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## 2.1 Introduction

Autism Spectrum Disorders (ASDs) are neurodevelopmental disabilities with a large heritable component. Concordance rate in monozygotic twins is 30–99% depending on the study, whereas concordance rates in dizygotic twins and siblings are 0–65% and 3–30%, with an estimated overall heritability of 0.7–0.8 [1]. ASD is clinically heterogeneous with respect to behavior, intellectual function, anthropometric traits (e.g., head size; BMI), and comorbid conditions [2]. The extreme clinical variability parallels the genetic heterogeneity, which is far to be completely identified. Indeed, even if epidemiological evidence from family and twin studies has convincingly demonstrated a strong genetic component to ASD, identifying the responsible genetic variants has been impaired by the lack of appropriate technical genomic tools. Only in recent years, we have rapidly developed novel and sensitive methods such as microarray analyses and next generation sequencing (NGS), which have allowed identifying several novel ASD-associated genetic and genomic lesions.

Several Mendelian diseases have been linked to ASD and genetic evidence suggests that up to 1500 genes are involved in ASD susceptibility [3]. Copy Number Variants (CNVs) explain 5–15% of ASD cases and pathogenic variants in single Mendelian genes likely account for a further 15–20%. Finally, oligogenic or polygenic inheritance may account for a still undetermined, but surely relevant group of patients.

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On the other hand, it is now clear that the same genetic determinant associated with ASD can also cause other neurodevelopmental anomalies, including isolated intellectual disability, or psychiatric disorders. The reason of this variable clinical expressivity is unknown and may be attributed to the genetic background, epigenetic or environmental factors.

The breakthrough of new genetic technologies has evidenced a determinant contribution of *de novo* genomic and genetic variants in ASD, which account for the rarity of familial cases of ASD. This means that these mutations arise in the parental germ cells or in somatic cells of the developing embryo. As for intellectual disability, the strong impact on phenotype associated with a reduced reproductive fitness of the severe ASD forms indicated *a priori* that *de novo* variants play an important role in ASD [4].

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## 2.2 CNVs Associated with Increased Risk for ASD

More than a decade ago, karyotyping and fluorescence in situ hybridization (FISH) have shown the role of rare genomic alterations in ASD [5], including the 7q11.23, 15q11–13, and 22q11.2 regions, already associated with micro-deletion and micro-duplication syndromes, characterized by autistic symptoms as a component [6, 7]. A breakthrough in the discovery of ASD genetic elements was determined by the development of microarray analyses, such as comparative genomic hybridization (CGH), which allowed a higher resolution—as low as 100 kb—compared with karyotype in the detection of CNVs [8]. The first analyses showed that individuals with ASD had 10–20 times the number of CNVs compared to healthy controls [9, 10]. Since then, a number of studies have consistently confirmed that individuals with ASD have more CNVs than non-related controls. In particular, the study of trios (parents and child) has revealed that part of ASD cases are caused by highly penetrant *de novo* CNVs [11, 12] (Table 2.1). The importance of CNVs in ASD is underlined by the fact that microarray methods to search for genomic deletions and duplications are now recommended as first-line genetic tests in ASD [13–16].

Part of the pathogenic CNVs are recurrent, i.e., involve the same genetic region in different affected subjects with *de novo* CNV. These are mediated by unequal crossing-over events due to a peculiar structure of the genomic region involved. On the other hand, many non-recurrent CNVs have been described, and are generated by different and more complex molecular mechanisms [17].

In both cases, a pathogenic CNV involves one or more dose-sensitive gene(s). This term indicates genes whose product amount is critical for the cell function. Its unbalance is thus associated with a genetic disease, both if decreased, such as in deletions, and increased, such as in duplications [18].

Some of the pathogenic CNVs can result in nearly opposite or mirror phenotypes depending on whether they are duplicated or deleted. This reciprocal impact of deletions/duplications is well-known for the 16p11.2 copy number variant. Severe obesity (deletion) and leanness (duplication) have mirror

**Table 2.1** CNVs frequently associated with ASD, and their counterparts

Alternative titles/related syndromes	Locus	CNV	Clinical features	OMIM number
Chromosome 1q21.1 deletion syndrome	1q21.1	Loss	Developmental delay, bone and cardiac anomalies	612474
Chromosome 1q21.1 duplication syndrome	1q21.1	Gain	ASD, schizophrenia	612475
Chromosome 2q37 deletion syndrome	2q37.2-q37.3	Loss	ASD	600430
	2q24.2	Loss	ASD	605556
Chromosome 2p16.3 deletion syndrome	2p16.3	Loss	ASD, MR, schizophrenia	614332
Chromosome 3q29 deletion syndrome	3q29	Loss	MR, ASD, schizophrenia, BD	609425
	3p14.2	Loss	ASD	
	3p14.2	Gain	ASD	
	6p23	Loss	ASD	600511
Williams–Beuren region duplication syndrome	7q11.23	Gain	ASD, speech delay, craniofacial anomalies	609757
	7q36.3	Gain	ID, schizophrenia	613959
	7q31.1	Loss		602081
Saethre–Chrotzen syndrome	7p21.1	Loss	ASD	101400
	10q11.23-q21.2	Gain	ASD	610987
	11q13.3-q13.4	Loss	ASD, ID	603290
	13q14.12-q14.13	Loss	ASD	615609
Chromosome 15q13.3 micro-deletion syndrome	15q13.3	Loss	ID, seizures, schizophrenia, ASD, BPD	612001
Duplication 15q11-q13 syndrome	15q11-q13.33	Gain	ASD	608636
Chromosome 15q11.2 deletion syndrome	15q11.2	Loss	ID, ADHD, schizophrenia, ASD	615656
Chromosome 16p13.3 deletion syndrome (Rubinstein–Taybi)	16p13.3	Loss	ASD	610543
	16p13.3	Gain	ID, speech problems, mild skeletal anomalies	613458
	16p13.1	Gain	ASD, ID, schizophrenia	
Chromosome 16p11.2 deletion syndrome, 220 kb	16p11.2	Loss	Obesity, obesity with developmental delay	613444
Chromosome 16p11.2 deletion syndrome, 593 kb	16p11.2	Loss	ASD	611913
Chromosome 16p11.2 duplication syndrome	16p11.2	Gain	ASD, schizophrenia, ADHD, microcephaly	614671
Chromosome 16p12.1 deletion syndrome, 520-kb	16p12.1	Loss	Developmental delay and learning disability	136570

(continued)

**Table 2.1** (continued)

Alternative titles/related syndromes	Locus	CNV	Clinical features	OMIM number
Smith–Magenis syndrome	17p11.2	Loss	ASD	182290
Potocki–Lupski syndrome	17p11.2	Gain	ASD	610883
Chromosome 17q12 deletion syndrome	17q12	Loss	ID, ASD, schizophrenia	614527
Chromosome 17q12 duplication syndrome	17q12	Gain	ID, behavioral abnormalities, psychomotor delay	614526
	22q13.33	Loss	ASD	606230
Chromosome 22q13 duplication syndrome	22q13.33	Gain	ASD, ID	615538
	20p13	Loss	ASD	
Chromosome 22q11.2 deletion syndrome	22q11.2	Loss	ID, schizophrenia, ADHD, other psychiatric disturbances	188400
Chromosome 22q11.2 duplication syndrome	22q11.2	Gain	ASD	608363
	Xp22.11	Loss	ASD, ADHD	300830
	Xq13.1	Loss	ASD, ID	300336

*ADHD* attention deficit hyperactivity disorder, *ID* intellectual disability

etiologies, possibly through contrasting effects on metabolism regulating energy balance [19]. A similar mirror phenotype is associated with the 7q11.23 deletion/duplication, both leading to multisystem neurodevelopmental disorders. The former is associated with the Williams–Beuren syndrome characterized by extreme friendliness and sociable traits lying at opposite ends of the same behavioral spectrum in duplication 7q11.23 which has language impairment, and autistic like features [20].

In other CNVs, two pathogenic different models have been proposed: (1) in the dominant model, gene expression changes in one direction only (decrease or increase) may contribute to a specific phenotype, with no effect (on the same trait) for a change in the other direction. An example is the immunodeficiency associated with the 22q11.2 deletion, but not observed in the corresponding duplication. (2) In the U-shaped model, genotype–phenotype correlations in reciprocal CNVs have allowed to demonstrate that a reduced or increased number in the number of copies of causal genes can lead to the same phenotype. Among many examples, the 15q13.3 deletion and duplication syndromes are overlapping (i.e., ID, DD, ASD, schizophrenia, ADHD) except for aggressive/impulsive behavior which are reported in deletion, but not duplication, carriers (up to 35%) [20].

Most recurrent CNVs are large (>400 kb), involving dozens of genes, and are individually rare (<0.1%). There are now several well-characterized rare CNVs, clearly associated with a high risk of ASD (Table 2.1). Very large CNVs seem to

be further enriched in individuals who have comorbidity with intellectual disability.

Recurrent CNVs can be distinguished in “syndromic,” where they are associated to a highly reproducible set of congenital anomalies, or “variable expressive CNVs,” resulting in a broad spectrum of disease phenotypes [21].

One emerging aspect of CNVs associated with ASD is that most manifest a wide range of clinical phenotypes. As an example, the 15q13.3 deletion and duplication are now clearly associated with ASD [22], ID [23], epilepsy [24], and schizophrenia [25].

Some CNVs are not only associated with a variable expressivity, but also with an incomplete penetrance. This is clear in families where the transmitting parent is apparently “unaffected” suggesting these CNVs are not sufficient to determine the disease. Indeed, recent works provide evidence for an oligogenic CNV model, where in addition to the primary CNV, a second CNV (inherited or *de novo*) is required at a different locus for a child to develop ASD. This phenomenon is exemplified by the 520-kb deletion on chromosome 16p12.1 (MIM# 136570), which is associated with developmental delay and extensive phenotypic variability [26]. Interestingly, in most cases, this deletion was inherited from a parent who also manifested mild neuropsychiatric features, and the severely affected children were more likely to carry another large (>500 kb) rare CNV.

These results suggest a contribution of rare variants in the genetic background toward neurodevelopmental disorders, depending upon the extent to which the primary variant sensitizes an individual toward a specific pathological phenotypic trajectory [26].

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### 2.3 Rare Highly Penetrant ASD Genes

CNVs are causative in only 5–15% of individuals with ASD, suggesting that other types of mutations must be operant in ASD as well. Rare Mendelian syndromes have been associated with ASD, showing that at least in part ASD is a monogenic disorder [27, 28]. Among these, fragile X syndrome (*FMR1* gene), Rett syndrome (*MECP2* gene), tuberous sclerosis (*TSC1* and *TSC2* genes), Timothy syndrome (*CACNA1C* gene), all display partial comorbidity with ASD [18].

The recent widespread availability of next generation sequencing (NGS) allowed a further increase in the resolution to detect genetic alteration in ASD. Starting from 2008, NGS has allowed to sequence the coding genes of an entire human genome, the so-called whole exome sequencing (WES) strategy, at affordable prices and without the need of an *a priori* hypothesis on the disease gene. A number of large WES studies have been completed in ASD, now encompassing several thousands of individuals [29–35]. Rare autosomal recessive disorders were identified in consanguineous families, affecting for instance the *AMT*, *BCKDK*, *C3ORF58*, *CNTNAP2*, *NHE9*, *PCDH10*, *PEX7*, *SYNE1*, *VPS13B*, *PAH*, *POMGNT1*, and *SLC9A9* genes [36–38]. These are associated with highly variable clinical presentation, and ASD

can be isolated in patients lacking the diagnostic criteria and features of the associated Mendelian disorders. A limited number of X-linked genes have also identified to contribute to ASD, among these, the already cited *FMR1* and *MECP2*, and neuroligins *NLGN3* and *NLGN4* [39, 40]. However, recently new important X-linked genes are emerging such as the X-linked dominant *DDX3X* gene, affecting females only [41].

The list of these genes is still limited and will surely expand in the next years thanks to the new sequencing methods.

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## 2.4 Novel Highly Penetrant ASD Genes

As for CNVs, also single-nucleotide pathogenic variations so far discovered are mainly *de novo* in highly penetrant ASD. These genes behave as autosomal dominant and are rarely found segregating in families (e.g., *SHANK1*, *SHANK2*, and *SHANK3*) often because their strong effect on reproductive fitness reduction. WES studies identified a number of high-confidence ASD candidate genes that likely may represent up to 20% of cases [42, 43]. Some of them are recurrently hit among families, such as *CHD8*, *DYRK1A*, *KATNAL2*, *GRIN2B*, *POGZNTNG1*, and *SCN2A* [44]. However, the general notion is that many genes associated with ASD phenotype are likely to be very rare or even “private,” unlikely to be found in many individuals. This suggests that rare variants have a larger than originally expected impact on ASD risk, although large cohorts of patients are needed to deepen the knowledge on this issue.

The list of candidate genes involved in ASD is continuously increasing as the complexity of data supporting their pathogenicity. Several groups have tried to develop criteria to rank and assess the strength of evidence associated with candidate genes. Among these one of the most complete databases is SFARI gene (<https://gene.sfari.org/>), built on information extracted from peer-reviewed scientific and clinical studies on the molecular genetics and biology of ASD [45]. SFARI gene integrates genetic, neurobiological, and clinical information about genes associated with ASD, reporting a total of 956 genes (version 3.0). The annotation criteria used allow dividing genes into seven categories: syndromic genes predisposing to autism in the context of a syndromic disorder (e.g., fragile X syndrome); categories 1 and 2 (high and strong confidence) contain genes with a genome-wide statistical significance, with independent replication; categories 3 and 4 (suggestive and minimal evidence) list genes reported in relatively small studies, whose evidence is still incomplete. Finally, in category 5 (hypothesized but untested) are reported genes that have been implicated solely by evidence in model organisms or other evidence of a marginal nature, and category 6 (evidence does not support a role) is for those genes that have been tested in a human cohort, but the weight of the evidence argues against a role in ASD (Table 2.2).

**Table 2.2** ASD genes reported in SFARI database (November 2018)

Category	Definition	N. of genes
Syndromic	Genes with a substantial degree of increased risk for ASD, and consistently linked to additional characteristics not required for an ASD diagnosis	79
Category 1, high confidence	Genes with evidence of recurrent and convincing mutations (functional or large pedigree segregation) accompanied by a rigorous statistical comparison with the mutation frequency in controls. This category also includes single genes that reached genome-wide significance in association studies independently replicated, or which reached genome-wide significance via meta-analysis of all current association studies	25
Category 2, strong candidate	Rare mutations that are recurrent and convincing (see above), accompanied by a rigorous statistical comparison with the mutation frequency in controls. Rare <i>de novo</i> variants, likely to be disruptive, in three or more unrelated cases. Results from association studies must reach genome-wide significance, uniquely implicating a single gene, but with no independent replication. Alternatively, consistently replicated association of the same allele, falling short of genome-wide significance, that must be accompanied by evidence that the risk variant has a relevant functional effect in humans	61
Category 3, suggestive evidence	Genes with consistently replicated association of the same allele, without functional support. Rare <i>de novo</i> variants, likely to be disruptive, in two or more unrelated cases. Genes within a CNV, or near a GWAS peak close to significance, with additional accessory evidence	184
Category 4, minimal evidence	Genes in an ASD-associated multi-genic CNV, proximal to genome-wide significant intergenic variants for which there is no other independent evidence. Any significant, convincing, but unreplicated association study data, along with any instances of multiple but inconsistent reports of association that are not overall significant by meta-analysis Genes with a series of two or more putative mutations identified (e.g., non-synonymous substitutions, single-gene deletion, duplication, disruption by translocation) for which there is not rigorous statistical comparison with controls Single rare <i>de novo</i> variants, likely to be disruptive	437
Category 5, hypothesized	Genes for which the only evidence comes from studies of model organisms. Genes in a region of linkage with no unique evidence for that gene versus others. Genes shown to functionally interact with category ASD strong candidates. Genes with a single rare variant observed in a single ASD case/family are placed here	170

This table reports a summary whose full text is available at <https://gene.sfari.org/about-gene-scoring/>

## 2.5 Common Variants Risk to ASD

High-throughput genotyping of single-nucleotide polymorphisms (SNP) allowed a large number of genome-wide association studies (GWAS) to identify common variants to ASD risk [46]. The potential number of genes likely able to confer moderately-sized risk for ASD is large. In fact, statistical modeling based on published results of both rare and common variation has predicted that up to 1000–1500 genes may ultimately be found to be associated with ASD [35, 47]. The comprehension of how such a large and varied number of genes can all be associated with one common clinical phenotype will be the major challenge to the field. The challenge of understanding how combinations of susceptibility genes interact during human brain development to cause disease (epistasis) has only begun to be explored.

Common variation throughout the genome exerts substantial additive genetic effects on ASD liability, with simplex/multiplex family status having an impact on the identified composition of that risk. As a fraction of the total variation in liability, the estimated narrow-sense heritability exceeds 60% for ASD individuals from multiplex families and is approximately 40% for simplex families. Genome-wide association studies demonstrate that a myriad of common variants of very small effect impacts ASD liability. The identification of such variants needs huge cohorts of patients to be analyzed and represents the challenge of ASD genetics for the next decades [48].

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## 2.6 Biological Insights into ASD

The genetic architecture of ASD has been proved to be complex and the large majority of cases still have no identifiable genetic cause [3, 49]. Despite these limitations, ASD-causing genes have started providing clues on functional pathways involved in the pathogenesis. WES studies have demonstrated grouping of protein–protein interaction networks, enriched for involvement in beta-catenin, p53 signaling, chromatin remodeling, ubiquitination, and neuronal function [29, 34, 43, 47, 50]. The analyses of convergent pathways integrated with experimental findings based on transcriptomic, and cellular and mouse models are now pointing toward three major cellular pathways interconnected through neuronal activity [44, 51].

1. Synapse development and function. The development and/or maintenance of synaptic function seem a critical factor in development of ASD [52]. Among the important genes are those encoding the presynaptic cell-adhesion molecules (CAMs) neuroligins (*NRXN1*) and their postsynaptic partners, neuroligins (*NLGN3* and *NLGN4*). Other molecules involved in pre-post synaptic anchoring are the SHANK family (*SHANK3*) and other molecules connected with the actin cytoskeleton (*CNTNAP2*). The most common electrophysiological and neuro-anatomical findings evidenced by mouse models of these genes are altered glutamatergic synaptic transmission, loss of inhibitory GABA interneurons, and impairment in synaptic plasticity attributable to dysfunction of NMDA and



AMPA receptors. Similar findings have been reported in the duplication 15q syndrome mouse models (*UBE3A* gene) which recapitulates the three core ASD features [53]. Glutamatergic transmission might represent a targetable pathway in ASD. Indeed, *Fmr1* knockout mice show a hyperactive mGluR5 signaling, leading to excessive protein synthesis at the synapse and increased trafficking of AMPA receptors [54, 55].

2. Growth, transcription regulation, and protein synthesis. Many ASD risk genes (e.g., *TSC1*, *TSC2*, and *PTEN*) lie downstream the signaling pathway containing mTOR, a key regulator of cell growth, proliferation, and survival. These genes are predicted to alter protein synthesis within synaptic spines, which is necessary for neuronal plasticity and thus proper cognitive function. Among the recently introduced genes in this list, *CDKL5* (Rett-like syndrome) has recently been shown to affect the mTOR pathway [56]. WNT pathway signaling is also considered to have a key role in the etiology of ASD [57, 58]. Defective synaptogenesis (or synaptic function), altered WNT signaling during brain development, and altered transcription and/or translation in neurons can influence neuronal circuit formation and activity [51].
3. Serotonin signaling and neuropeptides. Alterations in the serotonergic system were among the earliest evidence of abnormal brain function in ASD [59]. Serotonin mediates neurogenesis, cell migration and survival, synaptogenesis and plasticity [60]. Several variants in the serotonin system have been linked to ASD (*SERT/SLC6A4*, *MAOA*) [61].

A complementary approach to identify biological relationships among identified genes is to analyze the specific expression time window or molecular process. Two independent works showed that ASD genes are likely expressed in the mid-fetal brain (10–12 weeks of gestation), spatially corresponding to superficial glutamatergic neurons [62, 63]. Interestingly, ASD genes encode messenger RNAs interacting with FMRP, encoded by the *FMR1* gene, suggesting that convergence at common pathways of synaptic plasticity associated with gene regulation is mediated by this protein [47, 64].

Overall, a key role for fetal glutamatergic neuron development has been established for ASD, with a growing evidence for converging pathways in ASD-causing genes, with spatiotemporal specific expression pathways [44].

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## 2.7 Conclusion

In recent years, major progress in understanding the genetic architecture of ASD has been made. We now know that both rare and common variants contribute to ASD, with a number of genes and loci implicated. Much remains unknown: the penetrance and expressivity of many ASD genes is still to be determined, as well as the contribution of low penetrance genes in oligogenic forms. Several large-scale projects have just begun to understand both the genetic architecture and the pathophysiological mechanism of these heterogeneous disorders.

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## References

1. Ramaswami G, Geschwind DH. Genetics of autism spectrum disorder. *Handb Clin Neurol*. 2018;147:321–9.
2. American Psychiatric Association. 5th ed. Washington, DC: American Psychiatric Association; 2013.
3. Berg JM, Geschwind DH. Autism genetics: searching for specificity and convergence. *Genome Biol*. 2012;13(7):247.
4. Vissers LE, de Ligt J, Gilissen C, Janssen I, Steehouwer M, de Vries P, et al. A de novo paradigm for mental retardation. *Nat Genet*. 2010;42(12):1109–12.
5. Vorstman JA, Staal WG, van Daalen E, van Engeland H, Hochstenbach PF, Franke L. Identification of novel autism candidate regions through analysis of reported cytogenetic abnormalities associated with autism. *Mol Psychiatry*. 2006;11(1):1, 18–28
6. Somerville MJ, Mervis CB, Young EJ, Seo EJ, del Campo M, Bamforth S, et al. Severe expressive-language delay related to duplication of the Williams-Beuren locus. *N Engl J Med*. 2005;353(16):1694–701.
7. Mabb AM, Judson MC, Zylka MJ, Philpot BD. Angelman syndrome: insights into genomic imprinting and neurodevelopmental phenotypes. *Trends Neurosci*. 2011;34(6):293–303.
8. Alkan C, Coe BP, Eichler EE. Genome structural variation discovery and genotyping. *Nat Rev Genet*. 2011;12(5):363–76.
9. Jacquemont ML, Sanlaville D, Redon R, Raoul O, Cormier-Daire V, Lyonnet S, et al. Array-based comparative genomic hybridisation identifies high frequency of cryptic chromosomal rearrangements in patients with syndromic autism spectrum disorders. *J Med Genet*. 2006;43(11):843–9.
10. Sebat J, Lakshmi B, Malhotra D, Troke J, Lese-Martin C, Walsh T, et al. Strong association of de novo copy number mutations with autism. *Science*. 2007;316(5823):445–9.
11. Pinto D, Delaby E, Merico D, Barbosa M, Merikangas A, Klei L, et al. Convergence of genes and cellular pathways dysregulated in autism spectrum disorders. *Am J Hum Genet*. 2014;94(5):677–94.
12. Pinto D, Pagnamenta AT, Klei L, Anney R, Merico D, Regan R, et al. Functional impact of global rare copy number variation in autism spectrum disorders. *Nature*. 2010;466(7304):368–72.
13. Miller DT, Adam MP, Aradhya S, Biesecker LG, Brothman AR, Carter NP, et al. Consensus statement: chromosomal microarray is a first-tier clinical diagnostic test for individuals with developmental disabilities or congenital anomalies. *Am J Hum Genet*. 2010;86(5):749–64.
14. Manning M, Hudgins L, Professional P, Guidelines C. Array-based technology and recommendations for utilization in medical genetics practice for detection of chromosomal abnormalities. *Genet Med*. 2010;12(11):742–5.
15. Committee on Bioethics; Committee on Genetics, and; American College of Medical Genetics and; Genomics Social; Ethical; Legal Issues Committee. Ethical and policy issues in genetic testing and screening of children. *Pediatrics*. 2013;131(3):620–2.
16. Volkmar F, Siegel M, Woodbury-Smith M, King B, McCracken J, State M, et al. Practice parameter for the assessment and treatment of children and adolescents with autism spectrum disorder. *J Am Acad Child Adolesc Psychiatry*. 2014;53(2):237–57.
17. Zhang F, Gu W, Hurles ME, Lupski JR. Copy number variation in human health, disease, and evolution. *Annu Rev Genomics Hum Genet*. 2009;10:451–81.
18. Sztainberg Y, Zoghbi HY. Lessons learned from studying syndromic autism spectrum disorders. *Nat Neurosci*. 2016;19(11):1408–17.

19. Jacquemont S, Reymond A, Zufferey F, Harewood L, Walters RG, Kutalik Z, et al. Mirror extreme BMI phenotypes associated with gene dosage at the chromosome 16p11.2 locus. *Nature*. 2011;478(7367):97–102.
20. Deshpande A, Weiss LA. Recurrent reciprocal copy number variants: roles and rules in neurodevelopmental disorders. *Dev Neurobiol*. 2018;78(5):519–30.
21. Girirajan S, Rosenfeld JA, Coe BP, Parikh S, Friedman N, Goldstein A, et al. Phenotypic heterogeneity of genomic disorders and rare copy-number variants. *N Engl J Med*. 2012;367(14):1321–31.
22. Pagnamenta AT, Wing K, Sadighi Akha E, Knight SJ, Bolte S, Schmotzer G, et al. A 15q13.3 microdeletion segregating with autism. *Eur J Hum Genet*. 2009;17(5):687–92.
23. Sharp AJ, Mefford HC, Li K, Baker C, Skinner C, Stevenson RE, et al. A recurrent 15q13.3 microdeletion syndrome associated with mental retardation and seizures. *Nat Genet*. 2008;40(3):322–8.
24. Helbig I, Mefford HC, Sharp AJ, Guipponi M, Fichera M, Franke A, et al. 15q13.3 microdeletions increase risk of idiopathic generalized epilepsy. *Nat Genet*. 2009;41(2):160–2.
25. Stefansson H, Rujescu D, Cichon S, Pietilainen OP, Ingason A, Steinberg S, et al. Large recurrent microdeletions associated with schizophrenia. *Nature*. 2008;455(7210):232–6.
26. Pizzo L, Jensen M, Polyak A, Rosenfeld JA, Mannik K, Krishnan A, et al. Rare variants in the genetic background modulate cognitive and developmental phenotypes in individuals carrying disease-associated variants. *Genet Med*. 2019;21(4):816–25.
27. Abrahams BS, Geschwind DH. Advances in autism genetics: on the threshold of a new neurobiology. *Nat Rev Genet*. 2008;9(5):341–55.
28. Betancur C. Etiological heterogeneity in autism spectrum disorders: more than 100 genetic and genomic disorders and still counting. *Brain Res*. 2011;1380:42–77.
29. O’Roak BJ, Vives L, Girirajan S, Karakoc E, Krumm N, Coe BP, et al. Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations. *Nature*. 2012;485(7397):246–50.
30. O’Roak BJ, Deriziotis P, Lee C, Vives L, Schwartz JJ, Girirajan S, et al. Exome sequencing in sporadic autism spectrum disorders identifies severe de novo mutations. *Nat Genet*. 2011;43(6):585–9.
31. O’Roak BJ, State MW. Autism genetics: strategies, challenges, and opportunities. *Autism Res*. 2008;1(1):4–17.
32. O’Roak BJ, Stessman HA, Boyle EA, Witherspoon KT, Martin B, Lee C, et al. Recurrent de novo mutations implicate novel genes underlying simplex autism risk. *Nat Commun*. 2014;5:5595.
33. O’Roak BJ, Vives L, Fu W, Egerton JD, Stanaway IB, Phelps IG, et al. Multiplex targeted sequencing identifies recurrently mutated genes in autism spectrum disorders. *Science*. 2012;338(6114):1619–22.
34. Neale BM, Kou Y, Liu L, Ma’ayan A, Samocha KE, Sabo A, et al. Patterns and rates of exonic de novo mutations in autism spectrum disorders. *Nature*. 2012;485(7397):242–5.
35. Sanders SJ, Murtha MT, Gupta AR, Murdoch JD, Raubeson MJ, Willsey AJ, et al. De novo mutations revealed by whole-exome sequencing are strongly associated with autism. *Nature*. 2012;485(7397):237–41.
36. Musante L, Ropers HH. Genetics of recessive cognitive disorders. *Trends Genet*. 2014;30(1):32–9.
37. Yu TW, Chahrour MH, Coulter ME, Jiralerspong S, Okamura-Ikeda K, Ataman B, et al. Using whole-exome sequencing to identify inherited causes of autism. *Neuron*. 2013;77(2):259–73.
38. Morrow EM, Yoo SY, Flavell SW, Kim TK, Lin Y, Hill RS, et al. Identifying autism loci and genes by tracing recent shared ancestry. *Science*. 2008;321(5886):218–23.
39. Jamain S, Quach H, Betancur C, Rastam M, Colineaux C, Gillberg IC, et al. Mutations of the X-linked genes encoding neuroligins NLGN3 and NLGN4 are associated with autism. *Nat Genet*. 2003;34(1):27–9.
40. Nava C, Lamari F, Heron D, Mignot C, Rastetter A, Keren B, et al. Analysis of the chromosome X exome in patients with autism spectrum disorders identified novel candidate genes, including TMLHE. *Transl Psychiatry*. 2012;2:e179.

41. Wang X, Posey JE, Rosenfeld JA, Bacino CA, Scaglia F, Immken L, et al. Phenotypic expansion in DDX3X—a common cause of intellectual disability in females. *Ann Clin Transl Neurol.* 2018;5(10):1277–85.
42. Iossifov I, O’Roak BJ, Sanders SJ, Ronemus M, Krumm N, Levy D, et al. The contribution of de novo coding mutations to autism spectrum disorder. *Nature.* 2014;515(7526):216–21.
43. De Rubeis S, He X, Goldberg AP, Poultney CS, Samocha K, Cicek AE, et al. Synaptic, transcriptional and chromatin genes disrupted in autism. *Nature.* 2014;515(7526):209–15.
44. Chen JA, Penagarikano O, Belgard TG, Swarup V, Geschwind DH. The emerging picture of autism spectrum disorder: genetics and pathology. *Annu Rev Pathol.* 2015;10:111–44.
45. Abrahams BS, Arking DE, Campbell DB, Mefford HC, Morrow EM, Weiss LA, et al. SFARI gene 2.0: a community-driven knowledgebase for the autism spectrum disorders (ASDs). *Mol Autism.* 2013;4(1):36.
46. Gaugler T, Klei L, Sanders SJ, Bodea CA, Goldberg AP, Lee AB, et al. Most genetic risk for autism resides with common variation. *Nat Genet.* 2014;46(8):881–5.
47. Iossifov I, Ronemus M, Levy D, Wang Z, Hakker I, Rosenbaum J, et al. De novo gene disruptions in children on the autistic spectrum. *Neuron.* 2012;74(2):285–99.
48. Klei L, Sanders SJ, Murtha MT, Hus V, Lowe JK, Willsey AJ, et al. Common genetic variants, acting additively, are a major source of risk for autism. *Mol Autism.* 2012;3(1):9.
49. Stein JL, Parikshak NN, Geschwind DH. Rare inherited variation in autism: beginning to see the forest and a few trees. *Neuron.* 2013;77(2):209–11.
50. Michaelson JJ, Shi Y, Gujral M, Zheng H, Malhotra D, Jin X, et al. Whole-genome sequencing in autism identifies hot spots for de novo germline mutation. *Cell.* 2012;151(7):1431–42.
51. Quesnel-Vallieres M, Weatheritt RJ, Cordes SP, Blencowe BJ. Autism spectrum disorder: insights into convergent mechanisms from transcriptomics. *Nat Rev Genet.* 2019;20(1):51–63.
52. Zoghbi HY. Postnatal neurodevelopmental disorders: meeting at the synapse? *Science.* 2003;302(5646):826–30.
53. Smith SE, Zhou YD, Zhang G, Jin Z, Stoppel DC, Anderson MP. Increased gene dosage of Ube3a results in autism traits and decreased glutamate synaptic transmission in mice. *Sci Transl Med.* 2011;3(103):103ra97.
54. Bear MF, Huber KM, Warren ST. The mGluR theory of fragile X mental retardation. *Trends Neurosci.* 2004;27(7):370–7.
55. Huber KM, Gallagher SM, Warren ST, Bear MF. Altered synaptic plasticity in a mouse model of fragile X mental retardation. *Proc Natl Acad Sci U S A.* 2002;99(11):7746–50.
56. Wang IT, Allen M, Goffin D, Zhu X, Fairless AH, Brodtkin ES, et al. Loss of CDKL5 disrupts kinome profile and event-related potentials leading to autistic-like phenotypes in mice. *Proc Natl Acad Sci U S A.* 2012;109(52):21516–21.
57. Kwan V, Unda BK, Singh KK. Wnt signaling networks in autism spectrum disorder and intellectual disability. *J Neurodev Disord.* 2016;8:45.
58. Hormozdiari F, Penn O, Borenstein E, Eichler EE. The discovery of integrated gene networks for autism and related disorders. *Genome Res.* 2015;25(1):142–54.
59. Cook EH, Leventhal BL. The serotonin system in autism. *Curr Opin Pediatr.* 1996;8(4):348–54.
60. Azmitia EC. Serotonin and brain: evolution, neuroplasticity, and homeostasis. *Int Rev Neurobiol.* 2007;77:31–56.
61. Veenstra-VanderWeele J, Muller CL, Iwamoto H, Sauer JE, Owens WA, Shah CR, et al. Autism gene variant causes hyperserotonemia, serotonin receptor hypersensitivity, social impairment and repetitive behavior. *Proc Natl Acad Sci U S A.* 2012;109(14):5469–74.
62. Willsey AJ, Sanders SJ, Li M, Dong S, Tebbenkamp AT, Muhle RA, et al. Coexpression networks implicate human midfetal deep cortical projection neurons in the pathogenesis of autism. *Cell.* 2013;155(5):997–1007.
63. Parikshak NN, Luo R, Zhang A, Won H, Lowe JK, Chandran V, et al. Integrative functional genomic analyses implicate specific molecular pathways and circuits in autism. *Cell.* 2013;155(5):1008–21.
64. Steinberg J, Webber C. The roles of FMRP-regulated genes in autism spectrum disorder: single- and multiple-hit genetic etiologies. *Am J Hum Genet.* 2013;93(5):825–39.