Eco-Epidemiology of Vector-Borne Transmission of *Trypanosoma cruzi* **in Domestic Habitats**

Ricardo E. Gürtler, María del Pilar Fernández, and Marta Victoria Cardinal

Abstract Triatomine species largely differ in their degrees of adaptation to thrive in domestic habitats, blood-feed on humans, and transmit *Trypanosoma cruzi*. Pathogen transmission dynamics are shaped by ecological, biological, and social factors. Here we link housing quality and host availability to the host-feeding patterns of domestic triatomines and examine how their blood-feeding performance affects temperature-dependent vital rates and bug population dynamics. The stability/instability habitat divide connects with the large/small triatomine population size dichotomy and on whether bug population dynamics are density-dependent or density-independent and dominated by stochasticity. Seasonal variations in temperature acting on triatomine blood-feeding activity and human-vector contact rates determined the spring peak of symptomatic acute cases of Chagas disease in northern Argentina across four decades. The presence of domestic animals (dogs, cats, and chickens) and commensal rodents increases domestic infestation, vector infection, and parasite transmission across multiple settings and triatomine species. Both ecological and social factors contribute to human infection risk through social vulnerability, mobility patterns, and housing instability. Understanding the interactions among eco-bio-social factors may lead to the design and implementation of improved, sustainable disease control or elimination strategies.

M. del Pilar Fernández

Instituto de Ecología, Genética y Evolución de Buenos Aires, Consejo Nacional de Investigaciones Científcas y Técnicas, Ciudad Universitaria, Buenos Aires, Argentina

Earth Institute, Columbia University, New York, NY, USA e-mail: mpf2131@columbia.edu

© Springer Nature Switzerland AG 2021 447

R. E. Gürtler $(\boxtimes) \cdot M$. V. Cardinal

Laboratorio de Eco-Epidemiología, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Ciudad Universitaria, Buenos Aires, Argentina

Instituto de Ecología, Genética y Evolución de Buenos Aires, Consejo Nacional de Investigaciones Científcas y Técnicas, Ciudad Universitaria, Buenos Aires, Argentina e-mail: gurtler@ege.fcen.uba.ar[; mvcardinal@ege.fcen.uba.ar](mailto:mvcardinal@ege.fcen.uba.ar)

Laboratorio de Eco-Epidemiología, Facultad de Ciencias Exactas y Naturales, Universidad de Buenos Aires, Ciudad Universitaria, Buenos Aires, Argentina

A. Guarneri, M. Lorenzo (eds.), *Triatominae - The Biology of Chagas Disease Vectors*, Entomology in Focus 5, [https://doi.org/10.1007/978-3-030-64548-9_17](https://doi.org/10.1007/978-3-030-64548-9_17#DOI)

Keywords Habitat suitability · Vector host-feeding patterns · Population dynamics · Parasite transmission · Reservoir hosts · Socio-ecological systems

1 Background

The transmission of zoonotic and vector-borne diseases may be considered an inherently ecological process involving intraspecifc and interspecifc interactions between vectors, pathogens, and host populations (Keesing et al. [2006\)](#page-36-0). When human populations are implicated, pathogen transmission dynamics also involve socioeconomic, cultural, political, psychological, and ethical factors that pertain to the human dimension of disease (Spiegel et al. [2005;](#page-41-0) Briceño-León [2009](#page-30-0); Ellis and Wilcox [2009](#page-33-0)). These factors may be classified as intrinsic or extrinsic to the human population (Ehrenberg and Ault [2005\)](#page-33-1). Intrinsic factors are biological in nature (e.g., immune response) and can only be manipulated by advances in medical research and technology. Extrinsic factors include the environmental context, vector ecology and behavior, human activities, socioeconomic inequalities, and political factors, among others. These factors operate and interact at different scales within a complex system characterized by multiple interdependent components featuring feedback loops and nonlinear relations (Meadows [2008](#page-38-0)).

A thorough understanding of the combined effects of these factors is required to design more effective and sustainable disease control interventions (Charron [2012\)](#page-32-0). Control interventions have traditionally been crafted in a reductionist biomedical approach, which argues that the sum of information provided by separately studying each component of the system is sufficient to understand disease transmission dynamics (El Sayed et al. [2012\)](#page-33-2). Other more integrative approaches (ecohealth or eco-bio-social) focus on the interactions among multiple ecological, biological, and social factors and their combined effects on human health (Spiegel et al. [2005;](#page-41-0) Charron [2012](#page-32-0)). Social factors include large-scale forces such as poverty and social inequality; land tenure and agricultural development; public and private services such as water supply, sanitation, and garbage collection; demographic change and urbanization; vector control programs and other healthcare services; and community- and household-based knowledge, attitudes, and practices. The more integrative approaches pursue the design and implementation of sustainable, cost-effective disease control strategies to reduce social- and gender-associated inequalities related to health (Charron [2012;](#page-32-0) WHO [2008](#page-42-0)).

Chagas disease, caused by the protozoan *Trypanosoma cruzi* and mainly transmitted by triatomine bugs, is a major neglected tropical disease and a serious cause of human chronic disease in the Americas (WHO [2015](#page-42-1)). All triatomine species and mammals appear to be susceptible to the infection. Triatomine bugs are obligate hematophagous insects with opportunistic feeding habits on mammals and birds, and while doing so, they may contaminate their skin with urine or feces and transmit *T. cruzi* to mammalian hosts. The parasite may also be transmitted through food items contaminated by sylvatic triatomine bugs or sylvatic hosts (e.g., opossums). Other transmission routes (vertical, transfusional, organ transplant) exist. Some triatomine species may also transmit other trypanosomatids (e.g., *Trypanosoma rangeli*) to humans and mammals through their bites; *T. rangeli* is considered nonpathogenic for its mammalian hosts.

Here we selectively review the biological, ecological, and social factors involved in the domestic transmission of *T. cruzi* (Fig. [1](#page-2-0)). We frequently provide examples related to the Southern Cone of South America and to its main vector *Triatoma infestans*, partly because this is where Chagas disease attained the greatest prevalence in humans (WHO [2015](#page-42-1)) and partly because of the wealth of information on *T. infestans*. We also examine how ecological and social factors contribute to human infection risk. We defer consideration of vector control measures and their impacts to a separate chapter. Disease-related aspects and spatial dynamics fall outside of the scope of this review.

Fig. 1 Conceptual map of the relations between ecological, biological, and social factors affecting the domestic transmission of *T. cruzi*

2 Biological and Ecological Factors Related to the Vector

2.1 Species and Epidemiologic Relevance

Of the approximately 150 species of Triatominae (Heteroptera: Reduviidae) currently recognized, <10 species have become closely adapted to thrive in domestic premises and feed on humans and other domestic animals (i.e., domesticated), and < 20 species have been involved in the transmission of *T. cruzi* infection to humans (Gourbière et al. [2012](#page-34-0)). Noireau and Dujardin [\(2010](#page-38-1)) divided triatomines in domestic, domiciliary, intrusive, and sylvatic species based on their epidemiological relevance for transmission and control. Although several attempts have been made to classify triatomine species, the subject still remains controversial (e.g., Waleckx et al. [2015a;](#page-41-1) Abad-Franch [2016](#page-29-0)) (see chapter ["Chagas Disease Vector Control](https://doi.org/10.1007/978-3-030-64548-9_18)" on vector control). In this chapter, "domestic or domiciliary" refers to the set of contiguous human sleeping quarters and rooms that share a continuous roof structure.

2.2 Domesticity and Vector Abundance

Domesticity conveys several selective ftness advantages related to stable habitats (more protected from exposure to climatic extremes, such as human dwellings and structurally similar outhouses) with a stable supply of hosts (humans and domestic or synanthropic animals), balanced by the costs of adaptation to diverse types of habitats, blood-feeding on domestic hosts, and progressive genetic simplifcation (Dujardin and Schofeld [2004](#page-33-3)). The immediate gains of domesticity translate into much larger triatomine population sizes than in sylvatic habitats, with additional benefts derived from human-mediated passive dispersal and an enlarged geographic range. Nondomestic triatomine species occupying peridomestic and sylvatic ecotopes are well adapted to cope with habitat instability.

Triatoma infestans expresses the extreme of the evolutionary trend toward domesticity (Dujardin and Schofeld [2004\)](#page-33-3), with widespread sylvatic foci only in Bolivia and Chile (see chapter ["Chagas Disease Vector Control](https://doi.org/10.1007/978-3-030-64548-9_18)"). This dual feature is also shared by *T. dimidiata* in Central America versus Ecuador and by *Rhodnius prolixus* in Venezuela and Colombia versus Central America and southern Mexico. The intimate adaptation of *T. infestans* and *R. prolixus* to domestic habitats and epidemiological signifcance as vectors of human *T. cruzi* infection, combined with other technical aspects, justifed launching regional elimination programs (WHO [2002\)](#page-42-2).

Illustrating their relative success, the top domestic abundances recorded in a single human habitation reached 7900 (Rabinovich et al. [1979](#page-39-0)) to 11,403 *R. prolixus* (Sandoval et al. [2000a](#page-40-0), [b](#page-40-1)) and 8500 *T. infestans* over a 3-year period (Dias and Zeledón [1955\)](#page-33-4). An enclosed experimental population of *T. infestans* in a hut of \sim 1 m³ with one chicken reached 15,844 insects (excluding eggs) 2 years after being

founded with 5 females and 3 males (Cecere et al. [2003](#page-32-1)). *Panstrongylus megistus* and *T. dimidiata* reach much lower densities than *T. infestans* or *R. prolixus*. The greater nutritional value of human blood adds an unappreciated advantage to domesticity (see Sect. [2.5](#page-7-0)).

Domesticity carries with it several implications. The stability/instability habitat divide connects with the large/small population size dichotomy, whether bug population dynamics are density-dependent or density-independent (and dominated by stochastic events such as weather, host death, and exposure to predators) and modify the effectiveness of vector control efforts (Sect. [2.6](#page-12-0) and [2.7\)](#page-12-1).

2.3 Habitat Use and Quality for Triatomines

A unifed defnition of habitat includes "the resources and conditions present in an area that produce occupancy—including survival and reproduction—by a given organism (Hall et al. [1997\)](#page-36-1)." These include physical factors (such as temperature and moisture) and biotic factors (such as the availability of food and shelter and the absence of toxins or predators). This and other defnitions of habitat supersede the notion that it merely represents physical space. Habitat use is considered selective if it is used disproportionately to its availability or, more in point, to its accessibility. Accessibility depends on interspecifc interactions as well as social and environmental factors that might limit access to a given resource (Beyer et al. [2010\)](#page-30-1). Although many triatomine species have characteristic habitat types with associated host species, the mechanisms underlying the observed habitat selection patterns remain mostly unknown.

Each triatomine species selects refuges for optimal conditions of temperature, relative humidity, and darkness at the microhabitat level (Lorenzo and Lazzari [1999;](#page-37-0) Lazzari et al. [2013](#page-36-2)) and marks suitable refuges via assembling factors present in their feces (Lorenzo and Lazzari [1996](#page-37-1)). These factors induced aggregation in nymphs of *T. sordida* and *T. infestans*, thus acting as a pheromone and kairomone (Lorenzo Figueiras and Lazzari [1998\)](#page-37-2). Detailed site-level surveys before and after community-wide insecticide spraying in two rural areas of the Argentine Chaco have shown that sympatric triatomine species tended to occupy distinct peridomestic habitats (e.g., *Triatoma garciabesi* versus *T. guasayana* or *T. infestans*) or were positively associated both at site and house level (*T. sordida* and *T. infestans*) (Rodríguez-Planes et al. [2016](#page-40-2), [2018\)](#page-40-3). However, none of the secondary species were able to establish colonies and persist in domestic premises after the quasi-elimination of *T. infestans* despite recurrent invasion events (e.g., Rojas de Arias et al. [2012;](#page-40-4) Rodríguez-Planes et al. [2020](#page-40-5)).

Most assessments of habitat use have been derived from the frequency distribution of occurrence or relative abundance of triatomine bugs in feld surveys. This information was used to rank the suitability of multiple habitats for a given species and to identify which of them may function as key sources of triatomines for house reinfestation after control interventions. For example, initial assessments of habitat suitability for *T. infestans* in the Argentine Chaco were accomplished via surveys of habitat-specifc infestation and bug abundance. These surveys suggested that chicken coops, goat corrals, and pig corrals were the most important ecotopes (López et al. [1999](#page-37-3); Cecere et al. [2004](#page-32-2), [2006](#page-32-3), [2013](#page-32-4); Gürtler et al. [2004](#page-35-0); Ceballos et al. [2005](#page-32-5)). However, a broader survey measuring several ftness-related components in more habitat types showed that chicken coops and human sleeping quarters were relatively more important while goat corrals were at the other extreme (Gürtler et al. [2014b\)](#page-35-1). These ftness components (sex ratio, female fecundity, nutritional status, and vector abundance) were in theory better descriptors of habitat suitability. Bug abundance and fecundity were aggregated across ecotopes, suggesting that bug population growth was concentrated in a few productive, high-quality sites.

These patterns are consistent with the 80-20 rule, by which approximately 80% of the vectors or disease cases would occur in 20% of the sample sites (Woolhouse et al. [1997\)](#page-42-3). Using a different theory and methods, Taylor's law of fuctuation scaling described accurately the mean and variance of the habitat-specifc abundances of four triatomine species in the Argentine Chaco, with slopes indicating spatial aggregation or variation in habitat suitability (Cohen et al. [2017](#page-32-6)). Both spatial aggregation and variation in habitat suitability are relevant for improved vector control and surveillance. Extending these approaches to other triatomine species and settings may provide new perspectives on their high-quality habitats.

One dimension of habitat quality relates to physical structure. Housing design and construction materials combined with their degree of maintenance determine the availability of refuges for domestic bugs, such as cracked walls, tiled or thatched roofng, and earthen foors (Dumonteil et al. [2013;](#page-33-5) Bustamante et al. [2014](#page-31-0)), affect the susceptibility of a house to bug invasion and subsequent colonization (Monroy et al. [2009\)](#page-38-2), and may modify the effectiveness of insecticide applications (Gürtler et al. [2004;](#page-35-0) Cecere et al. [2013\)](#page-32-4). The surface structure of indoor walls (unplastered or with many crevices) and the existence of thatched roofs were signifcant predictors of domestic infestation and abundance of the main domestic vectors of *T. cruzi* (Cecere et al. [1998;](#page-32-7) Mott et al. [1978a;](#page-38-3) Rojas de Arias et al. [1999;](#page-40-6) Andrade et al. [1995\)](#page-30-2).

Experimental evidence showed that bug population size increased steadily with refuge availability over a 2-year period, with two out of three bug populations in low-refuge huts becoming extinct (Cecere et al. [2003](#page-32-1)). However, the exact way in which housing quality affects domestic infestation may vary between triatomine species. Dirt floors are highly important for *T. dimidiata* as it uses dirt for camoufage (Zeledón and Vargas [1984](#page-42-4); Bustamante et al. [2014](#page-31-0)) but appear to be of little utility to *T. infestans*, which lacks a camoufage behavior. Dirt foors are indicators of high social vulnerability and frequently are associated with other household features favoring house infestation and triatomine abundance, e.g., mud walls and overcrowding (Fernández et al. [2019a](#page-34-1)). Window screens greatly reduced the domestic invasion of *T. dimidiata* in Yucatan, Mexico (Waleckx et al. [2015b](#page-41-2)). Peridomestic structures also provide numerous shelters, as in chicken houses and animal enclosures built with mud, wood, rock, or piled thorny shrubs (Carcavallo et al. [1999;](#page-31-1) Cecere et al. [2004](#page-32-2); Walter et al. [2007;](#page-41-3) López et al. [1999;](#page-37-3) Diotaiuti et al. [2000;](#page-33-6) Gurevitz et al. [2013;](#page-35-2) Ramsey et al. [2003](#page-40-7)). If bug-proof chicken houses replaced the

current structures used in northeast Argentina, *T. infestans* and *T. sordida* would lose a key productive habitat (Gurevitz et al. [2013](#page-35-2); Rodriguez-Planes et al. [2018](#page-40-3)). In general, peridomestic structures function as host and triatomine breeding sites and serve as sources of insects that invade human habitations (Cecere et al. [2004,](#page-32-2) [2006\)](#page-32-3).

2.4 Host Availability

Host availability is quantifed by the local abundance of avian or mammalian hosts. However, a host may be available but not accessible if host-seeking triatomines cannot actually feed on it because of some actual or virtual barrier (e.g., bednets, repellents, distance). In domestic habitats, the most frequent hosts of triatomine bugs are humans, dogs, cats, chickens, and rodents. Domestic bug abundance increased with increasing numbers of human residents in rural communities infested with *T. infestans*, *P. megistus*, *R. prolixus*, or *T. dimidiata* (Marsden et al. [1982;](#page-37-4) Piesman et al. [1983;](#page-39-1) Gurevitz et al. [2011](#page-35-3); Campbell-Lendrum et al. [2007\)](#page-31-2). Moreover, the presence of indoor-resting chickens and dogs correlated positively with domestic bug abundance across settings and species (e.g., Lardeux et al. [2015;](#page-36-3) Dumonteil et al. [2013;](#page-33-5) Bustamante et al. [2009,](#page-31-3) [2014;](#page-31-0) Cecere et al. [1998](#page-32-7); Gurevitz et al. [2011\)](#page-35-3). These relationships tend to be consistent with the outcome of domestic triatomines' blood meal surveys (Sect. [2.5\)](#page-7-0).

Host demography and domestic animal management practices add another dimension to the links between host availability/accessibility, vector abundance, and socioeconomic position, as the habits of keeping chickens indoors for protection and letting domestic dogs wander freely or share domestic premises are frequent in rural areas under a subsistence economy. For example, in typical rural villages of the Argentine Chaco, seasonal breeding pulses determine a surge of host numbers with the appearance of juveniles and their enhanced exposure to triatomines. The occurrence of indoor-nesting chickens peaked in spring and decreased during the hot summer months toward reaching a minimum over fall-winter (Cecere et al. [1997](#page-32-8)). Permanent chicken coops were rare and nesting sites relatively instable over time; in the absence of the host, the starved bugs presumably dispersed and generated new foci. The household number of chickens varied widely over time depending on household acquisition, consumption, and disease outbreaks (Rodríguez-Planes et al. [2018\)](#page-40-3). In goat corrals, kids kept enclosed for protection were continually exposed to triatomines, while adult goats were allowed to roam and forage the little grass available in the dry Chaco (Ceballos et al. [2005\)](#page-32-5). The population size of domestic dogs remained stable from year to year though with high turnover rates (Gürtler et al. [1990\)](#page-35-4). These fne-scale spatial and temporal heterogeneities affect triatomine vital rates and population size.

Host behavior patterns also affect host accessibility. Although household size varies little from year to year, in practice householders exposed to hot weather or nuisance pests move their beds and hammocks to open air as an adaptive response (see Sect. [2.5](#page-7-0)) and thus may become less accessible to domestic triatomines (Rabinovich [1985;](#page-39-2) Gürtler et al. [1997](#page-35-5); Brenière et al. [2017](#page-30-3)). In doing so, however, householders may become more exposed to peridomestic or sylvatic vectors such as *R. pallescens*. Households also differed in letting domestic dogs and cats roam freely for all or part of their food. Dogs have a marked crepuscular free-ranging behavior, and male dogs wander in small packs at night during estrus periods (Matter and Daniels [2000\)](#page-37-5). In rural areas throughout Latin America, it is quite common that dogs are not neutered. These host activity patterns may reduce the likelihood of host-vector encounters. Thus, scoring the actual presence or absence of domestic hosts (past and current) presents special challenges. Site-level occupancies may vary widely over time depending on weather, host behavior, biting insects, and cultural patterns. Triatomines may respond fast to the sudden absence of a host and actively disperse (Castillo-Neyra et al. [2015](#page-31-4)).

2.5 Blood-Feeding Performance

Host-Feeding Patterns The host-feeding patterns of Triatominae are key to understanding the eco-epidemiology of Chagas disease. Most species of Triatominae show eclectic blood-feeding patterns on birds and mammals (Wisnivesky-Colli [1987;](#page-42-5) Rabinovich et al. [2011\)](#page-39-3), with very few triatomine species displaying a remarkable specifcity for a host species (e.g., *C. pilosa* for bats in caves) (Dujardin and Schofeld [2004\)](#page-33-3). Some triatomine species may occasionally feed on reptiles, on amphibians, and on insect hemolymph. The small nymphs may even feed on bloodengorged conspecifcs (cleptohematophagy). In domestic habitats, the main blood meal sources of triatomine bugs are humans, dogs, chickens, and cats. House mice and rats are also a relevant blood meal source for domestic populations of *T. dimidiata* and *Triatoma barberi* (De Urioste-Stone et al. [2015](#page-32-9)).

The species of Triatominae with signifcant public health relevance are those that feed on humans or which may contaminate foodstuffs. Table [1](#page-8-0) identifes 24 species of Triatominae with documented evidence of having fed on humans as determined by immunologic methods, estimated from the data compiled by the most recent review on triatomine blood-feeding patterns (Rabinovich et al. [2011\)](#page-39-3). From the 159 data sets, we selected those that included any species collected in domestic habitats only, or in domestic or peridomestic habitats only, and excluded all other habitat sources. The selected list includes 68 studies and 28,054 specimens tested.

The degree of human-vector contact, measured by the average human blood index (i.e., the fraction of tested insects having a human blood meal), peaked in *R. prolixus* (79.2%), *R. pallescens* (65.0%), *T. pallidipennis* (44%), *T. infestans* (40.3%), and *P. megistus* (36.6%). A second group of species had a mean human index between 10% and 20% (*P. herreri*, *T. barberi*, *R. pictipes*, *T. vitticeps*, *R. ecuadoriensis*, *T. dimidiata*, *T. brasiliensis*, and *T. pseudomaculata*). Chickens ranked high, followed by dogs and cats. The rodent blood index averaged 17.1% across all species in the list and frequently exceeded 40%. Even sylvatic populations of

Table 1 Host-feeding patterns of Triatominae collected in domestic or peridomestic habitats **Table 1** Host-feeding patterns of Triatominae collected in domestic or peridomestic habitats

T. infestans had human blood meals (Buitrago et al. [2013](#page-31-5), [2016](#page-31-6)), as did sylvatic *Mepraia spinolai* (Brenière et al. [2017](#page-30-3)) and *T. sanguisuga* (Waleckx et al. [2014\)](#page-41-4). Many other species have been reported to attack humans in the wilderness, such as *R. brethesi*, *T. guasayana*, and *T. brasiliensis*. In summary (i) many triatomine species other than the main domestic vectors blood-feed on or may attack humans under the appropriate circumstances; (ii) the rates of human-vector contact largely differed between and within species, habitats, and settings; (iii) the recorded patterns are fraught with a putative selection bias, as there is sparse information for the great majority of triatomine species; and (iv) mixing domestic and peridomestic bug collections defates human blood indices and increases chicken blood indices, for example. The fact that triatomine host-feeding patterns are seldom related to local host numbers (i.e., at site or house level), host, and vector infection hampers the full understanding of system dynamics.

The host-feeding choices of hematophagous insect vectors are affected by the host species composition in a given habitat, relative host abundance, host proximity, and host defensive behavior (Lehane [2005\)](#page-36-4). Host proximity has usually been considered more important than any intrinsic host preference for a host-seeking bug (Minter [1976\)](#page-38-4). However, there were very few controlled host choice experiments in Triatominae. For *T. sordida* (typically associated with birds), frst-instar nymphs signifcantly preferred birds to humans (Rocha e Silva et al. [1977](#page-40-8)), whereas the feeding success and blood meal size of ffth-instar nymphs were signifcantly larger on guinea pigs than on pigeons (Crocco and Catalá [1997](#page-32-10)). In laboratory-based trials, four caged vertebrate host species were simultaneously exposed to separate groups of ffth-instar nymphs of *T. infestans*, *T. dimidiata*, and *R. prolixus* (Jirón and Zeledón [1982](#page-36-5)); none of these species displayed defnite host-feeding preferences among dogs, chickens, and opossums, but toads were rarely selected. Dogs were highly preferred over chickens or cats in host choice experiments conducted in small huts where the released bugs could choose to feed on any of the two host species available (Gürtler et al. [2009](#page-35-6)). Bugs that fed on dog engorged signifcantly more than bugs that fed on chicken or cat, suggesting dogs were more tolerant to bites than chickens or cats. These results were mainly consistent with feld hostfeeding patterns.

Host defensive behavior in response to triatomine bites determines the selective utilization of some host species and individuals (Kelly and Thompson [2000\)](#page-36-6). Several triatomine species displayed negative density-dependent engorgement rates on non-anesthetized, unrestrained small chickens, pigeons, and rodents in laboratory settings (Crocco and Catalá [1997](#page-32-10); Rabinovich [1985;](#page-39-2) Piesman et al. [1983;](#page-39-1) Schofield [1982\)](#page-40-9). In the host choice trial described above, high vector densities tended to increase the shifts between hosts and signifcantly increased post-exposure bug weight though not to a large extent (Gürtler et al. [2009](#page-35-6)).

These basic processes combined with the rather limited dispersal range of Triatominae (especially of nymphs) determine that their host-feeding patterns tend to be spatially structured according to habitat type and correlate closely with the main local resident host(s). In rural villages in northwestern Argentina, the main or only blood meal sources of *T. infestans* in chicken coops, goat corrals, and pig

corrals were chickens, goats, and pigs, respectively (Gürtler et al. [2014b](#page-35-1)). The domestic populations of *T. infestans* displayed seasonal variations in feeding patterns related to the shifting resting sites of hosts during the hot season (Gürtler et al. [1997\)](#page-35-5). In spring-summer domestic bug collections, the human blood index decreased substantially with the increasing presence of chickens or dogs indoors and also decreased with increasing domestic bug abundance, refecting changes in human exposure. Moreover, the dog blood index increased signifcantly with increasing numbers of dogs and with domestic bug abundance and decreased as the chicken blood index rose. The presence of indoor-resting chickens correlated positively with domestic bug abundance and with an increasing chicken blood index (Cecere et al. [1997\)](#page-32-8). Underlying these patterns are the opportunistic nature of host choice (expressed in high rates of mixed blood meals on different host species) and the spatiotemporal variations in host availability and accessibility. The inverse relationship between the human and chicken blood indices in domestic *T. infestans* was verifed in at least two other rural settings in northern Argentina (Gürtler et al. [2014a](#page-35-7); Ordóñez-Krasnowski et al. [2020\)](#page-39-4). Host shifts affected domestic transmission (see Sect. [3.6](#page-19-0)).

Host Blood and Fitness The question on whether host blood type affects the ftness of triatomines has implications for understanding domesticity, population growth, and transmission risks. Host blood effects are well known in mosquitoes and tsetse fies (Lehane [2005\)](#page-36-4) and affected the vital rates and engorgement levels of triatomines (Gardiner and Maddrell [1972;](#page-34-2) Núñez and Segura [1987\)](#page-38-5). Human blood was better than sheep blood for vitellogenesis, general metabolism, and development in *R. prolixus* (Valle et al. [1987](#page-41-5)). Cohorts of *R. prolixus* that fed artifcially on citrated blood during their entire lives in the same artifcial environment achieved faster development rates and much larger blood meal size, body weight, and female fecundity when fed on human or rabbit blood rather than when bugs fed on chicken, sheep, or horse blood (Gomes et al. [1990](#page-34-3)). Avian blood usually has much lower hemoglobin and plasma protein than the blood of clinically healthy mammals and hence appears to be nutritionally inferior. However, the lower blood viscosity of avian blood allows increased ingestion rates and would reduce the risk of hostinduced death (Lehane [2005](#page-36-4)). Goat (or sheep or cow) blood reduced the feeding effciency of *T. brasiliensis* because triatomines are unable to agglutinate cattle erythrocytes (Araujo et al. [2009\)](#page-30-4). Gardiner and Maddrell [\(1972](#page-34-2)) reported that *R. prolixus* bugs tended to blood-feed much less or not at all on adult goats or sheep that had been exposed repeatedly to bites (i.e., developed acquired resistance) and hence laid fewer eggs. This suggested that the rather large feld populations of *T. infestans* in goat corrals most likely thrived at the expense of the immunologically naïve kids (Gürtler et al. [2004,](#page-35-0) [2017](#page-36-7)). When suitable hosts are not available, goat corrals become sources of dispersing triatomines as their nutritional state declines.

Blood-Feeding Rate and Blood meal Size These parameters affect the vital rates of triatomine bugs, including their propensity to initiate fight, the regulation of local population size (Schofield [1994](#page-41-6)), and parasite transmission rates. There are

few estimates of the blood-feeding rates, blood meal sizes, and nutritional status of Triatominae in feld settings (reviewed in Gürtler et al. [2014b](#page-35-1)). Preliminary estimates of the feeding rates of domestic *T. infestans* and *R. prolixus* based on the distributions of body weight and body length yielded 5–8 to 5–20 days, respectively (Schofeld [1980](#page-40-10); Rabinovich et al. [1979\)](#page-39-0). By measuring the temperature-adjusted occurrence of transparent urine assessed shortly after capture in experimental chicken coops, the blood-feeding rate of *T. infestans* proved to be temperature- and bug density-dependent (Catalá [1994\)](#page-31-7). Domestic populations of *T. infestans* bloodfed every 3–4 days over the spring-summer period (Catalá et al. [1997;](#page-31-8) Gürtler et al. [2014a](#page-35-7)). Mean feeding intervals varied widely across ecotopes and peaked in chicken coops (López et al. [1999](#page-37-3); Ceballos et al. [2005](#page-32-5); Gürtler et al. [2014b\)](#page-35-1). Some triatomine species may feed every 2 days in the insectary and withstand prolonged starvation over several months.

The only feld estimate of blood meal size comes from experimental chicken houses stationed outdoors (Catalá [1994\)](#page-31-7). The total blood meal contents of recently fed triatomines may be used as a proxy of blood meal size under assumed steadystate conditions (Gürtler et al. [2017\)](#page-36-7). Human- and chicken-fed *T. infestans* had signifcantly larger blood contents than bugs fed on other hosts, whereas goat-fed bugs ranked last, in consistency with their average blood-feeding rates. Female fecundity was also maximal in chicken-fed bugs from chicken coops and minimal in goat-fed bugs. The greater blood-feeding performance and nutritional status of bugs from chicken coops, closely followed by domestic bugs, refected in their having a larger body length than other peridomestic bugs across most life stages – a clear indication of habitat-associated ftness advantages.

Timing of Defecation Blood meal size determines the timing of defecation and hence the chance of eventual skin contamination with triatomine feces (Kirk and Schofeld [1987](#page-36-8); Trumper and Gorla [1991\)](#page-41-7). Blood meal size and the timing of defecation were both inversely density-dependent. As host irritation increases, its defensive reactions lead to interrupted (smaller) blood meals and prolong the time to the frst fecal drop. These mechanisms supported the hypothesis that the greatest chances of transmission of *T. cruzi* would occur at low bug densities. However, it does not necessarily follow from the above experiments that a feeding contact with a single bug at high bug population density has a lower risk of transmitting infection to an uninfected mammalian host than a feeding contact with a single bug at a low bug population density (Cohen and Gürtler [2001\)](#page-32-11). The time needed to secure a replete blood meal and the timing of defecation differed substantially among the main triatomine vectors (Zeledón et al. [1977\)](#page-42-6); both *T. infestans* and *R. prolixus* defecated during or shortly after blood-feeding while on the host. Although the North American species of Triatominae were once considered poor vectors based on their long defecation times, subsequent studies revealed that several of them would be effcient vectors (Zeledón et al. [2012\)](#page-42-7).

2.6 Environmental Variables

Temperature and precipitation affect the geographic distribution of Triatominae (Gorla and Noireau [2017\)](#page-34-4), their host-seeking and metabolic rates, body size, and vital rates. Before the onset of large-scale insecticide spraying campaigns in Argentina during the 1960s, house infestation with *T. infestans* occurred in areas where daily maximum temperatures exceeded 20 \degree C and vapor-pressure deficits were greater than 1100 hectopascals over at least 220 days a year (Curto de Casas et al. [1999\)](#page-32-12). The minimum temperature of the coldest month apparently defned the geographic range of *T. infestans* and *R. prolixus*, the latter being also infuenced by precipitation-related variables (Medone et al. [2015](#page-38-6)). At fner scales, triatomines are affected by variable microsite conditions. Different species display specifc patterns of thermopreference and hygropreference that vary substantially over the feeding and daily cycles and affect refuge selection (Lazzari et al. [2013\)](#page-36-2). For example, both egg hatching and molting success are severely reduced by very low relative humidity.

Environmental variables interact with habitat structure to generate heterogeneous microsite conditions. The domestic and peridomestic habitats of *T. infestans* damped external temperatures to different extents depending on their physical structure, ranging from a minimum in thorn shrub-fenced goat corrals to a maximum in typical mud-and-thatch human habitations (Vazquez-Prokopec et al. [2002](#page-41-8)). In general, bugs from animal enclosures with little capacity to dampen climatic extremes are exposed to increased risks of hyperthermia and desiccation, which likely trigger dispersal. Large swings in diurnal temperature prolong insect development times, reduce survival and female fecundity relative to constant temperatures (Nijhout et al. [2014\)](#page-38-7), and reduce body size when temperatures reach stressful levels (Colinet et al. [2015\)](#page-32-13). In contrast, goat corrals with thick fences made of piled shrubs and goat dung have a large damping capacity and are able to sustain large triatomine populations despite harsh winters (Schofeld [1985](#page-41-9)). Thus, the exact physical structure of the habitat rather than its main function or host determine whether local conditions are suitable for triatomines.

2.7 Population Dynamics and Vital Rates

Nearly all the available information on triatomine survival, fecundity, and development rates comes from laboratory-reared cohorts held under optimal, constant conditions (e.g., Rabinovich [1972](#page-39-5); Perlowagora-Szumlewicz [1975;](#page-39-6) Rabinovich and Feliciangeli [2015](#page-39-7)). Triatomine populations are stage-structured and have long life cycles (from 4–6 to 12 months or more). The most realistic approximations to feld settings were provided by experimental bug populations kept in huts that mimicked typical rural houses exposed to the temperate climate of central Argentina. The huts were caged with mosquito netting to prevent in- or out-migration and housed either two or four chickens in structurally homogeneous huts (Gorla and Schofeld [1989;](#page-34-5)

Gorla [1991](#page-34-6)) or one chicken under a three-level gradient of refuges (Cecere et al. [2003\)](#page-32-1).

In these experimental systems, the population abundance of *T. infestans* fuctuated seasonally following temperature variations and peaked once in early- or midsummer, as in rural houses from Brazil (Dias [1955](#page-33-7); Schofeld [1980\)](#page-40-10). These populations displayed two peaks of adult emergence of differing intensity (Cecere et al. [2003;](#page-32-1) Gorla [1991\)](#page-34-6) and peak numbers of eggs per female lagging by 1 month. During spring-summer, fecundity averaged approximately three eggs per female per day in huts with two to four chickens (Gorla and Schofeld [1989](#page-34-5)) and in huts with maximum refuge availability and one chicken (Cecere et al. [2003\)](#page-32-1). These estimates are lower than the average fecundity of *T. infestans* (4.07 eggs per female per day) fed twice a week and kept under optimal conditions (Núñez and Segura [1987](#page-38-5)). Both female fecundity and nymphal development rates steadily increased with increasing temperature above 16 °C (the threshold for development), whereas mortality was mainly determined by monthly mean minimum temperatures (Gorla [1992\)](#page-34-7). A stagestructured stochastic model of the population dynamics of *T. infestans* described well the temperature-dependent seasonal variations in bug abundance, stage structure, and the two peaks of female fecundity rates and total egg numbers observed in hut experiments (Castañera et al. [2003\)](#page-31-9).

In the experimental huts, the egg-to-adult mortality of *T. infestans* ranged from ~98.5% (Gorla and Schofeld [1989](#page-34-5)) to 94.8–97.6% (Cecere et al. [2003\)](#page-32-1) and exceeded a preliminary estimate for domestic *T. infestans* (86.3%) based on the recovery of dead bugs and exuviae (Schofeld [1980\)](#page-40-10). In one of the studies (Cecere et al. [2003\)](#page-32-1), a founder female had a maximum life span of 20 months, and the average for adult males (7.8) and females (5.6) exceeded other estimates of 4–5 months (Gorla and Schofeld [1989](#page-34-5)). Bug mortality was density-independent, and female fecundity was weakly density-dependent (Gorla and Schofeld [1989](#page-34-5); Gorla [1991\)](#page-34-6). Domestic populations of *R. prolixus* also failed to display density-dependent mortality (Rabinovich [1985\)](#page-39-2). Chickens most likely were a refuge-dependent mortality factor: both nymphs and adults from huts with plastered walls had lower survival rates than bugs from maximum-refuge huts (Cecere et al. [2003\)](#page-32-1). In the absence of migration, the net reproductive rate (R_0) decreased steadily from 3.91 to 0.25 in maximum- to minimum-refuge huts, respectively (Cecere et al. [2003](#page-32-1)). Schofeld [\(1980](#page-40-10)) estimated $R_0 = 8.7$ by combining laboratory and field data for domestic *T. infestans.* Rabinovich ([1972\)](#page-39-5) estimated $R_0 = 25.04$ for laboratory-reared cohorts of *T. infestans* having very large survivorship and very low fecundity. These estimates of R_0 reflect the species' large capacity for establishing new foci and for population recovery following insecticide treatment.

Insecticides are a major source of bug mortality even when they are applied by householders. In the absence of government-sponsored insecticide campaigns, domestic bug abundance was negatively related to the domestic application of insecticides carried out by householders (Gurevitz et al. [2011;](#page-35-3) Gaspe et al. [2015\)](#page-34-8). With increasing access to insecticides, especially in agricultural settings, the population size of domestic triatomines may rarely reach the typical high levels recorded in the past at which density-dependent effects on vital rates may set in. Rather,

domestic bug populations may be held at the lower limits imposed by insecticide use, housing improvements, and environmental stochasticity. These smaller bug populations occupying discrete patches connected by frequent active dispersal (zu Dohna et al. [2009](#page-42-8)), subject to stochastic events, may dominate the current and future eco-epidemiological scene.

Dispersal plays a key role in the invasion of domestic premises and establishment of new colonies, the recovery of bug population size after control interventions, and the spatial structure of triatomine populations. It may also contribute to the regulation of local population size, as suggested by the density-dependent loss of marked adult *R. prolixus* from domestic premises (Rabinovich [1985\)](#page-39-2). In contrast, fight initiation was inversely related to *T. infestans* density in a laboratory setting (McEwen et al. [1993](#page-38-8)), suggesting that other processes may be implicated (e.g., pheromone-mediated searches for mates).

The dispersal of triatomine bugs is accomplished by active (walking, fight) or passive means via carriage in clothes, luggage, and frewood or as eggs stuck to the feathers of some birds (Schofeld [1994\)](#page-41-6). Passive transport of domestic triatomines is always a possibility, as shown by the fnding of a *T. infestans* adult further south of its historical geographic range (Piccinali et al. [2010\)](#page-39-8). Flight dispersal is limited to a few 100 m up to a few km, whereas passive dispersal has been associated with the long-distance range expansion of *T. infestans* to northeast Brazil, *R. prolixus* to Central America, and *T. dimidiata* to Ecuador. Mark-recapture studies demonstrated an intense exchange of nymphs and adults of domestic *R. prolixus* between houses located 100 and 500 m apart, consistent with its well-known fight capacity (Rabinovich [1985\)](#page-39-2). The fight range of *T. infestans* (considered a poor fier) may exceed 2400 m, as suggested by sustained tethered fights at speeds of 2 m/s for at least 20 min (Ward and Baker [1982\)](#page-41-10). Walking dispersal of nymphs and adult bugs may play a substantial role in establishing new foci at fner scales and contribute to the spatial aggregation of infestation (Vazquez-Prokopec et al. [2006;](#page-41-11) Abrahan et al. [2011\)](#page-30-5). It may also explain the fnding of human-fed triatomines in nearby peridomestic habitats not used as human resting sites (Cecere et al. [1997;](#page-32-8) Gürtler et al. [2014b\)](#page-35-1).

Flight dispersal invariably includes unfed triatomines and occurs within the frst 2–3 h after sunset in most of the species investigated (Di Iorio and Gürtler [2017\)](#page-33-8). The initiation of fights is usually triggered by starving conditions (refected in low body weight-to-body length ratios) and mean temperatures in the range from 20 to 30 °C. The duration and detailed time structure of the dispersal season are relevant because they determine subsequent establishment events and eventual human exposures. Analysis of a long time series of site infestations in a rural village suggested increased dispersal of *T. infestans* during spring and a 6-month lag between a new bug establishment on a site and dispersal from this site (zu Dohna et al. [2009](#page-42-8)).

Artifcial light sources usually attract adult triatomines of many species and favor house invasion. Public streetlights were positively and signifcantly associated with domestic infestation with *T. dimidiata* in Yucatan, Mexico (Pacheco-Tucuch et al. [2012\)](#page-39-9). When the streetlight posts of a small village in western Argentina were systematically inspected for triatomines between sunset and midnight over spring-summer, the occurrence of fight-dispersing triatomines (from four species) steadily increased between 16.6 and 31.7 °C, suggesting a putative temperature threshold for fight initiation at 17–18 °C (Di Iorio and Gürtler [2017](#page-33-8)). Although the catch of adult *T. infestans* at the streetlight posts was sex-independent, that of *T. guasayana* was strongly male-biased – a pattern that has been recorded elsewhere and in other triatomine species. In an experimental setting, male *R. prolixus* bugs increased substantially their takeoff activity in response to female pheromones, but the reverse did not occur (Zacharías et al. [2010\)](#page-42-9). This suggests that a colonizing, unfertilized female triatomine may be able to recruit fight-dispersing males and thus increase the chances of establishing a viable bug colony. However, fight dispersal may also be female-biased, as suggested by feld observations of individually marked adult *P. megistus* (Forattini et al. [1977\)](#page-34-9) or *T. infestans* in open chicken coops and by microsatellite-based genetic studies (reviewed in Gürtler et al. [2014b\)](#page-35-1). Sexbiased fight dispersal may contribute to imbalanced adult sex ratios across ecotopes (Payet et al. [2009\)](#page-39-10).

3 Biological and Ecological Factors Related to Parasite Transmission

3.1 Parasite Diversity

Trypanosoma cruzi (Kinetoplastida: Trypanosomatidae) has a genetically diverse clonal structure classifed into six genotypes (TcI-TcVI) denominated discrete typing units, or DTUs (Zingales et al. [2012\)](#page-42-10). Whether TcBat, a recently described genotype mostly restricted to bats, is a different DTU is still under debate (Marcili et al. [2009;](#page-37-6) Zingales [2018](#page-42-11)). All DTUs are capable of infecting humans and mammals, and their geographical distribution and frequency of occurrence vary widely across the Americas (reviewed by Brenière et al. [2016\)](#page-30-6). Humans and > 150 species of nonhuman mammalian hosts have been found naturally infected with *T. cruzi*; other vertebrates are refractory to the infection (Jansen et al. [2017](#page-36-9)).

Transmission Cycles Domestic and sylvatic habitats sustain two main types of transmission cycles (Miles et al. [2003\)](#page-38-9). Sylvatic cycles involve sylvatic mammals and sylvatic triatomine species, whereas domestic cycles mainly include domestic triatomines, humans, and domestic or synanthropic animals. The two archetypical sylvatic cycles across the Americas are an arboreal cycle involving didelphid marsupials and TcI and a terrestrial cycle involving armadillos and TcIII (Yeo et al. [2005;](#page-42-12) Brenière et al. [2016](#page-30-6)). However, these associations are not absolute. TcI has the broadest distribution from southern United States to Argentina and Chile. It has been mainly found in sylvatic cycles across the Americas and in domestic cycles to the north of the Amazon basin. TcII has been mainly associated with domestic cycles but has also been isolated from sylvatic mammals. TcV and TcVI predominate

in domestic cycles across the Southern Cone countries. TcBat, TcIII, and TcIV have been mainly found in sylvatic cycles.

The degree of connectivity or overlap between domestic and sylvatic cycles may affect disease control and elimination efforts. If transmission cycles overlap, the introduction of sylvatic parasites may threaten efforts directed at curtailing domestic transmission, as recorded in Mexico (Ramsey et al. [2012](#page-40-11)). Overlapping transmission cycles of TcI and TcVI were also recorded in Yucatan, Mexico (López-Cancino et al. [2015](#page-37-7)), Guatemala (Pennington et al. [2015\)](#page-39-11), and Venezuela, where *Rhodnius* bugs infested both houses and palm trees (Miles et al. [2003\)](#page-38-9). A classic example of separate transmission cycles occurred in Bahia, Brazil, where TcII circulated in houses infested with *P. megistus* while TcI circulated between *T. tibiamaculata* and *Didelphis albiventris* opossums in bromeliad epiphytes (Miles et al. [2003\)](#page-38-9). Separate transmission cycles occurred in the Argentine Chaco region, where TcV/TcVI predominated in domestic habitats while TcI and TcIII were restricted to sylvatic hosts (Cardinal et al. [2008;](#page-31-10) Diosque et al. [2003](#page-33-9); Macchiaverna et al. [2015,](#page-37-8) [2018;](#page-37-9) Orozco et al. [2013;](#page-39-12) Enriquez et al. [2013;](#page-33-10) Lucero et al. [2016\)](#page-37-10).

3.2 Domestic Reservoir Hosts

Nonhuman Reservoir Hosts Dogs, cats, rodents, and domesticated guinea pigs are major domestic nonhuman reservoir hosts of *T. cruzi* (reviewed in Gürtler and Cardinal [2015](#page-35-8))*.* They are able to maintain *T. cruzi* in the absence of any other host species and play key roles as amplifying hosts and parasite sources in many domestic or peridomestic transmission cycles across ecoregions and triatomine species. House mice and rats contributed to domestic bug infection with *T. cruzi* in many settings (Bustamante et al. [2014](#page-31-0); Rosal et al. [2018\)](#page-40-12). Community-based rodent control measures signifcantly reduced rodent infestations and the prevalence of *T. cruzi* infection in early-stage nymphs of *T. dimidiata* (De Urioste-Stone et al. [2015\)](#page-32-9).

Human Hosts The prevalence of human infection with *T. cruzi* attests to the potential magnitude of the disease and to the size of the human reservoir. Humans constitute a many decade-long reservoir of *T. cruzi* unless an effective treatment is administered. In contrast, other nonhuman domestic reservoir hosts have large turnover rates, which combined with effective vector control actions produce a fast clearance of infected individuals, which are mostly replaced with uninfected ones (Gürtler et al. [1990;](#page-35-4) Gürtler and Cardinal [2015](#page-35-8)).

3.3 Human Infection

Humans typically acquire a vector-borne infection with *T. cruzi* while sleeping at night at their usual resting location or during short visits to other villages (Gürtler et al. [2007b](#page-35-9)). Incidental human infection may sporadically occur in campsites and as an occupational hazard (Brenière et al. [2017](#page-30-3)). Human exposure may eventually derive from infected triatomines dispersing from peridomestic outhouses (Cardinal et al. [2014\)](#page-31-11) or from the interface with sylvatic habitats, as in the dry-shrub fences harboring *T. eratyrusiformis* in northern Argentina (Cecere et al. [2016](#page-32-14)), stone piles infested with *T. pallidipennis* in Mexico (Brenière et al. [2017](#page-30-3)), and palm trees with intrusive *Rhodnius* sp. and other triatomine species (Jácome-Pinilla et al. [2015;](#page-36-10) Abad-Franch et al. [2015](#page-30-7)). Human susceptibility to *T. cruzi* infection appears to be independent of age and gender.

The infection involves a short acute phase and a lifelong chronic phase and is irreversible unless the patient is treated with nifurtimox or benznidazole. A very small fraction of *T. cruzi*-seropositive individuals spontaneously revert to a seronegative status in the absence of etiologic treatment (e.g., Morillo et al. [2015\)](#page-38-10). The death rate of untreated individuals during the acute phase ranged from 2 to 12% and was inversely related to patient age (Dias and Schofeld [2017](#page-33-11)), as does the chance of a symptomatic presentation (Romaña [1963](#page-40-13)). A variable fraction (20–30%) of the survivors develop cardiac disease and suffer increased death rates between 35 and 50 years of age, especially males.

The bug-to-human transmission probability (*b*) is the probability that, in one feeding contact between one infected triatomine and an uninfected human, the human acquires a *T. cruzi* infection. Using indirect methods, *b* was estimated to range between 0.00026 and 0.0011 (Rabinovich et al. [1990](#page-39-13); Nouvellet et al. [2013\)](#page-38-11). Combined with vectorial capacity and other variables, *b* was used to estimate the threshold density of infected vectors required to initiate transmission chains and allow the infection to persist in the community. Vector control programs have extrapolated this rationale to the notion of a threshold domestic infestation prevalence associated with vector-borne transmission (Aiga et al. [2012](#page-30-8)). In practice, measuring each of the variables involved in vectorial capacity with any accuracy is fraught with major diffculties (Dye [1992\)](#page-33-12). Several studies revealed the occurrence of prevalent or incident human cases at very low densities of *T. cruzi*-infected domestic triatomines per unit of search effort (Piesman et al. [1985;](#page-39-14) Rabinovich et al. [1990;](#page-39-13) Gürtler et al. [2005;](#page-35-10) Cardinal et al. [2018\)](#page-31-12). Furthermore, the relation between bug population density and the probability of transmission by contamination during a single feeding contact with an infected bug is hard to assess empirically. Estimation of a threshold density of infected bugs for domestic transmission, if there is any, is additionally hindered by the low sensitivity and imprecision of triatomine sampling methods (Abad-Franch et al. [2014;](#page-30-9) Rojas de Arias et al. [2012](#page-40-4)) and human mobility (see Sect. [4.3](#page-27-0)).

3.4 Host Infectiousness

This key parameter has usually been measured by xenodiagnosis in at least three partially related forms, including the proportion of uninfected vectors that become infected after a replete blood meal on an infected host (i.e., host infectiousness), and more lately by real-time PCR (Gürtler and Cardinal [2015](#page-35-8)). Dog and cat infectiousness determined by xenodiagnosis correlated closely with the concentration of *T. cruzi* DNA determined by quantitative real-time PCR (Enriquez et al. [2014](#page-33-13)).

Before the implementation of large-scale control campaigns in southeast Brazil, the prevalence of xenodiagnosis-positive dogs (28.6%) and cats (19.7%) largely exceeded that recorded in humans (5.7%) (Freitas [1950](#page-34-10)). A similar ranking was often recorded in areas infested with *R. prolixus*, *T. dimidiata*, and *T. infestans* (Gürtler et al. [1996](#page-35-11); Pifano [1973;](#page-39-15) Zeledón et al. [1975\)](#page-42-13). Nearly all the triatomines that fed on human acute cases usually became infected regardless of the species or instar used (Minter-Goedbloed et al. [1978\)](#page-38-12), whereas in a review of several studies *T. cruzi*-seropositive patients (presumably in the chronic stage) infected from 2–3% to 26% of xenodiagnosis nymphs (Gürtler et al. [1996](#page-35-11)). In a recent population-based survey, 60.5% of *T. cruzi*-seropositive humans were infectious to xenodiagnostic triatomines examined by optical microscopy and molecular methods; on average, they infected 5.2% of fourth-instar nymphs, and human infectiousness conformed to the 80-20 rule (Macchiaverna et al. [2020\)](#page-37-11). The infectiousness of *T. cruzi*-seropositive people declined with age (Maguire et al. [1982\)](#page-37-12), unlike the ageindependent pattern frequently recorded in dogs from endemic rural areas (Gürtler et al. [1996;](#page-35-11) Enriquez et al. [2014](#page-33-13)). Moreover, the mean infectiousness of seropositive dogs to seropositive humans differed by an order of magnitude. Thus, although all mammalian species may be considered potential hosts of *T. cruzi*, their reservoir competence may differ substantially.

3.5 Vector Competence

All nymphal instars and adult stages of Triatominae are susceptible to *T. cruzi*, although perhaps not to the same extent, and may become infected with *T. cruzi* when feeding on an infected mammal carrying bloodstream trypomastigotes. The infection may rarely occur by coprophagy or cannibalism and is mostly irreversible except when the insects are severely starved. Following the infectious blood meal and a short latent period ranging from 2 to 10 days, the intensity of infection increases exponentially up to a maximum reached by 45–60 days postinfection when the bugs are fed regularly on noninfected hosts (Garcia et al. [2007](#page-34-11)). Parasite multiplication and development of metacyclic trypomastigotes in *T. infestans* was optimal between 23 and 27 °C, nil below 10 °C, and declined at 28 or 36 °C (Neves [1971\)](#page-38-13), anticipating the strong seasonal forcing in human incidence of infection with *T. cruzi* (see Sect. [3.6\)](#page-19-0). The intensity of bug infection varied both with parasite DTU

and triatomine species (Campos et al. [2007](#page-31-13); Carvalho-Moreira et al. [2003](#page-31-14); de Lana and de Menezes-Machado [2017\)](#page-32-15). The interactions between *T. cruzi* and triatomines are affected by parasite strain, triatomine nutritional state, trypanolytic compounds, digestive enzymes, lectins, gut microbiota, and endocrine system (Garcia et al. [2007;](#page-34-11) Dumonteil et al. [2018](#page-33-14)).

The infection with *T. cruzi* reduced the life span of third-, fourth-, and ffth-instar nymphs of *T. infestans* by 14–17% when the insects were severely starved (Schaub [1992\)](#page-40-14), decreased the fecundity and fertility of *R. prolixus*, increased bug mortality in a temperature-dependent way, and prolonged development (Elliot et al. [2015;](#page-33-15) Marliére et al. [2015;](#page-37-13) Guarneri and Lorenzo [2017\)](#page-35-12). Whether *T. cruzi* affects the fight dispersal of *T. dimidiata* in a sex-dependent way requires further experimental research (Ramirez-Sierra et al. [2010\)](#page-39-16).

In feld settings, the cumulative chances of exposure to an infectious blood meal source are expected to increase over time or with increasing bug developmental stage and blood meal size; these increase the probability of ingesting at least one parasite and the total number of parasites ingested. Bug superinfections (i.e., a new *T. cruzi* infection of an already infected bug) may be quite frequent when the intensity of transmission is high. The prevalence of bug infection increased with bug stage, usually from third instars onward and reached maximum levels in ffth-instar nymphs or adult bugs (e.g., Albarracin-Veizaga et al. [1999;](#page-30-10) Cardinal et al. [2007](#page-31-15), [2014\)](#page-31-11). The intensity of *T. cruzi* infection in *T. infestans* also increased with each successive instar and peaked in late spring, including trypomastigote densities (Giojalas et al. [1990\)](#page-34-12). Only 5–20 to 2000 metacyclic trypomastigotes are needed to experimentally infect a susceptible mouse via the conjunctival, oral, or contaminative cutaneous routes (Eickhoff et al. [2013\)](#page-33-16).

3.6 Transmission Dynamics

Before the implementation of large-scale insecticide spraying campaigns in endemic areas, the seroprevalence of human infection with *T. cruzi* increased nonlinearly with age in stable populations subject to a rather constant risk of infection over time (Fig. [2\)](#page-20-0). Simple catalytic models of pathogen transmission can be ftted to agespecifc seroprevalence rates of infection to estimate the per capita rate at which a susceptible host acquired the infection in unit time over the age range covered by the curve (the force of infection) under several assumptions (Muench [1959;](#page-38-14) Nouvellet et al. [2015](#page-38-15)). Figure [2](#page-20-0) shows the close ft of the irreversible catalytic model to seroprevalence data obtained in two areas of central Argentina in 1956 (Rosenbaum and Cerisola [1961](#page-40-15)). The force of infection was −0.041 per year (95% confdence interval, −0.0254 and − 0.057) in Ojo del Agua, Santiago del Estero (Fig. [2a\)](#page-20-0), and − 0.045 (−0.053 and − 0.037) in Elcano, Córdoba (Fig. [2b\)](#page-20-0). Seroprevalence rates were predicted, and observed, to reach 100% in age groups >70 years; the 50% infection rates were reached by 20 years of age. More recent surveys also showed that nearly all old-age inhabitants were seropositive for *T. cruzi* in some rural

Fig. 2 Observed (dots) and expected (line) age-specifc seroprevalence rates for *T. cruzi* infection in humans from Ojo de Agua (**a**) and Sebastián Elcano (**b**), Argentina, May 1956. Data taken from Rosenbaum and Cerisola ([1961\)](#page-40-15); seropositivity determined by a complement-fxation test. The line is the ft of the catalytic model with constant force of infection over time and age estimated as in Gürtler et al. [\(2005](#page-35-10))

communities of the Bolivian Chaco (Samuels et al. [2013\)](#page-40-16) and in indigenous communities of Santa Marta, Colombia (Mejía-Jaramillo et al. [2014](#page-38-16)). At least 50% of those aged >40 years were *T. cruzi* seropositive in the Argentine Chaco (Cardinal et al. [2018;](#page-31-12) Fernández et al. [2019b](#page-34-13)).

Before large-scale insecticide campaigns, most *symptomatic* acute cases occurred before reaching 15 years of age, but the distribution displayed a long tail extending up to 52 years of age, with no gender-related asymmetry (Romaña [1963](#page-40-13)). Figure [3](#page-21-0) shows that while the expected seroprevalence of *T. cruzi* infection increased with increasing age in the absence of control actions, the observed frequency of *symptomatic* acute cases declined fast with age, although some occurred even after 50 years of age.

Following the implementation of control actions, the observed seroprevalence rates can be compared with those predicted by other candidate models to infer whether the efforts had effectively diminished the force of infection and by how much (Cardinal et al. [2007;](#page-31-15) Cucunubá et al. [2017;](#page-32-16) Feliciangeli et al. [2003](#page-34-14); Nouvellet et al. [2015;](#page-38-15) Samuels et al. [2013](#page-40-16)).

Seasonality strongly affects the intensity of transmission of *T. cruzi* in subtropical and temperate areas. The incidence of *symptomatic* human cases of Chagas disease displayed similar seasonal variations in northern Argentina across four decades (Romaña [1963;](#page-40-13) Rebosolán and Terzano [1958](#page-40-17); Ledesma Patiño et al. [1992;](#page-36-11) Lugones et al. [1994](#page-37-14)). Incidence steeply rose from nonzero minimum values in August (cold season) to peak by November (late spring) and then slightly decreased during the hot summer months (Fig. [4](#page-21-1)). The relative rate of change in the frequency of acute cases increased faster from late winter to spring than at other times in three of the four data sets. A similar pattern occurred in central Brazil in the 1950s (Schofeld [1994\)](#page-41-6). These patterns closely match seasonal variations in temperature acting on triatomine blood-feeding rates (Fig. [5](#page-22-0)).

Fig. 4 Monthly-specifc relative frequency of symptomatic acute cases of Chagas disease in humans from Tucumán and Santiago del Estero, Argentina. Data taken from Rebosolán and Terzano ([1958\)](#page-40-17), including 511 cases diagnosed over 1947–1956; Romaña [\(1963](#page-40-13)), including 272 cases; Lugones et al. [\(1994](#page-37-14)), including 289 cases, and Ledesma Patiño et al. ([1992\)](#page-36-11), including 128 cases over a 3-year period ca. 1990. The lines are the ft of a four-degree polynomial model. Alltime series start at the all-time minimum values recorded in August

The relations between domestic host availability, bug abundance, and bloodfeeding behavior described above, in interaction with local host infection and infectiousness, determine household-level variations in the prevalence and abundance of domestic bugs infected with *T. cruzi*. The strong and positive association between household infection (especially children) and the presence or abundance of infected *T. infestans* or *P. megistus* has been documented across multiple settings (Mott et al.

Fig. 5 Conceptual framework for the social determinants of health (SDHs) applied to Chagas disease. Adapted from Solar and Irwin ([2010\)](#page-41-12). The structural determinants are differentiated between those related to the socioeconomic and political context at national or regional levels and those pertaining to the individual and household level, as are the intermediary SDHs. The dashed light gray arrow indicates the possibility that negative health outcomes infuence policy; this feedback is less likely to occur in the case of marginalized diseases

[1976;](#page-38-17) Piesman et al. [1985;](#page-39-14) Gürtler et al. [1998a,](#page-35-13) [2005;](#page-35-10) Alroy et al. [2015](#page-30-11); Cardinal et al. [2018;](#page-31-12) Fernández et al. [2019a\)](#page-34-1). In a peri-urban community of Arequipa (Peru), child infection was instead associated with peridomestic infection in *T. infestans* (Levy et al. [2007\)](#page-37-15). In rural villages of northwestern Argentina and northeast Brazil, human and vector infections were strongly and positively associated with the house-hold presence and number of infected dogs (Cardinal et al. [2014](#page-31-11); Gürtler et al. [1998a](#page-35-13),[b,](#page-35-14) [2005;](#page-35-10) Mott et al. [1978b\)](#page-38-18).

In theory, the circulation of *T. cruzi* among multiple host species differing in reservoir competence might favor pathogen persistence (maintenance) or high pathogen abundance (amplifcation) or reduce both of them (the dilution effect) (Begon [2008\)](#page-30-12). The empirical relations described above were embodied in a mathematical model of domestic transmission mediated by *T. infestans*, which accounted for host-species effects and seasonality (Cohen and Gürtler [2001\)](#page-32-11). Having two or more *T. cruzi*-infected dogs disproportionately increased the prevalence of vector and human infection. Dogs acquired the infection within a few months of exposure, and 90–100% of dogs were infected by 3 years of age (Gürtler et al. [1996](#page-35-11), [2007a\)](#page-35-15). Although refractory to *T. cruzi* infection, the indoor occurrence of one or two nesting chickens boosted domestic bug populations beyond the constraints set by human and dog numbers and ultimately contributed to larger infected-bug abundances and human infection through frequent host shifts (Gürtler et al. [1998b](#page-35-14)) (see Sect. [2.5\)](#page-7-0). Other model specifcations reached similar conclusions: increasing the number of dogs would amplify the intensity of domestic transmission (Fabrizio et al. [2016;](#page-34-15)

Flores-Ferrer et al. [2019](#page-34-16); Nouvellet et al. [2015;](#page-38-15) Peterson et al. [2015](#page-39-17); Spagnuolo et al. [2012\)](#page-41-13). The net effects of adding transmission non-competent hosts (chickens) on bug and host infection may vary with the precise details on the relative number and location of chickens, humans, and triatomine species involved and thus cannot be generalized to all ecological settings. The key fnding is that chickens and selected domestic animals increase the equilibrium vector population size across triatomine species (e.g., Minter [1976;](#page-38-4) Dumonteil et al. [2018\)](#page-33-14) and ultimately increase human-vector contact rates and exposure.

4 Social Determinants of Domestic Transmission

The social determinants of health (SDHs) condition both disease distribution and the ways they are handled (Manderson et al. [2009\)](#page-37-16). Although the links between social factors and health outcomes have been widely recognized since the 1990s, the SDHs are often disregarded in biomedical research related to disease control because they fall outside the scope of traditional healthcare systems (Bizimana et al. [2015;](#page-30-13) Manderson et al. [2009](#page-37-16)). To address this issue, the World Health Organization has developed a conceptual framework to act upon the SDHs (Solar and Irwin [2010\)](#page-41-12). This framework is based on the concept that "the social position of individuals and population groups is the main determinant of health inequalities within a community" by determining differential exposures to health-adverse conditions among individuals, differential consequences resulting from exposure (socioeconomic or health outcomes), and/or differential capabilities to recover (Solar and Irwin [2010](#page-41-12)).

The SDHs of Chagas disease include interrelated socioeconomic and demographic factors such as ethnicity, gender, occupation, educational level, and income, i.e., structural determinants (Ehrenberg and Ault [2005;](#page-33-1) Hotez et al. [2008;](#page-36-12) Aagaard-Hansen and Claire [2010](#page-29-1); Solar and Irwin [2010](#page-41-12)). These determinants affect other factors more directly associated with disease exposure and outcome (i.e., intermediary determinants), such as household and dwelling characteristics.

4.1 Socioeconomic Factors

The social stratifcation of individuals and demographic groups results from, and also perpetuates, socioeconomic inequalities (Pluciński et al. [2013](#page-39-18)), leading some people to live in a state of relative or absolute poverty. Understanding poverty as a dynamic and multidimensional process (as opposed to a lack of resources) requires introducing the concept of social vulnerability, which considers the "defenselessness, insecurity, and exposure to risks, shocks and stress" experienced by households (Chambers [1989\)](#page-32-17). The notion of social vulnerability summarizes the multiple, interrelated structural and intermediary determinants associated with the

socioeconomic position (SEP) of individuals and groups in a population. In the context of low- and middle-income countries, socioeconomic inequalities were represented using surrogate indicators such as educational attainment and household ownership of assets (Houweling et al. [2016\)](#page-36-13). These indicators may only partially capture the full complexity of poverty. In particular, Chagas disease presents a disproportionately high disease burden on indigenous communities and poor rural peasants across Latin America (Hotez et al. [2008](#page-36-12); Hotez [2014](#page-36-14); Gürtler [2009\)](#page-35-16). Although poverty has long been acknowledged as the main driver of Chagas disease risk (Ault [2007](#page-30-14); Guhl et al. [2007;](#page-35-17) Briceño-León and Méndez Galván [2007](#page-31-16); Gürtler [2009\)](#page-35-16), evidence of the effects of socioeconomic inequalities is limited compared to other NTDs (Houweling et al. [2016](#page-36-13); Fernández et al. [2019a\)](#page-34-1).

The socioeconomic position of individuals and households is associated with other structural SDHs such as educational level, ethnicity, and income, which in turn will determine several intermediary SDHs associated with vector occurrence or invasion of human habitations, human exposure, and the resources required to deal with the consequences of vector exposure. However, fnding appropriate measures of household SEP in low- and middle-income countries and in contexts of structural poverty is not trivial. Traditional income-based indices may fail to capture the full range of heterogeneity across household socioeconomic status (Booysen et al. [2008;](#page-30-15) Howe et al. [2012\)](#page-36-15), particularly in communities where household monetary income is principally dependent on the scarce jobs available and on welfare support, as in many endemic areas for Chagas disease.

As an alternative, several studies have explored the effects of SEP-related intermediary and structural SDHs on house infestation. Most of them focused on housing quality and construction materials because of their association with refuge availability rather than as a measure of SEP (Levy et al. [2006](#page-36-16); Gurevitz et al. [2011;](#page-35-3) Dumonteil et al. [2013;](#page-33-5) Bustamante et al. [2014](#page-31-0); Gaspe et al. [2015](#page-34-8)). Other studies investigated the effects of other surrogates of SEP on house infestation with *T. infestans* and the occurrence of infected domestic vectors, sometimes using overcrowding and/or ownership of livestock (goat-equivalent index) as surrogate indices of household wealth (Gaspe et al. [2015;](#page-34-8) Cardinal et al. [2018](#page-31-12)). Overcrowding (i.e., human density in sleeping quarters) incorporates both household size and number of rooms and was closely and positively associated with domestic infestation and bug abundance (Gaspe et al. [2015](#page-34-8)). Overcrowding is expected to facilitate host fnding and blood-feeding success on humans and most likely underlies the positive relation between the number of human occupants and domestic infestation with triatomine bugs (Marsden et al. [1982](#page-37-4); Piesman et al. [1983](#page-39-1); Levy et al. [2006;](#page-36-16) Campbell-Lendrum and Woodruff [2007;](#page-31-17) Provecho et al. [2017](#page-39-19); Cardinal et al. [2018\)](#page-31-12). The goat-equivalent index was weakly and inversely associated with house infestation with *T. infestans* and the occurrence of infected vectors in the Argentine Chaco (Gaspe et al. [2015](#page-34-8); Cardinal et al. [2018](#page-31-12)); although it captured part of the variability between households and demographic groups, the effect was not statistically signifcant. Another study that included SEP as a risk factor for house infestation with *T. dimidiata* in Guatemala found a moderate association between house infestation and two summary indices representing household assets (cell phone and livestock)

and access to electricity (Bustamante et al. [2014](#page-31-0)). For intrusive species such as *T. dimidiata*, artifcial light in the house or in nearby streets was positively associated with house infestation (Pacheco-Tucuch et al. [2012;](#page-39-9) Dumonteil et al. [2013](#page-33-5)).

Educational level has also been considered a surrogate of SEP in studies of house infestation in Yucatan (Dumonteil et al. [2013](#page-33-5)) and in the Argentine Chaco (Gaspe et al. [2015\)](#page-34-8), where it showed a negative and signifcant association with domestic infestation. A direct causal pathway from increasing household educational levels to decreasing infestation may be related to access to information and receptivity to health education messages, which may translate into healthier practices (Solar and Irwin [2010](#page-41-12)). For example, education levels correlated directly with the severity of Chagas disease cardiomyopathy (Viotti et al. [2009\)](#page-41-14). However, educational level (as determined by the duration of formal instruction) does not specify its quality nor health education through informal channels (Gaspe et al. [2015](#page-34-8)) and most likely refects socioeconomic inequalities between households.

The association between human infection with *T. cruzi* and selected sociodemographic factors in rural endemic areas has been a recurrent focus of interest (e.g., Gürtler et al. [1998a](#page-35-13), [b](#page-35-14), [2005](#page-35-10); Levy et al. [2007](#page-37-15); Samuels et al. [2013;](#page-40-16) Alroy et al. [2015;](#page-30-11) Cardinal et al. [2018\)](#page-31-12). However, these studies did not address the combined effects of ecological and social variables due to limited data availability. While human infection increased with infected-bug abundance and the household presence or number of domestic dogs (see Sect. [3.5\)](#page-18-0), a less consistent association was found with house construction quality (i.e., thatched roofs and cracks in the walls): some studies reported an inverse relation between infection and selected aspects of housing quality (Mott et al. [1978a;](#page-38-3) Gürtler et al. [1998a,](#page-35-13) [b,](#page-35-14) [2005;](#page-35-10) Samuels et al. [2013\)](#page-40-16), whereas others did not (Levy et al. [2007](#page-37-15); Alroy et al. [2015](#page-30-11)).

The multiple SDHs related to poverty (e.g., poor-quality housing, household overcrowding, and low educational level) may be summarized into a social vulnerability index by means of multiple correspondence analysis (Fernández et al. [2019a\)](#page-34-1). The concept of social vulnerability may be taken as an ex ante risk that a household will fall below the poverty line or, if already poor, will remain in poverty (Chaudhuri et al. [2002\)](#page-32-18). When considered as a SDH, social vulnerability refers to a predisposition of certain individuals or groups to acquire the disease(s) in question, and their capacity to respond and access the healthcare system (Hagenlocher and Castro [2015;](#page-36-17) Bizimana et al. [2015\)](#page-30-13). In creole and indigenous households in the Argentine Chaco, the social vulnerability index was positively associated with house infestation and the abundance of *T. cruzi*-infected domestic triatomines and was negatively correlated with the asset index and domestic insecticide use (Fernández et al. [2019a\)](#page-34-1). This analysis clearly reveals the tight links between triatomine exposure and resource constraints, poor housing quality (suitable habitats), householders' prevention practices, and sociodemographic factors that reflect and perpetuate poverty. Most importantly, the social vulnerability index was positively associated with human infection after adjusting for other relevant demographic and ecological factors (Fernández et al. [2019b\)](#page-34-13).

Access to health services, an important determinant related to SEP, also depends on health and infrastructure-related policies (Solar and Irwin [2010\)](#page-41-12). Access to health services includes the availability of healthcare facilities and personnel and the distance and transportation means available to the household. In general, evidence of the association between socioeconomic status and access to health services is scattered and seems to be context-dependent (Raso et al. [2005;](#page-40-18) Fürst et al. [2009\)](#page-34-17). Other barriers to healthcare access in endemic rural communities include alleged discriminatory behaviors within the health system, especially of indigenous and other vulnerable groups (Dell'Arciprete et al. [2014;](#page-33-17) Brierley et al. [2014\)](#page-31-18). In the Argentine Chaco, domestic infestation was signifcantly lower in houses with greater access to health services, possibly refecting the aggregation of non-infested, new houses built around healthcare posts or their improved access to insecticides or capacity to demand vector control actions (Fernández et al. [2019a](#page-34-1)). Access to healthcare and other services were one of the main reasons for household mobility within rural communities and relocation in the periphery of the local town (Sect. [4.3](#page-27-0)).

4.2 Ethnicity

The intersection between SEP and ethnicity can further increase the inequalities observed in Chagas disease endemic areas, as explained by the intersectional paradigm (Hankivsky and Christoffersen [2008\)](#page-36-18). Multiple indigenous groups live in Chagas disease endemic areas (Hotez et al., [2008\)](#page-36-12). In the Gran Chaco region, the seroprevalence of *T. cruzi* in indigenous peoples tended to exceed that of creole residents (Basombrio et al. [1999](#page-30-16); Taranto et al. [2003;](#page-41-15) Biancardi et al. [2003](#page-30-17); Diosque et al. [2004;](#page-33-18) Alonso et al. [2009](#page-30-18); Sosa-Estani et al. [2009;](#page-41-16) Moretti et al. [2010;](#page-38-19) Lucero et al. [2016](#page-37-10); Cardinal et al. [2018](#page-31-12)). House infestation rates with *T. infestans* were higher in indigenous households, consistent with their more precarious living conditions (Gurevitz et al. [2011;](#page-35-3) Gaspe et al. [2015,](#page-34-8) [2018;](#page-34-18) Provecho et al. [2017](#page-39-19)) and greater social vulnerability (Fernández et al. [2019a](#page-34-1)). Their dogs and cats also displayed greater infection prevalence than those owned by creoles (Cardinal et al. [2014\)](#page-31-11). However, the statistical effects of ethnic background ceased to be signifcant when other ecological and socioeconomic variables more closely related to house infestation or domestic triatomine abundance were incorporated to these multimodelbased analyses (Gurevitz et al. [2011\)](#page-35-3). The effects of ethnicity on human infection with *T. cruzi* may be more complex than anticipated: while a twofold greater risk occurred among indigenous people after adjusting for social vulnerability and other factors in a section of Pampa del Indio (Fernández et al. [2019b\)](#page-34-13), no effects were observed in another rural section including a majority of creole households (Cardinal et al. [2018](#page-31-12)). Cultural factors associated with ethnic background (Arrom-Suhurt et al. [2018\)](#page-30-19) likely affected human exposure to triatomines.

4.3 Human Migration and Mobility

Human migration and mobility can affect domestic transmission patterns by facilitating vector dispersal and modifying exposure to the vector and by introducing *T. cruzi*-infected people and nonhuman reservoir hosts. Migration from endemic regions to non-endemic countries (driven mostly by economic/labor reasons) expanded Chagas disease to Europe and non-endemic areas in North America, Japan, and Australia (Schmunis and Yadon [2010;](#page-40-19) Lee et al. [2013\)](#page-36-19). There, vertical transmission became the main transmission route (Sicuri et al. [2011](#page-41-17); Howard et al. [2014\)](#page-36-20), followed by transmission via blood transfusion and organ transplantation before the implementation of prevention measures (Girolamo et al. [2011](#page-33-19)). The SEP of migrants in the receiving country and their access to health services created a new set of challenges for case detection and treatment (Ventura-Garcia et al. [2013\)](#page-41-18). The same argument holds for migration from endemic rural areas to non-endemic areas within Latin America.

The steady rural-to-urban migration recorded during the twentieth century and projected for future decades, combined with increasing travel and transportation of goods from rural to peri-urban or urban areas, provides multiple routes of entry of triatomine bugs into habitats wrongly assumed not to be at risk of infestation. Consequently, several species of triatomines (including *T. infestans*, *T. dimidiata*, *Triatoma pallidipennis*, and *Mepraia spinolai*) colonized peri-urban and urban habitats and even invaded the top stories of city buildings through fight dispersal (Vallvé et al. [1996](#page-41-19); Albarracin-Veizaga et al. [1999](#page-30-10); Cattan et al. [2002;](#page-32-19) Ramsey et al. [2005;](#page-40-20) Levy et al. [2006;](#page-36-16) Guzman-Tapia et al. [2007](#page-36-21); Medrano-Mercado et al. [2008;](#page-38-20) Lima et al. [2012](#page-37-17); Gaspe et al. [2020\)](#page-34-19). Peri-urban areas, defned as "the areas where the urban core intermingles with adjacent 'non-urban' systems" (MacGregor-Fors [2011\)](#page-37-18), provide a transition between urban and rural areas. In Latin America, periurban areas frequently include precarious settlements where new migrants usually frst settle, occupying vacant land with low land-tenure security (Levy et al. [2014\)](#page-37-19). Therefore, migration and settlement patterns may represent relevant risk factors for house infestation and transmission of *T. cruzi* (Bayer et al. [2009;](#page-30-20) Delgado et al. [2013;](#page-32-20) Levy et al. [2014](#page-37-19)). The combination of substandard housing quality and prox-imity between houses facilitates triatomine invasion (Levy et al. [2006\)](#page-36-16).

Rural communities in the Argentine Chaco displayed signifcant rates of rural-tourban migration, internal mobility (i.e., moving within the study area: local movers), and return of migrants from urban areas (Fernández et al. [2019a](#page-34-1)). These migration and mobility patterns differed between ethnic groups: creoles displayed the traditional rural-to-urban movement (Briceño-León [2009\)](#page-30-0), whereas indigenous (Qom) migration rates equaled internal mobility rates, with in-migration almost fully compensating out-migration. Qom mobility patterns are rooted in socioeconomic and cultural factors: nomadic traditions (Maidana [2011](#page-37-20)), formation of new families, household mobility to gain increased access to basic services (e.g., better water sources and school), and cultural reasons (decease of the head of family). Local mobility implied elevated house turnover rates causing substantial negative effects on extant house infestations (Gaspe et al. [2015,](#page-34-8) [2018](#page-34-18)) and possibly created

additional heterogeneities in human-vector contact rates (Stoddard et al. [2009\)](#page-41-20). Local movers were likely exposed to lower, more variable infestations over time and greater chances of occupying an infested house than non-movers (Fernández et al., [2019a\)](#page-34-1).

Human mobility in interaction with vector exposure created complex patterns in the seroprevalence rates of children from stable households (non-movers) and among movers (Fernández et al. [2019b\)](#page-34-13). Among non-mover children, infection prevalence was sixfold greater when domestic premises were infested (as expected from prolonged vector exposure), whereas no signifcant association between child infection and domestic infestation occurred among movers (Fernández et al. [2019b\)](#page-34-13). In a peri-urban area of Arequipa, domestic infestation was positively associated with land-tenure security (i.e., more established households, rather than more recent immigrants), suggesting that although land-tenure security may pave the way to improved house quality, it may also provide a more stable environment for triatomine establishment (Levy et al. [2014\)](#page-37-19). Further research is needed to disentangle the effects of internal mobility on triatomine and human exposure dynamics and gauge the relative magnitude of these effects in endemic communities with different socioeconomic and demographic profles.

4.4 Interactions Between Social and Ecological Factors

Understanding the domestic transmission of *T. cruzi* requires a better understanding of the interactions between ecological and social factors such as SEP, ethnicity, and household mobility. In rural communities of the Argentine Chaco, the intersection between ethnicity and SEP further combined with human mobility patterns: movers had higher social vulnerability than non-movers both within indigenous and creole households (Fernández et al. [2019a\)](#page-34-1). The relative odds of human infection varied signifcantly with the interaction between social vulnerability index and infectedvector abundance: infected-vector abundance exerted lower effects (on a per capita basis) in households with higher social vulnerability (Fernández et al. [2019b\)](#page-34-13). This unexpected effect may be explained by several nonexclusive reasons: (i) social vulnerability may also indicate past exposure to triatomines; (ii) households with higher social vulnerability had greater local mobility, suggesting putative vector exposures elsewhere; and (iii) more frequent human-triatomine contact given the greater overcrowding in households with higher social vulnerability.

5 Scaling Up from Householdto Population-Level Transmission

Although the domestic transmission of *T. cruzi* is clustered at a household level (Mott et al. [1976;](#page-38-17) Gürtler et al. [1998a,](#page-35-13) [b](#page-35-14); Levy et al. [2007](#page-37-15); Cardinal et al. [2014](#page-31-11), [2018;](#page-31-12) Fernández et al. [2019b\)](#page-34-13), the combination of human mobility, spatial distribution of houses, and community-level variables (e.g., insecticide spraying or housing improvement efforts) may generate new properties at higher spatial scales. By scaling up from household-level dynamics to the population level, we may gain insight into the ecological and social factors associated with human infection and the impact of community-based vector control actions. The joint analysis of the spatial distribution of human and vector infection can shed light into the processes and factors associated with vector-borne transmission of *T. cruzi*. By integrating the spatial component of infection with household-level and individual-based risk factor analysis, we may identify transmission hotspots, create risk maps of *T. cruzi* infection, and stratify the affected areas for targeted control (Fortin and Dale [2005;](#page-34-20) Wiegand and Moloney [2014](#page-41-21)).

By contrast to vector-related research, the spatial component of human *T. cruzi* infection has rarely been investigated (Levy et al. [2007,](#page-37-15) [2009;](#page-37-21) Delgado et al. [2011;](#page-32-21) Fernández et al. [2019b](#page-34-13)). As stated by Houweling et al. ([2016\)](#page-36-13), "spatial clustering of infection because of geographic conditions, among other causes, is typical for most NTDs." Spatial clustering may also be context-specifc and depend on the intersection between social and ecological factors. The spatial aggregation of house infestation (e.g., Cecere et al. [2004;](#page-32-2) Gaspe et al. [2015\)](#page-34-8) and of human cases of *T. cruzi* infection across settings and scales indicates that spatial heterogeneity is the rule rather than the exception and that transmission is not restricted to the household. The underlying community-level processes possibly emerge from a combination of active and passive vector dispersal (Levy et al. [2006](#page-36-16); Gurevitz et al. [2011:](#page-35-3) Gaspe et al. [2013,](#page-34-21) [2015;](#page-34-8) Provecho et al. [2017\)](#page-39-19), the aggregation of more vulnerable households, and the effects of human mobility (Gaspe et al. [2015](#page-34-8), [2018](#page-34-18): Fernández et al. [2019a](#page-34-1)). These in turn can be affected by social networks and public policies. Integrating the ecological and social determinants of human infection with the spatial component is key to the design of more cost-effective vector control strategies in resource-constrained areas to identify higher-priority areas for targeted interventions oriented to suppress house (re)infestations, treat infected children, and thus reduce the current and future burden of disease.

Acknowledgments We are grateful to Carla Cecere, Sol Gaspe, Gustavo Enriquez, Yael Provecho, Paula Sartor, and Natalia Macchiaverna for helpful discussions. This work was supported by the University of Buenos Aires (UBACYT 20020130100843BA), PICT 2014-2661 and PICT 2015-2921.

References

Aagaard-Hansen J, Claire LC (2010) Neglected tropical diseases: equity and social determinants. In: Blas E, Kurup AS (eds) Equity, social determinants and public health programmes. World Health Organization, Ginebra, pp 135–157

Abad-Franch F (2016) A simple, biologically sound, and potentially useful working classifcation of Chagas disease vectors. Mem Inst Oswaldo Cruz 111:649–651

- Abad-Franch F, Valença-Barbosa C, Sarquis O et al (2014) All that glisters is not gold: samplingprocess uncertainty in disease-vector surveys with false negative and false-positive detections. PLoS Negl Trop Dis 8:e3187
- Abad-Franch F, Lima MM, Sarquis O et al (2015) On palms, bugs, and Chagas disease in the Americas. Acta Trop 151:126–141
- Abrahan L, Gorla D, Catalá S (2011) Dispersal of *Triatoma infestans* and other Triatominae species in the arid Chaco of Argentina. Flying, walking or passive carriage? The importance of walking females. Mem Inst Oswaldo Cruz 106:232–239
- Aiga H, Sasagawa E, Hashimoto K et al (2012) Chagas disease: assessing the existence of a threshold for bug infestation rate. Am J Trop Med Hyg 86:972–979
- Albarracin-Veizaga H, Carvalho ME, Nascimento EM et al (1999) Chagas disease in an area of recent occupation in Cochabamba, Bolivia. Rev Saude Publ 33:230–236
- Alonso JM, Fabre AR, Galván M et al (2009) La enfermedad de Chagas en poblaciones aborígenes del Noreste de Argentina. Enf Emerg 11:115–118
- Alroy KA, Huang C, Gilman RH et al (2015) Prevalence and transmission of *Trypanosoma cruzi* in people of rural communities of the High Jungle of Northern Peru. PLoS Negl Trop Dis 9:e0003779
- Andrade AL, Zicker F, De Oliveira RM et al (1995) Evaluation of risk factors for house infestation by *Triatoma infestans* in Brazil. Am J Trop Med Hyg 53:443–447
- Araujo RN, Pereira MH, Soares AC et al (2009) Effect of intestinal erythrocyte agglutination on the feeding performance of *Triatoma brasiliensis* (Hemiptera: Reduviidae). J Insect Physiol 55:862–868
- Arrom-Suhurt CM, Arrom-Suhurt CH, Arrom-Suhurt MA et al (2018) Socioeconomic profle and perceptions of Chagas disease in indigenous communities of the Paraguayan Chaco. J Public Health:1–10
- Ault SK (2007) Chagas disease and neglected diseases: challenging poverty and exclusion. In: La enfermedad de Chagas a la puerta de los 100 años de conocimiento de una endemia americana ancestral. Organización Panamericana de la Salud-Fundación Mundo Sano, Buenos Aires, pp 13–18
- Basombrio MA, Segovia A, Esteban E et al (1999) Endemic *Trypanosoma cruzi* infection in indian populations of the Gran Chaco territory of South America: performance of diagnostic assays and epidemiological features. Ann Trop Med Parasitol 93:41–48
- Bayer AM, Hunter GC, Gilman RH et al (2009) Chagas disease, migration and community settlement patterns in Arequipa, Peru. PLoS Negl Trop Dis 3:e567
- Begon M (2008) Effects of host diversity on disease dynamics. In: Ostfeld RS, Keesing F, Eviner ET (eds) Infectious disease ecology. Effects of ecosystems on disease and of disease on ecosystems. Princeton University Press, Princeton, pp 12–29
- Beyer HL, Haydon DT, Morales JM et al (2010) The interpretation of habitat preference metrics under use-availability designs. Philos Trans R Soc B 365:2245–2254
- Biancardi MA, Conca Moreno M, Torres N et al (2003) Seroprevalencia de la enfermedad de Chagas en 17 parajes del "Monte impenetrable" de la provincia del Chaco. Medicina (Buenos Aires) 63:125–129
- Bizimana J-P, Twarabamenye E, Kienberger S (2015) Assessing the social vulnerability to malaria in Rwanda. Malar J 14:2
- Booysen F, van der Berg S, Burger R et al (2008) Using an asset index to assess trends in poverty in seven Sub-Saharan African countries. World Dev 36:1113–1130
- Brenière SF, Waleckx E, Barnabé C (2016) Over six thousand *Trypanosoma cruzi* strains classifed into Discrete Typing units (DTUs): attempt at an inventory. PLoS Negl Trop Dis 10:e0004792
- Brenière SF, Villacis A, Aznar C (2017) Vector transmission: how it works, what transmits, where it occurs. In: Tibayrenc M, Telleria J (eds) American trypanosomiasis Chagas disease: one hundred years of research, 2nd edn. Elsevier, Saint Louis, pp 497–516
- Briceño-León R (2009) La enfermedad de Chagas en las Américas: una perspectiva de ecosalud. Cad Saude Publica 25(Supl1):71–82
- Briceño-León R, Méndez Galván J (2007) The social determinants of Chagas disease and the transformations of Latin America. Mem Inst Oswaldo Cruz 102:109–112
- Brierley CK, Suarez N, Arora G et al (2014) Healthcare access and health beliefs of the indigenous peoples in remote Amazonian Peru. Am J Trop Med Hyg 90:180–183
- Buitrago NL, Bosseno MF, Waleckx E et al (2013) Risk of transmission of *Trypanosoma cruzi* by wild *Triatoma infestans* (Hemiptera: Reduviidae) in Bolivia supported by the detection of human blood meals. Infect Genet Evol 19:141–144
- Buitrago R, Bosseno MF, Depickère S et al (2016) Blood meal sources of wild and domestic *Triatoma infestans* (Hemiptera: Reduviidae) in Bolivia: connectivity between cycles of transmission of *Trypanosoma cruzi*. Parasit Vectors 9:214
- Bustamante DM, Monroy C, Pineda S et al (2009) Risk factors for intradomiciliary infestation by the Chagas disease vector *Triatoma dimidiata* in Jutiapa, Guatemala. Cad Saude Publica 25:S83–S92
- Bustamante DM, de Urioste-Stone SM, Cruz JG et al (2014) Ecological, social and biological risk factors for continued *Trypanosoma cruzi* transmission by *Triatoma dimidiata* in Guatemala. PLoS One 9:e104599
- Campbell-Lendrum D, Woodruff R (2007) Climate change – quantifying the health impact at national and local levels. In: Environmental burden of disease series, no.14. World Health Organization, Geneva
- Campbell-Lendrum DH, Angulo VM, Esteban L et al (2007) House-level risk factors for triatomine infestation in Colombia. Int J Epidemiol 36:866–872
- Campos R, Acuna-Retamar M, Botto-Mahan C et al (2007) Susceptibility of *Mepraia spinolai* and *Triatoma infestans* to different *Trypanosoma cruzi* strains from naturally infected rodent hosts. Acta Trop 104:25–29
- Carcavallo RU, Franca-Rodriguez ME, Salvatella R et al (1999) Habitats and related fauna. In: Carcavallo RU, Girón IG, Jurberg J, Lent H (eds) Atlas of Chagas disease vectors in the Americas, vol II. Editora Fiocruz, Rio de Janeiro, pp 561–620
- Cardinal MV, Lauricella MA, Marcet PL et al (2007) Impact of community-based vector control on house infestation and *Trypanosoma cruzi* infection in *Triatoma infestans*, dogs and cats in the Argentine Chaco. Acta Trop 103:201–211
- Cardinal MV, Lauricella MA, Ceballos LA et al (2008) Molecular epidemiology of domestic and sylvatic *Trypanosoma cruzi* infection in rural northwestern Argentina. Int J Parasitol 38:1533–1543
- Cardinal MV, Orozco MM, Enriquez GF et al (2014) Heterogeneities in the eco-epidemiology of *Trypanosoma cruzi* infection in rural communities of the Argentinean Chaco. Am J Trop Med Hyg 90:1063–1073
- Cardinal MV, Sartor PA, Gaspe MS et al (2018) High levels of human infection with *Trypanosoma cruzi* associated with the domestic density of infected vectors and hosts in a rural area of northeastern Argentina. Parasit Vectors 11:492
- Carvalho-Moreira CJ, Spata MC, Coura JR et al (2003) In vivo and in vitro metacyclogenesis tests of two strains of *Trypanosoma cruzi* in the triatomine vectors *Triatoma pseudomaculata* and *Rhodnius neglectus*: short/long-term and comparative study. Exp Parasitol 103:102–111
- Castañera MB, Aparicio JP, Gürtler RE (2003) A stage-structured stochastic model of the population dynamics of *Triatoma infestans*, the main vector of Chagas disease. Ecol Model 162:33–53
- Castillo-Neyra R, Barbu CM, Salazar R et al (2015) Host-seeking behavior and dispersal of *Triatoma infestans*, a vector of Chagas disease, under semifeld conditions. PLoS Negl Trop Dis 9:e3433
- Catalá SS (1994) Blood meal size and nutritional status of *Triatoma infestans* under natural climatic conditions. Med Vet Entomol 8:104–106
- Catalá S, Crocco LB, Morales GF (1997) *Trypanosoma cruzi* transmission risk index (TcTRI): an entomological indicator of Chagas disease vectorial transmission to humans. Acta Trop 68:285–295
- Cattan PE, Pinochet A, Botto-Mahan C et al (2002) Abundance of *Mepraia spinolai* in a periurban zone of Chile. Mem Inst Oswaldo Cruz 97:285–287
- Ceballos LA, Vazquez-Prokopec GM, Cecere MC et al (2005) Feeding rates, nutritional status and fight dispersal potential of peridomestic populations of *Triatoma infestans* in rural northwestern Argentina. Acta Trop 95:149–159
- Cecere MC, Gürtler RE, Chuit R et al (1997) Effects of chickens on the prevalence of infestation and population density of *Triatoma infestans* in rural houses of North-West Argentina. Med Vet Entomol 11:383–388
- Cecere MC, Gürtler RE, Canale DM et al (1998) Factors limiting the domiciliary density of *Triatoma infestans*, vector of Chagas' disease, in North-West Argentina: a longitudinal study. Bull World Health Organ 76:373–384
- Cecere MC, Canale DM, Gürtler RE (2003) Effects of refuge availability on the population dynamics of *Triatoma infestans* in Central Argentina. J Appl Ecol 40:742–756
- Cecere MC, Vazquez-Prokopec GM, Gürtler RE et al (2004) Spatiotemporal analysis of reinfestation by *Triatoma infestans* (Hemiptera: Reduviidae) following insecticide spraying in a rural community in northwestern Argentina. Am J Trop Med Hyg 71:803–810
- Cecere MC, Vazquez-Prokopec GM, Ceballos LA et al (2006) Comparative trial of the effectiveness of pyrethroid insecticides against peridomestic populations of *Triatoma infestans* in northwestern Argentina. J Med Entomol 43:902–909
- Cecere MC, Vazquez-Prokopec GM, Ceballos LA et al (2013) Improved chemical control of Chagas disease vectors in the dry Chaco region. J Med Entomol 50:394–403
- Cecere MC, Leporace M, Fernández MP et al (2016) Host-feeding sources and infection with *Trypanosoma cruzi* of *Triatoma infestans* and *Triatoma eratyrusiformis* (Hemiptera, Reduviidae) from the Calchaqui Valleys in Northwestern Argentina. J Med Entomol 53:666–673
- Chambers R (1989) Special issue on vulnerability: how the poor cope. IDS Bull 20:1–7
- Charron D (2012) Ecohealth research in practice. Innovative applications of an ecosystem approach to health. International Development Research Centre/Springer, Ottawa/New York
- Chaudhuri S, Jalan J, Suryahadi A (2002) Assessing household vulnerability to poverty from crosssectional data: a methodology and estimates from Indonesia. Columbia University, New York
- Cohen JE, Gürtler RE (2001) Modeling household transmission of American trypanosomiasis. Science 293:694–698
- Cohen JE, Rodríguez-Planes LI, Gaspe MS et al (2017) Taylor's law and Chagas disease vector control. PLoS Negl Trop Dis 11:6092
- Colinet H, Sinclair BJ, Vernon P et al (2015) Insects in fuctuating thermal environments. Annu Rev Entomol 60:123–140
- Crocco L, Catalá S (1997) Host preferences of *Triatoma sordida*. Ann Trop Med Parasitol 91:927–930
- Cucunubá ZM, Nouvellet P, Conteh L et al (2017) Modelling historical changes in the forceof-infection of Chagas disease to inform control and elimination programmes: application in Colombia. BMJ Glob Health 2:e000345
- Curto de Casas SI, Carcavallo RU, Galíndez Girón I et al (1999) Bioclimatic factors and zones of life. In: Carcavallo RU, Girón IG, Jurberg J, Lent H (eds) Atlas of Chagas disease vectors in the Americas, vol III. Editora Fiocruz, Rio de Janeiro, pp 793–838
- de Lana M, de Menezes-Machado EM (2017) Biology of *Trypanosoma cruzi* and biological diversity. In: Tibayrenc M, Telleria J (eds) American Trypanosomiasis Chagas disease: one hundred years of research, 2nd edn. Elsevier, Saint Louis, pp 339–363
- de Urioste-Stone SM, Pennington PM, Pellecer E et al (2015) Development of a community-based intervention for the control of Chagas disease based on peridomestic animal management: an eco-bio-social perspective. Trans R Soc Trop Med Hyg 109:159–167
- Delgado S, Castillo Neyra R, Quispe Machaca VR et al (2011) A history of Chagas disease transmission, control, and re-emergence in peri-rural La Joya, Perú. PLoS Negl Trop Dis 5:e970
- Delgado S, Ernst KC, Pumahuanca ML et al (2013) A country bug in the city: urban infestation by the Chagas disease vector *Triatoma infestans* in Arequipa, Peru. Int J Health Geogr 12:48
- Dell'Arciprete A, Braunstein J, Touris C et al (2014) Cultural barriers to effective communication between indigenous communities and health care providers in northern Argentina: an anthropological contribution to Chagas disease prevention and control. Int J Equity Health 13:6
- Di Girolamo C, Bodini C, Marta BL et al (2011) Chagas disease at the crossroad of international migration and public health policies: why a national screening might not be enough. Eur Secur 16:19965
- Di Iorio O, Gürtler RE (2017) Seasonality and temperature-dependent fight dispersal of *Triatoma infestans* and other vectors of Chagas disease in western Argentina. J Med Entomol 54:1285–1292
- Dias E (1955) Variações mensais da incidência das formas evolutivas do *Triatoma infestans* e do *Panstrongylus megistus* no município de Bambuí, Estado de Minas Gerais. Mem Inst Oswaldo Cruz 53:457–472
- Dias JCP, Schofeld CJ (2017) Social and medical aspects on Chagas disease management and control. In: Tibayrenc M, Telleria J (eds) American Trypanosomiasis Chagas disease: one hundred years of research, 2nd edn. Elsevier, Saint Louis, pp 47–57
- Dias E, Zeledón R (1955) Infestaçao domiciliária em grau extremo por *Triatoma infestans*. Mem Inst Oswaldo Cruz 53:473–486
- Diosque P, Barnabé C, Padilla AM et al (2003) Multilocus enzyme electrophoresis analysis of *Trypanosoma cruzi* isolates from a geographically restricted endemic area for Chagas' disease in Argentina. Int J Parasitol 33:997–1003
- Diosque P, Padilla AM, Cimino RO et al (2004) Chagas disease in rural areas of Chaco Province, Argentina: epidemiologic survey in humans, reservoirs, and vectors. Am J Trop Med Hyg 71:590–593
- Diotaiuti L, Faria Filho OF, Carneiro FC et al (2000) Aspectos operacionais do controle do *Triatoma brasiliensis*. Cad Saude Publica 16:S61–S67
- Dujardin JP, Schofeld CJ (2004) Triatominae: systematics, morphology and population biology. In: Maudlin I, Holmes PH, Miles MA (eds) The trypanosomiases. CABI, Wallingford, pp 181–200
- Dumonteil E, Nouvellet P, Rosecrans K et al (2013) Eco-bio-social determinants for house infestation by nondomiciliated *Triatoma dimidiata* in the Yucatan península, Mexico. PLoS Negl Trop Dis 7:e2466
- Dumonteil E, Ramirez-Sierra MJ, Pérez-Carrillo S et al (2018) Detailed ecological associations of triatomines revealed by metabarcoding and next-generation sequencing: implications for triatomine behavior and *Trypanosoma cruzi* transmission cycles. Sci Rep 8:4140
- Dye C (1992) The analysis of parasite transmission by bloodsucking insects. Annu Rev Entomol 37:1–19
- Ehrenberg JP, Ault SK (2005) Neglected diseases of neglected populations: thinking to reshape the determinants of health in Latin America and the Caribbean. BMC Public Health 5:119
- Eickhoff CS, Dunn BA, Sullivan NA et al (2013) Comparison of the infectivity of *Trypanosoma cruzi* insect–derived metacyclic trypomastigotes after mucosal and cutaneous contaminative challenges. Mem Inst Oswaldo Cruz 108:508–511
- Elliot SL, Rodrigues JO, Lorenzo MG et al (2015) *Trypanosoma cruzi*, etiological agent of Chagas disease, is virulent to its triatomine vector *Rhodnius prolixus* in a temperature-dependent manner. PLoS Negl Trop Dis 9:e0003646
- Ellis BR, Wilcox BA (2009) The ecological dimensions of vector-borne disease research and control. Cad Saude Publica 25(supl1):S155–S167
- El-Sayed AM, Scarborough P, Seemann L et al (2012) Social network analysis and agent-based modeling in social epidemiology. Epidemiol Perspect Innov 9:1
- Enriquez GF, Cardinal MV, Orozco MM et al (2013) Discrete typing units of *Trypanosoma cruzi* identifed in rural dogs and cats in the humid Argentinean Chaco. Parasitology 140:303–308
- Enriquez GF, Bua J, Orozco MM et al (2014) High levels of *Trypanosoma cruzi* DNA determined by qPCR and infectiousness to *Triatoma infestans* support dogs and cats are major sources of parasites for domestic transmission. Infect Genet Evol 25:36–43
- Fabrizio MC, Schweigmann NJ, Bartoloni NJ (2016) Analysis of the transmission of Trypanosoma cruzi infection through hosts and vectors. Parasitology 143:1168–1178
- Feliciangeli MD, Campbell-Lendrum D, Martinez C et al (2003) Chagas disease control in Venezuela: lessons for the Andean region and beyond. Trends Parasitol 19:44–49
- Fernández MP, Gaspe MS, Gürtler RE (2019a) Inequalities in the social determinants of health and Chagas disease transmission risk in indigenous and creole households in the argentine Chaco. Parasit Vectors 12:184
- Fernández MP, Gaspe MS, Sartor P et al (2019b) Human *Trypanosoma cruzi* infection is driven by eco-social interactions in rural communities of the Argentine Chaco. PLoS Negl Trop Dis 13:e0007430
- Flores-Ferrer A, Waleckx E, Rascalou G et al (2019) *Trypanosoma cruzi* transmission dynamics in a synanthropic and domesticated host community. PLoS Negl Trop Dis 13:e000790
- Forattini OP, Ferreira OA, Silva EOR et al (1977) Aspectos ecológicos da tripanossomíase americana: IX-Variação e mobilidade de *Panstrongylus megistus* em ecótopos artifciais. Rev Saúde Pública 11:199–213
- Fortin MJ, Dale MRT (2005) Spatial analysis: a guide for ecologist. Cambridge University Press, Cambridge
- Freitas JLP (1950) Observacões sôbre xenodiagnósticos practicados em reservatórios domésticos e silvestres do *Trypanosoma cruzi* em uma localidade endémica da moléstia de Chagas no estado de São Paulo. Hosp Rio Janeiro 38:63–71
- Fürst T, Raso G, Acka CA et al (2009) Dynamics of socioeconomic risk factors for neglected tropical diseases and malaria in an armed confict. PLoS Negl Trop Dis 3:e513
- Garcia ES, Ratcliffe NA, Whitten MM et al (2007) Exploring the role of insect host factors in the dynamics of *Trypanosoma cruzi*–*Rhodnius prolixus* interactions. J Insect Physiol 53:11–21
- Gardiner BO, Maddrell SH (1972) Techniques for routine and large-scale rearing of *Rhodnius prolixus* Stål (Hem., Reduviidae). Bull Entomol Res 61:505–515
- Gaspe MS, Gurevitz JM, Gürtler RE et al (2013) Origins of house reinfestation with *Triatoma infestans* after insecticide spraying in the Argentine Chaco using wing geometric morphometry. Infect Genet Evol 17:93–100
- Gaspe MS, Provecho YM, Cardinal MV et al (2015) Ecological and sociodemographic determinants of house infestation with *Triatoma infestans* in indigenous communities in the Argentine Chaco. PLoS Negl Trop Dis 9:e0003614
- Gaspe MS, Provecho YM, Fernández MP et al (2018) Beating the odds: sustained Chagas disease vector control in remote indigenous communities of the Argentine Chaco over a seven-year period. PLoS Negl Trop Dis 12:e0006804
- Gaspe MS, Fernández MP, Cardinal MV et al (2020) Urbanisation, risk stratifcation and house infestation with the main vector *Triatoma infestans* in an endemic municipality of the Argentine Chaco. Parasit Vectors 13:316
- Giojalas LC, Catalá SS, Asin SN et al (1990) Seasonal changes in infectivity of domestic populations of *Triatoma infestans*. Trans R Soc Trop Med Hyg 84:439–442
- Gomes JE, Azambuja P, Garcia ES (1990) Comparative studies on the growth and reproductive performances of *Rhodnius prolixus* reared on different blood sources. Mem Inst Oswaldo Cruz 85:299–304
- Gorla DE (1991) Recovery of *Triatoma infestans* populations after insecticide application: an experimental feld study. Med Vet Entomol 5:311–324
- Gorla DE (1992) Population dynamics and control of *Triatoma infestans*. Med Vet Entomol 6:91–97
- Gorla DE, Noireau F (2017) Geographic distribution of Triatominae vectors in America. In: Tibayrenc M, Telleria J (eds) American Trypanosomiasis Chagas disease: one hundred years of research, 2nd edn. Elsevier, Saint Louis, pp 197–222
- Gorla DE, Schofeld CJ (1989) Population dynamics of *Triatoma infestans* under natural climatic conditions in the Argentine Chaco. Med Vet Entomol 3:179–194
- Gourbière S, Dorn P, Tripet F et al (2012) Genetics and evolution of triatomines: from phylogeny to vector control. Heredity (Edinb) 108:190–202
- Guarneri AA, Lorenzo MG (2017) Triatomine physiology in the context of trypanosome infection. J Insect Physiol 97:66–76
- Guhl F, Lazdins-Helds J, Grupo de trabajo científco sobre la enfermedad de Chagas (2007) Reporte sobre la enfermedad de Chagas. World Health Organization, Geneva
- Gurevitz JM, Ceballos LA, Gaspe MS et al (2011) Factors affecting infestation by *Triatoma infestans* in a rural area of the humid Chaco in Argentina: a multi-model inference approach. PLoS Negl Trop Dis 5:e1365
- Gurevitz JM, Gaspe MS, Enriquez GF et al (2013) Intensifed surveillance and insecticide-based control of the Chagas disease vector *Triatoma infestans* in the Argentinean Chaco. PLoS Negl Trop Dis 7:e2158
- Gürtler RE (2009) Sustainability of vector control strategies in the Gran Chaco region: current challenges and possible approaches. Mem Inst Oswaldo Cruz 104:52–59
- Gürtler RE, Cardinal MV (2015) Reservoir host competence and the epidemiologic role of domestic and commensal hosts in the transmission of *Trypanosoma cruzi*. Acta Trop 151:32–50
- Gürtler RE, Kravetz FO, Petersen RM et al (1990) The prevalence of *Trypanosoma cruzi* and the demography of dog populations after insecticidal spraying of houses: a predictive model. Ann Trop Med Parasitol 84:313323
- Gürtler RE, Cecere MC, Castañera MB et al (1996) Probability of infection with *Trypanosoma cruzi* of the vector *Triatoma infestans* fed on infected humans and dogs in Northwest Argentina. Am J Trop Med Hyg 55:24–31
- Gürtler RE, Cohen JE, Cecere MC et al (1997) Shifting host choices of the vector of Chagas disease *Triatoma infestans* in relation to the availability of hosts in houses in north-West Argentina. J Appl Ecol 34:699–715
- Gürtler RE, Chuit R, Cecere MC et al (1998a) Household prevalence of seropositivity for *Trypanosoma cruzi* in three rural villages in Northwest Argentina: environmental, demographic, and entomologic associations. Am J Trop Med Hyg 59:741–749
- Gürtler RE, Cohen JE, Cecere MC et al (1998b) Infuence of humans and domestic animals on the household prevalence of *Trypanosoma cruzi* in *Triatoma infestans* populations in Northwest Argentina. Am J Trop Med Hyg 58:748–758
- Gürtler RE, Cecere MC, Canale DM et al (1999) Monitoring house reinfestation by vectors of Chagas disease: a comparative trial of detection methods during a four-year follow-up. Acta Trop 72:213–234
- Gürtler RE, Canale DM, Spillmann C et al (2004) Effectiveness of residual spraying with deltamethrin and permethrin on peridomestic populations of *Triatoma infestans* in rural western Argentina: a district-wide randomized trial. Bull World Health Organ 82:196–220
- Gürtler RE, Cecere MC, Lauricella MA et al (2005) Incidence of *Trypanosoma cruzi* infection among children following domestic reinfestation after insecticide spraying in rural northwestern Argentina. Am J Trop Med Hyg 73:95–103
- Gürtler RE, Cecere MC, Lauricella MA et al (2007a) Domestic dogs and cats as sources of *Trypanosoma cruzi* infection in rural northwestern Argentina. Parasitology 134:69–82
- Gürtler RE, Kitron U, Cecere MC et al (2007b) Sustainable vector control and management of Chagas disease in the Gran Chaco, Argentina. Proc Natl Acad Sci USA 104:16194–16199
- Gürtler RE, Ceballos LA, Ordóñez-Krasnowski P et al (2009) Strong host-feeding preferences of the Chagas disease vector *Triatoma infestans* modifed by vector density: implications for the epidemiology of Chagas disease. PLoS Negl Trop Dis 3:e447
- Gürtler RE, Cecere MC, Vázquez-Prokopec GM et al (2014a) Domestic animal hosts strongly infuence human-feeding rates of the Chagas disease vector *Triatoma infestans* in Argentina. PLoS Negl Trop Dis 8:e2894
- Gürtler RE, Cecere MC, Fernández MP et al (2014b) Key source habitats and potential dispersal of *Triatoma infestans* populations in Northwestern Argentina: implications for vector control. PLoS Negl Trop Dis 8:e3238
- Gürtler RE, Fernández MP, Cecere MC et al (2017) Body size and hosts of *Triatoma infestans* populations affect the size of blood meal contents and female fecundity in rural northwestern Argentina. PLoS Negl Trop Dis 11:6097
- Guzman-Tapia Y, Ramírez-Sierra MJ, Dumonteil E (2007) Urban infestation by *Triatoma dimidiata* in the City of Mérida, Yucatán, México. Vector-Borne Zoonotic Dis 7:597–606
- Hagenlocher M, Castro MC (2015) Mapping malaria risk and vulnerability in the United Republic of Tanzania: a spatial explicit model. Popul Health Metrics 13:1–14
- Hall LS, Krausman PR, Morrison ML (1997) The habitat concept and a plea for standard terminology. Wildlife Soc Bull 25:171–182
- Hankivsky O, Christoffersen A (2008) Intersectionality and the determinants of health: a Canadian perspective. Crit Public Health 18:271–283
- Hotez P (2014) Ten global "hotspots" for the neglected tropical diseases. PLoS Negl Trop Dis 8:e2496
- Hotez PJ, Bottazzi ME, Franco-Paredes C et al (2008) The neglected tropical diseases of Latin America and the Caribbean: a review of disease burden and distribution and a roadmap for control and elimination. PLoS Negl Trop Dis 2:e300
- Houweling TAJ, Karim-Kos HE, Kulik MC et al (2016) Socioeconomic inequalities in neglected tropical diseases: a systematic review. PLoS Negl Trop Dis 10:e0004546
- Howard EJ, Xiong X, Carlier Y et al (2014) Frequency of the congenital transmission of *Trypanosoma cruzi*: a systematic review and meta-analysis. Int J Obstet Gynaecol 121:22–33
- Howe LD, Galobardes B, Matijasevich A et al (2012) Measuring socio-economic position for epidemiological studies in low- and middle-income countries: a methods of measurement in epidemiology paper. Int J Epidemiol 41:871–886
- Jácome-Pinilla D, Hincapie-Peñaloza E, Ortiz MI et al (2015) Risks associated with dispersive nocturnal fights of sylvatic Triatominae to artifcial lights in a model house in the northeastern plains of Colombia. Parasit Vectors 8:600
- Jansen AM, Xavier SC, Roque AL (2017) Ecological aspects of *Trypanosoma cruzi*: wild hosts and reservoirs. In: Tibayrenc M, Telleria J (eds) American Trypanosomiasis Chagas disease: one hundred years of research, 2nd edn. Elsevier, Saint Louis, pp 243–264
- Jirón LF, Zeledón R (1982) Preferencias alimentarias de tres especies de Triatominae (Hemiptera: Reduviidae) en condiciones experimentales. Rev Biol Trop 30:151–159
- Keesing F, Holt RD, Ostfeld RS (2006) Effects of species diversity on disease risk. Ecol Lett 9:485–498
- Kelly DW, Thompson CE (2000) Epidemiology and optimal foraging: modelling the ideal-free distribution of insect vectors. Parasitology 120:319–327
- Kirk ML, Schofeld CJ (1987) Density-dependent timing of defaecation by *Rhodnius prolixus*, and its implications for the transmission of *Trypanosoma cruzi*. Trans R Soc Trop Med Hyg 81:348–349
- Lardeux F, Depickère S, Aliaga C et al (2015) Experimental control of *Triatoma infestans* in poor rural villages of Bolivia through community participation. Trans R Soc Trop Med Hyg 109:150–158
- Lazzari CR, Pereira MH, Lorenzo MG (2013) Behavioural biology of Chagas disease vectors. Mem Inst Oswaldo Cruz 108(Suppl 1):34–47
- Ledesma Patiño OSR, Ribas Meneclierr CA, Kalalo E et al (1992) Epidemiología, clínica y laboratorio de la enfermedad de Chagas aguda en Santiago del Estero. In: Madoery RJ, Madoery C, Cámera MI (eds) Actualizaciones de la enfermedad de Chagas. Libro del Organismo Ofcial del Congreso Nacional de Medicina, Córdoba, pp 39–50
- Lee BY, Bacon KM, Bottazzi ME et al (2013) Global economic burden of Chagas disease: a computational simulation model. Lancet Infect Dis 13:342–348
- Lehane MJ (2005) The biology of blood-sucking in insects, 2nd edn. Cambridge University Press, Cambridge
- Levy MZ, Bowman NM, Kawai V et al (2006) Periurban *Trypanosoma cruzi*-infected *Triatoma infestans*, Arequipa, Peru. Emerg Infect Dis 12:1345–1352
- Levy MZ, Kawai V, Bowman NM et al (2007) Targeted screening strategies to detect *Trypanosoma cruzi* infection in children. PLoS Negl Trop Dis 1:e103
- Levy MZ, Bowman NM, Kawai V et al (2009) Spatial patterns in discordant diagnostic test results for Chagas disease: links to transmission hotspots. Clin Infect Dis 48:1104–1106
- Levy MZ, Barbu CM, Castillo-Neyra R et al (2014) Urbanization, land tenure security and vectorborne Chagas disease. Proc R Soc B Biol Sci 281:20141003–20141003
- Lima MM, Sarquis O, de Oliveira TG et al (2012) Investigation of Chagas disease in four periurban areas in northeastern Brazil: epidemiologic survey in man, vectors, non-human hosts and reservoirs. Trans R Soc Trop Med Hyg 106:143–149
- López A, Crocco L, Morales G et al (1999) Feeding frequency and nutritional status of peridomestic populations of *Triatoma infestans* from Argentina. Acta Trop 73:275–281
- López-Cancino SA, Tun-Ku E, De la Cruz-Felix HK et al (2015) Landscape ecology of *Trypanosoma cruzi* in the southern Yucatan Peninsula. Acta Trop 151:58–72
- Lorenzo Figueiras AN, Lazzari CR (1998) Aggregation behaviour and interspecifc responses in three species of Triatominae. Mem Inst Oswaldo Cruz 93:133–137
- Lorenzo MG, Lazzari CR (1996) The spatial pattern of defaecation in *Triatoma infestans* and the role of faeces as a chemical mark of the refuge. J Insect Physiol 42:903–907
- Lorenzo MG, Lazzari CR (1999) Temperature and relative humidity affect the selection of shelters by *Triatoma infestans*, vector of Chagas disease. Acta Trop 72:241–249
- Lucero RH, Brusés BL, Cura CI et al (2016) Chagas' disease in aboriginal and creole communities from the Gran Chaco region of Argentina: Seroprevalence and molecular parasitological characterization. Infect Genet Evol 41:84–92
- Lugones H, Ledesma O, Storino R et al (1994) Chagas agudo. In: Storino R, Milei J (eds) Enfermedad de Chagas. Mosby Doyma, Buenos Aires, pp 209–234
- Macchiaverna NP, Gaspe MS, Enriquez GF et al (2015) *Trypanosoma cruzi* infection in *Triatoma sordida* before and after community-wide residual insecticide spraying in the Argentinean Chaco. Acta Trop 143:97–102
- Macchiaverna NP, Enriquez GF, Buscaglia CA (2018) New human isolates confrm the predominance of hybrid lineages in the Argentine domestic cycle of *Trypanosoma cruzi*. Infect Genet Evol 66:229–235
- Macchiaverna NP, Enriquez GF, Bua J et al (2020) Human infectiousness and parasite load in chronic patients seropositive for *Trypanosoma cruzi* in a rural area of the Argentine Chaco. Infect Genet Evol 78:104062
- MacGregor-Fors I (2011) Misconceptions or misunderstandings? On the standardization of basic terms and defnitions in urban ecology. Landsc Urban Plan 100:347–349
- Maguire JH, Mott KE, Hoff R et al (1982) A three-year follow-up study of infection with *Trypanosoma cruzi* and electrocardiographic abnormalities in a rural community in Northeast Brazil. Am J Trop Med Hyg 31:42–47
- Maidana CA (2011) Migrantes Toba (Qom). Procesos de territorización y construcción de identidades.<http://sedici.unlp.edu.ar/handle/10915/21132>
- Manderson L, Aagaard-Hansen J, Allotey P et al (2009) Social research on neglected diseases of poverty: continuing and emerging themes. PLoS Negl Trop Dis 3:e332
- Marcili A, Lima L, Cavazzana M et al (2009) A new genotype of *Trypanosoma cruzi* associated with bats evidenced by phylogenetic analyses using SSU rDNA, cytochrome b and Histone H2B genes and genotyping based on ITS1 rDNA. Parasitology 136:641–655
- Marliére NP, Latorre-Estivalis JM, Lorenzo MG et al (2015) Trypanosomes modify the behavior of their insect hosts: effects on locomotion and on the expression of a related gene. PLoS Negl Trop Dis 9:e0003973
- Marsden PD, Virgens D, Magalhães I et al (1982) Domestic ecology of *Triatoma infestans* in Mambaí, Goiás-Brazil. Rev Inst Med Trop São Paulo 24:364–373
- Matter HC, Daniels TJ (2000) Dog ecology and population biology. In: Macpherson CNL, Meslin FX, Wandeler AI (eds) Dogs, Zoonoses and public health. CABI Publication, New York, pp 17–50
- McEwen PK, Lehane MJ, Whitaker CJ (1993) The effect of adult population density on fight initiation in *Triatoma infestans* (Klug) (Hem., Reduviidae). J Appl Entomol 116:321–325
- Meadows DH (2008) Thinking in systems: a primer. Earthscan, London
- Medone P, Ceccarelli S, Parham PE et al (2015) The impact of climate change on the geographic distribution of two vectors of Chagas disease: implications for the force of infection. Philos Trans R Soc B 370:20130560
- Medrano-Mercado N, Ugarte-Fernandez R, Butrón V et al (2008) Urban transmission of Chagas disease in Cochabamba, Bolivia. Mem Inst Oswaldo Cruz 103:423–430
- Mejía-Jaramillo AM, Agudelo-Uribe LA, Dib JC et al (2014) Genotyping of *Trypanosoma cruzi* in a hyper-endemic area of Colombia reveals an overlap among domestic and sylvatic cycles of Chagas disease. Parasit Vectors 7:108
- Miles MA, Feliciangeli MD, Rojas de Arias A (2003) American trypanosomiasis (Chagas' disease) and the role of molecular epidemiology in guiding control strategies. Br Med J 326:1444–1448
- Minter DM (1976) Feeding patterns of some Triatominae vectors. In: New approaches in American Trypanosomiasis research, Pan American health organization scientifc publication no. 318, Washington, pp 33–47
- Minter-Goedbloed E, Minter DM, Marshall TF (1978) Quantitative comparison between xenodiagnosis and haemoculture in the detection of *Trypanosoma* (*Schizotrypanum*) *cruzi* in experimental and natural chronic infections. Trans R Soc Trop Med Hyg 72:217–225
- Monroy C, Bustamante DM, Pineda S et al (2009) House improvements and community participation in the control of *Triatoma dimidiate* re-infestation in Jutiapa, Guatemala. Cad Saude Publica 25:S168–S178
- Moretti E, Castro I, Franceschi C et al (2010) Chagas disease: serological and electrocardiographic studies in Wichi and Creole communities of Misión Nueva Pompeya, Chaco, Argentina. Mem Inst Oswaldo Cruz 105:621–627
- Morillo CA, Marin-Neto JA, Avezum A et al (2015) Randomized trial of benznidazole for chronic Chagas' cardiomyopathy. N Engl J Med 373:1295–1306
- Mott KE, Lehman JS, Hoff R et al (1976) The epidemiology and household distribution of seroreactivity to *Trypanosoma cruzi* in a rural community in Northeast Brazil. Am J Trop Med Hyg 25:552–562
- Mott KE, Muniz TM, Lehman JS Jr et al (1978a) House construction, triatomine distribution, and household distribution of seroreactivity to *Trypanosoma cruzi* in a rural community in Northeast Brazil. Am J Trop Med Hyg 27:1116–1122
- Mott KE, Mota EA, Sherlock I et al (1978b) *Trypanosoma cruzi* infection in dogs and cats and household seroreactivity to *T. cruzi* in a rural community in Northeast Brazil. Am J Trop Hyg 27:1123–1127
- Muench H (1959) Catalytic models in epidemiology. Harvard University Press, Cambridge
- Neves DP (1971) Infuencia da temperatura na evolucão do *Trypanosoma cruzi* em triatominos. Rev Inst Med Trop São Paulo 13:155–161
- Nijhout HF, Riddiford LM, Mirth C et al (2014) The developmental control of size in insects. Wiley Interdiscip Rev Dev Biol 3:113–134
- Noireau F, Dujardin JP (2010) Biology of Triatominae. In: Tibayrenc M, Telleria J (eds) American Trypanosomiasis Chagas disease: one hundred years of research. Elsevier, Saint Louis, pp 149–168
- Nouvellet P, Dumonteil E, Gourbière S (2013) The improbable transmission of *Trypanosoma cruzi* to human: the missing link in the dynamics and control of Chagas disease. PLoS Negl Trop Dis 7:e2505
- Nouvellet P, Cucunubá ZM, Gourbière S (2015) Ecology, evolution and control of Chagas disease: a century of neglected modelling and a promising future. Adv Parasitol 87:135–191
- Núñez JA, Segura EL (1987) Rearing of Triatominae. In: Brenner RR, Stoka AM (eds) Chagas' disease vectors, vol II. CRC Press, Florida, pp 31–40
- Ordóñez-Krasnovsky P, Lanatti L, Gaspe MS et al (2020) Domestic host availability modifes human-triatomine contact and host shifts of the Chagas disease vector *Triatoma infestans* in the humid Argentine Chaco. Med Vet Entomol 34:459–469
- Orozco MM, Enriquez GF, Alvarado-Otegui JA et al (2013) New sylvatic hosts of *Trypanosoma cruzi* and their reservoir competence in the Humid Chaco of Argentina: a longitudinal study. Am J Trop Med Hyg 88:872–882
- Pacheco-Tucuch FS, Ramirez-Sierra MJ, Gourbière S et al (2012) Public street lights increase house infestation by the Chagas disease vector *Triatoma dimidiata*. PLoS One 7:e36207
- Payet V, Ramirez-Sierra M, Rabinovich JE et al (2009) Variations in sex ratio, feeding, and fecundity of *Triatoma dimidiata* (Hemiptera: Reduviidae) among habitats in the Yucatan Peninsula, Mexico. Vector-Borne Zoonotic Dis 9:243–251
- Pennington PM, Messenger LA, Reina J et al (2015) The Chagas disease domestic transmission cycle in Guatemala: parasite-vector switches and lack of mitochondrial co-diversifcation between *Triatoma dimidiata* and *Trypanosoma cruzi* subpopulations suggest non-vectorial parasite dispersal across the Motagua valley. Acta Trop 151:80–87
- Perlowagora-Szumlewicz A (1975) Laboratory colonies of Triatominae, biology, and population dynamics. In: New approaches in American Trypanosomiasis research. Pan American Health Organization Scientifc Publication no. 318, Washington, pp 18–21
- Peterson JK, Bartsch SM, Lee BY et al (2015) Broad patterns in domestic vector-borne *Trypanosoma cruzi* transmission dynamics: synanthropic animals and vector control. Parasit Vectors 8:537
- Piccinali RV, Canale DM, Sandoval AE et al (2010) *Triatoma infestans* bugs in southern Patagonia. Emerg Infect Dis 16:887–889
- Piesman J, Sherlock IA, Christensen HA (1983) Host availability limits population density of *Panstrongylus megistus*. Am J Trop Med Hyg 32:1445–1450
- Piesman J, Sherlock IA, Mota E (1985) Association between household triatomine density and incidence of *Trypanosoma cruzi* infection during a nine-year study in Castro Alves, Bahia, Brazil. Am J Trop Med Hyg 34:866–869
- Pifano CF (1973) La dinámica epidemiológica de la enfermedad de Chagas en el Valle de los Naranjos, Estado Carabobo, Venezuela. II. La infección chagásica en la población rural del área. Arch Venez Med Trop Parasitol Med 5:31–45
- Pluciński MM, Ngonghala CN, Getz WM et al (2013) Clusters of poverty and disease emerge from feedbacks on an epidemiological network. J R Soc Interface 10:20120656
- Provecho YM, Gaspe MS, Fernández MP et al (2017) House reinfestation with *Triatoma infestans* (Hemiptera: Reduviidae) after community-wide spraying with insecticides in the Argentine Chaco: a multifactorial process. J Med Entomol 54:646–657
- Rabinovich JE (1972) Vital statistics of triatominae (Hemiptera: Reduviidae) under laboratory conditions. I *Triatoma infestans*. Klug J Med Entomol 9:351–370
- Rabinovich JE (1985) Ecología poblacional de los triatominos. In: Carcavallo RU, Rabinovich JE, Tonn RJ (eds) Factores Biológicos y Ecológicos en la Enfermedad de Chagas, 1. Servicio Nacional de Chagas (Argentina), Buenos Aires, pp 121–147
- Rabinovich JE, Feliciangeli MD (2015) Vital statistics of Triatominae (Hemiptera: Reduviidae) under laboratory conditions: IV. *Panstrongylus geniculatus*. J Med Entomol 52:797–805
- Rabinovich JE, Leal JA, Feliciangeli de Piñero D (1979) Domiciliary biting frequency and blood ingestion of the Chagas's disease vector *Rhodnius prolixus* Ståhl (Hemiptera: Reduviidae), in Venezuela. Trans R Soc Trop Med Hyg 73:272–283
- Rabinovich JE, Solarz ND, Gürtler RE et al (1990) Probability of transmission of Chagas disease by *Triatoma infestans* (Hemiptera: Reduviidae) in an endemic area of Santiago del Estero, Argentina. Bull World Health Organ 68:737–746
- Rabinovich JE, Kitron UD, Obed Y et al (2011) Ecological patterns of blood-feeding by kissingbugs (Hemiptera: Reduviidae: Triatominae). Mem Inst Oswaldo Cruz 106:479–494
- Ramirez-Sierra MJ, Herrera-Aguilar M, Gourbière S et al (2010) Patterns of house infestation dynamics by non-domiciliated *Triatoma dimidiata* reveal a spatial gradient of infestation in

rural villages and potential insect manipulation by *Trypanosoma cruzi*. Tropical Med Int Health 15:77–86

- Ramsey JM, Cruz-Celis A, Salgado L et al (2003) Effcacy of pyrethroid insecticides against domestic and peridomestic populations of *Triatoma pallidipennis* and *Triatoma barberi* (Reduviidae: Triatominae) vectors of Chagas' disease in Mexico. J Med Entomol 40:912–920
- Ramsey JM, Alvear AL, Ordoñez R et al (2005) Risk factors associated with house infestation by the Chagas disease vector *Triatoma pallidipennis* in Cuernavaca metropolitan area, Mexico. Med Vet Entomol 19:219–228
- Ramsey JM, Gutiérrez-Cabrera AE, Salgado-Ramírez et al (2012) Ecological connectivity of *Trypanosoma cruzi* reservoirs and *Triatoma pallidipennis* hosts in an anthropogenic landscape with endemic Chagas disease. PLoS One 7:e46013
- Raso G, Utzinger J, Silue KD et al (2005) Disparities in parasitic infections, perceived ill health and access to health care among poorer and less poor schoolchildren of rural Cote d'Ivoire. Tropical Med Int Health 10:42–57
- Rebosolán JB, Terzano R (1958) Quinientos casos de chagasosis aguda en Santiago del Estero. Mis Est Patol Reg Arg 87–88:55–57
- Rocha e Silva EO, Souza JMP, Andrade JCR et al (1977) Preferencia alimentar (entre sangue humano e ave) dos *Triatoma sordida* encontrados em casas habitadas da região norte do estado de São Paulo, Brasil. Rev Saude Publica 11:258–269
- Rodríguez-Planes LI, Vazquez-Prokopec GM, Cecere MC et al (2016) Selective insecticide applications directed against *Triatoma infestans* (Hemiptera: Reduviidae) affected a nontarget secondary vector of Chagas disease, *Triatoma garciabesi*. J Med Entomol 53:144–151
- Rodríguez-Planes LR, Gaspe MS, Enriquez GF et al (2018) Habitat-specifc occupancy and a metapopulation model of *Triatoma sordida* (Hemiptera: Reduviidae), a secondary vector of Chagas disease, in Northeastern Argentina. J Med Entomol 55:370–381
- Rodríguez-Planes LI, Gaspe MS, Enriquez GF et al (2020) Impacts of residual insecticide spraying on house infestation with *Triatoma sordida* and co-occurrence of *Triatoma infestans*: a threeyear follow-up in northeastern Argentina. Acta Trop 202:105251
- Rojas de Arias A, Ferro EA, Ferreira ME et al (1999) Chagas disease vector control through different intervention modalities in endemic localities of Paraguay. Bull World Health Organ 77:331–339
- Rojas de Arias A, Abad-Franch F, Acosta N et al (2012) Post-control surveillance of *Triatoma infestans* and *Triatoma sordida* with chemically-baited sticky traps. PLoS Negl Trop Dis 6:e1822
- Romaña C (1963) Enfermedad de Chagas. Lopez Libreros Editores, Buenos Aires
- Rosal GG, Nogueda-Torres B, Villagrán ME et al (2018) Chagas disease: importance of rats as reservoir hosts of *Trypanosoma cruzi* (Chagas, 1909) in western Mexico. J Infect Public Health 11:230–233
- Rosenbaum M, Cerisola JA (1961) Epidemiología de la Enfermedad de Chagas. O Hospital 60:75–123
- Samuels AM, Clark EH, Galdos-Cardenas G et al (2013) Epidemiology of and impact of insecticide spraying on Chagas disease in communities in the Bolivian Chaco. PLoS Negl Trop Dis 7:e2358
- Sandoval CM, Gutiérrez R, Luna S et al (2000a) High density of *Rhodnius prolixus* in a rural house in Colombia. Trans R Soc Trop Med Hyg 94:372–373
- Sandoval CM, Joya MI, Gutierez R et al (2000b) Cleptohaematophagy of the triatomine bug *Belminus herreri*. Med Vet Entomol 14:100–101
- Schaub GA (1992) The effects of trypanosomatids on insects. Adv Parasitol 31:255–319
- Schmunis GA, Yadon ZE (2010) Chagas disease: a Latin American health problem becoming a world health problem. Acta Trop 115:14–21
- Schofeld CJ (1980) Nutritional status of domestic populations of *Triatoma infestans*. Trans R Soc Trop Med Hyg 74:770–778
- Schofeld CJ (1982) The role of blood intake in density regulation of populations of *Triatoma infestans* (Klug) (Hemiptera: Reduviidae). Bull Entomol Res 72:617–629
- Schofeld CJ (1985) Population dynamics and control of *Triatoma infestans*. Ann Soc Belge Med Trop 65:149–164
- Schofeld CJ (1994) Triatominae—biology and control. Eurocommunica Publications, Bognor Regis
- Sicuri E, Muñoz J, Pinazo MJ et al (2011) Economic evaluation of Chagas disease screening of pregnant Latin American women and of their infants in a non endemic area. Acta Trop 118:110–117
- Solar O, Irwin A (2010) A conceptual framework for action on the social determinants of health. Social determinants of health discussion. Paper 2 (policy and practice). World Health Organization, Geneva
- Sosa-Estani S, Dri LL, Touris C et al (2009) Transmisión vectorial y congénita del *Trypanosoma cruzi* en Las Lomitas, Formosa. Medicina (Buenos Aires) 69:424–430
- Spagnuolo AM, Shillor M, Kingsland L et al (2012) A logistic delay differential equation model for Chagas disease with interrupted spraying schedules. J Biol Dyn 6:377–394
- Spiegel J, Bennett S, Hattersley L et al (2005) Barriers and bridges to prevention and control of dengue: the need for a social–ecological approach. EcoHealth 2:273–290
- Stoddard ST, Morrison AC, Vazquez-Prokopec GM et al (2009) The role of human movement in the transmission of vector-borne pathogens. PLoS Negl Trop Dis 3:e481
- Taranto NJ, Cajal SP, De Marzi MC et al (2003) Clinical status and parasitic infection in a Wichí aboriginal community in Salta, Argentina. Trans R Soc Trop Med Hyg 97:554–558
- Trumper E, Gorla DE (1991) Density dependent timing of defaecation by *Triatoma infestans*. Trans R Soc Trop Med Hyg 85:800–802
- Valle D, Lima Gomes JEP, Goldenberg S et al (1987) *Rhodnius prolixus* vitellogenesis: dependence upon the blood source. J Insect Physiol 33:249–254
- Vallvé SL, Rojo H, Wisnivesky-Colli C (1996) Urban ecology of *Triatoma infestans* in San Juan, Argentina. Mem Inst Oswaldo Cruz 91:405–408
- Vazquez-Prokopec GM, Ceballos LA, Cecere MC et al (2002) Seasonal variations of microclimatic conditions in domestic and peridomestic habitats of *Triatoma infestans* (Hemiptera: Reduviidae) in rural Northwest Argentina. Acta Trop 84:229–238
- Vazquez-Prokopec GM, Ceballos LA, Marcet PL et al (2006) Seasonal variations in active dispersal of natural populations of *Triatoma infestans* in rural North-Western Argentina. Med Vet Entomol 20:273–279
- Ventura-Garcia L, Roura M, Pell C et al (2013) Socio-cultural aspects of Chagas disease: a systematic review of qualitative research. PLoS Negl Trop Dis 7:e2410
- Viotti R, Vigliano CA, Álvarez MG et al (2009) El impacto de las condiciones socioeconómicas en la evolución de la enfermedad de Chagas crónica. Rev Española Cardiol 62:1224–1232
- Waleckx E, Suarez J, Richards B et al (2014) *Triatoma sanguisuga* blood meals and potential for Chagas disease, Louisiana, USA. Emerg Infect Dis 20:2141–2143
- Waleckx E, Gourbière S, Dumonteil E (2015a) Intrusive versus domiciliated triatomines and the challenge of adapting vector control practices against Chagas disease. Mem Inst Oswaldo Cruz 110:324–338
- Waleckx E, Camara-Mejia J, Ramirez-Sierra MJ et al (2015b) An innovative ecohealth intervention for Chagas disease vector control in Yucatan, Mexico. Trans R Soc Trop Med Hyg 109:143–149
- Walter A, Lozano-Kasten F, Bosseno MF et al (2007) Peridomicilary habitat and risk factors for *Triatoma* infestation in a rural community of the Mexican occident. Am J Trop Med Hyg 76:508–515
- Ward JP, Baker PS (1982) The tethered fight performance of a laboratory population of *Triatoma infestans* (Klug) (Hemiptera: Reduviidae). Bull Entomol Res 72:17–28
- Wiegand T, Moloney KA (2014) Handbook of spatial point-pattern analysis in ecology. CRC Press, Boca Raton
- WisniveskyColli C, Gürtler RE, Solarz ND et al (1985) Epidemiological role of humans, dogs and cats in the transmission of *Trypanosoma cruzi* in a central area of Argentina. Rev Inst Med Trop São Paulo 27:346352
- Wisnivesky-Colli C (1987) Feeding patterns of Triatominae in relation to transmission of American trypanosomiasis. In: Brenner RR, Stoka AM (eds) Chagas disease vectors, vol 1. CRC Press, Boca Raton, pp 99–117
- Woolhouse ME, Dye C, Etard JF et al (1997) Heterogeneities in the transmission of infectious agents: implications for the design of control programs. Proc Natl Acad Sci USA 94:338–342
- World Health Organization (2002) Control of Chagas disease. Second report of the WHO expert committee. WHO technical report series 905. World Health Organization, Geneva
- World Health Organization (2008) WHO position statement on integrated vector management. Wkly Epidemiol Rec 20:177–181
- World Health Organization (2015) Chagas disease in Latin America: an epidemiological update based on 2010 estimates. Wkly Epidemiol Rec 90:33–44
- Yeo M, Acosta N, Llewellyn MS et al (2005) Origins of Chagas disease: *Didelphis* species are natural hosts of *Trypanosoma cruzi* I and armadillos hosts of *Trypanosoma cruzi* II, including hybrids. Int J Parasitol 35:225–233
- Zacharias CA, Pontes GB, Lorenzo MG et al (2010) Flight initiation by male *Rhodnius prolixus* is promoted by female odors. J Chem Ecol 36:449–451
- Zeledón R, Vargas LG (1984) The role of dirt foors and of frewood in rural dwellings in the epidemiology of Chagas' disease in Costa Rica. Am J Trop Med Hyg 33:232–235
- Zeledón R, Solano G, Burstin L et al (1975) Epidemiological pattern of Chagas' disease in an endemic area of Costa Rica. Am J Trop Med Hyg 24:214–225
- Zeledón R, Alvarado R, Jirón LF (1977) Observations on the feeding and defecation patterns of three triatomine species (Hemiptera: Reduviidae). Acta Trop 34:65–77
- Zeledón R, Beard CB, Dias JP et al (2012) An appraisal of the status of Chagas disease in the United States. Elsevier, Oxford
- Zingales B (2018) *Trypanosoma cruzi* genetic diversity: something new for something known about Chagas disease manifestations, serodiagnosis and drug sensitivity. Acta Trop 184:38–52
- Zingales B, Miles MA, Campbell DA et al (2012) The revised *Trypanosoma cruzi* subspecifc nomenclature: rationale, epidemiological relevance and research applications. Infect Genet Evol 12:240–253
- zu Dohna H, Cecere MC, Gürtler RE et al (2009) Spatial re-establishment dynamics of local populations of vectors of Chagas disease. PLoS Negl Trop Dis e490:3