# **When the Race Is Lost: The Clinical Impact of Prosthetic Joint Infections**



**Justin Vaida and Matthew J. Dietz**

**Abstract** Joint arthroplasty, a procedure that can relieve patients of life-altering and debilitating pain, has proven to be so successful that over 1,000,000 procedures are performed annually in the USA alone. However, a prosthetic joint infection represents a devastating complication for patients which can lead to not only revision surgery but possible permanent loss of function, amputation, and even death. Infection can present not only in the immediate postoperative period but at any point for the duration of the implant's life.

The challenges confronting providers are numerous. Diagnostic testing has varying sensitivities and specificities depending on duration of infection meaning there is no true "gold standard" for testing. Additionally, once the diagnosis of infection is made, treatment options are limited and have high rates of morbidity and mortality.

Given the impending rise in total joint arthroplasty case volume and subsequent revision case volume due to PJI, an urgent need exists for continued work in the development of preventive, diagnostic, and therapeutic tools.

**Keywords** Joint arthroplasty · Infection · Biofilm · PJI · TJA · Diagnosis · Treatment

# **Epidemiology/Incidence**

The goal of total joint arthroplasty (TJA) is to relieve a patient of debilitating joint pain by replacing the biological joint surface with prosthetic implants. This surgery is most often performed in the setting of severe osteoarthritis although rheumatoid arthritis and osteonecrosis of the hip are other common indications. A patient becomes an appropriate surgical candidate only after conservative measures such as

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anti-inflammatory medications, physical therapy, and various injections have been exhausted and failed. TJA has proved to be one of the most successful surgeries performed, with consistently high patient satisfaction scores [\[1](#page-22-0), [2](#page-22-1)]. The success of the procedure has led to more than 370,000 primary total hip arthroplasties (THA) and 680,000 primary total knee arthroplasties (TKA) performed in 2014 in the USA alone [\[3](#page-22-2)].

Periprosthetic joint infection (PJI) is one of the most feared and devastating complications of TJA due to clinical challenges in diagnosis and treatment and the extreme financial, physical, and emotional costs to the patient [[4\]](#page-22-3). Despite evolving preoperative and intraoperative regimens to reduce infection risk, estimates of PJI incidence rates for both primary hip and knee arthroplasties range from approximately 0.5–2.0%, with a slightly higher rate of PJI in TKA compared to THA [[5–](#page-22-4)[8\]](#page-22-5). More troubling is that failure rates after the first line of treatment for PJI often exceed 25% with an increasing failure rate with repeated subsequent revision procedures. With an average cost of \$116,383 for an infected total hip arthroplasty and \$88,623 for an infected total knee arthroplasty, a substantial burden is placed on the healthcare system [[9,](#page-22-6) [10](#page-22-7)]. Additional costs are assumed by the patient in extended rehabilitation, prolonged hospital stays, and emotional costs [[11,](#page-22-8) [12\]](#page-22-9). Ultimately, PJI can lead to not only revision surgery but also possible is permanent loss of function, amputation (Fig. [1\)](#page-2-0), and even death, with the 1-year mortality of PJI in THA at 7% between stages of a two-stage revision and 33% within 5 years of completion of revision [[13,](#page-22-10) [14\]](#page-22-11).

Among the many challenges facing clinicians are shifting definitions of PJI, myriad diagnostic tests that, while helpful in aggregate, lack 100% accuracy, treatment regimens that have unacceptably high failure rates, and a spectrum of disease representing a moving and evolving target. With a projected 1.9 million combined primary total hip and knee arthroplasties expected to be performed annually by 2030 in the USA alone [[3\]](#page-22-2), there is an expected proportional increase in the infection burden. Moreover, some research has indicated this projection may prove too conservative and a more dramatically exponential increase in the TJA caseload may be borne out [[15\]](#page-22-12). Therefore, it is imperative that clinicians and basic scientists continue research into preventing, diagnosing, and treating this devastating complication.

## **Overview of Challenges**

The diagnosis and treatment of PJI depends largely on two factors: the time since index surgery and the duration of a patient's signs and symptoms. While in reality the infectious process occurs on a continuum and does not adhere to discrete intervals, the sensitivity and specificity of various diagnostic tests as well as treatment algorithms have historically been categorized by these two crucial factors. Because successful eradication of infection depends in part on the identification of where a patient falls on this spectrum, several categorizations have been proposed [[16–](#page-22-13)[19\]](#page-22-14).

<span id="page-2-0"></span>

**Fig. 1** (**a**–**c**) Progression in a single patient from infected revision total knee arthroplasty to knee fusion and ultimately to amputation due to recurrent infections. This patient eventually suffered multiorgan system failure and succumbed to his infection

For diagnostic and therapeutic purposes, PJI has been delineated into three broad categories [\[20](#page-23-0)]:

- Early postoperative infection: within 4 weeks of index surgery.
- Acute hematogenous infection (AHI): less than 3 weeks of symptoms. These infections typically cause an abrupt onset of symptoms that progress rapidly in severity.

• Chronic (late) hematogenous infection (CHI): seeded from a remote source and can be present at any point in the life of the patient after TJA, from months to years [[21,](#page-23-1) [22\]](#page-23-2). Symptoms develop more gradually.

Part of the rationale for temporal categorization is directly related to one of the unique challenges inherent in PJIs: the ability of infective bacteria to form a "biofilm"—a metabolically cooperative colony surrounded by an extracellular glycocalyx (Fig. [2\)](#page-3-0). Unfortunately, the same synthetic joint surface of the prosthesis which so successfully relieves pain in a patient makes the formation of this biofilm easier. Bacteria can adhere to prosthetic surfaces and resist mechanical disturbance making biofilm eradication especially challenging and often necessitates complete implant removal. Further, the glycocalyx protects bacteria not only from antibiotics and host antibodies but also detection from diagnostic testing. Biofilms have been found in samples from confirmed PJI cases in which preoperative cultures were negative [\[23](#page-23-3)]. It has been postulated that PJIs exhibit alternating periods of quiescent growth and acute exacerbations caused by the release of bacteria [\[24](#page-23-4)]. In the quiescent phases, few if any symptoms may be present, while during acute phases, symptoms may be limited locally to the affected joint or systemically manifesting as a fever, malaise, or frank septicemia. It is imperative that when a clinician suspects PJI, the patient is treated as soon as possible as a mature biofilm may be less likely to develop into acute PJI and implants may possibly be retained.

<span id="page-3-0"></span>

**Fig. 2** Scanning electron microscopy image of *Staphylococcus aureus* biofilm adherent to a stainless steel disc

It should be noted that this categorization is more to help in the development of treatment algorithms and research purposes than based on a demonstrated bacterial threshold as studies have shown biofilm formation within mere hours of joint inoculation [[25\]](#page-23-5). Therefore, the consideration of infection as a spectrum rather than a distinct acute/chronic dichotomy is likely more accurate.

# **Host Risk Factors**

Prevention remains the best step in getting a head start in the race to the surface and several risk factors have been identified which aid clinicians in identifying patients predisposed towards PJI (Table [1](#page-4-0)). These include characteristics considered both modifiable, such as elevated body mass index, poorly controlled diabetes, tobacco

Host risk factors for PJI/SSI in TJA		
Modifiable	Nonmodifiable	
Active infection	Age	
Alcoholism	$ASA$ score $> 2$	
Cardiovascular disease	Bariatric surgery	
Congestive heart failure		
Cardiac arrhythmia		
Chronic kidney disease	Chronic anticoagulation	
Chronic obstructive pulmonary disease	Hemiplegia/paraplegia	
Clotting disorders	<b>HBV</b>	
Depression	Osteonecrosis	
Diabetes mellitus	Previous joint infection	
H <sub>b</sub> A <sub>1</sub> c		
Serum glucose		
Drug abuse	Previous joint surgery	
Frailty	Previous infection	
<b>HIV/AIDS</b>	<b>Sex</b>	
Immunosuppression	Transplant	
Intra-articular steroid/viscosupplement injection		
Malnutrition		
MRSA colonization		
Obesity		
Peripheral vascular disease		
Psychosis		
Renal disease		
Rheumatoid arthritis		

<span id="page-4-0"></span>**Table 1** Host risk factors for PJI/SSI in TJA [[122](#page-28-0)]

Outlines the modifiable and nonmodifiable risk factors known to impact the risk for prosthetic joint infection as described by Cizmic et al.

*HbA1c* hemoglobin A1c, *HIV* human immunodeficiency virus, *AIDS* acquired immunodeficiency syndrome, *MRSA* Methicillin-resistant *Staphylococcus aureus*, *ASA* American Society of Anesthesiologists, *HBV* hepatitis B

use, alcohol consumption, and immunosuppression, as well as nonmodifiable, such as previous joint surgery and previous PJI [[26,](#page-23-6) [27\]](#page-23-7).

#### *Body Mass Index (BMI)*

BMI is recognized as a major risk factor with postoperative infection rates 6.7 times higher after TKA and 4.2 times higher after THA in patients with BMI  $\geq$  35 kg/m<sup>2</sup> compared to nonobese patients [[28\]](#page-23-8). This risk elevates incrementally with each point increase in BMI >  $25 \text{ kg/m}^2$  (hazard ratio of 1.09 per unit) [[29,](#page-23-9) [30\]](#page-23-10).

## *Diabetes*

For diabetic patients, a hemoglobin A1c level of 7.5 has been shown to almost triple the risk of PJI in TKA when compared to those below this threshold [\[31](#page-23-11)]. Studies have demonstrated PJI rates in both TKA and THA rising from 0.8 to 5.4% with an A1c of 7.7 or higher compared to an A1c  $\leq$  7.6 [\[32](#page-23-12)].

## *Lifestyle Factors*

Several other lifestyle factors have demonstrated an elevated risk of PJI. For instance, current tobacco users have more than double the risk of PJI than nonsmokers (OR 2.16 [1.57–2.97]) and this risk persists even after smoking cessation (OR 1.52 [1.16–1.99]) [[33\]](#page-23-13). Alcohol consumption in the perioperative period has been shown to increase the risk of PJI and alcohol cessation is recommended at least 4 weeks prior to surgery to reduce postoperative morbidity [\[34](#page-23-14)[–36](#page-23-15)]. Several of these lifestyle factors are related to effects on wound healing and coagulation.

#### *Modifiable Medical Risk Factors*

While some TJA patients suffer from chronic conditions such as rheumatoid arthritis (RA) which elevate the risk of PJI, these patients should be medically optimized prior to surgery [[37\]](#page-23-16). For instance, those on biologic disease-modifying antirheumatic drugs (DMARDs) should discontinue these medications in the perioperative period (approximately 2 weeks before and after surgery, based on drug half-life) as numerous studies have shown an increased risk of surgical site infections (SSIs)/ PJIs with perioperative use of these medications [\[38](#page-23-17)[–40](#page-24-0)]. This elevated risk is borne out in patients using glucocorticoids within 90 days of surgery as well due to immunosuppressive effects [[37\]](#page-23-16). However, those on nonbiologic DMARDs (e.g., methotrexate, leflunomide) can continue these medications throughout the perioperative period. Finally, anemia has consistently been found to be a factor in both the incidence and failure of treatment of PJI and is another example of a modifiable medical risk factor [\[34](#page-23-14), [35](#page-23-18), [41](#page-24-1), [42](#page-24-2)].

#### *Nonmodifiable Risk Factors*

Several significant nonmodifiable risk factors such as prior PJI and prior joint surgery have been identified. A history of prior joint infection increases the risk of PJI 5.0–21.0 times [[43–](#page-24-3)[45\]](#page-24-4). Patients with a history of previous joint surgery have almost tripled the risk of PJI [\[46](#page-24-5)]. While risk factors such as these cannot be modified, an awareness of these patient factors and heightened surveillance enables clinicians to detect PJI in its early stages thereby increasing the likelihood of treatment success. Further, it should be noted that, while many host factors exist, they should all be taken in the context of the patient to determine the overall risk of PJI with no single factor providing a definitive risk assessment.

To synthesize several of these risk factors, calculators are available online enabling clinicians to compute the overall risk of PJI. Risk factors utilized include BMI, sex, race, insurance status, smoking, drug abuse, prior surgeries, and various comorbidities [[47\]](#page-24-6). While these calculators do not yield an absolute risk assessment, they can be a useful aid in guiding diagnostic testing.

## **Diagnosis**

The diagnosis of PJI remains challenging due to the lack of a single test with 100% accuracy. Instead, a clinician must rely on a combination of clinical history, physical examination, imaging, laboratory testing including serological and synovial markers, microbiological culture, and intraoperative findings.

# *Clinical Presentation*

Most often, PJI is suspected initially due to patient symptomatology rather than incidental findings on imaging or laboratory testing in an asymptomatic patient. Because a patient can become infected even years after surgery, any patient with a history of a TJA presenting with a painful joint should raise a clinician's suspicion. Unlike an acute infection typically associated with an abrupt onset of rapidly progressing symptoms, chronic PJI represents an indolent infection with gradual progression of less severe symptoms. In the settings of both acute and chronic PJIs, patients most

<span id="page-7-0"></span>

**Fig. 3** (**a**, **b**) Clinical pictures of a suspected sinus tract and wound complication in the setting of a presumed PJI in a TKA. (**c**) Radiograph of a suspected infected THA. (**d**) Joint arthrogram confirming a sinus tract communicating from the joint directly to the skin surface. The presence of a sinus tract is diagnostic of a PJI

often present with a progressively painful joint [[48\]](#page-24-7). Fever, while specific for PJI, is inconsistently present and has been found in 32.5% of early postoperative infections, 75.5% of AHI, and 14.0% of CHI [[48\]](#page-24-7). Fever is also frequently present in the immediate postoperative period as a normal physiological response to surgery [[49\]](#page-24-8). Local signs of inflammation such as warmth or diffuse swelling are more easily visible at the knee due to the more superficial nature of the joint. The presence of warmth in TKA and THA was found in 50% and 14% cases of PJI, respectively, while effusions were found in 75% and 29%, respectively [\[50](#page-24-9)]. Local warmth or hyperemia can be

Major criteria (at least one of the following)			Decision	
Two positive growths of the same organism using standard culture methods			Infected	
Sinus tract with evidence of communication to the joint or				
visualization of the prosthesis				
Minor criteria	<b>Threshold</b>		<b>Score</b>	<b>Decision</b>
	Acute	Chronic		
Serum CRP (mg/L)	100	10	$\overline{c}$	Combined preoperative and postoperative score:
<i>or</i>			$\geq 6$ infected	
D-Dimer $(\mu g/L)$	Unknown	860		$3-5$ inconclusive <sup><math>a</math></sup>
Elevated serum ESR (mm/h)	No role	30	$\mathbf{1}$	<3 not infected
Elevated synovial WBC (cells/ $\mu$ L)	10,000	3000	$\mathcal{E}$	
<i>or</i>				
Leukocyte esterase	$^{++}$	$^{++}$		
or				
Positive alpha-defensin (signal/ cutoff)	1.0	1.0		
Elevated synovial PMN $(\%)$	90	70	2	
Single positive culture			$\overline{2}$	
Positive histology intraoperative (whites/HPF)		3		
Positive intraoperative purulence		3		

<span id="page-8-0"></span>**Table 2** International Consensus Meeting (ICM) on Musculoskeletal Infection criteria [[70](#page-25-0)]

International Consensus Meeting criteria for diagnosing a PJI. Acute infections defined as occurring less than 3 months from index arthroplasty, and acute hematogenous PJI, defined as symptoms occurring for less than 6 weeks but more than 3 months from index surgery

*CRP* C-reactive protein; *ESR* erythrocyte sedimentation rate; *WBC* white blood cell; *PMN* polymorphonuclear; *HPF* high-power field

Reprinted from The Journal of Arthroplasty, 34(2s), Shohat et al., Hip and Knee Section, What is the Definition of a Periprosthetic Joint Infection (PJI) of the Knee and the Hip? Can the Same Criteria be Used for Both Joints?: Proceedings of International Consensus on Orthopedic Infections, S325-s327, Copyright (2019), with permission from Elsevier

a Consider further molecular diagnosis

confusing in the acute postoperative period as this can often be related to increased blood flow in the area due to the normal healing response after surgery.

These signs may be accompanied by skin changes such as erythema, puckering, or obvious drainage of fluid. Drainage must be examined with particular scrutiny as it may be the result of a sinus tract or fistula communicating directly with the joint (Fig. [3a–d\)](#page-7-0). The presence of a sinus tract or abscess represents deep involvement and, due to its high sensitivity and specificity, represents one of the major criteria for diagnosis of PJI (Table [2](#page-8-0)).

On physical examination, a number of signs may be present with manipulation of the joint such as crepitus or bogginess. The patient may feel pain with either palpation or range of motion especially at extremes of motion. The reported presence of stiffness or restricted range of motion varies widely but has been reported as high as 74% in TKA and 85% in THA [\[50](#page-24-9)]. Conversely, laxity may be appreciated

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**Fig. 4** (**a**, **b**) Normal postoperative radiographs showing a well-fixed prosthesis in a TKA with normal cement fixation. (**c**, **d**) Radiographs of a prior revision total knee arthroplasty in the setting of a PJI; classic signs of infection are visible including osteolysis, bone remodeling, loss of cement fixation, heterotopic bone formation, and migration of the prosthesis within the intramedullary canal. (**e**, **f**) Intraoperative findings demonstrating infection and loose femoral and tibial components

as a function of component subsidence or loosening. Unfortunately, all of these findings are fairly nonspecific.

## *Imaging*

Initial imaging consists of anterior–posterior (AP) and lateral radiographs (X-rays) and should be compared to previous X-rays if available (Fig. [4a, b](#page-9-0)). Additional specialized views are not routinely acquired. Several signs are possible on X-ray imaging to support a diagnosis of PJI. Osteolysis, as represented by radiolucency, may be visible and may be indirectly seen as manifested by component loosening or subsidence, as the foundation upon which it rests has been compromised. Sinus tracts may be inferred by the appearance of a distinct interruption of the cortex. Finally, a generalized periosteal reaction may be seen (Fig. [4c–f](#page-9-0)). Unfortunately, X-rays are fraught with limitations due to their low sensitivity as a significant amount of cortical disruption must be present in order to be visible on an X-ray, diminishing their utility especially in early infections. Further, distinguishing loosening as a result of PJI vs. aseptic etiology is challenging [\[51\]](#page-24-10). Despite these limitations, the low cost and ease of acquisition make radiographs an ideal component of initial workup of a painful joint.

Advanced imaging modalities include bone scintigraphy, or "bone scan," which may be utilized when infection is strongly suspected but serological markers or synovial fluid analysis are equivocal. Leukocyte, antigranulocyte, and combined leukocyte and bone marrow scintigraphy are also available. All scintigraphy modalities are fairly sensitive (80–99%); however, the specificities among the tests vary widely due to false positives from conditions such as a fracture or bone remodeling. This trend is especially true in the first 12 months following surgery as periprosthetic bone remodeling continues. Therefore, these studies are typically reserved for the setting of chronic or low-grade infection and can be a useful adjunct in diagnosis. Positron emission tomography can be employed and has a sensitivity of 70% and specificity of 84%. However, due to its high cost, it is rarely used [\[52\]](#page-24-11).

Computerized tomography (CT), magnetic resonance (MR), and ultrasound imaging are not routinely employed in the diagnosis of PJI due to their significant limitations. CT and MR both suffer from beam hardening and projection data noise resulting in image artifacts, making it difficult to distinguish bony architecture surrounding the prosthesis. Ultrasound is most useful in evaluating small areas of soft tissue architecture or the presence of fluid collections or sinus tracts [\[53\]](#page-24-12), but does not provide useful information about bone morphology or implant position.

## *Criteria*

Once a thorough history and clinical examination raise suspicion for a PJI, the algorithm endorsed by the American Academy of Orthopaedic Surgeons (AAOS) begins with the noninvasive serum biomarkers C-reactive protein (CRP) and erythrocyte sedimentation rate  $(ESR)$  [\[54](#page-24-13)] (Fig. [5\)](#page-11-0). Due to their high sensitivities, they have been used as reliable predictors of the absence of infection [[55\]](#page-25-1). Recent metaanalysis, however, suggests sensitivities of 88% and 75% and specificities of 74% and 70%, respectively and debate continues as to their utility [[56\]](#page-25-2). It should be noted that CRP increases postoperatively from a baseline, peaking on postoperative day 2–3 and normalizing by day 21 [[57\]](#page-25-3). Therefore, the thresholds used are >100 mg/L in the acute phase and >10 mg/L in the chronic phase. Similarly, ESR

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Algorithm for patients with a higher probability of hip or knee periprosthetic joint infection.

a Perform repeat aspiration when a discrepancy exists between the probability of infection and the result of the initial aspiration culture

b Perform frozen section when the diagnosis has not been established at the time of surgery; synovial fluid white blood cell count and differential may also be obtained intraoperatively

c Nuclear imaging modalities: labeled-leukocyte imaging combined with bone or bone marrow imaging, F-18 fluorodeoxyglucose–positron emission tomography, gallium imaging, or label-leukocyte imaging

CRP = C-reactive protein, ESR = erythrocyte sedimentation rate

**Fig. 5** An algorithm endorsed by the American Academy of Orthopaedic Surgeons used in the diagnosis of a PJI. (Reprinted with permission from Della Valle et al., Diagnosis of Periprosthetic Joint Infections of the Hip and Knee, Journal of the American Academy of Orthopaedic Surgeons, 18(12), 762, [https://journals.lww.com/jaaos/Fulltext/2010/12000/Diagnosis\\_of\\_Periprosthetic\\_](https://journals.lww.com/jaaos/Fulltext/2010/12000/Diagnosis_of_Periprosthetic_Joint_Infections_of.6.aspx#pdf-link) [Joint\\_Infections\\_of.6.aspx#pdf-link](https://journals.lww.com/jaaos/Fulltext/2010/12000/Diagnosis_of_Periprosthetic_Joint_Infections_of.6.aspx#pdf-link))

increases immediately postoperatively, though it demonstrates a slow and irregular decline for several months after surgery and thus has more diagnostic utility in a chronic PJI with a threshold of  $>$ 30 m/h [[57,](#page-25-3) [58\]](#page-25-4). Ultimately CRP and ESR, as with other serum and synovial markers, must be interpreted with respect to time from index surgery.

Research is underway examining the utility of D-Dimer, a fibrinolytic byproduct. One study found that using a threshold of 850 ng/mL resulted in a sensitivity of 89% and a specificity of 93% compared to a combined CRP/ESR sensitivity and specificity of 84% and 47%, respectively [\[59](#page-25-5)]. Further studies are underway to validate its utility but, due to its success, it is included in the current International Consensus Meeting on Musculoskeletal Infection (ICM) criteria (Table [2\)](#page-8-0).

## *Joint Aspiration*

If either ESR or CRP is elevated, the clinician should consider aspiration of the concerned joint for synovial fluid analysis which provides the most direct non operative assessment. If no serological markers are elevated, PJI is unlikely though continued clinical suspicion warrants aspiration. Analysis of synovial fluid yields a variety of diagnostic tests including cell count, culture, and inflammatory biomarkers. Synovial fluid cell count with differential and leukocyte esterase is most commonly obtained. Leukocyte count (WBC), leukocyte esterase, and polymorphonuclear cell percentage (PMN%) have demonstrated sensitivities of 89%, 77%, and 89%, respectively, and specificities of 86%, 95%, and 86%, respectively. Alpha-defensin is a marker which has grown in popularity due to its reported 97% sensitivity and 96% specificity, though recently its sensitivity of point of care testing and sensitivity after prior treatment for PJI have both been called into question [[60–](#page-25-6)[62\]](#page-25-7). Regardless, initial promising studies have warranted its inclusion in the current ICM criteria. As with serological markers, synovial markers are affected by proximity to surgery. For example, the threshold for WBC in the acute period is 10,000 cell/μL and 3000 cells/ μL in the chronic period. Several other synovial markers, such as interleukin-6 (IL-6), IL-8, and CRP, demonstrate high sensitivities and specificities but are not routinely available at all institutions and therefore are not included in the standard workup for PJI.

## *Culture*

Joint aspiration also allows for bacterial and fungal culture of synovial fluid which has been found to be 94% specific and subsequently is part of the criteria for PJI, though two positive cultures are required as part of the major criteria. However, while culture may seem a likely candidate for a "gold standard" diagnostic test, it has also been found to have only 62% sensitivity [\[62](#page-25-7)]. Further, a review of the most

recent literature indicates a culture-negative PJI rate from 7.0% to as high as 42.1% [\[63](#page-25-8), [64](#page-25-9)]. The most common risk factors identified for culture-negative results were antecedent antibiotic use and presence of postoperative wound drainage. A culturenegative sample may be due to the presence of a biofilm in a quiescent state with a relative lack of planktonic bacteria to sample. Though the outcome of culturenegative PJIs is similar to that caused by known organisms, a negative culture hinders diagnosis and presents challenges in postoperative treatment as antibiotics cannot be tailored to specific sensitivities [\[65](#page-25-10)].

Various bacteria have been implicated in PJIs with a preponderance for grampositive organisms with *Staphylococcus aureus* (*S. aureus*) the most prevalent followed by coagulase-negative *Staphylococcus* [\[64\]](#page-25-9). Finally, in rare cases, PJI may result from fungal or mycobacterium species. In the case of fungal PJIs, which account for  $\langle 1\% \rangle$  of all PJIs, cultures may require up to 4 weeks of incubation [[66,](#page-25-11) [67\]](#page-25-12).

Next-generation (next-gen) sequencing, capable of sequencing all DNA present in a sample concurrently, allows for a more complete assessment of the microbes present. Studies have shown its ability to outperform traditional microbial culture with an 89.3% sensitivity vs. 60.7%. Moreover, next-gen sequencing was able to detect bacteria in 81.8% of culture-negative samples from PJIs and 25.0% in pre-sumed aseptic culture-negative revisions [[68\]](#page-25-13). Finally, next-gen sequencing has demonstrated an ability in some cases to reveal a polymicrobial infection, which on culture initially grew only a single organism [[69\]](#page-25-14). This finding may explain failure of some prior therapies and aid antibiotic selection in future cases. Further studies are underway but, due to the relatively high cost, next-gen sequencing is not a firstline test but may prove to be a useful adjunct.

Ultimately, these tests should be interpreted in the context of a patient's clinical history and presentation. To consolidate information and offer guidelines for clinicians, recommendations made by the Musculoskeletal Infection Society in 2011 and further amended at the International Consensus Meeting (ICM) in 2013, set forth guidelines to aid in the diagnosis of PJI. These guidelines were further amended and externally validated for chronic infections in 2018 and found to have a sensitivity of 97.7% and specificity of 99.5% [\[70](#page-25-0)] (Table [2](#page-8-0)). A robust debate continues within the orthopedic community as to the importance of each of the individual components; however, the establishment and revisiting of the Musculoskeletal Infection Society (MSIS) criteria provides a discussion and common roadmap for clinicians for the diagnosis of PJI.

#### **Treatment**

Once the diagnosis of PJI has been made, treatment is beset by its own challenges. There are a myriad of treatment regimens available with numerous surgical techniques, implant options, and antimicrobial therapies from which to choose. However, despite advancements and research over the last several decades, there remains a high recurrence rate. For instance, one study of 1.5 million infected TKA knees found a 26% recurrence rate of infection after first-line treatment [[71\]](#page-25-15). The selection of treatment and success of that regimen require a keen clinical acumen and continued surveillance.

## *Suppressive Antibiotic Therapy (SAT)*

For some patients, medical treatment alone via long-term antibiotic suppression may be the only choice. The goal of therapy is to reduce or at least keep in check the bioburden and thus the incidence of systemic effects cause by the bacterial infection. The scope of patients for whom this treatment would be considered is narrow, as it is not a curative option, but typically includes those medically unsuitable to undergo surgery, those who refuse surgery, and those for whom surgery would not improve functional outcomes. Antibiotic therapy should be continued for the remainder of the patient's life; however, adverse reactions may limit the patient's ability to tolerate such therapy. Several small studies indicate moderate success with 68.5–86.2% of patients maintaining a functioning prosthesis [\[72](#page-25-16)[–74](#page-26-0)]. However, complete resolution of a PJI requires surgical intervention; therefore, SAT should only be considered a palliative option.

### *Surgery*

Because of the presence of biofilm which prevents antibiotics from fully permeating and completely eradicating an infection, the only definitive procedure for the elimination of a PJI is open surgery.

## *Debridement and Irrigation with Implant Retention (DAIR)*

DAIR involves an open exposure of the joint to visualize and access the prosthetic implants. Most surgeons employ a surgical/mechanical debridement along with a chemical debridement. All surfaces are scrubbed with an antiseptic solution and irrigated with 6–9 L of sterile saline via low-pressure lavage while the easily exchanged components are removed and replaced, both to improve access to the joint and decrease the bioburden present. The indications for the procedure are narrow: early postoperative infections and acute hematogenous infections. Further, patients must be appropriate surgical candidates, have a microbial isolate of low virulence, and no sinus tract or wound complication present [\[75](#page-26-1)]. Several factors have been associated with treatment failure including presence of bacteremia and infection caused by *S. aureus* or *Enterococci* [[76\]](#page-26-2).

The advantage is a much faster, less expensive procedure with less morbidity and quicker recovery time. The operative time and blood loss are greatly increased when

components are removed which is avoided in this procedure. However, numerous studies have shown failure rates of 56–76% across multiple PJI chronicities caused by multiple organisms [[77–](#page-26-3)[79\]](#page-26-4). Outcomes appear to be improving, however, with time from surgery (<7 days) as a major factor in that success [\[80](#page-26-5)], and subsequent risk of failure rising with each additional day from the onset of symptoms [[81\]](#page-26-6). For the right patient population, DAIR is a viable option that can be considered.

## *Exchange Arthroplasty*

The current definitive treatment for PJI is an open procedure in which all components are removed and new components are implanted. There is an ongoing debate in the orthopedic community as to whether component exchange should be done in a single procedure or in two stages.

#### *Single-Stage Exchange*

In a single-stage exchange procedure, infected components and bone cement are removed, tissue is aggressively debrided, and definitive implants are placed in the same surgical setting. Local antibiotics are delivered during the procedure while postoperatively patients receive 4–6 weeks of intravenous antibiotics tailored to culture growth and sensitivities obtained from samples obtained during surgery. Indications include absence of bacteremia, positive isolation of causative organism with sensitivities, and minimal bone and soft tissue loss [[82,](#page-26-7) [83\]](#page-26-8). Relative contraindications include bacteremia, culture-negative PJI, poor bone stock for fixation of new components, presence of a sinus tract, and soft tissue deficiencies which would preclude adequate closure of the wound [[84–](#page-26-9)[88\]](#page-26-10).

The advantages of one-stage exchange compared to two-stage exchange are clear: decreased morbidity and mortality, reduced cost, and earlier functional return as only one surgery is required. However, research is conflicting with some data suggesting equivalence or superiority to two-stage exchange [\[89](#page-26-11)[–91](#page-27-0)] while others suggest an elevated risk of reinfection [[92\]](#page-27-1). However, research comparing the two treatment options suffer from heterogeneity of patient populations and are predominantly retrospective in nature. Therefore, several multicenter, prospective, randomized, controlled studies are underway to answer this important clinical question.

#### *Two-Stage Exchange*

In a two-stage exchange, the removal of infected components and reimplantation of new components occurs in two surgeries separated in time by the retention of an intra-articular polymethyl methacrylate (PMMA) spacer implanted during the first procedure. The PMMA spacer, which may be retained for several months or even years, is impregnated with antibiotics which passively elute over time. The patient additionally receives 4–6 weeks of intravenous and/or oral antibiotics. While there is no definitive threshold as to when it is safe to reimplant, most surgeons will monitor serological and/or synovial markers. When they feel it is clinically appropriate, the spacer is removed and new components are implanted in a second procedure.

The PMMA spacer utilized during a two-stage exchange may be static, preventing motion through the implantation of a joint-spanning rod, or dynamic which allows partial to full range of motion (Fig.  $6a$ , b). Due to the exothermic reaction (82–86 °C in femurs and 115 °C in tibias) that occurs during the curing of PMMA, the scope of antibiotics able to be incorporated is limited to those with heat stability, such as vancomycin or tobramycin [\[93](#page-27-2)]. Recent investigations suggest antibiotics once considered "heat-sensitive," such as ceftazadime, may be more resilient than previously thought but until more data are available, the antibiotic arsenal able to be incorporated into PMMA spacers remains limited [\[94](#page-27-3), [95\]](#page-27-4). Further, the antibiotics in a PMMA spacer elute in a burst fashion, peaking in concentration on postoperative days two and three and quickly decreasing over several weeks. Finally, the spacer cannot be redosed with antibiotics once implanted. Research has demonstrated low-dose intraosseous or intra-articular vancomycin administration results in equal or better tissue and synovial fluid concentrations when compared to systemic administration and minimizes side effects [[96,](#page-27-5) [97\]](#page-27-6). Methods to improve drug delivery options may provide improved treatment success allowing for minimization of both dosing concentrations and time needed for treatment.

Though the two-stage exchange procedure is associated with high rates of infection control for those patients who complete both stages (83–89.8%), it has also been shown to have substantial mortality [\[13](#page-22-10), [92](#page-27-1)]. Prior reports in the 90–95% success range likely did not account for the attrition due to death between stages [\[89](#page-26-11), [98\]](#page-27-7). After just the first stage for TKA, the 30-day readmission rate is 11.1% and 90-day mortality rate is 2.6% [\[99](#page-27-8)]. The 1-year and 5-year mortalities for completed TKA two-stage revision have been reported as 4.33% and 21.64%, respectively [\[100](#page-27-9)]. While considered the "gold standard" in the USA, two-stage revision leaves room for improvement of outcomes and places a substantial burden on the patient.

#### *Resection Arthroplasty*

When the race is truly lost in the face of recalcitrant infections, several procedures are available as salvage options to maximize the function and health of the patient. The decision to employ one of these options is a result of host factors, namely, poor soft tissue envelope or bone stock, a patient's physical and emotional exhaustion, and a reluctance or inability to continue with therapy.

<span id="page-17-0"></span>

**Fig. 6** (**a**) Lateral radiograph of an infected articulating spacer with an extensor mechanism disruption as demonstrated by the high-riding patella. (**b**) The same patient status post placement of a static spacer, in this case a humeral nail with high dose antibiotic cement. (**c**) Following explant of the static spacer, an intramedullary nail was placed for the knee fusion. (**d**) Due to the patient's poorly controlled diabetes and renal dysfunction requiring dialysis, 2 years later she presented with a reinfected implant though healed fusion. (**e**) The infected nail was removed and after debridement, the healed fusion was able to be retained. (**f**) If bony fusion is unobtainable, intercalary fusion presents another treatment option

## *Knee Arthrodesis and Above the Knee Amputation*

In the setting of an infected TKA, knee arthrodesis (KA) may be employed to provide a stable, painless knee via intramedullary nail or compression plating which may be augmented temporarily by external fixation once all infected TKA components have been removed (Fig. [6c–f\)](#page-17-0). The intramedullary nail may be coated with PMMA impregnated with antibiotics for further infection control. The knee is fused in full extension and therefore requires a rehabilitative period for gait training. Once fused, the patient can ambulate on their native leg and KA requires a lower energy expenditure (0.16 mL  $O_2$ /kg/min vs. 0.20 mL  $O_2$ /kg/min) when compared to an above the knee amputation (AKA) [[101,](#page-27-10) [102](#page-27-11)]. The retention of the native knee, however, does retain a possible nidus for latent infection and reinfection rates of 5.4–10.6% have been reported [[103\]](#page-27-12). Further, a significantly higher rate of postoperative complications has been associated with KA when compared to AKA [[104\]](#page-27-13). Conversely, KA has been associated with improved functional outcomes and lower mortality when compared to AKA [[105,](#page-27-14) [106\]](#page-27-15). Further, AKA has been associated with more systemic complications, longer hospital stays, and higher readmission rates [[107\]](#page-27-16). Barring poor host factors such as severe comorbidities and poor soft tissue envelope or bone stock, KA is the preferred salvage procedure compared to its lower incidence of complications, lower mortality, and superior functional outcomes [\[104](#page-27-13), [108](#page-27-17)].

#### *Girdlestone and Hip Disarticulation*

In the setting of recurrent THA infection, the Girdlestone procedure provides the surgeon with a salvage procedure in which prosthetic components are removed and infection is controlled at the expense of joint functionality (Fig. [7a–d\)](#page-19-0). After implant removal, an osteotomy is performed just above the greater trochanter resecting the femoral neck and head. The hip capsule remains and fibrous tissue fills in creating a pseudarthrosis. No bony fusion occurs, allowing for a limited range of motion [[109\]](#page-27-18). Patient satisfaction varies widely with the procedure (13–83%) with resolution of infection occurring in 80–100% of cases [\[110](#page-27-19)]. Conversely, a hip disarticulation, in which the entire lower extremity is amputated at the level of the hip joint, is considered a morbid procedure and reserved typically for life-threatening scenarios such as systemic sepsis or extreme soft tissue compromise [\[111](#page-28-1)]. In addition to limitations with weight bearing, special postoperative considerations exist such as wheelchair use requiring a special balance of weights to compensate for loss of the entire limb.

#### *Adjunctive Antibiotic Therapy*

Postoperative antibiotics are required regardless of procedure selected by the surgeon. Ideally, a causative organism with sensitivities is identified prior to surgery. Multiple samples are taken at the time of implant removal, and patients are started

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**Fig. 7** (**a**) Clinical picture of a draining wound in a male with a history of multiple hip surgeries and eventual partial hip exchange for MRSA infection. Given his medical comorbidities, it was decided to proceed with a Girdlestone procedure, as any attempt for reimplantation would likely result in repeat infection. (**b**) Radiograph of the patient's prior partial hip resection. The femoral stem and acetabular cup were retained with placement of nonabsorbable antibiotic-impregnated PMMA beads. (**c**) Postoperative radiograph after debridement and removal of all components with placement of absorbable calcium sulfate high-dose vancomycin beads. (**d**) Radiograph 5 months after a Girdlestone procedure demonstrating the greater trochanter articulating with the pelvis. The patient now ambulates with a shoe lift and crutches



**Fig. 7** (continued)

immediately postoperatively on empiric intravenous antimicrobials. Once sensitivities are obtained, drugs are then tailored to the specific organism(s). Multiple studies have demonstrated a success rate of 90–100% with the use of 6 weeks or less of intravenous antibiotics [[112,](#page-28-2) [113\]](#page-28-3) with one study directly examining a 1-week course vs. 6-week and finding no superiority [\[114](#page-28-4)]. Bernard et al. examined the outcomes of 144 patients treated with DAIR, one- and two-stage exchange, Girdlestone, or KA and found no advantage to antibiotic therapy longer than 6 weeks [\[115](#page-28-5)]. To date, no studies published have directly compared exclusively

oral vs. exclusively intravenous antibiotics. However, studies have demonstrated equivalent efficacy of beginning antibiotic therapy parenterally and then transitioning to oral [\[116](#page-28-6)[–118](#page-28-7)]. The Infectious Disease Society of America currently endorses a 4–6-week course of intravenous or highly bioavailable oral antibiotics with a sixweek course endorsed for more virulent organisms.

## *Negative Pressure Wound Therapy (NPWT)*

NPWT or "wound vac" is often applied as an adjunct for wound closure and to contend with postoperative drainage in both primary and revision cases. Studies have demonstrated reduced wound exudate, fewer dressing changes required postoperatively, and a decreased rate of superficial wound infections [[119,](#page-28-8) [120\]](#page-28-9). Given that the rate of infection increases 29% for TKA and 42% for THA for each additional day of postoperative drainage, many surgeons apply NPWT prophylactically [\[121](#page-28-10)].

## **Call to Action**

Despite surgical, technological, and medical advances, the infection rate in total joint arthroplasty has remained largely unchanged for the last 30 years. Some have speculated infection can never truly be prevented and subsequently healthcare providers must remain ever vigilant in the face of this devastating complication. The race for the surface may not be a winning vs. losing proposition but rather is likely a continuous spectrum. The treatment for each patient should be as individualized as the clinical scenario and depends on host factors, microbial isolates, and treatment regimens available.

As bacteria adapt to our arsenal, so too must we continue our efforts against them. There is a critical knowledge gap and growing need for continued treatment evolution in the way of improved osteoinductive/antimicrobial materials at time of a reimplant, in addition to prevention technology. Given the impending rise in total joint arthroplasty case volume and subsequent revision case volume due to PJI, an urgent need exists for continued work in the development of preventive, diagnostic, and therapeutic tools.

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# **References**

- <span id="page-22-0"></span>1. Scott CE, Bugler KE, Clement ND, MacDonald D, Howie CR, Biant LC (2012) Patient expectations of arthroplasty of the hip and knee. J Bone Joint Surg Br 94(7):974–981
- <span id="page-22-1"></span>2. Palazzo C, Jourdan C, Descamps S, Nizard R, Hamadouche M, Anract P, Boisgard S, Galvin M, Ravaud P, Poiraudeau S (2014) Determinants of satisfaction 1 year after total hip arthroplasty: the role of expectations fulfilment. BMC Musculoskelet Disord 15:53
- <span id="page-22-2"></span>3. Sloan M, Premkumar A, Sheth NP (2018) Projected volume of primary total joint arthroplasty in the U.S., 2014 to 2030. J Bone Joint Surg Am 100(17):1455–1460
- <span id="page-22-3"></span>4. Bozic KJ, Ries MD (2005) The impact of infection after total hip arthroplasty on hospital and surgeon resource utilization. J Bone Joint Surg Am 87(8):1746–1751
- <span id="page-22-4"></span>5. Namba RS, Inacio MC, Paxton EW (2013) Risk factors associated with deep surgical site infections after primary total knee arthroplasty: an analysis of 56,216 knees. J Bone Joint Surg Am 95(9):775–782
- 6. Edwards JR, Peterson KD, Mu Y, Banerjee S, Allen-Bridson K, Morrell G, Dudeck MA, Pollock DA, Horan TC (2009) National Healthcare Safety Network (NHSN) report: data summary for 2006 through 2008, issued December 2009. Am J Infect Control 37(10):783–805
- 7. Huotari K, Peltola M, Jamsen E (2015) The incidence of late prosthetic joint infections: a registry-based study of 112,708 primary hip and knee replacements. Acta Orthop 86(3):321–325
- <span id="page-22-5"></span>8. Ong KL, Kurtz SM, Lau E, Bozic KJ, Berry DJ, Parvizi J (2009) Prosthetic joint infection risk after total hip arthroplasty in the Medicare population. J Arthroplast 24(6 Suppl):105–109
- <span id="page-22-6"></span>9. Kapadia BH, McElroy MJ, Issa K, Johnson AJ, Bozic KJ, Mont MA (2014) The economic impact of periprosthetic infections following total knee arthroplasty at a specialized tertiarycare center. J Arthroplast 29(5):929–932
- <span id="page-22-7"></span>10. Kapadia BH, Banerjee S, Cherian JJ, Bozic KJ, Mont MA (2016) The economic impact of periprosthetic infections after total hip arthroplasty at a Specialized Tertiary-Care Center. J Arthroplast 31(7):1422–1426
- <span id="page-22-8"></span>11. Whitehouse JD, Friedman ND, Kirkland KB, Richardson WJ, Sexton DJ (2002) The impact of surgical-site infections following orthopedic surgery at a community hospital and a university hospital: adverse quality of life, excess length of stay, and extra cost. Infect Control Hosp Epidemiol 23(4):183–189
- <span id="page-22-9"></span>12. de Lissovoy G, Fraeman K, Hutchins V, Murphy D, Song D, Vaughn BB (2009) Surgical site infection: incidence and impact on hospital utilization and treatment costs. Am J Infect Control 37(5):387–397
- <span id="page-22-10"></span>13. Berend KR, Lombardi AV Jr, Morris MJ, Bergeson AG, Adams JB, Sneller MA (2013) Twostage treatment of hip periprosthetic joint infection is associated with a high rate of infection control but high mortality. Clin Orthop Relat Res 471(2):510–518
- <span id="page-22-11"></span>14. Choi HR, Beecher B, Bedair H (2013) Mortality after septic versus aseptic revision total hip arthroplasty: a matched-cohort study. J Arthroplast 28(8 Suppl):56–58
- <span id="page-22-12"></span>15. Kurtz S, Ong K, Lau E, Mowat F, Halpern M (2007) Projections of primary and revision hip and knee arthroplasty in the United States from 2005 to 2030. J Bone Joint Surg Am 89(4):780–785
- <span id="page-22-13"></span>16. Tsukayama DT, Goldberg VM, Kyle R (2003) Diagnosis and management of infection after total knee arthroplasty. J Bone Joint Surg Am 85-A(Suppl 1):S75–S80
- 17. Tsukayama DT, Estrada R, Gustilo RB (1996) Infection after total hip arthroplasty. A study of the treatment of one hundred and six infections. J Bone Joint Surg Am 78(4):512–523
- 18. Parvizi J, Gehrke T (2014) Definition of periprosthetic joint infection. J Arthroplast 29(7):1331
- <span id="page-22-14"></span>19. McPherson EJ, Tontz W Jr, Patzakis M, Woodsome C, Holtom P, Norris L, Shufelt C (1999) Outcome of infected total knee utilizing a staging system for prosthetic joint infection. Am J Orthop (Belle Mead NJ) 28(3):161–165
- <span id="page-23-0"></span>20. Li C, Renz N, Trampuz A (2018) Management of Periprosthetic Joint Infection. Hip Pelvis 30(3):138–146
- <span id="page-23-1"></span>21. Luthringer TA, Fillingham YA, Okroj K, Ward EJ, Della Valle C (2016) Periprosthetic joint infection after hip and knee arthroplasty: a review for emergency care providers. Ann Emerg Med 68(3):324–334
- <span id="page-23-2"></span>22. Cook JL, Scott RD, Long WJ (2007) Late hematogenous infections after total knee arthroplasty: experience with 3013 consecutive total knees. J Knee Surg 20(1):27–33
- <span id="page-23-3"></span>23. Costerton W, Veeh R, Shirtliff M, Pasmore M, Post C, Ehrlich G (2003) The application of biofilm science to the study and control of chronic bacterial infections. J Clin Invest 112(10):1466–1477
- <span id="page-23-4"></span>24. Costerton JW (2005) Biofilm theory can guide the treatment of device-related orthopaedic infections. Clin Orthop Relat Res (437):7–11
- <span id="page-23-5"></span>25. Barth E, Myrvik QM, Wagner W, Gristina AG (1989) In vitro and in vivo comparative colonization of Staphylococcus aureus and Staphylococcus epidermidis on orthopaedic implant materials. Biomaterials 10(5):325–328
- <span id="page-23-6"></span>26. Alamanda VK, Springer BD (2019) The prevention of infection: 12 modifiable risk factors. Bone Joint J  $101-b(1$  Supple A):3–9
- <span id="page-23-7"></span>27. Kee JR, Mears SC, Edwards PK, Barnes CL (2017) Modifiable risk factors are common in early revision hip and knee arthroplasty. J Arthroplast 32(12):3689–3692
- <span id="page-23-8"></span>28. Namba RS, Paxton L, Fithian DC, Stone ML (2005) Obesity and perioperative morbidity in total hip and total knee arthroplasty patients. J Arthroplast 20(7 Suppl 3):46–50
- <span id="page-23-9"></span>29. Smith JO, Frampton CMA, Hooper GJ, Young SW (2018) The impact of patient and surgical factors on the rate of postoperative infection after total hip arthroplasty—a New Zealand Joint Registry Study. J Arthroplast 33(6):1884–1890
- <span id="page-23-10"></span>30. Wagner ER, Kamath AF, Fruth KM, Harmsen WS, Berry DJ (2016) Effect of body mass index on complications and reoperations after total hip arthroplasty. J Bone Joint Surg Am 98(3):169–179
- <span id="page-23-11"></span>31. Cancienne JM, Werner BC, Browne JA (2017) Is there a threshold value of hemoglobin A1c that predicts risk of infection following primary total hip arthroplasty? J Arthroplast 32(9s):S236–s240
- <span id="page-23-12"></span>32. Tarabichi M, Shohat N, Kheir MM, Adelani M, Brigati D, Kearns SM, Patel P, Clohisy JC, Higuera CA, Levine BR, Schwarzkopf R, Parvizi J, Jiranek WA (2017) Determining the threshold for HbA1c as a predictor for adverse outcomes after total joint arthroplasty: a multicenter, retrospective study. J Arthroplasty 32(9s):S263–S267.e1
- <span id="page-23-13"></span>33. Bedard NA, DeMik DE, Owens JM, Glass NA, DeBerg J, Callaghan JJ (2019) Tobacco use and risk of wound complications and periprosthetic joint infection: a systematic review and meta-analysis of primary total joint arthroplasty procedures. J Arthroplast 34(2):385–396.e4
- <span id="page-23-14"></span>34. Bozic KJ, Lau E, Kurtz S, Ong K, Rubash H, Vail TP, Berry DJ (2012) Patient-related risk factors for periprosthetic joint infection and postoperative mortality following total hip arthroplasty in Medicare patients. J Bone Joint Surg Am 94(9):794–800
- <span id="page-23-18"></span>35. Bozic KJ, Lau E, Kurtz S, Ong K, Berry DJ (2012) Patient-related risk factors for postoperative mortality and periprosthetic joint infection in medicare patients undergoing TKA. Clin Orthop Relat Res 470(1):130–137
- <span id="page-23-15"></span>36. Poultsides LA, Ma Y, Della Valle AG, Chiu YL, Sculco TP, Memtsoudis SG (2013) In-hospital surgical site infections after primary hip and knee arthroplasty—incidence and risk factors. J Arthroplast 28(3):385–389
- <span id="page-23-16"></span>37. Cordtz RL, Zobbe K, Hojgaard P, Kristensen LE, Overgaard S, Odgaard A, Lindegaard H, Dreyer L (2018) Predictors of revision, prosthetic joint infection and mortality following total hip or total knee arthroplasty in patients with rheumatoid arthritis: a nationwide cohort study using Danish healthcare registers. Ann Rheum Dis 77(2):281–288
- <span id="page-23-17"></span>38. Goodman SM, Springer B, Guyatt G, Abdel MP, Dasa V, George M, Gewurz-Singer O, Giles JT, Johnson B, Lee S, Mandl LA, Mont MA, Sculco P, Sporer S, Stryker L, Turgunbaev M, Brause B, Chen AF, Gililland J, Goodman M, Hurley-Rosenblatt A, Kirou K, Losina E, MacKenzie R, Michaud K, Mikuls T, Russell L, Sah A, Miller AS, Singh JA, Yates A

(2017) 2017 American College of Rheumatology/American Association of Hip and Knee Surgeons guideline for the perioperative management of antirheumatic medication in patients with rheumatic diseases undergoing elective total hip or total knee arthroplasty. Arthritis Rheumatol 69(8):1538–1551

- 39. Momohara S, Kawakami K, Iwamoto T, Yano K, Sakuma Y, Hiroshima R, Imamura H, Masuda I, Tokita A, Ikari K (2011) Prosthetic joint infection after total hip or knee arthroplasty in rheumatoid arthritis patients treated with nonbiologic and biologic disease-modifying antirheumatic drugs. Mod Rheumatol 21(5):469–475
- <span id="page-24-0"></span>40. Suzuki M, Nishida K, Soen S, Oda H, Inoue H, Kaneko A, Takagishi K, Tanaka T, Matsubara T, Mitsugi N, Mochida Y, Momohara S, Mori T, Suguro T (2011) Risk of postoperative complications in rheumatoid arthritis relevant to treatment with biologic agents: a report from the Committee on Arthritis of the Japanese Orthopaedic Association. J Orthop Sci 16(6):778–784
- <span id="page-24-1"></span>41. Pruzansky JS, Bronson MJ, Grelsamer RP, Strauss E, Moucha CS (2014) Prevalence of modifiable surgical site infection risk factors in hip and knee joint arthroplasty patients at an urban academic hospital. J Arthroplast 29(2):272–276
- <span id="page-24-2"></span>42. Swenson RD, Butterfield JA, Irwin TJ, Zurlo JJ, Davis CM 3rd (2018) Preoperative anemia is associated with failure of open debridement polyethylene exchange in acute and acute hematogenous prosthetic joint infection. J Arthroplast 33(6):1855–1860
- <span id="page-24-3"></span>43. Pugely AJ, Martin CT, Gao Y, Schweizer ML, Callaghan JJ (2015) The incidence of and risk factors for 30-day surgical site infections following primary and revision total joint arthroplasty. J Arthroplast 30(9 Suppl):47–50
- 44. Bongartz T, Halligan CS, Osmon DR, Reinalda MS, Bamlet WR, Crowson CS, Hanssen AD, Matteson EL (2008) Incidence and risk factors of prosthetic joint infection after total hip or knee replacement in patients with rheumatoid arthritis. Arthritis Rheum 59(12):1713–1720
- <span id="page-24-4"></span>45. Bedair H, Goyal N, Dietz MJ, Urish K, Hansen V, Manrique J, Hamilton W, Deirmengian G (2015) A history of treated Periprosthetic joint infection increases the risk of subsequent different site infection. Clin Orthop Relat Res 473(7):2300–2304
- <span id="page-24-5"></span>46. Kunutsor SK, Whitehouse MR, Blom AW, Beswick AD (2016) Patient-related risk factors for periprosthetic joint infection after total joint arthroplasty: a systematic review and metaanalysis. PLoS One 11(3):e0150866
- <span id="page-24-6"></span>47. Prosthetic Joint Infection risk calculator. [https://icmphilly.com/ortho-applications/](https://icmphilly.com/ortho-applications/prosthetic-joint-infection-pji-risk-calculator/) [prosthetic-joint-infection-pji-risk-calculator/](https://icmphilly.com/ortho-applications/prosthetic-joint-infection-pji-risk-calculator/)
- <span id="page-24-7"></span>48. Amanatullah D, Dennis D, Oltra EG, Marcelino Gomes LS, Goodman SB, Hamlin B, Hansen E, Hashemi-Nejad A, Holst DC, Komnos G, Koutalos A, Malizos K, Martinez Pastor JC, McPherson E, Meermans G, Mooney JA, Mortazavi J, Parsa A, Pecora JR, Pereira GA, Martos MS, Shohat N, Shope AJ, Zullo SS (2019) Hip and knee section, diagnosis, definitions: proceedings of international consensus on orthopedic infections. J Arthroplasty 34(2s):S329–s337
- <span id="page-24-8"></span>49. Ghosh S, Charity RM, Haidar SG, Singh BK (2006) Pyrexia following total knee replacement. Knee 13(4):324–327
- <span id="page-24-9"></span>50. Zajonz D, Wuthe L, Tiepolt S, Brandmeier P, Prietzel T, von Salis-Soglio GF, Roth A, Josten C, Heyde CE, Ghanem M (2015) Diagnostic work-up strategy for periprosthetic joint infections after total hip and knee arthroplasty: a 12-year experience on 320 consecutive cases. Patient Saf Surg 9:20
- <span id="page-24-10"></span>51. Tigges S, Stiles RG, Roberson JR (1994) Appearance of septic hip prostheses on plain radiographs. AJR Am J Roentgenol 163(2):377–380
- <span id="page-24-11"></span>52. Verberne SJ, Sonnega RJ, Temmerman OP, Raijmakers PG (2017) What is the accuracy of nuclear imaging in the assessment of periprosthetic knee infection? A meta-analysis. Clin Orthop Relat Res 475(5):1395–1410
- <span id="page-24-12"></span>53. Sofka CM (2007) Current applications of advanced cross-sectional imaging techniques in evaluating the painful arthroplasty. Skelet Radiol 36(3):183–193
- <span id="page-24-13"></span>54. Della Valle C, Parvizi J, Bauer TW, Dicesare PE, Evans RP, Segreti J, Spangehl M, Watters WC 3rd, Keith M, Turkelson CM, Wies JL, Sluka P, Hitchcock K (2010) Diagnosis of periprosthetic joint infections of the hip and knee. J Am Acad Orthop Surg 18(12):760–770
- <span id="page-25-1"></span>55. Spangehl MJ, Masri BA, O'Connell JX, Duncan CP (1999) Prospective analysis of preoperative and intraoperative investigations for the diagnosis of infection at the sites of two hundred and two revision total hip arthroplasties. J Bone Joint Surg Am 81(5):672–683
- <span id="page-25-2"></span>56. Berbari E, Mabry T, Tsaras G, Spangehl M, Erwin PJ, Murad MH, Steckelberg J, Osmon D (2010) Inflammatory blood laboratory levels as markers of prosthetic joint infection: a systematic review and meta-analysis. J Bone Joint Surg Am 92(11):2102–2109
- <span id="page-25-3"></span>57. Larsson S, Thelander U, Friberg S (1992) C-reactive protein (CRP) levels after elective orthopedic surgery. Clin Orthop Relat Res (275):237-42
- <span id="page-25-4"></span>58. Nazem K, Motififard M, Yousefian M (2016) Variations in ESR and CRP in total knee arthroplasty and total hip arthroplasty in Iranian patients from 2009 to 2011. Adv Biomed Res 5:148
- <span id="page-25-5"></span>59. Shahi A, Kheir MM, Tarabichi M, Hosseinzadeh HRS, Tan TL, Parvizi J (2017) Serum D-Dimer test is promising for the diagnosis of periprosthetic joint infection and timing of reimplantation. J Bone Joint Surg Am 99(17):1419–1427
- <span id="page-25-6"></span>60. Stone WZ, Gray CF, Parvataneni HK, Prieto HA (2019) Clinical evaluation of alpha defensin test following staged treatment of prosthetic joint infections. J Arthroplast 34:1446
- 61. Carli AV, Abdelbary H, Ahmadzai N, Cheng W, Shea B, Hutton B, Sniderman J, Philip Sanders BS, Esmaeilisaraji L, Skidmore B, Gauthier-Kwan OY, Bunting AC, Gauthier P, Crnic A, Logishetty K, Moher D, Fergusson D, Beaule PE (2019) Diagnostic accuracy of serum, synovial, and tissue testing for chronic periprosthetic joint infection after hip and knee replacements: a systematic review. J Bone Joint Surg Am 101(7):635–649
- <span id="page-25-7"></span>62. Lee YS, Koo KH, Kim HJ, Tian S, Kim TY, Maltenfort MG, Chen AF (2017) Synovial fluid biomarkers for the diagnosis of periprosthetic joint infection: a systematic review and metaanalysis. J Bone Joint Surg Am 99(24):2077–2084
- <span id="page-25-8"></span>63. Yoon HK, Cho SH, Lee DY, Kang BH, Lee SH, Moon DG, Kim DH, Nam DC, Hwang SC (2017) A review of the literature on culture-negative periprosthetic joint infection: epidemiology, diagnosis and treatment. Knee Surg Relat Res 29(3):155–164
- <span id="page-25-9"></span>64. Tande AJ, Patel R (2014) Prosthetic joint infection. Clin Microbiol Rev 27(2):302–345
- <span id="page-25-10"></span>65. Berbari EF, Marculescu C, Sia I, Lahr BD, Hanssen AD, Steckelberg JM, Gullerud R, Osmon DR (2007) Culture-negative prosthetic joint infection. Clin Infect Dis 45(9):1113–1119
- <span id="page-25-11"></span>66. Phelan DM, Osmon DR, Keating MR, Hanssen AD (2002) Delayed reimplantation arthroplasty for candidal prosthetic joint infection: a report of 4 cases and review of the literature. Clin Infect Dis 34(7):930–938
- <span id="page-25-12"></span>67. Bosshard PP (2011) Incubation of fungal cultures: how long is long enough? Mycoses 54(5):e539–e545
- <span id="page-25-13"></span>68. Tarabichi M, Shohat N, Goswami K, Alvand A, Silibovsky R, Belden K, Parvizi J (2018) Diagnosis of periprosthetic joint infection: the potential of next-generation sequencing. J Bone Joint Surg Am 100(2):147–154
- <span id="page-25-14"></span>69. Ivy MI, Thoendel MJ, Jeraldo PR, Greenwood-Quaintance KE, Hanssen AD, Abdel MP, Chia N, Yao JZ, Tande AJ, Mandrekar JN, Patel R (2018) Direct detection and identification of prosthetic joint infection pathogens in synovial fluid by metagenomic shotgun sequencing. J Clin Microbiol 56(9)
- <span id="page-25-0"></span>70. Shohat N, Bauer T, Buttaro M, Budhiparama N, Cashman J, Della Valle CJ, Drago L, Gehrke T, Marcelino Gomes LS, Goswami K, Hailer NP, Han SB, Higuera CA, Inaba Y, Jenny JY, Kjaersgaard-Andersen P, Lee M, Llinas A, Malizos K, Mont MA, Jones RM, Parvizi J, Peel T, Rivero-Boschert S, Segreti J, Soriano A, Sousa R, Spangehl M, Tan TL, Tikhilov R, Tuncay I, Winkler H, Witso E, Wouthuyzen-Bakker M, Young S, Zhang X, Zhou Y, Zimmerli W (2019) Hip and knee section, what is the definition of a periprosthetic joint infection (PJI) of the knee and the hip? Can the same criteria be used for both joints? Proceedings of International Consensus on Orthopedic Infections. J Arthroplast 34(2s):S325–s327
- <span id="page-25-15"></span>71. Cochran AR, Ong KL, Lau E, Mont MA, Malkani AL (2016) Risk of reinfection after treatment of infected total knee arthroplasty. J Arthroplast 31(9 Suppl):156–161
- <span id="page-25-16"></span>72. Goulet JA, Pellicci PM, Brause BD, Salvati EM (1988) Prolonged suppression of infection in total hip arthroplasty. J Arthroplast 3(2):109–116
- 73. Rao N, Crossett LS, Sinha RK, Le Frock JL (2003) Long-term suppression of infection in total joint arthroplasty. Clin Orthop Relat Res 414:55–60
- <span id="page-26-0"></span>74. Siqueira MB, Saleh A, Klika AK, O'Rourke C, Schmitt S, Higuera CA, Barsoum WK (2015) Chronic suppression of periprosthetic joint infections with oral antibiotics increases infection-free survivorship. J Bone Joint Surg Am 97(15):1220–1232
- <span id="page-26-1"></span>75. Haasper C, Buttaro M, Hozack W, Aboltins CA, Borens O, Callaghan JJ, de Carvalho PI, Chang Y, Corona P, Da Rin F, Esposito S, Fehring TK, Sanchez XF, Lee GC, Martinez-Pastor JC, Mortazavi SM, Noiseux NO, Peng KT, Schutte HD, Schweitzer D, Trebse R, Tsiridis E, Whiteside L (2014) Irrigation and debridement. J Arthroplast 29(2 Suppl):100–103
- <span id="page-26-2"></span>76. Wouthuyzen-Bakker M, Sebillotte M, Lomas J, Taylor A, Palomares EB, Murillo O, Parvizi J, Shohat N, Reinoso JC, Sanchez RE, Fernandez-Sampedro M, Senneville E, Huotari K, Barbero JM, Garcia-Canete J, Lora-Tamayo J, Ferrari MC, Vaznaisiene D, Yusuf E, Aboltins C, Trebse R, Salles MJ, Benito N, Vila A, Toro MDD, Kramer TS, Petersdorf S, Diaz-Brito V, Tufan ZK, Sanchez M, Arvieux C, Soriano A (2019) Clinical outcome and risk factors for failure in late acute prosthetic joint infections treated with debridement and implant retention. J Infect 78(1):40–47
- <span id="page-26-3"></span>77. Koyonos L, Zmistowski B, Della Valle CJ, Parvizi J (2011) Infection control rate of irrigation and debridement for periprosthetic joint infection. Clin Orthop Relat Res 469(11):3043–3048
- 78. Fehring TK, Odum SM, Berend KR, Jiranek WA, Parvizi J, Bozic KJ, Della Valle CJ, Gioe TJ (2013) Failure of irrigation and debridement for early postoperative periprosthetic infection. Clin Orthop Relat Res 471(1):250–257
- <span id="page-26-4"></span>79. Odum SM, Fehring TK, Lombardi AV, Zmistowski BM, Brown NM, Luna JT, Fehring KA, Hansen EN (2011) Irrigation and debridement for periprosthetic infections: does the organism matter? J Arthroplast 26(6 Suppl):114–118
- <span id="page-26-5"></span>80. Tsang SJ, Ting J, Simpson A, Gaston P (2017) Outcomes following debridement, antibiotics and implant retention in the management of periprosthetic infections of the hip: a review of cohort studies. Bone Joint J 99-b(11):1458–1466
- <span id="page-26-6"></span>81. Volpin A, Sukeik M, Alazzawi S, Haddad FS (2016) Aggressive early debridement in treatment of acute periprosthetic joint infections after hip and knee replacements. Open Orthop J 10:669–678
- <span id="page-26-7"></span>82. Buchholz HW, Elson RA, Engelbrecht E, Lodenkamper H, Rottger J, Siegel A (1981) Management of deep infection of total hip replacement. J Bone Joint Surg Br 63-b(3):342–353
- <span id="page-26-8"></span>83. Jackson WO, Schmalzried TP (2000) Limited role of direct exchange arthroplasty in the treatment of infected total hip replacements. Clin Orthop Relat Res 381:101–105
- <span id="page-26-9"></span>84. Lange J, Troelsen A, Solgaard S, Otte KS, Jensen NK, Soballe K (2018) Cementless onestage revision in chronic periprosthetic hip joint infection. Ninety-one percent infection free survival in 56 patients at minimum 2-year follow-up. J Arthroplast 33(4):1160–1165.e1
- 85. Bori G, Navarro G, Morata L, Fernandez-Valencia JA, Soriano A, Gallart X (2018) Preliminary results after changing from two-stage to one-stage revision arthroplasty protocol using cementless arthroplasty for chronic infected hip replacements. J Arthroplast 33(2):527–532
- 86. Wolf M, Clar H, Friesenbichler J, Schwantzer G, Bernhardt G, Gruber G, Glehr M, Leithner A, Sadoghi P (2014) Prosthetic joint infection following total hip replacement: results of onestage versus two-stage exchange. Int Orthop 38(7):1363–1368
- 87. Jenny JY, Lengert R, Diesinger Y, Gaudias J, Boeri C, Kempf JF (2014) Routine one-stage exchange for chronic infection after total hip replacement. Int Orthop 38(12):2477–2481
- <span id="page-26-10"></span>88. Raut VV, Siney PD, Wroblewski BM (1994) One-stage revision of infected total hip replacements with discharging sinuses. J Bone Joint Surg Br 76(5):721–724
- <span id="page-26-11"></span>89. Leonard HA, Liddle AD, Burke O, Murray DW, Pandit H (2014) Single- or two-stage revision for infected total hip arthroplasty? A systematic review of the literature. Clin Orthop Relat Res 472(3):1036–1042
- 90. George DA, Logoluso N, Castellini G, Gianola S, Scarponi S, Haddad FS, Drago L, Romano CL (2016) Does cemented or cementless single-stage exchange arthroplasty of chronic periprosthetic hip infections provide similar infection rates to a two-stage? A systematic review. BMC Infect Dis 16(1):553
- <span id="page-27-0"></span>91. Haddad FS, Sukeik M, Alazzawi S (2015) Is single-stage revision according to a strict protocol effective in treatment of chronic knee arthroplasty infections? Clin Orthop Relat Res 473(1):8–14
- <span id="page-27-1"></span>92. Romano CL, Gala L, Logoluso N, Romano D, Drago L (2012) Two-stage revision of septic knee prosthesis with articulating knee spacers yields better infection eradication rate than one-stage or two-stage revision with static spacers. Knee Surg Sports Traumatol Arthrosc 20(12):2445–2453
- <span id="page-27-2"></span>93. Vaishya R, Chauhan M, Vaish A (2013) Bone cement. J Clin Orthop Trauma 4(4):157–163
- <span id="page-27-3"></span>94. Carli AV, Sethuraman AS, Bhimani SJ, Ross FP, Bostrom MPG (2018) Selected heatsensitive antibiotics are not inactivated during polymethylmethacrylate curing and can be used in cement spacers for periprosthetic joint infection. J Arthroplast 33(6):1930–1935
- <span id="page-27-4"></span>95. Samara E, Moriarty TF, Decosterd LA, Richards RG, Gautier E, Wahl P (2017) Antibiotic stability over six weeks in aqueous solution at body temperature with and without heat treatment that mimics the curing of bone cement. Bone Joint Res 6(5):296–306
- <span id="page-27-5"></span>96. Young SW, Zhang M, Freeman JT, Mutu-Grigg J, Pavlou P, Moore GA (2014) The Mark Coventry Award: higher tissue concentrations of vancomycin with low-dose intraosseous regional versus systemic prophylaxis in TKA: a randomized trial. Clin Orthop Relat Res 472(1):57–65
- <span id="page-27-6"></span>97. Roy ME, Peppers MP, Whiteside LA, Lazear RM (2014) Vancomycin concentration in synovial fluid: direct injection into the knee vs. intravenous infusion. J Arthroplast 29(3):564–568
- <span id="page-27-7"></span>98. Toulson C, Walcott-Sapp S, Hur J, Salvati E, Bostrom M, Brause B, Westrich GH (2009) Treatment of infected total hip arthroplasty with a 2-stage reimplantation protocol: update on "our institution's" experience from 1989 to 2003. J Arthroplasty  $24(7)$ :1051–1060
- <span id="page-27-8"></span>99. Browne JA, Cancienne JM, Novicoff WM, Werner BC (2017) Removal of an infected hip arthroplasty is a high-risk surgery: putting morbidity into context with other major nonorthopedic operations. J Arthroplast 32(9):2834–2841
- <span id="page-27-9"></span>100. Lum ZC, Natsuhara KM, Shelton TJ, Giordani M, Pereira GC, Meehan JP (2018) Mortality during total knee periprosthetic joint infection. J Arthroplast 33(12):3783–3788
- <span id="page-27-10"></span>101. Conway JD, Mont MA, Bezwada HP (2004) Arthrodesis of the knee. J Bone Joint Surg Am 86-a(4):835–848
- <span id="page-27-11"></span>102. Waters RL, Perry J, Antonelli D, Hislop H (1976) Energy cost of walking of amputees: the influence of level of amputation. J Bone Joint Surg Am 58(1):42–46
- <span id="page-27-12"></span>103. Balato G, Rizzo M, Ascione T, Smeraglia F, Mariconda M (2018) Re-infection rates and clinical outcomes following arthrodesis with intramedullary nail and external fixator for infected knee prosthesis: a systematic review and meta-analysis. BMC Musculoskelet Disord 19(1):361
- <span id="page-27-13"></span>104. Chen AF, Kinback NC, Heyl AE, McClain EJ, Klatt BA (2012) Better function for fusions versus above-the-knee amputations for recurrent periprosthetic knee infection. Clin Orthop Relat Res 470(10):2737–2745
- <span id="page-27-14"></span>105. Wu CH, Gray CF, Lee GC (2014) Arthrodesis should be strongly considered after failed twostage reimplantation TKA. Clin Orthop Relat Res 472(11):3295–3304
- <span id="page-27-15"></span>106. Son MS, Lau E, Parvizi J, Mont MA, Bozic KJ, Kurtz S (2017) What are the frequency, associated factors, and mortality of amputation and arthrodesis after a failed infected TKA? Clin Orthop Relat Res 475(12):2905–2913
- <span id="page-27-16"></span>107. Carr JB 2nd, Werner BC, Browne JA (2016) Trends and outcomes in the treatment of failed septic total knee arthroplasty: comparing arthrodesis and above-knee amputation. J Arthroplast 31(7):1574–1577
- <span id="page-27-17"></span>108. Khanna V, Tushinski DM, Soever LJ, Vincent AD, Backstein DJ (2015) Above knee amputation following total knee arthroplasty: when enough is enough. J Arthroplast 30(4):658–662
- <span id="page-27-18"></span>109. Castellanos J, Flores X, Llusa M, Chiriboga C, Navarro A (1998) The Girdlestone pseudarthrosis in the treatment of infected hip replacements. Int Orthop 22(3):178–181
- <span id="page-27-19"></span>110. Cordero-Ampuero J (2012) Girdlestone procedure: when and why. Hip Int 22(Suppl 8):S36–S39
- <span id="page-28-1"></span>111. Zalavras CG, Rigopoulos N, Ahlmann E, Patzakis MJ (2009) Hip disarticulation for severe lower extremity infections. Clin Orthop Relat Res 467(7):1721–1726
- <span id="page-28-2"></span>112. Mittal Y, Fehring TK, Hanssen A, Marculescu C, Odum SM, Osmon D (2007) Two-stage reimplantation for periprosthetic knee infection involving resistant organisms. J Bone Joint Surg Am 89(6):1227–1231
- <span id="page-28-3"></span>113. McKenna PB, O'Shea K, Masterson EL (2009) Two-stage revision of infected hip arthroplasty using a shortened post-operative course of antibiotics. Arch Orthop Trauma Surg 129(4):489–494
- <span id="page-28-4"></span>114. Hsieh PH, Huang KC, Lee PC, Lee MS (2009) Two-stage revision of infected hip arthroplasty using an antibiotic-loaded spacer: retrospective comparison between short-term and prolonged antibiotic therapy. J Antimicrob Chemother 64(2):392–397
- <span id="page-28-5"></span>115. Bernard L, Legout L, Zurcher-Pfund L, Stern R, Rohner P, Peter R, Assal M, Lew D, Hoffmeyer P, Uckay I (2010) Six weeks of antibiotic treatment is sufficient following surgery for septic arthroplasty. J Infect 61(2):125–132
- <span id="page-28-6"></span>116. Bassetti M, Cadeo B, Villa G, Sartor A, Cainero V, Causero A (2014) Current antibiotic management of prosthetic joint infections in Italy: the 'Udine strategy'. J Antimicrob Chemother 69(Suppl 1):i41–i45
- 117. Darley ES, Bannister GC, Blom AW, Macgowan AP, Jacobson SK, Alfouzan W (2011) Role of early intravenous to oral antibiotic switch therapy in the management of prosthetic hip infection treated with one- or two-stage replacement. J Antimicrob Chemother 66(10):2405–2408
- <span id="page-28-7"></span>118. Farhad R, Roger PM, Albert C, Pelligri C, Touati C, Dellamonica P, Trojani C, Boileau P (2010) Six weeks antibiotic therapy for all bone infections: results of a cohort study. Eur J Clin Microbiol Infect Dis 29(2):217–222
- <span id="page-28-8"></span>119. Redfern RE, Cameron-Ruetz C, O'Drobinak SK, Chen JT, Beer KJ (2017) Closed incision negative pressure therapy effects on postoperative infection and surgical site complication after total hip and knee arthroplasty. J Arthroplast 32(11):3333–3339
- <span id="page-28-9"></span>120. Karlakki SL, Hamad AK, Whittall C, Graham NM, Banerjee RD, Kuiper JH (2016) Incisional negative pressure wound therapy dressings (iNPWTd) in routine primary hip and knee arthroplasties: a randomised controlled trial. Bone Joint Res 5(8):328–337
- <span id="page-28-10"></span>121. Patel VP, Walsh M, Sehgal B, Preston C, DeWal H, Di Cesare PE (2007) Factors associated with prolonged wound drainage after primary total hip and knee arthroplasty. J Bone Joint Surg Am 89(1):33–38
- <span id="page-28-0"></span>122. Cizmic Z, Feng JE, Huang R, Iorio R, Komnos G, Kunutsor SK, Metwaly RG, Saleh UH, Sheth N, Sloan M (2019) Hip and knee section, prevention, host related: proceedings of international consensus on orthopedic infections. J Arthroplasty 34(2s):S255–s270