



# Age, Sex and Racial Differences in Cardiac Repolarization and Arrhythmogenesis

Arja Suzanne Vink, Sally-Ann B. Clur, Pieter G. Postema, Nico A. Blom, and Arthur A. M. Wilde

## Introduction

Willem Einthoven recorded the first electrocardiogram (ECG) in a healthy man in 1905, long before the understanding of the role of ion channels and the cardiac action potential. Over the last decades, extensive progress has been made in our understanding of the relationship between structure and

function of the cardiac ion channels and the effects of changes in expression and gating of these channels on the electrical substrate and the ECG. The function of ion channels is significantly modified by the subunit assembly and environmental conditions (i.e. hormones, electrolyte concentrations and pH), which have substantial effects on the cardiac depolarization and repolarization. Profound ECG changes in individuals with primary arrhythmia syndromes have helped to discover the role of subunits of ion channels in causing different types of cardiac channelopathies. Indeed, pathogenic mutations in the genes encoding these subunits are causal to some of these rare arrhythmia syndromes. However, despite this improved understanding of the physiology of the cardiac ion channels, there is still a lack of knowledge on age, sex and racial differences seen in the human ECG especially regarding the QTc-interval. This is due to incomplete and controversial information on age, sex and racial differences in the expression and properties of ion channels and the regulation of ion channels, i.e. by sex hormones.

This chapter outlines the recent developments in the study of age, sex and racial differences in QTc-interval in healthy individuals.

A. S. Vink (✉)

Department of Clinical and Experimental Cardiology, Amsterdam UMC, University of Amsterdam, Heart Center, Amsterdam, The Netherlands

Department of Pediatric Cardiology, Emma Children's Hospital, Amsterdam UMC, University of Amsterdam, Amsterdam, The Netherlands  
e-mail: [a.s.vink@amsterdamumc.nl](mailto:a.s.vink@amsterdamumc.nl)

S.-A. B. Clur

Department of Pediatric Cardiology, Emma Children's Hospital, Amsterdam UMC, University of Amsterdam, Amsterdam, The Netherlands  
e-mail: [s.a.clur@amsterdamumc.nl](mailto:s.a.clur@amsterdamumc.nl)

P. G. Postema · A. A. M. Wilde

Department of Clinical and Experimental Cardiology, Amsterdam UMC, University of Amsterdam, Heart Center, Amsterdam, The Netherlands  
e-mail: [p.g.postema@amsterdamumc.nl](mailto:p.g.postema@amsterdamumc.nl);  
[a.a.wilde@amsterdamumc.nl](mailto:a.a.wilde@amsterdamumc.nl)

N. A. Blom

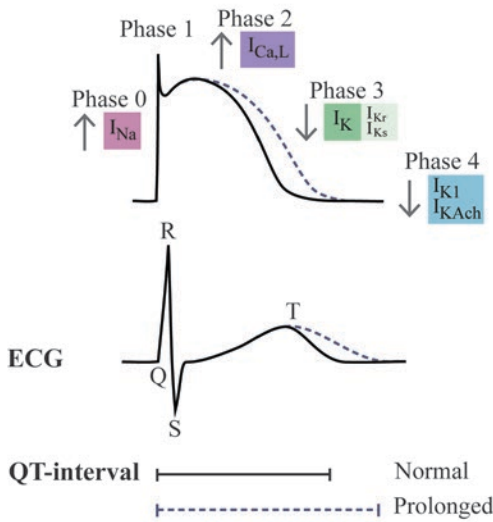
Department of Pediatric Cardiology, Emma Children's Hospital, Amsterdam UMC, University of Amsterdam, Amsterdam, The Netherlands

Department of Paediatric Cardiology, Willem-Alexander Children's Hospital, Leiden University Medical Centre, Leiden, The Netherlands  
e-mail: [n.a.blom@amsterdamumc.nl](mailto:n.a.blom@amsterdamumc.nl)

## General Aspects of Cardiac Repolarization

The role of cardiac ion channels in the generation of the ventricular cardiac action potential is

### Ventricular action potential



**Fig. 6.1** Electrophysiological basis of the ventricular action potential and prolongation of the QT-interval. See text for explanation

outlined in depth in Chap. 1. In short, the rapid depolarization (phase 0) is caused by the influx of sodium ions into the cell through voltage-gated sodium channels ( $I_{Na}$ ) (Fig. 6.1). Phase 1 repolarization is mainly caused by activation of the transient outward potassium currents ( $I_{to}$ ) together with a corresponding rapid decay of the sodium current, which is followed by phase 2. In this plateau phase, continued L-type late calcium ( $I_{Ca,L}$ ) and a small amplitude late sodium current into the cell balance the effect of potassium currents out of the cell. The decay of the calcium current and the increase in delayed rectifier potassium current ( $I_{Ks}$ ), in the rapid activation component of the delayed rectifier potassium current ( $I_{Kr}$ ), and particularly the late activation of the inward rectifier potassium current ( $I_{K1}$ ), are together responsible for the repolarization (phase 3) with ultimate return to the resting potential (phase 4).

The vectorial sum of the complex interactions of these different electrical currents in all the cardiomyocytes results in the physical manifestation of the cardiac waveform morphologies on the ECG that consist of a QRS complex and a T-wave.

The QT-interval is measured from the beginning of the QRS complex to the end of the T-wave and represents the duration of activation and recovery of the ventricular myocardium. The QT-interval is most affected by alterations in phase 2 and phase 3 of the ventricular action potential [1]. Upregulation of the  $I_{Ca,L}$  channel currents prolongs the QT-interval, whereas downregulation shortens the QT-interval. During phases 2 and 3, upregulation of  $I_{Ks}$ ,  $I_{Kr}$  and  $I_{K1}$  channel currents shorten the QT-interval, and downregulation prolong the QT-interval. Finally, an increased amplitude of the late sodium current lengthens the action potential and thus the QT-interval.

In the normal heart, the QT-interval shortens with an increase in heart rate and lengthens with a decrease in heart rate. Since the QT-interval adapts to the heart rate, the QT-interval should be corrected for heart rate using the preceding RR-interval (QTc-interval), which can be done using several formulas [2]. None of these formulas gives an optimal correction, but the Bazett correction formula is most frequently used in daily practice [3]. A prolonged QTc-interval is a marker for an increased risk of torsades de pointes (TdP), a malignant polymorphic ventricular tachyarrhythmia that precipitates syncope, sudden cardiac arrest (SCA) or sudden cardiac death (SCD) [4, 5].

### Age-Related QTc-Interval Changes in Healthy Individuals

The QTc-interval is remarkably long after birth, in both males and females [6–8]. The QTc-interval then decreases during the first weeks after birth in both sexes [6–8] and then remains relatively consistent over the years until the age of approximately 16 years [6, 8–14]. However, when individual age trends are taking into account, both males and females have longer QTc-intervals at the age of 12 years compared to the ages of 6 and 15 years [15, 16]. After puberty, the QTc-interval shortens in males but not in females [6, 17–21]. During adulthood, the QTc-interval gradually increases with age [11, 22–33]

in both sexes [17–20, 22, 34–45]. In contradiction, there is one study that showed QTc-interval shortening in older age groups (>60 years) [46].

---

### Sex Differences in QTc-Interval in Healthy Individuals

Nearly 100 years ago in the early ECG recordings by Bazett [3], differences in QTc-interval were described between healthy adult males and females. Decades later, it became clear that these sex differences are not present during the first month of life [6, 8, 47, 48] but arise at a later age. At the age of 1–3 months, females have a slightly longer QTc-interval compared to males [6, 13, 14]. Thereafter, no sex difference is seen until approximately the onset of puberty [6, 9, 10, 12–14, 16, 18, 20, 49, 50]. Fukushima et al. [15] described a shorter QTc-interval in 6-year-old females compared to males, but this difference was very small (384 ms versus 386 ms;  $P < 0.05$ ). After puberty, females have a longer QTc-interval compared to males [6, 9, 14–17, 20, 21, 50–59]. This difference has also been reported in adulthood by several studies [3, 15, 17, 19, 20, 22–27, 29–33, 35–46, 57, 60–74], although some other studies found no difference [34, 66] or even a longer QTc-interval in males [75]. The difference in QTc-interval between males and females decreases with age [25] because the QTc-interval increases more in time in males compared to females [37]. As a result, no clear sex differences are present in the highest age groups of approximately >60 years [19, 20, 27, 34, 37].

---

### Racial Differences in QTc-Interval in Healthy Individuals

The presence of racial differences in QTc-interval still requires clarification as the effect of race has been looked at in only a limited number of studies. Most studies report no clear racial differences [29, 48, 70, 76], although some small differences may be present. Blacks tend to have slightly shorter QTc-intervals compared to Caucasians [8, 31, 32, 74], and Asians, especially females [31,

62, 72, 73, 77], have slightly longer QTc-intervals compared to Caucasians [30, 48, 72, 78]. In addition, collective consideration of available pharmacogenetic and clinical information suggests that there may be inter-race differences in QT-prolonging effects of drugs and that Caucasians may be more sensitive than other populations [79]. These possible differences are most likely the result of the presence of considerable heterogeneity among race/ethnicity for multiple genetic loci that have an impact on the QT-interval. Some of these loci show a striking difference (i.e. order of magnitude 40–50%) between the highest and lowest frequencies between ethnicities [80]. Unfortunately, there is no data on racial-specific age-related sex differences.

---

### Possible Role of Sex Hormones on the QTc-Interval and Arrhythmogenesis

The age- and sex-related differences in QTc-interval, especially the change post-puberty, most likely are due to changes in sex hormones levels. The mechanisms underlying the influence of sex hormones on the repolarization are complex and still unresolved; however, mechanistic studies suggest that sex hormone has varying effects on the  $I_{CaL}$ ,  $I_{Kr}$ ,  $I_{Ks}$  and  $I_{K1}$  channel currents. Testosterone decreases the  $I_{CaL}$  current and increases the potassium channel currents, resulting in a shorter QTc-interval observed in both animal and human studies [81]. Progesterone decreases the  $I_{CaL}$  current and increases the  $I_{Ks}$  current and may therefore shorten the QTc-interval [81]. Conflicting results of endogenous oestrogen on the QTc-interval have been described. Oestrogen lengthens the QTc-interval in animals; however, this has not been supported by human studies. In animal studies, oestrogen decreases the potassium channel currents and may lengthen the QT-interval through this mechanism [81].

In children, concentrations of sex hormones are influenced by the activity of the hypothalamic-pituitary-gonadal (HPG) axis. The HPG axis is active during the (I) mid-gestational period in the

foetus, (II) first months of life and (III) pubertal period [82], and therefore higher concentrations of testosterone and oestrogen are found during these periods. As a consequence, during periods of sudden changes in sex hormone concentrations (i.e. the first months of life and the onset of puberty), a marked QTc-interval shortening in males would be expected based on the higher level of testosterone compared to females. This could explain the shorter QTc-intervals in males between 1 and 3 months and after the onset of puberty compared to females.

In adulthood, the level of testosterone gradually decreases with age in males [83], potentially explaining why with ageing, the QTc-interval in males gradually lengthens and approximates that of females. Changes in sex hormone levels in females are more complex due to the influence of the menstrual cycle, pregnancy and the menopause.

## Menstrual Cycle

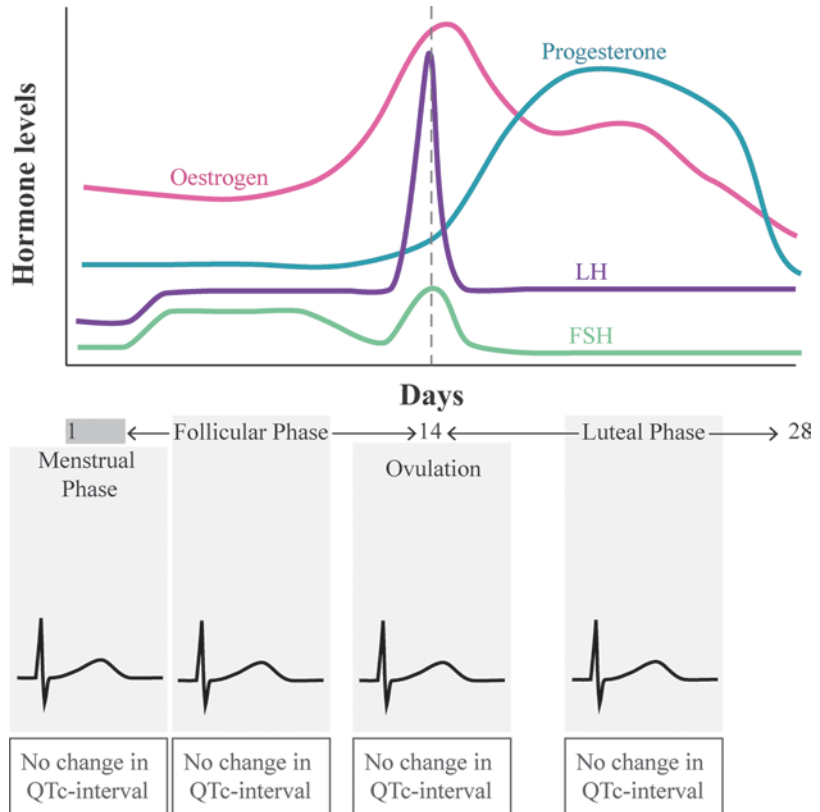
Oestrogen and progesterone levels fluctuate during the menstrual cycle and are lowest at the onset of menses (menstrual phase). After the cessation of the menstrual flow, there is a gradual increase in oestrogen (follicular phase). Oestrogen levels peak in the middle of the menstrual phase during the ovulation (ovulation phase), and after the ovulation, the oestrogen levels gradually decrease during the luteal phase. Progesterone levels, in contrast, are low during the menstrual and follicular phase and increase after the ovulation through the luteal phase (Fig. 6.2).

In females, there are no clear QTc-interval differences between the different phases of the menstrual cycle [84–89]. In addition, there are also no distinct differences in heart rate [86, 87, 89–92], so there is probably no effect of the method chosen to correct the QT-interval for the heart rate. Hulot et al. [84] found no relationship between the level of oestrogen and the length of the QTc-interval ( $P = 0.92$ ). In the study by Nakagawa et al. [85], the uncorrected QT-interval was shorter in the luteal phase compared to the follicular phase, although no difference in QTc-interval

was seen. They found no statistically significant difference in oestrogen level between the luteal and follicular phase, while progesterone ( $P < 0.001$ ) and noradrenaline levels ( $P < 0.05$ ) were higher in the luteal phase suggesting the role of progesterone and/or autonomic tone on the uncorrected QT-interval. When double autonomic blockade was given, e.g. atropine and propranolol administration, QTc-interval differences between the phases of the menstrual cycle were seen. Under these circumstances, Burke et al. [86] found a shorter QTc-interval in the luteal phase compared to the menstrual and follicular phase, whereas Endres et al. [87] observed a significantly longer QTc-interval during the follicular phase compared to the menstrual and luteal phase. Both studies indicate the important role of autonomic tone in alternations in the QTc-interval during the menstrual cycle. Rodriguez et al. [88] showed that ibutilide infusion, a known  $I_{Kr}$  blocker mimicking long QT syndrome (LQTS), causes differences in QTc-interval between the phases of the menstrual cycle, whereas the luteal phase seemed to be protective against the drug-induced QTc-interval prolongation. Hence, since there is an effect of the menstrual cycle on the QTc-interval during administration of QTc-interval prolonging medication, it could be argued that this effect is also present in patients with LQTS.

Despite the unclear changes in the QTc-interval between the phases of the menstrual cycle in the normal situation, there seems to be a cyclic variation in the occurrence of episodes of arrhythmia with the menstrual cycle. During the luteal phase, an increase in the number and duration of paroxysmal supraventricular tachycardia (SVT) has been reported compared to other phases of the menstrual cycle [93, 94], with a correlation with plasma concentrations of progesterone and an inverse correlation with plasma concentrations of oestradiol [93]. Also ventricular ectopic beats seems to be more frequent during the luteal phase of the menstrual cycle [93]. So it seems that there can be significant changes in the arrhythmogenic substrate of the heart throughout the menstrual cycle without dramatic changes in the QTc-interval. As we know, there is no simple relationship between absolute QTc-

**Fig. 6.2** Oestrogen, progesterone, luteinizing hormone (LH) and follicle-stimulating hormone (FSH) levels over a single menstrual cycle in females together with a schematic representation of the QTc-intervals during the phases of the menstrual cycle (i.e. menstrual phase, follicular phase, ovulation and luteal phase). See text for explanation. (Derived from data in: Sedlak et al. [81])



interval and arrhythmogenic potential or SCD [63, 88, 95], so perhaps the autonomic tone also plays an important role which is influenced by sex hormones [85, 96].

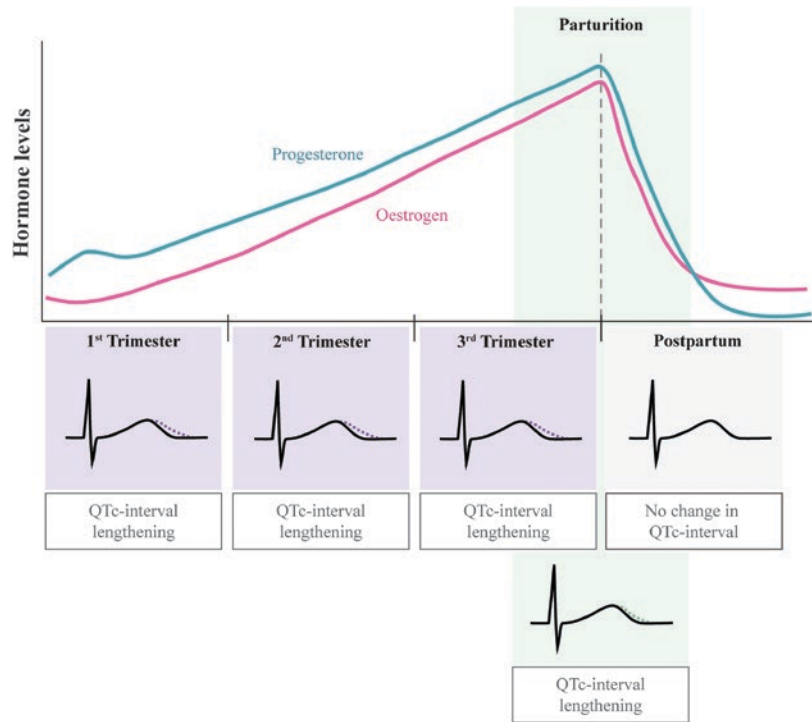
The fluctuations of oestrogen and progesterone levels during the menstrual cycle are controlled by the gonadotropin hormones, e.g. luteinizing hormone (LH) and follicle-stimulating hormone (FSH). A peak in LH and FSH is seen prior to ovulation in the normal menstrual cycle which is recognizable by an increase in body temperature (Fig. 6.2). Abehsira et al. [97] showed recently that the QTc-interval is probably influenced by a complex interaction between sex hormones and gonadotropins. In both males and females, FSH was positively correlated to QTc-interval ( $r = 0.39$  and  $r = 0.38$ , respectively, in males and females), while free testosterone in males ( $r = -0.34$ ) and progesterone/oestrogen ratio in women ( $r = -0.38$ ) were negatively correlated. LH was only correlated in females to the QTc-interval ( $r = 0.30$ ).

## Pregnancy

During gestation, there is a complex and varying combination of sex hormones. Oestrogen and progesterone levels gradually increase during pregnancy until labour when the levels drop sharply reaching pregravid levels by the fifth postpartum day (Fig. 6.3) [98–102].

Studies regarding changes in QT/QTc-interval during pregnancy used either longitudinal data during pregnancy [103], the postpartum state as a control [104] or a control group of non-pregnant women [105, 106]. There is a longer QTc-interval during the first trimester and late pregnancy, with a shortening of the QTc-interval after delivery [104–106]. The QTc-interval just after delivery is however longer compared to the postpartum period [104]. The heart rate is, on the contrary, higher during pregnancy and delivery compared to the control group [104–106] and seems to increase with gestation [103]. Due to these changes in heart rate, it is not unlikely that the

**Fig. 6.3** Oestrogen and progesterone levels during pregnancy and after parturition together with a schematic representation of the QTc-intervals during the trimesters, parturition and the postpartum period. See text for explanation. (Derived from data in: Bett [115])



observed differences in the QTc-interval are influenced by the correction method used to correct the QT-interval for the heart rate [107]. Anneken et al. [108] recently studied five healthy women who became pregnant after stimulation by clomiphene citrate therapy for infertility, observing shorter QTc-intervals during higher oestrogen levels. Progesterone did not affect the QTc-interval significantly in that study.

The gestational prolongation of the QTc-interval does not precipitate widespread fatal cardiac arrhythmias in pregnant women. However, there is a slight increase in arrhythmias during pregnancy regarding SVTs and even ventricular tachycardia (VT) compared to the postpartum period or non-pregnant controls [109, 110]. Although the occurrence of VT is usually uncommon, their presence should raise a suspicion of underlying cardiovascular disease.

## Menopausal Period

The menopause is defined as the permanent cessation of menstrual periods, determined retrospectively after a woman has experienced 12 months of

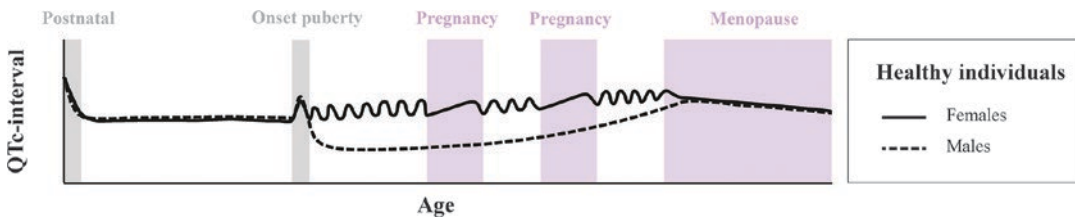
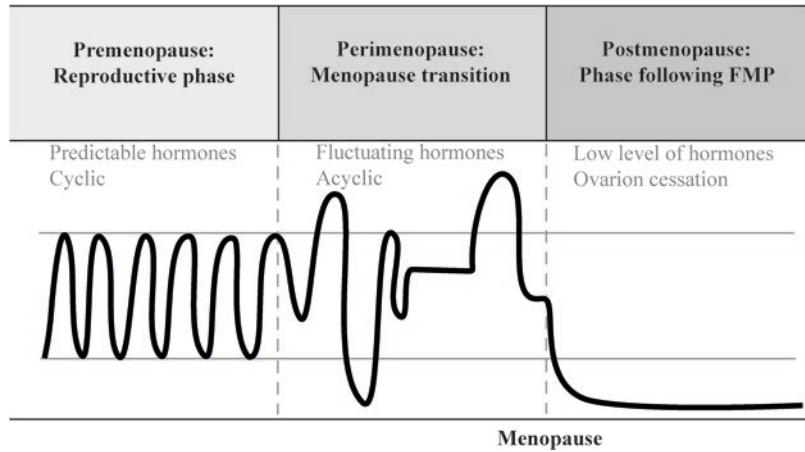
amenorrhoea without any other obvious pathological or physiological cause. The menopause is a reflection of complete, or near-complete, ovarian follicular depletion resulting in very low levels of oestrogen and progesterone and high levels of FSH concentrations (Fig. 6.4). The menopausal transition, or perimenopause, occurs after the reproductive years and before the menopause. This period is characterized by irregular menstrual cycles and significant hormonal variability [111, 112].

Two studies comparing premenopausal females to postmenopausal females showed no differences in QTc-interval and heart rate [113, 114]. One of the studies also measured the hormone levels and found no difference in oestrogen levels but a lower level of progesterone in the postmenopausal phase compared to the premenopausal phase [114].

The data regarding postmenopausal arrhythmogenesis is lacking; however the autonomic tone may also play a role in the peri-/postmenopausal period. Hence, the QTc-interval is shorter in women with hot flushes compared to those without, and the absence of menopausal hot flushes is associated with an elevated activity of the sympathetic nervous system.



**Fig. 6.4** Menstrual cycle patterns during menopause. FMP = Final menstrual period. See text for explanation. (Reprinted by permission from Springer Nature: Deecher and Dorries [116])



**Fig. 6.5** Schematic representation of hypothetical changes in QTc-interval in healthy individuals

## Conclusion

Age, sex and race have an influence on the QTc-interval. Although data regarding racial differences is lacking, the small differences seen are probably the result of the presence of considerable heterogeneity among race/ethnicity for multiple genetic loci that influence the QT-interval. Age- and sex-related differences in QTc-interval (Fig. 6.5) are most likely the result of changes in sex-specific hormones. Although the exact mechanisms and pathophysiology of the effect of sex hormones on the QTc-interval and the arrhythmogenesis are not known, testosterone appears to shorten the QTc-interval in males. In females, however, there is a more complex interaction between progesterone and oestrogen. In addition, the autonomic nervous system and gonadotropins may play an important part in this complex interaction.

**Acknowledgements** The authors are indebted to Larissa Groot and Jacqueline Limpens for their support.

## References

1. James AF, Choisy SC, Hancox JC. Recent advances in understanding sex differences in cardiac repolarization. *Prog Biophys Mol Biol.* 2007;94(3): 265–319.
2. Funck-Brentano C, Jaillon P. Rate-corrected QT interval: techniques and limitations. *Am J Cardiol.* 1993;72(6):17b–22b.
3. Bazett HC. An analysis of the time-relations of electrocardiograms. *Heart.* 1920;7:353–70.
4. van Noord C, Eijgelsheim M, Stricker BH. Drug- and non-drug-associated QT interval prolongation. *Br J Clin Pharmacol.* 2010;70(1):16–23.
5. Nachimuthu S, Assar MD, Schussler JM. Drug-induced QT interval prolongation: mechanisms and clinical management. *Ther Adv Drug Saf.* 2012;3(5):241–53.
6. Semizel E, Ozturk B, Bostan OM, Cil E, Ediz B. The effect of age and gender on the electrocardiogram in children. *Cardiol Young.* 2008;18(1):26–40.
7. Walsh SZ. Electrocardiographic intervals during the first week of life. *Am Heart J.* 1963;66:36–41.
8. Schaffer MS, Trippel DL, Buckles DS, Young RH, Dolan PL, Gillette PC. The longitudinal time course of QTc in early infancy. Preliminary results of a prospective sudden infant death syndrome surveillance program. *J Perinatol.* 1991;11(1):57–62.

9. Benatar A, Feenstra A. QT correction methods in infants and children: effects of age and gender. *Ann Noninvasive Electrocardiol.* 2015;20(2):119–25.
10. Dilaveris P, Roussos D, Giannopoulos G, Katinakis S, Maragiannis D, Raftopoulos L, et al. Clinical determinants of electrocardiographic and spatial vectorcardiographic descriptors of ventricular repolarization in healthy children. *Ann Noninvasive Electrocardiol.* 2011;16(1):49–55.
11. Braschi A, Abrignani MG, Francavilla VC, Abrignani V, Francavilla G. Age- and sex-based reference ranges for non-invasive ventricular repolarisation parameters. *Int J Clin Pract.* 2017;71(5).
12. Molinari G, Brunetti ND. Electrocardiograms of children and adolescents practicing non-competitive sports: normal limits and abnormal findings in a large European cohort evaluated by telecardiology. *Sports Med.* 2017;47(3):555–63.
13. Rijnbeek PR, Witsenburg M, Schrama E, Hess J, Kors JA. New normal limits for the paediatric electrocardiogram. *Eur Heart J.* 2001;22(8):702–11.
14. Yoshinaga M, Iwamoto M, Horigome H, Sumitomo N, Ushinohama H, Izumida N, et al. Standard values and characteristics of electrocardiographic findings in children and adolescents. *Circ J.* 2018;82(3):831–9.
15. Fukushige T, Yoshinaga M, Shimago A, Nishi J, Kono Y, Nomura Y, et al. Effect of age and overweight on the QT interval and the prevalence of long QT syndrome in children. *Am J Cardiol.* 2002;89(4):395–8.
16. Hazeki D, Yoshinaga M, Takahashi H, Tanaka Y, Haraguchi Y, Abe M, et al. Cut-offs for screening prolonged QT intervals from Fridericia's formula in children and adolescents. *Circ J.* 2010;74(8):1663–9.
17. Mason JW, Ramseth DJ, Chanter DO, Moon TE, Goodman DB, Mendzelevski B. Electrocardiographic reference ranges derived from 79,743 ambulatory subjects. *J Electrocardiol.* 2007;40(3):228–34.
18. Rautaharju PM, Mason JW, Akiyama T. New age- and sex-specific criteria for QT prolongation based on rate correction formulas that minimize bias at the upper normal limits. [Erratum appears in *Int J Cardiol.* 2015;178:299; PMID: 25639760]. *Int J Cardiol.* 2014;174(3):535–40.
19. Rijnbeek PR, van Herpen G, Bots ML, Man S, Verweij N, Hofman A, Hillege H, Numans ME, Swenne CA, Witteman JC, Kors JA. Normal values of the electrocardiogram for ages 16-90 years. *J Electrocardiol.* 2014;47(6):914–21.
20. Surawicz B, Parikh SR. Prevalence of male and female patterns of early ventricular repolarization in the normal ECG of males and females from childhood to old age. *J Am Coll Cardiol.* 2002;40(10):1870–6.
21. Santini M, Di Fusco SA, Colivicchi F, Gargaro A. Electrocardiographic characteristics, anthropometric features, and cardiovascular risk factors in a large cohort of adolescents. *Europace.* 2018;20(11):1833–40. <https://doi.org/10.1093/europace/euy073>.
22. Macfarlane PW, Lloyd SM, Singh D, Hamde S, Clark E, Devine B, et al. Normal limits of the electrocardiogram in Indians. *J Electrocardiol.* 2015;48(4):652–68.
23. de Bruyne MC, Hoes AW, Kors JA, Hofman A, van Bommel JH, Grobbee DE. Prolonged QT interval predicts cardiac and all-cause mortality in the elderly. The Rotterdam Study. *Eur Heart J.* 1999;20(4):278–84.
24. Dewhurst MJ, Di Marco LY, Dewhurst F, Adams PC, Murray A, Orega GP, et al. Electrocardiographic reference values for a population of older adults in sub-Saharan Africa. *Ann Noninvasive Electrocardiol.* 2014;19(1):34–42.
25. Rabkin SW, Cheng XJ, Thompson DJ. Detailed analysis of the impact of age on the QT interval. *J Geriatr Cardiol.* 2016;13(9):740–8.
26. Chen CY, Chiang BN, Macfarlane PW. Normal limits of the electrocardiogram in a Chinese population. *J Electrocardiol.* 1989;22(1):1–15.
27. Mangoni AA, Kinirons MT, Swift CG, Jackson SH. Impact of age on QT interval and QT dispersion in healthy subjects: a regression analysis. *Age Ageing.* 2003;32(3):326–31.
28. Baumert M, Czipelova B, Porta A, Javorka M. Decoupling of QT interval variability from heart rate variability with ageing. *Physiol Meas.* 2013;34(11):1435–48.
29. Benoit SR, Mendelsohn AB, Nourjah P, Staffa JA, Graham DJ. Risk factors for prolonged QTc among US adults: Third National Health and Nutrition Examination Survey. *Eur J Cardiovasc Prev Rehabil.* 2005;12(4):363–8.
30. Macfarlane PW, McLaughlin SC, Devine B, Yang TF. Effects of age, sex, and race on ECG interval measurements. *J Electrocardiol.* 1994;27 Suppl:14–9.
31. Macfarlane PW, Katibi IA, Hamde ST, Singh D, Clark E, Devine B, Francq BG, Lloyd S, Kumar V. Racial differences in the ECG-selected aspects. *J Electrocardiol.* 2014;47(6):809–14.
32. Ramirez AH, Schildcrout JS, Blakemore DL, Masys DR, Pulley JM, Basford MA, Roden DM, Denny JC. Modulators of normal electrocardiographic intervals identified in a large electronic medical record. *Heart Rhythm.* 2011;8(2):271–7.
33. Zerkiebel N, Perret F, Bovet P, Abel M, Jaggy C, Paccaud F, Kappenberger L. Electrocardiographic findings in a middle-aged African population in the Seychelles islands. *J Electrocardiol.* 2000;33(1):1–15.
34. Reardon M, Malik M. QT interval change with age in an overtly healthy older population. *Clin Cardiol.* 1996;19(12):949–52.
35. Katibi I, Clark EN, Devine B, Lloyd SM, Macfarlane PW. Normal limits of the electrocardiogram in Nigerians. *J Electrocardiol.* 2013;46(4):289–95.



36. Tran H, White CM, Chow MS, Kluger J. An evaluation of the impact of gender and age on QT dispersion in healthy subjects. *Ann Noninvasive Electrocardiol.* 2001;6(2):129–33.
37. Vicente J, Johannesen L, Galeotti L, Strauss DG. Mechanisms of sex and age differences in ventricular repolarization in humans. *Am Heart J.* 2014;168(5):749–56.
38. Wu J, Kors JA, Rijnbeek PR, van Herpen G, Lu Z, Xu C. Normal limits of the electrocardiogram in Chinese subjects. *Int J Cardiol.* 2003;87(1):37–51.
39. Tan ES, Yap J, Xu CF, Feng L, Nyunt SZ, Santhanakrishnan R, et al. Association of age, sex, body size and ethnicity with electrocardiographic values in community-based older Asian adults. *Heart Lung Circ.* 2016;25(7):705–11.
40. Sugao M, Fujiki A, Sakabe M, Nishida K, Tsuneda T, Iwamoto J, et al. New quantitative methods for evaluation of dynamic changes in QT interval on 24 hour Holter ECG recordings: QT interval in idiopathic ventricular fibrillation and long QT syndrome. *Heart.* 2006;92(2):201–7.
41. van der Ende MY, Siland JE, Snieder H, van der Harst P, Rienstra M. Population-based values and abnormalities of the electrocardiogram in the general Dutch population: The LifeLines Cohort Study. *Clin Cardiol.* 2017;40(10):865–72.
42. Heemskerk CPM, Pereboom M, van Stralen K, Berger FA, van den Bemt P, Kuijper AFM, et al. Risk factors for QTc interval prolongation. *Eur J Clin Pharmacol.* 2018;74(2):183–91.
43. Khumrin P, Srisuwan P, Lertprayoonmit W, Leelarphat L, Phumphuang C. Effective ECG reference ranges for Northern Thai people. *Heart Asia.* 2015;7(1):32–40.
44. Rautaharju PM, Zhou SH, Wong S, Calhoun HP, Berenson GS, Prineas R, Davignon A. Sex differences in the evolution of the electrocardiographic QT interval with age. *Can J Cardiol.* 1992;8(7):690–5.
45. Mizuno Y. Normal limits and variability of electrocardiographic items of the Japanese. *Jpn Circ J.* 1966;30(4):357–78.
46. Shinmura K, Ebihara Y, Kawamura M, Tani M, Nakamura Y. [Changes in electrocardiographic findings with aging in a longitudinal study of 500 apparently healthy persons aged 60 years and older]. *Nihon Ronen Igakkai Zasshi Jpn J Geriatr.* 1994;31(5):366–73.
47. Stramba-Badiale M, Spagnolo D, Bosi G, Schwartz PJ. Are gender differences in QTc present at birth? MISNES Investigators. Multicenter Italian Study on Neonatal Electrocardiography and Sudden Infant Death Syndrome. *Am J Cardiol.* 1995;75(17):1277–8.
48. Marti-Almor J, Berruero R, Garcia-Algar O, Mur A, Bazan V, Recasens L, Pérez-Rodón J, Bruguera J. QT interval in newborns of different ethnic origin: usefulness of neonatal ECG screening. *Rev Esp Cardiol.* 2008;61(9):980–2.
49. Eberle T, Hessling G, Ulmer HE, Brockmeier K. Prediction of normal QT intervals in children. *J Electrocardiol.* 1998;31(Suppl):121–5.
50. Pearl W. Effects of gender, age, and heart rate on QT intervals in children. *Pediatr Cardiol.* 1996;17(3):135–6.
51. Griffet V, Finet G, Di Filippo S, Lantelme P, Caignault JR, Guerard S. [Athlete's heart in the young: electrocardiographic and echocardiographic patterns in 107 French athletes]. *Ann Cardiol Angeiol (Paris).* 2013;62(2):116–21.
52. Kumar N, Saini D, Froelicher V. A gender-based analysis of high school athletes using computerized electrocardiogram measurements. *PLoS One.* 2013;8(1):e53365.
53. Arai K, Nakagawa Y, Iwata T, Horiguchi H, Murata K. Relationships between QT interval and heart rate variability at rest and the covariates in healthy young adults. *Auton Neurosci.* 2013;173(1–2):53–7.
54. Bessem BB, de Bruijn MM, Nieuwland WW. Gender differences in the electrocardiogram screening of athletes. *J Sci Med Sport.* 2017;20(2):213–7.
55. Mandic S, Fonda H, Dewey F, Le VV, Stein R, Wheeler M, et al. Effect of gender on computerized electrocardiogram measurements in college athletes. *Phys Sportsmed.* 2010;38(2):156–64.
56. Omiya K, Sekizuka H, Kida K, Suzuki K, Akashi YJ, Ohba H, Musha H. Influence of gender and types of sports training on QT variables in young elite athletes. *Eur J Sport Sci.* 2014;14 Suppl 1:S32–8.
57. Merri M, Benhorin J, Alberti M, Locati E, Moss AJ. Electrocardiographic quantitation of ventricular repolarization. *Circulation.* 1989;80(5):1301–8.
58. Tutar HE, Ocal B, Imamoglu A, Atalay S. Dispersion of QT and QTc interval in healthy children, and effects of sinus arrhythmia on QT dispersion. *Heart.* 1998;80(1):77–9.
59. Misigoj-Durakovic M, Durakovic Z, Prskalo I. Heart rate-corrected QT and JT intervals in electrocardiograms in physically fit students and student athletes. *Ann Noninvasive Electrocardiol.* 2016;21(6):595–603.
60. Haarmark C, Graff C, Andersen MP, Hardahl T, Struijk JJ, Toft E, et al. Reference values of electrocardiogram repolarization variables in a healthy population. *J Electrocardiol.* 2010;43(1):31–9.
61. Rautaharju PM, Zhang ZM, Haisty WK Jr, Gregg RE, Warren J, Horacek MB, et al. Race- and sex-associated differences in rate-adjusted QT, QTpeak, ST elevation and other regional measures of repolarization: the Atherosclerosis Risk in Communities (ARIC) Study. *J Electrocardiol.* 2014;47(3):342–50.
62. Santhanakrishnan R, Wang N, Larson MG, Magnani JM, Vasan RS, Wang TJ, et al. Racial differences in electrocardiographic characteristics and prognostic significance in whites versus Asians. *J Am Heart Assoc.* 2016;5(3):e002956.
63. Goldberg RJ, Bengtson J, Chen ZY, Anderson KM, Locati E, Levy D. Duration of the QT interval and

- total and cardiovascular mortality in healthy persons (The Framingham Heart Study experience). *Am J Cardiol.* 1991;67(1):55–8.
64. Extramiana F, Maisson-Blanche P, Badilini F, Pinoteau J, Deseo T, Coumel P. Circadian modulation of QT rate dependence in healthy volunteers: gender and age differences. *J Electrocardiol.* 1999;32(1):33–43.
  65. Zhang Y, Ouyang P, Post WS, Dalal D, Vaidya D, Blasco-Colmenares E, et al. Sex-steroid hormones and electrocardiographic QT-interval duration: findings from the third National Health and Nutrition Examination Survey and the Multi-Ethnic Study of Atherosclerosis. *Am J Epidemiol.* 2011;174(4):403–11.
  66. Sliwa K, Lee GA, Carrington MJ, Obel P, Okreglicki A, Stewart S. Redefining the ECG in urban South Africans: electrocardiographic findings in heart disease-free Africans. *Int J Cardiol.* 2013;167(5):2204–9.
  67. Akylbekova EL, Crow RS, Johnson WD, Buxbaum SG, Njemanze S, Fox E, et al. Clinical correlates and heritability of QT interval duration in blacks: the Jackson Heart Study. *Circ Arrhythm Electrophysiol.* 2009;2(4):427–32.
  68. Kassotis J, Costeas C, Bedi AK, Tolat A, Reiffel J. Effects of aging and gender on QT dispersion in an overtly healthy population. *Pacing Clin Electrophysiol.* 2000;23(7):1121–6.
  69. Marjamaa A, Salomaa V, Newton-Cheh C, Porthan K, Reunanen A, Karanko H, et al. High prevalence of four long QT syndrome founder mutations in the Finnish population. *Ann Med.* 2009;41(3):234–40.
  70. Bonny A, Bika Lele EC, Mandengue S, Larrazet F, Amara W. [Ethnic differences in electrocardiogram between a black African and a white European young adult population under 35 years]. *Presse Med.* 2013;42(4 Pt 1):e96–105.
  71. Brink AJ. The normal electrocardiogram in the adult South African Bantu. *S Afr J Lab Clin Med.* 1956;2(2):97–123.
  72. Grandinetti A, Seifried S, Mor J, Chang HK, Theriault AG. Prevalence and risk factors for prolonged QTc in a multiethnic cohort in rural Hawaii. *Clin Biochem.* 2005;38(2):116–22.
  73. Mansi IA, Nash IS. Ethnic differences in electrocardiographic intervals and axes. *J Electrocardiol.* 2001;34(4):303–7.
  74. Vitelli LL, Crow RS, Shahar E, Hutchinson RG, Rautaharju PM, Folsom AR. Electrocardiographic findings in a healthy biracial population. Atherosclerosis Risk in Communities (ARIC) Study Investigators. *Am J Cardiol.* 1998;81(4):453–9.
  75. Woods JB, Laurie W. The electrocardiogram of the South African Bantu. *Circulation.* 1959;19(2):251–6.
  76. Sgarbossa EB, Pinski SL, Williams D, Pavlovic-Surjancev B, Tang J, Trohman RG. Comparison of QT intervals in African-Americans versus Caucasians. *Am J Cardiol.* 2000;86(8):880–2.
  77. Rautaharju PM, Prineas RJ, Kadish A, Larson JC, Hsia J, Lund B. Normal standards for QT and QT subintervals derived from a large ethnically diverse population of women aged 50 to 79 years (the Women's Health Initiative [WHI]). *Am J Cardiol.* 2006;97(5):730–7.
  78. Munger RG, Prineas RJ, Crow RS, Changbumrung S, Keane V, Wangsuphachart V, et al. Prolonged QT interval and risk of sudden death in South-East Asian men. *Lancet.* 1991;338(8762):280–1.
  79. Shah RR. Drug-induced QT interval prolongation: does ethnicity of the thorough QT study population matter? *Br J Clin Pharmacol.* 2013;75(2):347–58.
  80. Seyerle AA, Young AM, Jeff JM, Melton PE, Jorgensen NW, Lin Y, Carty CL, et al. Evidence of heterogeneity by race/ethnicity in genetic determinants of QT interval. *Epidemiology.* 2014;25(6):790–8.
  81. Sedlak T, Shufelt C, Iribarren C, Merz CN. Sex hormones and the QT interval: a review. *J Womens Health (Larchmt).* 2012;21(9):933–41.
  82. Kuiri-Hanninen T, Sankilampi U, Dunkel L. Activation of the hypothalamic-pituitary-gonadal axis in infancy: minipuberty. *Horm Res Paediatr.* 2014;82(2):73–80.
  83. Harman SM, Metter EJ, Tobin JD, Pearson J, Blackman MR. Longitudinal effects of aging on serum total and free testosterone levels in healthy men. Baltimore Longitudinal Study of Aging. *J Clin Endocrinol Metab.* 2001;86(2):724–31.
  84. Hulot JS, Demolis JL, Riviere R, Strabach S, Christin-Maitre S, Funck-Brentano C. Influence of endogenous oestrogens on QT interval duration. *Eur Heart J.* 2003;24(18):1663–7.
  85. Nakagawa M, Ooie T, Takahashi N, Taniguchi Y, Anan F, Yonemochi H, Saikawa T. Influence of menstrual cycle on QT interval dynamics. *Pacing Clin Electrophysiol.* 2006;29(6):607–13.
  86. Burke JH, Ehlert FA, Kruse JT, Parker MA, Goldberger JJ, Kadish AH. Gender-specific differences in the QT interval and the effect of autonomic tone and menstrual cycle in healthy adults. *Am J Cardiol.* 1997;79(2):178–81.
  87. Endres S, Mayuga KA, Cristofaro A, Taneja T, Goldberger JJ, Kadish AH. Menstrual cycle and ST height. *Ann Noninvasive Electrocardiol.* 2004;9(2):121–6.
  88. Rodriguez I, Kilborn MJ, Liu XK, Pezzullo JC, Woosley RL. Drug-induced QT prolongation in women during the menstrual cycle. *JAMA.* 2001;285(10):1322–6.
  89. Dogan M, Yiginer O, Uz O, Kucuk U, Degirmencioglu G, Isilak Z, et al. The effects of female sex hormones on ventricular premature beats and repolarization parameters in physiological menstrual cycle. *Pacing Clin Electrophysiol.* 2016;39(5):418–26.
  90. Yildirim A, Kabakci G, Akgul E, Tokgozoglu L, Oto A. Effects of menstrual cycle on cardiac autonomic innervation as assessed by heart rate variability. *Ann Noninvasive Electrocardiol.* 2002;7(1):60–3.

91. Sato N, Miyake S, Akatsu J, Kumashiro M. Power spectral analysis of heart rate variability in healthy young women during the normal menstrual cycle. *Psychosom Med.* 1995;57(4):331–5.
92. Claydon VE, Younis NR, Hainsworth R. Phase of the menstrual cycle does not affect orthostatic tolerance in healthy women. *Clin Auton Res.* 2006;16(2):98–104.
93. Rosano GM, Leonardo F, Sarrel PM, Beale CM, De Luca F, Collins P. Cyclical variation in paroxysmal supraventricular tachycardia in women. *Lancet.* 1996;347(9004):786–8.
94. Myerburg RJ, Cox MM, Interian A Jr, Mitrani R, Gargis I, Dylewski J, Castellanos A. Cycling of inducibility of paroxysmal supraventricular tachycardia in women and its implications for timing of electrophysiologic procedures. *Am J Cardiol.* 1999;83(7):1049–54.
95. Roden DM. Drug-induced prolongation of the QT interval. *N Engl J Med.* 2004;350(10):1013–22.
96. Du XJ, Riemersma RA, Dart AM. Cardiovascular protection by oestrogen is partly mediated through modulation of autonomic nervous function. *Cardiovasc Res.* 1995;30(2):161–5.
97. Abehsira G, Bachelot A, Badilini F, Koehl L, Lebot M, Favet C, et al. Complex influence of gonadotropins and sex steroid hormones on QT interval duration. *J Clin Endocrinol Metab.* 2016;101(7):2776–84.
98. Boroditsky RS, Reyes FI, Winter JS, Faiman C. Maternal serum estrogen and progesterone concentrations preceding normal labor. *Obstet Gynecol.* 1978;51(6):686–91.
99. Tulchinsky D, Hobel CJ, Yeager E, Marshall JR. Plasma estrone, estradiol, estriol, progesterone, and 17-hydroxyprogesterone in human pregnancy. I. Normal pregnancy. *Am J Obstet Gynecol.* 1972;112(8):1095–100.
100. Dorr HG, Heller A, Versmold HT, Sippell WG, Herrmann M, Bidlingmaier F, Knorr D. Longitudinal study of progestins, mineralocorticoids, and glucocorticoids throughout human pregnancy. *J Clin Endocrinol Metab.* 1989;68(5):863–8.
101. Johansson ED. Plasma levels of progesterone in pregnancy measured by a rapid competitive protein binding technique. *Acta Endocrinol.* 1969;61(4):607–17.
102. Yannone ME, McCurdy JR, Goldfien A. Plasma progesterone levels in normal pregnancy, labor, and the puerperium. II. Clinical data. *Am J Obstet Gynecol.* 1968;101(8):1058–61.
103. Baumert M, Javorka M, Seeck A, Faber R, Sanders P, Voss A. Multiscale entropy and detrended fluctuation analysis of QT interval and heart rate variability during normal pregnancy. *Comput Biol Med.* 2012;42(3):347–52.
104. Lechmanova M, Kittnar O, Mlcek M, Slavicek J, Dohnalova A, Havranek S, et al. QT dispersion and T-loop morphology in late pregnancy and after delivery. *Physiol Res.* 2002;51(2):121–9.
105. Tanindi A, Akgun N, Pabuccu EG, Gursoy AY, Yuce E, Tore HF, Duvan CI. Electrocardiographic P-wave duration, QT interval, T peak to end interval and Tp-e/QT ratio in pregnancy with respect to trimesters. *Ann Noninvasive Electrocardiol.* 2016;21(2):169–74.
106. Carpenter RE, D’Silva LA, Emery SJ, Uzun O, Rassi D, Lewis MJ. Changes in heart rate variability and QT variability during the first trimester of pregnancy. *Physiol Meas.* 2015;36(3):531–45.
107. Luo S, Michler K, Johnston P, Macfarlane PW. A comparison of commonly used QT correction formulae: the effect of heart rate on the QTc of normal ECGs. *J Electrocardiol.* 2004;37 Suppl:81–90.
108. Anneken L, Baumann S, Vigneault P, Biliczki P, Friedrich C, Xiao L, et al. Estradiol regulates human QT-interval: acceleration of cardiac repolarization by enhanced KCNH2 membrane trafficking. *Eur Heart J.* 2016;37(7):640–50.
109. Gowda RM, Khan IA, Mehta NJ, Vasavada BC, Sacchi TJ. Cardiac arrhythmias in pregnancy: clinical and therapeutic considerations. *Int J Cardiol.* 2003;88(2–3):129–33.
110. Shotan A, Ostrzega E, Mehra A, Johnson JV, Elkayam U. Incidence of arrhythmias in normal pregnancy and relation to palpitations, dizziness, and syncope. *Am J Cardiol.* 1997;79(8):1061–4.
111. Buckler H. The menopause transition: endocrine changes and clinical symptoms. *J Br Menopause Soc.* 2005;11(2):61–5.
112. Hee J, MacNaughton J, Bangah M, Burger HG. Perimenopausal patterns of gonadotrophins, immunoreactive inhibin, oestradiol and progesterone. *Maturitas.* 1993;18(1):9–20.
113. Dogan U, Dogan NU, Basarir AO, Yildirim S, Celik C, Incesu F, Ozdemir K. P-wave parameters and cardiac repolarization indices: does menopausal status matter? *J Cardiol.* 2012;60(4):333–7.
114. Lavi S, Nevo O, Thaler I, Rosenfeld R, Dayan L, Hirshoren N, et al. Effect of aging on the cardiovascular regulatory systems in healthy women. *Am J Physiol Regul Integr Comp Physiol.* 2007;292(2):R788–93.
115. Bett GC. Hormones and sex differences: changes in cardiac electrophysiology with pregnancy. *Clin Sci (Lond).* 2016;130(10):747–59.
116. Deecher DC, Dorries K. Understanding the pathophysiology of vasomotor symptoms (hot flushes and night sweats) that occur in perimenopause, menopause, and postmenopause life stages. *Arch Womens Ment Health.* 2007;10(6):247–57.