# **Chapter 9 Role of Recombinant DNA Technology in Biofertilizer Production**



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**Abstract** Nutrients present in plants are important for the maintenance of crops and production of healthy food for the constantly rising population of the world. For this effcient means of maintaining soil fertility are required. Soil maintenance approaches nowadays are largely reliant on chemical fertilizers, which may pose a severe threat to human well-being and the environment. In this backdrop, biofertilizers have been recognized as a substitute for elevating soil richness and hence crop productivity in sustainable agricultural practices. The utilization of advantageous microbial organisms as biofertilizers is extremely signifcant in agriculture as it aids in maintenance of food security and elevating crop produce. Furthermore, biofertilizers are highly signifcant in maintaining the quality of soil. Microbes which are frequently utilized as biofertilizers include potassium solubilizers, nitrogen fxers, mycorrhiza, cyanobacteria or blue green algae, plant growth-promoting *Rhizobacteria*. Biofertilizers aid in nutrient uptake by plants, offer forbearance to biotic and abiotic situations to plants and also maintain plant growth. Biofertilizers maintain nutrient richness by means of nitrogen fxation, solubilization of potassium, production of antibiotics, disintegration of organic substances and release of plant growth-promoting agents. Biofertilizers, when given in the form of seed or soil inoculants, contribute in nutrient cycling and lead to enhanced crop production. Biofertilizers, therefore, play a vital role in maintaining soil nutrients and hence agricultural produce. Furthermore, biofertilizer production by using the tools of molecular biotechnology like recombinant DNA technology can improve the metabolic pathways of production of important plant growth-promoting factors like phytohormones,, if recognized and transmitted to the useful plant growth-promoting microbes. Recombinant DNA technology offers numerous benefts, as explicit biological pathways can be controlled with high accuracy and entirely novel functions can be engineered into the microorganisms for producing effcient biofertilizers.

**Keywords** Biofertilizers · Solubilizers · Mycorrhiza · Phytohormones · Cyanobacteria

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### **9.1 Introduction to Biotechnology**

The term "biotechnology" was coined by Karl Ereky and is blend of the two words, i.e. biology and technology. This feld is tremendously divergent, enormous and multidisciplinary. Thus, a clear-cut description of the subject is slightly hard. Biotechnology is fundamentally the utilization of biological substances like microbes or cellular constituents in controlled fashion for the advantage of mankind (Okeno et al. [2012\)](#page-19-0). In other words, biotechnology is an integrated utilization of biochemistry, microbiology and engineering knowledge for the utilization of microbes, cultured cells or tissues to their best. Human beings have sustained their search for enhancing the natural potential of microbes and making them competent for novel methods. In past, people exploited microbes for the production of cheese, bread production or brewing alcohol, even if the process of fermentation was not tacit comprehensively (Carpenter et al. [2002\)](#page-17-0). At present, application of biotechnology is highly complicated. Now, scientists can manipulate living organisms and transfer genetic matter among them, producing transgenic organisms. The present relevance of biotechnology is largely in the area of biomedicine and agriculture. Current methods allow the construction of novel and enhanced food products. In biomedicine area, novel vaccines, antibiotics, etc. have been produced against various diseases like AIDS, cancer and many hereditary diseases. Biotechnology is also used in the area of bio-fuel production, mining and pollution control. Genetically modifed microbes and plants are utilized to remove toxic chemicals from oil spoil spills or industrial effluents (Chen et al. [2007](#page-17-1)). Besides, improved superiority of life and still there exists a countless exhilarating opportunities in the feld of biotechnology (Figs. [9.1](#page-2-0) and [9.2](#page-2-1)).

### *9.1.1 Subfelds of Biotechnology*

Generally, biotechnology is categorized in three major subtypes:

- Green biotechnology
- White biotechnology
- Red biotechnology

### *9.1.2 Green Biotechnology*

Green biotechnology is a vital feld of contemporary biotechnology. Its foundation is on the crop enhancement and manufacture of new crop products (McAllister et al. [2012\)](#page-19-1). This is achieved by introducing foreign genes into the plants having huge economic importance. It comprises of three major areas which include:

<span id="page-2-0"></span>

<span id="page-2-1"></span>**Fig. 9.1** Applications of biotechnology



Fig. 9.2 Subfields of biotechnology

- (a) Plant tissue culture
- (b) Plant genetic engineering
- (c) Plant molecular marker assisted breeding

Plant tissue culture involves the production of the whole plant or part of it under laboratory conditions. Its main advantage is the quick manufacture of plant materials like citrus fruits, banana, etc. On the other hand, plant genetic engineering involves the introduction of benefcial genes from one living organism to other. This generates improved varieties of crops with enhanced production (Brookes and Barfoot [2009\)](#page-17-2).

In the case of plant molecular marker-assisted breeding, molecular markers (specifc short sequences of DNA) are accountable for a preferred attribute. Thus, improved properties, like disease resistance, can be achieved (Horvath et al. [2012\)](#page-18-0).

### *9.1.3 White Biotechnology*

This area is concerned with industries. It utilizes enzymes, bacteria, yeast or moulds to produce valuable products. It results in the manufacture of wide range of bioproducts like vitamins, antibiotics, detergents, etc. (Bueno et al. [2016\)](#page-17-3).

### *9.1.4 Red Biotechnology*

It is concerned with medical biotechnology. It involves genetic manipulation of organisms to create antibiotics. Herein, the human body's own tools are utilized to eliminate the pathogens. It is of immense signifcance in the conventional drug discovery and also aids in improving the potential for cure, anticipation and analysis of diseases (Becker et al. [2008](#page-17-4)).

### **9.2 Recombinant DNA Technology**

The growth and understanding of biological phenomena over the past few decades, both at molecular and cellular levels, is transfgured by the dawn of genetic engineering or recombinant DNA technology. This branch of biology is largely spawned under contemporary biotechnology that uses living organisms to generate enhanced and precious products for the betterment of human society. Chemical and biochemical engineering techniques are concerned with the production of recombinant DNA. Cultivation of microbes and their downstream processes rely on engineering techniques.

The history of recombinant DNA technology dates back to 1953, when the double helical structure of DNA was explicated by Watson and crick and the genetic code was cracked by Nirenberg. Afterwards, in 1973, the method of restriction digestion was invented by Cohen and Boyer which involve cut and paste of the DNA sequences (Ames and Martin [1964;](#page-17-5) Cohen et al. [1973](#page-18-1)).

Due to recombinant DNA technology, cloning of genes for production of polypeptides (growth factors, interferon, blood clotting factors, human insulin, viral coat proteins, etc.) has become achievable. Each of the polypeptide is unique in the context of its sequence or target. Now, with the advent of recombinant DNA technology, researchers can express a natural gene even in a very simple bacterium like *E. coli* (Brown et al. [2015](#page-17-6)).

Somatostatin was the frst human protein produced in *E.coli* in 1977. Later in 1982, the frst recombinant protein, i.e. human insulin, was available in the market. Kary Mullis in 1985 envisaged the idea of polymerase chain reaction (PCR) which revolutionized the feld of recombinant biotechnology. Bimolecular archaeology, DNA fngerprinting, molecular ecology, forensics, etc. are novel branches that have become achievable due to PCR (Kakumanu et al. [2012;](#page-18-2) Huang et al. [2001\)](#page-18-3).

### **9.3 Construction of the Recombinant DNA Molecule**

To construct a recombinant DNA molecule, a DNA fragment (restriction fragment) is inserted in cloning vector at the corresponding restriction site. Then sticky ends of the vector and the foreign DNA are allowed to anneal. Then, by means of DNA ligase, they are joined covalently to create a chimeric DNA (Fig. [9.3\)](#page-5-0).

### **9.4 Cloning of DNA by Recombinant DNA Technology**

Comprehensive study of the working and construction of the gene at molecular stage needs a huge amount of individual gene in purest form. Recombinant DNA technology offers great advantage in cloning that permits researchers to create a huge quantity of matching DNA molecules. The DNA molecule so produced has sequences derived from diverse sources. In DNA cloning one of the important steps is to link the desired DNA fragment to a vector DNA that could duplicate within the host cell. As a result, the recombinant DNA molecule is produced that replicates together with the vector, producing a huge quantity of matching DNA molecules (Bonneau and Laarved [1999\)](#page-17-7). The scheme of production of recombinant DNA is shown in the following diagram:

<span id="page-5-0"></span>

**Fig. 9.3** Construction of recombinant DNA molecule. (Adapted from Biochemistry, 4th edition, Donald Voet and Judith G. Voet, 2011)



# **9.5 Role of Restriction Enzymes in the Creation of Recombinant DNA Molecule**

The main purpose of DNA cloning is to generate distinct, tiny regions of DNA molecule that comprises of defnite genes. Practically very little amount of DNA molecules could be cloned in a vector. Thus, extremely lengthy DNA molecules must be cleaved into fragments that could be put easily into a vector DNA. To facilitate this process, restriction enzymes are used (Becker et al. [2008](#page-17-4)). Restriction enzymes are endonucleases that are obtained from bacteria and are characteristically able to distinguish and cleave specifc four- to eight-base pair sequences, called "restriction site"*.* These sites commonly are short palindromes (sequence same on each DNA strand when read in the  $5' \rightarrow 3'$  direction). One of the widely used restriction enzymes is from *E. coli* called *EcoR1* (Brown et al. [2015](#page-17-6)). This enzyme is able to make staggered cuts at the defnite six-base pair (palindromic sequence) as shown in Fig. [9.4.](#page-6-0)

For every restriction endonuclease, bacteria also create a modifcation enzyme that protects the bacterium's own DNA from cleavage. This modifcation enzyme adds a methyl group to one or more bases, generally within the restriction site. This methyl group prevents endonuclease from cleaving the DNA. Methylating enzyme and the restriction endonuclease together form the "restriction modifcation system"

<span id="page-6-0"></span>

**Fig. 9.4** Cleavage of DNA by *Eco*R1. (Adapted from Molecular Cell Biology, 4th edition, Lodish et al. 2002)

that protects the host DNA while it destroys incoming foreign DNA by cleaving it at all the restriction sites (Overton [2014\)](#page-19-2).

### **9.6 Inserting DNA Fragments into Vectors**

DNA ligases aid in the insertion of DNA fragments into the vector DNA. During normal DNA replication, these ligase take part in joining of short segments of DNA called *Okazaki* fragments. In case of DNA cloning purifed DNA ligases covalently join the ends of restriction fragments and vector DNA that possesses complementary ends. This linkage is done through standard  $3' \rightarrow 5'$  phosphodiester bonds of DNA. The DNA ligase from the bacteriophage T4 can ligate both the blunt ends and the complementary sticky ends. Nevertheless, blunt-end ligation is intrinsically ineffective and needs huge amount of both DNA and DNA ligase (He et al. [2000](#page-18-4)).

### **9.7 Role of Recombinant DNA Technology in Agriculture**

With the rise in demand for food, there is a huge demand to integrate biotechnology to enhance crop improvement strategies. The advent of biotechnology has revolutionized the whole crop improvement strategies by offering novel strains of crops, highly profcient, precise and selective pesticides and valuable fertilizers (Ewen and Pusztai [1999\)](#page-18-5). Ancient communities maintained the crops by implementing selective breeding programs. By gathering seeds from the most advantageous crops, they

were able to generate crops that were adapted to the changing environment and could offer higher yield. In agriculture, biotechnology has resulted in enhancement of crop productivity by controlling diseases through improved genetic resistance programs. Thus, it offers effcient tools for enhancing and sustaining food security. It offers an outstanding scenario for improving health by engineering the nutritional quality of food, bioremediation and genetic conservation (Buikema and Haselkorn [2001](#page-17-8)).

The development of protein engineering and synthetic biology has revolutionized the exploitation of proficient microbial systems for the generation of value added products. Environmentally friendly routes of manufacture, creation of smaller amounts of by-products and enhanced output of the target compounds are advantages of microbial biosynthesis over the chemical synthesis.

Extensive optimization of growth conditions is required while obtaining compounds from their native host. However, with the help of recombinant DNA technology, several natural products, their derivatives, or even unnatural compounds have been created within established microbial hosts. Alkaloids, terpenoids, favonoids, amino acids, peptides, antibiotics, organic acids and vitamins are some examples of microbially produced compounds through modern recombinant biotechnology (Chaurasia et al. [2008\)](#page-17-9).

### *9.7.1 Soil Fertility*

Soil is that portion of the earth on which plant grows. It comprises of three main layers (top soil, sub soil and parent soil). Uppermost layer, i.e. top soil, contains minerals, water, air and inorganic or organic matter. Minerals include silica, aluminium oxide, calcium, potassium, magnesium and extremely little quantity of nitrogen, sulphur, boron, zinc, molybdenum, etc. (Huang et al. [2004\)](#page-18-6). Among all the minerals, only 14 are essential minerals. Essential nutrients are further categorized into micronutrients and macronutrients. The macronutrients are further classifed into primary macronutrient and secondary macronutrients. Primary macronutrients are frequently limited in the soil, while secondary macronutrients are rarely limited (Dash et al. [2016](#page-18-7)). Soil quality decides the quality and quantity of agricultural production. Besides, it also offers niche to a wide range of living organisms. For this reason, proper management of soil is one of the key factors for enhanced crop productivity (Sanahuja et al. [2011](#page-19-3)).

### **9.8 Biofertilizers**

Chemical fertilizers offer convenient technique to supplement soil with valuable nutrients and therefore help to overcome the growing requirements of food. Nevertheless, they are reasonably very costly and risky to human well-being. On the

other hand, they not only supply necessary nutrients to crops but also supply them in an easy accessible mode. Thus, chemical fertilizers can rapidly improve the development and yield of crops and are hence gaining fame around the world (Raja [2013\)](#page-19-4). However, extensive employment of such fertilizers causes grave ecological problems. Nitrate leaching and contamination of groundwater are due to augmented exploitation of fertilizers. Inorganic fertilizers like calcium nitrate, ammonium chloride and sodium nitrate produce greenhouse gases that result in pollution. Elevated levels of greenhouse gases and heavy metal uptake by plants are major causes of environmental damage. Eutrophication of freshwater is also due to chemical fertilizers. Furthermore, chemical fertilizers can eradicate the advantageous microbial or insect community of the soil. Fortunately, nature has bestowed the soil with a variety of microbes with specifc mechanisms to overcome this challenge. This mechanism besides maintaining soil quality also works in tandem with plants as an element of ecosystem. Such mechanism is what constitutes "biofertilizers" (Khosro and Yousef [2012](#page-18-8)). Biofertilizers constitute a central part of green agriculture. Biofertilizers contain proficient strains of microbes, organic products and departed and rotten parts of plants which supply nutrients to soil. It progressively elevates crop yield by means of enhancing soil fertility. They change the unavailable form of nutrients to the accessible form by escalating the population of microbes in the rhizosphere (Leonardo et al. [2006\)](#page-18-9). Microbes are accountable for delivering soluble nutrients to crops (Chang and Yang [2009](#page-17-10)). These are helpful in a variety of ways that include solubilization of plant nutrients and fxing of atmospheric nitrogen. They also encourage the formation of growth-promoting phytohormones like cytokinins and auxins. They also defend the plant against various abiotic and biotic stresses (Mitragotri et al. [2014\)](#page-19-5).

Biofertilizers aid plants in accessing the nutrient present in its surroundings. The microbes frequently employed as the biofertilizers include *Rhizobium*, *Azotobacter*, *Anabaena* (nitrogen fxers), *Pseudomonas putida*, mycorrhizal fungi, etc. (Liu and Golden [2002](#page-18-10)). Likewise, phytohormone-/auxin-producing bacteria could also be utilized as biofertilizer (Somasegaran and Springer [1994](#page-19-6)). All of these microbes enhance growth and development of plants (Table [9.1](#page-8-0)). The grievance from agriculturalists regarding the effectiveness of biofertilizer is their improper storage and the larger time period between feld application and production. This restricts their employment due to compatibility and constancy issues under diverse soil environments. For this reason, improved shelf life is the basis for the popularization of biofertilizers (Adesemoye and Kloepper [2009](#page-17-11)).

Microbe	Plant growth promoting substance
Azotobacter	Vitamins, gibberellins
Azospirillum	Indole acetic acid, gibberellins, indole lactic acid
Cyanobacteria	<b>Vitamins</b>
Phosphate-solubilizing bacteria	Vitamins (thiamin, biotin, riboflavin)
Mycorrhizae	Cytokinin, gibberellins

<span id="page-8-0"></span>**Table 9.1** Plant growth-promoting substances associated with various microbes

Presently, a variety of marketable biofertilizers are obtainable and a variety of mechanisms have been formulated to guarantee maximum viability of the microbes used in such formulations (Bhattacharyya and Jha [2012\)](#page-17-12). These strategies include:

- Optimization of biofertilizer formulation
- Usage of thermo-resistant or drought-resistant and genetically modifed strains
- Employment of liquid biofertilizer

For dexterity, a carrier substance is utilized as a vehicle for the microbes which are to be used as biofertilizer. Carrier substances include clay, vermiculite, peat, seed, lignite powder, rice bran, charcoal, etc. For enhanced shelf life of biofertilizer formulation, a combination of these carriers is employed. Likewise, pre-sterilization of carriers is done to enhance the shelf life of microbes (Wani et al. [2013](#page-20-0); Liddycoat et al. [2009](#page-18-11)). Liquid biofertilizer formulation is an important aspect to improve shelf life. These formulations enclose an adequate amount of cell protectants and nutrients that are responsible for the extended shelf life of biofertilizers. Besides, these formulations can endure huge temperature range (Santos et al. [2012;](#page-19-7) Ruiz-Sanchez et al. [2010\)](#page-19-8).

Biofertilizers got commercialized with the launch of "nitrogen" by Hiltner and Nobbe. This preparation was for legumes. Later microbial inoculants for legumes were made like "Alnit". It proved advantageous for the development of nonleguminous plants. These bacteria were recognized to be local ammonifers. Discovery of *Azotobacter* and *Clostridium* developed a new feld for investigating economical bacterial fertilizers. The rhizosphere of these plants contains a range of species of soil bacteria that enhance plant growth by numerous ways. Such bacteria are jointly known as plant growth-promoting *Rhizobacteria* (PGPR). One of the ways is through fxing of atmospheric nitrogen which enhances the accessibility of exploitable form of nitrogen in the rhizosphere. They also promote symbiosis between plants and microorganisms (Mflinge and Mtei [2014](#page-19-9)).

There are diverse modes of interactions between biofertilizers and plants, taking into account the extent of association between microbes with plant roots which are mentioned as follows (Youssef and Eissa [2014\)](#page-20-1):

- Microbes living in the soil near the root, utilizing nitrogen and carbon metabolites leaking from the root (rhizosphere)
- Microbes colonizing the root surface (rhizoplane)
- Microbes colonizing the root tissue inhabiting intercellular spaces (endophytes)
- Microbes living inside cells in specialized root structures or nodules (symbionts)

### *9.8.1 Types of Biofertilizers*

Biofertilizers are categorized into various types on the basis of microorganisms they contain (Chun-Li et al. [2014](#page-17-13)). The different types of biofertilizers are discussed below:

#### **9.8.1.1 Symbiotic Biofertilizers**

Symbiotic microbes infect root tissues and form new structures. In many cases, the application of molecular biology tools allows the discovery of the genes and signals involved in the benefcial interaction between the microorganism and the plant. The main symbiosis relating to agricultural application as biofertilizers is considered below.

#### **9.8.1.2 Rhizobia**

Rhizobium is an illustration of a symbiotic association colonizing legume roots and fxes the atmospheric nitrogen. It has a capability to fx atmospheric nitrogen in leguminous and non-leguminous plants. The different genus and species inhabiting legume root nodules are usually referred to as *Rhizobia*. These involve *Alphaproteobacterias*, e.g. *Rhizobium*, *Bradyrhizobium*, *Sinorhizobium*, *Mesorhizobium*, *Azorhizobium*, *Allorhizobium* and *Agrobacterium*, and *Betaproteobacteria*, e.g. *Burkholderia*. The best model describing the interaction between rhizobia and legume roots includes favonoid/isofavonoid molecules released by the plants which induce bacterial genes and consequently the synthesis of the lipochitin oligosaccharide (LCO) molecules, which in turn control infection and nodule growth in the root tissue (Rajaram and Apte [2008\)](#page-19-10). Usually, it pierces the root hair and multiplies there in special root structures called root nodules. The quantity of nitrogen fxed depends on host, strain of *Rhizobium* and existing environmental conditions. They are very profcient biofertilizers for legumes as far as the magnitude of nitrogen fxation is concerned. The nod, nif and fx genes control the nodulation and nitrogen fxation by the bacterium (Lavakush et al. [2014\)](#page-18-12).

#### **9.8.1.3 Blue Green Algae**

Blue green algae (BGA) are the most ancient organisms possibly the frst among those that started evolving oxygen. These appear in numerous shapes (single celled, branched or unbranched with flaments). The majority of them possess special structure called heterocyst whose role is to fx nitrogen. The algae that are frequently applied in felds belong to *Anabaena*, *Nostoc*, *Scytonema*, *Tolypothrix*, etc. (Joseph and Meeks [1987](#page-18-13)). These are widely used in rice felds (Zhou et al. [1998\)](#page-20-2). BGA secrete numerous growth-promoting substances like amino acids, vitamins, polysaccharides, sugars, etc. which boost the yield of crops (Schiefer et al. [2002;](#page-19-11) Hussain et al. [2002\)](#page-18-14).

#### **9.8.1.4 Mycorrhiza**

Mycorrhiza is the best example of the symbiotic association between fungi and plant roots (higher plants). The fungi enhance the growth of plants and protect them from various stresses. These fungi colonize the root cortex and mycelia of the plants and help them to obtain nutrients from soil. These fungi are cosmopolitan in soil and are seen in the roots of thallophytes, gymnosperms, pteridophytes and angiosperms (Stewart [1980\)](#page-19-12). Plants, on the other hand, protect fungi from root pathogens and also provide them with carbohydrates, hormones, nutrients, etc. The mycorrhizal plants have better forbearance to poisonous metals, salinity, elevated soil temperatures and unfavourable pH. Such plants also resist transplantation shocks. They play a signifcant task by enhancing growth and nutrient uptake in plants (Vessey [2003\)](#page-19-13).

#### **9.8.1.5 Free-Living or Non-symbiotic Biofertilizers**

Since the description of PGPR by Kloepper and Schroth ([1978\)](#page-18-15), many different bacteria genera have been described as PGPR: *Pseudomonas*, *Azospirillum*, *Azotobacter*, *Gluconacetobacter*, Herbaspirillum, *Bacillus*, *Burkholderia*, *Erwinia*, *Caulobacter*, *Azotobacter*, *Chromobacterium*, *Serratia*, *Micrococcus*, *Flavobacterium*, *Actinobacteria*, *Enterobacter*, *Arthrobacter*, *Agrobacterium* and *Hyphomicrobium* and fungus such as *Trichoderma*, among others (Gupta [2004\)](#page-18-16).

Many PGPR have been described as endophytic bacteria. It is not clear if the plant growth promotion effects are a consequence of plant-microbe interaction in the external part of the rhizosphere or if an endophytic state is necessary (Hayat et al. [2010\)](#page-18-17). Many different mechanisms have been claimed to be responsible for the plant growth promotion effect after in vitro experiments under controlled conditions. In some cases, the use of appropriate mutants helps in the defnition of these mechanisms. But since different mechanisms are always present in a single strain, it is almost impossible to know which are the main mechanisms operating and driving the plant growth promotion. Irrespective of the real mechanisms operating in PGPR with a positive effect in the feld, the use of these micro-organisms has dramatically increased in recent years and will probably continue to grow because biofertilizers appear as a valuable opportunity for future sustainable agriculture (Gonzalez et al. [2015\)](#page-18-18). Many commercial products already exist which are based on *Pseudomonas* or *Azospirillum* strains in the market (Yang et al. [2009;](#page-20-3) Scalenghe et al. [2012\)](#page-19-14). The different mechanisms operating in PGPR can be classified as  $N_2$  (nitrogen) and P (phosphorus) nutrition effects and plant root development and ftness mediated by phytohormones **(**Fig. [9.5](#page-12-0)**).**

<span id="page-12-0"></span>

**Fig. 9.5** Mode of action of PGPR

## **9.9 Phytohormone-Mediated Mechanism of Plant Growth-Promoting Microorganisms/Bacteria (PGPB)**

One of the most visible effects on plants after inoculation with PGPB is the huge development – and sometimes changes in the architecture – of the root of the plant. This general improvement of root growth, including root-hair development, is one of the characteristic phenotypes of the interaction plant-PGPB.

It is likely that water and mineral uptake is consequently improved because of the increase in the root system, although the specifc mechanism is not completely clear. Changes in hormone balance, enhancement of proton-effux activity and modifcation in a wide range of related enzymatic activities would be part of the mechanisms behind this phenotype (Backman and Sikora [2008;](#page-17-14) Joo et al. [2005](#page-18-19)).

### *9.9.1 Auxins*

The general root improvement phenotype can be reproduced by replacing phytohormones with PGPB. Auxin-related substances, such as indole acetic acid (IAA), appear to be involved in one of the most important mechanisms regarding the general root development improvement. Nevertheless, bacterial production of IAA in plants has not yet been demonstrated. There are no IAA completely deficient mutants, but IAA attenuated mutants were ineffective as PGPB, compared to parental strains (Ahmed and Hasnain [2010](#page-17-15); Chen [2006](#page-17-16)).

### *9.9.2 Cytokinins*

The role of cytokinins in the promotion of root development is not clear, but cytokinin-producing PGPB stimulate nodulation in legumes when co-inoculated with *Rhizobia*. Besides, it has been demonstrated recently that there is a Nod factorindependent mechanism for infection and nodulation, probably mediated by rhizobial cytokinin. This particular area deserves more attention in the future (Riefer et al. [2006;](#page-19-15) Sokolova et al. [2011\)](#page-19-16).

### *9.9.3 Ethylene*

Ethylene is related to general plant responses when a stress condition appears, even if it is a very low stress situation. When this happens, the plant synthesizes ethylene and stops its growth temporarily. This is because of the regulatory effects of ethylene on different cell functions. 1-aminocyclopropane-1-carboxylate is a precursor of ethylene synthesis. The enzyme ACC deaminase is present in some bacteria which can even use ACC as C (carbon) and N sources. When ACC deaminase is expressed by rhizospheric bacteria, root growth and development are enhanced. It is probably because of the elimination of the inhibitory concentrations of ethylene produced by the plant. This enzyme is not present in every bacteria, and its activity is codifed by a single gene acdS. The introduction of this gene from *Pseudomonas putida* into other bacteria species confers plant growth-promoting functions to the recipient bacteria that are absent in the parental strain. This represents a potential biotechnology-based tool to improve microorganisms to be used as biofertilizers (Reid [1981\)](#page-19-17).

### *9.9.4 Nitric Oxide*

Nitric oxide (NO), a plant regulator volatile phytohormone, is also produced by some PGPB. Bacterial nitric oxide is a mediator in IAA-induced root development. NO can also mediate plant growth-promoting action in *Azospirillum brasilense* Sp245 inducing morphological alterations in tomato roots irrespective of the full bacterial capability for IAA biosynthesis (Butterbach-Bahl et al. [2013\)](#page-17-17).

### *9.9.5 Helper Bacteria*

In the studies of plant microbe interaction which induced some kind of plant growth promotion, there are other cases that do not ft into the previous defnitions but which can be considered as another kind of biofertilizer. That is the case of bacteria which improve a plant-microbe interaction as a third partner in the interaction. An example can be found in rhizospheric actinomycetes isolated from legumes or actinorhizal nitrogen-fxing nodules which are able to stimulate nodulation, consequently nitrogen fxation in the plant and fnally plant growth. This tripartite plant-microbe interaction is not well known in terms of mechanism. However, it clearly shows that biofertilizers can be improved by the use of more than one microorganism at a time (Egamberdiyeva [2007](#page-18-20)).

#### **9.10 Genetically Modifed Microbes as Biofertilizers**

Genetically modifed organisms (GMOs) symbolize a genetic reserve. Such microorganisms may fnd a use as donor or recipient of desirable genes. Microorganisms play an important role in various sectors of agriculture, food processing, environmental management and pharmaceutical industries. Genes of microbes can be optimized or improved by means of various genetic modifcations using recombinant DNA technology (Tabashnik et al. [2011\)](#page-19-18). Usually, this is dependable on the recognition and selection of the mutants with favourable traits. In numerous cases, the usage of molecular biology tools or recombinant DNA technology allows the discovery of the genes and signals concerned in the advantageous interaction (endophytic, mycorrhizal and diazotrophic) between the microbe and plant **(**Fig. [9.6](#page-15-0)**).** These symbiotic interactions can assist plant growth and development through nitrogen uptake, siderophore production, phosphate solubilization, etc. (Ritika and Uptal [2014\)](#page-19-19).

Recombinant biotechnology offers an advantage to decrease the employment of synthetic fertilizers. Biofertilizer technology has signifcantly developed in the market. The nature of multiple mechanisms discovered for PGPR actions and the option of genetically modifying a specifc strain relating to a particular plant growthpromoting activity imply that the use of genetically modifed organisms like biofertilizers will be an area of diverse potential in the coming times (Tabashnik et al. [2011\)](#page-19-18). Further, the knowledge of microbial ecology and its dynamics will surely enhance the biofertilizer technology. Microbes are particularly targeted for genetic improvement since they are given huge importance in modern agriculture as they are used as biofertilizers. Biofertilizers represent an alternative to synthetic fertilizers which are facing lots of disparagement due to their negative impact on the ecology and human well-being. There is an important requirement to build up ecofriendly control using existing microbes. Such microbes would offer protection to plants against pathogens and would be economical, reliable and effective (Pishchik et al. [2002\)](#page-19-20). To obtain this target, better-quality strains are needed. Thus, genetically modifed microbes could be used for this purpose. Efforts are in progress to formulate profcient biofertilizers compatible with a broad choice of plants and soil by means of genetically engineered techniques. For example, biofertilizers have been formulated based on nitrogen-fxing rhizobial bacteria occurring naturally in the nodules of leguminous plants. Nevertheless, these microbes are not competent

<span id="page-15-0"></span>

**Fig. 9.6** The use of recombinant DNA technology in the generation of products assisting in the symbiotic interactions. (Adapted from Vitorino and Bessa [2017](#page-20-4))

enough to supply nitrogen to non-legumes. In such cases, genetic engineering is of special importance, as it assists in the development of effcient delivery systems. In this way non-legumes could be grown together with symbiotic rhizobial root nodules devoid of externally applied nitrogen fertilizers (Aloni et al. [2006;](#page-17-18) Ruiz-Sanchez et al. [2010\)](#page-19-8). The foreign genes used for transforming microbes could be integrated into the host genome or plasmid. To express a heterologous gene in bacteria and fungi, the regulatory area of this gene should be modifed in promoter and terminator regions in order to optimize the function of the inserted gene in the new host. Adding specifc genes which can bestow biocontrol ability could improve the biocontrol ability of microbes lacking such genes (Dash et al. [2016](#page-18-7)). For example, many *Rhizobacteria* with biocontrol activity produce chitinases. However, few *Rhizobacteria* like *Rhizobium meliloti* and *Pseudomonas putida*, both of which are outstanding root colonizers, are defcient in synthesizing chitinase (Bagwan et al.

[2010\)](#page-17-19). Incorporation of chitinase gene into their genome has enabled them to defend the plant against fungi.

Nitrogen-fxing property of *Rhizobium* inoculants could be augmented by means of genetic engineering tools. An additional way is by planting the crops that use nitrogen more profciently. An example of such crops is genetically modifed Canola which exhibits a noteworthy decline in the amount of nitrogen fertilizer that is leached into soil or lost into the atmosphere, and hence it improves the economies of farmers through the enhanced proftability. Moreover, biofertilizers when formulated by means of molecular biotechnology can improve the biological pathways of production of phytohormones like auxin, cytokinin, etc. which assist in plant growth and development (Nautiyal et al. [2008](#page-19-21)). Similarly, many pseudomonads in the rhizosphere manufacture siderophores which can chelate iron ions and thus escalate iron uptake by the plants. The genetically modifed strain (RMBPC-2) of *Sinorhizobium meliloti* has added genes that control nitrogenase enzyme from the plant to the bacterium (Boccia and Sarnacchiaro [2015\)](#page-17-20). Likewise, Trichoderma species are extensively found in the soil and are antagonistic to other fungi. *Trichoderma harzianum* is an efficient rhizosphere colonizer and is able to parasitize pathogenic fungi. Many extracellular enzymes like glucanases, chitinases, lipases and proteases synthesized by *Trichoderma* species have been improved with the transfer of chitinase genes, notably from *Serratia marcescens* (Awais et al. [2010\)](#page-17-21). Thus, such genetically modified strains could act as efficient biofertilizers and will aid in crop improvement.

### **9.11 Conclusion**

Our reliance on chemical fertilizers has encouraged the fourishing of factories or industries that are generating lethal chemicals that are not only dangerous for human utilization but can also perturb the normal environmental equilibrium. Now, attention is diverting towards consuming food grown with organic fertilizers than with chemical fertilizers (Leonardo et al. [2006\)](#page-18-9). Biofertilizers can assist in solving the problem of food crises of the ever-rising worldwide population. It is essential to recognize the positive aspects of biofertilizers so as to apply it to modern agriculture. The employment of biofertilizers containing advantageous microbes improves the crop productivity to a larger extent (Kanchiswamy et al. [2015\)](#page-18-21). Biofertilizers play an important role in maintenance of soil quality. This would in turn protect the environment and would require less expenditure. Besides, biofertilizers when formulated using the tools of molecular biotechnology can improve the biological pathways of production of plant growth-promoting substances, if identifed and transferred to the useful plant growth-promoting microbes (Goswami et al. [2014\)](#page-18-22). Recombinant biotechnology offers numerous advantages in this area, as particular metabolic processes could be tackled with additional accuracy, and entirely novel functions could be introduced in microbes.

### **References**

- <span id="page-17-11"></span>Adesemoye AO, Kloepper JW (2009) Plant-microbes interactions in enhanced fertilizer use effciency. Appl Microbiol Biotechnol 85(1):1–12
- <span id="page-17-15"></span>Ahmed A, Hasnain S (2010) Auxin producing Bacillus sp.: auxin quantifcation and effect on the growth Solanumm tuberosum. Pure Appl Chem 82:313–319
- <span id="page-17-18"></span>Aloni R, Aloni E, Langhans M (2006) Role of cytokinin and auxin in shaping root architecture: regulating vascular differentiation, lateral root initiation, root apical dominance and root gravitropism. Ann Bot 97:883–893
- <span id="page-17-5"></span>Ames BN, Martin RG (1964) Biochemical aspects of genetics: the operon. Annu Rev Biochem 33:235–258
- <span id="page-17-21"></span>Awais MA, Pervez A, Yaqub R, Sarwar FA, Siraj S (2010) Current status of biotechnology in health. Am Eur J Agric Environ Sci 7(2):210–220
- <span id="page-17-14"></span>Backman PA, Sikora RA (2008) An emerging tool for biological control. Biol Control 46(1):1–3
- <span id="page-17-19"></span>Bagwan JD, Patil SJ, Mane AS, Kadam VV, Vichare S (2010) Genetically modifed crops: food of the future. Int J Adv Biotech Res 1(1):21–30
- <span id="page-17-4"></span>Becker PD, Noerder M, Guzmán CA (2008) Genetic immunization: bacteria as DNA vaccine delivery vehicles. Hum Vaccin 4:189–202
- <span id="page-17-12"></span>Bhattacharyya PN, Jha DK (2012) Plant growth-promoting rhizobacteria (PGPR): emergence in agriculture. World J Microbiotechnol 28(4):1327–1350
- <span id="page-17-20"></span>Boccia F, Sarnacchiaro P (2015) Genetically modifed foods and consumer perspective. Recent Pat Food Nutr Agric 7:28–34
- <span id="page-17-7"></span>Bonneau M, Laarved B (1999) Biotechnology in animal nutrition, physiology and health. Livest Prod Sci 59:223–241
- <span id="page-17-2"></span>Brookes G, Barfoot P (2009) Global impact of biotech crops. Income and production effect 1996-20017. AgBioforum 12:184–208
- <span id="page-17-6"></span>Brown CT, Hug LA, Thomas BC, Sharon I, Castelle CJ, Singh A et al (2015) Unusual biology across a group comprising more than 15% of domain Bacteria. Nature 523:208–211
- <span id="page-17-3"></span>Bueno MM, Thys RCS, Rodrigues RC (2016) Microbial enzymes as substitutes of chemical additives in baking wheat four-part II: combined effects of nine enzymes on dough rheology. Food Bioprocess Technol 9:1598–1611
- <span id="page-17-8"></span>Buikema WJ, Haselkorn R (2001) Expression of the Anabaena hetR gene from a copper-regulated promoter leads to heterocyst differentiation under repressing conditions. Proc Natl Acad Sci U S A 98:2729–2734
- <span id="page-17-17"></span>Butterbach-Bahl K, Baggs EM, Dannenmann M, Kiese R, Zechmeister-Boltenstern S (2013) Nitrous oxide emissions from soils: how well do we understand the processes and their controls? Philos Trans R Soc B 368
- <span id="page-17-0"></span>Carpenter J, Felsot A, Goode T, Hammig M, Onstad D, Sankula S (2002) Comparative environmental impacts of biotechnology-derived and traditional soybean, corn, and cotton crops. Council for Agricultural Science and Technology (CAST), Ames
- <span id="page-17-10"></span>Chang CH, Yang SS (2009) Thermotolerant phosphate solubilizing microbes for multifunctional biofertilizer preparation. Bio/Technology 100(4):1648–1658
- <span id="page-17-9"></span>Chaurasia AK, Parasnis A, Apte SK (2008) An integrative expression vector for strain improvement and environmental applications of the nitrogen fxing cyanobacterium, Anabaena sp. strain PCC7120. J Microbiol Methods 73:133–141
- <span id="page-17-16"></span>Chen J (2006) The combined use of chemical and organic fertilizer and or biofertilizer for crop growth and soil fertility. International workshop on sustained management of the soilrhizosphere system for effcient crop production and fertilizer use, pp 37–43
- <span id="page-17-1"></span>Chen Z, Wang H-G, Wen Z-J, Wang Y (2007) Life sciences and biotechnology in China. Philos Trans R Soc B 362:947–957
- <span id="page-17-13"></span>Chun-Li W, Shiuan-Yuh C, Chiu-Chung Y (2014) Present situation and future perspective of biofertilizer for environmentally friendly agriculture. Annu Rep:1–5
- <span id="page-18-1"></span>Cohen SN, Chang AC, Boyer HW, Helling RB (1973) Construction of biologically functional bacterial plasmids in vitro. Proc Natl Acad Sci U S A 70:3240–3244
- <span id="page-18-7"></span>Dash A, Kundu D, Das M, Bose D, Adak S, Banerjee R (2016) Food biotechnology: a step towards improving nutritional quality of food for asian countries. Recent Pat Biotechnol 10:43–57
- <span id="page-18-20"></span>Egamberdiyeva D (2007) The effect of plant growth promoting bacteria on growth and nutrient uptake of maize in two different soils. Appl Soil Ecol 36:184–189
- <span id="page-18-5"></span>Ewen SW, Pusztai A (1999) Effects of diets containing genetically modifed potatoes expressing Galanthusnivalis lectin on rat small intestine. Lancet 354:1314–1315
- <span id="page-18-18"></span>Gonzalez AJ, Larraburu EE, Llorente BE (2015) Azospirillum brasilense increased salt tolerance of jojoba during in vitro rooting. Ind Crop Prod 76:41–48
- <span id="page-18-22"></span>Goswami D, Pithwa S, Dhandhukia P, Thakker JN (2014) Delineating Kocuria turfanensis 2M4 as a credible PGPR: a novel IAA producing bacteria isolated from saline desert. J Plant Interact 9:566–576
- <span id="page-18-16"></span>Gupta AK (2004) The complete technology book on biofertilizer and organic farming. National Institute of Industrial Research Press, Delhi, pp 242–253
- <span id="page-18-17"></span>Hayat R, Ali S, Amara U (2010) Soil benefcial bacteria and their role in plant growth promotion: a review. Ann Microbiol 60:579–598
- <span id="page-18-4"></span>He Z, Wlazlo AP, Kowalczyk DW, Cheng J, Xiang ZQ, Giles-Daves W et al (2000) Viral recombinant vaccines to the E6 and E7 antigens of HPV-16. Virology 270:146–161
- <span id="page-18-0"></span>Horvath DM, Stall RE, Jones JB, Pauly MH, Vallad GE, Dahlbeck D, Staskawisz BJ, Scott JW (2012) Transgenic resistance confers effective feld level control of bacterial spot disease in tomato. PLoS One 7:e42036
- <span id="page-18-3"></span>Huang J, Rozelle S, Pray C, Wang Q (2001) Plant biotechnology in China. Science 295:674–677
- <span id="page-18-6"></span>Huang X, Dong Y, Zhao J (2004) HetR homodimer is a DNA-binding protein required for heterocyst differentiation, and the DNA-binding activity is inhibited by Pat S. Proc Natl Acad Sci U S A 101:4848–4853
- <span id="page-18-14"></span>Hussain N, Mujeeb F, Tahir M et al (2002) Effectiveness of Rhizobium under salinity stress. Asian J Plant Sci 4:124–129
- <span id="page-18-19"></span>Joo GJ, Kim YM, Kim JT (2005) Gibberellins-producing rhizobacteria increase endogenous gibberellins content and promote growth of red peppers. J Microbiol 43:510–515
- <span id="page-18-13"></span>Joseph CM, Meeks JC (1987) Regulation of expression of glutamine synthetase in a symbiotic Nostoc strain associated with Anthoceros punctatus. J Bacteriol 169:2471–2475
- <span id="page-18-2"></span>Kakumanu K, Ambavaram MMR, Klumas C, Krishnan A, Batlang U, Myers E, Grene R, Pereira A (2012) Effects of drought on gene expression in maize reproductive and leaf meristem tissue revealed by RNASeq. Plant Physiol 160:846–867
- <span id="page-18-21"></span>Kanchiswamy CN, Malnoy M, Maffei ME (2015) Chemical diversity of microbial volatiles and their potential for plant growth and productivity. Front Plant Sci 6:151
- <span id="page-18-8"></span>Khosro M, Yousef S (2012) Bacterial biofertilizers for sustainable crop production: A review. APRN J Agric Biol Sci 7(5):237–308
- <span id="page-18-15"></span>Kloepper JW, Schroth MN (1978). Plant growth promoting rhizobacteria on radishes. In: Station de Pathologie Vegetale et Phytobacteriologie, ed. Proceedings of the 4th international conference on plant pathogenic bacteria II. Tours: Gilbert-Clary, 879–882
- <span id="page-18-12"></span>Lavakush YJ, Verma JP, Jaiswal DK, Kumar A (2014) Evaluation of PGPR and different concentration of phosphorous level on plant growth, yield and nutrient content of rice (Oryza sativa). Ecol Eng 62:123–128
- <span id="page-18-9"></span>Leonardo D, Blanca LF, Landa B, Weller DM (2006) Host crop affects rhizosphere colonization and competitiveness of 2,4-diacetylphloroglucinol-producing Pseudomonas fuoresens. Phytopathology 96:751–762
- <span id="page-18-11"></span>Liddycoat SM, Greenberg BM, Wolyn DJ (2009) The effect of plant growth promoting rhizobacteria on Asparagus seedlings and germinating seeds subjected to water stress under green house conditions. Can J Microbiol 55:388–394
- <span id="page-18-10"></span>Liu D, Golden JW (2002) hetL overexpression stimulates heterocyst formation in Anabaena sp. strain PCC 7120. J Bacteriol 184:6873–6881
- <span id="page-19-1"></span>McAllister CH, Beatty PH, Good AG (2012) Engineering nitrogen use effcient crop plants: the current status. Plant Biotechnol 10:1011–1025
- <span id="page-19-9"></span>Mflinge A, Mtei K, Ndakidemi (2014) Effect of rhizobium inoculation and supplementation with phosphorus and potassium on growth leaf chlorophyll content and nitrogen fxation of bush bean varieties. Am J Res Commun 2(10):49–87
- <span id="page-19-5"></span>Mitragotri S, Burke PA, Langer R (2014) Overcoming the challenges in administering biopharmaceuticals: formulation and delivery strategies. Nat Rev Drug Discov 13:655–672
- <span id="page-19-21"></span>Nautiyal CS, Govindarajan R, Lavania M, Pushpangadan P (2008) Novel mechanisms of modulating natural antioxidants in functional foods: involvement of plant growth promoting rhizobacteria NRRL B-30488. J Agric Food Chem 56:4474–4481
- <span id="page-19-0"></span>Okeno JA, Wolt JD, Misra MK, Rodriguez L (2012) Africa's inevitable walk to genetically modifed (GM) crops: opportunities and challenges for commercialization. New Biotechnol 30:124–130
- <span id="page-19-2"></span>Overton TW (2014) Recombinant protein production in bacterial hosts. Drug Discov Today 19:590–601
- <span id="page-19-20"></span>Pishchik VN, Vorobyev NJ, Chernyaeva LI, Timofeeva SV, Kazhemyakov AP, Alexeev YV (2002) Experimental and mathematical simulation of plant growth promoting rhizobacteria and plant interaction under cadmium stress. Plant Soil 243:173–186
- <span id="page-19-4"></span>Raja N (2013) Bipesticides and biofertilizers: ecofriendly sources for sustainable agriculture. J Biofertil Biopest 3:112–115
- <span id="page-19-10"></span>Rajaram H, Apte SK (2008) Nitrogen status and heat-stress-dependent differential expression of the cpn60 chaperonin gene infuences thermotolerance in the cyanobacterium Anabaena. Microbiology 154:317–325
- <span id="page-19-17"></span>Reid MS (1981) The role of ethylene in fower senescene. Acta Hortic 261:157–169
- <span id="page-19-15"></span>Riefer M, Novak O, Strnad M, Schm ulling T (2006) Arabidopsis cytokinin receptor mutants reveal functions in shoot growth, leaf senescence, seed size, germination, rootdevelopment, and cytokinin metabolism. Plant Cell 18: 40–54
- <span id="page-19-19"></span>Ritika B, Uptal D (2014) Biofertilizer a way towards organic agriculture: a review. Acad J 8(24):2332–2342
- <span id="page-19-8"></span>Ruiz-Sanchez M, Aroca R, Monoz Y et al (2010) The arbuscular mycorrhiza symbiosis enhances the photosynthetic effciency and the antioxidative response of rice plants subjected to drought stress. J Plant Physiol 167:862–869
- <span id="page-19-3"></span>Sanahuja G, Banakar R, Twyman RM, Capell T, Christou P (2011) Bacillus Thuringiensis: a century of research, development and commercial applications. Plant Biotechnol J 9(3):283–300
- <span id="page-19-7"></span>Santos VB, Araujo SF, Leite LF et al (2012) Soil microbial biomass and organic matter fractions during transition from conventional to organic farming systems. Geodderma 170:227–231
- <span id="page-19-14"></span>Scalenghe R, Edwards AC, Barberis E et al (2012) Agricultural soil under a continental temperature climate susceptible to episodic reducing conditions and increased leaching of phosphorus. J Environ Manag 97:141–147
- <span id="page-19-11"></span>Schiefer W, Schutz K, Hachtel W, Happe T (2002) Molecular cloning mand characterization of hetR genes from flamentous cyanobacteria. Biochim Biophys Acta 1577:139–143
- <span id="page-19-16"></span>Sokolova MG, Akimova GP, Vaishlia OB (2011) Effect of phytohormones synthesized by rhizosphere bacteria on plants. Prikl Biokhim Mikrobiol 47:302–307
- <span id="page-19-6"></span>Somasegaran P, Springer H (1994) Carrier materials used in biofertilizer making . Nature publisher's. 2-6
- <span id="page-19-12"></span>Stewart WDP (1980) Some aspects of structure and function in nitrogenfxing cyanobacteria. Annu Rev Microbiol 34:497–536
- <span id="page-19-18"></span>Tabashnik BE, Huang F, Ghimire MN, Leonard BR, Siegfried BD, Rangasamy M, Yang Y, Wu Y, Gahan LJ, Heckel DG, Bravo A, Soberón M (2011) Effcacy of genetically modifed BT toxins against insects with different genetic mechanisms of resistance. Nat Biotechnol 12:1128–1131
- <span id="page-19-13"></span>Vessey JK (2003) Plant growth promoting Rhizobacteria as biofertilizers. J Plant Soil 225(43):571–586
- <span id="page-20-4"></span>Vitorino LC, Bessa LA (2017) Technological Microbiology: Development and Applications. Front Microbiol 8:827. <https://doi.org/10.3389/fmicb.2017.00827>
- <span id="page-20-0"></span>Wani SA, Chand S, Ali T (2013) Potential use of Azotobacter chroococcum in crop production: an overview. Curr Agric Resour J 1:35–38
- <span id="page-20-3"></span>Yang JW, Kloeppe JW, Ryu CM (2009) Rhizosphere bacteria help plants tolerate abiotic stress. Trends Plant Sci 14(1):1–4
- <span id="page-20-1"></span>Youssef MMA, Eissa MFM (2014) Biofertilizers and their role in management of plant parasitic nematodes: a review. Biotechnol Pharm Res 5(1):1–6
- <span id="page-20-2"></span>Zhou R, Cao Z, Zhao J (1998) Characterization of HetR protein turnover in Anabena sp. PCC7120. Arch Microbiol 169:417–423