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Pathophysiology of Obstructive Sleep Apnea Syndrome in Childhood

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Introduction

Obstructive sleep apnea syndrome (OSAS) refers to a breathing disorder characterized by recurrent, partial, or complete episodes of upper airway obstruction, commonly associated with intermittent hypoxemia and sleep fragmentation [[1,](#page-15-0) [2\]](#page-15-1). OSAS affects individuals of all ages, from neonates to the elderly. However, it is still not known whether OSAS represents a continuum of a disorder that places children at risk for the disorder also as adults [[3](#page-15-2)], or whether OSAS during different stages of life comprises distinct clinical entities [\[4](#page-15-3)–[7\]](#page-15-4).

The structure and the neural control of the upper airway have evolved to serve three important physiological functions: (1) respiration (2) deglutition, and (3) speech. The upper airway is collapsible in order to accommodate these functions. The anatomic factors predisposing to OSAS differ over the lifespan. However, a smaller upper airway is noted in patients with OSAS in all age groups, and probably predisposes to airway collapse during sleep. Anatomic factors such as craniofacial anomalies, obesity, and adenotonsillar hypertrophy contribute to OSAS throughout life, yet a clear anatomic factor cannot always be identifed. This suggests that functional factors altering upper airway neuromotor tone and the biomechanical properties of the upper airway could also play an important role in the etiology of OSAS [\[8](#page-15-5), [9](#page-15-6)]. This chapter focuses on the anatomic, functional, and biomechanical aspects of the upper airway in the pathophysiology of OSAS during childhood as summarized in Table [35.1](#page-1-0).

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Pharyngeal Development

The anatomy of the newborn pharynx is similar to the anatomy of other primates and mammals [[10\]](#page-15-7). The uvula and epiglottis are in close proximity, creating a secure airway that allows for independent suckling and breathing. This anatomic relationship is maintained in other mammals throughout life, with discrete pathways for respiration and deglutition. However, in the human, at about 18 months of development the larynx descends to the level of the ffth cervical vertebrae. This anatomic formation develops because of the additional role of the human pharynx of phonation. For the purpose of deglutition, the pharynx functions as a fexible tube. The pharyngeal muscles, namely, the pharyngeal constrictors and tongue, force food from the oral cavity into the esophagus. For phonation, the pharynx functions as a muscular tube that can change its length and shape to alter the sounds generated by the larynx and passing through the pharynx. For respiration, the pharynx must remain as rigid as possible in order to allow air passage without collapse. However, despite the importance of respiration to sustain life, when one considers the muscles of the upper airway in the human, it becomes apparent that not one of these muscles has a primary function of pharyngeal dilation. It is speculated that the lack of such pharyngeal dilators in humans resulted from the absence of an evolutionary need in mammals and primates, because the anatomic orientation of the structures securing their upper airway is maintained throughout development, whereas in the human it is not.

Pharyngeal Anatomy

The pharynx is generally divided into three anatomic regions (Fig. [35.1](#page-2-0)):

1. *The nasopharynx*, located superior to the level of the soft palate and continuous anteriorly, through the choanae, with the nasal cavities.

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	Infancy	Childhood	Adolescence	Adulthood
Demographics:				
Estimated prevalence	$\overline{?}$	2%	2%	$4 - 9\%$
Peak age (years)	<1	$2 - 8$	$12 - 18$	$30 - 60$
Gender	M > F	$M = F$	γ	M > F
Weight	Normal	Normal, may be underweight or obese	Mostly overweight and Obese obese	
Risk factors	Craniofacial anomalies Prematurity Gastroesophageal reflux Adenotonsillar Hypertrophy	Adenotonsillar hypertrophy Obesity	Obesity Adenotonsillar hypertrophy	Obesity Women-postmenopause
Level of obstruction	Nasopharyngeal Retropalatal	Nasopharyngeal Retropalatal	Nasopharyngeal Retropalatal Retroglossal?	Retropalatal Retroglossal
Anatomic findings				
Airway	Small?	Small	Small	Small
Craniofacial features	May have craniofacial anomalies: Midfacial hypoplasia Micrognathia	Majority normal	$\overline{\mathcal{L}}$	Retrognathia Micrognathia
Soft tissues	May have adenoidal hypertrophy; usually normal tonsils	Adenotonsillar hypertrophy Large soft palate	Adenotonsillar hypertrophy Other soft tissues?	Large lateral pharyngeal walls, tongue, soft palate, parapharyngeal fat pads
Functional findings				
Ventilatory drive: Normal subjects OSAS	High? $\overline{?}$	High Overall normal Some with subtle abnormalities	Moderate $\overline{\mathcal{L}}$	Lower Studies conflicting
Arousability: Normal subjects OSAS	Low Very low	Very low Very low	Moderate $\overline{\cdot}$	High High
Upper airway collapsibility: Normal subjects OSAS	Very low High	Very low High	Low High	Moderate High
Upper airway reflexes during sleep: Normal subjects OSAS	Brisk $\overline{\cdot}$	Active Blunted	High Low	Low Low
Biomechanical findings				
Nasal resistance	$\overline{\cdot}$	Higher in OSAS	$\overline{?}$	Higher in OSAS with obesity
Pharyngeal pressure drop	$\overline{\cdot}$	Higher in OSAS Lower after AT surgery	Higher in OSAS?	Lowered by MMA surgery
Pressure-area slope during tidal breathing	$\overline{?}$	Sedated: more positive in OSAS Awake: more negative in OSAS		
Treatment				
Treatment of choice	Craniofacial surgery CPAP	Adenotonsillectomy	CPAP Weight reduction Adenotonsillectomy	CPAP Weight reduction
Treatment success	High	High	Moderate	Moderate

Table 35.1 Developmental aspects of obstructive sleep apnea syndrome

From Arens et al. [\[7](#page-15-4)]. Reprinted with permission from Oxford University Press

2. *The orop*harynx, located between the level of the soft palate and the larynx, communicating anteriorly with the oral cavity, and having the posterior one-third of the tongue as its anterior border. Based on a midsagittal view (Fig. [35.1a, b\)](#page-2-0), the oropharynx is subdivided into retropalatal (bounded by the level of the hard palate and

the caudal margin of the soft palate) and retroglossal (bounded by the caudal margin of the soft palate to the tip of the epiglottis) regions. In infants and young children, the oropharynx includes mostly the retropalatal region, since the soft palate and the epiglottis are in close proximity. The anterior oropharyngeal wall is

Fig. 35.1 Normal airway. A midsagittal MRI of the head of a child (**a**) and an adult (**b**) are shown. The airway is shown in black. Note the three main anatomical regions of the upper airway in the child: nasopharynx (NP), oropharynx adjacent to the retropalatal region OP_{RP}), and hypopharynx (HP). The adult airway differs from that of the child by having, in addition, an oropharyngeal segment that is retroglossal (OP_{RG}). This anatomic difference is related to the descent of the larynx during the frst 18 months of life. An axial MRI of the head at the retropalatal level

of a child (**c**) and an adult (**d**) are shown. The airway is shown in black. The lateral pharyngeal walls (PW) in the child are formed mainly by the palatine tonsils (T). In the adult, the tonsils are usually absent or minimal in size and the PW is formed by a combination of muscles (see text for details). Ad, adenoid; SP, soft palate. (From Arens et al. [\[7](#page-15-4)]. Reprinted with permission from Oxford University Press)

formed primarily by the tongue and soft palate, while the posterior wall of the oropharynx is formed by the superior, middle, and inferior constrictor muscles [\[11](#page-15-8), [12\]](#page-15-9). The lateral pharyngeal walls (PW) (Fig. [35.1c, d\)](#page-2-0) are formed by several different soft tissues, including muscles (hyoglossus, styloglossus, stylohyoid, stylopharyngeus, palatoglossus, palatopharyngeus, and the lateral aspects of the superior, middle and inferior pharyngeal constrictors [[13](#page-15-10), [14\]](#page-15-11)); lymphoid tissue, primarily the palatine tonsils (noted more in children, Fig. [35.1c](#page-2-0)) [[15\]](#page-15-12); and adipose tissue (lateral parapharyngeal fat pads).

3. *The hypopharynx*, located posterolateral to the larynx, and communicating with the cavity of the larynx through the auditus. This includes the pyriform recesses and the valleculae.

Anatomic Considerations

Anatomic determinants of OSAS in children can be discussed in relation to broad age categories—infants, children, and adolescents.

Infancy

Infants are predisposed to obstructive events and oxygen desaturation during sleep because of high nasal resistance, reduced airway stiffness, and a highly compliant chest wall with reduced functional residual capacity [[7,](#page-15-4) [16](#page-15-13)]. Spontaneous neck fexion can also result in airway obstruction in premature infants [\[17](#page-15-14)]. Nasal occlusion results in a switch to oral breathing only in a minority of infants [\[18](#page-15-15)] and therefore obstruction of the nasal passages from respiratory infection, craniofacial syndromes or choanal stenosis can result in signifcant OSAS. Upper airway obstruction may also occur as a result of airway edema, laryngospasm, and airway edema from gastroesophageal refux disease (GERD). Intrinsic softness of the larynx from laryngomalacia in infants has been demonstrated to be associated with obstructive sleep apnea with improvement after supraglottoplasty [\[19](#page-15-16)]. OSAS in infancy is notable for its association with several important risk factors: (1) craniofacial anomalies, (2) altered soft tissue size, and (3) neurological disorders.

Common Pediatric Disorders Afecting Upper Airway Size and Associated with Obstructive Sleep Apnea Syndrome

I. *Craniofacial Anomalies*: Apert syndrome Crouzon syndrome Pfeiffer syndrome Treacher-Collins syndrome Robin sequence Stickler syndrome Nager syndrome Hallermann–Streiff syndrome Goldenhar syndrome Rubinstein–Taybi Down syndrome Beckwith–Wiedemann

- Achondroplasia Klippel–Feil syndrome Marfan syndrome Choanal stenosis Mucopolysaccharidoses (Hurler, Hunter) II. *Neurological Disorders:* Cerebral palsy Syringobulbia Syringomyelia Myasthenia gravis Möbius syndrome Arnold–Chiari malformation Poliomyelitis III. *Miscellaneous Disorders:* **Obesity** Polycystic ovary syndrome (PCOS) Prader Willi syndrome Melanocortin-4 receptor deficiency Congenital hypothyroidism Sickle cell disease Laryngomalacia Subglottic stenosis Airway papillomatosis Face and neck burns Gastroesophageal refux IV. *Postoperative Disorders*: Post-adenotonsillectomy leading to naso- and or
	- oropharyngeal stenosis Post-pharyngeal fap: leading to naso- and/or oropharyngeal stenosis

From Arens et al. [\[7](#page-15-4)]. Reprinted with permission from Oxford University Press.

Craniofacial Anomalies

The relationship between craniofacial structure and OSAS is most compelling in infants with distinct craniofacial anomalies seen with craniofacial synostosis, such as Crouzon, Pfeiffer, and Apert syndromes [\[20](#page-15-17), [21](#page-15-18)]; and with mandibulofacial dysostoses, such as Robin sequence [[22–](#page-15-19)[25\]](#page-15-20) and Treacher-Collins syndrome [[26\]](#page-15-21). Altered facial skeletal development, especially the association of maxillary and/or mandibular hypoplasia, may lead to airway narrowing due to crowding of adenoid, tonsils, and other soft tissues within the mid and lower face skeletal boundaries. Decreased neuromotor tone may further reduce airway size by inducing glossoptosis and hypopharyngeal collapse during sleep. Children with craniofacial anomalies may present with OSAS soon after birth and during the frst years of life. In some cases, OSAS does not occur until the child is older and develops adenotonsillar hypertrophy in conjunction with the narrow upper airway. Some craniofacial syndromes, such as Down syndrome, are also associated with hypotonia, which can contribute to upper airway obstruction. Children with associated central nervous system abnormalities may also have central hypoventilation.

Down syndrome is the most common genetic disorder associated with craniofacial anomalies. OSAS is present in 30–60% of these patients [\[27](#page-15-22)[–30](#page-15-23)]. Anatomic factors related to the Down syndrome phenotype, including midfacial and mandibular hypoplasia, glossoptosis, adenoid and tonsillar hypertrophy, laryngotracheal anomalies, and obesity, are the most common causes for OSAS in this group [[31,](#page-15-24) [32](#page-15-25)]. Reduction in neuromuscular tone may also play a role in the development of sleep-disordered breathing in these children.

Altered Soft Tissue Size

The size of the upper airway soft tissues (tonsils, adenoid, fat pads, and musculature) are determined by genetic factors. In addition, the size of these tissues may be affected by infammation, infection, and infltration by various metabolic or storage components. Finally, abnormal neuromotor tone may further alter the shape of upper airway musculature, predisposing to airway narrowing and collapse during sleep.

Infammatory changes leading to adenotonsillar hypertrophy are seen in some infants prior to 1 year of age, leading to the full clinical spectrum of OSAS [\[33](#page-16-0), [34\]](#page-16-1). Macroglossia can signifcantly reduce upper airway size. It commonly occurs in infants and children with Down syndrome, as well as in infants and children with various storage and metabolic disorders, such as mucopolysaccharidosis [\[35](#page-16-2)] and Beckwith–Wiedemann syndrome [[36\]](#page-16-3). In patients with glossoptosis, the tongue may prolapse posteriorly and occlude the airway. Glossoptosis is commonly seen in patients with a small and retroposed mandible as in the Robin sequence [\[22](#page-15-19)[–25](#page-15-20)], or in conditions associated with poor upper airway muscle tone such as Down syndrome [\[27](#page-15-22)[–32](#page-15-25)]. Anomalies of the soft palate, such as cleft palate and velopharyngeal insuffciency, are not usually associated with OSAS. However, the surgical repair of these malformations by palatoplasty and pharyngeal fap, respectively, are associated at times with a moderate degree of OSAS [[37,](#page-16-4) [38\]](#page-16-5).

Neurological Disorders

Various central nervous system disorders have been associated with OSAS in young infants. All induce pharyngeal hypotonia and predispose to sleep-disordered breathing and airway obstruction. Common causes include cerebral palsy, increased intracranial pressure, brain stem compression/dysplasia such as Arnold Chiari malformations, recurrent laryngeal nerve palsy, palsies of the cranial nerves, and syrinx [\[39](#page-16-6)[–42](#page-16-7)].

Childhood

In preschool children, the incidence of OSAS is estimated to be 2% [\[43](#page-16-8), [44\]](#page-16-9), whereas primary snoring is more common and is estimated as 6–9% in school-aged children [\[45](#page-16-10)]. Although the exact mechanism for OSAS in children is not fully understood, important anatomic risk factors have been identifed, and are linked to the anatomical structures surrounding the airway that affect airway size, shape, dynamics, and mechanics.

Anatomical Assessment of the Upper Airway

Physical examination of the upper airway is important and should be performed in each child as part of the general assessment. However, in order to more thoroughly evaluate the airway, endoscopy [\[46](#page-16-11)] and imaging techniques such as lateral neck radiographs, cephalometrics, fuoroscopy, acoustic refection, computerized tomography, and MRI are helpful [[47–](#page-16-12)[52\]](#page-16-13). The above modalities have all demonstrated that the upper airway of children with OSAS is smaller on average than that of the normal child.

MRI is a particular powerful tool because (1) it provides excellent upper airway and soft tissue resolution; (2) it provides accurate, reproducible quantifcation of the upper airway, and surrounding soft tissue structure; (3) imaging can be performed in the axial, sagittal, and coronal planes; (4) volumetric data analysis including three-dimensional reconstructions of upper airway soft tissue and craniofacial structures can be performed [\[15](#page-15-12), [47](#page-16-12), [53](#page-16-14)[–55](#page-16-15)]; (5) dynamic images provide four-dimensional data of the size and shape of the airway during breathing [[56,](#page-16-16) [57](#page-16-17)]; and (6) it does not expose subjects to ionized radiation. On the other hand, several limitations should be noted: (1) young children need to be sedated to avoid motion artifact; (2) studies in sleep are limited in the MRI environment because of noise, arousals, and movement artifact; and (3) MRI is expensive and not always available.

Airway Size

Using MRI, Arens et al. [[47\]](#page-16-12) studied the upper airway in 18 children with moderate OSAS (age 4.8 ± 2.1 years) with an apnea/hypopnea index of 11.2 ± 6.8 and compared these fndings to 18 matched controls. MRI was performed under sedation with intravenous pentobarbital, and axial and sagittal T1- and T2-weighted sequences were obtained (Fig. [35.2a,](#page-5-0) [b](#page-5-0)). The volume of the upper airway was smaller in subjects with OSAS in comparison to controls $(1.5 \pm 0.8 \text{ cm}^3 \text{ vs.})$ 2.5 ± 1.2 cm³, $p < 0.005$). This finding was later reproduced by other investigators [\[58](#page-16-18), [59](#page-16-19)] using similar techniques.

Region of Vulnerability and Overlap Region

In order to determine the anatomic region of maximal narrowing in children with OSAS, Isono et al. performed upper

Fig. 35.2 Mid-sagittal image of a non-obese young child with OSAS (**a**). Black arrow points at narrow nasopharynx and overlap region between adenoid (Ad), soft palate (SP), and tonsils (T). Axial image of the same subject (**b**); white arrow points to a narrowed oropharyngeal airway by two tonsils (T). Mid-sagittal image of an obese adolescent

with OSAS (**c**). Note complete occlusion of nasopharynx in the overlap region by large adenoid (Ad), soft-palate and tonsils (T). Axial image of the same subject (**d**); white arrow points to a very small oropharyngeal airway between both tonsils (T)

airway endoscopy under general anesthesia, evaluating discrete levels of the upper airway including the adenoid, soft palate, tonsil, and tongue [[46\]](#page-16-11). The minimum cross-sectional area was found to be at the level of the adenoid and the soft palate. These fndings, along with high closing pressures noted at these points in the same study, suggest that the superior upper airway segments are most involved in children

with OSAS. These fndings are supported by two recent studies evaluating upper airway shape with MRI. Arens et al. [[53\]](#page-16-14) showed that airway narrowing in children with OSAS occurred along the upper two thirds of the airway, and was maximal in the region where the adenoid overlap the tonsils and soft palate—"the overlap region" [\[53](#page-16-14)] (Fig. [35.3](#page-6-0)). Similar fndings were noted by Fregosi et al. [\[59](#page-16-19)], who **
T ** *** *

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Area (mm2)

Area (mm²)

**

**

Control Adenoid Extent

OSA Adenoid Extent

Overlap Region

nasopharynx **Airway Length (%)**

Fig. 35.3 Airway cross-sectional area from choana to epiglottis and the "overlap region." (**a**) Airway length versus cross sectional area in 20 control children (open circles) and 20 children with OSAS (closed circles). Data points are means \pm SD. Horizontal bars show the regions of the adenoid and tonsils adjacent to the airway. Gray = controls, black =

OSAS. **p* < 0.5, ***p* < 0.005; ****p* < 0.0005. Note that the *overlap region* of the adenoid and tonsils in both groups corresponds to the minimal airway cross sectional area. Modifed from reference [\[53\]](#page-16-14). (**b**) 3D reconstruction of the airway and overlap region where the adenoid and tonsils overlap the soft palate in Control and OSAS

OSAS

described maximal narrowing in the retropalatal region where the soft palate, adenoid, and tonsils overlap.

Airway Dynamics Depicted by MRI

Arens et al. used respiratory-gated MRI to demonstrate the dynamics of the upper airway during tidal breathing in sedated children with OSAS [[56\]](#page-16-16). They showed that the maximum restriction in patients with OSAS occurred in mid-inspiration (Fig. [35.4](#page-7-0)), and that dynamic fuctuations in the airway overlap region were sixfold higher than in controls. They have speculated that such changes may have been induced by one of the following: altered upper airway motor tone, increased airway compliance, or excessive inspiratory driving pressures caused by proximal airway narrowing.

The above study demonstrated different size and shape confguration of the airway in children with OSAS in both inspiration and expiration as compared with control subjects. Subjects with OSAS exhibited an airway shape narrowed across the A–P axis. This could be caused by anatomic features infuencing the width of the lateral pharyngeal wall and/or by neuromotor factors affecting upper airway dilator muscle activity along this axis (i.e., genioglossal activation).

These differences, together with the magnitude of area changes during tidal breathing, may contribute to a more collapsible airway in children with OSAS during sleep, as suggested by functional studies [[46,](#page-16-11) [50,](#page-16-20) [60\]](#page-16-21).

Soft Tissues

Adenoid and Tonsils In normal children without OSAS, the soft tissues, particularly the tonsils and adenoid, grow commensurate with age maintaining a constant proportionality with the pharyngeal airway [\[7](#page-15-4), [15\]](#page-15-12). It has been speculated that disproportional overgrowth of the adenoid and tonsils in children with OSAS results from infammation and/or infections but the mechanisms leading to this process have not been elucidated [[15\]](#page-15-12).

Arens et al. measured the size of the adenoid and tonsils in children with OSAS compared to controls [\[47](#page-16-12)]. They found that both were signifcantly larger in the OSAS group; 9.9 ± 3.9 cm³ and 9.1 ± 2.9 cm³ versus 6.4 ± 2.3 cm³ and 5.8 \pm 2.2 cm³ (p < 0.005; p < 0.0005, respectively). In addition, the combined size of the adenoid and tonsils correlated significantly with the apnea/hypopnea index $(p = 0.03,$ $r = 0.51$, suggesting that volumetric measurements of these

Fig. 35.4 Airway dynamics – cross-sectional area during tidal breathing in Control and OSAS. Dynamic changes in cross-sectional area at mid-tonsillar level during tidal breathing (TV), 5-vol increments of

inspiration (Ins), 5-vol increments of expiration (Exp) in a control child (*top panels*) and a child with OSAS (*bottom panels*)

tissues may be useful in predicting the severity of obstructive sleep apnea in children.

In most cases, large tonsils and/or adenoid can explain the clinical symptoms of children with OSAS, and surgical removal of these tissues cures or ameliorates the disorder in the majority of cases [\[33](#page-16-0), [61](#page-16-22)[–63](#page-16-23)]. However, it is estimated that in 10–15% of otherwise normal children with OSAS, this disorder is not resolved by the simple removal of the tonsils and adenoid [\[64](#page-16-24)[–66](#page-16-25)].

Although the importance of adenoidal and tonsillar hypertrophy in the pathogenesis of childhood OSAS is unquestioned, much remains to be learned. It is possible that the three-dimensional orientation of these tissues, and how they overlap in the airway, is a more important factor, and may significantly affect flow resistance during sleep [[47,](#page-16-12) [53,](#page-16-14) [59\]](#page-16-19).

Tongue Size The tongue is one of the largest structures defning the oropharyngeal airway and bounds its anterior aspect. It is composed of extrinsic muscles (genioglossus, hyoglossus, and styloglossus), which alter its position; and intrinsic muscles, which alter its shape; both of which can affect airway size and shape. Arens et al. found that the overall volume of the tongue in nonsyndromic children with OSAS did not differ from controls [\[47](#page-16-12)].

Soft Palate There are few data on the dimensions of the soft palate in children with OSAS. Using direct measurements, Brodsky et al. [[67\]](#page-16-26) did not fnd a correlation between soft palate length and severity of tonsillar hypertrophy in children with OSAS. Using MRI, Arens et al. [[47\]](#page-16-12) noted a 30% increase in the volume of the soft palate of children with mild to moderate OSAS compared to controls. They speculated that the larger palatal volume might have been due to edema and infammatory changes secondary to chronic snoring, as described in adults [[68–](#page-16-27)[70\]](#page-16-28).

Craniofacial Structure

Several studies using cephalometrics support the idea that children without distinct craniofacial anomalies have subtle craniofacial morphometric features associated with OSAS [[49,](#page-16-29) [71](#page-16-30)[–74\]](#page-17-0). Kawashima et al. [\[75](#page-17-1)] reported that children with OSAS and more pronounced tonsillar hypertrophy had retrognathic mandibles and increased posterior facial height compared to children with OSAS and less pronounced tonsillar hypertrophy. Shintani et al. [[71\]](#page-16-30) noted that the relationship of the mandible with respect to the cranial base was retrognathic in children with OSAS compared to normal children. Zucconi et al. [[76](#page-17-2)] noted that children with OSAS had increased craniomandibular, intermaxillary, goniac, and mandibular plane angles, indicating a hyperdivergent growth pattern (angle between nasionsella line and mandibular line >38°).

In contrast to the above, other investigators suggested that the craniofacial changes found in children with OSAS are mild, and are reversible following adenotonsillectomy [[74,](#page-17-0) [77](#page-17-3), [78\]](#page-17-4). This is supported by a study evaluating upper airway structure using MRI, showing no signifcant differences in

the size of the mandible and maxilla of children with OSAS versus controls [\[47](#page-16-12)]. Furthermore, in a more comprehensive evaluation of the mandible after three-dimensional reconstruction, the above authors found no difference in eight dimensions of the mandible between children with OSAS and controls, suggesting that mandibular size and shape does not play a signifcant role in the causation of childhood OSAS in nonsyndromic children [[79\]](#page-17-5).

Childhood Obesity

Earlier descriptions of childhood OSAS characterized children as being of normal weight, and failure to thrive was a common complication [33 m [80](#page-17-6)]. However, the dramatic increase in pediatric obesity [\[81](#page-17-7), [82](#page-17-8)] is not refected in most of literature characterizing risk for OSAS from early infancy to late childhood [[44,](#page-16-9) [82–](#page-17-8)[88\]](#page-17-9) although a large epidemiological study involving 399 children between 2 and 18 years of age found that obesity was the most signifcant risk factor for OSAS, with an odds ratio of 4.5 [\[44](#page-16-9)]. The prevalence of OSAS was reported to be 46% by Marcus et al. in unselected obese children undergoing polysomnography [[86\]](#page-17-10); Silvestri et al. reported a prevalence of 59% in obese children referred for evaluation of sleep disordered breathing [\[85](#page-17-11)]; and Kalra et al. reported a prevalence of 55% in morbidly obese children undergoing bariatric surgery [\[89](#page-17-12)]. The reason for such a high prevalence of OSAS in obese children compared to the 2% reported in the general pediatric population [\[44](#page-16-9)] is unknown. However, it may be related to a different underlying pathophysiology of the disorder distinguishing it from OSAS in nonobese children, and/or an augmented effect on regular causative factors, resulting from their obesity accelerating growth of upper airway lymphoid tissues [[54\]](#page-16-31).

In nonobese children with OSAS the most common treatment is adenotonsillectomy [\[90](#page-17-13)]. Adenotonsillectomy cures or ameliorates the disorder in the majority of cases [[33,](#page-16-0) [61–](#page-16-22) [63](#page-16-23)]. However, as noted earlier, it is estimated that in 10–20% of otherwise normal children, signifcant residual symptoms exist after surgery [\[64](#page-16-24)[–66](#page-16-25), [91](#page-17-14)]. Similarly, several investigators emphasize the role of adenoid and tonsillar hypertrophy in obese children with OSAS [\[86](#page-17-10), [92–](#page-17-15)[94\]](#page-17-16). A recent study suggests that 45% of morbidly obese children and adolescents with OSAS have evidence of adenotonsillar hypertrophy [\[95](#page-17-17)]. However, after adenotonsillectomy in obese children with OSAS, residual OSAS is noted in up to 50% of children [\[96](#page-17-18)]. This fnding suggests that other anatomical and/or functional factors play a signifcant role in the pathophysiology of OSAS in this group.

Obese children may have excess deposition of adipose tissue within the muscles and tissues surrounding the airway, limiting airway size and increasing airway resistance as observed in adults [\[97](#page-17-19)]. Additional factors that may predispose obese children to OSAS include altered chest wall mechanics and reduced lung volumes due to altered body composition [\[54](#page-16-31), [98](#page-17-20)], resulting in decreased oxygen reserves and decreased central ventilatory drive [\[83](#page-17-21), [99](#page-17-22)]. However, the exact effects of weight gain or weight loss on upper airway structure and function have not been studied in obese children. Moreover, as mentioned above, other mechanisms affecting upper airway neuromotor tone and increasing upper airway collapsibility could have a compound effect in these children with an anatomically compromised airway.

Adolescence

There are few data related to the epidemiology of OSAS in adolescence. Only one study assessed the prevalence of the disorder in this age group and estimated it at 1.9% [\[100](#page-17-23)]. It is not known whether OSAS appearing in adolescence is an extension of the clinical disorder of childhood, with adenotonsillar hypertrophy as a major risk factor, or whether it represents an early manifestation of the adult form of OSAS, with obesity as a major risk factor.

Several studies have addressed the relationship between childhood OSAS and OSAS during adolescence. In a retrospective study, Morton et al. [[101\]](#page-17-24) found that sleep-disordered breathing in adolescence was more common in those who had undergone adenotonsillectomy during early childhood. Tasker et al. [[102\]](#page-17-25) noted a significant increase in inspiratory effort and snoring during sleep in adolescents 12 years after adenotonsillectomy, compared to controls. The latter authors speculated that airway narrowing could have originated in childhood and predisposed to OSAS during adolescence. Guilleminault and colleagues noted alterations in craniofacial morphology in three adolescents with OSAS and a history of upper airway obstruction in childhood. They hypothesized that both genetic factors altering craniofacial growth, and secondary modifcation of craniofacial growth secondary to adenotonsillar hypertrophy, predisposed these patients to OSAS [[103,](#page-17-26) [104\]](#page-17-27).

Another possibility is that OSAS during adolescence represents an early manifestation of the adult form of OSAS, especially when associated with obesity. It is well established that the antecedents of adult obesity begin during childhood and adolescence [\[81](#page-17-7)]. Childhood obesity in all age groups is currently on the rise, and the highest prevalence (15.5%) is seen in adolescent children between 12 and 19 years of age [\[105](#page-17-28)] also having all components of the metabolic syndrome [\[106](#page-17-29)[–109](#page-17-30)]. Studies investigating the pathophysiology of OSA in this population have shown that upper airway lymphoid hypertrophy restricting airway size continue to be an important contributor to OSAS [[54,](#page-16-31) [110\]](#page-17-31) (Fig. [35.2c, d](#page-5-0)) though functional factors impacting airway collapsibility may also play an important role [[111\]](#page-18-0).

Polycystic ovary syndrome (PCOS) is one of the most common endocrine disorders associated with overweight

and obesity. It affects 5–10% of adolescent girls and women of reproductive age [\[112](#page-18-1)]. PCOS usually presents during adolescence with irregular menstruation and clinical signs of hyperandrogenism and is associated with obesity and cardiometabolic abnormalities [\[113](#page-18-2)]. In recent years it has been shown that adolescent girls [\[114](#page-18-3)] and adult women with the disorder have a signifcantly higher prevalence of OSAS compared to women without the disorder [\[115](#page-18-4)[–117](#page-18-5)], screening for OSAS has therefore been recommended in these subjects [\[118](#page-18-6)].

Summary of Anatomic Considerations

Various anatomic mechanisms may lead to OSAS in children. However, a smaller upper airway is noted in all age groups and probably predisposes to airway narrowing and collapse during sleep. OSAS is uncommon in infancy. However, children born with craniofacial anomalies are at increased risk for the development of a severe form of the disorder.

The most common type of childhood OSAS occurs in children between 2 and 8 years of age and is associated with adenotonsillar hypertrophy in most cases. Surgical removal of the adenoid and tonsils ameliorates the disorder in most but not all children, suggesting that other mechanisms such as those leading to altered upper airway neuromotor tone during sleep may contribute to OSAS in these children.

Recent data suggest that obesity may be a leading cause for OSAS during adolescent years. This form of OSAS probably shares much with the adult form of OSAS and particularly with the metabolic consequences of the disorder. However, in contrast to the adult form of OSAS, adenotonsillar hypertrophy commonly observed in early childhood plays an important anatomical contributor in this age group as well.

Functional Considerations

There are several arguments that suggest that functional attributes have an important role in limiting and infuencing OSAS in children. The frst is that the upper airway in children is smaller compared to adults. Since the prevalence of OSAS is much lower in children, it is probable that children have nonanatomical attributes that enhance airway stability during sleep. The second is that airway obstruction occurs during sleep and not during wakefulness, suggesting that neuromotor activation keeps the airway open during wakefulness but not during sleep, when activation is diminished. The third is that subjects with OSAS survive each night. Therefore, there must be overriding mechanisms that prevent unremitting airway obstruction and anoxia from leading to death.

Central Ventilatory Drive

The role of ventilatory drive in the pathophysiology of OSAS in adults and children with OSAS is not fully understood. The central ventilatory drive changes with age from infancy to adulthood. Methodological limitations in measuring drive and mechanical and anatomical differences across the age spectrum do not allow precise comparisons of ventilatory drive throughout the life span. However, it appears that ventilatory drive gradually declines from childhood to old age, possibly because of declining basal metabolic rate with age.

Some adults with OSAS have been reported to have increased ventilatory responses to hypercapnia and hypoxia leading to a high-gain ventilatory control system that could predispose some individuals to irregular or periodic breathing, ventilatory instability, and apnea [[119,](#page-18-7) [120](#page-18-8)]. This relationship is not frmly established; normal or even reduced ventilatory responses have also been reported in adults [\[121](#page-18-9)].

In comparison, nonobese children with OSAS were shown to have normal ventilatory responses to hypercapnia and hypoxia during wake and sleep [\[7](#page-15-4), [122](#page-18-10), [123\]](#page-18-11), and other studies in obese children showed a blunted ventilatory response to hypercapnia in sleep. Such studies suggest that OSAS in children is unlikely to be initiated through ventilatory instability and high chemical loop gain [[124,](#page-18-12) [125\]](#page-18-13).

Ventilatory Response to Inspiratory Resistive Loading

During wakefulness, addition of an external resistive load leads to an immediate compensatory increase in ventilatory effort that maintains gas exchange. In sleep, this compensatory response is not normally seen unless there is a complete airway occlusion. With partial occlusion, a decrease in minute ventilation ensues and compensation of ventilation is delayed; this eventual correction is believed to be in response to gas exchange abnormalities. In normal children, this compensation can be limited and delayed by 3 minutes or more as compared to adults [\[126](#page-18-14)]. Young children with OSAS have reduced arousal responses to inspiratory resistive loads during sleep that together with the aforementioned inadequate compensation of ventilation may explain the prolonged periods of obstructive hypoventilation observed in childhood OSAS [[126\]](#page-18-14). Similar abnormalities in upper airway refexes and diminished response to resistive loading have been described in adolescents with OSAS and particularly if were obese and during REM sleep [[127,](#page-18-15) [128\]](#page-18-16).

Arousals from Sleep

Arousals are a normal phenomenon of sleep and are defned as sudden shifts in EEG frequency lasting for 3 seconds. However, if arousals occur too often, they produce sleep disruption, and interfere with the restorative nature of sleep. It should also be pointed out that arousals may be considered protective to subjects with OSAS since they coincide with increased dilator muscle activity, reduced upper airway resistance, and restoration of normal ventilation.

Studies in children and adults have clearly shown that frequent arousals and sleep fragmentation often lead to decreased vigilance, sleepiness, and other neurocognitive impairments. Interestingly, children are much less prone to arousals due to respiratory events than adults and typically are less sleepy compared to adults with OSAS. The major stimuli for arousals from OSAS are thought to be mechanical stimulation of lung and chest wall stretch receptors due to increased respiratory effort. However, hypercapnia is also considered a potent arousal stimulus. Most obstructive events in adults are associated with arousals from non-REM sleep. In children most obstructive events occur during REM sleep, and associated arousals are less frequent than in adults. Normal children have a higher arousal threshold than adults; children with OSAS seem to have an even loftier threshold for arousal in response to inspiratory loading [\[126](#page-18-14)] and hypercapnia [[123\]](#page-18-11) compared to children without OSAS.

Upper Airway Neuromotor Tone

Flow through the upper airway depends not only on mechanical and anatomic factors but also the active dilation of the airway by neuromotor tone [\[8](#page-15-5), [9\]](#page-15-6). Pressure–fow relationships based on the Starling model provide an understanding of airway stability in the "active" state with neuromotor activation and in the "passive state" before neuromotor responses are activated [[7,](#page-15-4) [60](#page-16-21), [129](#page-18-17)[–131](#page-18-18)] (Fig. [35.5\)](#page-10-0). Plotting a range of airway pressures against the resulting maximal inspiratory flows of breaths generates a pressure–flow line with the critical closing pressure (Pcrit) being represented by the intercept on the pressure axis (Fig. [35.6,](#page-11-0) adult). Airway pressure is applied by a nasal mask with the subject in a supine position and airfow is measured by a pneumotach; the pressures applied range from positive pressures to negative (subatmospheric) pressures. When pressure is maintained in a steady state, neuromotor activation occurs and the airway is in the active state; this active Pcrit is considered a measure of airway collapsibility. The nasal pressure at which the airway closes or is estimated to close by the pressure-fow line is typically lower for the active airway compared to the passive airway (Fig. [35.6\)](#page-11-0). In children, the Pcrit tends to be very negative (i.e., motor tone is very high) such that extrapola-

Passive airway: brief sudden drops in pressure

Fig. 35.5 Active and passive critical closing pressure protocols. Schematic of active and passive airway critical closing pressure protocols. Holding pressure (horizontal solid line) is maintained at levels just enough to abolish fow limitation. (**a**) Active protocol: To determine active critical airway closing pressure, nasal pressure (Pn) is reduced in 1–2 cm H2O decrements (broken lines) and maintained for prolonged periods (1–10 min) to allow dynamic airway activation to occur and maximal inspiratory air flow is obtained at each pressure. Airway pressure is reduced until airfow approaches zero or arousal from sleep occurs. (**b**) Passive protocol: To determine passive airway critical pressure, pressure drops (broken lines) from holding pressure (horizontal sold line) are made for brief periods lasting fve breaths, before dynamic responses are activated. Airway pressure is then raised to holding pressure rapidly for 1 or more minutes before dropping it further in increments of 1-2 cm H₂O pressure till zero flow is approximated or arousal occurs. Maximal inspiratory fow (Vi max) at each pressure setting is determined. Pressures employed span a range of positive to negative (sub atmospheric) values to estimate critical closing pressure

tion of the pressure fow line can become unreliable; the slope of the pressure–fow line is taken as the best estimate of upper airway collapsibility (Fig. [35.6](#page-11-0), child). In addition, airfow in the frst few breaths following a sudden drop in pressure, before neuromotor responses can occur, represents the "passive airway"; this passive Pcrit estimates mechanical and structural properties of the airway. A passive airway closing pressure can also be estimated by the pressure–crosssectional area relationship endoscopically observed in anaesthetized subjects (in whom neuromotor activation is suppressed) analogous to the pressure–airflow relationship. Neuromotor activation can more directly be estimated by measuring the EMG activity of the genioglossus muscle, which is the major pharyngeal dilator.

The pediatric airway is very resistant to collapse compared to the adult airway, airway collapsibility increases with age during adolescence and is not a function of pubertal development [\[7](#page-15-4), [129](#page-18-17)]. In children and adolescents with OSAS the critical closing pressure is much higher than non-OSAS children [\[7](#page-15-4), [60](#page-16-21), [128\]](#page-18-16). Childhood OSAS is most prominent in REM sleep which is associated with reduced pharyngeal tone and wide fuctuations of airfow, both of which probably contribute to OSAS. While closing pressure

Flow (VI max)

Fig. 35.6 Active and passive critical closing pressure (Pcrit) in a child and adult. Schematic representation of plots of nasal pressure (on *x*-axis) and maximum inspiratory air fow (Vi max) (on *y*-axis). Pressure fow lines calculated from fow at each pressure setting in the active or passive condition are used to obtain critical closing pressures; the intercept on the *x*-axis is the critical closing pressure. The activated pressure fow line (dashed line) has a lower airway closing pressure than the passive airway pressure fow line (solid line). Children tend to have very stable airways with a zero slope (dashed and solid line at top)

is difficult to measure in REM sleep for practical reasons, reduced airway tone can be demonstrated by EMG studies of the tongue muscles. Awake children with OSAS have higher baseline EMG tone than normal children, most probably to compensate for their narrower airways. With sleep onset these children have a rapid decline in EMG tone [[132\]](#page-18-19), with a further decline in REM sleep, predisposing them to airway obstruction during sleep [[133\]](#page-18-20).

Upper Airway Sensation

The afferent (sensory) loop of the upper airway negative pressure refex also plays a role in promoting airway stability. During wakefulness, topical nasopharyngeal anesthesia results in increased upper airway collapsibility in both children [\[50](#page-16-20)] and adults [\[134](#page-18-21)]. Similarly, during sleep, the application of topical nasopharyngeal anesthesia in adults results in increased upper airway collapsibility, leading to obstructive apnea [[135–](#page-18-22)[138\]](#page-18-23). The resultant worsening of apnea appears to be due at least in part to changes in muscle tone [\[139](#page-18-24)], but also to blunting of the arousal response [[135,](#page-18-22) [138](#page-18-23)]. These studies are supported by the study of Tapia et al. showing impaired sensation along of the tongue and hard palate and possible primary sensory function abnormality in children with OSAS during wakefulness. The latter authors speculated that this has been secondary to nerve damage and/ or hypoxemia caused by OSAS [\[140](#page-18-25)].

Summary of Functional Consideration

The pediatric airway is very resistant to collapse compared to the adult airway, and airway collapsibility increases with age and during adolescence. This trend may explain the lower prevalence of OSAS in children compared to adults. Differences in ventilatory drive, arousal thresholds, airway refexes during sleep, and active and passive properties of the upper airway also suggest different pathophysiological mechanisms in childhood OSAS compared to the adult form.

Active and passive techniques assessing airway collapsibility in sleep in children with OSAS show that the critical closing pressure is higher than in non-OSAS children. However, the overall ventilatory drive in response to hypoxia and hypercapnia is probably normal in OSAS children, although infants have a strong biphasic response to hypoxemia, and are more likely to develop central apnea when exposed to prolonged hypoxemia. The central ventilatory drive also plays a role in augmenting upper airway neuromotor refexes and tone. Normal children have brisker upper airway refexes during sleep than adults, perhaps due to their greater central ventilatory drive. These refexes appear to be blunted in children with OSAS. Finally, children with OSAS are less likely to arouse in response to upper airway obstruction and do not compensate for prolonged increases in inspiratory resistive load. This may explain why patients in the pediatric age group often have obstructive hypoventilation rather than discrete, cyclic obstructive apneas.

Biomechanical Considerations

In recent years, biomechanical modeling of the upper airway has been developed to characterize the anatomical and functional mechanisms that play a role in the pathophysiology of OSAS in children. This approach, called image-based modeling, uses imaging modalities such as magnetic resonance imaging (MRI) with engineering tools such as computational fluid dynamics (CFD) to analyze individual subjects. Dynamic imaging and image-based fow models create a rich data set that may help to explain how anatomy, tissue properties, and muscle function contribute to fow limitation, hypopneas, and apneas. Upper airway pressure felds modeled by CFD have been validated in vivo [[141\]](#page-18-26) and in vitro [[142,](#page-18-27) [143](#page-18-28)]. The pharynx in OSAS is often restricted where the adenoid, soft palate, and tonsils overlap [[53\]](#page-16-14), and CFD models can identify the location and quantify severity of anatomical restrictions. CFD model outcomes based on the drop

Fig. 35.7 Image-based CFD as a tool to assess surgical outcomes in children with OSAS. Top: CFD pressure felds at peak inspiratory fow, before (left) and after (right) adenotonsillectomy surgery, in a subject with 100% improved AHI postsurgery. Maximum pressure drop from choanae through the "overlap region" where tonsils and adenoids overlap (T and A on lower right image), dP_{TAmax} , was reduced by 96% after

surgery. Bottom left: reduction in dP_{TAmax} after surgery correlates strongly with improvement in AHI, especially in patients with signifcant postsurgery improvement. Bottom right: sagittal centerline MR image before surgery showing enlarged tonsil (T) and adenoid (A) also overlapping with the soft palate

in pressure between the choanae and location of maximum restriction have been correlated to OSAS severity in obese children [[141,](#page-18-26) [142](#page-18-27), [144](#page-18-29)]. Such studies identifed the drop in airway pressure between the choanae and the point of maximal narrowing at maximum inspiratory flow, dP_{TAmax} , as the

CFD measurement that most consistently correlated with OSAS severity [[141,](#page-18-26) [145\]](#page-18-30). Changes in CFD pressure drop correlate strongly to improvements in OSAS after AT surgery $[145]$ $[145]$ (Fig. [35.7](#page-12-0)) or oral appliance placement $[146,$ $[146,$ [147](#page-19-0)]. Compared to CFD biomarkers, correlation to strictly

anatomical biomarkers such as cross-section area or airway volume is often weaker or not signifcant [\[141](#page-18-26), [148\]](#page-19-1). The above works show that CFD has matured as a robust and accurate method to compute the effect of anatomical restriction over the upper airway air pressure feld.

An early study of pressure drops and fow resistance in the pharynx of individual children with OSAS [[142\]](#page-18-27) showed signifcantly higher airfow resistance in the upper airway of mildly sedated young children with OSAS. The same trend was observed in wakeful older obese children imaged during relaxed tidal breathing [\[141](#page-18-26)]. In a minority of OSAS cases, airflow resistance was lower $[149]$ $[149]$, suggesting that in these children OSAS does not result from anatomical restriction, but rather from altered *functional* factors related to tissue compliance and neuromuscular tone during sleep. Deficits in these factors can be suggested by the Pcrit which measures collapsibility of the upper airway, but Pcrit cannot identify the location of collapse or the anatomical or mechanical causes of collapse. Thus, image-based CFD may help to identify different patient phenotypes (Fig. [35.8](#page-13-0)) if it is combined with a functional assessment.

Fig. 35.8 OSAS phenotypes by computational fuid dynamics. (**a**) Anatomically driven OSAS: signifcant anatomical restriction in the "overlap region" leads to high pressure drop between choanae and minimum area of the pharynx in OSAS compared to controls. (**b**) Functional factor driven OSAS: OSAS with area restriction and CFD pharyngeal pressure drop similar to controls suggests loss of airway function during sleep. (**c**) Functional protection in control subject with anatomical restriction and high CFD pharyngeal pressure drop compared to OSAS. (From Wootton et al. [[141\]](#page-18-26). ©American Physiological Society)

CFD analysis based on dynamic imaging may be used to characterize both anatomical and functional factors, using a novel noninvasive method to compute the effective regional mechanical airway compliance [[150\]](#page-19-3). The effective compliance is the slope of a plot of cross-sectional area versus the local airway pressure, computed from image-based CFD. Effective compliance measures the combined effects of passive tissue properties and active airway muscle tone similar to Pcrit, but with the advantage of providing this data at any location along the airway in reference to the phase of the breathing cycle.

In a study of obese adolescent girls that combined dynamic MRI, CFD, and Pcrit, the effective compliance in the nasopharynx correlated signifcantly and negatively with both AHI and Pcrit [\[151](#page-19-4)], suggesting strong phasic airway dilator activation while awake, compensates for a collapsible airway, and loss of muscle tone during sleep leads to obstructions. An early study of sedated sleeping subjects showed positive effective compliance in subjects with OSAS, consistent with a passive, compliant pharynx susceptible to collapse [[150](#page-19-3)]. CFD based on dynamic MRI of subjects before and after the onset of sleep has the potential to reveal both anatomical and functional factors to better identify OSAS phenotypes.

Airfow resistance of the nasal passages from nares to choanae is another important biomechanical factor that often determines the majority of the air pressure force in the pharyngeal airway prior to airway collapse. Nasal resistance measured using anterior rhinomanometry is signifcantly higher in children with OSAS than controls [[152\]](#page-19-5), and may be an important tool for screening and diagnosing OSAS.

Additional Considerations

Edema

Although increased soft tissue size surrounding the airway as a cause for OSAS is primarily related to obesity, chronic edema, and infammation of the upper airway soft tissues may further restrict the dimensions of the upper airway. The mechanism for this effect is speculated to be the effect of chronic vibratory effects of snoring, and of upper airway soft tissue being tugged caudally during fuctuation in intrathoracic pressure, resulting in trauma to the upper airway soft tissues [\[69](#page-16-32), [70\]](#page-16-28). Indeed, the therapeutic effect of CPAP is thought to be partially mediated through a reduction in upper airway soft tissue edema, and the use of CPAP has been shown to reduce soft palate volume [[153\]](#page-19-6).

Myopathy

It has been suggested that patients with OSAS have a primary myopathy. Several studies have demonstrated an increase in type II fast twitch fbers in the genioglossus of patients with OSAS [\[154](#page-19-7)[–156](#page-19-8)]. Type II fbers are less resistant to fatigue than type I fbers. It is possible that the increased number of type II fbers is secondary to chronic muscle injury which, in turn, may alter the size, length, and confguration of the affected muscles.

Gender

In adults, OSAS is far more common in males than females [\[157](#page-19-9)]. Considerable effort has been expended in trying to determine the mechanisms underlying this male predominance, but no clear explanation has emerged. Studies have not shown differences in pharyngeal anatomy resulting in a smaller pharyngeal lumen in males. On the contrary, females were found to have a smaller pharynx [\[158](#page-19-10), [159\]](#page-19-11), despite the presence of larger soft tissues in males [[160\]](#page-19-12). It is therefore possible that the reduced occurrence of OSAS in females is due to a stiffer and less collapsible upper airway despite its smaller size [[161–](#page-19-13)[163\]](#page-19-14). Speculated mechanisms mediating differences in airway collapsibility include hormonal differences, differences in chemosensitivity, and differences in tissue properties [[161,](#page-19-13) [162,](#page-19-15) [164\]](#page-19-16).

In contrast, no gender differences have been noted in children with OSAS. Pillar et al. evaluated the upper airway length in pre- and post-pubertal non-OSAS children using CT images. They noted that airway length after normalization was signifcantly greater in males in post-pubertal years and speculated that such changes in length after puberty may predispose males to OSAS later in life [\[165](#page-19-17)].

As mentioned above, PCOS is one of the most common endocrine disorders associated with overweight and obesity and affects 5–10% of adolescent girls and women of reproductive age [\[112](#page-18-1)]. Screening for OSAS has therefore been recommended in these subjects [[118\]](#page-18-6).

Genetics

Genetic factors most probably play an important role in both the pathophysiology and health outcomes of OSAS in children and adults. However, genetic studies in the feld of OSAS particularly in children lag behind other common medical disorders.

Evidence to suggest that OSAS is genetically mediated in children includes the following: (1) the strong association of OSAS with discrete craniofacial skeletal disorders restricting the upper airway such as Down syndrome, Treacher Collins, and Apert syndrome; (2) distinct genetic disorders associated with obesity that present with an extremely high prevalence of OSAS and sleep disordered breathing such as PWS, PCOS, and melanocortin-4 receptor

deficiency. The above examples suggest that distinct genes regulating upper airway morphology, body composition, or ventilatory control also contribute to the pathophysiology of OSAS in such groups. However, so far, such genes have not been identifed and it is unclear if such putative genes share similarities to genes responsible for OSAS on otherwise healthy children.

In adults, evidence to support that OSAS is a heritable disorder include the marked gender difference in disease prevalence and progression [[157\]](#page-19-9), familial aggregations of the disorder $[166]$ $[166]$, and twin studies $[167]$ $[167]$.

Genetic approaches to study OSAS have been utilized only in adults. Research in this area has been limited by the relatively small number of studies, small number of participants and the small number of replication studies. Standard genetic approaches include heritability studies to discover candidate genes using single nucleotide polymorphisms (SNPs) and OSAS phenotypes in case–control or cohort studies, or genome-wide linkage studies and genome wide association studies (GWAS) to identify causal genes without having a priori knowledge of functionality [\[168](#page-19-20)].

Nevertheless, several genetic association studies have identifed candidate polymorphisms in genes linked to OSAS: the tumor necrosis factor alpha (TNF- $α$) gene (−308G/A) [[169\]](#page-19-21), the rs1409986 SNP in the prostaglandin E2 receptor (PTGER3) gene [\[170](#page-19-22)], and the rs7030789 SNP in the lysophosphatidic acid receptor 1 (LPAR1) gene [\[170](#page-19-22)].

Large-scale GWAS have identifed loci for traits associated with OSAS such as: (1) polymorphism in the G-protein receptor gene (GPR83) [[171\]](#page-19-23) which is expressed in several areas of the brain including the hypoglossal nucleus, the dorsal motor nucleus of the vagus, and the nucleus of the solitary tract; (2) variants in the β-arrestin 1 (ARRB1) gene, which is an important regulator of hypoxia inducible factor 1 alpha (HIF-1 α) [[171\]](#page-19-23); (3) genes regulating obesity and body composition and insulin resistance $[172]$ $[172]$; and (4) genes regulating craniofacial structures [[173\]](#page-20-0). It still remains to examine if these genes are applicable to risk for OSAS in children.

Research Questions

There are many pressing unanswered questions regarding the developmental pathophysiology of OSAS. In particular:

- What is the pathophysiology of the "idiopathic" OSAS often seen in infants?
- What is the natural history of childhood OSAS? Is childhood OSAS a precursor of adult OSAS, or a separate disease process? If the former, what is the recurrence rate during later life, and what are the risk factors for recurrence?
- What is the effect of childhood OSAS on craniofacial growth and structure? Does adenotonsillar hypertrophy per se result in craniofacial changes that could result in obstructive apnea later in life, or is the apparent adenotonsillar hypertrophy a result of overcrowding from a narrower upper airway? What is the potential role of treatments such as intraoral appliances and rapid maxillary expansion in changing craniofacial growth in children?
- When does the childhood pattern of OSAS transition into the adult pattern? What are the effects of puberty on upper airway function and structure?
- What roles do genetic, ethnic, and anthropometric factors play in the pathophysiology of OSAS?
- How can biomechanical studies using novel imaging modalities such as MRI improve diagnosis and treatment for children with OSAS?

Addressing these questions may improve diagnosis and provide optimal treatment outcomes for various phenotype of childhood OSAS in the upcoming years.

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References

- 1. Marcus CL. Sleep-disordered breathing in children. Am J Respir Crit Care Med. 2001;164:16–30.
- 2. Cardiorespiratory sleep studies in children. Establishment of normative data and polysomnographic predictors of morbidity. American Thoracic Society. Am J Respir Crit Care Med. 1999;160:1381–7.
- 3. McNamara F, Sullivan CE. Pediatric origins of adult lung diseases. 3: the genesis of adult sleep apnoea in childhood. Thorax. 2000;55:964–9.
- 4. Rosen CL, D'Andrea L, Haddad GG. Adult criteria for obstructive sleep apnea do not identify children with serious obstruction. Am Rev Respir Dis. 1992;146:1231–4.
- 5. Carroll JL, Loughlin GM. Diagnostic criteria for obstructive sleep apnea syndrome in children. Pediatr Pulmonol. 1992;14:71–4.
- 6. Standards and indications for cardiopulmonary sleep studies in children. American Thoracic Society. Am J Respir Crit Care Med. 1996;153:866–78.
- 7. Arens R, Marcus CL. Pathophysiology of upper airway obstruction: a developmental perspective. Sleep. 2004;27:997–1019.
- 8. Mezzanotte WS, Tangel DJ, White DP. Waking genioglossal electromyogram in sleep apnea patients versus normal controls (a neuromuscular compensatory mechanism). J Clin Invest. 1992;89:1571–9.
- 9. Mezzanotte WS, Tangel DJ, White DP. Infuence of sleep onset on upper-airway muscle activity in apnea patients versus normal controls. Am J Respir Crit Care Med. 1996;153:1880–7.
- 10. Laitman JT, Crelin ES, Conlogue GJ. The function of the epiglottis in monkey and man. Yale J Biol Med. 1977;50:43–8.
- 11. van Lunteren E. Muscles of the pharynx: structural and contractile properties. Ear Nose Throat J. 1993;72:27–9, 33.
- 12. van Lunteren E, Strohl KP. The muscles of the upper airways. Clin Chest Med. 1986;7:171–88.
- 13. Kuna ST, Smickley JS, Vanoye CR. Respiratory-related pharyngeal constrictor muscle activity in normal human adults. Am J Respir Crit Care Med. 1997;155:1991–9.
- 14. Schwab RJ, Gupta KB, Gefter WB, Metzger LJ, Hoffman EA, Pack AI. Upper airway and soft tissue anatomy in normal subjects and patients with sleep-disordered breathing. Signifcance of the lateral pharyngeal walls. Am J Respir Crit Care Med. 1995;152:1673–89.
- 15. Arens R, McDonough JM, Corbin AM, Hernandez EM, Maislin G, Schwab RJ, Pack AI. Linear dimensions of the upper airway structure during development: assessment by magnetic resonance imaging. Am J Respir Crit Care Med. 2002;165:117–22.
- 16. Katz ES, Mitchell RB, D'Ambrosio CM. Obstructive sleep apnea in infants. Am J Respir Crit Care Med. 2012;185:805–16.
- 17. Thach BT, Stark AR. Spontaneous neck fexion and airway obstruction during apneic spells in preterm infants. J Pediatr. 1979;94:275–81.
- 18. Swift PG, Emery JL. Clinical observations on response to nasal occlusion in infancy. Arch Dis Child. 1973;48:947–51.
- 19. Zafereo ME, Taylor RJ, Pereira KD. Supraglottoplasty for laryngomalacia with obstructive sleep apnea. Laryngoscope. 2008;118:1873–7.
- 20. Mixter RC, David DJ, Perloff WH, Green CG, Pauli RM, Popic PM. Obstructive sleep apnea in Apert's and Pfeiffer's syndromes: more than a craniofacial abnormality. Plast Reconstr Surg. 1990;86:457–63.
- 21. Sculerati N, Gottlieb MD, Zimbler MS, Chibbaro PD, McCarthy JG. Airway management in children with major craniofacial anomalies. Laryngoscope. 1998;108:1806–12.
- 22. Spier S, Rivlin J, Rowe RD, Egan T. Sleep in Pierre Robin syndrome. Chest. 1986;90:711–5.
- 23. Abramson DL, Marrinan EM, Mulliken JB. Robin sequence: obstructive sleep apnea following pharyngeal fap. Cleft Palate Craniofac J. 1997;34:256–60.
- 24. Shprintzen RJ. Pierre Robin, micrognathia, and airway obstruction: the dependency of treatment on accurate diagnosis. Int Anesthesiol Clin. 1988;26:64–71.
- 25. Sher AE. Mechanisms of airway obstruction in Robin sequence: implications for treatment. Cleft Palate Craniofac J. 1992;29:224–31.
- 26. Johnston C, Taussig LM, Koopmann C, Smith P, Bjelland J. Obstructive sleep apnea in Treacher-Collins syndrome. Cleft Palate J. 1981;18:39–44.
- 27. Donaldson JD, Redmond WM. Surgical management of obstructive sleep apnea in children with Down syndrome. J Otolaryngol. 1988;17:398–403.
- 28. Marcus CL, Keens TG, Bautista DB, von Pechmann WS, Ward SL. Obstructive sleep apnea in children with Down syndrome. Pediatrics. 1991;88:132–9.
- 29. Southall DP, Stebbens VA, Mirza R, Lang MH, Croft CB, Shinebourne EA. Upper airway obstruction with hypoxaemia and sleep disruption in Down syndrome. Dev Med Child Neurol. 1987;29:734–42.
- 30. Stebbens VA, Dennis J, Samuels MP, Croft CB, Southall DP. Sleep related upper airway obstruction in a cohort with Down's syndrome. Arch Dis Child. 1991;66:1333–8.
- 31. Jacobs IN, Gray RF, Todd NW. Upper airway obstruction in children with Down syndrome. Arch Otolaryngol Head Neck Surg. 1996;122:945–50.
- 32. Uong EC, McDonough JM, Tayag-Kier CE, Zhao H, Haselgrove J, Mahboubi S, Schwab RJ, Pack AI, Arens R. Magnetic resonance imaging of the upper airway in children with Down syndrome. Am J Respir Crit Care Med. 2001;163:731–6.
- 33. Brouillette RT, Fernbach SK, Hunt CE. Obstructive sleep apnea in infants and children. J Pediatr. 1982;100:31–40.
- 34. Leiberman A, Tal A, Brama I, Sofer S. Obstructive sleep apnea in young infants. Int J Pediatr Otorhinolaryngol. 1988;16:39–44.
- 35. Leighton SE, Papsin B, Vellodi A, Dinwiddie R, Lane R. Disordered breathing during sleep in patients with mucopolysaccharidoses. Int J Pediatr Otorhinolaryngol. 2001;58:127–38.
- 36. Kotoku R, Kinouchi K, Fukumitsu K, Taniguchi A. A neonate with Beckwith-Wiedemann syndrome who developed upper airway obstruction after glossopexy. Masui. 2002;51:46–8.
- 37. Liao YF, Chuang ML, Chen PK, Chen NH, Yun C, Huang CS. Incidence and severity of obstructive sleep apnea following pharyngeal fap surgery in patients with cleft palate. Cleft Palate Craniofac J. 2002;39:312–6.
- 38. de Serres LM, Deleyiannis FW, Eblen LE, Gruss JS, Richardson MA, Sie KC. Results with sphincter pharyngoplasty and pharyngeal fap. Int J Pediatr Otorhinolaryngol. 1999;48:17–25.
- 39. Jennum P, Borgesen SE. Intracranial pressure and obstructive sleep apnea. Chest. 1989;95:279–83.
- 40. Guilleminault C, Korobkin R, Winkle R. A review of 50 children with obstructive sleep apnea syndrome. Lung. 1981;159:275–87.
- 41. Gozal D, Arens R, Omlin KJ, Jacobs RA, Keens TG. Peripheral chemoreceptor function in children with myelomeningocele and Arnold-Chiari malformation type 2. Chest. 1995;108:425–31.
- 42. Gilmore RL, Falace P, Kanga J, Baumann R. Sleep-disordered breathing in Mobius syndrome. J Child Neurol. 1991;6:73–7.
- 43. Ali NJ, Pitson DJ, Stradling JR. Snoring, sleep disturbance, and behaviour in 4–5 year olds. Arch Dis Child. 1993;68:360–6.
- 44. Redline S, Tishler PV, Schluchter M, Aylor J, Clark K, Graham G. Risk factors for sleep-disordered breathing in children. Associations with obesity, race, and respiratory problems. Am J Respir Crit Care Med. 1999;159:1527–32.
- 45. Corbo GM, Fuciarelli F, Foresi A, De Benedetto F. Snoring in children: association with respiratory symptoms and passive smoking. BMJ. 1989;299:1491–4.
- 46. Isono S, Shimada A, Utsugi M, Konno A, Nishino T. Comparison of static mechanical properties of the passive pharynx between normal children and children with sleep-disordered breathing. Am J Respir Crit Care Med. 1998;157:1204–12.
- 47. Arens R, McDonough JM, Costarino AT, Mahboubi S, Tayag-Kier CE, Maislin G, Schwab RJ, Pack AI. Magnetic resonance imaging of the upper airway structure of children with obstructive sleep apnea syndrome. Am J Respir Crit Care Med. 2001;164:698–703.
- 48. Monahan KJ, Larkin EK, Rosen CL, Graham G, Redline S. Utility of noninvasive pharyngometry in epidemiologic studies of childhood sleep-disordered breathing. Am J Respir Crit Care Med. 2002;165:1499–503.
- 49. Kulnis R, Nelson S, Strohl K, Hans M. Cephalometric assessment of snoring and nonsnoring children. Chest. 2000;118:596–603.
- 50. Gozal D, Burnside MM. Increased upper airway collapsibility in children with obstructive sleep apnea during wakefulness. Am J Respir Crit Care Med. 2004;169:163–7.
- 51. Croft CB, Brockbank MJ, Wright A, Swanston AR. Obstructive sleep apnoea in children undergoing routine tonsillectomy and adenoidectomy. Clin Otolaryngol. 1990;15:307–14.
- 52. Fernbach SK, Brouillette RT, Riggs TW, Hunt CE. Radiologic evaluation of adenoids and tonsils in children with obstructive sleep apnea: plain flms and fuoroscopy. Pediatr Radiol. 1983;13:258–65.
- 53. Arens R, McDonough JM, Corbin AM, Rubin NK, Carroll ME, Pack AI, Liu J, Udupa JK. Upper airway size analysis by magnetic resonance imaging of children with obstructive sleep apnea syndrome. Am J Respir Crit Care Med. 2003;167:65–70.
- 54. Arens R, Sin S, Nandalike K, Rieder J, Khan UI, Freeman K, Wylie-Rosett J, Lipton ML, Wootton DM, McDonough JM,

Shifteh K. Upper airway structure and body fat composition in obese children with obstructive sleep apnea syndrome. Am J Respir Crit Care Med. 2011;183:782–7.

- 55. Liu J, Udupa JK, Odhnera D, McDonough JM, Arens R. System for upper airway segmentation and measurement with MR imaging and fuzzy connectedness. Acad Radiol. 2003;10:13–24.
- 56. Arens R, Sin S, McDonough JM, Palmer JM, Dominguez T, Meyer H, Wootton DM, Pack AI. Changes in upper airway size during tidal breathing in children with obstructive sleep apnea syndrome. Am J Respir Crit Care Med. 2005;171:1298–304.
- 57. Wagshul ME, Sin S, Lipton ML, Shifteh K, Arens R. Novel retrospective, respiratory-gating method enables 3D, high resolution, dynamic imaging of the upper airway during tidal breathing. Magn Reson Med. 2013;70:1580–90.
- 58. Donnelly LF, Casper KA, Chen B. Correlation on cine MR imaging of size of adenoid and palatine tonsils with degree of upper airway motion in asymptomatic sedated children. AJR Am J Roentgenol. 2002;179:503–8.
- 59. Fregosi RF, Quan SF, Kaemingk KL, Morgan WJ, Goodwin JL, Cabrera R, Gmitro A. Sleep-disordered breathing, pharyngeal size and soft tissue anatomy in children. J Appl Physiol. 2003;95:2030–8.
- 60. Marcus CL, McColley SA, Carroll JL, Loughlin GM, Smith PL, Schwartz AR. Upper airway collapsibility in children with obstructive sleep apnea syndrome. J Appl Physiol. 1994;77:918–24.
- 61. Brodsky L, Adler E, Stanievich JF. Naso- and oropharyngeal dimensions in children with obstructive sleep apnea. Int J Pediatr Otorhinolaryngol. 1989;17:1–11.
- 62. Guilleminault C, Eldridge FL, Simmons FB, Dement WC. Sleep apnea in eight children. Pediatrics. 1976;58:23–30.
- 63. Suen JS, Arnold JE, Brooks LJ. Adenotonsillectomy for treatment of obstructive sleep apnea in children. Arch Otolaryngol Head Neck Surg. 1995;121:525–30.
- 64. Rosen GM, Muckle RP, Mahowald MW, Goding GS, Ullevig C. Postoperative respiratory compromise in children with obstructive sleep apnea syndrome: can it be anticipated? Pediatrics. 1994;93:784–8.
- 65. Tal A, Bar A, Leiberman A, Tarasiuk A. Sleep characteristics following adenotonsillectomy in children with obstructive sleep apnea syndrome. Chest. 2003;124:948–53.
- 66. Marcus CL, Ward SL, Mallory GB, Rosen CL, Beckerman RC, Weese-Mayer DE, Brouillette RT, Trang HT, Brooks LJ. Use of nasal continuous positive airway pressure as treatment of childhood obstructive sleep apnea. J Pediatr. 1995;127:88–94.
- 67. Brodsky L, Moore L, Stanievich JF. A comparison of tonsillar size and oropharyngeal dimensions in children with obstructive adenotonsillar hypertrophy. Int J Pediatr Otorhinolaryngol. 1987;13:149–56.
- 68. Ryan CF, Lowe AA, Li D, Fleetham JA. Magnetic resonance imaging of the upper airway in obstructive sleep apnea before and after chronic nasal continuous positive airway pressure therapy. Am Rev Respir Dis. 1991;144:939–44.
- 69. Hamans EP, Van Marck EA, De Backer WA, Creten W, Van de Heyning PH. Morphometric analysis of the uvula in patients with sleep-related breathing disorders. Eur Arch Otorhinolaryngol. 2000;257:232–6.
- 70. Sekosan M, Zakkar M, Wenig BL, Olopade CO, Rubinstein I. Infammation in the uvula mucosa of patients with obstructive sleep apnea. Laryngoscope. 1996;106:1018–20.
- 71. Shintani T, Asakura K, Kataura A. Adenotonsillar hypertrophy and skeletal morphology of children with obstructive sleep apnea syndrome. Acta Otolaryngol Suppl. 1996;523:222–4.
- 72. Shintani T, Asakura K, Kataura A. Evaluation of the role of adenotonsillar hypertrophy and facial morphology in children with obstructive sleep apnea. ORL J Otorhinolaryngol Relat Spec. 1997;59:286–91.
- 73. Kawashima S, Niikuni N, Chia-hung L, Takahasi Y, Kohno M, Nakajima I, Akasaka M, Sakata H, Akashi S. Cephalometric comparisons of craniofacial and upper airway structures in young children with obstructive sleep apnea syndrome. Ear Nose Throat J. 2000;79:499–502, 505–496.
- 74. Agren K, Nordlander B, Linder-Aronsson S, Zettergren-Wijk L, Svanborg E. Children with nocturnal upper airway obstruction: postoperative orthodontic and respiratory improvement. Acta Otolaryngol. 1998;118:581–7.
- 75. Kawashima S, Peltomaki T, Sakata H, Mori K, Happonen RP, Ronning O. Craniofacial morphology in preschool children with sleep-related breathing disorder and hypertrophy of tonsils. Acta Paediatr. 2002;91:71–7.
- 76. Zucconi M, Caprioglio A, Calori G, Ferini-Strambi L, Oldani A, Castronovo C, Smirne S. Craniofacial modifcations in children with habitual snoring and obstructive sleep apnoea: a case-control study. Eur Respir J. 1999;13:411–7.
- 77. Behlfelt K. Enlarged tonsils and the effect of tonsillectomy. Characteristics of the dentition and facial skeleton. Posture of the head, hyoid bone and tongue. Mode of breathing. Swed Dent J Suppl. 1990;72:1–35.
- 78. Hultcrantz E, Larson M, Hellquist R, Ahlquist-Rastad J, Svanholm H, Jakobsson OP. The infuence of tonsillar obstruction and tonsillectomy on facial growth and dental arch morphology. Int J Pediatr Otorhinolaryngol. 1991;22:125–34.
- 79. Schiffman PH, Rubin NK, Dominguez T, Mahboubi S, Udupa JK, O'Donnell AR, McDonough JM, Maislin G, Schwab RJ, Arens R. Mandibular dimensions in children with obstructive sleep apnea. Sleep. 2004;27(5):959–65.
- 80. Marcus CL, Carroll JL, Koerner CB, Hamer A, Lutz J, Loughlin GM. Determinants of growth in children with the obstructive sleep apnea syndrome. J Pediatr. 1994;125:556–62.
- 81. Dietz WH. Health consequences of obesity in youth: childhood predictors of adult disease. Pediatrics. 1998;101:518–25.
- 82. Wang G, Dietz WH. Economic burden of obesity in youths aged 6 to 17 years: 1979–1999. Pediatrics. 2002;109:E81–81.
- 83. Mallory GB Jr, Fiser DH, Jackson R. Sleep-associated breathing disorders in morbidly obese children and adolescents. J Pediatr. 1989;115:892–7.
- 84. Brooks LJ, Stephens BM, Bacevice AM. Adenoid size is related to severity but not the number of episodes of obstructive apnea in children. J Pediatr. 1998;132:682–6.
- 85. Silvestri JM, Weese-Mayer DE, Bass MT, Kenny AS, Hauptman SA, Pearsall SM. Polysomnography in obese children with a history of sleep-associated breathing disorders. Pediatr Pulmonol. 1993;16:124–9.
- 86. Marcus CL, Curtis S, Koerner CB, Joffe A, Serwint JR, Loughlin GM. Evaluation of pulmonary function and polysomnography in obese children and adolescents. Pediatr Pulmonol. 1996;21:176–83.
- 87. Rosen CL. Clinical features of obstructive sleep apnea hypoventilation syndrome in otherwise healthy children. Pediatr Pulmonol. 1999;27:403–9.
- 88. Kahn A, Mozin MJ, Rebuffat E, Sottiaux M, Burniat W, Shepherd S, Muller MF. Sleep pattern alterations and brief airway obstructions in overweight infants. Sleep. 1989;12:430–8.
- 89. Kalra M, Inge T, Garcia V, Daniels S, Lawson L, Curti R, Cohen A, Amin R. Obstructive sleep apnea in extremely overweight adolescents undergoing bariatric surgery. Obes Res. 2005;13:1175–9.
- 90. Clinical practice guideline: diagnosis and management of childhood obstructive sleep apnea syndrome. Pediatrics. 2002;109:704–12.
- 91. Guilleminault C, Li KK, Khramtsov A, Pelayo R, Martinez S. Sleep disordered breathing: surgical outcomes in prepubertal children. Laryngoscope. 2004;114:132–7.
- 92. Spector A, Scheid S, Hassink S, Deutsch ES, Reilly JS, Cook SP. Adenotonsillectomy in the morbidly obese child. Int J Pediatr Otorhinolaryngol. 2003;67:359–64.
- 93. Chay OM, Goh A, Abisheganaden J, Tang J, Lim WH, Chan YH, Wee MK, Johan A, John AB, Cheng HK, Lin M, Chee T, Rajan U, Wang S, Machin D. Obstructive sleep apnea syndrome in obese Singapore children. Pediatr Pulmonol. 2000;29:284–90.
- 94. Wing YK, Hui SH, Pak WM, Ho CK, Cheung A, Li AM, Fok TF. A controlled study of sleep related disordered breathing in obese children. Arch Dis Child. 2003;88:1043–7.
- 95. Gordon JE, Hughes MS, Shepherd K, Szymanski DA, Schoenecker PL, Parker L, Uong EC. Obstructive sleep apnoea syndrome in morbidly obese children with tibia vara. J Bone Joint Surg Br. 2006;88:100–3.
- 96. Mitchell RB, Kelly J. Adenotonsillectomy for obstructive sleep apnea in obese children. Otolaryngol Head Neck Surg. 2004;131:104–8.
- 97. Horner RL, Mohiaddin RH, Lowell DG, Shea SA, Burman ED, Longmore DB, Guz A. Sites and sizes of fat deposits around the pharynx in obese patients with obstructive sleep apnoea and weight matched controls. Eur Respir J. 1989;2:613–22.
- 98. Pillai S, Nandalike K, Kogelman Y, Muzumdar R, Balk SJ, Arens R. Severe obstructive sleep apnea in a child with melanocortin-4 receptor deficiency. J Clin Sleep Med. 2014;10:99-101.
- 99. Orenstein DM, Boat TF, Stern RC, Doershuk CF, Light MS. Progesterone treatment of the obesity hypoventilation syndrome in a child. J Pediatr. 1977;90:477–9.
- 100. Sanchez-Armengol A, Fuentes-Pradera MA, Capote-Gil F, Garcia-Diaz E, Cano-Gomez S, Carmona-Bernal C, Castillo-Gomez J. Sleep-related breathing disorders in adolescents aged 12 to 16 years: clinical and polygraphic fndings. Chest. 2001;119:1393–400.
- 101. Morton S, Rosen C, Larkin E, Tishler P, Aylor J, Redline S. Predictors of sleep-disordered breathing in children with a history of tonsillectomy and/or adenoidectomy. Sleep. 2001;24:823–9.
- 102. Tasker C, Crosby JH, Stradling JR. Evidence for persistence of upper airway narrowing during sleep, 12 years after adenotonsillectomy. Arch Dis Child. 2002;86:34–7.
- 103. Guilleminault C, Partinen M, Praud JP, Quera-Salva MA, Powell N, Riley R. Morphometric facial changes and obstructive sleep apnea in adolescents. J Pediatr. 1989;114:997–9.
- 104. Guilleminault C, Pelayo R, Leger D, Clerk A, Bocian RC. Recognition of sleep-disordered breathing in children. Pediatrics. 1996;98:871–82.
- 105. Ogden CL, Flegal KM, Carroll MD, Johnson CL. Prevalence and trends in overweight among US children and adolescents, 1999– 2000. JAMA. 2002;288:1728–32.
- 106. Steinberger J, Steffen L, Jacobs DR Jr, Moran A, Hong CP, Sinaiko AR. Relation of leptin to insulin resistance syndrome in children. Obes Res. 2003;11:1124–30.
- 107. Steinberger J. Diagnosis of the metabolic syndrome in children. Curr Opin Lipidol. 2003;14:555–9.
- 108. Steinberger J, Daniels SR. Obesity, insulin resistance, diabetes, and cardiovascular risk in children: an American Heart Association scientifc statement from the Atherosclerosis, Hypertension, and Obesity in the Young Committee (Council on Cardiovascular Disease in the Young) and the Diabetes Committee (Council on Nutrition, Physical Activity, and Metabolism). Circulation. 2003;107:1448–53.
- 109. Redline S, Storfer-Isser A, Rosen CL, Johnson NL, Kirchner HL, Emancipator J, Kibler AM. Association between metabolic syndrome and sleep-disordered breathing in adolescents. Am J Respir Crit Care Med. 2007;176:401–8.
- 110. Schwab RJ, Kim C, Bagchi S, Keenan BT, Comyn FL, Wang S, Tapia IE, Huang S, Traylor J, Torigian DA, Bradford RM,

Marcus CL. Understanding the anatomic basis for obstructive sleep apnea syndrome in adolescents. Am J Respir Crit Care Med. 2015;191:1295–309.

- 111. Marcus CL, Keenan BT, Huang J, Yuan H, Pinto S, Bradford RM, Kim C, Bagchi S, Comyn FL, Wang S, Tapia IE, Maislin G, Cielo CM, Traylor J, Torigian DA, Schwab RJ. The obstructive sleep apnoea syndrome in adolescents. Thorax. 2017;72:720–8.
- 112. Azziz R, Woods KS, Reyna R, Key TJ, Knochenhauer ES, Yildiz BO. The prevalence and features of the polycystic ovary syndrome in an unselected population. J Clin Endocrinol Metab. 2004;89:2745–9.
- 113. Ehrmann DA. Polycystic ovary syndrome. N Engl J Med. 2005;352:1223–36.
- 114. Nandalike K, Strauss T, Agarwal C, Coupey SM, Sin S, Rajpathak S, Cohen HW, Arens R. Screening for sleep-disordered breathing and excessive daytime sleepiness in adolescent girls with polycystic ovarian syndrome. J Pediatr. 2011;159:591–6.
- 115. Fogel RB, Malhotra A, Pillar G, Pittman SD, Dunaif A, White DP. Increased prevalence of obstructive sleep apnea syndrome in obese women with polycystic ovary syndrome. J Clin Endocrinol Metab. 2001;86:1175–80.
- 116. Tasali E, Van Cauter E, Ehrmann DA. Relationships between sleep disordered breathing and glucose metabolism in polycystic ovary syndrome. J Clin Endocrinol Metab. 2006;91:36–42.
- 117. Vgontzas AN, Legro RS, Bixler EO, Grayev A, Kales A, Chrousos GP. Polycystic ovary syndrome is associated with obstructive sleep apnea and daytime sleepiness: role of insulin resistance. J Clin Endocrinol Metab. 2001;86:517–20.
- 118. Subramanian S, Desai A, Joshipura M, Surani S. Practice patterns of screening for sleep apnea in physicians treating PCOS patients. Sleep Breath. 2007;11:233–7.
- 119. White DP. Pathogenesis of obstructive and central sleep apnea. Am J Respir Crit Care Med. 2005;172:1363–70.
- 120. Younes M, Ostrowski M, Thompson W, Leslie C, Shewchuk W. Chemical control stability in patients with obstructive sleep apnea. Am J Respir Crit Care Med. 2001;163:1181–90.
- 121. Foster GE, Hanly PJ, Ostrowski M, Poulin MJ. Ventilatory and cerebrovascular responses to hypercapnia in patients with obstructive sleep apnoea: effect of CPAP therapy. Respir Physiol Neurobiol. 2009;165:73–81.
- 122. Marcus CL, Gozal D, Arens R, Basinski DJ, Omlin KJ, Keens TG, Ward SL. Ventilatory responses during wakefulness in children with obstructive sleep apnea. Am J Respir Crit Care Med. 1994;149:715–21.
- 123. Marcus CL, Lutz J, Carroll JL, Bamford O. Arousal and ventilatory responses during sleep in children with obstructive sleep apnea. J Appl Physiol. 1998;84:1926–36.
- 124. Nava-Guerra L, Tran WH, Chalacheva P, Loloyan S, Joshi B, Keens TG, Nayak KS, Davidson Ward SL, Khoo MC. Modelbased stability assessment of ventilatory control in overweight adolescents with obstructive sleep apnea during NREM sleep. J Appl Physiol (1985). 2016;121:185–97.
- 125. Yuan H, Pinto SJ, Huang J, McDonough JM, Ward MB, Lee YN, Bradford RM, Gallagher PR, Shults J, Konstantinopoulou S, Samuel JM, Katz ES, Hua S, Tapia IE, Marcus CL. Ventilatory responses to hypercapnia during wakefulness and sleep in obese adolescents with and without obstructive sleep apnea syndrome. Sleep. 2012;35:1257–67.
- 126. Marcus CL, Moreira GA, Bamford O, Lutz J. Response to inspiratory resistive loading during sleep in normal children and children with obstructive apnea. J Appl Physiol. 1999;87:1448–54.
- 127. Huang J, Karamessinis LR, Pepe ME, Glinka SM, Samuel JM, Gallagher PR, Marcus CL. Upper airway collapsibility during REM sleep in children with the obstructive sleep apnea syndrome. Sleep. 2009;32:1173–81.
- 128. Huang J, Pinto SJ, Yuan H, Katz ES, Karamessinis LR, Bradford RM, Gallagher PR, Hannigan JT, Nixon T, Ward MB, Lee YN, Marcus CL. Upper airway collapsibility and genioglossus activity in adolescents during sleep. Sleep. 2012;35:1345–52.
- 129. Marcus CL, Lutz J, Hamer A, Smith PL, Schwartz A. Developmental changes in response to subatmospheric pressure loading of the upper airway. J Appl Physiol. 1999;87:626–33.
- 130. Schwartz AR, Smith PL, Wise RA, Bankman I, Permutt S. Effect of positive nasal pressure on upper airway pressure-fow relationships. J Appl Physiol. 1989;66:1626–34.
- 131. Schwartz AR, Smith PL, Wise RA, Gold AR, Permutt S. Induction of upper airway occlusion in sleeping individuals with subatmospheric nasal pressure. J Appl Physiol. 1988;64:535–42.
- 132. Katz ES, White DP. Genioglossus activity in children with obstructive sleep apnea during wakefulness and sleep onset. Am J Respir Crit Care Med. 2003;168:664–70.
- 133. Katz ES, White DP. Genioglossus activity during sleep in normal control subjects and children with obstructive sleep apnea. Am J Respir Crit Care Med. 2004;170:553–60.
- 134. Fogel RB, Malhotra A, Shea SA, Edwards JK, White DP. Reduced genioglossal activity with upper airway anesthesia in awake patients with OSA. J Appl Physiol. 2000;88:1346–54.
- 135. Berry RB, Kouchi KG, Bower JL, Light RW. Effect of upper airway anesthesia on obstructive sleep apnea. Am J Respir Crit Care Med. 1995;151:1857–61.
- 136. Chadwick GA, Crowley P, Fitzgerald MX, O'Regan RG, McNicholas WT. Obstructive sleep apnea following topical oropharyngeal anesthesia in loud snorers. Am Rev Respir Dis. 1991;143:810–3.
- 137. McNicholas WT, Coffey M, McDonnell T, O'Regan R, Fitzgerald MX. Upper airway obstruction during sleep in normal subjects after selective topical oropharyngeal anesthesia. Am Rev Respir Dis. 1987;135:1316–9.
- 138. Basner RC, Ringler J, Garpestad E, Schwartzstein RM, Sparrow D, Weinberger SE, Lilly J, Weiss JW. Upper airway anesthesia delays arousal from airway occlusion induced during human NREM sleep. J Appl Physiol. 1992;73:642–8.
- 139. Berry RB, McNellis MI, Kouchi K, Light RW. Upper airway anesthesia reduces phasic genioglossus activity during sleep apnea. Am J Respir Crit Care Med. 1997;156:127–32.
- 140. Tapia IE, Bandla P, Traylor J, Karamessinis L, Huang J, Marcus CL. Upper airway sensory function in children with obstructive sleep apnea syndrome. Sleep. 2010;33:968–72.
- 141. Wootton DM, Luo H, Persak SC, Sin S, McDonough JM, Isasi CR, Arens R. Computational fuid dynamics endpoints to characterize obstructive sleep apnea syndrome in children. J Appl Physiol (1985). 2014;116:104–12.
- 142. Xu C, Sin S, McDonough JM, Udupa JK, Guez A, Arens R, Wootton DM. Computational fuid dynamics modeling of the upper airway of children with obstructive sleep apnea syndrome in steady fow. J Biomech. 2006;39:2043–54.
- 143. Mylavarapu G, Murugappan S, Mihaescu M, Kalra M, Khosla S, Gutmark E. Validation of computational fuid dynamics methodology used for human upper airway fow simulations. J Biomech. 2009;42:1553–9.
- 144. Vos WG, De Backer WA, Verhulst SL. Correlation between the severity of sleep apnea and upper airway morphology in pediatric and adult patients. Curr Opin Allergy Clin Immunol. 2010;10:26–33.
- 145. Luo H, Sin S, McDonough JM, Isasi CR, Arens R, Wootton DM. Computational fuid dynamics endpoints for assessment of adenotonsillectomy outcome in obese children with obstructive sleep apnea syndrome. J Biomech. 2014;47:2498–503.
- 146. De Backer JW, Vanderveken OM, Vos WG, Devolder A, Verhulst SL, Verbraecken JA, Parizel PM, Braem MJ, Van de Heyning

PH, De Backer WA. Functional imaging using computational fuid dynamics to predict treatment success of mandibular advancement devices in sleep-disordered breathing. J Biomech. 2007;40:3708–14.

- 147. Zhao M, Barber T, Cistulli PA, Sutherland K, Rosengarten G. Simulation of upper airway occlusion without and with mandibular advancement in obstructive sleep apnea using fuidstructure interaction. J Biomech. 2013;46:2586–92.
- 148. Van Holsbeke C, Vos W, Van Hoorenbeeck K, Boudewyns A, Salgado R, Verdonck PR, Ramet J, De Backer J, De Backer W, Verhulst SL. Functional respiratory imaging as a tool to assess upper airway patency in children with obstructive sleep apnea. Sleep Med. 2013;14:433–9.
- 149. Wootton D, Guez A, Vaidyanathan P, McDonough JM, Udupa JK, Arens R. Imaged-based modeling of upper airway resistance in children with obstructive sleep apnea syndrome. APSS. Chicago: Sleep. 2003;26(Suppl):A233.
- 150. Persak SC, Sin S, McDonough JM, Arens R, Wootton DM. Noninvasive estimation of pharyngeal airway resistance and compliance in children based on volume-gated dynamic MRI and computational fuid dynamics. J Appl Physiol. 2011;111:1819–27.
- 151. Wootton DM, Sin S, Luo H, Yazdani A, McDonough JM, Wagshul ME, Isasi CR, Arens R. Computational fuid dynamics upper airway effective compliance, critical closing pressure, and obstructive sleep apnea severity in obese adolescent girls. J Appl Physiol (1985). 2016;121:925–31.
- 152. Sin S, Wootton DM, McDonough JM, Nandalike K, Arens R. Anterior nasal resistance in obese children with obstructive sleep apnea syndrome. Laryngoscope. 2014;124:2640–4.
- 153. Ryan CF, Lowe AA, Li D, Fleetham JA. Three-dimensional upper airway computed tomography in obstructive sleep apnea. A prospective study in patients treated by uvulopalatopharyngoplasty. Am Rev Respir Dis. 1991;144:428–32.
- 154. Carrera M, Barbe F, Sauleda J, Tomas M, Gomez C, Agusti AG. Patients with obstructive sleep apnea exhibit genioglossus dysfunction that is normalized after treatment with continuous positive airway pressure. Am J Respir Crit Care Med. 1999;159:1960–6.
- 155. Friberg D, Ansved T, Borg K, Carlsson-Nordlander B, Larsson H, Svanborg E. Histological indications of a progressive snorers disease in an upper airway muscle. Am J Respir Crit Care Med. 1998;157:586–93.
- 156. Series F, Cote C, Simoneau JA, Gelinas Y, St Pierre S, Leclerc J, Ferland R, Marc I. Physiologic, metabolic, and muscle fber type characteristics of musculus uvulae in sleep apnea hypopnea syndrome and in snorers. J Clin Invest. 1995;95:20–5.
- 157. Young T, Palta M, Dempsey J, Skatrud J, Weber S, Badr S. The occurrence of sleep-disordered breathing among middle-aged adults. N Engl J Med. 1993;328:1230–5.
- 158. Mohsenin V. Gender differences in the expression of sleepdisordered breathing: role of upper airway dimensions. Chest. 2001;120:1442–7.
- 159. Brooks LJ, Strohl KP. Size and mechanical properties of the pharynx in healthy men and women. Am Rev Respir Dis. 1992;146:1394–7.
- 160. Whittle AT, Marshall I, Mortimore IL, Wraith PK, Sellar RJ, Douglas NJ. Neck soft tissue and fat distribution: comparison between normal men and women by magnetic resonance imaging. Thorax. 1999;54:323–8.
- 161. Trinder J, Kay A, Kleiman J, Dunai J. Gender differences in airway resistance during sleep. J Appl Physiol. 1997;83:1986–97.
- 162. Pillar G, Malhotra A, Fogel R, Beauregard J, Schnall R, White DP. Airway mechanics and ventilation in response to resistive loading during sleep: infuence of gender. Am J Respir Crit Care Med. 2000;162:1627–32.
- 163. Martin SE, Mathur R, Marshall I, Douglas NJ. The effect of age, sex, obesity and posture on upper airway size. Eur Respir J. 1997;10:2087–90.
- 164. Zhou XS, Rowley JA, Demirovic F, Diamond MP, Badr MS. Effect of testosterone on the apneic threshold in women during NREM sleep. J Appl Physiol. 2003;94:101–7.
- 165. Ronen O, Malhotra A, Pillar G. Infuence of gender and age on upper-airway length during development. Pediatrics. 2007;120:e1028–34.
- 166. Redline S, Tosteson T, Tishler PV, Carskadon MA, Millman RP. Studies in the genetics of obstructive sleep apnea. Familial aggregation of symptoms associated with sleep-related breathing disturbances. Am Rev Respir Dis. 1992;145:440–4.
- 167. Koskenvuo M, Hublin C, Partinen M, Heikkila K, Kaprio J. Heritability of diurnal type: a nationwide study of 8753 adult twin pairs. J Sleep Res. 2007;16:156–62.
- 168. Mukherjee S, Saxena R, Palmer LJ. The genetics of obstructive sleep apnoea. Respirology. 2018;23:18–27.
- 169. Zhong A, Xiong X, Xu H, Shi M. An updated meta-analysis of the association between tumor necrosis factor-alpha -308G/A polymorphism and obstructive sleep apnea-hypopnea syndrome. PLoS One. 2014;9:e106270.
- 170. Patel SR, Goodloe R, De G, Kowgier M, Weng J, Buxbaum SG, Cade B, Fulop T, Gharib SA, Gottlieb DJ, Hillman D, Larkin EK, Lauderdale DS, Li L, Mukherjee S, Palmer L, Zee P, Zhu X, Redline S. Association of genetic loci with sleep apnea in European Americans and African-Americans: the Candidate Gene Association Resource (CARe). PLoS One. 2012;7:e48836.
- 171. Cade BE, Chen H, Stilp AM, Gleason KJ, Sofer T, Ancoli-Israel S, Arens R, Bell GI, Below JE, Bjonnes AC, Chun S, Conomos MP, Evans DS, Johnson WC, Frazier-Wood AC, Lane JM, Larkin EK, Loredo JS, Post WS, Ramos AR, Rice K, Rotter JI, Shah NA, Stone KL, Taylor KD, Thornton TA, Tranah GJ, Wang C, Zee PC, Hanis CL, Sunyaev SR, Patel SR, Laurie CC, Zhu X, Saxena R, Lin X, Redline S. Genetic associations with obstructive sleep apnea traits in Hispanic/Latino Americans. Am J Respir Crit Care Med. 2016;194:886–97.
- 172. Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, Powell C, Vedantam S, Buchkovich ML, Yang J, Croteau-Chonka DC, Esko T, Fall T, Ferreira T, Gustafsson S, Kutalik Z, Luan J, Magi R, Randall JC, Winkler TW, Wood AR, Workalemahu T, Faul JD, Smith JA, Zhao JH, Zhao W, Chen J, Fehrmann R, Hedman AK, Karjalainen J, Schmidt EM, Absher D, Amin N, Anderson D, Beekman M, Bolton JL, Bragg-Gresham JL, Buyske S, Demirkan A, Deng G, Ehret GB, Feenstra B, Feitosa MF, Fischer K, Goel A, Gong J, Jackson AU, Kanoni S, Kleber ME, Kristiansson K, Lim U, Lotay V, Mangino M, Leach IM, Medina-Gomez C, Medland SE, Nalls MA, Palmer CD, Pasko D, Pechlivanis S, Peters MJ, Prokopenko I, Shungin D, Stancakova A, Strawbridge RJ, Sung YJ, Tanaka T, Teumer A, Trompet S, van der Laan SW, van Setten J, Van Vliet-Ostaptchouk JV, Wang Z, Yengo L, Zhang W, Isaacs A, Albrecht E, Arnlov J, Arscott GM, Attwood AP, Bandinelli S, Barrett A, Bas IN, Bellis C, Bennett AJ, Berne C, Blagieva R, Bluher M, Bohringer S, Bonnycastle LL, Bottcher Y, Boyd HA, Bruinenberg M, Caspersen IH, Chen YI, Clarke R, Daw EW, de Craen AJM, Delgado G, Dimitriou M, Doney ASF, Eklund N, Estrada K, Eury E, Folkersen L, Fraser RM, Garcia ME, Geller F, Giedraitis V, Gigante B, Go AS, Golay A, Goodall AH, Gordon SD, Gorski M, Grabe HJ, Grallert H, Grammer TB, Grassler J, Gronberg H, Groves CJ, Gusto G, Haessler J, Hall P, Haller T, Hallmans G, Hartman CA, Hassinen M, Hayward C, Heard-Costa NL, Helmer Q, Hengstenberg C, Holmen O, Hottenga JJ, James AL, Jeff JM, Johansson A, Jolley J, Juliusdottir T, Kinnunen L, Koenig W, Koskenvuo M, Kratzer W, Laitinen J, Lamina C, Leander K, Lee NR, Lichtner P, Lind L, Lindstrom J, Lo KS,

Lobbens S, Lorbeer R, Lu Y, Mach F, Magnusson PKE, Mahajan A, McArdle WL, McLachlan S, Menni C, Merger S, Mihailov E, Milani L, Moayyeri A, Monda KL, Morken MA, Mulas A, Muller G, Muller-Nurasyid M, Musk AW, Nagaraja R, Nothen MM, Nolte IM, Pilz S, Rayner NW, Renstrom F, Rettig R, Ried JS, Ripke S, Robertson NR, Rose LM, Sanna S, Scharnagl H, Scholtens S, Schumacher FR, Scott WR, Seufferlein T, Shi J, Smith AV, Smolonska J, Stanton AV, Steinthorsdottir V, Stirrups K, Stringham HM, Sundstrom J, Swertz MA, Swift AJ, Syvanen AC, Tan ST, Tayo BO, Thorand B, Thorleifsson G, Tyrer JP, Uh HW, Vandenput L, Verhulst FC, Vermeulen SH, Verweij N, Vonk JM, Waite LL, Warren HR, Waterworth D, Weedon MN, Wilkens LR, Willenborg C, Wilsgaard T, Wojczynski MK, Wong A, Wright AF, Zhang Q, LifeLines Cohort S, Brennan EP, Choi M, Dastani Z, Drong AW, Eriksson P, Franco-Cereceda A, Gadin JR, Gharavi AG, Goddard ME, Handsaker RE, Huang J, Karpe F, Kathiresan S, Keildson S, Kiryluk K, Kubo M, Lee JY, Liang L, Lifton RP, Ma B, McCarroll SA, McKnight AJ, Min JL, Moffatt MF, Montgomery GW, Murabito JM, Nicholson G, Nyholt DR, Okada Y, JRB P, Dorajoo R, Reinmaa E, Salem RM, Sandholm N, Scott RA, Stolk L, Takahashi A, Tanaka T, van't Hooft FM, Vinkhuyzen AAE, Westra HJ, Zheng W, Zondervan KT, Consortium AD, Group A-BW, Consortium CAD, Consortium CK, Glgc, Icbp, Investigators M, Mu TC, Consortium MI, Consortium P, ReproGen C, Consortium G, International Endogene C, Heath AC, Arveiler D, Bakker SJL, Beilby J, Bergman RN, Blangero J, Bovet P, Campbell H, Caulfeld MJ, Cesana G, Chakravarti A, Chasman DI, Chines PS, Collins FS, Crawford DC, Cupples LA, Cusi D, Danesh J, de Faire U, den Ruijter HM, Dominiczak AF, Erbel R, Erdmann J, Eriksson JG, Farrall M, Felix SB, Ferrannini E, Ferrieres J, Ford I, Forouhi NG, Forrester T, Franco OH, Gansevoort RT, Gejman PV, Gieger C, Gottesman O, Gudnason V, Gyllensten U, Hall AS, Harris TB, Hattersley AT, Hicks AA, Hindorff LA, Hingorani AD, Hofman A, Homuth G, Hovingh GK, Humphries SE, Hunt SC, Hypponen E, Illig T, Jacobs KB, Jarvelin MR, Jockel KH, Johansen B, Jousilahti P, Jukema JW, Jula AM, Kaprio J, Kastelein JJP, Keinanen-Kiukaanniemi SM, Kiemeney LA, Knekt P, Kooner JS, Kooperberg C, Kovacs P, Kraja AT,

Kumari M, Kuusisto J, Lakka TA, Langenberg C, Marchand LL, Lehtimaki T, Lyssenko V, Mannisto S, Marette A, Matise TC, McKenzie CA, McKnight B, Moll FL, Morris AD, Morris AP, Murray JC, Nelis M, Ohlsson C, Oldehinkel AJ, Ong KK, Madden PAF, Pasterkamp G, Peden JF, Peters A, Postma DS, Pramstaller PP, Price JF, Qi L, Raitakari OT, Rankinen T, Rao DC, Rice TK, Ridker PM, Rioux JD, Ritchie MD, Rudan I, Salomaa V, Samani NJ, Saramies J, Sarzynski MA, Schunkert H, Schwarz PEH, Sever P, Shuldiner AR, Sinisalo J, Stolk RP, Strauch K, Tonjes A, Tregouet DA, Tremblay A, Tremoli E, Virtamo J, Vohl MC, Volker U, Waeber G, Willemsen G, Witteman JC, Zillikens MC, Adair LS, Amouyel P, Asselbergs FW, Assimes TL, Bochud M, Boehm BO, Boerwinkle E, Bornstein SR, Bottinger EP, Bouchard C, Cauchi S, Chambers JC, Chanock SJ, Cooper RS, de Bakker PIW, Dedoussis G, Ferrucci L, Franks PW, Froguel P, Groop LC, Haiman CA, Hamsten A, Hui J, Hunter DJ, Hveem K, Kaplan RC, Kivimaki M, Kuh D, Laakso M, Liu Y, Martin NG, Marz W, Melbye M, Metspalu A, Moebus S, Munroe PB, Njolstad I, Oostra BA, Palmer CNA, Pedersen NL, Perola M, Perusse L, Peters U, Power C, Quertermous T, Rauramaa R, Rivadeneira F, Saaristo TE, Saleheen D, Sattar N, Schadt EE, Schlessinger D, Slagboom PE, Snieder H, Spector TD, Thorsteinsdottir U, Stumvoll M, Tuomilehto J, Uitterlinden AG, Uusitupa M, van der Harst P, Walker M, Wallaschofski H, Wareham NJ, Watkins H, Weir DR, Wichmann HE, Wilson JF, Zanen P, Borecki IB, Deloukas P, Fox CS, Heid IM, O'Connell JR, Strachan DP, Stefansson K, van Duijn CM, Abecasis GR, Franke L, Frayling TM, MI MC, Visscher PM, Scherag A, Willer CJ, Boehnke M, Mohlke KL, Lindgren CM, Beckmann JS, Barroso I, North KE, Ingelsson E, Hirschhorn JN, RJF L, Speliotes EK. Genetic studies of body mass index yield new insights for obesity biology. Nature. 2015;518:197–206.

173. Shaffer JR, Orlova E, Lee MK, Leslie EJ, Raffensperger ZD, Heike CL, Cunningham ML, Hecht JT, Kau CH, Nidey NL, Moreno LM, Wehby GL, Murray JC, Laurie CA, Laurie CC, Cole J, Ferrara T, Santorico S, Klein O, Mio W, Feingold E, Hallgrimsson B, Spritz RA, Marazita ML, Weinberg SM. Genome-wide association study reveals multiple loci infuencing normal human facial morphology. PLoS Genet. 2016;12:e1006149.