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3.1 Introduction

Due to the ongoing shift in the distribution of the world's population towards old age, we recently experience a dramatic increase in comorbidities like diabetes or venous and arterial insufficiency. This results in a raising number of chronic wounds which have become not only an individual medical but also a significant economic burden, consuming 2-4% of health care budgets worldwide [1].

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Wound healing is a complex system depending on the timed coordination of several cell types, intra- and extracellular mechanisms, proteins, and pathways, but also on several external factors like infections or mechanical irritation (Fig. 3.1). Defect or dominance of one factor can cause to a sudden breakdown of localized healing capacity, leading to formation of chronic wounds. A famous example for the fragility of the cellular mechanism for tissue homeostasis and repair is the connection between vitamin C deficiency and scurvy resulting in nonhealing wounds and spontaneous bleeding known since the sixteenth century [2, 3]. Mentioned first in journey books of Christopher Columbus as a result of monotone diet, the pathomechanism remained unclear until the twentieth century. Then it could be demonstrated that vitamin C represents a main cofactor for collagen crosslinking and an important factor to reduce oxidative stress [4]. This example shows how impactful minimal alterations in our metabolism can be for tissue regeneration. Therefore, a complete understanding of all molecular and cellular players involved in wound healing is pivotal for developing treatment strategies and effective drugs.

In this chapter, we summarize the most promising recent advances in wound healing therapeutics with the corresponding challenges and shed light on possible solutions for effective application.

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Basic Principles and Current Approach for Soft Tissue Regeneration

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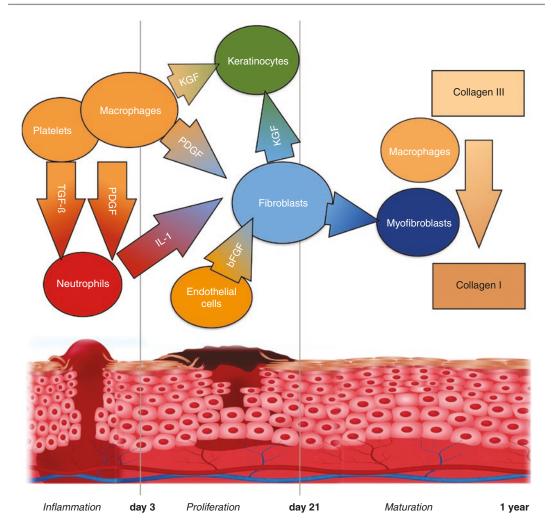


Fig. 3.1 Phases of adult wound healing with main affecting cells and signals. (Left) Coagulation and inflammation—day 0–3. Platelets: formation of platelet plug and secretion of platelet-derived growth factor and transforming growth factor for chemotaxis of neutrophils. Neutrophils: secrete interleukin 1 for the beginning of chemokine-cascade for chemotaxis of inflammatory cells; macrophages: phagocyte bacteria and secrete paracrine factors for keratinocyte-based epithelization and fibroblast activation. (Middle) Proliferation—day 4–21.

Beginning of angiogenesis, dependent on endothelial cells: activated by vascular endothelial growth factor. Formation of the extracellular matrix based on fibroblast activation: activated by basic fibroblast growth factor, interleukin 1, and platelet-derived growth factor. Epithelialization: keratinocyte based as a response to keratinocyte growth factor. (Right) Maturation phase—day 21–1 year. Wound contraction: transformation of fibroblasts to myofibroblast. Collagen remodeling: effected by myofibroblasts and macrophages

3.2 Recent Advances in Wound Therapeutics

3.2.1 Growth Factor Therapy

The wound healing promoting effect of growth factors is broadly known. Although they have shown to act beneficial in preclinical studies, large clinical studies supporting this are still missing. A meta-analysis based on the Cochrane database showed a general benefit of growth factor-induced wound healing without any significant general adverse effects [5]. The plateletderived growth factors (PDGF), vascular endothelial growth factor (VEGF), epidermal growth factor (EGF), fibroblast growth factor (FGF), and the transforming growth factor-beta (TGF- β) are in current focus of research.

PDGF-BB, administered in hydrogels and available as "Regranex" (Ortho-McNeil, Raritan, NJ), is the only growth factor therapy that is currently approved for treatment of nonhealing wounds [6]. Although there has been shown advantage in hypertensive leg ulcers, the application of this gel should be considered only as ultima-ratio treatment due to a higher rate of malignancies in patients treated with PDGF-BB [7].

As another growth factor VEGF was successfully used for accelerated wound closure in diabetic mice. Preclinical study showed less than half of resurfacing time in VEGF-treated group than in a non-treated group [8]. Also non-treated wounds on the contralateral site of animals with VEGF therapy showed an accelerated wound closure time. This leads to the suggestion of an additional systemic effect of local VEGF treatment with the possibility of interacting with tumor growth and of promoting malignant tendencies. There has been only one clinical trial comparing VEGF treatment and placebo showing no significant benefit for VEGF [9].

Several study groups investigated the effect of intralesional EGF injections on wound closure [10–12]. Although first clinical trials have been performed in Cuba in 2006 and have shown accelerated wound healing of high-grade diabetic foot healing and complete wound closure in up to 85% of cases, it is still no treatment option in western countries. A meta-analysis performed by Yang et al. [13] confirmed these first results strengthening the hopes for a new clinical treatment option for diabetic and avascular wounds.

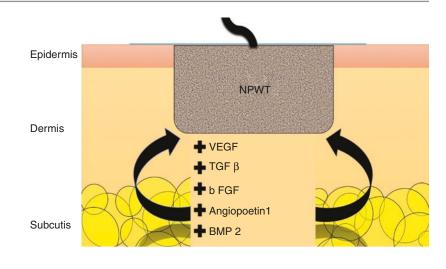
B-FGF is one of the first growth factors that has been investigated. A clinical study by Richard et al. in 1995 involved 17 patients and could find no promoting effect of b-FGF [14]. Up to now the efficacy of b-FGF in wound healing remains unclear. While there exist clinical studies showing a promotion of diabetic wound healing effected by injectable b-FGF, others deny a significant effect of b-FGF releasing sponges while preclinical trials have shown a promising effect [15–17]. Furthermore, not only b-FGF but also acid-FGF has been tested for wound healing and similarly showed inconclusive results [18].

Although the TGF- β family, including TGF- β -1, 2, and 3, has shown to be involved in both promotion of wound healing and scarring, it has not become a possible treatment option in clinical routine yet [19]. The wound healing ability of TGF- β -1 and TGF- β -2 in murine models are well described. Although first clinical phase I and II trials have shown efficacy and safety of a TGF- β -releasing scaffold for treatment of venous ulcers, there exist no supporting phase III study [20].

Despite the fact that growth factor therapy has shown to be effective in preclinical and some clinical trials, only few of these substances hold promise to enter the clinical routine. A therapy containing only one growth factor is most likely not sufficient to efficiently promote wound healing, especially compared to NPWT, which has shown to significantly enhance a plethora of autologous growth factor levels.

3.2.2 Negative-Pressure Wound Therapy

The use of negative-pressure wound therapies (NPWT = vacuum-assisted closure = VAC) provides an effective and elegant way to close wounds, prevent infections, and simultaneously increase local growth factor levels. NPWT has shown benefit on bacterial contamination rate. It also temporarily creates relative hypoxia in the wound region, resulting in significant higher levels of the main growth factors (VEGF, TGF β , and basic FGF), angiopoietin 1 (essential for neoangiogenesis), and bone morphogenetic protein 2 (BMP 2-involved in cartilage and bone metabolism) [21–25]. Several studies suggest that microdeformation of the wound surface leads to accelerated cell migration and matrix production (Fig. 3.2) [23–25]. Interestingly, the temporary hypoxia induces the osteogenetic differentiation of MSCs. Therefore NPWT seems to be the perfect option for treatment of soft tissue defects involving bone defects and infected or potentially infected wounds. Further research has to be carried out to confirm these hypotheses in a clinical setting.



Technical development has led to change in handling of NPWT systems. Dressing changes only three times a week instead of twice daily as recommended in the first trails using NPWT has led to the possibility of a long-term use. By using silver-coated foams, that additionally hinder bacterial growth, NPWT became even more successful [26]. In aggregate, NPWT provides an effective and elegant way to treat difficult wounds by enhancing local growth factor levels and decreasing bacterial contamination of wounds.

3.2.3 Antioxidants and Wnt Modulation

The human skin is constantly exposed to environmental factors such as UV light, radiation, ozone (O₃), or air pollution inducing reactive oxygen species (= ROS). Additionally, cellular metabolism leads to ROS as side products. The causality between free radicals and aging has been described by different groups within the last century [27–31]. By causing accumulation of oxidative toxic products in long-living molecules such as collagen, it leads to peroxidation and dysfunction of these molecules. ROSinduced damaging of the DNA is directly followed by base loss/modification or breakage. ROS can further lead to glycation of proteins followed by degradation. According to this, ROS can significantly inhibit endogenous ability for

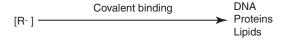


Fig. 3.3 Mechanism of reactive oxygen species (ROS) damage

wound healing by destruction of cells, key proteins, or parts of the ECM (Fig. 3.3).

Antioxidant treatment has been used since several years as a product improving the quality of skin. The most popular and most commonly used antioxidant is a polyphenol, also called aloe vera. As the main component of ointments or gels its therapeutic effect is related to the stimulation of collagen syntheses on the one hand and to antioxidative effects on the other hand [32]. But the use of anti-oxidants is not only limited on aesthetic treatment options. Some iron chelators like deferoxamine (= Desferal or more potent, desferriexochelin-772SM (D-Exo)) [33], deferiprone, and deferasirox are already in use in different medical fields [34]. Next to being well known as treatment option for beta-thalassemia [35, 36], anti-oxidant drugs have shown benefits mainly due to their anti-inflammatory potential to increase the retention rate of fat grafts, the survival rate of free flaps, and the healing process of diabetic wounds [37, 38]. This can be explained by the ability of free iron to induce the prolylhydroxylation of the hypoxia-inducible factor 1a (HIF-1 α), a process leading to inactivation and degradation of HIF-1 α [39]. Less free iron results in a higher expression of HIF-1 α and thereby

Fig. 3.2 Molecular mechanism of negative-pressure wound therapy

leads to benefits in local neovascularization and tissue regeneration. Harnessing these effects, a transdermal delivery system releasing DFO showed accelerated wound healing in diabetic ulcers. Prophylactic use of this system has a preventive effect on ulcer formation [40].

But not only antioxidative agents, also wnt pathway manipulation is a promising new alley of wound healing research. The wnt pathway represents a sequence of factors that can easily be targeted by nanoparticles. The wnt pathway was found to play an important role in embryonic vertebrae development, in the development of different malignancies, and additionally in tissue repair and scarring [41–44]. Pyrvinium, an antihelminthic drug, inhibits the wnt pathway by promoting the effectivity of the case in kinase 1α (CK1 α), leading to accelerated degradation of casein, a factor of wnt pathway [45, 46]. Studies using pyrvinium show a 1.4-fold increase of MSC proliferation by simultaneously inhibiting the osteogenic and chondrogenic differentiation. These changes were initiated by a pyrvinium-releasing sponge. Pyrvinium only affected the proliferation rate of MSCs. Other cell lines like HUVECs have shown no significant changes compared to nontreated cells [47].

Additionally, the wnt pathway has also shown to play a significant role in scar formation. By stimulation of the wnt pathway and the FGF pathway, the regeneration of hair follicles could be induced. Hair follicle secretes bone morphogenetic protein (BMP), which again stimulates myofibroblasts to differentiate into adipocytes. This pathway leads to inhibition of scar formation. Targeting and mimicking these three pathways to either prevent scarring or treat hypertrophic scars or keloids is a future goal of drug development [48–50].

3.2.4 RNA Interference-Based Therapy

Gene expression initially starts in the nucleus with transcription—the production of mRNA (messenger-RNA) followed by an export to the cytoplasm. Translation of mRNA leads to the production of proteins. After this process the mRNA is degraded.

The basic principle of RNA interference (RNAi-based therapy) is based on body's own mechanism for mRNA degradation: By binding to mRNAs, endogenous miRNAs (micro-RNAs) or synthetic siRNAs (small interfering RNAs) lead to mRNA degradation or to formation of double-stranded RNA and as a result to suppression of their translation [51, 52]. Synthetically produced siRNA can be used for knocking down factors which inhibit neo-angiogenesis and inhibit keratinocyte migration followed by re-epithelialization [53–55].

For example, endogenous miRNA-21 is one of the best studied miRNAs. It has been shown to regulate re-epithelialization, cell proliferation, wound contraction, and formation of granulation tissue [54, 56, 57]. RNAi is not limited to wound healing applications, but also offers new possibilities in cancer therapy or treatment of genetic diseases like amyotrophic lateral sclerosis (by targeting and destroying wild-type mRNAs) [58, 59].

Difficulties for the application of this novel therapy include delivery to specific cells and problems with internalization of i-RNA to certain cell types [60, 61]. Additionally, a high degradation rate through RNases leads to a short intracellular half-life. However, further development of this technique may lead to new ways to enhance tissue regeneration and to bring us closer to the holy grail of scarless wound healing [62, 63].

3.2.5 Stem Cell-Based Therapy

Mesenchymal stromal cells (MSCs) can be utilized to treat challenging wounds, such as wounds followed irradiation, ischemic or diabetic wounds. The basic principle is a potential differentiation of stromal cells and a higher local level of growth factors. Although preclinical and clinical studies showed promising results, there still remain several problems. Irregularities are caused by patient's individual factors, such as diabetes or age [64, 65]. These uncertainties still limit the clinical use. But stem cell-based therapy is not only limited to the regenerative potential of MSCs harvested from bone marrow or adipose tissue. Also peripheral blood cells (PBCs) have shown to secrete a mixture of the pro-angiogenetic factors VEGF and HIF-1, when being temporally conditioned under hypoxic stress [66]. These findings have been used for developing both an implantable and an injectable wound healing system and seem to be a promising approach to accelerate wound healing [67].

3.2.6 Scaffolds and Skin Equivalents

Bioactive dressings are engineered from components that are naturally present in the ECM or composed of polymers to mimic this unique matrix [68]. Biomimetic collagen hydrogels have been shown to accelerate early wound healing by modifying cell recruitment and augmenting granulation tissue formation [69]. Recently biologic matrices have evolved from being simple ECM replacements towards drug and cell delivery vehicles. Novel regenerative matrices are capable of both skin replacement and stimulation of endogenous cells [70]. For example, pullulan (a polysaccharide polymer)-collagen matrices was seeded with mesenchymal stem cells (MSCs) and showed to enhance cell survival [71]. MSCs delivered in such a structured matrix environment have demonstrated enhanced efficacy by increased angiogenic cytokine expression [72]. Therefore, scaffolds can represent an intelligent and efficient drug-delivery vehicle to overcome certain problems of cell-based therapies [73].

In addition to improving wound healing and skin regeneration by increased neovascularization, scaffold-seeded progenitor cells can also enhance tissue repair by inducing a specific immune response. Delivery in the correct niche environment can further enhance the immunemodulatory effects of MSCs and have positive impact on scar formation. ASCs delivered to cutaneous excisional wounds via an ECM patch attenuate wound fibrosis more effectively than ASCs applied without scaffold support [74].

Despite significant scientific advancements and early clinical trials, clinical translation of progenitor cell-seeded biomimetic scaffolds for skin regeneration still remains a challenge. However, innovative therapies based on emerging concepts arising from the intersection of engineering, molecular signaling, and stem cell biology will potentially result in the transformation of fibrotic healing into skin regeneration. Looking ahead, understanding the genetic and epigenetic indicators that might predispose a patient to impaired wound healing or excessive scarring may enhance tissue regenerative approaches further.

3.3 Conclusions

A growing number of patients suffering from chronic wounds have brought soft tissue regeneration into spotlight of current research. The ideal treatment for wound healing is cheap and effective for most wounds. Furthermore, it is essential for upcoming devices to avoid side effect, to provide long-lasting application and should not be impaired by patient-dependent factors such as chronic systemic diseases. Several different approaches have been developed and have shown promising results in preclinical studies. Nevertheless, only NPWT has made the step to clinical routine. Possible reasons for this big gap between basic science and clinical implementation include several uncertain factors such as possible malignancy (growth factor-based therapy), high degradation rate (RNAi, growth factor, and stem cell-based therapy), systemic side effects (pyrvinium), uncertain retention rates, and individual limitations (stem cell-based therapy). Further approached should strive for a holistic attempt to correct the plethora of molecular defects that lead to nonhealing wounds rather than a one-factor replacement therapy.

References

- Lindholm C, Searle R. Wound management for the 21st century: combining effectiveness and efficiency. Int Wound J. 2016;13(Suppl 2):5–15.
- Lanman TH, Ingalls TH. Vitamin C deficiency and wound healing: an experimental and clinical study. Ann Surg. 1937;105(4):616–25.

- Tiesler V, Coppa A, Zabala P, Cucina A. Scurvyrelated morbidity and death among Christopher Columbus' Crew at La Isabela, the first european town in the new world (1494–1498): an assessment of the skeletal and historical information. Int J Osteoarchaeol. 2014;26(2):191–202.
- Grinnell F, Fukamizu H, Pawelek P, Nakagawa S. Collagen processing, crosslinking, and fibril bundle assembly in matrix produced by fibroblasts in longterm cultures supplemented with ascorbic acid. Exp Cell Res. 1989;181(2):483–91.
- Marti-Carvajal AJ, Gluud C, Nicola S, Simancas-Racines D, Reveiz L, Oliva P, Cedeño-Taborda J. Growth factors for treating diabetic foot ulcers. Cochrane Database Syst Rev. 2015;10:CD008548.
- Senet P, Vicaut E, Beneton N, Debure C, Lok C, Chosidow O. Topical treatment of hypertensive leg ulcers with platelet-derived growth factor-BB: a randomized controlled trial. Arch Dermatol. 2011;147(8):926–30.
- Papanas N, Maltezos E. Benefit-risk assessment of becaplermin in the treatment of diabetic foot ulcers. Drug Saf. 2010;33(6):455–61.
- Galiano RD, Tepper OM, Pelo CR, Bhatt KA, Callaghan M, Bastidas N, Bunting S, Steinmetz HG, Gurtner GC. Topical vascular endothelial growth factor accelerates diabetic wound healing through increased angiogenesis and by mobilizing and recruiting bone marrow-derived cells. Am J Pathol. 2004;164(6):1935–47.
- Hanft JR, Pollak RA, Barbul A, van Gils C, Kwon PS, Gray SM, Lynch CJ, Semba CP, Breen TJ. Phase I trial on the safety of topical rhVEGF on chronic neuropathic diabetic foot ulcers. J Wound Care. 2008;17(1):30–2, 4–7
- Singla S, Garg R, Kumar A, Gill C. Efficacy of topical application of beta urogastrone (recombinant human epidermal growth factor) in Wagner's grade 1 and 2 diabetic foot ulcers: comparative analysis of 50 patients. J Nat Sci Biol Med. 2014;5(2):273–7.
- Fernandez-Montequin JI, Betancourt BY, Leyva-Gonzalez G, Mola EL, Galan-Naranjo K, Ramirez-Navas M, Bermúdez-Rojas S, Rosales F, García-Iglesias E, Berlanga-Acosta J, Silva-Rodriguez R, Garcia-Siverio M, Martinez LH. Intralesional administration of epidermal growth factor-based formulation (Heberprot-P) in chronic diabetic foot ulcer: treatment up to complete wound closure. Int Wound J. 2009;6(1):67–72.
- 12. Fernandez-Montequin JI, Infante-Cristia E, Valenzuela-Silva C, Franco-Perez N, Savigne-Gutierrez W, Artaza-Sanz H, Morejón-Vega L, González-Benavides C, Eliseo-Musenden O, García-Iglesias E, Berlanga-Acosta J, Silva-Rodríguez R, Betancourt BY, López-Saura PA, Cuban Citoprot-P Study Group. Intralesional injections of Citoprot-P (recombinant human epidermal growth factor) in advanced diabetic foot ulcers with risk of amputation. Int Wound J. 2007;4(4):333–43.
- Yang S, Geng Z, Ma K, Sun X, Fu X. Efficacy of topical recombinant human epidermal growth fac-

tor for treatment of diabetic foot ulcer: a systematic review and meta-analysis. Int J Low Extrem Wounds. 2016;15(2):120–5.

- 14. Richard JL, Parer-Richard C, Daures JP, Clouet S, Vannereau D, Bringer J, Rodier M, Jacob C, Comte-Bardonnet M. Effect of topical basic fibroblast growth factor on the healing of chronic diabetic neuropathic ulcer of the foot. A pilot, randomized, doubleblind, placebo-controlled study. Diabetes Care. 1995;18(1):64–9.
- 15. Kanda N, Morimoto N, Ayvazyan AA, Takemoto S, Kawai K, Nakamura Y, Sakamoto Y, Taira T, Suzuki S. Evaluation of a novel collagen-gelatin scaffold for achieving the sustained release of basic fibroblast growth factor in a diabetic mouse model. J Tissue Eng Regen Med. 2014;8(1):29–40.
- Uchi H, Igarashi A, Urabe K, Koga T, Nakayama J, Kawamori R, Tamaki K, Hirakata H, Ohura T, Furue M. Clinical efficacy of basic fibroblast growth factor (bFGF) for diabetic ulcer. Eur J Dermatol. 2009;19(5):461–8.
- Yang Y, Xia T, Chen F, Wei W, Liu C, He S, Li X. Electrospun fibers with plasmid bFGF polyplex loadings promote skin wound healing in diabetic rats. Mol Pharm. 2012;9(1):48–58.
- Tan Y, Xiao J, Huang Z, Xiao Y, Lin S, Jin L, Feng W, Cai L, Li X. Comparison of the therapeutic effects recombinant human acidic and basic fibroblast growth factors in wound healing in diabetic patients. J Health Sci. 2008;54(4):432–40.
- Pakyari M, Farrokhi A, Maharlooei MK, Ghahary A. Critical role of transforming growth factor beta in different phases of wound healing. Adv Wound Care. 2013;2(5):215–24.
- Robson MC, Phillip LG, Cooper DM, Lyle WG, Robson LE, Odom L, Hill DP, Hanham AF, Ksander GA. Safety and effect of transforming growth factorbeta(2) for treatment of venous stasis ulcers. Wound Repair Regen. 1995;3(2):157–67.
- Ingber DE. Mechanical control of tissue growth: function follows form. Proc Natl Acad Sci U S A. 2005;102(33):11571–2.
- Ingber DE. The mechanochemical basis of cell and tissue regulation. Mech Chem Biosyst. 2004;1(1):53–68.
- Nie B, Yue B. Biological effects and clinical application of negative pressure wound therapy: a review. J Wound Care. 2016;25(11):617–26.
- 24. Zhang YG, Wang X, Yang Z, Zhang H, Liu M, Qiu Y, Guo X. The therapeutic effect of negative pressure in treating femoral head necrosis in rabbits. PLoS One. 2013;8(1):e55745.
- 25. Li X, Liu J, Liu Y, Hu X, Dong M, Wang H, Hu D. Negative pressure wound therapy accelerates rats diabetic wound by promoting agenesis. Int J Clin Exp Med. 2015;8(3):3506–13.
- Valente PM, Deva A, Ngo Q, Vickery K. The increased killing of biofilms in vitro by combining topical silver dressings with topical negative pressure in chronic wounds. Int Wound J. 2016;13(1):130–6.
- 27. Dunn JA, McCance DR, Thorpe SR, Lyons TJ, Baynes JW. Age-dependent accumulation of N

epsilon-(carboxymethyl)lysine and N epsilon-(carboxymethyl)hydroxylysine in human skin collagen. Biochemistry. 1991;30(5):1205–10.

- 28. Rhie G, Shin MH, Seo JY, Choi WW, Cho KH, Kim KH, Park KC, Eun HC, Chung JH. Agingand photoaging-dependent changes of enzymic and nonenzymic antioxidants in the epidermis and dermis of human skin in vivo. J Invest Dermatol. 2001;117(5):1212–7.
- Kohen R. Skin antioxidants: their role in aging and in oxidative stress—new approaches for their evaluation. Biomed Pharmacother. 1999;53(4):181–92.
- Harman D. Aging: a theory based on free radical and radiation chemistry. J Gerontol. 1956;11(3):298–300.
- Harman D. The free radical theory of aging. Antioxid Redox Signal. 2003;5(5):557–61.
- Surjushe A, Vasani R, Saple D. Aloe vera: a short review. Indian J Dermatol. 2008;53(4):163.
- Hodges YK, Reese SM, Pahl PM, Horwitz LD. Paradoxical effects of iron chelation on growth of vascular endothelial cells. J Cardiovasc Pharmacol. 2005;45(6):539–44.
- 34. Pepe A, Meloni A, Capra M, Cianciulli P, Prossomariti L, Malaventura C, Putti MC, Lippi A, Romeo MA, Bisconte MG, Filosa A, Caruso V, Quarta A, Pitrolo L, Missere M, Midiri M, Rossi G, Positano V, Lombardi M, Maggio A. Deferasirox, deferiprone and desferrioxamine treatment in thalassemia major patients: cardiac iron and function comparison determined by quantitative magnetic resonance imaging. Haematologica. 2011;96(1):41–7.
- 35. Kuo KH, Mrkobrada M. A systematic review and meta-analysis of deferiprone monotherapy and in combination with deferoxamine for reduction of iron overload in chronically transfused patients with beta-thalassemia. Hemoglobin. 2014;38(6):409–21.
- Moayedi Esfahani BA, Reisi N, Mirmoghtadaei M. Evaluating the safety and efficacy of silymarin in beta-thalassemia patients: a review. Hemoglobin. 2015;39(2):75–80.
- 37. Ram M, Singh V, Kumawat S, Kumar D, Lingaraju MC, Uttam Singh T, Rahal A, Tandan SK, Kumar D. Deferoxamine modulates cytokines and growth factors to accelerate cutaneous wound healing in diabetic rats. Eur J Pharmacol. 2015;764:9–21.
- Temiz G, Sirinoglu H, Yesiloglu N, Filinte D, Kacmaz C. Effects of Deferoxamine on fat graft survival. Facial Plast Surg. 2016;32(4):438–43.
- 39. Lu H, Dalgard CL, Mohyeldin A, McFate T, Tait AS, Verma A. Reversible inactivation of HIF-1 prolyl hydroxylases allows cell metabolism to control basal HIF-1. J Biol Chem. 2005;280(51):41928–39.
- 40. Duscher D, Neofytou E, Wong VW, Maan ZN, Rennert RC, Inayathullah M, Januszyk M, Rodrigues M, Malkovskiy AV, Whitmore AJ, Walmsley GG, Galvez MG, Whittam AJ, Brownlee M, Rajadas J, Gurtner GC. Transdermal deferoxamine prevents pressure-induced diabetic ulcers. Proc Natl Acad Sci U S A. 2015;112(1):94–9.

- Miller JR, Moon RT. Signal transduction through beta-catenin and specification of cell fate during embryogenesis. Genes Dev. 1996;10(20):2527–39.
- Brennan KR, Brown AM. Wnt proteins in mammary development and cancer. J Mammary Gland Biol Neoplasia. 2004;9(2):119–31.
- Barham W, Frump AL, Sherrill TP, Garcia CB, Saito-Diaz K, VanSaun MN, Fingleton B, Gleaves L, Orton D, Capecchi MR, Blackwell TS, Lee E, Yull F, Eid JE. Targeting the Wnt pathway in synovial sarcoma models. Cancer Discov. 2013;3(11):1286–301.
- 44. Leavitt T, Hu MS, Marshall CD, Barnes LA, Lorenz HP, Longaker MT. Scarless wound healing: finding the right cells and signals. Cell Tissue Res. 2016;365(3):483–93.
- Saraswati S, Alfaro MP, Thorne CA, Atkinson J, Lee E, Young PP. Pyrvinium, a potent small molecule Wnt inhibitor, promotes wound repair and post-MI cardiac remodeling. PLoS One. 2010;5(11):e15521.
- 46. Thorne CA, Hanson AJ, Schneider J, Tahinci E, Orton D, Cselenyi CS, Jernigan KK, Meyers KC, Hang BI, Waterson AG, Kim K, Melancon B, Ghidu VP, Sulikowski GA, LaFleur B, Salic A, Lee LA, Miller DM 3rd, Lee E. Small-molecule inhibition of Wnt signaling through activation of casein kinase 1alpha. Nat Chem Biol. 2010;6(11):829–36.
- 47. Saraswati S, Deskins DL, Holt GE, Young PP. Pyrvinium, a potent small molecule Wnt inhibitor, increases engraftment and inhibits lineage commitment of mesenchymal stem cells (MSCs). Wound Repair Regen. 2012;20(2):185–93.
- 48. Plikus MV, Guerrero-Juarez CF, Ito M, Li YR, Dedhia PH, Zheng Y, Shao M, Gay DL, Ramos R, Hsi TC, Oh JW, Wang X, Ramirez A, Konopelski SE, Elzein A, Wang A, Supapannachart RJ, Lee HL, Lim CH, Nace A, et al. Regeneration of fat cells from myofibroblasts during wound healing. Science. 2017;355(6326):748–52.
- Plikus MV, Mayer JA, de la Cruz D, Baker RE, Maini PK, Maxson R, Chuong CM. Cyclic dermal BMP signalling regulates stem cell activation during hair regeneration. Nature. 2008;451(7176):340–4.
- Chan CK, Longaker MT. Fibroblasts become fat to reduce scarring. Science. 2017;355(6326):693–4.
- Banerjee J, Chan YC, Sen CK. MicroRNAs in skin and wound healing. Physiol Genomics. 2011;43(10):543–56.
- 52. Lai WF, Siu PM. MicroRNAs as regulators of cutaneous wound healing. J Biosci. 2014;39(3):519–24.
- 53. Yu J, Ryan DG, Getsios S, Oliveira-Fernandes M, Fatima A, Lavker RM. MicroRNA-184 antagonizes microRNA-205 to maintain SHIP2 levels in epithelia. Proc Natl Acad Sci USA. 2008;105(49):19300–5.
- 54. Wang XH, Qian RZ, Zhang W, Chen SF, Jin HM, Hu RM. MicroRNA-320 expression in myocardial microvascular endothelial cells and its relationship with insulin-like growth factor-1 in type 2 diabetic rats. Clin Exp Pharmacol Physiol. 2009;36(2):181–8.
- Dykxhoorn DM, Palliser D, Lieberman J. The silent treatment: siRNAs as small molecule drugs. Gene Ther. 2006;13(6):541–52.

- 56. Shaw TJ, Martin P. Wound repair at a glance. J Cell Sci. 2009;122(Pt 18):3209–13.
- 57. Liu X, Ma L, Liang J, Zhang B, Teng J, Gao C. RNAi functionalized collagen-chitosan/silicone membrane bilayer dermal equivalent for full-thickness skin regeneration with inhibited scarring. Biomaterials. 2013;34(8):2038–48.
- Brummelkamp TR, Bernards R, Agami R. Stable suppression of tumorigenicity by virus-mediated RNA interference. Cancer Cell. 2002;2(3):243–7.
- 59. Ding H, Schwarz DS, Keene A, Affar e B, Fenton L, Xia X, Shi Y, Zamore PD, Xu Z. Selective silencing by RNAi of a dominant allele that causes amyotrophic lateral sclerosis. Aging Cell. 2003;2(4):209–17.
- Toloue MM, Ford LP. Antibody targeted siRNA delivery. Methods Mol Biol. 2011;764:123–39.
- Gary DJ, Puri N, Won YY. Polymer-based siRNA delivery: perspectives on the fundamental and phenomenological distinctions from polymer-based DNA delivery. J Control Release. 2007;121(1–2):64–73.
- 62. Walmsley GG, Maan ZN, Wong VW, Duscher D, Hu MS, Zielins ER, Wearda T, Muhonen E, McArdle A, Tevlin R, Atashroo DA, Senarath-Yapa K, Lorenz HP, Gurtner GC, Longaker MT. Scarless wound healing: chasing the holy grail. Plast Reconstr Surg. 2015;135(3):907–17.
- 63. Scarabel L, Perrone F, Garziera M, Farra R, Grassi M, Musiani F, Russo Spena C, Salis B, De Stefano L, Toffoli G, Rizzolio F, Tonon F, Abrami M, Chiarappa G, Pozzato G, Forte G, Grassi G, Dapas B. Strategies to optimize siRNA delivery to hepatocellular carcinoma cells. Expert Opin Drug Deliv. 2017;14(6):797–810.
- 64. Rennert RC, Sorkin M, Januszyk M, Duscher D, Kosaraju R, Chung MT, Lennon J, Radiya-Dixit A, Raghvendra S, Maan ZN, Hu MS, Rajadas J, Rodrigues M, Gurtner GC. Diabetes impairs the angiogenic potential of adipose-derived stem cells by selectively depleting cellular subpopulations. Stem Cell Res Ther. 2014;5(3):79.
- 65. Duscher D, Rennert RC, Januszyk M, Anghel E, Maan ZN, Whittam AJ, Perez MG, Kosaraju R, Hu MS, Walmsley GG, Atashroo D, Khong S, Butte AJ, Gurtner GC. Aging disrupts cell subpopulation dynamics and diminishes the function of mesenchymal stem cells. Sci Rep. 2014;4:7144.

- 66. Hadjipanayi E, Schilling AF. Regeneration through autologous hypoxia preconditioned plasma. Organogenesis. 2014;10(2):164–9.
- 67. Hadjipanayi E, Bauer AT, Moog P, Salgin B, Kuekrek H, Fersch B, Hopfner U, Meissner T, Schlüter A, Ninkovic M, Machens HG, Schilling AF. Cell-free carrier system for localized delivery of peripheral blood cell-derived engineered factor signaling: towards development of a one-step device for autologous angiogenic therapy. J Control Release. 2013;169(1–2):91–102.
- Yildirimer L, Thanh NT, Seifalian AM. Skin regeneration scaffolds: a multimodal bottom-up approach. Trends Biotechnol. 2012;30(12):638–48.
- 69. Wong VW, Rustad KC, Galvez MG, Neofytou E, Glotzbach JP, Januszyk M, Major MR, Sorkin M, Longaker MT, Rajadas J, Gurtner GC. Engineered pullulan-collagen composite dermal hydrogels improve early cutaneous wound healing. Tissue Eng Part A. 2011;17(5–6):631–44.
- Rennert RC, Rodrigues M, Wong VW, Duscher D, Hu M, Maan Z, Sorkin M, Gurtner GC, Longaker MT. Biological therapies for the treatment of cutaneous wounds: phase III and launched therapies. Expert Opin Biol Ther. 2013;13(11):1523–41.
- Wong VW, Rustad KC, Glotzbach JP, Sorkin M, Inayathullah M, Major MR, Longaker MT, Rajadas J, Gurtner GC. Pullulan hydrogels improve mesenchymal stem cell delivery into high-oxidative-stress wounds. Macromol Biosci. 2011;11(11):1458–66.
- Rustad KC, Wong VW, Sorkin M, Glotzbach JP, Major MR, Rajadas J, Longaker MT, Gurtner GC. Enhancement of mesenchymal stem cell angiogenic capacity and stemness by a biomimetic hydrogel scaffold. Biomaterials. 2012;33(1):80–90.
- 73. Garg RK, Rennert RC, Duscher D, Sorkin M, Kosaraju R, Auerbach LJ, Lennon J, Chung MT, Paik K, Nimpf J, Rajadas J, Longaker MT, Gurtner GC. Capillary force seeding of hydrogels for adipose-derived stem cell delivery in wounds. Stem Cells Transl Med. 2014;3:1079–89.
- 74. Lam MT, Nauta A, Meyer NP, Wu JC, Longaker MT. Effective delivery of stem cells using an extracellular matrix patch results in increased cell survival and proliferation and reduced scarring in skin wound healing. Tissue Eng Part A. 2013;19(5–6):738–47.