

Biology of Conventional Chondrosarcoma

35

Maria Serena Benassi

Conventional chondrosarcoma (CS) (80–85% of all CS) includes central (85–90%) and peripheral (10–15%) CS subtypes that are characterized by complex karyotype and genetic instability. The presence in their benign precursors of specific gene mutations is considered an early event in tumorigenesis [1], and secondary molecular changes are required for the malignant transformation. Currently, the most clinical reliable prognostic factors are age and grade.

Central CS can arises from or enchondromatosis (up to 40%) in patients with somatic mutations in the isocitrate dehydrogenase genes, IDH1 (R132C and R132H) and IDH2 (R172S) [2].

Wild-type IDH1 and IDH2 are important metabolic enzymes involved in lipid metabolism and in Krebs cycle [3]. Heterozygous somatic IDH1/ IDH2 mutations, also seen in many other tumours, cause an abnormal production of the potential oncometabolite D-2-hydroxyglutarate (D-2HG) [4, 5] that leads to genome-wide alteration in DNA methylation, thus supporting the causal role for IDHR132H in driving epigenetic instability [6, 7].

Elevated D-2HG concentration has been detected in acute myeloid leukemia and glioma patients with mutant IDH, confirming the role of this potential biomarker for both diagnosis and therapy [8-10]. An antibody against IDH R132H

is currently widely used for the differential diagnosis of glioma.

Peripheral CS arises from osteochondroma (15-20% of cases) or multiple osteochondroma (up to 1-5% of cases) that have genetic abnormalities in EXT1 or EXT2 oncosuppressor genes [11, 12], although some data demonstrate that the pathogenesis of secondary peripheral CS may be independent of EXT mutations [13].

EXT1 and EXT2 are located on the chromosome bands 8q24 and 11p11–12 respectively and the loss of their activity impairs the heparan sulfate (HS) biosynthesis, essential for the diffusion of hedgehog proteins involved in chondrocyte differentiation. A disturbance of Indian Hedgehog signaling pathway (IHH) breaks the negative feedback loop with parathyroid hormone-related protein (PTHrP), resulting in an unbalance between chondrocyte proliferation and differentiation [14].

Reactivation of PTHrP signaling and antiapoptotic Bcl2 protein overexpression promote the progression toward low-grade and high-grade CS, progressively acquiring p53 mutations, defects in the most important cellular signaling pathways and environment structural changes [14].

Conventional CSs are drug-resistant tumours and surgery remains the primary treatment. However, the knowledge of additional genetic or epigenetic changes during malignant progression led to the identification of potential targets that may be useful for planning new

M. S. Benassi (🖂)

Laboratory of Experimental Oncology, IRCCS Istituto Ortopedico Rizzoli, Bologna, Italy

[©] Springer Nature Switzerland AG 2020

P. Picci et al. (eds.), *Diagnosis of Musculoskeletal Tumors and Tumor-like Conditions*, https://doi.org/10.1007/978-3-030-29676-6_35

adjuvant treatments [14]. In conventional CS, cytogenetic studies showed heterogeneity with respect to karyotype complexity. A positive relation between histological grade and degree of karyotype complexity was found associated with high cellular density, chondroid matrix destruction and vascularization. Chromosome 9p21 and 12q13–15 aberrations resulted in loss of CDKN2A tumour suppressor gene activity and amplification of CDK4 that inhibits pRb activity. P53 is inactivated, while overexpression of prosteoglandin COX2, metalloproteinases MMPs, and pro-hypoxia inducible factor HIF, make them candidate targets for therapeutic approaches [15].

While IDH and EXT mutations are no longer essential for tumour growth, representing the initial phase of malignant progression, the activation of key endpoints controlling tumour cell growth and survival differentiates high-grade CS. The evidence that altered IHH and kinasedependent signaling pathways drive the increase of malignancy has addressed the preclinical and clinical studies to the use of kinase inhibitors or HH antagonists [16–18].

HH, Bcl2, PDGFR, Src, P13K/Akt pathways are considered candidate targets in conventional CS, but the clinical results are not quite satisfactory [19]. Interestingly, Bcl2-dependent drugresistance of CS cells could be overcome by the treatment with Src kinase inhibitors and doxorubicin in p53-negative cells [20]. A phase II clinical trial using dasatinib observed a prolonged stable disease in more than 10% of patients, suggesting that further evaluations should be considered in future clinical trials [21].

In contrast, a previous study [22] failed in terms of objective response and disease-free survival by using imatinib mesylate, an anti-PDGFR agent.

Currently, phase I phase II clinical studies with agents targeting IDH mutations, PI3K-AktmTOR pathway and angiogenesisis are under investigation with the aim to improve the CS patient survival [23].

References

- Szuhai K, Cleton-Jansen AM, Hogendoorn PC, Bovée JV. Molecular pathology and its diagnostic use in bone tumors. Cancer Genet. 2012;205:193–204.
- Pansuriya TC, van Eijk R, d'Adamo P, van Ruler MA, Kuijjer ML, Oosting J, Cleton-Jansen AM, van Oosterwijk JG, Verbeke SL, Meijer D, et al. Somatic mosaic IDH1 and IDH2 mutations are associated with enchondroma and spindle cell hemangioma in Ollier disease and Maffucci syndrome. Nat Genet. 2011;43:1256–61.
- Waitkus MS, Diplas BH, Yan H. Biological role and therapeutic potential of IDH mutations in cancer. Cancer Cell. 2018;34(2):186–95. https://doi. org/10.1016/j.ccell.2018.04.011.
- Amary MF, Bacsi K, Maggiani F, Damato S, Halai D, Berisha F, Pollock R, O'Donnell P, Grigoriadis A, Diss T, Eskandarpour M, Presneau N, Hogendoorn PC, Futreal A, Tirabosco R, Flanagan AM. IDH1 and IDH2 mutations are frequent events in central chondrosarcoma and central and periosteal chondromasi but not in other mesenchymal tumours. J Pathol. 2011;224:334–43.
- Kato Kaneko M, Liu X, Oki H, Ogasawara S, Nakamura T, Saidoh N, Tsujimoto Y, Matsuyama Y, Uruno A, Sugawara M, Tsuchiya T, Yamakawa M, Yamamoto M, et al. Isocitrate dehydrogenase mutation is frequently observed in giant cell tumor of bone. Cancer Sci. 2014;105:744–8.
- Duncan CG, Barwick BG, Jin G, Rago C, Kapoor-Vazirani P, Powell DR, Jen-Tsan Chi J-T, Bigner DD, Vertino PM, Yan H. A heterozygous IDH1R132H/WT mutation induces genome-wide alterations in DNA methylation. Genome Res. 2012;22:2339–55.
- Turcan S, Rohle D, Goenka A, Walsh LA, Fang F, Yilmaz E, Campos C, Fabius AW, Lu C, Ward PS, Thompson CB, Kaufman A, Guryanova O, Levine R, Heguy A, Viale A, Morris LG, Huse JT, Mellinghoff IK, Chan TA. IDH1 mutation is sufficient to establish the glioma hypermethylator phenotype. Nature. 2012;483:479–83.
- Andronesi OC, Rapalino O, Gerstner E, Chi A, Batchelor TT, Cahill DP, Sorensen AG, Rosen BR. Detection of oncogenic IDH1 mutations using magnetic resonance spectroscopy of 2-hydroxyglutarate. J Clin Invest. 2013;123:3659–63.
- Dinardo CD, Propert KJ, Loren AW, Paietta E, Sun Z, Levine RL, Straley KS, Yen K, Patel JP, Agresta S, et al. Serum 2-hydroxyglutarate levels predict isocitrate dehydrogenase mutations and clinical outcome in acute myeloid leukemia. Blood. 2013;121:4917–24.
- Stein EM, DiNardo CD, Pollyea DA, Fathi AT, Roboz GJ, Altman JK, Stone RM, DeAngelo DJ, Levine RL, Flinn IW, et al. Enasidenib in mutant IDH2 relapsed or refractory acute myeloid leukemia. Blood. 2017;130:722–31.

- Szuhai K, Jennes I, de Jong D, Bovée JV, Wiweger M, Wuyts W, Hogendoorn PC. Tiling resolution array-CGH shows that somatic mosaic deletion of the EXT gene is causative in EXT gene mutation negative multiple osteochondromas patients. Hum Mutat. 2011;32:E2036–49.
- Reijnders CM, Waaijer CJ, Hamilton A, Buddingh EP, Dijkstra SP, Ham J, Bakker E, Szuhai K, Karperien M, Hogendoorn PC, Stringer SE, Bovée JV. No haploinsufficiency but loss of heterozygosity for EXT in multiple osteochondromas. Am J Pathol. 2010;177:1946–57.
- De Andrea CE, Reijnders CM, Kroon HM, et al. Secondary peripheral chondrosarcoma evolving from osteochondroma as a result of outgrowth of cells with functional EXT. Oncogene. 2012;31:1095.
- Bovée JV, Hogendoorn PC, Wunder JS, Alman BA. Cartilage tumours and bone development: molecular pathology and possible therapeutic targets. Nat Rev Cancer. 2010;10:481–8.
- Bovée JV, Cleton-Jansen AM, Taminiau AH, Hogendoorn PC. Emerging pathways in the development of chondrosarcoma of bone and implications for targeted treatment. Lancet Oncol. 2005;6:599–607.
- Tiet TD, Hopyan S, Nadesan P, Gokgoz N, Poon R, Lin AC, Yan T, Andrulis IL, Alman BA, Wunder JS. Constitutive hedgehog signaling in chondrosarcoma up-regulates tumor cell proliferation. Am J Pathol. 2006;168:321–30.
- Schrage YM, Briaire-de Bruijn IH, de Miranda NFCC, et al. Kinome profiling of chondrosarcoma reveals Src-pathway activity and dasatinib as option for treatment. Cancer Res. 2009;69:6216–22.

- Schrage YM, Machado I, Meijer D, Briaire-de Bruijn I, van den Akker BE, Taminiau AH, Kalinski T, Llombart-Bosch A, Bovée JV. COX-2 expression in chondrosarcoma: a role for celecoxib treatment? Eur J Cancer. 2010;46:616–24.
- van Oosterwijk JG, Anninga JK, Gelderblom H, Cleton-Jansen A-M, Bovée JVMG. Update on targets and novel treatment options for high-grade osteosarcoma and chondrosarcoma. Hematol Oncol Clin N Am. 2013;27:1021–48.
- 20. van Oosterwijk JG, Herpers B, Meijer D, Briaire-de Bruijn IH, Cleton-Jansen AM, Gelderblom H, van de Water B, Bovée JV. Restoration of chemosensitivity for doxorubicin and cisplatin in chondrosarcoma in vitro: BCL-2 family members cause chemoresistance. Ann Oncol. 2012;23:1617–26.
- 21. Schuetze SM, Bolejack V, Choy E, Ganjoo KN, Staddon AP, Chow WA, Tawbi HA, Samuels BL, Patel SR, von Mehren M, D'Amato G, Leu KM, Loeb DM, Forscher CA, Milhem MM, Rushing DA, Lucas DR, Chugh R, Reinke DK, Baker LH. Phase 2 study of dasatinib in patients with alveolar soft part sarcoma, chondrosarcoma, chordoma, epithelioid sarcoma, or solitary fibrous tumor. Cancer. 2017;123:90–7.
- 22. Grignani G, Palmerini E, Stacchiotti S, Boglione A, Ferraresi V, Frustaci S, Comandone A, Casali PG, Ferrari S, Aglietta M. A phase 2 trial of imatinib mesylate in patients with recurrent nonresectable chondrosarcomas expressing platelet-derived growth factor receptor-alpha or -beta: an Italian Sarcoma Group Study. Cancer. 2011;117:826–31.
- Polychronidou G, Karavasilis V, Pollack SM, Huang PH, Lee A, Jones RL. Novel therapeutic approaches in chondrosarcoma. Future Oncol. 2017;13:637–48.