Chapter 13 Cultivation and Utilization of Shiitake Mushroom



F. Atila

Abstract Shiitake (*Lentinula edodes*) is the third most commonly cultivated edible mushroom species in the world. It has attracted people's attention with its medical properties as well as taste and nutritional value. Shiitake which has been known and used in Chinese medicine for more than 2000 years is now considered a great resource for modern clinical and pharmacological research. This mushroom contains many biologically active compounds (polysaccharides, lentinan, LEM and KS–2, ergosterol, nucleic acid derivatives, water-soluble lignins, eritadenine, etc.) which possess different medicinal effects such as antitumor, immunomodulatory, hypocholesterolemic, antibacterial, antifungal, anti–inflammatory and antioxidant. The chapter presents an overview of the research on the shiitake mushroom including its taxonomy, cultivation techniques, biotechnological approach, functional compounds and medicinal properties.

Keywords Lentinula edodes \cdot Medicinal mushroom \cdot Lentinan \cdot Eritadenine \cdot Anticancer

13.1 Introduction

Shiitakae (*Lentinula edodes*) is the third most commonly cultivated edible mushroomin the world, ranking just behind *Agaricus bisporus* and *Pleurotus* spp. representing about 17% of worldwide production (Zervakis and Koutrotsios 2017).

Although it is also known by different names such as the oakwood mushroom, the golden oak mushroom and the black forest mushroom (USA), xiang-gu and dong-gugo (China), lectin (France) in different parts of the world, today the name shiitake is the most widely used name for this mushroom. "Shiitake" name was derived from two words, "shii" (shii tree (*Castanopsis cuspidata* (Thunb.) Schottky)) and "take" (mushroom in Japanese), so shiitake means "mushroom of the shii or oak tree" in Japanese.

F. Atila (🖂)

Faculty of Agriculture, Department of Horticulture, Kirsehir Ahi Evran University, 40200 Kırşehir, Turkey

e-mail: funda.atila@ahievran.edu.tr

[©] The Author(s), under exclusive license to Springer Nature Switzerland AG 2021 H. M. Ekiert et al. (eds.), *Medicinal Plants*, Sustainable Development and Biodiversity 28, https://doi.org/10.1007/978-3-030-74779-4_13

The shiitake mushroom has been a symbol of healthy and youth for thousands of years in Far Eastern culture. Although shiitake production started in China and Japan, today there is also a significant increase in the production and consumption of shiitake in other parts of the world.

Until the mid-1980s, Japan that grown shiitake on natural logs was the main producer of shiitake in the world, while China became the major producer of shiitake in a short time with the development of sawdust-based techniques. According to the data of China Edible Mushrooms Association, in 2015, shiitake mushroom production accounted for 20% of the total edible mushroom production in China (Li et al. 2018), and this amount is also estimated to be approximately 98% of worldwide shiitake production (Yamanaka 2017).

Shiitake is a mushroom species that take people's attention with its high nutritional value and taste. Besides nutritional value, shiitake is one of the most widely known medical mushrooms in the world. The shiitake mushroom was known and used in classical Chinese medicine since more than 2000 years ago (Mizuno 1995). Shiitake contains several bioactive compounds, including polysaccharides, lentinan, LEM and KS–2, ergosterol, nucleic acid derivatives, water-soluble lignins and eritade-nine. Shiitake has immune modulating, anticarcinogenic and antitumor, antioxidative, antilipidemic, hepatoprotective, antiviral, antibacterial and antiparasitic effects in relation to these bioactive compounds contained in fruitbody and mycelium.

Shiitake can be consumed directly as fresh and dried food as well as health supplements. Nowadays, different types of supplements obtained from shiitake are sold as capsules, tonics or tablet in many of countries in the world (Bisen et al. 2010).

The chapter focuses on cultivation and medicinal properties of shiitake which is an edible and medicinal mushroom. The aim of this article is to gather and summarize available information on the shiitake mushroom, including its taxonomy, enzyme production, cultivation techniques, functional compounds and medicinal properties.

13.2 Geographic Distribution

Lentinula genus includes five morphologically defined species that were identified on the basis of morphology characteristics and geographical distribution by Pegler (1983) (Table 13.1). However, through subsequent research, it was determined that the phylogenetic relationships of *Lentinula* genus were more complex than the Pegler's would suggest, and apart from these five species, new *Lentinula* species were also identified (Mata and Petersen 2000; Hibbett 2001; Mata et al. 2001).

Shiitake (*L. edodes*) grows naturally throughout Southeast Asia, but the exact limits are uncertain. Samgina (1981) reported that *L. edodes* was found in Kaza-khstan. This report notes that the mushroom was found on conifer wood. But *L. edodes* is usually grow on *Quercus, Castanopsis and Lithocarpus* (Pegler 1983). Therefore, the species idetification may have been incorrect.

L. edodes was first identified as *Agaricus edodes* by Miles Joseph Berkeley in 1877. Then, Singer placed shiitake in *Lentinus* genus in 1936. David Pegler suggested

Species	Geographic distribution
Lentinula boryana (Berk. & Mont.) Pegler	Central America, northern South America, and the Gulf Coast states of North America
Lentinula guarapiensis (Speg.) Pegler	Paraguai
Lentinula edodes (Berk.) Pegler	North-east Asia
Lentinula lateritia (Berk.) Pegler	Southeast Asia and Australasia (except New Zealand
Lentinula novaezelandieae (Stev.) Pegler	New Zealand

 Table 13.1 Geographical distribution of Lentinula genus (Pegler 1983)

transferring this species to genus *Lentinula* based on microscopic observations in 1975. Molecular phylogenetic studies also support relocation of shiitake from the genus *Lentinus* to genus *Lentinula* (Molina et al. 1992; Hibbett and Vilgalys 1993; Hibbett and Donoghue 1996). Today, shiitake is classified in the genus *Lentinula*, the family *Tricholomataceae*, the order *Agaricales* and the subphylum of *Basidioimycotina*. But even now, the shiitake is often still being misspelt as *Lentinus edodes* (Berk.).

Shiitake is a fleshy gilled mushroom. The mushroom produces white-colored spores and white mycelia. Shiitake pileus that are light tan to dark brown is convex to applanate, and size of pileus ranges from 5 to 25 cm. The stipe is usually attached to the pileus centrally. Deep cracks can occur that reveal the underlying white tissue on pileus when shiitake grown on hardwood logs.

Shiitake is a saprophytic white-rot fungi that has the ability to enzymatically degrade cellulose, lignin and other macromolecules (Asgher et al. 2008). In the nature, this mushroom grows in cutting or dead logs particularly of the oak family (*Quercus* spp.) and various deciduous or broad-leaved trees in warm and humids regions (Royse 1997).

13.3 Nutritional Properties of Shiitake

Shiitake has high nutritional content as well as excellent flavor. This high-quality mushroom has important nutrients including dietary fiber (Mattila et al. 2002), minerals (George et al. 2014), vitamin B_{12} (Bito et al. 2014) and vitamin D (Jasinghe and Perera 2006), while it does not have vitamins A and C.

Also, the shiitake mushroom represents an excellent protein supplement. Dried shiitake contains a level of protein comparable to that of several different types of meat. In addition, shitake has 18 different amino acids and almost ideal ratios of eight essential amino acids (Turło et al. 2008). The essential amino acid content of shiitake is better than soybeans, meat, milk or eggs (Vetter 1995), especially they are rich in arginine and lysine (Liu and Bau 1980).

Dietary fiber plays an *important* role in preventing type 2 diabetes, insüline resistance, obesity, hypertension and some type of cancers (Galisteo et al. 2008). The dietary fiber content of shiitake is significantly higher than meats, whereas its fat content is much lower. Moreover, 77.7% of fatty acid content of shiitake consist of unsaturated fatty acids (Bisen et al. 2010). High dietary fiber and unsaturated fatty acid content of shiitake may help protect against cardiovascular diseases by lowering cholesterol values.

13.4 Major Active Compounds Isolated from Shiitake

13.4.1 Lentinan

Lentinan (β -(1 \rightarrow 3)–D–glucan) is a polysaccaride isolated from the fruiting bodies or mycelium of shiitake. It is situated in cell wall and has a high molecular weight. The estimated molecular weight of lentinan was reported as 400– 800 × 10³ Da by Ooi and Liu (2000). The chemical structure of lentinan is consisting of five β -(1 \rightarrow 3)–D–glucopyranoside in a linear linkages and two β -(1 \rightarrow 6)–D– glucopyranoside branches in side chains. This structure results in a right-handed triple-helical form (Wang et al. 2020). The biological activity is assossiated with the position of the glucose molecules in the helix structure (Surenjav et al. 2006). The molecular weight of polysaccades has also an influence on the immune stimulating effect as well as the degree of branching and chain conformation of them (Kulicke et al. 1997).

The lentinan displays numerous bioactivities such as immunomodulator and antitumor (Chihara et al. 1970; Zheng et al. 2005), antivirus (Guo et al. 2009), stimulating the expression of cytokines (Kupfahl et al. 2006) and hypocholesterolemic (Gu and Belury 2005). It does not have direct cytotoxic effects on cancer cells, instead it displays antitumor activity by strengthening the host immune system (Chihara et al. 1970) Moreover, lentinan has been recognized as a promising compound for the formulation of new functional foods and nutraceuticals due to its minimal side effects in addition to medicinal properties mentioned above. The sulfated derivatives of lentinan also exhibit bioactivities similar to lentinan such as strengthening the immune system (Guo et al. 2009).

13.4.2 Lem

Lentinula edodes mycelium (LEM) is a bioactive substance derived from powdered mycelia of shiitake harvested before fructification. The major active compound od LEM is a heteroglycan protein conjugate, being a protein-bound polysaccaride and largely composed of sugar (44%) and protein (24.6%) (Sugano et al. 1982). In addition to heteroglycan protein complex in the structure of LEM, various nucleic acid

derivatives, thiamine (vitamin B1), riboflavin (vitamin B2), ergosterol and eritadenine (Breene 1990), water-soluble lignins (Suzuki et al. 1990) and KS–2 (Fujii et al. 1978) are also presented.

LEM extract has thus been used as a medicinal food for at least 30 years in Japan (Yoshioka et al. 2012). LEM has been determined to have some bioactivity such as antioxidant (Akamatsu et al. 2004), hepatoprotective (Watanabe et al. 2006; Yoshioka et al. 2012), immunoregulatory activity and anticancer (Sugano et al. 1982; Kojima et al. 2010) activity. The antitumor activity of LEM is also thought to be associated with strengthening the host immune system rather than direct cytotoxicity, similar to lentinan. Macrophages are vital components of immune system, and LEM may play a significant role in macrophage stimulation. This macrophage activation is thought to be related to the antitumor and immunoregulatory activity of LEM (Morinaga et al. 1992).

13.4.3 Ks-2

KS–2 is a peptide–polysaccharide complex isolated from shiitake. KS–2 is consist of a-linked mannose and a small amount of peptide which is composed of serine, threonine and alanine with residual amounts of the other amino acids (Bisen et al. 2010). KS–2 polysaccharides possess antitumor and antiviral properties, and the molecular weight of KS–2 was reported as between 6.0×10^4 and 9.5×10^4 (Fujii et al. 1978).

13.4.4 Eritadenine

Eritadenine (2(R),3(R)–dihydroxy–4–(9– adenyl)–butyric acid), also known as lentinacin or lentysine, is a nucleic acid derivative produced mainly by shiitake. It was isolated from shiitake first time by Chibata et al. (1969) and Rokujo et al. (1970). Eritadenine is considered to be one of the major active substances accountable for hypocholesterolemic activity of shiitake (Sugiyama et al. 1995; Shimada et al. 2003; Enman et al. 2008; Bisen et al. 2010). To date, a number of studies (Sugiyama et al. 1995) have reported on the mechanism by which eritadenine exerts its cholesterol and triglyceride lowering properties, but the detailed mechanism is not yet completely explained.

13.4.5 Lectins

Lectins are carbohydrate-binding proteins (glycoproteins) of non-immunoglobulin origin, with an ability to produce cell agglutination (Dixion 1981). Various biological activities of mushroom lectins such as immunomodulatory and anti-tumor (Wang et al. 1996; Zhang et al. 2010), antivirus (Li et al. 2008) antifungal (Chandrasekaran et al. 2016), antibacterial (Chandrasekaran et al. 2016) and hypotensive (Wang et al. 1996) have been reported. However, there are few studies on the lectins of shiitake mushrooms (Jeune et al. 1990; Wang et al. 1999; Vetchinkina et al. 2008). Extracellular lectin activity of shiitake grown in submerged cultures was reported in some studies (Tsivileva et al. 2005; Wang et al. 1999; Vetchinkina et al. 2008).

The saline extract of the fruiting bodies of shiitake contains an amount of lectin that represents about 10% of the protein content of the fruitbody (Li et al. 2018). Lectin activity of fruiting bodies of shiitake is usually higher that that of lectins of mycelia. Mitogenic effects of lectin isolated from shiitake in human and murine were determined by Jeune et al. (1990) and Moon et al. (1995). A lectin isolated from shiitake has ability to agglutinate L1210 cell lines and HeLa cells (Moon et al. 1995).

13.4.6 Lentin

The lentin, a antifungal protein, isolated from the fruiting bodies of shiitake, exhibiting strong antifungal activity. It also showed an ability of inhibit proliferation of HIV–1 reverse transcriptase and leukemia cells (Ngai and Ng 2003).

13.4.7 Ergesterol

Ergosterol, (ergosta–5,7,22–trien–3β–ol) is a sterol found in cell membranes of mushrooms. It is abundant in shiitake as most of edible mushrooms. Ergosterol is provitamin form of D₂, and ultraviolet irradiation can convert these bioactive sterols to vitamin D₂ (Jasinghe and Perera, 2006; Morales et al. 2017). Shiitake mushrooms containing about 0.5% ergosterol (dry weight) were able to produce 400 IU of vitamin D per gram after being exposed to a fluorescent sunlamp (Breene 1990). Although mushrooms are traditionally dried under the sun, today this process is mostly performed in mechanical dryers.

Vitamin D_2 content of shiitake mushroom can be increased up to 5 times by exposure to direct sunlight for 3 h/day. Exposure to sunlight also increases the free amino acid content of the fruitbodies, making them sweeter and less bitter (Kiribuchi 1991).

13.5 Shiitake Cultivation

Shiitake is a saprotrophic mushroom, which means that they obtain the nutrients they need by decomposing various lignocellulosic waste. The ability of shiitake in converting complex lignocellulosics into simple organic compounds has been allowed many agricultural waste to be used in the cultivation of shiitake. In the commercial production of shiitake mushroom, two different techniques are applied, namely natural log cultivation and bag cultivation techniques in the world.

13.5.1 Natural Log Cultivation Technique

Cultivation of shiitake on natural logs began in far east almost a thousand years ago (Chang and Miles 2004). The first traditional cultivation method used natural logs, usually from the oak family such as shii tree (*Castanopsis cuspidata*) under outdoor conditions. This method has been the most common cultivation method until the mid-1980s.

Shiitake can be grown on many kind of hardwood and softwood trees, but oak (*Quercus*) are the most widely used species in natural log cultivation. Although logs from hardwood trees have longer fruiting period, harvest starts later than those of softwood logs. One of the most important points in the selection of logs is that shiitake mycelia colonize easily on the sapwood. Sapwood is living portion of log and contains polysaccharides needed for mycelial development, whereas colonization is difficult in the heartwood, a dead portion. For this reason, logs with a wide sapwood portion should be preferred while selecting for the production of shiitake.

Spawn running period in the log cultivation of shiitake may take 6–18 months, whereas cultivation cycle takes approximately 6 years depending on spawn, tree species, log size, moisture content of logs and climate factors, etc. Maximum biological efficiency is around 33%. Approximately 75% of the total yield is obtained the 2nd and 3rd years (Royse 2001).

Production of shiitake on log has been steadily declining with the development of sawdust-based techniques that has the shorter crop cycle and quick return of the money invested. However, the natural log technique has also some advantages such as requiring less care and labor, less susceptibility to microorganisms, rich in flavor and bioactive content. High molecular weight polysaccharide contents of shiitake mushrooms grown on logs are higher than those of fruitbodies grown on bags (Brauer et al. 2002).

13.5.2 Bag Cultivation Technique

After the middle of the 80s, a new method that uses plastic bags which are filled with lignocellulosic substrates has gradually replaced the traditional system on tree logs (Chang and Miles 2004). The time between the start and end of the crop cycle of the bag cultivation system is approximately 3 months, which corresponds to approximately 6% of the that of the log system. Moreover, in this technique, the biological efficiency is on average 75 to 125%, depending on the substrates used (3 times higher than the log system). The bag cultivation technique has advantages such as a shorter cultivation cycle, higher yields and year-round mushroom production, even if a bag cultivation technology needs relatively high initial investment cost of installation.

13.5.2.1 Growing Substrates

Although hardwood sawdust is the most commonly substrate for shiitake cultivation, various studies have shown that different agricultural or agro-industrial by-products that are locally abundant and cheaper such as cotton straw (Levanon et al. 1993), wheat straw and corn cobs (Philippoussis et al. 2003), sunflower hulls (Curvetto et al. 2002), hazelnut husk (Özçelik and Pekşen 2007), chickpea straw, sunflower head residue, alfalfa hay, corn stalk (Atila 2019a) may be alternative substrates for shiitake cultivation.

Supplementation of sawdust with millet, rice bran, sugarcane molasses, maize powder, rye, soy flour, grape pomace has improved mushrooms yields considerably (Royse 1996; Rossi et al. 2003; Royse and Sanchez 2007; Moonmoon et al. 2011; Atila 2019b), and but the use of alternative substrates rich in phenolic content such as olive press cake (Gregory and Pohleven 2014 and green walnut husk (Atila 2019b) seem to affect negatively mycelia growth and yield of shiitake. The situation could be associated with the presence of phenolic compound inhibiting mycelium growth and fructification.

13.5.2.2 Substrate Preparation

Optimal environmental conditions and choosing the suitable substrates are essential for success of shiitake cultivation. The first step of shiitake production is the preparation of the growing medium. Substrate preparation is based on process involving shredding, wetting, mixing and sterilization.

If sawdust is used as basal substrate in the preparation of the growing medium, no shredding is required. However, straw and other agricultural wastes such as corn cobs, cotton stalk and corn stalk need to be shredded into pieces 3-5 cm in length to facilitate disinfection and bagging process. Then, the substrates are soaked in water 6-12 h at room temperature and drained. Shiitake mycelium needs nitrogen sources as well as carbon. Therefore, several supplements rich in nitrogen such as bran, soybean

flour and maize powder are added to basal substrate. Overuse of supplements may cause the growth of some competitive organisms such as green mold (*Trichoderma* sp). After the mixing process is completed, the moisture of the growing medium is adjusted to 60–65% and filled in bags. Substrate disinfection is an important stage for maximum yield and quality in shiitake production. Autoclave sterilization method is generally used for substrate disinfection. However, it is difficult to use this method by some farmers due to the high cost of production and the need for expensive equipment. For this reason, steam sterilization has been recommended by some researchers (Mata and Savoie 1998; Savoie et al. 2000). However, this method is not commercially available.

13.5.2.3 Spawning Substrate and Incubation

After cooling, sterilized bags are inoculated with spawn at 1-5% (w/w) ratio in sterile environment. Incubation period must be carried out at 25 °C \pm 2 with a 12hour light and 12-hour dark cycle. Unlike the other commercial mushroom species, in the production of shiitake, vegetative phase takes place in two stages, spawn run and browning. At the end of the spawn running period, the entire surface of the substrate turns from white to brown, and a hard hipha crust forms on the surface of the substrate, indicating that mycelium is ready for fructification. Although it is not a general rule, after the incubation period, generally the plastic bags are removed, the substrate blocks are soaked or sprinkled with cold water for fruiting induction, and then, bags are transferred to the production room at 17 to 19 °C with a humidity of 90%. Moreover, lighting should be provided 12 h daily. Mushrooms should be harvested when they are turgid and before the pileus extends fully. Harvest may be done by hand, grasping the mushrooms at the base and turning them slightly so that they can be removed without physical damage. After obtaining the first harvest, blocks can be rehydrated to induce a second flush by soaking them in water for 12 h (Gaitán-Hernández and Mata 2004).

13.5.2.4 Yield

Shiitake mushrooms yield better in substrates containing moderate amounts of N, hemicellulose and lignin and with a low cellulose: lignin ratio (Atila 2019a). Biological efficiency differs considerably by ranging from 2.8 to 124.1% in sawdust-based substrates (Diehle and Royse 1996; Pire et al. 2001; Atila 2019b), up to 99.3% in straw-based substrates (Gaitan-Hernandez and Mata 2004; Philippoussis et al. 2007; Elisashvili et al. 2015) or from 102 to 112% in sunflowers hulls (Curvetto et al. 2002) (Table 13.2).

Substrate	Biological efficiency (%)	References
Sawdust (maple and Birch) Rice bran, millet	6.11–124.1	Diehle and Royse (1986)
Cotton straw (CS) Cotton straw + wheat straw (CWS)	CS-46 CWS-82	Levanon et al. (1993)
Sawdust Wheat bran, white millet, rye, CO ₃	59.1–99.6	Royse (1996)
Sugarcane baggase, Sugarcane leaves Pineapple crown	36.3–133.0	Salmones et al. (1999)
Various formulations of rice straw, chestnut sawdust, pinus sawdust, soyflour, rice, barley, maize flour, some chemical fertilizer	42.3–59.5	Morais et al. (2000)
Different types of sawdust	2.8–52.3	Pire et al. (2001)
Sugarcane bagasse + rice bran (25–30%)	98.42–99.84	Rossi et al. (2003)
Oak sawdust, wheat straw, corn cobs	19.44–54.17	Philippoussis et al. (2003)
Pasteurized Wheat Straw	24.8–55.6	Gaitan-Hernandez and Mata (2004)
Sunflower hulls + wheat bran	102–112	Curvetto et al. (2002)
Different formulations of corn cob, Euclyptus sawdust and rice bran	18.88-43.87	Eira et al. (2005)
Vineyard pruning, barley straw, wheat straw	37.02–93.25	Gaitan-Hernandez et al. (2006)
Different mixtures of wheat straw (WS), corn-cobs (CC), and oak wood sawdust (OS) and millet, wheat bran, soybean flour	41.07-80.64	Philippoussis et al. (2007)
HH alone and its mixtures with wheat straw (WS), beech wood-chip (BWC) and wheat bran (WB) in different ratios	43.73-87.73	Özçelik and Pekşen (2007)
Wheat straw Oak sawdust Rye, wheat bran, white millet	80.4–98.9	Royse and Sanchez (2007)
Sawdust with 10%–40% of rice bran wheat bran and maize powder	53.5–153.3 g/500 g substrate	Moonmoon et al. (2011)

 Table 13.2
 Biological efficiency of shiitake grown on different substrates

(continued)

Substrate	Biological efficiency (%)	References
Various formulations of oak sawdust, corn cobs, wheat straw, maize stubble, chopped cardboard, cotton waste, peanut husk, wheat kernels, wheat bran, rice meal, MgSO ₄ , CaCO ₃ , thiamine, urea	4.9-61	Martínez-Guerrero et al. (2012)
Sawdust (S) and straw (St) with different ratio of wheat bran (Wb)	S + Wb = 7.20 - 81.0 St + Wb = 11.8 - 66.8	Sharma et al. (2013)
Pasteurized wheat straw	66–320	Gaitan-Hernandez et al. (2014)
Chickpea straw, sunflower head residue, alfalfa hay; corn stalk	20.1–51.0	Atila (2019a)
Oak sawdust, grape pomace, green walnut hulls, tea wastes, olive press cake	38.2–70.7	Atila (2019b)

Table 13.2 (continued)

13.6 Medicinal Properties and Usage

Shiitake has been considered as a medicament for the inhibition and treatment of many diseases in the Orient for thousands of years. Today, medicinal properties of the shiitake have been also confirmed by a large number of high-quality scientific researches involving in vitro, in vivo and clinical studies. Extracts from shiitake, entire or part of fruit body, have been stated as having anti-cancer and antitumor, anti-hypercholesterolemic, hypoglycemic, hepatoprotective, antioxidant and antimicrobial (Table 13.3). In the following sections, major published laboratory and clinical studies on the some medicinal properties of shiitake were summarized.

13.6.1 Antitumor and Immunostimulating Activity

Cancer is the second leading cause of death globally. Although treatment methods such as chemotherapy and radiotherapy are widely used in cancer treatment, they are not always effective and often cause a number of side effects. Therefore, alternative methods of treatment, such as medicinal mushrooms, have attracted great attention of people all over the world.

Shiitake is a mushroom species known for its antitumor properties. Antiproliferative activities of this mushroom against prostate cancer, gastric cancer, breast cancer, colon carcinoma, lung cancer, skin cancer lines have been explored, and promising results have been obtained from many clinical and experimental trials carried out in shiitake (Vere White et al. 2002; Ng et al. 2002; Gu and Belury 2005; Fang et al.

Table 13.3 E	3iologically active cons	stituents isolated	d from shiitake and the	eir medicinal properties		
Major	Source	Extract	Pharmacological	Applications		References
Bioactive Compounds			effects	In vitro	In vivo	
Lentinan	Fruitbody	Ethanol	Immunomodulator		Mice	Chihara et al. 1970
	Fruitbody	5% NaOH–0.05% NaBH4	Immunomodulator	A colorimetric 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) method	Rats	Zheng et al. (2005)
	Mushroom fruiting bodies, mushroom spores and mushroom cultured broth	Ethanol	Hypocholesterolemic	mouse skin carcinoma cell line, CH72, and the non-tumor cell line, C50 cultured in modified Eagle's Minimal Essential Medium		Gu and Belury (2005)
	Fruitbody	Boiling water and ethanol	Antiviral		Chickens	Guo et al. (2009)
	Fruitbody		Antitumor		Mice	Ng and Yap (2002)
LEM	Culturated mycelia of shiitake	Ethanol	Antiviral	Cultured medium		Tochikura et al. (1988)
	Culturated mycelia of shiitake	Hot water and ethanol	Antioxidant Hepatoprotective	İnvitro	Dimethyl nitrosamine-injured mice	Akamatsu et al. (2004)
	Mycelia	Hot water	Hepatoprotective	primary cultured rat hepatocytes		Watanabe et al. (2006)
	LEM extract powder	Hot water and ethanol (50%)	Hepatoprotective	primary cultures of rat hepatocytes exposed to CCl4		Yoshioka et al. (2012)
	Mycelia	Water	Anticancer		Rats	Sugano et al. (1982;)
						(continued)

394

Table 13.3 (continued)					
Major	Source	Extract	Pharmacological	Applications		References
Bioactive Compounds			effects	In vitro	In vivo	
	Cultured mycelia	Hot water	Anticancer, immunoregulatory activity	In vitro		Kojima et al. (2010)
KS-2	Cultured mycelia	Hot water	Antitumor		Mice	Fujii et al. (1978)
Eritadenine	Dry mushroom	Ethanol	Hypocholesterolemic		Rats	Chibata et al. (1969)
	Fruitbody		Hypocholesterolemic		Rats	Rokujo et al. (1970;)
	Fruitbody		Hypocholesterolemic		Rats	Sugiyama et al. (1995;)
	Pure eritadenine		Hypocholesterolemic		Rats	Shimada et al. (2003)
	Mycelia	Methanol	Hypocholesterolemic	Submerged culture		Enman et al. (2008)
Lectin	Fruitbody	Physiological saline	Antitumor		Animals and human	Moon et al. (1995)
	Mycelium, brown mycelial film, primordium and fruitbody	Water	Hemagglutinating activity	Several agar media		Tsivileva et al. (2001)
						(continued)

13 Cultivation and Utilization of Shiitake Mushroom

395

Table 13.3 (continued)					
Major	Source	Extract	Pharmacological	Applications		References
Bioactive Compounds			effects	In vitro	In vivo	
<i>O</i> -sulfonated α-D-glucan	Fruitbody	5% NaOH/0.05% NaBH4	Antitumor		Mice	Unursaikhan et al. (2006)
	Fruitbody	Ethanol	Antiviral		Tobacco plants infected tobacco mosaic viruses	Wang et al. (2015)
Mycelial extracts	Mycelia	Water	Antiviral	Culture media		Sasaki et al. (2001)
LEP	Fruitbody	Aqueous and ethanol extracts	Antiviral	Poliovirus, Bovine herpesvirus cells grown in Dulbecco's Modified Eagle Medium (DMEM)		Rincão et al. (2012)
Chitosan	Stripe of fruitbody	Aqueous NaOH	Antioxidant	The conjugated diene method		Yen et al. (2007)

 Table 13.3 (continued)

2006; Ina et al. 2013; Zhang et al. 2018). Biologically active substances isolated from shiitake exhibit numerous mechanisms of anticancer activity such as the inhibition of angiogenesis (Deocaris et al. 2005), stimulation of the cancer cells for apoptosis (Fang et al. 2006) or retarding the development of tumors (Ng et al. 2002).

Lentinan, known as immunomodulatory and anticancer agents, is the most broadly studied compound in the shiitake mushroom. The antitumour effect of lentinan is attributed to stimulation of the immune response in various investigations (Chihara et al. 1970; Fujii et al. 1978; Ina et al. 2013). Moreover, lentinan is reported to trigger hematopoietic stem cells, macrophages and natural killer cells (Akramiene et al. 2007). Similarly, macrophage achieved from the KS–2 treated mice. KS–2 strongly inhibited tumor growth in mices who administered orally in both doses between 1 and 100 mg/kg (Fuji et al. 1978).

Antitumor properties of shiitake are not only due to polysaccharides, and a lectin isolated from shiitake was an agglutinin of tumor cell lines tested by L1210 and HeLa cells (Moon et al. 1995). Arginine, a substance used in the supplement of cancer patients, is also abundant in shiitake (Eghianruwa et al. 2011). IA–a (a glycogen-like structure) and IA–b (arabinoxylan-like polysaccharide) isolated from LEM stimulate cytokine production and phagocytosis in RAW264.7 cells (Kojima et al. 2010). The O–sulfonated α –D–glucan of shiitake has higher antitumor activity than those of the native glucan (1 \rightarrow 3)– α –D –glucans and the antitumor activity of the native glukans against S–180 can be enhanced by O–sulfonation of these glukans (Unursaikhan et al. 2006).

Several researcher reported that use of the combination of lentinan and chemotherapy drugs has inhibited proliferation and induced apoptosis than use of chemoterapy drugs alone (Zhao et al. 2013; Liu et al. 2015; Sun et al. 2015). Clinical researches have revealed that lentinan is effective in prolonging survival in patients with stomach, ovarian or colorectal cancer (Borchers et al. 1999; Fujimoto et al. 2006) and prevents side effects such as nausea and asthenia, which are common in chemotherapeutic treatment of D. Moreover, administration of lentinan in combination with Bacillus Calmette–Guerin (BCG) vaccine which is used against tuberculosis induces activation of immune cells in the lung tissue (Drandarska et al. 2005).

13.6.2 Hypocholesterolemic Activity

Coronary artery disease (CAD) is the first cause of death worldwide. The most important risk factors of the disease are hypercholesterolemia, obesity, diabetes, high triglycerides and low density lipoprotein cholesterol (LDLc) levels, hypertension and cigarette smoking as well as genetic factors (Abdel-aziz and Mohamed 2013).

The ability of shiitake to lower cholesterol was described for the first time by Kamiya et al. (1969). The major active hypocholesterolemic component in the shiitake mushroom is a adenosine derivative eritadenine. (Takashima et al. 1973). In addition to eritadenine, nucleic acid compounds extracted from shiitake were found to be inhibitors of platelet agglutination (Sugiyama et al. 1995).

Eritadenine can reduce cholesterol level in plasma and expedite lipid accumulation in the liver by removing it from the circulations. LDL is converted into high-density lipoprotein (HDL) cholesterol which is beneficial for the human system in the liver (Kabir and Kimura 1989). Another suggestion regarding the activity of eritadenine is that high dosages of eritadenine may break down the secretion of very low density lipoprotein (VLDL) and reduce cholesterol by lowering the ratio of phosphatidylcholine (PC) to phosphatidylethanolamine (PE) in liver microsomes (Sugiyama et al. 1995). However, the mechanism that reveals the cholesterol-lowering effect of eritadenine has not been fully elucidated. Eritadenine signafically reduced serum cholesterol, phospholipids and triglycerides, both in intact rats and in animals fed a high-fat diet (Rokujo et al. 1970). The eritadenine content in the shiitake mushrooms was in the range 3.2-6.3 mg/g dried mushrooms (Enman et al. 2007). A diet containing eritadenine (0.005%) caused a 25% reduction in total cholesterol within 1 week (Chibata et al. 1969). The use of eritadenine 10-21 mg/kg/day in male and female rats decreased the atherogenic index (TC/HDL) in rat sera (Morales et al. 2018). Although the hypocholesterolemic effect of eritadenine has been investigated in several studies on rats (Sugiyama et al. 1995; Shimada et al. 2003), there are few human studies in the literature (Suzuki and Ohshima 1976).

The experimental and clinical data show that shiitake has beneficial effects on lowering low-density lipoproteins, total cholesterol and triglycerides, as well as in preventing the diseases such as arterial hypertension and high blood sugar levels which are effective in the development of cardiovascular diseases. (Kabir and Kamura 1989; Yang et al. 2002).

13.6.3 Antioxidant Activity

Oxidative stress occurs as a result of an imbalance caused by increased reactive oxygen species (ROS) and/or decreased antioxidant defense systems of the body. Numerous studies have demonstrated that oxidative stress plays a role in the emergence and development of some diseases such as atherosclerosis (Kattoor et al. 2017), muscle wasting (Moylan and Reid 2006), hypertension (Higashi et al. 2002), neurodegeneration (Brown 2005) and stroke (Cherubini et al. 2005).

Methanolic extract of shiitake is a promising alternative for use as an antioxidant (Sasidharan et al. 2010). LEP (polysaccharides isolated from shiitake), which can act as an antioxidant, may play a role in healing oral ulceration. LEP administration, in rats with oral ulceration, significantly was increased activities of serum antioxidant enzymes, whereas it was decreased levels of serum, mucosal interleukin–2 (IL–2) and tumor necrosis factor alpha (TNF–a) (Yu et al. 2009). Moreover, the administration of LEP can stimulate the expression of genes encoding antioxidant enzymes, reducing the increased oxidation stress-induced feeding by high-fat diet in rats. In addition, this treatment would decrease expression of VCAM–1mRNA of thoracic aorta endothelial cell in rats (Xu et al. 2008). Another substance that

has strong antioxidant and anti-inflammatory properties in the shiitake mushroom is ergothioneine, and it is an amino acid analog (Jang et al. 2016).

Heat treatment significantly increases the antioxidant activities of shiitake mushrooms. The antioxidant activity of raw shiitake mushroom is increased about 2.0–fold by heat treated at 121 °C for 30 min (Choi et al. 2006). Although freeze drying is suggested for the protection of eritadenine and protein content of fruitbody, hot air drying at 50 °C has been proposed for the stability or formation of total phenolics in shiitake (Zhang et al. 2013).

13.6.4 Hepatoprotective Activity

Shiitake has direct protective effects on hepatocytes. Methanolic extract of shiitake fruitbody can protect liver cells from paracetamol-induced liver damage, with its antioxidative effect on hepatocytes, thus reducing or eliminating the harmful effects of toxic metabolites of paracetamol. Administration of shiitake extract at a dose of 200 mg/kg for seven days to paracetamol-induced hepatatoxic mice reduces the activity of serum enzymes and bilirubin, resulting in significant hepatoprotective effects (Sasidharan et al. 2010). A significant reduction in liver injury was also noted when mice with severe liver damage were fed vitamin D–enriched shiitake mushroom extracts (Drori et al. 2016).

Not only the fruit bodies of shiitake, but also its mycelia has hepatoprotective activity. Polyphenolic compounds contained in the L.E.M. seemed to be responsible for the protective effect (Watanabe et al. 2006). Oral administration of the extracts of LEM has the protective effect against CCl_4 (Chen 2012) and D–galactosamine (Watanabe et al. 2006) induced hepatic injury in rats. Hot water and ethanol extracts of L.E.M. repress the development of liver fibrosis induced by dimethylnitrosamine (DMN) and inhibit proliferation and morphological change of isolated rat hepatic stellate cells (HSCs) (Akamatsu et al. 2004). LEM containing anti-oxidation and anti-inflammation activities might be used for alleviating side effects of chemotherapy and preventing the progression of liver cancer for patients with chronic hepatitis (Yagi 2012).

13.6.5 Antimicrobial Activity

Antibiotics are widely used as therapeutic agents in the treatment of many diseases. But, with increasing bacterial resistance to antibiotics, plants and mushrooms with antibacterial activity have attract attention. The superior abilities of fungi in improving host immunity can be very useful in fighting infection. Shiitake contains several compounds such as lentinan, which have the ability to stimulate humoral immunity to help prevent bacterial infections that are resistant to antibiotics (Markova et al. 2003; Hatvani 2001).

Sources	Target organisms	References
Aqueous extracts of dry and fresh mushroom	Bacillus subtilis, Esherichia coli	Casaril et al. (2011)
Chloroform, ethylavetate and water extracts of dried mushroom	Streptococcus spp., Actinomyces spp., Lactobacillus spp., Provotella spp., Porphyromonas spp.	Hirasawa et al, (1999)
The culture filtrate after 18–25 days of cultivation of shiitake	Bacillus subtilis	Ishikawa et al. (2001)
Mushroom extract	Aspergillus ochraceus and Penicillium verrucosum	Ricelli et al. (2002)
Lentin isolated from fruitbody of shiitake	Physalospora piricola, Botrytis cinereal, Mycosphaerella arachidicola	Ngai and Ng, (2003)
Mycelial extracts of shiitake	Helminthosporium euphorbiae, Helminthosporium sp, Fusarium solani and Phomopsis sojae	Sasaki et al. (2001)
Mushroom extract	Aspergillus parasiticus, Aspergillus flavus	Reverberi et al. (2011)

 Table 13.4
 Antimicrobial activity of shiitake in vitro

Extracts and pure compounds of shiitake exhibit high levels of antimicrobial activity, including the antibacterial and antifungal action. The effect of shiitake on the growth of various bacteria and fungi was revealed by several authors (Hirasawa et al. 1999; Ishikawa et al. 2001; Sasaki et al. 2001; Ricelli et al. 2002; Ngai and Ng 2003; Reverberi et al. 2011; Casaril et al. 2011) in vitro studies (Table 13.4).

13.6.6 Antiviral Activity

Various extracts of shiitake mushroom and some polysaccharides isolated from shiitake have been suggested as sources of potential antiviral agents. Aqueous and ethanol extracts and polysaccharide (LeP) from shiitake are effective in the replication of poliovirus type 1 (PV–1) and bovine herpes virus type 1 (BoHV–1) (Rincão et al. 2012). Improvement was observed in liver function tests in patients with cornic hepatitis B and seropositive for hepatitis B (Hbe) antigenemia who consumed 6 g of LEM orally daily for 4 months, while some patients undergo a change from HBeAgpositive to anti-HBe positive (Amagase 1987). A laccase isolated from fresh fruitbody of shiitake mushroom exhibited inhibitory activity to HIV–1 (Sun et al. 2011). LEM and ethanol extract of LEM blocked the HIV virus at the initial stage of its development (Tochikura et al. 1988). Clinical and in vitro studies shown that LEM has ability of inhibition of HIV infection of cultured T-cells. LEM increased the T-cell count in

HIV patients with AIDS symptoms from 1250/mm³ to 2550/mm³, and the symptoms were much improved after 60 days (Izuka 1990). Administration of aqueous extracts from shiitake in dose of 0.4–2 mg/mice protected mice against lethality induced by the herpes simplex type 2 virus (Razumov et al. 2013). The feeding of influenza virus-infected mice for 2 weeks with a mixture of glucans obtained mycelial mush-room powders of Shiitake significantly reduced the clinical symptoms of infection. (Vetvicka and Vetvickova 2015). The possibility of using shiitake polysaccarides and other compounds in the control and prevention of viral infections that affect plants and animals was also reported (Sasaki et al. 2001; Wang et al. 2015).

13.6.7 Dosage and Toxicity

It is important to know the efficacious and safety doses of dietary supplements and nutraceuticals in order to benefit effectively from them. The recommended dose is 6–16 g for dried shiitake and about 90 g for fresh fruitbody (Liu and Bau 1980). One study on mice showed that the daily intake of 100 mg/kg of shiitake mushroom could have potential health-improving effects (Grotto et al. 2016). Since an aqueous extract of shiitake fruitbody reduces the activity of blood platelets in the process of coagulation, especially people who are taking blood thinners should be careful when using shiitake or water-soluble fractions (Yang and Jong 1989).

The doses of compounds isolated from shiitake such as lentinan are lower than that of mushroom consumption. Lentinan has been found safe to be administered to humans by IV injection in a dose range of 1–5 mg/day once or twice a week, and greater doses can cause immune suppression (Taguchi et al. 1982; Aoki 1984). In the early stages of AIDS or chronic hepatitis, the best dose of LEM was recommended between 2–6 g per day in 2 or 3 divided doses orally, while the dose may be reduced to 1/2–1 g per day once the disease becomes more stable (Sharon 1988). Lentinan and LEM have no known serious side effects (Aoki 1984). Some people may experience minor side effects or allergic reactions, known as shiitake dermatitis, caused by glucan lentinan (Nguyen et al. 2017).

13.7 Biotechnological Approach

13.7.1 Submerged Liquid Fermentation with Shiitake

Submerged liquid fermentation (SLF) techniques are used in different areas such as liquid spawn production, enzyme production and biomass production for pharmaceutical and nutraceutical applications.

Several researchers reported that liquid has a shortened spawn running period and a higher yield in comparison with the grain spawn (Kawai et al. 1996; Leatham

and Griffin 1984; Lee et al. 2019). On the other hand, thanks to the liquid spawn technology, mycelia can be stored for a long time (Zilly et al. 2011). But there may be some problems in use of liquid spawn such as degeneration and mutation (Itävaara 1993). Submerged liquid cultivation can also be a promising alternative method for bioactive molecules to be obtained in a shorter time, with a higher amount and with less risk of contamination (Harvey et al. 2001; Tepwong et al. 2012). Moreover, various enzyme activities of shiitake have been determined in submerged liquid cultures (Buswell et al. 1996; Nagai et al. 2002). It is possible to eliminate the toxicity of wastewaters by using this feature of shiitake mycelia (D'Annibale et al. 2004). The medium composition and environmental parameters are crucial for optimal biomass, enzyme or metabolite production (Enman et al. 2008; Lee et al. 2019).

13.7.2 Utility of Spent Shiitake Mushroom Substrate

The total production of cultivated mushrooms in the world was approximately 34 million tons in 2013, and shiitake accounted for 7.48 million tons (Royse et al. 2017). Every kg of mushroom produced 5 kg of wet mass SMS (Medina et al. 2012). When 35–40% of the waste compost is calculated as dry matter, the resulting spent shiitake mushroom substrate (SSMS) in 2013 can be estimated about 13 million tons.

Shiitake mycelia secrete various enzymes capable of breaking down polyphenols, including lignin peroxidase, Mn-dependent peroxidase and laccase (Asgher et al. 2008). SSMS contains plenty of shiitake mycelia and can biodegrade organic xenobiotic compounds found in soil and water and adsorb some pollutants (Ahlawat and Sing 2009). This allows the use of SSMS in some biotechnological applications easily and cheaply. The biotechnological areas, in which SSMS can be used, are reviewed in the following section.

13.7.2.1 Bioremediation

The extracellular enzyme system of shiitake developed unique non-specific enzyme systems with the ability to attack not only lignin but also a broad spectrum aromatic compounds as well as some non-aromatic organopollutants such as pentachlorophenol (Okeke et al. 1993), 17α -ethinylestradiol (Eldridgea et al. 2017) and 2,4-dichlorophenol (Tsujiyama et al. 2013). The use of shiitake cultures for these remediation of contaminated soils could be less expensive and beneficial for human health and environment. Moreover, the biological degradation of pollutants using shiitake spent substrates would reduce the cost of disposal.

13.7.2.2 Dye Wastewater Degradation

A large amount of dye wastewaters, which are harmful to the environment and human health, are released into the environment from paint factories and other dye-using industries. Although some methods are applied to dispose of these wastewaters, they are expensive applications (Moreira et al. 2000). Biodegradation of colored wastewater by ligninolytic enzymatic system of white rot fungus appears to be an attractive alternative. Use of shiitake in the degredation processes has a important potential. Several strains of shiitake also have been reported as the dye- or colored material-degrading organisms (Hatvani and Mecs 2002; Boer et al. 2004). On the other hand, olive oil mill water (OMWW) has inhibitor effects of on plant growth, microbial activity and soil properties (Rusan et al. 2016; Mekki et al. 2013). The main cause of the harmful effects of OMWW is considered to be the presence of high concentrations of phenolic compounds (Azam et al. 2002). Some of strain of shiitake show high performance for treatment of OMW, resulting a significant decrease in the color and phenolics concentration of OMWW (Lakhtar et al. 2010).

13.7.2.3 Ethanol Production

In the traditional production of bioethanol, which attracts attention as an alternative fuel today, lignocellulosic materials with high sugar and starch content such as straw, corn, sugar cane, potatoes are used (Watanabe et al. 2010; Belal 2013; Patni et al. 2013). However, uses of most of these products as food cause significant cost increases.

Shiitake growing medium is prepared from various agricultural and forest wastes. SSMS remaining after mushroom production contains high amount of lignin. The use of this waste material with high lignocellulosic content as a raw material in ethanol production reduces the cost (Asada et al. 2011). The producing ethanol from SSMS has been demonstrated in the several studies (Asada et al. 2011; Hiyama et al. 2016; Xiong et al. 2019). However, further studies are needed to develop technologies that will integrate mushroom and biofuel production.

13.8 Future and Perspective

Shiitake is a type of mushroom that is appreciated not only for its unique flavor and nutritional value, but also for its health benefits. Despite these superior properties, some problems with the production and medicinal use of this mushroom need to be addressed in order for shiitake to reach its deserved place.

Although shiitake is widely consumed in some countries, the amount of consumption in some countries is very low or absent. The reasons for low consumption are high prices, lack of familiarity with the mushroom species and low production quantities. Increasing local production and developing new technologies to get higher yield of shiitake can lead to a decrease in the retail price. Moreover, shiitake production can be a good source of income for farmers, especially living in rural areas of less developed countries, and limited resources can be used beneficially.

In the rural areas, the biggest problem for the farmers could be the supply of growing substrate for shiitake cultivation. In countries where the shiitake production sector does not improve, it is not possible for farmers to obtain the growing substrates from a supplier. On the other hand, it is difficult to prepare the growing medium by himself, because the disinfection of the growing medium in the cultivation of shiitake is carried out by autoclave sterilization method and the establishment and implementation costs of this method are high. Therefore, it is important to develop alternative disinfection methods in order to expand cultivation of shiitake in rural areas. Selection of strains adapted to different substrates and disinfection methods is critically important for prolonged crop cycle and high mushroom yield. Moreover, researches for the selection and breeding of genotypes with high production capacity at high temperatures will be of great benefit in reducing energy consumption in tropical regions and summer production.

Consumers are increasingly interested in functional foods that are proven to help improve human health. Some bioactive substance of shiitake are especially effective at strengthening the immune system and lowering cholesterol and used as active compounds in the development of various functional foods. Although in vitro and in vivo studies conducted with identified bioactive compounds isolated from shiitake offer exciting results for this compounds to be qualified as a functional food and source of potential drugs, more information is required about the therapeutic effects of bioactive ingredients. Further studies, including clinical trials, to determine dose ranges that are both safe and useful in the treatment or prevention of diseases are required to obtain maximum benefit from bioactive compounds without negative consequences. Moreover, detailed studies on the chemical structures and mechanisms of action of some less researched bioactive substances isolated from shiitake may contribute to the determination of the new medicinal properties of the fungus.

Although the content of bioactive substance of shiitake may be affected by the strain, growing substrates and culture conditions, there are no protocols yet to ensure standardization of the quality and quantity of bioactives. For this reason, it is one of the conditions that should be taken into consideration that approved standard production protocols may be required to guarantee the quality and effectiveness of fungal products to be used for pharmaceutical applications. The production of high-quality products of standard quality and the provision of sustainable production under controlled conditions should be determined as the most important targets. The submerged liquid fermentation method appears to be much more useful in achieving these targets.

On the other hand, factors such as temperature and humidity applied in the cultivation of mushrooms provide optimum conditions for the development of harmful fungi species, bacteria and pests. For this reason, it is common to use pesticides to solve the problems related to diseases and pests that are frequently encountered in mushroom production. Considering the negative effects of these pesticides on human health, it should be preferred to apply controlled and certified production systems such as organic agriculture or good agricultural practices in the production of mushrooms that will be used for medical purposes. Another solution proposal in this regard is to use a submerged culture instead of fruitbody to produce bioactive metabolites for pharmaceutical applications. Mushroom mycelium is known to be very rich in bioactive content. While it is possible to produce mycelium without using chemicals in the production of submerged mycelia culture, it will be much easier to set this type of production to standards.

References

- Abdel-aziz TA, Mohamed RH (2013) Association of endothelial nitric oxide synthase gene polymorphisms with classical risk factors in development of premature coronary artery disease. Mol Biol Rep 40:3065–3071
- Ahlawat OP, Sing R (2009) Influence of pH, temperature and cultural media on decolorization of synthetic dyes through spent substrate of different mushrooms. J Sci End Res 68:1068–1074
- Akamatsu S, Watanabe A, Tamesada M, Nakamura R, Hayashi S, Kodama D, Kawase M, Yagi K (2004) Hepatoprotective effect of extracts from *Lentinus edodes* mycelia on dimethylni-trosamine—induced liver injury. Biol Pharm Bull 27(12):1957–1960
- Akramiene D, Kondrotas A, Didziapetriene J, Kevelaitis E (2007) Effects of beta-glucans on the immune system. Medicina (kaunas) 43(8):597–606
- Amagase H (1987) Treatment of hepatitis B patients with *Lentinus edodes* mycelium. In: Proceedings of the XII international congress of gastroenterology. Lisbon, p 197
- Aoki T (1984) Lentinan. In: Fenichel RL, Chirgis MA (eds) Immune, modulation agents and their mechanisms, vol 25. Immunol Studies, pp 62–77
- Asada C, Asakawa A, Sasaki C, Nakamura Y (2011) Characterization of the steam-exploded spent Shiitake mushroom medium and its efficient conversion to ethanol. Bioresour Tech 102:10052– 10056
- Asgher M, Bhatti HN, Ashraf M, Legge RL (2008) Recent developments in biodegradation of industrial pollutants by white rot fungi and their enzyme system. Biodegradation 19:771–783
- Atila F (2019a) Compositional changes in lignocellulosic content of some agro-wastes during the production cycle of shiitake mushroom. Sci Hort 245:263–268
- Atila F (2019b) The use of phenolic-rich agricultural wastes for *Hericium erinaceus* and *Lentinula edodes* cultivation. Ege Üniv Ziraat Fak Derg 56(4):417–425
- Azam F, Muller C, Weiske A, Benckiser G, Ottow J (2002) Nitrification and denitrification as sources of atmospheric nitrous oxide—role of oxidizable carbon and applied nitrogen. Biol Fert Soil 35(1):54–61
- Belal EB (2013) Bioethanol production from rice straw residues. Braz J Microbiol 44(1):225-234
- Berkeley MJ (1857) Introduction to cryptogamic botany. H.Bailliere, London
- Bisen PS, Baghel RK, Sanodiya BS, Thakur GS, Prasad GBKS (2010) *Lentinus edodes*: a macrofungus with pharmacological activities. Curr Med Chem 17:2419–2430
- Bito T, Teng F, Ohishi N, Takenaka S, Miyamoto E, Sakuno E, Terashima K, Yabuta Y, Watanabe F (2014) Characterization of vitamin B₁₂ compounds in the fruiting bodies of shiitake mushroom (*Lentinula edodes*) and bed logs after fruiting of the mushroom. Mycoscience 55:462–468
- Boer CG, Obici L, Souza CGM, Peralta RM (2004) Decolorization of synthetic dyes by solid state cultures of *Lentinula (Lentinus) edodes* producing manganese peroxidase as the main ligninolytic enzyme. Bioresour Technol 56:1384–1387
- Borchers AT, Stern JS, Hackman RM, Keen CL, Gershwin ME (1999) Mushrooms, tumors, and immunity. Proc Soc Exp Biol Med 221:281–293

- Brauer D, Kimmons T, Phillips M (2002) Effects of management on the yield and highmolecularweight polysaccharide content of shiitake (*Lentinula edodes*) mushrooms. J Agri Food Chem 50:5333–5237
- Breene W (1990) Nutritional and medicinal value of specialty mushrooms. J Food Prot 53:883-994
- Brown DR (2005) Neurodegeneration and oxidative stress: prion disease results from loss of antioxidant defence. Folia Neuropathol 43(4):229–243
- Buswell JA, Cai YJ, Chang ST, Peberdy JF, Fu SY, Yu H-S (1996) Lignocellulolytic enzyme profiles of edible mushroom fungi. World J Microbiol Biotechnol 12:537–542
- Casaril KBPB, Kasuya MCM, Vanetti MCD (2011) Antimicrobial activity and mineral composition of shiitake mushrooms cultivated on agricultural waste. Braz Arch Biol Technol 54(5):991–1002
- Chang S, Miles P (2004) Cultivation, nutritional value, medicinal effect, and environment impact. CRC Press, Florida
- Chandrasekaran G, Lee YC, Park H, Wu Y, Shin HJ (2016) Antibacterial and antifungal activities of lectin extracted from fruiting bodies of the Korean cauliflower medicinal mushroom, *Sparassis latifolia* (Agaricomycetes). Int J Med Mushroom 18(4):291–299
- Chen J (2012) Recent advance in the studies of β -Glucans for cancer therapy. AntiCancer Agents Med Chem 13:679–680
- Cherubini A, Ruggiero C, Polidori MC, Mecocci P (2005) Potential markers of oxidative stress in stroke. Free Radic Biol Med 39(7):841–852
- Chibata I, Okumura K, Takeyama S, Kotera K (1969) Lentinacin: a new hypocholesterolemic substance in *Lentinus edodes*. Experientia 25:1237–1238
- Chihara G, Hamuro J, Maeda Y, Arai Y, Fukuoka F (1970) Fractionation and purification of the polysaccharides with marked antitumour activity especially lentinan from *Lentinus edodes*. Cancer Res 30:2776–2781
- Choi Y, Lee SM, Chun J, Lee HB, Lee J (2006) Influence of heat treatment on the antioxidant activities and polyphenolic compounds of shiitake (*Lentinus edodes*) mushroom. Food Chem 99:381–387
- Curvetto N, Figlas D, Delmastro S (2002) Sunflower seed hulls as substrate for the cultivation of shiitake mushrooms. Hort Technol 12:652–655
- D'Annibale A, Casa R, Pieruccetti F, Ricci M, Marabottini R (2004) Lentinula edodes removes phenols from olive-mill wastewater: impact on durum wheat (Triticum durum Desf.) germinability. Chemosphere 54:887–894
- Deocaris CC, de Castro MC, Oabel AT, Co EL, Mojica ER (2005) Screening for anti-angiogenic activity in Shiitake mushroom (*Lentinus edodes* Berk) extracts. J Med Sci 5(1):43–46
- deVere White RW, Hackman RM, Soares SE, Beckett LA, Sun B (2002) Effects of a mushroom mycelium extract on the treatment of prostate cancer. Urology 60(4):640–644
- Diehle DA, Royse DJ (1986) Shiitake cultivation on sawdust: evaluation of selected genotypes for biological efficiency and mushroom size. Mycologia 78(6):929–933
- Dixion HBF (1981) Defining a lectin. Nature 292:192
- Drandarska I, Kussovski V, Nikolaeva S, Markova N (2005) Combined immunomodulating effects of BCG and *Lentinan* after intranasal application in guinea pigs. Int Immunopharm 5(4):795–803
- Drori A, Shabat Y, Ya'acov AB, Danay O, Levanon D, Zolotarov L, Ilan Y (2016) Extracts from *Lentinula edodes* (Shiitake) edible mushrooms enriched with vitamin D exert an antiinflammatory hepatoprotective effect. J Med Food 19(4):383–389
- Eghianruwa Q, Odekanyin O, Kuku A (2011) Physicochemical properties and acute toxicity studies of a lectin from the saline extract of the fruiting bodies of the shiitake mushroom, *Lentinula edodes* (Berk). Int J Biochem Mol Biol 2(4):309–317
- Eira FCD, Meirelles WF, Paccola-Meirelles LD (2005) Shiitake production in corncob substrates. Rev Bras Milho Sorgo 4(2):141–148
- Eldridgea HC, Milliken A, Farmerd C, Hamptone A, Wendlanda N, Cowardf L, Gregory DJ, Johnson CM (2017) Efficient remediation of 17α–ethinylestradiol by Lentinula edodes (shiitake) laccase. Biocatal Agric Biotechnol 10:64–68

- Elisashvili V, Kachlishvili E, Asatiani M (2015) Shiitake medicinal mushroom, *Lentinus edodes* (higher Basidiomycetes) productivity and lignocellulolytic enzyme profiles during wheat straw and tree leaf bioconversion. Int J Med Mushrooms 17:77–86
- Enman J, Rova U, Berglund KA (2007) Quantification of the bioactive compound eritadenine in selected strains of shiitake mushroom (*Lentinus edodes*). J Agric Food Chem 55:1177–1180
- Enman J, Hodge D, Berglund KA, Rova U (2008) Production of the bioactive compound eritadenine by submerged cultivation of Shiitake (*Lentinus edodes*) mycelia. J Agric Food Chem 56:2609– 2612
- Fang N, Li Q, Yu S, Zhang J, He L, Ronis MJJ, Badger TM (2006) Inhibition of growth and induction of apoptosis in human cancer cell lines by an ethyl acetate fraction from shiitake mushrooms. J Altern Complement Med 12(2):125–132
- Fujii T, Aeda H, Suzuki F, Shida N (1978) Isolation and characterization of a new antitumor polysaccharide, KS-2 extract from culture mycelia of *Lentinus edodes*. J Antibiot 11:1079–1090
- Fujimoto K, Tomonaga M, Goto S (2006) A case of recurrent ovarian cancer successfully treated with adoptive immunotherapy and lentinan. Anticancer Res 26:4015–4018
- Gaitan-Hernandez R, Mata G (2004) Cultivation of the edible mushroom *Lentinula edodes* (shiitake) in pasteurized wheat straw-alternative use of geothermal energy in Mexico. Eng Life Sci 4(4):363–367
- Gaitán-Hernández R, Esqueda M, Gutiérrez A, Sánchez A, Beltrán-García M, Mata G (2006) Bioconversion of agrowastes by *Lentinula edodes*: the high potential of viticulture residues. Appl Microbiol Biot 71:432–439
- Gaitan-Hernandez R, Cortes N, Mata G (2014) Improvement of yield of the edible and medicinal mushroom *Lentinula edodes* on wheat straw by use of supplemented spawn. Braz J Microbiol 45(2):467–474
- Galisteo M, Duarte J, Zarzuelo A (2008) Effects of dietary fibers on disturbances clustered in the metabolic syndrome. J Nutr Biochem 19:71–84
- George PL, Ranatunga TD, Reddy SS, Sharma GC (2014) A comparative analysis of mineral elements in the mycelia and the fruitbodies of shiitake mushrooms. Am J Food Tech 9(7):360–369
- Gregory A, Pohleven F (2014) Cultivation of three medicinal mushroom species (*Ganoderma lucidum*, Lentinula edodes and Grifola frondosa) on olive oil press cakes containing substrates and their bioactivity assessment. Ann Agrar Sci 12(3):35–38
- Grotto D, Bueno DCR, Ramos GK, da Costa ASR, Spim SRV, Gerenutti M (2016) Asse ssment of the safety of the shiitake culinary-medicinal mushroom, *Lentinus edodes* (Agaricomycetes), in rats: Biochemical, hematological, and antioxidative parameters. Int J Med Mushroom 18(10):861–870
- Gu Y, Belury MA (2005) Selective induction of apoptosis in murine skin carcinoma cells (CH72) by an ethanol extract of *Lentinula edodes*. Cancer Lett 220:21–28
- Guo Z, Hu Y, Wang D, Ma X, Zhao X, Zhao B, Wang J, Liu P (2009) Sulfated modification can enhance the adjuvanticity of lentinan and improve the immune effect of ND vaccine. Vaccine 27:660–665
- Harvey L, McNeil B, Kristiansen B, Smith JE, Kheng Tan K (2001) Production of lentinan by submerged cultivation of *Lentinus edodes* (Berk.) Sing. Int J Med Mush 3:161
- Hatvani N (2001) Antibacterial effect of the culture fluid of *Lentinula edodes* mycelium grown in submerged liquid culture. J Antibact Antifungal Ag 17(1):71–74
- Hatvani N, Mécs I (2002) Effect of the nutrient composition on dye decolorisation and extracellular enzyme production by *Lentinus edodes* on solid medium. Enzyme Microb Tech 30:381–386
- Hibbett DS (2001) Shiitake mushrooms and molecular clocks: historical biogeography of *Lentinula*. J Biogeog 28:231–241
- Hibbett DS, Donoghue MJ (1996) Implications of phylogenetic studies for conservation of genetic diversity in shiitake mushrooms. Conserv Biol 10:1321–1327
- Hibbett DS, Vilgalys R (1993) Phylogenetic relationships of *Lentinus* (Basidiomycotina) inferred from molecular and morphological characters. Syst Bot 18:409–433
- Higashi Y, Sasaki S, Nakagawa K, Matsuura H, Oshima T, Chayama K (2002) Endothelial function and oxidative stress in renovascular hypertension. N Engl J Med 346:1954–1962

- Hirasawa M, Shouji N, Neta T, Fukushima K, Takada K (1999) Three kinds of antibacterial substances from *Lentinus edodes* (Berk.) Sing. (shiitake, an edible mushroom). Int J Antimicrob Agents 11(2):151–157
- Hiyama R, Harada A, Seiki G, Orihashi K (2016) Ethanol production from unpretreated waste medium of Shiitake mushroom (*Lentinula edodes*) by semi-simultaneous saccharification and fermentation under high substrate concentration conditions. Cellulose Chem Technol 59(7–8):771–780
- Ina K, Kataoka T, Ando T (2013) The use of lentinan for treating gastric cancer. Anti-Cancer Agents in Med Chem 13:681–688
- Ishikawa NK, Kasuya MCM, Vanetti MCD (2001) Antibacterial activity of *Lentinula edodes*. Braz J Microbiol 32(3):206–210
- Itävaara M (1993) Problems associated with the liquid cultivation of shiitake, *Lentinula edodes* (Berk.) Pegler, vol 150. VTT Publications (Technical Research Centre of Finland), pp 11–62
- Izuka C (1990) Antiviral composition extracts from basidiomycetes. European Patent Application EP 464,311. In: CA, 116, 76351z
- Jang Y, Park J, Ryoo R, Park Y, Hyeon Ka K (2016) Ergothioneine contents of shiitake (*Lentinula edodes*) fruiting bodies on sawdust media with different nitrogen sources. Kor J Mycol 44(2):100–102
- Jasinghe VJ, Perera CO (2006) Ultraviolet irradiation: the generator of vitamin D_2 in edible mushrooms. Food Chem 95(4):638–643
- Jeune KH, Moon IJ, Kim MK, Chung SR (1990) Studies on lectins from Korean higher fungi; IV. A mitogenic lectin from the mushroom *Lentinus edodes*. Planta Med 56:592
- Kabir Y, Kimura S (1989) Dietary mushrooms reduce blood pressure in spontaneously hypertensive rats (SHR). J Nutr Sci Vitaminol 35:91–94
- Kamiya T, Saito Y, Hashimoto M, Seki H (1969) Structure and synthesis of lentysine, a new hypocholesterolemic substance. Tetrahedron Lett 10:4729–4732
- Kattoor AJ, Pothineni NVK, Palagiri D, Mehta JL (2017) Oxidative stress in atherosclerosis. Curr Atheroscler Rep 19:42
- Kawai G, Kobayashi H, Fukushima Y, Ohsaki K (1996) Effect of liquid mycelial culture used as a spawn on sawdust cultivation of shiitake (*Lentinula edodes*). Mycoscience 37:201–207
- Kiribuchi T (1991) Effective uses of fungi by UV irradiation. 3. Changes of free amino acid composition in fungi by sun or ultraviolet light irradiation. Nippon Kasei Gakk 42(5):415–21
- Kojima H, Akaki J, Nakajima S (2010) Structural analysis of glycogen–like polysaccharides having macrophage—activating activity in extracts of *Lentinula edodes* mycelia. J Nat Med 64:16–23
- Kulicke WM, Lettau AI, Thielking H (1997) Correlation between immunological activity, molar mass, and molecular structure of different $(1\rightarrow 3)\beta$ –D–glucans. Carbohydr Res 297:135–143
- Kupfahl C, Hof GH (2006) Lentinan has a stimulatory effect on innate and adaptive immunity against murine Listeria monocytogenes infection. Int Immunopharmacol 6:686–696
- Lakhtar H, Ismaili-Alaoui M, Philippoussis A, Perraud-Gaime I, Roussos S (2010) Screening of strains of *Lentinula edodes* grown on model olive mill wastewater in solid and liquid state culture for polyphenol biodegradation. Int Biodeterior Biodegradation 64:167–172
- Leatham GF, Griffin TJ (1984) Adapting liquid spawn *Lentinus edodes* to oak wood. Appl Microbiol Biotechnol 20:360–363
- Lee SJ, KimHH KSH, Kim SH, Sung NJ (2019) The effect of different culture conditions of liquid spawn on the quality characteristics of shiitake mushroom (*Lentinula edodes*). J Mushrooms 17(3):99–106
- Levanon D, Rothschild N, Danai O, Masaphy S (1993) Bulk treatment of substrate for the cultivation of shiitake mushrooms (*Lentinus edodes*) on straw. Bioresource Technol 45(1):63–64
- Li YR, Liu QH, Wang HX, Ng TB (2008) A novel lectin with potent antitumor, mitogenic and HIV– 1 reverse transcriptase inhibitory activities from the edible mushroom Pleurotus citrinopileatus. Biochim Biophys Acta 1780:51–57
- Li S, Wang A, Liu L, Tian G, Wei S, Xu F (2018) Evaluation of nutritional values of shiitake mushroom (*Lentinus edodes*) stipes. J Food Meas Charact 12:2012–2019

- Liu B, Bau YS (1980) Fungi pharmacopoeia. Kinoko, Oakland, CA. A translation of Chinese medicinal fungi, 2nd edn. Shansi People's Publishing House. With added material, pp 302
- Liu W, Gu J, Qi J, Zeng XN, Ji J, Chen ZZ, Sun XL (2015) Lentinan exerts synergistic apoptotic effects with paclitaxel in A549 cells via activating ROS–TXNIP–NLRP3 inflammasome. J Cell Mol Med 19(8):1949–1955
- Markova N, Kussovskia V, Drandarska I, Nikolaeva S, Georgieva N, Radoucheva T (2003) Protective activity of *Lentinan* in experimental tuberculosis. Int Immunopharm 3:1557–1562
- Martínez-Guerrero MA, Sihuanca D, Macías-López A, Pérez-López RI, Martínez-Madrigal JD, López-Olguín JF (2012) Characterization and production of shiitake (*Lentinula edodes*) in Mexico using supplemented sawdust. Afr J Biotechnol 11(46):10582–10588
- Mata JL, Petersen RH (2000) A new species of *Lentinula* (Agaricales) from Central America. Mycoscience 41:351–355
- Mata G, Savoie J-M (1998) Extracellular enzyme activities in six *Lentinula edodes* strains during cultivation in wheat straw. World J Microbiol Biotechnol 14(4):513–519
- Mata JL, Petersen RH, Hughes KW (2001) The genus *Lentinula* in the Americas. Mycologia 93(6):1102–1112
- Mattila P, Salo-Vaananen P, Konko K, Aro H, Jalava T (2002) Basic composition and amino acid contents of mushrooms cultivated in Finland. J Agric Food Chem 50:6419–6422
- Medina E, Paredes C, Pérez-Murcia MD, Bustamante MA, Moral R (2009) Spent mushroom substrates as component of growing media for germination and growth of horticultural plants. Bioresour Technol 100(18):4227–4232
- Medina E, Paredes C, Bustamante MA, Moral R, Moreno-Caselles J (2012) Relationships between soil physic-chemical, chemical and biological properties in a soil amended with spent mushroom substrate. Geoderma 173–174:152–161
- Mekki A, Dhouib A, Sayadi S (2013) Review: effects of olive mill wastewater application on soil properties and plants growth. Int J Recycl Org Waste Agricult 2:15
- Mizoguchi Y, Yamamoto S (1985) Immunotherapy in liver diseases. Rinsyo Menneki 17:452-460
- Mizuno T (1995) Shiitake, *Lentinus edodes*: functional properties for medicinal and food purposes. Food Rev Int 11:7–21
- Molina FI, Shen P, Jong SC, Orikono K (1992) Molecular evidence supports the separation of *Lentinula edodes* from *Lentinus* and related genera. Can J Bot 70:2446–2452
- Moon IJ, Chung SR, Jeune KH (1995) Mitotic Stimulation and cancer cell agglutination of the lectin from *Lentinus edodes*. Yakhak Hoeji 39(3):260–267
- Moonmoon M, Shelly NJ, Md Khan A, Md Uddin N, Hossain K, Tania M, Ahmed S (2011) Effects of different levels of wheat bran, rice bran and maize powder supplementation with sawdust on the production of shiitake mushroom (*Lentinus edodes* (Berk.) Singer). Saudi J Biol Sci 18(4):323–328
- Morais MH, Ramos AC, Matos N, Santos-Olivera EJ (2000) Production of shiitake mushroom (*Lentinula edodes*) on lignocellulosicresidues. Food Sci Tech Int 6:123–128
- Morales D, Gil-Ramirez A, Smiderle FR, Piris AJ, Ruiz-Rodriguez A, Soler-Rivas C (2017) Vitamin D—enriched extracts obtained from shiitake mushrooms (*Lentinula edodes*) by supercritical fluid extraction and UV–irradiation. Innov Food Sci Emerg Technol 41:330–336
- Morales D, Smiderle FR, Piris AJ, Soler-Rivas C, Prodanov M (2018) Production of a β–d–glucan– rich extract from Shiitake mushrooms (*Lentinula edodes*) by an ex-traction/microfiltration/reverse osmosis (nanofiltration) process. Innov Food Sci Emerg Technol 41:330–336
- Moreira MT, Mielgo I, Feijoo G, Lema JM (2000) Evaluation of different fungal strains in the decolourisation of synthetic dyes. Biotechnol Lett 22:1499–1503
- Morinaga H, Tazawa K, Sakamoto K, Katsuyama S (1992) Inhibitive effect of LEM by oral administration of AH–60C and RL1 for liver metastasis. Biotherapy 6:763–765
- Moylan JS, Reid M (2006) Oxidative stress, chronic disease, and muscle wasting. Muscle Nerve 35(4):411–429
- Ng TB, Yap AT (2002) Inhibition of human colon carcinoma development by lentinan from shiitake mushrooms (*Lentinus edodes*). J Altern Complement Med 8(5):581–589

- Nagai M, Sato T, Watanabe H, Saito K, Kawata M, Enei H (2002) Purification and characterization of an extracellular laccase from the edible mushroom *Lentinula edodes*, and decolorization of chemically different dyes. Appl Microbiol Biot 60:327–335
- Ngai PHK, Ng TB (2003) Lentin, a novel and potent antifungal protein from shiitake mushroom with inhibitory effects on activity of human immunodeficiency virus–1 reverse transcriptase and proliferation of leukemia cells. Life Sci 73:3363–3374
- Nguyen AH, Gonzaga MI, Lim VM, Adler MJ, Mitkov MV, Cappel MA (2017) Clinical features of shiitake dermatitis: A systematic review. Int J Dermatol 56:610–616
- Okeke BC, Smith JE, Paterson A, Watson-Craik IA (1993) Aerobic metabolism of pentachlorophenol by spent Sawdust culture of "shiitake" mushroom (*Lentinus edodes*) in soil. Biotechnol Lett 15(10):1077–1080
- Ooi VE, Liu F (2000) Immunomodulation and anti-cancer activity of polysaccharide–protein complexes. Curr Med Chem 7:715–729
- Özçelik E, Pekşen A (2007) Hazelnut husk as a substrate for the cultivation of shiitake mushroom (*Lentinula edodes*). Bioresource Technol 98:2652–2658
- Patni N, Pillai SG, Dwivedi AH (2013) Wheat as a promising substitute of corn for bioethanol production. Procedia Eng 51:355–362
- Pegler DN (1983) The genus Lentinula (Tricholomataceae tribe Collybieae). Sydowia 36:227-239
- Philippoussis AN, Diamantopoulou PA, Zervakis GI (2003) Correlation of the properties of several lignocellulosic substrates to the crop performance of the shiitake mushroom *Lentinula edodes*. World J Microbiol Biotechnol 19:551–557
- Philippoussis A, Diamantopoulou P, Israilides C (2007) Productivity of agricultural residues used for the cultivation of the medicinal fungus *Lentinula edodes*. Int Biodeterior Biodegradation 59:216–219
- Pire DG, Wright JE, Albertó E (2001) Cultivation of shiitake using sawdust from widely available local woods in Argentina. Micol Apl Int 13:87–91
- Razumov IA, Kazachinskaia EI, Puchkova LI, Kosogorova TA, Gorbunova IA, Loktev VB, Tepliakova TV (2013) Protective activity of aqueous extracts from higher mushrooms against Herpes simplex virus type–2 on albino mice model. Antibiot Chemotherapy 58:8–12
- Reverberi M, Zjalic S, Ricelli A, Di Meo C, Scarpari M, Fanelli C, Fabbri AA (2011) Mushroom versus fungi: natural compounds from *Lentinula edodes* inhibit aflatoxin biosynthesis by *Aspergillus parasiticus*
- Ricelli A, Fabbri AA, Trionfetti-Nisini P, Reverberi M, Zjalic S, Fanelli C (2002) Inhibiting effect of different edible and medicinal mushrooms on the growth of two Ochratoxigenic microfungi. Int J Med Mushrooms 4(2):173–179
- Rincão VP, Yamamoto KA, Ricardo NMPS, Soares SA, Meirelles LDP, Nozawa C, Linhares REC (2012) Polysaccharide and extracts from *Lentinula edodes*: structural features and antiviral activity. Virol J 9(37):1–6
- Rokujo T, Kikuchi H, Tensho A, Tsukitani Y, Takenawa T, Yoshida K, Kamiya T (1970) Lentysine: a new hypolipidemic agent from a mushroom. Life Sci 9:379–385
- Rossi IH, Monteiro AC, Machado JO, Andrioli JL, Barbosa JC (2003) Shiitake (*Lentinula edodes*) production on a sterilized bagasse substrate enriched with rice bran and sugarcane molasses. Braz J Microbiol 34(1):66–71
- Royse DJ (1996) Yield stimulation of shiitake by millet supplementation of wood chip substrate. In: Royse DJ (ed) Mushroom biology and mushroom products. University Park, PA, Penn State University Press, pp 277–283
- Royse DJ (1997) Speciality mushrooms and their cultivation. Hortic Rev 19:59-97
- Royse DJ (2001) Cultivation of shiitake on synthetic and natural logs. Cooperative Extension, Pennsylvania State University, University Park, PA, USA, College of Agricultural Sciences, p 12p
- Royse DJ, Sanchez JE (2007) Ground wheat straw as a substitute for portions of oak wood chips used in shiitake (*Lentinula edodes*) substrate formulae. Bioresource Technol 98:2137–2141

- Royse DJ, Baars J, Tan Q (2017) Current overview of mushroom production in the World. In: Diego CZ, Pardo-Gimenez A (eds). Edible and medicinal mushrooms: technology and applications pp 5–13
- Rusan MJM, Albalasmeh AA, Malkawi HI (2016) Treated olive mill wastewater effects on soil properties and plant growth. Water Air Soil Pollut 227:135
- Salmones D, Mata G, Ramos LM, Waliszewski KN (1999) Cultivation of shiitake mushroom, Lentinula edodes, in several lignocellulosic materials originating from subtropics. Agronomie 19(1):13–19
- Samgina DI (1981) Agailkovie gribi I. Agaricales. Fl Spor rast Kazakhst 18:1-268
- Sasaki SH, Linhares REC, Nozawa CM, Moltalván R, Paccola-Meireles LD (2001) Strains of Lentinula edodes suppress growth phytopathogenic fungi and inhibit Alagoas serotype of vesicular stomatitis virus. Braz J Microbiol 32:52–55
- Sasidharan S, Aravindran S, Latha LY, Vijenthi R, Saravanan D, Amutha S (2010) In vitro antioxidant activity and hepatoprotective effects of *Lentinula edodes* against paracetamol–induced hepatotoxicity. Molecules 15:4478–4489
- Savoie JM, Delpech P, Billette C, Mata G (2000) Inoculum adaptation changes the outcome of the competition between *Lentinula edodes* and Trichoderma spp. During shiitake cultivation on pasteurized wheat straw. Mush Sci 15:667–674
- Sharma S, Khanna PK, Kapoor S (2013) Effect of supplementation of wheat bran on the production of shiitake (*Lentinus edodes* (Berk.) Pegler) using wheat straw and saw dust substrates. Bioscan 8(3):817–820
- Sharon TM (1988) Personal observations: Lentinus edodes (shiitake) mycelial extract. Typescript
- Shimada Y, Morita T, Sugiyama K (2003) Eritadenine–induced alterations of plasma lipoprotein lipid concentrations and phosphatidylcholine molecular species profile in rats fed cholesterol–free and cholesterol–enriched diets. Biosci Biotech Bioch 67:996–1006b
- Sugano N, Hibino Y, Choji Y, Maeda H (1982) Anticarcinogenic actions of watersoluble and alcoholinsoluble fractions from culture medium of *Lentinus edodes* mycelia. Cancer Lett 17:109–114
- Sugiyama K, Akachi T, Yamakawa A (1995) Hypocholesterolemic action of eritadenine is mediated by a modification of hepatic phospholipid metabolism in rats. J Nutr 125:2134–2144
- Sugiyama K, Yamakawa A, Kawagishi H, Saeki S (1997) J Nutr 127:593-599
- Sun J, Wang H, Ng TB (2011) Antitumor activities of *O*-sulfonated derivatives of $(1\rightarrow 3)-\alpha-D-$ Glucan from different *Lentinus edodes*. Ind J Biochem Biophys 48:88–94
- Sun M, Zhao W, Xie Q, Zhan Y, Wu B (2015) Lentinan reduces tumor progression by enhancing gencitabine chemotherapy in urothelial bladder cancer. Surg Oncol 24:28–34
- Surenjav U, Zhang L, Xu X, Zhang X, Zeng F (2006) Effects of molecular structure on antitumor activities of β -(1 \rightarrow 3)–D–glucans from different *Lentinan edodes*. Carbohydr Polym 63:97–104
- Suzuki S, Ohshima S (1976) Influence of *L. edodes (Lentinus edodes)* on human serum cholesterol. Mushroom Sci 9(1):463–467
- Suzuki H, Iiyama K, Yoshida O, Yamazaki S, Yamamoto N, Toda S (1990) Structural characterization of the immunoactive and antiviral water–solubilized lignin in an extract of the culture medium of *Lentinus edodes* mycelia (LEM). Agric Biol Chem 54(2):479–487
- Taguchi T et al (1982) Clinical trials on lentinan (polysaccharide). In: Yamamura Y et al (eds) Immunomodulation by microbial products and related synthetic compounds. Elsevier, New York, pp 467–475
- Takashima K, Izumi K, Iwai H, Takeyame S (1973) The hypocholesterolemic action of eritadenine in the rat. Atherosclerosis 17(3):491–502
- Tepwong P, Giri A, Ohshima T (2012) Effect of mycelial morphology on ergothioneine production during liquid fermentation of *Lentinula edodes*. Mycoscience 53:102–112
- Tochikura TS, Nakashima H, Ohashi Y, Yamamoto N (1988) Inhibition (in vitro) of replication and of the cytopathic effect of human immunodeficiency virus by an extract of the culture medium of *Lentinus edodes* mycelia. Med Microbial Immunol 177:235–244
- Tsivileva OM, Nikitina VE, Garibova LV (2005) Effect of culture medium composition on the activity of extracellular lectins of *Lentinus edodes*. Appl Biochem Microbiol 41:174–176

- Tsujiyama S, Muraoka T, Takada N (2013) Biodegradation of 2,4–dichlorophenol by shiitake mushroom (*Lentinula edodes*) using vanillin as an activator. Biotechnol Lett 35:1079–1083
- Turło J, Gutkowska B, Herold F, Krzyczkowski W, Błazewicz A, Kocjan R (2008) Optimizing vitamin B12 biosynthesis by mycelial cultures of Lentinula edodes (Berk.) Pegl. Enzyme Microb Technol 43(4–5):369–374
- Unursaikhan S, Xu X, Zeng F, Zhang L (2006) Antitumor activities of *O*-sulfonated derivatives of $(1\rightarrow 3)-\alpha$ -D-glucan from different *Lentinus edodes*. Biosci Biotechnol Biochem 70(1):38–46
- Vetchinkina EP, Nikitina VE, Tsivileva OM, Garibova LV (2008) Activity of *Lentinus edodes* intracellular lectins at various developmental stages of the fungus. Appl Biochem Microbiol 44(1):66–72
- Vetter J (1995) Mineralstoff and aminosauregehalte des esbaren, kultivilten Pilzes Shii-take (*Lentinus edodes*). Z Lebensm Unters Forsch 201:17–19
- Vetvicka V, Vetvickova J (2015) Glucan supplementation enhances the immune response against an influenza challenge in mice. Ann Transl Med 3(2):22
- Wang HX, Liu WK, Ng TB, Ooi VEC, Chang ST (1996) The immunomodulatory and antitumor activities of lectins from the mushroom *Tricholoma mongolicum*. Immunopharmacol 31:205–211
- Wang HX, Ng TB, Ooi VEC (1999) Studies on purification of a lectin from fruiting bodies of the edible shiitake mushroom *Lentinus edodes*. Int J Biochem Cell Biol 31:595–599
- Wang J, Yu G, Li Y, Shen L, Qian Y, Yang J, Wang F (2015) Inhibitory effects of sulfated lentinan with different degree of sulfation against tobacco mosaic virus (TMV) in tobacco seedlings. Pestic Biochem Physiol 122:38–43
- Wang Y, Jin H, Yu J, Qu C, Wang Q, Yang S, Ma S, Ni J (2020) Quality control and immunological activity of lentinan samples produced in China. Int J Biol Macromol 159:129–136
- Watanabe A, Kobayashi M, Hayashi S, Kodama D, Isoda K, Kondoh M, Kawase M, Tamesada M, Yagi K (2006) Protection against D–galactosamine-induced acute liver injury by oral administration of extracts from *Lentinus edodes* mycelia. Biol Pharm Bull 29:1651–1654
- Watanabe T, Srichuwong S, Arakane M, Tamiya S, Yoshinaga M, Watanabe I, Yamamoto M, Ando A, Tokuyasu K, Nakamura T (2010) Selection of stress-tolerant yeasts for simultaneous saccharification and fermentation (SSF) of very high gravity (VHG) potato mash to ethanol. Bioresour Tech 101:9710–9714
- Xiong S, Martín C, Eilertsen L, Wei M, Myronycheva O, Larsson S, Lestander T, Atterhem L, Jönsson L (2019) Energy-efficient substrate pasteurisation forcombined production of shiitake mushroom (*Lentinula edodes*) and bioethanol. Bioresour Technol 274:65–72
- Xu C, HaiYan Z, Jian Hong Z, Jing G (2008) The pharmacological effect of polysaccharides from Lentinus edodes on the oxidative status and expression of VCAM–1mRNA of thoracic aorta endothelial cell in high–fat–diet rats. Carbohydr Polym 74:445–450
- Yagi K (2012) Liver protective effect of *Lentinula edodes* mycelia (LEM). Gan to Kagaku Ryoho 39:1099–1102
- Yamanaka K (2017) Cultivation of mushrooms in plastic bottles and small bags. In: Zied DC, Pardo-Gimenez A (eds) Edible and medicinal mushrooms technology and applications
- Yang QY, Jong SC (1989) Medicinal mushrooms in China. Mushr Sci 12:631–642
- Yang BK, Kim DH, Jeong SC, Das S, Choi YS, Shin JS, Lee SC, Song CH (2002) Hypoglycemic effect of a *Lentinus edodes* exopolymer produced from a submerged mycelial culture. Biosci Biotechnol Biochem 66:937–942
- Yen MT, Tseng YH, Li RC, Mau JL (2007) Antioxidant properties of fungal chitosan from shiitake stipes. LWT Food Sci Technol 40(2):255–261
- Yoshioka Y, Kojima H, Tamura A, Tsuji K, Tamesada M, Yagi K, Murakami N (2012) Lowmolecular-weight lignin-rich fraction in the extract of cultured *Lentinula edodes* mycelia attenuates carbon tetrachloride-induced toxicity in primary cultures of rat hepatocytes. J Nat Med 66:185–191
- Yu Z, LiHua Y, Qian Y, Yan L (2009) Effect of Lentinus edodes polysaccharide on oxidative stress, immunity activity and oral ulceration of rats stimulated by phenol. Carbohydr Polym 75:115–118

- Zervakis GI, Koutrotsios G (2017) Solid–State Fermentation of plant residues and Agro-industrial wastes for the production of medicinal mushrooms. In: Agrawal et al (eds) Medicinal plants and fungi: recent advances in research and Development, Medicinal and Aromatic Plants of the World, , pp 365–396
- Zhang Y, Ming G, Kaiping W, Zhixiang C, Liquan D, Jingyu L, Fang Z (2010) Structure, chain conformation and antitumor activity of a novel polysaccharide from *Lentinus edodes*. Fitoterapia 81(8):1163–1170
- Zhang N, Chen H, Zhang Yu, Ma L, Xu X (2013) Comparative studies on chemical parameters and antioxidant properties of stipes and caps of shiitake mushroom as affected by different drying methods. J Sci Food Agric 93:3107–3113
- Zhang Y, Liu W, Xu C, Huang W, He P (2017) Characterization and antiproliferative effect of novel acid polysaccharides from the spent substrate of shiitake culinary-medicinal mushroom *Lentinus* edodes (Agaricomycetes) cultivation. Int J Med Mushrooms 19(5):395–403
- Zhang Y, Zhang M, Jiang Y, Li X, He Y, Zeng P, Guo Z, Chang Y, Luo H, Liu Y, Hao C, Wang H, Zhang G, Zhang L (2018) Lentinan as an immunotherapeutic for treating lung cancer: a review of 12 years clinical studies in China. J Cancer Res Clin 144:2177–2186
- Zhao L, Xiao Y, Xiao N (2013) Effect of lentinan combined with docetaxel and cisplatin on the proliferation and apoptosis of BGC823 cells. Tumor Biol 34:1531–1536
- Zheng R, Jie S, Hanchuan D, Moucheng W (2005) Characterization and immunomodulating activities of polysaccharide from Lentinus edodes. Int Immunopharmacol 5:811–820
- Zilly A, Bracht A, Coelho-Moreira JS, Maciel GM, Soares AA, Bazanella GCS, Peralta RM (2011) Enzymes from Ganoderma lucidum: production and applications. Curr Top Biochem Res 13:1–11