# **Chapter 9 Emergence of Invasive Fungal Infection: Diagnosis and Treatment in Humans**



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# 9.1 Introduction

Fungi are opportunistic pathogens (endogenous or exogeneous) that are freely present in the environment and cause either no or very mild symptoms in healthy individuals, but manifest a fatal physical morbid state in immunocompromised individuals. Invasive fungal infections are those infections wherein fungal pathogens have invaded and colonized the deep tissues, which results in prolonged illness (Badiee and Hashemizadeh 2014). The Invasive Fungal Infections Cooperative group (IFICG) of the European Organisation for Research and Treatment of Cancer (EORTC) and the Mycology Study Group (MSG) of the National Institute of Allergy and Infectious Diseases (NIAID) define invasive fungal infections as the presence of fungal elements either as mold or yeast in deep tissues or needle aspiration biopsy that is further confirmed by culturing and histopathological examination (Ramana et al. 2013). The innate immune system of healthy individuals combats the invasion from fungal infection and does not acquire invasive fungal infections, except exhibiting mild symptoms. The reason for this is that the skin barrier of healthy individuals and the innate and adaptive immunities provide defense against these infections in most normal situations. In the case of patients with severe burns or severely impaired immune system, the chances of invasive fungal infections increase significantly. Invasive fungal infections can affect any part of the body and result in morbidity as well as enhanced mortality chances in immunocompromised individuals due to weakened immune system in response to these infections. These individuals can also have multiple comorbidities (Hope et al. 2013). The number of

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patients at risk of suffering from invasive fungal infections is increasing worldwide. Research in this area has attracted attention due to: (a) the complex nature of infections; (b) often there being multiple infections occurring at the same time; and (c) its diagnosis and treatment. In addition to the immunocompromised individuals, the patients undergoing immunosuppressive therapy, major surgery, hematopoietic stem cell transplantation (HSCT), and the patients with neoplastic diseases, acquired immune deficiency syndrome (AIDS), chronic pulmonary diseases, etc. form a high-risk group of individuals (Fig. 9.1) (Badiee and Hashemizadeh 2014; Person et al. 2010). Invasive fungal infections have also been increasingly reported in individuals with grafts, prosthetic devices, and also in the patients who underwent any major aggressive surgery or organ transplantation (Person et al. 2010). There are several factors that can increase the chances of invasive fungal infections in individuals. One of the major reasons is the use of broad-spectrum antibiotics and antineoplastic and immunosuppressive agents (Enoch et al. 2006; Ravikumar et al. 2015). The increased predisposition of patients with pancreatitis, human immunodeficiency virus (HIV) infection, burns, and neutropenia to fungal infection are other reasons for increased risk of invasive fungal infections (Ruping et al. 2008). Various reports have demonstrated that the chances of invasive fungal infections are also determined by several factors, which include both genetic as well as environmental conditions (Maskarinec et al. 2016; Benedict and Park 2014).

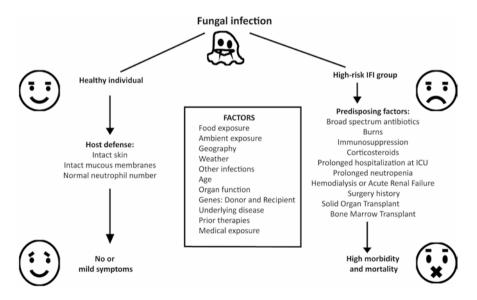


Fig. 9.1 Factors promoting invasive fungal infections and the formation of a high-risk invasive fungal infection group

# 9.2 Invasive Fungal Infections

The most important fungal pathogens causing invasive fungal infections are yeasts (Candida spp., Cryptococcus spp., and Pneumocystis spp.) and molds (Aspergillus spp., Fusarium spp., Scedosporium prolificans, Mucor, Rhizopus, Rhizomucor, and Absidia) (Ramana et al. 2013). Of these, Cryptococcus, Candida, Aspergillus, and Pneumocystis collectively cause more than 90% of the deaths due to invasive fungal infections globally (Brown et al. 2012). These infections can be transmitted by the inhalation of spores, penetration into the mucosa by commensal organisms, such as Candida albicans, and the ingestion of contaminated food (Badiee and Hashemizadeh 2014). The resulting infection might be mild or life-threatening, depending upon various factors, as shown in Fig. 9.1. Invasive fungal infections are associated with high morbidity and mortality rates. For instance, mortality associated with invasive candidiasis varies from 50 to 75%, while it can reach almost 100% in the case of invasive aspergillosis (Pfaller and Diekema 2007). Delay in antifungal therapy to treat invasive fungal infections also increases mortality and, therefore, requires early diagnosis and medical intervention, but both of which are very challenging (Badiee and Hashemizadeh 2014; Morrell et al. 2005). Table 9.1 lists the major invasive fungal infections and shows the statistics for invasive fungal infections worldwide.

In the following sections, the major invasive fungal infections and their diagnosis and treatment will be discussed.

### 9.2.1 Aspergillosis

Aspergillosis is the most common fungal infection in humans and is caused by a group of filamentous fungi, *Aspergillus* (Hartwick and Batsakis 1991; Schmiedel and Zimmerli 2016). *Aspergillus* is mostly found in soil, food, air, water, and

Disease	Causative agent	Mortality rates in infected populations (%)
Aspergillosis	Aspergillus fumigatus, Aspergillus niger, and Aspergillus flavus	30–95
Candidiasis	Candida albicans, Candida glabrata, Candida parapsilosis, Candida tropicalis, and Candida krusei	46–75
Cryptococcosis	Cryptococcus neoformans and Cryptococcus gattii	20–70
Mucormycosis	Rhizopus oryzae, Rhizopus microsporus, and Rhizomucor pusillus	30–90
Pneumocystis	Pneumocystis jirovecii	20-80

Table 9.1 Statistics of the most significant invasive fungal infections (Brown et al. 2012)

decomposing matter (Anaissie et al. 2002; Tamgadge et al. 2012) and cause a gradual destructive disease of the lungs called chronic pulmonary aspergillosis, which complicates other pulmonary-related conditions or diseases, such as tuberculosis, chronic obstructive pulmonary disease (COPD), and systemic inflammatory disease. There are several species of this genus but A. fumigatus, A. niger, and A. flavus are the most common causative agents of aspergillosis (Sethi et al. 2012). According to the guidelines of the Infectious Diseases Society of America (IDSA) for the treatment of aspergillosis, there are three major types of aspergillosis: invasive aspergillosis, pulmonary aspergilloma, and allergic bronchopulmonary aspergillosis (ABPA) (Stevens et al. 2000). Pulmonary aspergilloma, a saprophytic form of aspergillosis, usually develops in a pre-existing cavity in the lung and results from the ingrowth of colonized Aspergillus, while allergic bronchopulmonary aspergillosis is an immunological pulmonary disorder caused by hypersensitivity to Aspergillus fumigatus (Agarwal et al. 2013). However, there are also reports showing that, under certain circumstances, the colonization of airways by Aspergillus species can transform into an invasive disease (Shahi et al. 2015; Gago et al. 2018). Invasive aspergillosis is a devastating infection and is highly lethal in immunocompromised individuals. The lungs are the most common sites of primary invasive disease, while the central nervous system (CNS) is the secondary site of this invasive disease (Anagnostou et al. 2014; Mohammadi et al. 2015). Invasive aspergillosis leads to a mortality rate of around 50% even if diagnosed and treated, but delayed diagnosis may be 100% fatal (Latge 1999).

### 9.2.1.1 Diagnosis

Diagnosing aspergillosis remains an obstacle and early diagnosis in immunocompromised individuals is also one of the major challenges. Aspergillus is common in the environment and can be found in the saliva and sputum of healthy people. The symptoms of aspergillosis are similar to those of tuberculosis. Therefore, the isolation of Aspergillus spp. in sputum samples is highly determined by the immune status of the host (Garnacho-Montero et al. 2005). For instance, invasive aspergillosis cannot be ruled out by negative sputum samples; on the other hand, invasive disease can be easily established in immunocompromised patients. Apart from this, as high as 70% of negative sputum samples could still have invasive aspergillosis disease (Tang and Cohen 1992; Yu et al. 1986). Histopathological examination followed by bronchoalveolar lavage, transthoracic needle biopsy (TNB), or videoassisted thoracoscopic surgery are standard procedures for the diagnosis of invasive aspergillosis (Walsh et al. 2008). Chest X-ray and computed tomography (CT) scan are good tools to diagnose aspergilloma as well as the characteristic signs of invasive and allergic bronchopulmonary aspergillosis, but they have some limitations (Hope et al. 2013). Chest X-ray is of less significance in the early stages of invasive disease, while the CT scan in combination with high-resolution images (HRCT) is more useful (Caillot et al. 1997, 2001). A recent advancement in the diagnosis of invasive aspergillosis is the detection of *Aspergillus* antigens, such as galactomannan and  $(1 \rightarrow 3)$ -beta-D-glucan in the body fluids of patients. A double-sandwich enzyme-linked immunosorbent assay (ELISA) kit approved by the U.S. Food and Drug Administration (US FDA) for the detection of galactomannan that is released by *Aspergillus* during its growth in the patient's serum provides a non-culture-based diagnosis of invasive aspergillosis (Boutboul et al. 2002; Marr et al. 2004a). In this test, the patient's sputum is stained with a dye, which identifies the presence of *Aspergillus*.

### 9.2.1.2 Treatment

Oral corticosteroids are one of the best ways to treat bronchopulmonary aspergillosis (Rosenberg et al. 1978; Wang et al. 1979; Capewell et al. 1989). Antifungal medications are not very helpful in allergic bronchopulmonary aspergillosis, but combination with corticosteroids can improve lung condition and its functioning. In certain situations of aspergilloma, surgical resection is required to remove the fungal mass, but the surgical route should be avoided, as it is risky (Uflacker et al. 1985; Soltanzadeh et al. 1977; Massard et al. 1992).

The management of invasive pulmonary aspergillosis is difficult. Voriconazole is considered the first-line therapy for invasive aspergillosis infections caused by these pathogens (Johnson and Kauffman 2003; Sambatakou et al. 2006; Ghannoum and Kuhn 2002; Herbrecht et al. 2002; Limper et al. 2011; Walsh et al. 2008). Voriconazole, a broad-spectrum triazole, is available in both intravenous and oral formulations. Although voriconazole has a milder side-effect profile, it can potentially interact with many significant drugs, such as cyclosporine, warfarin, terfenadine, carbamazepine, quinidine, rifampin, statins, and sulfonylureas (Johnson and Kauffman 2003; Sambatakou et al. 2006; Ghannoum and Kuhn 2002; Herbrecht et al. 2006; Channoum and Kuhn 2002; Herbrecht et al. 2007; Limper et al. 2011).

Isavuconazole, a newer triazole, has been approved for the treatment of invasive aspergillosis and is more tolerable than voriconazole (Spitzer et al. 2017). Amphotericin B products can be used as second-line agents in patients who cannot tolerate or are failing voriconazole therapy (Cornely et al. 2007). To reduce the side effects associated with amphotericin, which include nephrotoxicity, electrolyte disturbances, and hypersensitivity, a new lipid-based preparation of amphotericin B, such as liposomal amphotericin B and lipid complex amphotericin B, has been introduced and used for treatment (Cornely et al. 2007). Echinocandin derivatives such as caspofungin, micafungin and anidulafungin are also effective agents in the treatment of invasive aspergillosis and can be used as salvage therapy. Another broad-spectrum triazole, posaconazole, is effective and safe as salvage therapy in patients with invasive pulmonary aspergillosis refractory to standard antifungal therapy (Pitisuttithum et al. 2005; Spanakis et al. 2006; Cohen-Wolkowiez et al. 2006). The combined therapy of caspofungin and liposomal amphotericin B is more successful as a primary rather than a salvage therapy (Koulenti et al. 2014). In case

of failure of this therapy, the combination of caspofungin and voriconazole is also recommended (Marr et al. 2004b). Combinations of antifungal therapies have also been used as salvage therapy in many high-risk patients (Elizabeth et al. 2015).

# 9.2.2 Candidiasis

Candida species are the most common causative agents of fungal infections, which can range from a non-life-threatening state such as mucocutaneous illnesses to invasive infection that may affect many organs. There are more than 17 different species of Candida that have been identified to cause invasive candidiasis and, of these species, only five species (C. albicans, C. glabrata, C. parapsilosis, C. tropicalis, and C. krusei) are responsible for more than 90% of invasive candidiasis (Pfaller and Diekema 2007). Among these *Candida* species, *C. albicans* is the most common infectious agent, while C. glabrata is the second most common species causing invasive candidiasis (Achkar and Fries 2010; Pfaller and Diekema 2007). Candida species form a highly structured biofilm under different environmental conditions. Surface-associated Candida can grow embedded in extracellular matrix or biofilm, which is composed of carbohydrates and proteins (Achkar and Fries 2010). Candida albicans is distinguished from many other fungal species by its ability to form both yeast cells and hyphae. Hyphae are an important structural component of C. albicans biofilm formation (Nobile and Johnson 2015). This is the most common member of the human microbiota, appearing in the oral cavity, gastrointestinal tract, female genital tract, or on the skin (Achkar and Fries 2010; Ganguly and Mitchell 2011; Nobile and Johnson 2015). In the healthy human population, C. albicans is harmless, while under the alteration of immune defenses, it overgrows and causes serious systemic disease and organ failure (Prieto and Pla 2015). Candida albicans also forms biofilm along with Staphylococcus aureus on the surface of implantable medical devices or organs (Peters et al. 2010). Such multispecies infection leads to higher mortality rates than single-species infection (Zago et al. 2015). In addition, infections caused by C. albicans can be acquired at hospitals and have become a cause of major health concerns. Candida albicans is identified as the fourth most common blood isolate in US hospitals, accounting for around 10% of hospitalacquired bloodstream infections (Horn et al. 2012).

### 9.2.2.1 Diagnosis

The diagnosis of invasive candidiasis is difficult and complicated because there are no specific clinical manifestations of the disease. Researchers have developed polymerase chain reaction (PCR)-based methods for the detection of *Candida* species in blood samples (Mirhendi and Makimura 2003), DNA-based fluorescence in-situ hybridization (FISH) (Bisha et al. 2011), as well as ELISA for the diagnosis of candidiasis infections (Lain et al. 2007).

#### 9.2.2.2 Treatment

The treatment of *Candida* infections varies substantially and is determined by the anatomic location of the infection and the immunity status of the infected individual. Polyenes, triazoles (fluconazole, itraconazole, voriconazole, and posaconazole), echinocandins (caspofungin, anidulafungin, and micafungin), and flucytosine are the most common antifungal agents for the effective treatment of candidiasis (Pappas et al. 2007, 2009; Pfaller and Diekema 2007). Fluconazole is considered a first-line agent in non-neutropenic patients with invasive candidiasis. Amphotericin B is used for non-albicans species (e.g., *C. krusei*).

### 9.2.3 Cryptococcosis

Cryptococcosis is the one of the invasive fungal infections that cause significant morbidity and mortality in both immunocompromised and immunocompetent individuals worldwide. Cryptococcosis is caused by basidiomycetous fungi, Cryptococcus species. The two most common species that cause cryptococcosis are C. neoformans, which mostly affects immunocompromised individuals, and C. gat*tii*, which can infect both immunocompromised as well as immunocompetent individuals (Oladele et al. 2017). These species are further divided into different genotypes (VNI, VII, VNB, VNIV, and VGI-IV) (Perfect and Bicanic 2015). The capsule of C. neoformans is made up of two polysaccharides, glucuronoxylomannan (GXM) and galactoxylomannan (GalXM), which act as a virulent factor (Zaragoza and Casadevall 2004; Bose et al. 2003; Zaragoza et al. 2009). Infection is caused by inhalation of these fungal spores, so the primary localization site is the lung, from where it spreads through the bloodstream to the CNS. Cryptococcus species have the propensity to locate in the CNS and cause fungal meningitis. Pulmonary cryptococcosis may be asymptomatic but meningitis is a common feature of infection, especially in HIV patients and solid-organ transplant recipients (Enoch et al. 2006). For instance, the HIV-infected patients are at higher risk of contracting cryptococcal infection, while the non-HIV patients also get infected by Cryptococcus spp., more often if have a history of transplantation or immunosuppressive therapies.

#### 9.2.3.1 Diagnosis

Histopathology varies according to the immunological status of the host. In immunocompetent individuals, typical granulomas with multinucleated giant cells are formed at the site of cryptococcal infection, while in immunosuppressed hosts, cryptococcosis infects the CNS, skin, and the oral mucous membrane. The definitive diagnosis of cryptococcosis is established with periodic acid Schiff (PAS), mucicarmine stain, and Fontana-Masson stain. The fungal wall stains with the PAS stain, mucicarmine stains the fungal capsule, while Fontana-Masson stains the fungal growth itself because it contains melanin, which is characteristic of cryptococci (Guarner and Brandt 2011). Diagnosis can also be done by the detection of fungal growth in cerebrospinal fluid (CSF) with India ink examination, isolation in tissue culture, and detection of cryptococcal polysaccharide capsular antigen (CrAg) in the serum and in the CSF through latex agglutination or ELISA (Oladele et al. 2017; Amaral et al. 2016).

### 9.2.3.2 Treatment

For many decades, amphotericin B has been used for treatment in transplant recipients and HIV patients (Sloan and Parris 2014). At present, the combination of liposomal amphotericin B and flucytosine (5-FC) treatment is considered as standard treatment due to its better performance in treating cryptococcal meningitis as compared to amphotericin B treatment alone (Larsen et al. 1990; de Gans et al. 1992; Brouwer et al. 2004; Dromer et al. 2008; Day et al. 2013; Sloan and Parris 2014). This combination is also a good treatment of choice for HIV-infected patients. This combination therapy improved early fungicidal activity (EFA) as well as lowered toxicity. However, although the high doses of amphotericin B and 5-FC have improved fungicidal activity, an increase in serious side effects has been reported (Bicanic et al. 2008). Fluconazole is also recommended as an alternative to flucytosine (Bellmann and Smuszkiewicz 2017; Perfect et al. 2010; Elizabeth et al. 2015). During combination of amphotericin B and fluconazole, higher fluconazole doses gave better outcomes compared to lower fluconazole doses (Yao et al. 2014; Elizabeth et al. 2015). Apart from this, other azoles, such as itraconazole, voriconazole, isavuconazole, and posaconazole, have also been tested for the treatment of cryptococcosis, but they seem unreliable (Chotmongkol and Jitpimolmard 1992; Pitisuttithum et al. 2005; Thompson 3rd et al. 2016; de Gans et al. 1992). Out of these, voriconazole has better absorption and penetration to the CNS than itraconazole and posaconazole and it has been used for cryptococcosis treatment in normal individuals (Yao et al. 2015). The survivors of cryptococcal meningitis HIV-infected patients have notably higher concentrations of cytokine stimulating factor, such as IFN- $\gamma$ , TNF- $\alpha$ , IL-6 and -8, than the non-survivors of cryptococcal meningitis HIVinfected patients (Siddiqui et al. 2005). Researchers also found that the combination of liposomal amphotericin B, voriconazole, and IFN- $\gamma$  is successful in the treatment of cryptococcal meningitis in HIV patients (Gamaletsou et al. 2012). In addition to all the antifungal treatments, a new technology is in progress for cryptococcal meningitis treatment. This technology, in combination with the antifungal therapies, would prove to be better for cryptococcal meningitis management (Cutshaw et al. 2016).

### 9.2.4 Mucormycosis

Mucormycosis is an infection caused by members of the order Mucorales (Ibrahim et al. 2012). Mucorales are distributed into six families and all members of the six families cause cutaneous and deep infections. *Rhizopus oryzae*, *R. microsporus*, and *R. pusillus* are the most common species isolated from patients with mucormycosis and are responsible for around 70% of all cases of mucormycosis. This causative agent is ubiquitous in nature and mostly found in soil and decaying organic matter. Because of the abundance of this fungus, the mouth and the nose inhale the spores very often. In a normal person, the macrophage cells phagocytize the spores, while in immunocompromised individuals, the spores germinate into the hyphae and cause infections in the sinuses and lungs.

### 9.2.4.1 Different Forms of Mucormycosis

Mucormycosis invades the blood vessels, producing thrombosis and tissue infarction. It causes five major forms of infections (Prabhu and Patel 2004): (i) rhinoorbitocerebral, (ii) pulmonary, (iii) disseminated, (iv) cutaneous, and (v) gastrointestinal. Rhino-orbitocerebral is the most common form of mucormycosis, while pulmonary mucormycosis is a rare pulmonary fungal infection and difficult to diagnose. After aspergillosis and candidiasis, mucormycosis is the third most common invasive fungal infection (Hammond et al. 2011).

#### 9.2.4.1.1 Rhino-orbitocerebral Mucormycosis

This is the most common form of mucormycosis and is mostly found in diabetic patients. The disease starts from the nose and progresses into the sinuses, orbit, and then intracranially. The initial symptoms are sinusitis or periorbital cellulitis, facial pain, blurry vision, and soft tissue swelling. If untreated, infection spreads from the ethmoid sinus to the orbit, resulting in loss of extraocular muscle function (Spellberg et al. 2005). This is also divided into two major forms: (i) highly fatal rhino-orbitocerebral and (ii) less fatal rhino-orbitocerebral. The former involves ophthalmic and internal carotid arteries, while the latter involves the sphenopalatine and greater palatine arteries, resulting in thrombosis of turbinates and necrosis of the palate (Yanagisawa et al. 1977).

#### 9.2.4.1.2 Pulmonary Mucormycosis

Pulmonary mucormycosis is the most common form found in neutropenic and stem cell-transplant patients. It may develop as a result of inhalation or by hematogenous or lymphatic spread. Its symptoms include dyspnea, cough, and chest pain. Patients

with untreated pulmonary mucormycosis usually die from disseminated disease before respiratory failure occurs (Tedder et al. 1994). The overall mortality rate of pulmonary mucormycosis is approximately 50–70%, but is >95% if the pulmonary mucormycosis is part of a disseminated process (Spellberg et al. 2005).

### 9.2.4.1.3 Disseminated Mucormycosis

Disseminated mucormycosis may originate from any primary site of infection. Pulmonary mucormycosis has the highest incidence of dissemination. The most common site of dissemination is the brain, but metastatic lesions may also be found in the spleen, heart, skin, and other organs. The mortality associated with dissemination to the brain is about 100%. Even without CNS involvement, disseminated mucormycosis has a mortality of >90%. The diagnosis of disseminated disease is difficult because patients are usually severely ill from multiple diseases. When disseminated mucormycosis is suspected, a careful search should be made for cutaneous lesions that can be biopsied for diagnostic purposes (Spellberg et al. 2005).

# 9.2.4.1.4 Cutaneous Mucormycosis

The mucormycosis agent cannot penetrate the skin but individuals with a disrupted protective cutaneous barrier are at high risk of cutaneous mucormycosis. Cutaneous mucormycosis can be very invasive and penetrate from the cutaneous and subcutaneous tissues into the adjacent fat, muscle, and even bone (Spellberg et al. 2005).

### 9.2.4.1.5 Gastrointestinal Mucormycosis

Gastrointestinal mucormycosis is a rare form of mucormycosis. It mainly occurs in extremely malnourished individuals, especially infants or children. The stomach, colon, and ileum are the most commonly involved sites during gastrointestinal mucormycosis. Non-specific abdominal pain and distention associated with nausea and vomiting are the most common symptoms (Spellberg et al. 2005).

# 9.2.4.2 Diagnosis

The diagnosis of mucormycosis is challenging and treatment should start as early as possible in order to decrease mortality. Although the diagnosis is difficult, some tests such as CT scan or magnetic resonance imaging (MRI) may be helpful to define the infections or tissue destruction. Histopathology is the gold standard for this diagnosis. The detection of fungal DNA in tissue samples by PCR is a non-culture-based method that may allow improved diagnosis of mucormycosis (Hammond et al. 2011).

#### 9.2.4.3 Treatment

There are some important factors for the eradication of mucormycosis : rapidity of diagnosis, appropriate surgical removal of infected tissue, and appropriate antifungal therapy. For patients with mucormycosis, surgical treatment plus antifungal therapy is better than the use of antifungal therapy alone (Spellberg et al. 2005). Polyene-based antifungal therapy, such as amphotericin B, is the primary therapy for mucormycosis treatment. Amphotericin B deoxycholate (d-AmB) is the only antifungal agent for the treatment of mucormycosis that has been approved by the US FDA. d-AmB has been replaced by the various lipid formulations of amphotericin. Posaconazole is better than itraconazole and isavuconazole. Fluconazole and voriconazole have no meaningful activity against agents of mucormycosis. No other azoles except for posaconazole are recommended in the treatment of mucormycosis. Flucytosine lacks activity against agents of mucormycosis (Skiada et al. 2013). Liposomal amphotericin B is frequently used and is the most effective drug against mucormycosis. Lipid amphotericin B is also combined with echinocandins to treat mucormycosis (Spellberg and Ibrahim 2010).

### 9.2.5 Pneumocystis Pneumonia

Pneumocystis pneumonia is an opportunistic infection that occurs in immunosuppressed individuals, primarily individuals with HIV infections, caused by the invasion of the unicellular fungus, *Pneumocystis jirovecii*. Initially, it was misclassified as protozoan, on the basis of its morphology, but, later, it was considered as fungi (Thomas Jr. and Limper 2004). *Pneumocystis* isolated from one species cannot infect another species, meaning that the microbe is host-specific. There are five major species-specific *Pneumocystis* species: *P. carinii* and *P. wakefieldiae* in rats, *P. murina* in mice, *P. oryctolagi* in rabbits, and *P. jirovecii* in humans (Aliouat-Denis et al. 2008; Sokulska et al. 2015). Transmission occurs via aerosols from patients with pneumonia or from early-life contact with family or community members who carry the organism in their lungs (Brown et al. 2012). Its presence in lungs is asymptomatic. Pneumocystis pneumonia has been one of the main causes of morbidity and mortality among HIV-infected people (Sokulska et al. 2015), but the death rate in non-HIV individuals (30–60%) is significant higher than that in HIV-infected individuals (Sokulska et al. 2015; Schmiedel and Zimmerli 2016).

### 9.2.5.1 Diagnosis

Pneumocystis pneumonia is difficult to diagnose because of the non-specific symptoms and signs of infection. The diagnosis of pneumocystis pneumonia requires microscopic detection of *P. jirovecii* in respiratory tract specimens, followed by staining with dyes or antibodies. Cysts or trophozoites are morphologically identified by methenamine silver nitrate or Giemsa stains (Sowden and Carmichael 2004). Pneumocystis from sputum, bronchoalveolar fluid, or lung tissue can also be identified by reverse transcription (RT)-PCR. Microscopic methods have limited sensitivity, while RT-PCR has high sensitivity and can be implemented as a rapid routine diagnostic test.  $(1 \rightarrow 3)$ -beta-D-glucan in association with lactate dehydrogenase has been used as a biomarker in the case of HIV-infected patients (Esteves et al. 2014; Held et al. 2011). Up to now, attempts to culture *P. jirovecii* have been failures, except for recently, when a three-dimensional air–liquid interface culture system made up of differentiated pseudostratified airway epithelial cell line named as CuFi-8 cells has been demonstrated for culturing *P. jirovecii* (Schildgen et al. 2014).

### 9.2.5.2 Treatment

Typical antifungal drugs have been found to be not effective for Pneumocystis infection. Folic acid is needed for the synthesis of purines, glycine, and thymidylate, which are necessary for the proper functioning of an organism. The lifecycle on P. jirovecii also depends on folic acid synthesis but disruption of the folic acid synthesis pathway results in failed acquisition of the folic acid from the environment. Thus, the folic acid synthesis pathway has been used as a target for therapeutic agents for pneumocystis pneumonia treatment. Dihydrofolate reductase (DHFR) and dihydropteroate synthase (DHPS) are two important enzymes involved in the folic acid synthesis pathway (Volpe et al. 1993). Trimethoprim (TMP) and a sulfa drug named sulfamethoxazole (SMZ) are inhibitors of the DHFR and DHPS enzymes, respectively, and have been used as agents to treat pneumocystis pneumonia (Lobo et al. 2013). The most effective treatment for pneumocystis pneumonia is the combination of these two drugs, as has been shown in HIV-negative individuals (Thomas Jr. and Limper 2004). This drug can be taken either orally or by intravenous infusion. However, in HIV-positive individuals, sulfamethoxazole causes allergic reactions, so sulfamethoxazole should be replaced by dapsone. Apart from this drug, pentamidine, prednisone, a combination of clindamycin and primaquine, and a combination of trimetrexate and leucovorin are also used. The inhibition effect of TMP-SMZ can be further enhanced by the addition of low doses of caspofungin, which is an inhibitor of the  $(1 \rightarrow 3)$ -beta-D-glucan synthase, an enzyme required for the synthesis of cell walls of many fungi. Caspofungin and clindamycin have been reported to be a salvage therapy to treat immunocompromised patients having allergies or adverse reactions to TMP-SMZ (Li et al. 2016).

# 9.3 Conclusion and Future Implications

Fungi are opportunistic infectious agents and most of them are usually not pathogenic to normal healthy individuals. However, when they infect individuals at high risk of invasive fungal infection, they cause a wide range of diseases, ranging from superficial infections to disseminated infections of the vital internal organs. Invasive fungal infections continue to be a major problem due to improved medical care among individuals at high risk of invasive fungal infections. Despite the recent efforts, early diagnosis and intervention still need to be addressed due to their high associated mortality and morbidity rates. There have been significant advances in therapeutic options for invasive infections caused by Aspergillus and Candida species. Successful treatment of these opportunistic infections requires prompt diagnosis and aggressive therapy with antifungal agents. Inhalation of spores of these microorganisms is the most common mode of infection in a susceptible host (Deepa et al. 2014). To better understand the problem, we need to define accurate epidemiological data, the socioeconomic impact of the disease, and also stimulate scientific interest in this field. To tackle these challenges, we need to focus on: (i) development of better and rapid diagnostics tools which have an immediate impact on the mortality rate and (ii) development of more effective and less toxic antifungal drugs/ vaccines, especially for the treatment of immunocompromised patients. By the use of combination drug therapy, we can increase the effectiveness of the drug and minimize the risk of development of resistance and decrease toxicity (Elizabeth et al. 2015). During vaccination development against invasive fungal infections, efforts should be focused on identifying epitopes with immunogenic T cell and adjuvants. Attention directed towards these goals would have a significant effect in reducing the global burden and negative impact of invasive fungal infections.

# References

- Achkar JM, Fries BC (2010) *Candida* infections of the genitourinary tract. Clin Microbiol Rev 23(2):253–273
- Agarwal R, Chakrabarti A, Shah A, Gupta D, Meis JF, Guleria R, Moss R, Denning DW, Group AcaIw (2013) Allergic bronchopulmonary aspergillosis: review of literature and proposal of new diagnostic and classification criteria. Clin Exp Allergy 43(8):850–873
- Aliouat-Denis CM, Chabe M, Demanche C, Aliouat el M, Viscogliosi E, Guillot J, Delhaes L, Dei-Cas E (2008) *Pneumocystis* species, co-evolution and pathogenic power. Infect Genet Evol 8(5):708–726
- Amaral DM, Rocha RC, Carneiro LE, Vasconcelos DM, Abreu MA (2016) Disseminated cryptococcosis manifested as a single tumor in an immunocompetent patient, similar to the cutaneous primary forms. An Bras Dermatol 91(5 suppl 1):29–31
- Anagnostou T, Arvanitis M, Kourkoumpetis TK, Desalermos A, Carneiro HA, Mylonakis E (2014) Nocardiosis of the central nervous system: experience from a general hospital and review of 84 cases from the literature. Medicine (Baltimore) 93(1):19–32

- Anaissie EJ, Stratton SL, Dignani MC, Summerbell RC, Rex JH, Monson TP, Spencer T, Kasai M, Francesconi A, Walsh TJ (2002) Pathogenic *Aspergillus* species recovered from a hospital water system: a 3-year prospective study. Clin Infect Dis 34(6):780–789
- Badiee P, Hashemizadeh Z (2014) Opportunistic invasive fungal infections: diagnosis & clinical management. Indian J Med Res 139(2):195–204
- Bellmann R, Smuszkiewicz P (2017) Pharmacokinetics of antifungal drugs: practical implications for optimized treatment of patients. Infection 45(6):737–779
- Benedict K, Park BJ (2014) Invasive fungal infections after natural disasters. Emerg Infect Dis 20(3):349–355
- Bicanic T, Wood R, Meintjes G, Rebe K, Brouwer A, Loyse A, Bekker LG, Jaffar S, Harrison T (2008) High-dose amphotericin B with flucytosine for the treatment of cryptococcal meningitis in HIV-infected patients: a randomized trial. Clin Infect Dis 47(1):123–130
- Bisha B, Kim HJ, Brehm-Stecher BF (2011) Improved DNA-FISH for cytometric detection of *Candida* spp. J Appl Microbiol 110(4):881–892
- Bose I, Reese AJ, Ory JJ, Janbon G, Doering TL (2003) A yeast under cover: the capsule of *Cryptococcus neoformans*. Eukaryot Cell 2(4):655–663
- Boutboul F, Alberti C, Leblanc T, Sulahian A, Gluckman E, Derouin F, Ribaud P (2002) Invasive aspergillosis in allogeneic stem cell transplant recipients: increasing antigenemia is associated with progressive disease. Clin Infect Dis 34(7):939–943
- Brouwer AE, Rajanuwong A, Chierakul W, Griffin GE, Larsen RA, White NJ, Harrison TS (2004) Combination antifungal therapies for HIV-associated cryptococcal meningitis: a randomised trial. Lancet 363(9423):1764–1767
- Brown GD, Denning DW, Gow NA, Levitz SM, Netea MG, White TC (2012) Hidden killers: human fungal infections. Sci Transl Med 4(165):165rv113
- Caillot D, Casasnovas O, Bernard A, Couaillier JF, Durand C, Cuisenier B, Solary E, Piard F, Petrella T, Bonnin A, Couillault G, Dumas M, Guy H (1997) Improved management of invasive pulmonary aspergillosis in neutropenic patients using early thoracic computed tomographic scan and surgery. J Clin Oncol 15(1):139–147
- Caillot D, Mannone L, Cuisenier B, Couaillier JF (2001) Role of early diagnosis and aggressive surgery in the management of invasive pulmonary aspergillosis in neutropenic patients. Clin Microbiol Infect 7(Suppl 2):54–61
- Capewell S, Chapman BJ, Alexander F, Greening AP, Crompton GK (1989) Corticosteroid treatment and prognosis in pulmonary eosinophilia. Thorax 44(11):925–929
- Chotmongkol V, Jitpimolmard S (1992) Itraconazole in the treatment of cryptococcal meningitis. J Med Assoc Thail 75(2):85–88
- Cohen-Wolkowiez M, Benjamin DK Jr, Steinbach WJ, Smith PB (2006) Anidulafungin: a new echinocandin for the treatment of fungal infections. Drugs Today (Barc) 42(8):533–544
- Cornely OA, Maertens J, Bresnik M, Ebrahimi R, Ullmann AJ, Bouza E, Heussel CP, Lortholary O, Rieger C, Boehme A, Aoun M, Horst HA, Thiebaut A, Ruhnke M, Reichert D, Vianelli N, Krause SW, Olavarria E, Herbrecht R, AmBiLoad Trial Study G (2007) Liposomal amphotericin B as initial therapy for invasive mold infection: a randomized trial comparing a high-loading dose regimen with standard dosing (AmBiLoad trial). Clin Infect Dis 44(10):1289–1297
- Cutshaw D, Premji A, Pagadala P, Giamberardino C, McCabe A, Lad SP, Perfect JR (2016) A novel therapeutic approach for cryptococcal meningitis. Open Forum Infect Dis 3(suppl\_1):1643–1643
- Day JN, Chau TTH, Wolbers M, Mai PP, Dung NT, Mai NH, Phu NH, Nghia HD, Phong ND, Thai CQ, Thai LH, Chuong LV, Sinh DX, Duong VA, Hoang TN, Diep PT, Campbell JI, Sieu TPM, Baker SG, Chau NVV, Hien TT, Lalloo DG, Farrar JJ (2013) Combination antifungal therapy for cryptococcal meningitis. N Engl J Med 368(14):1291–1302
- de Gans J, Portegies P, Tiessens G, Eeftinck Schattenkerk JK, van Boxtel CJ, van Ketel RJ, Stam J (1992) Itraconazole compared with amphotericin B plus flucytosine in AIDS patients with cryptococcal meningitis. AIDS 6(2):185–190

- Deepa A, Nair BJ, Sivakumar T, Joseph AP (2014) Uncommon opportunistic fungal infections of oral cavity: a review. J Oral Maxillofac Pathol 18(2):235–243
- Dromer F, Bernede-Bauduin C, Guillemot D, Lortholary O, French Cryptococcosis Study G (2008) Major role for amphotericin B-flucytosine combination in severe cryptococcosis. PLoS One 3(8):e2870
- Elizabeth SB, Eric Y, Graeme NF (2015) Combination antifungal therapy: when, where, and why. Curr Clin Microbiol Rep 2(2):67–75
- Enoch DA, Ludlam HA, Brown NM (2006) Invasive fungal infections: a review of epidemiology and management options. J Med Microbiol 55(Pt 7):809–818
- Esteves F, Lee CH, de Sousa B, Badura R, Seringa M, Fernandes C, Gaspar JF, Antunes F, Matos O (2014) (1-3)-beta-D-glucan in association with lactate dehydrogenase as biomarkers of *Pneumocystis* pneumonia (PcP) in HIV-infected patients. Eur J Clin Microbiol Infect Dis 33(7):1173–1180
- Gago S, Overton NLD, Ben-Ghazzi N, Novak-Frazer L, Read ND, Denning DW, Bowyer P (2018) Lung colonization by Aspergillus fumigatus is controlled by ZNF77. Nat Commun 9(1):3835
- Gamaletsou MN, Sipsas NV, Kontoyiannis DP, Tsiakalos A, Kontos AN, Stefanou I, Kordossis T (2012) Successful salvage therapy of refractory HIV-related cryptococcal meningitis with the combination of liposomal amphotericin B, voriconazole, and recombinant interferon-gamma. Diagn Microbiol Infect Dis 74(4):409–411
- Ganguly S, Mitchell AP (2011) Mucosal biofilms of *Candida albicans*. Curr Opin Microbiol 14(4):380–385
- Garnacho-Montero J, Amaya-Villar R, Ortiz-Leyba C, Leon C, Alvarez-Lerma F, Nolla-Salas J, Iruretagoyena JR, Barcenilla F (2005) Isolation of *Aspergillus* spp. from the respiratory tract in critically ill patients: risk factors, clinical presentation and outcome. Crit Care 9(3):R191–R199
- Ghannoum MA, Kuhn DM (2002) Voriconazole-better chances for patients with invasive mycoses. Eur J Med Res 7(5):242–256
- Guarner J, Brandt ME (2011) Histopathologic diagnosis of fungal infections in the 21st century. Clin Microbiol Rev 24(2):247–280
- Hammond SP, Bialek R, Milner DA, Petschnigg EM, Baden LR, Marty FM (2011) Molecular methods to improve diagnosis and identification of mucormycosis. J Clin Microbiol 49(6):2151–2153
- Hartwick RW, Batsakis JG (1991) Sinus aspergillosis and allergic fungal sinusitis. Ann Otol Rhinol Laryngol 100(5 Pt 1):427–430
- Held J, Koch MS, Reischl U, Danner T, Serr A (2011) Serum (1 -> 3)-beta-D-glucan measurement as an early indicator of *Pneumocystis jirovecii* pneumonia and evaluation of its prognostic value. Clin Microbiol Infect 17(4):595–602
- Herbrecht R, Denning DW, Patterson TF, Bennett JE, Greene RE, Oestmann JW, Kern WV, Marr KA, Ribaud P, Lortholary O, Sylvester R, Rubin RH, Wingard JR, Stark P, Durand C, Caillot D, Thiel E, Chandrasekar PH, Hodges MR, Schlamm HT, Troke PF, de Pauw B, Invasive Fungal Infections Group of the European Organisation for R, Treatment of C, the Global Aspergillus Study G (2002) Voriconazole versus amphotericin B for primary therapy of invasive aspergillosis. N Engl J Med 347(6):408–415
- Hope W, Natarajan P, Goodwin L (2013) Invasive fungal infections. Clin Med (Lond) 13(5):507-510
- Horn F, Heinekamp T, Kniemeyer O, Pollmacher J, Valiante V, Brakhage AA (2012) Systems biology of fungal infection. Front Microbiol 3:108
- Ibrahim AS, Spellberg B, Walsh TJ, Kontoyiannis DP (2012) Pathogenesis of mucormycosis. Clin Infect Dis 54(Suppl 1):S16–S22
- Johnson LB, Kauffman CA (2003) Voriconazole: a new triazole antifungal agent. Clin Infect Dis 36(5):630–637
- Koulenti D, Garnacho-Montero J, Blot S (2014) Approach to invasive pulmonary aspergillosis in critically ill patients. Curr Opin Infect Dis 27(2):174–183

- Lain A, Elguezabal N, Brena S, Garcia-Ruiz JC, Del Palacio A, Moragues MD, Ponton J (2007) Diagnosis of invasive candidiasis by enzyme-linked immunosorbent assay using the N-terminal fragment of *Candida albicans* hyphal wall protein 1. BMC Microbiol 7:35
- Larsen RA, Leal MA, Chan LS (1990) Fluconazole compared with amphotericin B plus flucytosine for cryptococcal meningitis in AIDS. A randomized trial. Ann Intern Med 113(3):183–187 Latge JP (1999) Aspergillus fumigatus and aspergillosis. Clin Microbiol Rev 12(2):310–350
- Li H, Huang H, He H (2016) Successful treatment of severe *Pneumocystis* pneumonia in an immunosuppressed patient using caspofungin combined with clindamycin: a case report and literature review. BMC Pulm Med 16(1):144
- Limper AH, Knox KS, Sarosi GA, Ampel NM, Bennett JE, Catanzaro A, Davies SF, Dismukes WE, Hage CA, Marr KA, Mody CH, Perfect JR, Stevens DA, American Thoracic Society Fungal Working G (2011) An official American Thoracic Society statement: treatment of fungal infections in adult pulmonary and critical care patients. Am J Respir Crit Care Med 183(1):96–128
- Lobo ML, Esteves F, de Sousa B, Cardoso F, Cushion MT, Antunes F, Matos O (2013) Therapeutic potential of caspofungin combined with trimethoprim-sulfamethoxazole for *Pneumocystis* pneumonia: a pilot study in mice. PLoS One 8(8):e70619
- Marr KA, Balajee SA, McLaughlin L, Tabouret M, Bentsen C, Walsh TJ (2004a) Detection of galactomannan antigenemia by enzyme immunoassay for the diagnosis of invasive aspergillosis: variables that affect performance. J Infect Dis 190(3):641–649
- Marr KA, Boeckh M, Carter RA, Kim HW, Corey L (2004b) Combination antifungal therapy for invasive aspergillosis. Clin Infect Dis 39(6):797–802
- Maskarinec SA, Johnson MD, Perfect JR (2016) Genetic susceptibility to fungal infections: what is in the genes? Curr Clin Microbiol Rep 3(2):81–91
- Massard G, Roeslin N, Wihlm JM, Dumont P, Witz JP, Morand G (1992) Pleuropulmonary aspergilloma: clinical spectrum and results of surgical treatment. Ann Thorac Surg 54(6):1159–1164
- Mirhendi SH, Makimura K (2003) PCR- detection of *Candida albicans* in blood using a new primer pair to diagnosis of systemic candidiasis. Iran J Public Health 32(1):1–5
- Mohammadi H, Sadeghi S, Zandi S (2015) Central nervous system aspergilloma in an immunocompetent patient: acase report. Iran J Public Health 44(6):869–872
- Morrell M, Fraser VJ, Kollef MH (2005) Delaying the empiric treatment of *Candida* bloodstream infection until positive blood culture results are obtained: a potential risk factor for hospital mortality. Antimicrob Agents Chemother 49(9):3640–3645
- Nobile CJ, Johnson AD (2015) *Candida albicans*biofilms and human disease. Annu Rev Microbiol 69:71–92
- Oladele RO, Bongomin F, Gago S, Denning DW (2017) HIV-associated cryptococcal disease in resource-limited settings: acase for "prevention is better than cure"? J Fungi (Basel) 3(4):67
- Pappas PG, Rotstein CM, Betts RF, Nucci M, Talwar D, De Waele JJ, Vazquez JA, Dupont BF, Horn DL, Ostrosky-Zeichner L, Reboli AC, Suh B, Digumarti R, Wu C, Kovanda LL, Arnold LJ, Buell DN (2007) Micafungin versus caspofungin for treatment of candidemia and other forms of invasive candidiasis. Clin Infect Dis 45(7):883–893
- Pappas PG, Kauffman CA, Andes D, Benjamin DK Jr, Calandra TF, Edwards JE Jr, Filler SG, Fisher JF, Kullberg BJ, Ostrosky-Zeichner L, Reboli AC, Rex JH, Walsh TJ, Sobel JD, Infectious Diseases Society of A (2009) Clinical practice guidelines for the management of candidiasis: 2009 update by the Infectious Diseases Society of America. Clin Infect Dis 48(5):503–535
- Perfect JR, Bicanic T (2015) Cryptococcosis diagnosis and treatment: What do we know now. Fungal Genet Biol 78:49–54
- Perfect JR, Dismukes WE, Dromer F, Goldman DL, Graybill JR, Hamill RJ, Harrison TS, Larsen RA, Lortholary O, Nguyen MH, Pappas PG, Powderly WG, Singh N, Sobel JD, Sorrell TC (2010) Clinical practice guidelines for the management of cryptococcal disease: 2010 update by the infectious diseases society of america. Clin Infect Dis 50(3):291–322
- Person AK, Kontoyiannis DP, Alexander BD (2010) Fungal infections in transplant and oncology patients. Infect Dis Clin N Am 24(2):439–459

- Peters BM, Jabra-Rizk MA, Scheper MA, Leid JG, Costerton JW, Shirtliff ME (2010) Microbial interactions and differential protein expression in *Staphylococcus aureus–Candida albicans* dual-species biofilms. FEMS Immunol Med Microbiol 59(3):493–503
- Pfaller MA, Diekema DJ (2007) Epidemiology of invasive candidiasis: a persistent public health problem. Clin Microbiol Rev 20(1):133–163
- Pitisuttithum P, Negroni R, Graybill JR, Bustamante B, Pappas P, Chapman S, Hare RS, Hardalo CJ (2005) Activity of posaconazole in the treatment of central nervous system fungal infections. J Antimicrob Chemother 56(4):745–755
- Prabhu RM, Patel R (2004) Mucormycosis and entomophthoramycosis: a review of the clinical manifestations, diagnosis and treatment. Clin Microbiol Infect 10(Suppl 1):31–47
- Prieto D, Pla J (2015) Distinct stages during colonization of the mouse gastrointestinal tract by *Candida albicans*. Front Microbiol 6:792
- Ramana KV, Sabitha K, Venkata BP, Sharada CV, Ratna R, Ratna M, Sanjeev DR (2013) Invasive fungal infections: a comprehensive review. Am J Infect Dis Microbiol 1(4):64–69
- Ravikumar S, Win MS, Chai LY (2015) Optimizing outcomes in immunocompromised hosts: understanding the role of immunotherapy in invasive fungal diseases. Front Microbiol 6:1322
- Rosenberg M, Patterson R, Roberts M, Wang J (1978) The assessment of immunologic and clinical changes occurring during corticosteroid therapy for allergic bronchopulmonary aspergillosis. Am J Med 64(4):599–606
- Ruping MJ, Vehreschild JJ, Cornely OA (2008) Patients at high risk of invasive fungal infections: when and how to treat. Drugs 68(14):1941–1962
- Sambatakou H, Dupont B, Lode H, Denning DW (2006) Voriconazole treatment for subacute invasive and chronic pulmonary aspergillosis. Am J Med 119(6):527 e517–527 e524
- Schildgen V, Mai S, Khalfaoui S, Lusebrink J, Pieper M, Tillmann RL, Brockmann M, Schildgen O (2014) *Pneumocystis jirovecii* can be productively cultured in differentiated CuFi-8 airway cells. MBio 5(3):e01186–e01114
- Schmiedel Y, Zimmerli S (2016) Common invasive fungal diseases: an overview of invasive candidiasis, aspergillosis, cryptococcosis, and Pneumocystis pneumonia. Swiss Med Wkly 146:w14281
- Sethi P, Saluja R, Jindal N, Singh V (2012) Invasive aspergillosis in an immunocompetent host. J Oral Maxillofac Pathol 16(2):297–300
- Shahi M, Ayatollahi Mousavi SA, Nabili M, Aliyali M, Khodavaisy S, Badali H (2015) *Aspergillus* colonization in patients with chronic obstructive pulmonary disease. Curr Med Mycol 1(3):45–51
- Siddiqui AA, Brouwer AE, Wuthiekanun V, Jaffar S, Shattock R, Irving D, Sheldon J, Chierakul W, Peacock S, Day N, White NJ, Harrison TS (2005) IFN-gamma at the site of infection determines rate of clearance of infection in cryptococcal meningitis. J Immunol 174(3):1746–1750
- Skiada A, Lanternier F, Groll AH, Pagano L, Zimmerli S, Herbrecht R, Lortholary O, Petrikkos GL, European Conference on Infections in L (2013) Diagnosis and treatment of mucormycosis in patients with hematological malignancies: guidelines from the 3rd European Conference on Infections in Leukemia (ECIL 3). Haematologica 98(4):492–504
- Sloan DJ, Parris V (2014) Cryptococcal meningitis: epidemiology and therapeutic options. Clin Epidemiol 6:169–182
- Sokulska M, Kicia M, Wesolowska M, Hendrich AB (2015) Pneumocystis jirovecii–from a commensal to pathogen: clinical and diagnostic review. Parasitol Res 114(10):3577–3585
- Soltanzadeh H, Wychulis AR, Sadr F, Bolanowski PJ, Neville WE (1977) Surgical treatment of pulmonary aspergilloma. Ann Surg 186(1):13–16
- Sowden E, Carmichael AJ (2004) Autoimmune inflammatory disorders, systemic corticosteroids and pneumocystis pneumonia: a strategy for prevention. BMC Infect Dis 4:42
- Spanakis EK, Aperis G, Mylonakis E (2006) New agents for the treatment of fungal infections: clinical efficacy and gaps in coverage. Clin Infect Dis 43(8):1060–1068
- Spellberg B, Ibrahim AS (2010) Recent advances in the treatment of mucormycosis. Curr Infect Dis Rep 12(6):423–429

- Spellberg B, Edwards J Jr, Ibrahim A (2005) Novel perspectives on mucormycosis: pathophysiology, presentation, and management. Clin Microbiol Rev 18(3):556–569
- Spitzer M, Robbins N, Wright GD (2017) Combinatorial strategies for combating invasive fungal infections. Virulence 8(2):169–185
- Stevens DA, Kan VL, Judson MA, Morrison VA, Dummer S, Denning DW, Bennett JE, Walsh TJ, Patterson TF, Pankey GA (2000) Practice guidelines for diseases caused by *Aspergillus*. Infectious Diseases Society of America. Clin Infect Dis 30(4):696–709
- Tamgadge AP, Mengi R, Tamgadge S, Bhalerao SS (2012) Chronic invasive aspergillosis of paranasal sinuses: a case report with review of literature. J Oral Maxillofac Pathol 16(3):460–464
- Tang CM, Cohen J (1992) Diagnosing fungal infections in immunocompromised hosts. J Clin Pathol 45(1):1–5
- Tedder M, Spratt JA, Anstadt MP, Hegde SS, Tedder SD, Lowe JE (1994) Pulmonary mucormycosis: results of medical and surgical therapy. Ann Thorac Surg 57(4):1044–1050
- Thomas CF Jr, Limper AH (2004) Pneumocystis pneumonia. N Engl J Med 350(24):2487-2498
- Thompson GR 3rd, Rendon A, Ribeiro Dos Santos R, Queiroz-Telles F, Ostrosky-Zeichner L, Azie N, Maher R, Lee M, Kovanda L, Engelhardt M, Vazquez JA, Cornely OA, Perfect JR (2016) Isavuconazole treatment of cryptococcosis and dimorphic mycoses. Clin Infect Dis 63(3):356–362
- Uflacker R, Kaemmerer A, Picon PD, Rizzon CF, Neves CM, Oliveira ES, Oliveira ME, Azevedo SN, Ossanai R (1985) Bronchial artery embolization in the management of hemoptysis: technical aspects and long-term results. Radiology 157(3):637–644
- Volpe F, Ballantine SP, Delves CJ (1993) The multifunctional folic acid synthesis fas gene of *Pneumocystis carinii* encodes dihydroneopterin aldolase, hydroxymethyldihydropterin pyrophosphokinase and dihydropteroate synthase. Eur J Biochem 216(2):449–458
- Walsh TJ, Anaissie EJ, Denning DW, Herbrecht R, Kontoyiannis DP, Marr KA, Morrison VA, Segal BH, Steinbach WJ, Stevens DA, van Burik JA, Wingard JR, Patterson TF, Infectious Diseases Society of A (2008) Treatment of aspergillosis: clinical practice guidelines of the Infectious Diseases Society of America. Clin Infect Dis 46(3):327–360
- Wang JL, Patterson R, Roberts M, Ghory AC (1979) The management of allergic bronchopulmonary aspergillosis. Am Rev Respir Dis 120(1):87–92
- Yanagisawa E, Friedman S, Kundargi RS, Smith HW (1977) Rhinocerebral phycomycosis. Laryngoscope 87(8):1319–1335
- Yao ZW, Lu X, Shen C, Lin DF (2014) Comparison of flucytosine and fluconazole combined with amphotericin B for the treatment of HIV-associated cryptococcal meningitis: a systematic review and meta-analysis. Eur J Clin Microbiol Infect Dis 33(8):1339–1344
- Yao Y, Zhang JT, Yan B, Gao T, Xing XW, Tian CL, Huang XS, Yu SY (2015) Voriconazole: a novel treatment option for cryptococcal meningitis. Infect Dis (Lond) 47(10):694–700
- Yu VL, Muder RR, Poorsattar A (1986) Significance of isolation of Aspergillus from the respiratory tract in diagnosis of invasive pulmonary aspergillosis. Results from a three-year prospective study. Am J Med 81(2):249–254
- Zago CE, Silva S, Sanita PV, Barbugli PA, Dias CM, Lordello VB, Vergani CE (2015) Dynamics of biofilm formation and the interaction between *Candida albicans* and methicillin-susceptible (MSSA) and -resistant *Staphylococcus aureus* (MRSA). PLoS One 10(4):e0123206
- Zaragoza O, Casadevall A (2004) Experimental modulation of capsule size in Cryptococcus neoformans. Biol Proced Online 6:10–15
- Zaragoza O, Rodrigues ML, De Jesus M, Frases S, Dadachova E, Casadevall A (2009) The capsule of the fungal pathogen *Cryptococcus neoformans*. Adv Appl Microbiol 68:133–216