

Tinospora cordifolia (Willd.) Miers ex Hook. F. & Thoms (Menispermaceae)

(**Syns.**: *T. sinensis* (Lour.) Merr.; *T. cordifolia* (D.C.) Miers, *T. cordifolia* (Willd.) Miers.; *Cocculus cordifolius* (Willd.) DC.; *Menispermum cordifolium* Willd.)

Abstract

A large, perennial, deciduous, climbing herbaceous vine, that is distributed throughout India, Myanmar, Sri Lanka and China. In Ayurveda, it is mentioned as rasayan and is traditionally used for the treatment of asthma, chronic cough, to improve immune system, as a general tonic, antiperiodic in fevers, antispasmodic, anti-inflammatory, antiarthritic and antidiabetic agent, and is also credited with aphrodisiac property. Fresh plant is said to be more efficient than the dried one. It is taken with milk in rheumatism, acidity of the urine and dyspepsia. The stem of this very bitter herbaceous vine is used medicinally in Unani medicine as a bitter tonic, astringent, stomachic, anthelmintic, blood purifier, diuretic, and antipyretic for all types of fevers, including tuberculous fever. Water extracted from fresh plant is more potent. It is also used for chronic diarrhea, and in diseases, such as syphilis, and leprosy. In the Philippines and Malaysia, this is the most popular medicinal plant, and is considered a universal medicine. Its aqueous extract is used as a remedy for stomach trouble, indigestion and diarrhea. A preparation with coconut oil is considered an effective cure for rheumatism and for flatulence in children. Various constituents, such as alkaloids, diterpenoid lactones, cardiac glycosides, steroids, sesquiterpenoid, phenolics, aliphatic compounds and polysaccharides have been reported from the plant. The yield and physicochemical profile of the starchy material extracted from stem used in Ayurvedic preparations vary due to the plant stem size, collection time, season and maturity of the plant. Total alkaloidal contents are a bit higher in rainy and spring seasons. Aqueous, alcohol and chloroform extracts exerted significant hypoglycemic and antihyperglycemic effects in normal and diabetic animals. Aqueous extract significantly stimulates glucose uptake in 3T3-L1 adipocytes, comparable to insulin and greater than pioglitazone. Aqueous extract also prevented hyperalgesia of diabetic neuropathy, and inhibited aldose reductase. Ethanol extract of aerial parts offered significant neuroprotection against 6-OHDA-induced Parkinson's disease-like lesions in rat model, and decreased locomotor activity but did not affect amphetamine-induced hyperactivity in mice.

Keywords

Gilo • Giloe • Guduchi • Guduchi-kräutertee • Gulancha tinospora • Gulbel • Guricha • Jivantika • Makabúhaí • Xin ye qing niu dan

Vernaculars: Urd.: Giloe; Hin.: Giloe, Guduchi, Gulach, Gulancha, Guruch; San.: Amurta, Bhishakpriya, Chinnaruha, Giloy, Guduchi, Jivantika, Nirjara, Pittaghni, Soma-valli; Ben.: Gadancha, Giloe, Gulach, Gulancha, Palo (extract); Guj.: Galo; Mal.: Amrita, Amruthu, Chitramruta; Mar.: Gharol, Guduchi, Gulavel, Guloe; Tam.: Amirthavalli, Kunali, Seenthil kodi, Shindilakodi, Shindil-shakkarai (extract); Tel.: Guluchi, Guricha, Manapala, Tippa-teega, Tippa-tige-satu (extract), Tippategeveru (root); Ara.: Gilo; Chi.: 心叶青牛胆, Xin ye qing niu dan; Eng.: Gulancha tinospora, Heartleaf moonseed; Fre.: Guduchi, Tinofolin; Ger.: Guduchi-kräutertee; Nep.: Gurjo; Per.: Gulbel; Tag.: Makabúhaí; Tha.: Ching cha chali.

Description: It is a large, glabrous, perennial, deciduous, climbing herbaceous vine of weak and fleshy stem spreading on trees of *Mangifera indica* and *Azadirachta indica*. It is distributed throughout India, Myanmar, Sri Lanka and China. Fresh stem has a green succulent bark, covered by a thin brown epidermis, which peels off in flakes. It is studded with warty prominences, and gives off roots here and there, and branches bearing smooth heart-shaped leaves, and bunches of red berries. When dry it shrinks very much, and the bark separates from the wood, and becomes of a dull-brown color; the latter consists of a number of wedge-shaped bundles; the taste is very bitter, the odor is not in any way peculiar.^{XL} Flowers are typically greenish-yellow, and the flowering season extends from summer to winter; male flowers are clustered, while the female flowers are solitary [99]. According to Narkhede et al. [51], *T. sinensis* closely resembles the description of *guduchi* in *Ayurvedic* literature rather than the commonly available *T. cordifolia*, but may be used as a substitute for *T. sinensis; T. cordifolia* growing on *Azadirachta indica* is called *Neem-guduchi* and has better immunomodulatory potential (Figs. 1 and 2).

Actions and Uses: In *Ayurveda*, it (temperament, cold and dry) is mentioned as *rasayan* and is traditionally used for the treatment of asthma, chronic cough, to improve immune system, as a general tonic, antiperiodic in fevers, antispasmodic, anti-inflammatory, antiarthritic and antidiabetic agent, and is also credited with aphrodisiac property [73, 98]. Fresh plant is said to be more efficient than the dried one. It is taken with milk in rheumatism, acidity of the urine and dyspepsia. It is said that if the stem is placed upon a bush in the open air, will retain its vitality through the hot season, and when the rains start, put forth leaves and long whipcord-like roots, which soon reach the ground, hence the Sanskrit synonym *Chinnaruha*, or growing when cut.^{XL} The stem (temperament, hot 1° and dry 1°) of this very bitter herbaceous vine is used medicinally in *Unani* medicine as a bitter tonic, astringent, stomachic, anthelmintic, blood purifier, diuretic, and antipyretic for all types of fevers, including tuberculous fever. Water extracted from fresh plant is more potent. It is also used for chronic diarrhea, and in diseases, such as syphilis, and leprosy.^{LXXVII} It is also



Fig. 1 *Tinospora cordifolia*, Plant, Tmd, WikimediaCommons, https://commons.wikimedia.org/ wiki/File:Tinospora_cordifolia.jpg



Fig. 2 *Tinospora cordifolia*, Fruits, Vinayaraj, WikimediaCommons; ShareAlike 4.0 International CC BY-SA 4.0, https://commons.wikimedia.org/wiki/File:Tinospora_cordifolia_fruits_03. JPG; https://creativecommons.org/licenses/by-sa/4.0/deed.en

rheumatism, skin diseases (such as impetigo), jaundice, debility caused by repeated attacks of fever, ^{LXXXI,CV} urinary disorders, diabetes and anemia [8]. Water extract is used as a febrifuge and is referred to as 'Indian quinine.'^{CV} Whole plant pounded with water is used for spermatorrhea and gonorrhea.^{CXVII} In Ramgiri, Koraput district of Orissa (India), tribals orally administer 8 g starch obtained from stem, mixed in water

with equal quantity of sugar, daily for seven days to treat jaundice [24]. The Boxa tribe of Nainital district (India), use stem decoction bath in postdelivery fever [84]. It is also considered antihepatotoxic, antistress, immunomodulatory, and antioxidant [52]. In the Philippines and Malaysia, this is the most popular medicinal plant, and is considered a universal medicine. The aqueous extract is used as a remedy for stomach trouble, indigestion and diarrhea. A preparation with coconut oil is considered an effective cure for rheumatism and for flatulence in children.^{CXVII}

Phytoconstituents: Various constituents, such as alkaloids, diterpenoid lactones, cardiac glycosides, steroids, sesquiterpenoid, phenolics, aliphatic compounds and polysaccharides have been reported from the plant [47, 99]. The yield and physicochemical profile of the starchy material extracted from stem used in Ayurvedic preparations vary due to the plant stem size, collection time, season and maturity of the plant. Total alkaloidal contents are a bit higher in rainy and spring seasons [79]. Ethanol leaf extract showed the presence of steroids, anthraquinones, flavonoids, cardiac glycosides, tannins and phenolics [94]. Seven compounds, 11-hydroxymustakone, N-methyl-2-pyrrolidone, N-formylannonain, magnoflorine, cordifolioside A, tinocordiside, and syringin, with immunomodulatory activity were isolated, and the activity is assumed to be due to their synergistic effect [80]. Bala et al. [8] also isolated jatrorrhizine, palmatine, and yangambin from stem. The isoquinoline alkaloids, jatrorrhizine, palmatine and magnoflorine demonstrated significant inhibitory activity against aldose reductase isolated from male rats [56]. A novel sulfur-containing clerodane diterpene glycoside, cordifolide A, and two diterpene glycosides, cordifolides B and C were also isolated from the stem [53]. Ahmad et al. [4] isolated tinosporafuranol, tinosporafurandiol, tinosporaclerodanol, and tinosporaclerodanoid, along with β -sitosterol from stem bark. Two aporphine alkaloids (Tinoscorside A and B), a clerodane diterpene, tinoscorside C and a phenylpropanoid, tinoscorside D were isolated from methanol extract of aerial parts [100], while four clerodane furanoditerpene glucosides (Amritosides A, B, C and D) [46], three norditerpene furanglycosides, cordifolisides A, B and C [16], and two diterpinoid furonolactones, tinosporide [92] and columbin [93] were isolated from the stem. An immunologically active arabinogalactan with polyclonal mitogenic activity against B-cells was also reported from the stem [13]. Several other immunomodulating compounds have been reported from the plant. Syringin and cordiol inhibit in vitro immunohaemolysis of antibody-coated sheep erythrocytes by guinea pig serum; while cordioside, cordiofolioside A and cordiol activate macrophages [36]. A polysaccharide from stem is composed of glucose (98%), xylose (0.8%), arabinose (0.5%), galactose (0.3%), rhamnose (0.2%) and mannose (0.2%) [30]. Jatrorrhizine is also reported from the root of the plant [74].

Pharmacology: Aqueous, alcohol and chloroform extracts exerted significant hypoglycemic and antihyperglycemic effects in normal and diabetic animals [19, 21, 37, 39, 48, 58, 103]. Aqueous extract also prevented hyperalgesia of diabetic neuropathy, and *in vitro* inhibited aldose reductase [49]. Overexpression of angiogenic and inflammatory mediators, markers of diabetic retinopathy, was inhibited, retinal oxidative stress reduced and antioxidant enzyme levels of diabetic rats was

restored [3]. Treatment of diabetic animals also prevents polyuria and reduces urinary albumin [20], rise in insulin, TGs and glucose-insulin index, improves antioxidant status [66, 67, 71], inhibits α -glucosidase [14], and significantly prevents cataract formation [3, 65]. Oral administration of an α -glucosidase inhibitor constituent, saponarin, to maltose-fed rats produced hypoglycemic activity in doses of 20-80 mg/kg, comparable to 100–200 mg/kg of acarbose [76]. The isoquinoline alkaloid rich fraction of the stem and three alkaloids viz., palmatine, jatrorrhizine and magnoflorine significantly decreased FBG, and increased serum insulin level in glucosefed rats [55]. Aqueous and ethanol root extracts also significantly reduced serum and tissue cholesterol, phospholipids, FFAs, and glucose of diabetic rats [60, 61, 90], and ethanol extract also improved antioxidant status [59, 62]. The plant is an immunostimulator [5, 40, 50, 81, 89, 91, 101]; the aqueous extract improved cellular immunity and significantly reduced rats' mortality following cholestasis and E. coli infection [69]; and the ethanol extract improved phagocytic function without affecting humoral or cell-mediated immune system [6], protected against CP-induced myelosuppression and leucopenia [44, 95], and against gamma radiation exposure [18, 52, 85]. Activation of macrophages by the extract [75] leads to increase in GM-CSF, resulting in leucocytosis and improved neutrophil function [96]. Various extracts exhibit analgesic and anti-inflammatory activities [17, 25, 57]. In a mouse model of asthma, hydroalcohol extract protected against oxidative stress, proinflammatory cytokines release and redox signaling, and reduced airway hyperresponsiveness [98].

Ethanol extract of aerial parts offered significant neuroprotection against 6-OHDA-induced Parkinson's disease-like lesions in rat model [41], and decreased locomotor activity but did not affect amphetamine-induced hyperactivity in mice [31]. Methanol stem extract significantly inhibited *in vitro* AChE [102], and petroleum ether extract at a relatively low dose produced significant antidepressant-like effect in mice, comparable to impramine and sertraline, without significantly affecting locomotor functions and reducing activities of MAOs of whole brain [15]. Pretreatment with ethanol extract of whole plant reduced the infarct size and lipid peroxide levels of serum and heart tissue in surgically-induced myocardial I/R injury in rats [63], and normalized calcium chloride-induced cardiac arrhythmia in rats, comparable to verapamil [77]. Aqueous extracts of stem and leaves also reversed hematological changes in lead-treated mice [82]. Pretreatment with stem and leaves extracts protects from lead nitrate-hepatotoxicity, increased activities of antioxidant enzymes [83], CCl₄-liver damage [9], and whole plant powder protected against antitubercular drugs-hepatotoxicity [1, 54]. Ethanol extract of stems and leaves also showed antioxidant activity and decreased LPO in NDEA-induced liver cancer in rats [32], in diabetic rats [88], and CP-induced toxicity in mice [45].

Exposure of HeLa cells to methanol, aqueous, methylene chloride and dichloromethane extracts caused significant dose-dependent increase in cell killing [27, 28]. Dichloromethane extract increased tumor-free survival of mice transplanted with Ehrlich ascites carcinoma, with optimum effect when the extract was administered within five days of tumor inoculation [29, 64]. Hydroethanol extract also increased survival time and decreased peritoneal ascitic fluid content of Dalton's lymphoma ascites in Swiss mice [2], due to augmentation of function of macrophages [86]. Significant reduction by the extract in cumulative number, tumor yield, tumor burden, and tumor weight, along with significant elevation of phase II detoxifying enzymes, and inhibition of LPO was reported in skin carcinogenesis model [12]. The extract also inhibits melanoma cell-induced capillary formation in animals [42]; octacosanol has been identified as the antiangiogenic compound [97]. A polysaccharide fraction produced 72% inhibition in metastases formation of melanoma cells in the lungs of syngeneic C57BL/6 mice [43].

Sequential petroleum ether, chloroform, ethyl acetate, acetone, and ethanol extracts exhibited activity against Pseudomonas spp., while acetone, ethanol and aqueous extracts were active against *K. pneumonia*; Proteus spp. were inhibited by petroleum ether and benzene extracts, and *E. coli* was susceptible to ethyl acetate and acetone extracts [47]. Ethanol extract was inhibitory against *E. coli*, *P. vulgaris*, *E. faecalis*, *S. typhi*, *S. aureus* and *S. marcesenses* [33], and clinical isolates of MRSA and carbapenemase-producing *K. pneumoniae* [10]. Oral administration of methanol extract of stem to male rats for 60-days significantly decreased weight of testes, epididymis, seminal vesicle and ventral prostate, significantly reduced sperm motility and density, and serum testosterone levels, resulting in complete infertility [23]. A standardized aqueous extract reversed effects of cisplatin on gastric emptying, normalized intestinal hypermotility and the phagocytic function irrespective to the direction of change, complying to the definition of an adaptogen [70]. Ethanol and aqueous extracts of stem-bark produced dose-dependent antidiarrheal effect, and gastric antiulcer activity in rats [38].

Clinical Studies: In thirty Indian patients with malignant obstructive jaundice, addition of aqueous extract to conventional treatment with vitamin K, antibiotics and biliary drainage in half of the patients normalized the neutrophils phagocytic activity, completely resolved clinical signs of septicemia, and improved postoperative survival to near complete, compared to the control group with 40% survival rate [68]. Supplementation with aqueous extract to chronic asymptomatic moderate alcohol drinker with no chronic liver disease was still significantly protective against alcohol-induced damage [78]. Addition of aqueous extract as adjunct to chloroquine in partially/slow responding three Indian patients with malarial splenomegaly significantly regressed spleen size by two-third after six-months of treatment [87]. Sixty percent HIV positive Indian participants treated with a standardized aqueous extract for six-months reported relief from various symptoms compared to 20% in the placebo group of a double-blind RCT [35]. In a double-blinded RCT of patients with allergic rhinitis, eight-weeks treatment with the extract was effective in completely relieving sneezing in 83% and in more than two-thirds from nasal discharge and nasal obstruction, compared to those treated with placebo, who showed no relief in more than 80% patients [7, 22]. Topical application of a T. cordifolia lotion was comparably effective with permethrin in scabies-infected pediatric patients [11].

Mechanism of Action: Aqueous extract significantly stimulates glucose uptake in 3T3-L1 adipocytes, comparable to insulin and greater than pioglitazone [34]. Dichloromethane extract of stem *in vitro* inhibited 100% of α -glucosidase, 75% of

salivary amylase and 83% pancreatic amylase [14]. Anti-inflammatory effect in rat adjuvant-induced arthritis is mediated via reduction of proinflammatory cytokines [72].

Human A/Es, Allergy and Toxicity: Commonly reported adverse effects of a standardized aqueous extract in HIV positive patients were anorexia, nausea, vomiting and weakness [35].

Animal Toxicity: Oral LD50 of ethanol extract in mice is reported to be 2,650 mg (range 2,209–3,091 mg/kg). Oral doses of hexane- and chloroform-soluble extracts of the stem produced no significant toxic or adverse effects in rabbits up to the highest dose of 1,600 mg/kg [26].

Commentary: Significant protective and therapeutic effects of the aqueous extract on liver, spleen and HIV have been documented in RCTs, that should be further investigated in larger clinical trials and diverse patient populations to firmly validate its therapeutic efficiency. Other significant effects observed in animal studies also need further exploration in systematic clinical trials.

References

- Adhvaryu MR, Reddy N, Parabia MH. Effects of four Indian medicinal herbs on isoniazid-, rifampicin- and pyrazinamide-induced hepatic injury and immunosuppression in guinea pigs. World J Gastroenterol. 2007;13: 3199–205.
- Adhvaryu MR, Reddy N, Parabia MH. Antitumor activity of four Ayurvedic herbs in Dalton lymphoma ascites bearing mice and their short-term *in vitro* cytotoxicity on DLA-cell-line. Afr J Tradit Complement Altern Med. 2008; 5:409–18.
- Agrawal SS, Naqvi S, Gupta SK, Srivastava S. Prevention and management of diabetic retinopathy in STZ diabetic rats by *Tinospora cordifolia* and its molecular mechanisms. Food Chem Toxicol. 2012;50:3126–32.
- Ahmad F, Ali M, Alam P. New phytoconstituents from the stem bark of *Tinospora cordifolia* Miers. Nat Prod Res. 2010;24:926–34.
- Aranha I, Clement F, Venkatesh YP. Immunostimulatory properties of the major protein from the stem of the Ayurvedic medicinal herb, guduchi (*Tinospora cordifolia*). J Ethnopharmacol. 2012;139:366–72.
- Atal CK, Sharma ML, Kaul A, Khajuria A. Immunomodulating agents of plant origin. I: preliminary screening. J Ethnopharmacol. 1986;18:133–41.
- Badar VA, Thawani VR, Wakode PT, et al. Efficacy of *Tinospora cordifolia* in allergic rhinitis. J Ethnopharmacol. 2005;96:445–9.
- Bala M, Pratap K, Verma PK, Singh B, Padwad Y. Validation of ethnomedicinal potential of *Tinospora cordifolia* for anticancer and immunomodulatory activities and quantification of bioactive molecules by HPTLC. J Ethnopharmacol. 2015;175:131–7.

- Bishayi B, Roychowdhury S, Ghosh S, Sengupta M. Hepatoprotective and immunomodulatory properties of *Tinospora cordifolia* in CCl4 intoxicated mature albino rats. J Toxicol Sci. 2002;27:139–46.
- Bonvicini F, Mandrone M, Antognoni F, Poli F, Gentilomi GA. Ethanolic extracts of *Tinospora cordifolia* and *Alstonia scholaris* show antimicrobial activity towards clinical isolates of methicillin-resistant and carbapenemaseproducing bacteria. Nat Prod Res. 2014;28:1438–45.
- 11. Castillo AL, Osi MO, Ramos JD, et al. Efficacy and safety of *Tinospora cordifolia* lotion in *Sarcoptes scabiei* var. *hominis*-infected pediatric patients: a single blind, randomized controlled trial. J Pharmacol Pharma-cother. 2013;4:39–46.
- Chaudhary R, Jahan S, Goyal PK. Chemopreventive potential of an Indian medicinal plant (*Tinospora cordifolia*) on skin carcinogenesis in mice. J Environ Pathol Toxicol Oncol. 2008;27:233–43.
- Chintalwar G, Jain A, Sipahimalani A, et al. An immunologically active arabinogalactan from *Tinospora cordifolia*. Phytochemistry. 1999;52: 1089–93.
- 14. Chougale AD, Ghadyale VA, Panaskar SN, Arvindekar AU. Alpha glucosidase inhibition by stem extract of *Tinospora cordifolia*. J Enzyme Inhib Med Chem. 2009;24:998–1001.
- 15. Dhingra D, Goyal PK. Evidences for the involvement of monoaminergic and GABAergic systems in antidepressant-like activity of *Tinospora cordifolia* in mice. Indian J Pharm Sci. 2008;70:761–7.
- Gangan VD, Pradhan P, Sipahimalani AT, Banerji A. Cordifolisides A, B, C: norditerpene furan glycosides from *Tinospora cordifolia*. Phytochemistry. 1994;37:781–6.
- 17. Goel B, Pathak N, Nim DK, et al. Clinical evaluation of analgesic activity of guduchi (*Tinospora cordifolia*) using animal model. J Clin Diagn Res. 2014;8:HC01–4.
- 18. Goel HC, Prasad J, Singh S, et al. Radioprotective potential of an herbal extract of *Tinospora cordifolia*. J Radiat Res (Tokyo). 2004;45:61–8.
- Grover JK, Rathi SS, Vats V. Amelioration of experimental diabetic neuropathy and gastropathy in rats following oral administration of plant (*Eugenia jambolana*, *Mucuna pruriens* and *Tinospora cordifolia*) extracts. Indian J Exp Biol. 2002;40:273–6.
- Grover JK, Vats V, Rathi SS, Dawar R. Traditional Indian antidiabetic plants attenuate progression of renal damage in streptozotocin induced diabetic mice. J Ethnopharmacol. 2001;76:233–8.
- Grover JK, Vats V, Rathi SS. Antihyperglycemic effect of *Eugenia jambolana* and *Tinospora cordifolia* in experimental diabetes and their effects on key metabolic enzymes involved in carbohydrate metabolism. J Ethnopharmacol. 2000;73:461–70.
- 22. Guo R, Pittler MH, Ernst E. Herbal medicines for the treatment of allergic rhinitis: a systematic review. Ann Allergy Asthma Immunol. 2007;99:483–95.

- 23. Gupta RS, Sharma A: Antifertility effect of *Tinospora cordifolia* (Willd.) stem extract in male rats. Indian J Exp Biol. 2003;41:885–9.
- 24. Hemadri K, Rao SS. Jaundice: tribal medicine. Ancient Sci Life. 1984;3: 209–12.
- Hussain L, Akash MS, Ain NU, Rehman K, Ibrahim M. The analgesic, anti-Inflammatory and antipyretic activities of *Tinospora cordifolia*. Adv Clin Exp Med. 2015;24:957–64.
- Ikram M, Khattak SG, Gilani SN. Antipyretic studies on some indigenous Pakistani medicinal plants: II. J Ethnopharmacol. 1987;19:185–92.
- Jagetia GC, Nayak V, Vidyasagar MS. Evaluation of the antineoplastic activity of guduchi (*Tinospora cordifolia*) in cultured HeLa cells. Cancer Lett. 1998;127:71–82.
- Jagetia GC, Rao SK. Evaluation of cytotoxic effects of dichloromethane extract of guduchi (*Tinospora cordifolia* Miers ex Hook. F. & Thoms.) on cultured HeLa cells. Evid Based Complement Alternat Med. 2006;3:267–72.
- Jagetia GC, Rao SK. Evaluation of the antineoplastic activity of guduchi (*Tinospora cordifolia*) in Ehrlich ascites carcinoma bearing mice. Biol Pharm Bull. 2006;29:460–6.
- Jahfar M. Glycosyl composition of polysaccharide from *Tinospora cordifolia*. Acta Pharm. 2003;53:65–9.
- Jain BN, Jain VK, Shete A. Antipsychotic activity of aqueous ethanolic extract of *Tinospora cordifolia* in amphetamine challenged mice model. J Adv Pharm Technol Res. 2010;1:30–3.
- Jayaprakash R, Ramesh V, Sridhar MP, Sasikala C. Antioxidant activity of ethanolic extract of *Tinospora cordifolia* on N-nitrosodiethylamine (diethylnitrosamine) induced liver cancer in male Wister albino rats. J Pharm Bioallied Sci. 2015;7 Suppl 1:S40–5.
- Jeyachandran R, Xavier TF, Anand SP. Antibacterial activity of stem extracts of *Tinospora cordifolia* (Willd.) Hook. f & Thomson. Anc Sci Life. 2003;23:40–3.
- Kalekar SA, Munshi RP, Bhalerao SS, Thatte UM. Insulin sensitizing effect of 3 Indian medicinal plants: an *in vitro* study. Indian J Pharmacol. 2013; 45:30–3.
- Kalikar MV, Thawani VR, Varadpande UK, et al. Immunomodulatory effect of *Tinospora cordifolia* extract in human immunodeficiency virus positive patients. Indian J Pharmacol. 2008;40:107–10.
- Kapil A, Sharma S. Immunopotentiating compounds from *Tinospora* cordifolia. J Ethnopharmacol. 1997;58:89–95.
- Kar A, Choudhary BK, Bandyopadhyay NG. Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. J Ethnopharmacol. 2003;84:105–8.
- Kaur M, Singh A, Kumar B. Comparative antidiarrheal and antiulcer effect of the aqueous and ethanolic stem bark extracts of *Tinospora cordifolia* in rats. J Adv Pharm Technol Res. 2014;5:122–8.

- Khan V, Najmi AK, Akhtar M, et al. A pharmacological appraisal of medicinal plants with antidiabetic potential. J Pharm Bioallied Sci. 2012;4: 27–42.
- 40. Koppada R, Norozian FM, Torbati D, et al. Physiological effects of a novel immune stimulator drug, (1,4)-alpha-d-glucan, in rats. Basic Clin Pharmacol Toxicol. 2009;105:217–21.
- Kosaraju J, Chinni S, Roy PD, et al. Neuroprotective effect of *Tinospora* cordifolia ethanol extract on 6-hydroxydopamine induced Parkinsonism. Indian J Pharmacol. 2014;46:176–80.
- 42. Leyon PV, Kuttan G. Effect of *Tinospora cordifolia* on the cytokine profile of angiogenesis-induced animals. Int Immunopharmacol. 2004;4:1569–75.
- 43. Leyon PV, Kuttan G. Inhibitory effect of a polysaccharide from *Tinospora cordifolia* on experimental metastasis. J Ethnopharmacol. 2004;90:233–7.
- Manjrekar PN, Jolly CI, Narayanan S. Comparative studies of the immunomodulatory activity of *Tinospora cordifolia* and *Tinospora sinensis*. Fitoterapia. 2000;71:254–7.
- Mathew S, Kuttan G. Antioxidant activity of *Tinospora cordifolia* and its usefulness in the amelioration of cyclophosphamide induced toxicity. J Exp Clin Cancer Res. 1997;16:407–11.
- Maurya R, Manhas LR, Gupta P, et al. Amritosides A, B, C and D: clerodane furano diterpene glucosides from *Tinospora cordifolia*. Phytochemistry. 2004;65:2051–5.
- Mishra A, Kumar S, Pandey AK. Scientific validation of the medicinal efficacy of *Tinospora cordifolia*. ScientificWorldJournal. 2013;2013: 292934.
- 48. Modak M, Dixit P, Londhe J, et al. Indian herbs and herbal drugs used for the treatment of diabetes. J Clin Biochem Nutr. 2007;40:163–73.
- 49. Nadig PD, Revankar RR, Dethe SM, et al. Effect of *Tinospora cordifolia* on experimental diabetic neuropathy. Indian J Pharmacol. 2012;44:580–3.
- 50. Nair PK, Melnick SJ, Ramachandran R, et al. Mechanism of macrophage activation by (1,4)-alpha-D-glucan isolated from *Tinospora cordifolia*. Int Immunopharmacol. 2006;6:1815–24.
- 51. Narkhede AN, Jagtap SD, Kasote DM, Kulkarni OP, Harsulkar AM. Comparative immunomodulation potential of *Tinospora cordifolia* (Willd.) Miers ex Hook. F., *Tinospora sinensis* (Lour.) Merrill and *Tinospora cordifolia* growing on *Azadirachta indica* A. Juss. Indian J Exp Biol. 2014;52:808–13.
- 52. Pahadiya S, Sharma J. Alteration of lethal effects of gamma rays in Swiss albino mice by *Tinospora cordifolia*. Phytother Res. 2003;17:552–4.
- 53. Pan L, Terrazas C, Lezama-Davila CM, et al. Cordifolide A, a sulfurcontaining clerodane diterpene glycoside from *Tinospora cordifolia*. Org Lett. 2012;14:2118–21.
- Panchabhai TS, Ambarkhane SV, Joshi AS, et al. Protective effect of *Tinospora cordifolia*, *Phyllanthus emblica* and their combination against antitubercular drugs induced hepatic damage: an experimental study. Phytother Res. 2008;22:646–50.

- 55. Patel MB, Mishra S. Hypoglycemic activity of alkaloidal fraction of *Tinospora cordifolia*. Phytomedicine. 2011;18:1045–52.
- 56. Patel MB, Mishra S. Isoquinoline alkaloids from *Tinospora cordifolia* inhibit rat lens aldose reductase. Phytother Res. 2012;26:1342–7.
- Patgiri B, Umretia BL, Vaishnav PU, et al. Anti-inflammatory activity of Guduchi Ghana (aqueous extract of *Tinospora cordifolia* Miers.). Ayu. 2014;35:108–10.
- Patil RN, Patil RY, Ahirwar B, Ahirwar D. Evaluation of antidiabetic and related actions of some Indian medicinal plants in diabetic rats. Asian Pac J Trop Med. 2011;4:20–3.
- Prince PS, Kamalakkannan N, Menon VP. Restoration of antioxidants by ethanolic *Tinospora cordifolia* in alloxan-induced diabetic Wistar rats. Acta Pol Pharm. 2004;61:283–7.
- 60. Prince PS, Menon VP, Gunasekaran G. Hypolipidaemic action of *Tinospora cordifolia* roots in alloxan diabetic rats. J Ethnopharmacol. 1999;64:53–7.
- 61. Prince PS, Menon VP. Hypoglycaemic and hypolipidaemic action of alcohol extract of *Tinospora cordifolia* roots in chemical induced diabetes in rats. Phytother Res. 2003;17:410–3.
- 62. Prince PS, Padmanabhan M, Menon VP. Restoration of antioxidant defence by ethanolic *Tinospora cordifolia* root extract in alloxan-induced diabetic liver and kidney. Phytother Res. 2004;18:785–7.
- 63. Rao PR, Kumar VK, Viswanath RK, Subbaraju GV. Cardioprotective activity of alcoholic extract of *Tinospora cordifolia* in ischemia-reperfusion induced myocardial infarction in rats. Biol Pharm Bull. 2005;28:2319–22.
- Rao SK, Rao PS, Rao BN. Preliminary investigation of the radiosensitizing activity of guduchi (*Tinospora cordifolia*) in tumor-bearing mice. Phytother Res. 2008;22:1482–9.
- Rathi SS, Grover JK, Vikrant V, Biswas NR. Prevention of experimental diabetic cataract by Indian Ayurvedic plant extracts. Phytother Res. 2002; 16:774–7.
- 66. Reddy SS, Ramatholisamma P, Karuna R, Saralakumari D. Preventive effect of *Tinospora cordifolia* against high fructose diet-induced insulin resistance and oxidative stress in male Wistar rats. Food Chem Toxicol. 2009;47:2224–9.
- 67. Reddy SS, Ramatholisamma P, Ramesh B, et al. Beneficiary effect of *Tinospora cordifolia* against high-fructose diet induced abnormalities in carbohydrate and lipid metabolism in Wistar rats. Horm Metab Res. 2009; 41:741–6.
- Rege N, Bapat RD, Koti R, et al. Immunotherapy with *Tinospora cordifolia*: a new lead in the management of obstructive jaundice. Indian J Gastroenterol. 1993;12:5–8.
- Rege NN, Nazareth HM, Bapat RD, Dahanukar SA. Modulation of immunosuppression in obstructive jaundice by *Tinospora cordifolia*. Indian J Med Res Sect A—Infect Dis. 1989;90:478–83.

- 70. Rege NN, Thatte UM, Dahanukar SA. Adaptogenic properties of six rasayana herbs used in Ayurvedic medicine. Phytother Res. 1999;13:275–91.
- Sangeetha MK, Balaji Raghavendran HR, Gayathri V, Vasanthi HR. *Tinospora cordifolia* attenuates oxidative stress and distorted carbohydrate metabolism in experimentally induced type 2 diabetes in rats. J Nat Med. 2011;65:544–50.
- Sannegowda KM, Venkatesha SH, Moudgil KD. *Tinospora cordifolia* inhibits autoimmune arthritis by regulating key immune mediators of inflammation and bone damage. Int J Immunopathol Pharmacol. 2015;28:521–31.
- Sarala M, Velu V, Anandharamakrishnan C, Singh RP. Spray drying of *Tinospora cordifolia* leaf and stem extract and evaluation of antioxidant activity. J Food Sci Technol. 2012;49:119–22.
- 74. Sarma DN, Khosa RL, Sahai M. Isolation of jatrorrhizine from *Tinospora cordifolia* roots. Planta Med. 1995;61:98–9.
- Sengupta M, Sharma GD, Chakraborty B. Effect of aqueous extract of *Tinospora cordifolia* on functions of peritoneal macrophages isolated from CCl4 intoxicated male albino mice. BMC Complement Altern Med. 2011; 11:102.
- Sengupta S, Mukherjee A, Goswami R, Basu S. Hypoglycemic activity of the antioxidant saponarin, characterized as alpha-glucosidase inhibitor present in *Tinospora cordifolia*. J Enzyme Inhib Med Chem. 2009;24: 684–90.
- Sharma AK, Kishore K, Sharma D, et al. Cardioprotective activity of alcoholic extract of *Tinospora cordifolia* (Willd.) Miers in calcium chlorideinduced cardiac arrhythmia in rats. J Biomed Res. 2011;25:280–6.
- Sharma B, Dabur R. Protective effects of *Tinospora cordifolia* on hepatic and gastrointestinal toxicity induced by chronic and moderate alcoholism. Alcohol Alcohol. 2016;51:1–10.
- Sharma R, Amin H, Prajapati PK. Seasonal variations in physicochemical profiles of Guduchi Satva (starchy substance from *Tinospora cordifolia* [Willd.] Miers). J Ayurveda Integr Med. 2013;4:193–7.
- Sharma U, Bala M, Kumar N, et al. Immunomodulatory active compounds from *Tinospora cordifolia*. J Ethnopharmacol. 2012;141:918–26.
- Sharma U, Bala M, Saini R, et al. Polysaccharide enriched immunomodulatory fractions from *Tinospora cordifolia* (Willd.) miers ax hook. f. & Thoms. Indian J Exp Biol. 2012;50:612–7.
- 82. Sharma V, Pandey D. Beneficial effects of *Tinospora cordifolia* on blood profiles in male mice exposed to lead. Toxicol Int. 2010;17:8–11.
- Sharma V, Pandey D. Protective role of *Tinospora cordifolia* against lead-induced hepatotoxicity. Toxicol Int. 2010;17:12–7.
- Sing H, Bisht GS. Some novel folk treatments among the tribes of Uttar Pradesh. Anc Sci Life. 1999;18:250–3.
- Singh L, Tyagi S, Rizvi MA, Goel HC. Effect of *Tinospora cordifolia* on gamma ray-induced perturbations in macrophages and splenocytes. J Radiat Res (Tokyo). 2007;48:305–15.

- Singh N, Singh SM, Shrivastava P. Immunomodulatory and antitumor actions of medicinal plant *Tinospora cordifolia* are mediated through activation of tumor-associated macrophages. Immunopharmacol Immunotoxicol. 2004;26:145–62.
- Singh RK. *Tinospora cordifolia* as an adjuvant drug in the treatment of hyperreactive malarious splenomegaly—case reports. J Vector Borne Dis. 2005;42:36–8.
- 88. Sivakumar V, Rajan MS. Antioxidant effect of *Tinospora cordifolia* extract in alloxan-induced diabetic rats. Indian J Pharm Sci. 2010;72:795–8.
- Spelman K, Burns J, Nichols D, et al. Modulation of cytokine expression by traditional medicines: a review of herbal immunomodulators. Altern Med Rev. 2006;11:128–50.
- Stanely P, Prince M, Menon VP. Hypoglycaemic and other related actions of *Tinospora cordifolia* roots in alloxan-induced diabetic rats. J Ethnopharmacol. 2000;70:9–15.
- Sudhakaran DS, Srirekha P, Devasree LD, et al. Immunostimulatory effect of *Tinospora cordifolia* Miers leaf extract in Oreochromis mossambicus. Indian J Exp Biol. 2006;44:726–32.
- Swaminathan K, Sinha UC, Bhatt RK, et al. Structure of tinosporide, a diterpenoid furanolactone from *Tinospora cordifolia* Miers. Acta Crystallographica—Section C—Crystal Struct Commun. 1989;45:134–6.
- Swaminathan K, Sinha UC, Ramakumar S, et al. Structure of columbin, a diterpenoid furanolactone from *Tinospora cordifolia* Miers. Acta Crystallographica—Section C—Crystal Struct Commun. 1989;45:300–3.
- Tambekar DH, Khante BS, Chandak BR, et al. Screening of antibacterial potentials of some medicinal plants from Melghat forest in India. Afr J Tradit Complement Altern Med. 2009;6:228–32.
- Thatte UM, Dahanukar SA. Comparative study of immunomodulating activity of Indian medicinal plants, lithium carbonate and glucan. Methods Findings Exp Clin Pharmacol. 1988;10:639–44.
- Thatte UM, Rao SG, Dahanukar SA. *Tinospora cordifolia* induces colony stimulating activity in serum. J Postgrad Med. 1994;40:202–3.
- Thippeswamy G, Sheela ML, Salimath BP. Octacosanol isolated from *Tinospora cordifolia* downregulates VEGF gene expression by inhibiting nuclear translocation of NF-κB and its DNA binding activity. Eur J Pharmacol. 2008;588:141–50.
- Tiwari M, Dwivedi UN, Kakkar P. *Tinospora cordifolia* extract modulates COX-2, iNOS, ICAM-1, proinflammatory cytokines and redox status in murine model of asthma. J Ethnopharmacol. 2014;153:326–37.
- Upadhyay AK, Kumar K, Kumar A, Mishra HS. *Tinospora cordifolia* (Willd.) Hook. f. and Thoms. (Guduchi)-validation of the Ayurvedic pharmacology through experimental and clinical studies. Int J Ayurveda Res. 2012;1: 112–21.

- Van Kiem P, Van Minh C, Dat NT, et al. Aporphine alkaloids, clerodane diterpenes, and other constituents from *Tinospora cordifolia*. Fitoterapia. 2010;81:485–9.
- 101. Velazquez EA, Kimura D, Torbati D, et al. Immunological response to (1,4)-alpha-D-glucan in the lung and spleen of endotoxin-stimulated juvenile rats. Basic Clin Pharmacol Toxicol. 2009;105:301–6.
- Vinutha B, Prashanth D, Salma K, et al. Screening of selected Indian medicinal plants for acetylcholinesterase inhibitory activity. J Ethnopharmacol. 2007;109:359–63.
- Wadood N, Wadood A, Shah SA. Effect of *Tinospora cordifolia* on blood glucose and total lipid levels of normal and alloxan-diabetic rabbits. Planta Med. 1992;58:131–6.