Tan C, Li G, Cowger C, Carver BF, Xu X (2019) Characterization of Pm63, a powdery mildew resistance gene in Iranian landrace PI 628024. Theor Appl Genet 132(4):1137–1144
- Tehseen MM, Tonk FA, Tosun M, Amri A, Sansaloni CP, Kurtulus E, Yazbek M, Al-Sham'aa K, Ozseven I, Safdar LB, Shehadeh A, Nazari K (2020) Genome Wide Association Study of Resistance to PstS2 and Warrior Races of Stripe (Yellow) Rust in Bread Wheat Landraces. bioRxiv
- Thakur P, Kumar S, Malik JA, Berger JD, Nayyar H (2010) Cold stress effects on reproductive development in grain crops: an overview. Environ Exp Bot 67(3):429–443
- Thirumalaikumar VP, Devkar V, Mehterov N, Ali S, Ozgur R, Turkan I, Mueller-Roeber B, Balazadeh S (2018) NAC transcription factor JUNGBRUNNEN 1 enhances drought tolerance in tomato. Plant Biotechnol J 16(2):354–366
- Thompson AL, Smiley RW, Paulitz TC, Garland-Campbell K (2016) Identifcation of resistance to Pratylenchus neglectus and Pratylenchus thornei in Iranian landrace accessions of wheat. Crop Sci 56(2):654–672
- Thompson JP, O'reilly MM, Clewett TG (2009) Resistance to the root-lesion nematode Pratylenchus thornei in wheat landraces and cultivars from the West Asia and North Africa (WANA) region. Crop Pasture Sci 60(12):1209–1217
- Thompson JP, Seymour NP (2011) Inheritance of resistance to root-lesion nematode (Pratylenchus thornei) in wheat landraces and cultivars from the West Asia and North Africa (WANA) region. Crop Pasture Sci 62(1):82–93
- Toor AK, Bansal UK, Bhardwaj S, Badebo A, Bariana HS (2013) Characterization of stem rust resistance in old tetraploid wheat landraces from the Watkins collection. Genet Resour Crop Evol 60(7):2081–2089
- Toreti A, Cronie O, Zampieri M (2019) Concurrent climate extremes in the key wheat producing regions of the world. Sci Rep 9:5493
- Touzy G, Rincent R, Bogard M, Lafarge S, Dubreuil P, Mini A, Deswarte JC, Beauchene K, Le Gouis J, Praud S (2019) Using environmental clustering to identify specifc drought tolerance QTLs in bread wheat (T. aestivum L.). Theor Appl Genet 132(10):2859–2880
- Turner AS, Faure S, Zhang Y, Laurie DA (2013) The effect of day-neutral mutations in barley and wheat on the interaction between photoperiod and vernalization. Theor Appl Genet 126:2267–2277
- Uauy C, Brevis JC, Dubcovsky J (2006a) The high grain protein content gene Gpc-B1 accelerates senescence and has pleiotropic effects on protein content in wheat. J Exp Bot 57:2785–2794
- Uauy C, Distelfeld A, Fahima T, Blechl A, Dubcovsky J (2006b) A NAC gene regulating senescence improves grain protein, zinc, and iron content in wheat. Science 314:1298–1301
- Ullah S, Bramley H, Daetwyler H, He S, Mahmood T, Thistlethwaite R, Trethowan R (2018) Genetic contribution of emmer wheat (Triticum dicoccon Schrank) to heat tolerance of bread wheat. Front Plant Sci 9:1529
- Valdez VA, Byrne PF, Lapitan NL, Peairs FB, Bernardo A, Bai G, Haley SD (2012) Inheritance and genetic mapping of Russian wheat aphid resistance in Iranian wheat landrace accession PI 626580. Crop Sci 52(2):676–682
- Valluru R, Reynolds MP, Davies WJ, Sukumaran S (2017) Phenotypic and genome-wide association analysis of spike ethylene in diverse wheat genotypes under heat stress. New Phytol 214(1):271–283
- Van Oosten MJ, Costa A, Punzo P, Landi S, Ruggiero A, Batelli G, Grillo S (2016) Genetics of drought stress tolerance in crop plants. In Drought Stress Tolerance in Plants, vol 2. Springer, Cham, pp 39–70
- Varella AC, Weaver DK, Blake NK, Hofand ML, Heo HY, Cook JP, Lamb PF, Jordan KW, Akhunov E, Chao S, Talbert LE (2019) Analysis of recombinant inbred line populations derived from wheat landraces to identify new genes for wheat stem sawfy resistance. Theor Appl Genet 132(8):2195–2207
- Varella AC, Weaver DK, Cook JP, Blake NK, Hofand ML, Lamb PF, Talbert LE (2017) Characterization of resistance to the wheat stem sawfy in spring wheat landrace accessions from targeted geographic regions of the world. Euphytica 213(7):153
- Vats P, Banerjee UC (2004) Production studies and catalytic properties of phytases (myo-inositol hexakisphosphate phosphohydrolases): an overview. Enzyme Microb Technol 35(1):3–14
- Veraverbeke WS, Delcour JA (2002) Wheat protein composition and properties of wheat glutenin in relation to breadmaking functionality. Crit Rev Food Sci Nutr 42:179–208
- Vikram P, Franco J, Burgueño-Ferreira J, Li H, Sehgal D, Saint Pierre C, Ortiz C, Sneller C, Tattaris M, Guzman C, Sansaloni CP (2016a) Unlocking the genetic diversity of Creole wheats. Sci Rep 6:23092
- Vikram P, Franco J, Burgueño-Ferreira J, Li H, Sehgal D, Saint Pierre C, Cynthia Ortiz C, Clay Sneller C, Maria Tattaris M, Carlos Guzman C, Carolina Paola Sansaloni CP, Ellis M, Fuentes-Davila G, Reynolds M, Sonder K, Singh P, Payne T, Wenzl P, Sharma A, Bains NS, Singh GP, Crossa J, Singh S (2016b) Unlocking the genetic diversity of Creole wheats. Sci Rep 6:23092
- Villa TC, Maxted N, Scholten M, Ford-Lloyd B (2005) Defning and identifying crop landraces. Plant Genet Res 3(3):373–384
- Vishwakarma MK, Arun B, Mishra VK, Yadav PS, Kumar H, Joshi AK (2016) Marker-assisted improvement of grain protein content and grain weight in Indian bread wheat. Euphytica 208:313–321
- Von Rünker K (1908) Die Systematischeeinteilung und Benen-ung der Getreidesortenfu¨rpr aktische Zwecke. Jahrbuch der Deutschenlandwirtschafts-Gesellschaft 23:137–167
- Wamalwa MN, Owuoche J, Ogendo J, Wanyera R (2019) Multi-Pathotype Testing of Selected Kenyan Wheat Germplasm and Watkin Landraces for Resistance to Wheat Stripe Rust (Puccinia striiformis f. sp tritici). Races Agronomy 9(11):770
- Wamalwa M, Tadesse Z, Muthui L, Yao N, Zegeye H, Randhawa M, Wanyera R, Uauy C, Shorinola O (2020) Allelic diversity study of functional genes in East Africa bread wheat highlights opportunities for genetic improvement. Mol Breedi 40(11):1–14
- Wang S, Asuke S, Vy TTP, Inoue Y, Chuma I, Win J, Kato K, Tosa Y (2018a) A new resistance gene in combination with Rmg8 confers strong resistance against Triticum isolates of Pyricularia oryzae in a common wheat landrace. Phytopathology 108(11):1299–1306
- Wang Y, Peng H, Liu G, Xie C, Ni Z, Yang T, Liu Z, Sun Q (2010) Identifcation and molecular mapping of a leaf rust resistance gene in spelt wheat landrace Altgold. Euphytica 174(3):371–375
- Wang Z, Huang L, Wu B, Hu J, Jiang Z, Qi P, Zheng Y, Liu D (2018b) Characterization of an integrated active Glu-1Ay allele in common wheat from wild emmer and its potential role in four improvement. Int J Mol Sci 19:E923
- Wang Z, Li H, Zhang D, Guo L, Chen J, Chen Y, Wu Q, Xie J, Zhang Y, Sun Q, Dvorak J, Luo M, Liu Z (2015) Genetic and physical mapping of powdery mildew resistance gene MlHLT in Chinese wheat landrace Hulutou. Theor Appl Genet 128(2):365–373
- Wang Z, Ren J, Du Z, Che M, Zhang Y, Quan W, Jiang X, Ma Y, Zhao Y, Zhang Z (2019) Identifcation of a major QTL on chromosome arm 2AL for reducing yellow rust severity from a Chinese wheat landrace with evidence for durable resistance. Theor Appl Genet 132(2):457–471
- Ward RW, Yang ZL, Kim HS, Yen C (1998) Comparative analyses of RFLP diversity in landraces of Triticum aestivum and collections of T. tauschii from China and Southwest Asia. Theor Appl Genet 96:312–318
- Wei YM, Hou YC, Yan ZH, Wu W, Zhang ZQ, Liu DC, Zheng YL (2005) Microsatellite DNA polymorphism divergence in Chinese wheat (Triticum aestivum L.) landraces highly resistant to Fusarium head blight. J Appl Genet 46(1):3–9
- White PJ, Broadley MR (2005) Biofortifying crops with essential mineral elements. Trends Plant Sci 10(12):586–593
- WHO (2009) Global health risks, mortality and burden of disease attributable to selected major risks. Geneva, Switzerland, WHO
- Witcombe JR, Joshi A, Joshi KD, Sthapit BR (1996) Farmer Participatory Crop Improvement. I. Varietal Selection and Breeding Methods and Their Impact on Biodiversity. Exp Agric 32(04):445–460
- Worland AJ, Korzun V, Röder MS, Ganal MW, Law CN (1998) Genetic analysis of the dwarfng gene Rht8 in wheat. Part II. The distribution and adaptive signifcance of allelic variants at the Rht8 locus of wheat as revealed by microsatellite screening. Theor Appl Genet 96:1110–1120
- Wu L, Xia X, Rosewarne GM, Zhu H, Li S, Zhang Z, He Z (2015) Stripe rust resistance gene Yr18 and its suppressor gene in Chinese wheat landraces. Plant Breed 134(6):634–640
- Wu XL, Wang JW, Cheng YK, Ye XL, Li W, Pu ZE, Jiang QT, Wei YM, Deng M, Zheng YL, Chen GY (2016) Inheritance and molecular mapping of an all-stage stripe rust resistance gene derived from the Chinese common wheat landrace "Yilongtuomai". J Hered 107(5):463–470
- Xiao J, Jia X, Wang H, Zhao R, Fang Y, Gao R, Wu Z, Cao A, Wang J, Xue Z, Zhao W, Kang J, Chen Q, Chen P, Wang X (2011) A fast-neutron induced chromosome fragment deletion of 3BS in wheat landrace Wangshuibai increased its susceptibility to Fusarium head blight. Chromosome Res 19(2):225–234
- Xiao J, Jin X, Jia X, Wang H, Cao A, Zhao W, Pei H, Xue Z, He L, Chen Q, Wang X (2013) Transcriptome-based discovery of pathways and genes related to resistance against Fusarium head blight in wheat landrace Wangshuibai. BMC Genomics 14(1):1–19
- Xu H, Yi Y, Ma P, Qie Y, Fu X, Xu Y, Zhang X, An D (2015) Molecular tagging of a new broadspectrum powdery mildew resistance allele Pm2c in Chinese wheat landrace Niaomai. Theor Appl Genet 128(10):2077–2084
- Xu X, Li Q, Ma Z, Fan J, Zhou Y (2018) Molecular mapping of powdery mildew resistance gene PmSGD in Chinese wheat landrace Shangeda using RNA-seq with bulk segregant analysis. Mol Breed 38(3):23
- Xu X, Liu W, Liu Z, Fan JR, Zhou Y (2020) Mapping powdery mildew resistance gene pmYBL on chromosome 7B of Chinese Wheat (Triticum aestivum L.) Landrace Youbailan. Plant Dis 104(9):2411–2417
- Xu Y, An D, Liu D, Zhang A, Xu H, Li B (2012) Molecular mapping of QTLs for grain zinc, iron and protein concentration of wheat across two environments. Field Crop Res 138:57–62
- Xue F, Wang C, Li C, Duan X, Zhou Y, Zhao N, Wang Y, Ji W (2012) Molecular mapping of a powdery mildew resistance gene in common wheat landrace Baihulu and its allelism with Pm24. Theor Appl Genet 125(7):1425–1432
- Xue F, Zhai WW, Duan XY, Zhou YL, Ji WQ (2009) Microsatellite mapping of powdery mildew resistance gene in wheat landrace Xiaobaidong. Acta Agron Sin 34:1193–1198
- Xynias IN, Kozub NA, Sozinov IA (2011) Analysis of hellenic durum wheat *Triticum turgidum* L. var. *durum* germplasm using gliadin and high-molecular-weight glutenin subunit loci. Cereal Res Commun 39:415–425
- Yan L, Loukoianov A, Tranquilli G, Helguera M, Fahima T, Dubcovsky J (2003) Positional cloning of the wheat vernalization gene VRN1. Proc Natl Acad Sci U S A 100:6263–6268
- Yeken MZ, Akpolat H, Karaköy T, Çiftçi V (2018) Assessment of Mineral Content Variations for Biofortifcation of the Bean Seed. Int J Agri Wild Sci 4(2):261–269
- Yen C, Luo MC, Yang JL (1988) The origin of the Tibetan weedrace of hexaploid wheat, Chinese Spring, Chengdu-guang-tou and other landraces of the white wheat complex from China. In: Miller TE, Koebner RMD (eds.) Proceedings of the 7th International Wheat Genetics Symposium, Cambridge, pp 175–179
- Yu S, Wu J, Wang M, Shi W, Xia G, Jia J, Kang Z, Han D (2020) Haplotype variations in QTL for salt tolerance in Chinese wheat accessions identifed by marker-based and pedigree-based kinship analyses. Crop J. <https://doi.org/10.1016/j.cj.2020.03.007>
- Yuan FP, Zeng QD, Wu JH, Wang QL, Yang ZJ, Liang BP, Kang ZS, Chen XH, Han DJ (2018) QTL mapping and validation of adult plant resistance to stripe rust in Chinese wheat landrace Humai 15. Front Plant Sci 9:968
- Zampieri M, Ceglar A, Dentener F, Toreti A (2017) Wheat yield loss attributable to heat waves, drought and water excess at the global, national and subnational scales. Environ Res Lett 12(6):064008
- Zeven AC (1998) Landraces: a review of defnitions and classifcations. Euphytica 104:127–139
- Zeven AC (1999) The traditional inexplicable replacement of seed and seed ware of landraces and cultivars: a review. Euphytica 110:181–191
- Zeven AC (2000) Traditional maintenance breeding of landraces: 1. Data by crop. Euphytica 116:65–85
- Zhang K, Wang J, Qin H, Wei Z, Hang L, Zhang P, Reynolds M, Wang D (2019a) Assessment of the individual and combined effects of Rht8 and Ppd-D1a on plant height, time to heading and yield traits in common wheat. The Crop Journal 7:845–856
- Zhang P, Gebrewahid TW, Zhou Y, Ll Q, LI Z, LIu D (2019b) Seedling and adult plant resistance to leaf rust in 46 Chinese bread wheat landraces and 39 wheat lines with known Lr genes. J Integr Agric 18(5):1014–1023
- Zhang X, Pan H, Bai G (2012) Quantitative trait loci responsible for Fusarium head blight resistance in Chinese landrace Baishanyuehuang. Theor Appl Genet 125(3):495–502
- Zhang X, Yang S, Zhou Y, He Z, Xia X (2006) Distribution of the Rht-B1b, Rht-D1b and Rht8 reduced height genes in autumn-sown Chinese wheats detected by molecular markers. Euphytica 152:109–116
- Zhang XK, Xiao YG, Zhang Y, Xia XC, Dubcovsky J, He ZH (2008) Allelic variation at the vernalization genes Vrn-A1, Vrn-B1, Vrn-D1, and Vrn-B3 in Chinese wheat cultivars and their association with growth habit. Crop Sci 48:458–470
- Zhao C, Cui F, Wang X, Shan S, Li X, Bao Y, Wang H (2012) Effects of 1BL/1RS translocation in wheat on agronomic performance and quality characteristics. Field Crops Res 127:79–84
- Zhao FJ, Su YH, Dunham SJ, Rakszegi M, Bedo Z, McGrath SP, Shewry PH (2009) Variation in mineral micronutrient concentrations in grain of wheat lines of diverse origin. J Cereal Sci 49:290–295
- Zhou Y, Chen Z, Cheng M, Chen J, Zhu T, Wang R, Liu Y, Qi P, Chen G, Jiang Q, Wei Y, Luo MC, Nevo E, Allaby RG, Liu D, Wang J, Dvorak J, Zheng Y (2018) Uncovering the dispersion history, adaptive evolution and selection of wheat in China. Plant Biotechnol J 16(1):280–291
- Ziv O, Eyal Z (1977) Assessment of yield component losses caused in plants of spring wheat cultivars by selected isolates of septoria tritici. Phytopathology 68:791–796
- Zurn JD, Newcomb M, Rouse MN, Jin Y, Chao S, Sthapit J, See DR, Wanyera R, Njau P, Bonman JM, Brueggeman R, Acevedo M (2014) High-density mapping of a resistance gene to Ug99 from the Iranian landrace PI 626573. Molecular breeding 34(3):1