

Chapter 12

Congenital and Developmental Causes of Cystic Lung Disease



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Abbreviations

CCAM	Congenital cystic adenomatoid malformation
CHAOS	Congenital high airway obstruction syndrome
CLO	Congenital lobar overinflation
CNLD	Chronic neonatal lung disease
CPAM	Congenital pulmonary airway malformation
ELS	Extralobar sequestrations
FGFR2	Fibroblast growth factor receptor 2
FLNA	Filamin A
ILS	Intralobar sequestrations
miRNAs	Micro-RNAs
NKX2-1	NK2 homeobox 1
PPB	Pleuropulmonary blastoma
TBX4	T-box transcription factor 4
TTF-1	Thyroid transcription factor 1

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Congenital Cystic Lung Malformations

The widespread use of antenatal ultrasound has led to increased in utero detection of congenital cystic lung malformations. These entities comprise a group of rare but clinically significant developmental abnormalities, including bronchogenic cysts, congenital pulmonary airway malformation, pulmonary sequestration, congenital lobar emphysema, and pulmonary hyperplasia. Although many of these lesions will remain asymptomatic, there is a wide range of clinical complications including hydrops fetalis, airway obstruction, infection, and rarely neoplasia [1]. Treatment of even asymptomatic congenital lung lesions is variable with some managed conservatively, while others surgically resected [2]. Many congenital lung abnormalities detected early in pregnancy will have regressed significantly by birth, further complicating management decisions [3]. While the conventional nomenclature of these malformations infers that congenital cystic lung malformations are distinct lesions, they frequently share common modes of pathogenesis and histological features [4–7]. Specifically, bronchogenic cysts and extralobar sequestration reflect abnormal foregut budding during embryonic development, while congenital cystic pulmonary airway malformations, intralobar sequestration, pulmonary hyperplasia, and congenital lobar overinflation are often related to airway obstruction, with or without secondary dysplastic changes. In an attempt to bring clarity to the origin of these lesions and guide clinical management decisions, revised classifications have been proposed based on the anatomical/pathological findings or imaging appearance of the entities [4, 8–10] (Table 12.1).

Bronchogenic Cyst

Bronchogenic or bronchial cysts arise from anomalous budding of the foregut during development. They present as unilocular fluid or mucin-filled cysts that histologically recapitulate the bronchial structure, without connection to the lung (Fig. 12.1).

Bronchogenic cysts typically occur in the mediastinum or subcarinal area but may be intraparenchymal or even subdiaphragmatic or cutaneous [11, 12]. On radiological imaging, bronchogenic cysts appear as round thin-walled cystic cavities with sharp margins that may have air-fluid levels. They do not communicate with the normal tracheobronchial tree unless they become secondarily infected, which commonly occurs in intraparenchymal cysts. The time of clinical presentation is variable and related to compression of adjacent structures (e.g., trachea, bronchi, esophagus), infection, or rupture. Patients usually present later in life with dyspnea, recurrent coughing, wheezing, dysphagia, or pneumonia, but neonatal presentation or incidental detection on imaging taken for unrelated reasons is not unusual. On pathological examination, bronchogenic cysts consist of a single cyst containing clear or turbid fluid, the latter reflective of secondary infection. Management consists of surgical resection.

Table 12.1 Langston classification and radiographic classification of congenital lung malformations

Langston classification^a
I. Bronchopulmonary malformations
Bronchogenic cyst
Bronchial atresia
Cystic pulmonary airway malformation, large cyst type (Stocker type 1)
Isolated
With systemic arterial/venous connection (hybrid lesion/intralobar sequestration)
Cystic pulmonary airway malformation, small cyst type (Stocker type 2)
Extralobar sequestration (with/without connection to gastrointestinal tract)
Isolated
With cystic pulmonary airway malformation, small cyst type (hybrid lesion)
II. Pulmonary hyperplasia and related lesions
Congenital high airway malformation
Solid congenital pulmonary airway malformation, small cyst type (Stocker type 3)
Polyalveolar lobe
III. Congenital lobar emphysema
Radiographic Classification^b
I. Group 1 (congenital solid/cystic lung malformation)
Cystic pulmonary airway malformation, small cyst type (Stocker type 2)
Extralobar and intralobar sequestration
Bronchial atresia
Bronchogenic cyst
Mixed malformation
II. Group 2 (congenital hyperlucent lobe)
Congenital lobar emphysema
Polyalveolar lobe
III. Group 3 (congenital small lung)
Lung/lobar agenesis
Pulmonary hypoplasia

^aAdapted from [4]

^bAdapted from [10]

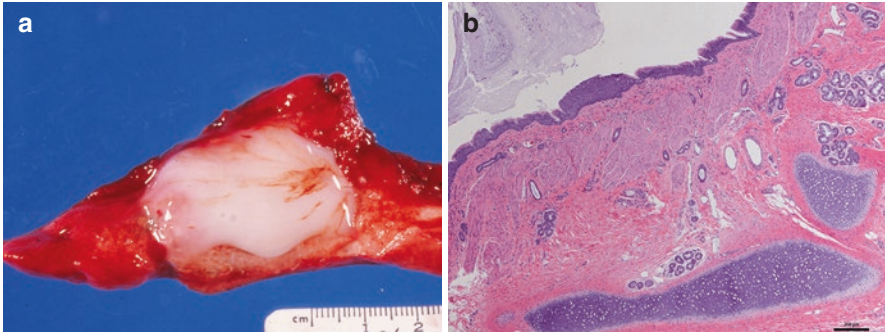


Fig. 12.1 Bronchogenic cyst. (a) An intraparenchymal bronchogenic cyst containing abundant mucus. (b) Bronchogenic cysts recapitulate the appearance of the normal bronchus lined by ciliated respiratory epithelium and containing smooth muscle, submucosal glands, and plates of cartilage in the wall (H&E)

Congenital Pulmonary Airway Malformation

Congenital pulmonary airway malformation (CPAM), previously known as congenital cystic adenomatoid malformation (CCAM), is the most common lung malformation of the lower respiratory tract [13]. CPAMs derive their blood supply from the pulmonary artery and drain via the pulmonary veins, with the exception of hybrid lesions (CPAM in pulmonary sequestrations) that have a systemic blood supply. CPAMs occur sporadically with no known genetic predilection. Mechanisms resulting in CPAM formation are debated, but the hypothesis that CPAMs arise from in utero obstruction during development has been substantiated by several studies [5–7].

Historically CPAMs have been categorized into separate subtypes in the Stocker classification, which is based upon cyst size and histological resemblance to segments of the normal respiratory tree, from trachea and large bronchi to peripheral acinar structures [14]. However, with further insights into CPAM pathogenesis, experience with the clinical behavior of this entity and frequent occurrence of hybrid lesions (CPAM in pulmonary sequestrations), the continued utility of the Stocker classification is contested [4]. Stocker type 0 CPAM is now recognized to be acinar dysplasia, a rare diffuse developmental lung disorder associated with mutations in the T-box transcription factor *TBX4* and fibroblast growth factor receptor 2 (*FGFR2*) [15, 16]. Stocker type 3 CPAM resembles pulmonary hyperplasia, and Stocker type 4 CPAM is thought to represent type I pleuropulmonary blastoma (discussed below). Langston divided the most frequent CPAMs into two types: a large cyst type (equivalent to Stocker type 1) and a small cyst type (equivalent to Stocker type 2). This terminology has in part been adapted in the radiographical literature to guide clinical management [17, 18] (see Table 12.1). By histological assessment, the small cyst type is the most common CPAM subtype [19].

Large cyst CPAM presents by radiographical imaging as a solitary lung mass containing variably sized cysts, usually measuring greater than 2 cm. The cysts replace the normal lung parenchyma and may be unilocular or multilocular and septated

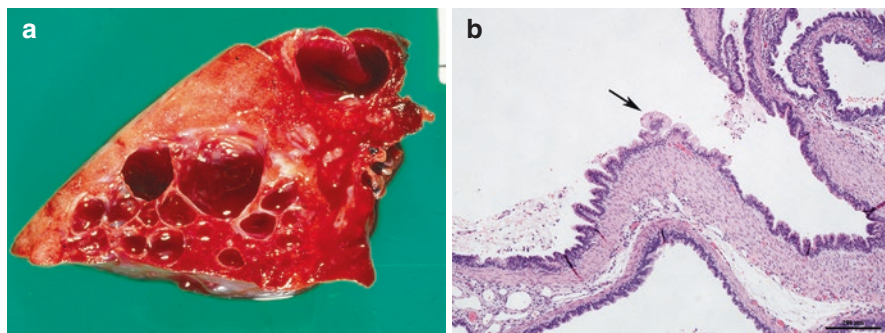


Fig. 12.2 Congenital pulmonary airway malformation (CPAM), large cyst type. (a) Large cyst type CPAM has cysts of variable size, usually measuring greater than 2 cm. (b) Microscopically, the cysts are lined by ciliated respiratory epithelium with smooth muscle in the wall; focal mucigenic epithelium is typically seen (*arrow*) (H&E)

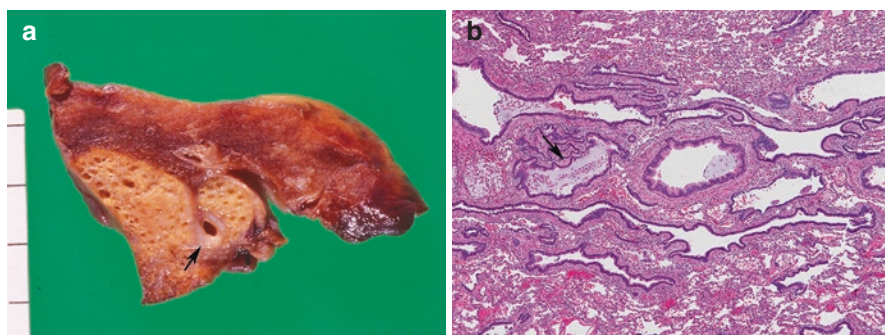


Fig. 12.3 Congenital pulmonary airway malformation (CPAM), small cyst type. (a) Small cyst type CPAM has multiple small cysts admixed with solid areas of parenchyma. A bronchus is present within the lesion (*arrow*). (b) Histologic sections show abnormally apposed bronchiole-like structures, many with intraluminal mucus providing evidence of obstruction (*arrow*) (H&E, 100 \times)

(Figure 12.2a). Histological examination demonstrates that the cysts are lined by ciliated respiratory epithelium, often containing clusters of mucigenic cells [19] (Figure 12.2b). Cartilage is rarely present in the cyst walls.

Small cyst CPAM is identified as an echogenic mass by ultrasound examination and is comprised of multiple small cysts, which can be heterogeneous in appearance based on the presence of cystic and solid components (Figure 12.3a). Small cyst CPAM may be associated with other congenital anomalies, including cardiovascular and renal malformations as well as diaphragmatic hernia [14]. Bronchial atresia is more frequently identified in small cyst than large cyst CPAM. Features of bronchial atresia are also commonly seen in pulmonary sequestrations with systemic blood supply [4]. Histologically a small cyst CPAM is composed of increased bronchiolar-like profiles lined by cuboidal to ciliated respiratory epithelium that, unlike normal bronchioles, are not accompanied by arteries (see Figure 12.3b).

Rhabdomyomatous differentiation with small bundles of mature skeletal muscle may be present around the cysts and is not reflective of rhabdomyosarcoma. CPAMs resected during fetal life demonstrate significant immaturity akin to the lung during early development and do not fit well into existing classification schemes [20].

The presentation of CPAMs is quite variable; patients may present with respiratory distress in early infancy or remain asymptomatic until later in life. The vast majority of cases are now detected by routine prenatal ultrasound examination. Large lesions typically cause symptoms by compromising alveolar growth during development leading to pulmonary hypoplasia. Mediastinal shift with subsequent development of polyhydramnios and hydrops may develop with rapidly enlarging lesions [21]. Expectant management is appropriate for the nonhydropic fetus, whereas survival of fetuses with hydrops may be dependent on fetal intervention, including thoracoamniotic shunt placement, percutaneous laser ablation, and rarely open fetal surgery [22]. Infants who are symptomatic postnatally are treated with surgical resection, which generally consists of lobectomy or segmental resection. Surgical intervention for a child with an asymptomatic CPAM is controversial. Recurrent infection and a small risk of neoplastic transformation have been cited as reasons for elective resection [19, 23–26]. Reported malignancies are primarily mucinous adenocarcinomas thought to arise from mucigenic epithelium which is frequently present in large cyst CPAM [27, 28]. KRAS mutations have been detected in several cases, similar to precursor adenocarcinoma lesions in the adult lung [29, 30].

Pulmonary Sequestrations

Pulmonary sequestrations are masses of nonfunctioning lung that do not connect to the normal tracheobronchial tree and have a systemic arterial blood supply, usually arising from the thoracic or abdominal aorta [31]. Occasionally, the systemic arterial supply originates from other arteries including the celiac, splenic, intercostal, subclavian, and even coronary arteries [32]. Venous drainage may occur through either a pulmonary or systemic vein. Pulmonary sequestrations are classified as intralobar sequestrations (ILS) when they occur within the visceral pleural lining of the adjacent surrounding lung, and extralobar sequestrations (ELS) when they are separate from the lung and have their own pleural lining. Sequestrations are the second most common lung lesion detected antenatally after CPAM.

ELS comprise the majority of sequestrations and are often identified in utero as a large echo-dense mass that may become smaller relative to the normal lung with advancing gestational age. ELS commonly present with respiratory distress shortly after birth. Antenatal presentation may include polyhydramnios and fetal hydrops. There is a frequent association of ELS with other congenital anomalies, especially diaphragmatic hernia, cardiac abnormalities, and other foregut duplications including bronchogenic cysts [33]. A connection to the gastrointestinal tract may be present (Figure 12.4a). Similar to bronchogenic cysts, ELS are believed to arise from a foregut-derived supernumerary lung bud, but contain distal lung parenchyma [4, 32]. While most are intrathoracic, they can also be found below the diaphragm. On gross

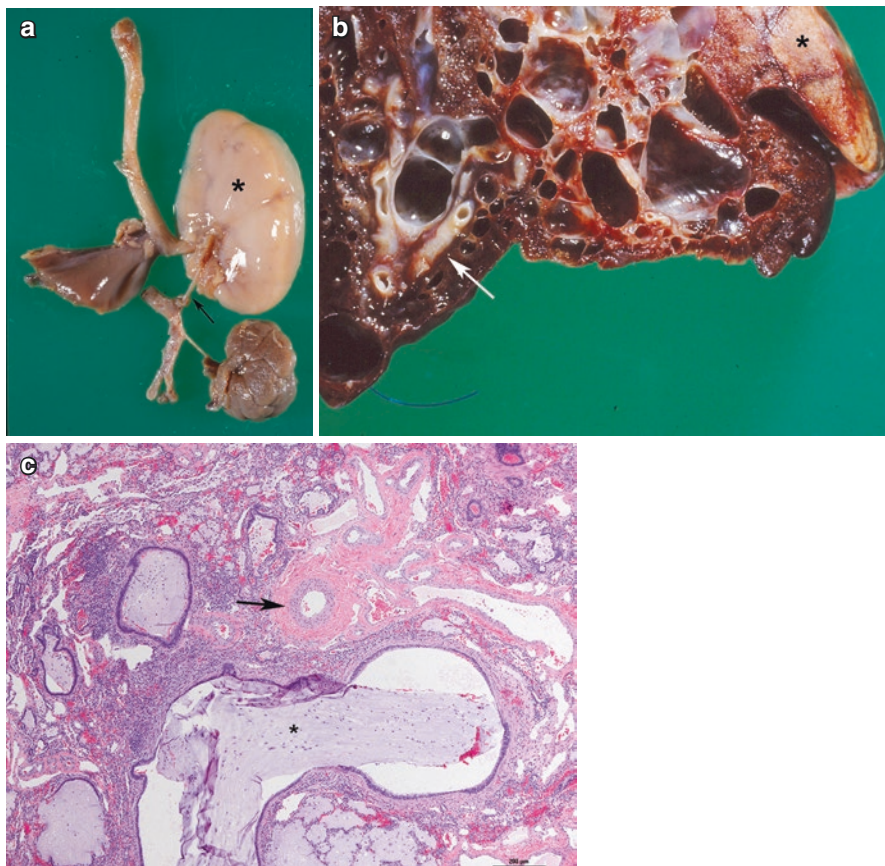


Fig. 12.4 Pulmonary sequestrations. (a) A large extralobar sequestration (ELS) (*) seen below the diaphragm with connection to the distal esophagus (stomach on left) and vascular supply (*arrow*) from the descending aorta; (right kidney lower right corner). (b) Intralobar sequestrations (ILS) are usually well-demarcated from the adjacent normal lung (*) and have thick-walled systemic arteries (*arrow*). They often resemble a congenital pulmonary airway malformation (CPAM) with cystic spaces. (c) Evidence of proximal airway obstruction is common in both ELS and ILS with dilated airways filled with mucus (*), identical to that seen in bronchial atresia. Arteries with medial thickening reflect systemic blood supply (*arrow*). Depending upon the age of presentation and presence of concurrent infection, inflammation and dense fibrosis may be present (H&E)

examination, ELS resemble an accessory lung lobe with its own pleura and a vascular pedicle containing a systemic artery and a draining vein. Pulmonary sequestrations are composed of lung parenchyma with enlarged and poorly subdivided airspaces, and containing abnormally large thick-walled vessels, reflective of the systemic vascular supply (Figure 12.4b). Notably, bronchial atresia with small cyst CPAM features is extremely common in both ELS and ILS [5, 34]. Depending on the presence of proximal airway obstruction, there may be associated mucus accumulation (Figure 12.4c).

ILS is defined by an isolated nonfunctioning lung segment contained within normal lung and with systemic arterial supply (see Figure 12.4b). Most ILS are located

in the lower lobes. If not detected by routine prenatal ultrasound, they present in childhood or adolescence with cough and recurrent pulmonary infections; in some cases, the lesion is detected as an incidental finding on chest radiograph [35]. Radiographically, ILS typically appears as a dense mass within the pulmonary parenchyma; infection can lead to cystic change with fluid levels.

Historically, there has been debate whether ILS is a congenital or acquired lesion. The late age of presentation and history of recurrent infection support ILS being an acquired lesion. In contrast, the frequent presence of bronchial atresia with CPAM features in ILS supports it being part of a spectrum of congenital lung malformations with or without systemic arterial supply [36].

Due to overlapping pathological features, ILS can be difficult to distinguish from a CPAM or isolated bronchial atresia in the absence of knowledge of an associated systemic arterial supply (Figure 12.4b,c). When infected, ILS may be confused with chronic pneumonia, although in chronic pneumonia there is a normal bronchial connection to the involved lung.

Pulmonary Hyperplasia and Congenital Lobar Overinflation

Pulmonary hyperplasia (also termed polyalveolar lobe) is the result of airway obstruction during development, which blocks outflow of fetal fluid leading to increased alveolar growth. Pulmonary hyperplasia has also been referred to as type 3 CPAM in the Stocker classification [37]. Upper airway obstruction, such as tracheal or laryngeal atresia, affects all lung lobes (also termed congenital high airway obstruction syndrome, or CHAOS) (Figure 12.5a,b), while bronchial atresia and stenosis result in focal hyperplasia (Figure 12.5c).

The pathogenesis of congenital lobar overinflation (CLO) is similar to pulmonary hyperplasia, in that intrinsic or extrinsic compression of an airway results in progressive lobar over expansion, albeit due to alveolar distension by air rather than increased alveolar growth as seen in pulmonary hyperplasia (Figure 12.5d). The underlying cause can be secondary to an intrinsic cartilaginous abnormality with resultant weak or absent bronchial cartilage or extrinsic compression of an airway (e.g., bronchogenic cyst or an enlarged or anomalous pulmonary artery). Intraluminal obstruction from meconium or mucus plugs is another potential etiology. In CLO, the compressed airway acts as a one-way valve, resulting in air trapping postnatally. While CLO may be referred to as congenital lobar emphysema, the term “emphysema” is inaccurate as alveolar wall destruction does not accompany the alveolar expansion.

Pulmonary hyperplasia may manifest in utero with polyhydramnios or fetal hydrops, from compression of venous return to the heart. In both pulmonary hyperplasia and CLO, newborns usually present with respiratory distress [38]. Tension pneumothorax may occur. Radiographical imaging demonstrates progressive hyperinflation or hyperlucency of a lobe. Marked overinflation may lead to compression of the adjacent lung and mediastinal shift, often necessitating lobectomy. In both

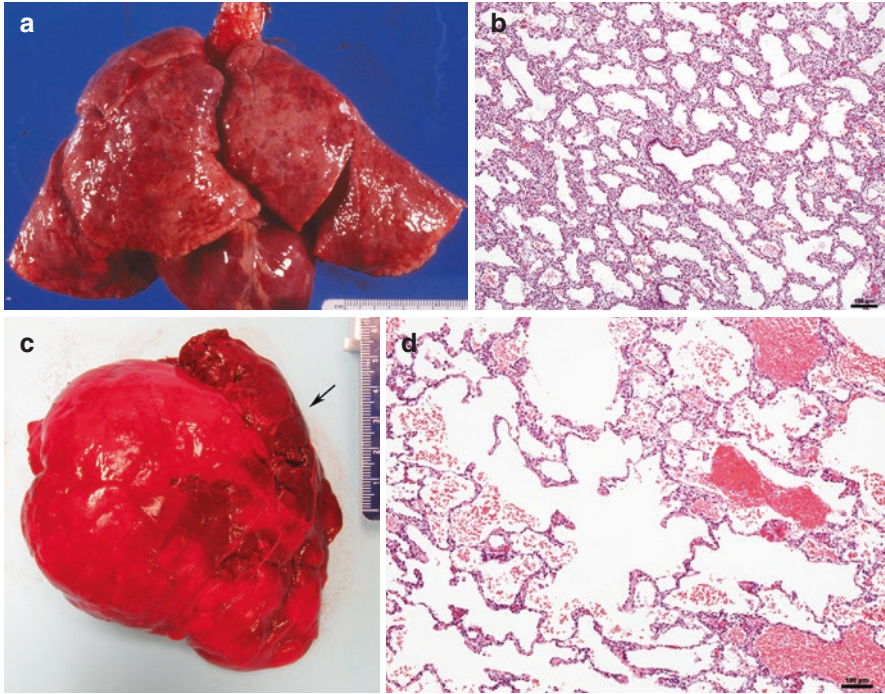


Fig. 12.5 Pulmonary hyperplasia and congenital lobar overinflation (CLO). (a) All lung lobes in an infant with congenital laryngeal stenosis are markedly enlarged (heart lower center). (b) Histologically the lungs have a striking increase in alveolar spaces compared to airways, that are often structurally abnormal (H&E). (c) Hyperinflation of the right middle lobe in a newborn with severe bronchial compression from a large aortopulmonary window (*arrow* denotes compressed right upper lobe). (d) CLO is characterized microscopically by diffuse dilatation of alveoli rather than an increase in alveolar spaces as seen in pulmonary hyperplasia (H&E)

pulmonary hyperplasia and CLO, the involved lung lobes are enlarged, bulky, and sometimes pale (see Figure 12.5a and c).

Developmental Diffuse Cystic Lung Disease

Alveolar Growth Abnormalities Presenting with Cystic Lung Disease

Alveolar growth abnormalities are characterized by impaired alveolarization with deficient alveolar septation and airspace enlargement that may be misinterpreted as “emphysematous change” on radiographical imaging and pathological evaluation [39]. While impaired lung growth is traditionally considered to occur in the context of prenatal onset pulmonary hypoplasia and chronic neonatal lung disease of

Table 12.2 Alveolar growth abnormalities presenting as diffuse cystic lung disease

I. Pulmonary hypoplasia (restriction of prenatal lung growth)
Oligohydramnios (e.g., prolonged rupture of membranes, renal dysfunction)
Restriction of thoracic volume (e.g., diaphragmatic hernia, pleural effusions, thoracic deformity from skeletal dysplasia)
Central nervous system and neuromuscular disorders resulting in decreased fetal breathing
II. Chronic neonatal lung disease of prematurity (clinical bronchopulmonary dysplasia)
III. Congenital heart disease
Cardiac anomalies with reduced pulmonary blood flow (e.g., tetralogy of Fallot, pulmonary artery stenosis/atresia, tricuspid atresia)
Cyanotic heart disease impairing postnatal alveolarization
IV. Chromosome disorders
Trisomy 21 with deficient postnatal alveolarization (often manifests with subpleural cysts on imaging)
Other chromosomal defects
V. Monogenetic disorders
<i>FLNA</i> (Filamin A; high phenotypic diversity including brain, cardiovascular, and gastrointestinal anomalies. Radiographical imaging may mimic congenital lobar overinflation)
<i>NKX2.1</i> (NKX2.1; associated with brain-thyroid-lung syndrome)
<i>DICER1</i> (DICER1; tumor predisposition syndrome with diverse manifestations most commonly including pleuropulmonary blastoma, cystic nephroma, ovarian Sertoli-Leydig cell tumors, and thyroid nodular hyperplasia/carcinoma. Radiographical imaging may mimic cystic pulmonary airway malformation)

prematurity (clinical bronchopulmonary dysplasia), it also occurs in infants with congenital heart disease as well as those with chromosomal disorders (Table 12.2) [40].

The majority of cases with pulmonary hypoplasia are secondary to congenital anomalies or pregnancy complications that inhibit lung development. As distension of the lung with liquid and fetal respiratory movements is required for prenatal lung growth, any mechanism that interferes with these processes can result in pulmonary hypoplasia. The severity of the lung growth abnormality depends on the mechanism of hypoplasia and the timing of the insult. Conditions that occur early in pregnancy (renal anomalies) may interfere with airway branching as well as alveolar formation, while later events (premature rupture of membranes) will impact only alveolar formation.

As lung maturation continues after birth, with most alveolarization occurring within the first 2 years of life [41], postnatal events can impact lung growth as well. Postnatal growth abnormalities predominate in infants with chronic neonatal lung disease (CNLD) related to prematurity, which is increasing as the survival of premature infants improves. Radiographical findings are variable based on etiology, age of the infant, and severity of the growth abnormality. Prior treatments involving high concentrations of oxygen and mechanical ventilation resulted in coarse reticular opacities, cystic lucencies, and distorted lung aeration reflecting alternating regions of

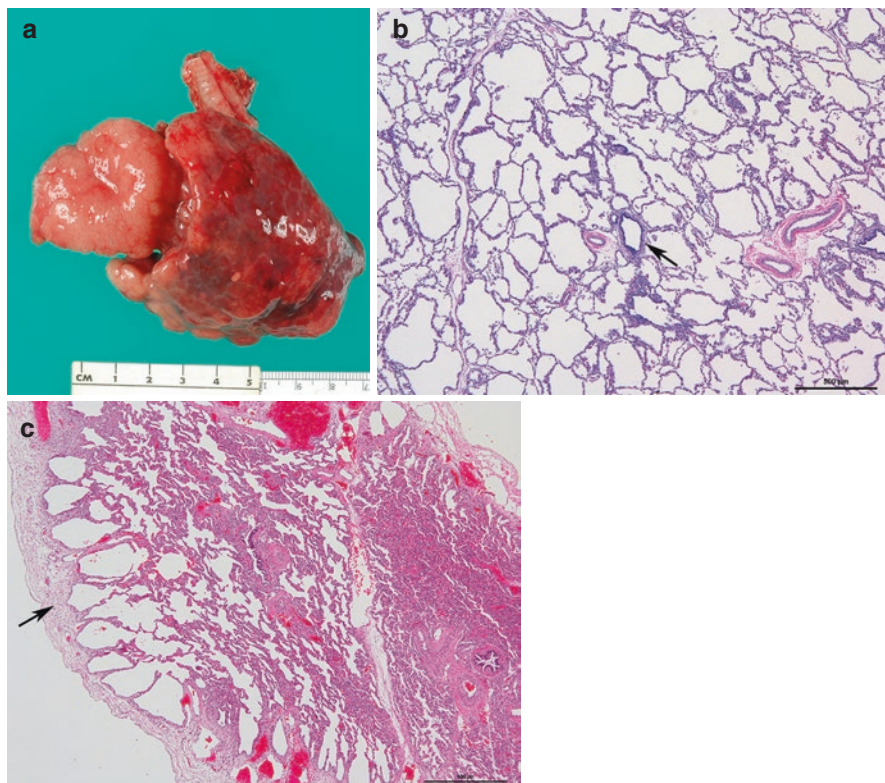


Fig. 12.6 Deficient lung growth manifesting as cystic lung disease. **(a)** Alternating areas of hyper-expansion and collapse are prominent in the left lung of an 11-week infant with severe pulmonary hypoplasia from bladder outlet obstruction. **(b)** Histologic assessment shows marked enlargement and simplification of alveoli, which normally are smaller in size than bronchioles (*arrow*) (H&E). **(c)** Deficient alveolarization is often most prominent in the subpleural space (*arrow*), as seen in the lungs from an infant with trisomy 21 (H&E)

interstitial fibrosis and atelectasis with hyperinflation on histology [42] (Figure 12.6a). Current neonatal practices including surfactant replacement and refined ventilatory strategies demonstrate more subtle radiographic imaging abnormalities, the most frequent being hyperlucent areas with linear and triangular opacities [43]. The hyperlucent areas correspond to enlarged alveoli and reduced distal vascularization (Figure 12.6b). In general, there is significant variability in the radiographical imaging findings associated with alveolar growth abnormalities, but subpleural cysts along the lung periphery, pulmonary fissures, and bronchovascular bundles can be quite characteristic, particularly in deficient lung growth associated with trisomy 21 [40, 44]. This finding correlates well with the subpleural accentuation of cystically dilated alveoli frequently seen on lung biopsy [45] (Figure 12.6c). The subpleural space is theorized to be the predominant site for late alveolar formation [46, 47].

Monogenetic Disorders Presenting with Diffuse Cystic Lung Disease

Heterozygous loss-of-function mutations in *FLNA*, which encodes the actin cross-linking protein filamin A, cause an X-linked dominant disorder with variable multi-organ involvement, including periventricular nodular heterotopias in the brain, cardiovascular anomalies, Ehlers-Danlos syndrome-like features, intestinal pseudo-obstruction, and pulmonary disease of varying severity [48–50]. Respiratory symptoms are often progressive and accompanied by severe clinical pulmonary hypertension. Age of onset of pulmonary disease associated with *FLNA* mutations occurs over a wide range but frequently presents in the first few months of life [50, 51]. Severe hyperinflation with hyperlucent cystic areas alternating with areas of atelectasis and thickening of the interlobular septa is a common appearance reported on computed tomography (CT) scan [50, 51]. The disorder may also present with severe lobar “emphysema” mimicking the localized lung malformation congenital lobar overinflation. Histologic assessment of lung tissue reveals marked alveolar enlargement and simplification reflective of a lung growth abnormality commonly combined with hypertensive changes of the pulmonary arteries, as well as abnormal muscularization of pulmonary veins and lymphatics [51, 52] (Figure 12.7a). The pathology can be similar to that of other lung disorders with deficient lung growth with pulmonary arteriopathy, which may delay diagnosis, especially in a premature infant, unless clinically suspected [52].

Diffuse cystic lung disease reflective of alveolar simplification can be the primary manifestation of pulmonary disease in individuals with heterozygous mutations in the *NKX2-1* gene (thyroid transcription factor 1, TTF-1). Cystically dilated airspaces may be accompanied by other pathologic findings including alveolar proteinosis, nonspecific interstitial pneumonitis, and fibrous remodeling [53–55]

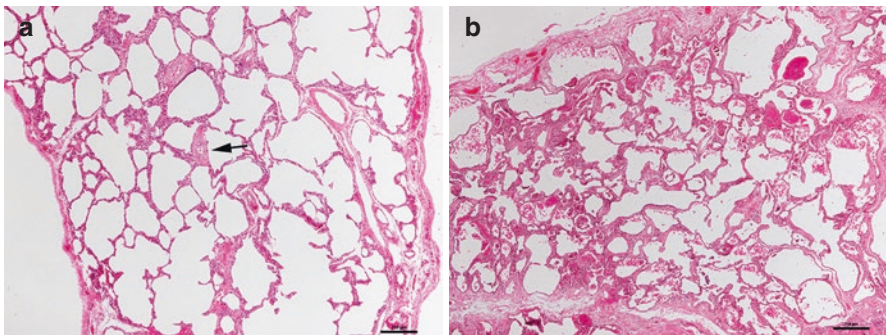


Fig. 12.7 Monogenetic disorders presenting with diffuse cystic lung disease. (a) A term infant with mild hypoplasia of the aortic arch and early onset respiratory distress was found to have a mutation in Filamin-A (*FLNA*) after lung biopsy (H&E). Medial hypertrophy of the pulmonary arteries (*arrow*) accompanies diffuse airspace enlargement. (b) Cystic dilatation of alveoli in a young child with an inherited *NKX2.1* mutation (H&E)

(Figure 12.7b). *NKX2-1* is a homeobox protein critical for development and function of the lung, brain, and thyroid [56, 57]. Haploinsufficiency of *NKX2-1* due to either gene deletions or loss-of-function mutations results in “Brain-Thyroid-Lung” syndrome with individuals having a variable degree of pulmonary disease, thyroid dysfunction, and neurologic abnormalities [53, 58]. The onset of presentation is often in the newborn period or early childhood with respiratory distress syndrome progressing to interstitial lung disease and frequent pulmonary infections [53]. Recurrent spontaneous pneumothorax has also been reported [53, 54]. CT imaging demonstrates a range of findings from mild to diffuse ground-glass opacities, cysts, infiltrates, and fibrosis. Patient prognosis is variable and dependent upon the extent of lung disease.

The *DICER1* tumor predisposition syndrome commonly presents as cystic lung disease. The *DICER1* syndrome is an autosomal dominant condition caused by mutations in the *DICER1* gene. *DICER1* is required for generation of mature microRNAs (miRNAs) that have essential regulatory roles in multiple processes including development and oncogenesis [59]. Pleuropulmonary blastoma (PPB) is the hallmark manifestation of the *DICER1* syndrome and one of the most important causes of *DICER1* associated morbidity and mortality [60]. Multiple other benign and malignant neoplasms are seen in the *DICER1* syndrome including cystic nephroma, sarcoma and Wilms tumor of the kidney, Sertoli-Leydig cell tumor of the ovary, adenomas and carcinomas of the thyroid gland, embryonal rhabdomyosarcoma, neuroblastoma, pituitary blastoma, and pineoblastoma [60, 61]. Heterozygous germline *DICER1* loss-of-function mutations were first identified in PPB in 2009 and have subsequently been identified in >70% of PPB patients [62, 63]. A somatic mutation is acquired in the second allele in *DICER1* associated benign and malignant lesions, characteristically occurring in one of five hotspot codons encoding the RNase IIIb domain resulting in a dysfunctional protein [59, 61]. Genetic studies in mice demonstrate that *DICER1* loss in the lung epithelium during a particular time frame during embryonic development recapitulates the PPB phenotype, and fibroblastic growth factor *FGF9* overexpression in the *DICER1* deficient lung epithelium promotes proliferation of the immature mesenchymal cells [64, 65]. Together, mouse models, the natural history of PPB, and tumor sequencing support *DICER1* loss during lung development initiating PPB pathogenesis with tumor progression to sarcoma resulting from accumulation of additional oncogenic mutations [66, 67].

PPB manifests as three tumor subtypes representing clinicopathologic progression from a cystic (type I PPB) to a combined cystic and solid malignancy (type II PPB) to a completely solid sarcoma (type III PPB) [68]. PPB primarily affects children with >95% of cases diagnosed by 7 years of age [69]. Median ages of diagnosis are 8, 35, and 41 months for type I, type II, and type III PPBs, respectively, supporting tumor progression over time. Progression from type I to types II and III PPB are well documented in individual patients; however, not all type I PPBs progress to the more malignant subtypes [69]. Lesions with the same multilocular cystic architecture as type I PPB but lacking the primitive mesenchymal cell component that progresses to malignant sarcoma that defines PPB types II and III are identified in children as well as adults with pathologic germline *DICER1* mutations [69, 70].

These lesions are presumed to be type I PPBs that have regressed and are designated type Ir PPB. Pulmonary cysts identified in adults with pathologic germline DICER1 variants are not typically resected for oncologic indications but can result in pneumothorax or symptoms related to superinfection [60]. Importantly, there are case reports of pediatric patients with type Ir PPB progressing to type II or III PPB as late as 53 months after diagnosis indicating the need for follow-up, especially when type Ir PPB is diagnosed in a young child [69].

PPB presents with variable clinical symptoms from respiratory distress to non-productive cough, fever, and chest pain with pneumothorax being a frequent presentation of cystic type I PPB [68, 69]. A primary consideration in the radiological differential diagnosis of cystic PPB is CPAM with intrapulmonary bronchogenic cyst, pneumatocele, and pleuropulmonary synovial sarcoma representing additional considerations [61]. Radiographical features favoring a diagnosis of CPAM include prenatal detection, presence of a systemic feeding vessel, asymptomatic presentation, and hyperinflated lung [71]. These distinguishing features form the basis for an algorithm to identify children at higher risk of PPB to aid in determining cystic lesions that require resection. Two critically important points need to be highlighted regarding the differential diagnosis of cystic PPB and CPAM. First, cystic PPB and CPAM can be clinically and radiographically indistinguishable [71, 72]. Thus, frequent careful follow-up of cystic lesions in children is recommended with development of high-risk features or growth prompting consideration for resection. Second, type I PPB and type 4 CPAM in the Stocker classification system have indistinguishable histologic and radiologic features leading to the conclusion that type I PPB and type 4 CPAM represent the same entity [73–76]. It is therefore critically important that type I PPB not be diagnosed as CPAM to ensure that patients receive appropriate postoperative care and follow-up for malignancy.

Distinguishing pathological features of cystic type I PPB are the multilocular architecture and the frequent well-circumscribed border with adjacent normal lung parenchyma (Figures 12.8a,b) [70]. Key diagnostic histopathological features are collections of small primitive mesenchymal cells within the cyst walls and the cuboidal to flattened epithelial cells resembling normal alveolar epithelial cells lining the septa that separate the cysts (Figures 12.8c,d). The abundance of immature mesenchymal cells varies greatly among cases consisting of a uniform cellular cambium layer underlying the surface epithelium (see Figures 12.8c,d) to very focal, small cellular aggregates requiring examination of multiple tissue sections for identification. Lesions with the same multilocular cystic architecture but lacking a primitive cell component are designated as regressed or nonprogressed PPB type Ir. PPB progression is characterized by expansion of the primitive mesenchymal cells to form a cystic malignancy with a solid sarcomatous component (type II PPB) or an entirely solid sarcoma (type III PPB). The PPB subtype is the strongest predictor of outcome with 5-year overall survival rates of 91%, 71%, and 53% for type I, II, and III PPB, respectively [69]. These outcome data highlight the need to detect and treat PPB in its earliest, most curable, cystic type I stage. To this end, testing to identify at-risk individuals and recommended surveillance strategies have been developed [60].

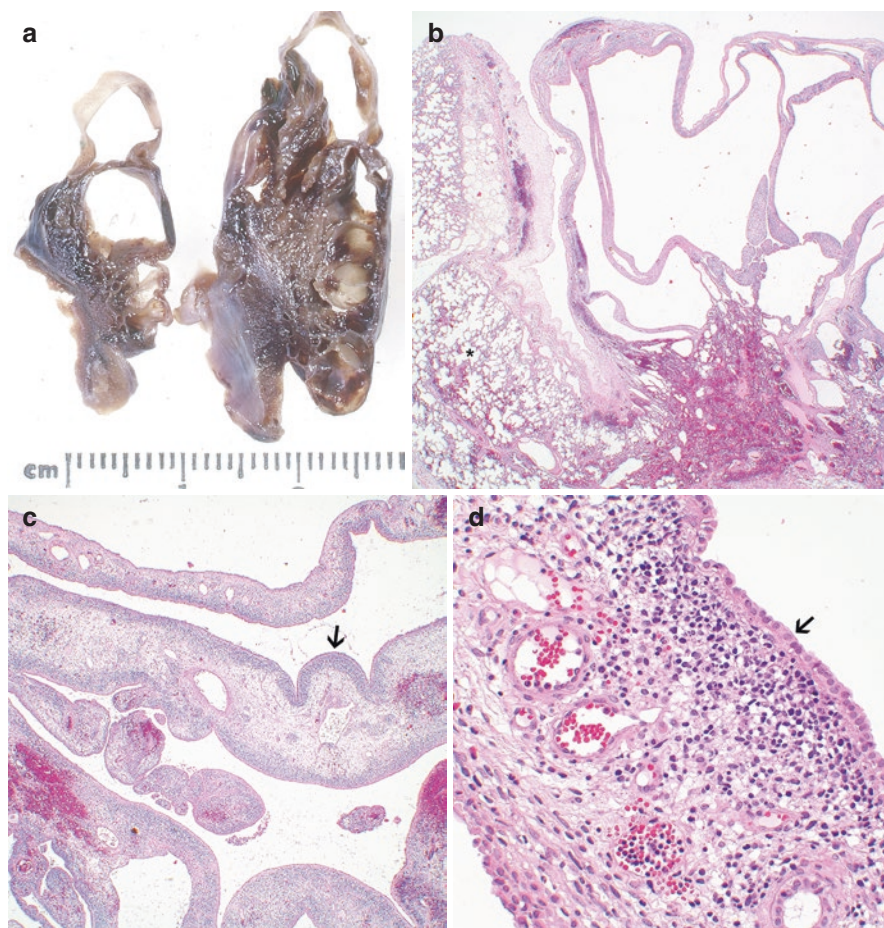


Fig. 12.8 Type I pleuropulmonary blastoma (PPB). (a) Lung tissue resected from a 2-year-old infant presenting with spontaneous pneumothorax containing a multiloculated cystic lesion well demarcated from the adjacent normal lung. (b) Microscopic examination confirms the multiloculated architecture and a well-circumscribed border with the adjacent normal lung (*) (H&E, 20 \times). (c) The cyst walls contain a cambium layer of mesenchymal cells underlying the surface epithelium (*arrow*) (H&E, 40 \times). (d) The cambium layer is comprised of small immature mesenchymal cells with the overlying epithelium (*arrow*) comprised of cuboidal cells resembling normal alveolar type 2 epithelial cells (H&E, 400 \times)

Conclusion

Diverse congenital and developmental entities present as cystic lung disease including lung malformations, growth abnormalities, and neoplasms. The widespread use of antenatal ultrasound has resulted in congenital lung malformations being increasingly detected in utero providing new challenges in clinical decision-making as well

as opportunities to observe lesion evolution over time. Identification of underlying mechanisms of disease, including airway obstruction and genetic alterations, has provided insights into the pathogenesis of multiple congenital and developmental lesions. A multidisciplinary approach is critical to arriving at a specific diagnosis, with integration of clinical, radiological, pathological, and frequently molecular testing required to optimize patient management.

Key Learning Points

- Airway obstruction is the etiology underlying many congenital cystic lung malformations.
- Revised pathology and radiology-based classifications have been developed for congenital cystic lung malformations to encompass pathogenesis and clinical behavior.
- Deficient lung growth can present as diffuse cystic lung disease.
- Genetic conditions can manifest as cystic lung disease.

References

1. Puligandla PS, Laberge JM. Congenital lung lesions. *Clin Perinatol*. 2012;39(2):331–47.
2. Stanton M, Njere I, Ade-Ajayi N, Patel S, Davenport M. Systematic review and meta-analysis of the postnatal management of congenital cystic lung lesions. *J Pediatr Surg*. 2009;44(5):1027–33.
3. Kunisaki SM, Ehrenberg-Buchner S, Dillman JR, Smith EA, Mychaliska GB, Treadwell MC. Vanishing fetal lung malformations: prenatal sonographic characteristics and postnatal outcomes. *J Pediatr Surg*. 2015;50(6):978–82.
4. Langston C. New concepts in the pathology of congenital lung malformations. *Semin Pediatr Surg*. 2003;12(1):17–37.
5. Riedlinger WF, Vargas SO, Jennings RW, Estroff JA, Barnewolt CE, Lillehei CW, et al. Bronchial atresia is common to extralobar sequestration, intralobar sequestration, congenital cystic adenomatoid malformation, and lobar emphysema. *Pediatr Dev Pathol*. 2006;9(5):361–73.
6. Kunisaki SM, Fauza DO, Nemes LP, Barnewolt CE, Estroff JA, Kozakewich HP, et al. Bronchial atresia: the hidden pathology within a spectrum of prenatally diagnosed lung masses. *J Pediatr Surg*. 2006;41(1):61–5. discussion-5
7. Imai Y, Mark EJ. Cystic adenomatoid change is common to various forms of cystic lung diseases of children: a clinicopathologic analysis of 10 cases with emphasis on tracing the bronchial tree. *Arch Pathol Lab Med*. 2002;126(8):934–40.
8. Achiron R, Hegesh J, Yagel S. Fetal lung lesions: a spectrum of disease. New classification based on pathogenesis, two-dimensional and color Doppler ultrasound. *Ultrasound Obstet Gynecol*. 2004;24(2):107–14.
9. Bush A. Congenital lung disease: a plea for clear thinking and clear nomenclature. *Pediatr Pulmonol*. 2001;32(4):328–37.
10. Seear M, Townsend J, Hoepker A, Jamieson D, McFadden D, Daigneault P, et al. A review of congenital lung malformations with a simplified classification system for clinical and research use. *Pediatr Surg Int*. 2017;33(6):657–64.
11. Nobuhara KK, Gorski YC, La Quaglia MP, Shamberger RC. Bronchogenic cysts and esophageal duplications: common origins and treatment. *J Pediatr Surg*. 1997;32(10):1408–13.
12. Zvulunov A, Amichai B, Grunwald MH, Avinoach I, Halevy S. Cutaneous bronchogenic cyst: delineation of a poorly recognized lesion. *Pediatr Dermatol*. 1998;15(4):277–81.

13. Gajewska-Knapik K, Impey L. Congenital lung lesions: prenatal diagnosis and intervention. *Semin Pediatr Surg.* 2015;24(4):156–9.
14. Stocker JT, Madewell JE, Drake RM. Congenital cystic adenomatoid malformation of the lung. Classification and morphologic spectrum. *Hum Pathol.* 1977;8(2):155–71.
15. Szafranski P, Coban-Akdemir ZH, Rupps R, Grazioli S, Wensley D, Jhangiani SN, et al. Phenotypic expansion of TBX4 mutations to include acinar dysplasia of the lungs. *Am J Med Genet A.* 2016;170(9):2440–4.
16. Barnett CP, Nataren NJ, Klingler-Hoffmann M, Schwarz Q, Chong CE, Lee YK, et al. Ectrodactyly and lethal pulmonary acinar dysplasia associated with homozygous FGFR2 mutations identified by exome sequencing. *Hum Mutat.* 2016;37(9):955–63.
17. Biyyam DR, Chapman T, Ferguson MR, Deutsch G, Dighe MK. Congenital lung abnormalities: embryologic features, prenatal diagnosis, and postnatal radiologic-pathologic correlation. *Radiographics.* 2010;30(6):1721–38.
18. Adzick NS, Harrison MR, Glick PL, Golbus MS, Anderson RL, Mahony BS, et al. Fetal cystic adenomatoid malformation: prenatal diagnosis and natural history. *J Pediatr Surg.* 1985;20(5):483–8.
19. Pogoriler J, Swarr D, Kreiger P, Adzick NS, Peranteau W. Congenital cystic lung lesions: redefining the natural distribution of subtypes and assessing the risk of malignancy. *Am J Surg Pathol.* 2019;43(1):47–55.
20. Kreiger PA, Ruchelli ED, Mahboubi S, Hedrick H, Scott Adzick N, Russo PA. Fetal pulmonary malformations: defining histopathology. *Am J Surg Pathol.* 2006;30(5):643–9.
21. Crombleholme TM, Coleman B, Hedrick H, Liechty K, Howell L, Flake AW, et al. Cystic adenomatoid malformation volume ratio predicts outcome in prenatally diagnosed cystic adenomatoid malformation of the lung. *J Pediatr Surg.* 2002;37(3):331–8.
22. Vu L, Tsao K, Lee H, Nobuhara K, Farmer D, Harrison M, et al. Characteristics of congenital cystic adenomatoid malformations associated with nonimmune hydrops and outcome. *J Pediatr Surg.* 2007;42(8):1351–6.
23. Ioachimescu OC, Mehta AC. From cystic pulmonary airway malformation, to bronchioloalveolar carcinoma and adenocarcinoma of the lung. *Eur Respir J.* 2005;26(6):1181–7.
24. Granata C, Gambini C, Balducci T, Toma P, Michelazzi A, Conte M, et al. Bronchioloalveolar carcinoma arising in congenital cystic adenomatoid malformation in a child: a case report and review on malignancies originating in congenital cystic adenomatoid malformation. *Pediatr Pulmonol.* 1998;25(1):62–6.
25. West D, Nicholson AG, Colquhoun I, Pollock J. Bronchioloalveolar carcinoma in congenital cystic adenomatoid malformation of lung. *Ann Thorac Surg.* 2007;83(2):687–9.
26. Abdallah HI, Karmazin N, Marks LA. Late presentation of misalignment of lung vessels with alveolar capillary dysplasia. *Crit Care Med.* 1993;21(4):628–30.
27. Lantuejoul S, Nicholson AG, Sartori G, Piolat C, Danel C, Brabencova E, et al. Mucinous cells in type 1 pulmonary congenital cystic adenomatoid malformation as mucinous bronchioloalveolar carcinoma precursors. *Am J Surg Pathol.* 2007;31(6):961–9.
28. Stacher E, Ullmann R, Halbwdel I, Gogg-Kammerer M, Boccon-Gibod L, Nicholson AG, et al. Atypical goblet cell hyperplasia in congenital cystic adenomatoid malformation as a possible preneoplasia for pulmonary adenocarcinoma in childhood: a genetic analysis. *Hum Pathol.* 2004;35(5):565–70.
29. Kim MY, Kang CH, Park SH. Multifocal synchronous mucinous adenocarcinomas arising in congenital pulmonary airway malformation: a case report with molecular study. *Histopathology.* 2014;65(6):926–32.
30. Sakamoto H, Shimizu J, Horio Y, Ueda R, Takahashi T, Mitsudomi T, et al. Disproportionate representation of KRAS gene mutation in atypical adenomatous hyperplasia, but even distribution of EGFR gene mutation from preinvasive to invasive adenocarcinomas. *J Pathol.* 2007;212(3):287–94.
31. Landing BH, Dixon LG. Congenital malformations and genetic disorders of the respiratory tract (larynx, trachea, bronchi, and lungs). *Am Rev Respir Dis.* 1979;120(1):151–85.

32. Newman B. Congenital bronchopulmonary foregut malformations: concepts and controversies. *Pediatr Radiol.* 2006;36(8):773–91.
33. Stocker JT, Kagan-Hallet K. Extralobar pulmonary sequestration: analysis of 15 cases. *Am J Clin Pathol.* 1979;72(6):917–25.
34. Conran RM, Stocker JT. Extralobar sequestration with frequently associated congenital cystic adenomatoid malformation, type 2: report of 50 cases. *Pediatr Dev Pathol.* 1999;2(5):454–63.
35. DeParedes CG, Pierce WS, Johnson DG, Waldhausen JA. Pulmonary sequestration in infants and children: a 20-year experience and review of the literature. *J Pediatr Surg.* 1970;5(2):136–47.
36. Holder PD, Langston C. Intralobar pulmonary sequestration (a nonentity?). *Pediatr Pulmonol.* 1986;2(3):147–53.
37. Stocker JT, Drake RM, Madewell JE. Cystic and congenital lung disease in the newborn. *Perspect Pediatr Pathol.* 1978;4:93–154.
38. Stigers KB, Woodring JH, Kanga JF. The clinical and imaging spectrum of findings in patients with congenital lobar emphysema. *Pediatr Pulmonol.* 1992;14(3):160–70.
39. Guillerman RP. Imaging of childhood interstitial lung disease. *Pediatr Allergy Immunol Pulmonol.* 2010;23(1):43–68.
40. Deutsch GH, Young LR, Deterding RR, Fan LL, Dell SD, Bean JA, et al. Diffuse lung disease in young children: application of a novel classification scheme. *Am J Respir Crit Care Med.* 2007;176(11):1120–8.
41. Cooney TP, Thurlbeck WM. The radial alveolar count method of Emery and Mithal: a reappraisal 2 – intrauterine and early postnatal lung growth. *Thorax.* 1982;37(8):580–3.
42. Northway WH Jr, Rosan RC, Porter DY. Pulmonary disease following respirator therapy of hyaline-membrane disease. Bronchopulmonary dysplasia *N Engl J Med.* 1967;276(7):357–68.
43. Agrons GA, Courtney SE, Stocker JT, Markowitz RI. From the archives of the AFIP: lung disease in premature neonates: radiologic-pathologic correlation. *Radiographics.* 2005;25(4):1047–73.
44. Biko DM, Schwartz M, Anupindi SA, Altes TA. Subpleural lung cysts in down syndrome: prevalence and association with coexisting diagnoses. *Pediatr Radiol.* 2008;38(3):280–4.
45. Cooney TP, Thurlbeck WM. Pulmonary hypoplasia in Down's syndrome. *N Engl J Med.* 1982;307(19):1170–3.
46. Massaro GD, Massaro D. Postnatal lung growth: evidence that the gas-exchange region grows fastest at the periphery. *Am J Phys.* 1993;265(4 Pt 1):L319–22.
47. Narayanan M, Owers-Bradley J, Beardsmore CS, Mada M, Ball I, Garipov R, et al. Alveolarization continues during childhood and adolescence: new evidence from helium-3 magnetic resonance. *Am J Respir Crit Care Med.* 2012;185(2):186–91.
48. Robertson SP, Twigg SR, Sutherland-Smith AJ, Biancalana V, Gorlin RJ, Horn D, et al. Localized mutations in the gene encoding the cytoskeletal protein filamin a cause diverse malformations in humans. *Nat Genet.* 2003;33(4):487–91.
49. Sole G, Coupury I, Rooryck C, Guerineau E, Martins F, Deves S, et al. Bilateral periventricular nodular heterotopia in France: frequency of mutations in FLNA, phenotypic heterogeneity and spectrum of mutations. *J Neurol Neurosur Ps.* 2009;80(12):1394–8.
50. Shelmerdine SC, Semple T, Wallis C, Aurora P, Moledina S, Ashworth MT, et al. Filamin A (FLNA) mutation—a newcomer to the childhood interstitial lung disease (ChILD) classification. *Pediatr Pulmonol.* 2017;52(10):1306–15.
51. Burrage LC, Guillerman RP, Das S, Singh S, Schady DA, Morris SA, et al. Lung transplantation for FLNA-associated progressive lung disease. *J Pediatr.* 2017;186:118–23. e6
52. Lord A, Shapiro AJ, Saint-Martin C, Claveau M, Melancon S, Wintermark P. Filamin A mutation may be associated with diffuse lung disease mimicking bronchopulmonary dysplasia in premature newborns. *Respir Care.* 2014;59(11):e171–7.
53. Hamvas A, Deterding RR, Wert SE, White FV, Dishop MK, Alfano DN, et al. Heterogeneous pulmonary phenotypes associated with mutations in the thyroid transcription factor gene NKX2-1. *Chest.*

54. Thorwarth A, Schnittert-Hubener S, Schruppf P, Muller I, Jyrch S, Dame C, et al. Comprehensive genotyping and clinical characterisation reveal 27 novel NKX2-1 mutations and expand the phenotypic spectrum. *J Med Genet.* 2014;51(6):375–87.
55. Galambos C, Levy H, Cannon CL, Vargas SO, Reid LM, Cleveland R, et al. Pulmonary pathology in thyroid transcription factor-1 deficiency syndrome. *Am J Respir Crit Care Med.* 2010;182(4):549–54.
56. Kimura S, Hara Y, Pineau T, Fernandez-Salguero P, Fox CH, Ward JM, et al. The T/ebp null mouse: thyroid-specific enhancer-binding protein is essential for the organogenesis of the thyroid, lung, ventral forebrain, and pituitary. *Genes Dev.* 1996;10(1):60–9.
57. Guillot L, Carre A, Szinnai G, Castanet M, Tron E, Jaubert F, et al. NKX2-1 mutations leading to surfactant protein promoter dysregulation cause interstitial lung disease in “brain-lung-thyroid syndrome”. *Hum Mutat.* 2010;31(2):E1146–62.
58. Krude H, Schutz B, Biebermann H, von Moers A, Schnabel D, Neitzel H, et al. Choreoathetosis, hypothyroidism, and pulmonary alterations due to human NKX2-1 haploinsufficiency. *J Clin Invest.* 2002;109(4):475–80.
59. Foulkes WD, Priest JR, Duchaine TF. DICER1: mutations, microRNAs and mechanisms. *Nat Rev Cancer.* 2014;14(10):662–72.
60. Schultz KAP, Williams GM, Kamihara J, Stewart DR, Harris AK, Bauer AJ, et al. DICER1 and associated conditions: identification of at-risk individuals and recommended surveillance strategies. *Clin Cancer Res.* 2018;24(10):2251–61.
61. Guillerman RP, Foulkes WD, Priest JR. Imaging of DICER1 syndrome. *Pediatr Radiol.* 2019;49(11):1488–505.
62. Brenneman M, Field A, Yang J, Williams G, Doros L, Rossi C, et al. Temporal order of RNase IIIb and loss-of-function mutations during development determines phenotype in pleuropulmonary blastoma / DICER1 syndrome: a unique variant of the two-hit tumor suppression model. *F1000Res.* 2015;4:214.
63. Hill DA, Ivanovich J, Priest JR, Gurnett CA, Dehner LP, Desruisseau D, et al. DICER1 mutations in familial pleuropulmonary blastoma. *Science.* 2009;325(5943):965.
64. Wagh PK, Gardner MA, Ma X, Callahan M, Shannon JM, Wert SE, et al. Cell- and developmental stage-specific Dicer1 ablation in the lung epithelium models cystic pleuropulmonary blastoma. *J Pathol.* 2015;236(1):41–52.
65. Yin Y, Castro AM, Hoekstra M, Yan TJ, Kanakamedala AC, Dehner LP, et al. Fibroblast growth factor 9 regulation by MicroRNAs controls lung development and links DICER1 loss to the pathogenesis of Pleuropulmonary Blastoma. *PLoS Genet.* 2015;11(5):e1005242.
66. Pugh TJ, Yu W, Yang J, Field AL, Ambrogio L, Carter SL, et al. Exome sequencing of pleuropulmonary blastoma reveals frequent biallelic loss of TP53 and two hits in DICER1 resulting in retention of 5p-derived miRNA hairpin loop sequences. *Oncogene.* 2014;33(45):5295–302.
67. Seki M, Yoshida K, Shiraishi Y, Shimamura T, Sato Y, Nishimura R, et al. Biallelic DICER1 mutations in sporadic pleuropulmonary blastoma. *Cancer Res.* 2014;74(10):2742–9.
68. Priest JR, McDermott MB, Bhatia S, Watterson J, Manivel JC, Dehner LP. Pleuropulmonary blastoma: a clinicopathologic study of 50 cases. *Cancer.* 1997;80(1):147–61.
69. Messinger YH, Stewart DR, Priest JR, Williams GM, Harris AK, Schultz KA, et al. Pleuropulmonary blastoma: a report on 350 central pathology-confirmed pleuropulmonary blastoma cases by the international Pleuropulmonary Blastoma registry. *Cancer.* 2015;121(2):276–85.
70. Hill DA, Jarzembowski JA, Priest JR, Williams G, Schoettler P, Dehner LP. Type I pleuropulmonary blastoma: pathology and biology study of 51 cases from the international pleuropulmonary blastoma registry. *Am J Surg Pathol.* 2008;32(2):282–95.
71. Feinberg A, Hall NJ, Williams GM, Schultz KA, Miniati D, Hill DA, et al. Can congenital pulmonary airway malformation be distinguished from type I pleuropulmonary blastoma based on clinical and radiological features? *J Pediatr Surg.* 2016;51(1):33–7.
72. Oliveira C, Himidan S, Pastor AC, Nasr A, Manson D, Taylor G, et al. Discriminating preoperative features of pleuropulmonary blastomas (PPB) from congenital cystic adenomatoid malformations (CCAM): a retrospective, age-matched study. *Eur J Pediatr Surg.* 2011;21(1):2–7.

73. Dehner LP, Messinger YH, Williams GM, Stewart DR, Harney LA, Schultz KA, et al. Type I Pleuropulmonary Blastoma versus congenital pulmonary airway malformation type IV. *Neonatology*. 2017;111(1):76.
74. Hill DA, Dehner LP. A cautionary note about congenital cystic adenomatoid malformation (CCAM) type 4. *Am J Surg Pathol*. 2004;28(4):554–5. author reply 5
75. MacSweeney F, Papagiannopoulos K, Goldstraw P, Sheppard MN, Corrin B, Nicholson AG. An assessment of the expanded classification of congenital cystic adenomatoid malformations and their relationship to malignant transformation. *Am J Surg Pathol*. 2003;27(8):1139–46.
76. Stocker JT. Cystic lung disease in infants and children. *Fetal Pediatr Pathol*. 2009;28(4):155–84.