

The Impact of Estrogen in the Tumor Microenvironment

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Abstract

Tumor immune escape is now a hallmark of cancer development, and therapies targeting these pathways have emerged as standard of care. Specifically, immune checkpoint signal blockade offers durable responses and increased overall survival. However, the majority of cancer patients still do not respond to checkpoint blockade immune therapy leading to an unmet need in tumor immunology research. Sexbased differences have been noted in the use of cancer immunotherapy suggesting that sex hormones such as estrogen may play an important

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role in tumor immune regulation. Estrogen signaling already has a known role in autoimmunity, and the estrogen receptor can be expressed across multiple immune cell populations and effect their regulation. While it has been well established that tumor cells such as ovarian carcinoma, breast carcinoma, and even lung carcinoma can be regulated by estrogen, research into the role of estrogen in the regulation of tumor-associated immune cells is still emerging. In this chapter, we discuss the role of estrogen in the tumor immune microenvironment and the possible immunotherapeutic implications of targeting estrogen in cancer patients.

Keywords

Tumor microenvironment (TME) · 17β-estradiol (E2) · Estrogen receptor α (ERα) or estrogen receptor β (ERβ) · Cancerassociated fibroblasts (CAFs) · Tumorassociated macrophages (TAMs) · Myeloid-derived suppressor cells $(MDSCs)$. Immune T cells · CD4+ T lymphocytes · CD8+ T lymphocytes · Regulatory T cells (Tregs) · Programmed death-1 (PD1) · Cytotoxic T-lymphocyte-associated protein 4 (CTLA4) · Immune B cells · Natural killer (NK) cells \cdot Dendritic cells (DCs) \cdot Fibroblast growth factor (FGF) · Epidermal growth factor (EGF) · Vascular endothelial growth factor (VEGF)

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2.1 Introduction

The tumor microenvironment (TME) is made up of multiple cell types beyond only tumor cells including immune cells, stromal cells including pericytes, and extracellular molecules all regulating tumor growth. These cells have been well established as mechanisms of resistance and have been targets for cancer therapy [\[1](#page-10-0)[–4](#page-10-1)]. While these therapeutic strategies have been promising, de novo, and acquired resistance leading to inevitable tumor progression remains an ongoing problem [\[5](#page-10-2)[–7](#page-10-3)]. Therefore, alternative regulatory pathways have become necessary to evaluate for possible avenues for future therapeutic research. Female gender has been suggested in retrospective meta-analyses to be associated with decreased response to checkpoint blockade therapy [[8–](#page-10-4)[10\]](#page-10-5). Given the known role of estrogen and other sex hormones effecting immune responses, these findings warrant the evaluation of estrogen signaling in the TME [\[11](#page-10-6)]. Estrogen is a steroid hormone that has many physiological functions associated with reproduction, metabolism, and even immune regulation [[12\]](#page-10-7). The main biological endogenous estrogen, 17β-estradiol (E2), is synthesized from androgens by aromatase (CYP19A1) and binds estrogen receptor α (ER α) or estrogen receptor β (ERβ) to exert its effects through both genomic and non-genomic mechanisms [\[12](#page-10-7)[–17](#page-11-0)]. Estrogen has been long established as a driver of malignancy in hormone-sensitive carcinomas such as ovarian, breast, endometrial, lung, colon, and even prostate [\[18](#page-11-1)]. The oncogenic function of ER is due to the ability of tumor cells to enable transcriptional upregulation of proliferation and cell-survival genes via growth factors such as insulin growth factor (IGF), fibroblast growth factor (FGF), and epidermal growth factor (EGF) [\[19](#page-11-2)[–23](#page-11-3)]. Therapy targeting these aspects of E2 signaling in cancer has been in use clinically for decades. These therapeutics include selective estrogen receptor modulators or degraders (SERMs or SERDs) and aromatase inhibitors (AIs), and are typically utilized in hormone-positive breast cancer, but their utility is being evaluated in other relevant solid tumors [[24\]](#page-11-4). However, most of the studies have

focused primarily on the tumoral signaling of E2, while the remainder of the TME has gone unexplored.

E2 signaling and ER expression are not limited to tumor cells but also found on immune cells where they have distinct functions of immune regulation [[25–](#page-11-5)[28\]](#page-11-6). The link between E2 and autoimmunity has been established since findings of sex disparities in patients with systemic lupus erythematosus (SLE), and multiple, current reviews outline E2 regulation of immune cell function and expansion [[29–](#page-11-7)[32\]](#page-11-8). While the link between E2 and immune regulation has been well characterized and tumor immunology is growing as a field, there is a missing connection between E2 pathways and tumor immunology. This chapter will discuss the current findings in the literature exploring the impact of E2 and tumor immunology, as well as the future therapeutic implications of targeting the E2 pathway in the cancer immunotherapy era.

2.2 E2 Signaling Pathways on Tumor Cells

While ER expression and E2 pathways are canonically associated with tumor cells from hormone-sensitive tumors such as ovarian, breast, and endometrial, there are almost 30 tumor types that are also associated with the E2 pathway [\[33](#page-11-9), [34\]](#page-11-10). These findings are also associated with changes in outcome for the disease further conveying the importance of understanding this pathway across multiple relevant tumor types. For example, nuclear ERα expression in breast cancer, ovarian cancer, or endometrial cancer is correlated with improved overall survival (OS) compared to cancer patients that are ERαnegative [[25,](#page-11-5) [35](#page-11-11)[–38](#page-12-0)], while some of the breast cancer patients that were $ER\alpha$ -positive also had increased disease burden. Conversely, cytoplasmic ERα expression in non-small-cell lung cancer (NSCLC) cells is correlated with worse OS [\[39](#page-12-1)[–41](#page-12-2)]. Aromatase and $ER\beta$ expression in tumor cells are more controversial with studies varying on whether they convey a survival benefit [[42–](#page-12-3) [47\]](#page-12-4). These mixed opinions in the literature are

possibly due to the lack of standardized and clinically validated staining ERβ antibody, as well as the multitude of ERβ splice variants and posttranslational modifications [[26,](#page-11-12) [48,](#page-12-5) [49\]](#page-12-6). While these findings are consistent with understanding E2 on tumor cells, there is still the need to evaluate the remainder of the TME.

2.3 The E2 Pathway in Tumor-Associated Stromal Cells and Immune Cells

Within the TME, ERs and aromatase are in notable concentrations in stromal and immune cells in addition to neoplastic cells (Table [2.1](#page-2-0)). A myriad of studies in the past decade have detailed key interactions between neoplastic cells and their recruited stromal cells that are responsible for tumorigenic potentiation (reviewed in [[4,](#page-10-1) [47](#page-12-4)]). Cellular architecture complicit in this potentiation is heterogeneous between and within tumor cells, but generally includes cancer-associated fibroblasts (CAFs), tumor-associated macrophages (TAMs), myeloid-derived suppressor cells (MDSCs), immune T and B cells, natural killer (NK) cells, and endothelial cells [[4\]](#page-10-1). Transitively, the association of hormonal protein expression in TME stromal and immune cells serves to underlie a potential immunomodulatory role of ER signaling in cancer biology, demonstrated by cell types listed in Table [2.1](#page-2-0).

2.3.1 Tumor-Infiltrating Lymphocytes (TIL)

There exists a notable interplay between cancer type and lymphocyte composition of the TME. It is often opposing immune infiltrates within a given primary tumor that promote neoplastic evolution and antitumor immunity [[65\]](#page-13-0). For example, CD4+ T-cell polarization has been identified as a mediator of tumor immune surveillance. Specifically, T helper 1 (Th1) T cell responses are associated with tumor suppression while T helper 2 (Th2) exhibit tumor activation via IFNγ and IL-12 upregulation and IL-4 expression, respec-

TME cell type	Cancer type			Human expression Murine expression Method of evaluation Reference	
Stromal	Breast	Aromatase	$ER\alpha$	PCR, IHC	[50, 51]
	Melanoma		$ER\alpha$	IHC	$\sqrt{511}$
	Lung		$ER\alpha$	IHC	$\left[51\right]$
	Endometrial	Aromatase		IHC	$\sqrt{52}$
CAF	Breast	$ER\alpha$		PCR	$\left[53\right]$
	Prostate	$ER\alpha$, $ER\beta$		IHC	[54, 55]
	Endometrial	$ER\alpha$, $ER\beta$		PCR	$\left[56\right]$
	Ovarian	$ER\alpha$		IHC	$\left[57\right]$
TAM	Ovarian	$ER\alpha$, $ER\beta$		IF.IHC	$\sqrt{58}$
	Breast	Aromatase		IHC, PCR	$\sqrt{59}$
	Lung	Aromatase	Aromatase	IHC	[17, 60]
MDSC	Ovarian	$ER\alpha$	$ER\alpha$	PCR, Western	$\left[57\right]$
NK cells	Breast	$ER\alpha, ER\beta$		IHC	[61]
Effector $CD4+/CD8+T$	Breast	$ER\alpha$, $ER\beta$		IHC	[27, 62]
cells	Nonmalignant				
Tregs	Cervical	$ER\alpha$		IHC	[63]

Table 2.1 Estrogen receptor (ER) and aromatase expression in stromal and immune cells in the tumor microenvironment

Table adapted from [[64](#page-13-9)]

Studies were identified by PubMed searches using keywords: ERα, ERβ, aromatase, stromal, CAF, TAM, MDSC, expression, cancer. *CAF* cancer-associated fibroblast, *TAM* tumor-associated macrophage, *MDSC* myeloid-derived suppressor cell, *IHC* immunohistochemistry, *PCR* polymerase chain reaction, *IF* immunofluorescence, *Western*: Western blotting analysis

tively [[66,](#page-13-10) [67](#page-13-11)]. Interestingly, several murine and human studies have reported an induction of Th2 response and IL-4 production in settings of elevated E2 [[29,](#page-11-7) [32](#page-11-8)]. Further support of ER's role in tumorigenesis was illuminated by a recent in silico study showing an increase in Th1 T cells, B cells, and cytotoxic T lymphocytes (CTLs) in ER-negative breast tumors relative to ER-positive breast tumors [\[68](#page-13-12)]. This study additionally saw an inverse correlation between ER activity and immune infiltration of these cell types in breast cancer tissues. The inverse correlation observed affirmed previous reports that increased TIL, specifically CD8+ T cells, in ER-negative tumors correlated with improved OS [\[68](#page-13-12), [69\]](#page-13-13). Additionally, post hoc analysis in ER-positive breast cancer patients treated with letrozole showed increased infiltration of B and Th1 cells both at the initiation and at the end of treatments [\[68](#page-13-12)].

2.3.1.1 Cytotoxic T Cells and Natural Killer Cells

Granule-mediated exocytosis of serine proteases, such as granzyme B, is a major pathway CTLs and NKCs initiate caspase-dependent apoptosis to eliminate pathogenic and tumor cells [[70,](#page-13-14) [71\]](#page-13-15). Jiang et al. cultured $ER\alpha$ -expressing human liver carcinoma cells with E2 resulting in upregulated expression of the granzyme B inhibitor, proteinase inhibitor-9 (PI-9). This upregulation protected the tumor cells against granule-mediated exocytosis by these cells per DNA fragmentation assays [\[72](#page-13-16)]. A similar study illustrated E2-induced PI-9 expression was also observed in ERαpositive MCF7 breast cancer cells with the same protection, while PI-9 knockdown blocked E2's protective effect [\[73](#page-13-17)]. Cumulatively, these studies suggest a component of E2 immunosuppression is via inhibition of NK- and CTL-mediated tumor cell elimination.

2.3.1.2 Regulatory T Cells

T cell activation and effector differentiation are integral to the adaptive immune response. FoxP3[−] expressing Tregs subdue neoplastic activity, as well as responder T cell expansion, through secretion of immunosuppressive cytokines [[74\]](#page-13-18).

Administration of physiologic doses of E2 to immunocompetent, ovariectomized mice has been observed to expand CD4+CD25+ Treg concentration, as well as Foxp3 expression in various tissue types [[75\]](#page-14-0). Furthermore, fluorescenceactivated cell sorting (FACs) assays revealed acquisition of CD25 in E2-incubated ERαexpressing CD4+CD25− cells [[75\]](#page-14-0). These transformed CD4+CD25+ T cells then exhibited an immunosuppressive Treg phenotype in vitro that significantly downregulated T cell concentration [\[75](#page-14-0)[–78](#page-14-1)]. Additional studies have reported E2-stimulated Foxp3 expression in murine Tregs, expression of which is vital to Treg functionality. High FoxP3+ Tregs in the TME is a negative prognostic indicator in a variety of cancers. For example, early-stage NSCLC with nuclear ERα expression has a relatively higher risk of both recurrence and FoxP3+ lymphocyte infiltrate [\[79](#page-14-2)]. Furthermore, a recent meta-analysis reported FoxP3+ Treg infiltration correlated negatively with OS in ER-positive breast cancer patients and positively in ER-negative patients [\[80](#page-14-3)]. Conversely, studies of $ER\alpha$ -positive breast tumors treated with letrozole in vivo demonstrated a resulting reduction of FoxP3+ Tregs [\[81](#page-14-4)].

E2 appears to suppress Treg expression in both physiologic and ERα/ERβ knockout mice, with the former group having increased expression of programmed-death 1 (PD-1) and the latter having decreased PD-1 expression [[82\]](#page-14-5). E2 treatment of ERα-positive endometrial and breast cancer cells also stimulates in vitro expression of the PD-1 ligand (PD-L1) via activation of PI3K signaling [[83\]](#page-14-6). PD-L1⁺ tumor cells exhaust PD-1⁺ cytotoxic T lymphocytes (CTLs) through this protein interaction, resulting in tumor immune evasion [\[84](#page-14-7)]. Given E2's upregulation of both PD-1 and PD-L1, the hormone appears to have an important influence on the pathway and its role in the TME.

2.3.2 Stromal Cells

Tumor evolution is heavily dependent on malignant tissue as well as recruited stromal cells that interact between and within the TME. Via an in vivo murine model, ERα expression in stromal cells was observed within the context of tumorcell-independent ER signaling in the TME. E2 interactions with stromal $ER\alpha$ has also been seen to accelerate neoplastic growth and blood vessel density in ovariectomized, syngeneic mice transplanted with ER-negative melanoma, breast, or lung cancer cells [[51\]](#page-12-8). The same study found this E2-stimulated tumor growth demonstrated a relative increase in immunocompromised mice, reflecting closer association with E2 modulation of innate immunity [[51\]](#page-12-8). Aromatase expression appears to also modulate the TME in certain neoplasms. Perineoplastic endometrial stromal cells' expression of aromatase also correlates with more advanced disease and, transitively, worse OS [[52,](#page-12-9) [85](#page-14-8)]. Similarly, perineoplastic breast adipocytes' expression of aromatase appears to be complicit in tumorigenesis in obese patients via inflammation and modification of the TME [\[50](#page-12-7), [86](#page-14-9), [87\]](#page-14-10). Additionally, type 2 pericytes have also been associated with tumorigenesis and vascular formation for tumors [\[88](#page-14-11)]. Pericytes recruited for vascular formation have been associated with $ER\alpha$ expression and E2-dependent signaling during function [\[89](#page-14-12), [90](#page-14-13)].

2.3.3 Cancer-Associated Fibroblasts

CAFs are one of the most integral stromal cell types in the TME for tumor survival and metastasis via paracrine-induced signaling pathways via chemokines and soluble growth factors [\[91](#page-14-14), [92\]](#page-14-15). ERα expression in breast CAFs have been observed in vivo through nuclear receptor arrays comparing gene expression between CAFs and normal human breast adipose fibroblasts [[53\]](#page-12-10). Interestingly, similar levels of ERα expression are seen in both malignant and physiologic fibroblasts, but with downstream upregulation of the direct transcriptional activator liver receptor homolog-1 (*LRH-1*) in the former [\[53](#page-12-10)]. The regulator serves to increase expression of the aromatase-encoding gene *CYP19A1* [[93–](#page-14-16)[95\]](#page-15-0). Co-expression of aromatase and LRH-1 in the breast TME suggests CAF-induced paracrine for-

mation of E2 and subsequent ER-mediated oncogenesis [[96\]](#page-15-1). Coculturing of endometrial CAFs with endometrial neoplastic cells have been seen to contribute to tumor progression, possibly attributed to CAFs' expression of ERα and ERβ [\[56](#page-13-1)]. This tumor progression mechanism is supported through in vitro upregulation of phosphatidylinositide 3-kinase (PI3K) and mitogen-activated protein kinase (MAPK) signaling networks, which are both well-known ER-mediated pathways in breast and lung cancer [\[56](#page-13-1), [97](#page-15-2)[–99](#page-15-3)].

Contrastingly, ER expression in prostate CAFs has contradicting evidence, with reports of ERα/ERβ expression portending advanced disease $[54]$ $[54]$ and others suggesting ER α expression is a protective factor again neoplastic invasion macrophage infiltration [[100,](#page-15-4) [101](#page-15-5)]. These latter in vitro studies conveyed that CAF ERα expression reduced murine and human prostate cancer cell invasion, as well as lymph node metastasis of orthotopically implanted human prostate cancer cells in mice [[101\]](#page-15-5). These $ER\alpha$ -positive CAFs appeared to halt invasion and metastasis of human prostate cancer cells through downstream downregulation of the C-C motif chemokine ligand 5 (CCL5) and IL-6 chemokines, whose roles are involved in growth factor signaling, inflammation, and tumor recruitment [[102,](#page-15-6) [103\]](#page-15-7).

2.3.4 Tumor-Associated Macrophages

In a physiologic setting, macrophages regulate tissue-specific innate immune responses to fight foreign invaders through polarization by varied cytokines. However, TAMs have been complicit in tumor proliferation and migration, as well as inflammation in the TME [\[104](#page-15-8), [105\]](#page-15-9). Physiologically, polarized M1 macrophages secrete the proinflammatory cytokines IFNγ, interleukin 12 (IL-12), and tumor necrosis factor (TNF)-α for tumor rejection and antigen presentation [\[106](#page-15-10)]. Alternatively, M2 macrophages produce interleukins 4, 5, 6, and 10 $[106]$ $[106]$, which are known promoters of tumor cell growth and immune evasion [\[107](#page-15-11)]. TAMs within the TME are often M2, with denser concentration demonstrating worse OS, thus offering a therapeutic opportunity for a variety of malignancies [[108\]](#page-15-12).

TAMs are an independent poor prognostic predictor for ovarian adenocarcinoma [[109\]](#page-15-13). Relatedly, co-localized expression of both ERα/ ER β is reported in human high-grade serous ovarian cystadenocarcinoma (HGSOC) TAMs. Interestingly, HGSOC in premenopausal women demonstrates elevated TAM infiltration relative to that of postmenopausal women. The highest concentration of TAMs in this TME can be found in ER α -positive tumors [\[58](#page-13-3)]. The mechanism of this was elucidated by an IHC analysis revealing aromatase expression in the TME of breast TAMs, which was observed to increase E2 production and breast cell proliferation [[59\]](#page-13-4). TAM proliferation, however, is relatively more prevalent in ER-negative breast malignancies [\[110](#page-15-14), [111](#page-15-15)]. It is important to note, however, that quantification of TAM polarization was not analyzed in these studies. Interestingly, aromatase and ERβ expression in NSCLC TAMs have also been observed, specifically in infiltrating macrophages of preneoplastic, tobacco carcinogen-induced murine lung lesions [[17,](#page-11-0) [60\]](#page-13-5).

Although direct observation of ER expression in TAMs has been limited, E2 induction of M2 polarization and subsequent tumor spread has been studied. A polyomavirus middle T (PyMT), ER-positive breast cancer murine model demonstrated direct E2 stimulation of tumoral M2 TAM infiltration and vascular endothelial growth factor (VEGF) [[112,](#page-15-16) [113\]](#page-15-17). Alternatively, untreated controls exhibited M1 TAM infiltration instead [\[112\]](#page-15-16). In a HGSOC, ovariectomized murine model, E2 induced growth of both ER-negative xenografts and M2 TAM infiltration [\[58](#page-13-3)]. In tobacco carcinogen-exposed mice, administration of E2 increased pulmonary TAM infiltration while mice receiving the aromatase inhibitor anastrozole had a significant reduction in pulmonary TAMs [\[114\]](#page-16-0). Further, E2-induced VEGF expression was also observed in this model [[114\]](#page-16-0). Of note, E2-mediated TAM infiltration has been observed in vitro to be fed forward via M2 TAM-induced epigenetic ER α upregulation via interleukin 17A (IL-17A) in endometrial malignancy [\[115](#page-16-1)]. This positive feedback mechanism between E2 and M2 TAMs provides a potential therapeutic target, a concept recently addressed via effects of the phytoestrogen SERM resveratrol in a lung cancer xenograft model [\[116](#page-16-2)]. Resveratrol treatment appeared to suppress tumor proliferation through decreased signal transducer and activator of transcription 3 (STAT3) signaling and M2 polarization [\[116](#page-16-2)].

2.3.5 Myeloid-Derived Suppressor Cells

MDSCs are another myeloid cell present in the TME known to interfere immune surveillance and facilitate tumor growth $[117]$ $[117]$. ER α expression in human ovarian adenocarcinoma MDSCs has been identified by IHC and confirmed through PCR and immunoblotting [[57\]](#page-13-2). In an E2-insensitive syngeneic ovarian cancer model, ovariectomized mice exhibited improved survival compared to non-ovariectomized mice following tumor challenge. Contrastingly, E2 supplementation in these mice accelerated tumor progression and reversed the protective effect found in estrogen-depleted mice [\[57](#page-13-2)]. Of note, this study found that T-cell-deficient mice lost survival benefit of estrogen depletion, suggesting adaptive immunocompetence to be mechanistically integral [\[57](#page-13-2)]. Estrogen's effect on the two legs of immunity was also observed in E2-treated mice, which were found to have notably decreased concentrations of helper and cytotoxic T cells, and significantly increased concentrations of granu-locytic MDSCs in spleen and tumor beds [[57\]](#page-13-2). ER-dependence of MDSC expansion was further studied with in vitro administration of the $ER\alpha$ antagonist methylpiperidino pyrazole (MPP) to inhibit MDSC proliferation [[57\]](#page-13-2). Ovarian tumorbearing mice treated with E2 had measurable JAK2 and SRC upregulation with downstream STAT3 signaling, a regulator of myeloid differentiation and development [[118\]](#page-16-4). In syngeneic lung and breast cancer murine models, E2-stimulated tumor growth was mitigated by MDSC depletion after treatment with anti-Gr1 antibodies [\[57](#page-13-2), [119](#page-16-5), [120\]](#page-16-6). Patients with cervical cancer that were pregnant with high E2 had increased expansion

of MDSCs and shorter PFS. These findings were further evaluated in mouse models [[120\]](#page-16-6).

2.3.6 Inflammatory Cytokines and Eicosanoids

Chronic inflammation has been accepted as a common factor in tumorigenesis and spread. TME facilitates neoplastic progression primarily through cytokine-induced oncogenic pathway activation, leading to cell proliferation, immune evasion, and infiltration [\[121](#page-16-7)]. IL-6 from TAFs has been observed to assist ERα-positive breast cancer proliferation and immune evasion [\[122](#page-16-8)] via STAT3 activation in vitro and in vivo [[123\]](#page-16-9). TNF α in ER α -positive breast cancer cells has been observed to regulate gene expression for metastasis [\[124](#page-16-10)]. This cytokine has also been shown to upregulate aromatase expression in cultured human adipose stromal cells [[125\]](#page-16-11). Neoplastic implication of these inflammatory markers is evidenced by data showing TNFα and IL-6 correlate closely with aromatase expression in human breast cancer tissue and not in adjacent noncancerous tissue [[126\]](#page-16-12). Aromatase has similar transcriptional correlation with cyclooxygenase-2 (COX-2) [\[126](#page-16-12)]. COX-2 mediates the inflammatory response by producing eicosanoids such as prostaglandin E2 (PGE2) [\[127](#page-16-13)], which upregulates aromatase expression through cyclic adenosine monophosphate (cAMP) in breast malignancy [\[128](#page-16-14)]. Despite conflicting reports, a case–control study demonstrated regular administration of the nonsteroidal anti-inflammatory drug (NSAID) aspirin reduced the risk of developing ERα-positive breast cancers (hazard ratio (HR) = 0.74; 95% CI, 0.60–0.93), but not $ER\alpha$ negative cancers (HR = 0.97; 95% CI, 0.67–1.40) [\[129](#page-16-15)].

ERα, TNFα, and NF-κB protein expression correlate closely in breast cancer tissues [[130\]](#page-16-16). NF-κB signaling, a proinflammatory cytokine associated with IL-6 and TNF α , is often constitutively activated in many tumor types [\[131](#page-16-17)]. High levels of the cytokine are also implicated in SERM resistance in ERα-expressing human breast cancer cells [[132,](#page-16-18) [133\]](#page-16-19). E2 also enhanced

pulmonary inflammation through increased NF-κB, VEGF, and IL-17A in a murine model evaluating tobacco carcinogen-induced lung cancer [\[114](#page-16-0)]. E2 inhibition with combined AI/ NSAID treatment served to noticeably decrease pulmonary malignancy in these mice. Notable pathways affected included IL-17A expression, IL-6 concentration, as well as STAT3 and MAPK [\[114](#page-16-0)]. Cumulatively, there appears to be a potential target for the E2 pathway as it interacts with tumorigenesis via inflammation.

2.3.7 The Impact of Supraphysiologic Estrogen

Esterified estrogen, specifically estrone, is significantly increased in the setting of obesity. Aromatase in adipocytes serves to increase estrone secretion in the setting of hypertrophy. The effect this supraphysiologic estrogen has on tumorigenesis has been controversial [\[134](#page-17-0), [135\]](#page-17-1). Recent findings suggest that while immune dysfunction and tumor progression are associated with obesity, improved response to immunotherapy may also be associated with obesity, supporting the immune-mediated link between obesity and cancer [\[136](#page-17-2)]. Chronic inflammation from obesity is integral to carcinogenesis and tumor evolution, as observed in postmenopausal, ERand progesterone receptor (PR)-expressing breast malignancy [[137\]](#page-17-3). It is important to note that studies suggesting protumor effects of estrogen in estrogen-depleted mammals have been performed primarily in the setting of hormone replacement therapy (HRT).

Tumorigenesis, progression, and infiltration in the setting of HRT in estrogen-depleted mammals remain controversial. There is a paucity of studies demonstrating proinflammatory changes with hormone replacement therapy in murine models. In contrast, there are many studies conveying a protective effect of exogenous estrogen. Specifically, ERβ-expression has been observed to prevent progression of human colorectal carcinoma (CRC) [\[134](#page-17-0), [138](#page-17-4)]. Mechanistic protection against carcinogenesis with exogenous estrogen in postmenopausal patients appears to primarily

be through a decrease in the natural postmenopausal increase in Th1/Th2 ratio [[139,](#page-17-5) [140\]](#page-17-6). Specifically, Th2 cytokines are quantifiably stable until late postmenopausal stage, while production of Th1 cytokines is progressively increased in women after menopause. HRT prevents this increased Th1/Th2 ratio, thereby improving the aberration of Th1/Th2 balance that is implicated in an inadequate immune response and neoplastic conditions [\[140](#page-17-6)]. Substantiation of this antitumoral concept was provided through an in vivo, placebo-controlled study regarding postmenopausal human breast cancer cell demonstrating estrogen's notable decrease in IL-6 production [\[141](#page-17-7)].

2.4 Clinical Implications of Targeting the Estrogen Pathway in the Tumor Microenvironment

Immunotherapy is a developing and effective treatment avenue in the world of cancer; yet the TME and its immunosuppressive mechanisms is a deterrent for large-scale success. As it stands, the immune checkpoint modulators of cytotoxic T-lymphocyte-associated antigen 4 (CTLA4) and PD-1/PD-L1 are the most studied immunotherapies [\[142](#page-17-8)]. These revolutionary options have had dramatic impacts on OS relative to standard-ofcare chemotherapies [[143–](#page-17-9)[146\]](#page-17-10). Even so, response rates are limited to 20–35% of cases, closely dependent on tumor type, stage, and PD-L1 expression [[147\]](#page-17-11). Moreover, 25–33% of melanoma patients often demonstrate delayed relapse during treatments attributed to tumor cell adaptation [[5,](#page-10-2) [6\]](#page-10-8).

There appears to be a balance of tumoral mutations and immunoediting that facilitate immune evasion, and subsequently, failure of checkpoint therapy. On the one hand, damaged DNA repair mechanisms, increased non-synonymous somatic mutational load, and neoantigen presentation cripple immune evasion and improve OS [\[2](#page-10-9), [3](#page-10-10), [148\]](#page-17-12). On the other hand, damage to antigen-presenting mechanisms, as well as recurrence of nonantigenic mutations,

appears to facilitate immune evasion [\[149](#page-17-13), [150\]](#page-17-14). Studies identifying these mechanisms provide insight into measurable biomarkers to assess tumor responsiveness to current and, inevitably necessary, novel immunotherapies. A potential investigative therapy is endocrinological agents that modulate E2 and its protumoral pathway to abrogate tumor immune evasion. Specifically, anti-estrogen therapy may reduce TME immunosuppression while increasing E2-sensitive tumor responsiveness.

Recently, a high-throughput screening assay in human lung cancer cells demonstrated fulvestrant, an anti-estrogen agent, as the most efficacious compound in increasing tumor sensitivity to immune-mediated lysis [[151\]](#page-18-0). Fulvestrant additionally has few interactions and overlapping toxicities with anti-PD-1/PD-L1 agents. Thus, anti-E2 therapies to target the immunosuppressive TME could increase efficacy and duration of response of current immune checkpoint inhibitors (ICI) [\[119](#page-16-5), [152](#page-18-1)] (Fig. [2.1](#page-8-0)).

Based on the well-established evidence of sex-driven dimorphism in immune function and response, patient sex has been postulated to have an influence on the efficacy of ICIs [[9\]](#page-10-11). This sexual dimorphism plays an important role in the disparity of cancer immunoediting in females and males and could not only explain differences in progression and mortality observed between male and female cancer patients but also sex differences in response rates, toxicity patterns, and outcomes to treatment with ICIs. In support of this concept, the PD-1/PD-L1 pathway is modulated by multiple X-linked microRNAs (miR-NAs), which crosstalk with the estrogen-ER α axis, suggesting an important role of the estrogen pathway and response to ICIs [\[153](#page-18-2), [154\]](#page-18-3). Further since estrogen modulation of the PD-1/PD-L1 pathway has been demonstrated in animal models [\[82](#page-14-5), [155\]](#page-18-4), it is reasonable to expect that immunotherapy efficacy may vary according to patient sex.

In an effort to identify patient characteristics linked to ICI effectiveness, several meta-analyses have been conducted to evaluate sex-differential effects in efficacy of ICIs. Conforti et al. evaluated the effect of patient's sex on the efficacy of

ICIs measured in terms of OS on different tumor types [\[156](#page-18-5)]. This study included 11,351 patients (67% men and 33% women) enrolled in 20 Phase II and III randomized controlled trials that evaluated CTLA4 inhibitors, as well as PD-1/PD-L1 inhibitors in patients with different tumor types, mostly melanoma and NSCLC. Results showed that male patients who received ICIs alone had a reduced risk of death compared to men in the control arms (HR = 0.72 , 95% CI 0.65-0.79). Similar findings were observed in female patients, but the difference in risk reduction was smaller between the treatment and the control arm $(HR = 0.86, 95\% \text{ CI } 0.79{\text -}0.93)$. Although there was a significant difference in the efficacy of ICIs between male and female patients, the heterogeneity test for this sex-related interaction was not quite significant.

A subsequent meta-analysis evaluated the differences in outcomes based on sex in lung cancer patients who received targeted therapy or immunotherapy [\[10\]](#page-10-5). This study included a total of 12 Phase III clinical trials evaluating EGFR, ALK, and PD-1 inhibitors versus chemotherapy. Of the 12 trials included in this meta-analysis, five compared PD-1 inhibitors

versus chemotherapy, two of which compared pembrolizumab versus chemotherapy (KEYNOTE 010 and KEYNOTE 024), and three compared nivolumab versus chemotherapy (CheckMate 017, CheckMate 026, CheckMate 057) [\[144,](#page-17-15) [157–](#page-18-6)[159\]](#page-18-7). The studies that compared ICIs versus chemotherapy included 1028 female and 1435 male lung cancer patients. While there was significant heterogeneity between studies, OS was favorable in male patients treated with ICIs compared to chemotherapy (HR = 0.76 ; 95% CI 0.068–0.86; $p < 0.00001$). There was no significant difference in survival in female lung cancer patients receiving chemotherapy compared to ICIs $(HR = 1.03; 95\% \text{CI } 0.89 \text{ to } 1.03; p = 0.69)$. In a separate study focused on metastatic NSCLC, El-Ostra et al. evaluated results from eight randomized clinical trials for predictors of benefit to single agent ICIs over chemotherapy [\[8\]](#page-10-4). NSCLC patients treated with ICIs had significant progression-free survival (PFS) superiority in ever-smokers, male patients, and patients with PD-L1-positive tumors. In contrast, female NSCLC patients had comparable PFS between ICIs and chemotherapy.

Wallis et al. also conducted a meta-analysis that included 23 randomized clinical trials (67.9% men and 32.1% women) that compared ICIs (both ICI alone and ICI plus chemotherapy trials) to standard-of-care treatment in advanced solid tumors (including NSCLC, SCLC, urothelial carcinoma, head and neck squamous carcinoma, melanoma, mesothelioma, clear cell renal carcinoma, and gastric or gastroesophageal carcinoma). In this study, no difference in OS between men and women who received immunotherapy was observed $(I^2 = 38\%; p = 0.6)$ [[160\]](#page-18-8). The conflicting results and limitations in these metaanalyses suggest that further investigation of the efficacy of ICIs and patients' sex is warranted in future studies. While the majority of the trials included in these studies were underpowered to detect clinically relevant sex differences in outcome, these results indicate that the hormonal milieu may have some effect on treatment response (Table [2.2\)](#page-9-0).

The current best predictive markers of therapeutic response to ICIs are high PD-L1 expression and high tumor mutational burden (TMB). The difference between PD-L1 expression between men and women has been evaluated in some cancer patient cohort with a reported increased PD-L1 expression in male patients [\[161](#page-18-9)[–163](#page-18-10)]. TMB has also been shown to be lower in women compared to men $(p = 0.0349)$, across multiple studies [\[164](#page-18-11), [165](#page-18-12)]. TMB is predictive of response to ICI in lung cancer and is lower in female lung cancer patients compared to male lung cancer patients [[165\]](#page-18-12). Similarly, sex differences in immune-related adverse events (irAEs) have also been noted in ICI trials [\[166](#page-19-0), [167\]](#page-19-1). The

gut microbiome and obesity are emerging areas of interest that may predict response to ICIs [\[168](#page-19-2)]. Whether or not these factors interact with sex hormones in the context of anti-cancer immunity is yet to be determined.

2.5 Conclusions and Perspective

The E2 pathway is an identified promoter of tumorigenesis in several cancers, largely for its genomic, epigenomic, and transcriptional effects on tumor cells and the TME. The reciprocal interactions of the peritumoral and tumoral environment are becoming more evident, with E2 playing a major role in modulation of primarily protumoral pathways. With immunoediting being a culprit in E2-mediated protumoral activity, it appears to be an important deterrent for checkpoint blockade immunotherapy success. Thus, inhibition of the E2 pathway may augment current immunotherapy response rates.

Carcinogenesis from obesity and its related illnesses are thought to be primarily driven through proinflammatory cytokine secretion. Supraphysiologic estrogen from adipocyte aromatase expression may also play a role, but as of now, it is difficult to distinguish. However, estrogen replacement therapy in postmenopausal women appears to have a relatively protective effect via immune modulation. Stabilization of immunologic aberrancies, notably in the adaptive immune system, is protective against age-related malignancies such as colorectal carcinoma and breast cancers. Based on the above discussion, future studies are war-

Malignancy	Selected study drugs	$n =$	Clinical trial number
ER+/Her2- Breast cancer	Exemestane and durvalumab/tremelimumab	240	NCT02997995
ER+/Her2- Breast cancer	Pembrolizumab, letrozole, and palbociclib	22	NCT02778685
ER+/Her2- Breast cancer	Atezolizumab and fulvestrant	126	NCT03280563
ER+/Her2- Breast cancer	Pembrolizumab and exemestane	25	NCT02990845
ER+/Her2- Breast cancer	Pembrolizumab and AI	37	NCT02971748
ER+/Her2- Breast cancer	Pembrolizumab and letrozole, exemestane anastrozole	56	NCT02648477
AR+/ER-Breast cancer	Pembrolizumab and enobosarm	29	NCT02971761

Table 2.2 Selected trials evaluating the combination of Estrogen pathway targeting agents with ICIs

Selected ongoing trials evaluating ICI in combination with therapeutic agents targeting the E2 pathway. Disease type, selected study agents, predicted accrual size, and clinical trial number are provided

ranted to assess responsiveness to current ICIs across sex, menopausal status, and BMI in order to isolate E2 pathway contribution to immune evasion.

References

- 1. Alexandrov LB, Nik-Zainal S, Wedge DC, Aparicio SA, Behjati S, Biankin AV, Bignell GR, Bolli N, Borg A, Borresen-Dale AL, Boyault S, Burkhardt B, Butler AP, Caldas C, Davies HR, Desmedt C, Eils R, Eyfjord JE, Foekens JA, Greaves M, Hosoda F, Hutter B, Ilicic T, Imbeaud S, Imielinski M, Jager N, Jones DT, Jones D, Knappskog S, Kool M, Lakhani SR, Lopez-Otin C, Martin S, Munshi NC, Nakamura H, Northcott PA, Pajic M, Papaemmanuil E, Paradiso A, Pearson JV, Puente XS, Raine K, Ramakrishna M, Richardson AL, Richter J, Rosenstiel P, Schlesner M, Schumacher TN, Span PN, Teague JW, Totoki Y, Tutt AN, Valdes-Mas R, van Buuren MM, van 't Veer L, Vincent-Salomon A, Waddell N, Yates LR, Australian Pancreatic Cancer Genome I, Consortium IBC, Consortium IM-S, PedBrain I, Zucman-Rossi J, Futreal PA, Mcdermott U, Lichter P, Meyerson M, Grimmond SM, Siebert R, Campo E, Shibata T, Pfister SM, Campbell PJ, Stratton MR (2013) Signatures of mutational processes in human cancer. Nature 500(7463):415–421. [https://doi.org/10.1038/](https://doi.org/10.1038/nature12477) [nature12477.](https://doi.org/10.1038/nature12477) PubMed PMID: 23945592; PMCID: PMC3776390
- 2. Rooney MS, Shukla SA, Wu CJ, Getz G, Hacohen N (2015) Molecular and genetic properties of tumors associated with local immune cytolytic activity. Cell 160(1–2):48–61. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cell.2014.12.033) [cell.2014.12.033.](https://doi.org/10.1016/j.cell.2014.12.033) PubMed PMID: 25594174; PMCID: PMC4856474
- 3. Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, Lee W, Yuan J, Wong P, Ho TS, Miller ML, Rekhtman N, Moreira AL, Ibrahim F, Bruggeman C, Gasmi B, Zappasodi R, Maeda Y, Sander C, Garon EB, Merghoub T, Wolchok JD, Schumacher TN, Chan TA (2015) Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science 348(6230):124–128. [https://doi.org/10.1126/sci](https://doi.org/10.1126/science.aaa1348)[ence.aaa1348](https://doi.org/10.1126/science.aaa1348). PubMed PMID: 25765070; PMCID: PMC4993154
- 4. Quail DF, Joyce JA (2013) Microenvironmental regulation of tumor progression and metastasis. Nat Med 19(11):1423–1437. [https://doi.org/10.1038/](https://doi.org/10.1038/nm.3394) [nm.3394](https://doi.org/10.1038/nm.3394). PubMed PMID: 24202395; PMCID: 3954707
- 5. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A (2017) Primary, adaptive, and acquired resistance to cancer immunotherapy. Cell 168(4):707–723. [https://doi.org/10.1016/j.cell.2017.01.017.](https://doi.org/10.1016/j.cell.2017.01.017) PubMed PMID: 28187290; PMCID: PMC5391692
- 6. Schachter J, Ribas A, Long GV, Arance A, Grob JJ, Mortier L, Daud A, Carlino MS, McNeil C, Lotem M, Larkin J, Lorigan P, Neyns B, Blank C, Petrella TM, Hamid O, Zhou H, Ebbinghaus S, Ibrahim N, Robert C (2017) Pembrolizumab versus ipilimumab for advanced melanoma: final overall survival results of a multicentre, randomised, open-label phase 3 study (KEYNOTE-006). Lancet 390(10105):1853–1862. [https://doi.org/10.1016/S0140-6736\(17\)31601-X](https://doi.org/10.1016/S0140-6736(17)31601-X). PubMed PMID: 28822576
- 7. Somasundaram A, Burns TF (2017) The next generation of immunotherapy: keeping lung cancer in check. J Hematol Oncol 10(1):87. [https://doi.](https://doi.org/10.1186/s13045-017-0456-5) [org/10.1186/s13045-017-0456-5.](https://doi.org/10.1186/s13045-017-0456-5) PubMed PMID: 28434399; PMCID: PMC5402056
- 8. El-Osta H, Jafri S (2019) Predictors for clinical benefit of immune checkpoint inhibitors in advanced non-small-cell lung cancer: a meta-analysis. Immunotherapy 11(3):189–99. Epub 2019/02/08. <https://doi.org/10.2217/imt-2018-0086>. PubMed PMID: 30730276
- 9. Capone I, Marchetti P, Ascierto PA, Malorni W, Gabriele L (2018) Sexual dimorphism of immune responses: a new perspective in cancer immunotherapy. Front Immunol 9:552. Epub 2018/04/06. [https://doi.org/10.3389/fimmu.2018.00552.](https://doi.org/10.3389/fimmu.2018.00552) PubMed PMID: 29619026; PMCID: PMC5871673
- 10. Pinto JA, Vallejos CS, Raez LE, Mas LA, Ruiz R, Torres-Roman JS, Morante Z, Araujo JM, Gomez HL, Aguilar A, Bretel D, Flores CJ, Rolfo C (2018) Gender and outcomes in non-small cell lung cancer: an old prognostic variable comes back for targeted therapy and immunotherapy? ESMO Open 3(3):e000344. Epub 2018/04/24. [https://doi.](https://doi.org/10.1136/esmoopen-2018-000344) [org/10.1136/esmoopen-2018-000344](https://doi.org/10.1136/esmoopen-2018-000344). PubMed PMID: 29682332; PMCID: PMC5905840
- 11. Klein SL, Flanagan KL (2016) Sex differences in immune responses. Nat Rev Immunol 16:626. <https://doi.org/10.1038/nri.2016.90>
- 12. Nilsson S, Gustafsson J (2010) Estrogen receptors: their actions and functional roles in health and human disease. In: Bunce C, Campbell MJ (eds) Nuclear receptors: current concepts and future challenges. Dordrecht, Netherlands, pp 91–141
- 13. Delaunay F, Pettersson K, Tujague M, Gustafsson JA (2000) Functional differences between the amino-terminal domains of estrogen receptors alpha and beta. Mol Pharmacol 58(3):584–590. PubMed PMID: 10953052
- 14. Zhu BT, Han GZ, Shim JY, Wen Y, Jiang XR (2006) Quantitative structure-activity relationship of various endogenous estrogen metabolites for human estrogen receptor alpha and beta subtypes: Insights into the structural determinants favoring a differential subtype binding. Endocrinology 147(9):4132–4150. <https://doi.org/10.1210/en.2006-0113>. PubMed PMID: 16728493
- 15. Brzozowski AM, Pike AC, Dauter Z, Hubbard RE, Bonn T, Engström O, Öhman L, Greene GL, Gustafsson J-Å, Carlquist M (1997) Molecular basis

of agonism and antagonism in the oestrogen receptor. Nature 389(6652):753

- 16. Klinge CM (2001) Estrogen receptor interaction with estrogen response elements. Nucleic Acids Res 29(14):2905–2919
- 17. Siegfried JM, Stabile LP (2014) Estrongenic steroid hormones in lung cancer. Semin Oncol 41(1):5–16. <https://doi.org/10.1053/j.seminoncol.2013.12.009>. PubMed PMID: 24565577; PMCID: PMC4001725
- 18. Folkerd EJ, Dowsett M (2010) Influence of sex hormones on cancer progression. J Clin Oncol 28(26):4038–4044. [https://doi.org/10.1200/](https://doi.org/10.1200/JCO.2009.27.4290) [JCO.2009.27.4290](https://doi.org/10.1200/JCO.2009.27.4290). PubMed PMID: 20644089
- 19. Frasor J, Danes JM, Komm B, Chang KC, Lyttle CR, Katzenellenbogen BS (2003) Profiling of estrogen up- and down-regulated gene expression in human breast cancer cells: insights into gene networks and pathways underlying estrogenic control of proliferation and cell phenotype. Endocrinology 144(10):4562–4574. [https://doi.org/10.1210/](https://doi.org/10.1210/en.2003-0567) [en.2003-0567](https://doi.org/10.1210/en.2003-0567). PubMed PMID: 12959972
- 20. Hershberger PA, Vasquez AC, Kanterewicz B, Land S, Siegfried JM, Nichols M (2005) Regulation of endogenous gene expression in human nonsmall cell lung cancer cells by estrogen receptor ligands. Cancer Res 65(4):1598–1605. [https://doi.](https://doi.org/10.1158/0008-5472.CAN-04-2694) [org/10.1158/0008-5472.CAN-04-2694.](https://doi.org/10.1158/0008-5472.CAN-04-2694) PubMed PMID: 15735050
- 21. Egloff AM, Rothstein ME, Seethala R, Siegfried JM, Grandis JR, Stabile LP (2009) Cross-talk between estrogen receptor and epidermal growth factor receptor in head and neck squamous cell carcinoma. Clin Cancer Res 15(21):6529–6540. [https://](https://doi.org/10.1158/1078-0432.CCR-09-0862) [doi.org/10.1158/1078-0432.CCR-09-0862.](https://doi.org/10.1158/1078-0432.CCR-09-0862) PubMed PMID: 19825947; PMCID: 2783886
- 22. Lanzino M, Morelli C, Garofalo C, Panno ML, Mauro L, Ando S, Sisci D (2008) Interaction between estrogen receptor alpha and insulin/IGF signaling in breast cancer. Curr Cancer Drug Targets 8(7):597–610. PubMed PMID: 18991569
- 23. Siegfried JM, Farooqui M, Rothenberger NJ, Dacic S, Stabile LP (2017) Interaction between the estrogen receptor and fibroblast growth factor receptor pathways in non-small cell lung cancer. Oncotarget 8(15):24063–24076. [https://doi.org/10.18632/onco](https://doi.org/10.18632/oncotarget.16030)[target.16030.](https://doi.org/10.18632/oncotarget.16030) PubMed PMID: 28445992; PMCID: 5421827
- 24. American Cancer Society. Cancer Facts & Figures 2019. Atlanta: American Cancer Society; 2019.
- 25. Dunnwald LK, Rossing MA, Li CI (2007) Hormone receptor status, tumor characteristics, and prognosis: a prospective cohort of breast cancer patients. Breast Cancer Res 9(1):R6. [https://doi.org/10.1186/](https://doi.org/10.1186/bcr1639) [bcr1639.](https://doi.org/10.1186/bcr1639) PubMed PMID: 17239243; PMCID: PMC1851385
- 26. Leung YK, Lee MT, Lam HM, Tarapore P, Ho SM (2012) Estrogen receptor-beta and breast cancer: translating biology into clinical practice. Steroids 77(7):727–737. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.steroids.2012.03.008) [steroids.2012.03.008](https://doi.org/10.1016/j.steroids.2012.03.008). PubMed PMID: 22465878; PMCID: PMC3356459
- 27. Phiel KL, Henderson RA, Adelman SJ, Elloso MM (2005) Differential estrogen receptor gene expression in human peripheral blood mononuclear cell populations. Immunol Lett 97(1):107–113. [https://](https://doi.org/10.1016/j.imlet.2004.10.007)
doi.org/10.1016/j.imlet.2004.10.007. PubMed doi.org/10.1016/j.imlet.2004.10.007. PMID: 15626482
- 28. Laffont S, Rouquie N, Azar P, Seillet C, Plumas J, Aspord C, Guery JC (2014) X-Chromosome complement and estrogen receptor signaling independently contribute to the enhanced TLR7-mediated IFN-alpha production of plasmacytoid dendritic cells from women. J Immunol 193(11):5444–5452. <https://doi.org/10.4049/jimmunol.1303400>. PubMed PMID: 25339659
- 29. Fish EN (2008) The X-files in immunity: sex-based differences predispose immune responses. Nat Rev Immunol 8(9):737–744. [https://doi.org/10.1038/](https://doi.org/10.1038/nri2394) [nri2394](https://doi.org/10.1038/nri2394). PubMed PMID: 18728636
- 30. Kovats S (2015) Estrogen receptors regulate innate immune cells and signaling pathways. Cell Immunol 294(2):63–69. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cellimm.2015.01.018) [cellimm.2015.01.018](https://doi.org/10.1016/j.cellimm.2015.01.018). PubMed PMID: 25682174; PMCID: 4380804
- 31. Kovats S (2012) Estrogen receptors regulate an inflammatory pathway of dendritic cell differentiation: mechanisms and implications for immunity. Horm Behav 62(3):254–262. [https://doi.](https://doi.org/10.1016/j.yhbeh.2012.04.011) [org/10.1016/j.yhbeh.2012.04.011](https://doi.org/10.1016/j.yhbeh.2012.04.011). PubMed PMID: 22561458; PMCID: 3415586
- 32. Khan D, Ansar AS (2015) The immune system is a natural target for estrogen action: opposing effects of estrogen in two prototypical autoimmune diseases. Front Immunol 6:635. [https://doi.org/10.3389/](https://doi.org/10.3389/fimmu.2015.00635) [fimmu.2015.00635.](https://doi.org/10.3389/fimmu.2015.00635) PubMed PMID: 26779182; PMCID: 4701921
- 33. Cerami E, Gao J, Dogrusoz U, Gross BE, Sumer SO, Aksoy BA, Jacobsen A, Byrne CJ, Heuer ML, Larsson E, Antipin Y, Reva B, Goldberg AP, Sander C, Schultz N (2012) The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov 2(5):401–404. [https://doi.org/10.1158/2159-8290.](https://doi.org/10.1158/2159-8290.CD-12-0095) [CD-12-0095](https://doi.org/10.1158/2159-8290.CD-12-0095). PubMed PMID: 22588877; PMCID: 3956037
- 34. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, Sun Y, Jacobsen A, Sinha R, Larsson E, Cerami E, Sander C, Schultz N (2013) Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 6(269):l1. [https://doi.org/10.1126/scisignal.2004088.](https://doi.org/10.1126/scisignal.2004088) PubMed PMID: 23550210; PMCID: 4160307
- 35. Li L, Wang Q, Lv X, Sha L, Qin H, Wang L, Li L (2015) Expression and localization of estrogen receptor in human breast cancer and its clinical significance. Cell Biochem Biophys 71(1):63–68. <https://doi.org/10.1007/s12013-014-0163-6>. PubMed PMID: 25113640
- 36. Grann VR, Troxel AB, Zojwalla NJ, Jacobson JS, Hershman D, Neugut AI (2005) Hormone receptor status and survival in a population-based cohort of patients with breast carcinoma. Cancer

103(11):2241–2251. [https://doi.org/10.1002/](https://doi.org/10.1002/cncr.21030) [cncr.21030.](https://doi.org/10.1002/cncr.21030) PubMed PMID: 15844176

- 37. Shen Z, Luo H, Li S, Sheng B, Zhao M, Zhu H, Zhu X (2017) Correlation between estrogen receptor expression and prognosis in epithelial ovarian cancer: a meta-analysis. Oncotarget 8(37):62400–13. [https://doi.org/10.18632/oncotarget.18253.](https://doi.org/10.18632/oncotarget.18253) PubMed PMID: 28977954; PMCID: PMC5617514
- 38. Zhang Y, Zhao D, Gong C, Zhang F, He J, Zhang W, Zhao Y, Sun J (2015) Prognostic role of hormone receptors in endometrial cancer: a systematic review and meta-analysis. World J Surg Oncol 13:208. <https://doi.org/10.1186/s12957-015-0619-1>. PubMed PMID: 26108802; PMCID: PMC4511445
- 39. Kawai H, Ishii A, Washiya K, Konno T, Kon H, Yamaya C, Ono I, Minamiya Y, Ogawa J (2005) Estrogen receptor alpha and beta are prognostic factors in non-small cell lung cancer. Clin Cancer Res 11(14):5084–5089. [https://doi.org/10.1158/1078-](https://doi.org/10.1158/1078-0432.CCR-05-0200) [0432.CCR-05-0200](https://doi.org/10.1158/1078-0432.CCR-05-0200). PubMed PMID: 16033821
- 40. Nose N, Sugio K, Oyama T, Nozoe T, Uramoto H, Iwata T, Onitsuka T, Yasumoto K (2009) Association between estrogen receptor-beta expression and epidermal growth factor receptor mutation in the postoperative prognosis of adenocarcinoma of the lung. J Clin Oncol 27(3):411–417. [https://doi.org/10.1200/](https://doi.org/10.1200/JCO.2008.18.3251) [JCO.2008.18.3251](https://doi.org/10.1200/JCO.2008.18.3251). PubMed PMID: 19064969
- 41. Stabile LP, Dacic S, Land SR, Lenzner DE, Dhir R, Acquafondata M, Landreneau RJ, Grandis JR, Siegfried JM (2011) Combined analysis of estrogen receptor beta-1 and progesterone receptor expression identifies lung cancer patients with poor outcome. Clin Cancer Res 17(1):154-164. [https://doi.](https://doi.org/10.1158/1078-0432.CCR-10-0992) [org/10.1158/1078-0432.CCR-10-0992.](https://doi.org/10.1158/1078-0432.CCR-10-0992) PubMed PMID: 21062926; PMCID: 3064257
- 42. Hsu LH, Chu NM, Kao SH (2017) Estrogen, estrogen receptor and lung cancer. Int J Mol Sci 18(8). [https://doi.org/10.3390/ijms18081713.](https://doi.org/10.3390/ijms18081713) PubMed PMID: 28783064; PMCID: PMC5578103
- 43. Mah V, Seligson DB, Li A, Marquez DC, Wistuba II, Elshimali Y, Fishbein MC, Chia D, Pietras RJ, Goodglick L (2007) Aromatase expression predicts survival in women with early-stage non small cell lung cancer. Cancer Res 67(21):10484–10490. <https://doi.org/10.1158/0008-5472.CAN-07-2607>. PubMed PMID: 17974992; PMCID: PMC3581354
- 44. Miller WR, Anderson TJ, Jack WJ (1990) Relationship between tumour aromatase activity, tumour characteristics and response to therapy. J Steroid Biochem Mol Biol 37(6):1055–1059. PubMed PMID: 2285581
- 45. Lipton A, Santen RJ, Santner SJ, Harvey HA, Sanders SI, Matthews YL (1992) Prognostic value of breast cancer aromatase. Cancer 70(7):1951–1955. PubMed PMID: 1525771
- 46. Esteban JM, Warsi Z, Haniu M, Hall P, Shively JE, Chen S (1992) Detection of intratumoral aromatase in breast carcinomas. An immunohistochemical study with clinicopathologic correlation. Am J

Pathol 140(2):337–343. PubMed PMID: 1739127; PMCID: PMC1886419

- 47. Miki Y, Suzuki T, Sasano H (2007) Controversies of aromatase localization in human breast cancer--stromal versus parenchymal cells. J Steroid Biochem Mol Biol 106(1–5):97–101. [https://doi.](https://doi.org/10.1016/j.jsbmb.2007.05.007) [org/10.1016/j.jsbmb.2007.05.007](https://doi.org/10.1016/j.jsbmb.2007.05.007). PubMed PMID: 17624762
- 48. Haldosen LA, Zhao C, Dahlman-Wright K (2014) Estrogen receptor beta in breast cancer. Mol Cell Endocrinol 382(1):665–672. [https://doi.](https://doi.org/10.1016/j.mce.2013.08.005) [org/10.1016/j.mce.2013.08.005.](https://doi.org/10.1016/j.mce.2013.08.005) PubMed PMID: 23954741
- 49. Leygue E, Murphy LC (2013) A bi-faceted role of estrogen receptor beta in breast cancer. Endocr Relat Cancer 20(3):R127–R139. [https://doi.org/10.1530/](https://doi.org/10.1530/ERC-12-0389) [ERC-12-0389.](https://doi.org/10.1530/ERC-12-0389) PubMed PMID: 23533249
- 50. Morris PG, Hudis CA, Giri D, Morrow M, Falcone DJ, Zhou XK, Du B, Brogi E, Crawford CB, Kopelovich L, Subbaramaiah K, Dannenberg AJ (2011) Inflammation and increased aromatase expression occur in the breast tissue of obese women with breast cancer. Cancer Prev Res (Phila) 4(7):1021–1029. [https://doi.org/10.1158/1940-](https://doi.org/10.1158/1940-6207.CAPR-11-0110) [6207.CAPR-11-0110](https://doi.org/10.1158/1940-6207.CAPR-11-0110). PubMed PMID: 21622727; PMCID: PMC3131426
- 51. Pequeux C, Raymond-Letron I, Blacher S, Boudou F, Adlanmerini M, Fouque MJ, Rochaix P, Noel A, Foidart JM, Krust A, Chambon P, Brouchet L, Arnal JF, Lenfant F (2012) Stromal estrogen receptoralpha promotes tumor growth by normalizing an increased angiogenesis. Cancer Res 72(12):3010– 3019. [https://doi.org/10.1158/0008-5472.CAN-11-](https://doi.org/10.1158/0008-5472.CAN-11-3768) [3768.](https://doi.org/10.1158/0008-5472.CAN-11-3768) PubMed PMID: 22523036
- 52. Segawa T, Shozu M, Murakami K, Kasai T, Shinohara K, Nomura K, Ohno S, Inoue M (2005) Aromatase expression in stromal cells of endometrioid endometrial cancer correlates with poor survival. Clin Cancer Res 11(6):2188–2194. [https://](https://doi.org/10.1158/1078-0432.CCR-04-1859) [doi.org/10.1158/1078-0432.CCR-04-1859.](https://doi.org/10.1158/1078-0432.CCR-04-1859) PubMed PMID: 15788666
- 53. Knower KC, Chand AL, Eriksson N, Takagi K, Miki Y, Sasano H, Visvader JE, Lindeman GJ, Funder JW, Fuller PJ, Simpson ER, Tilley WD, Leedman PJ, Graham J, Muscat GE, Clarke CL, Clyne CD (2013) Distinct nuclear receptor expression in stroma adjacent to breast tumors. Breast Cancer Res Treat 142(1):211–223. [https://doi.org/10.1007/s10549-](https://doi.org/10.1007/s10549-013-2716-6) [013-2716-6.](https://doi.org/10.1007/s10549-013-2716-6) PubMed PMID: 24122391
- 54. Daniels G, Gellert LL, Melamed J, Hatcher D, Li Y, Wei J, Wang J, Lee P (2014) Decreased expression of stromal estrogen receptor alpha and beta in prostate cancer. Am J Transl Res 6(2):140–146. PubMed PMID: 24489993; PMCID: PMC3902224
- 55. Leav I, Lau KM, Adams JY, McNeal JE, Taplin ME, Wang J, Singh H, Ho SM (2001) Comparative studies of the estrogen receptors beta and alpha and the androgen receptor in normal human prostate glands, dysplasia, and in primary and metastatic carci-

noma. Am J Pathol 159(1):79–92. PubMed PMID: 11438457; PMCID: PMC1850428

- 56. Subramaniam KS, Tham ST, Mohamed Z, Woo YL, Mat Adenan NA, Chung I (2013) Cancer-associated fibroblasts promote proliferation of endometrial cancer cells. PLoS One 8(7):e68923. [https://doi.](https://doi.org/10.1371/journal.pone.0068923) [org/10.1371/journal.pone.0068923](https://doi.org/10.1371/journal.pone.0068923). PubMed PMID: 23922669; PMCID: PMC3724864
- 57. Svoronos N, Perales-Puchalt A, Allegrezza MJ, Rutkowski MR, Payne KK, Tesone AJ, Nguyen JM, Curiel TJ, Cadungog MG, Singhal S, Eruslanov EB, Zhang P, Tchou J, Zhang R, Conejo-Garcia JR (2017) Tumor cell-independent Estrogen signaling drives disease progression through mobilization of myeloid-derived suppressor cells. Cancer Discov 7(1):72–85. [https://doi.org/10.1158/2159-8290.](https://doi.org/10.1158/2159-8290.CD-16-0502) [CD-16-0502.](https://doi.org/10.1158/2159-8290.CD-16-0502) PubMed PMID: 27694385; PMCID: 5222699
- 58. Ciucci A, Zannoni GF, Buttarelli M, Lisi L, Travaglia D, Martinelli E, Scambia G, Gallo D (2016) Multiple direct and indirect mechanisms drive estrogen-induced tumor growth in high grade serous ovarian cancers. Oncotarget 7(7):8155–8171. <https://doi.org/10.18632/oncotarget.6943>. PubMed PMID: 26797759; PMCID: PMC4884983
- 59. Mor G, Yue W, Santen RJ, Gutierrez L, Eliza M, Berstein LM, Harada N, Wang J, Lysiak J, Diano S, Naftolin F (1998) Macrophages, estrogen and the microenvironment of breast cancer. J Steroid Biochem Mol Biol 67(5–6):403–411. PubMed PMID: 10030689
- 60. Stabile LP, Rothstein ME, Cunningham DE, Land SR, Dacic S, Keohavong P, Siegfried JM (2012) Prevention of tobacco carcinogen-induced lung cancer in female mice using antiestrogens. Carcinogenesis 33(11):2181–2189
- 61. Curran EM, Berghaus LJ, Vernetti NJ, Saporita AJ, Lubahn DB, Estes DM (2001) Natural killer cells express estrogen receptor-α and estrogen receptor-β and can respond to estrogen via a non-estrogen receptor-α-mediated pathway. Cell Immunol 214(1):12–20
- 62. Michalek RD, Gerriets VA, Nichols AG, Inoue M, Kazmin D, Chang C-Y, Dwyer MA, Nelson ER, Pollizzi KN, Ilkayeva O (2011) Estrogen-related receptor-α is a metabolic regulator of effector T-cell activation and differentiation. Proc Natl Acad Sci 108(45):18348–18353
- 63. Adurthi S, Kumar MM, Vinodkumar H, Mukherjee G, Krishnamurthy H, Acharya KK, Bafna U, Uma DK, Abhishekh B, Krishna S (2017) Oestrogen receptor-α binds the FOXP3 promoter and modulates regulatory T-cell function in human cervical cancer. Sci Rep 7(1):17289
- 64. Rothenberger NJ, Somasundaram A, Stabile LP (2018) The role of the estrogen pathway in the tumor microenvironment. Int J Mol Sci 19(2):611
- 65. Fridman WH, Pages F, Sautes-Fridman C, Galon J (2012) The immune contexture in human tumours:

impact on clinical outcome. Nat Rev Cancer 12(4):298–306. <https://doi.org/10.1038/nrc3245>. PubMed PMID: 22419253

- 66. Haabeth OA, Lorvik KB, Hammarstrom C, Donaldson IM, Haraldsen G, Bogen B, Corthay A (2011) Inflammation driven by tumour-specific Th1 cells protects against B-cell cancer. Nat Commun 2:240. <https://doi.org/10.1038/ncomms1239>. PubMed PMID: 21407206; PMCID: PMC3072106
- 67. DeNardo DG, Barreto JB, Andreu P, Vasquez L, Tawfik D, Kolhatkar N, Coussens LM (2009) CD4(+) T cells regulate pulmonary metastasis of mammary carcinomas by enhancing protumor properties of macrophages. Cancer Cell 16(2):91–102. <https://doi.org/10.1016/j.ccr.2009.06.018>. PubMed PMID: 19647220; PMCID: PMC2778576
- 68. Dannenfelser R, Nome M, Tahiri A, Ursini-Siegel J, Vollan HKM, Haakensen VD, Helland A, Naume B, Caldas C, Borresen-Dale AL, Kristensen VN, Troyanskaya OG (2017) Data-driven analysis of immune infiltrate in a large cohort of breast cancer and its association with disease progression, ER activity, and genomic complexity. Oncotarget 8(34):57121–57133. [https://doi.org/10.18632/onco](https://doi.org/10.18632/oncotarget.19078)[target.19078.](https://doi.org/10.18632/oncotarget.19078) PubMed PMID: 28915659; PMCID: PMC5593630
- 69. Ali HR, Provenzano E, Dawson SJ, Blows FM, Liu B, Shah M, Earl HM, Poole CJ, Hiller L, Dunn JA, Bowden SJ, Twelves C, Bartlett JM, Mahmoud SM, Rakha E, Ellis IO, Liu S, Gao D, Nielsen TO, Pharoah PD, Caldas C (2014) Association between CD8+ T-cell infiltration and breast cancer survival in 12,439 patients. Ann Oncol 25(8):1536–1543. [https://doi.org/10.1093/annonc/mdu191.](https://doi.org/10.1093/annonc/mdu191) PubMed PMID: 24915873
- 70. Cullen SP, Martin SJ (2008) Mechanisms of granuledependent killing. Cell Death Differ 15(2):251–262. <https://doi.org/10.1038/sj.cdd.4402244>. PubMed PMID: 17975553
- 71. Lieberman J (2003) The ABCs of granule-mediated cytotoxicity: new weapons in the arsenal. Nat Rev Immunol 3(5):361–370. [https://doi.org/10.1038/](https://doi.org/10.1038/nri1083) [nri1083](https://doi.org/10.1038/nri1083). PubMed PMID: 12766758
- 72. Jiang X, Orr BA, Kranz DM, Shapiro DJ (2006) Estrogen induction of the granzyme B inhibitor, proteinase inhibitor 9, protects cells against apoptosis mediated by cytotoxic T lymphocytes and natural killer cells. Endocrinology 147(3):1419–1426. <https://doi.org/10.1210/en.2005-0996>. PubMed PMID: 16306080
- 73. Jiang X, Ellison SJ, Alarid ET, Shapiro DJ (2007) Interplay between the levels of estrogen and estrogen receptor controls the level of the granzyme inhibitor, proteinase inhibitor 9 and susceptibility to immune surveillance by natural killer cells. Oncogene 26(28):4106–4114. [https://doi.org/10.1038/](https://doi.org/10.1038/sj.onc.1210197) [sj.onc.1210197.](https://doi.org/10.1038/sj.onc.1210197) PubMed PMID: 17237823
- 74. Tanaka A, Sakaguchi S (2017) Regulatory T cells in cancer immunotherapy. Cell Res 27(1):109–

118. [https://doi.org/10.1038/cr.2016.151.](https://doi.org/10.1038/cr.2016.151) PubMed PMID: 27995907; PMCID: PMC5223231

- 75. Tai P, Wang J, Jin H, Song X, Yan J, Kang Y, Zhao L, An X, Du X, Chen X, Wang S, Xia G, Wang B (2008) Induction of regulatory T cells by physiological level estrogen. J Cell Physiol 214(2):456–464. [https://doi.org/10.1002/jcp.21221.](https://doi.org/10.1002/jcp.21221) PubMed PMID: 17654501
- 76. Polanczyk MJ, Carson BD, Subramanian S, Afentoulis M, Vandenbark AA, Ziegler SF, Offner H (2004) Cutting edge: estrogen drives expansion of the CD4+CD25+ regulatory T cell compartment. J Immunol 173(4):2227–2230. PubMed PMID: 15294932
- 77. Fontenot JD, Gavin MA, Rudensky AY (2003) Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. Nat Immunol 4(4):330–336. <https://doi.org/10.1038/ni904>. PubMed PMID: 12612578
- 78. Chaudhary B, Elkord E (2016) Regulatory T cells in the tumor microenvironment and cancer progression: role and therapeutic targeting. Vaccines (Basel) 4(3). [https://doi.org/10.3390/vac](https://doi.org/10.3390/vaccines4030028)[cines4030028](https://doi.org/10.3390/vaccines4030028). PubMed PMID: 27509527; PMCID: PMC5041022
- 79. Kadota K, Eguchi T, Villena-Vargas J, Woo KM, Sima CS, Jones DR, Travis WD, Adusumilli PS (2015) Nuclear estrogen receptor-alpha expression is an independent predictor of recurrence in male patients with pT1aN0 lung adenocarcinomas, and correlates with regulatory T-cell infiltration. Oncotarget 6(29):27505–27518. [https://doi.](https://doi.org/10.18632/oncotarget.4752) [org/10.18632/oncotarget.4752.](https://doi.org/10.18632/oncotarget.4752) PubMed PMID: 26318038; PMCID: PMC4695005
- 80. Shang B, Liu Y, Jiang SJ, Liu Y (2015) Prognostic value of tumor-infiltrating FoxP3+ regulatory T cells in cancers: a systematic review and meta-analysis. Sci Rep 5:15179.<https://doi.org/10.1038/srep15179>. PubMed PMID: 26462617; PMCID: PMC4604472
- 81. Generali D, Bates G, Berruti A, Brizzi MP, Campo L, Bonardi S, Bersiga A, Allevi G, Milani M, Aguggini S, Dogliotti L, Banham AH, Harris AL, Bottini A, Fox SB (2009) Immunomodulation of FOXP3+ regulatory T cells by the aromatase inhibitor letrozole in breast cancer patients. Clin Cancer Res 15(3):1046–1051. [https://doi.org/10.1158/1078-](https://doi.org/10.1158/1078-0432.CCR-08-1507) [0432.CCR-08-1507](https://doi.org/10.1158/1078-0432.CCR-08-1507). PubMed PMID: 19188178
- 82. Polanczyk MJ, Hopke C, Vandenbark AA, Offner H (2007) Treg suppressive activity involves estrogendependent expression of programmed death-1 (PD-1). Int Immunol 19(3):337–343. [https://doi.](https://doi.org/10.1093/intimm/dxl151) [org/10.1093/intimm/dxl151](https://doi.org/10.1093/intimm/dxl151). PubMed PMID: 17267414
- 83. Yang L, Huang F, Mei J, Wang X, Zhang Q, Wang H, Xi M, You Z (2017) Posttranscriptional control of PD-L1 expression by 17beta-Estradiol via PI3K/Akt signaling pathway in ERalpha-positive cancer cell lines. Int J Gynecol Cancer 27(2):196–205. [https://](https://doi.org/10.1097/IGC.0000000000000875) doi.org/10.1097/IGC.0000000000000875. PubMed PMID: 27870715; PMCID: PMC5258765
- 84. Jiang Y, Li Y, Zhu B (2015) T-cell exhaustion in the tumor microenvironment. Cell Death Dis 6:e1792. <https://doi.org/10.1038/cddis.2015.162>. PubMed PMID: 26086965; PMCID: PMC4669840
- 85. Matsumoto M, Yamaguchi Y, Seino Y, Hatakeyama A, Takei H, Niikura H, Ito K, Suzuki T, Sasano H, Yaegashi N, Hayashi S (2008) Estrogen signaling ability in human endometrial cancer through the cancer-stromal interaction. Endocr Relat Cancer 15(2):451–463. [https://doi.org/10.1677/ERC-07-](https://doi.org/10.1677/ERC-07-0227) [0227.](https://doi.org/10.1677/ERC-07-0227) PubMed PMID: 18508998
- 86. Subbaramaiah K, Morris PG, Zhou XK, Morrow M, Du B, Giri D, Kopelovich L, Hudis CA, Dannenberg AJ (2012) Increased levels of COX-2 and prostaglandin E2 contribute to elevated aromatase expression in inflamed breast tissue of obese women. Cancer Discov 2(4):356–365. [https://doi.org/10.1158/2159-](https://doi.org/10.1158/2159-8290.CD-11-0241) [8290.CD-11-0241](https://doi.org/10.1158/2159-8290.CD-11-0241). PubMed PMID: 22576212; PMCID: PMC3398487
- 87. Subbaramaiah K, Howe LR, Bhardwaj P, Du B, Gravaghi C, Yantiss RK, Zhou XK, Blaho VA, Hla T, Yang P, Kopelovich L, Hudis CA, Dannenberg AJ (2011) Obesity is associated with inflammation and elevated aromatase expression in the mouse mammary gland. Cancer Prev Res (Phila) 4(3):329–346. <https://doi.org/10.1158/1940-6207.CAPR-10-0381>. PubMed PMID: 21372033; PMCID: PMC3071249
- 88. Birbrair A, Zhang T, Wang Z-M, Messi ML, Olson JD, Mintz A, Delbono O (2014) Type-2 pericytes participate in normal and tumoral angiogenesis. Am J Phys Cell Phys 307(1):C25–C38
- 89. Sortino MA, Platania P, Chisari M, Merlo S, Copani A, Catania MV (2005) A major role for astrocytes in the neuroprotective effect of estrogen. Drug Dev Res 66(2):126–135
- 90. Bukovsky A, Cekanova M, Caudle MR, Wimalasena J, Foster JS, Henley DC, Elder RF (2003) Expression and localization of estrogen receptor-alpha protein in normal and abnormal term placentae and stimulation of trophoblast differentiation by estradiol. Reprod Biol Endocrinol 1(1):13
- 91. Xing F, Saidou J, Watabe K (2010) Cancer associated fibroblasts (CAFs) in tumor microenvironment. Front Biosci (Landmark Ed) 15:166–79. PubMed PMID: 20036813; PMCID: PMC2905156
- 92. Brechbuhl HM, Finlay-Schultz J, Yamamoto TM, Gillen AE, Cittelly DM, Tan A-C, Sams SB, Pillai MM, Elias AD, Robinson WA (2017) Fibroblast subtypes regulate responsiveness of luminal breast cancer to estrogen. Clin Cancer Res 23(7):1710–1721
- 93. Annicotte JS, Chavey C, Servant N, Teyssier J, Bardin A, Licznar A, Badia E, Pujol P, Vignon F, Maudelonde T, Lazennec G, Cavailles V, Fajas L (2005) The nuclear receptor liver receptor homolog-1 is an estrogen receptor target gene. Oncogene 24(55):8167–8175. [https://doi.org/10.1038/](https://doi.org/10.1038/sj.onc.1208950) [sj.onc.1208950.](https://doi.org/10.1038/sj.onc.1208950) PubMed PMID: 16091743; PMCID: PMC2259230
- 94. Clyne CD, Kovacic A, Speed CJ, Zhou J, Pezzi V, Simpson ER (2004) Regulation of aromatase expres-

sion by the nuclear receptor LRH-1 in adipose tissue. Mol Cell Endocrinol 215(1–2):39–44. [https://doi.](https://doi.org/10.1016/j.mce.2003.11.001) [org/10.1016/j.mce.2003.11.001.](https://doi.org/10.1016/j.mce.2003.11.001) PubMed PMID: 15026173

- 95. Chand AL, Herridge KA, Howard TL, Simpson ER, Clyne CD (2011) Tissue-specific regulation of aromatase promoter II by the orphan nuclear receptor LRH-1 in breast adipose stromal fibroblasts. Steroids 76(8):741–744. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.steroids.2011.02.024) [steroids.2011.02.024](https://doi.org/10.1016/j.steroids.2011.02.024). PubMed PMID: 21392518
- 96. Miki Y, Clyne CD, Suzuki T, Moriya T, Shibuya R, Nakamura Y, Ishida T, Yabuki N, Kitada K, Hayashi S, Sasano H (2006) Immunolocalization of liver receptor homologue-1 (LRH-1) in human breast carcinoma: possible regulator of insitu steroidogenesis. Cancer Lett 244(1):24–33. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.canlet.2005.11.038) [canlet.2005.11.038.](https://doi.org/10.1016/j.canlet.2005.11.038) PubMed PMID: 16427184
- 97. Guo RX, Wei LH, Tu Z, Sun PM, Wang JL, Zhao D, Li XP, Tang JM (2006) 17 beta-estradiol activates PI3K/Akt signaling pathway by estrogen receptor (ER)-dependent and ER-independent mechanisms in endometrial cancer cells. J Steroid Biochem Mol Biol 99(1):9–18. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.jsbmb.2005.11.013) [jsbmb.2005.11.013](https://doi.org/10.1016/j.jsbmb.2005.11.013). PubMed PMID: 16567092
- 98. Stabile LP, Lyker JS, Gubish CT, Zhang W, Grandis JR, Siegfried JM (2005) Combined targeting of the estrogen receptor and the epidermal growth factor receptor in non-small cell lung cancer shows enhanced antiproliferative effects. Cancer Res 65(4):1459–1470. [https://doi.org/10.1158/0008-](https://doi.org/10.1158/0008-5472.CAN-04-1872) [5472.CAN-04-1872](https://doi.org/10.1158/0008-5472.CAN-04-1872). PubMed PMID: 15735034
- 99. Keshamouni VG, Mattingly RR, Reddy KB (2002) Mechanism of 17-beta-estradiol-induced Erk1/2 activation in breast cancer cells. A role for HER2 AND PKC-delta. J Biol Chem 277(25):22558– 22565. <https://doi.org/10.1074/jbc.M202351200>. PubMed PMID: 11960991
- 100. Yeh CR, Slavin S, Da J, Hsu I, Luo J, Xiao GQ, Ding J, Chou FJ, Yeh S (2016) Estrogen receptor alpha in cancer associated fibroblasts suppresses prostate cancer invasion via reducing CCL5, IL6 and macrophage infiltration in the tumor microenvironment. Mol Cancer 15:7. [https://doi.org/10.1186/s12943-](https://doi.org/10.1186/s12943-015-0488-9) [015-0488-9.](https://doi.org/10.1186/s12943-015-0488-9) PubMed PMID: 26790618; PMCID: PMC4721150
- 101. Slavin S, Yeh CR, Da J, Yu S, Miyamoto H, Messing EM, Guancial E, Yeh S (2014) Estrogen receptor alpha in cancer-associated fibroblasts suppresses prostate cancer invasion via modulation of thrombospondin 2 and matrix metalloproteinase 3. Carcinogenesis 35(6):1301–1309. [https://](https://doi.org/10.1093/carcin/bgt488) doi.org/10.1093/carcin/bgt488. PubMed PMID: 24374826; PMCID: PMC4043239
- 102. Aldinucci D, Colombatti A (2014) The inflammatory chemokine CCL5 and cancer progression. Mediat Inflamm 2014:292376. [https://](https://doi.org/10.1155/2014/292376) doi.org/10.1155/2014/292376. PubMed PMID: 24523569; PMCID: PMC3910068
- 103. Kumari N, Dwarakanath BS, Das A, Bhatt AN (2016) Role of interleukin-6 in cancer progression and

therapeutic resistance. Tumour Biol 37(9):11553– 11572. <https://doi.org/10.1007/s13277-016-5098-7>. PubMed PMID: 27260630

- 104. Qian BZ, Pollard JW (2010) Macrophage diversity enhances tumor progression and metastasis. Cell 141(1):39–51. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cell.2010.03.014) [cell.2010.03.014.](https://doi.org/10.1016/j.cell.2010.03.014) PubMed PMID: 20371344; PMCID: PMC4994190
- 105. Liu Y, Cao X (2015) The origin and function of tumor-associated macrophages. Cell Mol Immunol 12(1):1–4. [https://doi.org/10.1038/](https://doi.org/10.1038/cmi.2014.83) [cmi.2014.83](https://doi.org/10.1038/cmi.2014.83). PubMed PMID: 25220733; PMCID: PMC4654376
- 106. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A (2002) Macrophage polarization: tumorassociated macrophages as a paradigm for polarized M2 mononuclear phagocytes. Trends Immunol 23(11):549–55. PubMed PMID: 12401408
- 107. Lee S, Margolin K (2011) Cytokines in cancer immunotherapy. Cancers (Basel) 3(4):3856–3893. <https://doi.org/10.3390/cancers3043856>. PubMed PMID: 24213115; PMCID: PMC3763400
- 108. Bingle L, Brown NJ, Lewis CE (2002) The role of tumour-associated macrophages in tumour progression: implications for new anticancer therapies. J Pathol 196(3):254–265. [https://doi.org/10.1002/](https://doi.org/10.1002/path.1027) [path.1027.](https://doi.org/10.1002/path.1027) PubMed PMID: 11857487
- 109. Wan T, Liu JH, Zheng LM, Cai MY, Ding T (2009) Prognostic significance of tumor-associated macrophage infiltration in advanced epithelial ovarian carcinoma. Ai Zheng 28(3):323–7. PubMed PMID: 19619451
- 110. Gwak JM, Jang MH, Kim DI, Seo AN, Park SY (2015) Prognostic value of tumor-associated macrophages according to histologic locations and hormone receptor status in breast cancer. PLoS One 10(4):e0125728. <https://doi.org/10.1371/journal.pone.0125728>. PubMed PMID: 25884955; PMCID: PMC4401667
- 111. Campbell MJ, Tonlaar NY, Garwood ER, Huo D, Moore DH, Khramtsov AI, Au A, Baehner F, Chen Y, Malaka DO, Lin A, Adeyanju OO, Li S, Gong C, McGrath M, Olopade OI, Esserman LJ (2011) Proliferating macrophages associated with high grade, hormone receptor negative breast cancer and poor clinical outcome. Breast Cancer Res Treat 128(3):703– 711. <https://doi.org/10.1007/s10549-010-1154-y>. PubMed PMID: 20842526; PMCID: PMC4657137
- 112. Svensson S, Abrahamsson A, Rodriguez GV, Olsson AK, Jensen L, Cao Y, Dabrosin C (2015) CCL2 and CCL5 Are novel therapeutic targets for Estrogen-dependent breast cancer. Clin Cancer Res 21(16):3794–3805. [https://doi.org/10.1158/1078-](https://doi.org/10.1158/1078-0432.CCR-15-0204) [0432.CCR-15-0204](https://doi.org/10.1158/1078-0432.CCR-15-0204). PubMed PMID: 25901081
- 113. Okizaki S, Ito Y, Hosono K, Oba K, Ohkubo H, Kojo K, Nishizawa N, Shibuya M, Shichiri M, Majima M (2016) Vascular endothelial growth factor receptor type 1 signaling prevents delayed wound healing in diabetes by attenuating the production of IL-1beta by recruited macrophages. Am J Pathol 186(6):1481– 1498. <https://doi.org/10.1016/j.ajpath.2016.02.014>. PubMed PMID: 27085138
- 114. Stabile LP, Farooqui M, Kanterewicz B, Abberbock S, Kurland BF, Diergaarde B, Siegfried JM (2017) Preclinical evidence for combined use of aromatase inhibitors and NSAIDs as preventive agents of tobacco-induced lung cancer. J Thorac Oncol. [https://doi.org/10.1016/j.jtho.2017.11.126.](https://doi.org/10.1016/j.jtho.2017.11.126) PubMed PMID: 29233790
- 115. Ning C, Xie B, Zhang L, Li C, Shan W, Yang B, Luo X, Gu C, He Q, Jin H, Chen X, Zhang Z, Feng Y (2016) Infiltrating macrophages induce ERalpha expression through an IL17A-mediated epigenetic mechanism to sensitize endometrial cancer cells to Estrogen. Cancer Res 76(6):1354–1366. [https://doi.](https://doi.org/10.1158/0008-5472.CAN-15-1260) [org/10.1158/0008-5472.CAN-15-1260.](https://doi.org/10.1158/0008-5472.CAN-15-1260) PubMed PMID: 26744532
- 116. Sun L, Chen B, Jiang R, Li J, Wang B (2017) Resveratrol inhibits lung cancer growth by suppressing M2-like polarization of tumor associated macrophages. Cell Immunol 311:86–93. [https://](https://doi.org/10.1016/j.cellimm.2016.11.002) [doi.org/10.1016/j.cellimm.2016.11.002.](https://doi.org/10.1016/j.cellimm.2016.11.002) PubMed PMID: 27825563
- 117. Umansky V, Blattner C, Gebhardt C, Utikal J (2016) The Role of Myeloid-Derived Suppressor Cells (MDSC) in cancer progression. Vaccines (Basel) 4(4). <https://doi.org/10.3390/vaccines4040036>. PubMed PMID: 27827871; PMCID: PMC5192356
- 118. Gabrilovich DI, Ostrand-Rosenberg S, Bronte V (2012) Coordinated regulation of myeloid cells by tumours. Nat Rev Immunol 12(4):253–268. [https://doi.org/10.1038/nri3175.](https://doi.org/10.1038/nri3175) PubMed PMID: 22437938; PMCID: PMC3587148
- 119. Márquez-Garbán DC, Deng G, Comin-Anduix B, Garcia AJ, Xing Y, Chen H-W, Cheung-Lau G, Hamilton N, Jung ME, Pietras RJ (2019) Antiestrogens in combination with immune checkpoint inhibitors in breast cancer immunotherapy. J Steroid Biochem Mol Biol 193:105415
- 120. Kozasa K, Mabuchi S, Matsumoto Y, Kuroda H, Yokoi E, Komura N, Kawano M, Takahashi R, Sasano T, Shimura K (2019) Estrogen stimulates female cancer progression by inducing myeloidderived suppressive cells: investigations on pregnant and non-pregnant experimental models. Oncotarget 10(20):1887
- 121. Yoshimura A (2006) Signal transduction of inflammatory cytokines and tumor development. Cancer Sci 97(6):439–447. [https://doi.](https://doi.org/10.1111/j.1349-7006.2006.00197.x) [org/10.1111/j.1349-7006.2006.00197.x](https://doi.org/10.1111/j.1349-7006.2006.00197.x). PubMed PMID: 16734720
- 122. Sasser AK, Sullivan NJ, Studebaker AW, Hendey LF, Axel AE, Hall BM (2007) Interleukin-6 is a potent growth factor for ER-alpha-positive human breast cancer. FASEB J 21(13):3763–3770. [https://](https://doi.org/10.1096/fj.07-8832com) [doi.org/10.1096/fj.07-8832com.](https://doi.org/10.1096/fj.07-8832com) PubMed PMID: 17586727
- 123. Studebaker AW, Storci G, Werbeck JL, Sansone P, Sasser AK, Tavolari S, Huang T, Chan MW, Marini FC, Rosol TJ, Bonafe M, Hall BM (2008) Fibroblasts isolated from common sites of breast cancer metastasis enhance cancer cell growth rates and invasiveness in an interleukin-6-dependent manner. Cancer Res 68(21):9087–9095. [https://doi.](https://doi.org/10.1158/0008-5472.CAN-08-0400)

[org/10.1158/0008-5472.CAN-08-0400.](https://doi.org/10.1158/0008-5472.CAN-08-0400) PubMed PMID: 18974155

- 124. Yin Y, Chen X, Shu Y (2009) Gene expression of the invasive phenotype of TNF-alpha-treated MCF-7 cells. Biomed Pharmacother 63(6):421–428. [https://](https://doi.org/10.1016/j.biopha.2009.04.032) [doi.org/10.1016/j.biopha.2009.04.032.](https://doi.org/10.1016/j.biopha.2009.04.032) PubMed PMID: 19564093
- 125. Zhao Y, Nichols JE, Valdez R, Mendelson CR, Simpson ER (1996) Tumor necrosis factor-alpha stimulates aromatase gene expression in human adipose stromal cells through use of an activating protein-1 binding site upstream of promoter 1.4. Mol Endocrinol 10(11):1350–1357. [https://doi.](https://doi.org/10.1210/mend.10.11.8923461) [org/10.1210/mend.10.11.8923461](https://doi.org/10.1210/mend.10.11.8923461). PubMed PMID: 8923461
- 126. Irahara N, Miyoshi Y, Taguchi T, Tamaki Y, Noguchi S (2006) Quantitative analysis of aromatase mRNA expression derived from various promoters (I.4, I.3, PII and I.7) and its association with expression of TNF-alpha, IL-6 and COX-2 mRNAs in human breast cancer. Int J Cancer 118(8):1915–1921. [https://doi.org/10.1002/ijc.21562.](https://doi.org/10.1002/ijc.21562) PubMed PMID: 16287071
- 127. Ricciotti E, FitzGerald GA (2011) Prostaglandins and inflammation. Arterioscler Thromb Vasc Biol 31(5):986–1000. [https://doi.org/10.1161/](https://doi.org/10.1161/ATVBAHA.110.207449) [ATVBAHA.110.207449.](https://doi.org/10.1161/ATVBAHA.110.207449) PubMed PMID: 21508345; PMCID: PMC3081099
- 128. Zhao Y, Agarwal VR, Mendelson CR, Simpson ER (1996) Estrogen biosynthesis proximal to a breast tumor is stimulated by PGE2 via cyclic AMP, leading to activation of promoter II of the CYP19 (aromatase) gene. Endocrinology 137(12):5739–5742. <https://doi.org/10.1210/endo.137.12.8940410>. PubMed PMID: 8940410
- 129. Terry MB, Gammon MD, Zhang FF, Tawfik H, Teitelbaum SL, Britton JA, Subbaramaiah K, Dannenberg AJ, Neugut AI (2004) Association of frequency and duration of aspirin use and hormone receptor status with breast cancer risk. JAMA 291(20):2433–2440. [https://doi.org/10.1001/](https://doi.org/10.1001/jama.291.20.2433) [jama.291.20.2433](https://doi.org/10.1001/jama.291.20.2433). PubMed PMID: 15161893
- 130. Zhou XL, Fan W, Yang G, Yu MX (2014) The clinical significance of PR, ER, NF- kappa B, and TNFalpha in breast cancer. Dis Markers 2014:494581. [https://doi.org/10.1155/2014/494581.](https://doi.org/10.1155/2014/494581) PubMed PMID: 24864130; PMCID: PMC4017837
- 131. Hoesel B, Schmid JA (2013) The complexity of NF-kappaB signaling in inflammation and cancer. Mol Cancer 12:86. [https://doi.org/10.1186/1476-](https://doi.org/10.1186/1476-4598-12-86) [4598-12-86.](https://doi.org/10.1186/1476-4598-12-86) PubMed PMID: 23915189; PMCID: PMC3750319
- 132. Johnston SR, Lu B, Scott GK, Kushner PJ, Smith IE, Dowsett M, Benz CC (1999) Increased activator protein-1 DNA binding and c-Jun NH2-terminal kinase activity in human breast tumors with acquired tamoxifen resistance. Clin Cancer Res 5(2):251– 256. PubMed PMID: 10037172
- 133. Zhou Y, Yau C, Gray JW, Chew K, Dairkee SH, Moore DH, Eppenberger U, Eppenberger-Castori S, Benz CC (2007) Enhanced NF kappa B and AP-1 transcriptional activity associated with antiestrogen

resistant breast cancer. BMC Cancer 7:59. [https://](https://doi.org/10.1186/1471-2407-7-59) [doi.org/10.1186/1471-2407-7-59.](https://doi.org/10.1186/1471-2407-7-59) PubMed PMID: 17407600; PMCID: PMC1852565

- 134. Guffey CR, Fan D, Singh UP, Murphy EA (2013) Linking obesity to colorectal cancer: recent insights into plausible biological mechanisms. Curr Opin Clin Nutr Metab Care 16(5):595–600. Epub 2013/06/08. <https://doi.org/10.1097/MCO.0b013e328362d10b>. PubMed PMID: 23743611
- 135. Lin JH, Morikawa T, Chan AT, Kuchiba A, Shima K, Nosho K, Kirkner G, Zhang SM, Manson JE, Giovannucci E, Fuchs CS, Ogino S (2012) Postmenopausal hormone therapy is associated with a reduced risk of colorectal cancer lacking CDKN1A expression. Cancer Res 72(12):3020–8. Epub 2012/04/19. [https://doi.org/10.1158/0008-5472.](https://doi.org/10.1158/0008-5472.CAN-11-2619) [CAN-11-2619](https://doi.org/10.1158/0008-5472.CAN-11-2619). PubMed PMID: 22511578; PMCID: PMC3377852
- 136. Wang Z, Aguilar EG, Luna JI, Dunai C, Khuat LT, Le CT, Mirsoian A, Minnar CM, Stoffel KM, Sturgill IR (2019) Paradoxical effects of obesity on T cell function during tumor progression and PD-1 checkpoint blockade. Nat Med 25(1):141
- 137. Crespi E, Bottai G, Santarpia L (2016) Role of inflammation in obesity-related breast cancer. Curr Opin Pharmacol 31:114–22. Epub 2016/11/28. [https://doi.](https://doi.org/10.1016/j.coph.2016.11.004) [org/10.1016/j.coph.2016.11.004](https://doi.org/10.1016/j.coph.2016.11.004). PubMed PMID: 27889687
- 138. Barzi A, Lenz AM, Labonte MJ, Lenz H-J (2013) Molecular pathways: estrogen pathway in colorectal cancer. Clin Cancer Res 19(21):5842–5848
- 139. Deguchi K, Kamada M, Irahara M, Maegawa M, Yamamoto S, Ohmoto Y, Murata K, Yasui T, Yamano S, Aono T (2001) Postmenopausal changes in production of type 1 and type 2 cytokines and the effects of hormone replacement therapy. Menopause 8(4):266–273. Epub 2001/07/13. PubMed PMID: 11449084
- 140. Kamada M, Irahara M, Maegawa M, Ohmoto Y, Murata K, Yasui T, Yamano S, Aono T (2001) Transient increase in the levels of T-helper 1 cytokines in postmenopausal women and the effects of hormone replacement therapy. Gynecol Obstet Investig 52(2):82–8. Epub 2001/10/05. [https://](https://doi.org/10.1159/000052948) [doi.org/10.1159/000052948.](https://doi.org/10.1159/000052948) PubMed PMID: 11586033
- 141. Berg G, Ekerfelt C, Hammar M, Lindgren R, Matthiesen L, Ernerudh J (2002) Cytokine changes in postmenopausal women treated with estrogens: a placebo-controlled study. Am J Reprod Immunol 48(2):63–69. Epub 2002/10/23. PubMed PMID: 12389594
- 142. Pardoll DM (2012) The blockade of immune checkpoints in cancer immunotherapy. Nat Rev Cancer 12(4):252–264. <https://doi.org/10.1038/nrc3239>. PubMed PMID: 22437870; PMCID: PMC4856023
- 143. Wolchok JD, Chiarion-Sileni V, Gonzalez R, Rutkowski P, Grob JJ, Cowey CL, Lao CD, Wagstaff J, Schadendorf D, Ferrucci PF, Smylie M, Dummer R, Hill A, Hogg D, Haanen J, Carlino MS,

Bechter O, Maio M, Marquez-Rodas I, Guidoboni M, McArthur G, Lebbe C, Ascierto PA, Long GV, Cebon J, Sosman J, Postow MA, Callahan MK, Walker D, Rollin L, Bhore R, Hodi FS, Larkin J (2017) Overall survival with combined Nivolumab and Ipilimumab in advanced melanoma. N Engl J Med 377(14):1345–1356. [https://doi.org/10.1056/](https://doi.org/10.1056/NEJMoa1709684) [NEJMoa1709684.](https://doi.org/10.1056/NEJMoa1709684) PubMed PMID: 28889792; PMCID: PMC5706778

- 144. Reck M, Rodriguez-Abreu D, Robinson AG, Hui R, Csoszi T, Fulop A, Gottfried M, Peled N, Tafreshi A, Cuffe S, O'Brien M, Rao S, Hotta K, Leiby MA, Lubiniecki GM, Shentu Y, Rangwala R, Brahmer JR (2016) Investigators K-. Pembrolizumab versus chemotherapy for PD-L1-positive non-small-cell lung cancer. N Engl J Med 375(19):1823–1833. [https://](https://doi.org/10.1056/NEJMoa1606774) doi.org/10.1056/NEJMoa1606774. PubMed PMID: 27718847
- 145. Brahmer J, Reckamp KL, Baas P, Crino L, Eberhardt WE, Poddubskaya E, Antonia S, Pluzanski A, Vokes EE, Holgado E, Waterhouse D, Ready N, Gainor J, Aren Frontera O, Havel L, Steins M, Garassino MC, Aerts JG, Domine M, Paz-Ares L, Reck M, Baudelet C, Harbison CT, Lestini B, Spigel DR (2015) Nivolumab versus docetaxel in advanced squamous-cell non-small-cell lung cancer. N Engl J Med 373(2):123–135. [https://doi.org/10.1056/](https://doi.org/10.1056/NEJMoa1504627) [NEJMoa1504627.](https://doi.org/10.1056/NEJMoa1504627) PubMed PMID: 26028407; PMCID: 4681400
- 146. Wang X, Bao Z, Zhang X, Li F, Lai T, Cao C, Chen Z, Li W, Shen H, Ying S (2017) Effectiveness and safety of PD-1/PD-L1 inhibitors in the treatment of solid tumors: a systematic review and meta-analysis. Oncotarget 8(35):59901–59914. [https://doi.](https://doi.org/10.18632/oncotarget.18316) [org/10.18632/oncotarget.18316.](https://doi.org/10.18632/oncotarget.18316) PubMed PMID: 28938692; PMCID: PMC5601788
- 147. Patel SP, Kurzrock R (2015) PD-L1 expression as a predictive biomarker in cancer immunotherapy. Mol Cancer Ther 14(4):847–856. [https://doi.](https://doi.org/10.1158/1535-7163.MCT-14-0983) [org/10.1158/1535-7163.MCT-14-0983](https://doi.org/10.1158/1535-7163.MCT-14-0983). PubMed PMID: 25695955
- 148. Green AR, Aleskandarany MA, Ali R, Hodgson EG, Atabani S, De Souza K, Rakha EA, Ellis IO, Madhusudan S (2017) Clinical impact of tumor DNA repair expression and T-cell infiltration in breast cancers. Cancer Immunol Res 5(4):292–299. <https://doi.org/10.1158/2326-6066.CIR-16-0195>. PubMed PMID: 28254786
- 149. McGranahan N, Rosenthal R, Hiley CT, Rowan AJ, Watkins TBK, Wilson GA, Birkbak NJ, Veeriah S, Van Loo P, Herrero J, Swanton C, Consortium TR (2017) Allele-specific HLA loss and immune escape in lung cancer evolution. Cell 171(6):1259–71 e11. <https://doi.org/10.1016/j.cell.2017.10.001>. PubMed PMID: 29107330; PMCID: PMC5720478
- 150. Marty R, Kaabinejadian S, Rossell D, Slifker MJ, van de Haar J, Engin HB, de Prisco N, Ideker T, Hildebrand WH, Font-Burgada J, Carter H (2017) MHC-I genotype restricts the oncogenic mutational landscape. Cell 171(6):1272–83 e15. [https://doi.](https://doi.org/10.1016/j.cell.2017.09.050)

[org/10.1016/j.cell.2017.09.050.](https://doi.org/10.1016/j.cell.2017.09.050) PubMed PMID: 29107334; PMCID: PMC5711564

- 151. Hamilton DH, Griner LM, Keller JM, Hu X, Southall N, Marugan J, David JM, Ferrer M, Palena C (2016) Targeting Estrogen receptor signaling with fulvestrant enhances immune and chemotherapy-mediated cytotoxicity of human lung cancer. Clin Cancer Res 22(24):6204–6216. [https://doi.org/10.1158/1078-](https://doi.org/10.1158/1078-0432.CCR-15-3059) [0432.CCR-15-3059](https://doi.org/10.1158/1078-0432.CCR-15-3059). PubMed PMID: 27267852; PMCID: 5143224
- 152. Welte T, Zhang XH, Rosen JM (2017) Repurposing antiestrogens for tumor immunotherapy. Cancer Discover 7(1):17–19. [https://doi.org/10.1158/2159-](https://doi.org/10.1158/2159-8290.CD-16-1308) [8290.CD-16-1308](https://doi.org/10.1158/2159-8290.CD-16-1308). PubMed PMID: 28062672; PMCID: PMC5224927
- 153. Smolle MA, Calin HN, Pichler M, Calin GA (2017) Noncoding RNAs and immune checkpointsclinical implications as cancer therapeutics. FEBS J 284(13):1952–66. Epub 2017/01/31. [https://doi.](https://doi.org/10.1111/febs.14030) [org/10.1111/febs.14030](https://doi.org/10.1111/febs.14030). PubMed PMID: 28132417
- 154. Dai R, Ahmed SA (2014) Sexual dimorphism of miRNA expression: a new perspective in understanding the sex bias of autoimmune diseases. Ther Clin Risk Manag 10:151–63. Epub 2014/03/14. <https://doi.org/10.2147/TCRM.S33517>. PubMed PMID: 24623979; PMCID: PMC3949753
- 155. Polanczyk MJ, Hopke C, Vandenbark AA, Offner H (2006) Estrogen-mediated immunomodulation involves reduced activation of effector T cells, potentiation of Treg cells, and enhanced expression of the PD-1 costimulatory pathway. J Neurosci Res 84(2):370–8. Epub 2006/05/06. [https://doi.](https://doi.org/10.1002/jnr.20881) [org/10.1002/jnr.20881.](https://doi.org/10.1002/jnr.20881) PubMed PMID: 16676326
- 156. Conforti F, Pala L, Bagnardi V, De Pas T, Martinetti M, Viale G, Gelber RD, Goldhirsch A (2018) Cancer immunotherapy efficacy and patients' sex: a systematic review and meta-analysis. Lancet Oncol 19(6):737–46. Epub 2018/05/21. [https://doi.](https://doi.org/10.1016/S1470-2045(18)30261-4) [org/10.1016/S1470-2045\(18\)30261-4.](https://doi.org/10.1016/S1470-2045(18)30261-4) PubMed PMID: 29778737
- 157. Herbst RS, Baas P, Kim DW, Felip E, Perez-Gracia JL, Han JY, Molina J, Kim JH, Arvis CD, Ahn MJ, Majem M, Fidler MJ, de Castro G Jr, Garrido M, Lubiniecki GM, Shentu Y, Im E, Dolled-Filhart M, Garon EB (2016) Pembrolizumab versus docetaxel for previously treated, PD-L1-positive, advanced non-small-cell lung cancer (KEYNOTE-010): a randomised controlled trial. Lancet 387(10027):1540– 50. Epub 2015/12/30. [https://doi.org/10.1016/](https://doi.org/10.1016/S0140-6736(15)01281-7) [S0140-6736\(15\)01281-7](https://doi.org/10.1016/S0140-6736(15)01281-7). PubMed PMID: 26712084
- 158. Vokes EE, Ready N, Felip E, Horn L, Burgio MA, Antonia SJ, Aren Frontera O, Gettinger S, Holgado E, Spigel D, Waterhouse D, Domine M, Garassino M, Chow LQM, Blumenschein G Jr, Barlesi F, Coudert B, Gainor J, Arrieta O, Brahmer J, Butts C, Steins M, Geese WJ, Li A, Healey D, Crino L (2018) Nivolumab versus docetaxel in previously treated advanced non-small-cell lung cancer (CheckMate 017 and CheckMate 057): 3-year update and out-

comes in patients with liver metastases. Ann Oncol 29(4):959–65. Epub 2018/02/07. [https://](https://doi.org/10.1093/annonc/mdy041) doi.org/10.1093/annonc/mdy041. PubMed PMID: 29408986

- 159. Carbone DP, Reck M, Paz-Ares L, Creelan B, Horn L, Steins M, Felip E, van den Heuvel MM, Ciuleanu TE, Badin F, Ready N, Hiltermann TJN, Nair S, Juergens R, Peters S, Minenza E, Wrangle JM, Rodriguez-Abreu D, Borghaei H, Blumenschein GR Jr, Villaruz LC, Havel L, Krejci J, Corral Jaime J, Chang H, Geese WJ, Bhagavatheeswaran P, Chen AC, Socinski MA (2017) CheckMate I. First-line Nivolumab in stage IV or recurrent non-small-cell lung cancer. N Engl J Med 376(25):2415–26. Epub 2017/06/22. [https://](https://doi.org/10.1056/NEJMoa1613493) doi.org/10.1056/NEJMoa1613493. PubMed PMID: 28636851
- 160. Wallis CJD, Butaney M, Satkunasivam R, Freedland SJ, Patel SP, Hamid O, Pal SK, Klaassen Z (2019) Association of patient sex with efficacy of immune checkpoint inhibitors and overall survival in advanced cancers: a systematic review and meta-analysis. JAMA Oncol . Epub 2019/01/04. PubMed PMID: 30605213. [https://doi.org/10.1001/](https://doi.org/10.1001/jamaoncol.2018.5904) [jamaoncol.2018.5904](https://doi.org/10.1001/jamaoncol.2018.5904)
- 161. Pan ZK, Ye F, Wu X, An HX, Wu JX (2015) Clinicopathological and prognostic significance of programmed cell death ligand1 (PD-L1) expression in patients with non-small cell lung cancer: a meta-analysis. J Thorac Dis 7(3):462–70. Epub 2015/04/30. [https://doi.org/10.3978/j.issn.2072-](https://doi.org/10.3978/j.issn.2072-1439.2015.02.13) [1439.2015.02.13](https://doi.org/10.3978/j.issn.2072-1439.2015.02.13). PubMed PMID: 25922726; PMCID: PMC4387432
- 162. Wu S, Shi X, Sun J, Liu Y, Luo Y, Liang Z, Wang J, Zeng X (2017) The significance of programmed cell death ligand 1 expression in resected lung adenocarcinoma. Oncotarget 8(10):16421–9. Epub 2017/02/02. [https://doi.org/10.18632/oncotar](https://doi.org/10.18632/oncotarget.14851)[get.14851.](https://doi.org/10.18632/oncotarget.14851) PubMed PMID: 28145884; PMCID: PMC5369973
- 163. Grassadonia A, Sperduti I, Vici P, Iezzi L, Brocco D, Gamucci T, Pizzuti L, Maugeri-Saccà M, Marchetti P, Cognetti G (2018) Effect of gender on the outcome of patients receiving immune checkpoint inhibitors for advanced cancer: a systematic review and meta-analysis of phase III randomized clinical trials. J Clin Med 7(12):542
- 164. Goodman AM, Kato S, Bazhenova L, Patel SP, Frampton GM, Miller V, Stephens PJ, Daniels GA, Kurzrock R (2017) Tumor mutational burden as an independent predictor of response to immunotherapy in diverse cancers. Mol Cancer Ther 16(11):2598–608. Epub 2017/08/25. [https://doi.](https://doi.org/10.1158/1535-7163.MCT-17-0386) [org/10.1158/1535-7163.MCT-17-0386](https://doi.org/10.1158/1535-7163.MCT-17-0386). PubMed PMID: 28835386; PMCID: PMC5670009
- 165. Wang S, Zhang J, He Z, Wu K, Liu XS (2019) The predictive power of tumor mutational burden in lung cancer immunotherapy response is influenced by patients' sex. Int J Cancer 145:2840
- 166. Unger JM, Moseley A, Ramsey SD, Osarogiagbon RU, Symington B, Hershman DL (2019) Socioeconomic deprivation and cancer outcomes in patients treated in clinical trials. Proc Am Soc Clin Oncol 37:162
- 167. Duma N, Abdel-Ghani A, Yadav S, Hoversten KP, Reed CT, Sitek AN, Enninga EAL, Paludo J, Aguilera JV, Leventakos K (2019) Sex differences in tolerabil-

ity to anti-programmed cell death protein 1 therapy in patients with metastatic melanoma and non-small cell lung cancer: are we all equal? Oncologist. <https://doi.org/10.1634/theoncologist.2019-0094>

168. Ozdemir BC, Dotto GP (2019) Sex hormones and anticancer immunity. Clin Cancer Res . Epub 2019/03/21. PubMed PMID: 30890551. [https://doi.](https://doi.org/10.1158/1078-0432.ccr-19-0137) [org/10.1158/1078-0432.ccr-19-0137](https://doi.org/10.1158/1078-0432.ccr-19-0137)