

The Liver and Insulin Resistance: The Important Convergence of Endocrinology and Hepatology

13

Charissa Y. Chang, Kerry Whitt, Zhenqi Liu, and Stephen H. Caldwell

Introduction

Recognition of a link between insulin resistance (IR) and liver disease dates back at least 100 years to the term "hepatogenous diabetes," which describes the association between cirrhosis and development of diabetes [\[1](#page-8-0)], and more recently to the term "diabetic fatty liver," which antedated the now more common terms "nonalcoholic steatohepatitis" (NASH) and "nonalcoholic fatty liver disease" (NAFLD). These terms were introduced in the 1980s and 1990s, respectively. Since their introduction, the ever-rising prevalence of obesity has brought increased attention to these disorders as the hepatic manifestation of "metabolic" or "insulin resistance" syndrome. Indeed, IR appears to be the common link among

Liver Disease Practice, Mount Sinai Health System, New York, NY, USA

e-mail[: charissa.chang@mssm.edu](mailto:charissa.chang@mssm.edu)

K. Whitt RMG Gastroenterology, Raleigh, NC, USA

Z. Liu

Division of Endocrinology and Metabolism, Department of Medicine, University of Virginia School of Medicine, Charlottesville, VA, USA

S. H. Caldwell

Division of Gastroenterology and Hepatology, Department of Medicine, University of Virginia Health System, Digestive Health Center, Charlottesville, VA, USA

metabolic syndrome, obesity, and nonalcoholic fatty liver. Metabolic alterations and hepatic steatosis can develop in the insulin-resistant state in the absence of and prior to diabetes mellitus. Moreover, it is now known that IR correlates with increasing fibrosis in other liver diseases including hepatitis C.

These pathological relationships have raised a now-common issue: "Does insulin resistance cause fatty liver or does fatty liver cause insulin resistance?" Below, we will discuss why the answer to this complicated "either $-$ or" metabolic question is actually "yes." In other words, fatty liver both results from IR and contributes to the problem. In order to better understand the relationship between IR and fatty liver disease, it is best to consider them in light of the most basic actions of insulin and other co-variables important in energy homeostasis and, perhaps, to consider the most fundamental disturbance in pathological IR states—disturbed intracellular fatty acid metabolism that leads to "lipotoxicity" or cellular injury due to the excessive accumulation of triglycerides and fatty acids and their subsequent oxidation. By understanding the normal flux of glucose and lipid between the major targets of insulin (adipose, muscle, and liver) and how IR relates to fatty liver disease, we can hopefully identify possibilities for early therapeutic intervention. Moreover, integration of the hepatologist's knowledge of human hepatic pathology and pathophysiology coupled to the

C. Y. Chang (\boxtimes)

[©] Springer Nature Switzerland AG 2020 207

P. S. Zeitler, K. J. Nadeau (eds.), *Insulin Resistance*, Contemporary Endocrinology, https://doi.org/10.1007/978-3-030-25057-7_13

endocrinologist's knowledge of insulin signaling and the associated metabolism of glucose and fat is essential in managing the growing problem of liver disease associated with the IR syndrome.

Normal Glucose and Lipid Flux in the Fed and Fasting States

The liver plays a central role in maintaining energy homeostasis. It is the major source of glucose production through glycogenolysis and gluconeogenesis. Overall, glucose homeostasis occurs through a balance between energy supply and demands from key organs (muscle, adipose, brain, and liver) and differs in the fed versus fasting state. In the fasting state, rates of glucose production and utilization are equal. In this state, 50% of glucose disposal occurs in the brain, 25% occurs in the splanchnic area, and 25% occurs in muscles [[2\]](#page-8-1). The liver meets its glucose demands through glycogenolysis and later through gluconeogenesis if fasting is prolonged beyond 10–18 h. While other organs are able to synthesize and hydrolyze glycogen, only the liver and kidney express glucose-6-phosphatase, the enzyme required for the release of glucose into the circulation.

Adipose tissue stores of triglycerides are an important source of energy during fasting through the release of stored fatty acids by hormonesensitive lipase (which is normally inhibited in the fed state by insulin). Once cleaved from glycerol, albumin-bound fatty acids are delivered to other tissues via the fatty acid-binding protein (FABP), which facilitates movement of fatty acids into the cell. There they can then undergo oxidation to provide adenosine triphosphate (ATP) for cellular activity, while glycerol is either used to resynthesize triglyceride or is converted to glucose through gluconeogenesis in the liver.

During periods of abundant calorie intake, excess glucose is converted to lipids, which are either stored as triglycerides or incorporated into lipoproteins (i.e., very low-density lipoproteins (VLDL)) to be exported out of the liver. De novo fatty acid synthesis from dietary carbohydrate occurs primarily in the liver, in lactating mammary glands, and to a lesser extent in the adipose tissue. In the fed state, triglycerides are synthesized within adipocytes through the action of lipoprotein lipase.

Hormonal Regulation: Insulin, Glucagon, and the Insulin Receptor Signaling Pathway

Under normal conditions, plasma glucose concentration is tightly maintained despite wide fluctuations in glucose supply and utilization during different states (fasting, fed, exercise, rest). Regulation is achieved through the competing hormones, insulin, glucagon, and epinephrine, and is heavily influenced by the activity of adipokines from adipose tissue and, more fundamentally, by the action of increased intracellular fatty acids that alter insulin signaling.

Insulin

Insulin is an anabolic hormone that regulates glucose homeostasis through actions on three integrated target tissues: liver, muscle, and adipose. It stimulates cell growth and differentiation, promotes storage of substrates in liver, fat, and muscle through lipogenesis as well as glycogen and protein synthesis, and inhibits lipolysis, glycogenolysis, and protein breakdown. Following a meal, one-third of the glucose is delivered to the liver, one-third to muscle and adipose, and one-third to non-insulin-dependent tissues (i.e., brain). A rise in plasma glucose concentration stimulates the release of insulin from pancreatic $β$ (beta)-cells. The liver removes 60% of the insulin that enters through the portal vein. Peripheral insulin mediates glucose uptake, glycolysis, and conversion to glycogen in muscle and adipose tissue. Most of the glucose delivered to peripheral tissues is utilized by muscle (80–85%), whereas only a small amount (4–5%) is metabolized in adipose [\[2](#page-8-1)]. While glucose disposal in adipose tissue is relatively low compared to muscle, adipose tissue plays a key role in overall glucose homeostasis through the release of free fatty acids (FFAs) and expression of adipokines, as will be discussed below.

Insulin regulates glucose metabolism in the liver through both direct and indirect means. Direct effects on the liver result in decreased glycogenolysis and gluconeogenesis by decreasing transcription and suppressing the activity of phosphoenolpyruvate carboxylase (PEPCK) and glucose-6-phosphatase (G-6-Pase), which are key enzymes involved in gluconeogenesis [\[3](#page-8-2)]. Indirect effects result from insulin's action on peripheral tissues. Insulin causes peripheral uptake of glucose in adipose and muscle tissue by stimulating translocation of the transporter GLUT 4 to the plasma membrane [\[4](#page-8-3)]. Insulin is anabolic in muscle by promoting glycogen and lipid synthesis while suppressing lipolysis and gluconeogenesis. Insulin promotes triglyceride storage and decreased FFA release in adipose tissue. The combined effects of insulin on muscle and adipose tissue result in decreased FFA influx into liver, which indirectly leads to less gluconeogenesis.

Glucagon and Epinephrine

Glucagon is a polypeptide secreted by α (alpha)cells of the pancreas. Under physiologic conditions, it acts on the liver by activating glycogenolysis and has a relatively small role in

gluconeogenesis. Glucagon favors partitioning of FFAs released by adipose tissue to oxidation in the liver to acetyl CoA, which can be used to form ketone bodies. Stress hormones including epinephrine, cortisol, and growth hormone increase glucose through stimulation of hepatic gluconeogenesis [[5\]](#page-8-4). Epinephrine is also especially important in activating adenylate cyclase in the adipocyte membrane, which results in the formation of 3,5′-cyclic adenosine monophosphate (AMP) that leads to lipolysis through activation of hormone-sensitive lipase.

Insulin Receptor Signaling Pathway

The insulin receptor is a tetramer composed of 2 extracellular α (alpha)-units bound to 2 membrane-spanning β (beta)-units (Fig. [13.1\)](#page-2-0). Activation begins with binding of insulin to the α (alpha)-subunit, which then triggers autophosphorylation of the β(beta)-subunit. Tyrosine kinase-mediated phosphorylation of the insulin receptor substrate (IRS) then triggers a cascade of pathways that differ depending upon the target tissue and the specific IRS type [[6,](#page-8-5) [7\]](#page-8-6). Four

Fig. 13.1 Insulin receptor signaling pathway. Insulin binds to a four-subunit membrane-spanning receptor, triggering tyrosine phosphorylation of the beta subunit, which results in a signaling cascade with combined endpoints: (a) translocation of the GLUT-4 glucose trans-

porter; (b) metabolic effects mediated through steroid regulatory element-binding protein (SREBP); and (c) mitogenic effects (growth, cell differentiation) through activation of the mitogen-activated protein kinase (MAPK) pathway

different types of IRS proteins have been identified: IRS 1 (skeletal muscle), IRS 2 (liver), IRS 3 (adipose, β[beta]-cells, liver), and IRS 4 (thymus, brain, kidney) [[8\]](#page-8-7). Binding of insulin to IRS proteins in muscle and adipose activates phosphatidylinositol 3-kinase (PI3K), which leads to translocation of GLUT 4 glucose transporter proteins to the cell membrane, resulting in an increase in transport of glucose into cells.

Unlike in muscle and adipose tissue, glucose transport in the liver occurs through the GLUT 2 transporter, which is not affected by insulin. This is an important distinction, as failure of insulin to suppress endogenous glucose production by the hepatocyte constitutes one of the pillars of the insulin-resistant state. This nondependence of the hepatocyte on insulin for glucose uptake may in fact allow the liver to function as something of a sink for excess glucose. Insulin signaling by PI3K in the liver appears to be important in activation of downstream expression of genes encoding enzymes involved in glycolysis, glycogen synthesis, and lipid synthesis through steroid regulatory element-binding protein (SREBP)-1c. Gluconeogenesis is inhibited by altering gene expression by the forkhead family of transcriptional factors (FoxA), while growth and cell differentiation effects of insulin are mediated through activation of the mitogen-activated protein kinase (MAPK) pathway via growth factor receptor-binding protein 2 (Grb2).

The Insulin-Resistant State

Insulin Resistance and Insulin Resistance Syndrome Defined

IR is defined as impaired response to normal or elevated insulin levels [\[9](#page-8-8)]. While it is often quantified through the euglycemic clamp model [\[10](#page-8-9)] and homeostasis model assessment (HOMA) [\[11](#page-8-10)] or quantitative insulin sensitivity check index (QUICKI) assessments [[12,](#page-8-11) [13](#page-9-0)], it is important to note that these only address the role of insulin in glucose metabolism. Using a modification of the fasting insulin and glucose-derived QUICKI, efforts have been made to incorporate fasting

fatty acid levels to improve the measurement of IR [\[14](#page-9-1)]. However, all these tests have some inherent limitations, although they remain clinically and experimentally very useful [[15\]](#page-9-2).

Insulin resistance syndrome has been defined by Reaven to encompass the multiple sequelae that result from the compensatory hyperinsulinemic state associated with IR. This includes dyslipidemia, endothelial dysfunction, and alterations in procoagulant factors and markers of inflammation [\[16](#page-9-3)]. Clinical manifestations include diabetes mellitus, cardiovascular disease, hypertension, polycystic ovary syndrome, and NAFLD.

Target Organ Alterations in IR

With the exception of specific genetic defects in the insulin receptor (leprechaunism, Rabson– Mendenhall syndrome, type A syndrome of IR), IR results from the typical combination of predisposing genetic and environmental factors (e.g., excessive calorie intake for the level of physical activity). In the early stages of IR, compensatory hyperinsulinemia maintains euglycemia. Progression to impaired glucose tolerance and later diabetes mellitus occurs when β(beta)-cells of the pancreas are no longer able to provide adequate insulin production. Thus, the development of overt diabetes in insulin resistance syndrome depends on the vitality, or lack thereof, of the islet cells, and other target organ damage resulting from hyperinsulinemia may occur in the absence of diabetes mellitus.

Phenotypic manifestations of insulin resistance syndrome are characterized by alterations in all 3 target tissues. In skeletal muscle, IR is associated with decreased glucose uptake (due to impaired translocation of GLUT4), decreased glycogen synthesis, and increased triglyceride accumulation [\[17](#page-9-4)]. In adipocytes, the major effect of IR is increased lipolysis with uncontrolled release of FFAs [\[18](#page-9-5)]. Excess FFAs released by adipose tissue plays a role in mediating hepatic IR both directly by interfering with insulin receptor signaling [\[6](#page-8-5)], or indirectly by promoting increased hepatic triglyceride accumulation

and subsequent hepatic steatosis. Hepatic IR, defined as impaired ability of insulin to suppress hepatic glucose output, can be viewed as a result of impaired response to insulin resulting from mediators released by peripheral tissue (FFAs and adipokines) or from causes within the liver itself (hepatic steatosis). The association of hepatic steatosis with IR may be bidirectional; products released by IR in the peripheral tissue (FFAs) contribute to hepatic steatosis; however, hepatic steatosis itself also contributes to IR.

Hepatic Steatosis: Is Fatty Liver a Cause, a Result, or Simply a Part of IR?

Epidemiologic studies support a direct association between IR and NAFLD [\[19](#page-9-6)[–21](#page-9-7)]. In nondiabetic individuals, hepatic steatosis correlates directly with IR as measured by HOMA-IR [[22\]](#page-9-8). A direct effect of hyperinsulinemia on hepatic steatosis is supported by the observation of a rim of subcapsular hepatic steatosis when insulin is added to peritoneal dialysate [[23\]](#page-9-9) and observation of hepatic steatosis following successful intraportal islet transplantation [[24\]](#page-9-10). This provides evidence of a direct effect of hyperinsulinemia on lipogenesis and is supported by the evidence of increased de novo lipogenesis in individuals with NAFLD [\[25](#page-9-11)]. Insulin promotes lipogenesis through the activation of sterol response elementbinding protein (SREBP)—a major transcription factor that activates genes involved in lipogenesis [\[26](#page-9-12)]. Hepatic steatosis associated with IR also results from excess FFAs released by peripheral lipolysis [[25,](#page-9-11) [27\]](#page-9-13).

Whereas there is a clear association between hepatic steatosis and IR, evidence suggesting that hepatic steatosis may itself cause IR is less clear. Evidence in rats with fatty liver in the absence of peripheral IR suggests a direct correlation between hepatic fat accumulation and hepatic IR through stimulation of gluconeogenesis and impaired activation of glycogen synthase [\[28](#page-9-14)]. In humans, increased liver fat is associated with impairment of insulin-induced suppression of hepatic glucose output, independent of obesity and fat distribution

[\[29](#page-9-15)]. However, hepatic steatosis is not invariably associated with IR. For example, individuals with hepatic steatosis in the setting of familial heterozygous hypobetalipoproteinemia (FHBL) have normal HOMA-IR. In addition, patients with exposure to petrochemicals have been clearly shown to develop fatty liver independent of IR, as measured by HOMA [[30\]](#page-9-16).

Insulin Resistance in Disease States

Nonalcoholic Fatty Liver Disease

NAFLD is common among individuals with obesity and IR [[31,](#page-9-17) [32\]](#page-9-18), and its prevalence in the general population is expected to increase with rising obesity [[33\]](#page-9-19). Pediatric cases of NAFLD are now being increasingly recognized and are an important public health concern [\[34](#page-9-20)]. The term NAFLD is used to include both simple hepatic steatosis without inflammation and NASH. The latter is defined by histologic findings of hepatic steatosis, inflammation, hepatocyte ballooning, and fibrosis [\[35](#page-9-21)], in conjunction with clinical and historical features, notably the absence of significant (>20 g daily) alcohol intake. Steatosis without inflammation is generally felt to carry a benign prognosis [[36\]](#page-9-22), whereas individuals with NASH can progress to cirrhosis and hepatocellular carcinoma [\[37](#page-9-23)]. IR and its metabolic consequences are the fundamental mechanisms leading to hepatic fat accumulation in NAFLD. However, only a subset of individuals with NAFLD has NASH. A "two-hit" hypothesis has been proposed as a pathophysiologic model to account for cellular injury, inflammation, and fibrosis beyond the simple hepatic steatosis observed in the subset of individuals with NASH [\[38](#page-9-24)].

Fat accumulation associated with IR is the "first hit" in NASH and results from a derangement in the physiologic mechanisms designed to maintain energy homeostasis. There is preferential activation of cellular pathways characteristic of calorie/macronutrient deficiency despite being in a state of excess. The liver, skeletal muscles, and adipose tissue all play roles in hepatic steatogenesis and promotion of IR. The resistance of visceral adipocytes to insulin's anti-lipolysis effects drives excess FFA accumulation. Additionally, escalated hepatic de novo lipid synthesis, reduced fatty acid β(beta)-oxidation by hepatocyte mitochondria, and impaired hepatic VLDL export, promote FFA accumulation [[39\]](#page-9-25). The cellular mechanisms by which these paradoxical and maladaptive events occur involve a complex interplay between the insulin/glucagon ratio and the cytokine milieu. These factors together modulate the transcription of the relevant genes and activity of their target enzymes. The sterol regulatory element-binding protein 1c (SREBP 1c), carbohydrate-responsive elementbinding protein (ChREBP), and peroxisome proliferator-activated receptors (PPAR) are three such transcription factors. PPARs promote lipid oxidation via increasing uptake of long-chain fatty acids in skeletal muscle and liver and promoting their β(beta)-oxidation in mitochondria. Hepatic lipogenesis is enhanced via SREBP 1c and ChREBP, which are stimulated by insulin and glucose, respectively [\[40](#page-9-26)]. The biology of SREBP 1c provides a relevant example of the interrelationship between diet, cytokines, and the pathogenesis of NAFLD. Glucose, sterol, and saturated fat consumption upregulates SREBP 1c synthesis, whereas intake of polyunsaturated fats has the opposite effect [\[41](#page-9-27)]. These nuclear transcription factors are also impacted by cytokines released from adipose tissue, the so-called "adipokines." Adiponectin is an insulin-sensitizing adipokine, and animal models have demonstrated an inverse correlation between adiponectin and SREBP 1c levels [[42\]](#page-9-28). The role of adipokines in NAFLD will be discussed in detail below in the context of obesity and IR.

The evolution of simple hepatic steatosis to NASH is characterized histologically by the appearance of cytologic ballooning, necroinflammation, and pericellular fibrosis. The two-hit hypothesis accounts for this transition by proposing additional insults that result in more severe hepatic manifestations of the metabolic syndrome. The inflammatory component of NASH is partially due to macrophage accumulation in visceral adipose tissue. These macrophages secrete pro-inflammatory cytokines, which fur-

ther promote IR, and contribute to trafficking of inflammatory cells to the steatotic liver [[43\]](#page-10-0). The ensuing hepatocyte apoptosis results from oxidative stress related to reactive oxygen species (ROS) accumulation and immune-mediated cytotoxicity. Mitochondrial dysfunction promotes ROS formation, resulting in both cellular necrosis and a self-propagating feedback cycle from ROS stimulation of tumor necrosis factoralpha (TNF- α [alpha]) synthesis. TNF- α (alpha) and other pro-inflammatory cytokines potentiate mitochondrial dysfunction and promote the lymphocyte infiltration typical of NASH [[40\]](#page-9-26).

The Role of Obesity in IR and NAFLD

While obesity is not essential for the development of IR and NAFLD, the strong association among these three conditions may provide some understanding of how the liver, adipose tissue, and other sites of insulin action are related in the development of clinical manifestations observed in the insulin-resistant state. Epidemiologic studies clearly demonstrate a positive correlation between increasing body mass index (BMI) and IR [[44,](#page-10-1) [45\]](#page-10-2). Similarly, a positive association between obesity and NAFLD exists [[46,](#page-10-3) [47\]](#page-10-4). Central to the understanding of how obesity plays a role in IR and NAFLD is the concept that adipose tissue is not only an energy store but also serves a role as an endocrine organ through the release of circulating FFAs and adipokines. One might even go so far as to think of these circulating factors as part of a "vicious cycle" whereby they are both a contributor to, and a result of, IR.

Obesity is associated with elevated circulating FFA levels [[48,](#page-10-5) [49](#page-10-6)] due to increased adipose mass as well as increased lipolysis due to IR. Increased FFA contributes to peripheral IR [[50\]](#page-10-7) via impairment of GLUT-4-mediated glucose transport [\[51](#page-10-8), [52\]](#page-10-9) and hepatic IR through competitive inhibition of IRS 2 signaling by diacylglycerol [\[53](#page-10-10)]. In vitro, FFAs promote hepatic IR by stimulating PEPCK and pyruvate carboxylase [\[54](#page-10-11)], key enzymes in gluconeogenesis, and by increasing the activity of glucose-6-phosphatase, the enzyme responsi-ble for the release of glucose from the liver [[55\]](#page-10-12). In vivo, increased serum FFA levels are associated with increased hepatic gluconeogenesis and decreased glycogenolysis [\[54](#page-10-11), [56](#page-10-13), [57\]](#page-10-14). Increased FFAs associated with obesity and IR result in hepatic steatosis through increased triglyceride formation, increased de novo lipogenesis, and decreased secretion of apolipoprotein B, which results in decreased export of triglycerides out of the hepatocyte as VLDL [\[39](#page-9-25)].

Altered expression of adipokines (leptin, adiponectin, resistin, TNF-α[alpha]) by adipose tissue in obesity also contributes to IR and hepatic steatosis. Adiponectin is a protein with insulin-sensitizing effects that is expressed exclusively by adipocytes in response to PPARγ(gamma) activation. Receptors for adiponectin have been identified in skeletal muscle and liver [[58\]](#page-10-15) and are downregulated in obesitylinked IR and diabetes [[59](#page-10-16)]. Serum adiponectin levels correlate inversely with BMI [[60](#page-10-17), [61](#page-10-18)] and liver fat content [[62–](#page-10-19)[64\]](#page-10-20), suggesting an inhibitory role in obesity and hepatic steatosis. Recombinant adiponectin improves IR in mouse models of obesity and type 2 diabetes [[65](#page-10-21)]. Insulin-sensitizing effects may occur through increased fatty acid oxidation in muscle and decreased fatty acid transport into the liver, resulting in a net decrease in triglyceride accumulation in both muscle and liver [[65\]](#page-10-21). Leptin is a protein expressed by mature adipocytes that acts on the hypothalamus to serve as a signal of energy sufficiency. It is produced in proportion to adipose tissue mass and improves IR and hepatic steatosis in patients with severe lipodystrophy [[66\]](#page-10-22). Serum leptin levels are increased in NASH [[67](#page-10-23)]; however, its role in NASH is debated [[68](#page-10-24)].

Unlike leptin and adiponectin, which are produced exclusively by adipocytes, other adipokines such as $TNF-\alpha$ (alpha) are derived mostly from macrophages in adipose tissue. Increased TNF- α (alpha) levels are found in obese individuals owing to increased macrophage infiltration in adipose tissue [[69](#page-10-25)[–71\]](#page-10-26) and overexpression of TNF-α(alpha) by enlarged adipocytes [[72](#page-10-27)]. Levels are elevated in individuals with NAFLD compared to controls matched for age, BMI, and sex [[73](#page-11-0)]. TNF- α (alpha) impairs insulin signaling through serine phosphorylation of IRS-1 [[74](#page-11-1)].

Lipodystrophy Syndromes

Paradoxically, loss of adipose tissue, as seen in patients with lipodystrophy, is also associated with IR. Lipodystrophies are disorders characterized by selective and variable loss of subcutaneous adipose tissue. They are clinically heterogeneous, and the affected patients are predisposed to IR, hypertriglyceridemia, hepatic steatosis, polycystic ovary syndrome, and type 2 diabetes. Lipodystrophies can be either acquired or inherited (familial).

Acquired lipodystrophies are much more common than the inherited forms. The most common form of acquired lipodystrophy is that seen in patients with human immunodeficiency virus (HIV) infection who are receiving treatment with highly active protease inhibitors. Patients typically present with loss of subcutaneous fat in the face, arms, and legs [\[75](#page-11-2)], with or without concomitant fat accumulation in the neck and trunk [\[76](#page-11-3)]. Patients may develop IR, hypertriglyceridemia, hepatic steatosis, low serum levels of high-density lipoprotein (HDL) cholesterol, and hyperglycemia [\[77](#page-11-4)[–79](#page-11-5)]. Possible mechanisms by which protease inhibitors cause lipodystrophy include impaired pre-adipocyte differentiation [\[80](#page-11-6)], increased apoptosis of subcutaneous adipocytes [[81\]](#page-11-7), and reduced mRNA expression of sterol regulatory elementbinding protein 1c (SREBP1c) and peroxisome proliferator-activated receptor γ (gamma) (PPARγ[gamma])—two key transcription factors involved in adipogenesis [[82\]](#page-11-8). In addition, protease inhibitors may directly induce IR by reducing the intrinsic transport activity of glucose transporter 4 [\[83](#page-11-9)].

Other forms of acquired lipodystrophies are rare [\[84](#page-11-10)]. Patients with acquired generalized lipodystrophy present with clinical features of loss of subcutaneous fat, muscular prominence, acanthosis nigricans, hepatic steatosis, autoimmune hepatitis, and cirrhosis. Patients with acquired partial lipodystrophy have fat loss affecting the face, neck, arms, thorax, and upper abdomen. In contrast, excess fat may be deposited in the hips and legs. Both acquired generalized and partial lipodystrophies occur during childhood and adolescence and occur approximately 3–4 times more often in women. Localized lipodystrophies refer to loss of subcutaneous adipose tissue in small areas and may be caused by local injection of medicines such as insulin and corticosteroids, recurrent pressure, trauma, inflammation, or other unknown mechanisms.

Inherited lipodystrophies are extremely rare and are caused by various genetic mutations [[84\]](#page-11-10). Congenital generalized lipodystrophy is an autosomal recessive disorder characterized by nearcomplete lack of adipose tissue since birth, with clinical features including acanthosis nigricans, hepatic steatosis, cirrhosis, splenomegaly, and umbilical hernia. Patients usually have severe IR/hyperinsulinemia, hypertriglyceridemia, and type 2 diabetes. Familial partial lipodystrophies are characterized by partial loss of subcutaneous fat in various parts of the body with distinct clinical features due to different genetic mutations. Patients may develop hypertriglyceridemia, fatty liver, and type 2 diabetes.

Metabolic complications such as IR, hypertriglyceridemia, hepatic steatosis, and type 2 diabetes increase in frequency and severity with the extent of fat loss. Patients initially develop compensatory hyperinsulinemia and later overt hyperglycemia and type 2 diabetes owing to gradual loss of β(beta)-cell function resulting from islet amyloidosis and cell atrophy [[85\]](#page-11-11). Though the underlying mechanisms remain unclear, it appears that ectopic accumulation of triglycerides may be the major culprit. Indeed, a major function of subcutaneous adipose depot is to store triglycerides during energy excess; and in patients with lipodystrophies the storage capacity decreases or even disappears, leading to the accumulation of excess triglycerides in the liver, intra-abdominal fat depot, and skeletal muscles [\[86](#page-11-12)[–88](#page-11-13)].

A large body of evidence has confirmed that accumulation of fat in these sites leads to IR [[89–](#page-11-14) [92](#page-11-15)]. Mice with congenital lipodystrophy manifest with severe IR in the liver and muscle, and hyperglycemia [\[93](#page-11-16)]. These mice have higher intracellular fatty acyl-CoA content in both muscle and liver than wild-type mice, and have defects in the insulin activation of IRS-1 and IRS-2 associated PI 3-kinase activity in muscle and liver [[94\]](#page-11-17). The importance of having subcutaneous fat depots is further demonstrated by the observation that transplanting adipose tissue from wild-type mice into the subcutaneous space in lipodystrophic mice reduces liver and muscle lipid contents and reverses IR as manifested by increased muscle glucose uptake and suppressed hepatic glucose production in response to insulin [[94\]](#page-11-17).

Patients with severe lipodystrophy have low plasma leptin levels. In mice with congenital lipodystrophy, chronic low-dose leptin treat-ment reverses IR and diabetes mellitus [[95\]](#page-11-18). This suggests that leptin deficiency may play an important role in the pathogenesis of IR and type 2 diabetes in patients with lipodystrophies. Indeed, recent studies have shown that leptin replacement in these patients results in improved insulin sensitivity in both muscle and liver and better glycemic and lipemic control [\[66](#page-10-22), [96,](#page-11-19) [97\]](#page-11-20). Reduced intramyocellular and liver fat contents and reduced appetite could be part of the mechanism [[66,](#page-10-22) [96\]](#page-11-19).

Hepatitis C

IR and the associated steatosis is an emerging aspect of chronic hepatitis C infection. Worsening IR affects chronic hepatitis C in several respects. For example, there appears to be an increased prevalence of type 2 diabetes in hepatitis C patients, especially with increasing age, even when adjusting for confounding variables such as weight [\[98](#page-11-21)], and an association between increased HOMA and chronic genotype 1 HCV infection has been observed [[99\]](#page-11-22). The occurrence of IR and the development of steatosis have a significant impact on the risk of disease progression [\[100](#page-11-23)[–102](#page-11-24)]. Finally, IR appears to have an impact on the response to interferon-based antiviral therapy, although this aspect remains somewhat uncertain, as it is unclear whether this represents the effects of obesity on drug delivery [\[103](#page-11-25)] or direct effects of IR itself. Moreover, more recent studies have failed to confirm results of earlier reports regarding the impact of steatosis and BMI on response, although the association with more

advanced histology seems consistent among studies [\[104](#page-12-0)]. It is possible and even likely that some of these relationships are obscured by the tendency of NASH to become decreasingly steatotic as the disease progresses and that fibrosis stage, which appears to be accelerated by steatosis in HCV, is also a predictor of sustained virological response. This may explain why another recent study revealed an association between steatosis and stage-3 fibrosis, but not stage-4 (cirrhosis) fibrosis $[105]$ $[105]$.

Steatosis when present in HCV is, indeed, often associated with typical findings of NASH in these patients and with increased activation of the hepatic stellate cells [\[106](#page-12-2)]. The mechanisms by which an HCV-infected liver accumulates excessive triglyceride stores are related to both host and viral factors. Not surprisingly, many such patients have independent risk factors for metabolic syndrome and therefore for NAFLD. However, there are also direct viral replication factors related especially to the metabolism of the nucleocapsid core protein and related to (but not restricted to) certain genotypes of the virus. For example, the prevalence of steatosis in genotype 3 is almost 2 times that of other genotypes [[107\]](#page-12-3). However, both in vitro and in vivo data have indicated that the core protein of other genotypes, including genotype 1, the most common genotype in the United States, alters intracellular fat metabolism [\[108](#page-12-4)]. These changes appear to involve significantly decreased levels of $PPAR\alpha(\text{alpha})$ and CPT-1, which therefore inhibit fatty acid oxidation. Other mechanisms may be simultaneously at work, including the indirect effects of increased TNFα(alpha) in HCV and HCV-mediated changes in IR phosphorylation [[109](#page-12-5), [110](#page-12-6)]. The potential role of insulin-sensitizing agents in conjunction with anti-HCV therapy and related concerns about lipid-lowering agents in HCV are areas in need of further investigation [\[111](#page-12-7)].

Summary and Conclusions

Insulin regulates energy homeostasis through its effects on key target organs: liver, adipose tissue, and muscle. Glucose and lipid metabolisms are closely linked via circulating FFAs and adipokines and their effects on insulin receptor signaling, glucose transport, and triglyceride accumulation within these organs. Paradoxically, IR is found in states associated with both adipose excess (obesity) and adipose loss (lipoatrophy). Adipokines released from both adipocytes and macrophages within adipose tissue play key roles in mediating IR and in inflammation. Further understanding of the complex interaction between key target organs and circulating mediators of IR may help guide therapy for NAFLD and other clinical manifestations of the insulin resistance syndrome.

References

- 1. Der NB. Diabetes mellitus. Wien: Nothangels Handbuch. A. Holder; 1906.
- 2. Bajaj M, Defronzo RA. Metabolic and molecular basis of insulin resistance. J Nucl Cardiol. 2003;10(3):311–23.
- 3. Weickert MO, Pfeiffer AF. Signalling mechanisms linking hepatic glucose and lipid metabolism. Diabetologia. 2006;49(8):1732–41.
- 4. Pessin JE, Saltiel AR. Signaling pathways in insulin action: molecular targets of insulin resistance. J Clin Invest. 2000;106(2):165–9.
- 5. Roden M, Bernroider E. Hepatic glucose metabolism in humans – its role in health and disease. Best Pract Res Clin Endocrinol Metab. 2003;17(3):365–83.
- 6. Choudhury J, Sanyal AJ. Insulin resistance and the pathogenesis of nonalcoholic fatty liver disease. Clin Liver Dis. 2004;8(3):575–94, ix.
- 7. White MF. Insulin signaling in health and disease. Science. 2003;302(5651):1710–1.
- 8. Wilcox G. Insulin and insulin resistance. Clin Biochem Rev. 2005;26(2):19–39.
- 9. Kahn CR. Insulin resistance, insulin insensitivity, and insulin unresponsiveness: a necessary distinction. Metabolism. 1978;27(12 Suppl 2):1893–902.
- 10. DeFronzo RA, Tobin JD, Andres R. Glucose clamp technique: a method for quantifying insulin secretion and resistance. Am J Phys. 1979;237(3):E214–23.
- 11. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. Diabetologia. 1985;28(7):412–9.
- 12. Katz A, Nambi SS, Mather K, Baron AD, Follmann DA, Sullivan G, et al. Quantitative insulin sensitivity check index: a simple, accurate method for assessing insulin sensitivity in humans. J Clin Endocrinol Metab. 2000;85(7):2402–10.
- 13. Chen H, Sullivan G, Yue LQ, Katz A, Quon MJ. QUICKI is a useful index of insulin sensitivity in subjects with hypertension. Am J Physiol Endocrinol Metab. 2003;284(4):E804–12.
- 14. Perseghin G, Caumo A, Caloni M, Testolin G, Luzi L. Incorporation of the fasting plasma FFA concentration into QUICKI improves its association with insulin sensitivity in nonobese individuals. J Clin Endocrinol Metab. 2001;86(10):4776–81.
- 15. Quon MJ. Limitations of the fasting glucose to insulin ratio as an index of insulin sensitivity. J Clin Endocrinol Metab. 2001;86(10):4615–7.
- 16. Reaven G. The metabolic syndrome or the insulin resistance syndrome? Different names, different concepts, and different goals. Endocrinol Metab Clin North Am. 2004;33(2):283–303.
- 17. Goodpaster BH, Thaete FL, Simoneau JA, Kelley DE. Subcutaneous abdominal fat and thigh muscle composition predict insulin sensitivity independently of visceral fat. Diabetes. 1997;46(10):1579–85.
- 18. Saltiel AR, Kahn CR. Insulin signalling and the regulation of glucose and lipid metabolism. Nature. 2001;414(6865):799–806.
- 19. Marchesini G, Brizi M, Morselli-Labate AM, Bianchi G, Bugianesi E, McCullough AJ, et al. Association of nonalcoholic fatty liver disease with insulin resistance. Am J Med. 1999;107(5):450–5.
- 20. Sanyal AJ, Campbell-Sargent C, Mirshahi F, Rizzo WB, Contos MJ, Sterling RK, et al. Nonalcoholic steatohepatitis: association of insulin resistance and mitochondrial abnormalities. Gastroenterology. 2001;120(5):1183–92.
- 21. Chitturi S, Farrell G, Frost L, Kriketos A, Lin R, Fung C, et al. Serum leptin in NASH correlates with hepatic steatosis but not fibrosis: a manifestation of lipotoxicity? Hepatology. 2002;36(2):403–9.
- 22. Angelico F, Del Ben M, Conti R, Francioso S, Feole K, Fiorello S, et al. Insulin resistance, the metabolic syndrome, and nonalcoholic fatty liver disease. J Clin Endocrinol Metab. 2005;90(3):1578–82.
- 23. Wanless IR, Bargman JM, Oreopoulos DG, Vas SI. Subcapsular steatonecrosis in response to peritoneal insulin delivery: a clue to the pathogenesis of steatonecrosis in obesity. Mod Pathol. 1989;2(2):69–74.
- 24. Bhargava R, Senior PA, Ackerman TE, Ryan EA, Paty BW, Lakey JR, et al. Prevalence of hepatic steatosis after islet transplantation and its relation to graft function. Diabetes. 2004;53(5):1311–7.
- 25. Donnelly KL, Smith CI, Schwarzenberg SJ, Jessurun J, Boldt MD, Parks EJ. Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. J Clin Invest. 2005;115(5):1343–51.
- 26. Horton JD, Shah NA, Warrington JA, Anderson NN, Park SW, Brown MS, et al. Combined analysis of oligonucleotide microarray data from transgenic and knockout mice identifies direct SREBP target genes. Proc Natl Acad Sci USA. 2003;100(21):12027–32.
- 27. Bugianesi E, Gastaldelli A, Vanni E, Gambino R, Cassader M, Baldi S, et al. Insulin resistance in non-diabetic patients with non-alcoholic fatty liver disease: sites and mechanisms. Diabetologia. 2005;48(4):634–42.
- 28. Samuel VT, Liu ZX, Qu X, Elder BD, Bilz S, Befroy D, et al. Mechanism of hepatic insulin resistance in non-alcoholic fatty liver disease. J Biol Chem. 2004;279(31):32345–53.
- 29. Seppala-Lindroos A, Vehkavaara S, Hakkinen AM, Goto T, Westerbacka J, Sovijarvi A, et al. Fat accumulation in the liver is associated with defects in insulin suppression of glucose production and serum free fatty acids independent of obesity in normal men. J Clin Endocrinol Metab. 2002;87(7):3023–8.
- 30. Cotrim HP, Carvalho F, Siqueira AC, Lordelo M, Rocha R, De Freitas LA. Nonalcoholic fatty liver and insulin resistance among petrochemical workers. JAMA. 2005;294(13):1618–20.
- 31. Gupte P, Amarapurkar D, Agal S, Baijal R, Kulshrestha P, Pramanik S, et al. Non-alcoholic steatohepatitis in type 2 diabetes mellitus. J Gastroenterol Hepatol. 2004;19(8):854–8.
- 32. Machado M, Marques-Vidal P, Cortez-Pinto H. Hepatic histology in obese patients undergoing bariatric surgery. J Hepatol. 2006;45(4):600–6.
- 33. Ogden CL, Yanovski SZ, Carroll MD, Flegal KM. The epidemiology of obesity. Gastroenterology. 2007;132(6):2087–102.
- 34. Schwimmer JB. Definitive diagnosis and assessment of risk for nonalcoholic fatty liver disease in children and adolescents. Semin Liver Dis. 2007;27(3):312–8.
- 35. Brunt EM. Nonalcoholic steatohepatitis: definition and pathology. Semin Liver Dis. 2001;21(1):3–16.
- 36. Dam-Larsen S, Franzmann M, Andersen IB, Christoffersen P, Jensen LB, Sorensen TI, et al. Long term prognosis of fatty liver: risk of chronic liver disease and death. Gut. 2004;53(5):750–5.
- 37. Ekstedt M, Franzen LE, Mathiesen UL, Thorelius L, Holmqvist M, Bodemar G, et al. Long-term follow-up of patients with NAFLD and elevated liver enzymes. Hepatology. 2006;44(4):865–73.
- 38. Day CP, James OF. Steatohepatitis: a tale of two "hits"? Gastroenterology. 1998;114(4):842–5.
- 39. Parekh S, Anania FA. Abnormal lipid and glucose metabolism in obesity: implications for nonalcoholic fatty liver disease. Gastroenterology. 2007;132(6):2191–207.
- 40. Sanyal AJ. Mechanisms of disease: pathogenesis of nonalcoholic fatty liver disease. Nat Clin Pract Gastroenterol Hepatol. 2005;2(1):46–53.
- 41. Mater MK, Thelen AP, Pan DA, Jump DB. Sterol response element-binding protein 1c (SREBP1c) is involved in the polyunsaturated fatty acid suppression of hepatic S14 gene transcription. J Biol Chem. 1999;274(46):32725–32.
- 42. Schaffler A, Scholmerich J, Buchler C. Mechanisms of disease: adipocytokines and visceral adipose tissue – emerging role in nonalcoholic fatty liver

disease. Nat Clin Pract Gastroenterol Hepatol. 2005;2(6):273–80.

- 43. Shoelson SE, Herrero L, Naaz A. Obesity, inflammation, and insulin resistance. Gastroenterology. 2007;132(6):2169–80.
- 44. Chan JM, Rimm EB, Colditz GA, Stampfer MJ, Willett WC. Obesity, fat distribution, and weight gain as risk factors for clinical diabetes in men. Diabetes Care. 1994;17(9):961–9.
- 45. Colditz GA, Willett WC, Rotnitzky A, Manson JE. Weight gain as a risk factor for clinical diabetes mellitus in women. Ann Intern Med. 1995;122(7):481–6.
- 46. Silverman JF, O'Brien KF, Long S, Leggett N, Khazanie PG, Pories WJ, et al. Liver pathology in morbidly obese patients with and without diabetes. Am J Gastroenterol. 1990;85(10):1349–55.
- 47. Wanless IR, Lentz JS. Fatty liver hepatitis (steatohepatitis) and obesity: an autopsy study with analysis of risk factors. Hepatology. 1990;12(5):1106–10.
- 48. Gordon ES. Lipid metabolism, diabetes mellitus, and obesity. Adv Intern Med. 1964;12:66–102.
- 49. Bjorntorp P, Bergman H, Varnauskas E. Plasma free fatty acid turnover rate in obesity. Acta Med Scand. 1969;185(4):351–6.
- 50. Boden G, Chen X. Effects of fat on glucose uptake and utilization in patients with non-insulin-dependent diabetes. J Clin Invest. 1995;96(3):1261–8.
- 51. Dresner A, Laurent D, Marcucci M, Griffin ME, Dufour S, Cline GW, et al. Effects of free fatty acids on glucose transport and IRS-1-associated phosphatidylinositol 3-kinase activity. J Clin Invest. 1999;103(2):253–9.
- 52. Shulman GI. Cellular mechanisms of insulin resistance. J Clin Invest. 2000;106(2):171–6.
- 53. Petersen KF, Shulman GI. Etiology of insulin resistance. Am J Med. 2006;119(5 Suppl 1):S10–6.
- 54. Bahl JJ, Matsuda M, DeFronzo RA, Bressler R. In vitro and in vivo suppression of gluconeogenesis by inhibition of pyruvate carboxylase. Biochem Pharmacol. 1997;53(1):67–74.
- 55. Massillon D, Barzilai N, Hawkins M, Prus-Wertheimer D, Rossetti L. Induction of hepatic glucose-6-phosphatase gene expression by lipid infusion. Diabetes. 1997;46(1):153–7.
- 56. Boden G, Cheung P, Stein TP, Kresge K, Mozzoli M. FFA cause hepatic insulin resistance by inhibiting insulin suppression of glycogenolysis. Am J Physiol Endocrinol Metab. 2002;283(1):E12–9.
- 57. Roden M, Stingl H, Chandramouli V, Schumann WC, Hofer A, Landau BR, et al. Effects of free fatty acid elevation on postabsorptive endogenous glucose production and gluconeogenesis in humans. Diabetes. 2000;49(5):701–7.
- 58. Yamauchi T, Kamon J, Ito Y, Tsuchida A, Yokomizo T, Kita S, et al. Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. Nature. 2003;423(6941):762–9.
- 59. Kadowaki T, Yamauchi T, Kubota N, Hara K, Ueki K, Tobe K. Adiponectin and adiponectin receptors in

insulin resistance, diabetes, and the metabolic syndrome. J Clin Invest. 2006;116(7):1784–92.

- 60. Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J, et al. Paradoxical decrease of an adiposespecific protein, adiponectin, in obesity. Biochem Biophys Res Commun. 1999;257(1):79–83.
- 61. Matsubara M, Maruoka S, Katayose S. Inverse relationship between plasma adiponectin and leptin concentrations in normal-weight and obese women. Eur J Endocrinol. 2002;147(2):173–80.
- 62. Westerbacka J, Corner A, Tiikkainen M, Tamminen M, Vehkavaara S, Hakkinen AM, et al. Women and men have similar amounts of liver and intraabdominal fat, despite more subcutaneous fat in women: implications for sex differences in markers of cardiovascular risk. Diabetologia. 2004;47(8):1360–9.
- 63. Bajaj M, Suraamornkul S, Piper P, Hardies LJ, Glass L, Cersosimo E, et al. Decreased plasma adiponectin concentrations are closely related to hepatic fat content and hepatic insulin resistance in pioglitazonetreated type 2 diabetic patients. J Clin Endocrinol Metab. 2004;89(1):200–6.
- 64. Bugianesi E, Pagotto U, Manini R, Vanni E, Gastaldelli A, de Iasio R, et al. Plasma adiponectin in nonalcoholic fatty liver is related to hepatic insulin resistance and hepatic fat content, not to liver disease severity. J Clin Endocrinol Metab. 2005;90(6):3498–504.
- 65. Yamauchi T, Kamon J, Waki H, Terauchi Y, Kubota N, Hara K, et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. Nat Med. 2001;7(8):941–6.
- 66. Petersen KF, Oral EA, Dufour S, Befroy D, Ariyan C, Yu C, et al. Leptin reverses insulin resistance and hepatic steatosis in patients with severe lipodystrophy. J Clin Invest. 2002;109(10):1345–50.
- 67. Uygun A, Kadayifci A, Yesilova Z, Erdil A, Yaman H, Saka M, et al. Serum leptin levels in patients with nonalcoholic steatohepatitis. Am J Gastroenterol. 2000;95(12):3584–9.
- 68. Chalasani N, Crabb DW, Cummings OW, Kwo PY, Asghar A, Pandya PK, et al. Does leptin play a role in the pathogenesis of human nonalcoholic steatohepatitis? Am J Gastroenterol. 2003;98(12):2771–6.
- 69. Permana PA, Menge C, Reaven PD. Macrophagesecreted factors induce adipocyte inflammation and insulin resistance. Biochem Biophys Res Commun. 2006;341(2):507–14.
- 70. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr. Obesity is associated with macrophage accumulation in adipose tissue. J Clin Invest. 2003;112(12):1796–808.
- 71. Xu H, Barnes GT, Yang Q, Tan G, Yang D, Chou CJ, et al. Chronic inflammation in fat plays a crucial role in the development of obesity-related insulin resistance. J Clin Invest. 2003;112(12):1821–30.
- 72. Peraldi P, Spiegelman B. TNF-alpha and insulin resistance: summary and future prospects. Mol Cell Biochem. 1998;182(1–2):169–75.
- 73. Hui JM, Hodge A, Farrell GC, Kench JG, Kriketos A, George J. Beyond insulin resistance in NASH: TNF-alpha or adiponectin? Hepatology. 2004;40(1):46–54.
- 74. Hotamisligil GS. The role of TNFalpha and TNF receptors in obesity and insulin resistance. J Intern Med. 1999;245(6):621–5.
- 75. Carr A, Samaras K, Burton S, Law M, Freund J, Chisholm DJ, et al. A syndrome of peripheral lipodystrophy, hyperlipidaemia and insulin resistance in patients receiving HIV protease inhibitors. AIDS. 1998;12(7):F51–8.
- 76. Lo JC, Mulligan K, Tai VW, Algren H, Schambelan M. Body shape changes in HIV-infected patients. J Acquir Immune Defic Syndr Hum Retrovirol. 1998;19(3):307–8.
- 77. Carr A, Samaras K, Thorisdottir A, Kaufmann GR, Chisholm DJ, Cooper DA. Diagnosis, prediction, and natural course of HIV-1 protease-inhibitor-associated lipodystrophy, hyperlipidaemia, and diabetes mellitus: a cohort study. Lancet. 1999;353(9170):2093–9.
- 78. Dube MP, Johnson DL, Currier JS, Leedom JM. Protease inhibitor-associated hyperglycaemia. Lancet. 1997;350(9079):713–4.
- 79. Sutinen J, Hakkinen AM, Westerbacka J, Seppala-Lindroos A, Vehkavaara S, Halavaara J, et al. Increased fat accumulation in the liver in HIV-infected patients with antiretroviral therapy-associated lipodystrophy. AIDS. 2002;16(16):2183–93.
- 80. Dowell P, Flexner C, Kwiterovich PO, Lane MD. Suppression of preadipocyte differentiation and promotion of adipocyte death by HIV protease inhibitors. J Biol Chem. 2000;275(52):41325–32.
- 81. Domingo P, Matias-Guiu X, Pujol RM, Francia E, Lagarda E, Sambeat MA, et al. Subcutaneous adipocyte apoptosis in HIV-1 protease inhibitor-associated lipodystrophy. AIDS. 1999;13(16):2261–7.
- 82. Bastard JP, Caron M, Vidal H, Jan V, Auclair M, Vigouroux C, et al. Association between altered expression of adipogenic factor SREBP1 in lipoatrophic adipose tissue from HIV-1-infected patients and abnormal adipocyte differentiation and insulin resistance. Lancet. 2002;359(9311):1026–31.
- 83. Murata H, Hruz PW, Mueckler M. The mechanism of insulin resistance caused by HIV protease inhibitor therapy. J Biol Chem. 2000;275(27):20251–4.
- 84. Garg A. Lipodystrophies. Am J Med. 2000;108(2):143–52.
- 85. Garg A, Chandalia M, Vuitch F. Severe islet amyloidosis in congenital generalized lipodystrophy. Diabetes Care. 1996;19(1):28–31.
- 86. Chandalia M, Garg A, Vuitch F, Nizzi F. Postmortem findings in congenital generalized lipodystrophy. J Clin Endocrinol Metab. 1995;80(10):3077–81.
- 87. Haque WA, Vuitch F, Garg A. Post-mortem findings in familial partial lipodystrophy, Dunnigan variety. Diabet Med. 2002;19(12):1022–5.
- 88. Szczepaniak LS, Babcock EE, Schick F, Dobbins RL, Garg A, Burns DK, et al. Measurement of intracellu-

lar triglyceride stores by H spectroscopy: validation in vivo. Am J Phys. 1999;276(5 Pt 1):E977–89.

- 89. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. Nature. 2006;444(7121):840–6.
- 90. Kelley DE, Goodpaster BH, Storlien L. Muscle triglyceride and insulin resistance. Annu Rev Nutr. 2002;22:325–46.
- 91. Savage DB, Petersen KF, Shulman GI. Disordered lipid metabolism and the pathogenesis of insulin resistance. Physiol Rev. 2007;87(2):507–20.
- 92. Caldwell SH, Ikura Y, Iezzoni JC, Liu Z. Has natural selection in human populations produced two types of metabolic syndrome (with and without fatty liver)? J Gastroenterol Hepatol. 2007;22(Suppl 1):S11–9.
- 93. Moitra J, Mason MM, Olive M, Krylov D, Gavrilova O, Marcus-Samuels B, et al. Life without white fat: a transgenic mouse. Genes Dev. 1998;12(20):3168–81.
- 94. Kim JK, Gavrilova O, Chen Y, Reitman ML, Shulman GI. Mechanism of insulin resistance in A-ZIP/F-1 fatless mice. J Biol Chem. 2000;275(12):8456–60.
- 95. Shimomura I, Hammer RE, Ikemoto S, Brown MS, Goldstein JL. Leptin reverses insulin resistance and diabetes mellitus in mice with congenital lipodystrophy. Nature. 1999;401(6748):73–6.
- 96. Oral EA, Simha V, Ruiz E, Andewelt A, Premkumar A, Snell P, et al. Leptin-replacement therapy for lipodystrophy. N Engl J Med. 2002;346(8):570–8.
- 97. Simha V, Szczepaniak LS, Wagner AJ, DePaoli AM, Garg A. Effect of leptin replacement on intrahepatic and intramyocellular lipid content in patients with generalized lipodystrophy. Diabetes Care. 2003;26(1):30–5.
- 98. Mehta SH, Brancati FL, Sulkowski MS, Strathdee SA, Szklo M, Thomas DL. Prevalence of type 2 diabetes mellitus among persons with hepatitis C virus infection in the United States. Ann Intern Med. 2000;133(8):592–9.
- 99. Fartoux L, Poujol-Robert A, Guechot J, Wendum D, Poupon R, Serfaty L. Insulin resistance is a cause of steatosis and fibrosis progression in chronic hepatitis C. Gut. 2005;54(7):1003–8.
- 100. Ortiz V, Berenguer M, Rayon JM, Carrasco D, Berenguer J. Contribution of obesity to hepatitis C-related fibrosis progression. Am J Gastroenterol. 2002;97(9):2408–14.
- 101. Adinolfi LE, Gambardella M, Andreana A, Tripodi MF, Utili R, Ruggiero G. Steatosis accelerates the progression of liver damage of chronic hepatitis C patients and correlates with specific HCV genotype and visceral obesity. Hepatology. 2001;33(6):1358–64.
- 102. Clouston AD, Jonsson JR, Purdie DM, Macdonald GA, Pandeya N, Shorthouse C, et al. Steatosis and chronic hepatitis C: analysis of fibrosis and stellate cell activation. J Hepatol. 2001;34(2):314–20.
- 103. Bressler BL, Guindi M, Tomlinson G, Heathcote J. High body mass index is an independent risk fac-

tor for nonresponse to antiviral treatment in chronic hepatitis C. Hepatology. 2003;38(3):639–44.

- 104. Hu KQ, Currie SL, Shen H, Cheung RC, Ho SB, Bini EJ, et al. Clinical implications of hepatic steatosis in patients with chronic hepatitis C: a multicenter study of US veterans. Dig Dis Sci. 2007;52(2):570–8.
- 105. Lok AS, Everhart JE, Chung RT, Padmanabhan L, Greenson JK, Shiffman ML, et al. Hepatic steatosis in hepatitis C: comparison of diabetic and nondiabetic patients in the hepatitis C antiviral long-term treatment against cirrhosis trial. Clin Gastroenterol Hepatol. 2007;5(2):245–54.
- 106. Sanyal AJ. Review article: non-alcoholic fatty liver disease and hepatitis $C -$ risk factors and clinical implications. Aliment Pharmacol Ther. 2005;22(Suppl 2):48–51.
- 107. Patton HM, Patel K, Behling C, Bylund D, Blatt LM, Vallee M, et al. The impact of steatosis on disease progression and early and sustained treatment

response in chronic hepatitis C patients. J Hepatol. 2004;40(3):484–90.

- 108. Dharancy S, Malapel M, Perlemuter G, Roskams T, Cheng Y, Dubuquoy L, et al. Impaired expression of the peroxisome proliferator-activated receptor alpha during hepatitis C virus infection. Gastroenterology. 2005;128(2):334–42.
- 109. Shintani Y, Fujie H, Miyoshi H, Tsutsumi T, Tsukamoto K, Kimura S, et al. Hepatitis C virus infection and diabetes: direct involvement of the virus in the development of insulin resistance. Gastroenterology. 2004;126(3):840–8.
- 110. Aytug S, Reich D, Sapiro LE, Bernstein D, Begum N. Impaired IRS-1/PI3-kinase signaling in patients with HCV: a mechanism for increased prevalence of type 2 diabetes. Hepatology. 2003;38(6):1384–92.
- 111. Lonardo A, Loria P, Bertolotti M, Carulli N. Statins and HCV: a complex issue. Hepatology. 2007;45(1):257.