Environmental Chemistry for a Sustainable World 60

Vineet Kumar · Praveen Guleria Shivendu Ranjan · Nandita Dasgupta Eric Lichtfouse *Editors*

Nanosensors for Environment, Food and Agriculture Vol. 1



Environmental Chemistry for a Sustainable World

Volume 60

Series Editors

Eric Lichtfouse , Aix-Marseille University, CNRS, IRD, INRAE, Coll France, CEREGE, Aix-en-Provence, France Jan Schwarzbauer, RWTH Aachen University, Aachen, Germany Didier Robert, CNRS, European Laboratory for Catalysis and Surface Sciences, Saint-Avold, France

Other Publications by the Editors

Books Environmental Chemistry http://www.springer.com/978-3-540-22860-8

Organic Contaminants in Riverine and Groundwater Systems http://www.springer.com/978-3-540-31169-0

Sustainable Agriculture Volume 1: http://www.springer.com/978-90-481-2665-1 Volume 2: http://www.springer.com/978-94-007-0393-3

Book series

Environmental Chemistry for a Sustainable World http://www.springer.com/series/11480

Sustainable Agriculture Reviews http://www.springer.com/series/8380

Journal

Environmental Chemistry Letters http://www.springer.com/10311

More information about this series at http://www.springer.com/series/11480

Vineet Kumar • Praveen Guleria Shivendu Ranjan • Nandita Dasgupta Eric Lichtfouse Editors

Nanosensors for Environment, Food and Agriculture Vol. 1



Editors Vineet Kumar Department of Biotechnology Lovely Professional University Jalandhar, Punjab, India

Shivendu Ranjan Faculty of Engineering and the Built Environment University of Johannesburg Johannesburg, South Africa

Eric Lichtfouse D Aix Marseille University, CNRS, IRD, INRAE, Coll France, CEREGE Aix-en-Provence, France Praveen Guleria Plant Biotechnology & Genetic Engineering Lab, Department of Biotechnology DAV University Jalandhar, Punjab, India

Nandita Dasgupta Department of Biotechnology Institute of Engineering and Technology Lucknow, Uttar Pradesh, India

 ISSN 2213-7114
 ISSN 2213-7122
 (electronic)

 Environmental Chemistry for a Sustainable World
 ISBN 978-3-030-63244-1
 ISBN 978-3-030-63245-8
 (eBook)

 https://doi.org/10.1007/978-3-030-63245-8

@ The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Switzerland AG 2021

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Switzerland AG The registered company address is: Gewerbestrasse 11, 6330 Cham, Switzerland

Preface

Food security is a major issue for human health in the context of climate change, global pollution, and rising population. In particular, the identification of contaminated food should improve food quality, yet classical analytical techniques are limited. This challenge has been solved by the recent development of nanosensors for food and the environment. Nanosensors enable ultrasensitive detection of multiple contaminants. Nanosensors are very specific because they are designed to bind selectively with some contaminants. Here, functional or biological recognition of the contaminant triggers an electric signal, which is converted into a readable computer graphic. This book gathers recent advances in the development of nanosensors for food, agriculture, and the environment. Nanosensors for the detection of heavy metals are reviewed in Chaps. 1, 2, and 3 by Bereli et al., He et al., and Xuyan et al., with focus on electrochemical, piezoelectric, optical, and colorimetric sensors and fluorescence sensors based on aggregationinduced emission (AIE). Biofilm-based nanosensors for monitoring pollutants such as toxins, hormones, and dioxins are presented by Nikoleli et al. in Chap. 4. Chapter 5 by Teodoro et al. reviews chemical sensors, including e-tongues (Fig. 1), and discuss the use of green composites, for example, cellulose and proteins.

Toxicity, risk assessment, and bioaccumulation of nanosensors are discussed in Chap. 6 by Heikal and Abdel-Aziz. Chap. 7 by Abdel-Aziz and Heikal reviews nanosensors and nanobiosensors in agriculture, with focus on the detection of bacteria, mycotoxins, pesticides, and drugs. Chapter 8 by Cimen et al. elaborates different sensing strategies for the detection of food contaminants. Chapter 9 by Manubolu focuses on sensing and removing mycotoxins from agricultural products and food samples. In Chap. 10, Karahalil discusses the use of nanomaterials for the detection of contaminants in food. Nanosensors based on quantum dots, carbon, dendrimers, liposomes, metal oxides, and dendrimers are reviewed in Chap. 11 by Sing and



Fig. 1 The experimental setup of a typical e-tongue: (i) interdigitated electrodes (IDEs) modified with distinct materials, (ii) measurement cell containing the sensor array immersed in the liquid under investigation, (iii) data analysis of trial solutions collected using the e-tongue generates a large amount of data, which can be classified and distinguished by statistical analysis.

Sing. In Chap. 12,, Nikoleli et al. review biosensor with emphasis on size reduction of lipid film devices, new sensing techniques, and transduction.

Jalandhar, Punjab, India	Vineet Kumar
Jalandhar, Punjab, India	Praveen Guleria
Johannesburg, South Africa	Shivendu Ranjan
Lucknow, Uttar Pradesh, India	Nandita Dasgupta
Aix-en-Provence, France	Eric Lichtfouse

Contents

1	Sensors for the Detection of Heavy Metal Contaminants in Water and Environment. Nilay Bereli, Duygu Çimen, Handan Yavuz, and Adil Denizli	1
2	Nanosensors for Heavy Metal Detection in EnvironmentalMedia: Recent Advances and Future TrendsXiaojia He, Hua Deng, and Huey-min Hwang	23
3	AIE-Based Fluorescent Nanosensors for Detection of Heavy Metal Ions Xueyan Huang, Bin Feng, Tang Gao, Anyao Bi, Liu Huang, Rong Song, Meihui Liu, and Wenbin Zeng	53
4	Nanosensors Based on Lipid Films for Environmental Applications . Georgia-Paraskevi Nikoleli, Christina G. Siontorou, Marianna-Thalia Nikolelis, Spyridoula Bratakou, and Vasillios N. Psychoyios	97
5	Novel Chemical Sensors Based on Green Composite Materials for Environmental Analysis K. B. R. Teodoro, F. L. Migliorini, M. H. M. Facure, R. C. Sanfelice, D. Martins, and D. S. Correa	109
6	Toxicology and Safety Aspects of Nanosensor on Environment, Food, and Agriculture Yasmin M. Heikal and Heba M. M. Abdel-Aziz	139
7	Nanosensors for the Detection of Fertilizers and OtherAgricultural ApplicationsHeba M. M. Abdel-Aziz and Yasmin M. Heikal	157
8	Sensors for the Detection of Food Contaminants Duygu Çimen, Nilay Bereli, Handan Yavuz, and Adil Denizli	169

9	Nanotechnology-Based Detection and Remediation of Mycotoxins for Food and Agriculture Application Manjunath Manubolu, Lavanya Goodla, Stuart A. Ludsin, Thanasekaran Jayakumar, Mike Fraker, and Kavitha Pathakoti	183
10	Detection of Toxic Contaminant in the Food Items Bensu Karahalil	213
11	Utility of Nanobiosensors in Environmental Analysis and Monitoring . Kshitij RB Singh and Ravindra Pratap Singh	229
12	Nanosensors Based on Lipid Membranes for the Rapid Detection of Food Toxicants	247
Ind	ex	261

About the Editors



Vineet Kumar is currently working as assistant professor of biotechnology in the School of Bioengineering and Biosciences, LPU, Jalandhar, Punjab, India. His areas of interest include natural plant product–based green synthesis of nanoparticles, nanotoxicity testing of nanoparticles, and application of nanoparticles in drug delivery, food technology, sensing, dye degradation, and catalysis. Vineet has published many articles in these areas featuring in peer-reviewed reputed journals. He is also serving as editorial board member and reviewer for international peer-reviewed journals. Vineet has received various awards such as senior research fellowship, best poster award, and postdoctoral fellowship.



Praveen Guleria is presently working as assistant professor in the Department of Biotechnology at DAV University, Jalandhar, Punjab, India. She has worked in the areas of plant biotechnology, plant metabolic engineering, and plant stress biology at CSIR -Institute of Himalayan Bioresource Technology, Palampur, H.P., India. Her research interests include plant stress biology, plant small RNA biology, plant epigenomics, and nanotoxicity. She has published several research articles in various peer-reviewed journals. Praveen is also serving as the editorial board member and reviewer for certain international peerreviewed journals. She has been awarded the SERB-Start Up Grant by DST, GOI. Praveen has also been awarded the prestigious "Bharat Gaurav Award" in 2016 by the India International Friendship Society, New Delhi. She has also received various awards like CSIR/ ICMR – junior research fellowship, CSIR – senior research fellowship, and state-level merit scholarship awards.

Nandita Dasgupta has vast working experience in nanoscience and is working as assistant professor in the Department of Biotechnology, Institute of Engineering and Technology, Lucknow, India. She has worked on mesenchymal stem cell–derived exosomes for the treatment of uveitis. Dr. Dasgupta has successfully engineered micro-vehicles for model drug molecules. Her areas of interest include nanomaterial fabrication and its applications in medicine, food, environment, agriculture, and biomedicine. She is the associate editor of *Environmental Chemistry Letters*.

Shivendu Ranjan is scientist at the Chaperon Biotech, SIDBI Centre, Indian Institute of Technology, Kanpur, India. He is also serving as a senior research associate, adjunct, in the Faculty of Engineering and Built Environment, University of Johannesburg, South Africa. His research interests include nanotechnology, nanomedicine, science policy, and diplomacy. He is associate editor of *Environmental Chemistry Letters*.

Eric Lichtfouse is an environmental chemist at the University of Aix-Marseille, France, and visiting professor at Xi'an Jiaotong University, China. He has invented carbon-13 dating. Dr. Lichtfouse is teaching scientific writing and communication, and has published the book *Scientific Writing for Impact Factors*. He is founder and chief editor of scientific journals and series on environmental chemistry and agriculture. He received the Analytical Chemistry Prize from the French Chemical Society, the Grand Prize of the Universities of Nancy and Metz, and a Journal Citation Award by the Essential Indicators.







Contributors

Heba M. M. Abdel-Aziz Botany Department, Faculty of Science, Mansoura University, Mansoura, Egypt

Department of Chemistry, Hacettepe University, Ankara, Turkey

Nilay Bereli Department of Chemistry, Hacettepe University, Ankara, Turkey

Anyao Bi Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

Spyridoula Bratakou Laboratory of Environmental & Sanitary Engineering, Department of Public Health, Faculty of Health and Caring Professions, University of West Attica, Athens, Attica, Greece

Duygu Çimen Department of Chemistry, Hacettepe University, Ankara, Turkey

D. S. Correa Nanotechnology National Laboratory for Agriculture, Embrapa Instrumentação, São Carlos, SP, Brazil

PPGQ, Department of Chemistry, Center for Exact Sciences and Technology, Federal University of São Carlos (UFSCar), São Carlos, SP, Brazil

Hua Deng Department of Chemistry, Morgan State University, Baltimore, MD, USA

Adil Denizli Department of Chemistry, Hacettepe University, Ankara, Turkey

M. H. M. Facure Nanotechnology National Laboratory for Agriculture, Embrapa Instrumentação, São Carlos, SP, Brazil

PPGQ, Department of Chemistry, Center for Exact Sciences and Technology, Federal University of São Carlos (UFSCar), São Carlos, SP, Brazil

Bin Feng Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

Mike Fraker Cooperative Institute for Great Lakes Research, University of Michigan, Ann Arbor, MI, USA

Tang Gao Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

Lavanya Goodla Department of Biochemistry and Molecular Biology, University of New Mexico, Albuquerque, NM, USA

Yasmin M. Heikal Botany Department, Faculty of Science, Mansoura University, Mansoura, Egypt

Xiaojia He Department of Marine Sciences, The University of Georgia, Athens, GA, Greece

Present Affiliation: Department of Medicine, Emory University School of Medicine, Atlanta, GA, USA

Liu Huang Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

Xueyan Huang Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

Huey-min Hwang Department of Biology, Jackson State University, Jackson, MS, USA

Thanasekaran Jayakumar Graduate Institute of Medical Sciences, College of Medicine, Taipei Medical University, Taipei, Taiwan

Bensu Karahalil Faculty of Pharmacy, Toxicology Department, Gazi University, Ankara, Turkey

Stephanos Karapetis Laboratory of Inorganic & Analytical Chemistry, School of Chemical Engineering, Dept 1, Chemical Sciences, National Technical University of Athens, Athens, Greece

Meihui Liu Hunan Huacheng Biotech Inc. and Jiangxi Haifu Biotech Inc., Changsha, PR China

Stuart A. Ludsin Aquatic Ecology Laboratory, Department of Evolution, Ecology and Organismal Biology, The Ohio State University, Columbus, OH, USA

Manjunath Manubolu Aquatic Ecology Laboratory, Department of Evolution, Ecology and Organismal Biology, The Ohio State University, Columbus, OH, USA

D. Martins Nanotechnology National Laboratory for Agriculture, Embrapa Instrumentação, São Carlos, SP, Brazil

F. L. Migliorini Nanotechnology National Laboratory for Agriculture, Embrapa Instrumentação, São Carlos, SP, Brazil

Georgia-Paraskevi Nikoleli Laboratory of Inorganic & Analytical Chemistry, School of Chemical Engineering, Dept 1, Chemical Sciences, National Technical University of Athens, Athens, Greece

Dimitrios P. Nikolelis Laboratory of Environmental Chemistry, Department of Chemistry, University of Athens, Athens, Greece

Marianna-Thalia Nikolelis Laboratory of Simulation of Industrial Processes, Department of Industrial Management and Technology, School of Maritime and Industry, University of Piraeus, Piraeus, Greece

Kavitha Pathakoti Interdisciplinary Center for Nanotoxicity, Department of Chemistry, Physics and Atmospheric Sciences, Jackson State University, Jackson, MS, USA

Department of Biology, Jackson State University, Jackson, MS, USA

Vasillios N. Psychoyios Laboratory of Simulation of Industrial Processes, Department of Industrial Management and Technology, School of Maritime and Industry, University of Piraeus, Piraeus, Greece

R. C. Sanfelice Department of Chemical Engineering, Institute of Technological and Exact Sciences – ICTE, Universidade Federal do Triangulo Mineiro – UFTM, Uberaba, MG, Brazil

Kshitij RB Singh Department of Biotechnology, Indira Gandhi National Tribal University, Amarkantak, Madhya Pradesh, India

Ravindra Pratap Singh Department of Biotechnology, Indira Gandhi National Tribal University, Amarkant, Madhya Pradesh, India

Christina G. Siontorou Laboratory of Simulation of Industrial Processes, Department of Industrial Management and Technology, School of Maritime and Industry, University of Piraeus, Piraeus, Greece

Rong Song Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

K. B. R. Teodoro Nanotechnology National Laboratory for Agriculture, Embrapa Instrumentação, São Carlos, SP, Brazil

Handan Yavuz Department of Chemistry, Hacettepe University, Ankara, Turkey

Wenbin Zeng Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

Chapter 1 Sensors for the Detection of Heavy Metal Contaminants in Water and Environment



Nilay Bereli, Duygu Çimen, Handan Yavuz, and Adil Denizli

Contents

1.1	Introdu	uction	2
1.2 Principle of Sensors.			3
	1.2.1	Electrochemical Sensors.	4
	1.2.2	Piezoelectric Sensors.	4
	1.2.3	Optical Sensors.	5
	1.2.4	Colorimetric Sensors.	6
1.3	Applic	ation of Sensors for Heavy Metal Detection in Environment	6
	1.3.1	Electrochemical Sensing for Heavy Metals.	6
	1.3.2	Piezoelectric Sensing for Heavy Metals.	7
	1.3.3	Optic Sensing for Heavy Metals.	10
	1.3.4	Colorimetric Sensing for Heavy Metals.	11
1.4	Conclu	isions	15
Refe	rences.		16

Abstract Environmental pollution by heavy metals is a global problem. The main sources of waste are industrial or domestic products and heavy metal–containing pesticides and fertilizers. Heavy metal contamination poses a serious danger to the human health and environment. Heavy metals are among the important environmental pollutants because of their toxic effects on plants, animals, and humans, even at very low concentrations. Moreover, they have non-biodegradable property and accumulation in the food chain. Currently available methods for determination of metal ion levels in water include chromatographic and spectroscopic methods. These chromatographic and spectroscopic methods are costly methods that require intense technical training because of their time-consuming and complicated proce-

N. Bereli · D. Çimen · H. Yavuz · A. Denizli (🖂)

Department of Chemistry, Hacettepe University, Ankara, Turkey e-mail: denizli@hacettepe.edu.tr

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_1

dures. The development of sensors for monitoring of toxic metals especially in water and environment is very important. The nano and functional material-based chemical sensors have been extensively employed for the detection of heavy metals. These sensors have advantages such as high selectivity, sensitivity, portability, onsite sensing capability, and improved performance of the devices. This chapter will outline the broad contours of heavy metals and the practical applications for their detection, with sensors, in the water and environment.

Keywords Heavy metals \cdot Sensors \cdot Sensitive determination \cdot Monitoring \cdot Waste water \cdot Environment \cdot Optical sensors \cdot Electrochemical sensors \cdot Colorimetric sensors \cdot Piezoelectric sensors

1.1 Introduction

The growing number of pollutants requires the development of sensitive, rapid, precise, specific, and easy-to-use innovative analytical devices to meet the increasing demand for the environmental pollution control and early warning. Quantitative analysis of samples is usually carried out by conventional analytical methods such as spectroscopic and chromatographic techniques to detect different environmental contaminants. These methods are accurate and sensitive, but they require expensive instrumentation and also expert staff to operate them with multistep sample preparation. It is difficult to achieve on-site, real-time, and high-frequency monitoring of contaminants by these labor-intensive and time-consuming methods (Bellan et al. 2011). The potential for environmental pollution requires the development of rapid and simple detection methods for organic, inorganic, and pathogenic pollutants' monitoring. The pollutant recognition is vitally important in some emergent situations. Therefore, novel devices with superior sensing capabilities and increased spatial localization are needed urgently. Researchers have been striving for the development of cost-effective, robust, and automated watermonitoring devices for sensitive and rapid analysis of environmental pollutants.

Heavy metal ions are released to the environment through industrial activities or by end-user products. In the environment, they ultimately come in contact with water and accordingly they contaminate the water, including ground water, lakes, oceans, and rivers. The water contaminated by heavy metal ions (such as lead, mercury, and arsenic ions) can cause acute and chronic health problems in humans and seriously affect aquatic life (Kim et al. 2012; Lin et al. 2011; Song et al. 2016; Weng et al. 2014). Therefore, it is extremely important to monitor the toxic metal ion levels in drinking water as well as other water sources to improve public health and protect aquatic life (Chen et al. 2013). Currently available methods for determination of metal ion levels in water include inductively coupled plasma mass spectrometry (ICP-MS), high-performance liquid chromatography (HPLC), and atomic absorption spectrometry—AAS (Kim et al. 2001; Udhayakumari et al. 2014; Bontidean et al. 1998; Erdem et al. 2018). While these chromatographic and spectroscopic methods offer high sensitivity and selectivity, these costly instruments require intense technical training because of their time-consuming and complicated procedures. Electrochemical or membrane-based methods have been also developed for detection of metal ions in water (Pesavento et al. 2009; Van Leeuwen et al. 2005). However, these methods often suffer from their poor selectivity.

Sensors that combine chemistry, biology, physics, nanotechnology, and electronics can follow developments in risk management approaches and environmental legislation because of their original characteristics such as specificity, sensitivity, real-time remote monitoring capability, and ease of use (Rogers 2006). Sensors provide fast, highly accurate, real-time, and high-frequency monitoring without any time-consuming sample enrichment, and sample pre-processing steps are effective alternatives to traditional analytical methods (Thevenot et al. 2001). Though their use in the field of environmental pollution control is still in the early stages, sensors have great potential applications in the areas of environmental monitoring, drug development, food safety, diagnosis, and biomedical research (Borisov and Wolfbeis 2008; Yılmaz et al. 2017a, b). Unfortunately, dispersed molecules that are diffusing freely into the solution are far from having sensitive surfaces in practical applications. In this regard, different types of sensors are promising devices, in that they combine their unique features such as high sensitivity, specificity, label-free and quantitative detection, inexpensiveness, and miniaturization.

1.2 Principle of Sensors

A sensor is a compact analytical device that consists of three parts: (i) a sensing receptor, (ii) a transducer, and (iii) a detector with a digital output (Goode et al. 2015). A sensor is a self-contained integrated device that is able to convert chemical signal depending on analyte concentration into a measurable analytical signal (Fig. 1.1). Figure 1.1 shows the basic components of the sensors. As can be concluded from the figure, sensors have unique advantages over conventional methods, like variety in the target structures, simple construct, and diversity in the detection techniques (Verma and Bhardwaj 2015; Altintas and Tothill 2013).

Sensors possess many advantages such as specificity, sensitivity, portability, quick response time, and so on, which are fabricated for various purposes, including heavy metal detection (Verma and Bhardwaj 2015). They are suitable for single analysis, multi analysis, short-term analysis, long-term implantable analysis, as disposable sensors, and also for environmental monitoring.



Fig. 1.1 The basic components of the sensor system

1.2.1 Electrochemical Sensors

In recent years, electrochemical sensors have exhibited several advantages over other analytical methods. Compared with the spectroscopic, chromatographic, and electrophoretic methods, electrochemical methods are more preferable and have attracted increasing attention because of their high sensitivity, low cost, fast response, portability, simple operation, and on-site operation (Kimmel et al. 2012; Yu et al. 2016a, b). Electrochemical sensors have been widely used for sensitive and selective determination of the environmental, clinical, and biotechnical targets. They are based on the control of electrical signal and its correlation to analyte concentration. These sensors represent a typical system for the construction of sensors, which include semi-conductors and screen-printed electrodes. These types of sensors can be classified into three major groups: (i) amperometric, (ii) potentiometric, and (iii) impedimetric transducers. In addition, these sensors have been employed to detect a variety of targets such as bacteria, pesticides, and heavy metal ions (Luo et al. 2013; Wang et al. 2015; Cui et al. 2015).

1.2.2 Piezoelectric Sensors

Mass-sensitive sensor devices work based on measuring the changes in properties of piezoelectric or magnetoelastic material via increased mass of molecules on sensor surface. Mass-sensitive sensors can work at both liquid and gas phases. Mass-sensitive devices can use quartz crystal microbalance (QCM), microcantilever, or magnetoelastic sensors (Nieuwenhuizen and Venema 1991; Li et al. 2010; Raiteri et al. 2001).

Figure 1.2 shows a schematic illustration of piezoelectric effect. In a piezoelectric material, the positive and negative charges are randomly distributed in each part of the surface. When some pressure on the material is applied, positive and negative



Fig. 1.2 Schematic illustration of piezoelectric effect

charges in the molecules are separated from each other. This polarization generates an electrical field and can be used to transform the mechanical energy used in the material deformation into electrical energy.

Piezoelectric biosensors, including the quartz crystal microbalance (QCM), utilize the sensitivity of piezoelectric crystal resonance to perturbations in the surrounding environment (Marx 2003; Cooper and Singleton 2007). The quartz crystal microbalance (QCM) as a piezoelectric sensor is a physical, nanogram-sensitive device that is able to detect variations in the resonance frequency of an electrically driven quartz crystal with changes in thickness or mass per unit area, when adding (loaded quartz) or removing (unloaded quartz) small masses, or when occurs oxide growth/decay, thin film growth/deposition, a biochemical interaction, or a catalysis at the surface of the quartz (Giamblanco et al. 2015; Karaseva et al. 2016).

1.2.3 Optical Sensors

Optical sensors focus on the measurement of a change in the optical characteristics of the transducer surface when target and recognition element form a complex. They have been widely used in fundamental biological studies, health science research, drug discovery, clinical diagnosis, food analysis, and environmental and agriculture monitoring (Shankaran et al. 2007; Saylan et al. 2017b; Yılmaz et al. 2017a, b; Saylan et al. 2017a). This sensor method of transduction has been used in many classes of sensors due to many different types of spectroscopy such as absorption, fluorescence, Raman Surface enhanced Raman spectroscopy (SERS), phosphorescence, refraction, and dispersion spectroscopy (Muslim et al. 2012). Optical-based biosensors are able to provide a label-free, real-time, and parallel detection.

1.2.4 Colorimetric Sensors

With the use of enzyme-linked sensitive thermistors, heat change was measured during enzyme substrate interaction for the first time by Danielson and Mosbach (1979). Colorimetric devices work on the basis of measurement of heat absorbed or evaluated during the chemical reactions between molecules (Mosbach 1991; Xie et al. 2000; Ramanathan and Danielsson 2001). Initially, the colorimetric transduction was employed for enzyme-based biosensor but has subsequently been applied in cells and immunosensors (Ahmad et al. 2010). The colorimetric sensors use thermistors that transform heat generated during a reaction into an electrical signal. The analyte is the substrate for the enzyme (Dinh and Cullum 2000). Recently, colorimetric sensor method has found uses in food industry and environmental monitoring (Kirchner et al. 2012).

1.3 Application of Sensors for Heavy Metal Detection in Environment

Usages of heavy metals have increased with industrialization, and this has caused heavy metal pollution, which is an important problem that needs to be solved. Heavy metals are non-degradable and accumulate in the environment. Arsenic, chromium, lead, copper, nickel, zinc, and cadmium are the most commonly found heavy metals in waste water, and all of these metals cause risk for human health. Heavy metal toxicity can damage the brain, liver, lungs, kidney, and other organs. Long-term exposure causes degeneration of muscular and neurological tissues (Zhao et al. 2012).

Accordingly, there is an expanding need for the development of innovative environmental monitoring and warning system enabling rapid, sensitive, and specific detection, cost-effective and field-portable screening methods. Recent developments in sensors, which allow for real-time, sensitive, and label-free detection, offer a promising alternative to conventional environmental analysis techniques, including spectroscopic, chromatographic, and electrochemical methods. Construction of the recognition element for interacting with the targets is the fundamental feature of sensors, with high affinity and high specificity used as recognition elements (Van Dorst et al. 2010; Wanekaya et al. 2008).

1.3.1 Electrochemical Sensing for Heavy Metals

Rudnitskaya et al. developed an array of potentiometric sensors with chalcogenide glass membranes for heavy metal ion detection. They reported 0.2, 0.4, 0.06, and 30 nmol/L for Cu(II), Pb(II), Cd(II), and Zn(II) in individual buffer solutions,

respectively. They studied the capability of the sensor to detect these ions in combined buffer solutions at the background of 0.5 M NaCl or artificial seawater. They reported 1 nmol/L detection concentration value for copper and zinc and 2 nmol/L for lead and cadmium. They showed suitable detection sensor array for Cu(II) and Zn(II) detection at the concentrations corresponding to the chronic seawater criteria, and Pb(II) and Cd(II) at the acute seawater criteria (Rudnitskaya et al. 2008).

Avuthu et al. prepared an electrochemical sensor, which was screen printed on polyethylene terephthalate. The response of the sensor showed that conventional printing processes are used for heavy metal ion detection. Carbon- and silver-based inks were used for metallization of the working, counter, and reference electrodes. Cyclic voltammetry response of the sensor resulted in reduction peaks at 0.2 and -0.6 eV for the selective detection of Hg²⁺ and Pb²⁺, respectively. An 87% and 9% change in the average peak currents were detected for the 50 µM concentration of Pb²⁺ and Hg²⁺, respectively (Avuthu et al. 2016).

Lee et al. prepared an electrode by using Fe₂O₃ nanoparticle/graphene composite for the sensitive and selective detection of Cd²⁺, Zn²⁺, and Pb²⁺. The linear range of the electrode was 1–100 µg L⁻¹. The detection limits were 0.08 µg L⁻¹ for Cd²⁺, 0.11 µg L⁻¹ for Zn²⁺, and 0.07 µg L⁻¹ for Pb²⁺. Repeatability was found to be 0.92% for Cd²⁺, 1.68% for Zn²⁺, and 1.69% for Pb²⁺ for single sensor with ten measurements and 1.15% for Cd²⁺, 0.89% for Zn²⁺, and 0.91% for Pb²⁺ for five different composite electrodes (Lee et al. 2016).

Shen et al. presented a highly selective and sensitive microfluidic electrochemical sensor for detection of heavy metals. Cd^{2+}/Pb^{2+} detection was achieved by this device using square-wave anodic stripping voltammetry. The detection limits were found to be 1.2 µg/L for Cd^{2+} and 1.8 µg/L for Pb^{2+} (Shen et al. 2017). Hwang et al. developed a modified nanoporous bismuth electrode sensor that measures successfully Cd^{2+} and Pb^{2+} in tap water even after over 40 times of measurements by a separate Bi/Sn electroplating process. The calibration curves showed strong correlations with respect to various concentrations of Cd^{2+} and Pb^{2+} with the limit of detection of 1.3 ppb for Cd^{2+} and 1.5 ppb for Pb^{2+} (Hwang et al. 2019). Liu et al. overviewed on recent advances in nanomaterial-enabled screen-printed sensors based on carbon, metal, and metal oxide nanomaterials. The modification strategies in heavy metal ion sensors and the application of these sensors for the analysis of heavy metal ions in real samples were discussed (Liu et al. 2019). Other studies for detection of heavy metal ions are summarized in Table 1.1.

1.3.2 Piezoelectric Sensing for Heavy Metals

Huang et al. developed a melanin-coated QCM sensor via cross-linking method to determine metal ions and they also examined the binding affinity of metal ions against melanin. The surface morphologies of melanin thin film upon binding to metal ions were characterized by atomic force microscopy (AFM), circular

Methods	Metal ions	Linear range	LOD	References
Voltammetric	Pb ²⁺ , Cu ²⁺ , Cr ²⁺ , As ²⁺	0.0071– 0.0083 M	7.12, 8.12, 7.17, 7.34 × 10 ⁻⁹ M	Vimala and Vedhi (2019)
Graphene oxide-based	Pb ²⁺ , Cu ²⁺ , Hg ²⁺	12.8–200 ppb	0.4, 0.8, 1.2 ppb	Gong et al. (2014)
Bimetallic PtAu alloy Nanoparticles/ electrospun carbon Nanofibers	Cd ²⁺ , Pb ²⁺ , Cu ²⁺	0.50–1.0 μΜ	0.10 μΜ	Zhang et al. (2017)
Conducting polymer based	$\begin{array}{l} Na^{*},K^{*},Mg^{2+},Sr^{2+},\\ Mn^{2+},Al^{3+},As^{5+},Zn^{2+},\\ Fe^{2+},Fe^{3+},Pb^{2+},Cu^{2+},\\ Cr^{6+} \end{array}$	100 ppb– 100 ppm	$2.19 \times 10^{-7} \mathrm{M}$	Dutta and Panda (2018)
Square-wave stripping voltammetry	Cu ²⁺	5–200 µg L ⁻¹	2 μg L ⁻¹	Wang et al. (2018)
Screen-printed electrode	Hg ²⁺	0.5–10 mg L ⁻¹	0.2 mg L ⁻¹	Fernández et al. (2015)
Square-wave voltammetry	Hg ²⁺	1.0×10^{-8} -1.0 × 10 ⁻⁵ M	$5.8 \times 10^{-9} \mathrm{M}$	Lahrich et al. (2016)

 Table 1.1
 Different electrochemical sensor applications for heavy metal detection in literature

dichroism, and infrared spectroscopy. Detection of metal ions including Hg²⁺, Sn²⁺, Li⁺, Zn²⁺, Cu²⁺, Ni²⁺, Ge⁴⁺, Bi³⁺, Co²⁺, Ag⁺, Al³⁺, and Fe³⁺ was carried out. The prepared piezoelectric sensor system showed sensitive detection for Hg²⁺ with 518 \pm 37 Hz/ppm (Huang et al. 2007). Cao et al. developed copolymer-coated sensor that was used for detecting heavy metal ions via the quartz crystal microbalance (QCM) method (Cao et al. 2011). The copolymer was synthesized by free radical copolymerization of MBTVBC as a hydrophobic monomer and VIM as a main monomer. The surface morphologies of the copolymer-coated quartz crystal microbalance were characterized by atomic force microscopy. The experimental results showed that the prepared sensor has high sensitivity, stability, and selectivity for the detection of Cu²⁺ in aqueous solution. The lowest limit of detection was obtained at 0.01 mg/mL Cu²⁺. They also studied repeatability and reliability of the QCM sensor and achieved stable adsorption equilibrium for each Cu²⁺ aqueous solution.

Sartore et al. reported an effective way to obtain surface-modified gold electrodes with high heavy metal ions' complexing capability. In order to achieve this, multifunctional polymers of polyamidoamine (PAA) type was grafted on preaminated gold via covalent bonding. For this purpose, they developed a flow-type quartz crystal microbalance (QCM) sensor. The unmodified and modified gold surfaces were also characterized by X-ray reflectivity (XRR) and total reflection X-ray fluorescence (TXRF). The developed polymer-grafted QCM sensors were managed to adsorb heavy metal ions, such as copper, chrome, cadmium, and lead, from solutions over a wide range, from 0.01 to 1000 ppm concentration selectively (Sartore et al. 2011).

Lou et al. designed a QCM sensor by using thiol-functionalized SBA-15 (SBA-15-SH) mesoporous silica materials, which have various pore sizes and specific surface areas, for the detection of Hg^{2+} ions in water (Lou et al. 2012). The QCM sensor loaded with thiol-functionalized SBA-15 (SBA-15-SH) was prepared via a post-grafting technique. All the materials were characterized by Fourier transform infrared (FT-IR) spectroscopy, small-angle X-ray scattering (SAXS), thermogravimetric analysis (TGA), transmission electron microscopy (TEM), and N₂ adsorption isotherms. The developed QCM sensor was able to detect 1 ppm Hg²⁺ in aqueous solution with the response frequency of 175 Hz and the response time was found to be as short as 1 min. Sun et al. aimed in their study to develop a quartz crystal microbalance (OCM) sensor with the fibrous polystyrene membranes functionalized by polyethyleneimine (PEI) for the determination of heavy metal ions in solution. The specific surface area (SSA) and morphology of the polystyrene membranes were controlled by way of adjusting the weight ratios of the blend solvents of tetrafuran, N,N-dimethylformamide (DMF). It was found that SSA of polystyrene membranes increased as DMF increased in mixture solvents. The experimental results indicated that QCM sensors have excellent selectivity for Cr3+ with a detection limit of 5 ppb. The sensor responses in the 5–200 ppb concentration range were obtained with good linearity. They also examined reproducibility of QCM sensors. More than 60% of the original response was maintained after five assays (Sun et al. 2012; Fig. 1.3).

Hüseynli et al. reported a new technique of quartz crystal microbalance nanosensor that is used for determination of Hg(II) ions in waste water (Hüseynli et al. 2019). For this purpose, they took advantage of the molecular imprinting method.



Fig. 1.3 Schematic diagram of a testing system for metal ions (Sun et al. 2012)

Firstly, a functional monomer and Hg(II) ions were co-complexed and then Hg(II) ions were imprinted into the polymeric film. Prepared QCM sensor for detecting Hg(II) ions was characterized by contact angle, Fourier transform infrared (FT-IR) spectroscopy, ellipsometer, and atomic force microscopy (AFM). In addition to selectivity and kinetic studies, reproducibility and recovery parameters were investigated. The linear working range was found to be $0.25-50 \times 10^{-3} \mu$ M. The limit of detection was determined as $0.21 \times 10^{-3} \mu$ M. The reproducibility of prepared sensor was reported as average %RSD 1.119 for 3 different days.

Sun et al. designed a QCM sensor, which was based on thioglycolic acidmodified CdTe nanoparticles, to determine heavy metal ions in aqueous solution. In this study, it was aimed to increase the detection limit of the sensor using CdTe nanoparticles and thioglycolic acid was the key factor in adsorption. It was also noted that the sensitivity of the prepared sensor was six times that of the quartz crystal microbalance sensor, which was based on the thioglycolic acid only. The limit of detection was obtained as $0.096 \ \mu g \ L^{-1}$ for Pb²⁺, $0.089 \ \mu g \ L^{-1}$ for Cd²⁺, and $0.189 \ \mu g \ L^{-1}$ for Cu²⁺. In order to investigate reproducibility, the relative standard deviations of Pb²⁺, Cd²⁺, and Cu²⁺ were calculated as 4.2%, 3.1%, and 2.7%, respectively (Sun et al. 2019).

1.3.3 Optic Sensing for Heavy Metals

Forzani et al. detected a heavy metal ion by high-resolution differential surface plasmon resonance (SPR) sensor. They reported Cu(II) and Ni(II) detection in the ppt-ppb ranges of concentration. They developed the SPR sensor by coating with peptides NH₂-Gly-Gly-His-COOH and NH₂-(His)6-COOH. Also, they reported Cu(II) in drinking water using this sensor (Forzani et al. 2005).

Verm and Gupta developed an optical fiber SPR sensor for detection of Cd^{2+} , Pb^{2+} , and Hg^{2+} ions in drinking water. They used silver metal and indium tin oxide for fabrication of the SPR probe, which was further modified with the coating of pyrrole and chitosan composites. They reported strong binding for Cd^{2+} ions than the other ions. They reported a highly sensitive detection for the low concentrations (Verm and Gupta 2015).

Sadrolhossein et al. prepared a polypyrrole-chitosan/nickel ferrite (PPy-Chi/NiFe₂O₄) nanoparticle composite layer in order to detect iron, nickel, cobalt, manganese, mercury, aluminum, and lead ions. The thickness of the layers was detected as 6.3-83.1 nm. A gold surface layer of SPR chip was improved with the polymer composite. The detection of diamagnetic ions was conducted with a limitation of roughly 0.5 ppm. The synthesized polymer composite decreased the response time of the sensor. Thus, the prepared layer could detect the heavy metals strongly (Sadrolhossein et al. 2017).

In another study, Ullah et al. (2018) discussed sensor systems for the detection of heavy metal ions in different matrices, especially in water. They reported about properties of nanomaterial-based chemical sensors such as high sensitivity,

portability, and selectivity. They severely engrossed on the recent progress with using various sensing strategies. The reported optical sensors focused and discussed in the review include fluorescent, surface plasmon resonance, and surface enhanced Raman scattering sensors.

Jalilzadeh et al. prepared Zn(II) ion-imprinted nanofilm on the SPR chip surface for detection of Zn(II) ions in aqueous solution and artificial plasma. Designed SPR sensor was applied for selective detection of Zn(II) ions in aqueous solution within the range of 0.5–1.0 mg mL⁻¹. The limit of detection and limit of quantification were calculated as 0.19 and 0.64 ng mL⁻¹, respectively (Jalilzadeh et al. 2019).

Safran et al. (2019) designed molecularly imprinted SPR biosensor for selective and label-free detection of Cu^{+2} in artificial urine and physiological serum samples to investigate the effects of metabolite residues. They synthesized Cu^{+2} imprinted poly(hydroxyethyl methacrylate-*N*-methacryloyl-(*L*)-cysteine methyl ester) nanoparticles, which they applied to the SPR sensor chip surface for the selective determination of Cu^{+2} ions.

1.3.4 Colorimetric Sensing for Heavy Metals

There is an ongoing research that utilizes metal nanoparticles and organic dyes to develop facile, rapid, and low-cost colorimetric assays for heavy metal ion detection (Du et al. 2013; Lin et al. 2011). In particular, gold and silver metal nanoparticles have attracted a great deal of interest due to their size- and shape-dependent optical properties and large absorption coefficients (Sepulveda et al. 2009; Saha et al. 2012). These assays are often based on aggregation of metal nanoparticles upon interaction of their surface functional groups (aptamers, peptides, proteins, small organic molecules, etc.) with metal ions (Lee et al. 2007; Ye and Yin 2008; Lee and Mirkin 2008; Li et al. 2008; Wang et al. 2010; Liu et al. 2008b, 2010, 2012). Aggregation changes the color of nanoparticle solution, which can be easily monitored by using a UV-Vis spectrophotometer or even with an untrained naked eye, without needing any complicated instruments. However, despite the extensive research in especially the last two decades, there is still need for improvements in the assay designs to improve their accuracy, reproducibility, as well as sensitivity and selectivity.

Mercury is widely released to the environment by industrial activities (e.g., gold mining) and combustion of fossil fuels and wastes. The most stable form of mercury in water is the solvated divalent mercuric ion (Hg^{2+}). Accumulation of mercury ions in the body can cause serious nervous system problems such as motor disorders and Minamata disease (Clarkson et al. 2003; Carvalho et al. 2008). Due to the serious effects of aqueous mercury ions on the human health and aquatic life, several colorimetric methods have been reported for its detection. One of the earlier methods for mercury ions was developed by Mirkin's group, where they prepared a method for the colorimetric detection of Hg^{2+} ions based on the thymidine– Hg^{2+} –thymidine bond formation using the melting point of DNA-functionalized gold nanoparticles.

The color change of the solution and the melting temperature of the DNA-gold nanoparticle aggregates are used to determine the concentration of Hg^{2+} ions. They reached a detection limit of 100 nM with good selectivity using this DNA approach (Lee et al. 2007).

Later, Merkoçi's group demonstrated a highly sensitive assay for the detection of Hg²⁺ ions by using gold nanorods. The gold nanorods were functionalized with *N*-alkylaminopyrazole, which can strongly and selectively interact with Hg²⁺ ions, and this interaction induced the formation of gold nanorod aggregates. They reported very low detection limit of 3 ppt for this method. In addition, the assay was very rapid and could be completed in a few minutes. Finally, they tested the selectivity of the assay against different metal ions: Cd²⁺, Zn²⁺, Cu²⁺, Pb²⁺, Hg²⁺, and As³⁺ (Placido et al. 2013).

Sener et al. reported a simple, sensitive colorimetric method, which does not require any surface modification step, for simple detection of mercury ion. The system was based on the aggregation of citrate-capped gold nanoparticles in the presence of Hg^{2+} ions and the positively charged lysine. The proposed mechanism of this system is the spontaneous reduction of Hg^{2+} ions onto the gold nanoparticle surface, which causes nanoparticle aggregation in the presence of lysine. Using this assay, Hg^{2+} ions can be detected with naked eye within a few minutes, and the detection limit of this assay is 2.9 nM. In addition, selectivity of the assay was tested by using several competing metal ions or a mixture of competing metal ions: Zn^{2+} , Ca^{2+} , Al^{3+} , Cu^{2+} , Ni^{2+} , Pb^{2+} , Ag^+ , Na^+Co^{2+} , K^+ , Fe^{3+} , Cd^{2+} , etc. (Sener et al. 2014b).

In an interesting approach, Wei et al. prepared a sensitive and cost-effective smartphone-based Hg^{2+} sensor with the detection limit of 3.5 ppb. They determined Hg^{2+} ion concentration using gold nanoparticles and aptamer interaction in disposable test tubes by using green and red LEDs at 523 and 625 nm to analyze the color of the solution. In addition, they employed this portable sensor to test over 50 water samples in California, USA (Wei et al. 2014).

Another excessively studied heavy metal ion is lead (Pb²⁺), which is released to the environment by end-user products such as gasoline, batteries, and dyes. Lead ions can get some serious health effects including cardiovascular, developmental, and neurological disorders, especially in children (Kim et al. 2012). Accordingly, several colorimetric assays were developed to detect aqueous lead ions. For instance, Chen et al. developed a label-free colorimetric method for the detection of Pb²⁺ ions in aqueous solution based on gold nanoparticles. This sulfate (S_2O^{2-3}) was used to form a complex with gold nanoparticles. In the presence of Pb2+ ions and 2-mercaptoethanol (2-ME), Pb-Au alloy was formed on the surface of gold nanoparticles, which resulted in the aggregation of nanoparticles. The detection limit of this selective method is 0.5 nM. They also utilized this assay to detect Pb2+ ions in real samples such as soil and river water (Chen et al. 2009). Yu et al. reported a sensitive colorimetric sensor based on glutathione-functionalized gold nanoparticles for the detection of Pb²⁺ leaking from perovskite solar cells in the rain water. They detected Pb²⁺ by naked eye with the detection limit of 15 nM. The results of this selective and sensitive method were also compared with ICP-MS (Yu et al. 2016a, b). On the other hand, Zhu et al. utilized bifunctionalized gold nanoparticles for colorimetric detection of Pb²⁺ ions in aqueous solution and living cells (HeLa cells). Glutathione and pentapeptide (CALNN) were chosen as a functional group for Pb²⁺ sensing and as a stabilizing ligand for gold nanoparticles under physiological conditions, respectively. They found that the Pb²⁺ sensor was selective over different metal cations (Na⁺, K⁺, Ca²⁺, Ba²⁺, Zn²⁺, Mg²⁺, Cu²⁺, Fe²⁺, Co²⁺, Ni²⁺, Cd²⁺ Mn²⁺, Hg²⁺Al³⁺, and Fe³⁺) in living cells (Zhu et al. 2012). Zheng et al. prepared a selective colorimetric method to detect and separate Pb²⁺ ions by using diethanolamine-modified gold nanoparticles (DEA-AuNPs) and *L*-(–)-malic acid-modified Fe₃O₄ nanoparticles (MA-NPs). They tested their sensor in different environmental sources such as drinking, river, and lake water. In the presence of Pb²⁺ ions, the color of the solution changed from red to blue after the interaction with modified gold nanoparticles in 10 min. Then with the addition of MA-Fe₃O₄ they successfully separated Pb²⁺-Au aggregates and MA-Fe₃O₄ with a magnet in 15 min (Zheng et al. 2010).

Another very toxic heavy metal ion is arsenic, which can cause some several diseases, including bloody diarrhea, acute renal failure, cardiovascular diseases, and lung, liver, bladder, and skin cancer (Song et al. 2016; Weng et al. 2014; Domínguez-González et al. 2014). Wu et al. reported a sensitive and selective colorimetric method for the detection of As^{3+} ions in aqueous solutions by using polydiallyldimethylammonium (PDDA), aptamer, and gold nanoparticles. They chose a cationic polymer PDDA, which could aggregate gold nanoparticles and hybridize the aptamer through electrostatic interaction. In the presence of As^{3+} ions, the aptamer bound to As^{3+} to form an As^{3+} -aptamer complex. Addition of cationic polymer aggregated gold nanoparticles that resulted in color change from red to blue. In the absence of As^{3+} ions, aptamers hybridized with cationic polymer so gold nanoparticles were stable in the solution and the color of the solution did not change. The limit of detection of this selective method is 5.3 ppb (Wu et al. 2012b).

In another work, Kalluri et al. employed glutathione, dithiothreitol, and cysteine (Cys) modified gold nanoparticles for detection of arsenic in ground water. Interestingly, the authors observed an excellent selectivity against arsenic in the presence of these three ligands; when they modified the gold surface only with glutathione and dithiothreitol, they reported weak affinity with some other metal ions tested. More importantly, utilization of dynamic light scattering instead of common UV-Vis adsorption measurements improved the sensitivity of the assay about two orders of magnitude. Accordingly, they obtained a very good sensitivity of 3 ppt, which is almost three orders of magnitude lower than the WHO limit. Finally, they tested the assay with real samples using drinking water from different origin and found high levels of arsenic in the tap water sample from Bangladesh and bottled from Jackson (Kalluri et al. 2009).

Later, inspired by nature (i.e., odor and taste differentiation mechanism of mammalians), several sensing units with broad sensitivity to several metal ions have been employed with data-recognition methods. These assays allow selective detection of multiple ions using a single assay (Fig. 1.4). For instance, Şener et al. developed a colorimetric sensor array using mercaptoundecanoic acid (MUA)capped gold nanoparticles to discriminate seven metal ions (Al³⁺, Cu²⁺, Hg²⁺, Cd²⁺,



Fig. 1.4 Colorimetric response of the colorimetric array against Hg^{2+} , Cd^{2+} , and Fe^{3+} ions (20 M) and their binary and ternary mixtures: (**a**) representative photograph; (**b**) hierarchical cluster analysis (Sener et al. 2014a)

Fe³⁺, Cr³⁺, and Pb²⁺) simultaneously with high selectivity. Amino acids (lysine, cysteine, histidine, tyrosine, and arginine) were employed to create several sensing units. Amino acids can bind to MUA-capped gold nanoparticles through their amino groups and form complexes with metal ions through their carboxyl and amino groups. The complex interactions between metal ions, amino acids, and MUA-capped gold nanoparticles resulted in enhanced or diminished nanoparticle aggregation and color changes from red to purple or blue (Şener et al. 2014a).

DNAzymes as an ideal recognition molecule toward specific targets with high selectivity are used for sensing. Because of their easy operation, high sensitivity, and easily detectable signals, numerous DNA aptamer-based sensors have been developed for the detection of heavy metals (Liu et al. 2008a; Wu et al. 2012a).

Kim et al. developed a high-affinity DNA aptamer for arsenic that can bind to arsenate [As(V)] and arsenite [As(III)] with a dissociation constant of 5 and 7 nM, respectively. The specific affinity interactions of the Ars-III aptamer to arsenic were verified against other heavy metals. They also applied Ars-III for removal of any arsenic present in the groundwater samples collected from the studied areas in Vietnam after obtaining successful removal results with a laboratory-prepared aqueous arsenic solution. They demonstrated that various arsenic concentrations ranging from 28.1 to 739.2 μ g/L were completely removed after 5 min of incubation with the arsenic-binding aptamer Ars-III (Kim et al. 2009).

In another recent study, a selective colorimetric method was developed by using green silver nanoparticles that were synthesized from plant extracts (mango, neem, pepper, and tea plants) to detect Hg^{2+} , Pb^{2+} , and Zn^{2+} ions in aqueous solutions across a wide range of pH (2.0–11). The complicated interactions between surfacebound plant extracts and metal ions enable detection of Hg^{2+} , Pb^{2+} , Zn^{2+} ions across a wide pH range (Kumar and Anthony 2014). Similarly, a colorimetric method was reported for the detection of Cd^{2+} , Hg^{2+} , and Pb^{2+} ions at ppm level in aqueous solution through metal ion interacting functionality. They synthesized silver nanoparticles functionalized with different organic ligands such as *N*-(2-hydroxybenzyl)-isoleucine (Slocik et al. 2008).

1.4 Conclusions

Recent advances in sensor technology have revolutionized our ability to characterize and quantify heavy metal contaminations, and undoubtedly offer benefits for environmental pollution control and early warning. Sensors have several significant and important advantages for such applications, for example: (1) the sensors provide rapid, sensitive, selective, and simple assay method; (2) novel materials may offer unique properties for real-time in-situ assays of binding kinetics between heavy metals and functional molecules; (3) rapid toxicity screening and multi-analyte testing of the environmental pollutants by more smaller, compact, adaptable, and robust sensors developed by sensing arrays can be enabled; (4) miniaturization of optical recognition elements by integration of microfluidics and microelectronics into sensors; and (5) emergence of environmental sensor networks facilitated by wireless-communication technology. Although various challenges still remain in creating improved, cost-effective, and more reliable sensors, sensors will provide the most productive means for heavy metal contamination control and early warning.

References

- Ahmad LM, Towe B, Wolf A, Mertens F, Lerchner J (2010) Binding event measurement using a chip calorimeter coupled to magnetic beads. Sensors Actuators B Chem 145:239–245. https:// doi.org/10.1016/j.snb.2009.12.012
- Altintas Z, Tothill I (2013) Biomarkers and biosensors for the early diagnosis of lung cancer. Sensors Actuators B Chem 188:988–998. https://doi.org/10.1016/j.snb.2013.07.078
- Avuthu SGR, Wabeke JT, Narakathu BB, Maddipatla D, Arachchilage JS, Obare SO, Atashbar MZ (2016) A screen printed phenanthroline-based flexible electrochemical sensor for selective detection of toxic heavy metal ions. IEEE Sensors J 16:8678–8684. https://doi.org/10.1109/JSEN.2016.2572184
- Bellan LM, Wu D, Langer RS (2011) Current trends in nanobiosensor technology. WIREs Nanomed Nanobiotechnol 3:229–246. https://doi.org/10.1002/wnan.136
- Bontidean I, Berggren C, Johansson G, Csöregi E, Mattiasson B, Lyoyd JR, Jakeman K, Brown NL (1998) Detection of heavy metal ions at femtomolar levels using protein-based biosensors. Anal Chem 70:4162–4169. https://doi.org/10.1021/ac9803636
- Borisov SM, Wolfbeis OS (2008) Optical biosensors. Chem Rev 108:423–461. https://doi. org/10.3390/s131013928.
- Cao Z, Guo J, Fan X, Xu J, Fan Z, Du B (2011) Detection of heavy metal ions in aqueous solution by P(MBTVBC-co-VIM)-coated QCM sensor. Sensors Actuators B Chem 157:34–41. https:// doi.org/10.1016/j.snb.2011.03.023
- Carvalho CML, Chew EH, Hashemy SI, Lu J, Holmgren A (2008) Inhibition of the human thioredoxin system-a molecular mechanism of mercury toxicity. J Biol Chem 283:11913–11923. https://doi.org/10.1074/jbc.M710133200
- Chen YY, Chang HT, Shiang YC, Hung YL, Chiang CK, Huang CC (2009) Colorimetric assay for lead ions based on the leaching of gold nanoparticles. Anal Chem 81:9433–9439. https://doi.org/10.1021/ac9018268
- Chen L, Fu XL, Lu WH, Chen LX (2013) Highly sensitive and selective colorimetric sensing of Hg²⁺ based on the morphology transition of silver nanoprisms. ACS Appl Mater Interfaces 5:284–290. https://doi.org/10.1021/am3020857
- Clarkson TW, Magos L, Myers GJ (2003) The toxicology of mercury-current exposures and clinical manifestations. N Engl J Med 349:1731–1737. https://doi.org/10.1056/NEJMra022471
- Cooper MA, Singleton VT (2007) A survey of the 2001 to 2005 quartz crystal microbalance biosensor literature: applications of acoustic physics to the analysis of biomolecular interactions. J Mol Recogn 20:154–184. https://doi.org/10.1002/jmr.826
- Cui L, Wu J, Ju H (2015) Electrochemical sensing of heavy metal ions with inorganic, organic and bio-materials. Biosens Bioelectron 63:276–286. https://doi.org/10.1016/j.bios.2014.07.052
- Danielsson B, Mosbach K (1979) Determination of enzyme activities with the enzyme thermistor unit. FEBS Lett 101:47–50. https://doi.org/10.1016/0014-5793(79)81292-2
- Dinh T, Cullum D (2000) Biosensors and biochips: advances in biological and medical diagnosis. J Anal Chem 366:540–551. https://doi.org/10.1007/s002160051549.
- Domínguez-González R, Varela LG, Bermejo-Barrera P (2014) Functionalized gold nanoparticles for the detection of arsenic in water. Talanta 118:262–269
- Du J, Jiang L, Shao Q, Liu X, Marks RS, Ma J, Chen X (2013) Colorimetric detection of mercury ions based on plasmonic nanoparticles. Small 9:1467–1481. https://doi.org/10.1002/ smll.201200811
- Dutta K, Panda S (2018) Identification of the levels of interference of ions toward heavy metal detection in electrochemical sensors using the barrier width technique. J Electrochem Soc 165:B378–B389. https://doi.org/10.1149/2.1331809jes
- Erdem Ö, Saylan Y, Andaç M, Denizli A (2018) Molecularly imprinted polymers for removal of metal ions: an alternative treatment method. Biomimetics 3:3040038. https://doi.org/10.3390/ biomimetics3040038

- Fernández E, Vidal L, Martín-Yerga D, CarmenBlanco M, Canals A, Costa-García A (2015) Screen-printed electrode based electrochemical detector coupled with ionic liquid dispersive liquid-liquid microextraction and microvolume back-extraction for determination of mercury in water samples. Talanta 135:34–40. https://doi.org/10.1016/j.talanta.2014.11.069
- Forzani E, Zhang H, Chen W, Tao AN (2005) Detection of heavy metal ions in drinking water using a high-resolution differential surface plasmon resonance sensor. Environ Sci Technol 39:1257–1262. https://doi.org/10.1021/es049234z
- Giamblanco N, Conoci S, Russoc D, Marletta G (2015) Single-step label-free hepatitis B virus detection by a piezoelectric biosensor. RSC Adv 5:38152–38158. https://doi.org/10.1039/ C5RA03467A
- Gong X, Bi Y, Zhao Y, Liu G, Teoh WY (2014) Graphene oxide-based electrochemical sensor: a platform for ultrasensitive detection of heavy metal ions. RSC Adv 4:24653–24658. https://doi.org/10.1039/c4ra02247e
- Goode JA, Rushworth JVH, Millner PA (2015) Biosensor regeneration: a review of common techniques and outcomes. Langmuir 31:6267–6276. https://doi.org/10.1021/la503533g
- Huang GS, Wang M, Su C, Chen Y, Hong M (2007) Picogram detection of metal ions by melaninsensitized piezoelectric sensor. Biosens Bioelectron 23:319–325. https://doi.org/10.1016/j. bios.2007.04.011
- Hüseynli S, Çimen D, Bereli N, Denizli A (2019) Molecular imprinted based quartz crystal microbalance nanosensors for mercury detection. Global Chall 3:1800071–1800077. https://doi. org/10.1002/gch2.201800071
- Hwang JH, Wang X, Zhao D, Rex MM, Cho HJ, Lee WH (2019) A novel nanoporous bismuth electrode sensor for in situ heavy metal detection. Electrochim Acta 298:440–448. https://doi.org/10.1016/j.electacta.2018.12.122
- Jalilzadeh M, Çimen D, Özgür E, Esen C, Denizli A (2019) Design and preparation of imprinted surface plasmon resonance (SPR) nanosensor for detection of Zn(II) ions. J Macromol Sci Pure Appl Chem 56:877–886. https://doi.org/10.1080/10601325.2019.1617634
- Kalluri JR, Khan SA, Neely A, Candice P, Varisli B, Washington M, McAfee S, Robinson B, Banerjee S, Singh AK, Senapati D (2009) Use of gold nanoparticles in a simple colorimetric and ultrasensitive dynamic light scattering assay: selective detection of arsenic in groundwater. Angew Chem Int Ed 48:1–5. https://doi.org/10.1002/anie.200903958
- Karaseva NA, Farafonova OV, Ermolaeva TN (2016) Highly sensitive detection of okadaic acid in seafood products via the unlabeled piezoelectric sensor. Food Anal Methods 9:1495–1501. https://doi.org/10.1007/s12161-015-0332-2
- Kim YJ, Johnson RC, Hupp JT (2001) Gold nanoparticle-based sensing of "spectroscopically silent" heavy metal ions. Nano Lett 1:165–167. https://doi.org/10.1021/nl0100116
- Kim M, Bang S, Min J, Kim YH (2009) Arsenic removal from Vietnamese groundwater using the arsenic-binding DNA aptamer. Environ Sci Technol 43:9335–9340. https://doi.org/10.1021/ es902407g
- Kim HN, Ren WX, Kim JS, Yoon J (2012) Fluorescent and colorimetric sensors for detection of lead, cadmium, and mercury ions. Chem Soc Rev 41:3210–3244. https://doi.org/10.1039/ C1CS15245A
- Kimmel DW, LeBlanc G, Meschievitz ME, Cliffel DE (2012) Electrochemical sensors and biosensors. Anal Chem 84:685–707. https://doi.org/10.1021/ac202878q
- Kirchner P, Oberländer J, Friedrich P, Berger J, Rysstad G, Keusgen M, Schöning MJ (2012) Realisation of a calorimetric gas sensor on polyimide foil for applications in aseptic food industry. Sensors Actuators B Chem 170:60–66. https://doi.org/10.1016/j.proeng.2010.09.098.
- Kumar VV, Anthony SP (2014) Silver nanoparticles based selective colorimetric sensor for Cd²⁺, Hg²⁺ and Pb²⁺ ions: tuning sensitivity and selectivity using co-stabilizing agents. Sensors Actuators 191:31–36. https://doi.org/10.1016/j.snb.2013.09.089
- Lahrich S, Hammani H, Boumya W, Loudiki A, Bouabi E, Elmoubarki R, Farahi A, Achak M, Bakasse M, El Mhammedi MA (2016) Effect of natural phosphate to remove silver interference

in the detection of mercury(II) in aquatic algae and seawater samples. Sens Biosensing Res 7:77–83. https://doi.org/10.1016/j.sbsr.2016.01.008

- Lee JS, Mirkin CA (2008) Chip-based scanometric detection of mercuric ion using DNAfunctionalized gold nanoparticles. Anal Chem 80:6805–6808. https://doi.org/10.1021/ ac801046a
- Lee JS, Han MS, Mirkin CA (2007) Colorimetric detection of mercuric ion (Hg²⁺) in aqueous media using DNA-functionalized gold nanoparticles. Angew Chem Int Ed 46:4093–4096. https://doi.org/10.1002/anie.200700269
- Lee S, Oh J, Kim D, Piao Y (2016) A sensitive electrochemical sensor using an iron oxide/graphene composite for the simultaneous detection of heavy metal ions. Talanta 160:528–536. https://doi.org/10.1016/j.talanta.2016.07.034
- Li D, Wieckowska A, Willner I (2008) Optical analysis of Hg²⁺ ions by oligonucleotide-goldnanoparticle hybrids and DNA-based machines. Angew Chem Int Ed 47:3927–3931. https:// doi.org/10.1002/anie.200705991
- Li S, Li Y, Chen H, Horikawa S, Shen W, Simonian A, Chin BA (2010) Direct detection of Salmonella typhimurium on fresh produce using phage-based magnetoelastic biosensors. Biosens Bioelectron 26:1313–1319. https://doi.org/10.1016/j.bios.2010.07.029
- Lin YW, Huang CC, Chang HT (2011) Gold nanoparticle probes for the detection of mercury, lead and copper ions. Analyst 136:863–871. https://doi.org/10.1039/c0an00652a
- Liu CW, Hsieh YT, Huang CC, Lin ZH, Chang HT (2008a) Detection of mercury(II) based on Hg²⁺-DNA complexes inducing the aggregation of gold nanoparticles. Chem Commun 19:2242–2244. https://doi.org/10.1039/b719856f
- Liu CW, Huang CC, Chang HT (2008b) Control over surface DNA density on gold nanoparticles allows selective and sensitive detection of mercury(II). Langmuir 24:8346–8350. https://doi.org/10.1021/1a800589m
- Liu DB, Qu WS, Chen WW, Zhang W, Wang Z, Jiang X (2010) Highly sensitive, colorimetric detection of mercury(II) in aqueous media by quaternary ammonium group-capped gold nanoparticles at room temperature. Anal Chem 82:9606–9610. https://doi.org/10.1021/ac1021503
- Liu DB, Wang SJ, Swierczewska M, Huang X, Bhirde AA, Sun J, Wang Z, Yang M, Jiang X, Chen X (2012) Highly robust, recyclable displacement assay for mercuric ions in aqueous solutions and living cells. ACS Nano 6:10999–11008. https://doi.org/10.1021/nn3046192
- Liu X, Yao Y, Ying Y, Ping J (2019) Recent advances in nanomaterial-enabled screen-printed electrochemical sensors for heavy metal detection. Trends Anal Chem 115:187–202. https://doi. org/10.1016/j.trac.2019.03.021
- Lou H, Zhang Y, Xiang Q, Xu J, Li H, Xu P, Li X (2012) The real-time detection of trace-level Hg²⁺ in water by QCM loaded with thiol-functionalized SBA-15. Sensors Actuators B Chem 166–167:246–252. https://doi.org/10.1016/j.snb.2012.02.053
- Luo C, Tang H, Cheng W, Yan L, Zhang D, Ju H, Ding S (2013) A sensitive electrochemical DNA biosensor for specific detection of Enterobacteriaceae bacteria by exonuclease III-assisted signal amplification. Biosens Bioelectron 48:132–137. https://doi.org/10.1016/j.bios.2013.03.084
- Marx KA (2003) Quartz crystal microbalance: a useful tool for studying thin polymer films and complex biomolecular systems at the solution-surface interface. Biomacromolecules 4:1099–1120. https://doi.org/10.1021/bm020116i
- Mosbach K (1991) Thermal biosensors. Biosens Bioelectron 6:179–182. https://doi. org/10.1016/0956-5663(91)80002-F
- Muslim NZM, Ahmad M, Heng LY, Saad B (2012) Optical biosensor test strip for the screening and direct determination of l-glutamate in food samples. Sensors Actuators B Chem 161:493–497. https://doi.org/10.1016/j.snb.2011.10.066
- Nieuwenhuizen MS, Venema A (1991) Mass-sensitive devices. Sensors Set A Compr Surv:647–680. https://doi.org/10.1002/9783527619269.ch13a
- Pesavento M, Alberti G, Biesuz R (2009) Analytical methods for determination of free metal ion concentration, labile species fraction and metal complexation capacity of environmental waters: a review. Anal Chim Acta 631:129–141. https://doi.org/10.1016/j.aca.2008.10.046

- Placido T, Aragay G, Pons J, Comparelli R, Curri ML, Merkoci A (2013) Ion-directed assembly of gold nanorods: a strategy for mercury detection. ACS Appl Mater Interf 5:1084–1092. https:// doi.org/10.1021/am302870b
- Raiteri R, Grattarola M, Butt HJ, Skládal P (2001) Micromechanical cantilever-based biosensors. Sens Actuators B 79:115–126. https://doi.org/10.1016/S0925-4005(01)00856-5
- Ramanathan K, Danielsson B (2001) Principles and applications of thermal biosensors. Biosens Bioelectron 16:417–423. https://doi.org/10.1016/s0956-5663(01)00124-5
- Rogers KR (2006) Recent advances in biosensor techniques for environmental monitoring. Anal Chim Acta 568:222–231. https://doi.org/10.1016/j.aca.2005.12.067
- Rudnitskaya A, Legin A, Seleznev B, Kirsanov D, Vlasov Y (2008) Detection of ultra-low activities of heavy metal ions by an array of potentiometric chemical sensors. Microchim Acta 163:71–80. https://doi.org/10.1007/s00604-007-0900-2
- Sadrolhossein AR, Naseri M, Rashid SA (2017) Polypyrrole-chitosan/nickel-ferrite nanoparticle composite layer for detecting heavy metal ions using surface plasmon resonance technique. Opt Laser Technol 93:216–223. https://doi.org/10.1016/j.optlastec.2017.03.008
- Safran V, Göktürk I, Derazshamshir A, Yılmaz F, Sağlam N, Denizli A (2019) Rapid sensing of Cu⁺² in water and biological samples by sensitive molecularly imprinted based plasmonic biosensor. Microchem J 148:141–150. https://doi.org/10.1016/j.microc.2019.04.069
- Saha K, Agasti SS, Kim C, Li XN, Rotello VM (2012) Gold nanoparticles in chemical and biological sensing. Chem Rev 112:2739–2779. https://doi.org/10.1021/cr2001178
- Sartore L, Barbaglio M, Borgese L, Bontempi E (2011) Polymer-grafted QCM chemical sensor and application to heavy metal ions real time detection. Sensors Actuators B Chem 155:538–544. https://doi.org/10.1016/j.snb.2011.01.003
- Saylan Y, Akgönüllü S, Çimen D, Derazshamshir A, Bereli N, Yılmaz F, Denizli A (2017a) Development of surface plasmon resonance sensors based on molecularly imprinted nanofilms for sensitive and selective detection of pesticides. Sensors Actuators B Chem 241:446–454. https://doi.org/10.1016/j.snb.2016.10.017
- Saylan Y, Yılmaz F, Derazshamshir A, Yılmaz E, Denizli A (2017b) Synthesis of hydrophobic nanoparticles for real-time lysozyme detection using surface plasmon resonance sensor. J Mol Recognit 2631:1–7. https://doi.org/10.1002/jmr.2631
- Şener G, Uzun L, Denizli A (2014a) Colorimetric sensor array based on gold nanoparticles and amino acids for identification of toxic metal ions in water. ACS Appl Mater Interf 6:18395–18400. https://doi.org/10.1021/am5071283
- Şener G, Uzun L, Denizli A (2014b) Lysine-promoted colorimetric response of gold nanoparticles: a simple assay for ultrasensitive mercury(II) detection. Anal Chem 86:514–520. https://doi. org/10.1021/ac403447a
- Sepulveda B, Angelome PC, Lechuga LM, Liz-Marzan LM (2009) LSPR-based nanobiosensors. Nano Today 4:244–251. https://doi.org/10.1016/j.nantod.2009.04.001
- Shankaran D, Gobi K, Miura N (2007) Recent advancements in surface plasmon resonance immunosensors for detection of small molecules of biomedical, food and environmental interest. Sensors Actuators B Chem 121:158–177. https://doi.org/10.1016/j.snb.2006.09.014
- Shen LL, Zhang GR, Li W, Biesalski M, Etzold BJM (2017) Modifier-free microfluidic electrochemical sensor for heavy-metal detection. ACS Omega 2:4593–4603. https://doi.org/10.1021/ acsomega.7b00611
- Slocik JM, Zabinski JS, Phillips DM, Naik RR (2008) Colorimetric response of peptidefunctionalized gold nanoparticles to metal ions. Small 4:548–551. https://doi.org/10.1002/ smll.200700920
- Song L, Mao K, Zhou X, Hu J (2016) A novel biosensor based on Au@Ag core-shell nanoparticles for SERS detection of arsenic (III). Talanta 146:285–290. https://doi.org/10.1016/j. talanta.2015.08.052
- Sun M, Ding B, Yu J (2012) Sensitive metal ion sensors based on fibrous polystyrene membranes modified by polyethyleneimine. RSC Adv 2:1373–1378. https://doi.org/10.1039/C1RA00673H

- Sun P, Chen Y, Yan M, Tang T (2019) Enhancement of QCM detection for heavy metal ions based on TGA modified CdTe nanospheres. J Inorg Organomet Polym. https://doi.org/10.1007/ s10904-019-01212-1
- Thevenot DR, Toth K, Durst RA, Wilson GS (2001) Electrochemical biosensors: recommended definitions and classification. Biosens Bioelectron 16:121–131. https://doi.org/10.1016/ s0956-5663(01)00115-4.
- Udhayakumari D, Suganya S, Velmathi S, MubarakAli D (2014) Naked eye sensing of toxic metal ions in aqueous medium using thiophene-based ligands and its application in living cells. J Mol Recognit 27:151–159. https://doi.org/10.1002/jmr.2343
- Ullah N, Mansha M, Khan I, Qurashi A (2018) Nanomaterial-based optical chemical sensors for the detection of heavy metals in water: recent advances and challenges. Trends Anal Chem 100:155–166. https://doi.org/10.1016/j.trac.2018.01.002
- Van Dorst B, Mehta J, Bekaert K, Rouah-Martin E, De Coen W, Dubruel P, Blust R, Robbens J (2010) Recent advances in recognition elements of food and environmental biosensors: a review. Biosens Bioelectron 26:1178–1194. https://doi.org/10.1016/j.bios.2010.07.033
- Van Leeuwen HP, Town RM, Buffle J, Cleven RFMJ, Davison W, Puy J, Van Riemsdijk WH, Sigg L (2005) Dynamic speciation analysis and bioavailability of metals in aquatic systems. Environ Sci Technol 39:8545–8556. https://doi.org/10.1021/es050404x
- Verm R, Gupta BD (2015) Detection of heavy metal ions in contaminated water by surface plasmon resonance based optical fibre sensor using conducting polymer and chitosan. Food Chem 166:568–575. https://doi.org/10.1016/j.foodchem.2014.06.045
- Verma N, Bhardwaj A (2015) Biosensor technology for pesticides a review. Appl Biochem Biotechnol 175:3093–3119. https://doi.org/10.1007/s12010-015-1489-2
- Vimala A, Vedhi C (2019) Electrochemical sensors for heavy metals detection in Gracilaria corticata using multiwalled carbon nanotubes modified glassy carbon electrode. J Anal Chem 74:276–285. https://doi.org/10.1134/S106193481903002X
- Wanekaya AK, Chen W, Mulchandani A (2008) Recent biosensing developments in environmental security. J Environ Monit 10:703–712. https://doi.org/10.1039/b806830p
- Wang Y, Yang F, Yang XR (2010) Colorimetric detection of mercury(II) ion using unmodified silver nanoparticles and mercury-specific oligonucleotides. ACS Appl Mater Interfaces 2:339–342. https://doi.org/10.1021/am9007243
- Wang Y, Jin J, Yuan C, Zhang F, Ma L, Qin D, Shana D, Lu X (2015) A novel electrochemical sensor based on zirconia/ordered macroporous polyaniline for ultrasensitive detection of pesticides. Analyst 140:560–566. https://doi.org/10.1039/c4an00981a
- Wang X, Sun J, Tong J, Guan X, Bian C, Xia S (2018) Paper-based sensor chip for heavy metal ion detection by SWSV. Micromachines 9:150–161. https://doi.org/10.3390/mi9040150
- Wei QS, Nagi R, Sadeghi K, Feng S, Yan E, Ki SJ, Caire R, Tseng D, Ozcan A (2014) Detection and spatial mapping of mercury contamination in water samples using a smart-phone. ACS Nano 8:1121–1129. https://doi.org/10.1021/nn406571t
- Weng C, Cang JS, Chang JY, Hsiung TM, Hung YL, Tseng YT, Li YJ, Shen YW, Huang CC (2014) Detection of As(III) through pulsed laser-induced desorption/ionization of gold nanoparticles on cellulose membranes. Anal Chem 86:3167–3173. https://doi.org/10.1021/ac500053e
- Wu Y, Liu L, Zhan S, Wang F, Zhou P (2012a) Ultrasensitive aptamer biosensor for arsenic (III) detection in aqueous solution based on surfactant-induced aggregation of gold nanoparticles. Analyst 137:4171–4178. https://doi.org/10.1039/c2an35711a
- Wu Y, Zhan S, Wang F, He L, Zhi W, Zhou P (2012b) Cationic polymers and aptamers mediated aggregation of gold nanoparticles for the colorimetric detection of arsenic(III) in aqueous solution. Chem Commun 48:4459–4461. https://doi.org/10.1039/c2cc30384a
- Xie B, Ramanathan K, Danielsson B (2000) Mini/micro thermal biosensors and other related devices for biochemical/clinical analysis and monitoring. Trends Anal Chem 19:340–349. https://doi.org/10.1016/S0165-9936(99)00211-3.

- Ye BC, Yin BC (2008) Highly sensitive detection of mercury(II) ions by fluorescence polarization enhanced by gold nanoparticles. Angew Chem Int Ed 47:8386–8389. https://doi.org/10.1002/ anie.200803069
- Yılmaz E, Özgür E, Bereli N, Türkmen D, Denizli A (2017a) Plastic antibody based surface plasmon resonance nanosensors for selective atrazine detection. Mater Sci Eng C 73:603–610. https://doi.org/10.1016/j.msec.2016.12.090
- Yılmaz F, Saylan Y, Akgönüllü S, Çimen D, Derazshamshir A, Bereli N, Denizli A (2017b) Surface plasmon resonance based nanosensors for detection of triazinic pesticides in agricultural foods. New Pestic Soil Sens 18:679–718. https://doi.org/10.1016/B978-0-12-804299-1.00019-9
- Yu Y, Hong Y, Gao P, Nazeeruddin MK (2016a) Glutathione modified gold nanoparticles for sensitive colorimetric detection of Pb²⁺ ions in rainwater polluted by leaking perovskite solar cells. Anal Chem 88:12316–12322. https://doi.org/10.1021/acs.analchem.6b03515
- Yu LL, Shi MX, Yue X, Qu LB (2016b) Detection of allura red based on the composite of poly (diallyldimethylammonium chloride) functionalized graphene and nickel nanoparticles modified electrode. Sensor Actuators B 225:398–404. https://doi.org/10.1016/j.snb.2015.11.061
- Zhang S, Zhu H, Ma P, Duan F, Dong W, Du M (2017) A self-supported electrochemical sensor for simultaneous sensitive detection of trace heavy metal ions based on PtAu alloy/carbon nanofibers. Anal Methods 9:6801–6808. https://doi.org/10.1039/C7AY02223A
- Zhao N, Chen C, Zhou J (2012) Surface plasmon resonance detection of ametryn using a molecularly imprinted sensing film prepared by surface-initiated atom transfer radical polymerization. Sensors Actuators B Chem 166:473–479. https://doi.org/10.1016/j.snb.2012.02.089.
- Zheng QL, Han CP, Li HB (2010) Selective and efficient magnetic separation of Pb²⁺ via gold nanoparticle-based visual binding enrichment. Chem Commun 46:7337–7339. https://doi.org/10.1039/c0cc02915g
- Zhu D, Li X, Liu X, Wang J, Wang Z (2012) Designing bifunctionalized gold nanoparticle for colorimetric detection of Pb²⁺ under physiological condition. Biosens Bioelectron 31:505–509. https://doi.org/10.1016/j.bios.2011.11.026
Chapter 2 Nanosensors for Heavy Metal Detection in Environmental Media: Recent Advances and Future Trends



Xiaojia He, Hua Deng, and Huey-min Hwang

Contents

2.1	Introduction			
2.2	Detection Mechanisms.			
	2.2.1	Surface Plasma Resonance.	26	
	2.2.2	UV-Visible Spectroscopy	29	
	2.2.3	Surface-Enhanced Infrared Absorption.	30	
	2.2.4	Surface-Enhanced Raman Scattering	32	
	2.2.5	Surface-Enhanced Fluorescence.	35	
	2.2.6	Fluorescence Quenching-Based Nanoprobes	37	
	2.2.7	Self-Propelled Nanojets.	39	
2.3	Application Examples and Detection Limit.			
	2.3.1	Hg ²⁺ Detection.	41	
	2.3.2	Pb ²⁺ Detection.	42	
	2.3.3	As ³⁺ Detection.	42	
	2.3.4	Cd ²⁺ Detection.	43	
	2.3.5	Cu ²⁺ Detection.	43	
	2.3.6	Cr ³⁺ and Cr ⁶⁺ Detection.	43	
2.4	Outlook			
	2.4.1	Green Synthesis.	44	
	2.4.2	Minimizing Nanotoxicity	45	
Refe	rences.	~ · ·	46	

X. He (🖂)

Department of Marine Sciences, The University of Georgia, Athens, GA, USA

Present Affiliation: Department of Medicine, Emory University School of Medicine, Atlanta, GA, USA e-mail: xiaojia.he25@uga.edu

H. Deng Department of Chemistry, Morgan State University, Baltimore, MD, USA

H.-m. Hwang (⊠) Department of Biology, Jackson State University, Jackson, MS, USA e-mail: huey-min.hwang@jsums.edu

© The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_2 23

Abstract Heavy metal pollution has long been an environmental and health problem for thousands of years. Although heavy metal contamination has been well studied and regulated nationally and internationally, remediation of heavy metal polluted environmental media remains a complicated challenge, as exposure to heavy metal continues. Novel heavy metal remediation has become a hot topic being vigorously researched in environmental science and engineering. The emergence of nanotechnology over the last two decades offers new solutions to the existing problems of the environment. Fabricated nanosensors are well recognized to be highly selective and sensitive toward certain heavy metal pollutants. Optical properties including light extinction, intensity, and scattering can be substantially altered and enhanced as a result of surface plasmon resonance phenomenon possessed by metallic nanostructures such as gold, platinum, and silver, and/or achieved by inner filter effect. Moreover, novel nanosensors could act as fluorescence quencher to produce strong fluorescence signals, or act as surface catalyzer to generate autonomous motion. These distinct mechanical, chemical, and electrical properties make many nanosensors promising candidates for environmental remediation. A variety of nanosensors, for example, surface-enhanced Raman spectroscopy and nanojets, has been designed and applied for heavy metal detection. In this review, we present a brief overview of the recent advances in the fabrication of novel nanosensors that provide potential solutions to heavy metal remediation. We also intend to provide perspective on strategies that are needed to assess and reduce the potential hazard of manufactured nanomaterials in a sustainable manner.

Keywords Heavy metal · Nanosensor · Surface plasmon resonance · Fluorescence quenching · Nanojets · Green synthesis · Nanotoxicology

2.1 Introduction

Heavy metal pollution has a long history associated with the development of human society (Hong et al. 1996; Nriagu 1996). Over the past century, due to the enormous mining activities and rapidly expanding industrial processing, environmental contamination with heavy metals has become a worldwide health issue. Main heavy metal pollutants reported in contaminated land and water include lead (Pb), mercury (Hg), arsenic (As), cadmium (Cd), copper (Cu), and chromium (Cr) (European Environment Agency 2016). According to the European Environment Agency (EEA), heavy metal in a total of 3.45×10^7 kg was released from Europe's industry (33,000 facilities in 33 countries including 28 European Union members) for the period 2007–2015 (European Environment Agency 2016). As shown in Fig. 2.1a, 1.55, 0.6, 7.99, 9.9, 13.82, and 0.65 million kg of As, Cd, Cr, Cu, Pb, and Hg were released, in which 50%, 48%, and 2% were directly released to water, air, and soil (Fig. 2.1b). By looking at the contribution to environmental pressure from releases of heavy metals, urban wastewater treatment plants, agriculture, and mining



Fig. 2.1 Heavy metal releases from Europe's industry for the period 2007–2016. Total release: 3.45×10^7 kg. (a) Distribution of heavy metals. (b) Environmental pressure to water, air, and soil. (c) Contribution to environmental pressure from release of heavy metals to water (note heavy metal releases to air are mainly from industrial processing). (d) Annual release from 2007 to 2015. Data sources: European Environment Agency. The European Pollutant Release and Transfer Register (E-PRTR), Member States reporting under Article 7 of Regulation (EC) No 166/2006

contributed 78.8% of the total release to water, whereas industrial processing largely contributed to the total release to air (Fig. 2.1c).

These heavy metals are known to cause severe heavy metal poisoning and illness at toxic doses to plants, animals, and humans, as a result of air or water contamination and heavy metal contaminated food products. Many of the heavy metal pollutants such as inorganic arsenic can accumulate in the vitals of organisms, including liver, tissues, spleen, and kidneys. Acute and chronic heavy metal poisoning may lead to symptoms like skin irritation, nausea, and seizure, and life-threatening complications such as brain damage, hemolysis, and even death. Additionally, heavy metals released by anthropogenic practices tend to persist in the environment for a long time with little microbial degradation. Heavy metal pollution thus poses great risks and hazards to our ecosystem and humans.

Heavy metal contamination has been observed in almost every type of environment and all locations on earth, from breathing air (Li et al. 2013a) to drinking water (Mohan et al. 1996), land (Wuana and Okieimen 2011) to ocean (McIntyre and He 2019; Millward et al. 2019), and Arctic (McConnell and Edwards 2008) to Antarctic (De Moreno et al. 1997). The effective monitoring and hazard assessment of heavy metal contaminants in a variety of environmental matrices is thus a prominent, yet challenging, section in ensuring environmental safety and human health (Gray et al. 2019). With the recent development in nanotechnology and nanoscience, engineered nanomaterials has received an ever-increasing attention with broad applications in many fields, including but not limited to biomedical researches (He et al. 2015b), environmental remediation (Ma et al. 2015; He et al. 2016a), transistor industry (Franklin 2015), energy application (Khan et al. 2017), and food and agriculture (He and Hwang 2016; He et al. 2019a). Nanotechnology has evolved to offer a potential solution to this challenging task: nanosensors (Su et al. 2012; Li et al. 2013b; Kumar et al. 2017). Recent market share forecasts that nanosensor market share is anticipated to grow from ~ 100 million USD to ~ 4 billion USD between 2017 and 2023, at a compound annual growth rate of 79.83% (Mordor Intelligence 2019). Nanosensors are engineered for chemical and mechanical applications because of the unique physicochemical properties at nanoscale. Successful examples have been reported in monitoring chemical pollution in our living environment (Su et al. 2012; Vikesland 2018) and agriculture practices (Baruah and Dutta 2009). Many nanosensors have also been designed and used to detect and monitor heavy metals in natural environmental media. Highly sensitive and selective nanosensors can be designed and modified to accommodate a large number of environmental applications.

Here we briefly present recent progress in nanosensor research and development for heavy metal contaminant detection. We aim to review mechanisms for detecting heavy metals and the potential contributions of nanotechnology-based sensors to human health in a sustainable manner.

2.2 Detection Mechanisms

2.2.1 Surface Plasma Resonance

Metallic nanoparticles are known to induce surface plasmon resonance as a result of electron oscillation due to light irradiation (incident electromagnetic wave E_o), as schematically illustrated in Fig. 2.2a. The electron cloud near nanoparticle's surface then displaces under the oscillating electric field relative to nuclei. The collective oscillation and displacement of the electron cloud of metallic nanoparticle induces electric dipole (Fig. 2.2b, c) for small nanoparticles or higher quadrupole modes (Fig. 2.2c, e) for large nanoparticles. Nanoparticle surface plasmon resonance ultimately leads to the amplification of the electric field near the metal surface (E_{out}), which can be characterized by (Kelly et al. 2003)



Fig. 2.2 (a) Schematic illustration of surface plasmon resonance showing the electron cloud displacement. (b–e) Electric field for Ag nanoparticles. (b and c) show the cross-section plane with propagation and polarization axes and (d and e) show the cross-section plane perpendicular to the propagation axis. (b and d) show 30 nm Ag at the main extinction peak of 369 nm and (c and f) show 60 nm Ag at the quadrupole peak of 358 nm. (f) shows the extinction spectra of nanoparticles with various shapes indicated by major to minor axis ratio (ranging from 1 to 10). Reproduced with permission from Kelly et al. (2003), American Chemical Society

$$E_{out} = E_o \vec{x} - \alpha E_o \left(\frac{\vec{x}}{r^3} - \frac{3x}{r^5} (x\vec{x} + y\vec{y} + z\vec{z}) \right)$$

where α is the nanoparticle polarizability, $x, y, z, \vec{x}, \vec{y}, \vec{z}$ are the Cartesian coordinates and unit vectors, and r is the radial distance. $E_{\alpha}\vec{x}$ represents the contribution of the

incident electromagnetic wave to electric field, and $\alpha E_o\left(\frac{\vec{x}}{r^3} - \frac{3x}{r^5}\left(x\vec{x} + y\vec{y} + z\vec{z}\right)\right)$

represents the contribution of the surface plasmon resonance to electric field. For quadrupole modes, E_{out} can be characterized by (Kelly et al. 2003)

$$E_{out} = E_o \vec{x} + ikE_o \left(x\vec{x} + z\vec{z} \right) - \alpha E_o \left(\frac{\vec{x}}{r^3} - \frac{3x}{r^5} \left(x\vec{x} + y\vec{y} + z\vec{z} \right) \right)$$
$$-\beta E_o \left(\frac{x\vec{x} + z\vec{z}}{r^5} - \frac{5z}{r^7} \left(x^2\vec{x} + y^2\vec{y} + xz\vec{z} \right) \right)$$

where β is the quadrupole polarizability.

Optical characteristics including extinction, absorption, and scattering spectra exhibit distinct features for nanoparticles with different chemical and physical properties. Hence, a variety of methodologies have been used for the detection of nanoparticle surface plasmon resonance features. Noticeably, surface-enhanced Raman spectroscopy (SERS) has been shown to be highly sensitive to the electromagnetic fields at or close to the surface of nanomaterials (Ding et al. 2017), exhibiting distinct extinction spectra in response to different particle sizes and shapes (Kelly et al. 2003). For instance, the relatively large dipole plasmon peak for small Ag nanoparticles (30 nm; Fig. 2.2b, d) can lead to strong electromagnetic enhancements in SERS. One more example is given to illustrate the particle shape dependent extinction properties in Fig. 2.2e. By a given nanoparticle volume equivalent to that for an 80 nm radius sphere, an increasing red shift of the surface plasmon resonance is expected as the shape changes from sphere (ratio of major to minor axes = 1) to highly oblate (ratio of major to minor axes = 10), indicating a substantial dependency of diploe plasmon resonance on particle shape.

Among many metallic nanoparticles, Au, Pt, and Ag nanostructures are mostly studied and applied as nanosensors due to their distinct plasmonic characteristics. They possess similar plasmonic characteristics aforementioned, with dependency on nanoparticle morphology, oxidation states, and aggregation/agglomeration status (Amirjani and Haghshenas 2018). The highly selective and sensitive surface plasmon resonance features then can be used for the heavy metal detection with careful manufacturing. Here we briefly discuss some recent progress on the surface plasmon resonance based nanosensors. Four subcategories are divided to represent different properties exhibited by the surface plasmon resonance, including ultraviolet (UV)-visible spectroscopy, surface-enhanced infrared absorption (SEIRA), surface-enhanced Raman scattering, and surface-enhanced fluorescence.

2.2.2 UV-Visible Spectroscopy

UV-visible spectroscopy is readily accessible in most research facilities, which makes it the common method for quantifying the characteristic changes in plasmonic nanostructures. Agglomeration or aggregation often occurs when plasmonic nanoparticles meet certain heavy metal ions. UV-visible spectrum shift as a result of agglomeration or aggregation then can be observed by UV-visible spectroscopy. Zhang and coworkers reported that a colloidal graphite-like nitride doped carbon quantum dots-capped gold nanoparticle (Au@g-CNQDs) for selective detection of Cd²⁺ ions (Zhang et al. 2018). They found that Cd²⁺ ions were adsorbed onto Au@g-CNQDs, likely due to surface functional groups such as heptazine, carboxyl, and hydroxyl (Fig. 2.3a), leading to the formation of Au@g-CNQDs aggregates



Fig. 2.3 (a) Schematic drawing that represents the Au@g-CNQD aggregation caused by Cd²⁺ ions, and (b and c) show the aggregation status of Au@g-CNQDs without (b) and with 1 mM Cd²⁺ ions (c). (D) UV-visible spectra of plasmonic characteristic shift of Au@g-CNQDs with Cd²⁺ ions. (e and f) Selectivity of Cd²⁺ ions to Au@g-CNQDs. (g) UV-visible spectra of the Au@g-CNQDs with different Cd²⁺ concentrations (0–12 μ M). (h) The colorimetric and spectroscopic observations with different Cd²⁺ concentrations (0–12 μ M). (i) shows a linear detection window with different Cd²⁺ concentrations (10 nM to 12 μ M). Reproduced with permission from Zhang et al. (2018), Elsevier

(Fig. 2.3b, c). The change in aggregation status then resulted in subsequent plasmonic characteristic change. The feature change was recorded by UV-visible spectroscopy with an apparent red shift at 650 nm/520 nm (Fig. 2.3d). Notably, Au@g-CNQDs showed strong selectivity and sensitivity to the Cd²⁺ ions (Fig. 2.3e, f) with a detection limit of 10 nM and linear detection window up to 3 μ M (Fig. 2.3g–i).

Similar UV-visible spectroscopy based on the aggregation or agglomeration of plasmonic nanoparticles was also employed by Ikram et al. (2019). They reported that epicatechin-capped silver nanoparticles (ECAgNPs) were highly selective to Pb²⁺ in blood and water matrices, among many other ions (Ikram et al. 2019). UV-visible spectroscopy shows that Pb²⁺ ion enhanced ECAgNP absorption at 412 nm with a detection limit of 1.52 μ M and up to 100 μ M (Ikram et al. 2019). Notably, UV-visible spectroscopy exhibits to be highly efficient and reliable in environmental samples, with a recovery rate of Cd²⁺ ranging from 97.7% to 99.8% in tap water samples and from 99.3% to 102.8% in lake water samples (Ikram et al. 2019).

2.2.3 Surface-Enhanced Infrared Absorption

Infrared (IR) absorption of the hybrid metallic plasmonic surface can also be enhanced in the presence of organic or biomolecules and heavy metal ions (Enders et al. 2006). The technique used for heavy metal detection is called surface-enhanced infrared absorption (SEIRA). Instead of inducing aggregation or agglomeration of nanoparticles, heavy metal ions could lead to a red shift in infrared plasmon of some nanomaterials, amplified by the nanostructure electromagnetic field (Enders et al. 2006). Literature survey shows little progress on the SEIRA-based nanosensor for heavy metal detection. We use the study from Hoang et al. (2013) as an example to illustrate how SEIRA-based nanosensor functions. Hoang et al. (2013) developed an Au plasmonic nanostructure with thiolated 15-base DNA oligonucleotides to monitor for the in-situ detection of Hg²⁺. The binding of Hg²⁺ with DNA aptamer induced vibrational signal in the infrared spectra. Notably, Au nanostructures functioned as electromagnetic field amplifier and further enhanced the vibrational signal of the DNA aptamer in the presence of Hg^{2+} , even at ppt level in situ. The schematic of the Hg²⁺ binding with DNA aptamer is shown in Fig. 2.4a. The Hg²⁺ ion can bridge two thymine bases by N-Hg-N bonds, resulting in structural and chemical change in the DNA aptamer. Such conformational changes in DNA aptamer in the presence of Hg²⁺ ions subsequently cause the shift in infrared optical spectra (Fig. 2.4b). Altogether, infrared optical spectra features then act as the fundamental mechanism to detect Hg²⁺ ions.

IR spectral features in Fig. 2.4c clearly indicate the development of infrared spectra profile of the Au-DNA nanostructure with Hg²⁺ concentrations increasing from 37 ppt to 37 ppb. Although the peak shape and position of the infrared spectrum peak at $\omega_T = 1400$ cm⁻¹ (red solid line) remains unchanged, the intensity, but not the



Fig. 2.4 (a) Schematic representation of the conformational changes in Au-DNA nanostructures caused by Hg²⁺. The N–Hg–N bonds in the imide structures (red dotted rectangular) then lead to the IR signal change. (b) Infrared spectra of Au-DNA plasmonic nanostructure in pure water (upper panel) and lake water (lower panel). (c) IR spectral features in the presence of Hg²⁺ in lake water samples. (d) IR spectra intensity at $\omega = 1400 \text{ cm}^{-1}$ (DNA signal) and at α : 1558 cm⁻¹, β : 1650 cm⁻¹, and γ : 3300 cm⁻¹ (Hg²⁺ induced signals) with different Hg²⁺ concentration. Vibrational IR signals from the DNA and Hg²⁺ are marked by red and blue lines, respectively. Reproduced with permission from Hoang et al. (2013), licensed under a Creative Commons Attribution-NonCommercial-NoDerivs 3.0 Unported License

peak shape, changes along with the Hg²⁺ concentration. Vibrational bands also occur at 1558 cm⁻¹ (α : C–N stretching and N–H bending modes, Amide II), 1650 cm⁻¹ (β : C=O Amide I), and 3300 cm⁻¹ (γ : N–H stretching mode) in the presence of Hg²⁺ ions. The intensity of the infrared spectrum peak at those three wavenumbers also increases as the Hg²⁺ concentration increases (Fig. 2.4d).

2.2.4 Surface-Enhanced Raman Scattering

The amplified electromagnetic field near the plasmonic nanostructure surface induced by light irradiation (visible and near-infrared radiation) can be recorded as Raman signal and is known as enhancement in surface-enhanced Raman scattering (SERS). Notably, the electromagnetic field enhancement can magnify the light intensity, which in turn further increases Raman scattering signals. The overall SERS measurements can be as much as 10¹⁰-10¹¹, making SERS possible for even single molecule detection. Plasmonic nanostructures such as Au and Ag with plasmon resonance frequencies within visible and near-infrared radiation are widely utilized for SERS measurement. The addition of heavy metals can reduce the SERS signal of plasmonic nanostructures, thus making SERS nanosensors possible to quantify the trace heavy metal concentrations. For example, the chemical interaction between methylthio moiety and plasmonic gold nanoparticles could be disrupted in the presence of copper(II) (Dugandžić et al. 2019). The conformational changes in the ligand can then be quantified by comparative SERS spectrum. Following the mechanism, trace metal ions can be detected as a function of SERS spectrum intensity at certain wavenumbers.

SERS-based nanosensors are rapidly growing in the last decade. Many other SERS nanosensors have been designed to monitor heavy metal ions in water samples with promising results (see selected studies in Table 2.1). Here we take a recent report from Zou et al. (2019) as an example. They reported a novel DNAzyme-Au nanocluster (AuNC) assembly coated on polystyrene microspheres (PSMPs) with DNA logic gates for theultrasensitive detection of Hg²⁺ and Pb²⁺ ion simultaneously (Zou et al. 2019). The newly constructed nanosensor uses Au nanostructured films as the SERS-active substrates. The 8–17 DNAzyme with enzyme strand (S1: 5'ACC TCTTCTTTGTTTCTTGCATCTCTTCTCCGAGCCGGTCGAAATAGTGAGT) and cleavage substrate strand (S2: 5'TGTCAACTCGTGACTCACTATGGAAGAG ATG) was conjugated with the short DNA sequence (S3: 5'GTTGTTTCTTTGTTC and S4: 5'CACGAGTTGACA) on the Au nanostructures. A number of 10 misaligned T-T base pairs in the short DNA sequence S3 were specifically engineered to bind Hg²⁺ by forming T–Hg–T bonds. Consequently, Raman reporter 5,5'-Dithiobis-(2nitrobenzoic acid) labeled AuNCs were bond into the PSMP substrates, exhibiting a magnified SERS intensity at 1330 cm⁻¹. However, in the presence of Pb²⁺ ions, Au nanoconjugates were removed from the PSMP substrates, leading to a lower Raman signal at 1580 cm⁻¹. The binding and detaching of Raman labeled-AuNCs from the PSMP substrates is thus controlled by the presence of Hg²⁺ or Pb²⁺ ions, respectively. The DNA logic gate system is regulated by (0,0), (0,1), (1,0), and (1,1), indicating no Hg²⁺ or Pb²⁺ ions, with Pb²⁺ ions, with Hg²⁺ ions, and with both ions, respectively (schematically shown in Fig. 2.5a). The conformation changes in SERS intensity for four modes were then captured by the Raman spectroscopy, as shown in Fig. 2.5b. Clearly, as Hg²⁺ and Pb²⁺ ions were simultaneously presented, the SERS features at different shift wavenumbers were distinctly observed as detected in Fig. 2.5c, d.

	Targeted	Reported	Detection	Recovery in	
Nanomaterials	metal	range	limits	samples	Reference
Gold nanoparticle	Hg ²⁺	Linear (log):	1.69 pg mL ⁻¹	100-109%	Li et al.
trimers	Ag ²⁺	0.002– 0.5 ng mL ⁻¹	1.71 pg mL ⁻¹	95–101%	(2015)
Citrate- functionalized gold nanoparticle	Pb ²⁺	Linear: 50–1000 ng/L	25 ng/L	98.6%	Frost et al. (2015)
Porous anodic alumina membrane functionalized with CoFe ₂ O ₄ nanoparticles and carbon nanotubes	Pb ²⁺ , Hg ²⁺ , and Cd ²⁺	1–100 ppb	1 ppb	N/A	Shaban and Galaly (2016)
Label-free Fe ₃ O ₄ @ ag	Hg ²⁺	Linear (log): 10 ⁻⁵ –10 ⁻¹¹ M	10 pM (2 ppt)	Tap water: 106–133%	Song et al. (2017)
				Regeneration water: 66.07–96.75%	-
				Secondary sedimentation effluent: 93.2–106%	
				Ultrafiltration membrane effluent: 76.25–95.28%	
Aptamer-modified SiO ₂ @Au core/shell	Hg ²⁺	1×10^{-8} to 1×10^{-3} M	10 nM	N/A	Lu et al. (2018)
nanoparticles		Linear (log): 1×10^{-7} to 1×10^{-3} M			
NaYF4@ Yb,Ho,Au- nanohybrid	Pb ²⁺	Linear (log): 10 ⁻⁵ –10 ³ µg/ mL	1.16×10^{-9} g/mL and	97.98–106.40	Annavaram et al. (2019)
NaYF4@ Yb,Ho,GO- nanohybrid			1.15 × 10 ⁻⁸ g/ mL	97.76–116.80	
Poly(vinylidene fluoride) membrane decorated with silver nanoparticles	Cd ²⁺	0.2–20 mM	$1 \times 10^{-10} \text{ M}$	N/A	Liu et al. (2019)
L-cysteine- functionalized Au@ Ag core-shell probes	Pb ²⁺	Linear range: 5 pM to 10 nM	1 pM	Drinking water: 96.2–103.5%	Xu et al. (2018)

 Table 2.1
 Selected reports on SERS nanosensors for the detection of heavy metals

(continued)

	Targeted	Reported	Detection	Recovery in	
Nanomaterials	metal	range	limits	samples	Reference
Oligonucleotide- functionalized and	Hg ²⁺	0–10 μΜ	0.1 nM	Lake water: 97.2–103.3%	Zou et al. (2019)
gold coated polystyrene microspheres				Tap water: 101.2–102.2% (Hg) (Pb) for	
	Pb ²⁺	0–100 μΜ	1 nM	Lake water: 94.6–106.3%	
				Tap water: 96.5–104.5%	
Aminobenzo-18- crown-6 (AB18C6)-AuNPs	Pb ²⁺	1 μM to 0.8 mM	0.69 pM	100–110% in the presence of other metal ions	Sarfo et al. (2018)
Dipicolylamine- based ligand plasmonic gold nanoparticles	Cu ²⁺	N/A	$5 \times 10^{-8} \mathrm{M}$	Drinking water: 88.83 ± 9.19%	Dugandžić et al. (2019)

Table 2.1 (continued)



Fig. 2.5 Schematic representation of DNAzyme-Au nanostructure (**a**) and detection of Hg^{2+} and Pb^{2+} with engineered DNA logic gates (**b**). (**c** and **d**) show SERS spectra for Hg^{2+} with 100 nM Pb^{2+} (**c**) and for Pb^{2+} with 100 nM Hg^{2+} (**d**). Reproduced with permission from Zou et al. (2019), Elsevier

Moreover, SERS spectra also showed an excellent correlation between the heavy metal ions and SERS intensity. Good linear correlations were observed for both Hg²⁺ ions and Pb²⁺ ions with $R^2 > 0.99$ and 0.91, respectively. The nanosensor assembly exhibited an ultra-sensitivity and selectivity toward Hg²⁺ ions and Pb²⁺ ions with detection limit of 0.1 nM and 1 nM, respectively, meeting the World Health Organization standard of 10.0 ppb (31 nM) and 10.0 ppm (3.02×10^4 nM), respectively, for drinking water. Noticeably, the proposed AuNC nanosensor also showed excellent recovery in environmental water samples, with 97.2–103.3% (Hg) and 94.6–106.3% (Pb) for lake water samples, and 101.2–102.2% (Hg) and 96.5–104.5% (Pb) for tap water samples.

2.2.5 Surface-Enhanced Fluorescence

In addition to enhanced light absorption and scattering, metal surface enhanced fluorescence has also drawn broad attention. Although fluorescence spectroscopy is ultrasensitive, detecting fluorophore has often suffered from its limited quantum yield and low photostability (Aslan et al. 2005). Over the past decades, the surface plasmon electrons of plasmonic metals have been found capable of interacting with fluorophores and thus enhancing the fluorescence spectral properties such as intensity (Ray et al. 2006; Aslan et al. 2007), photostability (Aslan et al. 2005; Lakowicz 2005), and decay lifetime (Ray et al. 2006; Lu et al. 2012). Although excellent examples have been reported for surface-enhanced fluorescence in many biomedical and environmental studies (Aslan et al. 2005), most of them are focused on biomolecule or organic pollutant detection. Examples are detection of microcystin-LR (Li et al. 2014), bacteria sensing (Abdulhalim et al. 2009), single molecule on cell membrane (Zhang et al. 2008), RNA sensing (Aslan et al. 2006), and cellular imaging (Cui et al. 2010). However, few examples are seen in heavy metal detections.

Here we briefly discuss three successful applications of surface-enhanced fluorescence-based nanosensors to monitor heavy metal ions. Sui et al. (2014) developed DNA-Cy3 aptamer immobilized Ag@SiO₂-DNA-fluorophore nanostructure that is highly sensitive for Hg²⁺ and Ag⁺(Sui et al. 2014), where Cy3 was used as the fluorophore. Hg²⁺ or Ag⁺ could lead to the binding of T–Hg²⁺–T or T–Ag⁺–T in the DNA aptamer, thus placing the fluorophore Cy3 near the metal surface and enhancing the fluorescence of Cy3 at 2.5-fold. The nanosensor assembly showed high selectivity and sensitivity for Hg²⁺ and Ag⁺ with a limit of detection at 1.4 nM. Taking advantage of a similar approach, a glutathione-stabilized gold nanoclusters/indiumbased metal–organic frameworks modified with cysteine (AuNCs/MIL-68(In)-NH₂/ Cys) was developed for the detection of Hg²⁺ ions (Wu et al. 2019), showing remarkably enhanced fluorescence emission. The red fluorescence intensity at 668 nm quenched in the addition of Hg²⁺, making it possible to detect trace Hg²⁺ with an extremely low limit of 6.7 pM. The fluorescent AuNCs/MIL-68(In)-NH₂/Cys



Fig. 2.6 (a) Schematic design of the metal surface enhanced fluorescence-based DNA aptamer-Ag@SiO₂ nanosensor for the detection of Hg²⁺. (b) Florescence spectra of the aptamer-Ag@SiO₂ with Hg²⁺ (0–900 nM). (c) The detection range of Hg²⁺ from 0 nM to 14 nM. (d) Selective detection of Hg²⁺. λ_{ex} = 482 nm, λ_{em} = 538 nm. Reproduced with permission from Panget al. (2015), licensed under a Creative Commons Attribution 4.0 International License

performed well in environmental water samples with a high recovery of 97.5–110.2% in lake water sample and 91.3–107.5% in tap water samples.

Another interesting study from Pang et al. (2015) demonstrated fluorescent coreshell Ag@SiO₂ nanosensors for the detection of Hg²⁺, as the first label-free metal surface enhanced fluorescence-based aptasensor to detect heavy metal pollutants (Pang et al. 2015). The single-strand structure of the T-rich ss-DNA aptamer bound to Ag@SiO₂ NPs was folded to hairpin duplex structure that resulted from the formation of T–Hg²⁺–T. Then the fluorescent reporter thiazole orange could bind the hairpin complex, resulting in higher fluorescence quantum yield (schematically illustrated in Fig. 2.6a). Moreover, the fluorescence intensity of the excited-state thiazole orange was further amplified by the surface-enhanced fluorescence of Ag@ SiO₂ nanosensors, therefore substantially increasing the sensitivity of the aptasensor. As shown in Fig. 2.6b, fluorescence intensity at 538 nm was enhanced by the increasing Hg²⁺ concentration (note the linear correlation window at 0–1.2 nM and 1.2–14 nM in Fig. 2.6c), with a limit at 0.33 nM, meeting the US EPA requirements of 10 nM. The hybrid fluorescence-enhanced $Ag@SiO_2$ nanosensors also showed high selectivity for Hg^{2+} among ten other metal ions (Fig. 2.6d), and great potential for in-situ environmental application.

2.2.6 Fluorescence Quenching–Based Nanoprobes

Besides the surface plasmon resonance enhanced nanosensors, many other properties have also been explored to engineer nanosensors. Fluorescence quenching based nanoprobes are among the noticeable nanosensors. In this case metallic nanostructures act as fluorescence quenchers for fluorophores, often at short distances (~0–5 nm; Aslan et al. 2005). The fluorescence quenching effect can then be quantified to indicate the presence of trace heavy metals. The small changes in the spatial distance between nanostructures and fluorophores can result in significant signal in fluorescence quenching. Heavy metal ions often can mobilize and detach nanoparticles from the hybrid fluorescent nanoprobes via disrupting electrostatic interaction between fluorophores and nanoparticles. The resulting fluorescence quenching is subsequently observed as the indicator for heavy metal concentration. Following this principle, metallic nanostructures have been successfully developed as fluorescent nanoprobes for a wide range of heavy metal ions (Wu et al. 2013; Kaewtong et al. 2014; Hormozi-Nezhad and Taghipour 2015).

Recently, Wang and coworkers engineered a *turn-on* fluorophore brilliant cresyl blue labeled-Au nanoparticle assembly to monitor Pb^{2+} in water samples (Wang et al. 2015). Au nanoparticles act as fluorescence quenchers that diminish brilliant cresyl blue fluorescence via electrostatic interactions. In the presence of Pb^{2+} , a chelating complex is formed between Pb^{2+} and negatively charged glutathione on the Au assembly, leading to the detachment of fluorophore brilliant cresyl blue from the Au nanoparticle assembly, and thus the recovery of fluorescence. The fluorophore labeled-Au nanoparticle assembly showed to be highly sensitive toward Pb^{2+} with a detection limit at 0.51 nM, among 12 other metal ions. Niu et al. (2018) also showed a *turn-on* graphene quantum dots and gold nanoparticle fluorescent sensor for detection of Pb^{2+} (Niu et al. 2018). A similar fluorescence-recovery mechanism was observed for the function of proposed graphene quantum dots and gold nanoparticle fluorescent sensor with Pb^{2+} . The proposed fluorescent nanosensor exhibited a high sensitivity with a wide detection range (50 nM to 4 μ M), and a detection limit at 16.7 nM.

Another example from Wang and coworkers also demonstrated a fluorescent DNA-Ag nanosensor for Pb²⁺ detection (Wang et al. 2019). Similarly, Pb²⁺ ions specifically interact with rA cleavage site of the ds-DNA-AgNC nanosensor (Fig. 2.7a). Thus, the DNA segment was released from the ds-DNA-AgNC assembly, leading to fluorescence spectra shift from red to green (Fig. 2.7b). The strong fluorescence intensity peaks at 560 nm and 685 nm are shown in Fig. 2.7c with a linear correlation between 0.001 nM and 10 nM. As shown in Fig. 2.7d, the



^aSquares and arrows drawn on DNA strands represent 5' termini and 3' termini, respectively.



Fig. 2.7 Schematic illustration of ds-DNA-AgNC nanosensor detecting Pb^{2+} ions (**a**) and the fluorescence spectra response (**b**). The strong fluorescence intensity peaks at 560 nm and 685 nm for Pb^{2+} (0.001–5000 nM) (**c**). (**d**) shows the high selectivity of reported nanosensor toward Pb^{2+} . Reproduced with permission from Wang et al. (2019), Elsevier

ds-DNA-AgNC nanosensor was ultrasensitive and selective to Pb^{2+} , while all the other metal ions did not exhibit strong fluorescence response. Furthermore, the fluorescent signal of Pb^{2+} was not interfered by any of other metal ions, due to the high affinity and selectivity between Pb^{2+} and DNAzyme. The ultrasensitive nanosensor showed an exceptionally low detection limit at 1.0 pM in environmental samples, with recovery of 82–108% in lake water samples and 89–104.4% in tap water samples.

2.2.7 Self-Propelled Nanojets

Self-propelled nanojets (or nanomotors) are relatively new tools with vast promising applications in biomedical researches, with examples seen in drug delivery and medical diagnosis (Sanchez and Pumera 2009; Sánchez and Soler 2015). Those nanojets are autonomously self-propelled nanomachines with relatively quick autonomous motion and sufficient power output (Yang et al. 2015). The commonly studied models of self-propelled nanojets are bubble self-propulsion by enzymatic reactions (e.g., O₂ bubbles generated by catalytic decomposition of H₂O₂ [Solovev et al. 2012]), or bubble-free propulsion powered by an enzyme-controlled catalysis (e.g., internal flows generated by turnover of urea \rightarrow NH_{3(aq)} + CO_{2(aq)} in the presence of enzyme urease [Ma et al. 2016]), as partly illustrated in Fig. 2.8a (Sanchez and



Fig. 2.8 (a) Mechanisms of self-propelled nanojets. Color scheme: Ni (green), Au (yellow), and Pt (blue). (b) Schematic illustration of photonic nanojets detecting a single nanoparticle by forming a potential well with maximum potential difference ΔU . (a) Reproduced with permission from Sánchez and Pumera (2009), John Wiley and Sons. (b) Reproduced with permission from Li et al. (2016), licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International License

Pumera 2009). Alternatively, photonic nanojets supplied by optical power also show great potential for optical operation and detection as nanosensors (Li et al. 2016). The example shown in Fig. 2.8b illustrates a potential well can be generated by the probe-microlens complex, which is able to trap a single nanoparticle or biomolecule, operating at low optical power (Yang et al. 2015; Li et al. 2016; Gu et al. 2018). Recent studies have demonstrated that novel self-propelled nanojets can be applied for rapid detection of pollutants, at the single nanoparticle (Yang et al. 2015; Li et al. 2016; Gu et al. 2018) or single biomolecule (Li et al. 2016) level, in the environment.

As an example to illustrate the application of self-propelled nanojets for the detection of heavy metal contaminants in aqueous samples, Maric and coworkers reported a novel, highly sensitive Pt-Halloysite nanojet to monitor Hg²⁺ (Maric et al. 2019). The proposed Pt-Hallovsite nanojets use Pt to catalyze the decomposition of H_2O_2 , while large Halloysite nanoclay surface allows high adsorption for metal ions (Fig. 2.9a). The positive charges of metal ions can interact with the negatively charged Halloysite nanoclay outer layer by electrostatic interactions. Pt-Halloysite nanojets exhibited high velocities with an average of 227 μ m s⁻¹, and preliminarily in the absence of heavy metal ions. However, the velocities of Pt-Hallovsite nanojets drastically decreased in the presence of Hg²⁺ (Fig. 2.9e), while moderate decrease was observed for other metal ions (Fig. 2.9b-d). The decreased velocity was largely due to the less-exposed active sites for H₂O₂ decomposition of the adsorbed metal ions onto the Pt surface, as shown in Fig. 2.9f, g. The difference observed among the four tested metal ions was possibly due to the difference in metal ion atomic size and electronegative values. Hg²⁺ has the largest atomic radius and relatively higher electronegative values, making it having higher affinity toward Pt surface of the nanojets. The proposed Pt-Halloysite nanojet assembly also indicated a broad detection range for Hg^{2+} (0.25–1000 ppb), with a detection limit at 3.24 ppb.

2.3 Application Examples and Detection Limit

With understanding of fundamental mechanisms that are widely applied in nanosensors for detecting heavy metals, we further show a few examples from recent reports from 2017 to present. We intend not to repeat what we have discussed in the previous section. But we may use the same studies to emphasize the detection mechanism of each specific type of heavy metals. Therefore, applications of nanosensors for six major heavy metal contaminants, including Hg²⁺, Pb²⁺, As³⁺, Cd²⁺, Cu²⁺, and Cr³⁺/Cr⁶⁺ ions, are carefully selected from literature and briefly discussed in this section.



Fig. 2.9 (a) Schematic representation of self-propelled Pt–Halloysite nanojets for the detection of heavy metal ions. (b–e) show velocity changes of Pt–Halloysite nanojets in the presence of (b) Cd^{2+} , (c) Zn^{2+} , (d) Pb^{2+} , and (e) Hg^{2+} . (f) The percentage of active nanojets in different heavy metal ions. (g) Representative images of nanojets with different heavy metal ions at 1 ppm. Reproduced with permission from Maric et al. (2019), John Wiley and Sons

2.3.1 Hg²⁺ Detection

Hg normally presents in elemental (Hg⁰), mercuric (Hg²⁺), or alkylated form (methyl/ethyl mercury) after entering natural environment. In particular, methylmercury is more toxic than inorganic Hg as it tends to be bio-accumulative in living tissue, while many researches focused on detecting inorganic Hg (Amini et al. 2017; Cai et al. 2017; Satapathi et al. 2018). It is therefore important to detect Hg in various forms. Notably, Yang and coworkers developed a fluorescent nanoprobe named folic-acid-capped gold nanoclusters to detect mercury speciation between inorganic (Hg²⁺) and organic (in form of CH₃Hg⁺) forms in water and fish

samples, showing a detection limit of 28 nM and 25 nM, respectively (Yang et al. 2018). The fluorescence quenching effect was observed for the Hg^{2+} , whereas CH_3Hg^+ exhibited strong fluorescence enhancement.

2.3.2 Pb^{2+} Detection

Pb is among the most heavy metals released in environment, with 13.82 million kg released from Europe's industry for the period 2007–2015 (European Environment Agency 2016). Ionic Pb²⁺, PbO, Pb(OH)₂, and lead-metal oxyanion complexes are usually observed in natural environment. Pb poisoning may cause severe damage to the brain, kidney, and central nervous system, posing significant risk to physical and mental development. Detecting of Pb²⁺ is mainly based on the "detaching" mechanism as a result of the interaction between Pb²⁺ ions and nanosensors. For instance, unlike Hg²⁺, the selectivity and sensitivity of Au nanoconjugates toward Pb²⁺ was due to the decreased Raman signal at 1580 cm⁻¹, as a result of mobilization of Au nanoconjugates from the PSMP substrates in the presence of Pb²⁺ (Zou et al. 2019). Similar detaching mechanism was also observed for many studies using fluorescence quenching nanosensors (Wang et al. 2015; Niu et al. 2018; Ikram et al. 2019; Wang et al. 2019). Fluorescence signal can be recovered for fluorophore by adding Pb²⁺, due to nanostructures, as the fluorescence quencher, removing from the fluorophore via chelating interaction between Pb²⁺ and nanostructure assembly.

2.3.3 As³⁺ Detection

Arsenic exists in environment in a variety of forms, including inorganic As^{3+} and organic methylated derivatives, such as $HAs(CH_3)_2$ and $As(CH_3)_3$. As^{3+} -detecting nanosensors often are based on the *turn-on* effect resulting from chemical interaction between the As^{3+} ions and specific regions of nanosensor assembly (Nath et al. 2014; Zhang et al. 2017a). For instance, As^{3+} ions specifically bind $(GT)_{29}$ region of the fluorescent DNA quantum dots (Zhang et al. 2017a). As a result, organized structure evolution and conformational changes were developed for the nanosensor. Subsequently, fluorescence signal was largely enhanced due to the phenomena occurring at the nanostructure surface at which the stacking of nucleic acid–base pairs promotes radiative decay and fluorescence emission.

2.3.4 Cd²⁺ Detection

 Cd^{2+} is believed to cause severe renal damage via chronic accumulation in kidney, with a well-known case of Cd^{2+} poisoning in the Jintsu River Valley, Japan. Severe bone diseases and kidney malfunction can occur as a result of Cd^{2+} poisoning. Cd^{2+} detection can also be achieved through recovering fluorescence signal. For instance, Cd^{2+} ions can break the complexation between the fluorescence quencher 1,10-phenanthroline and green emissive cadmium telluride quantum dots (CdTe QDs), thus turning on the green fluorescence signal (Qian et al. 2017). Similar mechanism was also reported in the L-cysteine-capped CdTe nanosensor. Adding Cd^{2+} can recover the fluorescence signal of CdTe QDs by replacing the fluorescence quencher dithizone (Li et al. 2017).

2.3.5 Cu^{2+} Detection

Copper is the second most released heavy metal from the European industry (Fig. 2.1a). The interaction between Cu²⁺ and environmental media is quite complex but usually in a form that is less toxic than other heavy metals. Nevertheless, it possesses high risks toward major organs such as kidney at high doses. Cu²⁺ detection can be achieved through many strategies and fluorescence is on the top list, followed by surface plasmon resonance and electrochemiluminescence (Wang et al. 2018). Cu²⁺ ions could act as fluorescence quencher (Elmizadeh et al. 2017; Shah et al. 2018), enhance surface plasmon resonance via aggregation (Deymehkar et al. 2018), and act as reaction catalyst in electrochemiluminescence ternary (e.g., PTC-NH₂ + S₂O₈²⁻ + CoFe₂O₄) system to substantially improve the detection sensitivity and limit (Lei et al. 2018).

2.3.6 Cr³⁺ and Cr⁶⁺ Detection

Detection of both Cr^{3+} and Cr^{6+} ions has been reported using a variety of methods. Carbon nanodots and cetyltrimethyl ammonium bromide capped-9,10-diphenyl anthracene nanosensors were used to determine Cr^{6+} concentration based on fluorescence quenching, showing a detection limit of 0.015 μ M (Liu et al. 2017) and 0.01392 μ g mL⁻¹ (Suryawanshi et al. 2019), respectively. Using colorimetric methods, the color change of 4-amino-5-methyl-4H-1,2,4-triazole-3-thiol modified gold nanoparticles (AuNPs), due to the aggregation induced by Cr³⁺, can be used as a quantitative tool for Cr³⁺ detection (Shahrivari et al. 2018). Similarly, Cr³⁺ could induce aggregation of methylene blue capped-AuNPs, which leads to surface plasmon absorption band red-shift signal with detection limit of 23.66 nM (Salimi et al. 2018).

2.4 Outlook

The increasing application of nanotechnology in many fields such as environmental remediation and pollution monitoring also comes with potential cost. The manufacturing, transportation, use, and disposal of nanomaterials may increase the exposure of nanoparticles to the environment and human beings. Yet, we do not have a full understanding of the fate and potential impact of nanomaterials on the environment and human, as the physicochemical properties of nanomaterials may be altered due to their dynamic interactions with surrounding environments. Many evidences have suggested nanomaterials may pose threat to non-target species and human with enhanced reactivity at the nanoscale, while others advocate the benefits of engineered nanomaterials. Questions on how we can design and manage nanoproducts in a sustainable manner remain unanswered at the present. Based on recent trends and progress on reducing adverse effects of nanomaterial production and use, we here provide insights on green synthesis and minimizing nanotoxicity.

2.4.1 Green Synthesis

Green synthesis of nanomaterials has evolved in efforts to reduce the use of toxic raw chemicals and adopt less extreme synthesis conditions (He et al. 2019b). Oftentimes the toxic reducing agent, stabilizing agent, and capping agent are replaced by non-toxic counterparts, particularly from natural products. In searching for sustainable and environment-friendly green synthesis method, biosynthesis has been long studied together with the development of nanomaterials. Biological hosts (e.g., plants and microorganisms) have been found suitable for synthesizing many types of nanomaterials, including Ag, Au, TiO₂, CuO, magnetic iron oxide, and ZnO nanostructures. Biological polymers, including chitosan, proteins, and lipids with functional groups, can serve as reducing agent, stabilizing agent, and capping agent (He et al. 2019b).

Notably, biosynthesized nanoparticles have been successfully used in environmental sensing for heavy metals. Maiti et al. (2016) reported biosynthesized silver nanoparticles for detection of Cu²⁺ ion. Furthermore, 3-mercapto-1,2-propanediol functionalized biosynthesized silver nanoparticles were able to detect Hg²⁺ ions at μ M level (Maiti et al. 2016). Additionally, natural polymers can be directly used as reducing agent, stabilizing agent, and capping agent for green synthesis of nanoparticles without a biological host. Muthivhi et al. (2018) reported that Au, Ag, and Pt nanoparticles could be produced using gelatin and maltose as capping agent and reducing agent, respectively. The biosynthesized noble metal nanoparticles showed high-sensitivity Hg ions in lake water (Muthivhi et al. 2018). However, the exact underlying mechanism for nanomaterial production in biological hosts remains unclear. Biosynthesis of nanomaterials is therefore less productive and controllable. Future research on systematic study of biosynthesis may provide a possible green chemistry solution to the present chemical engineering approach for nanomaterials.

2.4.2 Minimizing Nanotoxicity

Nanotoxicology is an emerging multidisciplinary field in toxicology that focuses on assessing the risks of manufactured nanomaterials to the environment and human health. Nanomaterials are often highly reactive compared to their bulk counterparts because of the large surface-area-to-volume ratio and they are always present as complicate systems (Deng et al. 2018). The enhanced chemistry may cause unintentional toxicity toward environmental species (Dasari and Hwang 2010; Hu and Zhou 2013; He et al. 2014a, 2016b, c; Hu et al. 2014, 2015; Chen et al. 2015; Zhang et al. 2017b; Zhou and Hu 2017; Yuan et al. 2018) and possibly the ecosystem (Hu and Zhou 2013; Ouyang et al. 2018). Thus, it is important to ensure the safe application of novel nanotechnology for commercial use (Hwang et al. 2013; He et al. 2014b, 2015a, 2018a, 2019a). It is imperative to study the complex nature of nano-bio-eco interface with an effort to deliver an accurate comprehension on the toxicological behavior of engineered nanomaterials (He et al. 2015a, 2018).

A comprehensive database is needed, which describes the physicochemical properties of the enormous amounts of nanomaterials being engineered. Then the toxicity can be studied and correlated to those properties in the database (Puzyn et al. 2011; Pathakoti et al. 2014). Traditional in vitro and in vivo toxicological studies on countless nanomaterials being generated everyday are often resourcelimited and time-consuming. Emerging omics studies, including genomics, transcriptomics, proteomics, and metabolomics, are powerful tools to reveal the potential metabolic pathways using model biological species. Lastly, recent trend on machine learning and data mining has revealed many previously unknown patterns hidden in the data collected. The comprehensive database can be useful to explore hidden patterns using data mining and predict new features using machine learning. One recent example is from Ban et al. (2018) who reported random forestbased machine learning approach to determine the priority factors underlying the reproductive toxicity of 18 nanomaterials. They found that Fe₃O₄, Fe₂O₃, and ZnO nanoparticles tend to accumulate in living organism at a much higher level than other nanoparticles, but induce less reproductive toxicity. Such finding somewhat contrasts to the intuitive as higher dose may cause severer damage. Their model also predicts that Ag nanoparticles have a weak dependency on dosage over a wide range (1-100 mg/kg), which is useful to assess the toxicity of Ag nanoparticles. Nevertheless, data heterogeneity and uncertainty is a challenge for the current machine learning algorithm to accurately represent the reality (Ban et al. 2018). Novel computational algorithms may aid in analyzing dataset with high diversities, uncertainties, and incompleteness of different degree, and discovering new features and patterns (Valdés 2018).

Acknowledgment H.M.H acknowledges the financial support from the National Science Foundation-Centers of Research Excellence in Science and Technology (NSF-CREST) program, with grant #HRD-157754 to Jackson State University.

References

- Abdulhalim I, Karabchevsky A, Patzig C, Rauschenbach B, Fuhrmann B, Eltzov E, Marks R, Xu J, Zhang F, Lakhtakia A (2009) Surface-enhanced fluorescence from metal sculptured thin films with application to biosensing in water. Appl Phys Lett 94:063106
- Amini MH, Faridbod F, Ganjali MR, Norouzi P (2017) Functionalized graphene quantum dots as a fluorescent "off–on" nanosensor for detection of mercury and ethyl xanthate. Res Chem Intermediat 43:7457–7470
- Amirjani A, Haghshenas DF (2018) Ag nanostructures as the surface plasmon resonance (SPR)based sensors: a mechanistic study with an emphasis on heavy metallic ions detection. Sens Actuat B-Chem 273:1768–1779
- Annavaram V, Kutsanedzie YHF, Agyekum A, Shah SA, Zareef M, Hassan MM, Waqas A, Ouyang Q, Chen Q (2019) NaYF4@ Yb, Ho, Au/GO-nanohybrid materials for SERS applications—Pb (II) detection and prediction. Sens Actuat B-Chem 174:598–606
- Aslan K, Gryczynski I, Malicka J, Matveeva E, Lakowicz J, Geddes C (2005) Metal-enhanced fluorescence: an emerging tool in biotechnology. Curr Opin Biotechnol 16:55–62
- Aslan K, Huang J, Wilson GM, Geddes CD (2006) Metal-enhanced fluorescence-based RNA sensing. J Am Chem Soc 128:4206–4207
- Aslan K, Wu M, Lakowicz JR, Geddes CD (2007) Fluorescent core– shell Ag@ SiO₂ nanocomposites for metal-enhanced fluorescence and single nanoparticle sensing platforms. J Am Chem Soc 129:1524–1525
- Ban Z, Zhou Q, Sun A, Mu L, Hu X (2018) Screening priority factors determining and predicting the reproductive toxicity of various nanoparticles. Environ Sci Technol 52:9666–9676
- Baruah S, Dutta J (2009) Nanotechnology applications in pollution sensing and degradation in agriculture: a review. Environ Chem Lett 7:191–204
- Cai K, Zeng M, Liu F, Liu N, Huang Z, Song Y, Wang L (2017) BSA–AuNPs@Tb–AMP metal– organic frameworks for ratiometric fluorescence detection of DPA and Hg²⁺. Luminescence 32:1277–1282
- Chen Y, Ren C, Ouyang S, Hu X, Zhou Q (2015) Mitigation in multiple effects of graphene oxide toxicity in zebrafish embryogenesis driven by humic acid. Environ Sci Technol 49:10147–10154
- Cui X, Tawa K, Hori H, Nishii J (2010) Tailored plasmonic gratings for enhanced fluorescence detection and microscopic imaging. Adv Funct Mater 20:546–553
- Dasari TP, Hwang H-M (2010) The effect of humic acids on the cytotoxicity of silver nanoparticles to a natural aquatic bacterial assemblage. Sci Total Environ 408:5817–5823
- De Moreno JEA, Gerpe MS, Moreno VJ, Vodopivez C (1997) Heavy metals in Antarctic organisms. Polar Biol 17:131–140
- Deng H, Zhang Y, Yu H (2018) Nanoparticles considered as mixtures for toxicological research. J Environ Sci Health C 36:1–20
- Deymehkar E, Taher MA, Karami C, Arman A (2018) Synthesis of SPR nanosensor using gold nanoparticles and its application to copper (II) determination. SILICON 10:1329–1336
- Ding SY, You EM, Tian ZQ, Moskovits M (2017) Electromagnetic theories of surface-enhanced Raman spectroscopy. Chem Soc Rev 46:4042–4076
- Dugandžić V, Kupfer S, Jahn M, Henkel T, Weber K, Cialla-May D, Popp J (2019) A SERS-based molecular sensor for selective detection and quantification of copper (II) ions. Sens Actuat B-Chem 279:230–237

- Elmizadeh H, Soleimani M, Faridbod F, Bardajee GR (2017) Ligand-capped CdTe quantum dots as a fluorescent nanosensor for detection of copper ions in environmental water sample. J Fluoresc 27:2323–2333
- Enders D, Nagao T, Pucci A, Nakayama T (2006) Reversible adsorption of Au nanoparticles on SiO₂/Si: an in situ ATR-IR study. Surf Sci 600:L71–L75
- European Environment Agency (2016) The European Pollutant Release and Transfer Register (E-PRTR), Member States reporting under Article 7 of Regulation (EC) No 166/2006
- Franklin AD (2015) Nanomaterials in transistors: from high-performance to thin-film applications. Science 349: aab2750
- Frost MS, Dempsey MJ, Whitehead DE (2015) Highly sensitive SERS detection of Pb²⁺ ions in aqueous media using citrate functionalised gold nanoparticles. Sens Actuat B-Chem 221:1003–1008
- Gray JS, Bewers JM, He X (2019) Pollution: approaches to pollution control. In: Cochran JK, Bokuniewicz JH, Yager LP (eds) Encyclopedia of ocean sciences, 3rd edn. Elsevier, Oxford, pp 231–235. https://doi.org/10.1016/B978-0-12-409548-9.11286-2
- Gu G, Song J, Chen M, Peng X, Liang H, Qu J (2018) Single nanoparticle detection using a photonic nanojet. Nanoscale 10:14182–14189
- He X, Hwang H-M (2016) Nanotechnology in food science: functionality, applicability, and safety assessment. J Food Drug Anal 24:671–681
- He X, Aker WG, Hwang H-M (2014a) An *in vivo* study on the photo-enhanced toxicities of S-doped TiO₂ nanoparticles to zebrafish embryos (*Danio rerio*) in terms of malformation, mortality, rheotaxis dysfunction, and DNA damage. Nanotoxicology 8:185–195
- He X, Aker WG, Leszczynski J, Hwang H-M (2014b) Using a holistic approach to assess the impact of engineered nanomaterials inducing toxicity in aquatic systems. J Food Drug Anal 22:128–146
- He X, Aker WG, Fu PP, Hwang H-M (2015a) Toxicity of engineered metal oxide nanomaterials mediated by nano-bio-eco-interactions: a review and perspective. Environ Sci Nano 2:564–582
- He X, Aker WG, Huang M-J, Watts JD, Hwang H-M (2015b) Metal oxide nanomaterials in nanomedicine: applications in photodynamic therapy and potential toxicity. Curr Top Med Chem 15:1887–1900
- He X, Aker WG, Pelaez M, Lin Y, Dionysiou DD, H-m H (2016a) Assessment of nitrogen–fluorinecodoped TiO₂ under visible light for degradation of BPA: implication for field remediation. J Photochem Photobiol A 314:81–92
- He X, McAlliser D, Aker WG, Hwang H-M (2016b) Assessing the effect of different natural dissolved organic matters on the cytotoxicity of titanium dioxide nanoparticles with bacteria. J Environ Sci 48:230–236
- He X, Sanders S, Aker WG, Lin Y, Douglas J, Hwang H-M (2016c) Assessing the effects of surface-bound humic acid on the phototoxicity of anatase and rutile TiO₂ nanoparticles *in vitro*. J Environ Sci 42:50–60
- He X, Fu P, Aker WG, Hwang H-M (2018) Toxicity of engineered nanomaterials mediated by nano-bio-eco interactions. J Environ Sci Health C Environ Carcinog Ecotoxicol Rev 36:21–42
- He X, Deng H, Aker WG, H-m H (2019a) Regulation and safety of nanotechnology in the food and agriculture industry. In: Molina G, Inamuddin, Pelissari FM et al (eds) Food applications of nanotechnology. CRC Press, Boca Raton, pp 525–536
- He X, Deng H, Hwang HM (2019b) The current application of nanotechnology in food and agriculture. J Food Drug Anal 27:1–21
- Hoang CV, Oyama M, Saito O, Aono M, Nagao T (2013) Monitoring the presence of ionic mercury in environmental water by plasmon-enhanced infrared spectroscopy. Sci Rep 3(1):1175
- Hong S, Candelone J-P, Patterson CC, Boutron CF (1996) History of ancient copper smelting pollution during Roman and medieval times recorded in Greenland ice. Science 272:246–249
- Hormozi-Nezhad MR, Taghipour M (2015) An ultrasensitive and selective turn-off fluorescent nanoprobe for the detection of copper ions. Anal Methods 7:5067–5073

Hu XG, Zhou QX (2013) Health and ecosystem risks of graphene. Chem Rev 113:3815-3835

- Hu X, Kang J, Lu K, Zhou R, Mu L, Zhou Q (2014) Graphene oxide amplifies the phytotoxicity of arsenic in wheat. Sci Rep 4:6122
- Hu XG, Zhou M, Zhou QX (2015) Ambient water and visible-light irradiation drive changes in graphene morphology, structure, surface chemistry, aggregation and toxicity. Environ Sci Technol 49:3410–3418
- Hwang H-M, Ray PC, Yu H, He X (2013) Toxicology of designer/engineered metallic nanoparticles. In: Luque R, Varma R (eds) Sustainable preparation of metal nanoparticles: methods and applications. Royal Society of Chemistry, Cambridge, pp 190–212
- Ikram F, Qayoom A, Aslam Z, Shah MR (2019) Epicatechin coated silver nanoparticles as highly selective nanosensor for the detection of Pb²⁺ in environmental samples. J Mol Liq 277:649–655
- Kaewtong C, Niamsa N, Wanno B, Morakot N, Pulpoka B, Tuntulani T (2014) Optical chemosensors for Hg²⁺ from terthiophene appended rhodamine derivatives: FRET based molecular and in situ hybrid gold nanoparticle sensors. New J Chem 38:3831–3839
- Kelly KL, Coronado E, Zhao LL, Schatz GC (2003) The optical properties of metal nanoparticles: the influence of size, shape, and dielectric environment. J Phys Chem B 107:668–677
- Khan AH, Ghosh S, Pradhan B, Dalui A, Shrestha LK, Acharya S, Ariga K (2017) Two-dimensional (2D) nanomaterials towards electrochemical nanoarchitectonics in energy-related applications. Bull Chem Soc Jpn 90:627–648
- Kumar P, Kim K-H, Bansal V, Lazarides T, Kumar N (2017) Progress in the sensing techniques for heavy metal ions using nanomaterials. J Ind Eng Chem 54:30–43
- Lakowicz J (2005) Radiative decay engineering 5: metal-enhanced fluorescence and plasmon emission. Anal Biochem 337:171–194
- Lei YM, Xiao BQ, Liang WB, Chai YQ, Yuan R, Zhuo Y (2018) A robust, magnetic, and selfaccelerated electrochemiluminescent nanosensor for ultrasensitive detection of copper ion. Biosens Bioelectron 109:109–115
- Li H, Qian X, Wang QG (2013a) Heavy metals in atmospheric particulate matter: a comprehensive understanding is needed for monitoring and risk mitigation. Environ Sci Technol 47:13210–13211
- Li M, Gou H, Al-Ogaidi I, Wu N (2013b) Nanostructured sensors for detection of heavy metals: a review. ACS Sustain Chem Eng 1:713–723
- Li Y, Sun J, Wu L, Ji J, Sun X, Qian Y (2014) Surface-enhanced fluorescence immunosensor using Au nano-crosses for the detection of microcystin-LR. Biosens Bioelectron 62:255–260
- Li S, Xu L, Ma W, Kuang H, Wang L, Xu C (2015) Triple Raman label-encoded gold nanoparticle trimers for simultaneous heavy metal ion detection. Small 11:3435–3439
- Li YC, Xin HB, Lei HX, Liu LL, Li YZ, Zhang Y, Li BJ (2016) Manipulation and detection of single nanoparticles and biomolecules by a photonic nanojet. Light-Sci Appl 5:e16176
- Li L, Liao L, Ding Y, Zeng H (2017) Dithizone-etched CdTe nanoparticles-based fluorescence sensor for the off-on detection of cadmium ion in aqueous media. RSC Adv 7:10361–10368
- Liu X, Li T, Wu Q, Yan X, Wu C, Chen X, Zhang G (2017) Carbon nanodots as a fluorescence sensor for rapid and sensitive detection of Cr(VI) and their multifunctional applications. Talanta 165:216–222
- Liu R, Tang J, Yang H, Jin W, Liu M, Liu S, Hu J (2019) In situ decoration of plasmonic silver nanoparticles on Poly (vinylidene fluoride) membrane for versatile SERS detection. New J Chem 43:6965–6972
- Lu G, Li W, Zhang T, Yue S, Liu J, Hou L, Li Z, Gong Q (2012) Plasmonic-enhanced molecular fluorescence within isolated bowtie nano-apertures. ACS Nano 6:1438–1448
- Lu Y, Zhong J, Yao G, Huang Q (2018) A label-free SERS approach to quantitative and selective detection of mercury (II) based on DNA aptamer-modified SiO₂@ Au core/shell nanoparticles. Sens Actuat B-Chem 258:365–372
- Ma S, Zhan S, Jia Y, Zhou Q (2015) Highly efficient antibacterial and Pb (II) removal effects of Ag-CoFe₂O₄-GO nanocomposite. ACS Appl Mater Interfaces 7:10576–10586

- Ma X, Hortelao AC, Miguel-López A, Sánchez S (2016) Bubble-free propulsion of ultrasmall tubular nanojets powered by biocatalytic reactions. J Am Chem Soc 138:13782–13785
- Maiti S, Barman G, Laha JK (2016) Detection of heavy metals (Cu²⁺, Hg²⁺) by biosynthesized silver nanoparticles. Appl Nanosci 6:529–538
- Maric T, Mayorga-Martinez CC, Nasir MZM, Pumera M (2019) Platinum-halloysite nanoclay nanojets as sensitive and selective mobile nanosensors for mercury detection. Adv Mater Technol 4:1800502
- McConnell JR, Edwards R (2008) Coal burning leaves toxic heavy metal legacy in the Arctic. PNAS 105:12140–12144
- McIntyre A, He X (2019) Global marine pollution-a brief history. In: Cochran JK, Bokuniewicz JH, Yager LP (eds) Encyclopedia of ocean sciences, 3rd edn. Elsevier, Oxford, pp 231–235. https://doi.org/10.1016/B978-0-12-409548-9.11429-0
- Millward GE, Turner A, He X (2019) Metal pollution. In: Cochran JK, Bokuniewicz JH, Yager LP (eds) Encyclopedia of ocean sciences, 3rd edn. Elsevier, Oxford, pp 342–349. https://doi. org/10.1016/B978-0-12-409548-9.11287-4
- Mohan SV, Nithila P, Reddy SJ (1996) Estimation of heavy metals in drinking water and development of heavy metal pollution index. J Environ Sci Health A 31:283–289
- Mordor Intelligence (2019) Nanosensors market segmented by type (physical nanosensor, chemical nanosensor, biological nanosensor), industry (healthcare, consumer electronics, automotive and industrial, aerospace and defense, power generation), and region – growth, trends, and forecast (2019–2024)
- Muthivhi R, Parani S, May B, Oluwafemi OS (2018) Green synthesis of gelatin-noble metal polymer nanocomposites for sensing of Hg²⁺ ions in aqueous media. Nano-Struct Nano-Objects 13:132–138
- Nath P, Arun RK, Chanda N (2014) A paper based microfluidic device for the detection of arsenic using a gold nanosensor. RSC Adv 4:59558–59561
- Niu X, Zhong Y, Chen R, Wang F, Liu Y, Luo D (2018) A "turn-on" fluorescence sensor for Pb²⁺ detection based on graphene quantum dots and gold nanoparticles. Sens Actuat B-Chem 255:1577–1581
- Nriagu JO (1996) A history of global metal pollution. Science 272:223-223
- Ouyang S, Hu X, Zhou Q, Li X, Miao X, Zhou R (2018) Nanocolloids in natural water: isolation, characterization, and toxicity. Environ Sci Technol 52:4850–4860
- Pang Y, Rong Z, Xiao R, Wang S (2015) "Turn on" and label-free core- shell Ag@ SiO₂ nanoparticles-based metal-enhanced fluorescent (MEF) aptasensor for Hg²⁺. Sci Rep 5:9451
- Pathakoti K, Huang MJ, Watts JD, He X, Hwang HM (2014) Using experimental data of Escherichia coli to develop a QSAR model for predicting the photo-induced cytotoxicity of metal oxide nanoparticles. J Photochem Photobiol B 130:234–240
- Puzyn T, Rasulev B, Gajewicz A, Hu X, Dasari TP, Michalkova A, Hwang HM, Toropov A, Leszczynska D, Leszczynski J (2011) Using nano-QSAR to predict the cytotoxicity of metal oxide nanoparticles. Nat Nanotechnol 6:175–178
- Qian J, Wang K, Wang C, Ren C, Liu Q, Hao N, Wang K (2017) Ratiometric fluorescence nanosensor for selective and visual detection of cadmium ions using quencher displacement-induced fluorescence recovery of CdTe quantum dots-based hybrid probe. Sensors Actuators B Chem 241:1153–1160
- Ray K, Badugu R, Lakowicz JR (2006) Metal-enhanced fluorescence from CdTe nanocrystals: a single-molecule fluorescence study. J Am Chem Soc 128:8998–8999
- Salimi F, Kiani M, Karami C, Taher MA (2018) Colorimetric sensor of detection of Cr (III) and Fe (II) ions in aqueous solutions using gold nanoparticles modified with methylene blue. Optik 158:813–825
- Sanchez S, Pumera M (2009) Nanorobots: the ultimate wireless self-propelled sensing and actuating devices. Chem Asian J 4:1402–1410
- Sánchez S, Soler L (2015) Chemically powered micro-and nanomotors. Angew Chem Int Ed 54:1414–1444

- Sarfo DK, Izake EL, O'Mullane AP, Ayoko GA (2018) Molecular recognition and detection of pb (II) ions in water by aminobenzo-18-crown-6 immobilised onto a nanostructured SERS substrate. Sens Actuat B-Chem 255:1945–1952
- Satapathi S, Kumar V, Chini MK, Bera R, Halder KK, Patra A (2018) Highly sensitive detection and removal of mercury ion using a multimodal nanosensor. Nano-Struct Nano-Objects 16:120–126
- Shaban M, Galaly AR (2016) Highly sensitive and selective in-situ SERS detection of Pb²⁺, Hg²⁺, and Cd²⁺ using nanoporous membrane functionalized with CNTs. Sci Rep 6:25307
- Shah MT, Alveroglu E, Balouch A (2018) Pyranine functionalized Fe₃O₄ nanoparticles for the sensitive fluorescence detection of Cu2+ ions. J Alloys Compd 767:151–162
- Shahrivari S, Faridbod F, Ganjali MR (2018) Highly selective and sensitive colorimetric determination of Cr3 + ion by 4-amino-5-methyl-4H-1,2,4-triazole-3-thiol functionalized Au nanoparticles. Spectrochimica Acta 191:189–194
- Solovev AA, Xi W, Gracias DH, Harazim SM, Deneke C, Sanchez S, Schmidt OG (2012) Selfpropelled nanotools. ACS Nano 6:1751–1756
- Song D, Yang R, Wang H, Li W, Wang H, Long H, Long F (2017) A label-free SERRS-based nanosensor for ultrasensitive detection of mercury ions in drinking water and wastewater effluent. Anal Methods 9:154–162
- Su S, Wu W, Gao J, Lu J, Fan C (2012) Nanomaterials-based sensors for applications in environmental monitoring. J Mater Chem 22:18101–18110
- Sui N, Wang L, Yan T, Liu F, Sui J, Jiang Y, Wan J, Liu M, William WY (2014) Selective and sensitive biosensors based on metal-enhanced fluorescence. Sens Actuat B-Chem 202:1148–1153
- Suryawanshi SB, Mahajan PG, Kolekar GB, Bodake AJ, Patil SR (2019) Selective recognition of Cr (VI) ion as $Cr_2O_7^{2-}$ in aqueous medium using CTAB-capped anthracene-based nanosensor: application to real water sample analysis. J Phys Org Chem 32:e3923
- Valdés JJ (2018) Extreme learning machines with heterogeneous data types. Neurocomputing 277:38–52
- Vikesland PJ (2018) Nanosensors for water quality monitoring. Nat Nanotechnol 13:651
- Wang S, Sun J, Gao F (2015) A turn-on near-infrared fluorescent chemosensor for selective detection of lead ions based on a fluorophore–gold nanoparticle assembly. Analyst 140:4001–4006
- Wang C, Yang F, Tang Y, Yang W, Zhong H, Yu C, Li R, Zhou H, Li Y, Mao L (2018) Graphene quantum dots nanosensor derived from 3D nanomesh graphene frameworks and its application for fluorescent sensing of Cu 2+ in rat brain. Sens Actuat B-Chem 258:672–681
- Wang J, Zhang Z, Gao X, Lin X, Liu Y, Wang S (2019) A single fluorophore ratiometric nanosensor based on dual-emission DNA-templated silver nanoclusters for ultrasensitive and selective Pb²⁺ detection. Sens Actuat B-Chem 282:712–718
- Wu YS, Huang FF, Lin YW (2013) Fluorescent detection of lead in environmental water and urine samples using enzyme mimics of catechin-synthesized Au nanoparticles. ACS Appl Mater Interfaces 5:1503–1509
- Wu XJ, Kong F, Zhao CQ, Ding SN (2019) Ratiometric fluorescent nanosensors for ultra-sensitive detection of mercury ions based on AuNCs/MOFs. Analyst 8:2523–2530
- Wuana RA, Okieimen FE (2011) Heavy metals in contaminated soils: a review of sources, chemistry, risks and best available strategies for remediation. ISRN Ecology 2011:402647
- Xu Z, Zhang L, Mei B, Tu J, Wang R, Chen M, Cheng Y (2018) A rapid surface-enhanced Raman scattering (SERS) method for Pb²⁺ detection using L-cysteine-modified ag-coated au nanoparticles with core–shell nanostructure. Coatings 8:394
- Yang H, Cornaglia M, Gijs MAM (2015) Photonic nanojet array for fast detection of single nanoparticles in a flow. Nano Lett 15:1730–1735
- Yang JY, Yang T, Wang XY, Chen ML, Yu YL, Wang JH (2018) Mercury speciation with fluorescent gold nanocluster as a probe. Anal Chem 90:6945–6951
- Yuan P, Zhou Q, Hu X (2018) The phases of WS_2 nanosheets influence uptake, oxidative stress, lipid peroxidation, membrane damage, and metabolism in algae. Environ Sci Technol 52:13543–13552

- Zhang J, Fu Y, Liang D, Zhao RY, Lakowicz JR (2008) Fluorescent avidin-bound silver particle: a strategy for single target molecule detection on a cell membrane. Anal Chem 81:883–889
- Zhang L, Cheng XZ, Kuang L, Xu AZ, Liang RP, Qiu JD (2017a) Simple and highly selective detection of arsenite based on the assembly-induced fluorescence enhancement of DNA quantum dots. Biosens Bioelectron 94:701–706
- Zhang X, Zhou Q, Zou W, Hu X (2017b) Molecular mechanisms of developmental toxicity induced by graphene oxide at predicted environmental concentrations. Environ Sci Technol 51:7861–7871
- Zhang Z, Zhang Z, Liu H, Mao X, Liu W, Zhang S, Nie Z, Lu X (2018) Ultratrace and robust visual sensor of Cd²⁺ ions based on the size-dependent optical properties of Au@ g-CNQDs nanoparticles in mice models. Biosens Bioelectron 103:87–93
- Zhou QX, Hu XG (2017) Systemic stress and recovery patterns of rice roots in response to graphene oxide nanosheets. Environ Sci Technol 51:2022–2030
- Zou Q, Li X, Xue T, Zheng J, Su Q (2019) SERS detection of mercury (II)/lead (II): a new class of DNA logic gates. Talanta 195:497–505

Chapter 3 AIE-Based Fluorescent Nanosensors for Detection of Heavy Metal Ions



Xueyan Huang, Bin Feng, Tang Gao, Anyao Bi, Liu Huang, Rong Song, Meihui Liu, and Wenbin Zeng

Contents

3.1	Introduction			
3.2 AIE-Based Nanosensors for the Detection of Hg ²⁺				
	3.2.1 Organic Molecule Nanoparticles	56		
	3.2.2 Metallic Nanoparticles/Clusters	62		
	3.2.3 Carbon Dots	64		
3.3	AIE-Based Nanosensors for the Detection of Cu ²⁺	65		
	3.3.1 Organic Molecule Nanoparticles	65		
	3.3.2 Metal-Organic Frameworks	71		
	3.3.3 Hydrogen-Bonded Organic Frameworks	72		
	3.3.4 Metallic Nanoparticles/Clusters.	72		
	3.3.5 Organic Dots	73		
3.4	AIE-Based Nanosensors for the Detection of Zn ²⁺	75		
	3.4.1 Organic Molecule Nanoparticles	75		
	3.4.2 Metal-Organic Frameworks	76		
	3.4.3 Metallic Nanoparticles/Clusters	76		
3.5	AIE-Based Nanosensors for the Detection of Pb ²⁺	78		
	3.5.1 Metallic Nanoparticles/Clusters	78		
	3.5.2 Carbon Dots	78		
3.6	AIE-Based Nanosensors for the Detection of Cr ³⁺ and Cr ⁶⁺	79		
3.7	AIE-Based Nanosensors for the Detection of Cd ²⁺	82		
3.8	AIE-Based Nanosensors for the Detection of As ³⁺ and As ⁵⁺	83		
3.9	AIE-Based Nanosensors for the Detection of Multiple Heavy Metals			
	3.9.1 Organic Molecule Nanoparticles	83		
	3.9.2 Metallic Nanoparticles/Clusters	86		
3.10	Conclusions and Future Perspectives	88		
Refer	rences	89		

X. Huang \cdot B. Feng \cdot T. Gao \cdot A. Bi \cdot L. Huang \cdot R. Song \cdot W. Zeng (\boxtimes)

Xiangya School of Pharmaceutical Sciences, Central South University, Changsha, PR China

Molecular Imaging Research Center, Central South University, Changsha, PR China

M. Liu

Hunan Huacheng Biotech Inc. and Jiangxi Haifu Biotech Inc., Changsha, PR China

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_3

Abstract Heavy metal pollution is a serious global environmental problem that causes irreversible damage to the ecological environment and harms the human health. It is clearly imminent to develop an efficient and simple method for detecting heavy metals in the environment. Aggregation-induced emission (AIE)-based nanomaterials are excellent as a sensing platform, which enables rapid detection of heavy metals with high sensitivity and selectivity in aqueous solution, overcoming the drawbacks of traditional sensors such as poor solubility in water, aggregationcaused quenching, and insufficient signal-to-noise ratio. Herein, we summarize the recent efforts to develop AIE-based nanosensors for detecting heavy metal ions (including mercury, lead, zinc, copper, cadmium, chromium, and arsenic) with a focus on the sensing materials and mechanisms. Organic nanoparticles, metallic nanoclusters or nanoparticles, metal-organic frameworks, hydrogen-bonded organic frameworks, carbon dots, and quantum dots have been applied in the construction of AIE-based nanosensors to achieve the trace analysis of heavy metal ions, acting more in a fluorescence enhancement or quenching behavior. Moreover, some ratiometric nanosensors based on coupling two or more nanomaterials have shown some superiorities in selectivity, sensitivity, and accuracy. Despite the report that AIEbased nanosensors exhibit sufficient sensitivity to detect heavy metal ions in aqueous environment, the insufficient selectivity and specificity affect the capability of specially sensing in complex matrices.

Keywords Nanosensor · AIE · Heavy metal · Detection · Fluorescence

3.1 Introduction

Heavy metal contamination is a major global environmental issue (Bashir et al. 2019). Owing to the non-biodegradability, long half-life, and the ability to accumulate in the body, heavy metals (mercury, lead, copper, cadmium, chromium, zinc, arsenic, etc.) often cause severe damage to the organs, tissues, bones, and nervous systems once they enter the human body (Jarup 2003; Wu et al. 2010). For instance, mercury, lead, and arsenic can cause schizophrenia and neurological disorders; copper, cadmium, mercury, and lead can cause kidney and liver failure; copper, lead, and cadmium can cause dermatosis; and so on (Aragay et al. 2011; Cui et al. 2015). Even in the presence of merely trace amounts of heavy metals in the environment, the human would be exposed to high levels of heavy metals ultimately due to their permanent retention in the ecosystem (Izah et al. 2016). In view of this, it is urgent to develop effective method for detecting heavy metal ions with high sensitivity, high selectivity, and simplicity in practical application (Soodan et al. 2014; Kumar et al. 2017). Fluorescent analysis provides an effective platform for heavy metal detection, which possesses unique physical and chemical properties, high specificity for heavy metal ions, cost-effectiveness, and portability, to break through the limitations of traditional methods such as the demand for expensive instruments and skilled professionals, long detection time, etc. (Zhang and Fang 2010; Ullah et al. 2018; De Acha et al. 2019). However, the traditional fluorophores suffer from aggregation-caused quenching (ACQ) in aqueous solution, which severely restricts the practical application in aqueous solution (Gao and Tang 2017). Fortunately, the concept of aggregation-induced emission (AIE) first proposed by Tang et al. can effectively alleviate the pressing problem (Luo et al. 2001; Hong et al. 2009). In the AIE process, structural rigidification in aggregation state blocks the non-radiative decay, thereby causing the excited electrons to return to the ground state mainly by radiation transition (Mei et al. 2014). The AIE material is capable of maintaining strong fluorescent emission in aggregated state, which makes it possible the assembly of those materials into nanoparticles (NPs) to construct the AIE-based nanosensors for metal detection (Zhao et al. 2019; Liu et al. 2019a, b). Actually, those nanosensors hold great benefits for heavy metals' detection: strong fluorescence emission in aqueous solution, anti-photobleaching, high sensitivity, low background interference, etc. (Zhang et al. 2015a, b; Huang et al. 2019). These fluorescent nanosensors are able to quantify heavy metal ions by analyte-induced fluorescence quenching or enhancement (Wang et al. 2018a, b). Furthermore, the aggregation process can be actively utilized to achieve the collective effect, which is definitely favorable to trace analysis (Gu et al. 2018; Yang et al. 2019a, b). This review will outline recent advances in nanosensors with AIE property in the detection of heavy metal ions, with a focus on Hg²⁺, Pb²⁺, Zn²⁺, Cu²⁺, Cr³⁺, Cd²⁺, and As³⁺, and pay special attention to the sensing materials and related mechanisms (Scheme 3.1).



Scheme 3.1 Schematics depicting the AIE-based nanosensors for heavy metals' detection. *MOFs* Metal-organic frameworks, *HOFs* Hydrogen-bonded organic frameworks, *NPs/NCs* nanoparticles/ clusters

3.2 AIE-Based Nanosensors for the Detection of Hg²⁺

Many different types of AIE nanosensors for Hg²⁺ detection have been reported, generally based on the following strategies: (1) complexation with ligands such as Schiff base, sulfhydryl, thymine-like group, sulfonamide group, aldoxime, and other sulfur atom-containing, oxygen atom-containing, or nitrogen atom-containing compounds; (2) utilizing specific reactions such as Hg²⁺-induced cleavage of the vinyl group, desulfurization reaction, dethioacetalization, ring-opening reaction of spirolactam, etc.; and (3) utilizing the ability to quench the metallic nanosensors' emission through metallophilic interactions. In this section, these nanosensors are reviewed according to the classification of materials in terms of organic molecule nanoparticles, supramolecular system, nanoparticles (NPs), nanoclusters (NCs), and carbon dots (CDs).

3.2.1 Organic Molecule Nanoparticles

Organic molecule nanoparticles refer to nanoparticles formed by aggregation of single organic molecules, multiple different molecules, or polymers. They usually involve typical fluorophores such as cyanostilbene derivative, rhodamine, rhodol, and the classical AIE skeleton tetraphenylethylene (TPE). Some fluorophores themselves can be used as ligands for complexing with Hg²⁺ while some need to incorporate specific recognition groups.

Wei et al. (2018) designed a TPE-based aggregation-induced enhanced emission (AIEE) nanosensor 1 for sensing Hg²⁺; it was composed of molecular polymer and introduced thiazole as the recognition unit through Pd-catalyzed Sonogashira coupling reaction. Similar to AIE effect, AIEE effect means stronger fluorescence in aggregation state compared with that in solution state. Since Hg²⁺ has a high affinity for sulfhydryl group, the reduction of the π -conjugation system of 1 was induced by the coordination with thiazole upon the addition of Hg²⁺. Consequently, the original bright yellow fluorescence of 1 was observed to gradually quench in tetrahydrofuran (THF) and water mixture (water fraction 90%). However, this "turn-off" behavior was nonlinear due to the failure to guarantee uniform distribution of coordination between Hg²⁺ and any polymer chain.

Schiff base has the advantage of simple synthetic procedure, and the nitrogen atoms of which can efficiently coordinate with Hg²⁺ (Bhalla et al. 2013; Ni et al. 2013). Wang et al. (2018a, b) prepared two TPE-based fluorescent nanosensors **2-1** and **2-2**, which formed by aggregation of small molecule functionalized with Schiff base (Fig. 3.1). The AIE effects of both sensors were investigated in THF-H₂O solution. The fluorescence intensity of **2-2** was stronger due to its possession of alkyl chain and longer conjugation. With the addition of Hg²⁺, the complexation of the diaminomaleonitrile moieties with Hg²⁺ resulted in the decrease in electron-withdrawing effect, which further inhibited the intramolecular charge transfer (ICT)



Fig. 3.1 Monomer chemical structures of AIE-based nanosensors 1-5 for Hg²⁺ detection

from TPE to the diaminomaleonitrile unit. In consequence, fluorescence intensity decreased gradually thereby achieving the quantification of Hg^{2+} . Further the detection limit of **2-2** was as low as 9.84 nM. These two sensors were shown to be cell permeable, leading to the success of detection of Hg^{2+} in living cells.

In another separate work, Wang et al. (2019a, b) synthesized seven different substituted Schiff base-based fluorescent sensors from **3-1** to **3-7** with AIE properties via a simple one-step reaction (Fig. 3.1). These sensors could coordinate to Hg^{2+} and aggregate into nanoparticles for achieving fluorescence "turn-on." They prepared *L*-coated test paper by uniformly coating the *L* powder on filter paper. The test paper showed good stability in sensing Hg^{2+} , which proved its potential for commercial use.

Besides, thymine-like amide groups are excellent specific binding sites for Hg^{2+} . Shi et al. (2016) prepared a triphenylamine-barbituric acid adduct **4** with AIEE features through a simple Knoevenagel reaction under mild conditions without a catalyst for sensing Hg^{2+} (Fig. 3.1). Exposure of **4** to Hg^{2+} resulted in the formation of polymer-like complex of thymine-like group in **4** with Hg^{2+} and the significant increase in fluorescence intensity at 600 nm. The formation process of the nanoaggregates exhibited good linearity over the range of Hg^{2+} concentration from 0.5 to 4μ M, and **4** possessed excellent anti-interference ability in selectivity experiments.

Further, Hg^{2+} shows high affinity for iodide ion due to the ability to form HgI_2 . Zhang et al. (2016) constructed an AIE nanosensor **5** with high sensitivity and selectivity containing a TPE fluorophore, which was functionalized by quinolinium salt with hexafluorophosphate as anion, for Hg^{2+} detection (Fig. 3.1). As a typical quencher, the addition of I⁻ could quench the red fluorescence emitted by **5** nanoaggregates in aqueous solution. However, the addition of Hg^{2+} could cause the formation of HgI_2 to restore fluorescence, making **5** a "turn-on" fluorescent sensor. The sensing behavior was observed having a good linear relationship at Hg^{2+} concentration of 0.5–4.0µM with a detection limit as low as 71.8 nM. These attributes made **5** suitable for the analysis of Hg^{2+} in water or urine.

The sensors based on special reaction appear to be more selective than those based on ligand complexation. Wang et al. (2015a, b) designed a "turn-off" fluorescent nanosensor **6** with AIE properties and good selectivity based on cyanostilbene derivative by taking advantage of the ability of Hg²⁺ to selectively cleave vinyl groups for sensing Hg²⁺ (Fig. 3.2). As the Hg²⁺ concentration increased, the fluorescence signal of **6** gradually changed from the original strong yellow fluorescence emission to weak emission. This quenching behavior showed good linearity in the range of Hg²⁺ concentration from 0 to 50µM with a low detection limit of 37 nM. Besides, the sensor has been successfully applied to the detection of Hg²⁺ in river, lake, and tap water samples.

Detection of Hg^{2+} based on Hg^{2+} -induced dethioacetalization has been documented, but it usually tends to aggregate and quench the fluorescence in aqueous solution (Gao et al. 2018; Cheng et al. 2010). Recently, our group developed a fluorescent nanosensor 7 functionalized with 2-mercaptoethanol for the detection of Hg^{2+} (Gao et al. 2019). In aqueous solution, 7 exhibited good solubility with weak



Fig. 3.2 Monomer chemical structures of AIE-based nanosensors 6-9 for Hg²⁺ detection

fluorescence emission. Besides, addition of Hg^{2+} caused the hydrolysis reaction to expose the aldehyde group, which changed its solubility and ICT efficiency, resulting in fluorescence "turn-on." The detection limit was as low as 1.45 nM, indicating it is of extremely high sensitivity. Good recovery rate was obtained in recovery experiments of real water samples and urine samples, and the prepared paper test strips were capable of detecting Hg^{2+} with the lowest concentration of 0.3µM. Moreover, **7** was able to image the Hg^{2+} in MDA-MB-231 cells and zebrafish models in a concentration-dependent manner.

Chen et al. (2017) combined TPE and rhodamine to prepare a highly efficient ratiometric nanosensor **8** for the detection of Hg^{2+} (Fig. 3.2). They firstly proposed the dark through-bond energy transfer (DTBET) mechanism; the characteristics of possessing rigid linker, dark donor with low quantum yield, and large torsional angle endowed DTBET-based system with the advantages of high energy transfer efficiency. Furthermore, it could effectively resolve the paradox of the conventional Förster resonance energy transfer (FRET) system that the requirement of large Stokes shift always decreases FRET efficiency. TPE as a dark donor did not leak fluorescence emissions exhibiting high energy transfer efficiency. In the absence of Hg^{2+} , only the blue fluorescence induced by TPE aggregate was observed in aqueous solution. Addition of Hg^{2+} would result in a significant ratiometric change in fluorescent emission due to the DTBET. The detection limit was as low as 0.3 ppb. It is worth mentioning that the nanosensor was applied to evaluate and visualize the bioaccumulation of Hg^{2+} in vivo within the rotifer in subsequent related work (Jiang et al. 2017).

Similarly, another ultrasensitive ratiometric fluorescent nanosensor **9** based on AIE and DTBET was developed by Jiang et al. (2019), which incorporated a rhodamine B thiolactone unit as acceptor (Fig. 3.2). The fluorescence enhancement could reach 30,000-fold when the reaction was saturated. Notably, the detection limit could be as low as 43 pM, which was the lowest among the currently reported ratiometric sensor for Hg²⁺. Moreover, the absorption spectrum showed that colorimetric detection was also feasible. Paper strips prepared using Whatman filter paper were also developed, which appeared purple when Hg²⁺ was as low as 400 nM and became darker as the concentration increased. The potential utility of **9** for biological application and environmental monitoring was demonstrated through fluorescence imaging in onion tissues and tap water sample recovery study.

Compared with conventional polymers, supramolecular polymers formed by directional and reversible noncovalent interactions have been paid high attention in sensor field due to their advantages of simple preparation, recyclability, and self-adjustability. Macrocycles that are intimately associated with supramolecular polymers play a critical role in providing cavities to accommodate guest molecules. In addition to some conventional macrocycles such as cyclodextrins (Harada et al. 2009), crown ethers (Zheng et al. 2012), calixarenes (Baldini et al. 2007), novel macrocycles such as pillararenes have also been developed recently (Ogoshi et al. 2016).

Wang et al. (2016) have exploited the self-assembled AIE micelles to construct a FRET-based ratiometric sensor 10 for sensing Hg^{2+} in water. They chose


Fig. 3.3 Self-assembled AIE micelles to construct FRET-based ratiometric sensor **10**. Cyanostilbene modified with hydrophilic methoxypolyethylene glycol and trifluoromethyl group (donor) acted as nano-carrier to form a rod-coil. Rhodamine B derivative (acceptor) was encapsulated inside the carrier. Adapted from Wang et al. (2016) with permission from Royal Society of Chemistry



Fig. 3.4 Nanosensor **11** was prepared by co-self-assembly of amphiphilic diketopyrrolopyrrole DPP1 and tetraphenylethylene TPE1 derivatives. The complexation of DPP1 with Hg^{2+} caused aggregation thus quenching its fluorescence but had no effect on the TPE1, thereby achieving ratiometric sensing. Adapted from Nie et al. (2018) with permission from Elsevier

cyanostilbene modified with hydrophilic methoxypolyethylene glycol and trifluoromethyl group capable of increasing hydrophobicity to form a rod-coil self-assembled system as donor and nano-carrier. The rhodamine B derivative was encapsulated inside the carrier (Fig. 3.3). The addition of Hg²⁺ caused the spirolactam ring to open and the maximum fluorescence intensity at 490 nm gradually decreased accompanying the increase in orange emission at 585 nm, which demonstrated the FRET process. The value of $I_{585 nm}/I_{490 nm}$ had a good linear correlation with the concentration of Hg²⁺ in the range of 0–1.2µM. Besides, the sensor exhibited superior selectivity toward Hg²⁺.

Interestingly, a novel approach for Hg^{2+} sensing was introduced by Nie et al. (2018). They prepared nanosensor **11** with two-photon property by co-self-assembly of two different molecules, amphiphilic furyl diketopyrrolopyrrole (DPP1) and TPE1, in water (Fig. 3.4). In aqueous solution, DPP1 could self-assemble into loose nanoaggregates and maintain stable fluorescence emission at 540 nm while TPE1



Fig. 3.5 Monomer chemical structures of AIE-based nanosensors 12-13 containing peptide backbone for Hg^{2+} detection

exhibited the AIE effect at 470 nm. The nanosensor, which possessed cavities provided for loading drugs, could be easily obtained by co-self-assembly. The complexation of DPP1 with Hg^{2+} caused aggregation thus quenching its fluorescence but had no effect on the fluorescence of TPE1, thereby achieving ratiometric sensing with a detection limit as low as 13 nM. In view of the fact that DPP1 endowed **11** two-photon absorption characteristic, they achieved the two-photon fluorescence sensing of Hg^{2+} with good linearity. The further recovery experiment in tap water sample made **11** potential for practical application.

Bioorganic compounds have been widely used for fluorescence sensing due to their good biocompatibility, biodegradability, and inherent optical properties (Zhang et al. 2014). Among them, biologically derived materials that can support the AIE effect have received strong interest in the field of materials. Singh et al. (2015) developed a fluorescent nanosensor **12** based on cephalexin self-assembly for Hg^{2+} detection (Fig. 3.5). Notably, the preparation was very convenient in which it only required a pulsed ultraviolet (UV) laser to irradiate the cephalexin aqueous solution. The sulfur moiety of the sensor could be combined with Hg^{2+} to cause fluorescence quenching, thus achieving the Hg^{2+} detection with good selectivity and the detection limit low up to 10^{-7} M.

The peptide is an attractive material in the sensing field due to their unique advantages, such as the simplicity of preparation, the adjustability of the peptide backbone, biocompatibility, degradability, and low toxicity. Besides, they possess multiple sites for metal building (Oliveira et al. 2011; Wang et al. 2015a, b). Given all that, Tomar et al. (2018) synthesized a self-assembling tetrapeptide **13** with an overlapping turn of type III/III for sensing Hg²⁺, the fluorophore of which was the dehydrophenylalanine residue (Fig. 3.5). The peptide monomers could self-assemble into spherical nanoparticles with a hydrophobic hollow core. The accumulation of Hg²⁺ in the hollow core induced aggregation accompanying enhanced

fluorescence, which illustrated the AIE property of the tetrapeptide. It is worth mentioning that the overlapping β -turn scaffold was essential for specific sensing of Hg²⁺.

3.2.2 Metallic Nanoparticles/Clusters

In recent years, metallic nanoclusters such as Au nanoclusters (Au NCs) and Ag nanoclusters (Ag NCs) have proven their application value in the field of material science due to their simplicity of preparation, modifiability, high surface area, and dielectric constant. They generally consist of a several to hundreds of Au/Ag atoms-containing metal core and a ligand shell, and they are smaller than 5 nm in size. Importantly, they possess molecular-like photoluminescence (PL) behavior adjusted by different size, oxidation states, and species of capping agents (Saha et al. 2012; Wilson 2008; Zhang et al. 2020). Considering the advantages of fluorescence stability, large Stokes shift, and biocompatibility, a large majority of researchers applied Au/Ag NCs to fluorescence detection of heavy metal ions based on metal ions induced fluorescence enhancement or quenching.

Au NCs have attracted tremendous attention in detecting mercury because of the high affinity from metallophilic Hg²⁺-Au interactions (Xie et al. 2010). AIE organic fluorescence nanoparticles (OFNs) and Au NCs were combined by electrostatic interaction to obtain a ratiometric fluorescent sensor 14 for detecting Hg^{2+} (Niu et al. 2015). A 9,10-stilbene-ruthenium (9,10-di((E)-styryl)anthracene [DSA]) derivative was applied to bind ε -polylysine to form positively charged nanoparticles Ply-9,10-bis(3-formylstyryl)anthracene (BFSA) OFNs as a reference material, while bovine serum albumin (BSA) stabilized nanoclusters as a responsive material with negative charge (Fig. 3.6). OFNs possessed stable green emission with the DSA derivative aggregates while red-emitting nanoclusters would quench when sensing Hg²⁺, resulting in the entire sensing system eventually turning greenemitting. Moreover, this change appeared observable to the naked eyes. The sensing process exhibited excellent linearity in the range of Hg²⁺ concentration from 0 to 1µM with detection limits as low as 22.7 nM. Cell experiments confirmed the good biocompatibility, and the nanosensor was found to be predominantly localized in the cytoplasm. Moreover, the stability verification experiment in human serum indicated the obvious practicability.

Ag NCs are also regarded as important sensing materials for Hg^{2+} detection due to their good stability and low cost. Besides, some possess the property of AIE such as thiolate-protected nanoclusters (Zhang et al. 2015a, b). Interestingly, hydrophilic ligand-protected nanoclusters have good solubility in aqueous solution and cannot maintain strong stable emission. Pan et al. (2018) demonstrated for the first time the hydrophobic interaction-induced AIE Ag NCs **15**. It was capped with thiosalicylic acid acting as hydrophobic capping ligand for detecting Hg^{2+} (Fig. 3.7). When the ratio of thiosalicylic acid to silver ion was 1:4, the optimum luminescence was obtained at 612 nm. The addition of Hg^{2+} could cause the fluorescence quenching,



Fig. 3.6 Schematic illustration of the synthesis of the dual-emission ratiometric nanosensor **14** for the detection of Hg²⁺. Ply-BFSA OFNs were synthesized through the reaction between ε -Ply and BFSA. Red-emitting Au NCs were prepared via a simple procedure using BSA. Through the electrostatic adsorption, Ply-BFSA OFNs with positive charge and Au NCs with negative charge assembled into **14**. Adapted from Niu et al. (2015) with permission from Royal Society of Chemistry



Fig. 3.7 Schematic composite of AIE Ag NC **15**. Sensor **15** was capped with thiosalicylic acid acting as hydrophobic capping ligand, for the detection of Hg^{2+} . Upon addition of Hg^{2+} , the fluorescence of **15** would be quenched in a linearity behavior from 0.3 to 2.2µM. Adapted from Pan et al. (2018) with permission from Royal Society of Chemistry

and the quenching behavior exhibited good linearity in the range of $0.3-2.2\mu$ M (limit of detection [LOD] = 91.3 nM). Moreover, **15** possessed good cell permeability and excellent anti-photobleaching ability.

Copper nanoparticles (Cu NPs) have also attracted considerable attention due to their relatively low price and large abundance in nature. Hatamie et al. (2016) synthesized **16** by using D-penicillamine (DPA) as stabilizing agent and ascorbic acid as reductant under dark condition, which overcame the obstacles of the instability and tendency of Cu NPs to oxidize (Fig. 3.8). The obtained aggregates exhibited strong red fluorescence emission, which benefited from the ligand-to-metal charge



Fig. 3.8 Synthetic strategy of AIE-based Cu NPs **16**. Red-emitting **16** was prepared by using DPA as a stabilizing agent and ascorbic acid as reductant. Adapted from Lin et al. (2016) with permission from Elsevier

transfer between the thiolate ligands in the DPA and Cu atoms. Upon the addition of Hg^{2+} , the high-affinity metallophilic interaction promoted the formation of the closed-shell complexes, which further inhibited the binding of the thiolate ligands with the Cu atoms. In consequence, the aggregates were dispersed into the small homogeneous particles accompanying the fluorescence quenching. The quenching behavior exhibited good linearity in the range from 1.0 to 30μ M of Hg^{2+} concentration (LOD = 32 nM). The superior properties made **16** capable of detecting Hg^{2+} in tap water.

3.2.3 Carbon Dots

Carbon dots, a newly emerged nanomaterial consisting of carbon (C), oxygen (O), nitrogen (N), hydrogen (H), and some doped elements, have been widely concerned in the field of sensing and bioimaging because of their simple synthesis, unique PL feature, excellent photostability, and good biocompatibility (Hola et al. 2014). Moreover, compared with Au/Ag nanoclusters, carbon dots are more environment friendly. The PL properties of carbon dots can be tuned by modulating the size and surface of chemical groups, and higher quantum yields can be obtained by doping other elements or inorganic salts (Wang et al. 2014).

As widely acknowledged, the carbon dots based on fluorescence enhancement generally showed better selectivity and higher accuracy in contrast with those sensors based on fluorescence quenching. Xu et al. (2018) studied the nitrogen and sulfur co-doped fluorescent carbon dots (N,S-CDs, **17**) prepared in glycerol for quantification of Hg^{2+} based on fluorescence "turn-on" method (Fig. 3.9). They took a one-step microblog-assisted method to obtain N,S-CDs with glycerol and cystine as reactants, and the average size was at 1–6 nm. Glycerol was used here not only as a solvent but also as a passivator proved by the presence of hydroxyl groups derived from glycerol on the surface of N,S-CDs. The specific interaction between sulfur-containing groups and Hg^{2+} led to aggregation, which induced AIEE at 385 nm thereby achieving detection with good selectivity (Table 3.1).



Fig. 3.9 Synthesis process and structure of nitrogen and sulfur co-doped fluorescent CDs (N,S-CDs, 17). Sensor 17 was obtained by a one-step microblog-assisted method using glycerol and cystine as reactants. It composited of two fragments: graphite morphology core and passivation layer. Adapted from Xu et al. (2018) with permission from Springer

3.3 AIE-Based Nanosensors for the Detection of Cu²⁺

Another promising direction of AIE-based nanosensors for practical application is the establishment of methods for Cu^{2+} detection. Unlike sensor for Hg^{2+} , nanosensor for tracing Cu^{2+} acts more in a "turn-off" behavior since Cu^{2+} itself is a fluorescence quencher due to its paramagnetic nature (Glaser et al. 2007). Besides, the chelation between Cu^{2+} and identification groups has effect on the nanoaggregates of nanosensors, leading to the attenuated intensity of fluorescence (Singh et al. 2016; Liu et al. 2017). As a result of both aspects, the fluorescence emission is quenched, also called chelation-enhanced quenching. Utilizing this strategy, numerous nanomaterials have been employed to construct "On-Off" nanosensors to detect Cu^{2+} , including organic molecule nanoparticles, metal-organic frameworks, hydrogen-bonded organic frameworks, and organic dots.

3.3.1 Organic Molecule Nanoparticles

Considering the difference in the identification units, it can be divided into several categories: (1) imine groups, for instance, Schiff bases, hydrazones, or salicylalde-hyde Schiff base scaffold; (2) pyridine units; (3) multiple carboxyl groups; and (4) aza-crown ether. Exploiting these units as ligands for complexing with Cu²⁺, the fluorescence emission of AIE fluorophores could be tuned after response behavior happens.

Among these recognition components, imine groups are the parts mainly deployed. Since the imine group has strong affinity to Cu^{2+} , coupling the AIE fluorophores and imine groups paves an interesting pathway to construct an AIE nanosensor for Cu^{2+} detection (Zou et al. 2012; Huo et al. 2012). Besides, some hydrazones derived from salicylaldehyde have even been reported to exhibit a typical AIE feature, meaning no need for further introduction of AIE units (Liu et al.

			-	1	0			
	Emission	Concentration of					Real water	
Nanosensor	wavelength	sensors	S:A	LOD	Linear range	System	examples	References
1	525 nm	10µM	1:1	Ι	I	THF/water (1:9 v/v)	I	Wei et al. (2018)
7	1	10µM	2:1	9.84 nM	I	THF/water (1:9 v/v)	1	Wang et al. (2018a,
								p)
3	550 nm	1	1:1	0.20 nM	0-120µM	Water	Y	Wang et al. (2019a,
								(q)
4	600 nm	50µM	I	3.4μM	0.5-4.0µM	THF/water (3:7 v/v)	I	Shi et al. (2016)
S	610 nm	10µM	I	71.8 nM	0.5-4.0µM	DMSO/water	I	Zhang et al. (2016)
						(1:99 v/v)		
9	537 nm	20µM	I	37 nM	0–50µM	PBS	Y	Wang et al. (2015a,
								p)
7	490 nm	10µM	I	1.45 nM	0–302.4 nM	DMSO/water	Y	Gao et al. (2019)
						(1:99 v/v)		
8	595/485 nm	10µM	I	0.3 ppb	I	CH ₃ CN/water	1	Chen et al. (2017)
						(4:6 v/v)		
6	592/485 nm	10µM	I	43 pM	2-10µM	CH ₃ CN/water	Y	Jiang et al. (2019)
						(4:6 v/v)		
10	585/490 nm	-	I	$0.11 \mu M$	$0.1.2\mu M$	Water	1	Wang et al. (2016)

Table 3.1 List of characteristics of nanosensors with AIE properties developed for Hg²⁺ detection

11	540/470 nm	1	1	Ca.	1	Water	Y	Nie et al. (2018)
				13 nM				~
12	460 nm	1	I	0.1μM	I	Water	1	Singh et al. (2015)
13	500 nm	50µM	1:1	10µM	I	Acetonitrile	1	Tomar et al. (2018)
14	625/525 nm	1	I	22.7 nM	50-	THF/water	1	Niu et al. (2015)
					1000 nM	(0.5:99.5 v/v)		
15	612 nm	6.6 ug/mL	I	91.3 nM	0.3–2.2µM	Water	1	Pan et al. (2018)
16	620 nm	1	I	23 nM	1.0–30 nM	Water	Y	Lin et al. (2016)
17	385 nm	1	I	0.5µM	$1-75\mu M$	Water	Y	Xu et al. (2018)

S:A stoichiometric ratio; LOD lowest limit of detection, THF tetrahydrofuran, DMSO dimethyl sulfoxide, PBS phosphate buffered saline; Y means successful practical application in real water samples

2015; Tiwari et al. 2018; Liu et al. 2017). Regrettably, the fluorescent intensity of such nanosensors sometimes showed abnormal fluctuation in the presence of Al^{3+} , Fe^{2+} , or Fe^{3+} , which limits its further practical application (Xu et al. 2019).

Feng et al. (2014) designed and synthesized a Schiff base macrocycle by the condensation reaction of TPE dialdehyde and 1,2-benzenediamine (Fig. 3.10). In an aqueous solution, the macrocycle (18) aggregated into nanofibers emitting strong fluorescence. After addition of Cu^{2+} at 13µM, the quench of fluorescence emission accompanying a color change from colorless to intense yellow-brown could be observed by the naked eyes. Sensor 18 was capable of achieving the nM level detection since the interspace in the macrocyclic framework between the two circumjacent imine groups just accommodated one Cu^{2+} ion. Moreover, the high selectivity toward Cu^{2+} among various metal ions allowed detection in various matrix, including real water and pork juice-containing water.

Singh et al. (2016) developed nanosensor **19** by taking advantage of the strategy involving excited state intramolecular proton transfer coupled AIE effect (Fig. 3.10). Sensor **19** could self-assemble into nanorods and exhibited intense green fluorescence in aqueous solution, but the presence of Cu^{2+} could disrupt the self-assembled aggregates via the metal chelation with Schiff base unit. Consequently, the nanorods were disassembled showing nearly no emission. Moreover, **19** could also be used for colorimetric assay along with the discernible color transition from light green to dark green.

Sharma et al. (2017) developed a Cu²⁺ nanosensor **20** based on the diimino dicarbazole derivatives with AIEE properties (Fig. 3.10). The aggregates of **20** could selectively sense Cu²⁺ in the nanomolar range and mediate the reduction of Cu²⁺ to Cu nanorods via the interaction with the nitrogen atoms of the imines. Along with the conversion, the fluorescence was quenched accompanying with the color change from light yellow to dark yellow. Further, ensembles of **20** and Cu nanorods were utilized for selective sensing of CN⁻ ions, which could itch the Cu nanorods and recover the aggregation state of **20**.

Huang et al. (2018) designed a chiral molecule **21** with AIEE characteristics by introducing a chiral alanine to hydrazone derivative (Fig. 3.10). Cu²⁺ exhibited high affinity to the nitrogen atom of the imine moiety and oxygen atom of the phenolic hydroxyl. Addition of Cu²⁺ resulted in fluorescence quenching due to the photo-induced electron or energy transfer process. Meanwhile, the self-assembly of **21** was transformed from long nano-helical fibers into irregular polyhedral particles. The "On-Off" behavior allowed detection of Cu²⁺ in concentration ranging from 0 to 0.6µM with an LOD calculated to be 67 nM. It is worth mentioning that the chiral component endowed **21** with the properties of aggregation-induced circular dichroism and circularly polarized luminescence.

The Cu²⁺-induced AIE fluorescence of 5-chlorosalicylaldehyde azine (**22**) quenching was first illustrated by Liu (2015) and the salicylaldehyde azine provided a new platform for Cu²⁺ detection. For instance, Liu et al. (2017) synthesized two Schiff base compounds via the condensation of salicylaldehyde or 2-hydroxy-1-naphthaldehyde with hydrazine hydrate (Fig. 3.10). The two AIE active compounds (**23** and **24**) could aggregate into needle-like particles in pure aqueous environment



Fig. 3.10 Monomer chemical structures of AIE-based nanosensors 18-25 containing Schiff bases or hydrazones for Cu²⁺ detection



Fig. 3.11 Novel AIE plus FRET strategy for Cu^{2+} detection with elevated sensitivity. Sensor 26-1 was combined with the Nile red to construct a FRET system (26) in which the energy of the excited state was transferred from 26-1 to Nile red. Upon addition of Cu^{2+} , the green fluorescence of 26-1 could be quenched thus disrupting the FRET process, making the sensitivity to be elevated hundreds of times. Adapted from Yang et al. (2019a, b) with permission from Elsevier

exhibiting intensive green fluorescence. Addition of Cu^{2+} promoted the further selfassembly to form more stable complexes, thus quenching the fluorescent emission. The "Off-to-On" response happened when the concentration of Cu^{2+} was as low as 10 ± 0.3 nM. More recently, Xu et al. (2019) reported an asymmetric hydrazone **25** based on salicylaldehyde for fluorometric "turn-off" detection of Cu^{2+} (Fig. 3.10). Owing to the introduction of an electron acceptor (nitro group), the fluorescence of **25** exhibited a little red-shift with a strong orange emission in the aggregate state. Sensor **25** was shown to have high sensitivity for Cu^{2+} (LOD = 74.5 nM).

Yang et al. (2019a, b) synthesized a material **26-1** derived from carbohydrazide displaying strong green emission in the nanoaggregates, which could be quenched upon addition of Cu^{2+} , as depicted in Fig. 3.11. Regrettably, the LOD of **26-1** was higher as 13.2μ M, which lacked sufficient sensitivity to meet the need for practical



Fig. 3.12 Monomer chemical structures of AIE-based nanosensors 27-30 for Cu²⁺ detection

application. In view of this, they combined **26-1** with Nile red to construct a FRET system (**26**). Consequently, the LOD of **26-1** was elevated into nano level (9.12 nM). Meanwhile, as the green emission of **26-1** acted as excitation energy for Nile red, the fluorescent signal of the mixed aggregates was red-shifted to 630 nm. This novel AIE plus FRET strategy provided a well-anticipated avenue to improve some poor properties and disadvantages of AIE active nanosensors.

Besides imine groups, some nanomaterials could also serve as a superior nanosensor for Cu^{2+} after introduction of multiple pyridine units (You et al. 2011; Zhang et al. 2013a, b), and act like the nanosensors containing Schiff bases in a chelationenhanced quenching behavior (Dasgupta et al. 2008). Li et al. (2015a, b) utilized AIE active binaphthol to develop a Cu^{2+} nanosensor 27 with the assistance of the pyridine moieties (Fig. 3.12). Upon addition of Cu^{2+} , the morphological structure of aggregates of 27 is transformed from nano-polyhedra into micro-branches, and the strong ligand-to-metal charge transfer also contributed to the quenching of fluorescence. Besides the fluorophore of 27, binaphthol is a well-known chiral fluorophore, thus allowing the circular dichroism signal as well.

Dipicolylamine unit is another multi-pyridine moiety with high affinity to Cu^{2+} and serves as an identification group in some sensors for Cu^{2+} detection (Choi et al. 2009). For example, recently a dipicolylamine unit was attached with TPE group to prepare a Cu^{2+} nanosensor **28** (Fig. 3.12) (Li et al. 2017). The nanoaggregates of **28** emitted intense green fluorescence in aqueous solution and could be effectively quenched by the presence of Cu^{2+} . This quenching process was highly selective toward Cu^{2+} over other metal ions thus guaranteeing its high anti-interference. The linear concentration of Cu^{2+} ranged from 0 to 8.0μ M while LOD was found as 4.29 nM, all validating its possibility in practical application in complex water matrix and living cells.

Crown ether group is another choice to constitute a moiety with high affinity to Cu^{2+} (Zheng et al. 2012). In Zhang's works, aza-crown ether was selected as a ligand to Cu^{2+} and coupled with Schiff base to construct a Cu^{2+} nanosensor **29** (Fig. 3.12;

Zhang et al. 2017a, b). In aqueous media, **29** aggregated into irregular globular-like aggregates emitting strong fluorescence but quenched in the presence of Cu²⁺. Although the sensitivity was not ideal (LOD = 2.32μ M), **29** exhibited rather higher selectivity while the interference of other coexisting metal ions was nearly negligible. The superior performance was ascribed to the appropriate characteristics of aza-crown ether such as the cavity of the macrocycle, the number of binding sites, or the variety of oxygen and nitrogen atoms on the crown. These all contributed to the practical detection of Cu²⁺ in real water.

TPE derivative **30** with quadruple carboxyl groups was illustrated having specific selectivity for Cu²⁺ ions in Geng's work, as shown in Fig. 3.12 (Geng et al. 2019). In aqueous solution, the aggregates of **30** exhibited intense blue emission but the addition of Cu²⁺ could lead to the fluorescence quenching. The quench efficiency of fluorescence exhibited linearity in the large range of $0-15\mu$ M with detection limit as low as 4.5 nM. Sensor **30** has been applied in fabricating test strips for detecting Cu²⁺ attributed to its strong emission in solid states.

3.3.2 Metal-Organic Frameworks

Although numerous nanosensors involving AIE scaffolds salicylaldehyde azines exhibited high affinity to Cu²⁺, their pH-dependent emission still limited their practical application (El-Sherif and Aljahdali 2013). Lu et al. (2016) provided a facile strategy to surmount this obstacle by confining them within metal-organic frameworks. The complex **31** was formed via the integration of **31-1** into a zirconium(IV) dicarboxylate porous material (UiO-66) exhibiting concentration and pHindependent fluorescence emission, as shown in Fig. 3.13. Owing to the nano-space confined effect, the space torsion of **31-1** was restricted, thereby exhibiting strong



Fig. 3.13 Formation process and structure of complex 31. Complex 31-1 was integrated into UiO-66 to obtain UiO-66-31-1 complex (31). Via the host-guest process, encapsulation of 31-1 into the metal-organic frameworks could enlarge its application in pH and concentration-independent behavior. Adapted from Lu et al. (2016) with permission from Royal Society of Chemistry



Fig. 3.14 Structural illustration of the formation of hydrogen-bonded organic frameworks **32** for Cu^{2+} sensing. With amide linkages, it was formed without employment of any guest molecules. Owing to the restriction of intra- and inter-molecular rotation, the blocked non-radiative channel led to the intensive emission of **32**. Adapted from Zhou et al. (2015) with permission from Royal Society of Chemistry

fluorescence over a wide pH range (pH = 4.0-8.0) as well as at low concentration, which could enlarge their application in some extreme environments. Since the guest **31-1** possessed a high affinity to Cu²⁺, **31** served as a sensitive and selective nanosensor for Cu²⁺ detection in aqueous solution (LOD = 50 nM).

3.3.3 Hydrogen-Bonded Organic Frameworks

Hydrogen-bonded organic framework is another pathway to detect Cu^{2+} with increasing interest as stimuli-sensitive nanomaterials. For example, **32** could form a stable structure without the employment of any guest molecules, and disperse well in organic solution (Zhou et al. 2015). Owing to the restriction of intra- and intermolecular rotation, the non-radiative channel was blocked, leading to high fluorescence (Fig. 3.14). Nevertheless, Cu^{2+} ions could coordinate with the amide groups and impair the interaction of hydrogen bond, thus causing the dissociation of frameworks. Subsequently, the intramolecular rotation of TPE moieties was allowed and the non-radiative channel was opened. Regrettably, the detection was performed in DMSO, which largely limited its practical application.

3.3.4 Metallic Nanoparticles/Clusters

As mentioned above, Cu NPs/NCs have been used widely in metal detection. Generally, highly fluorescent Cu NCs are formed by adding ligands into Cu²⁺ solution, with the "Off-to-On" fluorescence change (Kasana et al. 2017). In view of this

unique transformation, Li et al. (2015a, b) employed the fast formation process of Cu NCs (**33**), in which penicillamine was chosen as ligands, to detect Cu^{2+} . Due to the high specificity, no fluorescence could be triggered upon addition of metal ions, except for Cu^{2+} . Even the introduction of Ag⁺ and Au³⁺, two of noble metal ions often used for the construction of metal nanoparticles/clusters, had no impact on the detection process. In light of this strategy based on the formation process of Cu NCs, it may provide a new means for developing a low-cost, real-time, and easy-to-use sensor for evaluation of Cu²⁺.

3.3.5 Organic Dots

Similar to hydrogen-bonded organic frameworks that could modulate the property of AIE materials, organic dots provided another way to pave for the development of novel functional AIE materials (Dasgupta et al. 2017). Jiang et al. (2018) prepared poly(acrylamide-co-acrylic acid) (PSMA)-coated organic dots **34** in which 9,10-di((E)-styryl)anthracene (DSA) was employed as the AIE fluorescent material. As shown in Fig. 3.15, the hydrophobic moieties were encapsulated inside while the carboxyl groups were exposed on the surface in the self-assembly process of AIE dots. In water, the suspension of **34** displaying intense green-yellow fluorescence could be quenched by Cu²⁺ with LOD calculated as 107 nM. The response was attributed to the high-affinity coordination of carboxyl groups with metal ions. Nonetheless, similar efficiency of quenching also happened to Fe³⁺ and other ions (Table 3.2).



Fig. 3.15 Proposed self-assembly process and structure of organic dots 34. DSA was encapsulated inside while the carboxyl groups were exposed on the surface of 34. DSA was an AIE fluorescent material and these carboxyl groups were provided by PSMA. Adapted from Jiang et al. (2018) with permission from MDPI

				- 1				
	Emission	Concentration of					Real water	
Nanosensor	wavelength	sensors	S:A	LOD	Linear range	System	examples	References
19	526 nm	1μM	1:1	0.26 ppb	0-2.5μM	CH ₃ CN/water (1:99 v/v)	Y	Singh et al. (2016)
20	447 nm	10µM	I	Mu 06	I	THF/water (3:7 v/v)	I	Sharma et al. (2017)
21	586 nm	10µM	1:1	67 nM	0-0.6µM	THF/water (1:9 v/v)	I	Huang et al. (2018)
24	530 nm	10µМ	4:1	(10 ± 0.3) nM	1	CH ₃ CN/water (1:99 v/v)	I	Liu et al. (2017)
25	570 nm	10µМ	1:1	74.5 nM	0.5-5.0µM	CH ₃ CN/water (1:99 v/v)	1	Xu et al. (2019)
26	525 nm	10µМ	1:1	132 nM	1	CH ₃ CN/water (1:99 v/v)	1	Yang et al. (2019a, b)
27	511 nm	10µМ	1:1	148 nM	1	DMSO/water (2:8 v/v)	1	Li et al. (2015a, b)
28	500 nm	10µМ	1:1	4.29 nM	Мщ0.8-0	DMSO/water (1:4 v/v)	1	Li et al. (2017)
29	570 nm	10µМ	1:1	2.32µM	1	DMSO/water (1:99 v/v)	Y	Zhang et al. (2017a, b)
30	432 nm	10µМ	1:2	4.5 nM	0-15µM	Ethanol/water (1:9 v/v)	I	Geng et al. (2019)
31	560 nm	10µM	1:1	50 nM	0.1-4.0µM	Water	Y	Lu et al. (2016)
33	673 nm	I	I	0.3 ppm	0.95– 6.35 ppm	1	Υ	Li et al. (2015a, b)
34	524 nm	1	I	107 nM	0.025–6µM	1	I	Jiang et al. (2018)
S:A stoichior	netric ratio							

 Table 3.2
 List of characteristics of nanosensors with AIE properties developed for Cu²⁺ detection

74

LOD lowest limit of detection, THF tetrahydrofuran, DMSO dimethyl sulfoxide; Y means successful practical application in real water samples

3.4 AIE-Based Nanosensors for the Detection of Zn²⁺

Although it has been documented that dipicolylamine, 8-aminoquinoline, semicarbazide, and carboxyl were superior recognition components toward Zn^{2+} (Carol et al. 2007; Jia et al. 2012; Liu et al. 2013), their roles in AIE organic molecule nanoparticles were still rarely illustrated. By contrast, metallic NPs/NCs have been greatly developed. Au NCs, Ag NCs, and Cu NCs were all employed in the detection of Zn²⁺. Besides, the combination of Au NCs with metal-organic frameworks appeared as a good way to elevate the detection sensitivity and selectivity (Li et al. 2018).

3.4.1 Organic Molecule Nanoparticles

As one of the well-known building blocks of AIE fluorophores, the terpyridine moiety is also a metal-chelating group (Swiegers and Malefetse 2000; Schubert and Eschbaumer 2002). Jung et al. (2015) constructed a terpyridine-based nanosensor **35** for detecting Zn^{2+} (Fig. 3.16). The chelation between the terpyridine moieties and the Zn^{2+} induced the nanosensor **35** aggregation from the fibrillar structure into a spherical structure. This molecular aggregation by coordination bonding led to the restriction of intramolecular rotation (RIR) without π - π stacking. Therefore, the aggregated **35** could be utilized as a "turn-on" fluorescent nanosensor for Zn^{2+} . The fluorescence intensity showed tenfold enhancement when the concentration of Zn^{2+}



Fig. 3.16 Monomer chemical structures of AIE-based nanosensors 35 for Zn²⁺ detection



Fig. 3.17 Schematic representation of the preparation of metal-organic frameworks 36. Sensor 36 was formed by self-assembly due to the coordination of carboxylate groups to Tb^{3+} ions

was increased to 15 nM, while a strong fluorescence emission could still be observed as low as 1.1 nM. In addition, **35** was used to detect Zn^{2+} in urine samples successfully.

3.4.2 Metal-Organic Frameworks

Metal-organic frameworks have played an indispensable role in Zn^{2+} detection as well. For instance, Lin et al. (2017a, b) utilized an AIE fluorophore, **36-1**, to prepare a Zn^{2+} ratiometric fluorescent nanosensor **36** (Fig. 3.17). The nanosensor was formed by self-assembly due to the coordination of the carboxylate groups to metal ions. Interestingly, **36-1** exhibited different wavelength emission in combination with different ions, with Tb³⁺ emitting at 590 nm and Zn²⁺ at 470 nm. In view of the cation exchange process of **36** with Zn²⁺, the complex was developed as a ratiometric fluorescent nanosensor with high selectivity for the detection of Zn²⁺. The ratiometric biosensor could be applied for detection of Zn²⁺ in real water samples. Worth to mention, this strategy using the cation exchange process of metal-organic frameworks paved a way toward AIE nanomaterials with ratiometric fluorescence response.

3.4.3 Metallic Nanoparticles/Clusters

In the preparation process of metallic NPs/NCs, ethanol or some external stimulants were required to induce pre-aggregation (Kasana et al. 2017; Kong et al. 2017). For mercaptopropionic acid or glutathione (GSH)-capped metallic NPs/NCs, Zn²⁺ could act as cross-linker to bridge components, promoting further aggregation of these nanoparticles. As a result, the tighter aggregates yielded stronger fluorescence.

Additionally, elevated sensitivity, higher selectivity, or even ratiometric behavior could be achieved via combining with other active nanomaterials.

Recently, Kuppan, and Maitra (2017) reported rapid room temperature synthesis of 3-mercaptopropionic acid-capped Au NCs (**37**) in aqueous solution. Since the carboxylate groups of 3-mercaptopropionic acid acted as a ligand with high affinity to Zn²⁺, the electrostatic binding with Zn²⁺ could rigidify the Au NCs, causing the million-fold enhancement of yellow fluorescence ($\lambda_{em} = 580$ nm). Then, further self-assembly took place to yield highly ordered NCs, and the emission of yellow would transform into green ($\lambda_{em} = 500$ nm). According to the unique behavior, it has been applied for visual and fluorimetric detection of Zn²⁺ with rather high sensitivity (LOD = 9 nM).

In another work, Li et al. (2018) revealed that GSH-capped Au NCs/Zn-MOF composite (**38**) could be of analytical applications in Zn^{2+} detection (Fig. 3.18). In the mixture of Au NCs and 2-methylimidazole (**2-MIM**), the addition of Zn^{2+} could induce the generation of Zn-MOF in situ within which Au NCs were confined. Compared with sole Au NCs, the quantum yield, sensitivity, and selectivity were all elevated, presenting a relatively large linear range (12.3 nM to 24.6 mM) and considerably low LOD as 6 nM. In view of these superior characteristics, **38** was further applied to detect Zn^{2+} in various complex matrixes, including human serum, syrup samples, and milk. The success of unconventional combination of Au NCs with metal-organic frameworks provides a novel nanoscale platform for Zn^{2+} detection, although these metal-organic frameworks were not stable under acidic media.

In order to reduce the costs of metal nanoparticles/clusters using noble metals, Lin et al. (2017a, b) utilized Cu(NO₃)₂ to prepare Cu NCs (**39**) capped by GSH with red emission for "turn-on" detection of Zn²⁺. The presence of Zn²⁺ could cross-link the Cu NCs by electrophilic interaction to form aggregates, inducing the fluorescence enhancement. Compared with Au NCs mentioned above, the Cu NCs exhibit a faster response toward Zn²⁺ and a wider detection range (4.68–2240µM). Moreover, taking the advantages of red emission, large Stokes shift, and good biocompatibility, the attempt to develop a nanosensor based on Cu NCs other than Au NCs is meaningful. Besides, Ag NCs were also proved capable of detecting in Zn²⁺ as well. One



Fig. 3.18 Detection strategy of Zn^{2+} based on AIEE process of Au NCs-MOFs **38**. In a mixture of Au NCs and **2-MIM**, the addition of Zn^{2+} induced the generation of Zn-MOF in situ through enclosing Au NCs. The intensity of fluorescence was highly dependent on the amount of Zn^{2+} existing. Adapted from Li et al. (2018) with permission from Elsevier

attempt is GSH-capped Ag NCs (**40**), which were employed as a "turn-on" nanosensor with fluorescence enhancement of red emission (Liu et al. 2019a, b).

3.5 AIE-Based Nanosensors for the Detection of Pb²⁺

Although phosphate group could incorporate with Pb^{2+} (Khandare et al. 2014), its potential in constructing AIE-based nanosensors was still waiting for being explored. In recent years, compared with the limited progress in organic molecule nanoparticles, the sensor for tracing Pb^{2+} was mainly dependent on metallic NPs/NCs and carbon dots.

3.5.1 Metallic Nanoparticles/Clusters

Ji et al. (2015) firstly reported a kind of glutathione-capped Au NCs (**41**) with AIEE property for sensing Pb²⁺. The Au NCs showed ethanol-induced aggregation and emitted more intensively. Furthermore, addition of Pb²⁺ led to the aggregation of Au NCs accompanying with bright fluorescent emission due to the strong coordination between GSH and Pb²⁺. The "turn-on" response exhibited good linearity in the range of 5.0–50 mM. It is worth to mention that the sensing process could be completed just in 1 min. More importantly, **41** showed satisfactory selectivity toward Pb²⁺ over various divalent metal ions, guaranteeing its application in real water.

Based on the ethanol-induced further aggregation of the GSH-capped Au-NCs, our group optimized the working system and developed a Pb^{2+} nanosensor (42) with higher sensitivity and shorter response time (Zhang et al. 2017a, b). The detection sensitivity was largely enhanced with a higher fraction of ethanol due to the formation of dense Au nanoparticles other than loose clusters. The critical point of the fraction of ethanol was exploited. Upon the addition of Pb²⁺, the nanosensor would turn to be denser and more rigid, showing stronger fluorescence emission. Moreover, the detection of Pb²⁺ could be realized in not only real water but also human serum, with minimal interference from the other matrixes.

3.5.2 Carbon Dots

Tian et al. (2018) developed a "turn-on" nanosensor **43** via immobilizing carbon dots into the shell of the spherical polyelectrolyte brushes for detection of Pb^{2+} . The electrostatic interaction between carbon dots and brushes could induce the quenching of original fluorescence emission. Upon addition of Pb^{2+} , the AIEE-based fluorescence "turn-on" could be achieved due to the mutual contribution of aggregation of the spherical polyelectrolyte brushes particles and the aggregation of carbon dots in shell. Sensor **43** could detect Pb^{2+} in a wide range of concentration and has been applied in real water. Regrettably, the short-wavelength emission of **43**, which means its strong background signal interference, greatly limited its further application (Table 3.3).

3.6 AIE-Based Nanosensors for the Detection of Cr³⁺ and Cr⁶⁺

Chromium (Cr) is an ecological pollutant with valence states ranging from chromium(II) to chromium(VI), while elemental chromium (0) is not naturally occurring (Vincent 2000). As a naturally occurring element, Cr^{3+} exhibits fewer influences in toxicology (Tchounwou et al. 2012). However, excessive intake of Cr^{3+} will lead to genotoxic effects, and its deficiencies could also cause sugar metabolic disorders like engendering diabetes (Flora et al. 1990; Nickens et al. 2010). Accordingly, a number of nanosensors have been constructed for sensing of Cr^{3+} . Unfortunately, currently reported Cr^{3+} nanosensors with AIE characteristics are almost based on the imine bond, which shows high affinity to trivalent cations, which means, however, rough selectivity (Liu et al. 2014; Islam et al. 2019).

Recently, Yu et al. (2019) reported a novel AIE-active nanosensor 44 composed of a TPE unit and rhodamine spirolactam, in which thiophene ring was used as a conjugated electron-rich linker (Fig. 3.19). Compared with the conventional chelation-induced manner, the interaction of nano-aggregated 44 with Cr^{3+} was a comprehensive result of proton-induced ring-opening followed coordination interaction. The Lewis acid characteristics of Cr^{3+} induced the acidic hydrolysis of looplocked spirolactam and gave rise to loop-open response exhibiting AIEE. While, due to the perpendicular configuration and steric hindrance, the monovalent and divalent metal ions could not trigger the chelation-induced ring-opening, so 44 showed good selectivity to these metal ions. Regrettably, the response of 44 was inevitably triggered by other trivalent metal cations or H⁺ for the same reason. Nonetheless, AIE-active rhodamine B-based nanoaggregates were still promising for practical applications due to their high selectivity and visible color change.

Although chromium occurs mostly in the trivalent $[Cr^{3+}]$ form, the detection of hexavalent $[Cr^{6+}]$ form, which is less stable but more toxic, is still of significance (Tchounwou et al. 2012; Stohs and Bagchi 1995; Salnikow et al. 2008). Recent reported AIE nanosensors for detection of Cr^{6+} were mainly based on the oxidability of Cr^{6+} . The Cr^{6+} sensing ability of GSH-Au NCs was assumed to be ascribed to the reduction of Cr^{6+} under acidic conditions while ligand GSH behaved as a reducing agent (Fournier-Salaün and Salaün 2007). The "turn-off" effect was a result of the disruption of the inter-Au(I) interactions; subsequently, the destruction of core-shell structure happened to lead to the quenched fluorescence (Zhang et al. 2017a, b). The reduction process occurred under acidic condition, while there was no response at neutral condition due to the lower redox potential.

Yin et al. (2019a, b) provided a solution for this limiting condition utilizing the mechanism of polymer-salt aggregate self-assembly chemistry to entrap the GSH-Au NCs inside micron-sized capsules (Fig. 3.20). Polymer-salt aggregates **45** were formed via the electrostatic interactions between phosphate anions and the

		Emission	Concentration of					Real water	
Nanosensor	Analytes	wavelength	sensors	S:A	LOD	Linear range	System	examples	References
35	Zn^{2+}	442 nm	10µM	2:3	1.1 nM	1.1–15 nM	DMSO/water (1:99 v/v)	Y	Jung et al. (2015)
36	Zn ²⁺	470/590 nm	0.02 mg mL^{-1}	1	50 nM	0.1–60 nM	Water	Y	Lin et al. (2017a, b)
37	Zn^{2+}	580 nm	1	1:1	Mu 6	0.2-1.0µM	I	Y	Lin et al. (2017a, b)
38	Zn ²⁺	570 mm	20μM	1	40µM	12.3 nM to 504.6 mM	Water	Y	Li et al. (2018)
39	Zn ²⁺	630 mm	1	I	1.17μM	4.68–2240µM	Water	Y	Lin et al. (2017a, b)
41	Pb^{2+}	590 nm	1	I	5μM	5.0-50µM	Water	Y	Ji et al. (2015)
42	Pb^{2+}	595 nm	1	I	0.1µM	2.0–350μM	Ethanol/water (57:43 v/v)	Y	Zhang et al. (2017a, b)
43	Pb^{2+}	491 nm	I	I	22.8µM	0-1.67 mM	Water	Y	Tian et al. (2018)
S:A stoichior	metric ratio								

Table 3.3 List of characteristics of nanosensors with AIE properties developed for Zn²⁺ or Pb²⁺ detection

LOD lowest limit of detection, DMSO dimethyl sulfoxide; Y means successful practical application in real water samples



Fig. 3.19 Monomer chemical structure of AIE-based nanosensor **44** and its proposed response to Cr^{3+} . The interaction of nano-aggregated **44** with Cr^{3+} was a comprehensive result of protoninduced ring-opening followed coordination interaction. In the presence of Cr^{3+} , the Lewis acid characteristics of Cr^{3+} induced the acidic hydrolysis of loop-locked spirolactam and gave rise to loop-open response exhibiting AIEE. Adapted from Yu et al. (2019) with permission from Royal Society of Chemistry



Fig. 3.20 Schematic depiction of micro-encapsulation of Au NCs (**45**) for Cr^{6+} detection. The Au NCs were entrapped inside micro-capsules via polymer-salt aggregate self-assembly. SiO₂ NPs diffused and deposited within the surface to form the shells. Through the micro-encapsulation, the limit of detection was optimized from 52 ppb to 6 ppb. Adapted from Yin et al. (2019a, b) with permission from American Chemical Society

amine groups of poly(allylamine hydrochloride), and GSH-Au NCs were incorporated within their cross-linked network structure. Subsequently, SiO_2 NPs diffused and deposited within the surface to form the shells. Owing to the high density of poly(allylamine hydrochloride) throughout the micro-capsule wall and interior,

these components carrying protonated amine functional groups provided the acidic environment required for reduction of Cr^{6+} and no acidification was needed before testing. Besides, compared with these free Au NCs, the fluorescence of microcapsuled Au NCs was five times more intensive owing to the Mie scattering of multilayer-thick and semipermeable shell. Consequently, the sensitivity was elevated a lot after encapsulation. Furthermore, **45** could be immobilized on paper and successfully extended to a test strip system exhibiting similar Cr^{6+} -induced fluorescence quenching behavior with an LOD estimated to be 30 ppb.

3.7 AIE-Based Nanosensors for the Detection of Cd²⁺

Selective detection of Cd^{2+} has long been a challenge. The nanosensors based on the supramolecular systems such as metal-organic frameworks, nanoparticles, nanoclusters, and carbon dots are rarely reported, while the main papers developed are on the basis of organic molecule nanoparticles, which need be preorganized. For example, Mahajan et al. (2015) developed a small organic molecule **46** based on Isatin and further converted it into nanoparticles by reprecipitation method (Fig. 3.21). Due to the coordination of functional keto moiety with Cd^{2+} , these organic nanoparticles assembled more tightly, which restricted the free rotations of the carbonyl, which largely appeared as a significant enhancement of emission. Similar response has been observed in real water samples as well. Regrettably, **46** could be only used in a weak acid condition, otherwise, the ring-open of **46** would happen and the structural cavity would be disrupted.

Recently, Aguilar et al. (2018) reported a preorganized Schiff base 47, which exhibited an enhanced fluorescence response to Cd^{2+} ions at sub-micromolar concentration after it was converted to organic nanoparticles (Fig. 3.21). Sensor 47 showed high selectivity to Cd^{2+} from other metal ions owing to the stereochemistry of 47, which contained a molecular cavity suitable for chelation with Cd^{2+} ion. The interaction occurred between nitrogen atoms of 47 and the electron acceptor Cd^{2+} , which led to the formation of structurally rigid coordination complexes exhibiting "turn-on" behavior. Moreover, these organic nanoparticles of 47 showed less cytotoxicity in living cells than the free 47, which suggested that the conversion of organic nanoparticles modulated the elevated sensitivity and higher biocompatibility. The practical potential of 47 in Cd^{2+} detection has further been verified in real water and aqueous solution of cigarette tobacco leaves.

Fig. 3.21 Monomer chemical structures of AIE-based nanosensors 46-47 for Cd²⁺ detection



3.8 AIE-Based Nanosensors for the Detection of As³⁺ and As⁵⁺

Arsenic (As) is a ubiquitous metalloid whose contamination in groundwater is a serious global problem threatening the health of more than 150 million people (Jackson et al. 2012; Mohan et al. 2007). The major inorganic salts of arsenic are trivalent arsenite (AsO_3^{3-}) and the pentavalent arsenate (AsO_4^{3-}), while little exist in organic form (Cullen and Reimer 1989). As³⁺ has been reported with higher affinity to thiol or sulfhydryl groups of biomolecules and macromolecules (Mandal and Suzuki 2002; Smedley and Kinniburgh 2002), and some sensors based on the Cysmodified fluorophores have been elucidated (Tian et al. 2017). Unfortunately, similar strategies have not been employed in the development of AIE-based nanosensors for As^{3+} and As^{5+} detection.

Lately, Pathan et al. (2019) exploited Fe_3O_4 nanoparticle functionalized graphene quantum dots as a nanosensor (**48**) for tracing As^{3+} ions in contaminated water. The surface of magnetic dots was full of carboxyl, hydroxyl, and amide groups, which provided high affinity to heavy metal ions. After addition of As^{3+} ions, the coordination of the hydroxyl and Fe_3O_4 nanoparticles to As^{3+} ions via intramolecular or extra-molecular interactions or adsorption induced the formation of rigid structure of the ion-mediated complex. As a result, the fluorescence of **48** was enhanced largely. It is worth noting that **48** exhibited high selectivity for As^{3+} over a relatively wide pH range and the resultant LOD value was 5.1 ppb, which render this nanosensor of practical significance (Table 3.4).

3.9 AIE-Based Nanosensors for the Detection of Multiple Heavy Metals

Apart from these AIE nanosensors for single heavy metal ion detection, designing such nanomaterials with different responses toward multiple cations has also caused extensive concern (Srivastava et al. 2018). In view of the advantages like cost-effectiveness, labor reduction, and reduced analytical time, developing versatile nanomaterials with AIE characteristic for multiple heavy metals' detection is meaningful for practical application as well (Wang et al. 2013; Samanta et al. 2015). From the perspective of design strategy, the multi-response nanosensors could be implemented in various ways, such as varying work solvents, differential response behaviors or subsequent recognition processes.

3.9.1 Organic Molecule Nanoparticles

As mentioned earlier in the section concerning detection of Cu^{2+} , salicylaldehyde Schiff base scaffold showed high affinity to Cu^{2+} (Zou et al. 2012). In Yang's work, anthryl-contained Schiff base derivative **49** exhibited additional affinity toward Zn²⁺ (Yang et al. 2015). Due to the differential electronic configuration of Zn²⁺ and Cu²⁺,

		Emission	Concentration of			Linear		Real water	
Nanosensor	Analytes	wavelength	sensors	S:A	LOD	range	System	examples	References
44	Cr^{3+}	588 nm	10µM	1:6	11.9μM	1	CH ₃ CN/water (3:2 v/v)	1	Yu et al. (2019)
46	Cd ²⁺	417 nm	10µM	2:1	11.8μM	8.9– 443.6μΜ	Water	Y	Mahajan et al. (2015)
47	Cd ²⁺	436 nm	0.1µM	2:1	Mu 60.0	0-0.1μM	Water	Y	Aguilar et al. (2018)
48	As^{3+}	430 nm	10µM	1	5.1 ppb	1	1	1	Pathan et al. (2019)

ç
ste
ď
3+
As
Ţ.
0
5
2
~
1 ³⁺
C
or
Ē
e.
dc
Б
Š
q
es
÷E
je j
10.
Id
(TT)
н.
AIF
h AIF
vith AIF
s with AIF
ors with AIF
isors with AIF
ensors with AIF
osensors with AIF
anosensors with AIF
nanosensors with AIF
of nanosensors with AIF
s of nanosensors with AIF
tics of nanosensors with AIF
istics of nanosensors with AIF
teristics of nanosensors with AIF
acteristics of nanosensors with AIF
aracteristics of nanosensors with AIF
characteristics of nanosensors with AIF
f characteristics of nanosensors with AIF
of characteristics of nanosensors with AIF
ist of characteristics of nanosensors with AIF
List of characteristics of nanosensors with AIF
4 List of characteristics of nanosensors with AIF
3.4 List of characteristics of nanosensors with AIF

S:A stoichiometric ratio, LOD lowest limit of detection; Y means successful practical application in real water samples

49 served as an "Off-On" nanosensor for Cu^{2+} detection in methanol-H₂O mixture solution while acted as an "On-Off" nanosensor for Zn²⁺ sensing in pure methanol solution. Especially, other competing cations had nearly no effect on the fluorescence intensity of **49**.

Similarly, Yin et al. (2019a, b) reported a 2-hydroxyl phenyl imino-modified imidazole derivative **52** possessing salicylaldehyde Schiff base scaffold (Fig. 3.22). The coordination sites composed of the N atoms of imine group, imidazole ring, and the O atom of the hydroxyl group, which exhibited high affinity to Cu^{2+} . Upon addition of Cu^{2+} , the original regular aggregate of **52** was disrupted and then the emission was quenched along with the colorimetric change from colorless to light yellow. Subsequently, upon the sequential addition of Cr^{3+} to the solution, the quenched fluorescence was recovered because of the cation exchange. The whole detection procedure was validated in real water samples. The strategy based on subsequent recognition provided a wider pathway for construction of nanosensor for heavy metal ion detection.

In another work, Pannipara et al. (2018) developed a novel pyrazolyl derivative **50**, which was capable of detecting Cu²⁺, Ni²⁺, and Hg²⁺ ions in different behavior (Fig. 3.22). Similarly, the emission of **50** in nanoaggregates could be quenched upon addition of Cu²⁺, and Ni²⁺ displayed higher efficiency of quenching. Contrary to the fluorescent changes corresponding to Cu²⁺ and Ni²⁺ ions, the blue shift in the emission spectra occurred in the presence of Hg²⁺, which meant the ratiometric sensing was available. It was a result of this that the charge transfer from **50** to Hg²⁺ could lower the energy of lowest unoccupied molecular orbital (LUMO) but not cause fluorescence quench, whereby **50** could be applied for Cu²⁺ and Ni²⁺ detection via "On-Off" mode but could trace Hg²⁺ in ratiometric behavior.

As introduced earlier in the sections discussing Hg^{2+} and Cr^{3+} , the ring-open of spiro would be induced by both Hg^{2+} and Cr^{3+} , leading to the "turn-on" of rhodamine fluorescence (Wang et al. 2016; Yu et al. 2019). Similar situation arose with **51**,



Fig. 3.22 Monomer chemical structures of AIE-based nanosensors 49–52 for detection of multiple metal ions

which was based on the modulated naphthalimide linked with rhodamine through a flexible non-conjugated spacer (Fig. 3.22; Sun and Qian 2016). Prior to the addition of Cr^{3+} or Hg^{2+} , only a weak fluorescent emission of **51** could be observed due to the photo-induced electron transfer process from the N atom of spiro ring on the rhodamine to the naphthalimide moiety. Otherwise, since the electrons were absorbed to the metal cations in the chelation with the hetero atom, the electron transfer process was blocked and the emission of naphthalimide was recovered. On the basis of the above mechanism, **51** served as a multi-responsive nanosensor for detection of Hg^{2+} and Cr^{3+} simultaneously in ratiometric behavior and dual-channel.

3.9.2 Metallic Nanoparticles/Clusters

GSH-Au NC mentioned above is one of the typical luminophores with AIEE properties but needs some "triggers" to modulate its AIEE features, which restricts its application a lot (Zhang et al. 2017a, b; Tang et al. 2017; Guo et al. 2015). Qu et al. (2018) found that fluorescent amino-modified silicon nanoparticles (Si NPs) with positive surface charges could trigger the aggregation of GSH-Au NCs, which carry negative charge via the electrostatic interaction. They prepared self-assembled nanoarchitecture **53** constituting Si NPs and GSH-Au NCs with dual emissions (Fig. 3.23). Similar to free GSH-Au NCs, abundant carboxylic groups of GSH on the surface of **53** provided the binding sites for heavy metal ions while Si NPs had no effect on the chelation. Upon addition of Zn^{2+} or Cd^{2+} , unique second enhancement of fluorescence from GSH-Au NCs could be observed due to its bridge effect to form larger aggregates while the emission from Si NPs was unchanged acting more like an internal reference. Accordingly, the Si NP-capped GSH-Au NCs could be applied as a ratiometric sensor for sensing Zn^{2+} and Cd^{2+} , which has been validated both in real water samples and living cells (Table 3.5).



Fig. 3.23 Schematic illustration of the self-assembly of **53** (Si NPs@GSH-Au NCs) and the sensing process of Zn^{2+} or Cd^{2+} . Amino-modified Si NPs were positively surface charged while GSH-Au NCs were negatively charged. Via the electrostatic interaction, Si NPs could trigger the aggregation of GSH-Au NCs to obtain **53** with dual-emission. Upon addition of Zn^{2+} or Cd^{2+} , unique second enhancement of fluorescence from GSH-Au NCs could be observed but the emission from Si NPs was unchanged. Adapted from Qu et al. (2018) with permission from Royal Society of Chemistry

		Emission	Concentration of			Linear		Real water	
Nanosensor	Analytes	wavelength	sensors	S:A	LOD	range	System	examples	References
49	Cu ²⁺	523 nm	10µM	2:1	212µM	I	Methanol/water (4:1 v/v)	I	Yang et al. (2015)
	Zn^{2+}				71.9 nM		Methanol		
50	Cu ²⁺	424 nm	10µM	1:1	41.3 nM	1	DMF/water	Y	Yin et al. (2019a,
	Cr^{3+}				0.94µM	0-8μM	(50:50 v/v)		b)
51	Cu ²⁺	532 nm	20µM	I	1.61µM	I	THF/water	I	Pannipara et al.
	Hg^{2+}	532/481 nm		1			(15:85 v/v)		(2018)
52	Cr ³⁺	578/440 nm	10µM	I		I	Ethanol/water	I	Sun and Qian
	Hg ²⁺						(1:1 v/v)		(2016)
53	Zn^{2+}	570/450 nm	I	I	0.67 µM	1.5 -	Water	Y	Qu et al. (2018)
	Cd ²⁺				0.5µM	500µM			
S:A stoichion	netric ratic								

Table 3.5 List of characteristics of nanosensors with AIE properties developed for detection of multiple heavy metals

LOD lowest limit of detection, THF tetrahydrofuran, DMF dimethylformamide; Y means successful practical application in real water samples

3.10 Conclusions and Future Perspectives

For the purpose of guaranteeing environmental and human safety, it is critical and even urgent to develop the sensitive and selective sensors for detecting heavy metals. Nanomaterial-based sensors have achieved high sensitivity due to their large surface-to-volume ratio, high surface activity, and size-dependent property. In this chapter, we have summarized comprehensively and compared various AIE-based nanosensors developed in recent years with a focus on the types of nanomaterials and the related detecting mechanisms. These nanosensors provide stable and strong fluorescence and high sensitivity even at nanomolar concentration of metal ions, which is far below the minimum of environmental needs. Developing nanosensors, including organic nanoparticles, metallic nanoclusters or nanoparticles, metalorganic frameworks, hydrogen-bonded organic frameworks, and carbon dots, has grown noticeably and become a hot research area. Additionally, combining AIE material with other nanomaterials or even utilizing multiple AIE-based nanomaterials to construct a sensing system appears as an outbreak trend for heavy metal ions detection. Meanwhile, sensitivity and selectivity have also been elevated in numerous sensor systems reported. The sensing mechanisms mainly comprise two broad categories, fluorescence enhancement and quenching. Specifically, the addition of heavy metals often causes the sensor molecules to aggregate thereby turning on the fluorescence or disrupts the conjugated system to weaken or quench the fluorescence of the metallic NCs/NPs via metallophilic interaction. As to the AIE nanosensors based on fluorescence enhancement, by contrast, it exhibits lower detection limits and higher accuracy. Furthermore, some fluorescent nanosensors acting in ratiometric behavior can provide higher sensitivity and better selectivity, which makes them promising for real application.

For further design of AIE-based nanosensors for detecting heavy metals, the following trends are proposed: (1) to develop multifunctional sensors for simultaneously detecting multiple heavy metals; (2) to develop *all-in-one* sensors by combing multiple sensing signals to improve accuracy; (3) to develop *theranostic* sensors to achieve efficiently detection and the removal of heavy metals at the same time; and (4) to develop point-of-care testing sensor by integrating the AIE nanosensors into portable devices to provide novel tools in practical applications. We believe that fluorescent AIE-based nanosensors for heavy metal ions will play an increasingly important role in environment science and life of nature.

Acknowledgments We are grateful for the financial supports from National Natural Science Foundation of China (81671756 and 81971678), Science and Technology Foundation of Hunan Province (2020GK3019, 2019GK5012 and 2019SK2211) and Jiangxi Province (20192ACB60012 and 20192BCD40046), and the Fundamental Research Funds for the Central South Universities (2018zzts877 and 2018zzts772).

References

- Aguilar C, Narayanan J, Shanmuganathan R, Ricardo CT, Castrillo JIA, Miranda MYP, Pérez KMA (2018) FONs of highly preorganized N,N'-bis(3-aminobenzyl)-5,8-diiminequinoline with aggregation induced emission enhancement and metal-chelation for selective Cd²⁺ detection. J Photochem Photobiol A Chem 367:1–12. https://doi.org/10.1016/J. JPHOTOCHEM.2018.08.008
- Aragay G, Pons J, Merkoci A (2011) Recent trends in macro-, micro-, and nanomaterial-based tools and strategies for heavy-metal detection. Chem Rev 111(5):3433–3458. https://doi. org/10.1021/cr100383r
- Baldini L, Casnati A, Sansone F, Ungaro R (2007) Calixarene-based multivalent ligands. Chem Soc Rev 36(2):254–266. https://doi.org/10.1039/B603082N
- Bashir A, Malik LA, Ahad S, Manzoor T, Bhat MA, Dar GN, Pandith AH (2019) Removal of heavy metal ions from aqueous system by ion-exchange and biosorption methods. Environ Chem Lett 17:729–754. https://doi.org/10.1007/s10311-018-00828-y
- Bhalla V, Roopa KM, Sharma PR, Kaur T (2013) Hg²⁺ induced hydrolysis of pentaquinone based Schiff base: a new chemodosimeter for Hg²⁺ ions in mixed aqueous media. Dalton Trans 42:15063–15068. https://doi.org/10.1039/C3DT51273H
- Carol P, Sreejith S, Ajayaghosh A (2007) Ratiometric and near-infrared molecular probes for the detection and imaging of zinc ions. Chem Asian J 2:338–348. https://doi.org/10.1002/ asia.200600370
- Chen YC, Zhang WJ, Cai YJ, Kwok RTK, Hu YB, Lam JWY, Gu XG, He ZK, Zhao Z, Zheng XY, Chen B, Gui C, Tang BZ (2017) AIEgens for dark through-bond energy transfer: design, synthesis, theoretical study and application in ratiometric Hg²⁺ sensing. Chem Sci 8(3):2047–2055. https://doi.org/10.1039/C6SC04206F
- Cheng XH, Li QQ, Qin J, Li Z (2010) A new approach to design ratiometric fluorescent probe for mercury (II) based on the Hg²⁺-promoted deprotection of thioacetals. ACS Appl Mater Interfaces 2(4):1066–1072. https://doi.org/10.1021/am900840q
- Choi MG, Cha S, Lee H, Jeona HL, Chang SK (2009) Sulfide-selective chemosignaling by a Cu²⁺ complex of dipicolylamine appended fluorescein. Chem Commun 49:7390–7392. https://doi.org/10.1039/B916476F
- Cui L, Wu J, Ju HX (2015) Electrochemical sensing of heavy metal ions with inorganic, organic and bio-materials. Biosens Bioelectron 63:276–286. https://doi.org/10.1016/j.bios.2014.07.052
- Cullen WR, Reimer KJ (1989) Arsenic speciation in the environment. Chem Rev 89(4):713–764. https://doi.org/10.1021/cr00094a002
- Dasgupta M, Nag S, Das G, Nethaji M, Bhattacharya S (2008) N,N'-Bis(aryl)pyridine-2,6dicarboxamide complexes of ruthenium: synthesis, structure and redox properties. Polyhedron 27:139–150. https://doi.org/10.1016/j.poly.2007.08.042
- Dasgupta N, Ranjan S, Ramalingam C (2017) Applications of nanotechnology in agriculture and water quality management. Environ Chem Lett 15(4):591–605. https://doi.org/10.1007/ s10311-017-0648-9
- De Acha N, Elosua C, Corres JM, Arregui FJ (2019) Fluorescent sensors for the detection of heavy metal ions in aqueous media. Sensors 19(3):599–633. https://doi.org/10.3390/s19030599
- El-Sherif AA, Aljahdali MS (2013) Review: protonation, complex-formation equilibria, and metal–ligand interaction of salicylaldehyde Schiff bases. J Coord Chem 66(19):3423–3468. https://doi.org/10.1080/00958972.2013.839027
- Feng HT, Song S, Chen YC, Shen CH, Zheng YS (2014) Self-assembled tetraphenylethylene macrocycle nanofibrous materials for the visual detection of copper (II) in water. J Mater Chem C 2:2353–2359. https://doi.org/10.1039/C3TC32373K
- Flora SD, Bagnasco M, Serra D, Zanacchi P (1990) Genotoxicity of chromium compounds: a review. Mutat Res 238:99–172. https://doi.org/10.1016/0165-1110(90)90007-X

- Fournier-Salaün MC, Salaün P (2007) Quantitative determination of hexavalent chromium in aqueous solutions by UV-Vis spectrophotometer. Cent Eur J Chem 5:1084–1093. https://doi.org/10.2478/s11532-007-0038-4
- Gao M, Tang BZ (2017) Fluorescent sensors based on aggregation-induced emission: recent advances and perspectives. ACS Sensors 2:1382–1399. https://doi.org/10.1021/ acssensors.7b00551
- Gao YY, Ma TT, Ou ZZ, Cai WJ, Yang GQ, Li Y, Xu MH, Li QQ (2018) Highly sensitive and selective turn-on fluorescent chemosensors for Hg²⁺ based on thioacetal modified pyrene. Talanta 178:663–669. https://doi.org/10.1016/j.talanta.2017.09.089
- Gao T, Huang XY, Huang S, Dong J, Yuan K, Feng XP, Liu TT, Yu KQ, Zeng WB (2019) Sensitive water-soluble fluorescent probe based on umpolung and aggregation-induced emission strategies for selective detection of Hg²⁺ in living cells and zebrafish. J Agric Food Chem 67(8):2377–2383. https://doi.org/10.1021/acs.jafc.8b06895
- Geng L, Zhao Y, Kamya E, Guo JT, Sun B, Feng YK, Zhu MF, Ren XK (2019) Turn-off/on fluorescent sensors for Cu²⁺ and ATP in aqueous solution based on a tetraphenylethylene derivative. J Mater Chem C 7:2640–2645. https://doi.org/10.1039/C8TC06075D
- Glaser T, Heidemeier M, Strautmann JBH, Bçgge H, Stammler A, Krickemeyer E, Huenerbein R, Grimme S, Bothe E, Bill E (2007) Trinuclear copper complexes with triplesalen ligands: geometric and electronic effects on ferromagnetic coupling via the spin-polarization mechanism. Chem Eur J 13:9191–9206. https://doi.org/10.1002/chem.200700781
- Gu XG, Zhang XY, Ma HL, Jia SR, Zhang PF, Zhao YJ, Liu Q, Wang JG, Zheng XY, Lam J, Ding D, Tang BZ (2018) Corannulene-incorporated AIE nanodots with highly suppressed nonradiative decay for boosted cancer phototheranostics in vivo. Adv Mater 30(26):1801065. https:// doi.org/10.1002/adma.201801065
- Guo YH, Tong XY, Hu JM, Pei RJ (2015) Visual detection of Ca²⁺ based on aggregation-induced emission of Au(I)–Cys complexes with superb selectivity. Chem Commun 51:596–598. https:// doi.org/10.1039/C4CC07592G
- Harada A, Takashima Y, Yamaguchi H (2009) Cyclodextrin-based supramolecular polymers. Chem Soc Rev 38(4):875–882. https://doi.org/10.1039/B705458K
- Hola K, Zhang Y, Wang Y, Giannelis EP, Zboril R, Rogach AL (2014) Carbon dots-emerging light emitters for bioimaging, cancer therapy and optoelectronics. Nano Today 9(5):590–603. https://doi.org/10.1016/j.nantod.2014.09.004
- Hong YN, Lam J, Tang BZ (2009) Aggregation-induced emission: phenomenon, mechanism and applications. Chem Commun 29:4332–4353. https://doi.org/10.1039/b904665h
- Huang G, Wen R, Wang Z, Li BS, Tang BZ (2018) Novel chiral aggregation induced emission molecules: self-assembly, circularly polarized luminescence and copper (II) ion detection. Mater Chem Front 2:1884–1892. https://doi.org/10.1039/C8QM00294K
- Huang LT, Li SW, Ling X, Zhang J, Qin AJ, Zhuang J, Gao M, Tang BZ (2019) Dual detection of bioaccumulated Hg²⁺ based on luminescent bacteria and aggregation-induced emission. Chem Commun 55:7458–7461. https://doi.org/10.1039/c9cc02782c
- Huo FJ, Yin CX, Yang YT, Su J, Chao JB, Liu DS (2012) Ultraviolet-Visible light (UV–Vis)reversible but fluorescence-irreversible chemosensor for copper in water and its application in living cells. Anal Chem 84:2219–2223. https://doi.org/10.1021/ac202734m
- Islam MM, Hu Z, Wang Q, Redshaw C, Feng X (2019) Pyrene-based aggregation-induced emission luminogens and their applications. Mater Chem Front 3:762–781. https://doi.org/10.1039/ C9QM00090A
- Izah SC, Chakrabarty N, Srivastav AL (2016) A review on heavy metal concentration in potable water sources in Nigeria: human health effects and mitigating measures. Expos Health 8:285–304. https://doi.org/10.1007/s12403-016-0195-9
- Jackson BP, Taylor VF, Karagas MR, Punshon T, Cottingham KL (2012) Arsenic, organic foods, and brown rice syrup. Environ Health Perspect 120:623–626. https://doi.org/10.1289/ ehp.1104619

- Jarup L (2003) Hazards of heavy metal contamination. Br Med Bull 68:167–182. https://doi. org/10.1093/bmb/ldg032
- Ji L, Guo Y, Hong S, Wang Z, Wang K, Chen X, Zhang J, Hu J, Pei R (2015) Label-free detection of Pb²⁺ based on aggregation-induced emission enhancement of Au-nanoclusters. RSC Adv 5:36582–36586. https://doi.org/10.1039/C5RA03449C
- Jia J, Tang X, He YF, Zhang MY, Xing GW (2012) Recent progress in fluorescent probes for Zn²⁺/Cd²⁺ based on small organic molecules. Chin J Org Chem 32:1803–1811. https://doi.org/10.6023/cjoc1203103
- Jiang YS, He T, Chen YC, Ruan YL, Zhou YB, Tang BZ, Qin JG, Tang YH (2017) Quantitative evaluation and in vivo visualization of mercury ion bioaccumulation in rotifers by novel aggregation-induced emission fluorogen nanoparticles. Environ Sci Nano 4(11):2186–2192. https://doi.org/10.1039/c7en00599g
- Jiang R, Liu N, Li F, Fu W, Zhou Y, Zhang Y (2018) Novel PSMA-coated on-off-on fluorescent chemosensor based on organic dots with AIEgens for detection of copper (II), iron (III) and cysteine. Polymers 10:786–795. https://doi.org/10.3390/polym10070786
- Jiang Y, Duan QY, Zheng GS, Yang L, Zhang J, Wang YF, Zhang HT, He J, Sun HY, Ho D (2019) An ultra-sensitive and ratiometric fluorescent probe based on the DTBET process for Hg²⁺ detection and imaging applications. Analyst 144(4):1353–1360. https://doi.org/10.1039/ c8an02126k
- Jung SH, Kwon KY, Jung JH (2015) A turn-on fluorogenic Zn (II) chemoprobe based on a terpyridine derivative with aggregation-induced emission (AIE) effects through nanofiber aggregation into spherical aggregates. Chem Commun 51:952–955. https://doi.org/10.1039/C4CC05283H
- Kasana RC, Panwar NR, Kaul RK, Kumar P (2017) Biosynthesis and effects of copper nanoparticles on plants. Environ Chem Lett 15(2):233–240. https://doi.org/10.1007/s10311-017-0615-5
- Khandare DG, Joshi H, Banerjee M, Majik MS, Chatterjee A (2014) An aggregation-induced emission based "turn-on" fluorescent chemodosimeter for the selective detection of Pb²⁺ ions. RSC Adv 4:47076–47080. https://doi.org/10.1039/C4RA09451D
- Kong FY, Zhang JW, Li RF, Wang ZX, Wang WJ, Wang W (2017) Unique roles of gold nanoparticles in drug delivery, targeting and imaging applications. Molecules 22(9):1445–1458. https:// doi.org/10.3390/molecules22091445
- Kumar P, Kim KH, Bansal V, Lazarides T, Kumar N (2017) Progress in the sensing techniques for heavy metal ions using nanomaterials. J Ind Eng Chem 54:30–43. https://doi.org/10.1016/j. jiec.2017.06.010
- Kuppan B, Maitra U (2017) Instant room temperature synthesis of self-assembled emissiontunable gold nanoclusters: million-fold emission enhancement and fluorimetric detection of Zn²⁺. Nanoscale 9:15494–15504. https://doi.org/10.1039/C7NR05659A
- Li N, Feng H, Gong Q, Wu C, Zhou H, Huang Z, Yang J, Chen X, Zhao N (2015a) BINOL-based chiral aggregation-induced emission luminogens and their application in detecting copper (II) ions in aqueous media. J Mater Chem C 3:11458–11463. https://doi.org/10.1039/c5tc02778k
- Li D, Li B, Yang SI (2015b) A selective fluorescence turn-on sensing system for evaluation of Cu²⁺ polluted water based on ultrafast formation of fluorescent copper nanoclusters. Anal Methods 7:2278–2282. https://doi.org/10.1039/c5ay00219b
- Li PF, Liu YY, Zhang WJ, Zhao N (2017) A fluorescent probe for pyrophosphate based on tetraphenylethylene derivative with aggregation-induced emission characteristics. ChemistrySelect 2:3788–3793. https://doi.org/10.1002/slct.201700302
- Li Y, Hu X, Zhang X, Cao H, Huang Y (2018) Unconventional application of gold nanoclusters/ Zn-MOF composite for fluorescence turn-on sensitive detection of zinc ion. Anal Chim Acta 1024:145–152. https://doi.org/10.1016/J.ACA.2018.04.016
- Lin SM, Geng S, Li N, Li NB, Luo HQ (2016) D-penicillamine-templated copper nanoparticles via ascorbic acid reduction as a mercury ion sensor. Talanta 151:106–113. https://doi. org/10.1016/j.talanta.2016.01.028
- Lin L, Hu Y, Zhang L, Huang Y, Zhao S (2017a) Photoluminescence light-up detection of zinc ion and imaging in living cells based on the aggregation induced emission enhancement

of glutathione-capped copper nanoclusters. Biosens Bioelectron 94:523-529. https://doi.org/10.1016/J.BIOS.2017.03.038

- Lin N, Zhang Q, Xia X, Liang M, Zhang S, Zheng L, Cao Q, Ding Z (2017b) A highly zincselective ratiometric fluorescent probe based on AIE luminogen functionalized coordination polymer nanoparticles. RSC Adv 7:21446–21451. https://doi.org/10.1039/C6RA28551A
- Liu M, Tan H, Liu Z, Wang W, Zeng W (2013) Advances in fluorescent probes based on the small molecules for Zn²⁺. Chin J Org Chem 33:1655–1667. https://doi.org/10.6023/cjoc201301015
- Liu G, Yang M, Wang L, Zheng J, Zhou H, Wu J, Tian Y (2014) Schiff base derivatives containing heterocycles with aggregation-induced emission and recognition ability. J Mater Chem C 2:2684–2691. https://doi.org/10.1039/C3TC32591A
- Liu H, Wei R, Xiang Y, Tong A (2015) Fluorescence turn-on detection of pyrophosphate based on aggregation-induced emission property of 5-chlorosalicylaldehyde azine. Anal Methods 7:753–758. https://doi.org/10.1039/C4AY02341B
- Liu B, Zhou H, Yang B, Hu X (2017) Aggregation-induced emission activity and further Cu²⁺ induced self-assembly process of two Schiff compounds. Sensors Actuators B Chem 246:554–562. https://doi.org/10.1016/j.snb.2017.02.120
- Liu BB, He W, Lu H, Wang K, Huang MM, Kwok R, Lam J, Gao LC, Yang JP, Tang BZ (2019a) A facile design for multifunctional AIEgen based on tetraaniline derivatives. Sci China Chem 62:732–738. https://doi.org/10.1007/s11426-018-9440-0
- Liu X, Yu Y, Lin B, Cao Y, Guo M (2019b) A label-free fluorescent probe for the detection of adenosine 5'-triphosphate via inhibiting the aggregation-induced emission enhancement of glutathione modified silver nanoclusters triggered by zinc ion. Spectrochim Acta A 214:360–365. https://doi.org/10.1016/J.SAA.2019.02.040
- Lu Z, Wu M, Wu S, Yang S, Li Y, Liu X, Zheng L, Cao Q, Ding Z (2016) Modulating the optical properties of the AIE fluophor confined within UiO-66's nanochannels for chemical sensing. Nanoscale 8:17489–17495. https://doi.org/10.1039/C6NR05600H
- Luo J, Xie Z, Lam JWY, Cheng L, Chen HY, Qiu CF, Kwok HS, Zhan XW, Liu YQ, Zhu DB, Tang BZ (2001) Aggregation-induced emission of 1-methyl-1,2,3,4,5-pentaphenylsilole. Chem Commun 18:1740–1741. https://doi.org/10.1039/B105159H
- Mahajan PG, Bhopate DP, Kolekar GB, Patil SR (2015) N-methyl isatin nanoparticles as a novel probe for selective detection of Cd²⁺ ion in aqueous medium based on chelation enhanced fluorescence and application to environmental sample. Sensors Actuators B Chem 220:864–872. https://doi.org/10.1016/J.SNB.2015.05.119
- Mandal BK, Suzuki KT (2002) Arsenic round the world: a review. Talanta 58(1):201–235. https:// doi.org/10.1016/S0039-9140(02)00268-0
- Mei J, Hong YN, Lam JWY, Qin AJ, Tang YH, Tang BZ (2014) Aggregation-induced emission: the whole is more brilliant than the parts. Adv Mater 26:5429–5479. https://doi.org/10.1002/ adma.201401356
- Mohan D, Pittman CU, Bricka M, Smith F, Yancey B, Mohammad J, Steele PH, Alexandre-Franco MF, Gomez-Serrano V, Gong H (2007) Sorption of arsenic, cadmium, and lead by chars produced from fast pyrolysis of wood and bark during bio-oil production. J Colloid Interface Sci 310:57–73. https://doi.org/10.1016/j.jcis.2007.01.020
- Ni JK, Li QY, Li B, Zhang LM (2013) A novel fluorescent probe based on rhodamine B derivative for highly selective and sensitive detection of mercury(II) ion in aqueous solution. Sensors Actuators B Chem 186:278–285. https://doi.org/10.1016/j.snb.2013.06.011
- Nickens KP, Patierno SR, Ceryak S (2010) Chromium genotoxicity: a double-edged sword. Chem Biol Interact 188:276–288. https://doi.org/10.1016/j.cbi.2010.04.018
- Nie KX, Xu S, Duan XY, Shi HH, Dong B, Long MQ, Xu H, Jiang XF, Liu ZC (2018) Diketopyrrolopyrrole-doped hybrid FONs as two-photon absorbing and dual-emission fluorescent nanosensors for Hg²⁺. Sensors Actuators B Chem 265:1–9. https://doi.org/10.1016/j. snb.2018.03.026

- Niu CX, Liu QL, Shang ZH, Zhao L, Ouyang J (2015) Dual-emission fluorescent sensor based on AIE organic nanoparticles and au nanoclusters for the detection of mercury and melamine. Nanoscale 7:8457–8465. https://doi.org/10.1039/C5NR00554J
- Ogoshi T, Yamagishi TA, Nakamoto Y (2016) Pillar-shaped macrocyclic hosts pillar[*n*]arenes: new key players for supramolecular chemistry. Chem Rev 116(14):7937–8002. https://doi. org/10.1021/acs.chemrev.5b00765
- Oliveira E, Nuñez C, Rodríguez-González B, Capelo JL, Lodeiro C (2011) Novel small stable gold nanoparticles bearing fluorescent cysteine-coumarin probes as new metal-modulated chemosensors. Inorg Chem 50(18):8797–8807. https://doi.org/10.1021/ic200664z
- Pan SF, Liu WD, Tang JT, Yang YJ, Feng H, Qian ZS, Zhou J (2018) Hydrophobicity-guided selfassembled particles of silver nanoclusters with aggregation-induced emission and their use in sensing and bioimaging. J Mater Chem B 6:3927–3933. https://doi.org/10.1039/c8tb00463c
- Pannipara M, Al-Sehemi AG, Irfan A, Assiri M, Kalam A, Al-Ammari YS (2018) AIE active multianalyte fluorescent probe for the detection of Cu²⁺, Ni²⁺ and Hg²⁺ ions. Spectrochim Acta Part A Mol Biomol Spectrosc 201:54–60. https://doi.org/10.1016/j.saa.2018.04.052
- Pathan S, Jalal M, Prasad S, Bose S (2019) Aggregation-induced enhanced photoluminescence in magnetic graphene oxide quantum dots as a fluorescence probe for As (III) sensing. J Mater Chem C 7:8510–8520. https://doi.org/10.1039/C8TA11358K
- Qu F, Zhao L, Han W, You J (2018) Ratiometric detection of Zn²⁺ and Cd²⁺ based on self-assembled nanoarchitectures with dual emissions involving aggregation enhanced emission (AEE) and its application. J Mater Chem B 6:4995–5002. https://doi.org/10.1039/c8tb01046c
- Saha K, Agasti SS, Kim C, Li XN, Rotello VM (2012) Gold nanoparticles in chemical and biological sensing. Chem Rev 112(5):2739–2779. https://doi.org/10.1021/cr2001178
- Salnikow K, Zhitkovich A (2008) Genetic and epigenetic Mechanisms in metal carcinogenesis and cocarcinogenesis: nickel, arsenic, and chromium. Chem Res Toxicol 21:28–44. https://doi. org/10.1021/tx700198a
- Samanta S, Manna U, Raya T, Das G (2015) An aggregation-induced emission (AIE) active probe for multiple targets: a fluorescent sensor for Zn²⁺ and Al³⁺ & a colorimetric sensor for Cu²⁺ and F⁻. Dalton Trans 44:18902–18910. https://doi.org/10.1039/C5DT03186A
- Schubert US, Eschbaumer C (2002) Macromolecules containing bipyridine and terpyridine metal complexes: towards metallosupramolecular polymers. Angew Chem Int Ed 41:2892–2926. https://doi.org/10.1002/chin.200246226
- Sharma S, Virk TS, Pradeep CP, Dhir A (2017) ESIPT-induced carbazole-based AIEE material for nanomolar detection of Cu²⁺ and CN⁻ ions: a molecular keypad security device. Eur J Inorg Chem 18:2457–2463. https://doi.org/10.1002/ejic.201700091
- Shi W, Zhao SY, Su Y, Hui YH (2016) Barbituric acid-triphenylamine adduct as an AIEE-type molecule and optical probe for mercury(II). New J Chem 40(9):7814–7820. https://doi. org/10.1039/C6NJ00894A
- Singh PK, Prabhune A, Ogale S (2015) Pulsed laser-driven molecular self-assembly of cephalexin: aggregation-induced fluorescence and its utility as a mercury ion sensor. Photochem Photobiol 91:1340–1347. https://doi.org/10.1111/php.12526
- Singh P, Singh VV, Sharma R, Bhargava G, Kumar S (2016) Nanomolar Cu²⁺ detection in water based on disassembly of AIEgen: applications in blood serum, cell imaging and complex logic circuits. ChemistrySelect 1:6880–6887. https://doi.org/10.1002/slct.201601497
- Smedley PL, Kinniburgh DG (2002) A review of the source, behaviour and distribution of arsenic in natural waters. Appl Geochem 17(5):517–568. https://doi.org/10.1016/S0883-2927(02)00018-5
- Soodan RK, Pakade YB, Nagpal A, Katnoria JK (2014) Analytical techniques for estimation of heavy metals in soil ecosystem: a tabulated review. Talanta 125:405–410. https://doi. org/10.1016/j.talanta.2014.02.033
- Srivastava AK, Dev A, Karmakar S (2018) Nanosensors and nanobiosensors in food and agriculture. Environ Chem Lett 16:161–182. https://doi.org/10.1007/s10311-017-0674-7
- Stohs SJ, Bagchi D (1995) Oxidative mechanisms in the toxicity of metal ions. Free Radic Biol Med 18:321–336. https://doi.org/10.1016/0891-5849(94)00159-H

- Sun J, Qian Y (2016) A novel naphthalimide-rhodamine fluorescence sensor: synthesis, aggregation-induced emission enhancement and its dual-channel detection property. Chin J Org Chem 36:151–157. https://doi.org/10.6023/cjoc201506022
- Swiegers GF, Malefetse TJ (2000) New self-assembled structural motifs in coordination chemistry. Chem Rev 100(9):3483–3538. https://doi.org/10.1021/cr990110s
- Tang C, Feng H, Qian ZS (2017) Reversible luminescent nanoswitches based on aggregationinduced emission enhancement of silver nanoclusters for luminescence turn-on assay of inorganic pyrophosphatase activity. Anal Chem 89(9):4994–5002. https://doi.org/10.1021/acs. analchem.7b00319
- Tchounwou PB, Yedjou CG, Patlolla AK, Sutton DJ (2012) Heavy metals toxicity and the environment. EXS 101:133–164. https://doi.org/10.1007/978-3-7643-8340-4_6
- Tian X, Chen L, Li Y, Yang C, Nie Y, Zhou C, Wang Y (2017) Design and synthesis of a molecule with aggregation-induced emission effects and its application in the detection of arsenite in groundwater. J Mater Chem C 5:3669–3672. https://doi.org/10.1039/C7TC00363C
- Tian Y, Kelarakis A, Li L, Zhao F, Wang Y, Wang W, Yang Q, Ye Z, Guo X (2018) Facile fluorescence "turn on" sensing of lead ions in water via carbon nanodots immobilized in spherical polyelectrolyte brushes. Front Chem 6:470–483. https://doi.org/10.3389/FCHEM.2018.00470
- Tomar K, Kaur G, Verma S, Ramanathan G (2018) A self-assembled tetrapeptide that acts as a "turn-on" fluorescent sensor for Hg²⁺ ion. Tetrahedron Lett 59:3653–3656. https://doi.org/10.1016/j.tetlet.2018.08.043
- Tiwari K, Kumar S, Kumar V, Kaur J, Arora S, Mahajan RK (2018) An azine based sensor for selective detection of Cu 2+ ions and its copper complex for sensing of phosphate ions in physiological conditions and in living cells. Spectrochim Acta A 191:16–26. https://doi. org/10.1016/J.SAA.2017.09.072
- Ullah N, Mansha M, Khan I, Qurashi A (2018) Nanomaterial-based optical chemical sensors for the detection of heavy metals in water: recent advances and challenges. Trends Anal Chem 100:155–166. https://doi.org/10.1016/j.trac.2018.01.002
- Vincent JB (2000) Quest for the molecular mechanism of chromium action and its relationship to diabetes. Nutr Rev 58:67–72. https://doi.org/10.1111/j.1753-4887.2000.tb01841.x
- Wang L, Li H, Cao D (2013) A new photoresponsive coumarin-derived Schiff base: chemosensor selectively for Al³⁺ and Fe³⁺ and fluorescence "turn-on" under room light. Sensors Actuators B Chem 181:749–755. https://doi.org/10.1016/j.snb.2013.01.090
- Wang W, Cheng L, Liu WG (2014) Biological applications of carbon dots. Sci China Chem 57(4):522–539. https://doi.org/10.1007/s11426-014-5064-4
- Wang P, Wu J, Zhou PP, Liu W, Tang Y (2015a) A novel peptide-based fluorescent chemosensor for measuring zinc ions using different excitation wavelengths and application in live cell imaging. J Mater Chem B 3(17):3617–3624. https://doi.org/10.1039/C5TB00142K
- Wang AZ, Yang YX, Yu FF, Xue LW, Hu BW, Fan WP, Dong YJ (2015b) A highly selective and sensitive fluorescent probe for quantitative detection of Hg²⁺ based on aggregation-induced emission features. Talanta 132:864–870. https://doi.org/10.1016/j.talanta.2014.10.048
- Wang X, Gao ZC, Zhu JL, Gao Z, Wang F (2016) Aggregation induced emission of a cyanostilbene amphiphile as a novel platform for FRET-based ratiometric sensing of mercury ions in water. Polym Chem 7(33):5217–5220. https://doi.org/10.1039/C6PY01109H
- Wang K, Li JJ, Ji SM, Li LJ, Qiu ZP, Pan CQ, Zhang JY, Huo YP (2018a) Fluorescence probes based on AIE luminogen: application for sensing Hg²⁺ in aqueous media and cellular imaging. New J Chem 42(16):13836–13846. https://doi.org/10.1039/C8NJ02245C
- Wang ZM, Zhou F, Gui C, Wang J, Zhao ZJ, Qin AJ, Tang BZ (2018b) Selective and sensitive fluorescent probes for metal ions based on AIE dots in aqueous media. J Mater Chem C 6:11261–11265. https://doi.org/10.1039/c8tc03711f
- Wang A, Fan RQ, Zhou YZ, Zheng XB, Zhou XS, Hao S, Yang YL (2019a) Multiple-color aggregation-induced emission-based Schiff base sensors for ultrafast dual recognition of Hg²⁺ and pH integrating Boolean logic operations. J Coord Chem 72(1):102–118. https://doi.org/1 0.1080/00958972.2018.1546851

- Wang L, Li Y, Tian X, Yang C, Lu L, Zhou Z, Huang Y, Nie Y (2019b) Construction of salicylaldehyde analogues as turn-on fluorescence probes and their electronic effect on sensitive and selective detection of As (V) in groundwater. Anal Methods 11:955–964. https://doi. org/10.1039/c8ay02484g
- Wei G, Jiang Y, Wang F (2018) A novel AIEE polymer sensor for detection of Hg²⁺ and Ag⁺ in aqueous solution. J Photochem Photobiol A Chem 358:38–43. https://doi.org/10.1016/j. jphotochem.2018.03.006
- Wilson R (2008) The use of gold nanoparticles in diagnostics and detection. Chem Soc Rev 37(9):2028–2045. https://doi.org/10.1039/B712179M
- Wu G, Kang HB, Zhang XY, Shao HB, Chu LY, Ruan CJ (2010) A critical review on the bio-removal of hazardous heavy metals from contaminated soils: issues, progress, eco-environmental concerns and opportunities. J Hazard Mater 174:1–8. https://doi. org/10.1016/j.jhazmat.2009.09.113
- Xie JP, Zheng YG, Ying JY (2010) Highly selective and ultrasensitive detection of Hg²⁺ based on fluorescence quenching of Au nanoclusters by Hg²⁺-Au⁺ interactions. Chem Commun 46(6):961–963. https://doi.org/10.1039/B920748A
- Xu Y, Li HY, Wang B, Liu HC, Zhao L, Zhou TY, Liu MT, Huang N, Li Y, Ding L, Chen YH (2018) Microwave-assisted synthesis of carbon dots for "turn-on" fluorometric determination of Hg(II) via aggregation-induced emission. Microchim Acta 185:252. https://doi.org/10.1007/ s00604-018-2781-y
- Xu ZH, Wang Y, Wang Y, Li JY, Luo WF, Wu WN, Fan YC (2019) AIE active salicylaldehydebased hydrazone: a novel single-molecule multianalyte (Al³⁺ or Cu²⁺) sensor in different solvents. Spectrochim Acta A 212:146–154. https://doi.org/10.1016/J.SAA.2019.01.003
- Yang M, Zhang Y, Zhu W, Wang H, Jing H, Cheng L, Zhou H, Wu J, Tian Y (2015) Difunctional chemosensor for Cu (II) and Zn (II) based on Schiff base modified anthryl derivative with aggregation-induced emission enhancement and piezochromic characteristics. J Mater Chem C 3:1994–2002. https://doi.org/10.1039/c4tc02616k
- Yang J, Chai J, Yang B, Liu B (2019a) Achieving highly sensitive detection of Cu²⁺ based on AIE and FRET strategy in aqueous solution. Spectrochim Acta A 211:272–279. https://doi. org/10.1016/J.SAA.2018.12.020
- Yang L, Fang W, Ye YX, Wang ZK, Hu QL, Tang BZ (2019b) Redox-responsive fluorescent AIE bioconjugate with aggregation enhanced retention features for targeted imaging reinforcement and selective suppression of cancer cells. Mater Chem Front 3:1335–1340. https://doi. org/10.1039/c9qm00216b
- Yin YB, Coonrod CL, Heck KN, Lejarza F, Wong MS (2019a) Microencapsulated photoluminescent gold for ppb-level chromium (VI) sensing. ACS Appl Mater Interfaces 11:17491–17500. https://doi.org/10.1021/acsami.9b04699
- Yin H, Zhao B, Kan W, Liu T, Wang W, Yin G, Wang L, Gao Y, Wang J (2019b) Hydroxyl phenyl imino modified phenanthro[9,10-d] imidazole: an AIEE-active sensor for determination of Cu²⁺ in water samples and subsequent "turn-on" recognition of Cr³⁺ with logic gates. Spectrochim Acta Part A 217:18–26. https://doi.org/10.1016/J.SAA.2019.03.060
- You Y, Han Y, Lee YM, Park SY, Nam W, Lippard SJ (2011) Phosphorescent sensor for robust quantification of copper (II) ion. J Am Chem Soc 133(30):11488–11491. https://doi. org/10.1021/ja204997c
- Yu HX, Zhi J, Chang ZF, Shen T, Ding WL, Zhang X, Wang JL (2019) Rational design of aggregation-induced emission sensor based on Rhodamine B for turn-on sensing of trivalent metal cations, reversible data protection, and bioimaging. Mater Chem Front 3:151–160. https://doi.org/10.1039/C8QM00424B
- Zhang L, Fang M (2010) Nanomaterials in pollution trace detection and environmental improvement. Nano Today 5(2):128–142. https://doi.org/10.1016/j.nantod.2010.03.002
- Zhang H, Liu Q, Wang T, Yun Z, Li G, Liu J, Jiang G (2013a) Facile preparation of glutathionestabilized gold nanoclusters for selective determination of chromium (III) and chromium (VI)
in environmental water samples. Anal Chim Acta 770:140-146. https://doi.org/10.1016/j. aca.2013.01.042

- Zhang P, Pei L, Chen Y, Xu W, Lin Q, Wang J, Wu J, Shen Y, Ji L, Chao H (2013b) A dinuclear ruthenium (II) complex as a one - and two-photon luminescent probe for biological Cu²⁺ detection. Chem Eur J 19:15494–15503. https://doi.org/10.1002/chem.201302919
- Zhang XT, Lu XQ, Zhe YG, Liu J, Dong HL, Zhao GY, He P, Wang ZR, Jiang L, Hu WP (2014) Synthesis and aggregation-induced emissions of thienyl substituted cyclobutene derivatives. J Mater Chem C 2(26):5083–5086. https://doi.org/10.1039/C4TC00221K
- Zhang XY, Wang K, Liu MY, Zhang XQ, Tao L, Chen YW, Wei Y (2015a) Polymeric AIEbased nanoprobes for biomedical applications: recent advances and perspectives. Nanoscale 7(27):11486–11508. https://doi.org/10.1039/C5NR01444A
- Zhang KY, Yuan X, Kuah K, Luo ZT, Yao QF, Zhang QB, Xie JP (2015b) Boiling water synthesis of ultrastable thiolated silver nanoclusters with aggregation-induced emission. Chem Commun 51:15165–15168. https://doi.org/10.1039/C5CC04858C
- Zhang RX, Peng FL, Zhang WJ, Li N, Zhao N (2016) A highly sensitive fluorescent sensor with aggregation-induced emission characteristics for the detection of iodide and mercury ions in aqueous solution. J Mater Chem C 4(44):10479–10485. https://doi.org/10.1039/C6TC03696A
- Zhang X, Huang X, Gan X, Wu Z, Yu J, Zhou H, Tian Y, Wu J (2017a) Two multi-functional aggregation-induced emission probes: reversible mechanochromism and bio-imaging. Sensors Actuators B Chem 243:421–428. https://doi.org/10.1016/J.SNB.2016.11.161
- Zhang H, Wang S, Chen Z, Ge P, Jia R, Xiao E, Zeng W (2017b) A turn-on fluorescent nanoprobe for lead (II) based on the aggregation of weakly associated gold(I)-glutathione nanoparticles. Microchim Acta 184:4209–4215. https://doi.org/10.1007/S00604-017-2406-X
- Zhang H, Xu P, Zhang X, Cao X, Han W, Liu M, Liu X, Zeng W (2020) Au nanoparticles based ultra-fast "turn-on" fluorescent sensor for detection of biothiols and its application in living cell imaging. Chin Chem Lett. https://doi.org/10.1016/j.cclet.2019.10.005
- Zhao Z, Zheng XY, Du LL, Xiong Y, He W, Gao XX, Li CL, Liu YJ, Xu B, Zhang J, Song FY, Yu Y, Zhao XD, Cai YJ, He XW, Kwok R, Lam J, Huang XH, Phillips DL, Wang H, Tang BZ (2019) Non-aromatic annulene-based aggregation-induced emission system via aromaticity reversal process. Nat Commun 10:2952–2962. https://doi.org/10.1038/s41467-019-10818-5
- Zheng B, Wang F, Dong SY, Huang FH (2012) Supramolecular polymers constructed by crown ether-based molecular recognition. Chem Soc Rev 41(5):1621–1636. https://doi.org/10.1039/C1CS15220C
- Zhou H, Ye Q, Wu X, Song J, Cho CM, Zong Y, Tang BZ, Hor TSA, Yeow EKL, Xu J (2015) A thermally stable and reversible microporous hydrogen-bonded organic framework: aggregation induced emission and metal ion-sensing properties. J Mater Chem C 3:11874–11880. https:// doi.org/10.1039/c5tc02790j
- Zou Q, Li X, Zhang J, Zhou J, Suna B, Tian H (2012) Unsymmetrical diarylethenes as molecular keypad locks with tunable photochromism and fluorescence via Cu²⁺ and CN⁻ coordinations. Chem Commun 48:2095–2097. https://doi.org/10.1039/C2CC16942H

Chapter 4 Nanosensors Based on Lipid Films for Environmental Applications



Georgia-Paraskevi Nikoleli, Christina G. Siontorou, Marianna-Thalia Nikolelis, Spyridoula Bratakou, and Vasillios N. Psychoyios

Contents

4.1	Introd	uction	- 98
4.2	Metho	ds for Construction of Nanosensors Based on Lipid Membranes	100
	4.2.1	Metal-Supported Lipid Layers.	100
	4.2.2	Stabilized Lipid Films Formed on a Glass Fiber Filter	100
	4.2.3	Polymer-Supported Bilayer Lipid Membranes	100
	4.2.4	Polymeric Lipid Membranes Supported on Graphene Microelectrodes	100
4.3	Applications of Lipid-Film-Based Nanosensors for the Rapid Detection of		
	Environmental Pollutants.		102
4.4	Conclusions and Future Prospects.		
Refe	rences.		106

Abstract The recent advances in lipid membrane biosensors gave the ability to provide devices to detect trace of environmental pollutants in real samples. Nanotechnology provided sensors with increased selectivity and sensitivity, portability, ease of construction, rapid response times, etc. This chapter reviews recent advances in biosensor technologies for environmental pollutant monitoring. A large number of applications are described herein, clarifying some recent advances in the technology of these devices such as miniaturization, new sensing techniques, and nanotechnology-based transduction schemes. A wide range of environmental toxicants can be monitored and these compounds include toxins, hormones, diox-

C. G. Siontorou · M.-T. Nikolelis · V. N. Psychoyios

Laboratory of Simulation of Industrial Processes, Department of Industrial Management and Technology, School of Maritime and Industry, University of Piraeus, Piraeus, Greece

S. Bratakou

G.-P. Nikoleli (🖂)

Laboratory of Inorganic & Analytical Chemistry, School of Chemical Engineering, Dept 1, Chemical Sciences, National Technical University of Athens, Athens, Greece e-mail: dnikolel@chem.uoa.gr

Laboratory of Environmental & Sanitary Engineering, Department of Public Health, Faculty of Health and Caring Professions, University of West Attica, Athens, Attica, Greece

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_4

ins, PAHs, etc. As a result, the commercialization capabilities of these biosensors are given in the present chapter.

Keywords Lipid membrane biosensors · Environmental pollutants · Nanobiosensors · Environmental analysis · Nanotechnology

4.1 Introduction

A chemical sensor is a device that changes the concentration data of the convergence of a particular analyte into a scientifically helpful signal. Biosensors comprise of two parts: a chemical component ("receptor") and a physicochemical transducer. The acknowledgment framework deciphers the compound data (i.e., concentration of the analyte) into a quantifiable physical signal. The physical transducer gives the signal from the yield area of the component into an electrical, optical, or piezoelectric signal. A biosensor is an independent coordinated device that is fit for giving explicit quantitative diagnostic data utilizing an organic component (e.g., catalysts, antibodies, characteristic or fake receptors, and cells), which is held in direct spatial contact with a transduction component. Recent reports have appeared in the literature that describe the nanotechnological advances in biosensors (Pandit et al. 2009).

The early 1960s reconstitution of lipid bilayers has founded the "lipid membrane technology." The number of biosensors that were based on lipid membrane technology has increased during the last 20 years. During the last 10 years, the construction of stabilized lipid film biosensors was performed and the preparation of biosensors to monitor environmental pollutants in real samples and in the field was successful. These devices have a large number of advantages, which include biocompatibility, fast response times, a high sensitivity and selectivity, and their small size. The new class of stabilized lipid film nanobiosensors has the potentiality to provide analytical instrumentation with high selectivity, sensitivity, and stability.

Lipid film devices reduce the complexity of the surface interaction forces of membranes to well-defined surface chemistry (Fig. 4.1). The mechanism of signal generation still needs more investigations (Gould 2018); the investigations for drug permeability (Ebert et al. 2018) or protein–lipid interaction studies (Vacek et al. 2018) or even to manipulate processes on the lipid films by altering the macro-parameters are in a very good stage (Nikoleli et al. 2016). Biological "receptors" (enzymes, antibodies, receptors, ligands, DNA, etc.) can now be deposited on the lipid film surfaces (Mazur et al. 2017) or can be incorporated into the lipid structure (Vacek et al. 2018) by patterning (Raghunathan and Kenworthy 2018) or surface printing (Kalyankar et al. 2006). The lipid membrane represents a biocompatible structure for natural receptors to keep their function. The physical structure of the lipid film provides intrinsic signal transduction and amplification, which fits for biosensing—when the biological element interacts with the target analyte (Siontorou et al. 2017) providing current changes. Figure 4.1 shows the function and



Fig. 4.1 Simplified schematic of the function and composition of a biosensor based on lipid film technology, illustrating, also, the physicochemical mechanism for the signal recovery. (Source: Nikoleli et al. 2018)

composition of a biosensor based on lipid film technology, illustrating, also, the physicochemical mechanism for the transduction and other screening methods employed. A number of recent articles have appeared in the literature that mainly describe recent advances and applications of various platforms of lipid film-based biosensors (Nikoleli et al. 2016, 2018, 2019; Siontorou et al. 2017; Pandit et al. 2016; Aljerf and AlHamwi 2018).

4.2 Methods for Construction of Nanosensors Based on Lipid Membranes

4.2.1 Metal-Supported Lipid Layers

Tien and Salamon have reported in the literature a system for the construction of stabilized bilayer lipid membrane (s-BLM) at the end of Teflon-coated metallic wire that was freshly cut (Tien and Salamon 1989). This technique has been described in the literature in detail (Siontorou et al. 1997a, b; Nikolelis et al. 1996). Also, s-BLMs have been characterized (Siontorou et al. 1997a, b; Nikolelis et al. 1996). The diameter and composition of the wires are important for the time of the bilayer stabilization (Siontorou et al. 1997b; Nikolelis et al. 1996). A schematic setup for the s-BLM experimentation is shown in Fig. 4.2.

4.2.2 Stabilized Lipid Films Formed on a Glass Fiber Filter

The preparation of stabilized electrolyte lipid membranes was reported by Nikolelis' group and these membranes were constructed on GF/F glass microfiber (0.9 cm in diameter and 0.7 μ m nominal pore size) filters (Nikolelis et al. 1995; Andreou and Nikolelis 1998).

4.2.3 Polymer-Supported Bilayer Lipid Membranes

The use of a polymer-supported lipid film has been in practice to stabilize lipid membrane-based devices in the air for periods of over 1 month (Nikolelis and Mitrokotsa 2002).

The preparation of stable-in-air lipid membranes was reported by Nikolelis' group (Nikolelis and Mitrokotsa 2002; Nikolelis et al. 2006, 2008a, b). The lipids used were previously described. No receptor was used in these detections. The polymer stable-in-air lipid membranes were constructed as has been described in detail previously (Nikolelis and Mitrokotsa 2002; Nikolelis et al. 2006, 2008a, b).

4.2.4 Polymeric Lipid Membranes Supported on Graphene Microelectrodes

Our group has prepared an electrode that was composed from a lipid film on a copper wire that contained graphene nanosheets (Nikoleli et al. 2012). The experimental setup is shown in Fig. 4.3. These nanobiosensors were utilized for the fast



Fig. 4.2 Schematic of the sensor, measurement setup, and lipid self-assembly process (not drawn to scale). (a) The sensing electrode is tipped with a scalpel and immediately immersed in lipid solution before being dipped in the electrolyte solution. (b) The electrochemical setup consists of a 20-mL cell and a two-electrode configuration, that is, the sensing electrode and a Ag/AgCl reference electrode, placed in a grounded Faraday cage; an external DC potential of 25 mV is applied between the electrodes, and the ionic current through the BLM is measured with a digital electrometer; the cell is stirred using a magnetic stirrer. (c) Upon immersion, the lipid droplet attached to the wire is self-assembled into a bilayer that has one layer adsorbed on the metal surface and the other facing the aqueous solution. (d) Recording of the ion current decreases during the self-assembly process; recording started at the immersion of the sensing electrode in the electrolyte solution. (Source: Nikoleli et al. 2018)

monitoring of food and environmental toxicants (Nikoleli et al. 2012) such as urea, using the enzyme urease, cholesterol (using cholesterol oxidase), cholera toxin (using its receptor), and saxitoxin (using its receptor). The construction of graphene microelectrodes has been reported in the literature (Nikoleli et al. 2012).



Fig. 4.3 Schematic of the experimental setup; the micromachined chambers are separated by a thin (12.5 μ m thick) polyvinylidene chloride wrap and enclose the microfiber disk. (Source: Nikoleli et al. 2018)

4.3 Applications of Lipid-Film-Based Nanosensors for the Rapid Detection of Environmental Pollutants

Potentiometric and amperometric biosensors, which relied on modified gold electrodes, were constructed and used in pesticide analysis (Snejdarkova et al. 2004). Polyamidoamine (PAMAM) dendrimer (generation G4) was incorporated with 1-hexadecanethiol and used for the deposition of the following enzymes: acetylcholinesterase and choline oxidase for the construction of an amperometric device. Polyaniline doped with camphorsulfonic acid was utilized alternatively to have a potentiometric device. Trichlorfon, carbofuran, and eserine decreased the device response due to their inhibitory effects. The detection limits were on the following order: 0.003 and 200 nmol L^{-1} (trichlorfon), 0.04 and 6 nmol L^{-1} (carbofuran), and 0.1 and 700 nmol L^{-1} for amperometric and potentiometric devices, respectively.

A paper was reported in the literature using a synthetic "receptor" immobilized on supported lipid films on glass fiber filters. The supported lipid films were modified by calixarenes and proved adequate for the sensitive and rapid determination of various insecticides in fruit and vegetable samples (Nikolelis et al. 2008b). Other devices similarly developed include a disposable chemosensor for the selective and fast detection of food hormones such as naphthalene acetic acid (NAA) in fruits and vegetables (Nikolelis et al. 2008c) and a sensor for the detection of zinc in water (Nikolelis et al. 2009).

Bilayer lipid membranes (BLMs) were used as devices for the electrochemical detection of monocrotophos and carbofuran (Nikolelis and Krull 1994). Egg phosphatidylcholine (PC) and dipalmitoylphosphatidic acid (DPPA) were the lipids that were the basis of these detectors. A transient current signal of seconds was produced during interaction of these compounds with the lipid films. The sensitivity of the detectors was maximum when dipalmitoylphosphatidic acid was a component of the lipid mixture and by using calcium ions in the electrolyte solution. The peak heights of these transients were correlated to the concentration of monocrotophos or carbofuran in bulk solution with sub-micromolar detection limits.

An electrochemical device/minisensor for the rapid detection of ammonium ions was reported in the literature (Nikolelis et al. 1996), which was based on supported lipid membranes on metal wire. The ionophore that was used was gramicidin, and the lipid films were composed of egg phosphatidylcholine. The mechanical stability and longevity, sensitivity, and response times of this device were adequate, and the reversibility of this biosensor to ammonium ions was investigated and provided good results. The use of semisynthetic platelet-activating factor (PAF; 1-O-alkyl-2-acetyl-*sn*-gryceryl-3-phosphorylcholine, AGEPC) provided an increase of the biosensor and decreased the potassium interferences. The response times were rapid (to millisecond speeds) and the selectivity to ammonium ions in the presence of volatile amines was excellent. The analysis of small volumes of samples was possible using this sensor with detection limits on the order of ~1 × 10–6 M. Other advantages of this device were the uses of small volumes and that the sensor was disposable.

A device for the rapid detection of the triazine herbicides simazine, atrazine, and propazine was reported in the literature (Siontorou et al. 1997b). Egg phosphatidylcholine and dipalmitoylphosphatidic acid were the lipids that were used for the construction of the metal-supported lipid membranes. Ellipsometric data were used to explore the mechanism of signal generation. These triazines produced ion current increases within ~10 s after exposure of the lipid films to these compounds. The use of an anionic lipid such as dipalmitoylphosphatidic acid and calcium ions increased the magnitude of sensitivity. These magnitudes were linearly correlated to the insecticide, which could be detected at the nM concentration levels. The characteristics of the detection (i.e., mechanical stability, response times, and reproducibility) were excellent. These herbicides could be monitored in the presence of other insecticides and pesticides and the construction of the sensor could be easily performed and used in the field applications.

A report appeared in the literature that describes the changes in ion current through a lipid membrane with deposited DNA and it is the result of interactions of lipid film biosensors with hydrazines (Siontorou et al. 1998). These devices were based on the deposition of egg phosphatidylcholine on silver metal electrode. The DNAs were single stranded deoxyribonucleic acids: thymidylic acid icosanucleotide

that was terminated with a C-16 alkyl chain and deoxyadenylic acid icosanucleotide (dA_{20}) . These s-BLMs interact with hydrazines, and give a device to monitor parts per billion concentrations of hydrazines. These hydrazines included methylhydrazine, dimethylhydrazine, and phenylhydrazine.

A biosensor composed of supported polymeric lipid film with incorporated sheep anti-PCB antibody was described in the literature for the fast detection of Aroclor 1242 using flow injection analysis (FIA) (Michaloliakos et al. 2012). This antibody was deposited on the lipid film simultaneously with the lipid. Injections of polycyclic aromatic hydrocarbons (PAHs) were performed in the flowing solutions. A stoppedflow mode was used and the lipids were 15% (w/w) palmitic acid (PA) and 85% dipalmitoylphosphatidylcholine (DPPC); this lipid composition has provided one single signal. The peak height of the ion signal was correlated to the PAH concentration with detection limits at the nM concentration levels. The maximum number of cycles that could be performed was ca. 5.

The same principle was used to construct an electrochemical device for the monitoring of 3,3',4,4'-tetrachlorobiphenyl (PCB congener 77) (Nikolelis et al. 2013). The polymeric lipid membrane was composed of 85% (w/w) dipalmitoylphosphatidylcholine (DPPC) and 15% (w/w) dipalmitoylphosphatidic acid (DPPA), methacrylic acid, ethylene glycol dimethacrylate, azobisisobutyronitrile (AIBN), and sheep anti-congener 77 antiserum. A single transient signal was obtained and it was proportional to log (congener 77) from 10^{-8} to 10^{-5} M, with a detection limit of ca. 10^{-8} M. The maximum number of cycles that could be performed was ca. 5.

A carbon dioxide minisensor using hemoglobin as an ionophore and s-BLMs was reported in the literature (Siontorou et al. 1997a). The lipids used were egg phosphatidylcholine (egg PC) and dipalmitoylphosphatidic acid (DPPA). The parameters that were investigated included hemoglobin and DPPA concentration in BLMs; other factors exploited were pH of the electrolyte solution. The response times were of the order of 10s and the detection limits were ca. 0.4×10^{-6} M. The presence of semisynthetic platelet-activating factor (PAF; 1-*O*-alkyl-2-acetyl-*sn*-glyceryl-3-phosphorylcholine, AGEPC) in BLMs increased the sensitivity of the device. The device was found that could operate for periods of time of more than 48 h.

Shiratori et al. (2000) have suggested a gas biosensor composed of Langmuir-Blodgett (LB) membranes and fullerenes as spacers. The fullerenes provided a bilayer on a quartz crystal microbalance (QCM) electrode. PAHs were found to permeate more thoroughly through the membranes as molecular weight decreased, which was due to the interactions of the gas with the lipid film.

A biosensor appeared in the literature for the FIA analysis of hydrazine compounds and their mixtures (Siontorou et al. 2000). Filter-supported lipid films with incorporated DNA were used for the construction of this device. Hydrazines or their homologues were injected in the stream of a flowing electrolyte solution and ion current peaks were the result of the interactions of these hydrazines with BLMs. The signals were correlated with the concentration of hydrazines and the time appearance of these signals was different depending on the hydrophobicity of the analyte (i.e., the signal had longer time of appearance on the order of hydrazine, methylhydrazine or dimethylhydrazine, and phenylhydrazine).

Ishimori et al. (2002) suggested an environmental monitoring device that was constructed from lipid films. This device was able to continuously monitor underground water. A BLM device was proposed with an inkjet mechanism. Volatile organic chloride compounds (i.e., cis-1,2-dichloroethylene) could be monitored using this device with a sensitivity of the order of 10 ppb when using monoolein BLMs; these results were evaluated using real samples of underground water.

Novel electrochemical DNA aptasensors were developed on glassy carbon electrode that was altered using neutral red and polycarboxylated macrocyclic ligands polymerized onto the glassy carbon electrode (Evtugyn et al. 2014). The aptamers were deposited on the glassy carbon electrode using covalent immobilization techniques. The analyte was Aflatoxin B1 (AFB1). When AFB1 interacted with the electrode, the cathodic peak current was decreased and measured using cyclic voltametric and electrical impedance spectroscopy techniques. The detection limit was 0.1 nM for the cyclic voltametric method and 0.05 nM for impedance spectroscopy techniques. The aptasensor was used to monitor AFB1 in various samples of nuts, wine, and soy sauce.

A report was described in the literature for miniaturization of an electrochemical potentiometric naphthalene acetic acid (NAA) device on a graphene electrode in which a lipid membrane was deposited along with the auxin-binding protein 1 receptor (Bratakou et al. 2016). This sensor has given a good selectivity and sensitivity, fast response times, and detection limits for the hormone at 10 nM. The device could be easily constructed and has shown excellent reproducibility, reusability, and a slope of ca. 56 mV/decade of hormone concentration. An evaluation/validation of this device was made using real fruit and vegetable samples.

4.4 Conclusions and Future Prospects

The present review describes nanobiosensors that are based on lipid membranes for the fast detection and monitoring of biological compounds and toxicants such as herbicides, toxins, urea, and cholesterol and provides the advantages of lipid membrane devices (i.e., high sensitivity and selectivity, fast response times, and portability). It is of common sense that advances in nanotechnology will further provide biosensors with even improved characteristics.

References

- Aljerf L, AlHamwi B (2018) Carbon nanotubes synthesis developmental engineering demands will overcome the health challenge of nanotoxicity and its acute mortality for humans. Madridge J Nanotechnol Nanosci 3(2):116–118. https://doi.org/10.18689/mjnn-1000122
- Andreou VG, Nikolelis DP (1998) Flow injection monitoring of aflatoxin M₁ in milk and milk preparations using filter-supported bilayer lipid membranes. Anal Chem 70:2366–2371. https://doi.org/10.1021/ac9712097
- Bratakou S, Nikoleli G-P, Siontorou CG, Nikolelis DP et al (2016) Electrochemical biosensor for naphthalene acetic acid in fruits and vegetables based on lipid films with incorporated auxinbinding protein receptor using graphene electrodes. Electroanalysis 28(9):2171–2177. https:// doi.org/10.1002/elan.201500299
- Ebert A, Hannesschlaeger C, Goss K-U, Pohl P (2018) Passive permeability of planar lipid bilayers to organic anions. Biophys J 115:1931–1941. https://doi.org/10.1016/j.bpj.2018.09.025
- Evtugyn G, Porfireva A, Stepanova V, Sitdikov R, Stolokov I, Nikolelis D, Hianik T (2014) Electrochemical aptasensor based on polycarboxylic macrocycle modified with neutral red for aflatoxin B1 detection. Electroanalysis 26(10):2100–2109. https://doi.org/10.1002/ elan.201400328
- Gould SB (2018) Membranes and evolution. Curr Biol 28:381–385. https://doi.org/10.1016/ S0960-9822(06)00205-3
- Ishimori Y, Kawano K, Tamura H, Aoyama N, Mouri M, Kase T, Tamiya E, Ishizuka M (2002) Advanced environmental monitoring system using ecosensor based on bilayer lipid membrane. In: Proceedings of SPIE 2001, remote sensing for environmental monitoring, GIS applications, and geology, Toulouse, France. SPIE, volume 4545
- Kalyankar ND, Sharma MK, Vaidya SV, Calhoun D, Maldarelli C, Couzis A, Gilchrist L (2006) Arraying of intact liposomes into chemically functionalized microwells. Langmuir 22:5403–5411. https://doi.org/10.1021/la0602719
- Mazur F, Bally M, Städler B, Chandrawati R (2017) Liposomes and lipid bilayers in biosensors. Adv Colloid Interf Sci 249:88–99. https://doi.org/10.1016/j.cis.2017.05.020
- Michaloliakos AI, Nikoleli G-P, Siontorou CG, Nikolelis DP (2012) Rapid flow injection electrochemical detection of arochlor 1242 using stabilized lipid membranes with incorporated sheep anti-PCB antibody. Electroanalysis 24:495–501. https://doi.org/10.1002/elan.201100393
- Nikoleli G-P, Israr MQ, Tzamtzis N, Nikolelis DP, Willander M, Psaroudakis N (2012) Structural characterization of graphene nanosheets for miniaturization of potentiometric urea lipid film based biosensors. Electroanalysis 24:1285–1295. https://doi.org/10.1002/elan.201200104
- Nikoleli G-P, Nikolelis DP, Evtugyn G, Hianik T (2016) Advances in lipid film based biosensors. TrAC Trends Anal Chem 79:210–221. https://doi.org/10.1016/j.trac.2016.01.021
- Nikoleli GP, Nikolelis DP, Siontorou CG, Karapetis S (2018) Lipid membrane nanosensors for environmental monitoring: the art, the opportunities, and the challenges. Sensors 18:284–298. https://doi.org/10.3390/s18010284
- Nikoleli G-P, Siontorou CG, Nikolelis M-T, Bratakou S, Bendos DK (2019) Recent lipid membrane-based biosensing platforms. Appl Sci (Switzerland) 9(9):1745–1764. https://doi. org/10.3390/app9091745
- Nikolelis DP, Krull UJ (1994) Direct electrochemical sensing of insecticides by bilayer lipid membranes. Anal Chim Acta 288(3):187–192. https://doi.org/10.1016/0003-2670(94)80132-0
- Nikolelis DP, Mitrokotsa M (2002) Stabilized lipid film based biosensor for atenolol. Biosens Bioelectr 17(6–7):565–572. https://doi.org/10.1016/S0956-5663(02)00017-9
- Nikolelis DP, Siontorou CG, Andreou VG, Krull UJ (1995) Stabilized bilayer-lipid membranes for flow-through experiments. Electroanalysis 7:531–536. https://doi.org/10.1002/ elan.1140070605
- Nikolelis DP, Siontorou CG, Krull UJ, Katrivanos PL (1996) Ammonium ion minisensors from self-assembled bilayer lipid membranes using gramicidin as an ionophore. Modulation of

ammonium selectivity by platelet-activating factor. Anal Chem 68(10):1735–1741. https://doi. org/10.1021/ac950403v

- Nikolelis DP, Raftopoulou G, Nikoleli G-P, Simantiraki M (2006) Stabilized lipid membrane based biosensors with incorporated enzyme for repetitive uses. Electroanalysis 18:2467–2474. https://doi.org/10.1002/elan.200603702
- Nikolelis DP, Raftopoulou G, Chatzigeorgiou P, Nikoleli G-P, Viras K (2008a) Optical portable biosensors based on stabilized lipid membrane for the rapid detection of doping materials in human urine. Sens Actuators B Chem 130:577–582. https://doi.org/10.1016/j.snb.2007.10.011
- Nikolelis DP, Raftopoulou G, Simantiraki M, Psaroudakis N, Nikoleli G-P, Hianik T (2008b) Preparation of a selective receptor for carbofuran for the development of a simple optical spot test for its rapid detection using stabilized in air lipid films with incorporated receptor. Anal Chim Acta 620:134–141. https://doi.org/10.1016/j.aca.2008.05.011
- Nikolelis DP, Ntanos N, Nikoleli G-P, Tampouris K (2008c) Development of an electrochemical biosensor for the rapid detection of naphthalene acetic acid in fruits by using air stable lipid films with incorporated auxin-binding protein 1 receptor. Protein Pept Lett 15:789–794. https:// doi.org/10.2174/092986608785203692
- Nikolelis DP, Raftopoulou G, Psaroudakis N, Nikoleli G-P (2009) Development of an electrochemical chemosensor for the rapid detection of zinc based on air stable lipid films with incorporated calix4arene phosphoryl receptor. Int J Environ Anal Chem 89:211–222. https://doi. org/10.1080/03067310802578952
- Nikolelis DP, Psaroudakis N, Michaloliakos AI, Nikoleli G-P, Scoulos M (2013) Rapid flow injection electrochemical detection of 3,3',4,4' tetrachlorobiphenyl using stabilized lipid membranes with incorporated sheep antibody. Cent Eur J Chem 11:320–323. https://doi.org/10.2478/s11532-012-0157-4
- Pandit S, Dasgupta D, Dewan N, Ahmed P (2009) Nanotechnological advances in biosensors. Sensors 9(11):8907–8910. https://doi.org/10.3390/s91108907
- Pandit S, Dasgupta D, Dewan N, Ahmed P (2016) Nanotechnology based biosensors and its application. Pharm Innov J 5(6):18–25
- Raghunathan K, Kenworthy AK (2018) Dynamic pattern generation in cell membranes: current insights into membrane organization. BBA-Biomembranes 1860:2018–2031. https://doi. org/10.1016/j.bbamem.2018.05.002
- Shiratori SS, Kohno K, Yamada M (2000) High performance smell sensor using spatially controlled LB films with polymer backbone. Sens Actuators B 64:70–75. https://doi.org/10.1016/ S0925-4005(99)00486-4
- Siontorou CG, Nikolelis DP, Krull UJ (1997a) A carbon dioxide biosensor based on hemoglobin incorporated in metal supported bilayer lipid membranes (BLMs): investigations for enhancement of response characteristics by using platelet-activating factor. Electroanalysis 9(14):1043–1046. https://doi.org/10.1002/elan.1140091403
- Siontorou CG, Nikolelis DP, Krull UJ, Chiang KL (1997b) A triazine herbicide minisensor based on surface-stabilized bilayer lipid membranes. Anal Chem 69:3109–3114. https://doi.org/10.1021/ac970113+
- Siontorou CG, Nikolelis DP, Tarus B, Dumbrava J, Krull UJ (1998) DNA biosensor based on selfassembled bilayer lipid membranes for the detection of hydrazines. Electroanalysis 10:691–694. https://doi.org/10.1002/(SICI)1521-4109(199808)10:10<691::AID-ELAN691>3.0.CO;2-N
- Siontorou CG, Nikolelis DP, Krull UJ (2000) Flow injection monitoring and analysis of mixtures of hydrazine compounds using filter-supported bilayer lipid membranes with incorporated DNA. Anal Chem 72(1):180–186. https://doi.org/10.1021/ac990618v
- Siontorou CG, Nikoleli G-P, Nikolelis DP, Karapetis SK (2017) Artificial lipid membranes: past, present, and future. Membranes 7:38–61. https://doi.org/10.3390/membranes7030038
- Snejdarkova M, Svobodova L, Evtugyn G, Nikolelis DP et al (2004) Acetylcholinesterase sensors based on gold electrodes modified with dendrimer and polyaniline: a comparative research. Anal Chim Acta 514(1):79–88. https://doi.org/10.1016/j.aca.2004.03.019

- Tien HT, Salamon Z (1989) Formation of self-assembled lipid bilayers on solid substrates. J Electroan Chem Interfacial Electrochem 2:211–218. https://doi. org/10.1016/0022-0728(89)87265-1
- Vacek J, Zatloukalova M, Novak D (2018) Electrochemistry of membrane proteins and protein–lipid assemblies. Curr Opin Electrochem 12:73–80. https://doi.org/10.1016/j. coelec.2018.04.012

Chapter 5 Novel Chemical Sensors Based on Green Composite Materials for Environmental Analysis



K. B. R. Teodoro, F. L. Migliorini, M. H. M. Facure, R. C. Sanfelice, D. Martins, and D. S. Correa

Contents

5.1	Introduction			
5.2	Transduction Mechanism in Chemical Sensors: A Brief Introduction			
	5.2.1	Impedimetric Sensors (e-Tongues)	112	
	5.2.2	Electrochemical Sensors.	114	
	5.2.3	Optical Chemical Sensors.	115	
5.3	Applications of Green Composites in Sensors		117	
	5.3.1	Green Composite–Based Sensors for Detecting Heavy Metals in Water	117	
	5.3.2	Green Composites Applied in Sensors for Detecting Pathogens in Water	122	
	5.3.3	Green Composites Applied in Sensors for Detecting Hazardous Volatiles	126	
5.4	Final Remarks.		128	
Refe	References			

K. B. R. Teodoro · F. L. Migliorini · D. Martins

M. H. M. Facure · D. S. Correa (⊠) Nanotechnology National Laboratory for Agriculture, Embrapa Instrumentação, São Carlos, SP, Brazil

R. C. Sanfelice

© The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

109

Nanotechnology National Laboratory for Agriculture, Embrapa Instrumentação, São Carlos, SP, Brazil

PPGQ, Department of Chemistry, Center for Exact Sciences and Technology, Federal University of São Carlos (UFSCar), São Carlos, SP, Brazil e-mail: daniel.correa@embrapa.br

Department of Chemical Engineering, Institute of Technological and Exact Sciences – ICTE, Universidade Federal do Triangulo Mineiro – UFTM, Uberaba, MG, Brazil

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_5

Abstract In this book chapter, we report the recent advances in the development and application of chemical sensors based on green composite materials for environmental analysis. Emphasis is given on the sensor performance in terms of materials employed (including macromolecules from vegetal sources combined with varied nanostructures), structure-property relationships, and device fabrication technology. Specifically, the chapter focuses on green materials obtained from vegetal biomass including cellulose, lignin, starch, alginate, pectin, and vegetal protein, indicating the viability for the construction of non-toxic, recyclable, renewable, and biodegradable devices. Firstly, a brief overview on different types of chemical sensors, including electronic tongues, electrochemical sensors, and chemical optical sensors, is presented. Then, the general aspects of green composites as sensing layers are outlined and discussed with respect to their application in environmental analytical chemistry. Emphasis is placed on results reporting the detection of heavy metals and bacterial contamination in water as wells as hazardous volatile organic compounds. Lastly, a brief perspective on the challenges and upcoming opportunities toward the development of novel sensing devices based on green composites is discussed

Keywords Electronic tongue · Optical chemical sensor · Electrochemical sensor · Cellulose · Green materials · Nanostructures · Two-dimensional materials

5.1 Introduction

In the last decades, an increasing amount of organic and inorganic hazardous compounds, including heavy metal ions, pesticides, and pharmaceuticals, have been released into the environment without proper treatments (Antonacci et al. 2018; Wu et al. 2019; Patel et al. 2019). The identification and quantification of these substances play an important role in pollution monitoring and management, which requires the development of accurate and fast detection technologies (Arduini et al. 2017; Parmar et al. 2018; Zhang et al. 2019). Among these technologies, chemical sensors have been of special interest due to their potential applicability in the detection of a single analyte or simultaneous analytes in a fast and efficient manner, besides other advantages compared to traditional analytical techniques, including easy handling, reduction in cost per analysis, as well as the possibility of automation and miniaturization (Cui et al. 2015; Labib et al. 2016; Maduraiveeran and Jin 2017).

A chemical sensor is a device that combines a sensitive layer (part of the sensor that interacts chemically with the analyte under investigation) and a transducer, which is a component that converts/transforms the chemical information (e.g., composition or concentration) into a useful analytical signal (Hulanicki et al. 1991). Chemical sensors may be classified, based on the transduction principles, as electrical (e.g. impedimetric), electrochemical (e.g. impedimetric, potentiometric, and amperometric), optical (e.g. fluorescent, colorimetric), and gravimetric (e.g. quartz

crystal microbalance, magnetoelastic sensors, static and dynamic) sensors (Hulanicki et al. 1991; Correa et al. 2017; Kassal et al. 2018; Paladiya and Kiani 2018; Teodoro et al. 2019b).

A broad range of materials and their combinations have been proposed to be used as sensing layers in sensors, including metals and metal oxides, inorganic crystalline and amorphous materials, and polymers and composite materials (Mercante et al. 2019; Ray 2017). More recently, the number of publications describing the development of sensing layers prepared from ecofriendly solvents and reagents has exponentially increased (Yáñez-Sedeño et al. 2019). Several unique features of natural polymers make them attractive, including biodegradability, relatively low cost, non-toxicity, reactive functional groups, and abundant availability in nature (Maciel et al. 2019). Such features make them suitable for designing devices, including distinct types of sensors that are efficient, durable, inexpensive, simple, easy to prepare, and ecofriendly (La Mantia and Morreale 2011; Tingaut et al. 2012; Golmohammadi et al. 2017; Naveen et al. 2017; Teodoro et al. 2018, 2019a; Wang et al. 2018; Cichosz et al. 2018; Maciel et al. 2019; Andre et al. 2020).

A large variety of natural polymers such as cellulose, chitosan, chitin, and starch, as well as its derivatives, has been applied for the fabrication of ecofriendly sensors. Natural macromolecules as lignin and zein have also been used for this purpose (Sun et al. 2011; Suginta et al. 2013; Rudnitskaya et al. 2013; Saithongdee et al. 2014; Golmohammadi et al. 2017; Anichini et al. 2018; Maciel et al. 2019; Meng et al. 2019). It is well known that natural polymers and macromolecules show electrically insulating nature, which has stimulated researches based on the preparation of hybrid composite sensing layers via the combination with two-dimensional materials (2DMs) (Salavagione et al. 2014; Zhang et al. 2017; Maciel et al. 2019; Teodoro et al. 2019a; Yáñez-Sedeño et al. 2019; Yusoff 2019; Meng et al. 2019). For instance, 2DMs such as graphene, phosphorene, and transition metal dichalcogenides have rapidly emerged as attractive candidates for the fabrication of high-performance chemical sensors due to their remarkable electronic and optical properties combined with their high surface area-to-volume ratios, robustness, and flexibility, improving sensor performance in terms of specificity and sensitivity (Anichini et al. 2018; Meng et al. 2019).

In the next sections, we discuss on distinct types of sensors based on the plethora of green composite materials that can be successfully used in the design of such devices employed for monitoring heavy metals, microorganisms, as well as volatile organic compounds (VOCs).

5.2 Transduction Mechanism in Chemical Sensors: A Brief Introduction

As mentioned before, chemical sensors are devices capable to transform a physical, chemical, or biological stimulus into a measurable analytical signal (Kruse 2018; Vikesland 2018). The use of sensors and biosensors for health and environmental monitoring has been extensively explored in the last decades (Lahcen and Amine 2018; Jayathilaka et al. 2019; Reynoso et al. 2019). Compared to analytical techniques usually employed for these application as atomic absorption spectroscopy (AAS), inductively coupled plasma-mass spectrometry (ICP-MS), mass spectroscopy (MS), and X-ray fluorescence spectroscopy (XPS), chemical sensors offer some advantageous properties for analyte monitoring, which include experimental simplicity, expedite analysis, possibility of miniaturization, portability, and capability for on-site analysis (Wei et al. 2012; Arino et al. 2017; Golmohammadi et al. 2017; Mercante et al. 2017). In the next section we will focus on some types of chemical sensors, including impedimetric e-tongues, electrochemical, and optical chemical sensors.

5.2.1 Impedimetric Sensors (e-Tongues)

Electronic tongues (or e-tongues), as multicomponent analysis, appear as a suitable approach to perform on-site simultaneous evaluation and classification of trial solutions with very similar compositions (Riul-Jr et al. 2010; Karkra et al. 2017; (Kirsanov et al. 2019). Such devices consist of an array of distinct sensing units composed by interdigitated electrodes (IDEs) functionalized with different nanomaterials, and employ the concept of global selectivity and cross-sensitivity (Correa et al. 2014; Di Rosa et al. 2017; Podrazka et al. 2017) as illustrated in Fig. 5.1. Through electrical transduction, the e-tongues collect the sample's fingerprint, which is usually interpreted by varied statistical analysis (Riul-Jr et al. 2010).

The use of impedimetric e-tongues is advantageous for sample analysis once it does not require, in principle, the use of materials that provide specific interactions with the analyte, which is quite attractive for the analysis and classification of liquid samples of complex compositions. Additionally, unlike electrochemical measurements, the sensing array of non-specific electrodes used in impedimetric e-tongues does not require the use of electroactive species neither of reference electrode. The sensor-solution system can be described as an equivalent circuit in such a way that the electrostatic double layer formed at the interface electrode/solution governs the response at low frequencies, the conductance of solution and ultrathin films that coat the electrodes generate the responses at intermediate frequencies, while geometric capacitance is more influential at high frequencies (Taylor and Macdonald 1987; Riul-Jr et al. 2010; Giacometti et al. 2016).



Fig. 5.1 Schematic representation of an experimental setup of a typical e-tongue: (i) interdigitated electrodes (IDEs) modified with distinct materials; (ii) measurement cell containing the sensor array immersed in the liquid under investigation; (iii) data analysis of trial solutions collected using the e-tongue can generate a large amount of data, which can be classified and distinguished using the appropriated statistical analysis

This approach makes use of the impedance concept, once it represents the total opposition to the current in an AC circuit, in which resistance and reactances are provided by resistors, capacitors, and inductors (Bard and Faulkner 2000). In general, it can be assumed that impedance (Z) corresponds to the equivalent resistance of a DC circuit respecting the Ohm's Law. Considering that the impedance study of an AC circuit can be performed using graphical representations of a complex number in the complex plane, impedance spectroscopy can provide a graphical representation of the real and imaginary impedance data (Z and Z^* , respectively) in accordance with the frequency applied. Once an adequate AC voltage is applied to the electrode system, the corresponding response current is measured, and the data can then be collected as resistance, capacitance, or impedance (Bhand and Bacher 2017).

The impedance response is obtained by frequency variation over a wide range (e.g., mHz to MHz), in such a way that the assessment of information from a large amount of data is given. The use of advanced mathematical and statistical methods provides the capacity to distinguish samples with very similar chemical composition, providing useful qualitative and quantitative information. A suitable design of sensing units is a crucial strategy to develop e-tongues capable to discriminate varied analytes at concentrations as low as nanomolar levels (Alonso et al. 2012; Facure et al. 2017; Marques et al. 2017; Teodoro et al. 2019b).

5.2.2 Electrochemical Sensors

Electrochemical sensors constitute a powerful tool for chemical analysis, especially because of the high sensitivity and moderate cost when compared to other analytical techniques. Additionally, electrochemical sensors can provide fast analysis and depending on its design and materials employed, they can also be used as wearable and disposable sensors for in-situ and real-time analysis (Stradiotto et al. 2003; Bandodkar and Wang 2014; Patella et al. 2019). In addition, the versatility of electrochemical techniques also enables customizing and controlling electrode reactions by modifying the electrode–solution interface and potential applied to the electrochemical cell, combined to the ease of automation for the electrical signal measurements (Brett and Brett 1994; Bard and Faulkner 2000).

Electrochemical sensors are generally based on electrochemical cells with two or more electrodes in contact with the electrolyte, as displayed in Fig. 5.2. Such sensors can be classified, according to the mode of operation as amperometric, conductometric, potentiometric, or voltammetric sensors (Stradiotto et al. 2003). The choice of the measurement parameters is made in order to optimize the sensor sensitivity, selectivity, and accuracy, depending on the chemical species to be determined. In this way, the knowledge and limitations of each operational mode are essential in the planning and development of a sensor (Lowinsohn and Bertotti 2006). These types of sensors are usually employed for the evaluation and quantification of chemical or biological compounds, even when they are present at very low concentrations in the medium. The determination of these compounds occurs through redox reactions at the electrode/solution interface, which are later converted into electrical signals (Patella et al. 2019), that is, the output signal is a measurement of current, potential, or electrical resistance, depending on the mode of operation. As the redox reaction occurs at the surface of the working electrode, surface area is an important parameter that can enhance the output signal (Brett and Brett 1994).



Fig. 5.2 Schematic representation of an experimental setup employed for electrochemical sensor analysis composed of a three-electrode (working electrode, reference electrode, and counter electrode) cell, a potentiostat, and a computer

5.2.3 Optical Chemical Sensors

Optical chemical sensors are a subclass of chemical sensors that employ optical transduction to provide a useful analytical signal (McDonagh et al. 2008). An example of an experimental apparatus used in optical sensors for detection of gaseous analytes is shown in Fig. 5.3. The interaction of the electromagnetic radiation with the analyte of interest can cause significant changes in the optical parameters of the system and the degree of this interaction is related to the concentration of the analyte (Narayanaswamy 1993; Lobnik et al. 2012a; Lobnik et al. 2012b). The transduction can occur by different mechanisms, which include absorbance, scattering, reflection, refraction, and luminescence processes in the electromagnetic spectral region spanning from ultraviolet (UV) to infrared (IR). For the development of optical chemical sensors, the frequent methods of detection are based on the principles of absorption and light emission (luminescence), due to their good sensitivity, experimental simplicity, and sensitivity (Jeronimo et al. 2007; Z. Li et al. 2018).

The process of optical sensing occurs by a direct or indicator-mediated sensing. In the direct sensing, the absorption or emission (luminescence) of analyte is



Fig. 5.3 Schematic representation of an experimental setup employed for optical chemical sensor analysis, composed of a spectrophotometric equipment coupled to a hermetically sealed chamber and arranged to enter and exit the gas. Two cylinders are connected to the gas chamber, one filled with the gas being detected and the other filled with nitrogen gas used for the purge. Reprinted with permission from Usha et al. (2015)). Copyright (2015) MDPI

measured directly and quantitatively through a calibration curve. In the indicatormediated sensing, on the other hand, the use of an intermediate agent is necessary, which acts as an active layer or sensory platform (indicator) interacting with analyte and inducing the response (Hayashi 2015). Such process of optical sensing is interesting because it enables the analysis of a large number of analytes, including those that do not present intrinsic optical properties, the detection of which would be hindered by the direct method. In this case, the optical chemical sensors detect the chemical reactivity between the analyte and the indicator, instead of the physical properties of the analyte. Thus, it is possible to obtain sensors with high sensitivity and discrimination capacity between analytes of similar features in both gas and liquid phases (Lobnik et al. 2012b; Hayashi 2015; Schoolaert et al. 2017; Li et al. 2018).

The indicators usually employed in optical chemical sensors are thin films including those based on polymer owing to the thin film formation capacity at low temperature, adjustable properties, as well as its ability to form composite and hybrid materials with several nanostructures (Sanfelice et al. 2014; Christau et al. 2015; Sanfelice et al. 2017; Andre et al. 2018; Terra et al. 2018; Mahapure et al. 2019; Mao et al. 2019). To be used as a platform in an optical chemical sensor, the polymer material must have optical activity and interact with the analyte investigated. Conjugated polymers exhibit alternating π -bonds throughout the chain, which allows the optical activity of these materials for optical sensors (McQuade et al. 2000; Thomas et al. 2007), such as polypyrrole (PPy; Zhou et al. 2015; Liu et al. 2017b), polyaniline (PANI; Luo et al. 2017; Mohamad Ahad et al. 2018), and polythiophene (PT; Li and Shi 2013; Liu et al. 2017a).

When an optical sensor is based on the use of an indicator, the analyte quantification relies on changes observed in the analytical signal of the indicator. The two most widely employed types of optical chemical sensors are based on colorimetric (Schoolaert et al. 2017; Najarzadekan and Sereshti 2018) and luminescence properties (Najarzadekan and Sereshti 2018; Terra et al. 2018; O'Keeffe et al. 2018). In colorimetric detection, the indicators display color changes in the presence of the analyte of interest, which in many cases can be measured by UV-Vis absorption technique or even detected by naked eyes (Liu et al. 2017a; Maity and Ghosh 2018; Ma et al. 2018). Luminescence sensors usually present a higher sensitivity compared to other optical chemical sensors (Hayashi 2015). Two processes, namely fluorescence and phosphorescence, can explain the luminescence mechanism. Fluorescence occurs through the excitation of an electron to an excited state, and its subsequent radioactive relaxation to the ground state. In phosphorescence, first a forbidden transition occurs to an excited triplet state, and subsequently relaxation to the ground state (Lakowicz 2006). Fluorescence analyses are more commonly used because this technique can take advantage of different parameters to be investigated, such as fluorescence intensity and life time, anisotropy, and quantum yield.

5.3 Applications of Green Composites in Sensors

5.3.1 Green Composite–Based Sensors for Detecting Heavy Metals in Water

A worldwide environmental issue related to water degradation lies in the excessive proportion of heavy metals dumped into water resources, usually related to the inappropriate disposal of industrial wastes (Karkra et al. 2017; Peng et al. 2018). Since these pollutants are non-biodegradable, the occurrence of bio-accumulation in aquatic and terrestrial living organisms and human health triggers serious health disorders (Alkmim-Filho et al. 2014; Liu et al. 2017c; Jose and Ray 2018). Although some of these substances play important role in physiological mechanisms, most of them are poisonous even under low concentrations, resulting in serious diseases, mutation, and even death by intoxication (Tchounwou et al. 2012). Therefore, the development of reliable and expedite techniques capable to detect such hazardous compounds is highly required.

Recent works have shown the interesting synergism between green materials and hazardous heavy metal ions, which has led to the development of innovative strategies for heavy metal sensing detection. For instance, cellulose surface chemical modifications through carboxylation, amination, succinylation, and sulfonation have improved their interactions with varied pollutants (Seely and Hart 1974; Lee and Mooney 2000; Chin et al. 2017); lignin has a large number of active sites that are highly suitable for complexation with heavy metal ions (Garcia-Valls and Hatton 2003); and pectin can coordinate with some heavy metal ions due to electrostatic attraction by carboxyl and hydroxyl groups in its backbone (Kartel and Kupchik 1999; Juan Zhao and Qin Zhou 2015).

Cellulose is the most abundant green material available on Earth and can be tailored to design novel composites for a plethora of applications, including sensing and biosensing. In sensing platforms, cellulose is normally employed as paper substrate, membranes, or thin films. Paper-based platforms, as the one exemplified in Fig. 5.4, encompass features as flexibility, high specific stiffness, porous structure, compatibility with biological samples, printability, biodegradability, and low cost to fabricate disposable sensors (Chaiendoo et al. 2017; Li et al. 2019). Paper has been conveniently applied in order to further reduce the gap between complex laboratory testing and simple point-of-need testing (Cinti 2019). Cellulose can be applied in pristine form (Kuek Lawrence et al. 2014; Ferreira et al. 2015; Cui et al. 2019) or as part of composites (Rashid et al. 2016; Chaiendoo et al. 2017; Sahatiya et al. 2018), as substrate for screen-printed sensors (Fang et al. 2011; de Oliveira et al. 2019; Ruecha et al. 2019) and microfluidic platforms (Moraes et al. 2018; Ilacas et al. 2019; Ruecha et al. 2019). Bacterial cellulose can also be used as nanopaper as novel, flexible, and transparent substrate to optical sensor (Morales-Narváez et al. 2015; Abbasi-Moayed et al. 2018).

Although cellulose shows insulating characteristics, the advances in composite materials field have enabled to extend its application in electrical and



Fig. 5.4 Experimental setup related to electrochemical paper-based devices obtained by application of (**a**) high-adsorption substrates and (**b**) low-adsorption substrates (Cinti 2019). Reprinted with permission from Cinti (2019)). Copyright (2019) Springer Nature. (**c**) Example of paper-based screen-printed electrode proposed by Arduini et al. (2019)), which uses a paper-based device folded as three-dimensional origami combining different enzyme-inhibition biosensors for the detection of different classes of pesticides. Reprinted with permission from Arduini et al. (2019)). Copyright (2019) Elsevier

electrochemical sensors. For instance, Tan and coworkers (2018) studied the use of cellulosic fibers coated with polyaniline (PANI) as working electrode in the electrochemical detection of Pb(II). The composite fabrication required the control of PANI polymerization, in order to achieve a thin layer onto cellulose fibers. Firstly, the aniline monomers were adsorbed onto cellulose surface by H-bonding interactions and then the polymerizing agent was added. Such procedure allowed the formation of a PANI nanolayer of about 50 nm. The physicochemical characterization indicated that the presence of PANI afforded conductive properties for the paper. This disposable sensor responded linearly to Pb in the working range from 0.2 ppm to 1.0 ppm, yielding a limit of detection of 0.348 μ M (0.0721 ppm).

Also, potential and recently explored is the use of cellulose nanostructures in sensing platforms. Cellulose nanocrystals, nanofibrils, and nanofilms show distinctive properties that are superior to common paper (Abbasi-Moayed et al. 2018). Application of cellulose nanocrystals as fillers to mechanical reinforcement is very traditional. However, emerging technologies and the necessity of small, biodegradable, lightweight, and strong products have widened the application of this material. In a recent work, Teodoro and coworkers (2019) used a hybrid material based on cellulose nanocrystals and graphene oxide to functionalize fluorine-doped tin oxide (FTO) electrodes for electrochemical determination of Hg(II). Cellulose was explored as a green reducing and stabilizing agent for graphene oxide nanosheets. The combination of this hybrid material with electrospun nanofibers enabled enhancing the charge transfer properties and allowed to detect mercury(II) with a limit of detection of 520 nM. The good performance of this sensor was attributed to a good distribution of graphene onto the platform, as a consequence of synergistic interactions between cellulose nanocrystals and graphene.

Zinoubi et al. (2017) applied nanofibrillated cellulose (NFC) from *Eucalyptus* to modify carbon glass electrodes (CGE) to electrochemically evaluate heavy metal

trace in seawater. Nano-sized fibrils were produced by fibrillation of Eucalyptus fibers, yielding fibers with 5-10 nm of diameters. The fiber surface was modified using 2,2,6,6-tetramethyl-1-piperidinyloxy (TEMPO) reactant, by a procedure called TEMPO-mediated oxidation (Roy et al. 2009), leading to formation of carboxyl groups. The electrode modification was performed by adsorption of nanofibrillated cellulose onto CGE, which were highly adhered due to interactions between the long β -glucans of cellulose and the carbon surface, through hydrogen bonding. The authors also performed a study of accumulation time, once the electrochemical performance could be affected by interactions between NFC matrix and metallic ions. Differential pulse anodic stripping voltammetry (DPASV) was then applied to quantify trace amounts of Cd(II), Cu(II), Pb(II), and Hg(II). As a result, the proposed platform performed reproducible response, high sensitivity, and good stability for all analytes. A low limit of detection of 5 nM was obtained for Cd(II) and Hg(II), and 0.5 nM for Cu(II) and Pb(II). The higher sensitivity and the low limit of detection obtained for these metal ions were attributed to the good affinity with oxidized cellulose nanofibrils.

Chemical modification of cellulose is expanding the sensor fabrication methods and allowing interaction with chromophores, which enables the fabrication of optical heavy metal sensors. Azmi et al. (2018) produced an easy-to-interpret colorimetric platform by impregnation of chromophore dithizone in flat casting films of cellulose acetate. The composite was cut as strips and dipped into mercury(II) aqueous solutions, allowing naked eye detection at 3 ppm Hg(II). Zhang and coworkers (2018a) developed a sensor based on electrospun nanofibrous cellulose acetate/curcumin membranes for colorimetric detection of lead(II). The curcumin is a natural chromophore able to chelate with heavy metal ions to form complexes, which leads to color changes. The sensor demonstrated to be selective, but influenced by pH and membrane thickness.

Starch, another carbohydrate, was used as reducing and capping agent in gold nanoparticle (AuNP) green synthesis performed by Ban et al. (2015). The resultant composite could be applied in the plasmonic sensing determination of Cu(II) and Pb(II) in water. The sensing experiments were performed in aqueous solution, by simply mixing starch-AuNP suspension with known concentration of six different heavy metals: Cu(II), Ni(II), Zn(II), Pb(II), Hg(II), and As(II). The proposed material presented high sensitivity to Pb(II) and Cu(II), which could be seen by color change even under ppm concentrations. The color changed from pink (typically related to AuNP) to dark ruby red in the presence of Pb(II) and to violet due the presence of Cu(II). Simultaneous analysis of other metallic compounds revealed no significant color change, indicating the selectivity for these species. The authors reported that the affinity-based chelating activity of surface stabilizing agents were responsible for sensing the metal ions; therefore, starch might be responsible for the selectivity to Pb(II) and Cu(II). Similar work was performed by these authors using starch-capped silver nanoparticles (AgNPs; Ban and Paul 2018). Very small nanoparticles (2-10 nm) revealed optical (absorption and fluorescence) activity in UV-Vis spectra. The same six heavy metals were evaluated and a selective sensing response for Hg(II) was verified as consequence of the evident color change from yellow to colorless, under a more pronounced surface plasmon resonance (SPR) shift, varying in accordance with the concentration range of 1–10 ppm. The selectivity for Pb(II) and Cu(II) using starch-based optical sensors was also verified by Khachatryan and Khachatryan (2019). In their study, elastic films composed by starch-ZnS quantum dots (QDs) nanocomposites were produced. The sensing experiments showed a decrease in emission intensity when the films were exposed to Pb(II) and Cu(II) ions, which was attributed to the interaction between the cation metal and QDs surface that may reduce the electron-hole recombination.

Narayanan and Han (2017) have applied alginate-stabilized silver nanoparticles to label-free colorimetric sensing of manganese(II) in water. Alginate, a polysaccharide derived from algal biomass, acted in an ecofriendly reduction and stabilization of silver nanoparticles. The proposed nanoplatform showed selectivity and sensitivity for optical detection of Mn(II) in aqueous solution, in the range of $1-10 \mu$ M. The mechanism of detection involved the aggregation of silver nanoparticles, which led to color change from pale yellow to brownish yellow, followed by a gradual decrease and broadening of SPR absorbance band at 400 nm. The crosslinking of alginate polymer chains led to reduction in interparticle distance, inducing a very slight aggregation of AgNPs. For comparison, the chelating ability was compared with chelation caused by ethylenediaminetetraacetic acid (EDTA), and the effect caused by the presence of Mn(II) led to a more expressive colorimetric change (Fig. 5.5).

Carbon-based materials from biomass are reported as low-cost and readily available starting materials to the synthesis of interesting novel structures. Among them, lignin has attracted great attention due to its abundance in nature and because it has been applied, for instance, in environmentally friendly Carbon-Dots synthesis (Chen et al. 2016; Yang et al. 2018; Liu et al. 2019). Sources rich in organic compounds such as glucose, cellulose, and phenolic molecules are very applicable for the preparation of Carbon-Dots (Liu et al. 2019). Electrospun carbon-fibers obtained from lignin have also been explored in several applications (Jayawickramage and Ferraris 2019; Roman et al. 2019; Titirici et al. 2019).

Martins and coworkers (2008) performed sensing experiments with lignin extracted from sugarcane bagasse, which was applied in the fabrication of ultrathin Langmuir–Blodgett (LB) films. The sensitivity of lignin-based films to Cu(II) was evaluated by cyclic voltammetry and electrical impedance spectroscopy using an electronic tongue. For the electrochemical study, 21 layers of lignin films were deposited onto indium tin oxide (ITO) electrodes, and for impedimetric study, five layers of lignin films were deposited onto interdigitated gold electrodes. The results showed that the LB film could be used as sensor for detection of low concentrations of Cu (10 mg L^{-1}), the selectivity of which was attributed to the lignin complexation with metallic ions in solution.

Another interesting carbon-based material derived from biomass is the biochar, which has attracted attention in reducing ecological and health risks of hazardous substances, such as heavy metal compounds. Liu et al. (2016) fabricated a low-cost sensor depositing on carbon glass electrodes (CGEs) a thin film based in a biochar prepared from pyrolysis of pine for voltammetric determination of Cd(II). The



Fig. 5.5 (a) Photograph of aqueous suspension of alginate-stabilized AgNP obtained by Narayanan and Han (2017)). (b) Color changes caused by EDTA addition and (c) Mn(II) addition. (d) TEM images of alginate-AgNP and (e) alginate-AgNP agglomeration caused by Mn(II). In (f) is demonstrated the selectivity for Mn(II) by photographs and plot of A500/A400 for alginate-AgNP suspensions containing different metal ions. Reprinted with permission from Narayanan and Han (2017)). Copyright (2017) Springer Nature

biochar was mixed to montmorillonite for enhancing surface area, ion-exchange ability, dispersive stability, and adsorption capacity, and to carboxymethyl cellulose to improve water-dispersive, film-forming, and adhesive abilities. The as-obtained biochar has flaky aggregates morphology and the film montmorillonite-cellulose-biochar has a large specific surface area. The compositional analysis revealed the presence of several oxygen functional groups such as carboxylic, hydroxyl, phenol, and carbonyl in biochar. Films containing the biochar showed the highest stripping peak, indicating a significant enhanced synergistic effect of different components due to superior adsorption capacity, large specific surface area, and good exchange capacity. The selectivity of the sensor was verified toward other metals—Pb(II), Cu(II), Hg(II), Zn(II), Al(III), and Fe(III)—and the sensitivity was evaluated in linear range of Cd(II) concentration of 2.0 nM to 80 μ M, leading to a limit of detection of 6.7 nM.

5.3.2 Green Composites Applied in Sensors for Detecting Pathogens in Water

The monitoring of microorganisms in different environments and conditions is of great relevance nowadays because, in the same extent they play crucial role for the proper functioning of specific natural systems, some microorganisms can also be pathogenic and behave as infectious agents, posing high risks to human and animal health (Leonard et al. 2003; Deisingh and Thompson 2004; Duffy and Moore 2017). Infections with pathological agents can lead to serious illnesses, ultimately causing the death of the infected patient. In addition to health problems, infections caused by live pathogens represent a significant expenditure for the governments and can even threaten global security once they are used as biological weapons (Xu 2012).

A recurring concern in this regard is the quality of water, because infections caused by microbial pathogens can be acquired and transmitted by inadequate and unsafe water supply. Owing to the difficulty in identifying pathogenic microorganisms, combined to the precarious conditions of water in specific regions, waterborne infections are a major cause of concern worldwide (Cabral 2010; Aw and Rose 2012). The need to guarantee a safe water supply leads to the search and the development of ways to detect these pathogens in the water (Leonard et al. 2003). A very used and well-established method for the identification of pathogens is the preparation of a culture medium in which the microbial organisms will multiply and their determination will be carried out. However, these culture-based methods present some drawbacks, such as the long time required for the analysis, the impossibility of being used on-site, and the inability to detect traces of microorganisms. Therefore, accurate and fast methods capable to detect pathogens is crucial for preventing infections and outbreaks (Aw and Rose 2012; Xu 2012). In this way, the development of chemical sensors may represent a viable alternative for the detection of pathogens. The sensors must fulfill some important requirements, though. They

should present high sensitivity and low detection limit in order to detect a few cells of the pathogen, which is sufficient to cause health problems. In addition, the sensors must provide rapid results, assisting in the correct treatment choice and restricting the spread of pathogens. It is also desired a high specificity toward target cells and the possibility of performing in-situ detections (Heo and Hua 2009).

Immunosensors are sensors whose transduction mechanism is based on the specific interaction of a very stable complex formed between an antibody and an antigen (Felix and Angnes 2017). The detection is performed through the immobilization of the antibody or the antigen onto a transducer surface, which converts the complex formation into a measurable signal. Antigens can be biological pathogens, such as bacteria, viruses, and fungi, that form a stable complex with a specific antibody. Due to the specificity character, immunosensors present high selectivity and good sensitivity (Moina and Ybarra 2012; Felix and Angnes 2017).

Luo et al. developed an immunosensor for rapid and sensitive detection of the target pathogen Escherichia coli (E. Coli) O157:H7 and bovine viral diarrhea virus (BVDV) using direct electrical measurements (Luo et al. 2010). The biosensor was made based on electrospun cellulose nitrate (CN) nanofibers. The CN nanofibers were chosen because of their biocompatibility with proteins and the high surface area, which allows occurrence of more biological events, facilitating the assay. The sensor architecture consists of three pads: an application pad, a capture pad, and an absorption pad. The application pad was used to deposit the sample under investigation and to control the flow of the sample in the capture pad, while the absorption pad modulates the capillary action. Conductive magnetic nanoparticles were modified with antibodies and incubated with the sample to conjugate with the target pathogen. Next, a magnetic separation process was used for the sample purification. The CN nanofibers were plasma-treated to enhance the capillary performance and implemented as the capture pad after being immobilized with the specific secondary antibody. Silver electrodes were fabricated using spray deposition of silver paint and recognized the electron transport path through the formation of conductive sandwich complexes between antibodies and pathogen. After achieving capillary flow equilibrium, the direct-charge transfer to the electrode was measured. The amount of E. coli O157:H7 or BVDV in the sample was proportional to the captured sandwich complex that generated the electrical signal, leading to a change in the resistance values detected by the electrodes. The detection principle of the immunosensor is shown in the scheme of Fig. 5.6a. The CN nanofiber-based immunosensor showed a linear response for E. coli O157:H7 concentration from 0 to 10⁴ colonyforming unit per milliliter (CFU mL⁻¹) and a sensitivity of 61 CFU mL⁻¹. For the BVDV sample, the detection limit was estimated to be 10³ cell culture infectious dose per milliliter (CCDI mL⁻¹).

In a similar work, the detection of two influenza A viruses by a magnetic biosensor using CN membrane and magnetic beads was proposed by Hong et al. (2011)). Viruses were adsorbed to the CN membrane and specific antibodies were coupled to magnetic beads, enabling the formation of antigen–antibody complexes. The complexes were recognized based on the frequency mixing detection through the generation of signals that reflect the concentration of the magnetic beads and



Fig. 5.6 (a) Schematic illustration of the principle of detection of the immunosensor based on electrospun functionalized with antibodies. Reprinted with permission from Luo et al. (2010). Copyright (2010) Elsevier. (b) (i) Illustration of the process used to obtain the P-paper; (ii) cross-sectional SEM image of the P-paper; and (iii) the fabricated P-paper electrode for electrochemical detection of the pathogen. Adapted and reprinted with permission from Yang et al. (2016)). Copyright (2016) American Chemical Society. (c) Digital pictures of the cellulose paper and the distinct coloration developed after hybridization with (i) complementary (target) DNA, (ii) non-complementary DNA, and (iii) the control. Adapted and reprinted with permission from Saikrishnan et al. (2014)). Copyright (2014) Springer

consequently the amount of complex formation. The assay was demonstrated to be superior in some aspects (in terms of reduction of the assay time and robustness) to conventional enzyme-linked immunoflow assay (ELIFA).

Aptasensors have emerged as an interesting alternative for the detection of live pathogens. These biosensors are based on aptamers, that is, single-stranded DNA (ssDNA) or RNA molecules that mimic antibodies' properties. In the detection mechanism of pathogens, aptamers work as artificial antibodies recognizing target molecules through the formation of aptamer-analyte complexes. Aptasensors may present some advantages on immunosensors based on antibodies, namely high specificity, better stability, and ease of synthesis (Luppa et al. 2001; Felix and Angnes 2017). In this context, the selective detection of *Staphylococcus aureus* (S. aureus) was performed by an electrochemical aptasensor proposed by Ranjbar and Shahrokhian based on cellulose nanofibers (CNFs), carbon nanoparticles (CNPs), and gold nanoparticles (AuNPs; Ranjbar and Shahrokhian 2018). The desired features of CNFs for the biosensor were combined to the excellent conductivity of CNPs and AuNPs, besides the ability to capture a larger quantity of analytes due to the high surface area of the latter. Electrochemical impedance spectroscopy (EIS) measurements were conducted to perform the detection of S. aureus, in which an analytical signal was recorded and the presence of the pathogen caused a change in the charge transfer resistance (R_{ct}) . The limit of detection of the aptasensor was found to be 1 CFU mL⁻¹ with a wide linear dynamic range, which demonstrated the potential to detect *S. aureus* in human blood serum samples.

Lignin was used by Cerrutti et al. (2015) to fabricate an immunosensor for human immunodeficiency virus (HIV). In their work, an immunosensor based on layer-by-layer (LbL) films of lignin and an antigenic peptide (p-17) was proposed in the recognition of a specific antibody for HIV. The lignin was used due to its biocompatibility and availability and acted by immobilizing the p-17 in the LbL films. The lignin matrix preserved the structure of the peptide, which is essential for molecular recognition. Gold interdigitated electrodes were used in the electrical measurements conducted in phosphate-buffered saline (PBS) solution, and the lowest concentration of the specific antibody used, that is, 0.1 ng mL⁻¹, was comparable to other electrochemical biosensors.

Yang and coworkers used newspaper as a source of cellulose fiber to fabricate a flexible and disposable sensor by coating the fiber with parylene C in order to detect the pathogenic E. coli O157:H7 based on DNA hybridization (Yang et al. 2016). The parylene-C-coated paper (P-paper) was obtained by a dry chemical vapor deposition (CVD) method, which enabled an enhancement in the mechanical properties and an increase in the hydrophobicity of the sensor while maintaining its porous structure. Such features allowed its use in electrochemical measurements in aqueous media. The schematic illustration of the fabrication of the P-paper is shown in Fig. 5.6b(i), and the cross-sectional scanning electron microscope (SEM) image of P-paper is presented in Fig. 5.6b(ii). The paper electrodes were obtained via patterning of gold and silver layers on P-paper. To detect the pathogenic E. coli O157:H7, the single-stranded DNA (ssDNA) was immobilized onto the gold working electrode (Fig. 5.6c(iii)). The ssDNA is capable of recognizing the target complementary DNA (cDNA) of E. Coli O157:H7. EIS was employed and the increase in the $R_{\rm ct}$ values upon increasing target cDNA concentration was evaluated, yielding a detection limit of 0.16 nmol L^{-1} .

Detection of pathogenic bacteria was also conducted by Saikrishnan et al. using a cellulose-paper-based colorimetric bioassay through the detection of the target pathogen DNA (Saikrishnan et al. 2014). The cellulose was tosylated to achieve a very porous surface and then probe of oligonucleotides was immobilized covalently on the tosylated cellulose surface. The principle of detection was based on the manifestation of a blue color derived from the specific hybridization that occurs with a biotin-labeled oligonucleotide and the probe attached to the cellulose surface. A streptavidin-horseradish peroxidase (HRP) acted as a transducer providing the color change that indicated the biorecognition event. The detection of the target DNA resulted in a strong coloration of the cellulose paper, while lower intensities of blue spots were observed with non-complementary DNA and no color change was noticed in a negative control experiment (without DNA oligonucleotide), as can be seen in Fig. 5.6c. Polymerase chain reaction (PCR) was applied to test the biosensor with the pathogen DNA from *Mycobacterium tuberculosis*, and the results showed an expected blue color corresponding to the complementary probe. Visual detection of pathogenic bacteria was also performed by Tait et al. based on the generation of volatile organic compounds (VOCs) trapped in agarose gel (Tait et al. 2015). The detection was possible because the addition of an enzyme substrate to a bacteria sample generated VOCs, which were trapped in the agarose gel containing color-generating reagents. Agarose is inexpensive and was modified with specific and selective colorimetric reagents in order to show a change of color in the presence of target compounds. *Enterococcus faecium* and *Klebsiella pneumoniae* produce β -glucosidase and, when prepared with 2-nitrophenyl- β -Dglucoside, releasing 2-nitrophenol, generate a yellow color within the agarose gel. Other bacteria that do not have activity toward 2-nitrophenyl- β -D-glucoside showed no change of color. *Pseudomonas aeruginosa*, in turn, is a known β -alanyl aminopeptidase producer and hydrolyzes the trifluoroacetic acid (TFA) salt of 3-amino-*N*phenylpropanamide, changing the color of the gel to orange. The detection method allowed a simple, low-cost optical discrimination and identification of β -glucosidase producing bacteria and β -alanine aminopeptidase producing species.

5.3.3 Green Composites Applied in Sensors for Detecting Hazardous Volatiles

Concern about emission control and detection of hazardous volatiles, including volatile organic compounds (VOCs), has been increasing in the last years. VOCs are generally colorless (which hinders their direct detection by optical spectroscopy), often have low boiling point and are highly reactive and frequently mixed with interfering gases, which requires the development of sensors with high selectivity (Vaishnav et al. 2005). Some VOCs may be highly toxic and/or carcinogenic and may harm human health and the ecosystem as a whole. The common methods used for detecting hazardous volatiles are gas chromatography and mass spectrometry (De Gouw et al. 2003; Dincer et al. 2006), due to their low limit of detection. However, both apparatuses are expensive and analyses are time-consuming, which restrict their use for in-situ and real-time analysis. For this reason, there is an increasing interest in developing portable gas sensors that display high sensitivity, selectivity, and stability for specific applications (Khanna 2011).

Investigations involving gas sensor analyses have grown considerably in recent years. Approaches based on electrical (Amer et al. 2019), electrochemical (Pang et al. 2018), and optical (Terra et al. 2018) sensors employing different types of materials as active layers, such as polymers, metal nanoparticles, carbon nanotubes (CNTs), semiconductor metal oxides have been proposed (Andre et al. 2018). For instance, Khattab et al. (2019) proposed a solid-state reversible colorimetric sensor for detecting ammonia gas, which can be very harmful and cause serious health problems (Rousidou et al. 2013). In their work, bromocresol was added to a carboxymethylcellulose matrix for developing an easy-to-use and reversible ammonia gas sensor with excellent reversibility and short detection time. The sensor was

yellow before ammonia gas exposure, and changed to purple when exposed to ammonia, and the system exhibited a rapid response time and low detection limit of 9.0×10^{-2} ppb for aqueous ammonia.

Zhang et al. (2018b) used graphene/ethylcellulose (EC) nanocomposites to produce a low-cost and flexible VOC gas sensor by depositing a graphene/EC nanocomposite as a thin film onto a flexible polyethylene terephthalate (PET) substrate. The sensorial platform showed no sensitivity to deformation and yielded excellent responses to VOCs at room temperature with detection limits ranging from 37 to 167 ppm. The developed sensor showed potential to be employed in portable electronic products, labels, and intelligent packaging for the monitoring of perishable products.

Although PET and glass microfiber "paper" are known for producing flexible sensing platforms, both are of synthetic origin and display very long life cycle (Andre et al. 2017). Cellulose-based paper, on the other hand, is an ecologically favorable alternative, once it is derived from cellulose, and presents flexibility, versatility, porosity, and low cost, combined to easier degradability. Paper-based platforms used as optoelectronic devices are usually lighter, thinner, and can be easily moldable or cut using scissors, and these features enable their application in colorimetric, electrochemical, and fluorescence sensors (Khansili et al. 2018; Singh et al. 2018; Kanaparthi and Singh 2019). On the other hand, some paper substrates can present high roughness and absorption capacity, factors that can limit its use in flexible electronic devices. This problem can be overcome by treating the paper surface in order to limit its absorption and soften its surface (Bundy and Ishley 1991; Singh et al. 2018). Another possibility is the use of paper composed of cellulosic nanostructures, whose dimensions at the nanoscale exhibit high contact surface area and mechanical resistance (Kim et al. 2015).

Different types of materials can be associated to paper substrates to act as a sensitive layer in gas sensor assays, providing sensitivity and selectivity toward varied analytes. Among the materials employed for this purpose, we can highlight carbon nanotubes (CNTs), conductive polymers, metal oxides, graphene and graphene oxide, and metals. Several gas sensors based on flexible platforms have been reported in the literature, where usually the paper is the sensory platform, while a thin layer of sensitive material is deposited on it, enabling an efficient and economical detection of toxic gases (Moseley 2017). For instance, in the work of Niu et al. (2016), the authors developed an electronic gas sensor that detects ammonia at room temperature by depositing a thin film of polyaniline/reduced graphene oxide (PANI/ RGO) on a flexible green paper substrate. The porosity presented by the paper substrate was preserved even after deposition of the thin film, and its roughness is very important for the formation of thin PANI/RGO film coating responsible for the detection.

Maity and Ghosh (2018) reported the development of an inexpensive and disposable paper sensor that can be used at room temperature for ammonia gas sensing. The authors reported that the sensor was able to detect the presence of small amounts of ammonia leading to color changes. For detection, a thin film of perovskite halide CH₃NH₃PbI₃ (MAPI) was deposited on the paper, which showed black color when



Fig. 5.7 Color change response of the MAPI sensor, where (a) represents the original black color of the unexposed film of perovskite halide $CH_3NH_3PbI_3$ (MAPI) and (b) represents the yellow color in the presence of ammonia gas. Reprinted with permission from Maity and Ghosh (2018)). Copyright (2018) Nature

in contact with the paper, and yellowish coloring in the presence of the ammonia gas, as can be observed in the images of Fig. 5.7a and b. An important feature of this type of sensor is that there was no need for any extra equipment for its operation, since it is a visual sensor, where the color change could be easily detected with the naked eye.

In addition to being environmentally friendly, gas sensors that use paper as a substrate have become an important emerging class of electronic devices because of their flexibility. Salarbashi et al. (2019) constructed a gas sensor with high sensitivity and flexibility using PbS nanowires processed in solution as the detection material. The device developed demonstrated an excellent detection response after exposure to NO_2 at room temperature, and showed no decrease in performance under different angles of curvature throughout the detection cycles. The results suggested the promising use of paper substrates in flexible devices applied in gas sensor assays.

5.4 Final Remarks

A recent survey of different types of sensors, including electronic tongues and electrochemical and optical chemical sensors, for monitoring distinct substances, using green composite materials was here presented. Specifically, for the sensor design we focused on cellulose and other natural macromolecules combined to carbonbased, metallic, and oxide nanoparticles, to be used as active layers or in some cases as substrates for varied sensors aimed at monitoring heavy metals in water, bacterial contamination, and volatile organic compounds. The green composites produced for this purpose present remarkable properties and can be considered as an alternative to the use of single or isolated materials as active layers in sensors, improving sensor sensitivity, limit of detection, decreasing cost, etc. The use of low-cost and green materials for producing electrical and optical devices, including sensors, that are cheap, portable, and easy to operate appears to be a general approach that will push new developments in the area, including the scaled-up industrial production of such devices.

Acknowledgments The authors thank the financial support from Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) (grant numbers: 2017/12174-4, 2017/10582-8, 2017/20973-4, 2018/09414-6, 2018/22214-6), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) (155512/2018-8), MCTI-SisNano (CNPq/402.287/2013-4), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Código de Financiamento 001, and Rede Agronano-EMBRAPA from Brazil.

References

- Abbasi-Moayed S, Golmohammadi H, Bigdeli A, Hormozi-Nezhad MR (2018) A rainbow ratiometric fluorescent sensor array on bacterial nanocellulose for visual discrimination of biothiols. Analyst 143:3415–3424. https://doi.org/10.1039/c8an00637g
- Alkmim-Filho JF, Germano A, Dibai WLS, Vargas EA, Melo MM (2014) Heavy metals investigation in bovine tissues in Brazil. Food Sci Technol 34:110–115. https://doi.org/10.1590/ S0101-20612014005000013
- Alonso GA, Istamboulie G, Noguer T, Marty J, Mu R (2012) Rapid determination of pesticide mixtures using disposable biosensors based on genetically modified enzymes and artificial neural networks. Sensors Actuators B Chem 164:22–28. https://doi.org/10.1016/j.snb.2012.01.052
- Amer K, Elshaer AM, Anas M, Ebrahim S (2019) Fabrication, characterization, and electrical measurements of gas ammonia sensor based on organic field effect transistor. J Mater Sci Mater Electron 30:391–400. https://doi.org/10.1007/s10854-018-0303-7
- Andre RS, Chen J, Kwak D, Correa DS, Mattoso LHC, Lei Y (2017) A flexible and disposable poly(sodium 4-styrenesulfonate)/polyaniline coated glass microfiber paper for sensitive and selective detection of ammonia at room temperature. Synth Met 233:22–27. https://doi.org/10.1016/j.synthmet.2017.08.005
- Andre RS, Sanfelice RC, Pavinatto A, Mattoso LHC, Correa DS (2018) Hybrid nanomaterials designed for volatile organic compounds sensors: a review. Mater Des 156:154–166. https:// doi.org/10.1016/j.matdes.2018.06.041
- Andre RS, Dos Santos DM, Mercante LA, Facure MHM, Campana-Filho SP, Mattoso LHC, Correa DS (2020) Nanochitin-based composite films as a disposable ethanol sensor. J Environ Chem Eng 8(5):104163. https://doi.org/10.1016/j.jece.2020.104163
- Anichini C, Czepa W, Pakulski D, Aliprandi A, Ciesielski A, Samorì P (2018) Chemical sensing with 2D materials. Chem Soc Rev 47:4860–4908. https://doi.org/10.1039/C8CS00417J
- Antonacci A, Arduini F, Moscone D, Palleschi G, Scognamiglio V (2018) Nanostructured (Bio) sensors for smart agriculture. TrAC Trends Anal Chem 98:95–103. https://doi.org/10.1016/j. trac.2017.10.022
- Arduini F, Cinti S, Scognamiglio V, Moscone D, Palleschi G (2017) How cutting-edge technologies impact the design of electrochemical (bio)sensors for environmental analysis. A review. Anal Chim Acta 959:15–42. https://doi.org/10.1016/j.aca.2016.12.035
- Arduini F, Cinti S, Caratelli V, Amendola L, Palleschi G, Moscone D (2019) Origami multiple paper-based electrochemical biosensors for pesticide detection. Biosens Bioelectron 126:346–354. https://doi.org/10.1016/j.bios.2018.10.014

- Arino C, Serrano N, Diaz-Cruz JM, Esteban M (2017) Voltammetric determination of metal ions beyond mercury electrodes. A review. Anal Chim Acta 990:11–53. https://doi.org/10.1016/j. aca.2017.07.069
- Aw TG, Rose JB (2012) Detection of pathogens in water: from phylochips to qPCR to pyrosequencing. Curr Opin Biotechnol 23:422–430. https://doi.org/10.1016/j.copbio.2011.11.016
- Azmi NA, Ahmad SH, Low SC (2018) Detection of mercury ions in water using a membranebased colorimetric sensor. RSC Adv 8:251–261. https://doi.org/10.1039/c7ra11450h
- Ban DK, Paul S (2018) Rapid colorimetric and spectroscopy based sensing of heavy metal and cellular free oxygen radical by surface functionalized silver nanoparticles. Appl Surf Sci 458:245–251. https://doi.org/10.1016/j.apsusc.2018.07.069
- Ban DK, Pratihar SK, Paul S (2015) Controlled modification of starch in the synthesis of gold nanoparticles with tunable optical properties and their application in heavy metal sensing. RSC Adv 5:81554–81564. https://doi.org/10.1039/c5ra16473g
- Bandodkar AJ, Wang J (2014) Non-invasive wearable electrochemical sensors : a review. Trends Biotechnol 32:363–371. https://doi.org/10.1016/j.tibtech.2014.04.005
- Bard AJ, Faulkner LR (2000) Electrochemical methods: fundamentals and applications, 2nd edn. Wiley, New York
- Bhand S, Bacher G (2017) Impedimetric sensors in environmental analysis: An overview. 67–85. https://doi.org/10.1007/978-981-10-7751-7_4
- Brett CMA, Brett AM (1994) Electrochemistry- principles, methods and applications
- Bundy WM, Ishley JN (1991) Kaolin in paper filling and coating. Appl Clay Sci 5:397–420. https://doi.org/10.1016/0169-1317(91)90015-2
- Cabral JPS (2010) Water microbiology. Bacterial pathogens and water. Int J Environ Res Public Health 7:3657–3703. https://doi.org/10.3390/ijerph7103657
- Cerrutti BM, Moraes ML, Pulcinelli SH, Santilli CV (2015) Lignin as immobilization matrix for HIV p17 peptide used in immunosensing. Biosens Bioelectron 71:420–426
- Chaiendoo K, Tuntulani T, Ngeontae W (2017) A paper-based ferrous ion sensor fabricated from an ion exchange polymeric membrane coated on a silver nanocluster-impregnated filter paper. Mater Chem Phys 199:272–279. https://doi.org/10.1016/j.matchemphys.2017.07.013
- Chen W, Hu C, Yang Y, Cui J, Liu Y (2016) Rapid synthesis of carbon dots by hydrothermal treatment of lignin. Materials (Basel) 9. https://doi.org/10.3390/ma9030184
- Chin K, Ting SS, Ong HL, Omar M (2017) Surface functionalized nanocellulose as a veritable inclusionary material in contemporary bioinspired applications : A review. 46065: https://doi. org/10.1002/app.46065
- Christau S, Genzer J, von Klitzing R (2015) Polymer Brush/Metal Nanoparticle Hybrids for Optical Sensor Applications: from Self-Assembly to Tailored Functions and Nanoengineering. Zeitschrift für Phys Chemie:229. https://doi.org/10.1515/zpch-2014-0573
- Cichosz S, Masek A, Zaborski M (2018) Polymer-based sensors: a review. Polym Test 67:342–348. https://doi.org/10.1016/j.polymertesting.2018.03.024
- Cinti S (2019) Novel paper-based electroanalytical tools for food surveillance
- Correa DS, Medeiros ES, Oliveira JE, Paterno LG, Mattoso LHC (2014) Nanostructured conjugated polymers in chemical sensors: synthesis , properties and applications. J Nanosci Nanotechnol 14:1–19. https://doi.org/10.1166/jnn.2014.9362
- Correa DS, Pavinatto A, Mercante LA, Mattoso LHC, Oliveira JE, Riul A (2017) 6 Chemical sensors based on hybrid nanomaterials for food analysis. In: Grumezescu AMBT-N (ed). Academic Press, pp 205–244
- Cui L, Wu J, Ju H (2015) Electrochemical sensing of heavy metal ions with inorganic, organic and bio-materials. Biosens Bioelectron 63:276–286. https://doi.org/10.1016/j.bios.2014.07.052
- Cui Y, Wang X, Zhang Q, Zhang H, Li H, Meyerhoff M (2019) Colorimetric copper ion sensing in solution phase and on paper substrate based on catalytic decomposition of S-nitrosothiol. Anal Chim Acta 1053:155–161. https://doi.org/10.1016/j.aca.2018.11.050

- Carvalho LA De, Bueno PR, Química I De, Paulista UE, Sp A (2006) Espectroscopia De Impedância Eletroquímica Aplicada Ao Estudo Das Reações 29:796–804 . https://doi. org/10.1590/S0100-40422006000400029
- De Gouw J, Warneke C, Karl T, Eerdekens G, Van der Veen C, Fall R (2003) Sensitivity and specificity of atmospheric trace gas detection by proton-transfer-reaction mass spectrometry. Int J Mass Spectrom 223–224:365–382. https://doi.org/10.1016/S1387-3806(02)00926-0
- de Oliveira TR, Fonseca WT, de Oliveira SG, Faria RC (2019) Fast and flexible strategy to produce electrochemical paper-based analytical devices using a craft cutter printer to create wax barrier and screen-printed electrodes. Talanta 195:480–489. https://doi.org/10.1016/j. talanta.2018.11.047
- Deisingh AK, Thompson M (2004) Biosensors for the detection of bacteria. Can J Microbiol 50:69–77. https://doi.org/10.1139/W03-095
- Di Rosa AR, Leone F, Cheli F, Chiofalo V (2017) Fusion of electronic nose, electronic tongue and computer vision for animal source food authentication and quality assessment – a review. J Food Eng 210:62–75. https://doi.org/10.1016/j.jfoodeng.2017.04.024
- Dincer F, Odabasi M, Muezzinoglu A (2006) Chemical characterization of odorous gases at a landfill site by gas chromatography-mass spectrometry. J Chromatogr A 1122:222–229. https:// doi.org/10.1016/j.chroma.2006.04.075
- Duffy GF, Moore EJ (2017) Electrochemical Immunosensors for food analysis : a review of recent developments. Anal Lett 50:1–32. https://doi.org/10.1080/00032719.2016.1167900
- Facure MHM, Mercante LA, Mattoso LHC, Correa DS (2017) Detection of trace levels of organophosphate pesticides using an electronic tongue based on graphene hybrid nanocomposites. Talanta 167:59–66. https://doi.org/10.1016/j.talanta.2017.02.005
- Fang HL, Zheng HX, Ou MY, Meng Q, Fan DH, Wang W (2011) One-step sensing lead in surface waters with screen printed electrode. Sensors Actuators B Chem 153:369–372. https://doi. org/10.1016/j.snb.2010.10.049
- Felix FS, Angnes L (2017) Electrochemical immunosensors a powerful tool for analytical applications. Biosens Bioelectron 102:470–478
- Ferreira DCM, Giordano GF, Soares CCDSP, De Oliveira JFA, Mendes RK, Piazzetta MH, Gobbi AL, Cardoso MB (2015) Optical paper-based sensor for ascorbic acid quantification using silver nanoparticles. Talanta 141:188–194. https://doi.org/10.1016/j.talanta.2015.03.067
- Garcia-Valls R, Hatton TA (2003) Metal ion complexation with lignin derivatives. Chem Eng J 94:99–105. https://doi.org/10.1016/S1385-8947(03)00007-X
- Giacometti JA, Alves N, Teruya MY (2016) Impedance of aqueous solutions of KCl at the ultralow frequency range: use of Cole-Cole impedance element to account for the frequency dispersion peak at 20 mHz. Brazilian J Phys 46:50–55. https://doi.org/10.1007/s13538-015-0381-4
- Golmohammadi H, Morales-Narváez E, Naghdi T, Merkoçi A (2017) Nanocellulose in sensing and biosensing. Chem Mater 29:5426–5446. https://doi.org/10.1021/acs.chemmater.7b01170
- Hayashi K (2015) Optical chemical sensor. IEEJ Trans Sen Micromachines 135:299–304. https:// doi.org/10.1021/cr068102g
- Heo J, Hua SZ (2009) An overview of recent strategies in pathogen sensing. Sensors 9:4483–4502. https://doi.org/10.3390/s90604483
- Hong H-B, Krause H-J, Song K-B, Choi C-J, Chung M-A, Son S, Offenhäusser A (2011) Detection of two different influenza A viruses using a nitrocellulose membrane and a magnetic biosensor. J Immunol Methods 365:95–100. https://doi.org/10.1016/j.jim.2010.12.005
- Hulanicki A, Glab S, Ingman F (1991) Chemical sensors: definitions and classification. Pure Appl Chem 63:1247–1250. https://doi.org/10.1351/pac199163091247
- Ilacas GC, Sosa JD, Gomez FA, Nelms KJ, Basa A, Liu Y (2019) Paper-based microfluidic devices for glucose assays employing a metal-organic framework (MOF). Anal Chim Acta 1055:74–80. https://doi.org/10.1016/j.aca.2019.01.009
- Jayathilaka WADM, Qi K, Qin Y, Chinnappan A, Serrano-Garcia W, Baskar C, Wang H, He J, Cui S, Thmas SW, Ramakrishna S (2019) Significance of nanomaterials in wearables : a review on wearable. Actuators Sensors 1805921:1–21. https://doi.org/10.1002/adma.201805921
- Jayawickramage RAP, Ferraris JP (2019) High performance supercapacitors using lignin based electrospun carbon nanofiber electrodes in ionic liquid electrolytes. J Hazard Mater. https://doi. org/10.1088/1361-6528/aafe95 Manuscript
- Jeronimo P, Araujo A, Montenegro MCBSM (2007) Optical sensors and biosensors based on solgel films. Talanta 72:13–27. https://doi.org/10.1016/j.talanta.2006.09.029
- Jose A, Ray JG (2018) Toxic heavy metals in human blood in relation to certain food and environmental samples in Kerala, South India. Environ Sci Pollut Res 25:7946–7953. https://doi.org/10.1007/s11356-017-1112-x
- Juan Zhao X, Qin Zhou Z (2015) Synthesis and applications of pectin-based nanomaterials. Curr Nanosci 12:103–109. https://doi.org/10.2174/1573413711666150818224020
- Kanaparthi S, Singh SG (2019) Solvent-free fabrication of a room temperature ammonia gas sensor by frictional deposition of a conducting polymer on paper. Org Electron 68:108–112. https://doi.org/10.1016/j.orgel.2019.01.053
- Karkra R, Kumar P, Bansod BKS, Bagchi S, Sharma P, Krishna CR (2017) Classification of heavy metal ions present in multi-frequency multi-electrode potable water data using evolutionary algorithm. Appl Water Sci 7:3679–3689. https://doi.org/10.1007/s13201-016-0514-0
- Kartel MT, Kupchik LA (1999) Pii: s0045-6535 (98)00466-4. Science (80-) 38:2591-2596
- Kassal P, Steinberg MD, Steinberg IM (2018) Wireless chemical sensors and biosensors: a review. Sensors Actuators B Chem 266:228–245. https://doi.org/10.1016/j.snb.2018.03.074
- Khachatryan G, Khachatryan K (2019) Starch based nanocomposites as sensors for heavy metals – detection of Cu2+ and Pb2+ ions. Int Agrophysics 33:121–126. https://doi.org/10.31545/ intagr/104414
- Khanna VK (2011) Nanosensors: physical, chemical, and biological
- Khansili N, Rattu G, Krishna PM (2018) Label-free optical biosensors for food and biological sensor applications. Sensors Actuators B Chem 265:35–49. https://doi.org/10.1016/j. snb.2018.03.004
- Khattab TA, Dacrory S, Abou-Yousef H, Kamel S (2019) Development of microporous cellulosebased smart xerogel reversible sensor via freeze drying for naked-eye detection of ammonia gas. Carbohydr Polym 210:196–203. https://doi.org/10.1016/j.carbpol.2019.01.067
- Kim JH, Shim BS, Kim HS, Lee YJ, Min SK, Jang D, Abas Z, Kim J (2015) Review of nanocellulose for sustainable future materials. Int J Precis Eng Manuf - Green Technol 2:197–213. https://doi.org/10.1007/s40684-015-0024-9
- Kirsanov D, Correa DS, Gaal G, Riul A, Braunger ML, Shimizu FM, Oliveira ON, Liang T, Wan H, Wang P, Oleneva E, Legin A (2019) Electronic tongues for inedible media. Sensors 19:5113. https://doi.org/10.3390/s19235113
- Kruse P (2018) Review on water quality sensors. J Pysics D Appl Phys 51:203002
- Kuek Lawrence CS, Tan SN, Floresca CZ (2014) A "green" cellulose paper based glucose amperometric biosensor. Sensors Actuators B Chem 193:536–541. https://doi.org/10.1016/j. snb.2013.11.054
- La Mantia FP, Morreale M (2011) Green composites: a brief review. Compos Part A Appl Sci Manuf 42:579–588. https://doi.org/10.1016/j.compositesa.2011.01.017
- Labib M, Sargent EH, Kelley SO (2016) Electrochemical methods for the analysis of clinically relevant biomolecules. Chem Rev 116:9001–9090. https://doi.org/10.1021/acs.chemrev.6b00220
- Lahcen AA, Amine A (2018) Recent advances in electrochemical sensors based on molecularly imprinted polymers and nanomaterials. Electroanalysis 31:188–201. https://doi.org/10.1002/ elan.201800623
- Lakowicz JR (2006) Principles of fluorescence spectroscopy. Springer, Boston
- Lee KY, Mooney DJ (2000) Alginate: properties and biomedical applications. Prog Polym Sci 37:106–126. https://doi.org/10.1016/j.progpolymsci.2011.06.003.Alginate
- Leonard P, Hearty S, Brennan J, Dunne L, Quinn J, Chakraborty T, Kennedy RO (2003) Advances in biosensors for detection of pathogens in food and water. Enzym Microb Technol 32:3–13
- Li C, Shi G (2013) Polythiophene-based optical sensors for small molecules. ACS Appl Mater Interfaces 5:4503–4510. https://doi.org/10.1021/am400009d

- Li Z, Askim JR, Suslick KS (2018) The optoelectronic nose: colorimetric and Fluorometric sensor arrays. Chem Rev 119:231–292. https://doi.org/10.1021/acs.chemrev.8b00226
- Li S, Pan N, Zhu Z, Li R, Li B, Chu J, Li G, Chang Y, Pan T (2019) All-in-one Iontronic sensing paper. Adv Funct Mater 1807343:1–11. https://doi.org/10.1002/adfm.201807343
- Liu G, Li L, Zhang K, Wang X, Chang J, Sheng Y, Bai L, Wen Y (2016) Facile preparation of water-processable biochar based on pitch pine and its electrochemical application for cadmium ion sensing. Int J Electrochem Sci 11:1041–1054
- Liu M, Li J, Li B (2017a) A colorimetric aptamer biosensor based on cationic polythiophene derivative as peroxidase mimetics for the ultrasensitive detection of thrombin. Talanta 175:224–228. https://doi.org/10.1016/j.talanta.2017.07.003
- Liu X, Na W, Liu H, Su X (2017b) Fluorescence turn-off-on probe based on polypyrrole/graphene quantum composites for selective and sensitive detection of paracetamol and ascorbic acid. Biosens Bioelectron 98:222–226. https://doi.org/10.1016/j.bios.2017.06.044
- Liu Y, Deng Y, Dong H, Liu K, He N (2017c) Progress on sensors based on nanomaterials for rapid detection of heavy metal ions. Sci China Chem 60:329–337. https://doi.org/10.1007/ s11426-016-0253-2
- Liu ML, Bin CB, Li CM, Huang CZ (2019) Carbon dots: synthesis, formation mechanism, fluorescence origin and sensing applications. Green Chem 21:449–471. https://doi.org/10.1039/ c8gc02736f
- Lobnik A, Turel M, Urek ŠK (2012a) Optical chemical sensors : design and applications, advances in chemical sensors. Prof Wen Wang:4–28. https://doi.org/10.5772/1102
- Lobnik A, Turel M, Urek ŠK (2012b) Optical chemical sensors: design and applications. Adv Chem Sen:3–28. https://doi.org/10.5772/31534
- Lowinsohn D, Bertotti M (2006) Sensores Eletroquímicos: Considerações sobre mecanismos de funcionamento e aplicações no monitoramento de espécies químicas em ambientes microscópicos. Quim Nova 29:1318–1325
- Luo Y, Nartker S, Miller H, Hochhalter D, Wiederoder M, Wiederoder S, Setterington E, Drzal LT, Alocilja EC (2010) Surface functionalization of electrospun nanofibers for detecting E. coli O157:H7 and BVDV cells in a direct-charge transfer biosensor. Biosens Bioelectron 26:1612–1617. https://doi.org/10.1016/j.bios.2010.08.028
- Luo C, Wang Y, Li X, Jiang X, Gao P, Sun K, Zhou J, Zhang Z, Jiang Q (2017) An optical sensor with polyaniline-gold hybrid nanostructures for monitoring pH in saliva. Nano 7:67. https:// doi.org/10.3390/nano7030067
- Luppa PB, Sokoll LJ, Chan DW (2001) Immunosensors principles and applications to clinical chemistry. Clin Chim Acta 314:1–26
- Ma Y, Li Y, Ma K, Wang Z (2018) Optical colorimetric sensor arrays for chemical and biological analysis. Sci China Chem 61:643–655. https://doi.org/10.1007/s11426-017-9224-3
- Maciel JV, Durigon AMM, Souza MM, Quadrado RFN, Fajardo AR, Dias D (2019) Polysaccharides derived from natural sources applied to the development of chemically modified electrodes for environmental applications: A review. Trends Environ Anal Chem:e00062. https://doi. org/10.1016/j.teac.2019.e00062
- Maduraiveeran G, Jin W (2017) Nanomaterials based electrochemical sensor and biosensor platforms for environmental applications. Trends Environ Anal Chem 13:10–23. https://doi.org/10.1016/j.teac.2017.02.001
- Mahapure PD, Gangal SA, Aiyer RC, Gosavi SW (2019) Combination of polymeric substrates and metal-polymer nanocomposites for optical humidity sensors. J Appl Polym Sci 136:47035. https://doi.org/10.1002/app.47035
- Maity A, Ghosh B (2018) Fast response paper based visual color change gas sensor for efficient ammonia detection at room temperature. Sci Rep 8:1–10. https://doi.org/10.1038/ s41598-018-33365-3
- Mao Y, Akram M, Shi J, Wen J, Yang C, Jiang J, Lu Z, Zhou B, Tian Y (2019) Optical oxygen sensors based on microfibers formed from fluorinated copolymers. Sensors Actuators B Chem 282:885–895. https://doi.org/10.1016/j.snb.2018.11.143

- Marques I, Magalhâes-Mota G, Pires F, Sério S, Ribeiro PA, Raposo M (2017) Detection of traces of triclosan in water. Appl Surf Sci 421:142–147. https://doi.org/10.1016/j.apsusc.2016.12.170
- Martins GF, Pereira AA, Stracçalano BA, Antunes PA, Pasquini D, Curvelo AAS, Ferreira M, Riul A, Constatino CJL (2008) Ultrathin films of lignins as a potential transducer in sensing applications involving heavy metal ions. Sensors Actuators B Chem 129:525–530. https://doi. org/10.1016/j.snb.2007.08.051
- McDonagh C, Burke CS, MacCraith BD (2008) Optical chemical sensors. Chem Rev 108:400–422. https://doi.org/10.1021/cr068102g
- McQuade DT, Pullen AE, Swager TM (2000) Conjugated polymer-based chemical sensors. Chem Rev 100:2537–2574. https://doi.org/10.1021/cr9801014
- Meng Z, Stolz RM, Mendecki L, Mirica KA (2019) Electrically-transduced chemical sensors based on two-dimensional nanomaterials. Chem Rev 119:478–598. https://doi.org/10.1021/ acs.chemrev.8b00311
- Mercante LA, Scagion VP, Migliorini FL, Mattoso LHC, Correa DS (2017) Electrospinning-based (bio)sensors for food and agricultural applications: a review. Trends Anal Chem 91:91–103. https://doi.org/10.1016/j.trac.2017.04.004
- Mercante LA, Andre RS, Mattoso LHC, Correa DS (2019) Electrospun ceramic nanofibers and hybrid-nanofiber composites for gas sensing. ACS Appl Nano Mater 2(7):4026–4042. https:// doi.org/10.1021/acsanm.9b01176
- Mohamad Ahad IZ, Wadi Harun S, Gan SN, Phang SW (2018) Polyaniline (PAni) optical sensor in chloroform detection. Sen Actuators B Chem 261:97–105. https://doi.org/10.1016/j. snb.2018.01.082
- Moina C, Ybarra G (2012) Fundamentals and applications of Immunosensors. In: Advances in Immunoassay Technology. pp. 65–80
- Moraes FC, Cincotto FH, Fatibello-Filho O, Faria RC, Fava EL (2018) A new disposable microfluidic electrochemical paper-based device for the simultaneous determination of clinical biomarkers. Talanta 195:62–68. https://doi.org/10.1016/j.talanta.2018.11.022
- Morales-Narváez E, Golmohammadi H, Naghdi T, Yousefi H, Kostiv U, Horák D, Pourreza N, Merkoçi A (2015) Nanopaper as an optical sensing platform. ACS Nano 9:7296–7305. https:// doi.org/10.1021/acsnano.5b03097
- Moseley PT (2017) Progress in the development of semiconducting metal oxide gas sensors: a review. Meas Sci Technol 28. https://doi.org/10.1088/1361-6501/aa7443
- Najarzadekan H, Sereshti H (2018) Transparent Polycaprolactam electrospun nanofibers doped with 1,10-phenanthroline optical sensor for colorimetric determination of Iron (II) and vitamin C. Fibers Polym 19:2149–2156. https://doi.org/10.1007/s12221-018-7568-6
- Narayanan KB, Han SS (2017) Colorimetric detection of manganese(II) ions using alginatestabilized silver nanoparticles. Res Chem Intermed 43:5665–5674. https://doi.org/10.1007/ s11164-017-2954-z
- Narayanaswamy R (1993) Tutorial review: optical chemical sensors: transduction and signal processing. Analyst 4:317–322. https://doi.org/10.1039/AN9931800317
- Naveen MH, Gurudatt NG, Shim Y-B (2017) Applications of conducting polymer composites to electrochemical sensors: a review. Appl Mater Today 9:419–433. https://doi.org/10.1016/j. apmt.2017.09.001
- Niu Z, Sun X, Wan P, Guo Y, Yan H, Lai S (2016) Flexible room-temperature gas sensors of Nanocomposite network-coated papers. Chem Select 1:2816–2820. https://doi.org/10.1002/ slct.201600648
- O'Keeffe S, Sullivan F, Woulfe P (2018) Optical fibre luminescence sensor for real-time LDR brachytherapy dosimetry. In: Gannot I (ed) Optical fibers and sensors for medical diagnostics and treatment applications XVIII. SPIE, p 37
- Paladiya C, Kiani A (2018) Nano structured sensing surface: significance in sensor fabrication. Sensors Actuators B Chem 268:494–511. https://doi.org/10.1016/j.snb.2018.04.085

- Pang X, Shaw MD, Gillot S, Lewis AC (2018) The impacts of water vapour and co-pollutants on the performance of electrochemical gas sensors used for air quality monitoring. Sensors Actuators B Chem 266:674–684. https://doi.org/10.1016/j.snb.2018.03.144
- Parmar J, Vilela D, Villa K, Wang J, Sánchez S (2018) Micro- and Nanomotors as active environmental microcleaners and sensors. J Am Chem Soc 140:9317–9331. https://doi.org/10.1021/ jacs.8b05762
- Patel M, Kumar R, Kishor K, Mlsna T, Pittman CU, Mohan D (2019) Pharmaceuticals of Emerging Concern in aquatic systems: chemistry, occurrence, effects, and removal methods. Chem Rev 119:3510–3673. https://doi.org/10.1021/acs.chemrev.8b00299
- Patella B, Sunseri C, Inguanta R, Fisica C, Industriale I (2019) Nanostructured based electrochemical sensors. J Nanosci Nanotechnol 19:3459–3470. https://doi.org/10.1166/jnn.2019.16110
- Peng B, Tang L, Zeng G, Zhou Y, Zhang Y, Long B, Fang S, Chen S, Yu J (2018) Current progress in aptasensors for heavy metal ions based on photoelectrochemical method: a review. Curr Anal Chem 14:4–12. https://doi.org/10.2174/1573411013666170412111128
- Podrazka M, Báczyńska E, Kundys M, Jeleń PS, Nery EW (2017) Electronic tongue-A tool for all tastes? Biosensors 8:1–24. https://doi.org/10.3390/bios8010003
- Ranjbar S, Shahrokhian S (2018) Design and fabrication of an electrochemical aptasensor using Au nanoparticles/carbon nanoparticles/cellulose nanofibers nanocomposite for rapid and sensitive detection of Staphylococcus aureus. Bioelectrochemistry 123:70–76. https://doi.org/10.1016/j. bioelechem.2018.04.018
- Rashid MI, Mujawar LH, Ismail IMI, El-Shahawi MS (2016) Styrofoam modified paper as a lowcost platform for qualitative and semi-quantitative determination of Ni2+ions in wastewater. Anal Methods 8:1496–1504. https://doi.org/10.1039/c5ay02394g
- Ray AK (2017) Organic materials for chemical sensing. In: Springer handbook of electronic and photonic materials. Springer, Cham, pp 1–1
- Reynoso EC, Torres E, Bettazzi F (2019) Trends and perspectives in Immunosensors for determination of currently-used pesticides: the case of glyphosate, Organophosphates. Biosensors 9:1–20. https://doi.org/10.3390/bios9010020
- Riul-Jr A, Dantas CAR, Miyazaki CM, Oliveira-Jr ON (2010) Recent advances in electronic tongues. Analyst 135:2481–2495. https://doi.org/10.1039/c0an00292e
- Roman J, Neri W, Derré A, Poulin P (2019) Electrospun lignin-based twisted carbon nanofibers for potential microelectrodes applications. Carbon NY 145:556–564. https://doi.org/10.1016/j. carbon.2019.01.036
- Rousidou C, Papadopoulou ES, Kortsinidou M, Giannakou IO, Singh BK, Menkissoglu-Spiroudi U, Karpouzas DG (2013) Bio-pesticides: harmful or harmless to ammonia oxidizing microorganisms? The case of a Paecilomyces lilacinus-based nematicide. Soil Biol Biochem 67:98–105. https://doi.org/10.1016/j.soilbio.2013.08.014
- Roy D, Semsarilar M, Guthrie JT, Perrier S (2009) Cellulose modification by polymer grafting: a review. Chem Soc Rev 38:2046–2064. https://doi.org/10.1039/b808639g
- Rudnitskaya A, Evtuguin DV, Costa LC, Graça MPF, Fernandes AJS, Correia MRP, Gomes MTSR, Oliveira JABP (2013) Potentiometric chemical sensors from lignin–poly(propylene oxide) copolymers doped by carbon nanotubes. Analyst 138:501–508. https://doi.org/10.1039/ C2AN36390A
- Ruecha N, Shin K, Chailapakul O, Rodthongkum N (2019) Label-free paper-based electrochemical impedance immunosensor for human interferon gamma detection. Sensors Actuators B Chem 279:298–304. https://doi.org/10.1016/j.snb.2018.10.024
- Sahatiya P, Jones SS, Badhulika S (2018) 2D MoS2–carbon quantum dot hybrid based large area, flexible UV–vis–NIR photodetector on paper substrate. Appl Mater Today 10:106–114. https:// doi.org/10.1016/j.apmt.2017.12.013
- Saikrishnan D, Goyal M, Rossiter S, Kukol A (2014) A cellulose-based bioassay for the colorimetric detection of pathogen DNA. Anal Bioanal Chem 406:7887–7898. https://doi.org/10.1007/ s00216-014-8257-y

- Saithongdee A, Praphairaksit N, Imyim A (2014) Electrospun curcumin-loaded zein membrane for iron(III) ions sensing. Sensors Actuators B Chem 202:935–940. https://doi.org/10.1016/j. snb.2014.06.036
- Salarbashi D, Bazeli J, Tafaghodi M (2019) Environment-friendly green composites based on soluble soybean polysaccharide: a review. Int J Biol Macromol 122:216–223. https://doi. org/10.1016/j.ijbiomac.2018.10.110
- Salavagione HJ, Díez-Pascual AM, Lázaro E, Vera S, Gómez-Fatou MA (2014) Chemical sensors based on polymer composites with carbon nanotubes and graphene: the role of the polymer. J Mater Chem A 2:14289–14328. https://doi.org/10.1039/C4TA02159B
- Sanfelice RC, Gonçalves VC, Balogh DT (2014) Langmuir and Langmuir-Schaefer films of poly(3hexylthiophene) with gold nanoparticles and gold nanoparticles capped with 1-octadecanethiol. J Phys Chem C 118:12944–12951. https://doi.org/10.1021/jp503083k
- Sanfelice RC, Mercante LA, Pavinatto A, Tomazio NB, Mendonça CR, Ribeiro SJL, Mattoso LHC, Correa DS (2017) Hybrid composite material based on polythiophene derivative nanofibers modified with gold nanoparticles for optoelectronics applications. J Mater Sci 52:1919–1929. https://doi.org/10.1007/s10853-016-0481-8
- Schoolaert E, Hoogenboom R, De Clerck K (2017) Colorimetric nanofibers as optical sensors. Adv Funct Mater 27:1702646. https://doi.org/10.1002/adfm.201702646
- Seely GR, Hart RL (1974) The binding of alkaline earth metal ions to alginate. Macromolecules 7:706–710. https://doi.org/10.1021/ma60041a030
- Singh AT, Lantigua D, Meka A, Taing S, Pandher M, Camci-Unal G (2018) Paper-based sensors: emerging themes and applications. Sensors (Switzerland) 18:1–22. https://doi.org/10.3390/ s18092838
- Stradiotto NR, Yamanaka H, Zanoni MVB (2003) Review electrochemical sensors : a powerful tool in analytical chemistry. J Braz Chem Soc 14:159–173
- Suginta W, Khunkaewla P, Schulte A (2013) Electrochemical biosensor applications of polysaccharides chitin and chitosan. Chem Rev 113:5458–5479. https://doi.org/10.1021/cr300325r
- Sun B, Xie G, Jiang Y, Li X (2011) Comparative CO2-sensing characteristic studies of PEI and PEI/starch thin film sensors. Energy Procedia 12:726–732. https://doi.org/10.1016/j. egypro.2011.10.098
- Tait E, Stanforth SP, Reed S, Perry JD, Dean JR (2015) Analysis of pathogenic bacteria using exogenous volatile organic compound metabolites and optical sensor detection. RSC Adv 5:15494–15499. https://doi.org/10.1039/c4ra13914c
- Tan DSY, Impas MGW, Camacho DH, Palisoc ST (2018) Paper-based electrode using Cladophora cellulose- polyaniline composite for electrochemical quantification of toxic lead (II) 52:853–861
- Taylor DM, Macdonald AG (1987) AC admittance of the metal / insulator / electrolyte interface. J Phys D Appl Phys 20:1277–1283
- Tchounwou PB, Yedjou CG, Patlolla AK, Sutton DJ (2012) Heavy metals toxicity and the environment. Mol Clin Environ Toxicol 101:133–164. https://doi.org/10.1007/978-3-7643-8340-4
- Teodoro KBR, Sanfelice RC, Mattoso LHC, Correa DS (2018) Cellulose whiskers influence the morphology and antibacterial properties of silver nanoparticles composites. J Nanosci Nanotechnol 18:4876–4883. https://doi.org/10.1166/jnn.2018.15285
- Teodoro KBR, Migliorini FL, Facure MHM, Correa DS (2019a) Conductive electrospun nanofibers containing cellulose nanowhiskers and reduced graphene oxide for the electrochemical detection of mercury(II). Carbohydr Polym 207:747–754. https://doi.org/10.1016/j. carbpol.2018.12.022
- Teodoro KBR, Shimizu FM, Scagion VP, Correa DS (2019b) Ternary nanocomposites based on cellulose nanowhiskers, silver nanoparticles and electrospun nanofibers: use in an electronic tongue for heavy metal detection. Sensors Actuators B Chem 290:387–395. https://doi. org/10.1016/j.snb.2019.03.125

- Terra IAA, Sanfelice RC, Valente GT, Correa DS (2018) Optical sensor based on fluorescent PMMA/PFO electrospun nanofibers for monitoring volatile organic compounds. J Appl Polym Sci 135:46128. https://doi.org/10.1002/app.46128
- Thomas SW, Joly GD, Swager TM (2007) Chemical sensors based on amplifying fluorescent conjugated polymers. Chem Rev 107:1339–1386. https://doi.org/10.1021/cr0501339
- Tingaut P, Zimmermann T, Sèbe G (2012) Cellulose nanocrystals and microfibrillated cellulose as building blocks for the design of hierarchical functional materials. J Mater Chem 22:20105. https://doi.org/10.1039/c2jm32956e
- Titirici M-M, Baker D, Schlee P, Hosseinaei O, Cazorla-Amorós D, Herou S, Tomani P, Landmér A, Mostazo-López MJ (2019) From waste to wealth: from Kraft lignin to free-standing Supercapacitors. Carbon N Y 145:470–480. https://doi.org/10.1016/j.carbon.2019.01.035
- Usha S, Mishra S, Gupta B (2015) Fabrication and characterization of a SPR based Fiber optic sensor for the detection of chlorine gas using silver and zinc oxide. Materials (Basel) 8:2204–2216. https://doi.org/10.3390/ma8052204
- Vaishnav VS, Patel PD, Patel NG (2005) Indium tin oxide thin film gas sensors for detection of ethanol vapours. Thin Solid Films 490:94–100. https://doi.org/10.1016/j.tsf.2005.04.006
- Vikesland PJ (2018) Nanosensors for water quality monitoring. Nat Nanotechnol 13:651–660. https://doi.org/10.1038/s41565-018-0209-9
- Wang J, Jákli A, West JL (2018) Liquid crystal/polymer fiber mats as sensitive chemical sensors. J Mol Liq 267:490–495. https://doi.org/10.1016/j.molliq.2018.01.051
- Wei Y, Gao C, Meng FL, Li HH, Wang L, Liu JH, Huang XJ (2012) SnO2/reduced graphene oxide nanocomposite for the simultaneous electrochemical detection of cadmium(II), lead(II), copper(II), and mercury(II): An interesting favorable mutual interference. J Phys Chem C 116:1034–1041. https://doi.org/10.1021/jp209805c
- Wu Y, Pang H, Liu Y, Wang X, Yu S, Fu D, Chen J, Wang X (2019) Environmental remediation of heavy metal ions by novel-nanomaterials: a review. Environ Pollut 246:608–620. https://doi. org/10.1016/j.envpol.2018.12.076
- Xu S (2012) Electromechanical biosensors for pathogen detection. Microchim Acta 178:245–260. https://doi.org/10.1007/s00604-012-0831-4
- Yáñez-Sedeño P, Campuzano S, Pingarrón JM (2019) Electrochemical (bio)sensors: promising tools for green analytical chemistry. Curr Opin Green Sustain Chem 19:1–7. https://doi. org/10.1016/j.cogsc.2019.01.004
- Yang M, Jeong SW, Chang SJ, Kim KH, Jang M, Kim CH, Bae NH, Sim GS, Kang T, Lee SJ, Choi BG, Lee KG (2016) Flexible and disposable sensing platforms based on newspaper. ACS Appl Mater Inter 8:34978–34984. https://doi.org/10.1021/acsami.6b10298
- Yang Y, Li X, Sun Y, Zhang Z, Xue B, Fan J (2018) Photoluminescent lignin hybridized carbon quantum dots composites for bioimaging applications. Int J Biol Macromol 122:954–961. https://doi.org/10.1016/j.ijbiomac.2018.11.018
- Yusoff N (2019) Chapter 7 graphene–polymer modified electrochemical sensors. In: Pandikumar A, Rameshkumar P (eds) Graphene-based electrochemical sensors for biomolecules. Elsevier, pp 155–186
- Zhang K, Hu R, Fan G, Li G (2017) Graphene oxide/chitosan nanocomposite coated quartz crystal microbalance sensor for detection of amine vapors. Sensors Actuators B Chem 243:721–730. https://doi.org/10.1016/j.snb.2016.12.063
- Zhang N, Wang X, Shi C, Zhang N, Wang X, Qiao R, Zhang Y, Zhang H, Zhong J, Qiao Y, Zhou X, Ma C (2018a) Electrospun Nanofibrous cellulose acetate/Curcumin membranes for fast detection of Pb ions. J Nanosci Nanotechnol 19:670–674. https://doi.org/10.1166/jnn.2019.15893
- Zhang Q, An C, Fan S, Shi S, Zhang R, Zhang J, Li Q, Zhang D, Hu X, Liu J (2018b) Flexible gas sensor based on graphene/ethyl cellulose nanocomposite with ultra-low strain response for volatile organic compounds rapid detection. Nanotechnology 29. https://doi. org/10.1088/1361-6528/aabf2f

- Zhang W, Wang L, Yang Y, Gaskin P, Teng KS (2019) Recent advances on electrochemical sensors for the detection of organic disinfection byproducts in water. ACS Sensors acssensors.9b00272 . https://doi.org/10.1021/acssensors.9b00272
- Zhou X, Ma P, Wang A, Yu C, Qian T, Wu S, Shen J (2015) Dopamine fluorescent sensors based on polypyrrole/graphene quantum dots core/shell hybrids. Biosens Bioelectron 64:404–410. https://doi.org/10.1016/j.bios.2014.09.038
- Zinoubi K, Majdoub H, Barhoumi H, Boufi S, Jaffrezic-Renault N (2017) Determination of trace heavy metal ions by anodic stripping voltammetry using nanofibrillated cellulose modified electrode. J Electroanal Chem 799:70–77. https://doi.org/10.1016/j.jelechem.2017.05.039

Chapter 6 Toxicology and Safety Aspects of Nanosensor on Environment, Food, and Agriculture



Yasmin M. Heikal and Heba M. M. Abdel-Aziz

Contents

6.1	Introd	uction	140	
6.2	Nanosensors Development in Agri/Food Sector.			
	6.2.1	Nanosensors: Definition, Mechanism, Types, and Classification	141	
	6.2.2	Nanosensors for Food and Agriculture Application	142	
6.3	Enviro	onmental Dynamics of Nanosensors, Their Interactions with Target and		
	Non-ta	arget Organisms	146	
	6.3.1	Nanosensor Life-Cycle Assessment	146	
	6.3.2	Nanoparticles Fate and Agro-Environment	146	
	6.3.3	Nanosensors Environmental Toxicology Aspects	148	
6.4	Regulatory Frameworks for Nanomaterials/Nanosensors Linked to Environmental,			
	Food a	and Agricultural Hazard, Exposure, and Risk Evaluation	148	
	6.4.1	Current Guidelines for Assessing the Safety of Nanomaterials/Nanosensors	149	
	6.4.2	Regulatory Aspects and Legislation of Nanomaterials/Nanosensors	151	
6.5	5.5 Conclusion			
Refe	erences.		153	

Abstract The potential of nanotechnology is for innovation in the agricultural and food industries. Different nanosensors types are being created to satisfy the varying agri/food industry demands, comprising food components, smart packaging, and rapid detection systems. It is necessary to understand the assigned mechanisms or causes for nanoscale toxic impacts, their features and properties both externally and within the biological environment. Several nations around the globe have therefore been concerned in testing the suitability of their legislative outlines to address nanotechnologies. Here we review the latest nanosensor developments and address the most difficult assignments and promising possibilities from chosen latest research in the agri-food sector. The main points are (1) knowledge acquisition and

139

Y. M. Heikal (🖂) · H. M. M. Abdel-Aziz

Botany Department, Faculty of Science, Mansoura University, Mansoura, Egypt e-mail: Yasminheikal@mans.edu.eg

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_6

development of nanosensor life cycle techniques, risk assessment, toxicity, bioaccumulation, and beneficial and negative impact assessment. (2) The toxicological fundamentals and the exposure risk connected with the usage of nanosensors in agri/food and the environment are also addressed. (3) Regulation and laws are also essential for the regulation of nanosensor production, processing, implementation, and disposal.

Keywords Nanosensors \cdot Toxicology \cdot Agriculture \cdot Food security \cdot Nanotechnology \cdot Environment security \cdot Safety \cdot Regulation \cdot Legislation \cdot Public perception

6.1 Introduction

In view of the growing worldwide population, contemporary technologies such as nanotechnology in the food and agricultural industries must be used to increase agricultural production (Batsmanova et al. 2013). In most developing countries, agriculture is the cornerstone and directly and indirectly offers food for humans. As one of the leading drivers of the economy, agri/food manufacturing is of essential significance. It can also provide paths for value-added plants. Agricultural practices are often in the public eye because the limitations of climate change, power, and resources and the quickly increasing worldwide population place unprecedented pressure on food and water resources (Ghasemzadeh 2012). With restricted land and water resources available, agricultural development can only be accomplished via the use of more efficient fertilizers and pesticides, but also on hygiene by enhancing productivity through excellent agronomy and promoting it with the efficient use of contemporary technology.

Compared to its use in pharmaceuticals and drug delivery, nanotechnology applications in agriculture and food are comparatively recent (Garcia et al. 2010). Nanotechnology relates to the comprehension and control of nanoscale matter where a distinctive phenomenon allows for new applications (Committee on Technology 2014). However, limiting the size of nanotechnology to a range of 1–100 nm excludes countless products and devices, particularly in the agricultural and pharmaceutical areas, and some specialists are cautious against a strict definition based on a sub-100 nm size. Nanotechnology can revolutionize farming and related areas tremendously, comprising fisheries and aquaculture. Nanofarming presently focuses on targeted farming, using nanosized particles with distinctive characteristics to increase crop and livestock productivity (Scott and Chen 2014). Nanotechnology has the ability to safeguard crops, monitor the growth of plant, detect illnesses of plants and animals, boost worldwide food manufacture, improve food quality, and decrease waste for "sustainable intensification" (Gruère et al. 2011; Sonkaria et al. 2012).

Nanosensors are developing as a successful instrument for farming and food production application. Compared to conventional chemical and biological methods, they give important improvements in selectivity, velocity, and sensitivity. Microbes, pollutants, contaminants, and food freshness can be detected by nanosensors (Joyner and Kumar 2015).

Sustainable agro-production relies primarily on several of the securities listed such as land, food, power, and water safety. It should be noted that requests for nanotechnology can be used to attain these prior bonds (Siddiqui et al. 2015; Dubey and Mailapalli 2016; Rai and da Silva 2017). Food and nutrition safety becomes a basic human right and means that all individuals must have access to sufficient, accessible, and safe foods at all socio-economic and geographical levels to satisfy people's nutritional needs and food choices for active and healthy living (FAO 2009).

Research connected with the nanoparticles potential risk to living organisms has attracted interest from academia, business, and government regulatory agencies globally due to the growing amount of nanomaterials and the broad variety of applications. This chapter will therefore concentrate on various nanosensor applications in sustainable agro-food manufacturing. The risk and safety of these nanosensors on the environment, food, and agriculture will also be outlined. The state-of-the-art various legislative policies for nanomaterials in agri/food will be discussed as well as legislation and guidelines for safety evaluation.

6.2 Nanosensors Development in Agri/Food Sector

6.2.1 Nanosensors: Definition, Mechanism, Types, and Classification

The effect of nanosensors on the industrial, food, and environmental industries is anticipated. Portable nanodevices can detect illnesses, pathogens, insects, contaminants, and chemicals quickly and can lead to quicker treatment (Farrell et al. 2012). The entire agri/food industry can be revolutionized by inexpensive sensors, cloud computing, and smart software together. Precise agriculture becomes a possibility thanks to computers, worldwide satellite positions, and remote sensing equipment. The key features of a precise agricultural vision are the monitoring and assessment of environmental factors, the taking of appropriate and prompt decisions, targeted actions based on information gathered to boost crop production using ideal resources for pesticides, fertilizers, and herbicides (Rai and Ingle 2012).

Nanosensors have the structure as ordinary sensors, but their processing is on the nanoscale. The nanosensor therefore can be described as an extremely small device capable of connecting to anything that needs to be detected and transmitting a signal. These minisensors can recognize and react by transmitting this response to a signal or output that can be used by humans in the physicochemical (sensors) and biological (biosensors). Compared with conventional sensors and their limitations, nanosensors have a range of beneficial characteristics: high selectivity and sensitivity, proximity to real time detection, low price, and portability (Lu and Bowles 2013).

Specific nanosensor design methods include molecular self-assembly, lithography from the top down and bottom up-assembly approaches. Current devices for nanosensors can be split into (Liu 2003): nanostructured materials—for example, porous silicon, sensors based on nanoparticles, nanoprobes, nanosensors, nanosystems: nanoelectromechanical systems (NEMS), and cantilevers. Agriculture uses mainly two kinds of nanosensors: (i) electrical-nanosensors and (ii) bio-nanosensors. Biological organisms have significance in defining the environmental condition; the combination of biology and nanoparticles into sensors is capable of improving awareness and could reduce reaction time to feel a prospective problem (Scott and Chen 2014). Nanobiosensor is a modified version of a biosensor that can be defined as a compact analytical unit that includes a sensitized part associated with the transducer (Turner 2000).

6.2.2 Nanosensors for Food and Agriculture Application

Numerous nanosensors have been developed to detect threats rapidly in the case of suspected food poisoning for various applications in the agricultural and food industry or to be embedded in the packaging as nanotracers in order to demonstrate the history of the foodstuff and whether its consistency is always appropriate. The use of nanosensors in food packaging, for example, in microorganisms detection and the color change when a threshold is reached, as well as in Internet process control nanosensors, helps track storage conditions so as to prevent toxicity to food (Augustin and Sanguansri 2009). Scientists use gold nanoparticles to coat them with molecules that bind to compounds like pesticides. Farmers could spray these nanoparticles in their areas to detect a chemical such as a pesticide (Rathbun 2013). It is possible to summarize the list of prospective applications of nanosensors in the agri/food industry as shown in Figs. 6.1a and 6.1b.

Field-divided nanosensors can also detect the presence of corn and other plant pathogens and concentrations of soil nutrients (Jones 2006; Brock et al. 2011). Gas sensors and nanosmart dust (using small wireless detectors and transponders) (Mousavi and Rezaei 2011) can rapidly assess levels of environmental pollution. Nanobarcodes and nano-related processing could also be used to monitor agricultural production quality (Li et al. 2005). Hormone regulation based on nanotechnology such as auxin allows scientists to understand how plant roots, especially marginal soils, adapt to their environment (McLamore et al. 2010).

Nanobiosensors can be used efficiently at very minute concentrations to detect a broad range of fertilizers, herbicides, pesticides, insecticides, pathogens, moisture, soil pH, poisonous pollutants, and industrial pollutants (Rai et al. 2012; Sekhon 2014). Nanobiosensors are used in sectors other than food and agriculture. Nevertheless, after considering their significance, several sensors have been recently developed (Sekhon 2014; Jyoti and Tomar 2016). This technology can offer better fertilization management for farmers in the agro-production industry, decrease of inputs and better management of time and climate, as well as control of crop health,



Fig. 6.1a Schematic diagram outlining the nanosensors types: Metal-based nanosensors, singlewalled carbon nano tubes and DNA, carbon black and polyaniline, array biosensors, electronic noses, nano-test strips, and nanocantilevers, nano-smart dust, nanobarcodes, nanobiosensors, biomimetic sensors (protein & biomimetic membranes) and smart biosensors and their applications in different agri/food sector



Fig. 6.1b Schematic diagram outlining the nanosensors types: surface plasmon-coupled emission biosensors (with Au); cerium oxide immunosensors and chitosan-based nanocomposites; carbon nanotubes and silicon nanowire transistors; iSTrip of time-temperature indicator/integrator; abuse indicators; partial temperature history indicator; full-temperature history indicator; reflective interferometry and their applications in different agri/food sector

food quality, and security, thus improving agriculture productivity in addition to helping in the effective use of natural resources such as water, chemicals, and nutrients (Farrell et al. 2012; Ansari et al. 2016).

Some researchers use nanosensors to track the biological and ecological health effect of agricultural pollutants (Ansari et al. 2016). These nanosensors can also be used for gases such as nitrogen oxides, sulfur dioxide, ammonia, sulfur hydrogen, and unstable organics and metal oxide nanowire, either metal nanoparticles (palladium, platinum and or gold) or metal oxide nanoparticles and nanotubes. Without altering confirmation and the linkage of the enzyme molecules outside the active enzyme centers, the biosensor development showed excellent compatibility between membranes and proteins (Bäcker et al. 2017). Therefore, the design of sensors/biosensors depends on different interactions resulting in nuclear power spectroscopy (Shuai et al. 2017).

The recent advances have led in near-market nanosensors: nanosensors and nanoscale coatings to substitute thicker, more corrosion-preventing wasteful polymer coatings, nanoscale biopolymers to enhance heavy metal decontamination and recycling, aquatic toxin nanosensors, nanosensors capable of offering quality assurance through the food processing chain by tracking microbes, toxins and contaminants, including the use of data capture for automatic monitoring (Prasad et al. 2014).

Wireless nanosensors provide accurate, time-based information, including food and environmental pesticide detection, quality control, and environmental status. Salicylic acid is a plant-based phenolic phytohormone that promotes plant growth, photosynthesis, and transpiration. It is an essential plant component and it is important to sensitively detect the amount of salicylic acid. Wang et al. (2010) use copper nanoparticles with gold electric nanosensors. Copper nanoparticles detect salicylic acid electro-catalytic oxidation and electrochemical salicylic acid behavior. Nano gold nanoparticle electrode detects properly levels of salicylic acid in oilseed rape infected with the fungal pathogen *Sclerotinia sclerotiorum* (Wang et al. 2010). Electrochemical sensor for the detection of triazophos insecticide with altered carbon nanotube electrode with a gold nanoparticle deposition was used in post-harvest vegetables (Li et al. 2012).

It is essential to highlight multiple micro-nano biosystems that have been established as European Commission projects (2015) and used to implement intelligent agri-food technologies. The development of technology, in particular the Internet of Things (IoT), as more devices connected through the Internet to customers and other devices grows, contributes significantly to the realization of smart agriculture across sustainability, efficiency, quantity and cost-efficacy of agricultural production. Nanotechnology's innovative implementation in IoT generates a fresh paradigm, called the Nano Things Internet (IoNT).

6.3 Environmental Dynamics of Nanosensors, Their Interactions with Target and Non-target Organisms

The need for knowledge and detection of the presence of nanoparticles in the setting, measurement of emissions of nanoparticles, toxicity, life cycle, and effect on the environment and human health are essential to attaining all the advantages that nanotechnologies have to give. The environmental toxicity of nanoparticles relies on their size, type, and load. In addition, nanoparticles impact on the atmosphere also relies on environmental variables (moisture, temperature, wind flow rate, and light nature). However, nanomaterial characteristics, tiny size, and big surface make it simple to disperse and bond with human tissues in the setting (Rai and Ingle 2012).

6.3.1 Nanosensor Life-Cycle Assessment

The requirement that they be biocompatible with the organism or biological tissues and liquids being tested is implicit in any in vitro or in vivo use of nano-bio sensors. They must be nontoxic for in-vivo processes and the materials must not produce a biological reaction that interferes with the experiment. Sensors intended for noninvasive or in vitro use must be consistent with the investigated biological fluids and any reagents, particles, or elements used for measurement purposes. Nanoparticlesmediated sensors such as bio-imaging aids, drug delivery aids, or colorimetric in vitro sensors may have extra problems such as enhanced rates of dissolution, aggregation, and surface adsorption of biological molecules that may reduce their efficacy. The Biocompatibility Assessment Team is structured to provide an early assessment of suggested sensors recognizing and resolving nano-bio interactions that may influence center systems development and marketing. Expertise is also available to assure safety and effectiveness of sensor technology in testing more mature prototypes (CNBS 2016).

6.3.2 Nanoparticles Fate and Agro-Environment

One of the most important environmental issues may be the environmental fate and transportation of nanoparticles. Nanoparticles in various environmental compartments such as soil, air, water, crops, sediments, and animals are parts of this destination and transportation cycle. It is the place to produce our agricultural products for our milk, feed, fibers, and fuel. Thus, nanoparticles movement and translocation in terrestrial environments are a major problem and should be tackled. In addition, this conduct of nanoparticles in agro-ecosystems involves both the beneficial side and the adverse side as shown in Fig. 6.2 (Omara et al. 2019).



Fig. 6.2 Schematic diagram outlining the possible environmental fate/behavior of nanoparticles (NPs) includes the negative impact as well as the release, transport, and toxicity of nanoparticles in the soil environments, plants, microbes, and human health

Several studies (e.g., Miseljic and Olsen 2014; Soni et al. 2015; Hegde et al. 2016; Tolaymat et al. 2017) have been released on the discharge, transport, and environmental toxicity of nanoparticles. It therefore continues an open question to distribute and transport nanoparticles in distinct environment. Nanoparticles may enter or transport crops and/or other microorganisms from various settings through active or passive uptake into the food chain. Many variables affect this translocation of nanoparticles such as air, soil, water, and sediment and cause severe human and animal changes (Soni et al. 2015; Kwak and An 2016).

One of the most significant agro-environmental researches can be considered by the interaction between nanoparticles under the soil–plant scheme. Agroenvironmental studies include relationships between soil–plant nanoparticles (Soni et al. 2015; Zhang et al. 2017) and soil microbial society impacts of nanoparticles (Samarajeewa et al. 2017). Nanoparticles also have the possibility to carry toxic substances (e.g., heavy metals and lipophilic pollutants) due to their large surface area and highly reactive nature (Baun et al. 2008; Gil-Díaz et al. 2017). Additional types of nanoparticles to pass through the cell walls (Zhang et al. 2017). Thus, nanoparticles of lower sizes can migrate and achieve the plasma membrane than the diameter of the pore. There is also some proof that nanoparticles can enter cells via ion canals and carriers and interfere with normal metabolism, possibly through the manufacture of reactive oxygen species (ROS) (Tolaymat et al. 2017).

6.3.3 Nanosensors Environmental Toxicology Aspects

Nanomaterials only get into the atmosphere and human beings through agricultural production (Rico 2015). Nanomaterials communicate heavily with soils. Improved pesticides and plant products by nanomaterials can spread into soil, water, and atmosphere, bind more heavily to pollutants, and bring them through soil and water. Nanofertilizer and pesticide exposure may lead to risks to health (Sastry 2012). There is also a high risk of nanoparticles migration integrated into human food (Berekaa 2015). Consumers direct exposure to nanomaterials is a major problem for human health. Nanoparticles may enter the human body through the skin, respiratory system, or digestive system. However, the exposure is limited as long as the nanoparticles remain bound. Teow et al. (2011) provided detailed information on the health effect and safety of nanomaterial application.

Modern technologies have shown the potential for various characters of toxickinetics and toxic-dynamics of nanomaterials that are more reactive and capable of crossing membrane barriers. Some nanomaterials interact by causing oxidative stress to proteins and enzymes by producing ROS that cause mitochondrial destruction and apoptosis. In recent years, a variety of biological and toxicological interactions in nanomaterials have been documented in vitro and in vivo by experimental structures. A broad variety of endpoints of biochemistry and toxicology have been recorded within each scheme. Most of the inflammatory markers have been targeted, because current health data concerning the environmental effects of nanomaterials have recognized a key role in oxidative stress and inflammation as toxicological action modes for nanoparticles. Consequently, over the last few years, there have been significant advances of the biological and toxicological effects of nanomaterials. Much of this is related to what physical and toxicological characterization data are required for risk assessment and how to get it. In restricted applications of nanomaterials to measure the acute toxicity of these chemical agents, no serious adverse effects were found. Accessible to nanomaterials, such as long-term inhalation research, reproductive or developmental studies, still exist rudimentary toxicological information sets. The fact that nanoparticles can be kept in cells for long periods of time when absorbed into the systemic circulation makes it important to carry out ongoing studies for hazard and nanomaterial risk assessment (Omanović-Mikličanina and Maksimović 2016).

6.4 Regulatory Frameworks for Nanomaterials/Nanosensors Linked to Environmental, Food and Agricultural Hazard, Exposure, and Risk Evaluation

The government is concerned about the toxicity and environmental impact of nanosensors in agriculture and in the food industry, despite the trembling benefits. Longterm negative effects knowledge is very minimal on soil, plant life, and finally on people (Dubey and Mailapalli 2016). The primary elements of nanotechnology are enhanced quality and dietary value in terms of worldwide food and livestock production. Despite prospective advantages in the agri/food industry (livestock feed, food production, packaging and nano-based smart systems), little is known about safety elements of the implementation of nanotechnology in food production and the integration of nanoparticles into food. Furthermore, nanotechnology knowledge is still lacking for customers (Sekhon 2014).

The use of organic nanomaterials in the agriculture and the consumable sector has raised concerns about latent risks associated with the extensive use of nanoparticles on human health. Biotech tools are used for nanoscalable goods such as carbohydrates, fats, and proteins. The main concerns are inadequate knowledge of nanobiosensor factors such as toxicity, bioaccumulation, and exposure risk. The timely behavior of the particles in biological environments and the complexity of locally measured characteristics, such as size, form, and surface chemistry make it difficult to conduct reproducible and precise nanostructure biocompatibility tests. Due to the complexity, risk evaluation should include an early evaluation of anticipated sensors in order for nanobiological interactions to be recognized and fixed that may influence development and marketing center systems. Expertises should be made available to ensure the safety and efficiency of sensor technology during testing of more advanced prototypes. In order to build nanosensors in biological conditions, it is necessary to properly describe and test the nanoparticles and nanoscale materials and assess their toxicity (CNBS 2016). The potential threat to biological organizations has been identified by nanoparticles of significant interest from researchers, business, government ethics committees, and NGOs worldwide.

6.4.1 Current Guidelines for Assessing the Safety of Nanomaterials/Nanosensors

Risk assessment processes are not specific to agri-food nanomaterials, resulting in uncertainty about the nature and magnitude of future risks in most situations. Nanomaterials currently used for food use include the use of nanoparticles and nanomaterials as food ingredients/additives placed directly in food or as food packaging portions (Cushen et al. 2012). During the stage of product development, public views on new technologies in the food business need to be taken into account in order to avoid some of the pit breakdowns encountered by the genetically modified food industry (López-Vázquez et al. 2012). The release of nanoparticles may adversely affect edible crops (Suppan 2013).

The established risk assessment and testing methods are widely seen in nanomaterial (NMs) components but there is a need for further creation of standardized and validated approaches to certain components, including sample preparation, characterizing, dosimetry, exposure, and modeling data (SCENIHR 2007; OECD 2013). The EU Scientific Committee on Emerging and Newly Identified Health Risks (SCENIHR 2009) proposed a case-by-case technique for risk assessment of NMs. Official bodies have released risk assessment guidelines focusing on the specific features of NMs. In 2011, European Food Safety Authority (EFSA) published a "Risk Assessment Guideline for the Application of Nanoscience and Nanotechnology in the Food and Feed Chain" (EFSA Scientific Committee 2011) that offers a practical approach to assessment of potential threats for nanotechnological and nanoscience applications in food additives, enzymes, flavorings, food contact materials, new food ingredients, feed additives, and pesticides. This provides guidelines on data generation and testing techniques by candidates for the identification and characterization of threats of nanomaterials. In addition, six exposure scenarios and associated testing methods are presented to define and characterize risks resulting from NM's characteristics. These six scenarios are dependent on possible NM transformations before and after food/feed ingestion. The sort of information required to carry out a risk assessment is indicated for each situation.

The guidance provides for reduced information requirements when data indicating no migration from food contact materials or when complete degradation/dissolution is proven without viable NM absorption as such is not verified by NM exposure. The guideline states: the NMs used in food or feed, which are converted into non-nanoforms in a food/feed matrix, may be used as guidance as nonnanoforms before ingestion. For NMs fully transformed in the gastrointestinal tract into non-nanoforms, consideration should be given to local impacts and possible absorption prior to conversion. Existing data should otherwise be used or guidelines should be followed for non-nanoforms. A comparison should show whether, compared to the non-nanoform, the nanoform has increased, less, or comparable risk. In a systematic, staged approach based on a comparison with the non-nano equivalent for food safety evaluations of NMs, an expert group created by the International Life Science Institute (ILSI) (Cockburn et al. 2012) has established a comprehensive approach. The distinction is based on the velocity of dissolution in water or under gastric conditions of the bulk and nano form of the same substance. Although released by EU bodies, these guidelines, like any other guidelines, are not part of EU legislation and therefore not legally binding. EFSA has also created a Food and Feed Nanotechnology Risk Assessment Network (EFSA Nano Network) with general objectives to promote the harmonization of evaluation procedures and methodologies, to improve information and data exchange between EFSA and EU Member States, and to attain synergies in risk assessment operations. In 2012, the EFSA Nano Network applied for an "Inventory of Agricultural, Food and Food Nanotechnology Applications" to obtain more up-to-date data on state-of-the-art nanotechnology apps generated by Rikilt and JRC (2014).

6.4.2 Regulatory Aspects and Legislation of Nanomaterials/ Nanosensors

The risk of negative and unintentional effects of nanotechnology and the public opinion about new agro-production techniques during product development stages is crucially important (Zuverza-Mena et al. 2017). The small and large size of nanoparticles allow for fast dispersal without structural obstacles and possible toxicity are causing the risk (Rico et al. 2011). Millions of small-scale farmers in the agro-producing industry can pose a significant health threat by using nanofertilizers and nanopesticides that enable simple soil, water, and atmospheric dispersal (Tripathi et al. 2017). Nanoparticles are permitted into the food chain via nutrients/ pesticide systems or food processing (Mattsson and Simkó 2016). Therefore, before NMs usage as agricultural goods, there should be studies of life cycles, particulate plant ingestion, biodistribution, and entry into the food chains (Rico et al. 2011).

In addition, nanobiosensor marketing is also associated with several risk variables such as: (a) original applications "could act as substitutes for agricultural commodities" that could be "disastrous" on the developing nations economy (Gruère et al. 2011); (b) Secondly, variable import regulations in separate countries could hinder the development of nanoproducts (Gruère and Sengupta 2009; Gruère et al. 2011); (c) Finally, the use of nanotechnology may be "negative economic effects on the poor by increasing productivity in developed countries," which could result in lower commodity prices in developing nations (Gruère et al. 2011). Besides all the related hazards, marketing of any product is dependent on public acceptance of the same, in addition to all the associated risks. Therefore, risk opinions are crucial for future worldwide nanoproduct availability (Gruère et al. 2011). Customers are hitherto more unwilling to use nanofood than other nanotechnology uses (Lyndhurst 2010; Gruère et al. 2011) apparent "benefits and health risks affect acceptance," "meaning consumers do not perceive all products with the same risk levels" (Siegrist et al. 2009; Gruère et al. 2011). Strategies for risk communication should be articulated "attention to the messenger and the target of the message" (Sylvester et al. 2009; Gruère et al. 2011). Some "external factors" could also put an impact in "shaping future acceptance in the key markets" that will have a major impact on the global future of nanotechnologies (Gruère et al. 2011). Nongovernmental organizations supporting a "ban on nanotechnology use in food and agriculture" can be important, approaching condensed international marketing (Lyons 2010; Gruère et al. 2011). This might result in "technology divide" (Court et al. 2004; Gruère et al. 2011).

In some developed countries, the probable answer to this problem is appropriate nanoproduct labeling that could lead to legislation and technology consumption in developing countries. The Science and Technology Committee of the UK House of Lords has recommended including a mandatory "pre-commercialization assessment using the methods supported by a research investment effort in risk assessment and detection methods" (Gruère et al. 2011). "The European Food Safety Agency supports the use of conventional risk assessment while addressing the limited knowledge on exposure to nanofood applications" (Harrington 2010; Gruère et al. 2011). Countries should take into account the present institutional capacity while laying down legislation "Similarities between biotech regulatory systems and nanotech regulatory systems" should be taken as a benefit (Niosi and Reid, "Biotechnology and Nanotechnology) (Gruère et al. 2011)". Bearing in mind the lessons learned "from the challenges observed in biosafety issues of biotech regulatory system," the "need of public education, transparency and predictability" has increased (Falck-Zepeda et al. 2009). "The lack of risk assessment capacity" and permeable boundaries should also promote the formation of local organizations by nations (Gruère et al. 2011).

The risk assessment of produced nanomaterials should be evaluated for regulatory and risk management reasons (Mattsson and Simkó 2016). This risk assessment of nanomaterials requires an understanding of exposure time, potential hazards, dosage, and nanomaterial type. This risk assessment of nanomaterials requires an understanding of exposure time, potential hazards, dosage, type and their characteristic changes over time (Priester et al. 2017). Despite accepted advancement in creating nanomaterial risk evaluation (Mattsson and Simkó 2016), there are still open issues about the destiny and conduct of nanomaterials in distinct settings. According to the European Parliament, some agro-producing regulations are as follows. (Lee and Stokes 2016): Legislation on food additives (Regulation: EC no. 1333 /2008, Art 12); Legislation on restrictions on hazardous substances in electrical and electronic equipment (Directive 2011 /65/EU, Recital 16); Legislation on waste electrical and electronic equipment (Directive 2012/19/EU, Art 8 (4); Legislation on food information for consumers (Regulation: EU no. 1169 /2011, e.g., Art 18); Legislation on cosmetic products (Regulation: EC no. 1223 /2009, e.g., Art 13 (1); Legislation on biocidal products (Regulation: EU no. 528/2012, e.g., Art 19 (1) and 7. Legislation on food for infants (Regulation: EU no. 609/2013, Art 9 (2).

Therefore, in the near future, a science-based risk assessment will be able to be carried out for the use of nanoparticles in all manufacturing and government sectors, including farming, medical care, transportation, energy, and machinery.

6.5 Conclusion

It is worth mentioning that the nanotechnology can affect or penetrate nearly all agricultural practices. It is important to explore new opportunities for incorporating nanotechnologies into nanosensors/nanobiosensors, aware of any future risks to the environment or human health. As a result, the use of agro-production has led in many hazards and therefore excellent difficulties have been developed to seek safety strategies for sustainable agro-production. We conclude that nanotechnologies in agriculture can be transformed by concentrated research and development to the goals of sustainable agriculture by government and academia. We discovered that there is still a need for efforts to improve public knowledge and appreciation of the

novel nano-enabled food and agricultural products. The intelligent use of nanosensors in a quality and sustainable environment can contribute to achieving food security. A move towards green nanotechnology and green IoT will lead in their comprehensive applications to a whole new globe of safe nanoproducts with low or no health and environmental dangers.

References

- Ansari MIH, Hassan S, Qurashi A, Khanday FA (2016) Microfluidic-integrated DNA nanobiosensors. Biosens Bioelectron 85:247–260. https://doi.org/10.1016/j.bios.2016.05.009
- Augustin MA, Sanguansri P (2009) Nanostructured materials in the food industry. Adv Food Nutr Res 58:183–213. https://doi.org/10.1016/S1043-4526(09)58005-9
- Bäcker M, Koch C, Eiben S, Geiger F, Eber F, Gliemann H, Poghossian A, Wege C, Schöning MJ (2017) Tobacco mosaic virus as enzyme nanocarrier for electrochemical biosensors. Sensor Actuat B-CHEM 238:716–722. https://doi.org/10.3389/fpls.2015.01137
- Batsmanova LM, Gonchar LM, Taran NY, Okanenko AA (2013) Using a colloidal solution of metal nanoparticles as micronutrient fertiliser for cereals. Proceedings of the international conference on nanomaterials: applications and properties; September 16–21; Crimea, Ukraine. Available from: http://nap.sumdu.edu.ua/index.php/nap/nap2013/paper/view/1097/504. Accessed 18 Apr 2014
- Baun A, Sørensen SN, Rasmussen RF, Hartmann NB, Koch CB (2008) Toxicity and bioaccumulation of xenobiotic organic compounds in the presence of aqueous suspensions of aggregates of nano-C60. Aquat Toxicol 86:379–387. https://doi.org/10.1016/j.aquatox.2007.11.019
- Berekaa M (2015) Nanotechnology in food industry; advances in food processing, packaging and food safety. Int J Curr Microbiol App Sci 4(5):345–357
- Brock DA, Douglas TE, Queller DC, Strassmann JE (2011) Primitive agriculture in a social amoeba. Nature 469(7330):393–396. https://doi.org/10.1038/nature09668
- CNBS (2016). http://cnbs.centers.ufl.edu/research/sensors.asp
- Cockburn A, Bradford R, Buck N, Constable A, Edwards G, Haber B, Hepburn P, Howlett J, Kampers F, Klein C, Radomski M, Stamm H, Wijnhoven S, Wildemann T (2012) Approaches to the safety assessment of engineered nanomaterials (ENM) in food. Food Chem Toxicol 50(0):2224–2242. https://doi.org/10.1016/j.fct.2011.12.029
- Committee on Technology (2014) National nanotechnology initiative: strategic plan. National Science and Technology Council, Washington, DC; 2014. Available from: http://nano.gov/sites/default/files/pub_resource/2014_nni_strategic_plan.pdf. Accessed 18 Apr 2014
- Court E, Daar S, Martin E, Acharya T, Singer P (2004) Will Prince Charles et al. diminish the opportunities of developing countries in nanotechnology? http://nanotechweb.org/cws/article/ indepth/18909
- Cushen M, Kerry J, Morris M, Cruz-Romero M, Cummins E (2012) Nanotechnologies in the food industry – recent developments, risks and regulation. Trends Food Sci Technol 24(1):30–46
- Dubey A, Mailapalli DR (2016) Nanofertilisers, Nanopesticides, nanosensors of Pest and Nanotoxicity in agriculture. In: Lichtfouse E (ed) Sustainable agriculture reviews, sustainable agriculture reviews, vol 19. Springer, Cham, pp 307–330. https://doi. org/10.1007/978-3-319-26777-7_7
- EFSA Scientific Committee (2011) Guidance on the risk Assessment of the application of nanoscience and nanotechnologies in the food and feed chain from. http://www.efsa.europa.eu/en/ efsajournal/doc/2140.pdf
- European Comission (2015) Projects on Smart Agri-Food systems. ec.europa.eu/newsroom/horizon2020/document.cfm?doc_id=18158 (4/12/2016)

- Falck-Zepeda J, Cavialeri A, Zambrano P (2009) Delivering genetically engineered crops to poor farmers: recommendations for improved biosafety regulations in developing countries, IFPRI brief 14. International Food Policy Research Institute, Washington, DC
- FAO (2009) High level expert forum global agriculture towards 2050, how to feed the world-2050, Rome or http://www.fao.org/fileadmin/templates/wsfs/docs/Issues_papers/ HLEF2050_Global_Agriculture.pdf
- Farrell D, Hoover M, Chen H, Friedersdorf L (2012) Overview of resources and support for nanotechnology for sensors and sensors for nanotechnology: improving and protecting health, safety, and the environment. US National Nanotechnology Initiative, Arlington. Available from http://nano.gov/sites/default/files/pub_resource/nsi_nanosensors_resources_for_web.pdf
- Garcia M, Forbe T, Gonzalez E (2010) Potential applications of nanotechnology in the agro-food sector. Food Sci Technol (Campinas) 30(3):573–581
- Ghasemzadeh A (2012) Global issues of food production. Agrotechnol 1(2):1–2. https://doi.org/1 0.4172/2168-9881.1000-102
- Gil-Díaz M, Pinilla P, Alonso J, Lobo MC (2017) Viability of a nano remediation process in single or multi-metal(loid) contaminated soils. J Hazard Mater 321:812–819. https://doi.org/10.1016/j.jhazmat.2016.09.071
- Gruère G, Sengupta D (2009) GM-free private standards and their effects on biosafety decisionmaking in developing countries. Food Policy 34(5):399–406. https://doi.org/10.1016/j. foodpol.2009.04.002
- Gruère G, Narrod C, Abbott L (2011) Agriculture, food, and water nanotechnologies for the poor: opportunities and constraints. Policy brief 19. International Food Policy Research Institute, Washington, DC. Available from http://www.ifpri.org/sites/default/files/publications/bp019.pdf
- Harrington R (2010) Nano risk assessment a work in progress. http://www.foodnavigator.com/ Science-Nutrition/Nano-risk-assessment-a-work-in-progress
- Hegde K, Brar SK, Verma M, Surampalli RY (2016) Current understandings of toxicity, risks and regulations of engineered nanoparticles with respect to environmental microorganisms. Nanotechnol Environ Eng 1:5. https://doi.org/10.1007/s41204-016-0005-4
- Jones PBC (2006) A nanotech revolution in agriculture and the food industry. Information Systems for Biotechnology, Blacksburg. Available from http://www.isb.vt.edu/articles/jun0605.htm
- Joyner JJ, Kumar DV (2015) Nanosensors and their applications in food analysis: a review. Int J Sci Technol 3:80–90
- Jyoti A, Tomar RS (2016) Detection of pathogenic bacteria using nanobiosensors. Environ Chem Lett: 1–6. https://doi.org/10.1007/s10311-016-0594-y
- Kwak J, An Y-J (2016) Trophic transfer of silver nanoparticles from earthworms disrupts the locomotion of springtails (Collembola). J Hazard Mater 315:110–116. https://doi.org/10.1016/j. jhazmat.2016.05.005
- Lee RG, Stokes E (2016) Material uncertainty: nanomaterials, regulation and symbolic legislation. In: van Klink B et al (eds) Symbolic legislation theory and developments in biolaw, Legisprudence library, vol 4. Springer, Cham, pp 237–252. https://doi.org/10.1007/978-3-319-33365-6_14
- Li Y, Cu YT, Luo D (2005) Multiplexed detection of pathogen DNA with DNA-based fluorescence nanobarcodes. Nat Biotechnol 23(7):885–889
- Li H, Xie C, Li S, Xu K (2012) Electropolymerized molecular imprinting on gold nanoparticle carbon nanotube modified electrode for electrochemical detection of triazophos. Colloids and Surf B: Biointerfaces 89:175–181. https://doi.org/10.1016/j.colsurfb.2011.09.010
- Liu Y (2003) Nanosensors. http://www.slideserve.com/kim-johnston/nanosensors
- López-Vázquez E, Brunner TA, Siegrist M (2012) Perceived risks and benefits of nanotechnology applied to the food and packaging sector in México. Brit Food J 114(2):197–205. https://doi.org/10.1108/00070701211202386
- Lu J, Bowles M (2013) How will nanotechnology affect agricultural supply chains? Int Food Agribusiness Manag Rev 16(2):21–42

- Lyndhurst B (2010) An evidence review of public attitudes to emerging food technologies, social science research unit. Food Stand Agency 27(2):82–89. http://www.foodbase.org.uk/admin-tools/reportdocuments/3701647_Emerging_Food_Technologies_Final_Report_with_logo.pdf
- Lyons K (2010) Nanotechnology: transforming food and the environment. Food First Backgrounder 16(1):1–4
- Mattsson MO, Simkó M (2016) The changing face of nanomaterials: risk assessment challenges along the value chain. Regul Toxicol Pharmacol 84:105–115. https://doi.org/10.1016/j. yrtph.2016.12.008
- McLamore ES, Diggs A, Calvo Marzal P, Shi J, Blakeslee JJ, Peer WA, Porterfield DM (2010) Non-invasive quantification of endogenous root auxin transport using an integrated flux microsensor technique. Plant J 63(6):1004–1016. https://doi.org/10.1111/j.1365-313X.2010.04300.x
- Miseljic M, Olsen SI (2014) Life-cycle assessment of engineered nanomaterials: a literature review of assessment status. J Nanopart Res 16:24–27. https://doi.org/10.1007/s11051-014-2427-x
- Mousavi SR, Rezaei M (2011) Nanotechnology in agriculture and food production. J Appl Environ Biol Sci 1(10):414–419. https://doi.org/10.2136/vzj2011.0105br
- OECD (2013) Current developments/activities on the safety of manufactured nanomaterials; series on the safety of manufactured nanomaterials no. 37. http://search.oecd.org/officialdocuments/display2&doclanguage=en
- Omanović-Mikličanina E, Maksimović M (2016) Nanosensors applications in agriculture and food industry. Bull Chem Technol Bosnia Herzegovina 47:59–70
- Omara AED, Elsakhawy T, Alshaal T, El-Ramady H, Kovács Z, Fári M (2019) Nanoparticles: a novel approach for sustainable agro-productivity. Environ Biodiv Soil Sec 3:29–62
- Prasad R, Kumar V, Prasad KS (2014) Nanotechnology in sustainable agriculture: present concerns and future aspects. Afr J Biotechnol 13(6):705–713
- Priester JH, Moritz SC, Espinosa K, Ge Y, Wang Y, Nisbet RM, Schimel JP, Goggi AS, Gardea-Torresdey JL, Holden PA (2017) Damage assessment for soybean cultivated in soil with either CeO₂ or ZnO manufactured nanomaterials. Sci Total Environ 579:1756–1768. https://doi. org/10.1016/j.scitotenv.2016.11.149
- Rai M, da Silva SS (2017) Nanotechnology for bioenergy and biofuel production. Green chemistry and sustainable technology series. Springer, Cham. https://doi.org/10.1007/978-3-319-45459-7
- Rai M, Ingle A (2012) Role of nanotechnology in agriculture with special reference to management of insect pests. Appl Microbiol Biotechnol 94:287–293
- Rai V, Acharya S, Dey N (2012) Implications of nanobiosensors in agriculture. J Biomater Nanobiotchnol 3:315–324
- Rathbun LC (2013) Nanosensors and the food supply. http://www.nanooze.org/nanosensors-andthe-foodsupply-2/, (2/10/2016)
- Rico C (2015) nanomaterial implications for agricultural productivity and food safety. http://www. azonano.com/article.aspx?ArticleID=4076, (20/12/2016)
- Rico CM, Majumdar S, Duarte-Gardea M, Peralta- Videa JR, Gardea-Torresdey JL (2011) Interaction of nanoparticles with edible plants and their possible implications in the food chain. J Agric Food Chem 59(8):3485–3498. https://doi.org/10.1021/jf104517j
- Rikilt and Joint Research Centre (JRC) (2014) Inventory of nanotechnology applications in the agricultural, feed and food sector. Parma, RIKILT. European Commission Joint Res Centre EN-621, 125. http://www.efsa.europa.eu/en/supporting/doc/621e.pdf
- Samarajeewa AD, Velicogna JR, Princz JI, Subasinghe RM, Scroggins RP, Beaudette LA (2017) Effect of silver nano-particles on soil microbial growth, activity and community diversity in a sandy loam soil. Environ Pollut 220(Part A):504–513. https://doi.org/10.1016/j. envpol.2016.09.094
- Sastry RK (2012) Potential economic impacts of Agri-nanotechnology. International Symposium on Assessing the Economic Impact of Nanotechnology, Washington, DC
- SCENIHR (2007) Opinion in the appropriateness of the risk assessment methodology in accordance with the technical guidance documents for new and existing substances for assessing the risks of nanomaterials. Scientific Committee on Emerging and Newly Identified Health

Risks, Brussels from. http://ec.europa.eu/health/archive/ph_risk/committees/04_scenihr/docs/ scenihr_o_010. pdf

- SCENIHR (2009) Risk assessment of products of nanotechnologies. Scientific Committee on Emerging and Newly Identified Health Risks, Brussels from. http://ec.europa.eu/health/ph_risk/committees/04_scenihr/docs/scenihr_o_023.pdf
- Scott N, Chen H (2014) Nanoscale science and engineering for agriculture and food systems. Ind Biotechnol 8(6):340–343. https://doi.org/10.1089/ind.2013.1555
- Sekhon BS (2014) Nanotechnology in Agri-food production: an overview. Nanotechnol Sci Appl 7(2):31–53. https://doi.org/10.2147/NSA.S39406
- Shuai HL, Huang KJ, Zhang WJ, Cao X, Jia MP (2017) Sandwich-type microRNA biosensor based on magnesium oxide nanoflower and graphene oxide–gold nanoparticles hybrids coupling with enzyme signal amplification. Sensor Actuat B-Chem 243:403–411
- Siddiqui MH, Al-Whaibi MH, Mohammad F (2015) Nanotechnology and plant sciences: nanoparticles and their impact on plants. Springer International Publishing, Cham. https://doi. org/10.1007/978-3-319-14502-0
- Siegrist M, Stampfli N, Kastenholz H (2009) Acceptance of nanotechnology foods: a conjoint study examining consumers' willingness to buy. Br Food J 111(7):660–668. https://doi. org/10.1108/00070700910972350
- Soni D, Naoghare PK, Saravanadevi S, Pandey RA (2015) Release, transport and toxicity of engineered nanoparticles. In: Whitacre DM (ed) Reviews of environmental contamination and toxicology 234. Springer, Cham. https://doi.org/10.1007/978-3-319-10638-0_1
- Sonkaria S, Ahn SH, Khare V (2012) Nanotechnology and its impact on food and nutrition: a review. Recent Pat Food Nutr Agric 4(1):8–18. https://doi.org/10.2174/1876142911204010008
- Suppan S (2013) Nanomaterials in soil: our future food chain? Institute for Agriculture and Trade Policy, Minneapolis, Available from http://www.iatp.org/files/2013_04_23_Nanotech_SS.pdf
- Sylvester D, Abbott K, Marchant G (2009) Not again! Public perception, regulation, and nanotechnology. Regul Gov 3(2):165–185. https://doi.org/10.1111/j.1748-5991.2009.01049.x
- Teow Y, Asharani PV, Hand MP, Valiyaveettil S (2011) Health impact and safety of engineered nanomaterials. Chem Commun 47:7025–7038
- Tolaymat T, Genaidy A, Abdelraheem W, Dionysiou D, Andersen C (2017) The effects of metallic engineered nanoparticles upon plant systems: An analytic examination of scientific evidence. Sci Total Environ 579:93–106. https://doi.org/10.1016/j.scitotenv.2016.10.229
- Tripathi DK, Singh S, Singh S, Pandey R, Singh VP, Sharma NC, Prasad SM, Dubey NK, Chauhan DK (2017) An overview on manufactured nanoparticles in plants: uptake, translocation, accumulation and phytotoxicity. Plant Physiol Biochem 110:2–12. https://doi.org/10.1016/j. plaphy.2016.07.030
- Turner AP (2000) Biosensors-sense and sensitivity. Science 290(5495):1315–1317. https://doi. org/10.1126/science.290.5495.1315
- Wang Z, Wei F, Liu SY, Xu Q, Huang JY, Dong XY, Yu JH, Yang Q, Zhao YD, Chen H (2010) Electrocatalytic oxidation of phytohormone salicylic acid at copper nanoparticles-modified gold electrode and its detection in oilseed rape infected with fungal pathogen *Sclerotinia sclerotiorum*. Talanta 80:1277–1281. https://doi.org/10.1016/j.talanta.2009.09.023
- Zhang W, Dan Y, Shi H, Ma X (2017) Elucidating the mechanisms for plant uptake and in-planta speciation of cerium in radish (*Raphanus sativus* L.) treated with cerium oxide nanoparticles. J Environ Chem Eng 5(1):572–577
- Zuverza-Mena N, Martínez-Fernández D, Du W, Hernandez-Viezcas JA, Bonilla-Bird N, López-Moreno ML, Komárek M, Peralta-Videa JR, Gardea-Torresdey JL (2017) Exposure of engineered nanomaterials to plants: insights into the physiological and biochemical responses-a review. Plant Physiol Biochem 110:236–264. https://doi.org/10.1016/j.plaphy.2016.05.037

Chapter 7 Nanosensors for the Detection of Fertilizers and Other Agricultural Applications



Heba M. M. Abdel-Aziz and Yasmin M. Heikal

Contents

7.1	Introd	uction	158		
7.2	Why We Need Nanosensors?				
7.3	Types of Nanosensors and Bionanosensors				
7.4	Applications of Nanosensors in Agriculture.				
7.5	Applications of Bionanosensors in Agriculture		161		
	7.5.1	Detection of Bacteria.	161		
	7.5.2	Detection of Mycotoxins.	163		
	7.5.3	Detection of Pesticides and Herbicides.	163		
	7.5.4	Detection of Veterinary Drugs.	164		
7.6	Conclusion 1				
Refe	rences.		165		

Abstract Recently, nanotechnology has started to revolutionize all agricultural practices. Nanotechnology research has improved all aspects of current agriculture. It can enhance the growth of plants by improving uptake of nutrients through engineered nanofertilizers. Also, it plays a pioneer role to monitor plant and soil conditions through nanosensors and detect pathogens and toxins (herbicides, pesticides, and veterinary drugs) residues in the food we eat through bionanosensors. Here we review nanotechnology applications in the form of nanosensors and bionanosensors and how they are used to monitor plant and soil conditions and detect diseases and pathogens in the crops we eat.

Keywords Nanosensors · Bionanosensor · Pathogens · Bacteria · Mycotoxins · Herbicides · Pesticides · Veterinary drugs · Soil · Plant

H. M. M. Abdel-Aziz (🖂) · Y. M. Heikal

Botany Department, Faculty of Science, Mansoura University, Mansoura, Egypt e-mail: hebammabdelaziz@mans.edu.eg

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_7

7.1 Introduction

The massive increase in population all around the globe leads to the need to produce more food from the same limited farm lands. Previous approaches to improve crop productivity included: the use of biotechnology, genetic engineering, and synthetic chemicals. These approaches caused serious concerns with regard to health and safety of humans. A recent technology has evolved which is nanotechnology (Mura and Dudáš 2013; Chhipa and Joshi 2016).

Nanotechnology deals with synthesis and applications of nanostructures of sizes at the nanometer scale. The word "nano" comes from the Greek word which means "dwarf," which means one billionth of a meter. Nanoparticles are considered to be atomic or molecular aggregates which have one of their dimensions from 1 to 100 nm (Thakkar et al. 2010; Prasad 2014; Prasad et al. 2016; Kaushal and Wani 2017). Nanoparticles and nanomaterials could be classified into four groups: (1) carbon-based materials, that is, carbon nanotubes; (2) metal oxides such as TiO₂, Al₂O₃, and ZnO; (3) polymer nanoparticles to perform some chemical functions such as dendrimers; and (4) composites which are combinations between two types of nanomaterials or nanomaterials with bulk materials (Lin and Xing 2008; Nair et al. 2010; Kaushal and Wani 2017).

Nanotechnology has reached all aspects of everyday life. Recently, nanotechnology research has started to address agricultural problems in an attempt to find pioneer solutions to enhance productivity of crops and decrease pests, infections, and environmental pollution (He et al. 2019). In a recent application of nanotechnology research, along with nanopesticides, nanoherbicides, and nanofertilizers, nanosensors aroused as new devices set to detect bacteria, mycotoxins, pesticides, herbicides, veterinary drugs, and monitor soil and plant conditions (Chen and Yada 2011; He et al. 2019). Such detection allows better control of pests and pathogens and provides information to improve crop productivity by better uptake of nutrients and improve soil properties. The use of nanosensors in conventional agriculture converts it into precision agriculture (Chhipa and Joshi 2016).

There are two main forms of nanosensors used in agronomy: (1) electrical and (2) bionanosensors (Chhipa and Joshi 2016). A chemical sensor can be defined as a device which measures the concentration of a specific material and transfers it into a useful analytical signal. However, a biosensor is defined as a device which contains an element (biologically active) such as an enzyme or antibody used for the quantitative detection of a specific analyte (Sharma and Rogers 1994; Srivastava et al. 2018). The characteristics of the biological molecule construct the reaction of the biosensor with the measured material which then becomes translated into a measurable signal (Gomes et al. 2015).

Carbon nanomaterials were developed to act as a chemical sensor to detect pesticides accumulation in plants (Sharon and Sharon 2008). For the identification of *Xanthomonas axonopdis*, a plant pathogen causing bacterial spot diseases in Solanaceae plants, a bionanosensor made of silicon nano-probes attached to goat anti-rabbits antibody was used (Yao et al. 2009).



Fig. 7.1 Difference between nanosensor and bionanosensor

In the current work, we reviewed the use of nanotechnology applications: nanosensors and bionanaosensors in agriculture and their effects on crop productivity, diseases detection, and environmental assessment.

7.2 Why We Need Nanosensors?

Recent advances in agricultural practices led to massive use of herbicides, pesticides, veterinary drugs, and synthetic chemical fertilizers. This led to concerns on human health due to the accumulation of these toxic materials in plants and the food we eat. Thus, the need to have devices with precise ability to detect such chemicals and toxins was a priority to research labs all over the globe. Hence, the introduction of sensors at the nano scale was developed (Omanović-Mikličanina and Maksimocic 2016). A nanosensor is a very tiny device that can bind with what needs to be measured and send back a signal. Nanosensors were classified in two main categories: (1) sensors of chemical origin attached to an electrical transducer which give physicochemical signal are defined as electrical nanosensors and (2) sensors which contain a moiety of biological source along with nanoparticles attached to transducer and respond to a biological signal are bionanosensors (Srivastava et al. 2018; He et al. 2019) (Fig. 7.1).

7.3 Types of Nanosensors and Bionanosensors

Nanosensors have their arrangement like normal sensors but their production is at the nanoscale.



Fig. 7.2 Schematic diagram of a typical bionanosensor

After virtually changing the detection section for upgraded yield with nanostructures, the word nano is included in nanosensors and bionanosensors. The transducer is a sensor composed of nanomaterials such as metallic nanoparticles of gold and iron oxide, quantum dots, graphene oxide, and carbon nanotubes. Increased mechanical, thermal, optical, electrical, and highly specialized surface of the nanomaterial and flexible morphology improve the transfer capabilities in a certain degree (Srivastava et al. 2018). The nanosensors and bionanosensors can be categorized by transduction mechanism for output generation, as nanomaterial-dependent electrochemical nanosensors, and optical nanosensors which are connected to metallic nanoparticle optical properties and quantum dots. Mass nanosensors depend on mechanical and electrical properties of crystals (Srivastava et al. 2018) (Fig. 7.2).

7.4 Applications of Nanosensors in Agriculture

In order to monitor cultivation diseases, development, nutrient effectiveness, and environmental circumstances in the field, nanosensors have been created. In agricultural systems, nanosensors can trace quantities of chemicals such as pesticides, herbicides, and pathogens. This monitoring system provides solutions to prevent crop losses and improve crop productivity together with the aid of other nanostructures (nanofertilizers, nanopesticides, and nanoherbicides). A recent research showed that copper-doped montmorillonite can be used in water (both fresh and salty water) to monitor fungicides with a small detectable threshold of approximately 1 mM (Abbacia et al. 2014; He et al. 2019). Another research has shown that pathogens in wastewater can be detected using nanomaterials such as graphene (Wibowo et al. 2018) and purify it for use as drinking water (Deng et al. 2016), indicating potential application in aquaculture. Many other nanomaterials such as copper nanoparticles (Geszke-Moritz et al. 2012), carbon nanotubes (Esser et al. 2012), gold nanoparticles (Lin et al. 2011), and silver nanoparticles (Jokar et al. 2016) have also been developing as nanosensors for real-time environmental monitoring and crop health and growth (He et al. 2019). Nanosensors were used to measure soil properties (soil moisture, residual pesticides, nutrients, and pH), check for pathogen and

determination of nitrogen uptake which helps farmers to maintain crop productivity (Bellingham 2011; Prasad et al. 2017).

The promotion of nanotechnology can be used to monitor cultivation and farm status in real time, such as the level of moisture, soil fertility, temperature, crop nutrient capabilities, and pathogens. The use of nanosensors for accurate soil measurement of soil criteria (soil moisture, residual pesticides, nutrients, and pH), pathogens control, and nitrogen absorbing assessment enables farmers make better use of input and therefore foster sustainable farming (Bellingham 2011; Prasad et al. 2017). These nanosensors are suitable for inspection of water and agrochemical treatments over time and volume based on the need for temporal and spatial macronutrients on plants necessary for the precise agriculture mission. Modern work is being done to design, in the future, accurate irrigation water supply schemes. The key features of this development are storage of water, the ability to hold water in situ, distribution of water near the root, the capacity to consume water for crops, and enclosed water, freed from crops. Microelectromechanical system sensors (MEMS) based on nanotechnology have the capacity to detect and respond to changes in the environment via microelectronic circuits. Wireless nanosensing devices composed of MEMS beams covered with water-sensitive nanopolymers for humidity optimization were created effectively by researchers (Kaushal and Wani 2017).

7.5 Applications of Bionanosensors in Agriculture

7.5.1 Detection of Bacteria

Microorganisms which are food-borne are a major concern to human health, especially in countries where there is no system to monitor food quality (Kumar et al. 2012). Bacteria are the main cause of gastroenteritis, food poisoning, and other illnesses such as *Salmonella* and *Escherichia coli*. Traditional methods to detect pathogen in food are time-consuming and very expensive. The use of bionanosensors all through food production processes from growth in the field till processing and distributing offers a good way to monitor bacterial pathogens and toxins in food that is infected (Mufamadi and Sekhejane 2017).

In a research study by Viswanathan et al. (2006), they developed a bionanosensor from carbon nanotubes and liposomes to detect cholera toxin. The detection limit of such immunosensor was 10–16 g of cholera toxin. A nanoparticle-based quantum dot biosensor for the identification of *Salmonella* species in food products has been created by Kim et al. (2013) in another research study. An electrochemical biosensor for the identification of *Salmonella* was created in a further study led by Afonso et al. (2013). In a recent study by Wang and Alocijia (2015), they developed with the aid of a nano-particle-labeled biosensor a new method for rapid detection of *Escherichia coli* (Mufamadi and Sekhejane 2017) (Table 7.1).

	Type of biological		Biological							
Bionanosensor	moiety	Nanomaterials	application	References						
Detection of Bacteria										
Carbon nanotubes	Liposomes	Carbon	To detect cholera	Viswanathan						
and liposomes		nanotubes	toxin	et al. (2006)						
Silicon nano-probes	Goat anti-rabbits	Silicon	Identification of	Yao et al.						
attached to goat	antibody	nano-probes	Xanthomonas	(2009)						
anti-rabbits antibody			axonopdis							
Quantum dot	Salmonella	Quantum dots	To detect	Kim et al.						
biosensor	antibodies		Salmonella	(2013)						
Nano-particle-	Monoclonal	Magnetic and	Detection of	Wang and						
labeled biosensor	antibodies	gold	Escherichia coli	Alocijia						
		nanoparticles		(2015)						
Detection of mycotox	ins									
Nanoparticle- based	Polythionine	Gold	For determination	Owino et al.						
immunosensor		nanoparticles	of aflatoxin B1	(2008)						
Electrochemical	Anti-primary	Carbon	To examine for	Parker and						
immunosensor	antibody	nanoparticles	the affatoxin MI	Tothill						
	4 .91 1	<u> </u>	T1	(2009)						
Optical biological	Antibody	Gold-nanorod	Identification of	Xu et al.						
sensor		0.11		(2013)						
Electrochemical	Polyclonal antibodies	Gold	Detection of	Bone et al.						
Immunosensors		nanoparticles	ochratoxins	(2010)						
Biocompatible	Oligo(ethylene	Magnetic	Detection of	Turan and						
hioneneeeneer	glycol) monomethyl	nanoparticies	ochratoxin A	Sanin (2016)						
Detection of pesticide	es and nerdicides	0.11		37 1 4 1						
Acetylcholinesterase	Acetylcholinesterase	Gold	Detection of	Vimala et al.						
nanobiosensor	A . 1 1 1 .	nanoparticles	organophosphates	(2016)						
Enzyme	Acetylcholinesterase	Carbon	Detection of	Zhang et al.						
bionanosensor	A . 1 1 1 .	nanotubes	Carbaryi	(2008)						
Acetylcholinesterase	Acetylcholinesterase	Chitosan	Detection of	Guan et al.						
biosensors	A . 1 1 1 .	nanoparticles	organophosphates	(2012)						
Electrochemical	Acetylcholinesterase	Gold	Detection of	Song et al.						
Clubert		nanoparticles		(2013)						
Glyphosate biosensor	Horseradish	Gold	To detect the	Songa et al.						
	peroxidase	nanoparticies	alufosinate	(20090)						
Enzymatic biogeneer	Turacinaca	7:0	To dataat	Haddaani						
Enzymatic biosensor	Tyrosinase	nanonarticles	chlortoluron	and Reques						
		nanoparticies		(2015)						
				(-515)						

 Table 7.1
 Types of bionanosensors

7.5.2 Detection of Mycotoxins

Mycotoxins are toxic chemicals produced by fungi (molds). They are natural contaminants of human food and animal feed especially when the food is stored at humid areas favorable to maintain molds growth (Akbas and Ozdemir 2006). Mycotoxins which are known to be toxic to humans and animals are: ochratoxin, aflatoxins, and zearalenone (Hussein and Brasel 2001; Ventura et al. 2004). Ingestion of these mycotoxins poses a threat to human health as they are labeled as carcinogenic, mutagenic, hepatotoxic, and nephrotoxic (Hussein and Brasel 2001). Hence, many countries regulate the entry of such toxins through food. Because of these concerns, it is highly recommended to have sensors that detect such toxins even at the lowest possible concentrations. That is where bionanosensors become developed to achieve such a target (Mufamadi and Sekhejane 2017). Several nanoparticles (immunosense systems, optical, magnetic, quantum dots, and carbon nanotubes) have been suggested for detection or tracking of mycotoxins in contaminated food (Xu and Han 2004; Adanyi et al. 2007; Owino et al. 2008; Viswanathan and Radecki 2008; Kaushnik et al. 2008; Mufamadi and Sekhejane 2017). With regard to aflatoxin detection, an electrochemical immunosensor has been developed to examine the aflatoxin M1 contaminants in matrix and milk interference (Parker and Tothill 2009). In addition, Xu et al. (2013) have established an optical biological sensor made of gold-nanorod (GNR) conjugated antibody for the identification of aflatoxin B1 in peanut. For the detection of ochratoxins, Bone et al. (2010) have created electrochemical immunosensors made of polyclonal antibodies and gold nanoparticles (Table 7.1).

Recently, Turan and Sahin (2016) have created an additional method for the detection of ochratoxin A with biocompatible molecular magnetic nanoparticles (Mufamadi and Sekhejane 2017) (Table 7.1).

7.5.3 Detection of Pesticides and Herbicides

The toxic substances used to control agricultural pests, such as weeds, fungi, and insects, are pesticides. The benefits of pesticides are the protection of crops by killing farm pests and weeds by herbicides. Unfortunately, residues of pesticides in agricultural products concern human health. Thus, it is important to track and detect pesticides and herbicides.

Current methods of detection are hampered because they are time-consuming and very expensive (Mufamadi and Sekhejane 2017). Hence the need to develop bionanosensors to detect pesticides and herbicides is of great importance to human welfare. Several nano-based pesticide biosensors have been developed to identify and track food for pesticides like herbicides, fungicides, insecticides, and other contaminants (McGrath et al. 2012; Zhao et al. 2015; Vimala et al. 2016). Zhang et al. (2008) created a new multiwalled carbon nanotube-based enzyme biosensor for the identification of pesticide residues in food. Acetilcholinesterase biosensors for the detection of residues of dichlorvos pesticides were developed in another study by Guan et al. (2012). In a new study, Song et al. (2015) have developed a simple electrochemical biosensor to measure carbamate pesticide quantitatively in real samples. The new glyphosate biosensor to detect the herbicide glufosinate has been created by Songa et al. (2009a, b, c). A disposable enzymatic nanobiosensor for the identification of chlortoluron herbicides has been developed in another research by Haddaoui and Raouafi (2015) (Table 7.1).

7.5.4 Detection of Veterinary Drugs

Veterinary drugs or antibiotics are used in farming animals to prevent and treat animal diseases. They are also suitable for improving animal growth, animal health, and feed efficiency (McEwen and Fedorka-Cray 2002). However, the use of antibiotics for animals may pose concerns as being found as residues in the meat and milk which are consumed by humans, hence poses human health problems. Thus, there is a great need to monitor the residues of such drugs in the food we eat. From this point, the need to develop bionanosensors to monitor animal drugs was of great importance. In the context of bionanosensors, intelligent analytical devices can be developed to determine the level or track the presence in real time of veterinary drug products (Wu et al. 2014, 2015). It also promises to provide cheaper, extremely sensitive, specific and precise equipment (Huet et al. 2010). Many nano-organic biosensors are specifically created to determine the level and the concentration of various kinds of veterinary antibiotics and drug residue in meat, poultry, milk, and much more. Most current biosensors are electrical biosensors, visual and aptamer. In addition, nanosensors offered an opportunity to detect residues of veterinary drugs in food by means of single or multi-analyte methods (Mungroo and Neethirajan 2014).

7.6 Conclusion

Nanotechnology has revolutionized all agricultural practices at research labs. The use of nanofertilizers, nanopesticides, nanoherbicdes, and eventually nanosensors has led to a new form of agriculture which is precision agriculture. Nanosensors have been used to detect pathogens, pesticides residues in plants and monitor plant and soil conditions. Bionanosensors have been employed to detect bacteria and mycotoxins throughout all agricultural processes till production of food. Also, they were used to detect pesticides, herbicides, and veterinary drugs. The use of nanosensors and bionanosensors is still at research level. Concerns arise as how these monitors can be monitored in a way to protect food and environment from being

polluted with nanoparticles. Further work is needed to assess the safety of using nanosensors and bionanosensors on human and animal health.

References

- Abbacia A, Azzouz N, Bouznit Y (2014) A new copper doped montmorillonite modified carbon paste electrode for propineb detection. Appl Clay Sci 90:130e4. https://doi.org/10.1016/j. clay.2013.12.036
- Adanyi N, Levkovets IA, Rodriguez-Gil S, Ronald A, Váradi M, Szendro I (2007) Development of immunosensor based on OWLS technique for determining aflatoxin B1 and ochratoxin A. Biosens Bioelectron 22(6):797–802. https://doi.org/10.1016/j.bios.2006.02.015
- Afonso AS, Perez-Lopez FRC, Mattoso LHC, Hernandez M (2013) Electrochemical detection of *Salmonella* using gold nanoparticles. Biosens Bioelectron 40(1):121–126. https://doi.org/10.1016/j.bios.2012.06.054
- Akbas M, Ozdemir M (2006) Effect of different ozone treatments on aflatoxin degradation and physicochemical properties of pistachios. J Sci Food Agric 86(13):2099–2104. https://doi. org/10.1002/jsfa.2579
- Bellingham BK (2011) Proximal soil sensing. Vadose Zone J 10:1342–1342. https://doi. org/10.2136/vzj2011.0105br
- Bone L, Vidal J, Duato P, Castillo J (2010) Ochratoxin A nanostructured electrochemical immunosensors based on polyclonal antibodies and gold nanoparticles coupled to the antigen. Anal Methods 2:335–341. https://doi.org/10.1039/B9AY00297A
- Chen H, Yada R (2011) Nanotechnologies in agriculture: new tools for sustainable development. Trends Food Sci Technol 22:585–594. https://doi.org/10.1016/j.tifs.2011.09.004
- Chhipa H, Joshi P (2016) Nanofertilisers, nanopesticides and nanosensors in agriculture. In: Ranjan S, Dasgupta N, Lichtfouse E (eds) Nanoscience in food and agriculture 1. Sustainable agriculture reviews, vol 20. Springer, pp 247–282. https://doi.org/10.1007/978-3-319-39303-2_9
- Deng H, Gao Y, Dasari TPS, Ray PC, Yu H (2016) A facile 3D construct of graphene oxide embedded with silver nanoparticles and its potential application as water filter. J Miss Acad Sci 61:190e7
- Esser B, Schnorr JM, Swager TM (2012) Selective detection of ethylene gas using carbon nanotubebased devices: utility in determination of fruit ripeness. Angew Chem Int Ed 51:5752e6. https:// doi.org/10.1002/anie.201201042
- Geszke-Moritz M, Clavier G, Lulek J, Schneider R (2012) Copper-or manganese-doped ZnS quantum dots as fluorescent probes for detecting folic acid in aqueous media. J Lumin 132:987e91. https://doi.org/10.1016/j.jlumin.2011.12.014
- Gomes RC, Pastore VAA, Martins OA, Biondi GF (2015) Nanotechnology applications in the food industry: a review. Braz J Hyg Anim Sanity 9(1):1–8. https://doi. org/10.5935/1981-2965.20150001
- Guan H, Zhang F, Yu J, Chi D (2012) The novel acetylcholinesterase biosensors based on liposome bioreactors-chitosan nanocomposites film for detection of organophosphates pesticides. Food Res Int 49(1):15–21. https://doi.org/10.1016/j.foodres.2012.07.014
- Haddaoui M, Raouafi N (2015) Chlortoluron-induced enzymatic activity inhibition in tyrosinase/ ZnO NPs/SPCE biosensor for the detection of ppb levels of herbicide. Sensors Actuators B Chem 219:171–178. https://doi.org/10.1016/j.snb.2015.05.023
- He X, Deng H, H-m H (2019) The current application of nanotechnology in food and agriculture. J Food Drug Anal 27:1–21. https://doi.org/10.1016/j.jfda.2018.12.002
- Huet A, Fodey T, Haughey SA, Weigel S, Elliott C, Delahaut P (2010) Advances in biosensor-based analysis for antimicrobial residues in food. Trends Anal Chem 29(11):1281–1294. https://doi. org/10.1016/j.trac.2010.07.017

- Hussein HS, Brasel JM (2001) Toxicity, metabolism, and impact of mycotoxins on humans and animals. Toxicology 167(2):101–134. https://doi.org/10.1016/S0300-483X(01)00471-1
- Jokar M, Safaralizadeh MH, Hadizadeh F, Rahmani F, Kalani MR (2016) Design and evaluation of an apta-nano-sensor to detect Acetamiprid in vitro and in silico. J Biomol Struct Dyn 34:2505e17. https://doi.org/10.1080/07391102.2015.1123188
- Kaushal M, Wani SP (2017) Nanosensors: frontiers in precision agriculture. In: Prasad R, Kumar M, Kumar V (eds) Nanotechnology. Springer, Singapore, pp 279–291. https://doi. org/10.1007/978-981-10-4573-8_13
- Kaushnik A, Solanski P, Ansari A, Ahmad S, Malhorta B (2008) Chitosan-iron oxide nanobiocomposite based immunosensor for ochratoxin-a. Electrochem Commun 10(9):1364–1368. https:// doi.org/10.1016/j.elecom.2008.07.007
- Kim G, Park SB, Moon J, Lee S (2013) Detection of pathogenic Salmonella with nanobiosensors. Anal Methods 5:5717–5723. https://doi.org/10.1039/C3AY41351A
- Kumar S, Dilbaghi N, Barnela M, Bhanjana G, Kumar R (2012) Biosensors as novel platforms for detection of food pathogens and allergens. BioNanoSci 2(4):196–217. https://doi.org/10.1007/ s12668-012-0057-2
- Lin D, Xing B (2008) Root uptake and phytotoxicity of ZnO nanoparticles. Environ Sci Technol 42:5580–5585. https://doi.org/10.1021/es800422x
- Lin YW, Huang CC, Chang HT (2011) Gold nanoparticle probes for the detection of mercury, lead and copper ions. Analyst 136:863e71. https://doi.org/10.1039/c0an00652a
- McEwen SA, Fedorka-Cray PJ (2002) Antimicrobial use and resistance in animals. Clin Infect Dis 34(3):93–106. https://doi.org/10.1086/340246
- McGrath TF, Elliott CT, Fodey TL (2012) Biosensors for the analysis of microbiological and chemical contaminants in food. Anal Bioanal Chem 403:75–92. https://doi.org/10.1007/s00216-011-5685-9
- Mufamadi MS, Sekhejane PR (2017) Nanomaterial-based biosensors in agriculture application and accessibility in rural Small holding farms: food security. In: Prasad R, Kumar M, Kumar V (eds) Nanotechnology: an agricultural paradigm. Springer, pp 263–278. https://doi. org/10.1007/978-981-10-4573-8_12
- Mungroo NA, Neethirajan S (2014) Biosensors for the detection of antibiotics in poultry industrya review. Biosensors 4:472–493. https://doi.org/10.3390/bios4040472
- Mura L, Dudáš T (2013) Development of ecological agricultural production and its latest global trends before the economic crisis. Int J Agric Innov Res 2:253–257
- Nair R, Varghese SH, Nair BG, Maekawa T, Yoshida Y, Kumar DS (2010) Nanoparticulate material delivery to plants. Plant Sci 179:154–163. https://doi.org/10.1016/j.plantsci.2010.04.012
- Omanović-Mikličanina E, Maksimocic M (2016) Nanosensors applications in agriculture and food industry. Bull Chem Tech Bosnai Herzegov 47:45–70
- Owino J, Arotiba O, Hendricks N, Songa E, Jahed N, Waryo TT, Ngece R, Baker P, Iwuoha E (2008) Electrochemical immunosensor based on polythionine/gold nanoparticles for the determination of aflatoxin B1. Sensors 8(12):8262–8274. https://doi.org/10.3390/s8128262
- Parker CO, Tothill IE (2009) Development of an electrochemical immunosensor for aflatoxin M in milk with focus on matrix interference. Biosens Bioelectron 24(8):2452–2457. https://doi.org/10.1016/j.bios.2008.12.021
- Prasad R (2014) Synthesis of silver nanoparticles in photosynthetic plants. J Nanoparticles 2014:Article ID 963961. https://doi.org/10.1155/2014/963961
- Prasad R, Pandey R, Barman I (2016) Engineering tailored nanoparticles with microbes: quo vadis. WIREs Nanomed Nanobiotechnol 8:316–330. https://doi.org/10.1002/wnan.1363
- Prasad R, Bhattacharyya A, Nguyen QD (2017) Nanotechnology in sustainable agriculture: recent developments, challenges, and perspectives. Front Microbiol 8:1014. https://doi.org/10.3389/ fmicb.2017.01014
- Sharma A, Rogers KR (1994) Biosensors. Meas Sci Technol 5:461-472
- Sharon M, Sharon M (2008) Carbon nanomaterials: applications in physico-chemical systems and biosystems. Def Sci J 58(4):460–485. https://doi.org/10.14429/dsj.58.1668

- Song Y, Chen J, Wang LA (2015) Simple electrochemical biosensor based on AuNPs/MPS/Au electrode sensing layer for monitoring carbamate pesticides in real samples. J hazardous 304:103–109. https://doi.org/10.1016/j.jhazmat.2015.10.058
- Songa EA, Somerset S, Waryo T, Baker PG, Iwuoha EI (2009a) Amperometric nanobiosensor for quantitative determination of glyphosate and glufosinate residues in corn samples. Pure Appl Chem 81(1):123. https://doi.org/10.1351/PAC-CON-08-01-15
- Songa EA, Waryo T, Jahed N, Baker PGL, Kgarebe B, Iwuoha EI (2009b) Electrochemical nanobiosensor for glyphosate herbicide and its metabolite. Electroanalysis 21(3–5):671–674. https:// doi.org/10.1002/elan.200804452
- Songa EA, Arotiba OA, Owino JH, Jahed N, Baker PG, Iwuoha EI (2009c) Electrochemical detection of glyphosate herbicide using horseradish peroxidase immobilized on sulfonated polymer matrix. Bioelectrochemistry 75(2):117–123. https://doi.org/10.1016/j.bioelechem.2009.02.007
- Srivastava AK, Dev A, Karmakar S (2018) Nanosensors and nanobiosensors in food and agriculture. Environ Chem Lett 16:161–182. https://doi.org/10.1007/s10311-017-0674-7
- Thakkar MN, Mhatre S, Parikh RY (2010) Biological synthesis of metallic nanoparticles. Nanotechol Biol Med 6:257–262. https://doi.org/10.1016/j.nano.2009.07.002
- Turan E, Sahin F (2016) Molecularly imprinted biocompatible magnetic nanoparticles for specific recognition of Ochratoxin A. Sensors Actuators B Chem 227:668–676. https://doi. org/10.1016/j.snb.2015.12.087
- Ventura M, Gomez A, Anaya I, Diaz J, Broto F, Agut M, Comellas L (2004) Determination of aflatoxins B1, G1, B2 and G2 in medicinal herbs by liquid chromatography-tandem mass spectrometry. J Chromatography A 1048(1):25–29
- Vimala V, Clarke SK, Urvinder Kaur S (2016) Pesticides detection using acetylcholinesterase nanobiosensor. Biosens J 5:1–4. https://doi.org/10.4172/2090-4967.1000133
- Viswanathan S, Radecki J (2008) Nanomaterials in electrochemical biosensors for food analysis. Pol J Food Nutrition Sci 58(2):157–164
- Viswanathan S, Wu L, Huang M, Ho J (2006) Electrochemical immunosensor for cholera toxin using liposomes and poly(3,4-ethylenedioxythiophene)-coated carbon nanotubes. Anal Chem 78(4):1115–1121. https://doi.org/10.1021/ac051435d
- Wang Y, Alocijia EC (2015) Gold nanoparticle-labeled biosensor for rapid and sensitive detection of bacterial pathogens. J Biol Eng 9:16. https://doi.org/10.1186/s13036-015-0014-z
- Wibowo KM, Sahdan MZ, Ramli NI, Muslihati A, Rosni N, Tsen VH et al (2018) Detection of *Escherichia coli* bacteria in wastewater by using graphene as a sensing material. In: Journal of physics: conference series. IOP Publishing, p 012063. https://doi. org/10.1088/1742-6596/995/1/012063
- Wu Y, Tang L, Huang L, Han Z, Wang J, Pan H (2014) A low detection limit penicillin biosensor based on single graphene nanosheets preadsorbed with hematein-ionic liquids-penicillinase. Mater Sci Eng C Mater Biol Appl 1(39):92–99. https://doi.org/10.1016/j.msec.2014.02.012
- Wu S, Zhang H, Duan S, Fang CC, Dai WZ (2015) Aptamer-based fluorescence biosensor for chloramphenicol determination using upconversion nanoparticles. Food Control 50:597–604. https://doi.org/10.1016/j.foodcont.2014.10.003
- Xu S, Han X (2004) A novel method to construct a third-generation biosensor: self-assembling gold nanoparticles on thiol-functionalized poly (styrene-coacrylic acid) nanospheres. Biosens Bioelectron 19(9):1117–1120. https://doi.org/10.1016/j.bios.2003.09.007
- Xu X, Liu X, Li Y, Ying Y (2013) A simple and rapid optical biosensor for detection of aflatoxin B1 based on competitive dispersion of gold nanorods. Biosens Bioelectron 47:361–367. https:// doi.org/10.1016/j.bios.2013.03.048
- Yao KS, Li S, Tzeng K, Cheng TC, Chang CY, Chiu C et al (2009) Fluorescence silica nanoprobe as a biomarker for rapid detection of plant pathogens. In: Proceedings of the advanced materials research. Trans Tech Publication, pp 513–516. https://doi.org/10.4028/www.scientific.net/ AMR.79-82.513
- Zhang S, Shan L, Tian Z, Zheng Y, Shi L et al (2008) Study of enzyme biosensor based on carbon nanotubes modified electrode for detection of pesticides residue. Chin Chem Lett 19:592–594. https://doi.org/10.1016/j.cclet.2008.03.014
- Zhao G, Wang H, Liu G (2015) Advances in biosensor-based instruments for pesticide residues rapid detection. Int J Electrochem Sci 10:9790–9807

Chapter 8 Sensors for the Detection of Food Contaminants



Duygu Çimen, Nilay Bereli, Handan Yavuz, and Adil Denizli

Contents

8.1	Introduction.	170
8.2	Sensor Applications for Food Contaminants	170
	8.2.1 Types of Sensors Based on Transducers.	171
8.3	Conclusions and Future Perspectives	178
Refe	rences	179

Abstract Food safety is a public health concern. Food contaminants could be metal ions, residues of pesticides, mycotoxins, drug residues, allergens, preservatives, and contaminants from food processing and packaging. The contamination hazards in the agricultural food chain include residues of agrochemicals and natural toxins. These chemical pollutants have serious effects on human health such as liver, kidney, and neurological syndromes even in the minimum limits. The food contamination with toxins and environmental pollutants occurs during the processing, packaging, preparing, storage, and transportation. Traditionally, food contaminants are detected by conventional analytical techniques. The other approach for detection of food contaminants is sensors. Sensors offer various advantages including simple use, high sensitivity and selectivity, user-friendly, on-site detection, rapid and low-cost operation. This chapter summarizes the different sensor applications in food contaminant detection and analysis.

Keywords Sensor \cdot Food \cdot Contaminants \cdot Detection \cdot Hazards \cdot Application \cdot Residue \cdot Limit of detection \cdot Health \cdot Environment

D. Çimen · N. Bereli · H. Yavuz · A. Denizli (🖂)

Department of Chemistry, Hacettepe University, Ankara, Turkey e-mail: denizli@hacettepe.edu.tr

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_8

8.1 Introduction

Food contaminants are chemical or biological substances. Food contaminants may be present in foods as a result of different stages such as food production, processing, transport, and storage (Aceña et al. 2015; Liang et al. 2015; Rapini and Marrazza 2017). The continuous use of these compounds is a worldwide problem due to the risks for food safety. Some food contaminants can be very harmful to human health even at low concentrations (Sliwinska et al. 2014; Scognamiglio et al. 2014; Vasilescu and Jean-Louis Marty 2016; Farahi et al. 2012). The residue of these substances even at trace levels can be a reason of food contamination. Although most food contaminants exist in detection limits, the bioaccumulation effect and continuous exposure may increase the risks for human and animal health. For this reason, analysis of food contaminants is essential to protect the ecosystem, ensure food quality and safety, and protect human health from potential hazards. Excessive application of food contaminants such as pesticide, metal ions, pathogens, mycotoxins, drug residues, allergens, and bacteria may leave harmful residues, including metabolites and degradation products into the food (Yılmaz et al. 2017; Hüseynli et al. 2019; Kantiani et al. 2010; Yang et al. 2014; Zhang et al. 2018a; Llorent-Martínez et al. 2019; Freire et al. 2016; Suaifan et al. 2017; Castillo et al. 2015). Significant amounts of food contaminant residues are a serious concern for human and animal health. Until today, various techniques have been used for the detection of food contaminants. They are traditionally detected by conventional analytical techniques such as liquid chromatography-tandem mass spectrometry, gas chromatography (GS), high-performance liquid chromatography (HPLC), mass spectrometry (MS), capillary electrophoresis (CEC), and enzyme-linked immunosorbent assays (ELISA) (Fig. 8.1) (He et al. 2007; Lin et al. 2008, 2018; Tao et al. 2012; Aşır et al. 2015).

These existing techniques have strong trace analysis ability, high reproducibility, and sensitivity with low detection limits. These techniques have many disadvantages such as being time consuming and laborious and requiring sophisticated equipment and expensive instruments, and they also require sample preparation and high-skilledlabors. In addition, they do not allow on-site and real-time application, which is especially important in emergency situations. The other approach for the detection of food contaminants is sensors. Sensors offer various advantages including simple use, high sensitivity and selectivity, user-friendly, on-site detection, rapid and low-cost operation (Zhu et al. 2015a; Li et al. 2019; Dai et al. 2013; Silva et al. 2018; Wu et al. 2018; Moises and Schäferling 2009).

8.2 Sensor Applications for Food Contaminants

Food contaminations have potential carcinogenic properties. Therefore, they need to be determined by a simple, fast, precise, and cost-effective method. The analysis of chemical contaminants needs to be determined compliance with regulatory limits and consumer safety. The detection and measurement of contaminants of foods is



Fig. 8.1 Available techniques for food contaminant detection

generally conducted by purely laboratory-based procedures such as gas chromatography (GC), high performance liquid chromatography (HPLC), and liquid chromatography coupled with mass spectrometry (LC–MS). All of these techniques require laboratories with stringent conditions, and they are expensive and complicated. These are also time-consuming and require trained professionals. In recent years, sensors have several advantages over other analytical methods. Compared with analytical methods, they are more preferable and have attracted increasing attention because of their high sensitivity, low cost, fast response, portability, simple and onsite operation. The use of sensors possesses fast and simple analysis, in situ testing, and additional benefit of low cost. This chapter summarizes different types of sensors developed for the control of food contaminants.

8.2.1 Types of Sensors Based on Transducers

Transducers are based on many physicochemical phenomena like electrochemical, optical, and mass-sensitive sensors, which is a basic classification of sensors.

Electrochemical Sensors

Electrochemical devices need a basic necessity. During recognition event, a measurable change in electrical properties (potential, current, impedance, etc.) must be observed on the sensor surface or in the solution media. These devices need special electrodes to work: working/sensing, counter, and reference electrodes. Electrodes can be made from Ag/AgCl, platinum, gold, graphite, silicon, etc. Electrochemical sensors can be classified into different analytical techniques: amperometric, potentiometric, or conductometric (Yanase et al. 2013).

In potentiometric sensors, the changes in ionic concentrations are determined by use of ion-selective electrodes. Abdel-Ghany et al. (Abdel-Ghany et al. 2017) designed a novel potentiometric sensor for dinotefuran detection in real samples. They prepared four potentiometric sensors using methacrylic acid or acrylamide and carboxylated polyvinyl chloride (PVC) as functional monomer. The performance of the prepared potentiometric membrane sensor was demonstrated in spiked cucumber and soil samples.

Lu et al. (2016) fabricated an electrochemical immunosensor for deoxynivalenol (DON) and fumonisin B1 (FB1) detection. In this study, disposable screen-printed carbon electrode (SPE) with gold nanoparticles (AuNPs) and polypyrrole (PPy)-electrochemically reduced graphene oxide (ErGO) nanocomposite film was prepared for antitoxin antibody immobilization. Electrical conductivity and biocompatibility studies were performed. The limit of detection (LOD) and linear range of FB1 were found as 4.2 ppb, and 0.2 to 4.5 ppm (%RSD: 4.9%), respectively (Table 8.1). The corresponding values for DON were determined as 8.6 ppb, and 0.05 to 1 ppm (%RSD: 5.7%). The prepared immunosensor showed low-matrix interference and high sensitivity when extracts from spiked corn samples were used (Table 8.2).

One used food processing of the commonly contaminants is 3-monochloropropane-1,2-diol (3-MCPD). Fang et al. (2019) prepared a carbon dot-filter paper with a molecularly imprinted polymeric film to extract 3-MCPD from samples. The carbon dots have a high fluorescence intensity and can be easily adapted to sensor systems. The high adsorption capacity for this paper-based sensor was found to be 68.97 mg/g. The limit of detection (LOD) and an excellent selectivity of 3-MCPD were found as 0.6 ng/mL and imprinting factor of 4.5. The recoveries were determined from 97.2% to 105.3% with relative standard deviations <5.6%. These results are correlated linearly when compared to the results of the classical GC–MS method (R = 0.998) as shown in Fig. 8.2.

Ochratoxin A (OTA) is the most important mycotoxin known as a pollutant in grain and wine (Armutcu et al. 2018). The detection of OTA with sensitive and rapid detection methods is important for food safety monitoring. Zhang et al. (2018b) developed a lateral flow strip fluorescent aptasensor for one-step determination of OTA in corn samples. In the first experiment step, they immobilized biotin-cDNA on the surface of a nitrocellulose filter. Without OTA and in the presence of OTA, Cy5-labeled aptamer formed a stable double helix. The linear range of OTA was calculated as 1–1000 ng/mL. The limit of detection (LOD) was determined as

			Limit of	
Mycotoxins	Sensor format	Linear range	detection	References
DON	Ab-AuNPs-PPy/ErGO-SPE	0.05–1 ppm	8.6 ppb	Lu et al.
FB1		0.2–4.5 ppm	4.2 ppb	(2016)
DON	ELIME assay	100– 4500 ng/mL	63 ng/mL	Romanazzo et al. (2010)
DON	Label-free inhibition based electrochemical immunosensor	6-30 ng/mL	0.3 μg/mL	Sunday et al. (2015)
DON	96-well SPE plate	2–20 µg/g	1.1 μg/g	Ricci et al. (2009)
DON	AuNPs/ <i>p</i> -aminothiophenol/ folic acid/GC	0.1–20 μg/ mL	0.03 µg/mL	Gu et al. (2015)
FB1 + FB2 + FB3	Electrochemical magnetoimmunosensor	_	0.33 µg/L	Jodra et al. (2015)
FB1 + FB2	Direct competitive/ amperometry	1–1000 ng/ mL	5 ng/mL	Kadir and Tothill (2010)
FB1	Multichannel electrochemical sensor	0–54 μg/L	0.58 µg/L	Ezquerra et al. (2015)
FB1	Electrochemical immunosensor	0.01– 1000 ng/mL	0.002 ng/ mL	Yang et al. (2015)

Table 8.1 Comparison of the performance of immunosensor (Ab-AuNPs-PPy/ErGO-SPE) and those reported in literature (Lu et al. 2016)

 Table 8.2
 Recovery of FB1 and DON from spiked corn samples (Lu et al. 2016)

Toxins	Spiked (µg/kg)	Measured (µg/kg)	RSD (%)	Recovery (%)
FB1	100	94	8.51	94.1
	500	484.9	3.30	97.0
	2000	1904.2	2.21	95.2
	4000	4141.6	1.71	103.5
DON	100	93.1	9.76	93.1
	500	521.7	4.0	104.3
	1000	982.4	3.26	98.2

0.40 ng/mL. The recovery of OTA in nailed corn samples was between 96.4% and 104.67% (Table 8.3). These experimental results show that it will be used as strip sensors in the practical application for detection of the OTA level in grain samples.

Aflatoxin is one of the most common mycotoxins in food matrices. Aflatoxin B1 (AFB1) is the most toxic contaminant with teratogenic, mutagenic, immunosuppressive, and carcinogenic effects. Yao et al. (2019) investigated a chemiluminescent aptasensor for aflatoxin B1 (AFB1) detection in food samples. In this study, the detection sensitivity of prepared sensor improved by using the hybridization chain reaction (HCR) signal amplification strategy. After determining the optimal experimental conditions, the linear range of AFB1 showed a good linear correlation from 0.5 to 40 ng/mL. The limit of detection (LOD) was determined as 0.2 ng/mL. The practicability of the prepared aptasensor was analyzed with peanuts and milk samples from local market. The different concentrations AFB1 were spiked into peanuts



Fig. 8.2 A comparison of paper-grafted sensor and GC–MS methods for 3-MCPD determination in soy sauce 1 (A) and 2 (B) (Fang et al. 2019)

Spiked (ng/mL)	Detected (ng/mL)	Recovery (%)	CV (%)
3.00	3.14 ± 0.47	104.67	6.40
10.00	9.64 ± 1.15	96.40	4.80
30.00	29.17 ± 2.73	97.23	5.10

Table 8.3 Detection of the OTA levels in spiked corn samples (n:3) (Zhang et al. 2018a, b)

Table 8.4 Detection of AFB1 in peanuts and milk samples (n:3) (Yao et al. 2019)

Samples	Spiked (ng/mL)	Detected (ng/mL)	Total Found (ng/mL)	Recovery (%)
Peanut 1	10	5.76	16.02	102.6
Peanut 2	10	-	9.74	97.4
Milk 1	10	-	9.82	98.2
Milk 2	20	-	20.88	104.4

and milk sample solutions. The recovery of AFB1 in food samples was observed in the range of 97.4–104.4% (Table 8.4).

Zhang et al. (2015) developed fluorescence-labeled single-stranded DNAfunctionalized graphene oxide sensors for the detection of different types of chemical contaminants in food. The designed sensor device was analyzed for multiplex detection of Hg^{2+} , Ag^+ , and aminoglycoside antibiotic residues in food. Different concentrations of each analyte (0.5 mM, 1 mM, and 2 mM) were spiked into food samples. The recovery ratios (%) were found to be from 87.5% to 116% for Hg^{2+} , 91% to 126% for Ag^+ , and 95% to 101% for neomycin detection.

Sulfonamides are the synthetic bacteriostatic antibiotics commonly used in many European countries (Şanli et al. 2010). The maximum residue limits (MRL) of antibiotics in foods have been established by the European Union AS (100 μ g/L). Adrian et al. (Adrian et al. 2009) prepared an immunosensor for the sensitive detection of sulfonamide antibiotics in milk. The limit of detection (LOD) of sulfonamide with the immunosensor in buffer and in milk was found as 0.2 ± 0.1 and 0.5 ± 0.1 μ g/L, respectively. The analysis time for all the experiments was less than 30 min. As a result, the experimental results showed that prepared immunosensors could be succesfully applied for sensitive and easy detection of sulfonamide antibiotics in milk.

Bisphenol A (BPA) is a widely used hormone disrupter. BPA is harmful to human health. BPA is usually found in food and beverage packages (Üzek et al. 2019; Shaikh et al. 2015). Xu et al. (2016) prepared the molecular imprinted nanocomposite and graphene sheets with high conductivity for the detection of bisphenol A. In this study, the chemical functionality and the two-dimensional structure of the graphene increased the sensitivity of the electrochemical sensor. The linear range of BPA was calculated as 3.21×10^{-10} to 2.8×10^{-1} g/L (*R*: 0.995). The limit of detection (LOD) was found to be 9.63×10^{-11} g/L.

Melamine is a cyanamide triazine with an organic-based property and is an important contaminant in food products (Dursun et al. 2016). Rovina and coworkers (Rovina and Siddiquee 2016) developed a fast, novel, simple, and effective electrochemical method for the analysis of melamine in milk products and infant milk. The modified gold electrode was characterized with cyclic voltammetry and differential pulse voltammetry for detection of melamine. Under optimal experimental conditions (pH 7.0 and 0.3 V), the prepared electrochemical sensor indicated an optimal response time, that is, 30 s. The linear range of melamine concentrations was from 9.6×10^{-4} to 9.6×10^{-13} M, and the limit of detection was calculated as 9.6×10^{-14} M.

Optical Sensors

Optical sensors have two basic methods to obtain sensor data from target molecules, which are label-based and label-free methods. Optical devices use special transducers like surface plasmon resonance (SPR), interferometers, gratings, and refractometers. This diversity causes plenty of different constructed optical devices. Optical biosensors are divided into several subclasses such as resonance, dispersion, reflection, refraction, phosphorescence, infrared absorption, Raman scattering, fluorescence, and chemiluminescence. Surface plasmon resonance (SPR)- and quartz crystal microbalance (QCM)-based optical sensors are widely used because of their high sensitivity (Jalilzadeh et al. 2019; Kartal et al. 2019).

Surface plasmon resonance (SPR)-based sensors offer the advantage of rapid, label-free simultaneous monitoring of binding events due to refractive index changes near the sensor surface (Jalilzadeh et al. 2019). SPR sensors can characterize the binding of target molecules to be determined on a sensor chip surface coated with a metal film (gold, 50 nm) directly without any labeling. With SPR sensors, it is possible to study the interactions of biological molecules such as proteins, oligonucle-otides, and lipids, from macromolecules to small particles such as phages, viral particles, and cells.

Pesticides applied to a broad range of composition, such as insecticides, herbicides, and fungicides. Use of pesticides is hazardous to humans and environment due to their nonspecific toxicological effects. Saylan et al. reported surface plasmon resonance (SPR) sensors for fast, selective, sensitive, and real-time detection of pesticides, including the most well-known triazine pesticides, atrazine (ATZ), cyanazine (SNZ), and simazine (SMZ). The biomimetic nanofilms on the SPR gold surfaces are prepared, which consist of *N*-methacryloyl-L-phenylalanine methyl ester (MAPA) as a functional monomer, 1-vinyl imidazole (VIM) as a comonomer, and ethylene glycol dimethacrylate (EGDMA) as a crosslinker. The real-time analysis on SPR sensor was performed with a linearity range from 0.10 to 6.64 nM pesticide (Saylan et al. 2017).

Oh et al. (2017) investigated a portable plasmonic biosensor that led to high sensitive and selective detection of *Salmonella typhimurium* in pork meat samples. Under experimental conditions, the limit of detection (LOD) was calculated as 10^4 CFU/mL for *S. typhimurium* in pure culture. All analysis time during the experiments was 30–35 min. The limit of detection of *S. typhimurium* in the spiked pork meat samples was found to be 1.0×10^4 CFU/mL.

Ochratoxin A (OTA) is harmful to human and exists usually in plant origin food. An anti-OTA aptamer immobilized sensor chip was prepared to measure ochratoxin A (OTA). Zhu and co-worker immobilized the streptavidin as a crosslinker onto the sensor chip. The biotin-aptamer was captured through streptavidin–biotin interaction. A linear range of OTA detection from 0.094 to 100 ng/mL was shown with a lower detection limit of 0.005 ng/mL. The detection of OTA was performed in the surface plasmon resonance (SPR) biosensor using simple liquid–liquid extraction for sample pretreatments in wine and peanut oil. Ochratoxin A recovery from spiked samples were found from 86.9% to 116.5% and coefficients of variation (CVs) were from 0.2% to 6.9%. The designed SPR biosensor method showed good reproducibility, stability, and performances with limits of detection much lower than the maximum residue limit.

Antibiotics such as benzyl-penicillin (PenG) and cephalosporin, amoxicillin are used usually in animal therapy. Maximum residue limits (MRLs) have established to prevent the negative impact of β -lactam and PenG residues present in the milk on human health. Pennacchioet and co-worker proposed an effective alternative to quantify the penicillin G using surface plasmon resonance (SPR) method. The produced antibodies were used as molecular recognition elements for the detection of PenG by SPR experiments. The detection limit was found as 8.0 pM. The results showed that the prepared SPR results could be successfully applied for the accurate and easy detection of PenG (Pennacchio et al. 2015).

Ciprofloxacin (CPX) is the most commonly used synthetic antibiotics. Sari et al. (2018) prepared SPR nanosensor for detection of ciprofloxacin. In this study, ciprofloxacin-imprinted nanoparticles (MIP/NPs) were prepared for sensitive and selective detection of ciprofloxacin and characterized with Fourier transform infrared (FTIR) spectroscopy, zetasizer, and contact angle measurements. The limit of detection (LOD) was found to be 3.21 and 7.1 ppb in ultrapure water and synthetic wastewater (SWW) for detection of CPX, respectively. The selectivity studies were used as competitor agents: tetracycline and enrofloxacin.

Erythromycin produced by *Saccharopolyspora erythreas* during the fermentation process is a macrolide antibiotic. Sari et al. (2016) developed novel surface plasmon resonance (SPR) nanosensor for detection of erythromycin (ERY). First, erythromycin-imprinted nanoparticles were synthesized and characterized by Fourier transform infrared (FTIR) spectroscopy, zetasizer, and scanning electron microscope (SEM). After, the erythromycin-imprinted nanoparticles were immobilized onto the SPR chip surface for detection of erythromycin. The limit of detection was calculated as 0.29 ppm. The stability of erythromycin-imprinted SPR chip was checked by using of equilibration-binding regeneration for several times.

Staphylococcal enterotoxin B (SEB) is an extracellular toxin produced by *Staphylococcus aureus*, causing food poisoning. Dudak and Boyaci developed a label-free SPR sensor using peptide ligands as recognition molecules. A 24-mer peptide was used for the detection of SEB and immobilized onto the gold-coated surface. Limit of detection was found as $20 \ \mu g/mL$. The sensor was tested with milk samples spiked with toxin and the signal from this sample was observed to be higher than toxin in buffer (Dudak and Boyaci 2014).

Mass-sensitive sensor devices work based on measuring the changes of properties of piezoelectric or magnetoelastic material via increased mass of biomolecules onto the sensor surface. Mass is a common property of all ions, small molecules, proteins, viruses, cells, etc. These sensors are applicable to all target biological structures without exception. Mass-sensitive sensors can work at both liquid and gas phases. Mass-sensitive devices can use quartz crystal microbalance, microcantilever, or magnetoelastic sensors. Quartz crystal microbalance (QCM) sensors produce signal with the changes in the resonance frequency of piezoelectric crystal upon interaction with analyte and have been used for the detection of food contamination in the last 5 years. However, the application of mass-based sensors for the food contamination detection is less than the electrochemical and optical sensors (Li et al. 2010; Raiteri et al. 2001).

Tobramycin is an aminoglycoside (AG) antibiotic that has ototoxicity and nephrotoxicity effects when used in a high amount. Yola et al. (2014) investigated quartz crystal microbalance (QCM) nanosensor for the detection of tobramycin (TOB). First, tobramycin-imprinted and nonimprinted poly(2-hydroxyethyl methacrylatemethacryloylamidoglutamic acid) films onto the QCM nanosensor surface were synthesized and characterized by using spectroscopy, atomic force microscopy (AFM), Fourier transform infrared (FTIR), and ellipsometry. The linear range of tobramycin concentrations was from 1.7×10^{-11} to 1.5×10^{-10} M. The limit of detection was calculated as 5.7×10^{-12} M. The prepared nanosensors were applied to chicken egg white and milk extract for the determination of TOB. The recovery of TOB in milk and egg samples is between 98.22% and 102.43%.

Formaldehyde is a pungent gas and is considered to be one of the main toxic closed pollutants; it is broadly used as an additive for preventing food spoilage, enhancing the flavor, and brightening the color. Hussain et al. reported the detection of formaldehyde vapors in air streams. QCM sensor coated with a copolymer thin film consisting of styrene, methacrylic acid, and ethylene glycol dimethacrylate provided a detection limit of 500 ppb formaldehyde in dry air (Hussain et al. 2016).

Bisphenol A (BPA), which is broadly employed in diverse consumables and crops, has been generally used as a monomer in fabricating polycarbonate epoxy resins and plastics. It is claimed that it can be leached from plastic products such as food packaging materials and acts as an endocrine disrupting chemical. In this study, Matsumoto et al. prepared bisphenol A (BPA)-imprinted polypeptide gel layers based on cyclodextrin-modified poly(L-lysine) (CD-PLL) on QCM sensor chips (Matsumoto et al. 2016).

8.3 Conclusions and Future Perspectives

Over the past several decades, significant effort has been invested with the aim of developing sensing technologies that will impact the practice of food research. Combination of several new techniques, derived from physical chemistry, molecular biology, biochemistry, thick and thin film physics, materials science, and electronics with the necessary expertise has revealed the promise for development of viable food useful sensor. The use of sensor systems in food technology is presented especially for standardization of raw materials, process development, and detection

of batch-to-batch variations and quality control of the final product. Device stability in an industrial environment and the know-how of industrial actors will be a key issue here. However, still a lot of research efforts are needed. Continued development is warranted because the sensors have advantages over more complex analytical techniques such as mass spectrometry and optical techniques. The integration of all analytical processes, from sample collection to analyte detection, on the same device, lab-on-a-chip system, can be sought. Sensor platform able to perform sample collection and analysis on the same device could be designed. These techniques include time-consuming procedures, expensive instruments, and hazardous labels. The development of rapid, easy to use, and cost-effective food contamination detection methods based on different sensor systems has been reported. Sensors are a highly sensitive technique and therefore very convenient for use in food contamination detection. The improvement of new sensors is a growing area and improvement provided in the field of sensitivity (low LOD), speed (fast detection), and easy use (portable devices).

References

- Abdel-Ghany MF, Hussein LA, El Azab NF (2017) Novel potentiometric sensors for the determination of the dinotefuran insecticide residue levels in cucumber and soil samples. Talanta 164:518–528. https://doi.org/10.1016/j.talanta.2016.12.019
- Aceña J, Stampachiacchiere S, Pérez S, Barceló D (2015) Advances in liquid chromatography– high-resolution mass spectrometry for quantitative and qualitative environmental analysis. Anal Bioanal Chem 407:6289–6299. https://doi.org/10.1007/s00216-015-8852-6
- Adrian J, Pasche S, Diserens JM, Sánchez-Baeza F, Gao H, Marco MP, Voirin G (2009) Waveguide interrogated optical *immunos*ensor (WIOS) for detection of sulfonamide antibiotics in milk. Biosens Bioelectron 24:3340–3346. https://doi.org/10.1016/j.bios.2009.04.036
- Armutcu C, Uzun L, Denizli A (2018) Determination of Ochratoxin a traces in foodstuffs: comparison of an automated on-line two-dimensional high-performance liquid chromatography and off-line immunoaffinity-high-performance liquid chromatography system. J Chromatogr A 1569:139–148. https://doi.org/10.1016/j.chroma.2018.07.057
- Aşır S, Derazshamshir A, Yılmaz F, Denizli A (2015) Triazine herbicide imprinted monolithic column for capillary electrochromatography. Electrophoresis 36:2888–2895. https://doi. org/10.1002/elps.201500232
- Castillo G, Spinella K, Poturnayova A, Snejdarkova M, Mosiello L, Hianik T (2015) Detection of aflatoxin B₁ by aptamer-based biosensor using PAMAM dendrimers as immobilization platform. Food Control 52:9–18. https://doi.org/10.1016/j.foodcont.2014.12.008
- Dai H, Gong L, Xu G, Zhang S, Lu S, Jiang Y, Lin Y, Guo L, Chen G (2013) An electrochemical sensing platform structured with carbon nanohorns for detecting some food borne contaminants. Electrochim Acta 111:57–63. https://doi.org/10.1016/j.electacta.2013.08.047
- Dudak FC, Boyaci İH (2014) Peptide-based surface plasmon resonance biosensor for detection of staphylococcal enterotoxin b. Food Anal Method 7:506–511. https://doi.org/10.1007/ s12161-013-9739-9
- Dursun EM, Üzek R, Bereli N, Şenel S, Denizli A (2016) Synthesis of novel monolithic cartridges with specific recognition sites for extraction of melamine. React Funct Polym 109:33–41. https://doi.org/10.1016/j.reactfunctpolym.2016.09.002
- Ezquerra A, Vidal JC, Bonel L, Castillo JR (2015) A validated multi-channel electrochemical immunoassay for rapid fumonisin B1 determination in cereal samples. Anal Methods 7:3742–3749. https://doi.org/10.1039/c4ay02897j

- Fang M, Zhou L, Zhang H, Liu L, Zhi-Yong Gong ZY (2019) A molecularly imprinted polymers/ carbon dots-grafted paper sensor for 3-T monochloropropane-1,2-diol determination. Food Chem 274:156–161. https://doi.org/10.1016/j.foodchem.2018.08.133
- Farahi RH, Passian A, Tetard L, Thundat T (2012) Critical issues in sensor science to aid food and water safety. ACS Nano 6:4548–4556. https://doi.org/10.1021/nn204999j
- Freire PG, Montes RHO, Romeiro FC, Lemos SCS, Lima RC, Richter EM, Munoz RAA (2016) Morphology of ZnO nanoparticles bound to carbon nanotubes affects electrocatalytic oxidation of phenolic compounds. Sensors Actuators B Chem 223:557–565. https://doi.org/10.1016/j. snb.2015.09.086
- Gu W, Zhu P, Jiang D, He X, Li Y, Ji J, Zhang L, Sun Y, Sun X (2015) A novel and simple cellbased electrochemical impedance biosensor for evaluating the combined toxicity of DON and ZEN. Biosens Bioelectron 70:447–454. https://doi.org/10.1016/j.bios.2015.03.074
- He L, Su Y, Fang B, Shen X, Zeng Z, Liu Y (2007) Determination of Sudan dyes residues in eggs by liquid chromatography and gas chromatography–mass spectrometry. Anal Chim Acta 594:139. https://doi.org/10.1016/j.aca.2007.05.021
- Hüseynli S, Çimen D, Bereli N, Denizli A (2019) Molecular imprinted based quartz crystal microbalance nanosensors for mercury detection. Global Chall 3:1800071. https://doi.org/10.1002/ gch2.201800071
- Hussain M, Kotova K, Lieberzeit PA (2016) Molecularly imprinted polymer nanoparticles for formaldehyde sensing with QCM. Sensors 16:1011. https://doi.org/10.3390/s16071011
- Jalilzadeh M, Çimen D, Özgür E, Esen C, Denizli A (2019) Design and preparation of imprinted surface plasmon resonance (SPR) nanosensor for detection of Zn(II) ions. J Macromol Sci A 56(9):877–886. https://doi.org/10.1080/10601325.2019.1617634
- Jodra A, Lopez MA, Escarpa A (2015) Disposable and reliable electrochemical magnetoimmunosensor for Fumonisins simplified determination in maize-based foodstuffs. Biosens Bioelectron 64:633–638. https://doi.org/10.1016/j.bios.2014.09.054
- Kadir MK, Tothill IE (2010) Development of an electrochemical immunosensor for fumonisins detection in foods. Toxins (Basel) 2:382–398. https://doi.org/10.3390/toxins2040382
- Kantiani L, Llorca M, Sanchís J, Farre M, Barcele D (2010) Emerging food contaminants: a review. Anal Bioanal Chem 398:2413–2427. https://doi.org/10.1007/s00216-010-3944-9
- Kartal F, Çimen D, Bereli N, Denizli A (2019) Molecularly imprinted polymer based quartz crystal microbalance sensor for the clinical detection of insülin. Mater Sci Eng C 97:730–737. https:// doi.org/10.1016/j.msec.2018.12.086
- Li S, Li Y, Chen H, Horikawa S, Shen W, Simonian A, Chin BA (2010) Direct detection of Salmonella typhimurium on fresh produce using phage-based magnetoelastic biosensors. Biosens Bioelectron 26:1313–1319. https://doi.org/10.1016/j.bios.2010.07.029
- Li X, Liu B, Wang F, Ma X, Li Z, Guo D, Wang Y, Wan F, Deng L, Zhang S (2018) Determination of 16 mycotoxins in maize by ultrahigh-performance liquid chro- matography-tandem mass spectrometry. Anal Lett 51:702–716. https://doi.org/10.1080/00032719.2017.1344246
- Li Y, Wang Z, Sun L, Liu L, Xu C, Hua Kuang H (2019) Nanoparticle based sensors for food contaminants. Trends Analyt Chem 113:74–83. https://doi.org/10.1016/j.trac.2019.01.012
- Liang HC, Liang N, Hay BM (2015) Analytical methods for pesticide residues in the water environment. Water Environ Res 87:1923–1937. https://doi.org/10.217 5/106143015X14338845156542
- Lin YF, Wang YC, Chang SY (2008) Capillary electrophoresis of aminoglyco- sides with argon-ion laser-induced fluorescence detection. J Chromatogr A 1188:331–333. https://doi.org/10.1016/j.chroma.2008.01.088
- Llorent-Martínez EJ, Fernández-Poyatos MP, Ruiz-Medina A (2019) Automated fluorimetric sensor for the determination of zearalenone mycotoxin in maize and cereals feedstuff. Talanta 191:89–93. https://doi.org/10.1016/j.talanta.2018.08.049
- Lu L, Seenivasan R, Wang YC, Yu JH, Gunasekaran S (2016) An electrochemical Immunosensor for rapid and sensitive detection of Mycotoxins Fumonisin B1 and Deoxynivalenol. Electrochim Acta 213:89–97. https://doi.org/10.1016/j.electacta.2016.07.096

- Matsumoto K, Tiu BDB, Kawamura A, Advincula RC, Takashi Miyata T (2016) QCM sensing of bisphenol a using molecularly imprinted hydrogel/conducting polymer matrix. Poly J 48:525–532. https://doi.org/10.1038/pj.2016.23
- Moises SS, Schäferling M (2009) Toxin immunosensors and sensor arrays for food quality control. Bioanal Rev 1:73–104. https://doi.org/10.1007/s12566-009-0006-x
- Oh SY, Heo NS, Shukla S, Cho HJ, Vilian ATE, Kim J, Lee SY, Han YK, Yoo SM, Huh YS (2017) Development of gold nanoparticle- aptamer-based LSPR sensing chips for the rapid detection of *Salmonella typhimurium* in pork meat. Sci Rep 7:10130. https://doi.org/10.1038/ s41598-017-10188-2
- Pennacchio A, Varriale A, Esposito MG, Scala A, Marzullo VM, Staiano M, D'Auria S (2015) A rapid and sensitive assay for the detection of benzylpenicillin (peng) in milk. PLoS One 10:0132396. https://doi.org/10.1371/journal.pone.0132396
- Raiteri R, Grattarola M, Butt HJ, Skládal P (2001) Micromechanical cantilever-based biosensors. Sens Act B 79:115–126. https://doi.org/10.1016/S0925-4005(01)00856-5
- Rapini R, Marrazza G (2017) Electrochemical aptasensors for contaminants detection in food and environment: recent advances. Bioelectrochemistry 118:47–61. https://doi.org/10.1016/j. bioelechem.2017.07.004
- Ricci F, Flavio P, Abagnale M, Messia M, Marconi E, Volpe G, Moscone D, Palleschi G (2009) Direct electrochemical detection of trichothecenes in wheat samples using a 96-well electrochemical plate coupled with microwave hydrolysis. World Mycotoxin J 2:239–245. https://doi. org/10.3920/WMJ208.1133.
- Romanazzo D, Ricci F, Volpe G, Elliott CT, Vesco S, Kroeger K, Moscone D, Stroka J, Van Egmond H, Vehniainen M, Palleschi G (2010) Development of a recombinant fab-fragment based electrochemical immunosensor for deoxynivalenol detection in food samples. Biosens Bioelectron 25:2615–2621. https://doi.org/10.1016/j.bios.2010.04.029
- Rovina K, Siddiquee S (2016) Electrochemical sensor based rapid determination of melamine using ionic liquid/zinc oxide nanoparticles/chitosan/gold electrode. Food Control 59:801–808. https://doi.org/10.1016/j.foodcont.2015.07.009
- Şanli N, Şanli S, Özkan G, Denizli A (2010) Determination of pKa values of some Sulfonamides by LC and LC-PDA methods in acetonitrile-water binary mixtures. J Braz Chem Soc 21:1952–1960
- Sari E, Üzek R, Duman M, Denizli A (2016) Fabrication of surface plasmon resonance nanosensor for the selective determination of erythromycin via molecular imprinted nanoparticles. Talanta 150:607–614. https://doi.org/10.1016/j.talanta.2015.12.043
- Sari E, Üzek R, Duman M, Adil Denizli A (2018) Detection of ciprofloxacin through surface plasmon resonance nanosensor with specific recognition sites. J Biomater Sci Polym Ed 29:1302–1318. https://doi.org/10.1080/09205063.2018.1457417
- Saylan Y, Akgönüllü S, Cimen D, Derazshamshir A, Bereli N, Yılmaz F, Denizli A (2017) Development of surface plasmon resonance sensors based on molecularly imprinted nanofilms for sensitive and selective detection of pesticides. Sensor Actuator B Chem 241:446–454. https://doi.org/10.1016/j.snb.2016.10.017
- Scognamiglio V, Arduini F, Palleschi G, Rea G (2014) Biosensing technology for sustainable food safety. Trends Analyt Chem 62:1–10. https://doi.org/10.1016/j.trac.2014.07.007
- Shaikh H, Sener G, Memon N, Bhanger MI, Nizamani SM, Üzek R, Denizli A (2015) Molecularly imprinted surface plasmon resonance (SPR) based sensing of bisphenol a for its selective detection in aqueous systems. Anal Methods 7:4661–4670. https://doi.org/10.1039/c5ay00541h.
- Silva NFD, Magalhães JMCS, Freire C, Delerue-Matos C (2018) Electrochemical biosensors for Salmonella: state of the art and challenges in food safety assessment. Biosens Bioelectron 99:667–682. https://doi.org/10.1016/j.bios.2017.08.019
- Sliwinska M, Wisniewska P, Dymerski T, Namiesnik J, Wardencki W (2014) Food analysis using artificial senses. J Agric Food Chem 62:1423–1448. https://doi.org/10.1021/jf403215y

- Suaifan GARY, Alhogail S, Mohammed Zourob M (2017) Paper-based magnetic nanoparticlepeptide probe for rapid and quantitative colorimetric detection of Escherichia coli O157:H7. Biosens Bioelectron 92:702–708. https://doi.org/10.1016/j.bios.2016.10.023
- Sunday CE, Masikini M, Wilson L, Rassie C, Waryo T, Baker PG, Iwuoha EI (2015) Application on gold nanoparticles-dotted 4-nitrophenylazo graphene in a label-free impedimetric deoxynivalenol immunosensor. Sensors (Basel) 15:3854–3871. https://doi.org/10.3390/s150203854
- Tao Y, Chen D, Yu H, Huang L, Liu Z, Cao X, Yuan Z (2012) Simultaneous determination of 15 aminoglycoside(s) residues in animal derived foods by automated solid-phase extraction and liquid chromatography-tandem mass spectrometry. Food Chem 135:676–683. https://doi. org/10.1016/j.chroma.2008.08.033.
- Üzek R, Sari E, Şenel S, Denizli A, Merkoç A (2019) A nitrocellulose paper strip for fluorometric determination of bisphenol a using molecularly imprinted nanoparticles. Microchim Acta 186:218. https://doi.org/10.1007/s00604-019-3323-y
- Vasilescu A, Jean-Louis Marty JL (2016) Electrochemical aptasensors for the assessment of food quality and safety. Trends Analyt Chem 79:60–70. https://doi.org/10.1016/j.trac.2015.11.024
- Wu W, Zhu Z, Li B, Liu Z, Jia L, Zuo L, Chen L, Zhu Z, Shan G, Luo SZ (2018) A direct determination of AFBs in vinegar by aptamer-based surface plasmon resonance biosensor. Toxicon 146:24–30. https://doi.org/10.1016/j.toxicon.2018.03.006
- Xu W, Yuan F, Li C, Huang W, Wu X, Yin Z, Yang W (2016) Acetylene black paste electrode modified with molecularly imprinted polymers/graphene for the determination of bisphenol a. J Sep Sci 39:4851–4857. https://doi.org/10.1002/jssc.201600803
- Yanase Y, Hiragun T, Yanase T, Kawaguchi T, Ishii K, Hide M (2013) Application of spr imaging sensor for detection of individual living cell reactions and clinical diagnosis of type I allergy. Allergol Int 62:163–169. https://doi.org/10.2332/allergolint.12-RA-0505
- Yang G, Fan MH, Zhang GM (2014) Emerging contaminants in surface waters in China-a short review. Environ Res Lett 9:074018. https://doi.org/10.1088/1748-9326/9/7/074018
- Yang X, Zhou X, Zhang X, Qing Y, Luo M, Liu X, Li C, Li Y, Xia H, Qiu J (2015) A highly sensitive electrochemical Immunosensor for Fumonisin B1Detection in corn using single-walled carbon nanotubes/chitosan. Electroanalysis 27:2679–2687. https://doi.org/10.1002/elan.201500169
- Yao Y, Wang H, Wang X, Wang X, Li F (2019) Development of a chemiluminescent aptasensor for ultrasensitive and selective detection of aflatoxin B1 in peanut and milk. Talanta 201:52–57. https://doi.org/10.1016/j.talanta.2019.03.109
- Yılmaz F, Saylan Y, Akgönüllü S, Çimen D, Derazshamshir A, Bereli N, Denizli A (2017) Surface plasmon resonance based nanosensors for detection of triazinic pesticides in agricultural foods. In: Grumezescu AM (ed) New pesticides and soil sensors. Elsevier, New York, pp 679–718
- Yola ML, Uzun L, Özaltın N, Denizli A (2014) Development of molecular imprinted nanosensor for determination of tobramycin in pharmaceuticals and foods. Talanta 120:318–324. https:// doi.org/10.1016/j.talanta.2013.10.064
- Zhang Y, Zuo P, Ye BC (2015) A low-cost and simple paper-based microfluidic device for simultaneous multiplex determination of different types of chemical contaminants in food. Biosens Bioelectron 68:14–19. https://doi.org/10.1016/j.bios.2014.12.042
- Zhang L, Zhu C, Chen C, Zhu S, Zhou J, Wang M, Shang P (2018a) Determination of kanamycin using a molecularly imprinted SPR sensor. Food Chem 266:170–174. https://doi.org/10.1016/j. foodchem.2018.05.128
- Zhang G, Zhu C, Huang Y, Yan J, Chen A (2018b) A lateral flow strip based Aptasensor for detection of Ochratoxin a in corn samples. Molecules 23:291. https://doi.org/10.3390/ molecules23020291
- Zhu C, Zhang L, Chen C, Zhou J (2015a) Determination of Bisphenol a using a molecular imprinted polymer surface plasmon resonance sensor. Anal Lett 48:1537:1550. https://doi. org/10.1080/00032719.2014.996809
- Zhu Z, Feng M, Zuo L, Zhu Z, Wang F, Chen L, Li J, Shan G, Luo SZ (2015b) An aptamer based surface plasmon resonance biosensor for the detection of ochratoxin a in wine and peanut oil. Biosens Bioelectron 65:320–326. https://doi.org/10.1016/j.bios.2014.10.059

Chapter 9 Nanotechnology-Based Detection and Remediation of Mycotoxins for Food and Agriculture Applications



Manjunath Manubolu D, Lavanya Goodla D, Stuart A. Ludsin, Thanasekaran Jayakumar, Mike Fraker, and Kavitha Pathakoti D

Contents

9.1	Introd	uction	184
9.2	Nano-	sensors for the Detection of Mycotoxins	189
	9.2.1	Gold Nanoparticles for the Detection of Mycotoxins	189
	9.2.2	Magnetic Nanoparticles for the Detection of Mycotoxins	192
	9.2.3	Quantum Dots for the Detection of Mycotoxins	195
	9.2.4	Miscellaneous Nanomaterials Employed in Developing Sensors for the	
		Detection of Mycotoxins.	197
9.3	Reme	diation of Mycotoxins From Food and Agriculture Using Nanotechnology	200
	9.3.1	Removal of Mycotoxins by Using Adsorption	200
	9.3.2	Degradation of Mycotoxins Using Nanotechnology	202
9.4	Summ	nary and Conclusion	204
Refe	erences.		205

M. Manubolu · S. A. Ludsin

Aquatic Ecology Laboratory, Department of Evolution, Ecology and Organismal Biology, The Ohio State University, Columbus, OH, USA

L. Goodla

Department of Biochemistry and Molecular Biology, University of New Mexico, Albuquerque, NM, USA

T. Jayakumar Graduate Institu

Graduate Institute of Medical Sciences, College of Medicine, Taipei Medical University, Taipei, Taiwan

M. Fraker (⊠) Cooperative Institute for Great Lakes Research, University of Michigan, Ann Arbor, MI, USA

K. Pathakoti (⊠) Interdisciplinary Center for Nanotoxicity, Department of Chemistry, Physics and Atmospheric Sciences, Jackson State University, Jackson, MS, USA

Department of Biology, Jackson State University, Jackson, MS, USA e-mail: kavitha.pathakoti@jsums.edu

© The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021 V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_9 183

Abstract Mycotoxins are highly toxic secondary metabolites produced by fungi, which may contaminate a large variety of food and feed commodities. Among them, the aflatoxins, deoxynivalenol, zearalenone, fumonisins, and ochratoxins are the most common contaminants posing a serious threat to human and animal health. Given that traditional mycotoxin detection methods have been shown to be laborious and time consuming, interest in developing reliable and rapid mycotoxin detection methods has increased during recent decades. Herein, we review emerging nanotechnology-based methods, including gold nanoparticles, magnetic nanoparticles, and quantum dot-based sensors, which have been developed to reliably and efficiently detecting mycotoxins in food and feed commodities. We also summarize recent technologies used to remove mycotoxins via adsorption and photocatalytic degradation. As our review illustrates, the emerging use of nanotechnology offers a reliable and cost-effective means to prevent mycotoxin contamination in food and feed commodities, which could reduce health risks to consumers.

Keywords $Mycotoxins \cdot Food contamination \cdot Nanotechnology \cdot Nanosensors \cdot Gold nanoparticles \cdot Quantum dots \cdot Magnetic nanoparticles \cdot Photocatalytic degradation$

9.1 Introduction

Mycotoxins are highly toxic secondary metabolites produced by fungi. They can invade a wide variety of agricultural crops while still on the field or afterwards during their processing in food and feed production chains. The presence of mycotoxins in agricultural commodities is high, with an estimated 25% of the world's crops being contaminated by molds or fungi, especially toxigenic species that belong to the genera Penicillium, Aspergillus, and Fusarium (Alshannaq and Yu 2017). At present, about 175 different mycotoxins have been identified in the food and feed commodities grown in outdoor and indoor environments (Bhat et al. 2010). Given their prevalence, mycotoxins have become a health hazard to both humans and animals worldwide (Fig. 9.1).

Based on possible dietary exposures together with their toxicity, aflatoxins (AFs), citrinin, patulin, ochratoxins A (OTA), zearalenone (ZEA), fumonisins, and trichothecenes are of a high concern due to their known health effects (Santos Pereira et al. 2019; Smith et al. 2016). Typical structures, source of contamination, food products contamination, health effects on mammalian system, and countrywise maximum tolerable levels of a wide range of mycotoxins are represented in Table 9.1. Risk assessments of mycotoxins in food are governed by the Joint Expert Committee of the Food and Agriculture Organization (JECFA) of the United Nations (FAO) and World Health Organization (WHO). The international standards are established by the Codex Alimentarius Commission, which lists maximum levels for various mycotoxins in foods (Jukes 2000; Organization 1999).



Fig. 9.1 Estimated worldwide health risk caused by mycotoxins during 2018. Data source: World Nutrition Forum/Database (BIOMIN, 2019)

The traditional methods of mycotoxin analysis involve more complex sample preparation and longer detection times combined with the usage of highly sophisticated analytical instruments (Vidal et al. 2013; Zhang et al. 2018a). Nanotechnology has gone through a booming development since 1981. The usage of nanoparticles in sensing applications are accurate with high precision resulting in signal amplification due to their unique characteristics, such as high surface to volume ratio (Tothill 2011). Further the integration of nanomaterials not only enhances the sensitivity in detection of toxins, but they are highly selective, cost effective, and portable and a rapid response can be obtained (Malhotra et al. 2014). A wide-range of nanomaterials comprising gold nanoparticles, silver nanoparticles, quantum dots (QDs), magnetic nanoparticles, graphene oxide, carbon nanohorns, carbon nanotubes (CNTs), and TiO₂ nanoparticles, have been used for the detection of different kinds of mycotoxins.

Till date many reviews have been published in sensing and detection of mycotoxins using nanotechnology (Goud et al. 2018; Horky et al. 2018; Ingle et al. 2020; Zeng et al. 2016). This is a comprehensive review including the various sensing applications with some of extensively used nanoparticles, such as gold, QDs, and magnetic nanoparticles. Here, we review advances from the last 5 years in the use of nanomaterials for the detection of mycotoxins in food and agriculture. This review also summarizes the recent technologies employed for the removal and the photocatalytic degradation and removal of mycotoxins. A literature search for the years

				Refs.	chuan Li et al.	(2011), Wang et al.	(2011) and Ye et al. (2010)				(Arévalo et al. (2011) and Flais	and Peraica (2009)			Abdul Kadir and	Tothill (2010) and Ghali et al. (2009)	~				
ci mum		Tolerable	levels	$(\mu g k g^{-1})$	14-15				20	30					800-4000		2000-	4000	2000-	5000	
Country-wise may	tolerable levels			Country	Australia,	Canada, Europe	Nigeria, New Zealand	South Africa	USA, Brazil	India					Europe, Turkey,	Norway, Switzerland	USA		Brazil		
			Health effects on	mammalian system	Carcinogenic, acute	hepatitis, impaired	immune system				Nephrotoxicity, hepatotoxicity.	embry otoxicity,	cytotoxicity,	immunotoxicity, carcinogenicity	Carcinogenic,	nephrotoxic, hepatotoxic, teratogenic)				
			Food products	contaminated	Maize, oil seeds, nuts,	dried fruits, cereals,	spices, milk, and milk products including	infant food			Oats, rice, corn, beans. fruits. fruit and	vegetable juices,	herbs, and spices		Maize, rice, sorghum,	cereals, green gram					
				Source	Aspergillus flavus, Aspergillus	nominus, Aspergillus parasiticus					Aspergillus carneus, Aspergillus niveus. Aspergillus terreus.	Penicillium citrinum, Penicillium	verrucosum, Penicillium	expansum	Aspergillus alternaria, Fusarium	anthophilum, Fusarium dlamini, Fusarium moniliforme.	Fusarium naphiformel,	Fusarium nygama, Fusarium	proliferatum, Fusarium verticillioides		
			Mycotoxin type/	Typical structure	Aflatoxins (B1, B2,	M1, M2, G1, and G2)			HY CH3		Citrinin		CH3	CH, CH,	Fumonisins (B1, and	B2)	⁴ NO CON	~~		He have	40 40 HO 40H

Table 9.1Mycotoxin sources, occurrence in foods, and health effects

	Heurich et al.	(2011), Pfohl-	Leszkowicz et al.	(CUI2), Barthelmehs et al	(2011) and Wu	et al. (2011)	Chehri and Godini	(2017), Li et al.	(2018b) and Maragos (2012)			Authority et al.	(2017) and Pleadin	et al. (2018)	(continued)
	20–30	5	2-10	20	5	50	500-	1750a	750– 3000a	700-1000	1000	Not	permitted	50–100a	
	Brazil	China, Kenya	Europe, Egypt	India	Nigeria, Russia	Uruguay	Europe		Brazil	Russia	Canada, China, India, Japan, USA	Europe		Russia	
	Mild liver damage, nephrotoxicity, and immune suppression					Weight loss, diarrhea,	vomiting,	gastrointestinal hemorrhaging,	immune-depressants,	uerman necrosus mutagenic, neurotoxic	Weight loss, diarrhea,	vomiting,	gastrointestinal hemorrhaging, immune-depressants, dermal necrosis mutagenic, neurotoxic		
	Maize, rice, rye, cereals, dry fruits, coffee, wine, beer, grape, juice, spices, licorice					Cereals, bakery products			Corn, wheat, barley, oats, rice, rye						
	Aspergillus alliaceus, Lapergillus auricomus, Lapergillus carbonarius, Lapergillus glaucus, Aspergillus melleus, Aspergillus niger, hencellium cyclopium, enicillium verrucosum, enicillium viridicatum					Fusarium graminearum,	Fusarium sporotrichioides,	Fusarium poae, Fusarium equiseti			F. sporotrichioides				
>	OTA	H-CH ²		0 HO 0 0H			Trichothecenes DON,	3- or 15-Ac-DON,	NIV (type B)	°	ъ щи uð	Trichothecenes T-2,	HT-2 (type A)	H ₃ C ^{-CH3} O ^{-H}	

Table 9.1 (continued)						
				Country-wise max tolerable levels	imum	
Mycotoxin type/		Food products	Health effects on		Tolerable levels	
Typical structure	Source	contaminated	mammalian system	Country	$(\mu g \ kg^{-1})$	Refs.
Patulin	Aspergillus clavatus, Aspergillus longivesica, Aspergillus terreus, Penicillium expansum, Penicillium griseofulvum, Ruscochlamus, sn	Apple juice and solid apple products	Genotoxicity, teratogenicity, cancer	Brazil, China, Europe, India, Japan, Kenya, Nigeria, Russia, South Africa	50	Erdoğan et al. (2018) and Zhong et al. (2018)
;	de chummingecha			USA USA		
Zearalenone	Fusarium culmorum, Fusarium	Corn, wheat, wheat	Estrogenic activity	Europe	75-400a	Panini et al. (2010)
	crookwellense, F. equiseti, F. graminearum, F.	flour, bread, cereals, noodles, rice, barley,	(infertility, vulval edema, vaginal prolapse,	Brazil	200– 1000a	
ch ở ch	sporotrichtoides	oats, sorghum, walnuts, milk, corn beer, meat, animal feed products	mammary hypertrophy in females, feminization of males	China, Russia, Chile	200,000	

188

from 2014 to 2019 was conducted in the PubMed (https://www.ncbi.nlm.nih.gov) and Scopus (https://www.scopus.com) databases using the keywords "nanoparticles, mycotoxin detection." Additionally, three categories of nanomaterials were searched specifically: "gold nanoparticles," "magnetic nanoparticles," and "quantum dots."

9.2 Nano-sensors for the Detection of Mycotoxins

At present, there is an increasing attention on nanotechnology since it has generated novel advantages across the range of areas comprising the food industry. Accordingly, nanotechnology has revealed new tasks for novelty in food production at a quick rate. Quality and safety of foods are a key component of public health, and with growing public awareness, customers are now demanding foodstuffs that are devoid of any contaminants (Hamad et al. 2018; Pal 2017; Pathakoti et al. 2019; Pradhan et al. 2015).

Due to their staggeringly small size, nanomaterials show specific physical and chemical features. Nanosensors are described as any chemical, biological, or surgical sensory point utilized to expose the nanoparticles to the microscopic world (Yu et al. 2018). Nanosensors utilize several nanomaterials that can identify toxins in sustenance at precise low levels, in the midst of handling or processing of foods (Sonawane et al. 2014; Willner and Vikesland 2018). Thus, in nanosensors, nanomaterials are employed in an analytical device, which is an improved version of a chemical sensor or biosensor. A biosensor works on two basic principles, biological recognition, and sensing. In general, nanosensors or nanobiosensors enhance at the activity by the integration of nanostructures in the sensing component for enhanced output. The basic structure of nanosensor and working principal is schematically shown in Fig. 9.2. These nanostructures enhance the activity both at bioreceptor level and the transducer level. Moreover, the transducer aids in fabrications of sensor with the use of various nanoparticles, such as metal oxide nanoparticles, quantum dots, magnetic nanoparticles, and carbon nanoparticles. Due to their staggeringly small size, nanomaterials show specific physical and chemical features, such as superior optical, electrical, thermal properties, and high surface area, which improves the transducing capability to a large extent. Based on signal production, that is, based on transducer type, nanosensors can be three major types: (1) optical: as optical (colorimetry, fluorescence, luminescence, surface-enhanced Raman scattering (SERS), surface plasmon resonance (SPR) and others), (2) electrochemical (amperometry, conductimetry, potentiometry, and voltammetry), and (3)Piezoelectric (quartz crystal microbalance).

9.2.1 Gold Nanoparticles for the Detection of Mycotoxins

A variety of biochemical and electrochemical assays based on Au nanoparticles have been described for the detection of mycotoxins. The most important and advantageous properties of gold nanoparticles for their usage in designing the



Fig. 9.2 Schematic representation of nanosensor and its working. modified from Goud et al. (2018), Copyright (2018) Elsevier

sensors to detect toxins include localized surface plasmon resonance (SPR) bands and enhanced scattering properties (Lara and Perez-Potti 2018). SPR imaging (SPRi) is an evolving label free semi-quantitative method for the sensitive detection of mycotoxins, which is highly reliable (Joshi 2017). Hossain and Maragos Hossain and Maragos (2018) described a gold nanoparticle based SPRi for the detection of Fusarium toxins such as DON, ZEA, and T-2 in wheat samples. This biosensor was amplified more than 12–90-fold with use of secondary antibody Ab2 with Au nanoparticles. Further, these antigen coated biochips are highly durable with short analysis time, are cost effective, and can be used for at least 46 cycles. In another study, Kong et al. (2016) have developed an Au nanoparticle-based immunochromatographic strip system for the detection of 20 different types of mycotoxins including ZEAs, DONs, T-2 toxins, AFs, and fumonisins (Fig. 9.3).

Similarly, AFs including B1, B2, M1, G1, and G2 in sustenance were identified depending upon a lateral flow strip (Santos et al. 2017). A monoclonal antibody (mB6 mAb), which has high specificity to AFs, was created to develop an immunochromatographic strip test, where the gold nanoparticles were coated with this antibody for detection. This strip consists of a control and test line, where the control line is coated with goat antimouse IgG and the test line containing AFB1 bovine serum albumin. In the strip, red gold nanoparticles form 1–2 pigmented lines on the



Fig. 9.3 (A) Immunochromatographic strip structure (B) Negative sample (C) Positive sample; c1: ZEAs; c2: DONs; c3: T-2s Toxin; c4: Aflatoxins; c5: Fumonisin B. Copyright (2016) Elsevier, (Kong et al. 2016)

membrane and the nanoparticles coated with anti-AFB1 as the signaling reagent for basic identification of AFB1.

A surface-enhanced Raman scattering (SERS) aptamer-based sensor was developed by using the gold@gold-silver nanostructures for the selective detection of OTA (Fig. 9.4). Magnetic nanoparticles (Fe₃O₄) were used for separation of toxin from solution, which is reusable. After magnetic separation, Raman signals were collected and this SERS probe can be used for the real time detection of OTA (Shao et al. 2018). Table 9.2 represents a wide range of methodology for sensor development using gold nanomaterials for the detection of different types of mycotoxins.



Fig. 9.4 Synthesis of Au@AuAg NNSs (A). Schematic illustration of SERS aptsensor-based on Au@AuAg NNSs-magnetic nanoparticles for OTA detection. Copyright (2018) Elsevier (Shao et al. 2018)

9.2.2 Magnetic Nanoparticles for the Detection of Mycotoxins

Superparamagnetism is one of the most important properties of magnetic nanoparticles. The superparamagnetic nanoparticles can be easily dispersed in aqueous solution, remain stable when coated with an appropriate layer, and are able to form ferrofluid. Wide ranges of applications of superparamagnetic nanoparticles are reported. Various methods of detection of mycotoxins by employing magnetic nanoparticles as sensors are represented in Table 9.3. A study by Gan et al. (2013) described the electrochemiluminescent immunoassay for the detection of mycotoxin, aflatoxin-M1 in milk samples. In their study, magnetic nanocomposites were prepared initially by immobilization of Fe_3O_4 nanoparticles onto graphene oxide, which is used as an adsorbent. Later on, aflatoxin-M1 antibody was attached to

Mycotoxin	Electrode/sensor	LOD or recovery (%)	Sample or matrix	Refs.
Patulin	Molecular imprinted polymer- SERS technique	96–108%	Fruit samples	Wu et al. (2019)
Ochratoxin A	AuNP@Cu-Co Purssian blue analogue-based electrochemical conductivity assay	5.2 fg mL ⁻¹	Spiked watermelon juice	Gu et al. (2019)
Patulin	AuNP-Black phosphorus nanosheets-electrochemical aptasensing	0.03 nM	Apple juice samples	Xu et al. (2019)
Fuminosin B1 and DON	Indium Tin oxide @ Polydimethylsiloxane electrodes functionalized with Au nanoparticles and anti-FB1 and anti-DON antibodies	0.3–140 ppb 0.2–60 ppb	Ground corn extract	Lu and Gunasekaran (2019)
ZEA	Calorimetric assay: use of an aptamer and Au nanoparticles with peroxidase like activity	10 ng mL	Corn and corn oil	Sun et al. (2018)
AFB1	Optical waveguide light mode spectroscopy (OWLS): signal enhancement by increasing the sensor surface through immobilization of Au nanoparticles on OWLS immunosensor chip	0.01– 10 ng mL ⁻¹	Hungarian paprika samples	Adányi et al. (2018)
Fuminosin B1 and B2	Fluorescence quenching immunoassay based on Au nanoparticles and a recombinant epitope-mimicking fusion protein	1.1 ng mL ⁻¹	Wheat samples	Peltomaa et al. (2018)
Fuminosin B1, B2, and B3	Indirect competitive enzyme-linked immunosorbent assay: Au nanoparticles-mercaptoundecanoic acid@ horseradish peroxidase— goat anti-mouse IgA	0.078µg mL ⁻¹	Maize	Li et al. (2018c)
AFB ₁	L-arginine- 6-aza-2-thiothymine— Au nanocluster-based photoluminescence enzyme immunoassay	3.2 pg mL ⁻¹	Peanut	Wang et al. (2018)
ΟΤΑ	Detection based on signal amplification by Exonuclease III (Exo III) and fluorescence quenching by Au nanoparticles	4.82 nM	Spiked red wine	Zhao et al. (2018)
Fuminosins	Lateral flow immunoassay: CdSe/ ZnS QD-BSA-FMB1@AuNP-Ab	1 ng mL ⁻¹	Maize flour samples	Anfossi et al. (2018)
DON; ZEA; T2/ HT2-toxin	Multiplex lateral flow immunoassay: monoclonal antibodies (mAb) conjugated with CdSeS/ZnS QDs or colloidal Au nanoparticles	80– 1000µg kg ⁻¹	Wheat samples	Foubert et al. (2017)

 Table 9.2 Detection of mycotoxins by using gold nanoparticles as sensors

(continued)

		I OD or	Sample or	
Mycotoxin	Electrode/sensor	recovery (%)	matrix	Refs.
AFB1	Label-free electrochemical	8 pg mL ⁻¹	Corn flakes	Chauhan
	Quartz crystal microbalance			et al. (2016)
	Based immunoassay: α-AFB1/			
	cysteamine/Au nanoparticles/			
	immunosensor			
ZEA	Immunochromatographic assay:	0.1 ng mL ⁻¹	Standard	Urusov et al.
	antibody immobilization on ablative Au nanoparticles		ZEA	(2015)
OTA	Label-free aptamer-based assay:	0.009 ng mL^{-1}	Liquor	Luan et al.
	cationic polymer poly		samples with	(2015a)
	chloride@Au nanoparticles		UIA	
OTA	SERS labels embedded Ag@Au	0.006 ng mL ⁻¹	Spiked	Zhao et al.
AFB1	core-shell nanoparticles	0.03 ng mL ⁻¹	mycotoxins	(2015)
			maize meal	
DON	Electrochemiluminescence	1 pg mL ⁻¹	Wheat flour	Lv et al.
	electrodes based on nanoporous			(2015)
	Cobalt/Co ₃ O ₄ – Au/RuS1@ Ru(hpy) ₂ ²⁺			
AFB1	AFB1-BSA conjugate modified	7 pg mL^{-1}	AFB1	Wang et al.
	magnetic beads were employed as	10	standard	(2016b)
	capture probe and <i>anti</i> -AFB1		solutions	
	used as detection probe for the			
	immunological recognition			
AFB1	SERS-based immunoassay using	0.1 ng mL ⁻¹	AFB1	Ko et al.
	silica-encapsulated hollow Au		standard	(2015)
DON	The electrochemical impedance	0.3µg mL ⁻¹	Certified	Sunday et al.
	spectroscopy of tris(bipyridine)		corn, wheat,	(2015)
	ruthenium (II) chloride was used as		and roasted	
	a marker enhanced with Au nanoparticles-dotted		coffee	
	4-nitrophenylazo functionalized			
	graphene nanocatalyst mediated in			
	Nation on a glassy carbon electrode	0.025 ng mt -1		Luon at al
AFBI	un-modified Au nanoparticles	0.023 lig lilL .	standard	(2015b)
	···· • • • • • • • • • • • • • • • • •		solutions	
AFB2	NaCl-induced aggregation of	25 pg mL ⁻¹	AFB2	Luan et al.
	aptamer-modified Au nanoparticles		solutions	(2015a)

Table 9.2 (continued)

(continued)

		LOD or	Sample or	
Mycotoxin	Electrode/sensor	recovery (%)	matrix	Refs.
ΟΤΑ	Immunochromatographic assay: OTA monoclonal antibody conjugated Au nanoparticles	0.25 ng mL ⁻¹	Maize, wheat, soybean, and rice	Majdinasab et al. (2015)
AFB1 OTA ZEA	SPRi: antibody-conjugated Au nanoparticles on poly[oligo(ethylene glycol) methacrylate-co-glycidyl methacrylate]@modified SPRi chip	8 pg mL ⁻¹ 30 pg mL ⁻¹ 15 pg mL ⁻¹	Spiked peanut samples	Hu et al. (2014)
Fumonisin B1	Electrochemiluminescence aptosensor: Au NP–Iridium complex as nanoprobe	0.27 ng mL ⁻¹	Spiked wheat flour	Zhao et al. (2014)

Table 9.2 (continued)

*LOD: Limit of detection

cadmium telluride quantum dots, which can be used as a signal tag. Finally, magnetic nanocomposites were used for the separation of aflatoxin-M1 from milk samples and detected using an immunoassay using an electrochemiluminescent signal. Further, this immunosensor is stable and can be regenerated after storing for two weeks at 4°C and its accuracy was about 95%.

9.2.3 Quantum Dots for the Detection of Mycotoxins

Physicochemical properties including enhanced fluorescence, narrow emission peaks, and high photostability of QDs are advantageous in developing the sensors for highly sensitive detection of mycotoxins. Some of the recent applications of quantum dots for mycotoxin detection are presented in Table 9.4. Fang et al. (2014) developed a novel molecularly imprinted optosensing material for selective detection of mycotoxin, ZON in cereal samples. The molecular imprinted polymer is based on ionic liquid stabilized cadmium selenium/ZnS QDs and the mechanism is based on fluorescence quenching.

A highly sensitive aptasensor for the detection of ochratoxin was developed with a combination of nanocomposites consisting of gold nanoparticles functionalized with silica coated Fe_3O_4 magnetic nanoparticles and CdTe QDs modified with graphene/gold nanoparticles. This novel aptasensor is ultrasensitive with detection limit at sub-picomolar level (Hao et al. 2016).

Zhang et al. (2017a) developed an optical sensor based on molecular imprinted polymer capped Mn-doped ZnS QD for detection of the mycotoxin, patulin, from apple juice. This nanosensor is based on a phosphorescence method, for determination of patulin from aqueous solutions, including the intended matrix, apple juice. Recently, Duan et al. (2019) developed multicolor CdSe/ZnS QD nanobeads with yellow, orange, and red luminescence for concurrent recognition of numerous

		LOD or	Sample or	
Mycotoxin	xin Electrode/sensor		matrix	Refs.
OTA	SERS: Au@AuAg-magnetic nanoparticle	0.004 ng mL ⁻¹	Red-wine samples	Shao et al. (2018)
ZEA	Aptamer-functionalized MNP @ fluorescence NaYF4: Ce/Tb nanoparticles	0.21 pg mL ⁻¹	Maize and wheat	Niazi et al. (2018)
AFB1	Magnetic nanoparticle-filled	0.15 and	Wheat	Li et al.
ZEA	amino-modified multi-walled carbon nanotubes (Fe ₃ O ₄ -MWCNTs-NH ₂)	0.24 ng g ⁻¹	flour samples	(2018a)
Fumonisin B1	Magnetic reduced graphene oxide/nickel/platinum nanoparticles	Iagnetic reduced graphene 0.70 ng mL ⁻¹ Beer and wine anoparticles 0.75 0.75		Molinero- Fernández et al. (2018)
ZEA;	Core-shell polydopamine	0.55-	Cow, goat,	Socas-
α-Zearalanol	coated magnetic nanoparticle	11.8µg L ⁻¹	sheep, and	Rodríguez
α -Zearalenol;	_		human	et al. (2018)
β-Zearalanol	_		пшк	
β-Zearalenol;	_			
β-Zearalanol- 10,10,11,12,12- d5				
AFB1	SERS based aptasensor: the amino-terminal aptamer conjugated magnetic-bead and the gold nanotriangles-DTNB @ Ag-DTNB nanotriangles	0.54 pg mL ⁻¹	Peanut oil samples	Yang et al. (2017)
AFB1	Magnetic microspheres encoded with fluorescent nanocrystals	9 pg mL ⁻¹	Spiked corn samples	Zhang et al. (2017b)
ZEA,	Core-shell poly(dopamine)	0.2-4.8µg L ⁻¹	Milk	González-
α-zearalanol,	magnetic nanoparticles		Yogurt	Sálamo et al.
β-zearalanol,				(2017)
α -zearalenol,				
β-zearalenol zearalanone				
OTA	Iuorescence resonance energy ransfer: quantum dot-labeled intibody to rhodamine-coated magnetic silica nanoparticles 0.8 pg mL ⁻¹ Spiked human serum		Mahdi et al. (2016)	
ZEA	Magnetic nanoparticles and chemiluminescent detection	es and 0.04 ng mL^{-1} Wheat Hend tection et al.		Hendrickson et al. (2016)
OTA	Chemiluminescence immunoassay: magnetic nanoparticles with targeted inhibition	1.39 pg mL ⁻¹	Rice	Kim and Lim (2015)

 Table 9.3 Detection of mycotoxins by using magnetic nanoparticles as sensors

(continued)

		LOD or Sample or		
Mycotoxin	Electrode/sensor	recovery (%)	matrix	Refs.
AFB1	FRET-based fluorescence 2×10^{-12} immunoassay: magnetic/silicacore-shell as a signal intensifier		Spiked human serum	Kalarestaghi et al. (2015)
AFB1	Immunoenzyme assay: antibody immobilization on the surface of magnetic nanoparticles	20 pg mL ⁻¹	Barley and maize extracts	Urusov et al. (2014)

Table 9.3 (continued)

mycotoxins such as ZEA, OTA, and fumonisin B1 from a corn matrix (Fig. 9.5). This is a qualitative method based on an immunochromatographic assay for visual detection of mycotoxins, which can be inferred with the naked eye under UV light with color differentiation.

9.2.4 Miscellaneous Nanomaterials Employed in Developing Sensors for the Detection of Mycotoxins

Zinc oxide nanoparticles are recognized for applications in immunosensing, due to their distinct characteristics, such as a high isoelectric point and higher binding energy and moreover being cost-effective and biocompatible (Ansari et al. 2010). For OTA detection, ZnO nanofilm was immobilized with rabbit immunoglobulin antibodies placed onto glass plate made of Indium tin oxide and bovine serum albumin for blocking nonspecific binding of OTA. This nanozinc immunoelectrode has amplified electrochemical signal and has an application for OTA and other mycotoxins (Ansari et al. 2010).

Similar to ZnO nanoparticles, nanostructured CeO_2 is biocompatible due to its non-toxicity and has attracted interest in development of biosensors. In addition, nano-CeO₂ has high chemical stability and mechanical strength. Kaushik et al. (2009a) described the fabrication of chitosan-nano-CeO₂ nanocomposites film deposited on indium tin oxide-coated glass substrate for the immobilization of r-IgGs and BSA. This immunoelectrode has a large surface, including greater electron transport and potential application for OTA detection and other mycotoxins such as AFB1 and citrining.

Likewise, a study by Kaushik et al. (2009b) reported the detection of OTA by integrating nanosilicon-chitosan nanocomposites film for r-IgGs loading, and this immunoelectrode has enhanced sensing properties. This could be due to the presence of chitosan-nano-SiO₂ composites due to greater surface area and higher electrochemical behavior. Similarly, a chitosan-TiO₂-nanoparticle-based immunosensor was developed for the recognition of OTA from *Aspergillus ochraceus* (Khan and Dhayal 2008). Electrochemical impedance spectroscopy (EIS) was used to determine the electroactivity of the bioactive electrode. This matrix has several advantages in biosensor application. Chitosan is cost-effective and has higher mechanical

Mycotoxin	Electrode/sensor	LOD	Sample or matrix	Refs.
ZEA	Electrochemiluminescence aptasensor: Nitrogen doped graphene QDs on amine functionalized Ru(bpy)3 2+-doped silica nanoparticles surface	1 fg mL ⁻¹	Corn flour	Luo et al. (2019)
Fumonisin B1	Fluorescence Enzyme linked immunoassay: mercaptopropionic acid–modified CdTe QDs	0.33 ng mL ⁻¹	Corn	Lu et al. (2018)
DON	Fluorescent-labeled immunosorbent assay: monoclonal antibody+amino functionalized multishell QDs	12.2µg kg ⁻¹	Maize	Zhang et al. (2018b)
AFB1	Glycine-enhanced 0.17- Chinese photoluminescence assay: 0.35 ng mL ⁻¹ medicinal 3-mercaptopropionic acid and herbs herbs thiol-terminated methoxy polyethylene Glycol functionalized Herbs		Zhang et al. (2018a)	
ΟΤΑ	Fluorescence resonance energy transfer: cerium oxide nanoparticles and graphene QDs aptosensor2.5 pg mL^{-1}Peanuts		Tian et al. (2018)	
ΟΤΑ	Direct competitive fluorescence linked immunosorbent assay: QD beads	0.028 pg mL ⁻¹	Corn Coffee Red wine	Xiong et al. (2017)
OTA, Fumonisin	Electrochemical detection: CdTe or Pbs QDs coated on to silica sphere	10 pg mL ⁻¹ – 10 ng mL ⁻¹ 50 pg mL ⁻¹ –	Maize	Wang et al. (2017)
BI		50 ng mL^{-1}	C. 1. 1 1	The second set
Pirlimycin	green fluorescent CdSe/ZnS core/ shell QDs immobilized in liposomes	0.5µg kg ⁻¹	natural milk samples	(2017)
ZEA,	Multiplex immunochemical assay:	50 and	Maize	Beloglazova et al. (2017)
DON	InP/ZnS QDs encapsulated in silica shells	500µg kg ⁻¹	Wheat	
DON, ZEA,	Immunoassay: colloidal QDs enrobed into silica shell	6.1 and 5.3	Cereals	Beloglazova et al. (2016)
& AFB1		5.4 and 4.1		
		2.6 and 1.9		
OTA	Fluorescence enzyme linked immunosorbent assay: glucose oxidase-mediated fluorescence quenching of mercaptopropionic acid-capped CdTe QDs	2.2 pg mL ⁻¹	Corn	Liang et al. (2016)
ZEA	Fluorescence enzyme linked immunosorbent assay: hydrogen peroxide-sensitive QDs	4.1 pg mL ⁻¹	Corn	Zhan et al. (2016)

 Table 9.4
 Detection of mycotoxins by using quantum dots

(continued)

Mycotoxin	Electrode/sensor	LOD	Sample or matrix	Refs.
AFB1	Photoelectrochemical immunoassay: CdTe QDs-modified photosensitive electrode	3.0 pg mL ⁻¹	Spiked peanut samples	Lin et al. (2016)
DON	Immunoassay: silica-coated CdSe/ CdS/ZnS core-shell QDs	20 ng mL ⁻¹	Food and feed	Goftman et al. (2016)
ΟΤΑ	Electrochemiluminescence assay: RuSi nanoparticles/CdTe QDs	3.0 fg mL ⁻¹	Corn and human serum	Wang et al. (2016a)
OTA	Fluorescence immunoassay:	0.05 pg mL ⁻¹	Corn	Huang et al.
	hydrogen peroxide-sensitive CdTe		Wheat	(2016)
	QDs		Rice	
ZEA	Immunochromatography assay: CdSe/ZnS QDs submicro beads	3.6 mg kg ⁻¹	Corn	Duan et al. (2015)
DON, ZEA,	Fluorescent immunosorbent assay:	$3.2\mu g \ kg^{-1}$	Cereals	Beloglazova
AFB1,	QD nanolabels	$0.6\mu g \ kg^{-1}$		et al. (2014)
T2-toxin,		$0.2\mu g \ kg^{-1}$		
R1		10µg kg ⁻¹		
		$0.4 \mu g \ kg^{-1}$		
AFB1	Fluorescent quenching assay: graphene oxide-aptamer-CdTe QDs	1.4 nM	Peanut oil	Lu et al. (2015)

Table 9.4 (continued)

*LOD: Limit of detection

strength and permeability. TiO_2 provides the biocompatibility and thereby provides longer life and improved stability to the electrode. This technique of conjugating chitosan with nanoparticles also provided the fastest response (25 s) with the lowest detection limit.

Recently, Goftman et al. (2016) synthesized the silica coated CdSe quantum dots to detect DON by the microemulsion method. Further, to increase the bioapplicability of QD@SIO₂ nanoparticles, they were modified using three diverse functional groups including amino, carboxyl, and epoxy groups and polyethylene glycol fragments. This developed fluorescence-labeled immunosorbent assay was used for rapid detection of mycotoxins, especially DON.

Carbon nanotubes (CNTs) are allotropes of carbon with a cylindrical nanostructure (Saifuddin et al. 2013). Among them, single-walled CNTs (SWCNTs) due to their exceptional characteristics such as greater surface area, high electrical conductivity, and mechanical strength provide a compatible environment to preserve the enzyme activity, thereby enhancing the electrochemical signal amplification (Guo 2013; Singh et al. 2009). Zhang et al. (2016) fabricated an electrochemical immunosensor for detection of AFB1 from corn flour using the SWCNTs and chitosan composites. As SWCNTs are not water soluble, addition of chitosan enhanced the dispersion of SWCNTs. This immunosensor was centered on an indirect competitive assay using the primary antibody anti-AFB₁ and AFB₁-BSA immobilized on glass carbon electrode.



Fig. 9.5 Scheme 1. (a) Schematic representation for the synthesis of tricolor QB via using the emulsification evaporation method. (b) Schematic illustration of developed tricolor QB-based mICA, in which three T lines of T1, T2, and T3 were designed for the simultaneous detection of ZEN, OTA, and FB1, while one C line was set to indicate the validity of developed mICA, respectively. (C) Schematic presentation for the interpretation of test results. 1, negative; 2, ZEN (–), OTA (–), FB1 (+); 3, ZEN (–), OTA (+), FB1 (–); 4, ZEN (+), OTA (–), FB1 (–); 5, ZEN (–), OTA (+), FB1 (+); 6, ZEN (+), OTA (–), FB1 (+); 7, ZEN (+), OTA (+), FB1 (–); 8, ZEN (+), OTA (+), FB1 (+); 9, invalid. ("+" and "–" indicate the "positive" and "negative," respectively). Copyright from (2019) Elsevier (Duan et al. 2019)

9.3 Remediation of Mycotoxins From Food and Agriculture Using Nanotechnology

9.3.1 Removal of Mycotoxins by Using Adsorption

Traditional enzyme-based decontaminant methods for the removal of mycotoxins from food have several drawbacks, such that the toxin may remain in the food matrix or that it can also result in highly toxic secondary metabolites (Jard et al. 2011; Manubolu et al. 2018). Although nanotechnology has advanced substantially in the detection of mycotoxins, very few studies are available on the elimination or removal of mycotoxins from various food matrices. Among the various immobilization approaches, magnetic nanoparticles are the best solid carriers due to their exclusive high saturation magnetization for facile separation and reusability (Ansari and Husain 2012). Some of the recent applications for removal of mycotoxins from various biological matrices are described in this section (Table 9.5).

Surface active maghemite nanoparticles (SAMNS Fe_2O_3 , 11 ± 2 nm) were used for the removal of citrinin in Monascus treated foods, which can be used in the food industry (Magro et al. 2016). The SAMN@citrinin conjugate complex was thoroughly characterized structurally and was magnetically isolated. The offered magnetic separation system was based on the citrinin binding on SAMNS surface and is based on the firm iron chelating keto-enol group on the toxin. In another study, Luo

			Max.	a 1	
Mycotoxin	Nanomaterial	Method	degradation or recovery (%)	Sample	Refs.
Applications	for removal	lifetilde			100101
Citrinin	Surface active maghemite nanoparticles (Fe ₂ O ₃)	Magnetic separation	70% removal	Biological matrixes	Magro et al. (2016)
Patulin	Chitosan-coated Fe_3O_4 nanoparticles	Magnetic adsorbent	99% within 60 min	Fruit juice; water or others	Luo et al. (2017)
Patulin	Magnetic chitosan- Fe ₃ O ₄ nanoparticles coated with <i>Candida utilis</i>	Magnetic adsorbent	90%	Orange juice	Ge et al. (2017)
Patulin	Zirconium-based MOF UiO-66(NH ₂) @Au-Cys	Adsorption	4.38µg mg ⁻¹	Apple juice	Liu et al. (2019)
Patulin	Magnetic multi- walled carbon nanotubes (MWCNT)	Magnetic adsorbent	640.2µg g ⁻¹	Aqueous systems	Zhang et al. (2019)
ZEA; fumonisin B1 and B2; DON	Magnetic graphene oxide nanocomposites	Adsorption	37–69%	Animal feed: palm kernel cake	Pirouz et al. (2017)
Applications	for degradation	1			·
DON	Graphene/ZnO hybrids	Photocatalytic (UV light)	99% of DON (15 ppm) within 30 min	Aqueous suspension	Bai et al. (2017)
AFB1	Scandium doped SrTi _{0.7} Fe _{0.3} O ₃	Photocatalytic (visible light)	88% at pH 8 within 120 min	Water	Jamil et al. (2017)
AFB1	CdS/WO ₃ composites	Photocatalytic (visible light)	80%	Water	Mao et al. (2019)
AFB1; AFB2	TiO ₂ , immobilized on a glass support	Photocatalytic (high-pressure UV-vis irradiation)	99% with 4 min	Peanut oil	Magzoub et al. (2019)
AFB1	TiO ₂ (P25, Degussa) layer in a closed-loop reactor	Photocatalytic (UV light)	60%	Peanut oil	Xu et al. (2019)
AFB1	Activated carbon supported TiO ₂ composites	Photocatalytic (visible light)	98% bare TiO ₂ 76%	Water	Sun et al. (2019)
DON	Dendritic α-Fe ₂ O ₃	Photocatalytic (visible light)	90%	Aqueous solution	Wang et al. (2019)

 Table 9.5
 Degradation or removal of mycotoxins

et al. (2017) reported the synthesis of chitosan coated Fe_3O_4 nanoparticles as a magnetic adsorbent for the patulin removal from fruit juice. Further, magnetic chitosan Fe_3O_4 nanoparticles coated with deactivated *Candida utilis* cells and used as a new biosorbent for patulin removal from fruit juice (Ge et al. 2017). Similarly, for patulin removal, magnetic multi-walled carbon nanotube (MWCNT) adsorbent was fabricated, which can be regenerated using NaOH solution for recycling usage up to four cycles (Zhang et al. 2019).

9.3.2 Degradation of Mycotoxins Using Nanotechnology

Recently, there has been a growing interest in photocatalytic degradation of many organic pollutants, including mycotoxins. Photocatalytic degradation has numerous advantages, such as being environmentally friendly, low–cost, and requiring mild conditions (Bhatkhande et al. 2002). Various photocatalysts, both under the UV light and visible light irradiation, are presented in Table 9.5. Nevertheless, as UV light accounts for only 4% of the total sunlight, visible light photocatalysts are more beneficial for practical degradation of organic pollutants in natural systems (Pathakoti et al. 2013, 2018).

Bai et al. (2017) reported the synthesis of graphene/ZnO hybrids using a single one-step hydrothermal method for the photodegradation of DON under UV irradiation. As shown in Scheme 1a, the proposed photodegradation mechanism is due to the formation of superoxide radicals at the conduction band. It is recognized that the graphene increases the adsorption capability and the ZnO increases the photocatalytic achievement by graphene hybridization (Fig. 9.6). Additionally, the degradation products of DON were analyzed by using ESI/MS analysis.

Recently, Mao et al. (2019) reported the photocatalytic inactivation of AFB1 at hypertoxic site C8=C9 under visible light treatment using the Z-schematic composite, clew-like WO₃ adorned with CdS nanoparticles, synthesized by the microwave supported hydrothermal and precipitation procedure (Fig. 9.7). The reaction of hydroxyl radical formation with C8=C9 bond of AF for the photocatalytic inactivation was confirmed by radical trapping experiments and ¹⁸O isotope labeling. Further, the degraded products were identified by using High Resolution Mass Spectroscopy, and the theoretical calculations using the density functional theory were used to confirm the formation of hydroxyl radicals at the C9 site, thereby forming AFB1-9-hydroxy.

Similarly, another study reported the 10 mole% Scandanium doped SrTi_{0.7}Fe_{0.3}O₃ (band gap, 1.58 eV) was synthesized for the photodegradation of afltoxinB1 under visible light irradiation (Jamil et al. 2017). Further, the degraded products were identified using gas chromatography-mass spectrometry (GC-MS), which are non-toxic to *Vibrio fischeri* organism. Hence, it is safe for water treatment, and more-over, both the catalyst and treated water can be reused for up to eight times. The mechanism of oxidation is due to rupture of short chain aliphatic alcohol, which leads to total degradation.



Fig. 9.6 Schematic drawing illustrating synthetic route and the mechanism of charge separation and adsorption-photocatalytic process over graphene/ZnO hybrid photocatalysts under UV light irradiation. Obtained with permission from Elsevier, Ref: (Bai et al. 2017)



Fig. 9.7 Illustration of the proposed mechanism over CdS/WO₃ composites for inactivating the hypertoxic site [(C8=C9) in AFB1 under visible light irradiation. Obtained with permission from Elsevier, Ref. (Mao et al. 2019)

A study by Magzoub et al. (2019) reported the photocatalytic detoxification of AFB1 and AFB2 from its real sample matrix, peanut oil, using the TiO_2 that was immobilized on a glass support under high pressure UV–vis irradiation. It is note-worthy that the TiO_2 treatment did not alter the physicochemical parameters of the
oil, such as iodine value, peroxide value, saponification value, fatty acid composition, and free fatty acids. Furthermore, the photocatalyst can be recycled up to ten cycles without reducing its effectiveness. The proposed mechanism is the formation of hydroxyl radicals and is comparable to previous reports (Jamil et al. 2017; Mao et al. 2019).

Likewise, Xu et al. (2019) reported the closed loop photocatalytic reactor comprising a glass tube holding TiO_2 catalyst for the decontamination of AFB1 from peanut oil. Based on their experimental data on AF detoxification, a theoretical model, a Weibull distribution model, has been proposed, which offers a good depiction for the photocatalytic process kinetics.

A simple hydrothermal method was described by Sun et al. (2019) for the photocatalytic degradation of AFB1 using an activated charcoal/TiO₂ composite under UV–vis irradiation. This newly synthesized composite had higher degradation efficiency when compared to the bare TiO₂ and the catalyst can be regenerated and recycled up to four cycles with 80% degradation efficiency. Formation of hydroxyl radicals plays a vital role in the degradation of AF, whereas superoxide radicals do not have an effect.

Another effective approach is the improvement of the photocatalyst, dendritic-like α -Fe₂O₃, for the DON degradation under visible light irradiation (Wang et al. 2019). A simple hydrothermal has been proposed for dendritic-like α -Fe₂O₃, which has better photocatalytic activity (more than 90% in 2 h) than the commercial α -Fe₂O₃. After degradation the intermediate products formed were identified by high performance liquid chromatography-mass spectrophotometry (HPLC-MS) and the major toxicity group, epoxy group at C12 and C13 and hydroxyl groups are destroyed in DON, thereby providing an efficient and green technology for mycotoxin decontamination.

9.4 Summary and Conclusion

The occurrence of mycotoxins in the food chain is a major safety concern around the world. Control of mycotoxins commonly depends on the suitable care taken during pre- and post-harvest conditions. The potential and present uses of nanotechnology in farming and food industries offer various significant benefits to ensure the microbial food quality and safe of nourishment items. The use of gold, QDs, and magnetic nanoparticles has significantly enhanced the biosensor technology for the mycotoxin analyses. All the above discussed nano-based sensors developed for detection and sensing of mycotoxins can be easily applied in the fields and also can be operated by unskilled personnel. Further, the sensitive and early detection of mycotoxins in food will be beneficial in eliminating these toxins to enter into food chain and thereby protects human health. Although QDs have various advantages due to their optical properties in comparison to other nanoparticles in sensing applications, but they are not cost-effective and expensive till date. In order to overcome the overestimation, false positive or underestimation, usage of more than one technique such as multiple sensor devices, which can be facile to fulfill the rapid monitoring and control of mycotoxins is essential.

Acknowledgments The author (K.P.) is thankful to the National Science Foundation (NSF/ CREST HRD-1547754) for financial support.

References

- Abdul Kadir MK, Tothill IE (2010) Development of an electrochemical immunosensor for fumonisins detection in foods. Toxins 2:382–398
- Adányi N et al (2018) Sensitivity enhancement for mycotoxin determination by optical waveguide lightmode spectroscopy using gold nanoparticles of different size and origin. Food Chem 267:10–14. https://doi.org/10.1016/j.foodchem.2018.04.089
- Alshannaq A, Yu J-H (2017) Occurrence, toxicity, and analysis of major mycotoxins in food. Int J Environ Res Public Health 14:632. https://doi.org/10.3390/ijerph14060632
- Anfossi L et al (2018) A lateral flow immunoassay for straightforward determination of fumonisin mycotoxins based on the quenching of the fluorescence of CdSe/ZnS quantum dots by gold and silver nanoparticles. Microchim Acta 185:94. https://doi.org/10.1007/s00604-017-2642-0
- Ansari SA, Husain Q (2012) Potential applications of enzymes immobilized on/in nano materials: a review. Biotechnol Adv 30:512–523. https://doi.org/10.1016/j.biotechadv.2011.09.005
- Ansari AA, Kaushik A, Solanki PR, Malhotra B (2010) Nanostructured zinc oxide platform for mycotoxin detection. Bioelectrochemistry 77:75–81
- Arévalo FJ, Granero AM, Fernández H, Raba J, Zón MA (2011) Citrinin (CIT) determination in rice samples using a micro fluidic electrochemical immunosensor. Talanta 83:966–973
- Authority EFS, Arcella D, Gergelova P, Innocenti ML, Steinkellner H (2017) Human and animal dietary exposure to T-2 and HT-2 toxin. EFSA J 15:e04972
- Bai X et al (2017) Photocatalytic degradation of deoxynivalenol using graphene/ZnO hybrids in aqueous suspension. Appl Catal B Environ 204:11–20
- Barthelmebs L, Hayat A, Limiadi AW, Marty J-L, Noguer T (2011) Electrochemical DNA aptamerbased biosensor for OTA detection, using superparamagnetic nanoparticles. Sensors Actuators B Chem 156:932–937
- Beloglazova N et al (2014) Novel multiplex fluorescent immunoassays based on quantum dot nanolabels for mycotoxins determination. Biosens Bioelectron 62:59–65
- Beloglazova NV et al (2016) Sensitive QD@ SiO2-based immunoassay for triplex determination of cereal-borne mycotoxins. Talanta 160:66–71
- Beloglazova NV, Sobolev AM, Tessier MD, Hens Z, Goryacheva IY, De Saeger S (2017) Fluorescently labelled multiplex lateral flow immunoassay based on cadmium-free quantum dots. Methods 116:141–148
- Bhat R, Rai RV, Karim AA (2010) Mycotoxins in food and feed: present status and future concerns. Compr Rev Food Sci Food Saf 9:57–81
- Bhatkhande DS, Pangarkar VG, Beenackers AACM (2002) Photocatalytic degradation for environmental applications. J Chem Technol Biotechnol 77:102–116
- Chauhan R, Singh J, Solanki PR, Manaka T, Iwamoto M, Basu T, Malhotra B (2016) Label-free piezoelectric immunosensor decorated with gold nanoparticles: kinetic analysis and biosensing application. Sensors Actuators B Chem 222:804–814
- Chehri K, Godini R (2017) Detection of nivalenol and deoxynivalenol chemotypes produced by Fusarium graminearum species complex isolated from barley in Iran using specific PCR assays. J Plant Prot Res 57:237–242

- chuan Li S, hua Chen J, Cao H, sheng Yao D, ling Liu D (2011) Amperometric biosensor for aflatoxin B1 based on aflatoxin-oxidase immobilized on multiwalled carbon nanotubes. Food Control 22:43–49
- Duan H, Chen X, Xu W, Fu J, Xiong Y, Wang A (2015) Quantum-dot submicrobead-based immunochromatographic assay for quantitative and sensitive detection of zearalenone. Talanta 132:126–131
- Duan H, Li Y, Shao Y, Huang X, Xiong Y (2019) Multicolor quantum dot nanobeads for simultaneous multiplex immunochromatographic detection of mycotoxins in maize. Sensors Actuators B Chem 291:411–417. https://doi.org/10.1016/j.snb.2019.04.101
- Erdoğan A, Ghimire D, Gürses M, Çetin B, Baran A (2018) Patulin contamination in fruit juices and its control measures. Eur J Sci Technol:39–48
- Fang G, Fan C, Liu H, Pan M, Zhu H, Wang S (2014) A novel molecularly imprinted polymer on CdSe/ZnS quantum dots for highly selective optosensing of mycotoxin zearalenone in cereal samples. RSC Adv 4:2764–2771
- Flajs D, Peraica M (2009) Toxicological properties of citrinin. Arch Ind Hyg Toxicol 60:457-464
- Foubert A, Beloglazova NV, De Saeger S (2017) Comparative study of colloidal gold and quantum dots as labels for multiplex screening tests for multi-mycotoxin detection. Anal Chim Acta 955:48–57. https://doi.org/10.1016/j.aca.2016.11.042
- Gan N, Zhou J, Xiong P, Hu F, Cao Y, Li T, Jiang Q (2013) An ultrasensitive electrochemiluminescent immunoassay for Aflatoxin M1 in milk, based on extraction by magnetic graphene and detection by antibody-labeled CdTe quantumn dots-carbon nanotubes nanocomposite. Toxins 5:865–883. https://doi.org/10.3390/toxins5050865
- Ge N, Xu J, Li F, Peng B, Pan S (2017) Immobilization of inactivated microbial cells on magnetic Fe3O4@CTS nanoparticles for constructing a new biosorbent for removal of patulin in fruit juice. Food Control 82:83–90. https://doi.org/10.1016/j.foodcont.2017.06.027
- Ghali R, Ghorbel H, Hedilli A (2009) Fumonisin determination in tunisian foods and feeds. ELISA and HPLC methods comparison. J Agric Food Chem 57:3955–3960
- Goftman VV et al (2016) Synthesis, modification, bioconjugation of silica coated fluorescent quantum dots and their application for mycotoxin detection. Biosens Bioelectron 79:476–481
- González-Sálamo J, Socas-Rodríguez B, Hernández-Borges J, Rodríguez-Delgado MÁ (2017) Core-shell poly(dopamine) magnetic nanoparticles for the extraction of estrogenic mycotoxins from milk and yogurt prior to LC–MS analysis. Food Chem 215:362–368. https://doi. org/10.1016/j.foodchem.2016.07.154
- Goud KY, Kailasa SK, Kumar V, Tsang YF, Lee SE, Gobi KV, Kim K-H (2018) Progress on nanostructured electrochemical sensors and their recognition elements for detection of mycotoxins: A review. Biosens Bioelectron 121:205–222. https://doi.org/10.1016/j.bios.2018.08.029
- Gu C, Yang L, Wang M, Zhou N, He L, Zhang Z, Du M (2019) A bimetallic (Cu-Co) Prussian Blue analogue loaded with gold nanoparticles for impedimetric aptasensing of ochratoxin a. Mikrochim Acta 186:343. https://doi.org/10.1007/s00604-019-3479-5
- Guo X (2013) Single-molecule electrical biosensors based on single-walled carbon nanotubes. Adv Mater 25:3397–3408
- Hamad AF, Han J-H, Kim B-C, Rather IA (2018) The intertwine of nanotechnology with the food industry. Saudi J Biol Sci 25:27–30. https://doi.org/10.1016/j.sjbs.2017.09.004
- Hao N, Jiang L, Qian J, Wang K (2016) Ultrasensitive electrochemical Ochratoxin A aptasensor based on CdTe quantum dots functionalized graphene/Au nanocomposites and magnetic separation. J Electroanal Chem 781:332–338
- Hendrickson O, Chertovich J, Petrakova A, Zherdev A, Dzantiev B (2016) Immunoenzyme assay of zearalenone using magnetic nanoparticles and chemiluminescent detection. Int J Appl Chem 12:613–627
- Heurich M, Kadir MKA, Tothill IE (2011) An electrochemical sensor based on carboxymethylated dextran modified gold surface for ochratoxin A analysis. Sensors Actuators B Chem 156:162–168

- Horky P, Skalickova S, Baholet D, Skladanka J (2018) Nanoparticles as a solution for eliminating the risk of mycotoxins. Nanomaterials (Basel, Switzerland) 8. https://doi.org/10.3390/ nano8090727
- Hossain MZ, Maragos CM (2018) Gold nanoparticle-enhanced multiplexed imaging surface plasmon resonance (iSPR) detection of Fusarium mycotoxins in wheat. Biosens Bioelectron 101:245–252. https://doi.org/10.1016/j.bios.2017.10.033
- Hu W et al (2014) Sensitive detection of multiple mycotoxins by SPRi with gold nanoparticles as signal amplification tags. J Colloid Interface Sci 431:71–76. https://doi.org/10.1016/j. jcis.2014.06.007
- Huang X, Zhan S, Xu H, Meng X, Xiong Y, Chen X (2016) Ultrasensitive fluorescence immunoassay for detection of ochratoxin A using catalase-mediated fluorescence quenching of CdTe QDs. Nanoscale 8:9390–9397
- Ingle AP, Gupta I, Jogee P, Rai M (2020) Chapter 2 Role of nanotechnology in the detection of mycotoxins: a smart approach. In: Rai M, Abd-Elsalam KA (eds) Nanomycotoxicology. Academic Press, pp 11–33. https://doi.org/10.1016/B978-0-12-817998-7.00002-1
- Jamil TS, Abbas H, Nasr RA, El-Kady AA, Ibrahim MI (2017) Detoxification of aflatoxin B1 using nano-sized Sc-doped SrTi0. 7Fe0. 3O3 under visible light. J Photochem Photobiol A Chem 341:127–135
- Jard G, Liboz T, Mathieu F, Guyonvarc'h A, Lebrihi A (2011) Review of mycotoxin reduction in food and feed: from prevention in the field to detoxification by adsorption or transformation. Food Addit Contam A 28:1590–1609
- Jiang W, Beloglazova NV, Luo P, Guo P, Lin G, Wang X (2017) A dual-color quantum dots encoded frit-based immunoassay for visual detection of aflatoxin M1 and pirlimycin residues in milk. J Agric Food Chem 65:1822–1828. https://doi.org/10.1021/acs.jafc.6b05337
- Joshi S (2017) Nanostructured imaging surface plasmon resonance biosensing. Wageningen University
- Jukes D (2000) The role of science in international food standards. Food Control 11:181-194
- Kalarestaghi A, Bayat M, Hashemi SJ, Razavilar V (2015) Highly sensitive FRET-based fluorescence immunoassay for detecting of aflatoxin B1 using magnetic/silica core-shell as a signal intensifier. Iran J Biotechnol 13:25
- Kaushik A, Solanki P, Pandey M, Ahmad S, Malhotra B (2009a) Cerium oxide-chitosan based nanobiocomposite for food borne mycotoxin detection. Appl Phys Lett 95:173703–173703. https://doi.org/10.1063/1.3249586
- Kaushik A, Solanki PR, Sood K, Ahmad S, Malhotra BD (2009b) Fumed silica nanoparticles– chitosan nanobiocomposite for ochratoxin-A detection. Electrochem Commun 11:1919–1923
- Khan R, Dhayal M (2008) Nanocrystalline bioactive TiO2–chitosan impedimetric immunosensor for ochratoxin-A. Electrochem Commun 10:492–495. https://doi.org/10.1016/j. elecom.2008.01.013
- Kim S, Lim HB (2015) Chemiluminescence immunoassay using magnetic nanoparticles with targeted inhibition for the determination of ochratoxin A. Talanta 140:183–188. https://doi. org/10.1016/j.talanta.2015.03.044
- Ko J, Lee C, Choo J (2015) Highly sensitive SERS-based immunoassay of aflatoxin B1 using silicaencapsulated hollow gold nanoparticles. J Hazard Mater 285:11–17. https://doi.org/10.1016/j. jhazmat.2014.11.018
- Kong D et al (2016) A gold nanoparticle-based semi-quantitative and quantitative ultrasensitive paper sensor for the detection of twenty mycotoxins. Nanoscale 8:5245–5253. https://doi.org/10.1039/C5NR09171C
- Lara S, Perez-Potti A (2018) Applications of nanomaterials for immunosensing. Biosensors 8:104
- Li W-k, H-x Z, Y-p S (2018a) Simultaneous determination of aflatoxin B1 and zearalenone by magnetic nanoparticle filled amino-modified multi-walled carbon nanotubes. Anal Methods 10:3353–3363. https://doi.org/10.1039/C8AY00815A
- Li Y, Sun M, Mao X, You Y, Gao Y, Yang J, Wu Y (2018b) Mycotoxins contaminant in kelp: a neglected dietary exposure pathway. Toxins 10:481. https://doi.org/10.3390/toxins10110481

- Li Z, Sheng W, Liu Q, Li S, Shi Y, Zhang Y, Wang S (2018c) Development of a gold nanoparticle enhanced enzyme linked immunosorbent assay based on monoclonal antibodies for the detection of fumonisin B1, B2, and B3 in maize. Anal Methods 10:3506–3513. https://doi. org/10.1039/C8AY01036F
- Liang Y, Huang X, Yu R, Zhou Y, Xiong Y (2016) Fluorescence ELISA for sensitive detection of ochratoxin A based on glucose oxidase-mediated fluorescence quenching of CdTe QDs. Anal Chim Acta 936:195–201. https://doi.org/10.1016/j.aca.2016.06.018
- Lin Y, Zhou Q, Tang D, Niessner R, Yang H, Knopp D (2016) Silver nanolabels-assisted ionexchange reaction with CdTe quantum dots mediated exciton trapping for signal-on photoelectrochemical immunoassay of mycotoxins. Anal Chem 88:7858–7866
- Liu M et al (2019) Patulin removal from apple juice using a novel cysteine-functionalized metalorganic framework adsorbent. Food Chem 270:1–9
- Lu L, Gunasekaran S (2019) Dual-channel ITO-microfluidic electrochemical immunosensor for simultaneous detection of two mycotoxins. Talanta 194:709–716. https://doi.org/10.1016/j. talanta.2018.10.091
- Lu Z, Chen X, Wang Y, Zheng X, Li CM (2015) Aptamer based fluorescence recovery assay for aflatoxin B1 using a quencher system composed of quantum dots and graphene oxide. Microchim Acta 182:571–578
- Lu T et al (2018) Fluorescence ELISA based on CAT-regulated fluorescence quenching of CdTe QDs for sensitive detection of FB1. Anal Methods 10:5797–5802. https://doi.org/10.1039/ C8AY02065E
- Luan Y, Chen J, Li C, Xie G, Fu H, Ma Z, Lu A (2015a) Highly sensitive colorimetric detection of ochratoxin A by a label-free aptamer and gold nanoparticles. Toxins 7:5377–5385
- Luan Y et al (2015b) Rapid visual detection of aflatoxin B1 by label-free aptasensor using unmodified gold nanoparticles. J Nanosci Nanotechnol 15:1357–1361. https://doi.org/10.1166/ jnn.2015.9225
- Luo Y, Zhou Z, Yue T (2017) Synthesis and characterization of nontoxic chitosan-coated Fe3O4 particles for patulin adsorption in a juice-pH simulation aqueous. Food Chem 221:317–323
- Luo L et al (2019) Monitoring zearalenone in corn flour utilizing novel self-enhanced electrochemiluminescence aptasensor based on NGQDs-NH2-Ru@SiO2 luminophore. Food Chem 292:98–105. https://doi.org/10.1016/j.foodchem.2019.04.050
- Lv X et al (2015) Electrochemiluminescence modified electrodes based on RuSi@Ru(bpy)32+ loaded with gold functioned nanoporous CO/Co3O4 for detection of mycotoxin deoxynivalenol. Biosens Bioelectron 70:28–33. https://doi.org/10.1016/j.bios.2015.03.020
- Magro M et al (2016) Citrinin mycotoxin recognition and removal by naked magnetic nanoparticles. Food Chem 203:505–512. https://doi.org/10.1016/j.foodchem.2016.01.147
- Magzoub R, Yassin A, Abdel-Rahim A, Gubartallah E, Miskam M, Saad B, Sabar S (2019) Photocatalytic detoxification of aflatoxins in Sudanese peanut oil using immobilized titanium dioxide. Food Control 95:206–214
- Mahdi M, Mansour B, Afshin M (2016) Competitive immunoassay for Ochratoxin a based on FRET from quantum dot-labeled antibody to rhodamine-coated magnetic silica nanoparticles. Microchim Acta 183:3093–3099. https://doi.org/10.1007/s00604-016-1951-z
- Majdinasab M, Sheikh-Zeinoddin M, Soleimanian-Zad S, Li P, Zhang Q, Li X, Tang X (2015) Ultrasensitive and quantitative gold nanoparticle-based immunochromatographic assay for detection of ochratoxin A in agro-products. J Chromatogr B 974:147–154. https://doi. org/10.1016/j.jchromb.2014.10.034
- Malhotra BD, Srivastava S, Ali MA, Singh C (2014) Nanomaterial-based biosensors for food toxin detection. Appl Biochem Biotechnol 174:880–896. https://doi.org/10.1007/s12010-014-0993-0
- Manubolu M, Goodla L, Pathakoti K, Malmlöf K (2018) Enzymes as direct decontaminating agents—mycotoxins. In: Enzymes in human and animal nutrition. Elsevier, pp 313–330
- Mao J et al (2019) Insights into photocatalytic inactivation mechanism of the hypertoxic site in aflatoxin B1 over clew-like WO3 decorated with CdS nanoparticles. Appl Catal B Environ 248:477–486

- Maragos C (2012) Signal amplification using colloidal gold in a biolayer interferometry-based immunosensor for the mycotoxin deoxynivalenol. Food Addit Contam A 29:1108–1117
- Molinero-Fernández Á, Jodra A, Moreno-Guzmán M, López MÁ, Escarpa A (2018) Magnetic reduced graphene oxide/nickel/platinum nanoparticles micromotors for mycotoxin analysis. Chem Eur J 24:7172–7176
- Niazi S et al (2018) A novel bioassay based on aptamer-functionalized magnetic nanoparticle for the detection of zearalenone using time resolved-fluorescence NaYF4: Ce/Tb nanoparticles as signal probe. Talanta 186:97–103. https://doi.org/10.1016/j.talanta.2018.04.013
- Organization WH (1999) The application of risk communication to food standards and safety matters: report of a joint FAO/WHO expert consultation, Rome, 2–6 February 1998. vol 70. Food & Agriculture Org
- Pal M (2017) Nanotechnology: a new approach in food packaging. J Food e Microbiol Safety Hyg 2:121
- Panini NV, Bertolino FA, Salinas E, Messina GA, Raba J (2010) Zearalenone determination in corn silage samples using an immunosensor in a continuous-flow/stopped-flow systems. Biochem Eng J 51:7–13
- Pathakoti K, Morrow S, Han C, Pelaez M, He X, Dionysiou DD, Hwang H-M (2013) Photoinactivation of *Escherichia coli* by sulfur-doped and nitrogen–fluorine-codoped TiO2 nanoparticles under solar simulated light and visible light irradiation. Environ Sci Technol 47:9988–9996
- Pathakoti K, Manubolu M, Hwang H-M (2018) Nanotechnology applications for environmental industry. In: Handbook of nanomaterials for industrial applications. Elsevier, pp 894–907
- Pathakoti K, Goodla L, Manjunath M, Hwang H-M (2019) Nanoparticles and its potential applications in the agriculture, biological therapies, food, biomedical and pharmaceutical industries: a review. https://doi.org/10.1201/9780429425660-3
- Peltomaa R, Amaro-Torres F, Carrasco S, Orellana G, Benito-Peña E, Moreno-Bondi MC (2018) Homogeneous quenching immunoassay for fumonisin B1 based on gold nanoparticles and an epitope-mimicking yellow fluorescent protein. ACS Nano 12:11333–11342
- Pfohl-Leszkowicz A, Hadjeba-Medjdoub K, Ballet N, Schrickx J, Fink-Gremmels J (2015) Assessment and characterisation of yeast-based products intended to mitigate ochratoxin exposure using in vitro and in vivo models. Food Addit Contam A 32:604–616
- Pirouz A, Selamat J, Iqbal S, Mirhosseini H, Karjiban RA, Bakar FA (2017) The use of innovative and efficient nanocomposite (magnetic graphene oxide) for the reduction on of Fusarium mycotoxins in palm kernel cake. Sci Rep 7:12453
- Pleadin J, Vulić A, Babić J, Šubarić D (2018) The incidence of T-2 and HT-2 toxins in cereals and methods of their reduction practice by the food industry. In: Fusarium – plant diseases, pathogen diversity, genetic diversity, resistance and molecular markers. IntechOpen, London, pp 41–64
- Pradhan N, Singh S, Ojha N, Shrivastava A, Barla A, Rai V, Bose S (2015) Facets of nanotechnology as seen in food processing, packaging, and preservation industry. Biomed Res Int 2015:365672–365672
- Saifuddin N, Raziah AZ, Junizah AR (2013) Carbon nanotubes: a review on structure and their interaction with proteins. J Chem 2013:18. https://doi.org/10.1155/2013/676815
- Santos Pereira C, C Cunha S, Fernandes JO (2019) Prevalent mycotoxins in animal feed: occurrence and analytical methods. Toxins 11:290. https://doi.org/10.3390/toxins11050290
- Santos VO et al (2017) Development and validation of a novel lateral flow immunoassay device for detection of aflatoxins in soy-based foods. Anal Methods 9:2715–2722. https://doi.org/10.1039/C7AY00601B
- Shao B, Ma X, Zhao S, Lv Y, Hun X, Wang H, Wang Z (2018) Nanogapped Au(core) @ Au-Ag(shell) structures coupled with Fe3O4 magnetic nanoparticles for the detection of Ochratoxin A. Anal Chim Acta 1033:165–172. https://doi.org/10.1016/j.aca.2018.05.058
- Singh P, Campidelli S, Giordani S, Bonifazi D, Bianco A, Prato M (2009) Organic functionalisation and characterisation of single-walled carbon nanotubes. Chem Soc Rev 38:2214–2230

- Smith M-C, Madec S, Coton E, Hymery N (2016) Natural co-occurrence of mycotoxins in foods and feeds and their in vitro combined toxicological effects. Toxins 8(94)
- Socas-Rodríguez B, Hernández-Borges J, Herrera-Herrera AV, Rodríguez-Delgado MÁ (2018) Multiresidue analysis of oestrogenic compounds in cow, goat, sheep and human milk using core-shell polydopamine coated magnetic nanoparticles as extraction sorbent in microdispersive solid-phase extraction followed by ultra-high-performance liquid chromatography tandem mass spectrometry. Anal Bioanal Chem 410:2031–2042
- Sonawane SK, Arya SS, LeBlanc JG, Jha N (2014) Use of nanomaterials in the detection of food contaminants. Eur J Nutr Food Safety 4:301–317
- Sun S, Zhao R, Feng S, Xie Y (2018) Colorimetric zearalenone assay based on the use of an aptamer and of gold nanoparticles with peroxidase-like activity. Microchim Acta 185:535. https://doi.org/10.1007/s00604-018-3078-x
- Sun S, Zhao R, Xie Y, Liu Y (2019) Photocatalytic degradation of aflatoxin B1 by activated carbon supported TiO2 catalyst. Food Control 100:183–188
- Sunday CE, Masikini M, Wilson L, Rassie C, Waryo T, Baker PG, Iwuoha EI (2015) Application on gold nanoparticles-dotted 4-nitrophenylazo graphene in a label-free impedimetric deoxynivalenol immunosensor. Sensors (Basel) 15:3854–3871. https://doi.org/10.3390/s150203854
- Tian J, Wei W, Wang J, Ji S, Chen G, Lu J (2018) Fluorescence resonance energy transfer aptasensor between nanoceria and graphene quantum dots for the determination of ochratoxin A. Anal Chim Acta 1000:265–272
- Tothill I (2011) Biosensors and nanomaterials and their application for mycotoxin determination. World Mycotoxin J 4:361–374
- Urusov A, Petrakova A, Vozniak M, Zherdev A, Dzantiev B (2014) Rapid immunoenzyme assay of aflatoxin B1 using magnetic nanoparticles. Sensors 14:21843–21857
- Urusov AE, Petrakova AV, Kuzmin PG, Zherdev AV, Sveshnikov PG, Shafeev GA, Dzantiev BB (2015) Application of gold nanoparticles produced by laser ablation for immunochromatographic assay labeling. Anal Biochem 491:65–71. https://doi.org/10.1016/j.ab.2015.08.031
- Vidal JC, Bonel L, Ezquerra A, Hernández S, Bertolín JR, Cubel C, Castillo JR (2013) Electrochemical affinity biosensors for detection of mycotoxins: A review. Biosens Bioelectron 49:146–158
- Wang H, Zhou X-J, Liu Y-Q, Yang H-M, Guo Q-L (2011) Simultaneous determination of chloramphenicol and aflatoxin M1 residues in milk by triple quadrupole liquid chromatography– tandem mass spectrometry. J Agric Food Chem 59:3532–3538
- Wang B et al (2016a) Aptamer induced assembly of fluorescent nitrogen-doped carbon dots on gold nanoparticles for sensitive detection of AFB1. Biosens Bioelectron 78:23–30. https://doi. org/10.1016/j.bios.2015.11.015
- Wang Q, Chen M, Zhang H, Wen W, Zhang X, Wang S (2016b) Enhanced electrochemiluminescence of RuSi nanoparticles for ultrasensitive detection of ochratoxin A by energy transfer with CdTe quantum dots. Biosens Bioelectron 79:561–567
- Wang C et al (2017) Magneto-controlled aptasensor for simultaneous electrochemical detection of dual mycotoxins in maize using metal sulfide quantum dots coated silica as labels. Biosens Bioelectron 89:802–809. https://doi.org/10.1016/j.bios.2016.10.010
- Wang Y, Lu M, Tang D (2018) Novel photoluminescence enzyme immunoassay based on supramolecular host-guest recognition using L-arginine/6-aza-2-thiothymine-stabilized gold nanocluster. Biosens Bioelectron 109:70–74. https://doi.org/10.1016/j.bios.2018.03.007
- Wang H, Mao J, Zhang Z, Zhang Q, Zhang L, Li P (2019) Photocatalytic degradation of deoxynivalenol over dendritic-like α-Fe2O3 under visible light irradiation. Toxins 11:105
- Willner MR, Vikesland PJ (2018) Nanomaterial enabled sensors for environmental contaminants. J Nanobiotechnol 16:95–95. https://doi.org/10.1186/s12951-018-0419-1
- Wu S, Duan N, Zhu C, Ma X, Wang M, Wang Z (2011) Magnetic nanobead-based immunoassay for the simultaneous detection of aflatoxin B1 and ochratoxin A using upconversion nanoparticles as multicolor labels. Biosens Bioelectron 30:35–42

- Wu L, Yan H, Li G, Xu X, Zhu L, Chen X, Wang J (2019) Surface-imprinted gold nanoparticlebased surface-enhanced raman scattering for sensitive and specific detection of patulin in food samples. Food Anal Methods:1–10
- Xiong S, Zhou Y, Huang X, Yu R, Lai W, Xiong Y (2017) Ultrasensitive direct competitive FLISA using highly luminescent quantum dot beads for tuning affinity of competing antigens to antibodies. Anal Chim Acta 972:94–101. https://doi.org/10.1016/j.aca.2017.03.039
- Xu C, Ye S, Cui X, Song X, Xie X (2019) Modelling photocatalytic detoxification of aflatoxin B1 in peanut oil on TiO2 layer in a closed-loop reactor. Biosyst Eng 180:87–95
- Yang M, Liu G, Mehedi HM, Ouyang Q, Chen Q (2017) A universal sers aptasensor based on DTNB labeled GNTs/Ag core-shell nanotriangle and CS-Fe3O4 magnetic-bead trace detection of Aflatoxin B1. Anal Chim Acta 986:122–130
- Ye Y, Zhou Y, Mo Z, Cheng W, Yang S, Wang X, Chen F (2010) Rapid detection of aflatoxin B1 on membrane by dot-immunogold filtration assay. Talanta 81:792–798
- Yu H, Park J-Y, Kwon CW, Hong S-C, Park K-M, Chang P-S (2018) An overview of nanotechnology in food science: preparative methods, practical applications, and safety. J Chem 2018:10. https://doi.org/10.1155/2018/5427978
- Zeng Y, Zhu Z, Du D, Lin Y (2016) Nanomaterial-based electrochemical biosensors for food safety. J Electroanal Chem 781:147. https://doi.org/10.1016/j.jelechem.2016.10.030
- Zhan S, Huang X, Chen R, Li J, Xiong Y (2016) Novel fluorescent ELISA for the sensitive detection of zearalenone based on H2O2-sensitive quantum dots for signal transduction. Talanta 158:51–56
- Zhang X et al (2016) A novel electrochemical immunosensor for highly sensitive detection of aflatoxin B1 in corn using single-walled carbon nanotubes/chitosan. Food Chem 192:197–202. https://doi.org/10.1016/j.foodchem.2015.06.044
- Zhang W, Han Y, Chen X, Luo X, Wang J, Yue T, Li Z (2017a) Surface molecularly imprinted polymer capped Mn-doped ZnS quantum dots as a phosphorescent nanosensor for detecting patulin in apple juice. Food Chem 232:145–154. https://doi.org/10.1016/j.foodchem.2017.03.156
- Zhang X et al (2017b) Multiplex lateral flow immunoassays based on amorphous carbon nanoparticles for detecting three fusarium mycotoxins in maize. J Agric Food Chem 65:8063–8071
- Zhang L, Dou X-W, Zhang C, Logrieco AF, Yang M-H (2018a) A review of current methods for analysis of mycotoxins in herbal medicines. Toxins 10:65. https://doi.org/10.3390/ toxins10020065
- Zhang X, Yu X, Wang J, Wang Q, Meng H, Wang Z (2018b) One-step core/multishell quantum dots-based fluoroimmunoassay for screening of deoxynivalenol in maize. Food Anal Methods 11:2569–2578. https://doi.org/10.1007/s12161-018-1198-x
- Zhang Z, Zeng C, Peng B (2019) Adsorption properties of magnetic carbon nanotubes for patulin removal from aqueous solution systems. Food Control 102:1–10
- Zhao Y, Luo Y, Li T, Song Q (2014) Au NPs driven electrochemiluminescence aptasensors for sensitive detection of fumonisin B1. RSC Adv 4:57709–57714
- Zhao Y, Yang Y, Luo Y, Yang X, Li M, Song Q (2015) Double detection of mycotoxins based on SERS labels embedded Ag@ Au core-shell nanoparticles. ACS Appl Mater Interfaces 7:21780–21786
- Zhao Y, Liu R, Sun W, Lv L, Guo Z (2018) Ochratoxin A detection platform based on signal amplification by Exonuclease III and fluorescence quenching by gold nanoparticles. Sensors Actuators B Chem 255:1640–1645. https://doi.org/10.1016/j.snb.2017.08.176
- Zhong L, Carere J, Lu Z, Lu F, Zhou T (2018) Patulin in apples and apple-based food products: the burdens and the mitigation strategies. Toxins 10:475. https://doi.org/10.3390/toxins10110475

Chapter 10 Detection of Toxic Contaminant in the Food Items



Bensu Karahalil

Contents

10.1	Introduction	214
10.2	Food Quality Assurance.	215
10.3	Classification of Food Contaminants.	216
Refer	ences	225

Abstract Food safety and quality are important issues for public health. It is essential to ensure them. Foods can be contaminated by contaminants, such as pesticides, which have not been intentionally added to it. Veterinary food contaminants can be classified into two main groups: (i) first group consists of four sub-groups (environmental contaminants, pesticide residues, veterinary drug residues, and migrants from packaging materials); (ii) second group consists of three sub-groups (mycotoxins, food processing contaminants, and chemical reactions. Food contaminants are hazardous on human health, and the effects range from minor gastric problems to major health fatalities. For example, toxic metals such as cadmium and lead, which easily enter the food chain, deplete specific nutrients in the body, which leads to declining immunological defenses, and impair psycho-social faculties, and some pesticides cause neural and kidney damage and are carcinogenic. For this reason, many techniques need to be discovered to monitor food contaminants. These techniques should be reliable and cheap and provide high-throughput measurement. Food safety organizations support and encourage using analytical tools to monitor contaminants in foods and establish limits for food contaminants, called maximum residue limits (MRLs), thus providing food standardization. Sensitive sensors are needed for monitoring and controlling food contaminants. Nanotechnology, which is emerging technology, and nanoparticles nanotechnology used have many benefits due to their unique properties. This technology provides to create or discover very sensitive sensors to control food contaminants. In this chapter, it is summarized about food contaminants and their detection and checking of them with nanosensors.

B. Karahalil (🖂)

Faculty of Pharmacy, Toxicology Department, Gazi University, Ankara, Turkey

213

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_10

Nanotechnology and sensor combination provide sensitive and reliable tools in determining and monitoring food contaminants.

Keywords Food contaminants · Metals · Pesticides · Mycotoxins · Veterinary drug residues · Nanotechnology · Food

10.1 Introduction

Food safety is an essential issue because it is directly related to the human health. It needs to improve sensitive, rapid, selective, and reliable methods to detect those food hazards due to some food safety incidents and concerns about synthetic food additives and also chemical residues. Contaminants are chemical substances which are not intentionally added to food or feed. They are present in food in different stages such as production, processing, or transport of foods (European Food Safety Authority [EFSA]). Contaminants are important problems for both animals and humans and have risk to animal and important to human health. When contamination takes places, it generally leads to a negative impact on food quality, therefore it may be a risk to human health (Australian Institute of Food Safety). EFSA carries out risk assessments on various chemicals in food and feed. This work is carried out by EFSA's panel on contaminants in the food chain. EFSA collects data and monitor activities on food contaminants to carry out risk assessment. Following the risk assessment of EFSA, EFSA gives scientific advice to European Union risk managers. Risk managers use this recommendation and decide the safety of these substances for human, animal, and plant health (EFSA).

It is necessary to ensure food safety to protect human health. Sensitive sensors are needed to detect contaminants in foods. Thanks to the conveniences and benefits that nanotechnology brings to our lives, the advantages of nanotechnology are also used in the development of sensors. Nanosensors, sensors developed using nanotechnology, have become rapid, sensitive, and reliable sensors in determining food contaminants. A wide range of nanosensors are being developed to meet the various requirements to provide food safety (Thiruvengadam et al. 2018; Panigrahi et al. 2017). Pathogenic microorganisms, toxins (such as metal toxins) which contaminate, adulterants, fertilizers, pesticides, taste makers, various toxic chemicals or chemical interaction products, vitamins, dyes, and flavor enhancers can be detected by sensors. Time, temperature, and oxygen indicators are used for monitoring food freshness. Furthermore, invisible nanobarcodes are also used for brand protection and product authenticity (Kumar et al. 2017). In this context, nanosensors created using particles with unique properties make superior and successful contributions to the improvement of food quality.

10.2 Food Quality Assurance

Effective national food control systems are imperative to protect the human health and provide the safety for domestic consumers. They are also critical in enabling countries to assure the safety and quality of their foods entering international trade and also to ensure that imported foods conform to national requirements (Food and Agriculture Organization [FAO]). Globalization of food production, concerns about food quality and safety, and increase in food standards bring a new approach when compared to conventional one. There is no more the former end-of-line product inspection (Grunert 2005 and Institute of Physics (IOP); Institute of Physics Conference). In a new inspection approach, food quality assurance is provided from each stage of food production chain to ensure safe food and to show compliance with regulatory and customer needs. New inspection approach gives insurance to the customers by controlling each step of food production (Food and Safety Quality program). For this reason, sensors with improved properties (such as low detection limits, high sensitivity and specificity, simple sample preparation, rapid detection, and miniature tools suitable for portable use) need to be developed (Omanovic-Miklicanin and Maksimovic 2016).

There are public health laboratories to detect the microbiological contamination. Identification of harmful food contaminants and adulterants is provided by carrying out routine, accurate, and reliable testing, which is the only way. Public health laboratories make collaboration with Disease Control and Prevention which is a member of PulseNet. PulseNet is a national laboratory network and monitor foodborne diseases. Without this network, it is very difficult to detect widespread foodborne disease outbreaks. Get the latest information on current foodborne outbreaks on Centers of Disease Control and Prevention's (CDC) website (Public Health Labs' Role in Food Safety) (CDC).

There have already been various methods for assessing food safety at laboratory level, such as cell culture, sensitive instrumental analysis; however, these methods have many disadvantages, including long analysis time (hours/days) and needed pretreatment steps. There are various food contaminants and standard analytical techniques are not enough to detect them. These conventional methods are insufficient to detect food contaminants (such as pesticide residues and some pathogens), and improvement of new analytical methods or techniques are needed to overcome these deficits (Li and Sheng 2014). Food safety is an important issue for public health. There are many challenges which lead to risk in food safety due to contaminants, especially chemical contaminants are a major concern. Regulatory agencies responsible for food safety have established strict control programs to avoid contamination through the food chain (Food Regulation and Safety). Thus, we need sensors with a rapid, sensitive, high-throughput, and fast-operating system to be able to measure many samples. Nanotechnology-based method, called nanosensor, is one of the effective detection methods to be used. Nanotechnology is a promising new field and uses nano-sized materials which have unique properties to provide rapid, sensitive products having advantages (Christian et al. 2008). This technology

provides new sensors (detection of external and internal conditions in food packaging, carbon nanotubes based electrochemical sensors for detection of cations, anions, and organic compounds in food, various aptamers for detection of pesticides, antibiotics, heavy metals, microbial cells, and toxins) to provide food safety and thereby improving food functionality and quality. Nanosensors have advantage to detect food components in an easy and quick manner compared to conventional methods.

10.3 Classification of Food Contaminants

Scientists or experts in food field classify food contaminants in different manners. For example, Mastovska classified food contaminants as environmental contaminants, food processing contaminants, unapproved adulterants and food additives, and migrants from packaging materials (Mastovska 2013). Other classification for food contaminants which are commonly used is as physical, chemical, biological, and radiological contaminants. The most common physical contaminants in food are metals and plastic, ceramic, and glass pieces which may occur. Physical and radiological contaminants are not commonly encountered and they their hazard potential is more weak. Therefore, they do not lead to public concern. Therefore, these topics will not be included in this chapter. If biological contaminants occur in food, they may affect human health, either by infection or intoxication by pathogens such as bacteria, viruses, parasites, and prions and cause fever, headache, nausea, vomiting, abdominal pain, and diarrhea. Chemical contaminants include mycotoxins, phycotoxins, pesticides residues, fertilizers, antibiotics and veterinary drug residue, allergens, adulterants in food, and heavy metals (physical contaminants). These contaminants are discussed in detail below.

Different types of classification on food contaminants are present; however, generally they are similar and they contain same type of contaminants, of course, only the presentation of classification type of them is different. My classification is also a little different from others. Food/chemical contaminants on food items, especially organic chlorine substances and agrochemicals like pesticides, a large part of chemical contaminants, have been introduced into our life in the last 70–80 years. Other chemical contaminants have been in the food items for thousands of years; however, we are aware of them after analytical methods were developed. Before the 1950s it was measured only at mg levels; nowadays detection limits of chemicals have increased, and even ppt (part per trillion) and ppq (part per quadrillion) levels are measured (Karakaya 2018).

There are more than 1000 food contaminants in different. Food contaminants are divided into two main categories (Fig. 10.1).

A. Food contaminants that originate from developments in the chemical industry on foods due to rapid industrialization in the twentieth century. This category also consists of four sub-groups:



Fig. 10.1 Classification of food contaminants

- 1. Environmental contaminants
- 2. Pesticides residues
- 3. Veterinary drug residues
- 4. Migrants from packaging materials
- B. Food contaminants we have been exposed to for thousands of years but we are aware of them for the last 50–60 years:
 - 1. Mycotoxins (aflatoxins, patulin, ochratoxins)
 - 2. Food processing contaminants (polycyclic aromatic hydrocarbons, pyrolysis products, acrylamide)
 - 3. Chemical reactions (N-nitroso substances)
- 1. Environmental contaminants

a. Organochlorine substances, which are in the first place on the Fig. 10.1, are no longer used today. They are chemicals of global concern due to their persistence in the environment, ability to bio-magnify, and bio-accumulate in the environment. Persistent organic pollutants (POPs) are thus found in organisms at the top of the food chain since they biomagnify throughout the food chain and bioaccumulate in organisms. Consequently, background levels of POPs can be found in the human body (Rather et al. 2017; Goerke et al. 2004; Guo et al. 2019).

Humans are exposed to these chemicals in a variety of ways: mainly through food, air, and at the workplace. The most commonly encountered persistent organic pollutants are organochlorine pesticides (OCP), such as dichlorodiphenyltrichloroethane (DDT), aldrin, lindane, dieldrin, endrin, chlorine, toxaphene, mirex, heptachlor, hexachlorobenzene, industrial chemicals, most notably polychlorinated biphenyls (PCB), as well as unintentional by-products of many industrial processes, especially polychlorinated dibenzo-p-dioxins (PCDD) and dibenzofurans (PCDF), commonly known as "dioxins."

Since 1976, the World Health Organization (WHO) through its the Global Environment Monitoring System ; GEMS/Food Programme, which is the Global Environment Monitoring System—Food Contamination Monitoring and Assessment Programme, commonly known as GEMS/Food, informs governments and has collected and evaluated information on levels of persistent organic pollutants in foods, including human milk. Three international studies were made to assess the levels and trends of polychlorinated dibenzodioxins, polychlorinated dibenzofurans, and dioxin-like polychlorinated biphenyls on human milk, which is an ideal matrix to generally monitor levels of persistent organic pollutants in the environment. Biomonitoring of human milk data can provide information on the exposure of the mother as well as the infants. This information provides guidance on the need for measures to reduce levels of these substances in food (WHO). Milk is a very important food due to its content of fat, protein, vitamins, and minerals. However, the use of organochlorine pesticides (OCP) for decades has also contaminated the milk since they have contaminated the soil, water, and air. Meanwhile, while mentioning milk, sensitive sensors are used in the determination of melamine used as fertilizer. Melamine, which is a fertilizer and used as adulterant in protein-rich products in infant formulas and cow milk, is detected by gold nanoparticles-based fluorescence sensors. These sensors are very sensitive and are able to detect at picomolar levels of melamine (Vasimalai and John 2013). Although organochlorinated pesticides have been banned since the 1980s, these compounds or their metabolites have still been detected in many samples and even above the maximum allowed levels by the World Health Organization. Milk and other dairy based products are highly consumed worldwide, especially by young children. Various electrochemical sensors have been developed. They assess milk quality for melamine, antibiotics, bisphenol A, DNA bases, and hydrogen peroxide. Hydrogen peroxide activates the inherent lactoperoxidase enzyme system and help to improve the quality of raw dairy products. However, possible adverse health effects from over exposure take place. Hydrogen peroxide is 80 ppm according to Food and Drug Administration. Eggshell membranes (ESMs) have been used as a platform for supporting graphene oxide (GO) for hydrogen peroxide sensitive electrodes. The versatility of electrochemical nanosensors are preferred because of their versatility and especially are used for detecting compounds like sulfite in sugar, grapes, wine and water, staphylococcal enterotoxin B (a possible source of food poisoning), aflatoxin, and durabolin (Baryy and O'Riordan 2016; Mittal et al. 2016).

b. Metals

In the early nineteenth century, foods were stored in glass jars, but it was difficult to use or handle in battlefields because it was easily broken and metal cans/ containers had to be developed (Badilla et al. 2012). Nowadays, metal containers are widely used in packaging besides glass jars. Most people prefer glass jar because they think that they are safer than metallic cans, so they prefer to use them; and they consider that metal migration takes place from can to food content and it is hazardous to human health. Metallic cans are cheaper than glass ones; for these reasons, in some regions, the preference, whether glass or metal, changes according to the socio-economic status of people. Of course, the migration of metal ions from cans is also an issue that should not be ignored. Heavy metals (lead, arsenic, cadmium, mercury, chromium) are hazardous elements. Lead causes memory problems, abdominal pain, constipation, headaches, irritability, inability to have children, and tingling in the hands and feet (Wani et al. 2015). It is reported that Arsenic causes multisystem diseases and encephalopathy and peripheral neuropathy (Ratnaike 2003). Cadmium causes damage to liver and kidney (Satarug 2018). Mercury is neurotoxic (Alexeandrov et al. 2018). When corrosion takes place on metallic cans, metallic ions migrate to food and thus food contamination occurs. To avoid this, the inner side of cans is coated to save from corrosion. Coating materials are epoxy resins. Still in some minor by-products the epoxy resins (cyclo-di-BADGE, bisphenol A, or bisphenol A diglycidyl ether) can migrate to food. These by-products are endocrine disrupters which mimic to estrogens or testosterone or other hormones and change hormonal balance of the body and are hazardous to human health. These metals have been extensively studied and their effects on human health regularly reviewed by international bodies such as the WHO. They can damage and alter the functioning of organs such as the brain, kidney, lungs, liver, and blood (Rather et al. 2017). Heavy metal toxicity can either be acute or chronic effects. Exposure to these heavy metals might be from food, air, or water pollution, as well as medicine, food containers with improper coating, industrial exposure, or lead-based paint. Therefore, sensitive and selective determination of toxic heavy metals with costeffective and convenient procedures is required. For detection of toxic metal ions such as cadmium ions in foods, multi-walled carbon nanotubes-silica nanocompositebased potentiometer sensors are used (Bagheri et al. 2013).

Electrochemical sensors are used for detection of heavy metal ions for food samples. For many years, anodic stripping voltammetry (ASV) at the mercury and its modified electrode was commonly used to determine the trace metal ions for the extensive cathodic potential range. However, the improper use of the mercury-containing device (disposal of that devise, incorrect handling, etc.) leaks mercury vapors. It causes health problems for humans and also not good for ecosystem. Therefore, various mercury-free electrodes were developed such as a nanostructured bismuth film electrode (nsBiFE). This nanosensor is used for ASV detection of multiple heavy metals (detection limits of 0.4μ gL⁻¹ and 0.1μ gL⁻¹ for Cd2+ and Pb2, respectively). Bismuth and antimony are used similarly for similar purpose. More and more electrochemical sensors based on screen-printed carbon electrode (SPCE) have been fabricated for detection of heavy metals in food safety (Baryy and O'Riordan 2016).

2. Pesticide Residues

Pesticides are widely used to control pests that would otherwise destroy or reduce food. In this area of agriculture, pesticides, especially organophosphate and carbamate, are commonly used worldwide. They are highly toxic since they inhibit acetylcholinesterase which is key enzyme for the nervous system. This inhibition causes respiratory failure, weakness in muscles, miosis, unconsciousness, convulsion, and finally death. For these reasons, pesticide contamination in food is very important issue. It is essential to detect the contamination to protect human health (Judge et al. 2016).

The development of simple, sensitive, user-friendly, and cost-effective sensors using nanotechnology is required. Biosensors have more advantages than the existing techniques such as high-performance liquid chromatography or gas chromatography, because biosensor can provide rapid, sensitive, simple, and low-cost on-field detection. Biosensors (optical, electrochemical, and piezoelectric) have been used and are a powerful alternative for the detection and monitoring of pesticide residues. Biosensors are based on the intimate contact between a biological element (such as receptor, tissue cell, nucleic acid, enzyme, antibody) and a transducer system. The biological component interacts specifically with the target analyte while the transducer transforms the signal. After interaction target analyte to biological element cause a signal thereby, measured and quantified. For organophosphate and carbamate pesticide residues, biosensors based on acetylcholinesterase inhibition form a link with the active site of the enzyme. The electrochemical impedance spectroscopy (EIS) is transduction technology, which detects direct analyte by studying the electrical properties of the sensing device interface. The new label-free impedimetric affinity sensors are also developed based on AChE and detection limit is 2,5 and 10 ug/kg for organophosphate and carbamate (Sciencetrends.com). The detection of some pesticides, such as pesticides in organophosphorus and carbamate groups, in foods is provided by using colorimetric and fluorometric sensors, which are composed of gold (Liu et al. 2012).

3. Veterinary Drug Residues

Antibacterial drugs and hormonal growth promoters are the main veterinary drug residues that potentially contaminate foods of animal origin. Consumption of these residues in animal products may cause the development of antibiotic resistance and hypersensitivity reaction. Veterinary drug products and agricultural chemicals used according to label directions should not result in residues at slaughter. However, in some cases, veterinary drug residues are encountered. These situationsare not to follow the recommended label directions or dosage (extra-label usage) not to properly clean equipment and not to prevent animals from reaching the spilled drugs, dosage/measurement errors, the use of contaminated drug due to drug interactions when it is used more than two drugs and to use the improper use of pesticides, etc. The presence of veterinary drug residues in foods is one of the key issues for food safety, and it is a great health concern. It is essential to develop sensor for detecting veterinary drug residues in foods and also needs to be quantified these residues which are less than the maximum residue limits (MRL) on the basis of food safety.

Detection of veterinary residues in meat and dairy products ensures their quality and safety. Using bioreceptors from biological organisms or receptors, biosensors have been employed as a new mean of analytical and chemical analysis. The most frequently used biosensors are those based on antibody/antigen affinity pairs for detection of veterinary drug residues. Electrochemical biosensors composed of nanomaterials have better performance due to their unique electrical and chemical properties. The incorporation of nanomaterials can potentially increase the response speed, selectivity and sensitivity. Biosensors using an aptamer as a recognition element are also commonly used for the detection of antibiotic residues. Electrochemical can be categorized as potentiometric, amperometric, voltammetric, impedimetric and conductivity sensors. An amperometric biosensor measures the current produced, potentiometric devices measure the changes in pH and ion concentration and conductimetric biosensors which is a limited response for the detection of antibiotics due to its poor signal/noise ratio, quantify the electrical conductivity changes in a solution at constant voltage (Majdinasab et al. 2017; Verdian 2018). Enzymatic biosensors are commonly used for the analysis of herbicides contaminants. Enzymatic biosensor for penicillin, cellular biosensors for antibiotic residues such as tetracyclines, beta-lactam antibiotics, quinolones, chloramphenicol and quinolones, and transducer-biosensors for antibiotic residues are developed (Falowo and Akimoladun 2019).

4. Migrants from Packaging Materials

Polymer packaging with chemicals are called plastics. These chemicals improve functional properties of plastics. However, these substances might be concern on product safety and quality when amount of these migrating compounds in food exceed their specified limits. Migration means the diffusion of substances from the food-contact layer to the food surface. This process is influenced by food-packaging interactions and temperature (Bhunia et al. 2013). The five polymers most commonly applied in plastic packaging include polyethylene (PE), polypropylene (PP), polyethylene terephthal-ate (PET), polystyrene (PS), and PVC. Manyo ther polymers, including, for example, polycarbonate (PC), polyamides, (PA, nylon), acrylics, polylactic acid (PLA), polyurethanes (PU) and even more specialized polymer types, are also used for specific packaging applications (Groh et al. 2019; Omeroğlu et al. 2017). Phthalates are esters of phthalic acid and are used as plasticizers to increase the flexibility, transparency, durability, and longevity of plastics. Phthalates are used in a large variety of applications, from enteric coatings of pharmaceutical pills to food packaging materials. These compounds are easily released into the environment since they are not covalently bound to the plastics. Phthalates can cause many health problems such as reproductive abnormalities, low birth weight, asthma, endocrine disruption, cancer, autism (Street et al. 2018). Sensitive and reliable methods to rapidly detect the presence of phthalates in food are clearly needed. The selectivity of the sensor is important for complex organic matrices. Phthalates are detected in detection system in complex organic matrices like soups, milk, juice and drinks. Molecular imprinting technique has been used for this purpose and develop sensor (Asif et al. 2015). Gas Chromatography-Mass Spectrometry (GC/ MS) and Liquid Chromatography-Mass spectrometry (LC/MS) are currently used

and enable to provide sensitive detection of phthalate compounds (Agilent Technologies). Phthalates are known endocrine disrupters which are structurally diverse compounds that interfere with the endocrine system, cause a broad range of adverse effects. Nanosensors developed for endocrine disruptors can also be used for phthalates. A high throughput cell-based nanosensor provides highly sensitive identification of estrogenic agents.

- A. Food contaminants that originate from developments in the chemical industry on foods due to rapid industrialization in the twentieth century
- B. Food contaminants that we have been exposed to for thousands of years, but we aware of them for the last 50–60 years
- 1. Mycotoxins

Mycotoxins are naturally occurring toxins produced by certain molds. Aflatoxins are produced by certain kinds of fungi (Aspergillus: *Aspergillus flavus*, *A parasiticus*, and *A. nomius*) that are found naturally all over the world; they can contaminate food crops and pose a serious health threat to humans and livestock. Long-term or chronic exposure to aflatoxins has several health problems including carcinogenicity, mutagenic in bacteria, genotoxic and causing birth defects, chronic diarrhea, infectious diseases, malnutrition (WHO).

Generally, GC-MS and LC-MS techniques are used for detection of mycotoxins in foods as contaminants due to their selectivity and sensitivity; however, these techniques are time-consuming and not enough rapid assay and also pretreatment steps are needed before detection. Inconsistent results and poor sensitivity are from sample preparation and extraction steps (Daly et al. 2000). Surface plasmon resonance (SPR) has advantages compared to these analytical techniques. SPR is used for molecular interactions. Furthermore, it is also used to elucidate biospecific interaction analysis. The feature of SPR, which makes use of molecular interactions, is adapted for various sensing applications. There is a biorecognition layer on sensor. The action mechanism of SPR is based on to detect the changes in the refractive index of the biorecognition layer. Furthermore, SPR monitors the binding interactions of immuno-components in real time (Wangkam et al. 2009; Choi et al. 2009).

Immunoassays and biosensor are preferred to these conventional techniques for detection of mycotoxins such as aflatoxins due to their advantages. Biosensors based on the use of monoclonal or polyclonal antibodies are used in the mycotoxin's analyses. The immunosensor based on antibody-attached gold nanoparticles provide rapid and effective detection for aflatoxin and detection range of that method for aflatoxin is 10–100 ng/d Land response time is very short (60 s) (Torres-Pacheco 2011). Systems (dip-stick kits, hyperspectral imaging, electronic noses, molecularly imprinted polymers, and aptamer-based biosensors are used for detection of aflatox-ins (WHO).

The immunosensor based on competitive fluorescence immunoassay is used for ochratoxin and also aflatoxin B1. The immunosensor is used antigen-modified magnetic nanoparticles as immunosensing probes and antibody-functionalized rareearth-doped sodium yttrium fluoride up conversion nanoparticles as multicolor signal probes for detection of ochratoxin. Ochratoxin also can detect an immunosensor method based on nanostructured zinc-oxide film, which is deposited onto indium-tin-oxide glass plate for comobilization of rabbit immunoglobulin antibodies and bovine serum albumin. The action mechanism of this method is to bind positively charged nano-zinc oxide with carboxyl groups of r-IgG by electrostatic interaction. Therefore, free amino- terminal sites of r-IgG bind to the carboxylic group of ochratoxin molecules. Aptamers, which use single-walled carbon nanotubes, are also used for detection of ochratoxin as a fluorescent assay (Kuswandi et al. 2017).

2. Food Processing Contaminants

Acrylamide naturally forms in starchy food products during high-temperature cooking, including frying, baking, roasting and also industrial processing, at +120 °C and low moisture. Maillard Reaction is the main chemical process is the same reaction that 'browns' food. Acrylamide forms from sugars and amino acids (mainly one called asparagine) that are naturally present in many foods such as potato crisps, French fries, bread, biscuits, and coffee (EFSA). Acrylamide can create adducts with hemoglobin which form the reaction between the -NH2 group of N-terminal valine of Hb with acrylamide. Voltammetric sensor based on the reaction of Hb with acrylamide is used for detection. Electrochemical biosensor is also designated for detection. The reversible conversion of Fe³ to Fe² of heme is required for its electroactivity. Electromediators such as carbon nanotubes and gold nanoparticles provide intensification of electron transfer rate between Hb and electrode surface (Krajewska et al. 2009). Fluorescent sensors for detection of acrylamide is preferred to conventional analytical methods (GC-MS and LCMSMS) since fluorescent sensors are rapid and specific to analytes.

Polycyclic aromatic hydrocarbons (PAHs) are a large group of organic compounds with two or more fused aromatic rings. Polycyclic aromatic hydrocarbons. Polycyclic aromatic hydrocarbons (PAHs) are a group of contaminants produced by burning of carbon-based materials. They can get into food either from the environment or during food processing. After the first regulatory limits in food were set in 2006, the European Food Safety Authority (EFSA) published a Scientific Opinion and stated that 16 PAHs in food are a possible health concern. Some PAHs are carcinogenic because they damage DNA. It is therefore important to develop methods which are rapid, reliable, and sensitive for PAHs in food to help reduce their deleterious impacts on human health based on the accurate quantification. A conductometric sensor based on screen-printed interdigital gold electrodes are used for PAH detection. These electrodes are on glass and this glass is coated with molecularly imprinted polyurethane layers. Screen-printed interdigital electrodes are very suitable transducers. The working principle is based on the measurement of changes in resistance of PAH-imprinted layers. The sensor has good selectivity and high sensitivity, for example, detection limit is 1.3 nmol/L for anthracene in water (Latif et al. 2018).

3. Chemical Reactions

N-nitroso compounds (NOCs) include two chemical classes, nitrosamines and nitrosamides, which are formed by the reaction of amines and amides, respectively, with nitrosating agents derived from nitrite. Endogenous N-nitroso compound formation is estimated to account for 45-75% of total exposure to N-nitroso compounds. Processed meat is a major dietary source of nitrite, amines, and amides (Dubrow et al. 2010). Nitrosamides do not require metabolic activation and directly make alkylation. There are many ways of exposure, and we could be exposed to them through eating foods containing these compounds, including fish, cheeses, bacon, and other cured meats; breathing air containing tobacco smoke or the air inside cars; and drinking some alcoholic beverages like beer and other malt beverages (https://dhss.delaware.gov). The N-nitroso compounds studied by the Environmental Health Protection (EPA) are classified as probable human carcinogens. For the detection of N-nitroso compounds, the commonly used analytical techniques include gas chromatography-mass spectrometry or tandem mass spectrometry and liquid chromatography-tandem mass spectrometry. They are timeconsuming techniques. Rapid and efficient sensors are tried to develop for N-nitroso compounds.

Food safety is an essential public health issue for all countries. We cannot think of food quality separately from food safety. Food quality is also an extremely important issue. There are many factors or agents which affect food safety and food quality. Sustainable food safety is a key element, for this reason detection or quantification of factors affecting food safety and food quality are of major issue. Chemical, biological, and sensors integrated with nanotechnology are used for detection of contaminants. However, as mentioned below, nanosensors are preferred to conventional techniques due to their valuable advantageous. Nanotechnology is relatively new technology. It is providing new products in all industrial sectors (electricity, cosmetics, pharmaceutical industry, etc.) widely all over the World. Products produced by nanotechnology have many advantages since it uses nanoparticles and nanomaterials. Nanoparticles or nanomaterials possess unique physical and chemical properties due to their high surface area and nanoscale size. Their optical properties are dependent on the size, which imparts different colors due to absorption in the visible region. Nanosensors, whether used as online or off-line, can be integrated with wireless technology and used for real-time transmission of contaminant alarm or test results to remote servers, providing rapid screening and reporting. Due to perfect integration of nanotechnology with chemical and biological items, nanosensors provide tremendous benefits to detect various food contaminants. It, therefore, provides superior success in food safety and food quality. Sensors developed using nanotechnology are used both on-line and off-line. In both situations, they are integrated with wireless technology and this provides real-time transmission of contaminant alarm or test results to remote servers. All these above properties provide rapid screening and reporting. Of course, there is a need to develop more sensors in future to control the safety of foods.

References

- Alexeandrov PN, Pogue A, Lukiw WJ (2018) Synergism in aluminum and mercury neurotoxicity. Integr Food Nutr Metab 5(3):1–13. https://doi.org/10.15761/IFNM.1000214
- Asif I, Zia AI, Afsarimanesh N, Xie L, Nag A, Al-Bahadly IH, Yu PL, Kosel J (2015) Improved detection limits for phthalates by selective solid-phase micro-extraction. 2015 Ninth International Conference on Sensing Technology. Online available at https://ieeexplore.ieee. org/stamp/stamp.jsp?tp=&arnumber=7438493. Accessed Jan 2020
- Australian Institute of Food safety. Available online at https://www.foodsafety.com.au/resources/ articles/food-safety-and-the-different-types-of-food-contamination. Accessed Jan 2020
- Badilla GL, Salas BV, Wiener MS (2012) Micro and nano corrosion in steel cans used in the seafood industry. In: Scientific, health and social aspects of the food industry. InTech, pp 129–141. Online available at at: https://www.researchgate.net/publication/221923595. Accessed Jan 2020. https://doi.org/10.5772/37411
- Bagheri H, Afkhami A, Shirzadmehr A, Khoshsafar H, Khoshsafar H, Ghaedi H (2013) Novel potentiometric sensor for the determination of Cd2Y based on a new nano-composite. Int J Environ Anal Chem 93:578–591. https://doi.org/10.1080/03067319.2011.649741
- Baryy S, Riordan AO (2016) Electrochemical nanosensors: advances and applications. Rep Electrochem 6:1–14. https://doi.org/10.2147/RIE.S80550
- Bhunia K, Sablani S, Tang J, Rasco B (2013) Migration of chemical compounds from packaging polymers during microwave, conventional heat treatment, and storage. Compr Rev Food Sci Food Saf 12:523–545. https://doi.org/10.1111/1541-4337.12028
- Centers of Disease Control and Prevention (CDC). Available online at https://www.cdc.gov/ncezid/what-we-do/partnership-in-action/innovation-amd.html. Accessed Jan 2020
- Chen J, Suang F. Rapid, sensitive, and robust detection of phthalates in food using GC/MS or LC/ MS. Application note. Agilent Technologies. Available online at https://www.agilent.com/cs/ library/applications/5990-9510EN.pdf. Accessed January 2020
- Choi SW, Chang HJ, Lee N, Kim JH, Chun HS (2009) Detection of mycoestrogen zearalenone by a molecularly imprinted polypyrrole-based surface plasmon resonance (SPR) sensor. J Agric Food Chem 57(4):1113–1118
- Christian P, Von der Kammer F, Baalousha M, Hofmann T (2008) Nanoparticles: structure, properties, preparation and behaviour in environmental media. Ecotoxicology 17:326–343. https:// doi.org/10.1007/s10646-008-0213-1
- Daly SJ, Keating GJ, Dillon PP, Manning BM, O'Kennedy R, Lee HA, Morgan MRA (2000) Development of surface plasmon resonance-based immunoassay for aflatoxin B-1. J Agric Food Chem 48(11):5097–5104
- Dubrow R, Darefsky AM, Park Y, Mayne ST, Moore SC, Kilfoy B, Cross AJ, Sinha R, Hollenbeck AR, Schatzkin A, Ward MH (2010) Dietary components related to N-nitroso compound formation: a prospective study of adult glioma. Cancer Epidemiol Biomark Prev 19:1709–1722. https://doi.org/10.1158/1055-9965
- Environmental Protection Agency, EPA. Available online at https://www.epa.gov/sites/production/ files/2015-09/documents/method_607_1984.pdf. Accessed Jan 2020
- European Food Safety Agency (EFSA). Available online at https://www.efsa.europa.eu/en/topics/ topic/acrylamide. Accessed Jan 2020
- Falowo AB, Akimoladun OF (2019) Veterinary drug residues in meat and meat products: occurrence, detection and implications. Vet Pharm IntechOpen. https://doi.org/10.5772/. intechopen.83616. Available online at https://www.intechopen.com/online-first/veterinary-drug-residues-in-meat-and-meat-products-occurrence-detection-and-implications. Accessed Jan 2020
- Food and Agriculture Organisation of the United Nations (FAO), 5. Strengthening national food control systems. Available at http://www.fao.org/3/y8705e/y8705e05.htm. Accessed Jan 2020
- Food and Quality program. Improving food safety and quality along the chain. Online available at http://www.fao.org/3/a-i2797e.pdf. Accessed Jan 2020

- Food Regulation and Safety. Available online at https://www.nasda.org/policy/nasda-policystatements/food-regulation-and-safety. Accessed Jan 2020
- Food Safety and the Different Types of Food Contamination. Available at https://www.foodsafety.com.au/resources/articles/food-safety-and-the-different-types-of-food-contamination. Accessed Jan 2020
- Goerke H, Weber K, Bornemann H, Ramdohr S, Plötz J (2004) Increasing levels and biomagnification of persistent organic pollutants (POPs) in Antarctic biota. Pollut Bull 48(3–4):295–302
- Groh KJ, Backhaus T, Carney-Almroth B, Geueke B, Inostroza PA, Lennquist A, Leslie HA, Maffini M, Slunge D, Trasande L, Warhurst AM, Muncke J (2019) Overview of known plastic packaging-associated chemicals and their hazards. Sci Total Environ 651(Pt 2):3253–3268. https://doi.org/10.1016/j.scitotenv.2018.10.015
- Grunert KG (2005) Food quality and safety: consumer perception and demand. Eur Rev Agric Econ 32:369–391. https://doi.org/10.1093/eurrag/jbi011
- Guo W, Pan B, Sakkiah S, Yavas G, Ge W, Zou W, Tong W, Hong H (2019) Persistent organic pollutants in food: contamination sources, health effects and detection methods. Int J Environ Res Public Health 16(22):pii: E4361. https://doi.org/10.3390/ijerph16224361. Review
- IOP Conference Series (2017) Food quality assurance role of food quality assurance and product certification systems on marketing aspectt. Earth Environ Sci 85(1):012011. https://doi. org/10.1088/1755-1315/85/1/012011
- Judge SJ, Savy CY, Campbell M, Dodds R, Gomes LK, Laws G, Watson A, Blain PG, Morris CM, Gartside SE (2016) Mechanism for the acute effects of organophosphate pesticides on the adult 5-HT system. Chem Biol Interact 245:82–89. https://doi.org/10.1016/j.cbi.2015.12.014
- Karakaya AEK (2018) Kimyasaldan gidaya doz ve risk (From chemical to food dosage and risk). ISBN: 978-605-245-083-3, Artek Reklamcılık (Artek Advertising), pp 1–100
- Krajewska A, Radecki J, Radecka H (2009) Sensors and biosensors for determination of acrylamide and acrylic acid in potato food products. Food 3 (Special issue 2):113–120. Available online at http://www.globalsciencebooks.info/Online/GSBOnline/images/0906/FOOD_3(SI2)/ FOOD_3(SI2)113-1200.pdf. Accessed Jan 2020
- Kumar V, Guleria P, Mehta SK (2017) Nanosensors for food quality and safety assessment. Environ Chem Lett 15:165–177. https://doi.org/10.1007/s10311-017-0616-4
- Kuswandi B, Futra D, Heng LY (2017) Nanosensors for the detection of food contaminants:307–333.
 ISBN (electronic): 9780128119433. https://doi.org/10.1016/B978-0-12-811942-6.00015-7. Available online at https://ukm.pure.elsevier.com/en/publications/nanosensors-for-the-detection-of-food-contaminants. Accessed Jan 2020
- Latif U, Ping L, Dickert FL (2018) Conductometric sensor for PAH detection with molecularly imprinted polymer as recognition layer. Sensors (Basel) 18:767
- Li Z, Sheng C (2014) Nanosensors for food safety. J Nanosci Nanotechnol J14:905-912
- Liu D, Chen W, Wei J, Li X, Wang Z, Jiang X (2012) A highly sensitive, dual-readout assay based on gold nanoparticles for organophosphorus and carbamate pesticides. Anal Chem 84:4185–4191. https://doi.org/10.1021/ac300545p
- Majdinasab M, Yaqup M, Rahim A, Catanante G, Hayat A, . Marty JL (2017) An overview on recent progress in electrochemical biosensors for antimicrobial drug residues in animal-derived food. Sensors 17: 1947. https://doi.org/10.3390/s17091947
- Mastovska K (2013) Modern analysis of chemical contaminants in food. Food Saf Mag. Available online at http://www.foodsafetymagazine.com/magazine-archive1/februarymarch-2013/ modern-analysis-of-chemical-contaminants-in-food/. Accessed Jan 2020
- Mittal A, Teotia M, Soni RK, Mittal J (2016) Applications of egg shell and egg shell membrane as adsorbents: a review. J Mol Liq 223:376–387. https://doi.org/10.1093/ps/82.3.510
- N-Nitroso compounds. Available online at https://dhss.delaware.gov/dhss/dph/files/nnitrosofaq. pdf. Accessed Jan 2020
- Omanovic-Miklicanin E, Maksimovic M (2016) Nanosensors applications in agriculture and food industry. Bull Chem Tech Bosnia Herzegovina, special issue:59–70. Print ISSN: 0367-4444,

Online ISSN: 2232-72662016. Available at http://www.pmf.unsa.ba/hemija/glasnik/files/ Issue%2047/5-59-70-Omanovic.pdf

- Omeroğlu PY, Özdal T, Bulut R (2017) Chemical migration from plastic types of food. Eurasian J Food Sci Tech:22–32. Online available at https://dergipark.org.tr/tr/download/article-file/486886. Accessed Jan 2020
- Panigrahi SS, Sarkar P, Roy E, Banerjee P (2017) Nanosensors in food safety. In: Portable biosensors and point-of-care systems, pp 183–207. Book DOI: https://doi.org/10.1049/ PBHE003E. Chapter DOI: https://doi.org/10.1049/PBHE003E_ch10. e-ISBN: 9781849199636
- Public Health Labs' Role in Food Safety. Available online at https://www.aphl.org/programs/food_ safety/Pages/Public-Health-Labs-Role.aspx. Accessed Jan 2020
- Rapid detection of pesticide residues in food. Available online at https://sciencetrends.com/rapiddetection-pesticide-residues-food/. Accessed Jan 2020
- Rather IA, Koh WY, Paek WK, Lim J (2017) The sources of chemical contaminants in food and their health implications. Front Pharmacol 8:830. https://doi.org/10.3389/fphar.2017.00830
- Ratnaike RN (2003) Acute and chronic arsenic toxicity. Postgrad Med 79(933):391–396. https:// doi.org/10.1136/pmj.79.933.391
- Satarug S (2018) Dietary cadmium intake and its effects on kidneys. MDPI Toxics 6:15. https:// doi.org/10.3390/toxics6010015. Online available at www.mdpi.com/journal/toxics. Accessed Jan 2020
- Street ME, Angelini S, Bernasconi S, Burgio E, Cassio A, Catellani C, Cirillo C, Deodati et al (2018) Current knowledge on endocrine disrupting chemicals (EDCs) from animal biology to humans, from pregnancy to adulthood: highlights from a National Italian Meeting. Int J Mol Sci 19:167. https://doi.org/10.3390/ijms19061647. Online available at https://www.mdpi.com/ journal/ijms
- Thiruvengadam M, Rajakumar G, Chung IM (2018) Nanotechnology: current uses and future applications in the food industry. 3 Biotech 8:74. https://doi.org/10.1007/s13205-018-1104-1107
- Torres-Pacheco I (2011) Aflatoxins detection, measurement and control. In: Mosiello L, Lamberti I (eds) Biosensors for aflatoxins detection. InTech, pp 147–160. https://doi.org/10.5772/22095. ISBN 978-953-307-711-6
- Vasimalai N, John SA (2013) Picomolar melamine enhanced the fluorescence of gold nanoparticles: spectrofluorimetric determination of melamine in milk and infant formulas using functionalized triazole capped gold nanoparticles. Biosens Bioelectron 42:267–272. https://doi. org/10.1016/j.bios.2012.10.023
- Verdian A (2018) Apta-nanosensors for detection and quantitative determination of acetamiprid a pesticide residue in food and environment. Talanta 176:456–464. https://doi.org/10.1016/j. talanta.2017.08.070
- Wangkam T, Srikhirin T, Wanachantararak P, Baxi V, Sutapun B, Amarit R (2009) Investigation of enzyme reaction by surface plasmon resonance (SPR) technique. Sens Actuators B-Chem 139(2):274–279
- Wani AL, Ara A, Usmani JA (2015) Lead toxicity: a review. Interdisc Toxicol 8(2):55–64. https:// doi.org/10.1515/intox-2015-0009. ISSN: 1337-6853 (print version) | 1337-9569 (electronic version)
- World Health Organization (WHO) Available online at https://www.who.int/foodsafety/areas_ work/chemical-risks/pops/en/index1.html. Accessed Jan 2020

Chapter 11 Utility of Nanobiosensors in Environmental Analysis and Monitoring



Kshitij RB Singh and Ravindra Pratap Singh

Contents

11.1	Introduction	230
11.2	Quantum-Dot-Based Nanobiosensors	232
11.3	Carbon-Based Nanobiosensors.	233
11.4	Dendrimers-Based Nanobiosensors.	235
11.5	Liposomal Nanobiosensors	236
11.6	Metal and Metal Oxides-Based Nanobiosensors.	237
11.7	Polymeric-Based Nanobiosensors.	238
11.8	Conclusion.	238
Refere	ences	240

Abstract Nanotechnology has evolved very fast in recent years, and biosensors based on nanomaterials (nanobiosensors) are finding a variety of applications in the environmental, clinical, and agricultural domains just due to the versatile properties of the nanomaterials used for designing nanobiosensors. The major threat to environment is the release of heavy metal ions, pesticides, chemical toxins, etc., into water bodies, food, and the atmosphere. These sources are directly in contact with the living world and these environmental pollutants pose severe global threats to health. Using nanoscale-structured material for improving biosensing technology has brought in a novel and cost-effective technology for environmental analysis and monitoring. Due to the complexity of these pollutants, traditional biosensors; these types of sensors are very selective and sensitive. In this book chapter, different types of nanobiosensors based on quantum dot, carbon, dendrimers, liposomal, metal and

K. R. Singh \cdot R. P. Singh (\boxtimes)

229

Department of Biotechnology, Indira Gandhi National Tribal University, Amarkantak, Madhya Pradesh, India

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_11

metal oxides, and polymeric are elaborated in detail for their application in environmental analysis and monitoring.

Keywords Biosensors · Nanobiosensors · Quantum dot nanobiosensors · Dendrimers nanobiosensors · Metal and metal oxide nanobiosensors · Carbonbased nanobiosensors · Liposomal nanobiosensors · Polymeric nanobiosensors

11.1 Introduction

Nanotechnology and nanoscience deal with the scientific study and applicability of nanoscale structures (1–100 nanometer). Nanotechnology is a science that is based on the engineering, design, synthesis, characterization, and application of materials and its products, which range in the nanometer-scale dimensions (Singh et al. 2010a; Silva 2004). In simple terms, nanotechnology basically deals with the fact that very small assemblies of bulk matter have unique properties and behavior that the bulk matter lacks with the same composition. The development of nanosensors for application in the monitoring and analysis of environment pollutants has evolved very fast; different nanostructures are used for the above applications, namely quantum dots, carbon-based, dendrimers, and liposomal (Singh et al. 2012a; Riu et al. 2006; Singh 2016).

Biosensors are analytical devices used for the detection of analytes of interest; analytes are biological components such as biological structure, biomolecule, and microorganism. They have three main parts: recognition element (bioreceptor), transducer, and amplifier. The bioreceptor is the recognition element that binds with analytes to produce biological signal; this signal is the primary signal produced by any biosensor device. The transducer recognizes the biological signal and converts it to a suitable form that can be analyzed, stored, manipulated, amplified, stored, and displayed. The amplifier just amplifies the converted biological signal to an electrical signal and displays the data on a micro-electronic device, as shown in Fig. 11.1



Fig. 11.1 Shows schematic representation of basic biosensors along with examples of different bioreceptors and transducers used in biosensing

(Singh 2011a, 2012; Rasooly and Herold 2006). Biosensors are of various types such as tissue and enzyme based, immunosensors, DNA (Deoxyribonucleic acid), piezoelectric, and thermal. They are used in various fields for environmental monitoring, agriculture, and biomedical diagnostics for the detection of analytes (Singh et al. 2008; Mehrotra 2016). Peptide-nucleic-acid-based biosensors are widely used in the field of biomedical for diagnostic applications (Singh et al. 2010a, 2020), and its application in environmental monitoring and analysis is very less explored.

The heavy metal waste (e.g., Pb, Cd, Hg) from metallurgic industries mostly pollutes the biosphere because of its toxic nature (Yilmaz et al. 2007). Thus, there is a very urgent need to develop real-time monitoring devices, such as biosensors, nanosensors, and nanobiosensors, for the detection of industrial pollutants (Singh and Choi 2010; Singh et al. 2011a). The existing methods, such as atomic absorption spectrometer (AAS), inductively coupled plasma mass spectrometry, atomic emission spectrometers, etc., are very expensive and are not designed for on-site detection (Burlingame et al. 1996; Jackson et al. 1996), but these nanomaterial-based sensors (nanobiosensors) are more powerful analytical tools because of their high sensitivity, portability, selectivity, and on-site detection (Ullah et al. 2018). Much literature suggests that enzyme-based nanobiosensors are very sensitive for heavy metal ion detection, and this type of enzyme-based nanobiosensors use enzyme activity inhibition when the heavy metal ions form conjugates with enzymes thiol groups (Wanekaya et al. 2008).

The use of pesticides in large quantities to enhance the yield of agricultural produce is the main cause of environmental pollution. Residues of pesticides are found in water and food products, which are direct routes of human interaction; these affect human health directly. Thus, there is a very urgent need to develop a device that has the ability to detect these small environmental pollutants in real samples (Singh et al. 2009b; Nsibande and Forbes 2016).

Electrochemical biosensors are most suitable for sensing environmental pollutants as they are sensitive, selective, precise, rapid, and easy to operate (Singh et al. 2009a, 2010b; Badihi-Mossberg et al. 2007). Immunosensors and DNA-based biosensors are effective in the determination of very low concentrations of environmental pollutants at rapid speed. This is because of signal amplification and noise reduction techniques, along with miniaturization, which increases sensitivity (Connelly and Baeumner 2012). The design of biosensors has seen significant changes in recent years, and the use of nanomaterials for biosensor development has reached new heights. Nanomaterial-based sensors (nanobiosensors) are now a hot topic and are attracting enthusiastic researchers for their applicability in the clinical, agricultural, and environmental fields (Singh et al. 2012b, 2012d). This is because of the fact that nanomaterials have unique properties such as better electrical conductivity, piezoelectric, and better shock-bearing ability (Malik et al. 2013).

There is a very urgent need to develop novel methodology for continuous monitoring and analysis of the environment. Thus, in this regard, biosensors have emerged as the best analytical tool for the determination of analytes that pollute the environment (Singh and Pandey 2011; Rodriguez-Mozaz et al. 2006). In this book chapter, we have described in detail the use of quantum-dot-based, carbon-based, dendrimers-based, liposomal-based, metal- and metal-oxides-based, and polymericbased nanobiosensors in environmental analysis and monitoring (Singh et al. 2012c).

11.2 Quantum-Dot-Based Nanobiosensors

Quantum-dot-based nanobiosensors for environmental analysis and monitoring are gaining more importance and are being developed at a faster pace in recent years. Quantum dots have outstanding optical and electronic properties, and when compared with other fluorophores, they exhibit high-fluorescence quantum yields. Apart from these properties, they also display multifaceted surface functionalization and are highly photostable, which make them unique when compared to organic dyes. There is still a need to develop quantum-dot-based nanobiosensors that can be commercialized and used routinely for the detection and monitoring of environmental pollutants such as pesticides, heavy metals, and chemical toxins. Quantum-dot-based nanobiosensors as a metal sensor for the detection of toxic heavy metal ions, such as cadmium (II)(Cd²⁺), Hg²⁺, and Pb²⁺, are receiving great importance, and many researchers have developed this type of sensors and many are working in this domain because heavy metal contamination of water is the major cause of water pollution (Nsibande and Forbes 2016; Vazquez-Gonzalez and Carrillo-Carrion 2014).

Nanobiosensor systems based on water-soluble quantum dots capped with ligands such as mercaptoacetic acid (MAA), L-cysteine, and N-acetyl-L-cysteine (NAC) for the detection of heavy metal ions are being developed. In this type of systems, ligands are responsible for response to metal ions as well as solubility of quantum dots in aqueous condition. A group of researchers have capped MAA with CdS and CdS/ZnS quantum dots, that resulted in MAA-CdS and MAA-CdS/ZnS quantum dots. The resulting compound is a fluorescent probe that was used for the detection of Hg²⁺ (Mercury (II)). This detection was totally dependent on the fluorescence quenching of quantum dots in the presence of these ions, and the result signifies that this type of sensors show excellent sensitivity and selectivity (Koneswaran and Narayanaswamy 2009, 2012). Further, study by Duan et al. (2011) produced a less toxic sensor based on ZnS quantum dots; this nanoparticle-based sensor/nanobiosensor was applied to detect Hg²⁺ traces in water sample.

Wu et al. (2008) reported thioglycolic acid (TGA)-capped CdTe quantum dots for the detection of lead II (Pb²⁺) in food samples of popcorn and instant noodles. The major principle behind using this method was fluorescence quenching of CdTe quantum dots and the result obtained from this study was almost similar to the results obtained from flame atomic absorption spectrometry. Cai et al. (2012) synthesized highly luminescent l-glutathione (GSH)-capped ZnSe quantum dots in aqueous samples. Further, for the detection of Pb²⁺ in micromolar concentration, Goncalves et al. (2009) also used GHS-capped CdTe quantum dots; to study this system for fluorescence spectra, they carried out PARAFAC (parallel factor) analysis. Zhao et al. (2013) reported an ultrasensitive method for the detection of Pb²⁺ by using dithizone (Dz)-functionalized CdSe/CdS based on enhancement in fluorescence, as conjugates of Dz-quantum dots cause fluorescence quenching. This fluorescent probe was validated on real samples of soil, tap water, and river water for Pb²⁺ detection, and the result of this study showed excellent performance with very good sensitivity and selectivity.

The development of nanobiosensors for heavy metal ion detection or other pollutants are based on changes induced by analytes in quantum dots; these changes are dependent on Forster's theory, and this theory is the major principle behind the fluorescence resonance energy transfer (FRET). In this system, the quantum dots act as donors, and the light energy absorbed by the quantum dots is transferred to other nearby acceptors (example, other nanoparticles and organic fluorophore) (Medintz et al. 2003; Algar et al. 2010). Based on FRET systems using quantum dots, Li et al. in 2008 reported TGA CdTe quantum dots as donors and butyl-rhodamine B, which is an acceptor of light energy in the presence of cetyltrimethylammonium bromide (a surfactant) for the detection of Hg²⁺. Many researchers have used ZnS QDs and Mn²⁺ quantum dots to make nanobiosensors for the detection of different pesticides such as cyphenothrin (Ren and Chen 2015), pentachlorophenol (Wang et al. 2009; Yang et al. 2012), and acetamiprid (Lin et al. 2016).

11.3 Carbon-Based Nanobiosensors

Carbon-based nanomaterials such as carbon nanotubes (CNTs), graphene, and other allotropes of carbon have been used in the development of nanobiosensors because they are very selective and sensitive in the detection of several analytes. Carbon-based nanomaterial biosensors impart unique electrical and physical properties, which offer them a variety of applications for the detection and monitoring of environmental pollutants such as heavy metal ions, explosives, pathogens, and pesticides. This type of nanobiosensors are simple, cost-effective, and handy (used for on-site detection), and give immediate results (Ramnani et al. 2016).

Aptasensors (DNA-functionalized CNT biosensors) are used widely to study heavy metal ion interaction with DNA because DNA are highly stable at optimum pH condition. Liu and Wei (2008) reported a DNA-single-walled carbon nanotube (DNA-SWCNT)-modified glassy carbon electrode (GCE) for the detection of arsenic (III) (As³⁺). This type of aptasensors was highly sensitive and selective to As³⁺ with a limit of detection (LOD) of 0.05 μ g/L. Li et al. (2009) reported graphenemodified GCE for the detection of Pb²⁺ and Cd²⁺. Further, Morton et al. (2009) reported a carboxylated CNT-modified GCE for the detection of heavy metal ions Pb²⁺ and Cu²⁺ (Copper (II)). Gong et al. (2013) reported a label-free chemiresistive biosensor for Hg²⁺ heavy metal ion detection based on SWCNTs functionalized with duplex polyT:polyA.

By changing the surface chemistry of the carbon-based nanomaterial using antibodies that are specific to a pathogen which is to be identified, a highly sensitive and selective device for the detection of viruses and bacteria can be developed. Liu et al. (2013) reported field-effect transistor-reduced graphene oxide nanobiosensors for the detection of the rotavirus (water-borne virus), which causes diarrhea. They have examined real samples and the result showed a LOD of 10³pfu/ml, which means that this detection method was very sensitive and selective. García-Aljaro et al. (2010) added by reporting SWCNT-chemiresistive biosensors functionalized with antibodies for the detection of E. coli O157:H7 bacteria and T7 bacteriophage virus. There results signify that this type of nanobiosensors for pathogen (bacteria and viruses) detection are very sensitive and selective. Zelada-Guillen et al. (2009) reported a carboxylated CNT-modified GCE for the detection of the Salmonella Typhi bacteria. Further, Huang et al. (2011) designed a graphene-based field-effect transistor for the detection of E. coli K12. Bacillus anthracis (gram-positive bacterium) are toxic to humans and are used as an agent for bioterrorism. They produce the lethal toxin anthrax, made up of a protective agent that, on entering the host, causes death in no time (Dixon et al. 1999; Milne et al. 1994). Cella et al. (2010) reported a chemiresistive nanobiosensor with SWCNTs matrix for the detection of protective agent toxin using a single-stranded DNA aptamer (biorecognition molecule).

Lian et al. (2014) reported, DNA-wrapped metallic SWCNTs which was very novel for fabricating the electrochemical nanobiosensor for the detection of Pb^{2+} in water. The working mechanism of this sensor for Pb^{2+} detection is based on the wrapping of Pb^{2+} with negatively charged phosphate groups or to the bases of DNA (Oliveira et al. 2008; Liu et al. 2009). Thus, the testing of real samples from tap water and on-site testing of river water showed similarity with the results of AAS, which proves that these nanobiosensors are reliable and accurate.

Pesticides are widely used in agricultural practices to inhibit the growth of certain plants and animals, which in turn increases the yield of agricultural produces. They are also known as toxic pollutants, which are harmful to human health. The most used toxic substance in pesticides is organophosphates, which cause cholinergic dysfunction in humans and animals. Thus, rapid, cost-effective, and sensitive detection of organophosphate neurotoxins is very much needed to protect food supplies, water resources, etc., from getting contaminated by these toxins. For the detection of organophosphates, there are traditional techniques used in laboratories such as mass spectrometry and chromatography, which consume time and are costly (Sherma 1993). Enzyme-based biosensors using CNTs as transducers for highly selective, sensitive, and rapid detection of pesticides. This enzymatic biosensor can be used for direct detection by using the catalytic activity mechanism of organophosphorus hydrolase (Mulchandani et al. 2001) or by using the inhibitory activity mechanism of different enzymes found in pesticides such as butyrylcholinesterase, tyrosinase, and acetylcholinesterase (AChE) (Andreescu and Marty 2006; Tanimoto and Ferreira 2007). Du et al. (2007) reported a biosensor for rapid detection of triazophos (organophosphorus insecticide) using AChE. They have used multiwalled carbon nanotubes (MWCNTs)-chitosan composite as working electrodes. Joshi et al. (2006) reported highly sensitive and selective carbon nanotube-based amperometric enzyme electrode biosensors for the detection of V-type nerve agents.

11.4 Dendrimers-Based Nanobiosensors

Dendrimers are nanoscale-dimension structures that originate radially from the central core; they are man-made branched polymers and monodispersed macromolecules with star-like structures. In general, dendrimers have three components, namely central core, branches (interior structure), and exterior surface, with functional groups, as shown in Fig. 11.2 (Klajnert and Bryszewska 2001; Singh 2011a, b). Dendrimers have a variety of applications in the biomedical domain for in vitro diagnosis, gene therapy, regenerative medicine, therapeutics, and many more (Dufes et al. 2005; Oliveira et al. 2010; Singh 2007). The use of dendrimer-based nanobiosensors for environmental monitoring and analysis is very less explored and further research in this direction is required.

Castillo et al. (2015) reported a biosensor based on aptamer for the detection of aflatoxin B1 (AFB1); it is a mycotoxin which is a known food contaminant. They used cystamine dendrimer layers and compared with different immobilization platforms such as 11-mercaptoundecanoic acid (MUA), cystamine (Cys), and 11-mercaptoundecanoic acid-dendrimers. After the immobilization, they validated the nanobiosensor for real samples such as peanut extracts and peanut-corn snacks, and the result of the sensing response was very sensitive and selective toward the analyte. Singh et al. (2009) reported a new approach for the fabrication of nanobiosensors based on polyamidoamine (PAMAM) dendrimers (a fourth generation dendrimer) for the detection of explosive molecules like trinitrotoluene, and this type of sensors showed high sensitivity, stability, and reproducibility. Grabchev and Guittonneau (2006) synthesized poly(amidoamine) dendrimers of zero and second generation, which contain fluorescent 4-piperidino-1, 8-naphthalimide units inside their periphery; further, they have investigated these synthesized materials for the



Fig. 11.2 Shows detailed structure of dendrimers

detection of metal cations and protons, and from their studies, they found that these materials are very sensitive and selective in the detection of environment-polluting metal cations and protons such as Zn^{2+} , Ni^{2+} , Ce^{3+} , Ag^+ , Co^{2+} , and Cu^{2+} cations.

Snejdarkova et al. (2003) reported choline oxidase and AChE immobilized with PAMAM fourth generation dendrimer based biosensors for the detection of dimethyl-2,2-dichlorovinyl- phosphate, carbofuran, and eserine pesticides. From the results of this study, they found that this type of nanobiosensors have very low detection limits compared with the previously reported amperometric and potentio-metric nanobiosensors based on AChE. Hence, keeping the above viewpoint dendrimer based nanobiosensors are highly sensitivity and stabel which make them a suitable candidate for the detection of environmental pollutants in real/field samples.

11.5 Liposomal Nanobiosensors

Liposomes are developed to study cell membranes; they are artificial spherical vesicles composed of a phospholipid bilayer surrounded by an aqueous cavity. These cavities can bear different molecules inside them due to which liposomes have unique application in diagnostics, therapeutics, and environmental monitoring. Other molecules can associate with liposomes in different ways such as encapsulation, covalent bonding, partitioning inside the lipid bilayer tails, and by electrostatic interaction. Marker attached on the surface of liposome with antibodies, DNA, or haptens can be used for the detection of environmental pollutants and further after lysing of liposomes, the marker is released (Ahn-Yoon et al. 2003; Edwards and Baeumner 2006).

Triazines are pesticide groups which have a wide range of applicability in the agriculture sector. This herbicide member in their family have many other derivative such as atrazine, terbuthylazine, etc. and is most commonly used in the USA and Europe. These compounds show moderate toxicity but can undergo further transformation and become more toxic, carcinogenic, and mutagenic. These are classified as environmental pollutants and can be found in the human food chain or in drinking water (Jones and de Voogt 1999; Baran et al. 2003). Thus, understanding the need for the detection of such moderately toxic compounds which can in future become more toxic, Baumner and Schmid (1998) reported a liposome-based amperometric sensor for the detection of triazine pesticides in aqueous samples. The developed sensor was disposable and was based on the competition between tagged pollutants and free pollutants.

The *Vibrio cholerae* bacteria secrets cholera toxin, which causes diarrhea, cramps, vomiting, and sometimes leads to death. When the cholera toxin binds to intestinal cells and infects the intestine, it triggers endocytosis of toxins. Hence, for the rapid and sensitive detection of the cholera toxin, Viswanathan et al. (2006) reported liposome carrying potassium ferrocyanide labelled with molecules that are highly specific to the analytes. These type of devices have applications for

biomedical diagnostics and environmental monitoring of pollutants; can also be very much useful in controlling epidemic.

11.6 Metal and Metal Oxides–Based Nanobiosensors

Solanki et al. (2011) reported that nanostructured metal oxides (NMOs) are the future materials for the next generation of nano-sized biosensing (nanobiosensing) applications because they provide an effective surface for the immobilization of biomolecules. NMOs exhibit unique electrical, molecular, and optical properties; these properties provide the NMO-based biosensors with unique applications in the clinical, environmental, and agricultural fields (Singh 2019c, d).

Electrode surface adsorption capacity plays a very significant role in electrochemical activities (Wei et al. 2012; Zhao et al. 2012). Ferrite nanoparticles have super paramagnetic properties and high adsorption capacity; therefore, they have attracted a lot of attention (Xu et al. 2013). Iron (Fe³⁺/Fe²⁺) exhibits redox coupling that make them relevent for reduction of Arsenic (II) (As^{2+}) (Cepria et al. 2007; Guardia et al. 2012). Arsenic (As) can be removed from water using strong adsorption mechanisms of Fe_3O_4 monodispersed nanocrystals (Yavuz et al. 2006). Thus, ferrite nanoparticles have great potential for the electrochemical detection of heavy metal ions as well as inorganic As (Zhang et al. 2016). Wu et al. (2014)reported a nanobiosensor fabricated by electrodeposition of manganese oxides and gold nanoparticles on the glassy carbon electrode (GCE) for the detection of As (III) in alkaline solution by utilizing the cyclic voltammetry and this reported nanosensor was very selective and sensitive when examined in real samples. Dong et al. (2016) reported metal-organic frameworks (MOFs) based electrochemical sensor that was fabricated by utilizing 1,3,5-benzenetricarboxylic acid copper $[Cu_3(BTC)_2]$) on modified carbon paste electrode for the detection of 2,4-dichlorophenol (2,4-DCP) in real water samples and this type of nanobiosensors gave satisfactory results in the real time sensing of 2,4-DCP.. Thus, MOF-based sensors exhibit promising future applications for routine sensing of environmental contaminants/pollutants (Arora et al. 2006; Shukla et al. 2010; Singh et al. 2011b, c).

For the detection of Pb^{2+} in real samples of water, Zhao et al. (2016) synthesized gold nanoparticles-rGO as surface enhanced Raman scattering (SERS) substrates for sensitive and selective detection. Frost et al. (2015) reported functionalized gold nanoparticles from citrate for the ultrasensitive detection of Pb^{2+} based on SERS in aqueous conditions. Chen et al. (2014) reported SERS nanobiosensors based on Ag nanoparticles functionalized by 4-mercaptopyridine for the detection of Hg^{2+} in aqueous samples with the help of spermine, which binds with Ag nanoparticles by Ag-N bonds that result in Ag nanoparticle aggregation and this, in turn, provides enhanced Raman intensity and helps in the optical sensing of Hg^{2+} metal ions. Li et al. (2013) reported dye-coded Ag nanoparticle nanobiosensors based on SERS for the sensitive and selective detection of Cu^{2+} and Hg^{2+} heavy metal ions, which are major environment polluting agents.

11.7 Polymeric-Based Nanobiosensors

Polymer-based nanoparticles are large molecules ranging between 1 and 1000 nm (sub-micron range). They are colloidal particles with good encapsulation properties; due to this reason, they are excellent pharmaceutical ingredients. Nanotechnology is playing a crucial role in environmental analysis and monitoring (Singh et al. 2014). Polymer-based nanocomposites exhibit unique properties and have received great attention from industries and academia, as the properties of polymers offer the formed composite unique physical and chemical characteristics. The polymer nanocomposites have interesting and never-ending environmental applications for the detection of environmental pollutants (Zhao et al. 2011; Kreuter 2014). Guo and Chen (2005) reported iron oxyhydroxide nanoparticles on cellulose polymer matrixbased biosensors for the detection of As^{3+} and As^{5+} heavy metal ions in groundwater. Zouboulis and Katsoyiannis (2002) reported iron oxides-alginate beads-based biosensors for the detection and removal of arsenic.

Gas sensors are a type of sensors which can detect presence of gases; this type of sensors help in the continuous analysis and monitoring of different types of gases present in the environment, such as carbon monoxide, ethanol, acetone, trimethylamine, carbon dioxide, nitrogen, and methanol (Singh 2019b). Keeping the above viewpoint, Ram et al. (2005) reported an ultrathin conducting matrix of polystyrene (PSS) and polyaniline (PANI) polymers with metal oxide nanoparticles for the detection of carbon monoxide. Further, Geng et al. (2007) reported PANI-SnO₂-based biosensors prepared by using a hydrothermal method for the detection and control of ethanol and acetone fume. Zheng et al. (2008) synthesized titanium dioxide (TiO₂)-PANI using chemical polymerization and the sol-gel method for use as a biosensor for the detection of trimethylamine. Suri et al. (2002) investigated iron oxide-polypyrrole-based biosensors based on Pd-PANI synthesized by using oxidative polymerization for the detection of methanol.

There are many other polymer-based nanobiosensors for the detection of heavy metal ions, humidity changes, industrial wastes, fecal matter (Singh 2017, 2019a). Sugunan et al. (2005) reported a nanobiosensor based on chitosan-Au for the detection of Zn^{2+} and Cu^{2+} . Su and Huang (2007) reported a synthesis of TiO₂-polypyrrole by in-situ photopolymerization for the detection of changes in humidity for proper monitoring and maintenance of environmental pollutants.

11.8 Conclusion

Nanotechnology has evolved very fast in the analysis and monitoring of the environment using nanoparticles, namely liposomes, dendrimers, metal, and metal oxide. These nanoparticles have a wide range of properties that offer them a wide range of applications in the environmental, biomedical, and agricultural fields. Talking



Fig. 11.3 Shows graphical description of the detail work

specifically in this book chapter, we have discussed the application of these nanoparticles in the environmental field for the analysis and monitoring of environmental pollutants, as depicted in Fig. 11.3.

Environmental pollution is the major cause of health issues in humans and animals, and it is also leading to global warming. The major environmental pollutants affect living things by different mediums, namely water, food, and air, as they are directly in contact with these mediums. Biosensors are widely used for the detection of environmental pollutants, which helps to better monitor and control environmental pollutants. But biosensors have evolved rapidly, and now nanoparticle-based biosensors are being used for these applications; these types of biosensors are called nanobiosensors. As nanobiosensors are more precise, sensitive, optimized, selective, rapid, and best suitable for on-site detection of pollutants (single compound/ class of compounds).

Hence, the use of these nanoparticles for making more suitable biosensors for solving the problems related to environmental analysis and monitoring is the future, which can help overcome this global challenge. This book chapter elaborates the current use of these nanoparticles in making nanobiosensors for analysis and monitoring the environment. There are many nanoparticles, namely polymer-based nano-composites, and dendrimers-based, whose fabrication for the development of nanobiosensors with high selectivity, sensitivity, and cost-effectiveness is the hot topic and needs to be explored by researchers. Studies in this direction for the purpose of commercialization need to be carried out.

Acknowledgments The authors are thankful to Indira Gandhi National Tribal University, Amarkantak, M.P., India, for providing facilities to prepare this book chapter.

References

- Ahn-Yoon S, DeCory TR, Baeumner AJ, Durst RA (2003) Ganglioside-liposome immunoassay for the ultrasensitive detection of cholera toxin. Anal Chem 75(10):2256–2261. https://doi. org/10.1021/ac026428t
- Algar WR, Tavares AJ, Krull UJ (2010) Beyond labels: a review of the application of quantum dots as integrated components of assays, bioprobes, and biosensors utilizing optical transduction. Anal Chim Acta 673(1):1–25. https://doi.org/10.1016/j.aca.2010.05.026
- Andreescu S, Marty JL (2006) Twenty years research in cholinesterase biosensors: from basic research to practical applications. Biomol Eng 23(1):1–15. https://doi.org/10.1016/j. bioeng.2006.01.001
- Arora K, Chaubey A, Singhal R, Singh RP, Pandey MK, Samanta SB, Malhotra BD, Chand S (2006) Application of electrochemically prepared polypyrrole-polyvinyl sulphonate films to DNA biosensor. Biosens Bioelectron 21(9):1777–1783
- Athawale AA, Bhagwat SV, Katre PP (2006) Nanocomposite of Pd–polyaniline as a selective methanol sensor. Sensors Actuators B Chem 114(1):263–267. https://doi.org/10.1016/j. snb.2005.05.009
- Badihi-Mossberg M, Buchner V, Rishpon J (2007) Electrochemical biosensors for pollutants in the environment. Electroanalysis 19(19–20):2015–2028. https://doi.org/10.1002/elan.200703946
- Baran S, Oleszczuk P, Baranowska E (2003) Degradation of soil environment in the post-flooding area: content of polycyclic aromatic hydrocarbons (PAHs) and S-triazine herbicides. J Environ Sci Health Part B 38(6):799812. https://doi.org/10.1081/PFC-120025561
- Bäumner AJ, Schmid RD (1998) Development of a new immunosensor for pesticide detection: a disposable system with liposome-enhancement and amperometric detection. Biosens Bioelectron 13(5):519–529. https://doi.org/10.1016/S0956-5663(97)00131-0
- Burlingame AL, Boyd RK, Gaskell SJ (1996) Mass spectrometry. Anal Chem 68(12):599–652. https://doi.org/10.1021/a1960021u
- Cai Z, Shi B, Zhao L, Ma M (2012) Ultrasensitive and rapid lead sensing in water based on environmental friendly and high luminescent l-glutathione-capped-ZnSe quantum dots. Spectrochim Acta Part A Mol Biomol Spec 97:909–914. https://doi.org/10.1016/j.saa.2012.07.069
- Castillo G, Spinella K, Poturnayova A, Šnejdárková M, Mosiello L, Hianik T (2015) Detection of aflatoxin B1 by aptamer-based biosensor using PAMAM dendrimers as immobilization platform. Food Control 52:9–18. https://doi.org/10.1016/j.foodcont.2014.12.008
- Cella LN, Sanchez P, Zhong W, Myung NV, Chen W, Mulchandani A (2010) Nano aptasensor for protective antigen toxin of anthrax. Anal Chem 82(5):2042–2047. https://doi.org/10.1021/ac902791q
- Cepriá G, Hamida S, Laborda F, Castillo JR (2007) Direct reduction of As(V) physically attached to a graphite electrode mediated by Fe(III). J Appl Electrochem 37(10):1171–1176. https://doi.org/10.1007/s10800-007-9380-7
- Chen L, Qi N, Wang X, Chen L, You H, Li J (2014) Ultrasensitive surface-enhanced Raman scattering nanosensor for mercury ion detection based on functionalized silver nanoparticles. RSC Adv 4(29):15055–15060. https://doi.org/10.1039/C3RA47492E
- Connelly JT, Baeumner AJ (2012) Biosensors for the detection of waterborne pathogens. Anal Bioanal Chem 402(1):117127. https://doi.org/10.1007/s00216-011-5407-3
- Dixon TC, Meselson M, Guillemin J, Hanna PC (1999) Anthrax. N Engl J Med 341(11):815–826. https://doi.org/10.1056/NEJM199909093411107
- Dong S, Suo G, Li N, Chen Z, Peng L, Fu Y, Huang T (2016) A simple strategy to fabricate high sensitive 2,4-dichlorophenol electrochemical sensor based on metal organic framework Cu3(BTC)2. Sensors Actuators B Chem 222:972–979. https://doi.org/10.1016/j. snb.2015.09.035
- Du D, Huang X, Cai J, Zhang A (2007) Amperometric detection of triazophos pesticide using acetylcholinesterase biosensor based on multiwall carbon nanotube–chitosan matrix. Sensors Actuators B Chem 127(2):531–535. https://doi.org/10.1016/j.snb.2007.05.006

- Duan J, Jiang X, Ni S, Yang M, Zhan J (2011) Facile synthesis of N-acetyl-l-cysteine capped ZnS quantum dots as an eco-friendly fluorescence sensor for Hg2+. Talanta 85(4):1738–1743. https://doi.org/10.1016/j.talanta.2011.06.071
- Dufes C, Uchegbu I, Schatzlein A (2005) Dendrimers in gene delivery. Adv Drug Deliv Rev 57(15):2177–2202. https://doi.org/10.1016/j.addr.2005.09.017
- Edwards K, Baeumner A (2006) Analysis of liposomes. Talanta 68(5):1432–1441. https://doi. org/10.1016/j.talanta.2005.08.031
- Frost MS, Dempsey MJ, Whitehead DE (2015) Highly sensitive SERS detection of Pb2+ ions in aqueous media using citrate functionalised gold nanoparticles. Sensors Actuators B Chem 221:1003–1008. https://doi.org/10.1016/j.snb.2015.07.001
- García-Aljaro C, Cella LN, Shirale DJ, Park M, Muñoz FJ, Yates MV, Mulchandani A (2010) Carbon nanotubes-based chemiresistive biosensors for detection of microorganisms. Biosens Bioelectron 26(4):1437–1441. https://doi.org/10.1016/j.bios.2010.07.077
- Geng L, Zhao Y, Huang X, Wang S, Zhang S, Wu S (2007) Characterization and gas sensitivity study of polyaniline/SnO2 hybrid material prepared by hydrothermal route. Sensors Actuators B Chem 120:568–572. https://doi.org/10.1016/j.snb.2006.03.009
- Goncalves H, Mendonca C, Esteves da Silva JCG (2009) PARAFAC analysis of the quenching of EEM of fluorescence of glutathione capped CdTe quantum dots by Pb(II). J Fluoresc 19(1):141–149. https://doi.org/10.1007/s10895-008-0395-1
- Gong JL, Sarkar T, Badhulika S, Mulchandani A (2013) Label-free chemiresistive biosensor for mercury (II) based on single-walled carbon nanotubes and structure-switching DNA. Appl Phys Lett 102(1):013701. https://doi.org/10.1063/1.4773569
- Grabchev I, Guittonneau S (2006) Sensors for detecting metal ions and protons based on new green fluorescent poly (amidoamine) dendrimers peripherally modified with 1, 8-naphthalimides. J Photochem Photobiol A Chem 179:28–34. https://doi.org/10.1016/j.jphotochem.2005.07.008
- Guardia P, Di Corato R, Lartigue L, Wilhelm C, Espinosa A, Garcia-Hernandez M, Pellegrino T (2012) Water-soluble iron oxide nanocubes with high values of specific absorption rate for cancer cell hyperthermia treatment. ACS Nano 6(4):3080–3091. https://doi.org/10.1021/ nn2048137
- Guo X, Chen F (2005) Removal of arsenic by bead cellulose loaded with iron oxyhydroxide from groundwater. Environ Sci Technol 39(17):6808–6818. https://doi.org/10.1021/es048080k
- Huang Y, Dong X, Liu Y, Li LJ, Chen P (2011) Graphene-based biosensors for detection of bacteria and their metabolic activities. J Mater Chem 21(33):12358. https://doi.org/10.1039/ c1jm11436k
- Jackson C, Chen YJ, Mays JW (1996) Size exclusion chromatography with multiple detectors: solution properties of linear chains of varying flexibility in tetrahydrofuran. J Appl Pol Sci 61(5):865–874. https://doi.org/10.1002/(SICI)1097-4628(19960801)61:5<865:: AID-APP20>3.0.CO;2-V
- Jones KC, de Voogt P (1999) Persistent organic pollutants (POPs): state of the science. Environ Pollut 100(1-3):209-221. https://doi.org/10.1016/S0269-7491(99)00098-6
- Joshi KA, Prouza M, Kum M, Wang J, Tang J, Haddon R, Mulchandani A (2006) V-type nerve agent detection using a carbon nanotube-based amperometric enzyme electrode. Anal Chem 78(1):331–336. https://doi.org/10.1021/ac051052f
- Klajnert B, Bryszewska M (2001) Dendrimers: properties and applications. Acta Biochim Pol 48(1):199–208. https://doi.org/10.18388/abp.2001_5127
- Koneswaran M, Narayanaswamy R (2009) RETRACTED: mercaptoacetic acid capped CdS quantum dots as fluorescence single shot probe for mercury(II). Sensors Actuators B Chem 139(1):91–96. https://doi.org/10.1016/j.snb.2008.09.011
- Koneswaran M, Narayanaswamy R (2012) CdS/ZnScore-shell quantum dots capped with mercaptoacetic acid as fluorescent probes for Hg(II) ions. Microchim Acta 178(1–2):171–178. https:// doi.org/10.1007/s00604-012-0819-0
- Kreuter J (2014) Drug delivery to the central nervous system by polymeric nanoparticles: what do we know? Adv Drug Deliv Rev 71:2–14. https://doi.org/10.1016/j.addr.2013.08.008
- Li J, Mei F, Li WY, He XW, Zhang YK (2008) Study on the fluorescence resonance energy transfer between CdTe QDs and butyl-rhodamine B in the presence of CTMAB and its application on the detection of Hg(II). Spectrochim Acta Part A Mol Biomol Spec 70(4):811–817. https://doi. org/10.1016/j.saa.2007.09.017
- Li J, Guo S, Zhai Y, Wang E (2009) High-sensitivity determination of lead and cadmium based on the Nafion-graphene composite film. Anal Chim Acta 649(2):196–201. https://doi. org/10.1016/j.aca.2009.07.030
- Li F, Wang J, Lai Y, Wu C, Sun S, He Y, Ma H (2013) Ultrasensitive and selective detection of copper (II) and mercury (II) ions by dye-coded silver nanoparticle-based SERS probes. Biosens Bioelectron 39(1):82–87. https://doi.org/10.1016/j.bios.2012.06.050
- Lian Y, Yuan M, Zhao H (2014) DNA wrapped metallic single-walled carbon nanotube sensor for Pb (II) detection. Fullerenes Nanotubes Carbon Nanostruct 22(5):510–518. https://doi.org/1 0.1080/1536383X.2012.690462
- Lin B, Yu Y, Li R, Cao Y, Guo M (2016) Turn-on sensor for quantification and imaging of acetamiprid residues based on quantum dots functionalized with aptamer. Sensors Actuators B Chem 229:100–109. https://doi.org/10.1016/j.snb.2016.01.114
- Liu Y, Wei W (2008) Layer-by-layer assembled DNA functionalized single-walled carbon nanotube hybrids for arsenic(III) detection. Electrochem Commun 10(6):872–875. https://doi. org/10.1016/j.elecom.2008.03.013
- Liu CW, Huang CC, Chang HT (2009) Highly selective DNA-based sensor for lead(II) and mercury(II) ions. Anal Chem 81(6):2383–2387. https://doi.org/10.1021/ac8022185
- Liu F, Kim YH, Cheon DS, Seo TS (2013) Micropatterned reduced graphene oxide based fieldeffect transistor for real-time virus detection. Sensors Actuators B Chem 186:252–257. https:// doi.org/10.1016/j.snb.2013.05.097
- Malik P, Katyal V, Malik V, Asatkar A, Inwati G, Mukherjee TK (2013) Nanobiosensors: concepts and variations. ISRN Nanomater 2013:1–9. https://doi.org/10.1155/2013/327435
- Medintz IL, Clapp AR, Mattoussi H, Goldman ER, Fisher B, Mauro JM (2003) Self-assembled nanoscale biosensors based on quantum dot FRET donors. Nat Mater 2(9):630–638. https:// doi.org/10.1038/nmat961
- Mehrotra P (2016) Biosensors and their applications- a review. J Oral Biol Craniofacial Res 6(2):153–159. https://doi.org/10.1016/j.jobcr.2015.12.002
- Milne JC, Furlong D, Hanna PC, Wall JS, Collier RJ (1994) Anthrax protective antigen forms oligomers during intoxication of mammalian cells. J Biol Chem 269(32):20607–20612. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/8051159
- Morton J, Havens N, Mugweru A, Wanekaya AK (2009) Detection of trace heavy metal ions using carbon nanotube modified electrodes. Electroanalysis 21(14):1597–1603. https://doi.org/10.1002/elan.200904588
- Mulchandani A, Chen W, Mulchandani P, Wang J, Rogers KR (2001) Biosensors for direct determination of organophosphate pesticides. Biosens Bioelectron 16(4–5):225–230. https://doi. org/10.1016/S0956-5663(01)00126-9
- Nsibande SA, Forbes PBC (2016) Fluorescence detection of pesticides using quantum dot materials-a review. Anal Chim Acta 945:9–22. https://doi.org/10.1016/j.aca.2016.10.002
- Oliveira SCB, Corduneanu O, Oliveira-Brett AM (2008) In situ evaluation of heavy metal–DNA interactions using an electrochemical DNA biosensor. Bioelectrochemistry 72(1):53–58. https://doi.org/10.1016/j.bioelechem.2007.11.004
- Oliveira JM, Salgado AJ, Sousa N, Mano JF, Reis RL (2010) Dendrimers and derivatives as a potential therapeutic tool in regenerative medicine strategies-a review. Prog Poly Sci 35(9):1163–1194. https://doi.org/10.1016/j.progpolymsci.2010.04.006
- Ram MK, Ozlem Yavuz, Vitawat L, Idissi M (2005) CO gas sensing from ultrathin nanocomposite conducting polymer film. Sensors Actuators B Chem 106(2), 750–757. https://doi. org/10.1016/j.snb.2004.09.027

- Ramnani P, Saucedo NM, Mulchandani A (2016) Carbon nanomaterial-based electrochemical biosensors for label-free sensing of environmental pollutants. Chemosphere 143:85–98. https:// doi.org/10.1016/j.chemosphere.2015.04.063
- Rasooly A, Herold KE (2006) Biosensors for the analysis of food- and waterborne pathogens and their toxins. J AOAC Internat 89(3):873–883. Retrieved from http://www.ncbi.nlm.nih.gov/ pubmed/16792089
- Ren X, Chen L (2015) Quantum dots coated with molecularly imprinted polymer as fluorescence probe for detection of cyphenothrin. Biosens Bioelectron 64:182–188. https://doi. org/10.1016/j.bios.2014.08.086
- Riu J, Maroto A, Rius FX (2006) Nanosensors in environmental analysis. Talanta 69(2 SPEC. ISS.):288–301. https://doi.org/10.1016/j.talanta.2005.09.045
- Rodriguez-Mozaz S, Lopez De Alda MJ, Barcelo D (2006) Biosensors as useful tools for environmental analysis and monitoring. Anal Bioanal Chem 386:1025–1041. https://doi.org/10.1007/ s00216-006-0574-3
- Sherma J (1993) Pesticides. Anal Chem 65(12):40-54. https://doi.org/10.1021/ac00060a004
- Shukla VK, Singh RP, Pandey AC (2010) Black pepper assisted biomimetic synthesis of silver nanoparticles. J Alloys Compd 507(1):L13–L16
- Silva GA (2004) Introduction to nanotechnology and its applications to medicine. Surg Neurol 61(3):216–220. https://doi.org/10.1016/j.surneu.2003.09.036
- Singh P (2007) Dendrimers and their applications in immunoassays and clinical diagnostics. Biotechnol Appl Biochem 48(1):1. https://doi.org/10.1042/BA20070019
- Singh RP (2011a) Prospects of nanobiomaterials for biosensing. Int J Electrochem, Review article ID 125487, 30 pages. https://doi.org/10.4061/2011/125487
- Singh RP (2011b) A catechol biosensor based on a gold nanoparticles encapsulated-dendrimer. Analyst 136(6):1216–1221
- Singh RP (2012) Prospects of organic conducting polymer modified electrodes: enzymosensors. In: Scharifker BR (ed) Int J Electrochem, Article ID 502707, 14 pages. https://doi. org/10.1155/2012/502707
- Singh RP (2016) Nanobiosensors: potentiality towards bioanalysis. J Bioanal Biomed 8:e143. https://doi.org/10.4172/1948-593X.1000e143
- Singh RP (2017) Application of nanomaterials towards development of nanobiosensors and their utility in agriculture. In: Prasad R, Kumar M, Kumar V (eds) Nanotechnology: an agricultural paradigm. Springer, New York, Chapter 14, pp 293–303
- Singh RP (2019a) Nanocomposites: recent trends, developments and applications. In: Aliofkhazraei M (ed) Advances in nanostructure nanocomposites. CRC Press, Taylor and Francis, 575 Pages, Chap 2
- Singh RP (2019b) Utility of nanomaterials in food safety. In: Singh RL, Mondal SK (eds) Food safety and human health. Chap 11. Elsevier Inc
- Singh RP (2019c) Potential of biogenic plant-mediated iron and iron oxide nanostructured nanoparticles and their utility. In: Prasad R (ed) Nanotechnology in the life sciences: plant nanobionics, vol. 2. Approaches in nanoparticles biosynthesis and toxicity, Chap 04. Springer Nature, Switzerland AG
- Singh RP (2019d) Potentialities of biogenic plant-mediated copper and copper oxide nanostructured nanoparticles and their utility. In: Prasad R (ed) Nanotechnology in the life sciences: plant nanobionics, vol 2. Approaches in nanoparticles biosynthesis and toxicity, Chap 05. Springer Nature, Switzerland AG
- Singh RP, Choi JW (2010) Bio-nanomaterials for versatile bio-molecules detection technology. Adv Mat Lett 1(1):83–84
- Singh RP, Pandey AC (2011) Silver nano-sieve using 1, 2-benzenedicarboxylic acid as a sensor for detecting hydrogen peroxide. Anal Methods 3:586–592
- Singh RP, Oh BK, Koo KK, Jyoung JY, Jeong S, Choi JW (2008) Biosensor arrays for environmental pollutants detection. Biochip J 2(4):223–234

- Singh P, Onodera T, Mizuta Y, Matsumoto K, Miura N, Toko K (2009) Dendrimer modified biochip for detection of 2, 4, 6 trinitrotoluene on SPR immunosensor: fabrication and advantages. Sensors Actuators B Chem 137:403–409. https://doi.org/10.1016/j.snb.2008.12.027
- Singh RP, Kang DY, Oh BK, Choi JW (2009a) Polyaniline based catalase biosensor for the detection of hydrogen peroxide and azide. Biotech Bioprocess Eng 14(4):443–449
- Singh RP, Kim YJ, Oh BK, Choi JW (2009b) Glutathione-s-transferase based electrochemical biosensor for the detection of captan. Electrochem Commun 11:181–185
- Singh RP, Oh BK, Choi JW (2010a) Application of peptide nucleic acid towards development of nanobiosensor arrays. Bioelectrochemistry 79(2):153–161
- Singh RP, Kang DY, Choi JW (2010b) Electrochemical DNA biosensor for the detection of sanguinarine in adulterated mustard oil. Adv Mater Lett 1(1):48–54
- Singh RP, Kang DY, Choi JW (2011a) Nanofabrication of bio-self assembled monolayer and its electrochemical property for toxicant detection. J Nanosci Nanotechnol 11:408–412
- Singh RP, Shukla VK, Yadav RS, Sharma PK, Singh PK, Pandey AC (2011b) Biological approach of zinc oxide nanoparticles formation and its characterization. Adv Mater Lett 2(4):313–317
- Singh RP, Tiwari A, Pandey AC (2011c) Silver/polyaniline nanocomposite for the electrocatalytic hydrazine oxidation. J Inorg Organmetal Poly Mat 21:788–792
- Singh RP, Choi JW, Pandey AC (2012a) Smart nanomaterials for biosensors, biochips and molecular bioelectronics. In: Li S, Ge Y, Li H (eds) Smart nanomaterials for sensor application. Bentham Science Publisher (USA), Chapter 1, pp 3–41
- Singh RP, Choi JW, Tiwari A, Pandey AC (2012b) Biomimetic Materials Toward Application of Nanobiodevices, Editors: Tiwari A, Mishra A. K, Kobayashi H, Turner APF. In: Intelligent Nanomaterials: Processes, Properties, and Applications. John Wiley & Sons, Inc., Hoboken, NJ, USA. Chapter 20, pp.741–782
- Singh RP, Kumar K, Rai R, Tiwari A, Choi JW, Pandey (2012c) Synthesis, characterization of metal oxide based nanomaterials and its application in biosensing. In: Rai R (ed) Synthesis, characterization and application of smart material. Nova Science Publishers, New York. Chapter 11, pp 225–238
- Singh RP, Choi JW, Tiwari A, Pandey AC (2012d) Utility and potential application of nanomaterials in medicine. In: Tiwari A, Ramalingam M, Kobayashi H, Turner APF (eds) Biomedical materials and diagnostic devices. Wiley, Hoboken. https://doi.org/10.1002/9781118523025.ch7
- Singh RP, Choi JW, Tiwari A, Pandey AC (2014) Functional nanomaterials for multifarious nanomedicine. In: Tiwari A, Turner APF (eds) Biosensors nanotechnology. Wiley, Hoboken. https:// doi.org/10.1002/9781118773826.ch6
- Singh KRB, Sridevi P, Singh RP (2020) Potential applications of peptide nucleic acid in biomedical domain. 2(9): e12238. https://doi.org/10.1002/eng2.12238
- Snejdarkova M, Svobodova L, Nikolelis DP, Wang J, Hianik T (2003) Acetylcholine biosensor based on dendrimer layers for pesticides detection. Electroanalysis 15(14):1185–1191. https:// doi.org/10.1002/elan.200390145
- Solanki PR, Kaushik A, Agrawal VV, Malhotra BD (2011) Nanostructured metal oxide-based biosensors. NPG Asia Mat 3(1):17–24. https://doi.org/10.1038/asiamat.2010.137
- Su PG, Huang LN (2007) Humidity sensors based on TiO₂ nanoparticles/polypyrrole composite thin films. Sensors Actuators B Chem 123(1):501–507. https://doi.org/10.1016/j.snb.2006.09.052
- Sugunan A, Thanachayanont C, Dutta J, Hilborn JG (2005) Heavy-metal ion sensors using chitosan-capped gold nanoparticles. Sci Technol Adv Mat 6(3–4):335–340. https://doi. org/10.1016/j.stam.2005.03.007
- Suri K, Annapoorni S, Sarkar AK, Tandon RP (2002) Gas and humidity sensors based on iron oxide–polypyrrole nanocomposites. Sensors Actuators B Chem 81(2–3):277–282. https://doi. org/10.1016/S0925-4005(01)00966-2
- Tanimoto de Albuquerque YD, Ferreira LF (2007) Amperometric biosensing of carbamate and organophosphate pesticides utilizing screen-printed tyrosinase-modified electrodes. Anal Chim Acta 596(2):210–221. https://doi.org/10.1016/j.aca.2007.06.013

- Ullah N, Mansha M, Khan I, Qurashi A (2018) Nanomaterial-based optical chemical sensors for the detection of heavy metals in water: recent advances and challenges. TrAC Tr Anal Chem 100:155–166. https://doi.org/10.1016/j.trac.2018.01.002
- Vazquez-Gonzalez M, Carrillo-Carrion C (2014) Analytical strategies based on quantum dots for heavy metal ions detection. J Biomed Opt 19(10):101503. https://doi.org/10.1117/1. jbo.19.10.101503
- Viswanathan S, Wu L, Huang MR, Ho JA (2006) Electrochemical immunosensor for cholera toxin using liposomes and poly(3,4-ethylenedioxythiophene)-coated carbon nanotubes. Anal Chem 78(4):1115–1121. https://doi.org/10.1021/ac051435d
- Wanekaya AK, Chen W, Mulchandani A (2008) Recent biosensing developments in environmental security. J Environ Monit 10(6):703. https://doi.org/10.1039/b806830p
- Wang HF, He Y, Ji TR, Yan XP (2009) Surface molecular imprinting on Mn-doped ZnS quantum dots for room-temperature phosphorescence optosensing of pentachlorophenol in water. Anal Chem 81(4):1615–1621. https://doi.org/10.1021/ac802375a
- Wei Y, Yang R, Yu XY, Wang L, Liu JH, Huang XJ (2012) Stripping voltammetry study of ultratrace toxic metal ions on highly selectively adsorptive porous magnesium oxide nanoflowers. Analyst 137(9):2183. https://doi.org/10.1039/c2an15939b
- Wu H, Liang J, Han H (2008) A novel method for the determination of Pb²⁺ based on the quenching of the fluorescence of CdTe quantum dots. Microchim Acta 161(1–2):81–86. https://doi. org/10.1007/s00604-007-0801-4
- Wu S, Zhao Q, Zhou L, Zhang Z (2014) Stripping analysis of trace arsenic based on the MnOx / AuNPs composite film modified electrode in alkaline media. Electroanalysis 26(8):1840–1849. https://doi.org/10.1002/elan.201400219
- Xu WH, Wang L, Wang J, Sheng GP, Liu JH, Yu HQ, Huang XJ (2013) Superparamagnetic mesoporous ferrite nanocrystal clusters for efficient removal of arsenite from water. Cryst Eng Commun 15(39):7895. https://doi.org/10.1039/c3ce40944a
- Yang M, Han A, Duan J, Li Z, Lai Y, Zhan J (2012) Magnetic nanoparticles and quantum dots coloaded imprinted matrix for pentachlorophenol. J Hazard Mater 237–238:63–70. https://doi. org/10.1016/j.jhazmat.2012.07.064
- Yavuz CT, Mayo JT, Yu WW, Prakash A, Falkner JC, Yean S, Colvin VL (2006) Low-field magnetic separation of monodisperse Fe₃O₄ nanocrystals. Science 314(5801):964–967. https://doi. org/10.1126/science.1131475
- Yılmaz F, Ozdemir N, Demirak A, Tuna AL (2007) Heavy metal levels in two fish species Leuciscuscephalus and Lepomisgibbosus. Food Chem 100(2):830–835. https://doi. org/10.1016/j.foodchem.2005.09.020
- Zelada-Guillen GA, Riu J, Düzgün A, Rius FX (2009) Immediate detection of living bacteria at ultralow concentrations using a carbon nanotube based potentiometric aptasensor. Angew Chem Int Ed 48(40):7334–7337. https://doi.org/10.1002/anie.200902090
- Zhang X, Zeng T, Hu C, Hu S, QiulinTian Q (2016) Studies on fabrication and application of arsenic electrochemical sensors based on titanium dioxide nanoparticle modified gold strip electrodes. Anal Methods 8(5):1162–1169. https://doi.org/10.1039/C5AY02397A
- Zhao X, Lv L, Pan B, Zhang W, Zhang S, Zhang Q (2011) Polymer-supported nanocomposites for environmental application: a review. Chem Eng J 170(2–3):381–394. https://doi.org/10.1016/j. cej.2011.02.071
- Zhao ZQ, Chen X, Yang Q, Liu JH, Huang XJ (2012) Selective adsorption toward toxic metal ions results in selective response: electrochemical studies on a polypyrrole/reduced graphene oxide nanocomposite. Chem Commun 48(16):2180–2182. https://doi.org/10.1039/C1CC16735A
- Zhao Q, Rong X, Ma H, Tao G (2013) Dithizone functionalized CdSe/CdS quantum dots as turnon fluorescent probe for ultrasensitive detection of lead ion. J Hazard Mater 250–251:45–52. https://doi.org/10.1016/j.jhazmat.2013.01.062
- Zhao L, Gu W, Zhang C, Shi X, Xian Y (2016) In situ regulation nanoarchitecture of Au nanoparticles/reduced graphene oxide colloid for sensitive and selective SERS detection of lead ions. J Colloid Interf Sci 465:279–285. https://doi.org/10.1016/j.jcis.2015.11.073

- Zouboulis AI, Katsoyiannis IA (2002) Arsenic removal using iron oxide loaded alginate beads. Ind Eng Chem Res 41(24):6149–6155. https://doi.org/10.1021/ie0203835
- Zheng J, Li G, Ma X, Wang Y, Wu G, Cheng Y (2008) Polyaniline–TiO2 nano-composite-based trimethylamine QCM sensor and its thermal behavior studies. Sensors Actuators B Chem 133:374–380. https://doi.org/10.1016/j.snb.2008.02.037

Chapter 12 Nanosensors Based on Lipid Membranes for the Rapid Detection of Food Toxicants



Georgia-Paraskevi Nikoleli, Dimitrios P. Nikolelis, Christina G. Siontorou, Marianna-Thalia Nikolelis, Stephanos Karapetis, and Spyridoula Bratakou

Contents

12.1	Introduction		248
12.2	Methods for Construction of Nanosensors Based on Lipid Membranes		249
	12.2.1	Metal-Supported Lipid Layers	249
	12.2.2	Stabilized Lipid Films on a Glass Fiber Filter	251
	12.2.3	Polymer-Supported Bilayer Lipid Membranes	252
	12.2.4	Polymer Lipid Films Supported on Graphene Microelectrodes	253
12.3	Applications of Lipid-Film-Based Nanosensors for the Rapid Detection		
	of Food Toxicants		254
12.4	Conclusions and Future Prospects		258
Refere	References		

Abstract The application of lipid film biosensor devices has created the opportunity to reconstitute a considerable part of their functionality to detect traces of food toxicants. Nanotechnology-enabled sensor miniaturization has extended the range of biological moieties that could be immobilized within a lipid bilayer device. This chapter reviews the recent progress in biosensor technologies for food-quality monitoring. A large number of applications are described herein, placing emphasis on size reduction of novel systems of lipid film devices, new sensing techniques, and nanotechnology-based transduction schemes. The range of analytes that can be

D. P. Nikolelis (🖂) · M.-T. Nikolelis

Laboratory of Environmental Chemistry, Department of Chemistry, University of Athens, Athens, Greece

e-mail: nikolelis@chem.uoa.gr

C. G. Siontorou

G.-P. Nikoleli · S. Karapetis · S. Bratakou

Laboratory of Inorganic & Analytical Chemistry, School of Chemical Engineering, Dept 1, Chemical Sciences, National Technical University of Athens, Athens, Greece

Laboratory of Simulation of Industrial Processes, Department of Industrial Management and Technology, School of Maritime and Industry, University of Piraeus, Piraeus, Greece

[©] The Editor(s) (if applicable) and The Author(s), under exclusive licence to Springer Nature Switzerland AG 2021

V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8_12

currently detected include insecticides, pesticides, herbicides, toxins, hormones, dioxins, and genetically modified foods. Technology limitations and future prospects are discussed, with a focus on the commercialization capabilities of the proposed sensors.

Keywords Nanosensors · Enzyme-based systems · Antibody-based systems · Receptor-based systems · Food analysis · Nanotechnology

12.1 Introduction

A biosensor is a device that transforms chemical information into an analytically useful signal. Biosensors have two components: a biological recognition element ("receptor", enzyme, antibody, natural receptor, cell, etc.) and a physical transducer. The recognition system provides the chemical information (i.e., concentration of the analyte) as a physical output signal. The physical transducer transfers the signal to an electrical, optical, or piezoelectric output, among others; the biological recognition element is in direct contact with a transduction element. Biosensors have made a large impact in the area of food applications and provide many advantages as compared to standard analytical techniques (i.e., chromatographic) such as miniaturization, no sample preparation, real-time detection, rapid response times, portability, high sensitivity, and selectivity.

Recent progress in nanotechnology has created the opportunity to mass-produce affordable devices and to integrate these into systems for market applications for the detection and monitoring of food toxicants. These applications include a wide range of food toxicants such as toxins, insecticides, pesticides, herbicides, viruses and other microorganisms, dioxins and polycyclic aromatic hydrocarbons (PAHs), hydrazines, and hormones.

Lipids are amphiphilic molecules, which possess both hydrophilic head groups and hydrophobic chains. Since the discovery of model bilayer lipid membranes (BLM) by Mueller et al. (Mueller et al. 1962), there have been various attempts to use lipid films to construct biosensing devices for applications to detect and monitor food toxicants. However, free-standing BLMs were very fragile and, therefore, not suitable for long-term use. They had low mechanical and electrical stability, which was the main obstacle to their practical application. The recent advances in the construction of stabilized lipid film devices have created the opportunity to prepare devices to detect a large diversion of food toxicants in real samples. Lipidmembrane-based biosensors represent an appropriate biocompatible structure with fast response times (on the order of a few seconds) and high sensitivity (i.e., nanomolar detection limits) and selectivity, small size, and offers many advantages compared with the bulky analytical instrumentation such as liquid chromatographic units. Most of these biosensors are cost efficient, easy to use, fast responding portability, and are good alternative to the expensive, bulky, time-consuming standard analytical methods (i.e., chromatographic techniques). The new generation of lipid membrane nanosensors has improved analytical performance and operational stability.

This work reviews the construction of stabilized lipid film nanosensors that are used to detect food toxicants and facilitate state-of-the-art design and microfabrication of prototype lipid membrane nanosensing devices: for rapid, in-the-field detection of food toxicants and the challenges that lie ahead. It also describes a wide range of applications of lipid-film-based biosensors in food analysis for the rapid detection of urea, cholesterol, glycose, cholera toxin, aflatoxins, naphthalenic acid, dioxins, etc.

12.2 Methods for Construction of Nanosensors Based on Lipid Membranes

A large number of methods were reported in the literature in the last two decades for the construction of stabilized lipid films that are not prone to electrical or mechanical breakage and provide devices that can be used for practical applications. Most of these methods provide lipid membranes with a size less than 1 μ m and, therefore, can be considered nanosensors. These biosensors have been used for electrochemical experimentation and belong, therefore, in electrochemical devices. An exception is the development of stabilized polymerized lipid films on a filter paper that switch on and off their fluorescence and therefore belong to optical biosensors. Below we provide the various techniques for the preparation of nanosensors based on lipid membranes.

12.2.1 Metal-Supported Lipid Layers

A simple system for the preparation of a self-assembled lipid bilayer membrane (sBLM) at the freshly cut tip of a Teflon-coated metallic wire was reported by Tien and Salamon (Tien and Salamon 1989). The technique of formation of sBLMs is based on the interaction of an amphiphatic lipid molecule with a nascent metallic surface. One end of a Teflon-coated stainless steel metal wire (with diameter of 0.1–0.5 mm) was immersed in a lipid solution in chloroform and then, while still immersed, the tip is cut off with a miniature guillotine. Then, the fresh tip of the wire becomes coated with the lipid membrane and was placed in the electrolyte (usually 0.1 M KCl), whereupon the lipid film spontaneously thinned, forming a self-assembled lipid bilayer membrane (sBLM). A schematic of the sensor, measurement setup, and lipid self-assembly process is shown in Fig. 12.1.

sBLMs have been fully characterized (Tien and Salamon 1989; Nikolelis et al. 1996; Siontorou et al. 1997a). The diameter and composition of the wires that were used were found to have an important role in the time taken for device stabilization



Fig. 12.1 Schematic of the sensor, measurement setup, and lipid self-assembly process (not drawn to scale): (**a**) the sensing electrode is tipped with a scalpel and immediately immersed in the lipid solution before being dipped in the electrolyte solution. (**b**) The electrochemical setup consists of a 20-mL cell and a two-electrode configuration, that is, the sensing electrode and a Ag/AgCl reference electrode, placed in a grounded Faraday cage; an external DC potential of 25 mV is applied between the electrodes and the ionic current through the BLM is measured with a digital electrometer; the cell is stirred using a magnetic stirrer. (**c**) Upon immersion, the lipid droplet attached to the wire is self-assembled into a bilayer that has one layer adsorbed on the metal surface and the other facing the aqueous solution. (**d**) Recording of the ion current decreases during the self-assembly process; recording started at the immersion of the sensing electrode in the electrolyte solution [Reprinted from Nikoleli et al. 2018)

and on the magnitude and noise of the background ion current (Nikolelis et al. 1996; Siontorou et al. 1997a). The use of a wire of 0.25 mm diameter and decane as a solvent should be avoided because the lipid films are so-called "black lipid membrane" and these films do not provide reproducible results. Silver wires of 0.5 and 1.0 mm diameter provide BLMs which were mechanically and electrical stable for more than 48 hours.

12.2.2 Stabilized Lipid Films on a Glass Fiber Filter

The construction of stabilized lipid membranes on ultrafiltration glass fiber filters has been provided in the literature (Nikolelis et al. 1995). These stabilized lipid membranes on glass fiber filters have allowed for their practical application in real samples, for example, for the determination of aflatoxin M_1 in milk and milk preparations (Andreou and Nikolelis 1998). The lipid membrane is formed on a microporous filter glass fiber disk (Nikolelis et al. 1995; Andreou and Nikolelis 1998). The filters and (nominal) pore size used were GF/F glass microfiber, 0.7 µm (Whatman Scientific Ltd., Kent, U.K.).

The experimental setup which is used for the construction of stabilized BLMs consists of two Plexiglas chambers which are separated by a Saran-Wrap partition (thickness of ca. 10 µm) (Nikolelis et al. 1995; Andreou and Nikolelis 1998). This plastic partition was cut to more than twice the size of the contact area of the surfaces of the chambers and was folded in half; then, a hole having a diameter of 0.32 mm was punched through the double layer of the plastic film with a perforation tool. A microporous glass GF/F microfiber disk with a diameter of 0.9 cm and nominal pore size of 0.7 µm is placed between the two plastic layers. The partition containing the ultrafiltration glass fiber was then clamped between the two Plexiglas chambers. One of the chambers consisting an electrochemical cell had a circular shape of diameter 1.0 cm and depth 0.5 cm; this chamber was connected with plastic tubing which was used for the flow of the carrier solution. An Ag/AgCl reference electrode was immersed in the waste of the carrier electrolyte solution. The second chamber was cylindrical and had its longitudinal axis perpendicular to the flow of the carrier solution. The upper hole of this cell was circular with a surface area of about 0.2 cm² and the lower was elliptical with diameters 0.5 and 1.4 cm parallel and vertical to the flow of the carrier electrolyte solution, respectively. The lower hole was facing the opposing cell. An Ag/AgCl reference electrode was positioned at the center of the cylindrical cell. An external voltage of 25 mV d.c. is applied between the two reference electrodes. A Keithley digital electrometer is used as a current-to-voltage converter. A peristaltic pump is used for the flow of the carrier electrolyte. Injections of the samples are made with a Hamilton repeating dispenser. The electrochemical cell and electronic equipment were isolated in a grounded Faraday cage. A simple scheme of the apparatus used is presented in Fig. 12.2. The procedure for the formation of the stabilized BLMs is as follows: (Nikolelis et al. 1995; Andreou and Nikolelis 1998): Lipid solution (ca. 10 µL) is put dropwise with a microliter syringe to the water/air interface in the cylindrical cell near the partition. The level of the electrolyte solution is dropped below the 0.32 mm hole and then raised again. The formation of the bilayer structure of the lipid membranes has to be verified by the ion current magnitude and by the electrochemical characterization using gramicidin D. This peptide has the size of a bilayer and is able to cross through only in a layer having the bilayer structure.



Fig. 12.2 Schematic of the experimental setup; the micromachined chambers are separated by a thin (12.5 μ m thick) polyvinylidene chloride wrap and enclose the microfiber disk. For more details, see text (From Nikoleli et al. 2018)

12.2.3 Polymer-Supported Bilayer Lipid Membranes

The preparation of polymer stabilized was recently reported in the literature. These lipid-membrane-based nanosensors were constructed by using UV irradiation instead of heating the lipid mixture to 60 °C (Nikolelis et al. 2006, 2008a) Thus, this process retains the activity of a "receptor" (enzyme, antibody, ion-channel receptor), whereas heating to 60 °C can deactivate an enzyme or antibody or a receptor. The results have shown that the polymerization is completed within 4 hours; physicochemical methods such as DSC, IR, or Raman spectrophotometry were used to exploit the polymerization time. The preparation of these devices makes possible the practical use of biosensors based on lipid membranes for chemical sensing, because it allows for the incorporation of a "receptor" molecule in these lipid films and, more importantly, these devices are stable in the air for more than 48 h.

The preparation of these stabilized lipid films is as follows (Nikolelis et al. 2006, 2008a): Briefly, 0.8 mL of a mixture containing 4% w/v egg phosphatidylcholine (PC) in n-hexane (this solvent evaporates so that these films do not retain the solvent in their structure) were mixed with 0.07 mL of methacrylic acid, 0.8 mL of ethylene glycol dimethacrylate, 8 mg of 2,2'-azobis-(2-methylpropionitrile), and 1.0 mL of acetonitrile. The mixture was sparged with nitrogen for about 1 min and sonicated for 30 min. For the preparation of the stabilized lipid films, 0.15 mL of this mixture was spread on a microfilter (microporous glass GF/F microfiber disk with a diameter of ca. 0.9 cm and nominal pore size of 0.7 μ m). The filter with the mixture was then irradiated using a UV deuterium lamp. Raman spectrometry and differential scanning calorimetry (DSC) were used to monitor the kinetics of the polymerization

process. The measuring setup was similar to that presented in Fig. 12.2. These membranes were stable in storage in air for repetitive use.

12.2.4 Polymer Lipid Films Supported on Graphene Microelectrodes

Graphene nanomaterials have received tremendous interest in the field of basic research and in technological applications due to their unique physicochemical properties, that is, good sensing ability, and excellent mechanical, thermal and electrical properties; other advantages include large surface-to-volume ratio, excellent biocompatibility, high electron-transfer rates, non-toxicity, and bio-safety. The development of nanosensors based on graphene microelectrodes is an interesting application of this field of nanotechnology. Their implementation in electrochemical biosensing is quite beneficial as it allows for miniaturization, increases the speed of response, decreases the detection limits, and solves the biocompatibility and biofouling problems. Several examples in the development of nanobiosensors by integrating enzymes and antibodies were recently described in literature. Stabilized lipid films were wrapped around a copper wire containing graphene nanosheets (Nikoleli et al. 2012; Bratakou et al. 2015). These nanosensors have been implemented in the rapid detection of food toxicants in real samples, such insecticides (Bratakou et al. 2015), naphthalene acetic acid (Bratakou et al. 2016), cholera toxin (Karapetis et al. 2016), and saxitoxin (Bratakou et al. 2017).

The preparation of graphene microelectrodes was as follows (Nikoleli et al. 2012; Bratakou et al. 2015, 2016, 2017; Karapetis et al. 2016): A homogeneous graphene dispersion (~0.4 mg/mL) has been obtained in *N*-methyl-pyrrolidone (NMP) through mild sonication for 180 hours and centrifugation at 700 rpm for 2 h. This suspension has been poured onto a copper wire (d = 0.25 mm) mounted on a glass fiber filter and evaporation of the organic solvent has been carried out using a fan heater. This copper wire has been used to establish the connection for the extraction of voltage signals for the calibration curve. Thus, a simplistic approach of dropwise dispersion of graphene suspended in NMP solution has been used to scatter the graphene nanosheets on the copper wire. The extended sonication time results in a good fraction of monolayer sheets but with smaller lateral sizes.

The procedure of construction of these devices is in brief as follows (Nikoleli et al. 2012; Bratakou et al. 2015, 2016, 2017; Karapetis et al. 2016): Stabilized lipid films were prepared by polymerization with a procedure similar to that previously described (Nikolelis et al. 2006; 2008a). Briefly, 0.15 mL of a mixture containing 5 mg of a mixed lipid powder composed of 35 (w/w) DPPA and 65 (w/w) of DPPC (1.75 mg DPPA and 3.25 mg DPPC) was mixed with 0.070 mL of methacrylic acid, 0.8 mL of ethylene glycol dimethacrylate, 8 mg of 2,2'-azobis-(2-methylpropionitrile), and 1.0 mL of acetonitrile. DPPC is used as lipid and not PC, which can be oxidized by air and does not provide reproducible results. The

mixture was spumed with nitrogen for about 1 min and sonicated for 30 min. This mixture could be stored in the refrigerator. For the preparation of the stabilized lipid films, 0.15 mL of this mixture was spread on the glass filer microfilter. The filter with the mixture was then irradiated using a UV deuterium lamp. Raman spectrometry was used to monitor the kinetics of the polymerization process (Nikolelis et al. 2006; Nikolelis et al. 2008a).

The enzyme, antibody or receptor ("receptor") were incorporated in these BLMs before polymerization by spreading 15 μ L of the "receptor" suspension with the polymerization mixture (i.e., for the preparation of the stabilized lipid films, 0.15 mL of the polymerization mixture and 15 μ L of "receptor" suspension were spread on the microfilter). The preparation of the polymerized lipid film onto the copper wire containing graphene nanosheets.

12.3 Applications of Lipid-Film-Based Nanosensors for the Rapid Detection of Food Toxicants

A work was described for the rapid and sensitive electrochemical flow injection monitoring of aflatoxin M1(AFM1) in cheese samples (Andreou and Nikolelis 1998). Stabilized filter-supported bilayer lipid membranes (BLMs) were used as detectors. Single-stranded DNA oligomers terminated with alkyl chains (dT20-C16) were incorporated into the membranes to control surface electrostatic properties. The incorporation of dT20-C16in BLMs lowered the detection limit for the detection of this toxin by one to four orders of magnitude as compared with the detection limit obtained in the absence of DNA. Therefore, it is now possible to continuously monitor this toxin at concentrations that approached those that could be of interest as set by the U.S. Food and Drug Administration and most European countries. The work that was described takes a significant step toward the development of a detector of greater practical potential by demonstrating that the incorporation of C16ssDNA into lipid membranes results in a combination of properties that provides for a much more sensitive and robust detection system. Injections of AFM1 were made into flowing streams of a 0.1 M KC1 electrolyte solution, and a transient current signal with duration of seconds reproducibly appeared in about 12 s after exposure of the detector element to the toxin. The magnitude of this signal was linearly related to the concentration of AFM1 with detection limits at subnanomolar level. The effect of interferents such as proteins and lipids was investigated. It was determined that interferences from proteins could be eliminated by adjustment of the {low rate of the carrier electrolyte solution. The technique was applied for the rapid flow injection determination of aflatoxin MI in cheese samples. AFM1 could be determined in continuous flowing systems with a rate of at least 4 samples min⁻¹. Repetitive cycles of injection of AFM1 have shown no weakning of signal upto 30 cycles of detection.

A work that was reported in the literature describes a technique for the rapid and sensitive electrochemical flow injection monitoring and analysis of mixtures of the artificial sweeteners acesulfame-K, cyclamate, and saccharin using stabilized systems of filter-supported bilayer lipid membranes (BLMs) (Nikolelis and Pantoulias 2001). Injections of artificial sweeteners were made into flowing streams of a carrier electrolyte solution, and a transient current signal with duration of seconds reproducibly appeared in <1 min after exposure of the lipid membranes to the artificial sweeteners. The magnitude of this signal was linearly related to the concentration of artificial sweeteners, which could be determined at micromolar levels. Repetitive cycles of injection of artificial sweeteners have shown no signal degradation during each cycle (30 sequential injections). The time of appearance of the transient response was different for each artificial sweetener and increased in the order of cyclamic acid, acesulfame-K, and saccharin. The difference in time of response has allowed selective detection and analysis of these artificial sweeteners in mixtures. The effect of potent interferences, including a wide range of compounds usually found in foods, proteins, and lipids, was investigated. The results showed no interferences from these constituents of real food samples. The major interference from proteins (most common in lipid-film-based biosensors) can be eliminated by modulation of the carrier solution that does not allow adsorption of these compounds in BLMs. The technique was applied in real food samples, that is, in artificial sweetener tablets, diet soft drinks, wines, and yogurts that contain mixtures of these artificial sweeteners with aspartame and other compounds. A comparison of results using the present method and that of an official method of analysis showed good agreement between the two methods.

A work was reported for the rapid detection of vanillin in alcoholic beverages using stabilized lipid-membrane-based biosensors (Nikolelis and Theoharis 2002). Microporous filters composed of glass fibers (nominal pore sizes 0.7 and 1.0 μ m) were used as supports for the polymerization of the lipid film and stabilization of these devices. The lipid film is formed on the filter by polymerization before its use. Methacrylic acid was the functional monomer, ethylene glycol dimethacrylate was the crosslinker, and 2,2'-azobis-(2-methylpropionitrile) was the initiator. The response toward vanillin of the present stabilized lipid membrane biosensors composed of phosphatidylcholine was investigated. The stabilized lipid membranes provided artificial ion-gating events in the form of transient signals and can be used again after storage in air. This has allowed for the practical use of the technique for chemical sensing based on lipid films for the rapid detection of vanillin in wines and alcoholic beverages.

The stabilized supported on glass fiber lipid film nanosensors were used for the flow injection analysis (FIA) of pesticides (Nikolelis et al. 2005). Carbofuran was chosen as a typical pesticide. The determination of this pesticide is based on the degree of inhibition and the reactivation of enzyme by injections of substrate. Carbofuran was determined at concentration levels of 10^{-7} to 10^{-9} M. The investigation of the effect of interferences included compounds usually found in foods, proteins, and lipids. The results have shown no interferences from these compounds. The technique was applied to fruits, vegetables, and dairy samples. The recovery

rate ranged between 96 and 106%, which confirms there is no interferences due to matrix effects.

A paper was reported in the literature using a synthetic "receptor" immobilized on supported lipid films on glass fiber filters (Nikolelis et al. 2008b). The supported lipid films were modified by calixarenes and permitted sensitive and rapid determination of insecticides in fruits and vegetables (Nikoleli et al. 2013).

A potentiometric urea lipid-film-based minisensor on graphene nanosheets in which a polymeric lipid membrane was deposited has been recently reported in the literature [10]. The structural characteristics of graphene nanosheets have been studied through atomic force microscopy (AFM) and transmission electron microscopy (TEM) measurements. UV-Vis and Fourier transform IR (FTIR) spectroscopy have been used to study the pre- and postconjugated surfaces of graphene nanosheets. The presented potentiometric urea biosensor shows good reproducibility, reusability, selectivity, fast response times (~4 s), long shelf life, and high sensitivity with a slope of ca. 70 mV/decade over the urea logarithmic concentration range from 1×10^{-6} M to 1×10^{-3} M.

A novel potentiometric cholesterol biosensor has been fabricated by immobilizing the stabilized polymeric lipid membrane onto a graphene electrode (Nikoleli et al. 2013). The stabilized polymeric lipid membrane is composed of a cholesterol oxidase enzyme and polymerization mixture, which holds paramount influence in the properties of the cholesterol biosensor. The presented biosensor reveals appreciable reproducibility, good selectivity, and high sensing capability having a linear slope curve of ~64 mV per decade. The strong biocompatibility among stabilized polymeric lipid membranes and human biofluids offers the possibility to use for real blood samples and other biological applications.

The electrochemical interactions of naphthalene acetic acid (NAA) with stabilized lipid films supported on a methacrylate polymer on a glass fiber filter with incorporated auxin-binding protein 1 receptor were exploited to develop a nanosensor for the rapid determination of NAA in fruits and vegetables (Nikolelis et al. 2008c). An FIA technique was used; NAA was injected into the flowing streams of a carrier electrolyte solution, the flow of the electrolyte solution stops and an ion current transient was obtained, the peak height of which was correlated to the hormone concentration (with μ M detection limits). The response times were rapid (on the order of 5 min). The effect of interferences included a wide range of compounds. The results showed no interferences from these compounds in concentration levels usually found in real samples. The method was applied for the determination of NAA in fruits, and vegetables and the reproducibility of the method was satisfactory. Thus, a quantitative method for the detection of NAA in fruits and vegetables that can be complimentary to HPLC methods was attained.

A potentiometric carbofuran minisensor on graphene nanosheets with incorporated lipid membranes has been described in the literature (Bratakou et al. 2015). The graphene electrode was used to develop a selective and sensitive chemical sensor for the detection of carbofuran by incorporating an artificial selective receptor (resorcin[4]arene receptor) on stable lipid films. This chemical nanosensor can detect carbofuran concentrations at nM concentration range, with rapid response times of ca. 20 s, is easy to construct, and exhibits good reproducibility, reusability, selectivity, long shelf life, and a high electrode slope of ca. 59 mV/decade over the carbofuran logarithmic concentration range from 10^{-6} to 10^{-3} M.

A work that describes a method for the rapid and sensitive electrochemical flow injection monitoring (FIA) and analysis of mixtures of the triazine herbicides simazine, atrazine, and propazine using stabilized systems of filter-supported BLMs consisted of egg PC and DPPA has been described in the literature (Nikolelis and Siontorou 1996; Siontorou et al. 1997b). Injections of these herbicides were made into flowing streams of a carrier electrolyte solution, and a transient current signal with a duration of seconds reproducibly appeared in less than 2 min after exposure of the lipid membranes to the herbicides. The magnitude of the peak was linearly related to the concentration of the herbicide have shown no signal degradation during each cycle. The time of appearance of the transient signal was different for each triazine and increased to the order of simazine, atrazine, and propazine, which has allowed selective detection and analysis of these triazines in mixtures.

A paper appeared in the literature that describes the investigations of electrochemical interactions of cholera toxin with stabilized lipid films on a polymer over a glass fiber Whatman GF/F filter with incorporated ganglioside GM1; this has led to the development of a minisensor for cholera toxin (Karapetis et al. 2016). The analyte was injected into the flowing streams of a carrier electrolyte solution. The flow of the solution stopped for 5 min and an ion current transient was obtained; the peak height of this transient was correlated to the cholera toxin concentration and had detection limits of 0.06 μ M.

A potentiometric cholera toxin minisensor on graphene nanosheets with incorporated lipid films has been described in the literature (Nikoleli et al. 2011). Ganglioside GM1 (a natural cholera toxin receptor) was incorporated on stabilized lipid films on graphene electrodes, providing adequate selectivity for the detection of cholera toxin over a wide range of concentrations, rapid response time of ca. 5 min, and detection limits of 1 nM. The proposed sensor is easy to construct and exhibits good reproducibility, reusability, selectivity, long shelf life, and a slope of ca. 60 mV/ decade of toxin concentration. The method was evaluated, implemented, and validated in lake water samples. This novel ultrathin film technology is currently adapted to the rapid detection of other toxins and could be used as a weapon against bioterrorism.

A potentiometric saxitoxin minisensor based on graphene nanosheets with incorporated lipid films and immobilized anti-STX (which is the natural saxitoxin receptor) on stabilized lipid films was recently reported in the literature (Bratakou et al. 2017). A good selectivity and sensitivity for the detection of saxitoxin, fast response times of ca. 5–20 min, and detection limits of 1 nM were observed. The proposed minisensor is easy to construct and exhibits good reproducibility, reusability, selectivity, long shelf life, and a slope of ca. 60 mV/decade over saxitoxin concentration. The method was evaluated and validated in lake water and shellfish samples. This novel ultrathin film technology is currently adapted to the rapid detection of other toxins that could be used as weapons against bioterrorism.

12.4 Conclusions and Future Prospects

The present paper describes the various routes for the preparation of nanosensors based on a lipid film technology for environmental analytical applications. Recent technological advances include the construction of stabilized supported lipid films on graphene nanoelectrodes with an incorporated "receptor" (enzyme, antibody, or natural or artificial receptor) stable in air that can be portable for in-the-field applications. These sensors reveal detection limits in the nanomolar range. The most important aspect of the present efforts is to provide a commercial portable unit that can be used for in-field and market applications.

The results have shown that a diversity of lipid film-based nanosensors can be reused after storage in air even after a period of a couple of months, and can be reproducibly fabricated with simplicity and low cost. These nanosensors have rapid response times, are easy to construct, and have a lower cost than those based on chromatographic techniques and can be used as rapid portable detectors complimentary to these methods for in-field and market measurements in food and for environmental monitoring.

Recently advanced lithography will enable the fabrication of nanopores and the insertion of nanoparticles in thin membranes where artificial bilayers can be assembled. This process accommodates a wide range of lipid composition in stable form, with the inclusion of membrane proteins. Therefore, the application of nanotechnology to this field of lipid film technology will allow miniaturization and will result in mass production of sensors. Producing the smaller patterns will enable sensors to respond faster, with a higher degree of sensitivity, and at lower production costs. Development of sensors using the present technologies will offer improved sensitivity for detection with high specificity at the molecular level, with an increment of several order of magnitude over currently available techniques with a large number of applications in environmental monitoring.

References

- Andreou VG, Nikolelis DP (1998) Flow injection monitoring of aflatoxin M₁ in milk and milk preparations using filter-supported bilayer lipid membranes. Anal Chem 70:2366–2371
- Bratakou S, Nikoleli G-P, Nikolelis DP, Psaroudakis N (2015) Development of a potentiometric chemical sensor for the rapid detection of carbofuran based on air stable lipid films with incorporated calix[4]arene phosphoryl receptor using graphene electrodes. Electroanalysis 27:2608–2613
- Bratakou S, Nikoleli G-P, Siontorou CG, Nikolelis DP, Tzamtzis N (2016) Electrochemical biosensor for naphthalene acetic acid in fruits and vegetables based on lipid films with incorporated auxin-binding protein receptor using graphene electrodes. Electroanalysis 28:2171–2177
- Bratakou S, Nikoleli G-P, Siontorou GC, Nikolelis DP, Karapetis S, Tzamtzis N (2017) Development of an electrochemical biosensor for the rapid detection of saxitoxin based on air stable lipid films with incorporated Anti-STX using graphene electrodes. Electroanalysis 29:990–997

- Karapetis S, Nikoleli G-P, Siontorou CG, Nikolelis DP, Tzamtzis N, Psaroudakis N (2016) Development of an electrochemical biosensor for the rapid detection of cholera toxin based on air stable lipid films with incorporated ganglioside GM1 using graphene electrodes. Electroanalysis 28:1584–1590
- Mueller P, Rudin DO, Tien HT, Wescott WC (1962) Reconstitution of cell membrane structure in vitro and its transformation into an excitable system. Nature 194:979–980
- Nikoleli G-P, Nikolelis DP, Tzamtzis N (2011) Development of an electrochemical biosensor for the rapid detection of cholera toxin using air stable lipid films with incorporated ganglioside GM1. Electroanalysis 23(9):2182–2189
- Nikoleli G-P, Israr MQ, Tzamtzis N, Nikolelis DP, Willander M, Psaroudakis N (2012) Structural characterization of graphene nanosheets for miniaturization of potentiometric urea lipid film based biosensors. Electroanalysis 24:1285–1295
- Nikoleli G-P, Ibutopo Z, Nikolelis DP, Likodimos V, Psaroudakis N, Tzamtzis N, Willandre M, Hianik T (2013) Potentiometric cholesterol biosensing application of graphene electrode with stabilized polymeric lipid membrane. Eur J Chem 11(9). https://doi.org/10.2478/s11532-013-0285-5
- Nikoleli G-P, Nikolelis D, Siontorou CG, Karapetis S (2018) Lipid membrane nanosensors for environmental monitoring: the art, the opportunities, and the challenges. Sensors 18(1):284
- Nikolelis DP, Pantoulias S (2001) Selective continuous monitoring and analysis of mixtures of acesulfame-K, cyclamate, and saccharin in artificial sweetener tablets, diet soft drinks, yogurts, and wines using filter-supported bilayer lipid membranes. Anal Chem 73(24):5945–5952
- Nikolelis DP, Siontorou CG (1996) Flow injection monitoring and analysis of mixtures of simazine, atrazine, and propazine using filter-supported bilayer lipid membranes (BLMs). Electroanalysis 8:907–912
- Nikolelis DP, Theoharis G (2002) Rapid detection of vanillin in alcoholic beverages using stabilized polymerized lipid film based biosensors. Electroanalysis 14(23):1661–1667
- Nikolelis DP, Siontorou CG, Andreou VG, Krull UJ (1995) Stabilized bilayer-lipid membranes for flow-through experiments. Electroanalysis 7:531–536
- Nikolelis DP, Siontorou CG, Krull UJ, Katrivanos PL (1996) Ammonium ion minisensors from self-assembled bilayer lipid membranes using gramicidin as an ionophore. Modulation of ammonium selectivity by platelet-activating factor. Anal Chem 15:1735–1741
- Nikolelis DP, Simantiraki M, Siontorou GC, Toth K (2005) Flow injection analysis of carbofuran in foods using air stable lipid film based acetylcholinesterase biosensor. Anal Chim Acta 537:169–177
- Nikolelis DP, Raftopoulou G, Nikoleli G-P, Simantiraki M (2006) Stabilized lipid membrane based biosensors with incorporated enzyme for repetitive uses. Electroanalysis 18:2467–2474
- Nikolelis DP, Raftopoulou G, Chatzigeorgiou P, Nikoleli G-P, Viras K (2008a) Optical portable biosensors based on stabilized lipid membrane for the rapid detection of doping materials in human urine. Sens Actuators B Chem 130:577–582
- Nikolelis DP, Raftopoulou G, Simantiraki M, Psaroudakis N, Nikoleli G-P, Hianik T (2008b) Preparation of a selective receptor for carbofuran for the development of a simple optical spot test for its rapid detection using stabilized in air lipid films with incorporated receptor. Anal Chim Acta 620:134–141
- Nikolelis DP, Ntanos N, Nikoleli G-P, Tampouris K (2008c) Development of an electrochemical biosensor for the rapid detection of naphthalene acetic acid in fruits by using air stable lipid films with incorporated auxin-binding protein 1 receptor. Protein Peptide Lett 15:789–794
- Siontorou CG, Nikolelis DP, Krull UJ, Chiang KL (1997a) A triazine herbicide minisensor based on surface-stabilized bilayer lipid membranes. Anal Chem 69:3109–3114
- Siontorou CG, Nikolelis DP, Krull UJ, Chiang KL (1997b) A triazine herbicide minisensor based on surface-stabilized bilayer lipid membranes. Anal Chem 69(15):3109–3114
- Tien HT, Salamon Z (1989) Formation of self-assembled lipid bilayers on solid substrates. J Electroanal Chem Interfacial Electrochem 22:211–218

Index

A

Abdel-Aziz, H.M.M., v, 140-153, 158-165 Abdel-Ghany, M.F., 172 Afonso, A.S., 161 Aggregation-induced emission (AIE), v, 55-63, 65-71, 73-76, 79, 80, 83, 84, 87, 88 Agriculture, v, xiv, xv, 5, 24, 26, 139–153, 158, 159, 161, 164, 184, 185, 215, 220, 231 Aguilar, C., 82, 84 Ahmad, S.H., 119 Alocijia, E.C., 161, 162 Annavaram, V., 33 Applications, xiii, xiv, 3, 6–15, 26, 35, 37, 39-45, 54, 55, 59, 61, 62, 65, 67, 68, 70-72, 74, 77-80, 83, 84, 86-88, 99, 102-105, 112, 117-128, 140-144, 148, 150-153, 157-165, 170-178, 185, 192, 195, 197, 200, 201, 204, 221, 222, 230, 231, 233, 235-239, 248, 249, 251, 253-258 Arduini, F., 118 Athawale, A.A., 238 Avuthu, S.G.R., 7 Azmi, N.A., 119

B

Bacteria, v, 4, 35, 123, 125, 126, 158, 161, 162, 164, 170, 216, 222, 233, 234, 236 Bai, X., 202 Ban, D.K., 119 Ban, Z., 45 Bereli, N., v, 2, 3, 5–13, 15, 170–179 Bi, A., 54–88 Bionanosensors, 158–165 Biosensors, vi, 5, 6, 11, 98, 99, 102–105, 112, 118, 123–125, 141–145, 158, 161–164, 176, 177, 190, 197, 204, 220–223, 230, 231, 233–239, 248, 249, 252, 254–256 Bone, L., 162, 163 Boyaci, İ.H., 177 Bratakou, S., 98–105, 248–258

С

Cai, Z., 232 Carbon based nanobiosensors, 233-234 Castillo, G., 235 Cella, L.N., 234 Cellulose, v, 111, 117-120, 122-125, 127, 128, 238 Cerrutti, B.M., 125 Chen, B., 59 Chen, F., 238 Chen, L., 237 Chen, Y.C., 66-67 Chen, Y.Y., 12 Çimen, D., v, 2, 3, 5-13, 15, 170-179 Colorimetric sensors, v, 6, 126 Contaminants, v, 1-15, 26, 40, 141, 145, 163, 169-179, 189, 214-224, 237 Correa, D.S., 110-129

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Switzerland AG 2021 V. Kumar et al. (eds.), *Nanosensors for Environment, Food and Agriculture Vol. 1*, Environmental Chemistry for a Sustainable World 60, https://doi.org/10.1007/978-3-030-63245-8 261

D

Danielson, B., 6 Dasgupta, N., v–xv Dendrimers nanobiosensors, v Deng, H., 24–45 Denizli, A., 2, 3, 5–13, 15, 170–179 Detections, v, 1–15, 26–44, 53–88, 100, 102–105, 110, 115–120, 122–128, 141, 142, 145, 146, 151, 157–165, 169–179, 185, 189–200, 204, 213–224, 230–239, 248, 249, 253–258 Du, D., 234 Duan, H., 195 Duan, J., 232 Dudak, F.C., 177 Dugandžić, V., 34

E

- Electrochemical sensors, 4, 7, 8, 114–115, 118, 145, 172–176, 216, 218, 219 Electronic tongues, 112, 120, 128
- Environmental analysis, 6, 109–129, 232, 238, 239
- Environmental pollutants, 2, 15, 98, 231–233, 236, 238, 239
- Environments, v, xiv, 1–15, 24–26, 40–42, 44, 45, 54, 64, 68, 82, 88, 110, 122, 139–153, 161, 164, 176, 179, 199, 217, 218, 221, 223, 230, 231, 236–239

F

Facure, M.H.M., 110-129 Fang, G., 195 Fang, M., 172 Feng, B., 54-88 Feng, H.T., 68 Feng, T., 54-88 Fluorescence, v, 5, 28, 35-39, 42, 43, 55-65, 68-73, 75-79, 82, 83, 85, 86, 88, 112, 116, 119, 127, 172, 176, 190, 193, 195, 196, 198, 199, 218, 222, 232, 233, 249 Food analysis, 5, 249 Food contaminations, 170, 178, 179, 218, 219 Foods, v, xiii, xiv, 3, 6, 25, 26, 101, 103, 139-153, 158, 159, 161, 163, 164, 169-179, 184-190, 199, 200, 204, 214-224, 231, 232, 234-236, 248, 249, 253-255, 258 Food security, v, 153 Forzani, E., 10 Frost, M.S., 33, 237

G

Galaly, A.R., 33 Gan, N., 192 Gao. T., 66-67 García-Aljaro, C., 234 Geng, L., 74, 238 Ghosh, B., 127, 128 Goftman, V.V., 199 Gold, 8, 10-14, 29, 32-35, 37, 41, 43, 102, 119, 120, 124, 125, 142, 145, 160, 162, 163, 172, 176, 185, 189-196, 204, 218, 220, 222, 223, 237 Goncalves, H., 232 Goud, K.Y., 189 Grabchev, I., 235 Green materials, 117, 129 Green synthesis, xiii, 44, 119 Guan, H., 162, 164 Guittonneau, S., 235 Guleria, P., v-xv Guo, X., 238 Gupta, B.D., 10

H

Haddaoui, M., 162 Han, S.S., 120, 121 Hatamie, 63, 66-67 Hazards, 25, 26, 148, 151, 152, 170, 214 He, X., v, 24-45 Health, 2, 5, 6, 11, 12, 24, 26, 35, 45, 83, 112, 117, 120, 122, 123, 126, 142, 145-149, 151-153, 158-161, 163-165, 170, 175, 177, 184, 186-189, 204, 214-216, 218-224, 231, 234, 239 Heavy metals, v, 1-15, 24-26, 28-30, 32-37, 40-44, 53-88, 110, 111, 117-120, 128, 145, 147, 216, 219, 231-233, 237, 238 Heikal, Y.M., v. 140-153, 158-165 Herbicides, 103, 105, 141, 142, 158-160, 162-164, 176, 221, 248, 257 Hoang, C.V., 30, 31 Hong, H.-B., 123 Hossain, M.Z., 190 Huang, G., 68, 74 Huang, L.N., 238 Huang, L.T., 54-88 Huang, X.Y., 54-88 Huang, Y., 234 Hüseynli, S., 9 Hussain, M., 178 Hwang, H.-M., 24 Hwang, J.H., 7

I

Ikram, F., 30 Ishimori, Y., 105

J

Jalilzadeh, M., 11 Ji, L., 78, 80 Jiang, R., 73, 74 Jiang, Y., 59, 66–67 Joshi, K.A., 234 Jung, S.H., 80

K

Kalluri, J.R., 13 Karahalil, B., v, 214–224 Karapetis, S., 248–258 Katsoyiannis, I.A., 238 Kaushik, A., 197 Kelly, K.L., 27 Khachatryan, G., 120 Khachatryan, K., 120 Kim, G., 161, 162 Kim, M., 15 Kong, D., 190 Kumar, V., v–xv Kuppan, B., 77

L

Lee, S., 7 Legislation, 3, 141, 150-152 Li, F., 237 Li, J., 233 Li, N., 70, 73, 74 Li, P.F., 74 Li, S., 33 Li, Y., 77, 80 Lian, Y., 234 Lichtfouse, E., v-xv Limit of detection (LOD), 7, 8, 10, 11, 13, 35, 63, 64, 66–74, 77, 80–84, 87, 118, 119, 122, 124, 126, 129, 172, 173, 175-179, 193, 195, 196, 198, 199, 233, 234 Lin, L., 76, 77, 80 Lin, S.M., 64 Lipid membrane biosensors, 255 Liposomal nanobiosensors, 236–237 Liu, B., 68, 74 Liu, F., 234 Liu. G., 120 Liu, M., 54-88 Liu, R., 33

Liu, X., 7 Liu, Y., 233 Lou, H., 9 Low, S.C., 119 Lu, Y., 33 Lu, Z., 71, 74 Luo, Y., 123, 124, 202

Μ

Magnetic nanoparticles, 123, 162, 163, 185, 189-192, 195-197, 200, 204, 222 Magzoub, R., 203 Mahajan, P.G., 82, 84 Maitra, U., 77 Maity, A., 127, 128 Manubolu, M., v Mao, J., 202 Maragos, C.M., 190 Maric, T., 41 Martins, D., 110-129 Martins, G.F., 120 Matsumoto, K., 178 Migliorini, F.L., 110-129 Monitoring, v, 2, 3, 5, 6, 26, 44, 59, 101, 104, 105, 110-112, 122, 127, 128, 141, 145, 160, 172, 176, 205, 214, 218, 220, 229-239, 248, 254, 255, 257, 258 Morton, J., 233 Mosbach, K., 6 Mueller, P., 248 Mycotoxins, v, 158, 162-164, 170, 172, 173, 184-202, 204, 205, 216, 217, 222

Ν

Nanobiosensors, v, 98, 100, 105, 142, 149, 151, 152, 162, 164, 190, 229-239, 253 Nanoiets, 39-41 Nanoparticles (NPs), xiii, 10-15, 26-30, 32-34, 37, 43-45, 55-57, 61-63, 65, 73, 75–78, 82, 83, 86, 88, 119, 120, 124, 126, 128, 141, 142, 145–149, 151, 152, 158-160, 162, 163, 165, 172, 177, 185, 189-202, 204, 222-224, 233, 237-239, 258 Nanosensors, v, 9, 23-45, 53-88, 97-105, 139-153, 157-165, 177, 178, 189, 190, 195, 214–216, 218, 219, 222, 224, 230, 231, 248-258 Nanostructures, 28-32, 34, 35, 37, 42, 44, 116, 118, 127, 149, 158, 160, 190, 191, 199, 230

Nanotechnologies, xiv, 3, 26, 44, 45, 105, 140-142, 145, 146, 149-153, 158, 159, 161, 164, 185, 189, 200, 204, 214, 215, 220, 224, 230, 238, 248, 253, 258 Nanotoxicology, 45 Narayanan, K.B., 120, 121 Nie, K.X., 60, 66-67 Nikoleli, G.-P., v. vi, 98-105, 248-258 Nikolelis, D.P., 248-258 Nikolelis, M.-T., 98-105, 248-258 Niu, C.X., 63, 66-67 Niu. X., 37 Niu, Z., 127

0

Oh, S.Y., 176 Optical chemical sensors, 112, 115-116, 128 Optical sensors, 5, 11, 115-117, 120, 176-178, 195 Owino, J., 162

Р

Pan, S.F., 62, 63, 66-67 Pang, Y., 36 Pannipara, M., 85, 87 Parker, C.O., 162 Pathan, S., 83, 84 Pathogens, 122-125, 141, 142, 145, 158, 160, 161, 164, 170, 215, 216, 233, 234 Pennacchioet, 177 Pesticides, v, 4, 102, 103, 110, 118, 140-142, 145, 148, 150, 151, 158–164, 170, 176, 214-218, 220, 231-234, 236, 248, 255 Photocatalytic degradation, 185, 202, 204 Piezoelectric sensors, 4-5, 8 Plants, xiii, 15, 24, 25, 44, 140, 142, 145, 147, 148, 158, 159, 161, 164, 176, 214, 234 Psychovios, V.N., 98-105 Pumera, M., 39

0

Qu, F., 86, 87 Quantum dots (QDs), v, 37, 42, 43, 83, 120, 160, 162, 163, 185, 189, 190, 195, 198-199, 230, 232, 233 Quenching, 37, 42, 43, 55, 58, 60-65, 68-71, 73, 78, 82, 85, 88, 193, 195, 198, 199, 232, 233

R

Ram, M.K., 238

Ranjan, S., v-xv Raouafi, N., 162 Regulations, 25, 142, 151, 152, 215 Residues, 61, 163, 164, 170, 175, 177, 214-217, 220, 221, 231 Rovina, K., 175 Rudnitskaya, A., 6

S

Sadrolhosse, A.R., 10 Safety, 3, 26, 88, 139-153, 158, 165, 170, 172, 189, 204, 214-216, 219-221, 223, 224 Safran, V., 11 Şahin, F., 162 Saikrishnan, D., 124, 125 Salamon, Z., 100, 249 Salarbashi, D., 128 Sánchez, S., 39 Sanfelice, R.C., 110-129 Sarfo, D.K., 34 Sari, E., 177 Sartore, L., 8 Saylan, Y., 176 Sener, G., 12, 13 Sensors, v, vi, 1-15, 26, 37, 56-66, 68-71, 73, 74, 76, 78, 80, 82-84, 86-88, 98, 101, 103, 105, 109–129, 141–143, 145, 146, 149, 158-163, 169-179, 190-198, 204, 205, 214-216, 218-224, 231, 232, 234, 236-238, 249, 250, 256-258 Shaban, M., 33 Sharma, S., 68, 74 Shen, L.L., 7 Shi, W., 66-67 Shi, X., 57 Shiratori, S.S., 104 Singh, K.R.B., 230-239 Singh, P., 68, 74, 235 Singh, P.K., 61, 66-67 Singh, R.P., 230-239 Siontorou, C.G., 98-105, 248-258 Snejdarkova, M., 236 Soils, 12, 24, 25, 142, 146-148, 151, 158, 160, 161, 164, 172, 218, 233 Solanki, P.R., 237 Song, D., 33 Song, R., 54-88 Song, Y., 162, 164 Songa, E.A., 162, 164 Su, P.G., 238 Sugunan, A., 238 Sun, J., 87 Sun, P., 10 Sun, S., 204

Surface plasmon resonance (SPR), 10, 11, 26–28, 37, 43, 120, 176, 177, 190, 222 Suri, K., 238

Т

Tait, E., 126 Tan, D.S.Y., 118 Tang, C., 55 Teodoro, K.B.R., v, 110–129 Teow, Y., 148 Tian, Y., 78, 80 Tien, H.T., 100, 249 Tomar, K., 61, 66–67 Tothill, E., 162 Toxicology, 45, 79, 139–153 Turan, E., 162 Two-dimensional materials, 111

U

Ullah, N., 10 Usha, S., 115

V

Verm, R., 10 Veterinary drugs, 158, 159, 164, 216, 217, 220, 221 Vimala, V., 162 Viswanath, S., 162 Viswanathan, S., 161, 236

W

Wang, A., 66–67 Wang, H., 57 Wang, J., 38 Wang, K., 58, 66–67 Wang, P., 66–67 Wang, S., 37 Wang, W., 56 Wang, X., 59, 60, 66–67 Wang, Y., 161, 162 Wang, Z., 145 Waste water, 6, 9 Wei, G., 56, 66–67 Wei, Q.S., 12 Wei, W., 233 Wu, H., 232 Wu, Y., 13

Х

Xu, C., 204 Xu, X., 162 Xu, Y., 64–67 Xu, Z., 33 Xu, Z.H., 69, 74

Y

Yang, J., 69, 74 Yang, M., 87, 124 Yao, K.S., 162 Yavuz, H., 2, 3, 5–13, 15, 170–179 Yin, H., 85 Yin, Y.B., 79, 81, 87 Yola, M.L., 178 Yu, H.X., 79, 81, 84

Z

Zelada-Guillen, G.A., 234 Zhang, Q., 119, 127 Zhang, R.X., 66-67 Zhang, S., 162, 163 Zhang, W., 195 Zhang, W.J., 57 Zhang, X., 74, 80 Zhang, Y., 175 Zhang, Z., 29 Zhao, L., 237 Zhao, Q., 232 Zheng, J., 238 Zheng, W., 54-88 Zhou, H., 72 Zinoubi, K., 118 Zou, Q., 32, 34 Zouboulis, A.I., 238