

10

A Review and an Update of European LeukemiaNet Recommendations for the Management of Chronic Myeloid Leukemia

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10.1 Introduction

The first version of the European LeukemiaNet (ELN) recommendations for the treatment of chronic myeloid leukemia (CML) was published in 2006 [1], the second and the third ones were published in 2009 and in 2013, respectively [2, 3]. Over this period, the ELN recommendations have provided an internationally shared basis for the treatment and monitoring of CML, contributing to the improvement of the management of CML. Over this period, patients with CML have enjoyed a survival that is nearly identical to the survival of the general population [4, 5], with an acceptable quality of life, due to the high efficacy and the low toxicity of the targeted

treatment. Other ELN recommendations and reviews concerning the management of BCR-ABL kinase point mutations, the side-effects, and the toxicity of tyrosine kinase inhibitors (TKI) were published in 2011 and in 2016, respectively [6, 7]. Now the recommendations have been updated and published [8]. In this chapter we analyze and discuss the evolution of the ELN recommendations over a 15-year period, and we compare the last version with other recent recommendations and guidelines that have been proposed by the European Society of Medical Oncology (ESMO) [9], the Italian Group for Hematologic Diseases of Adults (GIMEMA) [10], the British Society of Haematology (BSH) [11], and the National Comprehensive Cancer Center Network (NCCN) [12].

10.2 The Phases of CML

More than 95% of patients are diagnosed in the chronic phase (CP), 2–3% are diagnosed in the accelerated phase (AP), and 2–3% are diagnosed in the blastic phase (BP) [1–3, 8, 9]. The patients who are diagnosed in AP or in BP require TKI as the patients who are diagnosed in CP, but, in many cases, they require also allogeneic stem cell transplantation (SCT) [8]. In the chemotherapy era, almost all patients were progressing to BP, either directly (blast crisis) or

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through a transient AP. In the TKI era, the rate of progression is much lower (<10%) than in the chemotherapy era [8], but the recognition of the disease phase is still important. It is noticeable that in spite of progress in molecular biology, the boundaries between the three phases are mostly based on the same clinical and hematologic criteria that were selected many years ago [13, 14].

The 2013 version of the ELN recommendations [3] proposed the following definitions: for AP 15–29% blast cells, or blasts plus promyelocytes in blood or marrow >30%, with blasts <30%, or a platelet count <100 × 10⁹/L unrelated to treatment, or another clonal chromosome abnormality (ACA) in Ph + cells (ACA/Ph+); for BP, a blast cell percentage \geq 30% in blood or marrow or blast cell involvement of non-hematopoietic tissues or organs, excluding spleen and liver.

In the latest 2020 ELN version [8], the boundaries between CP and AP are no longer specified. Therefore, one could still rely on the definition of AP given in the 2013 ELN recommendations [3] or rely on another definition, which includes provisional criteria of response to TKI, as proposed in the 2017 WHO classification [15]. This uncertainty reflects doubts on whether the term "AP" should be maintained and used in clinical studies, as it has been for so many years, or if the term "AP" should be removed. As a matter of fact, in the TKI era it is difficult, and it is not very useful to assess the status of the disease based on clinical and hematological findings because the status of the disease can be assessed earlier, based on molecular response. A patient who "fails" (being resistant or intolerant) four TKIs has already entered into a phase of the disease that puts him at a high-risk of dying of leukemia [10], without taking into consideration blood cell counts and differential, particularly without waiting for a progressive increase of blast cells or the development of splenomegaly. In addition, the emergence of high-risk additional chromosome abnormalities in Ph + cells (ACA/Ph+) during TKI treatment is another confirmed signal of progression [16-22], and the finding of other somatic mutations may be also important [23]. For these reasons, the ELN 2020 recommendations use the term "end-phase CML," which comprises "early progression with emerging high-risk ACA and late progression with failing hematopoiesis and blast cell proliferation." BP is a late feature of progression, defined by the ELN only by the blast cells count (\geq 30%) in blood or marrow. In contrast, in the 2017 WHO classification [15], the definition of BP is still based on a proportion of blast cells \geq 20% or on a blast cell involvement of other non-hematopoietic tissues or organs. Importantly, not all patients dying of CML reach the BP-defining blast levels.

10.3 Prognostic Factors

10.3.1 Baseline

Historically, CML was an almost always fatal disease, but it was well recognized that survival could range between few and many years, already in the chemotherapy era. The Sokal score [13], which was proposed as early as 36 years ago, based on the survival of patients treated with conventional chemotherapy, was found to predict response and survival also for patients treated with interferonalpha and for patients treated with TKIs, particularly with imatinib. By the Sokal score, about 40% of patients are classified low-risk, about 40% are classified intermediate risk, and about 20% are classified high-risk. Other prognostic scores have been developed based on patients treated by interferon-alpha (EURO or Hasford score) [24], and in patients treated with imatinib (EUTOS score) [25]. Both EURO and EUTOS scores predict response and survival also in patients treated with imatinib. However, in the TKI era the causes of death are changed: about 50% of the CML patients who die, die in remission and not of leukemia. Therefore, another prognostic score, the Eutos Long-Term Survival Score (ELTS) [26] has been proposed. It has the merit of distinguishing the patients according to the risk of dying of leukemia: by the ELTS score about 60% of patients fall in the low-risk group, with a probability of dying of leukemia of 1–2%, about 30% in the intermediate-risk group, with a probability of dying of leukemia of 5-10%, and only about 10% in the high-risk group, with a probability of dying of leukemia of 10-20%. The last ELN recommendations [8] and the GIMEMA [10] and the BSH [11] recommendations recommend to use, prospectively, the ELTS score. The ELTS score is calculated using age, spleen size,

platelet count, and blast cell percentage in blood as it was the Sokal score. It is noticeable, and a bit unexpected, that after two decades of molecular studies, the prognosis at baseline is still based on clinical and hematologic findings and that splenomegaly is still assessed by manual palpation of the spleen, expressed as the maximum distance below the costal margin.

The calculation of the Sokal and of the ELTS scores is reported in Table 10.1. Both scores provide valuable information of long-term survival, and both are currently used to plan treatment because it is believed that low-risk patients may have a maximum benefit using imatinib in the first line. This belief is supported by solid evidence. It is also believed that high- and also intermediaterisk patients may benefit more of the earlier, first line, use of the "more potent" second-generation TKIs, but this expectation is not supported by solid evidence [8-11]. As a matter of fact, the strategies of treatment of high-risk patients have never been specifically designed and tested, such as the choice of the TKI, the doses, the role of allogeneic SCT, and the degree of the molecular response, optimal, warning or failure at the conventional cornerstones (3, 6, 12 months, and later).

Sokal and ELTS are not the unique prognostic factors that have been identified and proposed. Several reports have highlighted the importance of ACA/Ph + (including trisomy 8, +Ph, isochromosome 17 [i(17q10], trisomy 19, -7/-7q, 11q23 or 3q26.2 aberrations, and complex abnormalities [16–22, 27]), so that ACA/Ph + have been now recognized as baseline high-risk factors irrespective of Sokal and ELTS [8, 10]. Other factors were proposed [28–41], including the low expression of the organic cation transporter (OCT1) that prevents the influx into the cells of imatinib, the high expression and some polymorphisms of the MDR1 (ABCB1) proteins that increase the efflux

of TKI from the cells, a high level of the cancerous inhibitor of PP2A (CIP2A), some polymorphisms or the deletion of BIM more frequently found in Asian countries, the KIR2DS1 genotype associated with resistance to imatinib, a high serum level of tryptase, the fiber content in bone marrow biopsies, and also the immunophenotype showing the simultaneous detection of lymphoid markers in blast cells. Although some of these factors could theoretically be useful to guide the choice beteween imatinib and second-generation TKIs, none of these factors have come into clinical use, and none were recommended so far.

The transcript type of the major BCR-ABL gene may influence to some extent the sensitivity to TKIs, the e13a2 (b2a2) type being less sensitive to TKIs than the e14a2 (b3a2) type and, accordingly, the probability of achieving a deep molecular response and a treatment-free remission [42]. Until now, the BCR-ABL transcript type has not been included as a prognostic parameter in the ELN 2020 recommendations.

Different gene expression profiles (GEP) associated with progression from CP to advanced phases, and with some degree of resistance to imatinib, were reported already years ago [43]. More recently the introduction of new, next-generation, molecular biotechnologies has called attention to the value of additional somatic genomic abnormalities [44], similar to those that have been detected in acute leukemia, in the myelodysplastic syndromes, and also in healthy, elderly people [45]. These studies could pave the way to new targeted therapies.

10.3.2 During TKI Treatment

The response to TKIs, including the time to response and the depth of the response, are more

Table 10.1 The two main risk scoring systems, at diagnosis

Sokal	$Exp 0.0116 \times (age - 43.4) + 0.0345 \times (spleen - 7.51)$
	$+ 0.188 \times ((\text{platelets}/700)^2 - 0.563) + 0.0887 \times (\text{blasts} - 2.10)$
	Low-risk < 0.80. intermediate 0.81–1.20 high > 1.21
	http://www.leukemia-net.org/content/leukemias/cml/cml_score index_eng.Html.
ELTS	$0.0025 \times (age/10)^3 + (0.0615 \times spleen) + (0.1052 \times blasts) + ((0.4104 \times (platelets/1000)^{-0.5}).$
	Low-risk ≤ 1.5680, intermediate risk 1.5680–2.2185, high-risk > 2.2185
	http://www.leukemianet.org/content/leukemias/cml/eutos_score/index_eng.html.

Age in years, spleen in cm below the costal margin; platelets × 1000; blasts in % (peripheral blood)

important than all baseline factors. Cytogenetics is still valuable, but the assessment of the BCR-ABL level by the international standard (IS) is more sensitive and more accurate than the cytogenetic response. The definition of molecular response during treatment, as proposed by the most recent recommendations and guidelines [8, 10-12], is discussed thoroughly in the next section. Still, the definition of the response and its interpretation for guiding the treatment are based on a single value, sometimes on two consecutive tests in case of borderline values. However, the value of the qPCR may be better assessed not by an absolute value, but by the time that is necessary to reach that value [46] and more generally by the dynamics of the decrease of the BCR-ABL transcript level [47]. There is some reluctancy to adopt these dynamic criteria in practice, which is regrettable, because they may help taking several important decisions, concerning both the early and the late switch from one TKI to another, particularly to improve the rate of treatment-free remission.

10.4 Response Definition: The Evolution of Treatment Recommendations

Imatinib (IMA) was approved in the first-line treatment of CML in 2003, and for a short period it was the only TKI that was available [1]. Soon after, due to the development and the approval of second-generation (2G) TKIs, nilotinib (NIL) and dasatinib (DAS), in second line (in the patients resistant to or intolerant of imatinib) [2], as well as in the first-line setting [3], the criteria for the assessment of response definition have been progressively modified, leading to a more flexible definition of the responses, either optimal or failure or suboptimal (warning), at several critical cornerstones. In the case of optimal response, the recommendation is to continue the same TKI at the same dose. In the case of failure, the recommendation is to switch to another TKI. When the response is suboptimal (warning), the recommendation is to consider another TKI, depending on several variables, including the patient's age, health conditions, comorbidities, tolerability, and

also on the goal of the treatment, either survival and quality of life or the achievement of a condition of treatment-free remission (TFR).

The evolving scenario of response definition and treatment recommendations can be better appreciated comparing the ELN recommendations that were published from 2006 (first version) [1] to 2020 (fourth and last version) [8] (Table 10.1). In 2006 and in 2009 the early (3, 6, and 12 months) response was based on hematologic and cytogenetic data [1, 2]. In 2013 the response was based on cytogenetic or on molecular data [3]. In 2020 only the molecular data were considered because molecular tests are more sensitive than cytogenetics [8]. Moreover, they are performed on blood cells, so avoiding a marrow aspirate. However, cytogenetics is still recommended in case of molecular failure because the detection of ACA/Ph highlights the danger of progression.

In the last version of the ELN recommendations [8], at 3 months the response is optimal if BCR-ABL is $\leq 10\%$, it is warning if BCR-ABL is >10% in one test, and it is failure, if BCR-ABL is >10%, and the value is confirmed within 1-3 months; at 6 months it is optimal if BCR-ABL is $\leq 1\%$, it is warning if BCR-ABL is >1-10%, and it is failure if BCR-ABL is >10%; at 12 months the response is optimal if BCR-ABL is $\leq 0.1\%$ (MMR or MR 3.0), it is warning if BCR-ABL is >0.1–1%, and it is failure if BCR-ABL is >1%. The detection of mutations during treatment is always a marker of failure. After 12 months, and later on, if the BCR-ABL transcript level is ≤0.1% (optimal response) ELN 2020 recommends to continue the same TKI, while in the case of BCR-ABL > 0.1-1% (warning) there is a choice, either to change or to continue. If the BCR-ABL transcript level is >1%(failure), ELN recommends changing the TKI. These definitions are the same as in the last but one version of 2013.

In Table 10.2, the last ELN recommendations [8] are compared with the last ESMO (2017) [9] and NCCN (2.2021) guidelines [12], and with the recommendations that were recently proposed by GIMEMA (2019) [10] and by the BSH (2020) [11]. At 3 months, the response is defined as optimal by all recommendations and guidelines if the

 Table 10.2
 ELN definitions

Opum	Opumal response (= conunue with the same 1 K1)	same INI)		-	-
Year	3 months	6 months	12 months	18 months	Later
2006	CHR	PCyR	CCyR	BCR-ABL $\leq 0.1\%$	NS
2009	CHR and Ph + $\leq 65\%$	$Ph + \leq 35\%$	CCyR	BCR-ABL $\leq 0.1\%$	Stable MMR
2013	PCyR	CCyR	BCR-ABL $\leq 0.1\%$	NS	MMR or better
	or	or			
	$BCR-ABL \le 10\%$	BCR-ABL $\leq 1\%$			
2020^{a}	2020^{a} BCR-ABL $\leq 10\%$	BCR-ABL $\leq 1\%$	BCR-ABL $\leq 0.1\%$	NS	BCR-ABL $\leq 0.1\%$
Subop	Suboptimal response or warning (= consider a switch to another TKI)	nsider a switch to another TKI)			
Year	3 months	6 months	12 months	18 months	Later
2006	CHR	Ph+ > 35%	Ph + 1–35%	BCR-ABL > 0.1%	ACA/Ph+, response loss,
					mutations
2009	Ph+ > 95%	Ph + 35-95%	Ph + 1-35%	BCR-ABL >0.1–1%	MMR loss, mutations
2013	Ph + 36–95% or	Ph + 1–35% or	$Ph + \ge 1\%$ or	NS	ACA/Ph+
	BCR-ABL > 10%	BCR-ABL > 1-10%	BCR-ABL > 0.1-1%		
2020	BCR-ABL > 10%	BCR-ABL > 1–10%	BCR-ABL > 0.1–1%	NS	BCR-ABL > 0.1–1%
Failur	Failure (= switch to another TKI)				
Year	3 months	6 months	12 months	18 months	Later
2006	No HR	< CHR or Ph+ $> 95%$	Ph+ > 35%	$Ph + \ge 1\%$	Response loss
2009	< CHR	Ph+ > 95%	Ph+ > 35%	Ph + ≥ 1%	Response loss, mutations, ACA/Ph+
2013	< CHR or Ph+ $> 95%$	Ph+ > 35% or BCR-ABL > 10%	Ph + $\ge 1\%$ or BCR-ABL > 1%	NS	Response loss, ACA/Ph+, mutations
2020	BCR-ABL > 10%, confirmed	BCR-ABL > 10%	BCR-ABL > 1%	NS	BCR-ABL > 1%, HR ACA/ Ph+, resistance mutations
NS not :	NS not specified, HR high-risk, ELN European LeukemiaNet. CHR = complete hematologic response (WBC count <10,000/µl, differential	NS not specified, HR high-risk, ELN European LeukemiaNet. CHR = complete hematologic response (WBC count <10,000/µl, differential without immature granulocytes and	hematologic response (WBC count	<10,000/µl, differential w	ithout immature granulocytes ar

-Cytogenetic response (CyR): Ph+ >95% (no CyR), Ph + 66-95% (minimal CyR), Ph + 36-65% (minor CyR), Ph + 1-35% (Partial CyR), Ph + 0 (Complete CyR). Molecular response (MR): BCR-ABL >10% (no MR), BCR-ABL >1-10% (MR 1.0), BCR-ABL >0.1-1% (MR 2.0), BCR-ABL ≤ 0.1% (MMR or MR 3.0), BCR-ABL ≤ 0.01% (MR 4.0) ^aFor patients aiming at TFR, the optimal response (at any time) is BCR-ABL1 \leq 0.01% (MR4) dindu 1

Inition -) settindes i imitind	Optimal response (= continue with the same TKI)		-	
31	3 months	6 months	12 months	Later
ESMO 2017 Ph	Ph + ≤ 35% or BCR-ABL < 10%	Ph + 0% or BCR-ABL < 1%	BCR-ABL < 0.1%	$BCR-ABL < 0.01\%^{a}$
GIMEMA 2019 BC	BCR-ABL $\leq 10\%$	BCR-ABL $\leq 1\%$	BCR-ABL $\leq 0.1\%$	BCR-ABL $\leq 0.01\%$
BSH 2020 BC	BCR-ABL $\leq 10\%$	BCR-ABL $\leq 1\%$	BCR-ABL $\leq 0.1\%$	BCR-ABL $\leq 0.1\%$
ELN 2020 ^b BC	BCR-ABL $\leq 10\%$	BCR-ABL $\leq 1\%$	BCR-ABL $\leq 0.1\%$	BCR-ABL $\leq 0.1\%$
NCCN 2.2021 BC	$BCR-ABL \le 10\%$	BCR-ABL $\leq 10\%$	$BCR-ABL \le 0.1\%^{a}$ $BCR-ABL > 0.1-1\%^{c}$	SN
uboptimal response or wa	Suboptimal response or warning (= consider a switch to another TKI)	another TKI)	_	
31	3 months	6 months	12 months	Later
ESMO 2017 Ph	Ph + 36–95% or	Ph + 1–65% or	Ph + 0% or	BCR-ABL ≥ 0.1–1%
B(BCR-ABL > 10%	$BCR-ABL \ge 1-10\%$	BCR-ABL $\ge 0.1-1\%$	
GIMEMA 2019 NS		BCR-ABL > 1-10%	BCR-ABL > 0.1–1%	BCR-ABL > 0.01–0.1%
BSH 2020 BC	BCR-ABL > 10%	BCR-ABL > 1-10%	BCR-ABL > 0.1–1%	BCR-ABL 0.1–1%
ELN 2020 BC	BCR-ABL > 10%	BCR-ABL > 1-10%	BCR-ABL > 0.1–1%	BCR-ABL > 0.1%
NCCN 2.2021 BC	BCR-ABL > 10%	NS	BCR-ABL > 1–10%	NS
Failure (= switch to another TKI)	r TKI)			
31	3 months	6 months	12 months	Later
ESMO 2017 <c< td=""><td><chr or<="" td=""><td>Ph+ > 35% or</td><td>$Ph + \ge 1\%$ or</td><td>Relapse, loss of MMR</td></chr></td></c<>	<chr or<="" td=""><td>Ph+ > 35% or</td><td>$Ph + \ge 1\%$ or</td><td>Relapse, loss of MMR</td></chr>	Ph+ > 35% or	$Ph + \ge 1\%$ or	Relapse, loss of MMR
Ph	Ph+ > 95%	BCR-ABL > 10%	BCR-ABL > 1%	
GIMEMA 2019 <c bc<="" td=""><td><pre><chr or<br="">BCR-ABL > 10%, confirmed</chr></pre></td><td>BCR-ABL > 10%</td><td>BCR-ABL > 1%</td><td>BCR-ABL > 0.1%, mutations</td></c>	<pre><chr or<br="">BCR-ABL > 10%, confirmed</chr></pre>	BCR-ABL > 10%	BCR-ABL > 1%	BCR-ABL > 0.1%, mutations
BSH 2020 NS		BCR-ABL > 10%	BCR-ABL > 1%	BCR-ABL > 1%
ELN 2020 BC	BCR-ABL > 10%, confirmed	BCR-ABL > 10%	BCR-ABL > 1%	BCR-ABL > 1%, HR ACA/Ph+, resistance mutations
NCCN 2.2021 NS		BCR-ABL > 10%	BCR-ABL > 10%	NS

Cytogenetic response (CyR): Ph+ >95% (no CyR), Ph + 66–95% (minimal CyR), Ph + 36–65% (Minor CyR), Ph + 1–35% (Partial CyR), Ph + 0 (Complete CyR). Molecular response (MR): BCR-ABL >10% (no MR), BCR-ABL >1–10% (MR 1.0), BCR-ABL >0.1–1% (MR 2.0), BCR-ABL >0.01–0.1% (MR 2.0), BCR-ABL $\leq 0.01\%$ (MR 4.0) alf treatment goal is TFR

^bFor patients aiming at TFR, the optimal response (at any time) is BCR-ABL1 \leq 0.01% (MR4) ^cif treatment goal is long-term survival

BCR-ABL transcript level is $\leq 10\%$, while it is defined as a failure if BCR-ABL is >10% (confirmed in two consecutive tests) by ESMO, GIMEMA, and ELN 2020. No definition of failure at 3 months is given by the BSH and by NCCN 2.2021. At 6 months, if BCR-ABL is $\leq 1\%$ the response is optimal by ESMO, GIMEMA, BSH, and ELN 2020, but not by the NCCN (BCR-ABL 1–10%), and the response is failure by all recommendations and guidelines if BCR-ABL is >10%. At 12 months, if BCR-ABL transcript level is $\leq 0.1\%$ (MMR), the response is optimal in all recommendations, while it is a failure if BCR-ABL is >1%, with the exception of NCCN 2.2021 (> 10%). After the first year of treatment, the response is optimal if BCR-ABL is $\leq 0.1\%$ by ESMO, BSH, and NCCN 2.2021, but only if BCR-ABL is $\leq 0.01\%$ by GIMEMA while the response is failure if BCR-ABL is >1% by ELN 2020, but if it is >0.1% by GIMEMA. The definition of later responses was not specified by ESMO, BSH, and NCCN 2.2021.

In conclusion, all recommendations agree on the definition of failure at 6 months. The major difference is in the level of the transcript at 12 months: a value >1% is a failure, by all the recommendations, European but not by GIMEMA that defines a failure even at lower BCR-ABL transcript level ($\leq 0.1\%$, or MMR), and not by the American guidelines that define failure only if that value is much higher at >10%. These are important differences that cannot easily be explained because although it is almost universally recognized that achieving a major molecular response (MMR) is required for a "normal" survival, there is no consensus on the cut-off value of transcript level, and there is no agreement on the time that may take to achieve the MMR.

Among the European recommendations, an important difference is in the level of the transcript after 1 year of treatment. For ELN 2020, ESMO, and the BHS, the response is optimal if BCR-ABL is $\leq 0.1\%$ (MMR), while for GIMEMA the response is optimal only if BCR-ABL is $\leq 0.01\%$ (MR 4.0). As already noticed, for ELN 2020 and the BSH the response at 12 months is a failure if BCR-ABL is >1%, while for GIMEMA it is already a failure if BCR-ABL is >0.1% (less

than MMR). The reason of these differences is that the ELN 2020 and the BSH recommendations privilege survival, although they highlight that achievement of TFR may be a valid aim of treatment for selected patients, using a shared decision-making policy, while the GIMEMA recommendations privilege always the achievement of a deeper molecular response for treatment-free remission. In any case, the existence of differences at several cornerstones warns that in case of borderline values of the BCR-ABL transcript, a second test should be performed before deciding to continue or to change a TKI.

10.5 Treatment

10.5.1 First-Line

Four TKIs are currently approved as the first-line treatment of newly diagnosed CP CML: imatinib, nilotinib (Tasigna, Novartis Pharma), dasatinib (Sprycel, Bristol-Myers Squibb), and bosutinib (Bosulif, Pfizer). The respective approved doses are 400 mg once daily (OD), 300 mg twice daily (BID), 100 mg OD, and 400 mg OD. The last version of ELN recommendations [8], as well as ESMO, BSH, and NCCN 2021 [9, 11, 12], do not give priority to a TKI over another one as firstline treatment. Dasatinib, nilotinib, and bosutinib have been tested against imatinib in companysponsored randomized trials [48-56]. The results of these trials have provided the basis for approval of these TKIs in the first-line setting. Dasatinib, nilotinib, and bosutinib have never been evaluated formally in comparative clinical trials. Furthermore, comparisons among different trials, either company sponsored or academic, are quite challenging because the patient selection and the endpoints are different and are differently evaluated. The choice of the first-line treatment, anyway, is mainly based on the final endpoint of the treatment and on patient's comorbidities. Imatinib remains the reference drug because most physicians have a long experience with it and because clinically relevant or life-threatening complications have not been reported so far. Therefore, imatinib remains, probably, the safest drug. During the last 20 years imatinib has been studied not only in company-sponsored trials but also in important academic trials [57, 58]. Moreover, imatinib is currently less expensive than dasatinib, nilotinib, and bosutinib. Recently, the brand product Gleevec has been substituted in most countries by generic products, which are further less expensive. By comparison with imatinib 400 mg OD, dasatinib and nilotinib induce faster responses 5-year and deeper but the progression-free survival and the OS were reported to give marginal improvement with respect to imatinib [51, 53]. The same considerations apply to bosutinib vs. imatinib but with a much shorter observation period of 2 years [56]. There is a consensus favoring imatinib in elderly patients, in case of comorbidities, and in case of CML low-risk (the 5-year LRS of low-risk patients is higher than 95%) and favoring nilotinib, dasatinib, or bosutinib in case of high-risk. Moreover, the choice between nilotinib, dasatinib and bosutinib is influenced by comorbidities (cardiovascular risk, lung disease) and cost, which differs from country to country. However, the most important guide to the choice is the goal of treatment. If the goal of treatment is OS, imatinib may be sufficient. If the goal of treatment is a condition of treatment-free remission (TFR), it is likely that more patients will achieve that condition if they are treated first line with a secondgeneration TKI, but this expectation must still be proven.

10.5.2 Second-Line and beyond

In the second-line treatment, imatinib, nilotinib, dasatinib and bosutinib can be used at different doses: up to 400 mg BID for imatinib and nilotinib, up to 140 mg OD for dasatinib, and up to 600 mg OD for bosutinib [3, 59–61]. Finally, ponatinib (Iclusig, Takeda/Incyte) is licensed at a dose of 45 mg OD [62, 63] as second line for patients failing previous TKIs (USA), while in most EU countries the second line use is licensed for patients failing nilotinib or dasatinib first line, or in patients harboring the T315I mutation.

In the second-line treatment, four main scenarios are recognizable. Scenario no. 1 is that of intolerance to first-line treatment (toxicity). In

that case, switching to nilotinib, dasatinib, or bosutinib should be prioritized over ponatinib because it is a situation very similar to first line. Scenario no. 2 is that of failure of first-line treatment (resistance). In that case, the choice of the second-line TKI will be guided by BCR-ABL1 mutations (if a mutation is found), by age, comorbidities, the type of side effects of first-line therapy, physician experience, and TKI availability and cost. Regrettably, there are no trials comparing the five available TKIs in second line. Ponatinib will always be the drug of choice in the case of T315I mutation [62, 63]. Scenario no. 3 is "warning" that of at early milestones (3-6 months), particularly in the case of BCR-ABL1 transcript level > $10\%^{IS}$ at 3 months (absence of early molecular response, EMR). EMR predicts the rate and the depth of late molecular response as well as progression-free survival (PFS) and overall survival, and EMR is achieved more frequently with second-generation TKIs than with imatinib [51, 53]. Not achieving EMR with imatinib suggests considering an early switch to a second-generation TKI; the absence of EMR with a second-generation TKIs in first line is a more worrisome situation, in which strict monitoring is mandatory and switching to ponatinib should be considered. Scenario no. 4 is that of the patient who is an optimal responder but never reaches a deep molecular response, so that he or she becomes a candidate for a late switch to another TKI, looking for treatment discontinuation and TFR [64, 65]. A careful patient selection is required to balance the benefit of a possible future TFR versus the potential new toxicities after switching to another TKI. In summary, in the case of scenarios no. 1 and 2, the TKI must be changed; in the case of scenarios no. 3 and 4, the TKI should or may be changed, and prospective studies are needed to assess the benefit and cost of the change.

An important and mostly uncovered issue is that of dose. All five TKIs were approved in second line at a specific dose: imatinib 400 mg OD to 400 mg BID, nilotinib 400 mg BID, dasatinib 100–140 mg OD, bosutinib 500–600 mg OD, and ponatinib 45 mg OD. Regrettably, there are no robust data with different doses, but there is a general consensus that in many patients all these TKIs are overdosed. Today, nilotinib and dasatinib in second line are mostly used at a dose of 300 mg TD and 100 mg OD, respectively. The dose of bosutinib is likely to be higher than required and unnecessarily toxic. Recent data suggest that in second line a lower starting dose of bosutinib with response-driven dose escalation up to 400 mg OD is effective and well tolerated, at least in elderly patients [66]. Ponatinib is used at 45 mg only in a minority of cases (those bearing a T315I mutation or showing a high level or resistance to previous TKIs); the starting dose of 30 mg OD is preferred in most instances [8]. Ponatinib is currently tested at 30 or 15 mg OD to better balance efficacy versus cardiovascular toxicity [67]. Moreover, once MMR is achieved, ponatinib dose can be reduced to 15 mg OD with careful monitoring of response.

The scenario of third-line treatment is very heterogeneous, including patients who can still be rescued to an optimal response and patients at high-risk of progression and death [68–72]. Third-line treatment has an important impact on survival and may provide an essential bridge to allogeneic stem cell transplantation (allo-SCT). For these reasons, the patients who fail two or more TKIs should be referred to a center specialized in the treatment of CML, and allogeneic stem cell transplantation should be considered [8].

There are no published studies comparing different TKIs in third line. Ponatinib may be the first, or the last choice, because it covers almost all known mutations. Asciminib (Novartis Pharma), a new BCR-ABL allosteric inhibitor, active also against the T315I mutation, is currently in advanced clinical development [73], and a trial of asciminib vs. bosutinib in third line is ongoing; therefore, asciminib may become a valid option in this setting in the near future.

10.6 Treatment Continuation or Discontinuation, Treatment-Free Remission (TFR), Cure

In the 2013 ELN recommendations, continuous treatment with TKIs at the approved dose was referred as the standard of care [3]. Indeed, this approach demonstrated that the life expectancy of newly diagnosed CML patients was similar to

that of the age-matched general population [4, 74]. As a matter of fact, TKIs can induce deep and stable responses with up to a 5-log reduction of the BCR-ABL1 transcript levels. Experimental and clinical data strongly suggest that TKIs cannot eliminate all BCR-ABL1+ stem cells [75-89]. However, there is evidence from several clinical studies [90-100] that 40-60% of the patients who discontinue treatment, having been treated with TKI for at least 3-5 years and being in stable deep MR (MR4 or better for at least 2 years), remain in remission. Monitoring of genomic mesurable residual disease may help to identify true cures or very long lasting TFR [101]. The risk of molecular relapse is higher during the first 6–12 months after discontinuation, then the risk decreases progressively. Whether they will relapse much later is not yet known, but it is known that discontinuation does not increase the risk of progression: almost all patients who have a molecular relapse can regain a molecular remission upon reassumption of the same TKI used prior to discontinuation.

Given the several available studies and the vast experience with TFR gained in the recent years, the ELN 2020 recommendation [8] recognized TFR as a new significant goal of CML management and provided for the first time a set of requirements for TKI discontinuation, distinguishing mandatory, minimal (stop allowed) and optimal (stop recommended for consideration) criteria. However, the best treatment strategy, in first or subsequent lines, to drive patients toward a successful TFR is yet to be defined, and the 2020 ELN recommendations could not provide specific recommendations on that. Indeed, only a few trials are currently ongoing with TFR as a primary objective, including the GIMEMA SUSTRENIM trial and the German CML TIGER study, that is, however, a trial of nilotinib versus a combination of nilotinib and interferon- α . Waiting for the results of these trials, the GIMEMA published in 2019 a set of proposals specifically designed to optimize the treatment strategy for TFR.

The biological mechanisms underlying TFR are not well understood, but TFR probably represents an "operational cure" rather than a "true cure" (disease eradication). Indeed, residual BCR-ABL1+ stem cells are quiescent, no longer BCR-ABL1 addicted and not sensitive to TKIs [76, 80, 81, 83, 85, 88, 89]. Other agents, targeting the stem cells [75, 78], may be necessary to attain CML eradication. Over the last 20 years, many studies have been dedicated to the identification of new targets in the stem cells [102], and many studies have shown that, experimentally, the combination of a TKI with anti-stem-cell agents can eliminate BCR-ABL1+ stem cells [103]. Regrettably, none of these combinations have been tested successfully in vivo, also because there is a legitimate concern of the use of potentially toxic agents in patients with minimal residual disease, with a normal life expectancy and a normal quality of life. For the time being, the search of a cure is theoretical and is addressed toward the immunologic control of minimal residual disease [104–108] which today mainly rely on the addition of IFNs to TKI treatment [109–111].

Certainly, TFR is a clinically relevant endpoint that influences on the well-being and the psychologic behavior of the patients because it limits the long-term toxicity of the treatment and spares the economical resources that are necessary for life-long TKI therapy.

With current treatment approaches it is expected that no more than 20–30% of all newly diagnosed CML patients will be able to achieve a stable TFR. The remaining 70–80% of patients. would need lifelong "standard" dose TKI as per current recommendation and guidelines. For these patients the main objectives are an improvement of quality of life [112] and minimization of long-term toxicities. With this regard, de-escalation of treatment through permanent dose reductions or even intermittent treatment has been investigated and results are promising [113, 114].

10.7 Conclusions

In the last 15 years ELN recommendations contributed to the improvement of the management of CML. Today the survival of CML patients is comparable to that of the general population; the next step is the achievement of a normal survival without continuous treatment. The ELN 2020 recommendations recognized TFR as an important goal of CML therapy and identified a set of requirements for TKI discontinuation. However, it was not possible to recommend specific treatment strategies, both in first or subsequent lines, to increase the proportion of patients reaching TFR. For these reasons, next prospective studies should be designed to consider TFR as the primary endpoint (instead of response rates at a time point). Hopefully, the next version of ELN recommendations will take advantage of the results of such trials to finally propose an evidencebased path to TFR.

References

- Baccarani M, Saglio G, Goldman J, et al. Evolving concepts in the management of chronic myeloid leukemia: recommendations from an expert panel on behalf of the European LeukemiaNet. Blood. 2006;108(6):1809–20.
- Baccarani M, Cortes J, Pane F, et al. Chronic myeloid leukemia: an update of concepts and management recommendations of European LeukemiaNet. J Clin Oncol. 2009;27(35):6041–51.
- Baccarani M, Deininger MW, Rosti G, et al. European LeukemiaNet recommendations for the management of chronic myeloid leukemia: 2013. Blood. 2013;122(6):872–84.
- Bower H, Bjorkholm M, Dickman PW, Hoglund M, Lambert PC, Andersson TM. Life expectancy of patients with chronic myeloid Leukemia approaches the life expectancy of the general population. J Clin Oncol. 2016;34(24):2851–7.
- Hehlmann R, Lauseker M, Saussele S, et al. Assessment of imatinib as first-line treatment of chronic myeloid leukemia: 10-year survival results of the randomized CML study IV and impact of non-CML determinants. Leukemia. 2017;31(11):2398–406.
- Soverini S, Hochhaus A, Nicolini FE, et al. BCR-ABL kinase domain mutation analysis in chronic myeloid leukemia patients treated with tyrosine kinase inhibitors: recommendations from an expert panel on behalf of European LeukemiaNet. Blood. 2011;118(5):1208–15.
- Steegmann JL, Baccarani M, Breccia M, et al. European LeukemiaNet recommendations for the management and avoidance of adverse events of treatment in chronic myeloid leukaemia. Leukemia. 2016;30(8):1648–71.
- 8. Hochhaus A, Baccarani M, Silver RT, et al. European LeukemiaNet 2020 recommendations

for treating chronic myeloid leukemia. Leukemia. 2020;34(4):966–84.

- Hochhaus A, Saussele S, Rosti G, et al. Chronic myeloid leukaemia: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. Ann Oncol. 2017;28(Suppl 4):iv41–51.
- Baccarani M, Abruzzese E, Accurso V, et al. Managing chronic myeloid leukemia for treatmentfree remission: a proposal from the GIMEMA CML WP. Blood Adv. 2019;3(24):4280–90.
- Smith G, Apperley J, Milojkovic D, et al. A British Society for Haematology guideline on the diagnosis and management of chronic myeloid leukaemia. Br J Haematol. 2020;191:171.
- Deininger MW, Shah NP, Altman JK, et al. Chronic myeloid Leukemia, version 2.2021, NCCN Clinical Practice Guidelines in Oncology. J Natl Compr Cancer Netw. 2020;18(10):1385–415.
- Sokal JE, Cox EB, Baccarani M, et al. Prognostic discrimination in "good-risk" chronic granulocytic leukemia. Blood. 1984;63(4):789–99.
- Spiers AS. Metamorphosis of chronic granulocytic leukaemia: diagnosis, classification, and management. Br J Haematol. 1979;41(1):1–7.
- Barbui T, Thiele J, Gisslinger H, Finazzi G, Vannucchi AM, Tefferi A. The 2016 revision of WHO classification of myeloproliferative neoplasms: clinical and molecular advances. Blood Rev. 2016;30(6):453–9.
- Hehlmann R, Voskanyan A, Lauseker M, et al. High-risk additional chromosomal abnormalities at low blast counts herald death by CML. Leukemia. 2020;34(8):2074–86.
- Fabarius A, Leitner A, Hochhaus A, et al. Impact of additional cytogenetic aberrations at diagnosis on prognosis of CML: long-term observation of 1151 patients from the randomized CML study IV. Blood. 2011;118(26):6760–8.
- Luatti S, Castagnetti F, Marzocchi G, et al. Additional chromosomal abnormalities in Philadelphia-positive clone: adverse prognostic influence on frontline imatinib therapy: a GIMEMA Working Party on CML analysis. Blood. 2012;120(4):761–7.
- Fabarius A, Kalmanti L, Dietz CT, et al. Impact of unbalanced minor route versus major route karyotypes at diagnosis on prognosis of CML. Ann Hematol. 2015;94(12):2015–24.
- Wang W, Cortes JE, Lin P, et al. Clinical and prognostic significance of 3q26.2 and other chromosome 3 abnormalities in CML in the era of tyrosine kinase inhibitors. Blood. 2015;126(14):1699–706.
- Wang W, Cortes JE, Tang G, et al. Risk stratification of chromosomal abnormalities in chronic myelogenous leukemia in the era of tyrosine kinase inhibitor therapy. Blood. 2016;127(22):2742–50.
- 22. Alhuraiji A, Kantarjian H, Boddu P, et al. Prognostic significance of additional chromosomal abnormalities at the time of diagnosis in patients with chronic myeloid leukemia treated with frontline tyrosine kinase inhibitors. Am J Hematol. 2018;93(1):84–90.

- Branford S, Wang P, Yeung DT, et al. Integrative genomic analysis reveals cancer-associated mutations at diagnosis of CML in patients with high-risk disease. Blood. 2018;132(9):948–61.
- 24. Hasford J, Pfirrmann M, Hehlmann R, et al. A new prognostic score for survival of patients with chronic myeloid leukemia treated with interferon alfa. Writing Committee for the Collaborative CML Prognostic Factors Project Group. J Natl Cancer Inst. 1998;90(11):850–8.
- 25. Hasford J, Baccarani M, Hoffmann V, et al. Predicting complete cytogenetic response and subsequent progression-free survival in 2060 patients with CML on imatinib treatment: the EUTOS score. Blood. 2011;118(3):686–92.
- Pfirrmann M, Baccarani M, Saussele S, et al. Prognosis of long-term survival considering diseasespecific death in patients with chronic myeloid leukemia. Leukemia. 2016;30(1):48–56.
- Wang W, Cortes JE, Lin P, et al. Impact of trisomy 8 on treatment response and survival of patients with chronic myelogenous leukemia in the era of tyrosine kinase inhibitors. Leukemia. 2015;29(11):2263–6.
- 28. White DL, Saunders VA, Dang P, et al. OCT-1mediated influx is a key determinant of the intracellular uptake of imatinib but not nilotinib (AMN107): reduced OCT-1 activity is the cause of low in vitro sensitivity to imatinib. Blood. 2006;108(2):697–704.
- 29. White DL, Saunders VA, Dang P, et al. Most CML patients who have a suboptimal response to imatinib have low OCT-1 activity: higher doses of imatinib may overcome the negative impact of low OCT-1 activity. Blood. 2007;110(12):4064–72.
- 30. White DL, Dang P, Engler J, et al. Functional activity of the OCT-1 protein is predictive of long-term outcome in patients with chronic-phase chronic myeloid leukemia treated with imatinib. J Clin Oncol. 2010;28(16):2761–7.
- Engler JR, Frede A, Saunders VA, Zannettino AC, Hughes TP, White DL. Chronic myeloid leukemia CD34+ cells have reduced uptake of imatinib due to low OCT-1 activity. Leukemia. 2010;24(4):765–70.
- Watkins DB, Hughes TP, White DL. OCT1 and imatinib transport in CML: is it clinically relevant? Leukemia. 2015;29(10):1960–9.
- 33. Augis V, Airiau K, Josselin M, Turcq B, Mahon FX, Belloc F. A single nucleotide polymorphism in cBIM is associated with a slower achievement of major molecular response in chronic myeloid leukaemia treated with imatinib. PLoS One. 2013;8(11):e78582.
- Thomas J, Wang L, Clark RE, Pirmohamed M. Active transport of imatinib into and out of cells: implications for drug resistance. Blood. 2004;104(12):3739–45.
- 35. Katagiri S, Tauchi T, Saito Y, et al. BCL2L11 (BIM) deletion polymorphism is associated with molecular relapse after ABL tyrosine kinase inhibitor discontinuation in patients with chronic myeloid Leukemia with complete molecular response. Blood. 2014;124(21):1797.

- 36. Sperr WR, Pfeiffer T, Kundi M, Sillaber C, Herndlhofer S, Valent P. Serum Tryptase is a strong predictive biomarker that improves prognostication in Ph+ chronic myeloid Leukemia. Blood. 2012;120(21):2783.
- Mahon FX, Belloc F, Lagarde V, et al. MDR1 gene overexpression confers resistance to imatinib mesylate in leukemia cell line models. Blood. 2003;101(6):2368–73.
- Lucas CM, Harris RJ, Giannoudis A, Copland M, Slupsky JR, Clark RE. Cancerous inhibitor of PP2A (CIP2A) at diagnosis of chronic myeloid leukemia is a critical determinant of disease progression. Blood. 2011;117(24):6660–8.
- 39. Marin D, Gabriel IH, Ahmad S, et al. KIR2DS1 genotype predicts for complete cytogenetic response and survival in newly diagnosed chronic myeloid leukemia patients treated with imatinib. Leukemia. 2012;26(2):296–302.
- 40. El Rassi F, Bergsagel JD, Arellano M, et al. Predicting early blast transformation in chronicphase chronic myeloid leukemia: is immunophenotyping the missing link? Cancer. 2015;121(6):872–5.
- 41. Hidalgo-Lopez JE, Kanagal-Shamanna R, Quesada AE, et al. Bone marrow core biopsy in 508 consecutive patients with chronic myeloid leukemia: assessment of potential value. Cancer. 2018;124(19):3849–55.
- Baccarani M, Castagnetti F, Gugliotta G, et al. The proportion of different BCR-ABL1 transcript types in chronic myeloid leukemia. An international overview. Leukemia. 2019;33(5):1173–83.
- Radich JP, Dai H, Mao M, et al. Gene expression changes associated with progression and response in chronic myeloid leukemia. Proc Natl Acad Sci U S A. 2006;103(8):2794–9.
- 44. Branford S, Kim DDH, Apperley JF, et al. Laying the foundation for genomically-based risk assessment in chronic myeloid leukemia. Leukemia. 2019;33(8):1835–50.
- 45. Ashley EA. Towards precision medicine. Nat Rev Genet. 2016;17(9):507–22.
- 46. Branford S, Yeung DT, Parker WT, et al. Prognosis for patients with CML and >10% BCR-ABL1 after 3 months of imatinib depends on the rate of BCR-ABL1 decline. Blood. 2014;124(4):511–8.
- 47. Hanfstein B, Shlyakhto V, Lauseker M, et al. Velocity of early BCR-ABL transcript elimination as an optimized predictor of outcome in chronic myeloid leukemia (CML) patients in chronic phase on treatment with imatinib. Leukemia. 2014;28(10):1988–92.
- Kantarjian H, Shah NP, Hochhaus A, et al. Dasatinib versus imatinib in newly diagnosed chronicphase chronic myeloid leukemia. N Engl J Med. 2010;362(24):2260–70.
- 49. Kantarjian HM, Hochhaus A, Saglio G, et al. Nilotinib versus imatinib for the treatment of patients with newly diagnosed chronic phase, Philadelphia chromosome-positive, chronic myeloid leukaemia: 24-month minimum follow-up of the

phase 3 randomised ENESTnd trial. Lancet Oncol. 2011;12(9):841–51.

- 50. Kantarjian HM, Shah NP, Cortes JE, et al. Dasatinib or imatinib in newly diagnosed chronic-phase chronic myeloid leukemia: 2-year follow-up from a randomized phase 3 trial (DASISION). Blood. 2012;119(5):1123–9.
- 51. Cortes JE, Saglio G, Kantarjian HM, et al. Final 5-year study results of DASISION: the Dasatinib versus Imatinib study in treatment-naive chronic myeloid Leukemia patients trial. J Clin Oncol. 2016;34(20):2333–40.
- Saglio G, Kim DW, Issaragrisil S, et al. Nilotinib versus imatinib for newly diagnosed chronic myeloid leukemia. N Engl J Med. 2010;362(24):2251–9.
- 53. Hochhaus A, Saglio G, Hughes TP, et al. Long-term benefits and risks of frontline nilotinib vs imatinib for chronic myeloid leukemia in chronic phase: 5-year update of the randomized ENESTnd trial. Leukemia. 2016;30:1044.
- 54. Wang J, Shen ZX, Saglio G, et al. Phase 3 study of nilotinib vs imatinib in Chinese patients with newly diagnosed chronic myeloid leukemia in chronic phase: ENESTchina. Blood. 2015;125(18):2771–8.
- 55. Cortes JE, Kim DW, Kantarjian HM, et al. Bosutinib versus imatinib in newly diagnosed chronic-phase chronic myeloid leukemia: results from the BELA trial. J Clin Oncol. 2012;30(28):3486–92.
- 56. Cortes JE, Gambacorti-Passerini C, Deininger MW, et al. Bosutinib versus Imatinib for newly diagnosed chronic myeloid leukemia: results from the randomized BFORE trial. J Clin Oncol. 2018;36(3):231–7.
- 57. Castagnetti F, Gugliotta G, Breccia M, et al. Long-term outcome of chronic myeloid leukemia patients treated frontline with imatinib. Leukemia. 2015;29(9):1823–31.
- 58. Hehlmann R, Muller MC, Lauseker M, et al. Deep molecular response is reached by the majority of patients treated with imatinib, predicts survival, and is achieved more quickly by optimized high-dose imatinib: results from the randomized CML-study IV. J Clin Oncol. 2014;32(5):415–23.
- 59. Shah NP, Guilhot F, Cortes JE, et al. Long-term outcome with dasatinib after imatinib failure in chronicphase chronic myeloid leukemia: follow-up of a phase 3 study. Blood. 2014;123(15):2317–24.
- 60. Giles FJ, le Coutre PD, Pinilla-Ibarz J, et al. Nilotinib in imatinib-resistant or imatinib-intolerant patients with chronic myeloid leukemia in chronic phase: 48-month follow-up results of a phase II study. Leukemia. 2013;27(1):107–12.
- 61. Kantarjian HM, Cortes JE, Kim DW, et al. Bosutinib safety and management of toxicity in leukemia patients with resistance or intolerance to imatinib and other tyrosine kinase inhibitors. Blood. 2014;123(9):1309–18.
- Cortes JE, Kantarjian H, Shah NP, et al. Ponatinib in refractory Philadelphia chromosome-positive leukemias. N Engl J Med. 2012;367(22):2075–88.

- Cortes JE, Kim DW, Pinilla-Ibarz J, et al. A phase 2 trial of ponatinib in Philadelphia chromosome-positive leukemias. N Engl J Med. 2013;369(19):1783–96.
- 64. Hughes TP, Hochhaus A, Kantarjian HM, et al. Safety and efficacy of switching to nilotinib 400 mg twice daily for patients with chronic myeloid leukemia in chronic phase with suboptimal response or failure on front-line imatinib or nilotinib 300 mg twice daily. Haematologica. 2014;99(7):1204–11.
- 65. Hughes TP, Lipton JH, Spector N, et al. Deep molecular responses achieved in patients with CML-CP who are switched to nilotinib after long-term imatinib. Blood. 2014;124(5):729–36.
- 66. Castagnetti F, Gugliotta G, Bocchia M, et al. Dose optimization in elderly CML patients treated with Bosutinib after intolerance or failure of first-line tyrosine kinase inhibitors. Blood. 2019;134(Supplement_1):496.
- Cortes JE, Lomaia E, Turkina A, et al. Interim analysis (IA) of OPTIC: a dose-ranging study of three ponatinib (PON) starting doses. J Clin Oncol. 2020;38(15_suppl):7502.
- 68. Quintas-Cardama A, Kantarjian H, Jones D, et al. Dasatinib (BMS-354825) is active in Philadelphia chromosome-positive chronic myelogenous leukemia after imatinib and nilotinib (AMN107) therapy failure. Blood. 2007;109(2):497–9.
- Garg RJ, Kantarjian H, O'Brien S, et al. The use of nilotinib or dasatinib after failure to 2 prior tyrosine kinase inhibitors: long-term follow-up. Blood. 2009;114(20):4361–8.
- Khoury HJ, Cortes JE, Kantarjian HM, et al. Bosutinib is active in chronic phase chronic myeloid leukemia after imatinib and dasatinib and/or nilotinib therapy failure. Blood. 2012;119(15):3403–12.
- Russo Rossi A, Breccia M, Abruzzese E, et al. Outcome of 82 chronic myeloid leukemia patients treated with nilotinib or dasatinib after failure of two prior tyrosine kinase inhibitors. Haematologica. 2013;98(3):399–403.
- 72. Lipton JH, Bryden P, Sidhu MK, et al. Comparative efficacy of tyrosine kinase inhibitor treatments in the third-line setting, for chronic-phase chronic myelogenous leukemia after failure of second-generation tyrosine kinase inhibitors. Leuk Res. 2015;39(1):58–64.
- Hughes TP, Mauro MJ, Cortes JE, et al. Asciminib in chronic myeloid leukemia after ABL kinase inhibitor failure. N Engl J Med. 2019;381(24):2315–26.
- 74. Sasaki K, Strom SS, O'Brien S, et al. Relative survival in patients with chronic-phase chronic myeloid leukaemia in the tyrosine-kinase inhibitor era: analysis of patient data from six prospective clinical trials. Lancet Haematol. 2015;2(5):e186–93.
- Elrick LJ, Jorgensen HG, Mountford JC, Holyoake TL. Punish the parent not the progeny. Blood. 2005;105(5):1862–6.
- Kavalerchik E, Goff D, Jamieson CH. Chronic myeloid leukemia stem cells. J Clin Oncol. 2008;26(17):2911–5.

- Chen Y, Peng C, Sullivan C, Li D, Li S. Critical molecular pathways in cancer stem cells of chronic myeloid leukemia. Leukemia. 2010;24(9):1545–54.
- Ross DM, Hughes TP, Melo JV. Do we have to kill the last CML cell? Leukemia. 2011;25(2):193–200.
- Egan JM. Targeting stem cells in chronic myeloid Leukemia with a PPAR-gamma agonist. N Engl J Med. 2015;373(20):1973–5.
- Graham SM, Jorgensen HG, Allan E, et al. Primitive, quiescent, Philadelphia-positive stem cells from patients with chronic myeloid leukemia are insensitive to STI571 in vitro. Blood. 2002;99(1):319–25.
- Copland M, Hamilton A, Elrick LJ, et al. Dasatinib (BMS-354825) targets an earlier progenitor population than imatinib in primary CML but does not eliminate the quiescent fraction. Blood. 2006;107(11):4532–9.
- 82. Jorgensen HG, Copland M, Allan EK, et al. Intermittent exposure of primitive quiescent chronic myeloid leukemia cells to granulocytecolony stimulating factor in vitro promotes their elimination by imatinib mesylate. Clin Cancer Res. 2006;12(2):626–33.
- Jiang X, Zhao Y, Smith C, et al. Chronic myeloid leukemia stem cells possess multiple unique features of resistance to BCR-ABL targeted therapies. Leukemia. 2007;21(5):926–35.
- 84. Jiang X, Forrest D, Nicolini F, et al. Properties of CD34+ CML stem/progenitor cells that correlate with different clinical responses to imatinib mesylate. Blood. 2010;116(12):2112–21.
- 85. Lemoli RM, Salvestrini V, Bianchi E, et al. Molecular and functional analysis of the stem cell compartment of chronic myelogenous leukemia reveals the presence of a CD34- cell population with intrinsic resistance to imatinib. Blood. 2009;114(25):5191–200.
- Chomel JC, Bonnet ML, Sorel N, et al. Leukemic stem cell persistence in chronic myeloid leukemia patients with sustained undetectable molecular residual disease. Blood. 2011;118(13):3657–60.
- Chu S, McDonald T, Lin A, et al. Persistence of leukemia stem cells in chronic myelogenous leukemia patients in prolonged remission with imatinib treatment. Blood. 2011;118(20):5565–72.
- Corbin AS, Agarwal A, Loriaux M, Cortes J, Deininger MW, Druker BJ. Human chronic myeloid leukemia stem cells are insensitive to imatinib despite inhibition of BCR-ABL activity. J Clin Invest. 2011;121(1):396–409.
- Hamilton A, Helgason GV, Schemionek M, et al. Chronic myeloid leukemia stem cells are not dependent on Bcr-Abl kinase activity for their survival. Blood. 2012;119(6):1501–10.
- Rousselot P, Huguet F, Rea D, et al. Imatinib mesylate discontinuation in patients with chronic myelogenous leukemia in complete molecular remission for more than 2 years. Blood. 2007;109(1):58–60.
- Mahon FX, Rea D, Guilhot J, et al. Discontinuation of imatinib in patients with chronic myeloid leukaemia who have maintained complete molecular

remission for at least 2 years: the prospective, multicentre stop imatinib (STIM) trial. Lancet Oncol. 2010;11(11):1029–35.

- Takahashi N, Kyo T, Maeda Y, et al. Discontinuation of imatinib in Japanese patients with chronic myeloid leukemia. Haematologica. 2012;97(6):903–6.
- 93. Tang M, Foo J, Gonen M, Guilhot J, Mahon FX, Michor F. Selection pressure exerted by imatinib therapy leads to disparate outcomes of imatinib discontinuation trials. Haematologica. 2012;97(10):1553–61.
- 94. Mori S, Vagge E, le Coutre P, et al. Age and dPCR can predict relapse in CML patients who discontinued imatinib: the ISAV study. Am J Hematol. 2015;90(10):910–4.
- Etienne G, Guilhot J, Rea D, et al. Long-term follow-up of the French stop imatinib (STIM1) study in patients with chronic myeloid leukemia. J Clin Oncol. 2017;35(3):298–305.
- 96. Saussele S, Richter J, Guilhot J, et al. Discontinuation of tyrosine kinase inhibitor therapy in chronic myeloid leukaemia (EURO-SKI): a prespecified interim analysis of a prospective, multicentre, non-randomised, trial. Lancet Oncol. 2018;19(6):747–57.
- 97. Fava C, Rege-Cambrin G, Dogliotti I, et al. Observational study of chronic myeloid leukemia Italian patients who discontinued tyrosine kinase inhibitors in clinical practice. Haematologica. 2019;104(8):1589–96.
- Ross DM, Masszi T, Gomez Casares MT, et al. Durable treatment-free remission in patients with chronic myeloid leukemia in chronic phase following frontline nilotinib: 96-week update of the ENESTfreedom study. J Cancer Res Clin Oncol. 2018;144(5):945–54.
- 99. Shah NP, Garcia-Gutierrez V, Jimenez-Velasco A, et al. Dasatinib discontinuation in patients with chronic-phase chronic myeloid leukemia and stable deep molecular response: the DASFREE study. Leuk Lymphoma. 2020;61(3):650–9.
- 100. Kimura S, Imagawa J, Murai K, et al. Treatment-free remission after first-line dasatinib discontinuation in patients with chronic myeloid leukaemia (first-line DADI trial): a single-arm, multicentre, phase 2 trial. Lancet Haematol. 2020;7(3):e218–25.
- 101. Machova Polakova K, Zizkova H, Zuna J, et al. Analysis of chronic myeloid leukaemia during deep molecular response by genomic PCR: a traffic light stratification model with impact on treatment-free remission. Leukemia. 2020;34(8):2113–24.
- 102. Houshmand M, Simonetti G, Circosta P, et al. Chronic myeloid leukemia stem cells. Leukemia. 2019;33(7):1543–56.

- 103. Bhatia R. Novel approaches to therapy in CML. Hematology Am Soc Hematol Educ Program. 2017;2017(1):115–20.
- 104. Butt NM, Rojas JM, Wang L, Christmas SE, Abu-Eisha HM, Clark RE. Circulating bcr-ablspecific CD8+ T cells in chronic myeloid leukemia patients and healthy subjects. Haematologica. 2005;90(10):1315–23.
- 105. Quintarelli C, Dotti G, De Angelis B, et al. Cytotoxic T lymphocytes directed to the preferentially expressed antigen of melanoma (PRAME) target chronic myeloid leukemia. Blood. 2008;112(5):1876–85.
- 106. Kreutzman A, Juvonen V, Kairisto V, et al. Mono/ oligoclonal T and NK cells are common in chronic myeloid leukemia patients at diagnosis and expand during dasatinib therapy. Blood. 2010;116(5):772–82.
- 107. Clapp GD, Lepoutre T, El Cheikh R, et al. Implication of the autologous immune system in BCR-ABL transcript variations in chronic myelogenous leukemia patients treated with imatinib. Cancer Res. 2015;75(19):4053–62.
- Hughes A, Yong ASM. Immune effector recovery in chronic myeloid Leukemia and treatment-free remission. Front Immunol. 2017;8:469.
- 109. Hjorth-Hansen H, Stentoft J, Richter J, et al. Safety and efficacy of the combination of pegylated interferon-alpha2b and dasatinib in newly diagnosed chronic-phase chronic myeloid leukemia patients. Leukemia. 2016;30(9):1853–60.
- 110. Nicolini FE, Etienne G, Dubruille V, et al. Nilotinib and peginterferon alfa-2a for newly diagnosed chronic-phase chronic myeloid leukaemia (NiloPeg): a multicentre, non-randomised, open-label phase 2 study. Lancet Haematol. 2015;2(1):e37–46.
- 111. Simonsson B, Gedde-Dahl T, Markevarn B, et al. Combination of pegylated IFN-alpha2b with imatinib increases molecular response rates in patients with low- or intermediate-risk chronic myeloid leukemia. Blood. 2011;118(12):3228–35.
- 112. Efficace F, Cannella L. The value of quality of life assessment in chronic myeloid leukemia patients receiving tyrosine kinase inhibitors. Hematology Am Soc Hematol Educ Program. 2016;2016(1):170–9.
- 113. Clark RE, Polydoros F, Apperley JF, et al. De-escalation of tyrosine kinase inhibitor therapy before complete treatment discontinuation in patients with chronic myeloid leukaemia (DESTINY): a non-randomised, phase 2 trial. Lancet Haematol. 2019;6(7):e375–83.
- 114. Russo D, Martinelli G, Malagola M, et al. Effects and outcome of a policy of intermittent imatinib treatment in elderly patients with chronic myeloid leukemia. Blood. 2013;121(26):5138–44.